Supporting information

Simultaneous Upcycling of PET Plastic Waste and CO₂ Reduction through

Co-electrolysis: A Novel Approach Integrating CO₂ Reduction and PET

Hydrolysate Oxidation

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Characterization

Scanning electron microscopy (SEM) images, energy-dispersive X-ray spectroscopy analysis (EDX) data and EDX mapping images were obtained using a JEOL JSM-7800F field-emission scanning electron microscope (SEM) operated at 15 kV. X-ray diffraction (XRD) patterns were collected on a high flux Rigaku Smartlab rotated anode, working with a copper Ka radiation (1.5418 Å) at an applied voltage of 45 kV and an anode current of 200 mA in the 2θ range of 5– 90°. Raman measurements were performed on a LabRam HR Micro-Raman system (Horiba Jobin Yvon) using a 473 nm laser diode as excitation source. The chemical composition of all materials was analyzed by X-ray photoelectron spectroscopy (XPS) using an ESCALAB 220 XL spectrometer from Vacuum generators featuring a monochromatic Al Ka X-ray source at 1486.6 eV. Ultraviolet-visible (UV-vis) spectra were recorded by using Safas Bio-UVmc² spectrophotometer using a quartz cell (1 cm path length) and tungsten-halogen source. Absorption spectra were recorded from 200 to 800 nm at room temperature. Fourier-transform infrared spectroscopy (FTIR) analysis was performed on a Thermo Fisher Scientific (Nicolet 8700) instrument equipped with IR solution software. FTIR spectra were recorded at 6 cm⁻¹ spectral resolution in the frequency range of 700–4000 cm⁻¹ at room temperature.

The ¹H and ¹³C NMR spectra were collected on a Bruker 300 MHz. The samples for NMR analysis were prepared by mixing 550 μ L of electrolyte with 50 μ L of D₂O. Water suppression method was used in all ¹H NMR experiments.

Mechanism of formation of Bi₂O₂CO₃ (BOC) nanosheets

When BiPO₄@rGO electrode was immersed in a 0.5M KHCO₃ solution, no morphological changes were observed. However, when a potential of -0.8 V vs. RHE was applied for 1 h, *in situ*

transformation was seen. The following series of chemical reactions (1-5) could be responsible for the formation of BOC nanosheets.

$$CO_2 + H_2O + 2e^- \rightarrow HCOO^- + OH^-$$
 (1)

$$CO_2 + H_2O + 2e^- \rightarrow CO + 2OH^-$$
 (2)

$$2H_20 + 2e^- \to H_2 + 20H^-$$
(3)

$$Bi^{3+} + 30H^{-} \rightarrow Bi(0H)_{3} \tag{4}$$

$$2Bi(OH)_3 + CO_2 \rightarrow Bi_2O_2CO_3 + 3H_2O$$
 (5)



Figure S1. Color change from dark brown to black before (left) and after (right) hydrothermal reaction at 160 °C for 16h.



Figure S2. X-ray diffraction (XRD) pattern of the as-prepared BiPO₄@rGO product.



Figure S3. XRD patterns of electrochemical activation of BiPO₄@rGO at different applied

potentials.



Figure S4. FESEM image and corresponding EDX mapping of Bi, P and O of BiPO₄@rGO.



Figure S5. FESEM image and corresponding EDX mapping of C, O and Bi of BOC@rGO electrode.



Figure S6. (a) Raman spectra of BOC@rGO and BiPO₄@rGO. (b) I_d/I_g band ratio determined from the Raman spectra of BiPO₄@rGO and GO.



Figure S7. UV-visible absorption spectra of BOC@rGO and GO.



Figure S8. High resolution XPS spectra of the P 2p of BOC@rGO (black) and BiPO₄@rGO (red).



Figure S9. FTIR spectra of BOC@rGO and BiPO₄@rGO.



Figure S10. Chronoamperometry analysis at different potentials in CO₂-saturated 0.1 M KHCO₃ solution of (**a**) BOC@rGO and (**b**) BOC.



Figure S11. Calibration curve for formate concentration determined by ¹H NMR for CO₂RR.



Figure S12. Partial current densities for HCOOH production of BOC@rGO and BOC.



Figure S13. (a) CV curves of BOC@rGO and (b) BOC performed at different scan rates in Arsaturated 0.1 M KHCO₃ aqueous solution in non-faradaic region; (c) C_{dl} curves of BOC@rGO and BOC.



Figure S14. Bi 4f XPS spectra and XRD patterns of BOC@rGO after stability test. The spectra of as-prepared samples are shown for comparison.



Figure S15. Tafel plots of BOC@rGO and BOC.



Figure S16. HR-TEM images of CuCoO@rGO nanocomposite.



Figure S17. XPS survey spectrum of CuCoO@rGO nanocomposite.



Figure S18. Calibration curves for (a) formate and (b) ethylene glycol (EG) concentrations determined by ¹H NMR for PET hydrolysate.



Figure S19. LSV curves of CuCoO@rGO, Cu@rGO and Co@rGO in 1 M KOH solution; scan rate =10 mV s⁻¹.



Figure S20. LSV curves of CuCoO@rGO in 1 M KOH, 0.1 M TPA and 0.1 M EG; scan rate =10 mV s⁻¹.



Figure S21. (a) LSV curves of CuCoO@rGO at varying concentrations of EG, scan rate =10 mV s⁻¹. (b) Linear curve of catalytic current for EG oxidation *vs*. EG concentration at 1.5 V *vs*.

RHE.



Figure S22. Controlled potential electrolysis of PET hydrolysate solution using CuCoO@rGO.



Figure S23. ¹H NMR before and after electrolysis of PET hydrolysate solution using CuCoO@rGO nanocomposite.



Figure S24. ¹³C NMR before, during and after electrolysis of PET hydrolysate solution using CuCoO@rGO.



Figure S25. Long-term electrolysis upon subsequent addition of 20 mM ethylene glycol (EG) at different intervals of time.



Figure S26. (a) Cu 2p and (b) Co 2p XPS spectra after electrolysis of PET hydrolysate solution using CuCoO@rGO nanocomposite.



Figure S27. (a) LSV curves of CuCoO@rGO(+) || BOC@rGO(-) at varying concentrations of EG. (b) Linear curve of catalytic current for formate synthesis *vs.* EG concentration at 1.9 V cell voltage.



Figure S28. CPE analysis at various potentials in a 2-electrode system.

 Table S1. Concentration of different elements in the electrolyte obtained from ICP-MS analysis

	Before electrolysis (mg/L)	After electrolysis (mg/L)
Bismuth (Bi)	0.16	0.96
Copper (Cu)	0.18	0.66
Cobalt (Co)	<0.005	<0.005

during the long-term stability tests.