Supplementary Materials for:

A Nucleotide-Copper (II) Complex Possessing Monooxygenase-Like Catalytic Function

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Table 1. Materials and regents

Materials	Abbreviation	CAS	Company
Adenosine monophosphate	AMP	4578-31-8	
Cytidine monophosphate	CMP	6757-06-8	Aladdia
Guanosine monophosphate	GMP	5550-12-9	
Uridine monophosphate	UMP	2287-36-8	
Adenosine		58-61-7	
Cytidine		65-46-3	
Guanosine		118-00-3	
Uridine		58-96-8	
Adenosine diphosphate	ADP	16178-48-6	
Cytidine diphosphate	CDP	34393-59-4	
Guanosine diphosphate	GDP	7415-69-2	
Uridine diphosphate	UDP	21931-53-3	Aladdin
Adenosine triphosphate	ATP	987-65-5	
Cytidine triphosphate	CTP	36051-68-0	
Guanosine triphosphate	GTP	56001-37-7	
Uridine triphosphate	UTP	108321-53-5	
Tyramine hydrochloride		60-19-5	
L-Tyrosine hydrochloride		16870-43-2	
Pyrene		129-00-0	
Copper sulfate pentahydrate	CuSO4·5H2O	7758-99-8	
Hydrogen peroxide	H_2O_2	7722-84-1	
Tyrosinase from mushroom	Tyrosinase	9002-10-2	
ААААААААААААААААА	poly A20		Hippobio

222222222222222222222222222222222222222	poly C20	
GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	poly G20	
ттттттттттттттттт	poly T20	

Water was deionized using a Milli-Q system (\geq 18.25 MQ·cm–1).

Figure S1 UV-vis spectra of tyramine oxidation reaction catalysed by Cu^{2+} and nucleotides. $[Cu^{2+}] = 0.1 \text{ mM}, \text{[nucleotides]} = 10 \text{ mM}, \text{[tyramine]} = 1 \text{ mM}, \text{[H}_2O_2\text{]} = 5 \text{ mM}, \text{ in water}$ (pH=8.0).



Figure S2 Structure of nucleoside monophosphate.



Adenosine monophosphate

Cytodine monophosphate

Guanosine monophosphate

Uridine monophosphate

Figure S3 Ligands' concentration effect on velocity of tyramine's oxidation reaction catalysed by Cu^{2+} and A/C/G/UMP. Cu^{2+} (0.1 mM), H₂O₂ (5mM), tyramine (1mM). The data are presented as the mean ± s.d., with the error bars representing the s.d. and N= 3.



Figure S4 KCl effect on tyramine's oxidation reaction catalysed by Cu^{2+} and AMP. AMP (20 mM), Cu^{2+} (50 μ M), H_2O_2 (5 mM), tyramine (1 mM). The data are presented as the mean \pm s.d., with the error bars representing the s.d. and N= 3.



Figure S5 6 K CW-EPR of Cu²⁺/AMP and Cu²⁺/AMP/H₂O₂/tyramine. CuSO4 (500 μ M), AMP (10 mM), H₂O₂ (1 mM), tyramine (1 mM), mwFreq (9.74GHz).



Figure S6 (A) Methanol concentration effect on the catalytic tyramine oxidation reaction. (B) EPR measurement of the system of Cu^{2+/} CMP/ H₂O₂. 60% MeOH, 100 mM DMPO, 20 mM CMP, 100 μ M Cu²⁺, 100 μ M H₂O₂. The data are presented as the mean ± s.d., with the error bars representing the s.d. and N= 3.



Figure S7 EPR measurement of the system of Cu^{2+/} GMP/ H_2O_2 . 60% MeOH, 100 mM DMPO, 20 mM GMP, 100 μ M Cu²⁺, 100 μ M H_2O_2 .



Figure S8 EPR measurement of the system of Cu^{2+/} UMP/ H₂O₂. 60% MeOH, 100 mM DMPO, 20 mM UMP, 100 μ M Cu²⁺, 100 μ M H₂O₂.



Figure S9 Free radical quenchereffect on tyramine's oxidation reaction catalysed by Cu^{2+}/AMP . Cu^{2+} (0.1mM), AMP (20 mM), H₂O₂ (5 mM), tyramine (1 mM). TBA stands for tertbutanol, while SOD stands for superoxidase dismutase. The data are presented as the mean \pm s.d., with the error bars representing the s.d. and N= 3.



Figure S10 Density functional theory model of the ternary complex intermediate of nucleotides/ $Cu^{2+}/tyramine/H_2O_2$, and the Isosurface map of IRI analyse, revealing both chemical bonds and weak interactions. IRI=1.2.



Figure S11 Free energy during the formation of HO₂• radical suggested by DFT calculations. Free energies and enthalpies were calculated at 298 K. The inserted figures were structure optimization and spin density computation during the formation of HO₂• radical suggested by DFT calculations.



Figure S12 Active energy measurements of A/C/G/UMP and copper's catalytic reaction. [nucleotide] = 20 mM, $[Cu^{2+}] = 50\mu M$, H_2O_2 (1 mM), tyramine (1 mM).



Figure S13 Deaeration effect on tyramine's oxidation reaction catalysed by Cu^{2+} and AMP. AMP (20 mM), Cu^{2+} (50 μ M), H_2O_2 (5 mM), tyramine (1 mM). Insert was the initial reaction rate. The data were presented as the mean \pm s.d., with the error bars representing the s.d. and N=3.





Figure S14 Proposal hydroxyl radical intermediate route.

Figure S15 Comparison of DNA's and nucleotide's catalytic reactivity with Cu^{2+} . Cu^{2+} (50 Mm), H₂O₂ (5 mM), tyrosine (1 mM), nucleobase in nucleotides and ssDNA (20 mM). The data are presented as the mean ± s.d., with the error bars representing the s.d. and N= 3.



Figure S16 AMP concentration effect on the catalytic oxidation of tyramine. Cu^{2+} (50 μ M), H_2O_2 (5 mM), tyramine (1 mM). The data are presented as the mean \pm s.d., with the error bars representing the s.d. and N= 3.



Figure S17 Oxidation of o-tert-butylphenol (A), dopamine (B) and tyrosine (C) catalysed by AMP/Cu²⁺. AMP (2 mM), Cu²⁺ (100 μ M), H₂O₂ (5 mM), tyrosine (1 mM), time interval (5 min).



Figure S18 Temperature effect on oxidation of tyramine catalysed by AMP/Cu²⁺, polyA20/Cu²⁺ and tyrosinase. AMP (20 mM), A20 (1 μ M), Cu²⁺ (50 μ M), H₂O₂ (1 mM), tyrosinase (10 μ M), tyramine (1 mM). The data are presented as the mean ± s.d., with the error bars representing the s.d. and N = 3.



Figure S19 Temperature effect on oxidation of tyramine catalysed by AMP/Cu²⁺, GMP/Cu²⁺, GMP/Cu²⁺, polyG20 (1 μ M), GMP (20 mM), Cu²⁺ (50 μ M), H₂O₂ (1 mM), tyramine (1 mM). The data are presented as the mean \pm s.d., with the error bars representing the s.d. and N = 3.



Figure S20 Oxidation of tyramine catalysed by Cu^{2+} and AMP/ Cu^{2+} at different temperatures. [AMP] = 20 mM, [Cu^{2+}] = 50 μ M, [tyramine] = 1 mM, [H_2O_2] = 1 mM.

