Electronic Supplementary Information

Fe(III) incorporated porphyrin-based conjugated organic polymer as peroxidase-mimic for sensitive determination of glucose and H₂O₂

Samanka Narayan Bhaduri, Debojit Ghosh, Sauvik Chatterjee, Rima Biswas, Asim Bhaumik,* Papu Biswas*



Fig. S1 Survey spectra of Fe-DMP-POR



Fig. S2 (a) PXRD and (b) FT-IR of Fe-DMP-POR after treatment with 3 m NaOH, 3 M HCl and boiling water



Fig. S3 (a) Optimisation of pH and (b) optimisation of temperature for peroxidase like activity

| Catalyst | Substrate | K _m | V _{max} | Reference |
|--|-----------|----------------|--------------------------------------|-----------|
| | | (mM) | (10 ⁻⁸ Ms ⁻¹) | |
| Fe-DMP-POR | TMB | 0.14 | 1.44 | This work |
| | H_2O_2 | 16.9 | 4.50 | |
| HRP | TMB | 100 | 5.71 | 1 |
| | H_2O_2 | 142 | 1.33 | |
| Au/OMC | TMB | 0.015 | 1.50 | 2 |
| | H_2O_2 | 38.11 | 4.00 | |
| CePO ₄ -CeO ₂ composite nanorods | TMB | 0.236 | 8.78 | 3 |
| | H_2O_2 | 4.76 | 29.79 | |
| Bi ₂ Te ₃ -Au ₅ | TMB | 0.261 | 2.15 | 4 |
| | H_2O_2 | 11.36 | 4.48 | |

Table S1. Comparison of kinetic parameters like K_{m} and V_{max} with other reported materials

Table S2. Comparisons of different glucose sensing parameters

| Catalyst | Linear range (mM) | LOD (µM) | Reference |
|--|-------------------|----------|-----------|
| Fe-DMP-POR | 0-0.150 | 4.84 | This work |
| PVP-MoS ₂ | 1 10 | 220 | 5 |
| nanoparticles | 1-10 | 320 | 5 |
| NiCo ₂ S ₄ | 0.02.1 | 6.24 | 6 |
| Microflowers | 0.02-1 | 0.24 | 0 |
| MoS ₂ nanoflakes | 0.1-1 | 33.51 | 7. |
| Bi ₂ Te ₃ -Au _{0.5} | 0-4.5 | 380 | 4 |
| CePO ₄ -CeO ₂ | 0-0.1 | 4.1 | 3 |
| Core-shell Cu/Au | 0.02.0.670 | 15 | 0 |
| NPs | 0.02-0.070 | 15 | 8 |
| Fe ₃ O ₄ @Au–Pt | 0.00005-0.140 | 0.025 | 9 |



Fig. S4 UV spectra of solutions containing different monosaccharides and glucose

Table S3. Detection of glucose in blood serum samples

| Sample No. | Spiked glucose/mM | Recovered glucose/mM | RSD/% | Recovery/% |
|------------|-------------------|----------------------|-------|------------|
| 1 | 0.05 | 0.051 | 3.1 | 102 |
| 2 | 0.075 | 0.074 | 2.5 | 98 |
| 3 | 0.1 | 0.103 | 2.8 | 103 |
| 4 | 0.15 | 0.16 | 3 | 106 |



Fig. S5 Optimisation of NaOH concentration for H₂O₂ detection

| Modified | Linear range | Sensitivity | Limit of | Reference |
|-------------------------------------|--------------|--|-----------------------|-----------|
| electrodes | (mM) | $(\mu A \text{ cm}^{-2} \text{ m} M^{-1})$ | detection (μM) | |
| Pd–TiO ₂ | 1-20 | 550 | 23 | 10 |
| Ag_{dahlia} | upto 15 mM | 330 | 2.2 | 11 |
| CuS/RGO | 0.001-1 | 7.96 | 0.3 | 12 |
| α-Fe ₂ O ₃ | 0.050-1.340 | 24 | 5 | 13 |
| Ag-MnO ₂ - MWCNTs/CGE | upto 10.4 mM | 82.5 | 1.7 | 14 |
| RGO/Fe ₃ O ₄ | upto 6 mM | 688 | 3.2 | 15 |
| Fe-DMP-POR | 0.005-2 | 947.67 | 3.16 | This work |

 Table S4. Comparison of different parameters for amperometric peroxide detection with other materials.



Fig. S6 (a) Repeatability study, (b) Fabrication reproducibility (five electrode), (c) long term stability study of Fe-DMP-POR in 0.1 M NaOH and H_2O_2 and (d) selectivity study of Fe-DMP-POR for uric acid glucose, cysteine and ascorbic acid at -0.78 V in 0.1 M NaOH

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