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## **Electronic Supplementary Information**

## Functional material-mediated wireless physical stimulation for neuro-modulation and regeneration

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| Stimulation type | Biomaterial                     | Feature  | Stimulation parameter  | Cell/animal response  | Ref. |
|------------------|---------------------------------|--|--|---|------|
|                  | PVDF<br>BZT-BCT/PVDF<br>KNN/PLA | A nanopillar array fabricated by hot-<br>pressing  | US, 5 min, 2 times a day, 300-<br>500 W  | Neuron-like differentiation<br>of rBMSCs  | 1    |
|                  |                                 | Thin films fabricated by<br>electrospinning of the composite<br>containing PVDF and BZT-BCT<br>nanowires | US, 1 MHz, pulse width 5 ms,<br>pulse interval 100 ms, acoustic<br>pressure 150 kPa  | The generation of CMAPs and the twitching of limbs  | 2    |
| Piezoelectric    |                                 | A biodegradable 3D multichannel<br>nanofiber scaffold  | US:<br>(1) <i>In vitro</i> : 20 min, 1 MHz, 0-<br>150 kPa;<br>(2) <i>In vivo</i> : 20 min every 2<br>days, lasted for 4 weeks  | <ol> <li>Neuronal differentiation<br/>of NSCs;</li> <li>Neuro-regeneration and<br/>functional recovery of SCI<br/>rats</li> </ol>                         | 3    |
| stimulation      | BN                              | BN nanotubes prepared by annealing can be internalized by cells  | US, 20 W, 40 kHz, for 5 s, 4<br>times a day for 9 days   | Promoted neurite outgrowth of PC 12 cells   | 4    |
|                  | BTNPs                           | Tetragonal BTNPs   | US, 1 MHz, 0.1-0.8 W/cm <sup>2</sup>   | US induced calcium<br>transient in SH-SY5Y cells  | 5    |
|                  | BTNP-PDA-BNN6                   | US-driven release of NO, allowing the<br>BBB penetration   | US:<br>(1) <i>In vitro</i> :2.2 MHz, 10 s, 0.4<br>W/cm <sup>2</sup> ,<br>(2) <i>In vivo</i> : 1.5 MHz, 462.4<br>W/cm <sup>2</sup> , duty cycle: 10%,<br>pulse repetition frequency:<br>10 Hz, 66.4 W/cm <sup>2</sup> | <ol> <li>Increased Ca<sup>2+</sup> influx<br/>and dopamine release of SH-<br/>SY5Y cells;</li> <li>Alleviated symptoms of<br/>the mice with PD</li> </ol> | 6    |

 $\label{eq:tables} \textbf{Table S1} \text{ Overview of the functional materials for wireless physical stimulation.}$ 

| ME stimulation                | CoFe <sub>2</sub> O <sub>4</sub> @BaTiO <sub>3</sub> | Core-shell MENPs can be stereotactically injected into deep brain   | <ol> <li>(1) In vitro: 225 mT (DC MF),<br/>6 mT, 140 Hz (AC MF);</li> <li>(2) In vivo: 220 mT (DC MF),<br/>6 mT, 140 Hz (AC MF), 180 s</li> </ol>   | <ol> <li>Induced Ca<sup>2+</sup> transient f<br/>SH-SY5Y;</li> <li>Increased expression of<br/>c-Fos <i>in vivo</i> and controlled<br/>mouse behavior</li> </ol>  | 7  |
|-------------------------------|--|---|---|---|----|
|                               | FO@BTO/AHA hydrogel                                  | Core-shell MENPs were combined<br>with biomimetic hydrogel with features<br>of native neural ECM  | Pulsed MF (13 mT, 60 Hz,<br>duty cycle of 50%), 0.5 h per<br>day  | <ul> <li>(1) Promoted neurogenesis<br/>of PC 12 cells <i>in vitro</i>;</li> <li>(2) facilitated neuro-<br/>regeneration in SCI rats</li> </ul>  | 8  |
|                               | GO/CFO/PVDF  | ME nanofiber films prepared by electrospinning  | EMFs, 1 mT, 50 Hz, 8 h/day<br>for 21 days   | Neuron-like differentiation<br>of MSCs  | 9  |
|                               | Metglas/PVDF or PZT                                  | The nanogenerators were implanted<br>into the STN of rats with PD, ensuring<br>ME stimulation in freely moving rats                         | AMF, 1.5 mT, 1 min, 200 Hz,<br>biphasic pulses  | Decreased rotation rate of<br>the rats with PD  | 10 |
|                               | Metaglas/PZT   | The ME film can be implanted directly<br>and delivered through a percutaneous<br>catheter to big animals                                    | AMF, <1 mT, 3 V, 1.5 ms<br>pulse width monophasic, 1-10<br>Hz   | Induced CMAPs of rats and pigs  | 11 |
| Optoelectrical<br>stimulation | Silicon  | Nanoporous/non-porous silicon<br>heterojunctions fabricated by metal-<br>free etching   | 532 nm laser: 10 ms, 8-68<br>mW/mm <sup>2</sup> ;<br>808 nm laser:10 ms, 15-209<br>mW/mm <sup>2</sup>   | <i>In vivo</i> sciatic nerve<br>simulation induced the<br>generation of APs and leg<br>displacement of rats   | 12 |
|                               | Silicon  | Silicon structures, including<br>nanowires, membranes and meshes for<br>multiscale stimulation <i>in vitro</i> and <i>in</i><br><i>vivo</i> | <ol> <li>(1) For cells: 592 nm, 14.4<br/>mW, 237 nm spot size, 1 ms;</li> <li>(2) For brain slice: 473 nm, 1<br/>ms, 2 mW, 57 μm spot size;</li> <li>(3) <i>In vivo</i> photo-stimulation:</li> <li>473 nm, 4-5 mW, 216 μm spot<br/>size</li> </ol> | <ol> <li>(1) Induced intracellular<br/>Ca<sup>2+</sup> influx in both DRG<br/>and glial cells;</li> <li>(2) Generation of excitatory<br/>postsynaptic currents in<br/>brain slices;</li> <li>(3) Evoked small ipsilateral<br/>forelimb movements</li> </ol> | 13 |

|                           | РЗНТ   | Photovoltaic layer prepared by spin-<br>coating  | <ul> <li>(1) <i>In vitro</i>: 532 nm, 20 ms, 15 mW/mm<sup>2</sup>, 1-20 Hz;</li> <li>(2) <i>In vivo</i>: 532 nm, 10 ms, 4 mW/mm<sup>2</sup></li> </ul> | <ol> <li>Induced the generation<br/>of APs in primary neurons;</li> <li>Restored light-responses<br/>in blind retinas</li> </ol>                      | 14 |
|---------------------------|--|--|--|---|----|
|                           | P3HT/collagen or<br>collagen/hyaluronan<br>hydrogels | Hydrophilic P3HT NPs were integrated<br>into biomimetic hydrogels to create a<br>neural interface and a 3D biomimetic<br>stem cell niche | 530 nm pulsed light, 6<br>mW/cm <sup>2</sup> , 500 ms duration, 1<br>Hz, 30 min/12 h   | <ol> <li>(1) Promoted growth of<br/>primary neurons;</li> <li>(2) Enhanced neuronal<br/>differentiation of BMSCs</li> </ol>                           | 15 |
| Magetothermal stimulation | MnFe <sub>2</sub> O <sub>4</sub>                     | The MnFe <sub>2</sub> O <sub>4</sub> NPs (d=6 nm) were<br>conjugated with streptavidin for<br>specific targeting                         | RF magnetic field (40 MHz,<br>8.4 G, 30 s)   | TRPV1 opening and activation of APs in HEK 293 cells;   | 16 |
|                           | Fe <sub>3</sub> O <sub>4</sub>                       | MNPs were modified with PEG shell  | AMF, <i>f</i> =500 kHz, H <sub>o</sub> =15 kA/m  | <ol> <li>(1) Evoked correlated trains<br/>of APs in neurons <i>in vitro</i>;</li> <li>(2) Increased expression of<br/>c-Fos <i>in vivo</i></li> </ol> | 17 |
|                           | Co-Mn-Ferrite  | Core-shell MNPs  | AMF:<br>(1) <i>In vitro</i> : 412.5 kHz, 22.4<br>kA/m, 5 s;<br>(2) <i>In vivo</i> : 570 kHz, 7.5<br>kA/m, 1 min  | <ul> <li>(1) Elicited AP firing in<br/>neurons;</li> <li>(2) Increased angular<br/>velocity of movement in a<br/>mouse model</li> </ul>               | 18 |
| Photothermal stimulation  | Gold   | Gold nanostars prepared by a modified seedless and surfactant free method  | 785 nm laser, 3-15 mW/mm <sup>2</sup> ,<br>10 s,   | Inhibition of neuronal activity   | 19 |
|                           | Graphene   | A nanowire-templated 3D fuzzy graphene   | 405 nm laser, 1.2 ms pulse<br>duration, 1.45-3.02 mW, 1-20<br>Hz   | Highly controlled activation of APs in DRG neurons  | 20 |
|                           | PDA/collagen   | PDA NPs were integrated with<br>collagen to prepare a photothermal 3D<br>foam  | 808 nm laser, 6 mW/mm <sup>2</sup> , 10-<br>30 s duration  | Suppressed spike rate of neurons  | 21 |

|                              | Gold  | Gold nanorods   | 980 nm laser, 1 ms duration,<br>0.159-1.046 J/cm <sup>2</sup> ,  | Elicited compound nerve<br>APs of rat sciatic nerves   | 22 |
|------------------------------|---|---|--|--|----|
| Photoacoustic<br>stimulation | ZnO/epoxy @<br>graphite/epoxy   | A fiber-optoacoustic convertor with tip<br>composed of ZnO/epoxy @<br>graphite/epoxy double-layer structure<br>used for photoacoustic stimulation | 1030 nm laser, 3 ns pulse<br>width, 100 μJ, 3.6 kHz, 200<br>ms   | <ul> <li>(1) Induced Ca<sup>2+</sup> transient<br/>in primary neurons;</li> <li>(2) Induced neural<br/>activation in mouse brain</li> </ul>  | 23 |
|                              | CNT/PDMS  | CNT/PDMS act as the light absorption/thermal expansion layer  | 1030 nm laser, 3 ns pulse<br>width, 11.4 mW, 1.7 kHz, 1<br>and 50 ms duration  | Activation of a single<br>neuron or subcellular<br>structures  | 24 |
|                              | Polystyrene-block-poly<br>(acryl acid)  | The photoacoustic nanotransducers can be conjugated with targeting antibodies   | 1030 nm laser, 3 ns pulse<br>width, 3.3 kHz, 3 ms, 2.1<br>mJ/cm <sup>2</sup> ( <i>in vitro</i> ), 21 mJ/cm <sup>2</sup><br>( <i>in vivo</i> )                  | <ol> <li>(1) Transient activation of<br/>neurons;</li> <li>(2) Induced local field<br/>potential in brain of mice</li> </ol>   | 25 |
|                              | PANIP-ES@AOT  | The nanovesicles were prepared by<br>HRP/H <sub>2</sub> O <sub>2</sub> -trigged polymerization on<br>AOT vesicles                                 | <ul> <li>(1) <i>In vitro</i>: 1064 nm, 40 μJ,<br/>10 ms pulse width,</li> <li>(2) <i>In vivo</i>: 1064 nm, 40 μJ, 10<br/>s pulse width, 20 Hz, 10 s</li> </ul> | <ol> <li>(1) Ca<sup>2+</sup> transient in DRG<br/>neurons;</li> <li>(2) Increased neuronal<br/>firing rates in mouse<br/>hippocampus and motor<br/>cortex and induced fast<br/>motion of mice</li> </ol> | 26 |
|                              | CNT/silk  | Photoacoustic scaffold was prepared<br>by embedding PEG-functionalized<br>CNTs into silk  | 1030 nm laser, 3 ns pulse<br>width, 1.7 kHz, 8.8-23.5 μJ, 5<br>ms duration, every 2 min<br>within total duration of 1 h  | Induced Ca <sup>2+</sup> transient,<br>promoted neurite outgrowth<br>of DRG neurons and<br>increased expression of<br>BDNF and NGF   | 27 |
| Optogenetic stimulation      | NaYF <sub>4</sub> :Yb <sup>3+</sup> /Tm <sup>3+</sup> @<br>NaYF <sub>4</sub> (UCNP)/PCL | The UCNPs were combined with collagen-modified PCL scaffold by electrospinning and electrospraying  | 980 nm laser, 800 mW/cm <sup>2</sup> ,<br>500 ms pulse width, 1 Hz, 10<br>min/12 h;  | Promoted neurite extension<br>of PC 12 cells expressing<br>CatCh   | 28 |

|                               | NaYF4:Yb/Tm@SiO2  | Silica was used to decorate UCNP to<br>optimize the biocompatibility and<br>long-term utility   | <ul> <li>980 nm laser:</li> <li>(1) <i>In vitro</i>: 0.35-8.22 W/mm<sup>2</sup>, 10-50 Hz;</li> <li>(2) <i>In vivo</i>: 15-ms pulses at 20 Hz, 700 mW peak power</li> <li>(3) <i>In vivo</i>: 15-ms pulses at 8 Hz, 3.0 W peak power, 360 mW average power</li> </ul> | <ol> <li>(1) Evoked spikes in<br/>neurons expression ChR2 <i>in</i><br/><i>vitro</i>;</li> <li>(2) Increased expression of<br/>c-Fos and release of<br/>dopamine <i>in vivo</i>;</li> <li>(3) Induced hippocampal<br/>local field potential <i>in vivo</i></li> </ol> | 29 |
|-------------------------------|---|---|---|---|----|
|                               | NaYF <sub>4</sub> :Yb/Er@ NaYF <sub>4</sub><br>(UCNP)/polypropylene<br>(PP)                                 | The UCNP was combine with PP and implanted at the spinal cord of mice   | 980 nm laser, 20 mW/mm <sup>2</sup> ,<br>0.5 and 1 Hz, 50 ms pulse<br>width,  | Induced muscular activity<br>in ChR2-transfected mice<br>and inhibited movement   | 30 |
| Magnetomechanical stimulation | Zinc doped iron oxide<br>(Zn <sub>0.4</sub> Fe <sub>2.6</sub> O <sub>4</sub> )                              | Octahedral $Zn_{0.4}Fe_{2.6}O_4$ NPs assemble<br>into m-Torquer that is used to generate<br>torque force when exposed to a<br>rotating magnetic field generated by a<br>circular magnet array (CMA) | Rotating CMA: 0.5 s pulse<br>duration, 0.5 Hz, 90 degree<br>turn  | <ol> <li>Induced Ca<sup>2+</sup> influx in<br/>neurons;</li> <li>In vivo stimulation<br/>activated mouse motility</li> </ol>  | 31 |
|                               | Fe <sub>3</sub> O <sub>4</sub>  | Magnetite nanodiscs were produced by reducing hematite to magnetite   | Magnetic field, 1-5 Hz, 7-28<br>mT, 4 pulses of 10 s at<br>intervals of 30s.  | Increased Ca <sup>2+</sup> influx in<br>HEK-293 cells expressing<br>TRPV4   | 32 |
|                               | Mercaptan-functionalized<br>magnetic particles/HA<br>hydrogel   | 3D magnetic hydrogel composed of<br>hyaluronan with similar biochemical<br>and biophysical properties to brain<br>tissue  | Magnetomechanical<br>stimulation (calculated): 0.136<br>Hz, 0.15 to 1 µN, 30 min/day,<br>4 days   | Increased Ca <sup>2+</sup> influx in<br>DRG neurons   | 33 |
|                               | Zn <sub>0.4</sub> Fe <sub>2.6</sub> O <sub>4</sub> @SiO <sub>2</sub> @pNiP<br>MAm hydrogel<br>nanoparticles | Zn <sub>0.4</sub> Fe <sub>2.6</sub> O <sub>4</sub> converts magnetic<br>anisotropic energy to thermal energy to<br>induce the pNiPMAm contraction   | AMF, 500 kHz at 500 Oe, 30 s<br>ON/2 min OFF cycle for 15<br>times  | Activation of the Notch<br>signaling in Notch1-U2OS<br>cells  | 34 |

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