Electronic Supplementary Material (ESI) for Journal of Materials Chemistry B

Tracking the effects of PLGA-based nanoparticles on protein

expression in living cells through quantitative proteomics

Yuwan Chen,^{ab} Wen Zhou,^{ab} Jianhui Liu,^a Xinwei Li,^{ac} Wenxin Fu,^{ad} Baofu Ma,^{ab} Zhen Liang, ^a Kaiguang Yang,^{*a} Lihua Zhang,^{*a} and Yukui Zhang^a

a. State Key Laboratory of Medical Proteomics, National Chromatographic R. & A. Center, CAS Key Laboratory of Separation Science for Analytical Chemistry, Dalian Institute of Chemical Physics, Chinese Academy of Sciences, Dalian 116023, China.

b. University of Chinese Academy of Sciences, Beijing 100049, China.

c. Zhang Dayu School of Chemistry, Dalian University of Technology, Dalian 116024, China.

d. Research Center for Analytical Sciences, Northeastern University, Shenyang 110819, China.

Supplementary Figures and Table

Supplementary Table 1: The loading efficiency of DSS in the 10DSS-DDAB@PLGA/Kolliphor EL NPs, 40DSS-DDAB@PLGA/Kolliphor EL NPs and 70DSS-DDAB@PLGA/Kolliphor EL NPs, measured using HPLC.

Sample	10DDAB@PLGA/Kollip	40DDAB@PLGA/Kollip	70DDAB@PLGA/Kollip
	hor EL NPs	hor EL NPs	hor EL NPs
Loading	0.0881	19.37	24.84
efficiency (%)			



Supplementary Figure 1: Reproducibility of measurements. a-c Protein intensity measurements of HepG2 cells treated with DDAB@PLGA/Kolliphor EL NPs at 1, 3, and 6 h were plotted against with blank, and correlation is determined between measurements by Pearson's coefficient. d-g Protein intensity measurements of HepG2 cells treated with DDAB@PLGA/Kolliphor EL NPs were plotted against themselves and correlations between measurements were determined by Pearson's coefficient.



Supplementary Figure 2: Reproducibility of measurements. a-c Protein intensity measurements of HepG2 cells treated with 10DSS-DDAB@PLGA/Kolliphor EL NPs at 1, 3, and 6 h were plotted against with blank, and correlation is determined between measurements by Pearson's coefficient. d-g Protein intensity measurements of HepG2 cells treated with 10DSS-DDAB@PLGA/Kolliphor EL NPs were plotted against themselves and correlations between measurements were determined by Pearson's coefficient.



Supplementary Figure 3: Reproducibility of measurements. a-b Protein intensity measurements of HepG2 cells and HepG2 cells treated with 40DSS-DDAB@PLGA/Kolliphor EL NPs for 6 h were plotted against themselves. c Protein intensity measurements from HepG2 cells treated with 40DSS-DDAB@PLGA/Kolliphor EL NPs for 6 h were plotted against blank. d-e Protein intensity measurements of HepG2 cells and HepG2 cells treated with 70DSS-

DDAB@PLGA/Kolliphor EL NPs for 6 h were plotted against themselves. f Protein intensity measurements from HepG2 cells treated with 70DSS-DDAB@PLGA/Kolliphor EL NPs for 6 h were plotted against blank, and the correlation between the measurements was determined by Pearson coefficient.



Supplementary Figure 4: GO analysis of significant changed proteins. GO cellular component analysis results of the changed proteins identified in DDAB@PLGA/Kolliphor EL NP-treated HepG2 cells at all three time points.



Supplementary Figure 5: Significant changes in phosphorylated protein abundance. a-c Label-free quantification was performed to calculate changes in the abundance of phosphorylated protein sites between native and different DDAB@PLGA/Kolliphor EL NP-treated HepG2 cells for different times. Phosphorylated protein sites with a significant change in intensity of more than 2-fold in the treated group compared to the untreated group are shown as red dots, while the remaining proteins are shown as black dots. d The percentage of phosphorylation site intensities that significantly changed by more than 2-fold in the treated group compared to the untreated group compared to the untreated streated group compared to the untreated streated group compared to the untreated group is shown in light pink, while the remaining proteins are shown in sky blue. *P* value was calculated by using Student's T-test (adjusted p < 0.01). The P value was adjusted for multiple tests using FDR (Permutation-based). e Venn diagram

of the changed proteins at three time points. f GO analysis results for proteins with changes identified in all three time points were classified by biological process.



Supplementary Figure 6: GO analysis of significant changed proteins. GO cellular component analysis results of the changed proteins identified in 10DSS-DDAB@PLGA/Kolliphor EL NPs -treated HepG2 cells at all three time points.



Supplementary Figure 7: Significant changes in phosphorylated protein abundance. a-c Label-free quantification was performed to calculate changes in the abundance of phosphorylated protein sites between native and different 10DSS-DDAB@PLGA/Kolliphor EL NPs-treated HepG2 cells for different times. Phosphorylated protein sites with a significant change in intensity of more than 2-fold in the treated group compared to the untreated group are shown as red dots, while the remaining proteins are shown as black dots. d The percentage of phosphorylation site intensities that significantly changed by more than 2-fold in the treated group compared to the untreated group compared to the untreated symplement of proteins are shown in light pink, while the remaining proteins are shown in light pink, while the remaining proteins are shown of the untreated by using Student's T-test (adjusted p < 0.01). The P value was adjusted for multiple tests using FDR (Permutation-based). e Venn diagram of the changed proteins at three time points. f GO analysis results for proteins with





Supplementary Figure 8: GO analysis of significant changed proteins. GO cellular component analysis results of the changed proteins identified in different NPs-treated HepG2 cells for 6h. Treated-1 (10DSS-DDAB@PLGA/Kolliphor EL NPs were incubated with HepG2 cells for 6 h), Treated-2 (40DSS-DDAB@PLGA/Kolliphor EL NPs were incubated with HepG2 cells for 6 h) and Treated-3 (70DSS-DDAB@PLGA/Kolliphor EL NPs were incubated with HepG2 cells for 6 h).