Supplementary Data

Isolation of acute myeloid leukemia blasts from blood using a microfluidic device

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Supplementary Figures





Figure S1: Images from the videos obtained in the simulations. In the left, from the top to the bottom, is represented a section of each type of design: 3 lines, 4 lines and full lines, respectively. These sections were subjected to simulations in order to access to theoretical information about which design and geometry can potentially display a higher capture efficiency. In the right side the same pillar devices combined with the herringbones.

Table	S1 :	Result	ts of	the	simu	ilation	s in	a section	of the	device	and t	then of	extra	polated	for	whole	devices

		3L	3L + HB	4L	4L + HB	FL	FL + HB
# partialag atuak in unit call	2 µL/min	66	71	71	61	108	93
# particles stuck in unit cen	20 µL∕min	60	68	69	61	113	90
	40 µL∕min	62	70	68	60	110	100
	300	3L	3L + HB	4L	4L + HB	FL	FL + HB
% partialaa atuak in unit aall	2 µL/min	22.00%	23.67%	23.67%	20.33%	36.00%	31.00%
% particles stuck in unit cen	20 µL∕min	20.00%	22.67%	23.00%	20.33%	37.67%	30.00%
	40 µL∕min	20.67%	23.33%	22.67%	20.00%	36.67%	33.33%
	300	3L	3L + HB	4L	4L + HB	FL	FL + HB
% particles stuck in whole chip,	2 µL/min	93.50%	94.87%	97.72%	95.85%	98.20%	96.45%
according # unit cells	20 µL/min	91.41%	94.08%	97.42%	95.85%	98.58%	95.96%
	40 µL/min	92.17%	94.62%	97.26%	95.60%	98.36%	97.40%



Scheme S1. Schematic representation of the two different functionalisation strategies. Both characterised by the preliminar treatment with oxygen plasma to covalently bind functional silanes. Functionalisation A anchors the antibody through APTES and Glutaraldehyde (top); while functionalisation B composed links the antibody through CTS, activated with EDC/NHS (bottom).





Figure S2: XPS results of the silicon wafer treated with the two different functionalisation protocols. Functionalisation A based in the application of APTES, Glutaraldehyde and antibody (A) and Functionalisation B consists in the utilisation of CTS, EDC/NHS and antibody (B). XPS analysis was performed at each of the steps of the functionalisation protocol: silicon wafer before all the process (A i and B i); after plasma cleaner (A ii and B ii); after ethanol (A iii and B iii); after silane APTES (A iv) and CTS (B iv); after Glutaraldehyde (A v) and after EDC/NHS (B v); antibody (A vi and B vi). Comparison before and after the incubation of the antibody (A vii and B vii).



Figure S3: Fluorescently labelled antibody (in green) are patterned onto functionalised and bare glass slides by microcontact printing using a PDMS stamp.



Figure S4: Capture efficiency (%) of target cells in devices with CD33 and CD34 antibodies immobilised on the surface, using pillars (gap every 4 lines) combined with herringbone, using a flow rate of 40 μ L/min. The results are presented as mean ± SD of 3 independent biological replicates. Student's t test was applied to compare KG-1 (CD33+ / CD34+) and Jurkat (CD33- / CD34 -) cells isolation efficiency. *p<0.05.



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Figure S5. FC analysis and microscopic images obtained from immunocytochemistry (ICC) assays, in well-plates, for aberrant surface markers in AML cells. CD34 positivity was significantly different and higher in the KG-1 cells (A). Phenotypical characterization using CD7 and CD56 markers in AML cells (B): KG-1 and (i) HL-60 (ii) cells seeded in a well-plate. All cells were stained for DAPI (1:10), and CD7 and CD56 (1:50), in blue, green and red, respectively.

The FC results presented as mean \pm SD of 3 independent biological replicates. For the flow rates and controls an one-way ANOVA and Tukey's post hoc test were used to compare the different flow rates in the same type of device (gap every 4 lines), as well as for the different controls using the same device and flow rate. For the analysis of FC results a 2 way ANOVA test was applied to compare the expression of CD7, CD56 and both in KG-1 and HL-60 cells. ****p < 0.0001.

Table S2. Table with NGS results from clinical samples. Genetic analysis of the samples through a panel selected for the study of AML patients. Multiple genes relevant to AML disease were considered, as well as genes not associated with AML, but potential targets for study in this disease

Sample	Gene	Variant allele frequency (VAF %) BM PB		c.DNA	Protein	Classification	Observations	
	FLT3	<i>FLT3</i> >5		c.1773_1793dup	p.(Tyr597_Glu598ins AspValAspPheArgGluTyr)	TIER I	Prognostic and therapeutic indication in AML	
	IDH2	>5	>5	c.419G>A	p.(Arg140Gln)	TIER I	Prognostic and therapeutic indication in AML	
	NPM1	>5 >5		c.860_863dup	p.(Trp288Cysfs*12)	TIER I	Prognostic and therapeutic indication in AML	
t 1	ETV6	>5 >5		c.602T>C	p.(Leu201Pro)	TIER III	Not associated with AML	
Patien	SRSF2	>5	>5	c.284C>G	p.(Pro95Arg)	TIER I	Associated with AML	
	SETBP1	>5	>5	c.691G>C	p.(Val231Leu)	TIER IV	Without clinical significance	
	TET2	>5	>5	c.5162T>G	p.(Leu1721Trp)	TIER IV	Without clinical significance	
	ASXL1	>5	>5	c.2444T>C	p.(Leu815Pro)	TIER IV	Without clinical significance	
	KIT	>5	>5	c.1621A>C	p.(Met541Leu)	TIER III	Associated with AML	
	TP53	>5	>5	c.215C>G	p.(Pro72Arg)	TIER IV	Without clinical significance	
	TP53	>5	>5	c.375+5G>	unknown	TIER III	Not associated with AML	
Patient 2	TP53	>5 >5		c.215C>G	p.(Pro72Arg)	TIER IV	Without clinical significance	
	CSF3R	>5	>5	c.1319G>A	p.(Arg440Gln)	TIER IV	Without clinical significance	
	SETBP1	>5	>5	c.3388C>A	p.(Pro1130Thr)	TIER III	Not associated with AML	
	TET2	>5	>5	c.1064G>A	p.(Gly355Asp)	TIER IV	Without clinical significance	
	TET2	>5	>5	c.5284A>G	p.(lle1762Val)	TIER IV	Without clinical significance	
	ASXL1	>5	>5 >5 c.5284A>G		p.(Leu815Pro)	TIER IV	Without clinical significance	