

## Supplementary Information

### A Disposable Paper-based Electrochemical Biosensor Decorated by Electrospun

#### Cellulose Acetate Nanofibers for highly Sensitive Bio-detection

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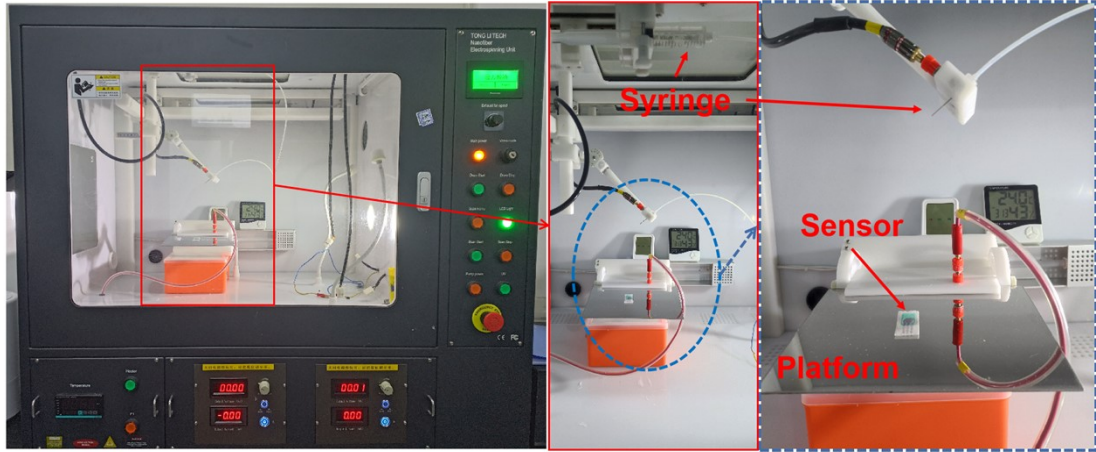
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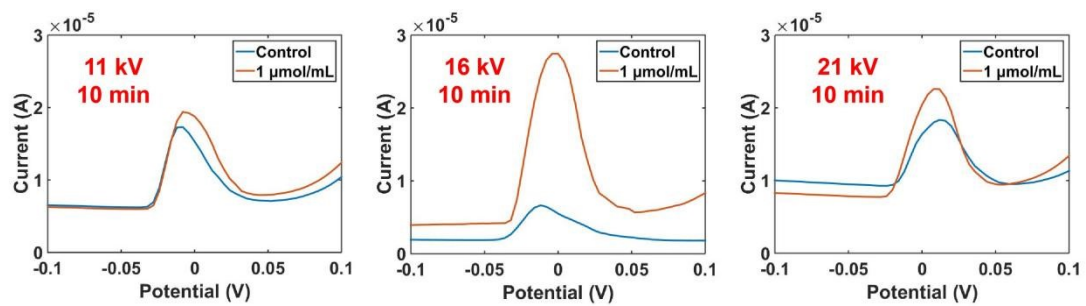
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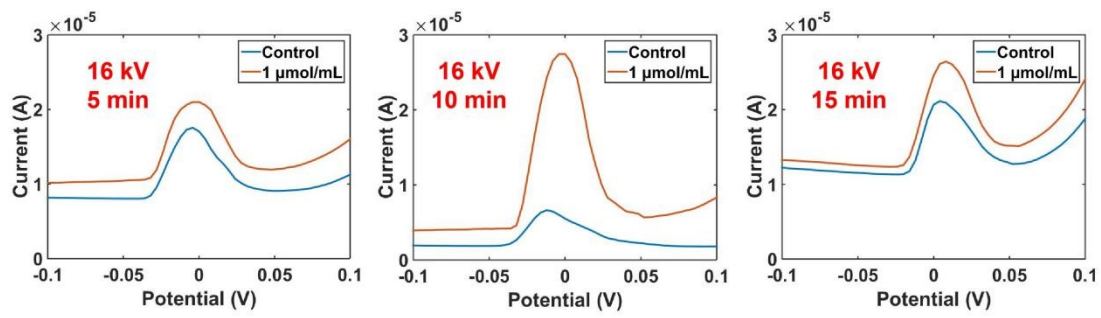
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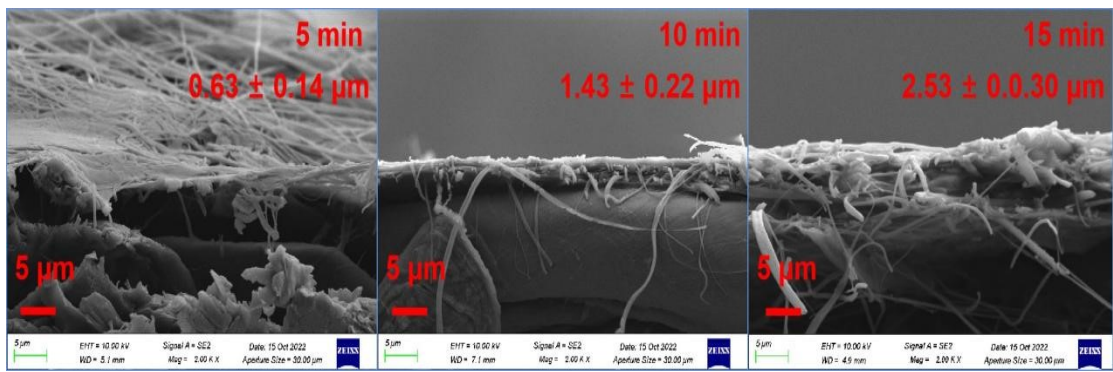
**Fig. S1** The details of electrospinning device and the sensor.



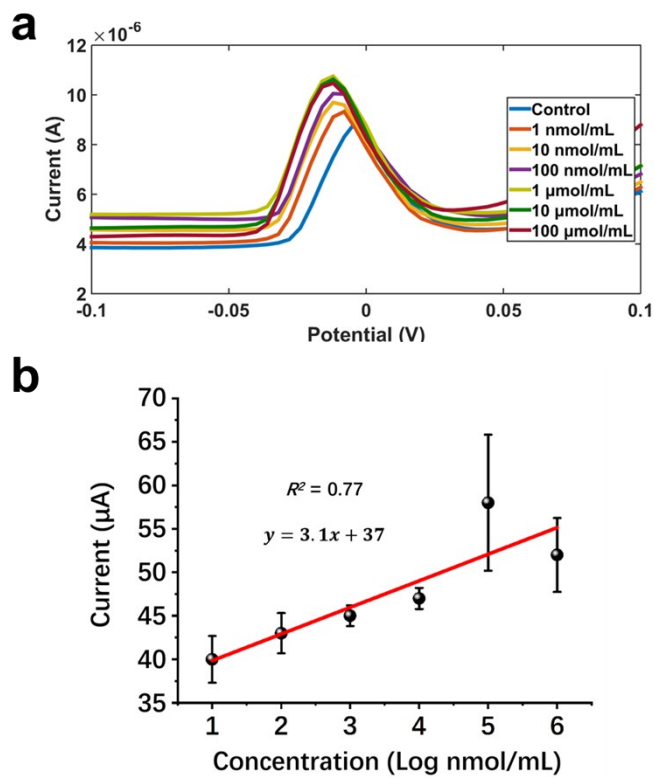
**Fig. S2** DPV curves of electrospinning CA nanofibers with different excitation voltage (11, 16, and 21 kV); coating time duration=10 min.



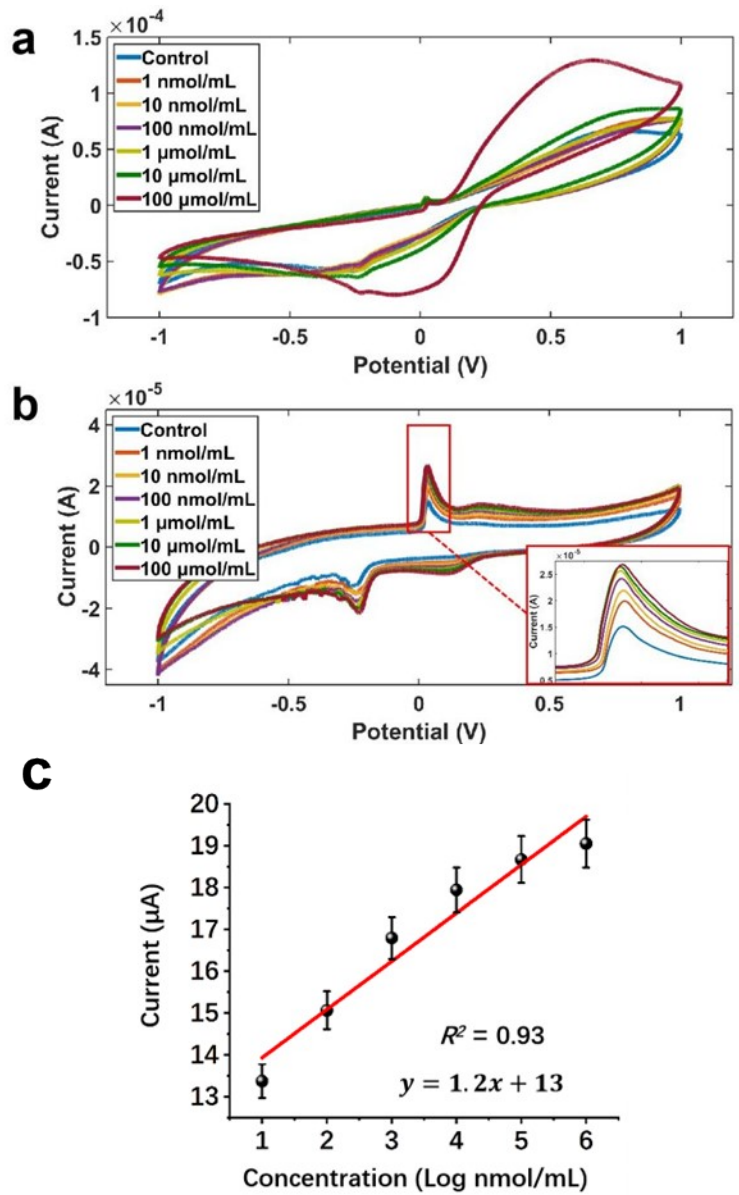
**Fig. S3** DPV curves of the CA nanofibers at different electrospinning duration. 5 (a), 10 (b), and 15 min (c) at voltage=16 kV.



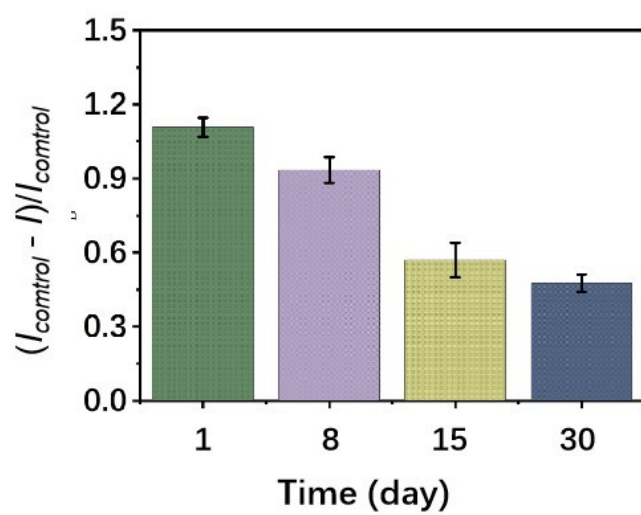
**Fig. S4** SEM images of the cross-section and the thickness of the CA nanofibers at different electrospinning duration. 5 (a), 10 (b), and 15 min (c) at voltage=16 kV.



**Fig. S5** (a) DPV curves and (b) calibration curve of glucose detection by the bare PBSP electrodes.

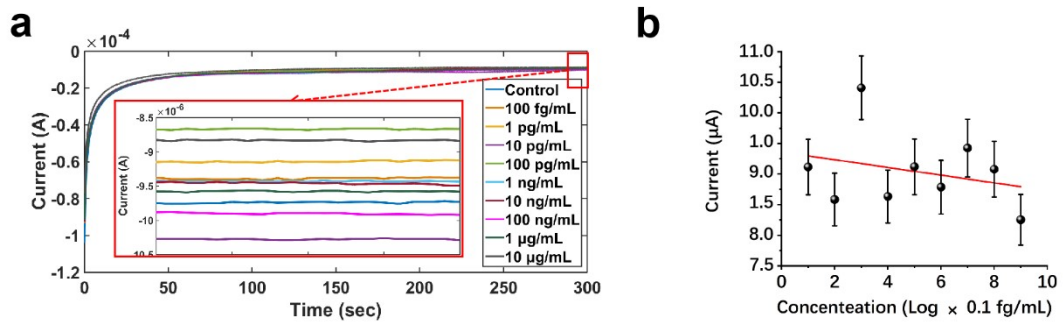


**Fig. S6** (a) CV curves of glucose detection by the bare PBSP electrodes; (b) CV curves and (c) calibration curve of glucose detection by CA NFs-decorated PBSP electrodes.

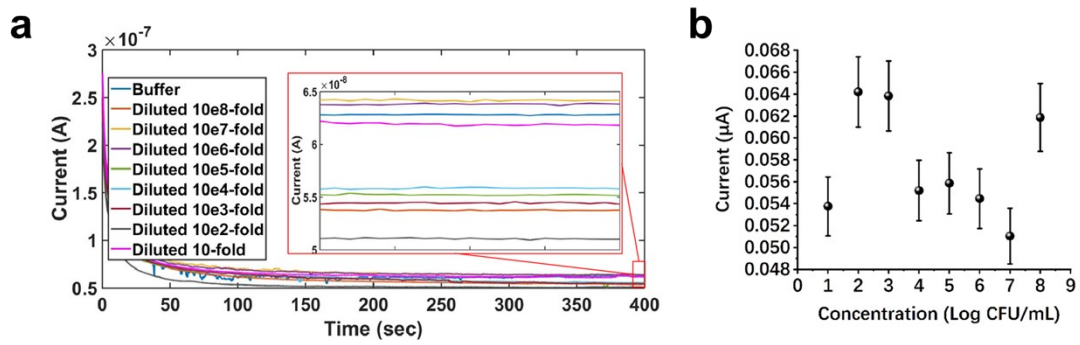


**Fig. S7** Stability of CA NFs-decorated PBSP electrodes after modification of glucose oxidase





**Fig. S8** (a)  $I$ - $t$  curves and (b) calibration curve of Ag85B protein detection by bare PBSP electrodes.



**Fig. S9** (a) I-t curves and (b) calibration curve of *E. Coli* O157:H7 detection by the CA NFs-decorated PBSP electrodes without the *E. coli* O157:H7 monoclonal antibody.

**Table S1.** Comparison of different electrochemical sensors for detecting glucose.

Electrode	Paper Structure	Technique	Range & LOD	Reference
CA/ZIF-8@enzyme/MWCNTs/AuNPs	No	-	1 – 10 $\mu\text{mol/mL}$ LOD: 5.347 $\text{nmol/mL}$	Xin L., et al. <sup>1</sup>
PVA-SbQ-MWCNTCOOHs	No	-	5 $\text{nmol/mL}$ - 4 $\mu\text{mol/mL}$ LOD: 2 $\text{nmol/mL}$	Eleni S., et al. <sup>2</sup>
Graphene/polyaniline/AuNPs/glucose oxidase SPCE	Yes	DPV	0.2 - 11.2 $\mu\text{mol/mL}$ LOD: 100 $\text{nmol/mL}$	Fen-Ying K., et al. <sup>3</sup>
PERs using a 3D printed BIA cell/SPEs	Yes	DPV	1 - 10 $\mu\text{mol/mL}$ LOD: 110 $\text{nmol/mL}$	Anderson A D., et al. <sup>4</sup>
CA NFs/paper-based SPEs	Yes	CV/DPV	1 $\text{nmol/mL}$ - 100 $\mu\text{mol/mL}$ LOD: 0.71 $\text{nmol/mL}$	<b>Present work</b>

**Table S2.** Comparison of different electrochemical sensors for detecting protein.

Electrode	Paper Structure	Technique	Range&LOD	Reference
Ab-25OHD/SPE/FMTAD	Yes	SPR	10 - 100 ng/mL LOD: 10 ng/mL	Chauhan D., et al. <sup>5</sup>
MWCNTs-doped Chitosan NFs	No	-	1 pg/mL - 1 ng/mL LOD: 0.05 pg/mL	Wang X., et al. <sup>6</sup>
PMPC-S/AuNPs-SPCE PADs	Yes	DPV	5 - 5000 ng/mL LOD: 1.6 ng/mL	Chanika P., et al. <sup>7</sup>
SPEs/rGO-TEPA/Au and simple paper-based microfluidic devices	Yes	SWV	0.01 - 100.0 ng/mL LOD: 0.005 ng/mL	Liangli C., et al. <sup>8</sup>
CA NFs/paper-based SPEs	Yes	Chronoamperometry	100 fg/mL - 10 µg/mL LOD: 89.1 fg/mL	<b>Present work</b>

**Table S3.** Comparison of different electrochemical sensors for detecting bacteria.

Electrode	Paper Structure	Technique	Range&LOD	Reference
Silica NPs on polyelectrolyte multilayer on Au electrode	No	CV	$8 \times 10^4 - 8 \times 10^6$ CFU/mL LOD: $2 \times 10^3$ CFU/mL	Mathelie G., et al. <sup>9</sup>
Fluoride-doped tin oxide electrode	No	DPV	$10^3 - 10^7$ CFU/mL LOD: $10^3$ CFU/mL	Divagar M., et al. <sup>10</sup>
Carbon paste, a mixture of multi-walled carbon nanotube (MWCNT)	Yes	DPV	$6.9 \times 10^2 - 10^6$ CFU/mL LOD: 690 CFU/mL	Chanhwi P., et al. <sup>11</sup>
CA NFs/paper-based SPEs	Yes	Chronoamperometry	$1.5 \times 10^2 - 1.5 \times 10^6$ CFU/mL, LOD: 30 CFU/mL	<b>Present work</b>

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