

## Characterization of Vaginal Lactobacillus in Biologically Relevant Fluid using Surface-Enhanced Raman Spectroscopy

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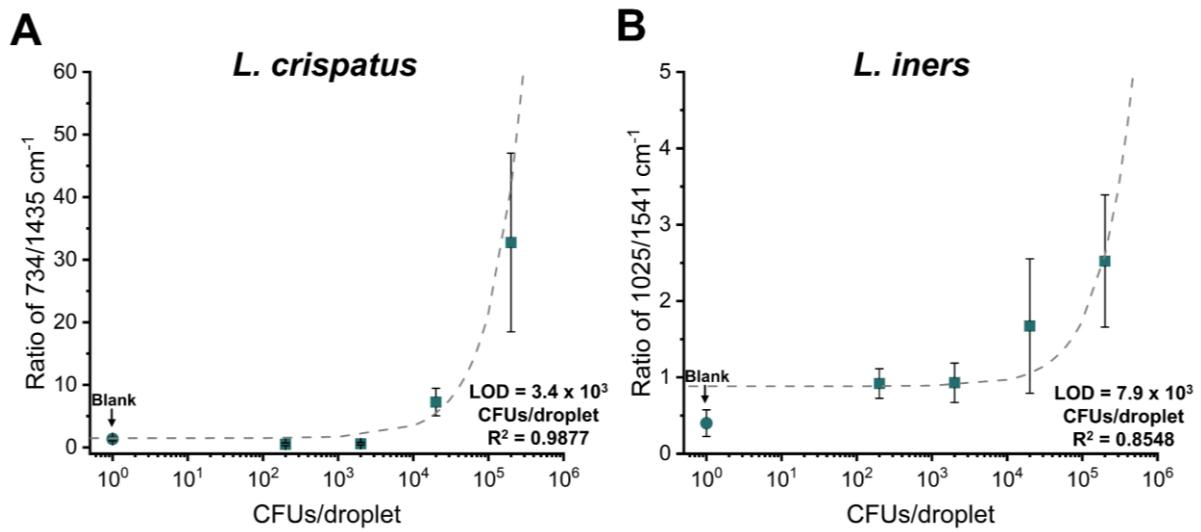
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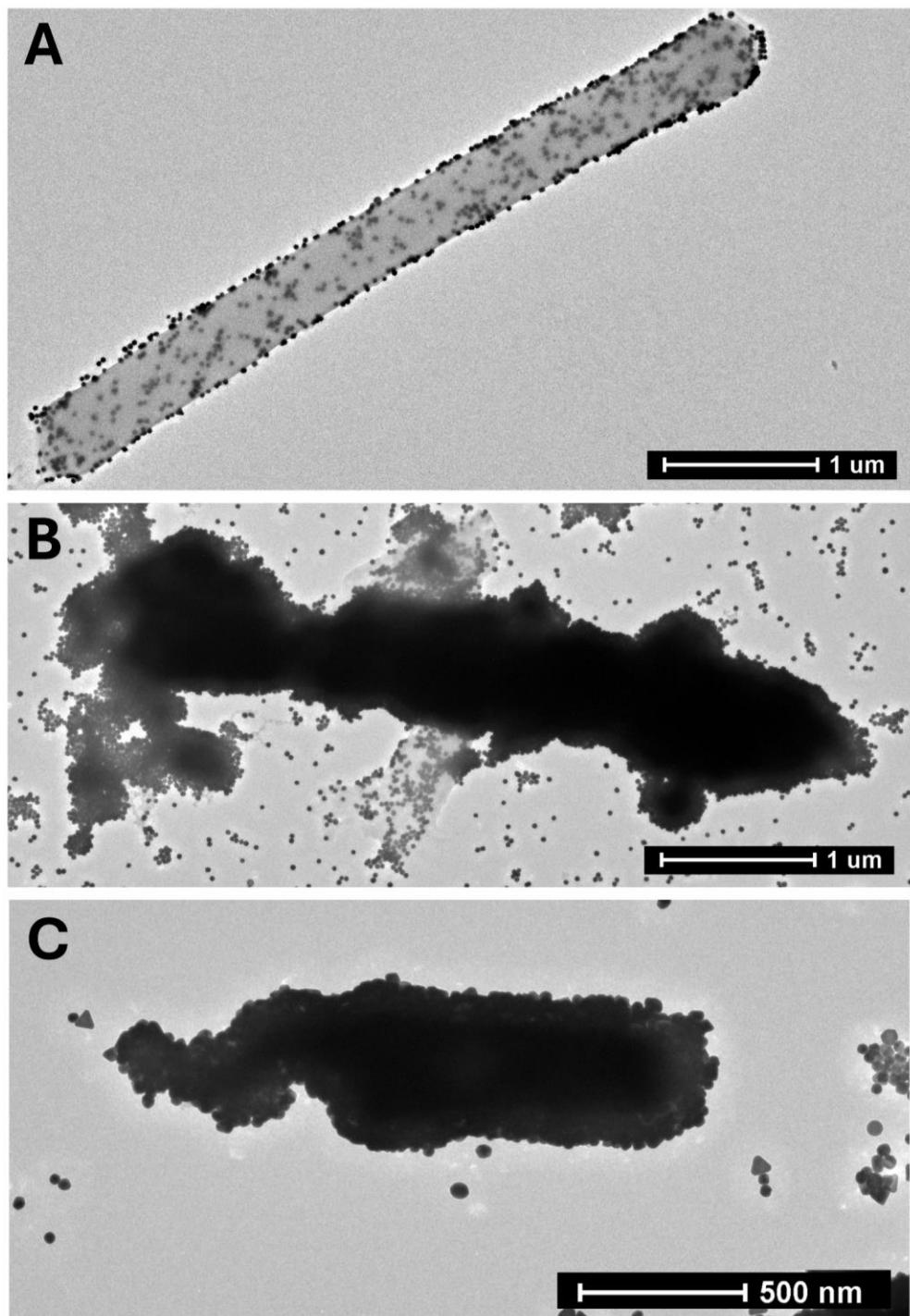
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### Transmission Electron Microscopy

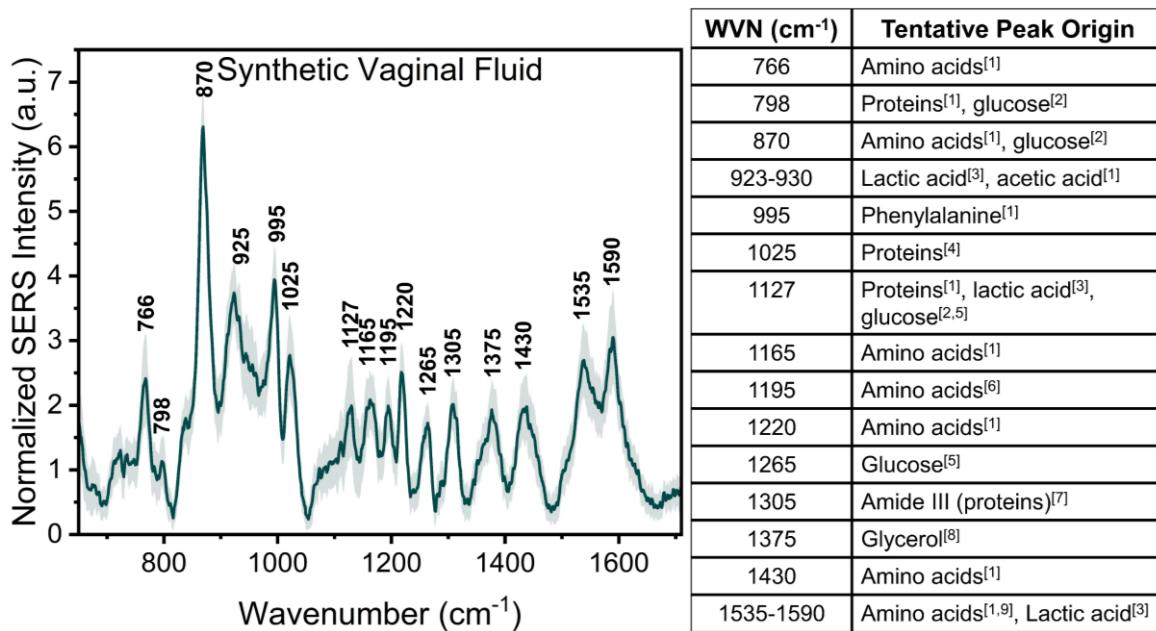
*L. crispatus* cultures were grown following procedures outlined previously. Liquid cultures were washed via centrifugation at 3,300g for 8 minutes and resuspended with sterile DI water. Washed cultures and AuNPs were mixed at a 1:4 volumetric ratio (bacteria:AuNPs) by pipetting and were vortexed for 2-3 seconds to ensure homogenous mixing. The solution was allowed to sit for 15 minutes to allow for AuNPs and bacteria interaction. Following the 15-minute interaction period, the solution was centrifuged at 725 RCF for 15 minutes and the supernatant was replaced with 2.5% glutaraldehyde for bacterial cell fixation. The solution was fixed in glutaraldehyde for 30 minutes before a 5 µL droplet was applied to a TEM grid (Ted Pella; Redding, CA). The grid was allowed to dry for 30 minutes then washed with DI water twice. The grid was allowed to fully dry before being imaged using a Tecnai Osiris TEM (Thermo Fisher Scientific, Waltham, MA) at 3,600X and 6,300X magnification with 200 kV accelerating voltage and a 1 second integration time.



**Supplemental Figure 1.** Scatter plots of ratiometric values used to calculate the limit of detection (LOD) of (A) *L. crispatus* and (B) *L. iners* with goodness of fit ( $R^2$ ) reported.



**Supplemental Figure 2.** TEM images of *L. crispatus* and *L. iners* cells at varying bacterial concentrations showing variations in the density of AuNPs attached to the bacterial cell wall and differing cell morphology: (A) *L. crispatus*,  $2 \times 10^5$  CFUs/droplet, 3600X magnification; (B) *L. crispatus*,  $2 \times 10^3$  CFUs/droplet, 3600X magnification; (C) *L. iners*,  $2 \times 10^3$  CFUs/droplet, 6300X magnification.



**Supplemental Figure 3.** SERS spectrum of SVF. Each spectra is the mean and standard deviation (represented by shaded error bars) of 3 experimental replicates [n=45 spectra]) and table of peak origins for the highlighted peaks.

**Supplemental Table 1.** Table of wavenumber assignments of peaks with VIP value greater than 1 in both PLSR models.

WVN (cm <sup>-1</sup> )	Pure or Complex	Tentative assignment
734	Both	C-N stretching, the glycosidic ring vibrations, or in plane breathing mode of adenine <sup>[10-13]</sup>
768	Pure	Indole ring breathing/stretching <sup>[6, 16, 17]</sup>
865	Complex	Indole ring scissoring, indole N-H displacement, indole ring vibration with N-H bending, C-H stretching <sup>[1,2,14]</sup>
962	Pure	C=C deformation, C-N stretching, C-O stretching, CH <sub>3</sub> symmetric stretching <sup>[11]</sup>
989-998	Both	Phenyl ring angular bending vibrations <sup>[15]</sup>
1025	Complex	C-N or C-C stretching, or C-H in plane bending <sup>[11]</sup>
1315-1320	Both	CH <sub>3</sub> CH <sub>2</sub> vibrations, Amide III <sup>[6]</sup>
1340	Pure	CH <sub>3</sub> CH <sub>2</sub> wagging, Amide III, CH <sub>2</sub> wagging, C-H deformation <sup>[6]</sup>
1380-1390	Complex	CH <sub>3</sub> , C-H, C-N vibrations <sup>[6]</sup>
1430-1440	Both	CH <sub>2</sub> scissoring and deformation <sup>[6]</sup>
1457-1468	Complex	CH <sub>2</sub> and C-H vibrations <sup>[11]</sup>
1535	Pure	Amide carbonyl group vibrations <sup>[6]</sup>
1560	Pure	Amide I, C=O stretching, C=C bending <sup>[11]</sup>

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