

Supporting Information

MOFs-mediated dual energy transfer nanoprobe integrated with Exonuclease III amplification strategy for highly sensitive detection of DNA

Xiaojing Xing,^{*a} Mengying Gao,^a Minglin Lei,^a Kunqi Cheng,^a Yifan Zhao,^a

Xianchao Du,^a Luyi Zong,^a Dongfang Qiu,^{*a} and Xueguo Liu^{*b}

^a *College of Chemistry and Pharmaceutical Engineering, Nanyang Normal University, Nanyang 473061, China.*

^b *Key Laboratory of Henan of Industrial Microbial Resources and Fermentation Technology, and Department of Biology and Chemical Engineering, Nanyang Institute of Technology, Nanyang 473004, China.*

^{*}Corresponding authors. Xiaojing Xing, *e-mail addresses:* xjxing@hnu.edu.cn; Dongfang Qiu, *e-mail addresses:* qiudf2008@163.com; Xueguo Liu, *e-mail addresses:* huanliu1987@126.com

Table of Contents

1. Materials.....	S1
2. Apparatus.....	S2
2. Fig. S1 EDS analysis of UiO-66-NH ₂	S3
3. Fig. S2 Variation of the fluorescence intensity of FAM-A20 with UiO-66-NH ₂ concentration.....	S4
4. Fig. S3 Variation of the fluorescence intensity of FAM-T20 with UiO-66-NH ₂ concentration.....	S5
5. Fig. S4 Variation of the fluorescence intensity of FAM-C20 with UiO-66-NH ₂ concentration.....	S6
6. Fig. S5 Variation of the fluorescence intensity of FAM-G20 with UiO-66-NH ₂ concentration.....	S7
7. Fig. S6 Construction of four control HBV-DNA sensing platforms and their response to HBV-DNA with different concentrations.....	S8

Materials

Zirconium tetrachloride (ZrCl₄, 98%) and 2-aminoterephthalic acid (H₂N-H₂BDC, 98%) were obtained from Aladdin (Shanghai, China). N,N'-dimethylformamide (DMF, AR), acetic acid(AR), sodium chloride (NaCl, AR), and magnesium chloride (MgCl₂, AR) were obtained from Sinopharm Chemical Reagent Co., Ltd (Shanghai, China). Agarose, 6×loading buffer, and GelRed (10000×) nucleic acid stain were purchased from Shanghai Sangon Biotech Co., Ltd (China). The DNA sequences purified by high-performance liquid chromatography (HPLC), were obtained from Shanghai Sangon Biotech Co., Ltd (China). All the sequences are listed as following:

FDNA: 5'-*AGA AGA GAA GTC CAT ATA ACT GAA AGC CAA*-FAM-3'

QDNA:5'-*BHQ1-GT TAT ATG GAC TTC CCCCCCCCCCCCCCCCCC*-3'

HBV-DNA: 5'-*TTG GCT TTC AGT TAT ATG GAT GAT GTG GTA*-3'

Mis-1: 5'-GTG GCT TTC AGT TAT ATG GAT GAT GTG GTA-3'

Mis-2: 5'-GTG GCT TTC AAT TAT ATG GAT GAT GTG GTA-3'

Mis-3: 5'-GTG GCT TTC AAT GAT ATG GAT GAT GTG GTA-3'

HPV-DNA: 5'-CTC ATA ACA GTA GAG ATC AGT T-3'

HIV-DNA: 5'-CCC TTT ACT GCT AGA GAT TTT CCA CAT TTT CCC G-3'

FAM-A20: 5'-FAM-AAAAAAAAAAAAAAAAAAAAA-3'

FAM-T20: 5'-FAM-TTTTTTTTTTTTTTTTTTTT-3'

FAM-C20: 5'-FAM-CCCCCCCCCCCCCCCCCCC-3'

FAM-G20: 5'-FAM-GGGGGGTGGGGGGTGGGGGG-3'

The buffer we used during the research was composed of 25 mM HEPES (pH 7.6), 150 mM NaCl and 1 mM MgCl₂. The ultrapure water used in all experiments was prepared on a Millipore milliq water purification system (Billerica, MA, USA).

Apparatus

Transmission electron microscopy (TEM) and scanning electron microscopy (SEM) were imaged using a JEM-2100F microscopy and Zeiss Sigma-500 FESEM, respectively. X-ray diffraction (XRD) analysis was performed using a Bruker D8 Advance X-ray diffractometer. Fourier-transform infrared (FTIR) spectra ranging from 500 to 4000 cm^{-1} were recorded on an IRTracer-100 instrument. DNA concentration was quantified using a UV-2600 spectrophotometer purchased from Shimadzu. Fluorescence spectra were measured using a Cary Eclipse fluorescence spectrophotometer (Varian, USA). The obtained data were processed using Origin software.

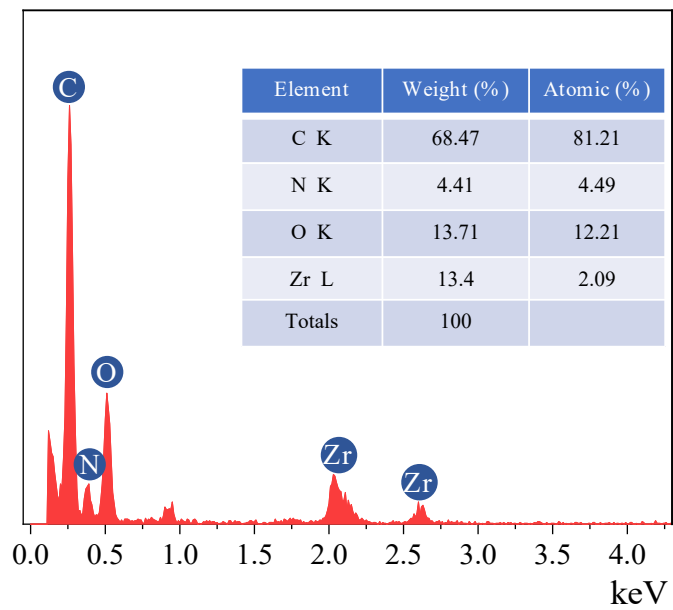


Fig. S1 EDS analysis of UiO-66-NH₂.

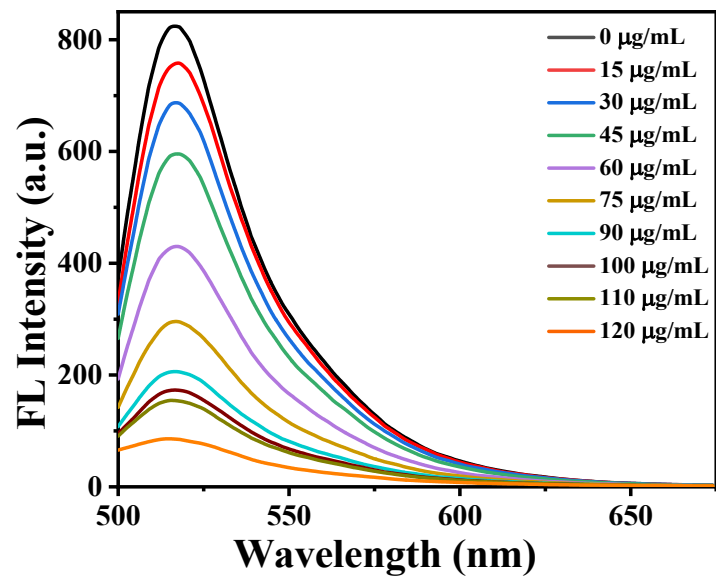


Fig. S2 Variation of the fluorescence intensity of FAM-A20 with UiO-66-NH₂ concentration.

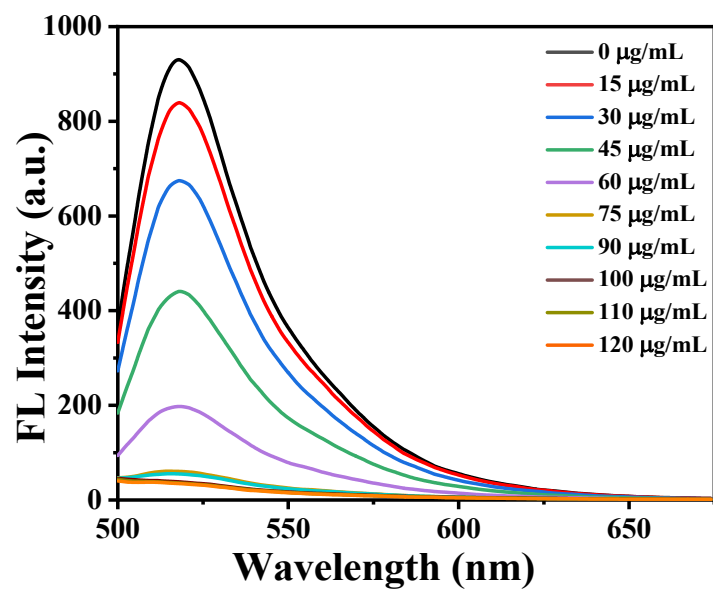


Fig. S3 Variation of the fluorescence intensity of FAM-T20 with UiO-66-NH₂ concentration.

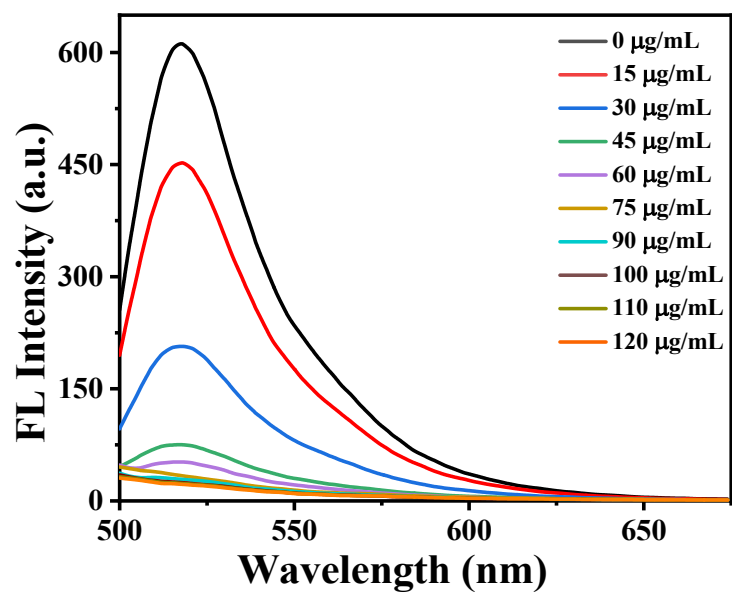


Fig. S4 Variation of the fluorescence intensity of FAM-C20 with UiO-66-NH₂ concentration.

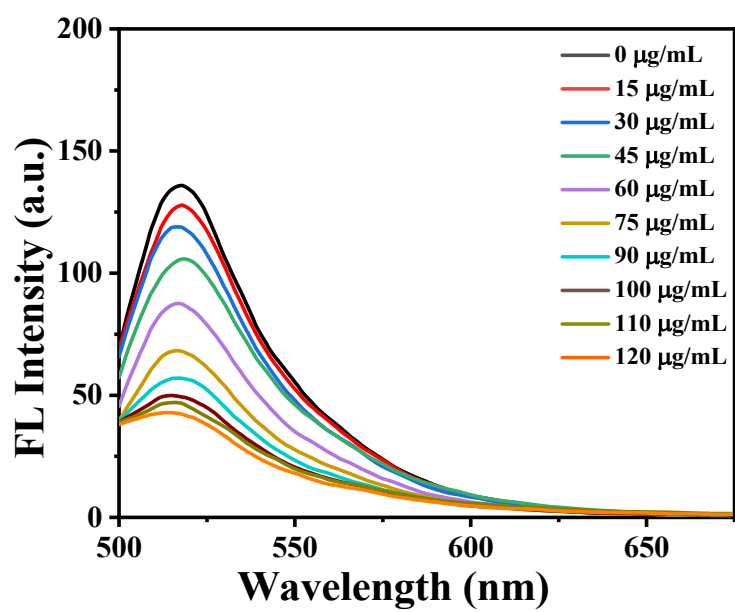


Fig. S5 Variation of the fluorescence intensity of FAM-G20 with UiO-66-NH₂ concentration.

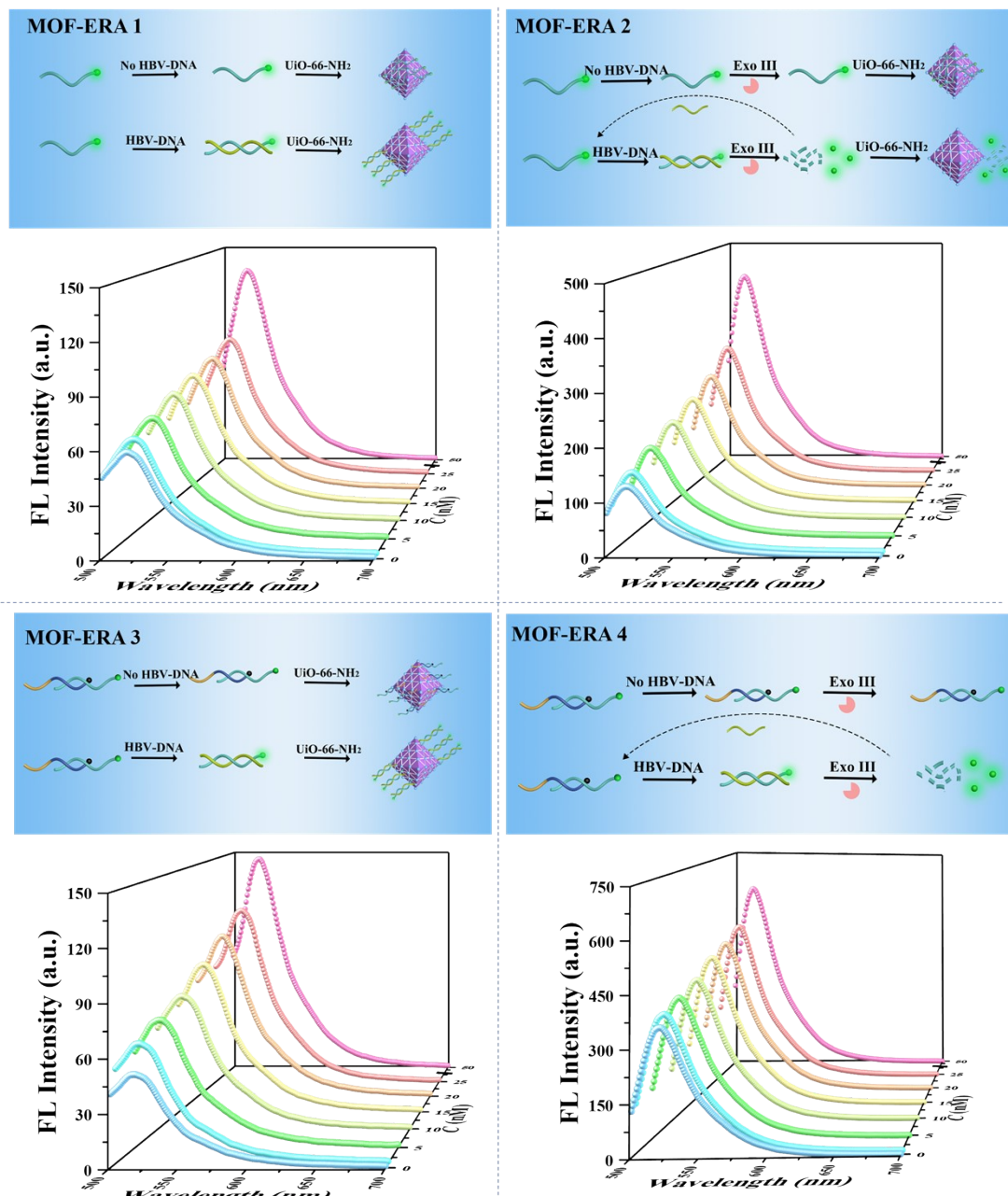


Fig. S6 Schematics of MOF-ERA1, MOF-ERA2, MOF-ERA3 or MOF-ERA4 platform for HBV-DNA detection and their fluorescence response to HBV-DNA with different concentrations.