

## SUPPLEMENTARY MATERIAL

### **A biphasic accelerated strand exchange amplification strategy for culture-independent and rapid detection of *Salmonella enterica* in food samples**

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**Table S1** Sequences information used in this work

Method	Name	Sequence (5'-3')
ASEA	<i>Salmonella enterica</i> strain <i>fimY</i> gene ( <sup>a</sup> CP051213.1 <sup>b</sup> 3426259-3426297)	<u>CGCTTAACCAGCTACGCGGCTCAGTT</u>
		<b>GGCAACAAAGC</b>
		CGCTTAACCAGCTACGCG
	R1	GCTTTGTTGCCAACTGAGC
<sup>c</sup> Real-time PCR	F2	TCGCACCGTCAAAGGAACCGTAAAGC
	R2	GCATTATCGATCAGTACCAGCCGTCT

<sup>a</sup> GenBank accession number. <sup>b</sup> The position of the specific sequence in the *fimY* gene. The underlined portion is the same sequence as that of primer F1. The sequence complementary to primer R1 is shown in bold. <sup>c</sup> The primers of real-time PCR were designed according to Chinese national standard (GB/T 28642-2012).

**Table S2** The performance of the biphasic ASEA method compared with other bacteria detection methods

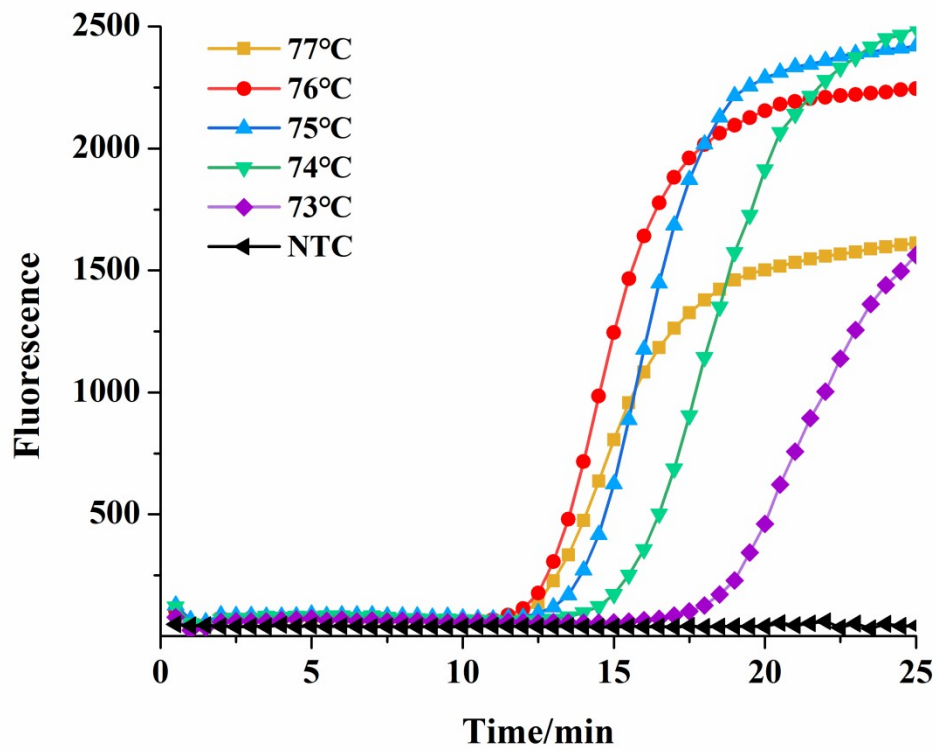
Detection method	Detection limit (CFU/mL)	Assay time (min)	Reference
DNA aptamer-enzyme-linked colorimetric method	$< 10^3$	30-40	1
LAMP-lateral flow dipstick	$3 \times 10^2$	90	2
Biacore surface plasmon resonance (SPR)-based biosensor method	23	$< 60$	3
Immunosensing method	10	30	4
Photosensitization colorimetric method	13 ( <i>Salmonella</i> in pure culture)	65	5
Fluorescent lateral flow immunoassay method	50 cells/mL	15	6
Biphasic ASEA method	50 ( <i>Salmonella</i> in milk sample)	50	This work

**Table S3** Biphasic ASEA and gold standard culture-based methods for detection of *S. enterica* in 82 food samples

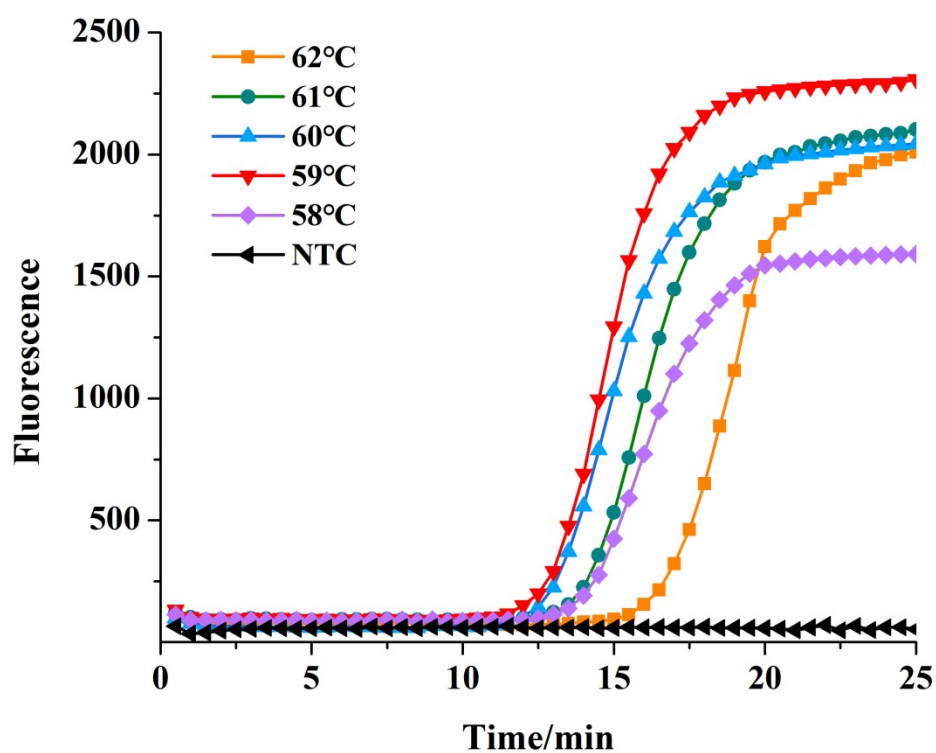
Kind of samples*	Samples number*	Detection results	
		Biphasic ASEA (this study)	Culture-based method
Milk	1-9	<input type="checkbox"/>	<input type="checkbox"/>
	10-14	<input type="checkbox"/>	<input type="checkbox"/>
	15-19	<input type="checkbox"/>	<input type="checkbox"/>
Scallop	20-23	<input type="checkbox"/>	<input type="checkbox"/>
	24-30	<input type="checkbox"/>	<input type="checkbox"/>
	31	<input type="checkbox"/>	<input type="checkbox"/>
	32-36	<input type="checkbox"/>	<input type="checkbox"/>
Oyster	37-39	<input type="checkbox"/>	<input type="checkbox"/>
	40	<input type="checkbox"/>	<input type="checkbox"/>
	41-43	<input type="checkbox"/>	<input type="checkbox"/>
	44-47	<input type="checkbox"/>	<input type="checkbox"/>
Duck	48	<input type="checkbox"/>	<input type="checkbox"/>
	49-55	<input type="checkbox"/>	<input type="checkbox"/>
	56-58	<input type="checkbox"/>	<input type="checkbox"/>
	59-61	<input type="checkbox"/>	<input type="checkbox"/>
Lettuce	62-66	<input type="checkbox"/>	<input type="checkbox"/>
	67	<input type="checkbox"/>	<input type="checkbox"/>
	68	<input type="checkbox"/>	<input type="checkbox"/>
Cucumber	69-70	<input type="checkbox"/>	<input type="checkbox"/>
	71-77	<input type="checkbox"/>	<input type="checkbox"/>
	78	-	<input type="checkbox"/>
	79-82	<input type="checkbox"/>	<input type="checkbox"/>

\*The 82 samples included 19 of milk, 12 of scallop, 12 of oyster, 15 of duck, 10 of lettuce, and 14 of cucumber.

represented that *S. enterica* was detected as positive;  represented that *S. enterica* was detected as negative.



**Fig. S1** Real-time fluorescence curves of biphasic ASEA reaction with serial initial Td values from 73 to 77°C. Non-targeted control (NTC) was conducted with DNase-free water as the target.



**Fig. S2** Real-time fluorescence curves of biphasic ASEA reaction with serial  $T_r$  values from 58 to 62°C. NTC was conducted with DNase-free water as the target.

## References

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