Supporting information

Microgel-based Modular 3D in vitro microfluidic cell culture platform

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Туре	Size
Small D: 3 cm F: 90 mL/min P: 10%	 515.625 μm 531.250 μm 580.423 μm 585.950 μm 606.794 μm Average size = 564 ± 38.71 μm
Medium D: 4.5 cm F: 60 mL/min P: 20%	 791.746 μm 835.938 μm 865.586 μm 882.809 μm 885.416 μm Average size = 852.29 ± 39.17 μm
Large D: 6 cm F: 45 mL/min P: 25%	 1. 1546.875 μm 2. 1572.917 μm 3. 1625 μm 4. 1651.042 μm 5. 1656.250 μm Average size = 1610.41 ± 48.49 μm

 Table S1: Average diameters for various sizes of microgels fabricated using

	Chip 1 (pixels)	Chip 2 (pixels)
Day 2	12.04 to 119.94 Av : 57.3	13.23 to 109.12 Av: 62.4
Day 6	5.83 to 84.77 Av: 34.1	7.89 to 77.42 Av: 36.2
Day 14	9.43 to 64.63 Av: 39.1	6.23 to 58.23 Av: 23.7

Table S2: Distance of Huh-7 (red) to nearest NIH-3T3 (green fluorescence) in twodifferent sets of microgels as chip 1 and chip 2 using FIJI image analyzer distance tool.

Day	1:1	1:2	1:3	2:3
2	2.18	4.23	3.43	4.51
6	3.92	4.39	4.64	27.86
8	3.94	4.63	4.29	25.74

Table S3: Albumin (pg/mL) production in HF conditions using different cell density ratios over 8 days time-period.



Figure S1: COMSOL simulation of diffusion across three sizes of microgels

	2 2 2 2 2 2 2 2 2 2 2 2 2 2	2495 2495 2495 2495 2495 2495 2495 2495	2484 2433 2447 243 2448 2448 2448 2448 2448 2448 2448
	Count	Area %	Min Feret Diameter (um)
Picture1-	157	16 179	19 /29

Figure S2: Binary for the SEM in Figure 1E to calculate the pore size.



Figure S3: (A) Fluorescence emission spectra at excitation 500nm for carbon dots (0.5mg/mL) dispersed in PBS at pH 9.5, pH 7.4 and pH 5.5; (B) Fluorescence microscopic images of Carbon dots encapsulated with cells for Cell proliferation and (C) Cytocompatibility assay of Carbon dots using various concentrations from 0.005-1mg/mL using Huh-7 cells

Optimization of the chip design:

The microfluidic chamber design was based on the intrinsic architecture of a hepatic lobule, the functional building block of the liver. Each lobule consists of a network of sinusoids linking the hepatic portal triad to the central vein. Initially, a hexagonal chamber (Figure S2A) was designed to emulate this structure, comprising six inlet ports converging into a common outlet port through several microfluidic channels. These channels were dimensioned to accommodate single medium-sized microgels linearly without overlap. At the end of each channel, a cylindrical post was constructed to sustain dynamic flow while preventing the microgels from escaping into the outlet. However, during manufacturing, the radial convergence of the channels at the outlet led to unintended inter-channel mixing at the outlet openings. To address this flaw, the chamber design was improved into a rectangular arrangement with 10 inlet ports connecting perpendicularly via microfluidic channels into a central outlet (Figure S2B). The new design retained all prior dimensions and features while providing the flexibility of including additional inlets, facilitating the inclusion of a greater amount of cell-encapsulated microgels, and thereby enhancing the efficiency of functionality evaluations within the chip.



Figure S4: **Evaluating the design of microfluidic chip:** (A) represents design comprising hexagonal lobular architecture and (B) The rectangular design with 10 inlet pores connecting perpendicularly via microfluidic channels into a central outlet.



Figure S5: TGF- β (Transforming growth factor- β) Assay for the combinations (H+F) and (HF) microgels in dynamic condition.



Figure S6: TNF- α (Tumor Necrosis factor- α) Assay for the combinations (H+F) and (HF) microgels in dynamic condition.



Figure S7: (A) Distance of Huh-7 to nearest NIH-3T3 over 14 days time-period in chip 1and (B) distance of Huh-7 to nearest NIH-3T3 over 14 days time period in chip 2.

The chip distance range refers to the minimum and maximum distance of Huh-7 to the NIH-3T3.