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**Figure S1: Number of PCR I cycles over the course of the selection.** Eleven subrounds were used in selection round 1 to increase the coverage of sequence space. Symbols of the same sub-round, with the same PCR I cycle number were positioned next to each other – with overlap – for clarity. For selection rounds 1-4, the number of PCR1 cycles was kept to ~18 by adding a small amount of random library (~1/30,000 of the main library) into the incubation with cTmp. In the absence of this addition, the number of PCR I cycles was at least 25 and led to the enrichment of PCR artifacts. In selection round 4, a parallel round was performed in which the addition of random library was omitted. At this selection round, no PCR artifacts were observed, and the addition of random library was omitted for selection rounds 4 and 5. In selection round 5, 22 sub-rounds were performed that differed in the presence of peptides: One subround contained no peptides, one sub-round contained all ten peptides, ten sub-rounds contained individual peptides at 1 mM concentration, and ten sub-rounds contained individual peptides at 10 mM concentration during the incubation with cTmp.

Figure S2



**Figure S2 Effect of peptides on RNA sequence enrichment. (A)** Correlation plot between two different peptide concentrations (1 mM and 10 mM) for the enrichment of RNA sequences in the presence of individual peptides. Each diamond represents a single RNA sequence (the peak sequence of a cluster) and its enrichment with one peptide at two different peptide concentrations. The majority of symbols is near the graph's origin, suggesting no strong influence of peptides or their concentration on RNA enrichment under selection conditions. (B) The table below the graph displays the number and percentage of sequence / peptide pairs in each quadrant of the plot in (A). Pairs that lie exactly on one of the axes (i.e. no peptide effect) are listed separately in the table. Most sequences showed lower enrichment at 10 mM peptide compared to 1 mM peptide. Since the enrichment of sequences is relative between sequences, it is not possible to state whether the absolute activity of RNAs would decrease at higher peptide concentration. However, the data suggest that on average, 10 mM peptide concentrations cause a larger difference in activity between RNA sequences than 1 mM peptide concentrations.

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C0 GAGACCGCAAGAGACGAAAAAAGCACCAGCATTTGGTTGCCATATACTATATCCACATGATATATGTGGTGTGTGGGGAAGACTCGTGTAGGCG
  AAAAGTATCAATATCGCTCCAATGTTTTCCCCGTAAGGAAAACCCTCGGAGCTGACTACTCCTTCTAGCCGACCAATGGTTCAGACTACAAC
TTGTATATATATACACGACAAGATGCACGATAATTGTGGAGATGCGTCACCGCTTGAATACAGGAACATAT CATGGTTCAGACTACAAC
C2 GAGACCGCAAGAGACTCTGCCGCTGTCTGTGGCGAGTGGATTAGGGAAGCCAAAACCCACTTGAGTTTCGTATGAATTATACGGGACCAAAATA
  AAATATCAGGAGTATTGGAAAATGTCAACATCGACAAAGACTGCTTTTATGAGAGGCACTCTTCAATTGCTCATGGTTCAGACTACAAC
C5 GAGACCGCAAGAGACGAAGTCAACTGATGGGGGGGATGGGCTGAACGACTTGAATGCCCCTGTTTGCATCTTGGAGCCTTGAAAAAACTGATAAGC
  C8 GAGACCGCAAGAGACACTAGATGGAAACACACACACACAAGAAGGGGTCTTCAGTGAACACTACCGAGCCTCCAACCGGACGCTGACGTATACGTT
  CGTGGAGAGTGGATTCCATCTGGAAACCTAGTCGGTACCACAATAACTGCATAACGTAAGGAATAAGTTTA CATGGTTCAGACTACAAC
C9 GAGACCGCAAGAGACCAAGGACACATGACAAGGATTTCGTTAAGTGCAACCGTTCGAAATGATAATTGCGGAGATGTTGTTTCACGATTTTCTG
  C10 GAGACCGCAAGAGACTAAGGCTTTGGTGTGGGCGTATACTAACCCTACGTTCCCTTACCAATATCTTGTCTGTAGTTATTGCCCAAACCTAAT
  AGAAAATTAAGGACAGATGCAACCACCCTAATGTTAGGAGGTCATCGGGAAAGCTAAGTGTGTCTGACTTTGCATGGTTCAGACTACAAC
C11 GAGACCGCAAGAGACAAAGGGGATTTGCCGGTCCCGGAATTGTCGAACCAAACGATTTGCGTAAACCGGGAAAAGAATAGCCGGAAATATTGA
  CGAATCTGTCGCGACAACTGCAAATCCCGCGTTTCAGTATATGAATCGGTGTTGTTTGAAATAG CATGGTTCAGACTACAAC
C12 GAGACCGCAAGAGACGTGAATGAGATTTCTTGGGGGGCAAAACCCCCACCACCACTGGTGAAGAGGAGGTGCATTTGTTCCCCAACGATGCTCAAAT
  TATTTTTGGAAAGAATAGCGATCGATTCCTGTGGCCTGATTTTGGAACTATAAGTTTGCTTTGGTATAGATCCATGGTTCAGACTACAAC
C36 GAGACCGCAAGAGACGTATGAACTTTGGATCTACGTAAAGTCATAGGCACGATAAAAACGTGTGCGAGAAGCGGTGTGCCCTTCTTACCTTTG
  TTTTTTGGATTAGGAAAATTACACAATTCAAGAAAATGAGAGTGAAGTACGCATCTAGATGATGATGAATAATCCCATGGTTCAGAACTAACAAC
C37 GAGACCGCAAGAGACTCTCAGAAAGGGGATCATGTTCCCCGATGTTTGAATCCATCAACTGATAAGCTGTACCGAGGAAAGGAAGACTCCGGTT
  GAAACGAAGATCGGAATTAGCATTTCAGTAGATACCTGCAATGAAAGGTTTTGGTCTATAAAGTGCAAGGAG CATGGTTCAGACTACAAC
\texttt{C48} \quad \texttt{GAGACCGCAAGAGAGC} \texttt{CAGTAATACATACATGGATCGCAGGGAATTTTGACCTGATAGGTTCAGTAAAAGCGCTTGGATAGGTTTTCGTATTAG}
  C54 GAGACCGCAAGAGACGAAGAGTTAAGTTGAATTGCCTAAACCTGAGCGACACAGGCACTGAAAACTTCGATCAAGAGATTCTTGAATCTAACC
  CATTCCGAGATCCATAGGACTCAGTTTCTAGATAGTGAAACGTGATTCCTAGGCGAAGTACCAGACTAACTCAGGTTCAGACTACAAC
C64 GAGACCGCAAGAGACTATGTAGCCGTTTAGGTGAAGAAACTGCCAACCATACCGGTCACAATTCACATTGTCAAACCGGCTAAGGTAATAGGT
  AAGAAATGCGCGGGAGAATCTAATACTCACTAAGGGAGTGATGGTCCTTGAACCACACGAACCCACCAGCTA <mark>CATGGTTCAGACTACAAC</mark>
C67 GAGACCGCAAGAGACTTAAAGTATAACAAACATGCGGAGTGTATTTGCTCGTTGTTTTAAGAGGGGTTTAAATGACGAGTCAGGCACGCCAGAA
  C74 GAGACCGCAAGAGAGACACAGATGGGGACTAGAACAAGGTCTGAACGACGTTCTCGGACAGAGGCAGTGCAAAAATCCGTGGGCATGCAATAACT
  C78 GAGACCGCAAGAGACAAATTAATTTCTCTTTAAACCACGTGTTTTAGGAGAGTGTCACAACAAAATGGATGTAGTCTGGATGTGCTAGAGGAACA
  C80 GAGACCGCAAGAGACGAAATAACTTTATTAAACGGATTTTAGGGATGATCTGTTTGTGTAGGGAAGACTTGTGTTGACTCGAGTTGTGAATTTC
  C84 GAGACCGCAAGAGACGTAGTGAACCGTCGGGTTATGATGGAATAAGAGCCATCTTCTGACTAGTTCGACCCCCTTATTCCGAAAATGGGTCAAA
  TAGGGGCGCAATGCTTCCGGGACATGGATGAAGCGAACGTGCTTCATGCAAACCCTGCTGAATGTTCCGTAGCATGGTTCAGACTACAAC
AGTACACGGATCTACAGTTAAAAGTGATCTGTGGAACACGATTGACGGAAAAAAGGTGAAGGACTCTAACCGTCATGGTTCAGACTACAAC
C175 GAGACCGCAAGAGACTGGAAGTTGTGGTGGCGAATCCGAAAACTAAATGGATCGAGTCACTGCCCCTATTTATAAGCAACCGGCGAGGGGAAA
  TAGTAGTACTGAAGGAGCGAATGAGAAGAATCAGGGTAGTAAAGGTAAGATTCACACTGGCCTGAAACTTCGTCATGGTTCAGACTACAAC
C181 GAGACCGCAAGAGACGGAACCTGATGGGCTTTGCAAGAGCTTCGTTTATGCGTAGCACAATATGACAATTGAAGCATATAGTGCTGGGCGCGCA
  ACTAAATGGGCTTGTGCCGTATGACAGTAAAGGACGCGATCCGTTGAAACTACGTACACTGTGTACGAAGGCCCATGGTTCAGACTACAAC
CTAGTTGCTCCTATTGGACCAAATCTTGGGCTGTGCAGAGAGCATAGTCGACATATGGAACATTTCCATAA CATGGTTCAGACTACAAC
C221 GAGACCGCAAGAGACAACGTAACGGTGACATTGCGAGTGGCAATGTGTTTTAGCTAAAAATGTTCCCAACATAAGATATCCGAGGAATTAATG
  GCACTCTGAAAATTTAGATGCCAGGCAGCGGACGGAAGTTGATGAAGTCAACTAATGGTAGAGATAACACTTACAGGTTCAGACTACAAC
AGGGGTCCCGGACGCCTTAATTGCCATCGAAGGCACTACCTGAAATAAAACTGACATAGCCAGAGTGGATAATG CATGGTTCAGACTACAAC
C238_GAGACCGCAAGAGACTATGTATTGGGAACTAGCCAGTGAGCCATAGACTGAAGGTCTGTCGGCTCTATAAGCTGGACTACAAAATCTAGTCC
  CTTGAAAAACTGTCTCTTGTCAACAAGGATTGTTGACAGGAGATGAAGAGCGGAGAAACACCAAGACTGACCTACATGGTTCAGACTACAAC
C308 GAGACCGCAAGAGACTGGAAGTTAGCGGTGGTACTTTATAAGGGGATGGTTAACTAGCATAAACTGTAGAGTGAATTTCTTAACAGGTCGAG
  CTACTTCTTTCTCACTTATAAAGCTAGACTCGAAGACGGGCAGTTTGAACCAATGTTACCATCAAATGGAGTTCAGACTACAAC
C337 GAGACCGCAAGAGACACTGGATTCGTGTTAAGCTAAAATCAGACATCTGCCCTGGCTTGGGACTGGGAAGTTGTCCCCAAAGACTATTTTTTG
  GGGTCTTCCGCGGATCAGGGCTAAACCTCTTTAGATATTAGTATTAATGCACCAAAACGGGCATTACTAAAACCATGGTTCAGACTACAAC
C368 GAGACCGCAAGAGACGAAGGAGTGTACACGTGGGGAGAACTTTCGGGTGTTTCGACTGGAACGTTTGATTGTAAAGACTGCCGATCAATTTT
  TATAAGAGCGCTAGTTCGCATACTCGAATCGTCTCCCTTTGTGAAAGCTGGTATGAACGACGCTAGGCACTCCCATGGTTCAGACTACAAC
C408 GAGACCGCAAGAGACAAAATTATTTTCGAAGACCATGCAGTTAACATACAAGATTATAAATTAGCTAAAATAACTCTTGATATACCTCGACG
  CTAGCACCCAGAACAAGTAGGAGTGATGGCCTCGAAGTAGCGGAAGAGCTAACAAAGTATTGCAAACGTAATACAAGGTACAAAC
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**Figure S3.** Ribozyme sequneces that were tested biochemically. Each sequence is the peak of a sequence cluster that was identified, and found promising for peptide benefit by HTS analysis. For each sequence, the 5'-constant regions (blue) and 3'-constant regions (red) are indicated, with the black sequence stemming from the N150 randomized region.



**Figure S4. Biochemical testing of peptide effects in ribozymes based on predictions from HTS analysis. (A)** Activity assay of 30 ribozymes in the absence, and presence of the one peptide that was predicted from HTS data (figure 2B) to increase ribozyme activity. The incubation with cTmp was done under selection conditions (50 mM Tris/HCl pH 8.3, 100 mM MgCl<sub>2</sub>, 50 mM Na<sub>3</sub>cTmp). Grey columns indicate te activity without peptide, colored columns indicate the activity with peptide. The identity of the peptide, and its concentration in the assay are given in the column labels. Error bars are standard deviations from triplicate experiments. **(B)** Activity assay of six promising ribozymes in the absence, and presence of the same peptide as in (A) but under suboptimal reaction conditions, i.e. at a decreased reaction pH. The incubation with cTmp was done under selection conditions (50 mM HEPES/NaOH pH 7.0, 100 mM MgCl<sub>2</sub>, 50 mM Na<sub>3</sub>cTmp). Grey columns indicate te activity without peptide, colored columns indicate the activity with peptide. The identity of the peptide, and its concentration in the assay are given in the column labels. Error bars are standard deviations from triplicate experiments.



**Figure S5:** Truncation analysis of ribozyme S2. (A) Ribozyme activity shown as function of its length, using truncations at its 3'-terminus. Blue columns indicate the activity with 10 mM peptide P4; gray columns indicate ribozyme activity without peptide. The chosen ribozyme S2T18 (180 nt length) is indicated by an arrow. Error bars are standard deviations from triplicate experiments.





Figure S6. Secondary structure probing of ribozyme S2, using SHAPE. (A) SHAPE profile of ribozyme S2 in the absence of peptide, in the presence of 100 mM MgCl<sub>2</sub>, 50 mM Na<sub>3</sub>cTmp, 50 mM MES/NaOH pH 6.0, 3.3 mM NaOH,1% DMSO (to dissolve 1M7, and preincubated for 2 minutes at 22°C. (B) As in (A) but preincubated for 2 hours. (C) As in (A) but in the presence of 10 mM peptide P4. Note that this peptide preparation increased the pH by about 1. (D) As in (A) but with 10 mM peptide P4, and pre-incubated for 2 hours. Error bars are standard deviations from triplicate experiments. Arrowheads indicate positions where error bars do not overlap with those from the data set (D). (E) Secondary structure model based on the constraints of a 2-hour pre-incubation with 10 mM peptide 4 (B) because we speculated that this data set was likely most consistent with the catalytically active structure.