# Copper(I)-free Syntheses of [<sup>11</sup>C/<sup>18</sup>F]Trifluoromethyl Ketones from Alkyl or Aryl Esters and [<sup>11</sup>C/<sup>18</sup>F]Fluoroform

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# **Supporting Information**

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### **General Information**

Septum-sealed glass vials were used for all reactions. Reagents and solvents were measured for reaction within a glove-box. All the reagents, precursors to be labeled, and reference standards were obtained from commercial sources and were used without further purification. Solvents were purchased in an anhydrous state and the DMF used for radiochemistry was obtained as anhydrous DMF stored on molecular sieves. Retention times ( $t_R$ ) of all standard compounds were determined by reverse-phase HPLC on a UFLC instrument (Shimadzu) using reverse phase analytical HPLC column with an UV absorbance (254 or 295 nm) detector. The column was eluted at 1.5–2.0 mL/min with a gradient elution of acetonitrile (B) and water (A). All the radiochemistry was performed within a lead-shielded hot-cell with remote control to protect personnel from radiation. The apparatus was not fully automated. The HPLC chromatograms were processed using GraphPad Prism 8.

#### **General Procedure for Carbon-11 Labeling**

NCA [<sup>11</sup>C]fluoroform (~ 200–500 MBq) was prepared and collected in DMF (~ 200–300  $\mu$ L) according to the reported method.<sup>1</sup> This was then added to a septum-sealed vial containing *t*-BuOK (15–125  $\mu$ mol) or KHMDS (15–75  $\mu$ mol) in DMF (100  $\mu$ L) at room temperature. The aryl or alkyl esters (from **1a–1o**, **1p**; 50  $\mu$ mol) in DMF (100  $\mu$ L) was then added, mixed vigorously, kept at room temperature for 10 min and shaken frequently. The reaction mixture was then quenched with 4M HCl (100  $\mu$ L), placed on a pre-heated heating block at 60 °C for 5 min and then diluted with H<sub>2</sub>O–MeCN (1:1 v/v; 300  $\mu$ L). An aliquot was then analyzed with reversed phase HPLC as described below. The formation of the radiolabeled compounds ([<sup>11</sup>C]**2a–2o** and [<sup>11</sup>C]**3i**) were confirmed by comparing the retention time of the reference standard compounds (**2a–2o** and **3i**) in analytical HPLC.

#### **General Procedure for Fluorine-18 Labeling**

NCA [<sup>18</sup>F]fluoroform (~ 100–150 MBq) in DMF (~ 200  $\mu$ L) was prepared according to the reported method.<sup>2</sup> This was then added to a septum-sealed vial containing *t*-BuOK (45  $\mu$ mol) in DMF (100  $\mu$ L) at room temperature. The aryl esters (**1a**, **1d** or **1j**; 50  $\mu$ mol) in DMF (100  $\mu$ L) was then added, mixed vigorously, kept at room temperature for 10 min and shaken frequently. The reaction mixture was then quenched with 4M HCl (100  $\mu$ L), placed on a preheated heating block at 60 °C for 5 min and then diluted with H<sub>2</sub>O–MeCN (1:1 v/v; 300  $\mu$ L). An aliquot was then analyzed with reversed phase HPLC as described below. The formation of the radiolabeled compounds ([<sup>18</sup>F]**2a**, [<sup>18</sup>F]**2d** and [<sup>18</sup>F]**2j**) were confirmed by comparing the retention time of the reference standard compounds (**2a**, **2d** and **2j**) in analytical HPLC.

### **Radio-HPLC Analysis**

HPLC was performed on a system comprising a Gold 126 solvent module (Beckman Coulter; Fullerton, CA) and a 166 or 168 UV absorbance detector (operating at 254 or 295 nm) plus a photomultiplier tube radioactivity detector (Flow-count; Bioscan, Washington, DC). For <sup>11</sup>C/<sup>18</sup>F-labeled compounds, an aliquot of the quenched reaction mixture was injected onto a reversed phase column to separate the product radioactive peak from its starting material using the HPLC conditions A–G as described below. Eluate was monitored for radioactivity and for absorbance at

254 nm. Radiochemical product identities were confirmed by comparison of HPLC retention times with the reference compounds. In some cases, appreciable tailing of product peaks was observed, likely as a result of the equilibrium between the ketone and ketal.

### **Condition A**:

Column: Luna C18(2), 10  $\mu$ m, 4.6 × 250 mm, i.d.; Phenomenex

Mobile phase: 10-80% MeCN (B)-H<sub>2</sub>O (A) for 15 min, then 80% B for 5 min; 2.0 mL/min flow rate.

### **Condition B:**

Column: Luna C18(2), 10  $\mu$ m, 4.6 × 250 mm, i.d.; Phenomenex

Mobile phase: 30-80% MeCN (B)-H<sub>2</sub>O (A) for 15 min, then 80% B for 5 min; 2.0 mL/min flow rate.

#### **Condition C:**

Column: XTerra RP<sub>18</sub>, 5  $\mu$ m, 4.6 × 250 mm, i.d.; Waters Corp. Mobile phase: 30–85% MeCN (B)–H<sub>2</sub>O (A) for 15 min, then 85% B for 5 min; 1.5 mL/min flow rate.

#### **Condition D**:

Column: Luna Phenyl-Hexyl, 5  $\mu$ m, 4.6 × 250 mm, i.d.; Phenomenex

Mobile phase: 10-80% MeCN (B)-H<sub>2</sub>O (A) for 15 min, then 80% B for 5 min; 2.0 mL/min flow rate.

## **Condition E**:

Column: Luna Phenyl-Hexyl, 5  $\mu$ m, 4.6 × 250 mm, i.d.; Phenomenex

Mobile phase: 10–50% MeCN (B)– $H_2O$  (A) for 10 min, followed by 50–80% for 5 min and then 80% B for 10 min; 2.0 mL/min flow rate.

# **Condition F**:

Column: Luna C18(2), 10  $\mu$ m, 4.6 × 250 mm, i.d.; Phenomenex

Mobile phase: 10–60% MeCN (B)– $H_2O$  (A) for 10 min, followed by 60–80% for 5 min and then 80% B for 10 min; 2.0 mL/min flow rate.

# **Condition G:**

Column: Luna C18(2), 10  $\mu$ m, 4.6 × 250 mm, i.d.; Phenomenex

Mobile phase: 10-80% MeCN (B)-0.5% TEA in H<sub>2</sub>O (A) for 15 min, then 80% B for 5 min; 2.0 mL/min flow rate.

# Experimental Procedure for the Determination of Molar Activity (A<sub>m</sub>) of [<sup>11</sup>C]2e

NCA [<sup>11</sup>C]fluoroform (~ 2.75 GBq) was prepared from a 15 x 15  $\mu$ A x min of <sup>11</sup>CH<sub>4</sub> (~ 14 GBq) and collected in DMF (~ 300  $\mu$ L) according to the reported method.<sup>1</sup> [<sup>11</sup>C]Fluoroform (~ 2.10 GBq, 250  $\mu$ L) was then added to a septum-sealed vial containing *t*-BuOK (45  $\mu$ mol) in DMF (100  $\mu$ L) at room temperature. The methyl 3-bromobenzoate (**1e**, 50  $\mu$ mol) in DMF (100  $\mu$ L) was added, mixed vigorously, kept at room temperature for 10 min, and shaken frequently. The reaction mixture was then quenched with 4M HCl (100  $\mu$ L), placed on a pre-heated

heating block at 60 °C for 5 min, diluted with H<sub>2</sub>O–MeCN (1:1 v/v; 3.4 mL, total volume 4 mL), and injected onto a reversed phase semi-preparative HPLC (XTerra RP<sub>18</sub>, 10  $\mu$ m, 7.8 × 300 mm, i.d.; Waters Corp.) with isocratic elution of 35% MeCN (B)–65% H<sub>2</sub>O (A) at 6.0 mL/min flow rate. Eluate was monitored for radioactivity and for absorbance at 254 nm. The desired product ([<sup>11</sup>C]**2e**) was collected between 9–12 min in an isolated yield of 83% (decay-corrected to the time of [<sup>11</sup>C]fluoroform addition into reaction vial; ~ 35 min; [<sup>11</sup>C]fluoromethane was excluded from calculation). The total synthesis time was 60 min.

An aliquot of the collected product was then analyzed with reversed phase analytical HPLC (XTerra RP<sub>18</sub>, 5  $\mu$ m, 4.6 × 250 mm, i.d.; Waters Corp.) with isocratic elution of 35% MeCN (B)–65% H<sub>2</sub>O (A) at 1.5 mL/min flow rate. Eluate was monitored for radioactivity and for absorbance at 200 nm. The radio-HPLC system for analysis of [<sup>11</sup>C]**2e** was calibrated for UV absorption response versus mass of reference compound in the analyte (Page S32). The radiolabeled product ([<sup>11</sup>C]**2e**) was obtained with a radiochemical purity of 97.5% and a molar activity ( $A_m$ ) of 56 ± 6 GBq/ $\mu$ mol (n = 2) at the end of synthesis (EOS) (see  $A_m$  calculation in Page S32). The formation of the radiolabeled compounds ([<sup>11</sup>C]**2e**) was confirmed by co-injection with the reference **2e** in analytical HPLC using the conditions described above.

### References

- 1. Haskali, M. B.; Pike, V. W. Chem. Eur. J. 2017, 23, 8156.
- 2. Yang, B. Y.; Telu, S.; Haskali, M. B.; Morse, C. L.; Pike, V. W. Sci. Rep. 2019, 9, 14835.

HPLC Chromatograms for <sup>11</sup>CHF<sub>3</sub> and CHF<sub>2</sub><sup>18</sup>F

















HPLC Chromatograms for [<sup>11</sup>C]2a-2o and 2a-2o























































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# HPLC Conditions:

Column: XTerra RP<sub>18</sub>, 10  $\mu$ m, 7.8 × 300 mm, i.d.; Waters Corp.

Mobile Phase: 35% MeCN (B)-65% H<sub>2</sub>O (A), isocratic, 6.0 mL/min flow rate.

Detection: Absorbance at 254 nm.



# Analytical HPLC Chromatogram for 2e

# HPLC Conditions:

Column: XTerra RP<sub>18</sub>, 5  $\mu$ m, 4.6 × 250 mm, i.d.; Waters Corp.

Mobile Phase: 35% MeCN (B)–65% H<sub>2</sub>O (A), isocratic, 1.5 mL/min flow rate.

Detection: Absorbance at 200 nm.



HPLC Conditions:

Column: XTerra RP<sub>18</sub>, 5  $\mu$ m, 4.6 × 250 mm, i.d.; Waters Corp.

Mobile Phase: 35% MeCN (B)–65% H<sub>2</sub>O (A), isocratic, 1.5 mL/min flow rate.

Detection: Absorbance at 200 nm.



# Analytical HPLC Chromatogram for Co-injection of [<sup>11</sup>C]2e and 2e

HPLC Conditions:

Column: XTerra RP<sub>18</sub>, 5  $\mu$ m, 4.6 × 250 mm, i.d.; Waters Corp.

Mobile Phase: 35% MeCN (B)–65% H<sub>2</sub>O (A), isocratic, 1.5 mL/min flow rate.

Detection: Absorbance at 200 nm.



# Calibration Curve and Molar Activity $(A_m)$ Calculation for $[^{11}C]2e$

Stock solution: 1 mg of 2e in 10 mL acetonitri	Stock solution: 1 mg of 2e in 10 mL acetonitrile:		0.1 mg/mL			
Solution for injection: 300 $\mu$ L stock diluted to	10 mL in 1:1	MeCN-water:	0.003 mg/mL			0
	1.00 million (1.00 million)				Br	
Analytical HPLC column: XTerra, RP <sub>18</sub> , 5 /	u, 250 x 4.6 r	nm (i.d. Waters	Corp.)		Ĩ	J 43
Mobile phase: 35%B (MeCN)-65%A (H <sub>2</sub> C	), isocratic, 1	.5 mL/min, 200	nm UV			
						2e
Weight (ng)	AUC					
30	138095.0		Calibration Curve to Determine	Molar Activit	y of [11C]	2e
60	261300.0	1200000	0			
90	401724.0	1200000.		y = 4017.1	x	
135	562293.0	1000000.	0	$R^2 = 0.998$	5	•
195	761180.0			1		
270	1070755.0	800000.	0			
	252.02	B 600000	0	and the second		2
MW of Ze (g/mol):	253.02	Ā				
QC area under the curve (AUC, 200 mm)	15584.00	400000.	0			
Injected activity ( $\mu$ Ci)	20.80	-				
Mass $(ng) = AUC / slope$	3.88	200000.	0			
Molar mass (nmol) = mass (ng) / MW	0.015					
Molar activity ( $A_m$ ) (mCi/ $\mu$ mol)	1356.60	0.	0 50 100 15	0 2.00	2.50	300
Molar activity ( $A_{\rm m}$ ) (GBq/ $\mu$ mol) at EOS	50.19		Weigh	t (ng)	0.000	
Molar activity (A m) (GBq/ $\mu$ mol) at EOB	379.00			(8/		
MW of <b>2e</b> (g/mol):	253.02					
QC area under the curve (AUC, 200 nm)	22007.00					
Injected activity (µ Ci)	36.30					
Mass (ng) = AUC / slope	5.48		Average A <sub>m</sub> (GBq/μmol) at EOS	SD (n = 2)		
Molar mass (nmol) = mass (ng) / MW	0.022		60 min after EOB			
Molar activity (A m) (mCi/ $\mu$ mol)	1676.54					
Molar activity (A $_{\rm m}$ ) (GBq/ $\mu$ mol) at EOS	62.03		56.11	5.92		
Molar activity (A m) (GBq/ $\mu$ mol) at EOB	495.00					