Supporting Information

Visible-light-induced synthesis of heteroaryl C-glycosides via

decarboxylative C-H glycosylation

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1. General information

Unless otherwise noted, all reagents were obtained from commercial suppliers and used without further purification. All of the reactions were carried out in pressure tubes. Except for the specially mentioned dry solvent, all the solvents were treated according to general methods. All the reactions were monitored by thin-layer chromatography (TLC) and were visualized using UV light. Product purification was done using silica gel column chromatography. Thin-layer chromatography (TLC) characterization was performed with precoated silica gel GF254 (0.2 mm), while column chromatography characterization was performed with silica gel (100-200 mesh). ¹H NMR and ¹³C NMR spectra were recorded with tetramethylsilane (TMS, $\delta = 0.00$ ppm) as the internal standard. ¹H NMR spectra were recorded at 400 or 600 MHz (Varian) and ¹³C NMR spectra were recorded at 100 or 150 MHz (Varian). Shifts are reported in ppm downfield from CDCl₃ (δ =7.26 ppm) or DMSO-d₆ (δ = 2.50 ppm; H₂O signal was found at δ = 3.34 ppm) for ¹H NMR and chemical shifts for ¹³C NMR spectra are reported in ppm relative to the central CDCl₃ $(\delta = 77.0 \text{ppm})$ or DMSO-d₆ ($\delta = 39.6 \text{ ppm}$). Coupling constants were given in Hz. The following notations were used: br-broad, s-singlet, d-doublet, t-triplet, q-quartet, m-multiplet, dd-doublet of doublet, dt-doublet of triplet, td-triplet of doublet. HRMS spectra were recorded a MicrOTOF-QIII(Bruker.Daltonics). Melting points were measured with YRT-3 melting point apparatus (Shantou Keyi Instrument &Equipment Co., Ltd., Shantou, China). Quinoxalin-2(1H)-ones and glycosyl NHP esters were synthesized according to the literature.

2. Condition Optimization

Table 1. Optimization of reaction conditions for 3aa.^a



Entry	Light source	Solvent	Time(h)	Yield(%) ^b
1	390-395nm	DMSO	24	65
2	420-425nm	DMSO	24	77
3	425-430nm	DMSO	24	79
4	450-455nm	DMSO	24	91
5	520-525nm	DMSO	24	82
6	427nm Kessil lamps	DMSO	24	75
7	456nm Kessil lamps	DMSO	24	84

8	Dark conditions	DMSO	24	N.R.	
9	450-455nm	DCE	24	47	
10	450-455nm	DMA	24	64	
11	450-455nm	DMF	24	71	
12	450-455nm	MeCN	24	41	
13	450-455nm	Dioxane	24	75	
14	450-455nm	THF	24	25	
15	450-455nm	Tol	24	36	
16 ^c	450-455nm	DMSO	24	77	
17	450-455nm	DMSO	36	91	
18	450-455nm	DMSO	12	71	
19 ^d	450-455nm	DMSO	24	91	

^{*a*} Reaction condition: **1a** (0.1 mmol), **2a** (0.12 mmol), solvent (1.0ml), at rt for 24 h under Ar. ^{*b*} Isolated yield. ^{*c*} Open to air. ^{*d*} **2a** (0.15 mmol). N.R.= No Reaction

3. Experimental Information

3.1 Synthesis of Quinoxalin-2(1H)-ones (1a as an example)

$$R_{1} \stackrel{H}{\stackrel{}_{\cup}} \stackrel{V}{\stackrel{}_{\vee}} \stackrel{V}{\stackrel{}_{\vee}} \stackrel{V}{\stackrel{}_{\vee}} \stackrel{K_{2}}{\stackrel{}_{\vee}} \stackrel{K_{2}}{\stackrel{}_{\vee} \stackrel{K_{2}}{\stackrel{}_{\vee}} \stackrel{K_{2}} \stackrel{K_{2}} \stackrel{K_{2}}{\stackrel{}_{\vee} } \stackrel{K_{2}} \stackrel{K_{2}} \stackrel{K_{2}} \stackrel{K_{2}} \stackrel{$$

Following a literature procedures ¹, to a 100 mL round bottom flask with a stir bar was added *N*-free quinoxalin-2(1*H*)-one derivatives (6.8 mmol), DMF (15 mL) and K₂CO₃ (8.2 mmol), followed by dropwise addition of alkyl halide (10.9 mmol). The reaction mixture was stirred for 1-12 h at room temperature. Then reaction mixture was partitioned in water and EtOAc, and extracted with EtOAc thrice. The combined EtOAc extracts were dried over Na₂SO₄, filtered and concentrated. The residue was purified by column chromatography (PET ether/ EtOAc) to afford the desired *N*-alkylated quinoxalin-2(1*H*)-ones.

3.2 Examples of general procedures for the synthesis of carboxylic acid



Scheme S1. Procedure for the synthesis of ribose-derived carboxylic acid.

Step 1^[2]: To a 250 mL round bottom flask was added D-ribose (10 g, 66 mmol, 1.0 equiv.), methanol (40 mL), and acetone (40 mL). Next, concentrated HCl (1.0 mL) was added at room temperature, the flask was heated to 75 °C for 4 h. The solution was cooled to room temperature and filtered to remove the solid then sat. Na₂CO₃ was added to adjust pH above 7. The solution was extracted by ethyl acetate (50 mL \times 2), washed by brine (50 mL \times 2) , dried by Na₂SO₄, combined the organic phase and concentrated with reduced pressure. The desired crude product was obtained as a faint yellow oil (10 g, 80% yield).

Step 2^[3]: The acid was prepared according to the reported paper. TEMPO (0.63 g, 4.0 mmol, 0.20 equiv.) and PhI(OAc)₂ (19 g, 60 mmol, 3.0 equiv.) were added to a solution of alcohol obtained from above step (4.1 g, 20 mmol, 1.0 equiv.) in MeCN/H₂O (50 mL/50 mL, 0.20 M) at room temperature. The reaction mixture was stirred for 9 h at room temperature, and then saturated aqueous Na₂S₂O₃ (5.0 mL) was added. After being stirred for 20 min at room temperature, the resultant mixture was extracted with CH₂Cl₂ (30 mL × 2). The combined organic layers were dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel using hexane/EtOAc as eluant to afford carboxylic acid (3.8 g, 89% yield).



Scheme S2. Procedure for the synthesis of Bn/Me-protected ribose-derived carboxylic acid. Step 1^[4]: To a solution of methyl beta D-ribofuranoside (5.0 g, 30 mmol, 1.0 equiv) in pyridine (60 mL, 0.50 M) was added trityl chloride (9.2 g, 33 mmol, 1.1 equiv) and the reaction mixture was stirred at room temperature overnight. The solvent was removed in vacuo and the residue was coevaporated with toluene, dissolve in EtOAc (90 mL) and washed with 1.0 M aqueous CuSO₄ solution (50 mL \times 3), brine (50 mL \times 2), dried over Na₂SO₄, filtered, and concentrated to afford desired product. Purification by flash column chromatography (cyclohexane/EtOAc) yielded product as a white foam (12g, 86% yield).

Step 2^[5]: The obtained compound (5.0 g, 12 mmol, 1.0 equiv) from above step was dissolved in DMF (60 mL, 0.20 M) and cooled to 0 °C. Sodium hydride (60% in mineral oil, 1.9 g, 48 mmol, 4.0 equiv) was added portion-wise. Then, BnCl/MeI (4.3 mL/2.3 mL, 36mmol, 3.0 equiv) was added dropwise. The mixture was stirred overnight and allowed to warm to room temperature. The reaction was quenched with MeOH and thiourea, and stirred for another 2 h at room temperature. The solvents were evaporated in vacuo and the residue was dissolved in EtOAc (50 mL) and then washed with water (50 mL \times 2). The aqueous phase was extracted with EtOAc (30 mL), the combined organic phases were washed with brine (50 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo to obtain crude product Purification by flash column chromatography (cyclohexane/CH₂Cl₂/EtOAc) yielded product as a white foam (5.6 g, 80% yield).

Step 3^[3]: The obtained compound (take the Bn-protected as an example, 5.0 g, 8.5 mmol, 1.0 equiv) from above step was dissolved in MeOH/Et₂O/H₂O (42 mL/4.2 mL/0.42 mL, 0.20 M) followed by addition of TsOH (0.73 g, 4.3 mmol, 0.50 equiv). After being stirred for 20 min at

room temperature, the resultant mixture was extracted with EtOAc (30 mL \times 3). The combined organic layers were dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel using hexane/EtOAc as eluent to get the alcohol (2.3 g, 80% yield).

Step 4^[3]: The acid was prepared according to the reported paper. TEMPO (0.18 g, 1.2 mmol, 0.20 equiv.) and PhI(OAc)₂ (5.6 g, 17 mmol, 3.0 equiv.) were added to a solution of alcohol obtained from above step (take the Bn-protected as an example, 2.0 g, 5.8 mmol, 1.0 equiv.) in MeCN/H₂O (15 mL/15 mL, 0.20 M) at room temperature. The reaction mixture was stirred for 9 h at room temperature, and then saturated aqueous Na₂S₂O₃ (5.0 mL) was added. After being stirred for 20 min at room temperature, the resultant mixture was extracted with CH₂Cl₂ (30 mL×3). The combined organic layers were dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel using hexane/EtOAc as eluant to afford carboxylic acid (1.6 g, 78% yield).



Scheme S3. Procedure for the synthesis of arabinose-derived carboxylic acid.

Step 1^[6]: To a solution of beta D-arabinose (8.0 g, 53 mmol, 1.0 equiv) in DMF(0.27 L, 0.20 M) was added imidazole (5.4 g, 80 mmol, 1.5 equiv) at 50 °C. Then TBDPSCl (21 mL,80 mmol, 1.5 equiv) was added dropwise. The mixture was stirred at 50 °C overnight . The solvents were evaporated in vacuo and the residue was dissolved in EtOAc (90 mL), and then washed with water (50 mL \times 2). The aqueous phase was extracted with EtOAc (30 mL), the combined organic phases were washed with brine (50 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo to obtain crude product Purification by flash column chromatography (cyclohexane/ EtOAc) yielded product as a colorless oil (17 g, 83% yield).

Step 2^[7]: The obtained compound (11 g, 28 mmol, 1.0 equiv) from above step was dissolved in acetone (30 mL) and 2, 2-dimethoxypropane (10 mL) and *p*-toluenesulfonic acid (0.20 g) was added. The reaction was neutralized by triethylamine and concentrated after stired 30 min at room temperature. The solvents were evaporated in vacuo, purification by flash column chromatography (cyclohexane / EtOAc). yielded product as a colorless oil (10 g, 84% yield).

Step 3^[5]: The obtained compound (10 g, 23 mmol, 1.0 equiv) from above step was dissolved in DMF (0.12 L, 0.20 M) and cooled to 0 °C. Sodium hydride (60% in mineral oil, 1.8 g, 46 mmol, 2.0 equiv) was added portion wise. Then, BnCl (4.1 mL, 35 mmol, 1.5 equiv) was added dropwise. The mixture was stirred overnight and allowed to warm to room temperature. The reaction was quenched with MeOH and thiourea, and stirred for another 2 h at room temperature. The solvents were evaporated in vacuo and the residue was dissolved in EtOAc (90 mL), and then washed with water (50 mL \times 2). The aqueous phase was extracted with EtOAc (30 mL), the combined organic phases were washed with water, dried over Na₂SO₄, filtered, and concentrated in vacuo to obtain crude product as an oil. Purification by flash column chromatography (cyclohexane / EtOAc). yielded product as a white solid (10 g, 87% yield).

Step 4^[6]: The obtained compound (10 g, 19 mmol, 1.0 equiv) from above step was dissolved in anhydrous THF (40 mL, 0.50 M), was reacted with TBAF (1.0 M in THF, 23 mL,1.2 equiv). The solution was stirred at room temperature for 2 h. The solvents were evaporated in vacuo, purification by flash column chromatography (cyclohexane / EtOAc). yielded product as a white solid (4.2 g, 79% yield).

Step 5^[3]: The acid was prepared according to the reported paper. TEMPO (0.17 g, 0.20 equiv.) and PhI(OAc)₂ (5.2 g, 3.0 equiv.) were added to a solution of alcohol (1.5 g, 5.4 mmol, 1.0 equiv.) obtained from above step (1.0 equiv.) in MeCN/H₂O (15 mL/15 mL, 0.20 M) at room temperature. The reaction mixture was stirred for 9 h at room temperature, and then saturated aqueous Na₂S₂O₃ (5.0 mL) was added. After being stirred for 20 min at room temperature, the resultant mixture was extracted with CH₂Cl₂ (30 mL \times 3). The combined organic layers were dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel using hexane/EtOAc as eluant to afford carboxylic acid 78% yield (white solid, 1.2 g).



Scheme S4. Procedure for the synthesis of xylose-derived carboxylic acid.

Step 1^[4]: To a solution of 1,2-O-Isopropylidene-alpha-D-xylofuranose (5.0 g, 26 mmol, 1.0 equiv) in pyridine (52 mL,0.50 M) was added trityl chloride (8.0 g, 29 mmol, 1.1 equiv) and the reaction mixture was stirred at room temperature overnight. The solvent was removed in vacuo and the residue was coevaporated with toluene, dissolve in EtOAc (90 mL) and washed with 1 M aqueous CuSO₄ solution (50 mL \times 3), brine (50 mL \times 2), dried over Na₂SO₄, filtered, and concentrated to afford desired product. Purification by flash column chromatography (cyclohexane/EtOAc) yielded product as a white foam (8.7 g, 83% yield).

Step 2^[5]: The obtained compound (6.0 g, 14 mmol, 1.0 equiv) from above step was dissolved in DMF (70 mL, 0.2 M) and cooled to 0 °C. Sodium hydride (60% in mineral oil, 1.1 g, 28 mmol, 2.0 equiv) was added portion-wise. Then, BnCl (2.5 mL, 21 mmol, 1.5 equiv) was added dropwise. The mixture was stirred overnight and allowed to warm to room temperature. The reaction was quenched with MeOH and thiourea, and stirred for another 2 h at room temperature. The solvents were evaporated in vacuo and the residue was dissolved in EtOAc (90 mL), and then washed with water (50 mL \times 2). The aqueous phase was extracted with EtOAc (30 mL), the combined organic phases were washed with brine (50 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo to obtain crude product Purification by flash column chromatography (cyclohexane/CH₂Cl₂/EtOAc) yielded product as a white foam (5.9 g, 81% yield).

Step 3^[3]: The obtained compound (5.4 g, 10 mmol, 1.0 equiv) from above step was dissolved in MeOH/Et₂O/H₂O (50 mL/5.0 mL/0.50 mL, 0.20 M) followed by addition of TsOH (0.86 g, 5.0 mmol, 0.50 equiv). After being stirred for 20 min at room temperature, the resultant mixture was extracted with EtOAc (30 mL \times 3). The combined organic layers were dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel using hexane/EtOAc as eluent to get the alcohol (2.3g, 85%).

Step 4^[3]: The acid was prepared according to the reported paper. TEMPO (0.17 g, 1.1 mmol, 0.20 equiv.) and PhI(OAc)₂ (5.2 g, 16 mmol, 3.0 equiv.) were added to a solution of alcohol (1.5 g, 5.4 mmol, 1.0 equiv.) obtained from above step in MeCN/H₂O (15 mL/15 mL, 0.20 M) at room temperature. The reaction mixture was stirred for 9 h at room temperature, and then saturated aqueous Na₂S₂O₃ (5.0 mL) was added. After being stirred for 20 min at room temperature, the resultant mixture was extracted with CH₂Cl₂ (30 mL × 3). The combined organic layers were dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel using hexane/EtOAc as eluant to afford carboxylic acid 78% yield (1.2 g).



Scheme S5. Procedure for the synthesis of Bn/Me-protected hexose-derived carboxylic acid. Step 1^[4]:To a solution of methyl- α -D-glucopyranoside (10 g, 51 mmol, 1.0 equiv) in pyridine (0.10 L, 0.50 M) was added trityl chloride (16 g, 56 mmol, 1.1 equiv) and the reaction mixture was stirred at room temperature overnight. The solvent was removed in vacuo and the residue was coevaporated with toluene, dissolve in EtOAc (90 mL) and washed with 1 M aqueous CuSO₄ solution (50 mL \times 3), brine (50 mL \times 2), dried over Na₂SO₄, filtered, and concentrated to afford desired product. Purification by flash column chromatography (cyclohexane/EtOAc) yielded product as a white foam (17 g, 80% yield).

Step.2^[2,5]: The obtained compound (8.0 g, 18 mmol, 1.0 equiv.) from above step was dissolved in DMF (90 mL, 0.20 M) and cooled to 0 °C. Sodium hydride (60% in mineral oil, 4.3 g, 0.11 mol 6.0 equiv) was added portion-wise. Then, BnCl (9.6 mL, 81 mmol, 4.5 equiv) /MeI was added dropwise. The mixture was stirred overnight and allowed to warm to room temperature. The reaction was quenched with MeOH and thiourea, and stirred for another 2 h at room temperature. The solvents were evaporated in vacuo and the residue was dissolved in EtOAc (90 mL), and then washed with water (50 mL \times 2). The aqueous phase was extracted with EtOAc (30mL), the combined organic phases were washed with brine (50mL), dried over Na₂SO₄, filtered, and concentrated in vacuo to obtain crude product Purification by flash column chromatography (cyclohexane/CH₂Cl₂/EtOAc) yielded product as a white foam (9.9 g, 78% yield).

Step.3^[3]: The obtained compound (8.0 g, 11 mmol, 1.0 equiv.) from above step was dissolved in MeOH/Et₂O/H₂O (50 mL/5.0 mL/0.50 mL, 0.20 M) followed by addition of TsOH (0.95 g, 5.5 mmol, 0.50 equiv). After being stirred for 20 min at room temperature, the resultant mixture was extracted with EtOAc (30 mL \times 3). The combined organic layers were dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel using hexane/EtOAc as eluent to get the alcohol (4.2 g, 82% yield).

Step.4^[3]: The acid was prepared according to the reported paper. TEMPO (0.20 g, 1.3 mmol, 0.20 equiv.) and PhI(OAc)₂ (6.3 g, 20 mmol, 3.0 equiv.) were added to a solution of alcohol (3.0g 6.5 mmol, 1.0 equiv.) obtained from above step in MeCN/H₂O (16 mL/16 mL, 0.20 M) at room temperature. The reaction mixture was stirred for 9 h at room temperature, and then saturated aqueous Na₂S₂O₃ (5.0 mL) was added. After being stirred for 20 min at room temperature, the

resultant mixture was extracted with CH_2Cl_2 (30 mL \times 3). The combined organic layers were dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel using hexane/EtOAc as eluant to afford carboxylic acid (2.4 g, 80% yield).



Scheme S6. Procedure for the synthesis of fructose-derived carboxylic acid.

(3aR,5aR,8aR,8bS)-2,2,7,7-tetramethyltetrahydro-3aH-bis([1,3]dioxolo)[4,5-b:4',5'-d]pyran-3a-carboxylic acid^[3]: To a 50 mL flask, the diacetonefructose (3.0 g, 12 mmol, 1.0 equiv) was treated with TEMPO (0.36 g, 2.3 mmol, 0.20 equiv.) and PhI(OAc)₂ (11 g, 35 mmol, 3.0 equiv.) in MeCN/H₂O (30 mL/30 mL, 0.20 M) at room temperature for 9 h, and then saturated aqueous Na₂S₂O₃ (5.0 mL) was added. After being stirred for 20 min at room temperature, the resultant mixture was extracted with CH₂Cl₂ (30 mL \times 3). The combined organic layers were dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel using hexane/EtOAc as eluant to afford carboxylic acid(2.6 g, 83%).



Scheme S7. Procedure for the synthesis of galactose-derived carboxylic acid.

((3aR,5R,5aS,8aS,8bR)-2,2,7,7-tetramethyltetrahydro-5H-bis([1,3]dioxolo)[4,5-b:4',5'-d]pyra n-5-yl)methanol^[8]:To a round bottom flask was added D-galactose (7.0 g, 39 mmol, 1.0 equiv.), acetone (0.25 L). Next, concentrated H₂SO₄ (7.7 mL) was added at 0 °C. The reaction mixtures were stirred at room temperature for 5 h and then neutralized by the addition of sat. Na₂CO₃ until pH = 7. The precipitate was removed by filtration and the filtrates were combined and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel using hexane/EtOAc as eluent to get the product as a colorless oil.(8.5 g, 84%).

(3aR,5S,5aR,8aS,8bR)-2,2,7,7-tetramethyltetrahydro-5H-bis([1,3]dioxolo)[4,5-b:4',5'-d]pyra n-5-carboxylic acid^[3]: The acid was prepared according to the reported paper. TEMPO (0.84 g, 5.4 mmol, 0.20 equiv.) and PhI(OAc)₂ (26 g, 81 mmol, 3.0 equiv.) were added to a solution of alcohol (7.0 g, 27 mmol, 1.0 equiv.) obtained from above step in MeCN/H₂O (67 mL/67 mL, 0.20 M) at room temperature. The reaction mixture was stirred for 9 h at room temperature, and then saturated aqueous Na₂S₂O₃ (5.0 mL) was added. After being stirred for 20 min at room temperature, the resultant mixture was extracted with CH₂Cl₂ (30 mL \times 3). The combined organic layers were dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel using hexane/EtOAc as eluant to afford carboxylic acid (5.1 g, 87% yield).

3.3 Synthetic procedure of N-hydroxyphthalimide esters



Scheme S8. Procedure for the synthesis of N-hydroxyphthalimide esters.

A round-bottom flask or culture tube was charged with carboxylic acid derived from sugar (1.0 equiv), N-hydroxyphthalimide (1.1 equiv) and 4-dimethylaminopyridine (0.050 equiv). Dichloromethane was added (0.10 - 0.20 M), and the mixture was stirred vigorously. Dicyclohexylcarbodiimide (DCC, 1.1 equiv) was added, and then the mixture was allowed to stir until the acid was consumed (determined by TLC). Typical reaction times were between 0.5 h to 12 h. The mixture was filtered through a thin pad of Celite and rinsed with additional CH_2Cl_2 . The solvent was removed under reduced pressure, and purification of the crude mixture by column chromatography (DCM/hexane/ethyl acetate eluent) afforded as the desired N-hydroxyphthalimide esters.



1,3-dioxoisoindolin-2-yl(3aS,4S,6R,6aR)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-d][1,3]di oxole-4-carboxylate (2a): ¹**H-NMR (400 MHz, Chloroform-***d***)** δ 7.89 (dd, J = 5.5, 3.1 Hz, 2H), 7.80 (dd, J = 5.5, 3.1 Hz, 2H), 5.36 (d, J = 5.8 Hz, 1H), 5.14 (s, 1H), 5.00 (s, 1H), 4.66 (d, J = 5.8 Hz, 1H), 3.50 (s, 3H), 1.52 (s, 3H), 1.35 (s, 3H). The compound was identified by spectral comparison with literature data.^[5]



1,3-dioxoisoindolin-2-yl(2S,3S,4R,5R)-3,4,5-trimethoxytetrahydrofuran-2-carboxylate (2b): ¹H-NMR (600 MHz, Chloroform-*d*) δ 7.93 – 7.85 (m, 2H), 7.85 – 7.76 (m, 2H), 5.05 (s, 1H), 4.85 (d, *J* = 4.5 Hz, 1H), 4.47 (s, 1H), 3.83 (s, 1H), 3.57 (s, 3H), 3.53 (s, 3H), 3.46 (s, 3H). The compound was identified by spectral comparison with literature data. ^[5]



1,3-dioxoisoindolin-2-yl(2S,3S,4R,5R)-3,4-bis(benzyloxy)-5-methoxytetrahydrofuran-2-carbo xylate (2c): ¹**H-NMR (400 MHz, Chloroform-***d***)** δ 7.90 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.80 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.42 – 7.28 (m, 10H), 5.00 (d, *J* = 6.6 Hz, 2H), 4.79 (d, *J* = 11.7 Hz, 1H), 4.71 –

4.60 (m, 4H), 3.87 (d, J = 4.5 Hz, 1H), 3.42 (s, 3H). The compound was identified by spectral comparison with literature data.^[5]



1,3-dioxoisoindolin-2-yl(3aS,5S,6S,6aS)-6-(benzyloxy)-2,2-dimethyltetrahydrofuro[2,3-d][1,3] dioxole-5-carboxylate (2d): ¹H-NMR (400 MHz, Chloroform-*d*) δ 7.90 (dd, J = 5.5, 3.1 Hz, 2H), 7.80 (dd, J = 5.5, 3.1 Hz, 2H), 7.39 – 7.28 (m, 5H), 6.07 (d, J = 3.4 Hz, 1H), 5.07 (d, J = 0.8 Hz, 1H), 4.78 – 4.72 (m, 2H), 4.71 – 4.65 (m, 2H), 1.50 (s, 3H), 1.33 (s, 3H). ¹³C-NMR (100 MHz, Chloroform-*d*) δ 166.3, 161.4, 136.5, 134.9, 128.8, 128.7, 128.3, 128.0, 124.1, 113.8, 107.0, 84.4, 82.9, 80.7, 72.5, 25.9, 25.7. HRMS (ESI) m/z [M + Na]⁺ calculated for 462.1159, found 462.1161.



1,3-dioxoisoindolin-2-yl(3aR,5S,6R,6aR)-6-(benzyloxy)-2,2-dimethyltetrahydrofuro[2,3-d][1, 3]dioxole-5-carboxylate (2e): ¹**H-NMR (400 MHz, Chloroform-***d***)** δ 7.91 (dd, J = 5.4, 3.1 Hz, 2H), 7.80 (dd, J = 5.4, 3.1 Hz, 2H), 7.46 – 7.27 (m, 5H), 6.11 (d, J = 3.4 Hz, 1H), 5.20 (d, J = 3.5 Hz, 1H), 4.92 (d, J = 11.9 Hz, 1H), 4.69 (d, J = 11.9 Hz, 1H), 4.56 (d, J = 3.4 Hz, 1H), 4.46 (d, J = 3.5 Hz, 1H), 1.51 (s, 3H), 1.32 (s, 3H). The compound was identified by spectral comparison with literature data. ^[5]



1,3-dioxoisoindolin-2-yl(3aR,5S,5aR,8aS,8bR)-2,2,7,7-tetramethyltetrahydro-5H-bis([1,3]dioxolo)[4,5-b:4',5'-d]pyran-5-carboxylate (2f): ¹H-NMR (400 MHz, Chloroform-*d*) δ 7.90 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.79 (dd, *J* = 5.5, 3.1 Hz, 2H), 5.70 (d, *J* = 5.0 Hz, 1H), 4.84 (d, *J* = 2.2 Hz, 1H), 4.74 (qd, *J* = 7.5, 2.4 Hz, 2H), 4.46 (dd, *J* = 5.0, 2.6 Hz, 1H), 1.61 (s, 3H), 1.54 (s, 3H), 1.42 (s, 3H), 1.37 (s, 3H). The compound was identified by spectral comparison with literature data.^[5]



1,3-dioxoisoindolin-2-yl(3aR,5aR,8aR,8bS)-2,2,7,7-tetramethyltetrahydro-3aH-bis([1,3]dioxo lo)[4,5-b:4',5'-d]pyran-3a-carboxylate (2g): ¹**H-NMR (400 MHz, Chloroform-***d***)** δ 7.89 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.79 (dd, *J* = 5.5, 3.1 Hz, 2H), 4.88 (d, *J* = 2.5 Hz, 1H), 4.69 (dd, *J* = 7.9, 2.5 Hz, 1H), 4.31 (d, *J* = 7.9 Hz, 1H), 3.96 (s, 2H), 1.62 (s, 3H), 1.57 (s, 3H), 1.51 (s, 3H), 1.37 (s, 3H). The compound was identified by spectral comparison with literature data.^[5]



1,3-dioxoisoindolin-2-yl(2S,3S,4S,5R,6S)-3,4,5,6-tetramethoxytetrahydro-2H-pyran-2-carbox ylate (2h): ¹**H-NMR (400 MHz, Chloroform-***d***) \delta 7.90 (dd, J = 5.5, 3.1 Hz, 2H), 7.81 (dd, J = 5.5, 3.1 Hz, 2H), 4.95 (d, J = 3.5 Hz, 1H), 4.42 (d, J = 9.9 Hz, 1H), 3.65 (d, J = 1.7 Hz, 6H), 3.57 (d, J = 9.6 Hz, 1H), 3.54 (s, 3H), 3.52 (s, 3H), 3.50 (d, J = 2.9 Hz, 1H), 3.30 (dd, J = 9.5, 3.5 Hz, 1H). The compound was identified by spectral comparison with literature data. ^[5]**



1,3-dioxoisoindolin-2-yl(2S,3S,4S,5R,6S)-3,4,5-tris(benzyloxy)-6-methoxytetrahydro-2H-pyr an-2-carboxylate (2i): ¹**H-NMR (400 MHz, Chloroform-***d***) \delta 7.90 (dd, J = 5.5, 3.1 Hz, 2H), 7.80 (dd, J = 5.5, 3.1 Hz, 2H), 7.43 – 7.26 (m, 15H), 4.99 (d, J = 10.9 Hz, 1H), 4.93 (d, J = 10.1 Hz, 1H), 4.86 (d, J = 11.0 Hz, 2H), 4.82 (d, J = 2.4 Hz, 1H), 4.71 (d, J = 3.4 Hz, 1H), 4.66 (d, J = 12.1 Hz, 1H), 4.58 (d, J = 10.0 Hz, 1H), 4.07 (t, J = 9.2 Hz, 1H), 3.96 – 3.90 (m, 1H), 3.63 (dd, J = 9.6, 3.5 Hz, 1H), 3.49 (s, 3H). The compound was identified by spectral comparison with literature data. ^[5]**



1,3-dioxoisoindolin-2-yl(2S,3S,4S,5R,6S)-6-methoxy-3,4,5-tris((4-methylbenzyl)oxy)tetrahyd ro-2H-pyran-2-carboxylate(2j): ¹**H NMR (400 MHz, Chloroform-***d***) \delta 7.88 (dd, J = 5.5, 3.1 Hz, 2H), 7.78 (dd, J = 5.5, 3.1 Hz, 2H), 7.26 – 7.07 (m, 12H), 4.91 (d, J = 10.6 Hz, 1H), 4.85 – 4.76 (m, 4H), 4.62 – 4.58 (m, 2H), 4.52 (d, J = 10.0 Hz, 1H), 4.01 (t, J = 9.2 Hz, 1H), 3.86 (t, J = 9.4 Hz, 1H), 3.56 (dd, J = 9.6, 3.5 Hz, 1H), 3.45 (s, 3H), 2.35 (s, 3H), 2.34 (s, 3H), 2.30 (s, 3H).**



1,3-dioxoisoindolin-2-yl(2S,3S,4S,5R,6S)-3,4,5-tris((4-bromobenzyl)oxy)-6-methoxytetrahydr o-2H-pyran-2-carboxylate(2k):¹H NMR (400 MHz, Chloroform-*d*) δ 7.90 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.81 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.47 – 7.40 (m, 6H), 7.19 (d, *J* = 8.4 Hz, 2H), 7.14 – 7.12 (m, 4H), 4.85 (d, *J* = 11.1 Hz, 2H), 4.78 (d, *J* = 3.4 Hz, 1H), 4.72 (d, *J* = 11.5 Hz, 1H), 4.69 – 4.65 (m, 2H), 4.60 (d, *J* = 12.2 Hz, 1H), 4.54 (d, *J* = 9.9 Hz, 1H), 3.98 (t, *J* = 9.2 Hz, 1H), 3.86 (dd, *J* = 10.0, 8.8 Hz, 1H), 3.57 (dd, *J* = 9.6, 3.5 Hz, 1H), 3.50 (s, 3H).



3.4 General procedure for the synthesis of desired products 3 and 4 (3aa as an example)

To an oven-dried 10 mL glass tube equipped with a stir bar, was added glycosyl NHP ester (0.12 mmol), *N*-methylquinoxalin-2(1*H*)-one (0.1 mmol). The tube was evacuated and back-filled with Ar (three times), then sealed with rubber stopper and parafilm. Then, anhydrous DMSO (1.0 mL) was added using a syringe. The solution was then stirred at room temperature under the

irradiation of 12 W Blue LEDs for 24 h. After completion of the reaction, 5.0 mL water was added and extracted by ethyl acetate (3×5.0 mL). The combined organic layer was washed with brine (5.0 mL) and then dried over anhydrous Na₂SO₄ and evaporated in vacuum. The desired products were obtained in the corresponding yields after purification by flash chromatography on silica gel eluting with hexane/ethyl acetate or hexane/dichloromethane.

4. Scale-Up Synthesis of 3aa



To an oven-dried 50ml schlenk tube equipped with a stir bar, was added glycosyl NHP ester (1.36 g, 3.75 mmol), *N*-methylquinoxalin-2(1*H*)-one (500 mg, 3.12 mmol). The tube was evacuated and back-filled with Ar (three times), then sealed with rubber stopper and parafilm. Then, anhydrous DMSO (20 mL) was added using a syringe. The solution was then stirred at room temperature under the irradiation of 12 W Blue LEDs for 24 h. After completion of the reaction, 50 mL water was added and extracted by ethyl acetate (3×10 mL). The combined organic layer was washed with brine (20 mL) and then dried over anhydrous Na₂SO₄ and evaporated in vacuum. The residue was directly purified by a silica gel column chromatography using ethyl acetate/petroleum as the eluent to afford product **3aa** (0.83 g, 86%).

5. Control experiments



To an oven-dried 10 mL glass tube equipped with a stir bar, was added glycosyl NHP ester (0.12 mmol), *N*-methylquinoxalin-2(1*H*)-one (0.10 mmol), TEMPO (0.45 mmol). The tube was evacuated and back-filled with Ar (three times), then sealed with rubber stopper and parafilm. Then, anhydrous DMSO (1.0 mL) was added using a syringe. The solution was then stirred at room temperature under the irradiation of 12 W 450nm Blue LEDs for 24 h using electronic fan to cool the tube. After 24 h, no corresponding product **3aa** was formed by TLC analysis, suggesting that the in-situ formed glycosyl radical might act as a key intermediate during this transformation.



To an oven-dried 10 mL glass tube equipped with a stir bar, was added glycosyl NHP ester (0.15 mmol), N-methylquinoxalin-2(1H)-one (0.10 mmol), 1,1-diphenylethylene (0.30 mmol).

The tube was evacuated and back-filled with Ar (three times), then sealed with rubber stopper and parafilm. Then, anhydrous DMSO (1.0 mL) was added using a syringe. The solution was then stirred at room temperature under the irradiation of 12 W Blue LEDs for 24 h using electronic fan to cool the tube. After completion of the reaction, corresponding product **3aa** was isolated in 51% yield and the heck-type product **5** was isolated in 26% yield, indicating that this photo-induced thioglycosidation protocol may proceed through a radical-based mechanism. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.40 – 7.21 (m, 10H), 6.08 (d, *J* = 10.5 Hz, 1H), 5.00 (s, 1H), 4.75 – 4.70 (m, 3H), 3.38 (s, 3H), 1.41 (s, 3H), 1.30 (s, 3H).

6. Stern-Volmer fluorescence quenching experiments

DMSO was degassed with a stream of argon for 30 min. Substrate **1a** or product **3aa** (5.0 μ mol) was dissolved in 1.0 mL DMSO to prepare a 5×10⁻³ M solution. 100 μ L of this solution was added to each of a set of 6 volumetric flasks (10 mL). Subsequently, the solution of quencher **2a** in DMSO (1.5 mL, 0.125 M) was added in increasing amounts (0, 100 μ L, 200 μ L, 300 μ L, 400 μ L, 500 μ L) to the volumetric flasks and the volume of volumetric flasks were adjusted to 10 mL by adding DMSO. All solutions were excited at 350 nm and the emission intensity at 418 nm was observed. All fluorescence measurements were recorded by a F-98 FL Spectrophotometer. Control experiments showed that the excited state substrate **1a** and product **3aa** were both mainly quenched by **2a**.



Figure S1 Emission spectra of 1a with varying concentrations 2a



Figure S2 Emission spectra of 3aa with varying concentrations 2a



Figure S3 Stern-Volmer plot of fluorescence quenching of 1a by 2a



Figure S4 Stern-Volmer plot of fluorescence quenching of 3aa Y by 2a

7. Investigation of potential EDA complex formation

The UV/VIS spectra of all the reaction components were measured in DMSO in a quartz cuvette using a GENESYS 180 UV-visible spectrophotometer in a 10.0 mm quartz cuvette with the aim of investigating the possible formation of an excited donor-acceptor (EDA) complex (exciplex). Spectra were measured for 1a, 2a, 1a+2a in different combinations at various concentrations mimicking the reaction concentration in DMSO. The following measurements were conducted: 1a (0.1 M); 2a (0.12 M); 1a(0.1 M) + 2a (0.12 M). In each case, no EDA complex (Electron-Donor Acceptor) was detected. These spectra are shown in (Figure S5) below along with the spectra for each component in isolation at these concentrations for comparison.



Figure S5 Absorption spectra of 1a, 2a, 1a+2a

8. EDA complex exclusion

The NMR signal of the mixture of **1a** and **2a** shows no shifting change. Combining with the results of investigation of potential EDA complex formation, we conclude that the pathway to generate C-centered radicals through formation of EDA complex is unlikely in this reaction.



Figure S6¹H NMR spectra of 1a, 2a and 1a+2a

9. Cyclic voltammetry measurements

Cyclic Voltammetry was performed using a CHI760E Electrochemical workstation with a glassycarbon as the working electrode, the Ag/AgCl electrode (3 M KCl) as the reference electrode, and a platinum electrode as the counter electrode. The testing solution of **1a**, **2a** were prepared by dissolving the sample (0.05 mmol) into DMSO (5 mL) with 0.1 M tetrabutylammonium hexafluorophosphate (TBAPF₆).



(a)



(b) Figure S7 Cyclic voltammograms for 1a and 2a (a)oxidation (b)reduction

10. Quantum yield measurement

Determination of the light intensity at 450 nm:

According to the literature, ^{9.10} the photon flux of the blue LED (λ max = 450 nm) was determined by standard ferrioxalate actinometry. Potassium ferrioxalate hydrate (2.21 g) was dissolved in H₂SO₄ (0.05 M, 30 mL) to prepare a solution of ferrioxalate (0.15 M). Phenanthroline (50 mg) and sodium acetate (11.25 g) were dissolved in 0.5 M H₂SO₄ (50 mL) to give a buffered solution of phenanthroline. The freshly prepared solutions were stored in dark. Then, the ferrioxalate solution (2.0 mL) was placed in a 3 mL cuvette and irradiated for 90.0 seconds at λ max = 450 nm to determine the photon flux of the blue LED. After irradiation, the phenanthroline solution (0.35 mL) was added to the cuvette, and the resulting solution was then allowed to stand for 1 h to ensure the complete coordination of the ferrous ions to the phenanthroline. The absorbance of the solution was measured at 510 nm. Similarly, a non-irradiated sample was prepared, whose absorbance at 510 nm was also measured. The results were shown as below:



Figure S8 UV-vis spectrum of irradiation and non-irradiation sample Conversion was calculated using equation 1.

$$mol \, Fe^{2+} = \frac{v \cdot \Delta A(510nm)}{l \cdot \varepsilon} = \frac{0.00235 \times 1.58}{1 \times 11100} = 3.3 \times 10^{-7} \tag{1}$$

Where V is the total volume (0.00235 L) of the solution after addition of phenanthroline, ΔA is the difference in absorbance at 510 nm between the irradiated and non-irradiated solutions, l is the path length (1.0 cm), and ε is the molar absorptivity at 510 nm (1.11x10⁴ L mol⁻¹ cm⁻¹). The photon flux can be calculated using equation 2.

photon flux =
$$\frac{mol Fe^{2+}}{\Phi \cdot t \cdot f} = \frac{3.3 \times 10^{-7}}{0.845 \times 90 \times 0.988} = 4.39 \times 10^{-9}$$
 (2)

Where Φ is the quantum yield for the ferrioxalate actinometer (0.845 for a 0.15 M solution at λ =

450 nm), ¹¹t is the time (90 s), and f is the fraction of light absorbed at $\lambda = 450$ nm. This value is calculated using equation 3 where A is the absorbance of the ferrioxalte solution at 450 nm. $f = 1 - 10^{-A} = 1 - 10^{-1.915} = 0.988$ (3)



Figure S9 UV-vis spectrum of ferrioxalate actinometer solution

Determination of the reaction quantum yield (Φ):

Three parallel standard reactions were proceeded on a 0.10 mmol scale according to the general procedure. The standard reaction was stirred and irradiated (24 W blue LEDs, $\lambda = 450$ nm) at room temperature for 10 h. The yield of three parallel standard reactions was determined by ¹H NMR. All of the following NMR yields were afford 64%, 65%, and 66% respectively, therefore, the average yield was obtained 65%. The quantum yield for the reaction was calculated using equation 4. The reaction quantum yield (Φ) was thus determined to be 0.41.

$$\Phi = \frac{\text{mol of product formed}}{\text{photon flux-t-f}} = \frac{6.5 \times 10^{-5}}{4.39 \times 10^{-9} \times 10 \times 60 \times 60 \times 0.999} = 0.41$$
(4)

where photon flux was determined as above described, t is the reaction time, f is the fraction of incident light absorbed by the reaction mixture. This value is calculated using equation 3 where A is the absorbance of the reaction mixture at 450 nm. The absorbance of the reaction mixture at 450 nm was measured to be 2.893, so the value of f is 0.999.

11. Configuration determination

11.1 X-ray diffraction parameters and data of 3aj Method for crystallization:

The purified compound **3aj** was dissolved in a mixed solvent of petroleum ether/n-Hexane (1:3), and placed in a dark cabinet for slowly evaporation. Colorless crystals were collected after few days for X-ray analysis.



Bond precision:	C-C = 0.0051 A	Wavelen	gth=1.54178
Cell:	a=9.9830(8)	b=9.6498(8)	c=10.6876(9)
Temperature:	alpha=90 150 K	beta=98.928(4)	gamma=90
	Calculated	Report	ed
Volume	1017.11(15)	1017.1	0(15)
Space group	P 21	P 1 21	1
Hall group	P 2yb	P 2yb	
Moiety formula	C20 H24 N2 O7	C20 H2	4 N2 O7
Sum formula	C20 H24 N2 O7	C20 H2	4 N2 O7
Mr	404.41	404.41	
Dx,g cm-3	1.321	1.320	
Z	2	2	
Mu (mm-1)	0.843	0.843	
F000	428.0	428.0	
F000'	429.47		
h,k,lmax	12,11,13	12,11,	13
Nref	4015[2135]	3936	
Tmin, Tmax	0.737,0.817	0.616,	0.754
Tmin'	0.668		
Correction metho AbsCorr = MULTI-	d= # Reported T L SCAN	imits: Tmin=0.616	Tmax=0.754
Data completenes	s= 1.84/0.98	Theta $(max) = 72$.204
R(reflections)=	0.0578(3785)		wR2(reflections)= 0.1510(3936)
S = 1.054	Npar= 2	267	Geological database 🔭 🗤 🦛 ja Giologic 🥸

We can conclude that H³ and H⁴ are on the different side of the sugar ring based on the X-ray crystallographic analysis of the molecular structure of product **3aj**. So it is easy to conclude that **3aj** was β isomer. The stereochemical outcome of other furanoses(**3ab-3pa**, **4aa-4ab**) was same to the compound **3aj**.

11.2 NOE experiments



3-((3aS,55,6R,6aS)-6-(benzyloxy)-2,2-dimethyltetrahydrofuro[2,3-*d***][1,3]dioxol-5-yl)-1-m ethylquinoxalin-2(1***H***)-one(4ac'): ¹H NMR (400 MHz, Chloroform-***d***, minor diastereomer) δ 8.08 (d,** *J* **= 8.0 Hz, 1H), 7.56 (t,** *J* **= 7.6 Hz, 1H), 7.36 (t,** *J* **= 7.7 Hz, 1H), 7.29 (d,** *J* **= 8.4 Hz, 1H), 7.14 – 7.11 (m, 1H), 7.07 (t,** *J* **= 7.3 Hz, 2H), 6.95 (d,** *J* **= 7.3 Hz, 2H), 6.27 (d,** *J* **= 3.8 Hz, H¹), 5.65 (d,** *J* **= 3.5 Hz, H⁴), 4.77 (d,** *J* **= 3.5 Hz, H³), 4.70 (d,** *J* **= 3.8 Hz, H²), 4.56 (d,** *J* **= 12.2 Hz, 1H), 4.32 (d,** *J* **= 12.2 Hz, 1H), 3.58 (s, 3H), 1.55 (s, 3H), 1.37 (s, 3H).**

We identified H¹ and H² through NOE experiment because they are on the same side. By comparing the data of another NOE experiment, it is clearly found H³ and H⁴ of the isomer **4ac'** are coupled to each other that they are on the same side. So we concluded that **4ac'** was α isomer and **4ac** was β isomer.



1D-NOE spectrum of compound 4ac'



1D-NOE spectrum of compound 4ac'



3-((3a*R*,5*R*,6*S*,6a*R*)-6-(benzyloxy)-2,2-dimethyltetrahydrofuro[2,3-*d*][1,3]dioxol-5-yl)-1methylquinoxalin-2(1*H*)-one(4ad'): ¹H NMR (600 MHz, Chloroform-*d*, minor diastereomer) δ 8.08 (dd, *J* = 8.0, 1.5 Hz, 1H), 7.57 (td, *J* = 7.9, 1.5 Hz, 1H), 7.37 (td, *J* = 7.7, 1.2 Hz, 1H), 7.30 (dd, *J* = 8.3, 1.2 Hz, 1H), 7.15 – 7.12 (m, 1H), 7.08 – 7.06 (m, 2H), 6.96 – 6.94 (m, 2H), 6.27 (d, *J* = 3.8 Hz, H¹), 5.66 (d, *J* = 3.5 Hz, H⁴), 4.77 (d, *J* = 3.6 Hz, H³), 4.70 (d, *J* = 3.8 Hz, H²), 4.56 (d, *J* = 12.3 Hz, 1H), 4.33 (d, *J* = 12.3 Hz, 1H), 3.58 (s, 3H), 1.55 (s, 3H), 1.37 (s, 3H).

We identified H¹ and H² through NOE experiment because they are on

the same side. Then it was clearly found H³ and H⁴ are coupled to each

other that they are on the same side through another NOE experiment. So we concluded that **4ad'** was α isomer and **4ad** was β isomer .



1D-NOE spectrum of compound 4ad'





1-methyl-3-((3a*R*,5*R*,5a*S*,8a*S*,8b*R*)-2,2,7,7-tetramethyltetrahydro-5*H*-bis([1,3]dioxolo)[4, 5-b:4',5'-d]pyran-5-yl)quinoxalin-2(1*H*)-one(4ae'): ¹H NMR (400 MHz, Chloroform-*d*) δ 8.08 (dd, *J* = 8.0, 1.5 Hz, 1H), 7.56 – 7.51 (m, 1H), 7.34 (td, *J* = 7.7, 1.2 Hz, 1H), 7.30 (dd, *J* = 8.4, 1.2 Hz, 1H), 5.90 (d, *J* = 5.1 Hz, H¹), 5.46 (d, *J* = 2.3 Hz, H⁵), 5.02 (dd, *J* = 7.7, 2.3 Hz, H⁴), 4.75 (dd, *J* = 7.7, 2.5 Hz, H³), 4.47 (dd, *J* = 5.2, 2.6 Hz, H²), 3.68 (s, 3H), 1.59 (s, 3H), 1.44 (s, 3H), 1.37 (s, 3H), 1.26 (s, 3H).

We identified H^1 and H^2 through NOE experiment because they are on the same side. Then it was clearly found H^4 and H^5 are coupled to each other that they are on the same side through another NOE experiment. So we concluded that **4ae'** was α isomer and **4ae** was β isomer .



1D-NOE spectrum of compound 4ae'







1-methyl-3-((2*R*,3*R*,4*S*,5*R*,6*S*)-3,4,5,6-tetramethoxytetrahydro-2*H*-pyran-2-yl)quinoxalin-2(1 *H*)-one(4ag): ¹H NMR (400 MHz, Chloroform-*d*) δ 7.94 (dd, J = 8.0, 1.6 Hz, 1H), 7.60 (td, J = 7.9, 1.5 Hz, 1H), 7.37 (td, J = 8.3, 1.2 Hz, 1H), 7.33 (dd, J = 8.5, 1.2 Hz, 1H), 5.29 (d, J = 10.1 Hz, H¹), 4.91 (d, J = 3.5 Hz, H⁵), 4.01 (dd, J = 10.1, 9.0 Hz, 1H), 3.76 (t, J = 9.3 Hz, H²), 3.72 (s, 3H), 3.68 (s, 3H), 3.62 (s, 3H), 3.56 (s, 3H), 3.46 (s, 3H), 3.39 (dd, J = 9.7, 3.6 Hz, H³).

We identified H¹ and H² through NOE experiment because they are on the same side. Then it was clearly found H³ and H⁵ are coupled to each other that they are on the same side through another NOE experiment. So we concluded that **4ag** was β isomer. The stereochemical outcome of benzyl-protected glucose products (**4ah-4aj**) was same to the compound **4ag**.



1D-NOE spectrum of compound **4ag**



1D-NOE spectrum of compound 4ag

12. References

[1] Ghosh, P.; Kwon, N. Y.; Kim, S.; Han, S.; Lee, S. H.; An, W.; Mishra, N. K.; Han, S. B.; Kim, I. S. C-H Methylation of Iminoamido Heterocycles with Sulfur Ylides**. *Angew. Chem. Int. Ed.* **2021**, 60 (1), 191-196.

[2] Ji, P.; Zhang, Y.; Wei, Y.; Huang, H.; Hu, W.; Mariano, P. A.; Wang, W. Visible-Light-Mediated, Chemo- and Stereoselective Radical Process for the Synthesis of C-Glycoamino Acids. *Org. Lett.* **201**9, 21 (9), 3086-3092.

[3] Masuda, K.; Nagatomo, M.; Inoue, M. Direct assembly of multiply oxygenated carbon chains by decarbonylative radical-radical coupling reactions. *Nat. Chem.* **2017**, 9 (3), 207-212.

[4] Agnihotri, G.; Misra, A. K. Synthesis of a di- and a trisaccharide related to the O-antigen of Escherichia coli O83:K24:H31. *Carbohydr. Res.* **2006**, 341 (14), 2420-5.

[5] Qi, R.; Wang, C.; Ma, Z.; Wang, H.; Chen, Q.; Liu, L.; Pan, D.; Ren, X.; Wang, R.; Xu, Z. Visible-Light-Promoted Stereoselective C(sp3) H Glycosylation for the Synthesis of C-Glycoamino Acids and C-Glycopeptides. *Angew. Chem. Int. Ed.* **2022**, 61 (24), e202200822.

[6] Takashima, K.; Sakano, M.; Kinouchi, E.; Nakamura, S.; Marumoto, S.; Ishikawa, F.; Ninomiya, K.; Nakanishi, I.; Morikawa, T.; Tanabe, G. Elongation of the side chain by linear alkyl groups increases the potency of salacinol, a potent α -glucosidase inhibitor from the Ayurvedic traditional medicine "Salacia," against human intestinal maltase. *Bioorg. Med. Chem. Lett.* **2021**, 33, 127751.

[7] Yamada, K.; Hayakawa, H.; Sakata, S.; Ashida, N.; Yoshimura, Y. Synthesis and antiviral evaluation of α -d-2',3'-didehydro-2',3'-dideoxy-3'-C-hydroxymethyl nucleosides. *Bioorg. Med Chem. Lett.* **2010**, 20 (20), 6013-6016.

[8] Annika, G.; Martin, Hiersemann. (-)-Lytophilippine A: Synthesis of a C1-C18 Building Block. *Org. Lett.* **2010**, 12 (22), 5258-5261.

[9] Lin, S.; Ischay, M. A.; Fry, C. G.; Yoon, T. P. Radical Cation Diels–Alder Cycloadditions by Visible Light Photocatalysis. J. Am. Chem. Soc. **2011**, 133(48), 19350.

[10] Cismesia, M. A.; Yoon, T. P. Characterizing chain processes in visible light photoredox catalysis. *Chem. Sci.* **2015**, 6(10), 5426.

[11] Demas, J. N.; W. D. Bowman. ; Zalewski, E. F.; Velapoldi, R. A. Determination of the quantum yield of the ferrioxalate actinometer with electrically calibrated radiometers. *J. Phys. Chem.* **1981**, 85(19), 2766–2771.

13. Characterization of the Products

3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-1-methylq uinoxalin-2(1*H*)-one(3aa)



Colorless wax; 30.2 mg, 91% yield. ¹H NMR (400 MHz, Chloroform-d) δ 7.87 (dd, J = 8.0, 1.5 Hz, 1H), 7.56 (td, J = 8.6, 1.5 Hz, 1H), 7.37 – 7.31 (m, 2H), 5.58 (d, J = 1.7 Hz, 1H), 5.41 (dd, J = 6.0, 1.7 Hz, 1H), 5.14 (s, 1H), 4.66 (d, J = 5.9 Hz, 1H), 3.72 (s, 3H), 3.37 (s, 3H), 1.59 (s, 3H), 1.39 (s, 3H). ¹³C NMR (100 MHz, Chloroform-d) δ 156.3, 153.9, 133.4, 132.3, 130.64, 130.59, 123.7, 113.6, 112.7, 109.9, 86.0, 85.2, 82.2, 55.3, 28.8, 26.8, 25.4. HRMS (ESI) m/z: [M+H]⁺ Calcd for

C₁₇H₂₁N₂O₅ 333.1450; Found 333.1453.

1-ethyl-3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)qui noxalin-2(1*H*)-one(3ab)



White solid, mp: 100-102 °C; 31.2 mg, 90% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.87 (dd, J = 8.0, 1.5 Hz, 1H), 7.55 (td, J = 8.6, 1.6 Hz, 1H), 7.34 – 7.30 (m, 2H), 5.58 (d, J = 1.6 Hz, 1H), 5.41 (dd, J = 6.0, 1.7 Hz, 1H), 5.14 (s, 1H), 4.65 (d, J = 5.9 Hz, 1H), 4.33 (q, J = 7.2 Hz, 2H), 3.38 (s, 3H), 1.58 (s, 3H), 1.39 (s, 3H), 1.36 (t, J = 7.2 Hz, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 155.3, 152.3, 131.6, 131.3, 129.9, 129.5, 122.5, 112.4, 111.7, 108.7, 85.0, 84.1,

81.3, 54.2, 36.0, 25.8, 24.4, 11.4. HRMS (ESI) m/z: $[M+H]^+$ Calcd for $C_{18}H_{23}N_2O_5$ 347.1607; Found 347.1605.

1-isobutyl-3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl) quinoxalin-2(1*H*)-one(3ac)



White solid, mp: 128-130 °C; 31.1 mg, 83% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.87 (dd, J = 8.3, 1.5 Hz, 1H), 7.55 – 7.51 (m, 1H), 7.34 – 7.30 (m, 2H), 5.58 (d, J = 1.6 Hz, 1H), 5.39 (dd, J = 5.9, 1.7 Hz, 1H), 5.15 (s, 1H), 4.66 (d, J = 5.9 Hz, 1H), 4.24 (dd, J = 13.7, 7.9 Hz, 1H), 4.05 (dd, J = 13.7, 7.1 Hz, 1H), 3.40 (s, 3H), 2.26 (dq, J = 13.8, 6.9 Hz, 1H), 1.59 (s, 3H), 1.40 (s, 3H), 1.00 (d, J = 6.8 Hz, 3H), 0.98 (d, J = 6.7 Hz, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.4, 154.1, 133.0, 132.5, 130.9, 130.3, 123.4, 114.1, 112.7, 109.9,

86.2, 85.2, 82.3, 55.3, 48.8, 27.2, 26.8, 25.4, 20.3, 20.1. HRMS (ESI) m/z: $[M+H]^+$ Calcd for $C_{20}H_{27}N_2O_5$ 375.1920; Found 375.1916.

1-butyl-3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)qui noxalin-2(1*H*)-one(3ad)



White solid, mp: 107-109 °C; 34.4 mg, 92% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.86 (dd, J = 8.2, 1.6 Hz, 1H), 7.54 (td, J = 7.7, 1.6 Hz, 1H), 7.33 – 7.29 (m, 2H), 5.57 (d, J = 1.7 Hz, 1H), 5.39 (dd, J = 5.9, 1.7 Hz, 1H), 5.14 (s, 1H), 4.64 (d, J = 5.9 Hz, 1H), 4.33 – 4.18 (m, 2H), 3.39 (s, 3H), 1.76 – 1.69 (m, 2H), 1.58 (s, 3H), 1.51 – 1.42 (m, 2H), 1.39 (s, 3H), 0.98 (t, J = 7.4 Hz,3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.4, 153.6, 132.61, 132.58, 130.8, 130.4, 123.5,

113.6, 112.7, 109.8, 86.2, 85.2, 82.3, 55.2, 41.9, 29.3, 26.8, 25.4, 20.3, 13.8. HRMS (ESI) m/z: $[M+H]^+$ Calcd for $C_{20}H_{27}N_2O_5$ 375.1920; Found 375.1925.

1-(cyclopropylmethyl)-3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3] dioxol-4-yl)quinoxalin-2(1*H*)-one(3ae)



White solid, mp: 108-110 °C; 31.3 mg, 84% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.88 (dd, J = 8.0, 1.6 Hz, 1H), 7.56 (td, J = 7.8, 1.6 Hz, 1H), 7.44 (d, J = 8.4 Hz, 1H), 7.35 – 7.31 (m, 1H), 5.57 (d, J = 1.7 Hz, 1H), 5.41 (dd, J = 5.9, 1.7 Hz, 1H), 5.15 (s, 1H), 4.65 (d, J = 5.9 Hz, 1H), 4.27 – 4.16 (m, 2H), 3.40 (s, 3H), 1.59 (s, 3H), 1.40 (s, 3H), 1.33 – 1.26 (m, 1H), 0.61 – 0.47 (m, 4H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 155.5, 152.9, 131.8, 131.5, 129.8, 129.4, 122.4, 112.9, 111.7, 108.8, 85.2, 84.1, 81.3, 54.2, 44.9, 25.8, 24.4, 8.6, 3.1, 3.0. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₂₀H₂₅N₂O₅

373.1763; Found 373.1758.

1-(cyclohexylmethyl)-3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3] dioxol-4-yl)quinoxalin-2(1*H*)-one(3af)



White solid, mp: 83-85 °C; 35.6 mg, 86% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.86 (dd, J = 8.2, 1.6 Hz, 1H), 7.53 (td, J = 8.6, 1.6 Hz, 1H), 7.34 – 7.29 (m, 2H), 5.57 (d, J = 1.7 Hz, 1H), 5.38 (dd, J = 6.0, 1.7 Hz, 1H), 5.15 (s, 1H), 4.65 (d, J = 5.9 Hz, 1H), 4.23 (dd, J = 13.7, 7.6 Hz, 1H), 4.06 (dd, J = 13.8, 7.1 Hz, 1H), 3.40 (s, 3H), 1.96 – 1.86 (m, 1H), 1.73 – 1.63 (m, 5H), 1.59 (s, 3H), 1.40 (s, 3H), 1.21 – 1.11 (m, 5H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.4, 154.1, 133.1, 132.5, 130.8, 130.3, 123.4, 114.1, 112.7, 109.9, 86.3, 85.2, 82.4, 55.3, 47.8, 36.5, 30.9, 30.8, 26.8, 26.2, 25.79, 25.75, 25.5.

HRMS (ESI) m/z: [M+H]⁺ Calcd for C₂₃H₃₁N₂O₅ 415.2233; Found 415.2239.

1-benzyl-3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)q uinoxalin-2(1*H*)-one(3ag)



Colorless oil; 38.4 mg, 94% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.87 (dd, J = 7.9, 1.6 Hz, 1H), 7.45 – 7.41 (m, 1H), 7.31 – 7.22 (m, 7H), 5.65 (d, J = 1.7 Hz, 1H), 5.58 (d, J = 15.6 Hz, 1H), 5.49 (dd, J = 5.9, 1.6 Hz, 1H), 5.44 (d, J = 15.6 Hz, 1H), 5.49 (dd, J = 5.9 Hz, 1H), 3.38 (s, 3H), 1.60 (s, 3H), 1.41 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.5, 154.0, 135.2, 132.8, 132.5, 130.7, 130.6, 128.9, 127.8, 127.0, 123.8, 114.4, 112.7, 109.9, 86.0, 85.3, 82.2, 55.3, 45.6, 26.8, 25.4. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₂₃H₂₅N₂O₅ 409.1763; Found 409.1766.

1-allyl-3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)qui noxalin-2(1*H*)-one(3ah)



White solid, mp: 139-141 °C; 33.3 mg, 93% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.87 (dd, J = 8.0, 1.5 Hz, 1H), 7.54 – 7.50 (m, 1H), 7.35 – 7.28 (m, 2H), 5.97 – 5.87 (m, 1H), 5.59 (d, J = 1.6 Hz, 1H), 5.44 (dd, J = 5.9, 1.7 Hz, 1H), 5.27 – 5.24 (m, 1H), 5.18 – 5.13 (m, 2H), 5.00 – 4.84 (m, 2H), 4.67 (d, J = 5.9 Hz, 1H), 3.37 (s, 3H), 1.58 (s, 3H), 1.39 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.4, 153.4, 132.6, 132.4,130.7, 130.6, 130.5, 123.7, 118.3, 114.2, 112.7, 109.8, 85.9, 85.2, 82.2, 55.2, 44.2, 26.8, 25.4. HRMS (ESI)

m/z: $[M+H]^+$ Calcd for $C_{19}H_{23}N_2O_5$ 359.1607; Found 359.1601.

3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-1-(prop-2-yn-1-yl)quinoxalin-2(1*H*)-one(3ai)



White solid, mp: 161-163 °C; 31 mg, 87% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.88 (dd, J = 8.0, 1.5 Hz, 1H), 7.59 (td, J = 7.8, 1.5 Hz, 1H), 7.47 (dd, J = 8.5, 1.2 Hz, 1H), 7.37 (td, J = 7.6, 1.2 Hz, 1H), 5.57 (d, J = 1.7 Hz, 1H), 5.44 (dd, J = 5.9, 1.7 Hz, 1H), 5.13 (s, 1H), 5.12 – 5.01 (m, 2H), 4.66 (d, J = 5.9 Hz, 1H), 3.36 (s, 3H), 2.27 (t, J = 2.5 Hz, 1H), 1.58 (s, 3H), 1.39 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 155.3, 151.8, 131.4, 130.8, 129.71, 129.66, 123.1, 113.2, 111.7, 108.8, 84.8, 84.1, 81.1, 75.7, 72.3, 54.2, 30.1, 25.8,

24.3. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₁₉H₂₁N₂O₅ 357.1450; Found 357.1458.

Ethyl2-(3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-2-oxoquinoxalin-1(2*H*)-yl)acetate(3aj)



White solid, mp: 159-161 °C; 32.4 mg, 80% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.88 (dd, J = 8.0, 1.5 Hz, 1H), 7.52 (td, J = 7.9, 1.5 Hz, 1H), 7.36 – 7.32 (m, 1H), 7.07 (d, J = 8.3 Hz, 1H), 5.57 (d, J = 1.6 Hz, 1H), 5.47 (dd, J = 5.9, 1.6 Hz, 1H), 5.13 (s, 1H), 5.08 – 4.99 (m, 2H), 4.67 (d, J = 5.9 Hz, 1H), 4.22 (q, J = 7.1 Hz, 2H), 3.34 (s, 3H), 1.58 (s, 3H), 1.39 (s, 3H), 1.25 (t, J = 7.1 Hz, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 165.9, 155.2, 152.4, 131.5, 131.3, 129.9, 129.7, 123.0, 112.1, 111.6, 108.8, 84.7, 84.2, 81.0, 61.1, 54.1, 42.2, 25.7, 24.3, 13.1. HRMS (ESI) m/z: [M+H]⁺ Calcd for

 $C_{20}H_{25}N_2O_7$ 405.1662; Found 405.1657.

3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-1-(2-oxo-2 -phenylethyl)quinoxalin-2(1*H*)-one(3ak)



White solid, mp: 191-193 °C; 35.4 mg, 87% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.07 – 8.04 (m, 2H), 7.90 (dd, J = 8.0, 1.5 Hz, 1H), 7.68 – 7.64 (m, 1H), 7.55 – 7.51 (m, 2H), 7.45 (td, J = 7.9, 1.6 Hz, 1H), 7.32 (td, J = 7.7, 1.2 Hz, 1H), 6.96 (dd, J = 8.5, 1.2 Hz, 1H), 5.80 – 5.70 (m, 2H), 5.58 (d, J = 1.5 Hz, 1H), 5.49 (dd, J = 5.9, 1.6 Hz, 1H), 5.14 (s, 1H), 4.69 (d, J = 5.9 Hz, 1H), 3.37 (s, 3H), 1.57 (s, 3H), 1.39 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 190.9, 156.1, 153.7, 134.5, 134.4, 132.9, 132.4, 130.9, 130.7, 129.1, 128.1, 123.9, 113.5, 112.6, 109.8, 85.7, 85.3, 82.1, 55.2, 48.2, 26.8, 25.3. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₂₄H₂₅N₂O₆ 437.1713; Found

437.1721.

2-(3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-2-oxoq uinoxalin-1(2*H*)-yl)acetonitrile(3al)



White solid, mp: 81-83 °C; 24 mg, 67% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.92 (dd, J = 8.0, 1.5 Hz, 1H), 7.65 (td, J = 8.3, 1.5 Hz, 1H), 7.44 (td, J = 7.7, 1.2 Hz, 1H), 7.34 (dd, J = 8.4, 1.2 Hz, 1H), 5.54 (d, J = 1.5 Hz, 1H), 5.45 (dd, J = 5.9, 1.6 Hz, 1H), 5.27 – 5.18 (m, 2H), 5.12 (s, 1H), 4.66 (d, J = 5.9 Hz, 1H), 3.33 (s, 3H), 1.58 (s, 3H), 1.39 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.0, 152.5, 132.3, 131.4, 131.3, 131.1, 125.0, 113.6, 113.1, 112.8, 110.0, 85.5, 85.2, 81.9, 55.3, 29.1, 26.8, 25.3. HRMS (ESI) m/z: [M+H]⁺

Calcd for C₁₈H₂₀N₃O₅ 358.1403; Found 358.1399.

3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-1,7-dimet hylquinoxalin-2(1*H*)-one(3ba)



Colorless oil; 27 mg, 78% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.74 (d, J = 8.1 Hz, 1H), 7.15 (dd, J = 8.2, 1.7 Hz, 1H), 7.10 (s, 1H), 5.56 (d, J = 1.6 Hz, 1H), 5.41 (dd, J = 6.0, 1.6 Hz, 1H), 5.13 (s, 1H), 4.66 (d, J = 5.9 Hz, 1H), 3.69 (s, 3H), 3.36 (s, 3H), 2.51 (s, 3H), 1.58 (s, 3H), 1.39 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 155.0, 154.1, 141.5, 133.3, 130.5, 130.3, 125.1, 113.8, 112.6, 109.8, 85.9, 85.2, 82.2, 55.2, 28.8, 26.8, 25.3,

22.2. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₁₈H₂₃N₂O₅ 347.1607; Found 347.1613.

3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-1,6-dimet hylquinoxalin-2(1*H*)-one(3ca)



Colorless oil; 27.7 mg, 80% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.66 (d, J = 1.9 Hz, 1H), 7.38 (dd, J = 8.5, 2.1 Hz, 1H), 7.21 (d, J = 8.5 Hz, 1H), 5.58 (d, J = 1.6 Hz, 1H), 5.41 (dd, J = 5.9, 1.6 Hz, 1H), 5.13 (s, 1H), 4.66 (d, J = 5.9 Hz, 1H), 3.69 (s, 3H), 3.36 (s, 3H), 2.44 (s, 3H), 1.59 (s, 3H), 1.39 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.2, 153.9, 133.6, 132.2, 131.8, 131.2, 130.4, 113.4, 112.6, 109.9, 85.9, 85.2, 82.2, 55.3,

28.9, 26.8, 25.3, 20.7. HRMS (ESI) m/z: $[M+H]^+$ Calcd for $C_{18}H_{23}N_2O_5$ 347.1607; Found 347.1603.

3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-1,6,7-trim ethylquinoxalin-2(1*H*)-one(3da)



Colorless oil; 32.1 mg, 89% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.60 (s, 1H), 7.07 (s, 1H), 5.56 (d, J = 1.9 Hz, 1H), 5.43 (dd, J = 5.9, 2.1 Hz, 1H), 5.11 (s, 1H), 4.66 (dd, J = 6.1, 1.9 Hz, 1H), 3.67 (s, 3H), 3.32 (s, 3H), 2.40 (s, 3H), 2.33 (s, 3H), 1.57 (s, 3H), 1.38 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 154.9, 154.0, 140.5, 132.6, 131.5, 130.7, 130.6, 114.2, 112.6, 109.9, 85.7, 85.3, 82.1, 55.1, 28.7, 26.8, 25.4, 20.6, 19.1. HRMS

(ESI) m/z: [M+H]⁺ Calcd for C₁₉H₂₅N₂O₅ 361.1763; Found 361.1771.

3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-1-methylb enzo[*g*]quinoxalin-2(1*H*)-one(3ea)



Yellow solid, mp: 79-81 °C; 36 mg, 94% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.36 (s, 1H), 7.95 (d, *J* = 8.2 Hz, 1H), 7.90 (d, *J* = 8.4 Hz, 1H), 7.59 (s, 1H), 7.56 (d, *J* = 8.0 Hz, 1H), 7.48 (t, *J* = 7.5 Hz, 1H), 5.63 (d, *J* = 1.7 Hz, 1H), 5.49 (dd, *J* = 5.9, 1.6 Hz, 1H), 5.16 (s, 1H), 4.71 (d, *J* = 5.9 Hz, 1H), 3.76 (s, 3H), 3.38 (s, 3H), 1.61 (s, 3H), 1.42 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.9, 153.8, 133.8, 131.8, 131.6, 130.0,

129.7, 128.5, 128.1, 127.2, 125.4, 112.7, 110.0, 109.9, 85.8, 85.3, 82.2, 55.3, 28.8, 26.8, 25.4. HRMS (ESI) m/z: $[M+H]^+$ Calcd for $C_{21}H_{23}N_2O_5$ 383.1607; Found 383.1601.

3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-1-methyl-6-phenylquinoxalin-2(1*H*)-one(3fa)



Yellow solid, mp: 99-101 °C; 38.1 mg, 93% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.09 (d, J = 2.2 Hz, 1H), 7.81 (dd, J = 2.2, 8.7 Hz, 1H), 7.65 – 7.62 (m, 2H), 7.49 – 7.45 (m, 2H), 7.40 – 7.36 (m, 2H), 5.61 (d, J = 1.6 Hz, 1H), 5.44 (dd, J = 5.9, 1.6 Hz, 1H), 5.16 (s, 1H), 4.68 (d, J = 5.9 Hz, 1H), 3.74 (s, 3H), 3.39 (s, 3H), 1.60 (s, 3H), 1.40 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.8, 153.8, 139.3,

136.9, 132.6, 132.5, 129.5, 129.1, 128.6, 127.7, 127.0, 114.1, 112.7, 109.9, 86.0, 85.2, 82.2, 55.3, 29.0, 26.8, 25.3. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₂₃H₂₅N₂O₅ 409.1763; Found 409.1767.

6-methoxy-3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl) -1-methylquinoxalin-2(1*H*)-one(3ga)



White solid, mp: 153-155 °C; 31.9 mg, 88% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.32 (d, J = 2.8 Hz, 1H), 7.23 (d, J = 9.1 Hz, 1H), 7.18 (dd, J = 9.1, 2.8 Hz, 1H), 5.58 (d, J = 1.6 Hz, 1H), 5.43 (dd, J = 5.9, 1.7 Hz, 1H), 5.13 (s, 1H), 4.67 (d, J = 5.9 Hz, 1H), 3.87 (s, 3H), 3.70 (s, 3H), 3.34 (s, 3H), 1.58 (s, 3H), 1.39 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.8, 156.0, 153.6, 133.0, 127.7, 119.7, 114.5, 112.7, 112.1, 109.9,

85.9, 85.2, 82.1, 55.8, 55.2, 29.0, 26.79, 25.4. HRMS (ESI) m/z: $[M+H]^+$ Calcd for $C_{18}H_{23}N_2O_6$ 363.1556; Found 363.1553.
7-bromo-3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-1 -methylquinoxalin-2(1*H*)-one(3ha)



White solid, mp: 109-111 °C; 33.7 mg, 82% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.71 (d, *J* = 8.4 Hz, 1H), 7.47 – 7.43 (m, 2H), 5.54 (d, *J* = 1.6 Hz, 1H), 5.39 (dd, *J* = 6.0, 1.7 Hz, 1H), 5.13 (s, 1H), 4.65 (d, *J* = 5.9 Hz, 1H), 3.68 (s, 3H), 3.35 (s, 3H), 1.58 (s, 3H), 1.39 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.8, 153.6, 134.4, 131.8, 131.1, 127.0, 124.8, 116.8, 112.8, 109.9, 85.9, 85.2, 82.1, 55.3, 29.0, 26.8, 25.3.

HRMS (ESI) m/z: $[M+H]^+$ Calcd for $C_{17}H_{20}BrN_2O_5$ 411.0556; Found 411.0562.

6-bromo-3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-1 -methylquinoxalin-2(1*H*)-one(3ia)



White solid, mp: 79-81 °C; 32.5 mg, 79% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.00 (d, J = 2.3 Hz, 1H), 7.64 (dd, J = 8.9, 2.3 Hz, 1H), 7.19 (d, J = 8.9 Hz, 1H), 5.56 (d, J = 1.6 Hz, 1H), 5.37 (dd, J = 6.0, 1.6 Hz, 1H), 5.12 (s, 1H), 4.64 (d, J = 5.9 Hz, 1H), 3.69 (s, 3H), 3.35 (s, 3H), 1.58 (s, 3H), 1.38 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.8, 152.5, 132.3, 132.0, 131.9, 131.5, 115.3, 114.1, 111.7, 108.9, 84.8, 84.1, 81.1,

54.3, 28.0, 25.7, 24.3. HRMS (ESI) m/z: $[M+H]^+$ Calcd for $C_{17}H_{20}BrN_2O_5$ 411.0556; Found 411.0551.

2-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-][1,3]dioxol-4-yl)-4-methyl-3-oxo-3,4-dihydroquinoxaline-6-carbonitrile(3ja)



White solid, mp: 152-154 °C; 23.2 mg, 65% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.95 (d, J = 8.1 Hz, 1H), 7.62 – 7.58 (m, 2H), 5.57 (d, J = 1.6 Hz, 1H), 5.36 (dd, J = 5.9, 1.7 Hz, 1H), 5.14 (s, 1H), 4.63 (d, J = 5.9 Hz, 1H), 3.72 (s, 3H), 3.37 (s, 3H), 1.58 (s, 3H), 1.38 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 159.0, 152.3, 133.3, 132.8, 130.4, 125.6, 117.0, 116.9, 112.7, 111.9, 108.9, 85.0, 84.0, 81.0, 54.4, 28.1, 25.7,

24.3. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₁₈H₂₀N₃O₅ 358.1403; Found 358.1407.

3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-1-methyl-2-oxo-1,2-dihydroquinoxaline-6-carbonitrile(3ka)



White solid, mp: 179-181 °C; 24.3 mg, 68% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.15 (d, J = 1.9 Hz, 1H), 7.78 (dd, J = 8.7, 1.9 Hz, 1H), 7.41 (d, J = 8.7 Hz, 1H), 5.54 (d, J = 1.6 Hz, 1H), 5.34 (dd, J = 5.9, 1.6Hz, 1H), 5.12 (s, 1H), 4.62 (d, J = 5.9 Hz, 1H), 3.72 (s, 3H), 3.35 (s, 3H), 1.57 (s, 3H), 1.38 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 158.9, 153.5, 136.7, 134.8, 133.1, 131.8, 117.8, 114.9, 112.9, 110.0, 107.3, 85.8,

85.0, 82.0, 55.4, 29.2, 26.7, 25.3. HRMS (ESI) m/z: $[M+H]^+$ Calcd for $C_{18}H_{20}N_3O_5$ 358.1403; Found 358.1396.

6,7-difluoro-3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-y l)-1-methylquinoxalin-2(1*H*)-one(3la)



White solid, mp: 77-79 °C; 31.3 mg, 85% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.66 (q, J = 10.1, 8.1 Hz, 1H), 7.12 (q, J = 11.2, 7.0 Hz, 1H), 5.53 (d, J = 1.6 Hz, 1H), 5.35 (dd, J = 5.9, 1.7 Hz, 1H), 5.12 (s, 1H), 4.63 (d, J = 5.9 Hz, 1H), 3.66 (s, 3H), 3.34 (s, 3H), 1.57 (s, 3H), 1.38 (s, 3H). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -129.92 – 129.81(m), -141.82 – 141.71(m). ¹³C NMR (100 MHz, Chloroform-*d*) δ 157.0 (d, J = 3.5 Hz), 153.4,

151.7(dd, J = 254.3 Hz, 14.4 Hz), 146.7 (dd, J = 247.5, 14.0 Hz), 130.7 (dd, J = 9.1, 1.9 Hz), 128.5 (dd, J = 9.2, 2.9 Hz), 118.1 (dd, J = 17.9, 2.3 Hz), 112.8, 109.9, 102.4 (d, J = 23.3 Hz), 85.8, 85.1, 82.1, 55.3, 29.4, 26.7, 25.3. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₁₇H₁₉F₂N₂O₅ 369.1262; Found 369.1269.

3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)quinoxalin -2(1*H*)-one(3ma)



White solid, mp: 230-232 °C; 27.4 mg, 86% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 12.49 (s, 1H), 7.85 (dd, J = 8.1, 1.4 Hz, 1H), 7.52 (td, J = 8.3, 1.4 Hz, 1H), 7.43 (dd, J = 8.3, 1.4 Hz, 1H), 7.35 (td, J = 7.6, 1.4 Hz, 1H), 5.68 (d, J = 6.2 Hz, 1H), 5.66 (d, J = 1.2 Hz, 1H), 5.12 (s, 1H), 4.76 (d, J = 5.8 Hz, 1H), 3.22 (s, 3H), 1.61 (s, 3H), 1.43 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.5, 155.9, 132.2, 131.3, 130.8, 129.7, 124.4, 116.0, 112.5, 110.1, 85.6, 84.5,

81.6, 55.0, 26.7, 25.3. HRMS (ESI) m/z: $[M+H]^+$ Calcd for $C_{16}H_{19}N_2O_5$ 319.1294; Found 319.1286.

3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)benzo[g]qu inoxalin-2(1*H*)-one(3na)



Yellow solid, mp: 219-221 °C; 33.9 mg, 92% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 12.04 (s, 1H), 8.36 (s, 1H), 7.95 (d, J = 8.3 Hz, 1H), 7.91 (d, J = 8.4 Hz, 1H), 7.81 (s, 1H), 7.56 (t, J = 7.5 Hz, 1H), 7.47 (t, J = 7.5 Hz, 1H), 5.75 (d, J = 5.7 Hz, 2H), 5.14 (s, 1H), 4.81 (d, J = 5.7 Hz, 1H), 3.22 (s, 3H), 1.65 (s, 3H), 1.46 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 157.1, 155.6, 134.1, 131.4, 130.3, 129.3, 129.1, 128.8, 128.1, 127.0,

125.3, 112.5, 111.9, 110.2, 85.7, 84.3, 81.5, 55.0, 26.8, 25.3. HRMS (ESI) m/z: $[M+H]^+$ Calcd for $C_{20}H_{21}N_2O_5$ 369.1450; Found 369.1457.

3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl)-6,7-dimet hylquinoxalin-2(1*H*)-one(3oa)



White solid, mp: 200-202 °C; 30.8 mg, 89% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 12.72 (s, 1H), 7.60 (s, 1H), 7.22 (s, 1H), 5.70 (d, *J* = 6.0 Hz, 1H), 5.68 (s, 1H), 5.10 (s, 1H), 4.77 (d, *J* = 5.8 Hz, 1H), 3.17 (s, 3H), 2.38 (s, 3H), 2.34 (s, 3H), 1.61 (s, 3H), 1.43 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.2, 155.0, 141.1, 133.5, 130.8, 129.5, 129.4, 116.3, 112.4, 110.1, 85.8, 84.2, 81.6, 54.8, 26.8, 25.3, 20.1, 19.5. HRMS (ESI) m/z: [M+H]⁺ Calcd for

C₁₈H₂₃N₂O₅ 347.1607; Found 347.1604.

6-methoxy-3-((3a*R*,4*R*,6*R*,6a*R*)-6-methoxy-2,2-dimethyltetrahydrofuro[3,4-*d*][1,3]dioxol-4-yl) quinoxalin-2(1*H*)-one(3pa)



Yellow solid, mp: 194-196 °C; 28.9 mg, 83% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 12.76 (s, 1H), 7.37 (d, *J* = 8.9 Hz, 1H), 7.29 (d, *J* = 2.8 Hz, 1H), 7.17 (dd, *J* = 8.9, 2.8 Hz, 1H), 5.70 (dd, *J* = 6.0, 1.2 Hz, 1H), 5.66 (d, *J* = 1.2 Hz, 1H), 5.11 (s, 1H), 4.77 (d, *J* = 5.9 Hz, 1H), 3.88 (s, 3H), 3.20 (s, 3H), 1.61 (s, 3H), 1.43 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 156.7, 156.6, 155.7, 133.0, 125.6, 120.8, 117.0, 112.5, 110.5, 110.1,

85.7, 84.3, 81.5, 55.8, 54.9, 26.7, 25.3. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₁₇H₂₁N₂O₆ 349.1400; Found 349.1405.

1-methyl-3-((2R,3R,4R,5R)-3,4,5-trimethoxytetrahydrofuran-2-yl)quinoxalin-2(1H)-one(4aa)



Colorless oil; 29.5 mg, 92% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.90 (dd, J = 8.0, 1.5 Hz, 1H), 7.56 (td, J = 7.9, 1.6 Hz, 1H), 7.36 – 7.29 (m, 2H), 5.57 (d, J = 5.1 Hz, 1H), 5.10 (d, J =2.8 Hz, 1H), 4.61 (t, J = 4.9 Hz, 1H), 3.92 (dd, J = 4.8, 2.8 Hz, 1H), 3.70 (s, 3H), 3.52 (s, 3H), 3.51 (s, 3H), 3.39 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 154.9, 153.5, 132.5, 131.4, 129.84, 129.81,

122.6, 112.6, 105.5, 81.6, 79.6, 77.9, 57.33, 57.28, 54.9, 28.0. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₁₆H₂₁N₂O₅ 321.1450; Found 321.1445.

3-((2*R*,3*R*,4*R*,5*R*)-3,4-bis(benzyloxy)-5-methoxytetrahydrofuran-2-yl)-1-methylquinoxalin-2(1*H*)-one(4ab)



Colorless oil; 36.4 mg, 77% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.85 (dd, J = 8.0, 1.6 Hz, 1H), 7.59 – 7.54 (m, 1H), 7.42– 7.39 (m, 2H), 7.37 – 7.19 (m, 10H), 5.69 (d, J = 5.5 Hz, 1H), 5.15 (d, J = 2.5 Hz, 1H), 4.85 (t, J = 5.1 Hz, 1H), 4.73 – 4.66(m, 4H), 4.08 (dd, J = 4.7, 2.4 Hz, 1H), 3.69 (s, 3H), 3.36 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 155.8, 154.5, 138.2, 138.0, 133.6, 132.4, 130.81,

130.78, 128.4, 128.2, 128.0, 127.9, 127.7, 127.6, 123.6, 113.6, 107.2, 80.7, 79.4, 78.6, 72.42, 72.39, 55.8, 29.0. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₂₈H₂₉N₂O₅ 473.2076; Found 473.2071.

3-((3a*S*,5*R*,6*R*,6a*S*)-6-(benzyloxy)-2,2-dimethyltetrahydrofuro[2,3-*d*][1,3]dioxol-5-yl)-1-meth ylquinoxalin-2(1*H*)-one(4ac)



Colorless oil; 19.9 mg, 49% yield. ¹H NMR (400 MHz, Chloroform-*d*, major diastereomer) δ 7.97 (dd, J = 8.0, 1.5 Hz, 1H), 7.57 (td, J = 8.6, 1.5 Hz, 1H), 7.43 (d, J = 7.2 Hz, 2H), 7.39 – 7.28 (m, 5H), 6.17 (d, J = 3.6 Hz, 1H), 5.65 (d, J = 1.9 Hz, 1H), 4.98 (d, J = 11.7 Hz, 1H), 4.85 (d, J = 11.7 Hz, 1H), 4.70 – 4.68 (m, 2H), 3.72 (s, 3H), 1.46 (s, 3H), 1.30 (s, 3H). ¹³C NMR (100 MHz,

Chloroform-*d*, major diastereomer) δ 154.4, 153.0, 136.9, 132.4, 131.3, 129.5, 129.4, 127.3, 126.9, 126.7, 122.9, 112.6, 112.1, 105.9, 83.9, 83.4, 82.6, 71.1, 27.8, 25.2, 25.0. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₂₃H₂₅N₂O₅ 409.1763; Found 409.1771.

3-((3aS,5S,6R,6aS)-6-(benzyloxy)-2,2-dimethyltetrahydrofuro[2,3-*d*][1,3]dioxol-5-yl)-1-methy lquinoxalin-2(1*H*)-one(4ac')



Colorless oil; 13.2 mg, 32% yield. ¹H NMR (400 MHz, Chloroform-*d*, minor diastereomer) δ 8.08 (d, J = 8.0 Hz, 1H), 7.56 (t, J = 7.6 Hz, 1H), 7.36 (t, J = 7.7 Hz, 1H), 7.29 (d, J = 8.4 Hz, 1H), 7.14 - 7.11 (m, 1H), 7.07 (t, J = 7.3 Hz, 2H), 6.95 (d, J = 7.3 Hz, 2H), 6.27 (d, J = 3.8 Hz, 1H), 5.65 (d, J = 3.5 Hz, 1H), 4.77 (d, J = 3.5 Hz, 1H), 4.70 (d, J = 3.8 Hz, 1H), 4.56 (d, J = 12.2 Hz, 1H),

4.32 (d, J = 12.2 Hz, 1H), 3.58 (s, 3H), 1.55 (s, 3H), 1.37 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*, minor diastereomer) δ 153.4, 152.4, 136.5, 131.8, 131.7, 129.8, 129.2, 127.0, 126.5, 126.4, 122.7, 112.4, 111.1, 104.1, 82.7, 82.0, 79.1, 71.3, 27.7, 26.0, 25.5. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₂₃H₂₅N₂O₅ 409.1763; Found 409.1760.

3-((3a*R*,5*S*,6*S*,6a*R*)-6-(benzyloxy)-2,2-dimethyltetrahydrofuro[2,3-*d*][1,3]dioxol-5-yl)-1-meth ylquinoxalin-2(1*H*)-one(4ad)



Colorless oil; 20.3 mg, 50% yield. ¹H NMR (600 MHz, Chloroform-*d*, major diastereomer) δ 7.97 (dd, J = 8.0, 1.5 Hz, 1H), 7.57 (td, J = 7.8, 1.5 Hz, 1H), 7.43 (d, J = 7.5 Hz, 2H), 7.38 – 7.33 (m, 4H), 7.28 (d, J = 7.5 Hz, 1H), 6.17 (d, J = 3.6 Hz, 1H), 5.65 (d, J = 1.9 Hz, 1H), 4.97 (d, J = 11.7 Hz, 1H), 4.85 (d, J = 11.7 Hz, 1H), 4.70 – 4.68 (m, 2H), 3.72 (s, 3H), 1.46 (s, 3H), 1.29 (s, 3H).

¹³C NMR (150 MHz, Chloroform-*d*, major diastereomer) δ 155.4, 154.0, 137.9, 133.4, 132.3, 130.51, 130.46, 128.4, 127.9, 127.7, 123.9, 113.7, 113.1, 106.9, 84.9, 84.4, 83.6, 72.2, 28.9, 26.2, 26.0. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₂₃H₂₅N₂O₅ 409.1763; Found 409.1756.

3-((3aR,5R,6S,6aR)-6-(benzyloxy)-2,2-dimethyltetrahydrofuro[2,3-d][1,3]dioxol-5-yl)-1-meth ylquinoxalin-2(1*H*)-one(4ad')



Colorless oil; 13.6 mg, 33% yield. ¹H NMR (600 MHz, Chloroform-*d*, minor diastereomer) δ 8.08 (dd, J = 8.0, 1.5 Hz, 1H), 7.57 (td, J = 7.9, 1.5 Hz, 1H), 7.37 (td, J = 7.7, 1.2 Hz, 1H), 7.30 (dd, J = 8.3, 1.2 Hz, 1H), 7.15 – 7.12 (m, 1H), 7.08 – 7.06 (m, 2H), 6.96 – 6.94 (m, 2H), 6.27 (d, J = 3.8 Hz, 1H), 5.66 (d, J = 3.5 Hz, 1H), 4.77 (d, J = 3.6 Hz, 1H), 4.70 (d, J = 3.8 Hz, 1H), 4.56 (d, J =

12.3 Hz, 1H), 4.33 (d, J = 12.3 Hz, 1H), 3.58 (s, 3H), 1.55 (s, 3H), 1.37 (s, 3H). ¹³C NMR (150 MHz, Chloroform-*d*, minor diastereomer) δ 154.3, 153.4, 137.5, 132.8, 132.7, 130.8, 130.3, 128.0, 127.5, 127.4, 123.8, 113.4, 112.1, 105.1, 83.7, 83.0, 80.1, 72.3, 28.7, 27.0, 26.5. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₂₃H₂₅N₂O₅ 409.1763; Found 409.1761.

1-methyl-3-((3a*R*,5*S*,5a*S*,8a*S*,8b*R*)-2,2,7,7-tetramethyltetrahydro-5*H*-bis([1,3]dioxolo)[4,5-b: 4',5'-d]pyran-5-yl)quinoxalin-2(1*H*)-one(4ae)



Colorless oil; 17.5 mg, 45% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.92 (dd, J = 8.0, 1.5 Hz, 1H), 7.58 – 7.53 (m, 1H), 7.35 – 7.30 (m, 1H), 7.28 (d, J = 8.8 Hz, 1H), 5.55 (d, J =2.5 Hz, 1H), 5.09 (dd, J = 9.9, 5.1 Hz, 1H), 5.01 (d, J = 10.0 Hz, 1H), 4.72 (d, J = 5.1 Hz, 1H), 4.37 (d, J = 2.2 Hz, 1H), 3.68 (s, 3H), 1.53 (s, 3H), 1.50 (s, 3H), 1.40 (s, 3H), 1.38 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 154.7, 154.0, 133.6, 132.5,

131.13, 131.08, 123.6, 113.5, 111.1, 109.1, 97.8, 77.3, 76.2, 74.7, 70.4, 69.2, 29.2, 28.0, 26.0, 25.9. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₂₀H₂₅N₂O₆ 389.1713; Found 389.1719.

1-methyl-3-((3aR,5R,5aS,8aS,8bR)-2,2,7,7-tetramethyltetrahydro-5H-bis([1,3]dioxolo)[4,5-b: 4',5'-d]pyran-5-yl)quinoxalin-2(1H)-one(4ae')



Colorless oil; 17.4 mg, 45% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.08 (dd, J = 8.0, 1.5 Hz, 1H), 7.56 – 7.51 (m, 1H), 7.34 (td, J = 7.7, 1.2 Hz, 1H), 7.30 (dd, J = 8.4, 1.2 Hz, 1H), 5.90 (d, J = 5.1 Hz, 1H), 5.46 (d, J = 2.3 Hz, 1H), 5.02 (dd, J = 7.7, 2.3 Hz, 1H), 4.75 (dd, J = 7.7, 2.5 Hz, 1H), 4.47 (dd, J = 5.2, 2.6 Hz, 1H), 3.68 (s, 3H), 1.59 (s, 3H), 1.44 (s, 3H), 1.37 (s, 3H), 1.26 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 154.5, 153.6,

133.0, 132.9, 130.8, 130.2, 123.7, 113.5, 109.8, 109.0, 97.1, 71.9, 71.2, 70.7, 68.2, 28.9, 26.1, 25.9, 25.0, 24.6. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₂₀H₂₅N₂O₆ 389.1713; Found 389.1712.

1-methyl-3-((3aS,5aR,8aR,8bS)-2,2,7,7-tetramethyltetrahydro-3aH-bis([1,3]dioxolo)[4,5-b:4', 5'-d]pyran-3a-yl)quinoxalin-2(1H)-one(4af)



Colorless oil; 33.8 mg, 87% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.92 (dd, J = 7.7, 1.7 Hz, 1H), 7.58 (t, J = 7.5 Hz, 1H), 7.35 – 7.30 (m, 2H), 5.73 (d, J = 2.4 Hz, 1H), 4.75 (dd, J = 8.0, 2.3 Hz, 1H), 4.32 (d, J = 8.0 Hz, 1H), 4.14 (dd, J = 12.9, 2.1 Hz, 1H), 3.96 (d, J = 13.0 Hz, 1H), 3.71 (s, 3H), 1.62 (s, 3H), 1.33 (s, 3H), 1.27 (s, 3H), 1.20 (s, 3H). ¹³C NMR (100 MHz,

Chloroform-*d*) & 153.2, 152.8, 134.0, 131.1, 130.9, 123.4, 113.5, 109.0, 108.7, 103.1, 70.9, 70.5, 70.2, 61.4, 29.0, 26.3, 25.9, 25.1, 24.4. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₂₀H₂₅N₂O₆ 389.1713; Found 389.1721.

1-methyl-3-((2*R*,3*R*,4*S*,5*R*,6S)-3,4,5,6-tetramethoxytetrahydro-2*H*-pyran-2-yl)quinoxalin-2(1 *H*)-one(4ag)



White solid, mp: 113-115 °C; 32.4 mg, 89% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.94 (dd, J = 8.0, 1.6 Hz, 1H), 7.60 (td, J = 7.9, 1.5 Hz, 1H), 7.37 (td, J = 8.3, 1.2 Hz, 1H), 7.33 (dd, J = 8.5, 1.2 Hz, 1H), 5.29 (d, J = 10.1 Hz, 1H), 4.91 (d, J = 3.5 Hz, 1H), 4.01 (dd, J = 10.1, 9.0 Hz, 1H), 3.76 (t, J = 9.3 Hz, 1H), 3.72 (s, 3H), 3.68 (s, 3H), 3.62 (s, 3H), 3.56 (s, 3H), 3.46 (s, 3H), 3.39 (dd, J)

J = 9.7, 3.6 Hz, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 155.1, 154.9, 133.7, 132.4, 131.1, 130.7, 123.7, 113.7, 98.2, 83.9, 81.9, 80.5, 67.1, 61.0, 60.3, 59.1, 55.7, 29.2. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₁₈H₂₅N₂O₆ 365.1713; Found 365.1718.

1-methyl-3-((2*R*,3*R*,4*S*,5*R*,6*S*)-3,4,5-tris(benzyloxy)-6-methoxytetrahydro-2*H*-pyran-2-yl)qui noxalin-2(1*H*)-one(4ah)



White solid, mp: 85-87 °C; 50.4 mg, 85% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.74 (dd, J = 8.0, 1.5 Hz, 1H), 7.49 – 7.45 (m, 1H), 7.33 – 7.29 (m, 4H), 7.27 – 7.13 (m, 8H), 6.86 (s, 5H), 5.31 (d, J = 8.9 Hz, 1H), 4.96 (d, J = 10.9 Hz, 1H), 4.82 (d, J = 10.9 Hz, 1H), 4.76 (dd, J = 11.9, 2.6 Hz, 2H),4.63 (d, J = 10.2 Hz, 1H), 4.61 (s, 1H), 4.50 (d, J = 11.6 Hz, 1H), 4.20 – 4.12 (m, 2H),

3.64 (dd, J = 9.3, 3.5 Hz, 1H), 3.52 (s, 3H), 3.47 (s, 3H). ¹³C NMR (150 MHz, Chloroform-*d*) δ 155.2, 154.6, 138.9, 138.4, 138.2, 133.6, 132.3, 131.0, 130.6, 128.5, 128.4, 128.2, 128.0, 127.9, 127.8, 127.59, 127.57, 127.0, 123.6, 113.6, 98.8, 82.5, 79.8, 79.6, 75.9, 74.7, 73.5, 55.6, 29.1. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₃₆H₃₇N₂O₆ 593.2652; Found 593.2645.

3-((2*R*,3*R*,4*S*,5*R*,6*S*)-6-methoxy-3,4,5-tris((4-methylbenzyl)oxy)tetrahydro-2*H*-pyran-2-yl)-1-methylquinoxalin-2(1*H*)-one(4ai)



White solid, mp: 93-95 °C; 55.8 mg, 88% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.83 (dd, J = 8.0, 1.5 Hz, 1H), 7.58 – 7.54 (m, 1H), 7.35 – 7.30 (m, 5H), 7.24 (d, J = 8.3 Hz, 1H), 7.16 (t, J = 7.3 Hz, 4H), 6.85 (d, J = 7.7 Hz, 2H), 6.75 (d, J = 7.7 Hz, 2H), 5.40 – 5.38 (m, 1H), 5.02 (d, J = 10.6 Hz, 1H), 4.90 – 4.81 (m, 3H), 4.71 – 4.67 (m, 2H), 4.56 (d, J = 11.5 Hz, 1H), 4.26 – 4.20 (m, 2H), 3.73 – 3.70 (m, 1H), 3.61 (s, 3H), 3.55 (s, 3H), 2.37 (s, 3H), 2.36 (s, 3H), 2.11 (s, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 155.4, 154.6, 137.6, 137.2, 136.7, 136.0, 135.5, 135.3,

133.6, 132.4, 130.9, 130.6, 129.2, 129.1, 128.5, 128.4, 128.3, 127.8, 123.5, 113.5, 99.0, 82.5, 79.58, 79.55, 77.4, 75.8, 74.5, 73.4, 55.6, 29.0, 21.3, 21.2, 21.1. HRMS (ESI) m/z: $[M+H]^+$ Calcd for C₃₉H₄₃N₂O₆ 635.3121; Found 635.3114.

1-methyl-3-((2*R*,3*R*,4*S*,5*R*,6*S*)-3,4,5-tris((4-bromobenzyl)oxy)-6-methoxytetrahydro-2*H*-pyra n-2-yl)quinoxalin-2(1*H*)-one(4aj)



White solid, mp: 101-103 °C; 68.8 mg, 83% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.80 (dd, J =8.0, 1.6 Hz, 1H), 7.61 – 7.56 (m, 1H), 7.43 (dd, J =8.2, 1.5 Hz, 4H), 7.37 – 7.33 (m, 1H), 7.28 – 7.26 (m, 1H), 7.23 – 7.18 (m, 4H), 7.01 (d, J = 7.9 Hz, 2H), 6.76 (d, J = 8.0 Hz, 2H), 5.30 (d, J = 9.0 Hz, 1H), 4.91 (d, J = 11.5 Hz, 1H), 4.79 (d, J = 11.5 Hz, 1H), 4.74 (d, J = 3.4 Hz, 1H), 4.71 – 4.63 (m, 3H), 4.47 (d, J = 12.3 Hz, 1H), 3.68 (dd, J = 9.5, 3.4 Hz, 1H), 3.60 (s, 3H), 3.54 (s, 3H). ¹³C NMR (100 MHz,

Chloroform-*d*) δ 154.8, 154.4, 137.8, 137.4, 137.1, 133.5, 132.2, 131.6, 131.5, 131.3, 130.9, 130.4, 129.7, 129.4, 129.2, 123.8, 121.9, 121.5, 121.1, 113.7, 98.5, 82.3, 79.8, 77.3, 74.9, 73.9, 72.5, 55.7, 29.1. HRMS (ESI) m/z: [M+H]⁺ Calcd for C₃₆H₃₄Br₃N₂O₆ 826.9967; Found 826.9976.

14. NMR spectra of compounds







S4. ¹H NMR spectra (400 MHz, CDCl₃) of compound 2d



S5. ¹H NMR spectra (400 MHz, CDCl₃) of compound 2e



S6. ¹H NMR spectra (400 MHz, CDCl₃) of compound 2f



S7. ¹H NMR spectra (400 MHz, CDCl₃) of compound 2g



S8. ¹H NMR spectra (400 MHz, CDCl₃) of compound **2h**



S9. ¹H NMR spectra (400 MHz, CDCl₃) of compound 2i



S10. ¹H NMR spectra (400 MHz, CDCl₃) of compound 2j



S11. ¹H NMR spectra (400 MHz, CDCl₃) of compound 2k



S12. ¹H NMR spectra (400 MHz, CDCl₃) of compound 5



100 90 f1 (ppm) S14. ^{13}C NMR spectra (100 MHz, CDCl_3) of compound $\boldsymbol{3aa}$



S16. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3ab**



S18. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3ac**



 $S20. \ {}^{13}C \ NMR \ spectra (100 \ MHz, CDCl_3) \ of \ compound \ 3ad$



S22. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3ae**





S24. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3af**



S26. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3ag**



S28. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3ah**





S30. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3ai**



S32. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3aj**



130 120 110 100 f1 (ppm)

S34. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3ak**



S35. ¹H NMR spectra (400 MHz, CDCl₃) of compound **3al**



S36. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3al**



S38. ^{13}C NMR spectra (100 MHz, CDCl_3) of compound 3ba



S40. 13 C NMR spectra (100 MHz, CDCl₃) of compound **3ca**



S42. ^{13}C NMR spectra (100 MHz, CDCl_3) of compound $\boldsymbol{3da}$



S44. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3ea**



S46. ^{13}C NMR spectra (100 MHz, CDCl_3) of compound 3fa



S48. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3ga**



S50. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3ha**



S52. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3ia**



S54. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3ja**



S56. ¹³C NMR spectra (100 MHz, CDCl₃) of compound 3ka


-100 -110 -120 -130 -140 -150 -160 -170 -180 f1 (ppm) -190 -200 -210 -30 -40 -50 -60 -70 -80 -90

S58. ¹⁹F NMR spectra (376 MHz, CDCl₃) of compound **3la**

-20



S59. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3la**



S60. ¹H NMR spectra (400 MHz, CDCl₃) of compound **3ma**



S61. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3ma**



S62. ¹H NMR spectra (400 MHz, CDCl₃) of compound **3na**



S63. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3na**



S64. ¹H NMR spectra (400 MHz, CDCl₃) of compound **30a**



S65. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **30a**



S66. ¹H NMR spectra (400 MHz, CDCl₃) of compound **3pa**



S67. ¹³C NMR spectra (100 MHz, CDCl₃) of compound **3pa**



S68. ¹H NMR spectra (400 MHz, CDCl₃) of compound 4aa



S69. ¹³C NMR spectra (100 MHz, CDCl₃) of compound 4aa



S70. ¹H NMR spectra (400 MHz, CDCl₃) of compound 4ab



S71. ¹³C NMR spectra (100 MHz, CDCl₃) of compound 4ab



S72. ¹H NMR spectra (400 MHz, CDCl₃) of compound 4ac



S73. ¹³C NMR spectra (100 MHz, CDCl₃) of compound 4ac



S74. ¹H NMR spectra (400 MHz, CDCl₃) of compound 4ac'



S75. ¹³C NMR spectra (100 MHz, CDCl₃) of compound 4ac'



S76. ¹H NMR spectra (600 MHz, CDCl₃) of compound 4ad



S77. ¹³C NMR spectra (150 MHz, CDCl₃) of compound 4ad



S78. ¹H NMR spectra (600 MHz, CDCl₃) of compound 4ad'



S79. ¹³C NMR spectra (150 MHz, CDCl₃) of compound 4ad'





S85



S84. ¹H NMR spectra (400 MHz, CDCl₃) of compound 4af



S85. ¹³C NMR spectra (100 MHz, CDCl₃) of compound 4af



S86. ¹H NMR spectra (400 MHz, CDCl₃) of compound 4ag



S87. ¹³C NMR spectra (100 MHz, CDCl₃) of compound 4ag



S88. ¹H NMR spectra (400 MHz, CDCl₃) of compound 4ah



S89. ¹³C NMR spectra (150 MHz, CDCl₃) of compound 4ah



S90. ¹H NMR spectra (400 MHz, CDCl₃) of compound 4ai



S91. ¹³C NMR spectra (100 MHz, CDCl₃) of compound 4ai



S92. ¹H NMR spectra (400 MHz, CDCl₃) of compound 4aj



S93. ¹³C NMR spectra (100 MHz, CDCl₃) of compound 4aj