

Supporting Information

Site-specific acetyl modification of 2'-OH of RNA by the oligonucleotide acetylating reagent

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1. General

High performance liquid chromatography (HPLC) was performed on a JASCO LC-2000 PLUS series equipped with YMC-Triart Bio C18 column (4.6 x 250 mm). UPLC/QTOF-MS and MSMS measurement were performed on a Waters ACQUITY UPLC Xevo G2-XS QToF equipped with Oligonucleotide BEH C18 column (2.1 x 50 mm). The UV spectra measurement and the determination of the concentration of oligonucleotides were performed by Thermo Scientific NanoDrop One. The DNA and RNA oligomer were purchased from Japan Bio Services Co., LTD. S-(bromomethyl) ethanethioate was purchased from Enamine Ltd.

2. General procedure of preparation of Ac-probe

To the 95 μ L solution of 50 μ M oligonucleotide probe containing a 6-thioguanine in 250 mM CAPS buffer pH10 was added 5 μ L 10 % S-(bromomethyl) ethanethioate in ACN (v/v, final 0.5 %), and the reaction mixture was incubated at 0 °C for 1h. After filtration, HPLC purification was performed (YMC-Triart Bio C18 column (4.6 x 250 mm), A: 0.1 M TEAA buffer pH7 containing 5 % ACN, B: ACN, B % [0 to 30 % over 20 min], UV: 254 nm, flow rate: 1 mL/min, column oven: 40 °C). The separated solution was concentrated with (Amicon® Ultra 3K, 25 °C, 14,000xg, 25 min). After ultrapure water exchange with Amicon, pure Ac-probe was obtained. The determination of the concentration of oligonucleotide was performed by the measurement of the 260 nm absorption.

3. General procedure of acetyl modification of RNA

The following 10 μ L solution was prepared on an ice bath and allowed to stand at respective temperatures. [Ac-probe (7.5 μ M), target RNA (5 μ M) and NaCl (100 mM) in buffer solution (50 mM, pH10: carbonate, pH8.5 and pH7.4: phosphate)]

4. UV spectra measurement

The following solutions were prepared and their UV absorption were measured by NanoDrop One (Thermo Scientific). [probe or Ac-probe (7.5 μ M) in buffer solution (50 mM, pH5.2: sodium acetate, pH7.4: HEPES-NaOH, pH9.0: Tris-HCl, pH10: carbonate)

5. Analysis of acetyl modification of 131 mer RNA

131 mer RNA (5'-ggg ucu aga guu uaa cuu uaa gaa gga gau aua cau aug gcu agc aug acu ggu gga cag caa aug ggu acc gaa uuc aag acc gcg caa gac uac aag gac gac gac gau aag uag uga aua acu aaU cc-3') was prepared with general transcription method using T7-RNA polymerase from the purchased DNA template containing T7 promoter. Acetylation reaction of 131 mer RNA was analyzed by UPLC/MS and the MS numbers were obtained as ions adduct.

ACQUITY UPLC: Oligonucleotide BEH C18 Column, 130Å, 1.7 μ m, 2.1x50 mm, A: 15 mM TEA, 400 mM HFIP pH7.9, B: A/MeOH=1/1, D:35% to 70% /10min, Flow rate: 0.2 mL/min, Column oven: 40 °C.

MS conditions: Xevo G2-XS Qtof, Mass range: 450 – 3000 Da, Internal Standard: LE, Mode: ESI negative resolution, continuum, Cone voltage: 40 V, Capillary voltage: 4 kV, desolvation: 600 °C, desolvation gas flow: 700 L/hr.

The target MS numbers were set as below; 131 mer RNA [927.797 - 928.140 (46 valent ion)], Ac-RNA [928.703 – 929.021 (46 valent ion)].

6. Supplemental data

Name	Sequence	calcd. M.W.	found
Ac-probe-1	5'd-GTA GTC TTX' CGC GGT	4719	4719
regenerated probe-1	5'd-GTA GTC TTX CGC GGT	4631	4631
Ac-probe-2	5'd-CTT TX'T TCT CCT TTC T	4859	4859
RNA (N=a)	3'r-cau cag aaa gcg cca	4781	4781
Ac-RNA (N=a)		4823	4823
RNA (N=g)	3'r-cau cag aag gcg cca	4797	4797
Ac-RNA (N=g)		4839	4839
RNA (N=c)	3'r-cau cag aac gcg cca	4757	4757
Ac-RNA (N=c)		4799	4799
RNA (N=u)	3'r-cau cag aaU gcg cca	4758	4758
Ac-RNA (N=u)		4800	4800

Table S1 Sequences and MS numbers.

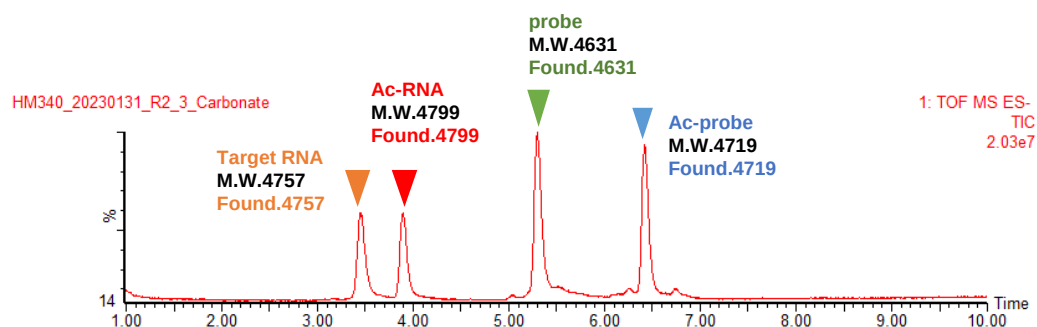


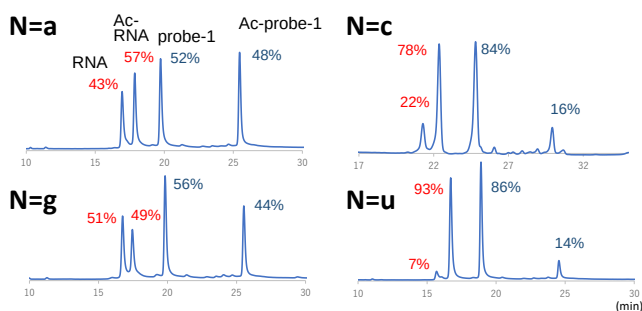
Fig. S1 TIC chart of the acetyl modification reaction under the condition of pH10 and 37 °C for 1h.

ACQUITY UPLC: Oligonucleotide BEH C18 Column, 130Å, 1.7µm, 2.1x50 mm, A: 15 mM TEA, 400 mM HFIP pH7.9, B: A/MeOH=1/1, D:35% to 70% /10min, Flow rate: 0.2 mL/min, Column oven: 40 °C.

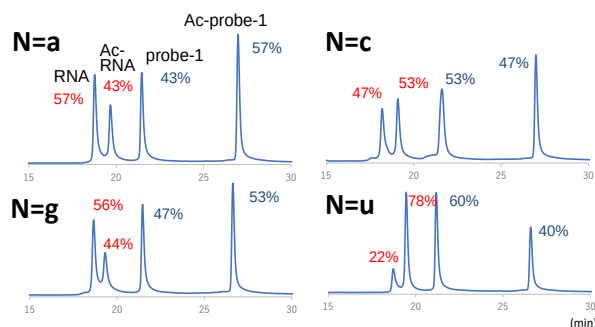
MS conditions: Xevo G2-XS Qtof, Mass range: 450 – 3000 Da, Internal Standard: LE, Mode: ESI negative resolution, continuum, Cone voltage: 40 V, Capillary voltage: 4 kV, desolvation: 600 °C, desolvation gas flow: 700 L/hr.

a)

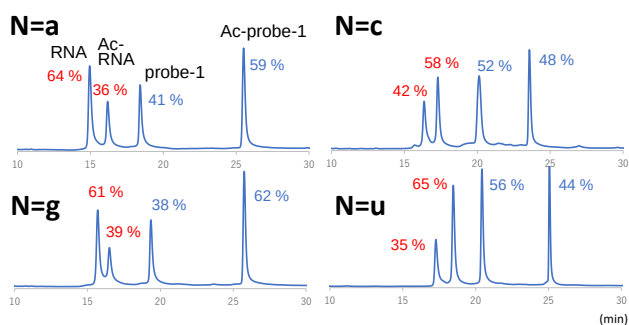
Carbonate Buffer pH10, 15 °C, 19 h



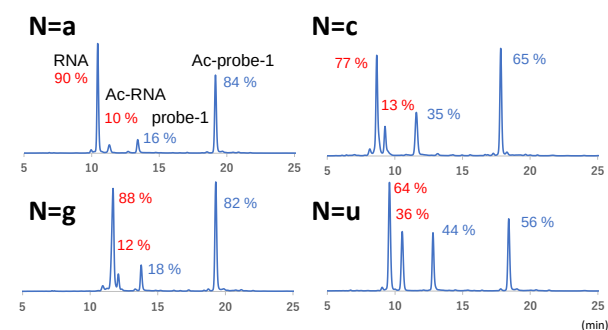
Carbonate Buffer pH10, 37 °C, 1 h



Phosphate Buffer pH8.5, 37 °C, 22 h



Phosphate Buffer pH7.4, 37 °C, 22 h



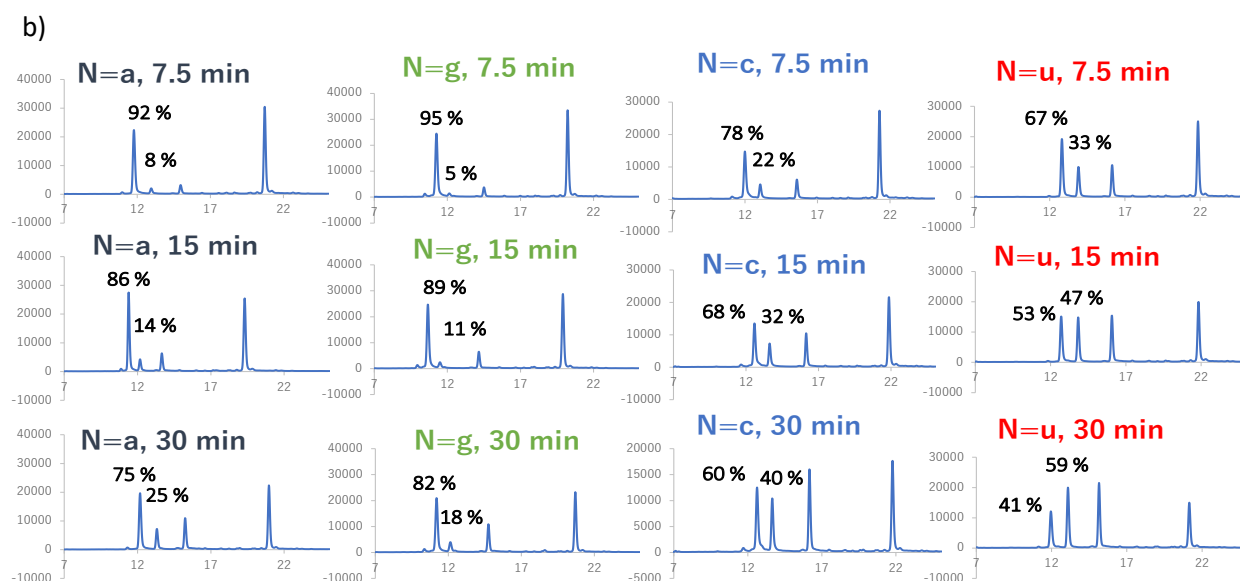


Fig. S2 HPLC charts of the acetyl modification. a) under various conditions, b) for time-course plots under pH10 at 37 °C.

YMC-Triart Bio C18 column (4.6 x 250 mm), A: 0.1 M TEAA buffer pH7 containing 5 % ACN, B: ACN, B % [0 to 10 % over 30 min], UV: 254 nm, flow rate: 1 mL/min, column oven: 40 °C.

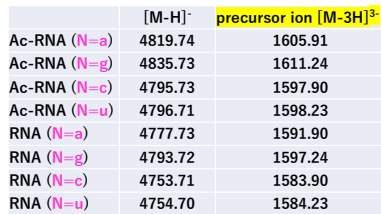


Fig. S3 MS/MS fragment analysis of the acetylated RNA and unmodified RNA.

LC conditions, ACQUITY UPLC: Oligonucleotide BEH C18 Column, 130Å, 1.7µm, 2.1x50 mm, A: 15 mM TEA, 400 mM HFIP pH7.9, B: A/MeOH=1/1, D:35% to 70% /10min, Flow rate: 0.2 mL/min, Column oven: 40 °C.

MS/MS conditions, MS system: Xevo G2-XS Qtof, Mass range: 450 – 2500 Da, Mode: ESI negative resolution, continuum, Capillary voltage: 2.7 kV, Cone voltage: 31 V, source: 120 °C, desolvation: 300 °C, desolvation gas flow: 500 L/hr, precursor ion: $[M-3H]^{3-}$, LockMass: 1685.765 ($Cs_6I_7^-$), scan time: 0.2sec, collision energy ramp: 25 to 55 V.

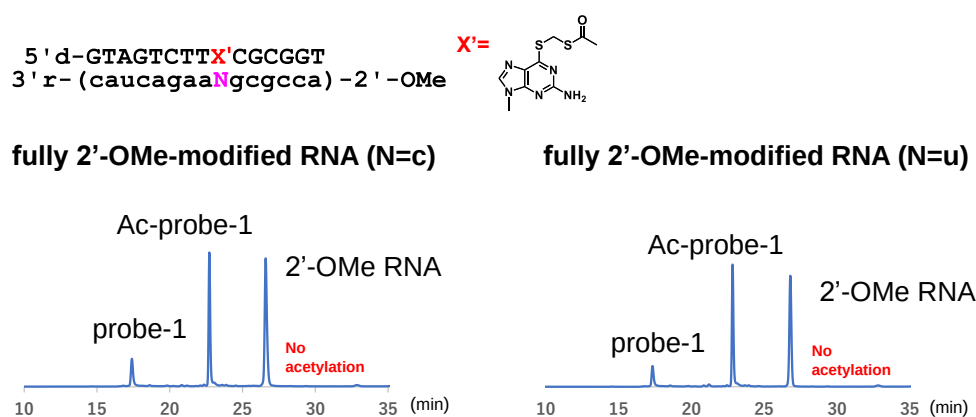


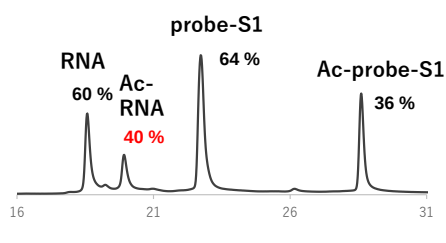
Fig. S4 The results of Ac-probe-1 vs. fully 2'-OMe-modified RNAs under the condition of pH10, 37°C for 1 h.

YMC-Triart Bio C18 column (4.6 x 250 mm), A: 0.1 M TEAA buffer pH7 containing 5 % ACN, B: ACN, B % [0 to 10 % over 30 min], UV: 254 nm, flow rate: 1 mL/min, column oven: 40 °C.

a)

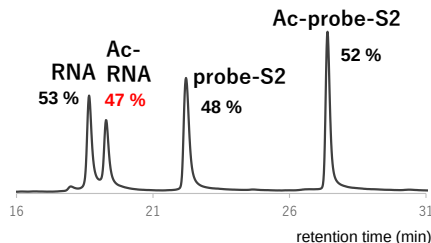
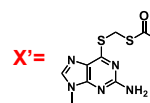
Ac-probe-S1

5' d-GTAX'**X'**TCTTGC GCGGT
3' r-cau**c** agaacgcgcca



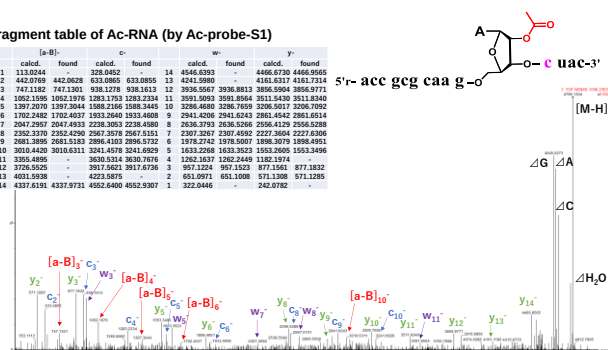
Ac-probe-S2

5' d-GTAGTCTTGC**X'**CGGT
3' r-caucagaacg**c**gcc



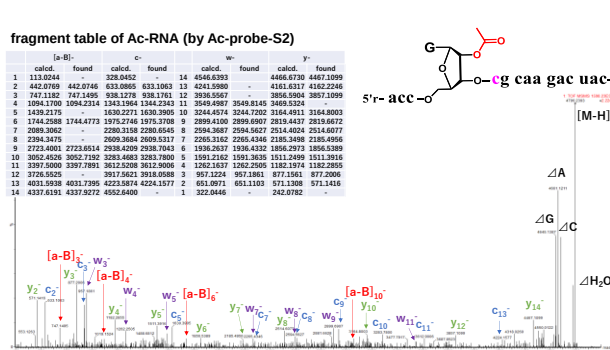
fragment table of Ac-RNA (by Ac-probe-S1)

	[a-B]		c-		w-		y-	
	calcd.	found	calcd.	found	calcd.	found	calcd.	found
1	113.0244	-	328.0452	-	454.6393	-	446.6720	446.6709
2	442.0789	442.0628	633.0895	633.0855	13	454.6398	-	416.6337
3	747.1182	747.1261	938.1278	938.1613	12	3936.0567	3936.0813	3856.9771
4	1052.1595	1052.1576	1353.1753	1353.2354	11	3593.5093	3593.5064	3513.5450
5	1397.2070	1397.2044	1688.2166	1688.2445	10	3286.4680	3286.4659	3206.5017
6	1702.2482	1702.4017	1933.2460	1933.4001	9	2943.4006	2943.4241	2863.4542
7	2047.2957	2047.4933	2238.3053	2238.4980	8	2636.3793	2636.5266	2556.4129
8	2352.3370	2352.4290	2587.3578	2587.5151	7	2307.3587	2307.4952	2227.3664
9	2681.3895	2681.5193	2896.4183	2896.5752	6	1978.2742	1978.5007	1898.3079
10	3010.4420	3010.6111	3241.4578	3241.6501	5	1633.2386	1633.3522	1553.2695
11	3355.4885	-	3630.5314	3630.7670	4	1282.1837	1282.2449	1182.1974
12	3726.5625	-	3917.5821	3917.6730	3	967.1224	967.1323	877.1561
13	4031.5938	-	4273.5875	-	2	653.6071	651.1088	571.1306
14	4337.6191	4337.7331	4552.6400	4552.9307	1	322.0446	-	242.0782



fragment table of Ac-RNA (by Ac-probe-S2)

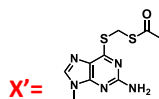
	[a-B]		c-		w-		y-	
	calcd.	found	calcd.	found	calcd.	found	calcd.	found
1	113.0244	-	328.0452	-	14	454.6393	-	446.6720
2	442.0789	442.0746	633.0895	633.1083	13	454.6398	-	416.6337
3	747.1182	747.1495	938.1278	938.1761	12	3936.0567	3936.0813	3856.9771
4	1052.1595	1054.2314	1343.1964	1344.2343	11	3549.6987	3549.8145	3469.5324
5	1409.2179	-	1630.2771	1630.3901	10	3244.4674	3244.7202	3164.4811
6	1744.2388	1744.4773	1975.2740	1975.3708	9	2899.4100	2899.6007	2819.4437
7	2089.3062	-	2300.3158	2300.6541	8	2554.3687	2554.5617	2474.4002
8	2394.3476	-	2609.3684	2609.5317	7	2205.3162	2205.4346	2125.3498
9	2723.4001	2723.6514	2938.4209	2938.7043	6	1856.2617	1856.4332	1776.2973
10	3052.4526	3052.7282	3263.4683	3263.7800	5	1501.2162	1501.3638	1421.2499
11	3397.5050	3397.7891	3612.5208	3612.9004	4	1146.1637	1146.2959	1066.1974
12	3726.5625	-	3917.5623	3918.0588	3	897.1224	897.1361	817.1561
13	4031.5938	4031.7389	4223.5874	4224.1577	2	651.0971	651.1103	571.1306
14	4337.6191	4337.8072	4552.6400	-	1	322.0446	-	242.0782



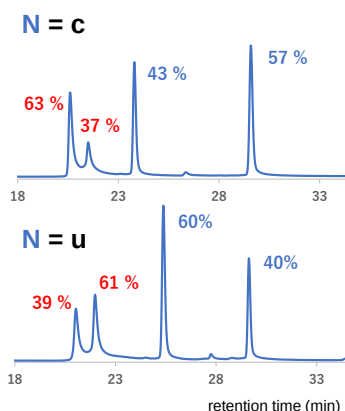
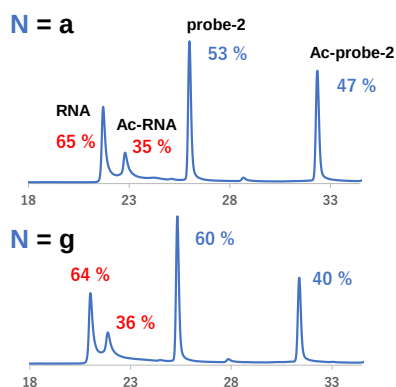
b)

Ac-probe-2

5' d-CTTT**X'**TTCTCCTTTCT
3' r-gaaa**N** aagaggaaaga



N = a, g, c or u



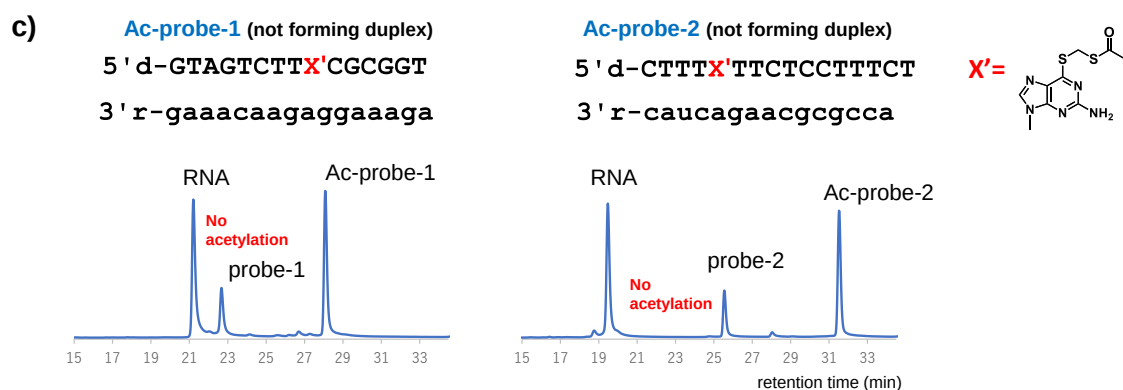


Fig. S5 The variation of modification reactions under the condition of pH10 and 37°C for 1 h. a) using Ac-probe-S with a 6-thioguanine reactive site (X') at different position (upper part), MS/MS analysis of the product RNAs (lower part), b) using Ac-probe-2 and its complementary target RNAs, c) using the pairs of Ac-probe and target RNA not forming a duplex.