

Electronic Supporting Information

Diselenide as novel effective fluorescence quenchers to construct a two-photon fluorescent probe for thiols in a mouse stroke model

Tao Ma^{a#}, Hao Gao^{c#}, Jun Wu^a, Jintao Zhao^a, Bingbing Chang^a, Zhibin Zhang^a, Shengxiang Zhang^{c*}, Baoxin Zhang^{a*}, and Jianguo Fang^{a, b}

^a State Key Laboratory of Applied Organic Chemistry, College of Chemistry and Chemical Engineering, Lanzhou University, Lanzhou 730000, China.

^b School of Chemistry and Chemical Engineering, Nanjing University of Science & Technology, Nanjing, Jiangsu 210094, China.

^c Gansu Key Laboratory of Biomonitoring and Bioremediation for Environmental Pollution, School of Life Sciences, Lanzhou University, Lanzhou 730000, China

[#]T.M. and H.G. contributed equally to this work.

*Corresponding authors: E-Mail: sxzhang@lzu.edu.cn

E-Mail:zhangbx@lzu.edu.cn

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1. Experimental Section

1.1 Materials:

All chemicals and reagents were used directly as obtained commercially unless otherwise stated. Solvents were purified and dried by standard procedures before use. MiliQ water was used in preparing aqueous systems⁰ if necessary, double distilled water was used throughout the other experiments. Dimethyl sulfoxide (DMSO) and esterase from porcine liver were obtained from Sigma-Aldrich (St. Louis, MO, USA). N-Acetylcysteine (NAC) and lipopolysaccharides (LPS) were obtained from Aladdin (Shanghai, China). Other reagents were analytical grade purchased from local supplier.

1.2 Instruments:

¹H NMR (400 MHz) and ¹³C NMR (151 MHz) analyses were performed on a Bruker Advance spectrometer. ESI-MS spectra were recorded on a Bruker Daltonics esquire 6000 mass spectrometer. High-resolution mass spectrometry (HRMS) was performed on an Orbitrap Q Exactive (Thermo Scientific). UV-vis spectra were recorded by a UV-vis spectrometer (Evolution 200, Thermo Scientific). Fluorescence spectroscopic studies were performed using a Cary Eclipse fluorescence spectrophotometer (Agilent Technologies). The fluorescence images of the cells were acquired on a Fluid cell imaging station (Life Technologies). The fluorescence images the brain of mice were taken with a two-photon fluorescence microscope (Olympus FV1000). All procedures for *in vivo* experiments were carried out in accordance with the institutional guidelines (Guidance of the Care and Use of Laboratory Animals), and all *in vivo* experiments were approved by the Ethics Committee of Lanzhou University, China.

1.3 Spectral measurements:

Stock solution of AMC, AMC-SS, AMC-SeSe, AFC, AFC-SS, AFC-SeSe, ANC, ANC-SS, ANC-SeSe, ARC, ARC-SS, ARC-SeSe (10 mM) was prepared in DMSO. Stock solutions of various amino acids and other analytes were prepared by direct dissolution in deionized water or DMSO. All chemicals used were of analytical grade. The detection experiments were measured in PBS buffer (pH =7.4, 10 mM).

Imaging of probes in Living Cells

Hela cells (2×10^4) were seeded in 12-well plates and incubated for 24 h at 37 °C. To determine the membrane permeability, HeLa cells were treated with **AFC-SeSe** (10 μ M), **AFC-SS** (10 μ M), **AMC-SeSe** (10 μ M) and **AMC-SS** (10 μ M) for 30 min or 60 min; Then the cells were washed three times with PBS, followed by imaging under a Fluid cell imaging station microscope.

To image probes reacted with intracellular thiols, First, Hela cells were pretreated with NEM (100 μ M) for 30 min, following treated with GSH (1 mM), Cys (100 μ M) and Hcy (100 μ M), respectively, and then incubated with **AFC-SeSe** (10 μ M), **AFC-SS** (10 μ M) for 30 min, Then the cells were washed three times with PBS, followed by imaging under a Fluid cell imaging station microscope.

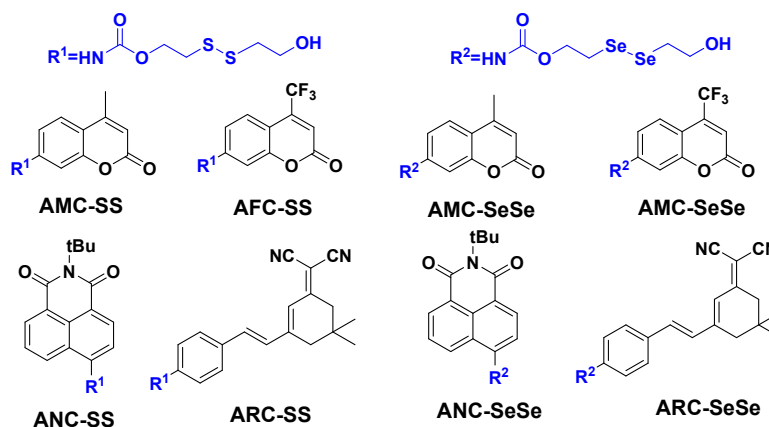
To image probes reacted with intracellular thiols under oxidation condition, First, Hela cells were pretreated with H₂O₂ (100 μ M) for 30 min or LPS (100 μ M) for 12 h, following treated with NAC (100 μ M), respectively, and then incubated with **AFC-SeSe** (10 μ M), **AFC-SS** (10 μ M) for 30 min, Then the cells were washed three times with PBS, followed by imaging under a Fluid cell imaging station microscope.

Two-Photon Imaging of AFC-SeSe and AFC-SS in Mouse Brains.

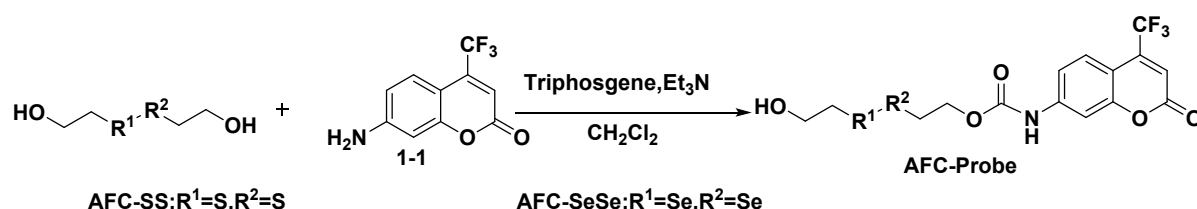
The cerebral tissue underwent incubation with **AFC-SeSe or AFC-SS** via a cranial window positioned above the mouse skull. A ϕ 3 mm area along the mouse skull perimeter was meticulously thinned using a dental drill. Subsequently, the bone flap was delicately removed using curved forceps. Throughout the surgical procedure, a sterilized artificial cortical-spinal solution (ACSF) was intermittently applied to the exposed brain tissue to prevent desiccation, while the animal's body temperature was maintained using a heating pad. Any residual blood was cleaned using a piece of moistened gelfoam. Following this, **AFC-SeSe or AFC-SS** (1 mM, 20 μ L) was administered to the brain tissue, and a cover glass (ϕ 4 mm) was affixed to the skull and sealed using dental cement. After a 30-minute incubation period, the mouse was gently repositioned beneath a $25 \times$ water-immersion objective (Olympus FV1000) for intravital imaging. The cortical region was imaged repeatedly through the cranial window, and 1024*1024 image stacks were acquired with a step size of 1 μ m. It was important to minimize exposure of the cortex to the laser to mitigate potential tissue

damage. The excitation wavelength for two-photon imaging was set at 800 nm.

2. Synthesis and Characterization:



Scheme S1. Chemical structures of fluorescence probes.



Scheme S2. The synthesis route of probe **AFC-SS** and **AFC-SeSe**.

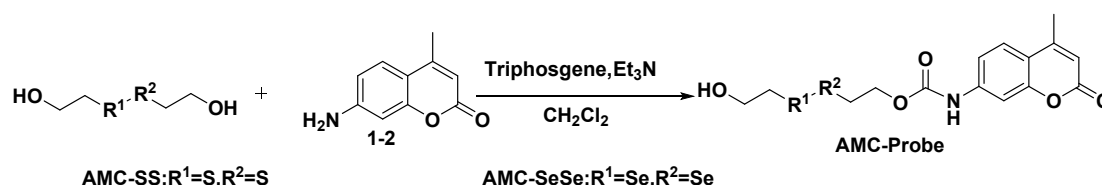
Synthesis details of **AFC-SS**:

Compounds **1-1** (100 mg, 0.37 mmol) was dissolved in DCM (5 mL) and then added to a 50 mL round-bottled flask, and triphosgene (56 mg, 0.19 mmol) was added to the system in batches at 0 °C followed with triethylamine (154 μ L, 1.11 mmol). After the reaction at room temperature for 30 min, bis (2-hydroxyethyl) disulfide (171 mg, 1.11 mmol) was added to the reaction system, and the reaction continued at room temperature for 6 h. After the reaction, the saturated sodium bicarbonate (5 mL) was added to quench the reaction, the organic phase was separated by DCM (3 x 6 mL), finally the solvent was removed, and yellow solid **AFC-SS** was obtained by column chromatography (PE/EA = 2/1) (36 mg, 42% yield). ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 10.41 (d, J = 13.2 Hz, 1H), 7.67-7.60 (m, 2H), 7.51-7.47 (m, 1H), 6.86 (d, J = 2.5 Hz, 1H), 4.90 (t, J = 5.4 Hz, 1H), 4.38 (t, J = 6.3 Hz, 2H), 3.63 (td, J = 6.4, 5.3 Hz, 2H), 3.04 (t, J = 6.3 Hz, 2H), 2.82 (t, J = 6.4 Hz, 2H).; ^{13}C NMR (151 MHz, $\text{DMSO-}d_6$) δ

159.10, 155.31, 153.48, 144.15, 125.97, 121.24, 115.60, 114.49, 108.09, 105.56, 63.13, 59.85, 41.51, 37.05.; ESI-MS (m/z): calculated for $C_{21}H_{25}N_2O_5S_2$ $[M-H]^+$ 408.01, found 408.05.

Synthesis details of **AFC-SeSe**:

The synthesis route is similar to that of compound **AFC-SS**, and yellow solid **AFC-SeSe** is obtained by column chromatography (PE/EA = 2/1) (35 mg, 39% yield). 1H NMR (400 MHz, DMSO- d_6) δ 10.40 (d, J = 17.5 Hz, 1H), 7.70-7.39 (m, 3H), 6.87 (s, 1H), 5.02-4.87 (m, 1H), 4.41 (t, J = 6.8 Hz, 2H), 3.78-3.59 (m, 2H), 3.29-3.19 (m, 2H), 3.15-2.91 (m, 2H).; ^{13}C NMR (151 MHz, DMSO- d_6) δ 158.03, 154.14, 152.39, 143.11, 124.92, 124.72, 107.03, 106.83, 104.52, 104.38, 32.39, 32.11.; calculated for $C_{15}H_{14}F_3NO_5Se_2$ $[M-H]^-$ 503.90, found 503.91.



Scheme S3. The synthesis route of probe **AMC-SS** and **AMC-SeSe**.

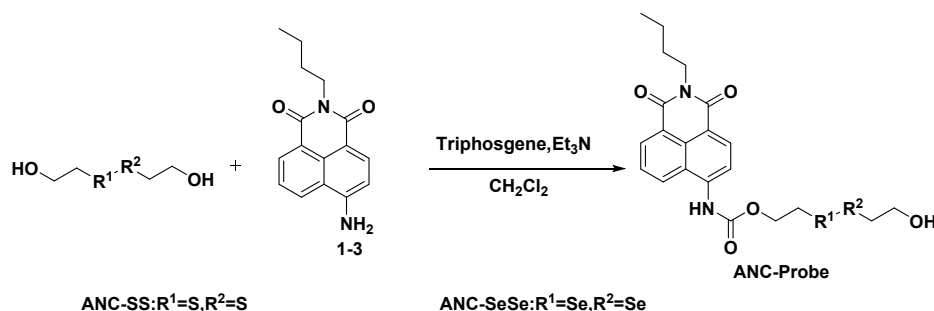
Synthesis details of **AMC-SS**:

The synthesis route is similar to that of compound **AFC-SS**, and yellow solid **AMC-SS** is obtained by column chromatography (PE/EA = 3/1) (47 mg, 51% yield). 1H NMR (400 MHz, DMSO- d_6) δ 10.20 (d, J = 14.3 Hz, 1H), 7.65 (d, J = 8.7 Hz, 1H), 7.51 (d, J = 2.0 Hz, 1H), 7.38 (dd, J = 8.7, 2.1 Hz, 1H), 6.18 (dd, J = 17.3, 1.5 Hz, 1H), 4.87 (t, J = 5.5 Hz, 1H), 4.33 (t, J = 6.2 Hz, 2H), 3.60 (q, J = 6.2 Hz, 2H), 3.00 (t, J = 6.3 Hz, 2H), 2.79 (t, J = 6.4 Hz, 2H), 2.32 (dd, J = 15.3, 1.2 Hz, 3H).; ^{13}C NMR (151 MHz, DMSO- d_6) δ 160.49, 154.25, 153.60, 153.51, 143.09, 126.42, 114.84, 114.75, 114.73, 112.39, 104.95, 62.96, 59.87, 41.53, 37.12, 18.45.

Synthesis details of **AMC-SeSe**:

The synthesis route is similar to that of compound **AFC-SS**, and yellow solid **AMC-SeSe** is obtained by column chromatography (PE/EA = 2/1) (34 mg, 29% yield). 1H NMR (400 MHz, DMSO- d_6) δ 10.24 (s, 1H), 7.68 (d, J = 9.0 Hz, 1H), 7.60-7.47 (m, 1H), 7.45-7.33 (m, 1H), 6.23 (s, 1H), 5.02-4.91 (m, 1H), 4.38 (s, 2H), 3.75-

3.63 (m, 2H), 3.30-2.94 (m, 4H), 2.37 (s, 3H).; ^{13}C NMR (151 MHz, $\text{DMSO-}d_6$) δ 160.51, 154.27, 153.66, 153.41, 143.07, 126.49, 126.30, 114.65, 112.40, 104.85, 64.88, 61.59, 33.44, 27.73, 18.47.; calculated for $\text{C}_{15}\text{H}_{14}\text{F}_3\text{NO}_5\text{Se}_2$ $[\text{M-H}]^-$ 449.94, found 449.90.



Scheme S4. The synthesis route of probe **ANC-SS** and **ANC-SeSe**.

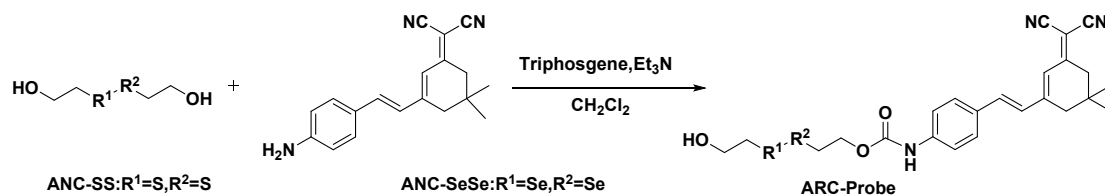
Synthesis details of **ANC-SS**:

The synthesis route is similar to that of compound **AFC-SS**, and yellow solid **ANC-SS** is obtained by column chromatography (PE/EA = 2/1) (47 mg, 39% yield). ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 10.37 (s, 1H), 8.69 (d, J = 8.5 Hz, 1H), 8.48 (dd, J = 19.6, 7.8 Hz, 2H), 8.16 (d, J = 8.3 Hz, 1H), 7.83 (t, J = 7.9 Hz, 1H), 4.94 (s, 1H), 4.46 (t, J = 6.5 Hz, 2H), 4.03 (t, J = 7.5 Hz, 2H), 3.66 (d, J = 6.6 Hz, 2H), 3.11 (t, J = 6.5 Hz, 2H), 2.86 (t, J = 6.4 Hz, 2H), 1.61 (p, J = 7.3 Hz, 2H), 1.35 (q, J = 7.5 Hz, 2H), 0.93 (t, J = 7.4 Hz, 3H).; ^{13}C NMR (151 MHz, $\text{DMSO-}d_6$) δ 163.92, 163.37, 154.35, 141.08, 132.09, 132.00, 131.37, 129.81, 128.77, 126.86, 124.43, 122.67, 118.97, 117.64, 63.46, 59.88, 41.53, 37.14, 30.14, 20.28, 14.21.

Synthesis details of **ANC-SeSe**:

The synthesis route is similar to that of compound **AFC-SS**, and yellow solid **ANC-SeSe** is obtained by column chromatography (PE/EA = 2/1) (27 mg, 19% yield). ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 10.34 (s, 1H), 8.71 (d, J = 8.5 Hz, 1H), 8.49 (dd, J = 20.4, 7.7 Hz, 2H), 8.17 (d, J = 8.2 Hz, 1H), 7.84 (dd, J = 8.6, 7.2 Hz, 1H), 5.00 (s, 1H), 4.47 (t, J = 7.0 Hz, 2H), 4.04 (t, J = 7.5 Hz, 2H), 3.68 (t, J = 6.8 Hz, 2H), 3.27 (t, J = 7.0 Hz, 2H), 3.09 (t, J = 6.8 Hz, 2H), 1.61 (dd, J = 8.5, 6.5 Hz, 2H), 1.37-1.33 (m, 2H), 0.93 (s, 3H).; ^{13}C NMR (151 MHz, $\text{DMSO-}d_6$) δ 163.95, 163.41, 154.33, 141.15, 132.11, 131.39, 129.86, 128.81, 126.88, 124.50, 122.70, 119.02, 117.65, 65.35, 61.72,

33.48, 33.19, 30.15, 22.56, 20.28, 14.18.; calculated for $C_{22}H_{23}N_2O_5Se_2$ $[M-H]^-$ 543.00, found 543.01.



Scheme S5. The synthesis route of probe **ARC-SS** and **ARC-SeSe**.

Synthesis details of **ARC-SS**:

The synthesis route is similar to that of compound **AFC-SS**, and red solid **ARC-SS** is obtained by column chromatography (PE/EA = 2/1) (36 mg, 42% yield). 1H NMR (400 MHz, $CDCl_3-d_4$) δ 7.48-7.42 (m, 4H), 7.10 (s, 1H), 7.03-6.88 (m, 2H), 6.81 (s, 1H), 4.46 (t, $J = 6.0$ Hz, 2H), 3.92 (t, $J = 6.0$ Hz, 2H), 3.00 (t, $J = 6.4$ Hz, 2H), 2.93 (t, $J = 6.0$ Hz, 2H), 2.59 (s, 2H), 2.45 (s, 2H), 1.07 (s, 6H); ^{13}C NMR (151 MHz, $CDCl_3-d_4$) δ 169.31, 154.02, 152.91, 139.13, 136.42, 130.95, 128.64, 127.92, 123.23, 118.75, 113.66, 112.82, 78.18, 63.21, 60.3, 43.0, 41.6, 39.1, 37.51, 32.03, 28.04.

Synthesis details of **ARC-SeSe**:

The synthesis route is similar to that of compound **AFC-SS**, and red gelatinous substance **ARC-SeSe** is obtained by column chromatography (PE/EA = 2/1) (16 mg, 12% yield). 1H NMR (400 MHz, $CDCl_3-d_4$) δ 7.45 (d, $J = 3.6$ Hz, 4H), 7.01 (d, $J = 16.0$ Hz, 1H), 6.89 (d, $J = 15.8$ Hz, 1H), 6.81 (s, 1H), 4.54 (t, $J = 23.5, 6.7$ Hz, 4H), 3.30 (t, $J = 77.0, 6.7$ Hz, 4H), 2.60 (s, 2H), 2.46 (s, 2H), 1.08 (s, 6H); ^{13}C NMR (151 MHz, $CDCl_3-d_4$) δ 169.24, 153.95, 136.35, 130.99, 129.66, 128.60, 128.57, 128.06, 127.69, 123.24, 118.87, 113.56, 112.84, 64.59, 43.00, 39.22, 32.05, 29.71, 28.12, 28.04.; calculated for $C_{15}H_{14}F_3NO_5Se_2$ $[M-H]^-$ 562.03, found 562.08.

3. Additional Test data:

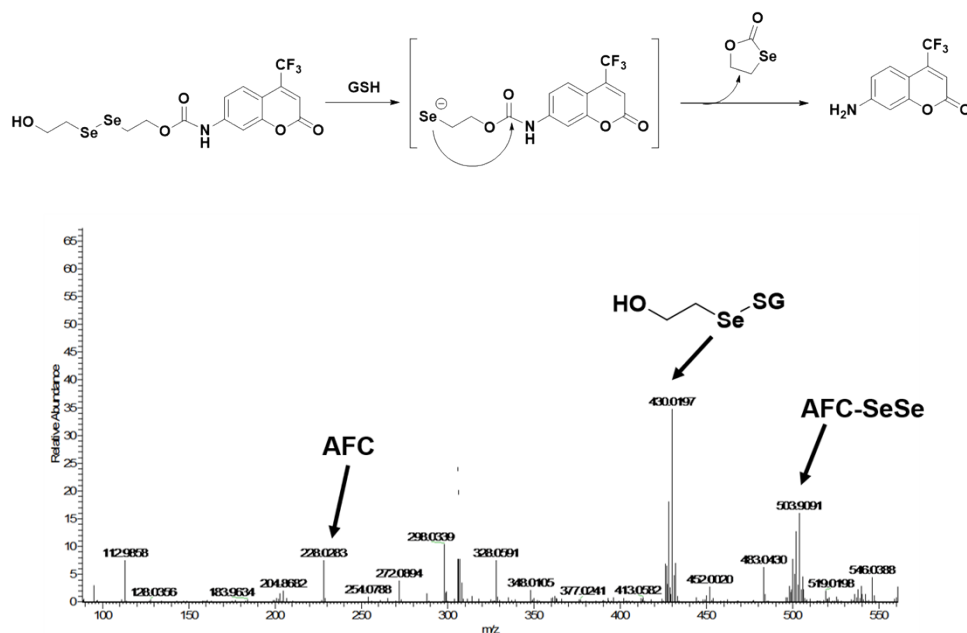


Figure S1. The mass after AFC-SeSe reacted with GSH.

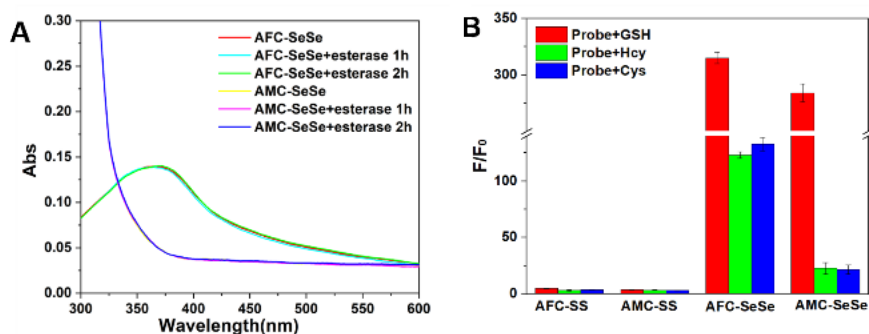


Figure S2. UV-vis absorption and fluorescence response of AFC-SeSe and AMC-SeSe. (A) UV-vis absorption spectra of AFC-SeSe and AMC-SeSe incubated with esterase (0.15 U/mL) in PBS buffer (10 mM, pH 7.4) at 37°C. (B) Fluorescence response of AFC-SeSe and AFC-SS ($\lambda_{ex}/\lambda_{em} = 368/490$ nm), and AMC-SeSe and AMC-SS ($\lambda_{ex}/\lambda_{em} = 330/440$ nm) towards GSH (1 mM), Hcy (100 μ M), and Cys (100 μ M) for 20 min in PBS buffer (10 mM, pH 7.4) at 37°C.

Table S1 Fluorescence response of probe with thiols for 20 min

Probe	GSH(F/F ₀)	Hcy(F/F ₀)	Cys(F/F ₀)
AFC-SS	4.67	3.03	3.73
AMC-SS	3.49	3.34	3.07
★AFC-SeSe	315.41	123.1	132.7
★AMC-SeSe	284.12	22.71	21.5

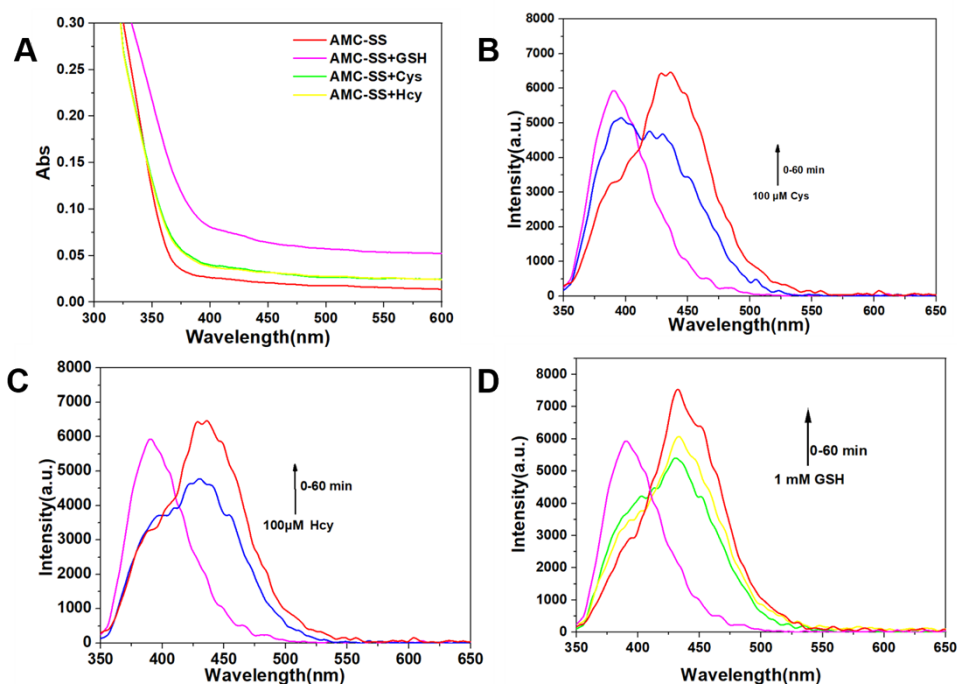


Figure S3. (A) UV-vis absorption of **AMC-SS** incubated with GSH (1 mM), Hcy (100 μ M), Cys (100 μ M) for 60 min in PBS buffer (10 mM, pH 7.4) at 37 $^{\circ}$ C.; (B-D) Fluorescence response of **AMC-SS** towards GSH (1 mM), Hcy (100 μ M), Cys (100 μ M) for 60 min in PBS buffer (10 mM, pH 7.4) at 37 $^{\circ}$ C (λ_{ex} = 368 nm).

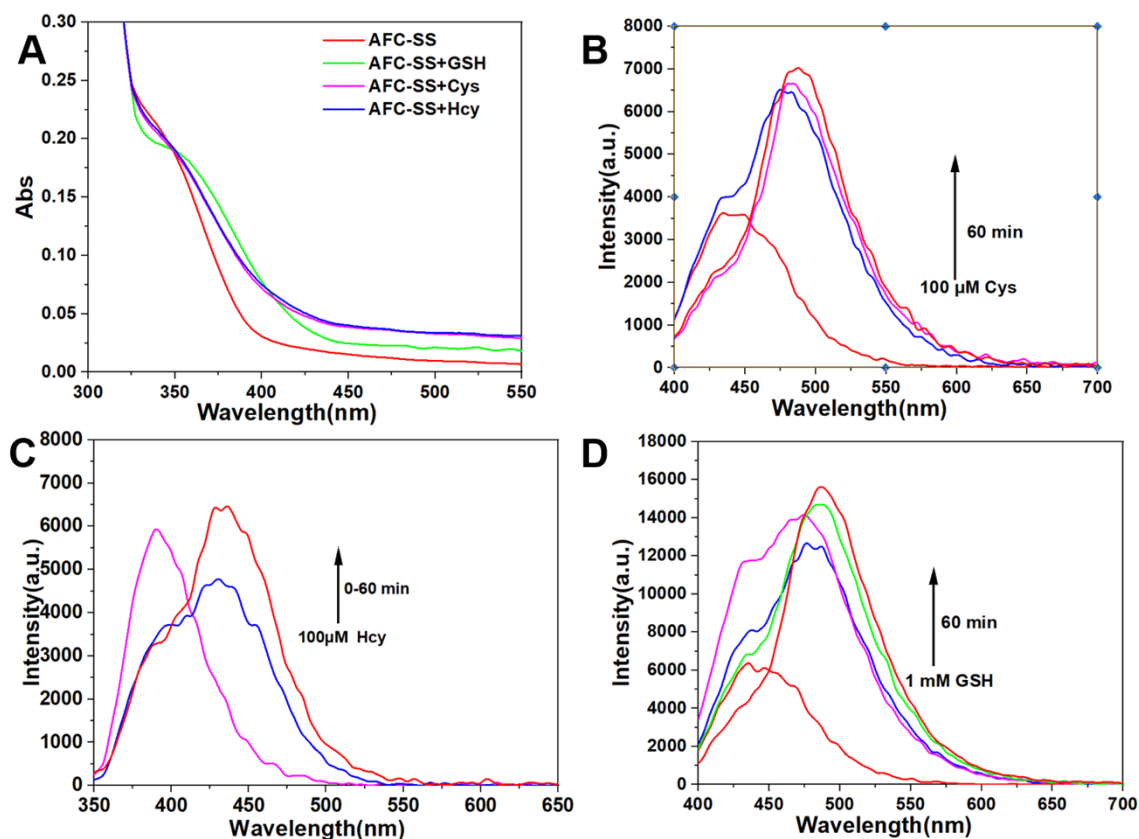


Figure S4. (A) UV-vis absorption of **AFC-SS** incubated with GSH (1 mM), Hcy (100 μ M), Cys (100 μ M) for 60 min in PBS buffer (10 mM, pH 7.4) at 37 $^{\circ}$ C.; (B-D) Fluorescence response of

before and after treated GSH (1 mM) under the different pH (3.0-11.0) at 37 °C;; (D) Fluorescence intensity of **AMC-SeSe** ($\lambda_{em}=490$ nm) after treated with various amino acids and ionic compounds (100 μ M) for 20 min in PBS buffer (10 mM, pH 7.4) at 37 °C, including Ala, Asp, Gln, Asn, Ile, Gly, Met, Val, Trp, Lys, Thr, His, Pro, Arg, Glu, Phe, Leu, NaCl, KO₂, NaClO, H₂O₂, KCl, Hcy(100 μ M), Cys(100 μ M), GSH (1 mM).

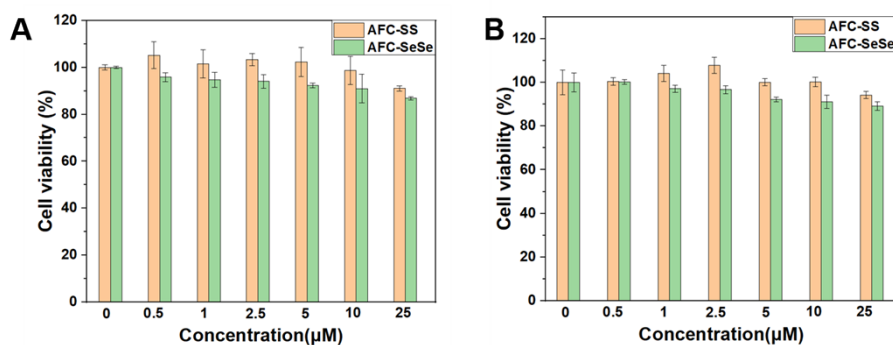


Figure S8. HeLa cells (A) and HeG2 cells (B) were treated with varying concentration of **AFC-SS** and **AFC-SeSe** for 12 h, and the cell viability was determined by CCK-8 assay.

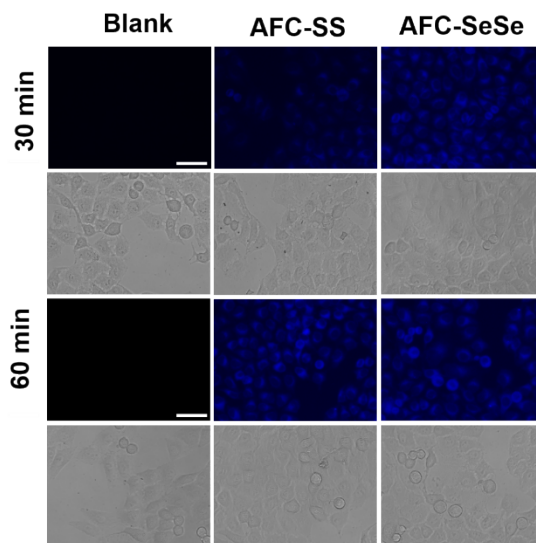


Figure S9. Response of probes to intracellular thiols. HeLa cells were treated with **AFC-SeSe** (10 μ M), **AFC-SS** (10 μ M) for 30 or 60 min. Scale bars: 20 μ m.

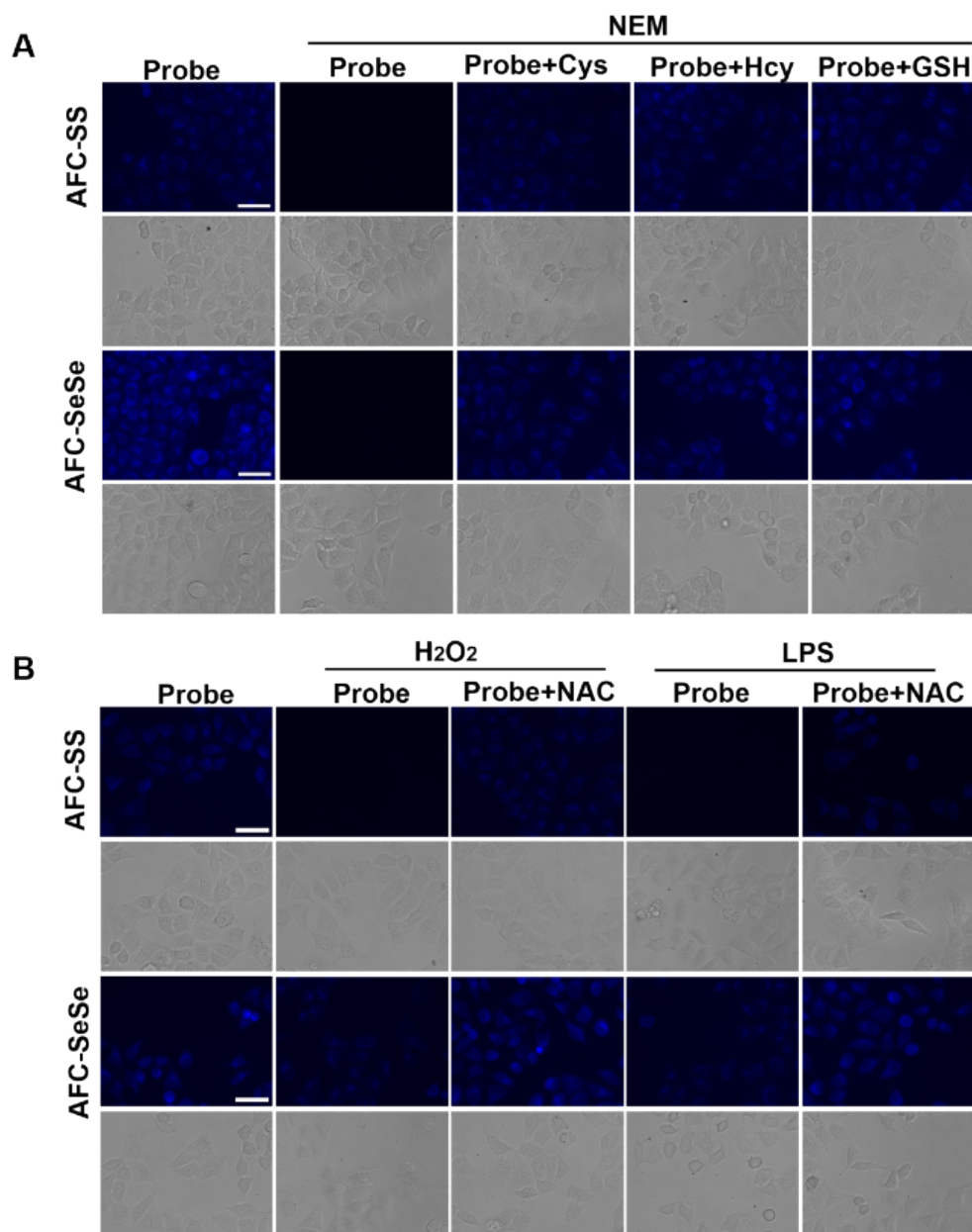


Figure S10. Response of probes to intracellular thiols under different stimuli. (A) HeLa cells were incubated with or without NEM (100 μ M) and then treated with GSH (1 mM), Hcy (100 μ M), or Cys (100 μ M) for 20 min, followed by treatment with AFC-SeSe (10 μ M) or AFC-SS (10 μ M) for 30 min. (B) HeLa cells were treated with H₂O₂ (100 μ M) for 30 min or LPS (100 μ M) for 12 hours, followed by incubation with NAC (100 μ M) for 30 min, and then treated with AFC-SeSe (10 μ M) or AFC-SS (10 μ M) for 30 min. Scale bars: 20 μ m.

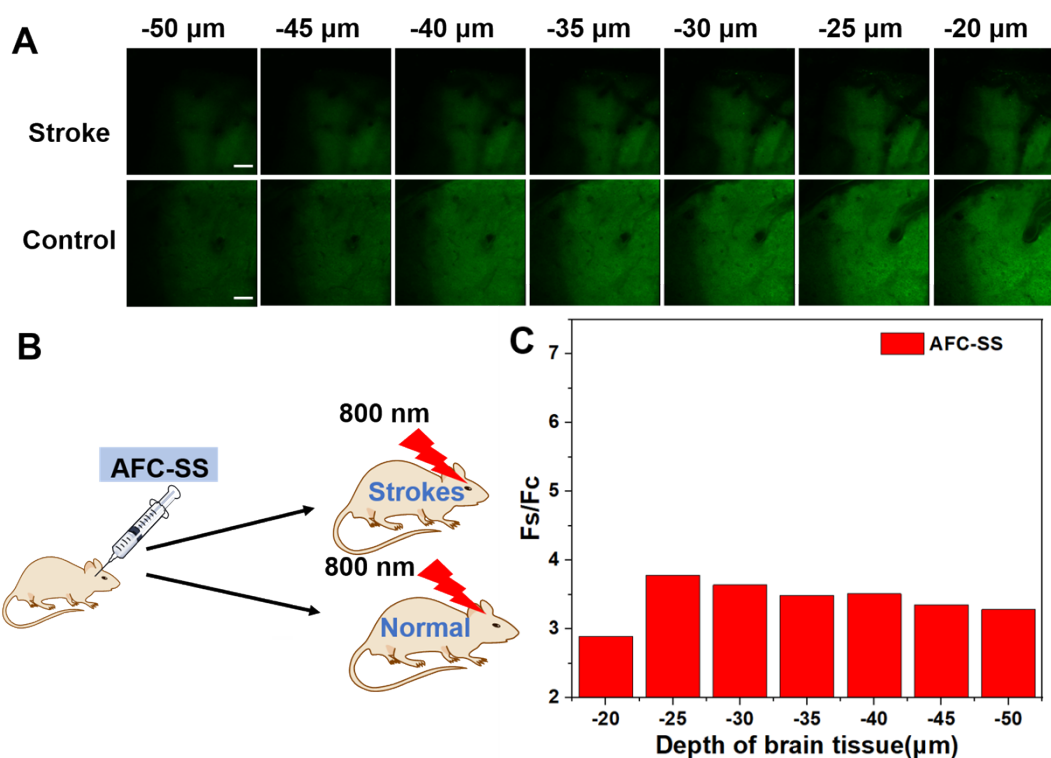


Figure S11. Two-photon fluorescence imaging of **AFC-SS**. (A) Two-photon fluorescence images of **AFC-SS** in the brains of healthy and stroke mice, respectively; (B) The diagram of mouse brain tissue imaging; (C) Comparison of fluorescence intensity at different depths of brain tissue. F_s represents the fluorescence intensity of the tissue of stroke mice; F_c represents the fluorescence intensity of the tissue of healthy mice; $\lambda_{\text{ex}}=800\text{ nm}$, Scale bars: $100\text{ }\mu\text{m}$.

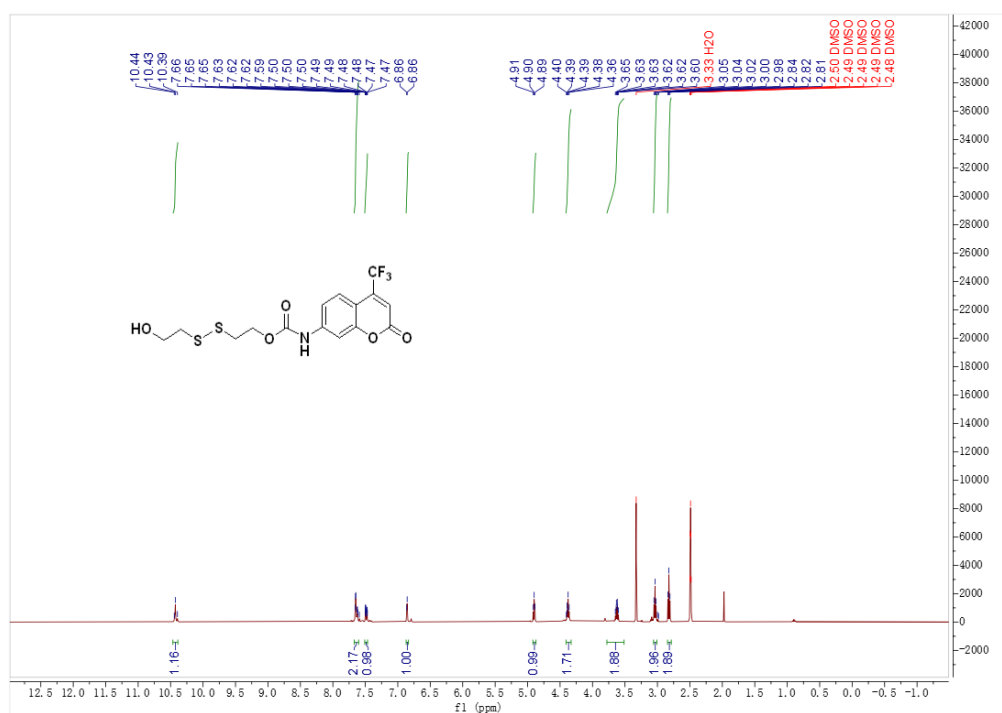


Figure S12. ^1H NMR Spectra of **AFC-SS** in $\text{DMSO}-d_6$ (400 MHz).

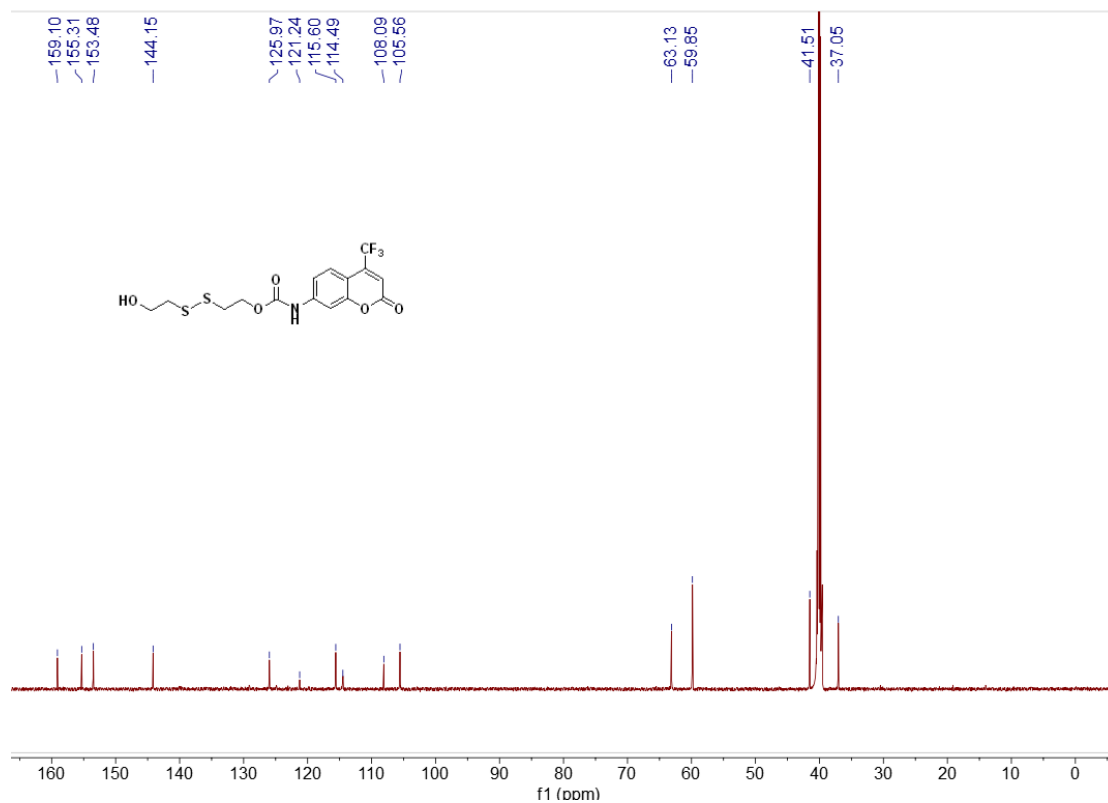


Figure S13. ¹³C NMR Spectra of AFC-SS in DMSO-*d*₆ (151 MHz).

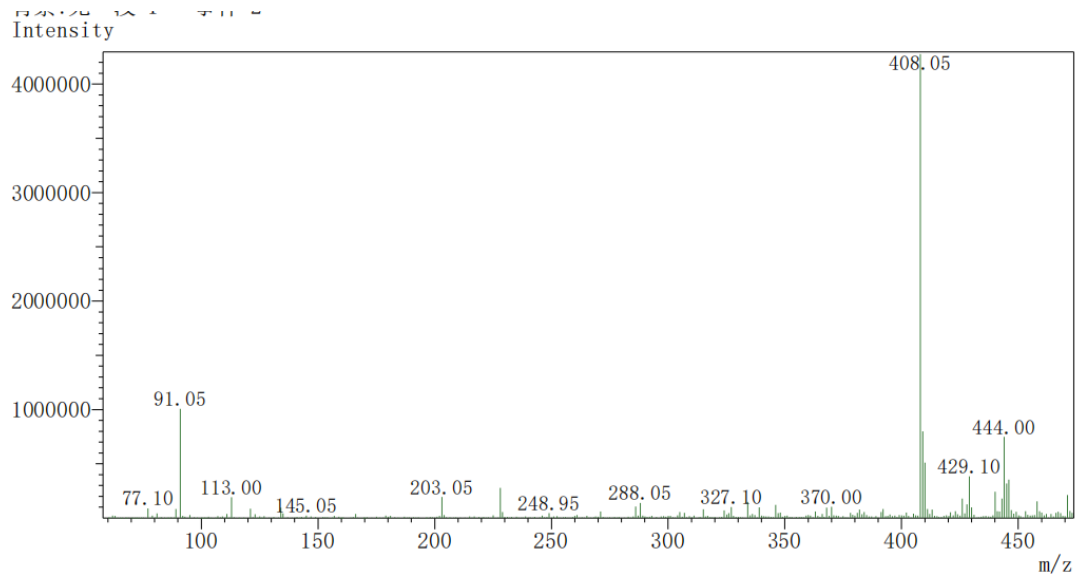


Figure S14. MS Spectra of AFC-SS.

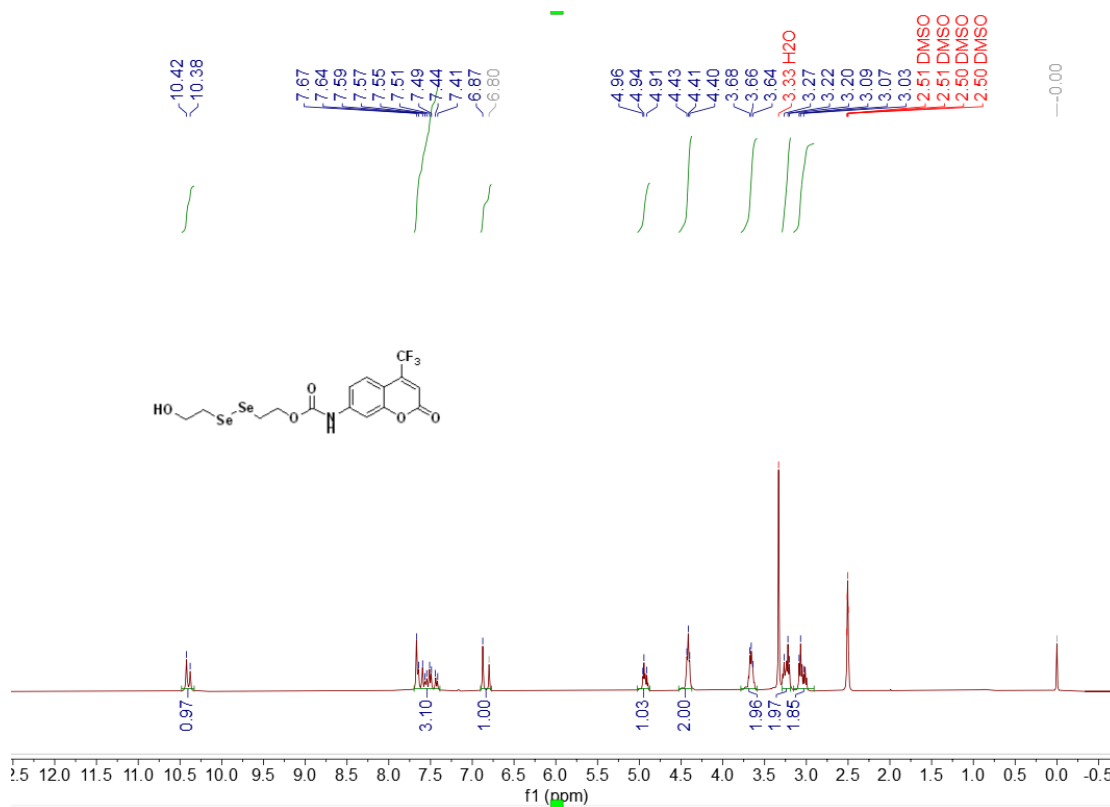


Figure S15. ¹H NMR Spectra of AFC-SeSe in DMSO-*d*₆ (400 MHz).

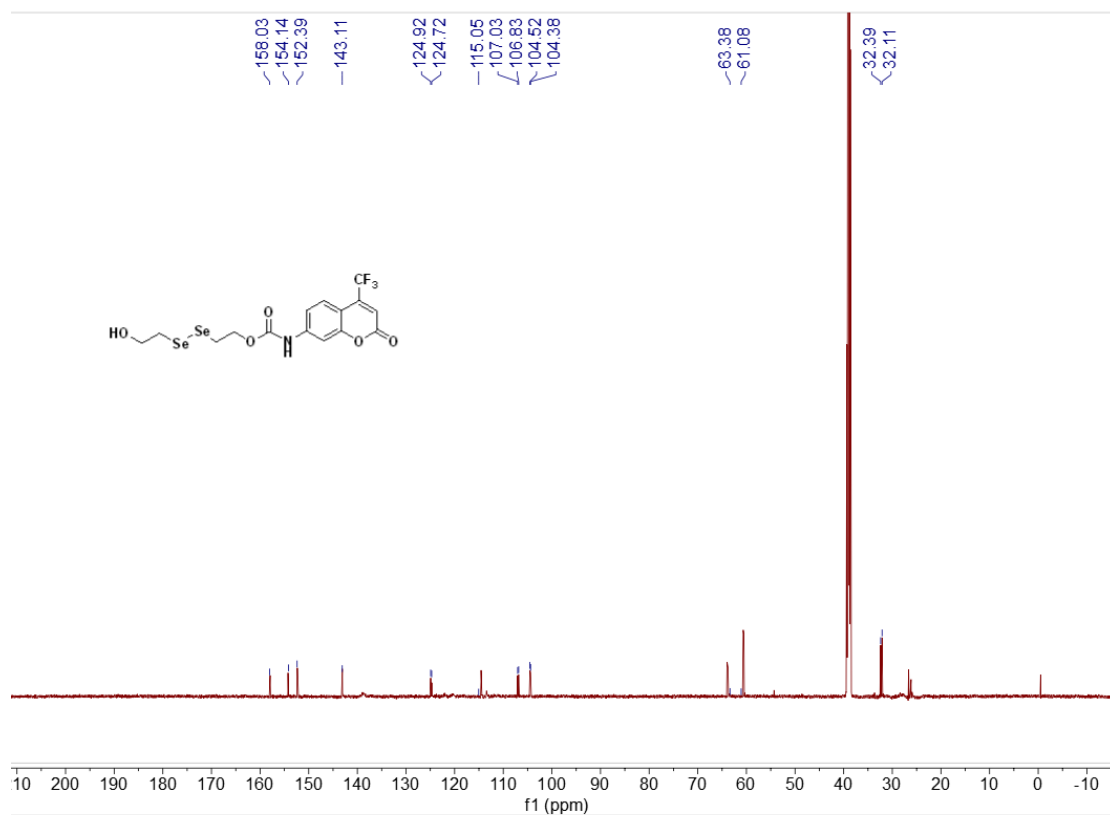


Figure S16. ¹³C NMR Spectra of AFC-SeSe in DMSO-*d*₆ (151 MHz).

mt 504 #203 RT: 0.45 AV: 1 NL: 6.33E5
T: FIMS-pESI Fullms [80.0000-1000.0000]

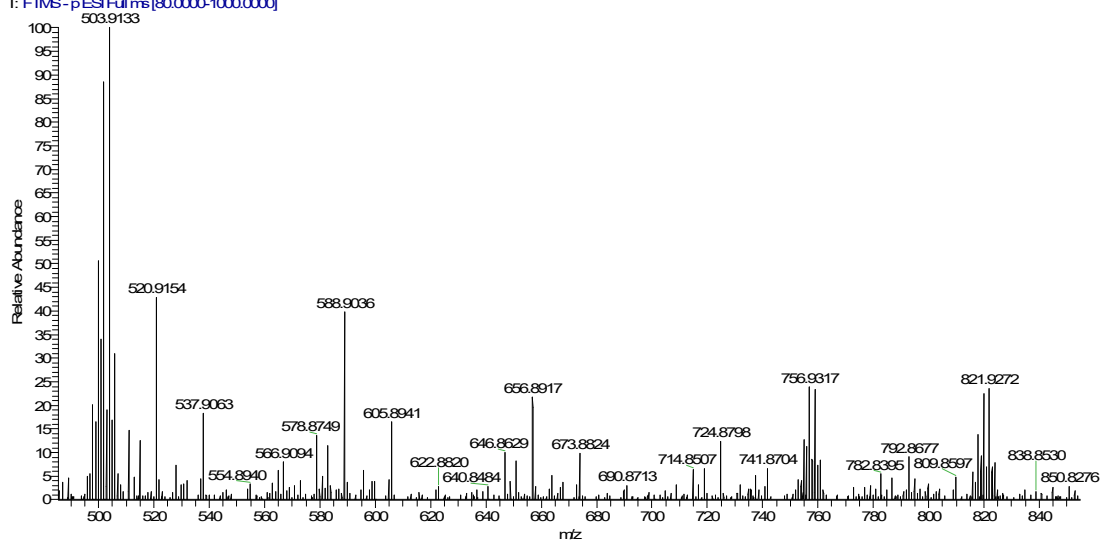


Figure S17. HR-MS Spectra of AFC-SeSe.

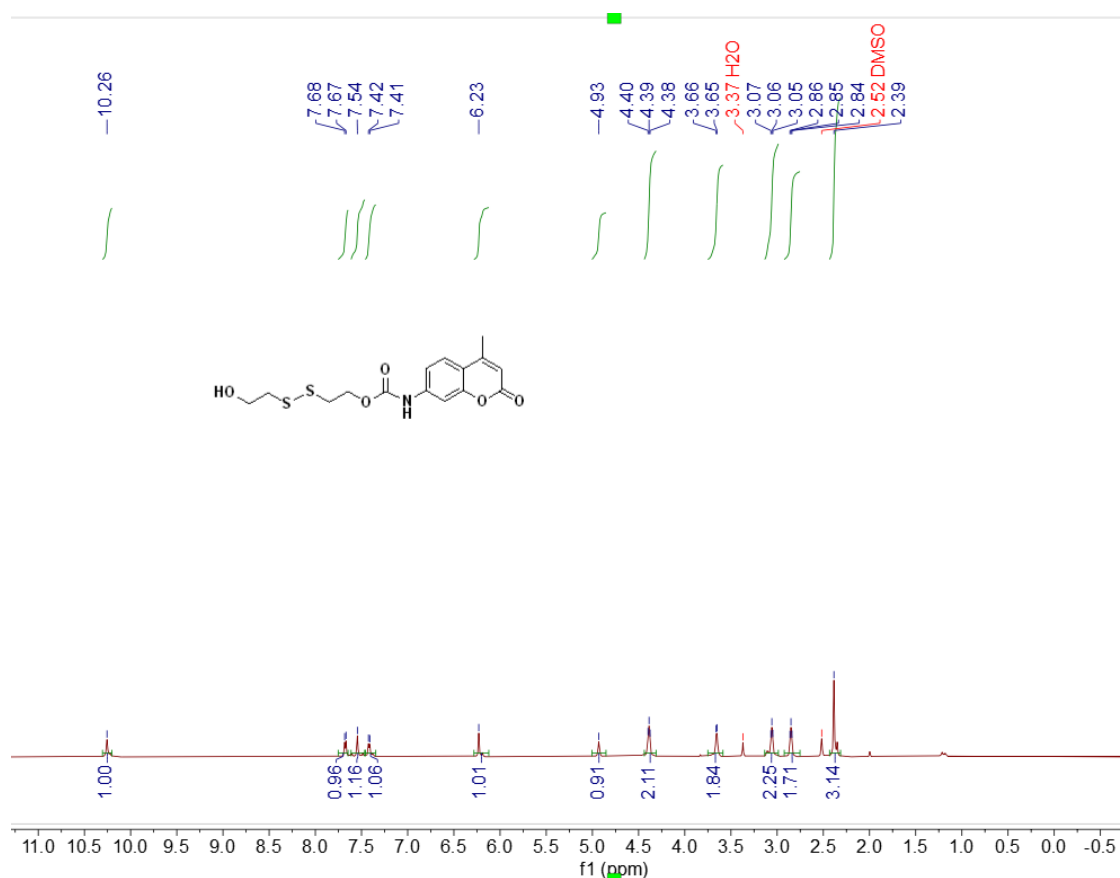
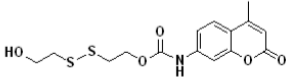


Figure S18. ¹H NMR Spectra of AMC-SS in DMSO-*d*₆ (400 MHz).



Chemical structure: CC1=C(C(=O)OCC[Se]CCOC(=O)N1)c2cc3c(cc2O)c(=O)oc4ccccc34

¹H NMR spectrum (ppm):

- 10.24 (s, 1H)
- 7.70 (d, 2H)
- 7.67 (d, 2H)
- 7.54 (d, 2H)
- 6.23 (s, 1H)
- 5.03 (s, 1H)
- 5.01 (s, 1H)
- 5.00 (s, 1H)
- 4.97 (s, 1H)
- 4.95 (s, 1H)
- 4.94 (s, 1H)
- 4.93 (s, 1H)
- 4.92 (s, 1H)
- 4.90 (s, 1H)
- 4.38 (s, 1H)
- 3.81 (s, 1H)
- 3.75 (s, 1H)
- 3.73 (s, 1H)
- 3.71 (s, 1H)
- 3.70 (s, 1H)
- 3.68 (s, 1H)
- 3.67 (s, 1H)
- 3.66 (s, 1H)
- 3.65 (s, 1H)
- 3.64 (s, 1H)
- 3.62 (s, 1H)
- 3.60 (s, 1H)
- 3.22 (s, 1H)
- 3.20 (s, 1H)
- 3.18 (s, 1H)
- 3.07 (s, 1H)
- 3.05 (s, 1H)
- 3.03 (s, 1H)
- 3.01 (s, 1H)
- 3.00 (s, 1H)
- 2.98 (s, 1H)
- 2.97 (s, 1H)

Integration values (from left to right): 1.03, 1.15, 1.02, 1.07, 0.98, 1.19, 2.17, 2.29, 4.10, 2.72.

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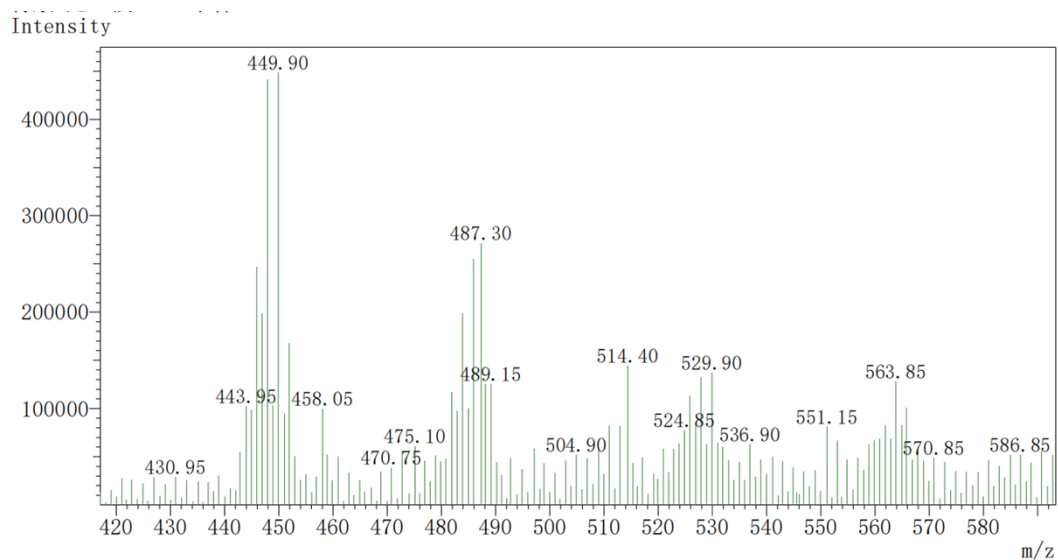


Figure S21. MS Spectra of AMC-SeSe.

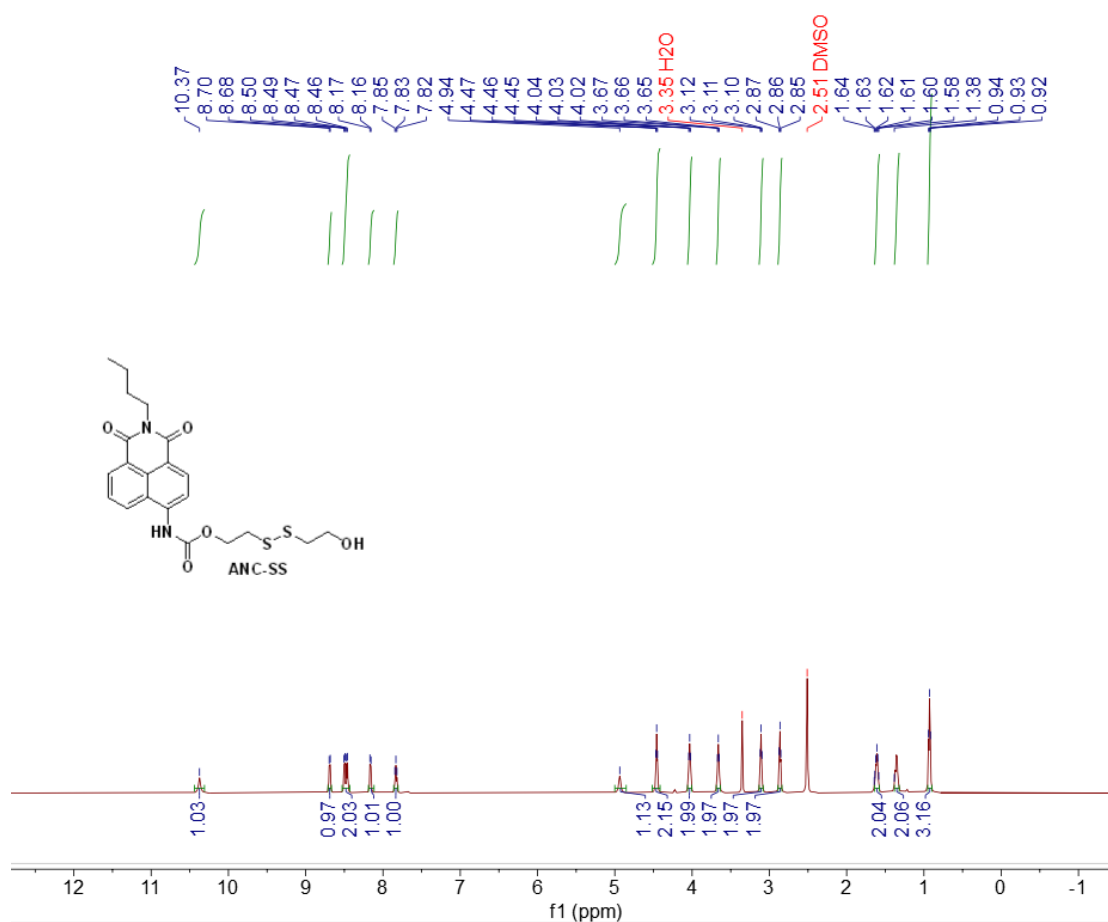


Figure S22. ¹H NMR Spectra of ANC-SS in DMSO-*d*₆ (400 MHz).

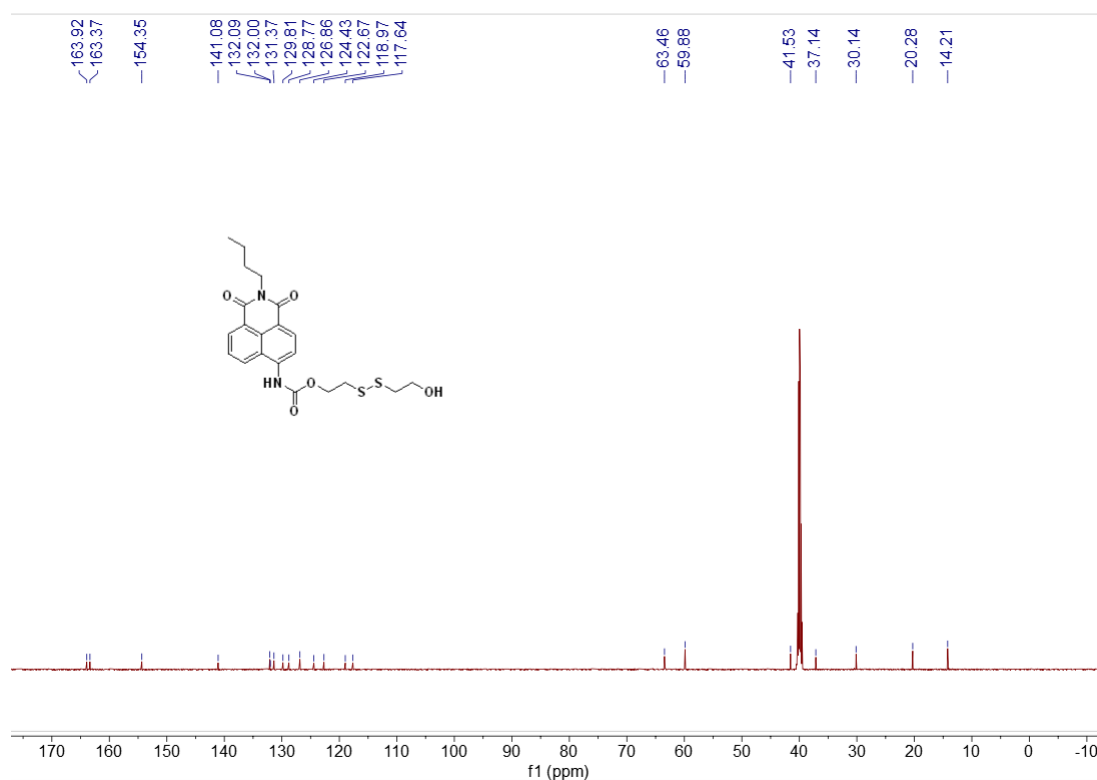


Figure S23. ¹³C NMR Spectra of ANC-SS in DMSO-*d*₆ (151 MHz).

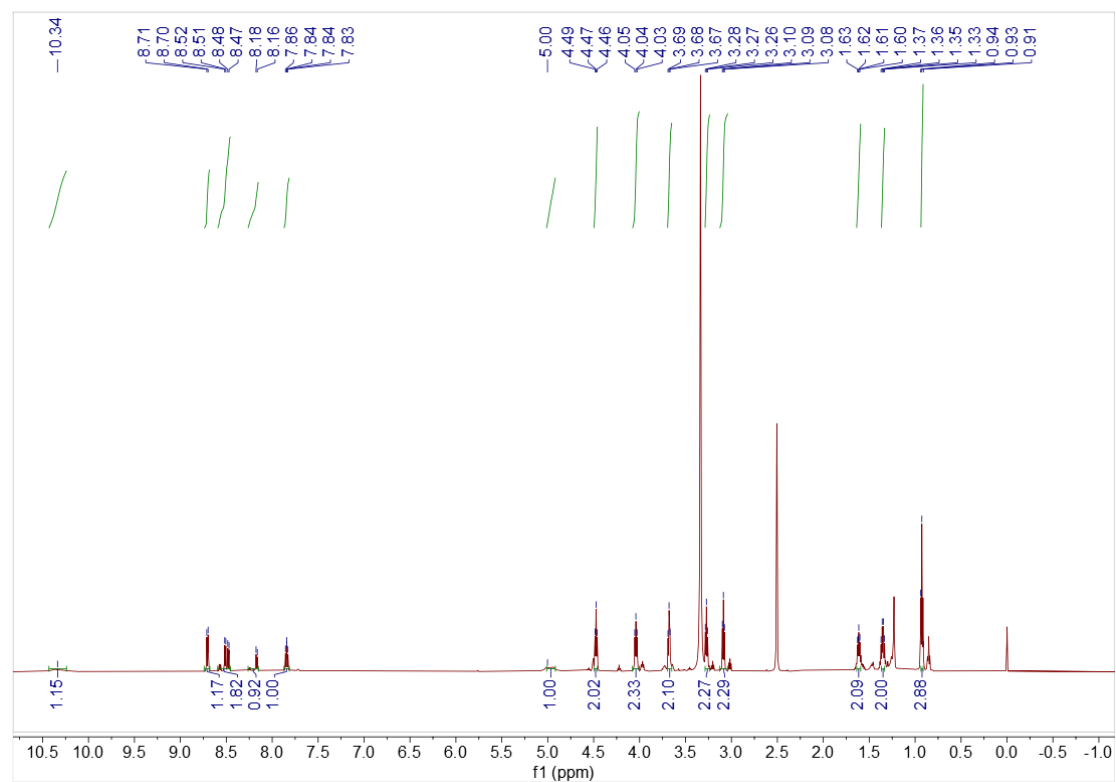


Figure S24. ¹H NMR Spectra of ANC-SeSe in DMSO-*d*₆ (400 MHz).

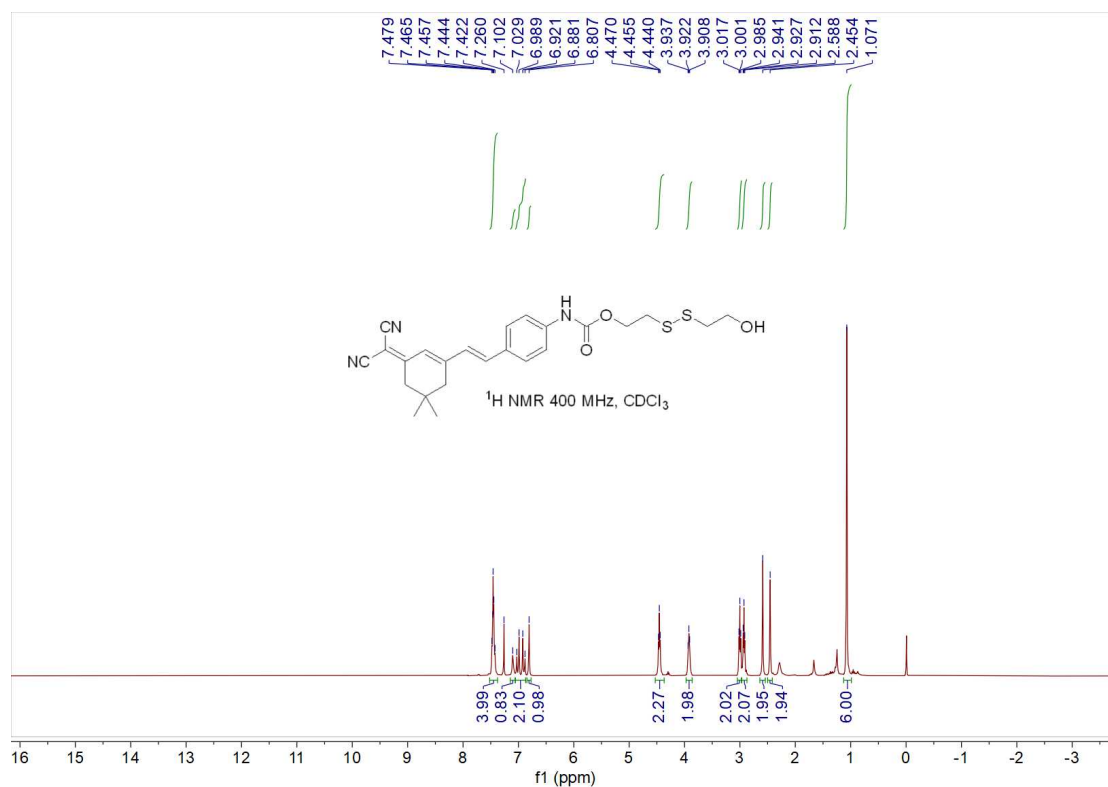


Figure S27. ¹H NMR Spectra of **ARC-SS** in CDCl₃-d₄ (400 MHz).

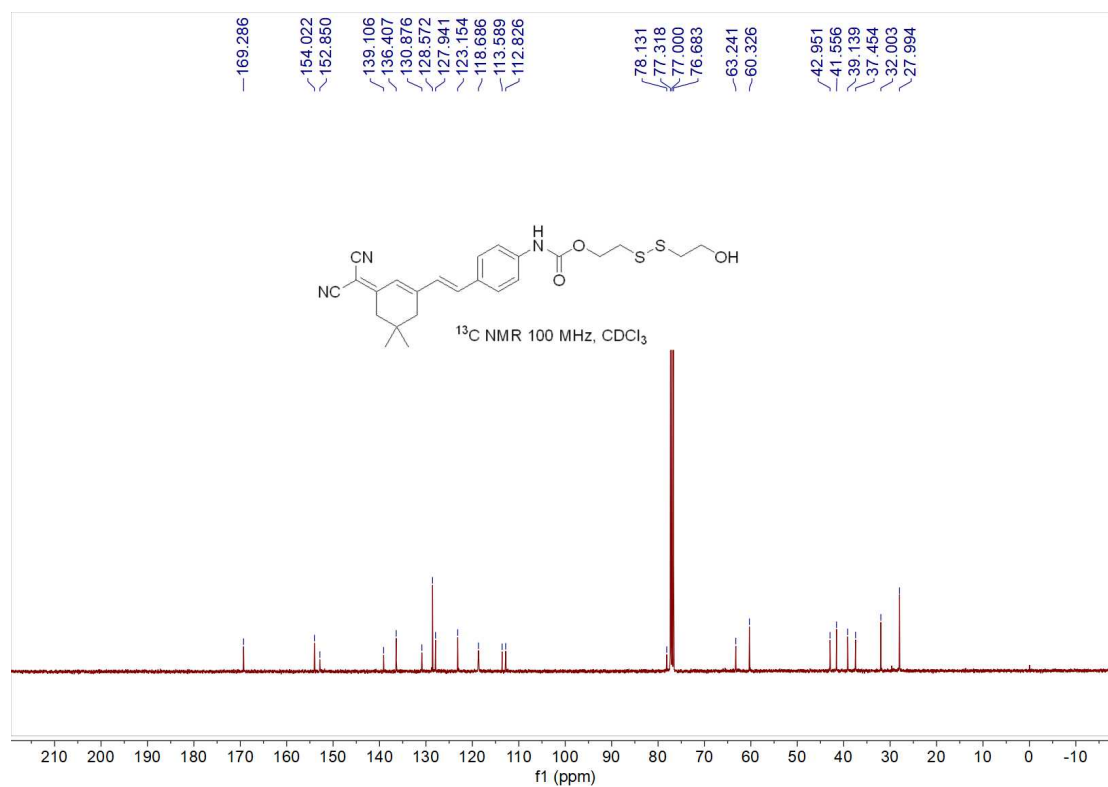


Figure S28. ¹³C NMR Spectra of **ARC-SS** in CDCl₃-d₄ (151 MHz).

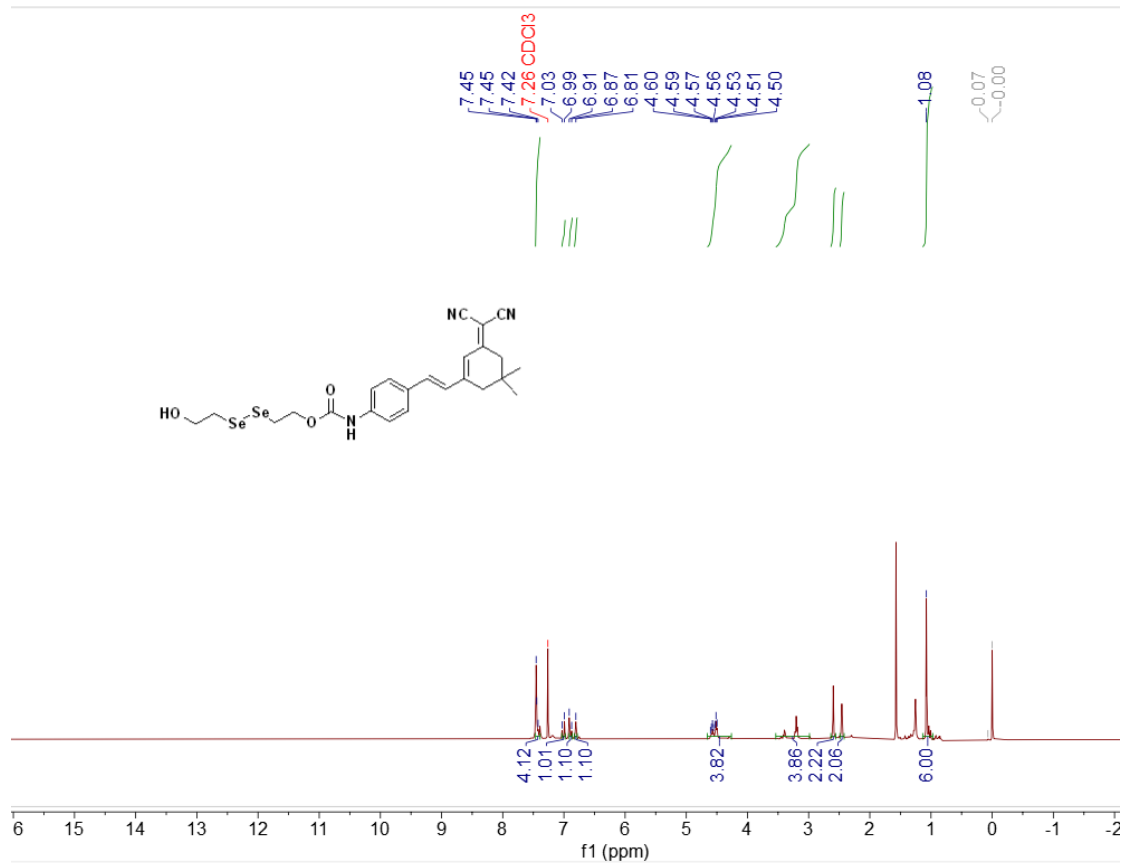


Figure S29. ¹H NMR Spectra of **ARC-SeSe** in CDCl₃-d₄ (400 MHz).

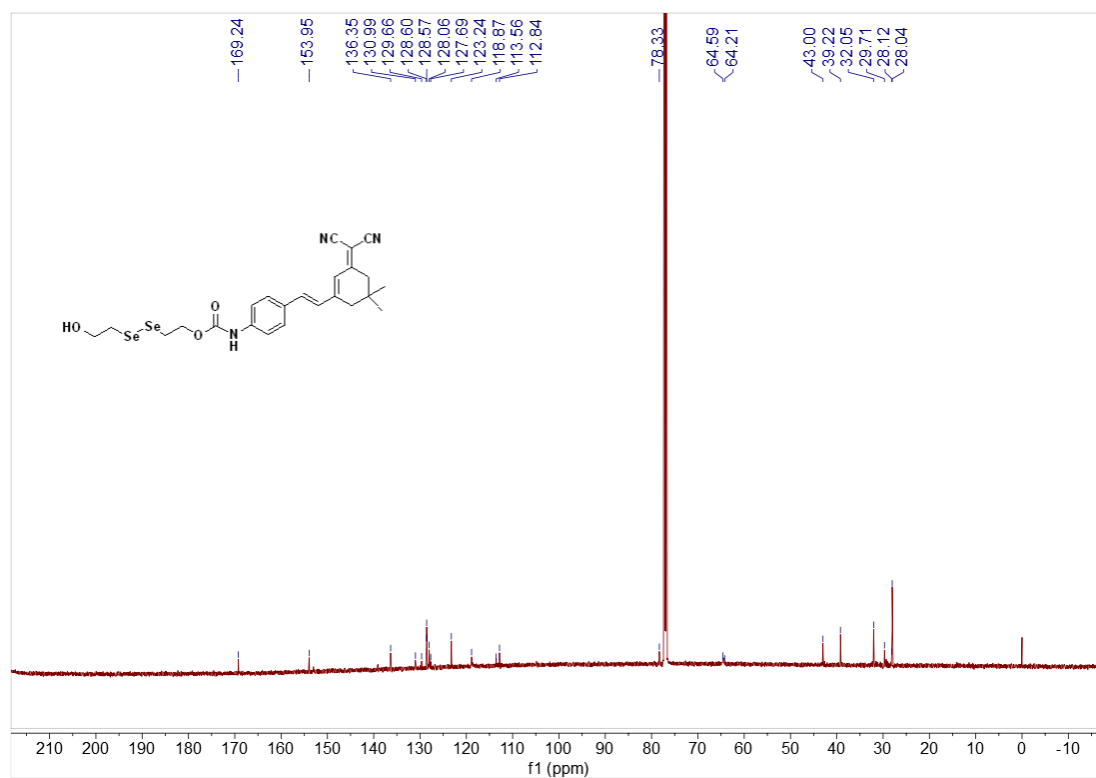


Figure S30. ¹³C NMR Spectra of **ARC-SeSe** in CDCl₃-d₄ (151 MHz).

