

Supporting Information

for

Ruthenium(II) N–Heterocyclic Carbene Polymer Based Sensors for Detection  
of Predatory Drugs like Ketamine and Scopolamine

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**Table SI1.** Crystal data and structure refinement parameters for salt ruthenium NHC organometallic polymer **6**.

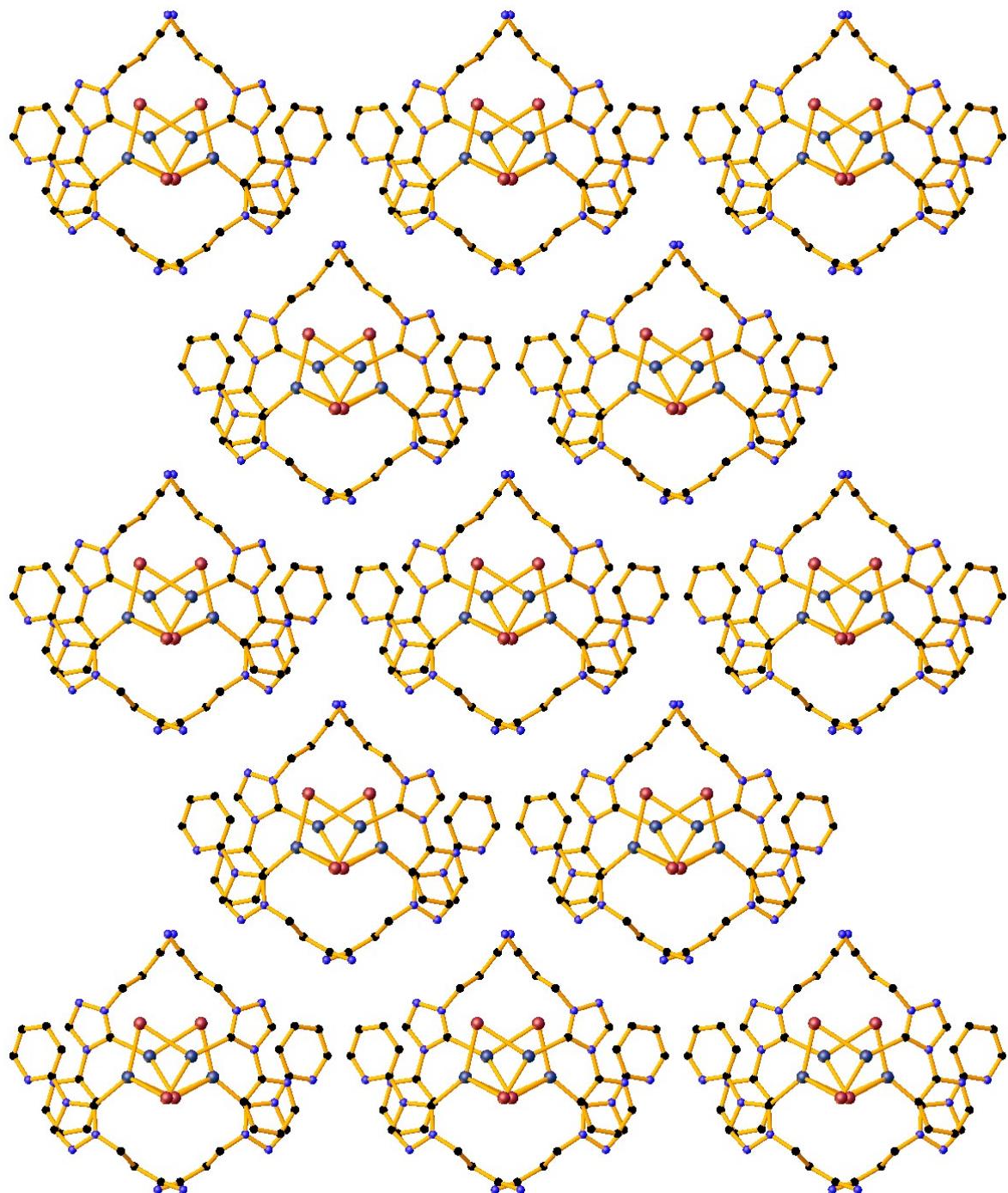
Empirical formula	C <sub>20</sub> H <sub>18</sub> Br <sub>2</sub> N <sub>10</sub> Ru <sub>2</sub>
Formula weight	760.4
Crystal system	monoclinic
Space group	Cc
a/Å	21.2545(14)
b/Å	15.6065(9)
c/Å	7.4968(4)
α/°	90
β/°	90.204(5)
γ/°	90
Volume/Å <sup>3</sup>	2486.7(3)
Z	4
ρ <sub>calc</sub> g/cm <sup>3</sup>	2.031
μ/mm <sup>-1</sup>	4.458
F(000)	1464
Crystal size/mm <sup>3</sup>	0.27 × 0.14 × 0.03
Radiation	Mo Kα ( $\lambda = 0.71073$ )
2Θ range for data collection/°	7.538 to 58.766
Index ranges	-26 ≤ h ≤ 29, -18 ≤ k ≤ 21, -9 ≤ l ≤ 10
Reflections collected	10829

**Table SI2.** Important bond distances and angles of ruthenium NHC organometallic polymer **6**.

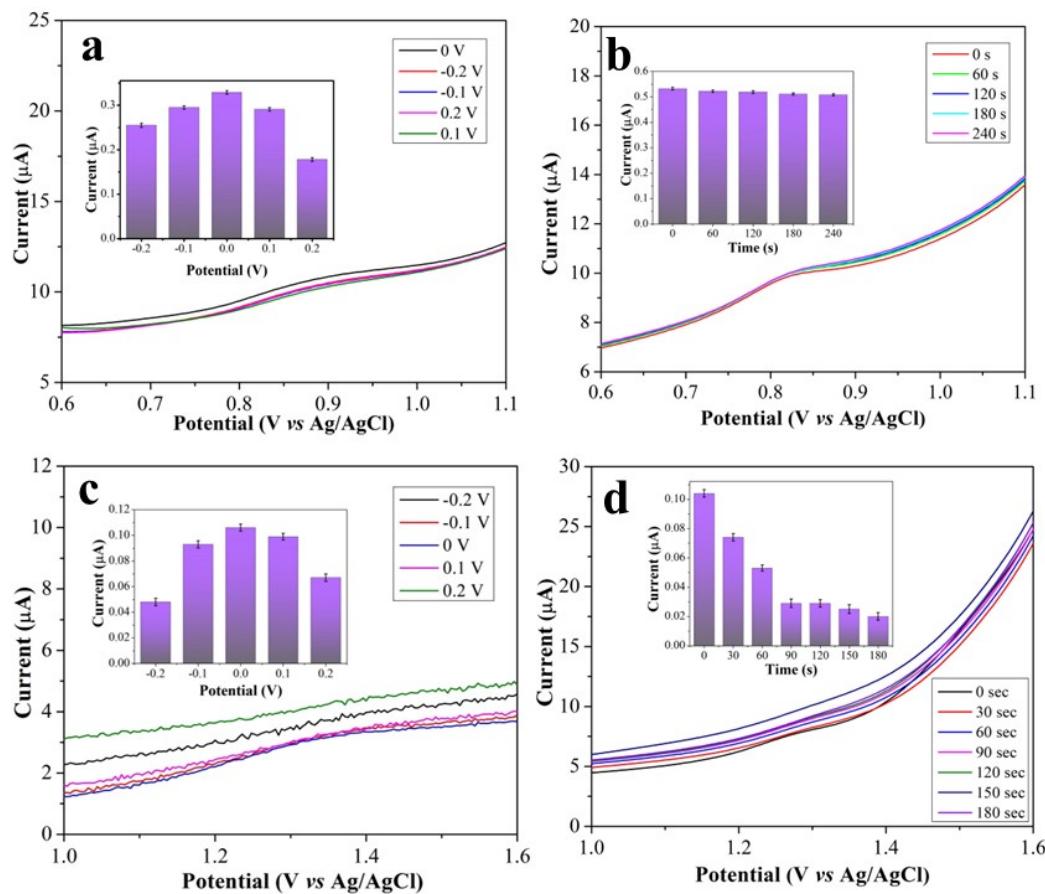
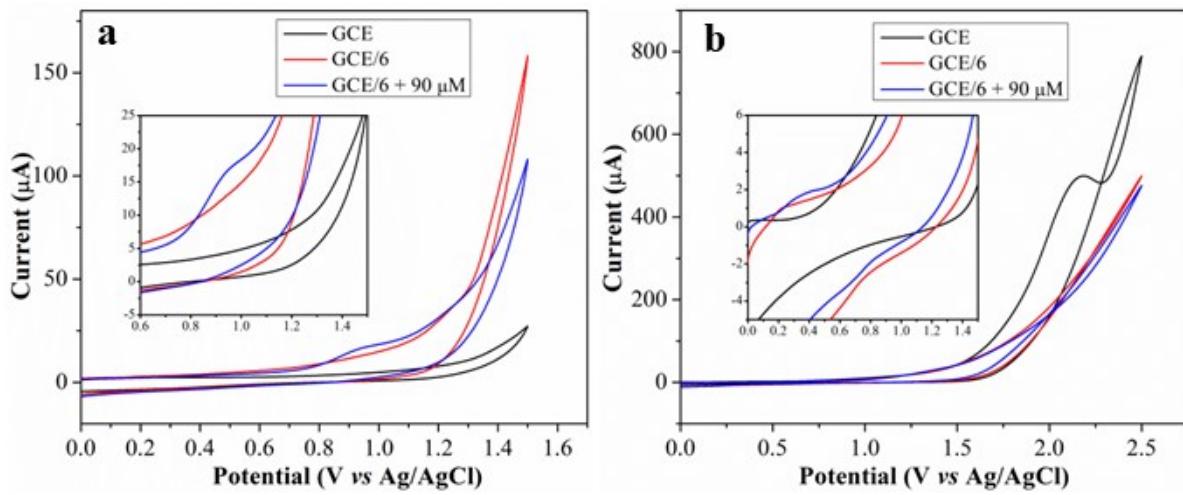
<b>Module</b>	<b>Angle/°</b>	<b>Module</b>	<b>Length /Å°</b>
Br1-Ru1-Br11	81.42(4)	Ru1-Br11	2.908(2)
Br2-Ru1-Br1	95.27(7)	Ru1-Br11	2.866(2)
Br2-Ru1-Br11	104.24(7)	Ru1-Br2	2.569(2)
C1-Ru1-Br11	102.2(4)	Ru1-C1	2.107(14)
C1-Ru1-Br1	114.4(4)	Ru2-Br1	2.563(3)
C1-Ru1-Br2	142.7(4)	Ru2-Br2	2.854(3)
Br1-Ru2-Br2	95.72(7)	Ru2-C11	2.149(17)
C11-Ru2-Br1	148.1(5)	N1-N2	1.359(18)
C11-Ru2-Br2	116.1(5)	N1-C1	1.343(19)
Ru1-Br1-Ru12	139.77(10)	N1-C8	1.47(2)
Ru2-Br1-Ru12	72.97(7)	N2-C2	1.29(2)
Ru2-Br1-Ru1	71.42(7)	N3-C1	1.385(19)
Ru1-Br2-Ru2	71.54(6)	N3-C2	1.39(2)
N2-N1-C8	117.8(13)	N3-C3	1.43(2)
C1-N1-N2	117.3(13)	N4-C3	1.32(2)
C1-N1-C8	125.0(13)	N4-C7	1.34(3)
C2-N2-N1	103.3(13)	N5-C10	1.12(3)
C1-N3-C2	110.1(13)	N6-N7	1.36(2)
C1-N3-C3	126.5(12)	N6-C11	1.26(2)
C2-N3-C3	123.5(13)	N6-C18	1.45(2)
C3-N4-C7	116.0(16)	N7-C12	1.28(2)
N7-N6-C18	119.5(14)	N8-C11	1.39(2)
C11-N6-N7	115.5(14)	N8-C12	1.39(2)
C11-N6-C18	125.0(16)	N8-C13	1.42(2)
C12-N7-N6	104.0(15)	N9-C13	1.32(2)
C11-N8-C13	130.8(15)	N9-C17	1.32(3)
C12-N8-C11	106.1(14)	N10-C20	1.15(2)
C12-N8-C13	122.9(13)	C3-C4	1.39(2)
C13-N9-C17	115.2(18)	C4-C5	1.34(3)
N1-C1-Ru1	125.2(10)	C5-C6	1.36(3)
N1-C1-N3	99.2(12)	C6-C7	1.34(3)
N3-C1-Ru1	135.4(10)	C8-C9	1.51(3)
N2-C2-N3	110.1(14)	C9-C10	1.48(3)
N4-C3-N3	112.6(14)	C13-C14	1.41(3)
N4-C3-C4	124.3(16)	C14-C15	1.35(3)
C4-C3-N3	123.1(15)	C15-C16	1.39(3)
C5-C4-C3	117.2(17)	C16-C17	1.36(3)
C4-C5-C6	119.9(19)	C18-C19	1.51(2)
C7-C6-C5	119.7(19)	C19-C20	1.49(2)

**Table SI3.** Comparative data for detection of predatory drugs with different techniques in current and past work.

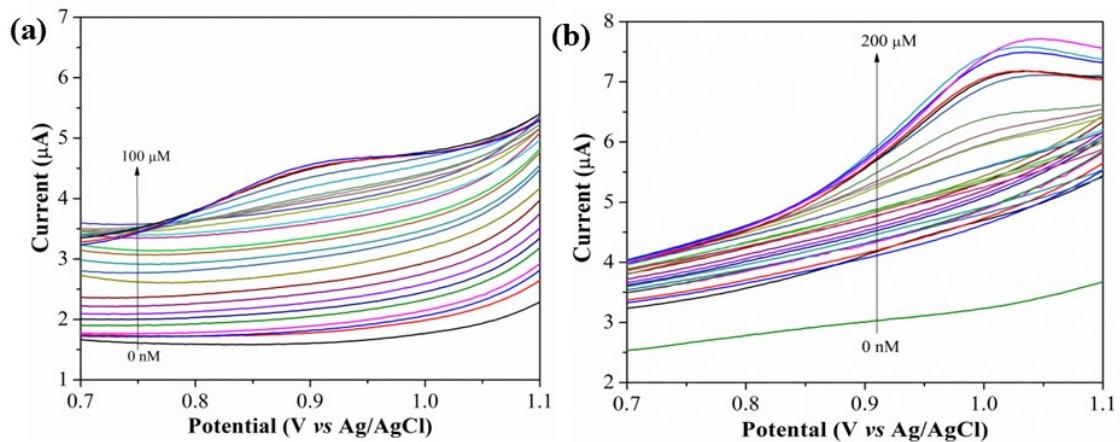
<b>Drugs</b>	<b>Technique</b>	<b>LOD</b>	<b>Matrix</b>	<b>Reference</b>
<b>Ketamine</b>	KT- MIM/MOFs@G/SPE with DPV	$4.0 \times 10^{-11}$ mol/L	PBS buffer	[1]
	PAMAM-CDs (Electrochemiluminescence)	0.067 ng/ mL	PBS buffer	[2]
	G4HTD/PtNPs/CPE with DPV	$1.0 \times 10^{-7}$ mol/L	Britton buffer	[3]
	GCMS	0.01 g/mL	Britton –Robinson buffer, urine and plasma	[4]
Ru(II)NHC+MWCNTs/ GCE with SWV	0.194 nM (0.0531 $\mu$ g/L)	Britton –Robinson buffer, alcoholic and non-alcoholic beverages	This work	
<b>Scopolamine</b>	GSH–Co <sub>3</sub> O <sub>4</sub> /GCE with DPV	0.001 $\mu$ M	Britton –Robinson buffer and urine	[5]
	SDBS/MWCNTs - CPE with DPV	0.449 ng/mL	PBS buffer, D. stramonium seeds, leaves and eye drops	[6]
	SPE with CV	3.9 $\mu$ M	Phosphate buffer, coca cola	[7]
	Activated 3D-graphene with SWV	1 $\mu$ M	Britton –Robinson buffer, vodka, white wine, whiskey, energy drink	[8]
Ru(II)NHC+MWCNTs/ GCE with SWV	3.81 nM (1.669 $\mu$ g/L)	Britton –Robinson buffer, beer, artificial serum	This work	



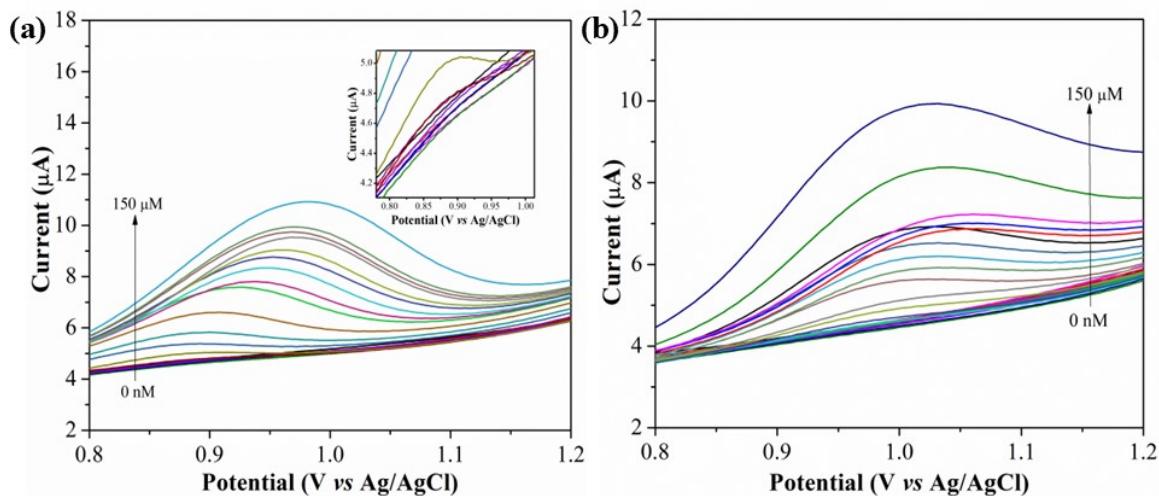
**Figure SII:** Crystal packing image of ruthenium organometallic polymer **6**. Hydrogen atoms are omitted for clarity.



**Figure SI3:** Optimisation with complex **6** modified GCE with 30  $\mu\text{M}$  ketamine (a) accumulation potential (b) accumulation time and with 50  $\mu\text{M}$  scopolamine (c) accumulation potential, (d) accumulation time.



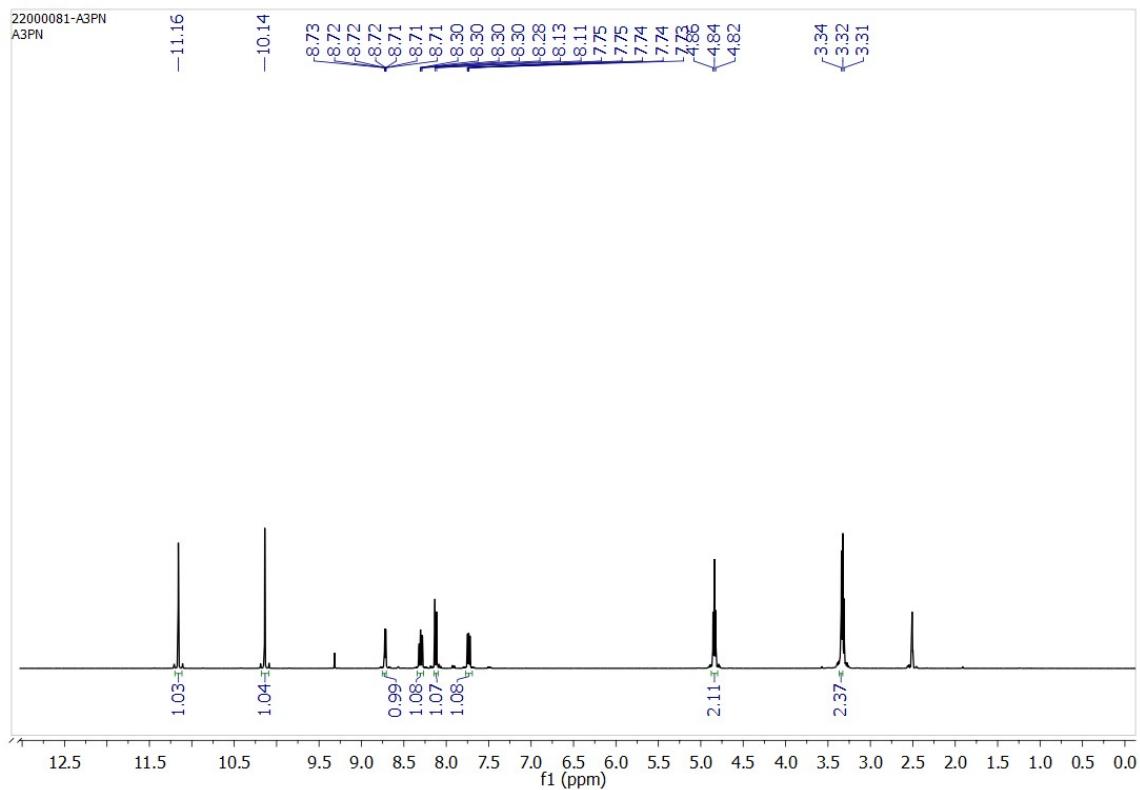
**Figure SI4:** Concentration dependent SWV responses of **6** modified GCE (a) and **7** modified GCE (b) towards ketamine quantification in Britton–Robinson buffer of pH 8 at room temperature.



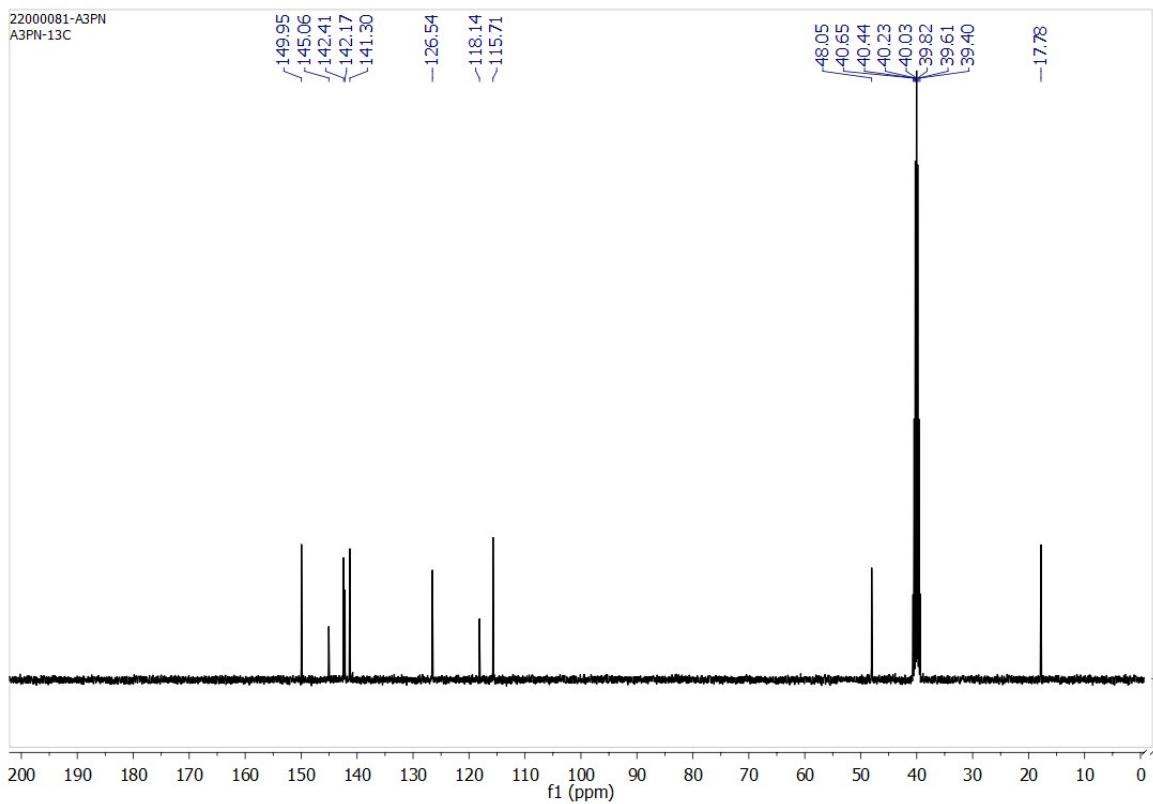
**Figure SI5:** Concentration dependent SWV responses of **6** modified GCE (a) and **7** modified GCE (b) towards scopolamine quantification in Britton–Robinson buffer of pH 7 at room temperature.

### NMR spectra

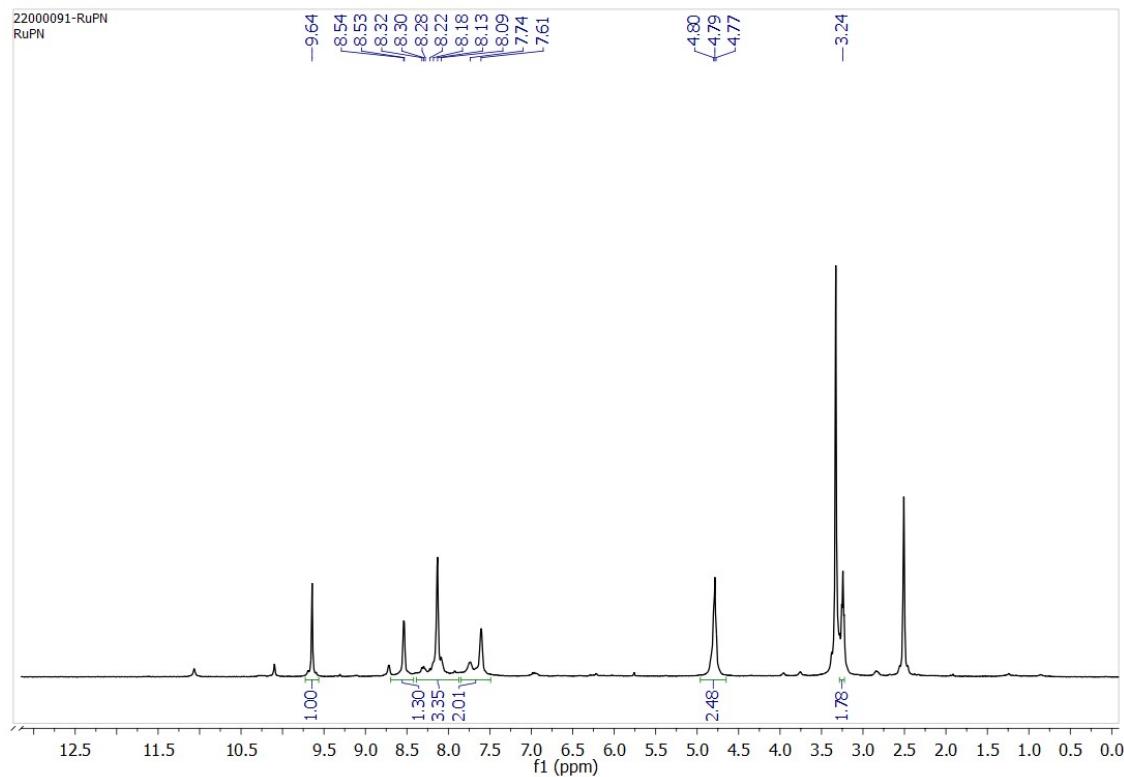
The following figures represent the NMR spectra of salts and their corresponding ruthenium NHC organometallic polymers [9].



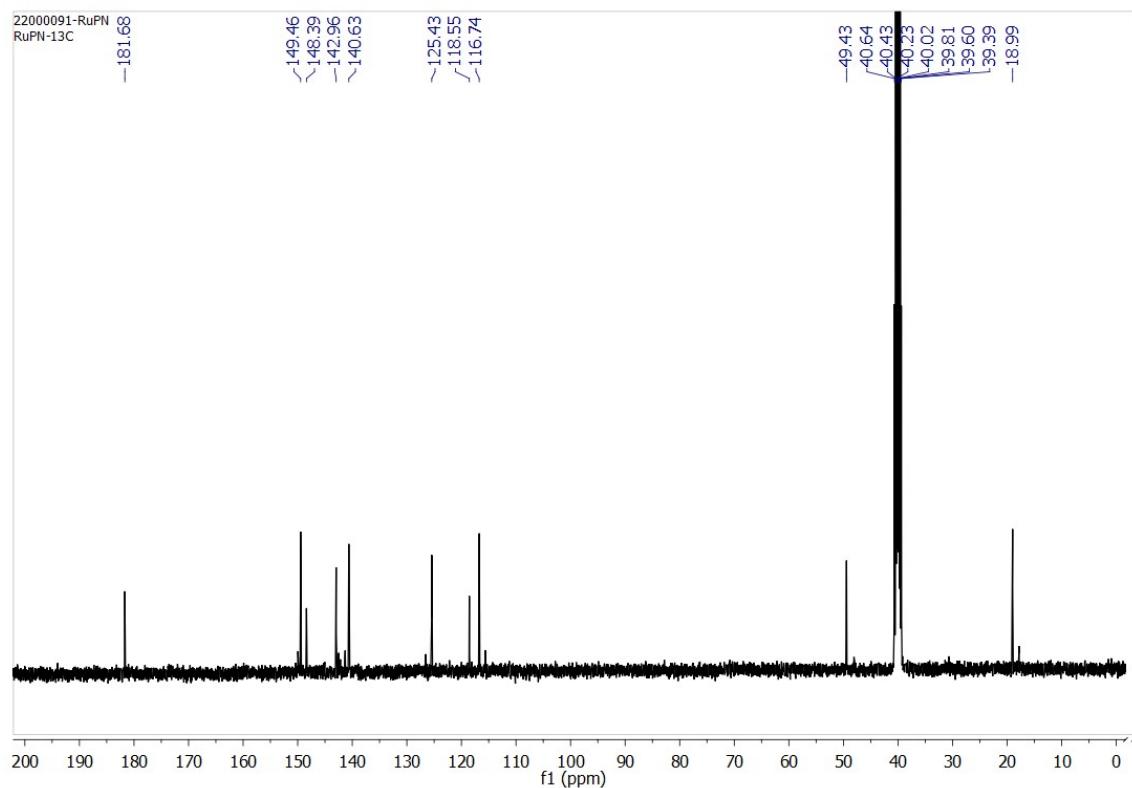
**Figure SI6:**  $^1\text{H}$ -NMR spectrum of salt **4** in  $d_6$ -DMSO at room temperature.



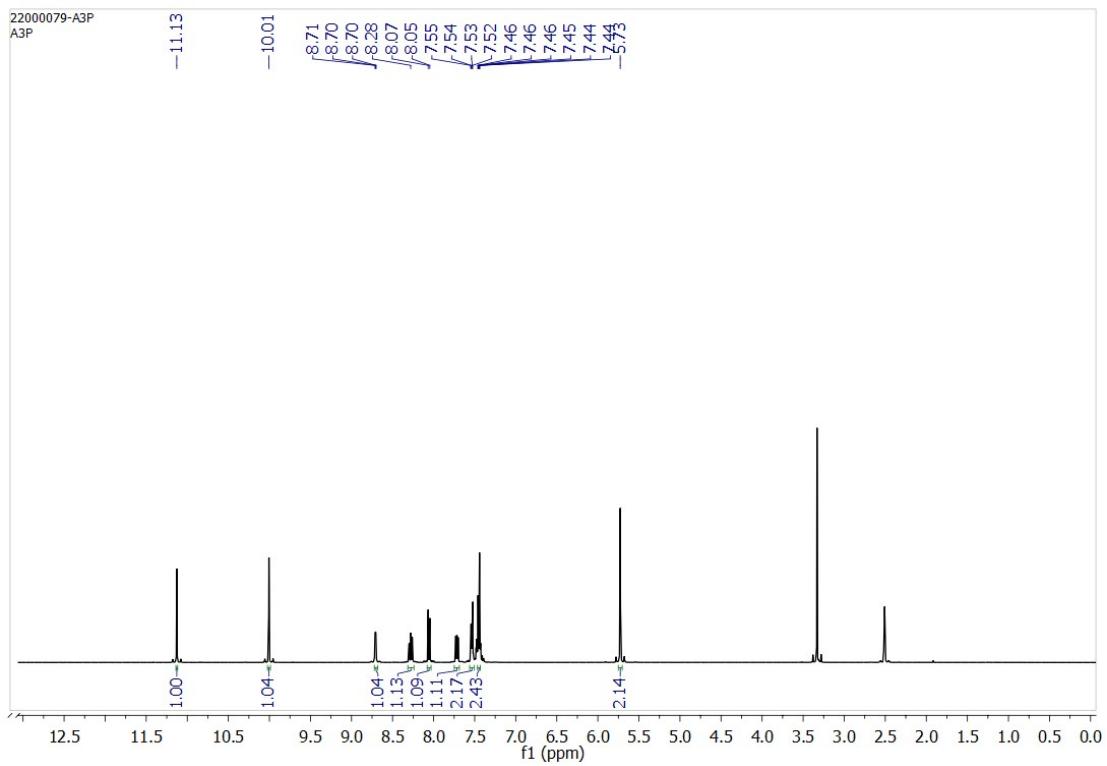
**Figure SI7:**  $^{13}\text{C}$ -NMR spectrum of **4** in  $d_6$ -DMSO at room temperature.



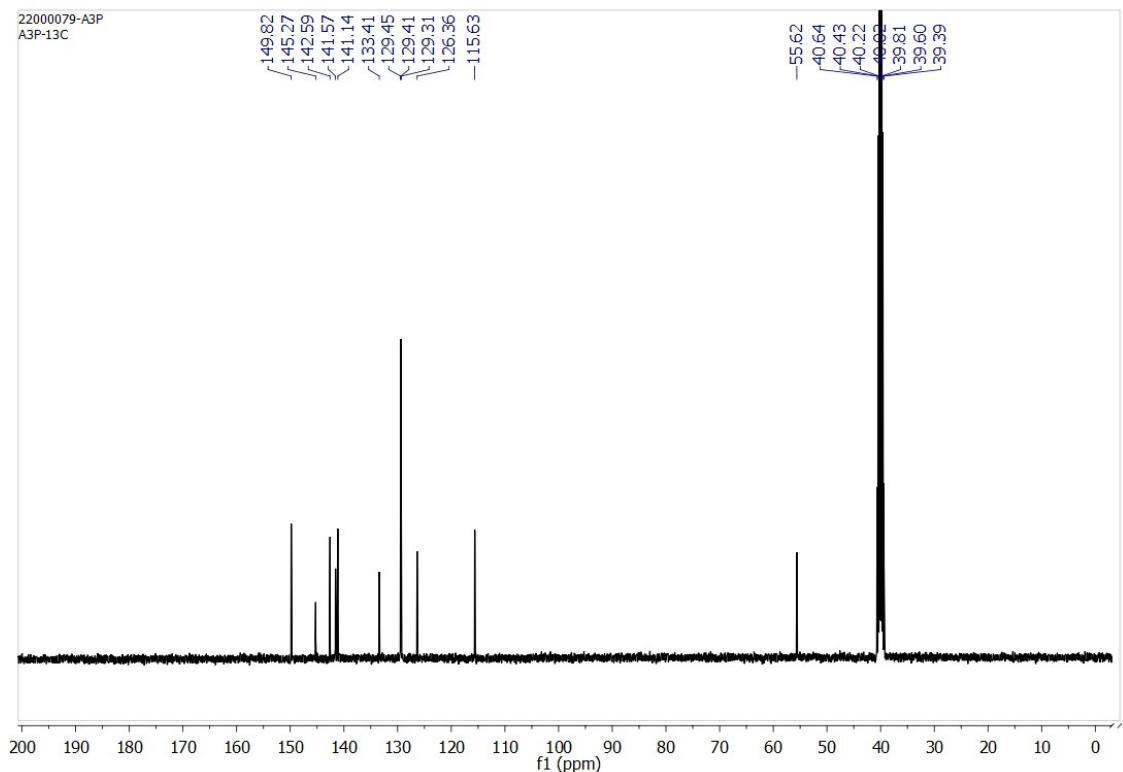
**Figure SI8:**  $^1\text{H}$ -NMR spectrum of **6** in  $d_6$ -DMSO at room temperature.



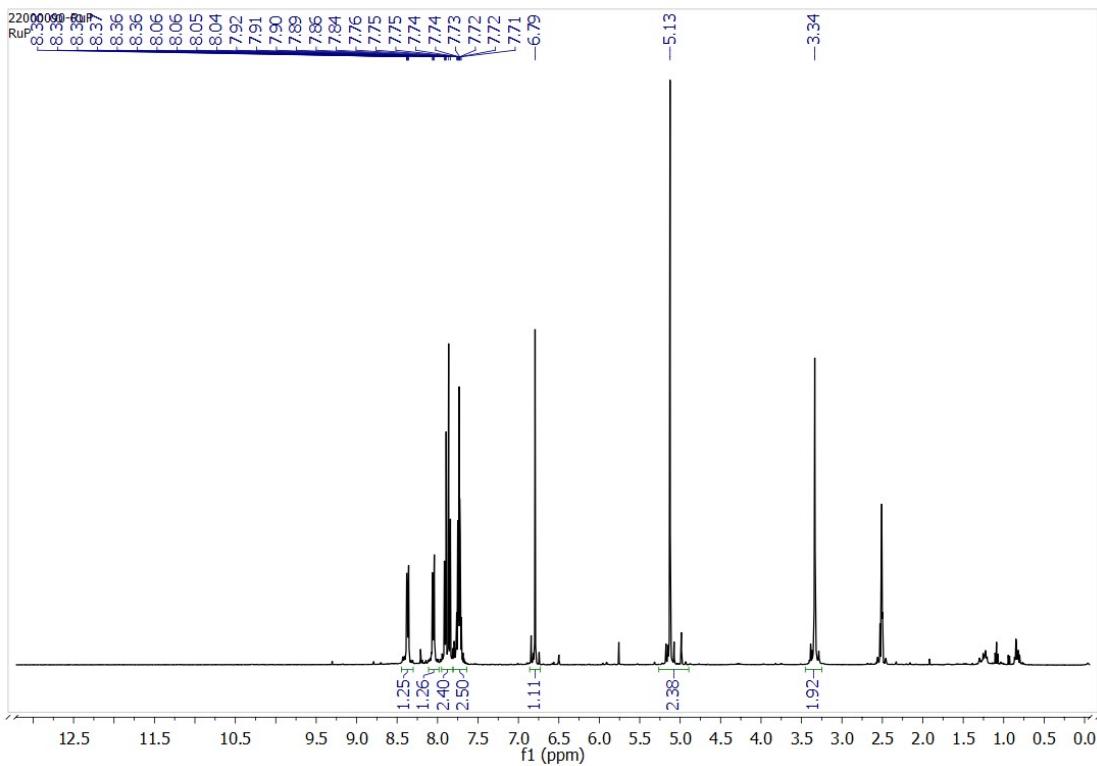
**Figure SI9:**  $^{13}\text{C}$ -NMR spectrum of **6** in  $d_6$ -DMSO at room temperature.



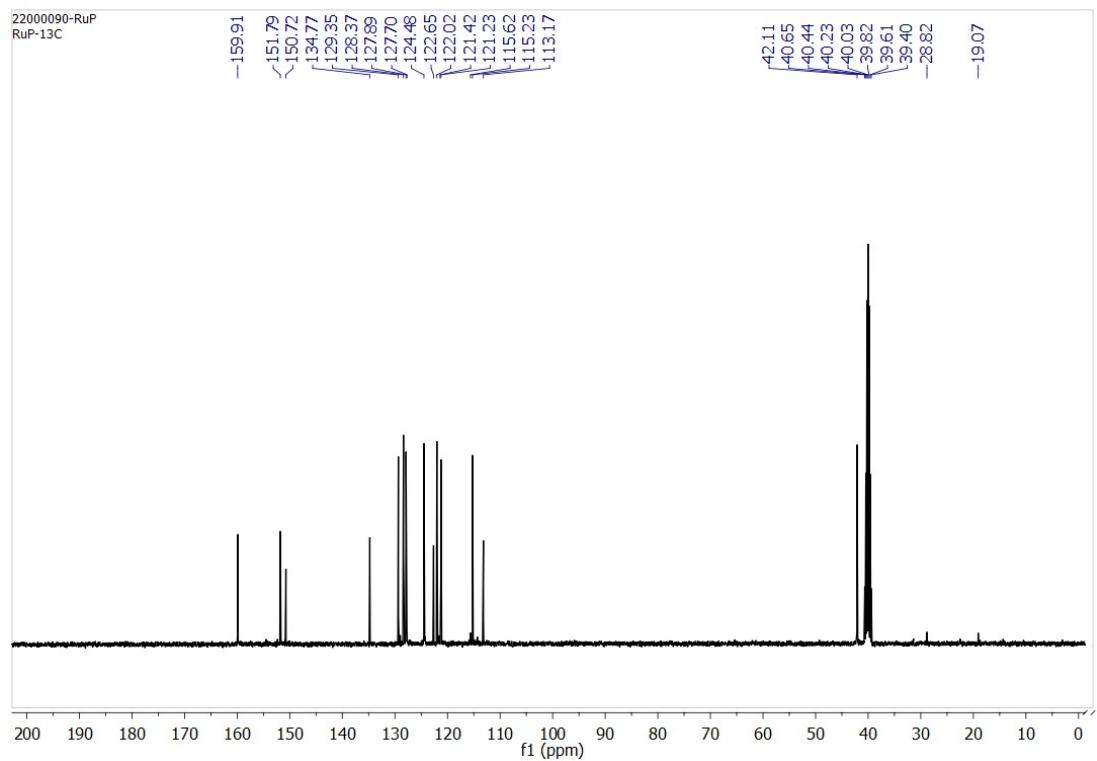
**Figure SI10:**  $^1\text{H}$ -NMR spectrum of **5** in  $d_6$ -DMSO at room temperature.



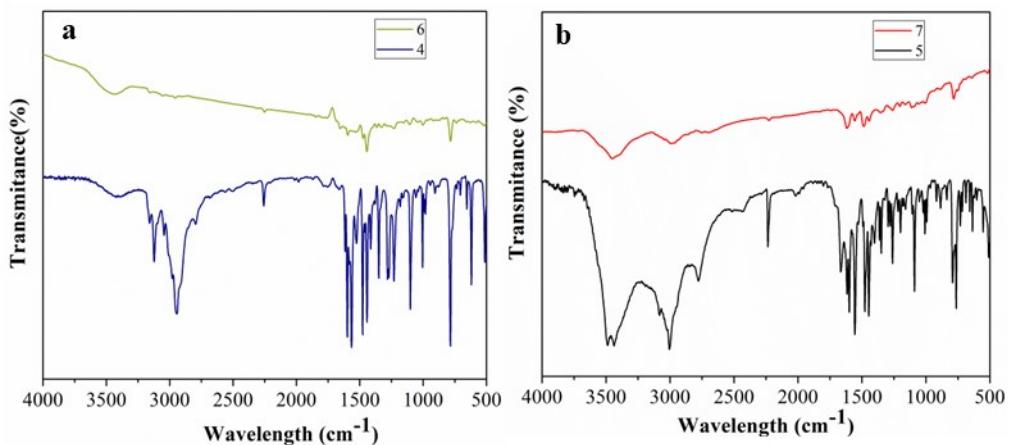
**Figure SI11:**  $^{13}\text{C}$ -NMR spectrum of **5** in  $d_6$ -DMSO at room temperature.



**Figure SI12:**  $^1\text{H}$ -NMR spectrum of **7** in  $d_6$ -DMSO at room temperature.



**Figure SI13:**  $^{13}\text{C}$ -NMR spectrum of **7** in  $d_6$ -DMSO at room temperature.



**Figure SI14:** IR spectra of salts **4** and **5** and their respective ruthenium organometallic polymers **6** and **7**.

### Equation 1

Limit of detection (LOD)

$$\text{LOD} = 3 * \text{SD/S}; \text{ where SD is the standard deviation of blank and S is the slope of calibration curve.}$$

### Equation 2

Limit of quantification (LOQ)

$$\text{LOQ} = 10 * \text{SD/S}; \text{ where SD is the standard deviation of blank and S is the slope of calibration curve.}$$

### Equation 3

Sensitivity = S/active surface area of working electrode; where S is the slope of the calibration curve.

### References

- [1] K. Fu, R. Zhang, J. He, H. Bai, and G. Zhang, “Sensitive detection of ketamine with an electrochemical sensor based on UV-induced polymerized molecularly imprinted membranes at graphene and MOFs modified electrode,” *Biosens. Bioelectron.*, vol. 143, no. May, p. 111636, 2019, doi: 10.1016/j.bios.2019.111636.
- [2] Q. Li, W. Tang, Y. Wang, J. Di, J. Yang, and Y. Wu, “Electrochemiluminescence immunosensor for ketamine detection based on polyamidoamine-coated carbon dot

- film," *J. Solid State Electrochem.*, vol. 19, no. 10, pp. 2973–2980, 2015, doi: 10.1007/s10008-015-2913-9.
- [3] Z. Bagheryan, J. B. Raoof, R. Ojani, and E. Hamidi-Asl, "Introduction of ketamine as a G-quadruplex-binding ligand using platinum nanoparticle modified carbon paste electrode," *Electroanalysis*, vol. 25, no. 12, pp. 2659–2667, 2013, doi: 10.1002/elan.201300418.
- [4] K. Lian, P. Zhang, L. Niu, S. Liu, L. Jiang, and W. Kang, "A novel derivatization approach for determination of ketamine in urine and plasma by gas chromatography-mass spectrometry," *J. Chromatogr. A*, vol. 1264, pp. 104–109, 2012, doi: 10.1016/j.chroma.2012.09.058.
- [5] R. A. Soomro, A. Nafady, K. R. Hallam, S. Jawaid, A. al Enizi, S. T. H. Sherazi, Sirajuddin, Z. H. Ibupoto, and M. Willander, "Highly sensitive determination of atropine using cobalt oxide nanostructures: Influence of functional groups on the signal sensitivity," *Anal. Chim. Acta*, vol. 948, pp. 30–39, 2016, doi: 10.1016/j.aca.2016.11.015.
- [6] R. A. Dar, P. K. Brahman, S. Tiwari, and K. S. Pitre, "Electrochemical determination of atropine at multi-wall carbon nanotube electrode based on the enhancement effect of sodium dodecyl benzene sulfonate," *Colloids Surfaces B Biointerfaces*, vol. 91, no. 1, pp. 10–17, 2012, doi: 10.1016/j.colsurfb.2011.10.020.
- [7] O. Ramdani, J. P. Metters, L. C. S. Figueiredo-Filho, O. Fatibello-Filho, and C. E. Banks, "Forensic electrochemistry: Sensing the molecule of murder atropine," *Analyst*, vol. 138, no. 4, pp. 1053–1059, 2013, doi: 10.1039/c2an36450f.
- [8] A. F. João, R. G. Rocha, T. A. Matias, E. M. Richter, J. Flávio S. Petrucci, and R. A. A. Muñoz, "3D-printing in forensic electrochemistry: Atropine determination in beverages using an additively manufactured graphene-polylactic acid electrode," *Microchem. J.*, vol. 167, no. February, 2021, doi: 10.1016/j.microc.2021.106324.
- [9] G. R. Fulmer, A. J. M. Miller, N. H. Sherden, H. E. Gottlieb, A. Nudelman, B. M. Bercaw, and K. Goldberg, "NMR chemical shifts of trace impurities: Common laboratory solvents, organics, and gases in deuterated solvents relevant to the organometallic chemist," *Organometallics*, vol. 29, no. 9, pp. 2176–2179, 2010, doi: 10.1021/om100106e.