

## Supporting Information

Detection and infectivity of SARS-CoV-2 in Korean municipal wastewater facilities and characterization of environmental factors influencing wastewater-bound SARS-CoV-2

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**SI Table S1.**

Primers and probes for RT-qPCR PEDV quantification, cycling conditions, and equations for calculating the recovery rate.

<b>Forward primer</b>	<b>Reverse primer</b>	<b>Probe</b>	<b>Cycling conditions</b>	<b>Recovery rate calculation</b>
5'-CGC AAA GAC TGA ACC CAC TAA TTT-3'	5'-TTG CCT CTG TTG TTA CTT GGA GAT-3'	5'-HEX- TGT TGC CAT TGC CAC GAC TCC TGC- BHQ2-3'	50°C for 20 min, 95°C for 10 min, 5 cycles of 95°C for 10s and 60°C for 30s, and 40 cycles of 95°C for 10s and 60°C for 30s.	Recovery rate =  <i>Concentration of quantified PEDV in wastewater after sample processing (gene copies/L)</i> <hr/> <i>Concentration of seeded PEDV in wastewater before sample processing (gene copies/L)</i>

**SI Table S2.**

Information of SARS-CoV-2 RT-qPCR detection kits.

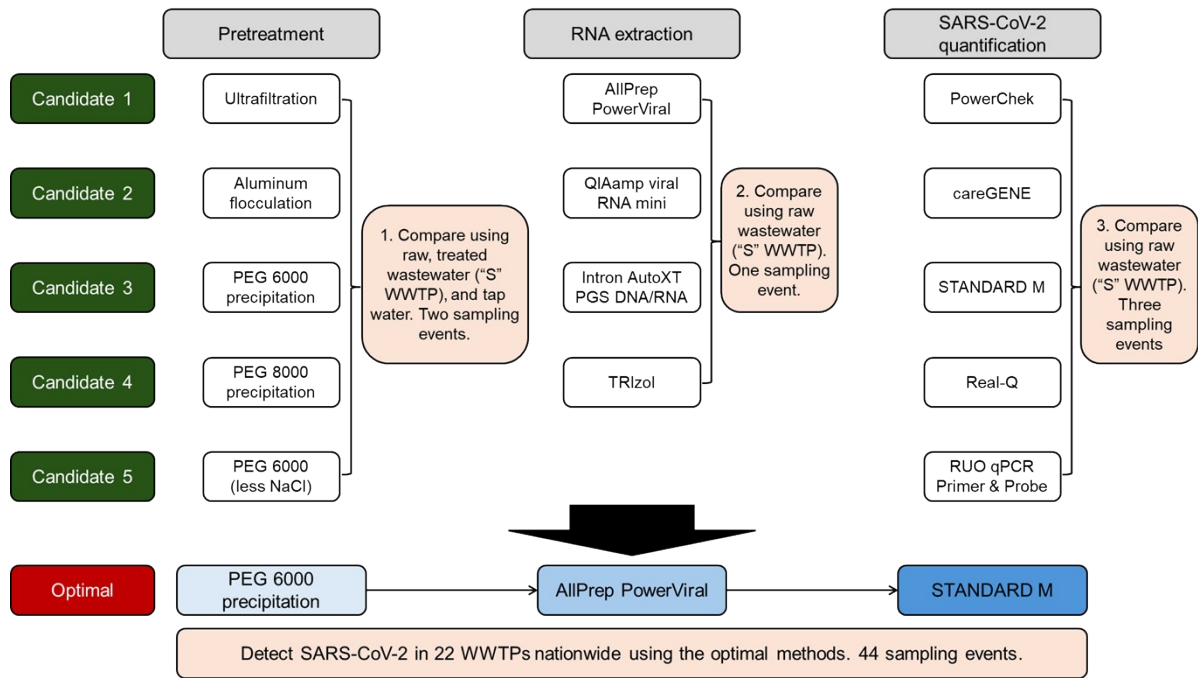
Kit	Target gene	Primers and probe (5'-3')	Cycling conditions	Limit of detection	Detection criteria for positive
PowerChek™ 2019-nCoV Real-time PCR		RdRp Forward: GTG ARA TGG TCA TGT GTG GCG G	50 °C for 30 min, 95 °C for 10 min, and 40 cycles of 95 °C for 15 s and 60 °C for 1 min.	18 gene copies /reaction	RdRp $C_t \leq 37$ , E gene $C_t \leq 37$ , and positive control $C_t \leq 28$
careGENE™ N-CoV RT-PCR		RdRp Reverse: CAR ATG TTA AAS ACA CTA TTA GCA TA	25 °C for 2 min, 55 °C for 10 min, 94 °C for 3 min, and 45 cycles of 94 °C for 15 s and 60 °C for 30 s.	50 gene copies /reaction	RdRp, E gene, and positive control $C_t \leq 45$
STANDAR D™ M nCoV Real-time Detection	RdRp and E gene	RdRp Probe: FAM- CAG GTG GAA CCT CAT CAG GAG ATG C-BHQ	50 °C for 15 min, 95 °C for 3 min, 5 cycles of 95 °C for 5 s and 60 °C for 40 s, and 40 cycles of 95 °C for 5 s and 60 °C for 40 s.	5 gene copies /reaction	RdRp $C_t \leq 36$ , E gene $C_t \leq 36$ , and positive control $C_t \leq 32$
Real-Q 2019-nCoV Detection		E gene Forward: ACA GGT ACG TTA ATA GTT AAT AGC GT	50 °C for 30 min, 95 °C for 15 min and 40 cycles at 95 °C for 15 s, and 62 °C for 45 s	31.25 gene copies /reaction	RdRp $C_t \leq 38$ , E gene $C_t \leq 38$ , and positive control $C_t \leq 28 \pm 5$
SARS-CoV-2 Research Use Only qPCR Primer & Probe Kit	RdRp	E gene Reverse: ATA TTG CAG CAG TAC GCA CAC A E gene Probe: FAM (HEX)-ACA CTA GCC ATC CTT ACT GCG CTT CG-BHQ	45 °C for 20 min, 95 °C for 2 min and 45 cycles at 95 °C for 10 s, and 55 °C for 30 s	35 gene copies/reaction (Barra et al., 2020; Kaya et al., 2022)	$C_t \leq 35$ and positive control $C_t \leq 35$

**SI Table S3.**

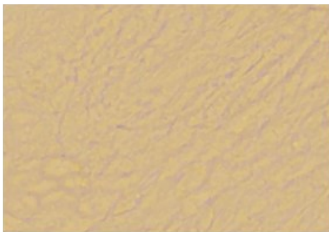

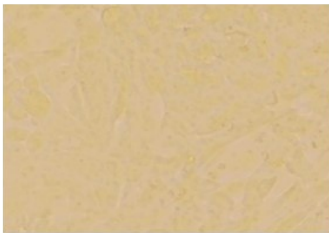
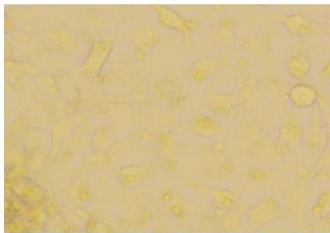

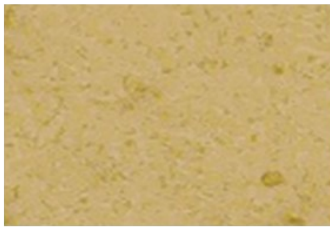
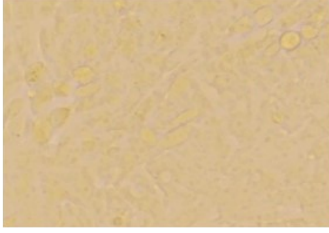

Cycle threshold results of RT-qPCR for SARS-CoV-2 detection in cell lysates from wastewater (WW) for infectivity testing.

Sample		Passage 1		Passage 2		Passage 3		Detection
		RdRp	E gene	RdRp	E gene	RdRp	E gene	
Raw	WW (Vero)	> 40	> 40	> 40	> 40	> 40	> 40	Negative
Frozen	WW (Vero)	> 40	> 40	> 40	> 40	> 40	> 40	Negative
Concentrated	WW (Vero)	> 40	> 40	> 40	> 40	> 40	> 40	Negative
Raw	WW (A549)	> 40	> 40	> 40	> 40	> 40	> 40	Negative
Frozen	WW (A549)	> 40	> 40	> 40	> 40	> 40	> 40	Negative
Concentrated	WW (A549)	> 40	> 40	> 40	> 40	> 40	> 40	Negative
Negative	control	> 40	> 40	> 40	> 40	> 40	> 40	Negative
Positive	control <sup>1</sup>	24.18	26.57	24.05	27.78	26.12	27.10	Positive

<sup>1</sup>Positive material corresponding to 100 gene copies/reaction was used for RT-qPCR.



**SI Fig. S1.** Summary of candidate virus-detection methods (pretreatment, RNA extraction, and SARS-CoV-2 quantification methods; see Section 2.2 for details) and an optimal method based on a comparative study.

	Vero	A549
Raw wastewater		
Frozen wastewater		
Concentrated wastewater		
Negative control		

**SI Fig. S2** Infectivity results of wastewater-bound SARS-CoV-2 in four different samples tested on two different cell types. No cytopathic effects were observed in any of the samples.

## References

Barra, G. B., Santa Rita, T. H., Mesquita, P. G., Jácomo, R. H., & Nery, L. F. A. (2020). Analytical Sensitivity and Specificity of Two RT-qPCR Protocols for SARS-CoV-2 Detection Performed in an Automated Workflow. *Genes*, 11, 1183. <https://doi.org/10.3390/genes11101183>

Kaya, D., Niemeier, D., Ahmed, W., & Kjellerup, B. V. (2022). Evaluation of multiple analytical methods for SARS-CoV-2 surveillance in wastewater samples. *Science of The Total Environment*, 808, 152033. [https://doi.org/https://doi.org/10.1016/j.scitotenv.2021.152033](https://doi.org/10.1016/j.scitotenv.2021.152033)