

Supplementary Information

Chemoenzymatic β -Specific Methylene C(sp³)-H Deuteration of Carboxylic Acids

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1. General Information

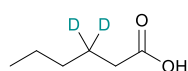
General. Analytical thin-layer chromatography was performed on glass plates coated with 0.25 mm 230-400 mesh silica gel containing a fluorescent indicator. Visualization was accomplished by exposure to a UV lamp, and/or treatment with a solution of phosphomolybdic acid (PMA) followed by brief heating with heating gun. Most of the products were compatible with standard silica gel chromatography. Column chromatography was performed on silica gel 60N (spherical and neutral, 200-300 mesh) using standard methods.

Structural analysis. NMR spectra were measured on a Bruker Avance-400 spectrometer and chemical shifts (δ) are reported in parts per million (ppm). ^1H NMR spectra were recorded at 400 MHz in NMR solvents (CDCl_3) and referenced internally to corresponding solvent resonance, and ^{13}C NMR spectra were recorded at 100 MHz, chemical shifts were reported in ppm on the δ scale relative to CHCl_3 ($\delta = 7.26$ for ^1H NMR, $\delta = 77.16$ for ^{13}C NMR). Coupling constants are reported in Hz with multiplicities denoted as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet) and br (broad). High-resolution mass Spectra (HRMS) were obtained on a WATERS I-Class VION IMS QToF using ESI.

Materials. Commercial reagents were purchased from J&K, Energy, Adamas, Sigma-Aldrich, Aladdin, Alfa Aesar, Acros Organics or TCI and used as received unless otherwise stated.

2. General procedure for one-pot β -Specific methylene C(sp³)-H deuteration of carboxylic acids

Carboxylic acids (5.0 mmol) and 8-aminoquinoline (6.0 mmol, 1.2 equiv.) were dissolved in toluene (5.0 mL), and then B(OCH₂CF₃)₃ (1.5 mmol, 0.3 equiv.) was added to the solution. The mixture was heated to reflux with a Dean-Stark setup for 48 h. After cooling down, to the required volume of solution (0.5 mL) was added acetic anhydride (0.20 mmol, 0.40 equiv.). The reaction mixture was stirred at room temperature for 2 h followed by additions of deuterium oxide (2.0 mL), palladium acetate (0.025 mmol, 5.0 mol%) and pivalic acid (0.50 mmol, 1.0 equiv.). After reaction at 90 °C for 24 h, to the cooled mixture were added NovoCor[®] ADL (2.5 mL) and water (200 mL). The resulting mixture was stirred at 50 °C for another 24 h. After cooling down to room temperature, the mixture was filtered through diatomite and extracted with ethyl acetate. The organic phase was concentrated to about 1 mL, then extracted three times with 1.0 M NaOH aq., then 1.0 M HCl aq. was added to acidification until pH = 2, and then the water phase was extracted three times with DCM. The combined organic phases were dried over anhydrous magnesium sulfate and concentrated in vacuo to obtain the deuterated carboxylic acids. The residue was purified by flash column chromatography (silica gel, ethyl acetate/petroleum ether or DCM/MeOH) if necessary. Deuterium incorporation was determined by ¹H NMR analysis of the corresponding benzyl ester or methyl ester.

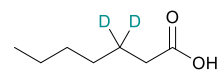


1b: The title compound was prepared on a 0.5 mmol scale and obtained as a yellow oil (50 mg, 83% yield, 94% deuterium incorporation).

¹H NMR (400 MHz, CDCl₃) δ 2.33 (s, 2H), 1.27-1.36 (m, 4H), 0.93-0.86 (t, 3H, J = 6.6 Hz).

¹³C NMR (100 MHz, CDCl₃) δ 180.4, 33.9, 31.0, 23.7, 22.4-23.7 (m, β -CD₂), 13.9.

HRMS (ESI-) m/z calcd. for C₆H₉D₂O₂⁻ [M-H]⁻: 117.0890, found: 117.0889

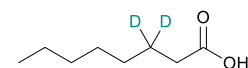


2b: The title compound was prepared on a 0.5 mmol scale and obtained as a yellow oil (65 mg, 98% yield, 98% deuterium incorporation).

¹H NMR (400 MHz, CDCl₃) δ 2.34 (s, 2H), 1.25-1.36 (m, 6H), 0.89 (t, 3H, J = 6.6 Hz).

¹³C NMR (100 MHz, CDCl₃) δ 180.7, 33.9, 31.4, 28.5, 23.7-24.1 (m, β -CD₂), 22.5, 14.0.

HRMS (ESI-) m/z calcd. for C₇H₁₁D₂O₂⁻ [M-H]⁻: 131.1046, found: 131.1044



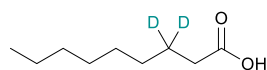
3b: The title compound was prepared on a 0.5 mmol scale and obtained as a yellow oil (62 mg, 85% yield, 96% deuterium

incorporation).

¹H NMR (400 MHz, CDCl₃) δ 2.34 (s, 2H), 1.25-1.33 (m, 8H), 0.88 (t, 3H, *J* = 6.6 Hz).

¹³C NMR (100 MHz, CDCl₃) δ 180.3, 33.9, 31.7, 28.9, 28.8, 23.8-24.2 (m, β-CD₂), 22.6, 14.1.

HRMS (ESI-) *m/z* calcd. for C₈H₁₃D₂O₂⁻ [M-H]⁻: 145.1203, found: 145.1205



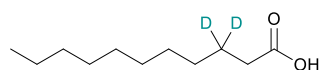
incorporation).

4b: The title compound was prepared on a 0.5 mmol scale and obtained as a yellow oil (65 mg, 81% yield, 96% deuterium

¹H NMR (400 MHz, CDCl₃): δ 2.33 (s, 2H), 1.22-1.35 (m, 10H), 0.87 (t, 3H, *J* = 6.6 Hz).

¹³C NMR (100 MHz, CDCl₃): δ 180.6, 33.9, 31.8, 29.2, 29.1, 28.9, 23.6-24.2 (m, β-CD₂), 22.7, 14.1.

HRMS (ESI-) *m/z* calcd. for C₉H₁₅D₂O₂⁻ [M-H]⁻: 159.1360, found: 159.1358



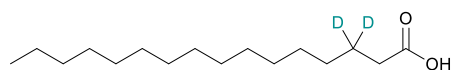
deuterium incorporation).

5b: The title compound was prepared on a 0.5 mmol scale and obtained as a colorless solid (46 mg, 49% yield, 97%

¹H NMR (400 MHz, CDCl₃): δ 2.33 (s, 2H), 1.23-1.32 (m, 14H), 0.88 (t, 3H, *J* = 6.6 Hz).

¹³C NMR (100 MHz, CDCl₃): δ 180.5, 33.9, 31.9, 29.5, 29.4, 29.3, 29.2, 28.8, 23.7-24.3 (m, β-CD₂), 22.7, 14.1.

HRMS (ESI-) *m/z* calcd. for C₁₁H₁₉D₂O₂⁻ [M-H]⁻: 187.1673, found: 187.1674



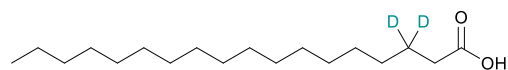
69% yield, 95% deuterium incorporation).

6b: The title compound was prepared on a 0.5 mmol scale and obtained as a white solid (89 mg,

¹H NMR (400 MHz, CDCl₃): δ 2.33 (s, 2H), 1.21-1.31 (m, 24H), 0.88 (t, 3H, *J* = 6.6 Hz).

¹³C NMR (100 MHz, CDCl₃): δ 180.0, 33.8, 31.9, 29.71, 29.70, 29.69, 29.68, 29.66, 29.61, 29.4, 29.3, 29.2, 28.8, 23.5 (m, β-CD₂), 22.7, 14.1.

HRMS (ESI-) *m/z* calcd. for C₁₆H₃₁D₂O₂⁻ [M-H]⁻: 257.2455, found: 257.2455

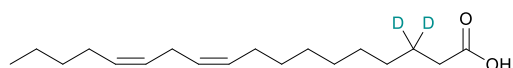


7b: The title compound was prepared on a 0.5 mmol scale and obtained as a white solid (61 mg, 43% yield, 97% deuterium incorporation).

¹H NMR (400 MHz, CDCl₃): δ 2.33 (s, 2H), 1.19-1.34 (m, 28H), 0.87 (t, 3H, *J* = 6.2 Hz).

¹³C NMR (100 MHz, CDCl₃): δ 180.4, 33.9, 31.9, 29.7, 29.69, 29.67, 29.62, 29.4, 29.3, 29.2, 28.8, 23.7-24.1 (m, β-CD₂), 22.7, 14.1.

HRMS (ESI-) *m/z* calcd. for C₁₈H₃₃D₂O₂⁻ [M-H]⁻: 285.2768, found: 285.2754

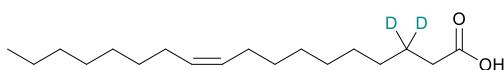


8b: The title compound was prepared on a 0.5 mmol scale and obtained as a yellow oil (87 mg, 62% yield, 95% deuterium incorporation).

¹H NMR (400 MHz, CDCl₃): δ 5.28-5.41 (m, 4H), 2.77 (t, 2H, *J* = 6.4 Hz), 2.33 (s, 2H), 2.01-2.08 (m, 4H), 1.24-1.40 (m, 14H), 0.89 (t, 3H, *J* = 6.7 Hz).

¹³C NMR (100 MHz, CDCl₃): δ 180.6, 130.2, 130.0, 128.0, 127.9, 33.9, 31.5, 29.6, 29.3, 29.12, 29.12, 28.8, 27.22, 27.20, 25.6, 23.5-24.3 (m, β-CD₂), 22.6, 14.1.

HRMS (ESI-) *m/z* calcd. for C₁₈H₂₉D₂O₂⁻ [M-H]⁻: 281.2455, found: 281.2453

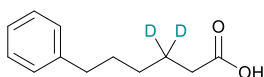


9b: The title compound was prepared on a 0.5 mmol scale and obtained as a yellow oil (130 mg, 61% yield, 94% deuterium incorporation).

¹H NMR (400 MHz, CDCl₃): δ 5.30-5.39 (m, 2H), 2.33 (s, 2H), 1.95-2.08 (m, 4H), 1.41-1.18 (m, 20H), 0.88 (t, 3H, *J* = 6.7 Hz).

¹³C NMR (100 MHz, CDCl₃): δ 180.4, 130.0, 129.7, 33.9, 31.9, 29.7, 29.6, 29.5, 29.4, 29.3, 29.2, 29.1, 28.8, 27.24, 27.17, 23.7-24.1 (m, β-CD₂), 22.7, 14.1.

HRMS (ESI-) *m/z* calcd. for C₁₈H₃₁D₂O₂⁻ [M-H]⁻: 283.2612, found: 283.2609

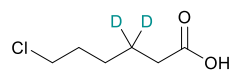


10b: The title compound was prepared on a 0.5 mmol scale and obtained as a yellow oil by flash column chromatography (ethyl acetate/petroleum ether = 1/5, *R_f* = 0.28) (81 mg, 84% yield, 96% deuterium incorporation).

¹H NMR (400 MHz, CDCl₃): δ 7.16-7.25 (m, 2H), 7.03-7.15 (m, 3H), 2.53 (t, 2H, *J* = 7.7 Hz), 2.27 (s, 2H), 1.50-1.62 (m, 2H), 1.31 (dt, 2H, *J* = 7.2, 6.0 Hz).

¹³C NMR (100 MHz, CDCl₃): δ 180.4, 142.4, 128.4, 128.3, 125.7, 35.7, 31.1, 31.1, 28.7, 28.5, 24.4-24.7 (m, β-CD₂).

HRMS (ESI-) m/z calcd. for $C_{12}H_{13}D_2O_2^-$ $[M-H]^-$: 193.1203, found: 193.1201

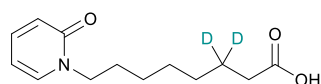


11b: The title compound was prepared on a 0.5 mmol scale and obtained as a yellow oil (32 mg, 42% yield, 91% deuterium incorporation).

1H NMR (400 MHz, $CDCl_3$): δ 3.53 (t, 2H, $J = 6.4$ Hz), 2.37 (t, 2H, $J = 6.5$ Hz), 1.74-1.84 (m, 2H), 1.45-1.53 (m, 2H).

^{13}C NMR (100 MHz, $CDCl_3$): δ 180.2, 44.7, 33.7, 32.1, 26.1, 23.2-23.6 (m, β - CD_2).

HRMS (ESI-) m/z calcd. for $C_6H_8D_2ClO_2^-$ $[M-H]^-$: 151.0500, found: 151.0498

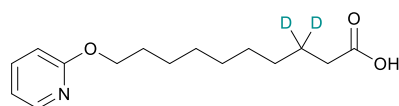


12b: The title compound was prepared on a 0.5 mmol scale and obtained as a white solid by flash column chromatography (DCM/ MeOH = 10: 1, $R_f = 0.25$) (38 mg, 32% yield, 93% deuterium incorporation).

1H NMR (400 MHz, $CDCl_3$): δ 7.29 (m, 2H), 6.57 (d, 1H, $J = 9.1$ Hz), 6.17 (dd, 1H, $J = 6.8$ Hz, 6.8 Hz), 3.88 (t, 2H, $J = 7.5$ Hz), 2.19 (s, 2H), 1.68 (dt, 2H, $J = 7.1$ Hz, 6.8 Hz), 1.39-1.17 (m, 6H).

^{13}C NMR (100 MHz, $CDCl_3$): δ 181.1, 162.8, 139.5, 137.6, 120.9, 106.3, 49.9, 35.1, 33.8, 29.0, 28.6, 26.2, 24.4-24.8 (m, β - CD_2).

HRMS (ESI-) m/z calcd. for $C_{13}H_{16}D_2NO_3^-$ $[M-H]^-$: 238.1418, found: 238.1416

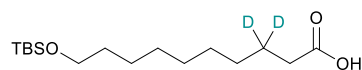


13b: The title compound was prepared on a 0.5 mmol scale and obtained as a colorless solid by flash column chromatography (DCM/ MeOH = 10: 1, $R_f = 0.30$) (89 mg, 67% yield, 95% deuterium incorporation).

1H NMR (400 MHz, $CDCl_3$): δ 8.15 (dd, 1H, $J = 5.1, 1.9$ Hz), 7.60-7.52 (m, 1H), 6.84 (dd, 1H, $J = 7.0, 5.1$ Hz), 6.72 (d, 1H, $J = 8.3$ Hz), 4.26 (t, 2H, $J = 6.7$ Hz), 2.33 (s, 2H), 1.76 (m, 2H), 1.43 (m, 2H), 1.26-1.35 (m, 8H).

^{13}C NMR (100 MHz, $CDCl_3$): δ 179.4, 164.0, 146.8, 138.6, 116.5, 111.0, 66.1, 33.8, 29.29, 29.24, 29.05, 29.03, 28.7, 25.9, 24.6-24.8 (m, β - CD_2).

HRMS (ESI-) m/z calcd. for $C_{15}H_{20}D_2NO_3^-$ $[M-H]^-$: 266.1730, found: 266.1729

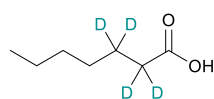


14b: The title compound was prepared on a 0.5 mmol scale and obtained as a yellow oil (44 mg, 29% yield, 95% deuterium incorporation).

¹H NMR (400 MHz, CDCl₃): δ 3.59 (t, 2H, *J* = 6.6 Hz), 2.33 (s, 2H), 1.50 (t, 2H, *J* = 6.8 Hz), 1.26-1.33 (m, 10H), 0.89 (s, 9H), 0.04 (s, 6H).

¹³C NMR (100 MHz, CDCl₃): δ 179.3, 63.3, 33.8, 32.8, 29.4, 29.3, 29.1, 28.8, 26.0, 25.6-25.9 (m, β-CD₂), 18.4, -5.2.

HRMS (ESI-) *m/z* calcd. for C₁₆H₃₁D₂O₃Si⁻ [M-H]⁻: 303.2330, found: 303.2329

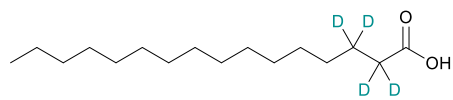


15b: The title compound was prepared on a 0.5 mmol scale and obtained as a yellow oil (66 mg, 98% yield, α-96%, β-98% deuterium incorporation).

¹H NMR (400 MHz, CDCl₃): δ 1.24-1.35 (m, 6H), 0.88 (t, 3H, *J* = 6.5 Hz).

¹³C NMR (100 MHz, CDCl₃): δ 180.7, 33.2-33.6 (m, α-CD₂), 31.3, 28.4, 23.6-23.9 (m, β-CD₂), 22.4, 14.0.

HRMS (ESI-) *m/z* calcd. for C₇H₉D₄O₂⁻ [M-H]⁻: 133.1172, found: 133.1170



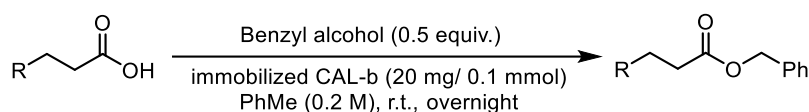
16b: The title compound was prepared on a 0.5 mmol scale and obtained as a yellow oil (87 mg, 67% yield, α-94%, β-96% deuterium incorporation).

¹H NMR (400 MHz, CDCl₃): δ 1.24-1.31 (m, 24H), 0.88 (t, 3H, *J* = 6.8 Hz).

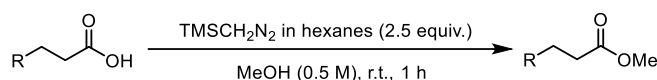
¹³C NMR (100 MHz, CDCl₃): δ 184.2, 37.4-37.7 (m, α-CD₂), 30.9, 28.68, 28.66, 28.64, 28.63, 28.5, 28.4, 28.3, 28.1, 27.7, 25.9, 23.3-23.6 (m, β-CD₂), 21.6, 13.1.

HRMS (ESI-) *m/z* calcd. for C₁₆H₂₇D₄O₂⁻ [M-H]⁻: 259.2580, found: 259.2574

3. Procedure for esterification



The carboxylic acid (0.10 mmol), alcohol (0.50 mmol) and Immobilized *Candida Antarctica* Lipase-B (20 mg) were added to toluene (0.50 mL), and the reaction was stirred at room temperature overnight. After filtration and concentration in vacuum, the corresponding esters were obtained quantitatively. Using the enzyme-catalyzed esterification method developed by us, no dedeuteration products were observed.



The carboxylic acid (0.10 mmol) was dissolved in 0.20 mL of MeOH, (trimethylsilyl)diazomethane (2.5 equiv., 2.0 M in hexanes) was added dropwise until the colorless reaction solution turned to bright yellow. The reaction mixture was stirred for another 30 min, and quenched with acetic acid. The reaction mixture became colorless. After dilution and extraction with ether, the organic layers were collected and dried with MgSO₄ and removed the solvent in vacuum. The corresponding methyl esters was obtained without further purification.¹

4. Procedure for gram scale reaction

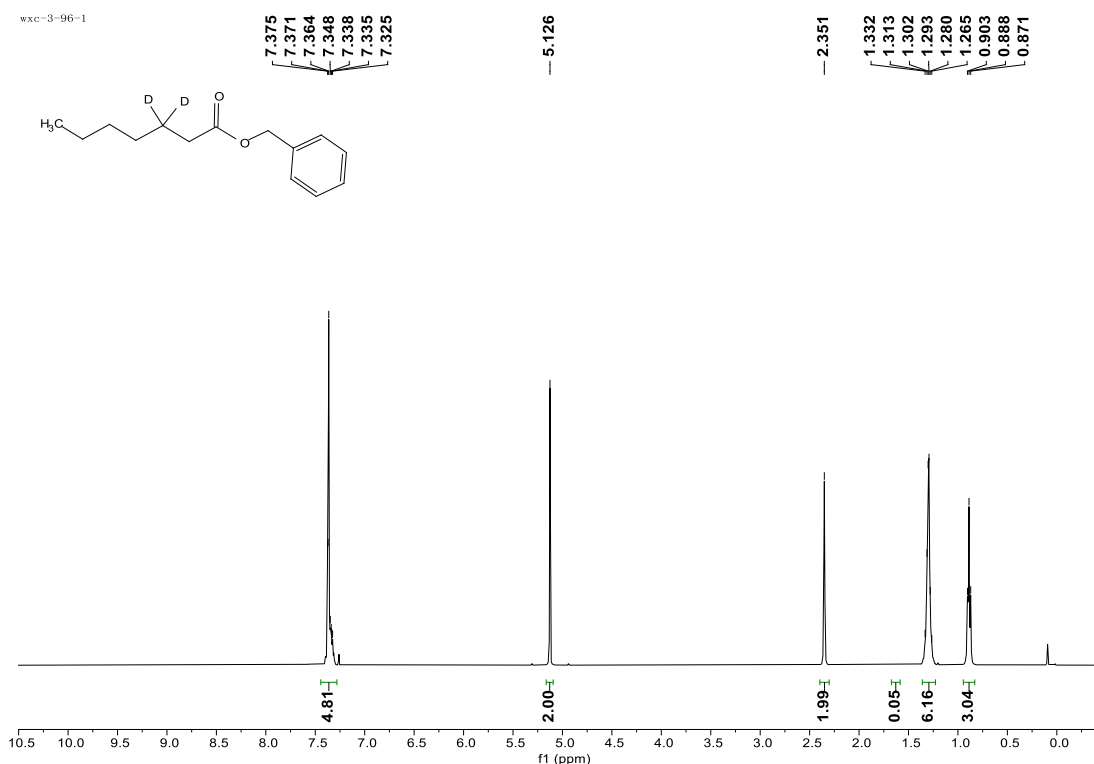
The carboxylic acid (5.0 mmol) and 8-aminoquinoline (6.0 mmol, 1.2 equiv.) were dissolved in 5.0 mL of toluene, and then B(OCH₂CF₃)₃ (1.5 mmol, 0.30 equiv.) was added to the solution. The mixture was stirred with a Dean-Stark system for 48 h in air. Then acetic anhydride (2.0 mmol, 0.4 equiv.) was added in the solution. The reaction mixture was stirred at room temperature for 2h, and deuterium oxide (20 mL), palladium acetate (0.25 mmol, 5.0 mol%) and trimethylacetic acid (5.0 mmol, 1.0 equiv.) was added to the solution. The solution was stirred at 90 °C for 24 h in air. Then 2.0 L water and 20 mL NovoCor[®] ADL was added in the tube. The mixture was stirred at 50 °C for 24 h in air. The mixture was filtered, and the water was concentrated to about 200 mL then extracted three times with ethyl acetate. The organic phase was concentrated to about 10 mL, then extracted three times with 1.0 M NaOH aq., then 1.0 M HCl solution was added to acidification until pH = 2, and then the water phase was extracted three times with DCM. The combined organic phases were dried over anhydrous magnesium sulfate and concentrated in vacuo to obtain the deuterated carboxylic acids.

5. Determination of deuteration rates by NMR spectroscopy

The decrease of the proton signal intensity was measured and compared with the values obtained from non-deuterated starting materials. The corresponding methyl- or benzyl ester of the carboxylic acid was synthesized, and the protons of methyl or benzyl group were used as internal standard to calculate the relative decrease in ^1H -signal intensity at the deuteration site.

$$\text{deuteration} = \left[100 - \left(\frac{\text{residual integral}}{\text{number of labelling sites}} \times 100 \right) \right] \%$$

Take product **2b** for example:



After esterification, we use the two proton signals at the benzyl position as an integral of 2 as the internal standard, the deuteration rate is:

$$\text{deuteration} = \left(100 - \frac{0.05}{2.00} \times 100 \right) \% = 98\%$$

6. General procedure for optimization experiments of amidation reaction, deuteration reaction and hydrolysis reaction.

Amidation reaction

To a 25 mL vial containing a magnetic stirrer were added *n*-octanoic acid (0.72 g, 5.00 mmol), 8-aminoquinoline (0.87 g, 6.00 mmol, 1.20 equiv.) and 5.00 mL of toluene, followed by addition of the selected amount of catalyst in open air. After heating for required reaction time, 1/10 of the reaction mixture was transferred to a vial and then dried under vacuum. To the residue was added 0.5 mL of CDCl₃ and 0.5 mmol of dibromomethane in sequence. After complete dissolution, the clear solution was transferred to an NMR tube and the ¹H NMR spectrum was recorded. The peak area of dibromomethane (4.95 ppm) was integrated as 2.00, then the carbonyl α-proton of the amide product was integrated as “x”. The yield was determined as:

$$\text{yield} = \frac{x}{2} \times 100\%$$

Deuteration reaction

N-(Quinoline-8-yl) octanamide (27.0 mg, 0.1 mmol) and the additives were added to a 10 mL Schlenk tube, and then the selected amount of Pd(OAc)₂, D₂O (0.4 mL) and an oven-dried PTFE-coated magnetic stirrer were added to the solution in open air. After heating for the required time, the reaction mixture was extracted 3 times by ethyl acetate. The solvent of the combined organic phases was removed under vacuum, and then 0.5 mL of CDCl₃ and 0.1 mmol of dibromomethane were added to the vial in sequence. The resulting solution was transferred to an NMR tube and the ¹H NMR spectrum was recorded. The peak area of dibromomethane (4.95 ppm) was integrated as 2.00, then the carbonyl β-proton of the amide product was integrated as “y”. The yield was determined as:

$$\text{deuteration} = \frac{2-y}{2} \times 100\%$$

Amide hydrolysis reaction

To a 10 mL vial containing a magnetic stirrer were added *n*-(quinoline-8-yl) octanamide (2.70 mg, 0.01 mmol), lipase, *n*-dodecane (1.70 mg, 0.01 mmol, 1.00 equiv.) and tap water in open air. The resulting mixture was heated for the required reaction time. The yield was determined by GC-FID method using *n*-dodecane as internal standard. After the reaction, the reaction mixture was ultrasonicated for 1 min. A sample of 0.01 mL solution was transferred to a centrifugal tube and 1.0 mL of ethyl acetate was added and centrifuged. Transfer 0.1 mL of supernatant into another centrifuge tube and add 1.0 mL of ethyl acetate for centrifugation. Next, 0.5 mL of supernatant was transferred to a chromatography tube, 1.0 mL ethyl acetate was added, and GC experiment was performed.

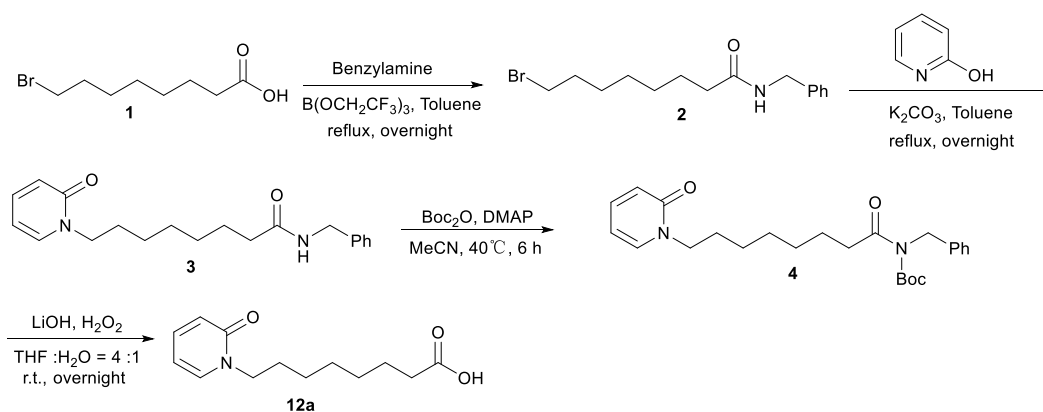
GC-FID analysis: Yields were determined by GC analysis on a Thermo Scientific Trace 1300 system using nitrogen as carrier gas and a TRACE™ TR-1MS GC column (100% dimethylpolysiloxane stationary phase, 30 m × 0.25 mm × 0.25 μm). The following

acquisition parameters were used: Oven program: 50 °C for 1 min, 20 °C/min to 210 °C for 5 min, 10 °C/min to 280 °C for 1 min, 5 °C/min to 320 °C for 1 min; total flow rate: 15 mL/min; injector temperature: 220 °C; detector temperature: 350 °C; splitless injection.

Retention time: *n*-dodecane (8.2 min), octanoic acid (9.3 min), 8-aminoquinoline (15.5 min), *N*-(quinoline-8-yl) octanamide (28.3 min)

7. Synthesis of starting materials

Synthesis of 8-(2-oxopyridin-1(2H)-yl)octanoic acid (12a)



Synthesis of Compound 2

1 (2.20 g, 10.0 mmol), benzylamine (0.963 g, 9.00 mmol, 0.900 equiv.) and $B(OCH_2CF_3)_3$ (0.308 g, 1.00 mmol, 10 mol%) was added in 10 mL toluene. The reaction mixture was refluxed overnight. After extraction, washed with 1 M NaOH (aq), dried over anhydrous Na_2SO_4 and concentrated in vacuum, the corresponding product **2** was afforded through silica gel column chromatography (ethyl acetate/hexanes) in 2.44 g (87% yield, m.p. : 83-85 °C).

¹H NMR (400 MHz, $CDCl_3$) δ 7.39 – 7.22 (m, 5H), 5.87 (s, 1H), 4.42 (d, $J = 5.7$ Hz, 2H), 3.39 (t, $J = 6.8$ Hz, 2H), 2.20 (t, $J = 7.6$ Hz, 2H), 1.83 (p, $J = 7.0$ Hz, 2H), 1.65 (p, $J = 7.3$ Hz, 2H), 1.42 (p, $J = 6.8$ Hz, 2H), 1.28 - 1.36 (m, 4H).

¹³C NMR (100 MHz, $CDCl_3$) δ 172.8, 138.3, 128.6, 127.7, 127.4, 43.5, 36.6, 33.9, 32.6, 29.0, 28.4, 27.9, 25.5.

HRMS (ESI+) m/z calcd. for $C_{15}H_{22}NO_2Br$ $[M+H]^+$: 312.0958, found: 312.0957.

Synthesis of Compound 3

Compound **2** (2.49 g, 8.00 mmol), 2-hydroxypyridine (0.912 g, 9.60 mmol, 1.20 equiv.) and K_2CO_3 (3.31 g, 24.0 mmol, 3.00 equiv.) were added to toluene (16.0 mL). The reaction mixture was refluxed overnight. The reaction mixture was extracted with DCM three times, and the combined extracts were washed with brine, dried over anhydrous Na_2SO_4 , and concentrated under vacuum. The corresponding product **3** was afforded

through silica gel column chromatography (methanol/dichloromethane) in 1.96 g (75% yield, m.p. : 65-69 °C).

¹H NMR (400 MHz, CDCl₃) δ 7.17-7.36 (m, 7H), 6.75 (d, *J* = 17.9 Hz, 1H), 6.45 (d, *J* = 9.1 Hz, 1H), 6.15 (t, *J* = 6.7 Hz, 1H), 4.40 (d, *J* = 5.4 Hz, 2H), 3.85 (d, *J* = 7.0 Hz, 2H), 2.20 (t, *J* = 7.5 Hz, 2H), 1.65 (dp, *J* = 21.8, 7.2 Hz, 4H), 1.25-1.36 (m, 6H).

¹³C NMR (100 MHz, CDCl₃) δ 173.2, 139.3, 138.7, 128.5, 127.7, 127.2, 49.6, 43.3, 36.3, 29.1, 28.9, 26.8, 26.2, 25.5.

HRMS (ESI+) *m/z* calcd. for C₂₀H₂₆N₂O₂ [M+H]⁺: 327.2067, found: 327.2064.

Synthesis of Compound 4

Compound **3** (1.96 mg, 6.00 mmol), Boc₂O (3.93 g, 18.0 mmol, 3.00 equiv.), DMAP (0.740 g, 0.600 mmol, 0.100 equiv.) were dissolved in MeCN (24.0 mL), and the reaction mixture was stirred at 40 °C for 6 h. After concentrated in vacuum, the corresponding product **4** was afforded through silica gel column chromatography (methanol/dichloromethane) in 2.38 g (93% yield).

Synthesis of Compound 12a

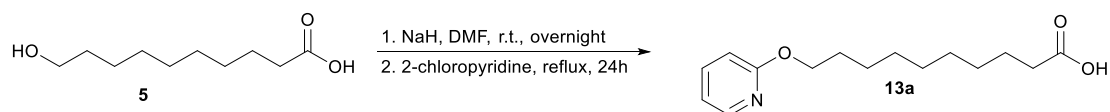
To a solution of compound **4** (2.35 g, 5.50 mmol) in THF/ H₂O (4:1, 110 mL), LiOH (0.360 g, 15.0 mmol, 2.72 equiv.) and H₂O₂ (3.30 mL, 30 % wt in H₂O) was added. The reaction mixture was stirred at room temperature overnight. Upon completion, saturated aqueous Na₂SO₃ solution was added, and the solution was stirred for another 1 h. After acidification with 1 M Hydrochloric acid solution, the reaction mixture was extracted with DCM three times. The combined extracts were washed with brine, dried over anhydrous Na₂SO₄, and concentrated under vacuum. The corresponding product **12a** was afforded through silica gel column chromatography (methanol/dichloromethane) in 1.19 g (91% yield, m.p. : 43-47 °C).

¹H NMR (400 MHz, CDCl₃) δ 11.42 (s, 1H), 7.36 (t, *J* = 8.1 Hz, 1H), 7.31 (d, *J* = 6.7 Hz, 1H), 6.68 (d, *J* = 9.1 Hz, 1H), 6.24 (t, *J* = 6.8 Hz, 1H), 3.94 (t, *J* = 7.5 Hz, 2H), 2.33 (t, *J* = 7.5 Hz, 2H), 1.69-1.80 (m, 2H), 1.57-1.68 (m, 2H), 1.29-1.41 (m, 6H).

¹³C NMR (100 MHz, CDCl₃) δ 177.9, 163.0, 139.8, 137.5, 120.7, 106.8, 50.1, 34.1, 29.1, 28.9, 28.8, 26.3, 24.6.

HRMS (ESI+) *m/z* calcd. for C₁₃H₁₉NO₃ [M+H]⁺: 238.1438, found: 238.1443

Synthesis of 10-(pyridin-2-yloxy)decanoic acid (13a)



To a solution of compound **5** (2.02 g, 10.0 mmol) in DMF (10.0 mL), NaH (20.0 mmol, 2.00 equiv.) was added, and the reaction mixture was stirred at room temperature

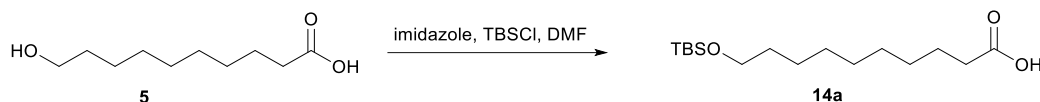
overnight. 2-chloropyridine (1.36 g, 12.0 mmol, 1.20 equiv.) was added, and the reaction mixture was refluxed for 24 h. Upon completion, the solution was quenched by water, and extracted with diethyl ether/hexanes (1:3) three times. The combined extracts were dried over anhydrous Na₂SO₄, and concentrated under vacuum. The corresponding product **13a** was afforded through silica gel column chromatography (ethyl acetate/hexanes) in 2.85 g (78% yield, m.p. : 48-50 °C).

¹H NMR (400 MHz, CDCl₃): δ 8.15 (dd, *J* = 5.1, 1.9 Hz), 1H, 7.60-7.52 (m, 1H), 6.84 (dd, *J* = 7.0, 5.1 Hz, 1H), 6.72 (d, 1H, *J* = 8.3 Hz), 4.26 (t, *J* = 6.7 Hz, 2H), 2.33 (s, 2H), 1.76 (p, *J* = 6.9 Hz, 2H), 1.63 (t, *J* = 7.3 Hz, 2H), 1.43 (q, *J* = 7.1 Hz, 2H), 1.26-1.35 (m, 8H).

¹³C NMR (100 MHz, CDCl₃) δ 179.6, 164.0, 146.8, 138.6, 116.5, 111.1, 66.1, 34.1, 29.30, 29.26, 29.1, 29.02, 29.01, 25.9, 24.7.

HRMS (ESI⁺) *m/z* calcd. for C₁₅H₂₀D₂NO₃⁻ [M+H]⁻: 266.1730, found: 266.1729

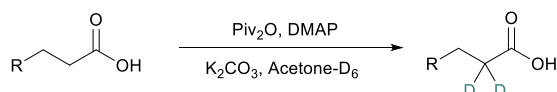
Synthesis of 10-OTBS-decanoic acid (**14a**)



To a solution of TBSCl (5.25 g, 35.0 mmol, 3.50equiv.) in DMF (20.0 mL), imidazole (6.80 g, 100 mmol, 10.0 equiv.) was added at 0 °C. After stirred at 0 °C for 1 h, compound **5** (2.02 g, 10.0 mmol) dissolved in DMF (5.00 mL) was added and stirred for another 4 h. Upon completion, brine was added to the mixture and extracted with diethyl ether/hexanes (1:3), the combined extracts were dried over Na₂SO₄ and concentrated in vacum. The mixture was solubilized with Methanol (150 mL), THF (80.0 mL). K₂CO₃ (4.50 g) in H₂O (50.0 mL) was added to the solution at 0 °C. The solution was stirred for 1 h, then 150 mL brine was added. The reaction mixture was acidified with 1 M HCl until pH reaches ~3, extracted with diethyl ether : hexanes = 1 : 3 three times, dried over Na₂SO₄, and evaporated. The corresponding product **14a** was afforded through silica gel column chromatography (ethyl acetate/hexane) in 2.82 g (93% yield).

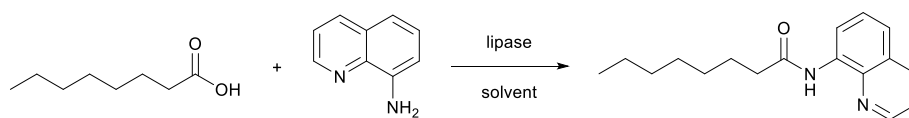
The NMR data was available according to the reported data.²

General procedure for Alpha-deuterated caboxylic acid



According to the known literature,³ we obtained α-deuterated carboxylic acid.

8. Enzymatic synthesis of acyl-quinoline-8-ammonia ^a



Entry	Enzyme	Temp °C	m(E)/(mg / 0.1 mmol)	Solvent	4Å MS (mg)	Yield (%)
1	ROL	60	20	dioxane	0	trace
2	ROL	80	20	toluene	0	trace
3	rugosa	60	20	dioxane	0	NR
4	rugosa	60	20	toluene	0	NR
5	cala	90	20	toluene	0	trace
6	cala	100	20	toluene	0	trace
7	calb	90	20	PBS buffer	0	trace
8	calb	90	20	acetonitrile	0	21
9	calb	90	20	solventless	0	2
10	calb	90	20	toluene	0	80
11	calb	100	20	toluene	0	47
12	calb	85	40	toluene	0	75
13	calb	90	40	toluene	0	83
14	calb	95	40	toluene	0	84
15	calb	90	40	benzene	0	28
16	calb	90	40	chlorobenzene	0	59
17	calb	90	40	DMF	0	0
18	calb	90	40	dioxane	0	78
19	calb	90	40	MTBE	0	75
20	calb	90	40	Benzotrifluoride	0	83
21	calb	90	40	Diphenyl ether	0	34
22	calb	90	40	tetrahydronaphthalene	0	87

23	calb	95	40	toluene	20	89
24	calb	90	40	tetrahydronaphthalene	20	91
25	calb	90	40	tetrahydronaphthalene	40	91
26	calb	90	40	tetrahydronaphthalene	60	88

^a 0.10 mmol acid, lipase and 0.12 mmol 8-AQ was added to 0.50 mL solvent. The reaction mixture was stirred for 24 h. Yield was detected by GC.

9. Complete environmental factor comparison of synthesis of α,β -tetradeuterated palmitic acid.

The **complete E-factor** (cEF) contains all material used for the synthesis, which includes all reagents, inorganics, organic solvents and water. The individual contributions to the cEF (reagents, inorganic, organic solvents and water) were calculated by adding up all material from this category and dividing this quantity by the amount of product generated. This calculation method was pioneered by Sheldon and Kaspar *et al.*⁴, which considers all materials used as:

$$\text{cEF} = \frac{\sum M_{\text{reagents}} + \sum M_{\text{auxillaries}} + \sum M_{\text{solvents}} - M_{\text{product}}}{M_{\text{product}}}$$

Synthesis of $\alpha,\alpha,\beta,\beta$ -tetradeuterated palmitic acid:

1. Present work:

The palmitic acid (5.0 mmol, 1.28 g), K_2CO_3 (0.07 g), Piv_2O (0.19 g), DMAP (0.06 g), acetone- d_6 (4.0 g), 48 h, 40 °C. The reaction mixture with D_2O (2.75 g) was stirred at room temperature for 1 h. To the reaction mixture was added 1 M HCl aq. (2.0 g, 0.5 g) and extracted with CH_2Cl_2 (42 g), and the solvent was removed under reduced pressure. Evaporation was carried out for the removal of pivalic acid.

The acid and 8-aminoquinoline (0.87 g, 6.0 mmol, 1.2 equiv.) were dissolved in 5.0 mL (4.35 g) of toluene, and then $\text{B}(\text{OCH}_2\text{CF}_3)_3$ (0.46 g, 1.5 mmol, 0.3 equiv.) was added to the solution. The mixture was stirred with a Dean-Stark system for 48 h in air. Then acetic anhydride (2.04 g, 2.0 mmol, 0.4 equiv.) was added to the solution. The reaction mixture was stirred at room temperature for 2 h, and deuterium oxide (20 mL, 22.1 g), palladium acetate (0.25 mmol, 5.0 mol%, 0.06 g) and pivalic acid (5.0 mmol, 1.0 equiv., 0.5 g) were added to the solution. The solution was stirred at 90 °C for 24 h in air. Then 2.0 L (2000.00 g) water and 20 mL NovoCor[®] ADL was added in the tube. The mixture was stirred at 50 °C for 48 h in air. The mixture was filtered, and the water was

concentrated to about 200 mL then extracted three times with ethyl acetate (300 g). The combined organic phases were concentrated to about 10 mL, then extracted three times with 1.0 M NaOH aq. (15 g water and 0.6 g NaOH). Then 1.0 M HCl (20 g water and HCl 1.5 g) solution was used to acidify the combined aqueous phases until pH = 2, and then the mixture was extracted three times with DCM (79.60 g). The combined organic phases were dried over anhydrous magnesium sulfate (1.2 g) and concentrated in vacuo to deliver the deuterated carboxylic acids (**0.86 g**).

$$\text{cEF} = (1.28 + 0.19 + 0.06 + 0.87 + 0.46 + 2.04 + 0.06 + 0.5 + 0.5 + 0.07 + 0.6 + 1.5 + 1.2 + 4 + 42 + 4.35 + 300 + 79.60 + 2.75 + 2.0 + 22.1 + 2000 + 15 + 20 - 0.86) / 0.86 = 6.3 + 4.5 + 500 + 2397.5 - 1 = \mathbf{2907.3}$$

with contributions from reagents (6.3), inorganic (4.5), organic solvents (500) and water (2397.5)

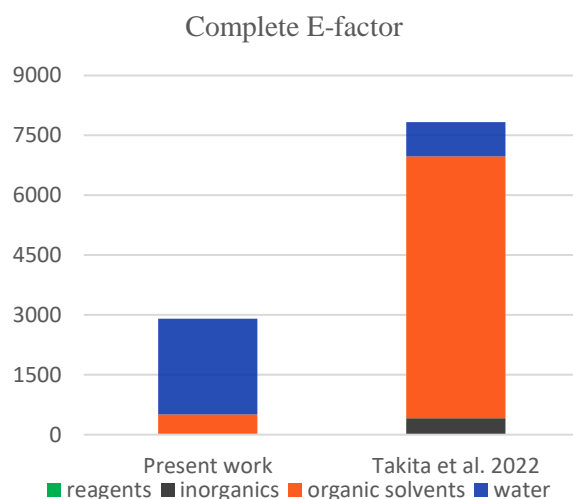
Takita's method (*Angew. Chem. Int. Ed.*, **2022**, *61*, e202202779)

Palmitic acid (1.28 g), 8-aminoquinoline (0.72 g), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide monohydrochloride (WSCD•HCl, 1.25 g), DMAP (0.61 g), and CH₂Cl₂ (70 g) were added to a Schlenk tube under an argon atmosphere at room temperature. After stirring overnight, the reaction was quenched with aqueous NH₄Cl (50 g, 3.6 g) and washed with brine (45 g, 16.2 g). The organic layer was dried over Na₂SO₄ (1.2 g), filtered, and evaporated under reduced pressure. The product was purified by silica gel (10 g) flash column chromatography (500 g). Amide, palladium acetate (0.045 g), and cesium pivalate (92 mg) were added to a 10-mL screw-cap tube under air. Toluene (43.6 g) was added to dissolve the amide, and then D₂O (5.5 g) was added to the mixture. The reaction was stirred at 80 °C for 18 h, and the progress was monitored by ESI-MS. The reaction mixture was cooled to room temperature, transferred to a separation funnel, diluted with EtOAc (50 g), and washed with brine (150 g, 54 g). The organic layer was dried over Na₂SO₄ (1.2 g), filtered, and evaporated under reduced pressure. The crude β-deuterated material was roughly purified by silica gel (10 g) flash column chromatography (500 g). This material, potassium carbonate (0.63 g), and methanol-d₁ (18 g) were added to a screw-cap tube under air. The reaction was stirred at 80 °C for 18 h, and the progress was monitored by ESI-MS. The reaction mixture was cooled to room temperature, transferred to a separation funnel, diluted with EtOAc (40 g), and washed with brine (150 g, 54 g). The organic layer was dried over Na₂SO₄ (1.2 g), filtered, and evaporated under reduced pressure. The obtained material was purified by silica gel (10 g) flash column chromatography (500 g) to afford amide-d₄. Amide-d₄, nickel bis(2,2,6,6-tetramethyl-3,5-heptanedionate) (0.4 g), and MeOH-d₁ (CH₃OD, 47 g) were added to a screw cap tube with N₂ purging. The reaction was stirred at 100 °C for 16 h, and the progress was monitored by TLC and ESI-MS. The reaction mixture was transferred to a separation funnel, diluted with EtOAc (100 g), and washed with brine. The organic layer was dried over Na₂SO₄ (1.2 g), filtered, and evaporated under reduced pressure. The crude material was purified by silica gel (10 g) flash column chromatography (500 g) to afford the ester-d₄. The obtained ester-d₄, THF

(28.5 g), and methanol-d₁ (CH₃OD, 13.7 g) were added to a screw-cap tube with N₂ purging. After cooling to 0 °C, NaOD/D₂O solution (40 wt.%, 1.2 g, 0.48 g) was added to the reaction mixture. The mixture was stirred at 0 °C for 5 min and then at room temperature for 2 h. The reaction progress was monitored by TLC. The reaction was quenched with 1 N HCl (10 g, 0.24 g), diluted with diethyl ether (285 g), and separated. The organic layer was dried over Na₂SO₄ (8.0 g), filtered, and evaporated under reduced pressure. The obtained material was purified by flash column chromatography (10 g, 500 g) to afford the corresponding acid-d₄ in 38% yield (0.48 g).

$$cEF = (1.28 + 0.72 + 1.25 + 0.61 + 0.045 + 0.4 + 3.6 + 16.2 + 1.2 + 10 + 54 + 1.2 + 10 + 0.63 + 54 + 1.2 + 10 + 1.2 + 10 + 0.48 + 0.24 + 8 + 10 + 70 + 500 + 50 + 500 + 18 + 40 + 500 + 47 + 100 + 500 + 28.5 + 13.7 + 285 + 500 + 45 + 50 + 5.5 + 150 + 150 + 1.2 + 10 - 0.48) / 0.48 = 9 + 400 + 6567 + 858 - 1 = 7833$$

with contributions from reagents (9), inorganic (400), organic solvents (6567) and water (858)

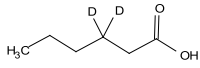


Reference

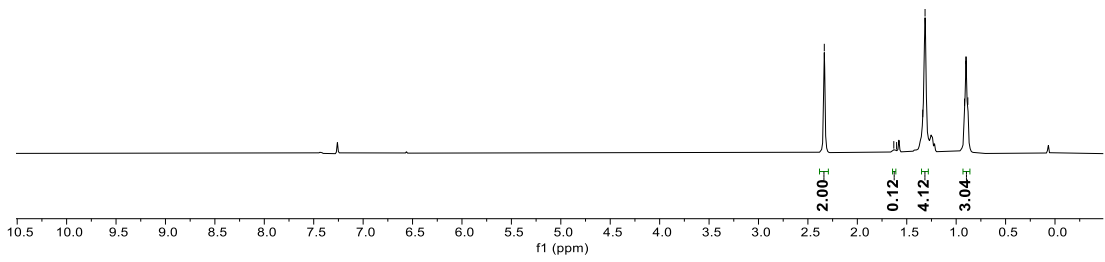
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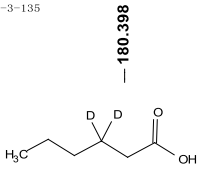
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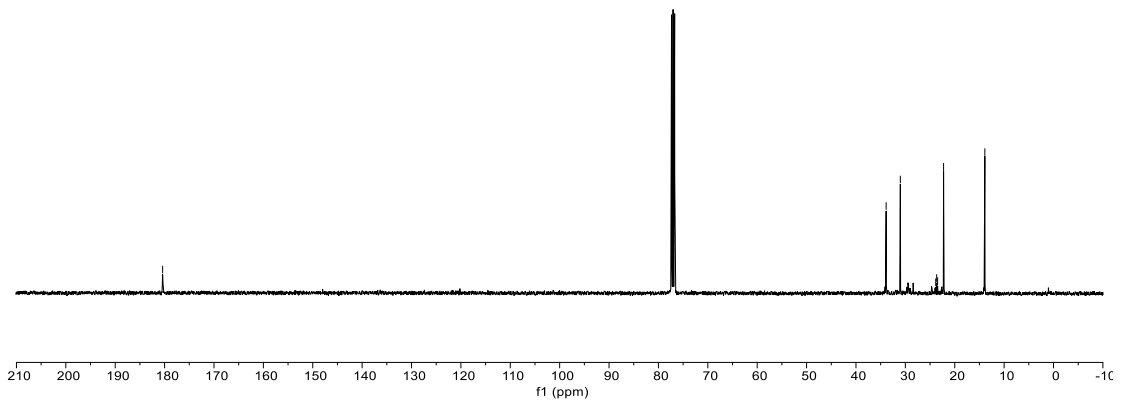
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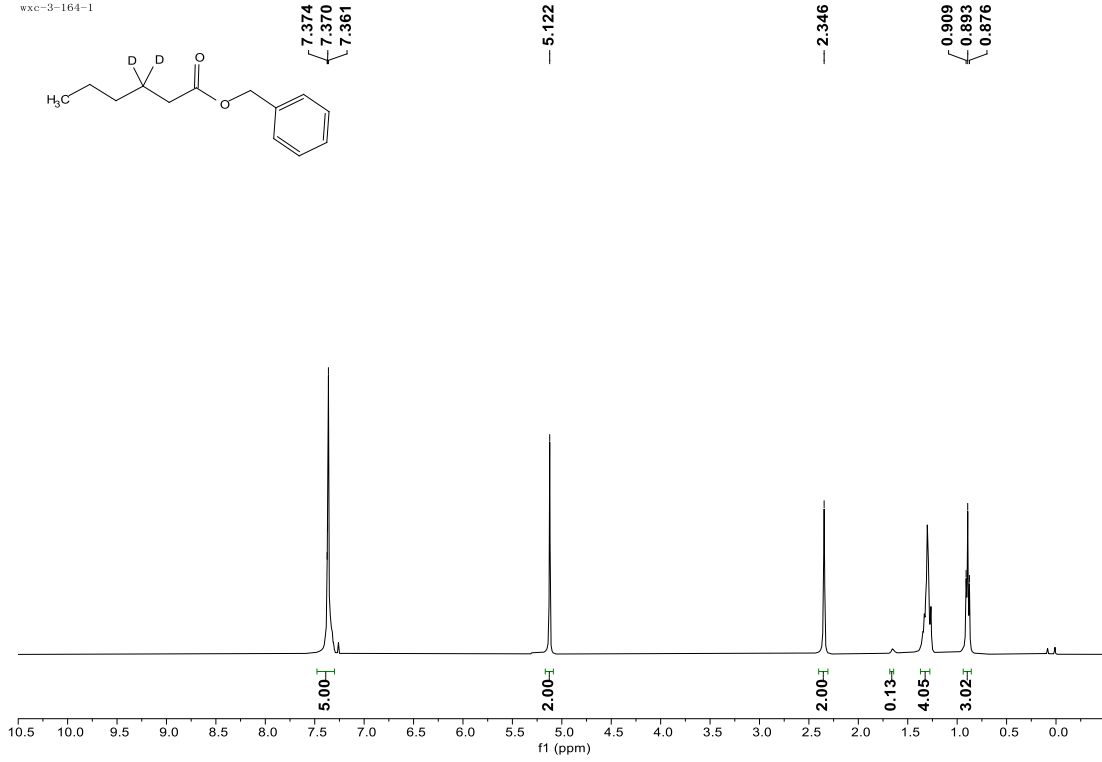
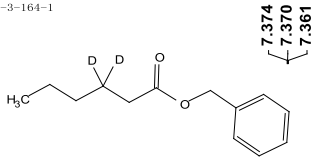
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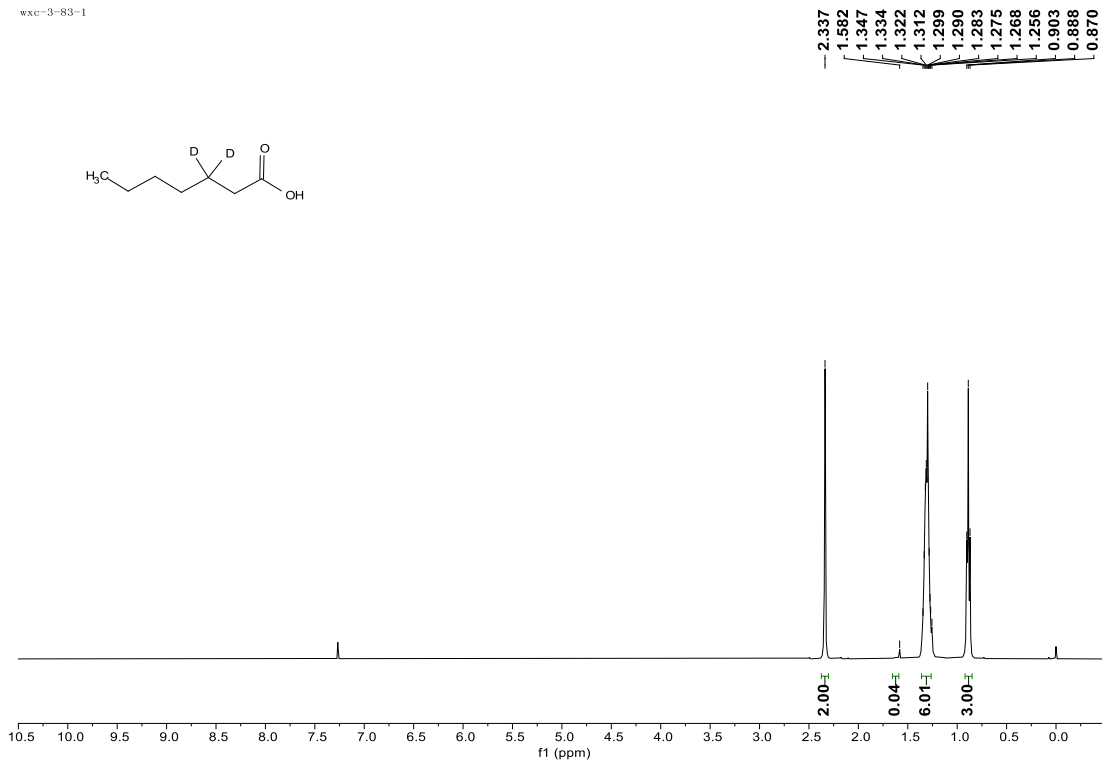
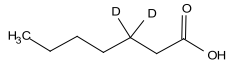
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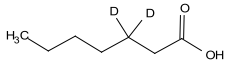


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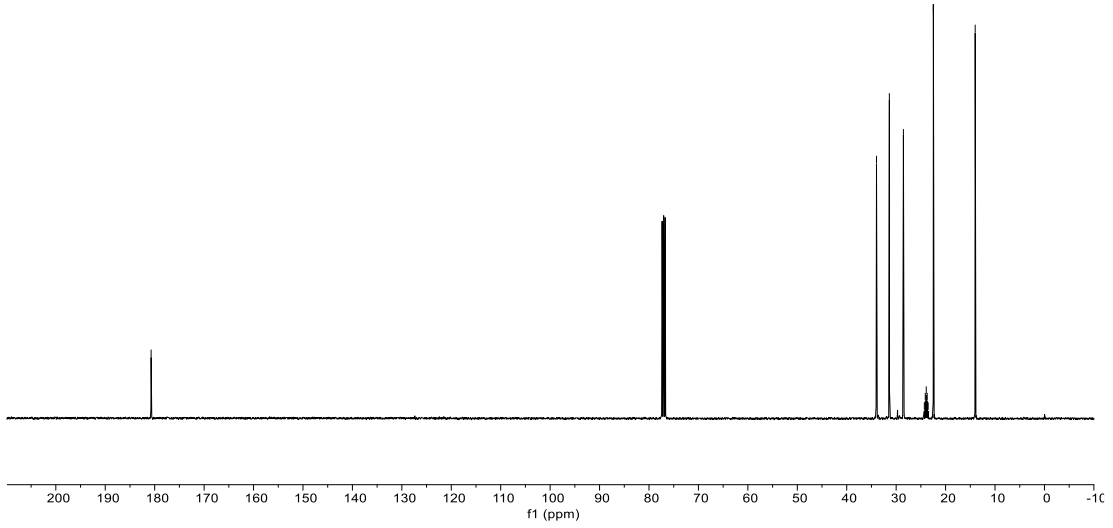


wxc-3-83-1

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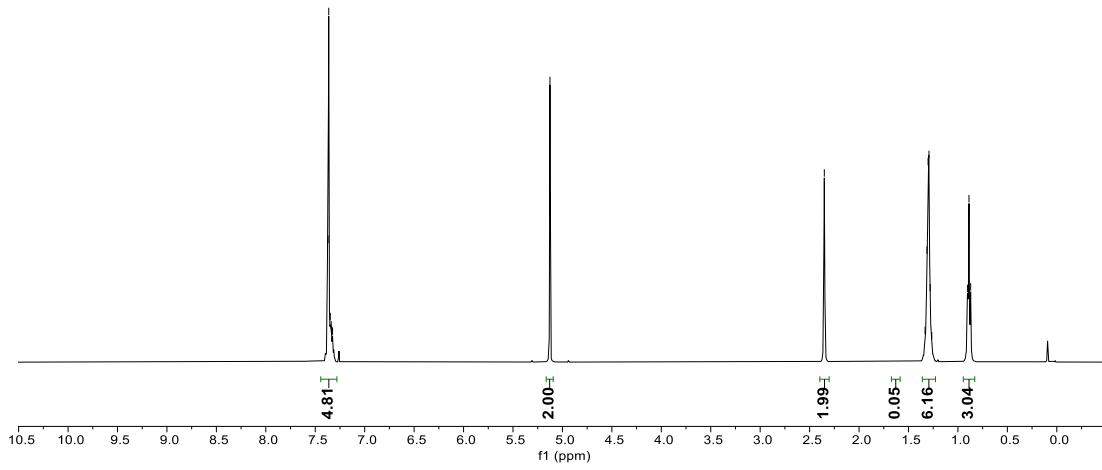
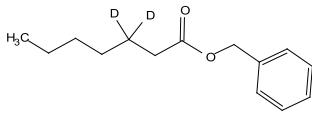
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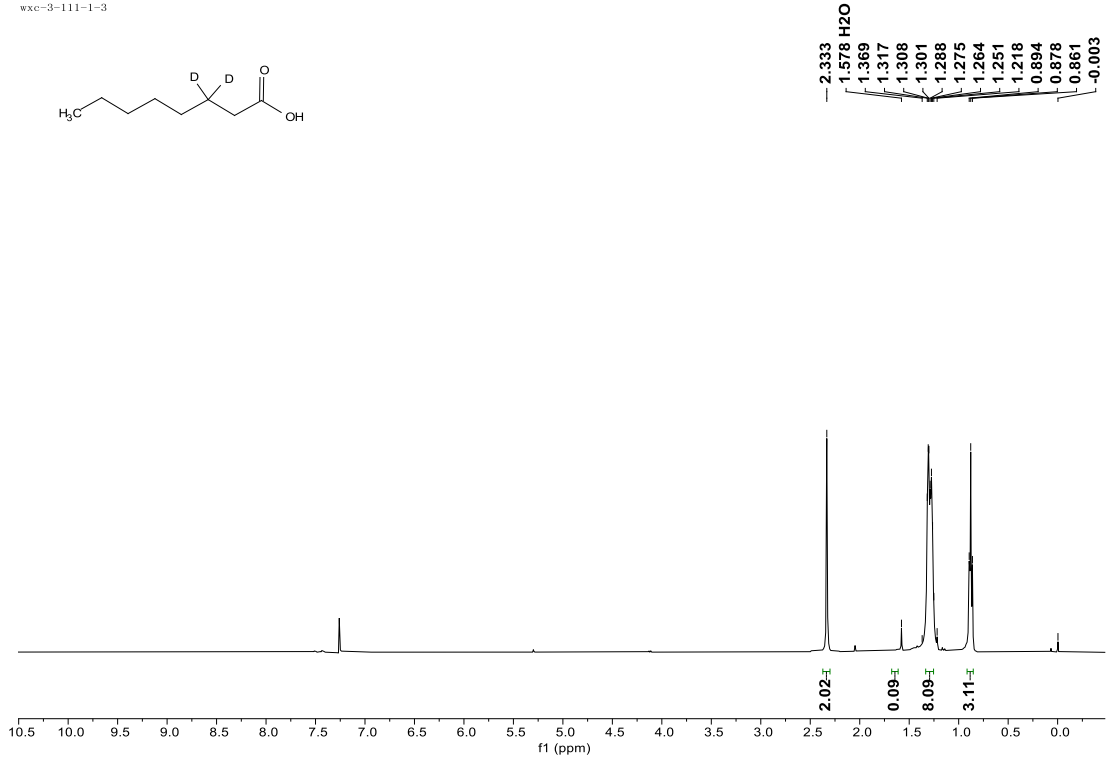
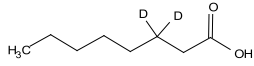
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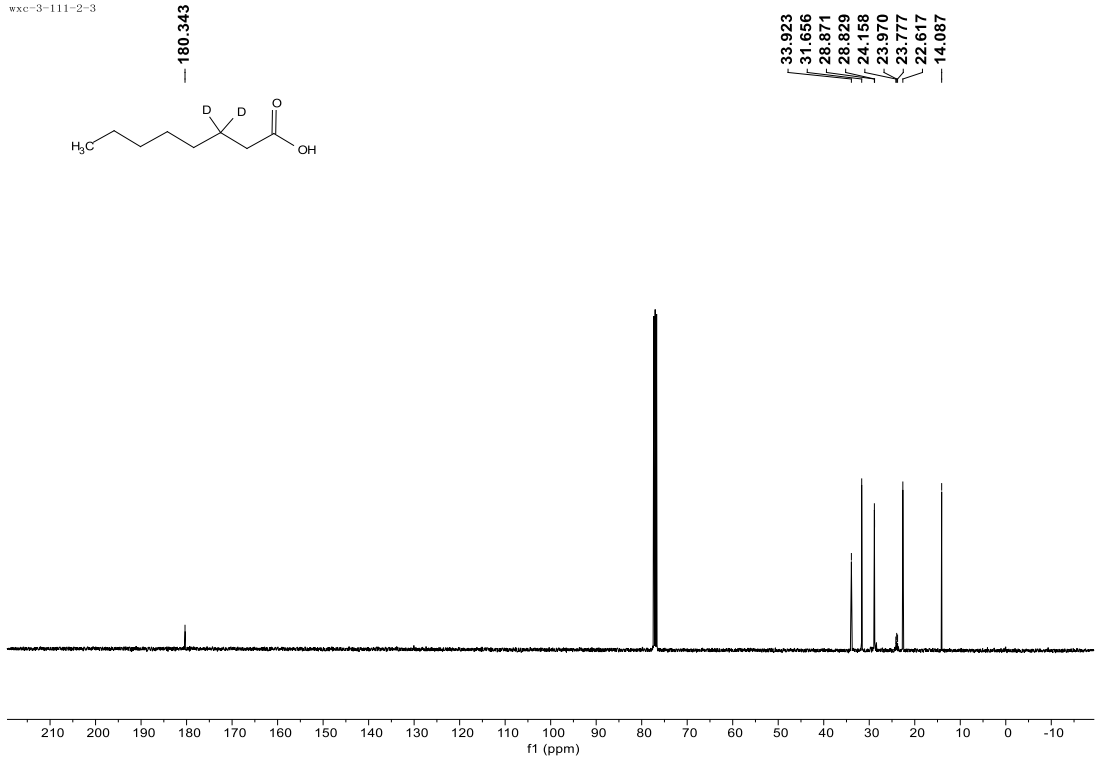
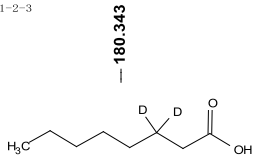
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0.871



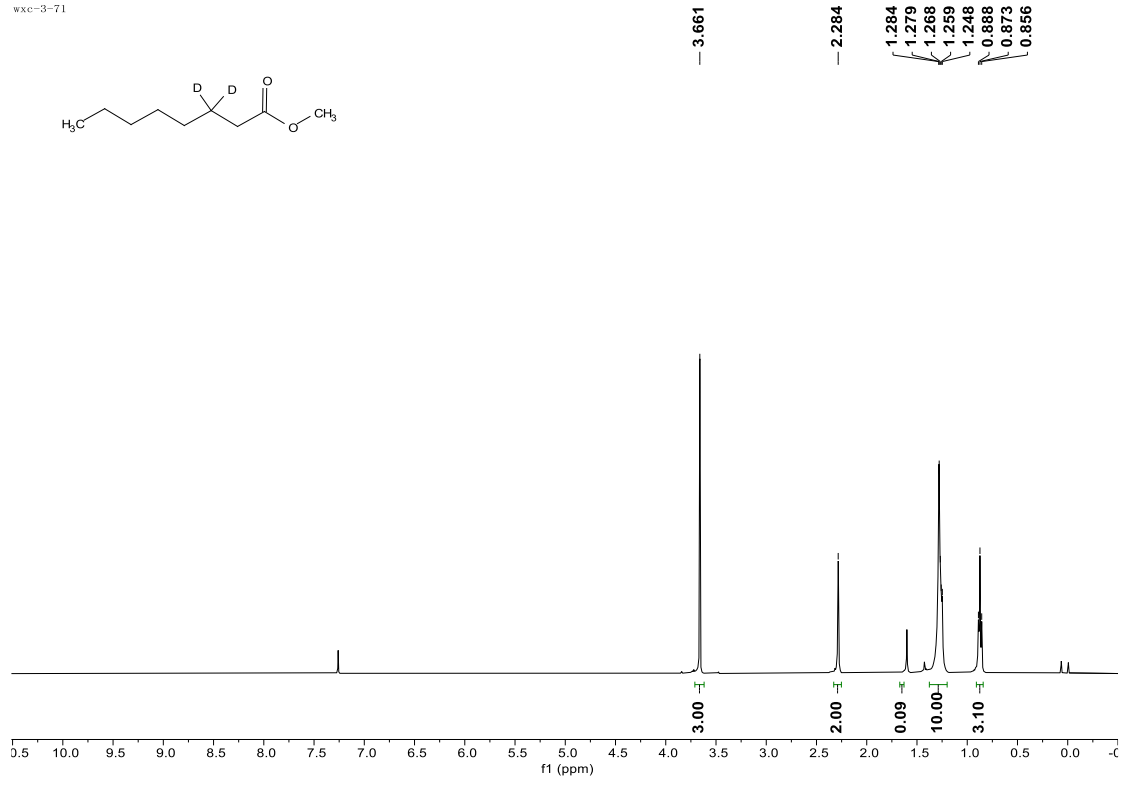
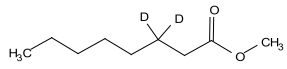
wxc-3-111-1-3



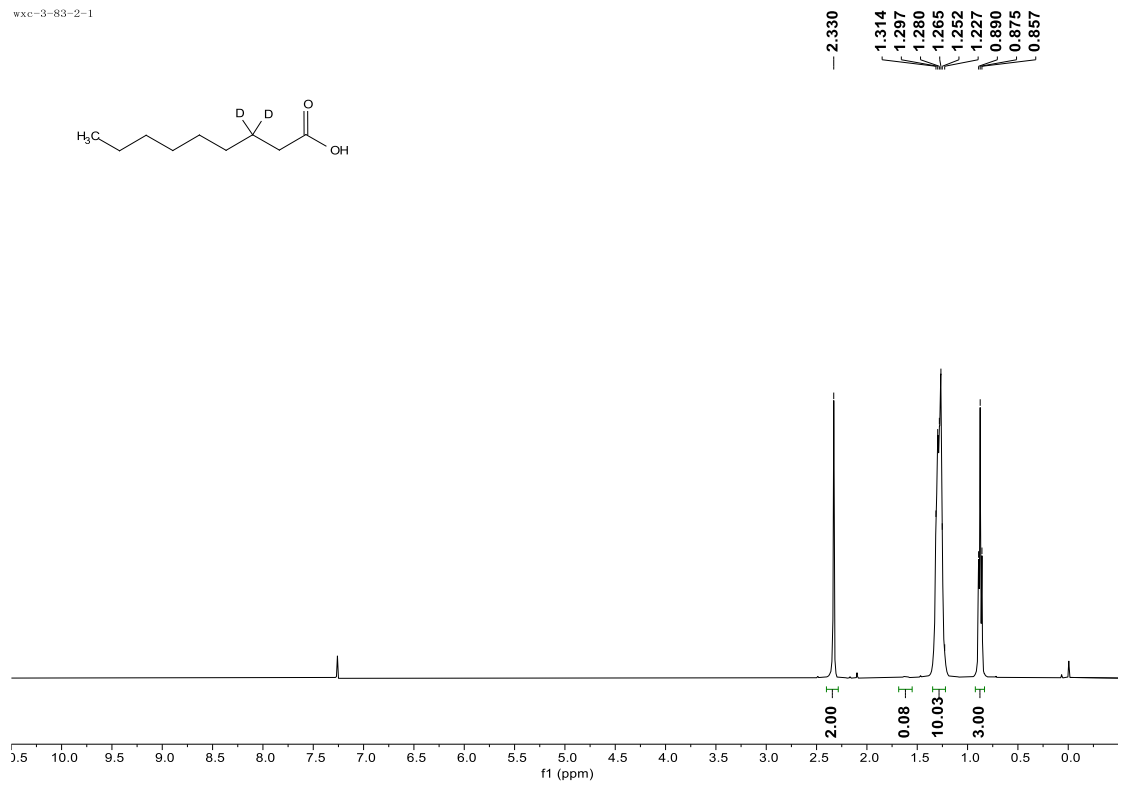
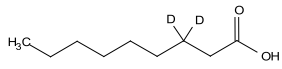
wxc-3-111-2-3



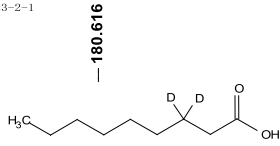
wxc-3-71



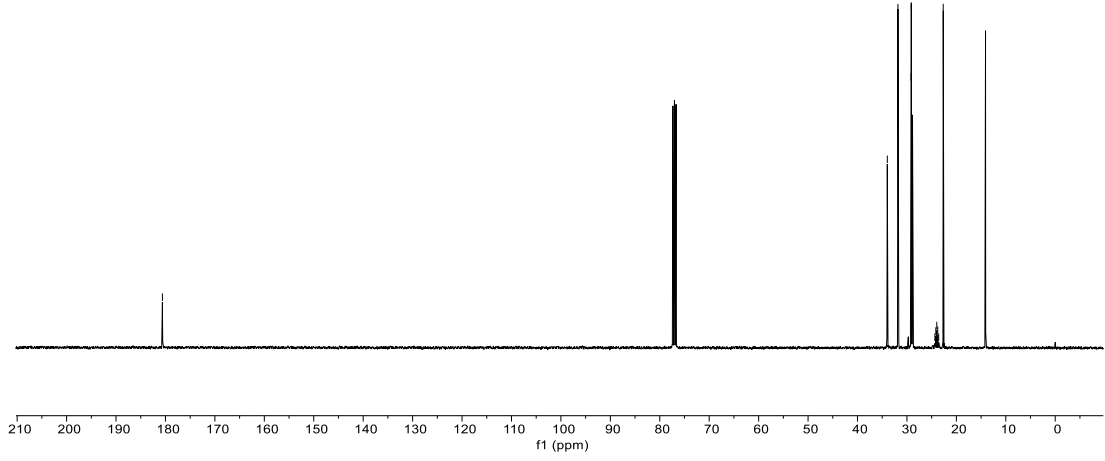
wxc-3-83-2-1



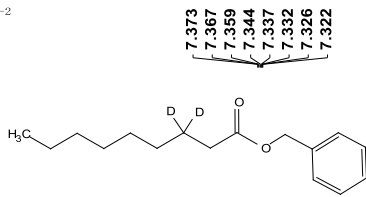
wxc-3-83-2-1



33.968
31.815
29.162
29.118
28.871
24.343
24.155
23.959
23.766
23.570
22.654
14.097



wxc-3-96-2

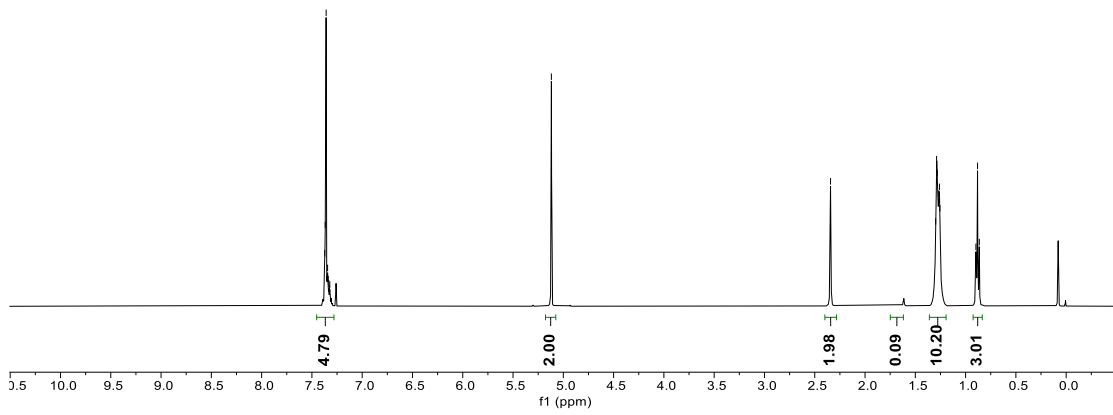


7.373
7.367
7.359
7.344
7.337
7.332
7.326
7.322

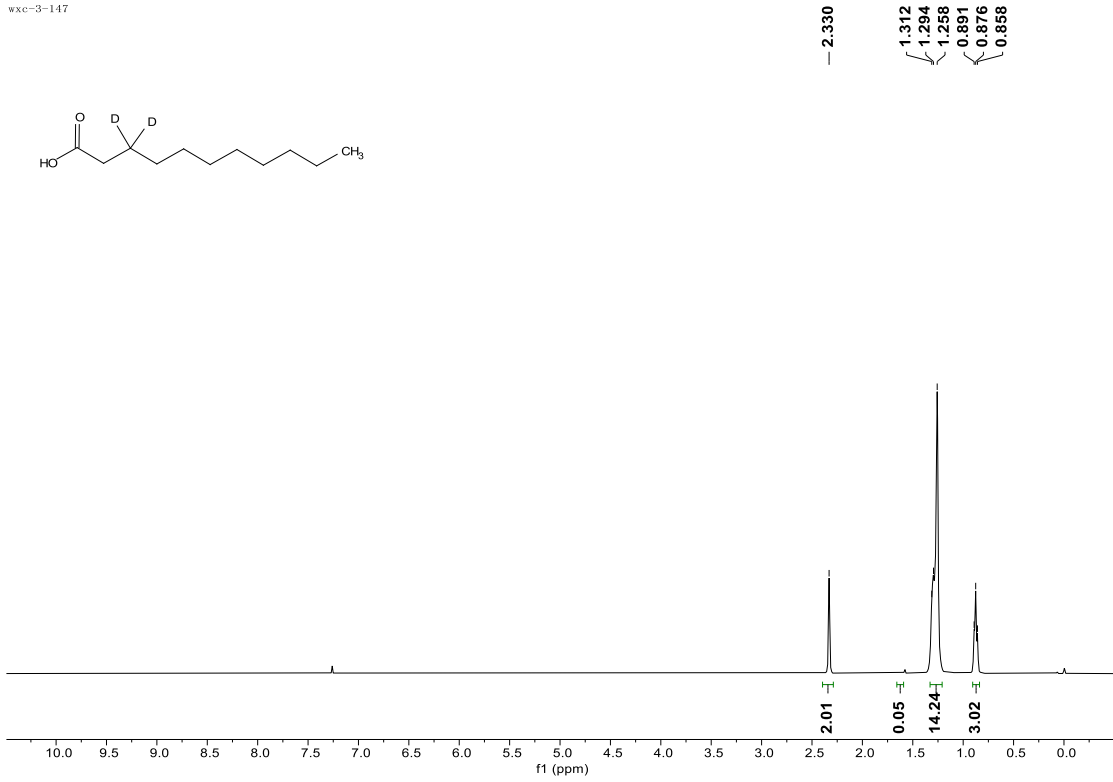
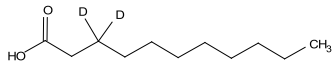
5.119

2.343

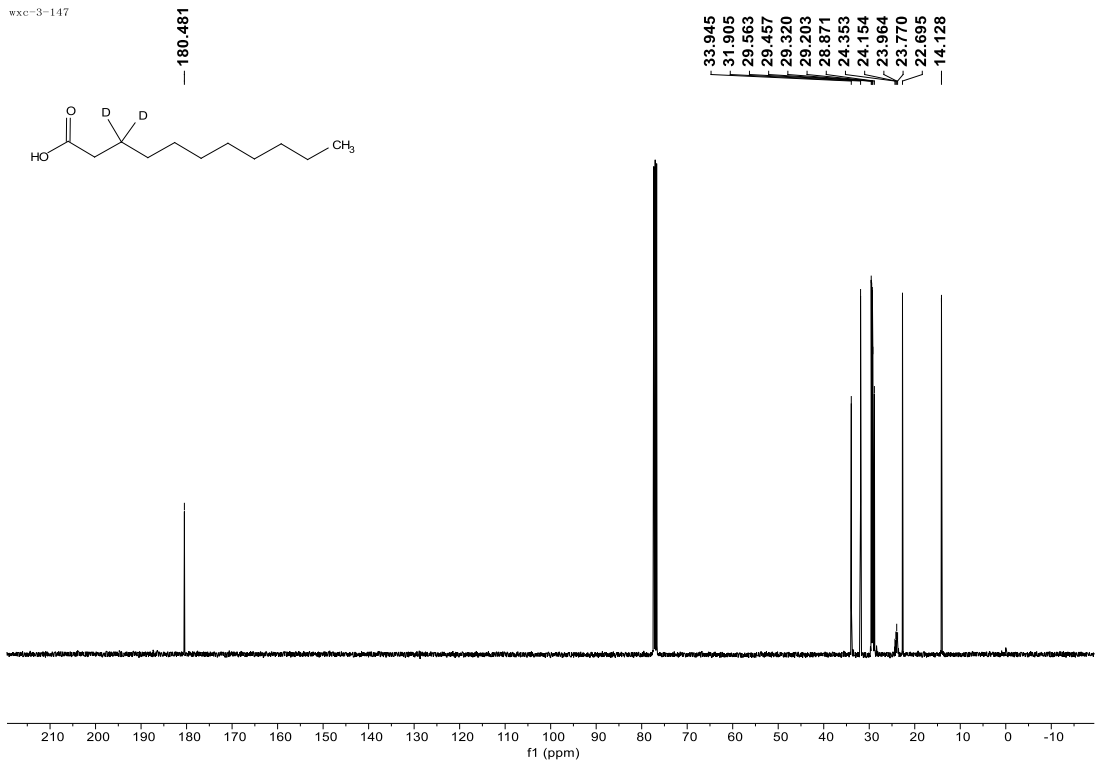
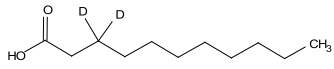
1.297
1.289
1.281
1.268
1.260
1.255
0.898
0.881
0.864



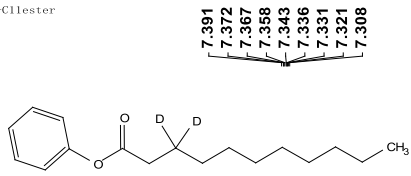
wxc-3-147



wxc-3-147



wxc-Cllester

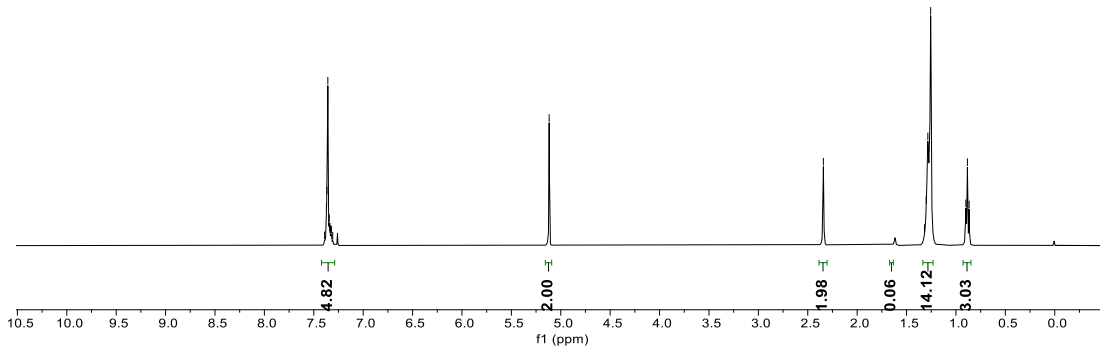


7.391
7.372
7.367
7.356
7.343
7.336
7.331
7.321
7.308

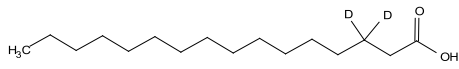
5.118

2.342

1.318
1.301
1.284
1.256
0.900
0.884
0.866

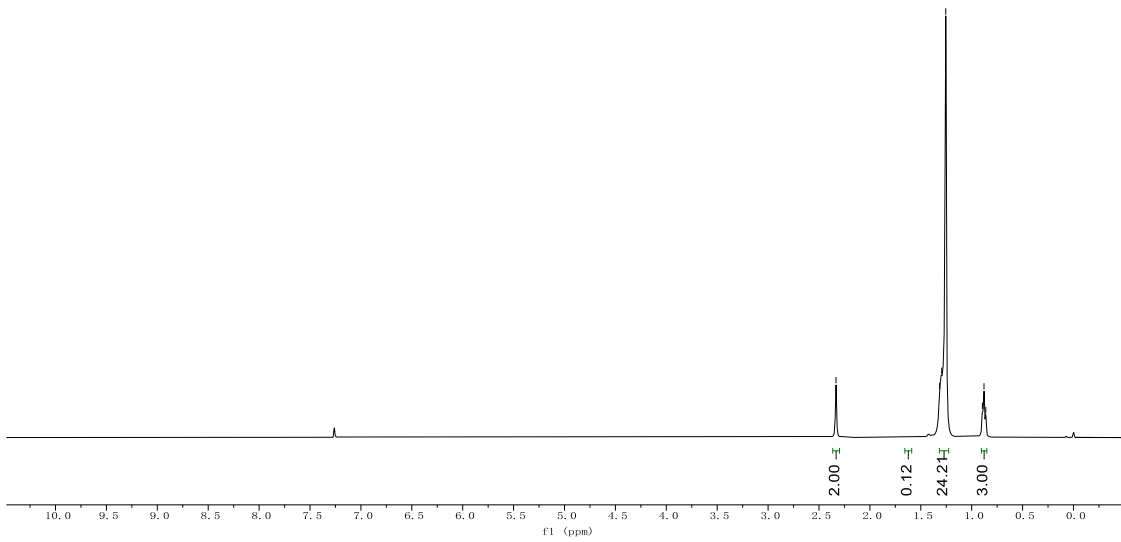


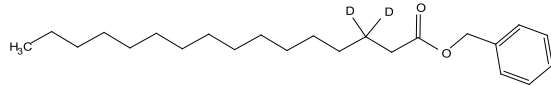
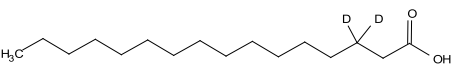
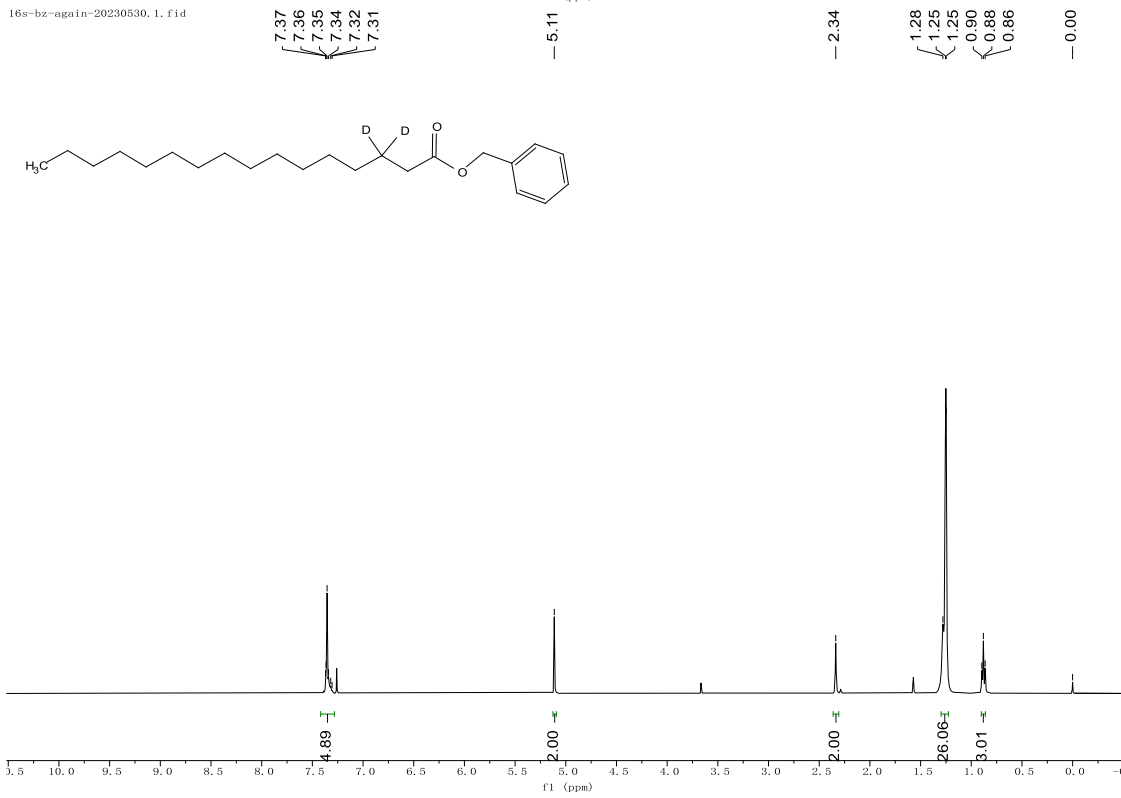
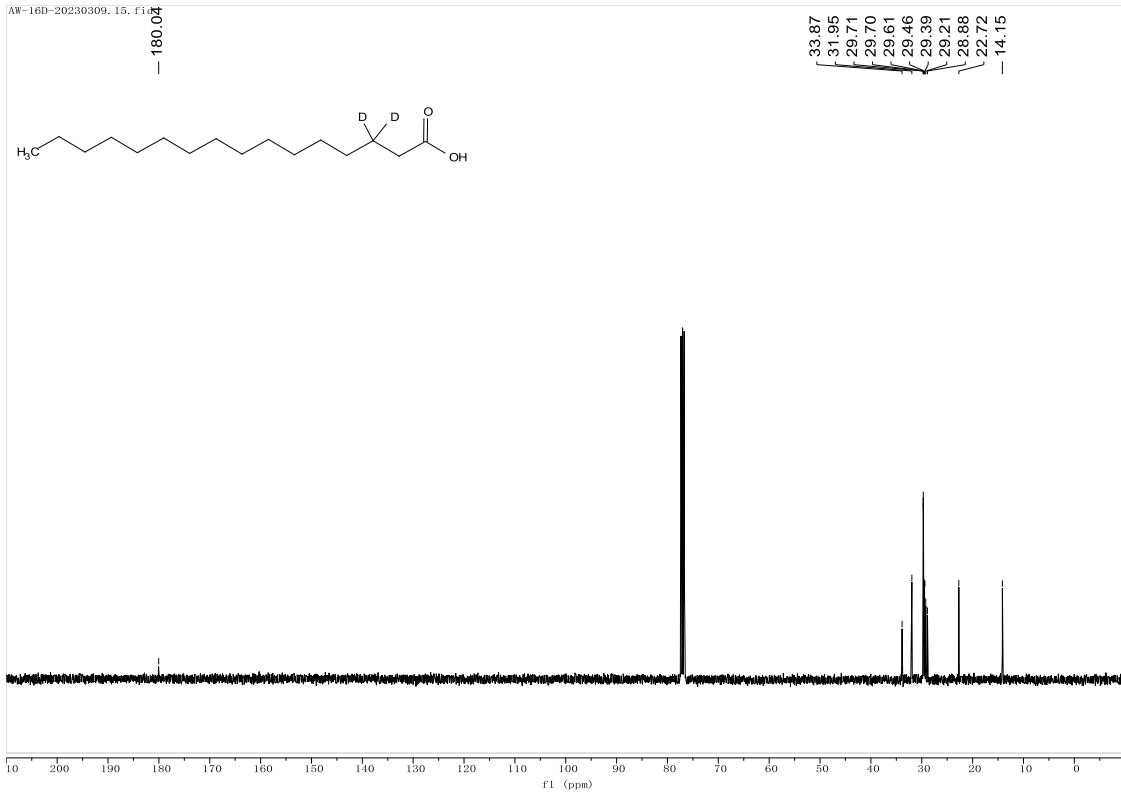
AW-16D-20230309_14.fid



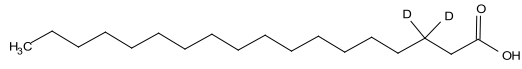
2.33

1.31
1.29
1.25
0.89
0.88
0.86

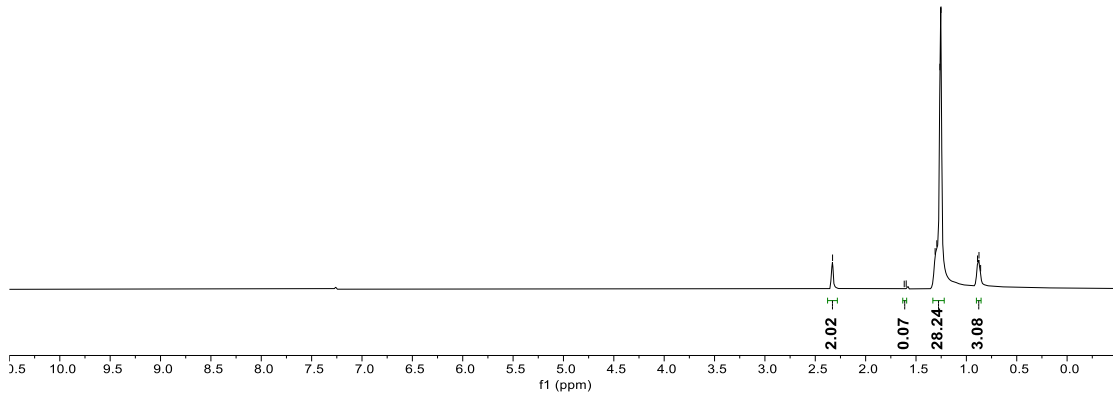




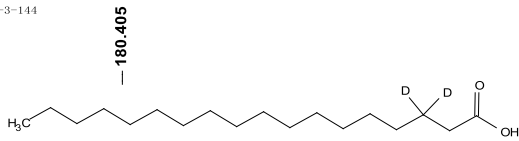
wxc-3-144



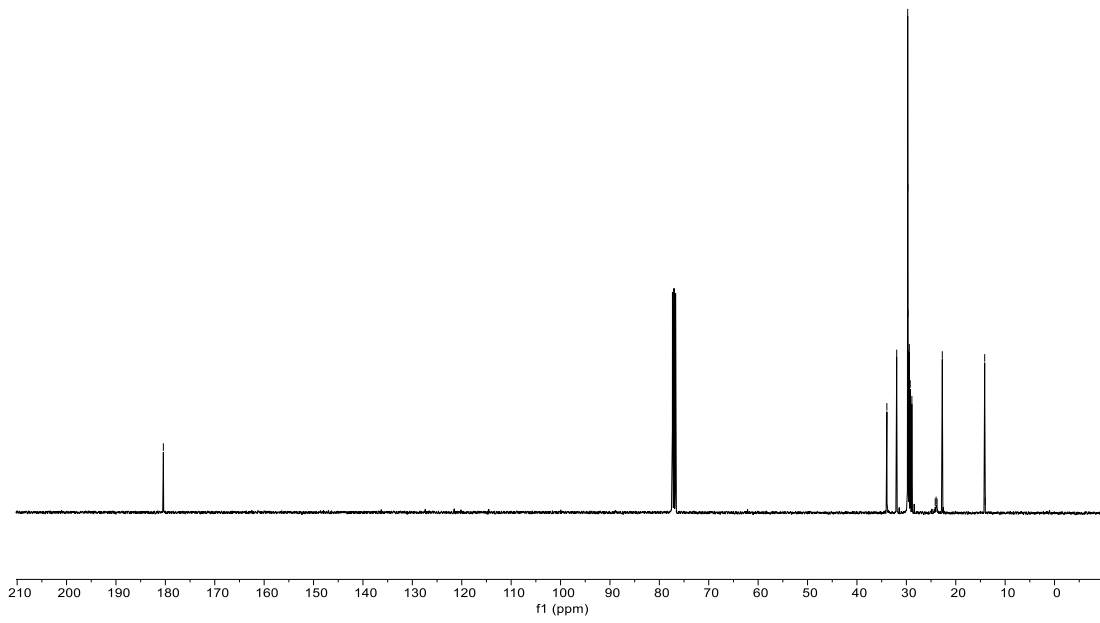
2.328
1.617
1.597
1.310
1.293
1.263
1.252
0.889
0.875
0.859



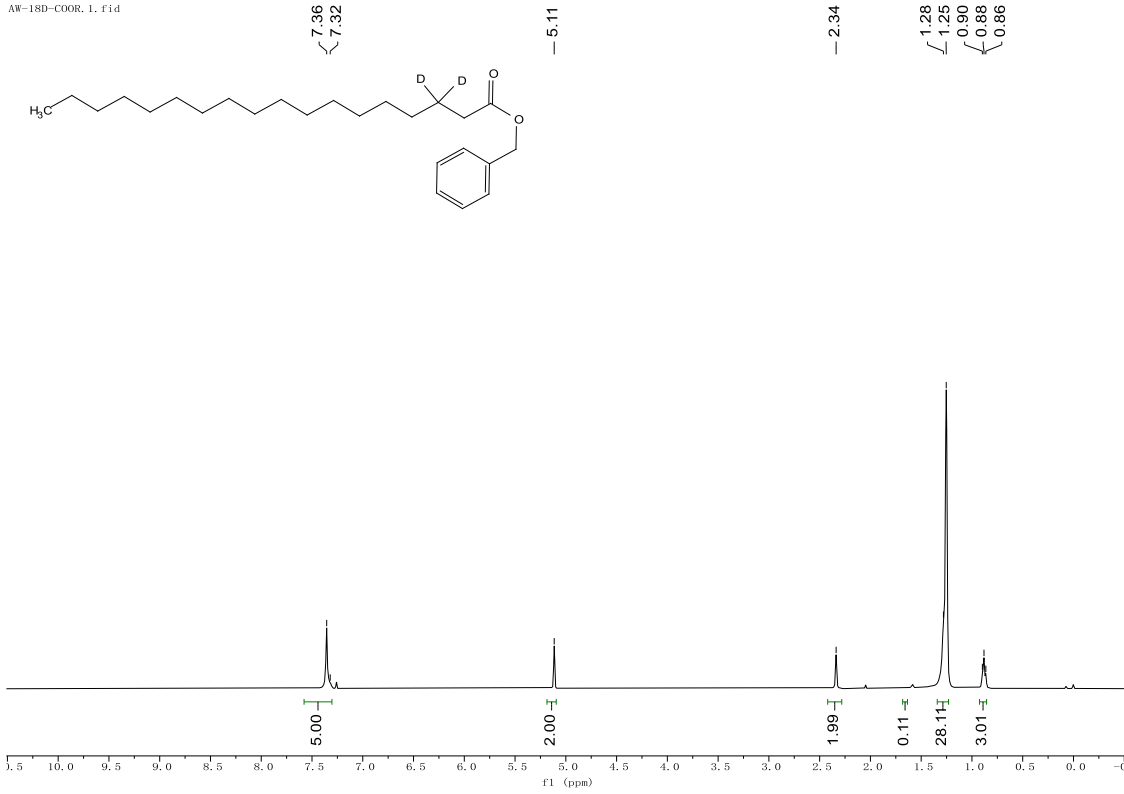
wxc-3-144



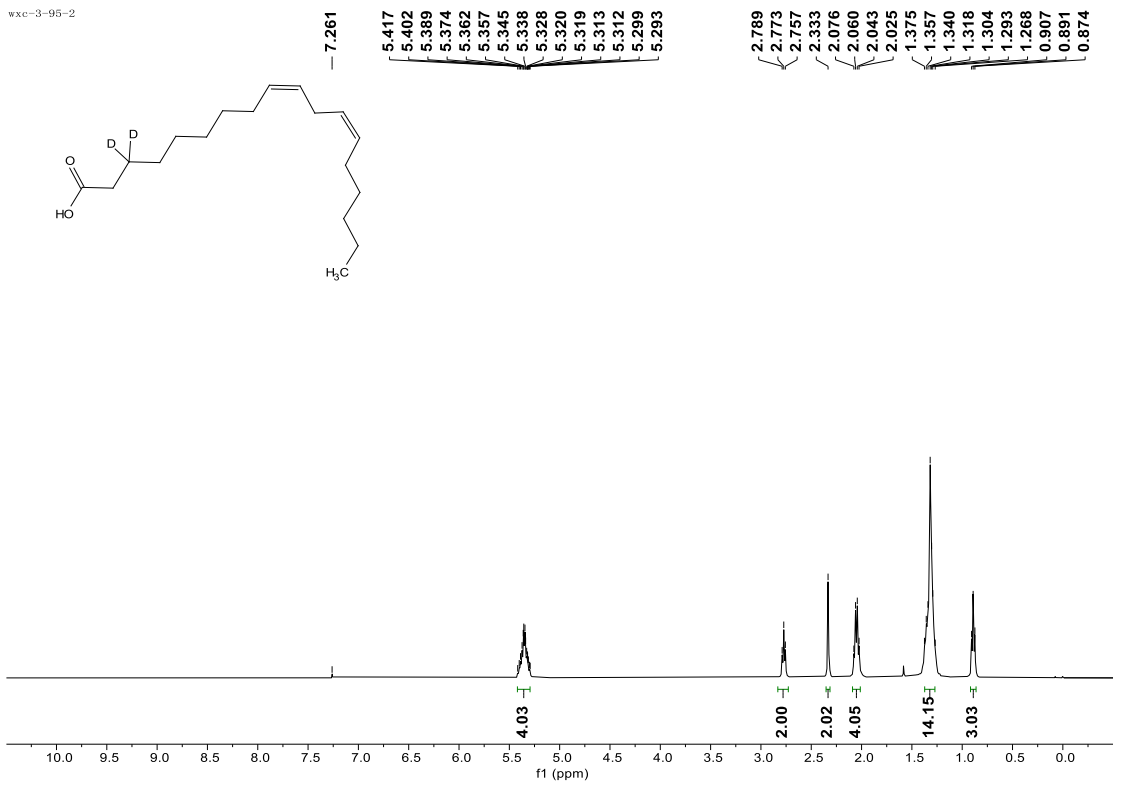
180.405
33.941
31.954
29.723
29.692
29.668
29.619
29.468
29.395
29.215
28.882
24.156
23.970
23.778
22.720
14.142
14.033



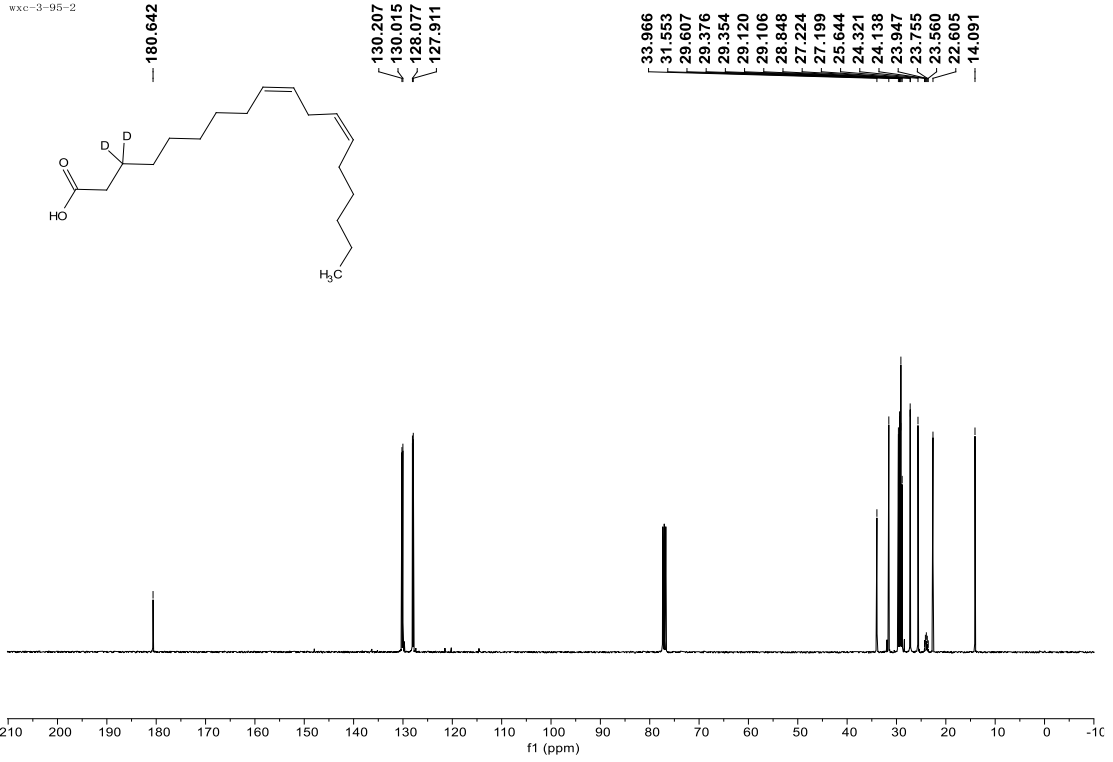
AW-18D-COOR, 1.fid



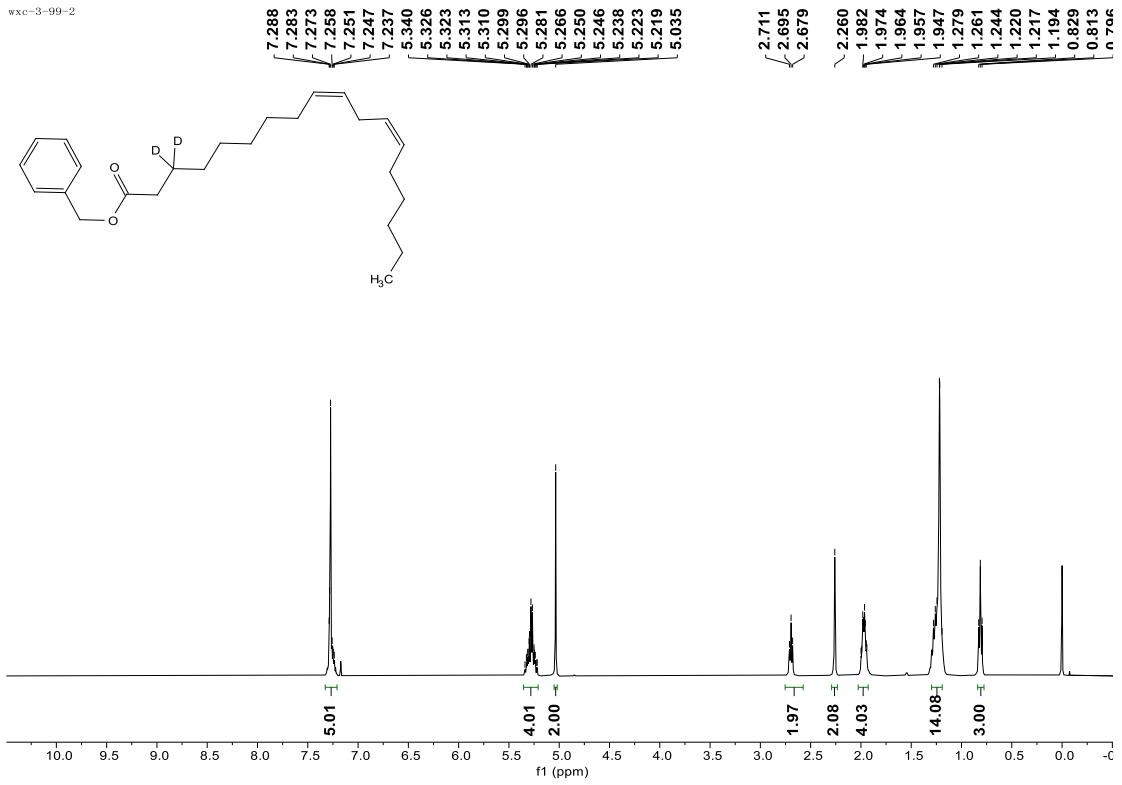
wxc-3-95-2



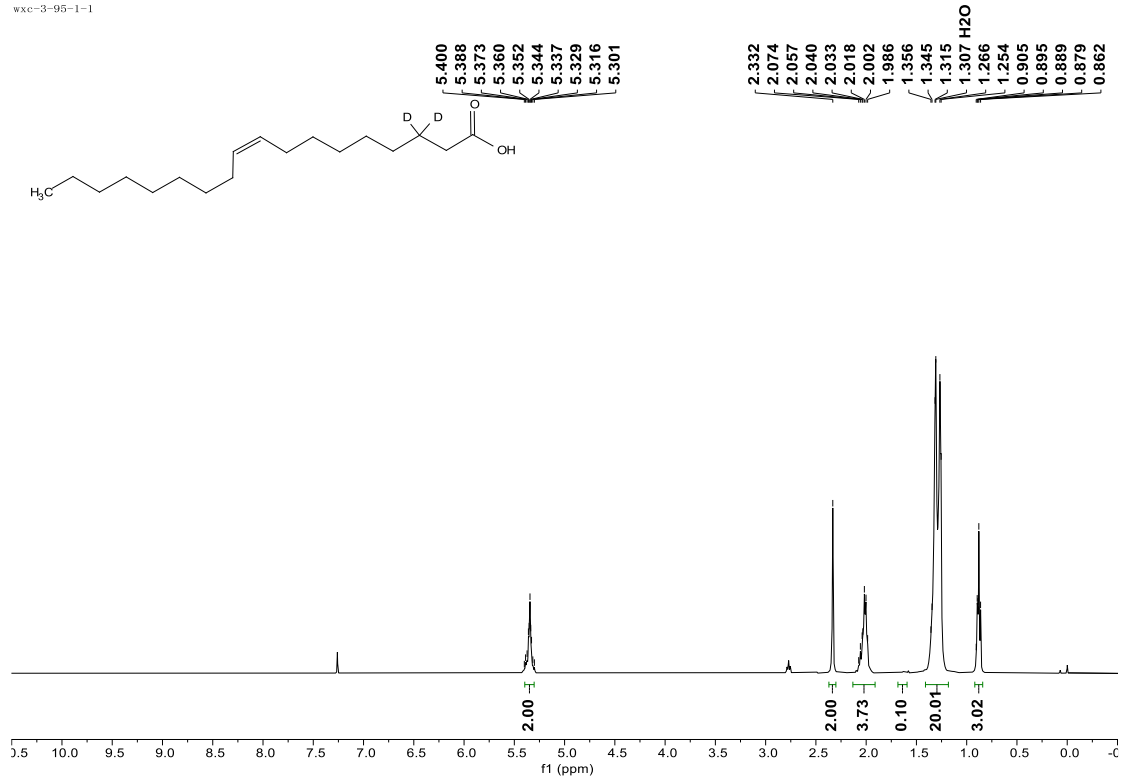
wxc-3-95-2



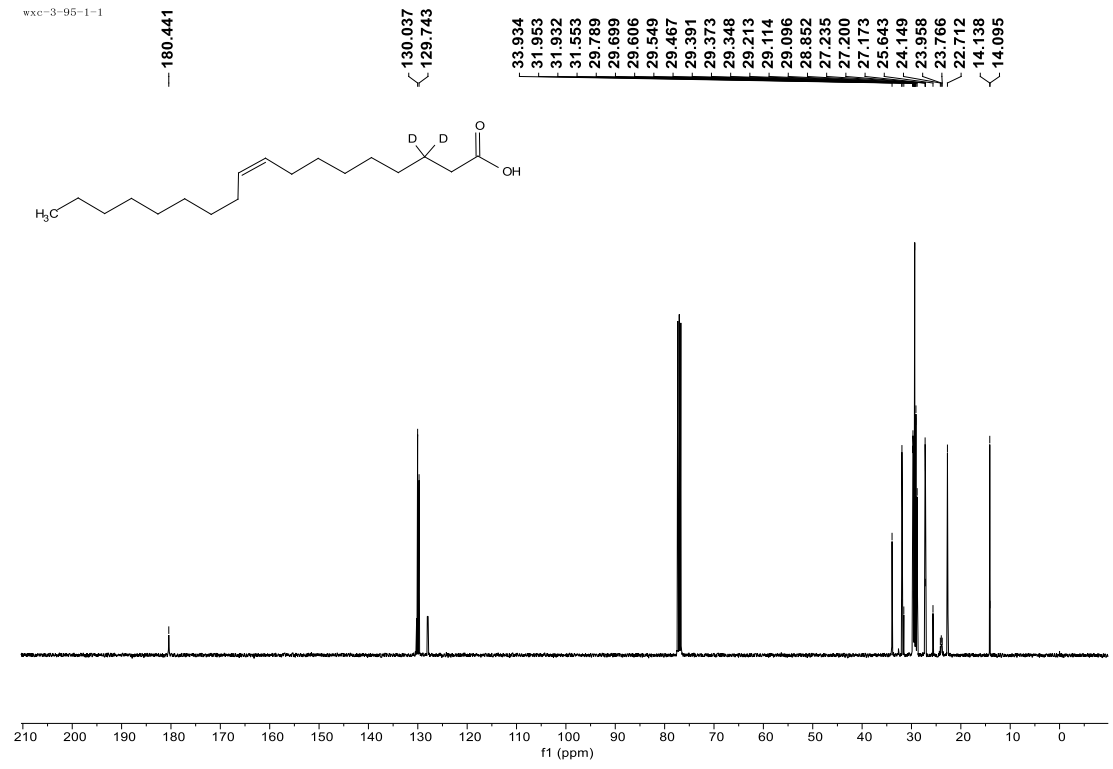
wxc-3-99-2



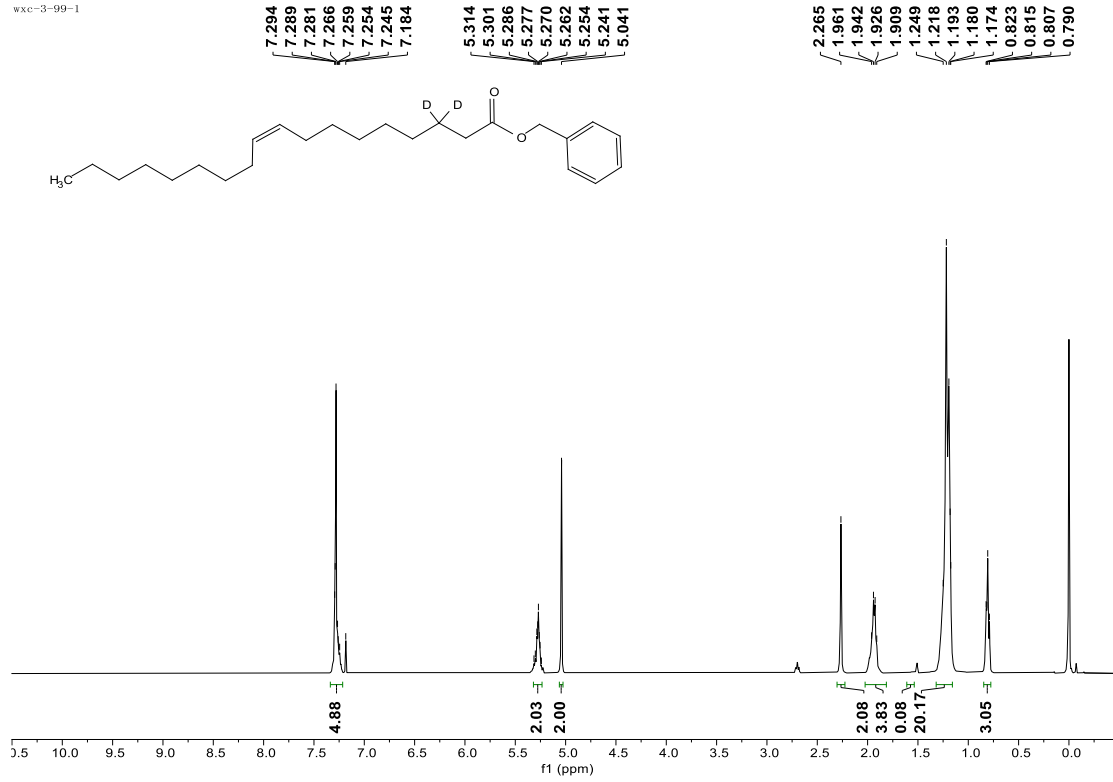
wxc-3-95-1-1



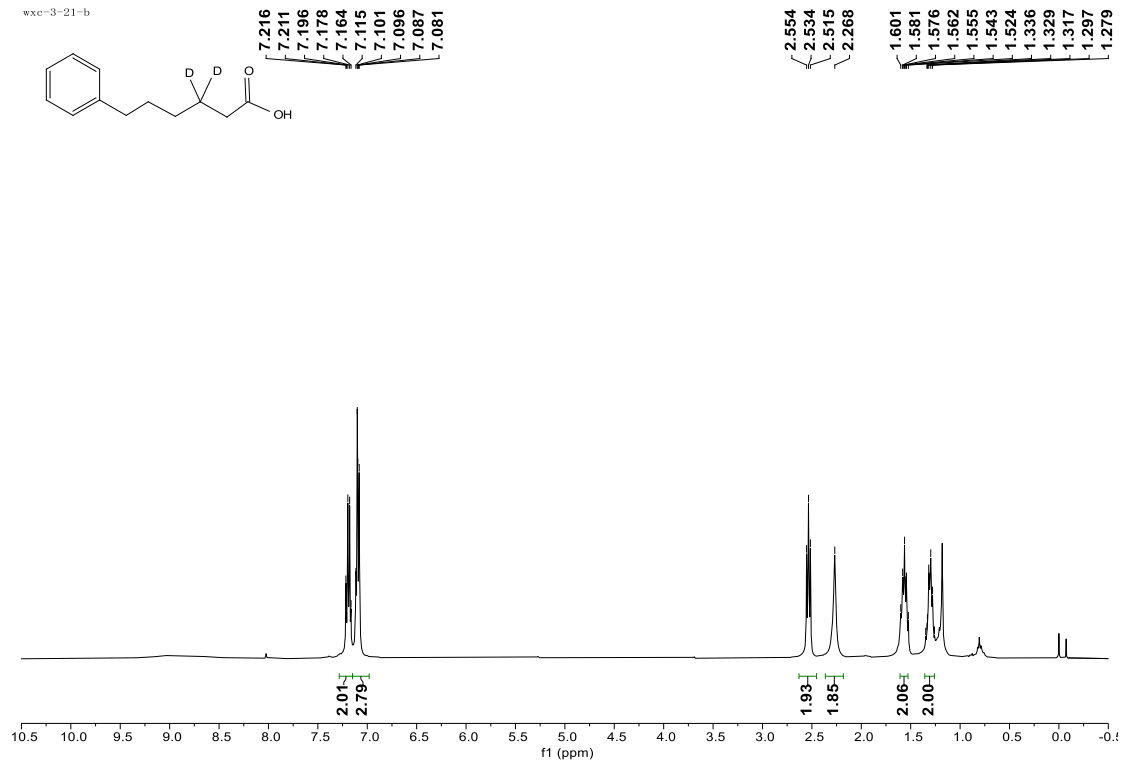
wxc-3-95-1-1



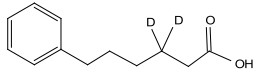
wxc-3-99-1



wxc-3-21-b



wxc-3-21-b



180.416

142.482

128.420

128.322

125.731

35.738

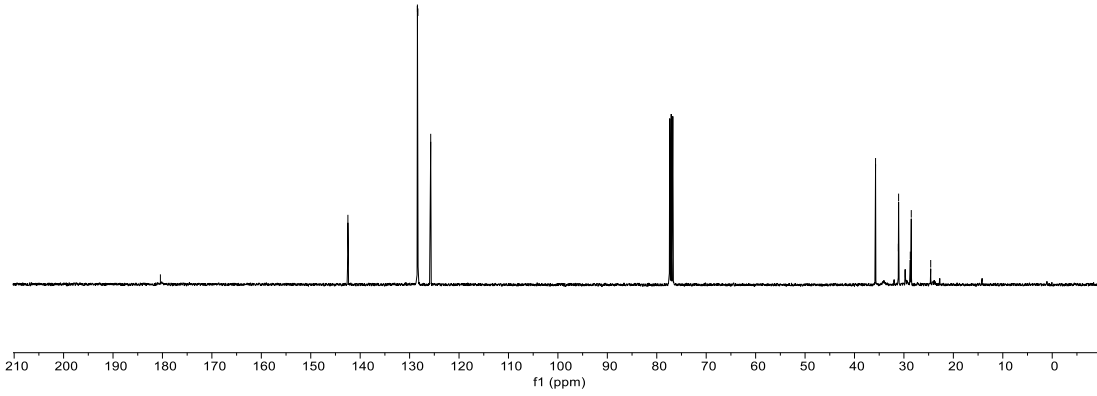
31.124

31.075

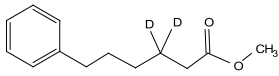
28.712

28.514

24.583



wxc-3-79



7.282

7.265

7.248

7.183

7.166

7.150

3.651

2.618

2.603

2.583

2.283

1.663

1.648

1.629

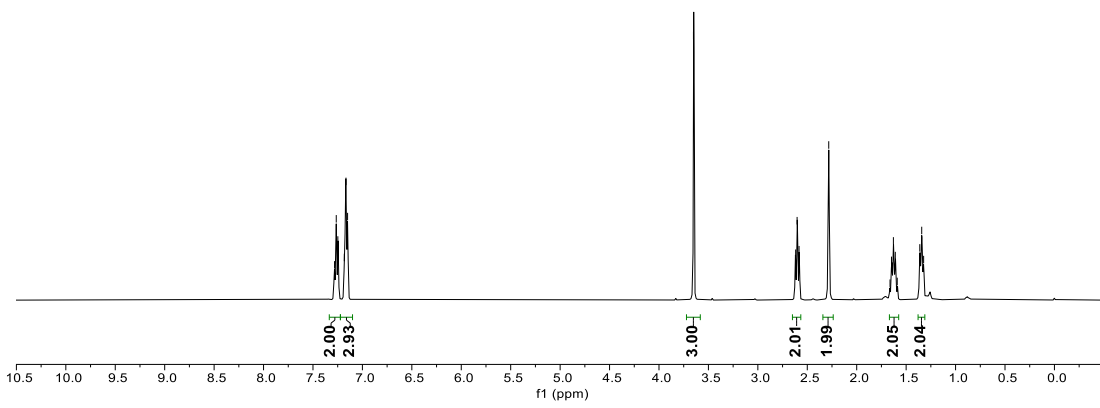
1.610

1.590

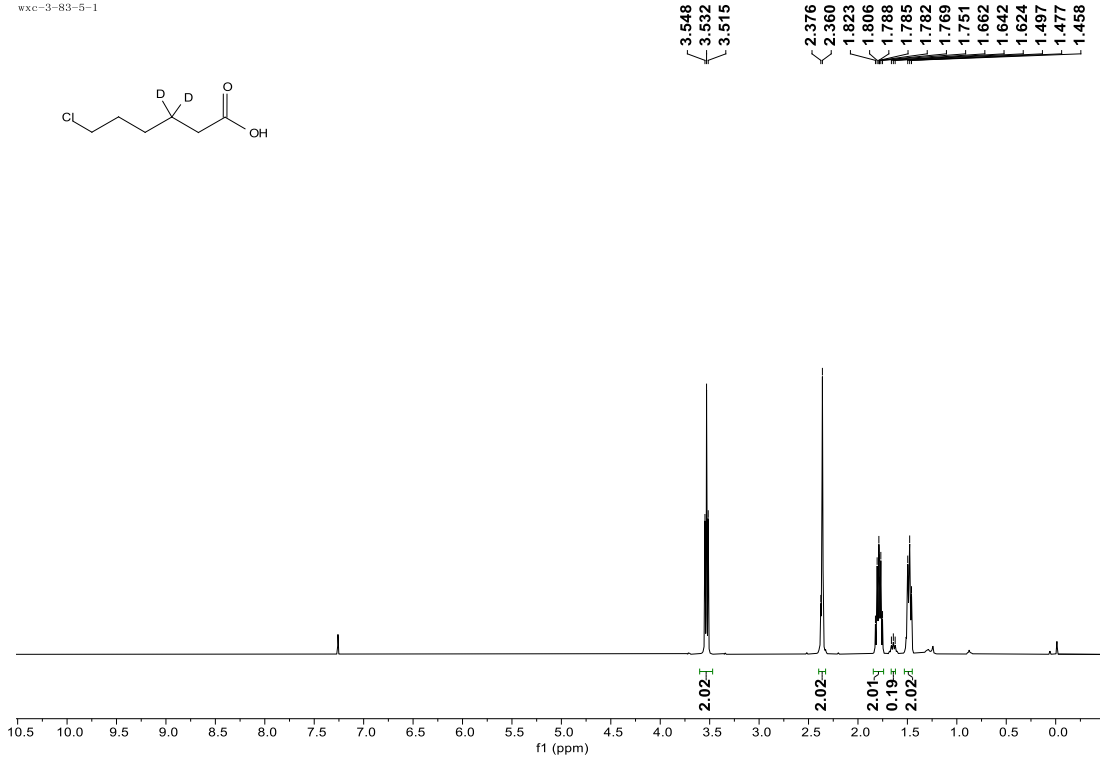
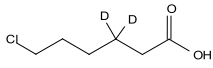
1.362

1.342

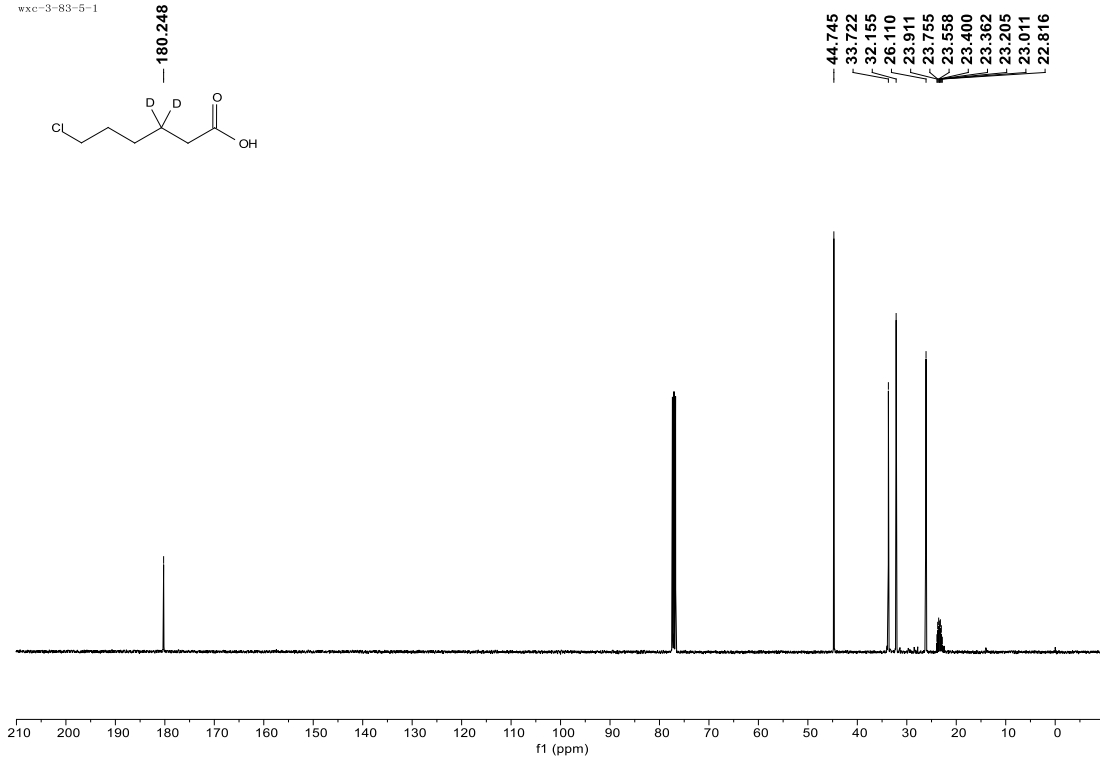
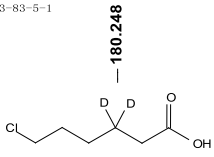
1.323



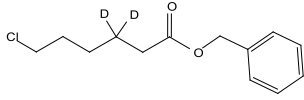
wxc-3-83-5-1



wxc-3-83-5-1



wxc-3-96-3

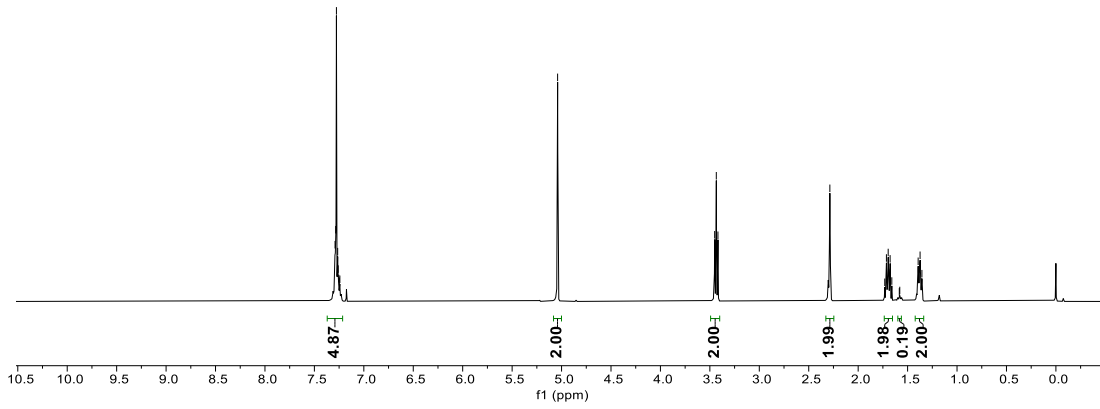


7.291
7.285
7.277
7.264
7.259
7.247
7.242

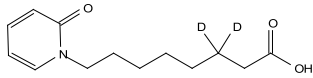
5.040

3.451
3.435
3.418

2.286
1.731
1.714
1.696
1.690
1.677
1.660
1.584
1.579
1.393
1.374
1.355



wxc-3-155-3.21.fid

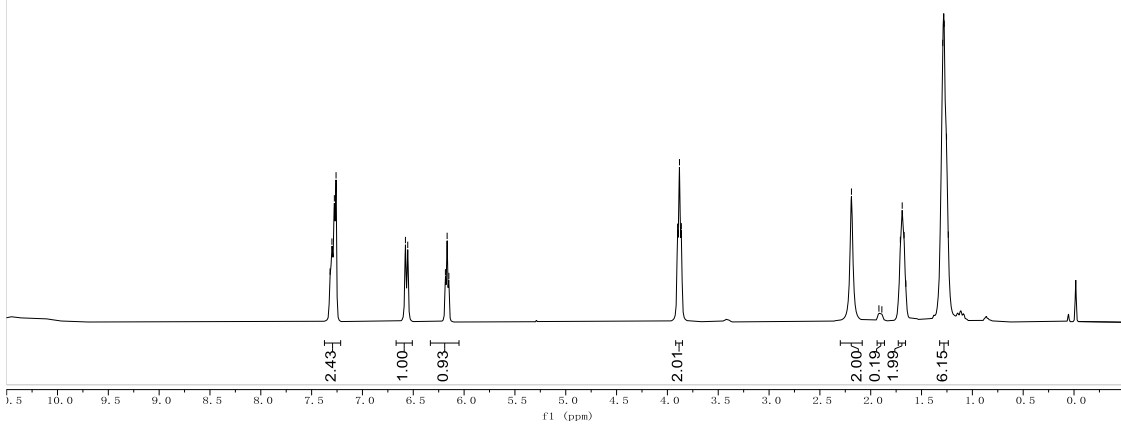


7.32
7.30
7.28
7.26

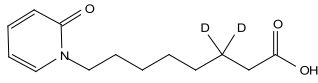
6.58
6.55
6.18
6.17
6.15

3.90
3.88
3.86

2.19
1.92
1.89
1.71
1.69
1.67
1.65
1.29
1.28
1.25
1.24



wxc-3-155-3



181.163

162.810

139.500

137.627

120.951

106.345

49.953

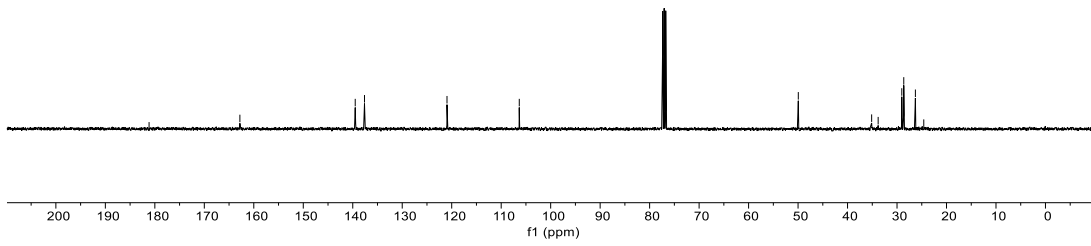
35.131

33.813

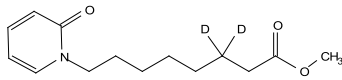
29.038

26.638

24.594



wxc-3-164-2



7.253

7.246

7.243

7.237

7.231

7.221

7.214

7.196

7.182

7.176

7.165

7.160

6.502

6.479

6.080

6.077

3.859

3.841

3.822

3.594

2.214

1.948

1.931

1.887

1.670

1.651

1.296

1.286

1.275

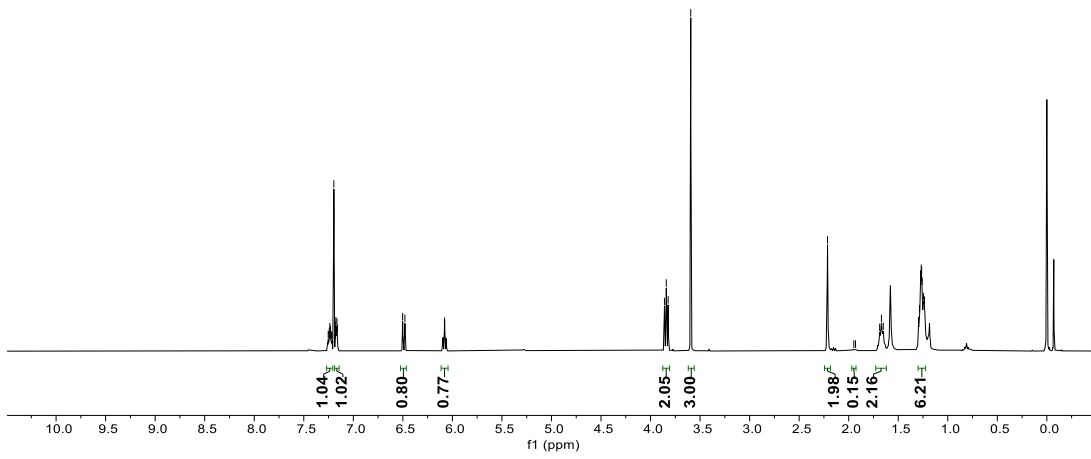
1.268

1.264

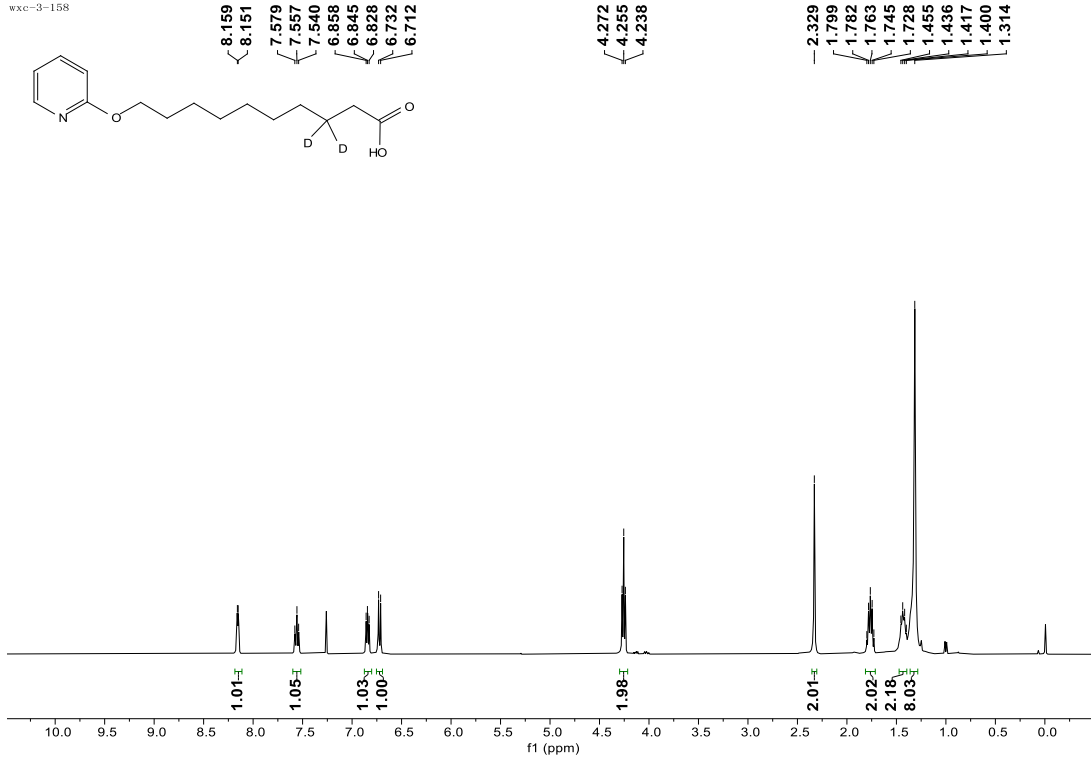
1.258

1.246

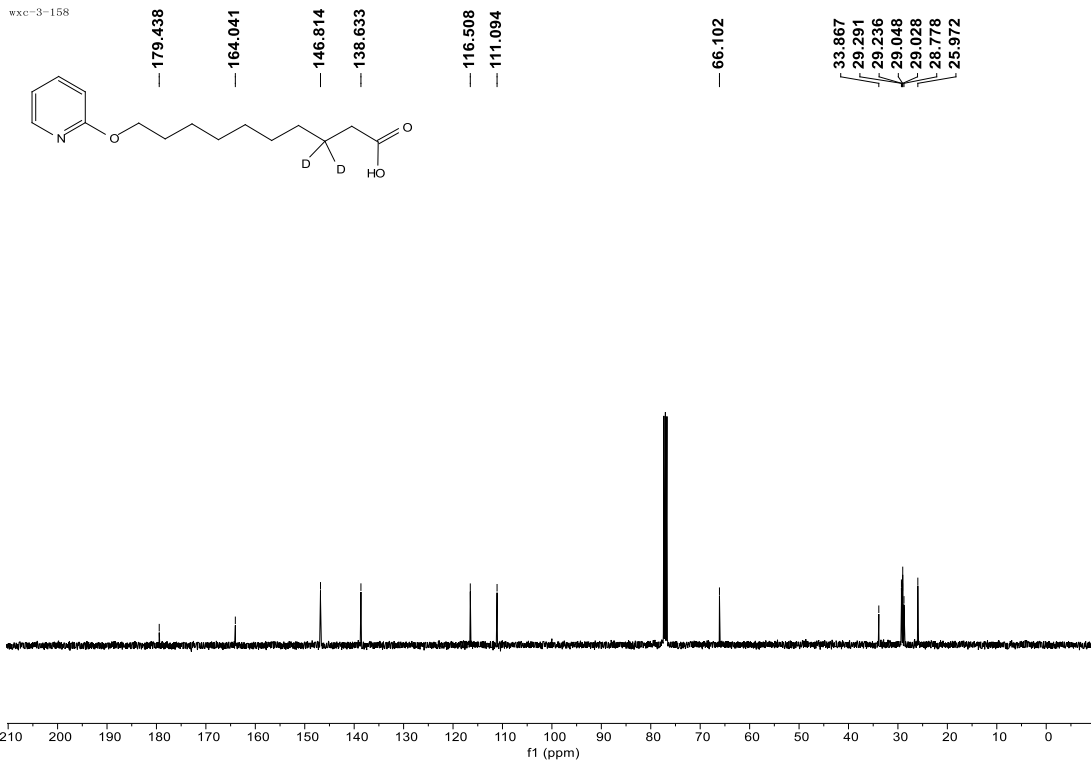
1.235



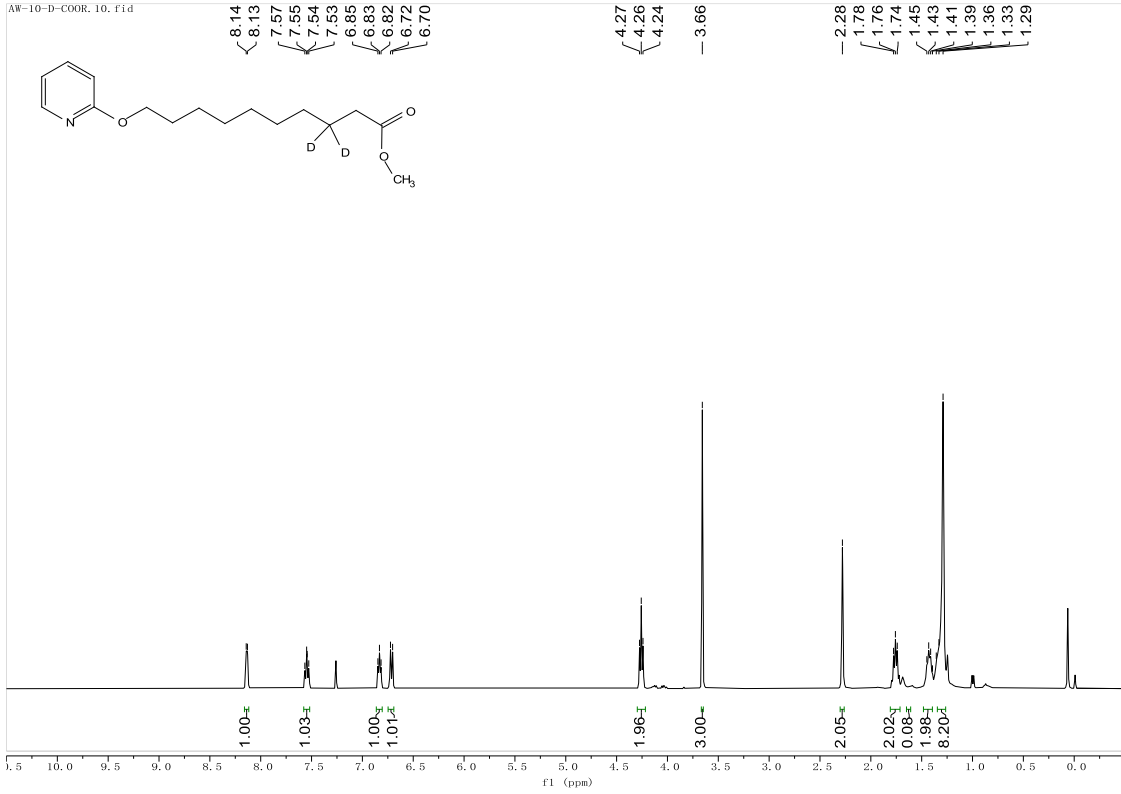
wxc-3-158



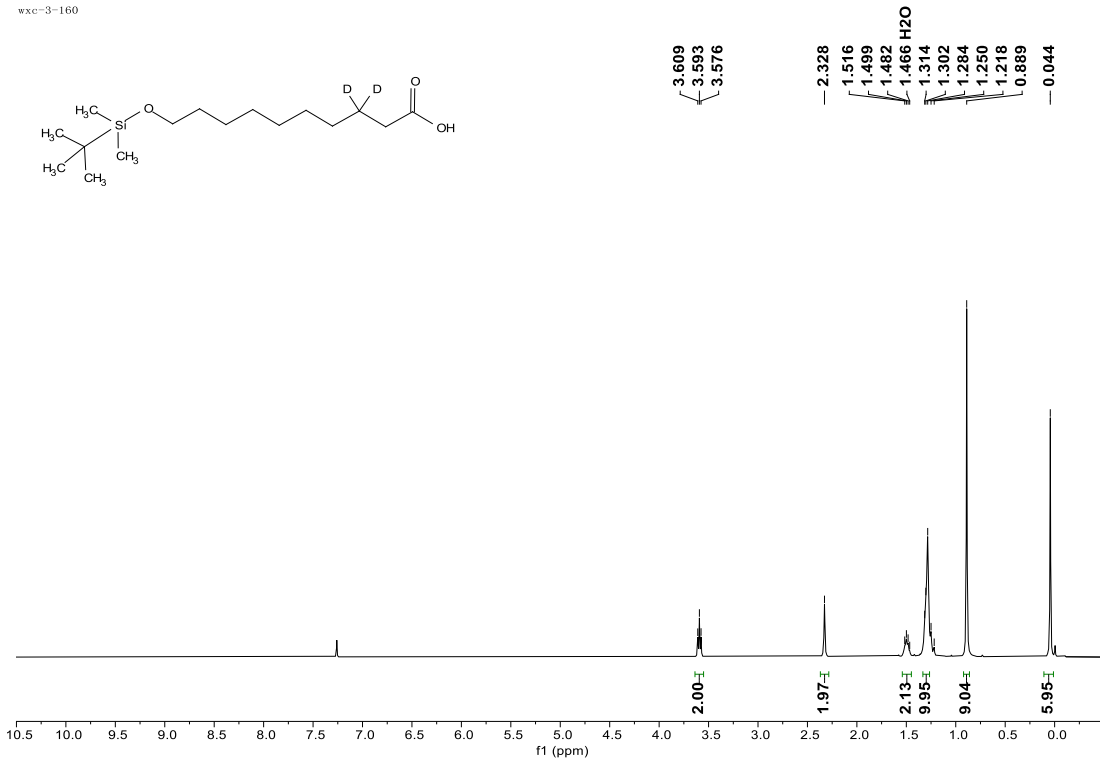
wxc-3-158



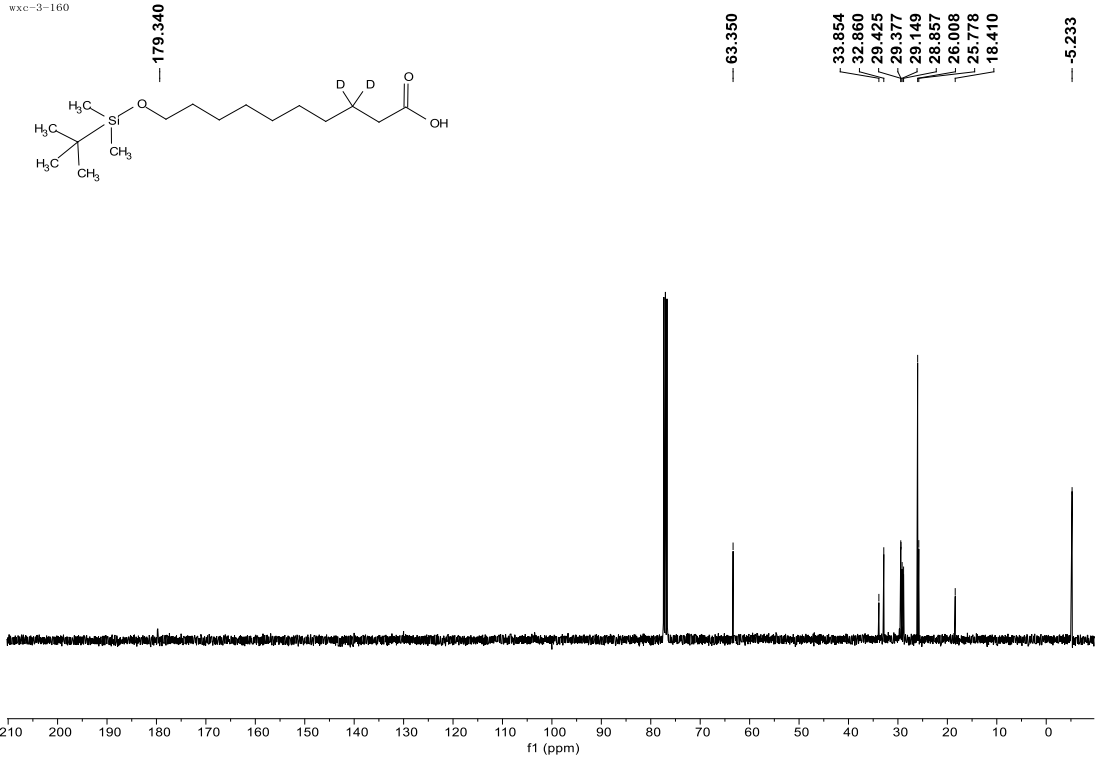
AW-10-D-COOR. 10. f1.d



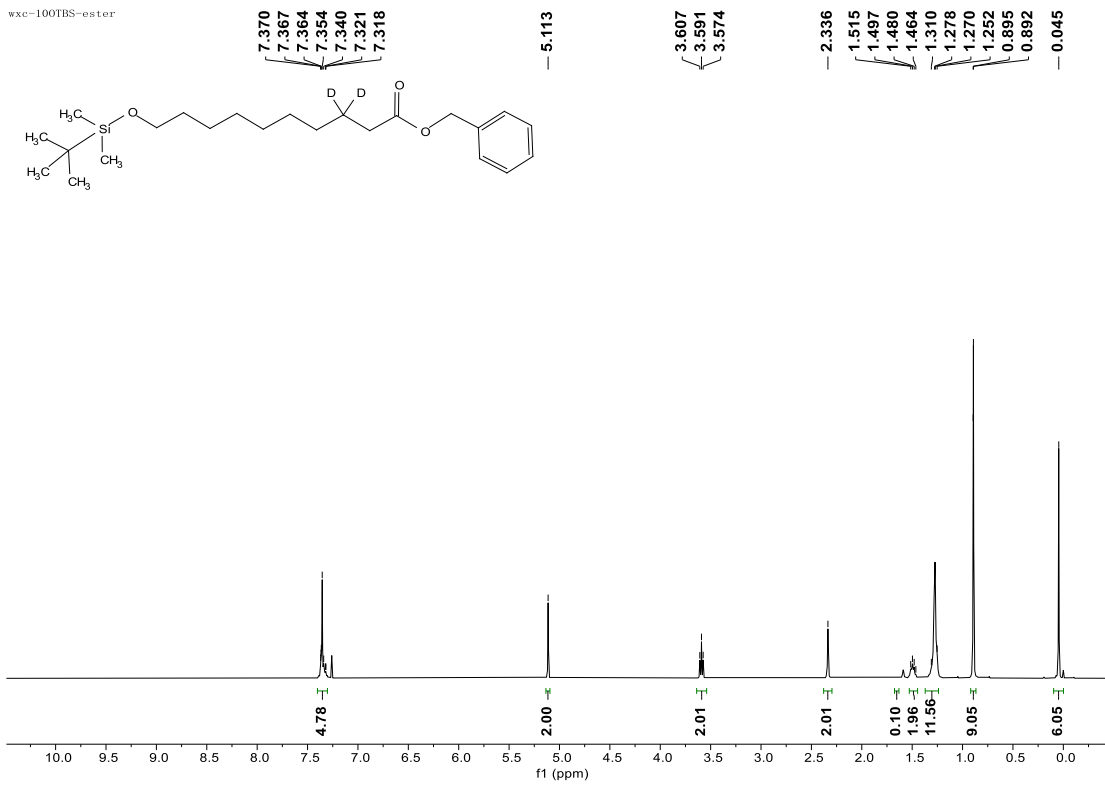
wxc-3-160



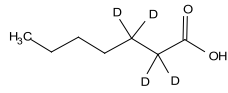
wxc-3-160



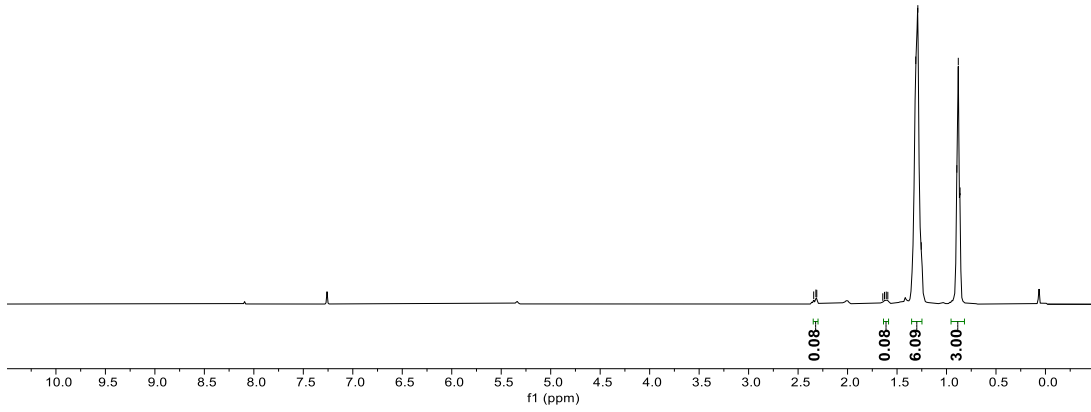
wxc-100TBS-ester



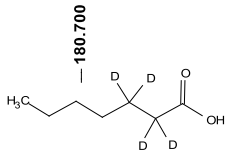
wxc-3-141



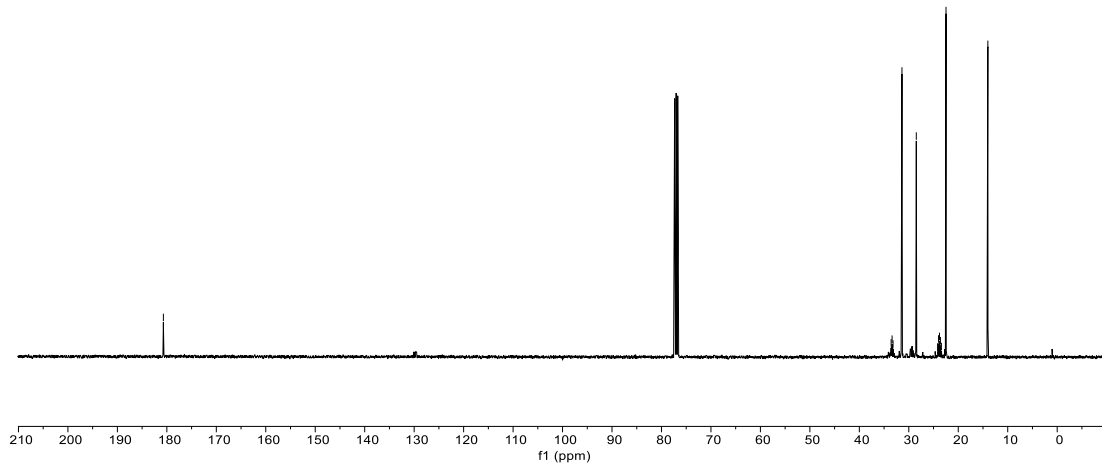
2.342
2.320
2.311
1.644
1.627
1.609
1.593
1.311
1.290
1.255
0.896
0.861
0.863



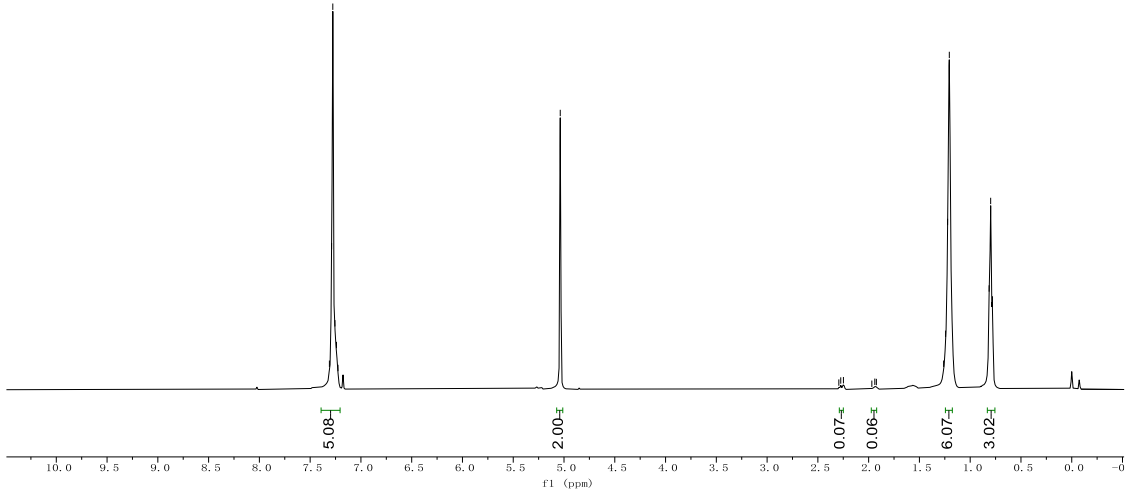
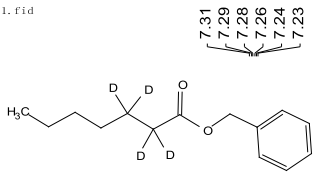
wxc-3-141



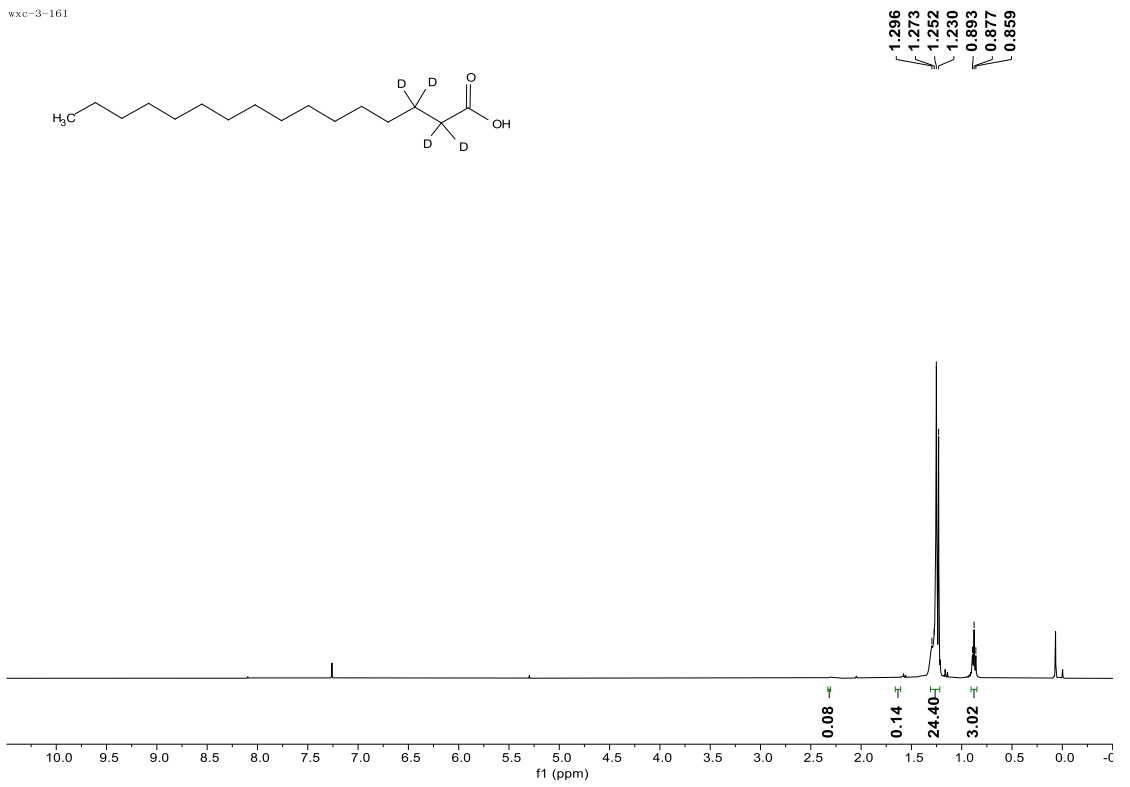
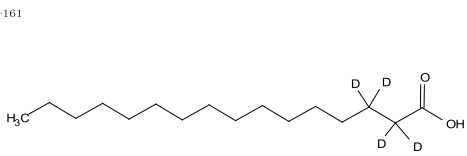
33.573
33.377
33.184
31.382
28.484
24.178
23.988
23.795
23.602
23.408
22.475
14.015



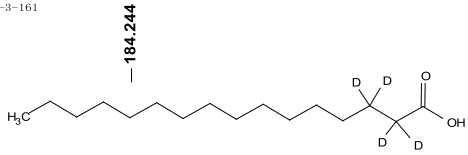
AW-7D-COOR. 1.fid



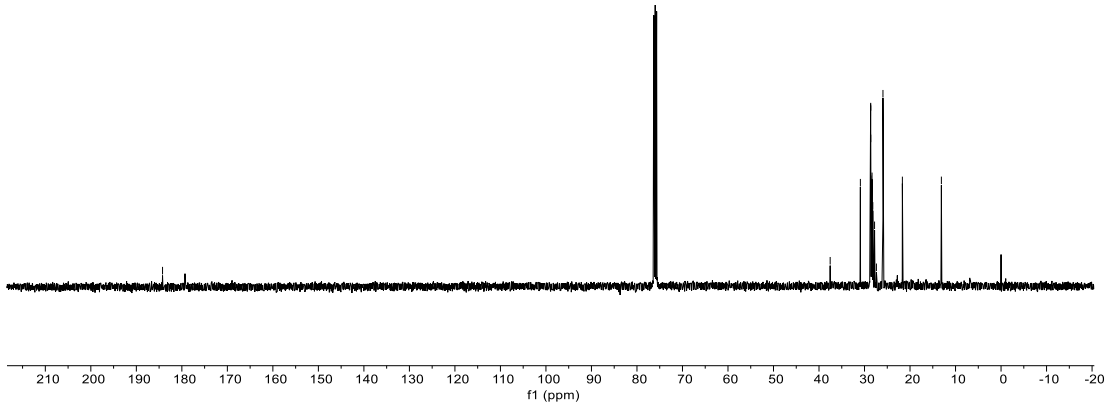
wxc-3-161



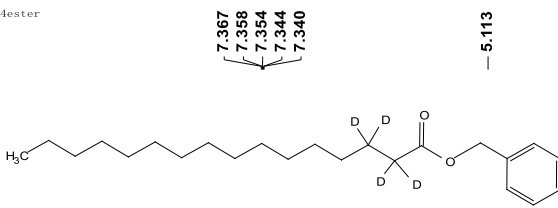
wxc-3-161



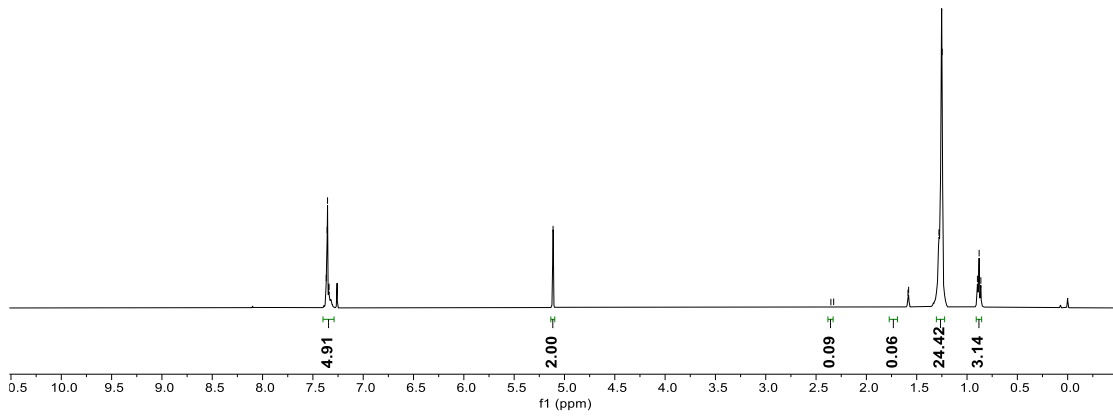
37.547
30.911
28.680
28.657
28.640
28.626
28.578
28.426
28.351
28.181
27.791
27.383
25.947
21.680
13.110



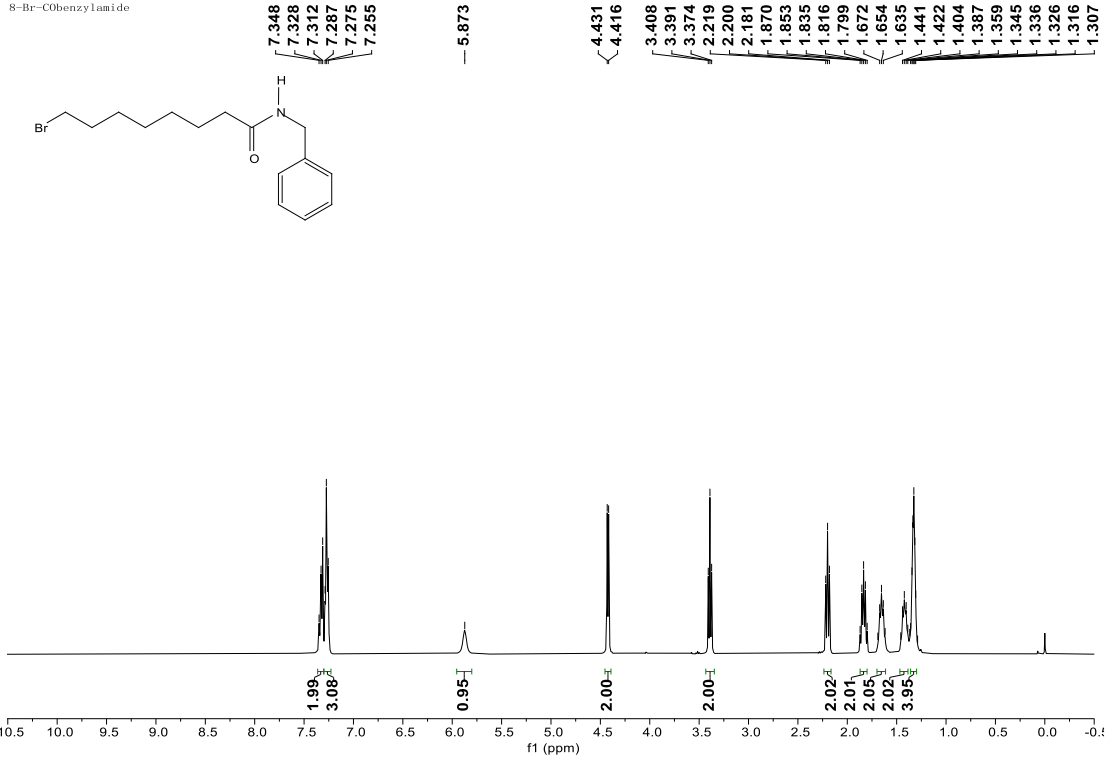
wxc-C16D4ester



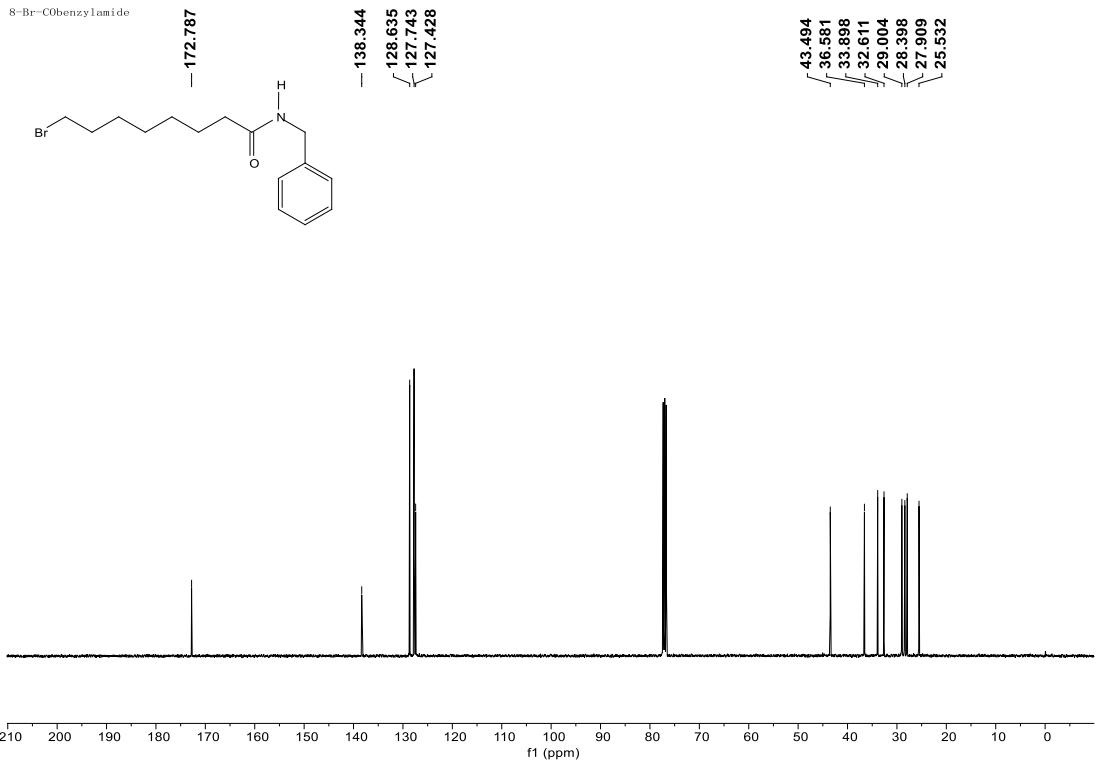
2.353
2.325
1.587
1.580
1.279
1.254
1.247
0.897
0.880
0.862



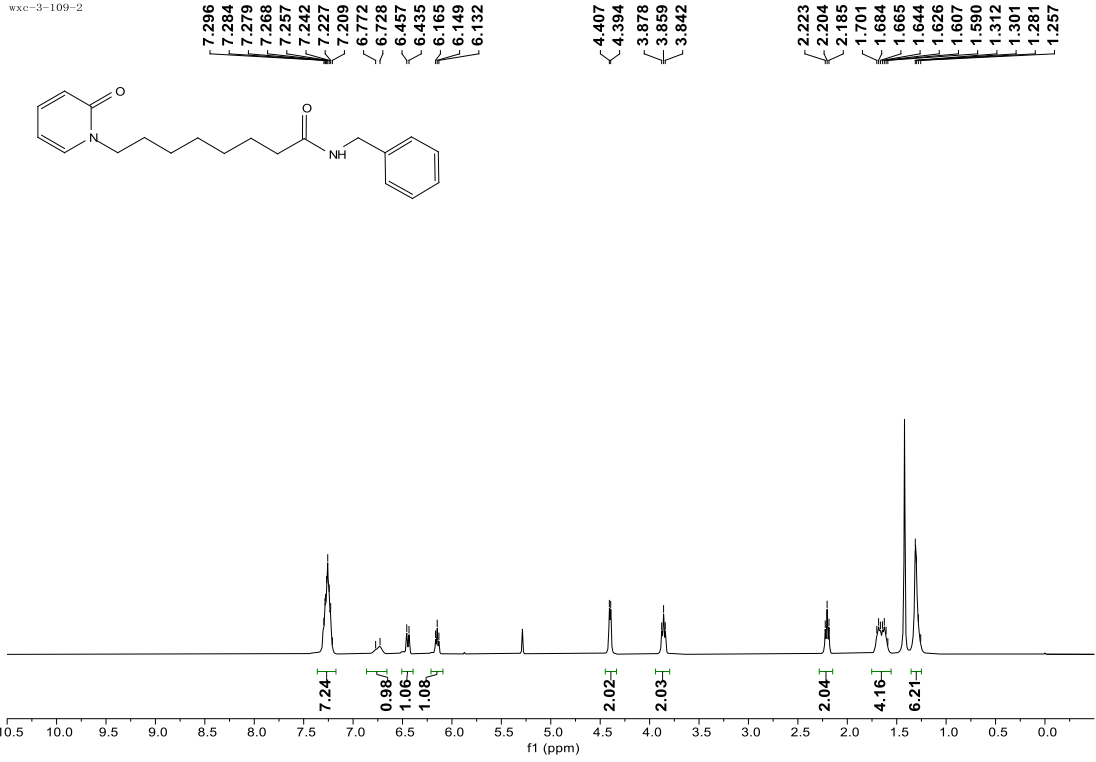
8-Br-Cobenzylamide



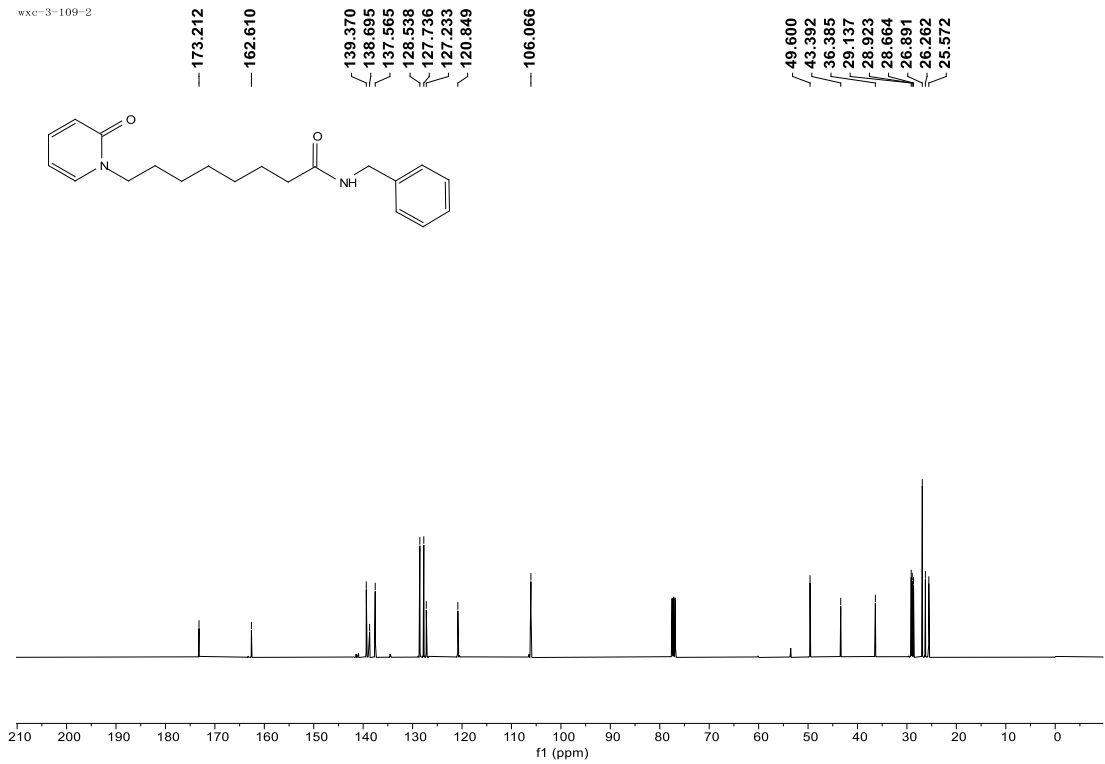
8-Br-Cobenzylamide



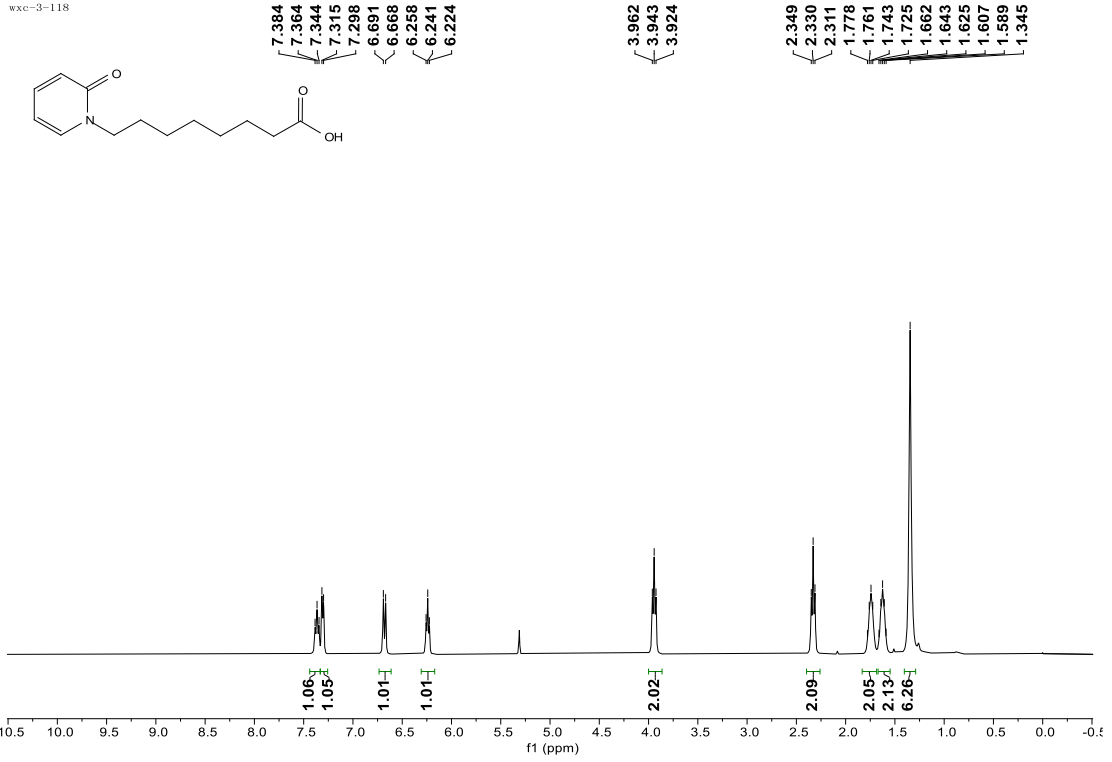
wxc-3-109-2



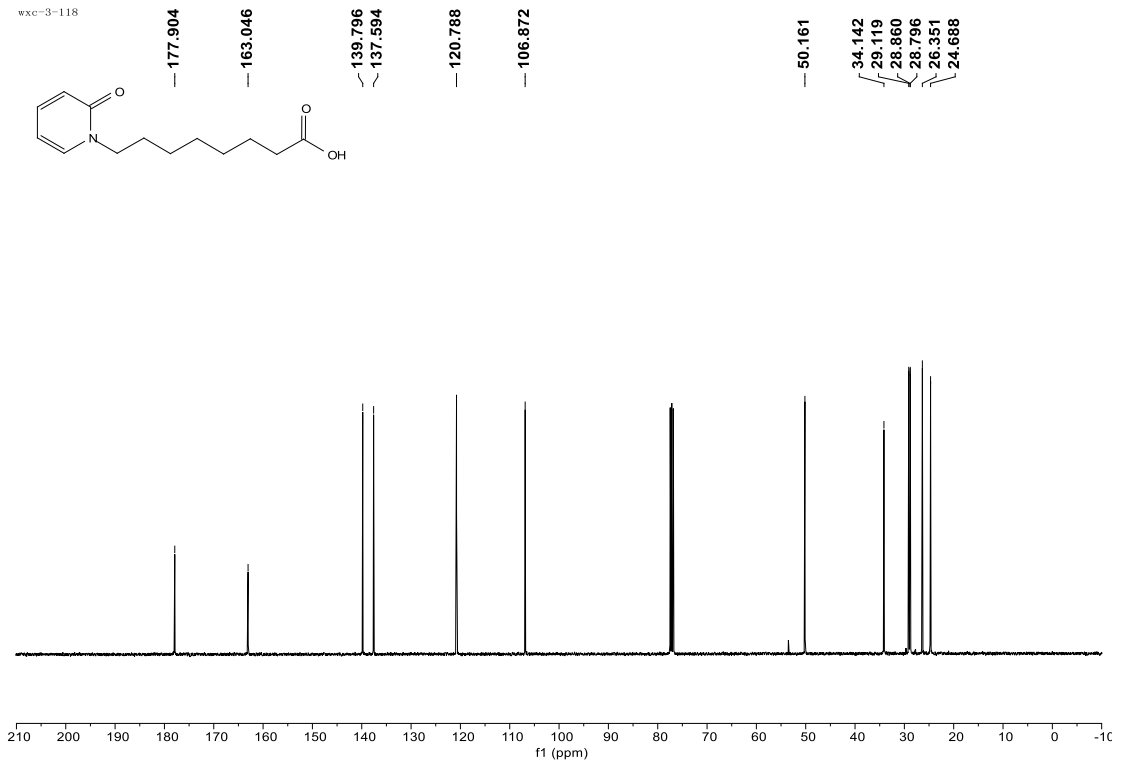
wxc-3-109-2



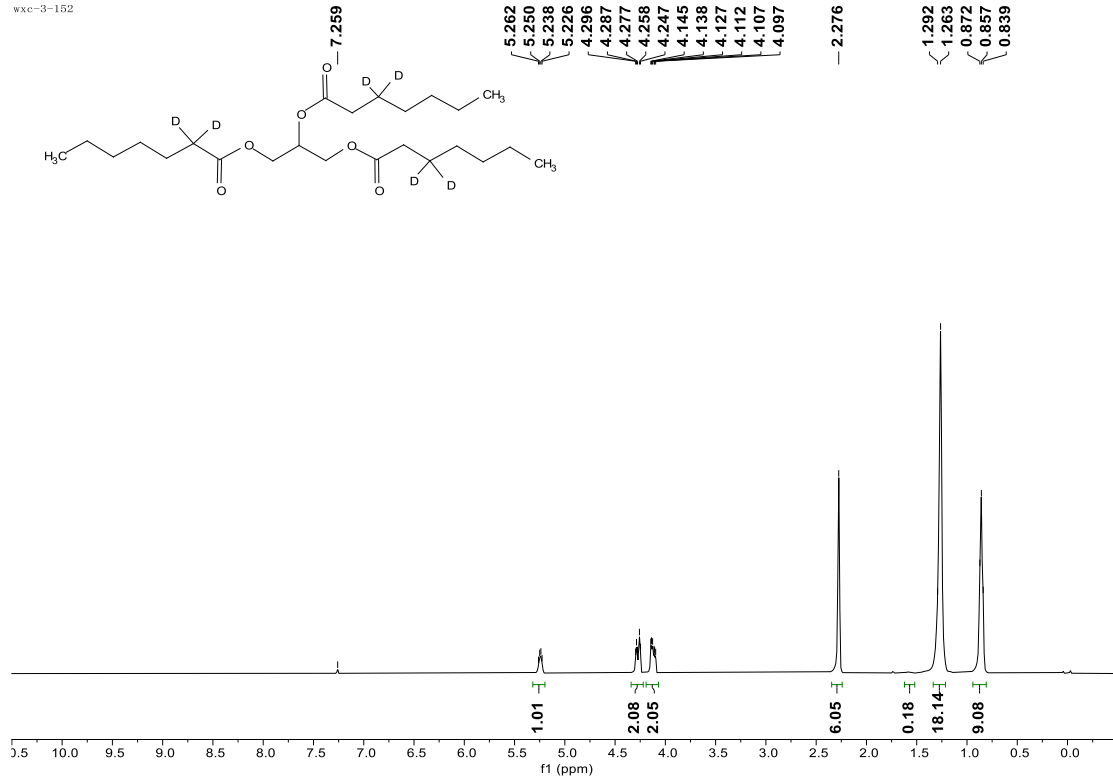
wxc-3-118



wxc-3-118



wxc-3-152



wxc-3-152

