Supplementary Information

Efficient preparation of fluorescent nanomaterials derived from chitin via

modification first strategy assisted by click chemistry

Yingyin Liu, Bowen Li, Chaoqun Xu, Zicong Shi, Liang Liu,

Yimin Fan, Juan Yu^{*}

Jiangsu Co-Innovation Center of Efficient Processing and Utilization of Forest Resources, International Innovation Center for Forest Chemicals and Materials, College of Chemical Engineering, Nanjing Forestry University, Nanjing 210037, China. E-mail: yujuannjfu@njfu.edu.cn

Experimental Section

(1) Deacetylation of Chitin

 α -Chitin (1 g) was suspended in 33% (w/w) NaOH (25 ml) and the slurry was heated at 90°C for 4 h with stirred. The partially deacetylated chitin was collected, and thoroughly washed with de-ionized water by repeated centrifugation at 8000 r/min for 5 min to neutrality. A portion of the wet NaOH-treated product was freeze-dried for further analyses, and the rest was kept in the wet state at 4°C.

(2) Preparation of TOChN-PPK and ChitinPPK

The preparation method of TOChN is as follows, the ratio of chitin to water was 1:100 (g/v) consisting of TEMPO (0.016 g/g chitin) and NaBr (0.10 g/g chitin), and then NaClO solution (8 mmol/g chitin) was mixed in the suspension in order to oxidize all the hydroxyl group at the C6 position. Moreover, the pH of the mixture was kept at 10 for 4 h at room temperature. Next, the sediments were cleaned to neutral with distilled water and homogenized under high pressure. The TOChN aqueous dispersion (the concentration was 1.01%, and the carboxyl content of TOChN was 0.93 mmol/g) was centrifuged (5 min, 8000 rpm) and stored at 4°C until use.

TOChN dispersion (0.10 g dry weight) was taken to adjust pH to 13, 0.081 g PPK (0.62 mmol) (dissolved in DMF) and DMAP (0.0075 g, 0.062 mmol) were added and stirred for 4 h. Finally, TOChN-PPK were obtained by multi centrifugal washing with

ethanol.

Chitin powder (0.10 g dry weight) was dispersed in water and the pH was adjusted pH to 13, 0.081 g PPK (0.62 mmol) (dissolved in DMF) and DMAP (0.0075 g, 0.062 mmol) were added and stirred for 4 h. Finally, ChitinPPK were obtained by multi centrifugal washing with ethanol.

Supplementary Figures



Strategy a: via modification firstStrategy b: via nanosizing firstFigure.S1 The left figure shows the dispersion and film of fluorescent chitin nanofibersprepared by modification first of strategy a. The right figure shows the dispersion andfilm of fluorescent chitin nanofibers prepared by nanosizing first of strategy b.



Figure.S2 Emission spectra of fluorescent chitin nanofibers prepared by two strategies.

Procedure	(Waste salt			
	NaOH/g	Acetic	Distilled	Sodium	
		acid/mL	water/mL	acetate/g	
b1	2.58	-	640	-	
b2	-	5.00	500	-	
b3	4.30	-	200	7.20	

Table S1 The amount of chemical reagents used and wasted in additional steps in

traditional methods strategy b

NOTE: The amount of the chemical reagents is calculated based on 1.00 g chitin and the resulted FChN dispersion with a concentration of 0.20 wt%.

Table C2	Commention	of managemention	musses of different	fluoressent non of home
Table SZ	Comparison	of preparation	brocesses of different	nuorescent nanombers.
	1	1 1	1	

_ Sample		Modification		Separation			
Entry	name	temperature time		steps	reagent	- Ref.	
1 dChNC- FITC		DТ	36 h		ethanol,	1	
		KI		centrilugation	water		
2	FChN	120℃	4-48 h	centrifugation,	ethanol	2	
				filtration	ethanor		
3	DTAF- CNF	RT	24 h	vacuum filtration,		3	
				centrifugation and	NaOH		
				ultrafiltration			
4	TOCNC	RT	16 h	centrifugation,	water	4	
	-AANI			dialysis			
5	CNC-	60°C	4 h+24 h	dialysis	water	5	
	RBITC			antrifugation	watan		
6	FCNC	RT	16 h	dialucia	water	6	
				centrifugation	water		
7	FCNF	60°C	4 h+24 h	filtration	water	7	
8	DTAP- CNF	RT	24 h	centrifugation	water		
				dialysis	PBS	8	
9	FChN	RT	4 h		ethanol	This	
				centrifugation		work	



Figure S3 DDA of different FCh samples.



Figure S4 The absorbance of TOChN and TOChN-PPK and digital photographs of ChitinPPK, TOChN and TOChNPPK under UV (365 nm).



Figure.S5 Hydrophobicity of DEChN and FChN6-9.

Sustainable materials.					
Entry	Sample name	Stress/ MPa	UV shielding/%	UV shielding to/nm	Ref.
1	ChNF/PVA	225.00	100.00	360	9
2	PVA-ChNF-LNPs	33.24	100.00	350	10
3	GL	70.00	100.00	400	11
4	CMC/Pal-D1	40.02	97.00	300	12
5	CNF-VE	127.78	100.00	400	13
6	C/PVA/CIP	54.00	100.00	355	14
7	BACNC	11.55	97.00	300	15
8	CS:hBNNSs	35.90	96.40	400	16
9	HEC/ANFs	55.60	100.00	375	17
10	ACN	73.50	87.00	400	18
11	PI/PDA-120	94.10	100.00	400	19
12	FChN	200.12	100.00	400	This work

Table S3 Summary of mechanical properties and UV shielding properties of different sustainable materials.



Figure S6 Thermal stability analysis of FChN9 and DEChN



Figure S7 Stability of DEChN and FChN9 at different temperatures

References

- 1 M. Zhou, Z. Liu, T. Liu, Y. Zhu and N. Lin, *ACS Sustain. Chem. Eng.*, 2022, **10**, 10327–10338.
- A. K. Dutta, H. Izawa, M. Morimoto, H. Saimoto and S. Ifuku, *J. Chitin Chitosan Sci.*, 2015, **3**, 53–56.
- 3 M. S. Reid, M. Karlsson and T. Abitbol, *Carbohydr. Polym.*, 2020, **250**, 116943.
- 4 W. Wu, R. Song, Z. Xu, Y. Jing, H. Dai and G. Fang, *Sensors Actuators B Chem.*, 2018, **275**, 490–498.
- 5 K. A. Mahmoud, J. A. Mena, K. B. Male, S. Hrapovic, A. Kamen and J. H. T. Luong, *ACS Appl. Mater. Interfaces*, 2010, **2**, 2924-2932.
- R. Song, Q. Zhang, Y. Chu, L. Zhang, H. Dai and W. Wu, *Cellulose*, 2019, 26, 9553–9565.
- 7 Q. Ding, J. Zeng, B. Wang, W. Gao, K. Chen, Z. Yuan, J. Xu and D. Tang, *Carbohydr. Polym.*, 2018, **186**, 73–81.
- M. Babi, A. Fatona, X. Li, C. Cerson, V. M. Jarvis, T. Abitbol and J. M. Moran-Mirabal, *Biomacromolecules*, 2022, 23, 1981–1994
- 9 D. Jiao, Z. Li, J. Y. Hu, X. N. Zhang, J. Guo, Q. Zheng and Z. L. Wu, *Compos. Sci. Technol.*, 2023, 244, 110295.
- 10 Q. Zhou, J. Chen, C. Wang, G. Yang, S. Janaswamy, F. Xu and Z. Liu, *Ind. Crops Prod.*, 2022, **188**, 115669.
- 11 M. J. Mehta and A. Kumar, *Chem. A Eur. J.*, 2019, **25**, 1269–1274.
- 12 D. Huang, Y. Zheng and Q. Quan, *Appl. Clay Sci.*, 2019, **183**, 105314.
- 13 B. Kumar, S. Adil, D. H. Pham and J. Kim, *Heliyon*, 2024, 10, e25272.
- M. Ahmed, A. R. Bhat, A. K. Verma and R. Patel, *ACS Appl. Bio Mater.*, 2023, 6, 663-673.
- 15 Y. Ren, J. Ma, W. Liu, C. Huang, C. Lai, Z. Ling and Q. Yong, *Int. J. Biol. Macromol.*, 2022, **204**, 41–49.
- K. Wang, F. Li, X. Sun, F. Wang, D. Xie and Y. Wei, *Int. J. Biol. Macromol.*, 2023, 251, 126308.
- 17 J. Huang, Z. Lu, J. Li, D. Ning, Z. Jin, Q. Ma, L. Hua, S. E and M. Zhang,

Carbohydr. Polym., 2021, 255, 117330.

- 18 M. M. Hasan, M. S. Hossain, M. D. Islam, M. M. Rahman, A. S. Ratna and S. Mukhopadhyay, ACS Appl. Mater. Interfaces, 2023, 15, 32011-32023.
- Q. Li, S. Liu, Y. Guo, Y. Liang, H. Peng, R. Chen, F. Lei, W. Wu, H. Zhao, Q. Zhang, R. K. Y. Li and W. Duan, ACS Appl. Polym. Mater., 2021, 3, 896-907.