

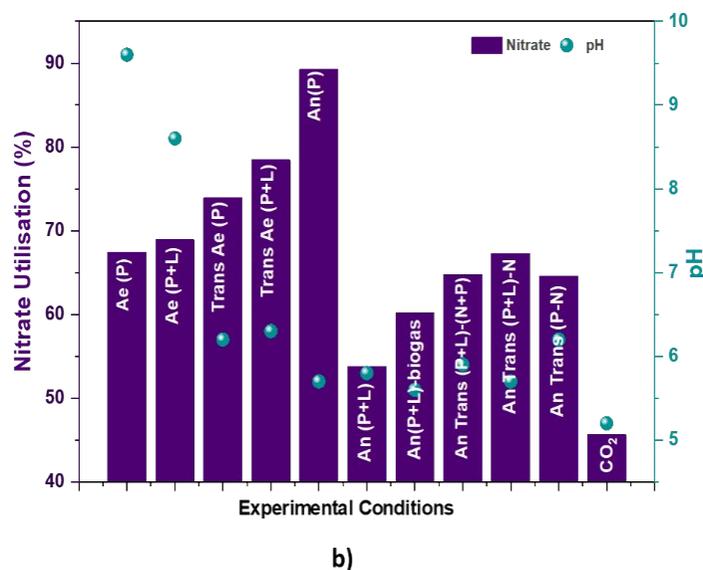
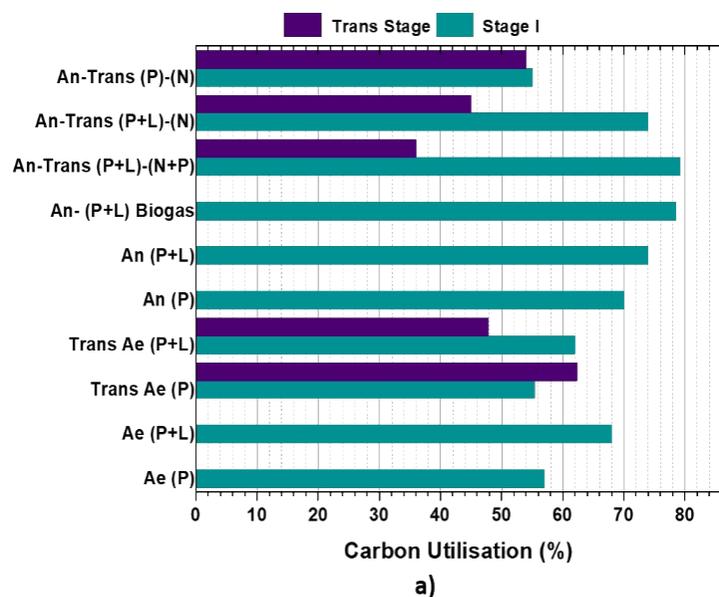
Metabolic Channelling in *Rhodospseudomonas palustris* for Hydroxyalkanoate Polymers Diversification and Sustainability Assessment

Poonam Kumari^{a,b} and S. Venkata Mohan^{a,b*}

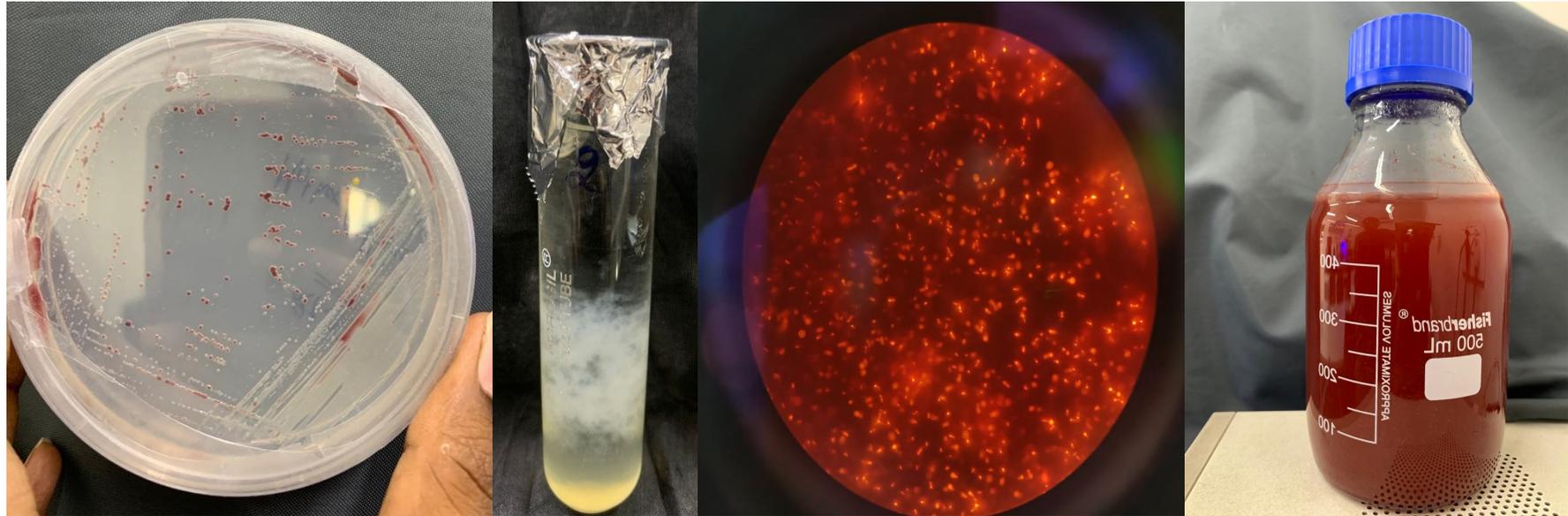
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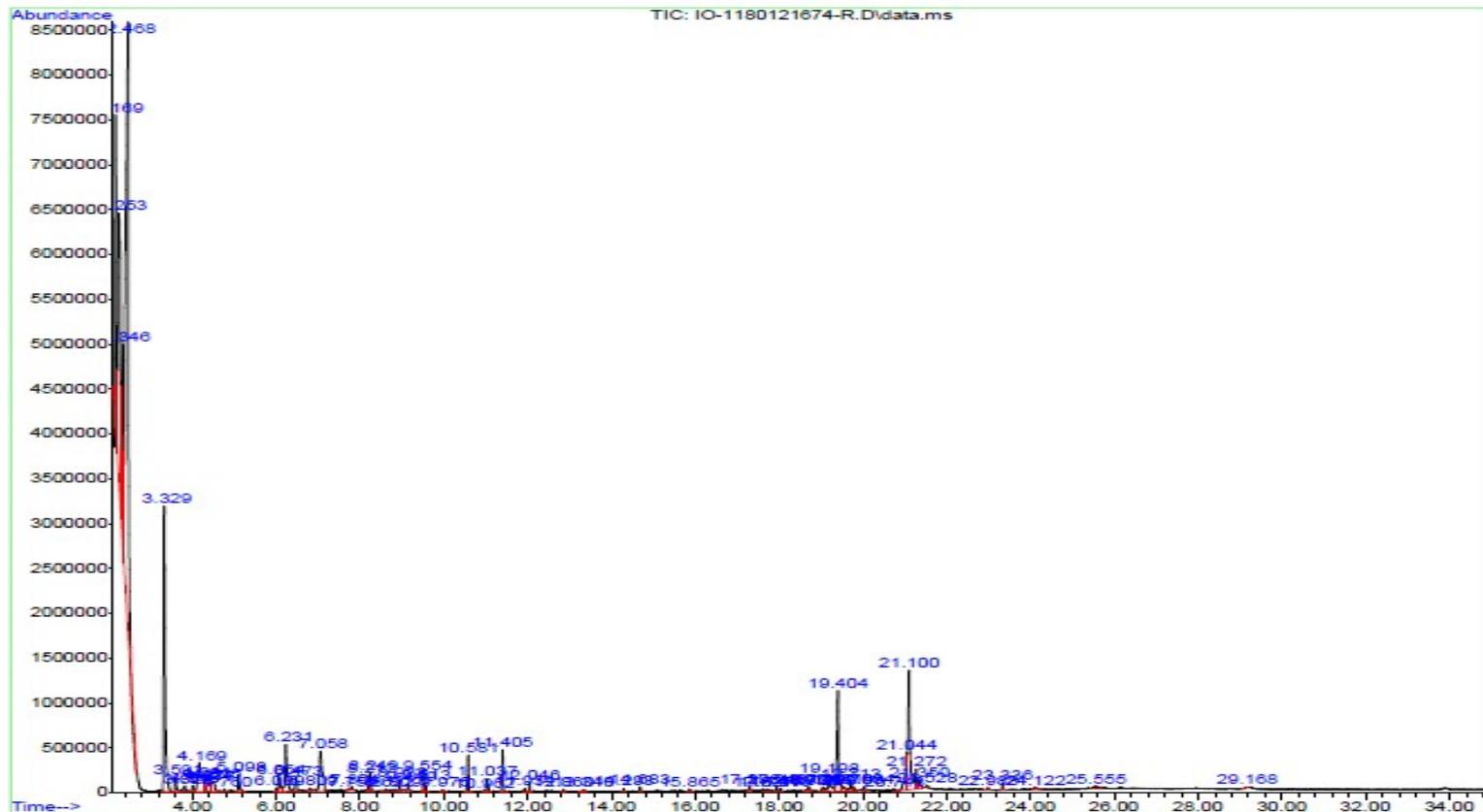


S Fig 1: a) Carbon utilisation in respective experimental conditions at the end of growth phases b) Nitrate concentration and variations at the end of growth phases



S Fig 2: Isolated *Rhodospirillum rubrum* SVMIICT10 strain (MTCC deposition no. 16028), PHA Precipitation during Extraction and Fluorescence Imaging

Sample : ST-1
Misc :
ALS Vial : 1 Sample Multiplier: 1
Search Libraries: C:\Database\NIST02.L Minimum Quality: 0
C:\Database\NIST14.L Minimum Quality: 0
C:\Database\W9N11.L
Unknown Spectrum: Apex
Integration Events: ChemStation Integrator - autoint1.e

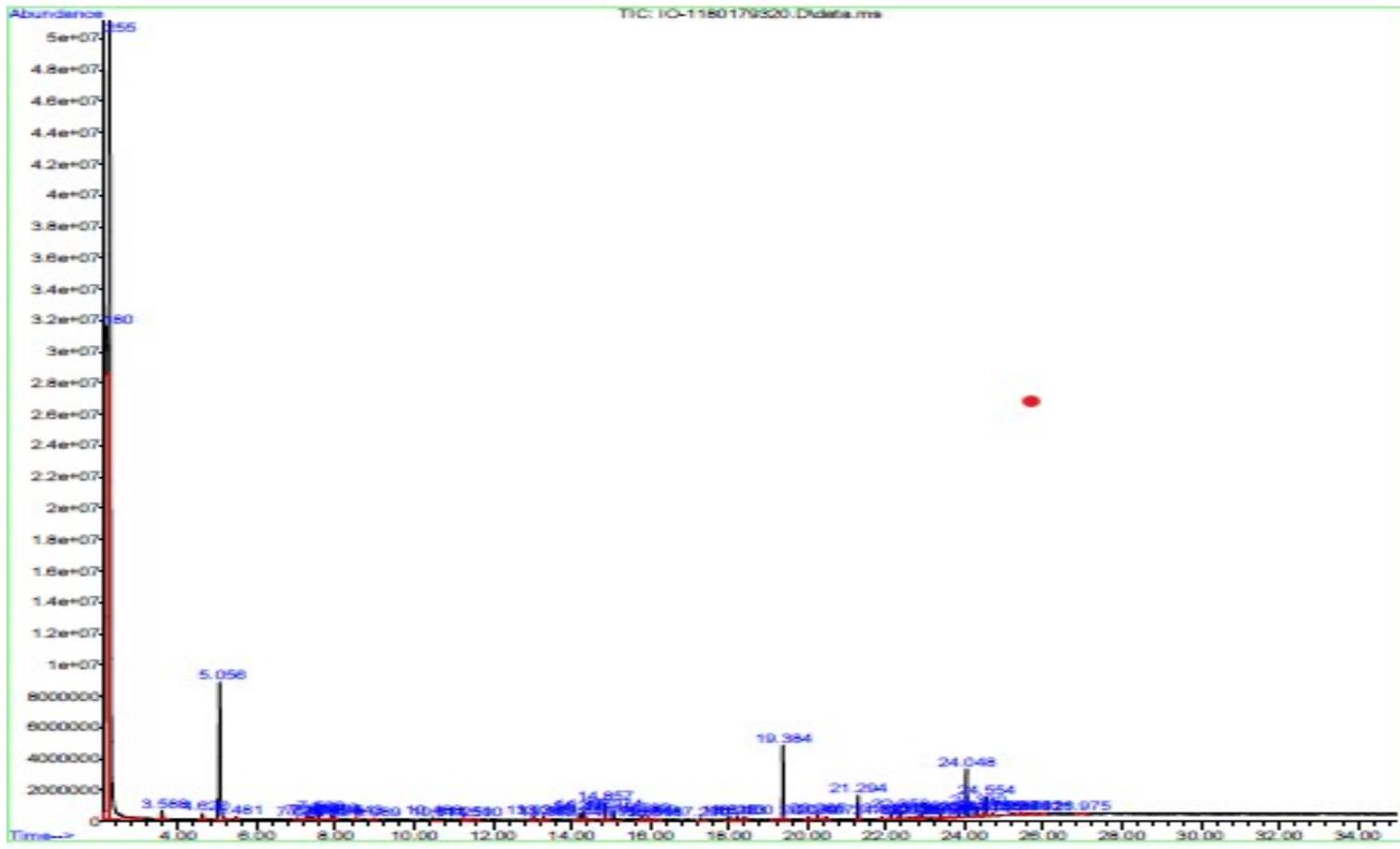


S Fig 3a: GC-MS Chromatogram of P-3HB Sigma Aldrich-Natural Origin Standard

Sample : AE-5
Misc :
ALS Vial : 6 Sample Multiplier: 1

Search Libraries: C:\Database\NIST14.L Minimum Quality: 0

Unknown Spectrum: Apex minus start of peak
Integration Events: ChemStation Integrator - autoint1.e

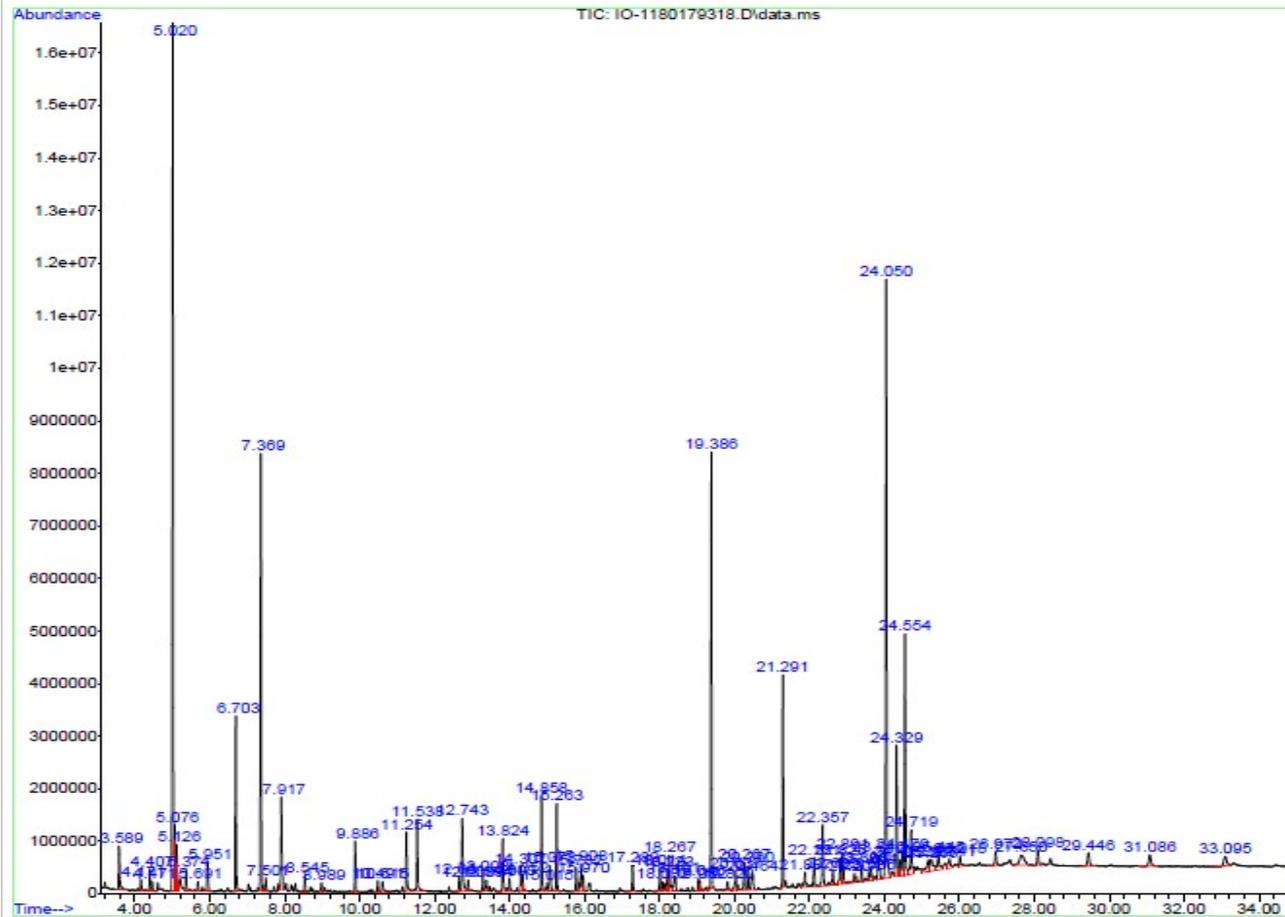


S Fig 3b: GC-MS Raw Chromatogram of Ae (P)

Sample : AE+L-3
Misc :
ALS Vial : 3 Sample Multiplier: 1

Search Libraries: C:\Database\NIST02.L Minimum Quality: 0

Unknown Spectrum: Apex minus start of peak
Integration Events: ChemStation Integrator - autoint1.e

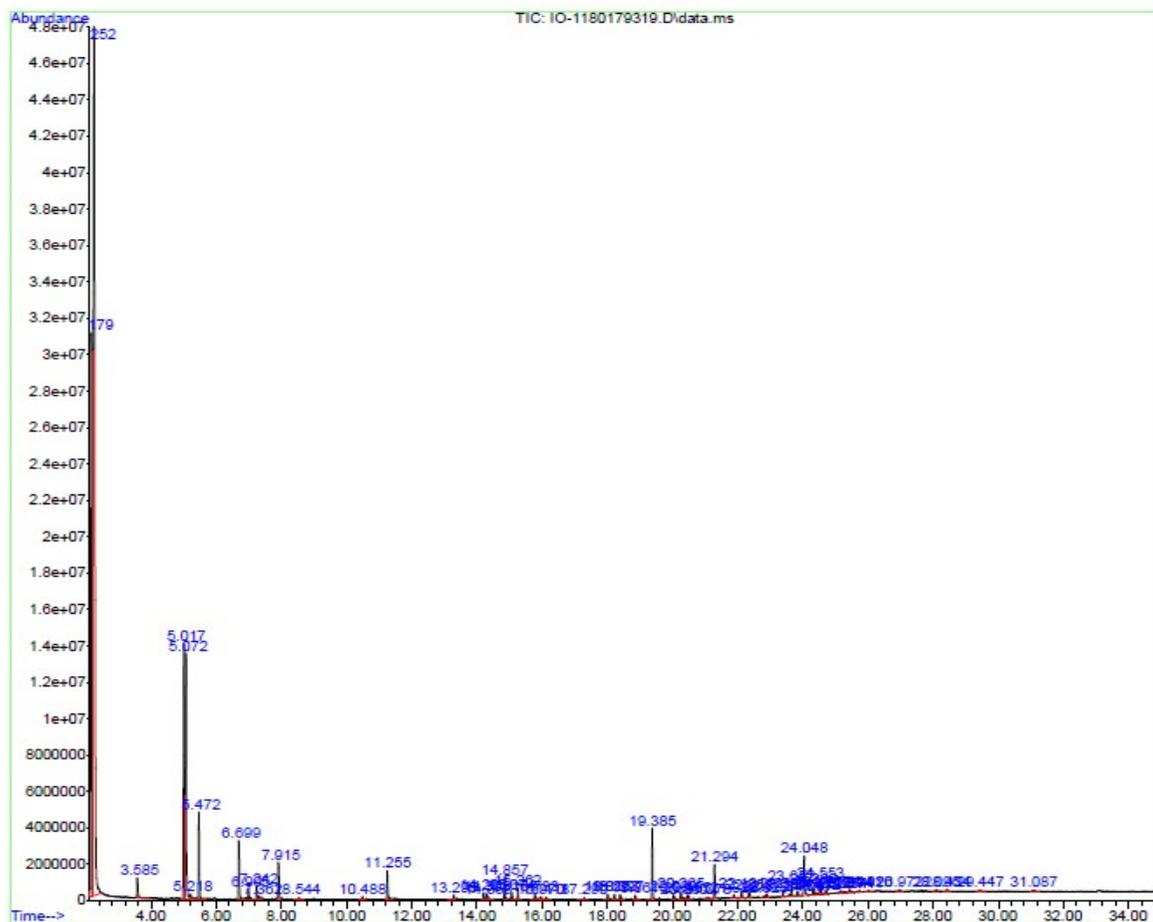


S Fig 3c: GC-MS Raw Chromatogram of Ae (P+L)

Sample : AE+L-TRANS-4
Misc :
ALS Vial : 4 Sample Multiplier: 1

Search Libraries: C:\Database\NIST14.L Minimum Quality: 0

Unknown Spectrum: Apex minus start of peak
Integration Events: ChemStation Integrator - autoint1.e

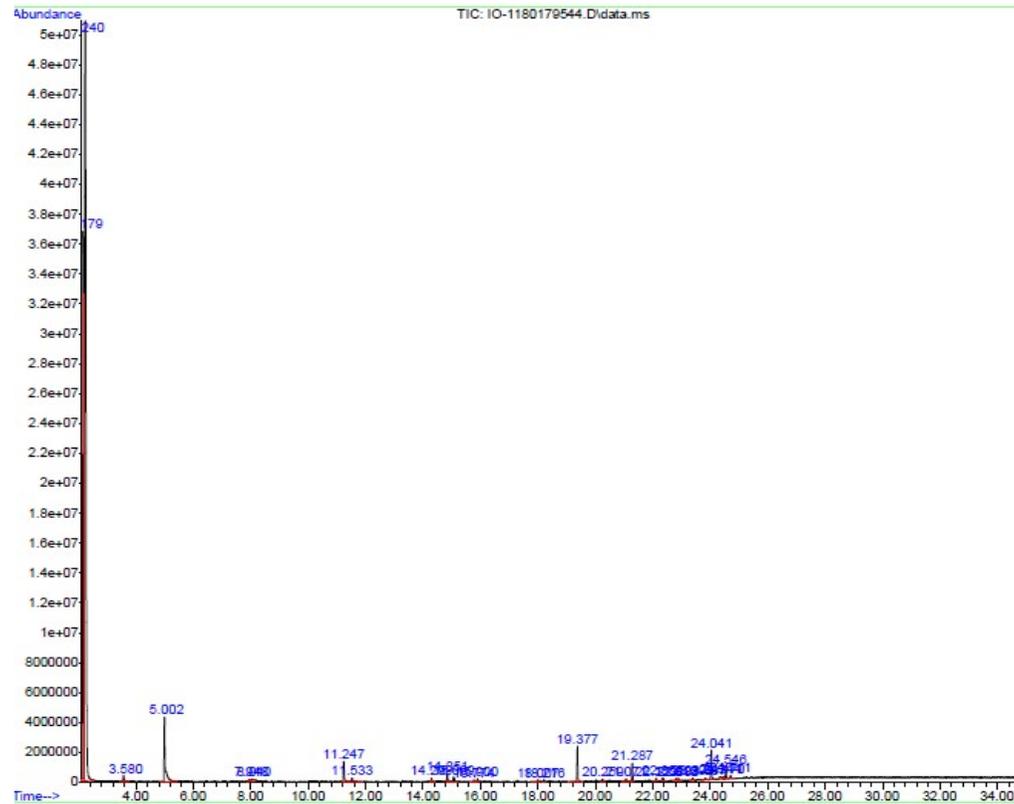


S Fig 3d: GC-MS Raw Chromatogram of Trans Ae (P+L)

Sample : AE-TRANS-6
Misc :
ALS Vial : 12 Sample Multiplier: 1

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C:\Database\NIST14.L Minimum Quality: 0
C:\Database\W9N11.L

Unknown Spectrum: Apex minus start of peak
Integration Events: ChemStation Integrator - autoint1.e

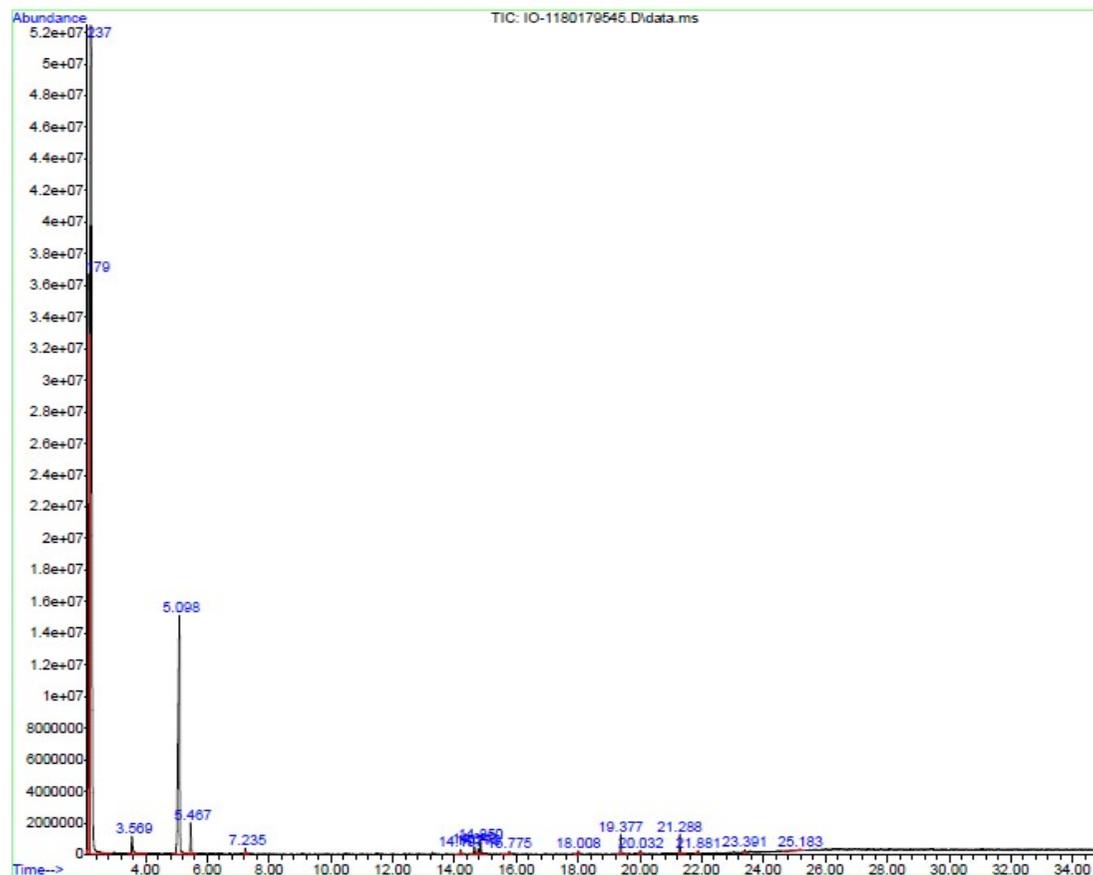


S Fig 3e: GC-MS Raw Chromatogram of Trans Ae (P)

Sample : AN-7
Misc :
ALS Vial : 13 Sample Multiplier: 1

Search Libraries: C:\Database\NIST02.L Minimum Quality: 0
C:\Database\NIST14.L Minimum Quality: 0
C:\Database\W9N11.L

Unknown Spectrum: Apex minus start of peak
Integration Events: ChemStation Integrator - autoint1.e

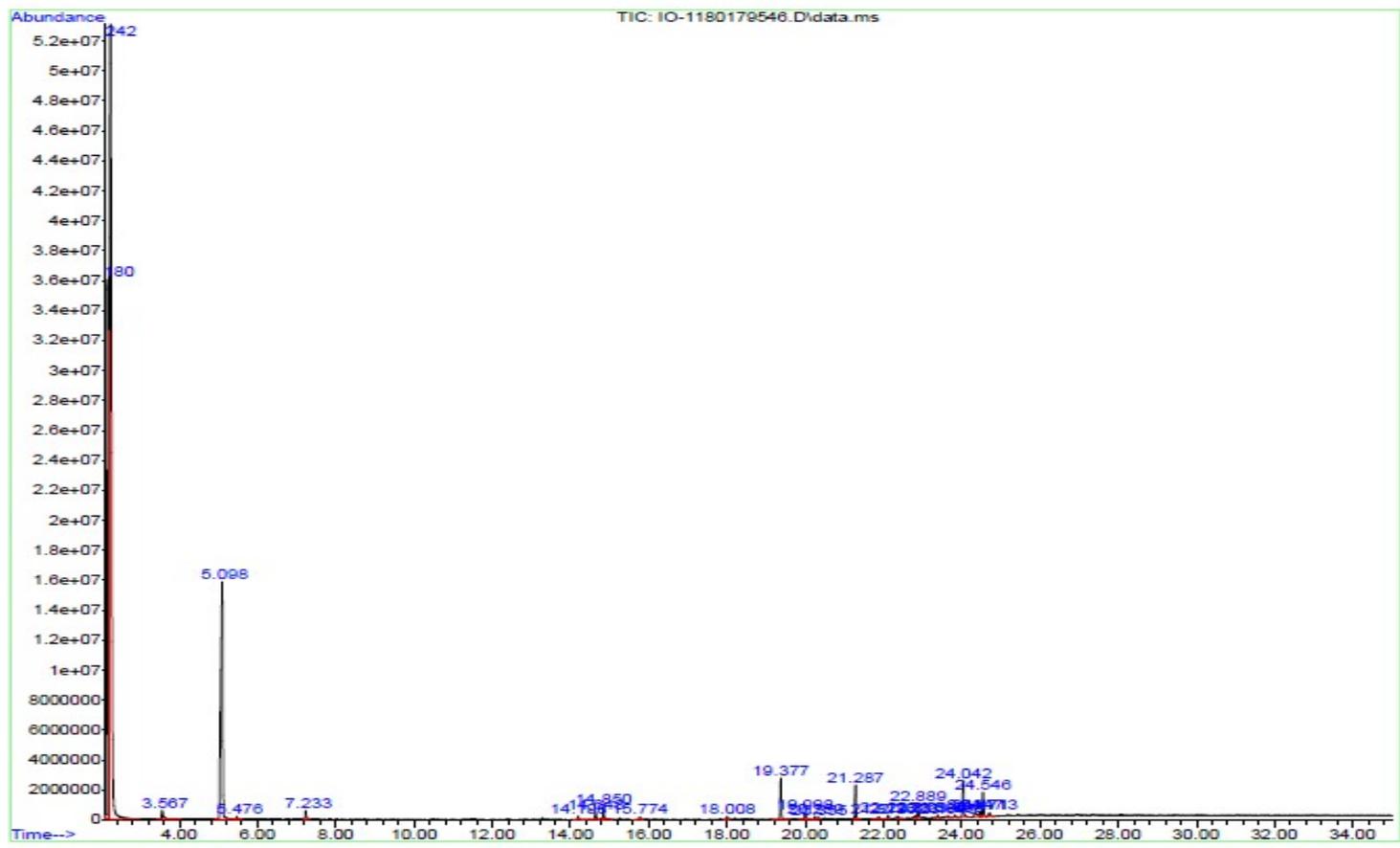


S Fig 3f: GC-MS Raw Chromatogram of An (P)

Operator : NONG LIAO
Sample : AN+L-8
Misc :
ALS Vial : 14 Sample Multiplier: 1

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C:\Database\NIST14.L Minimum Quality: 0
C:\Database\W9N11.L

Unknown Spectrum: Apex minus start of peak
Integration Events: ChemStation Integrator - autoint1.e

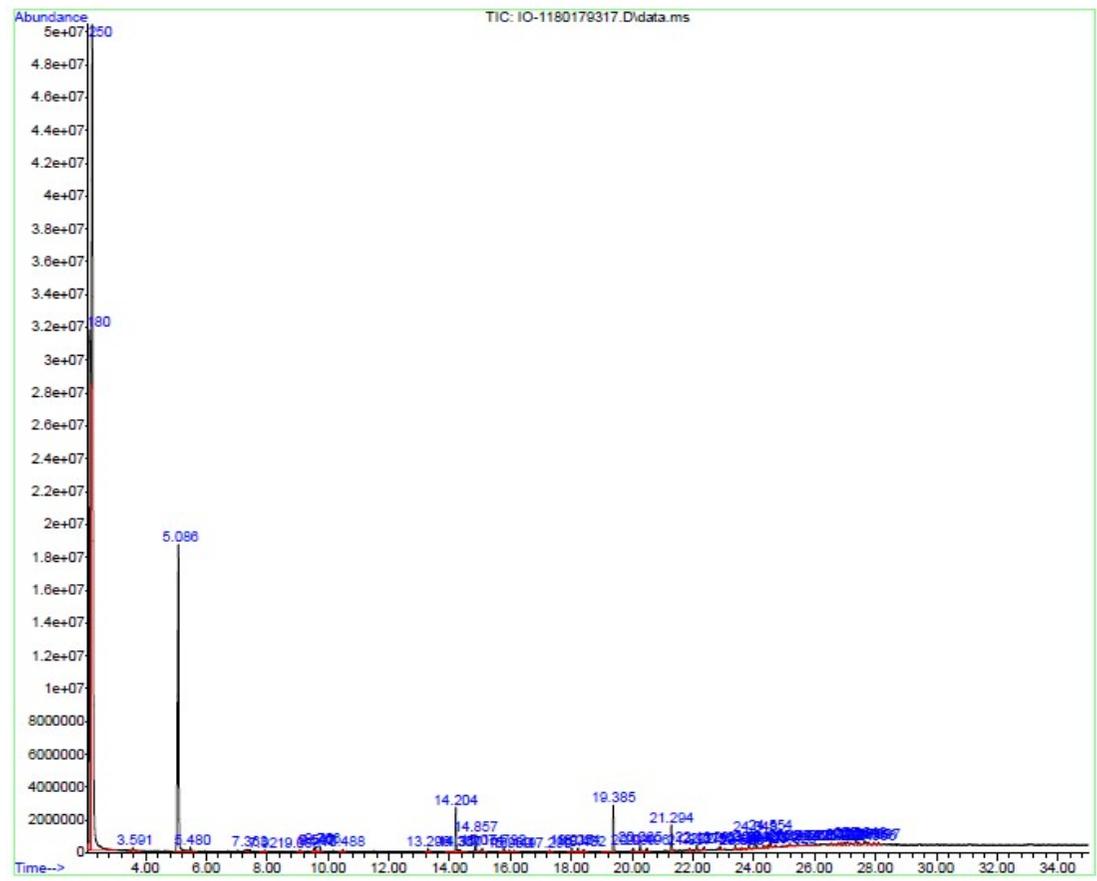


S Fig 3g: GC-MS Raw Chromatogram of An (P+L)

Sample : AN-H2-2
Misc :
ALS Vial : 5 Sample Multiplier: 1

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C:\Database\W9N11.L

Unknown Spectrum: Apex minus start of peak
Integration Events: ChemStation Integrator - autoint1.e

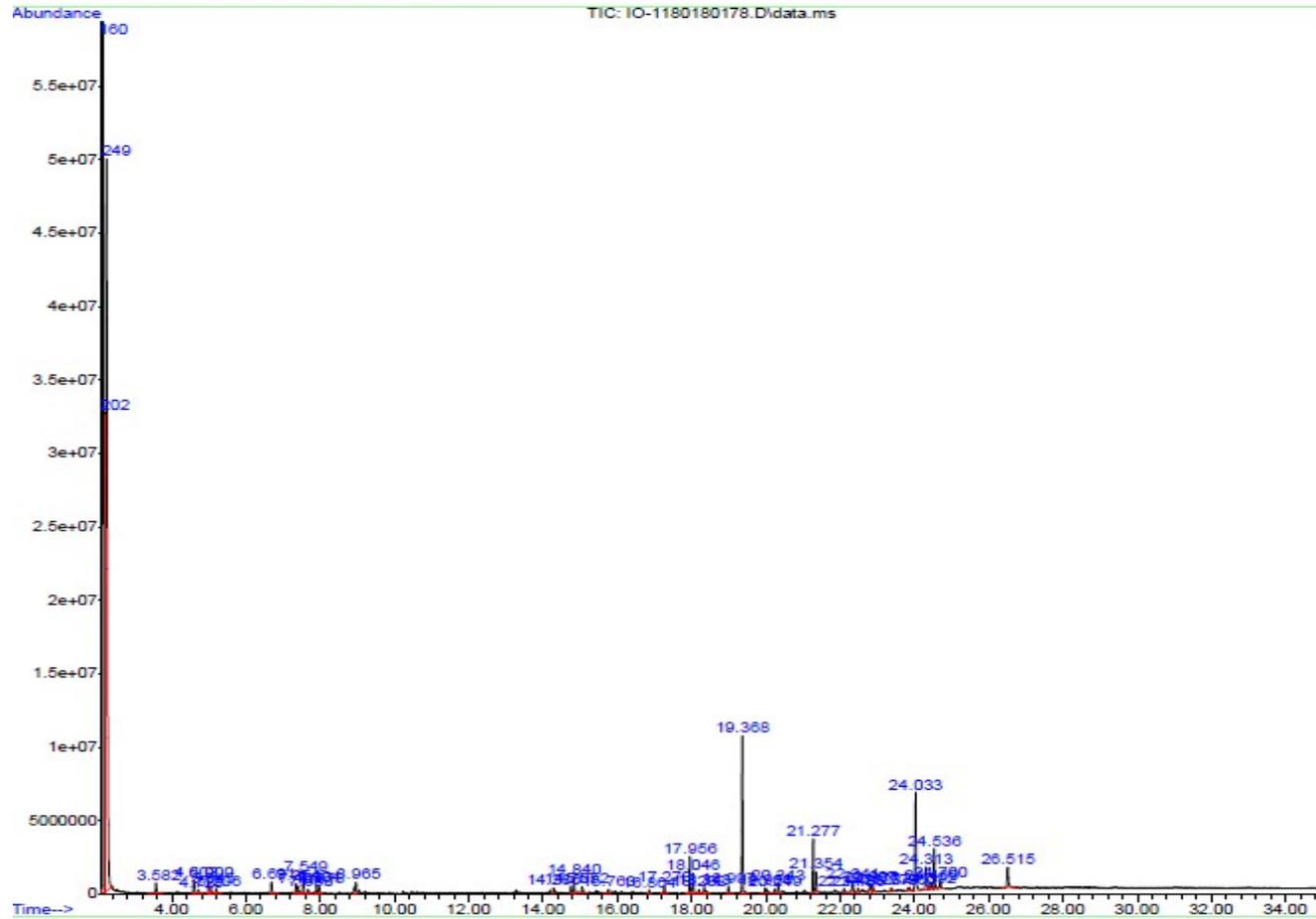


S Fig 3h: GC-MS Raw Chromatogram of An- (P+L) Biogas

Operator : NIMS Lab 10/10/2011
Sample : 11-TRANSL-N+P
Misc :
ALS Vial : 2 Sample Multiplier: 1

Search Libraries: C:\Database\NIST02.L Minimum Quality: 0

Unknown Spectrum: Apex minus start of peak
Integration Events: ChemStation Integrator - autoint1.e

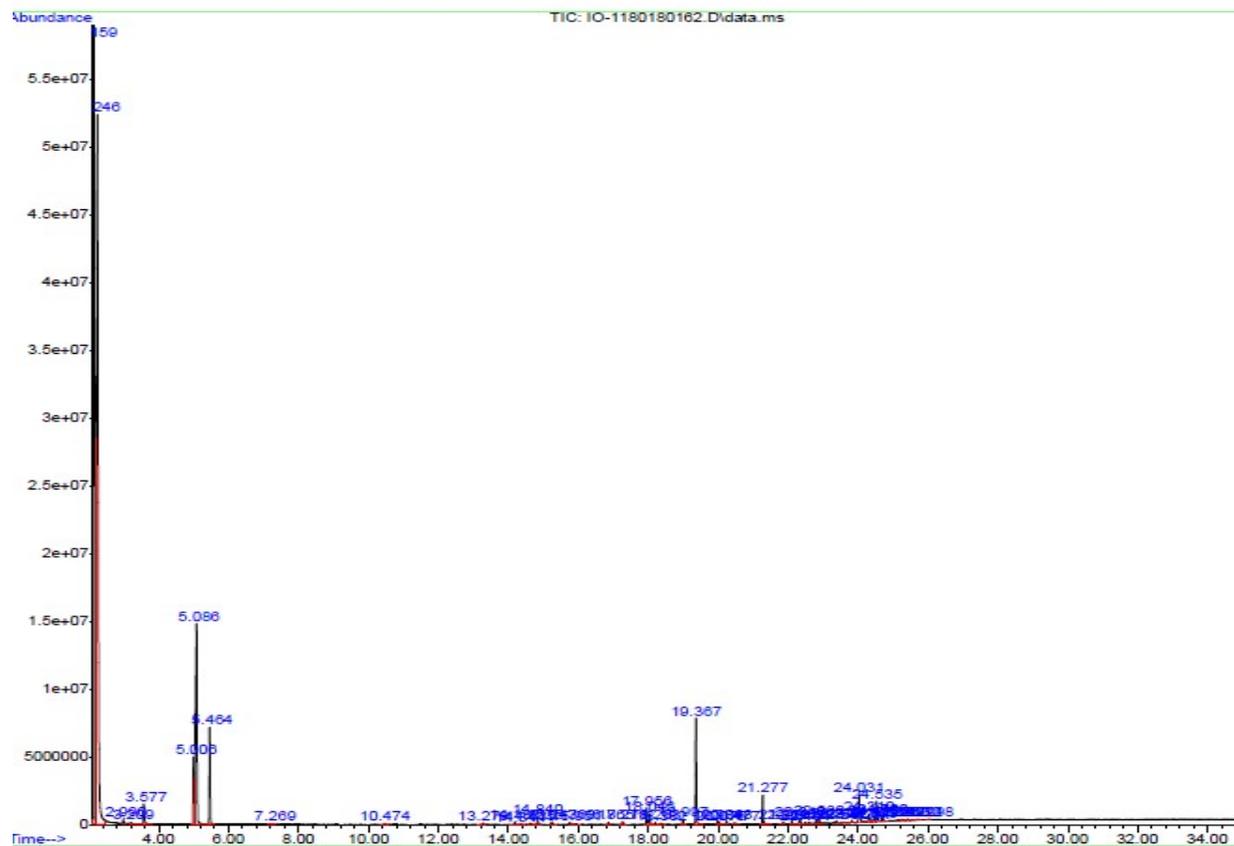


S Fig 3i: GC-MS Raw Chromatogram of An-Trans (P+L)-(N+Ph)

Sample : 10-TRANS-N
Misc :
ALS Vial : 3 Sample Multiplier: 1

Search Libraries: C:\Database\NIST02.L Minimum Quality: 0

Unknown Spectrum: Apex minus start of peak
Integration Events: ChemStation Integrator - autoint1.e

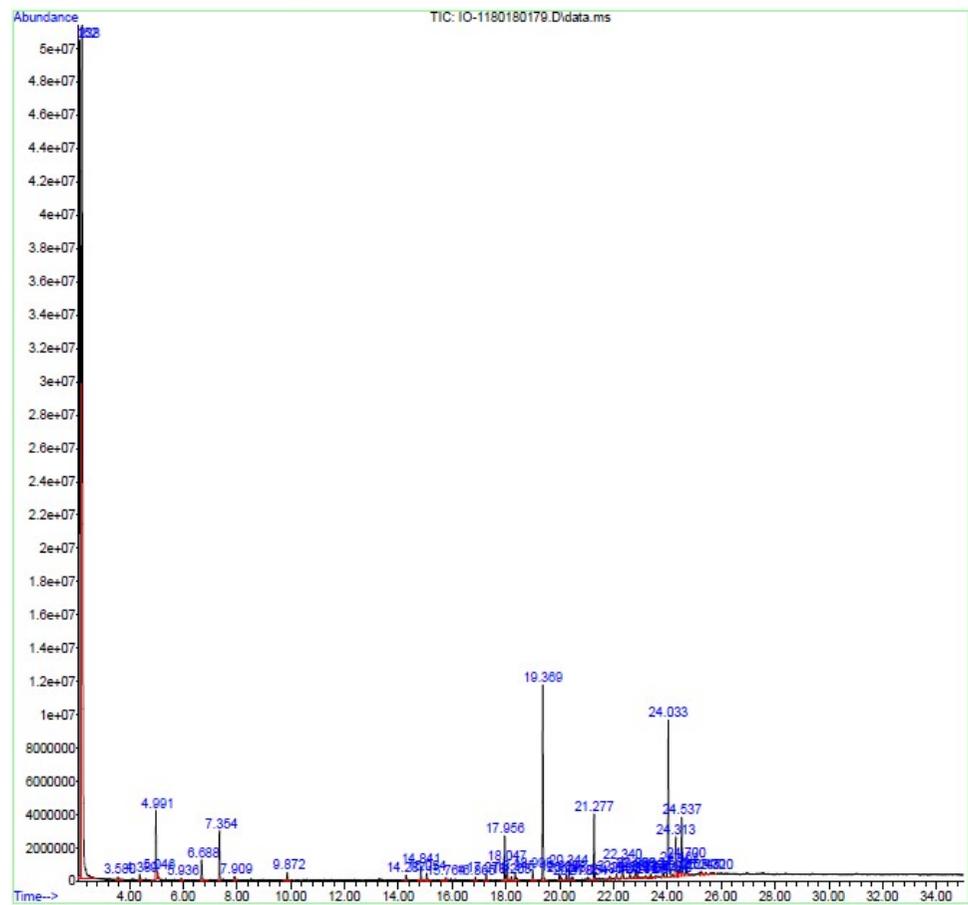


S Fig 3j: GC-MS Raw Chromatogram of An-Trans (P)-(N)

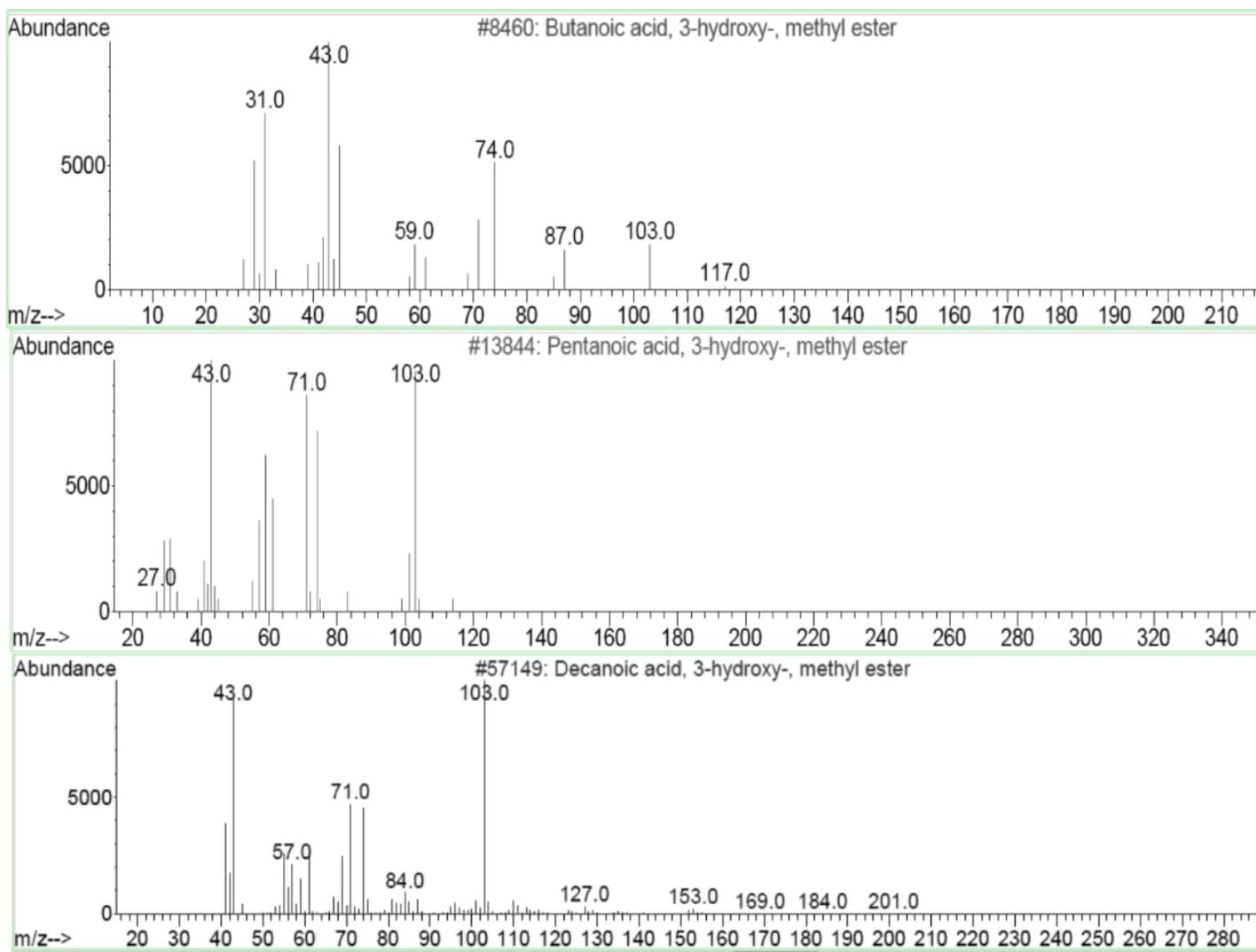
Sample : 12-TRANS-N
Misc :
ALS Vial : 4 Sample Multiplier: 1

Search Libraries: C:\Database\NIST02.L Minimum Quality: 0

Unknown Spectrum: Apex minus start of peak
Integration Events: ChemStation Integrator - autoint1.e



S Fig 3k: GC-MS Raw Chromatogram of An-Trans (P+L)-(N)



S Fig 4: Fragmentogram of the identified Alkyl groups as corresponding alkyl methyl esters in GCMS analysis

Table S 1: Back calculations for the relative absorbance of key functional group from FT-IR analysis

Condition	Transmittance % (C-H)	Transmittance % (C=O)	$A_{(C-H)} = -\log_{10}(T)$	$A_{(C=O)} = -\log_{10}(T)$
Ae (P)	90	41	0.045757491	0.387216
Ae (P+L)	82.007	58	0.086149075	0.236572
Trans Ae (P)	98.95	92.8	0.004584201	0.032452
Trans Ae (P+L)	92	75	0.036212173	0.124939
An (P)	93	85	0.031517051	0.070581
An (P+L)	98.7	94	0.005682847	0.026872
An- (P+L) -Biogas	96.5	92	0.015472687	0.036212
An-Trans (P+L)-(N+Ph)	97.74	92	0.009927665	0.036212
An-Trans (P+L)-(N)	96.29	82	0.016418813	0.086186
An-Trans (P)-(N)	90	65	0.045757491	0.187087
CO ₂	97.83	99.7	0.009527946	0.001

Table S 2: Prospective up-scale scenario of 25 l, with the assumptions

Process	Lab Scale Production	Process scale-up
Sterilisation	Auto-clave	In-tank Autoclaving (utilisation of the maximal efficiency)
Batch Cultivation	Bottles/Conical Flask	bio-reactor
Agitation	Magnetic stirrers	In-tank stirring
Illumination	Tube light	In tank LED lights with uniform illumination
Biomass Harvesting	Centrifuge	Filtration/Centrifuge
Drying	sunlight/oven	Vacuum drying/oven
Solvents	Evaporation	Recovery through distillation
Spent Biomass	Disposed	Multiproduct valorisation
Cell Disruption	Sonication	Sonication

Table S 3: Energy distribution for the lab scale and scale up production

Energy	Lab Scale Production (KWh)	Process scale-up 25 L (KWh)
Sterilisation	0.5	0.02 Kwh/L
Mixing	0.28	0.011
Light	1.108	0.044
Centrifuge	0.0583	0.0097
Sonicator	0.048	0.0016

The energy was calculated according to the power specifications of the instruments when used for its full efficiency for 25 L, Sterilisation (autoclave with 2kwh); Mixing (Magnetic stirrers with 2W); CFL lights (220 V and 80 mA specification for 6 days); Centrifuge (700 W); Sonicator (120 W).

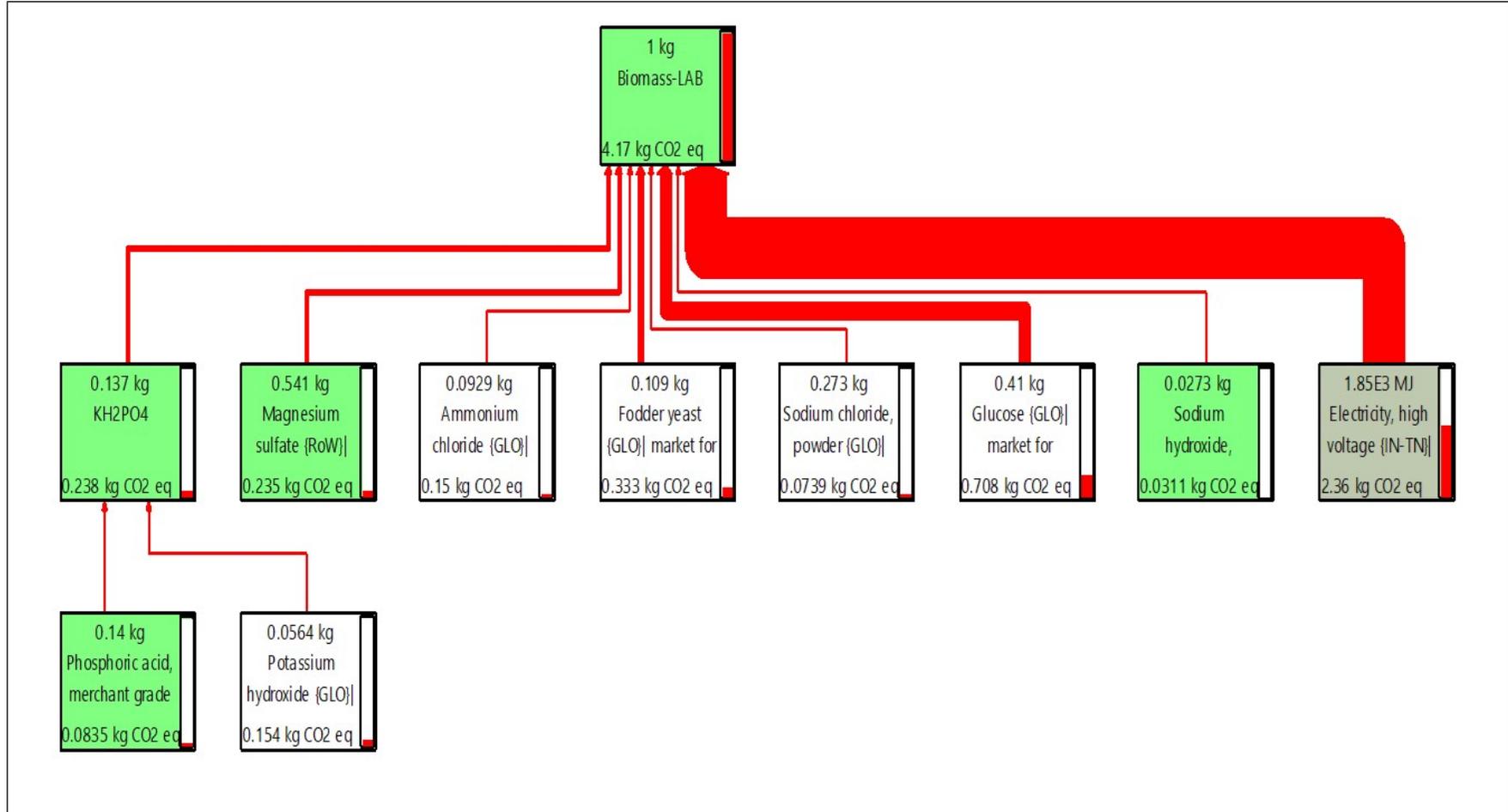
Table S 4: Unit specific Life Cycle Inventory for Lab scale and upscale production

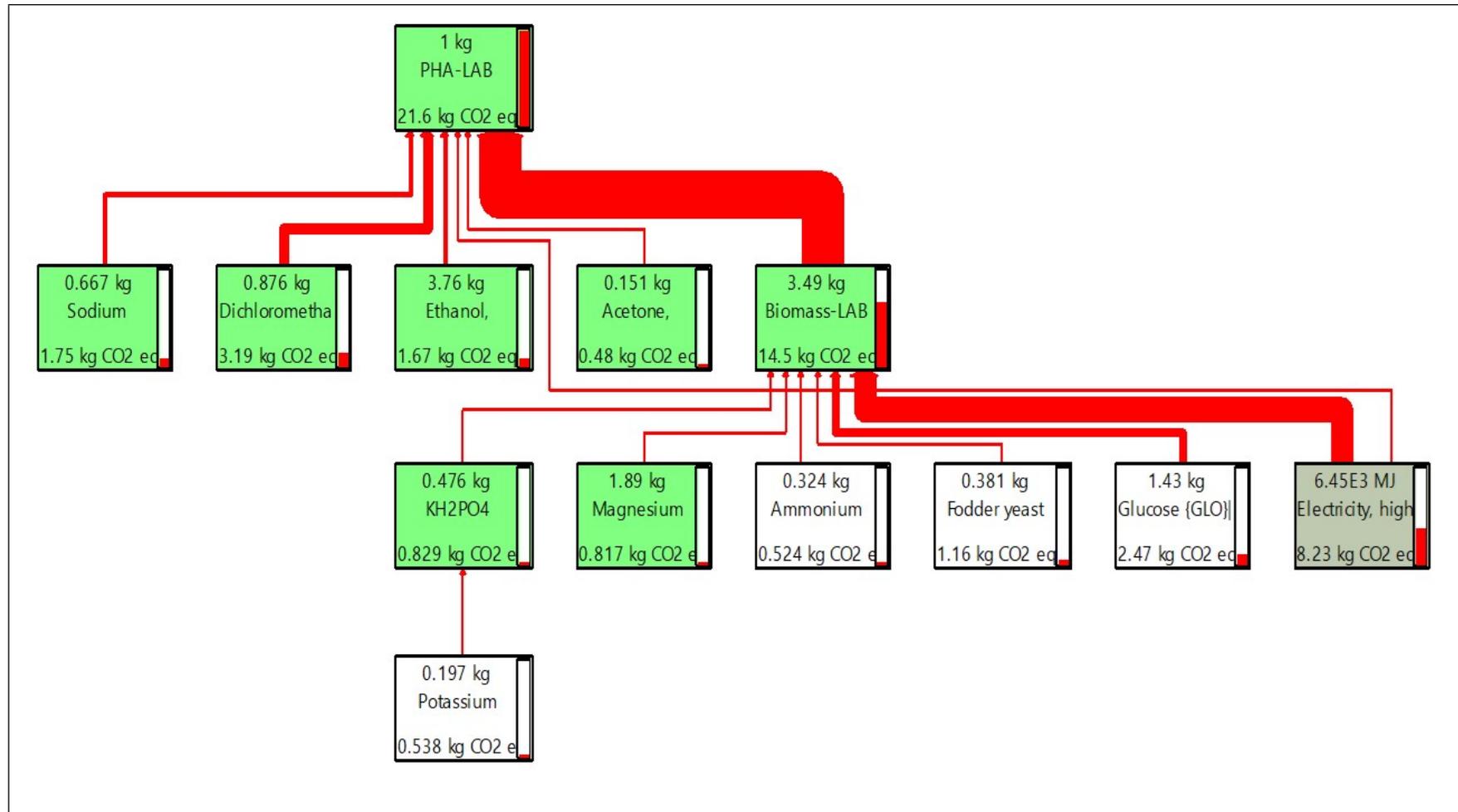
Lab Scale			Up-Scaled ¹		
BIOMASS PRODUCTION UNIT					
INPUTS	Unit	Value	INPUTS	Unit	Value
KH ₂ PO ₄	g	0.5	KH ₂ PO ₄	g	0.5
CaCl ₂ ·2H ₂ O	g	0.15	CaCl ₂ ·2H ₂ O	g	0.15
MgSO ₄ ·7H ₂ O	g	2	MgSO ₄ ·7H ₂ O	g	2
NH ₄ Cl	g	0.34	NH ₄ Cl	g	0.34
Yeast Extract	g	0.4	Yeast Extract	g	0.4
NaCl	g	1	NaCl	g	1
Ferric Citrate	g	0.005	Ferric Citrate	g	0.005
Sodium Pyruvate	g	3	Sodium Pyruvate	g	3

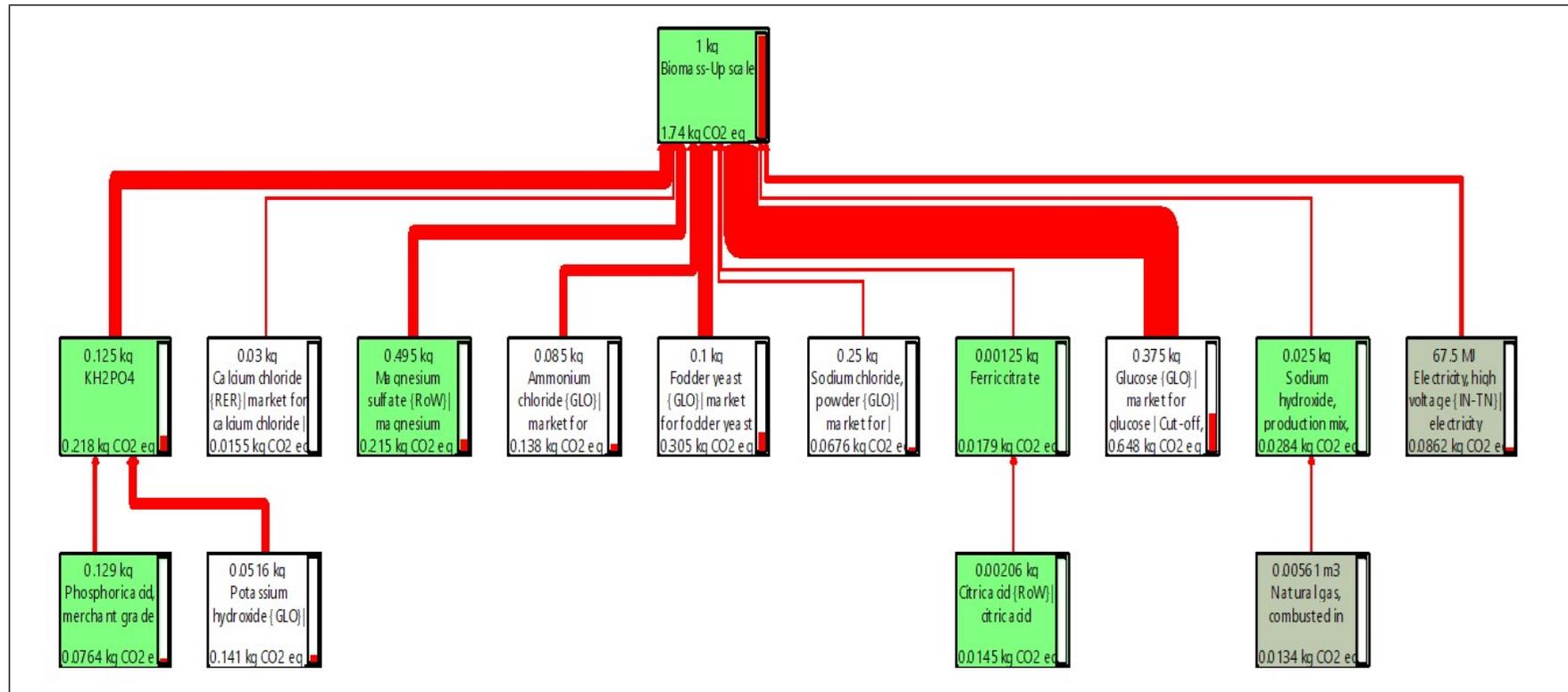
¹ The upscaled condition was assumed to be more efficient than a lab scale study under full sophisticated environmental conditions resulting to the biomass and PHA productivity in range of ± 0.4; the energy requirements are calculated based on the table S 3; the solvent recovery strategy is considered using distillation.

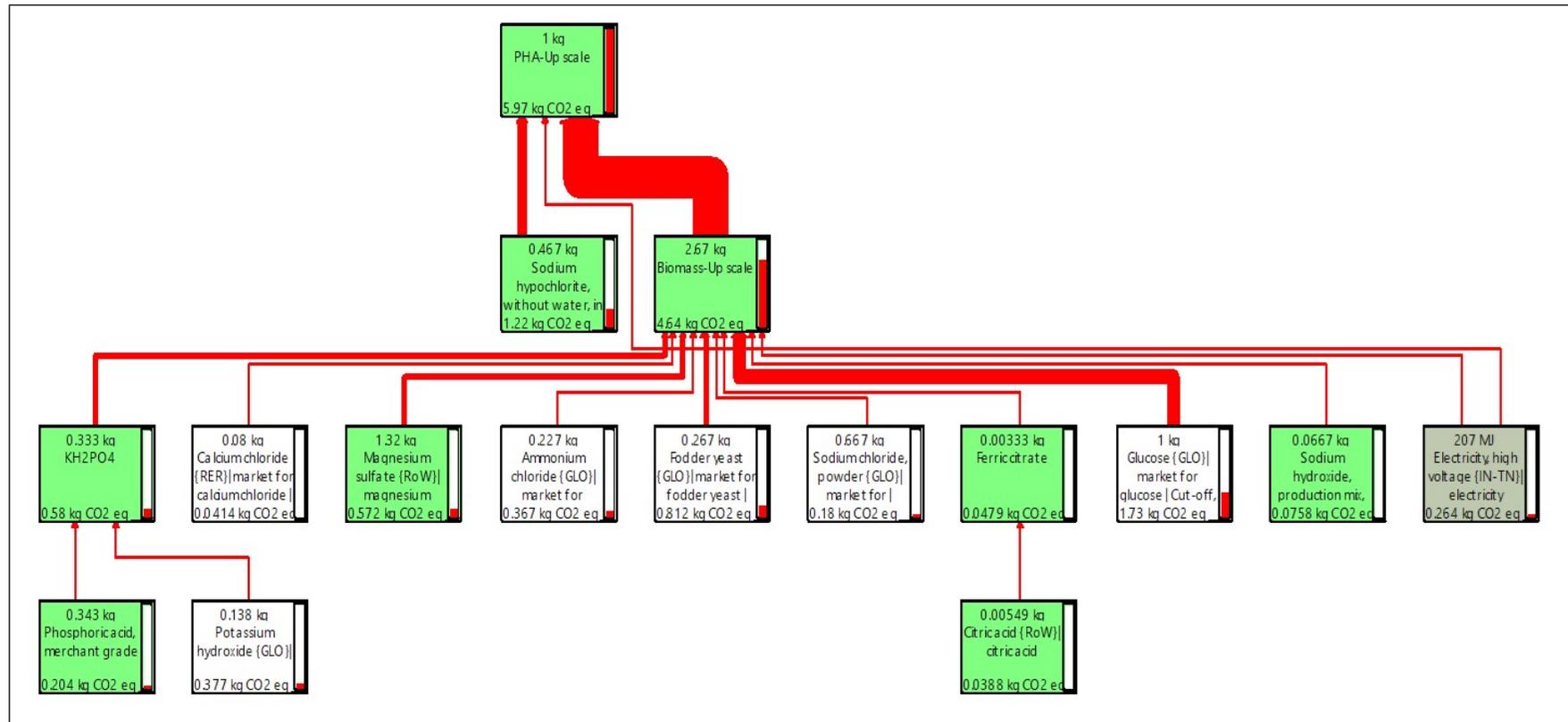
Water	L	1	Water	L	1
NaOH	g	1	NaOH	g	1
Energy	kWh	1.88	Energy	kWh	0.075
OUTPUTS	Unit	Value	OUTPUTS	Unit	Value
Biomass	g	3.66	Biomass	g	4
Wastewater	L	0.8	Wastewater	L	0
POLYMER DOWNSTREAMING UNIT					
INPUTS	Unit	Value	INPUTS	Unit	Value
Biomass	g	3.66	Biomass	g	4
NaOCl	g	0.7	NaOCl	g	0.7
DCM	g	0.92	DCM	g	0.01
Ethanol	g	3.945	Ethanol	g	0.1
Acetone	g	0.158	Acetone	g	0.01
Energy	kWh	0.106	Energy	kWh	0.0113
OUTPUTS	Unit	Value	OUTPUTS	Unit	Value
PHA	g	1.05	PHA	g	1.5
Wastewater	L	0.01	Wastewater	L	0.01
DCM Vapour	g	0.92	DCM Vapour	g	0.01
Ethanol Vapour	g	3.945	Ethanol Vapour	g	0.001

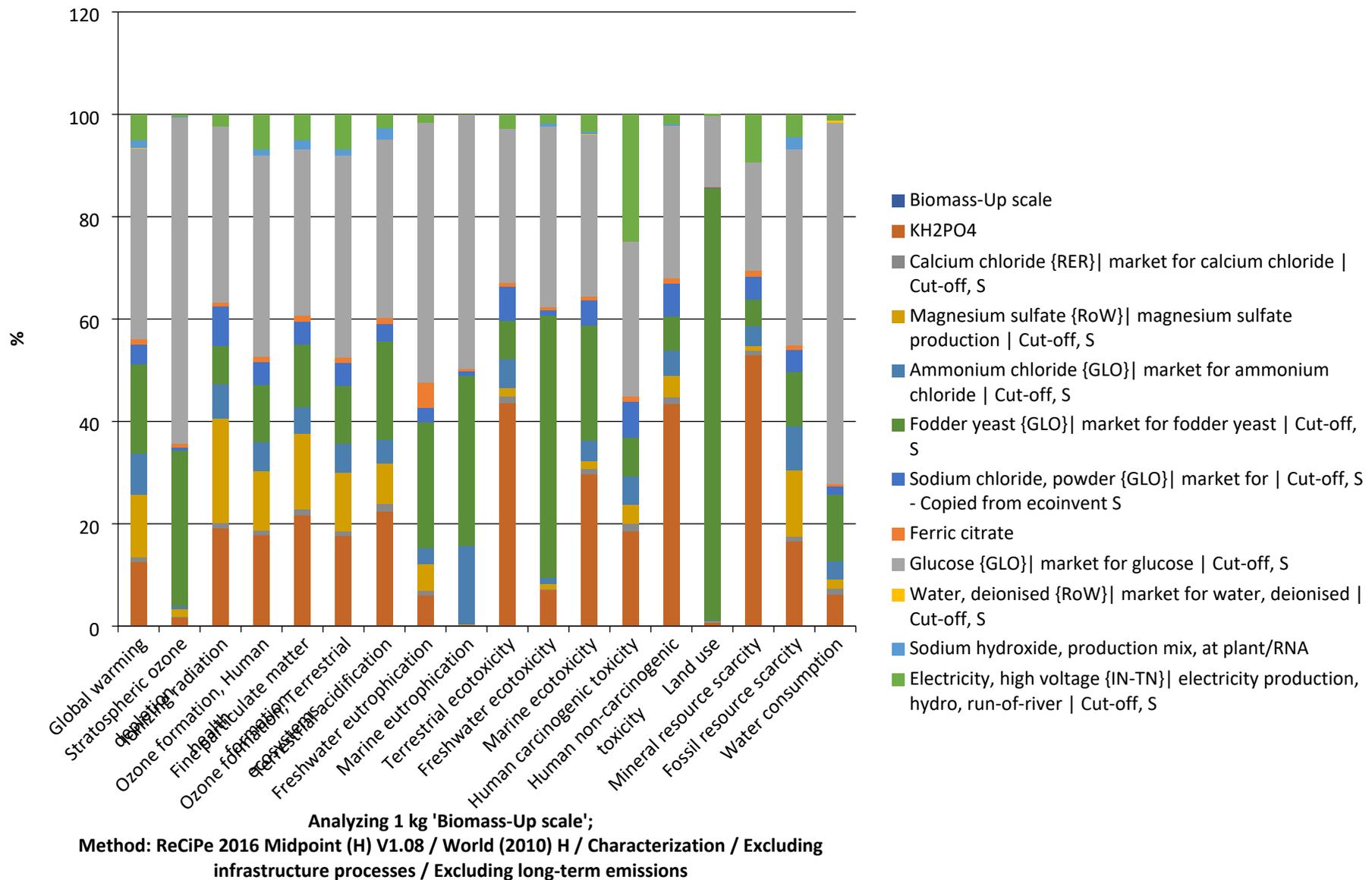
Table S5: CO₂ eq. emission from each unit process, Sankey representation of the contributory elements

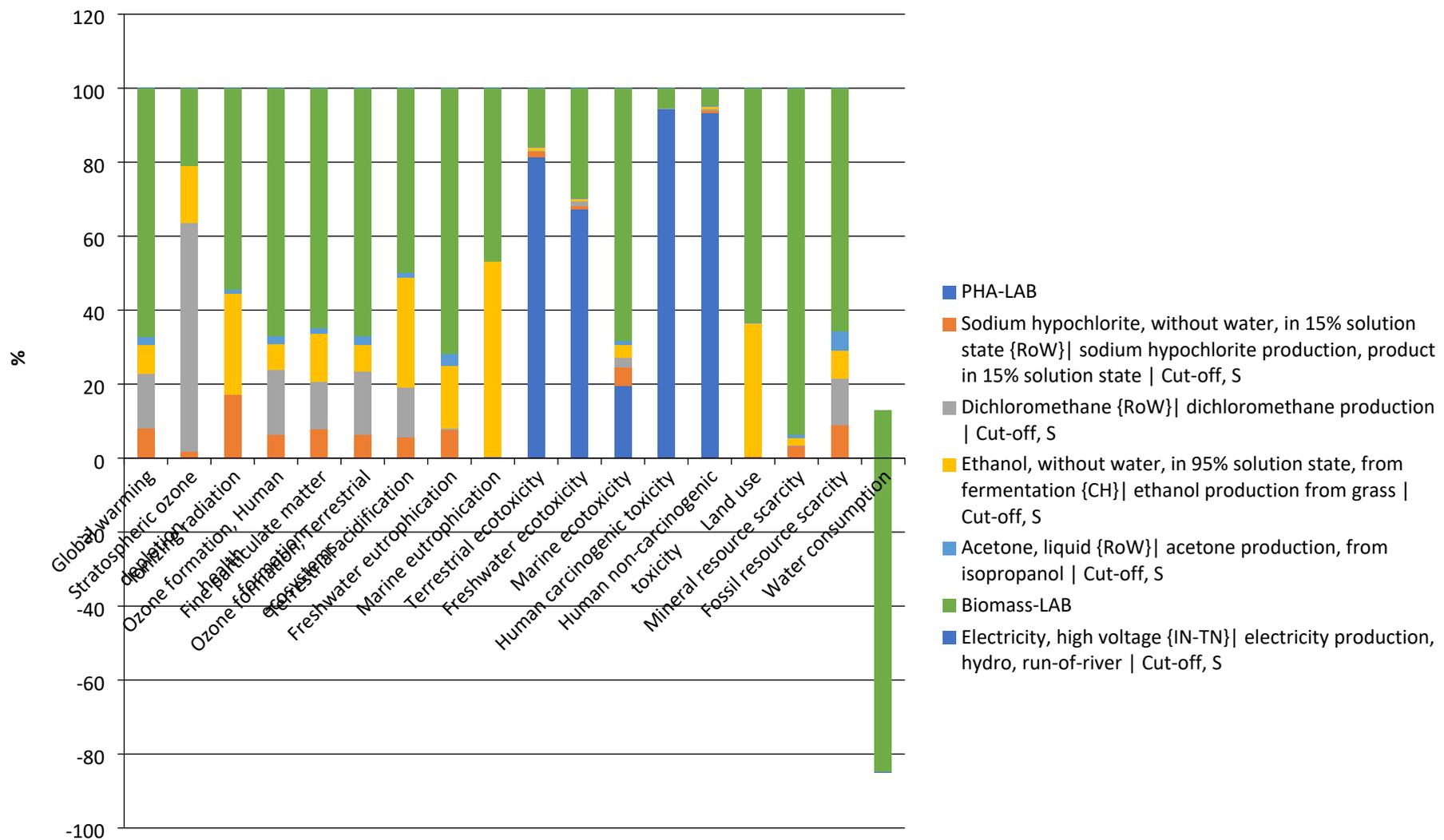




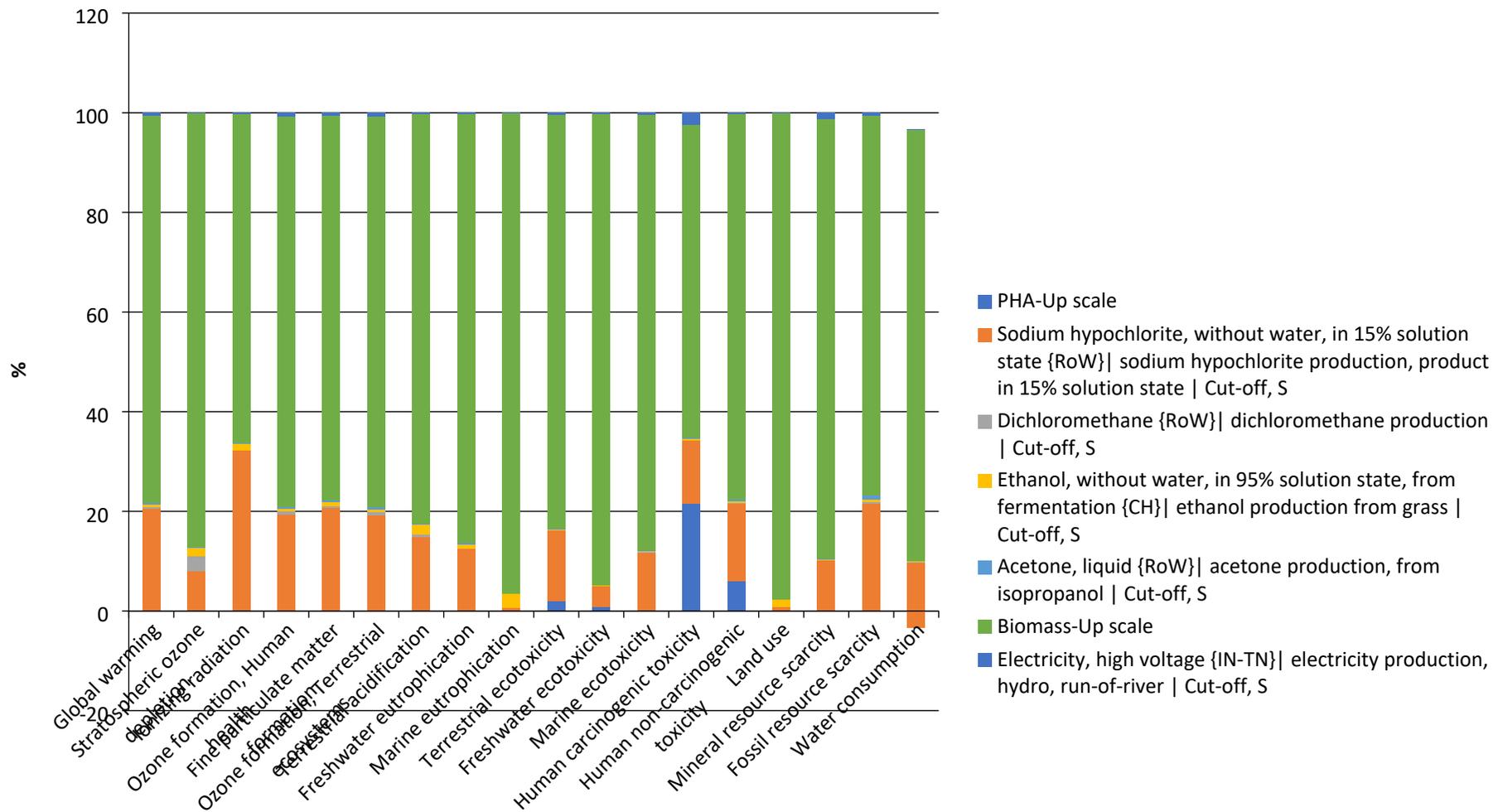








Analyzing 1 kg 'PHA-LAB';
Method: ReCiPe 2016 Midpoint (H) V1.08 / World (2010) H / Characterization / Excluding infrastructure processes / Excluding long-term emissions



Analyzing 1 kg 'PHA-Up scale';
Method: ReCiPe 2016 Midpoint (H) V1.08 / World (2010) H / Characterization / Excluding infrastructure processes / Excluding long-term emissions