

Supporting Information

Membrane lipid composition directs the cellular selectivity of antimicrobial metallohelices

Nicola J. Rogers,^{*a,b} Miles L. Postings,^b Ann M. Dixon,^b John Moat,^c Georgia Shreeve,^b Louise Stuart,^b Nick Waterfield,^d Peter Scott.^b

^a Department of Chemistry, Hong Kong Baptist University, Kowloon Tong, Hong Kong

^b Department of Chemistry, University of Warwick, Coventry CV4 7AL, UK

^c School of Life Sciences, University of Warwick, Gibbet Hill Campus, Coventry, UK

^d Warwick Medical School, University of Warwick, Coventry, UK

Contents

57Fe Compound characterisation data	2
Antimicrobial bactericidal data.....	10
Antimicrobial activity and cellular accumulation studies of ⁵⁷ Fe-metallohelices	10
DLS Data – Effect of metallohelices on model vesicle size.....	11
DLS data– Comparing the effects of enantiomers of 1	15
Zeta Potential Data – Effect of metallohelices on model vesicle surface charge	16
Fitting Zeta potential data – Binding constants	17

⁵⁷Fe Compound characterisation data

$\Delta_{\text{Fe}}\text{-}[\text{}^{57}\text{Fe}_2(\text{R,R-}\text{L}^1)_3]\text{Cl}_4\cdot 6\text{H}_2\text{O}$ ($\Delta\text{-1}'$)

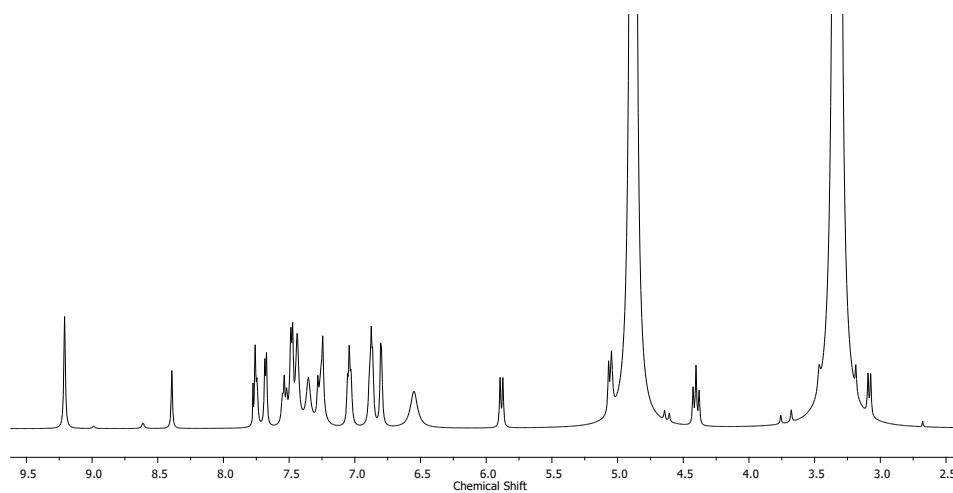
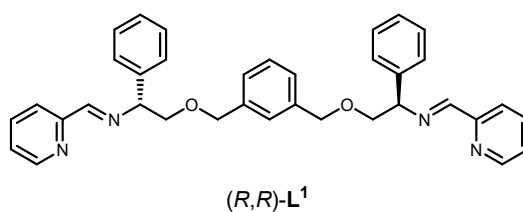
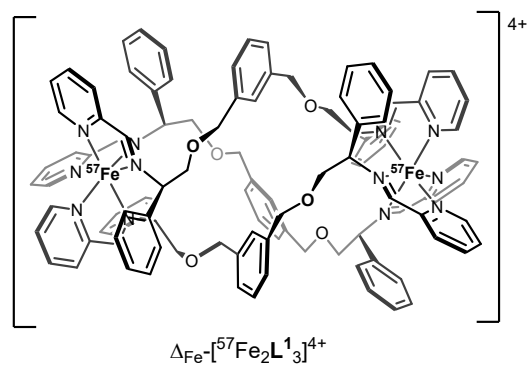


Figure S1. ^1H NMR (500 MHz, MeOD, 298 K) of $\Delta\text{-1}'$.

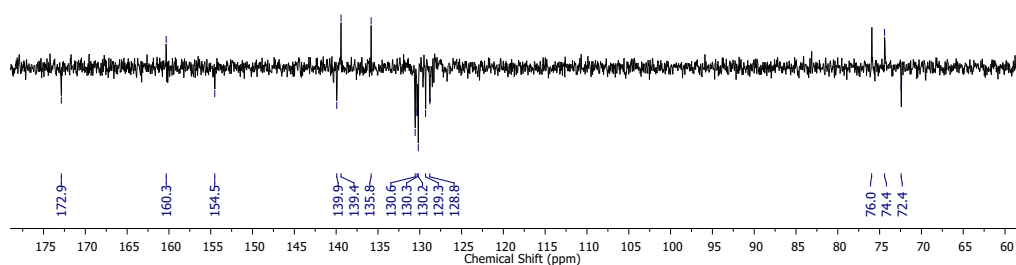


Figure S2. $^{13}\text{C}\{^1\text{H}\}$ APT NMR (126 MHz, MeOD, 298 K) of $\Delta\text{-1}'$.

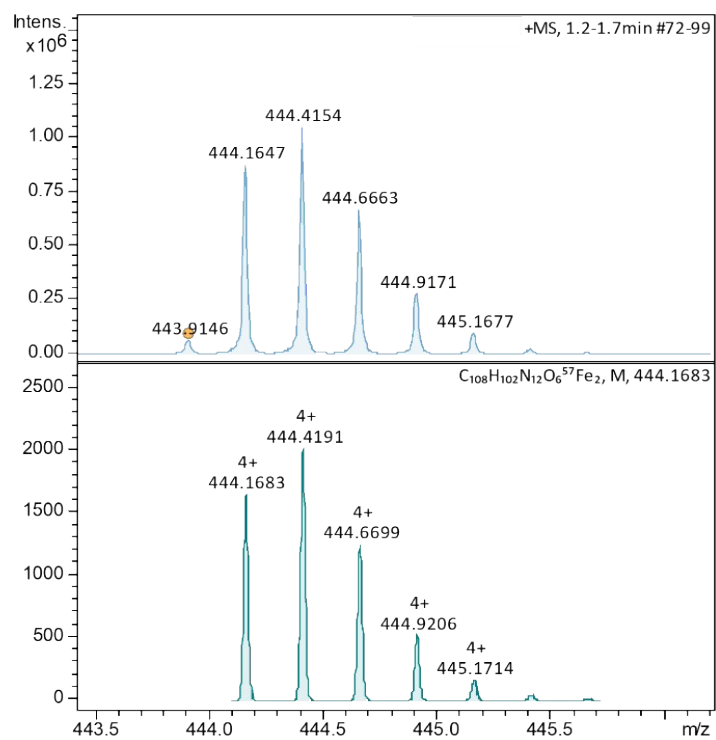


Figure S3. HRMS (+) of $\Delta-1'$, predicted (lower) and measured (upper) spectra.

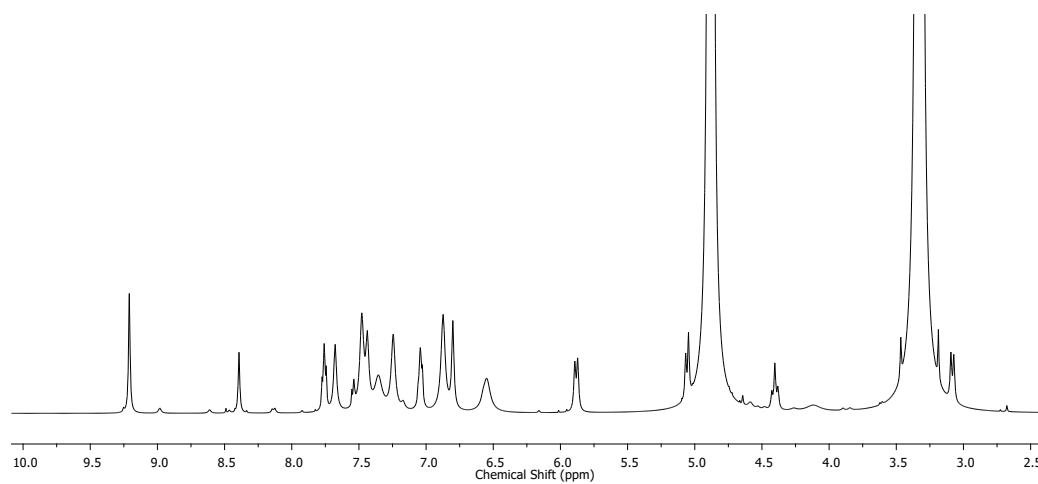
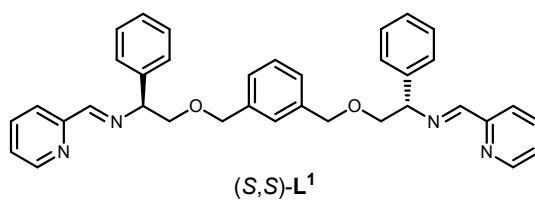
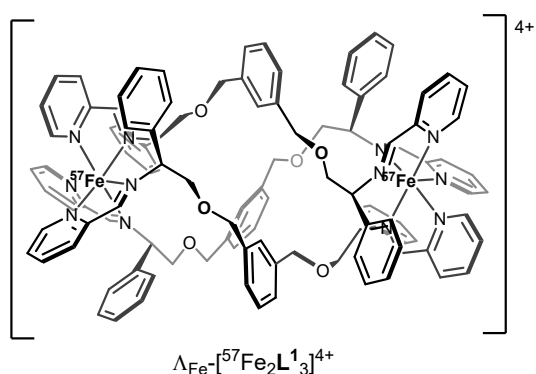
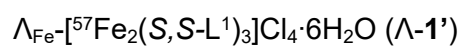


Figure S4. ^1H NMR (500 MHz, MeOD, 298 K) of $\Lambda\text{-1'}$.

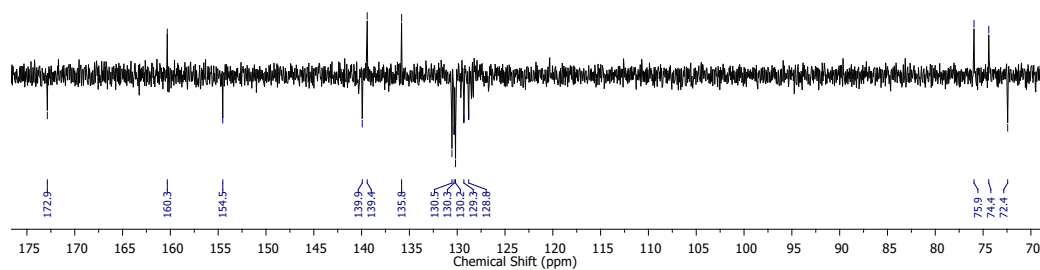


Figure S5. $^{13}\text{C}\{^1\text{H}\}$ APT NMR (126 MHz, MeOD, 298 K) of $\Lambda\text{-1'}$.

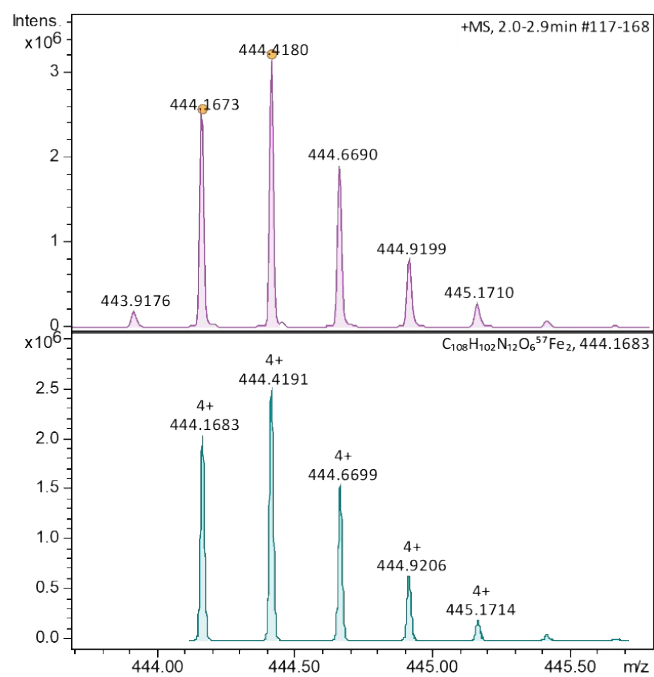


Figure S6. HRMS (+) of $\Lambda-1'$, predicted (lower) and measured (upper) spectra.

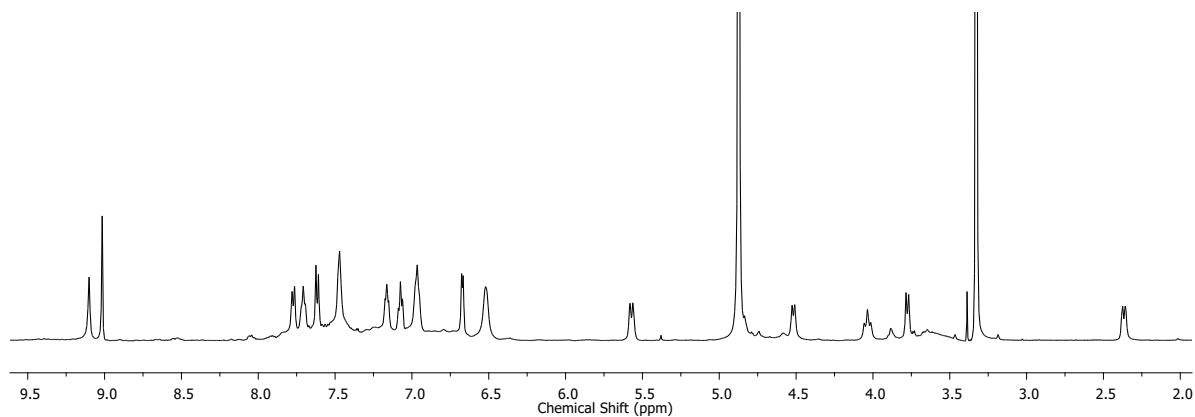
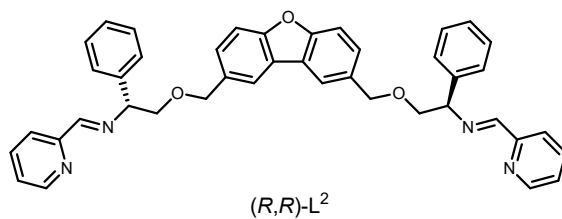
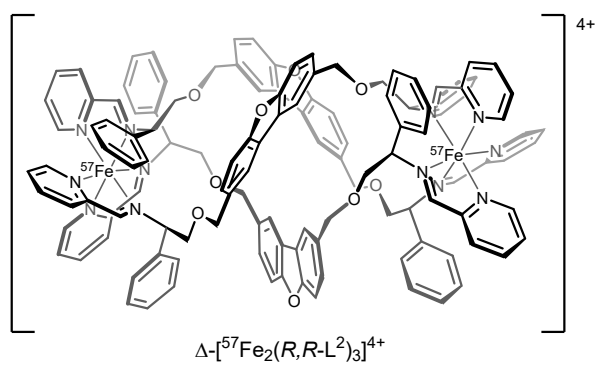
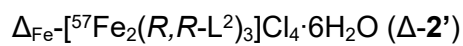


Figure S7. ^1H NMR (500 MHz, MeOD, 298 K) of $\Delta\text{-2'}$.

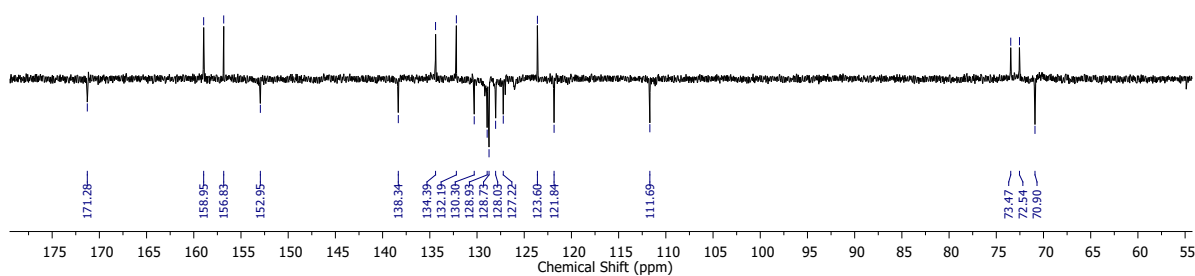


Figure S8. $^{13}\text{C}\{^1\text{H}\}$ APT NMR (126 MHz, MeOD, 298 K) of $\Delta\text{-2'}$.

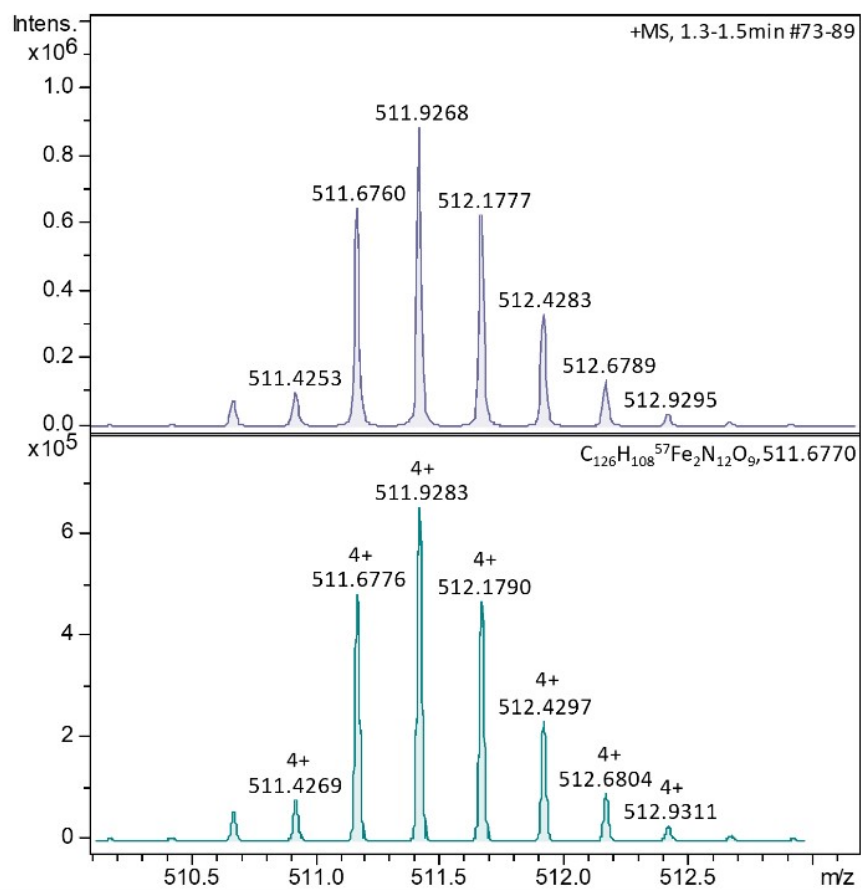


Figure S9. HRMS (+) of $\Delta-2'$, predicted (lower) and measured (upper) spectra.

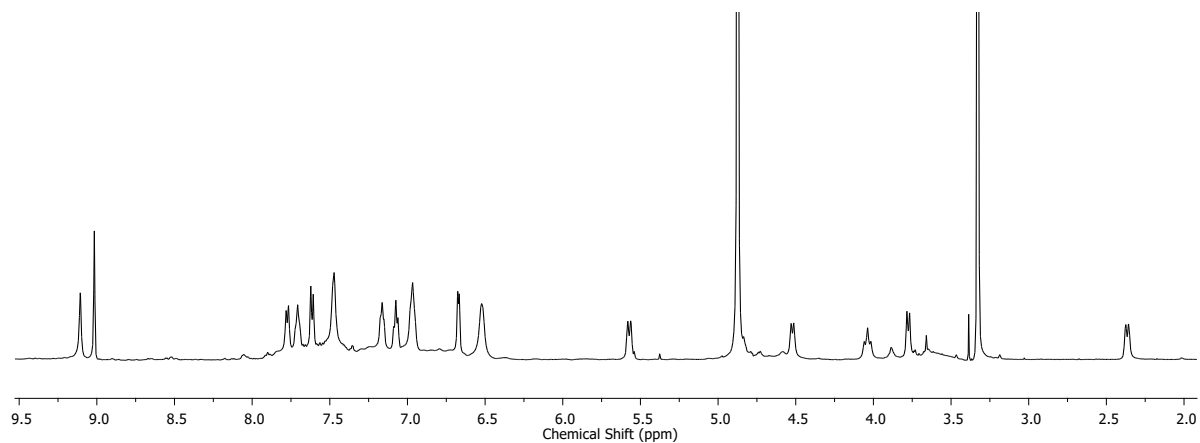
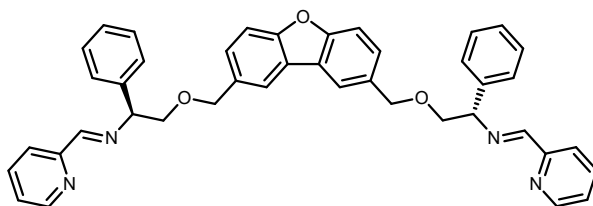
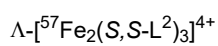
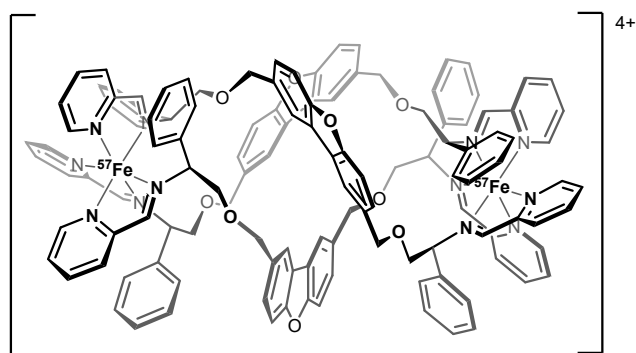
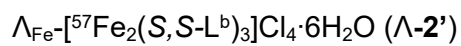


Figure S10. ^1H NMR (500 MHz, MeOD, 298 K) of $\Lambda\text{-2'}$.

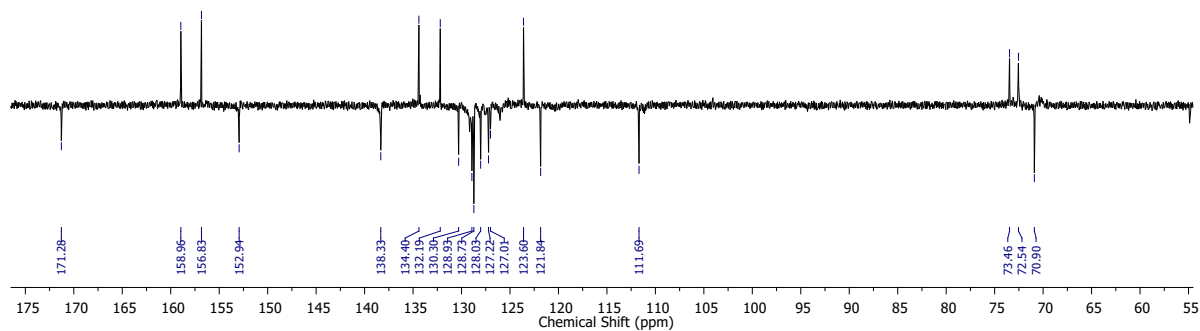


Figure S11. $^{13}\text{C}\{^1\text{H}\}$ APT NMR (126 MHz, MeOD, 298 K) of $\Lambda\text{-2'}$.

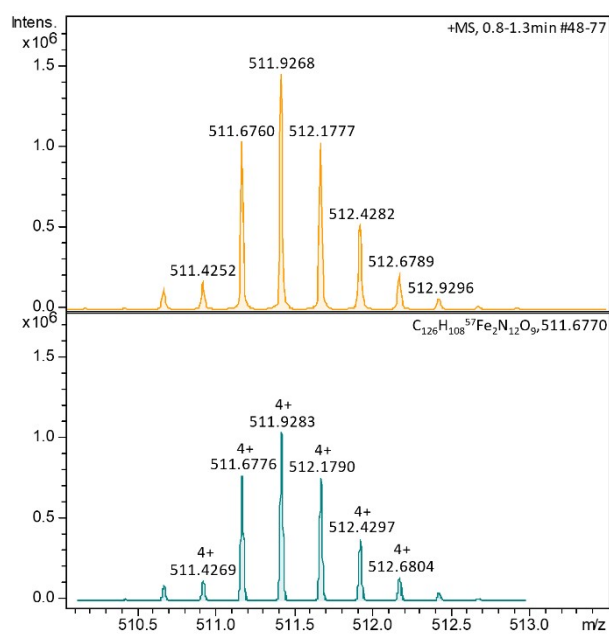


Figure S12. HRMS (+) of $\Lambda\text{-2'}$, predicted (lower) and measured (upper) spectra.

Antimicrobial bactericidal data

Table S1. Minimum bactericidal concentrations against Gram-negative – *E. coli* and Gram-positive – *S. aureus* bacteria, determined for Λ -1, Δ -1, Λ -2, and Δ -2.

compound	MBC/ $\mu\text{g mL}^{-1}$ (μM)	
	<i>E. coli</i> ATCC 25922	<i>S. aureus</i> 27853
Λ -1	8 (3.9)	32 (15.7)
Δ -1	32 (15.7)	64 (31.3)
Λ -2	64 (26.4)	8 (3.3)
Δ -2	64 (26.4)	8 (3.3)

Antimicrobial activity and cellular accumulation studies of ^{57}Fe -metallohelices

Table S2. Iron (^{57}Fe) cellular accumulation (ng ^{57}Fe per 10^8 cells) in *S. aureus* USA300 bacteria when dosed with at 8 mg mL^{-1} , MIC, and 0.5×MIC concentrations for 20 h.

Compound	MIC / $\mu\text{g mL}^{-1}$	Cellular accumulation of ^{57}Fe / ng per 10^8 cells		
		dose = 8 $\mu\text{g mL}^{-1}$	dose = MIC	dose = 0.5MIC
Λ -1'	16	10.9±0.2	23.1±0.2	11.9±0.3
Δ -1'	16	10.0±0.2	19.0±0.3	9.55±0.23
Λ -2'	2	186±6	47.7±1.4	25.0±0.8
Δ -2	2	195±1	51.3±1.5	25.6±0.6

Table S3. Iron (^{57}Fe) cellular accumulation (ng ^{57}Fe per 10^8 cells) in *E. coli* TOP10 bacteria when dosed with at 8 mg mL^{-1} , MIC, and 0.5×MIC concentrations for 20 h.

Compound	MIC / $\mu\text{g mL}^{-1}$	Cellular accumulation of ^{57}Fe / ng per 10^8 cells		
		dose = 8 $\mu\text{g mL}^{-1}$	dose = MIC	dose = 0.5MIC
Λ -1'	2	138±4	36.0±2.2	18.3±0.2
Δ -1'	4	77.3±4.3	37.2±2.2	18.8±0.1
Λ -2'	16	90.4±21.9	319±18	163±4
Δ -2'	16	98.1±8.4	335±12	178±7

DLS Data – Effect of metallohelices on model vesicle size

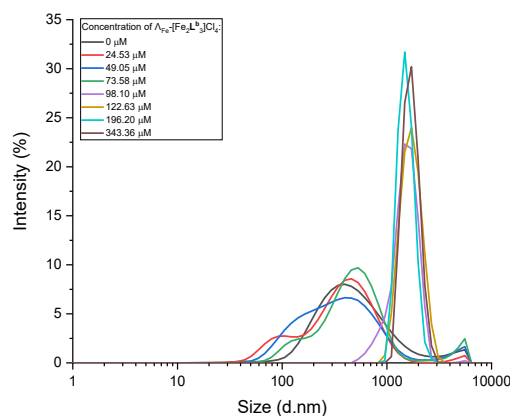


Figure S13 An overlay of the mean size (d/nm) distributions with regards to the scattering intensity (%) of *E. coli* cytosolic membrane-mimetic vesicles (~ 0.4 mM lipid) when 0 – 343 μ M of Λ -1 was added.

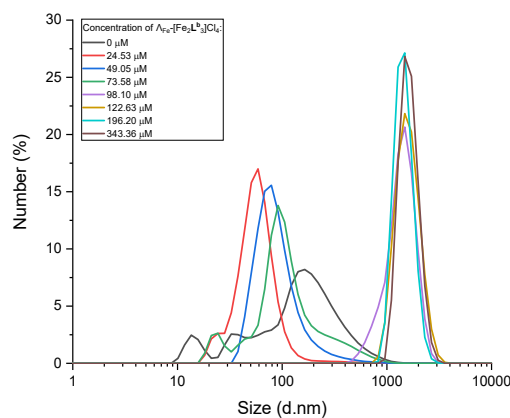


Figure S14 An overlay of the mean size (d/nm) distributions with regards to the calculated number distribution (%) of *E. coli* cytosolic membrane-mimetic vesicles (~ 0.4 mM lipid) when 0 – 343 μ M of Λ -1 was added.

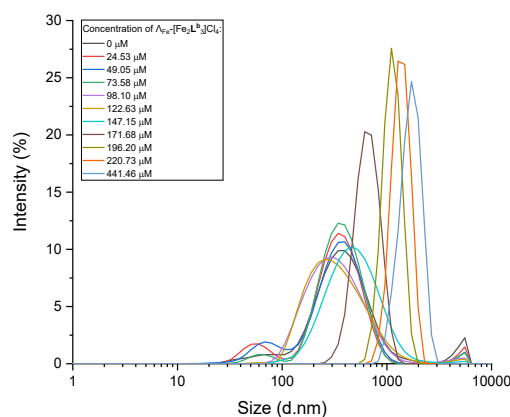


Figure S15 An overlay of the mean size (d/nm) distributions with regards to the scattering intensity (%) of *S. aureus* cytosolic membrane-mimetic vesicles (~ 0.4 mM lipid) when 0 – 441 μ M of Λ -1 was added.

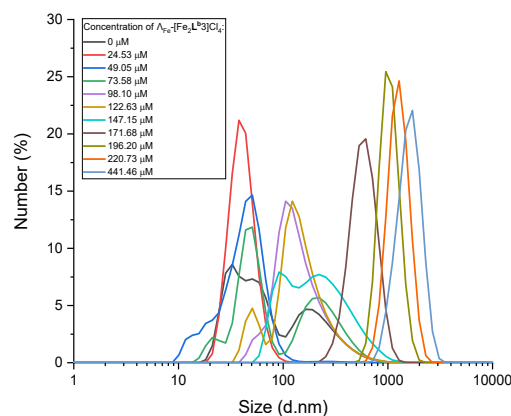


Figure S16 An overlay of the mean size (d/nm) distributions with regards to the calculated number intensity (%) of *S. aureus* cytosolic membrane-mimetic vesicles (~0.4 mM lipid) when 0 – 441 μM of Λ -1 was added.

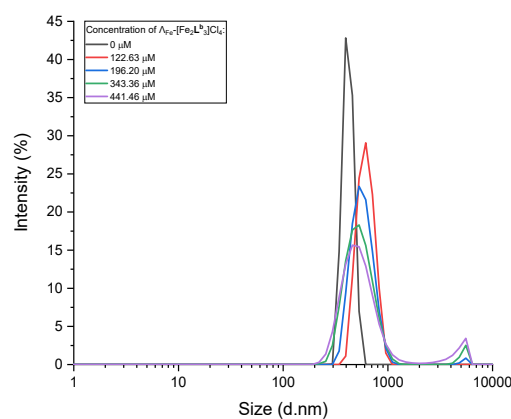


Figure S17 An overlay of the mean size (d/nm) distributions with regards to the scattering intensity (%) of mammalian plasma membrane-mimetic vesicles (~0.4 mM lipid) when 0 – 441 μM of Λ -1 was added.

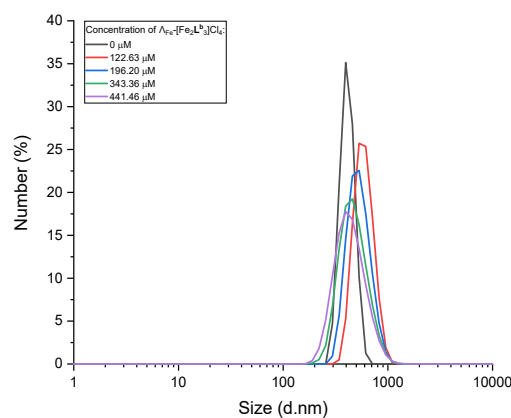


Figure S18 An overlay of the mean size (d/nm) distributions with regards to the calculated number distribution (%) of mammalian plasma membrane-mimetic vesicles (~0.4 mM lipid) when 0 – 441 μM of Λ -1 was added.

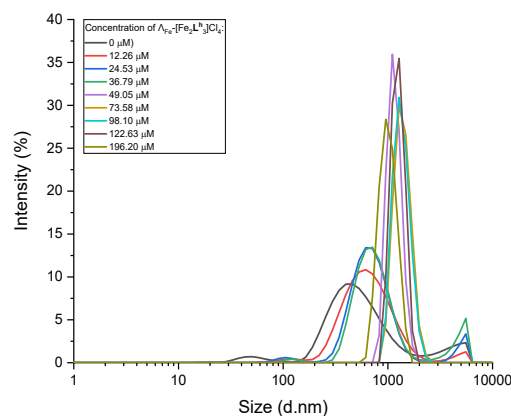


Figure S19 An overlay of the mean size (d/nm) distributions with regards to the scattering intensity (%) of *E. coli* cytosolic membrane-mimetic vesicles (~0.4 mM lipid) when 0 – 196 μ M of Λ -2 was added.

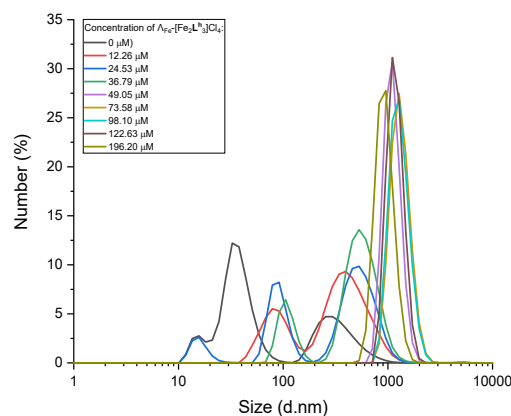


Figure S20 An overlay of the mean size (d/nm) distributions with regards to the calculated number distribution (%) of *E. coli* cytosolic membrane-mimetic vesicles (~0.4 mM lipid) when 0 – 196 μ M of Λ -2 was added.

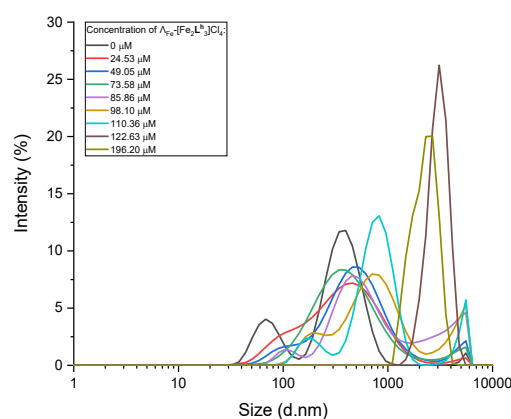


Figure S21 An overlay of the mean size (d/nm) distributions with regards to the scattering intensity (%) of *S. aureus* cytosolic membrane-mimetic vesicles (~0.4 mM lipid) when 0 – 196 μ M of Λ -2 was added.

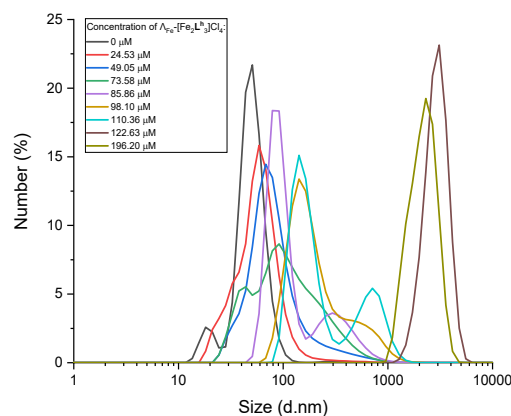


Figure S22 An overlay of the mean size (d/nm) distributions with regards to the calculated number distribution (%) of *S. aureus* cytosolic membrane-mimetic vesicles (~0.4 mM lipid) when 0 – 196 μM of Λ -2 was added.

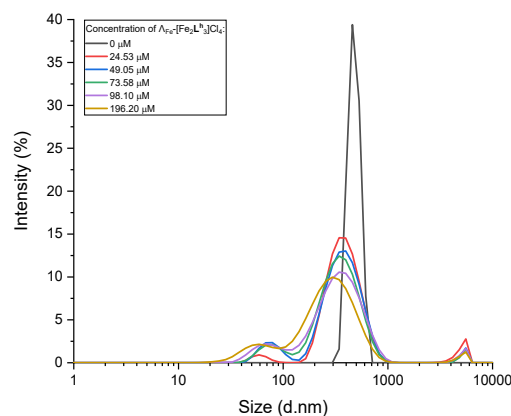


Figure S23 An overlay of the mean size (d/nm) distributions with regards to the scattering intensity (%) of mammalian plasma membrane-mimetic vesicles (~0.4 mM lipid) when 0 – 196 μM of Λ -2 was added.

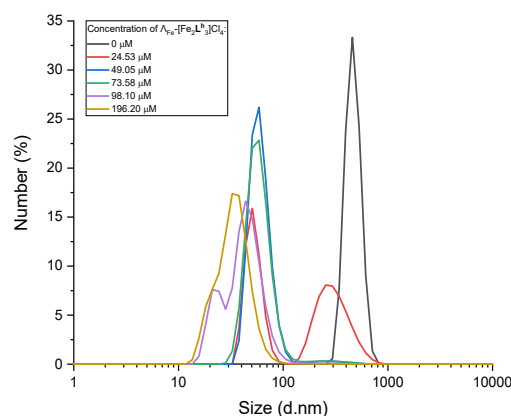


Figure S24 An overlay of the mean size (d/nm) distributions with regards to the calculated number distribution (%) of mammalian plasma membrane-mimetic vesicles (~0.4 mM lipid) when 0 – 196 μM of Λ -2 was added.

DLS data– Comparing the effects of enantiomers of 1

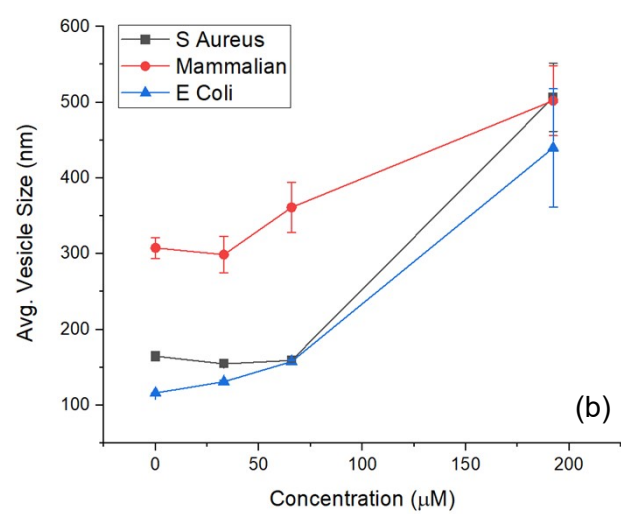
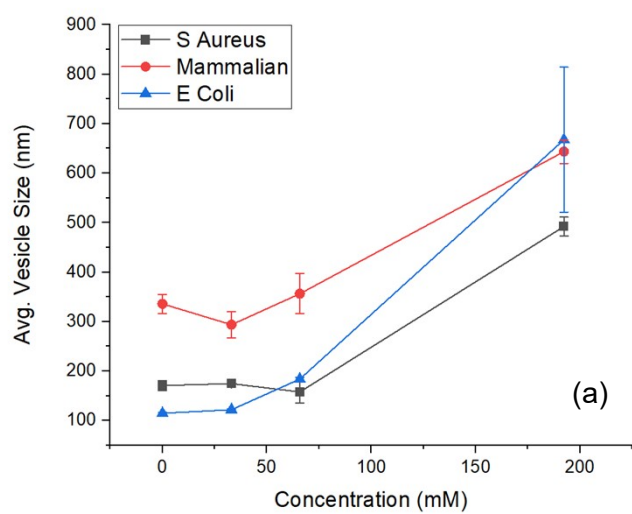


Figure S25 The mean Z-average size of membrane-mimetic vesicles (~0.6 mM lipid) when 0 – 192 μ M of Λ -1 (a) or Δ -1 (b) was added.

Similar effects on the vesicle size data was observed upon titration of Λ -1 and Δ -1.

Zeta Potential Data – Effect of metalloheliices on model vesicle surface charge

Table S4. *Bacteria model vesicles.* Zeta potential measurements of 0.5 mg mL⁻¹ unilamellar vesicles ([lipid] ~ 0.6 mM) plus 0-192 mM metalloheliix in sodium phosphate buffer (25 mM, pH 7.4).

	Zeta Potential / mV					
	<i>E. coli mimetic unilamellar vesicles</i>			<i>S. aureus mimetic unilamellar vesicles</i>		
[MH] / μ M	Λ -1	Δ -1	Λ -2	Λ -1	Δ -1	Λ -2
0	-50.2 $\pm$ 1.3	-52.3 $\pm$ 1.9	-48.5 $\pm$ 1.0	-64.1 $\pm$ 1.2	-64.1 $\pm$ 0.4	-62.8 $\pm$ 0.4
33	-25.1 $\pm$ 1.1	-26.8 $\pm$ 1.2	-28.0 $\pm$ 0.6	-52.4 $\pm$ 2.8	-53.9 $\pm$ 3.2	-50.8 $\pm$ 2.1
66	-19.8 $\pm$ 0.4	-17.0 $\pm$ 0.4	-8.4 $\pm$ 3.5	-46.4 $\pm$ 2.0	-41.5 $\pm$ 1.2	-47.3 $\pm$ 1.4
192	-10.8 $\pm$ 0.2	-12.9 $\pm$ 0.3	-8.3 $\pm$ 0.3	-27.6 $\pm$ 3.9	-21.2 $\pm$ 0.5	+6.9 $\pm$ 0.3

Table S5. *Mammalian model vesicles.* Zeta potential measurements of 0.5 mg mL⁻¹ unilamellar vesicles ([lipid] ~ 0.6 mM) plus 0-192 mM metalloheliix in sodium phosphate buffer (25 mM, pH 7.4).

	Zeta Potential / mV		
	<i>Mammalian mimetic unilamellar vesicles</i>		
[MH] / μ M	Λ -1	Δ -1	Λ -2
0	-5.7 $\pm$ 0.1	-8.1 $\pm$ 0.4	-5.5 $\pm$ 0.3
33	-4.7 $\pm$ 0.3	-6.2 $\pm$ 0.7	+14.7 $\pm$ 0.4
66	-1.5 $\pm$ 0.1	-5.0 $\pm$ 0.4	+18.3 $\pm$ 0.8
192	+4.6 $\pm$ 0.5	+2.0 $\pm$ 0.9	+21.3 $\pm$ 0.7

Fitting Zeta potential data – Binding constants

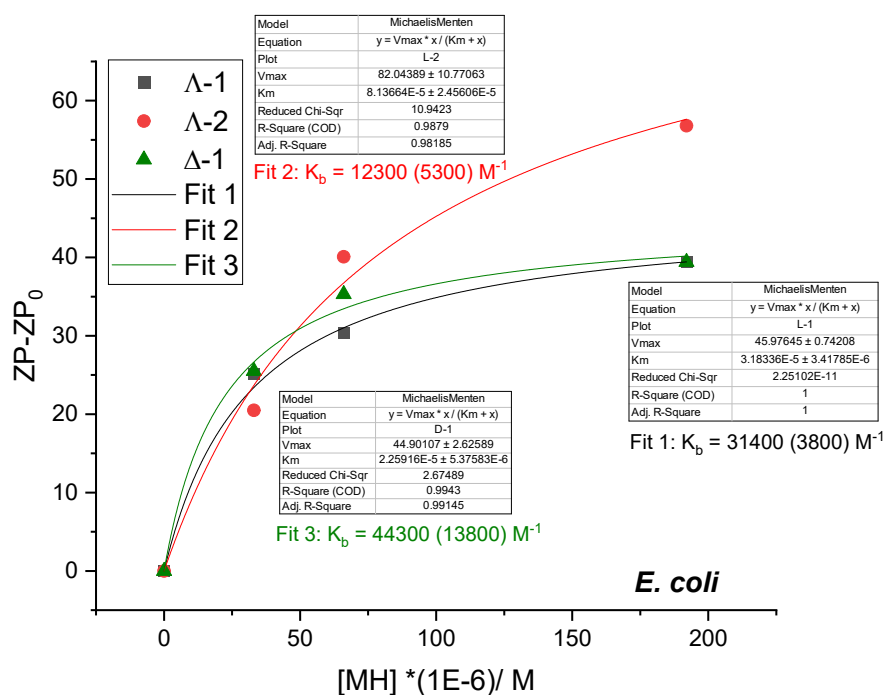


Figure S26 Fitting Zeta potential data. Zeta potential measurements of 0.5 mg mL^{-1} unilamellar *E. coli* vesicles ([lipid] $\sim 0.6 \text{ mM}$), following the addition of 0-192 mM Λ -1, Λ -2, and Δ -1, in sodium phosphate buffer (25 mM, pH 7.4).

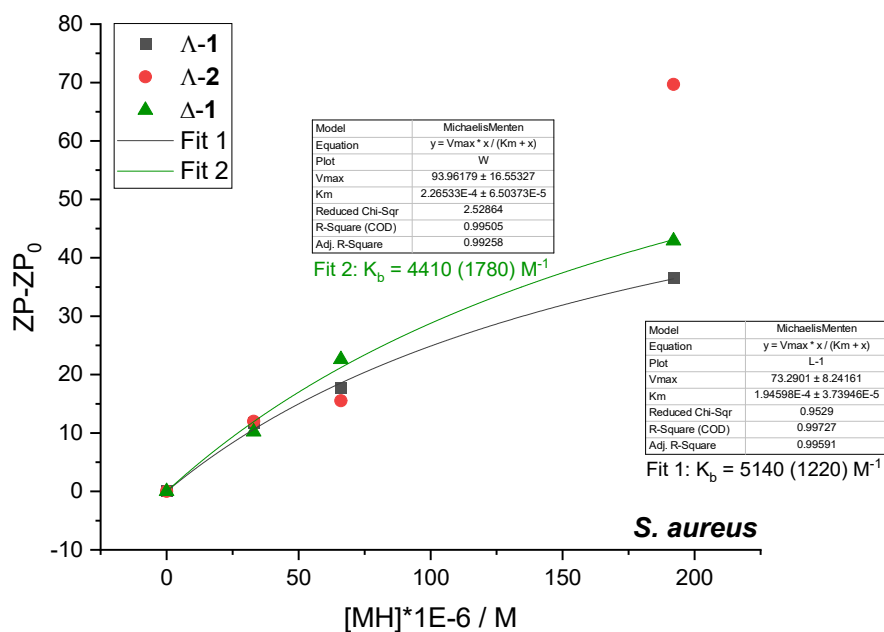


Figure S27 Fitting Zeta potential data. Zeta potential measurements of 0.5 mg mL^{-1} unilamellar *S. aureus* vesicles ([lipid] $\sim 0.6 \text{ mM}$), following the addition of 0-192 mM Λ -1 and Δ -1, in sodium phosphate buffer (25 mM, pH 7.4). It was not possible to fit the data following addition of Λ -2.

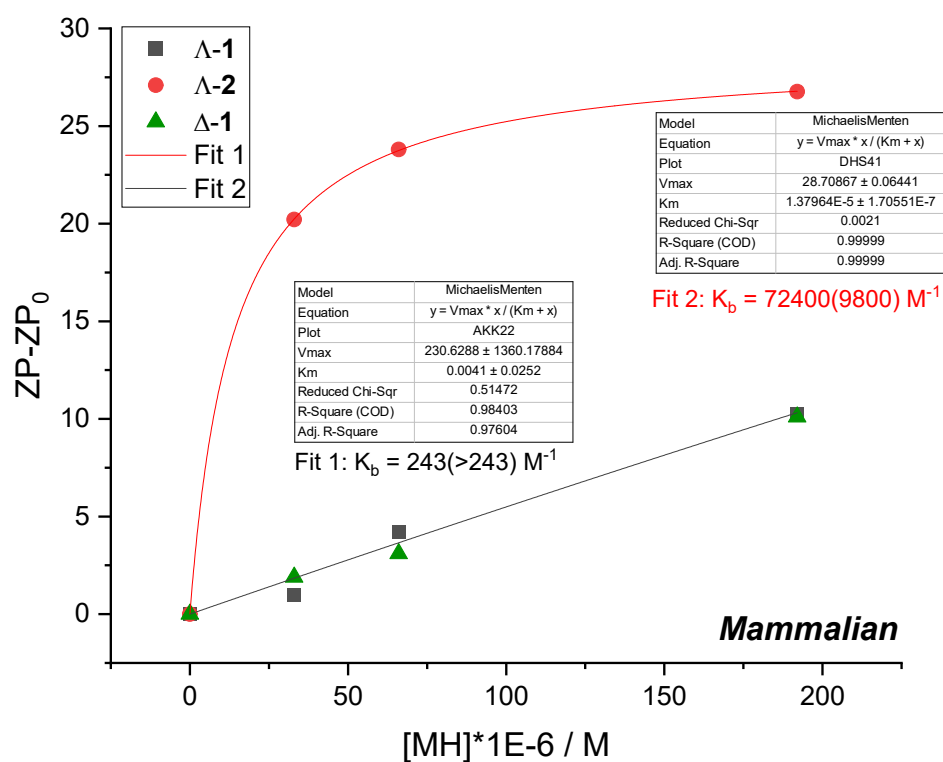


Figure S28 Fitting Zeta potential data. Zeta potential measurements of 0.5 mg mL⁻¹ unilamellar mammalian vesicles ([lipid] ~ 0.6 mM), following the addition of 0-192 mM Λ -1, Λ -2, and Δ -1, in sodium phosphate buffer (25 mM, pH 7.4).

Table S6. Apparent Binding constants (K_{app}) of metallohelicenes with model membranes, estimated from zeta potential titrations

	<i>E. coli</i>	<i>S. aureus</i>	Mammalian
Λ -1	$3.1 \pm 0.4 \times 10^4$	$5.1 \pm 1.2 \times 10^3$	- ^a
Δ -1	$4.4 \pm 1.4 \times 10^4$	$4.4 \pm 1.8 \times 10^3$	- ^a
Λ -2	$1.2 \pm 0.5 \times 10^4$	- ^b	$7.2 \pm 1.0 \times 10^4$

^a Cannot fit data due to negligible changes in zeta potential

^b Cannot fit data – looks like two binding events