SUPPORTING INFORMATION

Poly(3-hexylthiophene) nanoparticles as visible-light photoinitiators and photosensitizers in 3D printable acrylic hydrogels for photodynamic therapies

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SUPPLEMENTARY FIGURES



7.2 7.0 6.8 6.6 6.4 6.2 6.0 5.8 5.6 5.4 5.2 5.0 4.8 4.6 4.4 4.2 4.0 3.8 3.6 3.4 3.2 3.0 2.8 2.6 2.4 2.2 2.0 1.8 1.6 1.4 1.2 1.0 0.8 f1 (ppm)

Fig. S1 ¹H NMR spectrum of synthesized P3HT.



Fig. S2 (A) TEM image and (B) UV-vis absorption spectrum of synthesized P3HT SPNs.



Fig. S3 Elastic modulus (G') and Loss modulus (G'') of (A) PEGDA-SPNs_x and (B) PAAm-SPNs₈₀, PEGDA-SPNs₈₀, and PEGDA-co-HEA-SPNs₈₀ hydrogels as a function of the oscillation strain.



Fig. S4 (A) Representative pictures of PEGDA-co-HEA-SPNs_x hydrogels that show their flexibility properties. Elastic modulus (G') and Loss modulus (G'') of PEGDA-co-HEA-SPNs_x hydrog as a function of the (B) frequency and (C) oscillation strain.



Fig. S5 UV-vis spectra of PEGDA-co-HEA and PEGDA-co-HEA-SPNS_x ($x = 5, 80, and 105 mg mL^{-1}$) hydrogels.



Fig. S6 Pictures of PEGDA-co-HEA-SPNs₈₀ hydrogels (A) before and (B) after light excitation $(\lambda_{exc} = 250 \text{ nm})$ emitting fluorescence.



Fig. S7 Fluorescence microscopy images of mouse glioma 261 (GL261) cells in contact with silicone disks (CTRL) and PEGDA-co-HEA-SPNs_x hydrogels (A) before and (B) after LED irradiation ($\lambda = 467$ nm; 60 mW cm⁻², 40 J cm⁻²) for 10 min. Scale bars = 100 µm.



Fig. S8 Flow cytometry results of mouse glioma 261 (GL261) cells in contact with silicone disks (CTRL) and PEGDA-co-HEA-SPNs_x hydrogels, (A) before and (B) after LED irradiation ($\lambda = 467$ nm; 60 mW cm⁻², 40 J cm⁻²) for 10 min.



Fig. S9 Fluorescence microscopy images of mouse glioma 261 (GL261) cells in contact with silicone disks (CTRL) and PEGDA-co-HEA-SPNs_x hydrogels, and incubated with the fluorescence probe DCFDA, (A) before and (B) after LED irradiation ($\lambda = 467$ nm; 60 mW cm⁻², 40 J cm⁻²) for 10 min. Scale bars = 100 µm.