

**Supporting information**

**For**

**Homogenization Offers Access to Quinoxalines in Minutes: A Solvent-free, Catalyst-free Protocol with Near-zero E-factor**

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**Table of contents, spectral data, and copies of spectra**

<b>Table of contents</b>	<b>Page no.</b>
General experimental details	S2
General procedure for the synthesis of quinoxalines	S2
Procedure for Scale-up	S3
Reaction monitoring by solid-state infrared spectroscopy (Figure S1)	S3
Schematic representation of high-throughput synthesis of quinoxalines (Figure S2)	S3
Comparison with solution phase (Table S1)	S4
Comparative study of quinoxaline synthesis with existing literature ((Table S2 and S3))	S4-S5
E-factor and Ecoscale score calculations	S5-S6
Spectral characterization	S6-S14
References	S14-S16
Copies of $^1\text{H}$ NMR and $^{13}\text{C}$ NMR spectra of quinoxalines (Figure S3-S46)	S17-S60

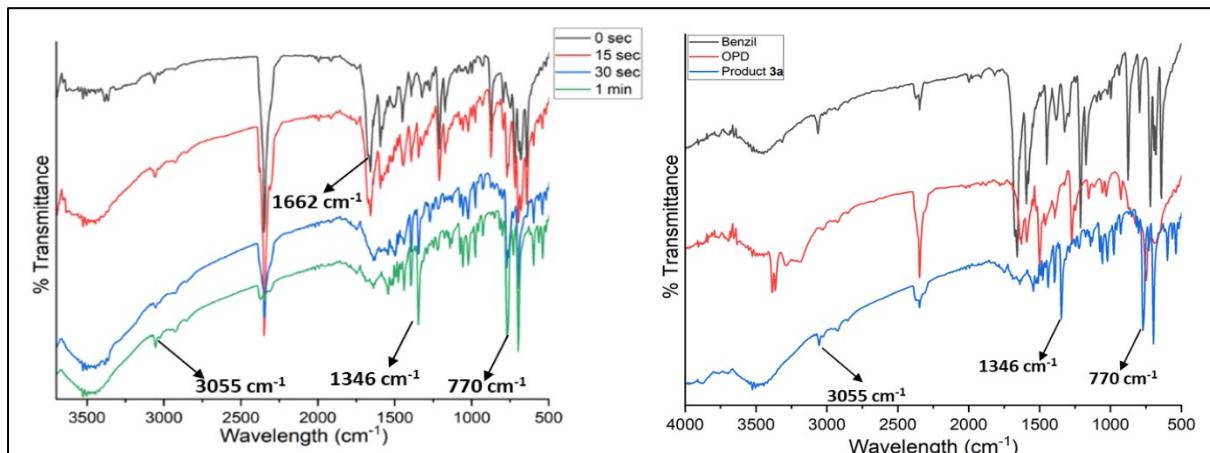
**General experimental details:** The chemicals were obtained from Sigma-Aldrich, Alfa Aesar, or TCI India and used directly without further purification. Common reagents and solvents of AR grade were obtained from local suppliers. The mechanochemical reactions were carried out in 2 mL homogenizing tubes made of polypropylene with 1 g of 2 mm SS ball in iRupt Jr, an indigenous mini-cell homogenizer (Neuation). The reactions were monitored by thin layer chromatography (TLC) carried out on 0.25 mm silica gel aluminum plates (60F-254) using UV light (254 or 365 nm) to monitor the progress of the reactions.  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR spectra were recorded on a Bruker Avance (500 MHz) NMR instrument with  $\text{CDCl}_3$  as the solvent. Tetramethylsilane (TMS) was used as an internal standard for  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopy. Chemical shifts are reported in parts per million ( $\delta$ ) units. Coupling constants are reported in hertz (Hz). Standard abbreviations are used for representing the multiplicity of NMR peaks, such as s (singlet), bs (broad singlet), d (doublet), dd (doublet of doublets), t (triplet), q (quartet), and m (multiplet). The melting point was measured in the EZ-Melt, Automatic Melting Point Apparatus (Stanford Research System). Mass spectra were recorded on a single quad LC-MS, Agilent using electrospray ionization (ESI) as an ion source.

**General procedure for the synthesis of quinoxalines:** *o*-Diaminoarene (**1**, 0.5 mmol) and 1,2-dicarbonyl compound (**2**, 0.5 mmol) was taken in a 2 mL polypropylene tube containing 1 g of 2 mm SS balls. The reaction was set to a speed of 4000 RPM in iRupt Jr instrument and was homogenized for 1 x 3 min with intermittent mixing (if required). The progress of the reaction was monitored by TLC. The product was taken out from the tube, the balls were separated using a magnetic retriever whenever possible, and the solid mass was air-dried to afford the corresponding quinoxaline derivative (**3**).

In selective cases, as mentioned in Table 2, the crude product was subjected to flash chromatography (silica gel, 230-400 mesh) and eluted by 10% EtOAc in petroleum ether to afford the pure product (**3**).

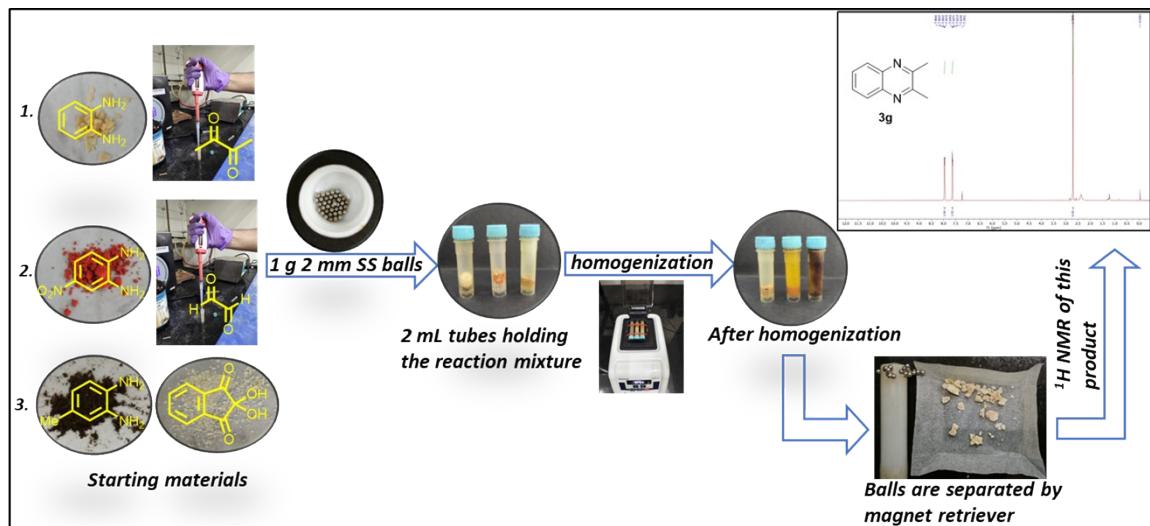
**Procedure for scale-up:** *o*-Diaminoarene (2.5 mmol) and 1,2-dicarbonyl (2.5 mmol) were taken in a 2 mL homogenizing tube containing 1 g of 2 mm SS balls. The reaction was set to a speed of 4000 RPM in the iRupt Jr instrument and was homogenized for 3 min. The product was isolated, as mentioned before.

### Reaction monitoring by solid-state infrared spectroscopy



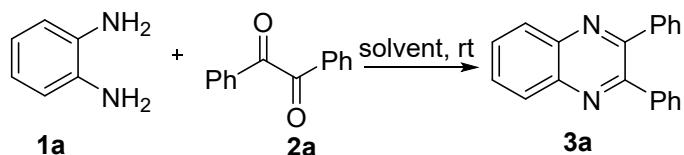
**Figure S1.** Monitoring of the reaction mixture in solid state by time-scale infrared spectroscopy. The disappearance of the carbonyl peak at  $1662\text{ cm}^{-1}$  and the gradual increase of the signature peaks at  $1346$ , and  $770\text{ cm}^{-1}$  just after 30 sec indicates the formation of **3a**.

### Schematic representation of high-throughput synthesis of quinoxalines



**Figure S2.** Schematic representation of reaction set-up for semi high-throughput synthesis of quinoxalines using homogenizer.

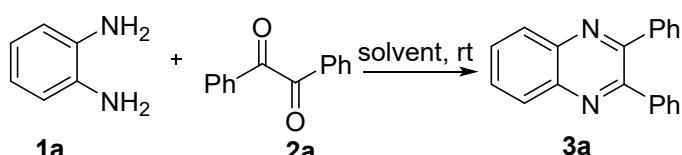
**Table S1: Comparison with solution phase:**



Entry	Solvent	Time (min)	Yield (%) <sup>b</sup>	% recovery of 2
1	Neat	120	11	69
2	Acetonitrile	120	82	12
3	Ethanol	60	91	--
4	Chloroform	120	68	18
5	Tetrahydrofuran	120	62	24

<sup>a</sup>In each case, 0.5 mmol each of **1a** and **2a** are taken in 2 mL of solvent (other than entry 1) and the mixture was stirred at room temperature. <sup>b</sup>All are isolated yields.

**Table S2: Comparative study of quinoxaline synthesis (selected methods)<sup>a</sup>**



Entr y	Media	Condition	Catalyst	Time (min)	Yield (%)	E-factor <sup>b</sup>	Eco-scal e	Referenc e
<b>Conventional solution phase</b>								
1	CH <sub>3</sub> CN	rt	Iodine	3	98	1.6	89	1
2	Ethanol	rt	CuSO <sub>4</sub> .5H <sub>2</sub> O	8	96	8.8	97	2
3	DMSO	rt	Iodine	35	95	4.2	97	3
4	Ethanol	rt	MnCl <sub>2</sub>	17	94	9	96	4
<b>Green-protocols</b>								
5	Water	rt	Itaconic acid	60	96	0.2	97	5
6	Solid phase	heating (100 °C)	CuO@g-C <sub>3</sub> N <sub>4</sub>	3	87	0.3	89	6
7	Water	ultrasonication	-	0.8	98	0.1 <sup>c</sup>	96	7
8	Glycero l	microwave	-	3	93	9.7	82	8
<b>9</b>	<b>Neat</b>	<b>homogenizer</b>	<b>-</b>	<b>3</b>	<b>99</b>	<b>0.01</b>	<b>97.5</b>	<b>Our work</b>

<sup>a</sup> Unless otherwise noted the comparisons are done taking **3a** as the model product. <sup>b</sup> Water formed as the side product of the reaction was not considered as waste during the E-factor calculation. <sup>c</sup> Ninhydrin was used instead of benzil to calculate the E-factor and eco-scale scores.

**Table S3: Mechanochemical methods using diamine and dicarbonyl<sup>a</sup>**

Entry	Mechanical technique	Catalyst	Milling auxiliary	Milling condition	Isolation/ Purification /time	Yield (%)	E-factor	Reference
1	Hand grinding	-	EtOH (LAG)	10 min	Recrystallization (EtOH)	95	0.16	9
2	Hand grinding	-	SiO <sub>2</sub>	15 min	Recrystallization (EtOH)	94	4	10
3	Hand grinding	nano-kaolin/ BF <sub>3</sub> / Fe <sub>3</sub> O <sub>4</sub>	-	20 min	Recrystallization (EtOH)	98	0.26	11
4	Hand grinding	Acetic acid	-	10 min	Recrystallization (acetone)	92 <sup>c</sup>	0.34	12
5	Hand grinding	-	Al <sub>2</sub> O <sub>3</sub>	10 min	Chromato- graphy	98	1.96	13

6	Planetary ball mill	La(DS) <sub>3</sub>	NaCl	600 RPM, 30 min	Chromato-graphy	99	2.1	14
7	Mixer mill	TCCA, <i>p</i> -TSA, K <sub>2</sub> CO <sub>3</sub>	-	20 Hz, 10 h	Chromato-graphy	78	1.8	15
8	<b>Homogenizer</b>	-	-	<b>4000 RPM, 3 min</b>	<b>no work-up, no purification</b>	<b>99</b>	<b>0.01</b>	<b>Our work</b>

<sup>a</sup> Unless otherwise noted the comparisons are done taking **3a** as the model product. <sup>b</sup>Water formed as the side product of the reaction was not considered as waste during the E-factor calculation. <sup>c</sup> Ninhydrin was used instead of benzil to calculate the E-factor. <sup>d</sup>The calculations are done using 4-bromoacetophenone and OPD as the reactants.

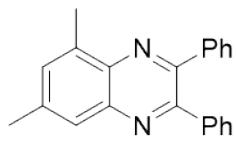
#### For a typical E-factor calculation:

Parameters	Values	Remarks
%Yield	99	Calculated for <b>3a</b>
Wt. of generated waste (mg)	3	
Wt. of end product (mg)	279	
<b>E-factor = Wt. of generated waste/Wt. of end product</b>	<b>0.01</b>	

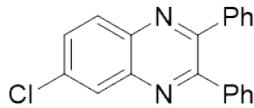
#### For a typical Ecoscale score calculation:

Parameters	Penalty points	Remarks
%Yield = (100-%Yield)/2	0.5	Calculated for <b>3a</b>
Price of the reaction component	0	Inexpensive (<10US\$)
Technical setup	2	Unconventional activation technique (Homogenizer)
Temperature/Time	0	Room temperature reaction/<1 h
Workup and purification	0	None
<b>Sum of penalty points</b>	<b>2.5</b>	
<b>Eco-scale score (100-Sum of penalty)</b>	<b>97.5</b>	

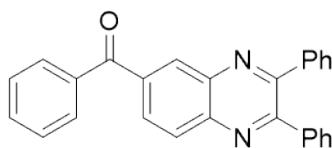
## Spectral characterization



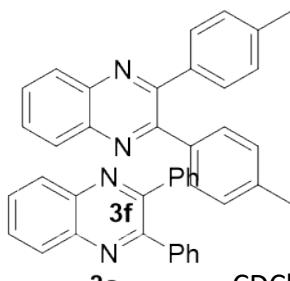
**2,3-Diphenylquinoxaline (3a):**<sup>9</sup> White solid, 140 mg (99%) m.p.: 124-125 °C (Lit. m.p.: 126-127 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.30-7.34 (m, 6H), 7.49-7.51 (m, 4H), 7.74-7.76 (m, 2H), 8.15-8.17 (m, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 128.3, 128.8, 129.2, 129.8, 130.0, 139.1, 141.2, 153.5. ESI-MS: *m/z* 283 [M + H]<sup>+</sup>.



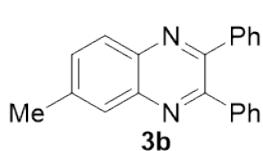
**6-Methyl-2,3-diphenylquinoxaline (3b):**<sup>9</sup> Light brown solid; 147 mg (99%), m.p.: 114-116 °C (Lit. m.p.: 118-119 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.59 (s, 3H), 7.28-7.34 (m, 6H), 7.49 (d, *J* = 6.8 Hz, 4H), 7.58 (dd, *J*<sub>1</sub> = 1.9 Hz, *J*<sub>2</sub> = 8.5 Hz, 1H), 7.93 (s, 1H), 8.04 (d, *J* = 8.5 Hz, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 21.9, 128.0, 128.2, 128.6, 128.7, 129.8, 129.9, 132.3, 139.2, 139.7, 140.5, 141.3, 152.6, 153.3; ESI-MS: *m/z* 297 [M + H]<sup>+</sup>.



**5,7-Dimethyl-2,3-diphenylquinoxaline (3c):**<sup>9</sup> Off-white solid, 153 mg (99%), m.p.: 94-96 °C (Lit. m.p.: 94-96 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.54 (s, 3H), 2.79 (s, 3H), 7.29-7.32 (m, 6H), 7.42 (s, 1H), 7.48-7.54 (m, 4H), 7.75 (s, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 17.0, 21.9, 125.7, 128.0, 128.2, 128.5, 128.6, 129.8, 130.1, 132.2, 137.0, 138.9, 139.5, 139.6, 140.1, 141.3, 150.9, 152.7; ESI-MS: *m/z* 311 [M + H]<sup>+</sup>.



**6-Chloro-2,3-diphenylquinoxaline (3d):**<sup>9</sup> White solid, 157 mg (99%), m.p.: 119-121 °C (Lit. m.p.: 124-125 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.30-7.36 (m, 6H), 7.48 (d, *J* = 7.4 Hz, 4H), 7.69 (d, *J* = 8.8 Hz, 1H), 8.08 (d, *J* = 8.9 Hz, 1H), 8.15 (s, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 128.1, 128.3, 129.0, 129.1, 129.81, 129.85, 130.4, 130.9, 135.6, 138.6, 138.7, 139.7, 141.5, 153.6, 154.3; ESI-MS: *m/z* 317 [M + H]<sup>+</sup>.

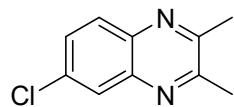


**6-Benzoyl-2,3-diphenylquinoxaline (3e):**<sup>10</sup> White solid, 183 mg (95%), m.p.: 149-151 °C (Lit. m.p.: 147-148 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.30-7.39 (m, 6H), 7.49-7.54 (m, 6H), 7.61 (t, *J* = 7.5 Hz, 1H), 7.89 (d, *J* = 7.3 Hz, 2H), 8.23-8.28 (m, 2H), 8.52 (s, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 128.4, 128.5, 129.1, 129.3, 129.7, 129.8, 129.9, 129.92, 130.1, 132.4, 132.8, 137.2, 138.3, 138.61, 138.64, 140.2, 143.0, 154.6, 155.1, 195.8; ESI-MS: *m/z* 387 [M + H]<sup>+</sup>.

**2,3-Di-p-tolylquinoxaline (3f):**<sup>9</sup> White solid, 153 mg (99%), m.p.: 146-147 °C; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.35 (s, 6H), 7.13 (d, *J* = 7.8 Hz, 4H), 7.41 (d, *J* =

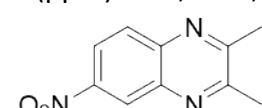
8.1 Hz, 4H), 7.72 (dd,  $J_1$  = 3.5 Hz,  $J_2$  = 6.5 Hz, 2H), 8.13 (dd,  $J_1$  = 3.5 Hz,  $J_2$  = 6.5 Hz, 2H);  $^{13}\text{C}$  { $^1\text{H}$ } NMR (125 MHz,  $\text{CDCl}_3$ ):  $\delta$  (ppm) 21.3, 129.0, 129.1, 129.6, 129.7, 136.3, 138.7, 141.1, 153.5; ESI-MS:  $m/z$  311 [M + H] $^+$ .

**2,3-Dimethylquinoxaline (3g):**<sup>9</sup> Light brown solid, 78 mg (99%), m.p.: 92-94 °C (Lit. m.p.: 105-106 °C);  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  (ppm) 2.70 (s, 6H), 7.63 (dd,  $J_1$  = 3.4 Hz,  $J_2$  = 6.4 Hz, 2H), 7.95 (dd,  $J_1$  = 3.6 Hz,  $J_2$  = 6.4 Hz, 2H);  $^{13}\text{C}$  { $^1\text{H}$ } NMR (125 MHz,  $\text{CDCl}_3$ ):  $\delta$  (ppm) 23.1, 128.2, 128.9, 140.9, 153.4; ESI-MS:  $m/z$  159 [M + H] $^+$ .



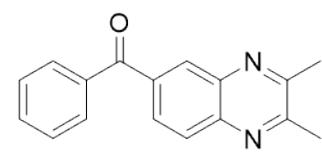
**3j**

**2,3,5-Trimethylquinoxaline (3h):**<sup>9</sup> Brown solid, 85 mg (99%), m.p.: 90-92 °C (Lit. m.p.: 90-91 °C);  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  (ppm) 2.51 (s, 3H), 2.66 (s, 6H), 7.44 (dd,  $J_1$  = 2.0 Hz,  $J_2$  = 8.6 Hz, 1H), 7.70 (s, 1H), 7.82 (d,  $J$  = 8.5 Hz, 1H);  $^{13}\text{C}$  { $^1\text{H}$ } NMR (125 MHz,  $\text{CDCl}_3$ ):  $\delta$  (ppm) 21.7, 23.0, 23.1, 127.2, 127.8, 131.0, 139.1, 139.4, 141.1, 152.4, 153.3; ESI-MS:  $m/z$  173 [M + H] $^+$ .



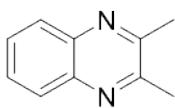
**3i**

**2,3,5,7-Tetramethylquinoxaline (3i):**<sup>9</sup> White solid, 92 mg (99%), m.p.: 68-69 °C (Lit. m.p.: 68-69 °C);  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  (ppm) 2.48 (s, 3H), 2.69 (s, 5H), 2.71 (s, 3H), 7.32 (s, 1H), 7.64 (s, 1H);  $^{13}\text{C}$  { $^1\text{H}$ } NMR (125 MHz,  $\text{CDCl}_3$ ):  $\delta$  (ppm) 17.0, 21.7, 22.4, 23.1, 124.2, 131.5, 136.2, 139.0, 139.4, 151.4, 151.9; ESI-MS:  $m/z$  187 [M + H] $^+$ .



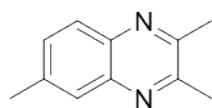
**3l**

**6-Chloro-2,3-dimethylquinoxaline (3j):**<sup>9</sup> Brown solid, 95 mg (99%), m.p.: 84-86 °C (Lit. m.p. : 84-86 °C);  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  (ppm) 2.69 (s, 6H), 7.57 (dd,  $J_1$  = 2.3 Hz,  $J_2$  = 8.9 Hz, 1H), 7.87 (d,  $J$  = 8.8 Hz, 1H), 7.94 (d,  $J$  = 2.4 Hz, 1H);  $^{13}\text{C}$  { $^1\text{H}$ } NMR (125 MHz,  $\text{CDCl}_3$ ):  $\delta$  (ppm) 23.1, 23.2, 127.3, 129.5, 129.8, 134.4, 139.5, 141.4, 153.7, 154.5; ESI-MS:  $m/z$  193 [M + H] $^+$ .



**3g**

**2,3-Dimethyl-6-nitroquinoxaline (3k):**<sup>9</sup> Brown solid, 100 mg (99%), m.p.: 134-135 °C (Lit. m.p: 135–136 °C);  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  (ppm) 2.76 (s, 6H), 8.06 (dd,  $J_1$  = 2.8 Hz,  $J_2$  = 9.1 Hz, 1H), 8.38 (dt,  $J_1$  = 3.0 Hz,  $J_2$  = 9.1 Hz, 1H), 8.84 (s, 1H);  $^{13}\text{C}$  { $^1\text{H}$ } NMR (125 MHz,  $\text{CDCl}_3$ ):  $\delta$  (ppm) 23.3, 23.5, 122.3, 124.8, 129.9, 139.9, 143.7, 147.1, 156.3, 157.2; ESI-MS:  $m/z$  204 [M + H] $^+$ .

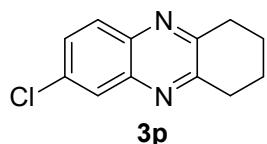


**3h**

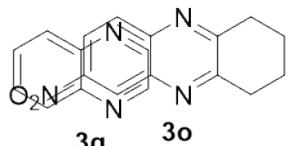
**6-Benzoyl-2,3-Dimethyl-quinoxaline (3l):**<sup>16</sup> Yellow solid, 126 mg (96%) m.p.: 122-124 °C (Lit. m.p.: 107-110 °C);  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  (ppm) 2.70 (s, 3H), 2.73 (s, 3H), 7.46 (t,  $J$  = 7.6 Hz, 2H), 7.57 (t,  $J$  = 7.5 Hz, 1H), 7.82 (d,  $J$  = 7.9 Hz, 2H), 8.04 (d,  $J$  = 8.7 Hz, 1H), 8.11 (dd,  $J_1$  = 1.8 Hz,  $J_2$  = 8.7 Hz, 1H), 8.31 (s, 1H);  $^{13}\text{C}$  { $^1\text{H}$ } NMR

(125 MHz, CDCl<sub>3</sub>): δ (ppm) 23.2, 23.4, 128.4, 128.8, 128.9, 130.0, 131.7, 132.7, 137.2, 137.3, 140.0, 142.9, 154.8, 155.7, 195.9. ESI-MS: *m/z* 263 [M + H]<sup>+</sup>.

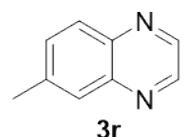
**1,2,3,4-Tetrahydronaphazine (3m):**<sup>5</sup> Brown solid, 98 mg (98%), m.p.: 87-88 °C (Lit. m.p.: 90 °C) <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 1.95 (s, 4H), 3.07 (s, 4H), 7.55-7.58 (m, 2H), 7.86-7.90 (m, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 22.6, 33.02, 33.04, 118.2, 128.1, 128.2, 128.7, 128.8, 141.02, 141.03, 153.9, 154.0. ESI-MS: *m/z* 185 [M + H]<sup>+</sup>.



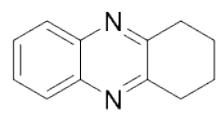
**7-Methyl-1,2,3,4-tetrahydronaphazine (3n):**<sup>5</sup> Yellow solid, 95 mg (96%), m.p.: 88-90 °C (Lit. m.p.: 92 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 1.97 (s, 4H), 2.50 (s, 3H), 3.08 (s, 4H), 7.43 (d, *J* = 8.5 Hz, 1H), 7.68 (s, 1H), 7.79 (d, *J* = 8.5 Hz, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 21.7, 22.81, 22.83, 33.0, 33.1, 127.1, 127.7, 131.2, 139.2, 139.6, 141.2, 153.0, 153.9. ESI-MS: *m/z* 199 [M + H]<sup>+</sup>.



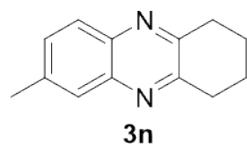
**7-Nitro-1,2,3,4-tetrahydronaphazine (3o):**<sup>5</sup> Light brown solid, 106 mg (93%), m.p.: 103-105 °C (Lit. m.p.: 101 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.04 (t, *J* = 3.6 Hz, 4H), 3.17 (t, *J* = 3.0 Hz, 4H), 8.04 (d, *J* = 9.1 Hz, 1H), 8.38 (dd, *J*<sub>1</sub> = 2.7 Hz, *J*<sub>2</sub> = 9.1 Hz, 1H), 8.82 (d, *J* = 2.6 Hz, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 22.4, 33.2, 33.5, 122.3, 124.9, 129.9, 139.9, 143.7, 147.1, 157.0, 158.0; ESI-MS: *m/z* 230 [M + H]<sup>+</sup>.



**7-Chloro-1,2,3,4-tetrahydronaphazine (3p):**<sup>5</sup> Yellow solid, 102 mg (94%), m.p.: 91-93 °C (Lit. m.p.: 94 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.00 (t, *J* = 3.6 Hz, 4H), 3.10-3.12 (m, 4H), 7.56 (d, *J* = 8.8 Hz, 1H), 7.86 (d, *J* = 8.8 Hz, 1H), 7.92 (s, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 22.6, 22.7, 33.1, 33.2, 127.3, 129.6, 129.9, 134.5, 139.7, 141.4, 154.4, 155.2; ESI-MS: *m/z* 219 [M + H]<sup>+</sup>.

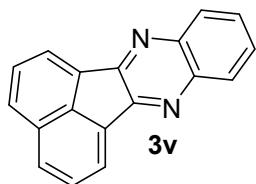


**Quinoxaline (3q):**<sup>9</sup> Light yellow liquid, 60 mg (92%); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.69 (dd, *J*<sub>1</sub> = 3.5 Hz, *J*<sub>2</sub> = 6.5 Hz, 2H), 8.03 (dd, *J*<sub>1</sub> = 3.6 Hz, *J*<sub>2</sub> = 6.5 Hz, 2H), 8.76 (s, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 129.3, 129.9, 142.8, 144.8; ESI-MS: *m/z* 131 [M + H]<sup>+</sup>.

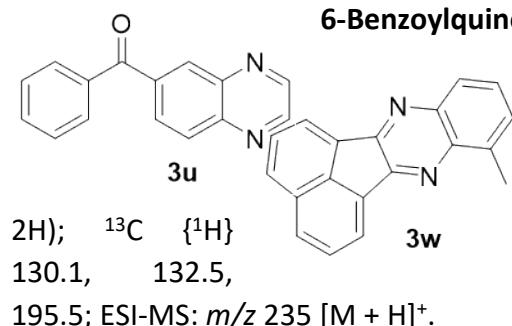


**6-Methylquinoxaline (3r):**<sup>9</sup> Colourless liquid, 68 mg (95%); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.55 (s, 3H), 7.56 (d, *J* = 8.6 Hz, 1H), 7.82 (s, 1H), 7.95 (d, *J* = 8.6 Hz, 1H), 8.74 (d, *J* = 11.9 Hz, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 21.8, 128.2, 128.9, 132.4, 140.6, 141.5, 143.0, 144.0, 144.8; ESI-MS: *m/z* 145 [M + H]<sup>+</sup>.

**6-Chloroquinoxaline (3s):**<sup>9</sup> Off-white solid, 77 mg (94%), m.p.: 63-65 °C (Lit. m.p.: 62-64 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.68 (dd, J<sub>1</sub> = 2.4 Hz, J<sub>2</sub> = 11.3 Hz, 1H), 8.01 (d, J = 8.9 Hz, 1H), 8.07 (d, J = 2.4 Hz, 1H), 8.81 (dd, J<sub>1</sub> = 1.9 Hz, J<sub>2</sub> = 6.8 Hz, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 128.4, 130.7, 131.1, 135.9, 141.5, 143.2, 145.0, 145.7; ESI-MS: m/z 165 [M + H]<sup>+</sup>.

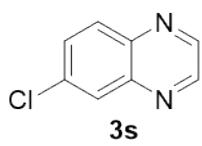


**6-Nitroquinoxaline (3t):**<sup>9</sup> Light yellow solid, 86 mg (98%), m.p.: 158-160 °C (Lit. m.p.: 158-160 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 8.25 (d, J = 9.2 Hz, 1H), 8.52 (dd, J<sub>1</sub> = 2.5 Hz, J<sub>2</sub> = 9.2 Hz, 1H), 8.91-9.07 (m, 3H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 123.5, 126.0, 131.4, 141.9, 145.3, 147.0, 147.7; ESI-MS: m/z 176 [M + H]<sup>+</sup>.

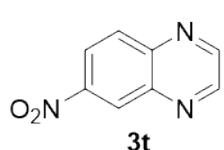


**6-Benzoylquinoxaline (3u):**<sup>17</sup> Light Brown solid, 106 mg (91%), m.p: 116-118 °C (Lit. m.p. 117 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.49 (t, J = 7.8 Hz, 2H), 7.60 (t, J = 7.5 Hz, 1H), 7.84 (d, J = 7.9 Hz, 2H), 8.20 (t, J = 10.8 Hz, 2H), 8.44 (s, 1H), 8.90 (dd, J<sub>1</sub> = 2.0 Hz, J<sub>2</sub> = 6.2 Hz, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 128.5, 129.9, 130.0, 133.0, 136.9, 138.5, 142.1, 144.6, 146.0, 146.6, 195.5; ESI-MS: m/z 235 [M + H]<sup>+</sup>.

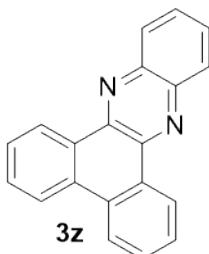
**Acenaphtho[1,2-b]quinoxaline (3v):**<sup>9</sup> Light yellow solid, 125 mg (99%), m.p.: 244-246 °C (Lit. m.p.: 243–245 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.70-7.72 (m, 2H), 7.76-7.80 (m, 2H), 8.04 (dd, J<sub>1</sub> = 2.4 Hz, J<sub>2</sub> = 8.2 Hz, 2H), 8.15-8.18 (m, 2H), 8.37 (d, J = 7.1 Hz, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 121.8, 128.6, 129.2, 129.4, 129.5, 130.0, 131.7, 136.4, 141.2, 154.0; ESI-MS: m/z 255 [M + H]<sup>+</sup>.



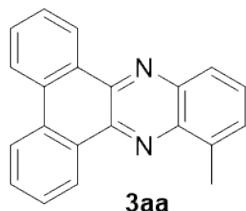
**5-Methylacenaphtho[1,2-b]quinoxaline (3w):**<sup>9</sup> Light yellow solid, 132 mg (99%), m.p.: 282-284 °C (Lit. m.p.: 282-284 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.93 (s, 3H), 7.57-7.64 (m, 2H), 7.82 (t, J = 8.4 Hz, 2H), 8.03 (d, J = 8.2 Hz, 1H), 8.08 (dd, J<sub>1</sub> = 3.1 Hz, J<sub>2</sub> = 8.4 Hz, 2H), 8.42 (t, J = 6.1 Hz, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 17.5, 121.7, 127.4, 128.60, 128.63, 128.8, 129.2, 129.3, 129.5, 130.0, 132.1, 132.4, 136.4, 138.0, 140.4, 141.3, 153.0, 153.6.; ESI-MS: m/z 269 [M + H]<sup>+</sup>.



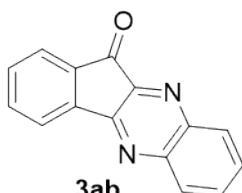
**6-Methylacenaphtho[1,2-*b*]quinoxaline (3x):**<sup>9</sup> Light yellow solid, 132 mg (99%), m.p.: 233-



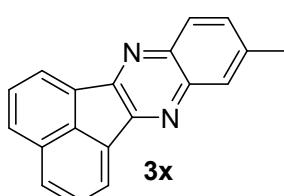
235 °C (Lit. m.p.: 233-235 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.57 (s, 3H), 7.51 (d, J = 8.4 Hz, 1H), 7.75 (t, J = 7.6 Hz, 2H), 7.91 (s, 1H), 7.99-8.03 (m, 3H), 8.31 (t, J = 6.5 Hz, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 21.7, 121.5, 121.7, 128.5, 128.6, 128.7, 129.0, 129.1, 129.3, 129.9, 131.3, 131.9, 132.0, 136.2, 139.5, 139.6, 141.2, 153.3, 154.0; ESI-MS: *m/z* 269 [M + H]<sup>+</sup>.



**6-Chloroacenaphtho[1,2-*b*]quinoxaline (3y):**<sup>9</sup> Dark brown solid, 142mg (99%) m.p.: 240-242 °C (Lit. m.p.: 240-242 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.61 (d, J = 8.8 Hz, 1H), 7.75-7.78 (m, 2H), 8.02-8.04 (m, 3H), 8.09 (s, 1H), 8.29-8.32 (m, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 122.0, 122.2, 128.5, 128.6, 129.6, 129.81, 129.88, 129.9, 130.5, 131.3, 131.3, 134.7, 136.5, 139.6, 141.5, 154.0, 154.6; ESI-MS: *m/z* 289 [M + H]<sup>+</sup>.

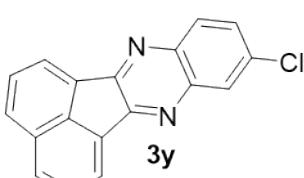


**Phenanthro[9,10-*b*]quinoxaline (3z):**<sup>9</sup> Yellow solid, 138 mg (99%), m.p.: 226-227 °C (Lit. m.p.: 225–226 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.68-7.76 (m, 4H), 7.80-7.82 (m, 2H), 8.27 (dd, J<sub>1</sub> = 3.5 Hz, J<sub>2</sub> = 6.4 Hz, 2H), 8.49 (d, J = 7.9 Hz, 2H), 9.34 (d, J = 6.3 Hz, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 122.8, 126.2, 127.9, 129.4, 129.7, 130.2, 130.3, 132.0, 142.1, 142.4; ESI-MS: *m/z* 281 [M + H]<sup>+</sup>.



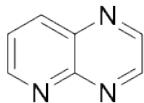
**5-Methylphenanthro[9,10-*b*]quinoxaline (3aa):**<sup>9</sup> Brown solid, 145 mg (99%), m.p.: 222-224 °C (Lit. mp.: 222-224 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.98 (s, 3H), 7.61-7.75 (m, 6H), 8.10 (d, J = 8.4 Hz, 1H), 8.48-8.50 (m, 2H), 9.32-9.36 (m, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 17.4, 122.83, 122.85, 126.1, 127.2, 127.7, 127.8, 129.3, 129.6, 130.0, 130.1, 130.3, 130.6, 131.9, 137.7, 141.0, 141.3, 141.8, 142.2; ESI-MS: *m/z* 295 [M + H]<sup>+</sup>.

**11*H*-Indeno[1,2-*b*]quinoxalin-11-one (3ab):**<sup>7</sup> Yellow solid, 115 mg (99%), m.p.: 216-218 °C (Lit. m.p.: 217–218 °C ); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.57 (s, 3H), 7.77 (t, J = 7.6 Hz, 2H), 7.77 (t, J = 7.6 Hz, 2H), 8.18 (d, δ (ppm) 122.5, 124.7, 129.6, 130.2, 131.5, 132.4, 132.5, 136.6, 136.8, 141.5, 142.6, 143.0, 149.2, 156.5, 189.8; ESI-MS: *m/z* 233 [M + H]<sup>+</sup>.

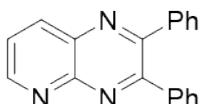


***b*]quinoxalin-11-one (3ab):**<sup>7</sup> Yellow solid, 115 216-218 °C (Lit. m.p.: 217–218 °C ); <sup>1</sup>H NMR (ppm) 7.55 (t, J = 7.4 Hz, 1H), 7.71 (q, J = 7.8 7.8 Hz, 1H), 7.87 (d, J = 7.6 Hz, 1H), 8.05,(t, J = J = 8.2 Hz, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>):

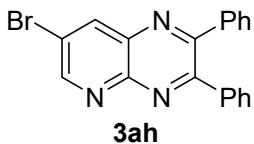
**6-Methyl-11*H*-indeno[1,2-*b*]quinoxalin-11-one (3ac):**<sup>18</sup> Yellow solid, 122 mg (99%), m.p.: 224-225 °C (Lit. m.p: 227-229 °C ); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.77 (s, 3H), 7.50-7.59 (m, 3H), 7.69 (t, *J* = 7.3 Hz, 1H), 7.84 (d, *J* = 7.5 Hz, 1H), 7.99-8.03 (m, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 17.3, 122.3, 124.5, 129.3, 129.8, 132.1, 132.7, 136.5, 136.6, 138.3, 141.8, 142.1, 142.7, 148.7, 155.4, 190.3; ESI-MS: *m/z* 247 [M + H]<sup>+</sup>.



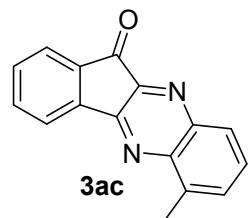
**2,3-di(pyridin-2-yl)quinoxaline (3ad):**<sup>19</sup> Brown solid, 141 mg (99%), m.p.: 185-187 °C (Lit. m.p.: 188-191 °C ); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.18-7.20 (m, 2H), 7.75-7.78 (m, 4H), 7.92 (d, *J* = 7.8 Hz, 2H), 8.18-8.20 (m, 2H), 8.34 (d, *J* = 4.5 Hz, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 122.9, 124.1, 129.2, 130.4, 136.5, 141.0, 148.4, 152.2, 157.2; ESI-MS: *m/z* 285 [M + H]<sup>+</sup>.



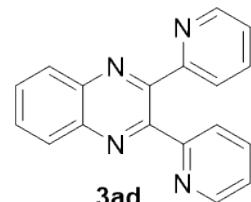
**2,3-Dimethylpyrido[2,3-*b*]pyrazine (3ae):**<sup>9</sup> Light brown solid, 79 mg (99%), m.p.: 133-135 °C (Lit. m.p.: 133-135 °C ); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.70 (s, 3H), 2.74 (s, 3H), 7.54-7.56 (m, 1H), 8.26 (dd, *J*<sub>1</sub> = 1.9 Hz, *J*<sub>2</sub> = 8.4 Hz, 1H), 8.97 (d, *J* = 2.5 Hz, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 23.0, 23.4, 124.3, 135.8, 137.3, 150.1, 152.5, 155.0, 157.3; ESI-MS: *m/z* 160 [M + H]<sup>+</sup>.



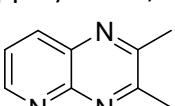
**Pyrido[2,3-*b*]pyrazine (3af):**<sup>9</sup> Light brown solid, 60 mg (91%), m.p.: 135-137 °C (Lit. m.p.: 133-135 °C ); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.71-7.74 (m, 1H), 8.46 (dd, *J*<sub>1</sub> = 1.9 Hz, *J*<sub>2</sub> = 8.3 Hz, 1H), 8.93 (s, 1H), 9.06 (s, 1H), 9.17 (dd, *J*<sub>1</sub> = 1.9 Hz, *J*<sub>2</sub> = 4.2 Hz, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 125.5, 138.4, 138.6, 146.1, 147.8, 151.4, 154.3; ESI-MS: *m/z* 132 [M + H]<sup>+</sup>.



**2,3-diphenylpyrido[2,3-*b*]pyrazine (3ag):**<sup>10</sup> Yellow solid, 140 mg (99%), m.p.: 141–143 °C (Lit. m.p.: 144-145 °C ); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.18-7.29 (m, 6H), 7.44 (d, *J* = 7.4 Hz, 2H), 7.53 (d, *J* = 7.6 Hz, 2H), 7.57-7.58 (m, 1H), 8.39 (d, *J* = 8.7 Hz, 1H), 9.04-9.05 (m, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 125.0, 127.9, 128.2, 129.1, 129.2, 129.6, 130.0, 135.9, 137.8, 137.9, 138.3, 149.6, 153.9, 154.5, 156.0; ESI-MS: *m/z* 284 [M + H]<sup>+</sup>.

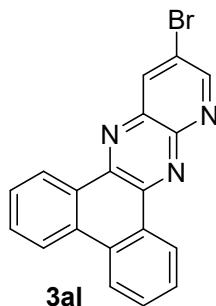


**Bromo-2,3-diphenylpyrido[2,3-*b*]pyrazine (3ah):**<sup>9</sup> White solid, 179 mg (99%), m.p.: 148-150 °C (Lit. m.p.: 148-150 °C ); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.28-7.39 (m, 6H), 7.50 (d, *J* = 6.8 Hz, 2H), 7.58 (d, *J* = 7.0 Hz, 2H), 8.63 (d, *J* = 2.4 Hz, 1H), 9.11 (d, *J* = 2.4 Hz, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 120.9, 128.2, 128.4, 129.6, 129.7, 129.8, 130.2, 136.4, 137.7, 138.0, 139.4, 148.2, 155.1, 155.4, 156.5; ESI-MS: *m/z* 364 [M + H]<sup>+</sup> (for <sup>79</sup>Br), 366 [M + H]<sup>+</sup> (for <sup>81</sup>Br).

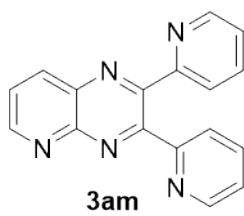




**Bromo-2,3-dimethylpyrido[2,3-*b*]pyrazine (3ai):**<sup>9</sup> Brown solid, 117 mg (99%), m.p.: 120-122 °C (Lit. m.p.: 120-122 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.72 (s, 3H), 2.74 (s, 3H), 8.44 (d, J = 2.6 Hz, 1H), 8.99 (d, J = 2.6 Hz, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 23.0, 23.5, 119.8, 136.2, 138.8, 148.6, 153.6, 156.1, 157.6; ESI-MS: *m/z* 238 [M + H]<sup>+</sup> (for <sup>79</sup>Br), 240 [M + H]<sup>+</sup> (for <sup>81</sup>Br).

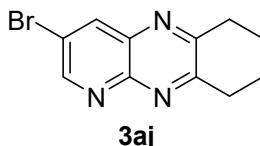


**3-Bromo-6,7,8,9-tetrahydropyrido[2,3-*b*]quinoxaline (3aj):** Yellow solid, 126 mg (96%), m.p.: 126-128 °C; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.01-2.04 (m, 4H), 3.13-3.21 (m, 4H), 8.44 (s, 1H), 9.01 (s, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 22.3, 22.5, 33.1, 33.5, 119.9, 136.4, 138.7, 148.5, 153.9, 156.8, 158.3; ESI-MS: *m/z* 264 [M + H]<sup>+</sup> (for <sup>79</sup>Br), 266 [M + H]<sup>+</sup> (for <sup>81</sup>Br).



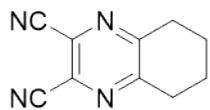
**7-Bromopyrido[2,3-*b*]pyrazine (3ak):**<sup>9</sup> Off-white solid, 101 mg (97%), m.p.: 158-160 °C (Lit. m.p.: 158-160 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 8.62 (d, J = 2.5 Hz, 1H), 8.91 (s, 1H), 9.05 (s, 1H), 9.15 (d, J = 2.5 Hz, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 121.4, 138.6, 139.9, 146.8, 147.9, 149.7, 155.5; ESI-MS: *m/z* 210 [M + H]<sup>+</sup> (for <sup>79</sup>Br), 212 [M + H]<sup>+</sup> (for <sup>81</sup>Br).

**12-Bromodibenzo[*f,h*]pyrido[2,3-*b*]quinoxaline (3al):**<sup>20</sup> Yellow solid, 178 mg (94%), m.p.: 216-216 °C; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.60-7.77 (m, 4H), 8.42 (d, J = 8.1 Hz, 2H), 8.67 (s, 1H), 9.08 (d, J = 7.9 Hz, 1H), 9.18 (s, 1H), 9.33 (d, J = 8.0 Hz, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 120.5, 122.8, 122.9, 126.5, 127.3, 128.1, 128.2, 129.1, 129.3, 131.2, 131.3, 132.4, 132.5, 137.3, 139.4, 143.9, 144.6, 148.0, 155.3; ESI-MS: *m/z* 347 [M + H]<sup>+</sup> (for <sup>79</sup>Br), 349 [M + H]<sup>+</sup> (for <sup>81</sup>Br).



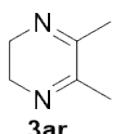
**2,3-Di(pyridin-2-yl)pyrido[2,3-*b*]pyrazine (3am):**<sup>19</sup> Brown solid, 134 mg (94%), m.p.: 267-269 °C; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.25-7.29 (m, 2H), 7.77-7.80 (m, 1H), 7.84-7.92 (m, 2H), 7.99 (d, J = 7.8 Hz, 1H), 8.28 (d, J = 4.5 Hz, 1H), 8.33 (d, J = 7.8 Hz, 1H), 8.40 (d, J = 4.8 Hz, 1H), 8.59 (dd, J<sub>1</sub> = 1.9 Hz, J<sub>2</sub> = 8.3 Hz, 1H), 9.23-9.24 (m, 1H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 123.2, 123.5, 124.1, 124.7, 125.7, 136.2, 136.7, 136.9, 138.3, 148.1, 148.6, 149.6, 153.9, 154.6, 155.2, 156.4, 157.0; ESI-MS: *m/z* 286 [M + H]<sup>+</sup>.

**5,6-Dimethylpyrazine-2,3-dicarbonitrile (3an):**<sup>21</sup> Gray solid, 78 mg (99%), m.p.: 168–170 °C; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.68 (s, 6H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 22.6, 113.1, 130.3, 157.8; ESI-MS: *m/z* 159 [M + H]<sup>+</sup>.



**3aq**

**Pyrazine-2,3-dicarbonitrile (3ao):**<sup>22</sup> White solid, 62 mg (95%), m.p.: 132–134 °C (Lit. m.p.: 133–134 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 8.92 (s, 2H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 112.6, 134.0, 147.1; ESI-MS: *m/z* 131 [M + H]<sup>+</sup>.

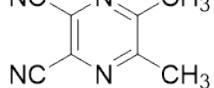


**3ar**

**5,6-Diphenylpyrazine-2,3-dicarbonitrile (3ap):**<sup>10</sup> Brown solid, 134 mg (95%), m.p.: 210–212 °C (Lit. m.p.: 209–210 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 7.35 (t, *J* = 7.6 Hz, 4H), 7.44 (t, *J* = 7.4 Hz, 2H), 7.51 (d, *J* = 7.7 Hz, 4H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 113.1, 128.8, 129.8, 131.1, 135.2, 155.4; ESI-MS: *m/z* 283 [M + H]<sup>+</sup>.

**5,6,7,8-Tetrahydroquinoxaline-2,3-dicarbonitrile (3aq):**<sup>23</sup> White solid, 91 mg (99%), m.p.: 140–142 °C (Lit. m.p.: 138–139 °C); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 1.96–1.99 (m, 4H), 3.05–3.08 (m, 4H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 21.5, 32.2, 113.1, 130.1, 158.6. ESI-MS: *m/z* 185 [M + H]<sup>+</sup>.

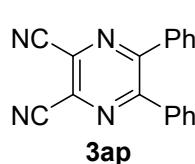
**5,6-Dimethyl-2,3-dihydropyrazine (3ar):**<sup>24</sup> Liquid, 100 mg (91%); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ (ppm) 2.09 (s, 6H), 3.30 (s, 4H); <sup>13</sup>C {<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>): δ (ppm) 23.1, 44.7, 159.3. ESI-MS: *m/z* 111 [M + H]<sup>+</sup>.



**3an**

## References:

1. S. V. More, M. N. V. Sastry, C.-C. Wang and C.-F. Yao, Molecular iodine: A powerful catalyst for the easy and efficient synthesis of quinoxalines, *Tetrahedron Lett.*, 2005, **46**, 6345–6348.
2. M. M. Heravi, S. Taheri, K. Bakhtiari and H. A. Oskooie, On Water: A practical and efficient synthesis of quinoxaline derivatives catalyzed by CuSO<sub>4</sub>.5H<sub>2</sub>O, *Catal. Commun.*, 2007, **8**, 211–214.
3. R. S. Bhosale, S. R. Sarda, S. S. Ardhapure, W. N. Jadhav, S. R. Bhusare and R. P. Pawar, An efficient protocol for the synthesis of quinoxaline

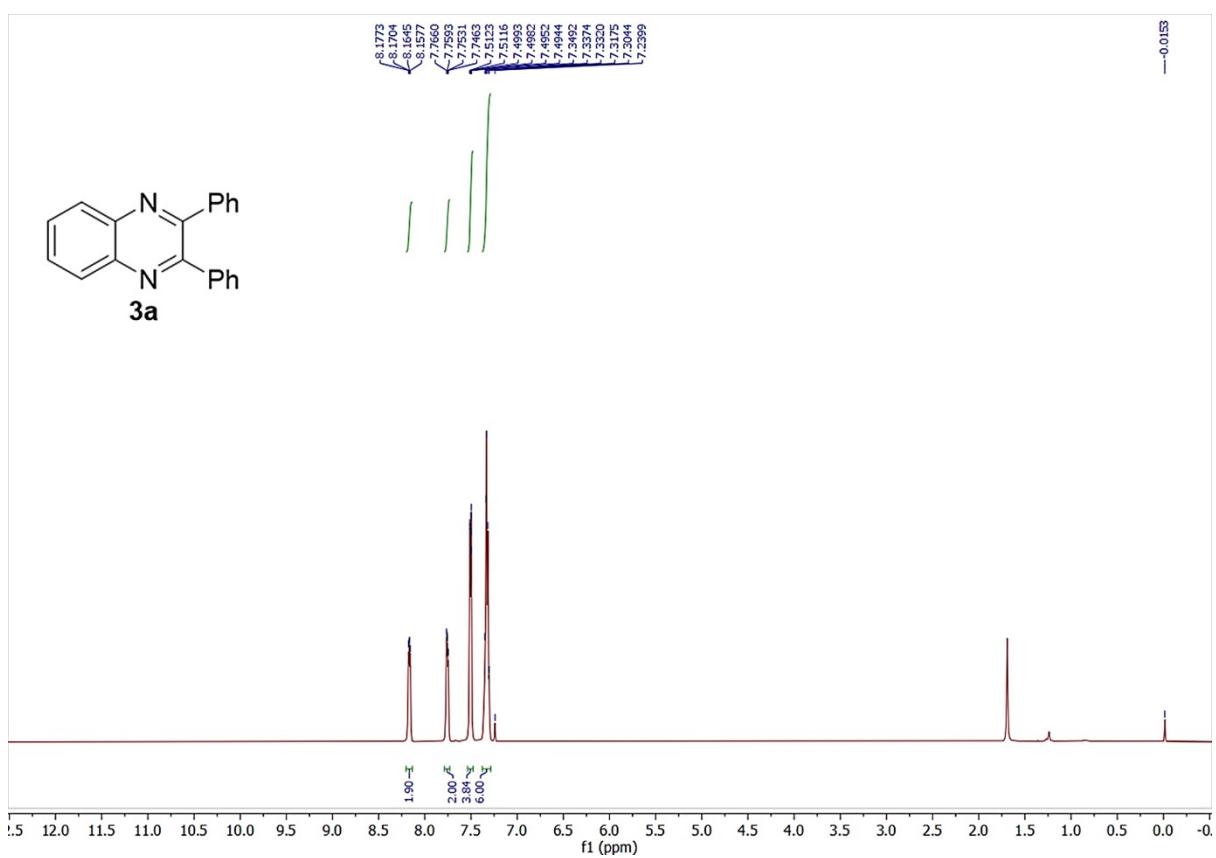


**3ap**

- derivatives at room temperature using molecular iodine as the catalyst, *Tetrahedron Lett.*, 2005, **46**, 7183–7186.
- 4. M. M. Heravi, K. Bakhtiari, H. A. Oskooie and S. Taheri, MnCl<sub>2</sub>-Promoted Synthesis of Quinoxaline Derivatives at Room Temperature, *Heteroat. Chem.*, 2008, **19**, 218–220.
  - 5. K. J. Tamuli, S. Nath and M. Bordoloi, In water organic synthesis: Introducing itaconic acid as a recyclable acidic promoter for efficient and scalable synthesis of quinoxaline derivatives at room temperature, *J. Heterocycl. Chem.*, 2021, **58**, 983–1002.
  - 6. S. Gajurel, R. Sarkar, F. K. Sarkar, L. Kyndiah and A. K. Pal, A sustainable route toward the synthesis of highly substituted imidazole and quinoxaline derivatives using CuO@g-C<sub>3</sub>N<sub>4</sub> as an efficient and reusable heterogeneous catalyst, *Appl. Organomet. Chem.*, 2023, **37**, e7159.
  - 7. A. Mishra, S. Singh, M. A. Quraishi and V. Srivastava, A catalyst-free expeditious green synthesis of quinoxaline, oxazine, thiazine, and dioxin derivatives in water under ultrasound irradiation, *Org. Prep. Proced.*, 2019, **51**, 345–356.
  - 8. W.-J. Zhou, X.-Zh. Zhang, X.-B. Sun, B. Wang, J.-X. Wang and L. Bai, Microwave-assisted synthesis of quinoxaline derivatives using glycerol as a green solvent, *Russ. Chem. Bull.*, 2013, **62**, 1244–1247.
  - 9. Z. T. Bhutia, G. Prasannakumar, A. Das, M. Biswas, A. Chatterjee and M. Banerjee, A Facile, catalyst-free mechano-synthesis of quinoxalines and their in-vitro antibacterial activity study *ChemistrySelect*, 2017, **2**, 1183–1187.
  - 10. G. C. Nandi, S. Samai, R. Kumar and M. S. Singh, Silica-Gel–Catalyzed Efficient Synthesis of quinoxaline derivatives under solvent-free conditions, *Synth. Commun.*, 2011, **41**, 417–425.
  - 11. B. F. Mirjalili and M. D. Tafti, Synthesis of quinoxalines promoted by eco-friendly nano-kaoline/BF<sub>3</sub>/Fe<sub>3</sub>O<sub>4</sub> nano-catalyst under grinding condition, *Sci. Iran.*, 2017, **24**, 3014–3021.
  - 12. S. Das, P. Das, S. Maity, P. Ghosh and A. Dutta, Mechanochemical reaction of ninhydrin with aromatics, enols and amines: Synthesis, crystal structure and supramolecular self-assembly of cyclic and acyclic adducts, *Results Chem.*, 2023, **5**, 100713.
  - 13. J. Li, D. Jiang, J. Chen, M. Liu, J. Ding and H. Wu, Eco-friendly synthesis of quinoxaline derivatives by grinding under solvent-free conditions, *J. Heterocycl. Chem.*, 2011, **48**, 403–406.
  - 14. Z. Wu, X. Li, P. Jiang, G. Feng, L.-A. Cao, X.-M. Wang, R. Tan and E. Feng, Mechanical milling promotes the preparation of catalyst lanthanum dodecyl sulfate and green solvent-free grinding for synthesis of N-heterocyclic derivatives, *Appl. Catal., A*, 2024, **684**, 119893.
  - 15. H. Nagarajaiah, A. K. Mishra and J. N. Moorthy, Mechanochemical solid-state synthesis of 2-aminothiazoles, quinoxalines and benzoylbenzofurans from ketones by one-pot sequential acid- and base-mediated reactions, *Org. Biomol. Chem.*, 2016, **14**, 4129–4135.
  - 16. J. Peralta-Cruz, M. Díaz-Fernández, A. Ávila-Castro, D. Ortegón-Reyna and A. Ariza-Castolo, An experimental and theoretical study of intramolecular regioselective

- oxidations of 6-substituted 2,3-dimethylquinoxaline derivatives, *New J. Chem.*, 2016, **40**, 5501–5515.
- 17. S. Samanta, A. Das Gupta and A. K. Mallik, An expedient “on-water” synthesis of quinoxalines, *Monatshefte Fur Chem. - Chem. Mon.*, 2014, **145**, 1669–1673.
  - 18. C.-H. Tseng, Y.-R. Chen, C.-C. Tzeng, W. Liu, C.-K. Chou, C.-C. Chiu and Y.-L. Chen, Discovery of indeno[1,2-b]quinoxaline derivatives as potential anticancer agents, *Eur. J. Med. Chem.*, 2016, **108**, 258–273.
  - 19. J. J. Morales-Castellanos, K. Ramírez-Hernández, N. S. Gómez-Flores, O. R. Rodas-Suárez and J. Peralta-Cruz, *Molecules*, 2012, **17**, 5164–5176.
  - 20. M. Jafarpour, A. Rezaeifard, M. Ghahramaninezhad and T. Tabibi, Reusable  $\alpha$ -MoO<sub>3</sub> nanobelts catalyzes the green and heterogeneous condensation of 1,2-diamines with carbonyl compounds, *New J. Chem*, 2013, **37**, 2087.
  - 21. G. R. Bardajee, R. Malakooti, F. Jami, Z. Parsaei and H. Atashin, Covalent anchoring of copper-schiff base complex into SBA-15 as a heterogeneous catalyst for the synthesis of pyridopyrazine and quinoxaline derivatives, *Catal. Commun.*, 2012, **27**, 49–53.
  - 22. O. Hordiyenko, I. Rudenko, I. Zamkova, O. Denisenko, A. Biitseva, A. Arrault and A. Tolmachev, Facile synthesis of hydrazine derivatives of 5H-pyrrolo[3,4-*b*]pyrazine and 1*h*-pyrrolo[3,4-*b*]quinoxaline, *Synthesis*, 2013, **45**, 3375–3382.
  - 23. S. Antoniotti and E. Duñach, Direct and catalytic synthesis of quinoxaline derivatives from epoxides and ene-1,2-diamines, *Tetrahedron Lett*, 2002, **43**, 3971–3973.
  - 24. M. Jafarpour, A. Rezaeifard, R. Haddad and S. Gazkar, A reusable zirconium(IV) schiff base complex catalyzes highly efficient synthesis of quinoxalines under mild conditions, *Transition Met Chem*, 2012, **38**, 31–36.

## **<sup>1</sup>H NMR and <sup>13</sup>C NMR of quinoxalines**



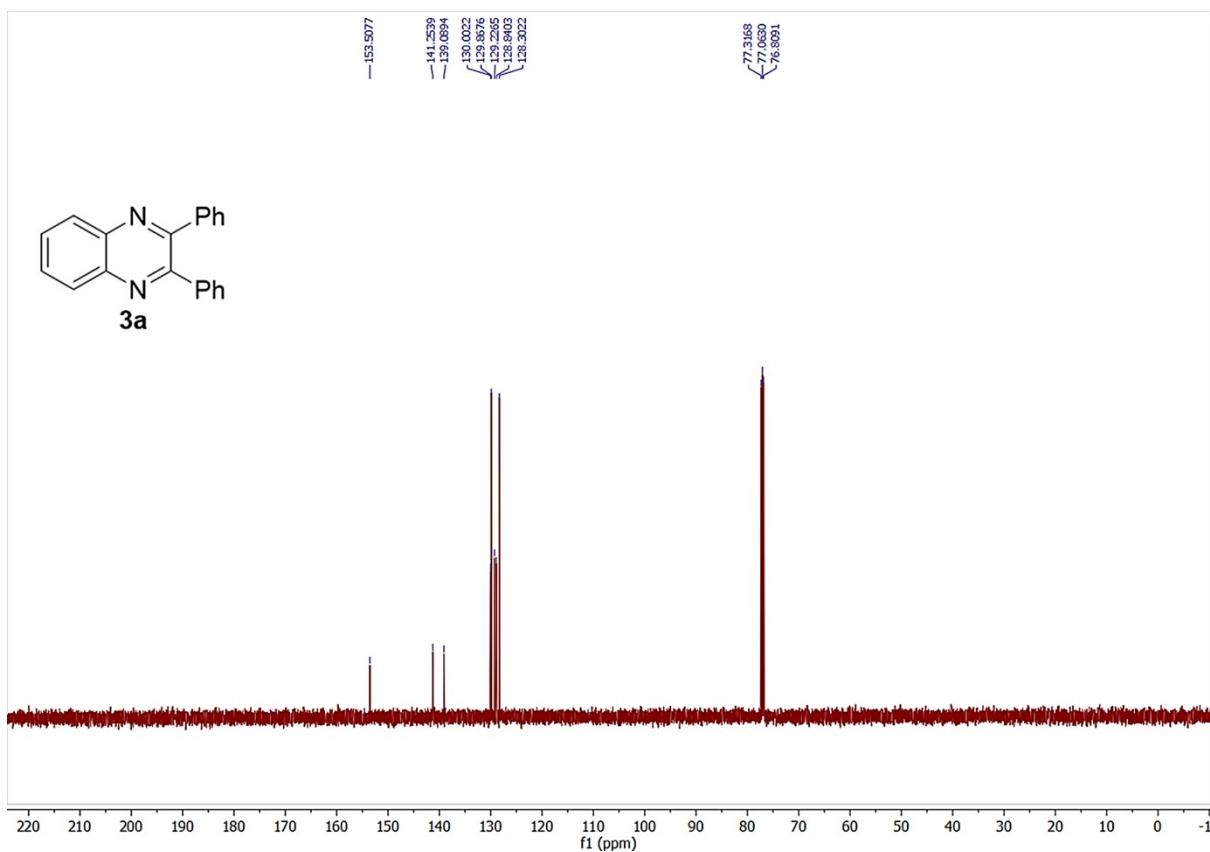
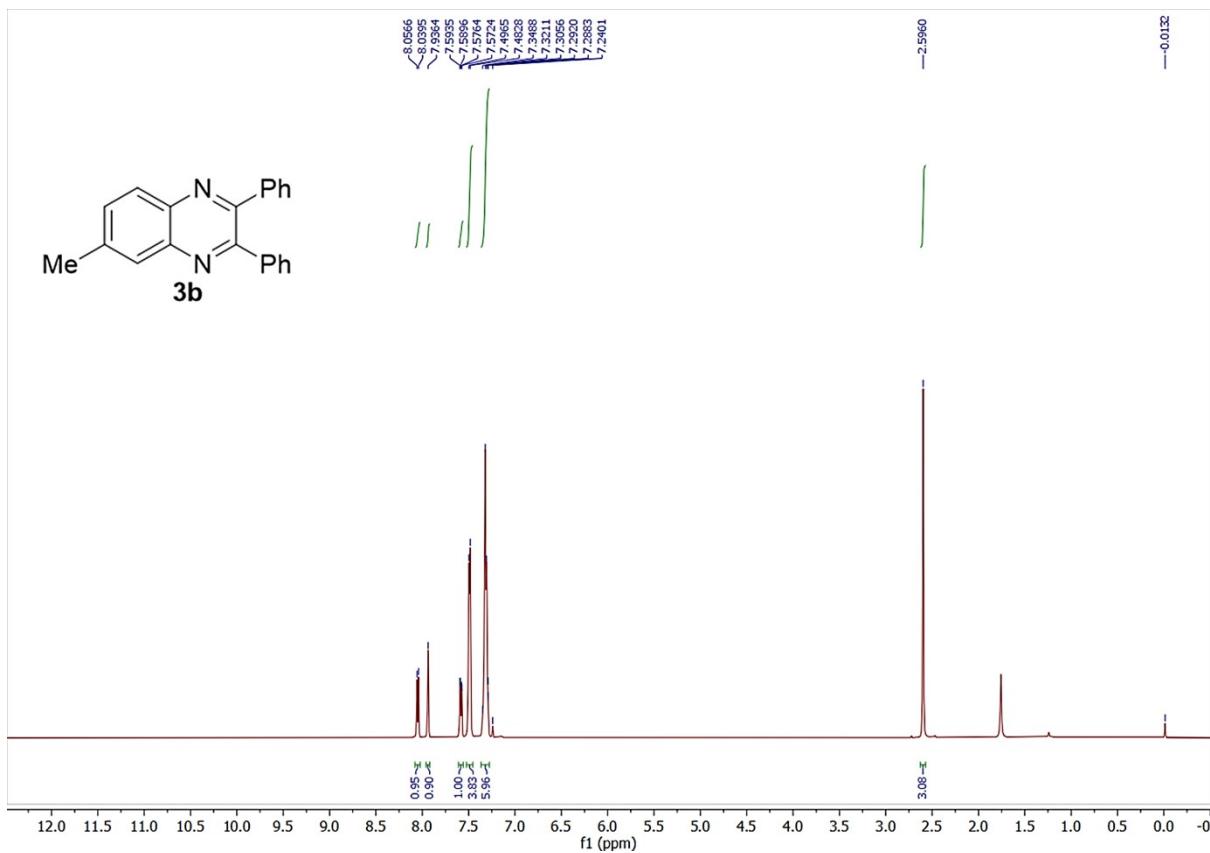


Figure S3:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3a**.



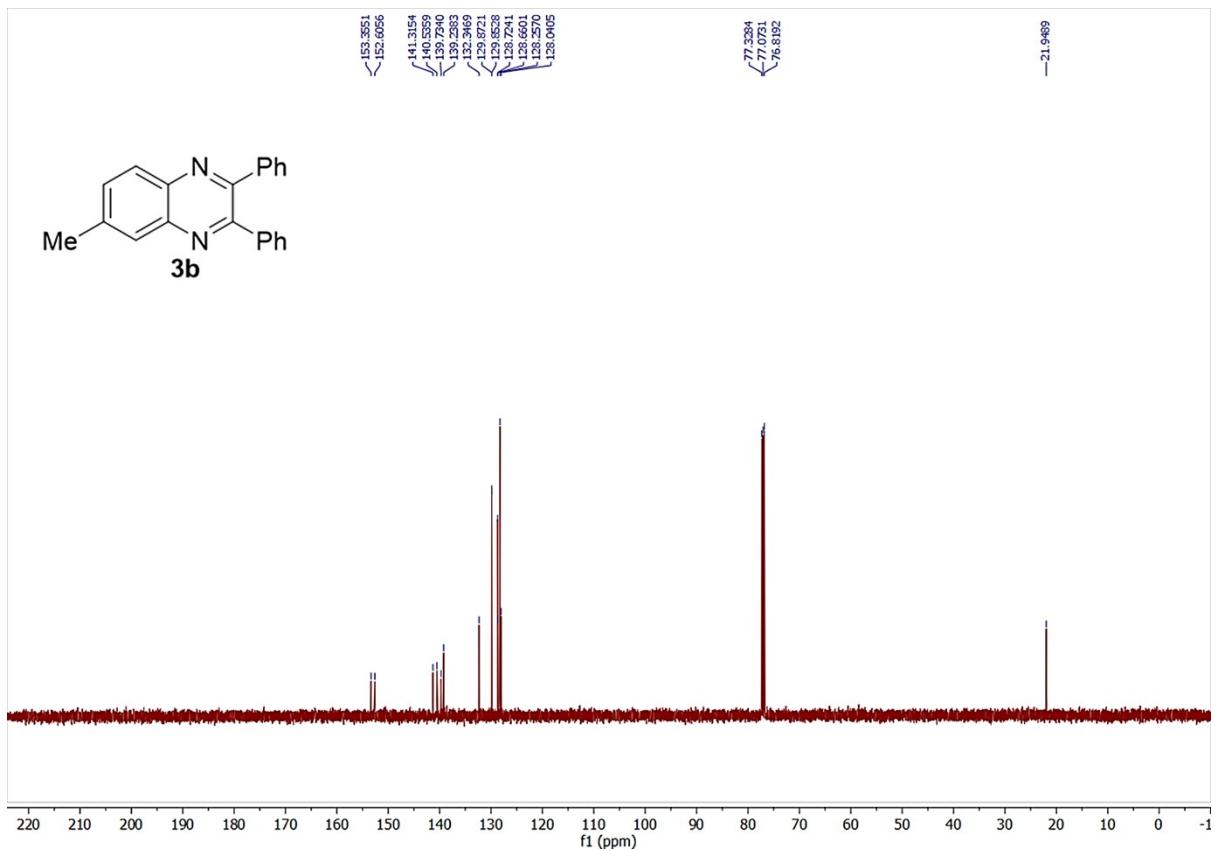
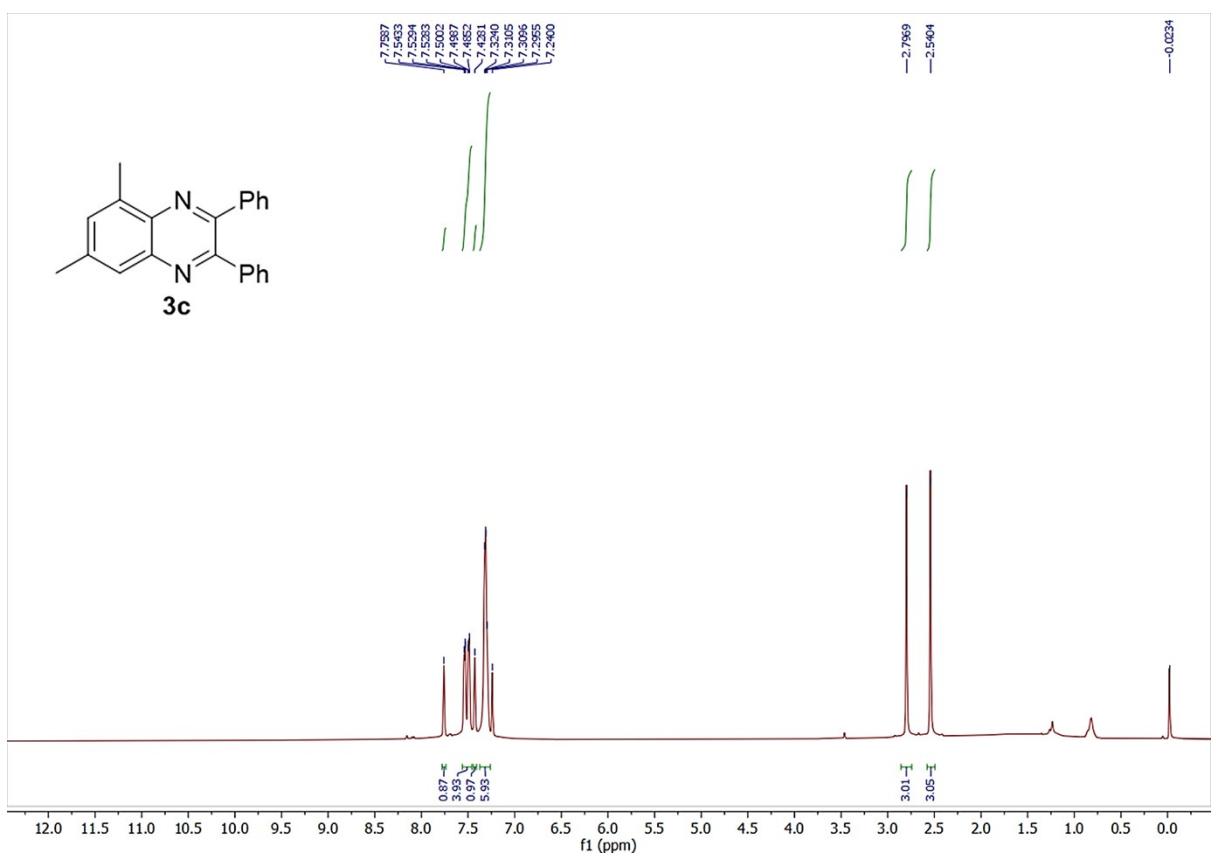


Figure S4:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3b**.



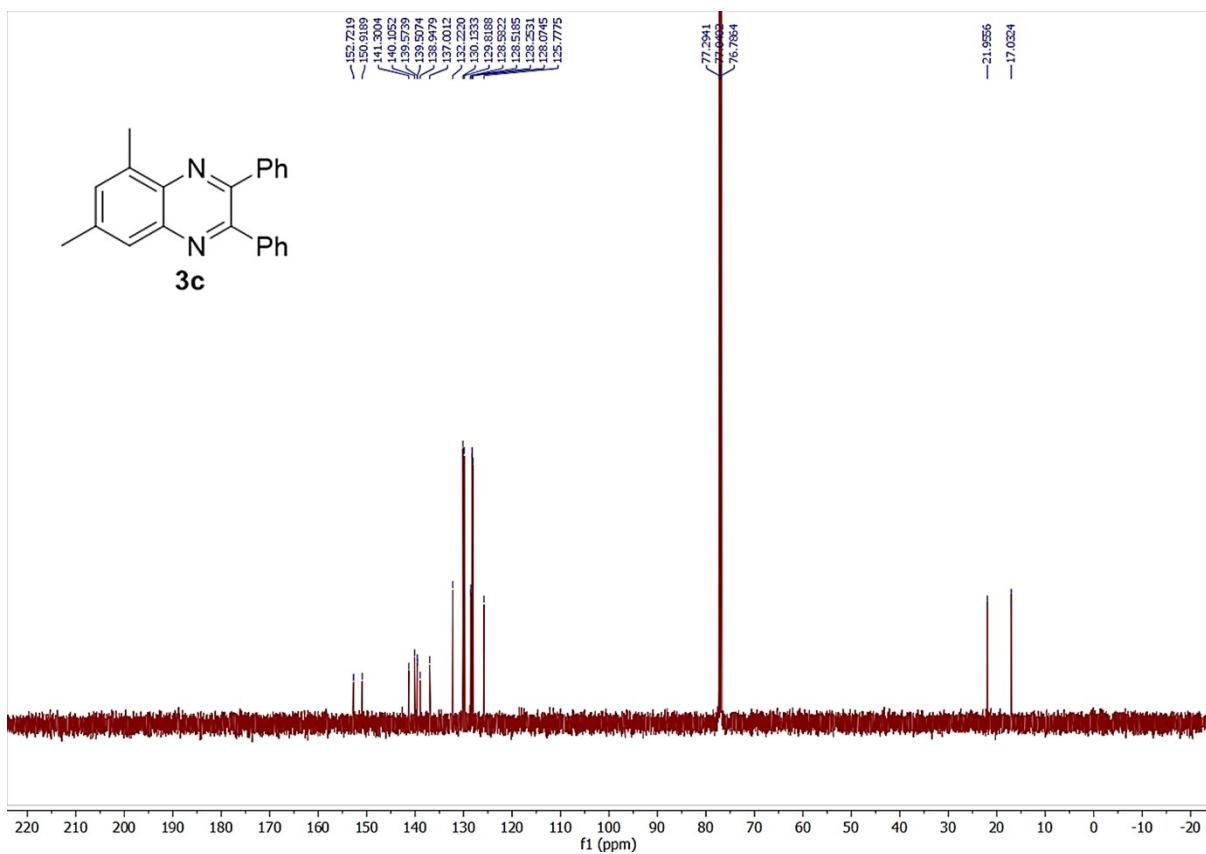
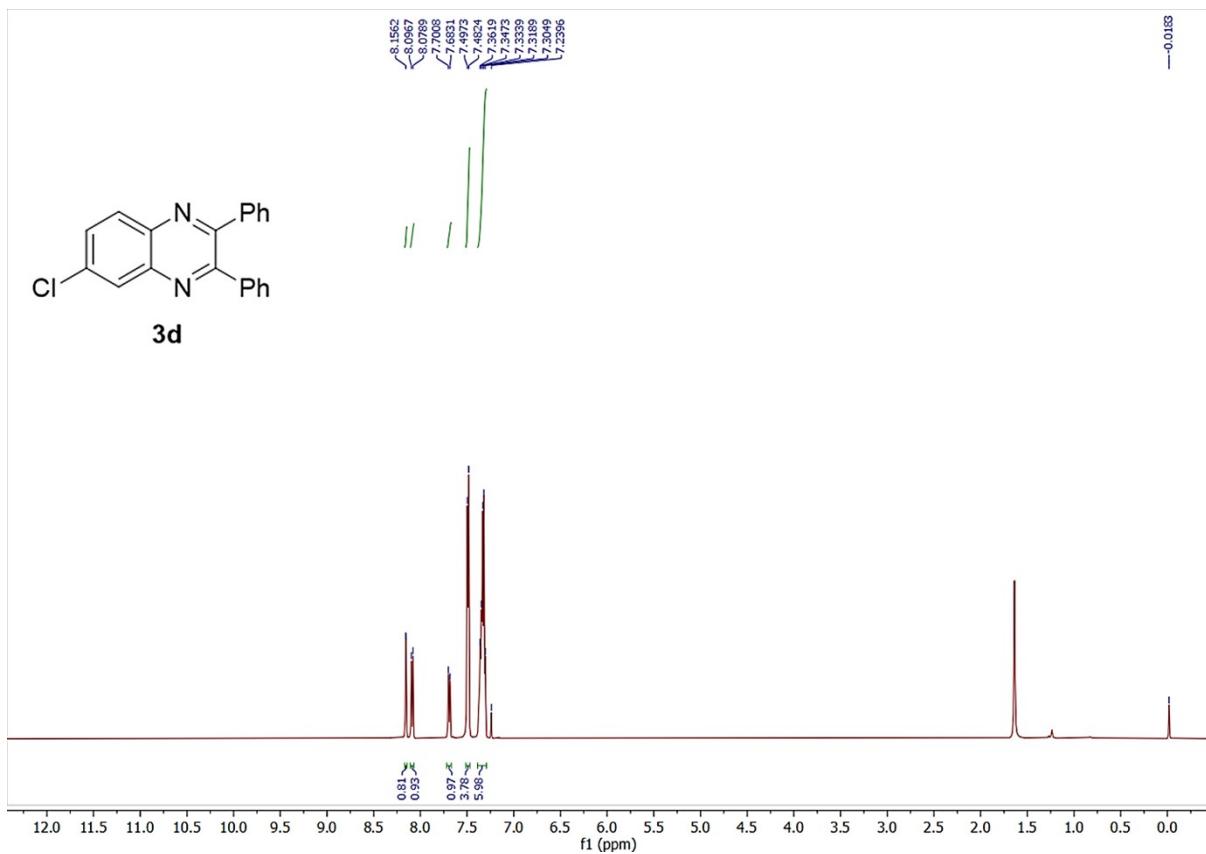


Figure S5: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound **3c**.



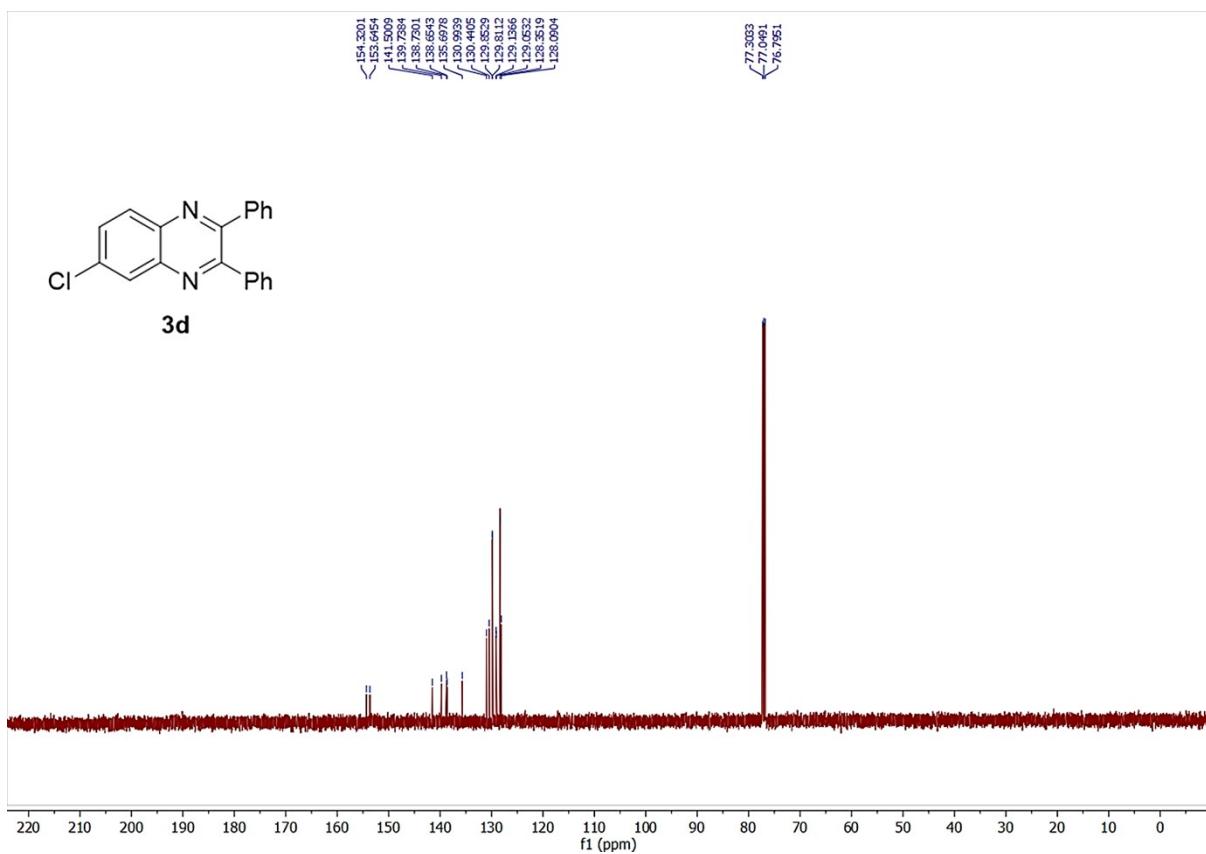
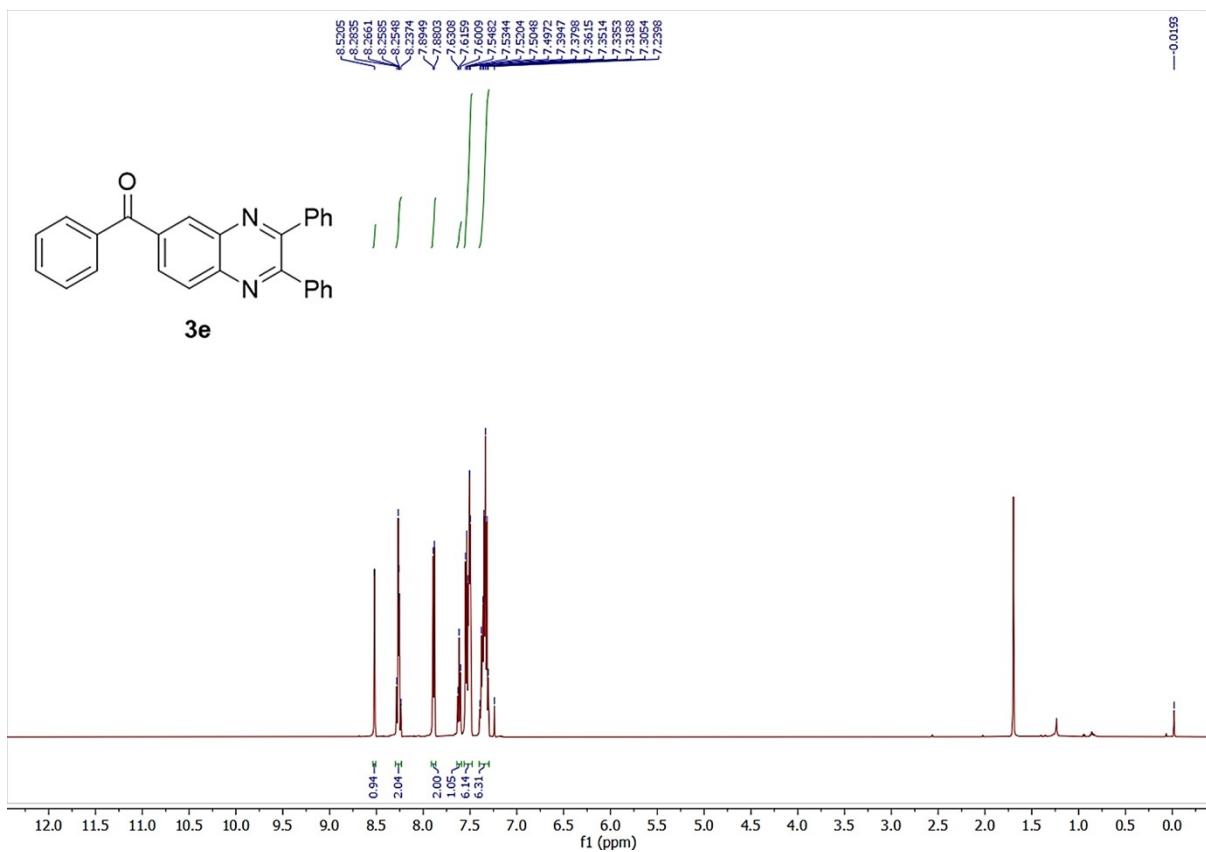


Figure S6: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound **3d**.



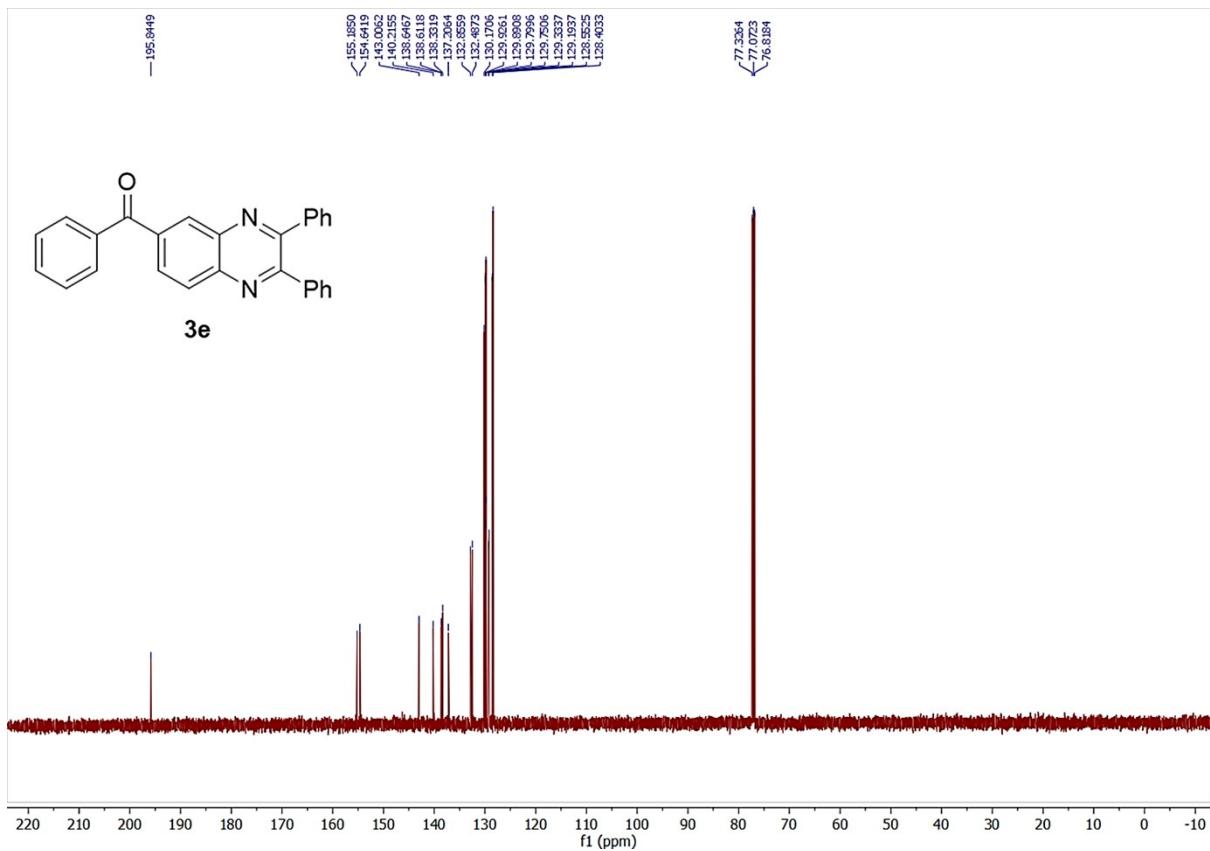
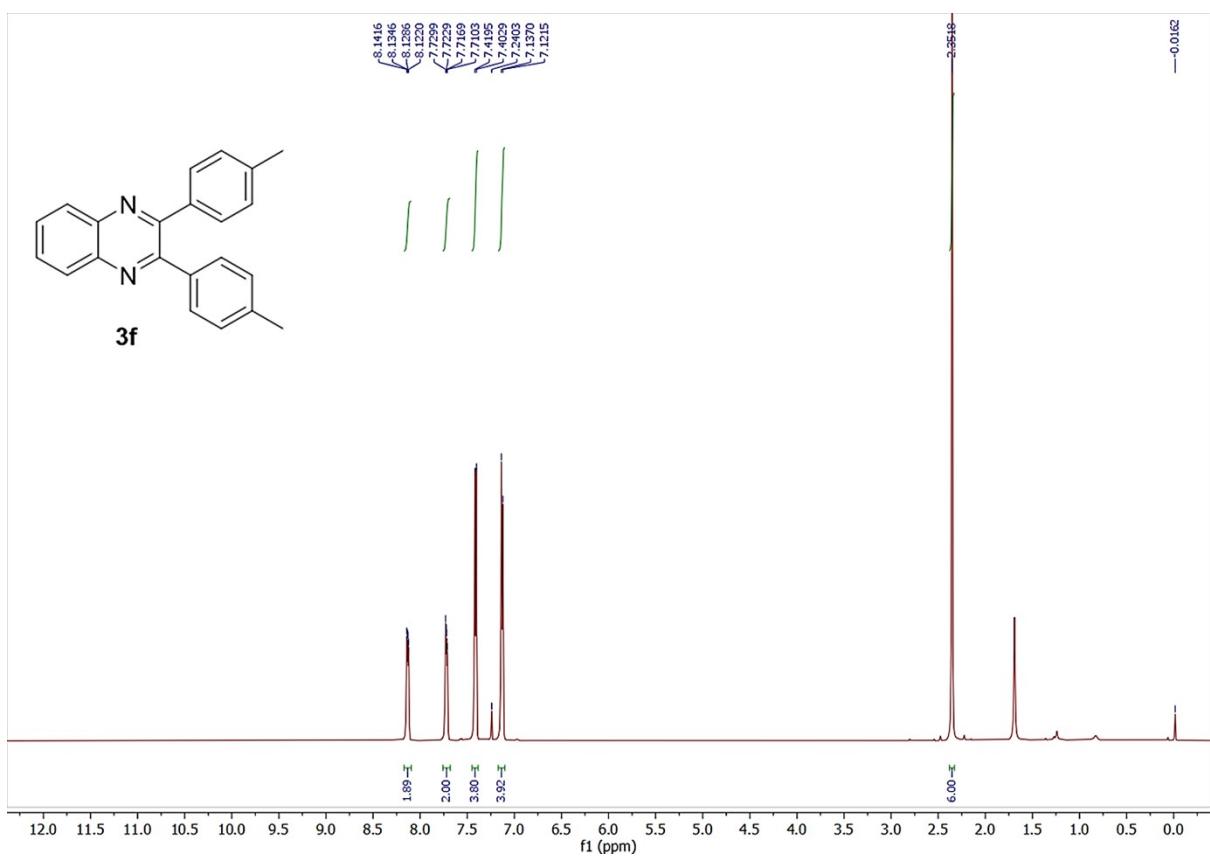


Figure S7:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3e**.



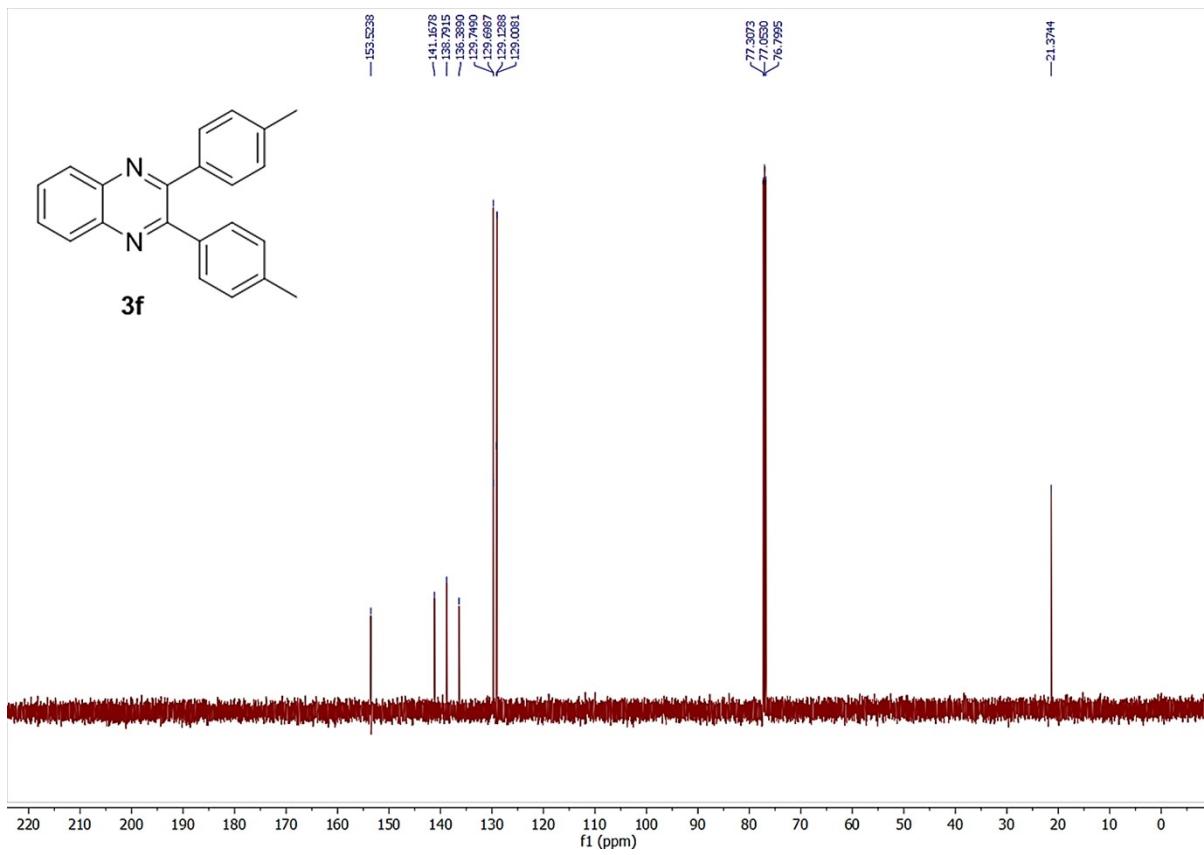
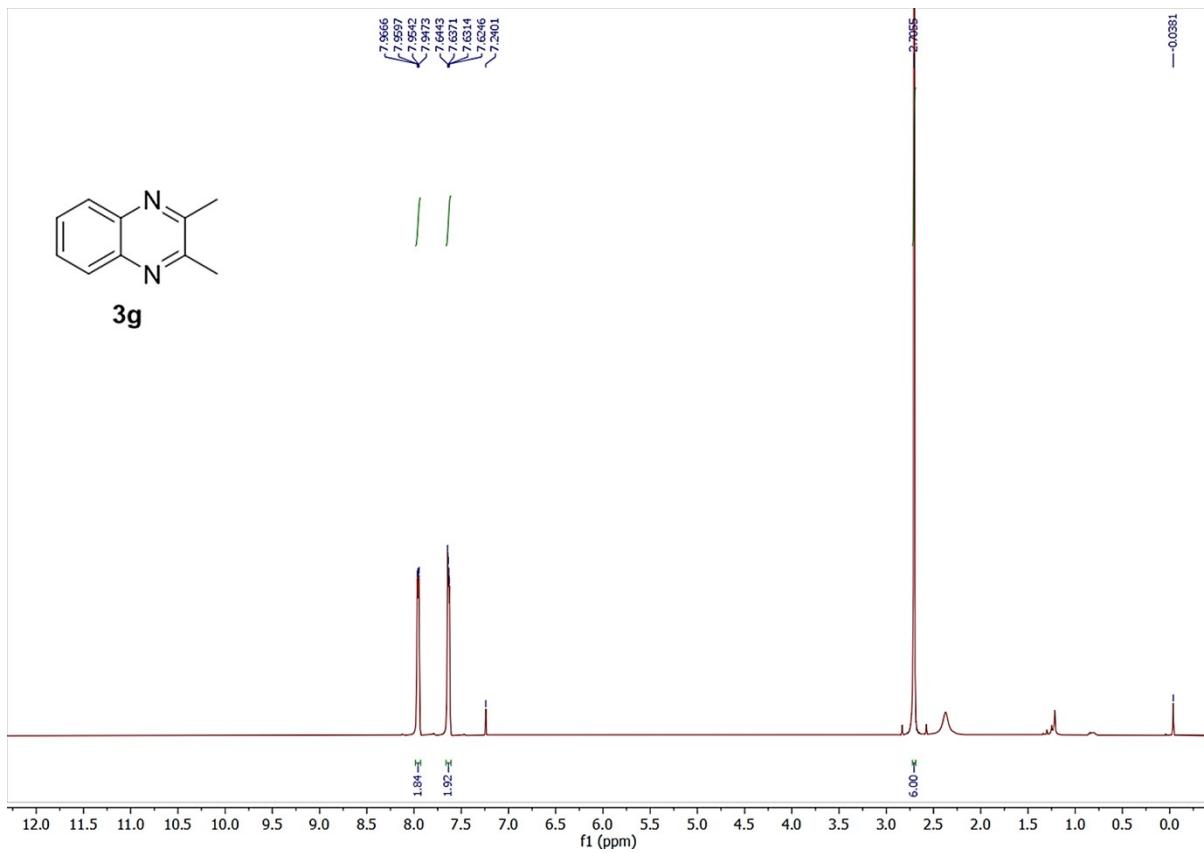


Figure S8:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3f**.



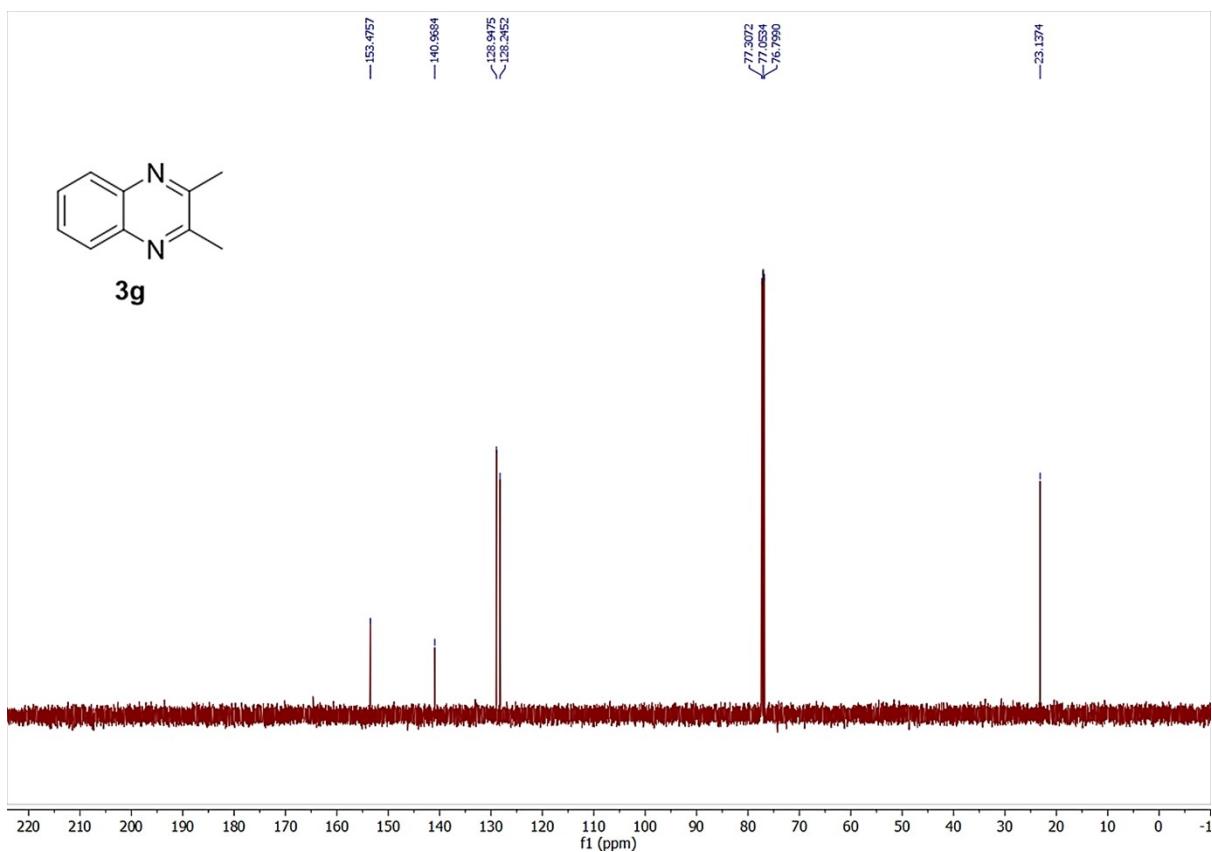
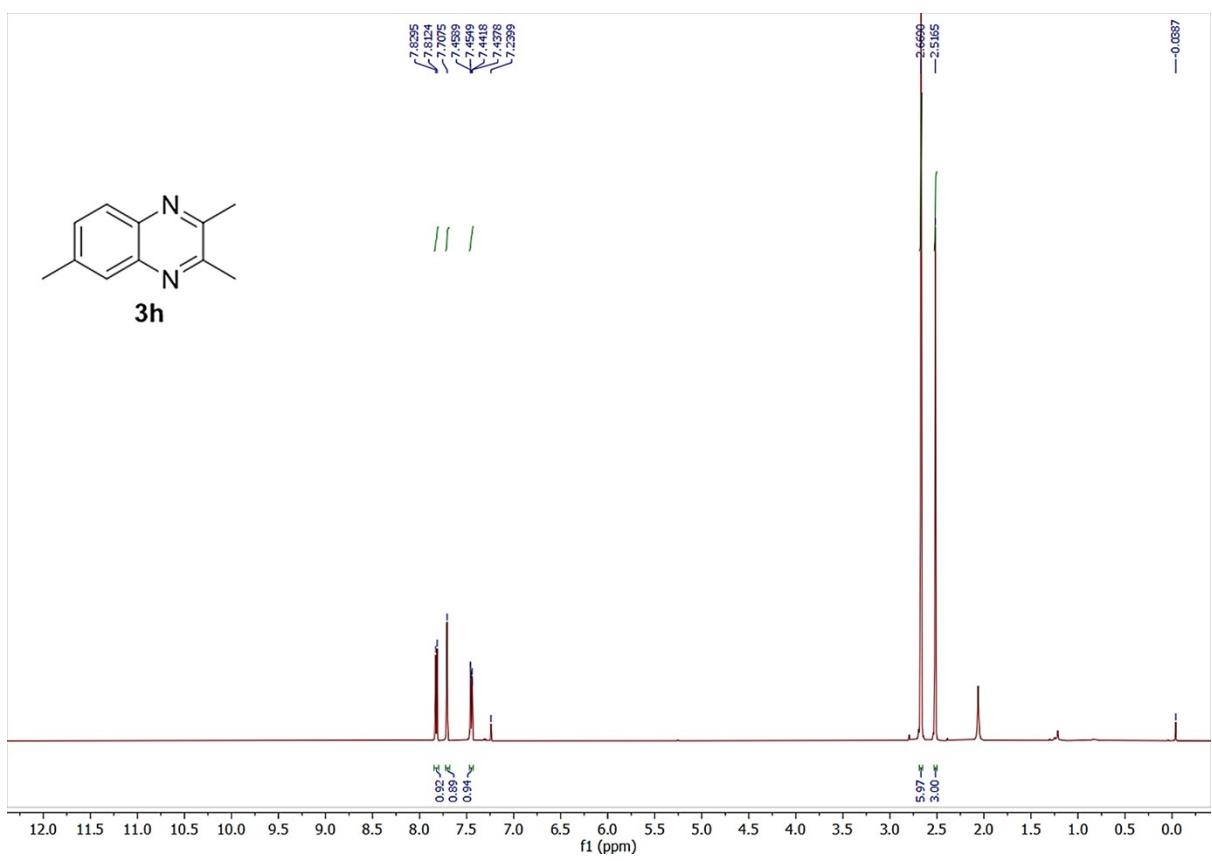


Figure S9:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3g**.



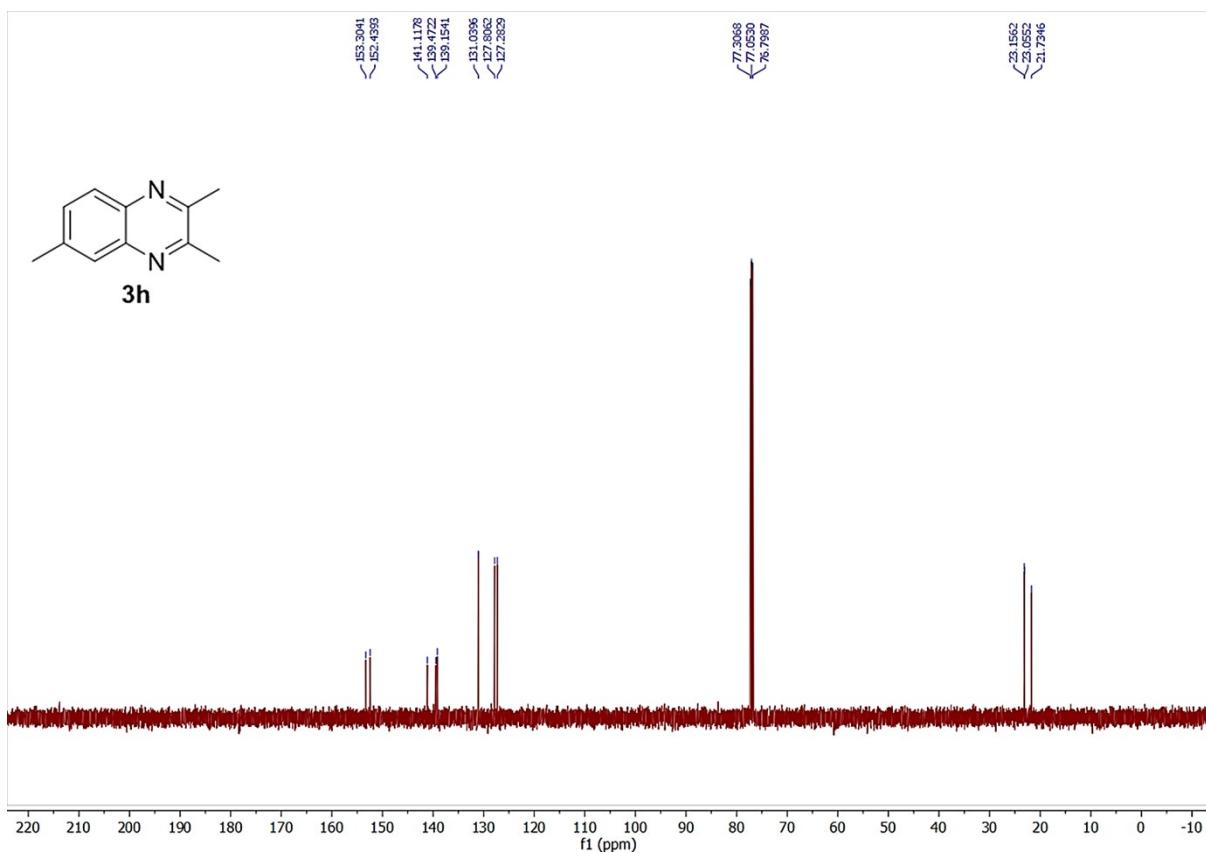
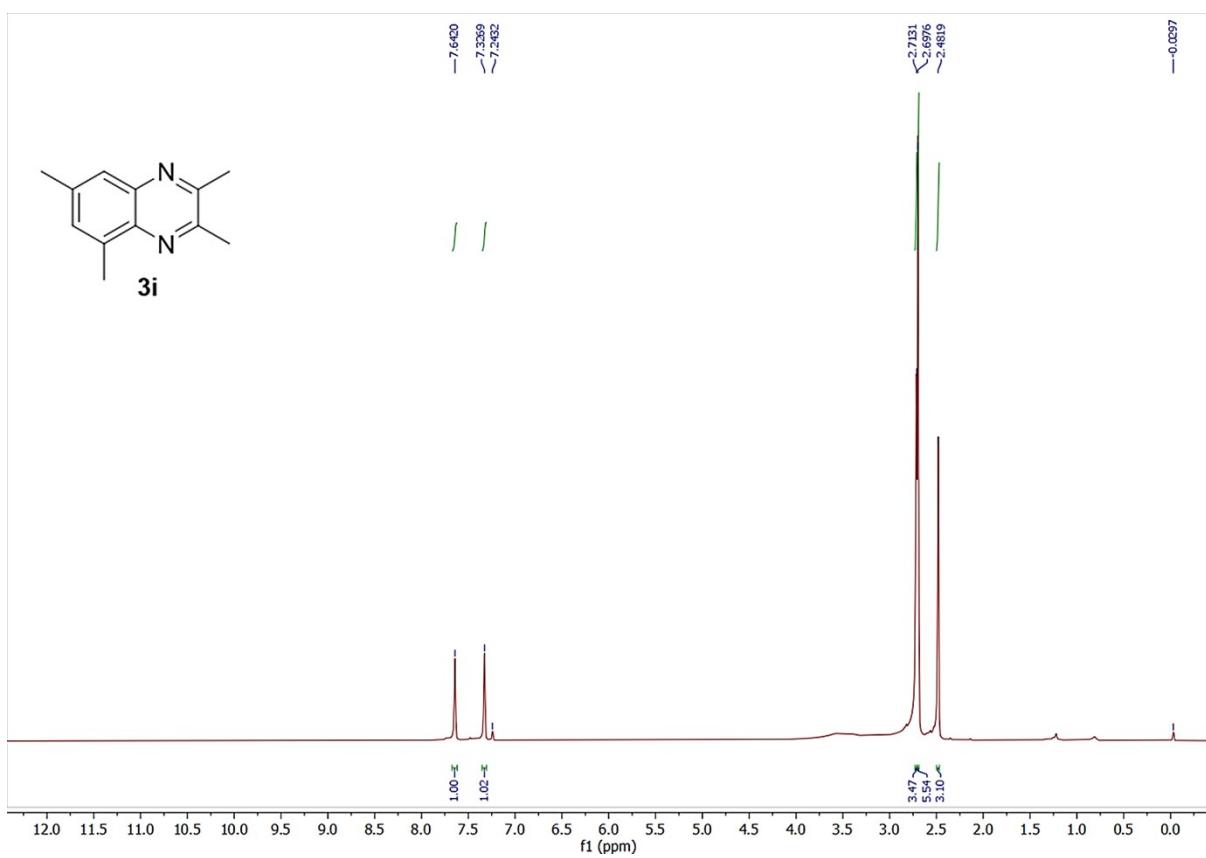


Figure S10: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound 3h.



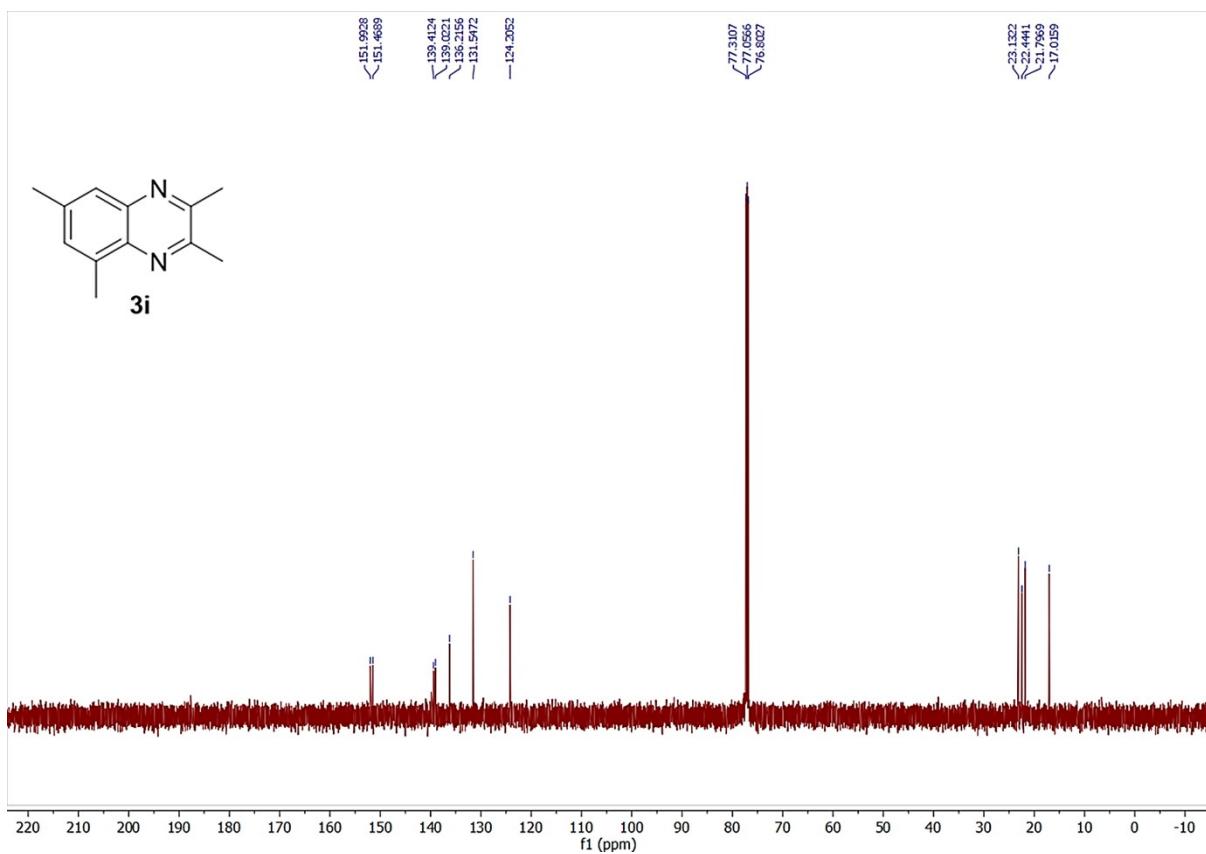
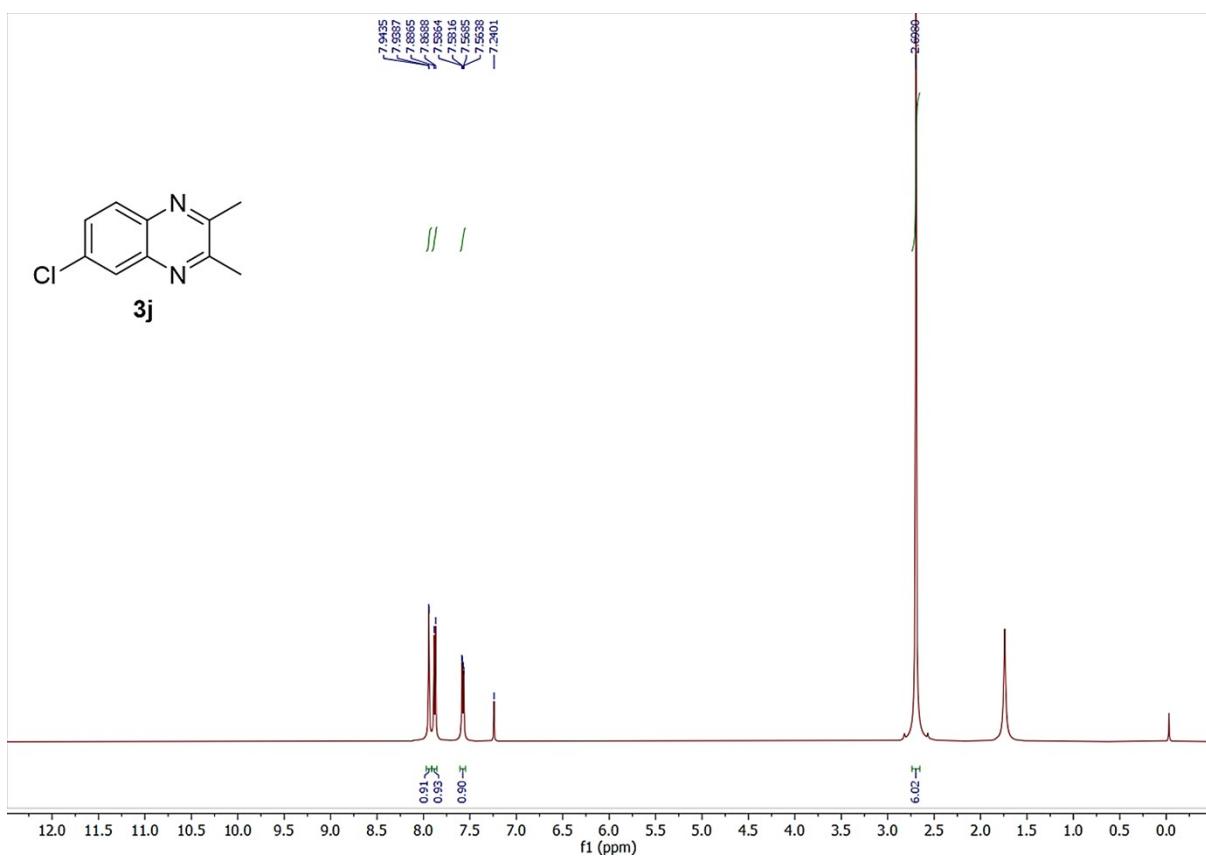


Figure S11:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3i**.



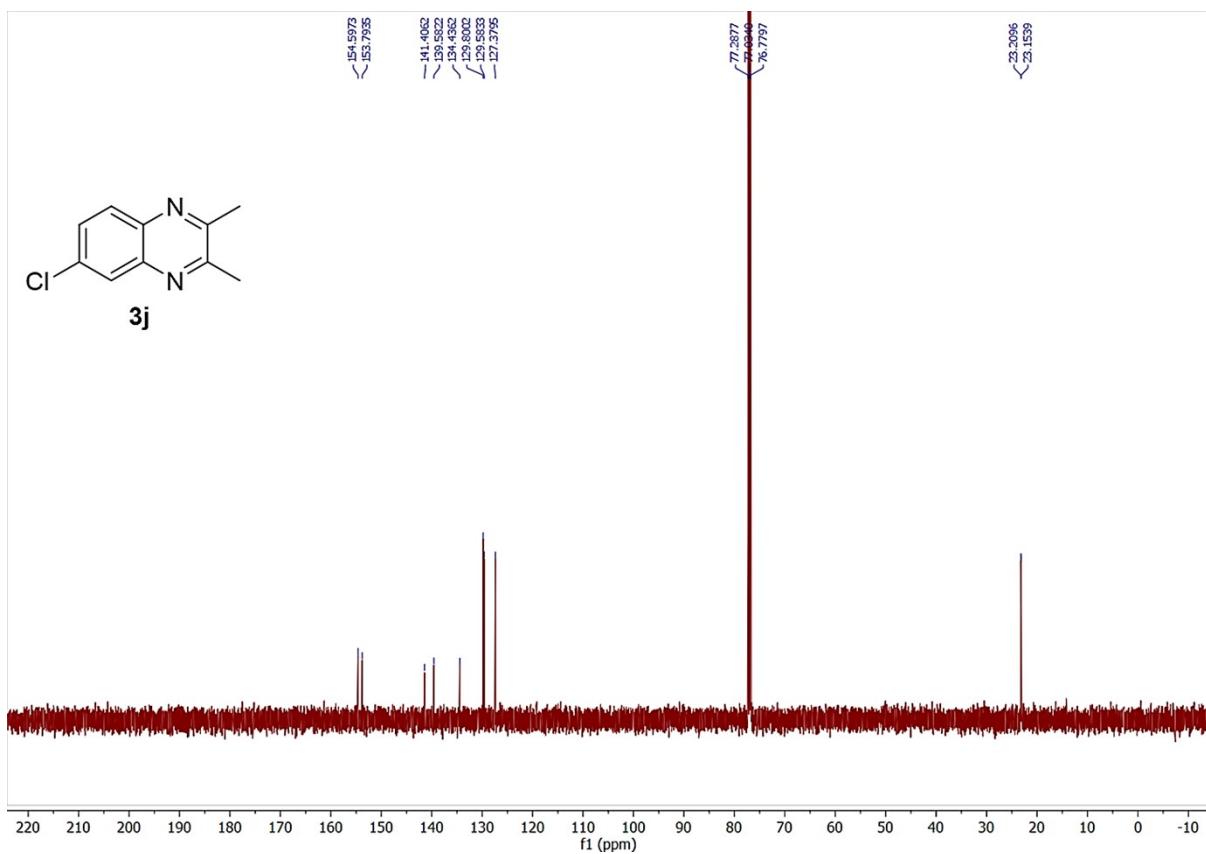
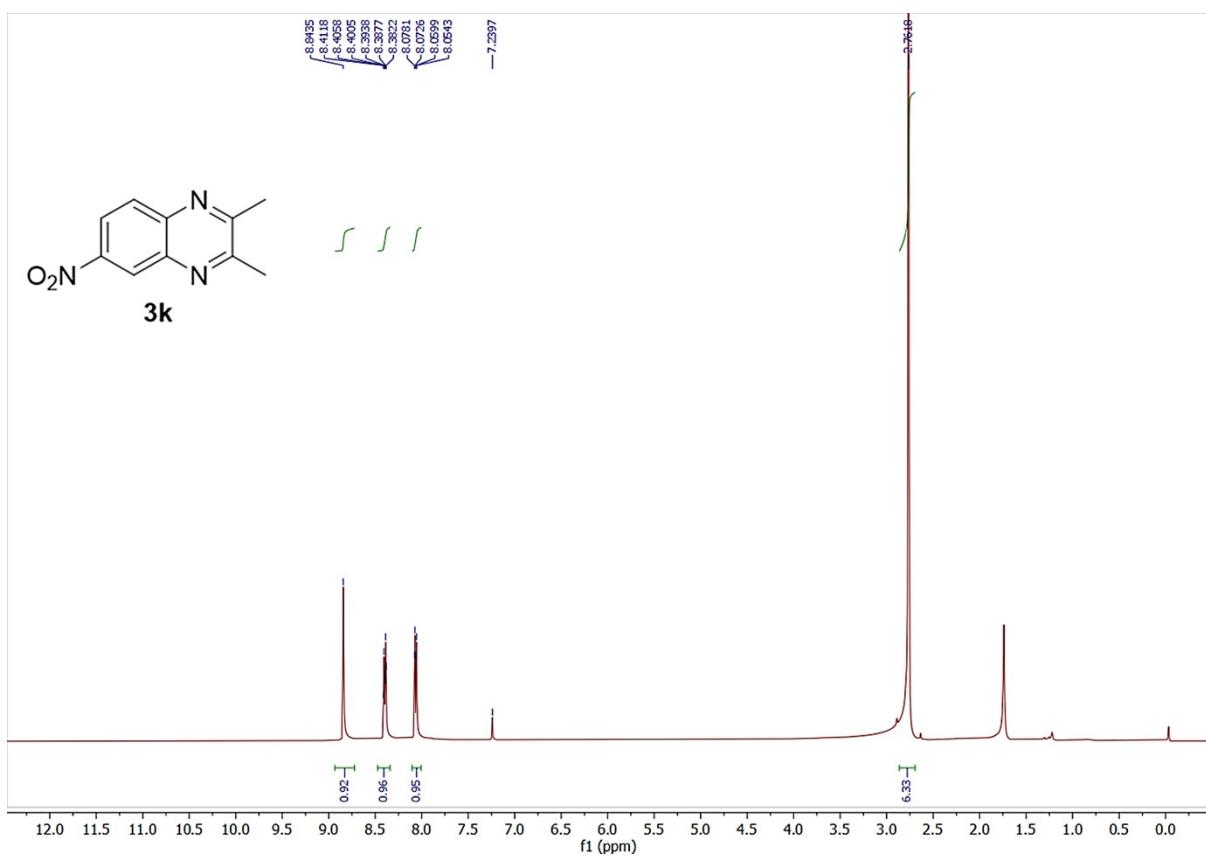


Figure S12:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3j**.



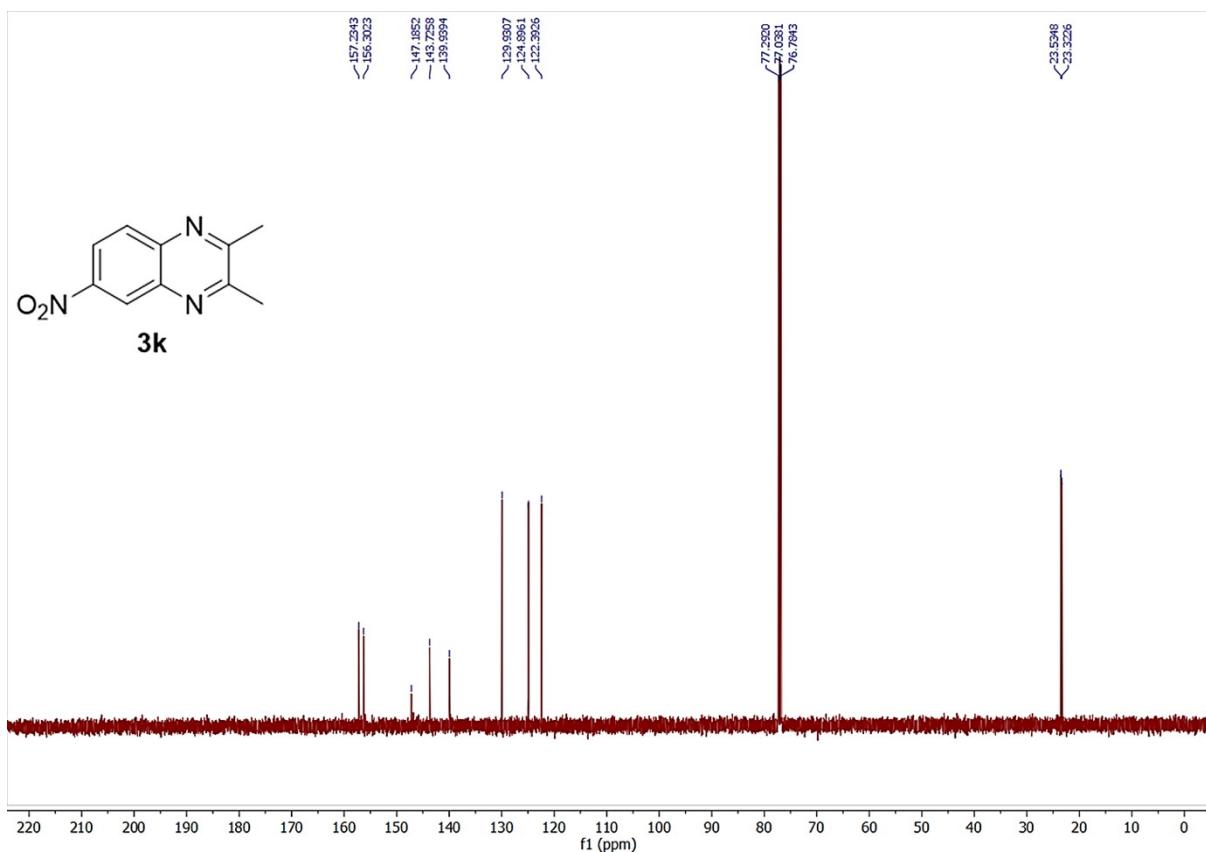
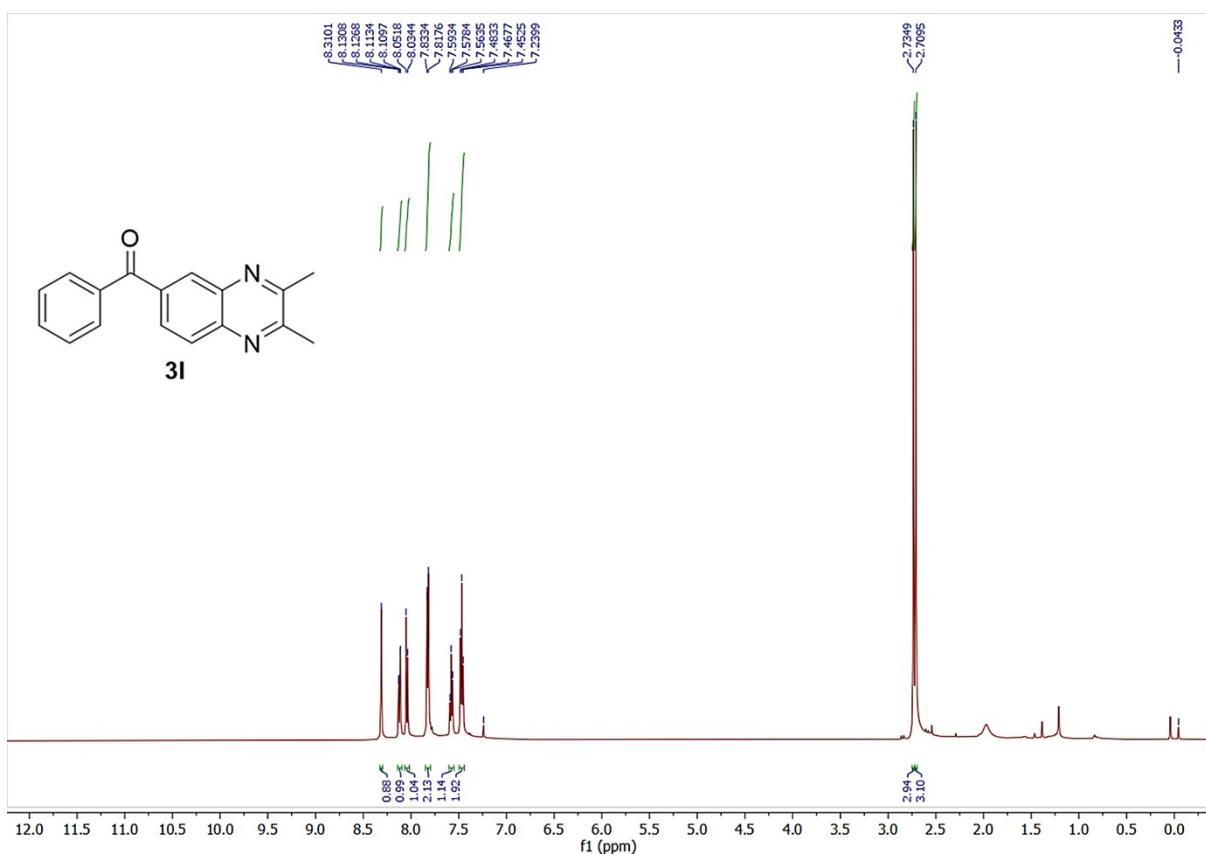


Figure S13:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3k**.



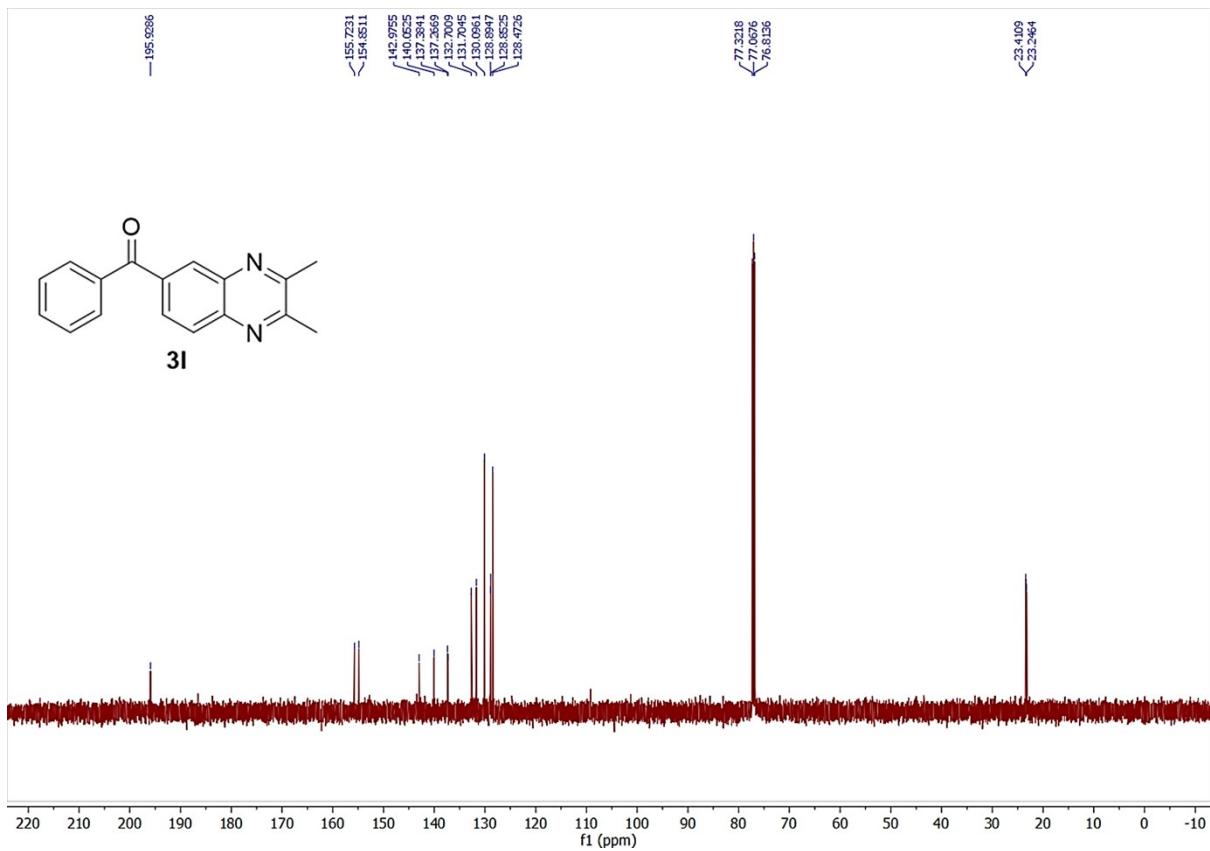
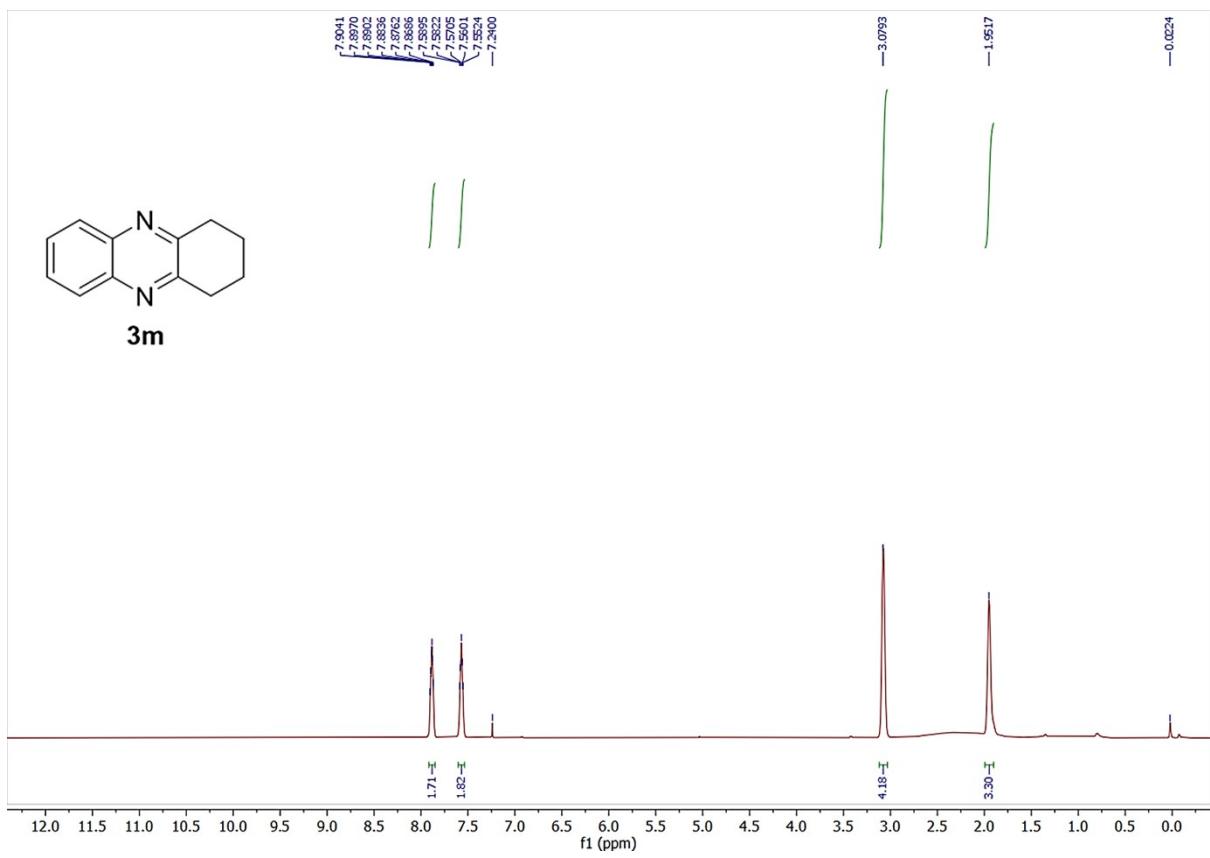


Figure S14: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound 3l.



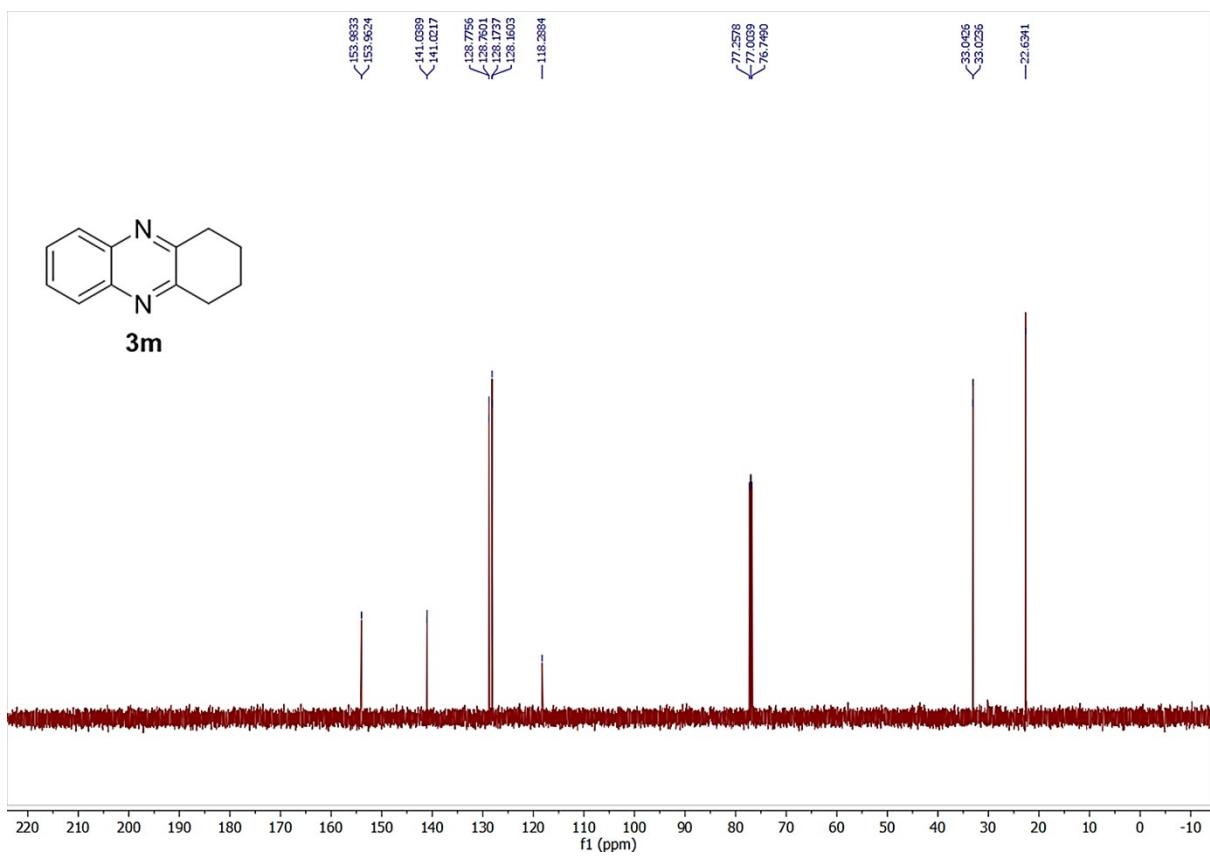
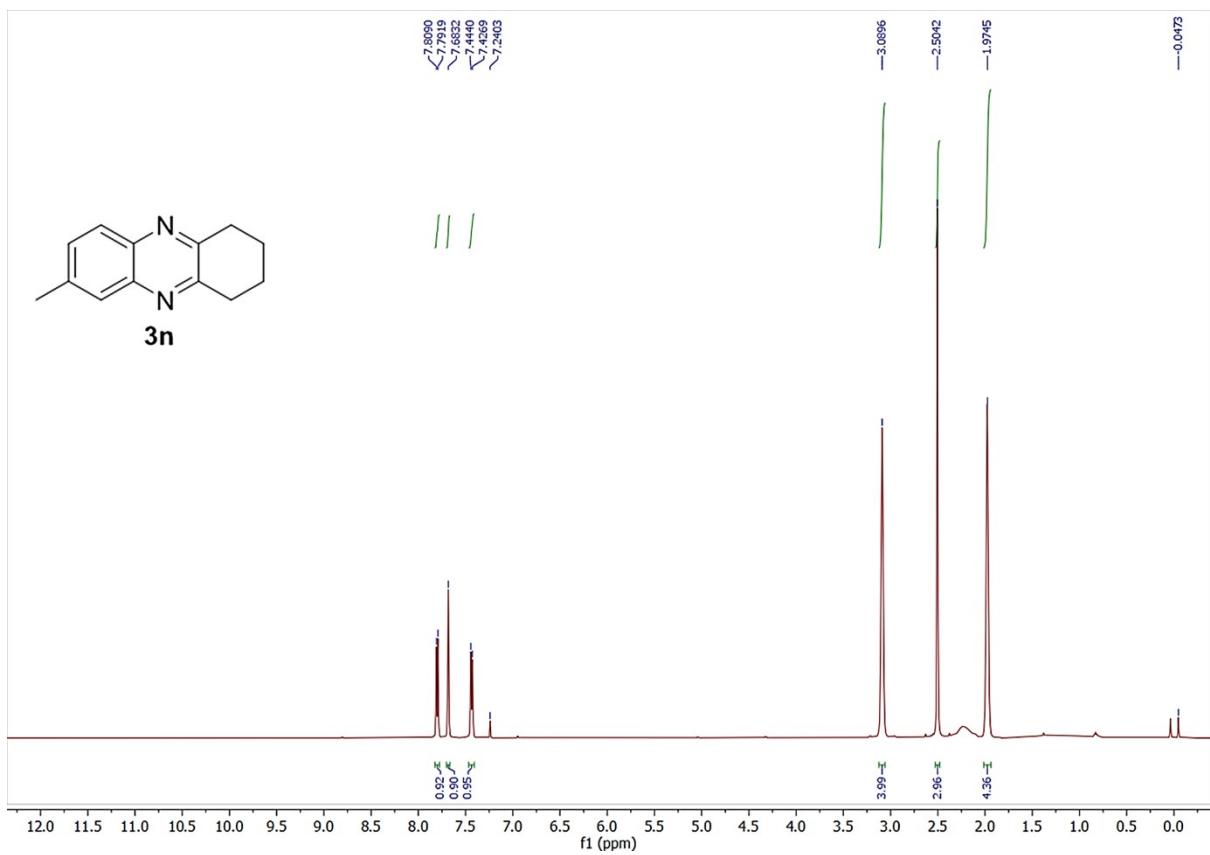


Figure S15:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3m**.



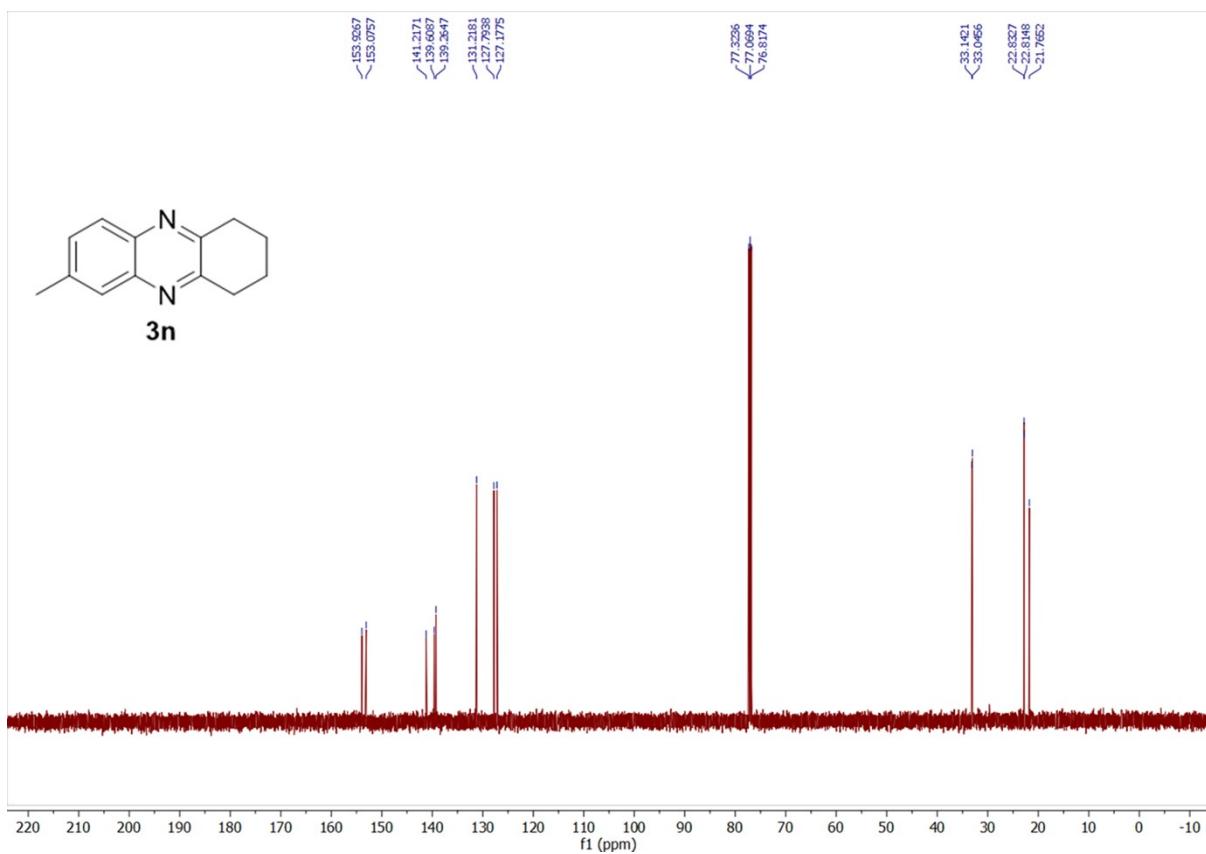
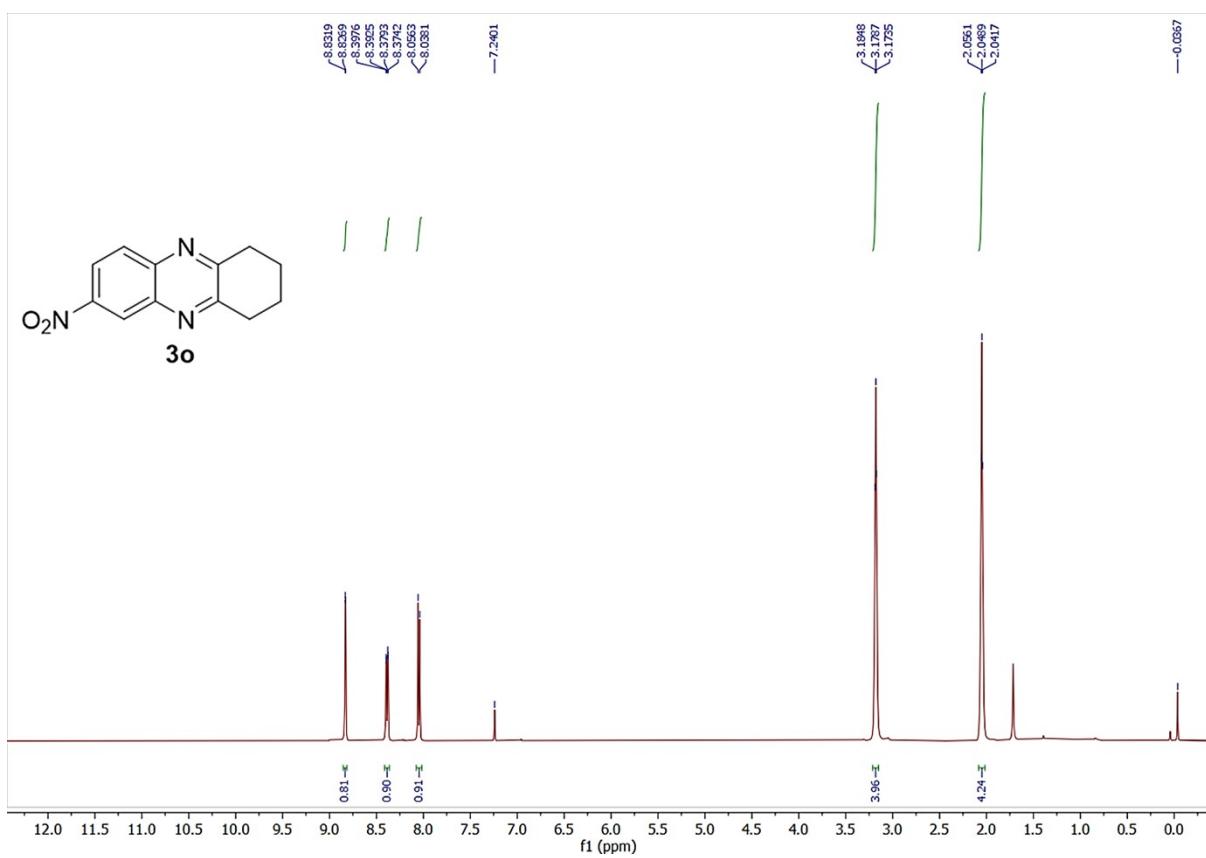


Figure S16: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound **3n**.



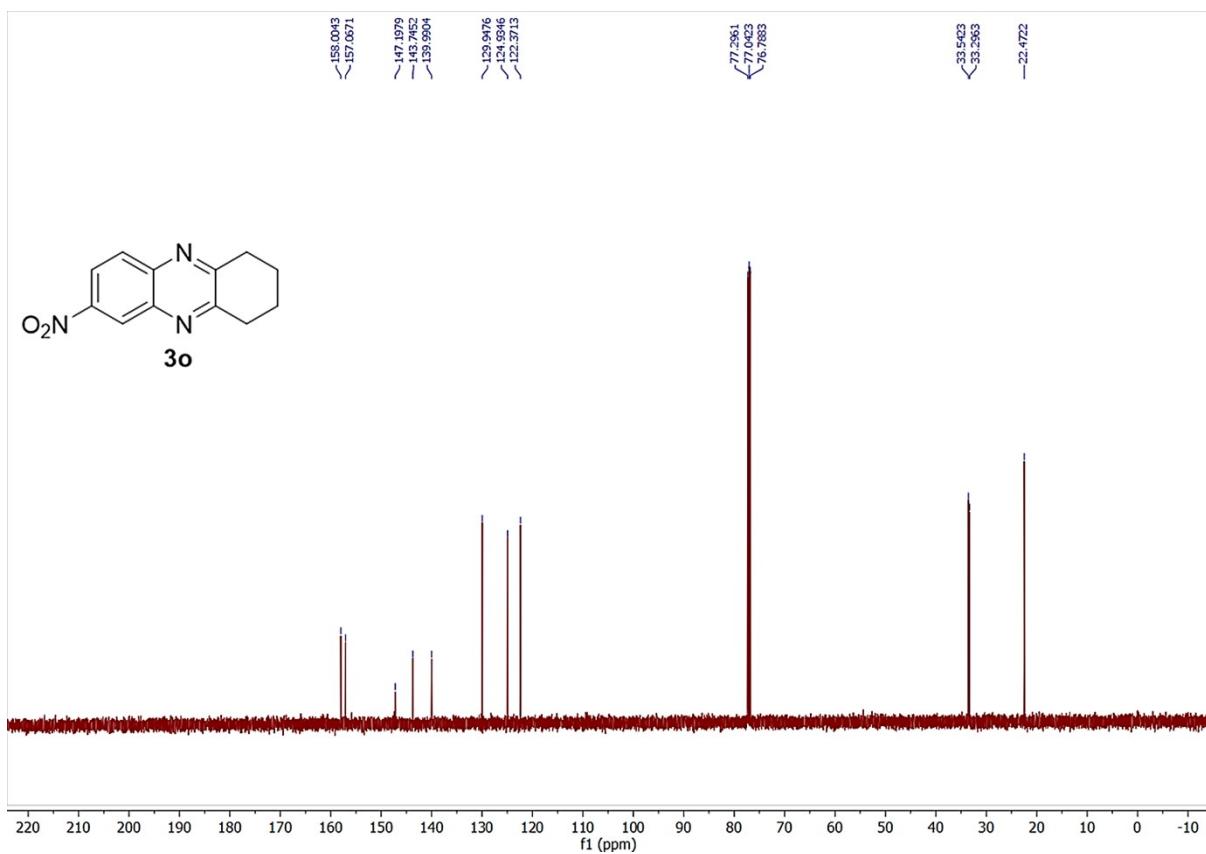
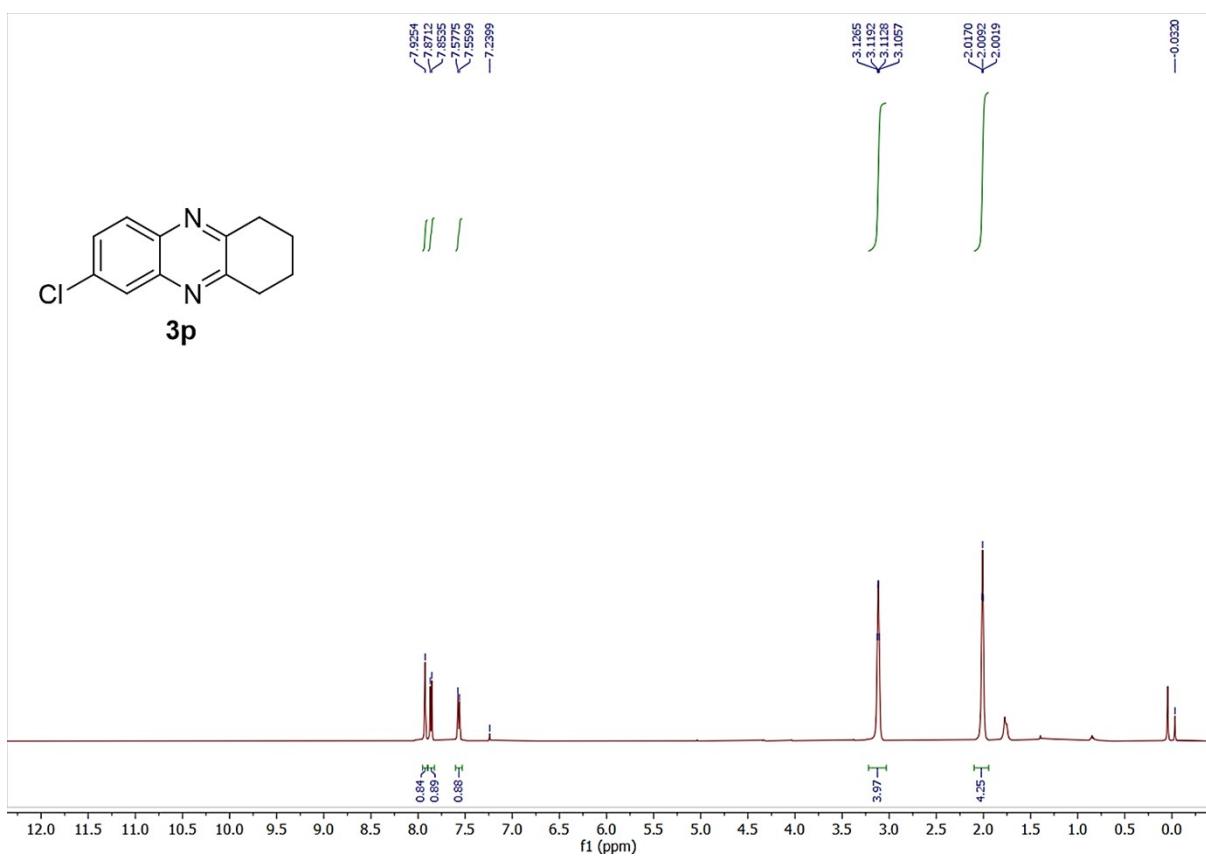


Figure S17:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3o**.



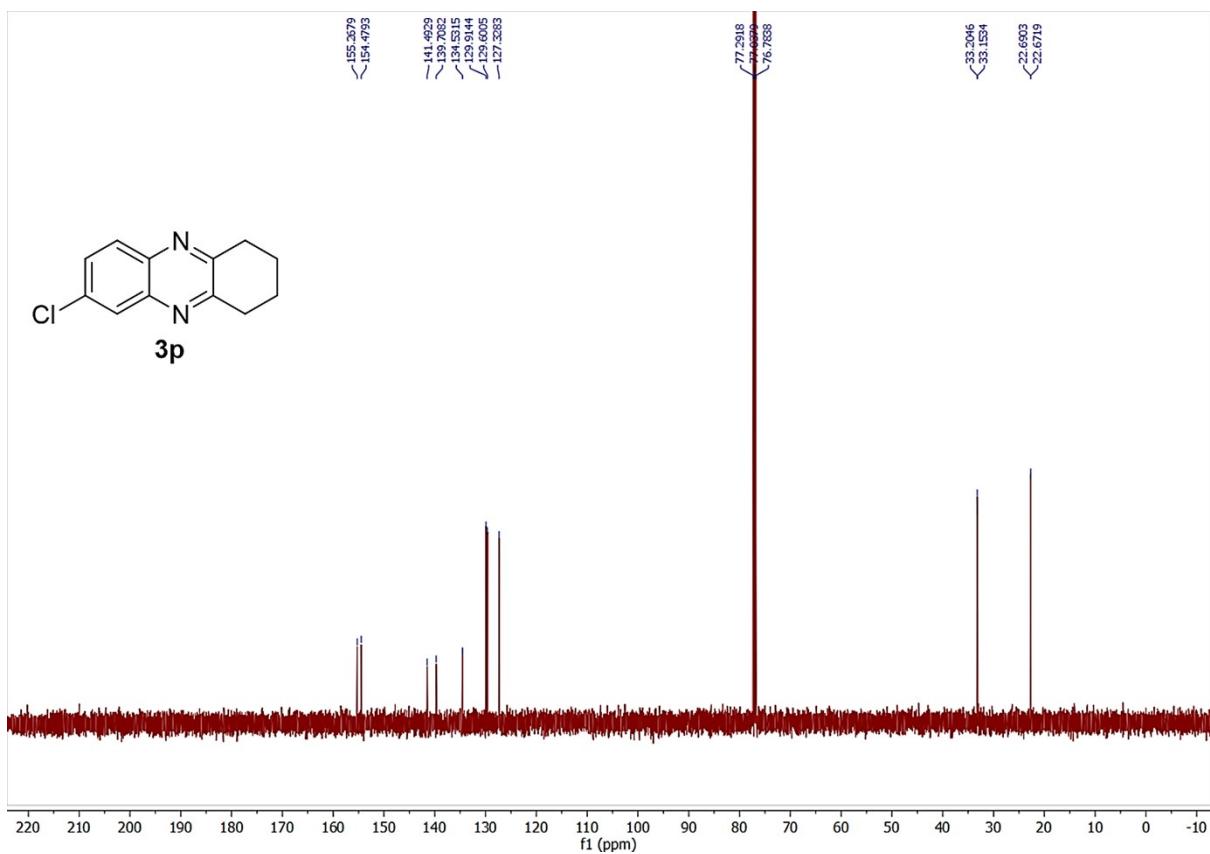
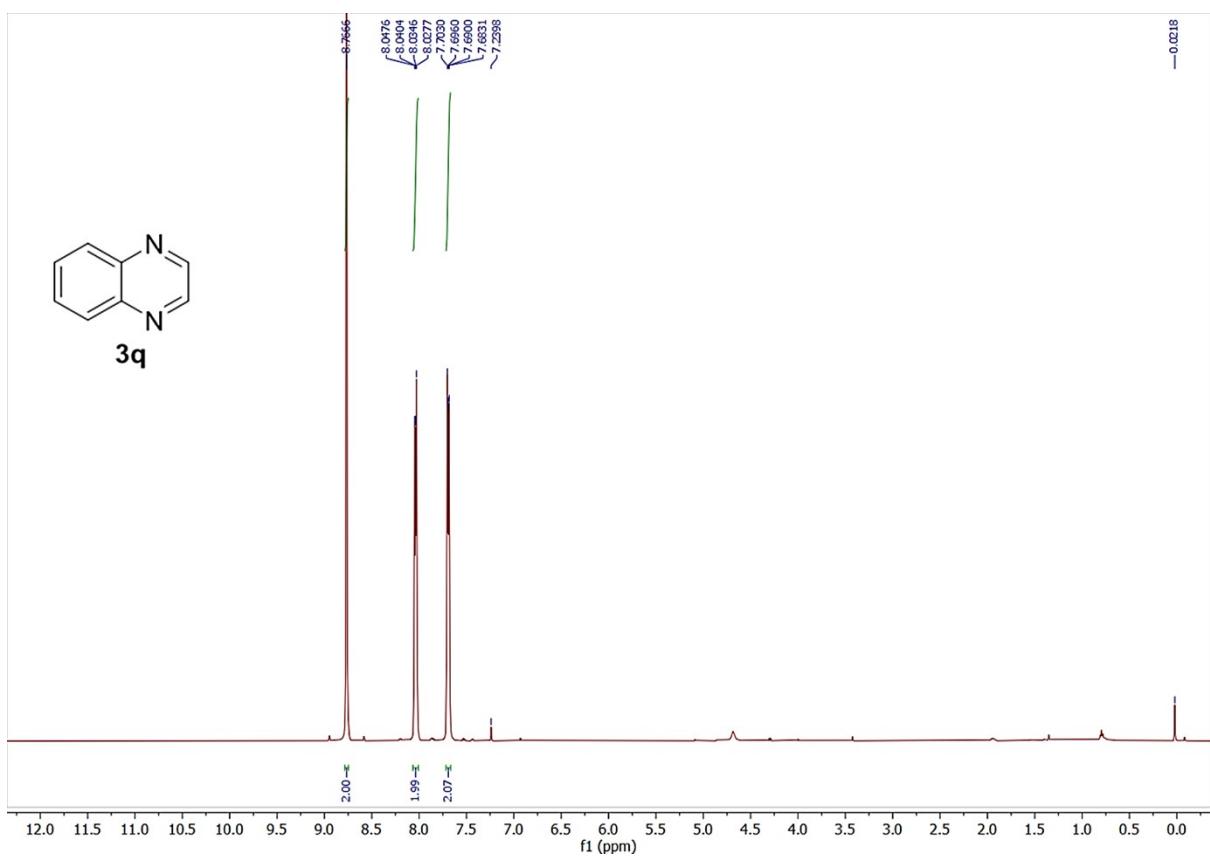


Figure S18:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3p**.



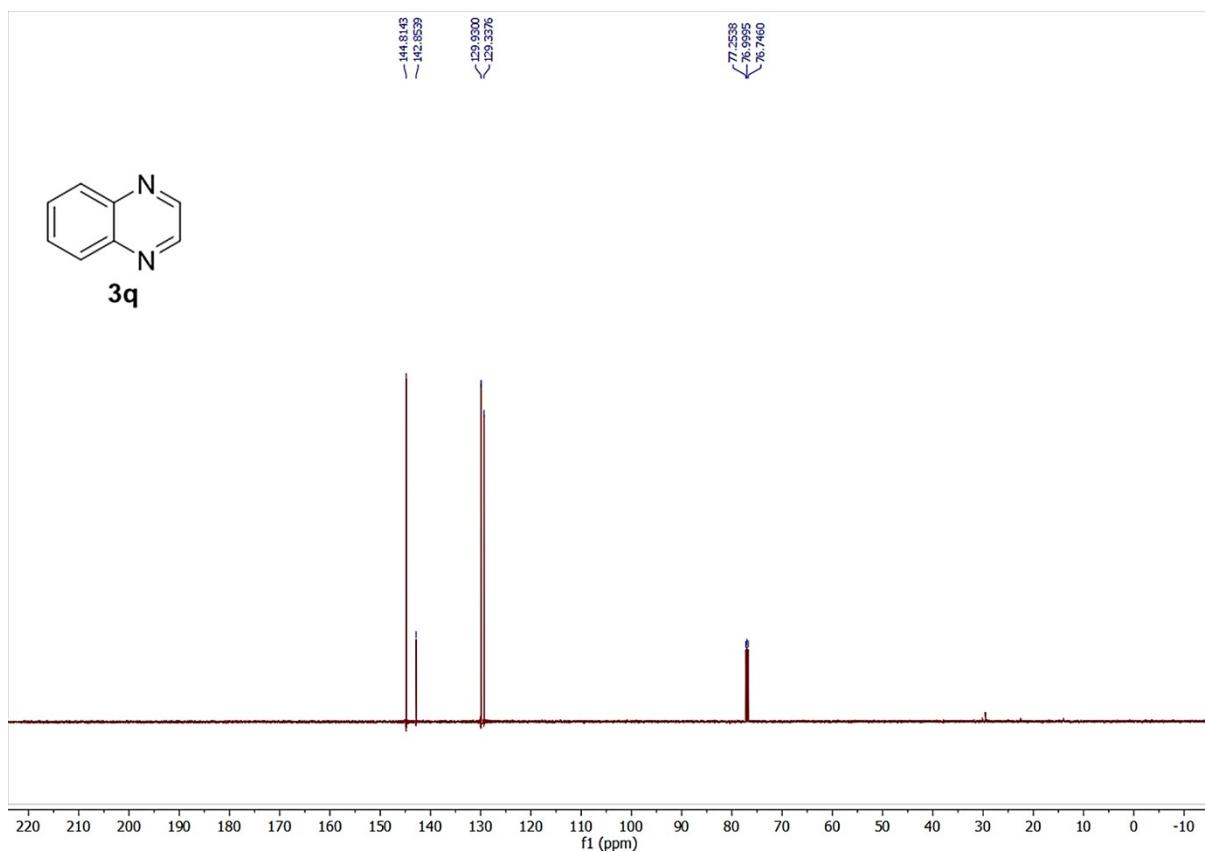
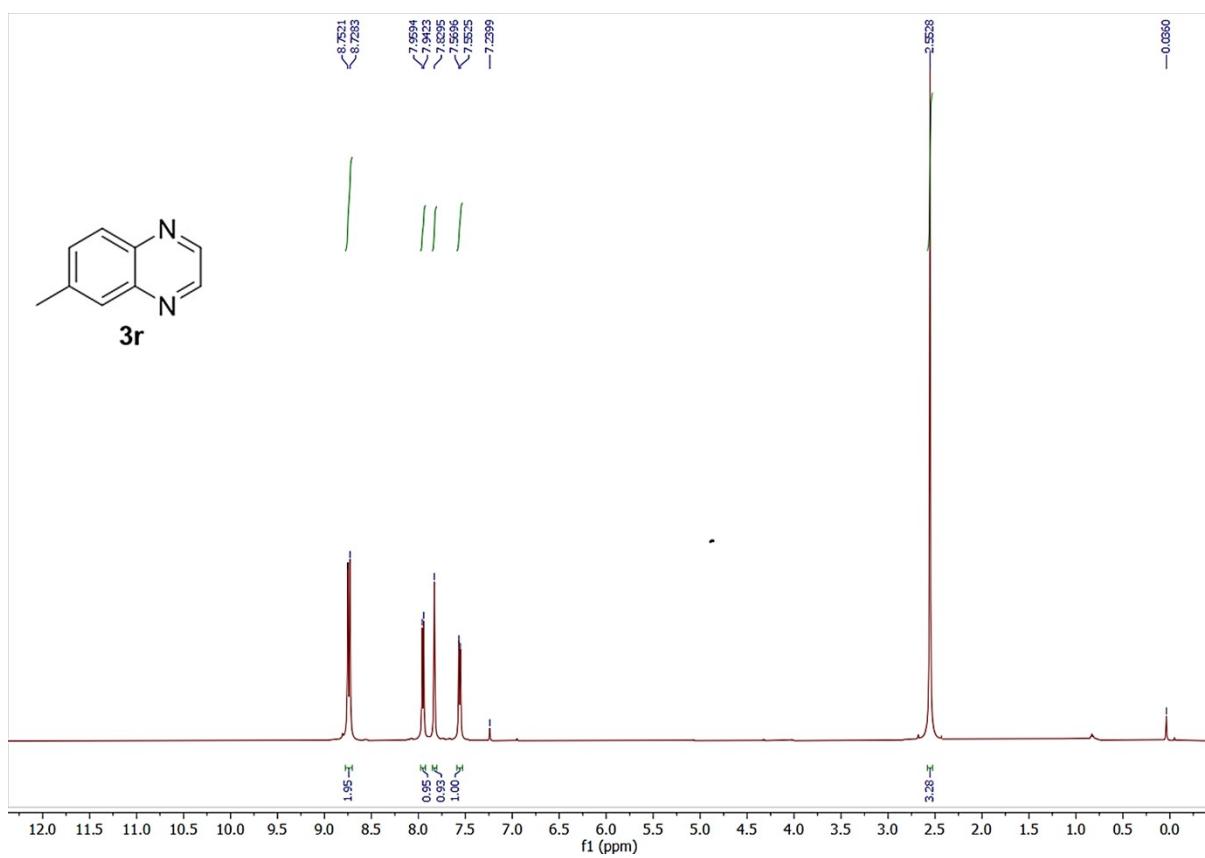


Figure S19: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound **3q**.



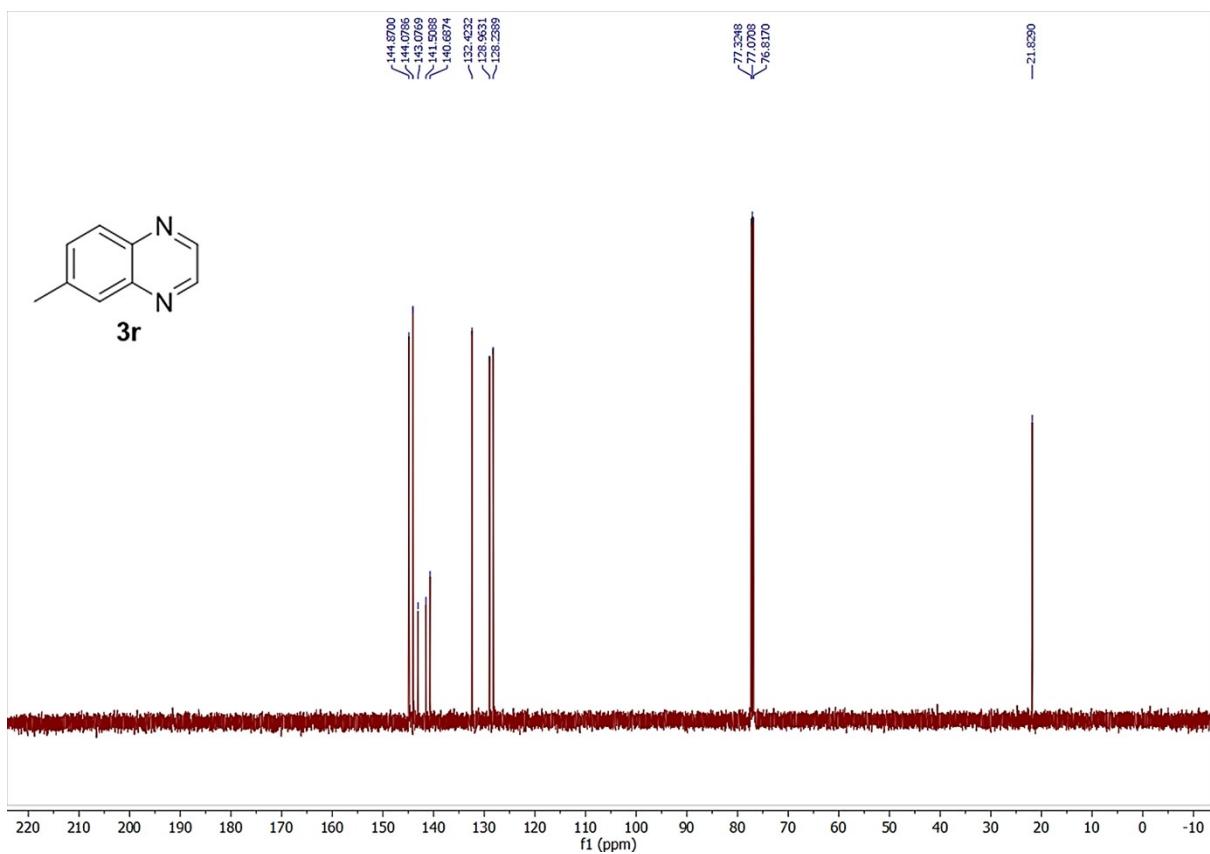
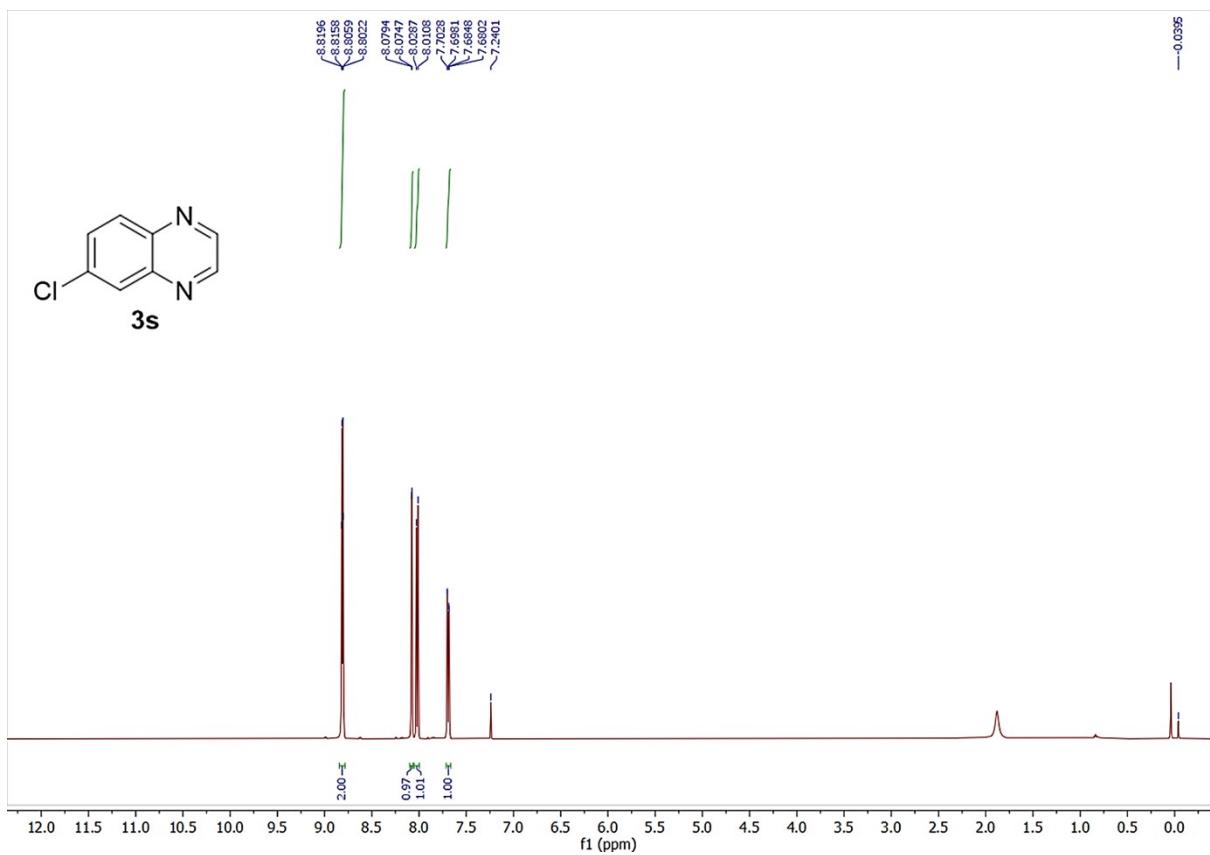


Figure S20: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound **3r**.



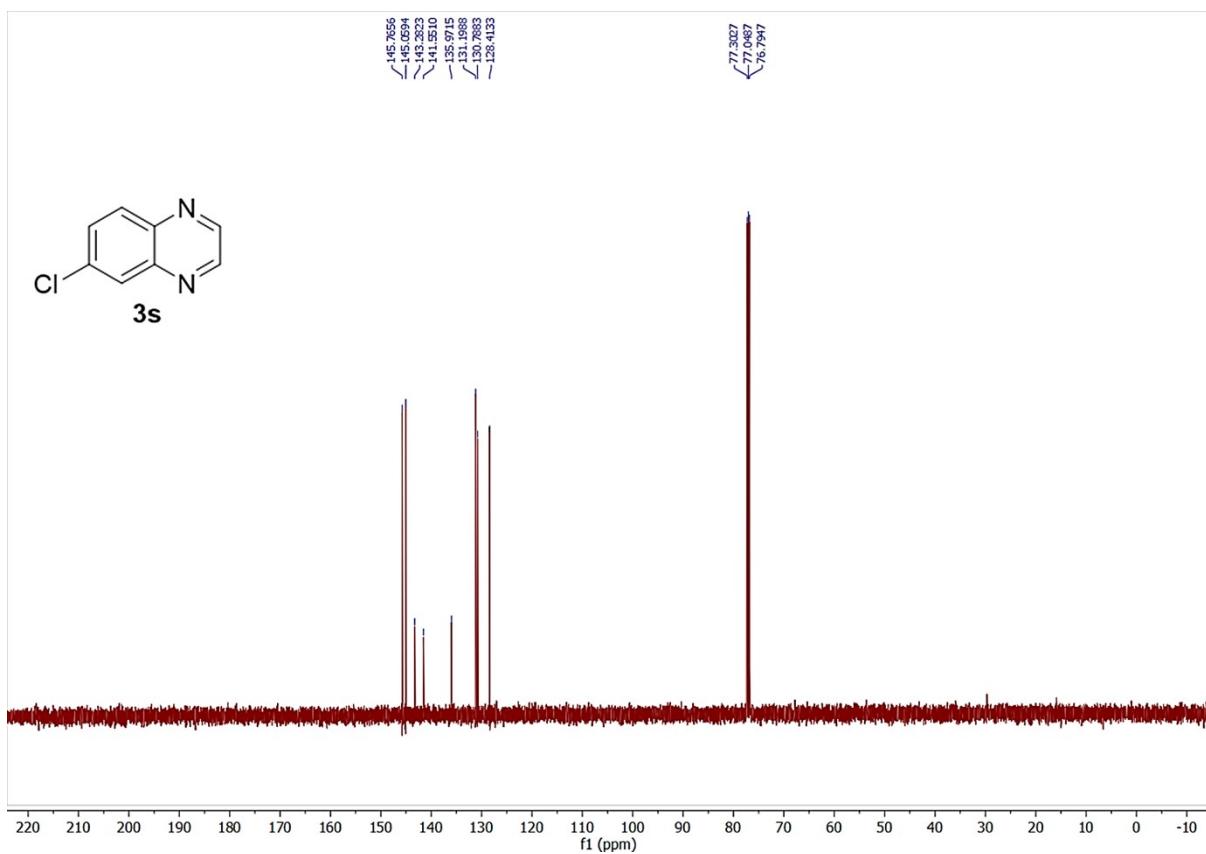
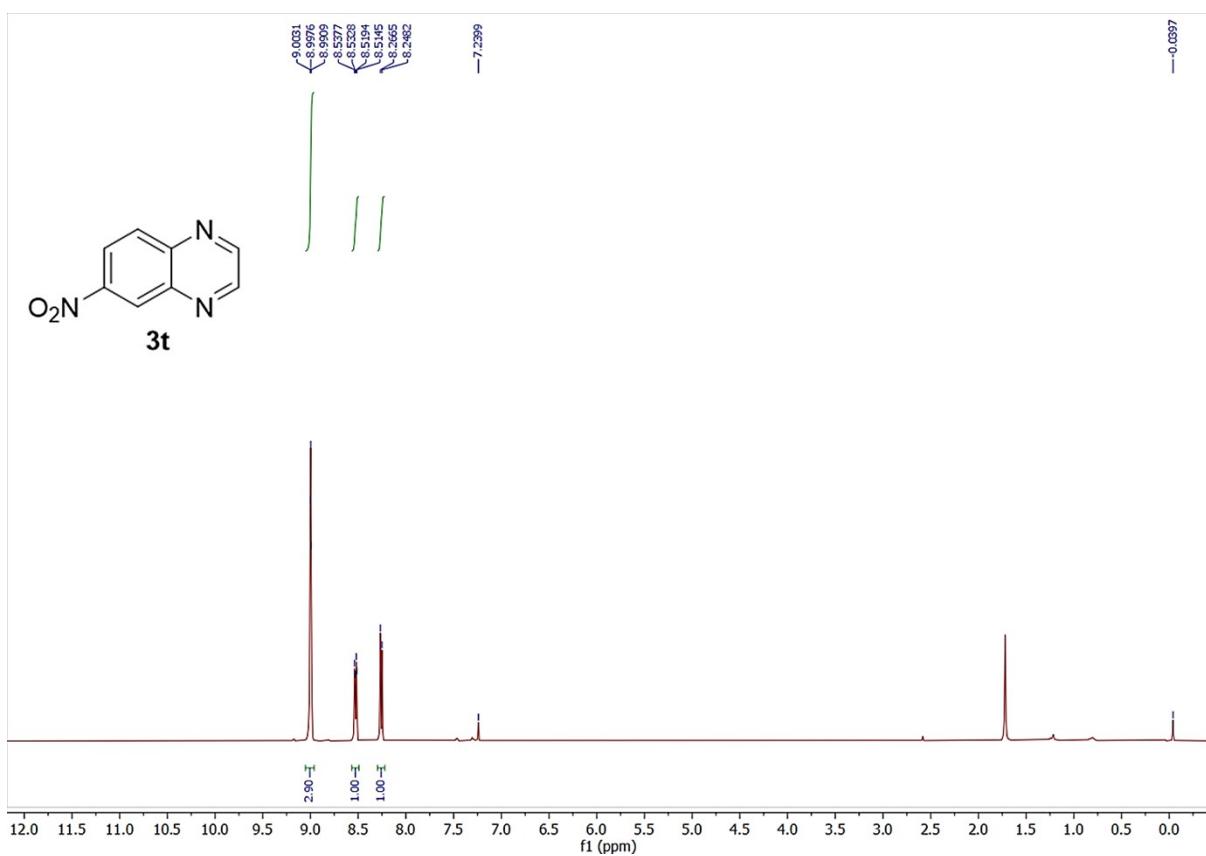


Figure S21: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound **3s**.



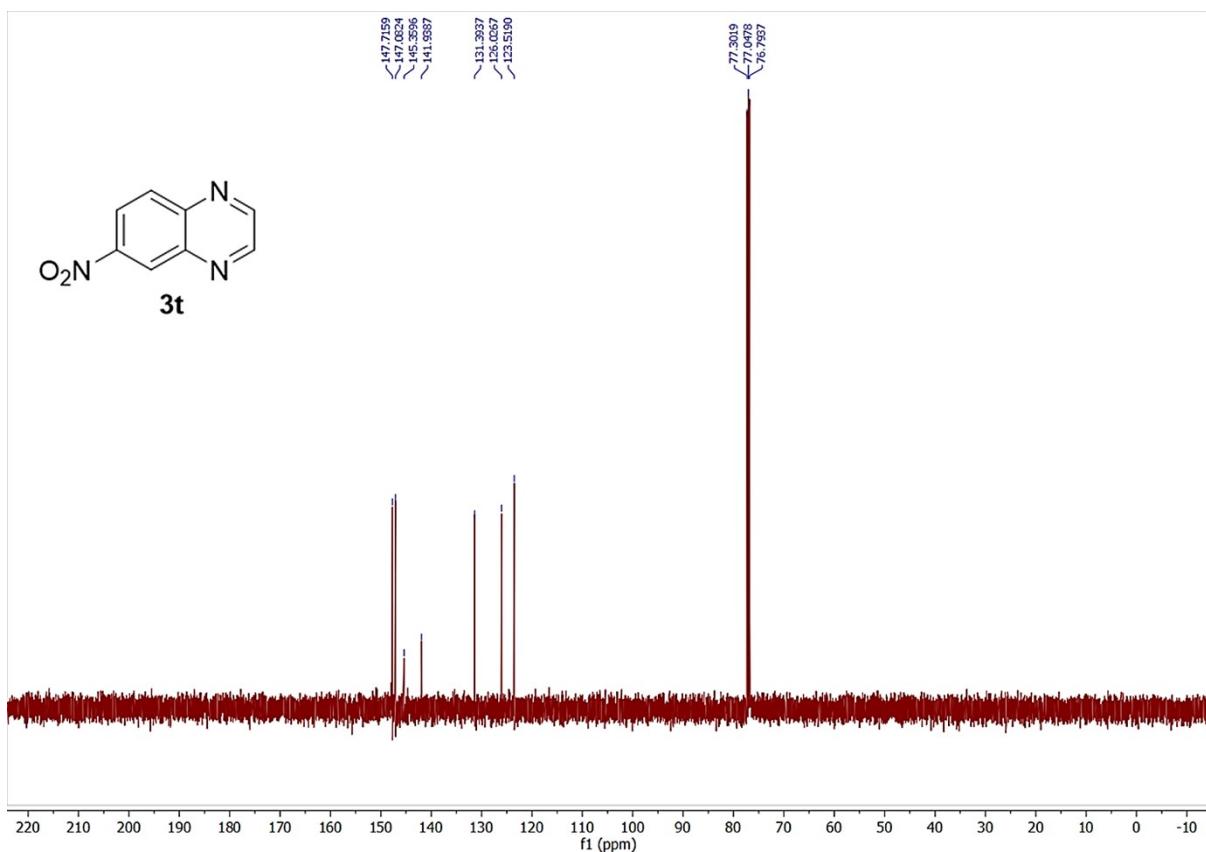
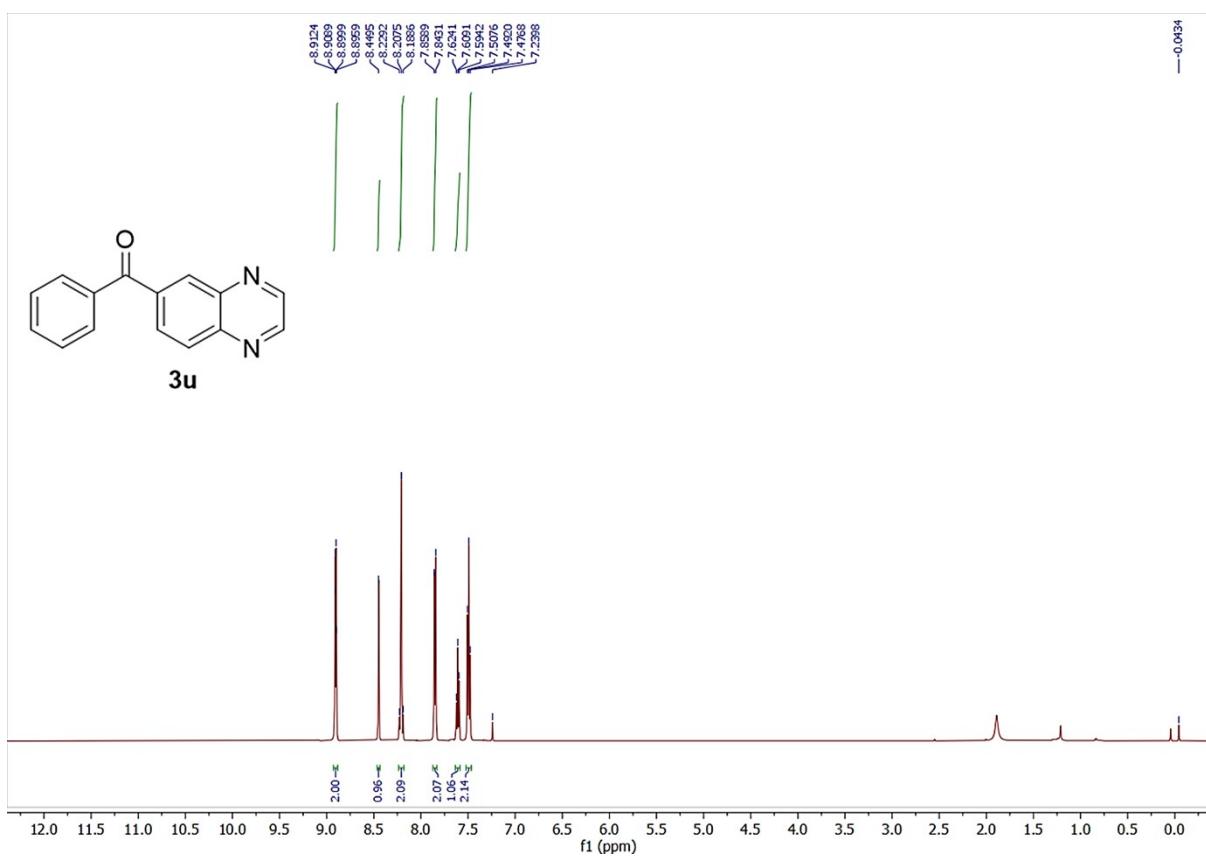


Figure S22: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound **3t**.



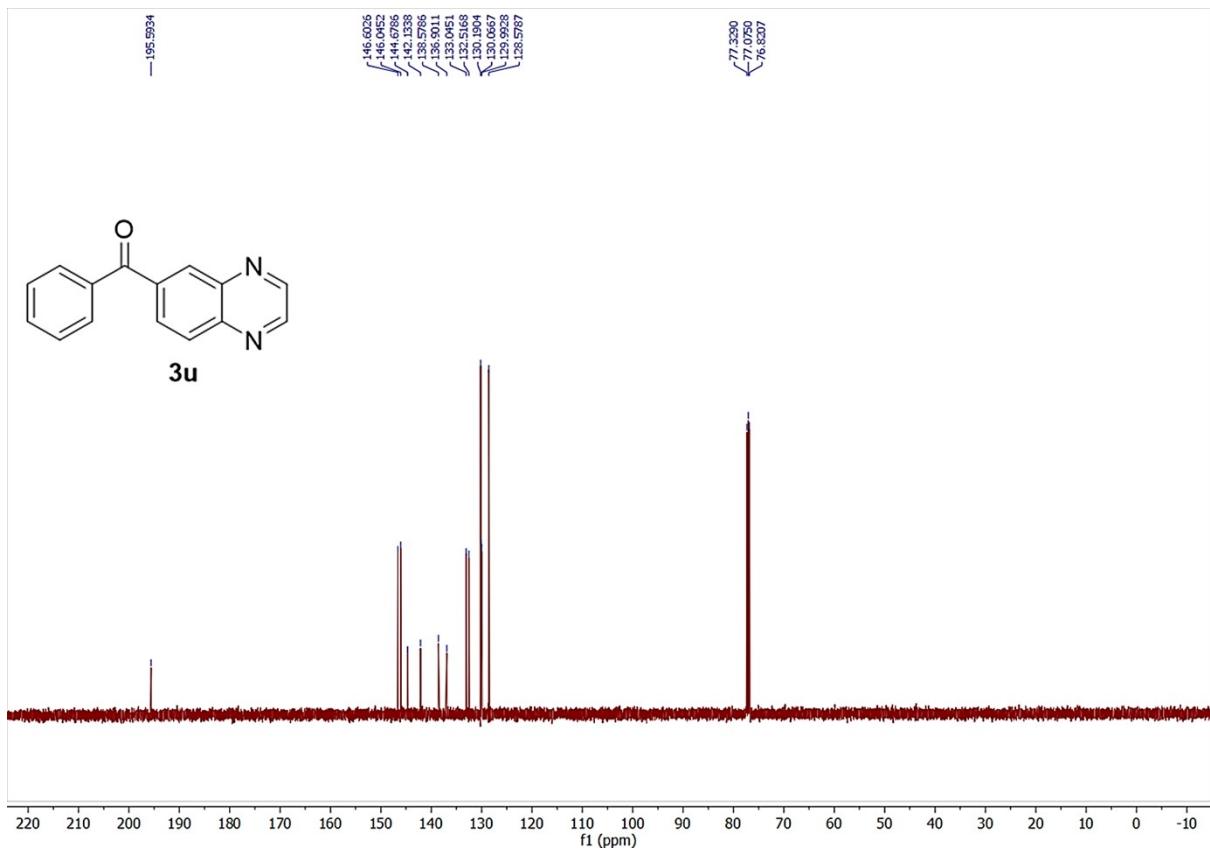
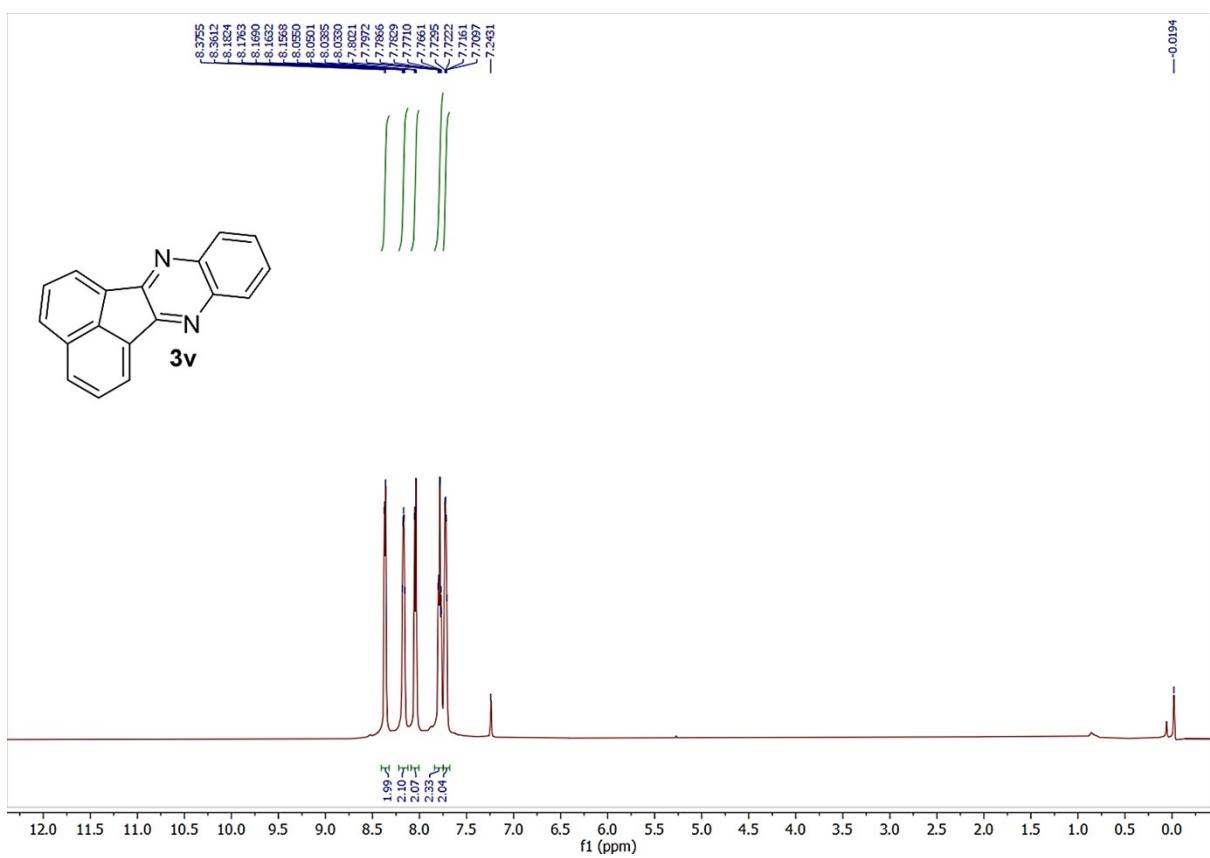


Figure S23:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3u**.



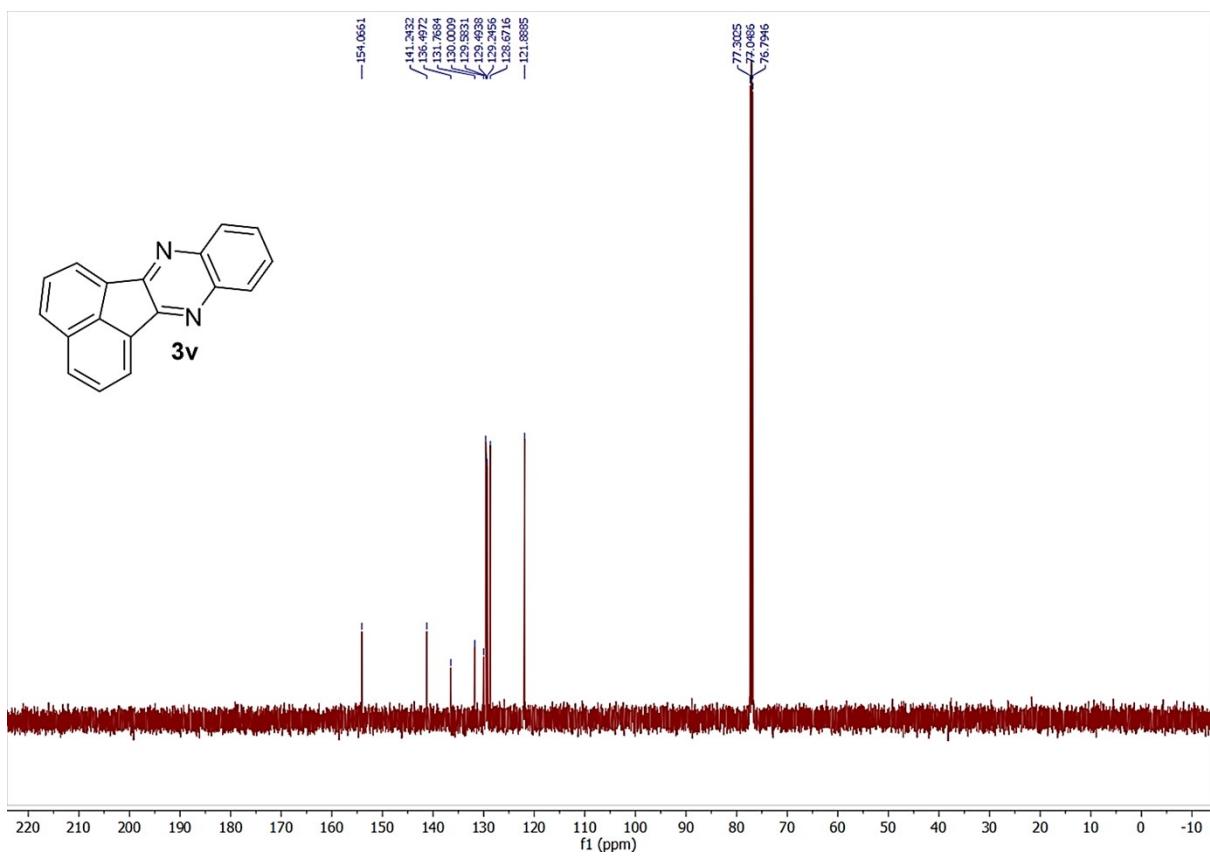
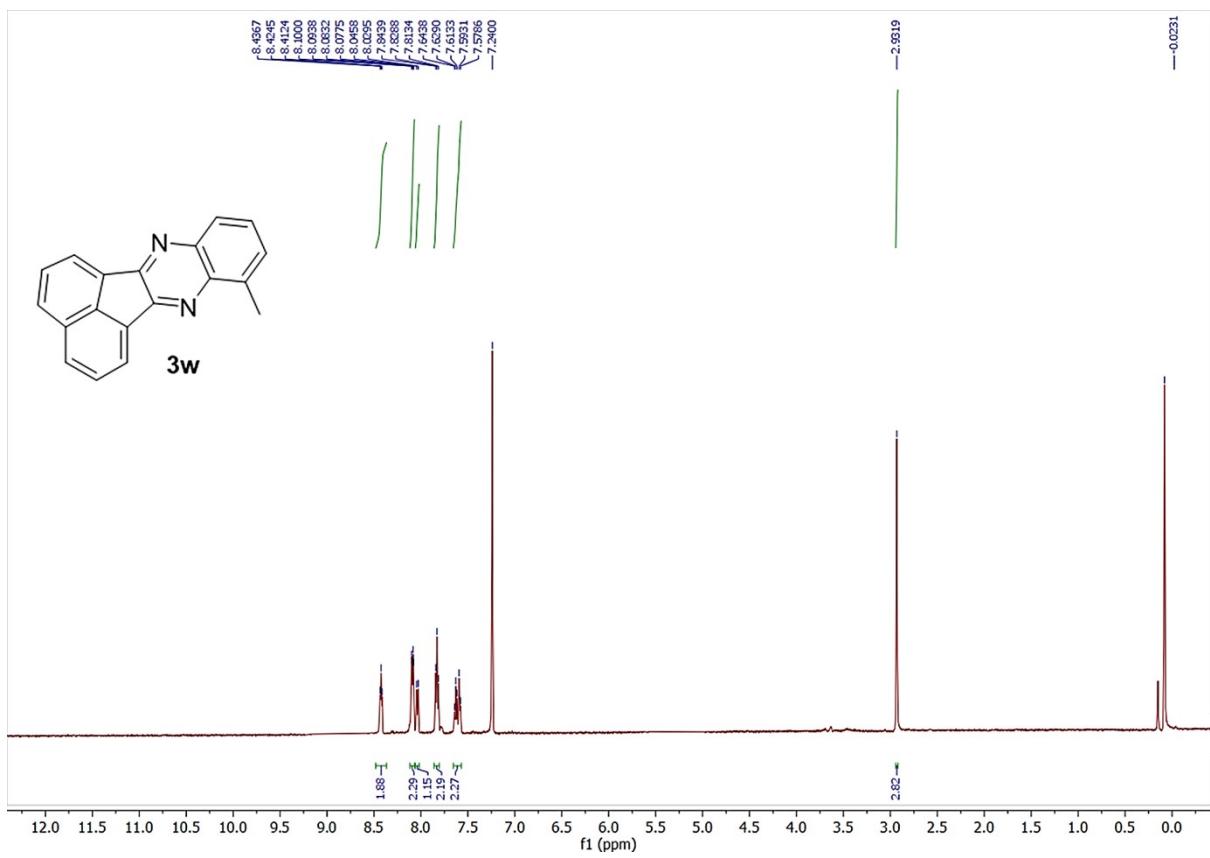


Figure S24:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3v**.



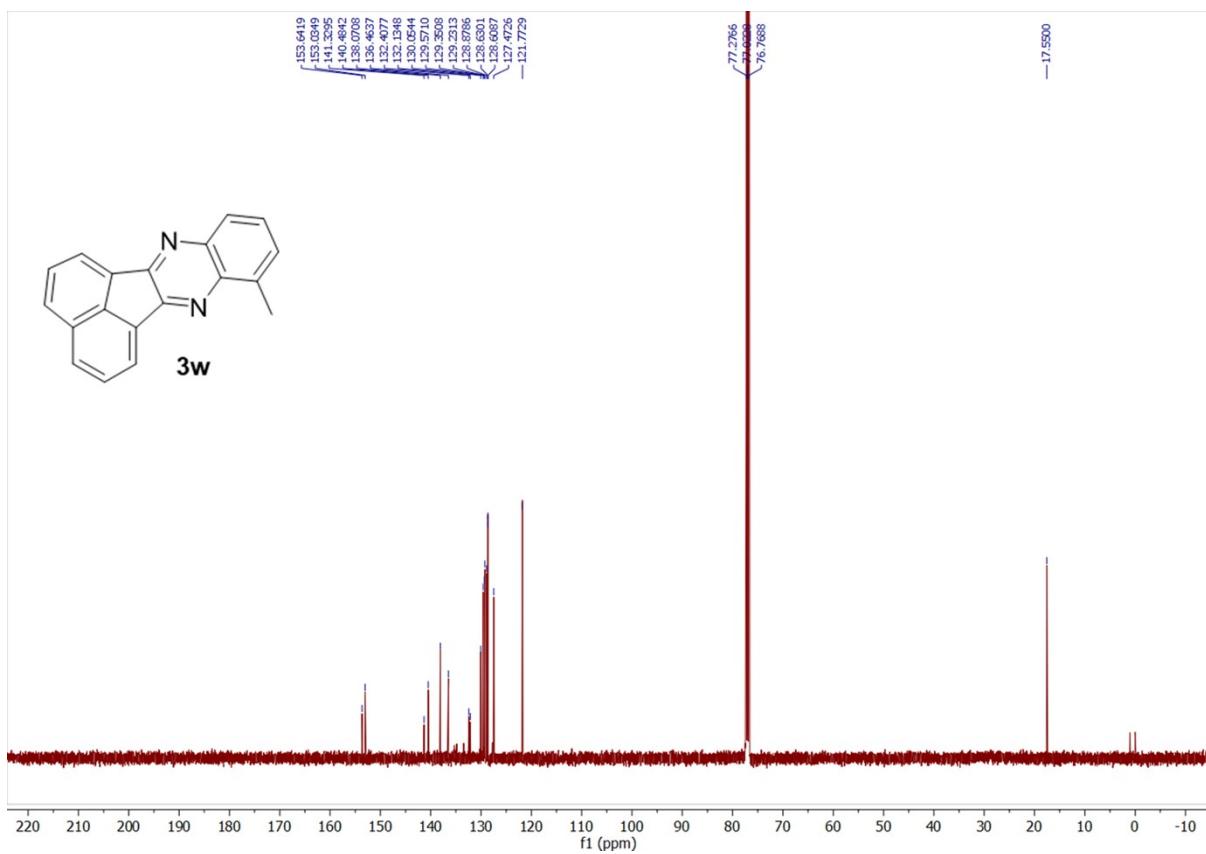
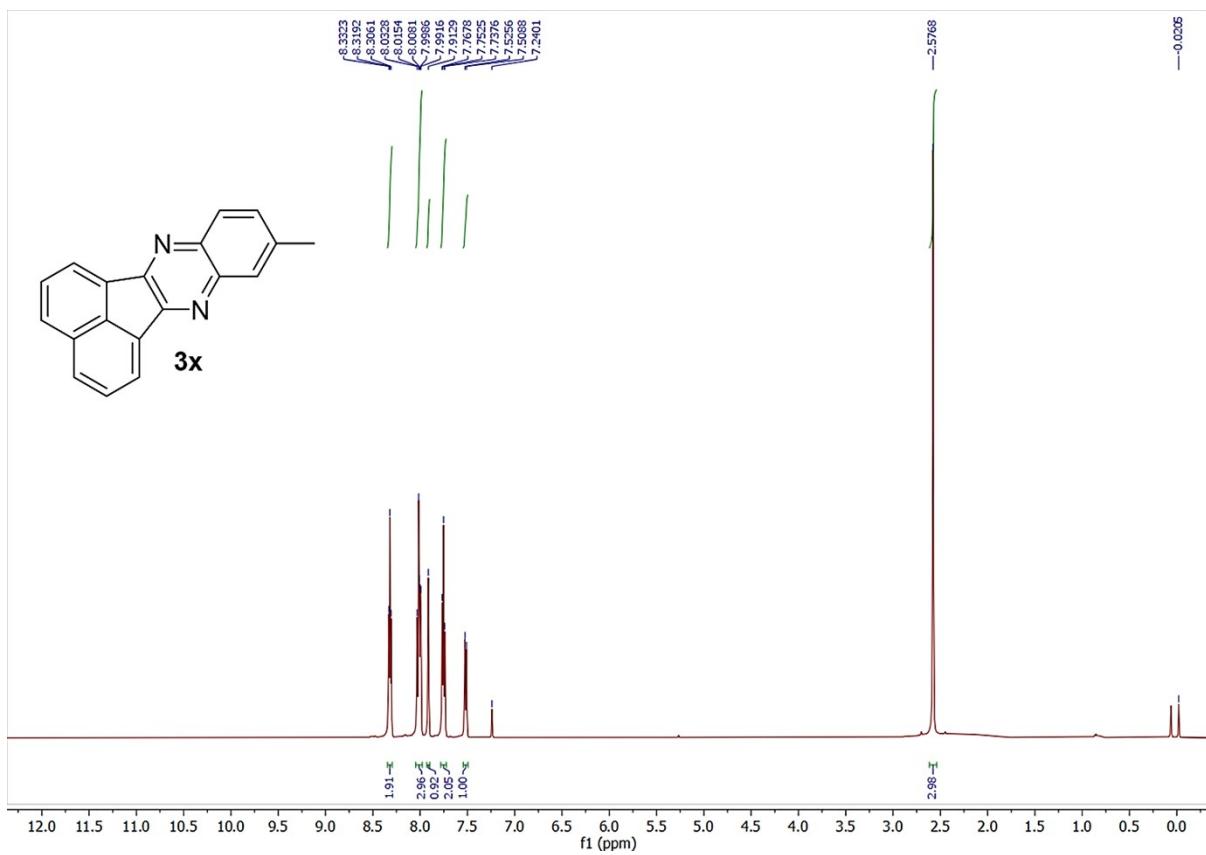


Figure S25:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3w**.



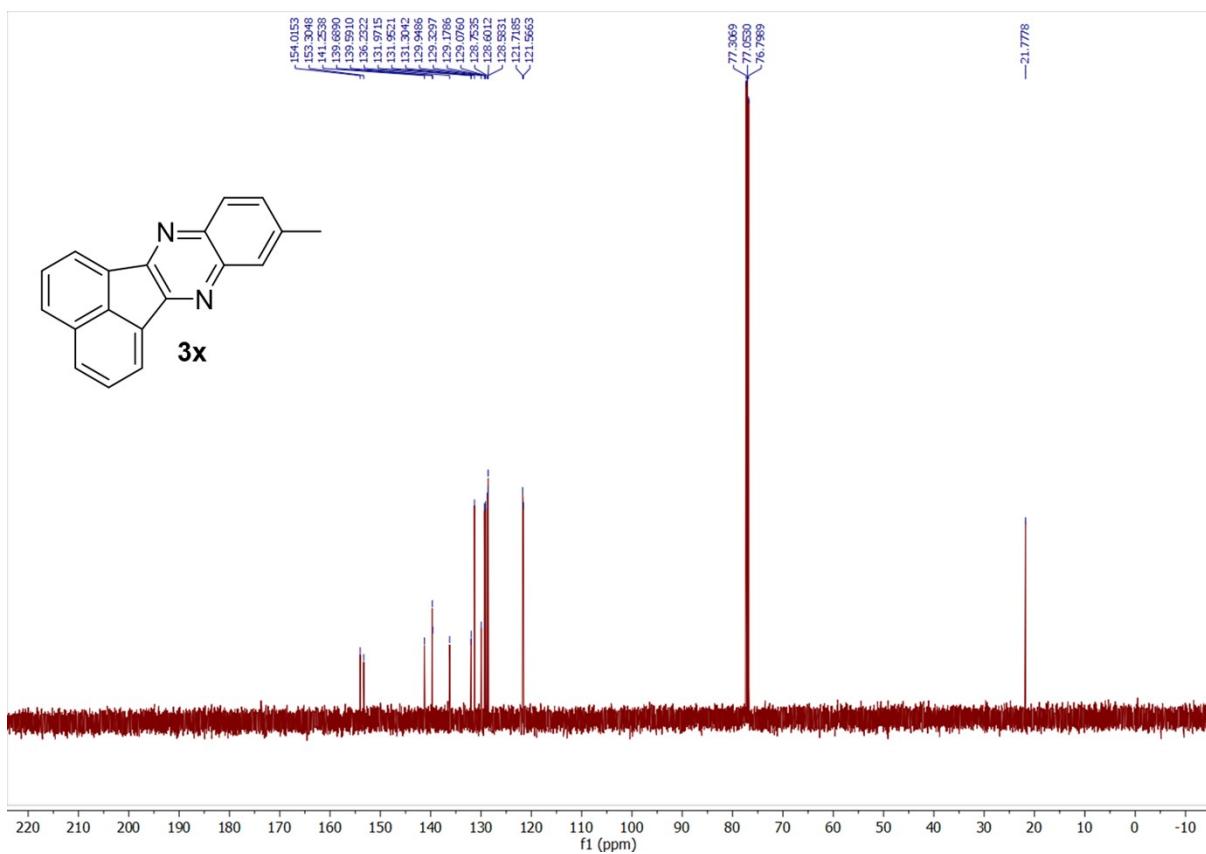
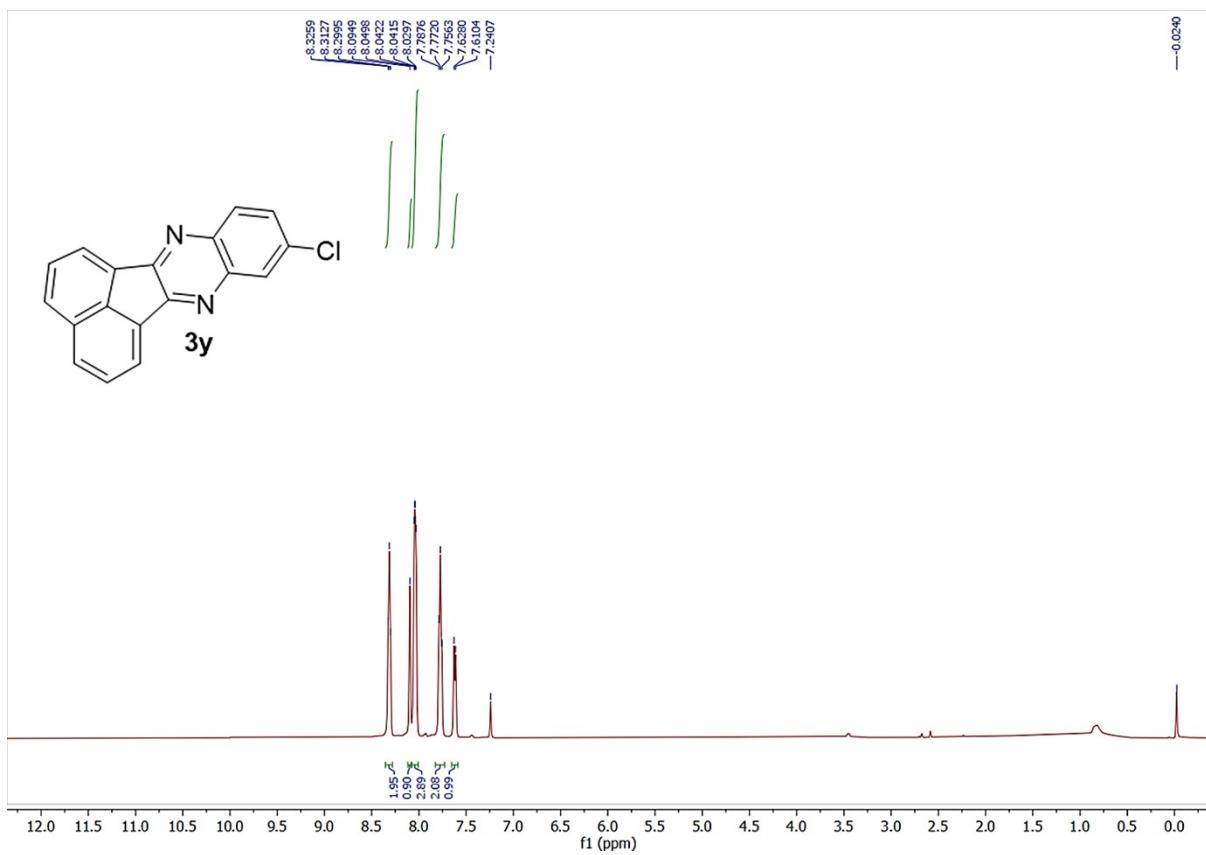


Figure S26:  $^1\text{H}$  (500 MHz, CDCl<sub>3</sub>) and  $^{13}\text{C}$  (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound **3x**.



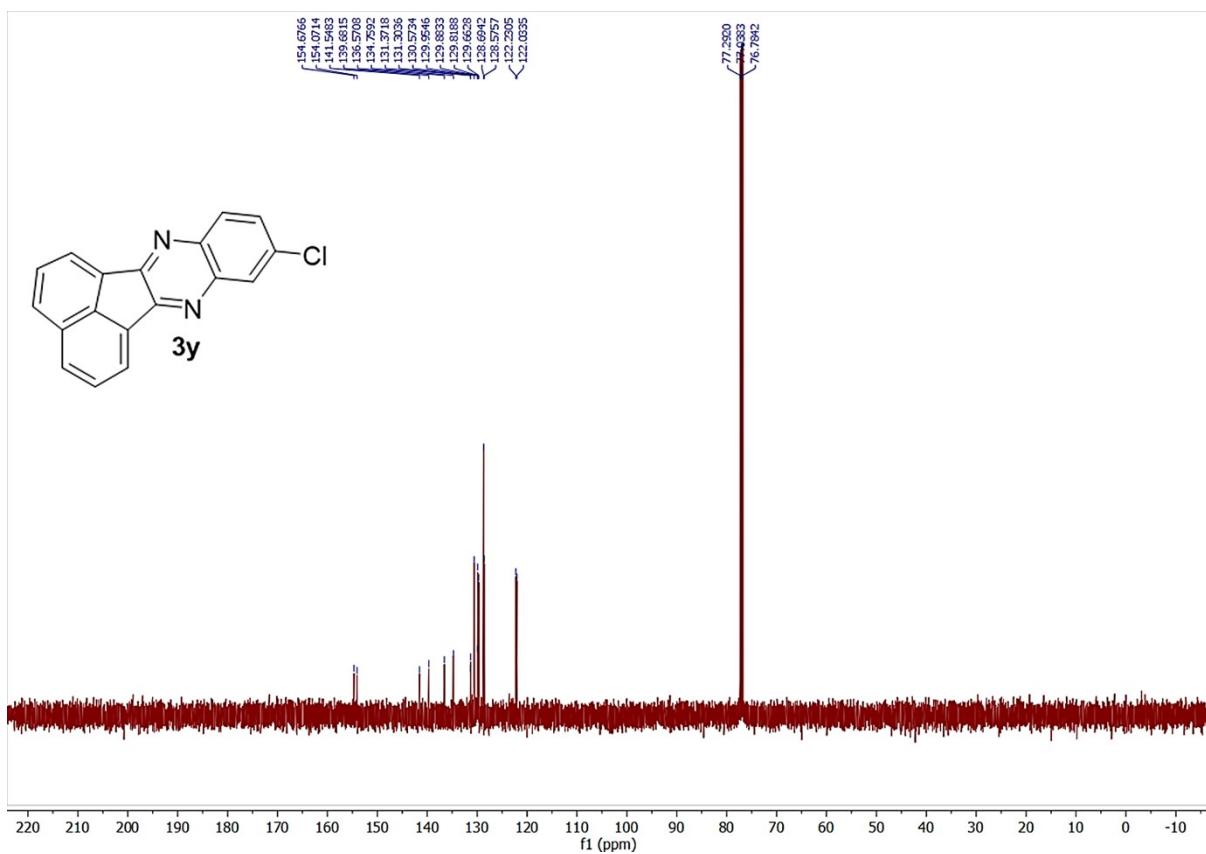
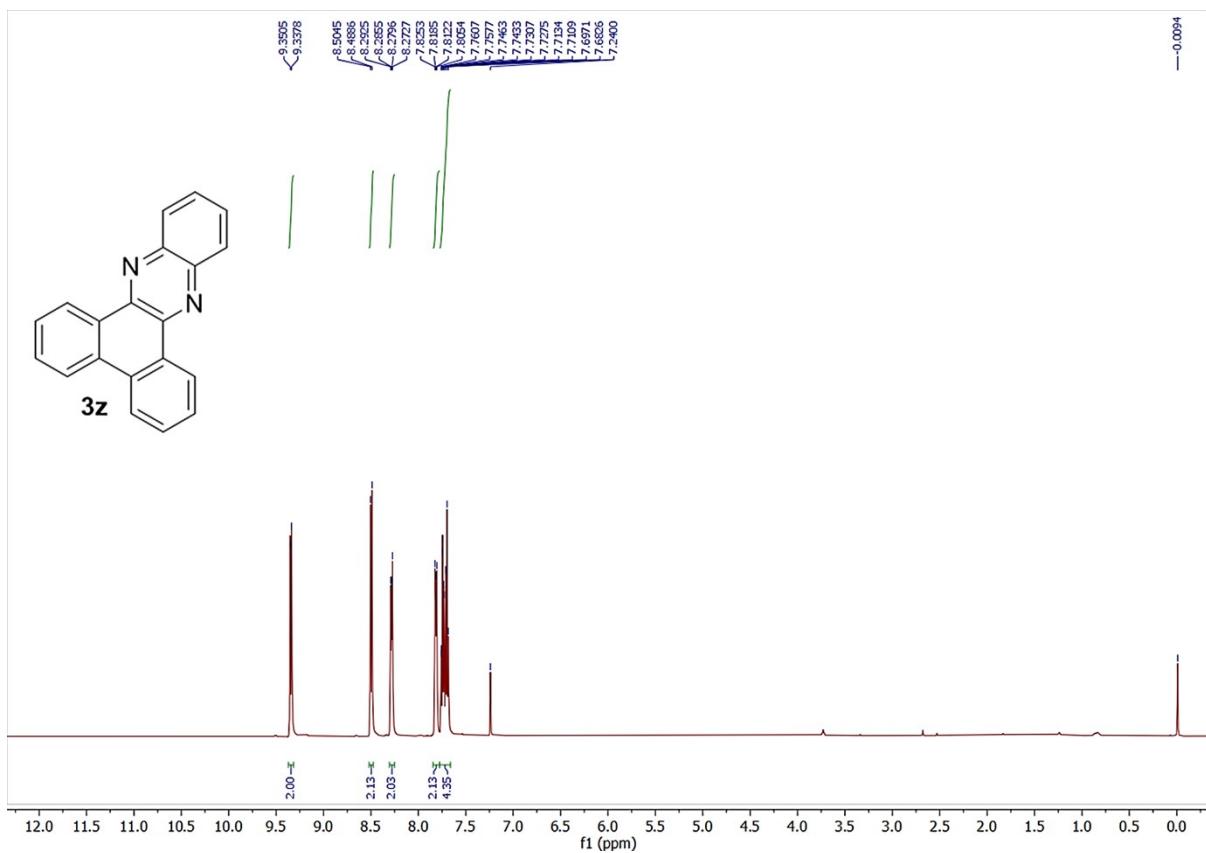


Figure S27:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3y**.



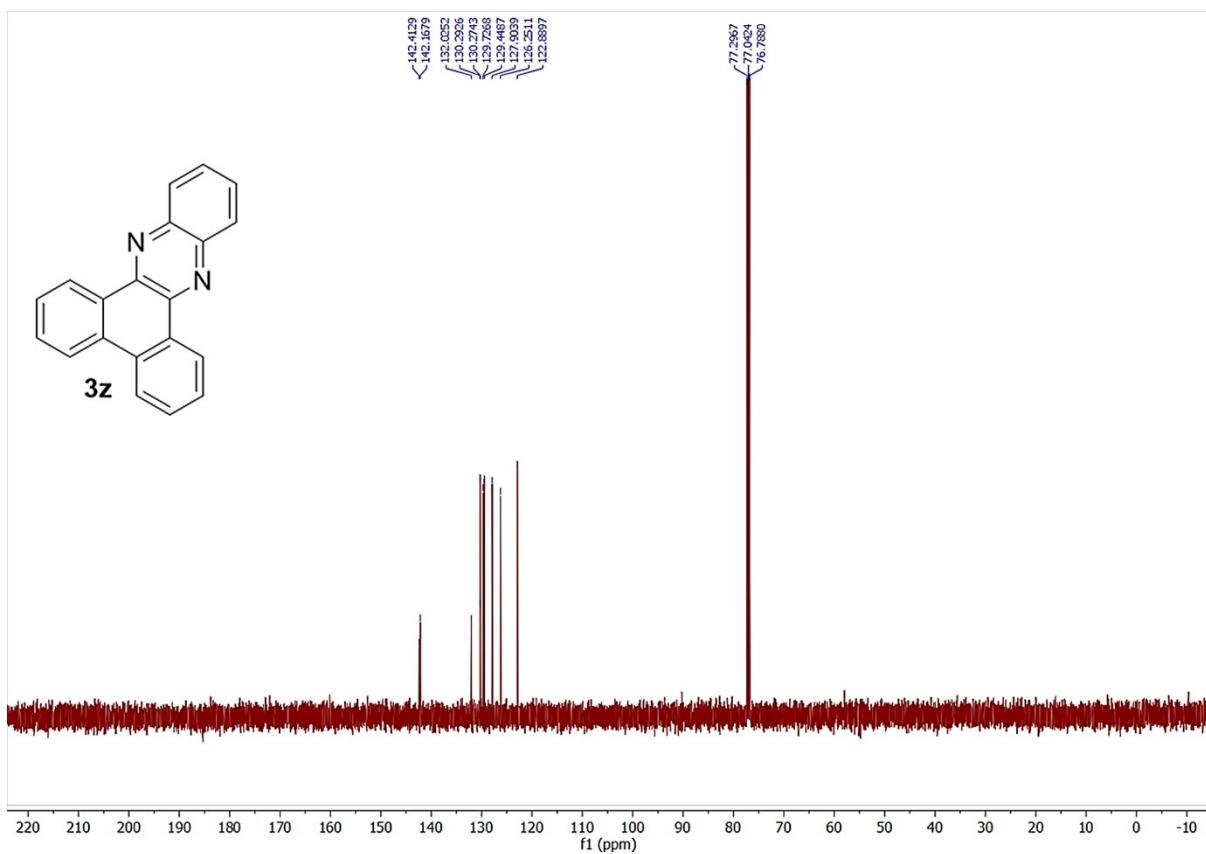
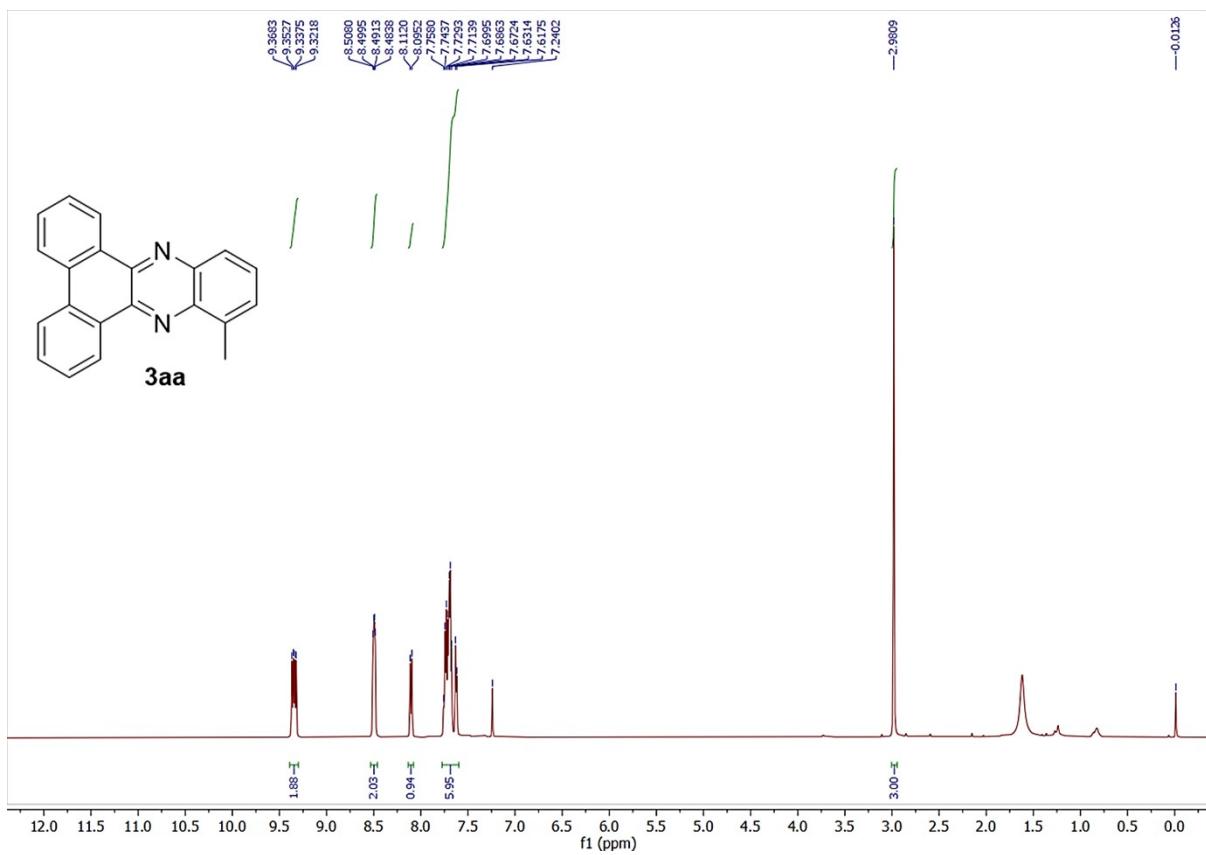


Figure S28:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3z**.



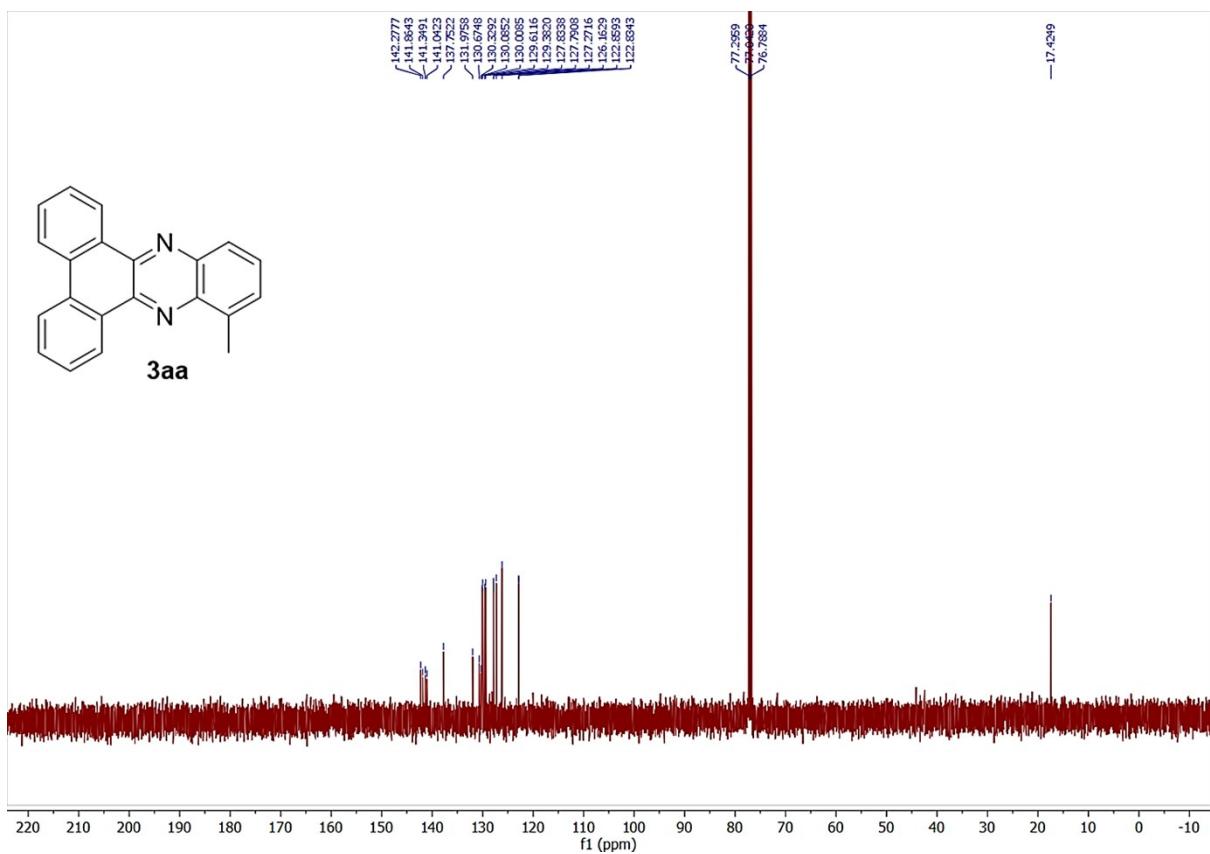
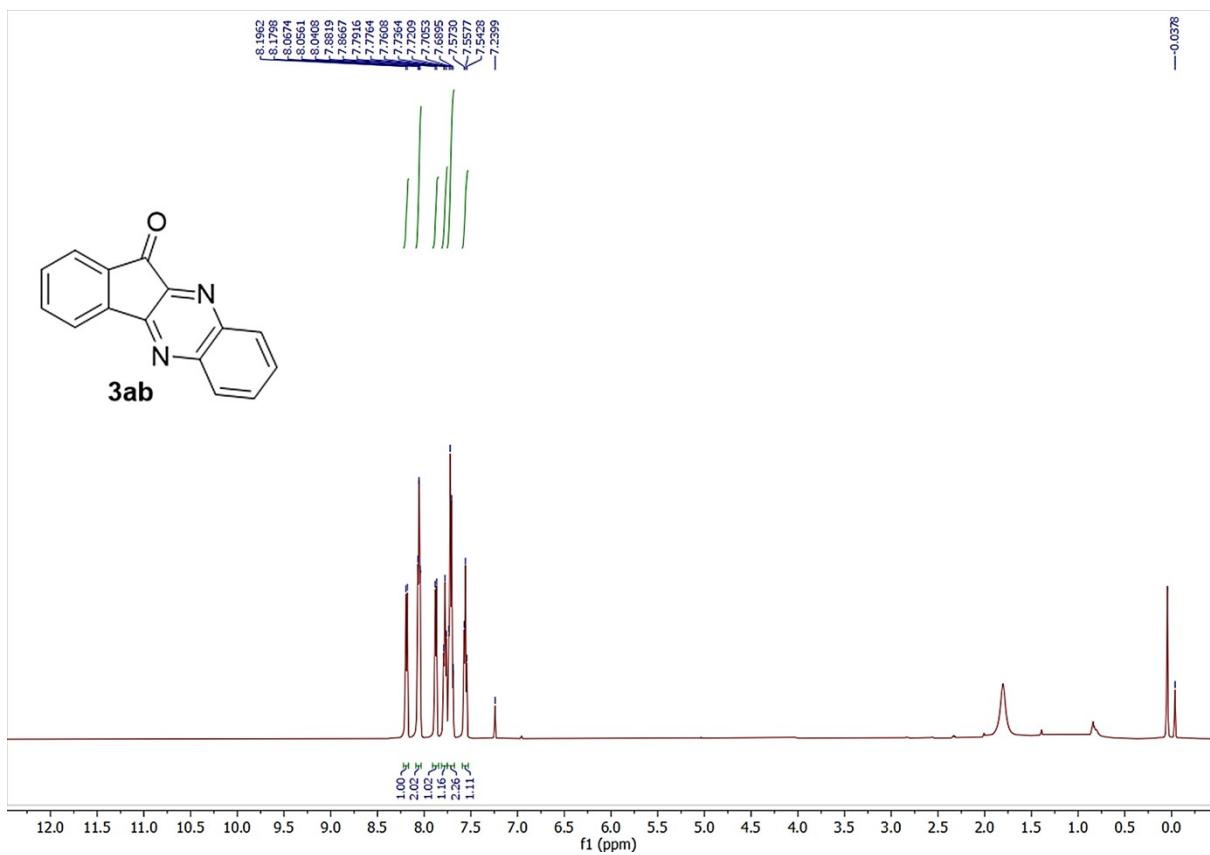


Figure S29:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3aa**.



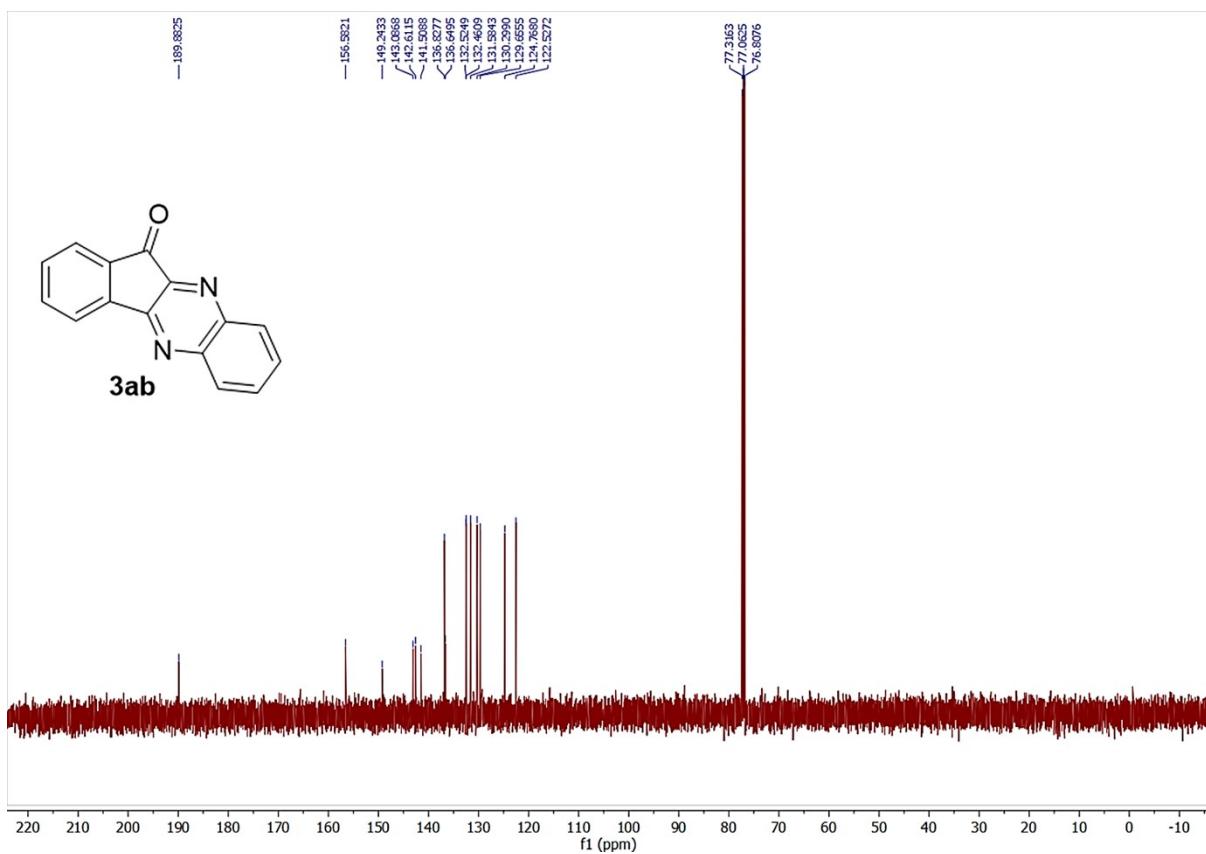
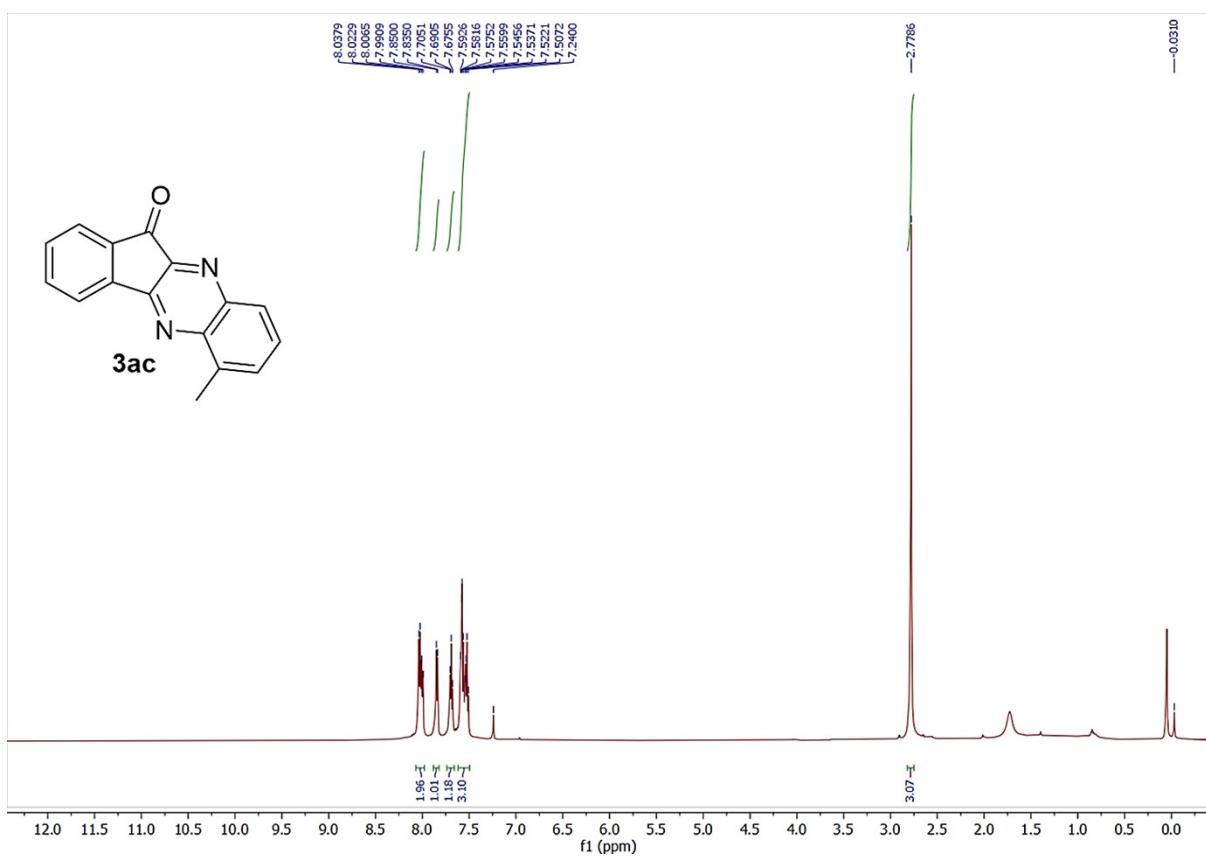


Figure S30:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3ab**.



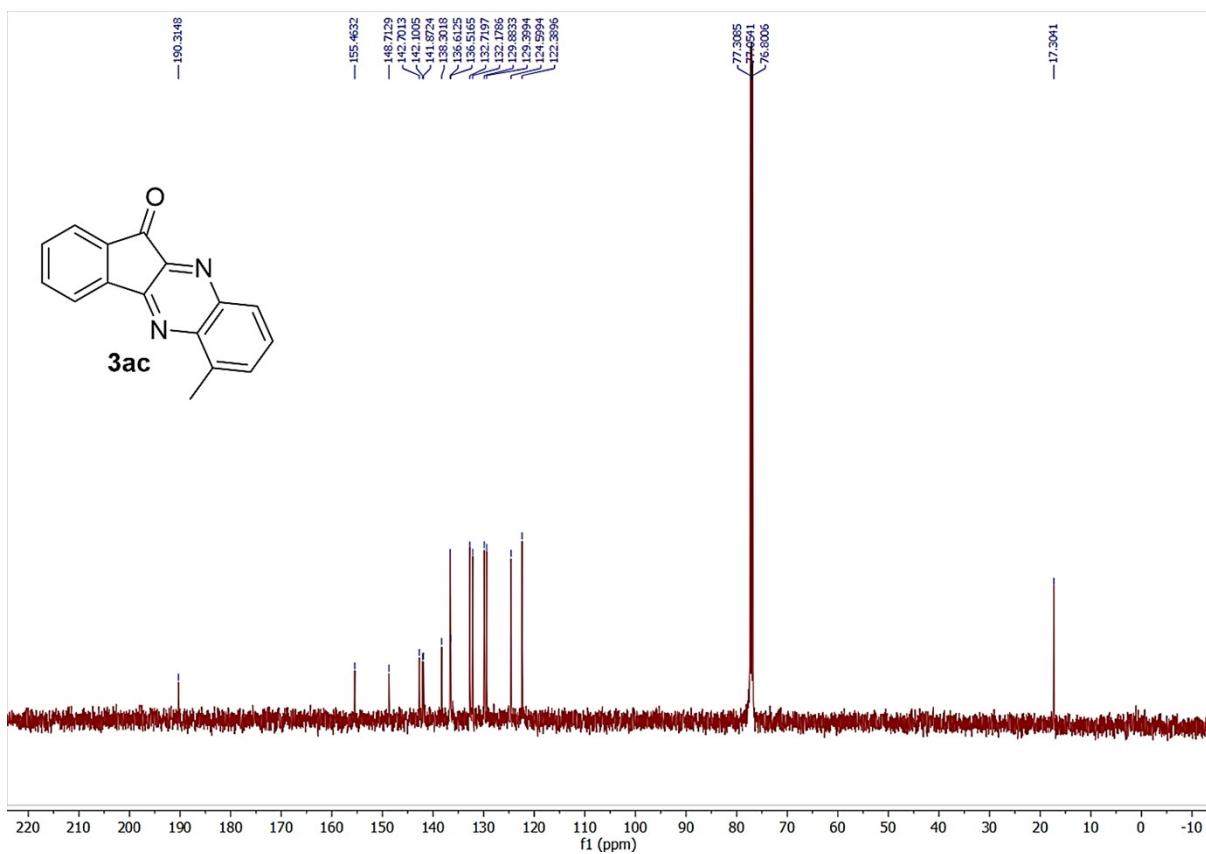
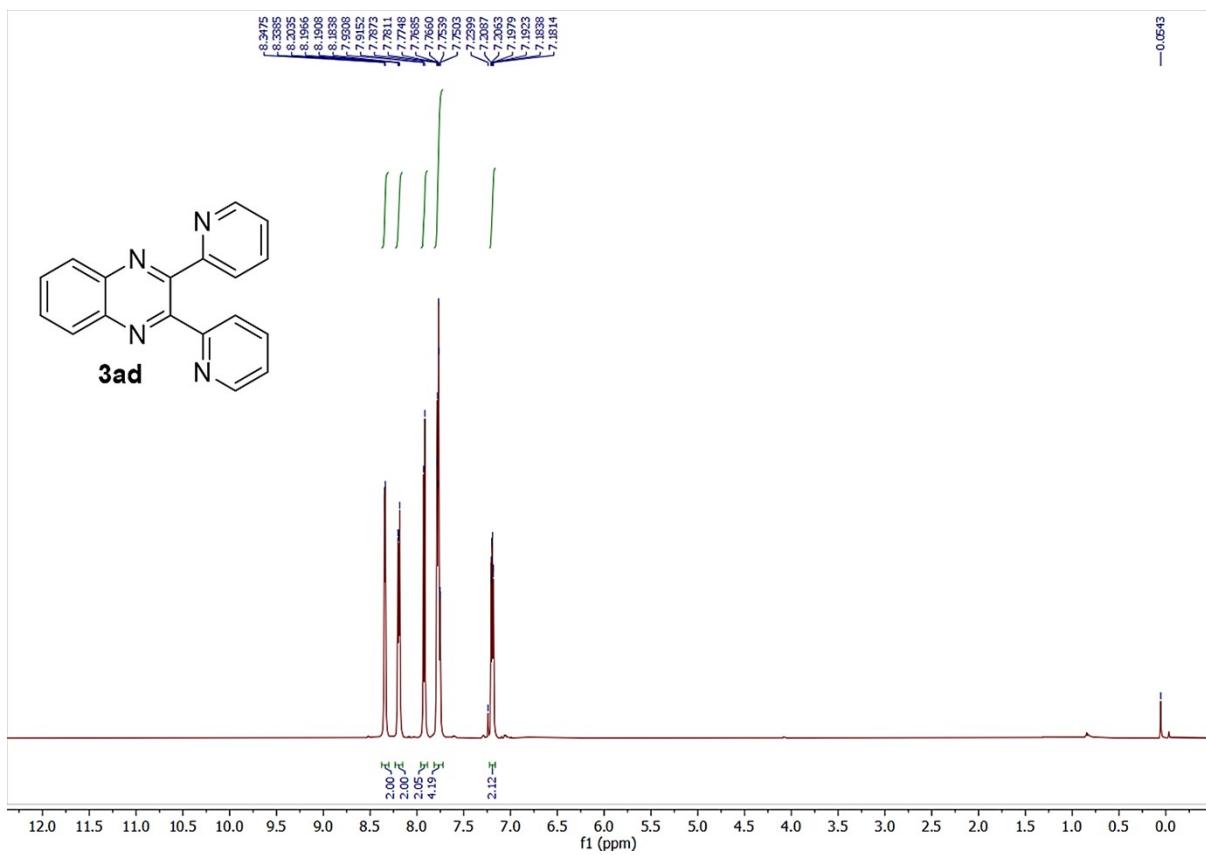


Figure S31:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3ac**.



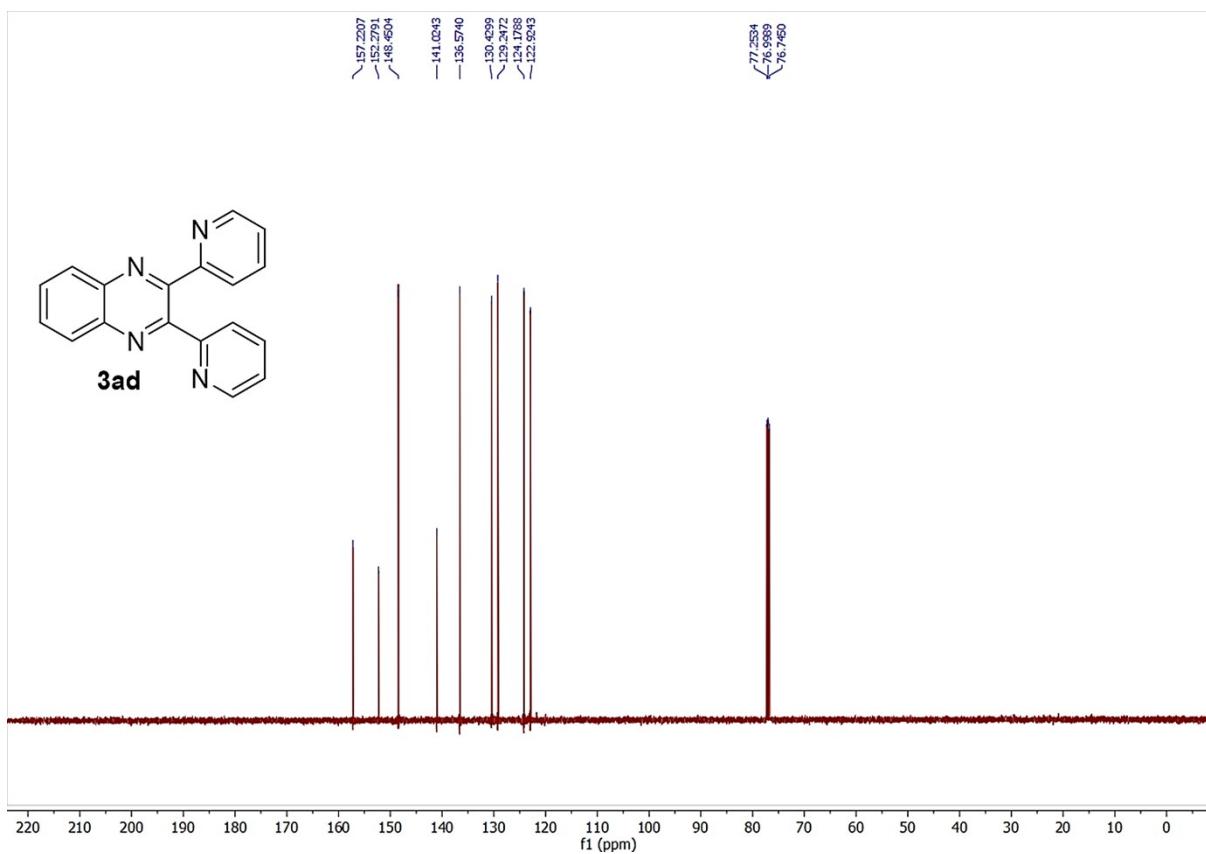
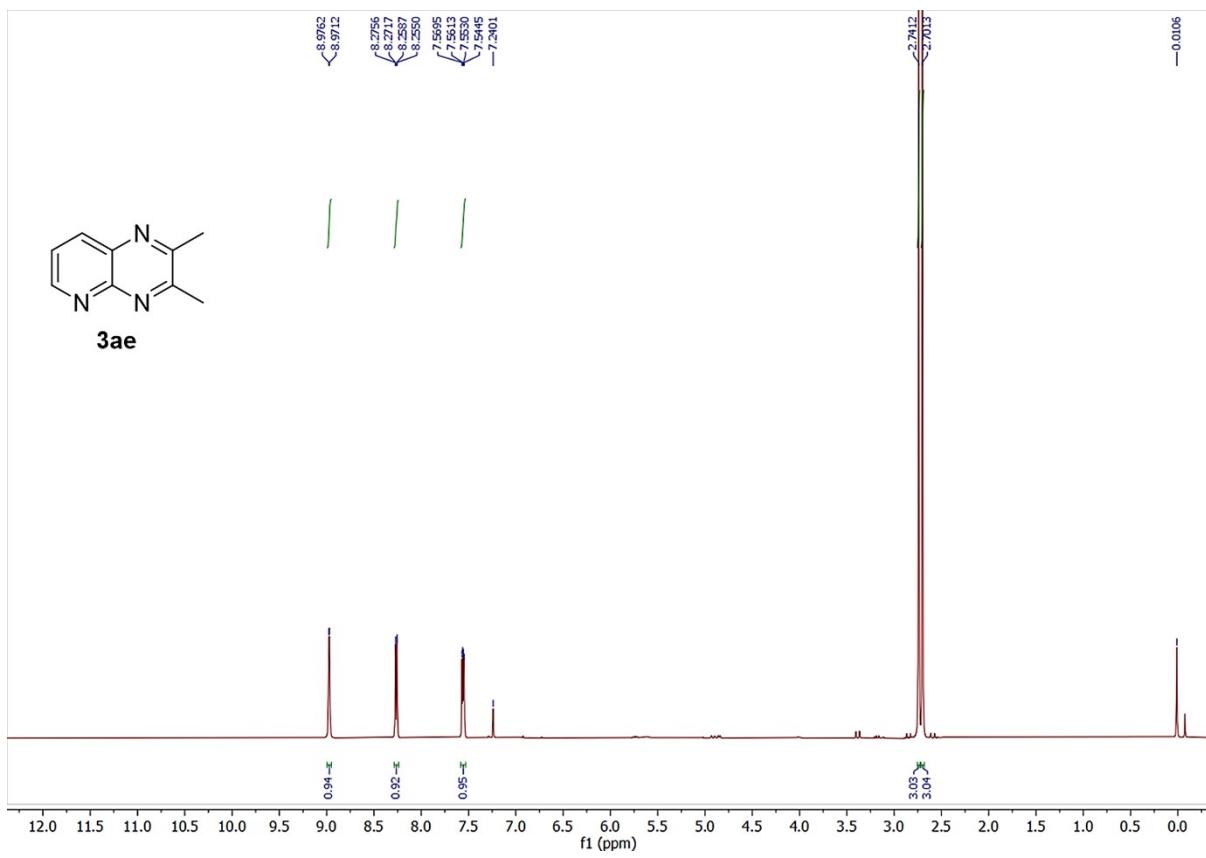


Figure S32:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3ad**.



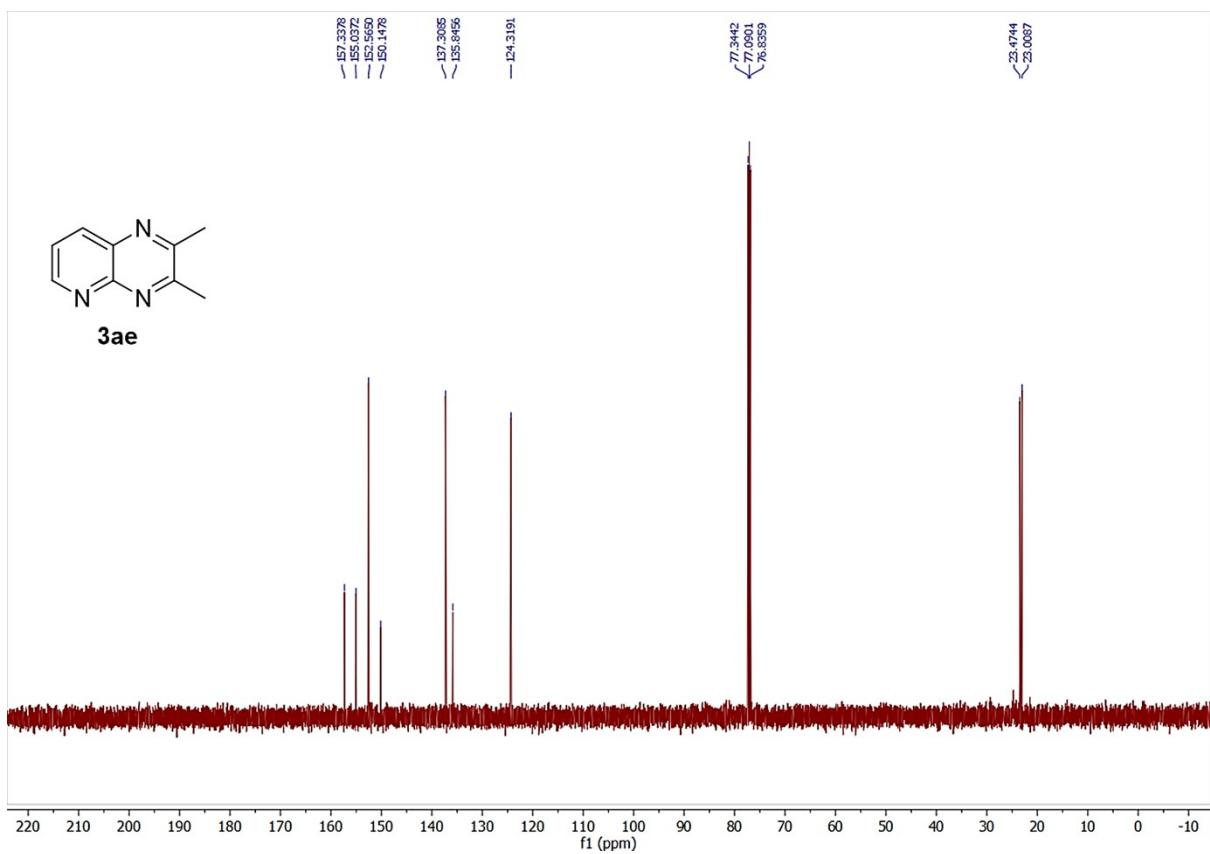
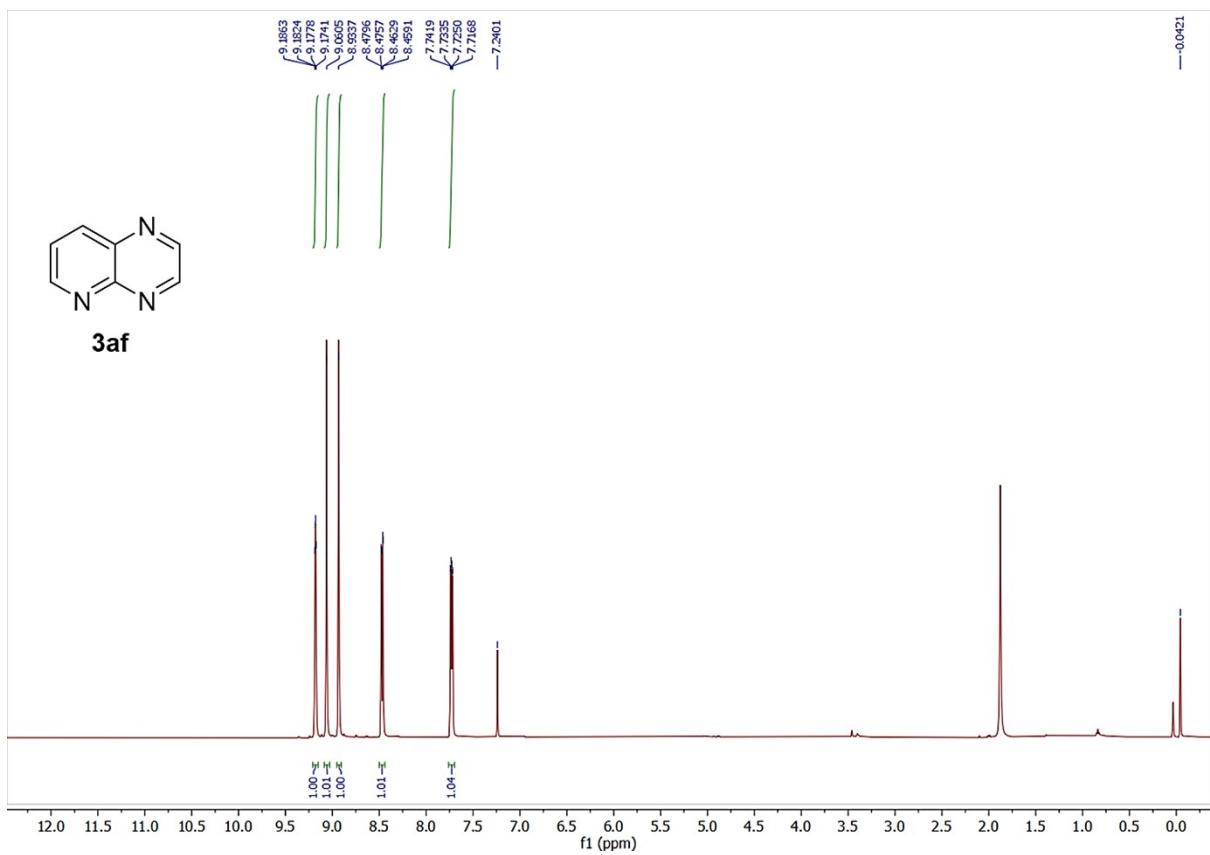


Figure S33: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound 3ae.



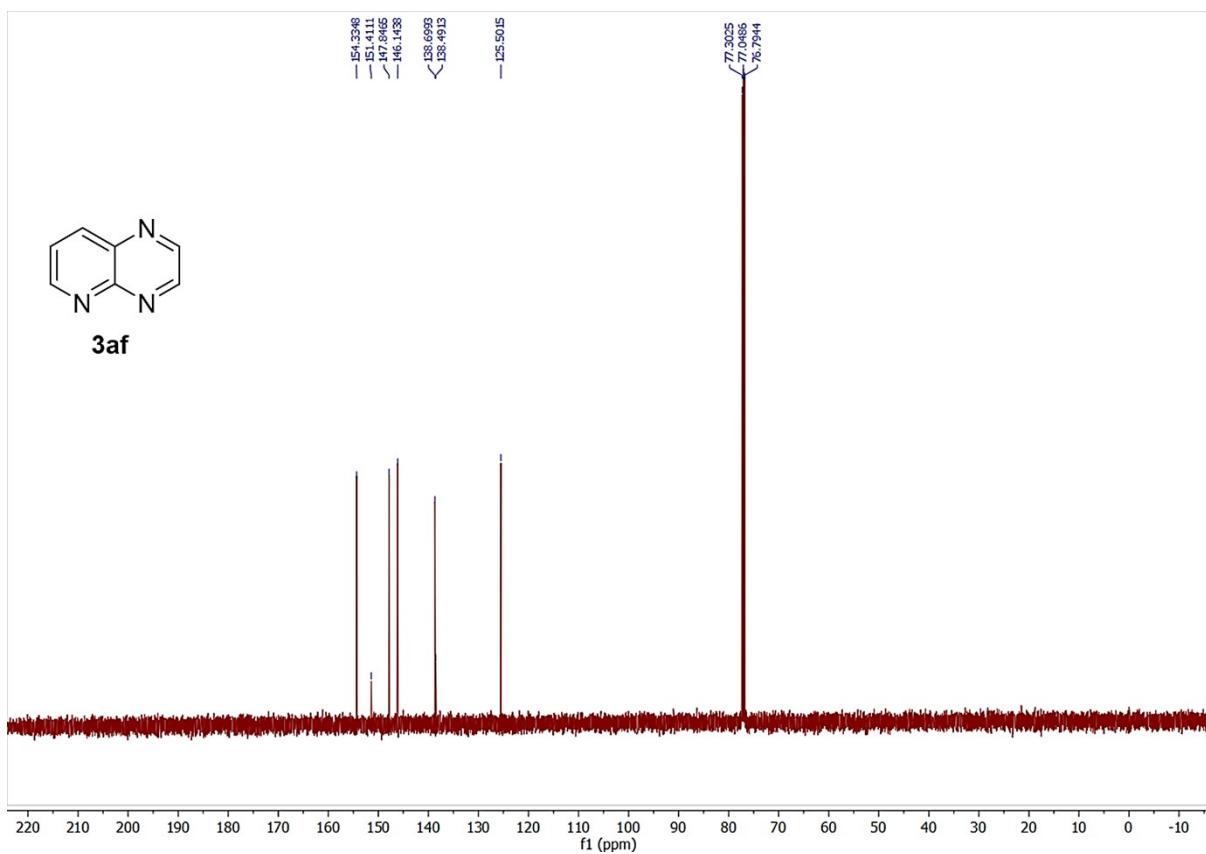
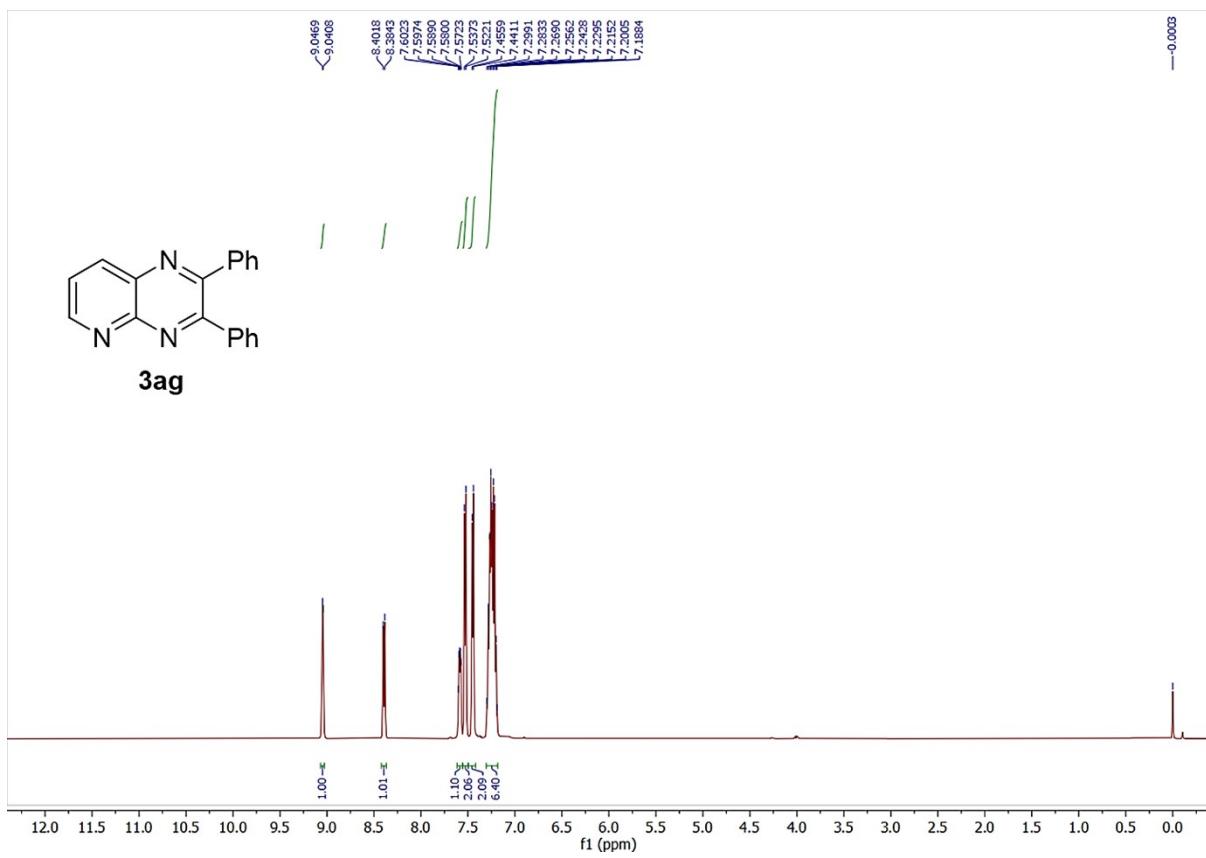


Figure S34:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound 3af.



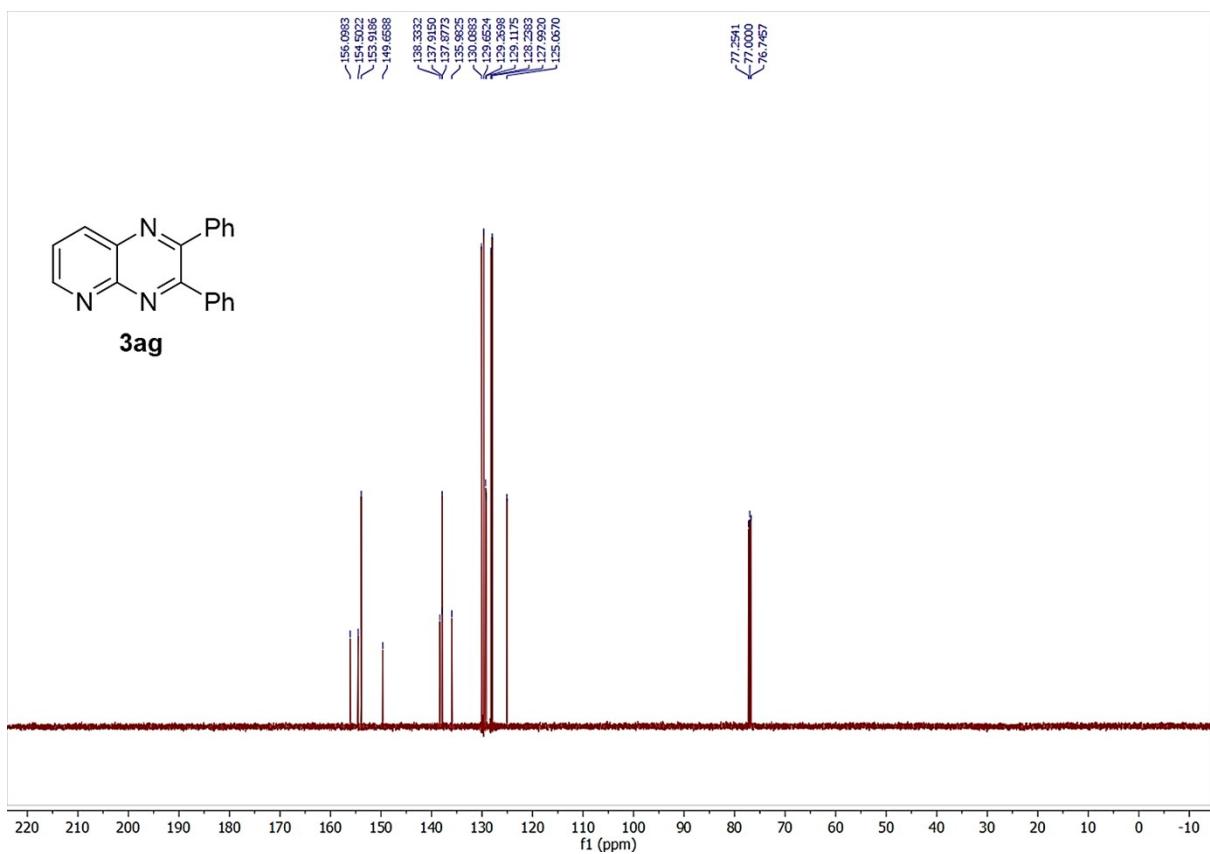
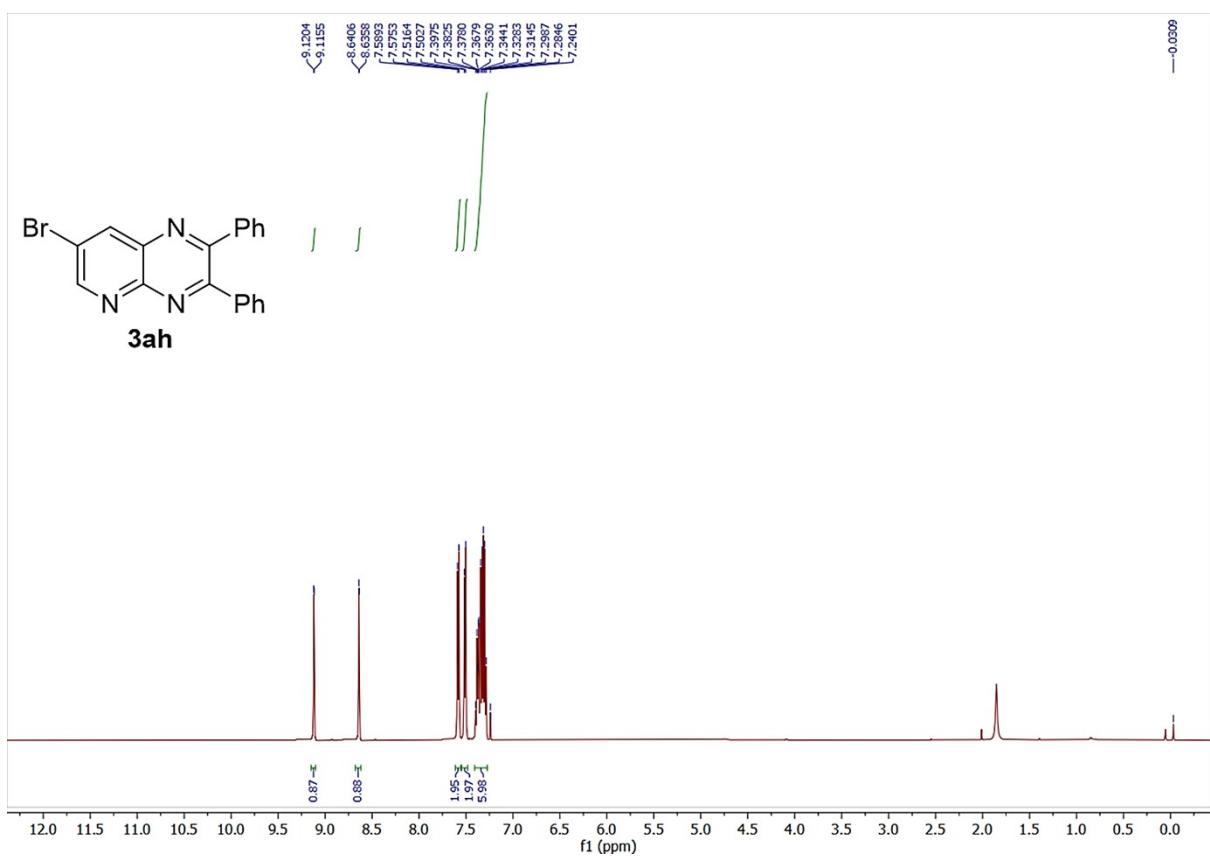


Figure S35:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3ag**.



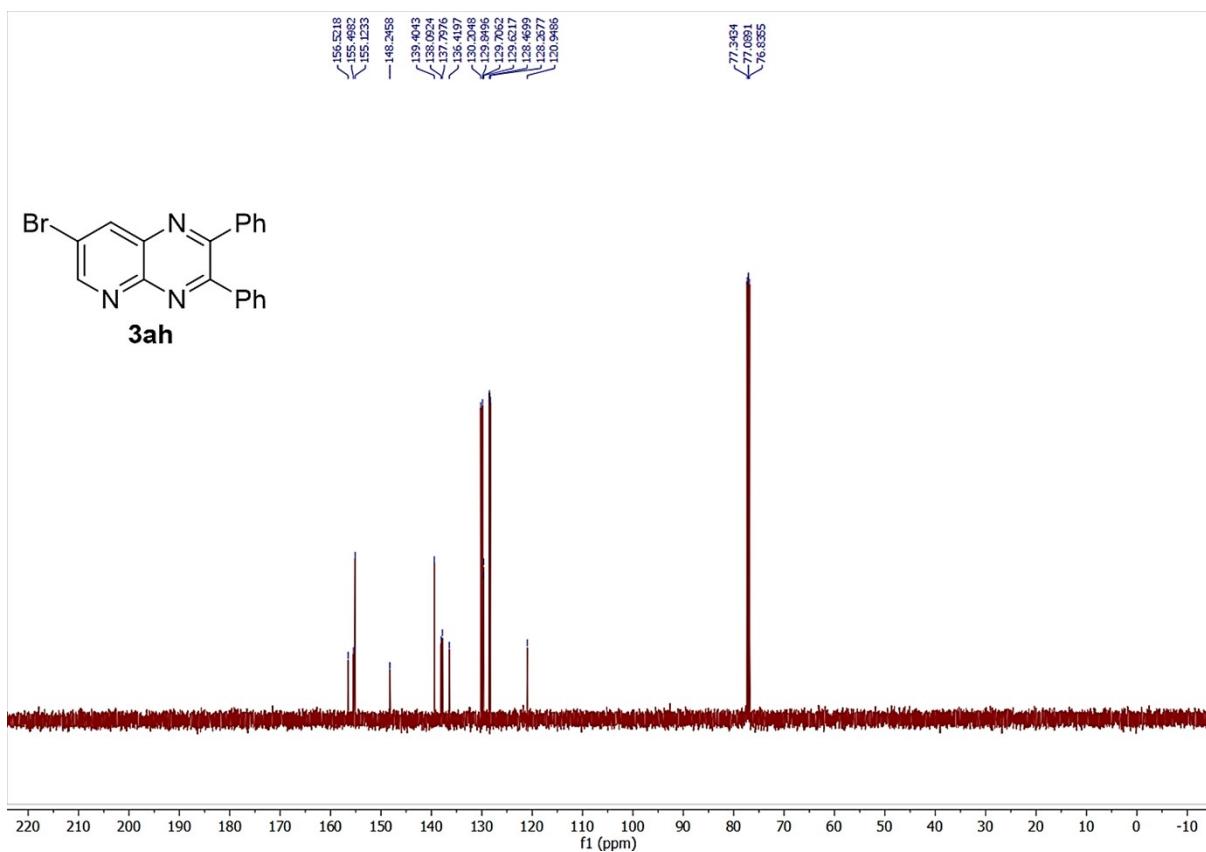
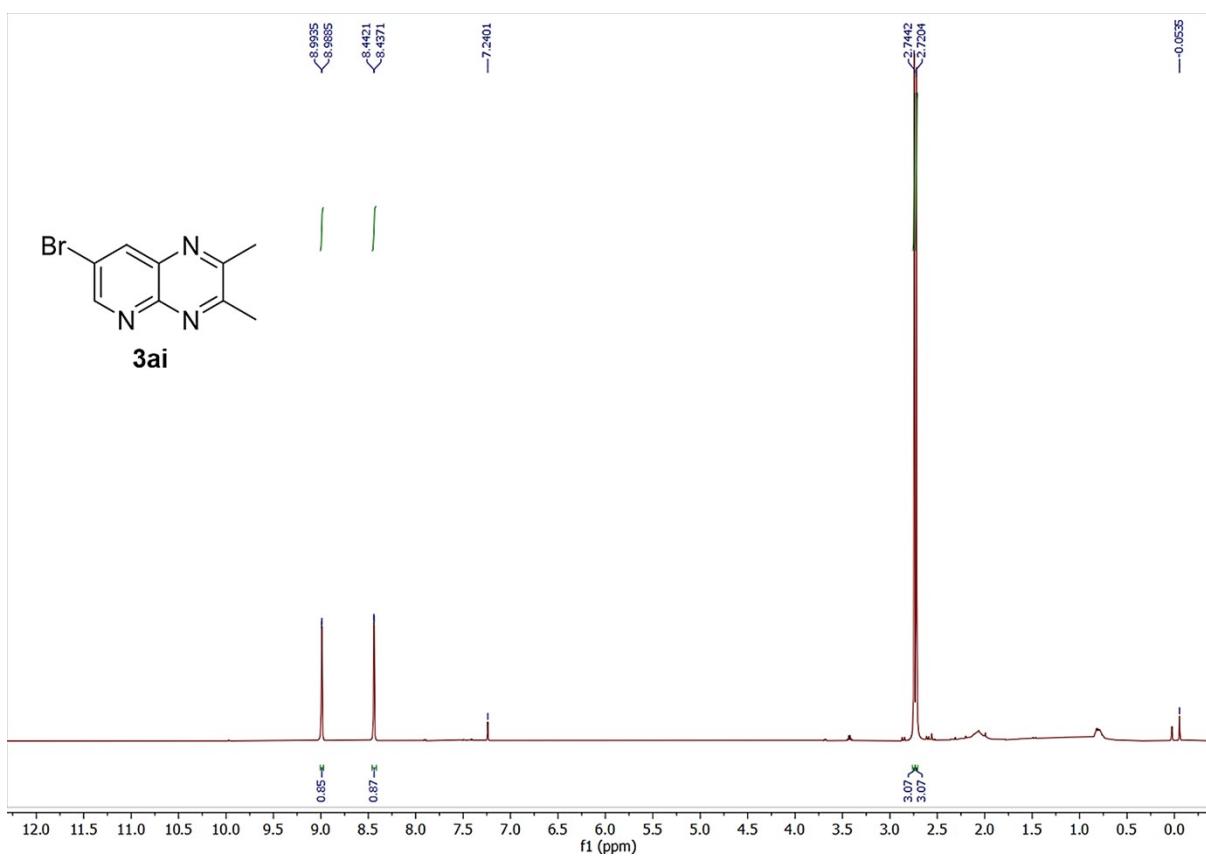


Figure S36:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3ah**.



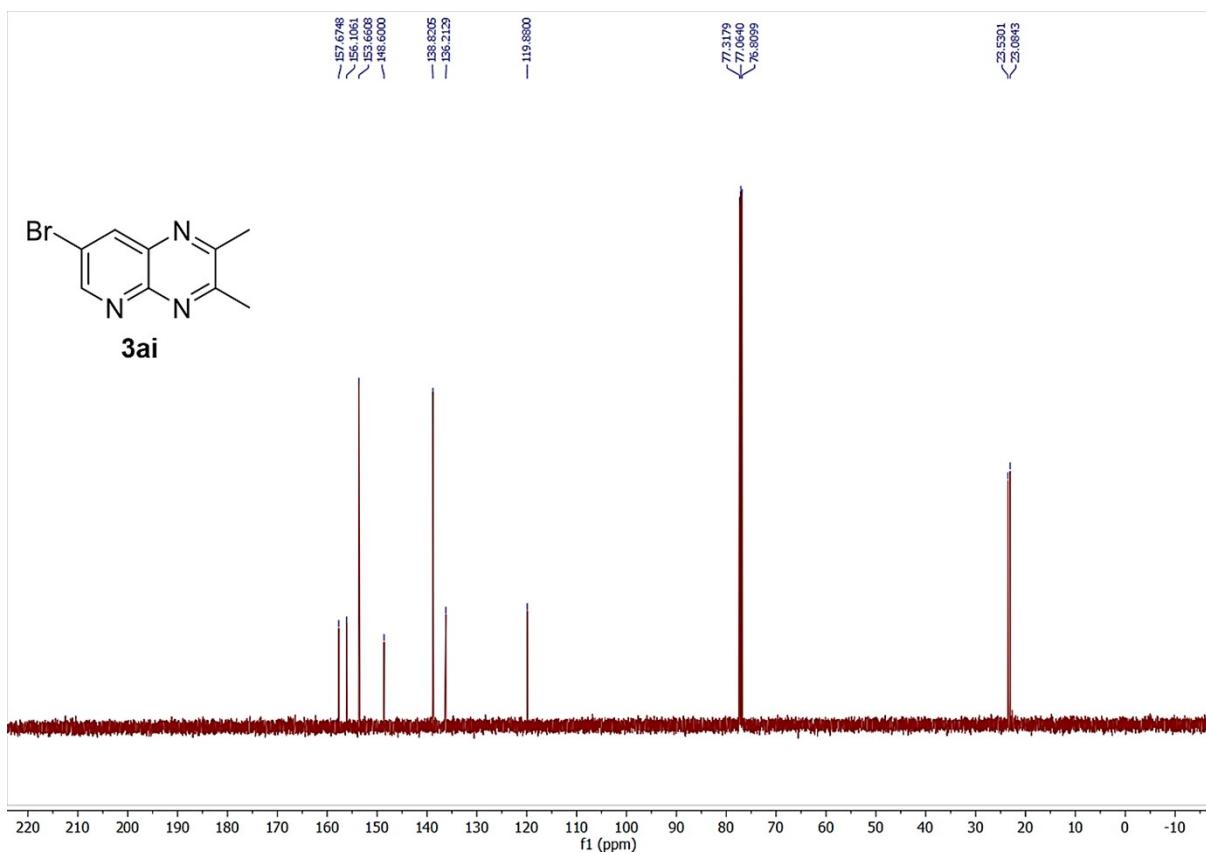
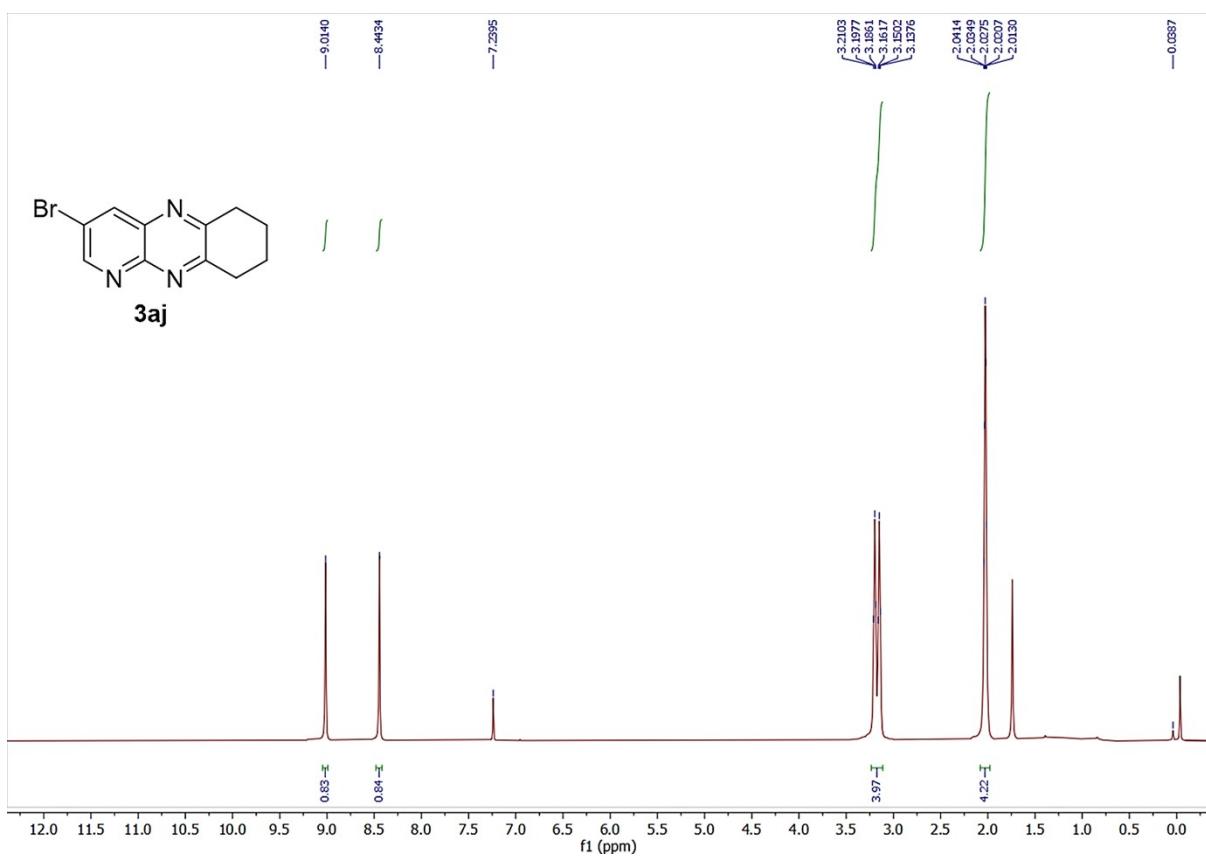


Figure S37:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3ai**.



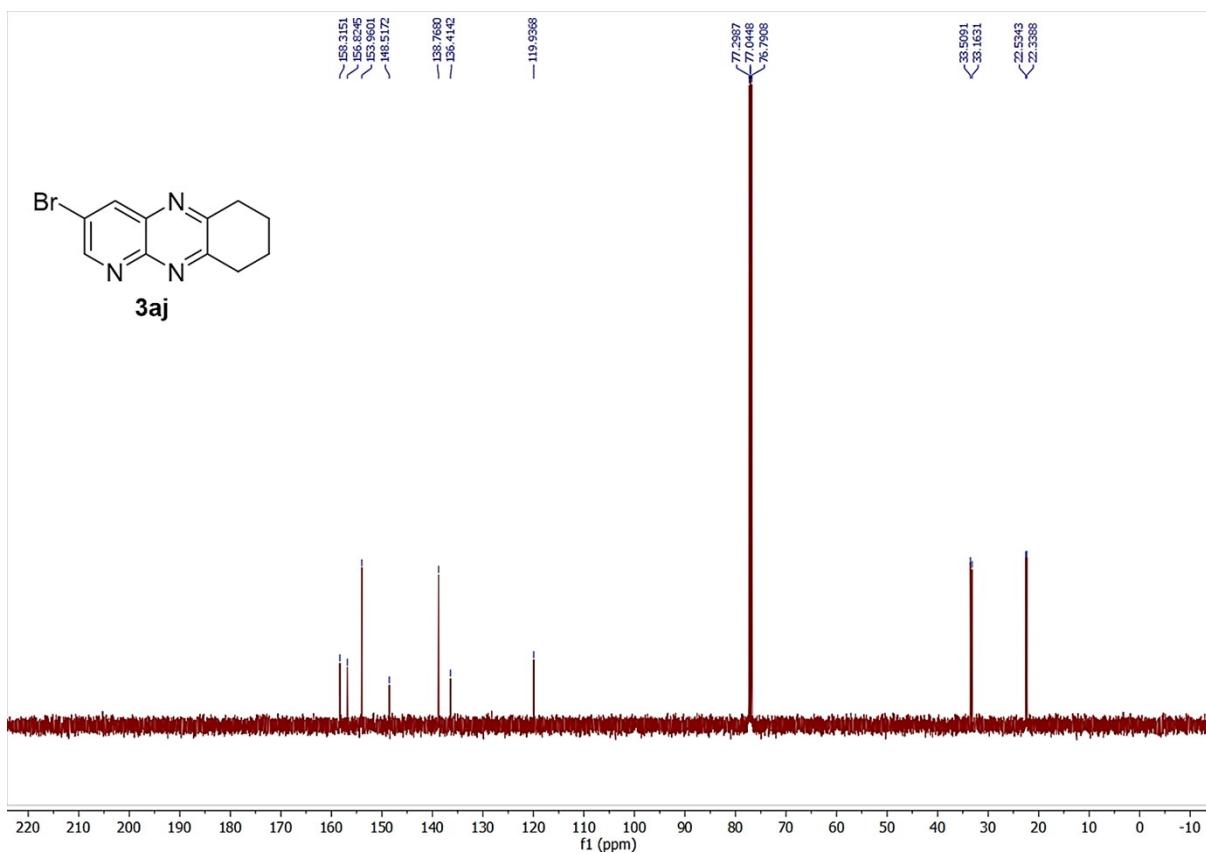
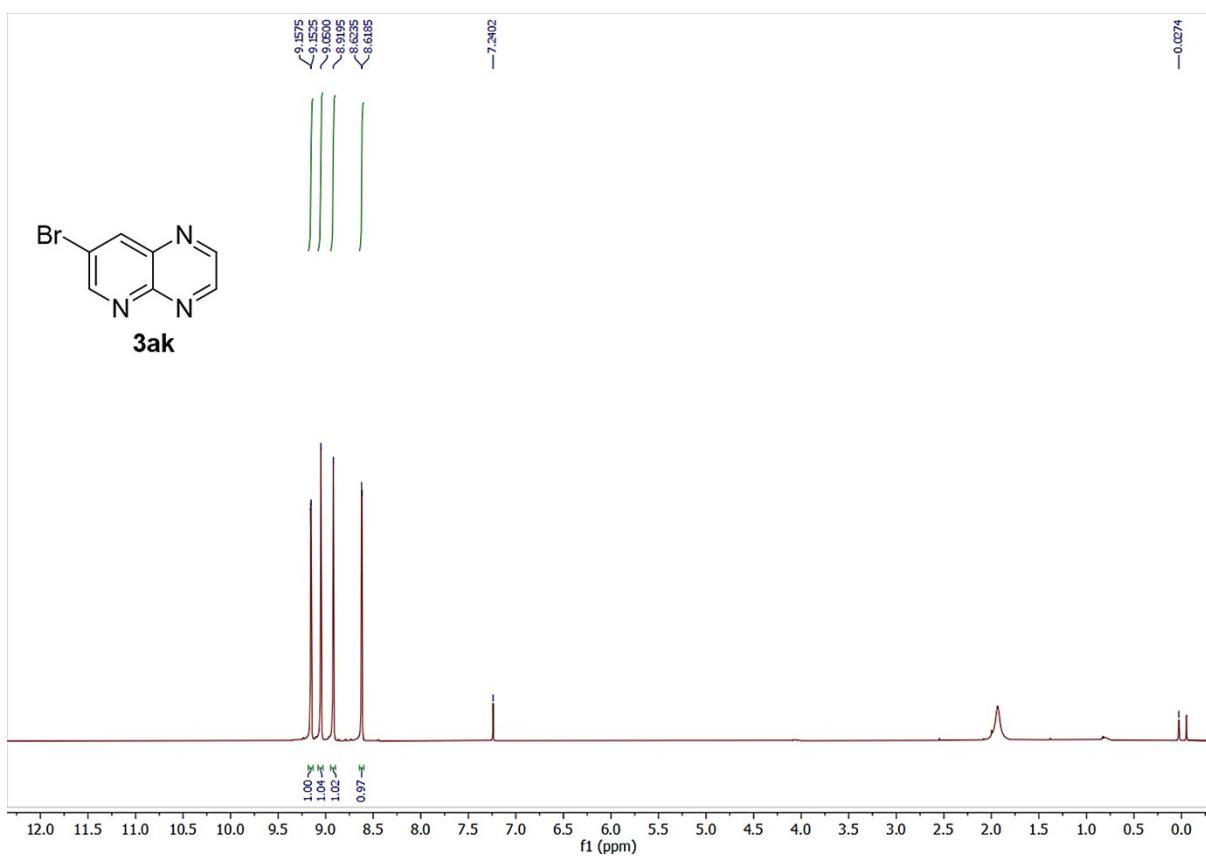


Figure S38:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3aj**.



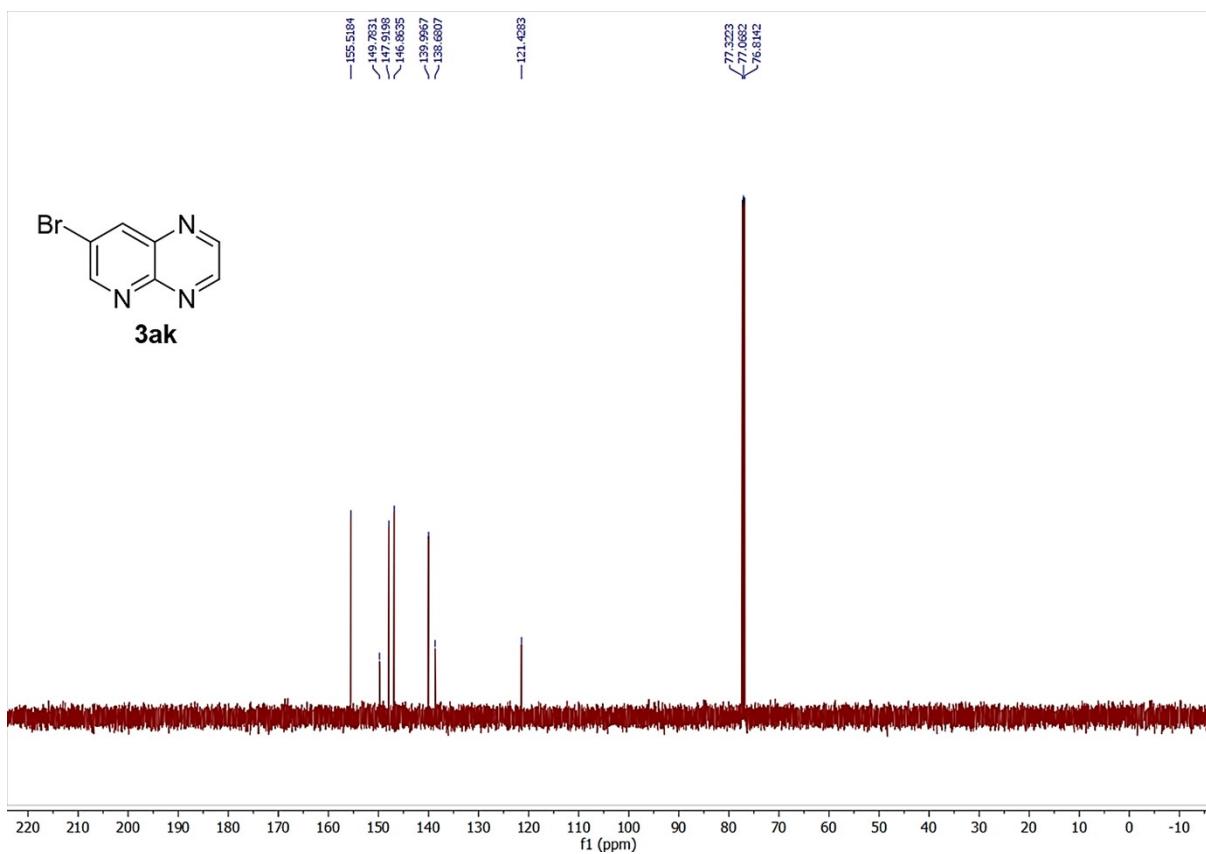
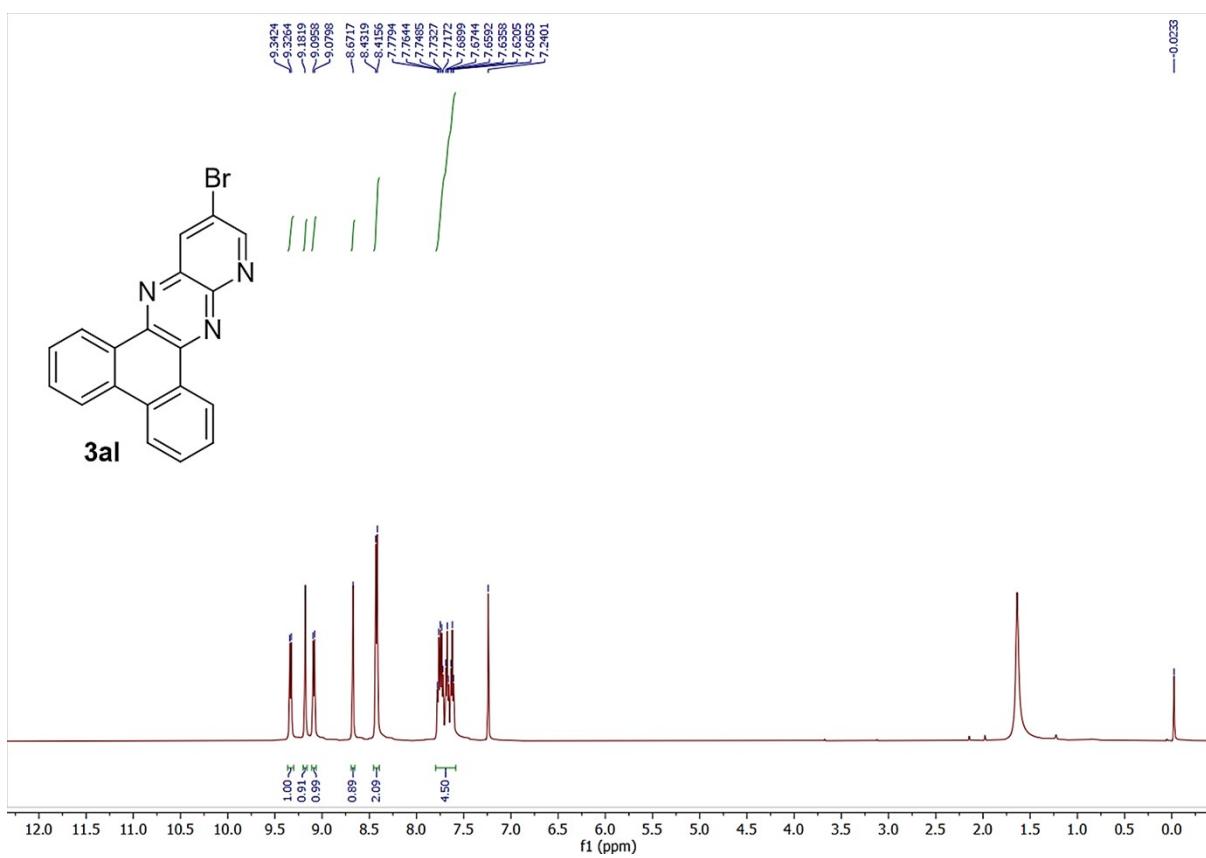


Figure S39: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound **3ak**.



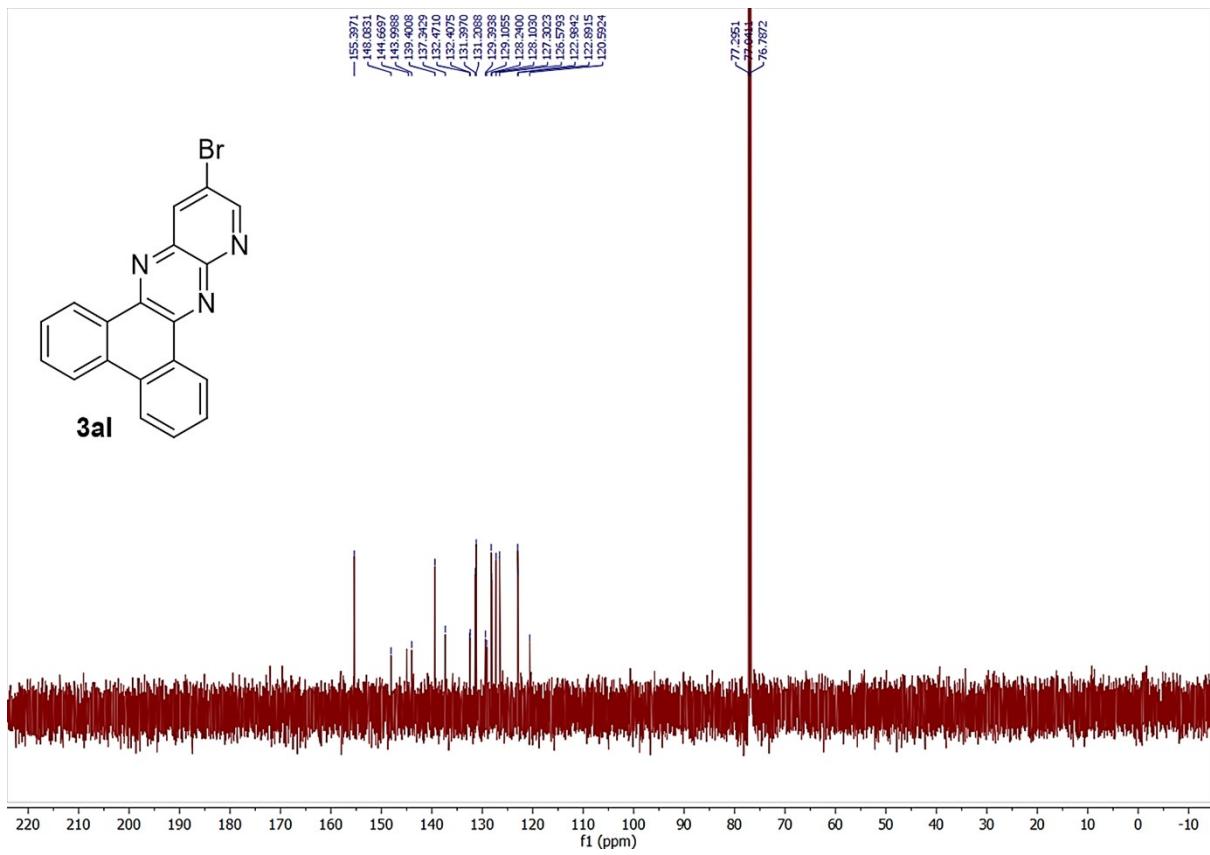
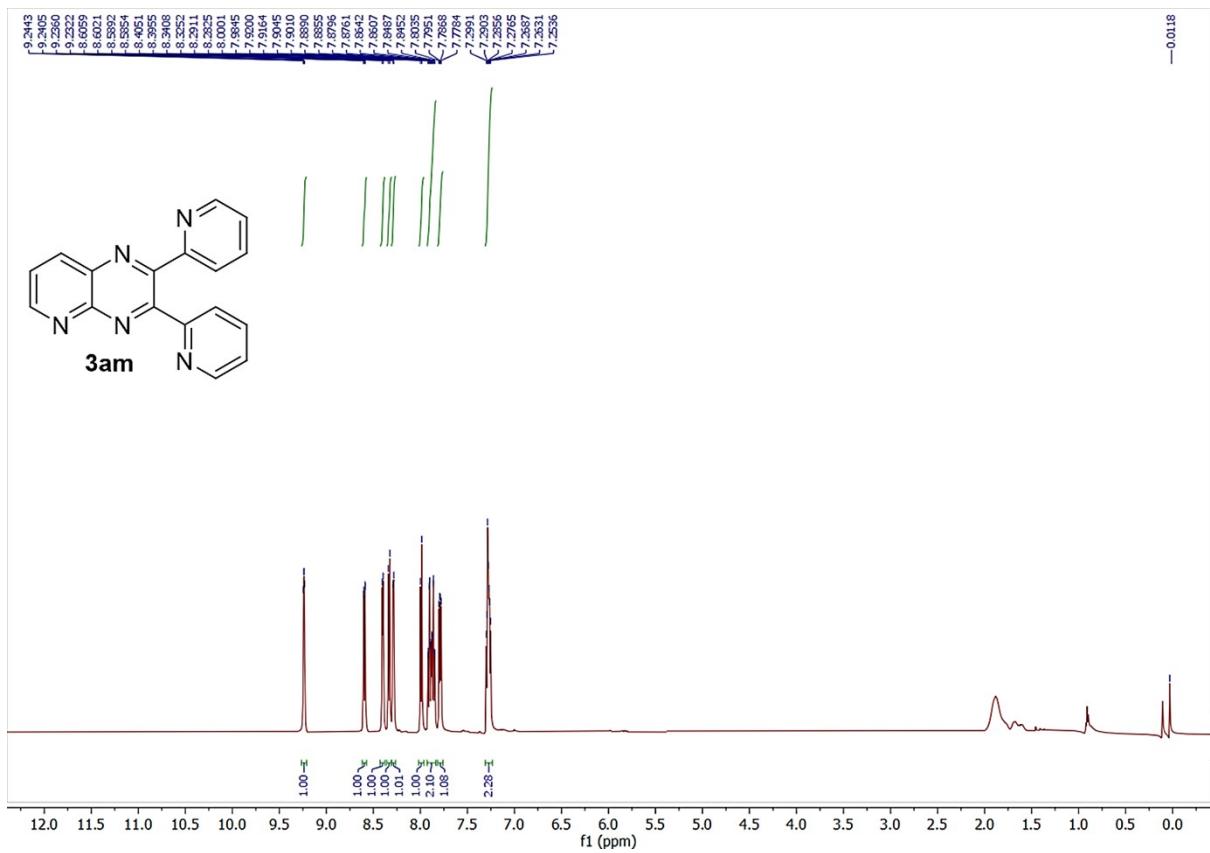


Figure S40:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound 3al.



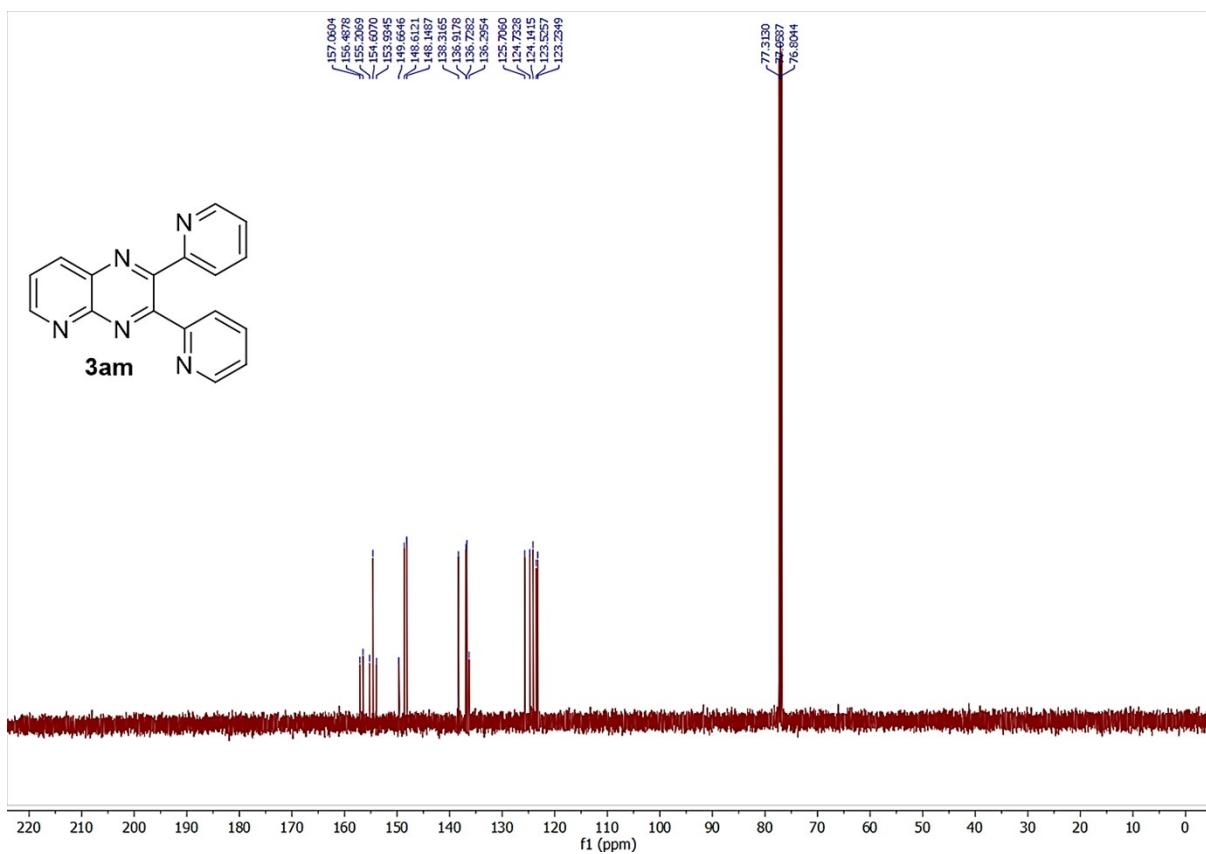
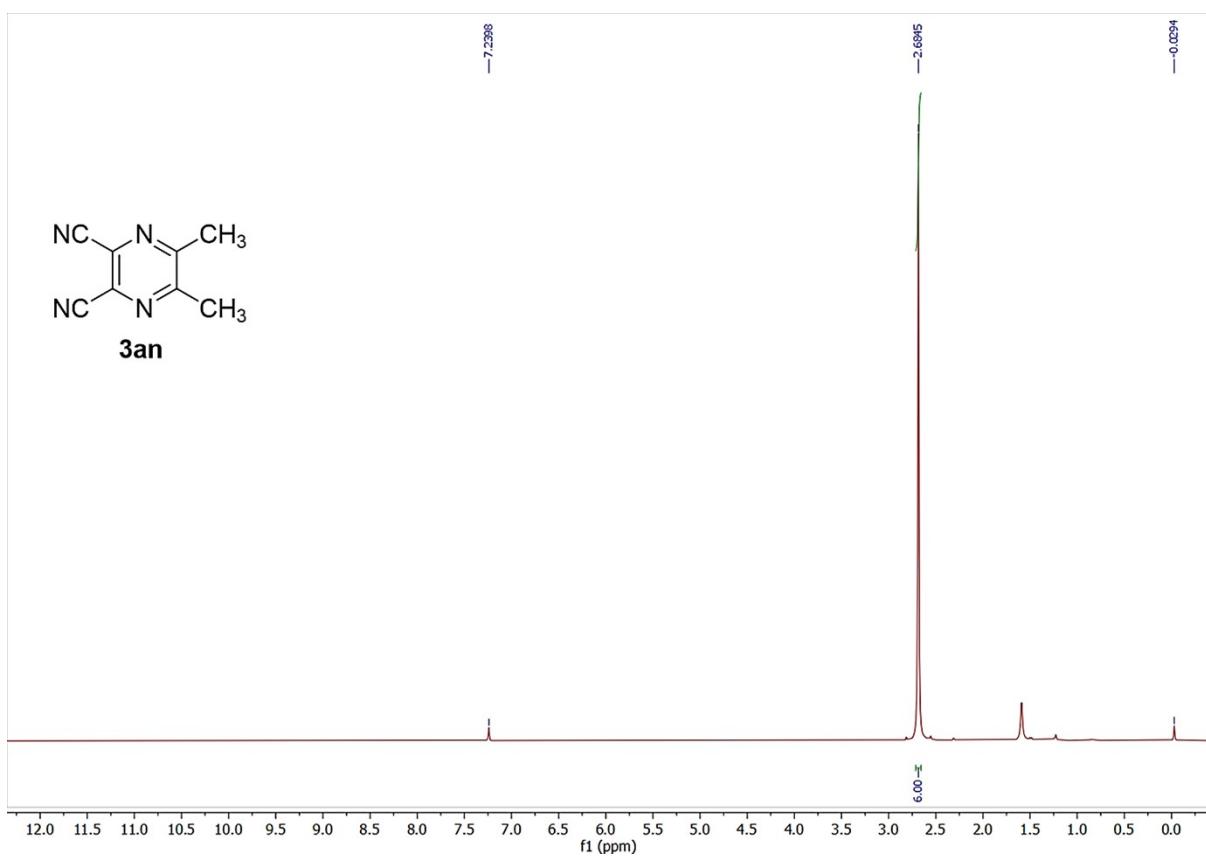


Figure S41:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3am**.



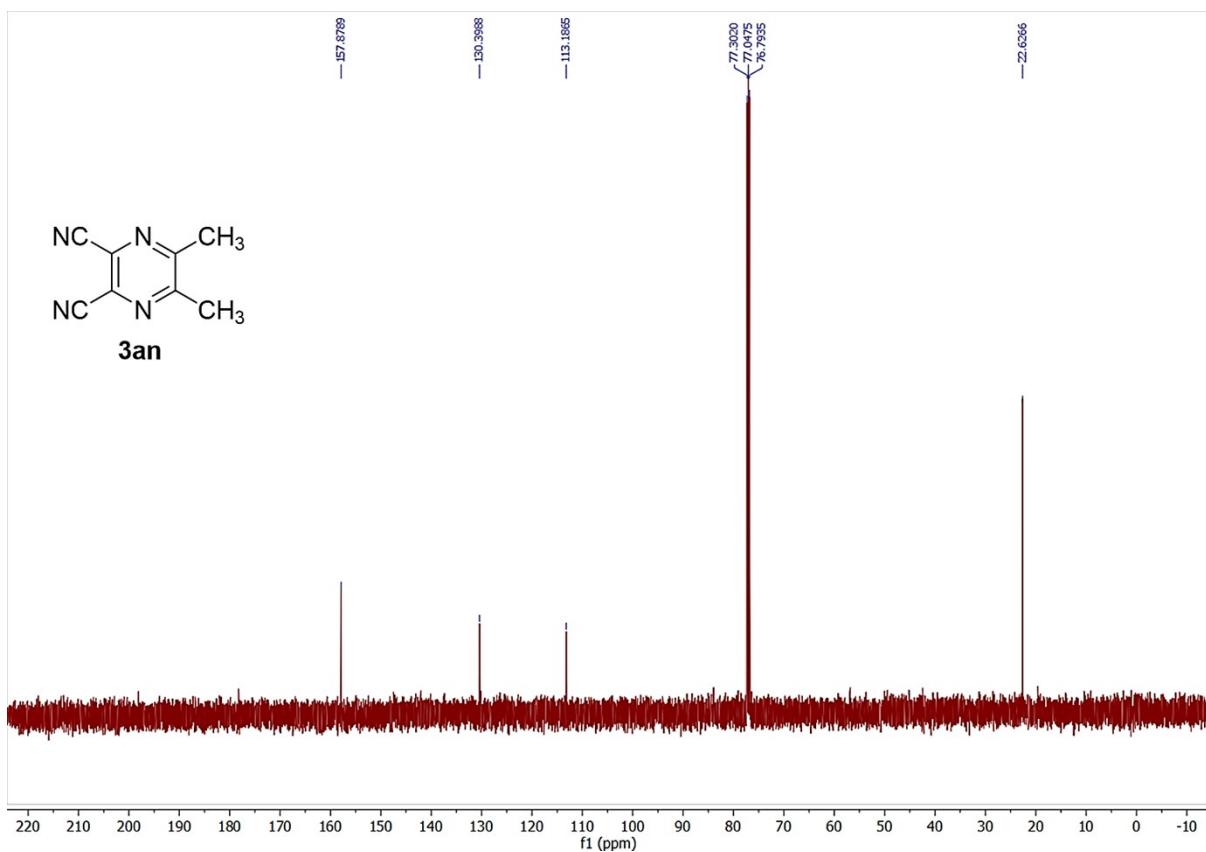
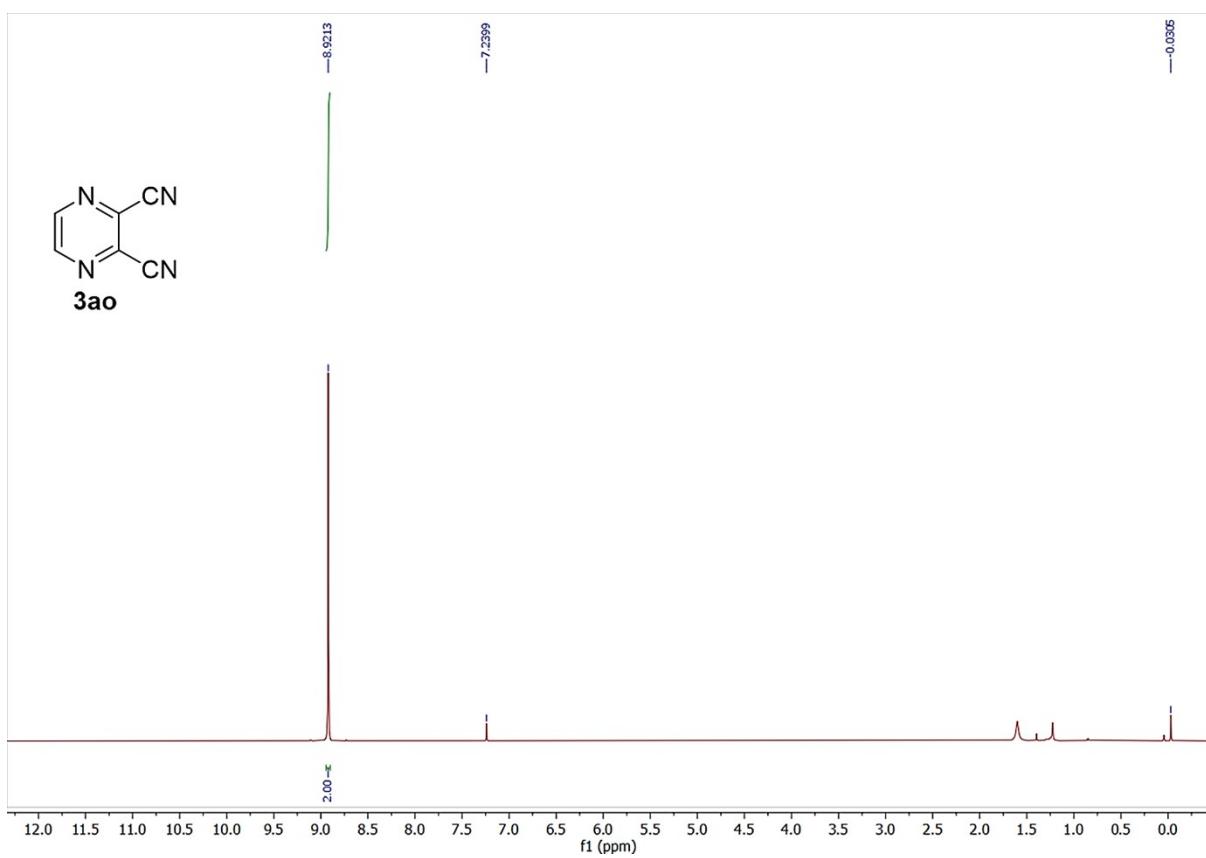


Figure S42: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound **3an**.



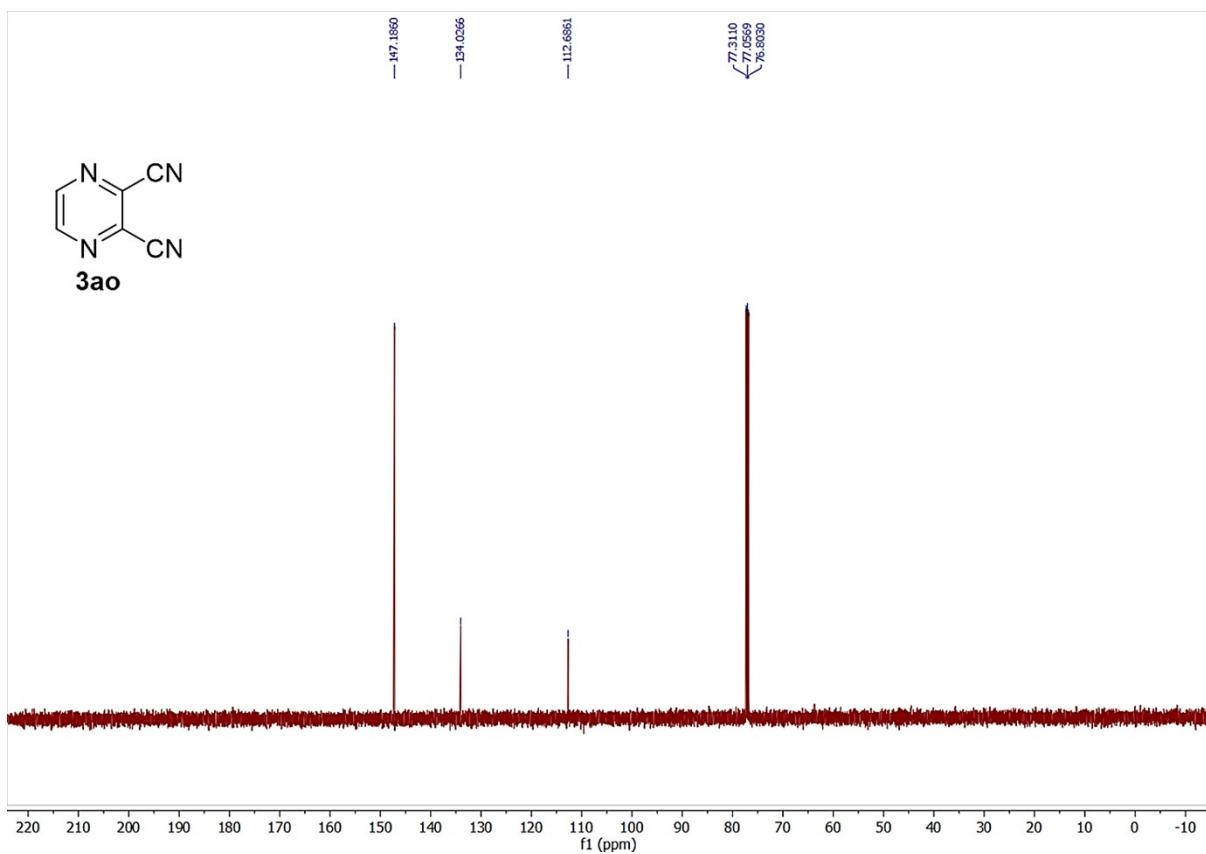
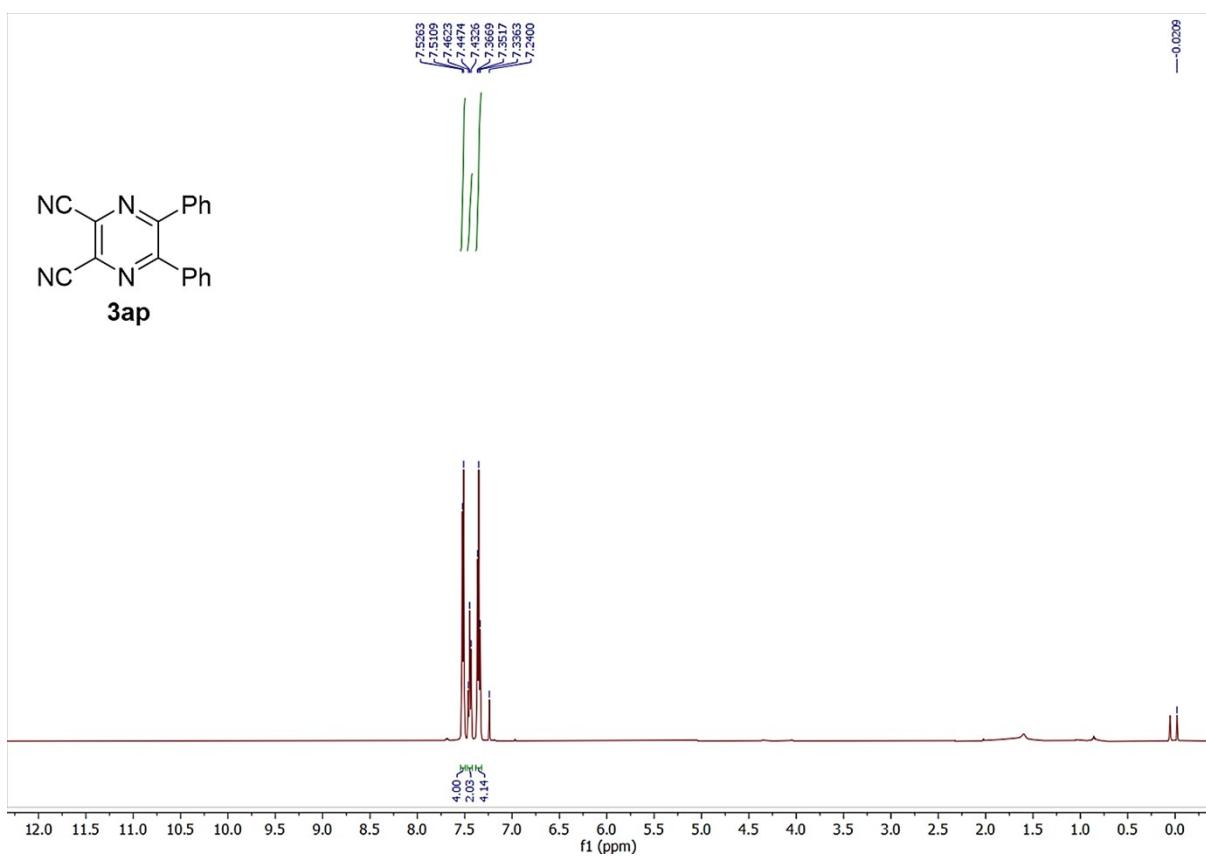


Figure S43: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound **3ao**.



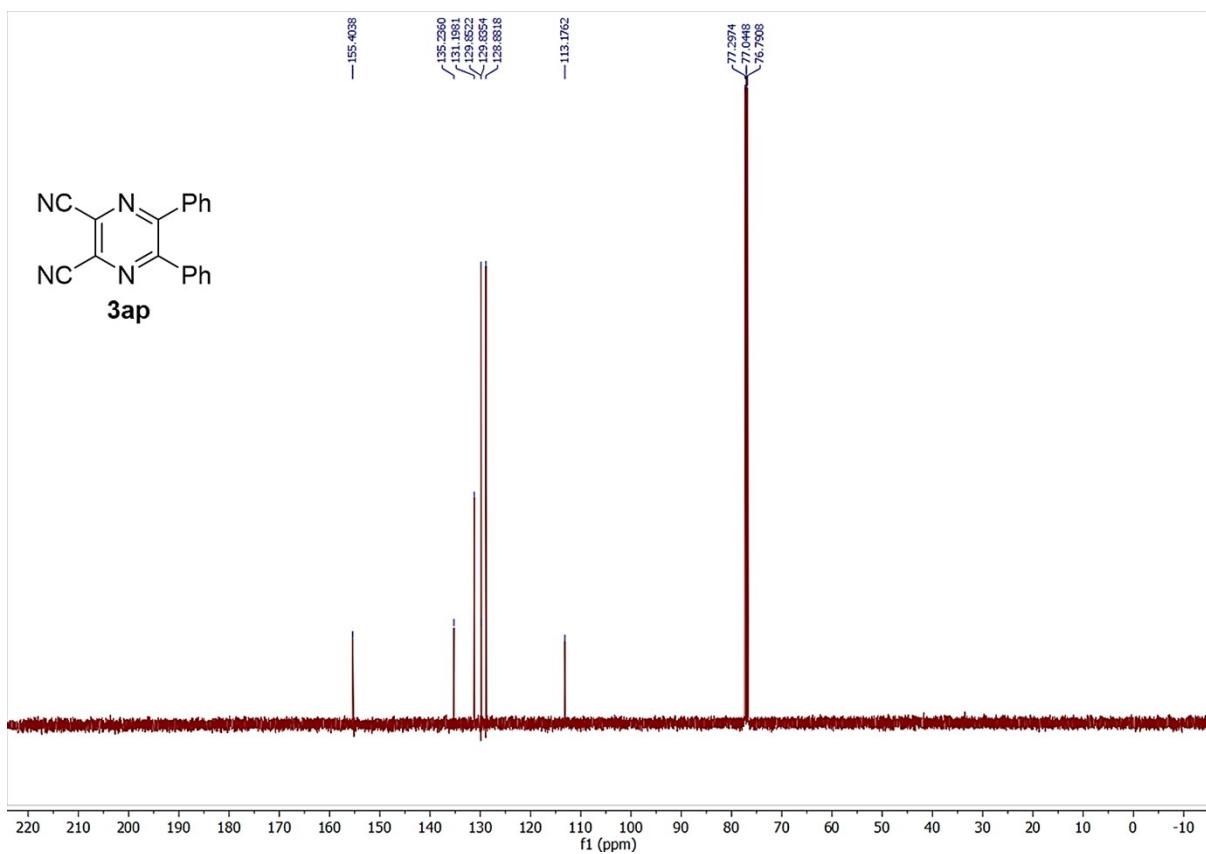
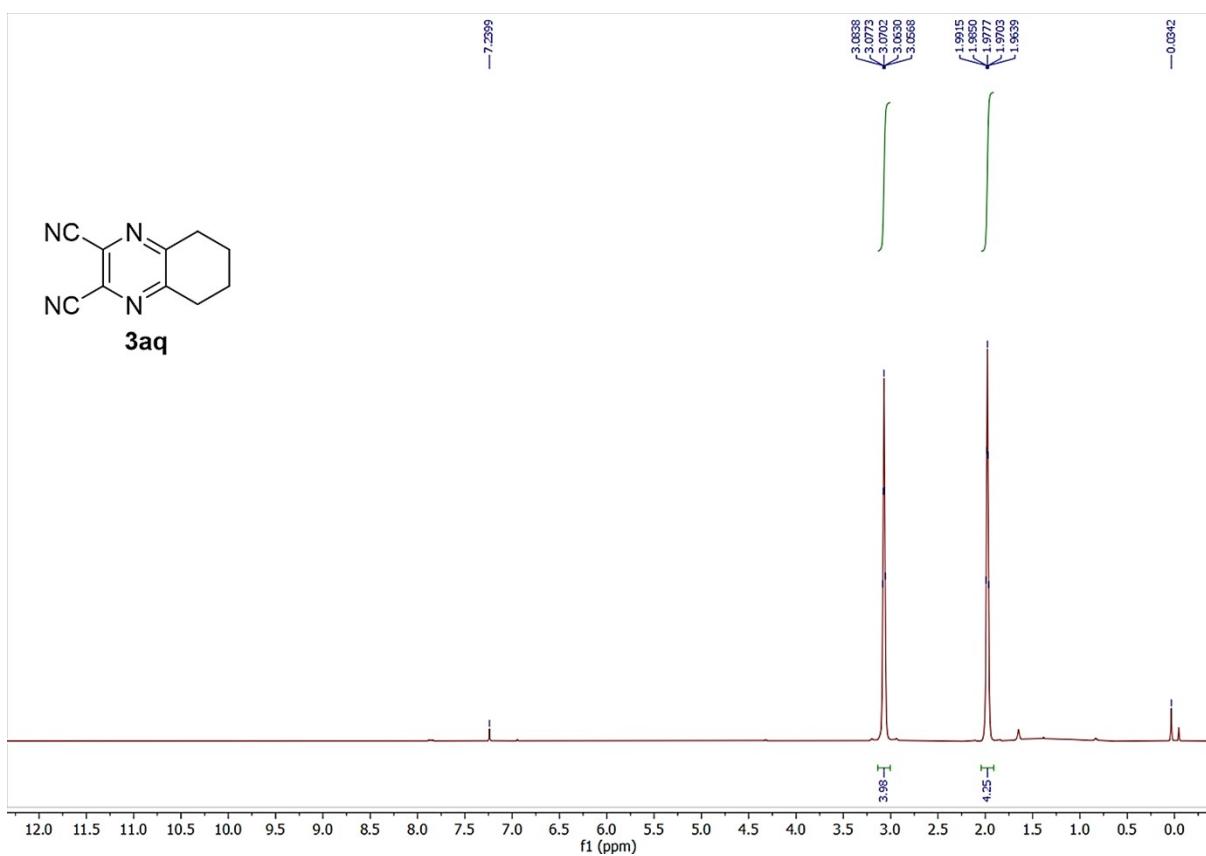


Figure S44: <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C (125 MHz, CDCl<sub>3</sub>) NMR spectra of compound 3ap.



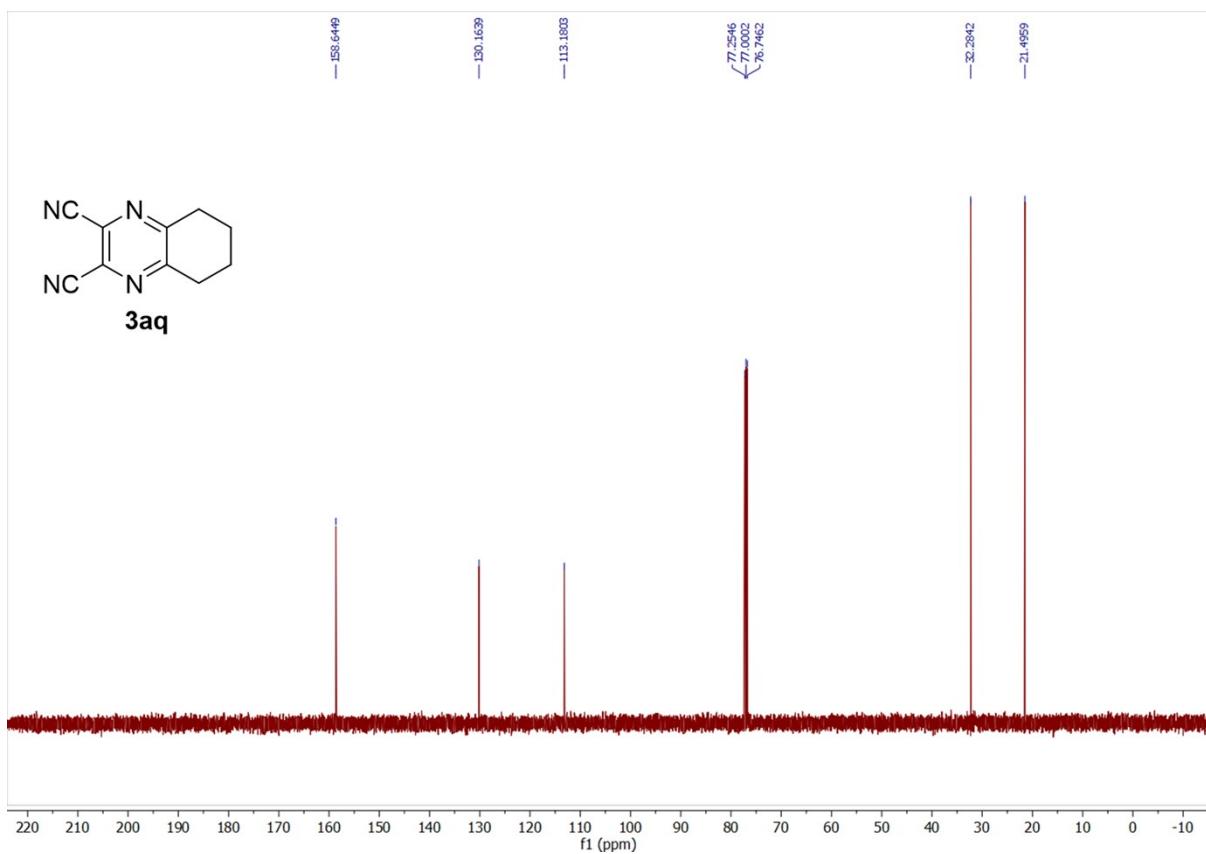
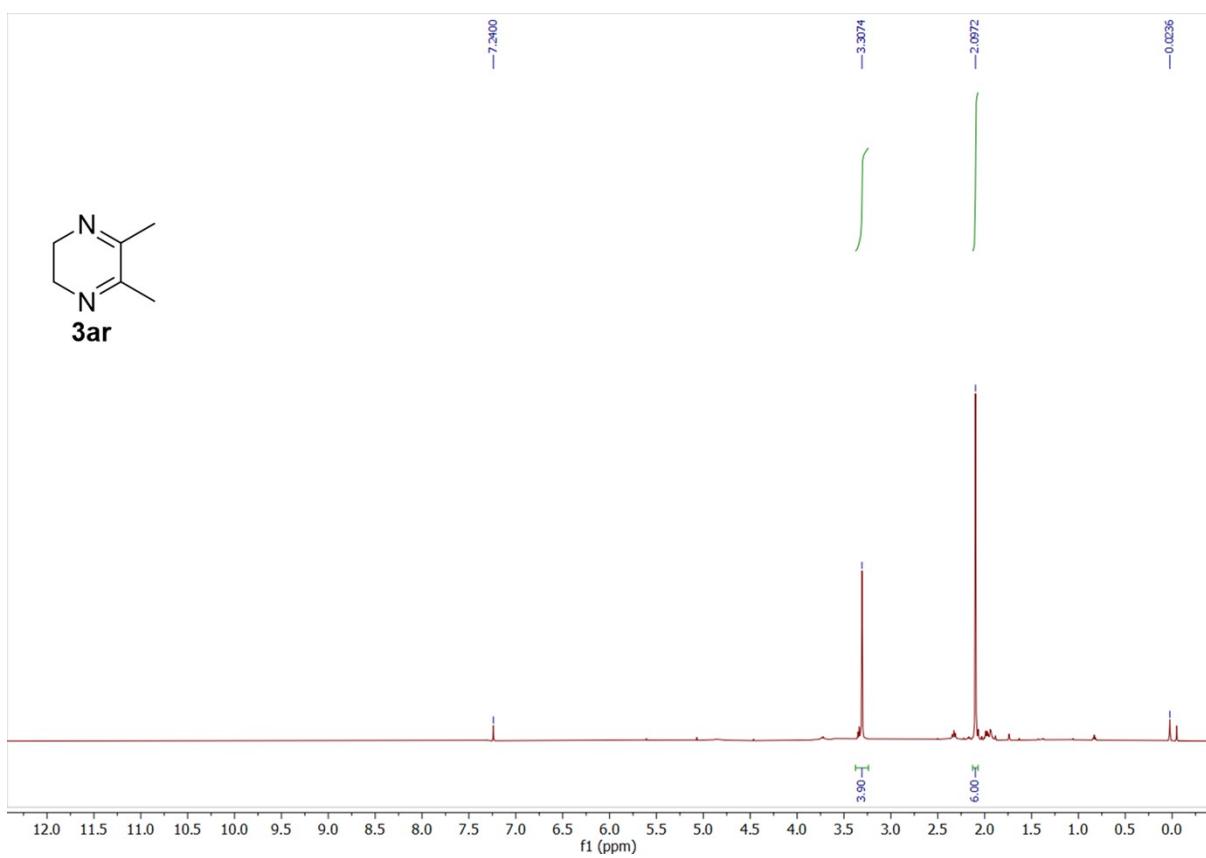


Figure S45:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3aq**.



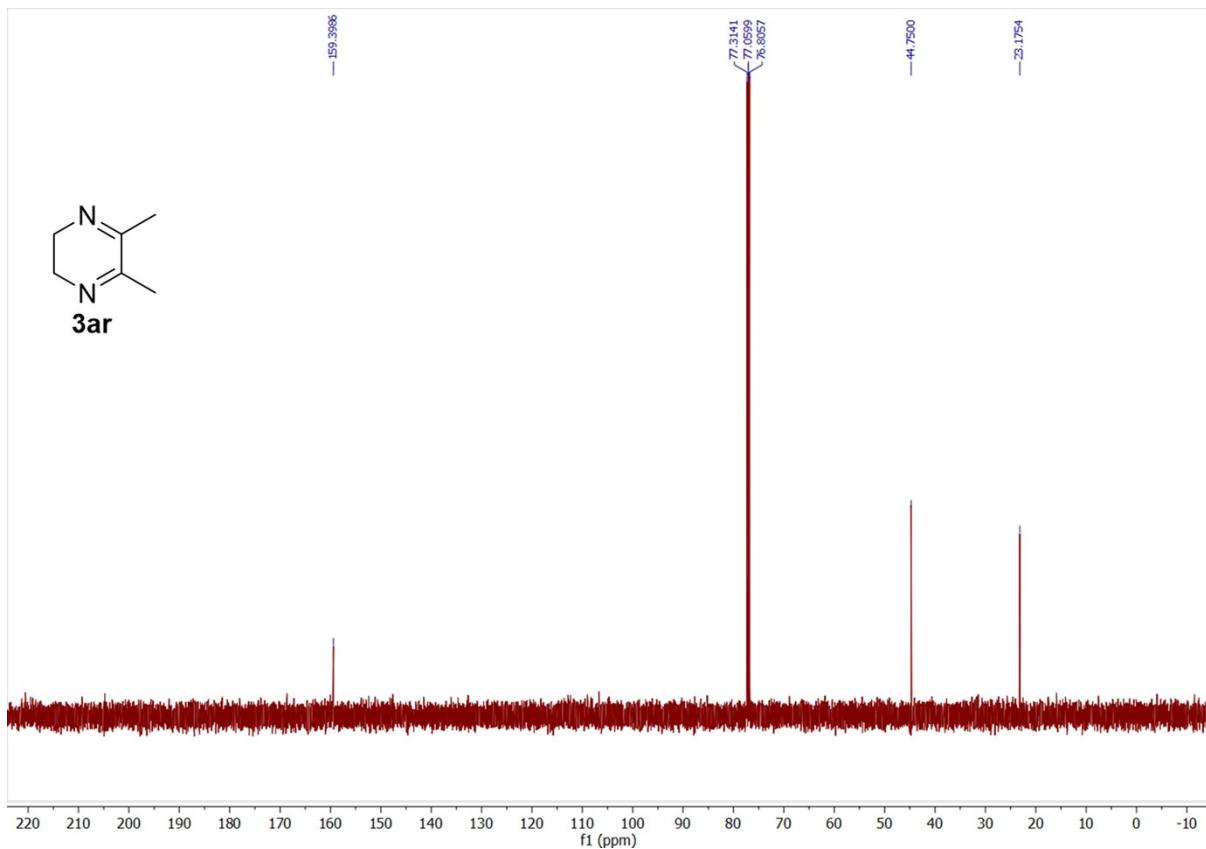


Figure S46:  $^1\text{H}$  (500 MHz,  $\text{CDCl}_3$ ) and  $^{13}\text{C}$  (125 MHz,  $\text{CDCl}_3$ ) NMR spectra of compound **3ar**.