

Supporting Information for:

Sesterterpenoids: sources, structural diversity, biological activity, and data management

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Table S1. Structures of sesterterpenoids

Table S2. Accepted name, family, species, synonyms, lifestyle, geographical distribution, and collection site of the selected Porifera

Table S3. Accepted name, family, species, synonyms, lifestyle, geographical distribution, and collection site of the selected Mollusca

Table S4. Accepted name, family, species, synonyms, lifestyle, geographical distribution, and collection site of the selected Cnidaria

Table S5. Accepted name, family, species, synonyms, and geographical distribution of the selected Insecta

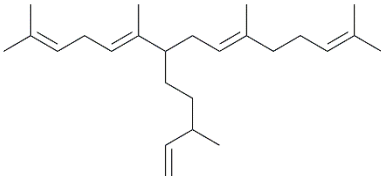
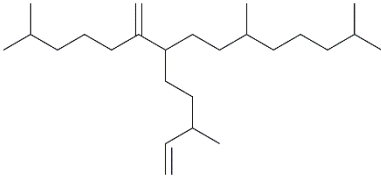
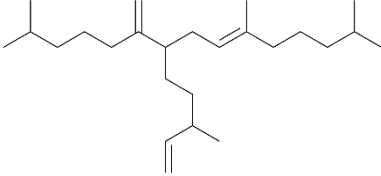
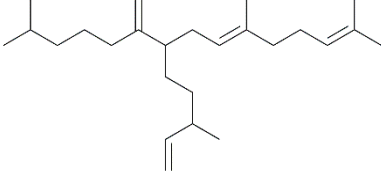
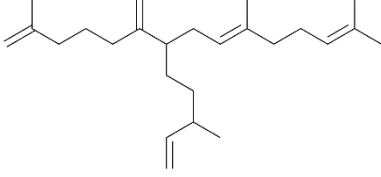
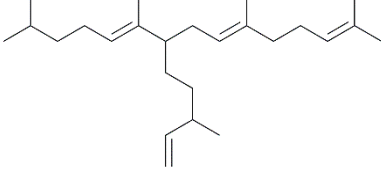
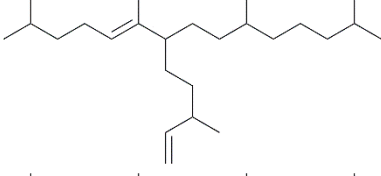
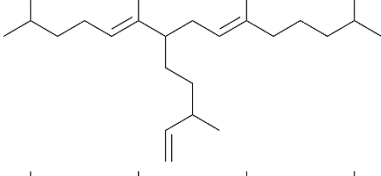
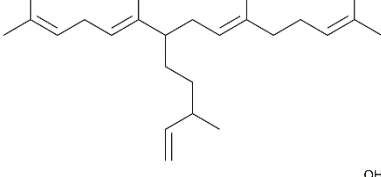
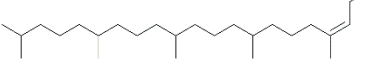
Table S6. Accepted name, family, species, synonyms, lifestyle, geographical distribution, and common environment of the selected microorganisms

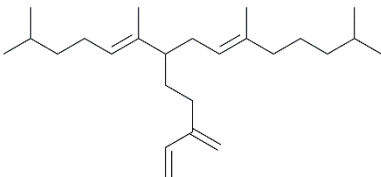
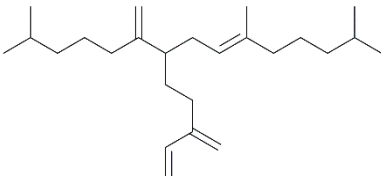
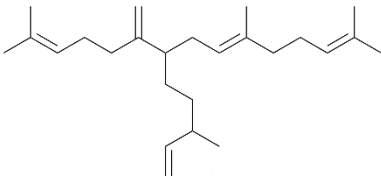
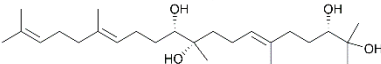
Table S7. Name of the species as reported in the publication, accepted name, family, synonyms, lifeform, native range, and ecology of the selected plants

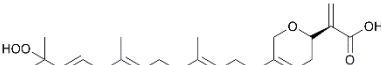
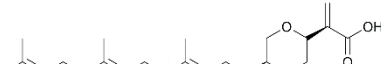
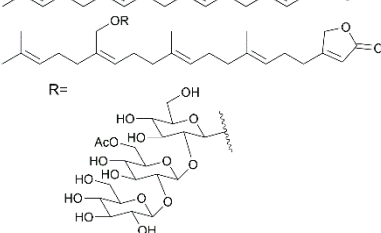
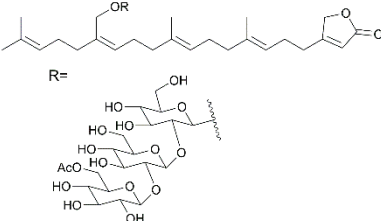
Table S8. Biological activities of sesterterpenoids

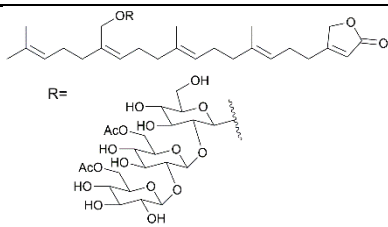
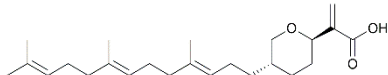
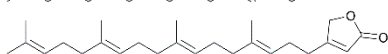
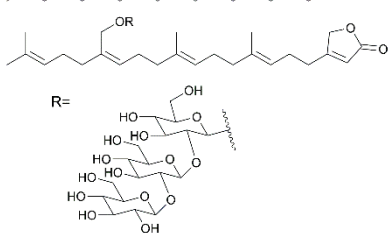
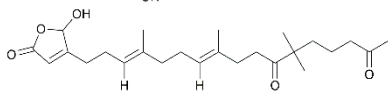
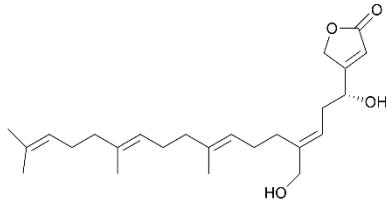
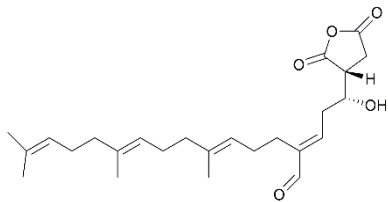
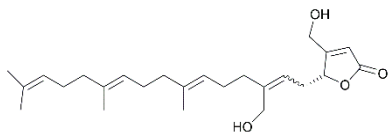
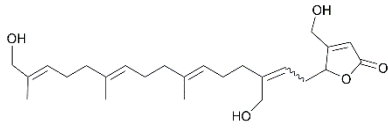
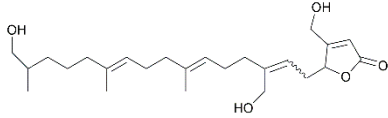
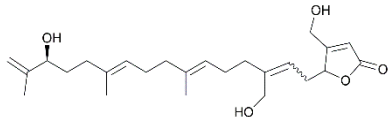
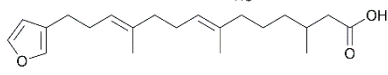
Table S1. Structures of sesterterpenoids (all the structures are reported as depicted in the cited papers)

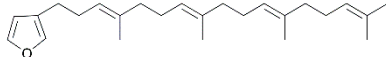
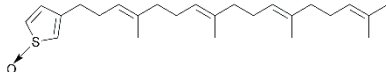
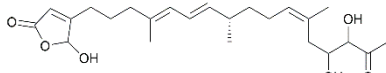
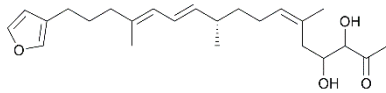
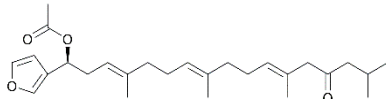
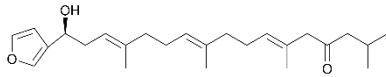
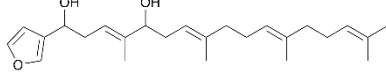
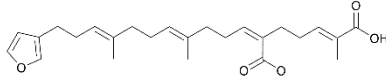
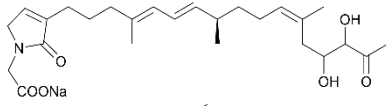
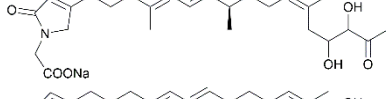
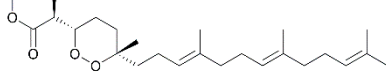
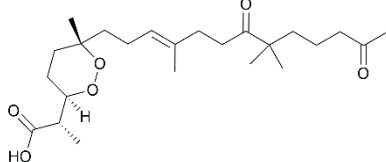
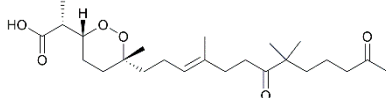
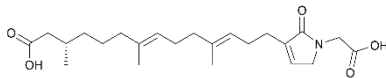
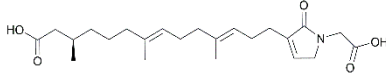
Section 1.1 Structures of linear acyclic (AI) sesterterpenoids lacking heterocyclic rings			
c. n.	IUPAC name or Trivial name	Structure	Organism as reported in the reference
AI-1	geranylfarnesol		<i>Ceroplastes albolineatus</i> ¹
AI-2	ω-hydroxygeranylfarnesol		<i>Fasciospongia fovea</i> ² <i>Ceroplastes albolineatus</i> ³
AI-3	geranylnerolidol		<i>Cochliobolus heterostrophus</i> ⁴
AI-4	3,7,11,15,19-pentamethyl-2-cis-6-trans-eicosadien-1-ol		<i>Solanum tuberosum</i> ⁵
AI-5	postrediene A (2,6,11,15,19-pentamethyl-6E,14E-eicosadiene2,3,10,11,18,19-hexanol)		<i>Pleurotus ostreatus</i> and <i>Trametes robiniophila</i> (coculture) ⁶
AI-6	postrediene B (2,6,11,15,19-pentamethyl-10,11,18,19-tetrahydroxyl-2E,6E,14E-eicosatrienoic acid)		<i>Pleurotus ostreatus</i> and <i>Trametes robiniophila</i> (coculture) ⁶
AI-7	postrediene C (2,6,11,15,19-pentamethyl-10,11,18,19-tetrahydroxyl-6E,14E-eicosadienoic acid)		<i>Pleurotus ostreatus</i> and <i>Trametes robiniophila</i> (coculture) ⁶
AI-8	(2Z,6Z,10E,14E)-geranylfarnesol		<i>Triticum aestivum</i> ⁷
AI-9	(2Z,6E,10E,14E)-geranylfarnesol		<i>Croton hieronymi</i> ⁸
AI-10	unnamed acyclic sesterterpene (reported as compound 5)		<i>Hippospongia lachne</i> ⁹
AI-11	2,6,10,14-tetramethyl-18-butanecarboxymethylenecarboxymethyl-17β-ol		<i>Oryza sativa</i> ¹⁰
AI-12	unnamed acyclic sesterterpene (reported as compound 3)		<i>Rhizosolenia setigera</i> ¹¹

AI-13	unnamed acyclic sesterterpene (reported as compound 4)		<i>Rhizosolenia setigera</i> ¹¹
AI-14	hasla-6(17),23-diene		<i>Haslea ostrearia</i> ^{12, 13}
AI-15	hasla-6(17),9,23-triene		<i>Haslea ostrearia</i> ^{12, 13}
AI-16	hasla-6(17),9,13,23-tetraene		<i>Haslea ostrearia</i> , <i>Haslea saltstonica</i> , <i>Haslea crucigera</i> ^{12, 13}
AI-17	hasla-2,6(17),9,13,23-pentaene		<i>Haslea ostrearia</i> ^{12, 13}
AI-18	hasla-5,9,13,23-tetraene		<i>Haslea ostrearia</i> , <i>Haslea pseudostrearia</i> ^{12, 13}
AI-19	hasla-5,23-diene		<i>Haslea ostrearia</i> ^{12, 13}
AI-20	hasla-5,9,23-triene		<i>Haslea ostrearia</i> ^{12, 13}
AI-21	hasla-2,5,9,13,23-pentaene		<i>Haslea ostrearia</i> ^{12, 13}
AI-22	2 <i>E</i> -3,7,11,15,19-pentamethyleicos-2-en-1-ol (Z isomer)		<i>Croton hieronymi</i> ⁸

	reported in the structure)		
AI-23	2,6,10,14-tetramethyl-7-(3-methylenepent-4-enyl)pentadec-5,9-diene		<i>Haslea ostrearia</i> ¹³
AI-24	2,6,14-trimethyl-10-methylene-9-(3methylenepent-4-enyl)pentadec-6-diene		<i>Haslea ostrearia</i> ¹³
AI-25	pentaene VIII		<i>Haslea crucigera</i> ^{12, 13}
AI-26	postrediene D		<i>Pleurotus ostreatus</i> SY10 ¹⁴

Section 1.2. Structures of linear monoheterocyclic (MH) sesterterpenoids			
c. n.	IUPAC name or Trivial name	Structure	Organism as reported in the reference
MH-1	rhopaloic acid H		<i>Hippospongia</i> sp. ¹⁵
MH-2	rhopaloic acid B		<i>Rhopaloeides</i> sp. ¹⁶
MH-3	rhopaloic acid C		<i>Rhopaloeides</i> sp. ¹⁶ <i>Hippospongia</i> sp. ¹⁵
MH-4	6''-O-acetylbalibaloside		<i>Oscarella balibaloj</i> ¹⁷
MH-5	6'''-O-acetylbalibaloside		<i>Oscarella balibaloj</i> ¹⁷

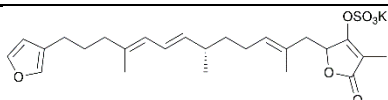
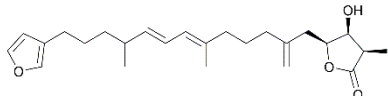
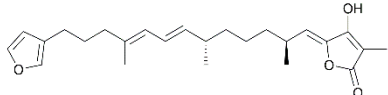
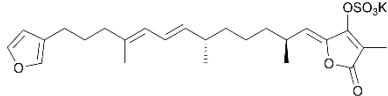
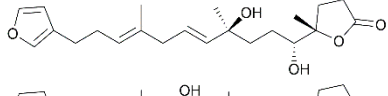
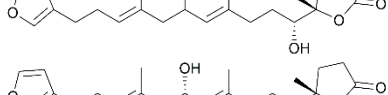
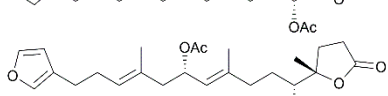
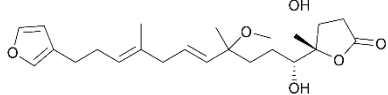
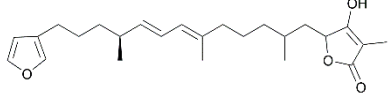
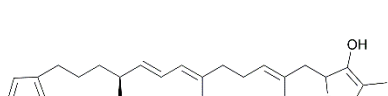
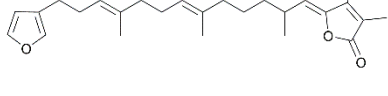
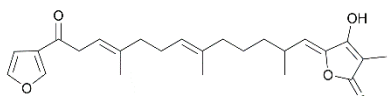
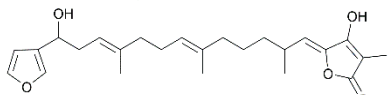
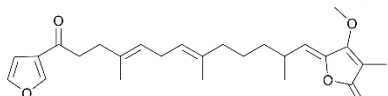
MH-6	6'',6'''-O-diacetylbalibaloside		<i>Oscarella balibaloï</i> ¹⁷
MH-7	rhopaloic acid A		<i>Rhopaloeides</i> sp. ¹⁸ <i>Hippospongia</i> sp. ¹⁵
MH-8	luffarin Q		<i>Thorecta horridus</i> ¹⁹ <i>Luffariella geometrica</i> ²⁰
MH-9	balibaloside		<i>Oscarella balibaloï</i> ¹⁷
MH-10	acantholide C		<i>Acanthodendrilla</i> sp. ²¹ <i>Hyrtios communis</i> ²²
MH-11	hippolide C		<i>Hippospongia lachne</i> ²³
MH-12	hippolide D		<i>Hippospongia lachne</i> ²³
MH-13	hippolide E		<i>Hippospongia lachne</i> ²³
MH-14	hippolide F		<i>Hippospongia lachne</i> ²³
MH-15	hippolide G		<i>Hippospongia lachne</i> ²³
MH-16	hippolide H		<i>Hippospongia lachne</i> ²³
MH-17	unnamed monoheterocyclic sesterterpene (reorted as C21-furanoterpene)		<i>Sarcotragus</i> sp. ²⁴

MH-18	furospinosulin-1		<i>Fasciospongia fovea</i> ² <i>Ircinia spinosula</i> ²⁵ <i>Spongia idia</i> ²⁶ <i>Dactylospongia elegans</i> ²⁷ <i>Xestospongia</i> sp. ²⁸ <i>Thorecta</i> sp. ²⁹ <i>Xestospongia</i> sp. ²⁸
MH-19	3-(4,8,12,16-tetramethylheptadeca-3,7,11,15-tetraenyl)-thiophene-1-oxide		
MH-20	sarcotin O		<i>Sarcotragus</i> sp. ³⁰
MH-21	sarcotin P		<i>Sarcotragus</i> sp. ³¹
MH-22	4-acetoxy-9-deoxoidiadione		<i>Smenospongia</i> sp. ³²
MH-23	4-hydroxy-9-deoxoidiadione		<i>Smenospongia</i> sp. ^{32, 33}
MH-24	5,10-dihydroxyfurospinulo sine-1		<i>Hyrtios erectus</i> ³⁴
MH-25	isofurospingin-4		<i>Spongia officinalis</i> ³⁵
MH-26	sarcotrine F		<i>Sarcotragus</i> sp. ³⁰
MH-27	isosarcotrine F		<i>Sarcotragus</i> sp. ³⁰
MH-28	idiadione		<i>Cadlina marginata</i> ²⁶ <i>Spongia idia</i> ²⁶
MH-29	unnamed cyclic peroxide ester		<i>Latrunculia</i> sp. ³⁶
MH-30	aikupikoxide A		<i>Diacarnus erythraenus</i> [sic!] ³⁷
MH-31	muqubilone B		<i>Latrunculia</i> sp. ³⁶
MH-32	ircinialactam C		<i>Ircinia</i> sp. CMB-03363 ³⁸
MH-33	ent-ircinialactam C		<i>Psammocinia</i> sp. CMB-03231 ³⁸

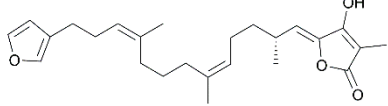
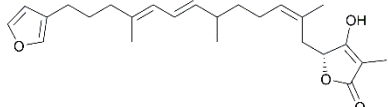
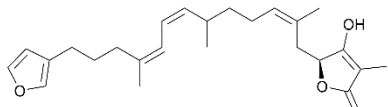
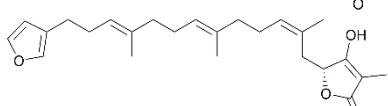
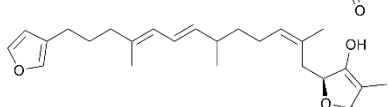
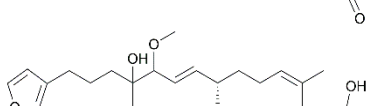
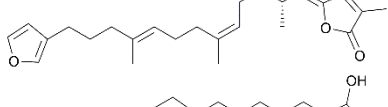
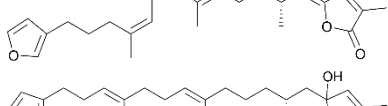
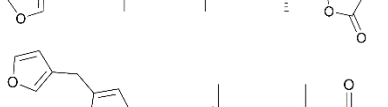
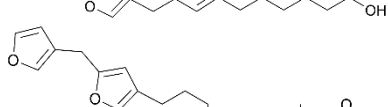
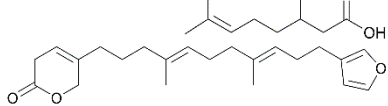
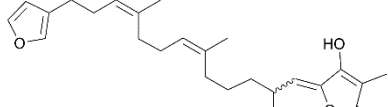
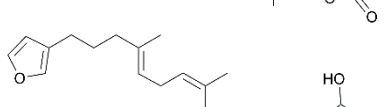
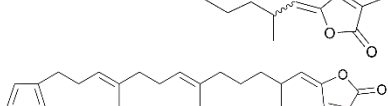
MH-34	ircinialactam D		<i>Ircinia</i> sp. CMB-03363 ³⁸
MH-35	unnamed monoheterocyclic sesterterpene (reported as compound 1)		<i>Ircinia</i> sp. ³⁹
MH-36	unnamed monoheterocyclic sesterterpene (reported as compound 3)		<i>Ircinia</i> sp. ³⁹
MH-37	barangcadoic acid		<i>Hippospongia</i> sp. ⁴⁰
MH-38	rhopaloic acid D		<i>Hippospongia</i> sp. ⁴⁰
MH-39	rhopaloic acid E		<i>Hippospongia</i> sp. ⁴⁰
MH-40	rhopaloic acid F		<i>Hippospongia</i> sp. ⁴⁰
MH-41	rhopaloic acid G		<i>Hippospongia</i> sp. ⁴⁰
MH-42	sarcotragin B		<i>Sarcotragus</i> sp. ⁴¹
MH-43	furospongins-3		<i>Spongia officinalis</i> ⁴²
MH-44	furospongins-4		<i>Spongia officinalis</i> ⁴²
MH-45	unnamed monoheterocyclic sesterterpene (reported as compound 2)		<i>Thorecta horridus</i> ¹⁹
MH-46	sarcotin I		<i>Sarcotragus</i> sp. ⁴³
MH-47	sarcotin J		<i>Sarcotragus</i> sp. ⁴³
MH-48	sigmosceptrillin D methyl ester		<i>Sigmosceptrella</i> sp. ⁴⁴
MH-49	sigmosceptrillin E methyl ester		<i>Sigmosceptrella</i> sp. ⁴⁴

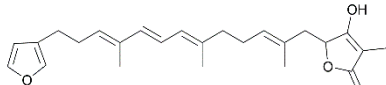
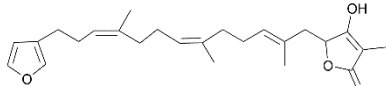
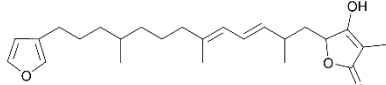
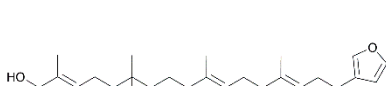
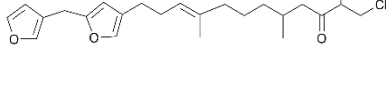
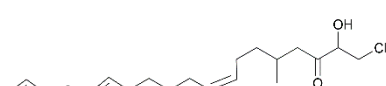
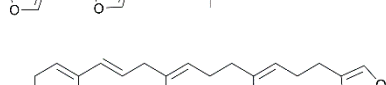
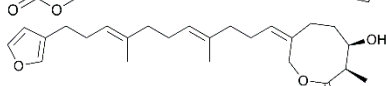
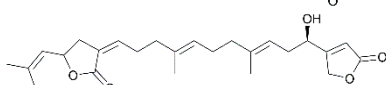
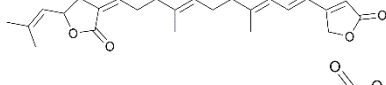
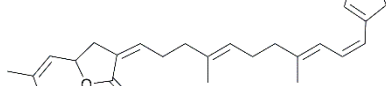
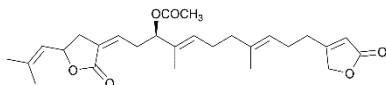
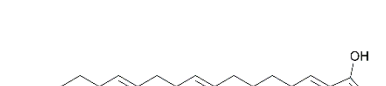
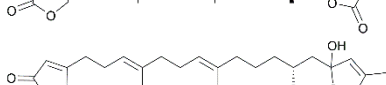
MH-50	coleifolide A		<i>Scutellaria coleifolia</i> ⁴⁵
MH-51	coleifolide B		<i>Scutellaria coleifolia</i> ⁴⁵
MH-52	granuloside		<i>Charcotia granulosa</i> ⁴⁶
MH-53	cacolic acid A		<i>Cacospongia</i> sp. CMB-03404 ⁴⁷
MH-54	sarcotragin C		<i>Sarcotragus</i> sp. ⁴⁸
MH-55	ircinialactam G		<i>Sarcotragus</i> sp. and <i>Psammocinia</i> sp. ⁴⁹
MH-56	8-hydroxy ircinialactam G		<i>Sarcotragus</i> sp. and <i>Psammocinia</i> sp. ⁴⁹
MH-57	dactylospene A		<i>Dactylospongia elegans</i> ⁵⁰
MH-58	furospinosulin B		<i>Dactylospongia elegans</i> ⁵⁰
MH-59	fasciospongide C		<i>Fasciospongia</i> sp. ⁵¹
MH-60	hippotulosa A		<i>Hippospongia fistulosa</i> ⁵²
MH-61	hippotulosa B		<i>Hippospongia fistulosa</i> ⁵²
MH-62	hippotulosa C		<i>Hippospongia fistulosa</i> ⁵²
MH-63	hippotulosa D		<i>Hippospongia fistulosa</i> ⁵²

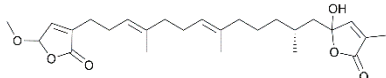
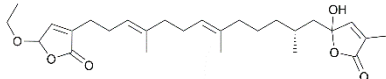
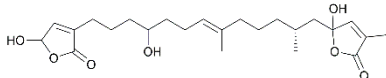
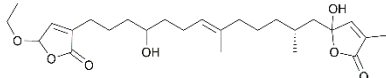
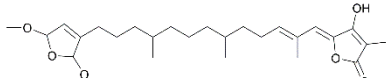
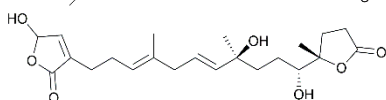
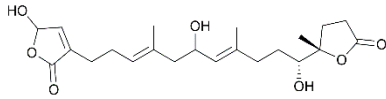
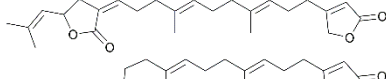
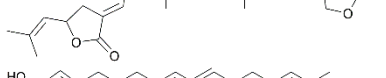
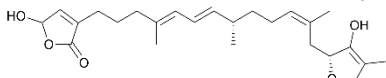
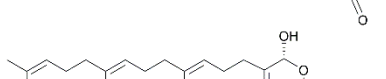
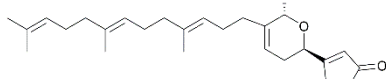
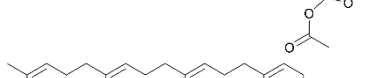
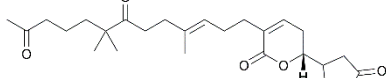
Section 1.3. Structures of linear diheterocyclic (DH) sesterterpenoids			
c. n.	IUPAC name or Trivial name	Structure	Organism as reported in the reference
DH-1	furospongins-1		<i>Spongia officinalis</i> ⁵³ <i>Hippospongia communis</i> ⁵³
DH-2	anhydrofurospongins-1		<i>Spongia officinalis</i> ⁵³ <i>Hippospongia communis</i> ⁵³
DH-3	palinurin		<i>Ircinia variabilis</i> ⁵⁴
DH-4	isopalnurin		<i>Psammocinia</i> sp. ^{55, 56}

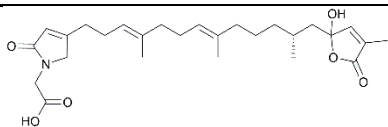
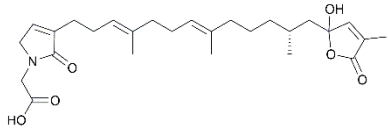
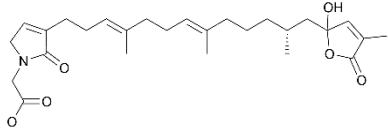
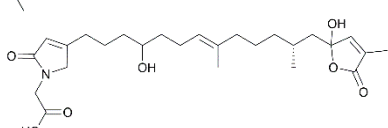
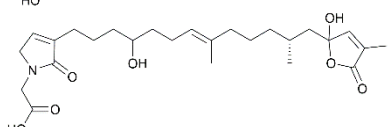
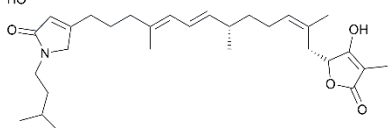
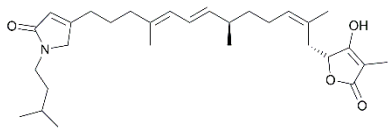
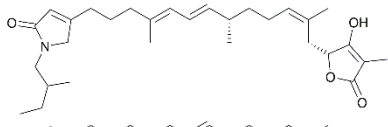
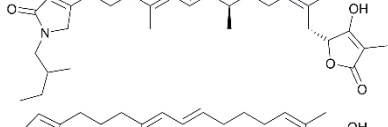
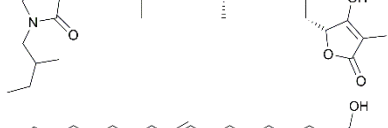
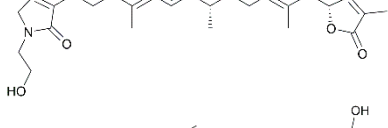
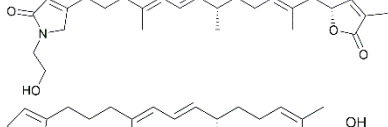
DH-5	palinurin sulfate ester		<i>Ircinia variabilis</i> ⁵⁷
DH-6	okinonellin B		<i>Spongionella</i> sp. ⁵⁸
DH-7	fasciculatin		<i>Ircinia fasciculata</i> ⁵⁹ <i>Ircinia variabilis</i> ⁵⁴
DH-8	fasciculatin sulfate ester		<i>Ircinia fasciculata</i> ⁵⁷
DH-9	irciformonin A* (revised as TH-19)		<i>Ircinia formosana</i> ⁶⁰
DH-10	irciformonin B		<i>Ircinia formosana</i> ⁶⁰
DH-11	15-acetyl-irciformonin B		<i>Ircinia</i> sp. ⁶¹
DH-12	10-acetyl-irciformonin B		<i>Ircinia</i> sp. ⁶¹
DH-13	irciformonin I		<i>Ircinia formosana</i> ⁶⁰
DH-14	5-(13-(3-furanyl)-2,6,10-trimethyl-6,8-tridecadienyl)-4-hydroxy-3-methyl-2(5H)-furanone		<i>Ircinia</i> sp. ⁶²
DH-15	5-(13-(3-furanyl)-2,6,10-trimethyl-2,6,8-tridecatrienyl)-4-hydroxy-3-methyl-2(5H)-furanone		<i>Psammocinia rugosa</i> ⁶³
DH-16	variabilin		<i>Fasciospongia</i> sp. ⁶⁴ <i>Ircinia</i> sp. ⁶⁵ <i>Ircinia dendroides</i> ⁶⁶ <i>Ircinia strobilina</i> ⁶⁷ <i>Ircinia variabilis</i> ⁶⁸ <i>Sarcotragus</i> sp. ²⁴ <i>Semitaspongia bactriana</i> ⁶⁹
DH-17	5-oxo-7E,12E,20Z-variabilin		<i>Ircinia</i> sp. ⁷⁰
DH-18	5-hydroxy-7E,12E,20Z-variabilin		<i>Ircinia</i> sp. ⁷⁰
DH-19	5-oxo-8(10)E,12E,20Z-variabilin		<i>Ircinia</i> sp. ⁷⁰

DH-20	5-oxo-8(10) <i>E</i> ,13,20 <i>Z</i> -variabilin		<i>Ircinia</i> sp. ⁷⁰
DH-21	8-hydroxy-12 <i>E</i> ,20 <i>Z</i> -variabilin		<i>Ircinia</i> sp. ⁷⁰ <i>Sarcotragus</i> sp. ⁷⁰
DH-22	25-hydroxy-variabilin		<i>Fasciospongia</i> sp., <i>Hypselodoris capensis</i> ⁷¹
DH-23	(18 <i>R</i>)-variabilin sulfate ester		<i>Ircinia variabilis</i> ⁷²
DH-24	(13 <i>Z</i> ,18 <i>R</i>)-variabilin sulfate ester		<i>Ircinia variabilis</i> ⁷²
DH-25	(12 <i>E</i> ,18 <i>S</i> ,20 <i>Z</i>)-8-hydroxyvariabilin		<i>Ircinia</i> sp. CMB-01064 ³⁸ <i>Ircinia</i> sp. CMB-03363 ³⁸
DH-26	(7 <i>E</i> ,12 <i>E</i> ,18 <i>S</i> ,20 <i>Z</i>)-variabilin		<i>Ircinia</i> sp. CMB-01064 ³⁸ <i>Ircinia</i> sp. CMB-03363 ³⁸ <i>Sarcotragus</i> sp. ²⁴
DH-27	(7 <i>E</i> ,12 <i>Z</i> ,18 <i>S</i> ,20 <i>Z</i>)-variabilin		<i>Ircinia</i> sp. CMB-01064 ³⁸ <i>Ircinia</i> sp. CMB-03363 ³⁸ <i>Ircinia oros</i> ⁷³
DH-28	(7 <i>E</i> ,12 <i>E</i> ,18 <i>R</i> ,20 <i>Z</i>)-variabilin		<i>Psammocinia</i> sp. ⁵⁶ <i>Sarcotragus</i> sp. ²⁴ <i>Ircinia dendroides</i> ⁶⁶
DH-29	variabilin fatty acid esters		<i>Ircinia felix</i> ⁷⁴
DH-30	psammocinin A ₁		<i>Psammocinia</i> sp. ⁵⁶
DH-31	psammocinin A ₂		<i>Psammocinia</i> sp. ⁵⁶
DH-32	strobilin		<i>Ircinia strobilina</i> ⁶⁷ <i>Ircinia oros</i> ⁷³
DH-33	(8 <i>Z</i> ,13 <i>Z</i> ,20 <i>Z</i>)-strobilin		<i>Ircinia felix</i> , <i>Ircinia strobilina</i> , <i>Ircinia campana</i> ⁶⁷ <i>Ircinia oros</i> ⁷³
DH-34	(8 <i>Z</i> ,13 <i>Z</i> ,18 <i>S</i> ,20 <i>Z</i>)-strobilin		<i>Ircinia oros</i> ⁷³

DH-35	(7Z,13Z,20Z)-felixinin		<i>Ircinia felix</i> , <i>Ircinia strobilina</i> , <i>Ircinia campana</i> ⁷⁵
DH-36	sarcotin A		<i>Sarcotragus</i> sp. ⁷⁶
DH-37	sarcotin B		<i>Sarcotragus</i> sp. ⁷⁶
DH-38	sarcotin C		<i>Sarcotragus</i> sp. ⁷⁶
DH-39	<i>epi</i> -sarcotin A		<i>Sarcotragus</i> sp. ⁴³
DH-40	sarcotin M		<i>Sarcotragus</i> sp. ⁴³
DH-41	(8E,13Z,18R,20Z)-strobilin		<i>Ircinia</i> sp. ⁷⁷
DH-42	(8Z,13E,18R,20Z)-strobilin		<i>Ircinia</i> sp. ⁷⁷
DH-43	cacolide A		<i>Cacospongia</i> sp. CMB-03404 ⁴⁷
DH-44	ircinin-3		<i>Spongia officinalis</i> ⁴²
DH-45	ircinin-4		<i>Spongia officinalis</i> ⁴²
DH-46	furodendin		<i>Phyllospongia dendyi</i> ⁷⁸
DH-47	unnamed diheterocyclic sesterterpene (reported as compound 6a)		<i>Ircinia dendroides</i> ⁶⁶
DH-48	unnamed diheterocyclic sesterterpene (reported as compound 8a)		<i>Ircinia dendroides</i> ⁶⁶
DH-49	22-deoxyvariabilin		<i>Thorecta</i> sp. ²⁹ <i>Hypselodoris capensis</i> and <i>Fasciospongia</i> sp. ⁷¹

DH-50	ircinic acid		<i>Ircinia</i> sp. ⁷⁹
DH-51	palomin		<i>Ircinia</i> sp. ⁸⁰
DH-52	unnamed diheterocyclic sesterterpene (reported as compound 5)		<i>Psammocinia</i> sp. ⁸¹
DH-53	unnamed diheterocyclic sesterterpene (reported as epoxide 12)		<i>Spongia</i> sp. ⁸²
DH-54	unnamed diheterocyclic sesterterpene (reported as compound 1)		<i>Ircinia oros</i> ⁸³
DH-55	unnamed diheterocyclic sesterterpene (reported as compound 3)		<i>Ircinia oros</i> ⁸³
DH-56	dehydrofurodendin		<i>Lendenfeldia</i> sp. ⁸⁴
DH-57	astakolactin		<i>Cacospongia scalaris</i> ⁸⁵
DH-58	luffarin R		<i>Luffariella geometrica</i> ²⁰ <i>Thorectandra</i> sp. ⁸⁶
DH-59	luffarin T		<i>Luffariella geometrica</i> ²⁰
DH-60	luffarin U		<i>Luffariella geometrica</i> ²⁰
DH-61	unnamed diheterocyclic sesterterpene (reported as compound 1a)		<i>Fasciospongia cavernosa</i> ⁸⁷
DH-62	ircinialactone A		<i>Sarcotragus</i> sp. ⁴⁹
DH-63	cacolide B		<i>Cacospongia</i> sp. CMB-03404 ⁴⁷
DH-64	cacolide C		<i>Cacospongia</i> sp. CMB-03404 ⁴⁷

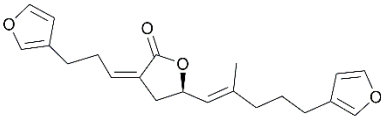
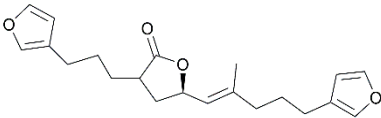
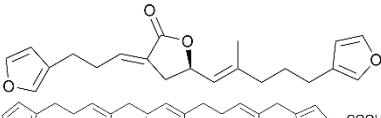
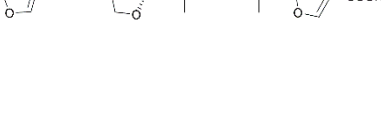
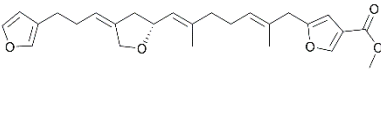
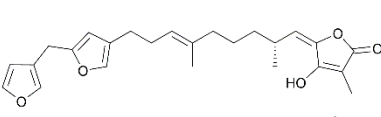
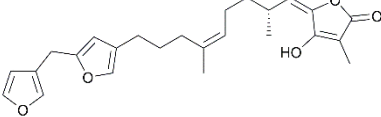
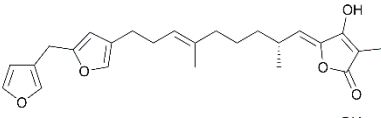
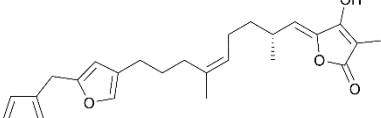
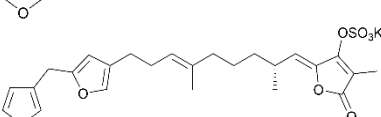
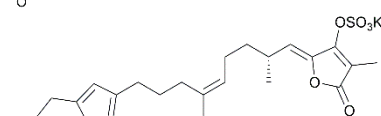
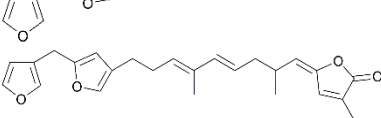
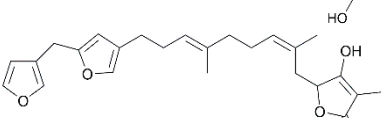
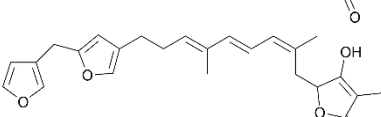
DH-65	cacolide D		<i>Cacospongia</i> sp. CMB-03404 ⁴⁷
DH-66	cacolide E		<i>Cacospongia</i> sp. CMB-03404 ⁴⁷
DH-67	cacolide F		<i>Cacospongia</i> sp. CMB-03404 ⁴⁷
DH-68	cacolide G		<i>Cacospongia</i> sp. CMB-03404 ⁴⁷
DH-69	psammocinin B		<i>Psammocinia</i> sp. ⁵⁶
DH-70	irciformonin C		<i>Ircinia formosana</i> ⁶⁰
DH-71	irciformonin D		<i>Ircinia formosana</i> ⁶⁰
DH-72	(6Z)-luffarin-V		<i>Fasciospongia cavernosa</i> ⁸⁸
DH-73	luffarin V		<i>Thorectandra</i> sp. ⁸⁶ <i>Luffariella geometrica</i> ²⁰
DH-74	sarcotin F		<i>Sarcotragus</i> sp. ⁴³
DH-75	epi-sarcotin F		<i>Sarcotragus</i> sp. ³⁰
DH-76	thorectolide		<i>Hyrtios</i> sp. ⁸⁹
DH-77	thorectolide monoacetate		<i>Thorectandra excavatus</i> ⁹⁰ <i>Hyrtios</i> sp. ⁸⁹
DH-78	cacospongionolide D		<i>Fasciospongia cavernosa</i> ⁸⁸
DH-79	erectusolide		<i>Hyrtios</i> sp. ⁹¹
DH-80	hippolide A		<i>Hippospongia lachne</i> ²³
DH-81	hippolide B		<i>Hippospongia lachne</i> ²³

DH-82	cacolide H		<i>Cacospongia</i> sp. CMB-03404 ⁴⁷
DH-83	cacolide I		<i>Cacospongia</i> sp. CMB-03404 ⁴⁷
DH-84	cacolide J		<i>Cacospongia</i> sp. CMB-03404 ⁴⁷
DH-85	cacolide K		<i>Cacospongia</i> sp. CMB-03404 ⁴⁷
DH-86	cacolide L		<i>Cacospongia</i> sp. CMB-03404 ⁴⁷
DH-87	sarcotrine A		<i>Sarcotragus</i> sp. ⁴³
DH-88	<i>epi</i> -sarcotrine A		<i>Sarcotragus</i> sp. ⁴³
DH-89	sarcotrine C		<i>Sarcotragus</i> sp. ⁴³
DH-90	<i>epi</i> -sarcotrine C		<i>Sarcotragus</i> sp. ⁴³
DH-91	sarcotrine D		<i>Sarcotragus</i> sp. ⁴³
DH-92	palinurine A		<i>Cunninghamella</i> sp. ⁹²
DH-93	palinurine B		<i>Cunninghamella</i> sp. ⁹²
DH-94	sarcotrine E		<i>Sarcotragus</i> sp. ³⁰

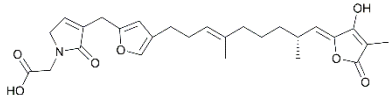
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DH-96	ircinialactam A		<i>Ircinia</i> sp. CMB-01064 ³⁸ <i>Ircinia</i> sp. CMB-03363 ³⁸ <i>Ircinia</i> sp. ³⁹
DH-97	8-hydroxyircinialactam A		<i>Ircinia</i> sp. CMB-01064 ³⁸
DH-98	8-hydroxyircinialactam B		<i>Ircinia</i> sp. CMB-01064 ³⁸ <i>Ircinia</i> sp. CMB-03363 ³⁸
DH-99	unnamed diheterocyclic sesterterpene (reported as compound 1)		<i>Hippospongia lachne</i> ⁹
DH-100	dihydrofurospongins-2		<i>Phyllospongia foliascens</i> ⁹³ <i>Spongia officinalis</i> ⁵³
DH-101	fasciospongide B		<i>Fasciospongia</i> sp. ⁵¹
DH-102	irciformonin F		<i>Ircinia formosana</i> ⁹⁴ <i>Ircinia</i> sp. ⁶¹
DH-103	irciformonin G		<i>Ircinia formosana</i> ⁹⁴
DH-104	irciformonin H		<i>Ircinia formosana</i> ⁹⁴
DH-105	irciformonin J		<i>Ircinia formosana</i> ⁹⁴
DH-106	luffarin S		<i>Luffariella geometrica</i> ²⁰
DH-107	(18 <i>R</i>)-variabilin		<i>Hypselodoris capensis</i> and <i>Fasciospongia</i> sp. ⁷¹
DH-108	(7 <i>E</i> ,13 <i>Z</i> ,18 <i>S</i> ,20 <i>Z</i>)-variabilin		<i>Sarcotragus</i> sp. ²⁴ <i>Ircinia oros</i> ⁷³
DH-109	22-deoxy-23-hydroxymethylvariabilin		<i>Hypselodoris capensis</i> and <i>Fasciospongia</i> sp. ⁷¹

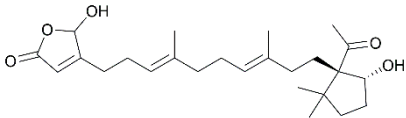
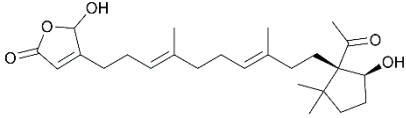
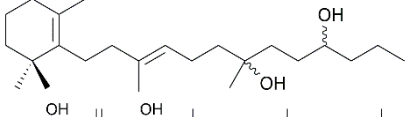
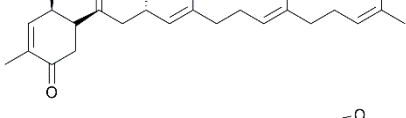
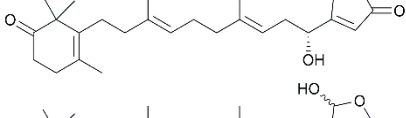
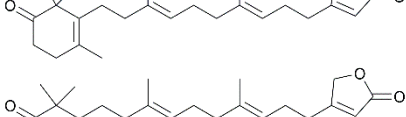
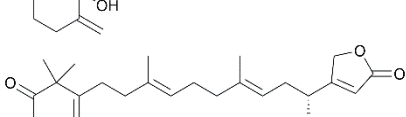
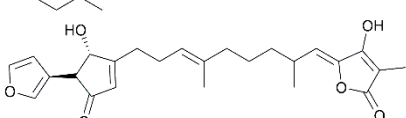
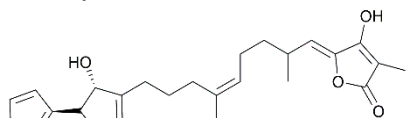
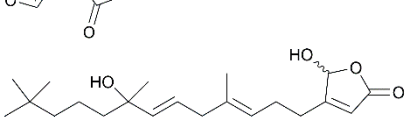
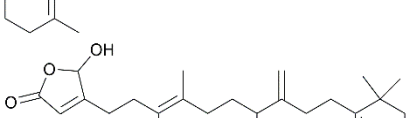
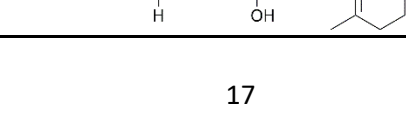
Section 1.4. Structures of linear triheterocyclic (TH) sesterterpenoids and related dimers

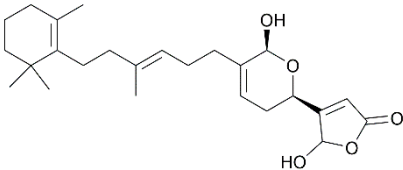
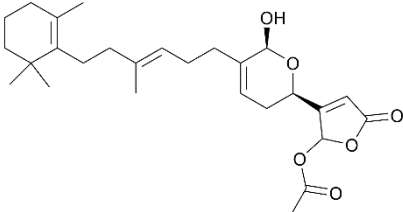
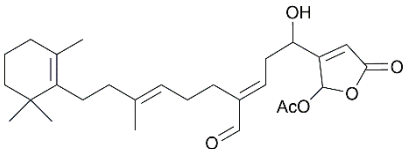
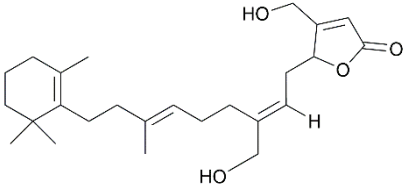
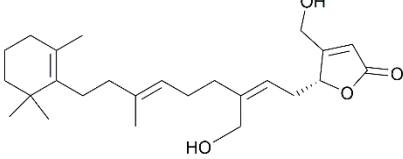
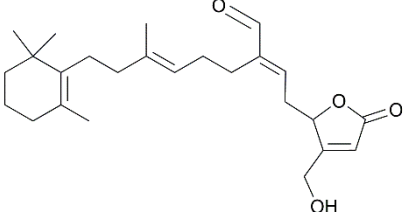
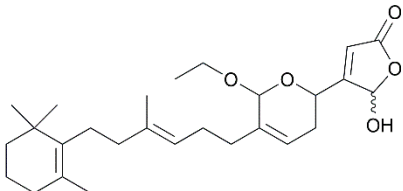
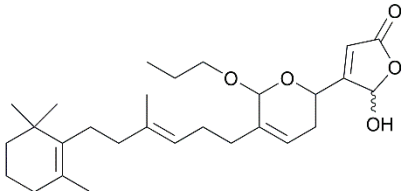
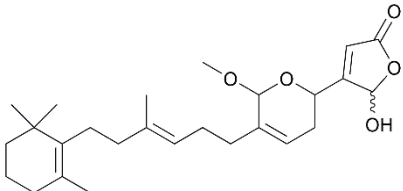
c. n.	IUPAC name or Trivial name	Structure	Organism as reported in the reference
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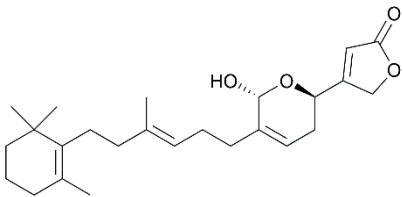
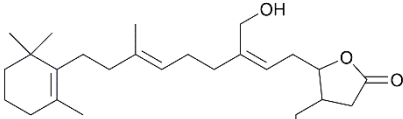
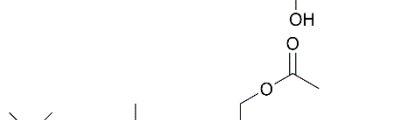
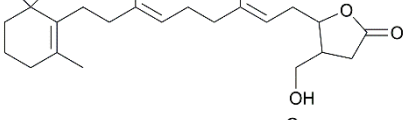
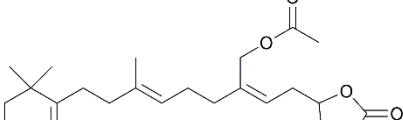
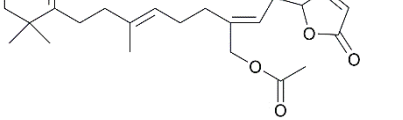
TH-1	nitenin		<i>Spongia nitens</i> ⁹⁵
TH-2	dihydronitenin		<i>Spongia nitens</i> ⁹⁵
TH-3	isonitenin		<i>Spongia officinalis</i> ⁹⁶
TH-4	unnamed triheterocyclic sesterterpene (reported as compound 1)		<i>Luffariella variabilis</i> ⁹⁷
TH-5	unnamed triheterocyclic sesterterpene (reported as compound 3)		<i>Luffariella variabilis</i> ⁹⁷
TH-6	sarcotin D		<i>Sarcotragus</i> sp. ⁷⁶
TH-7	sarcotin E		<i>Sarcotragus</i> sp. ⁷⁶
TH-8	ircinin-1		<i>Ircinia oros</i> ^{98, 99} <i>Sarcotragus</i> sp. ⁷⁶ <i>Psammocinia</i> sp. ¹⁰⁰
TH-9	ircinin-2		<i>Ircinia oros</i> ⁹⁸ <i>Sarcotragus</i> sp. ⁷⁶ <i>Psammocinia</i> sp. ¹⁰⁰
TH-10	ircinin-1 sulfate ester		<i>Ircinia oros</i> ⁷²
TH-11	ircinin-2 sulfate ester		<i>Ircinia oros</i> ⁷²
TH-12	24-hydroxyircinolide		<i>Thorecta marginalis</i> ²
TH-13	spongionellin		<i>Spongionella</i> sp. ¹⁰¹
TH-14	dehydro-spongionellin		<i>Spongionella</i> sp. ¹⁰¹

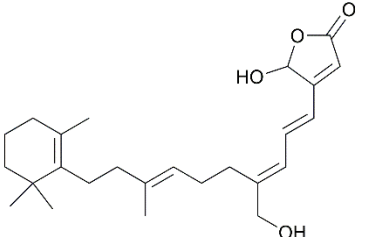
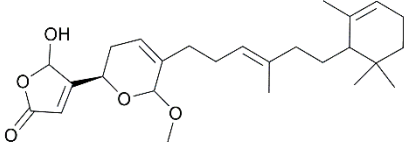
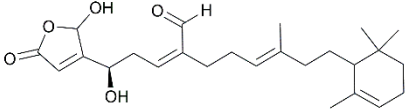
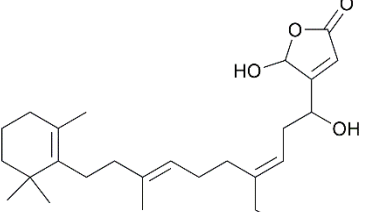
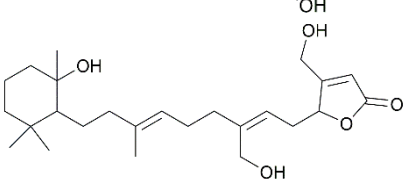
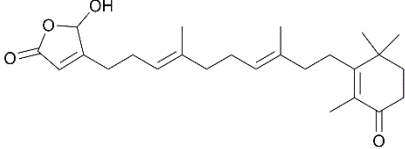
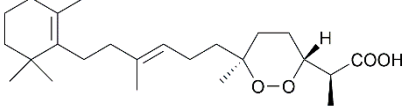
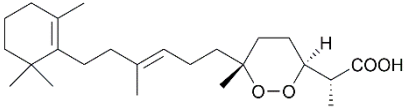
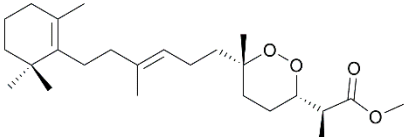
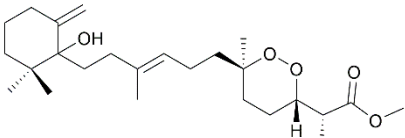
TH-15	cometin A		<i>Spongia</i> sp. ¹⁰²
TH-16	cometin B		<i>Spongia</i> sp. ¹⁰²
TH-17	cometin C		<i>Spongia</i> sp. ¹⁰²
TH-18	ircinolide		<i>Thorecta marginalis</i> ²
TH-19	irciformonin A (revised structure of DH-19)		<i>Ircinia formosana</i> ^{60, 94}
TH-20	irciformonin E		<i>Ircinia formosana</i> ⁹⁴
TH-21	irciformonin K		<i>Ircinia formosana</i> ⁹⁴
TH-22	sarcotin G		<i>Sarcotragus</i> sp. ⁴³
TH-23	sarcotin H		<i>Sarcotragus</i> sp. ⁴³
TH-24	ircinialactam F		<i>Ircinia oros</i> ⁹⁹
TH-25	sulawesin C		<i>Psammocinia</i> sp. ¹⁰⁰
TH-26	hippospongina A		<i>Hippospongia</i> sp. ¹⁰³
TH-27	hippospongina B		<i>Hippospongia</i> sp. ¹⁰³
TH-28	hippospongina C		<i>Hippospongia</i> sp. ¹⁰³
TH-29	hippospongina D		<i>Hippospongia</i> sp. ¹⁰³
TH-30	hippospongina E		<i>Hippospongia</i> sp. ¹⁰³
TH-31	hippospongina F		<i>Hippospongia</i> sp. ¹⁰³

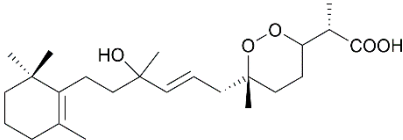
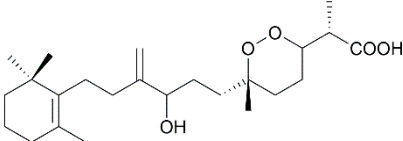
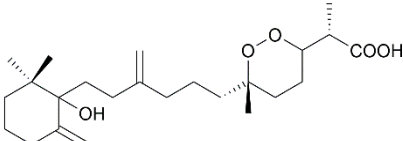
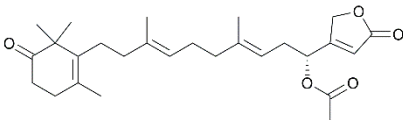
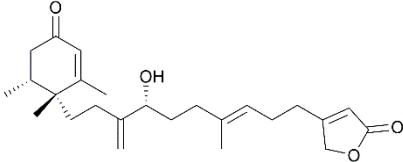
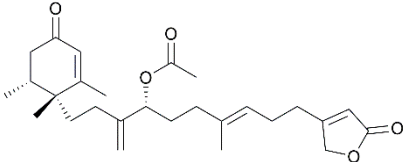
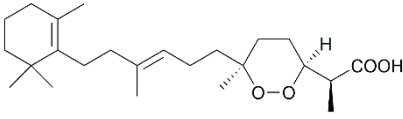
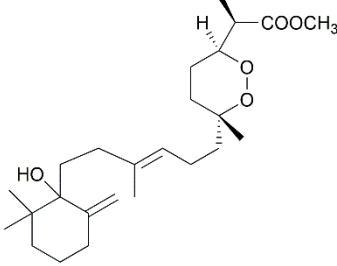
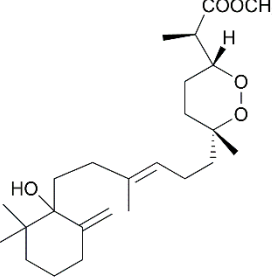
TH-32	(12 <i>E</i> ,18 <i>R</i> ,20 <i>Z</i>)- ircinialactam E		<i>Ircinia oros</i> ⁹⁹
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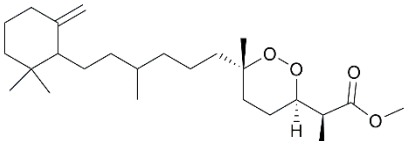
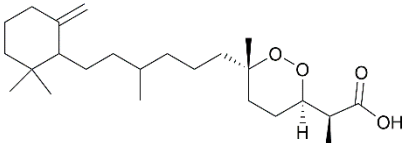
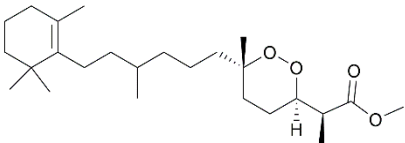
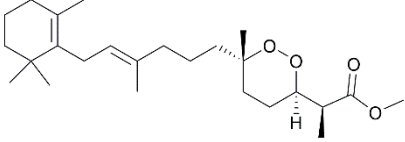
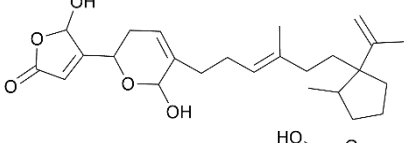
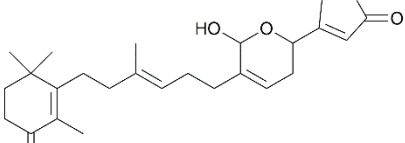
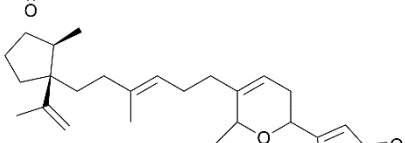
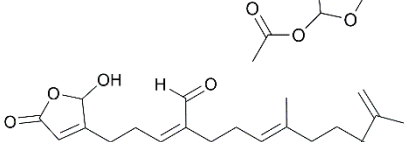
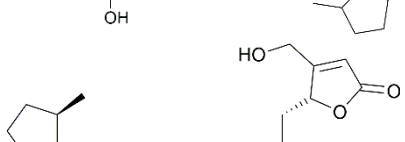
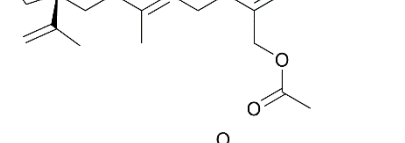
Section 1.5. Structures of monocarbocyclic (MC) sesterterpenoids (with or without hetherocycles)			
c. n.	IUPAC name or Trivial name	Structure	Organism as reported in the reference
MC-1	acantholide E		<i>Acanthodendrilla</i> sp. ²¹
MC-2	acantholide D		<i>Acanthodendrilla</i> sp. ²¹
MC-3	diacardiol A		<i>Diacarnus megaspinorhabdosa</i> [sic!] ¹⁰⁴
MC-4	phorbin A		<i>Monanchora</i> sp. ¹⁰⁵
MC-5	thorectidaelide A		<i>Hyrtios communis</i> ²²
MC-6	thorectidaelide B		<i>Hyrtios communis</i> ²²
MC-7	thorectidaelide D		<i>Hyrtios communis</i> ²²
MC-8	thorectidaelide E		<i>Hyrtios communis</i> ²²
MC-9	sulawesin A		<i>Psammocinia</i> sp. ¹⁰⁰
MC-10	sulawesin B		<i>Psammocinia</i> sp. ¹⁰⁰
MC-11	thorectidaelide C		<i>Hyrtios communis</i> ²²
MC-12	acantholide A		<i>Hyrtios communis</i> ²²

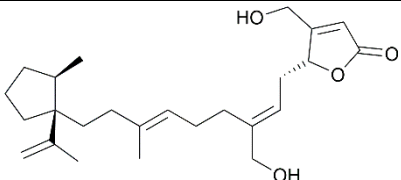
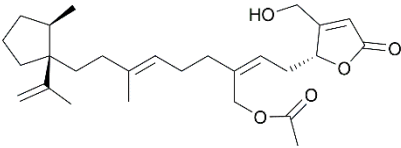
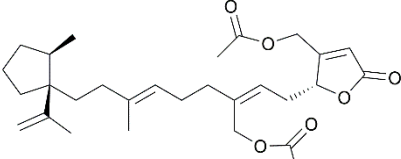
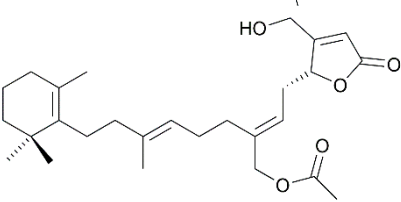
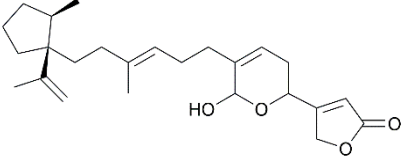
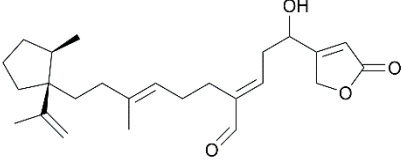
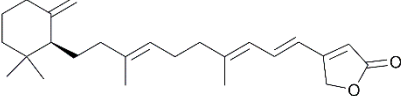
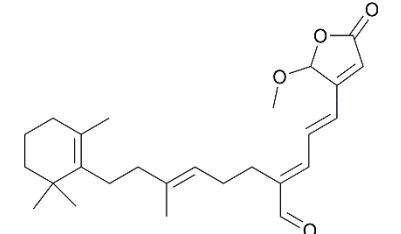
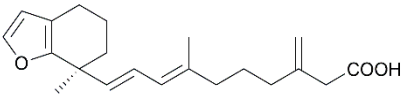
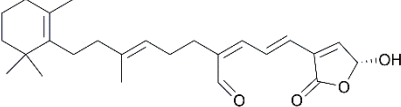
MC-13	manoalide		<i>Luffariella variabilis</i> ¹⁰⁶⁻¹⁰⁹ <i>Luffariella</i> cf. <i>variabilis</i> ¹¹⁰
MC-14	manoalide 25-monoacetate		<i>Thorectandra excavatus</i> ⁹⁰ <i>Brachiaster</i> sp. ¹¹¹ <i>Luffariella variabilis</i> ^{108, 112} <i>Luffariella</i> cf. <i>variabilis</i> ¹¹⁰
MC-15	25-acetoxyseco-manoalide		<i>Luffariella variabilis</i> ¹⁰⁸
MC-16	(6E)-neomanoalide		<i>Luffariella variabilis</i> ¹⁰⁶ <i>Hippospongia lachne</i> ²³
MC-17	(6Z)-neomanoalide		<i>Hippospongia lachne</i> ²³
MC-18	E-neomanoalide-24-al		<i>Luffariella</i> sp. ¹¹³
MC-19	24-O-ethylmanoalide		<i>Luffariella</i> cf. <i>variabilis</i> ¹¹⁰
MC-20	24-O-propylmanoalide		<i>Luffariella</i> sp. ¹¹⁴
MC-21	24-O-methylmanoalide		<i>Luffariella</i> cf. <i>variabilis</i> ¹¹⁰

MC-22	deoxymanoalide		<i>Chromodoris willani</i> ¹¹⁵
MC-23	Z-2,3-dihydroneomanoalide		<i>Luffariella</i> sp. ¹¹³
MC-24	Z-24-acetoxy-2,3-dihydroneomanoalide		<i>Luffariella</i> sp. ¹¹³
MC-25	Z-24-acetoxynemanoalide		<i>Luffariella</i> sp. ¹¹³
MC-26	6E-neomanoalide-24,25-diacetate		<i>Brachiaster</i> sp. ¹¹¹ <i>Luffariella</i> sp. ¹¹⁶
MC-27	6Z-neomanoalide-24,25-diacetate		<i>Brachiaster</i> sp. ¹¹¹
MC-28	secomanoalide		<i>Luffariella variabilis</i> ^{106, 108, 109} <i>Luffariella</i> cf. <i>variabilis</i> ¹¹⁰
MC-29	deoxysecomanoalide		<i>Chromodoris willani</i> ¹¹⁵

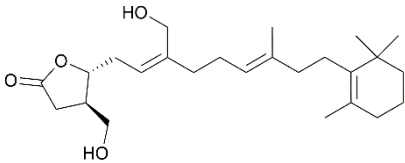
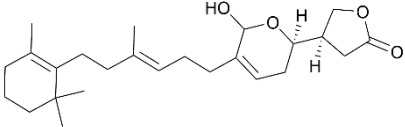
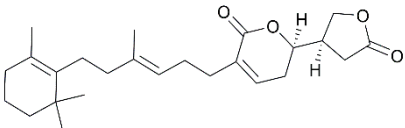
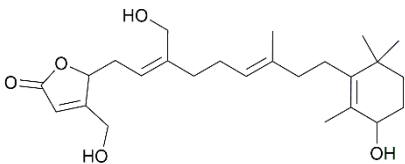
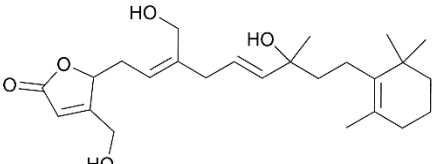
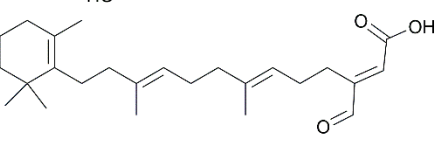
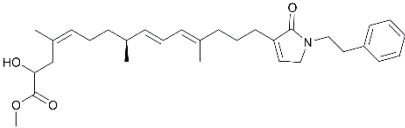
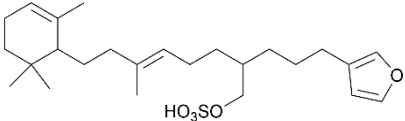
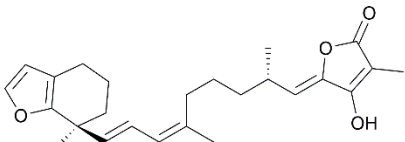
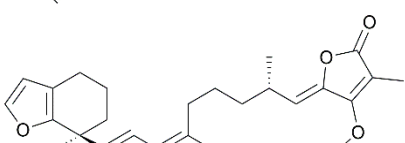
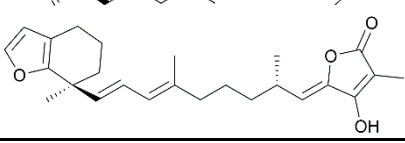
MC-30	aplysinoplide A		<i>Aplysinopsis digitata</i> ¹¹⁷
MC-31	luffariolide H		<i>Luffariella</i> sp. ¹¹⁶
MC-32	luffariolide J		<i>Luffariella</i> sp. ¹¹⁶
MC-33	aplysinoplide B		<i>Aplysinopsis digitata</i> ¹¹⁷
MC-34	aplysinoplide C		<i>Aplysinopsis digitata</i> ¹¹⁷
MC-35	acantholide B		<i>Hyrtios communis</i> ²²
MC-36	(+)-muquibilin (= prianicin A)		<i>Prianos</i> sp. ¹¹⁸⁻¹²⁰ Unidentified sponge ¹²¹ <i>Diacarnus spinopoculum</i> ¹²² <i>Diacarnus erythraenus</i> [sic!] ³⁷ <i>Diacarnus erythraeanus</i> ¹²³ <i>Diacarnus megaspinorhabdosa</i> [sic!] ¹⁰⁴ Synthesis ¹²⁴
MC-37	(-)-muquibilin ((-)-muquibilin A)		<i>Diacarnus spinopoculum</i> ¹²²
MC-38	muquibilin ester		<i>Prianos</i> sp. ¹¹⁸⁻¹²⁰
MC-39	hurghaperoxide		<i>Haplosclerida</i> sp. ¹²⁵

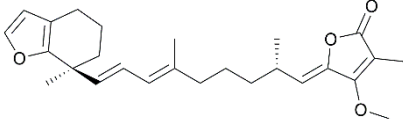
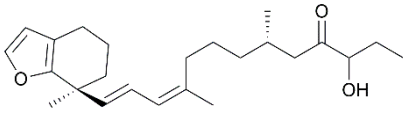
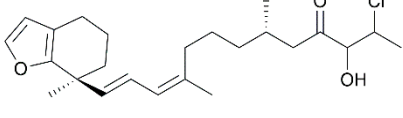
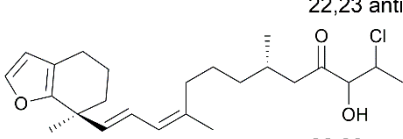
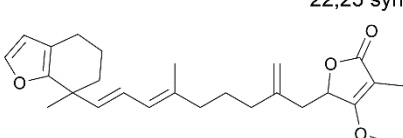
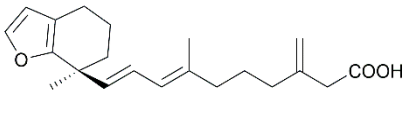
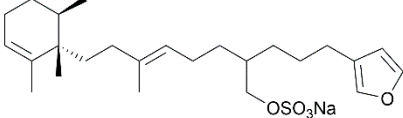
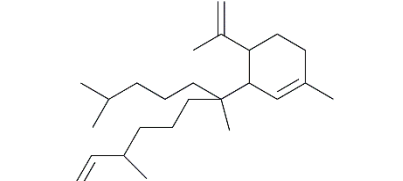
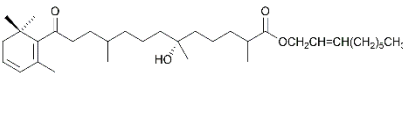
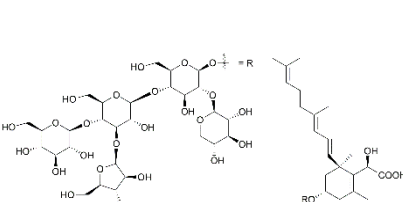
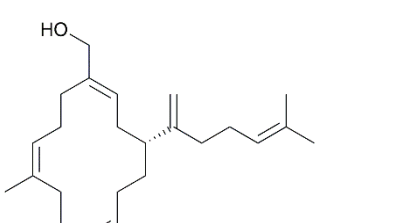
MC-40	tasnemoxide A		<i>Diacarnus erythraenus</i> [sic!] ¹²⁶
MC-41	tasnemoxide B		<i>Diacarnus erythraenus</i> [sic!] ¹²⁶
MC-42	tasnemoxide C		<i>Diacarnus erythraenus</i> [sic!] ¹²⁶
MC-43	4-acetoxythorectidaelide A		<i>Hyrtios communis</i> ²²
MC-44	cyclolinteinol		<i>Cacospongia</i> cf. <i>linteiformis</i> ¹²⁷
MC-45	cyclolinteinol acetate		<i>Cacospongia</i> cf. <i>linteiformis</i> ¹²⁷
MC-46	<i>epi</i> -muqubilin A (dubious configurational assignment)		<i>Diacarnus</i> cf. <i>spinopoculum</i> ¹²² <i>Latrunculia</i> sp. ³⁶
MC-47	muqubilin B (dubious configurational assignment)		<i>Diacarnus</i> cf. <i>spinopoculum</i> ¹²²
MC-48	<i>epi</i> -muqubilin B (dubious configurational assignment)		<i>Diacarnus</i> cf. <i>spinopoculum</i> ¹²² <i>Latrunculia</i> sp. ³⁶ <i>Diacarnus megaspinorhabdosa</i> [sic!] ¹⁰⁴

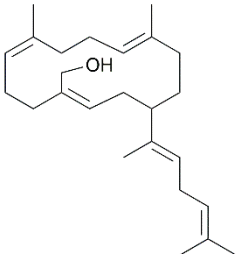
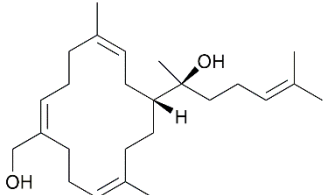
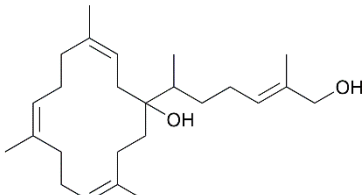
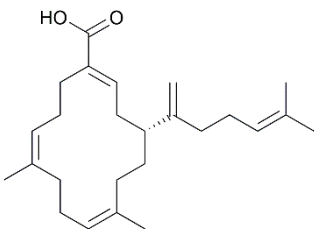
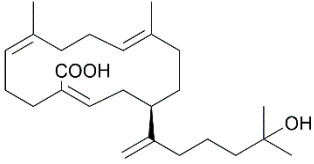
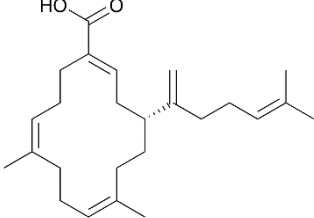
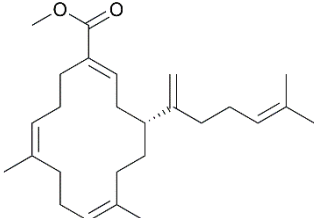
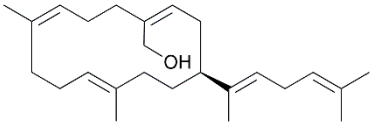
MC-49	diacarnoxide A		<i>Diacarnus levij</i> ¹²⁸
MC-50	diacarnoxide B		<i>Diacarnus levij</i> ¹²⁸
MC-51	diacarnoxide C		<i>Diacarnus levij</i> ¹²⁸
MC-52	diacarnoxide D		<i>Diacarnus levij</i> ¹²⁸
MC-53	luffariellin A		<i>Luffariella variabilis</i> ^{108, 109}
MC-54	fasciospongide A		<i>Fasciospongia</i> sp. ⁵¹
MC-55	25-acetoxyluffariellin A		<i>Luffariella variabilis</i> ¹⁰⁸
MC-56	luffariellin B		<i>Luffariella variabilis</i> ¹⁰⁹
MC-57	luffalide A		<i>Luffariella</i> sp. ¹²⁹
MC-58	luffalide B		<i>Luffariella</i> sp. ¹²⁹

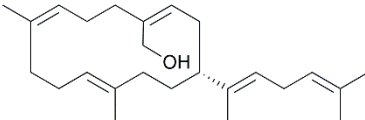
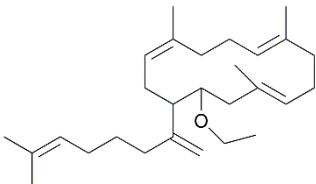
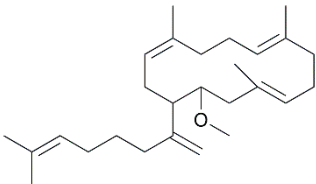
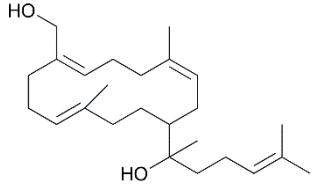
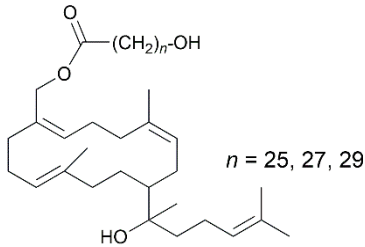
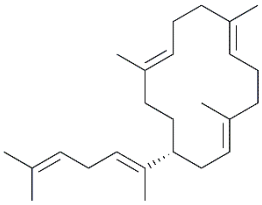
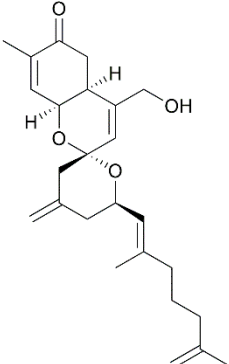
MC-59	luffalide C		<i>Luffariella</i> sp. ¹²⁹
MC-60	luffalide D		<i>Luffariella</i> sp. ¹²⁹
MC-61	luffalide E		<i>Luffariella</i> sp. ¹²⁹
MC-62	luffalide F		<i>Luffariella</i> sp. ¹²⁹
MC-63	luffariellin C		<i>Chromodoris funerea</i> ¹³⁰
MC-64	luffariellin D		<i>Chromodoris funerea</i> ¹³⁰
MC-65	luffarin P		<i>Luffariella geometrica</i> ²⁰
MC-66	(4E,6E)-dehydro-25-O-methylmanoalide		<i>Luffariella variabilis</i> ¹¹²
MC-67	untenic acid		Sponge NSO09 ¹³¹
MC-68	(4E,6E)-dehydromanoalide		<i>Luffariella variabilis</i> ¹³²

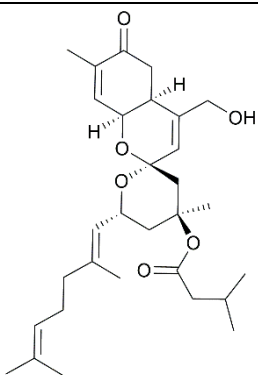
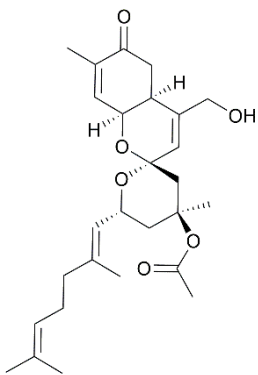
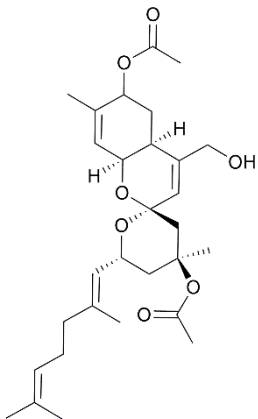
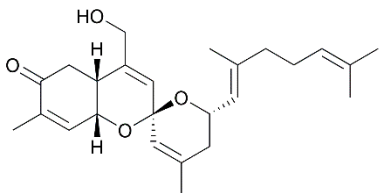
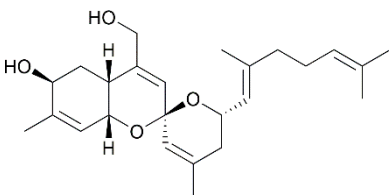
MC-69	25-acetoxyluffariellin B		<i>Luffariella variabilis</i> ¹⁰⁸
MC-70	luffariellolide		<i>Luffariella</i> sp. ¹⁰⁷ <i>Hyrtios communis</i> ²² <i>Fascaplysinopsis</i> sp. ¹³³ <i>Dactylospongia elegans</i> ⁵⁰ <i>Suberea</i> sp. ¹³⁴ <i>Acanthodendrilla</i> sp. ²¹ <i>Cacospongia</i> sp. ¹³⁵ <i>Thorectandra</i> sp. ¹³⁶
MC-71	igernellin		<i>Spongia irregularis</i> ¹³⁷ <i>Igernella</i> sp. ¹³⁸
MC-72	16-hydroxyluffariellolide		<i>Hyrtios communis</i> ²² <i>Thorectandra</i> sp. ¹³⁶
MC-73	dehydroluffariellolide diacid		<i>Fascaplysinopsis reticulata</i> ¹³⁹ <i>Thorectandra</i> sp. ¹³⁶
MC-74	isodehydrouffariellolide		<i>Fascaplysinopsis reticulata</i> ¹³⁹ <i>Hyrtios cf. erecta</i> [sic!] ¹⁴⁰
MC-75	unnamed monocarbocyclic sesterterpene (reported as compound 1)		<i>Hyrtios cf. erecta</i> [sic!] ¹⁴⁰
MC-76	unnamed monocarbocyclic sesterterpene (reported as compound 2)		<i>Hyrtios cf. erecta</i> [sic!] ¹⁴⁰
MC-77	luffariolide A		<i>Luffariella</i> sp. ¹⁴¹
MC-78	luffariolide B		<i>Luffariella</i> sp. ¹⁴¹

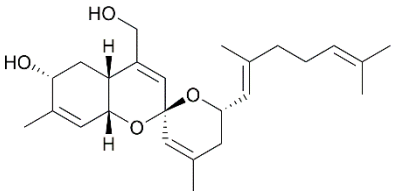
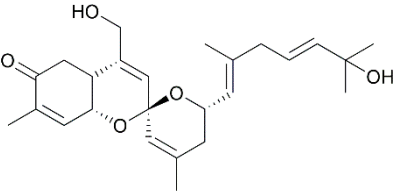
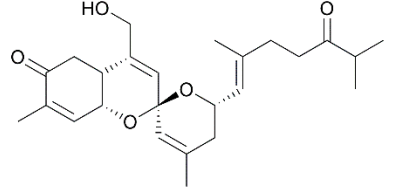
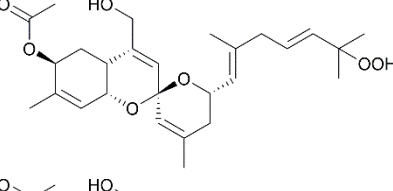
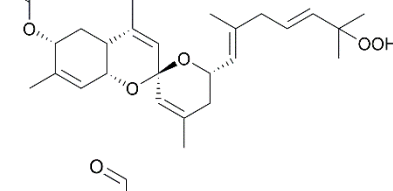
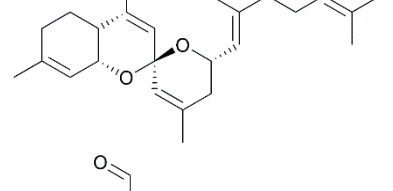
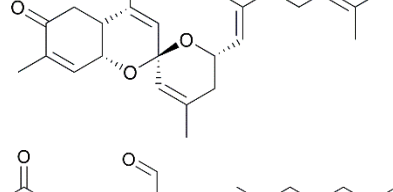
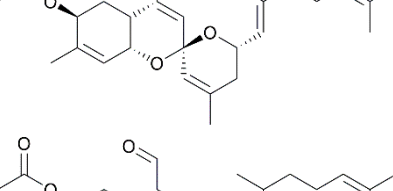
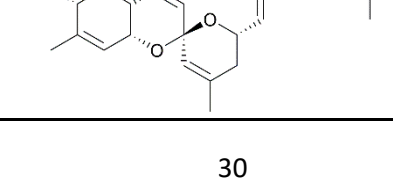
MC-79	luffariolide C		<i>Luffariella</i> sp. ¹⁴¹
MC-80	luffariolide D		<i>Luffariella</i> sp. ¹⁴¹
MC-81	luffariolide E		<i>Luffariella</i> sp. ¹⁴¹
MC-82	luffariolide F		<i>Luffariella</i> sp. ¹⁴²
MC-83	luffariolide G		<i>Luffariella</i> sp. ¹⁴²
MC-84	(2 <i>E</i> ,6 <i>E</i> ,10 <i>E</i>)-3-formyl-7,11-dimethyl-13-(2,6,6-trimethylcyclohex-1-enyl)trideca-2,6,10-trienoic acid		<i>Thorectandra</i> sp. ¹³⁶
MC-85	sarcotragin A		<i>Sarcotragus</i> sp. ⁴¹
MC-86	hipposulfate C		<i>Spongia irregularis</i> ¹³⁷
MC-87	unnamed monocarbocyclic sesterterpene (reported as compound 1)		<i>Ircinia</i> sp. ¹⁴³
MC-88	unnamed monocarbocyclic sesterterpene (reported as compound 2)		<i>Ircinia</i> sp. ¹⁴³
MC-89	unnamed monocarbocyclic sesterterpene (reported as compound 3)		<i>Ircinia</i> sp. ¹⁴³

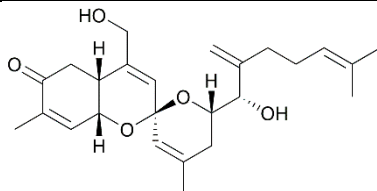
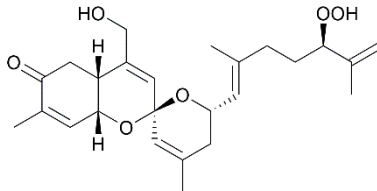
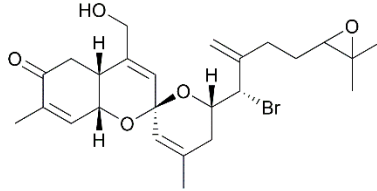
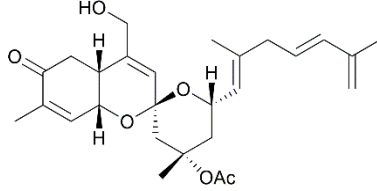
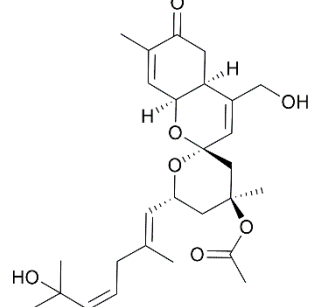
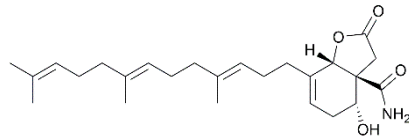
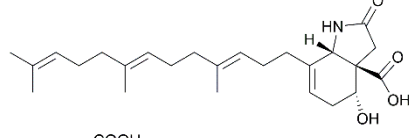
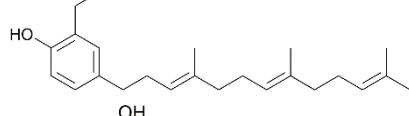
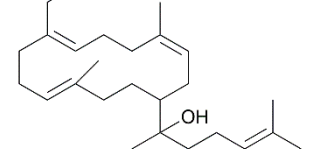
MC-90	unnamed monocarbocyclic sesterterpene (reported as compound 4)		<i>Ircinia</i> sp. ¹⁴³
MC-91	unnamed monocarbocyclic sesterterpene (reported as compound 5)		<i>Ircinia</i> sp. ¹⁴³
MC-92	unnamed monocarbocyclic sesterterpene (reported as compound 6)		<i>Ircinia</i> sp. ¹⁴³
MC-93	unnamed monocarbocyclic sesterterpene (reported as compound 7)		<i>Ircinia</i> sp. ¹⁴³
MC-94	unnamed monocarbocyclic sesterterpene (reported as compound 8)		<i>Ircinia</i> sp. ¹⁴³
MC-95	unnamed monocarbocyclic sesterterpene (reported as compound 9)		<i>Ircinia</i> sp. ¹⁴³
MC-96	halisulfate 2		<i>Halichondriidae</i> sp. ¹⁴⁴ <i>Dysidea</i> sp. ¹⁴⁵
MC-97	polyunsaturated sesterterpene I		<i>Rhizosolenia setigera</i> ¹¹
MC-98	<i>n</i> -non-2'-en-1'-yl-13(15,19,19-trimethylcyclohex-14, 16-dienyl)-2,6,10-trimethyltetradec-6-ol-13-on-1-oate		<i>Hemidesmus indicus</i> ¹⁴⁶
MC-99	3-[6-(4,8-dimethylnona-1,3,7-trienyl)-4-hydroxy-2,6-dimethylcyclohex-1-enyl]-3-hydroxypropionic acid 1 glucoside		<i>Woodwardia virginica</i> ¹⁴⁷
MC-100	cericerol-I		<i>Ceroplastes ceriferus</i> ¹⁴⁸⁻¹⁵¹

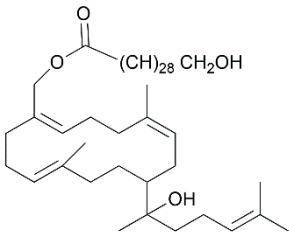
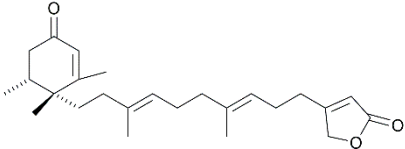
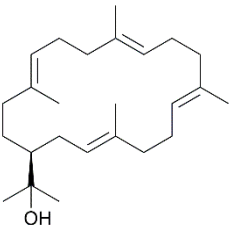
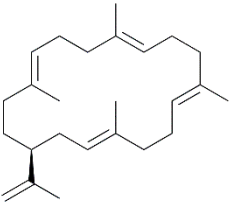
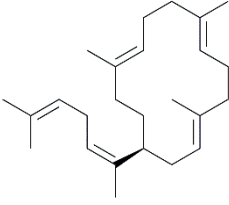
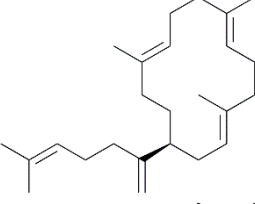
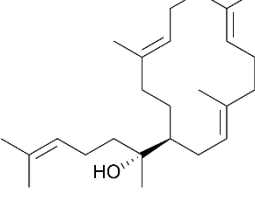
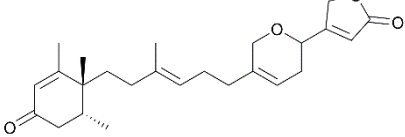
MC-101	α -cericerol-I		<i>Ceroplastes ceriferus</i> ¹⁴⁹
MC-102	cericerol-II		<i>Ceroplastes ceriferus</i> ¹⁴⁹
MC-103	albocerol		<i>Ceroplastes albolineatus</i> ¹⁵²
MC-104	cericeroic acid		<i>Ceroplastes ceriferus</i> ^{149, 151, 153}
MC-105	18-dihydro-19-hydroxycericeroic acid		<i>Ceroplastes ceriferus</i> ^{151, 153}
MC-106	ceriferic acid		<i>Ceroplastes ceriferus</i> ^{151, 153-155}
MC-107	ceriferic acid-I		<i>Ceroplastes ceriferus</i> ¹⁵¹
MC-108	ceriferol		<i>Ceroplastes ceriferus</i> ^{151, 153, 156}

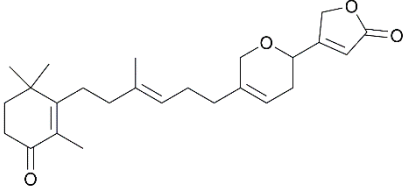
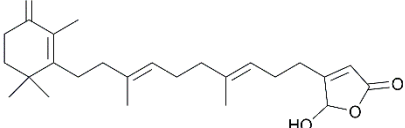
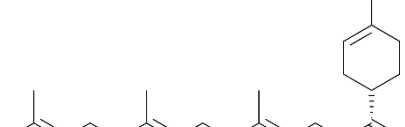
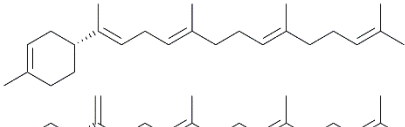
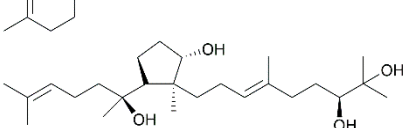
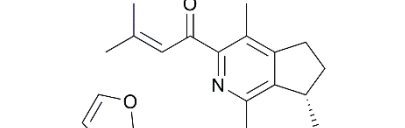
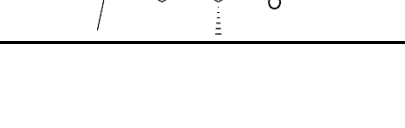
MC-109	ceriferol-I		<i>Ceroplastes ceriferus</i> ¹⁵¹
MC-110	13-ethoxycericerene		<i>Ceroplastes ceriferus</i> ^{149, 151}
MC-111	13-methoxycericerene		<i>Ceroplastes ceriferus</i> ^{149, 151}
MC-112	(2Z,6Z,10E)-cericerene-15,24-diol		<i>Ceroplastes pseudoceriferus</i> ¹⁵⁷
MC-113	24-(ω-hydroxy fatty acid) ester of (2Z,6Z,10E)-cericerene-15,24-diol		<i>Ceroplastes pseudoceriferus</i> ¹⁵⁷
MC-114	(2E)-α-cericerene		<i>Emericella variegata</i> ^{158, 159} <i>Escherichia coli</i> ¹⁶⁰ <i>Leucosceptrum canum</i> ¹⁵⁹
MC-115	(-)-alotaketal A		<i>Hamigera</i> sp. ¹⁶¹

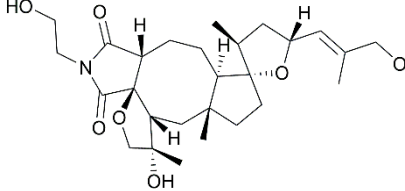
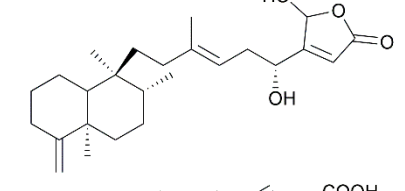
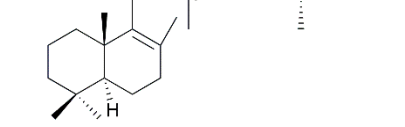
MC-116	alotaketal B		<i>Hamigera</i> sp. ¹⁶¹
MC-117	alotaketal C		<i>Hamigera</i> sp. ¹⁶¹ <i>Phorbas</i> sp. ^{162, 163}
MC-118	alotaketal D		<i>Hamigera</i> sp. ¹⁶¹ <i>Phorbas</i> sp. ¹⁶³
MC-119	phorbaketal A		<i>Monanchora</i> sp. ¹⁰⁵ <i>Phorbas</i> sp. ¹⁶⁴
MC-120	phorbaketal B		<i>Monanchora</i> sp. ¹⁰⁵ <i>Phorbas</i> sp. ¹⁶⁴

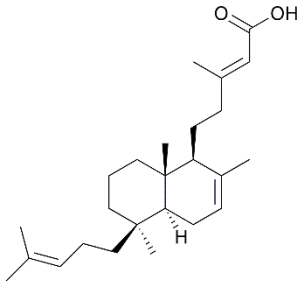
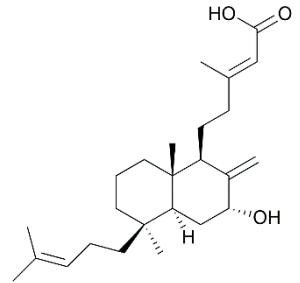
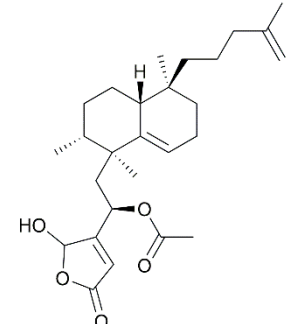
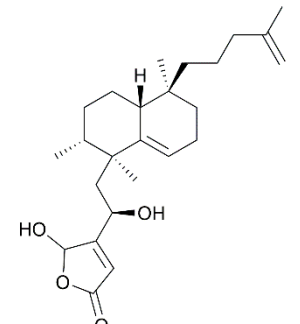
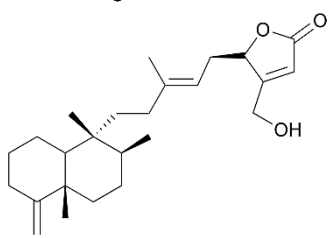
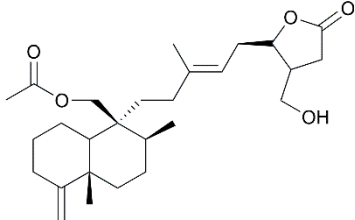
MC-121	phorbaketal C		<i>Monanchora</i> sp. ¹⁰⁵ <i>Phorbas</i> sp. ¹⁶⁴
MC-122	phorbaketal D		<i>Monanchora</i> sp. ¹⁰⁵
MC-123	phorbaketal E		<i>Monanchora</i> sp. ¹⁰⁵
MC-124	phorbaketal F		<i>Monanchora</i> sp. ¹⁰⁵
MC-125	phorbaketal G		<i>Monanchora</i> sp. ¹⁰⁵
MC-126	phorbaketal H		<i>Monanchora</i> sp. ¹⁰⁵
MC-127	phorbaketal I		<i>Monanchora</i> sp. ¹⁰⁵
MC-128	phorbaketal J		<i>Monanchora</i> sp. ¹⁰⁵
MC-129	phorbaketal K		<i>Monanchora</i> sp. ¹⁰⁵

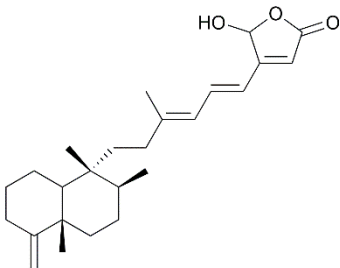
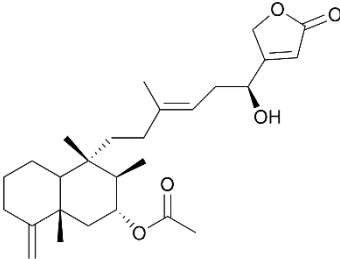
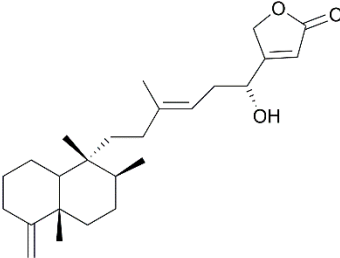
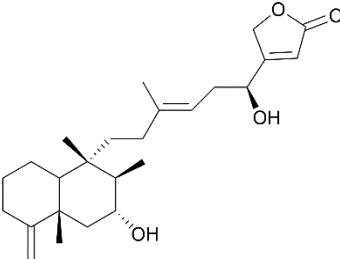
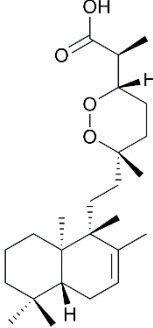
MC-130	phorbaketal L		<i>Phorbas</i> sp. ¹⁶⁵
MC-131	phorbaketal M		<i>Phorbas</i> sp. ¹⁶⁵
MC-132	phorbaketal N		<i>Phorbas</i> sp. ¹⁶⁵
MC-133	unnamed spiroketal sesterterpene (reported as spiroketal)		<i>Monanchora</i> sp. ¹⁶⁶
MC-134	alotaketal E		<i>Phorbas</i> sp. ¹⁶³
MC-135	unnamed monocarbocyclic sesterterpene (reported as compound 2)		<i>Hippospongia lachne</i> ⁹
MC-136	unnamed monocarbocyclic sesterterpene (reported as compound 3)		<i>Hippospongia lachne</i> ⁹
MC-137	unnamed monocarbocyclic sesterterpene (reported as compound 4)		<i>Hippospongia lachne</i> ⁹
MC-138	2Z,6Z,10E-cericerene-15,24-diol		<i>Ceroplastes pseudoceriferus</i> ¹⁵⁷

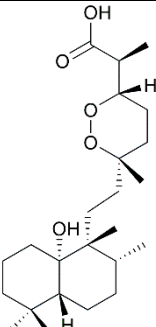
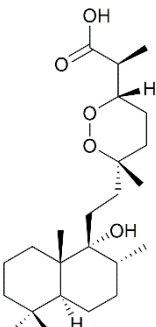
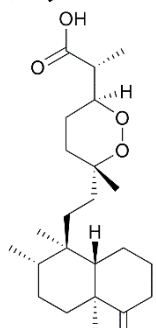
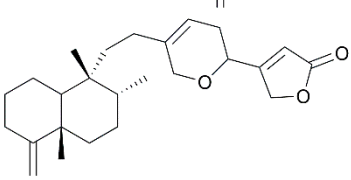
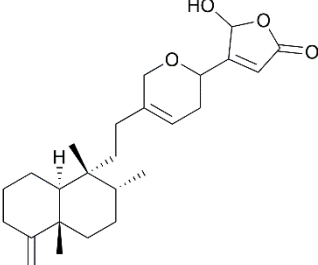
MC-139	2Z,6Z,10E-cericerene-15,24-diol-30-triacontanoate		<i>Ceroplastes pseudoceriferus</i> ¹⁵⁷
MC-140	cyclolinteinon		<i>Cacospongia</i> cf. <i>linteiformis</i> ¹⁶⁷
MC-141	(S,2E,6E,10E,14E)-3,7,11,15-tetramethyl-18-(1-hydroxyisopropyl)cyclooctadecatetraene		<i>Leucosceptrum canum</i> / <i>Nicotiana benthamiana</i> ¹⁶⁸
MC-142	(S,2E,6E,10E,14E)-3,7,11,15-tetramethyl-18-(1-methylethenyl)cyclooctadecatetraene		<i>Leucosceptrum canum</i> / <i>Nicotiana benthamiana</i> ¹⁶⁸
MC-143	(S,2E,15Z)-α-cericerene		<i>Leucosceptrum canum</i> / <i>Nicotiana benthamiana</i> ¹⁶⁸
MC-144	(S,2E)-cericerene		<i>Leucosceptrum canum</i> / <i>Nicotiana benthamiana</i> ¹⁶⁸
MC-145	(14S,15R,2E)-15-hydroxy-α-cericerene		<i>Leucosceptrum canum</i> / <i>Nicotiana benthamiana</i> ¹⁶⁸
MC-146	oshimalide A		<i>Luffariella</i> sp. ¹⁶⁹

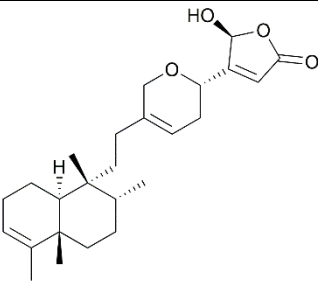
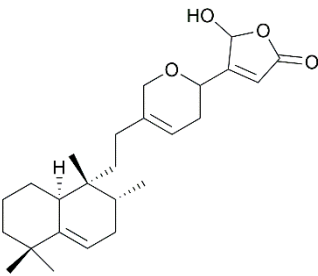
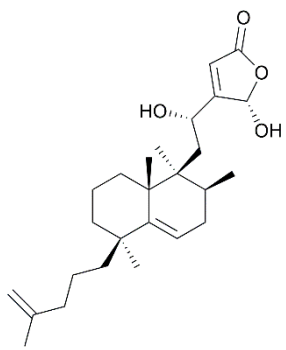
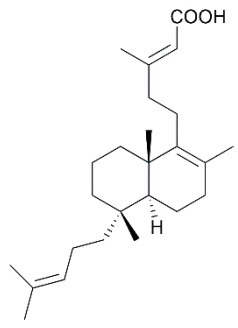
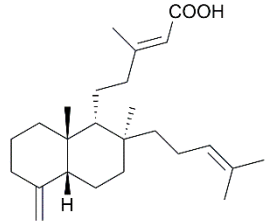
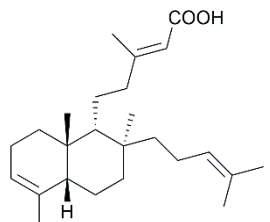
MC-147	oshimalide B		<i>Luffariella</i> sp. ¹⁶⁹
MC-148	16-oxoluffariellolide		<i>Thorectandra</i> sp. ¹³⁶
MC-149	(-)-somialensene B		<i>Streptomyces somaliensis</i> ¹⁷⁰
MC-150	(+)- α -geranylbisabolene		<i>Colquhounia coccinea</i> var. <i>mollis</i> /Escherichia coli ¹⁷¹
MC-151	(+)-somialensene B		<i>Colquhounia coccinea</i> var. <i>mollis</i> /Escherichia coli ¹⁷¹
MC-152	postrediene F		<i>Pleurotus ostreatus</i> SY10 ¹⁴
MC-153	leucosceptrodine		<i>Leucosceptrum canum</i> ¹⁷²

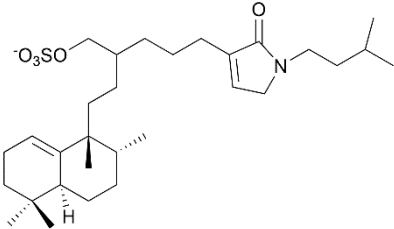
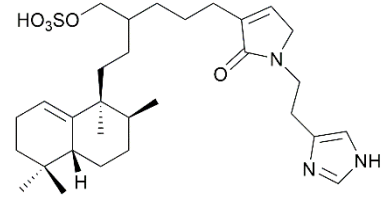
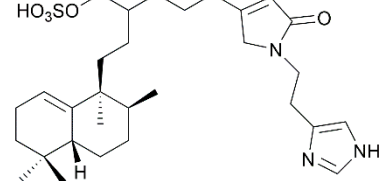
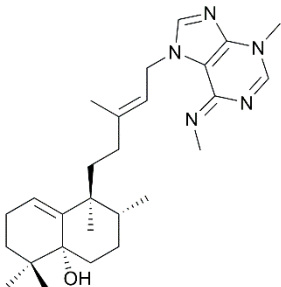
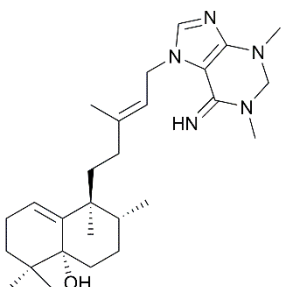
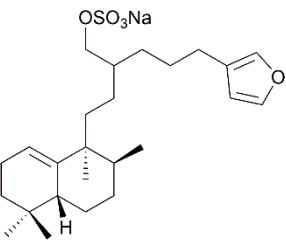
Section 1.6. Structures of dicarbocyclic (DC) sesterterpenoids (with and without heterocycles)			
c. n.	IUPAC name or Trivial name	Structure	Organism as reported in the reference
DC-1	bipolarolide E		<i>Bipolaris</i> sp. TJ403-B1 ¹⁷³
DC-2	palauolol		<i>Fascaplysinopsis</i> sp. ¹⁷⁴ <i>Thorectandra</i> sp. ¹⁷⁵
DC-3	unnamed dicarbocyclic sesterterpene (reported as compound 5)		<i>Latrunculia brevis</i> ¹⁷⁶

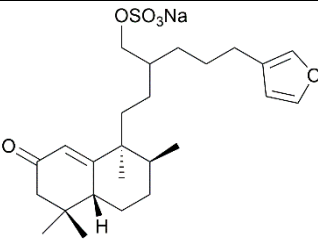
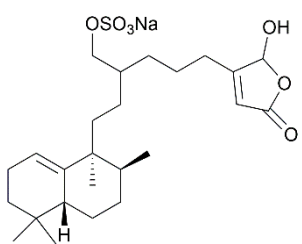
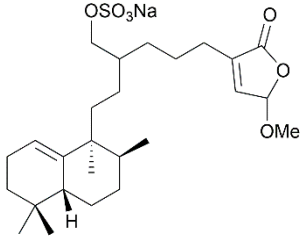
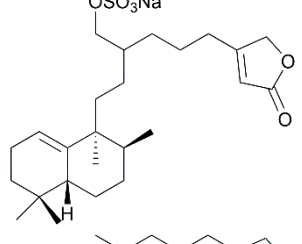
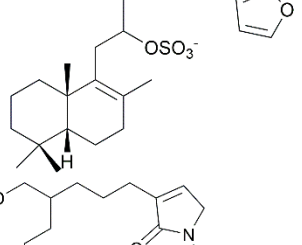
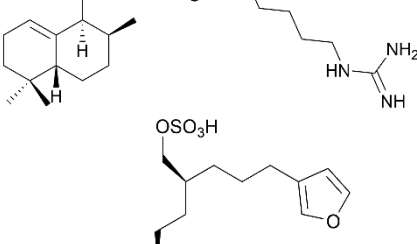
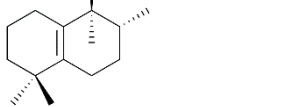
DC-4	kohamaic acid A		<i>Ircinia</i> sp. ¹⁷⁷
DC-5	kohamaic acid B		<i>Ircinia</i> sp. ¹⁷⁷
DC-6	cladocoran A		<i>Cladocora cespitosa</i> [sic!] ¹⁷⁸
DC-7	cladocoran B		<i>Cladocora cespitosa</i> [sic!] ¹⁷⁸
DC-8	thorectandrol A		<i>Thorectandra</i> sp. ¹⁷⁵
DC-9	thorectandrol B		<i>Thorectandra</i> sp. ¹⁷⁵

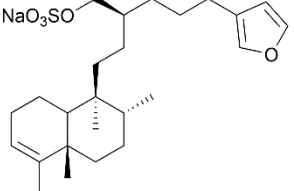
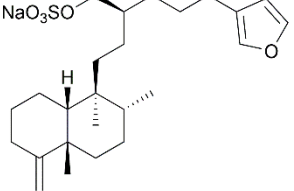
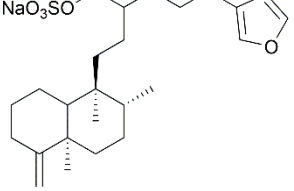
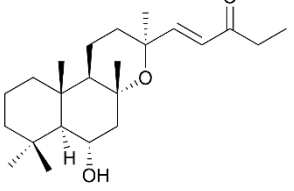
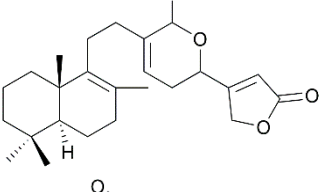
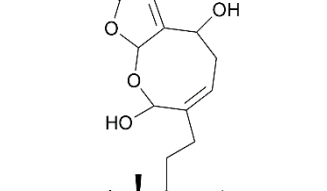
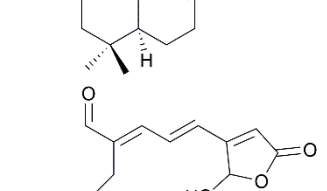
DC-10	palauolide		Unidentified sponge from Palau ¹⁷⁹ <i>Thorectandra</i> sp. ⁸⁶
DC-11	thorectandrol C		<i>Thorectandra</i> sp. ⁸⁶
DC-12	thorectandrol D		<i>Thorectandra</i> sp. ⁸⁶
DC-13	thorectandrol E		<i>Thorectandra</i> sp. ⁸⁶
DC-14	unnamed dicarbocyclic sesterterpene (reported as compound 1)		<i>Mycale (Carmia) cf. spongiosa</i> ¹⁸⁰

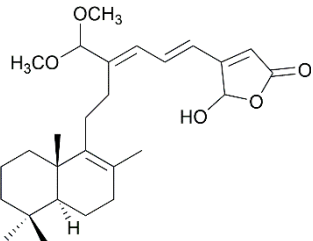
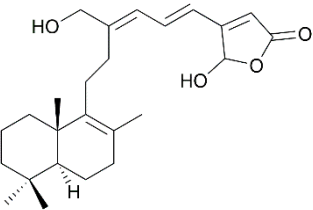
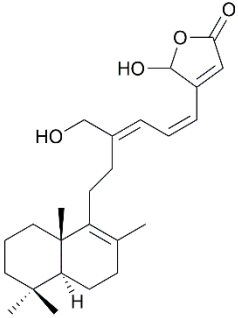
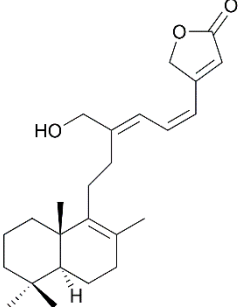
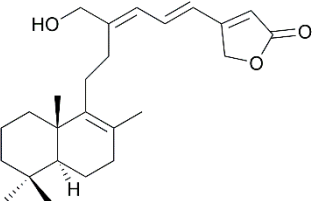
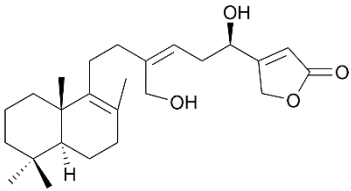
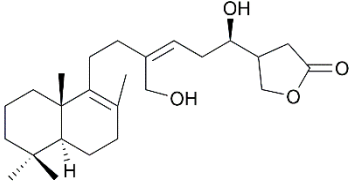
DC-15	mycaperoxide A		<i>Mycale</i> sp. ¹⁸¹
DC-16	mycaperoxide B		<i>Mycale</i> sp. ¹⁸¹
DC-17	sigmosceptrellin B (= prianicin B) (dubious configurational assignment) (reported as compound 2)		<i>Mycale</i> (<i>Carmia</i>) cf. <i>spongiosa</i> ¹⁸⁰ <i>Prianos</i> sp. ¹²⁰ <i>Sigmosceptrella laevis</i> ^{121, 182} <i>Diacarnus erythraeanus</i> ¹²³
DC-18	25-deoxycacospongionolide B		<i>Fasciospongia cavernosa</i> ¹⁸³
DC-19	cacospongionolide B		<i>Fasciospongia cavernosa</i> ¹⁸³

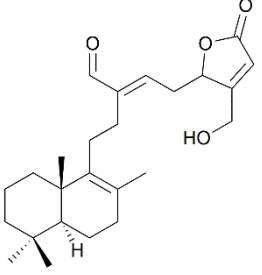
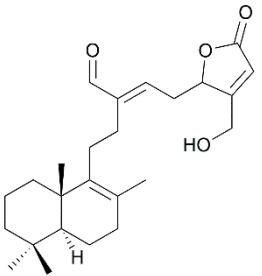
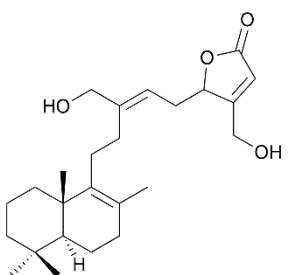
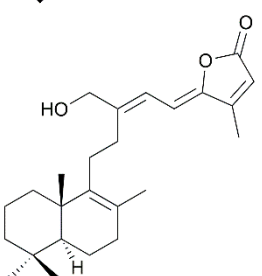
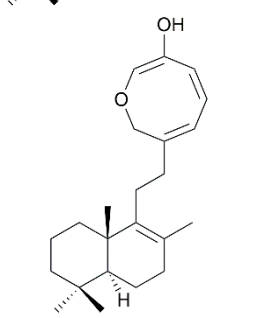
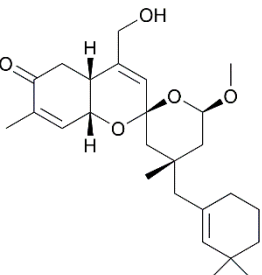
DC-20	cacospongionolide E		<i>Fasciospongia cavernosa</i> ¹⁸⁴
DC-21	cacospongionolide F		<i>Fasciospongia cavernosa</i> ¹⁸⁵
DC-22	dysidiolide		<i>Dysidea etheria</i> ¹⁸⁶
DC-23	dysideapalaunic acid		<i>Dysidea</i> sp. ¹⁸⁷
DC-24	bilosespene A		<i>Dysidea cinerea</i> ¹⁸⁸
DC-25	bilosespene B		<i>Dysidea cinerea</i> ¹⁸⁸

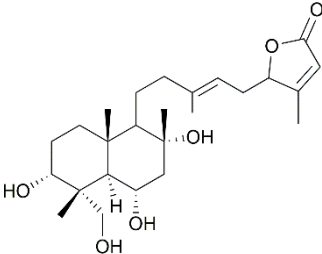
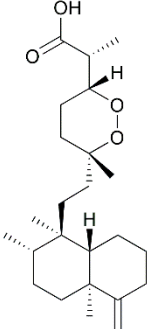
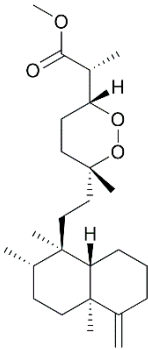
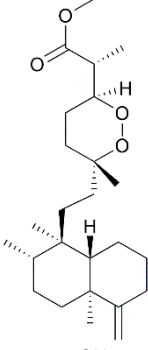
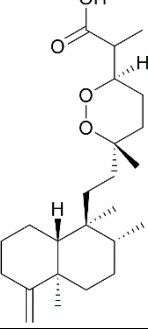
DC-26	irregularasulfate		<i>Spongia irregularis</i> ¹³⁷
DC-27	fasciospongine A		<i>Fasciospongia</i> sp. ¹⁸⁹
DC-28	fasciospongine B		<i>Fasciospongia</i> sp. ¹⁸⁹
DC-29	(+)-agelisamine-A		<i>Agelas mauritiana</i> ¹⁹⁰
DC-30	(+)-agelisamine-B		<i>Agelas mauritiana</i> ¹⁹⁰
DC-31	halisulfate 7		<i>Spongia irregularis</i> ¹³⁷ <i>Fasciospongia</i> sp. ¹⁹¹

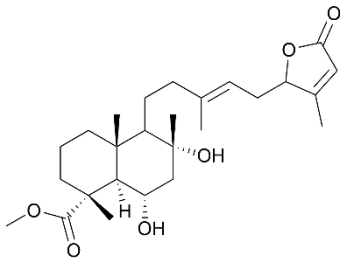
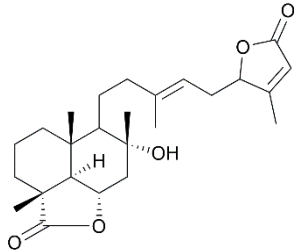
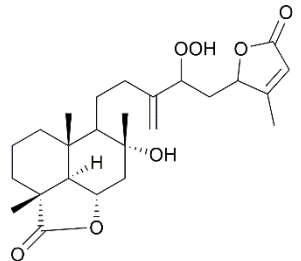
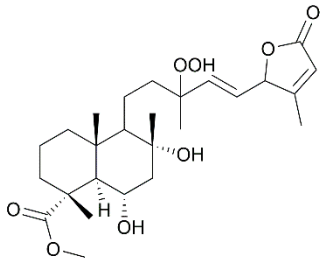
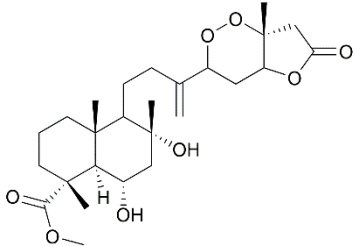
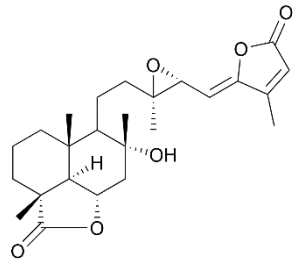
DC-32	halisulfate 8		<i>Darwinella australensis</i> ¹⁹²
DC-33	25-hydroxyhalisulfate 9		<i>Fasciospongia</i> sp. ¹⁹¹
DC-34	halisulfate 10		<i>Darwinella australensis</i> ¹⁹²
DC-35	halisulfate 9		<i>Darwinella australensis</i> ¹⁹² <i>Fasciospongia</i> sp. ¹⁹¹
DC-36	sch 599473		<i>Ircinia</i> sp. ¹⁹³
DC-37	fasciospongine C		<i>Fasciospongia</i> sp. ¹⁹¹
DC-38	halisulfate 3		Marine sponge of the family Halichondriidae ¹⁴⁴ <i>Igernella</i> sp. ¹³⁸ <i>Dysidea</i> sp. ¹⁴⁵

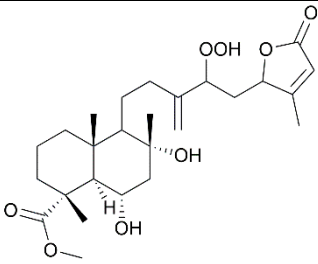
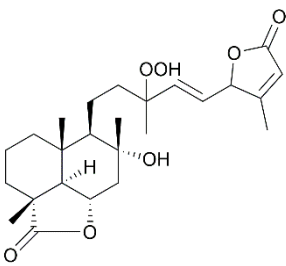
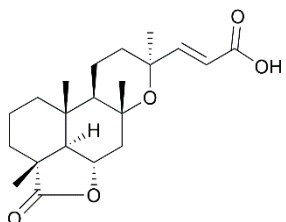
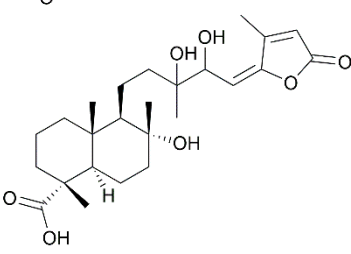
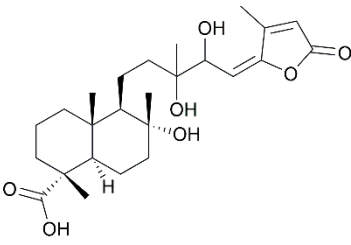
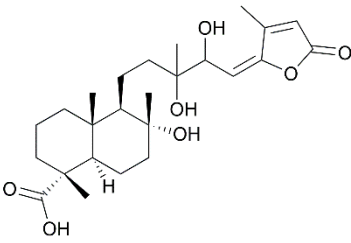
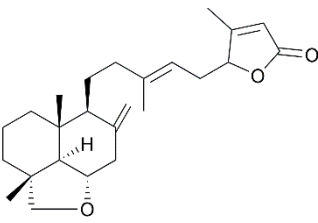
DC-39	halisulfate 4		Marine sponge of the family Halichondriidae ¹⁴⁴
DC-40	halisulfate 5		Marine sponge of the family Halichondriidae ¹⁴⁴ <i>Hippospongia</i> sp. ¹⁹⁴ <i>Dysidea</i> sp. ¹⁴⁵
DC-41	halisulfate 6		Marine sponge of the family Halichondriidae ¹⁴⁴
DC-42	13- <i>epi</i> -yosgadensenol		<i>Salvia yosgadensis</i> ¹⁹⁵
DC-43	luffarin A		<i>Luffariella geometrica</i> ²⁰
DC-44	luffarin B		<i>Luffariella geometrica</i> ²⁰
DC-45	luffarin C		<i>Luffariella geometrica</i> ²⁰

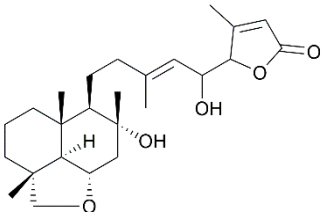
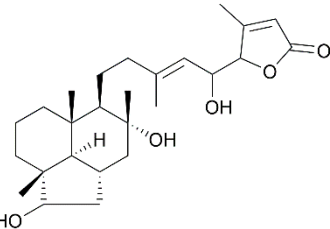
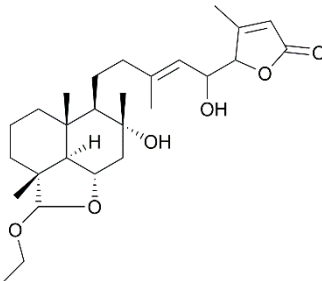
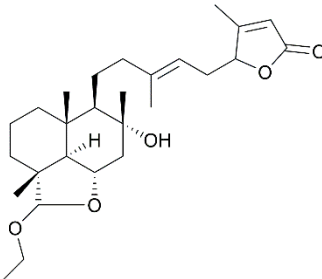
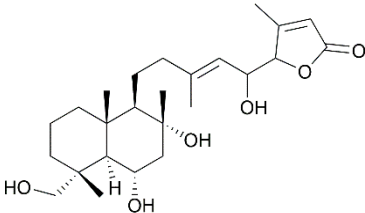
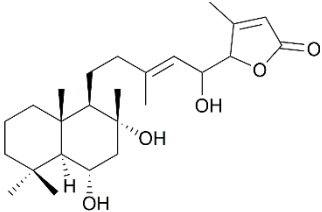
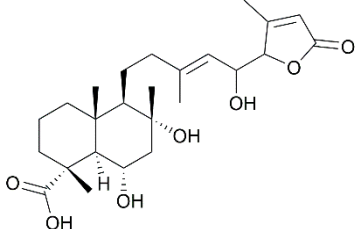
DC-46	luffarin D		<i>Luffariella geometrica</i> ²⁰
DC-47	luffarin E		<i>Luffariella geometrica</i> ²⁰
DC-48	luffarin F		<i>Luffariella geometrica</i> ²⁰
DC-49	luffarin G		<i>Luffariella geometrica</i> ²⁰
DC-50	luffarin H		<i>Luffariella geometrica</i> ²⁰
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DC-52	luffarin J		<i>Luffariella geometrica</i> ²⁰

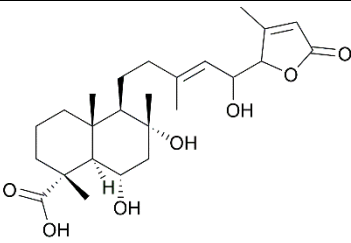
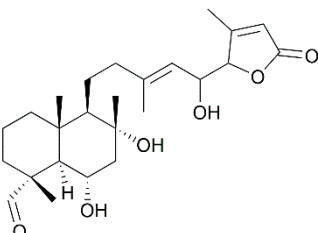
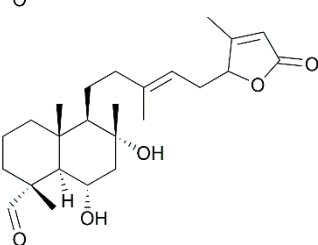
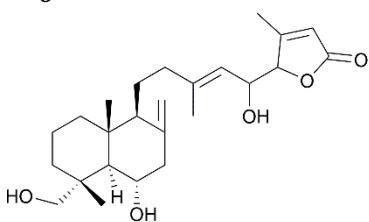
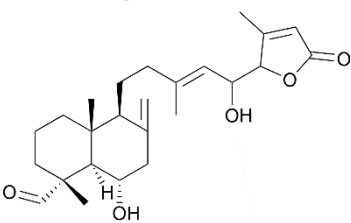
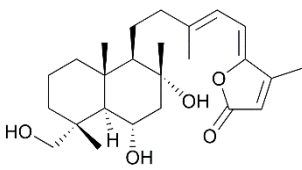
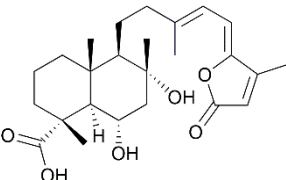
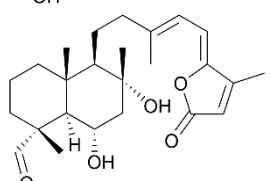
DC-53	luffarin K		<i>Luffariella geometrica</i> ²⁰
DC-54	luffarin M		<i>Luffariella geometrica</i> ²⁰
DC-55	luffarin L		<i>Luffariella geometrica</i> ²⁰
DC-56	luffarin N		<i>Luffariella geometrica</i> ²⁰
DC-57	luffarin O		<i>Luffariella geometrica</i> ²⁰
DC-58	unnamed dicarbocyclic sesterterpene (reported as compound 1)		<i>Haliclona</i> sp. ¹⁹⁶

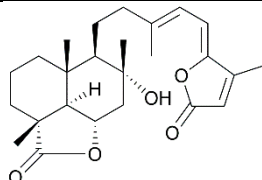
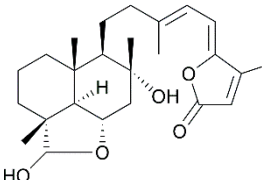
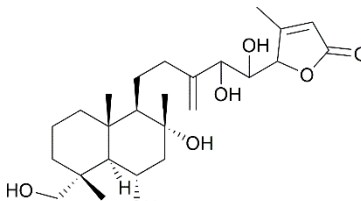
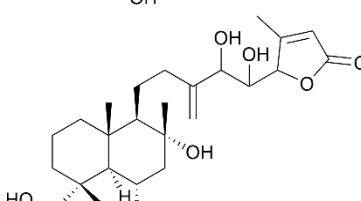
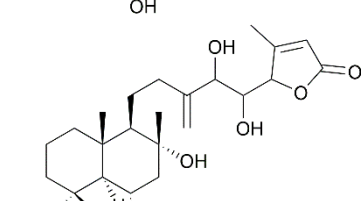
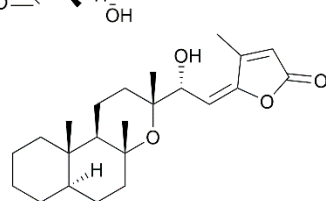
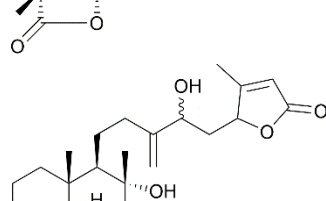
DC-59	salvisyriacolide		<i>Salvia syriaca</i> ¹⁹⁷
DC-60	sigmosceptrellin A		<i>Sigmosceptrella laevis</i> ^{182, 198} <i>Latrunculia</i> sp. ³⁶ unidentified sponge ⁶²
DC-61	sigmosceptrellin A methyl ester		<i>Sigmosceptrella laevis</i> ¹⁹⁸ <i>Latrunculia</i> sp. ³⁶
DC-62	sigmosceptrellin-B methyl ester		<i>Latrunculia</i> sp. ³⁶ <i>Sigmosceptrella laevis</i> ¹⁸² <i>Prianos</i> sp. ¹²⁰ unidentified sponge ⁶²
DC-63	sigmosceptrellin-C		<i>Sigmosceptrella laevis</i> ¹⁸²

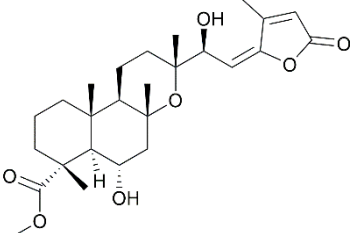
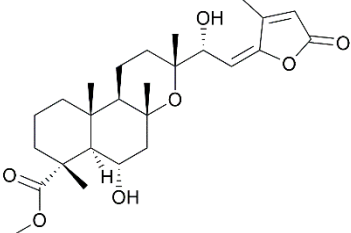
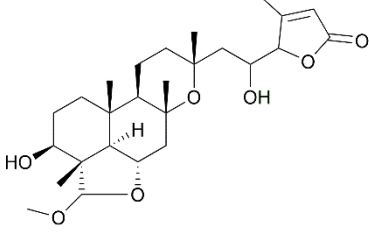
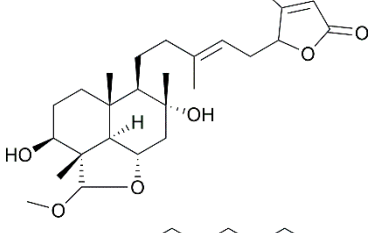
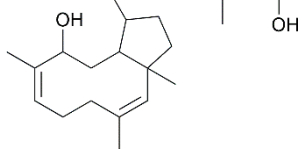
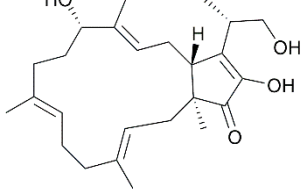
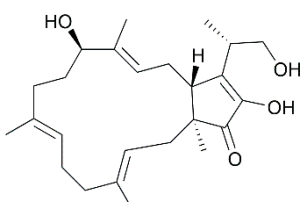
DC-64	salvileucolide methyl ester		<i>Salvia hypoleuca</i> ¹⁹⁹ <i>Salvia tingitana</i> ²⁰⁰
DC-65	salvileucolide-6,23-lactone		<i>Salvia hypoleuca</i> ¹⁹⁹ <i>Salvia tingitana</i> ²⁰⁰
DC-66	14-hydroperoxy-13(21)-dehydro-13,14-dihydrosalvileucolide-6,23-lactone		<i>Salvia hypoleuca</i> ²⁰¹
DC-67	salvileucolide-hydroperoxide 2		<i>Salvia hypoleuca</i> ²⁰¹
DC-68	salvileucolide-cyclic peroxide		<i>Salvia hypoleuca</i> ²⁰¹
DC-69	salvileucolide-lactone (15,16-deidrosalvileucolide-6,23-lactone-trans-epoxide)		<i>Salvia hypoleuca</i> ²⁰¹

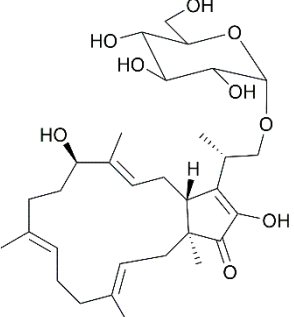
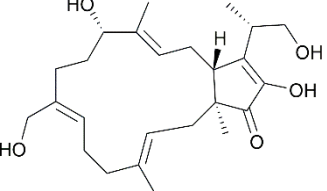
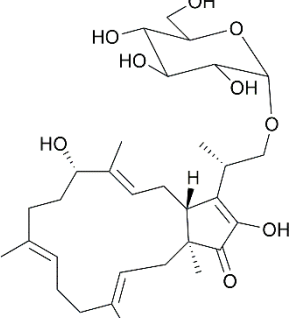
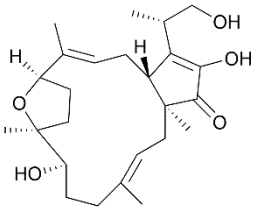
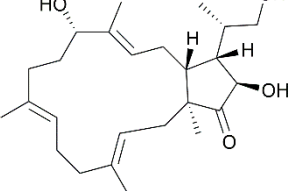
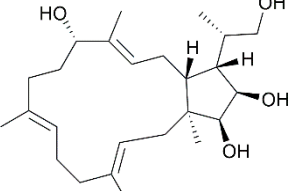
DC-70	unnamed dicarbocyclic sesterterpene (reported as hydroperoxide 4)		<i>Salvia hypoleuca</i> ²⁰¹
DC-71	8 <i>R</i> -hydroxy-13-hydroperoxylabd-14,17-dien-19,16,23,6 <i>α</i> -di-olide		<i>Salvia sahendica</i> ²⁰²
DC-72	17,18,19,20-tetranor-13- <i>epi</i> -manoyloxide-14-en-16-oic acid-23,6 <i>α</i> -olide		<i>Salvia sahendica</i> ²⁰²
DC-73	8 <i>α</i> ,13,14-threo-trihydroxy-labd-15,17-dien-16,19-olide-23 oic acid		<i>Salvia palaestina</i> ²⁰³
DC-74	8 <i>α</i> ,13,14-erythro-trihydroxy-labd-15,17-dien-16,19-olide-28-oic acid		<i>Salvia palaestina</i> ²⁰³
DC-75	3 <i>α</i> ,8 <i>α</i> ,13,14-erythro-tetrahydroxy-labd-15,17-dien-16,19-olide		<i>Salvia palaestina</i> ²⁰³
DC-76	23,6 <i>α</i> -epoxy-labd-8,13(14),17-trien-16(<i>R</i>),19-olide		<i>Salvia dominica</i> ²⁰⁴

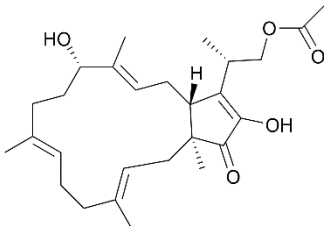
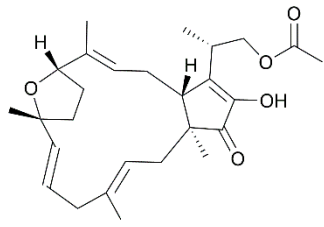
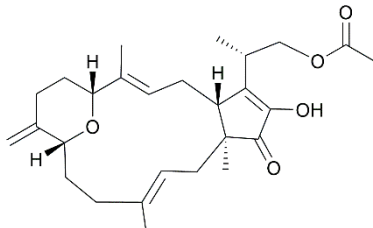
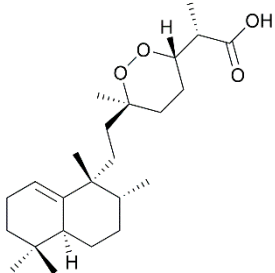
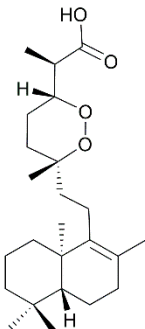
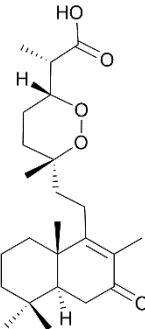
DC-77	8 α ,15(<i>S</i>)-dihydroxy-23,6 α -epoxy-labd-13(14),17-dien-16(<i>S</i>),19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-78	8 α ,15(<i>S</i>),23 α -trihydroxy-23,6 α -epoxy-labd-13(14),17-dien-16(<i>S</i>),19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-79	8 α ,15(<i>S</i>)-dihydroxy-23 α -O-ethyl-23,6 α -epoxy-labd-13(14),17-dien-16(<i>S</i>),19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-80	8 α -hydroxy-23 α -O-ethyl-23,6 α -epoxy-labd-13(14),17-dien-16(<i>R</i>),19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-81	6 α ,8 α ,15(<i>S</i>),23-tetrahydroxy-labd-13(14),17-dien-16(<i>S</i>),19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-82	6 α ,8 α ,15(<i>S</i>)-trihydroxy-labd-13(14),17-dien-16(<i>S</i>),19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-83	6 α ,8 α ,15(<i>S</i>)-trihydroxy-23-carboxyl-labd-13(14),17-dien-16(<i>S</i>),19-olide		<i>Salvia dominica</i> ²⁰⁴

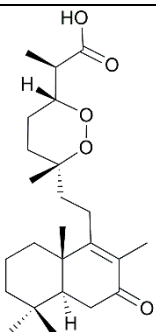
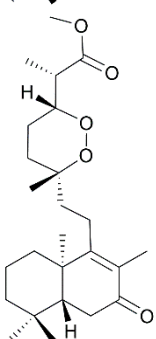
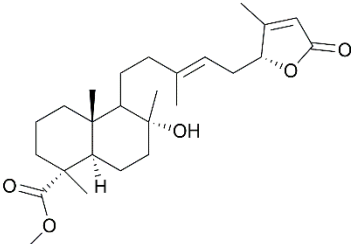
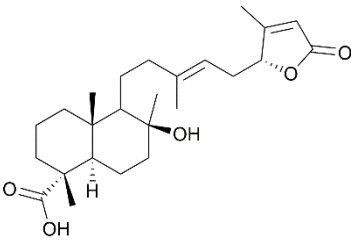
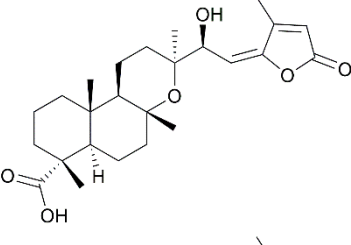
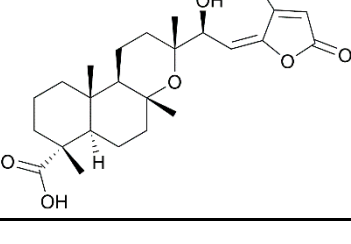
DC-84	6 α ,8 α -dihydroxy-23-carbossi-labd-13(14),17-dien-16,19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-85	6 α ,8 α ,15(S)-trihydroxy-23-oxo-labd-13(14),17-dien-16(S),19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-86	6 α ,8 α -dihydroxy-23-oxo-labd-13(14),17-dien-16(R),19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-87	6 α ,15(S),23-trihydroxy-labd-8(22),13(14),17-trien-16(S),19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-88	6 α ,15(S)-dihydroxy-23-oxo-labd-8(22),13(14),17-trien-16(S),19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-89	6 α ,8 α ,23-trihydroxy-labd-13(14),15,17-trien-16,19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-90	6 α ,8 α -dihydroxy-23-carbossi-labd-13(14),15,17-trien-16,19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-91	6 α ,8 α -dihydroxy-23-oxo-13(14),15,17-trien-16,19-olide		<i>Salvia dominica</i> ²⁰⁴

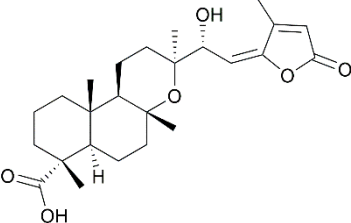
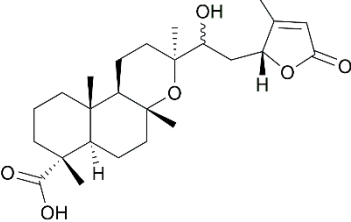
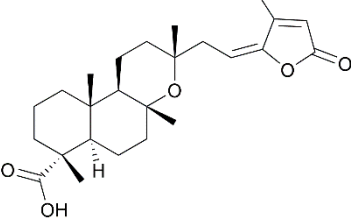
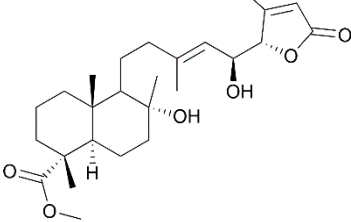
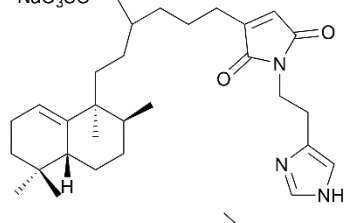
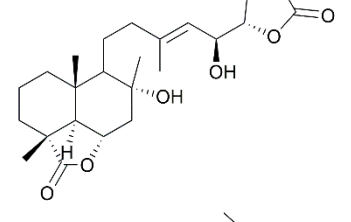
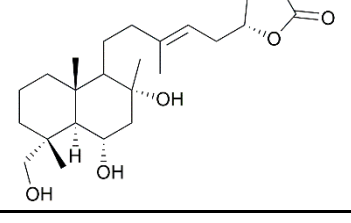
DC-92	8 α -hydroxylabd-13(14),15,17-trien-6R,23-16,19-diolide		<i>Salvia dominica</i> ²⁰⁴
DC-93	8 α -23-dihydroxy-23,6R-epoxy-labd-13(14),15,17-trien-16,19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-94	6 α ,8 α ,23,14,15-threo-pentahydroxy-labd-13(21),17-dien-16,19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-95	6 α ,8 α ,23,14,15-erythro-pentahydroxylabd-13(21),17-dien-16,19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-96	6 α ,8 α ,14,15-threotetrahydroxy-23-oxo-labd-13(21),17-dien-16,19-olide		<i>Salvia dominica</i> ²⁰⁴
DC-97	lachnocalyxolide A		<i>Salvia lachnocalyx</i> ²⁰⁵
DC-98	lachnocalyxolide B		<i>Salvia lachnocalyx</i> ²⁰⁵

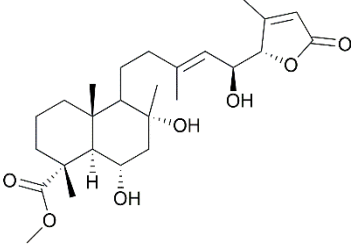
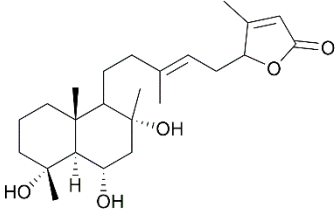
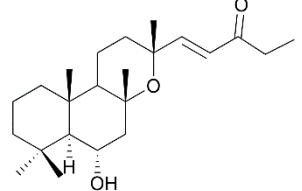
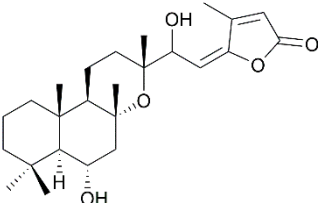
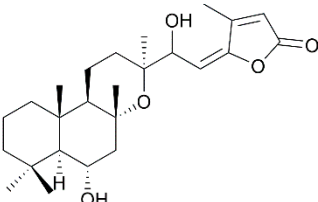
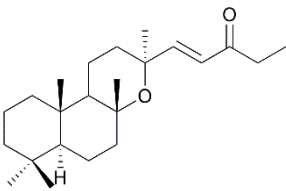
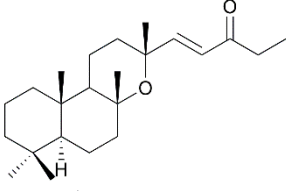
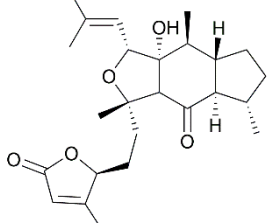
DC-99	lachnocalyxolide C		<i>Salvia lachnocalyx</i> ²⁰⁵
DC-100	lachnocalyxolide C'		<i>Salvia lachnocalyx</i> ²⁰⁵
DC-101	salvidominicolide A		<i>Salvia dominica</i> ²⁰⁶
DC-102	salvidominicolide B		<i>Salvia dominica</i> ²⁰⁶
DC-103	albolineol		<i>Ceroplastes albolineatus</i> ²⁰⁷
DC-104	terpestacin		<i>Alternaria alternata</i> ²⁰⁸ <i>Aplosporella javeedii</i> ²⁰⁹ <i>Arthrinium</i> spp. ²¹⁰⁻²¹³ <i>Bipolaris sorokiniana</i> BRIP10943 ²¹⁴ <i>Fusarium fujikuroi</i> ²¹⁵ <i>Fusarium proliferatum</i> ²¹⁶ <i>Phomopsis</i> sp. ²¹⁷ <i>Tolypocladium inflatum</i> ²¹⁸
DC-105	11- <i>epi</i> -terpestacin (siccanol)		<i>Bipolaris sorokiniana</i> NSDR-011 ²¹⁹ <i>Drechslera siccanis</i> ²²⁰

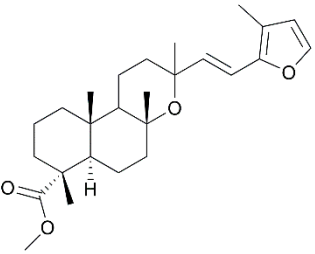
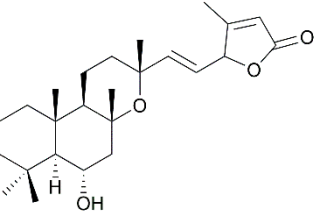
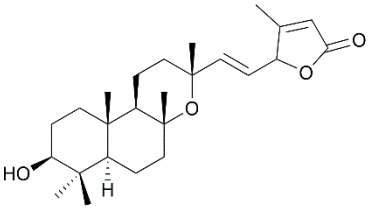
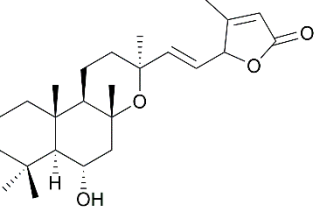
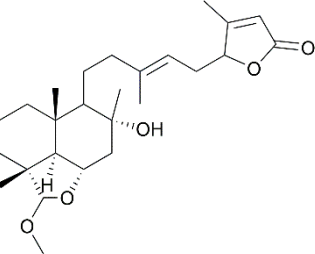
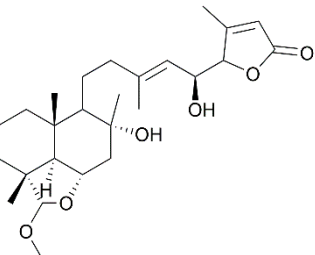
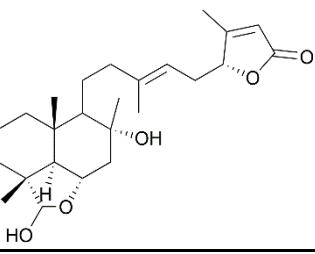
DC-106	11- <i>epi</i> -terpestacin glycoside		<i>Bipolaris sorokiniana</i> NSDR-011 ²²¹
DC-107	21-hydroxyterpestacin		<i>Arthrinium</i> sp. ²¹³
DC-108	24- α -D-glucosyl-(-)-terpestacin		<i>Alternaria alternata</i> ²⁰⁸
DC-109	terpestacin B		<i>Arthrinium</i> sp. ²¹³
DC-110	16,17-dihydro-(-)-terpestacin		<i>Aplosporella javeedii</i> ²⁰⁹
DC-111	terpestacin C		<i>Aplosporella javeedii</i> ²⁰⁹

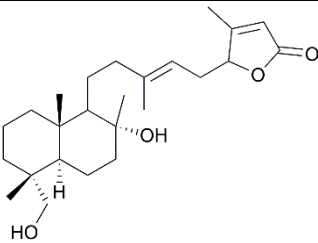
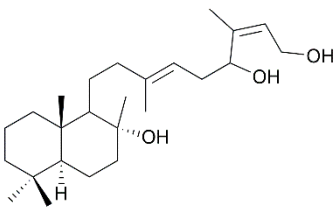
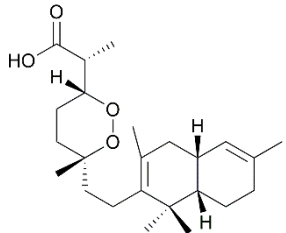
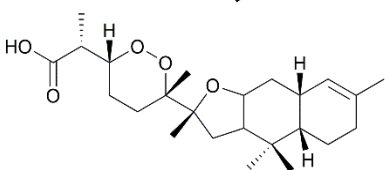
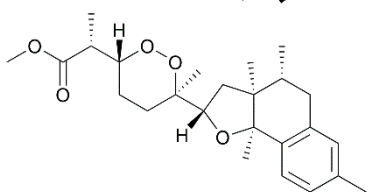
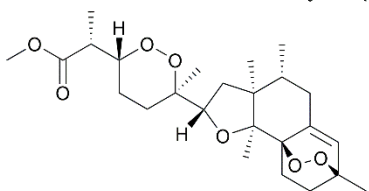
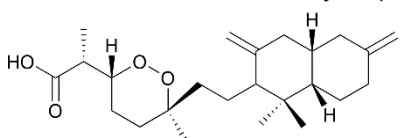
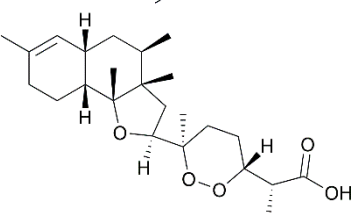
DC-112	fusaproliferin (proliferin)		<i>Alternaria alternata</i> ²⁰⁸ <i>Aplosporella javeedii</i> ²⁰⁹ <i>Bipolaris sorokiniana</i> NSDR-011 ²²¹ <i>Fusarium proliferatum</i> ^{216, 222-224} <i>Fusarium solani</i> ²²⁵
DC-113	fusaprolifin A		<i>Fusarium proliferatum</i> ²¹⁶
DC-114	fusaprolifin B		<i>Fusarium proliferatum</i> ²¹⁶
DC-115	mycaleperoxide		<i>Mycale</i> sp. ²²⁶
DC-116	diacarperoxide D		<i>Diacarnus megaspinorhabdosa</i> [sic!] ¹⁰⁴
DC-117	diacarperoxide E		<i>Diacarnus megaspinorhabdosa</i> [sic!] ¹⁰⁴

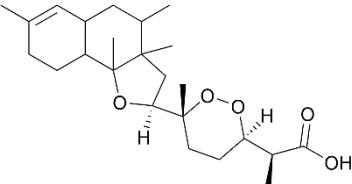
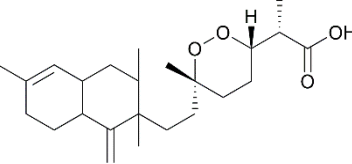
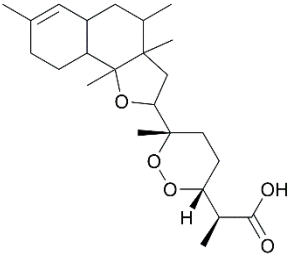
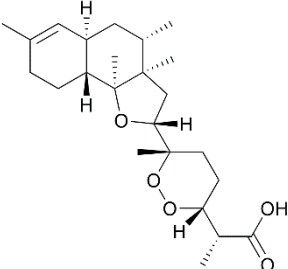
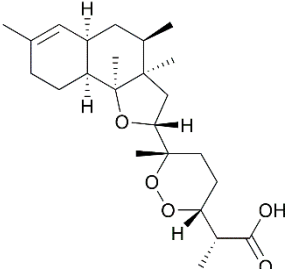
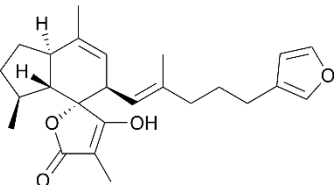
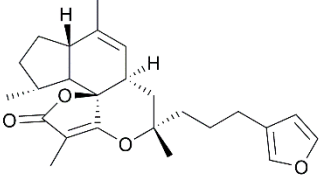
DC-118	diacarperoxide F		<i>Diacarnus megaspinorhabdosa</i> [sic!] ¹⁰⁴
DC-119	diacarperoxide G		<i>Diacarnus megaspinorhabdosa</i> [sic!] ¹⁰⁴
DC-120	(13 <i>E</i>)-8α-hydroxy-23-carboxymethylabd-13(14),17(18)-dien-16,19-olide		<i>Salvia tingitana</i> ²⁰⁰
DC-121	salvimirzacolide		<i>Salvia mirzayanii</i> ²²⁷
DC-122	4 <i>R</i> ,5 <i>R</i> ,8 <i>R</i> ,9 <i>R</i> ,10 <i>S</i> ,13 <i>S</i> ,14 <i>S</i>)-14-hydroxymanoyloxide-15,17-dien-15(<i>Z</i>)-16,19-olide		<i>Salvia mirzayanii</i> ²²⁸
DC-123	(4 <i>R</i> ,5 <i>R</i> ,8 <i>R</i> ,9 <i>R</i> ,10 <i>S</i> ,13 <i>R</i> ,14 <i>S</i>)-14-hydroxymanoyloxide-15,17-dien-15(<i>Z</i>)-16,19-olide		<i>Salvia mirzayanii</i> ²²⁸

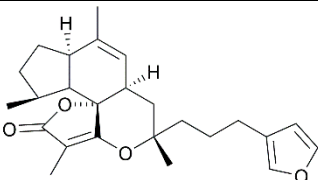
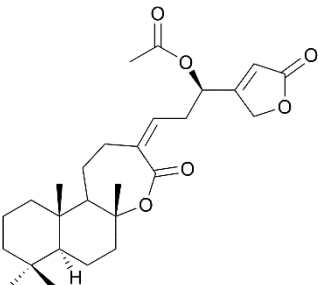
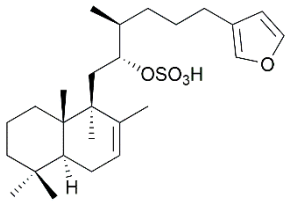
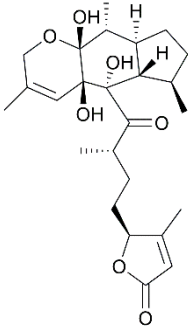
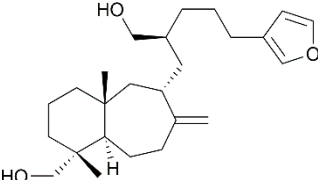
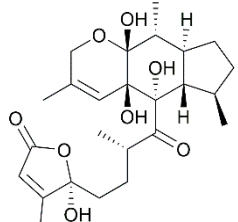
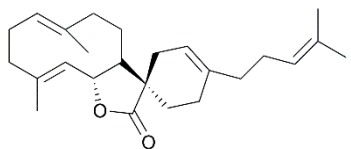
DC-124	(4 <i>R</i> ,5 <i>R</i> ,8 <i>R</i> ,9 <i>R</i> ,10 <i>S</i> ,13 <i>S</i> ,14 <i>R</i>)-14-hydroxymanoyloxide-15,17-dien-15(<i>Z</i>)-16,19-olide		<i>Salvia mirzayanii</i> ²²⁸
DC-125	(4 <i>R</i> ,5 <i>R</i> ,8 <i>R</i> ,9 <i>R</i> ,10 <i>S</i> ,13 <i>S</i> ,16 <i>R</i>)-14-hydroxymanoyloxide-17-en-16,19-olide		<i>Salvia mirzayanii</i> ²²⁸
DC-126	(4 <i>R</i> ,5 <i>R</i> ,8 <i>R</i> ,9 <i>R</i> ,10 <i>S</i> ,13 <i>R</i>)-manoyloxide-15,17-dien-15(<i>Z</i>)-16,19-olide		<i>Salvia mirzayanii</i> ²²⁸
DC-127	(13 <i>E</i>)-8α,15-dihydroxy-23-carboxymethylabd-13(14),17(18)-dien-16,19-olide		<i>Salvia tingitana</i> ²⁰⁰
DC-128	19-oxofasciospongine A		<i>Fasciospongia</i> sp. ¹⁹¹
DC-129	(15 <i>S</i> ,16 <i>S</i> ,13 <i>E</i>)-8α,15-dihydroxyabd-13(14),17(18)-dien-23,6α-16,19-diolide		<i>Salvia tingitana</i> ²⁰⁰ <i>Salvia dominica</i> ²⁰⁴
DC-130	(16 <i>R</i> ,13 <i>E</i>)-6α,8α,23-trihydroxyabd-13(14),17(18)-dien-16,19-olide		<i>Salvia tingitana</i> ²⁰⁰ <i>Salvia dominica</i> ²⁰⁴

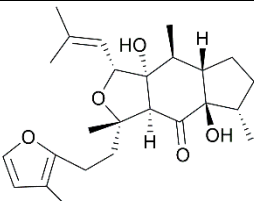
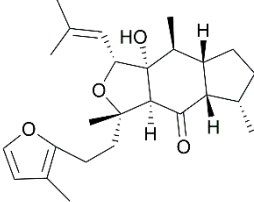
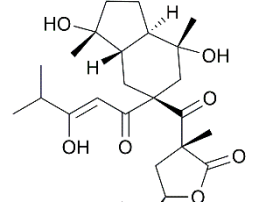
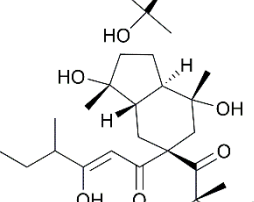
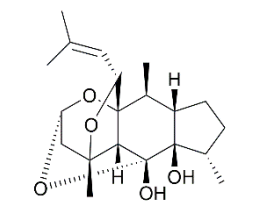
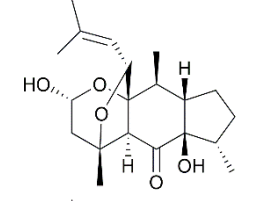
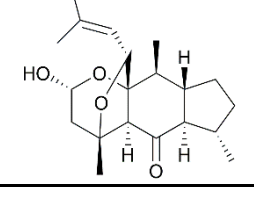
DC-131	(15 <i>S</i> ,16 <i>S</i> ,13 <i>E</i>)-6 α ,8 α ,15-trihydroxy-23-carboxymethylabd-13(14),17(18)-dien-16,19-olide		<i>Salvia tingitana</i> ²⁰⁰ <i>Salvia dominica</i> ²⁰⁴
DC-132	(13 <i>E</i>)-4 α ,6 α ,8 α -trihydroxyabd-13(14),17(18)-dien-16,19-olide		<i>Salvia tingitana</i> ²⁰⁰
DC-133	yosgadensenol		<i>Salvia yosgadensis</i> ¹⁹⁵
DC-134	yosgadensolide A		<i>Salvia yosgadensis</i> ²²⁹
DC-135	yosgadensolide B		<i>Salvia yosgadensis</i> ²²⁹
DC-136	6-dehydroxy-13- <i>epi</i> -yosgadensenol		<i>Salvia limbata</i> ²³⁰
DC-137	6-dehydroxyyosgadensenol		<i>Salvia limbata</i> ²³⁰
DC-138	leucosceptroid J		<i>Leucosceptрум canum</i> ²³¹

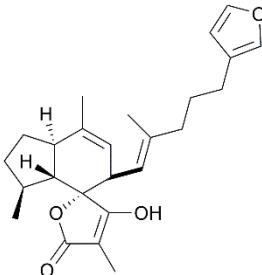
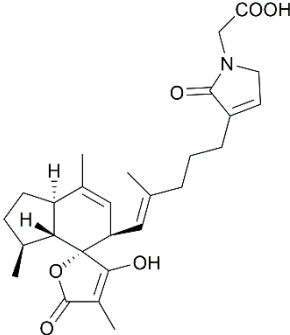
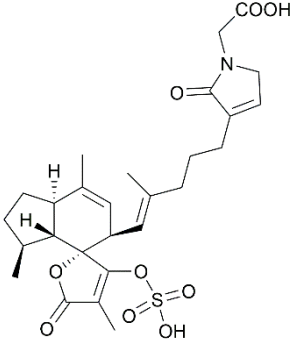
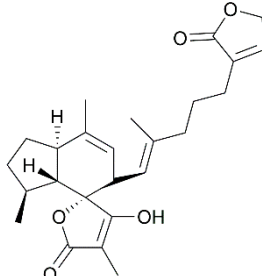
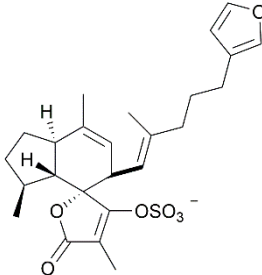
DC-139	(14 <i>E</i>)-methylmanoyloxide-14,16,18-trien-19,16-oxide-23-carboxylate		<i>Salvia tingitana</i> ²⁰⁰
DC-140	6α-hydroxymanoyloxide-14,17-dien-16,19-olide		<i>Salvia limbata</i> ²³⁰
DC-141	3β-hydroxymanoyloxide-14,17-dien-16,19-olide		<i>Salvia limbata</i> ²³⁰
DC-142	(13α),6α-hydroxymanoyloxide-14,17-dien-16,19-olide		<i>Salvia limbata</i> ²³⁰
DC-143	(13 <i>E</i>)-8α-hydroxy-23α- <i>O</i> -methyl-23,6α-epoxylabd-13(14),17-(18)-dien-16,19-olide		<i>Salvia tingitana</i> ²⁰⁰
DC-144	(13 <i>E</i>)-8α,15-dihydroxy-23α- <i>O</i> -methyl-23,6α-epoxylabd-13(14),17(18)-dien-16,19-olide		<i>Salvia tingitana</i> ²⁰⁰
DC-145	(13 <i>E</i>)-8α,23α-dihydroxy-23,6α-epoxylabd-13(14),17(18)-dien-16,19-olide		<i>Salvia tingitana</i> ²⁰⁰

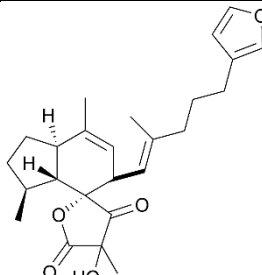
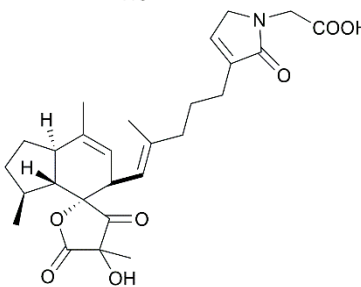
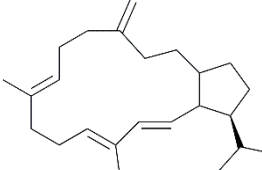
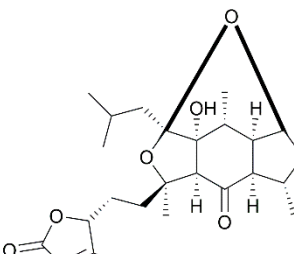
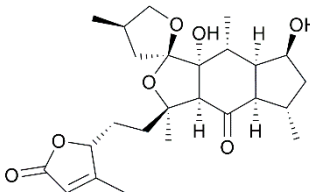
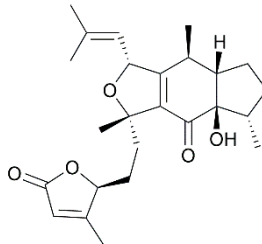
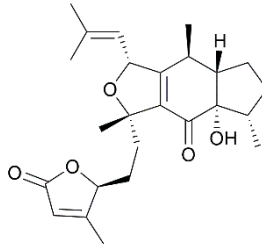
DC-146	(13 <i>E</i>)-8 α ,23-dihydroxyabd-13(14),17(18)-dien-16,19-olide		<i>Salvia tingitana</i> ²⁰⁰
DC-147	(13 <i>E</i>)-labd-13(14),17(18)-dien-8 α ,16,19-triol		<i>Salvia tingitana</i> ²⁰⁰
DC-148	trunculin A		<i>Latrunculia brevis</i> ²³²
DC-149	trunculin B		<i>Latrunculia brevis</i> ²³²
DC-150	trunculin C methyl ester		<i>Latrunculia brevis</i> ²³³
DC-151	trunculin D methyl ester		<i>Latrunculia brevis</i> ²³³
DC-152	trunculin E		<i>Latrunculia brevis</i> ²³³
DC-153	trunculin F		<i>Latrunculia conulosa</i> ²³⁴

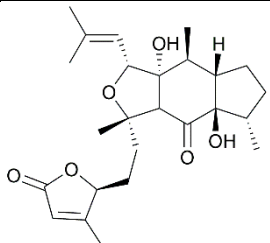
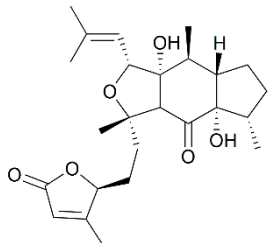
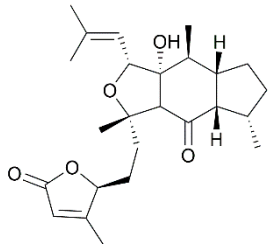
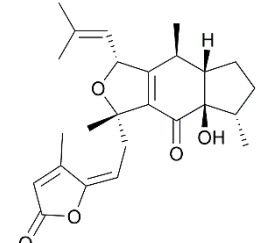
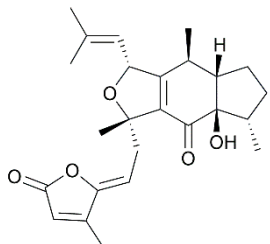
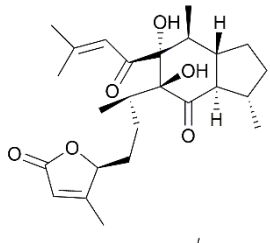
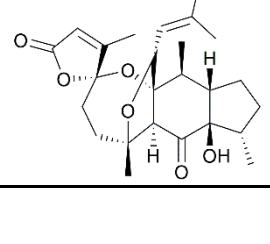
DC-154	trunculin H		<i>Latrunculia</i> sp. ²³⁵
DC-155	trunculin G		<i>Latrunculia</i> sp. ²³⁵
DC-156	trunculin I		<i>Latrunculia</i> sp. ²³⁵
DC-157	trunculin X		<i>Sigmosceptrella</i> sp. ²³⁶
DC-158	trunculin Y		<i>Sigmosceptrella</i> sp. ²³⁶
DC-159	ircinianin		<i>Ircinia wistarii</i> ^{237, 238}
DC-160	(+)-wistarin		<i>Ircinia wistarii</i> ²³⁷

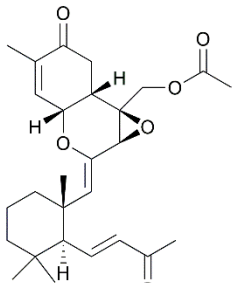
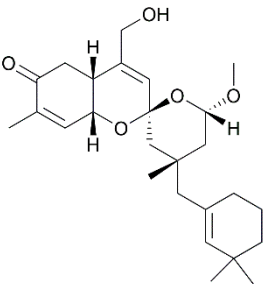
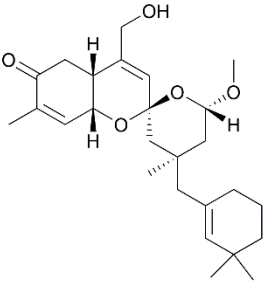
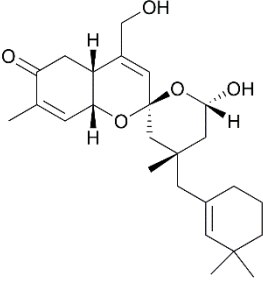
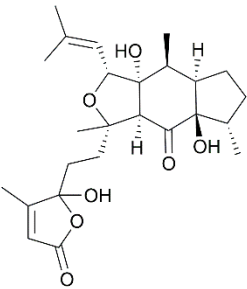
DC-161	(-)-wistarin		<i>Ircinia</i> sp. ²³⁹
DC-162	luffalactone		<i>Luffariella variabilis</i> ¹³²
DC-163	sulfircin		<i>Ircinia</i> sp. ²⁴⁰
DC-164	leucosesterlactone		<i>Leucosceptrum canum</i> ²⁴¹
DC-165	shinsonefuran		<i>Stoebea extensa</i> ²⁴²
DC-166	17 α -hydroxyleucosceptrine		<i>Leucosceptrum canum</i> ²⁴³
DC-167	genepolide		<i>Artemisia umbelliformis</i> ²⁴⁴

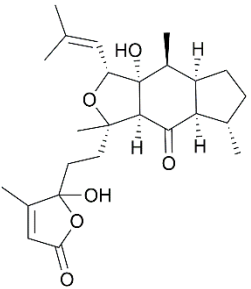
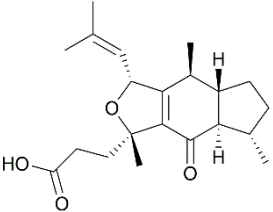
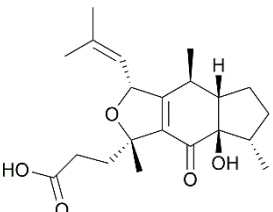
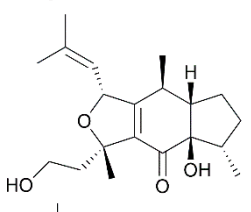
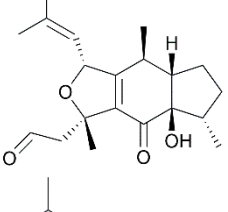
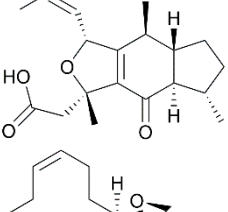
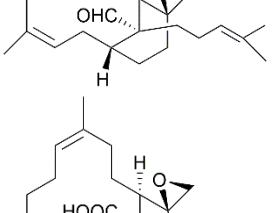
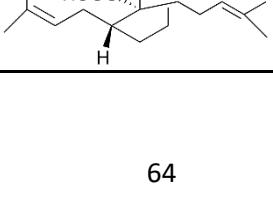
DC-168	leucosceptroid A		<i>Leucosceptrum canum</i> ²⁴⁵
DC-169	leucosceptroid B		<i>Leucosceptrum canum</i> ²⁴⁵
DC-170	biyoulactone D		<i>Hypericum chinense</i> ²⁴⁶
DC-171	biyoulactone E		<i>Hypericum chinense</i> ²⁴⁶
DC-172	norleucosceptroid A		<i>Leucosceptrum canum</i> ²⁴⁷
DC-173	norleucosceptroid B		<i>Leucosceptrum canum</i> ²⁴⁷
DC-174	norleucosceptroid C		<i>Leucosceptrum canum</i> ²⁴⁷

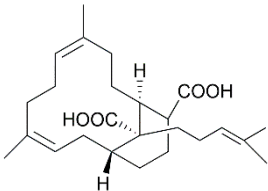
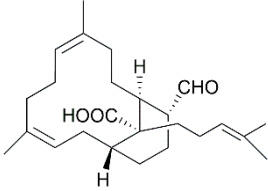
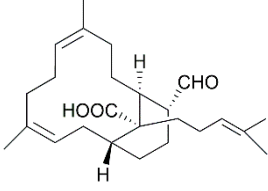
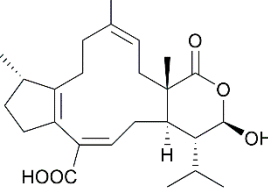
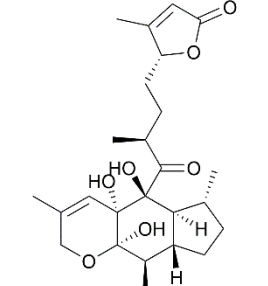
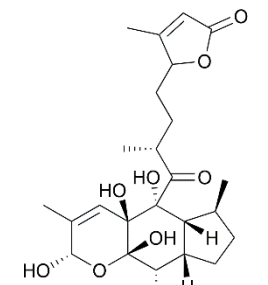
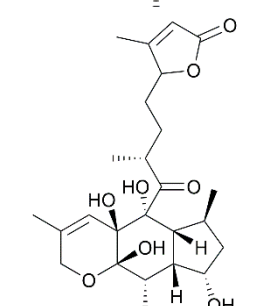
DC-175	(-)-ircinianin		<i>Ircinia wistarii</i> ^{237, 238} <i>Psammocinia</i> sp. CMB-01008, <i>Psammocinia</i> sp. CMB-03344 ²⁴⁸
DC-176	(-)-ircinianin lactam A		<i>Psammocinia</i> sp. CMB-03344 ²⁴⁸
DC-177	(-)-ircinianin lactam A sulfate		<i>Psammocinia</i> sp. CMB-01008 ²⁴⁸
DC-178	(-)-ircinianin lactone A		<i>Psammocinia</i> sp. CMB-02858 ²⁴⁸
DC-179	(-)-ircinianin sulfate		<i>Ircinia wistarii</i> ²⁴⁹ <i>Psammocinia</i> sp. CMB-01008, <i>Psammocinia</i> sp. CMB-03344 ²⁴⁸

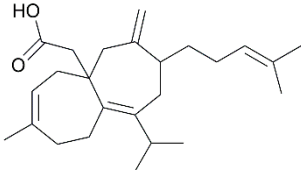
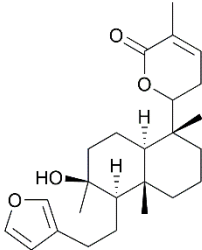
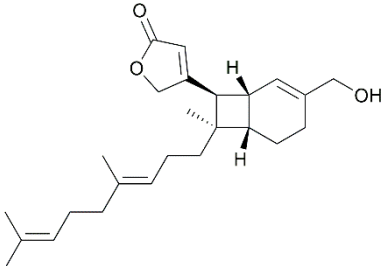
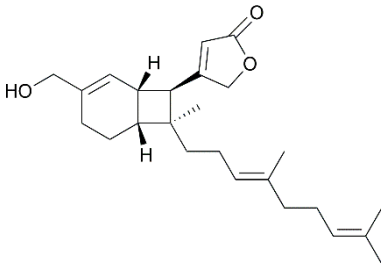
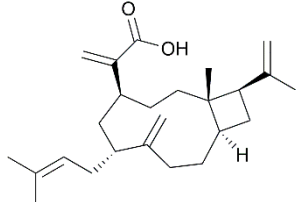
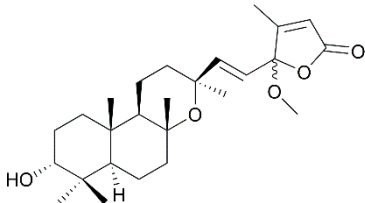
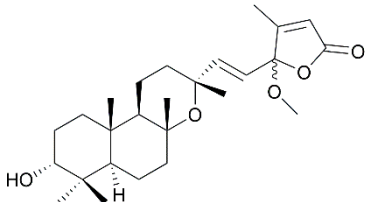
DC-180	(-)-oxoircinianin		<i>Psammocinia</i> sp. CMB-03344 ²⁴⁸
DC-181	(-)-oxoircinianin lactam A		<i>Psammocinia</i> sp. CMB-02858 ²⁴⁸
DC-182	sestermobaraene E		<i>Streptomyces mobaraensis</i> ²⁵⁰
DC-183	colquhounoid B		<i>Colquhounia coccinea</i> var. <i>mollis</i> ²⁵¹
DC-184	colquhounoid C		<i>Colquhounia coccinea</i> var. <i>mollis</i> ²⁵¹
DC-185	leucosceptroid E		<i>Leucosceptрум canum</i> ²³¹
DC-186	leucosceptroid F		<i>Leucosceptрум canum</i> ²³¹

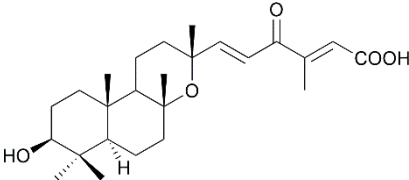
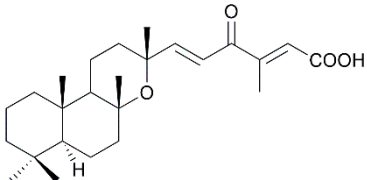
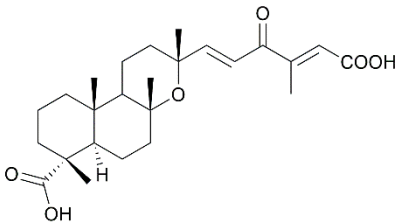
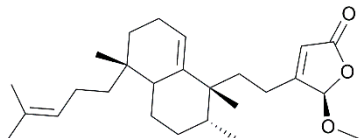
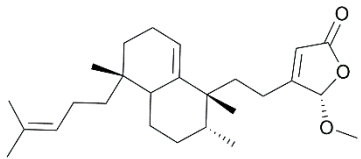
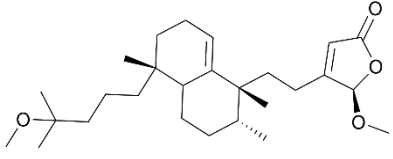
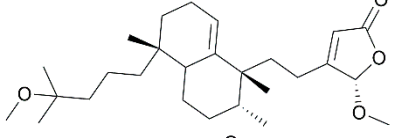
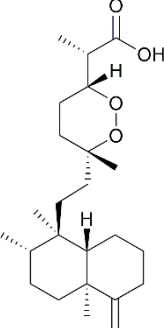
DC-187	leucosceptroid G		<i>Leucosceptrum canum</i> ²³¹
DC-188	leucosceptroid H		<i>Leucosceptrum canum</i> ²³¹
DC-189	leucosceptroid I		<i>Leucosceptrum canum</i> ²³¹
DC-190	leucosceptroid L		<i>Leucosceptrum canum</i> ²³¹
DC-191	leucosceptroid M		<i>Leucosceptrum canum</i> ²³¹
DC-192	leucosceptroid N		<i>Leucosceptrum canum</i> ²³¹
DC-193	leucosceptroid O		<i>Leucosceptrum canum</i> ²⁵²

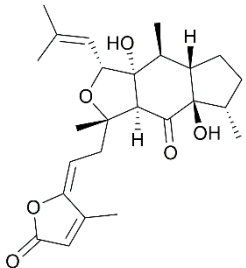
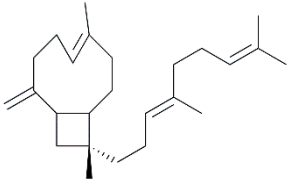
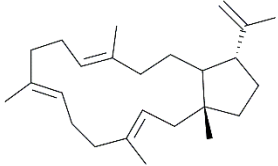
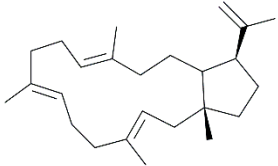
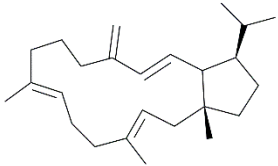
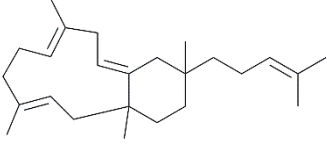
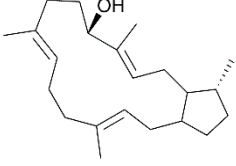
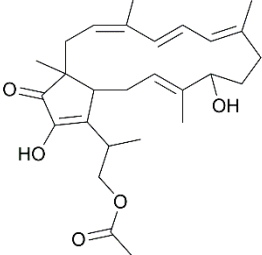
DC-194	secoepoxyansellone A		<i>Phorbas</i> sp. ¹⁶²
DC-195	gombaspiroketal A		<i>Clathria gombawuiensis</i> ²⁵³ <i>Haliclona</i> sp. ¹⁹⁶
DC-196	gombaspiroketal B		<i>Clathria gombawuiensis</i> ²⁵³ <i>Haliclona</i> sp. ¹⁹⁶
DC-197	gombaspiroketal C		<i>Clathria gombawuiensis</i> ²⁵³ <i>Haliclona</i> sp. ¹⁹⁶
DC-198	leucosceptroid P		<i>Leucosceptrum canum</i> ²⁵⁴

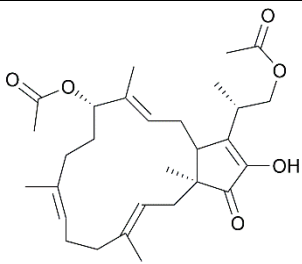
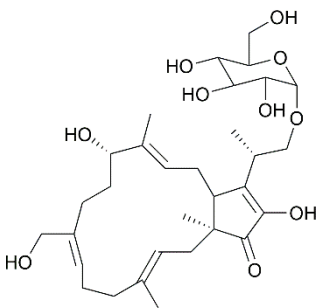
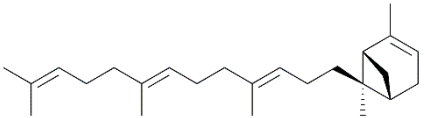
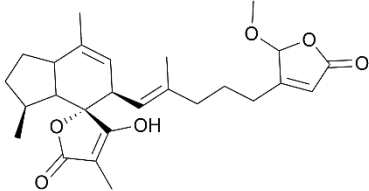
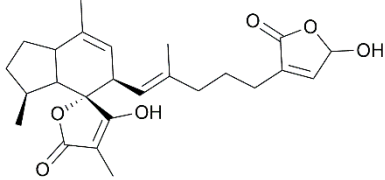
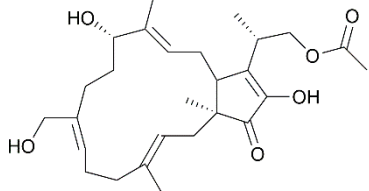
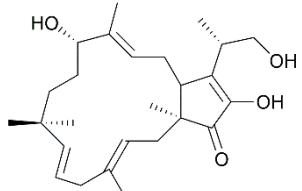
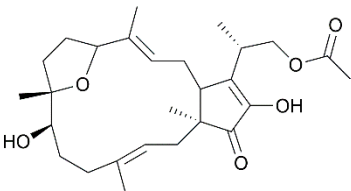
DC-199	leucosceptroid Q		<i>Leucosceptrum canum</i> ²⁵⁴
DC-200	norleucosceptroid D		<i>Leucosceptrum canum</i> ²⁵⁴
DC-201	norleucosceptroid E		<i>Leucosceptrum canum</i> ²⁵⁴
DC-202	norleucosceptroid F		<i>Leucosceptrum canum</i> ²⁵⁴
DC-203	norleucosceptroid G		<i>Leucosceptrum canum</i> ²⁵⁴
DC-204	norleucosceptroid H		<i>Leucosceptrum canum</i> ²⁵⁴
DC-205	emericellene A		<i>Emericella</i> sp. AST0036 ²⁵⁵
DC-206	emericellene B		<i>Emericella</i> sp. AST0036 ²⁵⁵

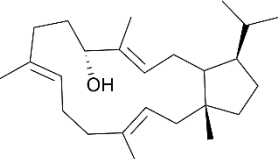
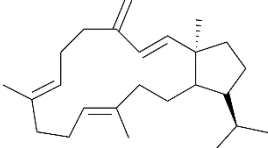
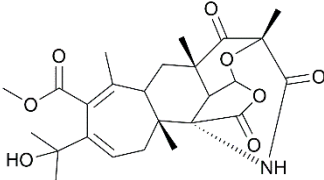
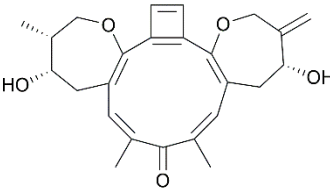
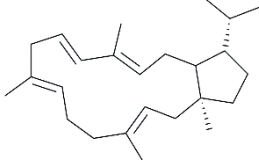
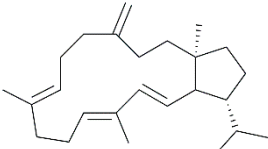
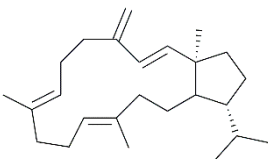
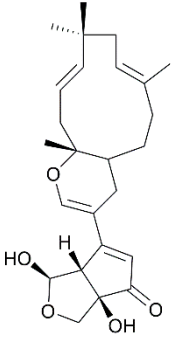
DC-207	emericlellene C		<i>Emericella</i> sp. AST0036 ²⁵⁵
DC-208	emericlellene D		<i>Emericella</i> sp. AST0036 ²⁵⁵
DC-209	emericlellene E		<i>Emericella</i> sp. AST0036 ²⁵⁵
DC-210	bipolarenic acid		<i>Lophiostoma bipolare</i> ²⁵⁶
DC-211	leucosceptrine		<i>Leucosceptrum canum</i> ²⁵⁷
DC-212	1 α -hydroxyleucosceptrine		From microbial transformation on leucosceptrine by <i>Rhizopus stolonifer</i> ²⁵⁸
DC-213	8 α -hydroxyleucosceptrine		From microbial transformation on leucosceptrine by <i>Rhizopus stolonifer</i> ²⁵⁸

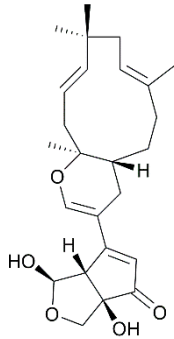
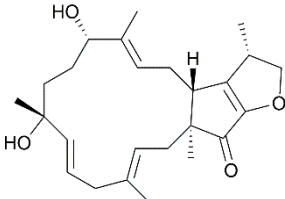
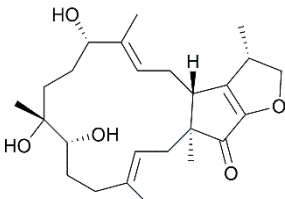
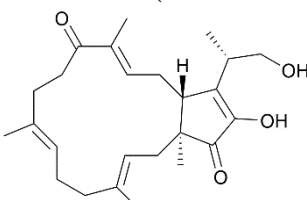
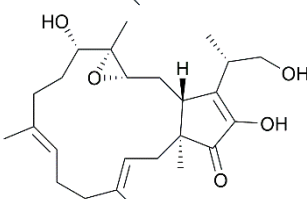
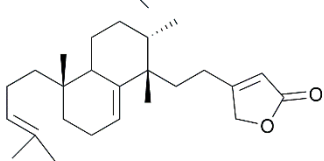
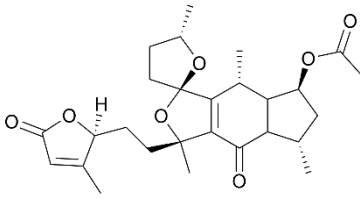
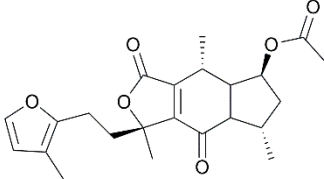
DC-214	goniocarpic acid		<i>Serjania goniocarpa</i> ²⁵⁹
DC-215	unnamed dicarbocyclic sesterterpene (reported as compound 2a)		<i>Luffariella variabilis</i> ⁹⁷
DC-216	(-)-hippolide J		<i>Hippospongia lachne</i> ²⁶⁰
DC-217	(+)-hippolide J		<i>Hippospongia lachne</i> ²⁶⁰
DC-218	raoulic acid		<i>Raoulia australis</i> ²⁶¹
DC-219	3- <i>epi</i> -salviaethiopisolid and C-16 epimer		<i>Salvia aethiopis</i> ²⁶²
DC-220	salviaethiopisolid and C-16 epimer		<i>Salvia aethiopis</i> ²⁶²

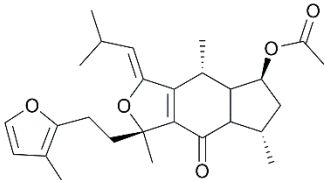
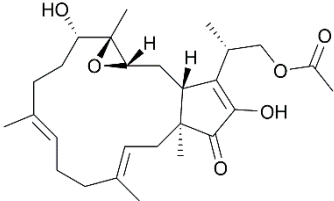
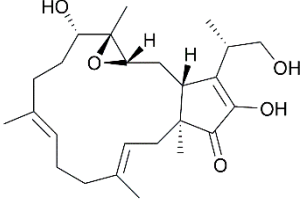
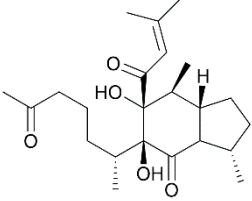
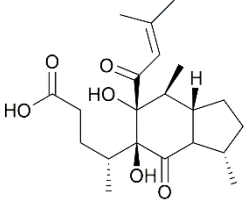
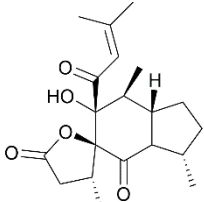
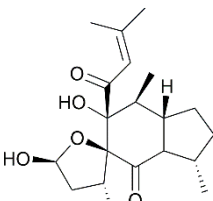
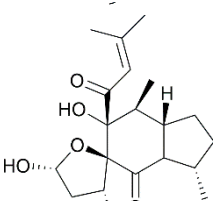
DC-221	3 α -hydroxymanoyloxide-14(<i>E</i>),17-dien-16-oxo-19-oic acid		<i>Salvia aethiopis</i> ²⁶³
DC-222	hydroxymanoyloxide-14,17-dien-16-oxo-19-oic acid		<i>Salvia aethiopis</i> ²⁶³
DC-223	hydroxymanoyloxide-14,17-dien-16-oxo-19,23-dioic acid		<i>Salvia aethiopis</i> ²⁶³
DC-224	dactylospene B		<i>Dactylosporgia elegans</i> ⁵⁰
DC-225	dactylospene C		<i>Dactylosporgia elegans</i> ⁵⁰
DC-226	dactylospene D		<i>Dactylosporgia elegans</i> ⁵⁰
DC-227	dactylospene E		<i>Dactylosporgia elegans</i> ⁵⁰
DC-228	enantio sigmosceptrellin-A methyl ester		Unidentified sponge ¹²¹

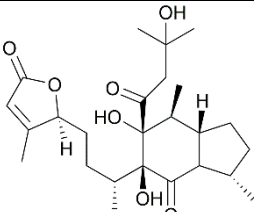
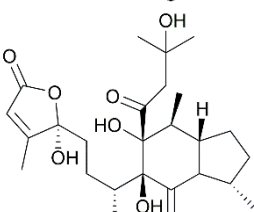
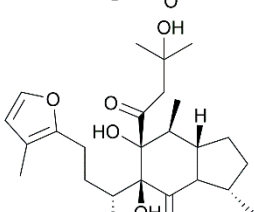
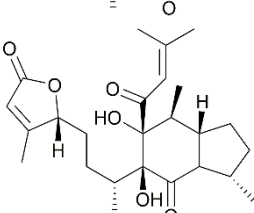
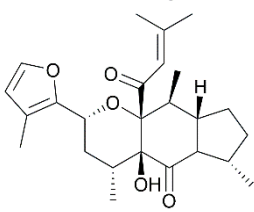
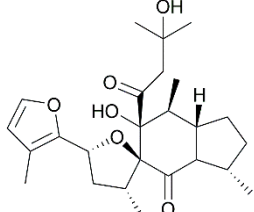
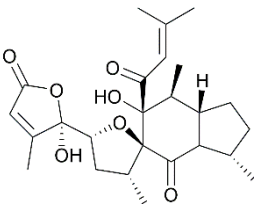
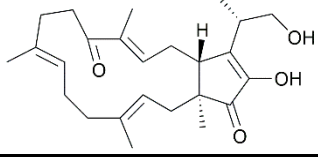
DC-229	leucosceptroid K		<i>Leucoscepttrum canum</i> ²³¹
DC-230	clavaphyllene		<i>Aspergillus clavatus</i> ²⁶⁴
DC-231	(+)-brassitetraene A		<i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁵
DC-232	(+)-brassitetraene B		<i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁵
DC-233	fusaproliferene		<i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁶
DC-234	variecoltetraene (reported as compound 2)		<i>Escherichia coli</i> ¹⁶⁰
DC-235	Bm3		<i>Bipolaris maydis</i> ²⁶⁷
DC-236	fusaproliferin. Wrong structure. See correct structure at DC-112		<i>Fusarium subglutinans</i> ²⁶⁸

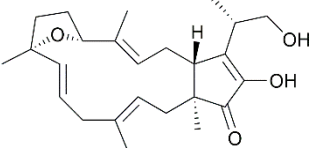
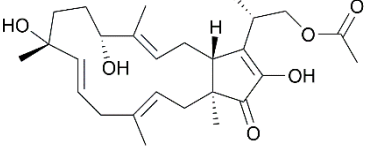
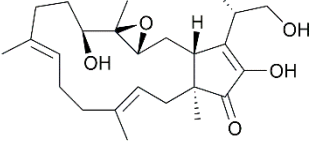
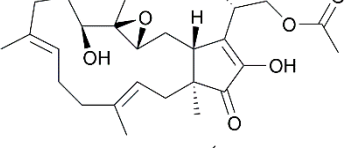
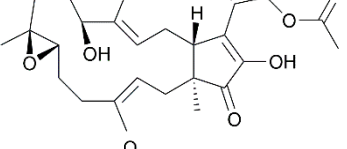
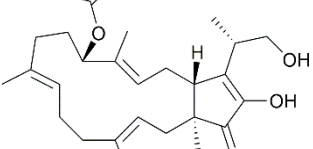
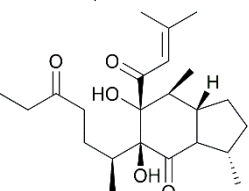
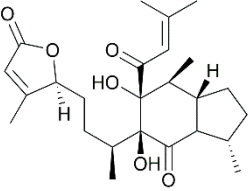
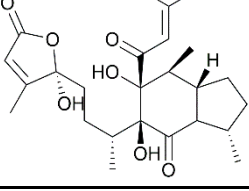
DC-237	sestersorokinicin A		<i>Bipolaris sorokiniana</i> / <i>Poa pratensis</i> ²⁶⁹
DC-238	sestersorokiniside A		<i>Bipolaris sorokiniana</i> / <i>Poa pratensis</i> ²⁶⁹
DC-239	somaliensene A		<i>Streptomyces somaliensis</i> ¹⁷⁰
DC-240	ircinianin lactone B		<i>Ircinia wistarii</i> ²⁷⁰
DC-241	ircinianin lactone C		<i>Ircinia wistarii</i> ²⁷⁰
DC-242	bipolariterpene A		<i>Bipolaris</i> sp. ²⁷¹
DC-243	bipolariterpene B		<i>Bipolaris</i> sp. ²⁷¹
DC-244	bipolariterpene C		<i>Bipolaris</i> sp. ²⁷¹

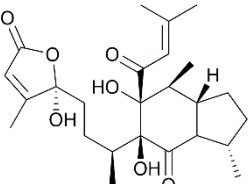
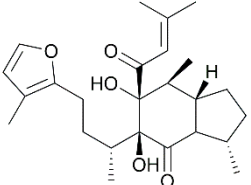
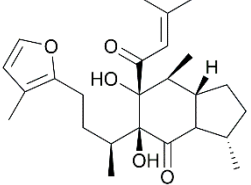
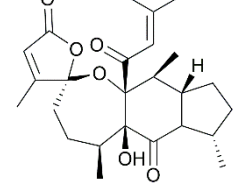
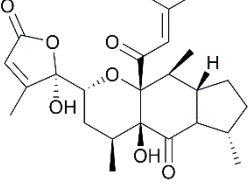
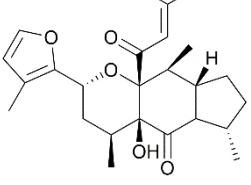
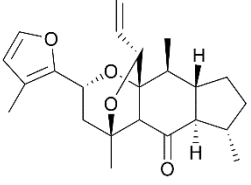
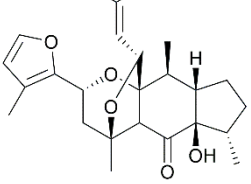
DC-245	preterpestacin I		<i>Alternaria alternata</i> MB-30/ <i>Leucosceptum canum</i> ²⁷²
DC-246	sesterviolene G		<i>Streptomyces violens</i> ²⁷³
DC-247	aspergstressin		<i>Aspergillus</i> sp. ²⁷⁴
DC-248	unnamed dicarbocyclic sesterterpene		<i>Artemisia frigida</i> ²⁷⁵
DC-249	sesterorbiculene		<i>Escherichia coli</i> strain DH10B/ <i>Saccharomyces cerevisiae</i> YZL141 ¹⁵⁹
DC-250	sesterviolene E		<i>Streptomyces violens</i> ²⁷³
DC-251	sesterviolene F		<i>Streptomyces violens</i> ²⁷³
DC-252	patriniaterpene C		<i>Patrinia scabra</i> ²⁷⁶

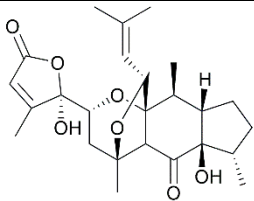
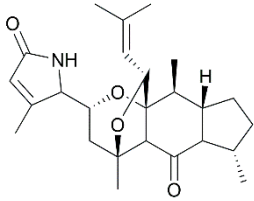
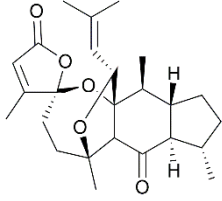
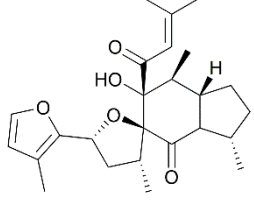
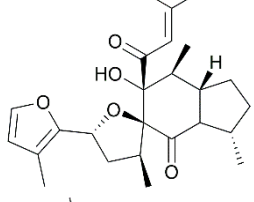
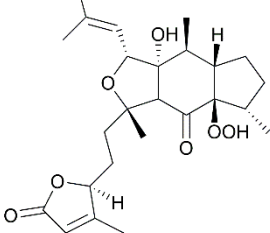
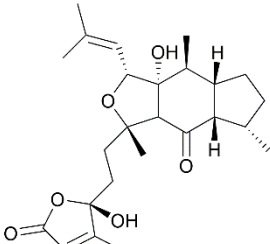
DC-253	patriniaterpene D		<i>Patrinia scabra</i> ²⁷⁶
DC-254	arthroliferin A		<i>Arthrinium</i> sp. SCSIO41221 ²⁷⁷
DC-255	arthroliferin B		<i>Arthrinium</i> sp. SCSIO41221 ²⁷⁷
DC-256	arthroliferin C		<i>Arthrinium</i> sp. SCSIO41221 ²⁷⁷
DC-257	arthroliferin D		<i>Arthrinium</i> sp. SCSIO41221 ²⁷⁷
DC-258	dactylospene F		<i>Sarcotragus</i> sp. ²⁷⁸
DC-259	colquhounoid E		<i>Colquhounia coccinea</i> var. <i>mollis</i> ²⁷⁹
DC-260	norcolquhounoid F		<i>Colquhounia coccinea</i> var. <i>mollis</i> ²⁷⁹

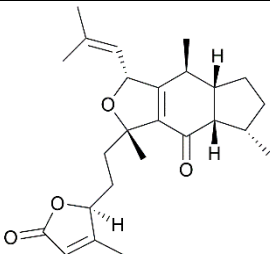
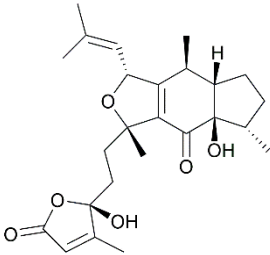
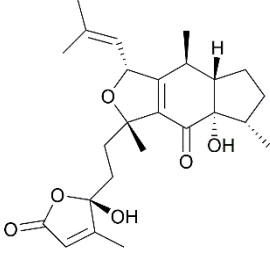
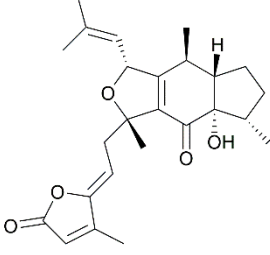
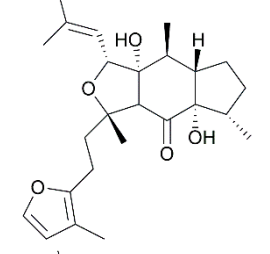
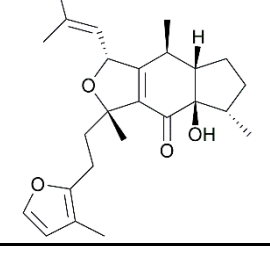
DC-261	colquhounoids F		<i>Colquhounia coccinea</i> var. <i>mollis</i> ²⁷⁹
DC-262	bipolariterpene D		<i>Bipolaris eleusines</i> ²⁸⁰
DC-263	bipolariterpene E		<i>Bipolaris eleusines</i> ²⁸⁰
DC-264	<i>nor</i> -leucosceptroid L		<i>Leucoscepttrum canum</i> ¹⁷²
DC-265	<i>nor</i> -leucosceptroid M		<i>Leucoscepttrum canum</i> ¹⁷²
DC-266	<i>epi-nor</i> -leucosceptroid J		<i>Leucoscepttrum canum</i> ¹⁷²
DC-267	<i>nor</i> -leucosceptroid N		<i>Leucoscepttrum canum</i> ¹⁷²
DC-268	16- <i>epi-nor</i> -leucosceptroid N		<i>Leucoscepttrum canum</i> ¹⁷²

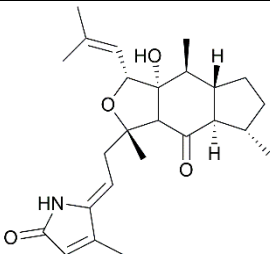
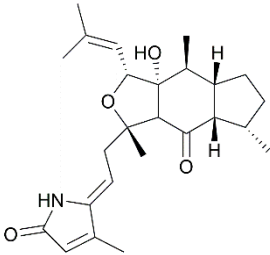
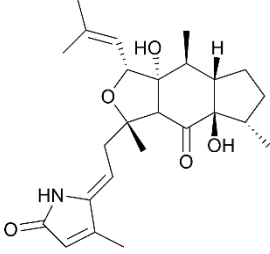
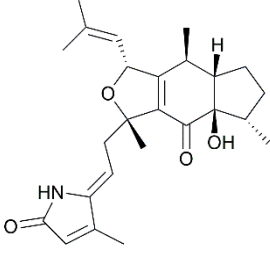
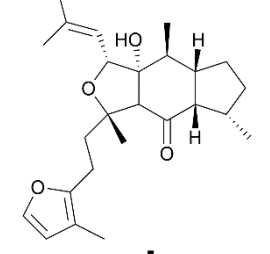
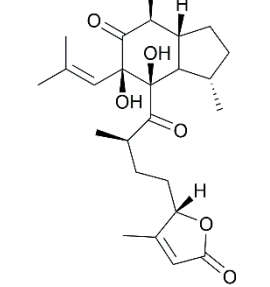
DC-269	3-H-2-hydroxy-leucosceptroid N		<i>Leucoscepttrum canum</i> ²⁸¹
DC-270	3-H-2,17 α -dihydroxy-leucosceptroid N		<i>Leucoscepttrum canum</i> ²⁸¹
DC-271	3-H-2-hydroxy-leucosceptroid R		<i>Leucoscepttrum canum</i> ²⁸¹
DC-272	17- <i>epi</i> -leucosceptroid N		<i>Leucoscepttrum canum</i> ²⁸¹
DC-273	5 α ,16 α -epoxy-leucosceptroid R		<i>Leucoscepttrum canum</i> ²⁸¹
DC-274	13 β ,16 α -epoxy-3-H-2-hydroxy-leucosceptroid R		<i>Leucoscepttrum canum</i> ²⁸¹
DC-275	13 β ,16 α -epoxy-17 α -hydroxy-leucosceptroid N		<i>Leucoscepttrum canum</i> ²⁸¹
DC-276	maydistacin A		<i>Bipolaris maydis</i> / <i>Hypericum longistylum</i> ²⁸²

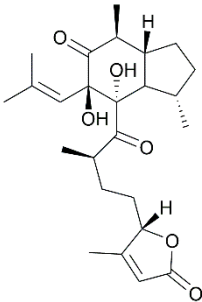
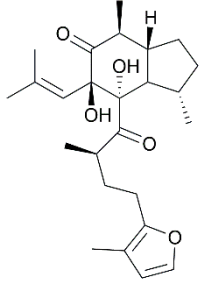
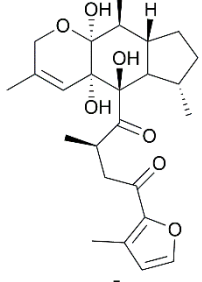
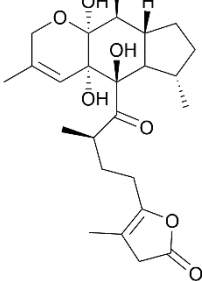
DC-277	maydistacin B		<i>Bipolaris maydis/Hypericum longistylum</i> 282
DC-278	maydistacin C		<i>Bipolaris maydis/Hypericum longistylum</i> 282
DC-279	maydistacin D		<i>Bipolaris maydis/Hypericum longistylum</i> 282
DC-280	maydistacin E		<i>Bipolaris maydis/Hypericum longistylum</i> 282
DC-281	maydistacin F		<i>Bipolaris maydis/Hypericum longistylum</i> 282
DC-282	maydistacin G		<i>Bipolaris maydis/Hypericum longistylum</i> 282
DC-283	norleucosceptroid I		<i>Leucoscepttrum canum</i> ²⁸¹
DC-284	14- <i>epi</i> -leucosceptroid N		<i>Leucoscepttrum canum</i> ²⁸¹
DC-285	17 α -hydroxy-leucosceptroid N		<i>Leucoscepttrum canum</i> ²⁸¹

DC-286	17 α -hydroxy-14- <i>epi</i> -leucosceptroid N		<i>Leucosceptrum canum</i> ²⁸¹
DC-287	leucosceptroid R		<i>Leucosceptrum canum</i> ²⁸¹
DC-288	14- <i>epi</i> -leucosceptroid R		<i>Leucosceptrum canum</i> ²⁸¹
DC-289	5 α ,17 β -epoxy-14- <i>epi</i> -leucosceptroid N		<i>Leucosceptrum canum</i> ²⁸¹
DC-290	5 α ,16 α -epoxy-17 α -hydroxy-14- <i>epi</i> -leucosceptroid N		<i>Leucosceptrum canum</i> ²⁸¹
DC-291	5 α ,16 α -epoxy-leucosceptroid R		<i>Leucosceptrum canum</i> ²⁸¹
DC-292	leucosceptroid S		<i>Leucosceptrum canum</i> ²⁸¹
DC-293	11 β -hydroxy-leucosceptroid S		<i>Leucosceptrum canum</i> ²⁸¹

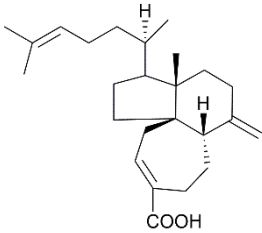
DC-294	leucosceptroid T		<i>Leucosceptrum canum</i> ²⁸¹
DC-295	leucosceplactam A		<i>Leucosceptrum canum</i> ²⁸¹
DC-296	5 α ,17 β -epoxy-leucosceptroid J		<i>Leucosceptrum canum</i> ²⁸¹
DC-297	13 β ,16 α -epoxy-leucosceptroid R		<i>Leucosceptrum canum</i> ²⁸¹
DC-298	13 β ,16 α -epoxy-14- <i>epi</i> -leucosceptroid R		<i>Leucosceptrum canum</i> ²⁸¹
DC-299	11 β -hydroperoxide-leucosceptroid I		<i>Leucosceptrum canum</i> ²⁸¹
DC-300	11- <i>epi</i> -leucosceptroid Q		<i>Leucosceptrum canum</i> ²⁸¹

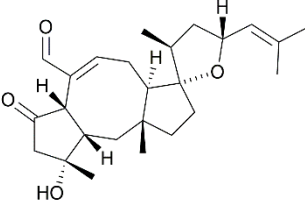
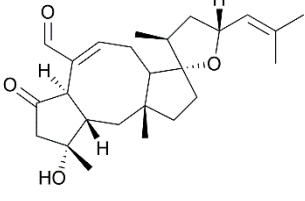
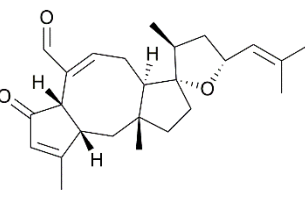
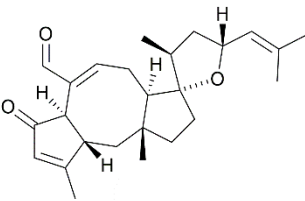
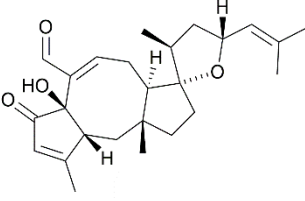
DC-301	13-dehydro-leucosceptroid I		<i>Leucosceptrum canum</i> ²⁸¹
DC-302	5,13-dehydro-leucosceptroid P		<i>Leucosceptrum canum</i> ²⁸¹
DC-303	5,13-dehydro-11- <i>epi</i> -leucosceptroid P		<i>Leucosceptrum canum</i> ²⁸¹
DC-304	11- <i>epi</i> -leucosceptroid M		<i>Leucosceptrum canum</i> ²⁸¹
DC-305	11- <i>epi</i> -leucosceptroid A		<i>Leucosceptrum canum</i> ²⁸¹
DC-306	5,13-dehydro-leucosceptroid A		<i>Leucosceptrum canum</i> ²⁸¹

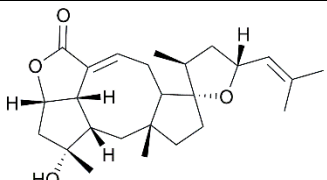
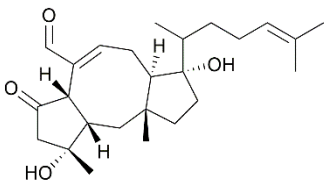
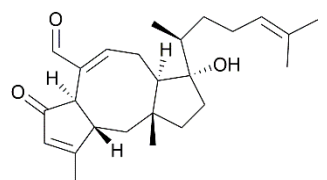
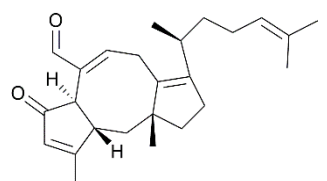
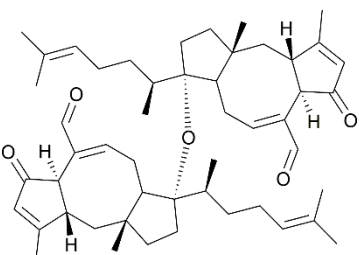
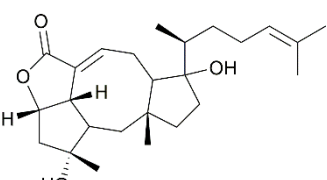
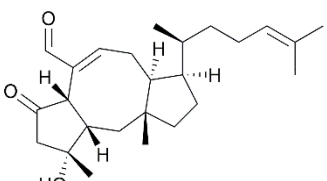
DC-307	leucosceplactam B		<i>Leucosceptrum canum</i> ²⁸¹
DC-308	leucosceplactam C		<i>Leucosceptrum canum</i> ²⁸¹
DC-309	leucosceplactam D		<i>Leucosceptrum canum</i> ²⁸¹
DC-310	leucosceplactam E		<i>Leucosceptrum canum</i> ²⁸¹
DC-311	11- <i>epi</i> -leucosceptroid B		<i>Leucosceptrum canum</i> ²⁸¹
DC-312	leucosceptrine A		<i>Leucosceptrum canum</i> ²⁸³

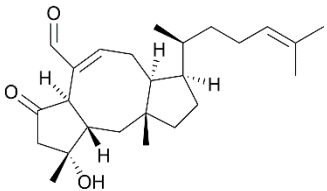
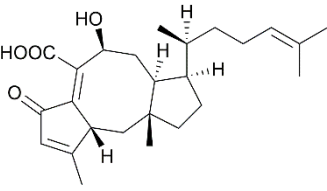
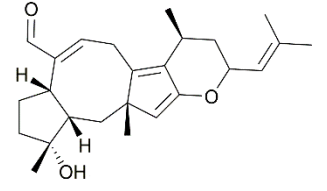
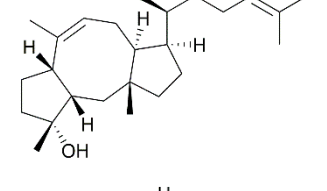
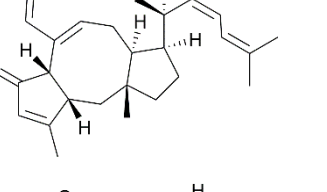
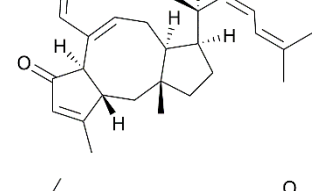
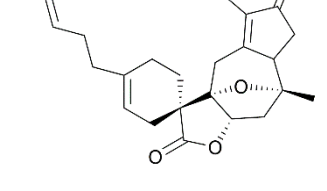
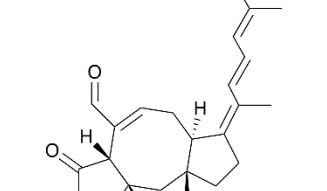
DC-313	leucosceptrine B		<i>Leucosceptrum canum</i> ²⁸³
DC-314	leucosceptrine C		<i>Leucosceptrum canum</i> ²⁸³
DC-315	leucosceptrine D		<i>Leucosceptrum canum</i> ²⁸³
DC-316	leucosceptrine E		<i>Leucosceptrum canum</i> ²⁸³

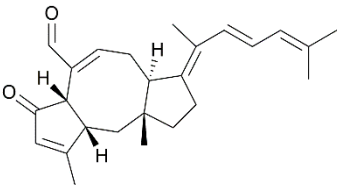
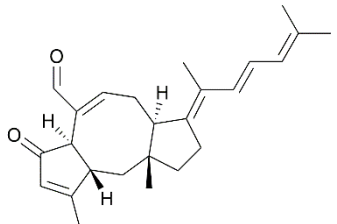
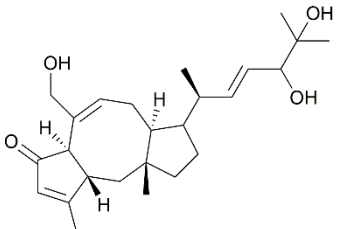
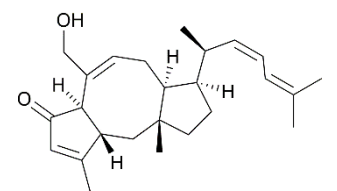
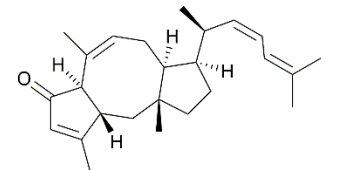
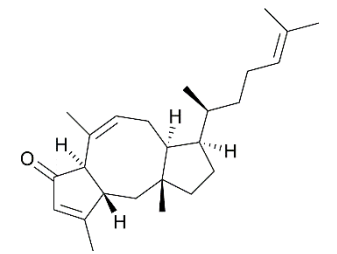
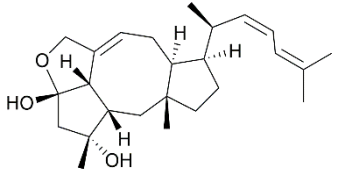
Section 1.7. Structures of tricarbocyclic (TrC) sesterterpenoids (with and without heterocycles)

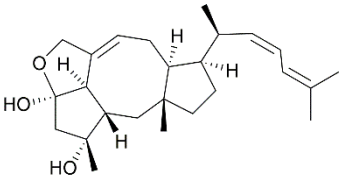
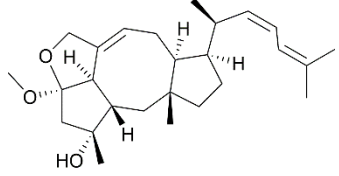
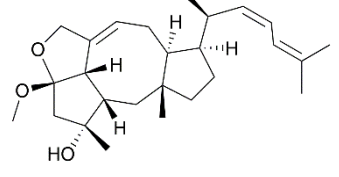
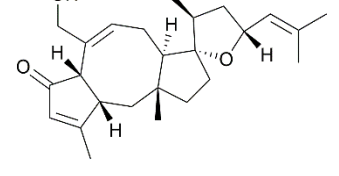
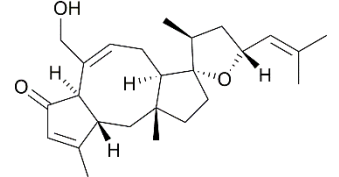
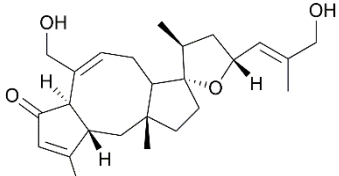
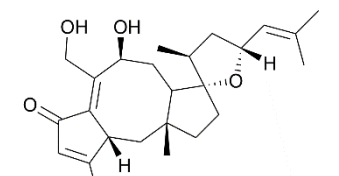
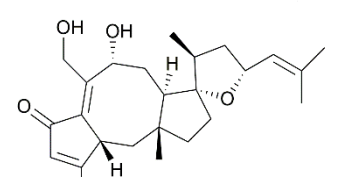
c. n.	IUPAC name or Trivial name	Structure	Organism as reported in the reference
TrC-1	gascardic acid		<i>Ceroplastes madagascariensis</i> (as <i>Gascardia madagascariensis</i>) ²⁸⁴⁻²⁸⁶

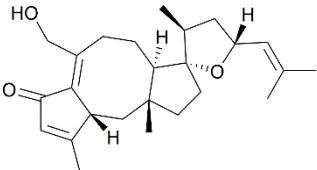
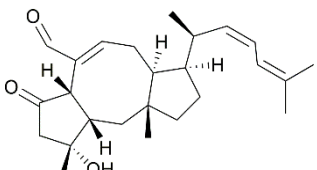
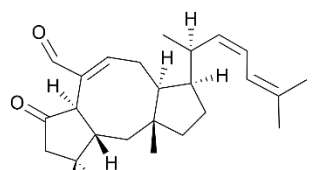
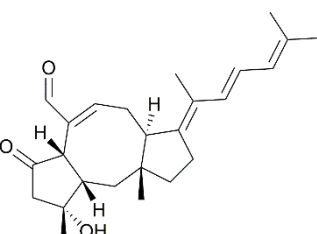
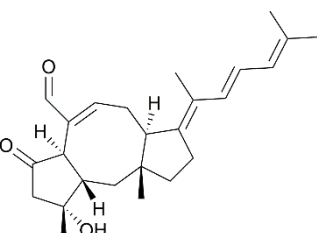
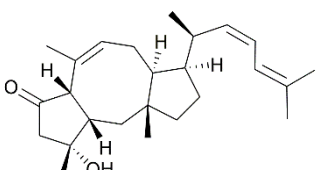
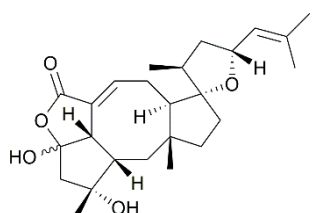
TrC-2	ophiobolin A (ophiobolin, ophiobalin, cochliobolin, cochliobolin A, zizanin) (ophiobola-7,18-dien-21-al-3α-ol-5-one-14α,17-oxide)		<i>Bipolaris leersiae</i> (as <i>Helminthosporium leersii</i>)²⁸⁷ <i>Bipolaris maydis</i> (as <i>Ophiobolus heterostrophus</i>, <i>Drechslera maydis</i>)^{287, 288} <i>Bipolaris oryzae</i> (as <i>Drechslera oryzae</i>, <i>Helminthosporium oryzae</i>, <i>Ophiobolus miyabeanus</i>)²⁸⁹⁻²⁹⁷ <i>Bipolaris panici-miliacei</i> (reported as <i>Helminthosporium panici-miliacei</i>)²⁸⁷ <i>Bipolaris setariae</i>²⁹⁸ <i>Bipolaris sorghicola</i> (also as <i>Drechslera sorghicola</i>)^{288, 299} <i>Cochliobolus heterostrophus</i>^{300, 301} <i>Cochliobolus miyabeanus</i>³⁰² <i>Drechslera gigantea</i> (from <i>Digitaria sanguinalis</i>)^{303, 304} <i>Drechslera zizaniae</i> (as <i>Helminthosporium zizaniae</i>)^{305, 306} <i>Exserohilum turcicum</i> (reported as <i>Helminthosporium turcicum</i>)³⁰⁷ <i>Helminthosporium</i> spp.³⁰⁸
TrC-3	6- <i>epi</i> -ophiobolin A		<i>Bipolaris maydis</i> (as <i>Drechslera maydis</i>)³⁰⁹ <i>Bipolaris oryzae</i> (also as <i>Drechslera oryzae</i>)^{295-297, 309} <i>Bipolaris sorghicola</i> (also as <i>Drechslera sorghicola</i>)^{288, 299} <i>Drechslera gigantea</i> (from <i>Digitaria sanguinalis</i>)^{303, 304} As microbial metabolite of ophiobolin A by <i>Penicillium patulum</i>³⁰⁰ <i>Helminthosporium</i> spp.³¹⁰
TrC-4	3-anhydrophiobolin A		<i>Bipolaris oryzae</i>^{295, 296, 311} <i>Bipolaris setariae</i>³¹² <i>Bipolaris sorghicola</i>²⁹⁹ <i>Cochliobolus heterostrophus</i>^{300, 301} <i>Helminthosporium</i> spp.³¹³
TrC-5	3-anhydro-6- <i>epi</i> -ophiobolin A		<i>Bipolaris</i> sp. TJ403-B1³¹⁴ <i>Bipolaris maydis</i> (as <i>Drechslera maydis</i>)^{288, 309} <i>Bipolaris oryzae</i> (also as <i>Drechslera oryzae</i>)^{295, 297, 311} <i>Bipolaris sorghicola</i>²⁹⁹ <i>Bipolaris sorghicola</i> (as <i>Drechslera sorghicola</i>)²⁸⁸ <i>Drechslera gigantea</i> (from <i>Digitaria sanguinalis</i>)³⁰⁴ <i>Cochliobolus heterostrophus</i>³⁰¹ <i>Helminthosporium</i> spp.³¹³ <i>Bipolaris oryzae</i>³¹¹
TrC-6	3-anhydro-6-hydroxyphiobolin A		

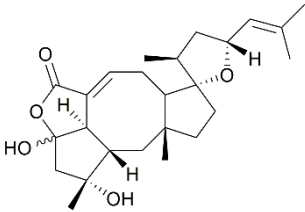
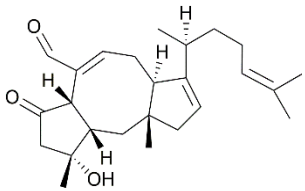
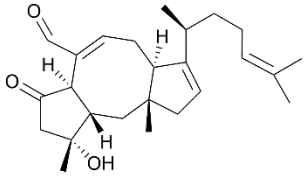
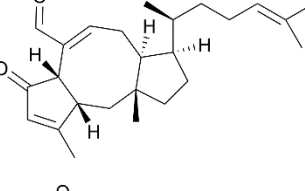
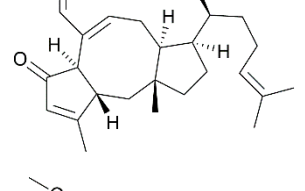
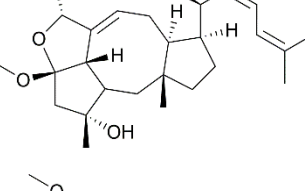
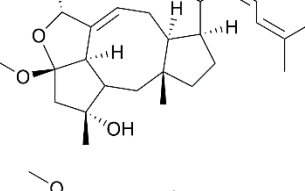
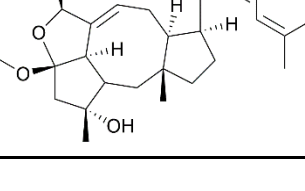
TrC-7	ophiobolin A lactone		As microbial metabolite of ophiobolin A by <i>Polyangium cellulorum</i> ³⁰⁰
TrC-8	ophiobolin B (cochliobolin B, ophiobolosin A, zizanin B) (ophiobola-7,18-dien-21-al-3 α ,14 α -diol-5-one)		<i>Bipolaris oryzae</i> (as <i>Drechslera oryzae</i> , <i>H. oryzae</i>) ^{295, 297, 315, 316} <i>Cochliobolus heterostrophus</i> ^{300, 301} <i>Cochliobolus miyabeanus</i> ³⁰² <i>Drechslera gigantea</i> ³⁰⁴ <i>Drechslera zizaniae</i> (as <i>Helminthosporium zizaniae</i>) ³⁰⁶ <i>Cochliobolus heterostrophus</i> ³⁰¹ <i>Drechslera gigantea</i> (from <i>Digitaria sanguinalis</i>) ^{303, 304}
TrC-9	3-anhydro-6- <i>epi</i> -ophiobolin B		<i>Cochliobolus heterostrophus</i> ³⁰¹ <i>Drechslera gigantea</i> (from <i>Digitaria sanguinalis</i>) ^{303, 304}
TrC-10	3-anhydro-6- <i>epi</i> - $\Delta^{10(14)}$ -ophiobolin B (degradation product)		<i>Cochliobolus heterostrophus</i> ³⁰¹
TrC-11	di-3-anhydro-6- <i>epi</i> -ophiobolin B (degradation product)		<i>Cochliobolus heterostrophus</i> ³⁰¹
TrC-12	ophiobolin B lactone		As microbial metabolite of ophiobolin A by <i>Pseudomonas aeruginosa</i> ³⁰⁰
TrC-13	ophiobolin C (zizanin A) (ophiobola-7,18-dien-21-al-3 α -ol-5-one)		<i>Aspergillus calidoustus</i> ³¹⁷ <i>Aspergillus insuetus</i> ³¹⁷ <i>Aspergillus flocculosus</i> ³¹⁸ <i>Bipolaris maydis</i> (as <i>Drechslera maydis</i>) ²⁸⁸ <i>Cochliobolus heterostrophus</i> (also as <i>Ophiobolus heterostrophus</i>) ^{306, 319} <i>Drechslera zizaniae</i> (as <i>Helminthosporium zizaniae</i>) ³⁰⁵ <i>Emericella varicolor</i> ³²⁰ <i>Mollisia</i> sp. ³²¹

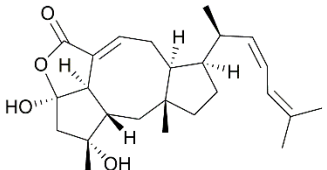
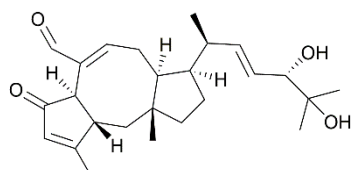
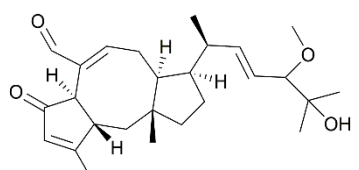
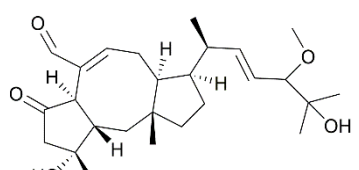
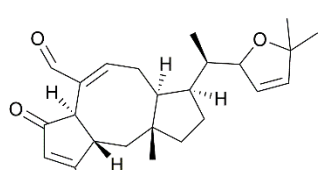
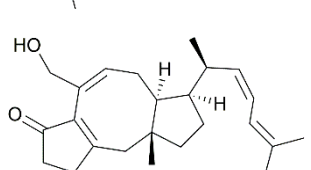
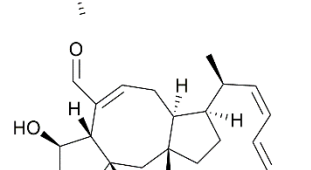
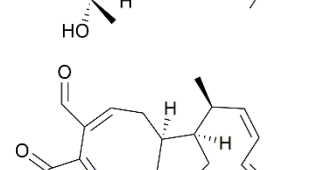
TrC-14	6- <i>epi</i> -ophiobolin C		<i>Aspergillus flocculosus</i> ³¹⁸ <i>Cochliobolus heterostrophus</i> ³¹⁹ <i>Emericella varicolor</i> ³²⁰
TrC-15	ophiobolin D (cephalonic acid) (ophiobola-3,6,18-trien-8 β -ol-21-oic acid)		<i>Cephalosporium caerulens</i> ³²²⁻³²⁴
TrC-16	ophiobolin E		<i>Drechslera gigantea</i> ^{303, 304}
TrC-17	ophiobolin F		<i>Bipolaris maydis</i> (as <i>Cochliobolus heterostrophus</i>) ^{4, 325} <i>Aspergillus ustus</i> ³²⁶
TrC-18	ophiobolin G		<i>Aspergillus</i> sp. ZJ-68 ³²⁷ <i>Aspergillus</i> sp. endophytic fungus from the body of <i>Zoanthus</i> ³²⁸ <i>Aspergillus ustus</i> ^{329, 330} <i>Emericella varicolor</i> ³²⁰
TrC-19	6- <i>epi</i> -ophiobolin G		<i>Aspergillus</i> sp. ZJ-68 ³²⁷ <i>Aspergillus calidoustus</i> ³¹⁷ <i>Aspergillus ustus</i> ³³⁰ <i>Emericella varicolor</i> ^{320, 331} <i>Ulocladium</i> sp. isolated from <i>Everniastrum</i> sp. ³³²
TrC-20	hedyosulide		<i>Hedyosmum brasiliense</i> ³³³
TrC-21	14,15-dehydro-(<i>Z</i>)-14-ophiobolin G		<i>Aspergillus flocculosus</i> ³¹⁸

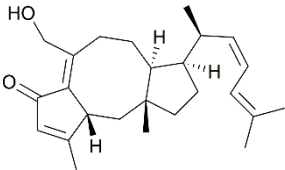
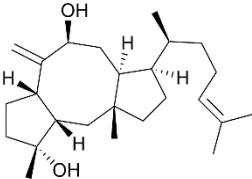
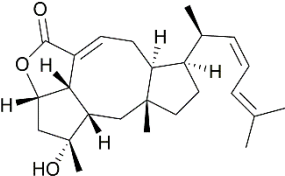
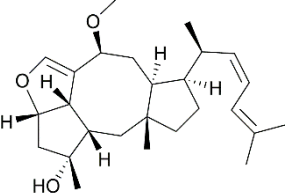
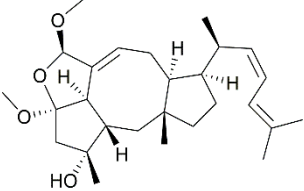
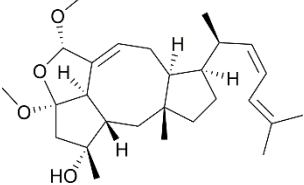
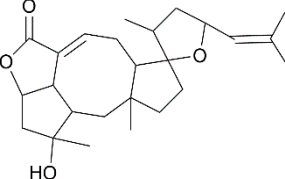
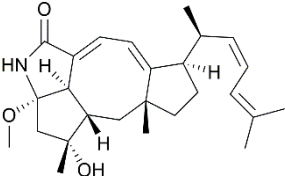
TrC-22	14,15-dehydroophiobolin G		<i>Aspergillus flocculosus</i> ³¹⁸
TrC-23	14,15-dehydro-6- <i>epi</i> - ophiobolin G		<i>Aspergillus flocculosus</i> ³¹⁸
TrC-24	(6 α)-18,19,21,21- <i>O</i> - tetrahydro-18,19- dihydroxyphiobolin G		<i>Aspergillus ustus</i> ³³⁴
TrC-25	(6 α)-21,21- <i>O</i> - dihydroophiobolin G or 21-deoxo-21-hydroxy-6- <i>epi</i> -ophiobolin G		<i>Aspergillus ustus</i> ^{326, 334} <i>Aspergillus</i> sp. ZJ-68 ³²⁷ <i>Ulocladium</i> sp. isolated from <i>Everniastrum</i> sp. ³³²
TrC-26	(6 α)-21-deoxyphiobolin G		<i>Aspergillus ustus</i> ³²⁶
TrC-27	(6 α)-16,17-dihydro-21- deoxyphiobolin G		<i>Aspergillus ustus</i> ³²⁶
TrC-28	ophiobolin H		<i>Aspergillus</i> sp. ZJ-68 ³²⁷ <i>Aspergillus</i> sp. endophytic fungus from the body of <i>Zoanthus</i> ³²⁸ <i>Aspergillus insuetus</i> ^{317, 335} <i>Aspergillus ustus</i> ^{326, 329, 330} <i>Aspergillus ustus</i> isolated from <i>Suberites</i> <i>domuncula</i> ³³⁶ <i>Emericella varicolor</i> ³²⁰

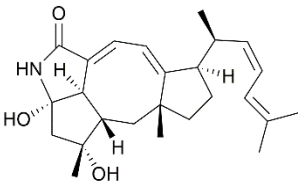
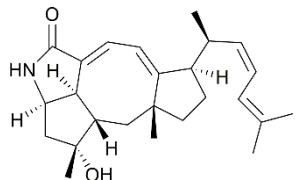
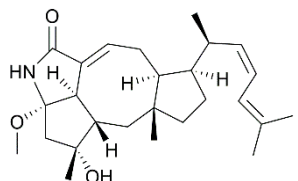
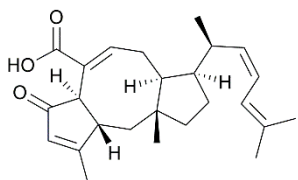
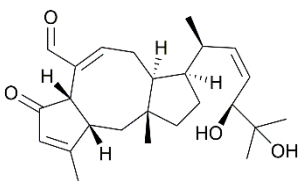
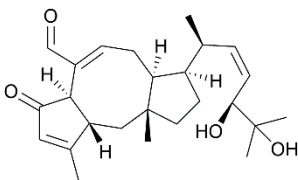
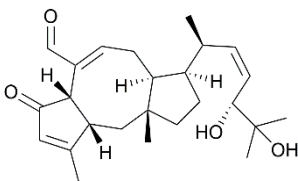
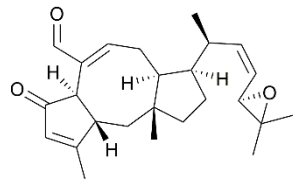
TrC-29	(5 α ,6 α)-ophiobolin H		<i>Aspergillus ustus</i> ^{326, 334} <i>Aspergillus</i> sp. ZJ-68 ³²⁷ <i>Aspergillus ustus</i> ³³⁰
TrC-30	(5 α ,6 α)-5- <i>O</i> -methylophiobolin H		<i>Aspergillus ustus</i> ³³⁴
TrC-31	5- <i>O</i> -methylophiobolin H		<i>Aspergillus ustus</i> ³³⁴
TrC-32	ophiobolin I		<i>Bipolaris oryzae</i> (also as <i>Drechslera oryzae</i>) ^{295-297, 337} <i>Bipolaris maydis</i> (as <i>Drechslera maydis</i>) ²⁸⁸ <i>Bipolaris setariae</i> ³¹² <i>Bipolaris sorghicola</i> (as <i>Drechslera sorghicola</i>) ²⁸⁸ <i>Cochliobolus heterostrophus</i> ³⁰¹ <i>Drechslera gigantea</i> (from <i>Digitaria sanguinalis</i>) ^{303, 304} As microbial metabolite of ophiobolin A by <i>Polyangium cellulosum</i> ³⁰⁰ <i>Bipolaris oryzae</i> (as <i>Drechslera oryzae</i>) ³³⁸
TrC-33	6- <i>epi</i> -ophiobolin I		
TrC-34	25-hydroxyophiobolin I		<i>Bipolaris oryzae</i> (as <i>Drechslera oryzae</i>) ²⁹⁷ <i>Bipolaris maydis</i> (as <i>Drechslera maydis</i>) ²⁸⁸ <i>Bipolaris sorghicola</i> (as <i>Drechslera sorghicola</i>) ²⁸⁸
TrC-35	ophiobolin J		<i>Bipolaris oryzae</i> (as <i>Drechslera oryzae</i>) ³³⁸ <i>Bipolaris</i> sp. TJ403-B1 ³¹⁴ <i>Drechslera gigantea</i> ³⁰⁴
TrC-36	8- <i>epi</i> -ophiobolin J		<i>Bipolaris</i> sp. TJ403-B1 ³¹⁴ <i>Drechslera gigantea</i> ³⁰⁴

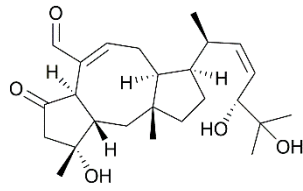
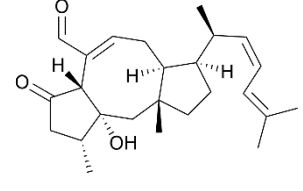
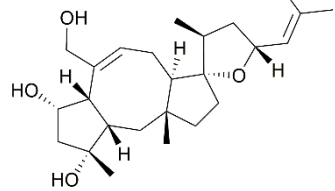
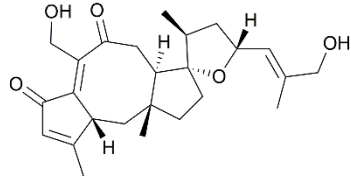
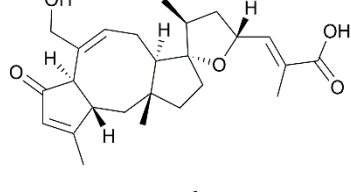
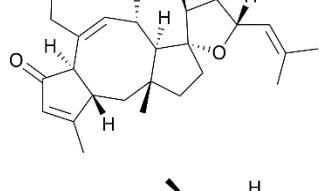
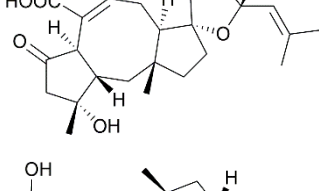
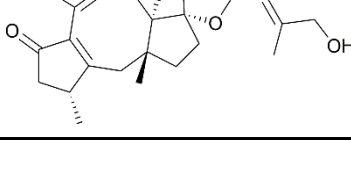
TrC-37	8-deoxyophiobolin J		<i>Bipolaris oryzae</i> (as <i>Drechslera oryzae</i>) ³³⁸ <i>Bipolaris</i> sp. TJ403-B1 ³³⁹
TrC-38	ophiobolin K		<i>Aspergillus</i> sp. ZJ-68 ³²⁷ <i>Aspergillus calidoustus</i> ^{317, 340} <i>Aspergillus insuetus</i> ³¹⁷ <i>Aspergillus</i> section <i>Usti</i> (IBT 18591) ³¹⁷ <i>Aspergillus ustus</i> ^{330, 341} <i>Emericella variecolor</i> ^{320, 331} <i>Aspergillus</i> sp. endophytic fungus from the body of <i>Zoanthus</i> ³²⁸
TrC-39	6- <i>epi</i> -ophiobolin K		<i>Aspergillus</i> sp. ZJ-68 ³²⁷ <i>Aspergillus</i> sp. endophytic fungus from the body of <i>Zoanthus</i> ³²⁸ <i>Aspergillus calidoustus</i> ^{317, 340} <i>Aspergillus insuetus</i> ³¹⁷ <i>Aspergillus</i> section <i>Usti</i> IBT 18591 ³¹⁷ <i>Aspergillus ustus</i> ^{330, 341} <i>Emericella variecolor</i> ^{320, 331}
TrC-40	14,15-dehydrophiobolin K		<i>Aspergillus flocculosus</i> ³¹⁸
TrC-41	14,15-dehydro-6- <i>epi</i> -phiobolin K		<i>Aspergillus flocculosus</i> ³¹⁸
TrC-42	21-deoxyphiobolin K		<i>Aspergillus ustus</i> ³³⁰
TrC-43	ophiobolin L		<i>Cochliobolus heterostrophus</i> ³⁰⁰

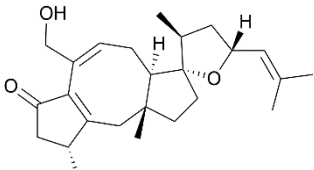
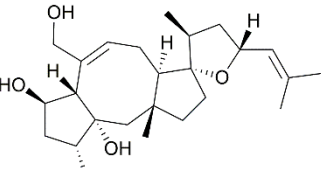
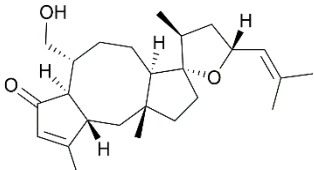
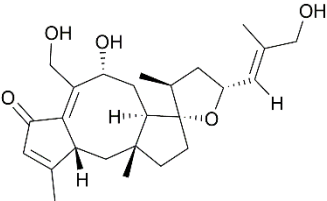
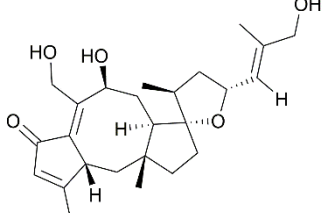
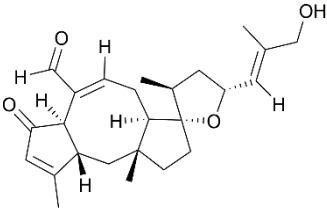
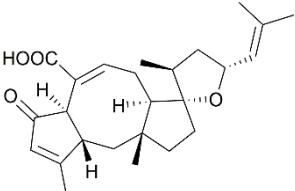
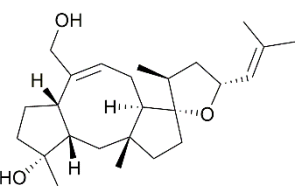
TrC-44	6- <i>epi</i> -ophiobolin L		As microbial metabolite of ophiobolin A by <i>Penicillium patulum</i> ³⁰⁰
TrC-45	ophiobolin M		<i>Cochliobolus heterostrophus</i> ³¹⁹
TrC-46	6- <i>epi</i> -ophiobolin M		<i>Cochliobolus heterostrophus</i> ³¹⁹
TrC-47	ophiobolin N (anhydrozizanin-A)		<i>Aspergillus flocculosus</i> ³¹⁸ <i>Ophiobolus heterostrophus</i> (<i>Bipolaris oryzae</i>) ³⁰⁶
TrC-48	6- <i>epi</i> -ophiobolin N		<i>Aspergillus insuetus</i> ³³⁵ <i>Emericella variegata</i> ³²⁰ <i>Aspergillus flocculosus</i> ³¹⁸
TrC-49	ophiobolin O		<i>Aspergillus</i> sp. endophytic fungus from the body of <i>Zoanthus</i> ³²⁸ <i>Aspergillus ustus</i> ^{330, 342} <i>Cephalosporium caerulens</i> ²⁹⁴
TrC-50	6- <i>epi</i> -ophiobolin O		<i>Aspergillus</i> sp. endophytic fungus from the body of <i>Zoanthus</i> ³²⁸
TrC-51	21- <i>epi</i> -ophiobolin O		<i>Aspergillus ustus</i> ³³⁰

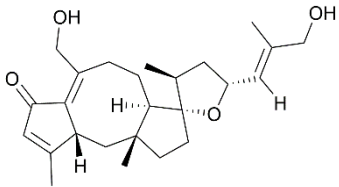
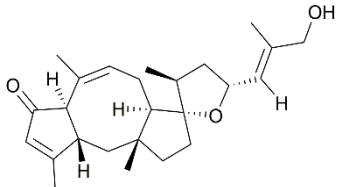
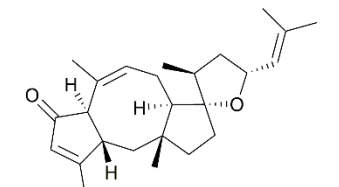
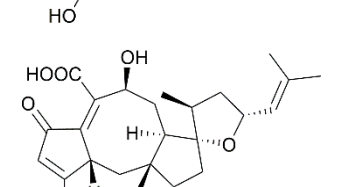
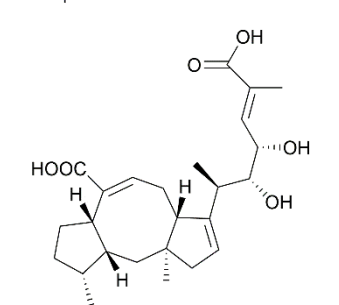
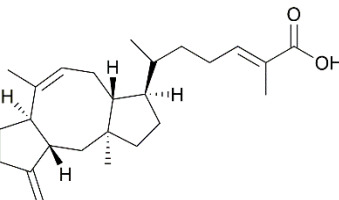
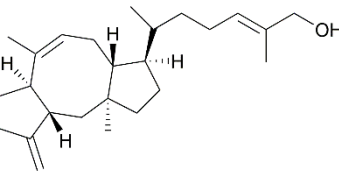
TrC-52	ophiobolin P		<i>Aspergillus</i> sp. ZJ-68 ³²⁷ <i>Aspergillus ustus</i> ³³⁰ <i>Ulocladium</i> sp. isolated from <i>Everniastrum</i> sp. ³³²
TrC-53	ophiobolin Q		<i>Aspergillus</i> sp. ZJ-68 ³²⁷ <i>Aspergillus ustus</i> ³³⁰ <i>Ulocladium</i> sp. isolated from <i>Everniastrum</i> sp. ³³²
TrC-54	ophiobolin R		<i>Ulocladium</i> sp. isolated from <i>Everniastrum</i> sp. ³³²
TrC-55	ophiobolin S		<i>Ulocladium</i> sp. isolated from <i>Everniastrum</i> sp. ³³²
TrC-56	ophiobolin T		<i>Ulocladium</i> sp. isolated from <i>Everniastrum</i> sp. ³³²
TrC-57	ophiobolin U*		<i>Aspergillus</i> sp. ZJ-68 ³²⁷ <i>Aspergillus ustus</i> ^{326, 330}
TrC-58	ophiobolin U*		<i>Aspergillus</i> sp. ZJ-68 ³²⁷ <i>Aspergillus insuetus</i> ^{317, 335}
TrC-59	21-dehydrophiobolin U		<i>Aspergillus ustus</i> ³³⁰

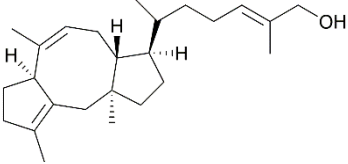
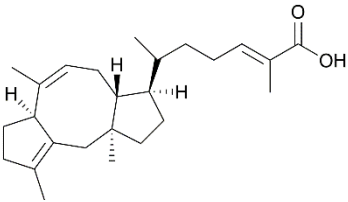
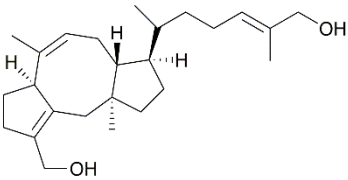
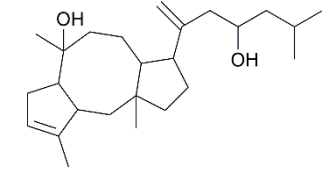
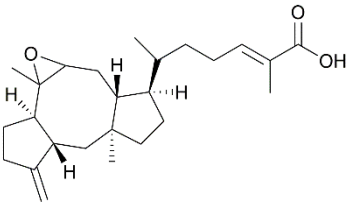
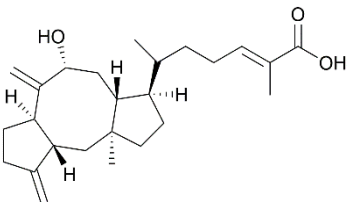
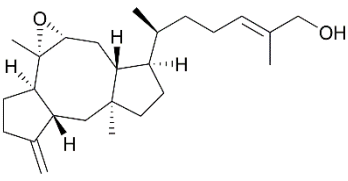
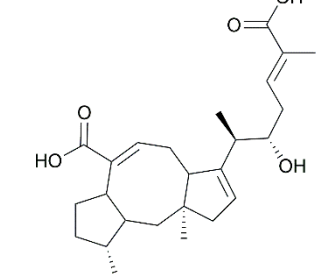
TrC-60	ophiobolin V		<i>Aspergillus ustus</i> ³²⁶
TrC-61	ophiobolin W		<i>Aspergillus ustus</i> ³²⁶
TrC-62	ophiobolin X		<i>Aspergillus</i> sp. ZJ-68 ³²⁷ <i>Aspergillus ustus</i> ³³⁰
TrC-63	ophiobolin Y		<i>Aspergillus ustus</i> ³³⁰
TrC-64	ophiobolin Z		<i>Aspergillus ustus</i> ³³⁰
TrC-65	21- <i>epi</i> -ophiobolin Z		<i>Aspergillus ustus</i> ³³⁰
TrC-66	ophiobola-7,19-dien-25-oic acid (14,18(<i>R</i>)-epoxy-3,5-dihydroxy-γ-lactone)		<i>Cochliobolus miyabeanus</i> ³⁴³
TrC-67	asperophiobolin A		<i>Aspergillus</i> sp. ZJ-68 ³²⁷

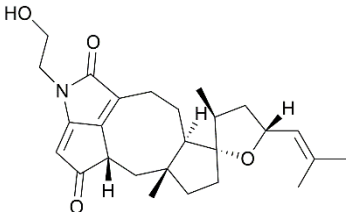
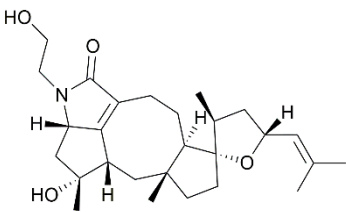
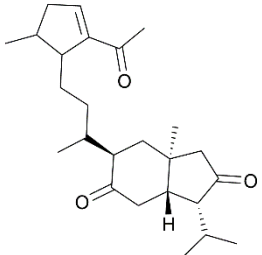
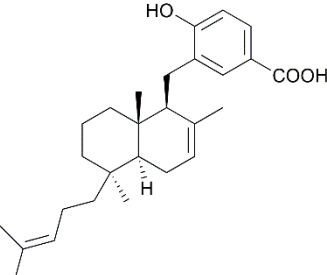
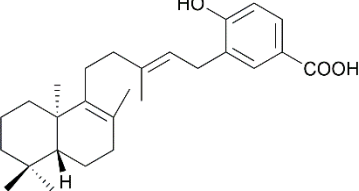
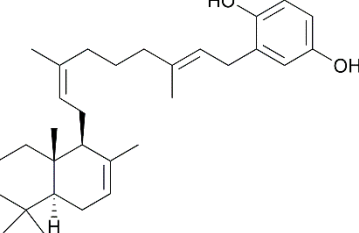
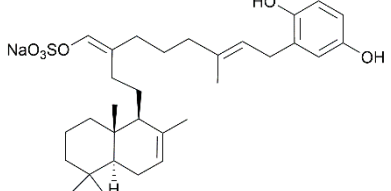
TrC-68	asperophiobolin B		<i>Aspergillus</i> sp. ZJ-68 ³²⁷
TrC-69	asperophiobolin C		<i>Aspergillus</i> sp. ZJ-68 ³²⁷
TrC-70	asperophiobolin D		<i>Aspergillus</i> sp. ZJ-68 ³²⁷
TrC-71	asperophiobolin E		<i>Aspergillus</i> sp. ZJ-68 ³²⁷
TrC-72	asperophiobolin F		<i>Aspergillus</i> sp. ZJ-68 ³²⁷
TrC-73	asperophiobolin G		<i>Aspergillus</i> sp. ZJ-68 ³²⁷
TrC-74	asperophiobolin H		<i>Aspergillus</i> sp. ZJ-68 ³²⁷
TrC-75	asperophiobolin I		<i>Aspergillus</i> sp. ZJ-68 ³²⁷

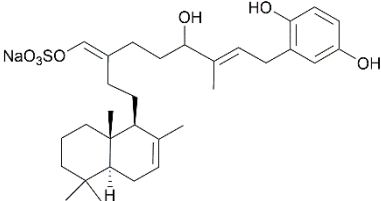
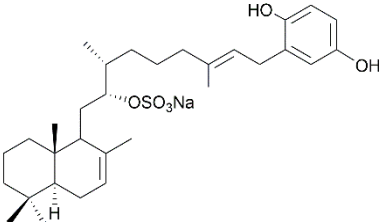
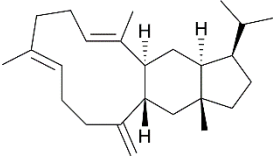
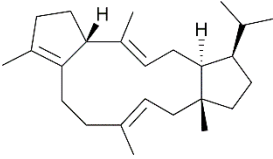
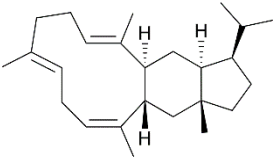
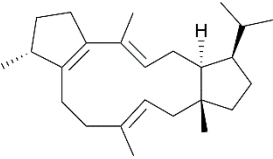
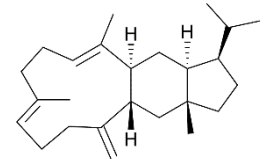
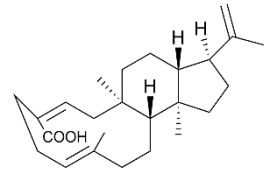
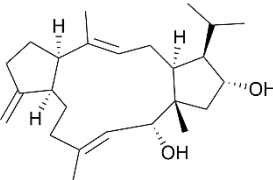
TrC-76	asperophiobolin J		<i>Aspergillus</i> sp. ZJ-68 ³²⁷
TrC-77	asperophiobolin K		<i>Aspergillus</i> sp. ZJ-68 ³²⁷
TrC-78	ophiobotriol		<i>Bipolaris</i> sp. TJ403-B1 ¹⁷³
TrC-79	bipolarin A		<i>Bipolaris</i> sp. TJ403-B1 ³³⁹
TrC-80	bipolarin B		<i>Bipolaris</i> sp. TJ403-B1 ³³⁹
TrC-81	bipolarin C		<i>Bipolaris</i> sp. TJ403-B1 ³³⁹
TrC-82	bipolarin D		<i>Bipolaris</i> sp. TJ403-B1 ³³⁹
TrC-83	bipolarin E		<i>Bipolaris</i> sp. TJ403-B1 ³³⁹

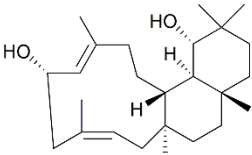
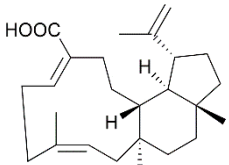
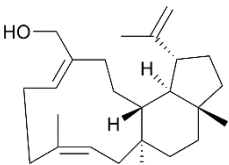
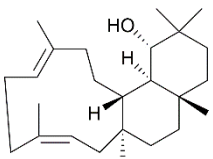
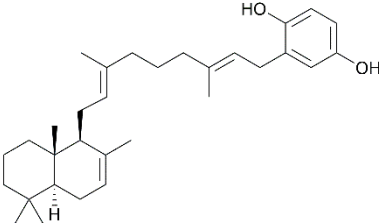
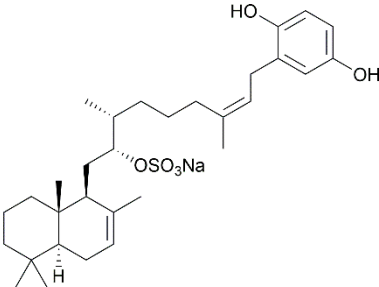
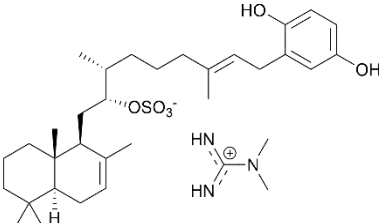
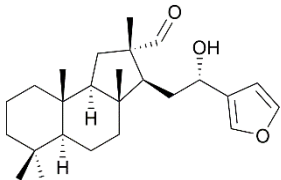
TrC-84	bipolarin F		<i>Bipolaris</i> sp. TJ403-B1 ³³⁹
TrC-85	bipolarin G		<i>Bipolaris</i> sp. TJ403-B1 ³³⁹
TrC-86	bipolarin H		<i>Bipolaris</i> sp. TJ403-B1 ³³⁹
TrC-87	bipolaricin A		<i>Bipolaris</i> sp. TJ403-B1 ³¹⁴
TrC-88	bipolaricin B		<i>Bipolaris</i> sp. TJ403-B1 ³¹⁴
TrC-89	bipolaricin C		<i>Bipolaris</i> sp. TJ403-B1 ³¹⁴
TrC-90	bipolaricin D		<i>Bipolaris</i> sp. TJ403-B1 ³¹⁴
TrC-91	bipolaricin E		<i>Bipolaris</i> sp. TJ403-B1 ³¹⁴

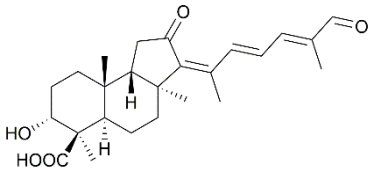
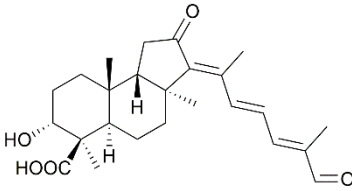
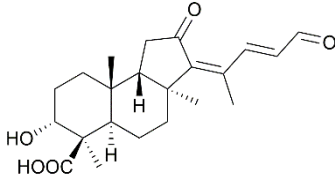
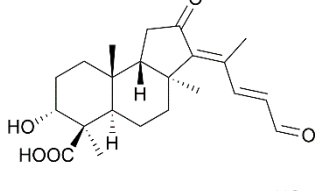
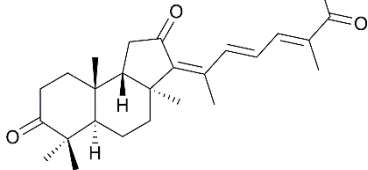
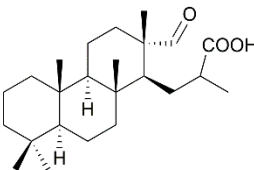
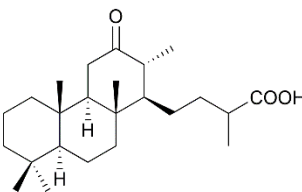
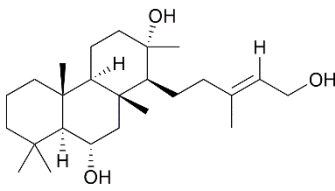
TrC-92	bipolaricin F		<i>Bipolaris</i> sp. TJ403-B1 ³¹⁴
TrC-93	bipolaricin G		<i>Bipolaris</i> sp. TJ403-B1 ³¹⁴
TrC-94	bipolaricin H		<i>Bipolaris</i> sp. TJ403-B1 ³¹⁴
TrC-95	bipolaricin I		<i>Bipolaris</i> sp. TJ403-B1 ³¹⁴
TrC-96	halorosellinic acid		<i>Halorosellinia oceanica</i> ³⁴⁴
TrC-97	ceroplasteric acid		<i>Ceroplastes albolineatus</i> ^{345, 346}
TrC-98	ceroplastol-I		<i>Ceroplastes albolineatus</i> ^{207, 345}

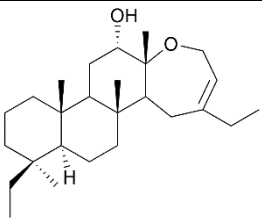
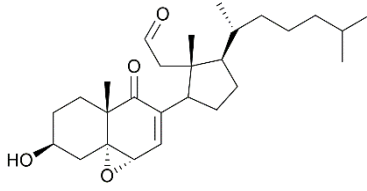
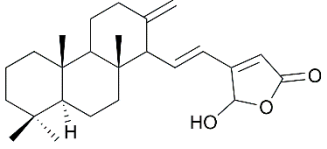
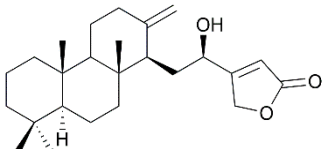
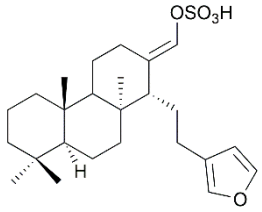
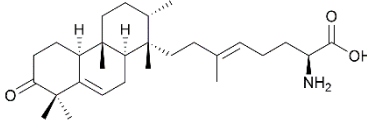
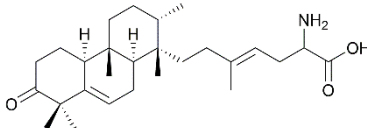
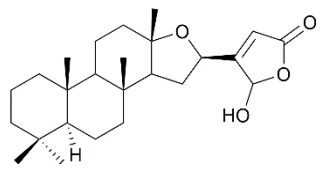
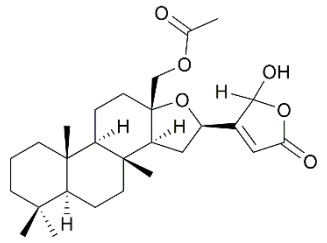
TrC-99	ceroplastol-II		<i>Ceroplastes albolineatus</i> ^{207, 347}
TrC-100	albolic acid		<i>Ceroplastes albolineatus</i> ³⁴⁶
TrC-101	ceroplastodiol		<i>Ceroplastes albolineatus</i> ³⁴⁸
TrC-102	cheilarinosin		<i>Cheilanthes farinosa</i> ³⁴⁹
TrC-103	ceralbic acid I		<i>Ceroplastes albolineatus</i> ³⁵⁰
TrC-104	ceralbic acid II		<i>Ceroplastes albolineatus</i> ³⁵⁰
TrC-105	ceralbol		<i>Ceroplastes albolineatus</i> ³⁵¹
TrC-106	17-dehydroxyhalorosellinic acid		<i>Halorosellinia oceanica</i> ³⁴⁴

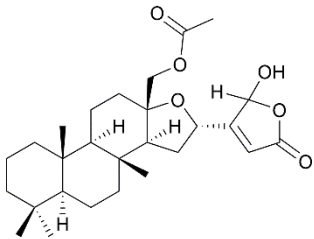
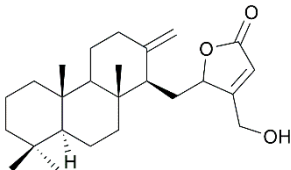
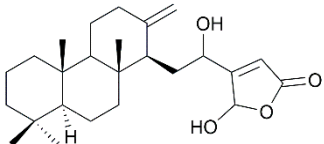
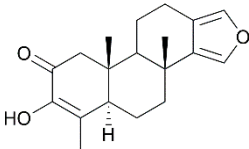
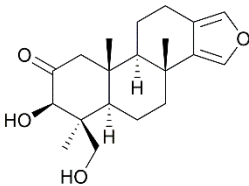
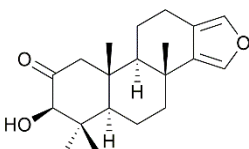
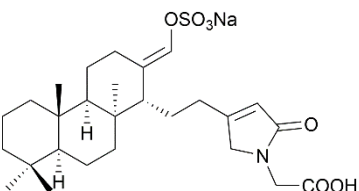
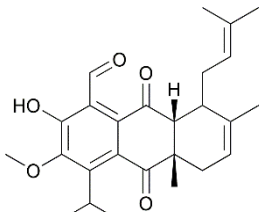
TrC-107	bipolarolide F		<i>Bipolaris</i> sp. TJ403-B1 ¹⁷³
TrC-108	bipolarolide G		<i>Bipolaris</i> sp. TJ403-B1 ¹⁷³
TrC-109	alborosin		<i>Gentianella alborosea</i> ³⁵²
TrC-110	jaspic acid		<i>Jaspis</i> cf. <i>johnstoni</i> ³⁵³
TrC-111	(-)-(5 <i>R</i> ,10 <i>R</i>)-subersic acid		<i>Suberea</i> sp. ³⁵⁴
TrC-112	coscinoquinol		<i>Coscinoidea</i> sp. ³⁵⁵ <i>Dysidea</i> sp. ¹⁴⁵
TrC-113	hipposulfate A		<i>Hippospongia</i> cf. <i>metachromia</i> ³⁵⁶

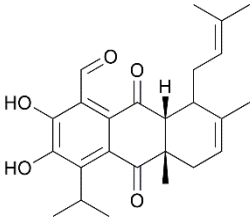
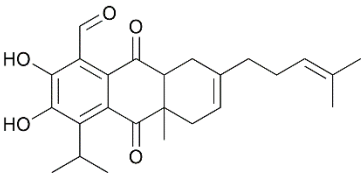
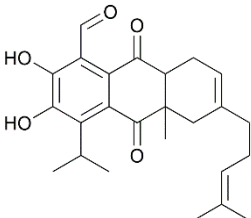
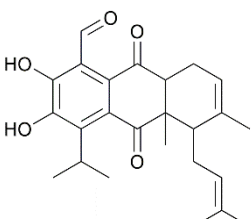
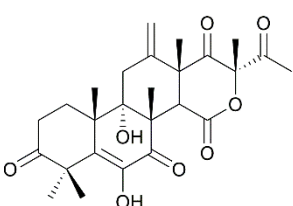
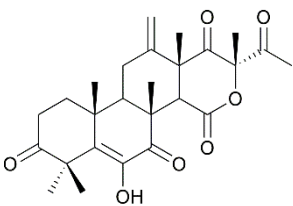
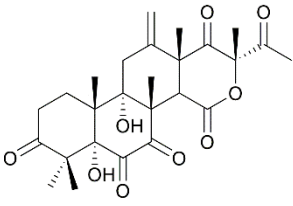
TrC-114	hipposulfate B		<i>Hippospongia</i> cf. <i>metachromia</i> ³⁵⁶
TrC-115	halisulfate 1		Marine sponge of the family <i>Halichondriidae</i> ¹⁴⁴ <i>Hippospongia</i> sp. ¹⁹⁴ <i>Dysidea</i> sp. ¹⁴⁵
TrC-116	(+)-arathanatriene (reported as (-)- caprutriene A by Huang et al. ²⁶⁵)		<i>Arabidopsis thaliana</i> ²⁶⁶ <i>Nicotiana benthamiana</i> / <i>Agrobacterium</i> <i>tumefaciens</i> ²⁶⁶
TrC-117	(-)-variculatriene A		<i>Nicotiana benthamiana</i> / <i>Agrobacterium</i> <i>tumefaciens</i> ²⁶⁶
TrC-118	(-)-caprutriene (reported as (+)-arathanatriene in Huang et al. ²⁶⁵)		<i>Nicotiana benthamiana</i> / <i>Agrobacterium</i> <i>tumefaciens</i> ²⁶⁶
TrC-119	(-)-variculatriene B		<i>Nicotiana benthamiana</i> / <i>Agrobacterium</i> <i>tumefaciens</i> ²⁶⁶
TrC-120	(+)-thalianatriene		<i>Arabidopsis thaliana</i> / <i>Escherichia coli</i> ³⁵⁷
TrC-121	stellatic acid		<i>Aspergillus stellatus</i> ^{358, 359}
TrC-122	variculanol		<i>Aspergillus varicolor</i> ³⁶⁰

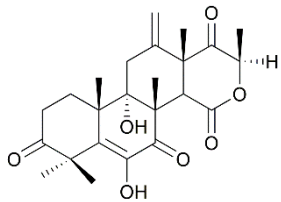
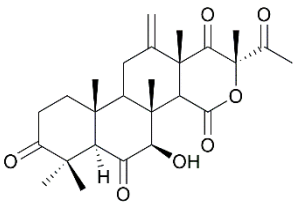
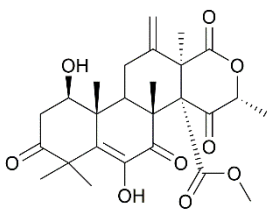
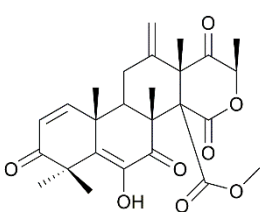
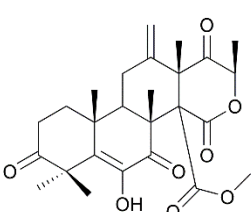
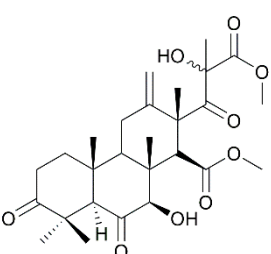
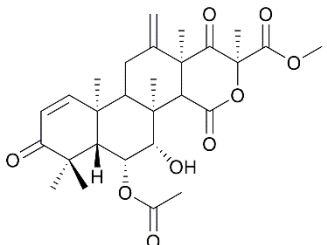
TrC-123	5 α -hydroxyfloridenol		<i>Ceroplastes floridensis</i> ³⁶¹
TrC-124	floceric acid		<i>Ceroplastes floridensis</i> ³⁶¹
TrC-125	flocerol		<i>Ceroplastes floridensis</i> ³⁶¹
TrC-126	floridenol		<i>Ceroplastes floridensis</i> ³⁶¹
TrC-127	(<i>E</i>)-halisulfate		<i>Coscinoderma</i> sp. ³⁶²
TrC-128	(17 <i>Z</i>)-halisulfate 1		<i>Coscinoderma</i> sp. ³⁶²
TrC-129	N,N-dimethylguanidinium halisulfate 1		<i>Dysidea</i> sp. ¹⁴⁵
TrC-130	hyrtiosal		<i>Hyrtios erectus</i> ³⁶³ <i>Spongia hispida</i> ³⁶⁴ <i>Hippospongia</i> sp. ³⁶⁵

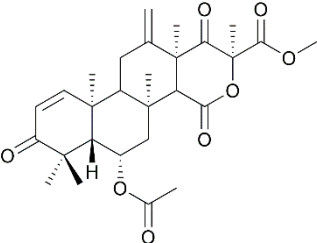
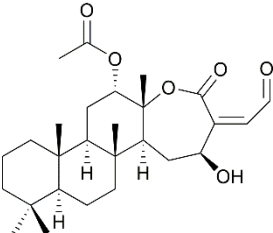
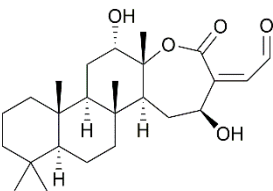
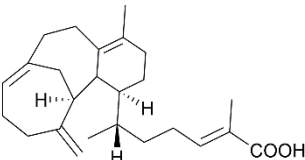
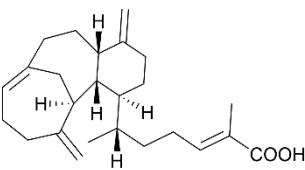
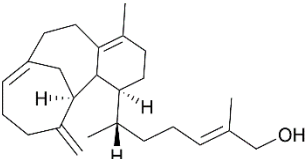
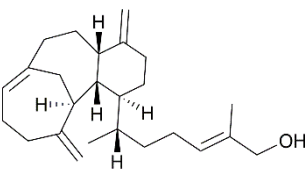
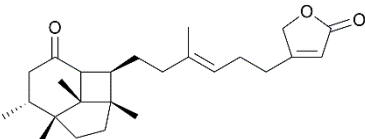
TrC-131	jaspiferal C		<i>Jaspis stellifera</i> ³⁶⁶
TrC-132	jaspiferal D		<i>Jaspis stellifera</i> ³⁶⁶
TrC-133	jaspiferal E		<i>Jaspis stellifera</i> ³⁶⁶
TrC-134	jaspiferal F		<i>Jaspis stellifera</i> ³⁶⁶
TrC-135	jaspolide F		<i>Jaspis</i> sp. ³⁶⁷
TrC-136	conulosin A		<i>Latrunculia conulosa</i> ³⁶⁸
TrC-137	conulosin B		<i>Latrunculia conulosa</i> ³⁶⁸
TrC-138	cheilanthatriol		<i>Cheilanthes farinosa</i> ³⁶⁹

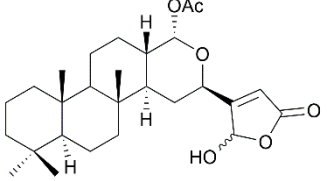
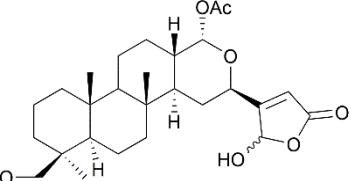
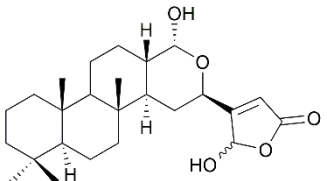
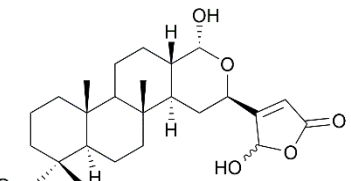
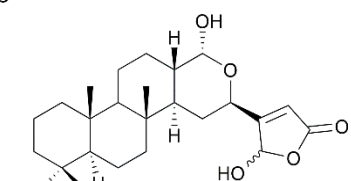
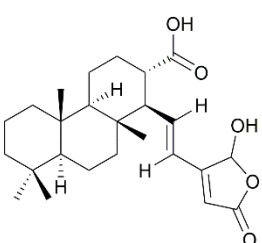
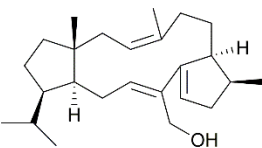
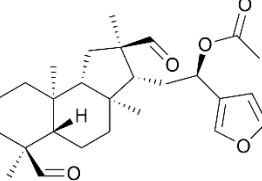
TrC-139	granulosane A		<i>Dysidea granulosa</i> ³⁷⁰
TrC-140	9,11-seco-3 α -hydroxy-5 α ,6 α -epoxy-9-oxo-9,11-seco-5 α -cholest-7-en-11-al		<i>Spongia matamata</i> ³⁷¹
TrC-141	25-hydroxy-13(24),15,17-cheilanthatrien-19,25-olide		<i>Ircinia</i> sp. ³⁷²
TrC-142	hamiltonin E		<i>Chromodoris hamiltoni</i> ³⁷³
TrC-143	suvanine		<i>Ircinia</i> sp. ³⁷⁴ <i>Coscinoderma mathewsi</i> ³⁷⁵
TrC-144	atolypene A		<i>Streptomyces albus</i> , <i>Amycolatopsis tolypomycina</i> ³⁷⁶
TrC-145	atolypene B		<i>Streptomyces albus</i> , <i>Amycolatopsis tolypomycina</i> ³⁷⁶
TrC-146	13,16-epoxy-25-hydroxy-17-cheilanthatrien-19,25-olide		<i>Ircinia</i> sp. ³⁷²
TrC-147	lintonolide A or spongianolide C		<i>Cacospongia</i> cf. <i>linteiformis</i> ³⁷⁷ <i>Spongia</i> sp. ³⁷⁸ <i>Semitaspongia bactriana</i> ⁶⁹

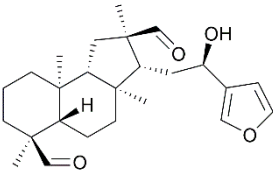
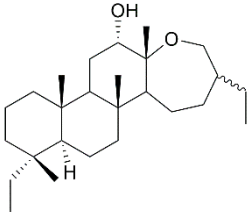
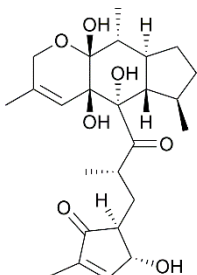
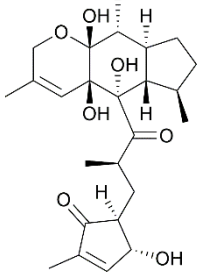
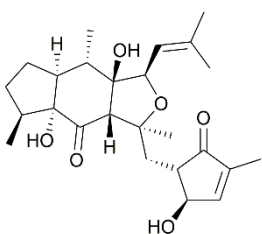
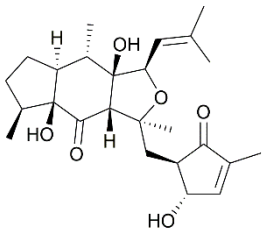
TrC-148	lintonolide B or spongianolide D		<i>Cacospongia</i> cf. <i>linteiformis</i> ³⁷⁷ <i>Spongia</i> sp. ³⁷⁸
TrC-149	25-hydroxy-13(24),17- cheilanthadien-16,19- olide		<i>Ircinia</i> sp. ³⁷²
TrC-150	16,25-dihydroxy- 13(24),17-cheilanthadien- 19,25-olide		<i>Ircinia</i> sp. ³⁷²
TrC-151	19-nor-3-hydroxyspongia- 3,13(16),14-trien-2-one		<i>Spongia tubulifera</i> ³⁷⁹
TrC-152	3β,19-dihydroxyspongia- 13(16),14-dien-2-one (epispongiadiol)		<i>Spongia tubulifera</i> ³⁷⁹
TrC-153	3β-hydroxyspongia- 13(16),14-dien-2-one		<i>Spongia tubulifera</i> ³⁷⁹
TrC-154	coscinolactam B		<i>Coscinoderma mathewsi</i> ³⁷⁵
TrC-155	heliocide B1		<i>Gossypium hirsutum</i> ¹⁵⁰

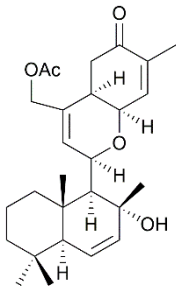
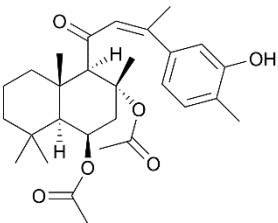
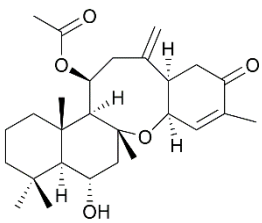
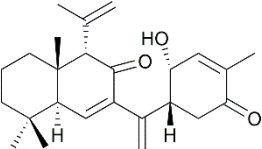
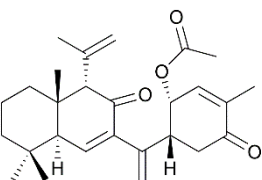
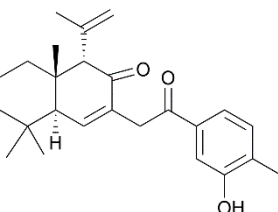
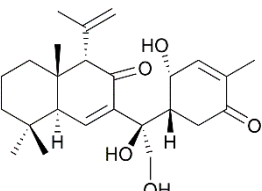
TrC-156	heliocide H1 also reported as heliocide 2 ³⁸⁰		<i>Gossypium hirsutum</i> ^{150, 381}
TrC-157	heliocide H2 also reported as heliocide 1 ³⁸⁰		<i>Gossypium hirsutum</i> ³⁸²
TrC-158	heliocide H3 also reported as heliocide 4 ³⁸⁰		<i>Gossypium hirsutum</i> ³⁸³
TrC-159	heliocide H4 also reported as heliocide 3 ³⁸⁰		<i>Gossypium hirsutum</i> ³⁸¹
TrC-160	terretonin		<i>Aspergillus terreus</i> NRRL 6273 ³⁸⁴
TrC-161	terretonin A		<i>Aspergillus terreus</i> ³⁸⁵
TrC-162	terretonin B		<i>Aspergillus terreus</i> ³⁸⁵

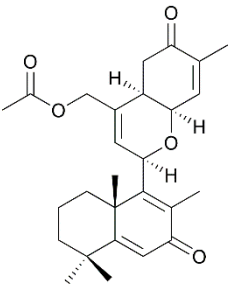
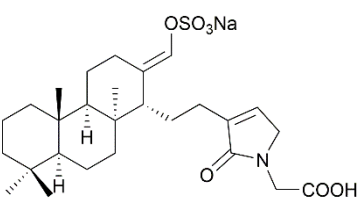
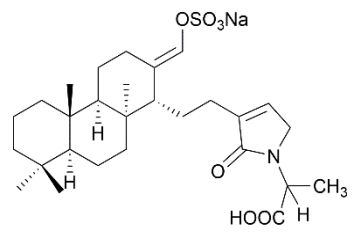
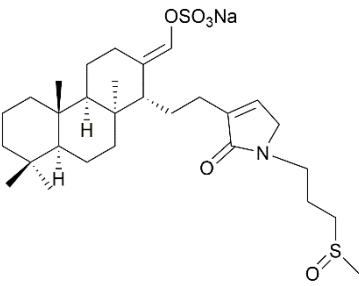
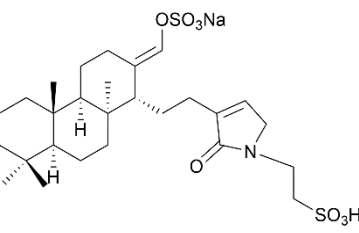
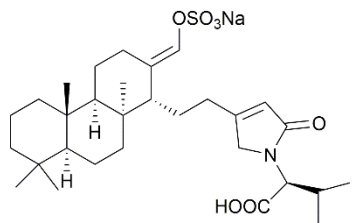
TrC-163	terretonin C		<i>Aspergillus terreus</i> ³⁸⁵
TrC-164	terretonin D		<i>Aspergillus terreus</i> ³⁸⁵
TrC-165	terretonin E		<i>Aspergillus insuetus</i> ³⁸⁶ <i>Aspergillus</i> sp. RR-YLW-12 (<i>Rhodomela confervoides</i>) ³⁸⁷
TrC-166	terretonin F		<i>Aspergillus insuetus</i> ³⁸⁶ <i>Aspergillus ustus</i> ³²⁶
TrC-167	1,2-dihydroterretonin F		<i>Aspergillus ustus</i> ³²⁶
TrC-168	terretonin G		<i>Aspergillus</i> sp. OPMF00272 ³⁸⁸
TrC-169	terretonin H		<i>Aspergillus ustus</i> ³⁸⁹

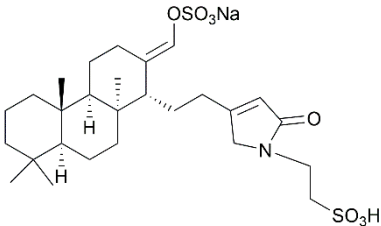
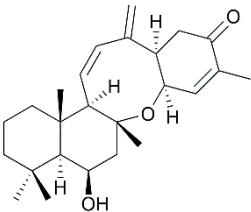
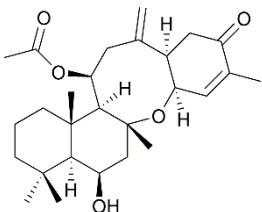
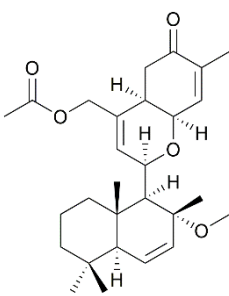
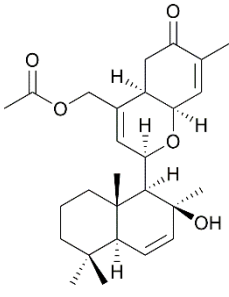
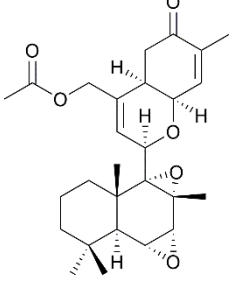
TrC-170	terretonin I		<i>Aspergillus ustus</i> ³⁸⁹
TrC-171	hyatolide A		<i>Hyattella intestinalis</i> ³⁹⁰
TrC-172	hyatolide B		<i>Hyattella intestinalis</i> ³⁹⁰
TrC-173	cerorubenic acid-I		<i>Ceroplastes rubens</i> ³⁹¹
TrC-174	cerorubenic acid-II		<i>Ceroplastes rubens</i> ³⁹¹
TrC-175	cerorubenol-I		<i>Ceroplastes rubens</i> ³⁹¹
TrC-176	cerorubenol-II		<i>Ceroplastes rubens</i> ³⁹¹
TrC-177	lintenone		<i>Cacospongia</i> cf. <i>linteiformis</i> ³⁹²

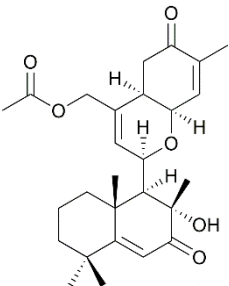
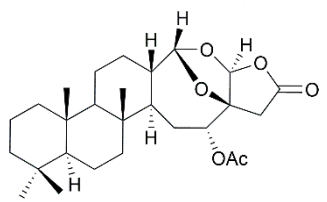
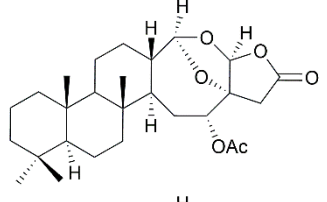
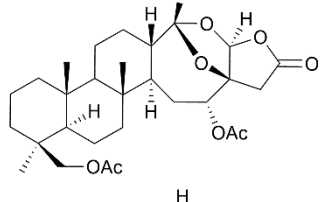
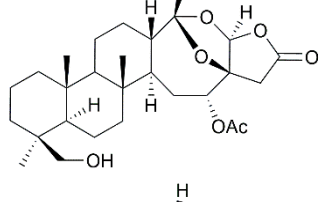
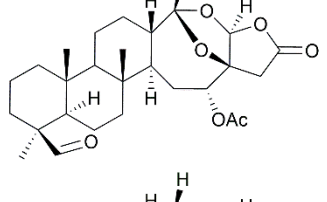
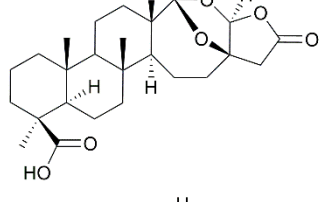
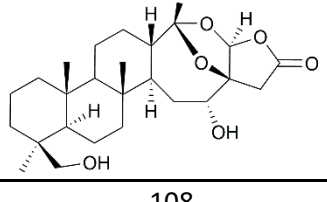
TrC-178	petrosaspongiolide M		<i>Petrosaspongia nigra</i> ³⁹³
TrC-179	petrosaspongiolide N		<i>Petrosaspongia nigra</i> ³⁹³
TrC-180	petrosaspongiolide P		<i>Petrosaspongia nigra</i> ³⁹³ <i>Spongia</i> sp. ³⁹⁴
TrC-181	21-hydroxy petrosaspongiolide P		<i>Spongia</i> sp. ³⁹⁴
TrC-182	petrosaspongiolide Q		<i>Petrosaspongia nigra</i> ³⁹³
TrC-183	petrosaspongiolide R		<i>Petrosaspongia nigra</i> ³⁹³
TrC-184	nitinol		<i>Gentianella nitida</i> ³⁹⁵
TrC-185	(+)-16-O-acetyl-20-formylhyrtiosal		<i>Hyrtios erectus</i> ³⁴

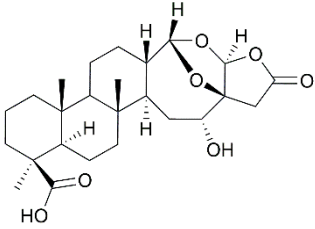
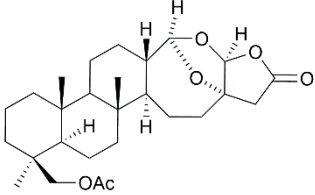
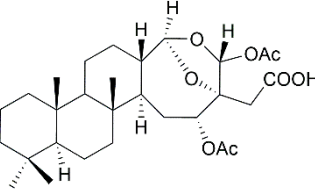
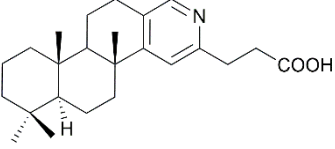
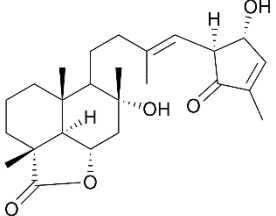
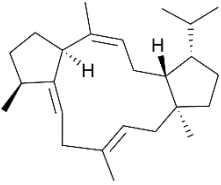
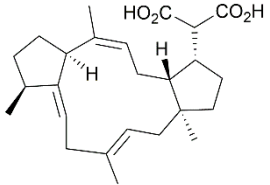
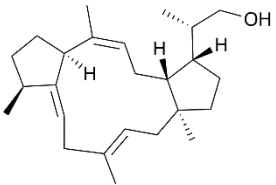
TrC-186	(+)-20-formylhyrtiosal		<i>Hyrtios erectus</i> ³⁴
TrC-187	12 β -hydroxy-20,24-dimethyl-13,18-oxa-25-norscalarane		<i>Phyllospongia madagascarensis</i> ³⁹⁶
TrC-188	leucosesterterpenone		<i>Leucosceptrum canum</i> ²⁴¹
TrC-189	14 β -methylleucosesterterpenone		<i>Leucosceptrum canum</i> ²⁴³
TrC-190	leucosceptroid C		<i>Leucosceptrum canum</i> ³⁹⁷
TrC-191	leucosceptroid D		<i>Leucosceptrum canum</i> ³⁹⁷

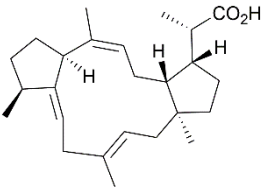
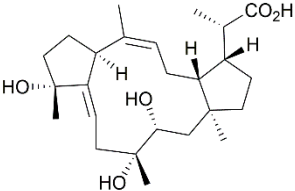
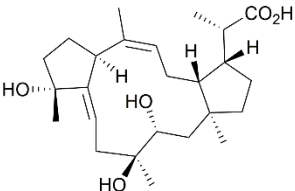
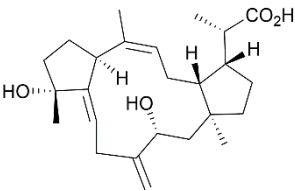
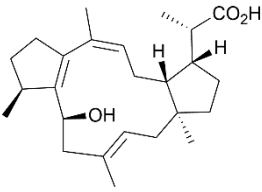
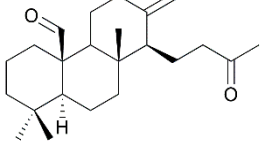
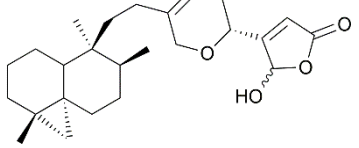
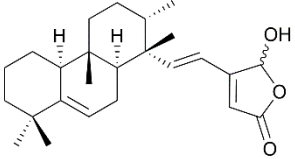
TrC-192	ansellone A		<i>Cadlina luteomarginata</i> ³⁹⁸ <i>Phorbas</i> sp. ³⁹⁸
TrC-193	ansellone B		<i>Phorbas</i> sp. ^{162, 399}
TrC-194	ansellone B1		<i>Phorbas</i> sp. ¹⁶²
TrC-195	phorbasone A		<i>Phorbas</i> sp. ⁴⁰⁰
TrC-196	phorbasone A acetate		<i>Phorbas</i> sp. ³⁹⁹
TrC-197	isophorbasone A		<i>Phorbas</i> sp. ³⁹⁹
TrC-198	phorbasone B		<i>Phorbas</i> sp. ⁴⁰⁰

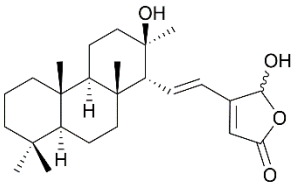
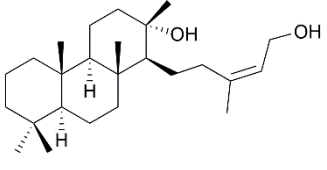
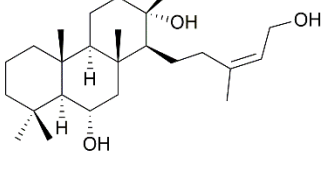
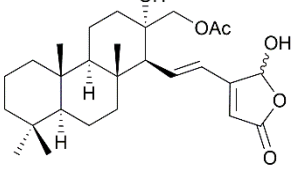
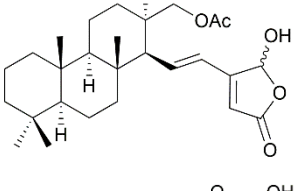
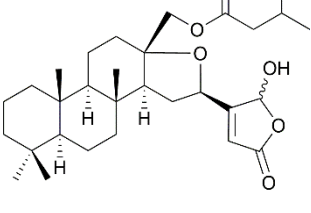
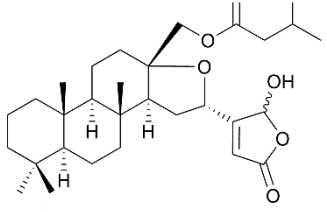
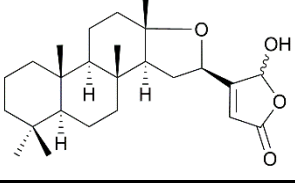
TrC-199	phorbadione		<i>Phorbas</i> sp. ¹⁶²
TrC-200	coscinolactam A		<i>Coscinoderma mathewsi</i> ³⁷⁵ <i>Coscinoderma</i> sp. ³⁰⁸
TrC-201	coscinolactam C		<i>Coscinoderma</i> sp. ³⁰⁸
TrC-202	coscinolactam D		<i>Coscinoderma</i> sp. ³⁰⁸
TrC-203	coscinolactam E		<i>Coscinoderma</i> sp. ³⁰⁸
TrC-204	coscinolactam F		<i>Coscinoderma</i> sp. ³⁰⁸

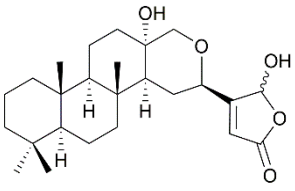
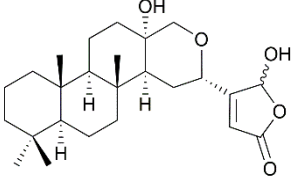
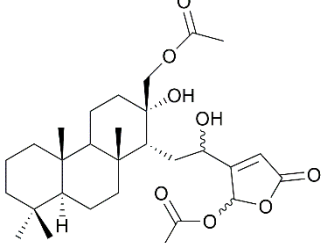
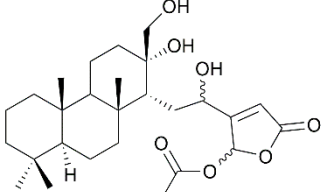
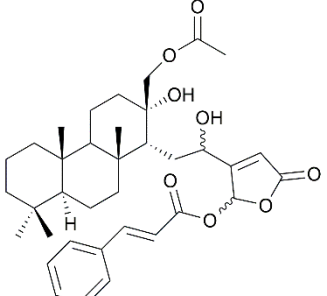
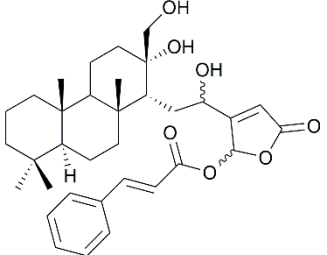
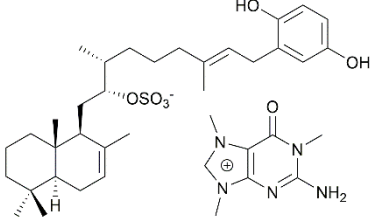
TrC-205	coscinolactam G		<i>Coscinoderma</i> sp. ³⁰⁸
TrC-206	ansellone C*		<i>Clathria gombawuiensis</i> ⁴⁰¹
TrC-207	ansellone C*		<i>Phorbas</i> sp. ¹⁶³
TrC-208	ansellone D		<i>Phorbas</i> sp. ¹⁶³
TrC-209	ansellone E		<i>Phorbas</i> sp. ¹⁶³
TrC-210	ansellone F		<i>Phorbas</i> sp. ¹⁶³

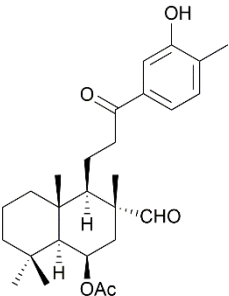
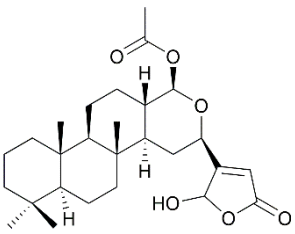
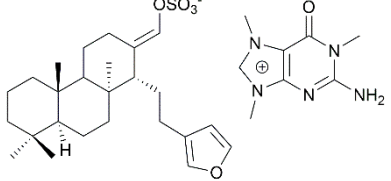
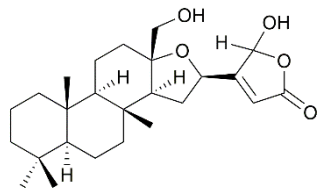
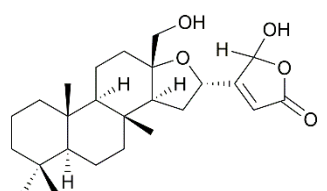
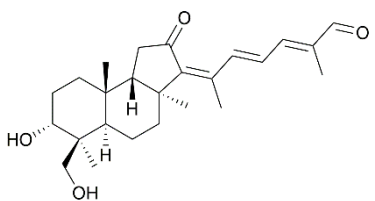
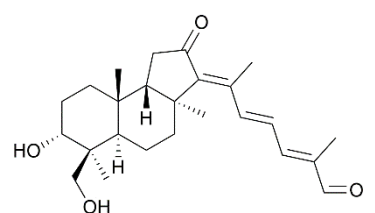
TrC-211	ansellone G		<i>Phorbas</i> sp. ¹⁶³
TrC-212	petrosaspongiolide A (reported also as unnamed tricycyclic sesterterpene, compound 2 ⁴⁰²)		<i>Dactylospongia</i> sp. ⁴⁰² <i>Petrosaspongia nigra</i> ⁴⁰³ <i>Spongia</i> sp. ⁴⁰⁴
TrC-213	petrosaspongiolide B (reported also as unnamed tricycyclic sesterterpene, compound 1 ⁴⁰²)		<i>Dactylospongia</i> sp. ⁴⁰² <i>Petrosaspongia nigra</i> ⁴⁰³ <i>Spongia</i> sp. ⁴⁰⁴
TrC-214	petrosaspongiolide C		<i>Petrosaspongia nigra</i> ⁴⁰⁵
TrC-215	petrosaspongiolide D		<i>Petrosaspongia nigra</i> ⁴⁰⁵ <i>Spongia</i> sp. ³⁹⁴
TrC-216	petrosaspongiolide E		<i>Petrosaspongia nigra</i> ⁴⁰⁵
TrC-217	petrosaspongiolide F		<i>Petrosaspongia nigra</i> ⁴⁰⁵
TrC-218	petrosaspongiolide G		<i>Petrosaspongia nigra</i> ⁴⁰⁵ <i>Spongia</i> sp. ³⁹⁴

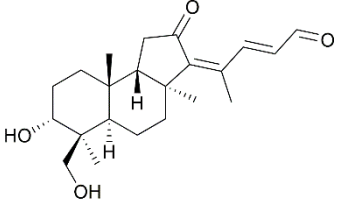
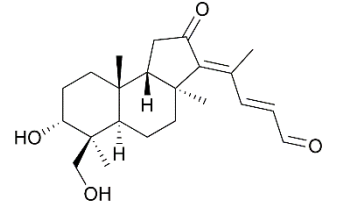
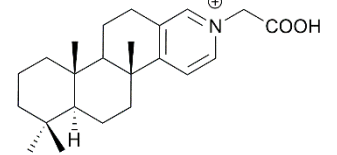
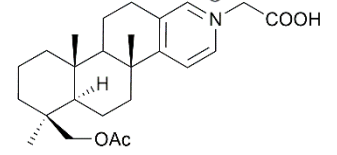
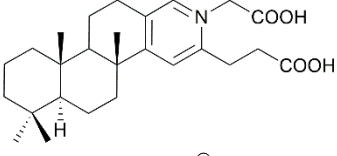
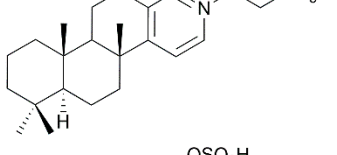
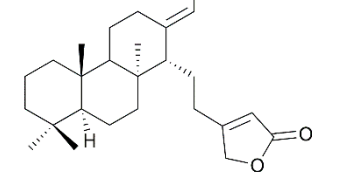
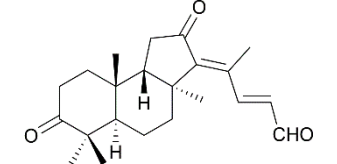
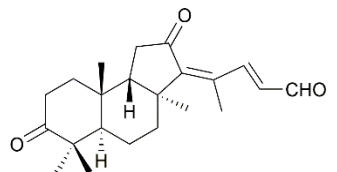
TrC-219	petrosaspongiolide H		<i>Petrosaspongia nigra</i> ⁴⁰⁵
TrC-220	petrosaspongiolide I		<i>Petrosaspongia nigra</i> ⁴⁰⁵ <i>Spongia</i> sp. ⁴⁰⁴
TrC-221	petrosaspongiolide J		<i>Petrosaspongia nigra</i> ⁴⁰⁵
TrC-222	petrosaspongiolide L		<i>Petrosaspongia nigra</i> ⁴⁰⁵
TrC-223	salvileucolidone		<i>Salvia hypoleuca</i> ²⁰¹
TrC-224	betaestacin I (Pb1)		<i>Phoma betae</i> ²⁶⁷ Genome mining of BFTS against <i>Phoma betae</i> and <i>Colletotrichum orbiculare</i> ⁴⁰⁶
TrC-225	betaestacin II		Genome mining of BFTS against <i>Phoma betae</i> and <i>Colletotrichum orbiculare</i> ⁴⁰⁶
TrC-226	betaestacin III		Genome mining of BFTS against <i>Phoma betae</i> and <i>Colletotrichum orbiculare</i> ⁴⁰⁶

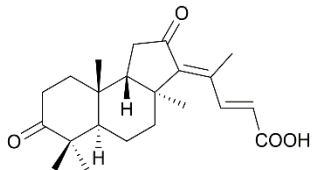
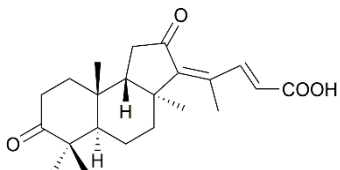
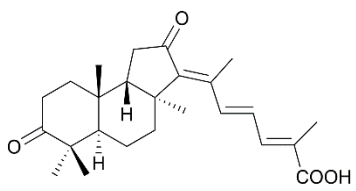
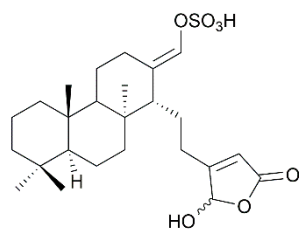
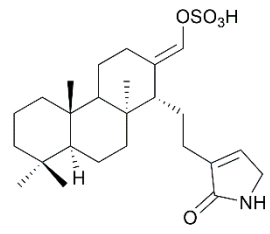
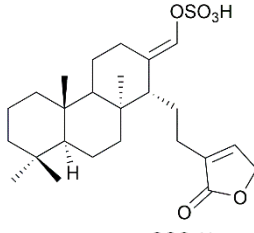
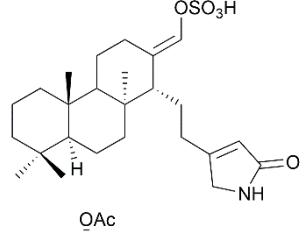
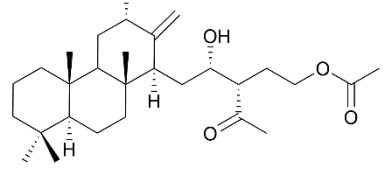
TrC-227	betaestacin IV		Genome mining of BFTS against <i>Phoma betae</i> and <i>Colletotrichum orbiculare</i> ⁴⁰⁶
TrC-228	betaestacin Va		Genome mining of BFTS against <i>Phoma betae</i> and <i>Colletotrichum orbiculare</i> ⁴⁰⁶
TrC-229	betaestacin Vb		Genome mining of BFTS against <i>Phoma betae</i> and <i>Colletotrichum orbiculare</i> ⁴⁰⁶
TrC-230	betaestacin Vc		Genome mining of BFTS against <i>Phoma betae</i> and <i>Colletotrichum orbiculare</i> ⁴⁰⁶
TrC-231	betaestacin VI		Genome mining of BFTS against <i>Phoma betae</i> and <i>Colletotrichum orbiculare</i> ⁴⁰⁶
TrC-232	luteone		<i>Cadlina luteomarginata</i> ⁴⁰⁷
TrC-233	cacospongionolide		<i>Cacospongia mollior</i> ⁴⁰⁸ <i>Fasciospongia cavernosa</i> ⁴⁰⁹
TrC-234	aplysolide A and B (25 α and 25 β)		<i>Aplysinopsis</i> sp. ⁴¹⁰

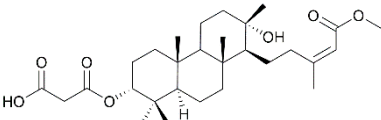
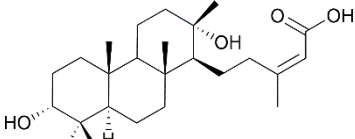
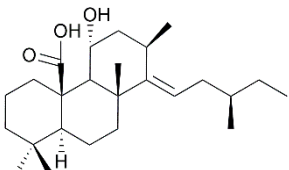
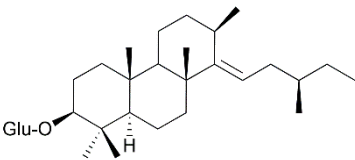
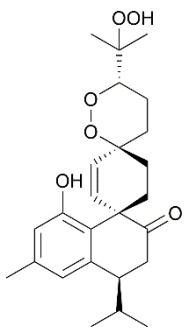
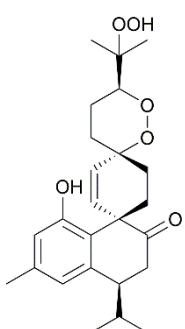
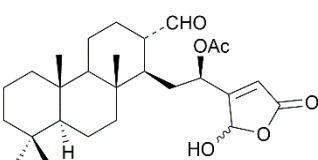
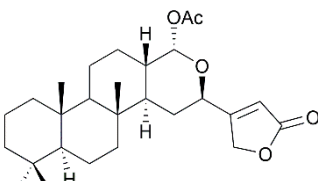
TrC-235	aplyolide		<i>Aplysinopsis</i> sp. ⁴¹⁰
TrC-236	cheilanthenediol		<i>Aleuritopteris khunii</i> ⁴¹¹
TrC-237	cheilanthenetriol		<i>Aleuritopteris khunii</i> ⁴¹¹
TrC-238	spongianolide A		<i>Spongia</i> sp. ³⁷⁸
TrC-239	spongianolide B		<i>Spongia</i> sp. ³⁷⁸
TrC-240	spongianolide E		<i>Spongia</i> sp. ³⁷⁸
TrC-241	spongianolide F		<i>Spongia</i> sp. ³⁷⁸
TrC-242	lintenolide C		<i>Cacospongia</i> cf. <i>linteiformis</i> ⁴¹²

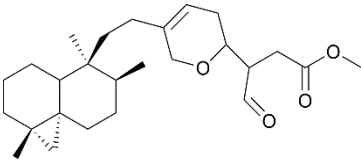
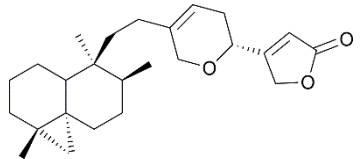
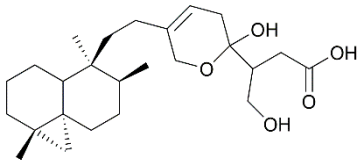
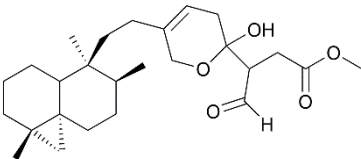
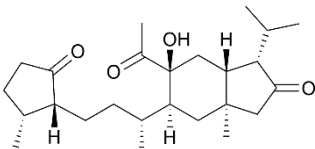
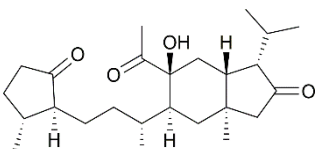
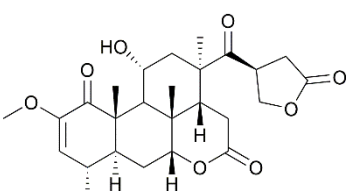
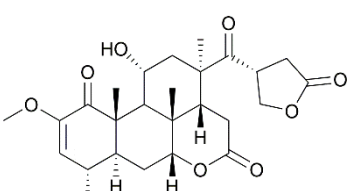
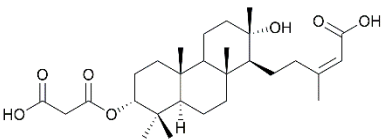
TrC-243	lintonolide D		<i>Cacospongia</i> cf. <i>linteiformis</i> ⁴¹²
TrC-244	lintonolide E		<i>Cacospongia</i> cf. <i>linteiformis</i> ⁴¹²
TrC-245	vulgaroside 1 (24,25-O-diacetylvulgaroside)		<i>Cydonia vulgaris</i> ⁴¹³
TrC-246	vulgaroside 2 (25-O-acetylvulgaroside)		<i>Cydonia vulgaris</i> ⁴¹³
TrC-247	vulgaroside 3 (24-O-acetyl-25-O-cinnamoylvulgaroside)		<i>Cydonia vulgaris</i> ⁴¹³
TrC-248	vulgaroside 4 (25-O-cinnamoylvulgaroside)		<i>Cydonia vulgaris</i> ⁴¹³
TrC-249	halisulfate 1 methylherbipoline		<i>Coscinoderma mathewsi</i> ⁴¹⁴

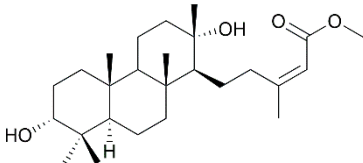
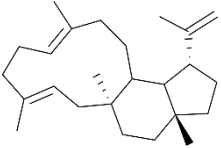
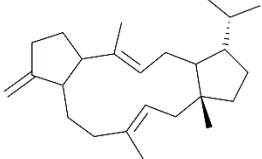
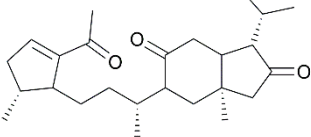
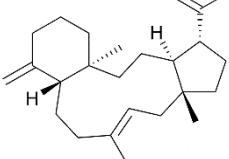
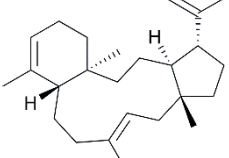
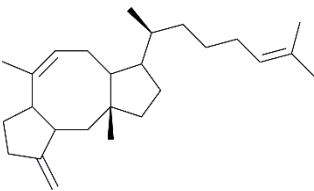
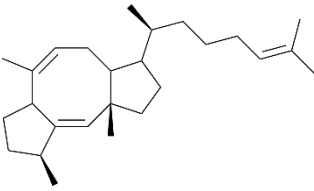
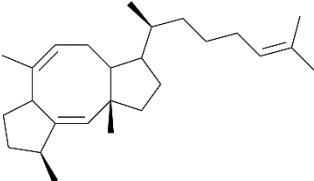
TrC-250	caminatal		<i>Suberites caminatus</i> ⁴¹⁵
TrC-251	cavernosolide		<i>Fasciospongia cavernosa</i> ⁴¹⁶ <i>Semitaspongia bactriana</i> ⁶⁹
TrC-252	suvanine methylherbipoline		<i>Coscinoderma mathewsi</i> ⁴¹⁴
TrC-253	lintenolide F		<i>Cacospongia</i> cf. <i>linteiformis</i> ⁴¹⁷
TrC-254	lintenolide G		<i>Cacospongia</i> cf. <i>linteiformis</i> ⁴¹⁷
TrC-255	auroral 1		<i>Rhabdastrella globostellata</i> ⁴¹⁸
TrC-256	auroral 2		<i>Rhabdastrella globostellata</i> ⁴¹⁸

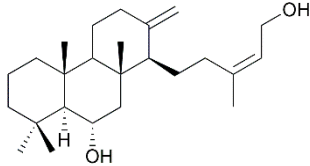
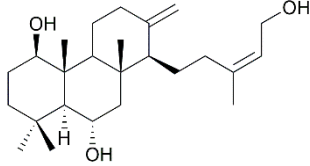
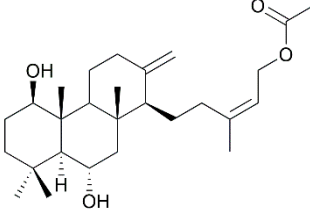
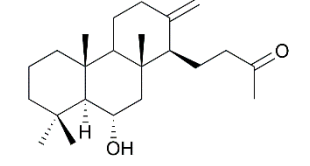
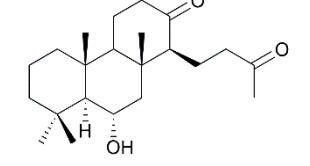
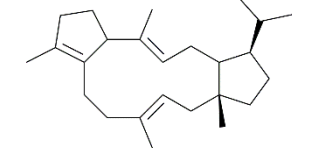
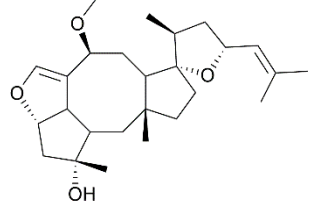
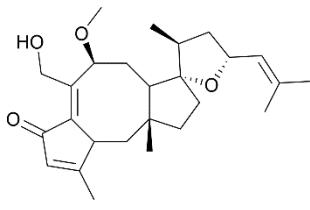
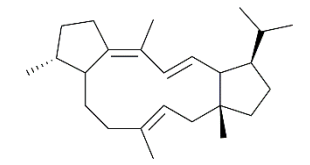
TrC-257	auroral 3		<i>Rhabdastrella globostellata</i> ⁴¹⁸
TrC-258	auroral 4		<i>Rhabdastrella globostellata</i> ⁴¹⁸
TrC-259	spongidine A		<i>Spongia</i> sp. ³⁹⁴
TrC-260	spongidine B		<i>Spongia</i> sp. ³⁹⁴
TrC-261	spongidine C		<i>Spongia</i> sp. ³⁹⁴
TrC-262	spongidine D		<i>Spongia</i> sp. ³⁹⁴
TrC-263	unnamed tricycyclic sesterterpene (reported as compound 10)		<i>Coscinoderma</i> sp. ³⁶²
TrC-264	globostelletin C		<i>Rhabdastrella globostellata</i> ⁴¹⁹
TrC-265	globostelletin D		<i>Rhabdastrella globostellata</i> ⁴¹⁹

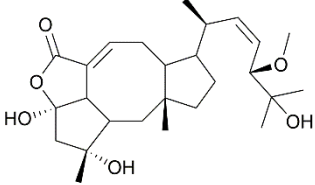
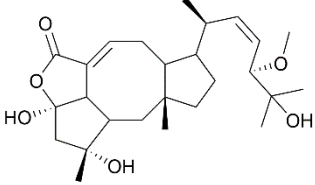
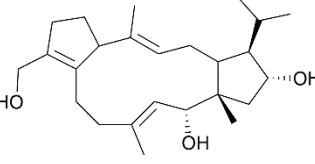
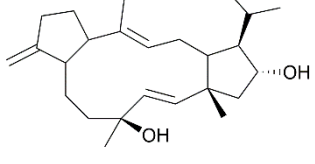
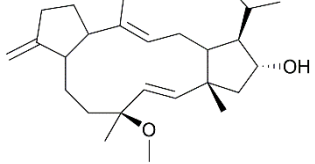
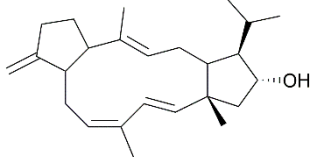
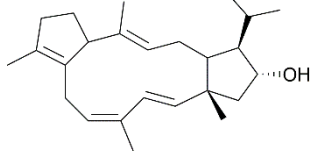
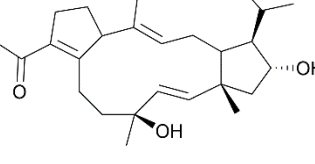
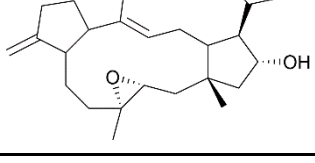
TrC-266	globostelletin E		<i>Rhabdastrella globostellata</i> ⁴¹⁹
TrC-267	globostelletin F		<i>Rhabdastrella globostellata</i> ⁴¹⁹
TrC-268	globostelletin G		<i>Rhabdastrella globostellata</i> ⁴¹⁹
TrC-269	unnamed tricycyclic sesterterpene (reported as compound 24)		<i>Coscinoderma mathewsi</i> ⁴²⁰
TrC-270	unnamed tricycyclic sesterterpene (reported as compound 25)		<i>Coscinoderma mathewsi</i> ⁴²⁰
TrC-271	unnamed tricycyclic sesterterpene (reported as compound 7)		<i>Coscinoderma</i> sp. ⁴²¹
TrC-272	unnamed tricycyclic sesterterpene (reported as compound 8)		<i>Coscinoderma</i> sp. ⁴²¹
TrC-273	flabelliferin A		<i>Carteriospongia flabellifera</i> ⁴²²

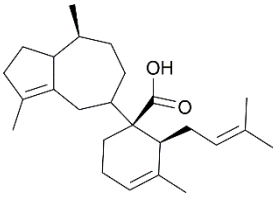
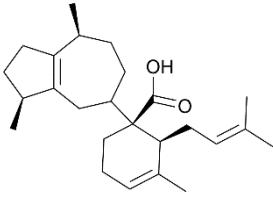
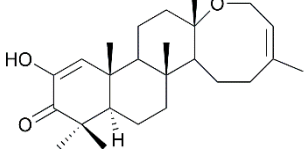
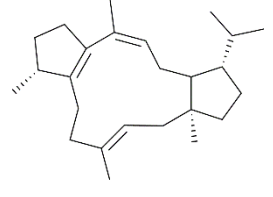
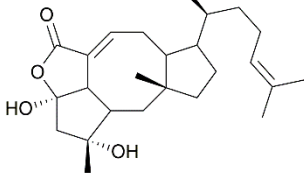
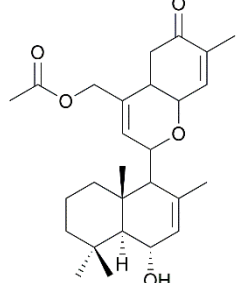
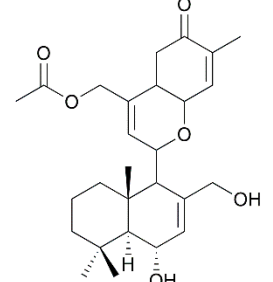
TrC-274	unnamed tricarbocyclic sesterterpene (reported as compound 1)		<i>Aletris farinosa</i> ⁴²³
TrC-275	unnamed tricarbocyclic sesterterpene (reported as compound 4)		<i>Aletris farinosa</i> ⁴²³
TrC-276	cristasesterterpenoic acid		<i>Caesalpinia crista</i> ⁴²⁴
TrC-277	cristasesterterpinol glucoside		<i>Caesalpinia crista</i> ⁴²⁴
TrC-278	involutispirone A		<i>Stahlianthus involucratus</i> ⁴²⁵
TrC-279	involutispirone B		<i>Stahlianthus involucratus</i> ⁴²⁵
TrC-280	luffolide		<i>Luffariella</i> sp. ⁴²⁶
TrC-281	inorolide C		<i>Chromodoris inornata</i> ⁴²⁷

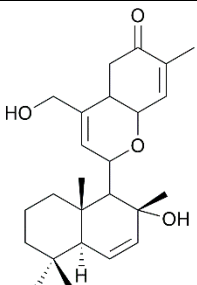
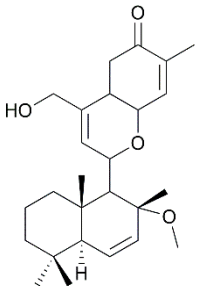
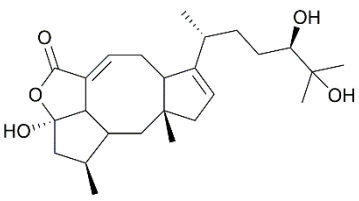
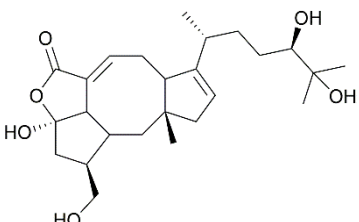
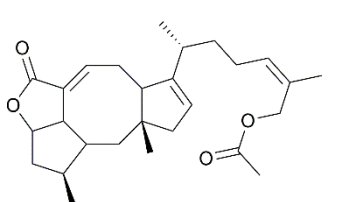
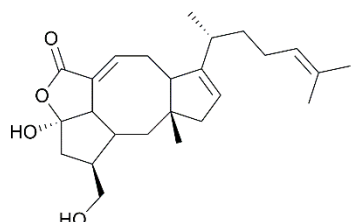
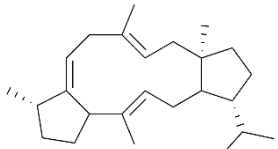
TrC-282	unnamed tricarbocyclic sesterterpene (reported as compound 4)		<i>Fasciospongia cavernosa</i> ⁴⁰⁹
TrC-283	25-deoxycacospongionolide		<i>Fasciospongia cavernosa</i> ⁴⁰⁹
TrC-284	unnamed tricarbocyclic sesterterpene (reported as compound 3)		<i>Fasciospongia cavernosa</i> ⁴⁰⁹
TrC-285	unnamed tricarbocyclic sesterterpene (reported as compound 4)		<i>Fasciospongia cavernosa</i> ⁴⁰⁹
TrC-286	gentianelloid A		<i>Gentianella turkestanorum</i> ⁴²⁸
TrC-287	gentianelloid B		<i>Gentianella turkestanorum</i> ⁴²⁸
TrC-288	2'-isopicrasin A		<i>Picrasma quassioides</i> ⁴²⁹
TrC-289	picrasin A		<i>Picrasma quassioides</i> ⁴³⁰
TrC-290	unnamed tricarbocyclic sesterterpene (reported as compound 2)		<i>Aletris farinosa</i> ⁴³¹

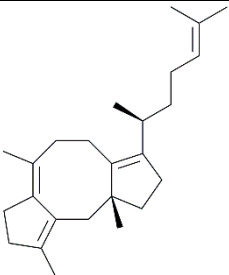
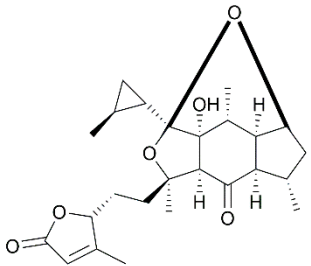
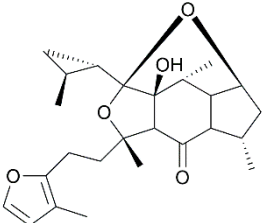
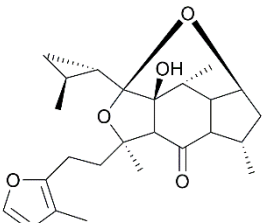
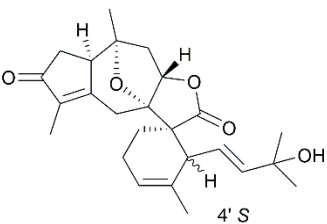
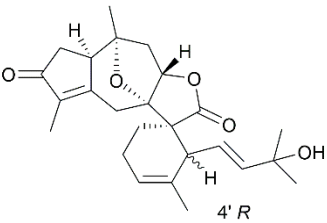
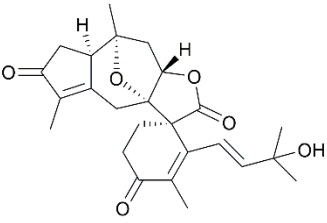
TrC-291	unnamed tricarbocyclic sesterterpene (reported as compound 3)		<i>Aletris farinosa</i> ⁴³¹
TrC-292	(2 <i>E</i>)-flocerene		<i>Arabidopsis thaliana</i> / <i>Escherichia coli</i> ⁴³²
TrC-293	sesterbrasiliatriene		<i>Penicillium brasilianum</i> ⁴³³
TrC-294	18- <i>epi</i> -alborosin		<i>Gentianella turkestanorum</i> ⁴³⁴
TrC-295	(-)-caprutriene B		<i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁵
TrC-296	(+)-caprutriene C		<i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁵
TrC-297	3,20-anhydrophiobolin F (ophiobolane 2)		<i>Aspergillus clavatus</i> ^{264, 435}
TrC-298	ophiobola-1,7,18-triene (ophiobolane 1)		<i>Aspergillus clavatus</i> ^{264, 435}
TrC-299	stellata-2,6,19-triene		<i>Aspergillus stellatus</i> ³⁵⁹

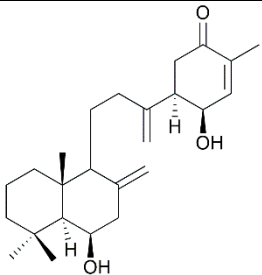
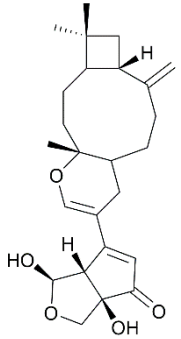
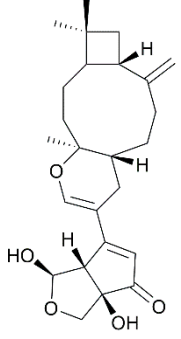
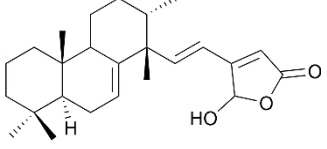
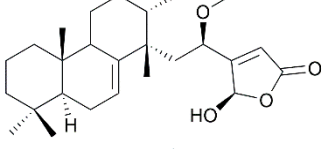
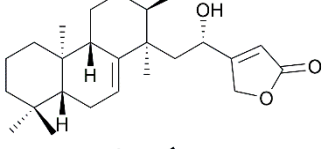
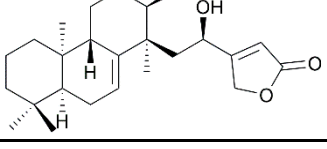
TrC-300	(17Z)-cheilantha-13(24),17-diene-6 α ,19-diol		<i>Aleuritopteris agetae</i> ⁴³⁶
TrC-301	(17Z)-cheilantha-13(24),17-diene-1 β ,6 α ,19-triol		<i>Aleuritopteris agetae</i> ⁴³⁶
TrC-302	(17Z)-19-acetoxycheilantha-13(24),17-diene-1 β ,6 α -diol		<i>Aleuritopteris agetae</i> ⁴³⁶
TrC-303	17-oxo-18,19-bisnorcheilanth-13(24)-en-6 α -ol		<i>Aleuritopteris agetae</i> ⁴³⁶
TrC-304	13,17-dioxo-18,19,24-trisnorcheilanth-6 α -ol		<i>Aleuritopteris agetae</i> ⁴³⁶
TrC-305	(-)-variculartriene A		<i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁵
TrC-306	maydispenoid A		<i>Bipolaris maydis</i> ⁴³⁷
TrC-307	maydispenoid B		<i>Bipolaris maydis</i> ⁴³⁷
TrC-308	Bm1		<i>Bipolaris maydis</i> ²⁶⁷ <i>Aspergillus oryzae</i> ²⁶⁷

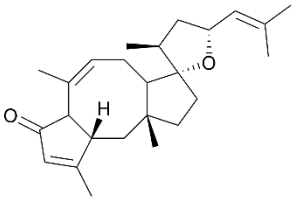
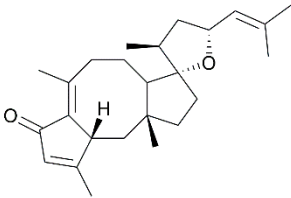
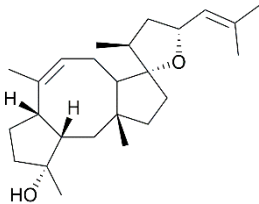
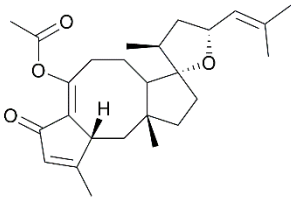
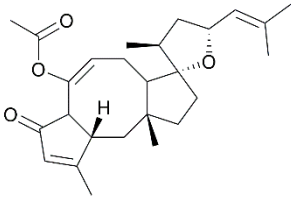
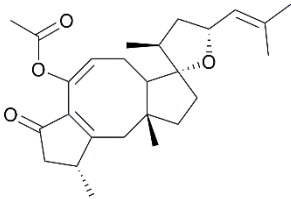
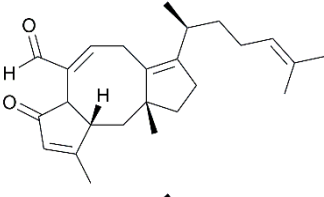
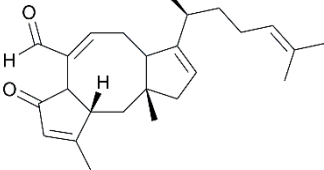
TrC-309	18,19-dihydro-19-hydroxy-18-methoxyphiobolin P		<i>Aspergillus</i> sp. RR-YLW-12/ <i>Rhodomela confervoides</i> ³⁸⁷
TrC-310	18,19-dihydro-19-hydroxy-18- <i>epi</i> -methoxyphiobolin P		<i>Aspergillus</i> sp. RR-YLW-12/ <i>Rhodomela confervoides</i> ³⁸⁷
TrC-311	spectanoid A		<i>Aspergillus spectabilis</i> ⁴³⁸
TrC-312	spectanoid B		<i>Aspergillus spectabilis</i> ⁴³⁸
TrC-313	spectanoid C		<i>Aspergillus spectabilis</i> ⁴³⁸
TrC-314	spectanoid D		<i>Aspergillus spectabilis</i> ⁴³⁸
TrC-315	spectanoid E		<i>Aspergillus spectabilis</i> ⁴³⁸
TrC-316	spectanoid F		<i>Aspergillus spectabilis</i> ⁴³⁸
TrC-317	spectanoid G		<i>Aspergillus spectabilis</i> ⁴³⁸

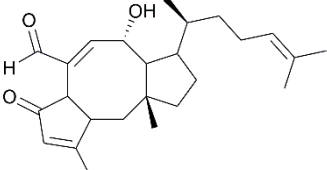
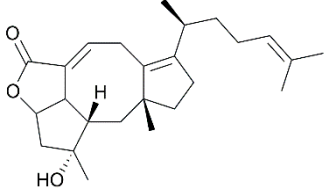
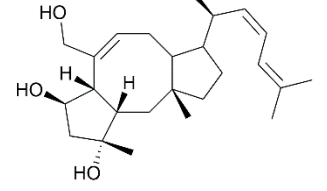
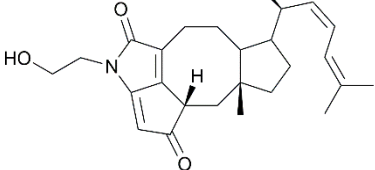
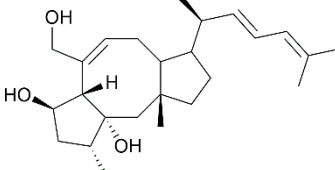
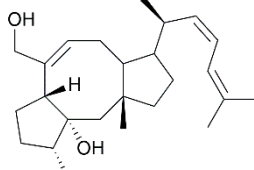
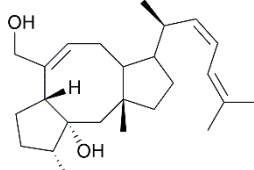
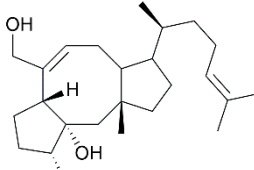
TrC-318	linderasesterterpenoid A		<i>Lindera glauca</i> ⁴³⁹
TrC-319	linderasesterterpenoid B		<i>Lindera glauca</i> ⁴³⁹
TrC-320	ancepsone A		<i>Aleuritopteris anceps</i> ⁴⁴⁰
TrC-321	schultriene		<i>Aspergillus oryzae</i> ⁴⁴¹
TrC-322	ophiobolin P1		<i>Aspergillus calidoustus</i> ⁴⁴²
TrC-323	ansellone H		<i>Phorbas</i> sp. ⁴⁴³
TrC-324	ansellone I		<i>Phorbas</i> sp. ⁴⁴³

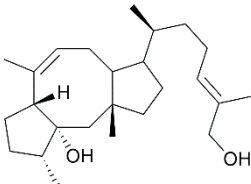
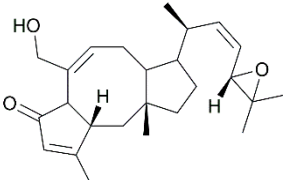
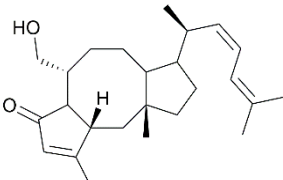
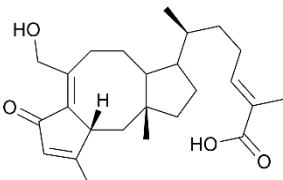
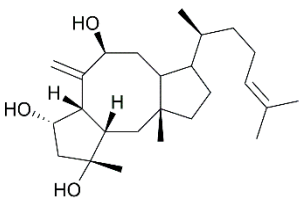
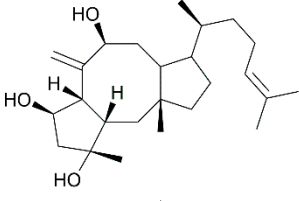
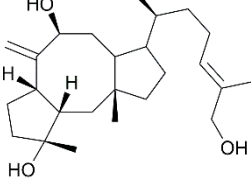
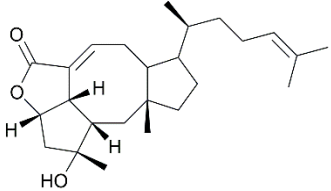
TrC-325	ansellone J		<i>Phorbas</i> sp. ⁴⁴³
TrC-326	ansellone K		<i>Phorbas</i> sp. ⁴⁴³
TrC-327	cyophiobiolin A		<i>Cytospora rhizophorae</i> A761/ <i>Gynochthodes officinalis</i> ⁴⁴⁴
TrC-328	cyophiobiolin B		<i>Cytospora rhizophorae</i> A761/ <i>Gynochthodes officinalis</i> ⁴⁴⁴
TrC-329	cyophiobiolin C		<i>Cytospora rhizophorae</i> A761/ <i>Gynochthodes officinalis</i> ⁴⁴⁴
TrC-330	cyophiobiolin D		<i>Cytospora rhizophorae</i> A761/ <i>Gynochthodes officinalis</i> ⁴⁴⁴
TrC-331	sesterviolene B		<i>Streptomyces violens</i> ²⁷³

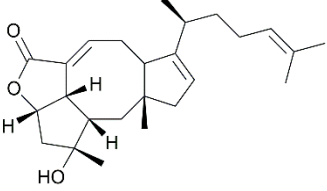
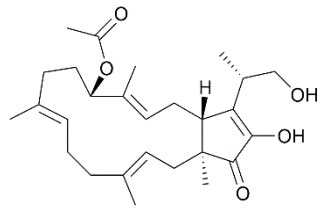
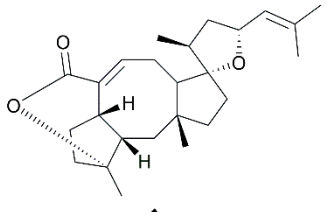
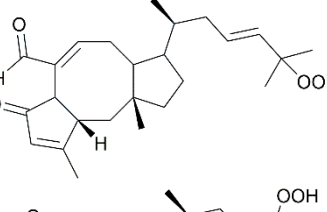
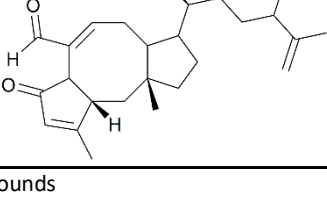
TrC-332	nigtetraene		<i>Saccharomyces cerevisiae</i> ⁴⁴¹
TrC-333	colquhounoid A		<i>Colquhounia coccinea</i> var. <i>mollis</i> ²⁵¹
TrC-334	colquhounoid D		<i>Colquhounia coccinea</i> var. <i>mollis</i> ⁴⁴⁵
TrC-335	14- <i>epi</i> -colquhounoid D		<i>Colquhounia coccinea</i> var. <i>mollis</i> ⁴⁴⁵
TrC-336	orientanoid A		<i>Hedyosmum orientale</i> ⁴⁴⁶
TrC-337	orientanoid B		<i>Hedyosmum orientale</i> ⁴⁴⁶
TrC-338	orientanoid C		<i>Hedyosmum orientale</i> ⁴⁴⁶

TrC-339	secosuberitenone A		<i>Suberites</i> sp. ⁴⁴⁷
TrC-340	patriniaterpene A		<i>Patrinia scabra</i> ²⁷⁶
TrC-341	patriniaterpene B		<i>Patrinia scabra</i> ²⁷⁶
TrC-342	sarcotragusolide A		<i>Sarcotragus</i> sp. ²⁷⁸
TrC-343	sarcotragusolide B		<i>Sarcotragus</i> sp. ²⁷⁸
TrC-344	sarcotragusolide C		<i>Sarcotragus</i> sp. ²⁷⁸
TrC-345	sarcotragusolide D		<i>Sarcotragus</i> sp. ²⁷⁸

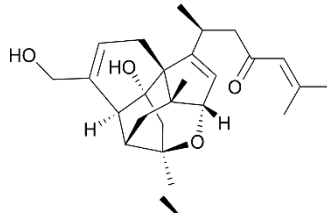
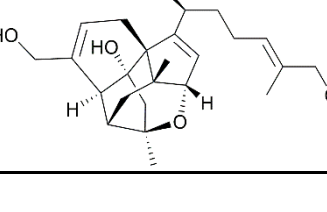
TrC-346	bipolaricin J		<i>Bipolaris eleusines</i> ⁴⁴⁸
TrC-347	bipolaricin K		<i>Bipolaris eleusines</i> ⁴⁴⁸
TrC-348	bipolaricin L		<i>Bipolaris eleusines</i> ⁴⁴⁸
TrC-349	bipolaricin M		<i>Bipolaris eleusines</i> ⁴⁴⁸
TrC-350	bipolaricin N		<i>Bipolaris eleusines</i> ⁴⁴⁸
TrC-351	bipolaricin O		<i>Bipolaris eleusines</i> ⁴⁴⁸
TrC-352	bipolaricin P		<i>Bipolaris eleusines</i> ⁴⁴⁸
TrC-353	bipolaricin Q		<i>Bipolaris eleusines</i> ⁴⁴⁸

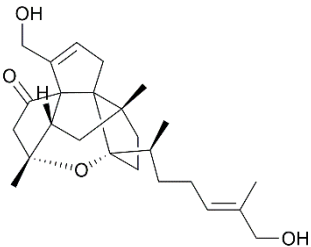
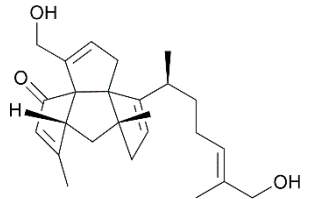
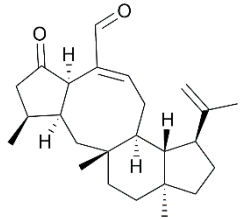
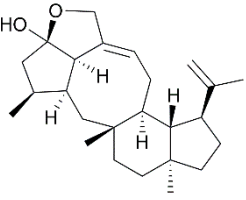
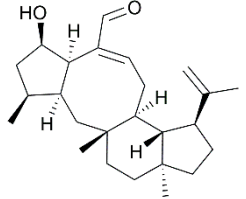
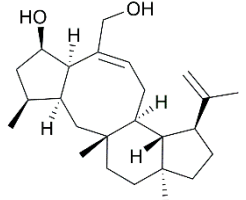
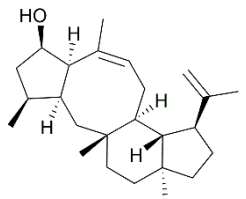
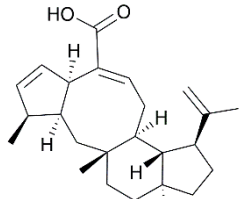
TrC-354	bipolaricin R		<i>Bipolaris eleusines</i> ⁴⁴⁸
TrC-355	bipolaricin S		<i>Bipolaris eleusines</i> ⁴⁴⁸
TrC-356	21-deoxo-21-hydroxyphiobolin U		<i>Aspergillus</i> sp. RR-YLW-12 ⁴⁴⁹
TrC-357	undobolin A		<i>Aspergillus undulatus</i> ⁴⁵⁰
TrC-358	undobolin B		<i>Aspergillus undulatus</i> ⁴⁵⁰
TrC-359	undobolin C		<i>Aspergillus undulatus</i> ⁴⁵⁰
TrC-360	undobolin D		<i>Aspergillus undulatus</i> ⁴⁵⁰
TrC-361	undobolin E		<i>Aspergillus undulatus</i> ⁴⁵⁰

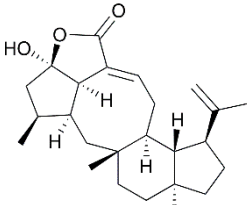
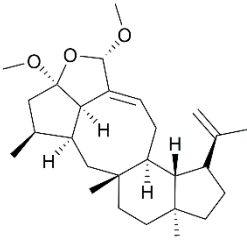
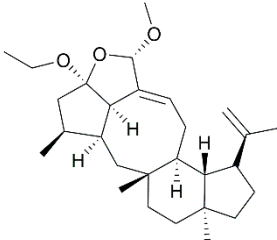
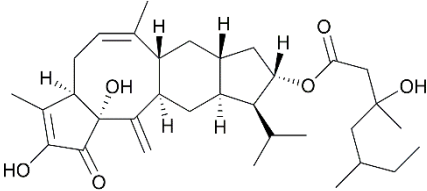
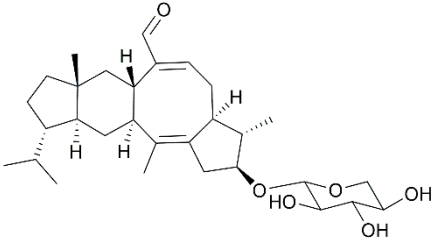
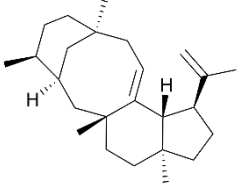
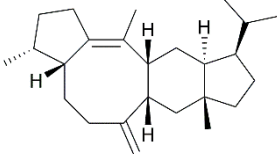
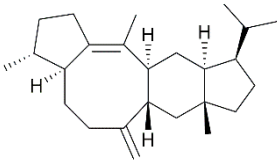
TrC-362	undobolin F		<i>Aspergillus undulatus</i> ⁴⁵⁰
TrC-363	undobolin G		<i>Aspergillus undulatus</i> ⁴⁵⁰
TrC-364	undobolin H		<i>Aspergillus undulatus</i> ⁴⁵⁰
TrC-365	undobolin I		<i>Aspergillus undulatus</i> ⁴⁵⁰
TrC-366	undobolin J		<i>Aspergillus undulatus</i> ⁴⁵⁰
TrC-367	undobolin K		<i>Aspergillus undulatus</i> ⁴⁵⁰
TrC-368	undobolin L		<i>Aspergillus undulatus</i> ⁴⁵⁰
TrC-369	bipoladien C		<i>Bipolaris maydis</i> ⁴⁵¹

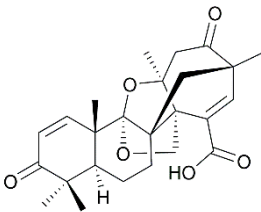
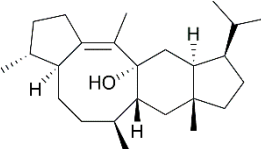
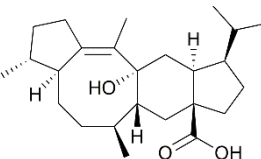
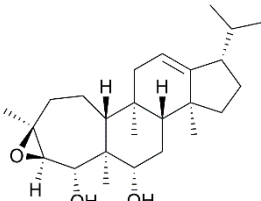
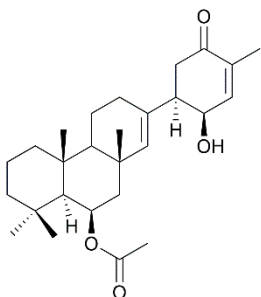
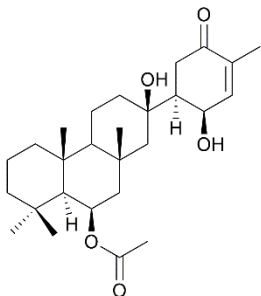
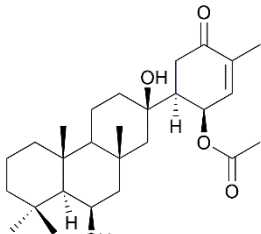
TrC-370	bipoladien D		<i>Bipolaris maydis</i> ⁴⁵¹
TrC-371	bipoladien E		<i>Bipolaris maydis</i> ⁴⁵¹
TrC-372	unnamed ophiobolin analog		<i>Bipolaris eleusines</i> ⁴⁵²
TrC-373	unnamed ophiobolin analog		<i>Bipolaris eleusines</i> ⁴⁵²
TrC-374	unnamed ophiobolin analog		<i>Bipolaris eleusines</i> ⁴⁵²

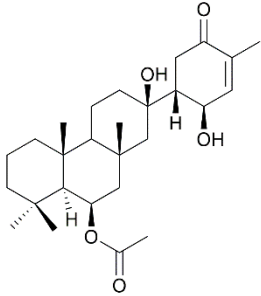
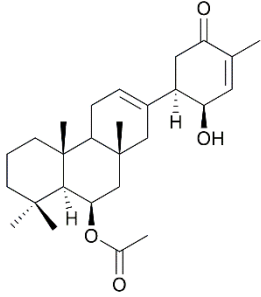
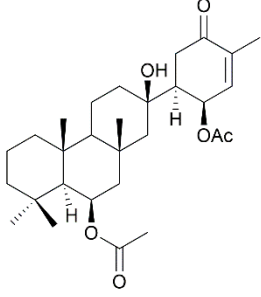
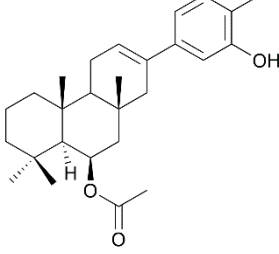
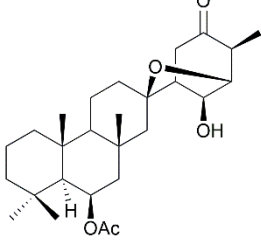
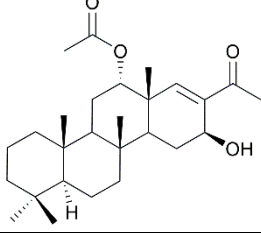
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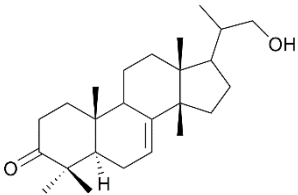
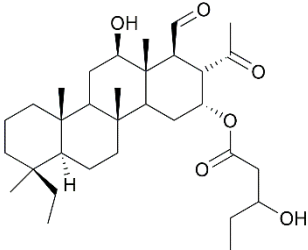
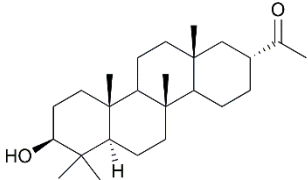
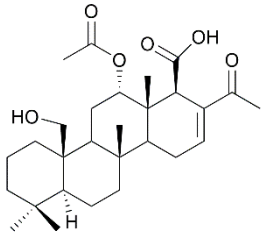
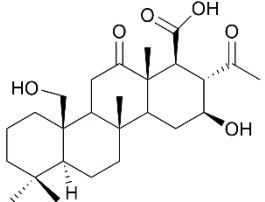
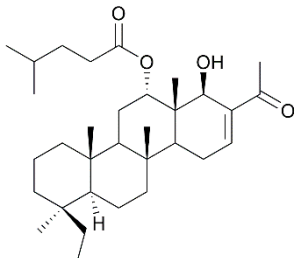
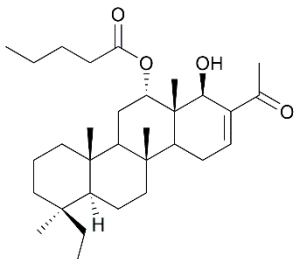
Section 1.8. Structures of tetracarboxylic (TeC) sesterterpenoids (with and without hetherocycles) and their dimers			
c. n.	IUPAC name or Trivial name	Structure	Organism as reported in the reference (reference)
TeC-1	bipolarolide A		<i>Bipolaris</i> sp. TJ403-B1 ¹⁷³
TeC-2	bipolarolide B		<i>Bipolaris</i> sp. TJ403-B1 ¹⁷³

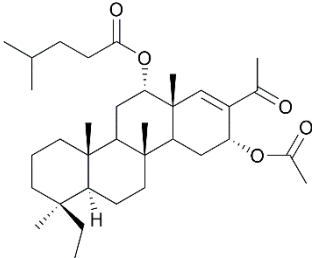
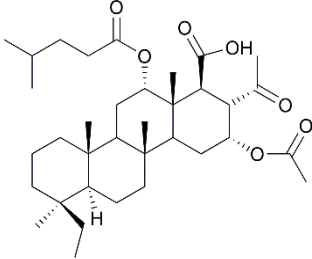
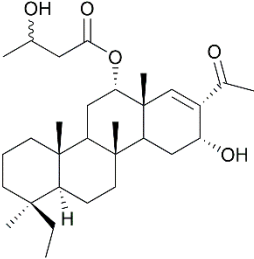
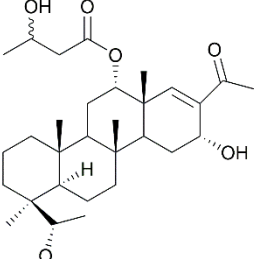
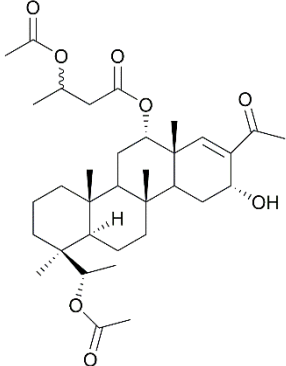
TeC-3	bipolarolide C		<i>Bipolaris</i> sp. TJ403-B1 ¹⁷³
TeC-4	bipolarolide D		<i>Bipolaris</i> sp. TJ403-B1 ¹⁷³
TeC-5	variecolin		<i>Aspergillus variecolor</i> ⁴⁵³ <i>Emericella aurantiobrunnea</i> ^{454, 455} <i>Emericella purpurea</i> ⁴⁵⁶ <i>Phoma</i> sp. ⁴⁵⁷
TeC-6	variecolol		<i>Emericella aurantiobrunnea</i> ⁴⁵⁴ <i>Emericella purpurea</i> ⁴⁵⁸
TeC-7	emericolin A		<i>Emericella aurantiobrunnea</i> ⁴⁵⁴
TeC-8	emericolin B		<i>Emericella aurantiobrunnea</i> ⁴⁵⁴
TeC-9	emericolin C		<i>Emericella aurantiobrunnea</i> ⁴⁵⁴
TeC-10	emericolin D		<i>Emericella aurantiobrunnea</i> ⁴⁵⁴

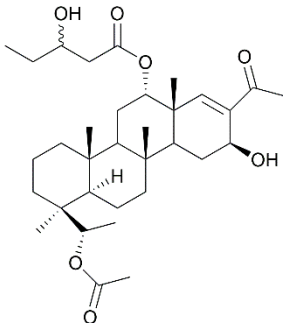
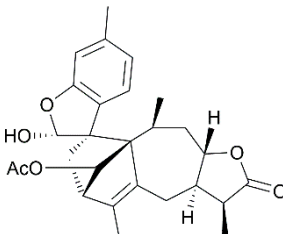
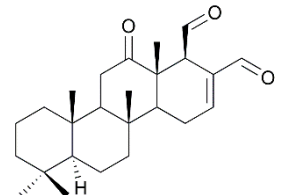
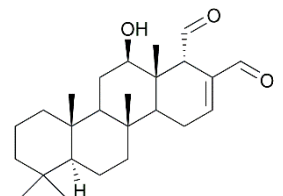
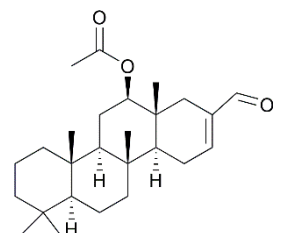
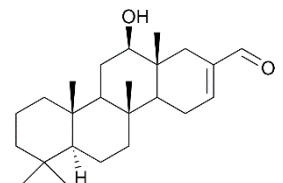
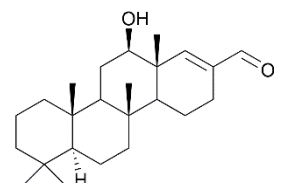
TeC-11	variecolactone		<i>Emericella aurantiobrunnea</i> ^{455, 459} <i>Emericella purpurea</i> ⁴⁵⁸
TeC-12	variecoacetal A		<i>Emericella aurantiobrunnea</i> ⁴⁵⁵
TeC-13	variecoacetal B		<i>Emericella aurantiobrunnea</i> ⁴⁵⁵
TeC-14	YW3699		<i>Codinaea simplex</i> ⁴⁶⁰
TeC-15	aleurodiscal		<i>Aleurodiscus mirabilis</i> ⁴⁶¹
TeC-16	astellifadiene		<i>Emericella variecolor/Aspergillus oryzae</i> ⁴⁶²
TeC-17	(-)-aleurodiscalene A		<i>Nicotiana benthamiana/Agrobacterium tumefaciens</i> ²⁶⁶
TeC-18	(+)-aleurodiscalene B		<i>Nicotiana benthamiana/Agrobacterium tumefaciens</i> ²⁶⁶

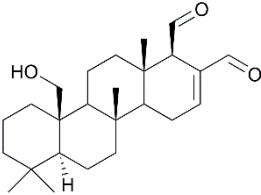
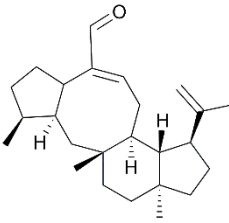
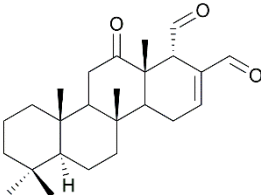
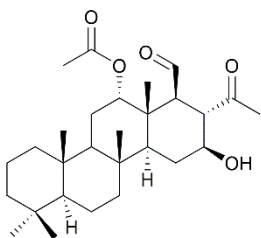
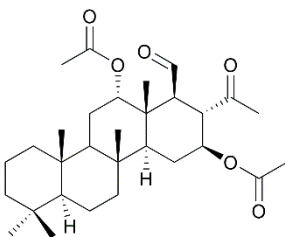
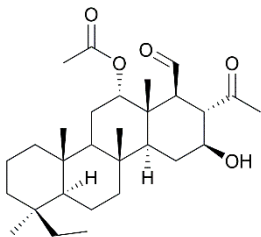
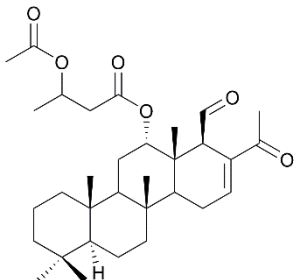
TeC-19	emericellic acid		<i>Emericella variecolor</i> ⁴⁶³
TeC-20	sesterfisherol		<i>Aspergillus oryzae</i> / <i>Neosartorya fischeri</i> ⁴⁶⁴
TeC-21	sesterfisheric acid		<i>Aspergillus oryzae</i> / <i>Neosartorya fischeri</i> ⁴⁶⁴
TeC-22	aspergilloxide		<i>Aspergillus</i> spp. ⁴⁶⁵ <i>Aspergillus unguis</i> ⁴⁶⁶
TeC-23	Suberitenone A		<i>Phorbas areolatus</i> ⁴⁶⁷ <i>Suberites</i> sp. ⁴⁶⁸
TeC-24	suberitenone B		<i>Phorbas areolatus</i> ⁴⁶⁷ <i>Suberites</i> sp. ⁴⁶⁸
TeC-25	isosuberitenone B		<i>Phorbas areolatus</i> ⁴⁶⁷

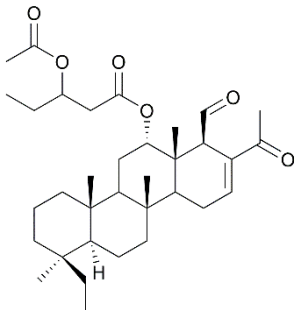
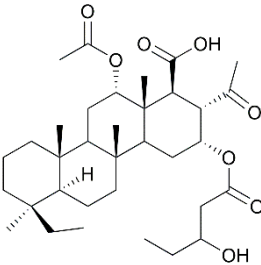
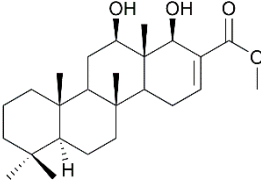
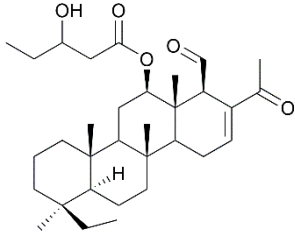
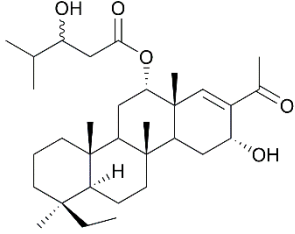
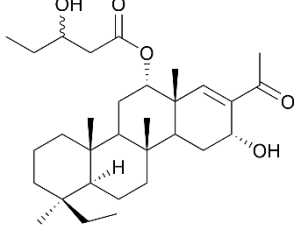
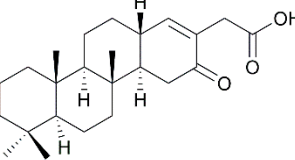
TeC-26	19- <i>epi</i> -suberitenone		<i>Phorbas areolatus</i> ⁴⁶⁷ <i>Suberites caminatus</i> ⁴⁶⁹
TeC-27	suberitenone C		<i>Suberites</i> sp. ⁴⁷⁰
TeC-28	suberitenone D		<i>Suberites</i> sp. ⁴⁷⁰
TeC-29	suberiphenol		<i>Suberites</i> sp. ⁴⁷⁰
TeC-30	oxaspirosuberitenone		<i>Phorbas areolatus</i> ⁴⁶⁷ <i>Suberites caminatus</i> ⁴⁶⁹
TeC-31	flabelliferin B		<i>Cateriospongia flabellifera</i> ⁴²²

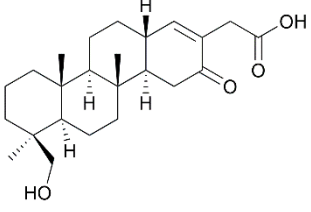
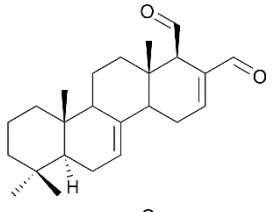
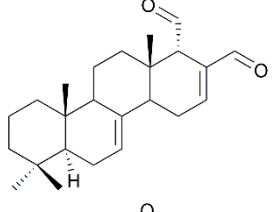
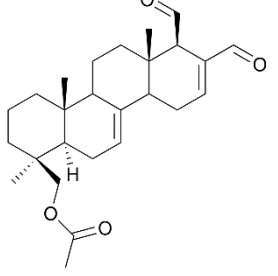
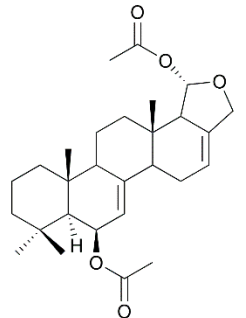
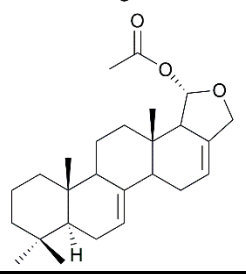
TeC-32	phellogine		<i>Phellodendron chinense</i> var. <i>glabriusculum</i> ⁴⁷¹
TeC-33	foliaspongin		<i>Carteriospongia foliascens</i> ⁴⁷² reported as <i>Phyllospongia foliascens</i> ⁴⁷³
TeC-34	perisomalien A		<i>Periploca somaliensis</i> ⁴⁷⁴
TeC-35	lendenfeldarane A		<i>Lendenfeldia</i> sp. ⁴⁷⁵
TeC-36	lendenfeldarane B		<i>Lendenfeldia</i> sp. ⁴⁷⁵
TeC-37	carteriofenone A		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁴⁷⁶
TeC-38	carteriofenone B		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁴⁷⁶

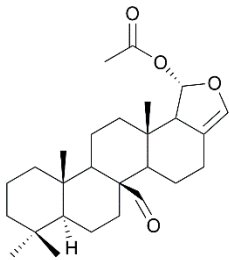
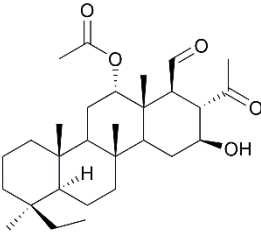
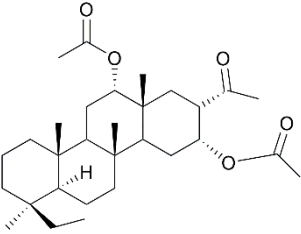
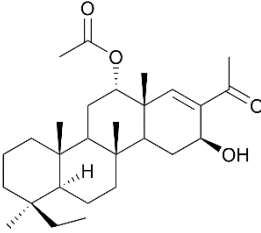
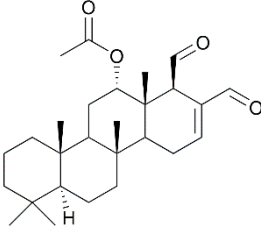
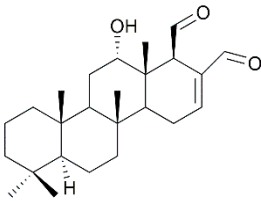
TeC-39	carteriofenone C		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁴⁷⁶
TeC-40	carteriofenone D		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁴⁷⁶
TeC-41	unnamed tetracycyclic sesterterpene (reported as compound 3)		<i>Dysidea granulosa</i> ³⁷⁰
TeC-42	unnamed tetracycyclic sesterterpene (reported as compound 5)		<i>Dysidea granulosa</i> ³⁷⁰
TeC-43	unnamed tetracycyclic sesterterpene (reported as compound 6)		<i>Dysidea granulosa</i> ³⁷⁰

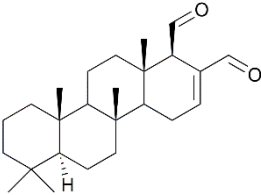
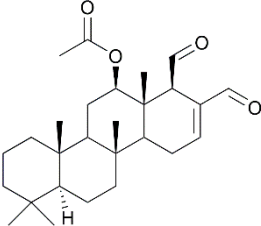
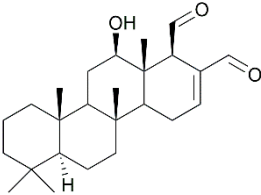
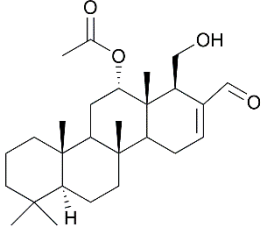
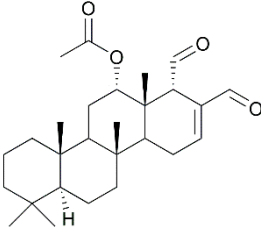
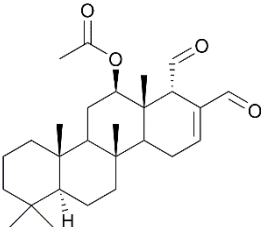
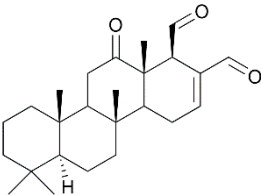
TeC-44	unnamed tetracarboxylic sesterterpene (reported as compound 7)		<i>Dysidea granulosa</i> ³⁷⁰
TeC-45	dibritannilactone A		<i>Inula britannica</i> ⁴⁷⁷
TeC-46	12-deacetyl-12-oxoscalaradial		<i>Glossodoris rufomarginata</i> ⁴⁷⁸ <i>Glossodoris pallida</i> ⁴⁷⁹
TeC-47	12-deacetyl-12,18-di-epi-scalaradial		<i>Spongia idia</i> ²⁶ <i>Hyrtios erectus</i> ⁴⁸⁰
TeC-48	hyrtial		<i>Hyrtios erecta</i> [sic!] ⁴⁸¹
TeC-49	12-deacetylhyrtial		<i>Collospongia auris</i> ⁴⁸² <i>Hyrtios erectus</i> ⁴⁸³
TeC-50	12-deacetyl- Δ^{17} -hyrtial		<i>Hyrtios erectus</i> ⁴⁸³

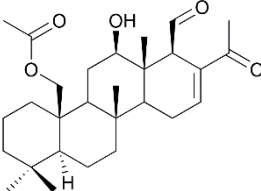
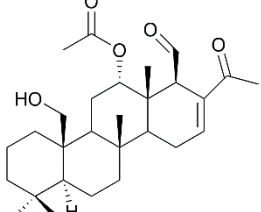
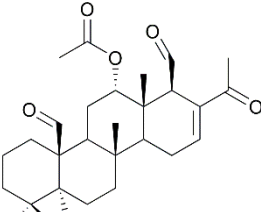
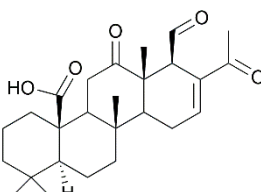
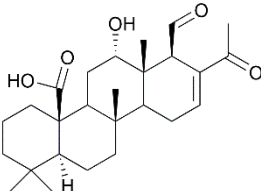
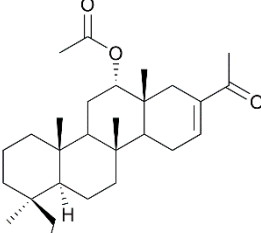
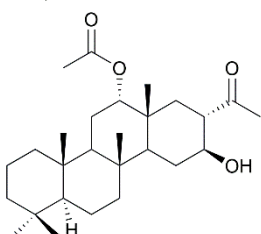
TeC-51	12-deacetoxy-23-hydroxyscalaradial		<i>Psammocinia</i> sp. ⁴⁸⁴
TeC-52	5-deoxovariecolin		<i>Aspergillus aegyptiacus</i> [sic!] ⁴⁸⁵
TeC-53	12-deacetyl-18- <i>epi</i> -12-oxoscalaradial		<i>Glossodoris rufomarginata</i> (= <i>Chromodoris youngbleuthi</i>) ⁴⁸⁶
TeC-54	unnamed tetracycyclic sesterterpene (reported as compound 1)		<i>Phyllospongia</i> sp. ⁴⁸⁷
TeC-55	unnamed tetracycyclic sesterterpene (reported as compound 1a)		<i>Phyllospongia</i> sp. ⁴⁸⁷
TeC-56	unnamed tetracycyclic sesterterpene (reported as compound 2)		<i>Phyllospongia</i> sp. ⁴⁸⁷
TeC-57	12 α -(3'-acetobutanoyloxy)-20,24-dimethyl-24-oxoscalar-16-en-25-al		<i>Strepsichordaia lendenfeldi</i> ⁴⁸⁸

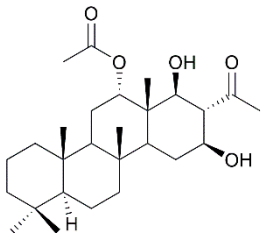
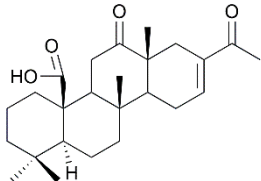
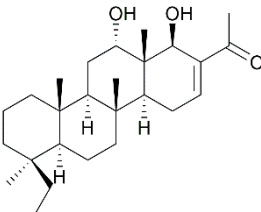
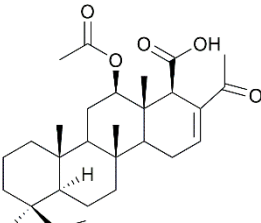
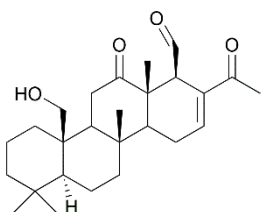
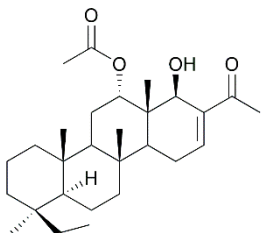
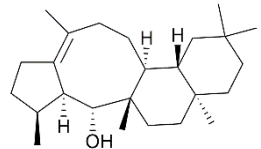
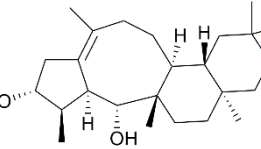
TeC-58	12 α -(3'-acetoxy-pentanoyloxy)-20,24-dimethyl-24-oxoscalar-16-en-25-al		<i>Strepsichordaia lendenfeldi</i> ⁴⁸⁸
TeC-59	12 α -acetoxy-16 α -(3'-hydroxypentanoyloxy)-20,24-dimethyl-24-oxoscalaran-25 β -oic acid		<i>Phyllospongia papyracea</i> ⁴⁸⁹
TeC-60	methyl 18-hydroxy-19-norscalar-16-en-20-carboxylate		<i>Collospongia auris</i> ⁴⁸² <i>Spongia</i> sp. ⁴⁹⁰
TeC-61	12 β -(3' β -hydroxypentanoyloxy)-20,24-dimethyl-24-oxoscalara-16-en-25-al		<i>Carteriospongia</i> sp. ⁴⁹¹
TeC-62	unnamed tetracycyclic sesterterpene (reported as compound 1)		<i>Carteriospongia foliascens</i> ⁴⁹²
TeC-63	unnamed tetracycyclic sesterterpene (reported as compound 2)		<i>Carteriospongia foliascens</i> ⁴⁹²
TeC-64	petrosaspongiolide K		<i>Petrosaspongia nigra</i> ⁴⁰⁵

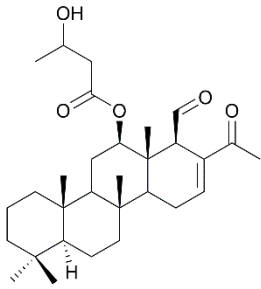
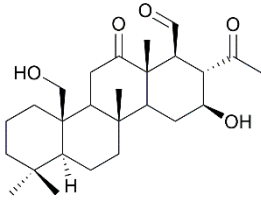
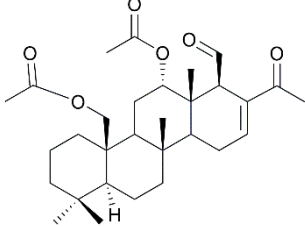
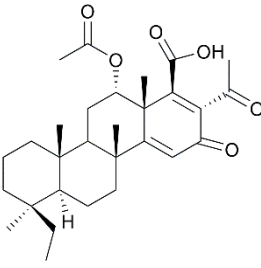
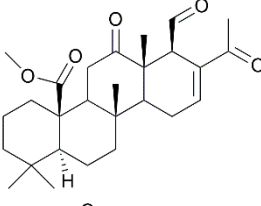
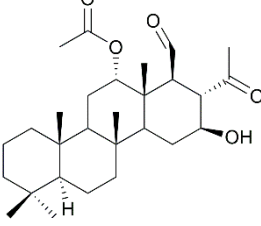
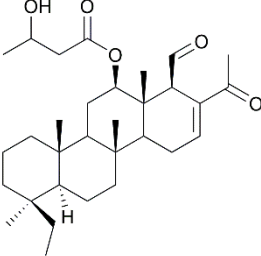
TeC-65	21-hydroxy- petrosaspongiolide K		<i>Spongia</i> sp. ³⁹⁴
TeC-66	mooloolabene A		<i>Hyattella intestinalis</i> ⁴⁹³
TeC-67	mooloolabene B		<i>Hyattella intestinalis</i> ⁴⁹³
TeC-68	mooloolabene C		<i>Hyattella intestinalis</i> ⁴⁹³
TeC-69	mooloolabene D		<i>Hyattella intestinalis</i> ⁴⁹³ <i>Dorisprismatica</i> (= <i>Glossodoris</i>) <i>atromarginata</i> ⁴⁹⁴
TeC-70	mooloolabene E		<i>Hyattella intestinalis</i> ⁴⁹³

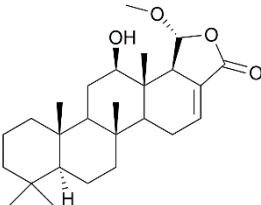
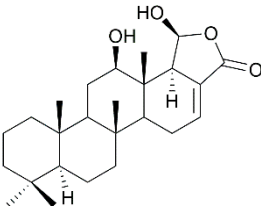
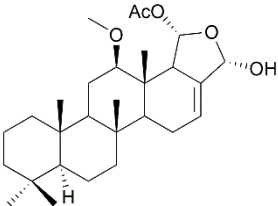
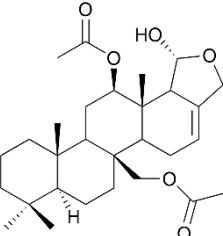
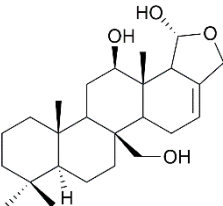
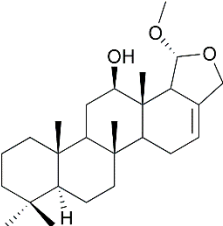
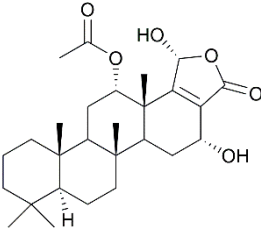
TeC-71	mooloolaldehyde		<i>Hyattella intestinalis</i> ⁴⁹³ <i>Dorisprismatica</i> (= <i>Glossodoris</i>) <i>atromarginata</i> ⁴⁹⁴
TeC-72	12 α -acetoxy-16 β -hydroxy-20,24-dimethyl-24-oxo-25-norscalarane		<i>Carteriospongia foliascens</i> ⁴⁹⁵
TeC-73	12 α ,16 α -diacetoxy-20,24-dimethyl-25-norsclaran-24-one		<i>Carteriospongia foliascens</i> ⁴⁹⁶
TeC-74	(12 α ,16 β)-12-acetoxy-16-hydroxy-20,24-dimethyl-25-norscalar-17-en-24-one		<i>Phyllospongia papyracea</i> ⁴⁹⁷
TeC-75	scalaradial		<i>Cacospongia mollior</i> ^{498, 499} <i>Cacospongia scalaris</i> ^{500, 501} <i>Dysidea herbacea</i> ⁵⁰² <i>Dysidea</i> sp. ⁵⁰³ <i>Glossodoris rufomarginata</i> ⁴⁷⁸ <i>Glossodoris tricolor</i> ⁵⁰⁴ <i>Hyrrios erecta</i> [sic!] ⁵⁰⁵ <i>Chromodoris youngbleuthi</i> and <i>Spongia oceanica</i> ⁴⁸⁶ <i>Spongia officinalis</i> ³⁵ <i>Glossodoris pallida</i> ^{479, 506} <i>Glossodoris cincta</i> ⁵⁰⁶ <i>Glossodoris hikuerensis</i> ⁵⁰⁷ <i>Glossodoris vespa</i> ⁵⁰⁷ <i>Cacospongia scalaris</i> ⁵⁰⁰ <i>Dysidea</i> sp. ⁵⁰³ <i>Glossodoris pallida</i> and <i>Cacospongia</i> sp. ⁴⁷⁹
TeC-76	desacetylscalaradial (deacetylscalaradial)		

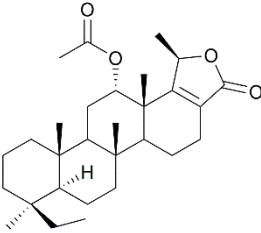
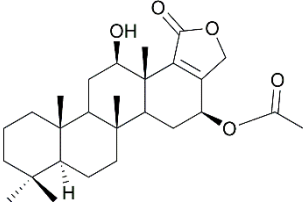
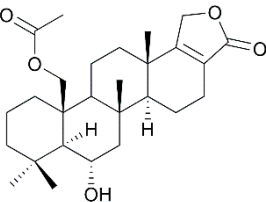
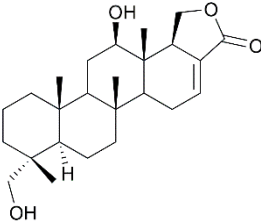
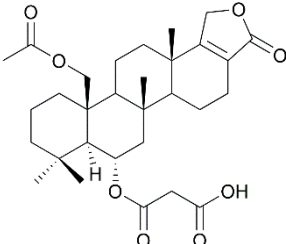
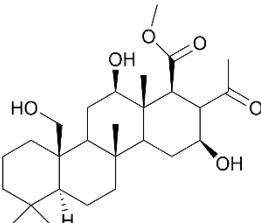
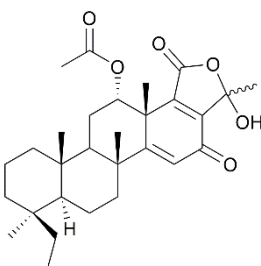
TeC-77	12-deacetoxyscalaradial (scalarenedial)		<i>Cacospongia mollior</i> ⁵⁰⁸ <i>Cacospongia scalaris</i> ⁵⁰¹ <i>Dysidea</i> sp. ⁵⁰³ <i>Psammocinia</i> sp. ⁴⁸⁴
TeC-78	12- <i>epi</i> -scalaradial		<i>Spongia nitens</i> ⁵⁰⁹ <i>Hyrtios erecta</i> [sic!] ⁵⁰⁵ <i>Collospongia auris</i> ⁴⁸² <i>Glossodoris cincta</i> ⁵⁰⁶
TeC-79	12-deacetyl-12- <i>epi</i> - scalaradial		<i>Chromodoris youngbleuthi</i> (<i>Glossodoris rufomarginata</i>) ⁴⁸⁶ <i>Collospongia auris</i> ⁴⁸² <i>Hyrtios erecta</i> [sic!] ⁵⁰⁵ <i>Spongia oceanica</i> ⁴⁸⁶
TeC-80	19-dihydroscalaradial		<i>Cacospongia scalaris</i> ⁵⁰¹
TeC-81	18- <i>epi</i> -scalaradial		<i>Cacospongia scalaris</i> ⁵⁰¹
TeC-82	12,18-di- <i>epi</i> -scalaradial		<i>Collospongia auris</i> ⁴⁸² <i>Spongia nitens</i> ⁵⁰⁹ <i>Hyrtios erecta</i> [sic!] ⁵⁰⁵
TeC-83	12-deacetoxy-12-oxo- scalaradial		<i>Glossodoris rufomarginata</i> ⁴⁷⁸ <i>Glossodoris pallida</i> ⁴⁷⁹ <i>Glossodoris averni</i> ⁴⁷⁹ <i>Glossodoris vespa</i> ⁴⁷⁹ <i>Glossodoris hikuerensis</i> ⁵⁰⁷

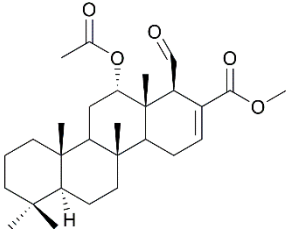
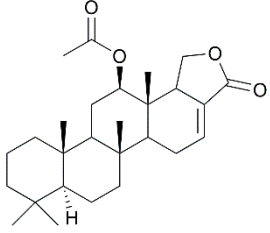
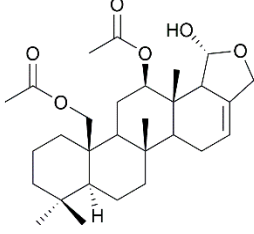
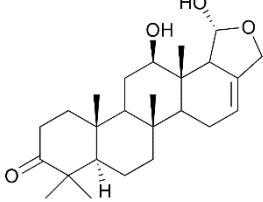
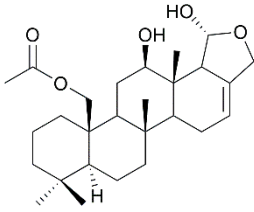
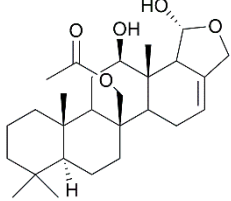
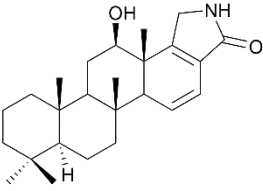
TeC-84	12-deacetyl-23-acetoxy-20-methyl-12- <i>epi</i> -scalaradial		<i>Glossodoris sedna</i> ⁵¹⁰
TeC-85	12 α -acetoxy-22-hydroxy-24-methyl-24-oxoscalar-16-en-25-al or PHC-4		<i>Dictyoceratida</i> sp. and <i>Halichondria</i> sp. ⁵¹¹ <i>Carteriospongia</i> sp. ⁵¹² <i>Lendenfeldia</i> sp. ⁴⁷⁵ <i>Phyllospongia chondrodes</i> ⁵¹³
TeC-86	12 α -acetoxy-24-methyl-24-oxoscalar-16-en-22,25-dial or PHC-2		<i>Dictyoceratida</i> sp. and <i>Halichondria</i> sp. ⁵¹¹ <i>Lendenfeldia chondrodes</i> ⁵¹⁴ <i>Phyllospongia chondrodes</i> ⁵¹³
TeC-87	24-methyl-12,24,25-trioxoscalar-16-en-22-oic acid or PHC-1		<i>Dictyoceratida</i> sp. and <i>Halichondria</i> sp. ⁵¹¹ <i>Lendenfeldia chondrodes</i> ⁵¹⁴ <i>Lendenfeldia</i> sp. ^{84, 475, 515} <i>Lendenfeldia</i> sp. ⁵¹⁶ <i>Phyllospongia chondrodes</i> ⁵¹³
TeC-88	12 α -hydroxy-24-methyl-24,25-dioxoscalar-16-en-22-oic acid		<i>Dictyoceratida</i> sp. and <i>Halichondria</i> sp. ⁵¹¹
TeC-89	12 α -acetoxy-20,24-dimethyl-25-norscalar-16-en-24-one		<i>Carteriospongia foliascens</i> ⁴⁹⁶ <i>Phyllospongia lamellosa</i> ⁵¹⁷
TeC-90	unnamed tetracyclic sesterterpene (reported as compound 6)		<i>Phyllospongia</i> sp. ⁴⁸⁷

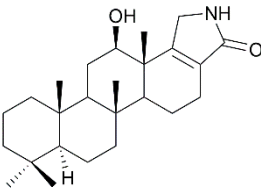
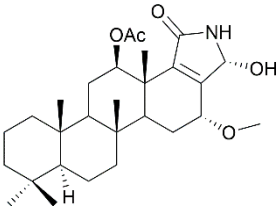
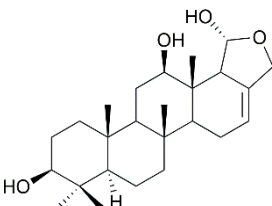
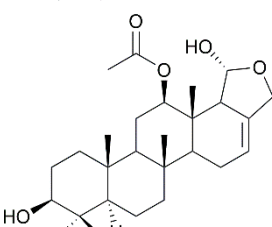
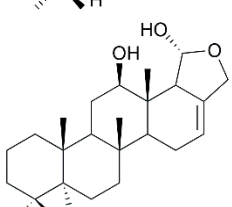
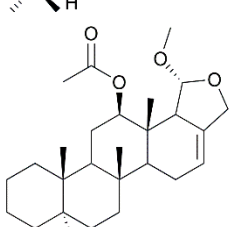
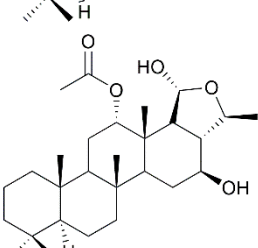
TeC-91	unnamed tetracarboxylic sesterterpene (reported as compound 7)		<i>Phyllospongia</i> sp. ⁴⁸⁷
TeC-92	25-nor-24-methyl-12,24-dioxoscalar-16-en-22-oic acid		<i>Dictyoceratida</i> sp. and <i>Halichondria</i> sp. ⁵¹¹
TeC-93	phyllphenone E		<i>Phyllospongia foliascens</i> ⁵¹⁸
TeC-94	12 α -acetoxy-20,24-dimethyl-24-oxoscalar-16-en-25 β -oic acid		<i>Phyllospongia papyracea</i> ⁴⁸⁹
TeC-95	22-hydroxy-24-methyl-12,24-dioxoscalar-16-en-25-al or PCH-3		<i>Lendenfeldia</i> sp. ⁵¹⁹ <i>Phyllospongia chondrodes</i> ⁵¹³
TeC-96	25-nor-12 α -acetoxy-20,24-dimethyl-24-oxoscalar-16-en-18 β -ol		<i>Phyllospongia papyracea</i> ⁴⁸⁹
TeC-97	asperterpenol A		<i>Aspergillus</i> sp. 085242 ⁵²⁰
TeC-98	asperterpenol B		<i>Aspergillus</i> sp. 085242 ⁵²⁰

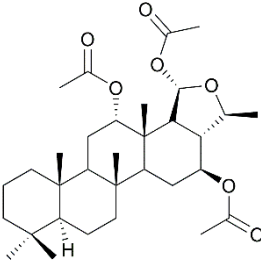
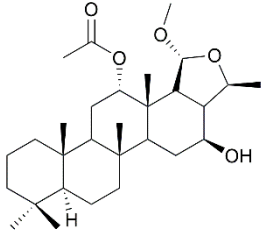
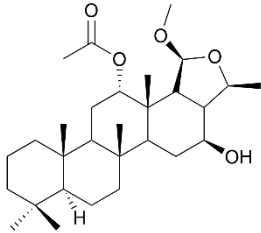
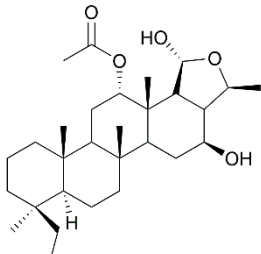
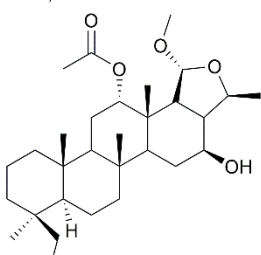
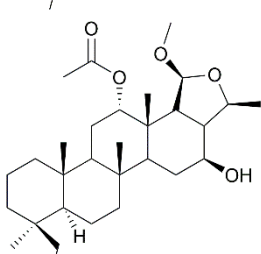
TeC-99	24-methyl-24,25-dioxoscalar-16-en-12 β -yl-3-hydroxybutanoate		<i>Phyllospongia dendyi</i> ⁵²¹
TeC-100	felixin F		<i>Ircinia felix</i> ⁵²² <i>Lendenfeldia</i> sp. ⁴⁷⁵
TeC-101	felixin G		<i>Ircinia felix</i> ⁵²²
TeC-102	12 α -acetoxy-20,24-dimethyl-16,24-dioxoscalar-14,17-dien-25-oic acid		<i>Carteriospongia foliascens</i> ⁴⁹⁵
TeC-103	24-methyl-12,24,25-trioxoscalar-16-en-22-oate		<i>Lendenfeldia</i> sp. ⁵¹⁹
TeC-104	12 α -acetoxy-16 β -hydroxy-24-methyl-24-oxoscalarane-25-al		<i>Lendenfeldia</i> sp. ⁸⁴
TeC-105	12 β -(3'β-hydroxybutanoyloxy)-20,24-dimethyl-24-oxoscalara-16-en-25-al		<i>Carteriospongia</i> sp. ⁴⁹¹

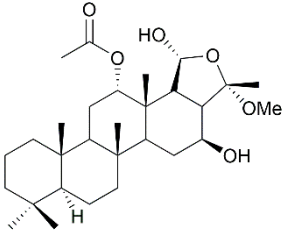
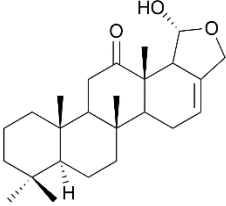
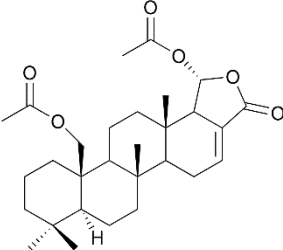
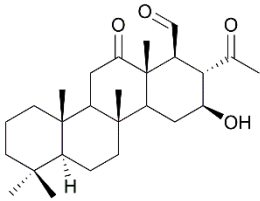
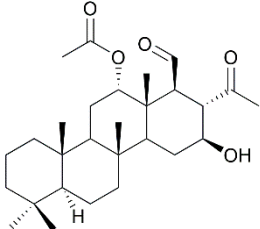
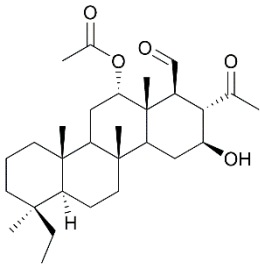
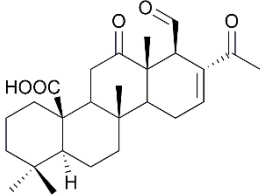
TeC-106	12- <i>O</i> -deacetyl-12- <i>epi</i> -19- <i>O</i> -methylsclarin		<i>Spongia</i> sp. ^{490, 523}
TeC-107	12- <i>O</i> -deacetyl-12,19-di- <i>epi</i> -sclarin		<i>Hyrtios erectus</i> ⁵²⁴
TeC-108	scalarinol		<i>Hyrtios erecta</i> [sic!] ⁵²⁵
TeC-109	12,24-diacetoxy-deoxosclarin		<i>Spongia</i> sp. ⁵²⁶
TeC-110	12- <i>O</i> -deacetoxy-24-hydroxyl-deoxosclarin		<i>Spongia</i> sp. ⁵²⁶
TeC-111	12- <i>O</i> -deacetoxy-19- <i>O</i> -methydeoxosclarin		<i>Hyattella</i> sp. ⁵²⁷ <i>Spongia</i> sp. ⁵²⁶ <i>Glossodoris hikuerensis</i> ⁵²⁸
TeC-112	12 α - <i>O</i> -acetylhyrtiolide		<i>Hyrtios erectus</i> ³⁴

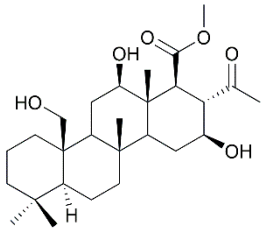
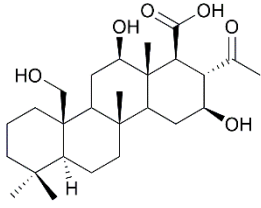
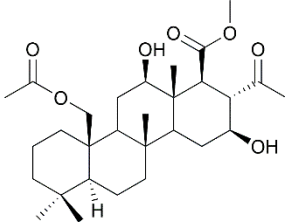
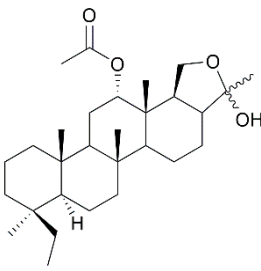
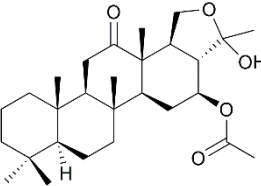
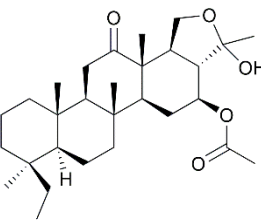
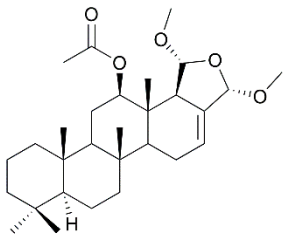
TeC-113	12 α -acetoxy-20,24 β -dimethylsclar-17-eno-25,24-lactone		<i>Phyllospongia papyracea</i> ⁴⁸⁹
TeC-114	sesterstatin 7		<i>Hyrtios erecta</i> [sic!] ⁵²⁹ <i>Hyrtios gumminae</i> ⁵³⁰
TeC-115	unnamed tetracycyclic sesterterpene (reported as compound 1)		<i>Pleurotus ostreatus</i> ⁵³¹
TeC-116	12- <i>O</i> -deacetyl-12- <i>epi</i> -19-deoxy-21-hydroxysclarin		<i>Hyattella</i> sp. ⁵²⁷ <i>Spongia</i> sp. ⁵²³
TeC-117	unnamed tetracycyclic sesterterpene (reported as compound 2)		<i>Pleurotus ostreatus</i> ⁵³¹
TeC-118	12 β ,16 β ,22-trihydroxy-24-methyl-24-oxosclaran-25-oxo methyl ester		<i>Lendenfeldia</i> sp. ⁸⁴
TeC-119	12 α -acetoxy-20,24-dimethyl-16,24-dioxosclara-14,17-dien-24-ol-25,24-olide		<i>Phyllospongia papyracea</i> ⁴⁸⁹

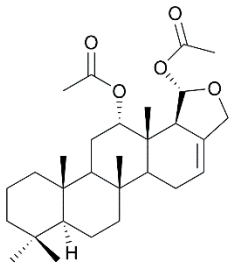
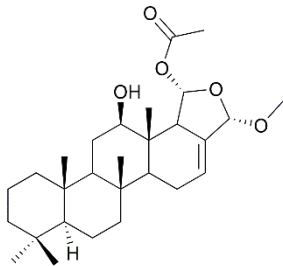
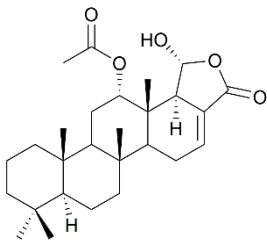
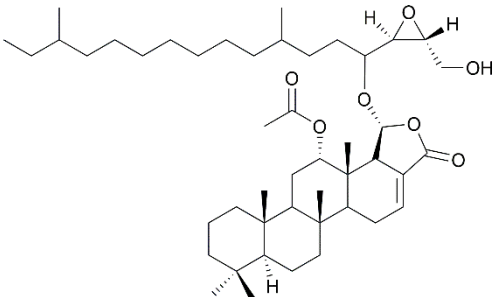
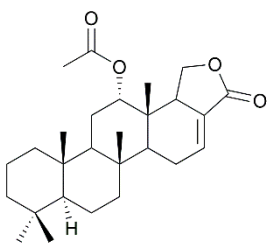
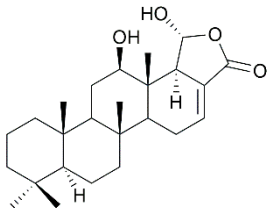
TeC-120	scalarester		<i>Dysidea</i> sp. ⁵⁰³
TeC-121	12- <i>epi</i> -19-deoxyscalarin		<i>Hyrtios erectus</i> ⁵²⁴ <i>Brachiaster</i> sp. ¹¹¹
TeC-122	unnamed tetracycyclic sesterterpene (reported as compound 7)		<i>Hyattella</i> sp. ⁵²⁷
TeC-123	salmahyrtisol C		<i>Hyrtios erecta</i> [sic!] ⁵³²
TeC-124	unnamed tetracycyclic sesterterpene (reported as compound 8)		<i>Hyattella</i> sp. ⁵²⁷ <i>Hyrtios erecta</i> [sic!] ⁵³³
TeC-125	24-acetoxy-12-deacetyl-12- <i>epi</i> -deoxyscalarin		<i>Hyattella intestinalis</i> ⁵³⁴
TeC-126	petrosapsongiolactam A		<i>Petrosaspongia</i> sp. ⁵³⁵

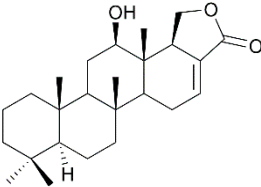
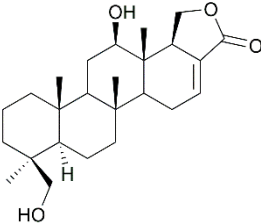
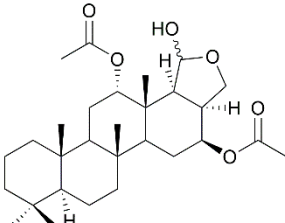
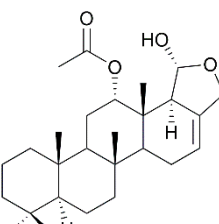
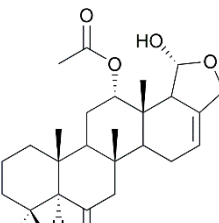
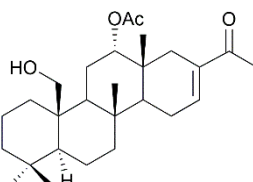
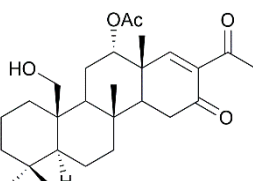
TeC-127	petrosapsongiolactam B		<i>Petrosaspongia</i> sp. ⁵³⁵
TeC-128	petrosapsongiolactam C		<i>Petrosaspongia</i> sp. ⁵³⁵
TeC-129	unnamed tetracycyclic sesterterpene (reported as compound 2a and as compound 6)		<i>Hyrtios erecta</i> [sic!] ⁵³³ <i>Spongia</i> sp. ⁵³⁶
TeC-130	unnamed tetracycyclic sesterterpene (reported as compound 2b)		<i>Hyrtios erecta</i> [sic!] ⁵³³
TeC-131	unnamed tetracycyclic sesterterpene (reported as compound 3)		<i>Hyrtios erecta</i> [sic!] ⁵³³
TeC-132	unnamed tetracycyclic sesterterpene (reported as compound 9)		<i>Hyattella</i> sp. ⁵²⁷
TeC-133	unnamed tetracycyclic sesterterpene (reported as compound 3)		<i>Phyllospongia</i> sp. ⁴⁸⁷

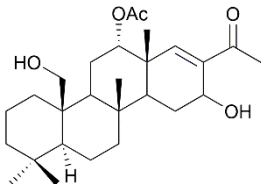
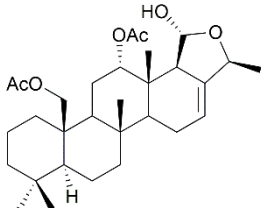
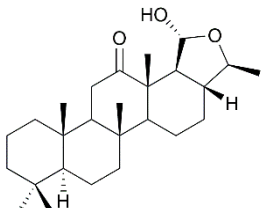
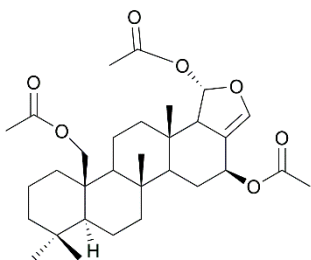
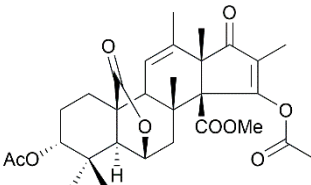
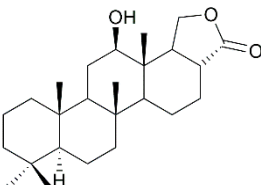
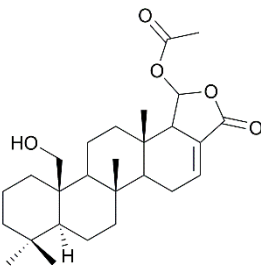
TeC-134	unnamed tetracarbocyclic sesterterpene (reported as compound 3a)		<i>Phyllospongia</i> sp. ⁴⁸⁷
TeC-135	unnamed tetracarbocyclic sesterterpene (reported as compound 3b)		<i>Phyllospongia</i> sp. ⁴⁸⁷
TeC-136	unnamed tetracarbocyclic sesterterpene (reported as compound 3c)		<i>Phyllospongia</i> sp. ⁴⁸⁷
TeC-137	unnamed tetracarbocyclic sesterterpene (reported as compound 4)		<i>Phyllospongia</i> sp. ⁴⁸⁷
TeC-138	unnamed tetracarbocyclic sesterterpene (reported as compound 4b)		<i>Phyllospongia</i> sp. ⁴⁸⁷
TeC-139	unnamed tetracarbocyclic sesterterpene (reported as compound 4c)		<i>Phyllospongia</i> sp. ⁴⁸⁷

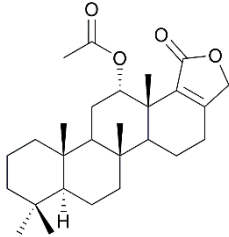
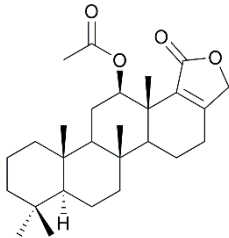
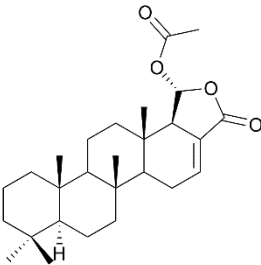
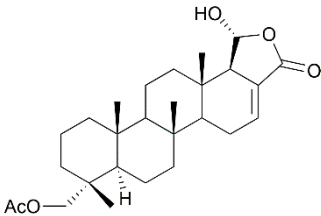
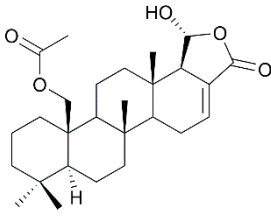
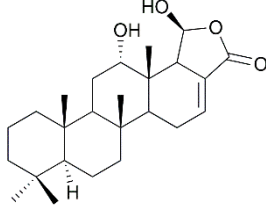
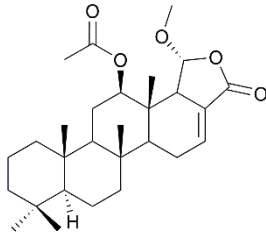
TeC-140	unnamed tetracarboxylic sesterterpene (reported as compound 5)		<i>Phyllospongia</i> sp. ⁴⁸⁷
TeC-141	12-deacetoxy-12-oxo-deoxoscalarin		<i>Glossodoris rufomarginata</i> ⁴⁷⁸ <i>Dorisprismatica</i> (= <i>Glossodoris</i>) <i>atromarginata</i> ⁴⁹⁴ <i>Glossodoris atromarginata</i> ⁵³⁷ <i>Glossodoris pallida</i> ⁴⁷⁹ <i>Glossodoris hikuerensis</i> ⁵⁰⁷ <i>Glossodoris vespa</i> ⁵⁰⁷
TeC-142	12-deacetoxy-23-acetoxy-19-O-acetylscalarin		<i>Smenospongia</i> sp. ³² <i>Psammocinia</i> sp. ⁴⁸⁴ <i>Smenospongia</i> sp. ³³
TeC-143	unnamed tetracarboxylic sesterterpene (reported as compound 2)		<i>Lendenfeldia</i> sp. ⁸⁴
TeC-144	unnamed tetracarboxylic sesterterpene (reported as compound 3)		<i>Lendenfeldia</i> sp. ⁸⁴
TeC-145	unnamed tetracarboxylic sesterterpene (reported as compound 4)		<i>Lendenfeldia</i> sp. ⁸⁴
TeC-146	unnamed tetracarboxylic sesterterpene (reported as compound 5)		<i>Lendenfeldia</i> sp. ⁸⁴

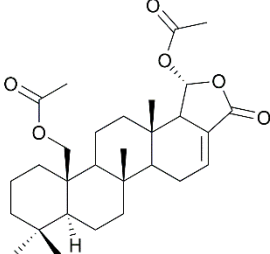
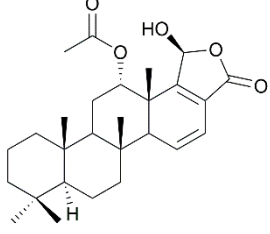
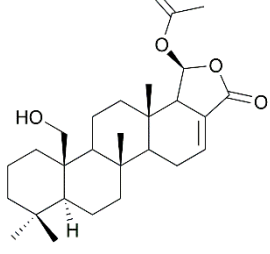
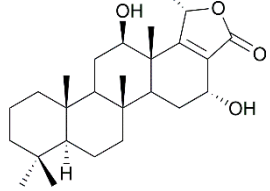
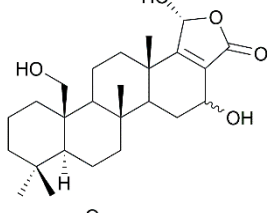
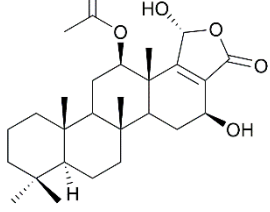
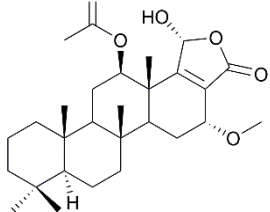
TeC-147	unnamed tetracyclic sesterterpene (reported as compound 6)		<i>Lendenfeldia</i> sp. ⁸⁴
TeC-148	unnamed tetracyclic sesterterpene (reported as compound 7)		<i>Lendenfeldia</i> sp. ⁸⁴
TeC-149	unnamed tetracyclic sesterterpene (reported as compound 8)		<i>Lendenfeldia</i> sp. ⁸⁴
TeC-150	12 α -acetoxy-24,25-epoxy-24-hydroxy-20,24-dimethylsclarane		<i>Carteriospongia foliascens</i> ⁴⁹⁵ <i>Phyllospongia lamellosa</i> ⁵¹⁷
TeC-151	scaldardysin-A		<i>Dysidea herbacea</i> ⁵⁰² <i>Phyllospongia lamellosa</i> ⁵¹⁷
TeC-152	scaldardysin-B		<i>Dysidea herbacea</i> ⁵⁰²
TeC-153	12-acetyl-19,20-dimethoxy-deoxoscalarin		<i>Collospongia auris</i> ⁴⁸²

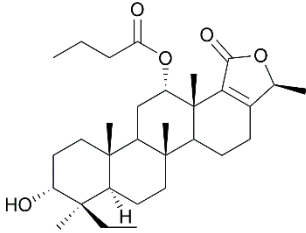
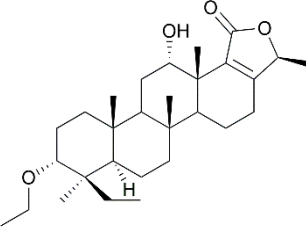
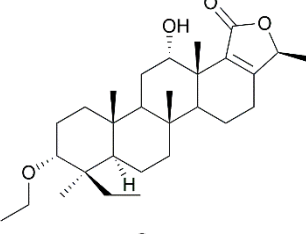
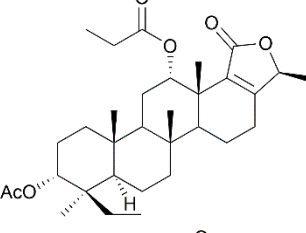
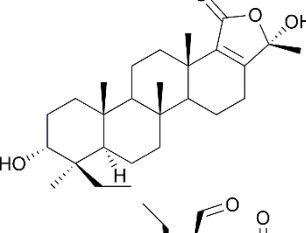
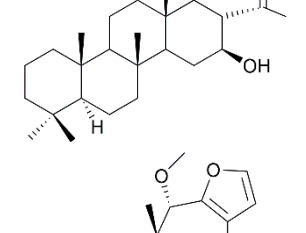
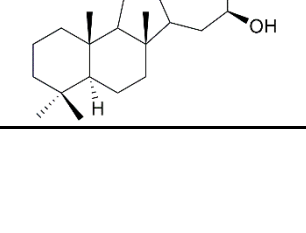
TeC-154	deoxo-scalarin acetate		<i>Spongia officinalis</i> ⁵³⁸
TeC-155	12- <i>epi</i> -deacetyl-19 α -acetoxymethoxyscalaran		<i>Hyrtios</i> sp. ⁵³⁹
TeC-156	scalarin		<i>Cacospongia scalaris</i> ⁵⁴⁰ <i>Dysidea</i> sp. ⁵⁰³ <i>Ircinia</i> sp. ⁵⁴¹ <i>Hyattella intestinalis</i> ³⁹⁰ <i>Spongia matamata</i> ³⁷¹ <i>Spongia tubulifera</i> ³⁷⁹ <i>Spongia virgultosa</i> ⁵⁴² <i>Hyrtios erectus</i> ⁵⁴³ <i>Scalarispongia</i> sp. ⁵⁴⁴ <i>Hyrtios erectus</i> ⁵⁴³
TeC-157	scalarinether		
TeC-158	19-deoxyscalarin		<i>Cacospongia scalaris</i> ⁵⁰¹ <i>Glossodoris pallida</i> ⁵⁰⁶
TeC-159	12- <i>O</i> -deacetyl-12- <i>epi</i> -scalarin		<i>Hyrtios erectus</i> ⁵⁴⁵ <i>Spongia</i> sp. ^{526, 536, 546}

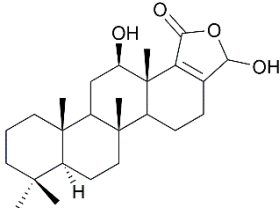
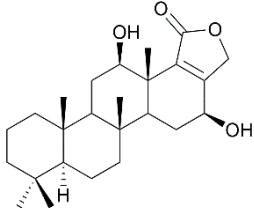
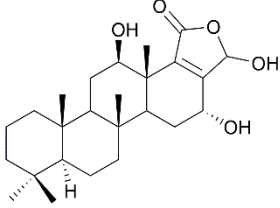
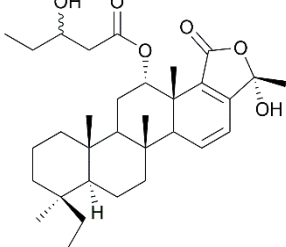
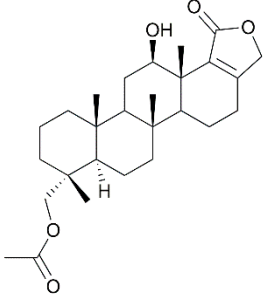
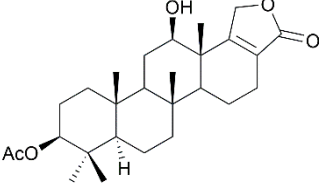
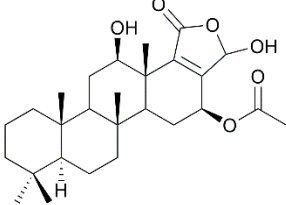
TeC-160	12-deacetyl-12- <i>epi</i> -19-deoxyscalarin		<i>Brachiaster</i> sp. ¹¹¹ <i>Glossodoris dalli</i> ⁵¹⁰ <i>Hyattella cribriformis</i> ⁵⁴⁷ <i>Hyrtios erecta</i> [sic!] ⁵⁴⁸ <i>Spongia</i> sp. ^{526, 536, 546}
TeC-161	12- <i>O</i> -deacetyl-12- <i>epi</i> -19-deoxy-22-hydroxyscalarin		<i>Spongia</i> sp. ⁵²³
TeC-162	16-acetoxy-dihydrodeoxoscalarin		<i>Cacospongia scalaris</i> ⁸⁵
TeC-163	deoxoscalarin		<i>Cacospongia mollior</i> (association with <i>Hypselodoris orsini</i> [sic!]) ⁵⁴⁹ <i>Cacospongia scalaris</i> ⁸⁵ <i>Glossodoris dalli</i> ⁵¹⁰ <i>Glossodoris tricolor</i> ⁵⁰⁴ <i>Glossodoris rufomarginata</i> ⁴⁷⁸ <i>Hyattella intestinalis</i> ³⁹⁰ <i>Spongia officinalis</i> ⁵⁵⁰ <i>Glossodoris pallida</i> ⁴⁷⁹ <i>Glossodoris averni</i> ⁴⁷⁹ <i>Cacospongia mollior</i> (association with <i>Hypselodoris orsini</i> [sic!]) ⁵⁴⁹ <i>Chromodoris funerea</i> ¹⁴⁴
TeC-164	6-keto-deoxoscalarin		<i>Cacospongia mollior</i> (association with <i>Hypselodoris orsini</i> [sic!]) ⁵⁴⁹ <i>Chromodoris funerea</i> ¹⁴⁴
TeC-165	felixin A		<i>Ircinia felix</i> ⁵²²
TeC-166	felixin B		<i>Ircinia felix</i> ⁵²²

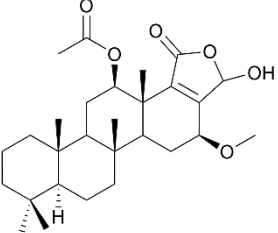
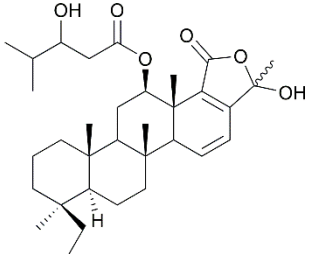
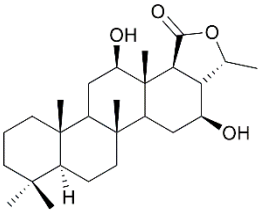
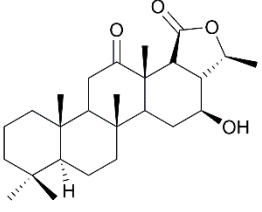
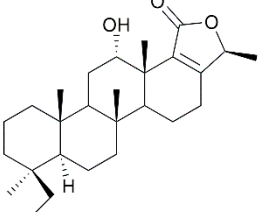
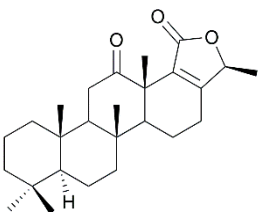
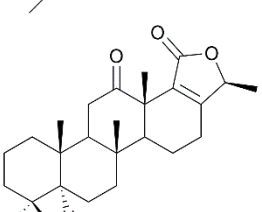
TeC-167	felixin C		<i>Ircinia felix</i> ⁵²²
TeC-168	felixin D		<i>Ircinia felix</i> ⁵²²
TeC-169	felixin E		<i>Ircinia felix</i> ⁵²²
TeC-170	12-deacetoxy-23-O-acetoxysteronemin		<i>Psammocinia</i> sp. ⁴⁸⁴
TeC-171	citreoahydnone A		Hybrid strain KO 0031 derived from <i>Penicillium citreo-viride</i> B. IFO 6200 and 4692 ⁵⁵¹
TeC-172	hippospongide B		<i>Hippospongia</i> sp. ³⁶⁵
TeC-173	scalarolide		<i>Cacospongia</i> ⁵⁴¹ <i>Hyrtios erectus</i> ⁴⁸³ <i>Spongia</i> sp. ⁴⁹⁰ <i>Spongia idia</i> ²⁶ <i>Hyrtios gumminae</i> ⁵³⁰ <i>Hyattella cribriformis</i> ⁵⁴⁷

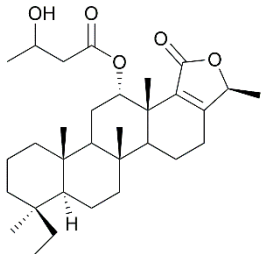
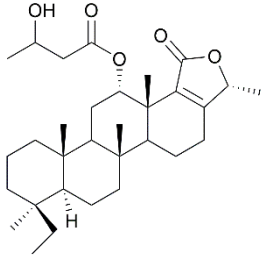
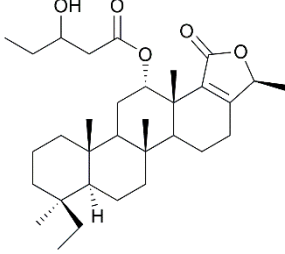
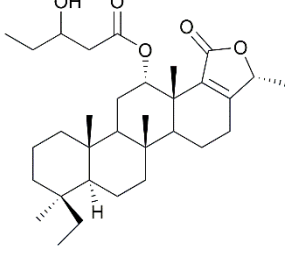
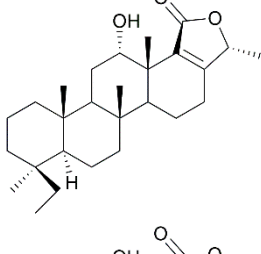
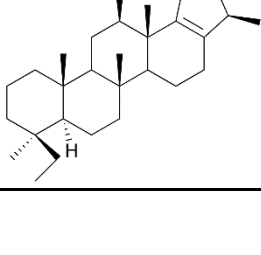
TeC-174	12- <i>epi</i> -12- <i>O</i> -acetylscalarolide		<i>Cacospongia scalaris</i> ⁵⁰¹ <i>Hyattella intestinalis</i> ³⁹⁰ <i>Scalarispongia</i> sp. ⁵⁴⁴
TeC-175	scalarolide acetate		<i>Collospongia auris</i> ⁴⁸² <i>Hyrtios erectus</i> ⁴⁸⁰ <i>Spongia</i> sp. ^{490, 536}
TeC-176	12-deacetoxy-21-acetoxyscalarin 19-acetate		<i>Hippospongia</i> sp. ³⁶⁵ <i>Brachiaster</i> sp. ¹¹¹
TeC-177	12-deacetoxy-21-acetoxyscalarin		<i>Hyrtios</i> cf. <i>erectus</i> ⁵⁵²
TeC-178	12-deacetoxy-22-acetoxyscalarin		<i>Smenospongia</i> sp. ³²
TeC-179	12- <i>O</i> -deacetyl-25- <i>epi</i> -scalarin		<i>Hyattella intestinalis</i> ³⁹⁰
TeC-180	12- <i>epi</i> -19- <i>O</i> -methylscalarin		<i>Hyattella</i> sp. ⁵²⁷

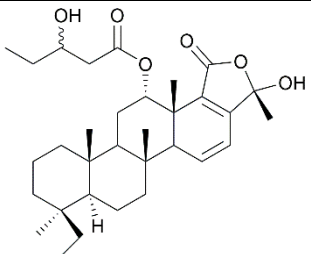
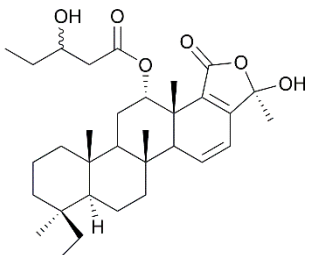
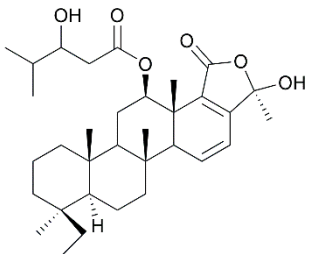
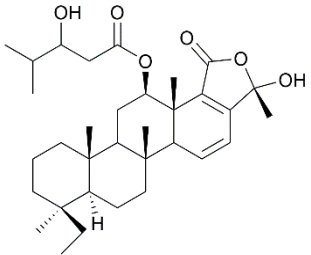
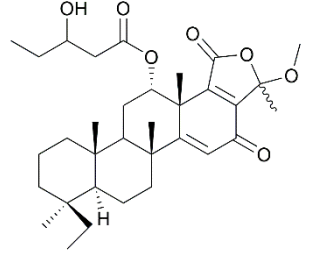
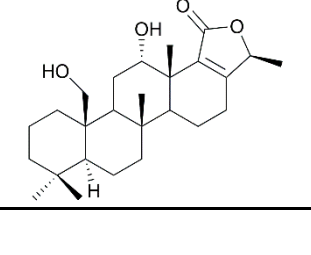
TeC-181	12-deacetoxy-22,25-diacetoxy-scalarin		<i>Smenospongia</i> sp. ³²
TeC-182	12 α -acetoxy-19 β -hydroxyscalara-15,17-dien-20,19-olide		<i>Spongia matamata</i> ³⁷¹ <i>Hyattella intestinalis</i> ³⁹⁰
TeC-183	12-deacetoxy-22-hydroxy-25-acetoxyscalarin		<i>Smenospongia</i> sp. ³³
TeC-184	hyrtiolide		<i>Hyrtios erectus</i> ⁴⁸³ <i>Hyrtios gumminae</i> ⁵³⁰
TeC-185	12-dehydroxy-23-hydroxyhyrtiolide		<i>Psammocinia</i> sp. ⁴⁸⁴
TeC-186	12-acetoxy,16- <i>epi</i> -hyrtiolide		<i>Hyrtios erectus</i> ^{480, 524}
TeC-187	12- <i>O</i> -acetyl-16- <i>O</i> -methylhyrtiolide		<i>Spongia</i> sp. ⁵⁴⁶ <i>Hyattella</i> sp. ⁵²⁷ <i>Hyrtios erectus</i> ⁵²⁴

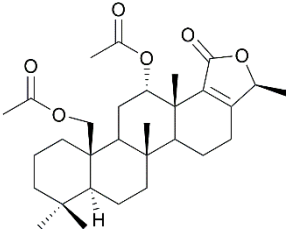
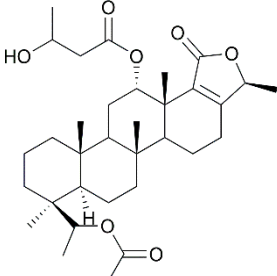
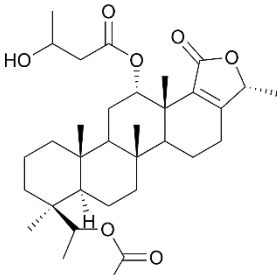
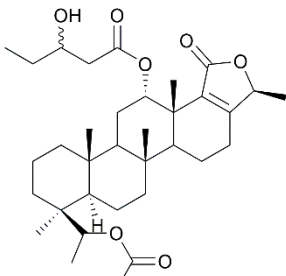
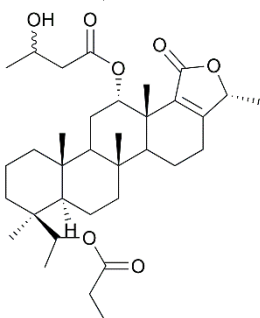
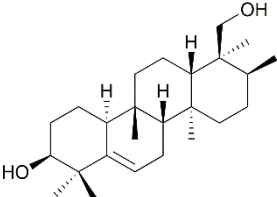
TeC-188	phyllolactone A		<i>Phyllospongia lamellosa</i> ⁵⁵³
TeC-189	phyllolactone B		<i>Phyllospongia lamellosa</i> ^{553, 554}
TeC-190	phyllolactone C		<i>Phyllospongia lamellosa</i> ^{553, 554}
TeC-191	phyllolactone D		<i>Phyllospongia lamellosa</i> ⁵⁵³
TeC-192	phyllolactone E		<i>Phyllospongia lamellosa</i> ⁵⁵³
TeC-193	unnamed tetracycyclic sesterterpene (reported as compound 2)		<i>Cacospongia scalaris</i> ⁵⁵⁵
TeC-194	similan A		<i>Hyrtios gumminae</i> ⁵³⁰

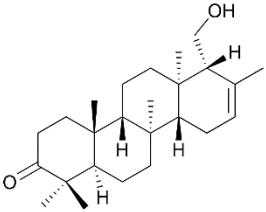
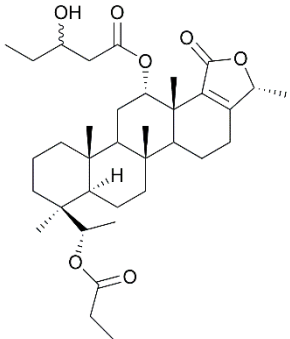
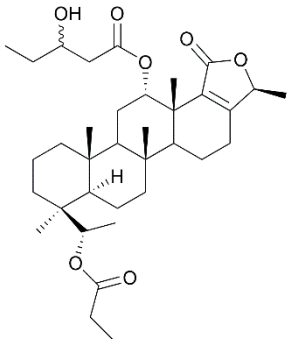
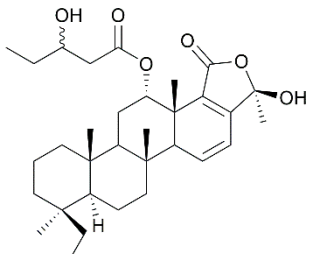
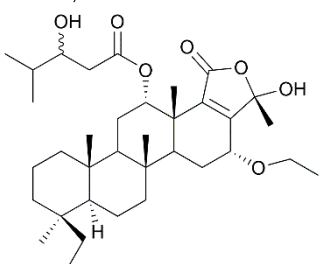
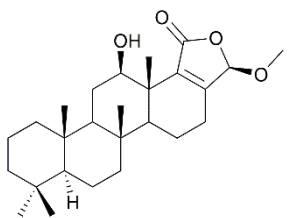
TeC-195	20-hydroxyscalarolide		<i>Collospongia auris</i> ⁴⁸²
TeC-196	16-hydroxyscalarolide		<i>Hyrtios erectus</i> ⁴⁸³
TeC-197	12 β 16 α ,20-trihydroxy-17-scalar-19,20-olide		<i>Hyrtios gumminae</i> ⁵³⁰
TeC-198	(12 α ,24S)-12-[(3-hydroxypentanoyl)oxy]-20,24-dimethyl-25-oxoscalar-15,17-dien-25,24-olide		<i>Phyllospongia papyracea</i> ⁴⁹⁷ <i>Carteriospongia foliascens</i> ⁴⁹²
TeC-199	19-acetylsesterstatin 3		<i>Hyrtios erecta</i> [sic!] ⁵³² <i>Hyrtios erectus</i> ⁵²⁴
TeC-200	3-acetylsesterstatin 1		<i>Hyrtios erecta</i> [sic!] ⁵³²
TeC-201	12 β ,20-dihydroxy-16 β -acetoxy-17-scalar-19,20-olide		<i>Hyrtios erectus</i> ⁴⁸⁰

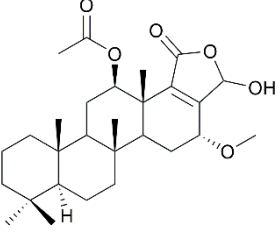
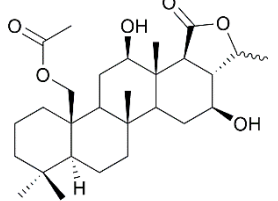
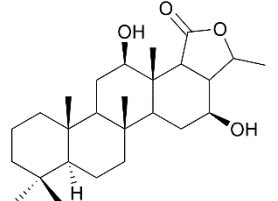
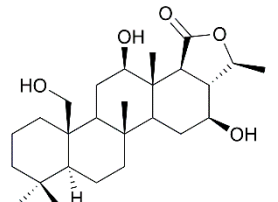
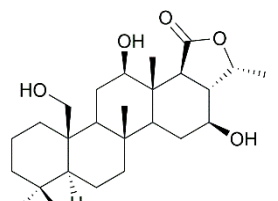
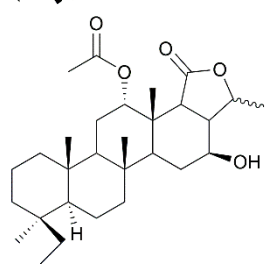
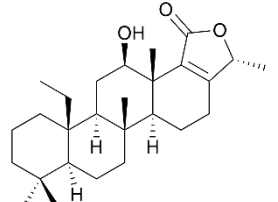
TeC-202	12 β -acetoxy,16 β -methoxy,20 α -hydroxy-17-scalarin-19,20-olide		<i>Hyrtios erectus</i> ⁴⁸⁰
TeC-203	12 α ,24-dihydroxy-20,24-dimethyl-15,17-scalarin-25,24-olide		<i>Phyllospongia papyracea</i> ⁵⁵⁶
TeC-204	12,16-dihydroxy-24-methylscalarin-25,24-olide		<i>Lendenfeldia</i> sp. ⁵¹⁵
TeC-205	phyllofolactone L		<i>Phyllospongia foliascens</i> ⁵¹⁸
TeC-206	phyllofolactone B		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁵⁵⁷ <i>Phyllospongia madagascarensis</i> ³⁹⁶ <i>Phyllospongia foliascens</i> ⁵⁵⁸
TeC-207	phyllofolactone C		<i>Phyllospongia foliascens</i> ⁵⁵⁹
TeC-208	phyllofolactone D		<i>Phyllospongia foliascens</i> ⁵⁵⁹

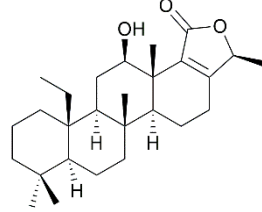
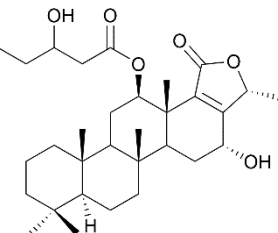
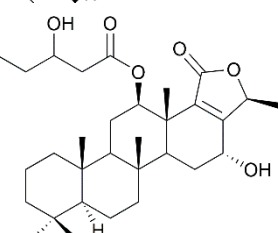
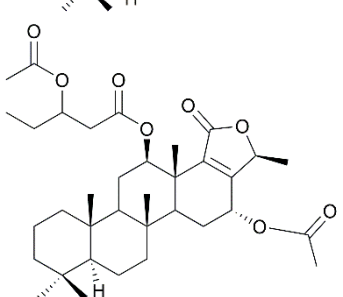
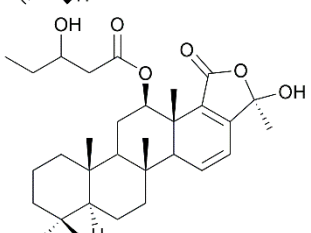
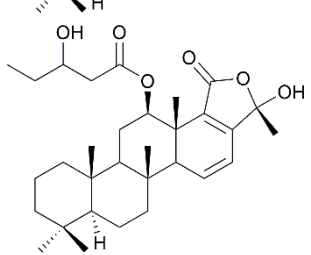
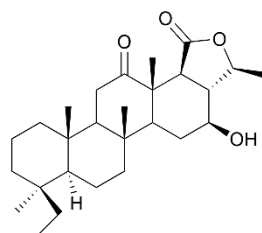
TeC-209	phyllofolactone H		<i>Strepsichordaia aliena</i> ⁵⁶⁰
TeC-210	phyllofolactone I		<i>Strepsichordaia aliena</i> ⁵⁶⁰
TeC-211	phyllofolactone J		<i>Strepsichordaia aliena</i> ⁵⁶⁰
TeC-212	phyllofolactone K		<i>Strepsichordaia aliena</i> ⁵⁶⁰
TeC-213	phyllofolactone M		<i>Phyllospongia foliascens</i> ⁵⁵⁸
TeC-214	12- <i>epi</i> -phyllofolactone-B		<i>Phyllospongia foliascens</i> ⁵⁶¹

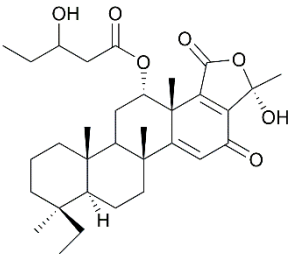
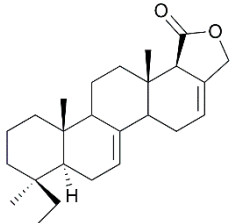
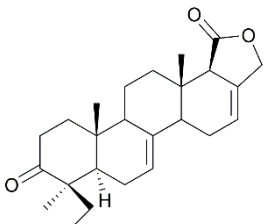
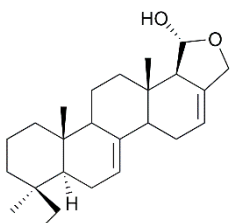
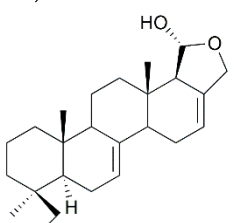
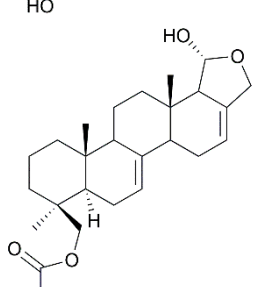
TeC-215	12- <i>epi</i> -phyllactone D		<i>Carteriospongia foliascens</i> ⁴⁹²
TeC-216	12- <i>epi</i> -phyllactone E		<i>Carteriospongia foliascens</i> ⁴⁹²
TeC-217	phyllactone F		<i>Phyllospongia foliascens</i> ⁵⁶² <i>Phyllospongia papyracea</i> ⁵⁵⁶
TeC-218	phyllactone G		<i>Phyllospongia foliascens</i> ⁵⁶² <i>Phyllospongia papyracea</i> ⁵⁵⁶
TeC-219	phyllactone H		<i>Phyllospongia papyracea</i> ⁵⁵⁶
TeC-220	lendenfeldarane C		<i>Lendenfeldia</i> sp. ⁴⁷⁵

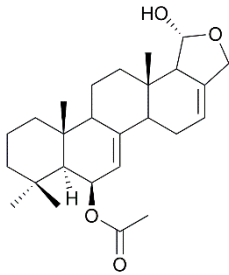
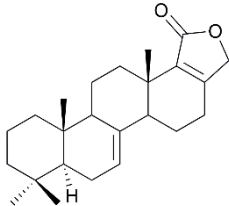
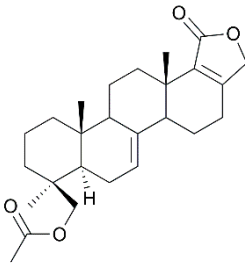
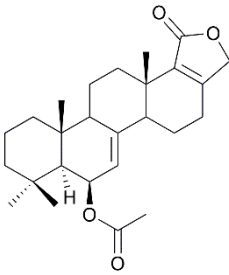
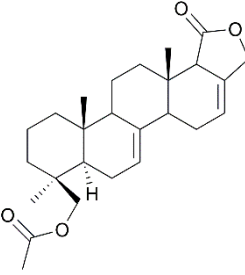
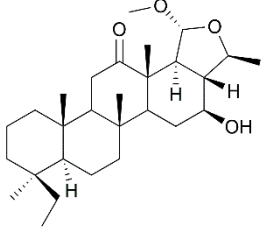
TeC-221	lendenfeldarane D		<i>Lendenfeldia</i> sp. ⁴⁷⁵
TeC-222	honulactone C		<i>Strepsichordaia aliena</i> ⁵⁶⁰
TeC-223	honulactone D		<i>Strepsichordaia aliena</i> ⁵⁶⁰
TeC-224	honulactone I		<i>Strepsichordaia aliena</i> ⁵⁶⁰
TeC-225	honulactone L		<i>Strepsichordaia aliena</i> ⁵⁶⁰
TeC-226	unnamed tetracycyclic sesterterpene (reported as compound 5)		<i>Aletris farinosa</i> ⁴³¹

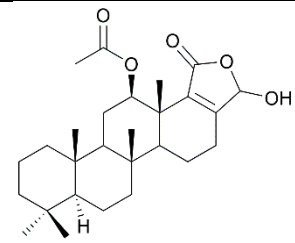
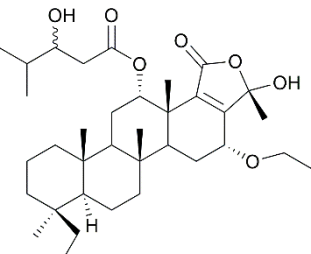
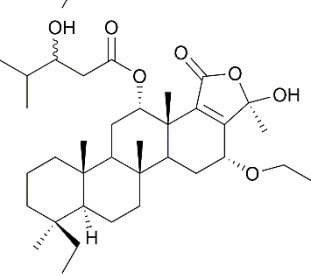
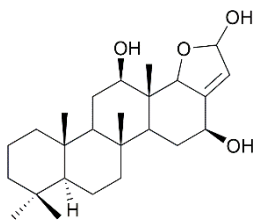
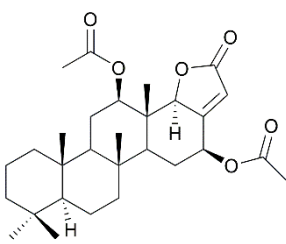
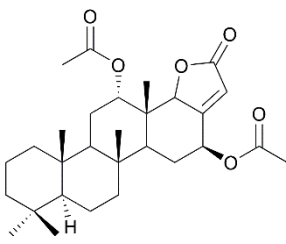
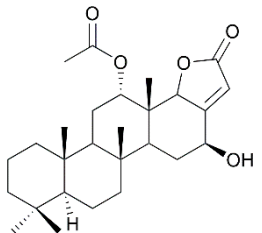
TeC-227	unnamed tetracarbocyclic sesterterpene (reported as compound 6)		<i>Aletris farinosa</i> ⁴³¹
TeC-228	unnamed tetracarbocyclic sesterterpene (reported as compound 13)		<i>Dysidea granulosa</i> ³⁷⁰
TeC-229	unnamed tetracarbocyclic sesterterpene (reported as compound 14)		<i>Dysidea granulosa</i> ³⁷⁰
TeC-230	(12 α ,24 <i>R</i>)-12-[(3-hydroxypentanoyl) oxy]-20,24-dimethyl-25-oxoscalar-15,17-dien-25,24-olide		<i>Phyllospongia papyracea</i> ⁴⁹⁷ <i>Carteriospongia foliascens</i> ⁴⁹²
TeC-231	unnamed tetracarbocyclic sesterterpene (reported as compound 3)		<i>Carteriospongia foliascens</i> ⁴⁹²
TeC-232	24 β -methoxyscalarolide		<i>Hyattella cribriformis</i> ⁵⁴⁷

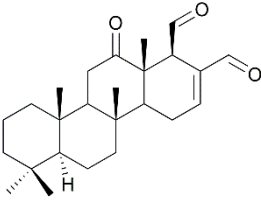
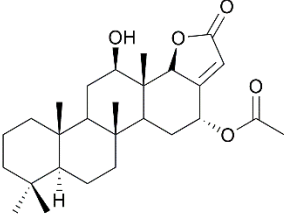
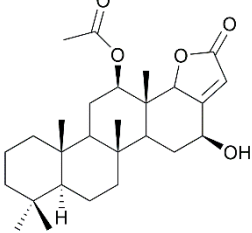
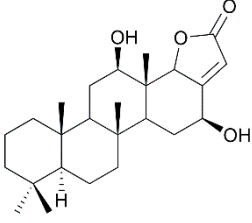
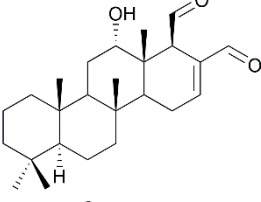
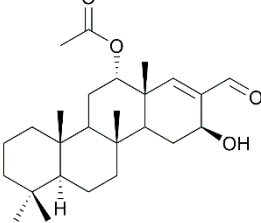
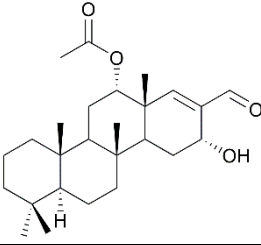
TeC-233	unnamed tetracarboxylic sesterterpene (reported as compound 10)		<i>Hyattella</i> sp. ⁵²⁷
TeC-234	22-acetoxy-12 β ,16 β -dihydroxy-24-methylscalaran-25,24-olide (reported also as homoscalaralactone IIB ⁵⁶³)		<i>Lendenfeldia</i> sp. ⁵¹⁶ <i>Lendenfeldia frondosa</i> ⁵⁶³
TeC-235	12 β ,16 β -dihydroxy-24-methylscalaran-25,24-olide		<i>Phyllospongia dendyi</i> ⁵⁶⁴
TeC-236	12 β ,16 β ,22-trihydroxy-24-methylscalaran-25 β ,24 β -olide		<i>Phyllospongia dendyi</i> ⁵⁶⁴ <i>Lendenfeldia frondosa</i> ⁵⁶⁵
TeC-237	12 β ,22-dihydroxy-24 α -methylscalar-17-en-24 α ,25 β -olide		<i>Lendenfeldia frondosa</i> ⁵⁶⁵
TeC-238	12 α -acetoxy-23,25-cyclo-16 β ,25Z-dihydroxy-20,24-dimethyl-24-oxoscalarane		<i>Carteriospongia foliascens</i> ⁴⁹⁵
TeC-239	hyattellactone A		<i>Hyattella</i> sp. ⁵⁶⁶

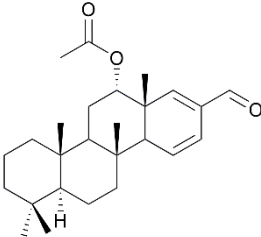
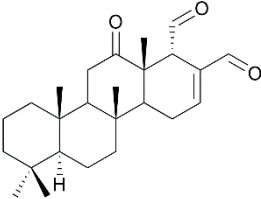
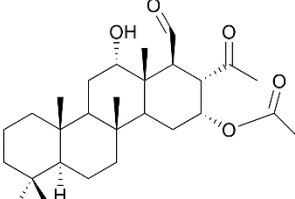
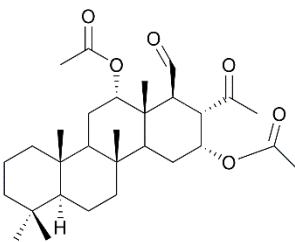
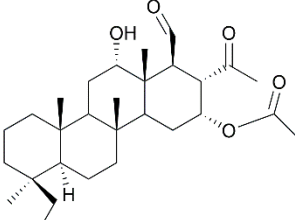
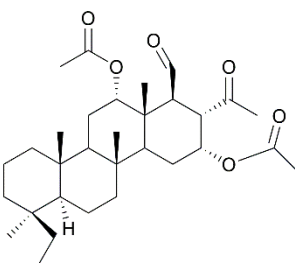
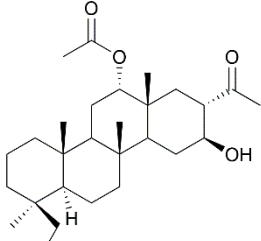
TeC-240	hyattellactone B		<i>Hyattella</i> sp. ⁵⁶⁶
TeC-241	phyllactone A		<i>Phyllospongia</i> (syn. <i>Carteriospongia</i>) <i>foliascens</i> ⁵⁶⁷ <i>Phyllospongia papyracea</i> ⁵⁵⁶
TeC-242	phyllactone B		<i>Phyllospongia</i> (syn. <i>Carteriospongia</i>) <i>foliascens</i> ⁵⁶⁷ <i>Phyllospongia papyracea</i> ⁵⁵⁶
TeC-243	phyllactone C		<i>Phyllospongia</i> (syn. <i>Carteriospongia</i>) <i>foliascens</i> ⁵⁶⁷
TeC-244	phyllactone D		<i>Phyllospongia</i> (syn. <i>Carteriospongia</i>) <i>foliascens</i> ⁵⁶⁷ <i>Phyllospongia papyracea</i> ⁵⁵⁶
TeC-245	phyllactone E		<i>Phyllospongia</i> (syn. <i>Carteriospongia</i>) <i>foliascens</i> ⁵⁶⁷ <i>Phyllospongia papyracea</i> ⁵⁵⁶
TeC-246	phylofolactone A		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁵⁵⁷

TeC-247	carteriofenone I		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁴⁷⁶
TeC-248	mooloolabene F		<i>Dorisprismatica</i> (= <i>Glossodoris</i>) <i>atromarginata</i> ⁴⁹⁴
TeC-249	mooloolabene G		<i>Dorisprismatica</i> (= <i>Glossodoris</i>) <i>atromarginata</i> ⁴⁹⁴
TeC-250	mooloolabene H		<i>Dorisprismatica</i> (= <i>Glossodoris</i>) <i>atromarginata</i> ⁴⁹⁴
TeC-251	mooloolabene I		<i>Dorisprismatica</i> (= <i>Glossodoris</i>) <i>atromarginata</i> ⁴⁹⁴
TeC-252	mooloolabene J		<i>Dorisprismatica</i> (= <i>Glossodoris</i>) <i>atromarginata</i> ⁴⁹⁴

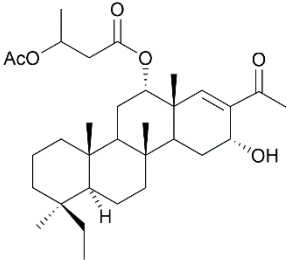
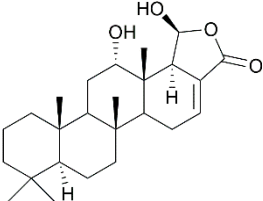
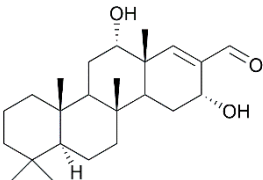
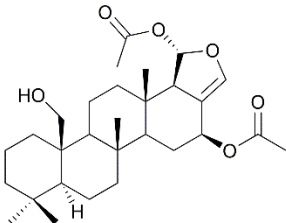
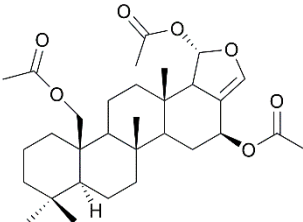
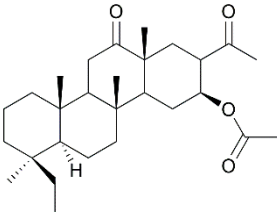
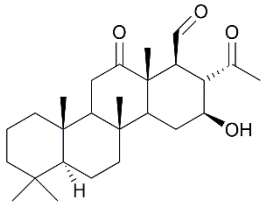
TeC-253	mooloolabene K		<i>Dorisprismatica</i> (= <i>Glossodoris</i>) <i>atromarginata</i> ⁴⁹⁴
TeC-254	mooloolabene L		<i>Dorisprismatica</i> (= <i>Glossodoris</i>) <i>atromarginata</i> ⁴⁹⁴
TeC-255	mooloolabene M		<i>Dorisprismatica</i> (= <i>Glossodoris</i>) <i>atromarginata</i> ⁴⁹⁴
TeC-256	mooloolabene N		<i>Dorisprismatica</i> (= <i>Glossodoris</i>) <i>atromarginata</i> ⁴⁹⁴
TeC-257	mooloolabene O		<i>Dorisprismatica</i> (= <i>Glossodoris</i>) <i>atromarginata</i> ⁴⁹⁴
TeC-258	carteriofenone K		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁴⁷⁶

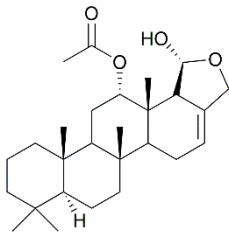
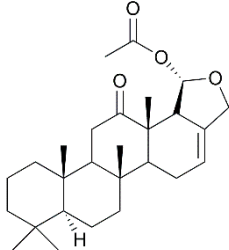
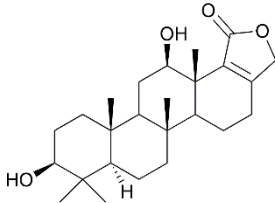
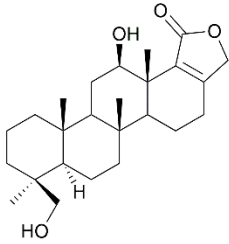
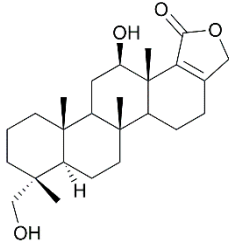
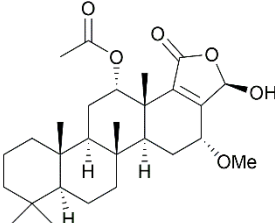
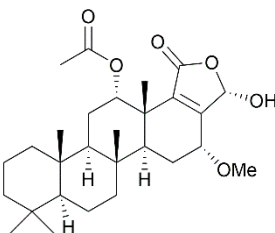
TeC-259	12 β -acetoxy-20-hydroxy-17-scalaren-19,20-olide		<i>Hyrtios gumminae</i> ⁵³⁰
TeC-260	unnamed tetracycyclic sesterterpene (reported as compound 3)		<i>Carteriospongia foliascens</i> ⁴⁹²
TeC-261	unnamed tetracycyclic sesterterpene (reported as compound 4)		<i>Carteriospongia foliascens</i> ⁴⁹²
TeC-262	unnamed tetracycyclic sesterterpene (reported as compound 4)		<i>Hyrtios erecta</i> [sic!] ⁵³³
TeC-263	12 β ,16 β -diacetoxyscalarolbutenolide		<i>Hyrtios erectus</i> ⁵⁴⁵
TeC-264	12 α ,16 β -diacetoxyscalarolbutenolide		<i>Spongia matamata</i> ³⁷¹ <i>Scalarispongia</i> sp. ⁵⁴⁴ <i>Glossodoris pallida</i> ⁴⁷⁹
TeC-265	12 α -acetoxy-16 β -hydroxyscalarolbutenolide		<i>Spongia matamata</i> ³⁷¹ <i>Scalarispongia</i> sp. ⁵⁴⁴

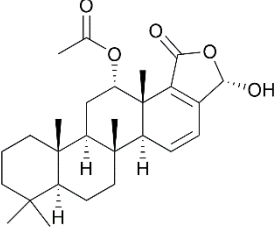
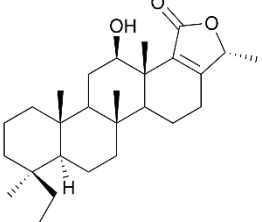
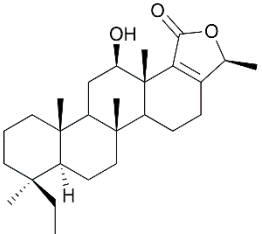
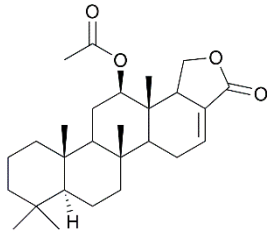
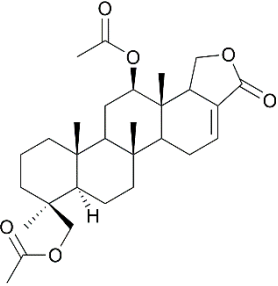
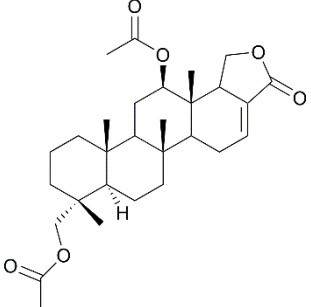
TeC-266	12-deacetoxy-12-oxo-deoxysclarin		<i>Glossodoris</i> sp. ⁴⁷⁹
TeC-267	scalarolbutenolide		<i>Spongia nitens</i> ⁵⁶⁸
TeC-268	12-O-acetyl-16-O-deacetyl-16- <i>epi</i> -scalarolbutenolide		<i>Hyrtios</i> cf. <i>erectus</i> ⁵⁵²
TeC-269	16-O-deacetyl-16- <i>epi</i> -scalarolbutenolide		<i>Hyrtios</i> cf. <i>erectus</i> ⁵⁵²
TeC-270	12-deacetyl-sclaradial		<i>Spongia oceanica</i> ⁴⁸⁶ <i>Glossodoris rufomarginata</i> ⁴⁷⁸
TeC-271	norscalaral A		<i>Cacospongia scalaris</i> ⁵⁰¹ <i>Hyattella intestinalis</i> ³⁹⁰
TeC-272	norscalaral B		<i>Cacospongia scalaris</i> ⁵⁰¹ <i>Hyattella intestinalis</i> ³⁹⁰

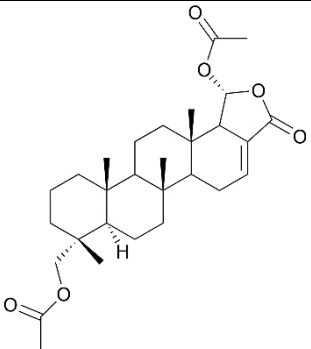
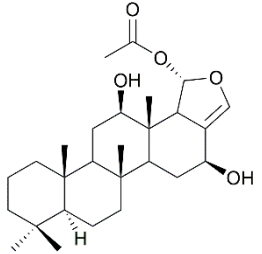
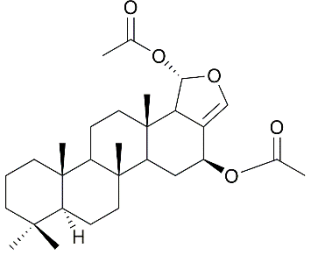
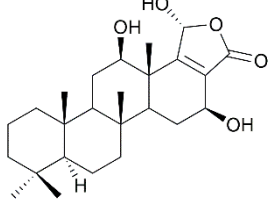
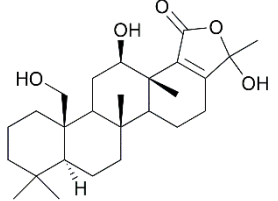
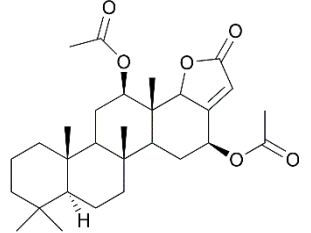
TeC-273	norscalaral C		<i>Cacospongia scalaris</i> ⁵⁰¹
TeC-274	12-deacetyl-18- <i>epi</i> -12-oxo-sclaradial		<i>Spongia oceanica</i> ⁴⁸⁶
TeC-275	scalarherbacin-A		<i>Dysidea herbacea</i> ⁵⁰²
TeC-276	scalarherbacin-A acetate		<i>Dysidea herbacea</i> ⁵⁰²
TeC-277	scalarherbacin-B		<i>Dysidea herbacea</i> ⁵⁰²
TeC-278	scalarherbacin-B acetate		<i>Dysidea herbacea</i> ⁵⁰²
TeC-279	unnamed tetracyclic sesterterpene (reported as compound 8)		<i>Phyllospongia</i> sp. ⁴⁸⁷

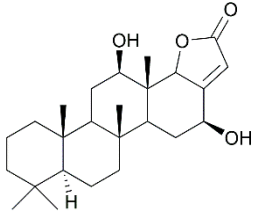
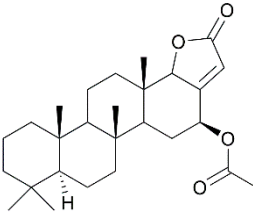
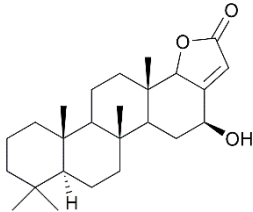
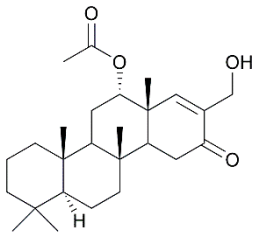
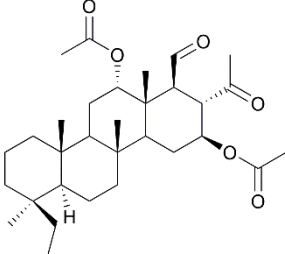
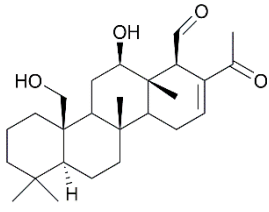
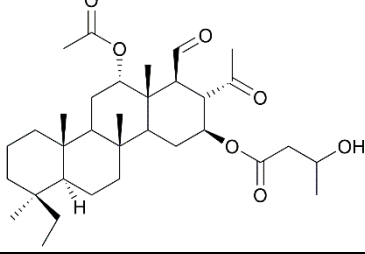
TeC-280	12- <i>epi</i> -deoxoscalarin		<i>Collospongia auris</i> ⁴⁸² <i>Glossodoris dalli</i> ⁵¹⁰ <i>Hyattella</i> sp. ⁵²⁷ <i>Hyattella intestinalis</i> ⁵³⁴ <i>Hyattella cribriformis</i> ⁵⁴⁷ <i>Spongia</i> sp. ⁵⁴⁶ <i>Spongia idia</i> ²⁶ <i>Spongia nitens</i> ⁵⁴² <i>Spongia officinalis</i> ⁵³⁸ <i>Glossodoris hikuerensis</i> ⁵²⁸
TeC-281	12- <i>epi</i> -sclarin		<i>Hyattella</i> sp. ⁵²⁷ <i>Hyrtios erecta</i> [sic!] ⁵⁰⁵ <i>Spongia</i> sp. ⁵⁴⁶ <i>Spongia</i> sp. ⁵²³ <i>Spongia nitens</i> ⁵⁴² <i>Chromodoris funerea</i> ¹³⁰ <i>Spongia</i> sp. ⁵³⁶
TeC-282	12-deacetyl-20-methyl-12- <i>epi</i> -deoxoscalarin		<i>Glossodoris sedna</i> ⁵¹⁰
TeC-283	12-deacetyl-23-acetoxy-20-methyl-12- <i>epi</i> -deoxoscalarin		<i>Glossodoris sedna</i> ⁵¹⁰
TeC-284	12-deacetoxy-23-acetoxysclarin		<i>Smenospongia</i> sp. ^{32, 33}
TeC-285	3-hydroxy-20,22-dimethyldeoxoscalarin		<i>Phyllospongia vermicularis</i> or <i>Dysidea vermicularis</i> ⁵⁶⁹
TeC-286	3-keto-20,22-dimethyl-20-deoxoscalarin		<i>Carteriospongia</i> sp. ⁵⁶⁹

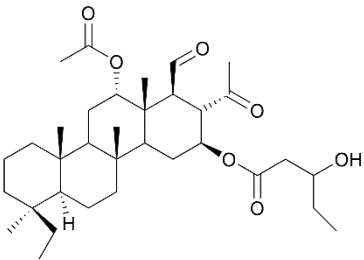
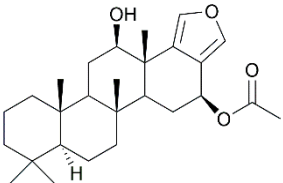
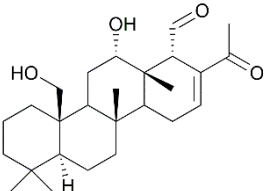
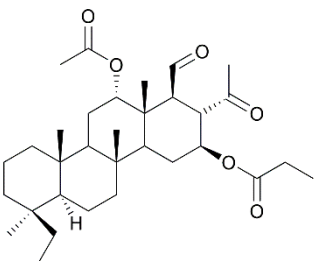
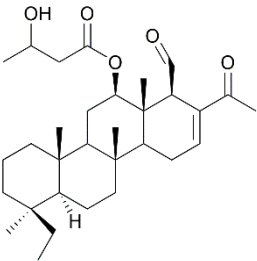
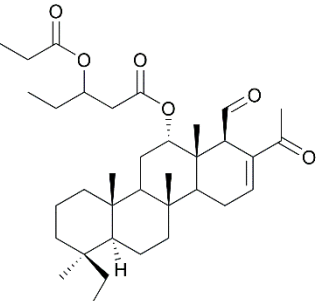
TeC-287	phyllofenone C		<i>Strepsichordaia aliena</i> ⁵⁷⁰
TeC-288	12-O-deacetyl-19- <i>epi</i> -sclarin		<i>Hyattella intestinalis</i> ³⁹⁰
TeC-289	12-O-deacetylnorscalaral B		<i>Hyattella intestinalis</i> ³⁹⁰
TeC-290	12-deacetoxy-23-hydroxyheteronemin		<i>Smenospongia</i> sp. ^{32, 33} <i>Psammocinia</i> sp. ⁴⁸⁴
TeC-291	unnamed tetracycyclic sesterterpene (reported as compound 4)		<i>Smenospongia</i> sp. ³³
TeC-292	16 β -acetoxy-20,24-dimethyl-12,24-dioxo-25-norscalarane		<i>Phyllospongia madagascarensis</i> ³⁹⁶
TeC-293	16 β -hydroxy-24-methyl-12,24-dioxoscalarane-25-al		<i>Lendenfeldia</i> sp. ⁸⁴

TeC-294	2 α -deoxoscalarin		<i>Glossodoris rufomarginata</i> ⁴⁷⁸
TeC-295	19-acetyl-12-deacetoxy-12-oxo-deoxoscalarin		<i>Glossodoris rufomarginata</i> ⁴⁷⁸ <i>Glossodoris pallida</i> ⁴⁷⁹ <i>Glossodoris hikuerensis</i> ⁵²⁸
TeC-296	sesterstatin 1		<i>Hyrtios erecta</i> [sic!] ⁵⁴⁸
TeC-297	sesterstatin 2		<i>Hyrtios erecta</i> [sic!] ⁵⁴⁸
TeC-298	sesterstatin 3		<i>Hyrtios erecta</i> [sic!] ⁵⁴⁸ <i>Hyrtios erectus</i> ⁵²⁴
TeC-299	hyatolide C		<i>Hyattella intestinalis</i> ³⁹⁰
TeC-300	hyatolide D		<i>Hyattella intestinalis</i> ³⁹⁰

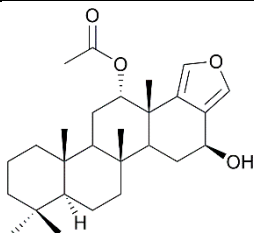
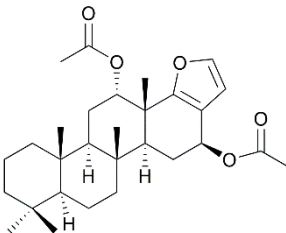
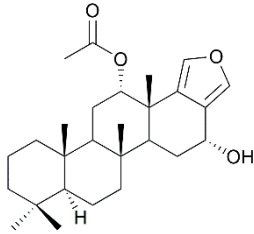
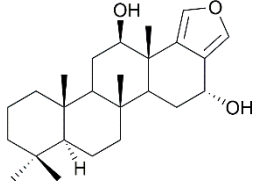
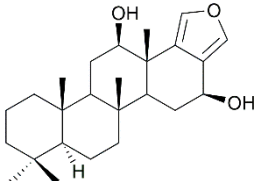
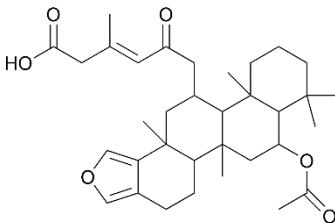
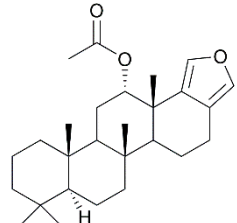
TeC-301	hyatolide E		<i>Hyattella intestinalis</i> ³⁹⁰
TeC-302	phyllofolactone F		<i>Hyattella</i> sp. ⁵⁶⁶ <i>Phyllospongia madagascarensis</i> ³⁹⁶
TeC-303	phyllofolactone G		<i>Hyattella</i> sp. ⁵⁶⁶ <i>Phyllospongia madagascarensis</i> ³⁹⁶
TeC-304	25-dehydroxy-12- <i>epi</i> -sclarin		<i>Hyrtios erecta</i> [sic!] ⁵⁷¹
TeC-305	hyrtiosin A		<i>Hyrtios erecta</i> [sic!] ⁵⁷¹
TeC-306	hyrtiosin B		<i>Hyrtios erecta</i> [sic!] ⁵⁷¹

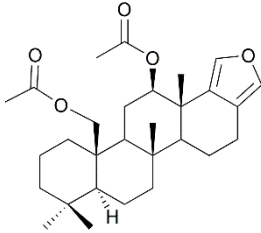
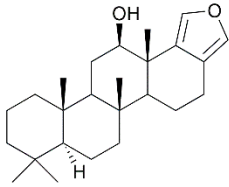
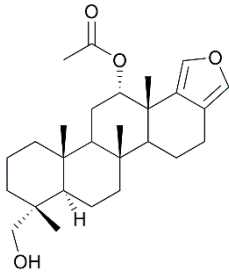
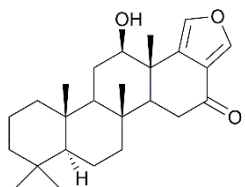
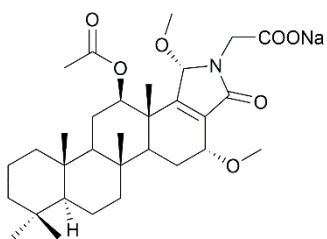
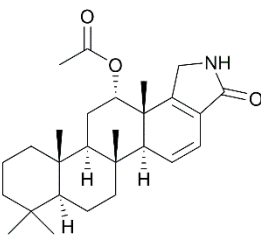
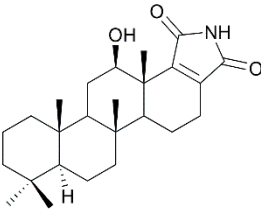
TeC-307	hyrtiosin C		<i>Hyrtios erecta</i> [sic!] ⁵⁷¹
TeC-308	hyrtiosin D		<i>Hyrtios erecta</i> [sic!] ⁵⁷¹
TeC-309	hyrtiosin E		<i>Hippospongia</i> sp. ³⁶⁵ <i>Hyrtios erecta</i> [sic!] ⁵⁷¹
TeC-310	sesterstatin 6		<i>Hyrtios erecta</i> [sic!] ⁵⁷²
TeC-311	12 β ,22,24 ϵ -trihydroxy-24-methylscalar-17-ene-18,24-carbolactone		<i>Carteriospongia</i> sp. ⁵¹²
TeC-312	12-O-acetyl-12,16- <i>epi</i> -scalarolbutenolide		<i>Scalarispongia</i> sp. ⁵⁴⁴

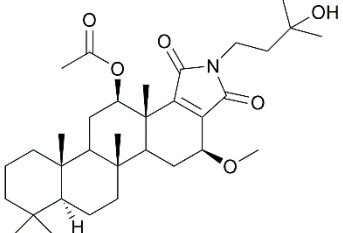
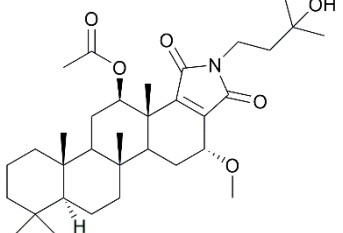
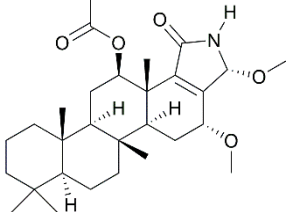
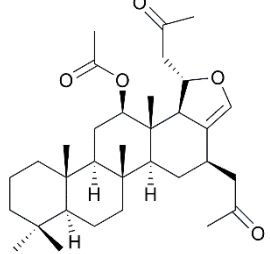
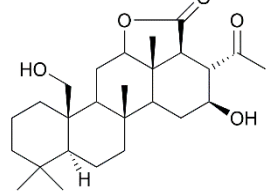
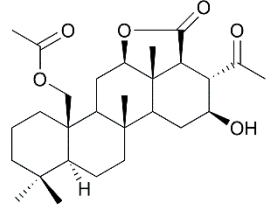
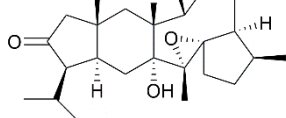
TeC-313	16- <i>O</i> -deacetyl-12,16- <i>epi</i> -scalarolbutanolide		<i>Scalarispongia</i> sp. ⁵⁴⁴
TeC-314	unnamed tetracycyclic sesterterpene (reported as compound 6)		<i>Scalarispongia</i> sp. ⁵⁴⁴
TeC-315	unnamed tetracycyclic sesterterpene (reported as compound 7)		<i>Scalarispongia</i> sp. ⁵⁴⁴
TeC-316	unnamed tetracycyclic sesterterpene (reported as compound 8)		<i>Scalarispongia</i> sp. ⁵⁴⁴
TeC-317	12 α ,16 β -diacetoxy-20,24-dimethyl-24-oxoscalar-25-al		<i>Strepsichordaia lendenfeldi</i> ⁴⁸⁸
TeC-318	12 β ,22-dihydroxy-24-methyl-24-oxoscalar-16-en-25 β -al		<i>Carteriospongia</i> sp. ⁵¹²
TeC-319	12 α -acetoxy-16 β -3'- <i>R</i> -hydroxybutanoyloxy)-20,24-dimethyl-24-oxoscalar-25-al		<i>Strepsichordaia lendenfeldi</i> ⁴⁸⁸

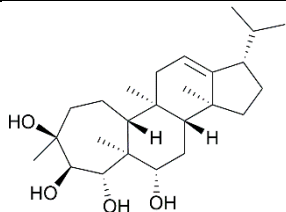
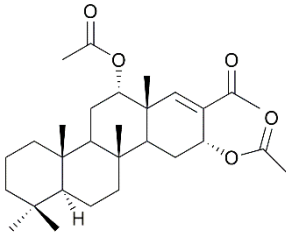
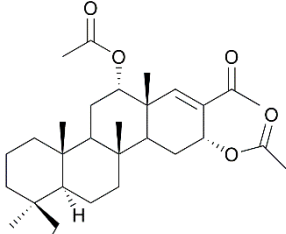
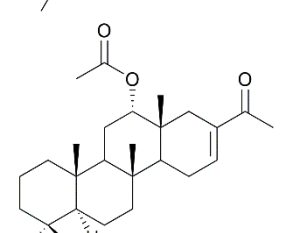
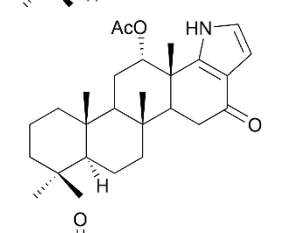
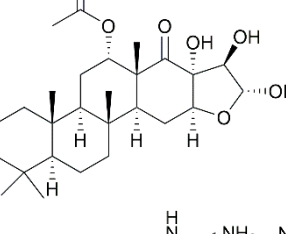
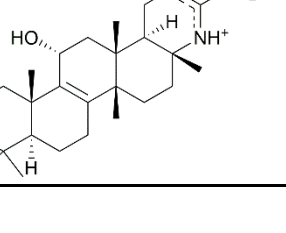
TeC-320	12 α -acetoxy-16 β -(3' <i>R</i> -hydroxypentanoyloxy)-20,24-dimethyl-24-oxoscalaran-25-al		<i>Strepsichordaia lendenfeldi</i> ⁴⁸⁸
TeC-321	12 α -acetoxy-16 β -hydroxy-20,24-dimethyl-24-oxoscalaran-25-al		<i>Strepsichordaia lendenfeldi</i> ⁴⁸⁸ <i>Lendenfeldia</i> sp. ⁸⁴
TeC-322	12 α ,22-dihydroxy-24-methyl-24-oxoscalar-16-en-25 α -al		<i>Carteriospongia</i> sp. ⁵¹²
TeC-323	12 α -acetoxy-16 β -propanoyloxy-20,24-dimethyl-24-oxoscalaran-25-al		<i>Strepsichordaia lendenfeldi</i> ⁴⁸⁸
TeC-324	12 α -(3' <i>R</i> -hydroxypentanoyloxy)-20,24-dimethyl-24-oxoscalar-16-en-25-al		<i>Strepsichordaia lendenfeldi</i> ⁴⁸⁸
TeC-325	12 α -(3'-propanoyloxypentanoyloxy)-20,24-dimethyl-24-oxoscalar-16-en-25-al		<i>Strepsichordaia lendenfeldi</i> ⁴⁸⁸

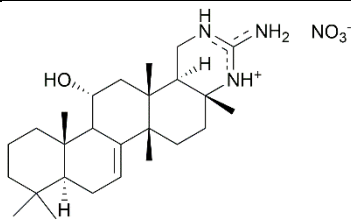
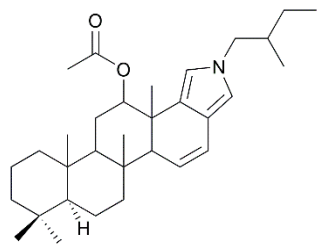
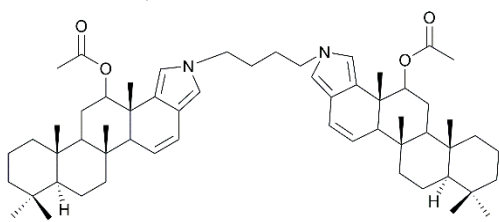
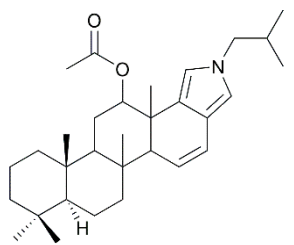
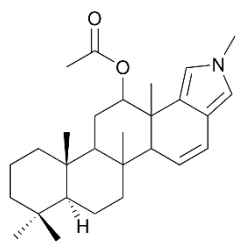
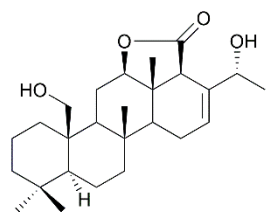
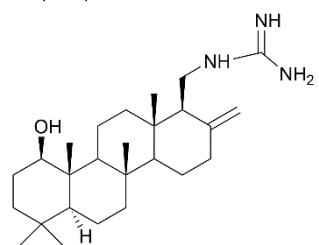
TeC-326	heteronemin		<i>Brachiaster</i> sp. ¹¹¹ <i>Cacospongia scalaris</i> ⁵⁰⁰ <i>Collospongia auris</i> ⁴⁸² <i>Heteronema erecta</i> ^{573, 574} <i>Hippospongia</i> sp. ³⁶⁵ <i>Hyattella intestinalis</i> ⁵³⁴ <i>Hyrtios erecta</i> [sic!] ⁵⁰⁵ <i>Hyrtios cf. erectus</i> ⁵⁵² <i>Hyrtios erectus</i> ⁵²⁴ <i>Spongia idia</i> ²⁶ <i>Ircinia</i> sp. ²³⁹ <i>Glossodoris cincta</i> ⁵⁰⁶ <i>Glossodoris hikeurensis</i> ⁵⁰⁶ <i>Glossodoris atromarginata</i> ⁵³⁷ <i>Hyrtios altum</i> [sic!] ⁵⁰⁶ <i>Glossodoris vespa</i> ⁵⁰⁷ <i>Hyrtios erecta</i> [sic!] ⁵⁷⁵
TeC-327	12- <i>epi</i> -heteronemin		
TeC-328	12- <i>epi</i> -heteronemin acetate		<i>Hyrtios erecta</i> [sic!] ⁵⁷⁵
TeC-329	furoscalarol		<i>Cacospongia mollior</i> ^{576, 577} <i>Hyattella intestinalis</i> ³⁹⁰ <i>Cacospongia scalaris</i> ^{85, 501} <i>Glossodoris tricolor</i> ⁵⁰⁴
TeC-330	12,16-di- <i>epi</i> -12- <i>O</i> -deacetyl-16- <i>O</i> -acetylfuroscalarol		<i>Spongia agaricina</i> ⁵⁷⁸
TeC-331	scalarafuran		<i>Spongia idia</i> ²⁶ <i>Hippospongia</i> sp. ³⁶⁵ <i>Hyrtios gumminae</i> ⁵³⁰

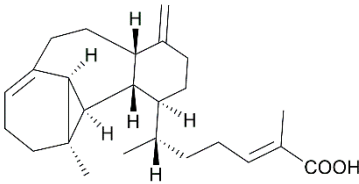
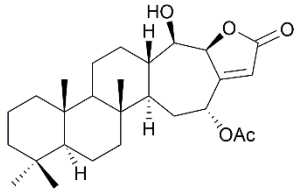
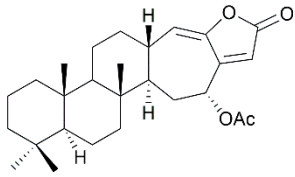
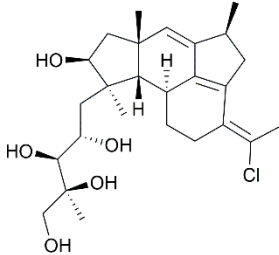
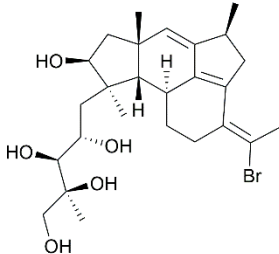
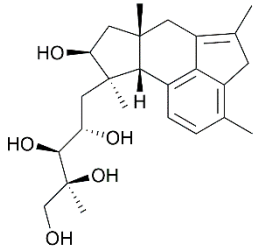
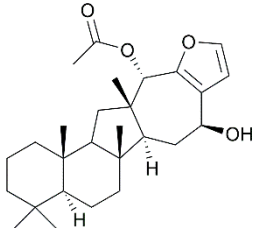
TeC-332	isoscalarafuran-A		<i>Spongia hispida</i> ³⁶⁴ <i>Hyrtios erectus</i> ⁵⁴³
TeC-333	16-acetylfuroscalarol		<i>Cacospongia scalaris</i> ⁵⁰¹
TeC-334	isoscalarafuran-B		<i>Spongia hispida</i> ³⁶⁴
TeC-335	sesterstatin 4 (16- <i>epi</i> -deacetyl-scalarafuran)		<i>Hyrtios erecta</i> [sic!] ⁵⁷⁹
TeC-336	sesterstatin 5		<i>Hyrtios erecta</i> [sic!] ⁵⁷⁹
TeC-337	nambiscalarane		<i>Neonothopanus nambj</i> ⁵⁸⁰
TeC-338	16-deacetoxy-12- <i>epi</i> -scalarafuranacetate		<i>Spongia officinalis</i> ⁵³⁸ <i>Glossodoris averni</i> ⁴⁷⁹

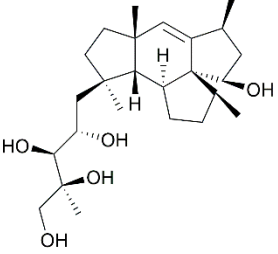
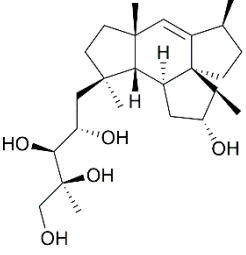
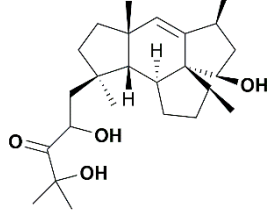
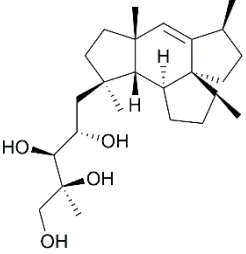
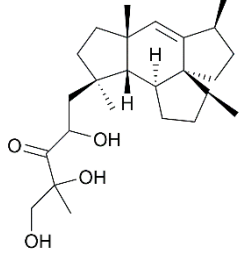
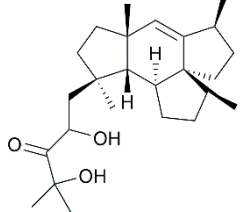
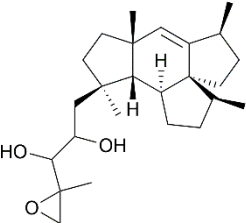
TeC-339	12- <i>O</i> -acetyl-16-deacetoxy-23-acetoxyscalarafuran		<i>Psammocinia</i> sp. ⁴⁸⁴
TeC-340	12- <i>O</i> -deacetylscalarafuran		<i>Spongia</i> sp. ⁵⁴⁶
TeC-341	21-hydroxy-16-deacetyl-12- <i>epi</i> -scalarafuran acetate		<i>Hyrtios erectus</i> ⁵⁴³
TeC-342	salmahyrtisol B*		<i>Hyrtios erecta</i> [sic!] ⁵³²
TeC-343	unnamed tetracyclobicyclic sesterterpene (reported as compound 11)		<i>Hyattella</i> sp. ⁵²⁷
TeC-344	hyatelactam		<i>Hyattella intestinalis</i> ³⁹⁰
TeC-345	sesterstamide		<i>Hyrtios</i> sp. ⁵⁸¹

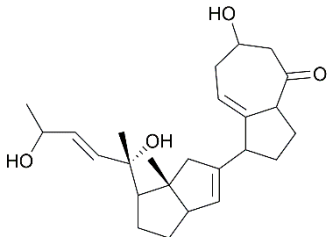
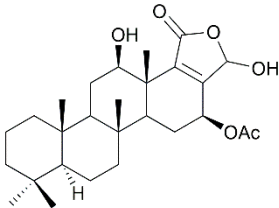
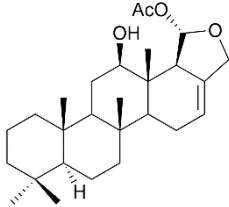
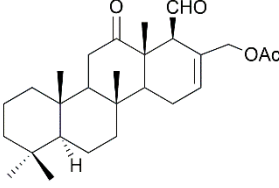
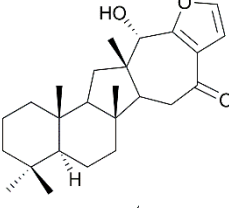
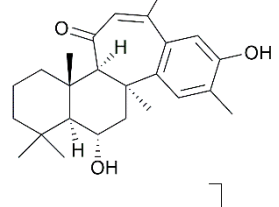
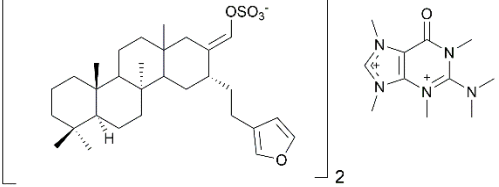
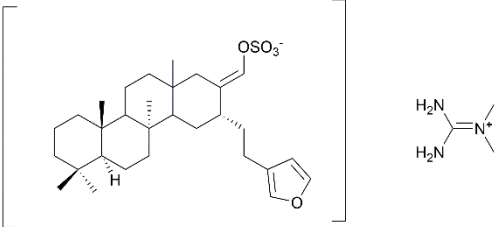
TeC-346	scalalactam A		<i>Spongia</i> sp. ⁵⁸²
TeC-347	scalalactam B		<i>Spongia</i> sp. ⁵⁸²
TeC-348	24-methoxypetrosaspongia C		<i>Hirtios erectus</i> ^{480, 524}
TeC-349	heteronemin acetate		<i>Hirtios erecta</i> [sic!] ⁵⁰⁵ <i>Hippospongia</i> sp. ³⁶⁵ <i>Brachiaster</i> sp. ¹¹¹
TeC-350	16 β ,22-dihydroxy-24-methyl-24-oxoscalaran-25,12 β -olactone (also reported as 16 β ,22-dihydroxy-24-methyl-24-oxoscalaran-25,12 β -olide ⁵⁶⁴ and as homoscalaralactone IIA ⁵⁶³)		<i>Dictyoceratida</i> sp. and <i>Halichondria</i> sp. ⁵¹¹ <i>Lendenfeldia</i> sp. ⁸⁴ <i>Carteriospongia</i> sp. ⁵¹² <i>Lendenfeldia</i> sp. ⁵¹⁶ <i>Phyllospongia dendyi</i> ⁵⁶⁴ <i>Lendenfeldia frondosa</i> ⁵⁶³
TeC-351	22-acetoxy-16 β -hydroxy-24-methyl-24-oxoscalarano-25,12 β -lactone		<i>Lendenfeldia chondrodes</i> ⁵¹⁴ <i>Lendenfeldia</i> sp. ⁸⁴
TeC-352	nitidasin		<i>Gentianella nitida</i> ⁵⁸³

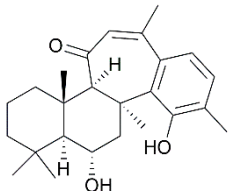
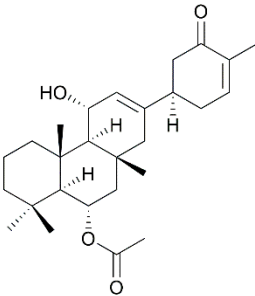
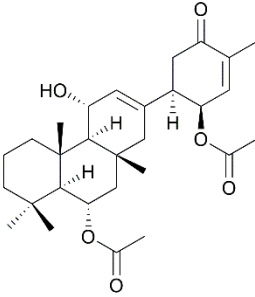
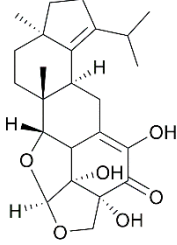
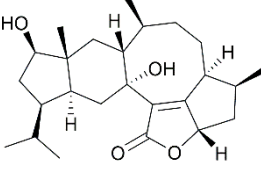
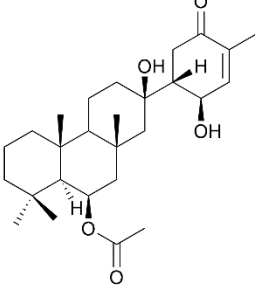
TeC-353	asperunguisin F		<i>Aspergillus unguis</i> ⁴⁶⁶
TeC-354	phyllospingin A		<i>Phyllospongia lamellosa</i> ⁵¹⁷
TeC-355	phyllospingin B		<i>Phyllospongia lamellosa</i> ⁵¹⁷
TeC-356	phyllospingin C		<i>Phyllospongia lamellosa</i> ⁵¹⁷
TeC-357	unnamed tetracarboxylic sesterterpene (reported as compound 4)		<i>Scalarispongia</i> sp. ⁵⁸⁴
TeC-358	hyatellone C		<i>Hyattella intestinalis</i> ³⁹⁰
TeC-359	cybastacine A		<i>Nostoc</i> sp. ⁵⁸⁵

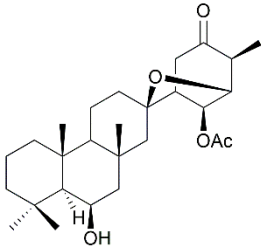
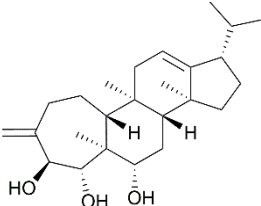
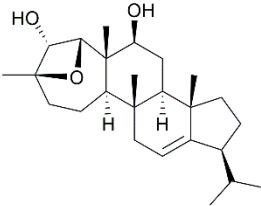
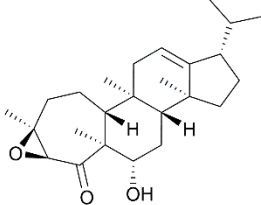
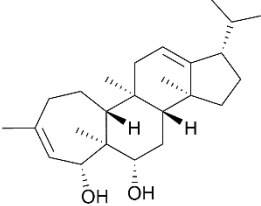
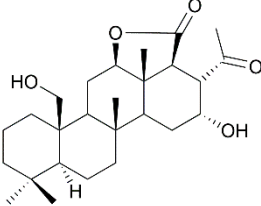
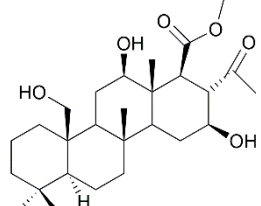
TeC-360	cybastacine B		<i>Nostoc</i> sp. ⁵⁸⁵
TeC-361	molliorin-a		<i>Cacospongia mollior</i> ⁵⁸⁶
TeC-362	molliorin-b		<i>Cacospongia mollior</i> ⁵⁸⁷
TeC-363	molliorin-c		<i>Cacospongia mollior</i> ⁵⁸⁸
TeC-364	molliorin-e		<i>Cacospongia mollior</i> ⁵⁸⁹
TeC-365	sednolide		<i>Chromodoris sedna</i> ⁵⁹⁰
TeC-366	scytoscalarol		<i>Scytonema</i> sp. UTEX 1163 ⁵⁹¹

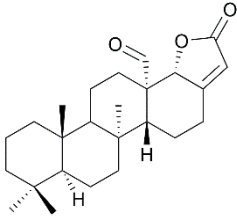
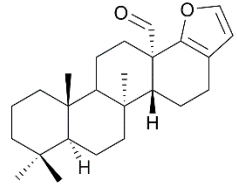
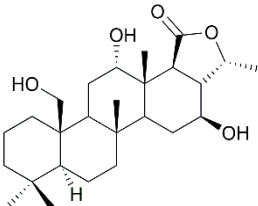
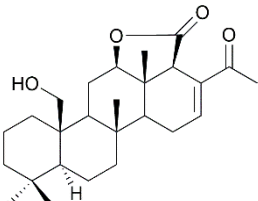
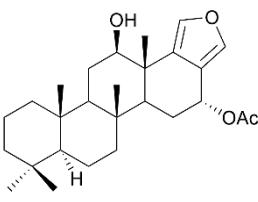
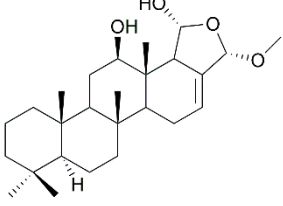
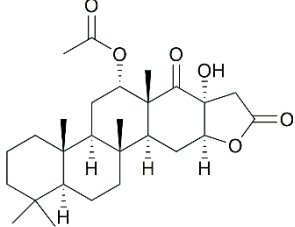
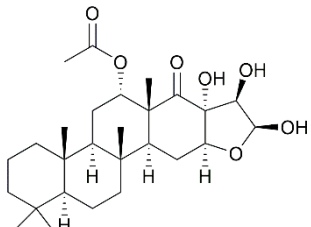
TeC-367	cerorubenic acid-III		<i>Ceroplastes rubens</i> ³⁹¹
TeC-368	inorolide A		<i>Chromodoris inornata</i> ⁴²⁷
TeC-369	inorolide B		<i>Chromodoris inornata</i> ⁴²⁷
TeC-370	neomangicol A		<i>Fusarium</i> spp. strain CNC-477 ⁵⁹²
TeC-371	neomangicol B		<i>Fusarium</i> spp. strain CNC-477 ⁵⁹²
TeC-372	neomangicol C		<i>Fusarium</i> spp. strain CNC-477 ⁵⁹²
TeC-373	salmahyrtisol A		<i>Hyrtios erecta</i> [sic!] ⁵³²

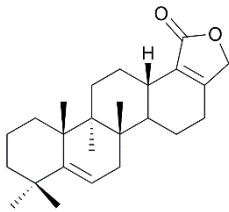
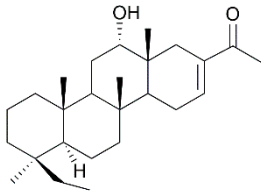
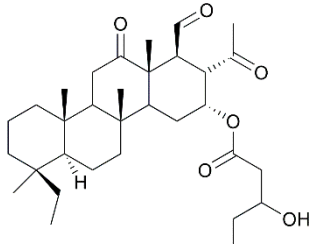
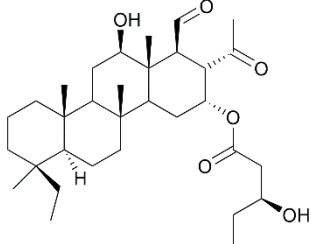
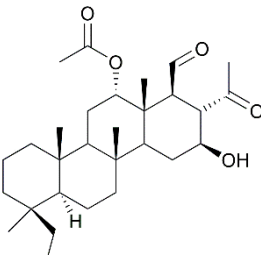
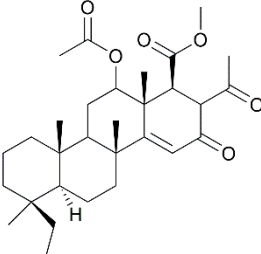
TeC-374	mangicol A		<i>Fusarium heterosporum</i> ⁵⁹³
TeC-375	mangicol B		<i>Fusarium heterosporum</i> ⁵⁹³
TeC-376	mangicol C		<i>Fusarium heterosporum</i> ⁵⁹³
TeC-377	mangicol D		<i>Fusarium heterosporum</i> ⁵⁹³
TeC-378	mangicol E		<i>Fusarium heterosporum</i> ⁵⁹³
TeC-379	mangicol F		<i>Fusarium heterosporum</i> ⁵⁹³
TeC-380	mangicol G		<i>Fusarium heterosporum</i> ⁵⁹³

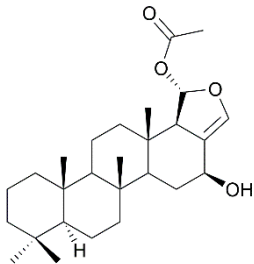
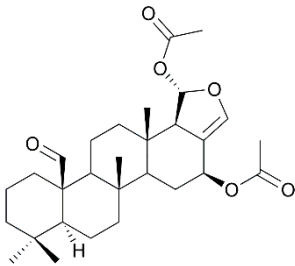
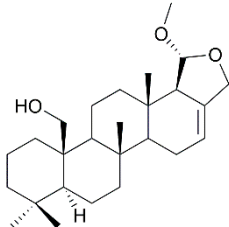
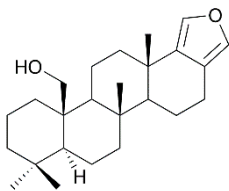
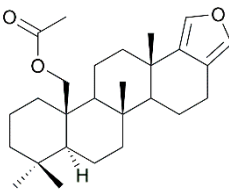
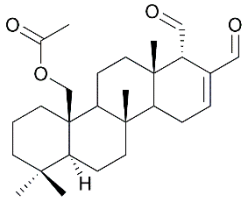
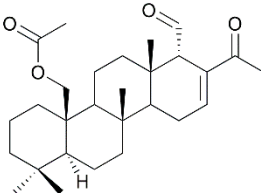
TeC-381	cyclocitrinol		<i>Penicillium citrinum</i> ⁵⁹⁴
TeC-382	12 β ,20-dihydroxy-16 β -acetoxy-17-scalaren-19,20-olide		<i>Hyrtios gumminae</i> ⁵³⁰
TeC-383	19-acetyl-12-deacetyl-12- <i>epi</i> -deoxoscalarin		<i>Glossodoris hikuensis</i> ⁵²⁸
TeC-384	unnamed tetracycyclic sesterterpene (reported as compound 2)		<i>Glossodoris hikuensis</i> ⁵²⁸
TeC-385	hippospongide A		<i>Hippospongia</i> sp. ³⁶⁵
TeC-386	phorone A		<i>Phorbas</i> sp. ³⁹⁹
TeC-387	suvanine N,N-dimethyl-1,3-dimethylherbipoline salt		<i>Coscinoderma</i> sp. ³⁰⁸
TeC-388	suvanine N,N-dimethylguanidium salt		<i>Coscinoderma</i> sp. ³⁰⁸

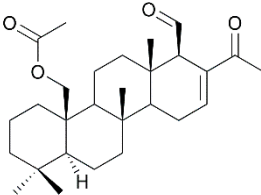
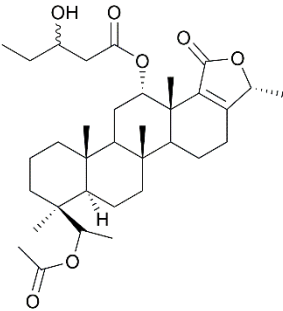
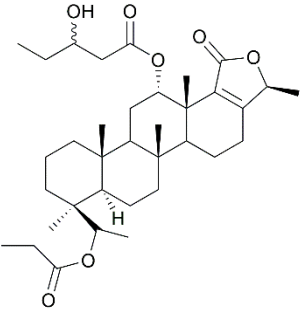
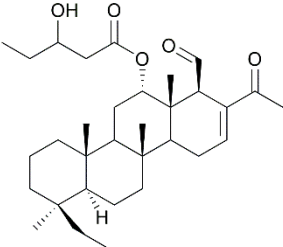
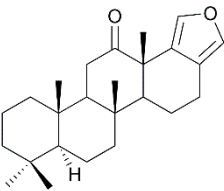
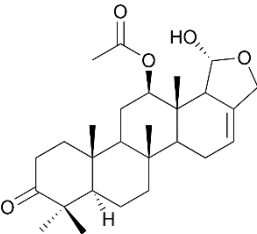
TeC-389	phorone B		<i>Clathria gombawuiensis</i> ⁴⁰¹
TeC-390	anvilone A		<i>Phorbas</i> sp. ¹⁶³
TeC-391	anvilone B		<i>Phorbas</i> sp. ¹⁶³
TeC-392	erinacine S		<i>Hericium erinaceus</i> ⁵⁹⁵
TeC-393	sesteralterin		<i>Alternaria alternata</i> ⁵⁹⁶
TeC-394	19- <i>epi</i> -isosuberitenone B		<i>Phorbas areolatus</i> ⁴⁶⁷

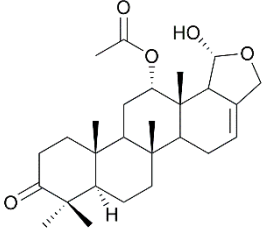
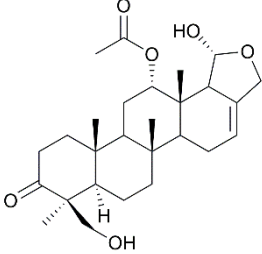
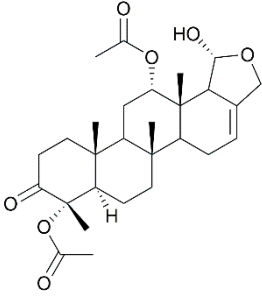
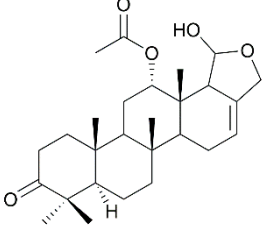
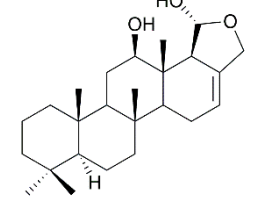
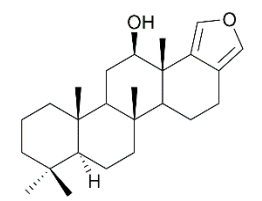
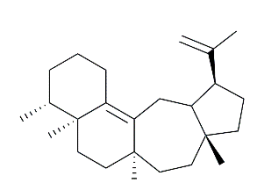
TeC-395	isoxaspirosuberitenone		<i>Phorbas areolatus</i> ⁴⁶⁷
TeC-396	asperunguisin A		<i>Aspergillus unguis</i> ⁴⁶⁶
TeC-397	asperunguisin C		<i>Aspergillus unguis</i> ⁴⁶⁶
TeC-398	asperunguisin D		<i>Aspergillus unguis</i> ⁴⁶⁶
TeC-399	asperunguisin E		<i>Aspergillus unguis</i> ⁴⁶⁶
TeC-400	epihomoscalaralactone IIA		<i>Lendenfeldia frondosa</i> ⁵⁶³
TeC-401	homoscalarate II		<i>Lendenfeldia frondosa</i> ⁵⁶³

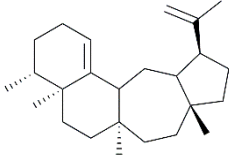
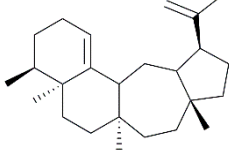
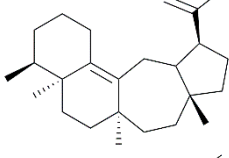
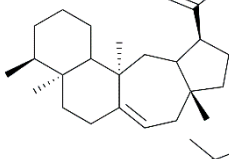
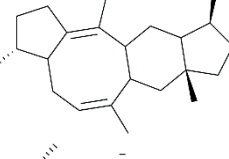
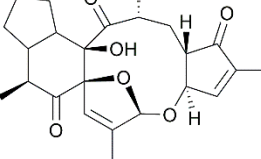
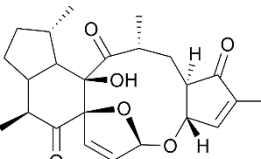
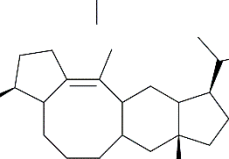
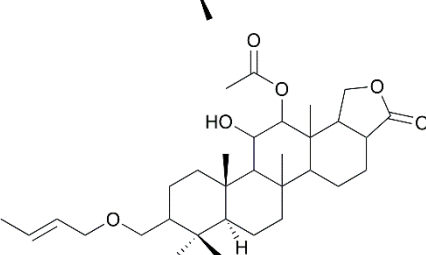
TeC-402	coscinalactone		<i>Coscinoderma mathewsi</i> ⁴¹⁴
TeC-403	coscinafuran		<i>Coscinoderma mathewsi</i> ⁴¹⁴
TeC-404	12 β ,16 β ,22-trihydroxy-24 <i>R</i> -methylscalar-25 β ,24 α -olide		<i>Lendenfeldia frondosa</i> ⁵⁶⁵
TeC-405	22-hydroxy-24-methylsedn-16-en-24-one-12 β ,25 β -olide		<i>Lendenfeldia frondosa</i> ⁵⁶⁵
TeC-406	unnamed tetracycyclic sesterterpene (reported as compound 1)		<i>Hyrtios</i> sp. ⁵³⁵
TeC-407	unnamed tetracycyclic sesterterpene (reported as compound 2)		<i>Hyrtios</i> sp. ⁵³⁵
TeC-408	hyatellone A		<i>Hyattella intestinalis</i> ³⁹⁰
TeC-409	hyatellone B		<i>Hyattella intestinalis</i> ³⁹⁰

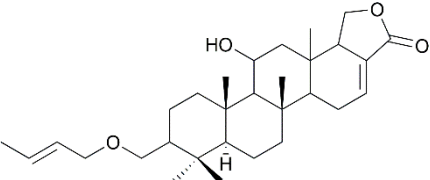
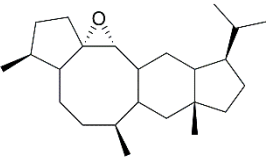
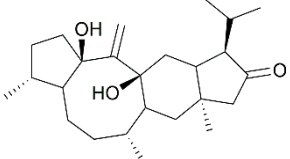
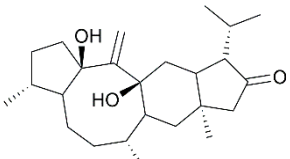
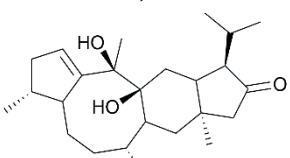
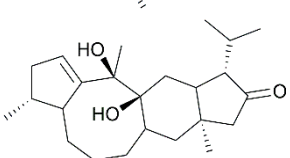
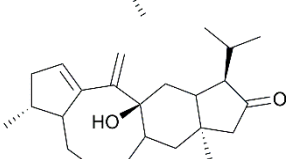
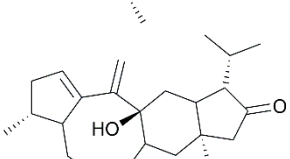
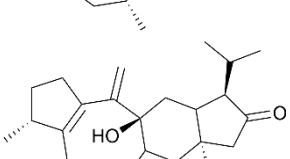
TeC-410	corallocarpsalarolide		<i>Corallocarpus epigaeus</i> ⁵⁹⁷
TeC-411	phyllofenone A		<i>Phyllospongia foliascens</i> ⁵⁹⁸
TeC-412	dehydrofoliaspongin		<i>Phyllospongia foliascens</i> ⁹³
TeC-413	phyllofoliaspongin		<i>Phyllospongia foliascens</i> ⁹³
TeC-414	12 α -acetoxy-16 β -hydroxy-20,24-dimethyl-24-oxoscalar-25-al		<i>Carteriospongia foliascens</i> ^{495, 599}
TeC-415	methyl 12 α -acetoxy-20,24-dimethyl-16,24-dioxoscalara-14,17-dien-25-oate		<i>Carteriospongia foliascens</i> ⁶⁰⁰

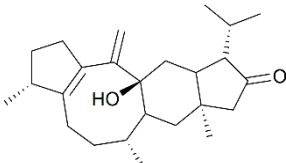
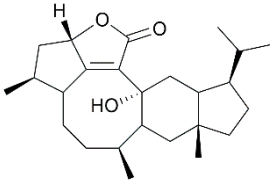
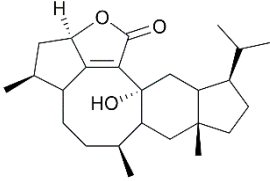
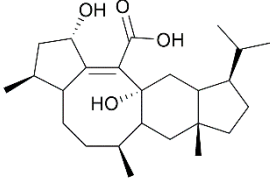
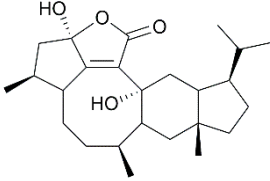
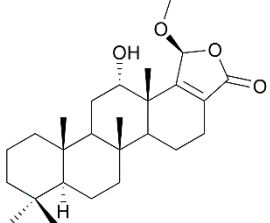
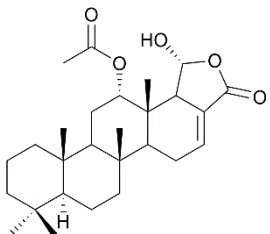
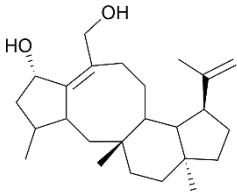
TeC-416	unnamed tetracyclic sesterterpene (12-deacetyl derivative of scalarin) (reported as compound 5)		<i>Smenospongia</i> sp. ³³
TeC-417	unnamed tetracyclic sesterterpene (reported as compound 6)		<i>Smenospongia</i> sp. ³³
TeC-418	unnamed tetracyclic sesterterpene (reported as compound 8)		<i>Smenospongia</i> sp. ³³
TeC-419	unnamed tetracyclic sesterterpene (reported as compound 9)		<i>Smenospongia</i> sp. ³³
TeC-420	unnamed tetracyclic sesterterpene (reported as compound 10)		<i>Smenospongia</i> sp. ³³
TeC-421	unnamed tetracyclic sesterterpene (reported as compound 11)		<i>Smenospongia</i> sp. ³³
TeC-422	unnamed tetracyclic sesterterpene (reported as compound 12)		<i>Smenospongia</i> sp. ³³

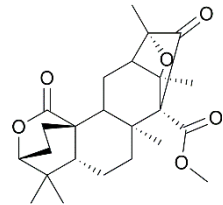
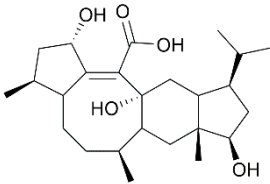
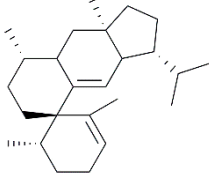
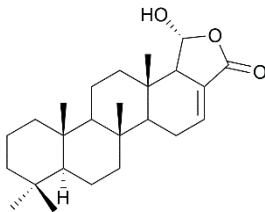
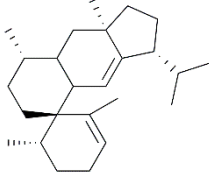
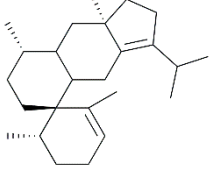
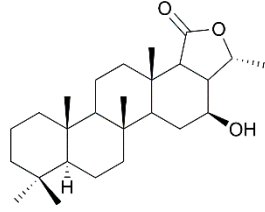
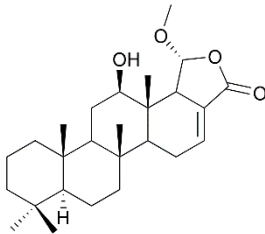
TeC-423	unnamed tetracarboxylic sesterterpene (reported as compound 13)		<i>Smenospongia</i> sp. ³³
TeC-424	honulactone J		<i>Strepsichordaia aliena</i> ⁵⁶⁰
TeC-425	honulactone K		<i>Strepsichordaia aliena</i> ⁵⁶⁰
TeC-426	12 α -(3 <i>R</i> '-hydroxybutanoyloxy)-20,24-dimethyl-24-oxoscalar-16-en-25-al		<i>Strepsichordaia lendenfeldi</i> ⁴⁸⁸
TeC-427	12,16-deacetoxy-12-oxo-scalarafuran		<i>Glossodoris pallida</i> ⁴⁷⁹ <i>Glossodoris averni</i> ⁴⁷⁹
TeC-428	12- <i>epi</i> -deoxoscalarin-3-one		<i>Chromodoris inornata</i> ⁶⁰¹

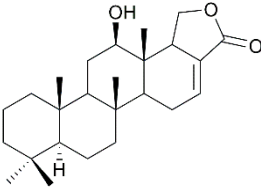
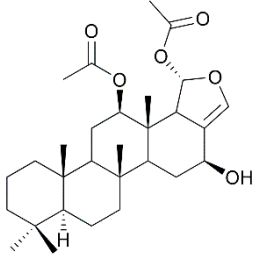
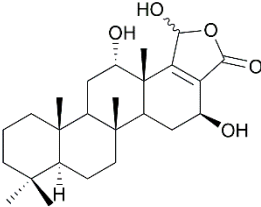
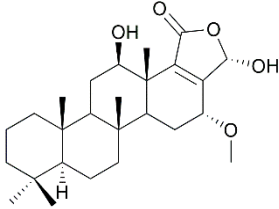
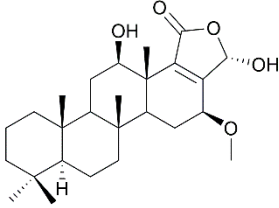
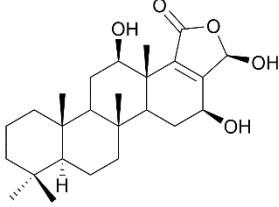
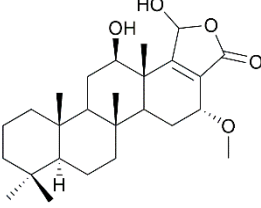
TeC-429	deoxoscalarin-3-one		<i>Chromodoris inornata</i> ⁶⁰¹
TeC-430	21-hydroxydeoxoscalarin		<i>Chromodoris inornata</i> ⁶⁰¹
TeC-431	21-acetoxyscalarin		<i>Chromodoris inornata</i> ⁶⁰¹
TeC-432	ketodeoxoscalarin		<i>Chromodoris funerea</i> ¹³⁰
TeC-433	12-deacetyl-12- <i>epi</i> -deoxoscalarin		<i>Glossodoris atromarginata</i> ⁵³⁷ <i>Glossodoris pallida</i> ⁴⁷⁹ <i>Glossodoris hikuensis</i> ⁵²⁸
TeC-434	16-deacetoxy-scalarafuran		<i>Glossodoris vespa</i> ⁴⁷⁹
TeC-435	(-)-caprudiene A		<i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁵

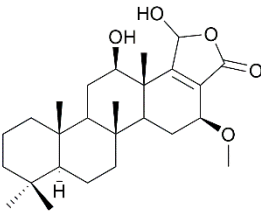
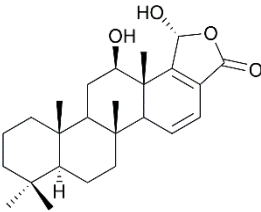
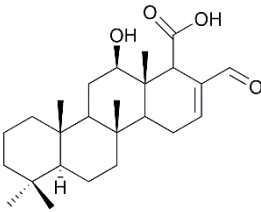
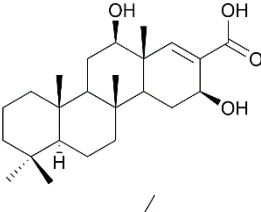
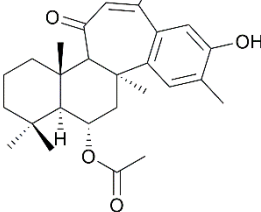
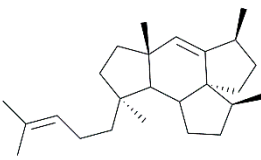
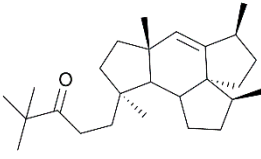
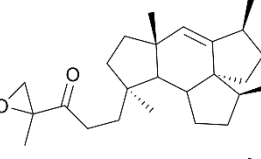
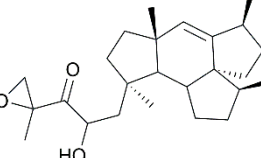
TeC-436	(+)-braparadiene A		<i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁵
TeC-437	(-)-braparadiene B		<i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁵
TeC-438	(-)-arathanadiene A		<i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁵
TeC-439	(-)-arathanadiene B		<i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁵
TeC-440	sesterevisene		<i>Escherichia coli</i> strain DH10B/ <i>Saccharomyces cerevisiae</i> YZL141 ⁶⁰²
TeC-441	eurysoloid A		<i>Eurysolen gracilis</i> ⁶⁰³
TeC-442	eurysoloid B		<i>Eurysolen gracilis</i> ⁶⁰³
TeC-443	unnamed tetracycyclic sesterterpene (reported as sesteralterinderivative, compound 1)		<i>Alternaria alternata</i> ⁶⁰⁴
TeC-444	erectascalarane A		<i>Hyrtios erectus</i> ⁶⁰⁵

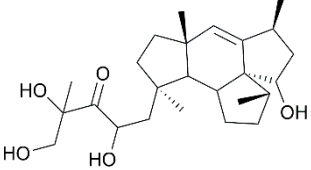
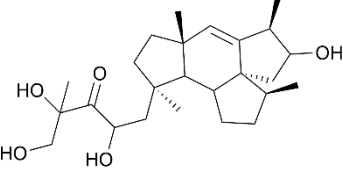
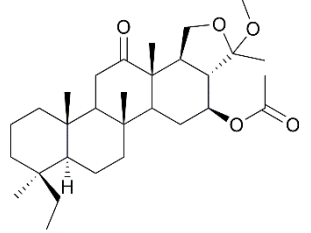
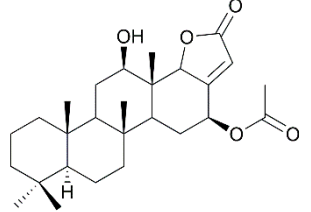
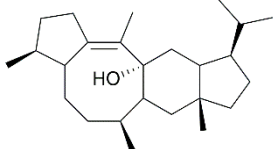
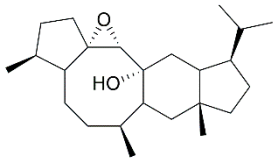
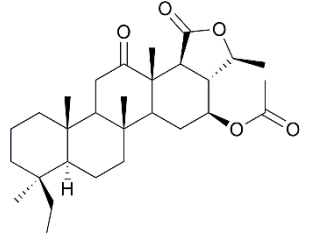
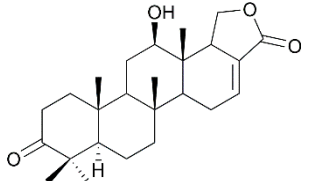
TeC-445	erectasclarane B		<i>Hyrtios erectus</i> ⁶⁰⁵
TeC-446	unnamed tetracyclic sesterterpene (reported as sesteralterinderivative, compound 2)		<i>Alternaria alternata</i> ⁶⁰⁴
TeC-447	gentianelloid C		<i>Gentianella turkestanorum</i> ⁴³⁴
TeC-448	18- <i>epi</i> -gentianelloid C		<i>Gentianella turkestanorum</i> ⁴³⁴
TeC-449	gentianelloid D		<i>Gentianella turkestanorum</i> ⁴³⁴
TeC-450	18- <i>epi</i> -gentianelloid D		<i>Gentianella turkestanorum</i> ⁴³⁴
TeC-451	gentianelloid E		<i>Gentianella turkestanorum</i> ⁴³⁴
TeC-452	18- <i>epi</i> -gentianelloid E		<i>Gentianella turkestanorum</i> ⁴³⁴
TeC-453	gentianelloid F		<i>Gentianella turkestanorum</i> ⁴³⁴

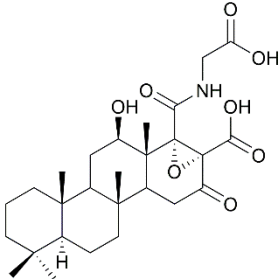
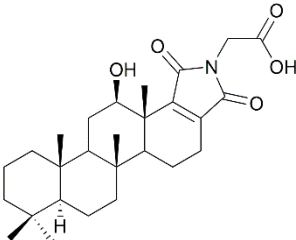
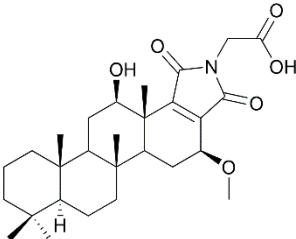
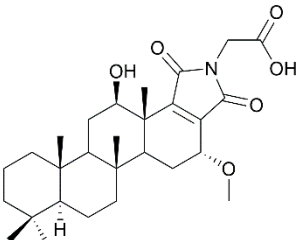
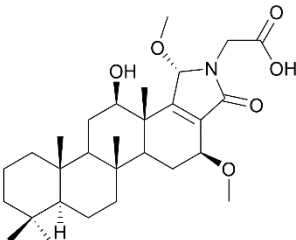
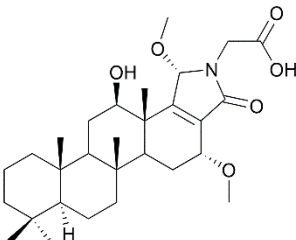
TeC-454	18- <i>epi</i> -gentianelloid F		<i>Gentianella turkestanorum</i> ⁴³⁴
TeC-455	unnamed tetracycyclic sesterterpene (reported as sesteralterinderivative, compound 3)		<i>Alternaria alternata</i> ⁶⁰⁴
TeC-456	unnamed tetracycyclic sesterterpene (reported as sesteralterinderivative, compound 4)		<i>Alternaria alternata</i> ⁶⁰⁴
TeC-457	unnamed tetracycyclic sesterterpene (reported as sesteralterinderivative, compound 5)		<i>Alternaria alternata</i> ⁶⁰⁴
TeC-458	unnamed tetracycyclic sesterterpene (reported as sesteralterinderivative, compound 6)		<i>Alternaria alternata</i> ⁶⁰⁴
TeC-459	dysiscalarone D		<i>Dysidea granulosa</i> ⁶⁰⁶
TeC-460	dysiscalarone E		<i>Dysidea granulosa</i> ⁶⁰⁶
TeC-461	spectanoid H		<i>Aspergillus spectabilis</i> ⁴³⁸

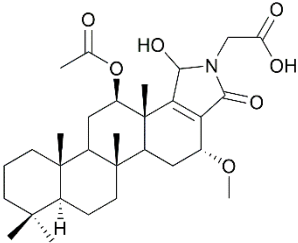
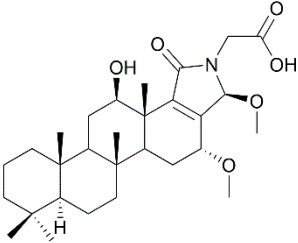
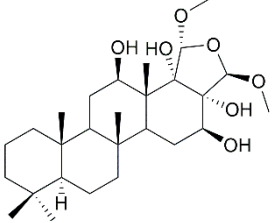
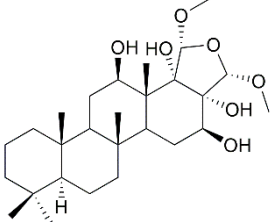
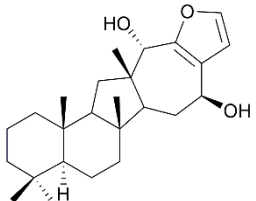
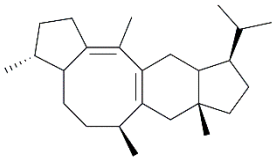
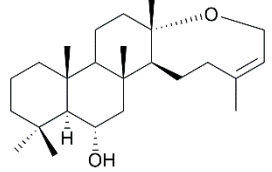
TeC-462	unnamed tetracarboxylic sesterterpene (reported as compound 1)		Endophytic fungus of the La Encrucijada tropical mangrove of Tapachula (Chiapas, Mexico) ⁶⁰⁷
TeC-463	unnamed tetracarboxylic sesterterpene (reported as sesteralterinderivative, compound 7)		<i>Alternaria alternata</i> ⁶⁰⁴
TeC-464	sesterviolene A		<i>Streptomyces violens</i> ²⁷³
TeC-465	deacetoxyscalarin		<i>Spongia</i> sp. ⁵³⁶
TeC-466	sesterviolene C		<i>Streptomyces violens</i> ²⁷³
TeC-467	sesterviolene D		<i>Streptomyces violens</i> ²⁷³
TeC-468	12 α -acetoxy-16 β -hydroxy-20,24-dimethyl-25-norscalarane-18,24-carbolactone		<i>Carteriospongia foliascens</i> ⁴⁹⁵
TeC-469	12-O-deacetyl-19 α -methoxy-12- <i>epi</i> -scalarin		<i>Hyrtilios erectus</i> ⁵⁴⁵

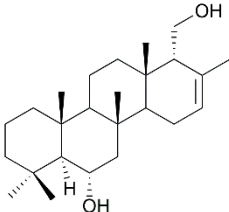
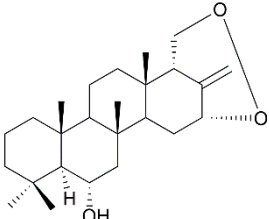
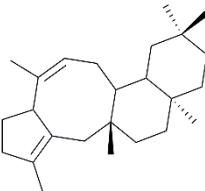
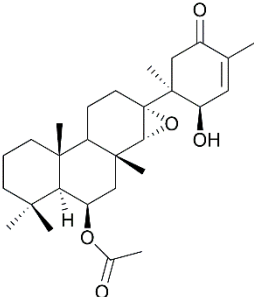
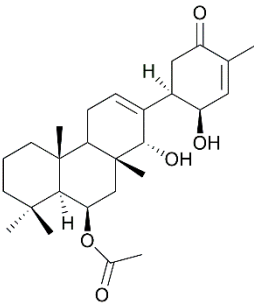
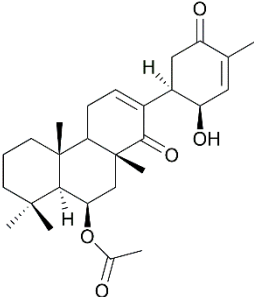
TeC-470	25-dehydroxy-12- <i>epi</i> -deacetylscalarin		<i>Heteronema erecta</i> ⁶⁰⁸ <i>Hyatella cribriformis</i> ⁵⁴⁷
TeC-471	hyrtiosin F		<i>Hyrtios erectus</i> ⁶⁰⁹
TeC-472	hyrtiosin G		<i>Hyrtios erectus</i> ⁶⁰⁹
TeC-473	12 β ,20 β -dihydroxy-16 α -methoxy-17-scalaren-19,20-olide		<i>Hyrtios erectus</i> ⁶¹⁰
TeC-474	12 β ,20 α -dihydroxy-16 β -methoxy-17-scalaren-19,20-olide		<i>Hyrtios erectus</i> ⁶¹⁰
TeC-475	12 β ,16 β ,20 β -trihydroxy-17-scalaren-19,20-olide		<i>Hyrtios erectus</i> ⁶¹⁰
TeC-476	12 β ,19 α (β)-dihydroxy-16 α -methoxy-17-scalaren-19,20-olide		<i>Hyrtios erectus</i> ⁶¹⁰

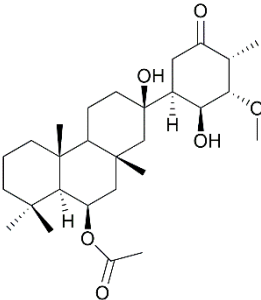
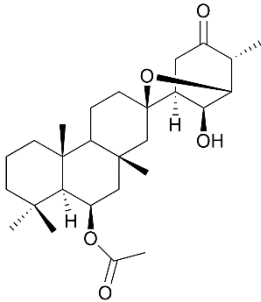
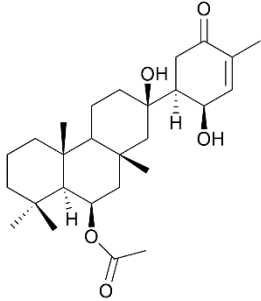
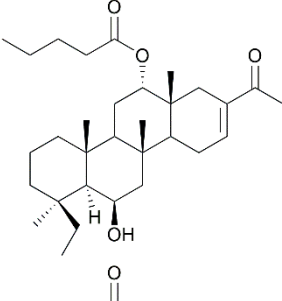
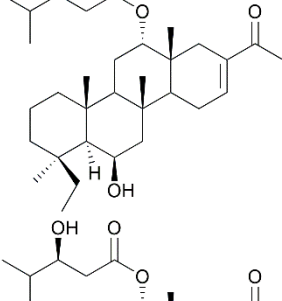
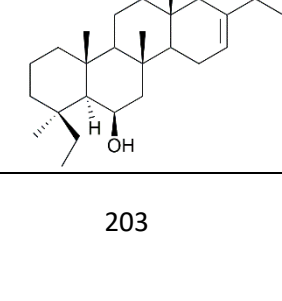
TeC-477	12 β ,19 α (β)-dihydroxy-16 β -methoxy-17-scalaren-19,20-olide		<i>Hyrtios erectus</i> ⁶¹⁰
TeC-478	12 β ,19 α -dihydroxy-14,15-dehydrate-17-scalaren-19,20-olide		<i>Hyrtios erectus</i> ⁶¹⁰
TeC-479	12-deacetyl-18- <i>epi</i> -carboxylic-12- <i>epi</i> -scalaral		<i>Hyrtios erectus</i> ⁶¹⁰
TeC-480	2- <i>O</i> -deacetyl-12,16-di- <i>epi</i> -norscalaral B		<i>Hyrtios erectus</i> ⁶¹⁰
TeC-481	phorone C		<i>Phorbas</i> sp. ⁴⁴³
TeC-482	mangicdiene		<i>Aspergillus oryzae</i> ⁶¹¹ <i>Escherichia coli</i> ¹⁶⁰
TeC-483	mangicol H		<i>Aspergillus oryzae</i> ⁶¹¹
TeC-484	mangicol I		<i>Aspergillus oryzae</i> ⁶¹¹
TeC-485	mangicol J		<i>Aspergillus oryzae</i> ⁶¹¹

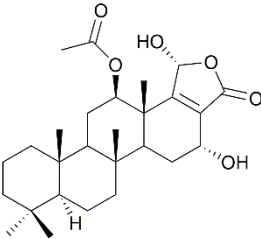
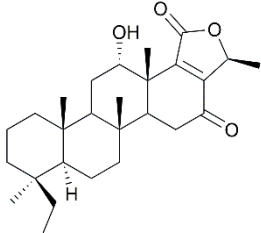
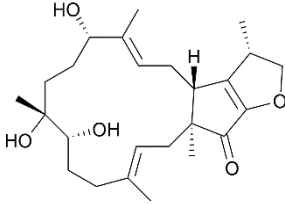
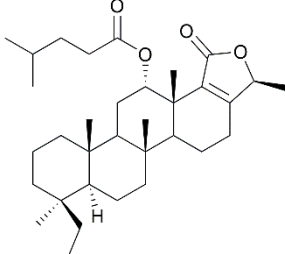
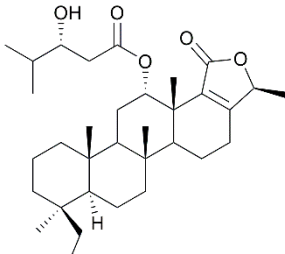
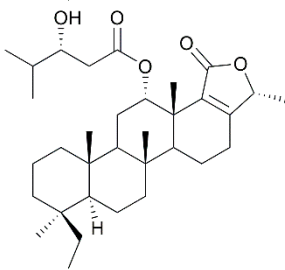
TeC-486	mangicol K		<i>Aspergillus oryzae</i> ⁶¹¹
TeC-487	mangicol L		<i>Aspergillus oryzae</i> ⁶¹¹
TeC-488	phylloketal		<i>Phyllospongia foliascens</i> ⁶¹²
TeC-489	16- <i>epi</i> -scalarole (16- <i>epi</i> -scalarolbutenolide)		<i>Hyrtios erecta</i> ⁵²⁹
TeC-490	sesteraltererol		<i>Alternaria alternata</i> MB-30/ <i>Leucosceptrum canum</i> ²⁷²
TeC-491	10,11-epoxysesteraltererol		<i>Alternaria alternata</i> MB-30/ <i>Leucosceptrum canum</i> ²⁷²
TeC-492	acetoxypyllofolactone A		<i>Phyllospongia foliascens</i> ⁶¹³
TeC-493	12-deacetyl-3-oxoscalarin		<i>Hyrtios erectus</i> ⁶¹⁴

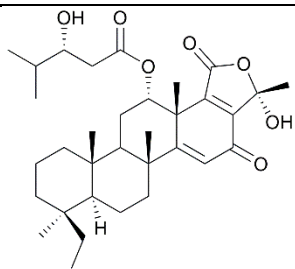
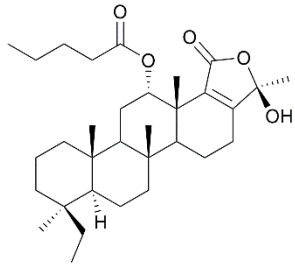
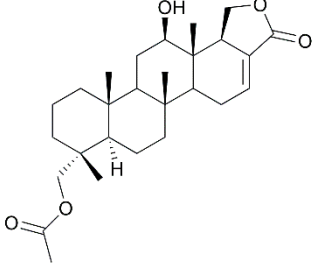
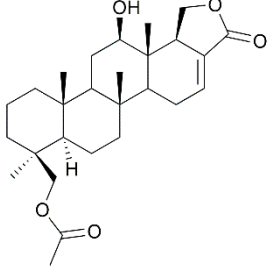
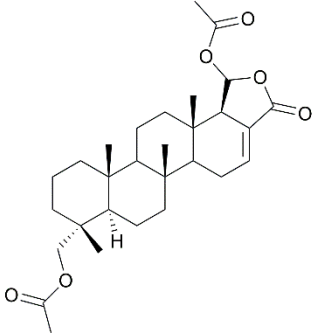
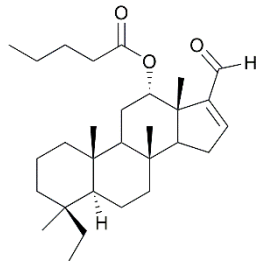
TeC-494	hyrtioscalarin A		<i>Hyrtios erectus</i> ⁶¹⁴
TeC-495	hyrtioscalarin B		<i>Hyrtios erectus</i> ⁶¹⁴
TeC-496	hyrtioscalarin C		<i>Hyrtios erectus</i> ⁶¹⁴
TeC-497	hyrtioscalarin D		<i>Hyrtios erectus</i> ⁶¹⁴
TeC-498	hyrtioscalarin E		<i>Hyrtios erectus</i> ⁶¹⁴
TeC-499	hyrtioscalarin F		<i>Hyrtios erectus</i> ⁶¹⁴

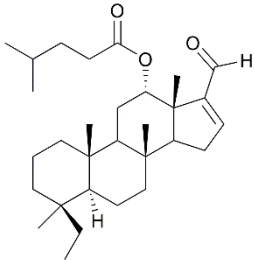
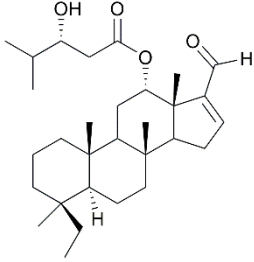
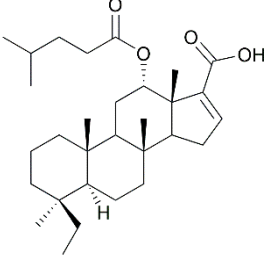
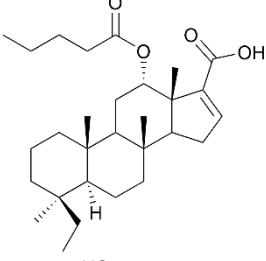
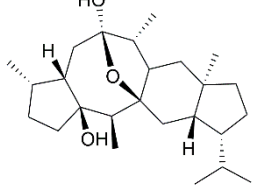
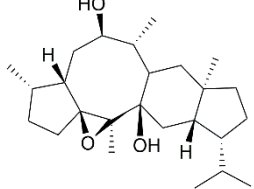
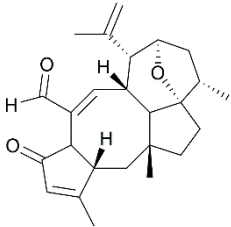
TeC-500	hyrtioscalarin G		<i>Hyrtios erectus</i> ⁶¹⁴
TeC-501	hyrtioscalarin H		<i>Hyrtios erectus</i> ⁶¹⁴
TeC-502	17(<i>R</i>),18(<i>S</i>)-dihydroxy-19(<i>R</i>),20(<i>S</i>)-dimethoxysesterstatin 5		<i>Hyrtios erectus</i> ⁶¹⁴
TeC-503	17(<i>R</i>),18(<i>S</i>)-dihydroxy-19(<i>R</i>),20(<i>R</i>)-dimethoxysesterstatin 5		<i>Hyrtios erectus</i> ⁶¹⁴
TeC-504	salmahyrtisol B*		<i>Hyrtios erectus</i> ⁶¹⁴
Tec-505	Bm2		<i>Aspergillus oryzae</i> ²⁶⁷
Tec-506	(17 <i>Z</i>)-13,19-epoxycheilanth-17-en-6 α -ol		<i>Aleuritopteris mexicana</i> ⁶¹⁵

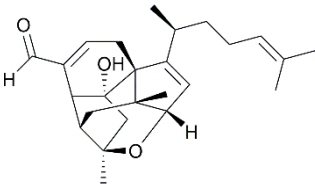
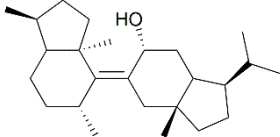
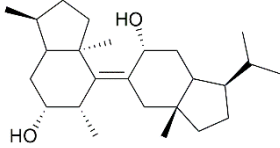
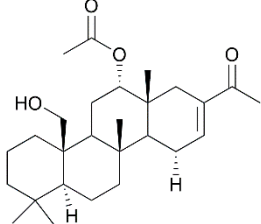
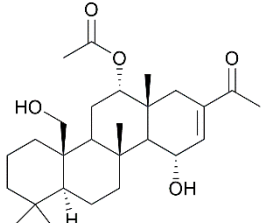
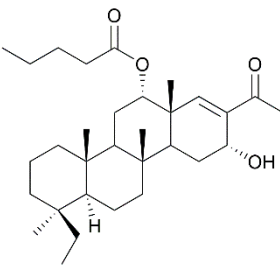
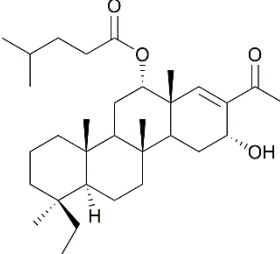
Tec-507	18-episcalar-16-ene-6 α ,19-diol		<i>Aleuritopteris mexicana</i> ⁶¹⁵
Tec-508	16 α ,19-epidioxy-18-episcalar-17(25)-en-6 α -ol		<i>Aleuritopteris mexicana</i> ⁶¹⁵
Tec-509	unamed 5-8-6-6 tetracyclic sesterterpene (reported as compound 56)		<i>Escherichia coli</i> ¹⁶⁰
Tec-510	suberitenone E		<i>Suberites</i> sp. ⁴⁴⁷
Tec-511	suberitenone F		<i>Suberites</i> sp. ⁴⁴⁷
Tec-512	suberitenone G		<i>Suberites</i> sp. ⁴⁴⁷

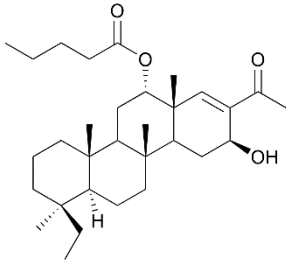
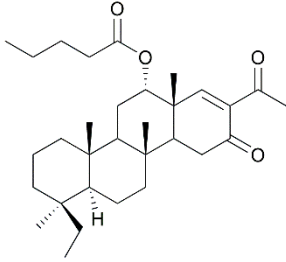
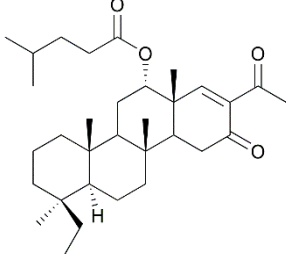
Tec-513	suberitenone H		<i>Suberites</i> sp. ⁴⁴⁷
Tec-514	suberitenone I		<i>Suberites</i> sp. ⁴⁴⁷
Tec-515	suberitenone J		<i>Suberites</i> sp. ⁴⁴⁷
Tec-516	phyllofenone F		<i>Phyllospongia foliascens</i> ⁶¹⁶
Tec-517	phyllofenone G		<i>Phyllospongia foliascens</i> ⁶¹⁶
Tec-518	phyllofenone H		<i>Phyllospongia foliascens</i> ⁶¹⁶

Tec-519	12-β-O-acetylhyrtiolide		<i>Sarcotragus</i> sp. ²⁷⁸
Tec-520	phyllofolactone N		<i>Phyllospongia foliascens</i> ⁶¹⁷
Tec-521	phyllofolactone O		<i>Phyllospongia foliascens</i> ⁶¹⁷
Tec-522	phyllofolactone P		<i>Phyllospongia foliascens</i> ⁶¹⁷
Tec-523	phyllofolactone Q		<i>Phyllospongia foliascens</i> ⁶¹⁷
Tec-524	phyllofolactone R		<i>Phyllospongia foliascens</i> ⁶¹⁷

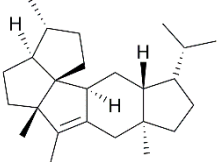
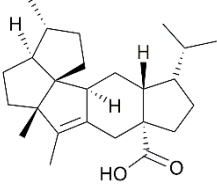
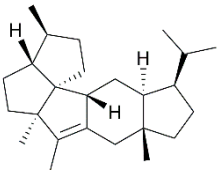
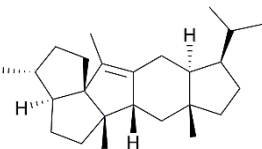
Tec-525	phyllofolactone S		<i>Phyllospongia foliascens</i> ⁶¹⁷
Tec-526	phyllofolactone T		<i>Phyllospongia foliascens</i> ⁶¹⁷
Tec-527	hyrtiosin H		<i>Hyrtios erecta</i> [sic!] ⁶¹⁸
Tec-528	hyrtiosin I		<i>Hyrtios erecta</i> [sic!] ⁶¹⁸
Tec-529	hyrtiosin J		<i>Hyrtios erecta</i> [sic!] ⁶¹⁸
Tec-530	phyllospongiane A		<i>Phyllospongia foliascens</i> ⁶¹⁶

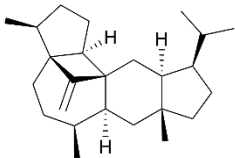
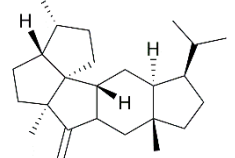
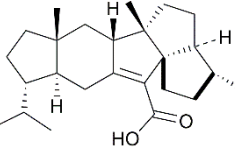
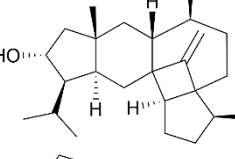
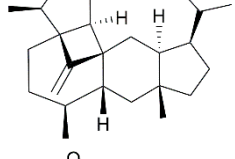
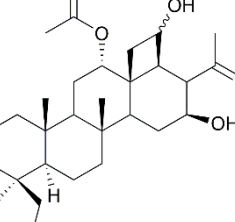
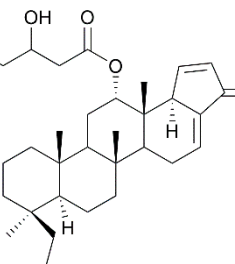
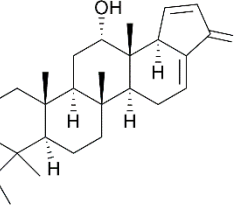
Tec-531	phyllospongiane B		<i>Phyllospongia foliascens</i> ⁶¹⁶
Tec-532	phyllospongiane C		<i>Phyllospongia foliascens</i> ⁶¹⁶
Tec-533	phyllospongiane D		<i>Phyllospongia foliascens</i> ⁶¹⁶
Tec-534	phyllospongiane E		<i>Phyllospongia foliascens</i> ⁶¹⁶
Tec-535	sesterchaetin A		<i>Chaetomium globosum</i> SD-347 ⁶¹⁹
Tec-536	sesterchaetin B		<i>Chaetomium globosum</i> SD-347 ⁶¹⁹
Tec-537	bipoladien A		<i>Bipolaris maydis</i> ⁴⁵¹

Tec-538	bipoladien B		<i>Bipolaris maydis</i> ⁴⁵¹
Tec-539	emerindanol A		<i>Emericella</i> sp. XL-029 ⁶²⁰
Tec-540	emerindanol B		<i>Emericella</i> sp. XL-029 ⁶²⁰
Tec-541	felixin A		<i>Lendenfeldia</i> sp. ⁶²¹
Tec-542	lendenfeldarane V		<i>Lendenfeldia</i> sp. ⁶²¹
Tec-543	phyllofenone I		<i>Phyllospongia foliascens</i> ⁶¹⁶
Tec-544	phyllofenone J		<i>Phyllospongia foliascens</i> ⁶¹⁶

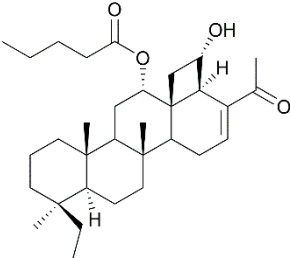
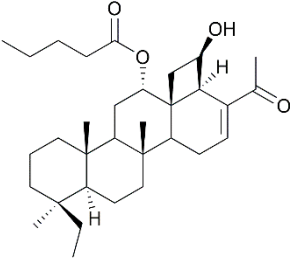
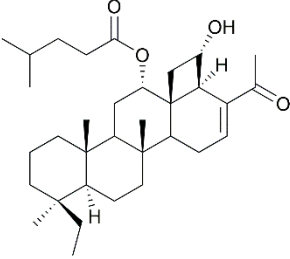
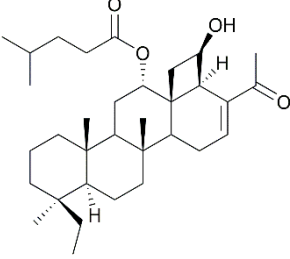
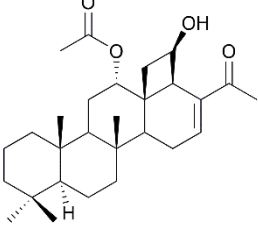
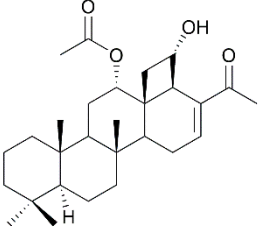
Tec-545	phyllofenone K		<i>Phyllospongia foliascens</i> ⁶¹⁶
Tec-546	phyllofenone L		<i>Phyllospongia foliascens</i> ⁶¹⁶
Tec-547	phyllofenone M		<i>Phyllospongia foliascens</i> ⁶¹⁶

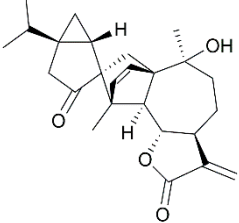
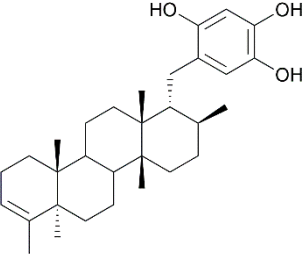
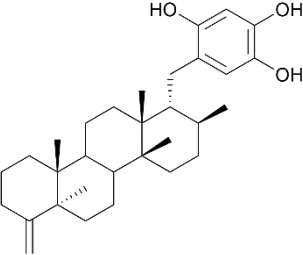
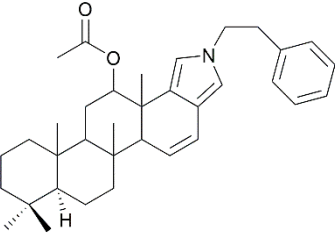
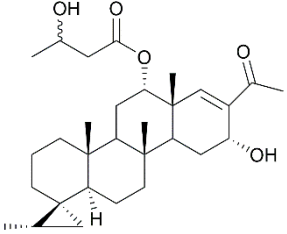
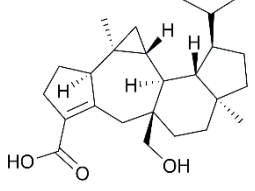
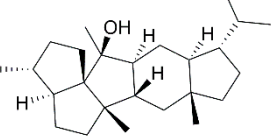
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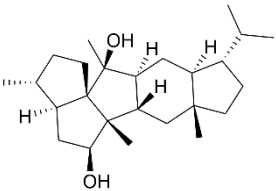
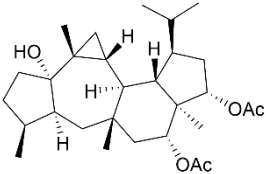
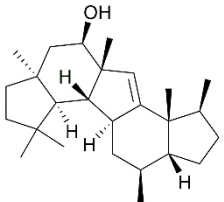
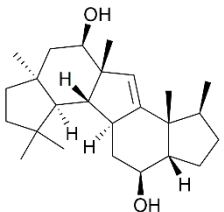
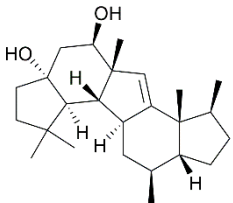
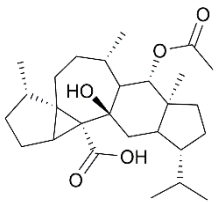
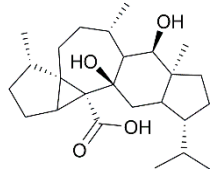
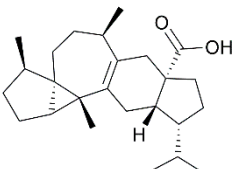
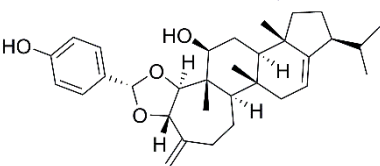
Section 1.9. Structures of pentacarbocyclic (PC) sesterterpenoids (with and without hetherocycles)			
c. n.	IUPAC name or Trivial name	Structure	Organism as reported in the reference (reference)
PC-1	quiannulatene		<i>Emericella variecolor</i> ⁶²²
PC-2	quiannulatic acid		<i>Emericella variecolor</i> ⁶²²
PC-3	(-)- <i>ent</i> - quiannulatene		<i>Arabidopsis thaliana</i> ²⁶⁶ <i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁶
PC-4	(-)-retigeranin B		<i>Arabidopsis thaliana</i> / <i>Escherichia coli</i> ^{266, 357} <i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁶

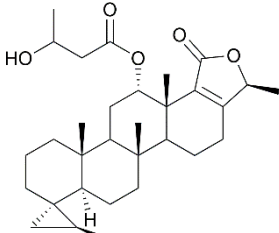
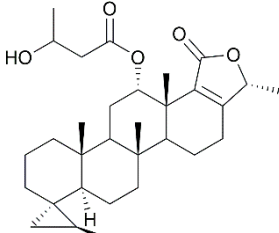
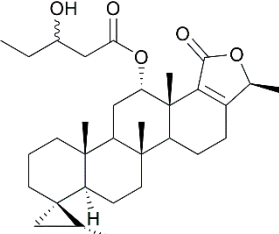
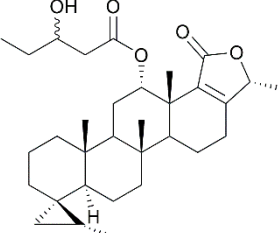
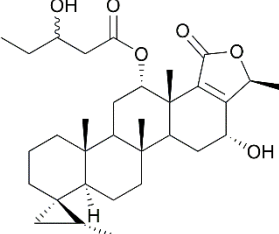
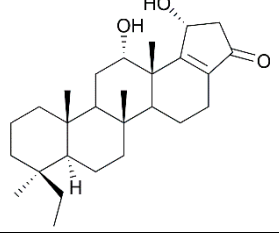
PC-5	(+)-astallatene		<i>Arabidopsis thaliana</i> / <i>Escherichia coli</i> ⁶²³ <i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁶
PC-6	(+)-boleracene		<i>Nicotiana benthamiana</i> / <i>Agrobacterium tumefaciens</i> ²⁶⁶
PC-7	retigeranic acid A		<i>Lobaria isidiosa</i> var. <i>subsidiosa</i> ^{624, 625} <i>Lobaria retigera</i> ⁶²⁶ <i>Lobaria subretigera</i> ⁶²⁶
PC-8	astellatol		<i>Aspergillus variegatus</i> (syn. <i>A. stellatus</i>) ⁶²⁷
PC-9	(+)-astellatene		<i>Nicotiana benthamiana</i> / <i>Arabidopsis thaliana</i> ²⁶⁶
PC-10	12 α -acetoxy-23,25-cyclo-16 β ,25Z-dihydroxy-20,24-dimethyl-oxoscalarane		<i>Carteriospongia foliascens</i> ⁴⁹⁵
PC-11	carteriofenone J		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁴⁷⁶
PC-12	phyllofenone B		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁵⁵⁷

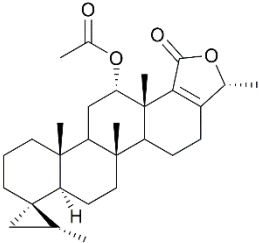
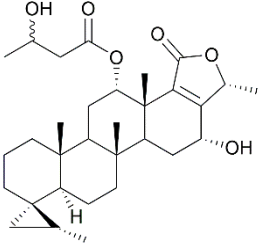
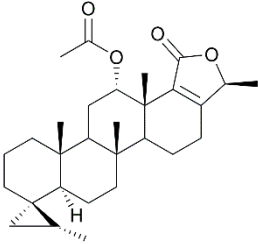
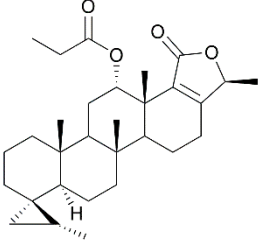
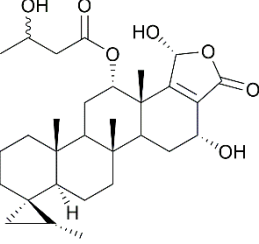
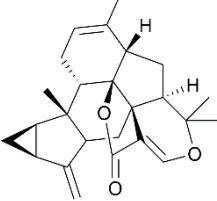
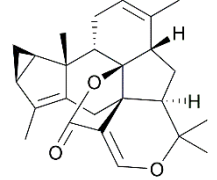
PC-13	phyllofenone D		<i>Phyllospongia foliascens</i> ⁵¹⁸
PC-14	12 β ,22-dihydroxy-24-oxo-24-homoscalara-16,25(26)-diene		<i>Carteriospongia</i> sp. ⁵¹²
PC-15	scalalactam C		<i>Spongia</i> sp. ⁵⁸²
PC-16	scalalactam D		<i>Spongia</i> sp. ⁵⁸²
PC-17	unnamed pentacarboxylic sesterterpene (reported as compound 2)		<i>Dysidea granulosa</i> ³⁷⁰
PC-18	12 α -acetoxy-13 β ,18 β -cyclobutane-20,24-dimethyl-24-oxoscalar-16-en-25 α -ol		<i>Phyllospongia papyracea</i> ⁴⁸⁹
PC-19	12 α -acetoxy-13 β ,18 β -cyclobutane-20,24-dimethyl-24-oxoscalar-16-en-25 β -ol		<i>Phyllospongia papyracea</i> ⁴⁸⁹ <i>Phyllospongia lamellosa</i> ⁵¹⁷

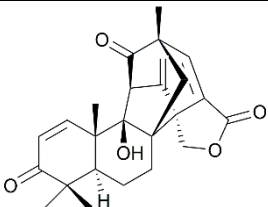
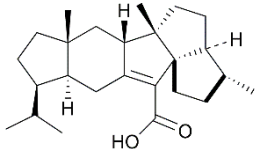
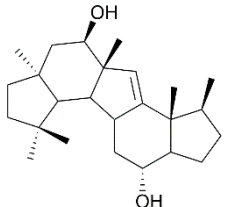
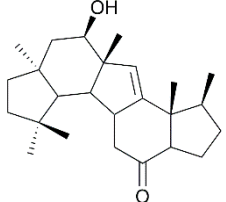
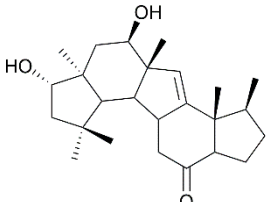
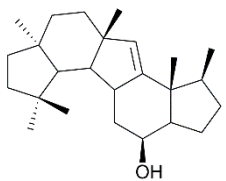
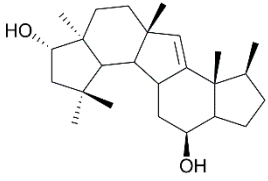
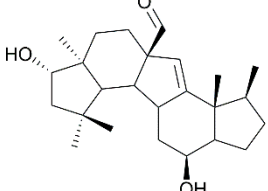
PC-20	carteriofenone E		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁴⁷⁶
PC-21	carteriofenone F		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁴⁷⁶
PC-22	carteriofenone G		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁴⁷⁶
PC-23	carteriofenone H		<i>Carteriospongia</i> (syn. <i>Phyllospongia</i>) <i>foliascens</i> ⁴⁷⁶
PC-24	phyllospongine D		<i>Phyllospongia lamellosa</i> ⁵¹⁷
PC-25	phyllospongine E		<i>Phyllospongia lamellosa</i> ⁵¹⁷

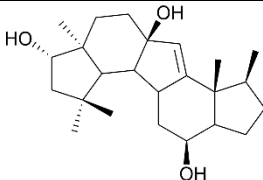
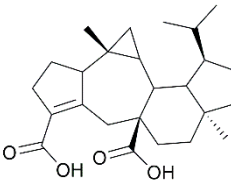
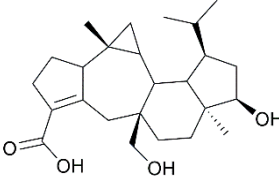
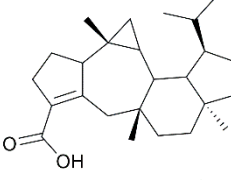
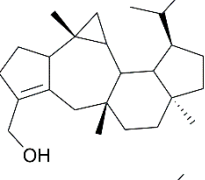
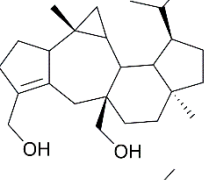
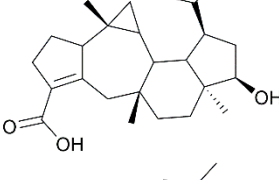
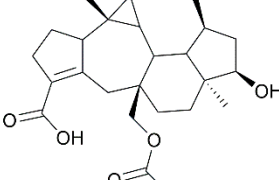
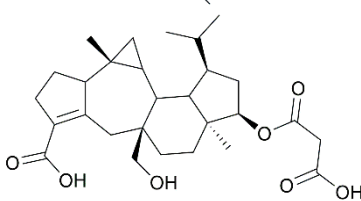
PC-26	isoartemisolid		<i>Artemisia argyi</i> ⁶²⁸
PC-27	toxistylide-A		<i>Microciona toxistyla</i> ⁶²⁹
PC-28	toxistylide-B		<i>Microciona toxistyla</i> ⁶²⁹
PC-29	molliorin-d		<i>Cacospongia mollior</i> ⁵⁸⁹
PC-30	unnamed pentacyclic sesterterpene (reported as compound 4)		<i>Dysidea granulosa</i> ³⁷⁰
PC-31	asperterpenoid A		<i>Aspergillus</i> sp. 16-5c ⁶³⁰
PC-32	retigeran-11-ol		<i>Leprocaulon microscopicum</i> ⁶³¹

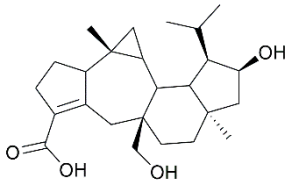
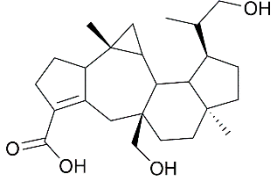
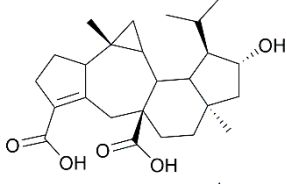
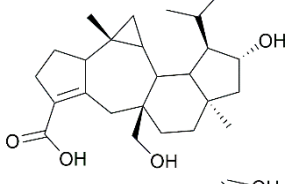
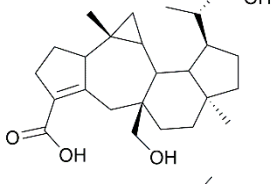
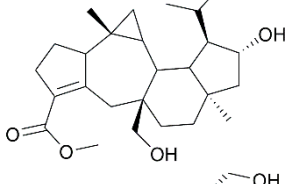
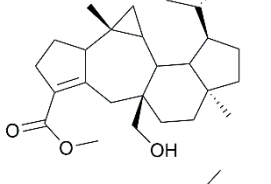
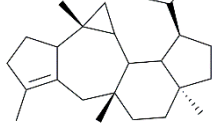
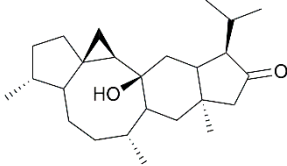
PC-33	4-hydroxyretigeran-11-ol		<i>Leprocaulon microscopicum</i> ⁶³¹
PC-34	gypmacrophin A		<i>Gypsoplaca macrophylla</i> ⁶³²
PC-35	peniroquesine A		<i>Penicillium roqueforti</i> ⁶³³
PC-36	peniroquesine B		<i>Penicillium roqueforti</i> ⁶³³
PC-37	peniroquesine C		<i>Penicillium roqueforti</i> ⁶³³
PC-38	aspterpenacid A		<i>Aspergillus terreus</i> H010/ <i>Kandelia obovata</i> ⁶³⁴
PC-39	aspterpenacid B		<i>Aspergillus terreus</i> H010/ <i>Kandelia obovata</i> ⁶³⁴
PC-40	aspterpenacid C		<i>Swertia bimaculata</i> ⁶³⁵
PC-41	asperunguisin B		<i>Aspergillus unguis</i> ⁴⁶⁶

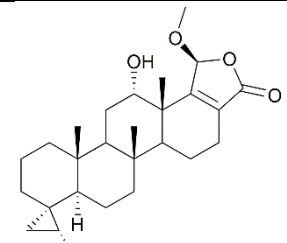
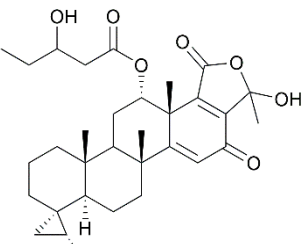
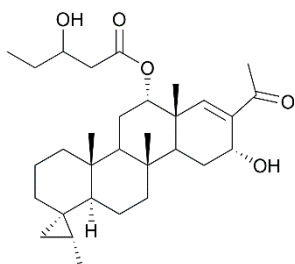
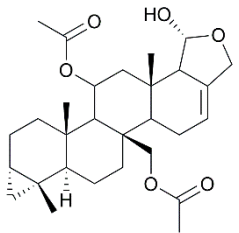
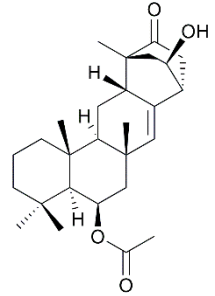
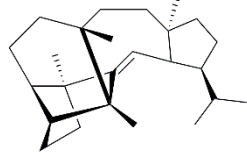
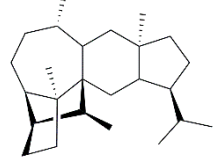
PC-42	honulactone A		<i>Strepsichordaia aliena</i> ⁵⁶⁰
PC-43	honulactone B		<i>Strepsichordaia aliena</i> ⁵⁶⁰
PC-44	honulactone E		<i>Strepsichordaia aliena</i> ⁵⁶⁰
PC-45	honulactone F		<i>Strepsichordaia aliena</i> ⁵⁶⁰
PC-46	honulactone G		<i>Strepsichordaia aliena</i> ⁵⁶⁰
PC-47	unnamed pentacycyclic sesterterpene (reported as compound 8)		<i>Dysidea granulosa</i> ³⁷⁰

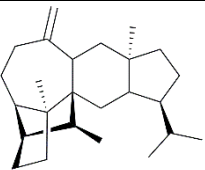
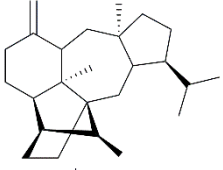
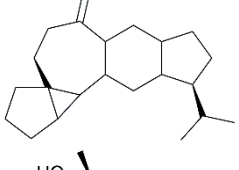
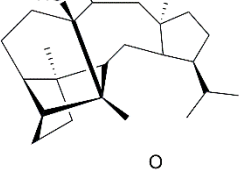
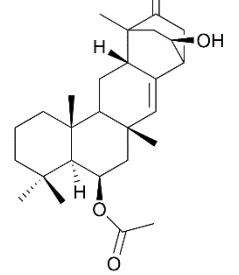
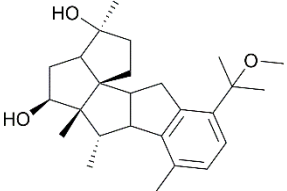
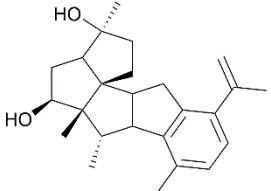
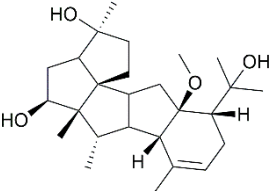
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PC-49	honulactone H		<i>Strepsichordaia aliena</i> ⁵⁶⁰
PC-50	unnamed pentacarbocyclic sesterterpene (reported as compound 11)		<i>Dysidea granulosa</i> ³⁷⁰
PC-51	unnamed pentacarbocyclic sesterterpene (reported as compound 12)		<i>Dysidea granulosa</i> ³⁷⁰
PC-52	unnamed pentacarbocyclic sesterterpene (reported as compound 9)		<i>Dysidea granulosa</i> ³⁷⁰
PC-53	bolivianine		<i>Hedyosmum angustifolium</i> ⁶³⁶
PC-54	isobolivianine		<i>Hedyosmum angustifolium</i> ⁶³⁶

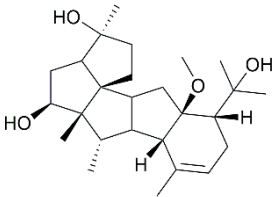
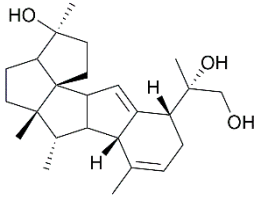
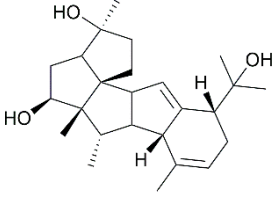
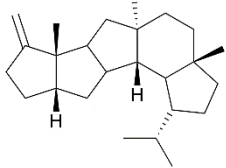
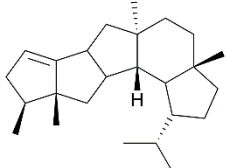
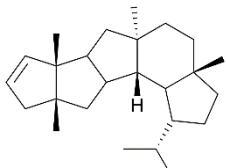
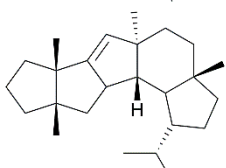
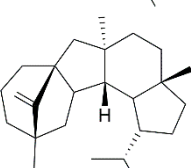
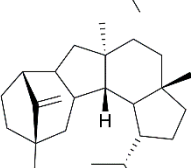
PC-55	emervardione		<i>Emericella variecolor</i> ⁶³⁷
PC-56	retigeranic acid B		<i>Lobaria isidiosa</i> var. <i>subsidiosa</i> ⁶²⁵
PC-57	8- <i>epi</i> -peniroquesine A		<i>Penicillium roqueforti</i> ⁶³⁸
PC-58	8-oxo-eniroquesine B		<i>Penicillium roqueforti</i> ⁶³⁸
PC-59	12-deoxy-16S-hydroxypeniroquesine B		<i>Penicillium roqueforti</i> ⁶³⁸
PC-60	12-deoxypeniroquesine A		<i>Penicillium roqueforti</i> ⁶³⁸
PC-61	12-deoxy-16S-hydroxypeniroquesine A		<i>Penicillium roqueforti</i> ⁶³⁸
PC-62	12-deoxy-22-oxo-16S-hydroxypeniroquesine A		<i>Penicillium roqueforti</i> ⁶³⁸

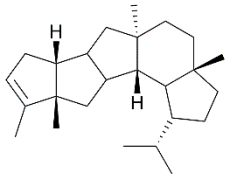
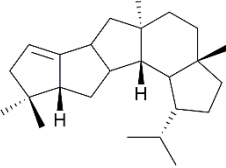
PC-63	8-deoxy-16S-hydroxypeniroquesine A		<i>Penicillium roqueforti</i> ⁶³⁸
PC-64	asperterpenoid B		<i>Aspergillus oryzae</i> ⁶³⁹
PC-65	asperterpenoid C		<i>Aspergillus oryzae</i> ⁶³⁹
PC-66	asperterpenoid D		<i>Aspergillus oryzae</i> ⁶⁴⁰
PC-67	asperterpenoid E		<i>Aspergillus oryzae</i> ⁶⁴⁰
PC-68	asperterpenoid F		<i>Aspergillus oryzae</i> ⁶⁴⁰
PC-69	asperterpenoid G		<i>Aspergillus oryzae</i> ⁶⁴¹
PC-70	asperterpenoid H		<i>Aspergillus oryzae</i> ⁶⁴¹
PC-71	asperterpenoid I		<i>Aspergillus oryzae</i> ⁶⁴¹

PC-72	asperterpenoid J		<i>Aspergillus oryzae</i> ⁶⁴¹
PC-73	asperterpenoid K		<i>Aspergillus oryzae</i> ⁶⁴¹
PC-74	asperterpenoid L		<i>Aspergillus oryzae</i> ⁶⁴¹
PC-75	asperterpenoid M		<i>Aspergillus oryzae</i> ⁶⁴¹
PC-76	asperterpenoid N		<i>Aspergillus oryzae</i> ⁶⁴¹
PC-77	asperterpenoid O		<i>Aspergillus oryzae</i> ⁶⁴¹
PC-78	asperterpenoid P		<i>Aspergillus oryzae</i> ⁶⁴¹
PC-79	preasperterpenoid A		<i>Penicillium verruculosum</i> ⁴³³
PC-80	18- <i>epi</i> -nitidasin		<i>Gentianella turkestanorum</i> ⁴³⁴

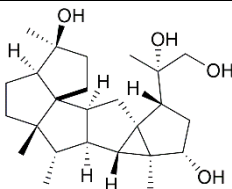
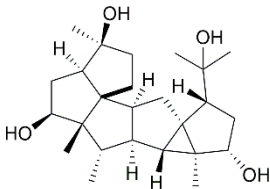
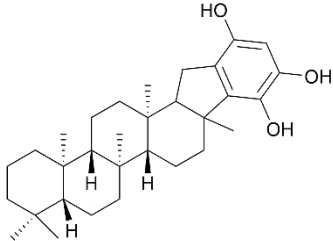
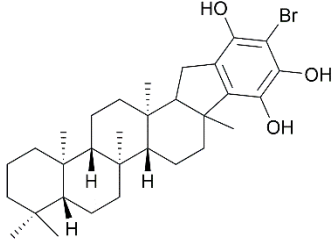
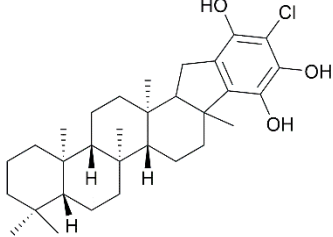
PC-81	dysiscalarone A		<i>Dysidea granulosa</i> ⁶⁰⁶
PC-82	dysiscalarone B		<i>Dysidea granulosa</i> ⁶⁰⁶
PC-83	dysiscalarone C		<i>Dysidea granulosa</i> ⁶⁰⁶
PC-84	12-deacetoxy-4-demethyl-11,24-diacetoxy-3,4-methylenedioxoscalarin		<i>Doriprismatica stellata</i> (egg ribbons), and <i>Spongia</i> cf. <i>agaricina</i> ⁶⁴²
PC-85	neosuberitenone		<i>Suberites</i> sp. ⁴⁴⁷
PC-86	sestermobaraene A		<i>Streptomyces mobaraensis</i> ²⁵⁰
PC-87	sestermobaraene B		<i>Streptomyces mobaraensis</i> ²⁵⁰

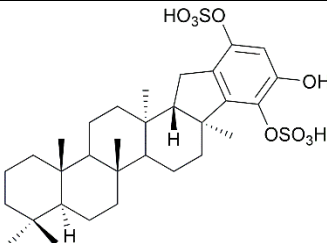
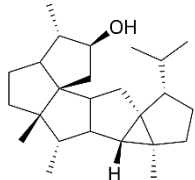
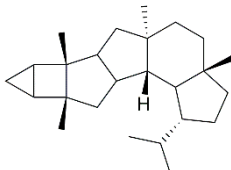
PC-88	sestermobaraene C		<i>Streptomyces mobaraensis</i> ²⁵⁰
PC-89	sestermobaraene D		<i>Streptomyces mobaraensis</i> ²⁵⁰
PC-90	sestermobaraene F		<i>Streptomyces mobaraensis</i> ²⁵⁰
PC-91	sestermobaraol		<i>Streptomyces mobaraensis</i> ²⁵⁰
PC-92	neosuberitenone		<i>Suberites</i> sp. ⁴⁴⁷
PC-93	niduene A		<i>Aspergillus nidulans</i> ⁶⁴³
PC-94	niduene B		<i>Aspergillus nidulans</i> ⁶⁴³
PC-95	niduene C		<i>Aspergillus nidulans</i> ⁶⁴³

PC-96	niduene D		<i>Aspergillus nidulans</i> ⁶⁴³
PC-97	niduene E		<i>Aspergillus nidulans</i> ⁶⁴³
PC-98	niduene F		<i>Aspergillus nidulans</i> ⁶⁴³
PC-99	unnamed pentacyclic sesterterpene		<i>Streptomyces subutilus</i> ⁶⁴⁴
PC-100	unnamed pentacyclic sesterterpene		<i>Streptomyces subutilus</i> ⁶⁴⁴
PC-101	unnamed pentacyclic sesterterpene		<i>Streptomyces subutilus</i> ⁶⁴⁴
PC-102	unnamed pentacyclic sesterterpene		<i>Streptomyces subutilus</i> ⁶⁴⁴
PC-103	unnamed pentacyclic sesterterpene		<i>Streptomyces subutilus</i> ⁶⁴⁴
PC-104	unnamed pentacyclic sesterterpene		<i>Streptomyces subutilus</i> ⁶⁴⁴

PC-105	unnamed pentacyclic sesterterpene		<i>Streptomyces subrutilus</i> ⁶⁴⁴
PC-106	unnamed pentacyclic sesterterpene		<i>Streptomyces subrutilus</i> ⁶⁴⁴

Section 1.10 Structures of hexacarbocyclic (HC) sesterterpenoids (with and without hetherocycles)

c. n.	IUPAC name or Trivial name	Structure	Organism (reference)
HC-1	niduterpenoid A		<i>Aspergillus nidulans</i> ⁶⁴⁵
HC-2	niduterpenoid B		<i>Aspergillus nidulans</i> ⁶⁴⁵
HC-3	disidein		<i>Dysidea pallescens</i> ⁶⁴⁶
HC-4	6'-bromo-disidein		<i>Disidea [sic!] pallescens</i> ⁶⁴⁷
HC-5	6'-cloro-disidein		<i>Disidea [sic!] pallescens</i> ⁶⁴⁷

HC-6	acanthosulfate		<i>Acanthodendrilla</i> sp. ⁶⁴⁸
HC-7	oxaliterpenoid		<i>Penicillium oxalicum</i> M893 ⁶⁴⁹
HC-8	subrutilane		<i>Streptomyces subrutilus</i> ⁶⁴⁴

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Table S2. Accepted name, family, species, synonyms, lifestyle, geographical distribution, and collection site of the selected Porifera¹

Accepted genus name and protologue	Family	Species	Synonyms	Lifeform / Ecology	Geographical distribution	Collection site (specimens with sesterterpenoids)	Environment
<i>Acanthodendrilla</i> Bergquist, 1995	Dictyodendrillidae Bergquist, 1980	<i>Acanthodendrilla</i> sp. 1		Benthic sessile aquatic animal holobiont	Pacific	Indonesia	Marine
		<i>Acanthodendrilla</i> sp. 2		Benthic sessile aquatic animal holobiont	Pacific	Philippines	Marine
<i>Agelas</i> Duchassaing & Michelotti, 1864	Agelasidae Verrill, 1907	<i>Agelas mauritiana</i> (Carter, 1883)	Orig. name <i>Ectyon mauritanus</i> Carter, 1883; Synonymised names: <i>Agelas mauritanus</i> (Carter, 1883) (agreement in gender); <i>Ectyon mauritanus</i> Carter, 1883 (genus transfer);	Benthic sessile aquatic animal holobiont	Indic, South Pacific, South Atlantic	Okinawa, Japan	Marine
<i>Aplysinopsis</i> Lendenfeld, 1888	Thorectidae Bergquist, 1978	<i>Aplysinopsis elegans</i> Lendenfeld, 1888	<i>Aplysinopsis digitata</i> Lendenfeld, 1888; <i>Aplysinopsis pedunculata</i> Lendenfeld, 1889; <i>Thorecta elegans</i> Lendenfeld, 1888	Benthic sessile aquatic animal holobiont	Indo-Pacific: Australia, Great Barrier Reef (GBR), Manning-Hawkesbury; Sea of Japan	Sea of Japan	Marine, coral reefs (tropical, subtropical)
		<i>Aplysinopsis</i> sp.		Benthic sessile aquatic animal holobiont	Indo-Pacific	Fiji	Marine
<i>Brachiaster</i> Wilson, 1925	Pachastrellidae Carter, 1875	<i>Brachiaster</i> sp.		Benthic sessile aquatic animal holobiont	Pacific: Philippines; New Zealand; Thailand.	Thailand	Marine, tropical, subtropical
<i>Cacospongia</i> Schmidt, 1862	Thorectidae Bergquist, 1978	<i>Cacospongia mollior</i> Schmidt, 1862	<i>Aplysina massa</i> (Szymanski, 1904); <i>Aplysinopsis massa</i> Szymanski, 1904; <i>Spongia irregularis</i> var. <i>mollior</i> Schmidt, 1862	Benthic sessile aquatic animal holobiont	Mediterranean: cosmopolitan; Atlantic: Canary, Madeira, Azores Islands, Indian: South India and Sri Lanka (inaccurate)	Mediterranean	Marine, template, cave-dwelling, coralligenous, rocky/detritic/muddy bottom, <i>Posidonia oceanica</i> meadow, lagoons, epibiotic on <i>Pinna nobilis</i>

<i>Clathria</i> Schmidt, 1862	Microcionidae Carter, 1875	<i>Cacospongia</i> sp. (CMB-03404)		Benthic sessile aquatic animal holobiont	Indo-Pacific	Australia (Deep sea)	Marine, deep sea
		<i>Cacospongia</i> sp. 1		Benthic sessile aquatic animal holobiont	South Pacific	New Zealand	Marine
		<i>Cacospongia</i> sp. 2		Benthic sessile aquatic animal holobiont	Mediterranean	Italy/Spain	Marine
		<i>Cacospongia</i> sp. 3		Benthic sessile aquatic animal holobiont	Pacific	Phillipines	Marine
		<i>Clathria (Clathria) gombawuiensis</i> Kim & Sim, 2005		Benthic sessile aquatic animal holobiont	South Korea	Korea	Marine, vertical surfaces of rocks or wrecks, encrusting pebbles in areas of strong current
		<i>Clathria (Clathria) toxistyla</i> (Sarà, 1959)	<i>Clathria toxistyla</i> (Sarà, 1959); <i>Microciona toxistyla</i> Sarà, 1959	Benthic sessile aquatic animal holobiont	Mediterranean; North Atlantic	Mediterranean	Marine, vertical surfaces of rocks or wrecks, encrusting pebbles in areas of strong current
		<i>Clathria</i> sp.		Benthic sessile aquatic animal holobiont	Pacific	Great Australian Bight (Australia)	Marine, vertical surfaces of rocks or wrecks, encrusting pebbles in areas of strong current
<i>Collosporgia</i> Bergquist, Cambie & Kernan, 1990	Thorectidae Bergquist, 1978	<i>Collosporgia auris</i> Bergquist, Cambie & Kernan, 1990	<i>Euspongia mathewsi</i> Lendenfeld, 1886	Benthic sessile aquatic animal holobiont	Australia: GBR, Tween-Moreton	GBR (Australia)	Marine, tropical, reef slopes, often grown by aquarists
<i>Coscinoderma</i> Carter, 1883	Spongiidae Gray, 1867	<i>Coscinoderma mathewsi</i> (Lendenfeld, 1886)		Benthic sessile aquatic animal holobiont	Pacific: Micronesia, New Caledonia, East Caroline Islands, Eastern Australia	Mooloolaba, S.E. Queensland (Australia)	Marine, tropical, lagoons, sandy bottoms
		<i>Coscinoderma</i> sp. 1		Benthic sessile aquatic animal holobiont	South Pacific	Great Barrier Reef, Australia	Marine, coral reefs (tropical, subtropical)
		<i>Coscinoderma</i> sp. 2		Benthic sessile aquatic animal holobiont	South Pacific	Chuuk Island, Micronesia.	Marine

<i>Dactylospongia</i> Bergquist, 1965	Thorectidae Bergquist, 1978		<i>Coscinoderma</i> sp. 3	Benthic sessile aquatic animal holobiont	Indo-Pacific	Chuuk Island, Micronesia	Marine
			<i>Coscinoderma</i> sp. 4	Benthic sessile aquatic animal holobiont	Indo-Pacific	Chuuk State, Micronesia.	Marine
		<i>Dactylospongia</i> <i>elegans</i> (Thiele, 1899)	<i>Aplysinopsis thielei</i> Topsent, 1934; <i>Luffariella elegans</i> Thiele, 1899	Benthic sessile aquatic animal holobiont	Indo-Pacific	Pacific: Banda Sea, East Caroline Islands, Malacca Strait, Philippines, Sulawesi Sea, Makassar Strait Okinawa (Japan)	Marine, tropical, reef, lagoon
		<i>Dactylospongia</i> <i>metachromia</i> (de Laubenfels, 1954)	<i>Hippospongia metachromia</i> de Laubenfels, 1954	Benthic sessile aquatic animal holobiont	Pacific: Solomon Islands, West Caroline Islands; Palau; Koror, NW of Koror Island, Lagoon near Ngarebgal Islet; Society Islands, French Polynesia		Marine, reef, rocks, crevices, lagoons
		<i>Dactylospongia</i> sp.		Benthic sessile aquatic animal holobiont	Pacific: New Caledonia	New Caledonia	Marine, reef, rocks, crevices
<i>Darwinella</i> Müller, 1865	Darwinellidae Merejkowsky, 1879	<i>Darwinella</i> <i>australiensis</i> Carter, 1885		Benthic sessile aquatic animal holobiont	Indo-West Pacific: Australia and Indonesia	Australia	Marine
<i>Dercitus (Stoebea)</i> Sollas, 1888	Ancorinidae Schmidt, 1870	<i>Dercitus (Stoebea)</i> <i>extensus</i> (Dendy, 1905)	<i>Calcabrina</i> Sollas, 1888; <i>Dercitancorina</i> Topsent, 1902; <i>Stoebea</i> Sollas, 1888; <i>Halina extensa</i> (Dendy, 1905); <i>Stoebea extensa</i> (Dendy, 1905)	Benthic sessile aquatic animal holobiont	Indian Ocean: Madagascar, South India, Sri Lanka	Japan (deep-sea)	Marine, encrusting calcareous algae,
<i>Diacarnus</i> Burton, 1934	Podospongiidae Laubenfels, 1936	<i>Diacarnus</i> <i>erythraeanus</i> Kelly- Borges & Vacelet, 1995		Benthic sessile aquatic animal holobiont	Red Sea	Red Sea	Marine, on coral substrate or rocks, or attached to small corals fragments in sand

		<i>Diacarnus laevis</i> (Lindgren, 1897)	<i>Sigmosceptrella laevis</i> (Lindgren, 1897); <i>Latrunculia laevis</i> Lindgren, 1897	Benthic sessile aquatic animal holobiont	Indo-Pacific: Seychelles, Bismarck Sea	Papua New Guinea	Marine, reefs, rocks
		<i>Diacarnus levii</i> Kelly-Borges & Vacelet, 1995		Benthic sessile aquatic animal holobiont	Pacific: New Caledonia, northeastern GBR Australia)	Papua New Guinea	Marine, fringing coral reef slopes down to 35m
		<i>Diacarnus megaspinorhabdosa</i> [sic!] Kelly-Borges & Vacelet, 1995		Benthic sessile aquatic animal holobiont	Pacific: Northern (Madang), southern (Motupore Island) Papua New Guinea; Batangas, Philippines		Marine, silty, fringing coral reefs down to 10m
		<i>Diacarnus spinipoculum</i> (Carter, 1879)	Orig. Name: <i>Axos spinipoculum</i> Carter, 1879; Synonymised names: <i>Axos spinipoculum</i> Carter, 1879 (genus transfer); <i>Latrunculia</i> (<i>Latrunculia</i>) <i>hallmanni</i> Wiedenmayer, 1989; <i>Latrunculia hallmanni</i> Wiedenmayer, 1989; <i>Latrunculia spinipoculum</i> (Carter, 1879)	Benthic sessile aquatic animal holobiont	Pacific	Solomon Islands	Marine
<i>Dictyodendrilla</i> Bergquist, 1980	Dictyodendrillidae Bergquist, 1980	<i>Dictyodendrilla</i> sp.		Benthic sessile aquatic animal holobiont	Pacific	Port Phillip Bay, Victoria (Australia)	Marine
<i>Dysidea</i> Johnston, 1842	Dysideidae Gray, 1867	<i>Dysidea cinerea</i> Keller, 1889	<i>Spongelia cinerea</i> (Keller, 1889)	Benthic sessile aquatic animal holobiont	Red Sea; Indian: Madagascar, East African coast, Western India; Pacific: South China Sea	Dahlak archipelago (Eritrea)	Marine, tropical reefs
		<i>Dysidea etheria</i> Laubenfels, 1936		Benthic sessile aquatic animal holobiont	Atlantic: South America East coast, Brazil, Guianan, Caribbean, Florida (USA), Bermuda	Caribbean	Marine, unconsolidated sediment, attached to algae or blades of turtle grass, rocky bottom, or over hard corals or other sponges

<i>Dysidea fragilis</i> (Montagu, 1814)	<i>Duseideia fragilis</i> (Montagu, 1814); <i>Dysidea coriacea</i> Bowerbank, 1874; <i>Spongelia fistularis</i> Schmidt, 1864; <i>Spongelia fragilis</i> (Montagu, 1814); <i>Spongelia fragilis</i> var. <i>irregularis</i> Lendenfeld, 1889; <i>Spongelia pallescens fragilis</i> (Montagu, 1814); <i>Spongia fragilis</i> Montagu, 1814	Benthic sessile aquatic animal holobiont	Mediterranean; Atlantic: Celtic Sea, Azores, Brazil, Bermuda; North Sea; Inaccurate: Caribbean; Black Sea, White Sea Indian: Africa, Maldives, Kerguelen, India		Marine, sandy bottoms, tidal, rocky estuarine areas, cryptic in rock crevices, on stones, shell and gravel, embedded in muddy sand.
<i>Dysidea granulosa</i> Bergquist, 1965		Benthic sessile aquatic animal holobiont	Pacific: West Caroline Islands, Palau, China, GBR (Australia)	South China Sea	Marine, tropical, reefs
<i>Dysidea gumminea</i> Ridley, 1884	<i>Dysidea gumminea</i> Ridley, 1884, <i>Dysideopsis gumminea</i> (Ridley, 1884); <i>Hyrrios gumminae</i> [sic!], in Mahidol et al., 2009]	Benthic sessile aquatic animal holobiont	Indian: Seychelles, African Coral Sea	Similan Island, Andaman Sea (Thailand)	Marine, tropical reefs, rocks
<i>Dysidea pallescens</i> (Schmidt, 1862)	<i>Dysidea avara</i> var. <i>pallescens</i> (Schmidt, 1862); <i>Dysidea pallescens</i> var. <i>pontica</i> (Czerniavsky, 1880); <i>Spongelia elastica</i> Schulze, 1879; <i>Spongelia elastica</i> var. <i>lobosa</i> Schulze, 1879; <i>Spongelia elastica</i> var. <i>massa</i> Schulze, 1879; <i>Spongelia pallescens</i> Schmidt, 1862; <i>Spongelia pallescens elastica</i> Schulze, 1879; <i>Spongelia pallescens elastica</i> var. <i>lobosa</i> Schulze, 1879; <i>Spongelia pallescens elastica</i> var. <i>massa</i> Schulze, 1879; <i>Spongelia pallescens</i> f. <i>pontica</i> Czerniavsky, 1880; <i>Spongelia pallescens</i> var. <i>lobosa</i> Schulze, 1879;	Benthic sessile aquatic animal holobiont	Mediterranean; Atlantic: Macaronesia, Cape Verde, Sahelian Upwelling, South European Atlantic Shelf	Mediterranean	Marine, cryptic in rock crevices, on stones

			<i>Spongelia putrescens</i> Nardo, 1834				
		<i>Dysidea</i> sp. 1		Benthic sessile aquatic animal holobiont	Pacific	Palau	Marine
		<i>Dysidea</i> sp. 2		Benthic sessile aquatic animal holobiont	Indian, South Pacific	Chuuk Islands, Micronesia	Marine
		<i>Dysidea</i> sp. 3		Benthic sessile aquatic animal holobiont	Pacific	Hainan, China	Marine
<i>Fascaplysinopsis</i> Bergquist, 1980	Thorectidae Bergquist, 1978	<i>Fascaplysinopsis reticulata</i> (Hentschel, 1912)	<i>Aplysinopsis reticulata</i> Hentschel, 1912	Benthic sessile aquatic animal holobiont	Pacific: Arafura Sea, Banda Sea, Coral Sea, New Caledonia, Palau; Indian: East Africa; Red Sea: Pacific: China Sea Pacific	Benga Lagoon (Fiji Islands)	Marine, shallow-water; on dead coral substrates and in mangrove communities on pneumatophores of <i>Avicennia marina</i> .
		<i>Fascaplysinopsis</i> sp.		Benthic sessile aquatic animal holobiont		Palau	Marine
<i>Fasciospongia</i> Burton, 1934	Thorectidae Bergquist, 1978	<i>Fasciospongia cavernosa</i> (Schmidt, 1862)	<i>Cacospongia aspergillum</i> Schmidt, 1868; <i>Cacospongia cavernosa</i> Schmidt, 1862; <i>Fasciospongia aspergillum</i> (Schmidt, 1868); <i>Stelospongia aspergillum</i> (Schmidt, 1868); <i>Stelospongia cavernosa</i> (Schmidt, 1862); <i>Stelospongia cavernosa</i> var. <i>mediterranea</i> Lendenfeld, 1889	Benthic sessile aquatic animal holobiont	Mediterranean; Atlantic: South European Atlantic Shelf, Macaronesia, Madeira, Canary Islands, Azores; Red Sea; Indian: Delagoa, Seychelles, Bismarck Sea	Tyrrhenian	Marine, cave, coralligenous community, rocky/detritic/muddy bottom, <i>Posidonia oceanica</i> meadow, burrowing behaviour, 1–367 m.
		<i>Fasciospongia turgida</i> (Lamarck, 1814)	<i>Fasciospongia penicillosa</i> (Lamarck, 1814); <i>Polyfibrospongia australis</i> (Lendenfeld, 1888); <i>Polyfibrospongia australis</i> var. <i>conulata</i> (Lendenfeld, 1888); <i>Spongia penicillosa</i> Lamarck, 1814; <i>Spongia penicillosa</i> var.	Benthic sessile aquatic animal holobiont	Indo-Pacific: Australia, Great Barrier Reef (GBR), Manning-Hawkesbury; New Zealand	Australia	Marine, cryptic in rock crevices, on stones

			<i>brevior</i> Lamarck, 1814; <i>Spongia penicillosa</i> var. <i>clavata</i> Lamarck, 1814; <i>Spongia turgida</i> Lamarck, 1814; <i>Stelospongia australis</i> Lendenfeld, 1888; <i>Stelospongia australis</i> var. <i>canaliculata</i> Lendenfeld, 1888; <i>Stelospongia australis</i> var. <i>conulata</i> Lendenfeld, 1888; <i>Stelospongia australis</i> var. <i>conulissima</i> Lendenfeld, 1888; <i>Stelospongia australis</i> var. <i>fovea</i> Lendenfeld, 1888; <i>Stelospongia australis</i> var. <i>villosa</i> Lendenfeld, 1888; <i>Stelospongia crassa</i> Lendenfeld, 1889; <i>Stelospongia vallata</i> Lendenfeld, 1889; <i>Stelospongos levis</i> Hyatt, 1877				
		<i>Fasciospongia</i> sp. 1		Benthic sessile aquatic animal holobiont	South Pacific	Palau	Marine
		<i>Fasciospongia</i> sp. 2					
		<i>Fasciospongia</i> sp. 3			Indian Pacific	South Africa New Caledonia	Marine
<i>Geodia</i> Lamarck, 1815	Geodiidae Gray, 1867	<i>Geodia japonica</i> (Sollas, 1888)	<i>Cydonium japonicum</i> Sollas, 1888	Benthic sessile aquatic animal holobiont	Pacific: Alaska, Japan, Philippines, China Sea	China Sea	Marine, encrusting on rocks, stones and gravel, with aggregating coral rubble and shells
<i>Halichondria</i> Fleming, 1828	Halichondriidae Gray, 1867	Halichondriidae sp.		Benthic sessile aquatic animal holobiont	Pacific: USA Western coast	Scripps Canyon California (USA)	Marine
		<i>Halichondria</i> sp.		Benthic sessile aquatic animal holobiont	South Pacific	Palau	Marine

<i>Haliclona (Reniera)</i> Schmidt, 1862	Chalinidae Gray, 1867	<i>Haliclona (Reniera)</i> sp. 1	<i>Haliclona (Reniclona)</i> de Laubenfels, 1954; <i>Kallypilidion</i> de Laubenfels, 1954; <i>Philotia</i> Gray, 1867; <i>Prianos</i> Gray, 1867; <i>Reniclona</i> de Laubenfels, 1954; <i>Reniera</i> Schmidt, 1862; <i>Toxadocia</i> Laubenfels, 1936	Benthic sessile aquatic animal holobiont	Pacific: Korea	Gageo Island (Korea)	Marine, coral reefs
		<i>Haliclona (Reniera)</i> sp. 2		Benthic sessile aquatic animal holobiont	Red Sea	Marsa el Muqubila in the Gulf of Eilat, Israel (Red Sea)	Marine
<i>Hamigera</i> Gray, 1867	Hymedesmiidae Topsent, 1928	<i>Hamigera</i> sp.		Benthic sessile aquatic animal holobiont	Indo-Pacific: Papua New Guinea	Milne Bay, Papua New Guinea	Marine, tropical reefs, rocks
<i>Hippospongia</i> Schulze, 1879	Spongiidae Gray, 1867	<i>Hippospongia</i> <i>lachne</i> (Laubenfels, 1936)	<i>Hippiospongia lachne</i> Laubenfels, 1936; sheep's wool sponge	Benthic sessile aquatic animal holobiont	Atlantic Brazil, Gulf of Mexico, Bahamas, Antilles, Florida; Caribbean	South China Sea	Marine, reef, rocks, sandy bottoms, rubble, lagoons
		<i>Hippospongia</i> <i>communis</i> (Lamarck, 1814)	<i>Spongia communis</i> Lamarck, 1814	Benthic sessile aquatic animal holobiont	Mediterranean	Italy, Mediterranean	Marine
		<i>Hippospongia</i> <i>fistulosa</i> (Lamarck, 1814)	<i>Euspongia officinalis</i> var. <i>cavernosa</i> Ridley, 1884 (genus transfer & junior homonym); <i>Hyattella fistulosa</i> (Lendenfeld, 1889) unaccepted (reverted genus transfer)	Benthic sessile aquatic animal holobiont	Pacific: Australia, Borneo, Vietnam	Vietnam	Marine
		<i>Hippospongia</i> sp. 1		Benthic sessile aquatic animal holobiont	Pacific	Taiwan	Marine, coral reef
		<i>Hippospongia</i> sp. 2		Benthic sessile aquatic animal holobiont	Indo-Pacific	Barangcadi Island, Ujung Pandang, Sulawesi, Indonesia	Marine
		<i>Hippospongia</i> sp. 3		Benthic sessile aquatic animal holobiont	Pacific	Australia	Marine reefs, rocks, dead coral, living coral

<i>Hyattella</i> Lendenfeld, 1888	Spongiidae Gray, 1867	<i>Hippospongia</i> sp. 4		Benthic sessile aquatic animal holobiont	South Pacific	Chuuk Islands, Micronesia	Marine
		<i>Hippospongia</i> sp. 5		Benthic sessile aquatic animal holobiont	South Pacific	Taiwan	Marine
		<i>Hyattella</i> <i>cribriformis</i> (Hyatt, 1877)	<i>Stelospongia cribriformis</i> (Hyatt, 1877); <i>Stelospongos</i> <i>cribriformis</i> Hyatt, 1877	Benthic sessile aquatic animal holobiont. Prey of heterobranch <i>Glossodoris</i> <i>atromarginata</i> , that accumulates diet sesterterpenoids.	Indian: Maldives, Seychelles, Sri Lanka, South India	India	Marine, reef, rocks, sandy bottoms, rubble, lagoons
		<i>Hyattella</i> <i>intestinalis</i> (Lamarck, 1814)	<i>Carteriospongia clathrata</i> (Carter, 1881); <i>Carterispongia</i> <i>clathrata</i> (Carter, 1881); <i>Hippospongia anomala</i> Poléjaeff, 1884; <i>Hippospongia</i> <i>clathrata</i> (Carter, 1881); <i>Hippospongia intestinalis</i> (Lamarck, 1814); <i>Hircinia</i> <i>clathrata</i> Carter, 1881; <i>Hyattella clathrata</i> (Carter, 1881); <i>Hyattella murrayi</i> Lendenfeld, 1889; <i>Hyattella</i> <i>velata</i> (Hyatt, 1877); <i>Ircinia</i> <i>clathrata</i> (Carter, 1881); <i>Spongelia velata</i> Hyatt, 1877; <i>Spongia cariosa</i> Lamarck, 1814; <i>Spongia intestinalis</i> Lamarck, 1814; <i>Spongia</i> <i>tubulosa</i> Lamarck, 1814; <i>Stelospongia kingii</i> Lendenfeld, 1889	Benthic sessile aquatic animal holobiont	Indo-Pacific: Australia, New Zealand, Solomon, Seychelles, India, Indonesia; Caribbean: Antilles, Bahamas	Gulf of California (Mexico); Australia; Mooloolaba; Southeast Queensland (Australia)	Marine, reef, rocks, sandy bottoms, rubble, lagoons

Hyrtios Duchassaing & Michelotti, 1864	Thorectidae Bergquist, 1978	<i>Hyattella</i> sp. 1		Benthic sessile aquatic animal holobiont	Pacific	Soheuksan-do (Korea)	Marine, reefs, rocks
		<i>Hyattella</i> sp. 2			Indo-Pacific	Lembah Strait, North Sulawesi, Indonesia	
		<i>Hyrtios communis</i> (Carter, 1885)		Benthic sessile aquatic animal holobiont	Indo-Pacific: Bassian, East Caroline Islands; Red Sea	Palau	Marine, on coral substrate or rocks, or attached to small corals, fragments in sand
		<i>Hyrtios erectus</i> (Keller, 1889)	<i>Duriella nigra</i> Row, 1911; <i>Dysidea fusca</i> Ridley, 1884, <i>Dysidea nigra</i> Keller, 1889; <i>Dysideopsis fusca</i> (Ridley, 1884); <i>Heteronema erecta</i> Keller, 1889; <i>Heteronema erectum</i> Keller, 1889; <i>Hyrtios erecta</i> (Keller, 1889); <i>Hyrtios fusca</i> (Ridley, 1884); <i>Inodes erecta</i> (Keller, 1889); <i>Thorectopsamma irregularis</i> Burton, 1934; <i>Thorectopsamma mela</i> de Laubenfels, 1954	Benthic sessile aquatic animal holobiont	Indo-Pacific: India, Sri Lanka, Caroline Islands, Seychelles, Banda Sea, Bismarck Sea, Australia, Indonesia; Red Sea	Australia; Hainan, China; Saudi Arabia; Tonga; Okinawa, Japan; Egypt; Papua, New Guinea; Republic of Maldives; New Caledonia; Bohol Island, Philippines	Marine, on coral substrate or rocks, or attached to small corals, fragments in sand
		<i>Hyrtios cf. erectus</i> (Keller, 1889)		Benthic sessile aquatic animal holobiont	Pacific	Fiji	Marine
		<i>Hyrtios cf. erectus</i> (Keller, 1889)		Benthic sessile aquatic animal holobiont	Pacific	Yakushima Island, Japan	Marine
		<i>Hyrtios</i> sp.1		Benthic sessile aquatic animal holobiont	South Pacific	New Caledonia	Marine
		<i>Hyrtios</i> sp.2		Benthic sessile aquatic animal holobiont	Indo-Pacific	Thailand	Marine

		<i>Hyrtios</i> sp.3		Benthic sessile aquatic animal holobiont	Indo-Pacific	Paracel Islands, Vietnam	Marine
		<i>Hyrtios</i> sp.4		Benthic sessile aquatic animal holobiont	Pacific	Fiji	Marine
<i>Igernella</i> Topsent, 1905	Dictyodendrillidae Bergquist, 1980	<i>Igernella</i> sp.		Benthic sessile aquatic animal holobiont	Pacific	Palau	Marine, tropical reefs
<i>Ircinia</i> Nardo, 1833	Irciniidae Gray, 1867	<i>Ircinia campana</i> (Lamarck, 1814)	Orig. Name: <i>Spongia campana</i> Lamarck, 1814. Synonymised names: <i>Hircinia (Sarcotragus) campana</i> (Lamarck, 1814); <i>Hircinia campana</i> (Lamarck, 1814) (genus name correction); <i>Hircinia campana</i> var. <i>fixa</i> (Duchassaing & Michelotti, 1864) (genus transfer and junior synonym); <i>Hircinia campana</i> var. <i>typica</i> Hyatt, 1877; <i>Polythereses campana</i> (Lamarck, 1814) (genus transfer); <i>Polythereses campana</i> var. <i>dimidiata</i> Duchassaing de Fonbressin, 1870 (nomen nudum, junior synonym and genus transfer); <i>Polythereses campana</i> var. <i>fixa</i> Duchassaing & Michelotti, 1864 (genus transfer and junior synonym); <i>Spongia campana</i> Lamarck, 1814 (genus transfer); <i>Stematumenia scyphus</i> Bowerbank, 1845 (genus transfer and junior synonym)	Benthic sessile aquatic animal holobiont	Atlantic, Caribbean	Caribbean, Colombia	Marine

<i>Ircinia dendroides</i> (Schmidt, 1862)	<i>Euricinia variabilis</i> var. <i>dendroides</i> (Schmidt, 1862); <i>Hircinia</i> (<i>Euricinia</i>) <i>variabilis</i> var. <i>dendroides</i> Schmidt, 1862; <i>Hircinia dendroides</i> Schmidt, 1862; <i>Hircinia variabilis</i> var. <i>dendroides</i> Schmidt, 1862; <i>Ircinia fasciculata</i> var. <i>dendroides</i> (Schmidt, 1862); <i>Ircinia variabilis</i> var. <i>dendroides</i> (Schmidt, 1862)	Benthic sessile aquatic animal holobiont	Mediterranean; Atlantic: Macaronesia, Canary Islads Cape Verde, Azores	Mogan, Grand Canary Islands (Spain)	Marine, cave, detritic and rocky bottom, coralligenous community. Bathymetric range 1–110 m
<i>Ircinia felix</i> (Duchassaing & Michelotti, 1864)	<i>Hircinia armata</i> (Duchassaing & Michelotti, 1864); <i>Hircinia</i> <i>armata</i> var. <i>fistularis</i> Verrill, 1907; <i>Hircinia campana</i> var. <i>felix</i> (Duchassaing & Michelotti, 1864); <i>Hircinia</i> <i>filamenta</i> Hyatt, 1877; <i>Hircinia</i> <i>fistularis</i> Verrill, 1907; <i>Hircinia</i> <i>nigra</i> Hyatt, 1877; <i>Hircinia</i> <i>variabilis</i> sensu Laubenfels, 1936; <i>Ircinia fasciculata</i> sensu de Laubenfels, 1949; <i>Ircinia</i> <i>felix</i> f. <i>acuta</i> Duchassaing & Michelotti, 1864; <i>Ircinia felix</i> f. <i>felix</i> Duchassaing & Michelotti, 1864; <i>Ircinia felix</i> f. <i>fistularis</i> (Verrill, 1907); <i>Ircinia fistularis</i> (Verrill, 1907); <i>Polythereses</i> <i>acuta</i> Duchassaing & Michelotti, 1864; <i>Polythereses</i> <i>armata</i> Duchassaing & Michelotti, 1864; <i>Polythereses</i> <i>columnaris</i> Duchassaing & Michelotti, 1864; <i>Polythereses</i> <i>felix</i> Duchassaing & Michelotti, 1864	Benthic sessile aquatic animal holobiont	Atlantic: South America East coast, Brazil, Guianan, Bahamas; Caribbean: Florida (USA), Bermuda	Southern Taiwan	Marine, shallow to mid- range reefs, near living corals, lagoons

<i>Ircinia oros</i> (Schmidt, 1864)	<i>Hircinia (Euricinia) variabilis</i> var. <i>oros</i> Schmidt, 1864; <i>Hircinia oros</i> Schmidt, 1864; <i>Hircinia variabilis</i> var. <i>oros</i> Schmidt, 1864	Benthic sessile aquatic animal holobiont	Mediterranean; Atlantic: Macaronesia	Naples (Italy), Malta	Marine, cave, detritic and rocky bottom, coralligenous, frequently covered by <i>Haliclona</i> (<i>Reniera</i>) <i>cratera</i> ; Bathymetric range 1–150 m Marine
<i>Ircinia ramosa</i> (Keller, 1889)	<i>Hippospongia frondosa</i> Hentschel, 1912; <i>Hircinia</i> <i>ramosa</i> Keller, 1889; <i>Ircinia</i> (<i>Sarcotragus</i>) <i>ramosa</i> Keller, 1889	Benthic sessile aquatic animal holobiont	Atlantic: Brazil, Gulf of Mexico, Antilles, Bermuda; Caribbean: Bahamas; Indian: Mozambique, Kenya; Pacific: Arafura Sea, Australia	Malaysia	
<i>Ircinia strobilina</i> (Lamarck, 1816)	<i>Dysidicinia longispina</i> (Duchassaing & Michelotti, 1864); <i>Filifera verrucosa</i> Lieberkühn, 1859; <i>Hircinia</i> (<i>Dysidicinia</i>) <i>longispina</i> (Duchassaing & Michelotti, 1864); <i>Hircinia (Psammocinia)</i> <i>verrucosa</i> (Lieberkühn, 1859); <i>Hircinia acuta</i> (Duchassaing & Michelotti, 1864); <i>Hircinia</i> <i>acuta</i> var. <i>filamentosa</i> Hyatt, 1877; <i>Hircinia acuta</i> var. <i>longispina</i> (Duchassaing & Michelotti, 1864); <i>Hircinia</i> <i>acuta</i> var. <i>nigra</i> Hyatt, 1877; <i>Hircinia strobilina</i> (Lamarck, 1814); <i>Hircinia verrucosa</i> (Lieberkhün, 1859); <i>Ircinia</i> <i>acuta</i> (Duchassaing & Michelotti, 1864); <i>Ircinia acuta</i> var. <i>longispina</i> (Duchassaing & Michelotti, 1864); <i>Ircinia</i> <i>linguiformis</i> (Duchassaing &	Benthic sessile aquatic animal holobiont	Atlantic: Brazil, Bermuda; Caribbean: Bahamas, Antilles, Florida, Guianan, Gulf of Mexico, Venezuela; Western Mediterranean (Inaccurate)	Brazil, Caribbean	Marine, reefs, but also on muddy sands and at bottoms at greater depths, prefer brightly lit areas

	Michelotti, 1864); <i>Ircinia longispina</i> (Duchassaing & Michelotti, 1864); <i>Ircinia verrucosa</i> (Lieberkühn, 1869); <i>Polythereses capitata</i> Duchassaing & Michelotti, 1864; <i>Polythereses cylindrica</i> Duchassaing & Michelotti, 1864; <i>Polythereses ignobilis</i> Duchassaing & Michelotti, 1864; <i>Polythereses linguiformis</i> Duchassaing & Michelotti, 1864; <i>Polythereses longispina</i> Duchassaing & Michelotti, 1864; <i>Spongia strobilina</i> Lamarck, 1816; <i>Stelospongos longispinus</i> (Duchassaing & Michelotti, 1864)				
<i>Ircinia variabilis</i> (Schmidt, 1862)	<i>Euricinia variabilis</i> (Schmidt, 1862); <i>Hircinia (Euricinia) variabilis</i> Schmidt, 1862; <i>Hircinia (Euricinia) variabilis</i> var. <i>flavescens</i> Schmidt, 1862; <i>Hircinia (Euricinia) variabilis</i> var. <i>galea</i> Lendenfeld, 1889; <i>Hircinia (Euricinia) variabilis</i> var. <i>hirsuta</i> (Schmidt, 1862); <i>Hircinia (Euricinia) variabilis</i> var. <i>lingua</i> Schmidt, 1868; <i>Hircinia (Euricinia) variabilis</i> var. <i>mammillaris</i> Schmidt, 1868; <i>Hircinia (Euricinia) variabilis</i> var. <i>typica</i> Nardo, 1847; <i>Hircinia flavescens</i> Schmidt, 1862; <i>Hircinia hebes</i> Schmidt, 1862; <i>Hircinia hirsuta</i> Schmidt, 1862; <i>Hircinia lingua</i> Schmidt, 1868; <i>Hircinia</i>	Benthic sessile aquatic animal holobiont	Cosmopolitan; Atlantic: Canary, Madeira, Azores Islands, Gulf of Guinea, Namibia; Black Sea	Mediterranean	Marine, cave, coralligenous community, detritic and rocky bottom, <i>Posidonia oceanica</i> meadow, lagoons, epibiotic on <i>Pinna nobilis</i> . Bathymetric range 0–450 m

	<i>mamillaris</i> Schmidt, 1868; <i>Hircinia panicea</i> Schmidt, 1862; <i>Hircinia typica</i> Nardo, 1847; <i>Hircinia variabilis</i> Schmidt, 1862; <i>Hircinia</i> <i>variabilis</i> var. <i>fistulata</i> Szymanski, 1904; <i>Hircinia</i> <i>variabilis</i> var. <i>galea</i> Lendenfeld, 1889; <i>Hircinia</i> <i>variabilis</i> var. <i>hirsuta</i> Schmidt, 1862; <i>Hircinia variabilis</i> var. <i>mammillaris</i> Schmidt, 1868; <i>Hircinia variabilis</i> var. <i>typica</i> Nardo, 1847; <i>Ircinia fasciculata</i> sensu Vacelet, 1959; <i>Ircinia</i> <i>fasciculata</i> var. <i>variabilis</i> (Schmidt, 1862); <i>Ircinia</i> <i>flavescens</i> (Schmidt, 1862); <i>Ircinia hebes</i> (Schmidt, 1862); <i>Ircinia hirsuta</i> (Schmidt, 1862); <i>Ircinia hospitum</i> Nardo, 1833; <i>Ircinia lingua</i> (Schmidt, 1868); <i>Ircinia panicea</i> (Schmidt, 1862); <i>Ircinia rigida</i> Nardo, 1833; <i>Ircinia spongiastrum</i> Nardo, 1833; <i>Ircinia tenax</i> Nardo, 1833; <i>Ircinia typica</i> (Nardo, 1847); <i>Ircinia variabilis</i> var. <i>typica</i> Nardo, 1847; <i>Spongia cavernosa</i> sensu Nardo, 1833; <i>Spongia</i> <i>licheniformis</i> var. <i>gamma</i> Lamarck, 1814				
<i>Ircinia wistarii</i> Wilkinson, 1978		Benthic sessile aquatic animal holobiont	Indo-Pacific	Heron Island, Wistari Reef, GBR (Australia)	Marine, coral reefs

<i>Ircinia</i> sp. CMB-01064		Benthic sessile aquatic animal holobiont	South Pacific	Durras Island, Australia	Marine
<i>Ircinia</i> sp. CMB-03363		Benthic sessile aquatic animal holobiont	South Pacific	Australia	Marine
<i>Ircinia</i> sp. 1		Benthic sessile aquatic animal holobiont	Atlantic	Canary Islands (Spain) Atlantic	Marine
<i>Ircinia</i> sp. 2		Benthic sessile aquatic animal holobiont	Atlantic	Gulf of Mexico, Atlantic	Marine
<i>Ircinia</i> sp. 3		Benthic sessile aquatic animal holobiont	Red Sea	Hurgada (Egypt) Red Sea	Marine
<i>Ircinia</i> sp. 4		Benthic sessile aquatic animal holobiont	Pacific	Iriomote Island, Okinawa (Japan) Pacific	Marine
<i>Ircinia</i> sp. 5		Benthic sessile aquatic animal holobiont	South Pacific	Australia	Marine
<i>Ircinia</i> sp. 6		Benthic sessile aquatic animal holobiont	Atlantic	Ouakam, Dakar (Senegal)	Marine
<i>Ircinia</i> sp. 7		Benthic sessile aquatic animal holobiont	Pacific	Okinawa, Japan	Marine
<i>Ircinia</i> sp. 8		Benthic sessile aquatic animal holobiont	South Pacific	Pacific Conquest Porpoise Cay, Wreck Reef, Queensland (Australia)	Marine
<i>Ircinia</i> sp. 9	<i>Ircinia formosana</i> Shen, Lo, Lin, Khalil, Kuo & Shih, 2006 (nomen nudum)	Benthic sessile aquatic animal holobiont	Pacific: East China Sea, Taiwan	Taiwan	Marine
<i>Ircinia</i> sp. 10		Benthic sessile aquatic animal holobiont	South Pacific	New Zealand	Marine

				<i>Ircinia</i> sp. 11	Benthic sessile aquatic animal holobiont	Pacific	Fiji	Marine
				<i>Ircinia</i> sp. 12	Benthic sessile aquatic animal holobiont	Atlantic, Caribbean	Palomino Island, Puerto Rico	Marine
				<i>Ircinia</i> sp. 13	Benthic sessile aquatic animal holobiont	Pacific	Okinawa, Japan	Marine
				<i>Ircinia</i> sp. 14	Benthic sessile aquatic animal holobiont	South Pacific	New Zealand	Marine
<i>Jaspis</i> Gray, 1867	Ancorinidae Schmidt, 1870	<i>Jaspis</i> cf. <i>johnstonii</i> (Schmidt, 1862)	<i>Asteropus incrustans</i> Lendenfeld, 1898; <i>Vioa johnstonii</i> Schmidt, 1862; <i>Xenospongia johnstonii</i> (Schmidt, 1862)	Benthic sessile aquatic animal holobiont	Mediterranean: cosmopolitan; Atlantic: Canary, Madeira, Azores Islands, Gulf of Guinea, Sahelian Upwelling, Caribbean	New Guinea	New Guinea	Marine, rocks,
		<i>Jaspis stellifera</i> (Carter, 1879)	<i>Amorphina stellifera</i> Carter, 1879; <i>Coppatias stellifera</i> (Carter, 1879); <i>Jaspis coriacea</i> (Carter, 1886); <i>Stellettinopsis coriacea</i> Carter, 1886	Benthic sessile aquatic animal holobiont	Indo-Pacific: Bassian, Palau, Australia		Okinawa (Japan)	Marine, rocks, shallow lagoons encrusting dead corals, coral rubble and generally available hard substrates
		<i>Jaspis</i> sp.		Benthic sessile aquatic animal holobiont	Pacific		South China	Marine
<i>Lamellodysidea</i> Cook & Bergquist, 2002	Dysideidae Gray, 1867	<i>Lamellodysidea herbacea</i> (Keller, 1889)	<i>Carteriospongia cordifolia</i> Keller, 1889; <i>Dysidea herbacea</i> (Keller, 1889); <i>Dysideopsis palmata</i> Topsent, 1897; <i>Dysideopsis topsenti</i> Hentschel, 1912; <i>Phyllospongia</i> complex de Laubenfels, 1954; <i>Phyllospongia cordifolia</i> (Keller, 1889); <i>Spongelia delicatula</i> Row, 1911;	Benthic sessile aquatic animal holobiont	Indo-Pacific: Australia, Afuran Sea, Coral Sea, Bismarck Sea, Indonesia, New Caledonia, Maldives, Hawai'i, New Caledonia, Singapore, Caroline Islands; Red Sea	Gulf of Suez (Red Sea)		Marine, lagoons, on the inner reef barrier slope or on the pinnacles

<i>Latrunculia</i> du Bocage, 1869	Latrunculiidae Topsent, 1922	<i>Latrunculia</i> (<i>Latrunculia</i>) <i>brevis</i> Ridley & Dendy, 1886	<i>Spongelia herbacea</i> Keller, 1889	Benthic sessile aquatic animal holobiont	Pacific: New Caledonia; Atlantic: Namaqua, Malvinas; Indian: Heard and McDonnald Islands Indo-Pacific	Australia	Marine, rocks
			<i>Latrunculia brevis</i> Ridley & Dendy, 1886				
		<i>Latrunculia</i> sp. 1		Benthic sessile aquatic animal holobiont		Indo-Polynesian	Marine
		<i>Latrunculia</i> sp. 2		Benthic sessile aquatic animal holobiont	Indo-Pacific	Jervis Bay, Australia	Marine
		<i>Latrunculia</i> sp. 3		Benthic sessile aquatic animal holobiont	Indo-Pacific	Australia	Marine
		<i>Latrunculia</i> sp. (C006879)		Benthic sessile aquatic animal holobiont	Indo-Pacific	Indo-Polynesian	Marine
<i>Leiosella</i> Lendenfeld, 1888	Spongiidae Gray, 1867	<i>Leiosella idia</i> (de Laubenfels, 1932)	<i>Spongia</i> (<i>Spongia</i>) <i>idia</i> de Laubenfels, 1932; <i>Spongia idia</i> de Laubenfels, 1932	Benthic sessile aquatic animal holobiont. Is prey of mollusc nudibranch <i>Cadlina luteomarginata</i> , that accumulates sponge-derived sesterterpenes for own defense.	North Pacific: California (USA)	San Diego, California (USA)	Marine, rocks
<i>Lendenfeldia</i> Bergquist, 1980	Thorectidae Bergquist, 1978	<i>Lendenfeldia chondrodes</i> (Laubenfels, 1954)	<i>Fasciospongia chondrodes</i> (de Laubenfels, 1954) unaccepted (genus transfer); <i>Spongionella chondrodes</i> de Laubenfels, 1954 unaccepted (genus transfer)	Benthic sessile aquatic animal holobiont	Indian and Pacific: Malacca Strait, Caroline Islands, Singapore, Micronesia, Japan	Milky Way Lake (Palau), Japan (Okinawa)	Marine, encrusts coral rubble sometimes, also living coral, lake, lagoons

		<i>Lendenfeldia dendyi</i> (Lendenfeld, 1889)	<i>Phyllospongia (Antheroplax) dendyi</i> Lendenfeld, 1889; <i>Phyllospongia dendyi</i> Lendenfeld, 1889	Benthic sessile aquatic animal holobiont	Indo-Pacific: Caroline Islands, Australia, Maldives, East African Coast, Madagascar Pacific	Great Barrier Reef; Nicobar Islands Australia; Andaman Islands, India	Marine, rocks, reef, dead coral on sand
		<i>Lendenfeldia frondosa</i> (Lendenfeld, 1889)	<i>Phyllospongia dendyi</i> var. <i>frondosa</i> Lendenfeld, 1889	Benthic sessile aquatic animal holobiont	Indian	Solomon Islands; Papua New Guinea	Marine, rocks, reef, dead coral on sand
		<i>Lendenfeldia</i> sp. 1		Benthic sessile aquatic animal holobiont		Barren Islands (Madagascar)	Marine
		<i>Lendenfeldia</i> sp. 2		Benthic sessile aquatic animal holobiont		Barren Islands (Madagascar)	Marine
		<i>Lendenfeldia</i> sp. 3		Benthic sessile aquatic animal holobiont	South Pacific	Taiwan	Marine
		<i>Lendenfeldia</i> sp. 4		Benthic sessile aquatic animal holobiont	Pacific	Mariana Islands	Marine
		<i>Lendenfeldia</i> sp. 5		Benthic sessile aquatic animal holobiont	Indo-Pacific	Indonesia	Marine
		<i>Lendenfeldia</i> sp. 6		Benthic sessile aquatic animal holobiont	Indo-Pacific	Australia	Marine
		<i>Luffariella</i> Thiele, 1899	Thorectidae Bergquist, 1978	Benthic sessile aquatic animal holobiont	Pacific: Australia, and Easter Islands, Tuvalu	Australia	Marine, rocks, reef
		<i>Luffariella geometrica</i> Kirkpatrick, 1900		Benthic sessile aquatic animal holobiont	Indo-Pacific: Malacca Strait, Caroline Islands, Singapore, Society Islands, Philippines Indian	GBR (Australia); Sepanggar Island, Sabah (Malaysia); Palau	Marine, rocks, reef
		<i>Luffariella variabilis</i> (Poléjaeff, 1884)		Benthic sessile aquatic animal holobiont		Mayotte Island (Indian Ocean)	
		<i>Luffariella</i> cf. <i>variabilis</i>		Benthic sessile aquatic animal holobiont			

		<i>Luffariella</i> sp. 1		Benthic sessile aquatic animal holobiont	Indo-Pacific	Australia	
		<i>Luffariella</i> sp. 2		Benthic sessile aquatic animal holobiont	Indo-Pacific	Pohnpei (Federated States of Micronesia)	
		<i>Luffariella</i> sp. 3		Benthic sessile aquatic animal holobiont	Pacific	Okinawa, Japan	
		<i>Luffariella</i> sp. 4		Benthic sessile aquatic animal holobiont	Pacific	Taiwan	
		<i>Luffariella</i> sp. 5		Benthic sessile aquatic animal holobiont	Pacific	Palau	
		<i>Luffariella</i> sp. 6		Benthic sessile aquatic animal holobiont	Pacific	Palau	
<i>Monanchora</i> Carter, 1883	Crambeidae Lévi, 1963	<i>Monanchora</i> sp. 1		Benthic sessile aquatic animal holobiont	Pacific	Gageo Island (Korea)	Marine, encrust dead areas of reefs, encrusting form especially around the bases of living coral heads
		<i>Monanchora</i> sp. 2		Benthic sessile aquatic animal holobiont	North Pacific	Aleutian Islands of Alaska, EEUU	Marine
<i>Mycale</i> Gray, 1867	Mycalidae Lundbeck, 1905	<i>Mycale</i> (<i>Arenochalina</i>) <i>laxissima</i> (Duchassaing & Michelotti, 1864)	<i>Acamas laxissima</i> Duchassaing & Michelotti, 1864; <i>Esperella nuda</i> Ridley & Dendy, 1886; <i>Hircinia cartilaginea</i> var. <i>horrida</i> Hyatt, 1877; <i>Hircinia horrida</i> Hyatt, 1877; <i>Hircinia purpurea</i> Whitfield, 1901; <i>Ircinia cartilaginea</i> var. <i>horrida</i> (Hyatt, 1877); <i>Mycale</i> (<i>Acamasina</i>) <i>laxissima</i> (Duchassaing & Michelotti, 1864); <i>Mycale angulosa</i> sensu Laubenfels, 1936; <i>Mycale</i>	Benthic sessile aquatic animal holobiont	Atlantic: Brazil, Florida, Bahamas; Caribbean: Antilles.	San-Felipe Island (Cuba)	Marine, harbor poles and reefs

	<i>hyatti</i> Pulitzer-Finali, 1986; <i>Mycale jamaicaensis</i> Pulitzer-Finali, 1986; <i>Mycale laxissima</i> (Duchassaing & Michelotti, 1864); <i>Mycale mucifluens</i> Pulitzer-Finali, 1986; <i>Mycale whitfieldi</i> Pulitzer-Finali, 1986; <i>Strongylacidon horridum</i> (Hyatt, 1877); <i>Thorecta horrida</i> (Hyatt, 1877)				
<i>Mycale (Arenochalina) mirabilis</i> (Lendenfeld, 1887)	<i>Mycale (Carmia) cf. spongiosa</i> ; <i>Arenochalina mirabilis</i> Lendenfeld, 1887; <i>Esperella spongiosa</i> Dendy, 1896; <i>Mycale (Arenochalina) tylostrongyla</i> Pulitzer-Finali, 1982; <i>Mycale (Naviculina) mirabilis</i> (Lendenfeld, 1887); <i>Mycale fistulata</i> Hentschel, 1911; <i>Mycale fistulata</i> var. <i>macrochela</i> Hentschel, 1911; <i>Mycale mirabilis</i> (Lendenfeld, 1887); <i>Mycale spongiosa</i> (Dendy, 1896); <i>Mycale tylostrongyla</i> Pulitzer-Finali, 1982; <i>Naviculina mirabilis</i> (Lendenfeld, 1887)	Benthic sessile aquatic animal holobiont	Indo-Pacific: Australia, Seychelles, Coral Sea, India, Sri Lanka	Wasp Head, Durras (Australia)	Marine reefs, rocks, dead coral, living coral
<i>Mycale</i> sp. 1		Benthic sessile aquatic animal holobiont	Indian	Thailand	Marine
<i>Mycale</i> sp. 2		Benthic sessile aquatic animal holobiont	Indian	Thailand	Marine
<i>Mycale</i> sp. 3		Benthic sessile aquatic animal holobiont	Indian	Thailand	Marine

<i>Oscarella</i> Vosmaer, 1884	Oscarellidae Lendenfeld, 1887	<i>Oscarella balibaloï</i> Pérez, Ivanišević, Dubois, Pedel, Thomas, Tokina & Ereskovsky, 2011		Benthic sessile aquatic animal holobiont	Mediterranean	Marseille (France) Mediterranean	Marine, overgrowing massive sponges, gorgonians or some erected bryozoans
<i>Petrosaspongia</i> Bergquist, 1995	Thorectidae Bergquist, 1978	<i>Petrosaspongia</i> <i>nigra</i> Bergquist, 1995		Benthic sessile aquatic animal holobiont	Pacific: New Caledonia	New Caledonia	Marine, reefs, rocks, sandy, dead coral, sediments, lagoons
<i>Phorbas</i> Duchassaing & Michelotti, 1864	Hymedesmiidae Topsent, 1928	<i>Petrosaspongia</i> sp. <i>Phorbas areolatus</i> (Thiele, 1905)	<i>Anchinoe areolata</i> (Thiele, 1905); <i>Hymedesmia areolata</i> Thiele, 1905	Benthic sessile aquatic animal holobiont	South Pacific Southern Atlantic: Maldivas, South Georgia, Uruguay, Argentina; Antarctic: South Shetland Pacific	Fiji South Shetland Islands (Antarctica)	Marine, rocks
		<i>Phorbas</i> sp. 1		Benthic sessile aquatic animal holobiont		Howe Sound British Columbia (Canada)	Marine
		<i>Phorbas</i> sp. 2		Benthic sessile aquatic animal holobiont	Pacific	Korea	Marine
		<i>Phorbas</i> sp. 3		Benthic sessile aquatic animal holobiont	Pacific	Howe Sound British Columbia (Canada)	Marine
		<i>Phorbas</i> sp. 4		Benthic sessile aquatic animal holobiont	Pacific	Korea	Marine
		<i>Phorbas</i> sp. 5		Benthic sessile aquatic animal holobiont	Pacific	Gageo Island, Korea	Marine
		<i>Phorbas</i> sp. 6		Benthic sessile aquatic animal holobiont	Pacific	British Columbia (Canada)	Marine
<i>Phyllospongia</i> Ehlers, 1870	Thorectidae Bergquist, 1978	<i>Phyllospongia</i> <i>foliascens</i> (Pallas, 1766)	<i>Spongia foliascens</i> Pallas, 1766; <i>Carteriospongia</i> <i>foliascens</i> (Pallas, 1766); <i>Hircinia</i> (<i>Polyfibrospongia</i>) <i>flabellifera</i> (Bowerbank, 1877); <i>Spongia othaitica</i> Lamarck,	Benthic sessile aquatic animal holobiont	Indo-Pacific: East South-East African coast, Madagascar, Maldives, Seychelles, New Zealand, Australia,	Okinawa (Japan), Papua New Guinea, Indonesia, GBR (Australia); South China;	Marine, tropical, subtropical reefs, shallow sandy lagoons

	1814; <i>Spongia penicillata</i> Esper, 1794		Solomon Islands, Banda Sea, Bismarck Sea; Red Sea.	Chidiapatu Andamans, India.	
<i>Phyllospongia lamellosa</i> (Esper, 1794)	<i>Cacospongia lamellosa</i> (Esper, 1794); <i>Carteriospongia lamellosa</i> (Esper, 1794); <i>Carteriospongia radiata</i> Hyatt, 1877; <i>Carteriospongia radiata</i> var. <i>complexa</i> Hyatt, 1877; <i>Carteriospongia radiata</i> var. <i>dulsiana</i> Hyatt, 1877; <i>Carteriospongia radiata</i> var. <i>radiata</i> Hyatt, 1877; <i>Mauricea lacinulosa</i> Carter, 1877; <i>Phyllospongia papyracea polyphylla</i> (Lamarck, 1814); <i>Phyllospongia radiata</i> (Hyatt, 1877); <i>Spongia laciniata</i> Lamarck, 1814; <i>Spongia lamellosa</i> Esper, 1794; <i>Spongia polyphylla</i> Lamarck, 1814; <i>Strepsichordaia radiata</i> (Hyatt, 1877)	Benthic sessile aquatic animal holobiont	Indian-Pacific: Australia, African coast; Red Sea.	Hurghada (Egypt) Red Sea; Indo- West Pacific Ocean	Marine, reefs, rocks
<i>Phyllospongia papyracea</i> (Esper, 1806)	<i>Carteriospongia madagascarensis</i> Hyatt, 1877; <i>Phyllospongia (Spongionella) distans</i> Lendenfeld, 1889; <i>Phyllospongia (Spongionella) madagascarensis</i> (Hyatt, 1877); <i>Phyllospongia (Spongionella) papyracea</i> (Esper, 1806); <i>Phyllospongia coriacea</i> Thiele, 1899); <i>Phyllospongia distans</i> Lendenfeld, 1889; <i>Phyllospongia holdsworthi</i> (Bowerbank, 1873); <i>Phyllospongia</i>	Benthic sessile aquatic animal holobiont	Indo-Pacific: Australia, New Zealand, India, African Coast, Madagascar; Red Sea	Madagascar; Hainan Island, China; Papua New Guinea; Sangihe Island; Indonesia	Marine, clear water marine habitats such as reefs, lagoons and intertidal flats

			<i>madagascarensis</i> (Hyatt, 1877); <i>Spongia papyracea</i> Esper, 1806; <i>Spongionella holdsworthii</i> Bowerbank, 1873				
		<i>Phyllospongia vermicularis</i> Lendenfeld, 1889	<i>Phyllospongia</i> (<i>Carteriospongia</i>) <i>vermicularis</i> Lendenfeld, 1889; <i>Carteriospongia vermicularis</i> (Lendenfeld, 1889); <i>Carterispongia vermicularis</i> (Lendenfeld, 1889); <i>Phyllospongia</i> (<i>Carteriospongia</i>) <i>vermicularis</i> Lendenfeld, 1889	Benthic sessile aquatic animal holobiont	Indo-Pacific	Solomon Islands	Marine
		<i>Phyllospongia</i> sp. 1		Benthic sessile aquatic animal holobiont	Indo-Pacific	Makassar (Indonesia)	Marine
		<i>Phyllospongia</i> sp. 2		Benthic sessile aquatic animal holobiont	Indo-Pacific	Australia	Marine
		<i>Phyllospongia</i> sp. 3	<i>Carteriospongia</i> sp.	Benthic sessile aquatic animal holobiont	Indo-Pacific	Tai-tung, Taiwan	Marine
		<i>Phyllospongia</i> sp. 4	<i>Carteriospongia</i> sp.	Benthic sessile aquatic animal holobiont	Indian	Madagascar	Marine
		<i>Phyllospongia</i> sp. 5	<i>Carteriospongia</i> sp.	Benthic sessile aquatic animal holobiont	Indo-Pacific	Indonesia	Marine
		<i>Phyllospongia</i> sp. 6	<i>Phyllospongia chondrodes</i> Wrong taxonomy	Benthic sessile aquatic animal holobiont	Pacific	Okinawa (Japan)	Marine
<i>Pleraplysilla</i> Topsent, 1905	Dysideidae Gray, 1867	<i>Pleraplysilla spinifera</i> (Schulze, 1879)	<i>Pleraplysilla minchini</i> Topsent, 1905; <i>Spongelia spinifera</i> Schulze, 1879	Benthic sessile aquatic animal holobiont	Mediterranean; Atlantic: Macaronesia, Canary, Cape Verde, Azores; North Sea	Mediterranean	Marine, growing on <i>Paramuricea chamaeleon</i>

<i>Polyfibrospongia</i> Bowerbank, 1877	Thorectidae Bergquist, 1978	<i>Polyfibrospongia flabellifera</i> Bowerbank, 1877	Orig. Name: <i>Polyfibrospongia flabellifera</i> Bowerbank, 1877. Synonymised names: <i>Carteriospongia flabellifera</i> (Bowerbank, 1877) (reverted genus)	Benthic sessile aquatic animal holobiont	South Pacific	Vanuatu	Marine
<i>Psammocinia</i> Lendenfeld, 1889	Irciniidae Gray, 1867	<i>Psammocinia halmiformis</i> (Lendenfeld, 1888)	<i>Hircinia (Psammocinia) halmiformis</i> Lendenfeld, 1888; <i>Hircinia halmiformis</i> Lendenfeld, 1888	Benthic sessile aquatic animal holobiont	Indo-Pacific: Australia	Australia	Marine, reef, rocks
		<i>Psammocinia</i> sp. 1		Benthic sessile aquatic animal holobiont	Indo-Pacific	Durras Island, Australia	Marine
		<i>Psammocinia</i> sp. 2		Benthic sessile aquatic animal holobiont	Indo-Pacific	Australia	Marine
		<i>Psammocinia</i> sp. 3		Benthic sessile aquatic animal holobiont	Indo-Pacific	Australia	Marine
		<i>Psammocinia</i> sp. 4		Benthic sessile aquatic animal holobiont	Indo-Pacific	Korea	Marine
		<i>Psammocinia</i> sp. 5		Benthic sessile aquatic animal holobiont	Indo-Pacific	Australia	Marine
		<i>Psammocinia</i> sp. 6		Benthic sessile aquatic animal holobiont	Indo-Pacific	North Sulawesi, Indonesia	Marine
		<i>Psammocinia</i> sp. 7		Benthic sessile aquatic animal holobiont	Indo-Pacific	Korea	Marine
		<i>Psammocinia</i> sp. (CMB-03344)		Benthic sessile aquatic animal holobiont	Indo-Pacific	Port Phillip Heads, Victoria, Australia	Marine
		<i>Psammocinia</i> sp. (CMB-01008)		Benthic sessile aquatic animal holobiont	Indo-Pacific	Durras Island, Australia	Marine

		<i>Psammocinia</i> sp. (CMB-02858)		Benthic sessile aquatic animal holobiont	Indo-Pacific	Barwon Heads, Victoria, Australia	Marine
<i>Rhabdastrella</i> Thiele, 1903	Ancorinidae Schmidt, 1870	<i>Rhabdastrella</i> cf. <i>distincta</i> (Thiele, 1900) <i>Rhabdastrella</i> <i>globostellata</i> (Carter, 1883)	<i>Coppatias distinctus</i> Thiele, 1900 <i>Aurora globostellata</i> (Carter, 1883); <i>Coppatias carteri</i> (Ridley, 1884); <i>Geodia</i> <i>globostellata</i> (Carter, 1883); <i>Jaspis carteri</i> (Ridley, 1884); <i>Stelletta globostellata</i> Carter, 1883; <i>Stellettinopsis carteri</i> Ridley, 1884; <i>Stellettinopsis</i> <i>purpurea</i> Carter, 1886	Benthic sessile aquatic animal holobiont Benthic sessile aquatic animal holobiont	Indo-Pacific: Sulawesi and Makassar Sea Indo-Pacific: Australia, New Zealand, African coast, Madagascar, Seychelles, China Sea, New Caledonia, Caroline Islands, Vietnam, Jaba Sea, Coral Sea, Maldives, Papua, Micronesia, Fiji, India Pacific	Hainan, the South China Sea New Caledonia; Hainan Island, South China	Marine, reefs, rocks, lagoons Marine, reefs, rocks, lagoons
<i>Rhopaloeides</i> Thompson, Murphy, Bergquist & Evans, 1987	Spongiidae Gray, 1867	<i>Rhopaloeides</i> sp.		Benthic sessile aquatic animal holobiont		Sada-misaki, Ehime Prefecture, (Japan)	Marine, rocks
<i>Sarcotragus</i> Schmidt, 1862	Irciniidae Gray, 1867	<i>Sarcotragus</i> <i>fasciculatus</i> (Pallas, 1766) <i>Sarcotragus</i> <i>spinosulus</i> Schmidt, 1862	<i>Spongia fasciculata</i> Pallas, 1766; <i>Hircinia</i> (<i>Sarcotragus</i>) <i>fasciculata</i> (Pallas, 1766); <i>Hircinia fasciculata</i> (Pallas, 1766); <i>Ircinia fasciculata</i> (Pallas, 1766); <i>Spongia</i> <i>fasciculata</i> Pallas, 1766; <i>Spongia fasciculata</i> sensu Esper, 1794; stinker sponge <i>Filifera</i> (<i>Sarcotragus</i>) <i>spinosulus</i> Schmidt, 1862; <i>Hircinia</i> (<i>Sarcotragus</i>) <i>spinosula</i> Schmidt, 1862; <i>Hircinia spinosula</i> (Schmidt, 1862); <i>Hircinia spinulosa</i> ; <i>Ircinia</i> (<i>Sarcotragus</i>) <i>spinosula</i> Schmidt, 1862; <i>Ircinia</i>	Benthic sessile aquatic animal holobiont Benthic sessile aquatic animal holobiont	Caribbean: Venezuela; Atlantic: Brazil, Cape Verde. Mediterranean; Atlantic: Gulf of Guinea, South European Atlantic Shelf, Macaronesia, Madeira, Canary Islands, Azores	Naples (Italy) Italy	Marine, cave, rocky bottom, <i>Posidonia oceanica</i> meadow, coralligenous community. Bathymetric range 1-100 m. Marine, cave, rocky, detritic and muddy bottom, coralligenous community, lagoons, <i>Posidonia oceanica</i> meadow, epibiotic on <i>Pinna</i> <i>nobilis</i> . Bathymetric range 1–60 m

			<i>spinosula</i> (Schmidt, 1862); <i>Sarcotragus spinosula</i> Schmidt, 1862				
		<i>Sarcotragus</i> sp. 1		Benthic sessile aquatic animal holobiont	Pacific	Chuja-do (Korea)	Marine
		<i>Sarcotragus</i> sp. 2		Benthic sessile aquatic animal holobiont	Pacific	Cheju Island (Korea)	Marine
		<i>Sarcotragus</i> sp. 3		Benthic sessile aquatic animal holobiont	Pacific	New Zealand	Marine
		<i>Sarcotragus</i> sp. 4		Benthic sessile aquatic animal holobiont	Pacific	Jaeju Island, Korea	Marine
		<i>Sarcotragus</i> sp. 5		Benthic sessile aquatic animal holobiont	Pacific	Durras, Wales, Australia	Marine
		<i>Sarcotragus</i> sp. 6		Benthic sessile aquatic animal holobiont	Pacific	Zew Zealand	Marine
<i>Scalarispongia</i> Cook & Bergquist, 2000	Thorectidae Bergquist, 1978	<i>Scalarispongia</i> <i>scalaris</i> (Schmidt, 1862	<i>Aplysinopsis massa</i> f. <i>lobulosa</i> Topsent, 1934; <i>Aplysinopsis</i> <i>massa</i> f. <i>osculata</i> Topsent, 1934; <i>Aplysinopsis massa</i> f. <i>porrecta</i> Topsent, 1934; <i>Cacospongia scalaris</i> Schmidt, 1862; <i>Stelospongia scalaris</i> (Schmidt, 1862); leather sponge	Benthic sessile aquatic animal holobiont. Prey of nudibranch <i>Hypselodoris</i> <i>fontandraui</i> (Pruvot-Fol, 1951) actively grazing	Mediterranean; Atlantic Macaronesia, Canary Islands, Cape Verde, Azores; Pacific	Gulf of Astakos (Greece); Tarifa, Spain	Marine, cave, rocky/detritic/muddy bottom, coralligenous community, <i>Posidonia</i> <i>oceanica</i> meadow, lagoons, artificial reefs, epibiotic on <i>Pinna nobilis</i> . Bathymetric range 1–250 m
		<i>Scalarispongia</i> cf. <i>scalaris</i> (Schmidt, 1863		Benthic sessile aquatic animal holobiont	Pacific	Japan	Marine
		<i>Scalarispongia</i> cf. <i>linteriformis</i> (Lamarck, 1814)	<i>Spongia linteriformis</i> Lamarck, 1814; <i>Cacospongia linteriformis</i> (Lamarck, 1814)	Benthic sessile aquatic animal holobiont	Atlantic, Caribbean	Grand Bahama Island, Bahamas (Caribbean)	Marine

		<i>Scalarispongia</i> sp.		Benthic sessile aquatic animal holobiont	Pacific	Dokdo (Korea)	Marine, reefs, rocks
<i>Semitaspongia</i> Cook & Bergquist, 2000	Thorectidae Bergquist, 1978	<i>Semitaspongia bactriana</i> Cook & Bergquist, 2000	<i>Semitaspongia bactriana</i> Cook & Bergquist, 2000	Benthic sessile aquatic animal holobiont	New Zealand	New Zealand	Marine
<i>Sigmatosceptrella</i> Dendy, 1922	Podospongiidae Laubenfels, 1936	<i>Sigmatosceptrella fibrosa</i> (Dendy, 1897)	<i>Latrunculia conulosa</i> Hallmann, 1912; <i>Spirastrella fibrosa</i> Dendy, 1897	Benthic sessile aquatic animal holobiont	Indo-Pacific: Australia	Flinders, Victoria (Australia)	Marine, rocks
		<i>Sigmatosceptrella</i> sp. 1		Benthic sessile aquatic animal holobiont	Pacific	Okinawa (Japan)	Marine, coral reef
		<i>Sigmatosceptrella</i> sp. 2		Benthic sessile aquatic animal holobiont	Pacific	Great Australian Bight, Australia	Marine
<i>Smenospongia</i> Wiedenmayer, 1977	Thorectidae Bergquist, 1978	<i>Smenospongia</i> sp. 1		Benthic sessile aquatic animal holobiont	Pacific	Gagu-D (Korea)	Marine, reefs, rocks
		<i>Smenospongia</i> sp. 2		Benthic sessile aquatic animal holobiont	Pacific	Soheuksan Island, (Korea)	Marine, reefs, rocks
<i>Spongia</i> Linnaeus, 1759	Spongiidae Gray, 1867	<i>Spongia</i> sp. 1		Benthic sessile aquatic animal holobiont	Atlantic	Ohio and Grassy Keys, Florida (USA)	Marine, rocks
		<i>Spongia</i> sp. 2		Benthic sessile aquatic animal holobiont	Pacific	Toyama Bay, Japan Sea	Marine, rocks
		<i>Spongia</i> sp. 3		Benthic sessile aquatic animal holobiont	Pacific	Toyama Bay, Japan Sea	Marine reefs, rocks, dead coral, living coral
		<i>Spongia</i> sp. 4		Benthic sessile aquatic animal holobiont	Pacific	Tong-Yong City (South Korea)	Marine reefs, rocks, dead coral, living coral
		<i>Spongia</i> sp. 5		Benthic sessile aquatic animal holobiont	Pacific	Tong-Yong City (South Korea)	Marine, rocks

<i>Spongia</i> sp. 6		Benthic sessile aquatic animal holobiont	Pacific	Vanuatu Islands, Australia	Marine reefs, rocks, dead coral, living coral, lagoons; growing on sides of coral heads, in gullies, and on walls of coral reefs
<i>Spongia</i> sp. 7		Benthic sessile aquatic animal holobiont	Pacific	Vanuatu Islands, Australia	
<i>Spongia</i> sp. 8		Benthic sessile aquatic animal holobiont	Pacific	Australia	
<i>Spongia</i> sp. 9		Benthic sessile aquatic animal holobiont	Pacific	Australia	
<i>Spongia</i> sp. 10		Benthic sessile aquatic animal holobiont	Pacific	Great Australian Bight, (Australia)	
<i>Spongia</i> sp. 11		Benthic sessile aquatic animal holobiont	Pacific	Borneo	
<i>Spongia</i> sp. 12		Benthic sessile aquatic animal holobiont	South Pacific	Tong-Yong City, Korea	
<i>Spongia (Spongia) agaricina</i> Pallas, 1766	<i>Spongia agaricina</i> Pallas, 1766 <i>Hippospongia agaricina</i> (Pallas, 1766); <i>Spongia agaricina</i> Pallas, 1766; <i>Spongia thienemanni</i> Arndt, 1943	Benthic sessile aquatic animal holobiont	Mediterranean	Cádiz, Spain	
<i>Spongia (Spongia) hispida</i> Lamarck, 1814	<i>Cacospongia mollior</i> sensu Ridley, 1884; <i>Euspongia irregularis</i> Lendenfeld, 1886; <i>Euspongia irregularis</i> var. <i>areolata</i> Whitelegge, 1901; <i>Euspongia irregularis</i> var. <i>dura</i> Lendenfeld, 1889; <i>Euspongia irregularis</i> var. <i>frondosa</i> Lendenfeld, 1889; <i>Euspongia</i>	Benthic sessile aquatic animal holobiont	Indo-Pacific: Sri Lanka, India, Australia, New Zealand, Madagascar, Banda Sea, Hawai'i; Red Sea	GBR (Australia); Papua New Guinea	Marine, coral reefs, rocks, lagoons

	<i>irregularis</i> var. <i>jacksoniana</i> Lendenfeld, 1886; <i>Euspongia</i> <i>irregularis</i> var. <i>lutea</i> Lendenfeld, 1886; <i>Euspongia</i> <i>irregularis</i> var. <i>mollior</i> (sensu Ridley, 1884); <i>Euspongia</i> <i>irregularis</i> var. <i>silicata</i> Lendenfeld, 1886; <i>Euspongia</i> <i>irregularis</i> var. <i>tenuis</i> Lendenfeld, 1886; <i>Euspongia</i> <i>irregularis</i> var. <i>villosa</i> Lendenfeld, 1889; <i>Spongia</i> (<i>Spongia</i>) <i>irregularis</i> (Lendenfeld, 1889); <i>Spongia</i> <i>hispida</i> Lamarck, 1814; <i>Spongia irregularis</i> (Lendenfeld, 1889); <i>Spongia</i> <i>irregularis</i> var. <i>mollior</i> (sensu Ridley, 1884)				
<i>Spongia</i> (<i>Spongia</i>) <i>matamata</i> de Laubenfels, 1954	<i>Spongia matamata</i> de Laubenfels, 1954; <i>Spongia</i> <i>officinalis matamata</i> de Laubenfels, 1954	Benthic sessile aquatic animal holobiont	Pacific: Marshall Islands, Palau	Marine lake at Palau, Western Caroline Islands	Marine reefs, rocks, dead coral, living coral, lagoons
<i>Spongia</i> (<i>Spongia</i>) <i>nitens</i> (Schmidt, 1862)	<i>Ditela nitens</i> Schmidt, 1862; <i>Euspongia nitens</i> (Schmidt, 1862); <i>Euspongia officinalis</i> var. <i>nitens</i> (Schmidt, 1862); <i>Spongia nitens</i> (Schmidt, 1862); <i>Spongia officinalis</i> <i>nitens</i> (Schmidt, 1862)	Benthic sessile aquatic animal holobiont	Mediterraenan; Atlantic Macaronesia, Canary Islands, Cape Verde, Azores	Italy Mediterranean	Marine, cave, coralligenous community. Bathymetric range 0–15 m.
<i>Spongia</i> (<i>Spongia</i>) <i>oceanica</i> de Laubenfels, 1950	<i>Spongia oceanica</i> de Laubenfels, 1950	Benthic sessile aquatic animal holobiont. Prey of <i>Glossodoris</i> <i>rufomarginata</i> (before <i>C.</i> <i>youngbleuthi</i>) that takes	Pacific: Hawai'i	Pupukea (Hawai'i) Pacific	Marine, reefs, rocks, dead- coral

<i>Spongia (Spongia) officinalis</i> Linnaeus, 1759	<i>Euspongia officinalis</i> (Linnaeus, 1759); <i>Euspongia officinalis</i> var. <i>adriatica</i> Schmidt, 1862; <i>Spongia adriatica</i> Schmidt, 1862; <i>Spongia officinalis</i> Linnaeus, 1759; <i>Spongia officinalis mediterranea</i> Hyatt, 1877; <i>Spongia officinalis mediterranea</i> var. <i>tubuliformis</i> Hyatt, 1877; <i>Spongia officinalis mediterranea</i> var. <i>zimociformis</i> Hyatt, 1877; <i>Spongia officinalis</i> var. <i>adriatica</i> Schmidt, 1862; <i>Spongia quarnerensis</i> Schmidt, 1862	sesterterpenes for own defense Benthic sessile aquatic animal holobiont	Mediterranean; Atlantic Macaronesia, Canary Islands, Cape Verde, Azores; Caribbean (inaccurate)	Naples (Italy); Adriatic; Sicily Mediterranean	Marine, cave, coralligenous community, rocky/detritic/muddy/sandy bottom, lagoons, coralligenous community, <i>Posidonia oceanica</i> meadow. Bathymetric range 1–70 m
<i>Spongia (Spongia) tubulifera</i> Lamarck, 1814	<i>Euspongia officinalis</i> var. <i>rotunda</i> (Hyatt, 1877); <i>Euspongia tubulifera</i> (Lamarck, 1814); <i>Luffaria rupicola</i> Duchassaing & Michelotti, 1864; <i>Spongia officinalis tubulifera</i> Lamarck, 1814; <i>Spongia officinalis tubulifera</i> var. <i>aperta</i> Hyatt, 1877; <i>Spongia officinalis tubulifera</i> var. <i>disciformis</i> Hyatt, 1877; <i>Spongia officinalis tubulifera</i> var. <i>duplex</i> Hyatt, 1877; <i>Spongia officinalis tubulifera</i> var. <i>exotica</i> Hyatt, 1877; <i>Spongia officinalis tubulifera</i> var. <i>rotunda</i> Hyatt, 1877; <i>Spongia tubulifera</i> Lamarck, 1814; <i>Spongia tubulifera</i> var.	Benthic sessile aquatic animal holobiont	Caribbean; Atlantic: Bermuda	Mexican Caribbean	Marine reefs, rocks, dead coral, living coral

			<i>disciformis</i> Hyatt, 1877; <i>Spongia tubulifera</i> var. <i>osculata</i> Duchassaing & Michelotti, 1864; <i>Spongia</i> <i>tubulifera</i> var. <i>porosa</i> Duchassaing & Michelotti, 1864; <i>Spongia tubulifera</i> var. <i>rotunda</i> Hyatt, 1877				
		<i>Spongia (Spongia)</i> <i>virgultosa</i> (Schmidt, 1868)	<i>Euspongia officinalis</i> var. <i>tubulosa</i> Schulze, 1879; <i>Euspongia virgultosa</i> Schmidt, 1868; <i>Spongia officinalis</i> <i>tubulosa</i> (Schulze, 1879); <i>Spongia virgultosa</i> (Schmidt, 1868)	Benthic sessile aquatic animal holobiont	Mediterranean; Atlantic Macaronesia, Canary Islands, Cape Verde, Azores	Italy, Spain Mediterranean	Marine, cave, coralligenous, detritic/muddy bottom, lagoons, artificial reef, <i>Posidonia oceanica</i> meadow, epibiotic on <i>Pinna</i> <i>nobilis</i> . Generally covered by epibionts in turbulent superficial water. Bathymetric range 1–50 m Marine
<i>Spongionella</i> Bowerbank, 1862	Dictyodendrillidae Bergquist, 1980	<i>Spongionella</i> sp. 1		Benthic sessile aquatic animal holobiont			
		<i>Spongionella</i> sp. 2		Benthic sessile aquatic animal holobiont	Pacific	Japan	Marine
<i>Stelletta</i> Schmidt, 1862	Ancorinidae Schmidt, 1870	<i>Stelletta tenuis</i> Lindgren, 1897		Benthic sessile aquatic animal holobiont	Indo-Pacific: Java		Marine, coral reefs, rocks
<i>Strepsichordaia</i> Bergquist, Ayling & Wilkinson, 1988	Thorectidae Bergquist, 1978	<i>Strepsichordaia</i> <i>aliena</i> (Wilson, 1925) <i>Strepsichordaia</i> <i>lendenfeldi</i> Bergquist, Ayling & Wilkinson, 1988	<i>Phyllospongia aliena</i> Wilson, 1925	Benthic sessile aquatic animal holobiont Benthic sessile aquatic animal holobiont	Indo-Pacific: Palawan, Northern Borneo Pacific: GBF (Australia)	Indonesia GBR (Australia)	Marine, coral reefs, rocks, lagoons Marine reefs, rocks, dead coral, sandy bottom, lagoons
<i>Suberea</i> Bergquist, 1995	Aplysinellidae Bergquist, 1980	<i>Suberea</i> sp. 1		Benthic sessile aquatic animal holobiont	Pacific	Philippines	Marine reefs, rocks, dead coral, sandy bottom, lagoons

		<i>Suberea</i> sp. 2		Benthic sessile aquatic animal holobiont	Pacific	Madnag, Papua New Guinea	Marine
<i>Suberites</i> Nardo, 1833	Suberitidae Schmidt, 1870	<i>Suberites caminatus</i> Ridley & Dendy, 1886		Benthic sessile aquatic animal holobiont	Antarctic Ocean: Antarctica; Atlantic: South Georgia Island, East coast of South America; Indian: Marion and Prince Edward Islands	Antarctica	Marine, rocks, sandy bottoms
		<i>Suberites</i> sp. 1		Benthic sessile aquatic animal holobiont	Antarctic	Antarctica	Marine
		<i>Suberites</i> sp. 2		Benthic sessile aquatic animal holobiont	Antarctic	Antarctica	Marine
<i>Taonura</i> Carter, 1882	Thorectidae Bergquist, 1978	<i>Taonura marginalis</i> (Lendenfeld, 1888)	<i>Cacospongia exemplum</i> var. <i>terminus</i> Carter, 1886; <i>Thorecta exemplum</i> var. <i>marginalis</i> Lendenfeld, 1888; <i>Thorecta marginalis</i> Lendenfeld, 1888	Benthic sessile aquatic animal holobiont	Pacific: Australia	New South Wales (Australia)	Marine, reefs, rocks,
<i>Thorecta</i> Lendenfeld, 1888	Thorectidae Bergquist, 1978	<i>Thorecta</i> sp.		Benthic sessile aquatic animal holobiont	Pacific	New Zealand	Marine, rocks
<i>Thorectandra</i> Lendenfeld, 1889	Thorectidae Bergquist, 1978	<i>Thorectandra</i> sp. 1		Benthic sessile aquatic animal holobiont			Marine
		<i>Thorectandra</i> sp. 2		Benthic sessile aquatic animal holobiont		Palau	Marine, reefs, rocks
		<i>Thorectandra excavatus</i> (Ridley, 1884)	<i>Stelospongia excavata</i> (Ridley, 1884); <i>Stelospongos excavatus</i> Ridley, 1884	Benthic sessile aquatic animal holobiont	Pacific: Australia, New Zealand	Darwin (Australia)	Marine, reefs, rocks
Xestospongia Laubenfels, 1932	Petrosiidae van Soest, 1980	<i>Xestospongia</i> sp.		Benthic sessile aquatic animal holobiont	Pacific	Sikao Bay (Thailand)	Marine, rocks

Dictyoceratida sponge	Order Dictyoceratida Minchin, 1900	Dictyoceratida sp.	Benthic sessile aquatic animal holobiont	South Pacific	Palau	Marine
Unidentified sponge		Unidentified sponge 1	Benthic sessile aquatic animal holobiont	Pacific	Australia	Marine
Unidentified sponge		Unidentified sponge 2	Benthic sessile aquatic animal holobiont	Red Sea	Egypt (Red Sea)	Marine
Unidentified sponge		Sponge NS009	Benthic sessile aquatic animal holobiont	Pacific	Okinawa, Japan	Marine

References

1. R. W. M. Van Soest, N. Boury-Esnault, J. N. A. Hooper, K. Rützler, N. J. de Voogd, B. Alvarez, E. Hajdu, A. B. Pisera, R. Manconi, C. Schönberg, M. Klautau, M. Kelly, J. Vacelet, M. Dohrmann, M.-C. Díaz, P. Cárdenas, J. L. Carballo, P. Ríos, R. Downey and C. C. Morrow, World porifera database, <http://www.marinespecies.org/porifera>, (accessed 24_04_2020).

Table S3. Accepted name, family, species, synonyms, lifestyle, geographical distribution, and collection site of the selected Mollusca ¹

Accepted genus name and protologue	Family	Species	Synonyms	Lifeform / Ecology	Geographical distribution	Collection site (specimens with sesterterpenoids)	Environment
<i>Ardeadoris</i> Rudman, 1984	Chromodorididae Bergh, 1892	<i>Ardeadoris averni</i> (Rudman, 1985)	<i>Glossodoris averni</i> Rudman, 1985	Nudibranch benthic motile mollusk, brilliant red margins and white body. The rhinophores and gills are the same shade as the margin, the depth of colour, frilliness and width of which is variable; feed on Dictyoceratidae sponges; simultaneous hermaphrodites	Australia	Gneerings Reef and at Mudjimba Island offshore from Mooloolaba, Queensland (Australia)	Marine, coral reefs
<i>Cadlina</i> Bergh, 1879	Cadlinidae Bergh, 1891	<i>Cadlina luteomarginata</i> MacFarland, 1966	<i>Cadlina marginata</i> MacFarland, 1905 (invalid: junior secondary homonym of <i>Doris marginata</i> Montagu, 1804, a species of <i>Cadlina</i> synonymous with <i>C. laevis</i> ; <i>C. luteomarginata</i> is a replacement name)	Nudibranch benthic motile mollusk, white with yellow capped tubercles, a yellow border to mantle and foot, and yellow tipped gills and rhinophores, edges of gill and rhinophore pockets also lined yellow; feed on a variety of spiculate sponges and accumulate terpenoid defense; simultaneous hermaphrodites	North Atlantic	Howe Sound, British Columbia (Canada); San Diego, California (USA)	Marine, feeding on <i>Phorbas</i> sp. Most of the terpenoids from <i>C. luteomarginata</i> are sequestered from sponge diet, but a subset is biosynthesized de novo by the nudibranch.
Chromodorididae Bergh, 1891	<i>Ceratosoma brevicaudatum</i> Abraham, 1876	<i>Ceratosoma brevicaudatum</i> Abraham, 1876	<i>Ceratosoma adelaidae</i> Basedow & Hedley, 1905; <i>Ceratosoma oblongum</i> Abraham, 1876	Nudibranch benthic motile mollusk, shape slug-like with gill structures visible as a single cluster on the back of the animal, pattern includes shades or colours of white, orange and/or red; feeds on sponges and accumulate terpenoid defense; simultaneous hermaphrodites	Pacific	Australia	Marine, living on sponge prey. These derivatives of thiofurodysinin acetate may derive from <i>Dysidea</i> sp. and <i>Dysidea avara</i> , where they have been also detected
		<i>Chromodoris aspersa</i> (Gould, 1852)	<i>Chromodoris inornata</i> Pease, 1871; <i>Chromodoris pallescens</i> Bergh, 1875; <i>Doris amabilis</i> Kelaart, 1859; <i>Doris aspersa</i> Gould, 1852; <i>Doris punctulifera</i> Bergh, 1874; <i>Glossodoris inornata</i> Pease, 1871	Nudibranch benthic motile mollusk, white body and foot covered with magenta spots, most surrounded by a translucent ring. The rhinophores are yellow-orange and the gills vary from pale yellow to orange; feed on orange thin sponges; simultaneous hermaphrodite	Pacific	Koino-ura, Fukuoka prefecture, (Japan)	Marine, rocks, reefs
		<i>Chromodoris hamiltoni</i> Rudman, 1977		Nudibranch benthic motile mollusk, blue background colour, can be very pale, usually three thin dark blue-black lines down the mantle but sometimes there	Indic: Kenya, Madagascar, Tanzania	Aliwal Shoal, Durban (South Africa)	Marine, living on their sponge prey

				can be extra lines and sometimes the inner lines can be replaced by broader diffuse brown or orange lines; feed on sponges and accumulate terpenoid defenses; simultaneous hermaphrodites			
		<i>Chromodoris lineolata</i> (van Hasselt, 1824)	unaccepted: <i>Chromodoris funerea</i> Collingwood, 1881			Kaibakku Lake, Palau	Marine
		<i>Chromodoris willani</i> Rudman, 1982		Nudibranch benthic motile mollusk, bluish white or translucent white background and dark blue or black longitudinal stripes, white spots or specks on both gills and rhinophores, which can range from a watery translucent to a straw yellow-brown, feeds on sponges in the family Thorectidae like <i>Cacospongia mycofijiensis</i> and <i>Semitaspongia</i> , and other (<i>e.g.</i> , <i>Luffariella variabilis</i>); simultaneous hermaphrodite	<i>Cacospongia mycofijiensis</i> and <i>Semitaspongia</i> , and other sponges in the family Thorectidae (<i>e.g.</i> , <i>Luffariella variabilis</i>),	Okinawa (Japan)	Marine, feeds on a sponge containing precursor compounds the sesterterpenes manoalide and secomanoalide (<i>e.g.</i> , <i>Luffariella variabilis</i>), and is likely to biotransform them into the sesterterpenes
<i>Curnon d'Udekem d'Acoz</i> , 2017	Curnonidae d'Udekem d'Acoz, 2017	<i>Curnon granulosa</i> (Vayssière, 1906)	<i>Charcotia granulosa</i> Vayssière, 1906	Nudibranch benthic motile mollusk, translucent white with branches of pink digestive gland showing through the skin, mantle is bordered by an opaque white band and the dorsal surface is covered by pointed tubercles which are tipped with white pigment, rhinophores are translucent and tipped with white; simultaneous hermaphrodite	Antarctica	Deception and Livingston Islands (South Shetland Islands) Antarctica	Marine, rocks
<i>Doriprismatica</i> d'Orbigny, 1839	Chromodorididae Bergh, 1891	<i>Chromolaichma sedna</i> (Ev. Marcus & Er. Marcus, 1967)	<i>Casella sedna</i> Ev. Marcus & Er. Marcus, 1967; <i>Casella sedna</i> Ev. Marcus & Er. Marcus, 1967 (original combination); <i>Chromodoris fayae</i> Lance, 1968; <i>Chromodoris sedna</i> (Ev. Marcus & Er. Marcus, 1967); <i>Doriprismatica sedna</i> (Ev. Marcus & Er. Marcus, 1967);	Nudibranch benthic motile mollusk, body and mantle translucent white with both foot and mantle bordered with three colour bands: an inner opaque white, then red and an outer yellow band, upper half of the rhinophore clubs and the tips of the gills are red; simultaneous hermaphrodites	Caribbean: Gulf of Mexico; Atlantic; Pacific: Mexican Tropical Pacific	Natural Park of Osa Ballena in Costa Rica	Marine, feeds exclusively on spiculated demosponges, includes 17 different prey species known.

			<i>Glossodoris sedna</i> (Ev. Marcus & Er. Marcus, 1967)				
	<i>Doriprismatica atromarginata</i> (Cuvier, 1804)		<i>Doris atromarginata</i> Cuvier, 1804; <i>Casella atromarginata</i> (Cuvier, 1804); <i>Casella atromarginata</i> var. <i>pallida</i> Bergh, 1905; <i>Casella maccarthyi</i> (Kelaart, 1859); <i>Casella philippinensis</i> Bergh, 1874; <i>Doris atromarginata</i> Cuvier, 1804 (original combination); <i>Doris maccarthyi</i> Kelaart, 1859; <i>Glossodoris atromarginata</i> (Cuvier, 1804); <i>Glossodoris maccarthyi</i> (Kelaart, 1858); <i>Goniodoris atromarginata</i> (Cuvier, 1804)	Nudibranch benthic motile mollusk, hard lemon-yellow body with an ruffled margin edged in black, portion of its body raised, black edging is also on the feathery gills and feathery rhinopores, black rings where the rhinophores emerge from the body; feeds on sponges: <i>Hyattella</i> sp., <i>Fasciospongia</i> sp., <i>Spongia</i> sp. and <i>Luffariella</i> sp.; simultaneous hermaphrodites; Cheesecake nudibrach	Red Sea, Indo-Pacific: Asutralia	Mandapam (India); Mudjimba Island and the Gneerings Reef in Mooloolaba, South East Queensland (Australia)	Marine, living on sponge preys: <i>Hyattella</i> sp., <i>Fasciospongia</i> sp., <i>Spongia</i> sp. and <i>Luffariella</i> sp
<i>Felimare</i> Ev. Marcus & Er. Marcus, 1967	Chromodorididae Bergh, 1891	<i>Felimare orsinii</i> (Vérany, 1846)	<i>Chromodoris orsinii</i> (Vérany, 1846); <i>Doris orsinii</i> Vérany, 1846; <i>Glossodoris coelestis</i> (Deshayes in Fredol, 1865); <i>Glossodoris orsinii</i> (Vérany, 1846); <i>Goniodoris coelestis</i> Deshayes in Fredol, 1865; <i>Hypselodoris coelestis</i> (Deshayes in Fredol, 1865); <i>Hypselodoris orsinii</i> (Vérany, 1846)	Nudibranch benthic motile mollusk, blue to dark blue with a yellow to white submarginal line, a central white line, blue rhinophores and posterior dermic mantle glands; feeds on <i>Cacospongia</i> spp. sponges, from which it obtains sesterterpene compounds and derivatives; simultaneous hermaphrodite	Mediterranean Atlantic: Spain, Cantabric	Spain, Italy	Marine, living on sponges prey, including <i>Cacospongia mollior</i> , <i>C. scalaris</i> , <i>Dysidea fragilis</i> , <i>Ircinia fasciculata</i> , <i>Petrosia</i> spp.
	<i>Felimare tricolor</i> (Cantraine, 1835)		<i>Chromodoris tricolor</i> (Cantraine, 1835); <i>Doris tricolor</i> Cantraine, 1835 (original combination); <i>Felimare midatlantica</i> (Gosliner, 1990); <i>Glossodoris tricolor</i> (Cantraine, 1835); <i>Hypselodoris midatlantica</i> Gosliner, 1990; <i>Hypselodoris tricolor</i> (Cantraine, 1835); <i>Mexichromis tricolor</i> (Cantraine, 1835)	Nudibranch benthic motile mollusk, blue to dark blue with a yellow submarginal line, a central yellow line and two discontinuous lateral white lines; feeds on sponge <i>Cacospongia mollior</i> ; simultaneous hermaphrodites	Mediterranean; Atlantic Macaronesia, Canary Islands, Cape Verde, Azores	Pupukea (Hawai'i) Hainan Island Pacific; Hainan Island, South China Sea	Marine, roaming on a black sponge (prey <i>Cacospongia mollior</i>)

		<i>Chromolaichma dalli</i> (Bergh, 1879)	<i>Felimida dalli</i> (Bergh, 1879), <i>Glossodoris dalli</i> (Bergh, 1879), <i>Chromodoris banksi</i> Farmer, 1963; <i>Chromodoris banksi banksi</i> Farmer, 1963; <i>Chromodoris banksi sonora</i> Er. Marcus & Ev. Marcus, 1967; <i>Chromodoris dalli</i> Bergh, 1879 (original combination); <i>Chromodoris sonora</i> Er. Marcus & Ev. Marcus, 1967; <i>Felimida dalli</i> (Bergh, 1879); <i>Glossodoris dalli</i> (Bergh, 1879)	Nudibranch benthic motile mollusk, translucent white with black-brown spots, sometimes orange to red spots or greenish spots on a blackish mantle. Rhinophores and gills are white with orange or light red tips. The mantle margin is orange or light red; feeds on sponges; simultaneous hermaphrodites	Pacific	Natural Park of Osa Ballena in Costa Rica	Marine, feeds exclusively on spiculated demosponges: <i>e.g. Hyrtios erectus</i>
<i>Glossodoris</i> Ehrenberg, 1831	Chromodorididae Bergh, 1891	<i>Glossodoris cincta</i> (Rüppell & Leuckart, 1830)	<i>Casella cincta</i> Bergh, 1888; <i>Casella cincta</i> Bergh, 1888; <i>Casella foxi</i> O'Donoghue, 1929	Nudibranch benthic motile mollusk, mottled brown body with bluish white (outer), then black, then yellow (inner) bands on the heavily folded mantle edge, and the foot. Philippines - Indonesia colour form bluish outer band on the mantle edge but the inner black and yellow lines tend to merge into a dull khaki band. The East African animals lack a distinct bluish white band at the mantle edge. Feeds on sponge <i>Cacospongia</i> spp.; simultaneous hermaphrodite	Red Sea; Pacific: Guam, Australia; Indic: India	Guam	Marine, feeding on sponge <i>Cacospongia</i> sp
		<i>Glossodoris hikuerensis</i> (Pruvot-Fol, 1954)		Nudibranch benthic motile mollusk, pale-brown body which is covered by speckled white dots. It has a very frilly mantle edged with brown-black-brown lines. Its gills are semi-translucent white, and its rhinophores have the same speckled pale-brown pattern as its body; feed on sponges, like <i>Hyrtios erectus</i> ; simultaneous hermaphrodite	Pacific: Tuamotu Islands	Lizard Island, GBR (Australia)	Marine, reefs; on <i>Hyrtios erectus</i> . When disturbed, releases a milky-white substance which is a form of chemical defence
		<i>Glossodoris pallida</i> (Rüppell & Leuckart, 1830)	<i>Doris pallida</i> Rüppell & Leuckart, 1830; <i>Chromodoris pallida</i> (Rüppell & Leuckart, 1830); <i>Doris pallida</i> Rüppell & Leuckart,	Nudibranch benthic motile mollusk, white mantle and yellow or orange border; feeds on sponge <i>Cacospongia</i> spp.; simultaneous hermaphrodite	Red Sea; Pacific: Guam, Australia	Guam	Marine, feeding on sponge <i>Cacospongia</i> sp.

			1830 (original combination); <i>Doris xantholeuca</i> Ehrenberg, 1831; <i>Glossodoris xantholeuca</i> (Ehrenberg, 1831)				
		<i>Glossodoris rufomarginata</i> (Bergh, 1890)	<i>Casella rufomarginata</i> Bergh, 1890; <i>Chromodoris rufomarginata</i> (Bergh, 1890); <i>Chromodoris youngbleuthi</i> Kay & Young, 1969 (original combination); <i>Chromolaichma youngbleuthi</i> (Kay & Young, 1969); <i>Glossodoris youngbleuthi</i> (Kay & Young, 1969)	Nudibranch benthic motile mollusk, externally by the orange-brown speckled pattern on the mantle, the orange-brown mantle border and the broad white submarginal band; feeds on Dictyoceratida sponge; simultaneous hermaphrodites		Pupukea (Hawai'i) Hainan IslandPacific; Hainan Island, South China Sea	Marine, In Hawaii this species take sesterterpenes for own defense from <i>Spongia oceanica</i> ; <i>Glossodoris rufomarginata</i> from China feed on sponges Dictyoceratida containing scalaradial.
		<i>Glossodoris vespa</i> Rudman, 1990		Nudibranch benthic motile mollusk, mantle is black with fine white specks and a broad yellow submarginal band around the mantle skirt both dorsally and ventrally. The body and foot are also black and there is a paler yellow submarginal band around the edge of the foot. The gills and the rhinophores are also black, the gill lamellae being translucent; feed on Dictyoceratidasponges; simultaneous hermaphrodites	Australia	Gneerings Reef and at Mudjimba Island offshore from Mooloolaba, Queensland (Australia); Mooloolaba, Queensland (Australia)	Marine reefs
<i>Hypselodoris</i> Stimpson, 1855	Chromodorididae Bergh, 1891	<i>Hypselodoris capensis</i> (Barnard, 1927)	<i>Glossodoris capensis</i> Barnard, 1927	Nudibranch benthic motile mollusk, white-bodied dorid with a smooth skin, opaque white lines along the notum and irregular reddish-orange spots, broken blue-purple margin. It has eight gills arranged around the anus and its rhinophores are perfoliate. The rhinophores and gill edges are orange;feeds on sponges <i>Fasciospongia</i> sp. and <i>Dysidea</i> sp.; simultaneous hermaphrodites	South Africa	Tsitsikamma Marine Reserve (South Africa)	Marine, living on prey sponges, <i>Fasciospongia</i> sp. and <i>Dysidea</i> sp.

References

1. WoRMS Editorial Board, World register of marine species, <http://www.marinespecies.org>, (accessed 05_05_2024).

Table S4. Accepted name, family, species, synonyms, lifestyle, geographical distribution, and collection site of the selected Cnidaria¹

Accepted genus name and protologue	Family	Species	Synonyms	Lifeform / Ecology	Geographical distribution	Collection site (specimens with sesterterpenes)	Environment
<i>Cladocora</i> Ehrenberg, 1834	Cladocoridae Milne Edwards & Haime, 1857	<i>Cladocora caespitosa</i> (Linnaeus, 1767)	<i>Madrepora caespitosa</i> Linnaeus, 1767 Orig. Name; <i>Caryophyllia caespitosa</i> (Linnaeus, 1767)-superseded combination; <i>Cladocora astraearia</i> Sars, 1857-junior subjective synonym; <i>Cladocora caespitosa</i> var. <i>megastoma</i> Ehrenberg, 1834-junior subjective synonym; <i>Cladocora caespitosa</i> var. <i>microstoma</i> Ehrenberg, 1834-junior subjective synonym; <i>Cladocora cespitosa</i> (Linnaeus, 1767)-incorrect subsequent spelling; <i>Cladocora cespitosa</i> var. <i>astraearia</i> Sars, 1857-junior subjective synonym; <i>Cladocora cespitosa</i> var. <i>stellaria</i> Milne Edwards & Haime, 1848 junior subjective synonym; <i>Cladocora cespitosa</i> var. <i>typica</i> Pax & Müller, 1954-junior subjective synonym; <i>Cladocora stellaria</i> Milne Edwards & Haime, 1848-junior subjective synonym; <i>Hoplangia pallaryi</i> Joubin, 1930-junior subjective synonym; <i>Madrepora caespitosa</i> Linnaeus, 1767-superseded combination (basonym); <i>Matrepora caespitosa</i> (Linnaeus, 1767)-superseded combination	Cushion coral forms the only true coral reef in the Mediterranean, polyps are a clear maroon colour, 5 mm in diameter, lives in symbiosis with zooxanthellae algae, produce deposits of calcium carbonate; spawner, gonochoric.	Mediterranean	Águilas, Murcia (Spain)	Marine, rocks, coralligenous, maximum depth of 40m

References

1. WoRMS Editorial Board, World register of marine species, <http://www.marinespecies.org>, (accessed 05_05_2020).

Table S5. Accepted name, family, species, synonyms, and geographical distribution of the selected Insecta

Accepted name ¹	Family	Synonyms	Geographical distribution	References
<i>Ceroplastes albolineatus</i> Cockerell, 1894	Coccidae Fallén, 1814	<i>Ceroplastes albolineatus</i> Cockerell, 1894: 157. Type data: JAMAICA: Kingston, on ornamental shrub. Syntypes, female, by subsequent designation. Type depository: Washington: United States National Entomological Collection, U.S. National Museum of Natural History, District of Columbia, USA. accepted valid name; <i>Ceroplastes albolineatus vulcanicus</i> Cockerell, 1903: 160. Type data: MEXICO: Volcan de Colima, on low bush below pines. Syntypes, female, Type depository: London: The Natural History Museum, England, UK Washington: United States National Entomological Collection, U.S. National Museum of Natural History, District of Columbia, USA. accepted valid name	Mexico, Jamaica, Brazil (described from Jamaica)	1, 2
<i>Ceroplastes ceriferus</i> Fabricius, 1798	Coccidae Fallén, 1814	<i>Coccus ceriferus</i> Fabricius, 1798: 546. Type data: INDIA: Coromandel Coast, probably collected on <i>Maytenus emarginatus</i> . Syntypes, female, accepted valid name Notes: For details on the type material see De Lotto, 1971; <i>Coccus (Ceroplastes) chilensis</i> Gray, 1828: 7. Type data: CHILE: on branches and peduncles of unidentified tree. Syntypes, preadult female, Type depository: London: The Natural History Museum, England, UK. junior synonym (discovered by Green 1899: 191); <i>Ceroplastes ceriferus</i> (Fabricius, 1798); Walker 1852: 1087. change of combination; <i>Ceroplastes australiae</i> Walker, 1852: 1087. Type data: AUSTRALIA: Sydney. Syntypes, female, Type depository: London: The Natural History Museum, England, UK. junior synonym (discovered by Green 1899: 191); <i>Columnea cerifera</i> (Fabricius, 1798); Targioni Tozzetti 1866: 144. change of combination; <i>Columnea chilensis</i> (Gray, 1828); Targioni Tozzetti 1866: 145. change of combination; <i>Lacca alba</i> Signoret, 1869: 848. nomen nudum (discovered by Ben-Dov 1993, xxiii); <i>Ceroplastes ceriferus</i> Signoret 1869: 848. subsequent use Notes: Author incorrectly cited as Anderson; <i>Ceroplastes ceriferus</i> Signoret 1872: 40. subsequent use Notes: Author incorrectly cited as Anderson; <i>Ceroplastes australae</i> Green 1904: 375. Misidentification; <i>Seroplastes ceriferus</i> Seabra 1925: 32. misspelling of genus name; <i>Ceroplastes ceriferus</i> Kiritchenko 1932: 248. subsequent use Notes: Author incorrectly cited as Anderson; <i>Ceroplastes vayssierei</i> Mahdihassan, 1933: 561. Type depository: São Paulo: Museu de Zoologia, Universidade de São Paulo, Brazil. replacement name that is unjustified; <i>Gascardia cerifera</i> (Fabricius, 1798); De Lotto 1965: 198. change of	Global (described from India)	1, 2

		combination Notes: Author incorrectly cited; <i>Ceroplastes pseudoceriferus</i> Paik 1978. Misidentification; <i>Ceroplastes ceriferens</i> (Fabricius, 1798); Tao 1978: 79. misspelling of species epithet Notes: Author incorrectly cited; <i>Ceroplastes ceriferens</i> (Fabricius, 1798); Su 1982: 61. misspelling of species epithet Notes: Author incorrectly cited; <i>Ceroplastes cerifera</i> (Fabricius, 1798); Gill 1988: 18. emendation that is unjustified		
<i>Ceroplastes floridensis</i> Comstock, 1881	Coccidae Fallén, 1814	<i>Ceroplastes vinsonii</i> Signoret, 1872: 38. Type data: MAURITIUS: on <i>Eriobotrya japonica</i> . Syntypes, female, by subsequent designation junior synonym (discovered by Ben-Dov et al. 2000: 424). Notes: Type material probably lost. No type material was found in MNHW, by Ben-Dov and Matile-Ferrero, during a visit (December 1994) at the Vienna Museum of Natural History; <i>Ceroplastes floridensis</i> Comstock, 1881: 331. Type data: U.S.A.: Florida, Jacksonville, on Tangerine orange. Lectotype, female, by subsequent designation (Gimpel et al. 1974: 44). Type depository: Washington: United States National Entomological Collection, U.S. National Museum of Natural History, District of Columbia, USA. accepted valid name; <i>Ceroplastes floridensis</i> Comstock, 1881; Bodenheimer 1953: 112. misspelling of species epithet Notes: Erroneous citation of Maskell as author; <i>Cerostegia floridensis</i> (Comstock, 1881) De Lotto 1969: 211. change of combination; <i>Paracerostegia floridensis</i> (Comstock, 1881); Tang 1991: 306. change of combination; <i>Ceroplastes vinsoni</i> Signoret, 1872; Ben-Dov 1993: 59, 60, 535. misspelling of species epithet.	Global (described from Florida, USA)	1, 2
<i>Ceroplastes madagascariensis</i> Targioni Tozzetti, 1893	Coccidae Fallén, 1814	<i>Gascardia madagascariensis</i> Targioni Tozzetti, 1893: 88. Type data: MADAGASCAR: on an undetermined tree of the Lauraceae. Syntypes, female, Type depository: Paris: Muséum National d'Histoire naturelle, France. accepted valid name Notes: Type material probably lost; G. Pellizzari Scaltriti, 1990, personal communication to Yair Ben-Dov. Lectotype (designated in Hodgson & Peronti, 2012): South Africa: Left label: Coll. No. / Natal. (S. Africa) / Richmond / ex.: / Coll. Mr. Fuller; right label states: Lectotype (USDA); <i>Ceroplastes candela</i> Cockerell & King in Cockerell, 1902: 113. Type data: SOUTH AFRICA: Natal, Richmond. Syntypes, female, Type depository: Washington: United States National Entomological Collection, U.S. National Museum of Natural History, District of Columbia, USA. junior synonym (discovered by Hodgson & Peronti 2012: 76). Notes: Host plant of type material not given; <i>Ceroplastes combreti</i> Brain, 1920: 27. Type data: South Africa: De Wildt, Pretoria District, on <i>Combretum</i> sp. Syntypes, female, Type depository:	Madagascar, South Africa, Brazil (described from Madagascar)	1, 2

		Pretoria: South African National Collection of Insects, South Africa. junior synonym (discovered by Hodgson & Peronti 2012: 76); <i>Ceroplastes madagascariensis</i> (Targioni Tozzetti, 1893); MacGillivray 1921: 154. change of combination; <i>Gascardia combreti</i> (Brain, 1920); De Lotto 1970: 145. change of combination; <i>Ceroplastes candella</i> Cockerell & King in Cockerell, 1902; Ben-Dov 1993: 24. misspelling of species epithet		
<i>Ceroplastes pseudoceriferus</i> Green, 1935	Coccidae Fallén, 1814	<i>Ceroplastes ceriferus</i> ; Green 1921b: 259. Misidentification; <i>Ceroplastes pseudoceriferus</i> Green 1935: 180. Type data: SRI LANKA: on undetermined plant, and INDIA: on <i>Azidarachta indica</i> and <i>Diospyros montana</i> . Syntypes, female, Type depository: London: The Natural History Museum, England, UK; accepted valid name	Asia (described from Sri Lanka and India)	1, 2
<i>Ceroplastes rubens</i> Maskell, 1893	Coccidae Fallén, 1814	<i>Ceroplastes rubens</i> Maskell, 1893: 214. Type data: AUSTRALIA: Queensland, Brisbane, on <i>Mangifera indica</i> and <i>Ficus</i> sp. Syntypes, female, Type depository: Auckland: New Zealand Arthropod Collection, Landcare Research, New Zealand. accepted valid name; <i>Ceroplastes rubens minor</i> Maskell, 1897: 309. Type data: HONG KONG: on <i>Pinus sinensis</i> and <i>P. thunbergii</i> . Syntypes, female, Type depository: Washington: United States National Entomological Collection, U.S. National Museum of Natural History, District of Columbia, USA. junior synonym (discovered by Gimpel et al. 1974: 57); <i>Ceroplastes myricae</i> Green 1900: 8. Misidentification.	Global (described from Australia)	1, 2

References

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2. EPPO, *Journal*, 2024.

Table S6. Accepted name, family, species, synonyms, lifestyle, geographical distribution, and common environment of the selected microorganisms

Accepted genus name and protologue	Family	Species	Synonyms	Lifestyle	Geographical distribution	Environment	References
<i>Agrobacterium</i> Conn (1942) emend. Sawada et al. (1993)	Rhizobiaceae Conn (1938)	<i>Agrobacterium tumefaciens</i> Smith and Townsend (1907); Conn (1942)	Synonym: <i>Bacterium tumefaciens</i> Smith and Townsend (1907); <i>Pseudomonas tumefaciens</i> Smith and Townsend (1907), Duggar (1909); <i>Phytomonas tumefaciens</i> Smith and Townsend (1907), Bergey et al. (1923); <i>Polymonas tumefaciens</i> Smith and Townsend (1907), Lieske (1928)	Symbiont, parasite	Cosmopolitan	Dicotyledons, roots	¹
<i>Aleurodiscus</i> Rabenh., Hedwigia 13: 184 (1874)	Stereaceae Pilát, Hedwigia 70: 34 (1930)	<i>Gloeosoma mirabile</i> (Berk. & M.A. Curtis) Rajchenb., Pildain & C. Riquelme, Mycologia 113 (6): 1272 (2021)	Basionym: <i>Psilopezia mirabilis</i> Berk. & M.A.Curtis, Journal of the Linnean Society. Botany 10: 364 (1869); Synonyms: <i>Acanthophysium mirabile</i> (Berk. & M.A.Curtis) Parmasto, Eesti NSV Teaduste Akadeemia Toimetised 16: 378 (1967); <i>Aleurodiscus mirabilis</i> (Berk. & M.A.Curtis) Höhn., Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften Math.-naturw. Klasse Abt. I 118: 818 (1906); <i>Aleurodiscus alboroseus</i> Bres., Annales Mycologici 18 (1-3): 46 (1920); <i>Aleurodiscus apiculatus</i> Burt, Annals of the Missouri Botanical Garden 5: 186 (1918); <i>Aleurodiscus japonicus</i> Yasuda, Botanical Magazine Tokyo 33: 33 (1919); <i>Aleurodiscus pallideroseus</i> Litsch., Symb. sinica, II: 41 (1937); <i>Corticium peradeniae</i> Berk. & Broome, Journal of the Linnean Society. Botany 14: 69 (1875); <i>Aleurodiscus peradeniae</i> (Berk. & Broome) Henn., Monsunia 1: 139 (1900); <i>Aleurodiscus peradeniyae</i> Höhn. & Litsch., Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften Math.-naturw. Klasse Abt. I 117: 1096 (1908); <i>Terana peradeniae</i> (Berk. & Broome) Kuntze, Revisio generum plantarum 2: 872 (1891); <i>Aleurodiscus petelotii</i> Pat., Bulletin de la Société Mycologique de France 40: 31 (1924); <i>Aleurodiscus salmoneus</i> Pat., Bulletin du Muséum d'Histoire Naturelle Paris 30: 407 (1924); <i>Aleurodiscus sinensis</i> Teng & L. Ling, Contributions from the Biological Laboratory of the Science Society of China 8: 273 (1933); <i>Aleurodiscus spinulosus</i> Henn., Botanische Jahrbücher für Systematik Pflanzengeschichte und Pflanzengeographie 38: 107 (1905); <i>Corticium</i>	Parasite	Cosmopolitan	Wood, roots	²

			<i>spinulosum</i> (Henn.) Sacc. & Trotter, Sylloge Fungorum 21: 403 (1912); <i>Aleurodiscus usambarensis</i> Henn., Botanische Jahrbücher für Systematik Pflanzengeschichte und Pflanzengeographie 34: 43 (1904); <i>Corticium usambarensis</i> (Henn.) Sacc. & D. Sacc., Sylloge Fungorum 17: 168 (1905)				
<i>Alternaria</i> Nees, System der Pilze und Schwämme: 72 (1817)	Pleosporaceae Nitschke, Verh. Naturhist. Vereines Preuss. Rheinl.: 74 (1869)	<i>Alternaria alternata</i> (Fr.) Keissl., Beihefte zum Botanischen Centralblatt 29: 433 (1912)	Basionym: <i>Torula alternata</i> Fr., Systema Mycologicum 3: 500 (1832). Obligate or homotypic synonyms: <i>Alternaria tenuis</i> Nees, System der Pilze und Schwämme: 68, t. 5:68 (1817); <i>Torula alternata</i> Fr., Systema Mycologicum 3: 500 (1832). Facultative or heterotypic synonyms: <i>Macrosporium erumpens</i> Cooke, Grevillea 12 (61): 32 (1883); <i>Macrosporium fasciculatum</i> Cooke & Ellis, Grevillea 6 (37): 6 (1877); <i>Macrosporium meliloti</i> Peck, Annual Report on the New York State Museum of Natural History 33: 26 (1883); <i>Macrosporium polytrichi</i> Peck, Annual Report on the New York State Museum of Natural History 43: 77 (1890); <i>Macrosporium seguierii</i> Allesch., Hedwigia 33: 75 (1894) Synonymy: <i>Ulocladium</i> Preuss, Linnaea 24: 111 (1851). Obligate or homotypic synonyms: <i>Ulocladium</i> Preuss, Linnaea 24: 111 (1851)	Biotroph, mycorrhizae, saprotroph	Cosmopolitan	Fruits, leaves, roots	3
<i>Amycolatopsis</i> Lechevalier et al. (1986)	Pseudonocardiaceae Embley et al. (1989)	<i>Amycolatopsis tolypomycina</i> Wink et al. (2003)	Synonym: <i>Streptomyces tolypophorus</i> Shibata et al. (1971); <i>Amycolatopsis tolypomycina</i> Wink et al. (2003)	Saprotroph	Cosmopolitan	Soil	4
<i>Aplosporella</i> Speg., Anales Soc. Ci. Argent. 10 (4): 157 (1880)	Aplosporellaceae Slippers, Boissin & Crous, fam. nov., Studies in Mycology 76: 41 (2013)	<i>Aplosporella javeedii</i> Jami, Slippers, M.J.Wingf. & Gryzenh., Fungal Biology 118 (2): 174 (2013)	Synonym: <i>Aplosporella javeedii</i> Jami, Slippers, M.J.Wingf. & Gryzenh., Fungal Biology 118 (2): 174 (2013)	Parasite, necrotroph	China	Wood	5
<i>Arthrinium</i> Kunze, Mykologische Hefte 1: 9 (1817)	Apiosporaceae K.D.Hyde, J.Fröhl., J.E.Taylor & M.E.Barr, Sydowia 50 (1): 23 (1998)	<i>Arthrinium</i> Kunze, Mykologische Hefte 1: 9 (1817)	Basionym: <i>Arthrinium</i> Kunze, Mykologische Hefte 1: 9 (1817)	Endophyte, saprotroph	Cosmopolitan	Air, plants, soil, marine sponge	6-9

<i>Aspergillus</i> P.Micheli ex Haller, Historia stirpium indigenarum Helvetiae inchoata: 113 (1768)	Eurotiomycetida e Geiser & Lutzoni, Mycological Research 111 (5): 528 (2007)	<i>Aspergillus</i> <i>aurantiobrunneus</i> (G.A.Atkins, Hindson & A.B.Russell) Raper & Fennell, The Genus <i>Aspergillus</i> : 511 (1965)	Synonym: <i>Emericella nidulans</i> var. <i>aurantiobrunnea</i> G.A.Atkins, Hindson & A.B.Russell, Transactions of the British Mycological Society 41 (4): 504 (1958); <i>Aspergillus aurantiobrunneus</i> (G.A.Atkins, Hindson & A.B.Russell) Raper & Fennell, The Genus <i>Aspergillus</i> : 511 (1965); Basionym: <i>Emericella nidulans</i> var. <i>aurantiobrunnea</i> G.A.Atkins, Hindson & A.B.Russell, Transactions of the British Mycological Society 41 (4): 504 (1958); Obligate or homotypic synonyms: <i>Aspergillus aurantiobrunneus</i> (G.A.Atkins, Hindson & A.B.Russell) Raper & Fennell, The Genus <i>Aspergillus</i> : 511 (1965), <i>Emericella nidulans</i> var. <i>aurantiobrunnea</i> G.A.Atkins, Hindson & A.B.Russell, Transactions of the British Mycological Society 41 (4): 504 (1958)	Saprotroph, lichenized, mycorrhizae, biotroph	Cosmopolitan	Rot, seed, leaf surface	10-13
		<i>Aspergillus</i> <i>calidoustus</i> Varga, Houbraken & Samson, Eukaryotic Cell 7 (4): 636 (2008)	Basionym: <i>Aspergillus calidoustus</i> Varga, Houbraken & Samson, Eukaryotic Cell 7 (4): 636 (2008)	Parasite, necrotroph, saprotroph	Cosmopolitan	Wood, human pathogen	14-16
		<i>Aspergillus</i> <i>clavatus</i> Desm., Ann. Sci. Nat., Bot. 2: 71 (1834)	Basionym: <i>Aspergillus clavatus</i> Desm., Ann. Sci. Nat., Bot. 2: 71 (1834)	Saprotroph	Cosmopolitan	Soil	14-16
		<i>Aspergillus</i> <i>egyptiacus</i> Moub. & Moustafa, Egypt. J. Bot.: 153 (1972)	Synonyms: <i>Aspergillus egyptianus</i> Moub. & Moustafa (1972)	Saprotroph, parasite	Cosmopolitan	Sandy soil, decaying vegetation	17
		<i>Aspergillus</i> <i>fischeri</i> Wehmer, Centbl. Bakt. ParasitKde, Abt. I 18: 390 (1907)	Synonyms: <i>Aspergillus fischeri</i> var. <i>glaber</i> Fennell & Raper, Mycologia 47(1): 74 (1955); <i>Aspergillus</i> <i>fischeri</i> var. <i>spinosus</i> Raper & Fennell, The Genus <i>Aspergillus</i> : 256 (1965); <i>Aspergillus fischeri</i> var. <i>thermomutatus</i> Paden, Mycopath. Mycol. appl. 36: 161 (1968); <i>Aspergillus fischeri</i> var. <i>verrucosus</i> Udagawa & H. Kawas., Trans. Mycol. Soc. Japan 8(3): 118 (1968); <i>Aspergillus thermomutatus</i> (Paden) S.W. Peterson, Mycol. Res. 96(7): 549 (1992); <i>Neosartorya</i> <i>fischeri</i> (Wehmer) Malloch & Cain, Can. J. Bot. 50: 2620 (1973); <i>Neosartorya fischeri</i> var. <i>glabra</i> Fennell	Parasite	Cosmopolitan	Humans	18

	& Raper, Can. J. Bot. 50(12): 2621 (1973) [1972]; <i>Neosartorya fischeri</i> var. <i>spinosa</i> Raper & Fennell, The Genus <i>Aspergillus</i>: 256 (1965); <i>Neosartorya fischeri</i> var. <i>verrucosa</i> (Udagawa & H. Kawas.) Malloch & Cain, Can. J. Bot. 50(12): 2621 (1973) [1972]; <i>Neosartorya glabra</i> (Fennell & Raper) Kozak., Mycol. Pap. 161: 56 (1989); <i>Neosartorya pseudofischeri</i> S.W. Peterson, Mycol. Res. 96(7): 549 (1992); <i>Neosartorya spinosa</i> (Raper & Fennell) Kozak., Mycol. Pap. 161: 58 (1989); <i>Sartorya fumigata</i> var. <i>glabra</i> (Fennell & Raper) Udagawa & H. Kawas., Trans. Mycol. Soc. Japan 8(3): 115 (1968); <i>Sartorya fumigata</i> var. <i>spinosa</i> (Raper & Fennell) Udagawa & H. Kawas., Trans. Mycol. Soc. Japan 8(3): 117 (1968); <i>Sartorya fumigata</i> var. <i>verrucosa</i> Udagawa & H. Kawas., Trans. Mycol. Soc. Japan 8(3): 118 (1968)				
<i>Aspergillus flavus</i> var. <i>oryzae</i> (Ahlb.) Kurtzman, M.J.Smiley, Robnett & Wicklow, Mycologia 78 (6): 957 (1986)	Basionym: <i>Eurotium oryzae</i> Ahlb., Dingler's Polytechn. J.: 330 (1878) Synonyms: <i>Aspergillus oryzae</i> (Ahlb.) Cohn, Jahresber. Schles. Ges. Vaterl. Cult. 61: 226 (1884) Basionym	Saprotroph	Cosmopolitan, hot and humid climate	Rots, decaying vegetation	19-23
<i>Aspergillus flocculosus</i> Frisvad & Samson, Studies in Mycology 50 (1): 33 (2004)	Basionym: <i>Aspergillus flocculosus</i> Frisvad & Samson, Stud. Mycol. 50 (1): 33 (2004)	Saprotroph	Venezuela, Slovenia, Greece, Costa Rica, Brazil	Decaying vegetation	24
<i>Aspergillus insuetus</i> (Bainier) Thom & Church, Manual of the Aspergilli: 153 (1929)	Basionym: <i>Sterigmatocystis insueta</i> Bainier, Bulletin de la Société Mycologique de France 24: 85 (1908). Obligate or homotypic synonyms: <i>Sterigmatocystis insueta</i> Bainier, Bulletin de la Société Mycologique de France 24: 85 (1908)	Saprotroph, parasite	Cosmopolitan	Wood, soil, <i>Theobroma cacao</i>	14
<i>Aspergillus nidulans</i> (Eidam) G.Winter,	Basionym: <i>Sterigmatocystis nidulans</i> Eidam, Beiträge zur Biologie der Pflanzen 3: 392 (1883). Obligate or homotypic synonyms: <i>Aspergillus nidulans</i> mut. <i>nidulans</i> (Eidam) G. Winter: 62 (1884);	Saprotroph	Cosmopolitan	Fruits, leaf, seed	25

Rabenhorst's Kryptogamen-Flora, Pilze - Ascomyceten 1(2): 62 (1884)	<i>Diplostephanus nidulans</i> (Eidam) Neveu-Lem., Précis de Parasitologie Humaine: Parasites Végétaux et Animaux: 101 (1921); <i>Emericella nidulans</i> (Eidam) Vuill., Comptes Rendues des Séances Hebdomadaires de l'Académie des Sciences Paris 184: 137 (1927); <i>Sterigmatocystis nidulans</i> Eidam, Beiträge zur Biologie der Pflanzen 3: 392 (1883). Teleomorph synonyms: <i>Emericella nidulans</i> (Eidam) Vuill., Comptes Rendues des Séances Hebdomadaires de l'Académie des Sciences Paris 184: 137 (1927). Facultative or heterotypic synonyms: <i>Aspergillus nidulans</i> var. <i>cesarii</i> Pinoy, Bull. Soc. Pathol. Exot.: 11 (1915); <i>Aspergillus nidulans</i> var. <i>dentatus</i> D.K.Sandhu & R.S.Sandhu, Mycologia 55 (3): 297 (1963); <i>Aspergillus nidulellus</i> Samson & W.Gams, Advances in Penicillium and Aspergillus Systematics: 44 (1986); <i>Emericella dentata</i> (D.K.Sandhu & R.S.Sandhu) Y.Horie, Transactions of the Mycological Society of Japan 21: 491 (1980); <i>Emericella nidulans</i> var. <i>dentata</i> (D.K.Sandhu & R.S.Sandhu) Subram., Current Science 41: 758 (1972); <i>Sterigmatocystis nidulans</i> var. <i>nicollei</i> Pinoy, Arch. Parasitol.: 437 (1906)				
<i>Aspergillus purpureus</i> Samson & Mouch., Antonie van Leeuwenhoek 41 (3): 350 (1975)	Synonyms: <i>Emericella purpurea</i> Samson & Mouch., Antonie van Leeuwenhoek 41 (3): 350 (1975) Taxon synonyms	Saprotroph	Egypt	Desert soil, Western Desert	26, 27
<i>Aspergillus spectabilis</i> M.Chr & Raper, Mycologia 70: 333 (1978)	Basionym: <i>Aspergillus spectabilis</i> M.Chr & Raper, Mycologia 70: 333 (1978) Synonyms: <i>Emericella spectabilis</i> M.Chr & Raper, Mycologia 70: 333 (1978)	Saprotroph	Cosmopolitan	Soil	28
<i>Aspergillus stellatus</i> Curzi, Rendic. Accad. naz. Lincei: 428 (1934)	Facultative or heterotypic synonyms: <i>Aspergillus variegator</i> Thom & Raper, Mycologia 31 (6): 663 (1939); <i>Aspergillus variegator</i> var. <i>major</i> Bat. & H.Maia, Anais da Sociedade de Biologia de Pernambuco 15 (1): 235 (1957); <i>Emericella variegator</i> var. <i>variegator</i> (?); <i>Inzengaea asterospora</i> Borzí (?); <i>Inzengaea erythrospora</i> Borzí, Jahrbücher für Wissenschaftliche Botanik 16: 450 (1885)	Saprotroph, parasite	Cosmopolitan	Decaying leaves	29

		<i>Aspergillus terreus</i> Thom, American Journal of Botany 5 (2): 85 (1918)	Synonyms: <i>Aspergillus terrestris</i> Thom, American Journal of Botany 5 (2): 85 (1918)	Saprotroph	Cosmopolitan	Numerous plant substrates	30
		<i>Aspergillus unguis</i> (Émile-Weill & L.Gaudin) Dodge, Medical mycology. Fungous diseases of men and other mammals: 637 (1935)	Basionym: <i>Sterigmatocystis unguis</i> Émile-Weill & L.Gaudin, Arch. Med. Exp. Anat. Pathol.: 463 (1918). Obligate or homotypic synonyms: <i>Sterigmatocystis unguis</i> Émile-Weill & L.Gaudin, Arch. Med. Exp. Anat. Pathol.: 463 (1918). Teleomorph synonyms: <i>Emericella unguis</i> Malloch & Cain, Canadian Journal of Botany 50 (1): 62 (1972). Facultative or heterotypic synonyms: <i>Aspergillus laokiashanensis</i> Y.K.Shih, Lingnan Sci. J.: 369 (1936); <i>Aspergillus mellinus</i> Novobr., Novosti Sistematiki Nizshikh Rastenii 9: 171 (1972)	Saprotroph	Cosmopolitan	Blossoms, corn, fruit	31
		<i>Aspergillus undulatus</i> H.Z. Kong & Z.T. Qi, Acta Mycol. Sin.: 211 (1985)	Basionym: <i>Aspergillus undulatus</i> H.Z. Kong & Z.T. Qi, Acta Mycol. Sin.: 211 (1985). Taxonomic synonym(s): <i>Emericella undulata</i> H.Z. Kong & Z.T. Qi, Acta mycol. sin.: 211 (1986)	saprotroph	China, Germany	Leaves, woods	32
		<i>Aspergillus ustus</i> (Bainier) Thom & Church, The Aspergilli: 152 (1926)	Basionym: <i>Sterigmatocystis usta</i> Bainier, Bulletin de la Société Botanique de France 28: 78 (1882). Obligate or homotypic synonyms: <i>Sterigmatocystis usta</i> Bainier, Bulletin de la Société Botanique de France 28: 78 (1882); <i>Sterigmatocystis ustus</i> Bainier (1881). Facultative or heterotypic synonyms: <i>Aspergillus minutus</i> Abbott, Iowa State Coll. Journ. Sci. (1927)	Saprotroph	Cosmopolitan, mo common in subtropical and tropical regions	Nut, leaf, corn	33
	<i>Bipolaris</i> Shoemaker, Canadian Journal of Botany 37 (5): 882 (1959)	<i>Bipolaris eleusines</i> (Alcorn & R.G. Shivas, 1990)	Basionym: <i>Bipolaris crotonis</i> Sivan., Transactions of the British Mycological Society 84: 404 (1985). <i>Bipolaris eleusines</i> Alcorn & R.G. Shivas, Mycotaxon 39: 367 (1990). <i>Cochliobolus eleusines</i> Alcorn, Mycotaxon 39: 367 (1990)	Saprotroph, parasite	Cosmopolitan	Potato	34
		<i>Bipolaris leersiae</i> (G.F. Atk.) Shoemaker, Canadian Journal of	Basionym: <i>Helminthosporium leersiae</i> G.F.Atk., Bulletin of the Cornell University (Science) 3 (1): 47 (1897). Obligate or homotypic synonyms: <i>Bipolaris leersii</i> (G.F.Atk.) Shoemaker (1959); <i>Drechslera leersiae</i> (G.F.Atk.) Subram. & B.L.Jain, Current Science 35 (14): 354 (1966); <i>Drechslera leersii</i> (G.F. Atk.) Subram. & B.L.Jain (1966); <i>Helminthosporium</i>	Saprotroph, parasite	Indopacif	Leaves, wood	35, 36

Botany 37 (5): 883 (1959)	<i>leersiae</i> G.F.Atk., Bulletin of the Cornell University (Science) 3 (1): 47 (1897); <i>Helminthosporium leersii</i> G.F.Atk. (1897)				
<i>Bipolaris maydis</i> (Y.Nisik. & C.Miyake) Shoemaker, Canadian Journal of Botany 37 (5): 882 (1959)	Basionym: <i>Helminthosporium maydis</i> Y.Nisik. & C.Miyake, Ber. Ohara Inst. landw. Biol.: 243 (1926). Obligat or homotypic synonyms: <i>Drechslera maydis</i> (Y.Nisik. & C.Miyake) Subram. & B.L.Jain, Current Science 35 (14): 354 (1966); <i>Helminthosporium maydis</i> Y.Nisik. & C.Miyake, Ber. Ohara Inst. landw. Biol.: 243 (1926). Teleomorph synonyms: <i>Cochliobolus heterostrophus</i> (Drechsler) Drechsler, Phytopathology 24: 973 (1934); <i>Ophiobolus heterostrophus</i> Drechsler, Journal of Agricultural Research 31: 701 (1925).	Saprotroph, lichen, biotroph,	Warm temperate, subtropical, and tropical regions, Indopacific	Living leaves. Multiple hosts but primarily Poaceae	35, 37, 38
<i>Bipolaris oryzae</i> (Breda de Haan) Shoemaker, Canadian Journal of Botany 37 (5): 883 (1959)	Synonymy: <i>Helminthosporium oryzae</i> Breda de Haan, Bulletin de l'Institut Botanique de Buitenzorg 6: 11 (1900); <i>Drechslera oryzae</i> (Breda de Haan) Subram. & B.L.Jain, Current Science 35 (14): 354 (1966); <i>Luttrellia oryzae</i> (Breda de Haan) Gornostai, Vodorosli, Griby i Mkhii Dal'nego Vostoka: 81 (1978); <i>Helminthosporium macrocarpum</i> Grev., Scott. crypt. fl. (Edinburgh): pl. 148 (1825); <i>Cochliobolus</i> Drechsler, Phytopathology 24: 973 (1934)	Saprotroph, parasite	Indopacific	Leaves, glumes, coleoptile leaf sheaths, panicle branches, roots of young seedlings, and stems	39-47
<i>Bipolaris panici- miliacei</i> (Y.Nisik.) Shoemaker, Canadian Journal of Botany 37 (5): 884 (1959)	Basionym: <i>Helminthosporium panici-miliacei</i> Y.Nisik., Spec. Rep. Ohara Inst. Agric. Res.: 120 (1929). Obligat or homotypic synonyms: <i>Drechslera panici-miliacei</i> (Y.Nisik.) Subram. & B.L. Jain, Current Science 35 (14): 354 (1966); <i>Helminthosporium panici-miliacei</i> Y.Nisik., Spec. Rep. Ohara Inst. Agric. Res.: 120 (1929)	Parasite	Cosmopolitan	Leaves, roots, stems	35
<i>Bipolaris setariae</i> Shoemaker, Canadian Journal of Botany 37 (5): 884 (1959)	Obligat or homotypic synonyms: <i>Drechslera setariae</i> (Sawada) S.Ito, Proceedings of the Imperial Academy Japan 6 (8): 355 (1930); <i>Helminthosporium setariae</i> Sawada, Report of the Department of Agriculture Government Research Institute of Formosa 19: 656 (1919). Teleomorph synonyms: <i>Cochliobolus setariae</i> (S.Ito & Kurib.) Drechsler ex Dastur, Indian Journal of Agricultural Sciences 12: 733 (1942); <i>Ophiobolus setariae</i> S. Ito & Kurib., Proceedings of the Imperial Academy Japan 6 (8): 352 (1930).	Saprotroph, parasite	Indopacific	Leaves, stems	48 49
<i>Bipolaris sorghicola</i>	Basionym: <i>Helminthosporium sorghicola</i> Lefebvre & Sherwin, Mycologia 40 (6): 714 (1948). Obligat or	Saprotroph, parasite	Cosmopolitan	Leaves	38, 50

		(Lefebvre & Sherwin) Alcorn, Mycotaxon 17: 69 (1983)	homotypic synonyms: <i>Drechslera sorghicola</i> (Lefebvre & Sherwin) M.J.Richardson & E.M.Fraser, Transactions of the British Mycological Society 51 (1): 148 (1968); <i>Helminthosporium sorghicola</i> Lefebvre & Sherwin, Mycologia 40 (6): 714 (1948); <i>Bipolaris cookei</i> (Sacc.) Shoemaker, Canadian Journal of Botany 37 (5): 884 (1959)				
		<i>Bipolaris sorokiniana</i> (Sacc.) Shoemaker, Canadian Journal of Botany 37 (5): 884 (1959)	Basionym: <i>Helminthosporium sorokinianum</i> Sacc.: 238 (1891). Obligate or homotypic synonyms: <i>Drechslera sorokiniana</i> (Sacc.) Subram. & B.L.Jain, Transactions of the British Mycological Society 47 (4): 613 (1964); <i>Helminthosporium sorokinianum</i> Sacc.: 238 (1891). Teleomorph synonyms: <i>Cochliobolus sativus</i> (S.Ito & Kurib) Drechsler ex Dastur, Indian Journal of Agricultural Sciences 12: 733 (1942); <i>Ophiobolus sativus</i> S.Ito & Kurib, Trans. Sapporo nat. Hist. Soc.: 138 (1929). Facultative or heterotypic synonyms: <i>Helminthosporium sativum</i> Pammel, C.M. King & Bakke, Bulletin of Iowa State College: 180 (1910).	Saprotroph, parasite	Cosmopolitan	Leaves, seedlings, roots	51 52
<i>Cephalosporium</i> Corda, Icones fungorum hucusque cognitorum 3: 11 (1839)	Hypocreomycetidae O.E.Erikss. & Winka, Myconet 1: 6 (1997)	<i>Cephalosporium caerulens</i> Matsumae, Kamio & Hata, The Journal of Antibiotics 16: 236 (1963)	Basionym: <i>Cephalosporium caerulens</i> Matsumae, Kamio & Hata, The Journal of Antibiotics 16: 236 (1963)	Saprotroph	Cosmopolitan	Leaves	53-55
<i>Chaetomium</i> Kunze, Mykologische Hefte 1: 16 (1817)	Chaetomiaceae G. Winter, 1885	<i>Chaetomium globosum</i> Kunze, Mykologische Hefte 1: 16 (1817)	Basionym: <i>Chaetomium globosum</i> Kunze, Mykologische Hefte 1: 16 (1817). Taxonomic synonym(s): <i>Chaetomidium barbatum</i> Traaen, Nytt Magazin for Naturvidenskapene 52: 19 (1914); <i>Chaetomium fibripilium</i> L.M. Ames, Mycologia 42 (5): 642 (1950); <i>Chaetomium mollipilium</i> L.M. Ames, Mycologia 42 (5): 644 (1950); <i>Chaetomium ochraceum</i> Tschudy, American Journal of Botany 24: 472 (1937); <i>Chaetomium olivaceum</i> Cooke & Ellis, Grevillea 6 (39): 96, Pl. 100, fig. 38 (1878); <i>Chaetomium rectum</i> Sergeeva, Not. syst. Pl. non-vasc.: 143 (1961); <i>Chaetomium spiculipilium</i> L.M. Ames, A monograph of the Chaetomiaceae Ser. 2: 37 (1963); <i>Chaetomium subglobosum</i> Sergeeva, Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk S.S.S.R. 13: 172 (1960); <i>Chaetomium</i>	Saprotroph	Cosmopolitan	Woods	56

			<i>subterraneum</i> Swift & Povah, Mycologia 21 (4): 210 (1929); <i>Chaetomium lusitanicum</i> M.R.M. Gomes, Estudos e Informação: 3 (1953); <i>Chaetomium affine</i> Corda, Icones fungorum hucusque cognitorum 4: 37, tab. 8, fig. 101 (1840); <i>Chaetomium globosum</i> var. <i>affine</i> Tschudy; <i>Chaetomium kunzeanum</i> Zopf (1881); <i>Chaetomium kunzeanum</i> var. <i>chlorina</i> Sacc.; <i>Chaetomium spirale</i> Zopf, Nova Acta Academiae Caesareae Leopoldino-Carolinae Germanicae Naturae Curiosorum 42 (5): 275 (1881)				
<i>Codinaea</i> Maire, Publicacions del Instituto Botánico Barcelona 3 (4): 15 (1937)	Chaetosphaeriaceae Réblová, M.E.Barr & Samuels, Sydowia 51: 56 (1999)	<i>Codinaea simplex</i> S. Hughes & W.B.Kendr., New Zealand Journal of Botany 6: 362 (1968)	Basionym: <i>Tainosphaeria simplex</i> (S.Hughes & W.B.Kendr.) Réblová & Hern.-Restr., Journal of Fungi 7 (12, no. 1097): 85 (2021); <i>Dictyochaeta simplex</i> (S.Hughes & W.B.Kendr.) Hol.-Jech., Folia Geobot. Phytotax. 19 (4): 434 (1984)	Saprotroph	Cosmopolitan, mostly on tropical regions	Soil	57
<i>Cunninghamella</i> Matr., Ann. Mycol. 1: 47 (1903)	Cunninghamellaceae Naumov ex R.K.Benj, Aliso 4: 415 (1959)	<i>Cunninghamella echinulata</i> (Thaxt.) Thaxt. ex Blakeslee, Bot. Gaz. (London) 40: 161 (1905)	Basionym: <i>Oedocephalum echinulatum</i> Thaxt., Botanical Gazette Crawfordsville 16: 17 (1891); Synonyms: <i>Cunninghamella echinulata</i> (Thaxt.) Thaxt., Rhodora 5: 98 (1903); <i>Cunninghamella bainieri</i> Naumov, Encyclopédie Mycologique 9: 107 (1939); <i>Cunninghamella dalmatica</i> Pispék, Acta Botanica Instituti Botanici Regalis Universitatis Zagrebensis 4: 100 (1929); <i>Cunninghamella echinata</i> Pispék, Acta Botanica Instituti Botanici Regalis Universitatis Zagrebensis 4: 101 (1929); <i>Muratella elegans</i> Bainier & Sartory, Bulletin de la Société Mycologique de France 29: 129 (1913); <i>Cunninghamella verticillata</i> F.S.Paine, Mycologia 19: 253 (1927); <i>Cunninghamella blakesleeana</i> var. <i>verticillata</i> (F.S.Paine) Baijal & B.S.Mehrotra, Sydowia 33: 10 (1980); <i>Cunninghamella echinulata</i> var. <i>verticillata</i> (F.S.Paine) R.Y.Zheng & G.Q.Chen, Mycosystema: 7 (1996)	Saprtroph	Temperate regions	Soil	58
<i>Cytospora</i> Ehrenb. Ex Fries, 1823	Valsaceae Tul. & C.Tul., 1861	<i>Cytospora rhizophorae</i> Kohlm. & E.Kohlm., Mycologia 63(4): 847 (1971).		Saprotroph	Guatemala	Roots	59

<i>Diaporthe</i> Nitschke, Pyrenomyces Germanici 2: 240 (1870)	Diaporthaceae Höhn. ex Wehm., American Journal of Botany 13: 638 (1926)	<i>Diaporthe</i> <i>amygdali</i> (Delacroix) Udayanga, P.W.Crous & K.D.Hyde, Fungal Diversity 56: 166 (2012)	Basionym: <i>Fusicoccum amygdali</i> Delacr., Bulletin de la Société Mycologique de France 21: 182 (1905). Obligate or homotypic synonyms: <i>Fusicoccum</i> <i>amygdali</i> Delacr., Bulletin de la Société Mycologique de France 21: 182 (1905); <i>Phomopsis amygdali</i> (Delacr.) J.J. Tuset & M.T. Portilla, Canadian Journal of Botany 67 (5): 1280 (1989)	Parasite	Indopacific	Fruits, wood	60, 61
<i>Drechslera</i> S.Ito, Proceedings of the Imperial Academy Japan 6 (8): 355 (1930)	Pleosporaceae Nitschke, Verh. Naturhist. Vereines Preuss. Rheinl.: 74 (1869)	<i>Drechslera</i> <i>gigantea</i> S.Ito, Proceedings of the Imperial Academy Japan 6 (8): 355 (1930)	Basionym: <i>Helminthosporium giganteum</i> Heald & F.A. Wolf, Mycologia 3 (1): 21 (1911). Obligate or homotypic synonyms: <i>Helminthosporium</i> Heald & F.A.Wolf, Bot. Misc. 11 (1): 223 + pl. 12, fig. 5 (1911); <i>Helminthosporium giganteum</i> Heald & F.A.Wolf, Mycologia 3 (1): 21 (1911)	Saprotroph, lichen, biotroph, mycorrhizae	South Pacific	Fruits, leaves	62, 63
<i>Everniastrum</i> Hale ex Sipman, Mycotaxon 26: 237 (1986)	Parmeliaceae Eschw., Systema Lichenum, Genera Exhibens rite distincta, Pluribus Novis Adaucta: 19 (1824)	<i>Everniastrum</i> <i>rhizodendroideu</i> <i>m</i> (J.C.Wei & Y.M.Jiang) Sipman, Mycotaxon 26: 242 (1986)	Basionym: <i>Hypotrachyna rhizodendroidea</i> (J.C.Wei & Y.M.Jiang) Divakar, A.Crespo, Sipman, Elix & Lumbsch, Phytotaxa 132 (1): 32 (2013); <i>Cetrariastrum rhizodendroideum</i> J.C.Wei & Y.M.Jiang, Acta Phytotaxonomica Sinica 20: 496 (1982)	Saprotroph	China	Roots, soil	64
<i>Escherichia</i> Castellani and Chalmers 1919	Enterobacteriaceae Rahn 1937, Ewing et al. 1980, Adeolu et al. 2016	<i>Escherichia coli</i> Castellani and Chalmers 1919	Homotypic synonym: <i>Bacillus coli</i> (Migula 1895), <i>Bacterium coli</i> (Migula 1895), Lehmann and Neumann 1896, <i>Enterococcus coli</i> , <i>Achromobacter</i> sp., <i>Bacterium coli</i> commune (Escherich 1885)	Saprotroph, Parasite	Cosmopolitan	Water, soil	65
<i>Exserohilum</i> K.J.Leonard & Suggs, Mycologia 66: 289 (1974)	Pleosporaceae Nitschke, Verh. Naturhist. Vereines Preuss. Rheinl.: 74 (1869)	<i>Exserohilum</i> <i>turcicum</i> (Pass.) K.J.Leonard & Suggs, Mycologia 66: 291 (1974)	Basionym: <i>Helminthosporium turcicum</i> Pass., Boln Comiz. Agr. Parmense: 3 (1876). Obligate or homotypic synonyms: <i>Bipolaris turcica</i> (Pass.) Shoemaker, Canadian Journal of Botany 37 (5): 884 (1959); <i>Drechslera turcica</i> (Pass.) Subram. & B.L.Jain, Current Science 35 (14): 355 (1966); <i>Helminthosporium turcicum</i> Pass., Boln Comiz. Agr. Parmense: 3 (1876); <i>Luttrellia turcica</i> (Pass.) Khokhr., Vodorosli, Griby i Mkhii Dal'nego Vostoka: 81 (1978). Teleomorph synonyms: <i>Keissleriella turcica</i> (Luttr.) Arx, The genera of fungi sporulating in pure culture: 126 (1970); <i>Setosphaeria turcica</i> (Luttr.) K.J.Leonard & Suggs, Mycologia 66: 295 (1974);	Parasite	Cosmopolitan	Leaves	66, 67

			<i>Trichometasphaeria turcica</i> Luttr., Phytopathology 48: 282 (1958). Facultative or heterotypic synonyms: <i>Helminthosporium inconspicuum</i> Cooke & Ellis, Grevillea 6 (39): 88 (1878); <i>Keissleriella turcica</i> (Luttr.) Arx, The genera of fungi sporulating in pure culture: 126 (1970); <i>Setosphaeria turcica</i> (Luttr.) K.J.Leonard & Suggs, Mycologia 66: 295 (1974); <i>Trichometasphaeria turcica</i> Luttr., Phytopathology 48: 282 (1958)				
<i>Fusarium</i> Link, Magazin der Gesellschaft Naturforschenden Freunde Berlin 3 (1): 10 (1809)	Nectriaceae Tul. & C.Tul., Selecta Fungorum Carpologia: Nectriei- Phacidiei- Pezizei 3: 3 (1865)	<i>Fusarium</i> <i>fujikuroi</i> Nirenberg, Mitt. Biol. Bundesanst. Land- Forstw. 169: 32 (1976)	Basionym: <i>Fusarium celosiae</i> Abe, Memoirs of the College of Agriculture Kyoto University 7 (4): 51 (1928); <i>Gibberella fujikuroi</i> var. <i>intermedia</i> Kuhlman, Mycologia 74: 766 (1982); <i>Lisea fujikuroi</i> Sawada, Report of the Department of Agriculture Government Research Institute of Formosa 19: 251 (1919); <i>Gibberella fujikuroi</i> (Sawada) Wollenw., Zeitschrift für Parasitenkunde 3: 514 (1931); <i>Gibberella fujikuroi</i> f. <i>oryzae</i> Saccas, Rev. Path. Vég.: 77 (1951)	Parasite	Cosmopolitan	Soil, seeds	9
		<i>Fusarium</i> <i>heterosporum</i> Nees & T.Nees, Nova Acta Physico-Medica Academiae Caesareae Leopoldino- Carolinae Naturae Curiosorum 9 (1): 235 (1818)	Basionym: <i>Fusarium heterosporum</i> Nees & T.Nees Nova Acta Phys.-Med. Acad. Caes. Leop.-Carol. Nat. Cur. 9: 235 (1818). Teleomorph synonyms: <i>Botryosphaeria cyanea</i> (Sollm.) Weese, Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften Math.-naturw. Klasse Abt. I 128: 707 (1919); <i>Gibberella cyanea</i> (Sollm.) Wollenw., Fusaria Autographice Delineata 1: no. 39 (1916); <i>Gibberella gordonii</i> C. Booth, The genus <i>Fusarium</i> : 177 (1971); <i>Sphaeria cyanea</i> Sollm., Bot. Ztg.: 193 (1863). Facultative or heterotypic synonyms: <i>Fusisporium lolii</i> Wm.G. Sm., Diseases of field and garden crops, chiefly as are caused by fungi: 213 (1884).	Biotroph	North America; Europe; Africa; Australia; India	Soil, seeds	68
		<i>Fusarium</i> <i>proliferatum</i> (Matsush.) Nirenberg ex Gerlach & Nirenberg, Mitteilungen der Biologischen	Basionym: <i>Cephalosporium proliferatum</i> Matsush., Microfungi of the Solomon Islands and Papua-New Guinea: 11 (1971). Obligate or homotypic synonyms: <i>Cephalosporium proliferatum</i> Matsush., Microfungi of the Solomon Islands and Papua-New Guinea: 11 (1971); <i>Fusarium proliferatum</i> (Matsush.) Nirenberg, Mitteilungen der Biologischen Bundesanstalt für Land- und Forstwirtschaft 169: 38 (1976)	Parasite	Cosmopolitan	Leaves, roots, fruit	69-72

Bundesanstalt für Land- und Forstwirtschaft 209: 309 (1982) <i>Fusarium solani</i> (Mart.) Sacc., Michelia 2 (7): 296 (1881)	Basionym: <i>Fusisporium solani</i> Mart., Die Kartoffel-Epidemie der letzten Jahre oder die Stockfäule und Räude der Kartoffeln: 20 (1842). Obligate or homotypic synonyms: <i>Fusarium solani</i> (Mart.) Appel & Wollenw., Arbeiten aus der Kaiserlichen Biologischen Anstalt für Land- und Forstwirtschaft 8: 64-78 (1910); <i>Fusisporium solani</i> Mart., Die Kartoffel-Epidemie der letzten Jahre oder die Stockfäule und Räude der Kartoffeln: 20 (1842); <i>Neocosmospora solani</i> (Mart.) L.Lombard & Crous, Studies in Mycology 80: 228 (2015). Teleomorph synonyms: <i>Hypomyces haematococcus</i> var. <i>breviconus</i> Wollenw., Fusaria Autographice Delineata 3: 828 (1930); <i>Nectria haematococca</i> var. <i>brevicon</i> (Wollenw.) Gerlach, Fusarium: Diseases, Biology, and Taxonomy: 422 (1981). Facultative or heterotypic synonyms: <i>Cylindrocarpon vaginiae</i> C. Booth, Y.M.Clayton & Usherw., Proceedings of the Indian Academy of Sciences (Plant Sciences) 94 (2-3): 436 (1985); <i>Fusarium solani</i> f. 2 W.C.Snyder, Zentralblatt für Bakteriologie und Parasitenkunde, Abteilung 2 91: 174 (1934); <i>Fusarium solani</i> var. <i>minus</i> Wollenw., Fusaria Autographice Delineata 1: 403 (1916); <i>Fusarium striatum</i> Sherb., Memoirs of the Cornell University Agricultural Experimental Station 6: 255 (1915); <i>Nectria cancri</i> Rutgers, Ann. Jard. Bot. Buitenzorg, II: 59 (1913)	Biotroph	Cosmopolitan	Leaves, wood, roots	73
<i>Fusarium subglutinans</i> (Wollenw. & Reinking) P.E.Nelson, Toussoun & Marasas, <i>Fusarium</i> species, an illustrated manual for	Basionym: <i>Fusarium moniliforme</i> var. <i>subglutinans</i> Wollenw. & Reinking, Phytopathology 15 (3): 163 (1925) Synonyms: <i>Fusarium sacchari</i> var. <i>subglutinans</i> (Wollenw. & Reinking) Nirenberg, Mitteilungen der Biologischen Bundesanstalt für Land- und Forstwirtschaft 169: 53 (1976); <i>Fusarium neoceras</i> var. <i>subglutinans</i> (Wollenw. & Reinking) Raillo, Fungi of the genus <i>Fusarium</i>: 263 (1950); <i>Gibberella subglutinans</i> (Wollenw. & Reinking) P.E.Nelson, Toussoun & Marasas, <i>Fusarium</i> species, an illustrated	Parasite	Cosmopolitan, wa regions	Corn, cereals, fruits	74

<i>Gypsoplaca</i> Timdal, Bibliotheca Lichenologica 38: 423 (1990)	Gypsoplacaceae Timdal, Bibliotheca Lichenologica 38: 423 (1990)	identification: 135 (1983) <i>Gypsoplaca</i> <i>macrophylla</i> (Zahlbr.) Timdal, Bibliotheca Lichenologica 38: 424 (1990)	manual for identification: 135 (1983); <i>Fusarium</i> <i>subglutinans</i> var. <i>subglutinans</i> Basionym: <i>Lecidea macrophylla</i> Zahlbr., Symbolae Sinica 3: 110 (1930); Synonyms: <i>Gypsoplaca</i> <i>macrophylla</i> f. <i>macrophylla</i> (Zahlbr.) Timdal (1990)	Squamulose	Euroasia, especially in arid and semi-arid habitats, from Austria to Mongolia and western North America, Greenland Malaysia and South East Asia.	Wood, stones	75
<i>Halorosellinia</i> Whalley, E.B.G.Jones, K.D.Hyde & Læssøe, Mycological Research 104 (3): 368 (2000)	Xylariaceae Tul. & C.Tul., Selecta Fungorum Carpologia, Tomus Secundus. Xylariei - Valsei - Sphaeriei 2: 3 (1863)	<i>Halorosellinia</i> <i>oceanica</i> (S.Schatz) Whalley, E.B.G.Jones, K.D.Hyde & Læssøe, Mycological Research 104 (3): 370 (2000)	Basionym: <i>Hypoxylon oceanicum</i> S.Schatz, Mycotaxon 33: 413 (1988). Obligate or homotypic synonyms: <i>Hypoxylon oceanicum</i> S.Schatz, Mycotaxon 33: 413 (1988)	Endophyte		Mangroves	76
<i>Haslea</i> R. Simonsen, 1974	<i>Naviculaceae</i> Kützinger	<i>Haslea ostrearia</i> (Gaillon) Simonsen, Meteor. Forschungserge bnisse, Reihe D: Biologie. 19: 47 (1974) <i>Haslea</i> <i>pseudostrearia</i> Massé, Rincé & E.J.Cox, C. R. Acad. Sci. Paris, Sciences de la Vie 324: 617 (2001) <i>Haslea</i> <i>salstonica</i> Massé, Rincé & E.J.Cox, C. R. Acad. Sci. Paris, Sciences de la	Homotypic synonyms: <i>Vibrio ostrearius</i> Gaillon 1820; <i>Navicula ostrearia</i> (Gaillon) Bory 1827 Heterotypic synonym: <i>Amphipleura danica</i> var. <i>ostrearia</i> (Bory) Kützinger 1849	Marine pennate diatom	Cosmopolitan	Euryhaline, well adapted to shallow waters, such as oyster ponds	77-80
			-	Marine epibenthic diatom	Kingsbridge estuary (UK)	Intertidal sediments	78, 81
				Marine epibenthic diatom	Kingsbridge estuary (UK)	Intertidal sediments	78, 81

<i>Helminthosporium</i> Link, Magazin der Gesellschaft Naturforschenden Freunde Berlin 3 (1): 10 (1809)	Massarinaceae Munk, Friesia 5 (3-5): 305 (1956)	Vie 324: 617 (2001) <i>Helminthosporium abietis</i> W.B.Cooke & C.G.Shaw, Mycologia 44: 808 (1952)	Basionym: <i>Helminthosporium abietis</i> W.B.Cooke & C.G.Shaw, Mycologia 44: 808 (1952)	Saprotroph, parasite	Cosmopolitan	Woods, leaves	67
<i>Hericium</i> Pers., Neues Magazin für die Botanik 1: 109 (1794)	Hericiaceae Donk, Persoonia 3 (2): 269 (1964)	<i>Hericium erinaceus</i> (Bull.) Pers., Commentatio de Fungis Clavaeformibus: 27 (1797)	Basionym: <i>Hydnum erinaceus</i> Bull., Herbar de la France 1: t. 34 (1781). Obligate or homotypic synonyms: <i>Clavaria erinaceus</i> (Bull.) Paulet, Traité des champignons 2: index (1793); <i>Dryodon erinaceus</i> (Bull.) P.Karst., Bidrag till Kännedom av Finlands Natur och Folk 37: 92 (1882); <i>Hericium commune</i> Roques, Histoire des champignons comestibles et vénéneux: 47 (1832); <i>Hericium erinaceum</i> (Bull.) Pers. (1797); <i>Hydnum erinaceus</i> Bull., Herbar de la France 1: t. 34 (1781); <i>Steccherinum quercinum</i> Gray, A natural arrangement of British plants 1: 651 (1821). Facultative or heterotypic synonyms: <i>Clavaria caput-medusae</i> Bull., Herbar de la France 9: t. 412 (1789); <i>Clavaria conferta</i> Paulet, Traité des champignons 2: 427, index (1793); <i>Dryodon juranus</i> Quél., Comptes Rendus de l'Association Française pour l'Avancement des Sciences 30 (2): 496 (1902); <i>Hericium grande</i> Raf., Journal de Botanique (Desvaux) 3: 236 (1813); <i>Hericium hystrix</i> Pers., Commentatio de Fungis Clavaeformibus: 27 (1797); <i>Hericium strictum</i> Pers., Traité sur les Champignons Comestibles: 252 (1818); <i>Hericium unguiculatum</i> Pers., Mycologia Europaea 2: 153 (1825); <i>Hydnum hystricinum</i> Batsch, Elenchus fungorum: 113 (1783); <i>Hydnum notarisii</i> Inzenga, Giorn. Sci. nat. econ. Palermo: 2 (1866); <i>Hydnum omasum</i> Panizzi, Commentario della Società Crittogamologica Italiana 1 (3): 175 (1862); <i>Manina cordiformis</i> Scop., Dissertationes ad scientiam naturalem pertinentes: 97, t. 10 (1772); <i>Martella echinus</i> Scop., Annus Historico-Naturalis 4: 151 (1770)	Saprotroph	America, Europe, Asia	Woods	82
<i>Leprocaulon</i> Nyl., Bulletin de la Société Botanique	Leprocaulaceae Lendemer & Hodkinson,	<i>Leprocaulon quisquiliare</i> (Leers) M.Choisy,	Basionym: <i>Lichen quisquiliaris</i> Leers, Flora Herbornensis: 264 (1775); Synonyms: <i>Stereocaulon quisquiliare</i> (Leers) Hoffm., Deutschlands Flora oder botanisches Taschenbuch. Zweyter Theil für das Jahr	Fruticose	Mild-temperate to Mediterranean	On basic siliceous rocks covered by a thin film of soil;	59, 83, 84

de France 25: 352 (1878)	Mycologia 105 (4): 1007 (2013)	Bulletin Mensuel de la Société Linnéenne de Lyon 19: 166 (1950)	1795. Cryptogamie: 130 (1796); <i>Lichen microscopicus</i> Vill., Histoire des plantes du Dauphiné 3: 946 (1789); <i>Leprocaulon microscopicum</i> (Vill.) Gams ex D.Hawks., Trans. Proc. Torquay nat. Hist. Soc. 1972-3: 128 (1974); <i>Leprocaulon microscopicum</i> (Vill.) Gams, Trans. Proc. Torquay nat. Hist. Soc. 1972-3: 128 (1974); <i>Stereocaulon microscopicum</i> (Vill.) Frey: 89 (1932)			on brick walls in archaeological areas, also found on bark (e.g. of <i>Olea</i>).	
<i>Lobaria</i> (Schreb.) Hoffm., Deutschlands Flora oder botanisches Taschenbuch. Zweyter Theil für das Jahr 1795. Cryptogamie: 138 (1796)	Lobariaceae Chevall., Flore Générale des Environs de Paris 1: 609 (1826) OPPURE <i>Peltigeraceae</i> Dumort.????	<i>Lobaria isidiosa</i> (Müll. Arg.) Vain., Philippine Journal of Science Section C Botany 8 (2): 129 (1913)	Basionym: <i>Sticta retigera</i> f. <i>isidiosa</i> Müll. Arg., Flora (Regensburg) 65 (19): 300 (1882). Obligate or homotypic synonyms: <i>Cetraria braunsiana</i> f. <i>isidiosa</i> (Müll. Arg.) Zahlbr. (1929); <i>Lobaria retigera</i> f. <i>isidiosa</i> (Müll. Arg.) Vain., Hedwigia Beiblätter 37: 35 (1898); <i>Lobarina isidiosa</i> (Müll. Arg.) Inumaru (1943); <i>Sticta retigera</i> f. <i>isidiosa</i> Müll. Arg., Flora (Regensburg) 65 (19): 300 (1882)	Broad-lobed foliose	Tropical and subtropical areas of North America, Australia, Africa, Papua New Guinea and New Zealand	On bark and more rarely on moist rocks.	59, 85, 86
		<i>Lobaria retigera</i> (Bory) Trevis., Lichenotheca Veneta 1-2: no. 75 (1869)	Basionym: <i>Lichen retiger</i> Bory, Voyage dans les quatre principales îles des mers d'Afrique fait par ordre du gouvernement pendant les années IX et X de la république (1801 bis (1802) 1: 391 (1804). Obligate or homotypic synonyms: <i>Lichen retiger</i> Bory, Voyage dans les quatre principales îles des mers d'Afrique fait par ordre du gouvernement pendant les années IX et X de la république (1801 bis (1802) 1: 391 (1804); <i>Lobarina retigera</i> (Bory) Nyl., Lichenes Insularum Guineensium: 10 (1889); <i>Phycodiscis retigera</i> (Bory) Clem., The genera of Fungi: 175 (1909); <i>Sticta retigera</i> (Bory) Ach., Lichenographia Universalis: 455 (1810); <i>Stictina retigera</i> (Bory) Müll. Arg., Flora (Regensburg) 61 (31): 484 (1878)	Broad-lobed foliose	Tropical and subtropical areas of North America, Australia, Africa, Papua New Guinea and New Zealand	On bark and more rarely on moist rocks	59, 85
		<i>Lobaria subretigera</i> Inumaru, Acta phytotax. geobot., Kyoto 10: 214 (1941)	-	Broad-lobed foliose	Broad-lobed foliose	On bark and more rarely on moist rocks	59
<i>Lophiostoma</i> Ces. & De Not., Comment. Soc.	Lophiostomataceae Sacc., Syll. Fung. 2: 672 (1883)	<i>Lophiostoma bipolare</i> (K.D.Hyde) E.C.Y.Liew,	Basionym: <i>Lentistoma bipolare</i> (K.D.Hyde) A.Hashim., K.Hiray. & Kaz. Tanaka, Studies in Mycology 90: 171 (2018); <i>Massarina bipolaris</i> K.D. Hyde, Nova Hedwigia 61 (1-2): 131 (1995)	Saprotroph	Indopacific	Woods, decaying vegetable	87

Crittog. Ital. 1 (4): 219 (1863)		Aptroot & K.D.Hyde, Mycologia 94 (5): 812 (2002)					
<i>Mollisia</i> (Fr.) P. Karst., Bidrag Kännedom Finlands Natur Folk 19: 15, 189 (1871)	Mollisiaceae Rehm, Rabenhorst's Kryptogamen-Flora, Pilze - Ascomyceten 1(3): 503 (1891)	<i>Mollisia betulina</i> (Alb. & Schwein.) Rehm, Berichte der Bayerischen Botanischen Gesellschaft 14: 96 (1914)	Basionym: <i>Mollisia betulina</i> (Alb. & Schwein.) Rehm, Berichte der Bayerischen Botanischen Gesellschaft 14: 96 (1914) Synonyms: <i>Pyrenopeziza betulina</i> (Alb. & Schwein.) Rauschert, Haussknechtia 4: 54 (1988); Basionym: <i>Peziza betulina</i> Alb. & Schwein., Conspectus Fungorum in Lusatiae superioris: 339 (1805); <i>Perisporium betulinum</i> (Alb. & Schwein.) Fr., Systema Mycologicum 3 (1): 249 (1829); <i>Sclerotium betulinum</i> (Alb. & Schwein.) Fr., Systema Mycologicum 2 (1): 262 (1822); <i>Melampsora betulina</i> (Alb. & Schwein.) Desm., Plantes cryptogames du nord de la France Fasc. 41: no. 2047 (1850)	Saprotroph	Cosmopolitan	Decaying vegetable	88
<i>Neonothopanus</i> R.H.Petersen & Krisai, Persoonia 17 (2): 210 (1999)	Omphalotaceae Bresinsky, Plant Systematics and Evolution 150: 113 (1985)	<i>Neonothopanus nambi</i> (Speg.) R.H.Petersen & Krisai, Persoonia 17 (2): 210 (1999)	Basionym: <i>Agaricus nambi</i> Speg., Anales de la Sociedad Científica Argentina 16 (5): 247 (1883); <i>Dendrosarcus nambi</i> (Speg.) Kuntze, Revisio generum plantarum 3 (3): 463 (1898); <i>Pleurotus nambi</i> (Speg.) Speg., Sylloge Fungorum 5: 372 (1887)	Saprotroph, bioluminescent fungi	South America, Australia	Plant	89
<i>Nostoc</i> Lyngbye ex Bornet and Flahault 1886	Nostocaceae Eichler 1886	<i>Nostoc commune</i> Vaucher ex Bornet & Flahault, 1888	Basionym: <i>Nostoc commune</i> Vaucher ex Bornet & Flahault, 1888	Filamentous cyanobacteria that can form macroscopic or microscopic colonies	Cosmopolitan	Marine, fresh, terrestrial. Different species grow in freshwater biotopes, epipellic, epilithic and epiphytic in unpolluted lakes, ponds and pools, many species grow in different soils (especially the subgenus <i>Amorphonostoc</i>) or on their surface	78, 79, 90, 91

<i>Penicillium</i> Link, Magazin der Gesellschaft Naturforschenden Freunde Berlin 3 (1): 16 (1809)	Aspergillaceae Link, Abhandlungen der Königl. Akademie der Wissenschaften zu Berlin 1824: 165 (1826)	<i>Penicillium brasilianum</i> Bat., Anais Soc. Biol. Pernambuco 15 (1): 162 (1957)	Basionym: <i>Penicillium brasilianum</i> Bat., Anais Soc. Biol. Pernambuco 15 (1): 162 (1957)	Saprotrroph, parasite	Cosmopolitan	Soil, cereal, fruits	92
		<i>Penicillium citreoviride</i> Biourge, La Cellule 33: 297 (1923)	Basionym: <i>Penicillium citreoviride</i> Biourge, La Cellule 33: 297 (1923) Synonyms: <i>Penicillium citreonigrum</i> Dierckx, Ann. Soc. Sci. Bruxelles 25 (1): 86 (1901); Basionym: <i>Penicillium citreonigrum</i> Dierckx, Ann. Soc. Sci. Bruxelles 25 (1): 86 (1901); <i>Penicillium citreoviride</i> var. <i>aeneum</i> S. Abe, Journal of General and Applied Microbiology Tokyo 2 (1-2): 58 (1956); <i>Penicillium aeneum</i> G. Sm., Transactions of the British Mycological Society 46 (3): 334 (1963); <i>Penicillium subcinereum</i> Westling, Arkiv för Botanik 11 (1): 137 (1911); <i>Penicillium bertai</i> Talice & J.A.Mackinnon, Annls Parasit. Hum. Comp.: 97 (1929)	Saprotrroph, parasite	Cosmopolitan	Soil, cereal	93
		<i>Penicillium citrinum</i> Thom, U.S.D.A. Bureau of Animal Industry Bulletin 118: 61 (1910)	Basionym: <i>Penicillium citrinum</i> Thom, U.S.D.A. Bureau of Animal Industry Bulletin 118: 61 (1910) Facultative or heterotypic synonyms: <i>Citromyces subtilis</i> Bainier & Sartory, Bulletin de la Société Mycologique de France 28: 46 (1912); <i>Penicillium aurifluum</i> Biourge, La Cellule 33: 250 (1923); <i>Penicillium botryosum</i> Bat. & H.Maia, Anais da Sociedade de Biologia de Pernambuco 15 (1): 159 (1957); <i>Penicillium citrinum</i> var. <i>pseudopaxilli</i> Martínez & Ramírez (?); <i>Penicillium phaeojanthinellum</i> Biourge, La Cellule 33: 289 (1923); <i>Penicillium sartorii</i> Thom (1930); <i>Penicillium sartoryi</i> Thom, The Penicillia: 233 (1930).	Saprotrroph, parasite	Cosmopolitan	Soil, fruits	94
		<i>Penicillium griseofulvum</i> Dierckx, Ann. Soc. Sci. Bruxelles 25 (1): 88 (1901)	Basionym: <i>Penicillium griseofulvum</i> Dierckx, Ann. Soc. Sci. Bruxelles 25 (1): 88 (1901) Synonyms: <i>Penicillium patulum</i> Bainier, Bulletin de la Société Mycologique de France 22: 208 (1906); <i>Penicillium flexuosum</i> E.Dale, Annales Mycologici 24: 137 (1924); <i>Penicillium urticae</i> Bainier, Bulletin de la Société Mycologique de France 23 (1): 15 (1907); <i>Penicillium duninii</i> Sidibe, Mikol. Fitopatol.: 371	Saprotrroph, parasite	Cosmopolitan	Fruits	95

	(1974); <i>Penicillium maltum</i> M.Hori & T.Yamam., Jap. J. Bacteriol.: 1105 (1954)				
<i>Penicillium oxalicum</i> Currie & Thom, 1915	Basionym: <i>Penicillium oxalicum</i> Currie & Thom, J. Biol. Chem. 22: 289 (1915). Taxonomic synonym(s): <i>Penicillium asturianum</i> C. Ramírez & A.T. Martínez, Mycopathologia 74 (1): 42 (1981)	Saprotroph	Cosmopolitan	Soil	96
<i>Penicillium roqueforti</i> Thom, U.S.D.A. Bur. Animal Industr. Bull. 82: 35 (1906)	Basionym: <i>Penicillium roqueforti</i> Thom, U.S.D.A. Bur. Animal Industr. Bull. 82: 35 (1906) Synonyms: <i>Penicillium conservandi</i> Novobr., Novosti Sistematiki Nizshikh Rastenii 11: 233 (1974); <i>Penicillium roqueforti</i> var. <i>viride</i> Datt.-Rubbo, Transactions of the British Mycological Society 22 (1-2): 178 (1938); <i>Penicillium stilton</i> Biourge, La Cellule 33: 204 (1923); <i>Penicillium aromaticum-casei</i> Sopp ex Sacc., Sylloge Fungorum 22: 1278 (1913); <i>Penicillium atroviride</i> Sopp, Skrifter udgivne af Videnskabs-Selskabet i Christiania. Matematisk-Naturvidenskabelig Klasse 11: 149 (1912); <i>Penicillium biourgei</i> Arnaud, Boll. Ist. Sieroter. Milan.: 27 (1928); <i>Penicillium roqueforti</i> var. <i>weidemannii</i> Westling (1911); <i>Penicillium suaveolens</i> Biourge, La Cellule 33: 200 (1923); <i>Penicillium vesiculosum</i> Bainier, Bulletin de la Société Mycologique de France 23 (1): 10 (1907); <i>Penicillium virescens</i> Sopp, Skrifter udgivne af Videnskabs-Selskabet i Christiania. Matematisk-Naturvidenskabelig Klasse 11: 157 (1912); <i>Penicillium weidemannii</i> var. <i>fuscum</i> Arnaud, Boll. Ist. Sieroter. Milan.: 27 (1928); <i>Penicillium conservandi</i> Novobr., Nauchnye Doklady Vyshe Shkoly: 105 (1972); <i>Penicillium roqueforti</i> var. <i>weidemannii</i> Westling, Arkiv för Botanik 11 (1): 71 (1911); <i>Penicillium gorgonzolae</i> Weid., La Cellule 33: 204 (1923); <i>Penicillium weidemannii</i> (Westling) Biourge, La Cellule 33: 204 (1923); <i>Penicillium gorgonzolae</i> Weid (1923)	Saprotroph	Cosmopolitan	Soil	97
<i>Penicillium verruculosum</i> Peyronel, I germi astmosferici dei fungi con micelio: 22 (1913)	Basionym: <i>Penicillium verruculosum</i> Peyronel, I germi astmosferici dei fungi con micelio: 22 (1913) Synonyms: <i>Penicillium aculeatum</i> var. <i>apiculatum</i> S.Abe, Journal of General and Applied Microbiology Tokyo 2 (1-2): 124 (1956)	Saprotroph, psychrophilic fungus	Northern Europe, Canada, South America	Grains, seeds, decaying vegetation	92

<i>Phoma</i> Sacc., Michelia 2 (6): 4 (1880)	Pleosporomyceti dae C.L.Schoch, Spatafora, Crous & Shoemaker, Mycologia 98 (6): 1048 (2007)	<i>Phoma</i> Sacc., Michelia 2 (6): 4 (1880)	Basionym: <i>Phoma</i> Sacc., Michelia 2 (6): 4 (1880) Synonyms: <i>Chlamydosporium</i> Peyronel, I germi astmosferici dei funghi con micelio: 18 (1913); <i>Leptophoma</i> Höhn., Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften Math.-naturw. Klasse Abt. I 124: 73 (1915); <i>Macropodiella</i> Speg., Anales del Museo Nacional de Historia Natural Buenos Aires 17: 134 (1908); <i>Paraphoma</i> Morgan-Jones & J.F.White, Mycotaxon 18 (1): 58 (1983); <i>Phomopsina</i> Petr., Annales Mycologici 20: 145 (1922); <i>Pseudosclerophoma</i> Petr., Annales Mycologici 21 (3- 4): 283 (1923); <i>Rhizosphaerella</i> Höhn., Hedwigia 59: 254 (1917); <i>Sclerophomina</i> Höhn., Hedwigia 59: 240 (1917); <i>Vialina</i> Curzi, Bolletino della Stazione di Patologia Vegetale di Roma 15: 252 (1935); <i>Peyronellaea</i> Goid., Atti della Accademia Nazionale dei Lincei Sér. 8, 1: 451 (1946)	Saprotroph	Pacific	Soil, plants	98
<i>Phomopsis</i> (Sacc.) Sacc., Ann. Mycol. 3 (6): 166 (1905)	Diaporthaceae Höhn. ex Wehm., American Journal of Botany 13: 638 (1926)	<i>Phomopsis</i> <i>lactucae</i> (Sacc.) Bubák, Österreichische Botanische Zeitschrift 55: 78 (1905)	Basionym: <i>Phoma lactucae</i> Sacc., Michelia 2 (6): 94 (1880)	Parasite	Indopacific	Leaves, fruits, rachises, shoots	61
<i>Pleurotus</i> (Fr.) P. Kumm., Der Führer in die Pilzkunde: 24 (1871)	Pleurotaceae Kühner, Bulletin Mensuel de la Société Linnéenne de Lyon 49: 184 (1980)	<i>Pleurotus</i> <i>ostreatus</i> (Jacq.) P.Kumm., Der Führer in die Pilzkunde: 105 (1871) <i>Pleurotus</i> <i>ostreatus</i> (Jacq.) P.Kumm., Der Führer in die Pilzkunde: 105 (1871)	Basionym: <i>Agaricus ostreatus</i> Jacq., Flora Austriaca 2: 3, t. 104 (1774). Obligate or homotypic synonyms: <i>Agaricus ostreatus</i> Jacq., Flora Austriaca 2: 3, t. 104 (1774); <i>Crepidopus ostreatus</i> (Jacq.) Gray, A natural arrangement of British plants 1: 616 (1821); <i>Dendrosarcus ostreatus</i> (Jacq.) Kuntze, Revisio generum plantarum 3 (2): 463 (1898). Facultative or heterotypic synonyms: <i>Agaricus ochraceus</i> Pers., Abb. Schwämme (1793); <i>Pleurotus columbinus</i> Quél., Fungi Tridentini 1 (1): 10 (1881); <i>Pleurotus floridanus</i> Singer, Papers of the Michigan Academy of Sciences 32: 134 (1948).	Saprotroph	Pacific	Soil, wood	99
<i>Pseudomonas</i> Migula 1894	Pseudomonadac eae Winslow et al. 1917	<i>Pseudomonas</i> <i>aeruginosa</i> (Schroeter 1872) Migula 1900	Homotypic synonym: <i>Bacterium aeruginosum</i> Cohn 1872; <i>Bacillus aeruginosus</i> (Schroeter 1872) Trevisan 1885; probable synonym or variety <i>Pseudomonas</i> <i>polycolor</i> Clara 1930 Heterotypic synonym: <i>Micrococcus pyocyaneus</i> Zopf 1884; <i>Pseudomonas pyocyanea</i> (Zopf 1884) Migula 1895; <i>Pseudomonas polycolor</i> ; <i>Bacillus pyocyaneus</i>	Parasite	Cosmopolitan	Humans	100

<i>Pyrenophora</i> Fr., Summa vegetabilium Scandinaviae 2: 397 (1849)	Pleosporaceae Nitschke, Verh. Naturhist. Vereines Preuss. Rheinl.: 74 (1869)	<i>Pyrenophora lolii</i> Dovaston, Transactions of the British Mycological Society 31 (3-4): 251 (1948)	(Zopf 1884) Flugge 1886; <i>Bacterium pyocyaneum</i> (Zopf 1884) Lehmann and Neumann 1896 Basionym: <i>Pyrenophora lolii</i> Dovaston, Transactions of the British Mycological Society 31 (3-4): 251 (1948)	Parasite	Pacific	Leaves, seeds	101
<i>Schizonema</i> C.A. Agardh, 1824	Bacillariophyceae familia incertae sedis	<i>Schizonema cruciger</i> f. <i>cruciger</i> W. Smith, 1856. Synopsis of British Diatomaceae. John Van Voorst, London 1856., 2: 107 pp., pls. 32-60, 61-62, A-E.	<i>Haslea crucigera</i> (W.Smith) Simonsen, 1974; <i>Navicula crucigera</i> (W.Smith) Cleve, 1894; <i>Schizonema crucigerum</i> W.Smith, 1856; <i>Stauroneis crucigera</i> (<i>crucigerum</i> , <i>cruciger</i>) (W. Smith) Heiberg 1863; <i>Dickieia crucigera</i> (W. Smith) De Toni 1891	Pennate, benthic diatom	Australia, North America, North Europe	Marine	78, 79
<i>Scytonema</i> C.Agardh ex Bornet & Flahault (1887)	Scytonematacea e Rabenhorst ex Bornet et Flahault 1886	<i>Scytonema varium</i> Kützing ex Bornet & Flahault, 1886	Basionym: <i>Scytonema varium</i> Kützing ex Bornet & Flahault, 1886	Saprotroph, ilamentous cyanobacteria	Cosmopolitan	Marine, fresh, terrestrial. Numerous species grow only in tropical and ecologically distinctly delimited habitats, such as lateritic soils, dripping rocks and reservoirs with water vegetation	78, 79, 102, 103
<i>Sorangium</i> E.Jahn (1924): Beitrage zur botanischen Protistologie. I. Die Polyangiden. pp. 1-107. Verlag von Gebruder	Polyangiaceae E.Jahn (1924): Beitrage zur botanischen Protistologie. I. Die Polyangiden. pp. 1-107. Verlag	<i>Sorangium cellulosum</i> A.A.Imshenetski & L.Solntseva (1936). Izv. Akad. Nauk S.S.S.R. Ser.	Basionym: <i>Sorangium cellulosum</i> A.A.Imshenetski & L.Solntseva (1936). Izv. Akad. Nauk S.S.S.R. Ser. Viol. (1936) 6:1115-1172. Homotypic synonym: <i>Polyangium cellulosum</i> (ex Imshenetski and Solntseva 1936) Brockman 1989	Saprotroph, parasite	Cosmopolitan	Soil, water, animal feces, and tree bark	95

Borntraeger, Leipzig	von Gebruder Borntraeger, Leipzig	Viol. (1936) 6:1115-1172.					
<i>Streptomyces</i> Waksman and Henrici 1943	Streptomycetaceae Waksman and Henrici 1943	<i>Streptomyces albus</i> (Rossi Doria 1891) Waksman and Henrici 1943 (Approved Lists 1980) emend. Labeda et al. 2014	Basionym: <i>Streptomyces albus</i> (Rossi Doria 1891) Synonym: <i>Actinomyces almquisti</i> Duche 1934; <i>Actinomyces Gibsoni</i> Dodge 1935; <i>Actinomyces Gibsonii</i> Erikson 1935; <i>Streptomyces flocculus</i> (Duche 1934) Waksman and Henrici; <i>Streptomyces almquistii</i> (Duche 1934) Pridham et al. 1958; <i>Streptomyces rangoonensis</i> corrig. (Erikson 1935) Pridham et al. 1958; <i>Streptomyces griseus</i> subsp. <i>solivifaciens</i> Pridham 1970; <i>Actinomyces rangoon</i> Erikson 1935	Saprotroph	Cosmopolitan	Soil, air	4
		<i>Streptomyces somaliensis</i> S.A.Waksman & A.T.Henrici (1948b)	Basionym: <i>Streptomyces somaliensis</i> S.A.Waksman & A.T.Henrici (1948b) Synonym: <i>Indiella somaliensis</i> Brumpt 1906,	Saprotroph	Cosmopolitan	Soil, air	104
		<i>Streptomyces subutilus</i> Arai et al. 1964	Basionym: <i>Streptomyces</i> Waksman & Henrici, Journal of Bacteriology 46 (4): 339 (1943)	Saprotroph	Cosmopolitan	Soil	105
		<i>Streptomyces violens</i> L.V.Kalakutskii & N.A.Krasil'nikov (1966)	Basionym: <i>Streptomyces violens</i> LV.Kalakutskii & N.A.Krasil'nikov (1966)	Saprotroph	Cosmopolitan	Soil	106
<i>Sundstroemia</i> Medlin, Boonprakob, Lundholm & Moestrup, 2021	<i>Rhizosoleniaceae</i> De Toni	<i>Streptomyces mobaraensis</i> (Brightwell) Medlin, Nova Hedwigia Beiheft 151: 223 (2021)	Homotypic synonym: <i>Rhizosolenia setigera</i> Brightwell 1858	Diatom	Arctic, Atlantic Islands, Europe, N. America, Western Atlantic, S. America, Indian Ocean Islands, Middle East, SW Asia, SE Asia, Asia, Australia and New Zealand	Marine species	78, 79, 107
<i>Tainosphaeria</i> F.A.Fernández & Huhndorf, Fungal Diversity 18: 44 (2005)	Chaetosphaeriaceae Réblová, M.E.Barr & Samuels, Sydowia 51: 56 (1999)	<i>Tainosphaeria simplex</i> (S.Hughes & W.B.Kendr.) Réblová & Hern.-Restr., Journal of Fungi	Basionym: <i>Codinaea simplex</i> S.Hughes & W.B.Kendr., New Zealand Journal of Botany 6: 362 (1968); Synonyms: <i>Dictyochaeta simplex</i> (S.Hughes & W.B.Kendr.) Hol.-Jech., Folia Geobot. Phytotax. 19 (4): 434 (1984)	Saprotroph	Cosmopolitan	Woods	57

		7 (12, no. 1097): 85 (2021)					5
<i>Tolypocladium</i> W.Gams, Persoonia 6 (2): 185 (1971)	Ophiocordycipita ceae G.H.Sung, J.M.Sung, Hywel- Jones & Spatafora, Studies in Mycology 57: 35 (2007)	<i>Tolypocladium</i> <i>inflatum</i> W.Gams, Persoonia 6 (2): 185 (1971)	Basionym: <i>Tolypocladium inflatum</i> W.Gams, Persoonia 6 (2): 185 (1971) Synonyms: <i>Pachybasium niveum</i> O.Rostr., Dansk botanisk Arkiv 2 (5): 41 (1916); <i>Tolypocladium niveum</i> (O.Rostr.) Bissett, Canadian Journal of Botany 61 (5): 1312 (1983); <i>Beauveria nivea</i> (O.Rostr.) Arx, Mycotaxon 25 (1): 156 (1986)	Saprotroph	Pacific	Woods	

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Table S7. Name of the species as reported in the publication, accepted name, family, synonyms, lifeform, native range, and ecology of the selected plants

Name of the species as reported in the publication	Accepted name, protologue ¹	Family ¹	Synonyms ¹	Lifeform ¹	Native range ¹	Ecology	References (and other literature therein)
<i>Aletris farinosa</i> L.	<i>Aletris farinosa</i> L., Sp. Pl.: 319 (1753)	<i>Nartheciaceae</i> Fr. ex Bjurzon	Heterotypic Synonyms: <i>Aletris alba</i> Michx., Fl. Bor.-Amer. 1: 189 (1803); <i>Aletris lucida</i> Raf., Autik. Bot.: 136 (1840)	Perennial or rhizomatous geophyte	SE. Canada to Central & E. U.S.A.	Temperate biome. Open sites of various habitats, moist bogs, dry to mesic prairies, and dry, upland woods and thickets. Alt. 0-2000+ m	1-4
<i>Aleuritopteris agetae</i> Saiki	<i>Hemionitis krameri</i> (Franch. & Sav.) Christenh., Global Fl. 4: 16 (2018)	<i>Pteridaceae</i> E.D.M.Kirchn	Homotypic Synonyms: <i>Aleuritopteris krameri</i> (Franch. & Sav.) Ching in Hong Kong Naturalist 10: 202 (1941); <i>Cheilanthes krameri</i> Franch. & Sav. in Enum. Pl. Jap. 2: 212, 619 (1877). Heterotypic Synonyms: <i>Aleuritopteris agetae</i> Saiki in J. Phytogeogr. Taxon. 32: 1 (1984); <i>Cheilanthes agetae</i> (Saiki) C.M.Kuo in Taiwania 30: 57 (1985); <i>Hemionitis agetae</i> (Saiki) Christenh. in Global Fl. 4: 9 (2018)	Lithophyte	Central Japan, Taiwan to Philippines	Subtropical biome. Rock crevices on slopes or cliffs. Alt. 1800-2700 m	1, 5-7
<i>Aleuritopteris anceps</i> (Blanf.) Panigrahi	<i>Hemionitis anceps</i> (Blanf.) Christenh., Global Fl. 4: 9 (2018)	<i>Pteridaceae</i> E.D.M.Kirchn	Homotypic Synonyms: <i>Aleuritopteris anceps</i> (Blanf.) Panigrahi in Bull. Bot. Surv. India 2: 321 (1961); <i>Aleuritopteris farinosa</i> var. <i>anceps</i> (Blanf.) Ching in Hong Kong Naturalist 10: 201 (1941); <i>Cheilanthes anceps</i> Blanf. in J. Simla Naturalists' Soc. 1(2): 21 (1886); <i>Cheilanthes farinosa</i> var. <i>anceps</i> (Blanf.) Blanf. in J. Asiat. Soc. Bengal 57: 301 (1888). Heterotypic Synonyms: <i>Aleuritopteris javanensis</i> Saiki in J. Phytogeogr. Taxon. 32: 7 (1984); <i>Aleuritopteris pseudofarinosa</i> Ching & S.K.Wu in Acta Phytotax. Sin. 19: 72 (1981); <i>Aleuritopteris pseudofarinosa</i> var. <i>glandulosa</i> H.G.Zhou in Guihaia 12: 323 (1992); <i>Aleuritopteris wuyishanensis</i> Ching in Wuyi Sci. J. 1: 2 (1981); <i>Cheilanthes candida</i> Zoll. in Natuur- Geneesk. Arch. Ned.-Indië 2: 203 (1845), nom. illeg.; <i>Cheilanthes pseudofarinosa</i> (Ching & S.K.Wu) K.Iwats. in Fl. Thailand 3(4): 618 (1989)	Rhizomatous geophyte	Tropical & Subtropical Asia	Wet tropical biome. Rock crevices on slopes. Alt. 300-2600 m	1, 5, 7
<i>Aleuritopteris kuhnii</i> (Milde) Ching	<i>Hemionitis kuhnii</i> (Milde) Christenh., Global Fl. 4: 16 (2018)	<i>Pteridaceae</i> E.D.M.Kirchn	Homotypic Synonyms: <i>Aleuritopteris kuhnii</i> (Milde) Ching in Hong Kong Naturalist 10: 202 (1941); <i>Cheilanthes kuhnii</i> Milde in Bot. Zeitung (Berlin) 25: 149 (1867); <i>Leptolepidium kuhnii</i> (Milde) K.H.Shing & S.K.Wu in Acta Bot. Yunnan. 1(1): 117 (1979); <i>Oeosporangium kuhnii</i> (Milde) Fraser-Jenk. in Annot. Checkl. Ind. Pterid. 1: 261 (2016)	Lithophyte	S. Russian Far East to Korea and China, Central Japan	Temperate biome. Rock crevices on dry slopes, forests. Alt. 1000-3500 m	1, 2, 5

			Heterotypic Synonyms: <i>Aleuritopteris caesia</i> (Christ) Ching in Hong Kong Naturalist 10: 202 (1941); <i>Aleuritopteris caesia</i> var. <i>efarinosa</i> H.S.Kung, Li Bing Zhang & X.S.Guo in Acta Bot. Yunnan. 17: 422 (1995); <i>Aleuritopteris kuhnii</i> f. <i>efarinosa</i> (Makino) Tagawa in Acta Phytotax. Sin. 14: 191 (1952); <i>Aleuritopteris tenella</i> (Ching & S.K.Wu) Saiki in J. Phytogeogr. Taxon. 32: 11 (1984); <i>Cheilanthes brandtii</i> Franch. & Sav. in Enum. Pl. Jap. 2: 212, 620 (1877); <i>Cheilanthes brandtii</i> var. <i>efarinosa</i> Makino in J. Jap. Bot. 1: 26 (1917); <i>Cheilanthes caesia</i> Christ in Bull. Acad. Int. Géogr. Bot., sér. 3, 16: 133 (1906); <i>Cheilanthes kuhnii</i> var. <i>caesia</i> (Christ) C.Chr. in Acta Horti Gothob. 1: 90 (1924); <i>Cheilanthes kuhnii</i> f. <i>efarinosa</i> (Makino) Nakaike in J. Nippon Fernist Club 3(Suppl. 2): 40 (2004); <i>Cheilanthes lanceolata</i> C.Chr. in Bot. Gaz. 56: 334 (1913); <i>Hemionitis brandtii</i> (Franch. & Sav.) Christenh. in Global Fl. 4: 10 (2018); <i>Hemionitis caesia</i> (Christ) Christenh. in Global Fl. 4: 11 (2018); <i>Leptolepidium caesium</i> (Christ) K.H.Shing & S.K.Wu in Acta Bot. Yunnan. 1(1): 117 (1979); <i>Leptolepidium kuhnii</i> var. <i>brandtii</i> (Franch. & Sav.) K.H.Shing & S.K.Wu in Acta Bot. Yunnan. 1(1): 118 (1979); <i>Leptolepidium tenellum</i> Ching & S.K.Wu in Acta Bot. Yunnan. 1(1): 117 (1979)				
<i>Aleuritopteris mexicana</i> Fée	<i>Hemionitis farinosa</i> (Forssk.) Christenh., Global Fl. 4: 13 (2018)	<i>Pteridaceae</i> E.D.M.Kirchn	Homotypic Synonyms: <i>Aleuritopteris farinosa</i> (Forssk.) Fée in Mém. Foug., 5. Gen. Filic.: 154 (1852); <i>Allosorus farinosus</i> (Forssk.) C.Presl in Tent. Pterid.: 153 (1836); <i>Cassebeera farinosa</i> (Forssk.) J.Sm. in J. Bot. (Hooker) 4: 159 (1841); <i>Cheilanthes farinosa</i> (Forssk.) Kaulf. in Enum. Filic.: 212 (1824); <i>Pteris farinosa</i> Forssk. in Fl. Aegypt.-Arab.: 187 (1775) Heterotypic Synonyms: <i>Aleuritopteris chihuahuensis</i> Saiki in J. Phytogeogr. Taxon. 32: 85 (1984); <i>Aleuritopteris mexicana</i> Fée in Mém. Foug., 5. Gen. Filic.: 154 (1852); <i>Aleuritopteris peruviana</i> Saiki in J. Phytogeogr. Taxon. 32: 89 (1984); <i>Cheilanthes chihuahuensis</i> (Saiki) Fraser-Jenk. in New Sp. Syndr. Indian Pteridol.: 81 (1997); <i>Cheilanthes mexicana</i> (Fée) Punetha & Kholia in New Botanist, Int. Quart. J. Pl. Sci. Res. 16: 119 (1989), nom. illeg.; <i>Cheilanthes pulveracea</i> C.Presl in Reliq. Haenk. 1: 64 (1825); <i>Cheilanthes rigidula</i> Wall. in Numer. List: n.° 2175 (1830), not validly publ.; <i>Hemionitis chihuahuensis</i> (Saiki) Christenh. in Global Fl. 4: 11 (2018); <i>Pellaea pulcherrima</i> Rovirosa in Pteridogr. Sur Mexico: t. 14 (1909), nom. nud.	Terrestrial / epipetric	Mexico to Venezuela and Peru, Africa to Arabian Peninsula	Wet tropical biome. Tropical deciduous and semideciduous forests. Pine - Oak Forest. Alt. 1500 to 2000 m	8
<i>Arabidopsis thaliana</i> (L.) Heynh.	<i>Arabidopsis thaliana</i> (L.) Heynh., F.Holl & H.Heynhold, Fl. Sachsen: 538 (1842)	<i>Brassicaceae</i> Burnett	Homotypic Synonyms: <i>Arabis muralis</i> Salisb. in Prodr. Stirp. Chap. Allerton: 272 (1796), nom. superfl.; <i>Arabis thaliana</i> L. in Sp. Pl.: 665 (1753); <i>Conringia thaliana</i> (L.) Rchb. in Icon. Fl. Germ. Helv. 2: t. 60, f. 4380 (1838); <i>Crucifera thaliana</i> (L.) E.H.L.Krause in J.W.Sturm, Deutschl. Fl. Abbild. ed. 2. 6: 86 (1902); <i>Erysimum thalianum</i> (L.) Kitt. in Taschenb. Fl. Deutschl., ed. 2: 899 (1844); <i>Hesperis thaliana</i> (L.) Kuntze in Revis. Gen. Pl. 2: 935 (1891); <i>Nasturtium thaliana</i> (L.) Andr. ex DC. in Syst. Nat. 2: 226 (1821); <i>Pilosella thaliana</i> (L.) Kostel. in Ind. Hort. Bot. Prag.: 104 (1844); <i>Sisymbrium thalianum</i> (L.) J.Gay in Ann. Sci. Nat. (Paris) 7:	Annual	Temp. Eurasia to Tropical African Mountains	Temperate biome. Plains, mountain slopes, river banks, roadsides. Alt. near sea level to 2000 m	1, 2, 5

			399 (1826); <i>Stenophragma thalianum</i> (L.) Čelak. in Arch. Naturwiss. Landesdurchf. Böhmen 2: 445 (1875). Heterotypic Synonyms: <i>Arabidopsis thaliana</i> var. <i>apetala</i> O.E.Schulz in H.G.A.Engler (ed.), Pflanzenr., IV, 105(86): 274 (1924); <i>Arabidopsis thaliana</i> var. <i>brachycarpa</i> Andr. in Ind. Horti Bot. Univ. Budapest 3: 25 (1939); <i>Arabidopsis thaliana</i> var. <i>burnatii</i> (Briq.) Briq. in Prodr. Fl. Corse 2(1): 38 (1913); <i>Arabidopsis thaliana</i> var. <i>pusilla</i> (Hochst. ex A.Rich.) O.E.Schulz in H.G.A.Engler (ed.), Pflanzenr., IV, 105(86): 274 (1924); <i>Arabis arcuata</i> Dulac in Fl. Hautes-Pyrénées: 200 (1867), nom. illeg.; <i>Arabis parviflora</i> Raf. in Amer. Monthly Mag. & Crit. Rev. 2: 43 (1817); <i>Arabis pubicalyx</i> Miq. in Ann. Mus. Bot. Lugduno-Batavi 2: 72 (1865); <i>Arabis pubicalyx</i> var. <i>soyensis</i> H.Boissieu in Bull. Herb. Boissier 7: 788 (1899); <i>Arabis ramosa</i> Lam. in Fl. Franç. 2: 510 (1779); <i>Arabis scabra</i> Gilib. in Fl. Lit. Inch. 2: 61 (1782), opus utique oppr.; <i>Arabis thaliana</i> var. <i>dentata</i> Hartm. in Handb. Skand. Fl.: 256 (1820); <i>Arabis thaliana</i> var. <i>pubicalyx</i> (Miq.) Makino in Y.linuma, Somoku-Dzusetsu, ed. 3, 3: 12 (1912); <i>Arabis thaliana</i> var. <i>vulgaris</i> Hartm. in Handb. Skand. Fl.: 256 (1820); <i>Arabis thaliana</i> var. <i>vulgaris</i> Hartm. in Handb. Skand. Fl.: 256 (1820); <i>Arabis zeyheriana</i> Turcz. in Bull. Soc. Imp. Naturalistes Moscou 27(II): 292 (1854 publ. 1855); <i>Cardamine pusilla</i> Hochst. ex A.Rich. in Tent. Fl. Abyss. 1: 18 (1848); <i>Erysimum pubicalyx</i> (Miq.) Kuntze in Revis. Gen. Pl. 2: 933 (1891); <i>Phryne gesneri</i> Bubani in Fl. Pyren. 3: 176 (1901); <i>Sisymbrium pumilio</i> Oliv. in Fl. Trop. Afr. 1: 64 (1868); <i>Sisymbrium simplicissimum</i> S.G.Gmel. in Reise Russland 3: 303 (1774); <i>Sisymbrium thalianum</i> var. <i>pusillum</i> (Hochst. ex A.Rich.) Durand & Schinz in Consp. Fl. Afric. 1: 99 (1898); <i>Stenophragma thalianum</i> var. <i>burnatii</i> Briq. in Annuaire Conserv. Jard. Bot. Genève 9: 132 (1905); <i>Stenophragma thalianum</i> var. <i>pusilla</i> (Hochst. ex A.Rich.) Engl. in Hochgebirgsfl. Afrika: 226 (1895)				
<i>Artemisia argyi</i> H.Lév. & Vaniot	<i>Artemisia argyi</i> H.Lév. & Vaniot, Repert. Spec. Nov. Regni Veg. 8: 138 (1910)	<i>Asteraceae</i> Bercht. & J.Presl	<i>Artemisia argyi</i> f. <i>argyi</i>; <i>Artemisia argyi</i> f. <i>gracilis</i> (Pamp.) Kitag. in Neolin. Fl. Manshur.: 612 (1979); <i>Artemisia argyi</i> f. <i>microcephala</i> Pamp. in Nuovo Giorn. Bot. Ital., n.s., 36: 453 (1930); <i>Artemisia argyi</i> var. <i>gracilis</i> Pamp. (Nuovo Giorn. Bot. Ital., n.s., 36: 453 (1930)); <i>Artemisia argyi</i> var. <i>incana</i> (Maxim.) Pamp. in Nuovo Giorn. Bot. Ital., n.s., 36: 451 (1930); <i>Artemisia chiarugii</i> Pamp. in Nuovo Giorn. Bot. Ital., n.s., 36: 486 (1930); <i>Artemisia handel-mazzettii</i> Pamp. in Nuovo Giorn. Bot. Ital., n.s., 36: 455 (1930); <i>Artemisia nutans</i> Nakai in Bot. Mag. (Tokyo) 23: 187 (1909), nom. illeg.; <i>Artemisia nutantiflora</i> Nakai in Fl. Sylv. Kor. 14: 101 (1923); <i>Artemisia nutantiflora</i> Nakai ex Pamp., 1930; <i>Artemisia vulgaris</i> var. <i>incana</i> Maxim. in Mém. Acad. Imp. Sci. St.-Pétersbourg Divers Savans 9: 160 (1859); <i>Artemisia vulgaris</i> var. <i>incanescens</i> Franch. in Nouv. Arch. Mus. Hist. Nat., sér. 2, 6: 49 (1884)	Perennial or subshrub	Russian Far East to China	Temperate biome. Waste places, roadsides, slopes, hills, steppes, forest steppes. Alt. low elevations to 1500 m	1-3, 5

<i>Artemisia frigida</i> Willd.	<i>Artemisia frigida</i> Willd., Sp. Pl. ed. 4. 3: 1838 (1803)	<i>Asteraceae</i> Bercht. & J.Presl	Homotypic Synonym: <i>Absinthium frigidum</i> (Willd.) Besser in Bull. Soc. Imp. Naturalistes Moscou 1: 251 (1829)	Perennial or subshrub	E. Europe to Temp. Asia, N. America	Subalpine or subarctic biome. On rubbly and stony slopes in steppes, standing fallow land, dry hillsides, stable dunes, dry waste areas. Alt. 1000-4000 m	1, 2, 4, 5, 9
<i>Artemisia umbelliformis</i> Lam.	<i>Artemisia umbelliformis</i> Lam., Encycl. 1: 262 (1783)	<i>Asteraceae</i> Bercht. & J.Presl	Homotypic Synonyms: <i>Artemisia glacialis</i> var. <i>umbelliformis</i> (Lam.) Rouy in G.Rouy & J.Foucaud, Fl. France 8: 290 (1903). <i>Artemisia laxa</i> Fritsch, (synonym of <i>Artemisia umbelliformis</i> subsp. <i>umbelliformis</i>), 6: 88 (1893) [in Flora d'Italia reported as synonym of <i>Artemisia umbelliformis</i> ¹⁰]; <i>Artemisia mutellina</i> Vill., non S.G.Gmelin, (synonym of <i>Artemisia umbelliformis</i> subsp. <i>umbelliformis</i>), Prosp. Hist. Pl. Dauphiné: 31 (1779) [in Flora d'Italia reported as synonym of <i>Artemisia umbelliformis</i> ¹⁰]	Subshrub	Europe	Mountain rocks, moraines, and river gravels. Alt. 2400-3000 m	1, 3, 6, 10-12
<i>Caesalpinia crista</i> L.	<i>Ticanto crista</i> (L.) R.Clark & Gagnon, PhytoKeys 205: 70 (2022)	<i>Fabaceae</i> Lindl.	Homotypic Synonyms: <i>Caesalpinia crista</i> L. in Sp. Pl.: 380 (1753); <i>Guilandina crista</i> (L.) Small in Fl. S.E. U.S.: 591 (1903) Heterotypic Synonyms: <i>Butea loureiroi</i> Spreng. in Syst. Veg., ed. 16. 3: 186 (1826), nom. superfl.; <i>Caesalpinia axillaris</i> (Lam.) DC. in Prodr. 2: 481 (1825); <i>Caesalpinia crista</i> var. <i>parvistipula</i> Urb. in Symb. Antill. 2: 271 (1900); <i>Caesalpinia kwangtungensis</i> Merr. in J. Arnold Arbor. 8: 7 (1927); <i>Caesalpinia laevigata</i> Perr. in Mém. Soc. Linn. Paris 3: 104 (1825); <i>Caesalpinia nuga</i> (L.) W.T.Aiton in Hortus Kew. 3: 32 (1811); <i>Caesalpinia paniculata</i> (Lam.) Roxb. in Hort. Bengal.: 32 (1814); <i>Caesalpinia scandens</i> B.Heyne ex Roth in Nov. Pl. Sp.: 209 (1821); <i>Caesalpinia scandens</i> J.Koenig ex Baker in J.D.Hooker, Fl. Brit. India 2: 255 (1878), pro syn.; <i>Genista scandens</i> Lour. in Fl. Cochinch.: 428 (1790); <i>Guilandina axillaris</i> Lam. in Encycl. 1: 435 (1785); <i>Guilandina nuga</i> L. in Sp. Pl., ed. 2.: 545 (1762); <i>Guilandina paniculata</i> Lam. in Encycl. 1: 435 (1785); <i>Guilandina parvifolia</i> Stokes in Bot. Mat. Med. 2: 466 (1812); <i>Guilandina rotunda</i> Noronha in Verh. Batav. Genootsch. Kunsten 5(4): 16 (1790), nom. nud.; <i>Ticanto nuga</i> (L.) Medik. in Theodora: 52 (1786)	Climbing shrub or liana	Tropical & Subtropical Asia to W. Pacific	Seasonally dry tropical biome. On sandy beaches, mangrove swamp and river margins, on limestone rocks or in secondary scrub. Mountain slopes, forests. Alt. 400-1500 m	1, 2, 5, 13
<i>Capsella rubella</i> Reut.	<i>Capsella rubella</i> Reut., Compt.-Rend. Trav. Soc. Hallér 1853-	<i>Brassicaceae</i> Burnett	Homotypic Synonyms: <i>Bursa rubella</i> (Reut.) Druce in Rep. Bot. Soc. Exch. Club Brit. Isles 7: 864 (1925 publ. 1926); <i>Capsella bursa-pastoris rubella</i> (Reut.) E.B.Almq. in Acta Horti Berg. 7: 53 (1921); <i>Capsella bursa-pastoris</i> subsp. <i>rubella</i> (Reut.) Hobk. in Bull. Soc. Roy. Bot. Belgique 8: 455 (1869); <i>Capsella bursa-pastoris</i> var. <i>rubella</i> (Reut.) Batt.	Annual or biennial	S. Central Europe to Medit	Temperate biome. Arid crops. Alt. 0-1300 m	1, 2, 10

1854: 18
(1853)

in J.A.Battandier & L.C.Trabut, Fl. Algérie, Dicot.: 42 (1888); *Crucifera rubella* (Reut.) E.H.L.Krause in J.W.Sturm, Deutschl. Fl. Abbild. ed. 2. 6: 145 (1902); *Thlaspi bursa-pastoris* var. *rubellum* (Reut.) Loret in H.Loret & A.Barrandon, Fl. Montpellier, éd. 2: 47 (1886); *Thlaspi rubellum* (Reut.) Loret & Barrandon in Fl. Montpellier: 60 (1876)
Heterotypic Synonyms: *Bursa alpestris* Miégev. in Bull. Soc. Bot. France 3: 10 (1863); *Bursa occidentalis* Shull in Proc. Int. Congr. Pl. Sci. Ithaca: 847 (1929); *Capsella bursa-pastoris* var. *alpestris* (Miégev.) Rouy & Foucaud in Fl. France 2: 95 (1895); *Capsella bursa-pastoris concava* E.B.Almq. in Acta Horti Berg. 4(6): 12 (1907); *Capsella bursa-pastoris* var. *crispata* Rouy & Foucaud in Fl. France 2: 95 (1895); *Capsella bursa-pastoris* proles *humilis* Rouy & Foucaud in Fl. France 2: 96 (1895); *Capsella bursa-pastoris* subsp. *occidentalis* (Shull) Maire in É.Jahandiez & al., Cat. Pl. Maroc 2: 297 (1932); *Capsella concava* (E.B.Almq.) E.B.Almq. in Rep. Bot. Soc. Exch. Club Brit. Isles 6: 200 (1920 publ. 1921); *Capsella rubella* var. *sabulosa* (Jord.) Caldesi in Nuovo Giorn. Bot. Ital. 11: 331 (1879); *Capsella rubescens* Personnat in Bull. Soc. Bot. France 7: 511 (1860); *Capsella sabulosa* Jord. in Diagn. Esp. Nouv.: 341 (1864)

Cheilanthes farinosa (Forssk.) Kaulf.
Hemionitis farinosa (Forssk.) Christenh., Global Fl. 4: 13 (2018)
Pteridaceae
E.D.M.Kirchn

Homotypic Synonyms: *Aleuritopteris farinosa* (Forssk.) Fée in Mém. Foug., 5. Gen. Filic.: 154 (1852); *Allosorus farinosus* (Forssk.) C.Presl in Tent. Pterid.: 153 (1836); *Cassebeera farinosa* (Forssk.) J.Sm. in J. Bot. (Hooker) 4: 159 (1841); *Cheilanthes farinosa* (Forssk.) Kaulf. in Enum. Filic.: 212 (1824); *Pteris farinosa* Forssk. in Fl. Aegypt.-Arab.: 187 (1775)
Heterotypic Synonyms: *Aleuritopteris chihuahuensis* Saiki in J. Phytogeogr. Taxon. 32: 85 (1984); *Aleuritopteris mexicana* Fée in Mém. Foug., 5. Gen. Filic.: 154 (1852); *Aleuritopteris peruviana* Saiki in J. Phytogeogr. Taxon. 32: 89 (1984); *Cheilanthes chihuahuensis* (Saiki) Fraser-Jenk. in New Sp. Syndr. Indian Pteridol.: 81 (1997); *Cheilanthes mexicana* (Fée) Punetha & Kholia in New Botanist, Int. Quart. J. Pl. Sci. Res. 16: 119 (1989), nom. illeg.; *Cheilanthes pulveracea* C.Presl in Reliq. Haenk. 1: 64 (1825); *Cheilanthes rigidula* Wall. in Numer. List: n.° 2175 (1830), not validly publ.; *Hemionitis chihuahuensis* (Saiki) Christenh. in Global Fl. 4: 11 (2018); *Pellaea pulcherrima* Rovirosa in Pteridogr. Sur Mexico: t. 14 (1909), nom. nud.

Lithophyte
Mexico to Venezuela and Peru, Africa to Arabian Peninsula
Wet tropical biome. Moist, wooded, rocky slopes.
Alt. 1450-3100 m
1, 2

Colquhounia coccinea var. *mollis* (Schltdl.) Prain
Colquhounia coccinea var. *mollis* (Schltdl.) Prain, J. Asiat. Soc. Bengal, Pt. 2, Nat.
Lamiaceae
Martinov

Homotypic Names: Basionym/Replaced Synonym *Colquhounia mollis* Schltdl., Linnaea 24: 681 (1852); Heterotypic Synonyms: *Colquhounia tomentosa* Houlet, Rev. Hort. (Paris) 45: 131 (1873); *Colquhounia vestita* var. *rugosa* C.B.Clarke ex Prain, J. Asiat. Soc. Bengal, Pt. 2, Nat. Hist. 62: 37 (1893).

Shrub
Central Himalaya to N. Thailand
Subtropical biome. Stony, grassy slopes, thickets, rarely forests.
Alt. 1400-3000 m
5

	Hist., 62: 37, 1893						
<i>Corallocarpus epigaeus</i> (Roettl.) Hook. f.	<i>Corallocarpus epigaeus</i> (Roettl.) Hook. f., Hook. f., Fl. Brit. Ind. 628. 1879	<i>Cucurbitaceae</i> Juss.	Homotypic name: <i>Bryonia epigaea</i> Rottler, Neue Schriften Ges. Naturf. Freunde Berlin 4: 223 (1803); Heterotypic Synonyms: <i>Aechmandra epigaea</i> (Rottler) Arn., J. Bot. (Hooker) 3: 274 (1841); <i>Coniandra corallina</i> Fenzl ex Naudin, Ann. Sci. Nat., Bot., sér. 4, 16: 180 (1862); <i>Corallocarpus bequaertii</i> De Wild., Rev. Zool. Bot. Africaines 9: 91 (1921); <i>Corallocarpus corallinus</i> (Naudin) Cogn., A.L.P.P.de Candolle & A.C.P.de Candolle, Monogr. Phan. 3: 647 (1881); <i>Corallocarpus elegans</i> Gilg, Bot. Jahrb. Syst. 34: 364 (1904); <i>Corallocarpus gracilipes</i> (Naudin) Cogn., A.L.P.P.de Candolle & A.C.P.de Candolle, Monogr. Phan. 3: 650 (1881); <i>Corallocarpus hildebrandtii</i> Gilg, Bot. Jahrb. Syst. 34: 362 (1904); <i>Corallocarpus palmatus</i> Cogn., A.L.P.P.de Candolle & A.C.P.de Candolle, Monogr. Phan. 3: 648 (1881); <i>Corallocarpus tavetensis</i> Gilg, Bot. Jahrb. Syst. 34: 362 (1904); <i>Rhynchocarpa corallina</i> Naudin, Ann. Sci. Nat., Bot., sér. 4, 16: 180 (1861); <i>Rhynchocarpa epigaea</i> (Rottler) Naudin, Ann. Sci. Nat., Bot., sér. 4, 16: 178 (1861)	Climbing caudex geophyte	Senegal to Eritrea and Tanzania, Arabian Peninsula, Indian Subcontinent	Seasonally dry tropical biome. Margins of rain-forest, wooded grassland, deciduous (<i>Commiphora</i>) bushland; grassy savanna. Alt. 100-2100 m	1-3, 6, 14, 15
<i>Croton hieronymi</i> Griseb.	<i>Croton hieronymi</i> Griseb., Abh. Königl. Ges. Wiss. Göttingen 24: 54 (1879)	<i>Euphorbiaceae</i> Juss.	Homotypic Names: <i>Oxydectes hieronymi</i> (Griseb.) Kuntze, Revis. Gen. Pl. 3(2): 289 (1898)	Shrub	Bolivia to NW. Argentina	Desert or dry shrubland biome. Lowlands. Alt. 300-1100 m	1-3, 6, 16, 17
<i>Cydonia vulgaris</i> Pers.	<i>Cydonia oblonga</i> Mill., Gard. Dict. ed. 8: n.º1 (1768)	<i>Rosaceae</i> Juss.	Homotypic Synonyms: <i>Cydonia communis</i> Loisel. in H.L.Duhamel du Monceau, Traité Arbr. Arbust., nouv. éd., 4: 135 (1809), nom. superfl.; <i>Cydonia communis</i> var. <i>oblonga</i> (Mill.) Loisel. in H.L.Duhamel du Monceau, Traité Arbr. Arbust., nouv. éd., 4: 136 (1809), not validly publ.; <i>Cydonia cydonia</i> (L.) Pers. in Syn. Pl. 2: 40 (1806), not validly publ.; <i>Cydonia cydonia</i> var. <i>oblonga</i> (Mill.) Asch. & Graebn. in Fl. Nordostdeut. Flachl.: 420, 803 (1898), not validly publ.; <i>Cydonia cydonia</i> var. <i>pyriformis</i> (Hayne) Asch. & Graebn. in Syn. Mitteleur. Fl. 6(2): 115 (1906), not validly publ.; <i>Cydonia silvestris</i> Risso in Hist. Nat. Prod. Eur. Mérid. 2: 296 (1826), nom. superfl.; <i>Cydonia silvestris</i> var. <i>oblonga</i> (Mill.) Risso in Hist. Nat. Prod. Eur. Mérid. 2: 298 (1826), not validly publ.; <i>Cydonia vulgaris</i> Tourn. ex Delarbre in Fl. Auvergne, ed. 2, 1: 384 (1800), nom. superfl.; <i>Cydonia vulgaris</i> var. <i>oblonga</i> (Mill.) DC. in Prodr. 2: 638 (1825), not validly publ.; <i>Cydonia vulgaris</i> var. <i>oblonga</i> (Mill.) Risso in Hist. Nat. Prod. Eur. Mérid. 2: 298 (1826), not validly publ.; <i>Cydonia vulgaris pyriformis</i> (Hayne) Dierb. in Syst. Uebers. Heidelb. Gewächse: 148 (1827), not validly publ.; <i>Cydonia vulgaris</i> f. <i>pyriformis</i> (Hayne) G.Kirchn. in E.A.Petzold & G.Kirchner, Arbor. Muscav.: 311 (1864), not validly publ.; <i>Cydonia vulgaris</i> var. <i>pyriformis</i> Hayne in	Tree	Caucasus to Central Asia.	Temperate biome. Cultivated throughout a large part of Europe; naturalized in hedges and thickets in South Europe and more locally in Central Europe. Alt. 0-1500 m	1, 3, 18

Getreue Darstell. Gew. 4: t. 47 (1816), not validly publ.; *Cydonia vulgaris silvestris* (Risso) M.Roem. in Fam. Nat. Syn. Monogr. 3: 217 (1847); *Pyrus cydonia* L. in Sp. Pl.: 480 (1753); *Pyrus cydonia oblonga* (Mill.) Kuntze in Taschen-Fl. Leipzig: 272 (1867), not validly publ.; *Pyrus cydonia* var. *oblonga* (Mill.) Martyn in P.Miller, Gard. Dict., ed. 9. 2: [s.p.] (1797), not validly publ.; *Pyrus cydonia* var. *pyriformis* (Hayne) Roth in Enum. Pl. Phaen. Germ. 1(2): 433 (1827), not validly publ.; *Pyrus oblonga* (Mill.) Steud. in Nomencl. Bot. 1: 670 (1821)

Heterotypic Synonyms: *Crataegus serotina* Salisb. in Prodr. Stirp. Chap. Allerton: 357 (1796); *Cydonia communis* var. *lusitanica* (Mill.) Loisel. in H.L.Duhamel du Monceau, Traité Arbr. Arbust., nouv. éd., 4: 136 (1809); *Cydonia communis* var. *maliformis* (Mill.) Loisel. in H.L.Duhamel du Monceau, Traité Arbr. Arbust., nouv. éd., 4: 136 (1809); *Cydonia cydonia* subvar. *lusitanica* (Mill.) Asch. & Graebn. in Syn. Mitteleur. Fl. 6(2): 115 (1906), not validly publ.; *Cydonia cydonia* var. *lusitanica* (Mill.) Pers. in Syn. Pl. 2: 40 (1806), not validly publ.; *Cydonia cydonia* var. *maliformis* (Mill.) Asch. & Graebn. in Syn. Mitteleur. Fl. 6(2): 115 (1906), not validly publ.; *Cydonia cydonia* monstr. *marmorata* (Dippel) Asch. & Graebn. in Syn. Mitteleur. Fl. 6(2): 115 (1906), not validly publ.; *Cydonia cydonia* *lusus pyramidalis* (Dippel) Asch. & Graebn. in Syn. Mitteleur. Fl. 6(2): 115 (1906), not validly publ.; *Cydonia europaea* Savi in Tratt. Alb. Toscana 1: 99 (1811); *Cydonia lusitanica* Mill. in Gard. Dict., ed. 8.: *Cydonia* n.° 3 (1768); *Cydonia maliformis* Mill. in Gard. Dict., ed. 8.: *Cydonia* n.° 2 (1768); *Cydonia media* Poit. & Turpin in H.L.Duhamel du Monceau, Traité Arbr. Fruit. 6: t. 328 (1829); *Cydonia minor* Poit. & Turpin in H.L.Duhamel du Monceau, Traité Arbr. Fruit. 6: t. 328 (1829); *Cydonia oblonga* var. *biserrulata* Kakhadze in Trudy Tbilissk. Univ. 109: 42 (1965), nom. illeg.; *Cydonia oblonga* subsp. *integerrima* Lobachev in Byull. Vsesoyuzn. Ordena Lenina Ordena Druzhby Narodov Nauchno-Issl. Inst. Rasteniev. N. I. Vavilova 113: 37 (1981); *Cydonia oblonga* var. *integerrimosepala* Lobachev in Byull. Vsesoyuzn. Ordena Lenina Ordena Druzhby Narodov Nauchno-Issl. Inst. Rasteniev. N. I. Vavilova 75: 70 (1977); *Cydonia oblonga* var. *integerrimosepala* Kakhadze in Trudy Tbilissk. Univ. 109: 43 (1965), nom. illeg.; *Cydonia oblonga* f. *lusitanica* (Mill.) Rehder in Bibliogr. Cult. Trees: 278 (1949); *Cydonia oblonga* subsp. *lusitanica* (Mill.) D.Rivera in D.Rivera & al., Varied. Trad. Frut. Cuenca Río Segura Cat. Etnobot.: 130 (1997); *Cydonia oblonga* var. *lusitanica* (Mill.) C.K.Schneid. in Ill. Handb. Laubholzk. 1: 654 (1906); *Cydonia oblonga* var. *maliformis* (Mill.) C.K.Schneid. in Ill. Handb. Laubholzk. 1: 654 (1906); *Cydonia oblonga* subsp. *maliformis* (Mill.) Thell. in Mitt. Bot. Mus. Univ. Zürich 58: 289 (1912); *Cydonia oblonga* f. *marmorata* (Dippel) C.K.Schneid. in Ill. Handb. Laubholzk. 1: 654 (1906); *Cydonia oblonga* var. *obpyricarpa* Lobachev in Byull. Vsesoyuzn. Ordena

Lenina Ordena Druzhby Narodov Nauchno-Issl. Inst. Rasteniev. N. I. Vavilova 113: 37 (1981); *Cydonia oblonga* var. *obpyriformis* Lobachev in Byull. Vsesoyuzn. Ordena Lenina Ordena Druzhby Narodov Nauchno-Issl. Inst. Rasteniev. N. I. Vavilova 113: 37 (1981); *Cydonia oblonga* var. *orbiculatocomplanata* Lobachev in Byull. Vsesoyuzn. Ordena Lenina Ordena Druzhby Narodov Nauchno-Issl. Inst. Rasteniev. N. I. Vavilova 113: 37 (1981); *Cydonia oblonga* var. *ovalicarpa* Lobachev in Byull. Vsesoyuzn. Ordena Lenina Ordena Druzhby Narodov Nauchno-Issl. Inst. Rasteniev. N. I. Vavilova 113: 37 (1981); *Cydonia oblonga* var. *ovalis* Lobachev in Byull. Vsesoyuzn. Ordena Lenina Ordena Druzhby Narodov Nauchno-Issl. Inst. Rasteniev. N. I. Vavilova 113: 36 (1981); *Cydonia oblonga* var. *plano-cyclocarpa* Lobachev in Byull. Vsesoyuzn. Ordena Lenina Ordena Druzhby Narodov Nauchno-Issl. Inst. Rasteniev. N. I. Vavilova 113: 36 (1981); *Cydonia oblonga* var. *pomiformis* Lobachev in Byull. Vsesoyuzn. Ordena Lenina Ordena Druzhby Narodov Nauchno-Issl. Inst. Rasteniev. N. I. Vavilova 113: 37 (1981); *Cydonia oblonga* f. *pyramidalis* (Dippel) Rehder in Bibliogr. Cult. Trees: 278 (1949); *Cydonia oblonga* var. *pyramidalis* (Dippel) C.K.Schneid. in Ill. Handb. Laubholzk. 1: 654 (1906); *Cydonia oblonga* f. *pyriformis* (Thell.) Rehder in Bibliogr. Cult. Trees: 278 (1949); *Cydonia oblonga* subsp. *pyriformis* Thell. in Mitt. Bot. Mus. Univ. Zürich 58: 289 (1912); *Cydonia oblonga* var. *pyriformis* (Thell.) Buia in T.Săvulescu, Fl. Republ. Popul. Române 4: 201 (1956); *Cydonia oblonga* var. *rotundata* Kakhadze in Trudy Tbilissk. Univ. 109: 44 (1965); *Cydonia oblonga* var. *serrulata* C.K.Schneid. in unknown publication; *Cydonia oblonga* var. *typica* C.K.Schneid. in Ill. Handb. Laubholzk. 1: 654 (1906); *Cydonia oblonga* var. *typica* Kakhadze in Trudy Tbilissk. Univ. 109: 42 (1965), comb. not validly publ.; *Cydonia oblonga* var. *urceolata* Lobachev in Byull. Vsesoyuzn. Ordena Lenina Ordena Druzhby Narodov Nauchno-Issl. Inst. Rasteniev. N. I. Vavilova 113: 37 (1981); *Cydonia piriformis* Medik. ex B.D.Jacks. in Index Kew. 1: 681 (1893), not validly publ.; *Cydonia pyriformis* M.Roem. in Fam. Nat. Syn. Monogr. 3: 217 (1847), pro syn.; *Cydonia silvestris* var. *communis* Risso in Hist. Nat. Prod. Eur. Mérid. 2: 297 (1826); *Cydonia silvestris* var. *lusitanica* (Mill.) Risso in Hist. Nat. Prod. Eur. Mérid. 2: 297 (1826); *Cydonia silvestris* var. *maliformis* (Mill.) Risso in Hist. Nat. Prod. Eur. Mérid. 2: 297 (1826); *Cydonia silvestris* var. *microcarpa* Risso in Hist. Nat. Prod. Eur. Mérid. 2: 298 (1826); *Cydonia sumboshia* Buch.-Ham. ex D.Don in Prodr. Fl. Nepal.: 237 (1825); *Cydonia vulgaris* var. *algeriensis* Lavallée in Énum. Arbres: 111 (1877), nom. nud.; *Cydonia vulgaris* var. *anglica* M.Roem. in Fam. Nat. Syn. Monogr. 3: 218 (1847); *Cydonia vulgaris* var. *aurantia* M.Roem. in Fam. Nat. Syn. Monogr. 3: 218 (1847); *Cydonia vulgaris* var. *bourgeana* Lavallée in Énum. Arbres: 111 (1877), nom. nud.; *Cydonia vulgaris* var. *brusvicensis* M.Roem. in Fam. Nat. Syn.

			Monogr. 3: 218 (1847); <i>Cydonia vulgaris communis</i> (Risso) M.Roem. in Fam. Nat. Syn. Monogr. 3: 217 (1847); <i>Cydonia vulgaris</i> var. <i>constantinopolitana</i> Lavallée in Énum. Arbres: 111 (1877), nom. nud.; <i>Cydonia vulgaris</i> var. <i>georgica</i> M.Roem. in Fam. Nat. Syn. Monogr. 3: 218 (1847); <i>Cydonia vulgaris</i> f. <i>lusitanica</i> (Mill.) G.Kirchn. in E.A.Petzold & G.Kirchner, Arbor. Muscav.: 311 (1864); <i>Cydonia vulgaris</i> var. <i>lusitanica</i> (Mill.) Hayne in Getreue Darstell. Gew. 4: t. 47 (1816); <i>Cydonia vulgaris</i> var. <i>macrocarpa</i> Lavallée in Énum. Arbres: 111 (1877), nom. nud.; <i>Cydonia vulgaris macrocarpa</i> M.Roem. in Fam. Nat. Syn. Monogr. 3: 218 (1847); <i>Cydonia vulgaris</i> f. <i>maliformis</i> (Mill.) G.Kirchn. in E.A.Petzold & G.Kirchner, Arbor. Muscav.: 311 (1864); <i>Cydonia vulgaris</i> var. <i>maliformis</i> (Mill.) Hayne in Getreue Darstell. Gew. 4: t. 47 (1816); <i>Cydonia vulgaris</i> var. <i>marmorata</i> Dippel in Handb. Laubholz. 3: 357 (1893); <i>Cydonia vulgaris</i> var. <i>pyramidalis</i> Dippel in Handb. Laubholz. 3: 357 (1893); <i>Pyrus cydonia</i> var. <i>dulcis</i> Weston in Bot. Univ. 1: 230 (1770); <i>Pyrus cydonia</i> subsp. <i>lusitanica</i> (Mill.) Ehrh. in Beitr. Naturk. Verw. Wiss. 7: 52 (1792), not validly publ.; <i>Pyrus cydonia</i> var. <i>lusitanica</i> (Mill.) Weston in Bot. Univ. 1: 230 (1770); <i>Pyrus cydonia</i> var. <i>major</i> Weston in Bot. Univ. 1: 230 (1770); <i>Pyrus cydonia</i> var. <i>maliformis</i> (Mill.) Roth in Enum. Pl. Phaen. Germ. 1(2): 433 (1827); <i>Pyrus cydonia</i> subsp. <i>maliformis</i> (Mill.) Ehrh. in Beitr. Naturk. Verw. Wiss. 7: 52 (1792), not validly publ.; <i>Pyrus cydonia maliformis</i> (Mill.) Kuntze in Taschen-Fl. Leipzig: 272 (1867); <i>Pyrus cydonia</i> var. <i>minor</i> Weston in Bot. Univ. 1: 230 (1770); <i>Pyrus cydonia</i> subsp. <i>oblonga</i> (Mill.) Ehrh. in Beitr. Naturk. Verw. Wiss. 7: 52 (1792), not validly publ.; <i>Pyrus cydonia</i> var. <i>rotunda</i> Weston in Bot. Univ. 1: 230 (1770); <i>Pyrus cydonia</i> var. <i>sylvestris</i> Duhamel ex DC. in J.B.A.M.de Lamarck & A.P.de Candolle, Fl. Franç., ed. 3, 5: 430 (1805); <i>Pyrus lusitanica</i> (Mill.) Steud. in Nomencl. Bot. 1: 670 (1821); <i>Pyrus maliformis</i> (Mill.) Steud. in Nomencl. Bot. 1: 670 (1821); <i>Pyrus samboshia</i> Spreng. in Syst. Veg., ed. 16. 4(2): 196 (1827); <i>Pyrus-cydonia cydonia</i> Weston in Bot. Univ. 1: 230 (1770); <i>Pyrus-cydonia cydonia lusitanica</i> (Mill.) Weston in Bot. Univ. 1: 230 (1770); <i>Sorbus cydonia</i> Crantz in Stirp. Austr. Fasc. 2: 57 (1763)				
<i>Eurysolen gracilis</i> Prain	<i>Eurysolen gracilis</i> Prain, Sci. Mem. Off. Med. Dept. Gov. India 11: 44 (1898)	<i>Lamiaceae</i> Martinov	-	Scrambling subshrub or shrub	China (Yunnan) to W. & Central Malesia	Wet tropical biome. Open places in moist mountain valleys. Alt. 700-1800 m	1, 2, 19
<i>Gentianella alborosea</i> (Gilg) Fabris	<i>Gentianella alborosea</i> (Gilg) Fabris, Bol. Soc.	<i>Gentianaceae</i> Juss.	Homotypic Synonyms: <i>Gentiana alborosea</i> Gilg, Beibl. Bot. Jahrb. Syst. 118: 55 (1916)	Perennial	Peru (Junín)	Subalpine or subarctic biome. Bunch grass formation.	1, 2, 20-22

<i>Gentianella nitida</i> (Griseb.) Fabris	Argent. Bot. 6: 48 (1955) <i>Gentianella nitida</i> (Griseb.) Fabris, Bol. Soc. Argent. Bot. 7: 91 (1958)	<i>Gentianaceae</i> Juss.	Homotypic Synonyms: <i>Gentiana nitida</i> Griseb. in Gen. Sp. Gent.: 216 (1838)	Perennial	Peru	Alt. 4100-4700 m Subalpine or subarctic biome. Grassy uplands. Alt. 4200-4300 m	1, 2, 20-22
<i>Gentianella turkestanorum</i> (Gand.) Holub	<i>Gentianella turkestanorum</i> (Gand.) Holub, Folia Geobot. Phytotax. 2: 118 (1967)	<i>Gentianaceae</i> Juss.	Homotypic Synonyms: <i>Gentiana turkestanorum</i> Gand. in Bull. Soc. Bot. France 65: 60 (1918) Heterotypic Synonyms: <i>Aloitis stoliczkae</i> (Kurz ex C.B.Clarke) Omer, Qaiser & Ali in Pakistan J. Bot. 20: 159 (1988); <i>Gentiana stoliczkae</i> Kurz ex C.B.Clarke in J. Linn. Soc., Bot. 14: 433 (1875); <i>Gentianella stoliczkae</i> (Kurz ex C.B.Clarke) Holub in Folia Geobot. Phytotax. 8: 171 (1973)	Annual or biennial	Afghanistan to Mongolia and W. Himalaya	Subalpine or subarctic biome. Moist places around lakes, river banks, shady meadow slopes, forests. Alt. 1500-3100 m	1, 2, 5
<i>Gossypium hirsutum</i> L.	<i>Gossypium hirsutum</i> L., Sp. Pl. ed. 2: 975 (1763)	<i>Malvaceae</i> Juss.	Homotypic Synonyms: <i>Gossypium barbadense</i> var. <i>hirsutum</i> (L.) Hook.f. & Benth. in W.J.Hooker, Niger Fl.: 229 (1849); <i>Gossypium herbaceum</i> var. <i>hirsutum</i> (L.) Mast. in J.D.Hooker, Fl. Brit. India 1: 347 (1874); <i>Xylon hirsutum</i> (L.) Medik. in Malvenfam.: 43 (1787)	Shrub	Pacific, Mexico to Ecuador and NE. Brazil	Seasonally dry tropical biome. seasonally dry tropical biome. From the cool temperate moist to wet through tropical very dry to moist forest life zones. Widely cultivated. Alt. 0-1250 m	1-3
<i>Gynochthodes officinalis</i> (F.C.How) Razafim. & B.Bremer	<i>Gynochthodes officinalis</i> (F.C.How) Razafim. & B.Bremer, Adansonia, sér. 3, 33: 293 (2011)	<i>Rubiaceae</i> Juss.	Homotypic Synonym: <i>Morinda officinalis</i> F.C.How in Acta Phytotax. Sin. 7: 326 (1958). Heterotypic Synonym: <i>Morinda officinalis</i> var. <i>hirsuta</i> F.C.How in Acta Phytotax. Sin. 7: 328 (1958)	Climber	SE. China to Hainan	Wet tropical biome. Sparse or dense forests and thickets on mountains, also cultivated; 100-500 m.	1, 23
<i>Hedyosmum angustifolium</i> (Ruiz & Pav.) Solms	<i>Hedyosmum angustifolium</i> (Ruiz & Pav.) Solms	<i>Chloranthaceae</i> R.Br. ex Sims	Homotypic Synonyms: <i>Tafalla angustifolia</i> Ruiz & Pav. in Syst. Veg. Fl. Peruv. Chil.: 272 (1798) Heterotypic Synonyms: <i>Hedyosmum laciniatum</i> (Ruiz & Pav.) Solms, A.P.de Candolle, Prodr. 16(1): 485 (1869); <i>Hedyosmum pavonii</i> (Solms) Diels, Notizbl. Bot. Gart. Berlin-Dahlem 14: 331 (1939); <i>Hedyosmum</i>	Shrub or tree	SW. Colombia to Bolivia	Wet tropical biome. Disturbed and undisturbed montane forest.	1, 2, 6, 24

<i>Hedyosmum brasiliense</i> Mart.	<i>Hedyosmum brasiliense</i> Mart., Fl. Bras. 4(1): 3 (1852)	<i>Chloranthaceae</i> R.Br. ex Sims	<i>scabrum</i> var. <i>pavonii</i> Solms, A.P.de Candolle, Prodr. 16(1): 480 (1869); <i>Tafalla angustifolia</i> Ruiz & Pav., Syst. Veg. Fl. Peruv. Chil.: 272 (1798); <i>Tafalla laciniata</i> Ruiz & Pav., Syst. Veg. Fl. Peruv. Chil.: 272 (1798) Homotypic Synonyms: <i>Tafalla brasiliensis</i> (Mart.) Kuntze in Revis. Gen. Pl. 2: 566 (1891) Heterotypic Synonyms: <i>Hedyosmum acutifolium</i> Cordem. ex Baill. in Adansonia 3: 306 (1863); <i>Hedyosmum bonplandianum</i> Mart. in Syst. Mat. Med. Bras.: 100 (1843), sensu auct.; <i>Hedyosmum grandifolium</i> Occhioni in Contr. Estud. Fam. Chloranth.: 34 (1954); <i>Hedyosmum weddellianum</i> Cordem. ex Baill. in Adansonia 3: 306 (1863); <i>Tafalla weddelliana</i> (Cordem. ex Baill.) Kuntze in Revis. Gen. Pl. 2: 566 (1891); <i>Coccoloba brasiliensis</i> Spreng. in Syst. Veg. ed. 16. 2: 252 (1825), nom. Illeg.	Shrub or tree	Venezuela to Brazil and Paraguay	Alt. 900-2990 m Wet tropical biome. Gallery forest and adjacent grassy campo or rocky cerrado (central Brazil); sandy soils in campinarana (Parà); restinga vegetation near sea level to montane rainforest (Serra do Mar).	6, 24, 25
<i>Hedyosmum orientale</i> Merr. & Chun	<i>Hedyosmum orientale</i> Merr. & Chun, Sunyatsenia 5: 36 (1940)	<i>Chloranthaceae</i> R.Br. ex Sims	-	Subshrub or shrub	China (Guangdong) to Vietnam, Sumatra to Sulawesi	Alt. 0-1500 m Subtropical biome. Forests, thickets, forested ravines. Alt. ca. 500 m	1, 3, 23
<i>Hemidesmus indicus</i> (L.) R.Br.	<i>Hemidesmus indicus</i> (L.) R.Br., W.T.Aiton, Hortus Kew., ed. 2, 2: 75 (1811)	<i>Apocynaceae</i> Juss	Homotypic Names: Basionym/Replaced Synonym: <i>Periploca indica</i> L., Sp. Pl.: 211 (1753) Heterotypic Synonyms: <i>Periploca cordata</i> Dennst., Schlüssel Hortus Malab., 14 (1818), nom. illeg.; <i>Hemidesmus pubescens</i> Wight & Arn., Contr. Bot. India: 63 (1834); <i>Periploca malabarica</i> Burm. ex Decne. in A.P.de Candolle, Prodr. 8: 499 (1844), not validly publ.; <i>Hemidesmus indicus</i> var. <i>pubescens</i> (Wight & Arn.) Hook.f., Fl. Brit. India 4: 5 (1883)	Climbing shrub	Indian Subcontinent to Indo-China and Peninsula Malaysia	Seasonally dry tropical biome. Mesophytic to semi dry conditions, open scrub jungles, hedges, uncultivated soil (coal mine wastes).	1, 2, 6, 25-28
<i>Hypericum chinense</i> L.	<i>Hypericum monogynum</i> L., Sp. Pl. ed. 2.: 1107 (1763)	<i>Hypericaceae</i> Juss.	Homotypic Names: <i>Hypericum chinense</i> L., Syst. Nat. ed. 10, 2: 1184 (1759), nom. illeg.; <i>Ascyrum monogynum</i> (L.) Moench, Suppl. Meth.: 42 (1802); <i>Norysca chinensis</i> Spach, Hist. Nat. Vég. 5: 427 (1836), nom. superfl. Heterotypic Synonyms: <i>Hypericum aureum</i> Lour., Fl. Cochinch.: 472 (1790); <i>Hypericum sinense</i> J.F.Gmel., Syst. Nat. ed. 13[bis]: 1157 (1792), orth. var.; <i>Hypericum</i>	Shrub	Central & S. China, Taiwan	Alt. 0-600 m Temperate biome. Mountain slopes, roadsides, thickets in dry habitats.	1, 2, 5

			<i>salicifolium</i> Siebold & Zucc., Abh. Math.-Phys. Cl. Königl. Bayer. Akad. Wiss. 4(2): 162 (1845); <i>Norysca salicifolia</i> (Siebold & Zucc.) K.Koch, Hort. Dendrol.: 65 (1853); <i>Hypericum chinense</i> var. <i>salicifolium</i> (Siebold & Zucc.) Choisy in H.Zollinger, Syst. Verz. Ind. Archip. 2: 150 (1854); <i>Norysca aurea</i> (Lour.) Blume, Mus. Bot. 2: 23 (1856); <i>Norysca punctata</i> Blume, Mus. Bot. 2: 23 (1856); <i>Hypericum monogynum</i> var. <i>salicifolium</i> (Siebold & Zucc.) André, Rev. Hort. (Paris) 61: 464 (1889); <i>Hypericum chinense</i> subsp. <i>latifolium</i> Kuntze, Revis. Gen. Pl. 1: 60 (1891); <i>Hypericum chinense</i> subsp. <i>obtusifolium</i> Kuntze, Revis. Gen. Pl. 1: 60 (1891); <i>Hypericum chinense</i> subsp. <i>salicifolium</i> (Siebold & Zucc.) Kuntze, Revis. Gen. Pl. 1: 60 (1891); <i>Komana salicifolia</i> (Siebold & Zucc.) Y.Kimura in M.Honda, Nom. Pl. Jap.: 222 (1939); <i>Norysca chinensis</i> var. <i>salicifolia</i> (Siebold & Zucc.) Y.Kimura in T.Nakai & M.Honda, Nov. Fl. Jap. 10: 107 (1951).			Alt. 0-1500 m	
<i>Hypericum longistylum</i> Oliv.	<i>Hypericum longistylum</i> Oliv., Hooker's Icon. Pl. 16: t. 1534 (1886)	<i>Hypericaceae</i> Juss.	Homotypic Synonyms: <i>Norysca longistyla</i> (Oliv.) Y.Kimura in T.Nakai & M.Honda, Nov. Fl. Jap. 10: 98 (1951)	Shrub	Central & S. China	Temperate biome. Open sunny places, cliffs, dry banks and slopes, streamsides. Alt. 200-2100 m.	1, 3, 23
<i>Inula britannica</i> L.	<i>Pentanema britannica</i> (L.) D.Gut.Larr., Santos-Vicente, Anderb., E.Rico & M.M.Mart.Ort., Taxon 67: 159 (2018). Also reported as <i>Pentanema britannicum</i> ²	<i>Asteraceae</i> Bercht. & J.Presl	Homotypic Synonyms: <i>Aster britannicus</i> (L.) All. in Fl. Pedem. 1: 197 (1785); <i>Conyza britannica</i> (L.) Moris ex Rupr. in Fl. Ingrica: 569 (1856); <i>Helenium britannica</i> (L.) Kuntze in Revis. Gen. Pl. 1: 342 (1891); <i>Inula britannica</i> L. in Sp. Pl.: 882. (1753) Heterotypic Synonyms: <i>Aster orientalis</i> S.G.Gmel. in Reise Russland 1: 155 (1770); <i>Aster undulatus</i> Moench in Methodus: 600 (1794), nom. illeg.; <i>Helenium macrolepis</i> Kuntze in Revis. Gen. Pl. 1: 343 (1891); <i>Helenium microcephalum</i> Kuntze in Revis. Gen. Pl. 1: 343 (1891), nom. illeg.; <i>Helenium repandum</i> Kuntze in Revis. Gen. Pl. 1: 343 (1891); <i>Inula britannica</i> var. <i>angustifolia</i> Becker in Denkschr. Kaiserl. Akad. Wiss., Wien. Math.-Naturwiss. Kl. 44: 318 (1882), nom. illeg.; <i>Inula britannica</i> var. <i>angustifolia</i> Boenn. in Prodr. Fl. Monast. Westphal.: 256 (1824); <i>Inula britannica</i> var. <i>comosa</i> (Lam.) Mérat in Nouv. Fl. Env. Paris: 329 (1812); <i>Inula britannica</i> var. <i>eglandulosa</i> R.Nabiev in Opred. Rast. Sred. Azii 10: 632 (1993); <i>Inula britannica</i> subsp. <i>hispanica</i> (Pau) O.Bolòs & Vigo in Fontqueria 14: 9 (1987); <i>Inula britannica</i> var. <i>intermedia</i> Rosén & Wahlenb. in Nova Acta Regiae Soc. Sci. Upsal., ser. 2, 8: 248 (1821); <i>Inula britannica</i> subsp. <i>latifolia</i> U.P.Pratov & R.Nabiev in Opred. Rast. Sred. Azii 10: 632 (1993); <i>Inula britannica</i> var. <i>longilepis</i> R.Nabiev in Opred. Rast. Sred. Azii 10: 633 (1993); <i>Inula britannica</i> var. <i>microcephala</i> Velen. in Abh. Königl. Böhm. Ges. Wiss., ser. 7, 1(8): 22 (1886); <i>Inula britannica</i> var. <i>oelandica</i> Ahlq. in Kongl. Vetensk. Acad.	Perennial	Europe to Russian Far East and Himalaya	Temperate biome. Wet habitats: river and stream margins, marshes, ditches, wet grassland, and wet woods. Alt. 300-1700 m	1, 2, 5, 29

Handl. 1821: 301 (1821); *Inula britannica* var. *ramosissima* Ledeb. in Fl. Ross. 2: 506 (1845); *Inula britannica* var. *sericans* Zalewski in Kosmos (Lvov) 5/6: 17 (1896); *Inula britannica* var. *stricta* Rosén & Wahlenb. in Nova Acta Regiae Soc. Sci. Upsal., ser. 2, 8: 248 (1821); *Inula britannica* f. *sublanata* (Kom.) Kitag. in Neolin. Fl. Manshur.: 648 (1979); *Inula britannica* var. *sublanata* Kom. in Trudy Imp. S.-Peterburgsk. Bot. Sada 25: 626 (1907); *Inula britannica* var. *tymiensis* Kudô in Contrib. Fl. North Sachal.: 58 (1923); *Inula britannica* var. *uniflora* Gaudin in Fl. Helv. 5: 320 (1829); *Inula britannica* var. *viridis* Rosén & Wahlenb. in Nova Acta Regiae Soc. Sci. Upsal., ser. 2, 8: 248 (1821); *Inula chinensis* (Kom.) Kom. in Key Pl. Far East. Reg. USSR 2: 1024 (1954); *Inula comosa* Lam. in Fl. Franç. 2: 147 (1779); *Inula dichotoma* Zuccagni in Cent. Observ. Bot.: 40 (1806); *Inula encelioides* Hornem. ex Ledeb. in Fl. Ross. 2: 506 (1845); *Inula hirta* Pollich in Hist. Pl. Palat. 2: 467 (1777), nom. illeg.; *Inula hispanica* Pau in Not. Bot. Fl. Españ. 6: 62 (1895); *Inula micranthos* DC. in Prodr. 5: 467 (1836); *Inula microcephala* Borbás in Bot. Jahrb. Syst. 8: 235 (1887); *Inula oetteliana* Rchb. in J.C.Mössler, Handb. Gewächsk., ed. 2. 2: 1506 (1829); *Inula orientalis* d'Urv. ex Boiss. in Fl. Orient. 3: 193 (1875), not validly publ.; *Inula serrata* Gilib. in Fl. Lit. Inch. 1: 207 (1782), opus utique oppr.; *Inula squarrosa* Krock. in Fl. Siles. 2(2): 442 (1790), sensu auct.; *Inula tymensis* Kudô in Contrib. Fl. North Sachal.: 58 (1923), not validly publ.; *Inula vaillantii* Schur ex Nyman in Consp. Fl. Eur.: 393 (1879)

<i>Kandelia obovata</i> Sheue, H.Y.Liu & J.W.H.Yong	<i>Kandelia obovata</i> Sheue, H.Y.Liu & J.W.H.Yong, Taxon 52: 291 (2003)	<i>Rhizophoraceae</i> Pers.		Shrub or tree.	SE. China to Vietnam, Japan (S. Kyushu) to Taiwan	Subtropical biome. Margins of mangrove swamps and muddy or sandy tidal flats; sea level.	1, 23
<i>Leucosceptrum canum</i> Sm.	<i>Leucosceptrum canum</i> Sm., Exot. Bot. 2: 113 (1806)	<i>Lamiaceae</i> Martinov	Homotypic Names: <i>Clerodendrum leucosceptrum</i> D.Don, Prodr. Fl. Nepal.: 103 (1825); <i>Teucrium leucosceptrum</i> (D.Don) Voigt, Hort. Suburb. Calcutt.: 463 (1845). Heterotypic Synonyms: <i>Teucrium macrostachyum</i> Wall. ex Benth., Labiat. Gen. Spec.: 664 (1835); <i>Comanthosphace nepalensis</i> Kitam. & Murata, Acta Phytotax. Geobot. 15: 109 (1954).	Shrub or tree	Himalaya to S. Central China and Indochina	Subtropical biome. Dry open waste areas, forest margins, valley streamsides, second growth forests, thickets. Alt. 1000-2600 m	1-3, 5, 6
<i>Lindera glauca</i> (Siebold & Zucc.) Blume	<i>Lindera glauca</i> (Siebold & Zucc.) Blume,	<i>Lauraceae</i> Juss.	Homotypic Synonyms: <i>Benzoin glaucum</i> Siebold & Zucc. in Abh. Math.-Phys. Cl. Königl. Bayer. Akad. Wiss. 4(3): 205 (1846).	Shrub or tree	Central & S. China to Indo-China	Temperate biome. Forests, roadsides on	1, 2, 5

	Mus. Bot. 1: 325 (1851)		Heterotypic Synonyms: <i>Benzoin glaucum</i> var. <i>kawakamii</i> (Hayata) Sasaki in List Pl. Formosa: 191 (1928); <i>Lindera glauca</i> f. <i>glabella</i> (Nakai) C.M.Pak in Fl. Coreana 2: 118 (1996); <i>Lindera glauca</i> var. <i>kawakamii</i> Hayata in J. Coll. Sci. Imp. Univ. Tokyo 30(1): 255 (1911); <i>Lindera glauca</i> var. <i>parvifolia</i> K.M.Li & W.D.Han in J. Nanjing Forest. Univ. 13(4): 88 (1989)			and Korea, S. Central & S. Japan, Taiwan	mountain slopes. Alt. 0-900 m	
<i>Nicotiana benthamiana</i> Domin	<i>Nicotiana benthamiana</i> Domin, Biblioth. Bot. 22(89): 591 (1929)	<i>Solanaceae</i> Juss.	Homotypic Synonyms: <i>Nicotiana suaveolens</i> var. <i>cordifolia</i> Benth. in Fl. Austral. 4: 470 (1868), nom. cons. prop.	Annual	N. Western Australia	Desert or dry shrubland biome. Sheltered sites, usually on the south-facing sides of rock outcrops and cliffs and in gorges of various compositions. Alt. ...	1, 2, 30, 31	
<i>Oryza sativa</i> L.	<i>Oryza sativa</i> L., Sp. Pl.: 333 (1753)	<i>Poaceae</i> Barnhart	Homotypic Synonyms: <i>Oryza palustris</i> Salisb. in Prodr. Stirp. Chap. Allerton: 25 (1796), nom. superfl. Heterotypic Synonyms: <i>Oryza aristata</i> Blanco in Fl. Filip.: 274 (1837), nom. illeg.; <i>Oryza communissima</i> Lour. in Fl. Cochinch.: 215 (1790); <i>Oryza denudata</i> (Desv.) Steud. in Nomencl. Bot. 1: 577 (1821), nom. provis.; <i>Oryza elongata</i> (Desv.) Steud. in Nomencl. Bot. 1: 577 (1821), nom. provis.; <i>Oryza formosana</i> Masam. & S.Suzuki in Trans. Nat. Hist. Soc. Formosa 25: 320 (1935); <i>Oryza glutinosa</i> Lour. in Fl. Cochinch.: 215 (1790); <i>Oryza marginata</i> (Desv.) Steud. in Nomencl. Bot. 1: 577 (1821), nom. provis.; <i>Oryza montana</i> Lour. in Fl. Cochinch.: 215 (1790); <i>Oryza mutica</i> Steud. in Nomencl. Bot. 1: 577 (1821), nom. nud.; <i>Oryza nepalensis</i> G.Don ex Steud. in Syn. Pl. Glumac. 1: 3 (1853), nom. nud.; <i>Oryza parviflora</i> P.Beauv. in Ess. Agrostogr.: 27 (1812), nom. nud.; <i>Oryza perennis</i> Moench in Methodus: 197 (1794); <i>Oryza plena</i> (Prain) N.P.Chowdhury in Indian Forester 75: 497 (1949); <i>Oryza praecox</i> Lour. in Fl. Cochinch.: 215 (1790); <i>Oryza pubescens</i> (Desv.) Steud. in Nomencl. Bot. 1: 577 (1821), nom. provis.; <i>Oryza pumila</i> Steud. in Nomencl. Bot., ed. 2, 2: 234 (1841), pro syn.; <i>Oryza repens</i> Buch.-Ham. ex Steud. in Syn. Pl. Glumac. 1: 3 (1853), nom. nud.; <i>Oryza rubriarbis</i> (Desv.) Steud. in Nomencl. Bot. 1: 577 (1821), nom. provis.; <i>Oryza sativa</i> var. <i>acutiligula</i> Miq. in Fl. Ned. Ind. 3: 370 (1855); <i>Oryza sativa</i> var. <i>adversoseluana</i> Portères in J. Agric. Trop. Bot. Appl. 2: 573, 579 (1955); <i>Oryza sativa</i> var. <i>affinis</i> Körn. in F.A.Körnicke & H.Werner, Handb. Getreidebaus 1: 234 (1885); <i>Oryza sativa</i> var. <i>alaotrana</i> Portères in J. Agric. Trop. Bot. Appl.	Annual or helophyte	Cultigen from China	Temperate biome. Cultivated, mainly in flooded fields. Alt. 0-2000 m	1, 2, 23	

2: 573, 595 (1955); *Oryza sativa* f. *alba* (Miq.) Makino in Pract. Hort. 25: 1261 (1940); *Oryza sativa* var. *alba* Miq. in Fl. Ned. Ind. 3: 369 (1855); *Oryza sativa* var. *allachabadica* Gustchin in Riz & Rizic. 8: 34 (1934); *Oryza sativa* var. *alterogoensis* Portères in J. Agric. Trop. Bot. Appl. 2: 573, 595 (1955); *Oryza sativa* var. *amaura* Alef. in Landw. Fl.: 318 (1866); *Oryza sativa* var. *anandica* Gustchin in Riz & Rizic. 8: 33 (1934); *Oryza sativa* var. *angusta* Portères in J. Agric. Trop. Bot. Appl. 2: 574, 595 (1955); *Oryza sativa* var. *anjanica* Gustchin in Riz & Rizic. 8: 33 (1934); *Oryza sativa* var. *anthropophaga* Portères in J. Agric. Trop. Bot. Appl. 2: 574, 595 (1955); *Oryza sativa* var. *aquatica* Roshev. in Trudy Prikl. Bot. Selektiv. 27(4): 41 (1931); *Oryza sativa* subvar. *aristata* A.Camus in Bull. Lab. Agron. Coloniale 1: 23 (1913), nom. illeg.; *Oryza sativa* subvar. *aristata* A.Camus in Bull. Lab. Agron. Coloniale 1: 28 (1913), nom. illeg.; *Oryza sativa* var. *aristata* A.Rich. in R.de la Sagra, Hist. Fis. Cuba, Bot. 11: 296 (1850), nom. illeg.; *Oryza sativa* var. *aristida* Blanco in Fl. Filip.: 273 (1837); *Oryza sativa* var. *aromatica* Gustchin in Riz & Rizic. 8: 33 (1934); *Oryza sativa* var. *assamica* Gustchin in Riz & Rizic. 8: 31 (1934); *Oryza sativa* var. *astaranica* Gustchin in Riz & Rizic. 8: 30 (1934); *Oryza sativa* var. *astarinica* Jabrova in Trudy Azerbaidzhansk. Otd. Zakavkazsk. Fil. Akad. Nauk S.S.S.R. 2: 50 (1935); *Oryza sativa* var. *atra* Körn. in F.A.Körnische & H.Werner, Handb. Getreidebaus 1: 234 (1885); *Oryza sativa* var. *atricolorata* Gustchin in Riz & Rizic. 8: 34 (1934); *Oryza sativa* var. *atrobrunnea* Gustchin in Riz & Rizic. 8: 29 (1934); *Oryza sativa* var. *atrofusca* Körn. in F.A.Körnische & H.Werner, Handb. Getreidebaus 1: 233 (1885); *Oryza sativa* var. *atrogrisea* Gustchin in Riz & Rizic. 8: 37 (1934); *Oryza sativa* var. *atropurpurea* Makino in Pract. Hort. 25: 1259 (1940); *Oryza sativa* f. *atropurpurea* (Makino) Makino in Acta Phytotax. Geobot. 9: 103 (1940); *Oryza sativa* var. *aurica* Gustchin in Riz & Rizic. 8: 37 (1934); *Oryza sativa* var. *auriculata* Portères in J. Agric. Trop. Bot. Appl. 2: 574, 595 (1955); *Oryza sativa* var. *azerbaidjanica* Gustchin in Riz & Rizic. 8: 35 (1934); *Oryza sativa* var. *badia* Portères in J. Agric. Trop. Bot. Appl. 2: 574, 595 (1955); *Oryza sativa* var. *balatensis* Portères in J. Agric. Trop. Bot. Appl. 2: 574, 595 (1955); *Oryza sativa* var. *bandica* Gustchin in Riz & Rizic. 8: 31 (1934); *Oryza sativa* var. *bansica* Gustchin in Riz & Rizic. 8: 30 (1934); *Oryza sativa* var. *bansmatica* Gustchin in Riz & Rizic. 8: 29 (1934); *Oryza sativa* var. *bastica* Gustchin in Riz & Rizic. 8: 35 (1934); *Oryza sativa* var. *baumannii* Körn. & K.Schum. in H.G.A.Engler, Pflanzenw. Ost-Afrikas, C: 61 (1895); *Oryza sativa* var. *bawako-mochii* Gustchin in Riz & Rizic. 8: 32 (1934); *Oryza sativa* var. *bicolorata* Gustchin in Riz & Rizic. 8: 35 (1934); *Oryza sativa* var. *binamban* Blanco in Fl. Filip.: 273 (1837); *Oryza sativa* var. *birainica* Gustchin in Riz & Rizic. 8: 37 (1934); *Oryza sativa* var. *breviaristata* Vavilov in Trudy Prikl. Bot., Prilozh. 33: 313 (1929); *Oryza*

sativa proles *brevijaponica* Portères in Rev. Int. Bot. Appl. Agric. Trop. 30: 151 (1950); *Oryza sativa* subsp. *brevindica* (Portères) Vasc. in Trab. Centro Bot. Junta Invest. Ultramar 8: ? (1953); *Oryza sativa* proles *brevindica* Portères in Rev. Int. Bot. Appl. Agric. Trop. 30: 151 (1950); *Oryza sativa* subsp. *brevis* Gustchin in Riz & Rizic. 8: 28 (1934); *Oryza sativa* var. *brjezitsky* Jabrova in Trudy Azerbaidzhansk. Otd. Zakavkazsk. Fil. Akad. Nauk S.S.S.R. 2: 47, 50 (1935); *Oryza sativa* var. *brunnea* Körn. in F.A.Körnische & H.Werner, Handb. Getreidebaus 1: 233 (1885); *Oryza sativa* var. *brunneopunctata* Gustchin in Riz & Rizic. 8: 37 (1934); *Oryza sativa* var. *calvescens* Miq. in Fl. Ned. Ind. 3: 369 (1855); *Oryza sativa* var. *caroliniana* Postiglione in Relaz. Intorno Condizioni Agric.: 266 (1876); *Oryza sativa* var. *caspica* Körn. in F.A.Körnische & H.Werner, Handb. Getreidebaus 1: 233 (1885); *Oryza sativa* var. *castanea* Portères in J. Agric. Trop. Bot. Appl. 2: 575, 595 (1955); *Oryza sativa* var. *castroi* Vasc. in Anais Inst. Super. Agron. 19: 3 (1952); *Oryza sativa* var. *catalonica* Körn. in F.A.Körnische & H.Werner, Handb. Getreidebaus 1: 233 (1885); *Oryza sativa* var. *caucasica* Batalin ex Tutteff in Bot. Arch. 26: 154 (1929); *Oryza sativa* var. *ceylonica* Gustchin in Riz & Rizic. 8: 30 (1934); *Oryza sativa* var. *chambhanica* Gustchin in Riz & Rizic. 8: 31 (1934); *Oryza sativa* var. *chlamydantha* Portères in J. Agric. Trop. Bot. Appl. 2: 575, 595 (1955); *Oryza sativa* var. *chlorocarpa* Körn. in Mém. Inst. Égypt. 4: 305 (1901); *Oryza sativa* f. *chrysocarpa* Portères in J. Agric. Trop. Bot. Appl. 3: 55 (1956); *Oryza sativa* var. *cinnamomea* Gustchin in Riz & Rizic. 8: 33 (1934); *Oryza sativa* *communis* Körn. in F.A.Körnische & H.Werner, Handb. Getreidebaus 2: 938 (1885); *Oryza sativa* subsp. *communis* (Körn.) Gustchin in Riz & Rizic. 8: 28 (1934); *Oryza sativa* var. *costivorensis* Portères in J. Agric. Trop. Bot. Appl. 2: 575, 595 (1955); *Oryza sativa* var. *culinaris* Alef. in Landw. Fl.: 318 (1866); *Oryza sativa* var. *cyclina* Alef. in Landw. Fl.: 319 (1866); *Oryza sativa* var. *daloaensis* Portères in J. Agric. Trop. Bot. Appl. 2: 575, 595 (1955); *Oryza sativa* var. *denudata* Desv. in J. Bot. Agric. 1: 77 (1813); *Oryza sativa* var. *desvauxii* Körn. in F.A.Körnische & H.Werner, Handb. Getreidebaus 1: 233 (1885); *Oryza sativa* var. *dextrinosa* Portères in J. Agric. Trop. Bot. Appl. 2: 576, 595 (1955); *Oryza sativa* var. *dichroa* Gustchin in Riz & Rizic. 8: 34 (1934); *Oryza sativa* var. *dicolorata* Gustchin in Riz & Rizic. 8: 30 (1934); *Oryza sativa* var. *dubia* Körn. in F.A.Körnische & H.Werner, Handb. Getreidebaus 1: 234 (1885); *Oryza sativa* var. *dura* Crevost & Lemarié in Cat. Prod. Indochine 1: 13 (1947); *Oryza sativa* var. *eedeniana* Körn. in F.A.Körnische & H.Werner, Handb. Getreidebaus 1: 234 (1885); *Oryza sativa* var. *elegans* Jabrova in Trudy Azerbaidzhansk. Otd. Zakavkazsk. Fil. Akad. Nauk S.S.S.R. 2: 48, 50 (1935); *Oryza sativa* var. *elongata* Desv. in J. Bot. Agric. 1: 76 (1813); *Oryza sativa* var. *erythroceros* Körn. in F.A.Körnische & H.Werner, Handb.

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2: 582, 596 (1955); *Oryza sativa* var. *paragoensis* Portères in J. Agric. Trop. Bot. Appl. 2: 582, 596 (1955); *Oryza sativa* var. *paraguanensis* Portères in J. Agric. Trop. Bot. Appl. 2: 582, 596 (1955); *Oryza sativa* var. *parajavanica* Portères in J. Agric. Trop. Bot. Appl. 2: 583, 596 (1955); *Oryza sativa* var. *paramutica* Portères in J. Agric. Trop. Bot. Appl. 2: 583, 593 (1955); *Oryza sativa* var. *paraovoidea* Portères in J. Agric. Trop. Bot. Appl. 2: 583, 597 (1955); *Oryza sativa* var. *paraphilippensis* Portères in J. Agric. Trop. Bot. Appl. 2: 583, 597 (1955); *Oryza sativa* var. *parasepica* Portères in J. Agric. Trop. Bot. Appl. 2: 583, 597 (1955); *Oryza sativa* var. *parazeravschanica* Portères in J. Agric. Trop. Bot. Appl. 2: 584, 597 (1955); *Oryza sativa* var. *parjattica* Gustchin in Riz & Rizic. 8: 31 (1934); *Oryza sativa* var. *parnellii* Gustchin in Riz & Rizic. 8: 38 (1934); *Oryza sativa* var. *parva* Alef. in Landw. Fl.: 317 (1866); *Oryza sativa* var. *peradenica* Gustchin in Riz & Rizic. 8: 34 (1934); *Oryza sativa* var. *persica* Gustchin in Riz & Rizic. 8: 30 (1934); *Oryza sativa* var. *phaneroleuca* Portères in J. Agric. Trop. Bot. Appl. 2: 581, 596 (1955); *Oryza sativa* var. *philippensis* Gustchin in Riz & Rizic. 8: 30 (1934); *Oryza sativa* var. *picta* Portères in J. Agric. Trop. Bot. Appl. 2: 581, 596 (1955); *Oryza sativa* var. *pilosa* Blanco in Fl. Filip.: 275 (1837); *Oryza sativa* var. *plena* Prain in Bengal Pl. 2: 1184 (1903); *Oryza sativa* var. *plumbea* Gustchin in Riz & Rizic. 8: 35 (1934); *Oryza sativa* var. *praecox* Blanco in Fl. Filip.: 274 (1837); *Oryza sativa* var. *pseudocinnamomea* Gustchin in Riz & Rizic. 8: 34 (1934); *Oryza sativa* var. *pseudodaloensis* Portères in J. Agric. Trop. Bot. Appl. 2: 584, 597 (1955); *Oryza sativa* var. *pseudogoensis* Portères in J. Agric. Trop. Bot. Appl. 2: 584, 597 (1955); *Oryza sativa* var. *pseudoguineensis* Portères in J. Agric. Trop. Bot. Appl. 2: 584, 597 (1955); *Oryza sativa* var. *pseudojarhanica* Portères in J. Agric. Trop. Bot. Appl. 2: 584, 597 (1955); *Oryza sativa* var. *pseudolimnicola* Portères in J. Agric. Trop. Bot. Appl. 2: 585, 597 (1955); *Oryza sativa* var. *pseudomulayana* Portères in J. Agric. Trop. Bot. Appl. 2: 585, 597 (1955); *Oryza sativa* var. *pseudoovoidea* Portères in J. Agric. Trop. Bot. Appl. 2: 585, 597 (1955); *Oryza sativa* var. *pseudosepica* Portères in J. Agric. Trop. Bot. Appl. 2: 585, 597 (1955); *Oryza sativa* var. *pubescens* Desv. in J. Bot. Agric. 1: 76 (1813); *Oryza sativa* var. *pudica* Portères in J. Agric. Trop. Bot. Appl. 2: 581, 596 (1955); *Oryza sativa* var. *pygmaea* Makino in Pract. Hort. 25: 1259 (1940); *Oryza sativa* var. *pyrocarpa* Alef. in Landw. Fl.: 318 (1866); *Oryza sativa* var. *quinanda* Blanco in Fl. Filip.: 274 (1837); *Oryza sativa* var. *ramica* Gustchin in Riz & Rizic. 8: 34 (1934); *Oryza sativa* var. *ratoonica* Gustchin in Riz & Rizic. 8: 30 (1934); *Oryza sativa* var. *rhodosperma* Miq. in Fl. Ned. Ind. 3: 370 (1855); *Oryza sativa* var. *rubescens* Gustchin in Riz & Rizic. 8: 35 (1934); *Oryza sativa* var. *rubra* Blanco in Fl. Filip.: 275 (1837); *Oryza sativa* f. *rubra* (Blanco) Makino in Pract. Hort. 25: 1261 (1940); *Oryza sativa* var.

rubribarbis Desv. in J. Bot. Agric. 1: 76 (1813); *Oryza sativa* var. *rubriglumella* Gustchin in Riz & Rizic. 8: 29 (1934); *Oryza sativa* var. *rubrostellata* Jabrova in Trudy Azerbaidzhansk. Otd. Zakavkazsk. Fil. Akad. Nauk S.S.S.R. 2: 50 (1935); *Oryza sativa* var. *rufibarbis* Gustchin in Riz & Rizic. 8: 30 (1934); *Oryza sativa* var. *rufibrunnea* Gustchin in Riz & Rizic. 8: 37 (1934); *Oryza sativa* var. *rufiglumella* Gustchin in Riz & Rizic. 8: 30 (1934); *Oryza sativa* var. *saginata* Portères in J. Agric. Trop. Bot. Appl. 2: 585, 597 (1955); *Oryza sativa* var. *sarica* Gustchin in Riz & Rizic. 8: 30 (1934); *Oryza sativa* var. *sathi-nigrescens* Gustchin in Riz & Rizic. 8: 36 (1934); *Oryza sativa* var. *savannae* Körn. in F.A.Körnische & H.Werner, Handb. Getreidebaus 1: 233 (1885); *Oryza sativa* var. *saxenii* Gustchin in Riz & Rizic. 8: 38 (1934); *Oryza sativa* var. *seulica* Gustchin in Riz & Rizic. 8: 39 (1934); *Oryza sativa* var. *sickenbergeri* Körn. in Mém. Inst. Égypt. 4: 306 (1901); *Oryza sativa* var. *sitapurica* Gustchin in Riz & Rizic. 8: 31 (1934); *Oryza sativa* var. *sonesardica* Gustchin in Riz & Rizic. 8: 35 (1934); *Oryza sativa* var. *sordida* Gustchin in Riz & Rizic. 8: 34 (1934); *Oryza sativa* var. *sorgoidea* Desv. in J. Bot. Agric. 1: 77 (1813); *Oryza sativa* f. *spontanea* Roshev. in Trudy Prikl. Bot. Selektiv. 27: 37 (1931); *Oryza sativa* var. *striata* Gustchin in Riz & Rizic. 8: 35 (1934); *Oryza sativa* var. *subalba* Jabrova in Trudy Azerbaidzhansk. Otd. Zakavkazsk. Fil. Akad. Nauk S.S.S.R. 2: 49, 50 (1935), nom. illeg.; *Oryza sativa* var. *subalba* Gustchin in Riz & Rizic. 8: 37 (1934); *Oryza sativa* var. *subamaura* L.K.Kara-Mursa in Zap. Sem. Kontr. Stantsii Narkomzema Azerb. SSR 3: 28, 35 (1929); *Oryza sativa* var. *subastarinica* Jabrova in Trudy Azerbaidzhansk. Otd. Zakavkazsk. Fil. Akad. Nauk S.S.S.R. 2: 47, 50 (1935); *Oryza sativa* var. *subazerbaidjanica* Gustchin in Riz & Rizic. 8: 36 (1934); *Oryza sativa* var. *subbrunnea* Gustchin in Riz & Rizic. 8: 36 (1934); *Oryza sativa* var. *subdichroa* L.K.Kara-Mursa in Zap. Sem. Kontr. Stantsii Narkomzema Azerb. SSR 3: 26, 35 (1929); *Oryza sativa* var. *suberythroceros* Gustchin in Riz & Rizic. 8: 36 (1934); *Oryza sativa* var. *subflavoacies* Volozh. in Trudy Opytn. Uchrezhd. Daln. Vostoka 1: 141, 148 (1930); *Oryza sativa* var. *subjanthoceros* Gustchin in Riz & Rizic. 8: 36 (1934); *Oryza sativa* var. *subjarhanica* Vasc. in Anais Inst. Super. Agron. 19: 2 (1952); *Oryza sativa* var. *subleucoceros* Volozh. in Trudy Opytn. Uchrezhd. Daln. Vostoka 1: 140, 148 (1930); *Oryza sativa* var. *submasenderano* Jabrova in Trudy Azerbaidzhansk. Otd. Zakavkazsk. Fil. Akad. Nauk S.S.S.R. 2: 47, 50 (1935); *Oryza sativa* var. *submelanotrix* Volozh. in Trudy Opytn. Uchrezhd. Daln. Vostoka 1: 143, 148 (1930); *Oryza sativa* var. *submovlanica* Vasc. in Anais Inst. Super. Agron. 19: 2 (1952); *Oryza sativa* var. *submutica* Miq. in Fl. Ned. Ind. 3: 369 (1855); *Oryza sativa* var. *subpyrocarpa* Gustchin in Riz & Rizic. 8: 36 (1934); *Oryza sativa* var. *subratoonica* Vasc. in Anais Inst. Super. Agron. 19: 1 (1952); *Oryza sativa* var. *subsubulata* Miq. in Fl. Ned. Ind. 3: 369 (1855);

<i>Patrinia scabra</i> Bunge	<i>Patrinia scabra</i> Bunge, Uchen. Zap. Imp. Kazansk. Univ. 1835(4): 171 (1835)	<i>Caprifoliaceae</i> Juss.	<i>Oryza sativa</i> var. <i>subvavilovii</i> Jabrova in Trudy Azerbaidzhansk. Otd. Zakavkazsk. Fil. Akad. Nauk S.S.S.R. 2: 48, 50 (1935); <i>Oryza sativa</i> var. <i>subvulgaris</i> Gustchin in Riz & Rizic. 8: 36 (1934); <i>Oryza sativa</i> var. <i>subzomica</i> Gustchin in Riz & Rizic. 8: 37 (1934); <i>Oryza sativa</i> var. <i>sultanpurica</i> Gustchin in Riz & Rizic. 8: 33 (1934); <i>Oryza sativa</i> var. <i>taurina</i> Alef. in Landw. Fl.: 318 (1866); <i>Oryza sativa</i> var. <i>tawnica</i> Gustchin in Riz & Rizic. 8: 29 (1934); <i>Oryza sativa</i> var. <i>tcharinica</i> Gustchin in Riz & Rizic. 8: 34 (1934); <i>Oryza sativa</i> var. <i>terrestris</i> Makino in J. Jap. Bot. 8: 31 (1932); <i>Oryza sativa</i> var. <i>trichoviolacea</i> Gustchin in Riz & Rizic. 8: 37 (1934); <i>Oryza sativa</i> var. <i>usitatissima</i> Körn. in F.A.Körnicke & H.Werner, Handb. Getreidebaus 2: 938 (1885); <i>Oryza sativa</i> var. <i>ussurica</i> Volozh. in Trudy Opytn. Uchrezhd. Daln. Vostoka 1: 146, 148 (1930); <i>Oryza sativa</i> var. <i>utilissima</i> A.Camus in Bull. Lab. Agron. Coloniale 1: 9 (1913); <i>Oryza sativa</i> var. <i>vavilovii</i> Gustchin in Riz & Rizic. 8: 35 (1934); <i>Oryza sativa</i> proles <i>verindojaponicoides</i> Portères in Rev. Int. Bot. Appl. Agric. Trop. 30: 153 (1950); <i>Oryza sativa</i> proles <i>verohumilojaponicoides</i> Portères in Rev. Int. Bot. Appl. Agric. Trop. 30: 153 (1950); <i>Oryza sativa</i> proles <i>verojaponicoides</i> Portères in Rev. Int. Bot. Appl. Agric. Trop. 30: 153 (1950); <i>Oryza sativa</i> var. <i>violacea</i> Blanco in Fl. Filip.: 275 (1837); <i>Oryza sativa</i> var. <i>violacea</i> Gustchin in Riz & Rizic. 8: 35 (1934), nom. illeg.; <i>Oryza sativa</i> var. <i>violapunctata</i> L.K.Kara-Mursa in Zap. Sem. Kontr. Stantsii Narkomzema Azerb. SSR 3: 29, 35 (1929); <i>Oryza sativa</i> var. <i>violeobrevis</i> Vasc. in Anais Inst. Super. Agron. 19: 3 (1952); <i>Oryza sativa</i> var. <i>viridescens</i> Körn. in F.A.Körnicke & H.Werner, Handb. Getreidebaus 1: 232 (1885); <i>Oryza sativa</i> var. <i>vulgaris</i> Postiglione in Relaz. Intorno Condizioni Agric.: 265 (1876); <i>Oryza sativa</i> var. <i>wightiana</i> Körn. in F.A.Körnicke & H.Werner, Handb. Getreidebaus 1: 232 (1885); <i>Oryza sativa</i> var. <i>xanthoxeros</i> Körn. in F.A.Körnicke & H.Werner, Handb. Getreidebaus 2: 943 (1885); <i>Oryza sativa</i> var. <i>zeravschanica</i> Gustchin in Riz & Rizic. 8: 33 (1934); <i>Oryza sativa</i> var. <i>zhukovskii</i> Gustchin in Riz & Rizic. 8: 32 (1934); <i>Oryza sativa</i> var. <i>zomica</i> Körn. in F.A.Körnicke & H.Werner, Handb. Getreidebaus 1: 234 (1885); <i>Oryza segetalis</i> Russell ex Steud. in Syn. Pl. Glumac. 1: 3 (1853), nom. nud.; <i>Oryza sorghoidea</i> (Desv.) Steud. in Nomencl. Bot. 1: 577 (1821), nom. provis. Homotypic Synonyms: <i>Fedia scabra</i> (Bunge) Kuntze in Revis. Gen. Pl. 1: 302 (1891); <i>Patrinia rupestris</i> subsp. <i>scabra</i> (Bunge) H.J.Wang in Acta Phytotax. Sin. 23: 382 (1985)	Perennial	N. China (to W. Henan)	Temperate biome. Sunny grassy slopes, forest margins; Alt. 300-1700 m	1, 23
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<i>Periploca somaliensis</i> Browicz	<i>Periploca somaliensis</i> Browicz, Arbor. Kórnické 9: 53 (1966)	<i>Apocynaceae</i> Juss.,	Heterotypic Synonyms: <i>Periploca brevicoronata</i> Goyder & Boulos in Kew Bull. 46: 133 (1991)	Scrambling shrub	N. Somalia, SW. Arabian Peninsula	Subtropical biome. Semi-arid mountain scrub. Alt. 2000-2500 m	1, 2, 32-34
<i>Phellodendron chinense</i> var. <i>glabriusculum</i> C.K.Schneid.	<i>Phellodendron chinense</i> var. <i>glabriusculum</i> C.K.Schneid., Ill. Handb. Laubholz. 2: 126 (1907)	<i>Rutaceae</i> Juss.	Heterotypic Synonyms: <i>Phellodendron amurense</i> var. <i>wilsonii</i> (Hayata & Kaneh.) C.E.Chang in Quart. J. Chin. Forest. 7(4): 58 (1974); <i>Phellodendron chinense</i> var. <i>falcatum</i> C.C.Huang in Acta Phytotax. Sin. 7: 336 (1958); <i>Phellodendron chinense</i> var. <i>omeiense</i> C.C.Huang in Acta Phytotax. Sin. 7: 335 (1958); <i>Phellodendron chinense</i> var. <i>yunnanense</i> C.C.Huang in Acta Phytotax. Sin. 7: 336 (1958); <i>Phellodendron sinense</i> Dode in Bull. Soc. Bot. France 55: 649 (1909); <i>Phellodendron wilsonii</i> Hayata & Kaneh. in Icon. Pl. Formosan. 9: 8 (1920)	Tree	Central & S. China	Temperate biome. Mixed broad-leaved forests, open to dense forests. Alt. 800-1500 (-3000) m	1, 2, 35
<i>Picrasma quassioides</i> (D.Don) Benn.	<i>Picrasma quassioides</i> (D.Don) Benn., Pl. Jav. Rar.: 198 (1844)	<i>Simaroubaceae</i> DC.	Homotypic Synonyms: <i>Simaba quassioides</i> D.Don, Prodr. Fl. Nepal.: 248 (1825). Heterotypic Synonyms <i>Rhus ailanthoides</i> Bunge, Mém. Acad. Imp. Sci. St.-Pétersbourg Divers Savans 2: 89 (1835); <i>Nima quassioides</i> Buch.-Ham. ex A.Juss., Mém. Mus. Hist. Nat. 12: 517 (1825); <i>Picrasma ailanthoides</i> (Bunge) Planch., London J. Bot. 5: 573 (1846); <i>Picrasma japonica</i> A.Gray, Mem. Amer. Acad. Arts, n.s., 6: 383 (1859); <i>Picrasma quassioides</i> f. <i>glabrescens</i> (Pamp.) Kitag., Neolin. Fl. Manshur.: 424 (1979); <i>Zwingera quassioides</i> Spreng., Syst. Veg. 4(2): 163 (1827)	Tree	Himalaya to Temp. E. Asia	Temperate biome. Mountainous mixed forests. Alt. 1400-3200 m	1, 2, 35
<i>Poa pratensis</i> L.	<i>Poa pratensis</i> L., Sp. Pl.: 67 (1753)	<i>Poaceae</i> Barnhart	Homotypic Synonyms: <i>Paneion pratense</i> (L.) Lunell in Amer. Midl. Naturalist 4: 222 (1915); <i>Poa angustifolia</i> subsp. <i>pratensis</i> (L.) K.Richt. in Pl. Eur. 1: 87 (1890); <i>Poa angustifolia</i> var. <i>pratensis</i> (L.) Simonk. in Enum. Pl. Transsilv.: 580 (1896); <i>Poa pratensis</i> var. <i>major</i> Roth in Enum. Pl. Phaen. Germ. 1(1): 334 (1827), not validly publ; <i>Poa pratensis</i> subsp. <i>vulgaris</i> Gaudin in Fl. Helv. 1: 258 (1828), not validly publ.	Perennial or rhizomatous geophyte	Subarctic to Temp. Northern Hemisphere and N. Mexico	Temperate biome. Moderately moist to wet conditions, from coastal meadows to forest shade, to alpine and tundra, often in disturbed sites; Alt. 500–4400 m	1, 23
<i>Raoulia australis</i> Hook.f. ex Raoul	<i>Raoulia australis</i> Hook.f. ex Raoul, Choix pl. Nouv.-Zél. 20, t. 15 (1846)	<i>Asteraceae</i> Bercht. & J.Presl	Heterotypic Synonym: <i>Raoulia lutescens</i> Beauverd in Bull. Soc. Bot. Genève, sér. 2, 2: 221 (1910)	Subshrub	New Zealand	Subalpine or subarctic biome. Dry grassland and stable riverbed, forming hard, dense, blue-grey, or yellow-green mats.	1, 2, 36-39

<i>Rhodomela confervoides</i> (Hudson) P.C.Silva 1952	Rhodomela confervoides (Hudson) P.C.Silva 1952, Silva, P.C. (1952). University of California Publications in Botany 25: 241-323	<i>Rhodomelaceae</i> Horaninow	Basionym: <i>Fucus confervoides</i> Hudson. Homotypic synonyms: <i>Fucus confervoides</i> Hudson 1762; <i>Sphaerococcus confervoides</i> (Hudson) C.Agardh 1817. Heterotypic synonyms: <i>Rhodomela subfusca</i> var. <i>firmior</i> J.Agardh null; <i>Fucus subfuscus</i> Woodward 1791; <i>Fucus variabilis</i> Goodenough & Woodward 1797; <i>Fuscaria variabilis</i> Stackhouse 1809; <i>Gigartina subfusca</i> (Woodward) J.V.Lamouroux 1813; <i>Sphaerococcus subfuscus</i> (Woodward) C.Agardh 1817; <i>Sphaerococcus subfuscus</i> var. <i>tenuior</i> C.Agardh 1817; <i>Gigartina subfusca</i> var. <i>tenuior</i> (C.Agardh) Lyngbye 1819; <i>Gigartina subfusca</i> var. <i>flaccida</i> Lyngbye 1819; <i>Gigartina subfusca</i> var. <i>racemosa</i> Lyngbye 1819; <i>Rhodomela subfusca</i> (Woodward) C.Agardh 1822; <i>Rhodomela subfusca</i> var. <i>tenuior</i> (C.Agardh) C.Agardh 1822; <i>Rhodomela subfusca</i> var. <i>flaccida</i> (Lyngbye) C.Agardh 1822; <i>Rhodomela subfusca</i> var. <i>penicillata</i> C.Agardh 1822; <i>Lophura gracilis</i> Kützting 1843; <i>Rhodomela rochei</i> Harvey 1853; <i>Rhodomela subfusca</i> f. <i>gracilior</i> J.Agardh 1863; <i>Rhodomela subfusca</i> f. <i>extrataeniensis</i> Areschoug 1875; <i>Rhodomela subfusca</i> f. <i>gracilis</i> (Kützting) Areschoug 1875; <i>Rhodomela subfusca</i> f. <i>intrataeniensis</i> Areschoug 1875; <i>Rhodomela subfusca</i> f. <i>tenuior</i> (C.Agardh) Svedelius 1901; <i>Rhodomela subfusca</i> f. <i>abyssicola</i> Rosenvinge 1924; <i>Rhodomela confervoides</i> f. <i>gracilior</i> (J.Agardh) W.R.Taylor 1957; <i>Rhodomela confervoides</i> f. <i>rochei</i> (Harvey) W.R.Taylor 1957; <i>Rhodomela confervoides</i> f. <i>abyssicola</i> (Rosenvinge) Pankow 1971; <i>Rhodomela confervoides</i> f. <i>gracilis</i> (Kützting) Pankow 1971; <i>Rhodomela confervoides</i> f. <i>tenuior</i> (C.Agardh) Pankow 1971	Marine species	Widely distributed	Alt. 0-1680 m On rocks and shells, intertidal pools at all levels	40
<i>Salvia aethiopis</i> L.	<i>Salvia aethiopis</i> L., Sp. Pl.: 27 (1753)	<i>Lamiaceae</i> Martinov	Homotypic Names: <i>Sclarea aethiopis</i> (L.) Mill., Gard. Dict. ed. 8: n.º 2 (1768); <i>Sclarea lanata</i> Moench, Methodus: 374 (1794), nom. superfl.; <i>Salvia lanata</i> Stokes, Bot. Mat. Med.: 52 (1812), nom. illeg. Heterotypic Synonyms: <i>Salvia kochiana</i> Kunze, Index Seminum (LZ, Lipsiensis) 1847: 4 (1847); <i>Salvia leuconeura</i> Boiss., Diagn. Pl. Orient., ser. 2, 4: 20 (1859); <i>Aethiopis vera</i> Fourr., Ann. Soc. Linn. Lyon, n.s., 17: 134 (1869); <i>Salvia idanensis</i> Gand., Fl. Lyon.: 171 (1875).	Biennial or perennial	Europe to Central Asia	Temperate biome. Steppe, igneous and limestone slopes, fallow fields, roadside banks. Alt. 0-2100 m	1-3, 41-43
<i>Salvia dominica</i> L.	<i>Salvia dominica</i> L., Sp. Pl.: 25 (1753)	<i>Lamiaceae</i> Martinov	Heterotypic Synonyms: <i>Salvia graveolens</i> Vahl, Enum. Pl. Obs. 1: 273 (1804); <i>Salvia commutata</i> Benth., Labiat. Gen. Spec.: 222 (1833); <i>Salvia syriaca</i> Gouan ex Benth. in A.P.de Candolle, Prodr. 12: 279 (1848).	Perennial	E. Medit. to Arabian Peninsula	Subtropical biome. Batha vegetation in chalk rocks and white-grayish soil.	1, 2, 44, 45

<i>Salvia hypoleuca</i> Benth.	<i>Salvia hypoleuca</i> Benth., A.P.de Candolle, Prodr. 12: 279 (1848)	<i>Lamiaceae</i> Martinov	-	Perennial	N. and Central Iran	Alt. 25-250 m Temperate biome. In the arid sub-alpine, in rocky detritus.	1, 2, 46, 47
<i>Salvia lachnocalyx</i> Hedge	<i>Salvia lachnocalyx</i> Hedge, Fl. Iranica 150: 455 (1982)	<i>Lamiaceae</i> Martinov	-	Perennial	S. Iran	Alt. 25-3000 m Subtropical biome. Endemic.	1-3, 41
<i>Salvia limbata</i> C.A.Mey.	<i>Salvia limbata</i> C.A.Mey., Verz. Pfl. Casp. Meer.: 86 (1831)	<i>Lamiaceae</i> Martinov	Heterotypic Synonyms: <i>Salvia flexuosa</i> Schrank, Syll. Pl. Nov. 2: 57 (1826), provisionally listed as a synonym; <i>Salvia polyadenia</i> Boiss. & Heldr. in P.E.Boissier, Diagn. Pl. Orient. 5: 7 (1844); <i>Salvia chrysadenia</i> Freyn, Bull. Herb. Boissier, sér. 2, 1: 279 (1901).	Perennial	E. Turkey to Iran	Alt. 2000-3000 m Temperate biome. Stony slopes, steppe, sandy loam, roadsides, cornfields.	1-3, 41
<i>Salvia mirzayanii</i> Rech.f. & Esfand.	<i>Salvia mirzayanii</i> Rech.f. & Esfand., Oesterr. Bot. Z. 99: 53 (1952)	<i>Lamiaceae</i> Martinov	-	Subshrub	NE. Arabian Peninsula, W. & S. Iran	Alt. 1060-2340 m Subtropical biome. In mountains.	1-3, 47-50
<i>Salvia palaestina</i> Benth.	<i>Salvia palaestina</i> Benth., Labiat. Gen. Spec.: 718 (1835)	<i>Lamiaceae</i> Martinov	Heterotypic Synonyms: <i>Salvia sinaica</i> Delile ex Benth., Labiat. Gen. Spec.: 718 (1835); <i>Salvia lorentii</i> Hochst. in J.A.Lorent, Wanderungen: 333 (1845); <i>Salvia sieberi</i> C.Presl, Abh. Königl. Böhm. Ges. Wiss., ser. 5, 3: 530 (1845); <i>Salvia rassamii</i> Boiss., Fl. Orient. 4: 615 (1879); <i>Salvia alliaria</i> Parsa, Kew Bull. 3: 224 (1948).	Perennial	E. Medit. to Iran	Alt. 1100-1700 m Subtropical biome. Limestone and igneous rocky slopes, cliffs, in Quercus scrub, vineyards, fallow fields.	1, 2, 41, 42
<i>Salvia sahendica</i> Boiss. & Buhse	<i>Salvia sahendica</i> Boiss. & Buhse, Nouv. Mém. Soc. Imp. Naturalistes	<i>Lamiaceae</i> Martinov	-	Perennial	NW. Iran	Alt. 300-1500 m Temperate biome. Endemic.	1, 2, 46, 51

<i>Salvia syriaca</i> L.	Moscou 12: 172 (1860) <i>Salvia syriaca</i> L., Syst. Nat. ed. 10, 2: 854 (1759)	Lamiaceae Martinov	Homotypic Names: <i>Sclarea syriaca</i> (L.) Mill., Gard. Dict. ed. 8: n.º 5 (1768). Heterotypic Synonyms: <i>Salvia parviflora</i> Vahl, Enum. Pl. Obs. 1: 268 (1804), nom. illeg.; <i>Salvia varia</i> Vahl, Enum. Pl. Obs. 1: 273 (1804).	Perennial	E. Medit. to Iran	Temperate biome. Steppe, marly banks, fallow and cultivated fields. Alt. 350-2180 m	1, 2, 42, 46, 51, 52
<i>Salvia tingitana</i> Etl.	<i>Salvia tingitana</i> Etl., Salv.: 35 (1777)	Lamiaceae Martinov	Homotypic Names: <i>Sclarea tingitana</i> (Etl.) Raf., Fl. Tellur. 3: 94 (1837).	Perennial or subshrub	W. Saudi Arabia	Subtropical biome. Rocky slopes, juniper zone. Alt. 1676-1800 m	1, 2, 53-57
<i>Salvia yosgadensis</i> Freyn & Bornm.,	<i>Salvia yosgadensis</i> Freyn & Bornm., Oesterr. Bot. Z. 42: 348 (1892)	Lamiaceae Martinov	-	Perennial	W. Central Turkey	Temperate biome. <i>Quercus</i> scrub, disturbed steppe, fields. Alt. 800-1635 m	1, 2, 42, 51, 52
<i>Scutellaria coleifolia</i> H.Lév.	<i>Scutellaria violacea</i> var. <i>violacea</i>	Lamiaceae Martinov	Heterotypic Synonyms: <i>Scutellaria coleifolia</i> H.Lév. in Repert. Spec. Nov. Regni Veg. 13: 343 (1914); <i>Scutellaria floribunda</i> Benth. in A.P.de Candolle, Prodr. 12: 418 (1848); <i>Scutellaria indica</i> Roxb. in Fl. Ind., ed. 1832. 3: 24 (1832), nom. illeg.; <i>Scutellaria nodulosa</i> A.Ham. in Esq. Monogr. Scutellaria: 26 (1832); <i>Scutellaria violacea</i> var. <i>floribunda</i> (Benth.) Hook.f. in Fl. Brit. India 4: 668 (1885); <i>Scutellaria violacea</i> var. <i>glabra</i> Trimen in Syst. Cat. Fl. Pl. Ceylon: 70 (1885); <i>Scutellaria violacea</i> var. <i>glabrior</i> Benth. in N.Wallich, Pl. Asiat. Rar. 1: 67 (1830); <i>Scutellaria violacea</i> var. <i>sikkimensis</i> Hook.f. in Fl. Brit. India 4: 668 (1885)	perennial	S. India, Himalaya to S. Central China and Thailand	Wet tropical biome. Pine forests, grassy slopes. Alt. 1900-3200 m	1, 2, 35
<i>Serjania goniocarpa</i> Radlk.	<i>Serjania goniocarpa</i> Radlk., Monogr. Serjania: 309 (1875)Radlk.	Sapindaceae Juss.	Heterotypic Synonyms: <i>Serjania racemosa</i> Seem. in Bonplandia (Hannover) 5: 74 (1857), nom. illeg.	Liana	Mexico to Honduras	Seasonally dry tropical biome. <i>Pinus-Quercus</i> and <i>Juniperus</i> forest, ecotonia with tropical deciduous forest with crassicaules, in black rocky soil, with limestone outcrops. Alt. 2030 m	1, 2, 58

<i>Solanum tuberosum</i> L.	<i>Solanum tuberosum</i> L., Sp. Pl.: 185 (1753)	<i>Solanaceae</i> Juss.	Homotypic Synonyms: <i>Lycopersicon tuberosum</i> (L.) Mill., Gard. Dict. ed. 8: n.º 7 (1768); <i>Solanum tuberosum</i> var. <i>cultum</i>, Arch. Sci. Phys. Nat., III, 15: 437 (1886); <i>Solanum tuberosum</i> var. <i>vulgare</i> Hook.f., Bot. Antarct. Voy. 1(2): 329 (1768) Heterotypic Synonyms: <i>Battata tuberosa</i> (L.) Hill, Hort. Kew. 148; <i>Larnax sylvarum</i> subsp. <i>novogranatensis</i> N.W.Sawyer, 11(4): 461 (-462) (2001); <i>Parmentiera edulis</i> Raf., Autik. Bot.: 108 (1840); <i>Solanum andigenum</i> Juz. & Bukasov, Trudy Vsesoyusn. S'ezda Gen. Selektiv. Semenov. Plemen. Zhivotnov. 3: 609 (1929); <i>Solanum andigenum</i> convar. <i>acutifolium</i> Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 47 (1983); <i>Solanum andigenum</i> convar. <i>adpressipilosum</i> Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 48 (1983); <i>Solanum andigenum</i> f. <i>alccai-huarmi</i> Bukasov & Lechn., 58: 54 (1933); <i>Solanum andigenum</i> f. <i>ancacc-maquín</i> Bukasov & Lechn., 58: 63 (1933); <i>Solanum andigenum</i> f. <i>arcuatum</i> Bukasov & Lechn., 58: 73 (1933); <i>Solanum andigenum</i> subsp. <i>argentinicum</i> Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 48 (1983); <i>Solanum andigenum</i> subsp. <i>australiperuvianum</i> Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 46 (1983); <i>Solanum andigenum</i> subsp. <i>aya-papa</i> Bukasov & Lechn., 58: 45, 164 (1933); <i>Solanum andigenum</i> var. <i>aymaranum</i> Bukasov & Lechn., 58: 68 (1933); <i>Solanum andigenum</i> f. <i>baiscopum</i> Bukasov & Lechn., 58: 69 (1933); <i>Solanum andigenum</i> f. <i>bifidum</i> Bukasov & Lechn., 58: 71 (1933); <i>Solanum andigenum</i> var. <i>bolivianum</i> Bukasov & Lechn., 58: 70 (1933); <i>Solanum andigenum</i> subsp. <i>bolivianum</i> Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 48 (1983); <i>Solanum andigenum</i> convar. <i>brachistylum</i> Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 48 (1983); <i>Solanum andigenum</i> convar. <i>brevicalyces</i> Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 48 (1983); <i>Solanum andigenum</i> var. <i>brevicalyx</i> Bukasov & Lechn., 58: 75 (1933); <i>Solanum andigenum</i> convar. <i>brevipilosum</i> Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 46 (1983); <i>Solanum andigenum</i> f. <i>caesium</i> Bukasov & Lechn., 58: 74 (1933); <i>Solanum andigenum</i> f. <i>caiceda</i> Bukasov, 47: 208 (1930); <i>Solanum andigenum</i> var. <i>carhua</i> Vargas, 2: 49 (1954); <i>Solanum andigenum</i> f. <i>ccompetillo</i> Bukasov & Lechn., 58: 56 (1933); <i>Solanum andigenum</i> f. <i>ccompis</i> Bukasov & Lechn., 58: 67 (1933); <i>Solanum andigenum</i> var. <i>ccusi</i> Bukasov & Lechn., 58: 62 (1933); <i>Solanum andigenum</i> subsp. <i>centraliperuvianum</i> Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 46 (1983); <i>Solanum andigenum</i> f. <i>cevallosii</i> Bukasov & Lechn., 58: 74 (1933); <i>Solanum andigenum</i> f. <i>chalconense</i> Bukasov, 47: 204, 517. (1930); <i>Solanum andigenum</i> f. <i>chimaco</i> Bukasov & Lechn., 58: 58 (1933); <i>Solanum andigenum</i> var. <i>ckello-huaccoto</i> Bukasov & Lechn., 58: [53] (1933); <i>Solanum andigenum</i> f. <i>coeruleum</i> Lechn. ex Bukasov, 58: 48 (1933); <i>Solanum andigenum</i> var. <i>colombianum</i> Bukasov, Trudy Prikl. Bot. 47: 205 (1930); <i>Solanum andigenum</i> subsp. <i>colombianum</i>	Tuberous geophyte	W. and S. South America to NW. Venezuela	Subtropical biome. Montane (highlands). Widely cultivated. Alt. 1650-3600 m	1-3, 26
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(Bukasov) Lechn., Trudy Prikl. Bot. 47: 205 (1930); *Solanum andigenum* f. *conicocolumnatum* Bukasov & Lechn., 58: 68 (1933); *Solanum andigenum* f. *cryptostylum* Bukasov & Lechn., 58: 75 (1933); *Solanum andigenum* convar. *curtibaccatum* Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 47 (1983); *Solanum andigenum* var. *cuzcoense* Bukasov & Lechn., 58: 59 (1933); *Solanum andigenum* var. *digitotuberosum* Vargas, 2: 49 (1954); *Solanum andigenum* f. *dilatatum* Bukasov & Lechn., 58: 72 (1933); *Solanum andigenum* f. *discolor* Bukasov & Lechn., 58: 69 (1933); *Solanum andigenum* subsp. *ecuatorianum* Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 46 (1983); *Solanum andigenum* convar. *elongatibaccatum* Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 48 (1983); *Solanum andigenum* f. *elongatipedicellatum* Lechn., 1: 17 (1940); *Solanum andigenum* f. *globosum* Bukasov & Lechn., 58: 69 (1933); *Solanum andigenum* var. *grauense* Vargas, 2: 48 (1954); *Solanum andigenum* f. *guatemalense* Bukasov, 47: 205, 518 (1930); *Solanum andigenum* var. *hederiforme* Bukasov; *Solanum andigenum* var. *herreriae* Bukasov & Lechn., 58: 60 (1933); *Solanum andigenum* f. *huaca-layra* Bukasov & Lechn., 58: 68 (1933); *Solanum andigenum* var. *huairuru* Bukasov & Lechn., 58: 57 (1933); *Solanum andigenum* f. *huallata* Bukasov & Lechn., 58: 61 (1933); *Solanum andigenum* f. *huaman-uma* Bukasov & Lechn., 58: 54 (1933); *Solanum andigenum* var. *imilla* Bukasov & Lechn., 58: 66 (1933); *Solanum andigenum* f. *incrassatum* Bukasov & Lechn., 58: 71 (1933); *Solanum andigenum* var. *juninum* Bukasov, 58: 46 (1933); *Solanum andigenum* f. *lanciacuminatum* Bukasov & Lechn., 58: 69 (1933); *Solanum andigenum* f. *lapazense* Bukasov & Lechn., 58: 70 (1933); *Solanum andigenum* var. *latius* Bukasov & Lechn., 58: 51 (1933); *Solanum andigenum* f. *leckeumo* Bukasov & Lechn., 58: 61 (1933); *Solanum andigenum* f. *lilacinoflorum* Bukasov, 58: 38 (1933); *Solanum andigenum* f. *lisarassa* Bukasov, 47: 209, 520 (1930); *Solanum andigenum* f. *llutuc-runtum* Lechn. ex Bukasov, 58: 60 (1933); *Solanum andigenum* convar. *longiacuminatum* Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 47 (1983); *Solanum andigenum* var. *longibaccatum* Bukasov & Lechn., 58: 73 (1933); *Solanum andigenum* convar. *macron* Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 48 (1983); *Solanum andigenum* f. *magnicorollatum* Bukasov & Lechn., 58: 71 (1933); *Solanum andigenum* var. *mexicanum* Bukasov, 47: 202, 516 (1930); *Solanum andigenum* f. *microstigma* Bukasov & Lechn., 58: 63 (1933); *Solanum andigenum* convar. *microstigmatum* Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 47 (1983); *Solanum andigenum* f. *nodosum* Bukasov, 58: 49 (1933); *Solanum andigenum* convar. *nudiculum* Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 47 (1983); *Solanum andigenum* convar. *obtusiacuminatum* Lechn., Trudy Prikl. Bot., Ser. 2, Genet. Rast. 79: 47

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			<i>strobilinum</i> Alef., Landw. Fl.: 141 (1866); <i>Solanum tuberosum</i> f. <i>surico</i> Ochoa, Phytologia 65: 110 (1988); <i>Solanum tuberosum</i> var. <i>taraco</i> Ochoa, Phytologia 65: 112 (1988); <i>Solanum tuberosum</i> var. <i>tener</i> Alef., Landw. Fl.: 139 (1866); <i>Solanum tuberosum</i> f. <i>tenuipedicellatum</i> Bukasov & Lechn., 58: 84 (1933); <i>Solanum tuberosum</i> f. <i>thalassinum</i> Alef., Landw. Fl.: 146 (1866); <i>Solanum tuberosum</i> f. <i>tinguipaya</i> Ochoa, Phytologia 65: 110 (1988); <i>Solanum tuberosum</i> var. <i>ulmense</i> Alef., Landw. Fl.: 147 (1866); <i>Solanum tuberosum</i> var. <i>versicolor</i> Alef., Landw. Fl.: 148 (1866); <i>Solanum tuberosum</i> var. <i>villaroella</i> Bukasov & Lechn., 58: 86 (1933); <i>Solanum tuberosum</i> f. <i>viride</i> Bukasov & Lechn., 58: 81 (1933); <i>Solanum tuberosum</i> var. <i>vuchefeldicum</i> Alef., Landw. Fl.: 146 (1866); <i>Solanum tuberosum</i> var. <i>vulgare</i> Macloskie, 8: 707 (1905); <i>Solanum tuberosum</i> f. <i>wila-huaycku</i> Ochoa, Phytologia 65: 110 (1988); <i>Solanum tuberosum</i> f. <i>wila-imilla</i> Ochoa, Phytologia 65: 109 (1988); <i>Solanum tuberosum</i> f. <i>wila-k'oyu</i> Ochoa, Phytologia 65: 112 (1988); <i>Solanum tuberosum</i> f. <i>wila-monda</i> Ochoa, Phytologia 65: 109 (1988); <i>Solanum tuberosum</i> f. <i>wila-pala</i> Ochoa, Phytologia 65: 109 (1988); <i>Solanum tuberosum</i> var. <i>xanthoceras</i> Alef., Landw. Fl.: 140 (1866); <i>Solanum tuberosum</i> f. <i>yurac-taraco</i> Ochoa, Phytologia 65: 113 (1988); <i>Solanum tuberosum</i> var. <i>yutuense</i> Bukasov & Lechn., 58: 79, 89, 180 (1933); <i>Solanum utile</i> Klotzsch, Allg. Gartenzeitung 17: 315 (1849); <i>Solanum yabari</i> Hawkes, Potato Collect. Exped. Mexico & S. Amer. 2 Syst. Classific. Collect. 65: 129 (1944); <i>Solanum yabari</i> var. <i>cuzcoense</i> Hawkes, Potato Collect. Exped. Mexico & S. Amer. 2 Syst. Classific. Collect. 65: 129 (1944); <i>Solanum yabari</i> var. <i>pepino</i> Hawkes, 2: 65 (1944); <i>Solanum zykinii</i> Lechn., Trudy Prikl. Bot. 62(1): 44 (1978) Homotypic Synonyms: <i>Kaempferia involucrata</i> King ex Baker in J.D.Hooker, Fl. Brit. India 6: 221 (1890); <i>Stahlianthus involucratus</i> (King ex Baker) Craib ex Loes. in H.G.A.Engler, Nat. Pflanzenfam., ed. 2. 15a: 564 (1930). Heterotypic Synonyms: <i>Kaempferia hainanensis</i> Hayata in Icon. Pl. Formosan. 5: 213 (1915)	Rhizomatous geophyte	Darjeeling to S. China	Subtropical biome. Deciduous dipterocarp forest. Alt. 500 m	1, 2, 59-61
<i>Stahlianthus involucratus</i> (King ex Baker) Craib ex Loes.	<i>Curcuma involucrata</i> (King ex Baker) Škorničk., Taxon 64: 369 (2015)	Zingiberaceae Martinov					
<i>Swertia bimaculata</i> (Siebold & Zucc.) Hook.f. & Thomson ex C.B.Clarke	<i>Swertia bimaculata</i> (Siebold & Zucc.) Hook.f. & Thomson ex C.B.Clarke, Sp. Pl.: 226 (1753)	Gentianaceae Juss.	Homotypic Synonyms: <i>Frasera bimaculata</i> (Siebold & Zucc.) Toyok. in Symb. Asahikaw. 1: 156 (1965); <i>Ophelia bimaculata</i> Siebold & Zucc. in Abh. Math.-Phys. Cl. Königl. Bayer. Akad. Wiss. 4(3): 159 (1846) Heterotypic Synonyms: <i>Silene esquirolii</i> H.Lév. in Fl. Kouy-Tchéou: 68 (1914); <i>Swertia biauriculata</i> H.Lév. in Cat. Pl. Yun-Nan: 116 (1916), nom. nud.; <i>Swertia bimaculata</i> var. <i>impunctata</i> Makino in Bot. Mag. (Tokyo) 24: 16 (1910); <i>Swertia bimaculata</i> var. <i>macrocarpa</i> Nakai in Bot. Mag. (Tokyo) 47: 262 (1933); <i>Swertia mairei</i> H.Lév. in Bull. Acad. Int. Géogr.	Annual or biennial	Nepal to Japan and N. Indochina	Temperate biome. On sandy loam in evergreen oak forest. Alt. 1520-3660 m	1, 2, 62

<i>Triticum aestivum</i> L.	<i>Triticum aestivum</i> L., Sp. Pl.: 85 (1753)	<i>Poaceae</i> Barnhart	Bot. 25: 22 (1915); <i>Swertia plantaginifolia</i> Griff. in Itin. Pl. Khasyah Mts.: 171 (1848); <i>Swertia platyphylla</i> Merr. in Lingnan Sci. J. 15: 424 (1936) Homotypic Synonyms: <i>Frumentum triticum</i> E.H.L.Krause in Bot. Centralbl. 73: 339 (1898), nom. superfl.; <i>Triticum aestivum</i> proles vulgare Thell. in Naturwiss. Wochenschr., n.f., 17: 471 (1918), not validly publ.; <i>Triticum aestivum</i> subsp. <i>vulgare</i> Thell. in Fl. Adv. Montpellier: 142 (1912), not validly publ.; <i>Triticum aristatum</i> Schübl. in Diss. Char. Descr. Cereal.: 11 (1818), nom. superfl.; <i>Triticum aristatum aestivum</i> (L.) Schübl. in Diss. Char. Descr. Cereal.: 12 (1818), nom. inval.; <i>Triticum cereale</i> var. <i>aestivum</i> (L.) Klett & Richt. in Fl. Leipzig: 118 (1830); <i>Triticum sativum</i> Lam. in Fl. Franç. 3: 625 (1779), nom. superfl.; <i>Triticum sativum</i> var. <i>aestivum</i> (L.) Pers. in Syn. Pl. 1: 109 (1805), not validly publ.; <i>Triticum sativum</i> var. <i>vulgare</i> Desv. in Opusc. Sci. Phys. Nat.: 162 (1833), not validly publ.; <i>Triticum sativum</i> subsp. <i>vulgare</i> (Vill.) P.Fourn. in Quatre Fl. France: 89 (1934), not validly publ.; <i>Triticum vulgare</i> Vill. in Hist. Pl. Dauphiné 2: 153 (1787), nom. superfl.; <i>Triticum vulgare</i> var. <i>aestivum</i> (L.) Willd. in Enum. Pl. 1: 133 (1809), not validly publ.; <i>Triticum vulgare</i> subsp. <i>aestivum</i> (L.) Arcang. in Comp. Fl. Ital.: 803 (1882), nom. illeg.; <i>Triticum vulgare</i> convar. <i>aristatum</i> Alef. in Landw. Fl.: 330 (1866), not validly publ.; <i>Triticum vulgare</i> var. <i>aristatum</i> Döll in Rhein. Fl.: 70 (1843), not validly publ.	Annual or biennial	Cultigen from Transcaucasus to Israel and NW. Iran, S. Pakistan to NW. India	Temperate biome. Cultivated. Alt. 0-1500 m	1-3
<i>Woodwardia virginica</i> (L.) Sm.	<i>Woodwardia virginica</i> (L.) Sm., Mém. Acad. Roy. Sci. (Turin) 5: 412 (1793)	<i>Aspleniaceae</i> Newman	Heterotypic Synonyms: <i>Anchistea virginica</i> C.Presl in Abh. Königl. Böhm. Ges. Wiss., ser. 5, 6: 71 (1851); <i>Anchistea virginica</i> f. <i>fertilis</i> (Farw.) M.Broun in Index No. Amer. Ferns: 17 (1938); <i>Blechnum banisterianum</i> Poir. in J.B.A.M.de Lamarck, Encycl., Suppl. 1: 644 (1811); <i>Blechnum carolinianum</i> Walter in Fl. Carol.: 257 (1788); <i>Blechnum virginicum</i> L. in Mant. Pl. 2: 307 (1771); <i>Doodia virginica</i> C.Presl in Tent. Pterid.: 99 (1836); <i>Lorinseria thelypteroides</i> C.Presl in Abh. Königl. Böhm. Ges. Wiss., ser. 5, 6: 72 (1851); <i>Woodwardia bannisteriana</i> Michx. in Fl. Bor.-Amer. 2: 263 (1803); <i>Woodwardia thelypteroides</i> Pursh in Fl. Amer. Sept. 2: 670 (1814); <i>Woodwardia virginica</i> f. <i>fertilis</i> Farw. in Pap. Michigan Acad. Sci. 3: 89 (1924); <i>Woodwardia virginica</i> f. <i>thelypteroides</i> Gilbert in List N. Amer. Pterid.: 21, 40 (1901)	Perennial or helophyte	E. Canada to E. Central & E. U.S.A.	Temperate biome. Swamps, marshes, bogs, and roadside ditches over non-calcareous substrates. Alt. 0-300 m	1, 6, 23

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Table S8. Biological activities of sesterterpenoids

Section 1. Biological activities of linear acyclic (AI) sesterterpenoids lacking heterocyclic rings

c. n.	IUPAC name or Trivial name	Bioactivity (reference)
AI-1	geranylfarnesol	Anti-cancer ¹ , Anti-protozoal ²
AI-2	ω -hydroxygeranylfarnesol	
AI-3	geranylnerolidol	
AI-4	3,7,11,15,19-pentamethyl-2-cis-6-trans-eicosadien-1-ol	Anti-fungal ³
AI-5	postrediene A (2,6,11,15,19-pentamethyl-6 <i>E</i> ,14 <i>E</i> -eicosadiene2,3,10,11,18,19-hexanol)	
AI-6	postrediene B (2,6,11,15,19-pentamethyl-10,11,18,19-tetrahydroxyl-2 <i>E</i> ,6 <i>E</i> ,14 <i>E</i> -eicosatrienoic acid)	
AI-7	postrediene C (2,6,11,15,19-pentamethyl-10,11,18,19-tetrahydroxyl-6 <i>E</i> ,14 <i>E</i> -eicosadienoic acid)	Anti-fungal ³
AI-8	(2 <i>Z</i> ,6 <i>Z</i> ,10 <i>E</i> ,14 <i>E</i>)-geranylfarnesol	
AI-9	(2 <i>Z</i> ,6 <i>E</i> ,10 <i>E</i> ,14 <i>E</i>)-geranylfarnesol	
AI-10	unnamed acyclic sesterterpene (reported as compound 5)	Enzyme Inhibitor (Protein tyrosine phosphatase 1B inhibition) ⁴
AI-11	2,6,10,14-tetramethyl-18-butanecarboxymethylene-henecos-12-en-17 β -ol	
AI-12	unnamed acyclic sesterterpene (reported as compound 3)	
AI-13	unnamed acyclic sesterterpene (reported as compound 4)	
AI-14	hasla-6(17),23-diene	
AI-15	hasla-6(17),9,23-triene	
AI-16	hasla-6(17),9,13,23-tetraene	
AI-17	hasla-2,6(17),9,13,23-pentaene	
AI-18	hasla-5,9,13,23-tetraene	
AI-19	hasla-5,23-diene	
AI-20	hasla-5,9,23-triene	
AI-21	hasla-2,5,9,13,23-pentaene	
AI-22	2 <i>E</i> -3,7,11,15,19-pentamethyleicos-2-en-1-ol (<i>Z</i> isomer reported in the structure)	
AI-23	2,6,10,14-tetramethyl-7-(3-methylenepent-4-enyl)pentadec-5,9-diene	
AI-24	2,6,14-trimethyl-10-methylene-9-(3-methylenepent-4-enyl)pentadec-6-diene	
AI-25	pentaene VIII	
AI-26	postrediene D	Anti-microbial ⁵

Section 2. Biological activities of linear monoheterocyclic (MH) sesterterpenoids

c. n.	IUPAC name or Trivial name	Bioactivity (reference)
MH-1	rhopaloic acid H	Anti-gastrulation ⁶
MH-2	rhopaloic acid B	
MH-3	rhopaloic acid C	
MH-4	6''- <i>O</i> -acetylbalibaloside	Anti-gastrulation ⁶
MH-5	6'''- <i>O</i> -acetylbalibaloside	
MH-6	6'',6'''- <i>O</i> -diacetylbalibaloside	
MH-7	rhopaloic acid A	Anti-cancer ^{7,8} , Anti-gastrulation ⁶
MH-8	luffarin Q	
MH-9	balibaloside	
MH-10	acantholide C	
MH-11	hippolide C	
MH-12	hippolide D	
MH-13	hippolide E	Anti-inflammatory ¹⁰
MH-14	hippolide F	
MH-15	hippolide G	
MH-16	hippolide H	
MH-17	unnamed monoheterocyclic sesterterpene (reported as C21-furanoterpene)	
MH-18	furospinosulin-1	
MH-19	3-(4,8,12,16-tetramethylheptadeca-3,7,11,15-tetraenyl)-thiophene-1-oxide	Anti-cancer ¹¹⁻¹³
MH-20	sarcotin O	Anti-cancer ¹⁴
MH-21	sarcotin P	Anti-cancer ¹⁵
MH-22	4-acetoxy-9-deoxoidiadione	Anti-cancer ¹⁶

MH-23	4-hydroxy-9-deoxoidiadione	Anti-cancer ¹⁶
MH-24	5,10-dihydroxyfurospinulosine-1	
MH-25	isofurospingin-4	
MH-26	sarcotrine F	
MH-27	isosarcotrine F	
MH-28	idiadione	Anti-cancer ¹⁷ Anti-viral ¹⁸
MH-29	unnamed cyclic peroxide ester	
MH-30	aikupikoxide A	
MH-31	muqubilone B	
MH-32	ircinialactam C	
MH-33	<i>ent</i> -ircinialactam C	
MH-34	ircinialactam D	
MH-35	unnamed monoheterocyclic sesterterpene (reported as compound 1)	
MH-36	unnamed monoheterocyclic sesterterpene (reported as compound 3)	
MH-37	barangcadoic acid	Enzyme Inhibitor (RCE-protease) ¹⁹
MH-38	rhopaloic acid D	
MH-39	rhopaloic acid E	
MH-40	rhopaloic acid F	
MH-41	rhopaloic acid G	
MH-42	sarcotragin B	
MH-43	furospingin 3	
MH-44	furospingin 4	
MH-45	unnamed monoheterocyclic sesterterpene (reported as compound 2)	
MH-46	sarcotin I	Anti-cancer ²⁰
MH-47	sarcotin J	
MH-48	sigmosceptrellin D methyl ester	
MH-49	sigmosceptrellin E methyl ester	
MH-50	coleifolide A	
MH-51	coleifolide B	
MH-52	granuloside	
MH-53	cacolic acid A	
MH-54	sarcotragin C	
MH-55	ircinialactam G	Anti-cancer ²¹
MH-56	8-hydroxy ircinialactam G	
MH-57	dactylospene A	
MH-58	furospinosulin B	
MH-59	fasciospongide C	
MH-60	hippotulosa A	
MH-61	hippotulosa B	
MH-62	hippotulosa C	
MH-63	hippotulosa D	
MH-64	rhopaloic acid H	

Section 3. Biological activities of linear diheterocyclic (DH) sesterterpenoids

c. n.	IUPAC name or Trivial name	Bioactivity (reference)
DH-1	furospingin 1	Anti-contraction ²² , Enzyme Inhibitor (Protein tyrosine phosphatase (PTP) 1B) ²³
DH-2	anhydrofurospingin 1	Enzyme Inhibitor (GSK-3 β Inhibitor), Anti-cancer ²⁴ , Anti-cancer ²⁵ , Anti-microbial ²⁶ , Anti-cancer ²⁵
DH-3	palinurin	
DH-4	isopalinurin	
DH-5	palinurin sulfate ester	
DH-6	okinonellin B	Anti-cancer ²⁷
DH-7	fasciculatin	
DH-8	fasciculatin sulfate ester	
DH-9	irciformonin A* revised as TH-19	
DH-10	irciformonin B	Anti-cancer ²⁸
DH-11	15-acetyl-irciformonin B	Anti-cancer ²⁸
DH-12	10-acetyl-irciformonin B	Anti-cancer ²⁹
DH-13	irciformonin I	

DH-14	5-(13-(3-furanyl)-2,6, 10-trimethyl-6,8-tridecadienyl)-4-hydroxy-3-methyl-2(5H)-furanone	
DH-15	5-(13-(3-furanyl)-2,6, 10-trimethyl-2,6,8-tridecatrienyl)-4-hydroxy-3-methyl-2(5H)-furanone	
DH-16	variabilin	Anti-cancer ³⁰ , Anti-inflammatory ³¹
DH-17	5-oxo-7 <i>E</i> ,12 <i>E</i> ,20 <i>Z</i> -variabilin	
DH-18	5-hydroxy-7 <i>E</i> ,12 <i>E</i> ,20 <i>Z</i> -variabilin	
DH-19	5-oxo-8(10) <i>E</i> ,12 <i>E</i> ,20 <i>Z</i> -variabilin	
DH-20	5-oxo-8(10) <i>E</i> ,13,20 <i>Z</i> -variabilin	
DH-21	8-hydroxy-12 <i>E</i> ,20 <i>Z</i> -variabilin	
DH-22	25-hydroxy-variabilin	
DH-23	(18 <i>R</i>)-variabilin sulfate ester	
DH-24	(13 <i>Z</i> ,18 <i>R</i>)-variabilin sulfate ester	
DH-25	(12 <i>E</i> ,18 <i>S</i> ,20 <i>Z</i>)-8-hydroxyvariabilin	Anti-inflammatory ³²
DH-26	(7 <i>E</i> ,12 <i>E</i> ,18 <i>S</i> ,20 <i>Z</i>)-variabilin	
DH-27	(7 <i>E</i> ,12 <i>Z</i> ,18 <i>S</i> ,20 <i>Z</i>)-variabilin	
DH-28	(7 <i>E</i> ,12 <i>E</i> ,18 <i>R</i> ,20 <i>Z</i>)-variabilin	Anti-cancer ²⁵
DH-29	variabilin fatty acid esters	
DH-30	psammocinin A ₁	Anti-cancer ²⁵
DH-31	psammocinin A ₂	Anti-cancer ²⁵
DH-32	strobilin	
DH-33	(8 <i>Z</i> ,13 <i>Z</i> ,20 <i>Z</i>)-strobilin	
DH-34	(8 <i>Z</i> ,13 <i>Z</i> ,18 <i>S</i> ,20 <i>Z</i>)-strobilin	
DH-35	(7 <i>Z</i> ,13 <i>Z</i> ,20 <i>Z</i>)-felixinin	
DH-36	sarcotin A	Anti-cancer ²⁰
DH-37	sarcotin B	Anti-cancer ²⁰
DH-38	sarcotin C	Anti-cancer ²⁰
DH-39	<i>epi</i> -sarcotin A	Anti-cancer ²⁰
DH-40	sarcotin M	Anti-cancer ²⁰
DH-41	(8 <i>E</i> ,13 <i>Z</i> ,18 <i>R</i> ,20 <i>Z</i>)-strobilin	
DH-42	(8 <i>Z</i> ,13 <i>E</i> ,18 <i>R</i> ,20 <i>Z</i>)-strobilin	
DH-43	cacolide A	
DH-44	ircinin-3	
DH-45	ircinin-4	
DH-46	furodendin	
DH-47	unnamed diheterocyclic sesterterpene (reported as compound 6a)	
DH-48	unnamed diheterocyclic sesterterpene (reported as compound 8a)	
DH-49	22-deoxyvariabilin	
DH-50	ircinic acid	
DH-51	palomin	
DH-52	unnamed diheterocyclic sesterterpene (reported as compound 5)	
DH-53	unnamed diheterocyclic sesterterpene (reported as epoxide 12)	
DH-54	unnamed diheterocyclic sesterterpene (reported as compound 1)	
DH-55	unnamed diheterocyclic sesterterpene (reported as compound 3)	
DH-56	dehydrofurodendin	Anti-viral (HIV-1 inhibitor) ³³
DH-57	astakolactin	
DH-58	luffarin R	
DH-59	luffarin T	
DH-60	luffarin U	
DH-61	unnamed diheterocyclic sesterterpene (reported as compound 1a)	
DH-62	ircinialactone A	
DH-63	cacolide B	
DH-64	cacolide C	
DH-65	cacolide D	
DH-66	cacolide E	
DH-67	cacolide F	
DH-68	cacolide G	
DH-69	psammocinin B	Anti-cancer ²⁵
DH-70	irciformonin C	Anti-cancer ³⁴
DH-71	irciformonin D	Anti-cancer ³⁴
DH-72	(6 <i>Z</i>)-luffarin-V	
DH-73	luffarin V	
DH-74	sarcotin F	Anti-cancer ²⁰
DH-75	<i>epi</i> -sarcotin F	

DH-76	thorectolide	Anti-viral (HIV-1 inhibitor) ³⁵
DH-77	thorectolide monoacetate	Anti-cancer ³⁵
DH-78	cacospongionolide D	
DH-79	erectusolide	
DH-80	hippolide A	Anti-cancer ¹⁰
DH-81	hippolide B	Anti-cancer ¹⁰
DH-82	cacolide H	
DH-83	cacolide I	
DH-84	cacolide J	
DH-85	cacolide K	
DH-86	cacolide L	
DH-87	sarcotrine A	Anti-cancer ²⁰
DH-88	<i>epi</i> -sarcotrine A	Anti-cancer ²⁰
DH-89	sarcotrine C	Anti-cancer ²⁰
DH-90	<i>epi</i> -sarcotrine C	
DH-91	sarcotrine D	
DH-92	palinurine A	
DH-93	palinurine B	
DH-94	sarcotrine E	
DH-95	isosarcotrine E	
DH-96	ircinialactam A	
DH-97	8-hydroxyircinialactam A	
DH-98	8-hydroxyircinialactam B	
DH-99	unnamed diheterocyclic sesterterpene (reported as compound 1)	
DH-100	dihydrofurospingin 2	
DH-101	fasciospongide B	
DH-102	irciformonin F	Anti-cancer ²⁸
DH-103	irciformonin G	
DH-104	irciformonin H	
DH-105	irciformonin J	
DH-106	luffarin S	
DH-107	(18 <i>R</i>)-variabilin	
DH-108	(7 <i>E</i> ,13 <i>Z</i> ,18 <i>S</i> ,20 <i>Z</i>)-variabilin	
DH-109	22-deoxy-23-hydroxymethylvariabilin	

Section 4. Biological activities of linear triheterocyclic (TH) sesterterpenoids and related dimers

c. n.	IUPAC name or Trivial name	Bioactivity (reference)
TH-1	nitenin	Anti-plasmodial ³⁶
TH-2	dihydronitenin	
TH-3	isonitenin	
TH-4	unnamed triheterocyclic sesterterpene (reported as compound 1)	
TH-5	unnamed triheterocyclic sesterterpene (reported as compound 3)	
TH-6	sarcotin D	Anti-cancer ²⁰
TH-7	sarcotin E	Anti-cancer ²⁰
TH-8	ircinin-1	Anti-cancer ^{37, 38}
TH-9	ircinin-2	Anti-cancer ³⁸
TH-10	ircinin-1 sulfate ester	
TH-11	ircinin-2 sulfate ester	
TH-12	24-hydroxyircinolide	
TH-13	spongionellin	
TH-14	dehydro-spongionellin	
TH-15	cometin A	
TH-16	cometin B	
TH-17	cometin C	
TH-18	ircinolide	
TH-19	irciformonin A (revised structure of DH-19)	
TH-20	irciformonin E	
TH-21	irciformonin K	
TH-22	sarcotin G	
TH-23	sarcotin H	
TH-24	ircinialactam F	

TH-25	sulawesin C
TH-26	hippospongins A
TH-27	hippospongins B
TH-28	hippospongins C
TH-29	hippospongins D
TH-30	hippospongins E
TH-31	hippospongins F
TH-32	(12 <i>E</i> ,18 <i>R</i> ,20 <i>Z</i>)-ircinialactam E

Section 5. Biological activities of monocarbocyclic (MC) sesterterpenoids (with or without heterocycles)

c. n.	IUPAC name or Trivial name	Bioactivity (reference)
MC-1	acantholide E	Anti-cancer ³⁹
MC-2	acantholide D	
MC-3	diacardiol A	Anti-cancer ⁴⁰
MC-4	phorbin A	Anti-cancer ⁴¹
MC-5	thorectidaelide A	Anti-cancer ⁴²
MC-6	thorectidaelide B	
MC-7	thorectidaelide D	
MC-8	thorectidaelide E	
MC-9	sulawesin A	Anti-cancer ³⁸
MC-10	sulawesin B	Anti-cancer ³⁸
MC-11	thorectidaelide C	
MC-12	acantholide A	
MC-13	manoalide	Quorum Sensing inhibitors ⁴³ , Anti-microbial ⁴⁴ , Anti-inflammatory ⁴⁵ , Anti-cancer ^{46, 47}
MC-14	manoalide 25-monoacetate	Quorum Sensing inhibitors ⁴³
MC-15	25-acetoxyseco-manoalide	
MC-16	(6 <i>E</i>)-neomanoalide	Anti-microbial ⁴⁸
MC-17	(6 <i>Z</i>)-neomanoalide	Anti-microbial ⁴⁸
MC-18	<i>E</i> -neomanoalide-24-al	Anti-microbial ⁴⁹
MC-19	24- <i>O</i> -ethylmanoalide	
MC-20	24- <i>O</i> -propylmanoalide	
MC-21	24- <i>O</i> -methylmanoalide	
MC-22	deoxymanoalide	Anti-microbial ⁵⁰ , Anti-inflammatory ⁵¹
MC-23	<i>Z</i> -2,3-dihydroneomanoalide	Anti-microbial ⁴⁹
MC-24	<i>Z</i> -24-acetoxy-2,3-dihydroneomanoalide	Anti-microbial ⁴⁹
MC-25	<i>Z</i> -24-acetoxyneomanoalide	Anti-microbial ⁴⁹
MC-26	6 <i>E</i> -neomanoalide-24,25-diacetate	
MC-27	6 <i>Z</i> -neomanoalide-24,25-diacetate	
MC-28	secomanoalide	Anti-microbial ⁵²
MC-29	deoxysecomanoalide	Anti-microbial ⁵⁰ Anti-inflammatory ⁵¹
MC-30	aplysinoplide A	Anti-cancer ⁵³
MC-31	luffariolide H	Anti-cancer ⁵⁴
MC-32	luffariolide J	Anti-cancer ⁵⁴
MC-33	aplysinoplide B	Anti-cancer ⁵³
MC-34	aplysinoplide C	Anti-cancer ⁵³
MC-35	acantholide B	Anti-microbial, Anti-fungal ³⁹
MC-36	(+)-muquibilin (= prianicin A)	
MC-37	(-)-muquibilin ((-)-muquibilin A)	Anti-cancer ⁵⁵
MC-38	muquibilin ester	
MC-39	hurghaperoxide	
MC-40	tasnemoxide A	Anti-cancer ⁵⁶
MC-41	tasnemoxide B	Anti-cancer ⁵⁶
MC-42	tasnemoxide C	Anti-cancer ⁵⁶
MC-43	4-acetoxythorectidaelide A	
MC-44	cyclolinteinol	Anti-inflammatory ⁵⁷
MC-45	cyclolinteinol acetate	Anti-inflammatory ⁵⁷
MC-46	<i>epi</i> -muquibilin A (dubious configurational assignment)	Anti-inflammatory ^{58, 59}
MC-47	muquibilin B (dubious configurational assignment)	

MC-48	<i>epi</i> -muquubilin B (dubious configurational assignment)	Anti-cancer ⁴⁰
MC-49	diacarnoxide A	Anti-cancer ⁶⁰
MC-50	diacarnoxide B	Anti-cancer ⁶⁰
MC-51	diacarnoxide C	Anti-cancer ⁶⁰
MC-52	diacarnoxide D	Anti-cancer ⁶⁰
MC-53	luffariellin A	
MC-54	fasciospongide A	
MC-55	25-acetoxyluffariellin A	
MC-56	luffariellin B	
MC-57	luffalide A	
MC-58	luffalide B	
MC-59	luffalide C	
MC-60	luffalide D	
MC-61	luffalide E	
MC-62	luffalide F	
MC-63	luffariellin C	
MC-64	luffariellin D	
MC-65	luffarin P	
MC-66	(4 <i>E</i> ,6 <i>E</i>)-dehydro-25- <i>O</i> -methylmanoalide	Anti-cancer ⁶¹
MC-67	untenic acid	
MC-68	(4 <i>E</i> ,6 <i>E</i>)-dehydromanoalide	
MC-69	25-acetoxyluffariellin B	
MC-70	luffariellolide	Anti-inflammatory ⁶²
MC-71	igernellin	
MC-72	16-hydroxyluffariellolide	
MC-73	dehydroluffariellolide diacid	
MC-74	isodehydrouffariellolide	
MC-75	unnamed monocarbocyclic sesterterpene (reported as compound 1)	
MC-76	unnamed monocarbocyclic sesterterpene (reported as compound 2)	
MC-77	luffariolide A	Anti-cancer ⁶³
MC-78	luffariolide B	Anti-cancer ⁶³
MC-79	luffariolide C	Anti-cancer ⁶³
MC-80	luffariolide D	Anti-cancer ⁶³
MC-81	luffariolide E	Anti-cancer ⁶³
MC-82	luffariolide F	
MC-83	luffariolide G	
MC-84	(2 <i>E</i> ,6 <i>E</i> ,10 <i>E</i>)-3-formyl-7,11-dimethyl-13-(2,6,6-trimethylcyclohex-1-enyl)trideca-2,6,10-trienoic acid	
MC-85	sarcotragin A	
MC-86	hipposulfate C	Enzyme Inhibitor (Calcineurin Inhibitor) ⁶⁴
MC-87	unnamed monocarbocyclic sesterterpene (reported as compound 1)	
MC-88	unnamed monocarbocyclic sesterterpene (reported as compound 2)	
MC-89	unnamed monocarbocyclic sesterterpene (reported as compound 3)	
MC-90	unnamed monocarbocyclic sesterterpene (reported as compound 4)	
MC-91	unnamed monocarbocyclic sesterterpene (reported as compound 5)	Anti-cancer ⁶⁵
MC-92	unnamed monocarbocyclic sesterterpene (reported as compound 6)	Anti-cancer ⁶⁵
MC-93	unnamed monocarbocyclic sesterterpene (reported as compound 7)	Anti-cancer ⁶⁵
MC-94	unnamed monocarbocyclic sesterterpene (reported as compound 8)	
MC-95	unnamed monocarbocyclic sesterterpene (reported as compound 9)	
MC-96	halisulfate 2	
MC-97	polyunsaturated sesterterpene I	
MC-98	<i>n</i> -non-2'-en-1'-yl-13(15,19,19-trimethyl-cyclohex-14, 16-dienyl)- 2,6,10-trimethyl-tetradec-6-ol-13-on-1-oate	
MC-99	3-[6-(4,8-dimethyl-nona-1,3,7-trienyl)-4-hydroxy2,6-dimethyl-cyclohex-1-enyl]-3-hydroxypropionic acid 1 glucoside	
MC-100	cericerol-I	
MC-101	α -cericerol-I	
MC-102	cericerol-II	
MC-103	albocerol	
MC-104	cericerioic acid	
MC-105	18-dihydro-19-hydroxycericerioic acid	
MC-106	ceriferic acid	
MC-107	ceriferic acid-I	

MC-108	ceriferol	
MC-109	ceriferol-I	
MC-110	13-ethoxycericerene	
MC-111	13-methoxycericerene	
MC-112	(2Z,6Z,10E)-cericerene-15,24-diol	
MC-113	24-(ω-hydroxy fatty acid) ester of (2Z,6Z,10E)-cericerene-15,24-diol	
MC-114	(2E)-α-cericerene	
MC-115	(-)-alotaketol A	Anti-cancer ⁶⁶
MC-116	alotaketol B	Anti-cancer ⁶⁶
MC-117	alotaketol C	Pro-viral ⁶⁷
MC-118	alotaketol D	Pro-viral ⁶⁷
MC-119	phorbaketol A	Anti-osteoporosis ⁶⁸ , Anti-inflammatory ⁶⁹ , Anti-cancer ⁷⁰
MC-120	phorbaketol B	Anti-cancer ⁷⁰
MC-121	phorbaketol C	Anti-cancer ⁷⁰
MC-122	phorbaketol D	
MC-123	phorbaketol E	Anti-cancer ⁷¹
MC-124	phorbaketol F	
MC-125	phorbaketol G	
MC-126	phorbaketol H	Anti-cancer ⁷¹
MC-127	phorbaketol I	Anti-cancer ⁷¹
MC-128	phorbaketol J	
MC-129	phorbaketol K	
MC-130	phorbaketol L	
MC-131	phorbaketol M	
MC-132	phorbaketol N	Anti-cancer ⁷²
MC-133	unnamed spiroketol sesterterpene (reported as spiroketol)	
MC-134	alotaketol E	
MC-135	unnamed monocarbocyclic sesterterpene (reported as compound 2)	
MC-136	unnamed monocarbocyclic sesterterpene (reported as compound 3)	
MC-137	unnamed monocarbocyclic sesterterpene (reported as compound 4)	
MC-138	2Z,6Z,10E-cericerene-15,24-diol	
MC-139	2Z,6Z,10E-cericerene-15,24-diol-30-triacontanoate	
MC-140	cyclolinteinon	Anti-inflammatory ⁷³
MC-141	(S,2E,6E,10E,14E)-3,7,11,15-tetramethyl-18-(1-hydroxyisopropyl)cyclooctadecatetraene	
MC-142	(S,2E,6E,10E,14E)-3,7,11,15-tetramethyl-18-(1-methylethenyl)cyclooctadecatetraene	
MC-143	(S,2E,15Z)-α-cericerene	
MC-144	(S,2E)-cericerene	
MC-145	(14S,15R,2E)-15-hydroxy-α-cericerene	
MC-146	oshimalide A	Anti-microbial ⁷⁴
MC-147	oshimalide B	
MC-148	16-oxoluffariellolide	
MC-149	(-)-somialensene B	
MC-150	(+)-a-geranylbisabolene	
MC-151	(+)-somialensene B	
MC-152	postrediene F	
MC-153	leucosceptrodine	Immuno-suppressive ⁵

Section 6. Biological activities of dicarbocyclic (DC) sesterterpenoids (with and without hetherocycles)

c. n.	IUPAC name or Trivial name	Bioactivity (reference)
DC-1	bipolarolide E	
DC-2	palauolol	Anti-cancer ⁷⁵
DC-3	unnamed dicarbocyclic sesterterpene (reported as compound 5)	
DC-4	kohamaic acid A	Anti-cancer ^{76, 77}
DC-5	kohamaic acid B	
DC-6	cladocoran A	Anti-inflammatory ^{78, 79}
DC-7	cladocoran B	Anti-inflammatory ⁷⁸
DC-8	thorectandrol A	Anti-cancer ⁷⁵
DC-9	thorectandrol B	Anti-cancer ⁷⁵
DC-10	palauolide	Anti-cancer ⁸⁰

DC-11	thorectandrol C	Anti-cancer ⁸⁰
DC-12	thorectandrol D	Anti-cancer ⁸⁰
DC-13	thorectandrol E	Anti-cancer ⁸⁰
DC-14	unnamed dicarbocyclic sesterterpene (reported as compound 1)	
DC-15	mycaperoxide A	Anti-cancer, Anti-microbial, Anti-viral ⁸¹
DC-16	mycaperoxide B	Anti-cancer, Anti-microbial, Anti-viral ⁸¹
DC-17	sigmosceptrellin B (= prianicin B) (dubious configurational assignment) (reported as compound 2)	
DC-18	25-deoxycacospongionolide B	
DC-19	cacospongionolide B	Anti-inflammatory ⁸²⁻⁸⁴
DC-20	cacospongionolide E	Anti-inflammatory ⁸⁵
DC-21	cacospongionolide F	Anti-microbial ⁸⁶
DC-22	dysidiolide	Anti-cancer ⁸⁷
DC-23	dysideapalaunic acid	
DC-24	bilosespene A	
DC-25	bilosespene B	
DC-26	irregularasulfate	Immunosuppressive ⁶⁴
DC-27	fasciospongine A	Anti-fungal ⁸⁸
DC-28	fasciospongine B	Anti-fungal ⁸⁸
DC-29	(+)-agelisamine-A	
DC-30	(+)-agelisamine-B	
DC-31	halisulfate 7	Enzyme Inhibitor (Calcineurin Inhibitor) ⁶⁴
DC-32	halisulfate 8	
DC-33	25-hydroxyhalisulfate 9	Anti-fungal ⁸⁹
DC-34	halisulfate 10	
DC-35	halisulfate 9	
DC-36	sch 599473	
DC-37	fasciospongine C	Anti-fungal ⁸⁹
DC-38	halisulfate 3	Anti-viral ⁹⁰
DC-39	halisulfate 4	
DC-40	halisulfate 5	Enzyme Inhibitor (β -lactamase) ⁹¹
DC-41	halisulfate 6	
DC-42	13- <i>epi</i> -yosgadensenol	
DC-43	luffarin A	
DC-44	luffarin B	
DC-45	luffarin C	
DC-46	luffarin D	
DC-47	luffarin E	
DC-48	luffarin F	
DC-49	luffarin G	
DC-50	luffarin H	
DC-51	luffarin I	Anti-cancer ⁹²
DC-52	luffarin J	
DC-53	luffarin K	
DC-54	luffarin M	
DC-55	luffarin L	
DC-56	luffarin N	
DC-57	luffarin O	
DC-58	unnamed dicarbocyclic sesterterpene (reported as compound 1)	
DC-59	salvisyriacolide	
DC-60	sigmosceptrellin A	Anti-microbial ⁹³
DC-61	sigmosceptrellin A methyl ester	Anti-microbial ⁹³ , Anti-cancer ⁹⁴
DC-62	sigmosceptrellin-B methyl ester	
DC-63	sigmosceptrellin-C	
DC-64	salvileucolide methyl ester	Anti-microbial ⁹⁵
DC-65	salvileucolide-6,23-lactone	Anti-microbial ⁹⁵
DC-66	14-hydroperoxy-13(21)-dehydro-13,14-dihydrosalvileucolide-6,23-lactone	
DC-67	salvileucolide-hydroperoxide 2	
DC-68	salvileucolide-cyclic peroxide	
DC-69	salvileucolide-lactone (15,16-deidrosalvileucolide-6,23-lactone-trans-epoxide)	
DC-70	unnamed dicarbocyclic sesterterpene (reported as hydroperoxide 4)	
DC-71	8 <i>R</i> -hydroxy-13-hydroperoxylabd-14,17-dien-19,16,23,6 α -di-olide	

DC-72	17,18,19,20-tetranor-13- <i>epi</i> -manoyloxide-14-en-16-oic acid-23,6 α -olide	
DC-73	8 α ,13,14-threo-trihydroxy-labd-15,17-dien-16,19-olide-23 oic acid	
DC-74	8 α ,13,14-erythro-trihydroxy-labd-15,17-dien-16,19-olide-28-oic acid	
DC-75	3 α ,8 α ,13,14-erythro-tetrahydroxy-labd-15,17-dien-16,19-olide	
DC-76	23,6 α -epoxy-labd-8,13(14),17-trien-16(<i>R</i>),19-olide	
DC-77	8 α ,15(<i>S</i>)-dihydroxy-23,6 α -epoxy-labd-13(14),17-dien-16(<i>S</i>),19-olide	
DC-78	8 α ,15(<i>S</i>),23 α -trihydroxy-23,6 α -epoxy-labd-13(14),17-dien-16(<i>S</i>),19-olide	Anti-cancer ⁹⁶
DC-79	8 α ,15(<i>S</i>)-dihydroxy-23 α - <i>O</i> -ethyl-23,6 α -epoxy-labd-13(14),17-dien-16(<i>S</i>),19-olide	Anti-cancer ⁹⁶
DC-80	8 α -hydroxy-23 α - <i>O</i> -ethyl-23,6 α -epoxy-labd-13(14),17-dien-16(<i>R</i>),19-olide	Anti-cancer ⁹⁶
DC-81	6 α ,8 α ,15(<i>S</i>),23-tetrahydroxy-labd-13(14),17-dien-16(<i>S</i>),19-olide	Anti-cancer ⁹⁶
DC-82	6 α ,8 α ,15(<i>S</i>)-trihydroxy-labd-13(14),17-dien-16(<i>S</i>),19-olide	
DC-83	6 α ,8 α ,15(<i>S</i>)-trihydroxy-23-carbossi-labd-13(14),17-dien-16(<i>S</i>),19-olide	Anti-cancer ⁹⁶
DC-84	6 α ,8 α -dihydroxy-23-carbossi-labd-13(14),17-dien-16,19-olide	Anti-cancer ⁹⁶
DC-85	6 α ,8 α ,15(<i>S</i>)-trihydroxy-23-oxo-labd-13(14),17-dien-16(<i>S</i>),19-olide	Anti-cancer ⁹⁶
DC-86	6 α ,8 α -dihydroxy-23-oxo-labd-13(14),17-dien-16(<i>R</i>),19-olide	Anti-cancer ⁹⁶
DC-87	6 α ,15(<i>S</i>),23-trihydroxy-labd-8(22),13(14),17-trien-16(<i>S</i>),19-olide	Anti-cancer ⁹⁶
DC-88	6 α ,15(<i>S</i>)-dihydroxy-23-oxo-labd-8(22),13(14),17-trien-16(<i>S</i>),19-olide	Anti-cancer ⁹⁶
DC-89	6 α ,8 α ,23-trihydroxy-labd-13(14),15,17-trien-16,19-olide	Anti-cancer ⁹⁶
DC-90	6 α ,8 α -dihydroxy-23-carbossi-labd-13(14),15,17-trien-16,19-olide	Anti-cancer ⁹⁶
DC-91	6 α ,8 α -dihydroxy-23-oxo-13(14),15,17-trien-16,19-olide	Anti-cancer ⁹⁶
DC-92	8 α -hydroxylabd-13(14),15,17-trien-6 <i>R</i> ,23-16,19-diolide	Anti-cancer ⁹⁶
DC-93	8 α -23-dihydroxy-23,6 <i>R</i> -epoxy-labd-13(14),15,17-trien-16,19-olide	Anti-cancer ⁹⁶
DC-94	6 α ,8 α ,23,14,15-threo-pentahydroxy-labd-13(21),17-dien-16,19-olide	
DC-95	6 α ,8 α ,23,14,15-erythro-pentahydroxylabd-13(21),17-dien-16,19-olide	
DC-96	6 α ,8 α ,14,15-threotetrahydroxy-23-oxo-labd-13(21),17-dien-16,19-olide	
DC-97	lachnocalyxolide A	Anti-cancer ⁹⁷
DC-98	lachnocalyxolide B	
DC-99	lachnocalyxolide C	
DC-100	lachnocalyxolide C'	
DC-101	salvidominicolide A	
DC-102	salvidominicolide B	
DC-103	albolineol	
DC-104	terpestacin	Anti-cancer ^{98, 99}
DC-105	11- <i>epi</i> -terpestacin (siccanol)	
DC-106	11- <i>epi</i> -terpestacin glycoside	
DC-107	21-hydroxyterpestacin	
DC-108	24- α -D-glucosyl(-)-terpestacin	
DC-109	terpestacin B	
DC-110	16,17-dihydro(-)-terpestacin	
DC-111	terpestacin C	
DC-112	fusaproliferin (proliferin)	Anti-cancer ¹⁰⁰
DC-113	fusaprolifin A	Anti-microbial ¹⁰¹
DC-114	fusaprolifin B	
DC-115	mycaleperoxide	
DC-116	diacarperoxide D	Anti-cancer ⁴⁰
DC-117	diacarperoxide E	
DC-118	diacarperoxide F	Anti-cancer ⁴⁰
DC-119	diacarperoxide G	
DC-120	(13 <i>E</i>)-8 α -hydroxy-23-carboxymethylabd-13(14),17(18)-dien-16,19-olide	
DC-121	salvimirzacolide	
DC-122	4 <i>R</i> ,5 <i>R</i> ,8 <i>R</i> ,9 <i>R</i> ,10 <i>S</i> ,13 <i>S</i> ,14 <i>S</i>)-14-hydroxymanoyloxide-15,17-dien-15(<i>Z</i>)-16,19-olide	
DC-123	(4 <i>R</i> ,5 <i>R</i> ,8 <i>R</i> ,9 <i>R</i> ,10 <i>S</i> ,13 <i>R</i> ,14 <i>S</i>)-14-hydroxymanoyloxide-15,17-dien-15(<i>Z</i>)-16,19-olide	
DC-124	(4 <i>R</i> ,5 <i>R</i> ,8 <i>R</i> ,9 <i>R</i> ,10 <i>S</i> ,13 <i>S</i> ,14 <i>R</i>)-14-hydroxymanoyloxide-15,17-dien-15(<i>Z</i>)-16,19-olide	
DC-125	(4 <i>R</i> ,5 <i>R</i> ,8 <i>R</i> ,9 <i>R</i> ,10 <i>S</i> ,13 <i>S</i> ,16 <i>R</i>)-14-hydroxymanoyloxide-17-en-16,19-olide	
DC-126	(4 <i>R</i> ,5 <i>R</i> ,8 <i>R</i> ,9 <i>R</i> ,10 <i>S</i> ,13 <i>R</i>)-manoyloxide-15,17-dien-15(<i>Z</i>)-16,19-olide	
DC-127	(13 <i>E</i>)-8 α ,15-dihydroxy-23-carboxymethylabd-13(14),17(18)-dien-16,19-olide	
DC-128	19-oxofasciospongine A	Anti-fungal ⁸⁹ , Anti-cancer ⁸⁹
DC-129	(15 <i>S</i> ,16 <i>S</i> ,13 <i>E</i>)-8 α ,15-dihydroxylabd-13(14),17(18)-dien-23,6 α -16,19-diolide	
DC-130	(16 <i>R</i> ,13 <i>E</i>)-6 α ,8 α ,23-trihydroxylabd-13(14),17(18)-dien-16,19-olide	

DC-131	(15 <i>S</i> ,16 <i>S</i> ,13 <i>E</i>)-6 α ,8 α ,15-trihydroxy-23-carboxymethylabd-13(14),17(18)-dien-16,19-olide	
DC-132	(13 <i>E</i>)-4 α ,6 α ,8 α -trihydroxylabd-13(14),17(18)-dien-16,19-olide	
DC-133	yosgadensenol	
DC-134	yosgadensolide A	
DC-135	yosgadensolide B	
DC-136	6-dehydroxy-13- <i>epi</i> -yosgadensenol	
DC-137	6-dehydroxyyosgadensenol	
DC-138	leucosceptroid J	
DC-139	(14 <i>E</i>)-methylmanoyloxide-14,16,18-trien-19,16-oxide-23-carboxylate	
DC-140	6 α -hydroxymannoyloxide-14,17-dien-16,19-olide	
DC-141	3 β -hydroxymannoyloxide-14,17-dien-16,19-olide	
DC-142	(13 α),6 α -hydroxymannoyloxide-14,17-dien-16,19-olide	
DC-143	(13 <i>E</i>)-8 α -hydroxy-23 α - <i>O</i> -methyl-23,6 α -epoxylabd-13(14),17-(18)-dien-16,19-olide	
DC-144	(13 <i>E</i>)-8 α ,15-dihydroxy-23 α - <i>O</i> -methyl-23,6 α -epoxylabd-13(14),17(18)-dien-16,19-olide	
DC-145	(13 <i>E</i>)-8 α ,23 α -dihydroxy-23,6 α -epoxylabd-13(14),17(18)-dien-16,19-olide	
DC-146	(13 <i>E</i>)-8 α ,23-dihydroxylabd-13(14),17(18)-dien-16,19-olide	
DC-147	(13 <i>E</i>)-labd-13(14),17(18)-dien-8 α ,16,19-triol	
DC-148	trunculin A	
DC-149	trunculin B	
DC-150	trunculin C methyl ester	
DC-151	trunculin D methyl ester	
DC-152	trunculin E	
DC-153	trunculin F	
DC-154	trunculin H	
DC-155	trunculin G	
DC-156	trunculin I	
DC-157	trunculin X	Anti-cancer ¹⁰²
DC-158	trunculin Y	Anti-cancer ¹⁰²
DC-159	ircinianin	Anti-microbial ¹⁰³
DC-160	(+)-wistarin	
DC-161	(-)-wistarin	
DC-162	luffalactone	Anti-inflammatory ⁴⁵
DC-163	sulfircin	Anti-cancer ¹⁰⁴
DC-164	leucosesterlactone	
DC-165	shinsonofuran	Anti-cancer ¹⁰⁵
DC-166	17 α -hydroxyleucosceptrine	
DC-167	genepolide	
DC-168	leucosceptroid A	Anti-feedant ¹⁰⁶
DC-169	leucosceptroid B	Anti-fungal ¹⁰⁶ , Anti-microbial ¹⁰⁷ , Anti-feedant ¹⁰⁶ , Immuno-suppressive ¹⁰⁸
DC-170	biyoulactone D	
DC-171	biyoulactone E	
DC-172	norleucosceptroid A	Anti-feedant ^{109, 110}
DC-173	norleucosceptroid B	Anti-feedant ^{109, 110}
DC-174	norleucosceptroid C	Anti-feedant ^{109, 110}
DC-175	(-)-ircinianin	
DC-176	(-)-ircinianin lactam A	Enzyme Positive Modulator (glycine-gated chloride channel receptors (GlyRs) ¹¹¹
DC-177	(-)-ircinianin lactam A sulfate	
DC-178	(-)-ircinianin lactone A	
DC-179	(-)-ircinianin sulfate	
DC-180	(-)-oxoircinianin	
DC-181	(-)-oxoircinianin lactam A	Enzyme Positive Modulator (glycine-gated chloride channel receptors (GlyRs) ¹¹¹
DC-182	colquhounoid A	Anti-insect ⁴²
DC-183	colquhounoid B	Anti-insect ⁴²
DC-184	colquhounoid C	Anti-insect ⁴²
DC-185	leucosceptroid E	

DC-186	leucosceptroid F	
DC-187	leucosceptroid G	
DC-188	leucosceptroid H	
DC-189	leucosceptroid I	
DC-190	leucosceptroid L	
DC-191	leucosceptroid M	
DC-192	leucosceptroid N	Immuno-suppressive ¹⁰⁸
DC-193	leucosceptroid O	
DC-194	secoepoxyansellone A	
DC-195	gombaspiroketal A	Anti-cancer ¹¹²
DC-196	gombaspiroketal B	Anti-cancer ¹¹²
DC-197	gombaspiroketal C	Anti-cancer ¹¹²
DC-198	leucosceptroid P	
DC-199	leucosceptroid Q	
DC-200	norleucosceptroid D	Anti-feedant ¹¹⁰
DC-201	norleucosceptroid E	Anti-feedant ¹¹⁰
DC-202	norleucosceptroid F	Anti-feedant ¹¹⁰
DC-203	norleucosceptroid G	Immuno-suppressive ¹⁰⁸
DC-204	norleucosceptroid H	
DC-205	emericlellene A	
DC-206	emericlellene B	
DC-207	emericlellene C	
DC-208	emericlellene D	
DC-209	emericlellene E	
DC-210	bipolarenic acid	
DC-211	leucosceptrine	
DC-212	1 α -hydroxyleucosceptrine	Enzyme Inhibitor (prolylendopeptidase) ¹¹³
DC-213	8 α -hydroxyleucosceptrine	
DC-214	goniocarpic acid	
DC-215	unnamed dicarbocyclic sesterterpene (reported as compound 2a)	Anti-cancer ¹¹⁴
DC-216	(-)-hippolide J	Anti-cancer ¹¹⁵
DC-217	(+)-hippolide J	
DC-218	raoulic acid	
DC-219	3- <i>epi</i> -salviaethiopisolid and C-16 epimer	Anti-viral ¹¹⁶
DC-220	salviaethiopisolid and C-16 epimer	
DC-221	3 α -hydroxymanoyloxide-14(<i>E</i>),17-dien-16-oxo-19-oic acid	
DC-222	hydroxymanoyloxide-14,17-dien-16-oxo-19-oic acid	
DC-223	hydroxymanoyloxide-14,17-dien-16-oxo-19,23-dioic acid	
DC-224	dactylospene B	Anti-inflammatory ²¹
DC-225	dactylospene C	Anti-cancer ²¹
DC-226	dactylospene D	
DC-227	dactylospene E	
DC-228	enantio sigmosceptrellin-A methyl ester	
DC-229	leucosceptroid K	
DC-230	clavaphyllene	
DC-231	(+)-brassitetraene A	
DC-232	(+)-brassitetraene B	
DC-233	fusaproliferene	
DC-234	unnamed dicarbocyclic sesterterpene (reported as compound 2)	
DC-235	Bm3	
DC-236	fusaproliferin	Anti-inflammatory ¹¹⁷
DC-237	sestersorokinicin A	
DC-238	sestersorokiniside A	
DC-239	somaliensene A	
DC-240	ircinianin lactone B	
DC-241	ircinianin lactone C	
DC-242	bipolariterpene A	
DC-243	bipolariterpene B	
DC-244	bipolariterpene C	
DC-245	preterpestacin I	
DC-246	colquhounoid D	
DC-247	14- <i>epi</i> -colquhounoid D	

DC-248	unnamed tetracarboxylic sesterterpene	
DC-249	sesterorbiculene	
DC-250	sesterviolene E	
DC-251	sesterviolene F	
DC-252	patriniaterpene C	
DC-253	patriniaterpene D	
DC-254	arthproliferin A	
DC-255	arthproliferin B	
DC-256	arthproliferin C	
DC-257	arthproliferin D	
DC-258	dactylospene F	
DC-259	colquhounoid E	Immuno-suppressive ¹¹⁸
DC-260	norcolquhounoid F	
DC-261	colquhounoids F	Immuno-suppressive ¹¹⁸
DC-262	bipolariterpene D	Anti-fungal ¹¹⁹
DC-263	bipolariterpene E	
DC-264	<i>nor</i> -leucosceptroid L	Immuno-suppressive ¹²⁰
DC-265	<i>nor</i> -leucosceptroid M	
DC-266	<i>epi-nor</i> -leucosceptroid J	
DC-267	<i>nor</i> -leucosceptroid N	
DC-268	16- <i>epi-nor</i> -leucosceptroid N	
DC-269	3-H-2-hydroxy-leucosceptroid N	
DC-270	3-H-2,17 α -dihydroxy-leucosceptroid N	Immuno-suppressive ^{108, 120, 121}
DC-271	3-H-2-hydroxy-leucosceptroid R	
DC-272	17- <i>epi</i> -leucosceptroid N	
DC-273	5 α ,16 α -epoxy-leucosceptroid R	
DC-274	13 β ,16 α -epoxy-3-H-2-hydroxy-leucosceptroid R	
DC-275	13 β ,16 α -epoxy-17 α -hydroxy-leucosceptroid N	
DC-276	maydistacin A	Anti-inflammatory ¹²²
DC-277	maydistacin B	
DC-278	maydistacin C	
DC-279	maydistacin D	
DC-280	maydistacin E	
DC-281	maydistacin F	
DC-282	maydistacin G	
DC-283	norleucosceptroid I	
DC-284	14- <i>epi</i> -leucosceptroid N	
DC-285	17 α -hydroxy-leucosceptroid N	
DC-286	17 α -hydroxy-14- <i>epi</i> -leucosceptroid N	
DC-287	leucosceptroid R	Immuno-suppressive ¹⁰⁸
DC-288	14- <i>epi</i> -leucosceptroid R	
DC-289	5 α ,17 β -epoxy-14- <i>epi</i> -leucosceptroid N	
DC-290	5 α ,16 α -epoxy-17 α -hydroxy-14- <i>epi</i> -leucosceptroid N	
DC-291	5 α ,16 α -epoxy-leucosceptroid R	Immuno-suppressive ¹⁰⁸
DC-292	leucosceptroid S	
DC-293	11 β -hydroxy-leucosceptroid S	
DC-294	leucosceptroid T	
DC-295	leucosceplactam A	
DC-296	5 α ,17 β -epoxy-leucosceptroid J	Immuno-suppressive ¹⁰⁸
DC-297	13 β ,16 α -epoxy-leucosceptroid R	Immuno-suppressive ¹⁰⁸
DC-298	13 β ,16 α -epoxy-14- <i>epi</i> -leucosceptroid R	
DC-299	11 β -hydroperoxide-leucosceptroid I	
DC-300	11- <i>epi</i> -leucosceptroid Q	
DC-301	13-dehydro-leucosceptroid I	
DC-302	5,13-dehydro-leucosceptroid P	
DC-303	5,13-dehydro-11- <i>epi</i> -leucosceptroid P	
DC-304	11- <i>epi</i> -leucosceptroid M	
DC-305	11- <i>epi</i> -leucosceptroid A	Immuno-suppressive ¹⁰⁸
DC-306	5,13-dehydro-leucosceptroid A	Immuno-suppressive ¹⁰⁸
DC-307	leucosceplactam B	
DC-308	leucosceplactam C	Immuno-suppressive ¹⁰⁸
DC-309	leucosceplactam D	
DC-310	leucosceplactam E	
DC-311	11- <i>epi</i> -leucosceptroid B	Immuno-suppressive ¹⁰⁸

DC-312	leucosceptrine A
DC-313	leucosceptrine B
DC-314	leucosceptrine C
DC-315	leucosceptrine D
DC-316	leucosceptrine E

Section 7. Biological activities of tricarbocyclic (TrC) sesterterpenoids (with and without heterocycles)

c. n.	IUPAC name or Trivial name	Bioactivity (reference)
TrC-1	gascardic acid	
TrC-2	ophiobolin A (ophiobolin, ophiobalin, cochliobolin, cochliobolin A, zizanin) (ophiobola-7,18-dien-21-al-3 α -ol-5-one-14 α ,17-oxide)	Anti-cancer ¹²³ , Phytotoxic ¹²⁴
TrC-3	6- <i>epi</i> -ophiobolin A	Phytotoxic ¹²⁵
TrC-4	3-anhydrophiobolin A	
TrC-5	3-anhydro-6- <i>epi</i> -ophiobolin A	Anti-inflammatory ¹²⁶
TrC-6	3-anhydro-6-hydroxyphiobolin A	Anti-microbial, Anti-cancer ¹²⁷ , Anti-neurodegeneration ¹²⁸
TrC-7	ophiobolin A lactone	Anti-microbial ¹²⁹
TrC-8	ophiobolin B (cochliobolin B, ophiobolosin A, zizanin B) (ophiobola-7,18-dien-21-al-3 α ,14 α -diol-5-one)	Phytotoxic ¹³⁰
TrC-9	3-anhydro-6- <i>epi</i> -ophiobolin B	
TrC-10	3-anhydro-6- <i>epi</i> - $\Delta^{10(14)}$ -ophiobolin B (degradation product)	
TrC-11	di-3-anhydro-6- <i>epi</i> -ophiobolin B (degradation product)	
TrC-12	ophiobolin B lactone	Anti-fungal ¹³¹
TrC-13	ophiobolin C (zizanin A) (ophiobola-7,18-dien-21-al-3 α -ol-5-one)	Anti-cancer ¹³²
TrC-14	6- <i>epi</i> -ophiobolin C	Anti-cancer ¹³²
TrC-15	ophiobolin D (cephalonic acid) (ophiobola-3,6,18-trien-8 β -ol-21-oic acid)	
TrC-16	ophiobolin E	
TrC-17	ophiobolin F	
TrC-18	ophiobolin G	Anti-cancer ^{133, 134}
TrC-19	6- <i>epi</i> -ophiobolin G	Anti-cancer ^{133, 134}
TrC-20	hedyosulide	Anti-microbial ¹³⁵
TrC-21	14,15-dehydro-(Z)-14-ophiobolin G	Anti-cancer ¹³²
TrC-22	14,15-dehydrophiobolin G	Anti-cancer ¹³²
TrC-23	14,15-dehydro-6- <i>epi</i> -ophiobolin G	Anti-cancer ¹³²
TrC-24	(6 α)-18,19,21,21- <i>O</i> -tetrahydro-18,19-dihydroxyphiobolin G	
TrC-25	(6 α)-21,21- <i>O</i> -dihydrophiobolin G or 21-deoxo-21-hydroxy-6- <i>epi</i> -ophiobolin G	
TrC-26	(6 α)-21-deoxyphiobolin G	
TrC-27	(6 α)-16,17-dihydro-21-deoxyphiobolin G	
TrC-28	ophiobolin H	Anti-inflammatory ¹³⁶
TrC-29	(5 α ,6 α)-ophiobolin H	Anti-microbial ¹³⁷
TrC-30	(5 α ,6 α)-5- <i>O</i> -methylophiobolin H	
TrC-31	5- <i>O</i> -methylophiobolin H	
TrC-32	ophiobolin I	
TrC-33	6- <i>epi</i> -ophiobolin I	
TrC-34	25-hydroxyphiobolin I	
TrC-35	ophiobolin J	Anti-inflammatory ¹²⁶
TrC-36	8- <i>epi</i> -ophiobolin J	Anti-inflammatory ¹²⁶
TrC-37	8-deoxyphiobolin J	
TrC-38	ophiobolin K	Anti-fungal, Anti-microbial ¹³⁸
TrC-39	6- <i>epi</i> -ophiobolin K	Anti-cancer ¹³⁹
TrC-40	14,15-dehydrophiobolin K	Anti-cancer ¹³⁹
TrC-41	14,15-dehydro-6- <i>epi</i> -ophiobolin K	Anti-cancer ¹³²
TrC-42	21-deoxyphiobolin K	Anti-cancer ¹³²
TrC-43	ophiobolin L	Anti-microbial ¹³⁴
TrC-44	6- <i>epi</i> -ophiobolin L	Anti-microbial ¹³¹
TrC-45	ophiobolin M	
TrC-46	6- <i>epi</i> -ophiobolin M	
TrC-47	ophiobolin N	Anti-cancer ¹³²
TrC-48	6- <i>epi</i> -ophiobolin N	Anti-cancer ¹³²
TrC-49	ophiobolin O	Anti-cancer ^{132, 140}

TrC-50	6- <i>epi</i> -ophiobolin O	Anti-cancer ¹⁴¹
TrC-51	21- <i>epi</i> -ophiobolin O	Anti-cancer ¹⁴¹
TrC-52	ophiobolin P	Anti-microbial ⁴¹
TrC-53	ophiobolin Q	Anti-cancer ¹³⁴
TrC-54	ophiobolin R	
TrC-55	ophiobolin S	
TrC-56	ophiobolin T	Anti-microbial ⁴¹
TrC-57	ophiobolin U*	Anti-microbial, Anti-fungal ^{134, 137}
TrC-58	ophiobolin U*	Anti-microbial ^{134, 137}
TrC-59	21-dehydroophiobolin U	
TrC-60	ophiobolin V	
TrC-61	ophiobolin W	
TrC-62	ophiobolin X	
TrC-63	ophiobolin Y	
TrC-64	ophiobolin Z	Anti-cancer ¹³⁴
TrC-65	21- <i>epi</i> -ophiobolin Z	
TrC-66	ophiobola-7,19-dien-25-oic acid (14,18(<i>R</i>))-epoxy-3,5-dihydroxy- γ -lactone)	
TrC-67	asperophiobolin A	
TrC-68	asperophiobolin B	
TrC-69	asperophiobolin C	
TrC-70	asperophiobolin D	
TrC-71	asperophiobolin E	
TrC-72	asperophiobolin F	
TrC-73	asperophiobolin G	
TrC-74	asperophiobolin H	Anti-inflammatory, Anti-microbial ¹³⁶
TrC-75	asperophiobolin I	Anti-inflammatory ¹³⁶
TrC-76	asperophiobolin J	Anti-inflammatory ¹³⁶
TrC-77	asperophiobolin K	
TrC-78	ophiobotriol	
TrC-79	bipolarin A	
TrC-80	bipolarin B	
TrC-81	bipolarin C	
TrC-82	bipolarin D	
TrC-83	bipolarin E	Anti-microbial ¹⁴²
TrC-84	bipolarin F	
TrC-85	bipolarin G	
TrC-86	bipolarin H	
TrC-87	bipolaricin A	
TrC-88	bipolaricin B	Anti-inflammatory ¹²⁶
TrC-89	bipolaricin C	Anti-inflammatory ¹²⁶
TrC-90	bipolaricin D	
TrC-91	bipolaricin E	
TrC-92	bipolaricin F	
TrC-93	bipolaricin G	
TrC-94	bipolaricin H	
TrC-95	bipolaricin I	
TrC-96	halorosellinic acid	Anti-tubercular, Antimalarial ¹⁴³
TrC-97	ceroplastic acid	
TrC-98	ceroplastol-I	
TrC-99	ceroplastol-II	
TrC-100	alcoholic acid	
TrC-101	ceroplastodiol	
TrC-102	cheilarinosin	
TrC-103	ceralbic acid I	
TrC-104	ceralbic acid II	
TrC-105	ceralbol	
TrC-106	fusicoccin A	Anti-cancer ¹⁴⁴
TrC-107	bipolarolide F	Anti-microbial ¹⁴⁵
TrC-108	bipolarolide G	
TrC-109	alborosin	
TrC-110	jaspic acid	Anti-inflammatory ¹⁴⁶
TrC-111	(-)-(5 <i>R</i> ,10 <i>R</i>)-subersic acid	Anti-inflammatory ¹⁴⁶
TrC-112	coscinoquinol	Anti-microbial ^{147, 148}
TrC-113	hipposulfate A	Anti-cancer ¹⁴⁹

TrC-114	hipposulfate B	
TrC-115	halisulfate 1	Anti-microbial ¹⁴⁷
TrC-116	(+)-arathanatriene (reported as (-)-caprutriene A by Huang et al. ¹⁵⁰)	
TrC-117	(-)-variculatriene A	
TrC-118	(-)-caprutriene (reported as (+)-arathanatriene in Huang et al. ¹⁵⁰)	
TrC-119	(-)-variculatriene B	
TrC-120	(+)-thalianatriene	
TrC-121	stellatic acid	
TrC-122	variculanol	
TrC-123	5 α -hydroxyfloridenol	
TrC-124	floceric acid	
TrC-125	flocerol	
TrC-126	floridenol	
TrC-127	(E)-halisufate	
TrC-128	(17Z)-halisulfate 1	
TrC-129	N,N-dimethylguanidinium halisulfate 1	
TrC-130	hyrtiosal	Anti-cancer ¹⁵¹ , Anti-viral ¹⁵²
TrC-131	jaspiferal C	
TrC-132	jaspiferal D	
TrC-133	jaspiferal E	
TrC-134	jaspiferal F	
TrC-135	jaspolide F	
TrC-136	conulosin A	
TrC-137	conulosin B	
TrC-138	cheilanthatriol	
TrC-139	granulosane A	
TrC-140	9,11-secoesterol-3 α -hydroxy-5 α ,6 α -epoxy-9-oxo-9,11-seco-5 α -cholest-7-en-11-al	
TrC-141	25-hydroxy-13(24),15,17-cheilanthatrien-19,25-olide	Anti-cancer ¹⁵³
TrC-142	hamiltonin E	
TrC-143	suvanine	Anti-viral ⁹⁰ , Anti-cancer ¹⁵⁴ , Anti-thrombotic ¹⁵⁵
TrC-144	atolypene A	Anti-cancer ¹⁵⁶
TrC-145	atolypene B	
TrC-146	13,16-epoxy-25-hydroxy-17-cheilanththen-19,25-olide	Anti-cancer ¹⁵³
TrC-147	lintonolide A or spongianolide C	
TrC-148	lintonolide B or spongianolide D	
TrC-149	25-hydroxy-13(24),17-cheilanthdien-16,19-olide	Anti-cancer ¹⁵³
TrC-150	16,25-dihydroxy-13(24),17-cheilanthdien-19,25-olide	Anti-cancer ¹⁵³
TrC-151	19-nor-3-hydroxyspongia-3,13(16),14-trien-2-one	
TrC-152	3 β ,19-dihydroxyspongia-13(16),14-dien-2-one (epispongiadiol)	
TrC-153	3 β -hydroxyspongia-13(16),14-dien-2-one	Anti-cancer ^{153, 157}
TrC-154	coscinolactam B	Anti-inflammatory ¹⁵⁸
TrC-155	heliocide B1	
TrC-156	heliocide H1 also reported as heliocide 2 ¹⁵⁹	
TrC-157	heliocide H2 also reported as heliocide 1 ¹⁵⁹	
TrC-158	heliocide H3 also reported as heliocide 4 ¹⁵⁹	
TrC-159	heliocide H4 also reported as heliocide 3 ¹⁵⁹	
TrC-160	terretonin	
TrC-161	terretonin A	
TrC-162	terretonin B	
TrC-163	terretonin C	
TrC-164	terretonin D	
TrC-165	terretonin E	Enzyme Inhibitor (NADH oxidase) ¹⁶⁰
TrC-166	terretonin F	Enzyme Inhibitor (NADH oxidase) ¹⁶⁰
TrC-167	1,2-dihydroterretonin F	
TrC-168	terretonin G	
TrC-169	terretonin H	Embryotoxic ¹⁶¹
TrC-170	terretonin I	Embryotoxic ¹⁶¹
TrC-171	hyatolide A	Anti-cancer ¹⁶²
TrC-172	hyatolide B	
TrC-173	cerorubenic acid-I	
TrC-174	cerorubenic acid-II	
TrC-175	cerorubenol-I	

TrC-176	cerorubenol-II	
TrC-177	lintonene	
TrC-178	petrosaspongiolide M	Anti-inflammatory ^{163, 164} , Anti-cancer ¹⁶⁵⁻¹⁶⁷
TrC-179	petrosaspongiolide N	Anti-inflammatory ¹⁶⁴
TrC-180	petrosaspongiolide P	Anti-inflammatory ¹⁶⁴
TrC-181	21-hydroxy petrosaspongiolide P	
TrC-182	petrosaspongiolide Q	
TrC-183	petrosaspongiolide R	Anti-inflammatory ¹⁶⁴
TrC-184	nitinol	Anti-cancer ¹⁶⁸
TrC-185	(+)-16- <i>O</i> -acetyl-20-formylhyrtiosal	
TrC-186	(+)-20-formylhyrtiosal	
TrC-187	12 β -hydroxy-20,24-dimethyl-13,18-oxa-25-norscalarane	
TrC-188	leucosesterterpenone	Anti-cancer ^{107, 169}
TrC-189	14 β -methylleucosesterterpenone	
TrC-190	leucosceptroid C	Anti-insect ¹⁷⁰ , Immuno-suppressive ¹⁰⁸
TrC-191	leucosceptroid D	Anti-insect ¹⁷⁰ , Immuno-suppressive ¹⁰⁸
TrC-192	ansellone A	Pro-viral ⁶⁷
TrC-193	ansellone B	Anti-inflammatory ¹⁷¹
TrC-194	ansellone B1	
TrC-195	phorbasone A	Anti-feedant ⁴²
TrC-196	phorbasone A acetate	
TrC-197	isophorbasone A	Anti-inflammatory ¹⁷¹
TrC-198	phorbasone B	
TrC-199	phorbadione	
TrC-200	coscinolactam A	
TrC-201	coscinolactam C	Anti-inflammatory ¹⁵⁸
TrC-202	coscinolactam D	Anti-cancer ¹⁷²
TrC-203	coscinolactam E	Anti-cancer ¹⁷²
TrC-204	coscinolactam F	Anti-cancer ¹⁷²
TrC-205	coscinolactam G	Anti-cancer ¹⁷²
TrC-206	ansellone C*	Anti-cancer ¹⁷²
TrC-207	ansellone C*	Anti-cancer ^{112, 173, 174}
TrC-208	ansellone D	
TrC-209	ansellone E	
TrC-210	ansellone F	
TrC-211	ansellone G	
TrC-212	petrosaspongiolide A (reported also as unnamed tricarbocyclic sesterterpene, compound 2 ¹⁷⁵)	
TrC-213	petrosaspongiolide B (reported also as unnamed tricarbocyclic sesterterpene, compound 1 ¹⁷⁵)	Anti-cancer ^{176, 177}
TrC-214	petrosaspongiolide C	Anti-cancer ¹⁷⁶
TrC-215	petrosaspongiolide D	Anti-cancer ¹⁷⁶
TrC-216	petrosaspongiolide E	Anti-cancer ¹⁷⁶
TrC-217	petrosaspongiolide F	Anti-cancer ¹⁷⁶
TrC-218	petrosaspongiolide G	Anti-cancer ¹⁷⁶
TrC-219	petrosaspongiolide H	
TrC-220	petrosaspongiolide I	Anti-cancer ¹⁷⁶
TrC-221	petrosaspongiolide J	Anti-cancer ¹⁷⁶
TrC-222	petrosaspongiolide L	Anti-cancer ¹⁷⁶
TrC-223	salvileucolidone	
TrC-224	betaestacin I (Pb1)	
TrC-225	betaestacin II	
TrC-226	betaestacin III	
TrC-227	betaestacin IV	
TrC-228	betaestacin Va	
TrC-229	betaestacin Vb	
TrC-230	betaestacin Vc	
TrC-231	betaestacin VI	
TrC-232	luteone	
TrC-233	cacospongionolide	

TrC-234	aplysolide A and B (25 α and 25 β)	Anti-cancer ¹⁷⁸
TrC-235	aplyolide	
TrC-236	cheilanthenediol	
TrC-237	cheilanthenetriol	
TrC-238	spongianolide A	
TrC-239	spongianolide B	Anti-cancer ¹⁷⁹
TrC-240	spongianolide E	
TrC-241	spongianolide F	
TrC-242	lintonolide C	
TrC-243	lintonolide D	Anti-cancer ¹⁸⁰
TrC-244	lintonolide E	Anti-cancer ¹⁸⁰
TrC-245	vulgaroside 1 (24,25- <i>O</i> -diacetylvulgaroside)	Anti-cancer ¹⁸⁰
TrC-246	vulgaroside 2	
TrC-247	vulgaroside 3	
TrC-248	vulgaroside 4	
TrC-249	halisulfate 1 methylherbipoline	
TrC-250	caminatal	Enzyme Inhibitor (Serine protease) ¹⁵⁵
TrC-251	cavernosolide	
TrC-252	suvanine methylherbipoline	Antifouling ¹⁸¹
TrC-253	lintonolide F	Anti-cancer ¹⁷²
TrC-254	lintonolide G	Anti-cancer ¹⁸⁰
TrC-255	auroral 1	Anti-cancer ¹⁸⁰
TrC-256	auroral 2	Anti-cancer ¹⁸²
TrC-257	auroral 3	Anti-cancer ¹⁸²
TrC-258	auroral 4	Anti-cancer ¹⁸²
TrC-259	spongidine A	Anti-cancer ¹⁸²
TrC-260	spongidine B	Anti-inflammatory ¹⁸³
TrC-261	spongidine C	Anti-inflammatory ¹⁸³
TrC-262	spongidine D	Anti-inflammatory ¹⁸³
TrC-263	unnamed tricarbocyclic sesterterpene (reported as compound 10)	Anti-inflammatory ¹⁸³
TrC-264	globostelletin C	
TrC-265	globostelletin D	Anti-cancer ¹⁸⁴
TrC-266	globostelletin E	Anti-cancer ¹⁸⁴
TrC-267	globostelletin F	
TrC-268	globostelletin G	Anti-cancer ¹⁸⁴
TrC-269	unnamed tricarbocyclic sesterterpene (reported as compound 24)	Anti-cancer ¹⁸⁴
TrC-270	unnamed tricarbocyclic sesterterpene (reported as compound 25)	
TrC-271	unnamed tricarbocyclic sesterterpene (reported as compound 7)	
TrC-272	unnamed tricarbocyclic sesterterpene (reported as compound 8)	
TrC-273	flabelliferin A	
TrC-274	unnamed tricarbocyclic sesterterpene (reported as compound 1)	
TrC-275	unnamed tricarbocyclic sesterterpene (reported as compound 4)	
TrC-276	cristasesterterpenoic acid	
TrC-277	cristasesterterpinol glucoside	
TrC-278	involudispirone A	
TrC-279	involudispirone B	
TrC-280	luffolide	
TrC-281	inorolide C	Anti-inflammatory ¹⁸⁵
TrC-282	unnamed tricarbocyclic sesterterpene (reported as compound 4)	
TrC-283	25-deoxycacospongionolide	
TrC-284	unnamed tricarbocyclic sesterterpene (reported as compound 3)	
TrC-285	unnamed tricarbocyclic sesterterpene (reported as compound 4)	
TrC-286	gentianelloid A	
TrC-287	gentianelloid B	Immunosuppressive ¹⁸⁶
TrC-288	2'-isopicrasin A	Immunosuppressive ¹⁸⁶
TrC-289	picrasin A	
TrC-290	unnamed tricarbocyclic sesterterpene (reported as compound 2)	
TrC-291	unnamed tricarbocyclic sesterterpene (reported as compound 3)	
TrC-292	(2 <i>E</i>)-flocerene	
TrC-293	sesterbrasiliatriene	
TrC-294	18- <i>epi</i> -alborosin	
TrC-295	(-)-caprutriene B	
TrC-296	(+)-caprutriene C	
TrC-297	ophiobolane 2 (3,20-anhydrophiobolin F)	

TrC-298	ophiobolane 1 (ophiobola-1,7,18-triene)	
TrC-299	stellata-2,6,19-triene	
TrC-300	(17Z)-cheilantha-13(24),17-diene-6a,19-diol	
TrC-301	(17Z)-cheilantha-13(24),17-diene-1b,6a,19-triol	
TrC-302	(17Z)-19-acetoxycheilantha-13(24),17-diene-1b,6a-diol	
TrC-303	17-oxo-18,19-bisnorcheilanth-13(24)-en-6a-ol	
TrC-304	13,17-dioxo-18,19,24-trisnorcheilanth-6a-ol	
TrC-305	(-)-variculartriene A	
TrC-306	maydispenoid A	Immunosuppressive ¹⁸⁷
TrC-307	maydispenoid B	Immunosuppressive ¹⁸⁷
TrC-308	Bm1	
TrC-309	18,19-dihydro-19-hydroxy-18-methoxyophiobolin P	
TrC-310	18,19-dihydro-19-hydroxy-18- <i>epi</i> -methoxyophiobolin P	
TrC-311	spectanoid A	Anti-cancer ¹⁸⁸
TrC-312	spectanoid B	
TrC-313	spectanoid C	Anti-cancer ¹⁸⁸
TrC-314	spectanoid D	
TrC-315	spectanoid E	
TrC-316	spectanoid F	Anti-cancer ¹⁸⁸
TrC-317	spectanoid G	
TrC-318	linderasesterterpenoid A	Anti-inflammatory ¹⁸⁹
TrC-319	linderasesterterpenoid B	Anti-inflammatory ¹⁸⁹
TrC-320	ancepsone A	Anti-cancer ¹⁸⁸
TrC-321	schultriene	
TrC-322	ophiobolin P1	
TrC-323	ansellone H	
TrC-324	ansellone I	
TrC-325	ansellone J	Anti-viral ¹⁹⁰
TrC-326	ansellone K	
TrC-327	cyophiobolin A	Anti-inflammatory ¹⁹¹
TrC-328	cyophiobolin B	Anti-inflammatory ¹⁹¹
TrC-329	cyophiobolin C	
TrC-330	cyophiobolin D	
TrC-331	sesterviolene B	
TrC-332	nigtetraene	
TrC-333	colquhounoid A	
TrC-334	colquhounoid D	
TrC-335	14- <i>epi</i> -colquhounoid D	
TrC-336	orientanoid A	Anti-cancer ¹⁹²
TrC-337	orientanoid B	
TrC-338	orientanoid C	
TrC-339	secosuberitenone A	
TrC-340	patriniaterpene A	Anti-inflammatory ¹⁹³
TrC-341	patriniaterpene B	Anti-inflammatory ¹⁹³
TrC-342	sarcotragusolide A	Anti-cancer ¹⁹⁴
TrC-343	sarcotragusolide B	Anti-cancer ¹⁹⁴
TrC-344	sarcotragusolide C	
TrC-345	sarcotragusolide D	Anti-cancer ¹⁹⁴
TrC-346	bipolaricin J	
TrC-347	bipolaricin K	Anti-cancer ¹⁹⁵
TrC-348	bipolaricin L	
TrC-349	bipolaricin M	Anti-cancer ¹⁹⁵ , Phytotoxic ¹⁹⁵
TrC-350	bipolaricin N	Phytotoxic ¹⁹⁵
TrC-351	bipolaricin O	Phytotoxic ¹⁹⁵
TrC-352	bipolaricin P	Anti-cancer ¹⁹⁵ , Phytotoxic ¹⁹⁵
TrC-353	bipolaricin Q	Anti-cancer ¹⁹⁵ , Phytotoxic ¹⁹⁵
TrC-354	bipolaricin R	Anti-cancer ¹⁹⁵ , Phytotoxic ¹⁹⁵
TrC-355	bipolaricin S	Phytotoxic ¹⁹⁵
TrC-356	21-deoxo-21-hydroxyophiobolin U	
TrC-357	undobolin A	
TrC-358	undobolin B	
TrC-359	undobolin C	
TrC-360	undobolin D	Immuno-suppressive ¹⁹⁶
TrC-361	undobolin E	

TrC-362	undobolin F	Immuno-suppressive ¹⁹⁶
TrC-363	undobolin G	
TrC-364	undobolin H	Anti-inflammatory ¹⁹⁶
TrC-365	undobolin I	
TrC-366	undobolin J	
TrC-367	undobolin K	
TrC-368	undobolin L	Immuno-suppressive ¹⁹⁶
TrC-369	bipoladien C	
TrC-370	bipoladien D	Anti-microbial ¹⁹⁷
TrC-371	bipoladien E	Anti-microbial ¹⁹⁷
TrC-372	unnamed ophiobolin analog	
TrC-373	unnamed ophiobolin analog	Anti-cancer ¹⁹⁸
TrC-374	unnamed ophiobolin analog	Anti-cancer ¹⁹⁸

*The same name was used for two different compounds

Section 8. Biological activities of tetracarboxylic (TeC) sesterterpenoids (with and without hetherocycles) and their dimers

c. n.	IUPAC name or Trivial name	Bioactivity (reference)
TeC-1	bipolarolide A	Enzyme Inhibitor (HMG-CoA reductase) ¹⁴⁵
TeC-2	bipolarolide B	
TeC-3	bipolarolide C	
TeC-4	bipolarolide D	
TeC-5	variecolin	Immunosuppressive ¹⁹⁹
TeC-6	variecolol	Immunosuppressive ¹⁹⁹
TeC-7	emericolin A	
TeC-8	emericolin B	
TeC-9	emericolin C	
TeC-10	emericolin D	
TeC-11	variecolactone	Immunosuppressive ²⁰⁰ , Anti-cancer ²⁰¹
TeC-12	variecoacetal A	Immunosuppressive ²⁰⁰
TeC-13	variecoacetal B	Immunosuppressive ²⁰⁰
TeC-14	YW3699	
TeC-15	aleurodiscal	Anti-fungal ²⁰²
TeC-16	astellifadiene	
TeC-17	(-)-aleurodiscalene A	
TeC-18	(+)-aleurodiscalene B	
TeC-19	emericellic acid	
TeC-20	sesterfisherol	
TeC-21	sesterfisheric acid	
TeC-22	aspergilloxide	
TeC-23	suberitenone A	Anti-cancer ²⁰³ , Anti-RSV (Respiratory syncytial virus) ²⁰⁴
TeC-24	suberitenone B	Anti-cancer ²⁰³ , Anti-RSV (Respiratory syncytial virus) ²⁰⁴
TeC-25	isosuberitenone B	Anti-cancer ²⁰⁵
TeC-26	19- <i>epi</i> -suberitenone	Anti-cancer ²⁰⁵
TeC-27	suberitenone C	
TeC-28	suberitenone D	
TeC-29	suberiphenol	
TeC-30	oxaspirosuberitenone	Anti-cancer ²⁰⁵
TeC-31	flabelliferin B	
TeC-32	phellogine	
TeC-33	foliaspongine	
TeC-34	perisomalien A	Anti-cancer ²⁰⁶
TeC-35	lendenfeldarane A	
TeC-36	lendenfeldarane B	
TeC-37	carteriofenone A	
TeC-38	carteriofenone B	
TeC-39	carteriofenone C	
TeC-40	carteriofenone D	Anti-cancer ²⁰⁷
TeC-41	unnamed tetracarboxylic sesterterpene (reported as compound 3)	

TeC-42	unnamed tetracarbocyclic sesterterpene (reported as compound 5)	
TeC-43	unnamed tetracarbocyclic sesterterpene (reported as compound 6)	
TeC-44	unnamed tetracarbocyclic sesterterpene (reported as compound 7)	
TeC-45	dibritannilactone A	Anti-inflammatory ²⁰⁸
TeC-46	12-deacetyl-12-oxoscalaradial	Ichthyotoxic ²⁰⁹
TeC-47	12-deacetyl-12,18-di- <i>epi</i> -scalaradial	Anti-cancer ²¹⁰
TeC-48	hyrtial	Anti-inflammatory ²¹¹
TeC-49	12-deacetylhyrtial	Anti-cancer ²¹²
TeC-50	12-deacetyl- Δ^{17} -hyrtial	Anti-cancer ²¹²
TeC-51	12-deacetoxy-23-hydroxyscalaradial	Anti-cancer ²¹³
TeC-52	5-deoxovaricocolin	Anti-cancer ²¹⁴
TeC-53	12-deacetyl-18- <i>epi</i> -12-oxoscalaradial	
TeC-54	unnamed tetracarbocyclic sesterterpene (reported as compound 1)	Anti-cancer ²¹⁵
TeC-55	unnamed tetracarbocyclic sesterterpene (reported as compound 1a)	Anti-cancer ²¹⁵
TeC-56	unnamed tetracarbocyclic sesterterpene (reported as compound 2)	Anti-cancer ²¹⁵
TeC-57	12 α -(3'-acetobutanoyloxy)-20,24-dimethyl-24-oxoscalar-16-en-25-al	Anti-cancer ²¹⁵
TeC-58	12 α -(3'-acetoxypentanoyloxy)-20,24-dimethyl-24-oxoscalar-16-en-25-al	Anti-cancer ²¹⁵
TeC-59	12 α -acetoxy-16 α -(3'-hydroxypentanoyloxy)-20,24-dimethyl-24-oxoscalar-25 β -oic acid	
TeC-60	methyl 18-hydroxy-19-norscalar-16-en-20-carboxylate	Anti-microbial ²¹⁶
TeC-61	12 β -(3'-hydroxypentanoyloxy)-20,24-dimethyl-24-oxo-scalar-16-en-25-al	Anti-cancer ²¹⁷
TeC-62	unnamed tetracarbocyclic sesterterpene (reported as compound 1)	Anti-cancer ²¹⁸
TeC-63	unnamed tetracarbocyclic sesterterpene (reported as compound 2)	
TeC-64	petrosaspongiolide K	Anti-cancer ¹⁷⁶
TeC-65	21-hydroxy petrosaspongiolide K	Anti-inflammatory ¹⁸³
TeC-66	mooloolabene A	Anti-cancer ²¹⁹
TeC-67	mooloolabene B	Anti-cancer ²¹⁹
TeC-68	mooloolabene C	Anti-cancer ²¹⁹
TeC-69	mooloolabene D	Anti-cancer ²¹⁹
TeC-70	mooloolabene E	Anti-cancer ²¹⁹
TeC-71	mooloolaldehyde	Anti-cancer ²¹⁹
TeC-72	12 α -acetoxy-16 β -hydroxy-20,24-dimethyl-24-oxo-25-norscalarane	Anti-cancer ²²⁰
TeC-73	12 α ,16 α -diacetoxy-20,24-dimethyl-25-norsclaran-24-one	
TeC-74	(12 α ,16 β)-12-acetoxy-16-hydroxy-20,24-dimethyl-25-norscalar-17-en-24-one	Anti-cancer ²²⁰
TeC-75	scalaradial	Anti-inflammatory ²²¹ , Anti-cancer ¹⁷⁸ , Enzyme Inhibitor (Transient receptor potential cation channel, subfamily M, member 2) ²²²
TeC-76	desacetylscalaradial (deacetylscalaradial)	
TeC-77	12-deacetoxyscalaradial (scalarenedial)	Anti-cancer ²¹³
TeC-78	12- <i>epi</i> -scalaradial	Anti-inflammatory ²²³ , Anti-microbial ²¹⁶
TeC-79	12-deacetyl-12- <i>epi</i> -scalaradial	Anti-cancer ²²⁴
TeC-80	19-dihydroscalaradial	
TeC-81	18- <i>epi</i> -scalaradial	Anti-cancer ²²⁵
TeC-82	12,18-di- <i>epi</i> -scalaradial	Anti-microbial ²¹⁶
TeC-83	12-deacetoxy-12-oxo-scalaradial	
TeC-84	12-deacetyl-23-acetoxy-20-methyl-12- <i>epi</i> -scalaradial	Anti-inflammatory ²²⁶
TeC-85	12 α -acetoxy-22-hydroxy-24-methyl-24-oxoscalar-16-en-25-al or PHC-4	Anti-cancer ²²⁷
TeC-86	12 α -acetoxy-24-methyl-24-oxoscalar-16-en-22,25-dial or PHC-2	Anti-cancer ²²⁸
TeC-87	24-methyl-12,24,25-trioxoscalar-16-en-22-oic acid or PHC-1	Anti-inflammatory ²²⁹
TeC-88	12 α -hydroxy-24-methyl-24,25-dioxoscalar-16-en-22-oic acid	
TeC-89	12 α -acetoxy-20,24-dimethyl-25-norscalar-16-en-24-one	Anti-cancer ²³⁰
TeC-90	unnamed tetracarbocyclic sesterterpene (reported as compound 6)	Anti-cancer ²¹⁵
TeC-91	unnamed tetracarbocyclic sesterterpene (reported as compound 7)	Anti-cancer ²¹⁵
TeC-92	25-nor-24-methyl-12,24-dioxoscalar-16-en-22-oic acid	
TeC-93	phylophenone E	
TeC-94	12 α -acetoxy-20,24-dimethyl-24-oxoscalar-16-en-25 β -oic acid	
TeC-95	22-hydroxy-24-methyl-12,24-dioxoscalar-16-en-25-al or PCH-3	Anti-cancer ²³¹

TeC-96	25-nor-12 α -acetoxy-20,24-dimethyl-24-oxoscalar-16-en-18b-ol	
TeC-97	asperterpenol A	Enzyme Inhibitor (Acetylcholinesterase) ²³²
TeC-98	asperterpenol B	Enzyme Inhibitor (Acetylcholinesterase) ²³²
TeC-99	24-methyl-24,25-dioxoscalar-16-en-12b-yl-3-hydroxybutanoate	Anti-inflammatory ²³³
TeC-100	felixin F	
TeC-101	felixin G	
TeC-102	12 α -acetoxy-20,24-dimethyl-16,24-dioxoscalar-14,17-dien-25-oic acid	
TeC-103	24-methyl-12,24,25-trioxoscalar-16-en-22-oate	Anti-cancer ²³¹
TeC-104	12 α -acetoxy-16b-hydroxy-24-methyl-24-oxoscalarane-25-al	Anti-cancer ³³
TeC-105	12 β -(3' β -hydroxybutanoyloxy)-20,24-dimethyl-24-oxo-scalar-16-en-25-al	Anti-cancer ²¹⁷
TeC-106	12- <i>O</i> -deacetyl-12- <i>epi</i> -19- <i>O</i> -methylscalarin	Anti-cancer ²³⁴
TeC-107	12- <i>O</i> -deacetyl-12,19-di- <i>epi</i> -scalarin	
TeC-108	scalarinol	Anti-cancer ²³⁵
TeC-109	12,24-diacetoxy-deoxoscalarin	Anti-hypercholesterolemic ²³⁶
TeC-110	12- <i>O</i> -deacetoxyl-24-hydroxyl-deoxoscalarin	Anti-hypercholesterolemic ²³⁶
TeC-111	12- <i>O</i> -deacetoxyl-19- <i>O</i> -methydeoxoscalarin	Anti-hypercholesterolemic ²³⁶
TeC-112	12 α - <i>O</i> -acetylhyrtiolide	
TeC-113	12 α -acetoxy-20,24 β -dimethylscalar-17-eno-25,24-lactone	
TeC-114	sesterstatin 7	Anti-microbial ²³⁷
TeC-115	unnamed tetracarbocyclic sesterterpene (reported as compound 1)	Anti-microbial ²³⁸
TeC-116	12- <i>O</i> -deacetyl-12- <i>epi</i> -19-deoxy-21-hydroxyscalarin	
TeC-117	unnamed tetracarbocyclic sesterterpene (reported as compound 2)	Anti-microbial ²³⁸
TeC-118	12 β ,16 β ,22-trihydroxy-24-methyl-24-oxoscalar-25-oxo methyl ester	
TeC-119	12 α -acetoxy-20,24-dimethyl-16,24-dioxoscalar-14,17-dien-24-ol-25,24-olide	
TeC-120	scalarester	
TeC-121	12- <i>epi</i> -19-deoxyscalarin	Anti-cancer ²³⁹
TeC-122	unnamed tetracarbocyclic sesterterpene (reported as compound 7)	
TeC-123	salmahyrtisol C	
TeC-124	unnamed tetracarbocyclic sesterterpene (reported as compound 8)	
TeC-125	24-acetoxy-12-deacetyl-12- <i>epi</i> -deoxoscalarin	
TeC-126	petrosapsongiolactam A	Enzyme Inhibitor (TDP-43) ²⁴⁰
TeC-127	petrosapsongiolactam B	
TeC-128	petrosapsongiolactam C	
TeC-129	unnamed tetracarbocyclic sesterterpene (reported as compound 2a and as compound 6)	
TeC-130	unnamed tetracarbocyclic sesterterpene (reported as compound 2b)	
TeC-131	unnamed tetracarbocyclic sesterterpene (reported as compound 3)	
TeC-132	unnamed tetracarbocyclic sesterterpene (reported as compound 9)	
TeC-133	unnamed tetracarbocyclic sesterterpene (reported as compound 3)	
TeC-134	unnamed tetracarbocyclic sesterterpene (reported as compound 3a)	
TeC-135	unnamed tetracarbocyclic sesterterpene (reported as compound 3b)	
TeC-136	unnamed tetracarbocyclic sesterterpene (reported as compound 3c)	
TeC-137	unnamed tetracarbocyclic sesterterpene (reported as compound 4)	
TeC-138	unnamed tetracarbocyclic sesterterpene (reported as compound 4b)	
TeC-139	unnamed tetracarbocyclic sesterterpene (reported as compound 4c)	
TeC-140	unnamed tetracarbocyclic sesterterpene (reported as compound 5)	
TeC-141	12-deacetoxy-12-oxo-deoxoscalarin	
TeC-142	12-deacetoxy-23-acetoxy-19- <i>O</i> -acetylscalarin	Anti-cancer ²¹³
TeC-143	unnamed tetracarbocyclic sesterterpene (reported as compound 2)	Anti-cancer ³³
TeC-144	unnamed tetracarbocyclic sesterterpene (reported as compound 3)	
TeC-145	unnamed tetracarbocyclic sesterterpene (reported as compound 4)	
TeC-146	unnamed tetracarbocyclic sesterterpene (reported as compound 5)	
TeC-147	unnamed tetracarbocyclic sesterterpene (reported as compound 6)	Anti-cancer ³³
TeC-148	unnamed tetracarbocyclic sesterterpene (reported as compound 7)	
TeC-149	unnamed tetracarbocyclic sesterterpene (reported as compound 8)	
TeC-150	12 α -acetoxy-24,25-epoxy-24-hydroxy-20,24-dimethylscalarane	Anti-cancer ²³⁰
TeC-151	scaldysin-A	Anti-microbial ²³⁰
TeC-152	scaldysin-B	
TeC-153	12-acetyl-19,20-dimethoxy-deoxoscalarin	
TeC-154	deoxo-scalarin acetate	
TeC-155	12- <i>epi</i> -deacetyl-19 α -acetoxy-20 α -methoxyscalarin	

TeC-156	scalarin	
TeC-157	scalarinether	
TeC-158	19-deoxyscalarin	
TeC-159	12- <i>O</i> -deacetyl-12- <i>epi</i> -scalarin	Anti-cancer ²⁴¹
TeC-160	12-deacetyl-12- <i>epi</i> -19-deoxyscalarin	
TeC-161	12- <i>O</i> -deacetyl-12- <i>epi</i> -19-deoxy-22-hydroxyscalarin	
TeC-162	16-acetoxy-dihydrodeoxoscalarin	
TeC-163	deoxoscalarin	Anti-cancer ²⁴²
TeC-164	6-keto-deoxoscalarin	
TeC-165	felixin A	Anti-cancer ²⁴³
TeC-166	felixin B	Anti-cancer ²⁴³
TeC-167	felixin C	Anti-cancer ²⁴³
TeC-168	felixin D	Anti-cancer ²⁴³
TeC-169	felixin E	Anti-cancer ²⁴³
TeC-170	12-deacetoxy-23- <i>O</i> -acetoxyheteronemin	Anti-cancer ²¹³
TeC-171	citreoahybidone A	
TeC-172	hippospongide B	
TeC-173	scalarolide	Anti-cancer ²⁴⁴
TeC-174	12- <i>epi</i> -12- <i>O</i> -acetylscalarolide	
TeC-175	scalarolide acetate	Anti-cancer ²⁴⁴
TeC-176	12-deacetoxyscalarin 19-acetate	Anti-cancer ²⁴⁵
TeC-177	12-deacetoxy-21-acetoxyscalarin	Anti-cancer ²⁴⁶
TeC-178	12-deacetoxy-22-acetoxyscalarin	Anti-cancer ¹⁶
TeC-179	12- <i>O</i> -deacetyl-25- <i>epi</i> -scalarin	
TeC-180	12- <i>epi</i> -19- <i>O</i> -methylscalarin	
TeC-181	12-deacetoxy-22,25- diacetoxy-scalarin	Anti-cancer ¹⁶
TeC-182	12 α -acetoxy-19 β -hydroxyscalara-15,17-dien-20,19-olide	
TeC-183	12-deacetoxy-22-hydroxy-25-acetoxyscalarin	Anti-cancer ²⁴⁷
TeC-184	hyrtiolide	Anti-cancer ²⁴⁸
TeC-185	12-dehydroxy-23-hydroxyhyrtiolide	Anti-cancer ²¹³
TeC-186	12-acetoxy,16- <i>epi</i> -hyrtiolide	Anti-cancer ²⁴⁹ , Anti-microbial ²³⁹
TeC-187	12- <i>O</i> -acetyl-16- <i>O</i> -methylhyrtiolide	Anti-cancer ²⁴¹
TeC-188	phyllolactone A	
TeC-189	phyllolactone B	
TeC-190	phyllolactone C	
TeC-191	phyllolactone D	
TeC-192	phyllolactone E	
TeC-193	unnamed tetracarbocyclic sesterterpene (reported as compound 2)	
TeC-194	similan A	Anti-cancer ²⁴⁸
TeC-195	20-hydroxyscalarolide	Anti-microbial ²¹⁶
TeC-196	16-hydroxyscalarolide	
TeC-197	12 β 16 α ,20-trihydroxy-17-scalaren-19,20-olide	Anti-cancer ²⁴⁸
TeC-198	(12 α ,245)-12-[(3-hydroxypentanoyl)oxy]-20,24-dimethyl-25-oxoscalar-15,17-dien-25,24-olide	
TeC-199	19-acetylsesterstatin 3	Anti-microbial ²³⁹
TeC-200	3-acetylsesterstatin 1	
TeC-201	12 β ,20-dihydroxy-16 β -acetoxy-17-scalaren-19,20-olide	
TeC-202	12 β -acetoxy,16 β -methoxy,20 α -hydroxy-17-scalaren-19,20-olide	
TeC-203	12 α ,24-dihydroxy-20,24-dimethyl-15,17-scalaradien-25,24-olide	
TeC-204	12,16-dihydroxy-24-methylscalaran-25,24-olide	Anti-cancer ²⁵⁰
TeC-205	phyllolactone L	
TeC-206	phyllolactone B	
TeC-207	phyllolactone C	
TeC-208	phyllolactone D	
TeC-209	phyllolactone H	
TeC-210	phyllolactone I	
TeC-211	phyllolactone J	
TeC-212	phyllolactone K	
TeC-213	phyllolactone M	
TeC-214	12- <i>epi</i> -phyllolactone-B	
TeC-215	12- <i>epi</i> -phyllactone D	
TeC-216	12- <i>epi</i> -phyllactone E	
TeC-217	phyllactone F	Anti-cancer ²⁵¹
TeC-218	phyllactone G	Anti-cancer ²⁵¹

TeC-219	phyllactone H	
TeC-220	lendenfeldarane C	Anti-cancer ²⁵²
TeC-221	lendenfeldarane D	
TeC-222	honulactone C	Anti-cancer ²⁵³
TeC-223	honulactone D	Anti-cancer ²⁵³
TeC-224	honulactone I	
TeC-225	honulactone L	
TeC-226	unnamed tetracarbocyclic sesterterpene (reported as compound 5)	
TeC-227	unnamed tetracarbocyclic sesterterpene (reported as compound 6)	
TeC-228	unnamed tetracarbocyclic sesterterpene (reported as compound 13)	
TeC-229	unnamed tetracarbocyclic sesterterpene (reported as compound 14)	
TeC-230	(12 α ,24R)-12-[(3-hydroxypentanoyl) oxy]-20,24-dimethyl-25-oxoscalar-15,17-dien-25,24-olide	
TeC-231	unnamed tetracarbocyclic sesterterpene (reported as compound 3)	
TeC-232	24 β -methoxyscalarolide	
TeC-233	unnamed tetracarbocyclic sesterterpene (reported as compound 10)	Anti-microbial ²³⁴
TeC-234	22-acetoxy-12 β ,16 β -dihydroxy-24-methylsclaran-25,24-olide (reported also as homosclalaralactone IIB) ²⁵⁴	
TeC-235	12 β ,16 β -dihydroxy-24-methylsclaran-25,24-olide	
TeC-236	12 β ,16 β ,22-trihydroxy-24-methylsclaran-25 β ,24 β -olide	
TeC-237	12 β ,16 β ,22- trihydroxy-24 α -methylsclalar-25 β ,24 α -olide	
TeC-238	12 α -acetoxy-23,25-cyclo-16 β ,25Z-dihydroxy-20,24-dimethyl-24-oxosclaranane	
TeC-239	hyattellactone A	Enzyme Inhibitor (Protein tyrosine phosphatase 1B) ²⁵⁵
TeC-240	hyattellactone B	
TeC-241	phyllactone A	Anti-cancer ²⁵⁶
TeC-242	phyllactone B	Anti-cancer ²⁵⁶
TeC-243	phyllactone C	
TeC-244	phyllactone D	
TeC-245	phyllactone E	
TeC-246	phyllofolactone A	
TeC-247	carteriofenone I	Anti-cancer ²⁰⁷
TeC-248	mooloolabene F	
TeC-249	mooloolabene G	
TeC-250	mooloolabene H	
TeC-251	mooloolabene I	
TeC-252	mooloolabene J	
TeC-253	mooloolabene K	
TeC-254	mooloolabene L	
TeC-255	mooloolabene M	
TeC-256	mooloolabene N	
TeC-257	mooloolabene O	
TeC-258	carteriofenone K	Anti-cancer ²⁰⁷
TeC-259	12 β -acetoxy-20- hydroxy-17-sclaren-19,20-olide	
TeC-260	unnamed tetracarbocyclic sesterterpene (reported as compound 3)	Enzyme Inhibitor (RCE-Protease) ²¹⁸
TeC-261	unnamed tetracarbocyclic sesterterpene (reported as compound 4)	Enzyme Inhibitor (RCE-Protease) ²¹⁸
TeC-262	unnamed tetracarbocyclic sesterterpene (reported as compound 4)	
TeC-263	12 β ,16 β -diacetoxysclarolbutenolide	
TeC-264	12 α ,16 β -diacetoxysclarolbutenolide	
TeC-265	12 α -acetoxy-16 β -hydroxysclarolbutenolide	
TeC-266	12-deacetoxy-12-oxo-deoxysclarin	
TeC-267	sclarolbutenolide	
TeC-268	12- <i>O</i> -acetyl-16- <i>O</i> -deacetyl-16- <i>epi</i> -sclarolbutenolide	Anti-cancer ²⁴⁶
TeC-269	16- <i>O</i> -deacetyl-16- <i>epi</i> -sclarolbutenolide	Anti-cancer ²⁴⁶
TeC-270	12-deacetyl-sclaradial	Anti-cancer ²⁵³
TeC-271	norsclalaral A	Anti-cancer ²²⁵
TeC-272	norsclalaral B	Anti-cancer ²²⁵
TeC-273	norsclalaral C	Anti-cancer ²²⁵
TeC-274	12-deacetyl-18- <i>epi</i> -12-oxo-sclaradial	
TeC-275	sclarherbacin-A	
TeC-276	sclarherbacin-A acetate	
TeC-277	sclarherbacin-B	
TeC-278	sclarherbacin-B acetate	

TeC-279	unnamed tetracarbocyclic sesterterpene (reported as compound 8)	Anti-cancer ²¹⁵
TeC-280	12- <i>epi</i> -deoxoscalarin	Anti-microbial ²¹⁶
TeC-281	12- <i>epi</i> -scalarin	Anti-cancer ^{241, 257} , Anti-microbial ²³⁴
TeC-282	12-deacetyl-20-methyl-12- <i>epi</i> -deoxoscalarin	
TeC-283	12-deacetyl-23-acetoxy-20-methyl-12- <i>epi</i> -deoxoscalarin	
TeC-284	12-deacetoxo-23-acetoxyscalarin	Anti-cancer ¹⁶
TeC-285	3-hydroxy-20,22-dimethyldeoxoscalarin	
TeC-286	3-keto-20,22-dimethyl-20-deoxoscalarin	
TeC-287	phylofenone C	
TeC-288	12- <i>O</i> -deacetyl-19- <i>epi</i> -scalarin	
TeC-289	12- <i>O</i> -deacetylnorscalaral B	Anti-cancer ¹⁶²
TeC-290	12-deacetoxo-23-hydroxyheteronemin	Anti-cancer ²¹³
TeC-291	unnamed tetracarbocyclic sesterterpene (reported as compound 4)	
TeC-292	16 β -acetoxy-20,24-dimethyl-12,24-dioxo-25-norscalarane	
TeC-293	16 β -hydroxy-24-methyl-12,24-dioxoscalar-25-al	Anti-cancer ³³
TeC-294	2 α -deoxoscalarin	
TeC-295	19-acetyl-12-deacetoxo-12-oxo-deoxoscalarin	
TeC-296	sesterstatin 1	Anti-cancer ²⁵⁸
TeC-297	sesterstatin 2	Anti-cancer, Anti-microbial ²⁵⁸
TeC-298	sesterstatin 3	Anti-cancer ²⁵⁸
TeC-299	hyatolide C	
TeC-300	hyatolide D	
TeC-301	hyatolide E	
TeC-302	phyllolactone F	Enzyme inhibitor (Protein tyrosine phosphatase 1B) ²⁵⁵
TeC-303	phyllolactone G	
TeC-304	25-dehydroxy-12- <i>epi</i> -scalarin	Anti-cancer ²⁴⁵
TeC-305	hyrtiosin A	
TeC-306	hyrtiosin B	Anti-fungal ²⁵⁹
TeC-307	hyrtiosin C	
TeC-308	hyrtiosin D	
TeC-309	hyrtiosin E	Anti-cancer ²⁶⁰
TeC-310	sesterstatin 6	Anti-cancer ²⁶¹
TeC-311	12 β ,22,24 ϵ -trihydroxy-24-methylscalar-17-ene-18,24-carbolactone	Anti-cancer ²²⁷
TeC-312	12- <i>O</i> -acetyl-12,16- <i>epi</i> -scalarolbutanolide	Anti-cancer ²⁶²
TeC-313	16- <i>O</i> -deacetyl-12,16- <i>epi</i> -scalarolbutanolide	
TeC-314	unnamed tetracarbocyclic sesterterpene (reported as compound 6)	Anti-cancer ²⁶²
TeC-315	unnamed tetracarbocyclic sesterterpene (reported as compound 7)	Anti-cancer ²⁶²
TeC-316	unnamed tetracarbocyclic sesterterpene (reported as compound 8)	Anti-cancer ²⁶²
TeC-317	12 α ,16 β -diacetoxo-20,24-dimethyl-24-oxoscalar-25-al	
TeC-318	12 β ,22-dihydroxy-24-methyl-24-oxoscalar-16-en-25 β -al	Anti-cancer ²²⁷
TeC-319	12 α -acetoxo-16 β -3' <i>R</i> -hydroxybutanoyloxy)-20,24-dimethyl-24-oxoscalar-25-al	Anti-cancer ²⁶³
TeC-320	12 α -acetoxo-16 β -(3' <i>R</i> -hydroxypentanoyloxy)-20,24-dimethyl-24-oxoscalar-25-al	Anti-cancer ²⁶³ , Anti-microbial ²⁶³
TeC-321	12 α -acetoxo-16 β -hydroxy-20,24-dimethyl-24-oxoscalar-25-al	Anti-cancer ²⁶³
TeC-322	12 α ,22-dihydroxy-24-methyl-24-oxoscalar-16-en-25 α -al	Anti-cancer ²⁶³
TeC-323	12 α -acetoxo-16 β -propanoyloxy-20,24-dimethyl-24-oxoscalar-25-al	Anti-cancer ²⁶³
TeC-324	12 α -(3' <i>R</i> -hydroxypentanoyloxy)-20,24-dimethyl-24-oxoscalar-16-en-25-al	Anti-cancer ²⁶³
TeC-325	12 α -(3'-propanoyloxy-pentanoyloxy)-20,24-dimethyl-24-oxoscalar-16-en-25-al	Anti-cancer ²⁶³
TeC-326	heteronemin	Anti-cancer, Anti-microbial ^{245, 264-266} , Enzyme Inhibitor (TDP-43) ^{267, 268}
TeC-327	12- <i>epi</i> -heteronemin	
TeC-328	12- <i>epi</i> -heteronemin acetate	
TeC-329	furoscalarol	
TeC-330	12,16-di- <i>epi</i> -12- <i>O</i> -deacetyl-16- <i>O</i> -acetylfuroscalarol	Anti-cancer ²⁶⁹
TeC-331	scalarafuran	
TeC-332	isoscalarafuran-A	
TeC-333	16-acetylfuroscalarol	Anti-cancer ²⁶⁹
TeC-334	isoscalarafuran-B	
TeC-335	sesterstatin 4 (16- <i>epi</i> -deacetyl-scalarafuran)	Anti-cancer ²⁷⁰
TeC-336	sesterstatin 5	Anti-cancer ²⁷⁰

TeC-337	nambiscalarane	
TeC-338	16-deacetoxy-12- <i>epi</i> -scalarafuranacetate	
TeC-339	12- <i>O</i> -acetyl-16-deacetoxy-23-acetoxyscalarafuran	Anti-cancer ²¹³
TeC-340	12- <i>O</i> -deacetylscalarafuran	Anti-cancer ²⁴¹
TeC-341	21-hydroxy-16-deacetyl-12- <i>epi</i> -scalarafuran acetate	
TeC-342	salmahyrtisol B*	Anti-cancer ²⁷¹
TeC-343	unnamed tetracarbocyclic sesterterpene (reported as compound 11)	Anti-microbial ²³⁴
TeC-344	hyatelactam	Anti-cancer ¹⁶²
TeC-345	sesterstamide	
TeC-346	scalalactam A	
TeC-347	scalalactam B	
TeC-348	24-methoxypetrosaspongia C	Anti-cancer, Anti-proliferative ²⁴⁹
TeC-349	heteronemin acetate	Anti-cancer ²⁴⁵
TeC-350	16 β ,22-dihydroxy-24-methyl-24-oxoscalaran-25,12 β -olactone (also reported as 16 β ,22-dihydroxy-24-methyl-24-oxoscalaran-25,12 β -olide ²⁷² and as homoscalaralactone IIA ²⁵⁴)	Anti-cancer ²²⁷
TeC-351	22-acetoxy-16 β -hydroxy-24-methyl-24-oxoscalarano-25,12 β -lactone	Anti-cancer ²⁷³ , Anti-microbial ²⁷⁴
TeC-352	nitidasin	Immunosuppressive ²⁷⁵
TeC-353	asperunguisin F	
TeC-354	phyllospongin A	Anti-cancer, Anti-microbial ²³⁰
TeC-355	phyllospongin B	Anti-cancer, Anti-microbial ²³⁰
TeC-356	phyllospongin C	Anti-cancer, Anti-microbial ²³⁰
TeC-357	unnamed tetracarbocyclic sesterterpene (reported as compound 4)	Anti-cancer ²⁷⁶
TeC-358	hyatellone C	
TeC-359	cybastacine A	Anti-microbial ²⁷⁷
TeC-360	cybastacine B	Anti-microbial ²⁷⁷
TeC-361	molliorin-a	
TeC-362	molliorin-b	
TeC-363	molliorin-c	
TeC-364	molliorin-e	
TeC-365	sednolide	Anti-microbial ²⁷⁸
TeC-366	scytoscalarol	Anti-microbial ²⁷⁹
TeC-367	cerorubenic acid-III	
TeC-368	inorolide A	
TeC-369	inorolide B	
TeC-370	neomangicol A	Anti-cancer ²⁸⁰
TeC-371	neomangicol B	Anti-cancer ²⁸⁰
TeC-372	neomangicol C	
TeC-373	salmahyrtisol A	Anti-cancer ²⁷¹
TeC-374	mangicol A	Anti-inflammatory ²⁸¹
TeC-375	mangicol B	Anti-inflammatory ²⁸¹
TeC-376	mangicol C	
TeC-377	mangicol D	
TeC-378	mangicol E	
TeC-379	mangicol F	
TeC-380	mangicol G	
TeC-381	cyclocitrinol	Induces the production of cAMP ²⁸²
TeC-382	12 β ,20-dihydroxy-16 β -acetoxy-17-scalaren-19,20-olide	
TeC-383	19-acetyl-12-deacetyl-12- <i>epi</i> -deoxoscalarin	
TeC-384	unnamed tetracarbocyclic sesterterpene (reported as compound 2)	
TeC-385	hippospongide A	
TeC-386	phorone A	
TeC-387	suvanine N,N-dimethyl-1,3-dimethylherbipoline salt	Anti-cancer, Anti-microbial ¹⁷²
TeC-388	suvanine N,N-dimethylguanidium salt	Anti-cancer ¹⁷²
TeC-389	phorone B	Anti-cancer ²⁸³
TeC-390	anvilone A	Pro-viral ⁶⁷
TeC-391	anvilone B	
TeC-392	erinacine S	Anti-neurodegeneration ²⁸⁴
TeC-393	sesteralterin	Anti-microbial ²⁸⁵
TeC-394	19- <i>epi</i> -isosuberitenone B	Anti-cancer ²⁰⁵
TeC-395	isoxaspirosuberitenone	Anti-cancer ²⁰⁵
TeC-396	asperunguisin A	Anti-cancer ²⁸⁶
TeC-397	asperunguisin C	Anti-cancer ²⁸⁶
TeC-398	asperunguisin D	

TeC-399	asperunguisin E	
TeC-400	epihomoscalaralactone IIA	
TeC-401	homoscalarate II	
TeC-402	coscinalactone	
TeC-403	coscinafuran	
TeC-404	12 β ,16 β ,22-trihydroxy-24 <i>R</i> -methylscalar-25 β ,24 α -olide	
TeC-405	22-hydroxy-24-methylsedn-16-en-24-one-12 β ,25 β -olide	
TeC-406	unnamed tetracarbocyclic sesterterpene (reported as compound 1)	Anti-neurodegeneration ²⁴⁰
TeC-407	unnamed tetracarbocyclic sesterterpene (reported as compound 2)	Anti-neurodegeneration ²⁴⁰
TeC-408	hyatelone A	Anti-cancer ¹⁶²
TeC-409	hyatelone B	
TeC-410	corallocarpsalarolide	
TeC-411	phylofenone A	
TeC-412	dehydrofoliaspongin	
TeC-413	phylofoliaspongin	Anti-cancer ²⁸⁷
TeC-414	12 α -acetoxy-16 β -hydroxy-20,24-dimethyl-24-oxoscalar-25-al	
TeC-415	methyl 12 α -acetoxy-20,24-dimethyl-16,24-dioxoscalara-14,17-dien-25-oate	
TeC-416	unnamed tetracarbocyclic sesterterpene (12-deacetyl derivative of scalarin) (reported as compound 5)	Anti-microbial, Anti-cancer ²⁴⁷
TeC-417	unnamed tetracarbocyclic sesterterpene (reported as compound 6)	Anti-cancer ²⁴⁷
TeC-418	unnamed tetracarbocyclic sesterterpene (reported as compound 8)	Anti-microbial, Anti-cancer ²⁴⁷
TeC-419	unnamed tetracarbocyclic sesterterpene (reported as compound 9)	Anti-microbial, Anti-cancer ²⁴⁷
TeC-420	unnamed tetracarbocyclic sesterterpene (reported as compound 10)	Anti-microbial, Anti-cancer ²⁴⁷
TeC-421	unnamed tetracarbocyclic sesterterpene (reported as compound 11)	Anti-cancer ²⁴⁷
TeC-422	unnamed tetracarbocyclic sesterterpene (reported as compound 12)	Anti-cancer ²⁴⁷
TeC-423	unnamed tetracarbocyclic sesterterpene (reported as compound 13)	Anti-microbial, Anti-cancer ²⁴⁷
TeC-424	honulactone J	
TeC-425	honulactone K	
TeC-426	12 α -(3 <i>R'</i> -hydroxybutanoyloxy)-20,24-dimethyl-24-oxoscalar-16-en-25-al	
TeC-427	12,16-deacetoxy-12-oxo-scalarafuran	
TeC-428	12- <i>epi</i> -deoxoscalarin-3-one	Anti-cancer ²⁴²
TeC-429	deoxoscalarin-3-one	Anti-cancer ²⁴²
TeC-430	21-hydroxydeoxoscalarin	Anti-cancer ²⁴²
TeC-431	21-acetoxydeoxoscalarin	Anti-cancer ²⁴²
TeC-432	ketodeoxoscalarin	
TeC-433	12-deacetyl-12- <i>epi</i> -deoxoscalarin	
TeC-434	16-deacetoxy-scalarafuran	
TeC-435	(-)-capradiene A	
TeC-436	(+)-braparadiene A	
TeC-437	(-)-braparadiene B	
TeC-438	(-)-arathanadiene A	
TeC-439	(-)-arathanadiene B	
TeC-440	sesterevisene	
TeC-441	eurysoloid A	
TeC-442	eurysoloid B	
TeC-443	unnamed tetracarbocyclic sesterterpene (reported as sesteralterinderivative, compound 1)	
TeC-444	erectasclarane A	
TeC-445	erectasclarane B	
TeC-446	unnamed tetracarbocyclic sesterterpene (reported as sesteralterinderivative, compound 2)	
TeC-447	gentianelloid C	
TeC-448	18- <i>epi</i> -gentianelloid C	
TeC-449	gentianelloid D	
TeC-450	18- <i>epi</i> -gentianelloid D	
TeC-451	gentianelloid E	
TeC-452	18- <i>epi</i> -gentianelloid E	
TeC-453	gentianelloid F	Immunosuppressive ²⁷⁵
TeC-454	18- <i>epi</i> -gentianelloid F	
TeC-455	unnamed tetracarbocyclic sesterterpene (reported as sesteralterinderivative, compound 3)	
TeC-456	unnamed tetracarbocyclic sesterterpene (reported as sesteralterinderivative, compound 4)	Anti-inflammatory ²⁸⁸

TeC-457	unnamed tetracarbocyclic sesterterpene (reported as sesteralterinderivative, compound 5)	Anti-inflammatory ²⁸⁸
TeC-458	unnamed tetracarbocyclic sesterterpene (reported as sesteralterinderivative, compound 6)	
TeC-459	dysiscalarone D	
TeC-460	dysiscalarone E	
TeC-461	spectanoid H	
TeC-462	unnamed tetracarbocyclic sesterterpene (reported as compound 1)	
TeC-463	unnamed tetracarbocyclic sesterterpene (reported as sesteralterinderivative, compound 7)	Anti-microbial ²⁸⁹
TeC-464	sesterviolene A	
TeC-465	deacetoxyscalarin	
TeC-466	sesterviolene C	
TeC-467	sesterviolene D	
TeC-468	12 α -acetoxy-16 β -hydroxy-20,24-dimethyl-25-norscalarane-18,24-carbolactone	
TeC-469	12- <i>O</i> -deacetyl-19 α -methoxy-12- <i>epi</i> -scalarin	
TeC-470	25-dehydroxy-12- <i>epi</i> -deacetylscalarin	
TeC-471	hyrtiosin F	
TeC-472	hyrtiosin G	
TeC-473	12 β ,20 β -dihydroxy-16 α -methoxy-17-scalaren-19,20-olide	
TeC-474	12 β ,20 α -dihydroxy-16 β -methoxy-17-scalaren-19,20-olide	
TeC-475	12 β ,16 β ,20 β -trihydroxy-17-scalaren-19,20-olide	
TeC-476	12 β ,19 α (β)-dihydroxy-16 α -methoxy-17-scalaren-19,20-olide	
TeC-477	12 β ,19 α (β)-dihydroxy-16 β -methoxy-17-scalaren-19,20-olide	
TeC-478	12 β ,19 α -dihydroxy-14,15-dehydrate-17-scalaren-19,20-olide	
TeC-479	12-deacetyl-18- <i>epi</i> -carboxylic-12- <i>epi</i> -scalaral	
TeC-480	2- <i>O</i> -deacetyl-12,16-di- <i>epi</i> -norscalaral B	
TeC-481	phorone C	Pro-viral ¹⁹⁰
TeC-482	mangicdiene	
TeC-483	mangicol H	
TeC-484	mangicol I	
TeC-485	mangicol J	
TeC-486	mangicol K	
TeC-487	mangicol L	
TeC-488	phylloketal	
TeC-489	16- <i>epi</i> -scalarole (16- <i>epi</i> -scalarolbutenolide)	
TeC-490	sesteraltererol	
TeC-491	10,11-epoxysesteraltererol	
TeC-492	acetoxiphyllolofactone A	
TeC-493	12-deacetyl-3-oxoscalarin	
TeC-494	hyrtioscalarin A	
TeC-495	hyrtioscalarin B	
TeC-496	hyrtioscalarin C	
TeC-497	hyrtioscalarin D	
TeC-498	hyrtioscalarin E	
TeC-499	hyrtioscalarin F	
TeC-500	hyrtioscalarin G	
TeC-501	hyrtioscalarin H	
TeC-502	17(<i>R</i>),18(<i>S</i>)-dihydroxy-19(<i>R</i>),20(<i>S</i>)-dimethoxysesterstatin 5	
TeC-503	17(<i>R</i>),18(<i>S</i>)-dihydroxy-19(<i>R</i>),20(<i>R</i>)-dimethoxysesterstatin 5	
TeC-504	salmahyrtisol B*	
Tec-505	Bm2	
Tec-506	(17Z)-13,19-epoxycheilanth-17-en-6 α -ol	
Tec-507	18-episcalar-16-ene-6 α ,19-diol	
Tec-508	16 α ,19-epidioxy-18-episcalar-17(25)-en-6 α -ol	
Tec-509	unamed 5-8-6-6 tetracyclic sesterterpene (reported as compound 56)	
Tec-510	suberitenone E	
Tec-511	suberitenone F	Anti-viral ²⁰⁴
Tec-512	suberitenone G	
Tec-513	suberitenone H	
Tec-514	suberitenone I	
Tec-515	suberitenone J	
Tec-516	phyllifenone F	

Tec-517	phylofenone G	Anti-cancer ²⁹⁰
Tec-518	phylofenone H	
Tec-519	12-β- <i>O</i> -acetylhyrtiolide	Anti-cancer ¹⁹⁴
Tec-520	phylofolactone N	
Tec-521	phylofolactone O	
Tec-522	phylofolactone P	
Tec-523	phylofolactone Q	
Tec-524	phylofolactone R	
Tec-525	phylofolactone S	Anti-cancer ²⁹¹
Tec-526	phylofolactone T	
Tec-527	hyrtiosin H	
Tec-528	hyrtiosin I	
Tec-529	hyrtiosin J	
Tec-530	phyllospongiane A	Anti-cancer ²⁹² , Anti-microbial ²⁹²
Tec-531	phyllospongiane B	Anti-cancer ²⁹² , Anti-microbial ²⁹²
Tec-532	phyllospongiane C	Anti-cancer ²⁹²
Tec-533	phyllospongiane D	Anti-cancer ²⁹² , Anti-microbial ²⁹²
Tec-534	phyllospongiane E	Anti-cancer ²⁹²
Tec-535	sesterchaetin A	Anti-microbial ²⁹³
Tec-536	sesterchaetin B	Anti-microbial ²⁹³
Tec-537	bipoladien A	
Tec-538	bipoladien B	
Tec-539	emerindanol A	
Tec-540	emerindanol B	
Tec-541	felixin A	Anti-cancer ²⁹⁴
Tec-542	lendenfeldarane V	
Tec-543	phylofenone I	Anti-cancer ²⁹⁰ , Anti-microbial ²⁹⁰
Tec-544	phylofenone J	Anti-cancer ²⁹⁰
Tec-545	phylofenone K	Anti-microbial ²⁹⁰
Tec-546	phylofenone L	Anti-cancer ²⁹⁰
Tec-547	phylofenone M	Anti-cancer ²⁹⁰

Section 9. Biological activities of pentacarbocyclic (PC) sesterterpenoids (with and without hetherocycles)

c. n.	IUPAC name or Trivial name	Bioactivity (reference)
PC-1	quiannulatene	
PC-2	quiannulatic acid	
PC-3	(-)- <i>ent</i> -quiannulatene	
PC-4	(-)-retigeranin B	
PC-5	(+)-astallatene	
PC-6	(+)-boleracene	
PC-7	retigeranic acid A	
PC-8	astellatol	
PC-9	(+)-astellatene	
PC-10	12α-acetoxy-23,25-cycle-16β,25Z-dihydroxy-20,24-dimethyl-oxoscalarane	
PC-11	carteriofenone J	Anti-cancer ²⁰⁷
PC-12	phylofenone B	Anti-cancer ²⁹⁵
PC-13	phylofenone D	Anti-cancer ²⁹⁶
PC-14	12β,22-dihydroxy-24-oxo-24-homoscalara-16,25(26)-diene	Anti-cancer ²²⁷
PC-15	scalalactam C	
PC-16	scalalactam D	
PC-17	unnamed pentacarbocyclic sesterterpene (reported as compound 2)	
PC-18	12α-acetoxy-13β,18β-cyclobutane-20,24-dimethyl-24-oxoscalar-16-en-25α-ol	
PC-19	12α-acetoxy-13β,18β-cyclobutane-20,24-dimethyl-24-oxoscalar-16-en-25β-ol	Anti-cancer, Anti-microbial ²³⁰
PC-20	carteriofenone E	
PC-21	carteriofenone F	
PC-22	carteriofenone G	
PC-23	carteriofenone H	
PC-24	phyllospongine D	Anti-cancer, Anti-microbial ²³⁰
PC-25	phyllospongine E	Anti-cancer, Anti-microbial ²³⁰
PC-26	isoartemisolid	Anti-inflammatory ²⁹⁷
PC-27	toxistylide-A	

PC-28	toxistilyde-B	
PC-29	molliorin-d	
PC-30	unnamed pentacarboxylic sesterterpene (reported as compound 4)	Anti-cancer ²⁹⁸
PC-31	asperterpenoid A	Anti-microbial ²⁹⁹
PC-32	retigeran-11-ol	
PC-33	4-hydroxyretigeran-11-ol	
PC-34	gypmacrophin A	
PC-35	peniroquesine A	
PC-36	peniroquesine B	
PC-37	peniroquesine C	
PC-38	aspterpenacid A	
PC-39	aspterpenacid B	
PC-40	aspterpenacid C	Anti-viral, Anti-inflammatory ³⁰⁰
PC-41	asperunguisin B	
PC-42	honulactone A	Anti-cancer ^{253, 288}
PC-43	honulactone B	Anti-cancer ²⁵³
PC-44	honulactone E	
PC-45	honulactone F	
PC-46	honulactone G	
PC-47	unnamed pentacarboxylic sesterterpene (reported as compound 8)	
PC-48	unnamed pentacarboxylic sesterterpene (reported as compound 10)	
PC-49	honulactone H	
PC-50	unnamed pentacarboxylic sesterterpene (reported as compound 11)	
PC-51	unnamed pentacarboxylic sesterterpene (reported as compound 12)	
PC-52	unnamed pentacarboxylic sesterterpene (reported as compound 9)	
PC-53	bolivianine	
PC-54	isobolivianine	
PC-55	emervaridione	
PC-56	retigeranic acid B	
PC-57	8- <i>epi</i> -peniroquesine A	Anti-cancer ³⁰¹
PC-58	8-oxo-eniroquesine B	
PC-59	12-deoxy-16S-hydroxypeniroquesine B	
PC-60	12-deoxypeniroquesine A	
PC-61	12-deoxy-16S-hydroxypeniroquesine A	Anti-cancer ³⁰¹
PC-62	12-deoxy-22-oxo-16S-hydroxypeniroquesine A	
PC-63	8-deoxy-16S-hydroxypeniroquesine A	Anti-microbial ³⁰¹
PC-64	asperterpenoid B	Anti-microbial ³⁰²
PC-65	asperterpenoid C	
PC-66	asperterpenoid D	
PC-67	asperterpenoid E	
PC-68	asperterpenoid F	
PC-69	asperterpenoid G	Anti-microbial ³⁰²
PC-70	asperterpenoid H	Anti-microbial ³⁰²
PC-71	asperterpenoid I	Anti-microbial ³⁰²
PC-72	asperterpenoid J	
PC-73	asperterpenoid K	
PC-74	asperterpenoid L	
PC-75	asperterpenoid M	
PC-76	asperterpenoid N	
PC-77	asperterpenoid O	
PC-78	asperterpenoid P	
PC-79	preasperterpenoid A	
PC-80	18- <i>epi</i> -nitidasin	
PC-81	dysiscalarone A	
PC-82	dysiscalarone B	
PC-83	dysiscalarone C	
PC-84	12-deacetoxy-4-demethyl-11,24-diacetoxy-3,4-methylenedioxoscalarin	
PC-85	neosuberitenone	Anti-viral ²⁰⁴
PC-86	sestermobaraene A	
PC-87	sestermobaraene B	
PC-88	sestermobaraene C	
PC-89	sestermobaraene D	
PC-90	sestermobaraene F	
PC-91	sestermobaraol	

PC-92	niduene A	Anti-cancer ³⁰³
PC-93	niduene B	
PC-94	niduene C	
PC-95	niduene D	
PC-96	niduene E	
PC-97	niduene F	
PC-98	unnamed pentacyclic sesterterpene	
PC-99	unnamed pentacyclic sesterterpene	
PC-100	unnamed pentacyclic sesterterpene	
PC-101	unnamed pentacyclic sesterterpene	
PC-102	unnamed pentacyclic sesterterpene	
PC-103	unnamed pentacyclic sesterterpene	
PC-104	unnamed pentacyclic sesterterpene	
PC-105	unnamed pentacyclic sesterterpene	

Section 10. Biological activities of hexacarbocyclic (HC) sesterterpenoids (with and without hetherocycles)

c. n.	IUPAC name or Trivial name	Bioactivity (reference)
HC-1	niduterpenoid A	Anti-cancer ³⁰⁴
HC-2	niduterpenoid B	Anti-cancer ³⁰⁴
HC-3	disidein	
HC-4	6'-bromo-disidein	
HC-5	6'-cloro-disidein	
HC-6	acanthosulfate	Enzyme Inhibitor (Proteasome) ³⁰⁵
HC-7	oxaliterpenoid	
HC-8	subrutilane	

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