

## SUPPLEMENTARY INFORMATION

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### Synthesis of Carbon Dots from Spent Coffee Grounds: Transforming Waste into Potential Biomedical Tools

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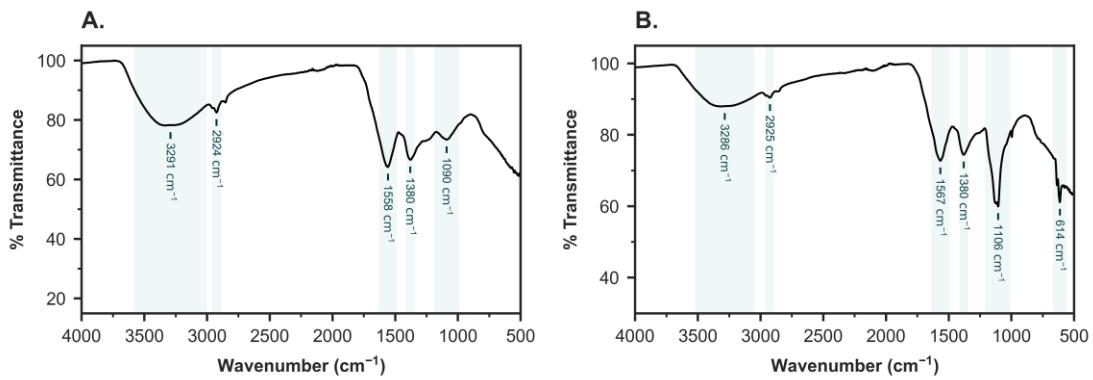
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## SUPPLEMENTARY INFORMATION



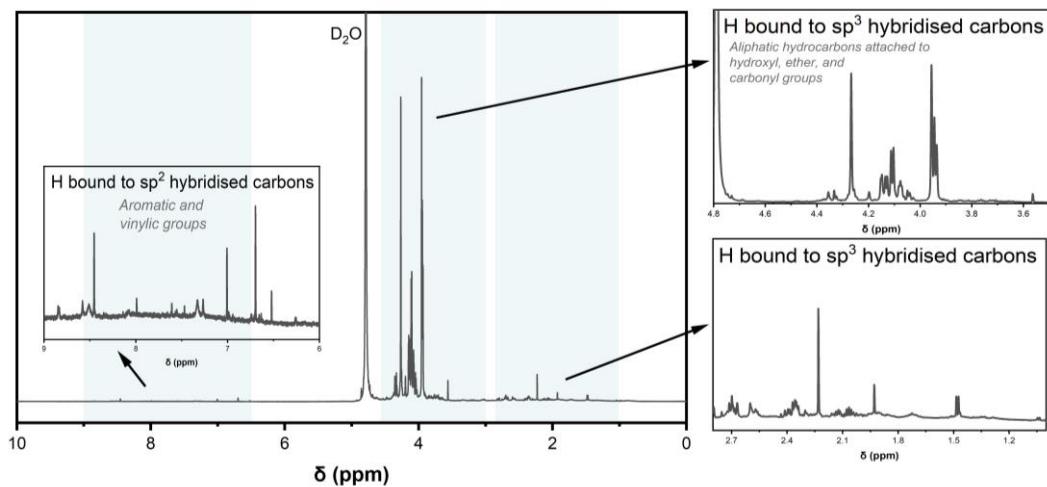
**Fig S1.** ATR-FTIR spectra of **A.** D-LAV-CDs and **B.** D-KEN-CDs. Prominent bands have been highlighted and labelled with the corresponding wavenumber.

**Tab S1.** Wavenumbers corresponding to key functional groups observed in the ATR-FTIR spectra of AP-LAV-CDs, D-LAV-CDs and D-KEN-CDs. The symbol  $\nu$  denotes stretching vibrations and *n.p.* denotes 'not present'.

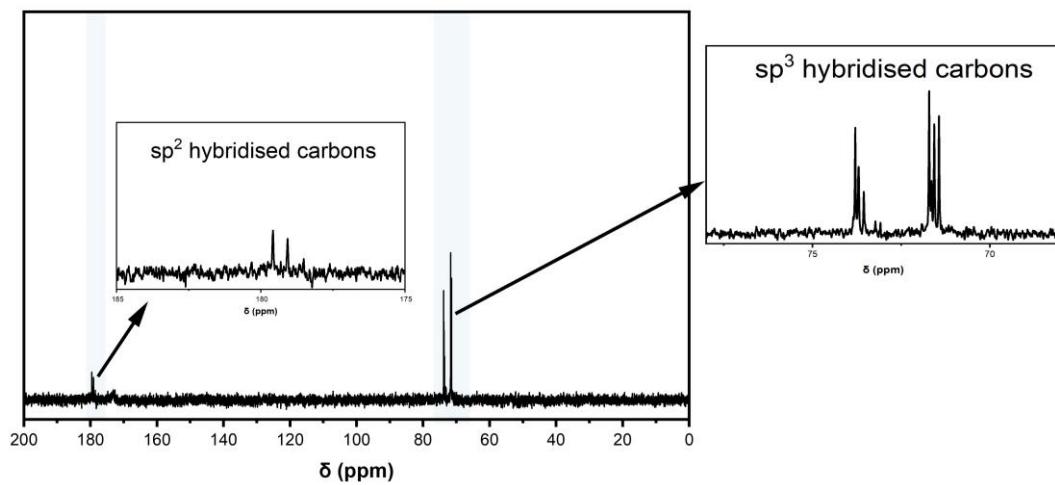
	AP-LAV-CDs (cm <sup>-1</sup> )	D-LAV-CDs (cm <sup>-1</sup> )	D-KEN-CDs (cm <sup>-1</sup> )
$\nu(\text{OH})$	3322	3291	3286
$\text{sp}^3 \nu(\text{C-H})$	<i>n.p.</i>	2924	2925
$\nu(\text{C=O})$	1598	1558	1567
$\nu(\text{C=C})$	1402	1380	1380
$\nu(\text{C-O-C})$	1099	1106	1106
<b>fingerprint region</b>	611	<i>n.p.</i>	614

## SUPPLEMENTARY INFORMATION

**A.**



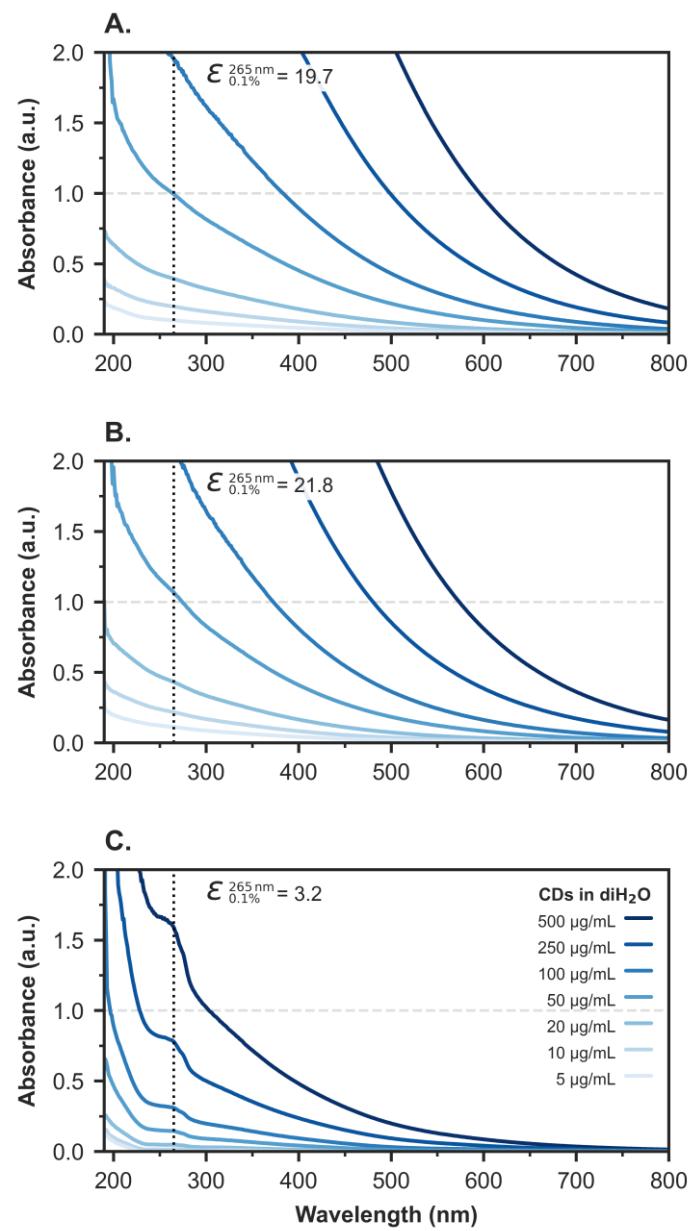
**B.**



**Fig S2.** NMR spectra of AP-LAV-CDs. **A.**  $^1\text{H}$  NMR spectrum (1024 scans), showing signals from  $\text{sp}^3$ - and  $\text{sp}^2$ -hybridised carbon atoms. *Insets:* Expanded regions from 9.6 ppm, 4.8-4.5 ppm, and 2.8-1 ppm. **B.**  $^{13}\text{C}$  NMR spectrum (4096 scans), displaying  $\text{sp}^3$ - and  $\text{sp}^2$ -hybridised carbon atoms, with peaks around 70 ppm and 180 ppm, respectively. *Inset:* Expanded regions from 185-175 ppm and 78-68 ppm.

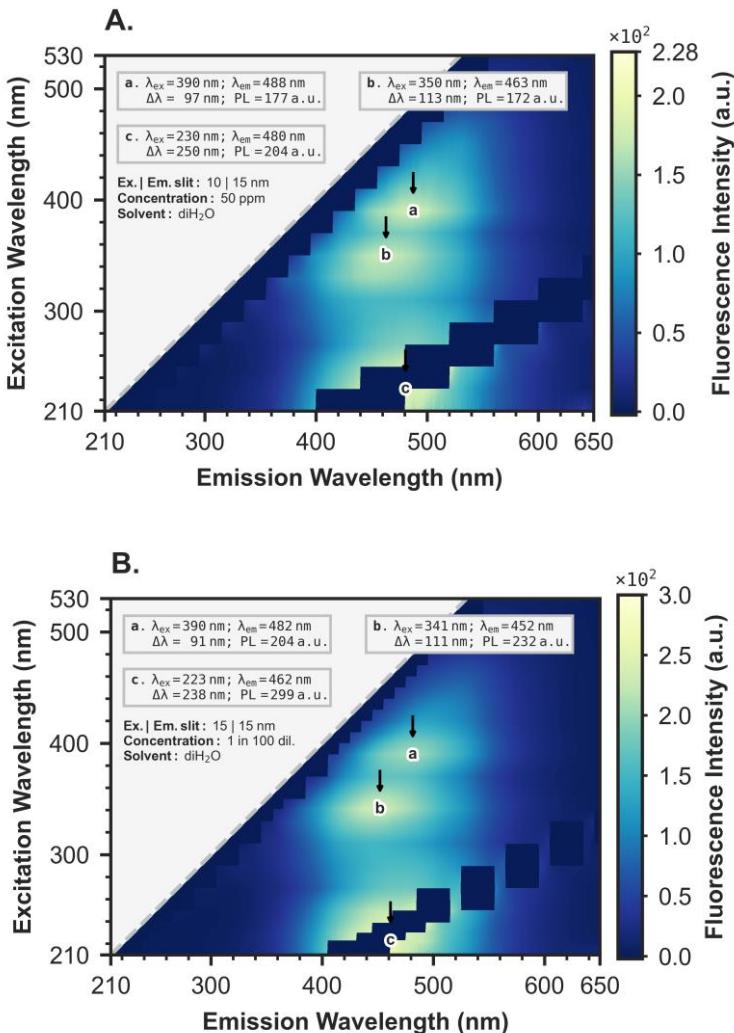
## SUPPLEMENTARY INFORMATION

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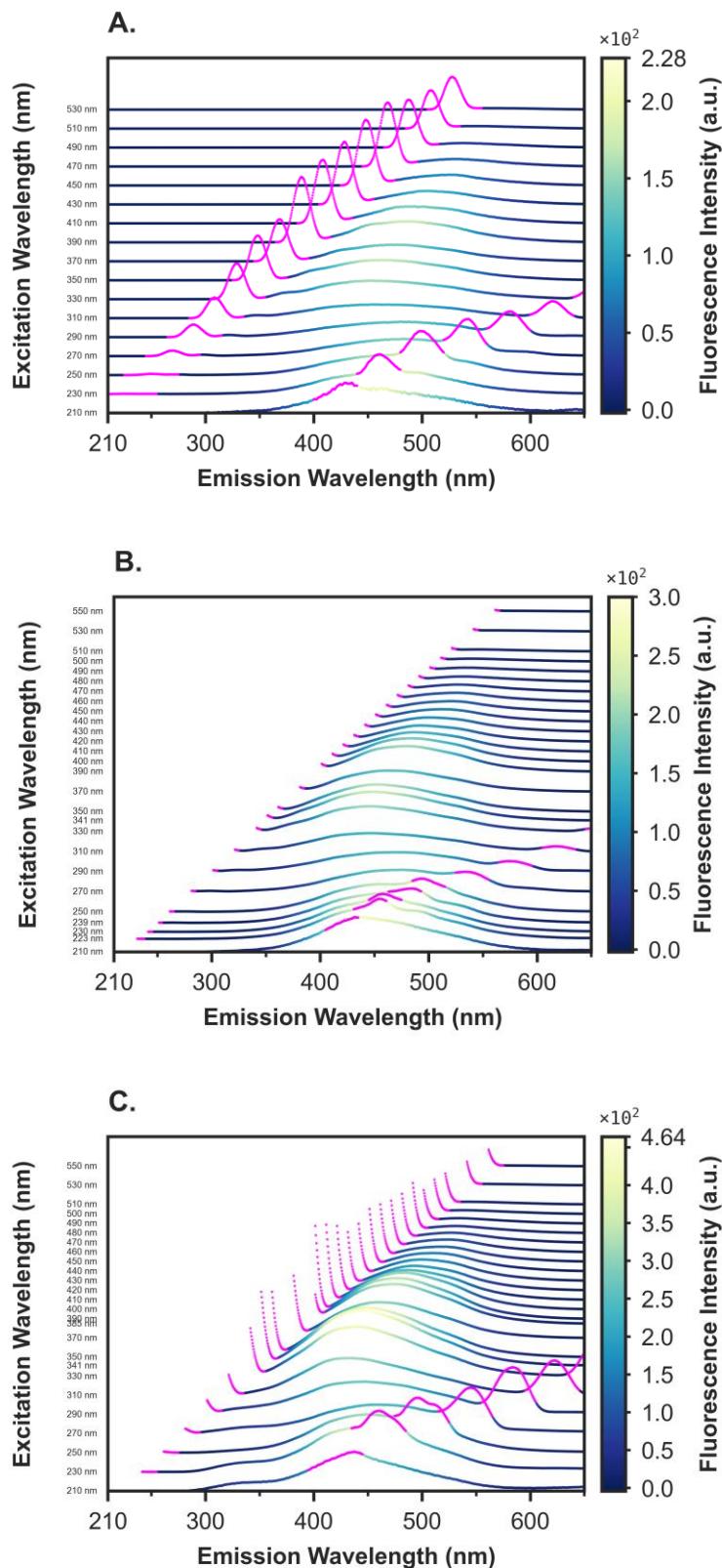
**Fig S3.** UV-Vis absorption spectra of **A.** D-LAV-CDs, **B.** D-KEN-CDs and **C.** AP-LAV-CDs dispersed in  $\text{diH}_2\text{O}$  at 5, 10, 20, 50, 100, 250 and 500  $\mu\text{g/mL}$ . The 0.1% excitation coefficient calculated at 265 nm for concentrations  $\leq 50 \mu\text{g/mL}$  is shown.

## SUPPLEMENTARY INFORMATION



**Fig S4.** Fluorescence excitation emission matrix (EEM) spectra of **A.** D-KEN-CDs and **B.** D-LAV-CDs. Relevant experimental conditions have been noted, and significant emission regions have been annotated as **a**, **b** & **c**, with the excitation wavelength ( $\lambda_{\text{ex}}$ ), emission wavelength ( $\lambda_{\text{em}}$ ) and Stokes shift ( $\Delta\lambda$ ) at the point of highest intensity emission (PL). First- and second-order Rayleigh scattering signals have been excluded (dark spaces).

## SUPPLEMENTARY INFORMATION



**Fig S5.** Fluorescence emission spectroscopy data of **A.** D-KEN-CDs; **B.** D-LAV-CDs; and **C.** AP-LAV-CDs visualised in an alternative approach to the EEM. First- and second-order Rayleigh scattering signals have been highlighted in pink.

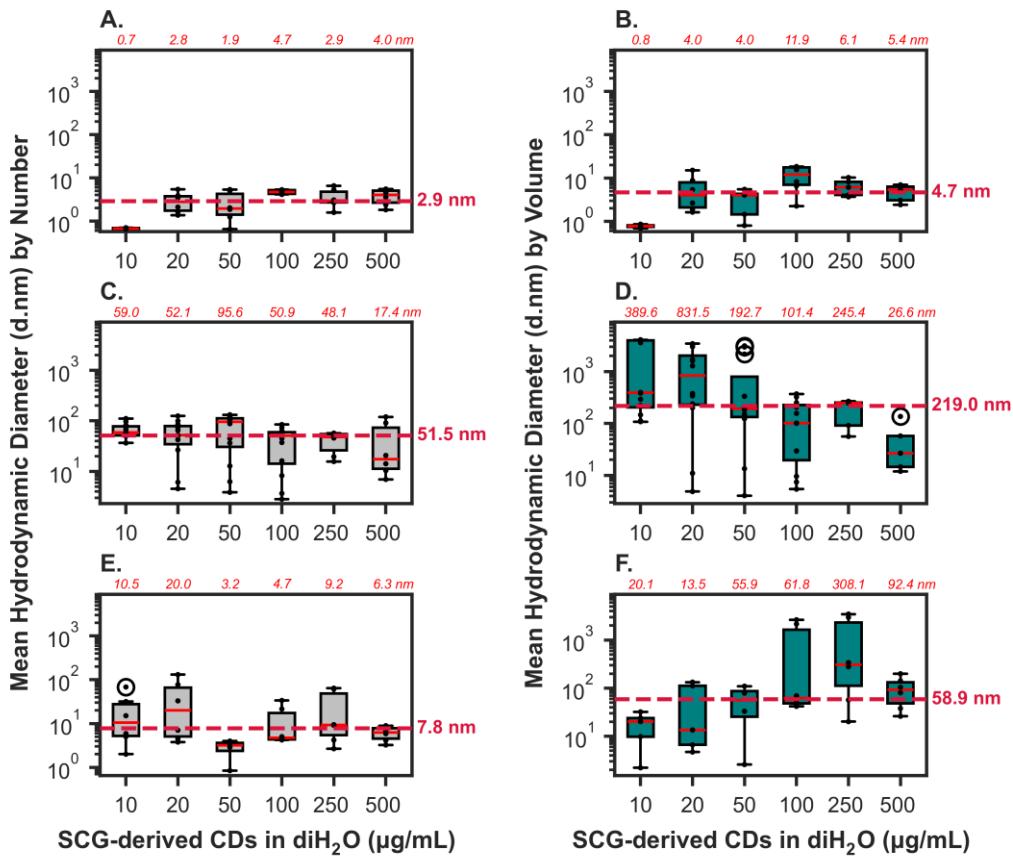
## SUPPLEMENTARY INFORMATION

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**Tab S2.** Summary of SCG-derived CDs' fluorescence emission results; included are the excitation wavelength ( $\lambda_{\text{ex}}$ ), emission wavelength ( $\lambda_{\text{em}}$ ), Stokes shift ( $\Delta\lambda$ ), and highest intensity emission (PL) for the annotated regions **a**, **b**, and **c** in the respective fluorescence excitation emission matrix (EEM) spectra of AP-LAV-CDs, D-LAV-CDs and D-KEN-CDs.

Region	Parameter	AP-LAV-CDs	D-LAV-CDs	D-KEN -CDs
<b>a</b>	$\lambda_{\text{ex}}$ (nm)	390	390	390
	$\lambda_{\text{em}}$ (nm)	478	482	488
	$\Delta\lambda$ (nm)	88	91	97
	PL (a.u.)	345	204	177
<b>b</b>	$\lambda_{\text{ex}}$ (nm)	341	341	350
	$\lambda_{\text{em}}$ (nm)	445	452	463
	$\Delta\lambda$ (nm)	104	111	113
	PL (a.u.)	462	232	172
<b>c</b>	$\lambda_{\text{ex}}$ (nm)	230	223	230
	$\lambda_{\text{em}}$ (nm)	434	462	480
	$\Delta\lambda$ (nm)	204	238	250
	PL (a.u.)	368	299	204

## SUPPLEMENTARY INFORMATION



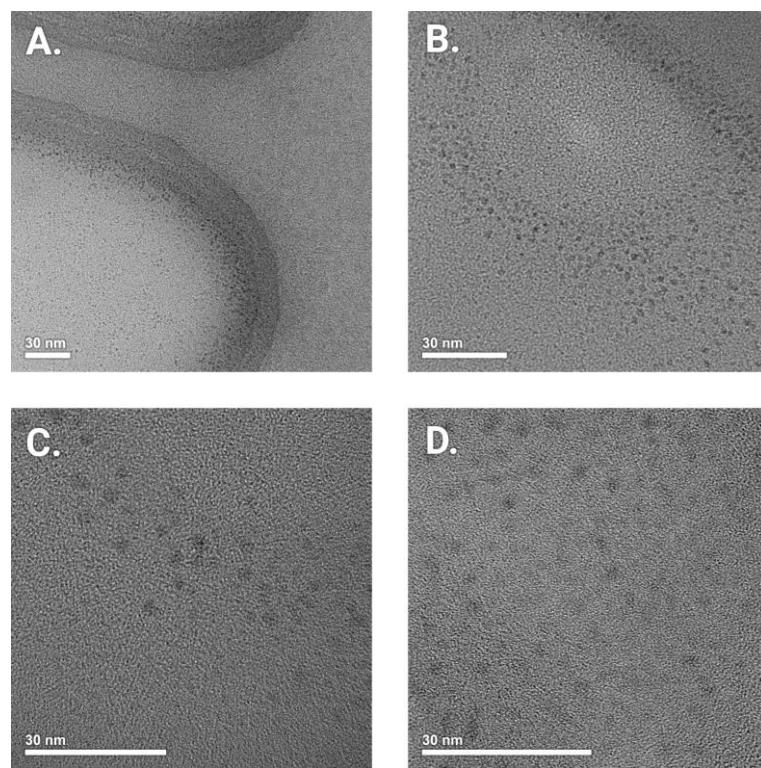
**Fig S6.** DLS hydrodynamic diameter (d.nm) data of **A. & B.** AP-LAV-CDs represented by number and by volume, respectively; **C. & D.** D-LAV-CDs represented by number and by volume, respectively; **E. & F.** D-KEN-CDs represented by number and by volume, respectively. The SCG-derived CDs have been dispersed in diH<sub>2</sub>O at 10, 20, 50, 100, 250 and 500 µg/mL, and the data represents  $n \geq 6$  runs. The data for each concentration has been represented as a box and whisker plot. The individual median values for each concentration, as well as the overall median across all concentrations, have been annotated in red.

**Tab S3.** DLS mean hydrodynamic diameter (d.nm) data for AP-LAV-CDs, D-LAV-CDs, and D-KEN-CDs, represented by number distribution. The SCG-derived CDs were dispersed in diH<sub>2</sub>O at concentrations of 10, 20, 50, 100, 250, and 500 µg/mL. Data is presented as mean  $\pm$  standard deviation from  $n \geq 6$  runs for each concentration.

Concentration (µg/mL)	AP-LAV-CDs (nm)	D-LAV-CDs (nm)	D-KEN-CDs (nm)
10	0.7 $\pm$ 0.1	66.8 $\pm$ 22.4	21.3 $\pm$ 25.4
20	3.0 $\pm$ 1.6	57.1 $\pm$ 37.5	42.7 $\pm$ 51.6
50	2.7 $\pm$ 2.0	73.5 $\pm$ 49.1	2.8 $\pm$ 1.4
100	4.7 $\pm$ 0.5	42.3 $\pm$ 28.4	12.2 $\pm$ 12.5
250	3.7 $\pm$ 1.9	40.7 $\pm$ 18.5	25.1 $\pm$ 29.2
500	3.8 $\pm$ 1.5	43.8 $\pm$ 48.7	6.2 $\pm$ 2.2

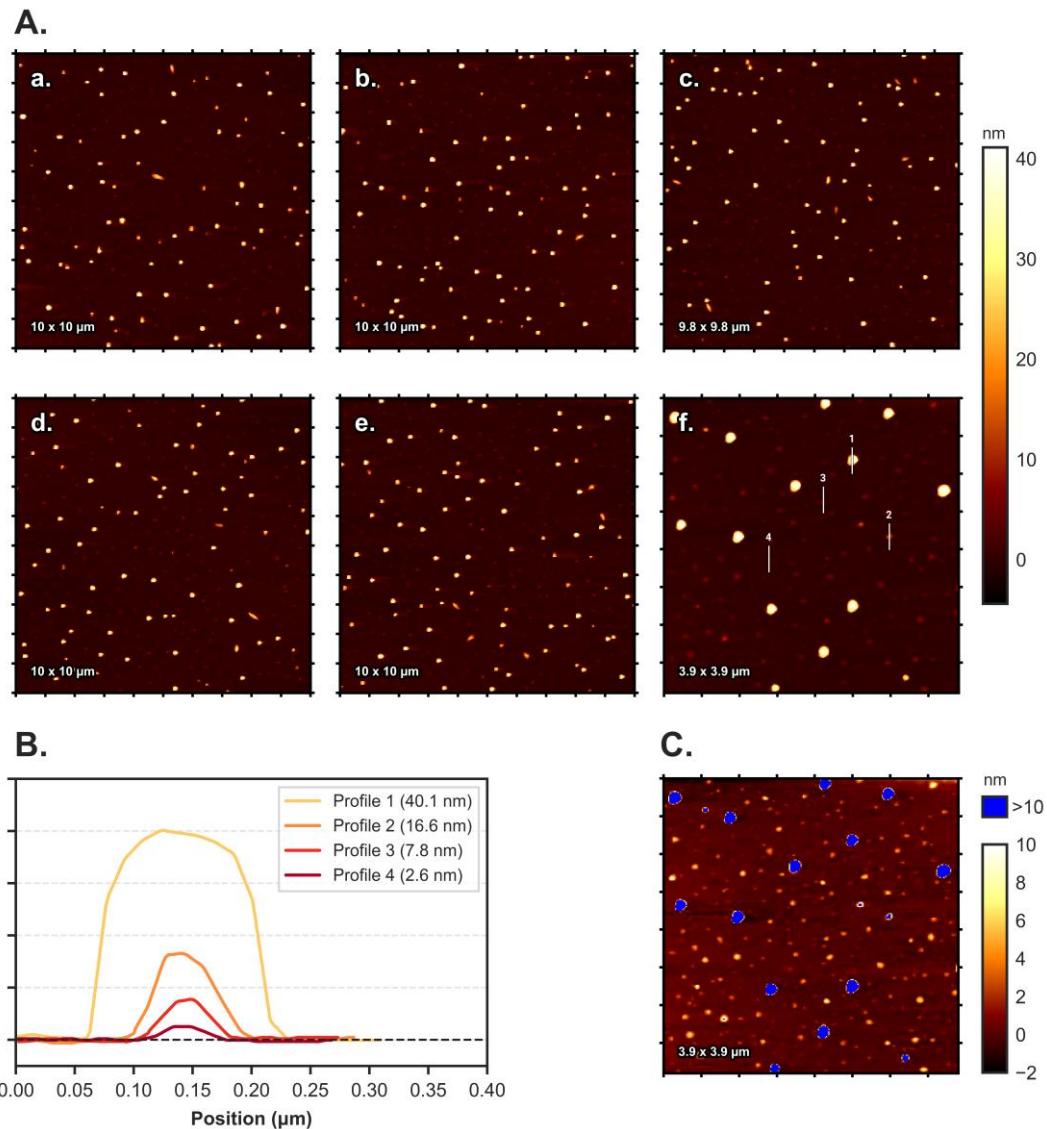
## SUPPLEMENTARY INFORMATION

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**Fig S7.** TEM images of AP-LAV-CDs at four different magnifications (**A.** 80,000x, **B.** 150,000x, **C.** 250,000x, and **D.** 300,000x); all images have been fitted with a 30 nm scale bar.

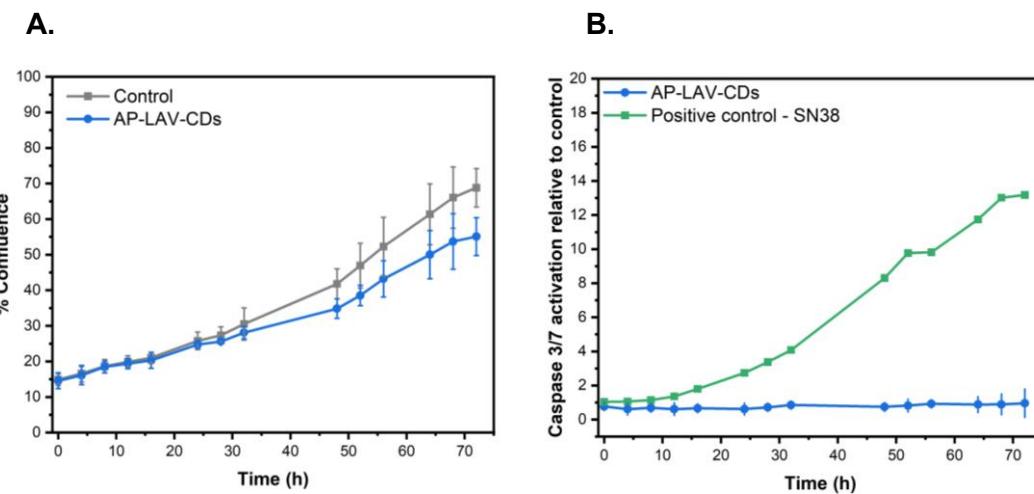
## SUPPLEMENTARY INFORMATION



**Fig S8.** **A.** AFM images of AP-LAV-CDs at different scan sizes. All AFM images **a-f**, have been represented using a common colour-scale for comparative purposes. **B.** Line profiles 1 to 4, respectively marked in subfigure **f.**, highlighting NPs of different heights; the maximum height of each line profile is provided in its respective legend entry. **C.** AFM image **f.** with the colour-scale normalised to a maximum height of 10 nm to better visualise small ( $< 10$  nm height) NPs. Height data clipping  $> 10$  nm has been highlighted in blue.

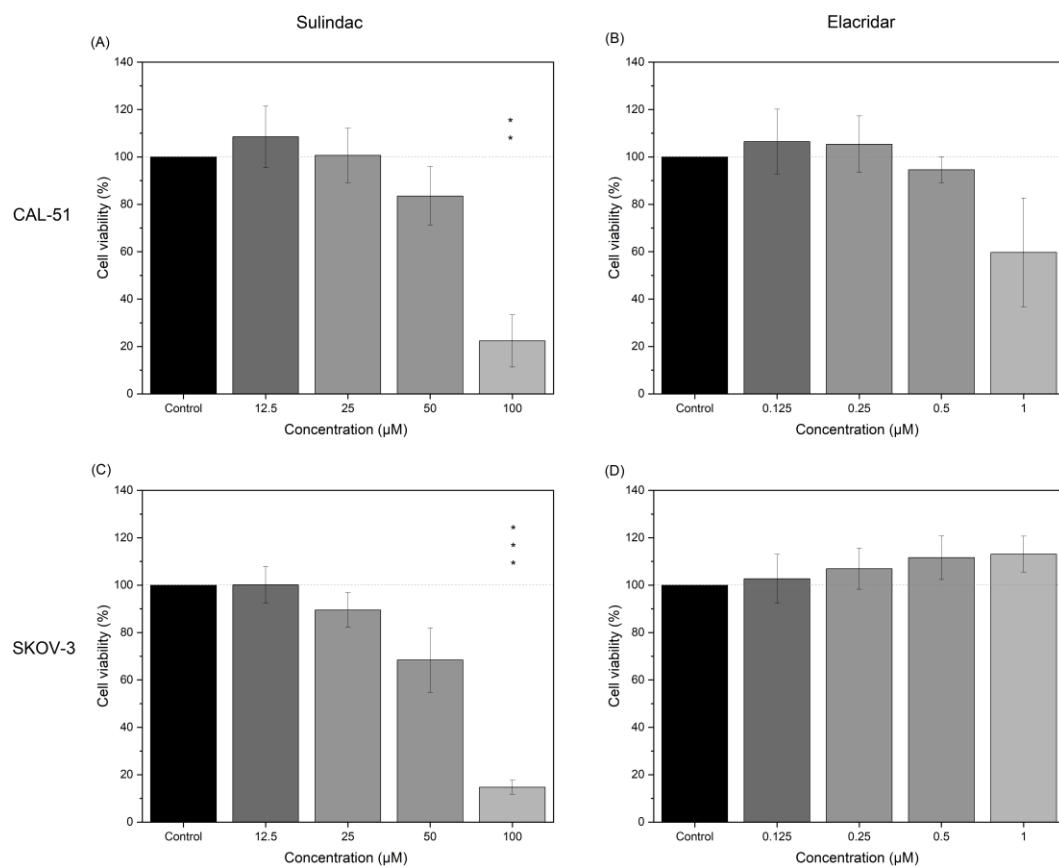
## SUPPLEMENTARY INFORMATION

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**Fig S9.** Apoptosis study on AP-LAV-CDs with InCucyte® Live-Cell Analysis System **A.** Cell confluence of CAL-51 cells treated with 250  $\mu$ g/mL AP-LAV-CDs; **B.** Apoptosis signal in CAL-51 cells treated with 250  $\mu$ g/mL AP-LAV-CDs compared to the positive control SN38. Data is normalized to untreated controls to account for baseline apoptosis levels.

## SUPPLEMENTARY INFORMATION



**Fig S10.** Cellular viability of **A.** CAL-51 cells treated with sulindac; **B.** CAL-51 cells treated with elacridar; **C.** SKOV-3 cells treated with sulindac; and **D.** SKOV-3 cells treated with elacridar. Cells were exposed to sulindac at concentrations of 0 (control), 12.5, 25, 50, and 100  $\mu$ M, and to elacridar at concentrations of 0 (control), 0.125, 0.25, 0.50, and 1  $\mu$ M, for 120 h. Data represent the mean  $\pm$  standard deviation of three replicates.