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Supporting Information

for

Iron-catalyzed Selective Construction of Indole Derivatives via Oxidative C(sp3)-H Functionalization of Indolin-2-ones

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1. General Procedure

NMR spectrum were recorded on Varian Inova 400, Bruker 400M, 500M and 600 M spectrometers in CDCl₃ or DMSO- d_6 and chemical shifts are expressed in parts per million (δ , ppm). Proton chemical shifts are referenced to 7.26 ppm (CHCl₃) or 2.50 ppm (DMSO- d_6) and carbon chemical shifts are referenced to 77.2 ppm (CHCl₃) or 39.5 ppm (DMSO- d_6). High resolution mass spectrum (HRMS) were recorded on a Q-TOF mass spectrometry equipped with Z-spray ionization source. Infrared spectrum (IR) were measured using a Nicolet NEXUS FT-IR spectrophotometer. Commercially available chemicals were purchased from TCI, Sigma-Aldrich, Alfa Aesar, Acros, adamas, Energy Chemicalor Innochem and used directly without further purification.

2. Iron-catalyzed oxidative reaction of indolin-2-ones to isatins.

A 10 mL flask containing a stir bar was charged with oxindolin-2-one (1.0 mmol), $FeCl_2$ (0.01 mmol, 1.0 mmol%) and EtOH (1.0 mL), respectively. The reaction mixture was stirred at 60 °C in open air for 12 h, then cooled to room temperature. The solvent was evaporated at reduced pressure and the crude product was purified by column chromatography on silica gel (petroleum ether/EtOAc: 10/1-3/1) to give desired products.



3. Iron-catalyzed oxidative coupling of indolin-2-ones to symmetrical isoindigos.

A 10 mL flask containing a stir bar was charged with oxindolin-2-one (1.0 mmol), $FeCl_2$ (0.001 mmol, 0.1 mmol%) and 1-pentanol (1.0 mL), respectively. The reaction mixture was stirred at 100 °C in open air for 24 h, then cooled to room temperature. The solvent was evaporated at reduced pressure and the crude product was purified by washing with dichloromethane or column chromatography on silica gel (petroleum ether/EtOAc: 10/1-5/1 or dichloromethane) to give desired products.



4. Iron-catalyzed cascade reaction of indolin-2-ones to non-symmetrical isoindigos.

In an argon atmosphere, a 10 mL oven-dried flask containing a stir bar was charged with oxindolin-2one (0.5 mmol), isatin (0.5 mmol), FeCl₂ (0.0005 mmol, 0.1 mmol%) and 1-pentanol (1.0 mL), respectively. The reaction mixture was stirred at 100 °C in argon for 2 h, then stirred for 1 h in open air. Subsequently, the reaction mixture was cooled to room temperature. The solvent was evaporated at reduced pressure and the crude product was purified by column chromatography on silica gel (petroleum ether/EtOAc: 10/1-1/1) to give desired product.



5. Mechanistic studies

5.1 Control experiments

In order to investigate reaction mechanism, several control experiments were performed as depicted in Scheme S1. In the absence of $FeCl_2$ or air, no desired product **2a** or **3a** was observed (Scheme S1. I, II, III and IV), which indicated iron salts and air were essential for this process.

(I) Reaction in argon at 65 °C

(III) Reaction in argon at 100 °C



Scheme S1. Control experiments

5.2 Intermediate capture experiments

In air atmosphere, a 10 mL flask containing a stir bar was charged with oxindolin-2-one 1a (1.0 mmol), isatin 2k (1.0 mmol), FeCl₂ (0.001 mmol 0.1 mmol%) and 1-pentanol (1.0 mL), respectively. The reaction mixture was stirred at 100 °C for 1 h under open air. Then the reaction mixture was cooled to room temperature, and the solvent was evaporated under reduced pressure. The crude product was filtered and the residue was washed with dichloromethane to give 3a in 8% yield. The filtrate was further purified by column chromatography on silica gel (dichloromethane) to gain 4a in 90% yield.



In argon atmosphere, a 10 mL oven-dried flask containing a stir bar was charged with oxindolin-2-one **1a** (1.0 mmol), isatin **2k** (1.0 mmol), FeCl₂ (0.001 mmol 0.1 mmol%) and 1-pentanol (1.0 mL), respectively. The reaction mixture was stirred at 100 °C for 1 h under argon atmosphere. Then the reaction mixture was cooled to room temperature. The solvent was evaporated under reduced pressure and the crude product was purified by washing with dichloromethane to give aldol product **5** in 62% yield.



Aldol product (5). White solid. ¹H NMR (600 MHz, DMSO- d_6) δ 10.12 (s, 1H), 7.56 (t, J = 13.1 Hz, 1H), 7.24 (m, 2H), 7.02 (t, J = 7.5 Hz, 1H), 6.93 (d, J = 7.7 Hz, 1H), 6.77 – 6.68 (m, 2H), 6.66 (s, 1H), 6.15 (d, J = 7.2 Hz, 1H), 4.02 (d, J = 10.8 Hz, 1H), 3.09 (d, J = 6.2 Hz, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 175.6, 173.9, 144.1, 143.3, 129.7, 128.5, 127.6, 126.6, 125.7, 123.1, 121.7, 121.1, 109.1, 108.4; IR (KBr cm⁻¹): 3250, 1705, 1686, 1614, 1472, 1380, 1336, 1105, 750; HRMS(ESI): calcd. for C₁₇H₁₅N₂O₃: [M+H]⁺ m/z 295.1083, found: 295.1084.



Figure S1. ¹H NMR spectrum of 5



Figure S2. ¹³C NMR spectrum of 5

A 10 mL flask containing a stir bar was charged with aldol product 5 (1.0 mmol), FeCl₂ (0.001 mmol 0.1 mmol%) and 1-pentanol (1.0 mL), respectively. The reaction mixture was stirred at 100 °C under open air for 1 h. Then the reaction mixture was cooled to room temperature. The solvent was evaporated at reduced pressure and the crude product was purified by column chromatography on silica gel (dichloromethane) to gain **4a** in 90% yield.



When the same experiment was conducted in absence of $FeCl_2$, only trace amounts of **4a** was detected in reaction mixture.



In order to further verify whether a free-radical process was involved in the dehydration of aldol product 5, 2, 2, 6, 6-tetramethylpiperdiny-l-oxy (TEMPO) was introduced. The results showed that no 4a was formed during this process. The detailed experimental process is as follows.

A 10 mL flask containing a stir bar was charged with aldol product 5 (1.0 mmol), TEMPO (2.0 mmol),

FeCl₂ (0.001 mmol, 0.1 mmol%) and 1-pentanol (1 mL), respectively. The reaction mixture was stirred at 100 °C in open air for 1 h. Then the reaction mixture was cooled to room temperature. The solvent was evaporated at reduced pressure, and the crude mixture was detected by ¹H-NMR spectrum.



A 10 mL oven-dried flask containing a stir bar was charged with oxindolin-2-one **1a** (1.0 mmol), isatin **2k** (1.0 mmol), FeCl₂ (0.001 mmol, 0.1 mmol%), TEMPO (2.0 mmol) and 1-pentanol (1 mL), respectively. The reaction mixture was stirred at 100 °C in open air for 1 h. Then the reaction mixture was cooled to room temperature. The solvent was evaporated at reduced pressure and the crude product was purified by column chromatography on silica gel (petroleum ether/EtOAc: 5/1) to obtain the radical adduct **6** in 10% yield.



Radical adduct (6). Pale yellow solid. ¹**H NMR** (400 MHz, CDCl₃) δ 7.91 (dd, J = 8.0, 1.3 Hz, 1H), 7.29 (dd, J = 11.6, 4.6 Hz, 1H), 6.73 – 6.55 (m, 2H), 5.77 (s, 1H), 1.82 – 1.62 (m, 4H), 1.58 (d, J = 2.6 Hz, 1H), 1.49 – 1.42 (m, 1H), 1.27 (s, 6H), 1.12 (s, 6H); ¹³**C NMR** (151 MHz, CDCl₃) δ 167.1, 150.1, 133.1, 129.5, 116.0, 115.3, 108.9, 59.5, 38.3, 31.1, 20.1; **IR** (neat): 2930, 1703, 1619, 1589,1487, 1455, 1381, 1231, 1042, 749, 700; **HRMS**(ESI): calcd. for C₁₇H₂₅N₂O₂: [M+H]⁺ m/z 288.1916, found: 288.1901.



Figure S3. ¹H NMR spectrum of 6



Figure S4. ¹³C NMR spectrum of 6

5.3 Control experiment on dehydration mechanism

In order to gain insight of the dehydration mechanism, the corresponding control experiment was conducted. A 10 mL flask containing a stir bar was charged with 7 (1.0 mmol), TEMPO (2.0 mmol), FeCl₂ (0.001 mmol, 0.1 mmol%) and 1-pentanol (1.0 mL), respectively. The reaction mixture was stirred at 100 °C in open air for 1 h. Meanwhile, the reaction process was investigated by high-resolution mass spectrometry every 5 min. To our delight, the radical adduct of **8** was detected, as shown in Figure S5, which further suggested a free radical process could be involved in the process of dehydration.



Figure S5. The HRMS spectrum of control experiment

6. Large-scale synthesis of valuable indole derivatives

6.1 Large-scale synthesis of 2a

A 250 mL round-bottom flask containing a stir bar was charged with indolin-2-ones **1a** (100 mmol, 13.3 g), $FeCl_2$ (1 mmol, 126 mg) and EtOH (100 mL), The reaction mixture was stirred at 60 °C in air for 12 h. Then the reaction mixture was cooled to room temperature. The solvent was evaporated at reduced pressure and the crude product was purified by column chromatography on silica gel (petroleum ether/EtOAc: 3/1) to give red solid 11.3 g in 76% yield.



6.2 Large-scale synthesis of 3a

A 250 mL round-bottom flask containing a stir bar was charged with indolin-2-ones **1a** (100 mmol, 13.3 g), $FeCl_2$ (0.1 mmol, 13 mg) and 1-pentanol (100 mL), respectively. The reaction mixture was stirred at 100 °C in air for 24 h. Then the reaction mixture was cooled to room temperature. The solvent was evaporated at reduced pressure and the crude product was washed by dichloromethane to give dark red solid 11.8 g in 90% yield.



6.3 Large-scale synthesis of 4a

A 500 mL round-bottom flask containing a stir bar was charged with indolin-2-ones **1a** (50 mmol, 6.7 g), 1-methylisatin **2k** (50 mmol, 8.0 g), FeCl₂ (0.05 mmol, 6 mg) and 1-pentanol (100 mL), respectively. The reaction mixture was stirred at 100 °C in argon for 2 h, then stirred at 100 °C in air for 1 h. Then the reaction mixture was cooled to room temperature. The solvent was evaporated at reduced pressure and the crude product was purified by column chromatography on silica gel (dichloromethane) to give dark red solid 10.1 g in 73% yield .



7. Characterization data of isatins

2a

2b

2c

2d

2e

2f

Red solid. ¹**H** NMR (400 MHz, DMSO- d_6) δ 11.04 (s, 1H), 7.58 (t, J = 7.7, 1H), 7.50 (d, J = 7.5 Hz, 1H), 7.06 (t, J = 7.5 Hz, 1H), 6.90 (d, J = 7.9 Hz, 1H); ¹³C NMR (151 MHz, DMSO- d_6) δ 185.0, 160.0, 151.1, 139.1, 125.3, 123.5, 118.2, 112.8. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹

Red solid. ¹**H NMR** (400 MHz, DMSO- d_6) δ 11.04 (s, 1H), 7.52 – 7.37 (m, 2H), 6.91 (dd, J = 8.5, 3.9 Hz, 1H); ¹³**C NMR** (151 MHz, DMSO- d_6) δ 184.1 (d, J = 2.0 Hz), 159.4 (d, J = 92.6 Hz), 157.5, 147.1, 124.7 (d, J = 24.2 Hz), 118.6 (d, J = 7.5 Hz), 113.7 (d, J = 6.0 Hz), 111.6 (d, J = 24 Hz). All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹

Orange-red solid. ¹H NMR (400 MHz, DMSO- d_6) δ 11.13 (s, 1H), 7.61 (dd, J = 8.4, 2.2 Hz, 1H), 7.56 (d, J = 2.1 Hz, 1H), 6.92 (d, J = 8.4 Hz, 1H); ¹³C NMR (151 MHz, DMSO- d_6) δ 183.6, 159.4, 149.4, 137.5, 127.1, 124.3, 119.2, 114.1. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹

Yellow solid. ¹H NMR (400 MHz, DMSO- d_6) δ 11.17 (s, 1H), 7.72 (d, J = 8.4 Hz, 1H), 7.62 (s, 1H), 6.88 (d, J = 8.3 Hz, 1H); ¹³C NMR (101 MHz, DMSO- d_6) δ 183.7, 159.5, 149.9, 140.7, 127.4, 119.9, 114.9, 114.9. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹

Orange solid. ¹H NMR (400 MHz, DMSO- d_6) δ 10.93 (s, 1H), 7.40 (d, J = 7.9 Hz, 1H), 7.32 (s, 1H), 6.81 (d, J = 8.0 Hz, 1H), 2.25 (s, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 184.8, 159.7, 148.7, 139.2, 132.4, 125.1, 117.9, 112.3, 20.3. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹

Orange solid. ¹H NMR (400 MHz, DMSO- d_6) δ 10.84 (s, 1H), 7.18 (dd, J = 8.5, 2.7 Hz, 1H), 7.08 (d, J = 2.5 Hz, 1H), 6.85 (d, J = 8.5 Hz, 1H), 3.75 (s, 3H); ¹³C NMR (151 MHz, DMSO- d_6) δ 184.9, 159.8, 155.5, 144.8, 125.1, 118.3, 113.4, 108.9, 55.9. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹

2g

2i

2i

2k

Orange solid. ¹H NMR (400 MHz, DMSO- d_6) δ 11.66 (s, 1H), 8.44 (dd, J = 8.7, 2.4 Hz, 1H), 8.21 (d, J = 2.3 Hz, 1H), 7.09 (d, J = 8.7 Hz, 1H); ¹³C NMR (101 MHz, DMSO- d_6) δ 182.8, 160.4, 155.6, 143.1, 133.6, 131.5, 120.1, 118.6, 113.0. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹



Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 10.07 (s, 1H), 7.97 (d, J = 8.2 Hz, 2H), 7.92 – 7.82 (m, 2H), 7.70 (d, J = 8.1 Hz, 2H), 7.01 (t, J = 10.5 Hz, 1H), 3.77 (t, J = 7.3 Hz, 2H), 1.73 (dd, J = 14.6, 7.5 Hz, 2H), 1.41 – 1.26 (m, 6H), 0.91 – 0.85 (m, 3H); ¹³**C** NMR (101 MHz, CDCl₃) δ ¹³**C** NMR (101 MHz, CDCl₃) δ 191.7, 183.5, 158.3, 151.1, 145.1, 137.1, 135.7, 135.7, 130.7, 127.2, 124.2, 118.3, 110.9, 77.5, 77.2, 76.8, 40.7, 31.5, 27.4, 26.7, 22.65, 14.12; **IR** (neat cm⁻¹): 2921 1738 1698 1620 1602 1483 1384 1344 1215 1170 820 737; **HRMS**(ESI): calcd. for C₂₁H₂₁NNaO₃: 358.1419 [M+Na]⁺, found: 358.1412.

Yellow solid. ¹H NMR (400 MHz, DMSO- d_6) δ 10.96 (s, 1H), 7.48 (d, J = 8.5 Hz, 1H), 6.59 (d, J = 8.5 Hz, 1H), 6.40 (s, 1H), 3.87 (s, 3H); ¹³C NMR (151 MHz, DMSO- d_6) δ 181.7, 168.0, 160.8, 153.7, 127.5, 111.3, 109.1, 98.0, 56.3. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.²

Yellow solid. ¹**H NMR** (600 MHz, CDCl₃) δ 7.86 (s, 1H), 7.60 (d, *J* = 7.5 Hz, 1H), 7.51 (d, *J* = 8.4 Hz, 1H), 7.18 (t, *J* = 7.9 Hz, 1H); ¹³**C NMR** (151 MHz, CDCl₃) δ 181.6, 158.1, 141.6, 133.7, 131.1, 124.6, 124.4, 121.5, 120.0; **IR** (neat cm⁻¹): 3212 1751 1633 1593 1491 1385 1279 1255 1234 1198 692; **HRMS**(ESI): calcd. for C₉H₃F₃NO₃: 230.0065 [M-H]⁻, found: 230.0065.



Orange solid. ¹H NMR (400 MHz, CDCl₃) δ 7.66 – 7.53 (m, 2H), 7.13 (t, J = 7.5 Hz, 1H), 6.94 – 6.85

(m, 1H), 3.26 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ ¹³C NMR (151 MHz, CDCl₃) δ 183.5, 158.3, 151.6, 138.6, 125.4, 124.0, 117.5, 110.1, 26.4. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹



Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 7.70 (d, J = 7.4 Hz, 1H), 7.60 – 7.51 (m, 3H), 7.46 (t, J = 6.3 Hz, 1H), 7.44 – 7.40 (m, 2H), 7.18 (t, J = 7.5 Hz, 1H), 6.90 (d, J = 8.0 Hz, 1H); ¹³**C** NMR (101 MHz, CDCl₃) δ 183.0, 157.4, 151.7, 138.5, 132.9, 130.1, 128.9, 126.1, 125.7, 124.4, 117.5, 111.4. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹



2m

2n

20

2p

21

Orange solid. ¹H NMR (400 MHz, CDCl₃) δ 7.61 (dd, J = 7.5, 0.8 Hz, 1H), 7.48 (m, 1.3 Hz, 1H), 7.39 – 7.29 (m, 5H), 7.09 (t, J = 7.5 Hz, 1H), 6.77 (d, J = 8.0 Hz, 1H), 4.93 (s, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 183.4, 158.4, 150.8, 138.8, 134.6, 129.1, 128.3, 127.5, 125.5, 124.0, 117.7, 111.1, 44.1. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹



Red solid. ¹**H NMR** (400 MHz, CDCl₃) δ 7.57 (dd, J = 18.4, 7.7 Hz, 2H), 7.11 (t, J = 7.5 Hz, 1H), 6.89 (d, J = 7.9 Hz, 1H), 5.83 (m, 1H), 5.30 (t, J = 12.7 Hz, 2H), 4.36 (d, J = 5.2 Hz, 2H); ¹³**C NMR** (101 MHz, CDCl₃) δ 183.3, 157.9, 150.8, 138.4, 130.4, 125.3, 123.8, 118.6, 117.5, 111.0, 42.5. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹



Orange solid. ¹**H NMR** (400 MHz, CDCl₃) δ 7.67 – 7.62 (m, 2H), 7.18 (t, *J* = 7.6 Hz, 1H), 7.13 (d, *J* = 8.3 Hz, 1H), 4.54 (d, *J* = 2.5 Hz, 2H), 2.31 (t, *J* = 2.5 Hz, 1H); ¹³**C NMR** (151 MHz, CDCl₃) δ 182.7, 157.3, 149.7, 138.6, 125.7, 124.4, 117.8, 111.2, 75.8, 73.5, 29.6. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.³



Orange solid. ¹H NMR (400 MHz, CDCl₃) δ 7.55 (t, J = 7.3 Hz, 2H), 7.08 (t, J = 7.5 Hz, 1H), 6.80 (d, J

= 8.1 Hz, 1H), 3.20 (s, 2H), 0.15 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 183.8, 158.0, 152.0, 138.2, 125.3, 123.6, 118.0, 110.3, 31.8, -1.3. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁴

8. Characterization data of symmetrical isoindigos

3a



Red solid. ¹**H** NMR (400 MHz, DMSO- d_6) δ 10.90 (s, 2H), 9.05 (d, J = 7.1 Hz, 2H), 7.38 – 7.30 (m, 2H), 6.96 (t, J = 7.5 Hz, 2H), 6.84 (d, J = 6.6 Hz, 2H); ¹³C NMR (126 MHz, DMSO- d_6) δ 169.0, 144.1, 133.3, 132.6, 129.3, 121.7, 121.1, 109.5. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹



Red solid. ¹**H NMR** (400 MHz, DMSO- d_6) δ 10.99 (s, 1H), 8.99 (dd, J = 11.3, 2.4 Hz, 1H), 7.24 (td, J = 8.6, 2.4 Hz, 1H), 6.84 (dd, J = 8.5, 4.9 Hz, 1H); ¹³**C NMR** (126 MHz, DMSO- d_6) δ 169.0, 156.1 (d, J = 233.2 Hz), 140.8, 133.9 (d, J = 2.5 Hz), 122.1 (d, J = 10.2 Hz), 119.6 (d, J = 24.2 Hz), 116.3 (d, J = 28.2 Hz), 110.2 (d, J = 8.3 Hz). All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹



Red solid. ¹**H** NMR (400 MHz, DMSO- d_6) δ 11.11 (s, 2H), 9.18 (d, J = 1.7 Hz, 2H), 7.42 (dd, J = 8.3, 2.0 Hz, 2H), 6.87 (d, J = 8.3 Hz, 2H); ¹³C NMR (126 MHz, DMSO- d_6) δ 168.7, 143.1, 133.5, 132.5, 129.0, 125.2, 122.7, 111.1. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁵



Red solid. ¹H NMR (400 MHz, DMSO- d_6) δ 11.06 (s, 2H), 9.31 (s, 2H), 7.54 (d, J = 6.5 Hz, 2H), 6.83 (d, J = 5.7 Hz, 2H). All the resonances in ¹H spectrum were in good agreement with literature values.⁶



Dark red solid. ¹H NMR (400 MHz, DMSO- d_6) δ 10.73 (s, 2H), 8.89 (s, 2H), 7.15 (d, J = 7.9 Hz, 2H), 6.73 (d, J = 7.9 Hz, 2H), 2.26 (s, 6H); ¹³C NMR (101 MHz, DMSO- d_6) δ 168.6, 141.4, 132.8, 132.2, 129.1, 121.5, 108.6, 20.2. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁶



Blue-green solid. ¹H NMR (400 MHz, DMSO- d_6) δ 10.69 (s, 2H), 8.85 (d, J = 2.2 Hz, 2H), 6.97 (dd, J = 8.4, 2.4 Hz, 2H), 6.75 (d, J = 8.5 Hz, 2H), 3.73 (s, 6H); ¹³C NMR (101 MHz, DMSO- d_6) δ 168.6, 153.8, 137.7, 133.5, 122.0, 118.3, 115.3, 109.1, 55.3. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁶



Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.18 (d, J = 8.0 Hz, 2H), 7.35 (t, J = 7.8 Hz, 2H), 7.04 (t, J = 7.8 Hz, 2H), 6.79 (d, J = 7.9 Hz, 2H), 3.77 (t, J = 7.4 Hz, 4H), 1.75 – 1.66 (m, 4H), 1.35 (dt, J = 42.6, 15.9 Hz, 16H), 0.88 (t, J = 6.5 Hz, 6H); ¹³**C** NMR (151 MHz, CDCl₃) δ 168.0, 144.9, 133.7, 132.5, 130.1, 122.3, 121.9, 108.0, 40.2, 31.7, 27.6, 26.9, 22.7, 14.2; **IR** (neat cm⁻¹): 2921 1733 1694 1609 1468 1384 1356 1099 776 748; **HRMS**(ESI): calcd. for C₂₈H₃₃N₄O₆; 521.2400 [M+H]⁺, found: 521.2332.



Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.45 (s, 2H), 7.59 (d, J = 8.5 Hz, 4H), 7.53 (d, J = 8.1 Hz, 2H), 6.99 (d, J = 8.5 Hz, 4H), 6.81 (d, J = 8.1 Hz, 2H), 3.86 (s, 6H), 3.77 (dd, J = 15.1, 7.7 Hz, 4H), 1.70 (dd, J = 14.8, 7.5 Hz, 4H), 1.46 – 1.23 (m, 12H), 0.89 (t, J = 6.7 Hz, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 167.9, 159.0, 143.7, 135.1, 134.0, 133.8, 130.7, 128.5, 128.0, 122.2, 114.4, 108.1, 55.5, 40.2, 31.7, 27.6, 26.8, 22.7, 14.2. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁷



Red solid. ¹**H** NMR (600 MHz, CDCl₃) δ 9.49 (s, 2H), 7.59 (t, J = 7.7 Hz, 6H), 7.48 (d, J = 8.2 Hz, 4H), 6.84 (d, J = 8.1 Hz, 2H), 3.80 (t, J = 7.4 Hz, 4H), 1.77 – 1.65 (m, 4H), 1.41 (dd, J = 14.3, 6.7 Hz, 4H), 1.37 (s, 18H), 1.35 – 1.29 (m, 8H), 0.89 (t, J = 6.8 Hz, 6H); ¹³**C** NMR (101 MHz, CDCl₃) δ 168.0, 149.9, 144.1, 138.4, 135.4, 134.1, 131.1, 128.9, 126.7, 125.9, 122.3, 108.2, 40.3, 34.7, 31.7, 31.6, 27.7, 26.9, 22.7, 14.2; **IR** (neat cm⁻¹): 2922 1694 1609 1481 1384 1333 1116 811 743; **HRMS**(ESI): calcd. for C₄₈H₅₉N₂O_{2:} 695.4577 [M+H]⁺, found: 695.4561.



Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 10.06 (s, 2H), 9.63 (d, J = 1.6 Hz, 2H), 7.98 (d, J = 8.2 Hz, 4H), 7.83 (d, J = 8.2 Hz, 4H), 7.68 (dd, J = 8.2, 1.7 Hz, 2H), 6.92 (d, J = 8.2 Hz, 2H), 3.90 – 3.75 (m, 4H), 1.80 – 1.67 (m, 4H), 1.37 (dd, J = 28.9, 9.4 Hz, 12H), 0.89 (t, J = 6.9 Hz, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 191.9, 167.8, 147.0, 145.1, 134.9, 133.9, 133.8, 131.5, 130.4, 129.2, 127.3, 122.2, 108.4, 40.3, 31.5, 27.5, 26.7, 22.6, 14.0; **IR** (neat cm⁻¹): 2922 1687 1601 1482 1385 1349 1319 1273 1224 1166 1113 809; **HRMS**(ESI): calcd. for C₄₂H₄₃N₂O₄: 639.3223 [M+H]⁺, found: 639.3212.



Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.58 (s, 2H), 7.75 (d, J = 8.1 Hz, 4H), 7.68 (dd, J = 15.1, 8.1 Hz, 10H), 7.47 (t, J = 7.6 Hz, 4H), 7.36 (t, J = 7.6 Hz, 2H), 6.89 (d, J = 8.0 Hz, 2H), 3.84 (t, J = 7.2 Hz, 4H), 1.74 (d, J = 7.5 Hz, 4H), 1.35 (t, J = 34.6 Hz, 16H), 0.90 (t, J = 6.6 Hz, 6H); ¹³C NMR (151 MHz, CDCl₃) δ 167.9, 144.2, 140.9, 140.0, 139.7, 134.8, 134.0, 131.0, 128.8, 128.7, 127.6, 127.2, 127.0, 122.2, 108.2, 40.2, 39.8, 31.6, 27.5, 26.7, 22.6, 14.1; **IR** (neat cm⁻¹): 2928 1693 1609 1487 1385 1110 810 770; **HRMS**(ESI): calcd. for C₅₂H₅₁N₂O₂: 735.3951 [M+H]⁺, found: 735.3942.



Red solid. ¹H NMR (400 MHz, CDCl₃) δ 9.55 (s, 1H), 8.91 (s, 1H), 8.58 (d, J = 3.8 Hz, 1H), 7.96 (d, J

31

S15

= 7.3 Hz, 1H), 7.61 (d, J = 7.4 Hz, 1H), 7.35 (t, J = 24.0 Hz, 1H), 6.91 (d, J = 7.9 Hz, 1H), 3.81 (t, J = 6.6 Hz, 2H), 1.73 (s, 2H), 1.38 (d, J = 33.4 Hz, 6H), 0.89 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.0, 156.7, 143.9, 133.8, 132.5, 131.0, 130.9, 130.8, 128.4, 121.7, 121.1, 111.4, 107.5, 40.2, 31.7, 27.7, 26.8, 22.7, 14.2. **IR** (neat cm⁻¹): 2951 1687 1609 1747 1396 1349 1177 1113 826 794; **HRMS**(ESI): calcd. for C₃₈H₄₁N₄O₂: 585.3230 [M+H]⁺, found: 585.3216.



3m

Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.59 (d, J = 1.7 Hz), 7.60 (dd, J = 8.1, 1.8 Hz), 7.34 (dd, J = 3.5, 1.0 Hz), 7.24 (dd, J = 5.1, 0.9 Hz), 7.09 (dd, J = 5.1, 3.6 Hz), 6.79 (d, J = 8.2 Hz), 3.80 (t, J = 7.4 Hz), 1.77 – 1.65 (m), 1.45 – 1.27 (m), 0.89 (t, J = 7.0 Hz); ¹³C NMR (101 MHz, CDCl₃) δ 167.9, 144.7, 144.3, 134.0, 130.2, 129.0, 128.1, 128.0, 124.1, 122.8, 122.3, 108.2, 40.3, 31.7, 27.6, 26.8, 22.7, 14.2; IR (neat cm⁻¹): 2921 1686 1607 1385 1352 1114 801 682; HRMS(ESI): calcd. for C₃₆H₃₉N₂O₂S_{2:} 595.2453 [M+H]⁺, found: 595.2444.



Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.65 (s, 2H), 8.10 (s, 2H), 7.94 (d, J = 8.1 Hz, 4H), 7.86 (dd, J = 13.8, 8.2 Hz, 4H), 7.73 (d, J = 7.7 Hz, 2H), 7.50 (t, J = 7.2 Hz, 4H), 6.90 (d, J = 8.0 Hz, 2H), 3.83 (s, 4H), 1.75 (s, 4H), 1.39 (d, J = 35.7 Hz, 12H), 0.89 (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 167.9, 144.3, 138.5, 135.3, 132.5, 131.4, 129.1, 128.5, 128.2, 127.6, 126.2, 125.7, 125.6, 125.2, 122.3, 108.2, 40.2, 31.5, 27.5, 26.7, 22.6, 14.0; **IR** (neat cm⁻¹): 2920 1697 1636 1558 1458 1385 1075 761; **HRMS**(ESI): calcd. for C₄₈H₄₇N₂O₂: 683.3638 [M+H]⁺, found: 683.3622.



Red solid. ¹**H** NMR (400 MHz, DMSO- d_6) δ 10.79 (s, 2H), 9.00 (d, J = 6.3 Hz, 2H), 6.45 (d, J = 58.2 Hz, 4H), 3.81 (s, 6H); ¹³**C** NMR (101 MHz, DMSO- d_6) δ 170.0, 162.4, 145.7, 130.8, 129.7, 115.1, 106.6, 95.4, 55.4. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁷



3p

3q

3r

3s

Red solid. ¹**H** NMR (400 MHz, DMSO- d_6) δ 11.65 (s, 2H), 9.06 (d, J = 8.1 Hz, 2H), 7.43 (d, J = 8.3 Hz, 2H), 7.09 (t, J = 8.2 Hz, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 168.5, 137.3, 133.6, 131.1, 128.5, 125.6, 123.9, 121.6, 120.3 (d, J = 257.6 Hz). All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁷



Red solid. ¹H NMR (400 MHz, CDCl₃) δ 9.22 (d, J = 7.4 Hz, 2H), 7.38 (td, J = 7.7, 1.1 Hz, 2H), 7.07 (td, J = 7.9, 1.0 Hz, 2H), 6.79 (d, J = 7.7 Hz, 2H), 3.29 (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 168.1, 145.3, 133.5, 132.5, 130.0, 122.5, 121.7, 107.8, 26.2. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹



Red solid. ¹H NMR (400 MHz, CDCl₃) δ 9.23 (d, J = 7.4 Hz, 2H), 7.36 – 7.26 (m, 12H), 7.09 – 7.01 (m, 2H), 6.72 (d, J = 7.8 Hz, 2H), 5.02 (s, 4H); ¹³C NMR (101 MHz, CDCl₃) δ 168.0, 144.5, 135.7, 133.5, 132.5, 130.0, 128.8, 127.7, 127.2, 122.5, 121.7, 108.7, 43.7. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹



Red solid. ¹H NMR (400 MHz, CDCl₃) δ 9.22 (d, J = 8.0 Hz, 2H), 7.57 (dd, J = 9.9, 5.6 Hz, 4H), 7.49 – 7.42 (m, 6H), 7.30 – 7.24 (m, 2H), 7.08 – 7.01 (m, 2H), 6.74 (d, J = 7.8 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 167.6, 145.5, 134.3, 133.9, 132.7, 130.4, 129.9, 128.5, 127.3, 123.0, 121.7, 109.2. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹



3t

3u

3v

3w

Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.20 (d, J = 8.0 Hz, 2H), 7.34 (t, J = 7.6 Hz, 2H), 7.06 (t, J = 7.6 Hz, 2H), 6.79 (d, J = 7.8 Hz, 2H), 5.88 (m, 2H), 5.25 (t, J = 13.0 Hz, 4H), 4.44 (d, J = 5.1 Hz, 4H); ¹³**C** NMR (101 MHz, CDCl₃) δ 167.8, 144.6, 133.5, 132.5, 131.3, 130.1, 122.5, 121.7, 117.7, 108.6, 42.4. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁷



Red solid. ¹**H** NMR (400 MHz, DMSO- d_6) δ 9.08 (d, J = 8.0 Hz, 2H), 7.48 (t, J = 7.7 Hz, 2H), 7.14 – 7.06 (m, 4H), 4.63 (d, J = 2.1 Hz, 4H), 3.27 (s, 2H); ¹³C NMR (151 MHz, DMSO- d_6) δ 166.4, 143.3, 133.1, 132.6, 129.3, 122.4, 120.8, 109.3, 77.9, 74.5, 29.0. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁷



Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.19 (d, J = 8.0 Hz, 1H), 7.34 (t, J = 7.7 Hz, 1H), 7.04 (t, J = 7.8 Hz, 1H), 6.72 (d, J = 7.8 Hz, 1H), 3.32 (s, 2H), 0.16 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 166.6, 144.6, 132.6, 131.2, 128.6, 121.3, 121.1, 107.3, 30.6, -2.2. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁶



Red solid. ¹**H NMR** (600 MHz, DMSO- d_6) δ 11.59 (s, 2H), 9.31 (d, J = 7.7 Hz, 2H), 8.21 (d, J = 4.7 Hz, 2H), 7.12 – 7.03 (m, 2H); ¹³**C NMR** (151 MHz, DMSO- d_6) δ 168.7, 168.7, 158.1, 150.6, 136.8, 132.5, 118.0, 116.1. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁷

9. Characterization data of non-symmetrical isoindigos



Dark red solid. ¹H NMR (400 MHz, DMSO- d_6) δ 10.89 (s, 1H), 9.07 (d, J = 7.7 Hz, 2H), 7.41 (t, J = 7.4 Hz, 1H), 7.34 (t, J = 7.4 Hz, 1H), 7.06 – 6.92 (m, 3H), 6.83 (d, J = 7.6 Hz, 1H), 3.21 (s, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 168.9, 167.2, 145.0, 144.2, 133.8, 132.9, 132.6, 132.3, 129.5, 129.0, 121.8, 121.7, 121.2, 120.8, 109.6, 108.5, 26.1. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁵



Dark red solid. ¹H NMR (400 MHz, DMSO- d_6) δ 10.89 (s, 1H), 9.08 (d, J = 7.9 Hz, 1H), 8.94 (dd, J = 11.4, 2.4 Hz, 1H), 7.41 (t, J = 7.6 Hz, 1H), 7.18 (td, J = 8.7, 2.5 Hz, 1H), 7.04 – 6.90 (m, 2H), 6.78 (dd, J = 8.5, 4.9 Hz, 1H), 3.18 (s, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 168.7, 167.2, 158.1, 155.8, 145.2, 140.6, 133.4, 133.0, 129.3, 122.3, 121.9, 120.6, 119.1, 118.9, 116.2, 115.9, 110.0, 110.0, 108.5, 26.0. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁸



Red solid. ¹H NMR (400 MHz, DMSO- d_6) δ 11.06 (s, 1H), 9.20 (s, 1H), 9.10 (d, J = 8.0 Hz, 1H), 7.49 - 7.33 (m, 2H), 7.04 (dd, J = 11.5, 7.8 Hz, 2H), 6.85 (d, J = 8.3 Hz, 1H), 3.22 (s, 3H); ¹³C NMR (151 MHz, DMSO- d_6) δ 168.6, 167.2, 145.3, 142.9, 133.7, 133.2, 132.4, 132.0, 129.3, 128.7, 125.1, 122.9, 121.9, 120.6, 110.9, 108.6, 26.1. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁵



Red solid. ¹**H** NMR (400 MHz, DMSO-*d*₆) δ 11.07 (s, 1H), 9.35 (d, J = 2.0 Hz, 1H), 9.11 (d, J = 7.8 Hz, 1H), 7.53 (dd, J = 8.3, 2.1 Hz, 1H), 7.48 – 7.43 (m, 1H), 7.08 – 7.02 (m, 2H), 6.82 (d, J = 8.3 Hz, 1H), 3.23 (s, 3H); ¹³**C** NMR (151 MHz, DMSO-*d*₆) δ 168.5, 167.3, 145.4, 143.3, 134.9, 133.8, 133.3, 132.3,

131.5, 129.3, 123.4, 122.0, 120.7, 112.9, 111.5, 108.7, 26.2. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁸



Red solid. ¹**H** NMR (400 MHz, DMSO- d_6) δ 10.79 (s, 1H), 9.08 (d, J = 7.0 Hz, 1H), 7.50 – 7.30 (m, 1H), 7.23 – 6.95 (m, 3H), 6.72 (d, J = 6.5 Hz, 1H), 3.22 (s, 3H), 2.27 (s, 3H); ¹³C NMR (151 MHz, DMSO- d_6) δ 168.9, 167.2, 144.9, 142.0, 134.1, 133.3, 132.4, 132.0, 129.8, 129.7, 128.9, 121.7, 120.8, 109.3, 108.4, 26.0, 21.0; **IR** (neat cm⁻¹): 2927 1688 1610 1472 1375 1343 1159 1095 814 772 751; **HRMS**(ESI): calcd. for C₁₈H₁₅N₂O₂: 291.1134 [M+H]⁺, found: 291.1130.



Red solid. ¹**H** NMR (400 MHz, DMSO- d_6) δ 10.72 (s, 1H), 9.11 (d, J = 7.7 Hz, 1H), 8.86 (d, J = 2.5 Hz, 1H), 7.43 (td, J = 7.7, 1.0 Hz, 1H), 7.03 (t, J = 8.4 Hz, 2H), 6.97 (dd, J = 8.5, 2.6 Hz, 1H), 6.74 (d, J = 8.5 Hz, 1H), 3.74 (s, 3H), 3.23 (s, 3H); ¹³**C** NMR (151 MHz, DMSO- d_6) δ 168.9, 167.3, 154.0, 145.0, 138.2, 134.4, 132.6, 132.4, 129.0, 122.3, 121.8, 120.8, 118.8, 115.2, 109.8, 108.5, 55.5, 26.1. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁸



Red solid. ¹**H** NMR (400 MHz, DMSO-*d*₆) δ 11.84 – 11.44 (m, 1H), 10.13 (d, J = 2.3 Hz, 1H), 9.11 (d, J = 7.7 Hz, 1H), 8.29 (dd, J = 8.7, 2.3 Hz, 1H), 7.48 (t, J = 7.7 Hz, 1H), 7.14 – 6.97 (m, 3H), 3.79 (t, J = 7.0 Hz, 2H), 1.63 (s, 2H), 1.30 (s, 6H), 0.85 (t, J = 6.9 Hz, 3H); ¹³**C** NMR (151 MHz, DMSO-*d*₆) δ 169.5, 167.6, 150.0, 145.5, 142.0, 135.3, 134.3, 131.7, 130.1, 128.9, 125.5, 122.5, 121.9, 121.0, 110.2, 109.4, 31.4, 31.2, 27.3, 26.4, 22.5, 19.7, 14.3; **IR** (neat cm⁻¹): 2929 1697 1626 1529 1401 1339 1164 771; **HRMS**(ESI): calcd. for C₂₂H₂₂N₃O₄: 392.1610 [M+H]⁺, found: 392.1607.



4h

Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.07 (t, J = 8.3 Hz, 2H), 7.80 (s, 1H), 7.31 (dd, J = 16.0, 7.8 Hz, 2H), 7.00 (dt, J = 34.4, 7.9 Hz, 2H), 6.79 (d, J = 7.8 Hz, 1H), 4.26 – 4.10 (m, 2H), 1.74 (s, 2H), 1.33

(s, 4H), 0.89 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 169.2, 168.1, 142.7, 140.5, 134.8, 134.5, 133.2, 132.7, 130.4, 128.1, 124.6, 122.9, 122.6, 122.6, 114.8, 109.5, 42.2, 31.6, 29.7, 26.6, 22.7, 14.2; **IR** (neat cm⁻¹): 2926 1728 1691 1620 1484 1467 1384 1332 1102 804 745; **HRMS**(ESI): calcd. for C₂₂H₂₀ClN₂O₂: 379.1213 [M-H]⁻, found: 379.1219.



4i

Red solid. ¹**H** NMR (400 MHz, DMSO- d_6) δ 11.59 (s, 1H), 10.96 (s, 1H), 9.11 – 8.98 (m, 2H), 7.48 – 7.25 (m, 2H), 7.05 (t, J = 8.2 Hz, 1H), 7.01 – 6.95 (m, 1H), 6.84 (d, J = 7.5 Hz, 1H); ¹³C NMR (151 MHz, DMSO- d_6) δ 168.8, 144.6, 137.0, 135.2, 133.4, 131.8, 131.1, 129.7, 128.2, 125.1, 124.2, 121.5, 121.4, 121.3, 119.6, 109.8; **IR** (neat cm⁻¹): 3084 1697 1637 1620 1489 1465 1325 1255 1201 1176 789; **HRMS**(ESI): calcd. for C₁₇H₈F₃N₂O₃: 345.0487 [M-H]⁻, found: 345.0491.



Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.53 (d, J = 1.7 Hz, 1H), 9.29 (s, 1H), 9.19 (d, J = 8.0 Hz, 1H), 7.63 – 7.58 (m, 3H), 7.46 (d, J = 8.4 Hz, 2H), 7.26 (m, 1H), 7.08 – 6.98 (m, 1H), 6.88 – 6.75 (m, 2H), 3.76 (t, J = 7.4 Hz, 2H), 1.72 (dt, J = 15.0, 7.6 Hz, 2H), 1.41 – 1.33 (m, 12H), 0.89 (t, J = 7.0 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 170.6, 167.9, 150.0, 144.0, 143.0, 138.2, 135.1, 134.2, 133.9, 132.6, 131.0, 130.4, 128.9, 126.5, 125.9, 122.6, 122.5, 122.1, 109.7, 108.2, 40.4, 34.6, 31.7, 31.5, 27.6, 26.9, 22.7, 14.2; **IR** (neat cm⁻¹): 2922 1691 1618 1608 1482 1385 1353 1333 1116 1020 811 744; **HRMS**(ESI): calcd. for C₃₂H₃₅N₂O₂: 479.2699 [M+H]⁺, found: 479.2702.



4k

Red solid. ¹**H** NMR (400 MHz, DMSO- d_6) δ 10.97 (s, 1H), 9.17 (d, J = 8.0 Hz, 1H), 8.98 (d, J = 8.0 Hz, 1H), 7.66 – 7.57 (m, 2H), 7.55 – 7.47 (m, 3H), 7.36 (dd, J = 13.6, 7.1 Hz, 2H), 7.10 (t, J = 7.8 Hz, 1H), 6.95 (t, J = 7.8 Hz, 1H), 6.86 (d, J = 7.8 Hz, 1H), 6.69 (d, J = 7.9 Hz, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 169.3, 167.7, 145.5, 142.6, 134.3, 134.0, 133.5, 132.9, 132.6, 130.6, 130.0, 129.9, 128.6, 127.3, 123.0, 122.6, 122.6, 121.7, 109.4, 109.2. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁵



Red solid. ¹H NMR (400 MHz, DMSO- d_6) δ 10.95 (s, 1H), 9.11 (dd, J = 7.9, 4.6 Hz, 2H), 7.42 – 7.30 (m, 6H), 7.30 – 7.22 (m, 1H), 7.00 (m, 3H), 6.85 (d, J = 7.8 Hz, 1H), 5.02 (s, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 168.8, 167.4, 144.4, 143.9, 136.3, 134.3, 133.1, 132.5, 131.8, 129.6, 129.2, 128.7, 127.5, 127.3, 121.9, 121.6, 121.2, 120.9, 109.7, 109.0, 42.7. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁵



4m

4n

Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.19 (d, J = 8.0 Hz, 1H), 9.09 (d, J = 8.0 Hz, 1H), 7.81 (s, 1H), 7.32 (dt, J = 11.5, 7.7 Hz, 2H), 7.04 (dd, J = 17.4, 8.1 Hz, 2H), 6.81 (d, J = 7.7 Hz, 1H), 6.71 (d, J = 7.8 Hz, 1H), 3.30 (s, 2H), 0.14 (s, 9H); ¹³**C** NMR (101 MHz, CDCl₃) δ 169.6, 167.4, 145.7, 142.4, 134.2, 133.1, 132.5, 132.5, 130.1, 129.6, 122.7, 122.6, 122.3, 121.9, 109.4, 108.3, 31.5, -1.2; **IR** (neat cm⁻¹): 2916 1691 1608 1464 1384 1354 1330 1103 1049 855 742; **HRMS**(ESI): calcd. for C₂₀H₂₁N₂O₂Si: 349.1372 [M+H]⁺, found: 349.1367.



Red solid. ¹**H** NMR (400 MHz, DMSO- d_6) δ 10.91 (s, 1H), 9.07 (dd, J = 16.7, 7.5 Hz, 2H), 7.42 (td, J = 7.7, 1.1 Hz, 1H), 7.34 (m, 1H), 7.08 (d, J = 7.7 Hz, 1H), 7.06 – 7.00 (m, 1H), 6.99 – 6.94 (m, 1H), 6.84 (d, J = 7.5 Hz, 1H), 3.79 (t, J = 7.0 Hz, 2H), 3.54 (s, 3H), 2.41 (t, J = 7.2 Hz, 2H), 1.88 (m, 2H); ¹³C NMR (151 MHz, DMSO- d_6) δ 172.9, 168.9, 167.2, 144.3, 144.2, 133.9, 132.9, 132.6, 132.2, 129.5, 129.1, 121.7, 121.7, 121.2, 120.9, 109.6, 108.5, 51.3, 38.6, 30.5, 22.3; **IR** (neat cm⁻¹): 2916 1728 1693 1608 1464 1384 1366 1329 1199 1107 776; **HRMS**(ESI): calcd. for C₂₁H₁₉N₂O₄: 363.1345 [M+H]⁺, found: 363.1342.



Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.20 (td, J = 8.0, 0.7 Hz, 2H), 7.34 (m, 2H), 7.12 – 6.99 (m, 2H), 6.77 (d, J = 7.8 Hz, 2H), 5.88 (m, 1H), 5.31 – 5.15 (m, 2H), 4.41 (dt, J = 5.2, 1.6 Hz, 2H), 3.27 (d, J = 2.2 Hz, 3H); ¹³**C** NMR (101 MHz, CDCl₃) δ 168.1, 167.8, 145.4, 144.5, 133.7, 133.3, 132.6, 132.4, 131.3, 130.1, 130.0, 122.5, 122.5, 121.7, 121.7, 117.7, 108.5, 107.8, 42.4, 26.3; **IR** (neat cm⁻¹): 3282 1692 1608 1468 1358 1108 772; **HRMS**(ESI): calcd. for C₂₀H₁₇N₂O_{2:} 317.1290 [M+H]⁺, found: 317.1285.



4p

Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.14 (d, J = 8.1 Hz, 1H), 8.94 (d, J = 8.0 Hz, 1H), 7.82 (d, J = 8.4 Hz, 2H), 7.43 (t, J = 7.9 Hz, 1H), 7.32 (t, J = 7.6 Hz, 1H), 7.18 (t, J = 7.8 Hz, 1H), 7.04 (t, J = 7.8 Hz, 1H), 6.80 (d, J = 7.8 Hz, 1H), 1.68 (s, 9H); ¹³**C** NMR (101 MHz, CDCl₃) δ 169.0, 166.2, 148.9, 142.8, 141.1, 136.3, 134.1, 133.1, 133.0, 130.4, 129.1, 124.2, 122.6, 122.4, 122.0, 114.3, 109.6, 84.9, 28.3; **IR** (neat cm⁻¹): 2920 1698 1619 1462 1385 1254 1099 774 742; **HRMS**(ESI): calcd. for C₂₁H₁₈N₂NaO₄; 385.1164 [M+Na]⁺, found: 385.1159.



Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.22 (m, 2H), 7.91 – 7.85 (m, 2H), 7.78 – 7.72 (m, 2H), 7.42 – 7.34 (m, 2H), 7.28 (s, 1H), 7.13 – 7.04 (m, 2H), 6.80 (t, *J* = 6.0 Hz, 1H), 5.77 (d, *J* = 5.6 Hz, 2H), 3.79 – 3.74 (m, 2H), 1.71 (dd, *J* = 14.6, 7.1 Hz, 2H), 1.35 – 1.31 (m, 4H), 0.91 – 0.87 (m, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 167.7, 167.5, 144.9, 142.8, 134.6, 134.5, 134.4, 132.7, 132.7, 132.4, 131.8, 130.4, 130.0, 123.9, 123.0, 122.4, 121.8, 121.7, 108.9, 107.9, 42.4, 40.2, 31.6, 27.5, 26.8, 22.6, 14.1; **IR** (neat cm⁻¹): 2928 1725 1694 1609 1467 1328 1098 776 749; **HRMS**(ESI): calcd. for C₃₁H₂₈N₃O₄: 506.2080 [M+H]⁺, found: 506.2074.



4r

4s

Red solid. ¹**H** NMR (400 MHz, DMSO- d_6) δ 11.53 (s, 1H), 9.32 (d, J = 7.2 Hz, 1H), 9.10 (d, J = 7.1 Hz, 1H), 8.18 (s, 1H), 7.45 (s, 1H), 7.03 (s, 3H), 3.21 (s, 3H); ¹³**C** NMR (151 MHz, DMSO- d_6) δ 168.5, 167.2, 157.7, 150.1, 145.4, 136.6, 133.6, 133.2, 131.4, 129.3, 122.0, 120.5, 117.7, 116.2, 108.6, 26.1. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁸



Red solid. ¹**H** NMR (600 MHz, CDCl₃) δ 9.32 – 9.24 (m, 1H), 9.03 (d, J = 8.0 Hz, 1H), 7.50 – 7.43 (m, 1H), 7.43 – 7.36 (m, 1H), 7.24 – 7.19 (m, 1H), 7.15 – 7.02 (m, 2H), 6.80 (dd, J = 15.3, 7.9 Hz, 1H), 3.29 (d, J = 5.3 Hz, 3H); ¹³**C** NMR (151 MHz, CDCl₃) δ 167.6, 167.6, 155.6, 145.9, 135.5, 133.7, 133.5, 130.1, 130.0, 127.3, 124.3, 122.9, 110.6, 108.2, 26.3. **IR** (neat cm⁻¹): 3132 1779 1699 1613 1474 1457 1326 1078 774 744; **HRMS**(ESI): calcd. for C₂₀H₁₇N₂O₂: 278.0817 [M+H]⁺, found: 278.0811.



Red solid. ¹**H** NMR (600 MHz, DMSO- d_6) δ 10.87 (s, 1H), 9.07 (dd, J = 30.7, 8.4 Hz, 2H), 7.39 (t, J = 7.7 Hz, 1H), 7.02 (dd, J = 13.3, 7.7 Hz, 2H), 6.54 (dd, J = 9.0, 2.5 Hz, 1H), 6.39 (d, J = 2.4 Hz, 1H), 3.83 (s, 3H), 3.23 (s, 3H); ¹³**C** NMR (151 MHz, DMSO- d_6) δ 169.7, 167.5, 163.3, 146.6, 144.2, 133.5, 131.6, 128.6, 128.3, 121.6, 121.0, 114.8, 108.3, 106.9, 95.6, 55.6, 26.0. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁸



4u

Red solid. ¹**H NMR** (400 MHz, DMSO- d_6) δ 10.97 (s, 1H), 9.32 (d, J = 7.8 Hz, 1H), 9.11 (d, J = 8.0 Hz, 1H), 8.25 (d, J = 4.9 Hz, 1H), 7.38 (t, J = 7.6 Hz, 1H), 7.11 – 7.05 (m, 1H), 6.99 (d, J = 7.5 Hz, 1H), 6.86 (d, J = 7.7 Hz, 1H), 3.82 (t, J = 7.0 Hz, 2H), 1.68 (dd, J = 14.7, 7.3 Hz, 2H), 1.33 (dd, J = 14.0, 6.7 Hz, 2H), 0.92 (t, J = 6.9 Hz, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 168.9, 167.0, 156.7, 149.5, 144.7, 136.3, 135.2, 133.5, 129.8, 129.6, 121.4, 121.3, 118.0, 115.7, 109.8, 29.2, 19.6, 13.6. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁹



4v

Red solid. ¹**H** NMR (400 MHz, CDCl₃) δ 9.18 (d, J = 8.2 Hz, 1H), 9.10 (d, J = 8.0 Hz, 1H), 7.61 (d, J = 11.1 Hz, 1H), 7.36 – 7.26 (m, 4H), 7.22 – 7.18 (m, 3H), 7.05 (ddd, J = 7.9, 5.6, 1.1 Hz, 2H), 6.81 (d, J = 7.3 Hz, 1H), 6.69 (d, J = 7.5 Hz, 1H), 3.88 – 3.80 (m, 2H), 2.80 – 2.71 (m, 2H), 2.11 – 2.03 (m, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 169.5, 167.9, 144.6, 142.5, 141.0, 133.8, 133.4, 132.5, 132.5, 130.1, 129.9, 128.5, 128.3, 126.1, 122.5, 122.4, 122.3, 121.7, 109.4, 107.9, 39.7, 33.3, 28.9. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.⁸



Red solid. ¹**H NMR** (600 MHz, CDCl₃) δ 9.14 (d, *J* = 8.1 Hz, 1H), 9.02 (d, *J* = 8.1 Hz, 1H), 7.57 (s, 1H), 7.40 (t, *J* = 7.7 Hz, 1H), 7.33 (t, *J* = 7.6 Hz, 1H), 7.19 (d, *J* = 7.5 Hz, 1H), 7.12 (t, *J* = 7.8 Hz, 1H), 7.05 (t, *J* = 7.8 Hz, 1H), 6.81 (d, *J* = 7.7 Hz, 1H), 5.80 (d, *J* = 69.0 Hz, 2H), 5.42 (t, *J* = 9.4 Hz, 1H), 5.36 – 5.24 (m, 1H), 4.29 – 4.20 (m, 2H), 4.02 – 3.86 (m, 1H), 2.10 (d, *J* = 6.8 Hz, 6H), 2.02 (s, 3H), 1.86 (s, 3H); ¹³**C NMR** (151 MHz, CDCl₃) δ 170.7, 170.2, 169.7, 169.3, 169.1, 167.7, 142.8, 134.4, 133.1, 132.8, 132.3, 130.1, 129.8, 123.3, 122.5, 122.5, 121.7, 109.6, 79.7, 74.9, 73.7, 68.1, 67.7, 61.9, 31.1, 20.9, 20.8, 20.7, 20.5. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹⁰



Red solid. ¹H NMR (400 MHz, DMSO- d_6) δ 11.56 (s, 1H), 9.15 (t, J = 9.0 Hz, 2H), 8.20 (d, J = 4.5 Hz, 1H), 7.49 (d, J = 6.9 Hz, 2H), 7.09 (dt, J = 12.7, 7.2 Hz, 2H), 6.09 (s, 1H), 5.58 (d, J = 9.0 Hz, 2H), 5.32 (s, 1H), 4.37 (s, 1H), 4.15 (s, 2H), 2.03 (d, J = 8.9 Hz, 6H), 1.97 (d, J = 9.8 Hz, 3H), 1.77 (s, 3H); ¹³C NMR (151 MHz, DMSO- d_6) δ 170.1, 169.6, 169.4, 168.9, 168.3, 167.0, 157.9, 150.5, 136.5, 133.2, 132.5, 132.2, 129.1, 122.6, 120.7, 117.8, 116.1, 78.4, 73.1, 72.6, 67.8, 67.3, 61.9, 20.5, 20.5, 20.3, 20.0. All the resonances in ¹H and ¹³C NMR spectrum were in good agreement with literature values.¹¹

10. References

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11. NMR spectrum









































































































S78













S84











