

Supporting Information

**Regioselective Synthesis of 4-Arylamino-1,2-Naphthoquinones in
Eutectogel as a Confined Reaction Media using LED Light**

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Experimental Section

1. General materials and methods

All reagents and solvents used for the synthesis of 4-(hydroxymethyl)-1,3:2,4-dibenzylidene-D-sorbitol and 4-arylamino-1,2-naphthoquinones were purchased from commercial suppliers such as Merck, Aldrich, Finar, SRL and Avra chemicals and were used without purification. For the compound purification and recrystallization, LR-grade solvents were employed. AR-grade solvents were used for the gelation studies. The pre-coated Merck silica gel plates (TLC Silica gel 60 F₂₅₄) were used to monitor the reaction progress and visualized the spots using any one or the combination of the following visualizing agents such as UV detection, KMnO₄, p-anisaldehyde, H₂SO₄ spray, or molecular iodine.

2. NMR and HRMS

¹H and ¹³C NMR spectra were recorded on a Bruker Avance 400 MHz instrument either in CDCl₃ or DMSO-d₆ at room temperature. Chemical shifts (δ) are reported in parts per million (ppm) with reference to the internal standard TMS and coupling constants (J) are given in Hz. Proton multiplicity is assigned using the following abbreviations: singlet (s), doublet (d), triplet (t), quartet (q), and multiplet (m). High-resolution MS analysis were performed on an Agilent 6520 Q-TOF instrument by dissolving the solid sample in methanol or any other suitable solvent.

3. Gelation Studies

To a known quantity of HMDBS taken in a glass vial, an appropriate amount of solvent was added and sealed tightly, and the system was heated until the solid was completely dissolved. The homogeneous solution thus obtained is allowed to cool slowly to rt. The gelation ability of HMDBS can be observed by naked eye by inverting the test tube. In the inverted test tube, a sample exhibiting no gravitational flow is referred as gel “G”, instead it remains as solution is referred as “S”. If the compound is insoluble, it is referred as “I”. Precipitation and crystallization observed in the test tube is denoted as “P” and “Crys” respectively.

4. Morphological study

The morphology of the gels was investigated using optical microscopy and scanning electron microscopy. The gel was prepared at a critical gelation concentration and placed over a glass plate and viewed using a Carl Zeiss Axio Scope A1 fluorescent/phase-contrast microscope. The xerogel was used for SEM analysis, which

is prepared by taking a suitable quantity of eutectogel and disperse in water. DES in the 3D fibrous network is exchanged slowly with water and the resultant gel is dried and used.

5. Rheological measurements

The viscoelastic behaviour of the gels was identified using a stress-controlled rheometer (Anton Paar 302 rheometer) equipped with a steel-coated parallel plate geometry of 25 mm diameter. The rheological behaviour of the gels was investigated by keeping the gel sample over the parallel plate at 23 °C. A 1 mm gap has been maintained between the two parallel plates and the excess gel squeezing out of the parallel plate was trimmed and the measurements were done.

6. FT-IR studies

Infrared (IR) spectra were recorded from 400 – 4000 cm⁻¹ using KBr in Perkin Elmer spectrum 100 spectrophotometer. Agilent Q-TOF 6230 instrument were used.

7. Formation of deep eutectic gel

Deep eutectic solvent is prepared my mixing a suitable quantity of choline chloride ChCl (HBA) and Ethylene glycol/ Glycerol/ Urea (HBD) in a round bottom flask and stirred at 70 °C till a homogeneous solution is formed. To the formed DES, HMBDS gelator is added and heated till the gelator dissolves and left to stand for the gel formation.

2 Synthesis

(a) Procedure for the synthesis of 4-hydroxymethyl benzaldehyde (2)

To a stirred solution of terephthalaldehyde **1** (1eq) in dry THF at 0 °C, NaBH₄ (0.3 eq) is added in portions. The mixture was stirred for 6h. After completion of the reaction as identified by TLC, 1N HCl is added to the mixture and further stirred for 20 min. Solvent was removed by under reduced pressure and the residue is extracted with ethyl acetate. Column chromatography was performed to obtain the pure product.⁷

(b) Procedure for the synthesis of 4-(hydroxymethyl)-1,3:2,4-dibenzylidene-D-sorbitol (4)

To a stirred solution of D-Sorbitol **3** (1 mmol) in acetonitrile (10 mL) at rt under N₂ atmosphere, 4-hydroxymethyl benzaldehyde **2** (2.1 mmol) and Dowex 50WX8 were added. The reaction mixture was stirred for about 24 h at 70 °C. After completion of the reaction as identified by TLC, the reaction mixture was allowed to cool down to room temperature. The formed precipitate is dissolved in hot cyrene and separate the catalyst by filtration. The cyrene solution is refrigerated for crystallization. The product is

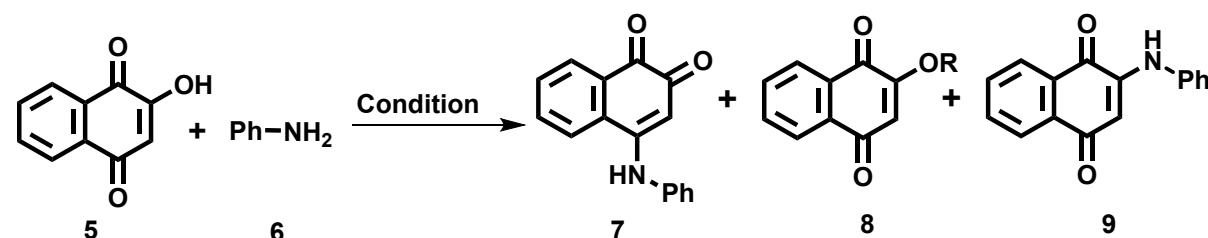
filtered, washed with water and dried under vacuum. The product was obtained as a white solid.

(c) General Procedure for the synthesis of 4-arylamino-1,2-naphthoquinones (7)

Lawsone (1 mmol) and amine (1.2 mmol) was incorporated in the HMDBS eutectogel by ether co-gelation method or cast on gel surface method. In cast on gel surface method, a complete permeation of reactant was achieved after 12h at room temperature. After incorporation of reactants, the gel vial is irradiated under blue LED for about 24 h. In order follow the reaction progress, a small piece of gel is taken, dispersed in DCM and analyzed with TLC. After completion of the reaction, the gel is placed in chloroform or hexane-chloroform or chloroform-methanol mixture to extract the product, this process is repeated 3 times. The extracted mixture is evaporated under vacuum. Brown product thus obtained is crystallized in a mixture of solvents (Hexane-Ethylacetate; Chloroform-Methanol, or a suitable solvents) and directly subjected to NMR analysis. Column chromatography is not performed..

3 Optimization studies

Table S1: Optimization studies for 4-arylamino-1,2-naphthoquinones using conventional method



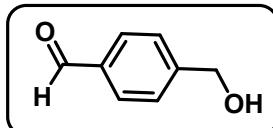
S.No	Catalyst	Condition	Solvent	Time (h)	7 (%)	8 (%)	9 (%)	observation
1.	pTSA	Reflux	MeOH	24	-	12	45	Isolated as a mixture
2.	TFA	Reflux	MeOH	12	traces	21	42	Isolated as a mixture
3.	CuSO ₄	Reflux	MeOH/EtOH/BuOH, iPrOH	24	traces	57	traces	Isolated as a mixture*

4.	ZnSO ₄	Reflux	MeOH	48	traces	traces	traces	-\$
5.	MgSO ₄	Reflux	MeOH	48	-	-	-	-\$
6.	No catalyst	Reflux	MeOH	24	13	52	32	Isolated yield
9.	CuSO ₄	Reflux	BuOH	48				98

* among all protic solvents MeOH rendered maximum yield, and hence yield mentioned are for MeOH in entry 3. \$ multiple spots were observed in TLC and starting materials are not consumed completely even after prolonged reaction. 5 and 6a (1 mmol) were used as model substrate for the conventional reaction.

4 Characterization studies

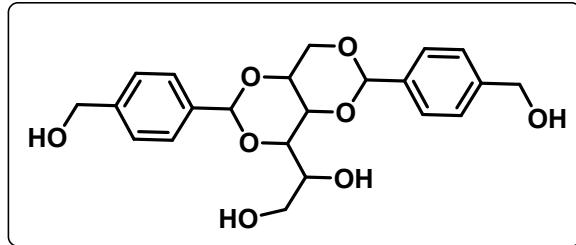
Compound 1: 4-hydroxymethyl benzaldehyde



The product was obtained as a colour less liquid with 70% yield

¹H NMR (400 MHz, CDCl₃) δ: 9.86 (d, *J* = 1.6 Hz, 1H, OH), 7.74 (d, *J* = 7.9 Hz, 2H, Ar-H), 7.41 (d, *J* = 7.9 Hz, 2H, Ar-H), 4.68 (s, 2H, Ar-CH₂).

Compound 2: 4-(hydroxymethyl)-1,3:2,4-dibenzylidene-D-sorbitol



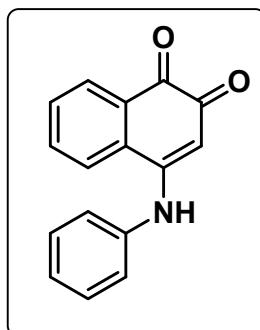
The product was obtained as a white solid with 86% yield

¹H NMR (400 MHz, DMSO-*d*₆) δ: 7.41 (t, *J* = 8.1 Hz, 4H, Ar-H), 7.31 (dd, *J* = 8.1, 1.6 Hz, 4H, Ar-H), 5.64 (s, 2H, acetal-H), 5.21 (s, 2H, benzylic-OH), 4.83 (s, 1H, Sac-H), 4.50 (s, 4H, Ar-CH₂), 4.21 – 4.11 (m, 3H, Sac-H), 3.93 (d, *J* = 1.7 Hz, 1H, Sac-H), 3.83 (dd, *J* = 9.2, 1.7 Hz, 1H, Sac-H), 3.76 (d, *J* = 6.4 Hz, 1H, Sac-H), 3.59 (dd, *J* = 11.3, 2.3 Hz, 1H, Sac-H), 3.43 (dd, *J* = 11.2, 5.1 Hz, 2H, Sac-H).

¹³C NMR (100 MHz, DMSO-*d*₆) δ: 143.39, 143.29, 137.64, 137.38, 126.40, 126.38, 126.36, 126.34, 99.84, 99.78, 78.20, 78.12, 70.55, 69.78, 68.89, 68.20, 63.12.

HRMS (ESI, m/z): C₂₂H₂₆O₈, [M+H]⁺ Calculated: 419.1706; [M+H]⁺ found: 419.1704.

Compound 7a: 4-(phenylamino)-naphthalene-1,2-dione



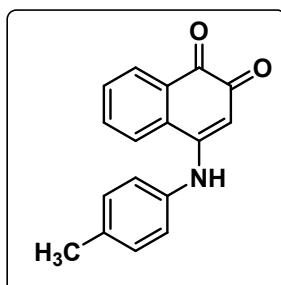
The product was obtained as a brown solid with a 92% yield.

¹H NMR (400 MHz, CDCl₃) δ: 8.05 (t, *J* = 6.3 Hz, 2H, Ar-H), 7.69 (t, *J* = 7.6 Hz, 1H, Ar-H), 7.63 – 7.57 (m, 1H, Ar-H), 7.50 (s, 1H, NH), 7.35 (t, *J* = 7.8 Hz, 2H, Ar-H), 7.21 (dd, *J* = 8.0 Hz, 2H, Ar-H), 7.15 (t, *J* = 7.4 Hz, 1H Ar-H), 6.35 (s, 1H, Olefin-H). **¹³C NMR (100 MHz, CDCl₃) δ:** 183.97, 182.09, 144.75, 137.45, 134.95, 133.24, 132.38, 130.38, 129.72, 126.55, 126.19, 125.65, 122.64, 103.41.

HRMS(ESI m/z):

C₁₆H₁₁NO₂, [M+H]⁺ Calculated: 250.0868, [M+H]⁺ Found: 250.0869.

Compound 7b: 4-(p-tolylamino)naphthalene-1,2-dione

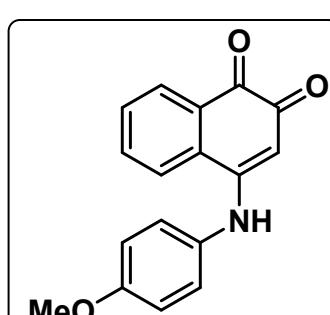


The product was obtained as a brown solid with a 88% yield.

¹H NMR (400 MHz, CDCl₃) δ: 8.10 (m, *J* = 7.6, 3.0, 1.3 Hz, 2H, Ar-H), 7.75 (td, *J* = 7.6, 1.3 Hz, 1H, Ar-H), 7.65 (td, *J* = 7.5, 1.4 Hz, 1H, Ar-H), 7.52 (s, 1H, NH), 7.22 (d, *J* = 8.2 Hz, 2H, Ar-H), 7.16 (d, *J* = 8.4 Hz, 2H, Ar-H), 6.35 (s, 1H, Olefin-H), 2.36 (s, 3H, Ar-CH₃).

¹³C NMR (100 MHz, CDCl₃) δ: 183.86, 182.16, 145.07, 135.66, 134.89, 134.77, 133.34, 132.26, 130.42, 130.25, 126.49, 126.16, 122.75, 103.04, 20.99.

Compound 7c: 4-((4-methoxyphenyl)amino)naphthalene-1,2-dione

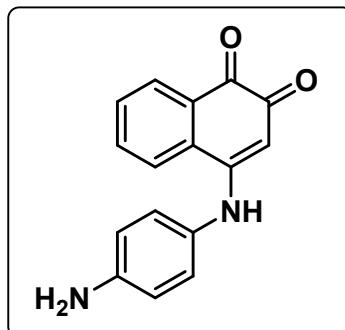


The product was obtained as a brown solid with a 86% yield.

¹H NMR (400 MHz, CDCl₃) δ: 8.11 (m, *J* = 7.6, 4.0, 1.3 Hz, 2H, Ar-H), 7.75 (td, *J* = 7.5, 1.4 Hz, 1H, Ar-H), 7.66 (td, *J* = 7.5, 1.4 Hz, 1H, Ar-H), 7.44 (s, 1H, NH), 7.20 (d, *J* = 8.9 Hz, 2H, Ar-H), 6.95 (d, *J* = 9.0 Hz, 2H, Ar-H), 6.22 (s, 1H, Olefin-H), 3.83 (s, 3H, O-CH₃).

¹³C NMR (100 MHz, CDCl₃) δ: 183.75, 182.19, 157.70, 145.70, 134.89, 133.42, 132.21, 130.44, 130.04, 126.45, 126.16, 124.87, 114.93, 102.55, 55.58.

Compound 7d: 4-((4-aminophenyl)amino)naphthalene-1,2-dione

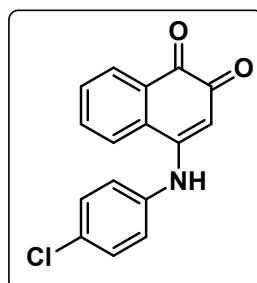


¹H NMR (400 MHz, DMSO-d₆) δ: 9.03 (s, 1H, NH), 8.09 (d, *J* = 7.5 Hz, 1H, Ar-H), 7.99 (d, *J* = 7.5 Hz, 1H, Ar-H), 7.89 (t, *J* = 7.6 Hz, 1H, Ar-H), 7.81 (t, *J* = 7.5 Hz, 1H, Ar-H), 7.07 (d, *J* = 8.7 Hz, 2H, Ar-H), 6.68 (d, *J* = 8.7 Hz, 2H, Ar-H), 5.91 (s, 1H, Olefin-H), 5.28 (s, 2H, NH₂).

¹³C NMR (100 MHz, DMSO-d₆) δ: 182.36, 147.64, 135.20, 133.66, 132.65, 131.05, 130.44, 126.41, 125.70, 122.53, 119.77, 114.92, 101.27.

HRMS(ESI m/z): C₁₆H₁₂N₂O₂, [M+H]⁺ Calculated: 265.0977, [M+H]⁺ Found: 265.0979.

Compound 7e: 4-((4-chlorophenyl)amino)naphthalene-1,2-dione



The product was obtained as a brown solid with a 91% yield.

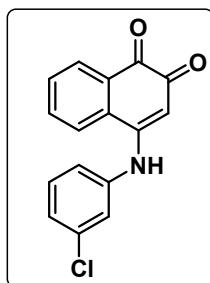
¹H NMR (400 MHz, CDCl₃) δ: 8.13 (d, *J* = 8.2 Hz, 2H, Ar-H), 7.78 (s, 1H, Ar-H), 7.69 (s, 1H, Ar-H), 7.52 (s, 1H, NH), 7.40 (d, *J* = 8.1 Hz, 2H, Ar-H), 7.27 – 7.17 (m, 2H, Ar-H), 6.37 (s, 1H, Olefin-H).

¹³C NMR (100 MHz, CDCl₃+DMSO-d₆) δ: 183.26, 181.85, 146.09, 137.37, 135.01, 133.03, 132.76, 130.77, 129.73, 129.49, 126.47, 125.73, 125.28, 103.03.

HRMS(ESI m/z): C₁₆H₁₀ClNO₂, [M+H]⁺ Calculated: 284.0478, [M+H]⁺ Found: 284.0477.

Compound 7f: 4-((3-dione

chlorophenyl)amino)naphthalene-1,2-



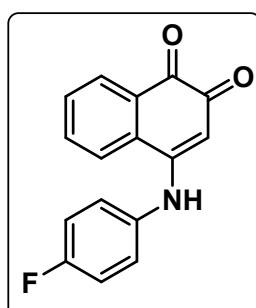
The product was obtained as a brown solid with a 93% yield.

¹H NMR (400 MHz, CDCl₃) δ: 8.05 (m, *J* = 7.5, 4.9, 1.3 Hz, 2H, Ar-H), 7.71 (td, *J* = 7.6, 1.4 Hz, 1H, Ar-H), 7.62 (td, *J* = 7.6, 1.4 Hz, 1H, Ar-H), 7.47 (s, 1H, NH), 7.28 (t, *J* = 8.0 Hz, 1H, Ar-H), 7.22 (t, *J* = 2.1 Hz, 1H, Ar-H), 7.14 – 7.09 (m, 2H, Ar-H), 6.36 (s, 1H, Olefin-H).

¹³C NMR (100 MHz, CDCl₃+DMSO-d₆) δ: 181.72, 134.81, 132.96, 132.55, 130.59, 126.48, 125.79, 125.17, 124.06, 123.21, 122.27, 103.55.

HRMS(ESI m/z): C₁₆H₁₀ClNO₂, [M+H]⁺ Calculated: 284.0475, [M+H]⁺ Found: 284.0477.

Compound 7g: 4-((4-fluorophenyl)amino)naphthalene-1,2-dione

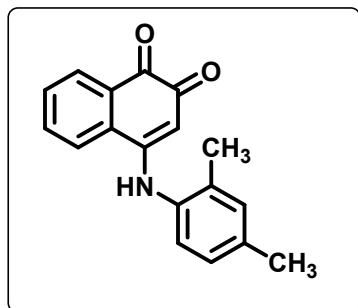


The product was obtained as a brown solid with a 84% yield.

¹H NMR (400 MHz, CDCl₃) δ: 8.11 (t, *J* = 7.0 Hz, 2H, Ar-H), 7.72 (dt, *J* = 36.7, 7.5 Hz, 2H, Ar-H), 7.46 (s, 1H, NH), 7.29 – 7.22 (m, 2H, Ar-H), 7.13 (t, *J* = 8.4 Hz, 2H, Ar-H), 6.25 (s, 1H, Olefin-H).

¹³C NMR (100 MHz, CDCl₃) δ: 183.86, 181.97, 145.32, 135.00, 133.22, 132.43, 130.36, 126.55, 126.24, 125.07, 124.99, 116.77, 116.55, 103.14.

Compound 7h: 4-((2,4-dimethylphenyl)amino)naphthalene-1,2-dione

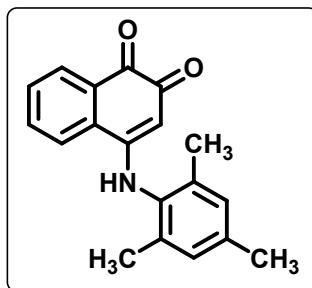


The product was obtained as a brown solid with a 90% yield.

¹H NMR (400 MHz, CDCl₃) δ: 8.11 (m, *J* = 9.4, 7.7, 1.0 Hz, 2H, Ar-H), 7.75 (td, *J* = 7.5, 1.3 Hz, 1H, Ar-H), 7.66 (td, *J* = 7.5, 1.3 Hz, 1H, Ar-H), 7.27 (s, 1H, NH), 7.17 – 7.04 (m, 3H, Ar-H), 5.90 (s, 1H, Olefin-H), 2.34 (s, 3H, Ar-CH₃), 2.24 (s, 3H, Ar-CH₃).

¹³C NMR (100 MHz, CDCl₃) δ: 183.71, 182.24, 146.16, 136.90, 134.87, 133.49, 133.13, 132.70, 132.20, 132.05, 130.53, 127.74, 126.39, 126.18, 124.93, 102.90, 20.99, 17.67.

Compound 7i: 4-((2,4,6-trimethylphenyl)amino)naphthalene-1,2-dione

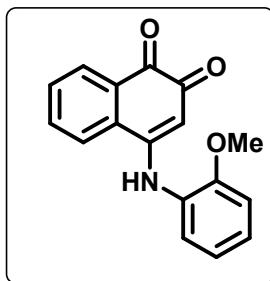


The product was obtained as a brown solid with a 92% yield.

¹H NMR (400 MHz, CDCl₃) δ: 8.15 – 8.07 (m, 2H, Ar-H), 7.74 (d, *J* = 1.3 Hz, 1H, Ar-H), 7.66 (td, *J* = 7.5, 1.2 Hz, 1H, Ar-H), 7.04 (s, 1H, NH), 6.95 (s, 2H, Ar-H), 5.38 (s, 1H, Olefin-H), 2.31 (s, 3H, Ar-CH₃), 2.16 (s, 6H, Ar-CH₃).

¹³C NMR (100 MHz, CDCl₃) δ: 183.51, 182.10, 146.92, 137.93, 135.43, 134.82, 133.65, 132.13, 130.92, 130.69, 129.48, 126.30, 126.21, 102.66, 20.96, 17.92.

Compound 7j: 4-((2-methoxyphenyl)amino)naphthalene-1,2-dione

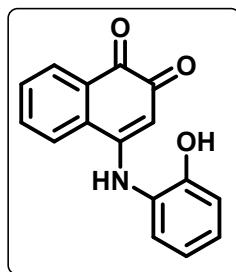


The product was obtained as a brown solid with a 86% yield.

¹H NMR (400 MHz, CDCl₃) δ: 8.13 (s, 2H, Ar-H), 7.72 (dt, *J* = 36.5, 7.0 Hz, 2H, Ar-H), 7.49 – 7.39 (m, 1H, Ar-H), 7.22 – 7.11 (m, 1H, Ar-H), 7.01 (dt, *J* = 13.2, 5.7 Hz, 2H, Ar-H), 6.48 (s, 1H, Olefin-H), 3.91 (s, 3H, Ar-OCH₃).

¹³C NMR (100 MHz, CDCl₃) δ: 184.00, 182.14, 151.18, 143.99, 134.78, 133.33, 132.30, 126.98, 126.54, 126.13, 125.48, 121.08, 120.92, 111.17, 103.63, 55.78.

Compound 7k: 4-((2-hydroxyphenyl)amino)naphthalene-1,2-dione

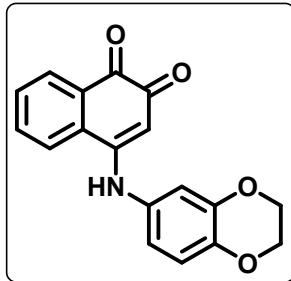


The product was obtained as a brown solid with a 62% yield.

¹H NMR (400 MHz, CDCl₃+DMSO) δ: 9.65 (s, 1H, NH), 8.15 – 8.03 (m, 3H, Ar-H), 7.81 – 7.73 (m, 1H, NH), 7.71 – 7.65 (m, 1H, Ar-H), 7.33 (d, *J* = 7.1 Hz, 1H, Ar-H), 7.07 – 6.97 (m, 2H, Ar-H), 6.93 – 6.85 (m, 1H Ar-H), 6.29 (s, 1H, Olefin-H).

¹³C NMR (100 MHz, CDCl₃+DMSO) δ: 183.52, 181.97, 149.92, 144.70, 134.72, 133.29, 132.27, 130.55, 126.40, 126.04, 125.83, 125.51, 122.17, 119.71, 116.32, 102.93.

Compound 7l: 4-((3,4-ethylenedioxyphenyl)amino)naphthalene-1,2-dione

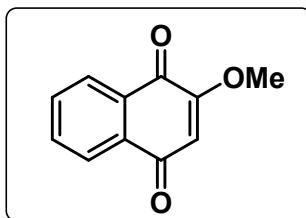


The product was obtained as a brown solid with a 75% yield.

¹H NMR (400 MHz, CDCl₃) δ: 8.10 (dt, *J* = 7.5, 1.7 Hz, 2H, Ar-H), 7.75 (td, *J* = 7.6, 1.3 Hz, 1H, Ar-H), 7.65 (td, *J* = 7.6, 1.3 Hz, 1H, Ar-H), 7.40 (s, 1H, NH), 6.89 (d, *J* = 8.6 Hz, 1H, Ar-H), 6.81 (d, *J* = 2.5 Hz, 1H, Ar-H), 6.74 (dd, *J* = 8.6, 2.5 Hz, 1H, Ar-H), 6.29 (s, 1H, Olefin-H), 4.28 (s, 4H, Ar-O-CH₂).

¹³C NMR (100 MHz, CDCl₃) δ: 183.78, 182.11, 145.38, 144.07, 141.75, 134.86, 133.35, 132.22, 130.72, 130.42, 126.45, 126.13, 118.02, 116.58, 112.47, 102.89, 64.41, 64.33.

Compound 8a: 2-methoxynaphthalene-1,4-dione



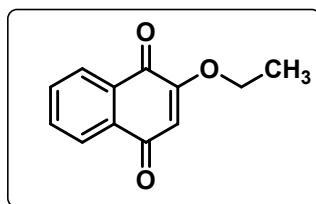
The product was obtained as a brown solid with a 92% yield.

¹H NMR (400 MHz, CDCl₃+DMSO-d₆) δ: 8.00 (dd, *J* = 17.0, 7.1 Hz, 2H, Ar-H), 7.70 (m, *J* = 7.5 Hz, 2H, Ar-H), 6.16 (s, 1H, Olefin-H), 3.85 (s, 3H, O-CH₃).

¹³C NMR (100 MHz, CDCl₃+DMSO-d₆) δ: 184.58, 179.95, 160.39, 134.39, 133.40, 131.95, 130.98, 126.49, 126.00, 109.95, 56.52.

HRMS(ESI m/z): C₁₁H₁₈O₃, [M+H]⁺ Calculated: 189.0551, [M+H]⁺ Found: 189.0559.

Compound 8b: 2-ethoxynaphthalene-1,4-dione



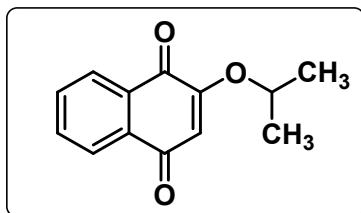
The product was obtained as a brown solid with a 80% yield.

¹H NMR (400 MHz, CDCl₃) δ: 8.08 – 7.99 (m, 2H, Ar-H), 7.70 – 7.61 (m, 2H, Ar-H), 6.08 (s, 1H, Olefin-H), 4.03 (q, *J* = 7.0 Hz, 2H, O-CH₂), 1.46 (t, *J* = 7.0 Hz, 3H, CH₃).

¹³C NMR (100 MHz, CDCl₃) δ: 185.08, 180.21, 159.72, 134.25, 133.28, 132.01, 131.15, 126.71, 126.12, 110.21, 65.35, 13.93.

HRMS(ESI m/z): C₁₂H₁₀O₃, [M+H]⁺ Calculated: 203.0708, [M+H]⁺ Found: 203.0708.

Compound 8c: 2-isopropoxynaphthalene-1,4-dione

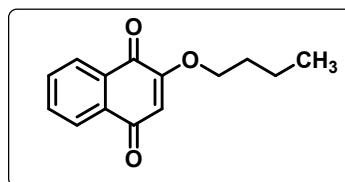


The product was obtained as a brown solid with a 78% yield.

¹H NMR (400 MHz, CDCl₃) δ: 8.03 (m, *J* = 16.8, 7.2, 1.1 Hz, 2H, Ar-H), 7.69 – 7.61 (m, 2H, Ar-H), 6.07 (s, 1H, Olefin-H), 4.50 (m, *J* = 6.1 Hz, 1H, O-CH), 1.38 (d, *J* = 6.1 Hz, 6H, CH₃).

¹³C NMR (100 MHz, CDCl₃) δ: 185.25, 180.53, 158.70, 134.18, 133.21, 131.96, 131.29, 126.70, 126.04, 110.50, 72.46, 21.18.

Compound 8d: 2-butoxynaphthalene-1,4-dione

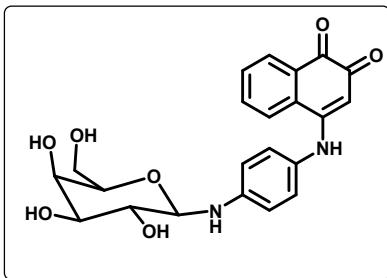


The product was obtained as a brown solid with a 98% yield.

¹H NMR (400 MHz, CDCl₃) δ: 8.02 (m, *J* = 17.0, 7.2, 1.3 Hz, 2H, Ar-H), 7.69 – 7.60 (m, 2H, Ar-H), 6.08 (s, 1H, Olefin-H), 3.94 (t, *J* = 6.6 Hz, 2H, O-CH₂), 1.81 (m, *J* = 9.1, 7.8, 6.5 Hz, 2H, alkyl(CH₂)), 1.50 – 1.38 (m, 2H, Alkyl(CH₂)), 0.92 (t, *J* = 7.4 Hz, 3H, Alkyl(CH₃)).

¹³C NMR (100 MHz, CDCl₃) δ: 185.09, 180.16, 159.92, 134.21, 133.25, 132.02, 131.18, 126.66, 126.10, 110.17, 69.38, 30.24, 19.12, 13.70.

Compound 10: 4-((4-Phenyl-β-D-galactose)amino)naphthalene-1,2-dione



The product was obtained as a brown solid with a 92% yield.

1H NMR (400 MHz, DMSO-d₆) δ: 9.10 (s, 1H, NH), 8.10 (d, J = 7.6 Hz, 1H, Ar-H), 7.99 (d, J = 7.6 Hz, 1H, Ar-H), 7.90 (t, J = 7.5 Hz, 1H, Ar-H), 7.82 (t, J = 7.5 Hz, 1H, Ar-H), 7.16 (d, J = 8.5 Hz, 2H, Ar-H), 6.83 (d, J = 8.4 Hz, 2H, Ar-H), 5.94 (s, 1H, Olefin-H), 5.37 (dd, J = 5.5 Hz, 1H, Sac-H), 5.15 – 4.95 (m, 1H, Sac-H), 4.81 (dd, J = 5.4 Hz, 1H, Sac-H), 4.62 (dq, J = 13.4, 5.9 Hz, 1H, Sac-H), 4.46 – 4.36 (m, 1H, Sac-H), 3.93 (m, J = 6.5, 6.0 Hz, 1H, Sac-H), 3.78 (d, J = 3.9 Hz, 1H, Sac-H), 3.53 (s, 2H, Sac-H). **13C NMR (100 MHz, DMSO-d₆) δ:** 182.29, 147.46, 146.00, 135.34, 133.42, 132.80, 130.92, 127.57, 126.48, 125.70, 125.62, 113.99, 100.95, 86.00, 76.08, 74.78, 70.60, 68.91, 61.05. **HRMS(ESI m/z):** C₂₂H₂₂N₂O₇, [M+H]⁺ Calculated : 427.1505, [M+H]⁺ Found: 427.1497 .

5 Gelation studies

Table S2: Gelation studies of HMDBS in various solvents

S.No	Solvent	Gelation (CGC, % (wt/v))
1	Water	I
2	Methanol	I
3	Ethanol	I
4	Glycerol	G(0.4)
5	Ethylene glycol	G(0.6)
6	Tetrahydrofuran	S
7	Dimethyl sulfoxide	S
8	Dimethyl sulfoxide + water	G(0.4)

9	Acetone	I
10	1,4-dioxane	PS
11	Dimethyl formamide	G(0.6)
12	Polyethylene glycol	G(0.5)
13	Acetic acid	I
14	N-methyl pyrrolidone	I
15	1,2-Dichlorobenzene	I
16	pyridine	I
17	n-Butanol	G(0.4)
18	Toulene	I
19	Benzyl alcohol	G(0.4)
20	Olive oil	I
21	Linseed oil	I
22	Dimethyl carbonate	I
24	Cyrene	I

I – Insoluble, G – Gelation, S – solution, PS – Partially soluble

Table S3: Gelation studies of HMDBS in Deep eutectic solvents

S.NO	Deep Eutectic solvents	Gelation(CGC wt/v)
1	Choline Chloride + Ethylene glycol (1:4)	G(0.3)
2	Choline Chloride + Ethylene glycol (1:3)	G(0.3)
3	Choline Chloride + Ethylene glycol (1.5:2)	G(1)
4	Choline Chloride + Glycerol (1:4)	G(0.4)
5	Choline Chloride + Glycerol (1:3)	G(0.5)
6	Choline Chloride + Glycerol (1.5:2)	G(1.3)
7	Choline Chloride + Urea (1:4)	G(0.6)
8	Choline Chloride + Urea (1:3)	G(0.6)

9	Choline Chloride + Urea (1.5:2)	G(1.5)
10	Choline Chloride + Polyethylene glycol (1:4)	Dense precipitation is observed
11	Choline Chloride + Polyethylene glycol (1:3)	Dense precipitation is observed
12	Choline Chloride + Polyethylene glycol (1.5:2)	Dense precipitation is observed

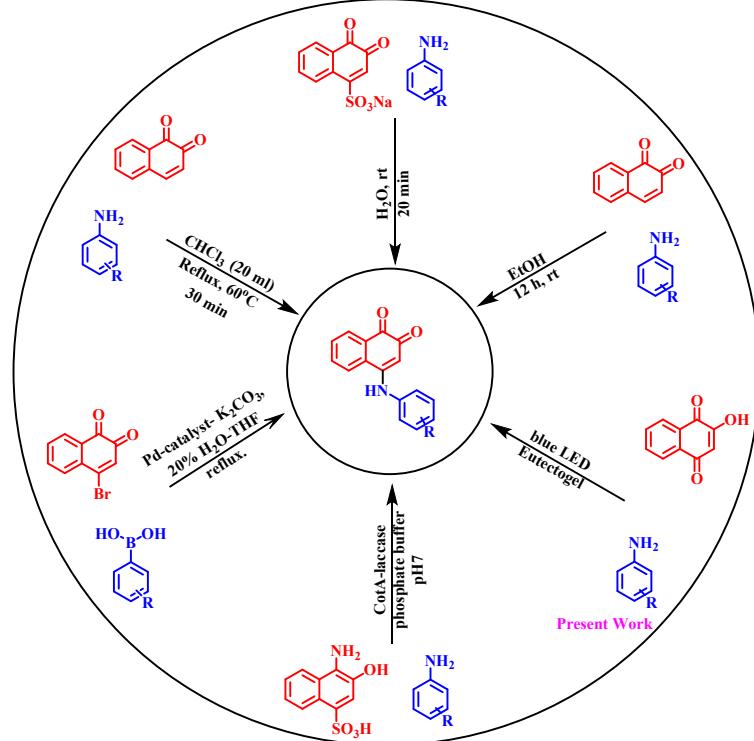


Figure S1: Various reaction conditions for the synthesis of 4-arylamino-1,2-naphthoquinones present in literature^{1,2,3,4,5,6}

6 NMR and HRMS spectra

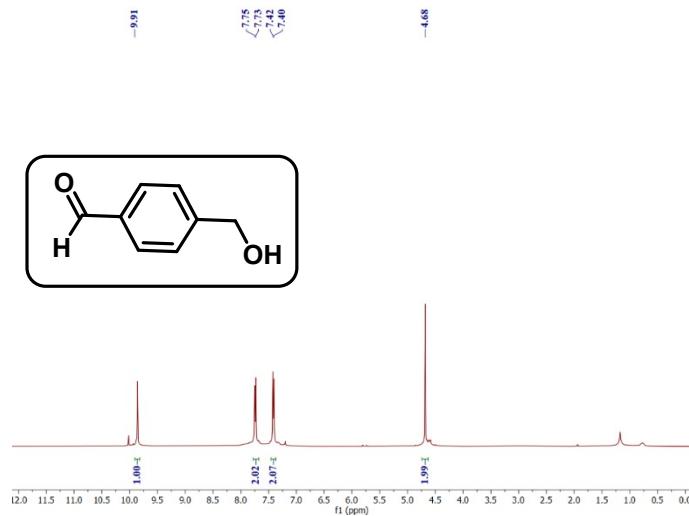


Figure S2: ¹H NMR (400MHz) Spectrum of HMBA **2** in CDCl₃ at 28 °C.

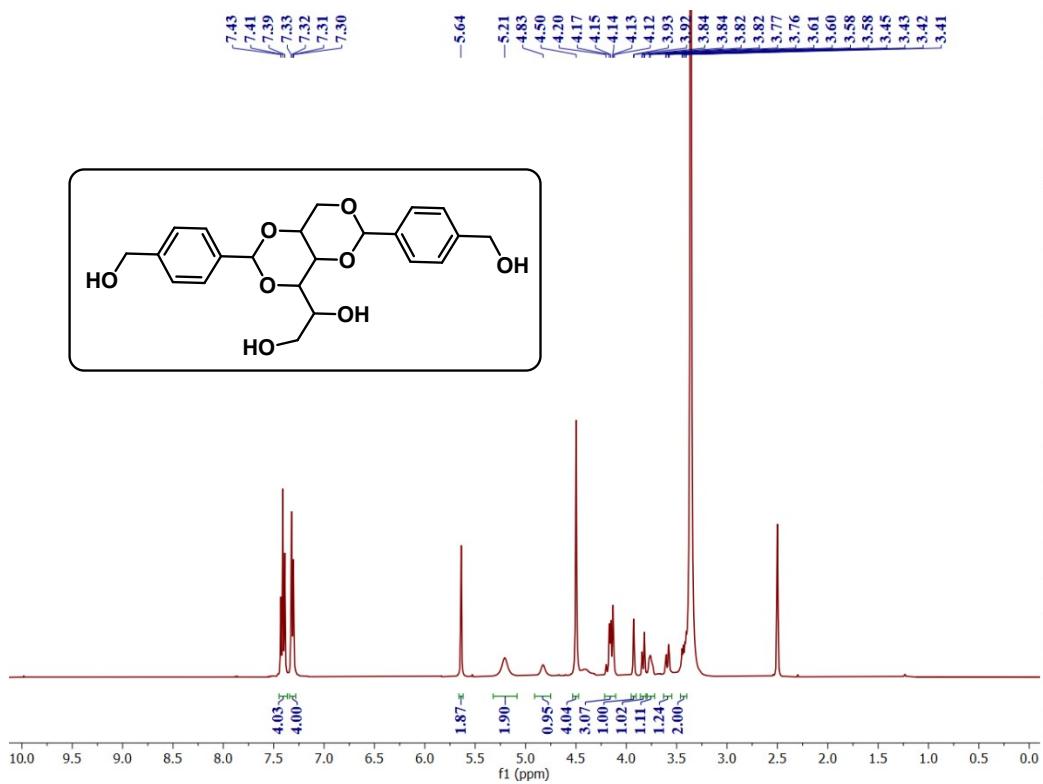


Figure S3: ¹H NMR (400MHz) Spectrum of HMDBS **4** in DMSO-d₆ at 28 °C.

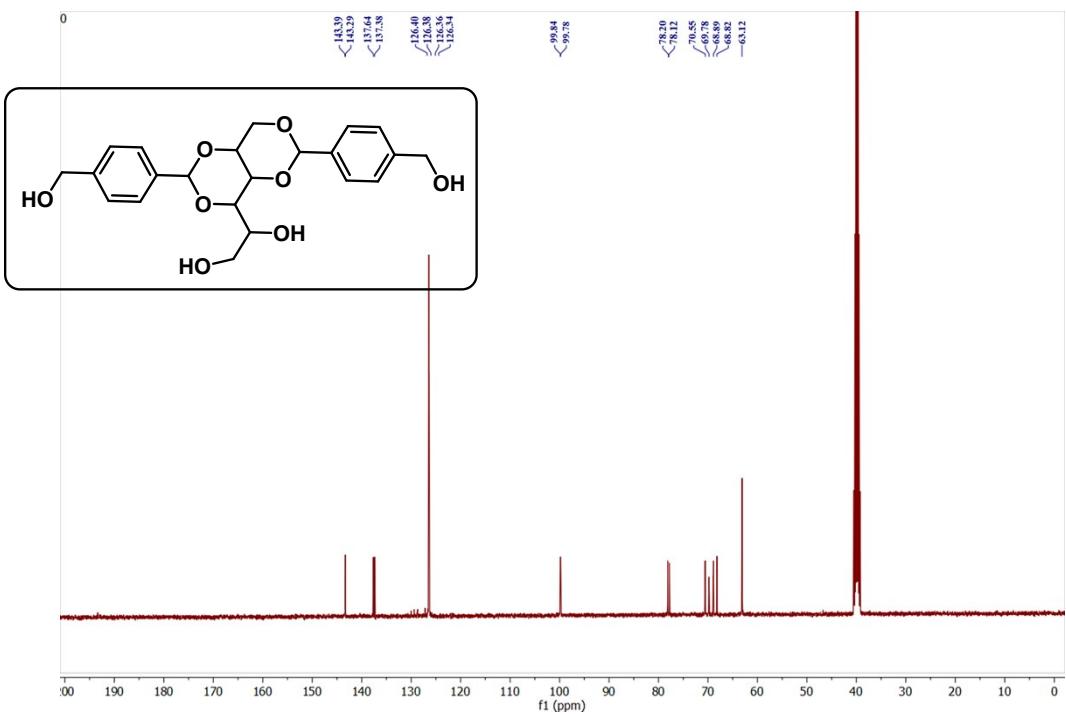


Figure S4: ^{13}C NMR (100 MHz) Spectrum HMDBS 4 in $\text{DMSO}-d_6$ 28 °C.

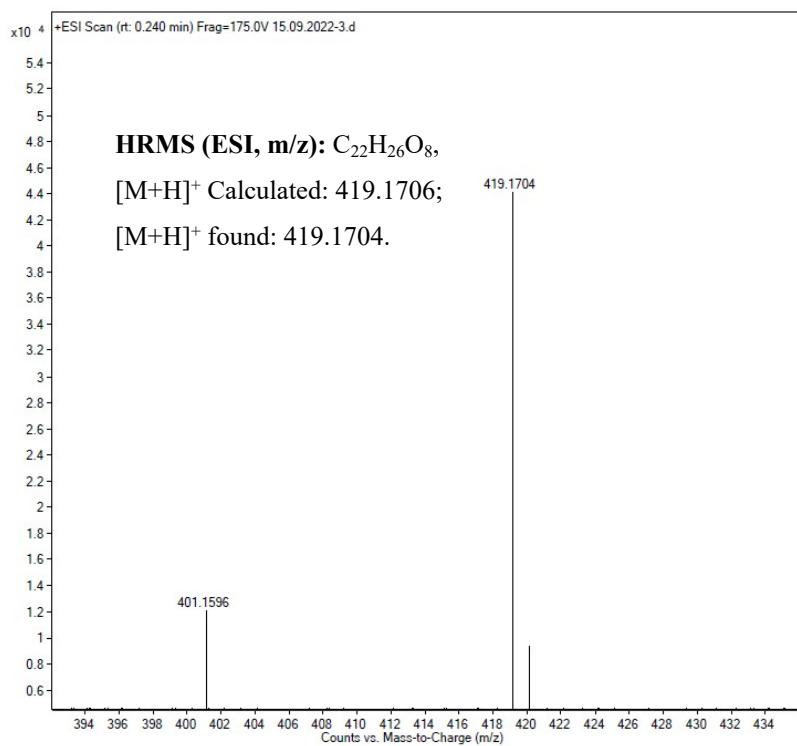


Figure S5: HRMS Spectrum of HMDBS 4.

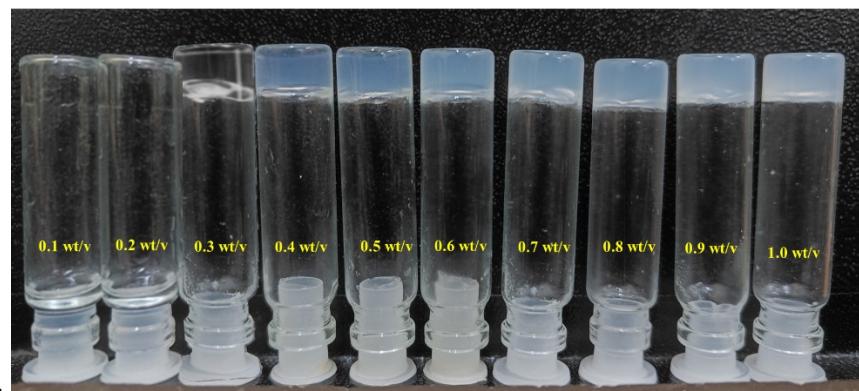


Figure S6: Images of gel formed by HMDBS in ChCl:EG (1:3) ratio with increasing gelator concentration in wt/v.

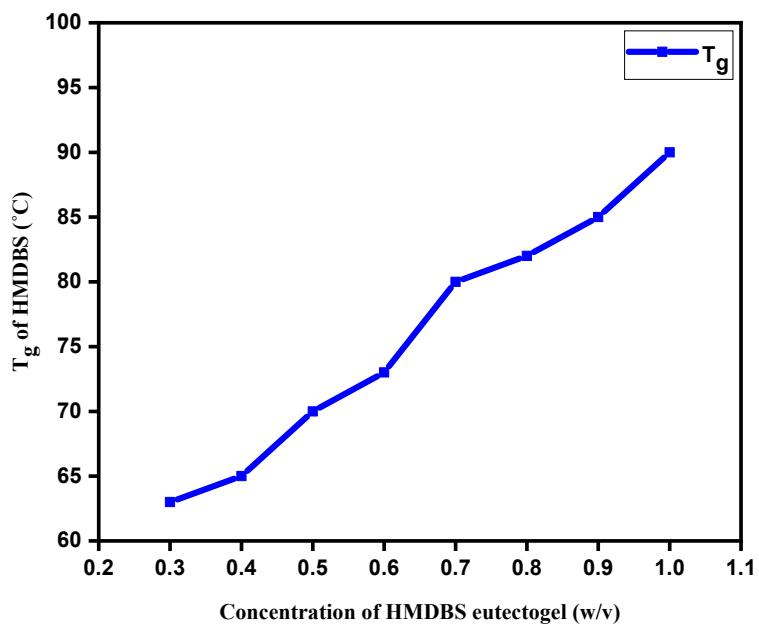


Figure S7: Transition temperature (T_g) values of HMDBS- ChCl: EG gels determined via reproducible tube inversion experiments.

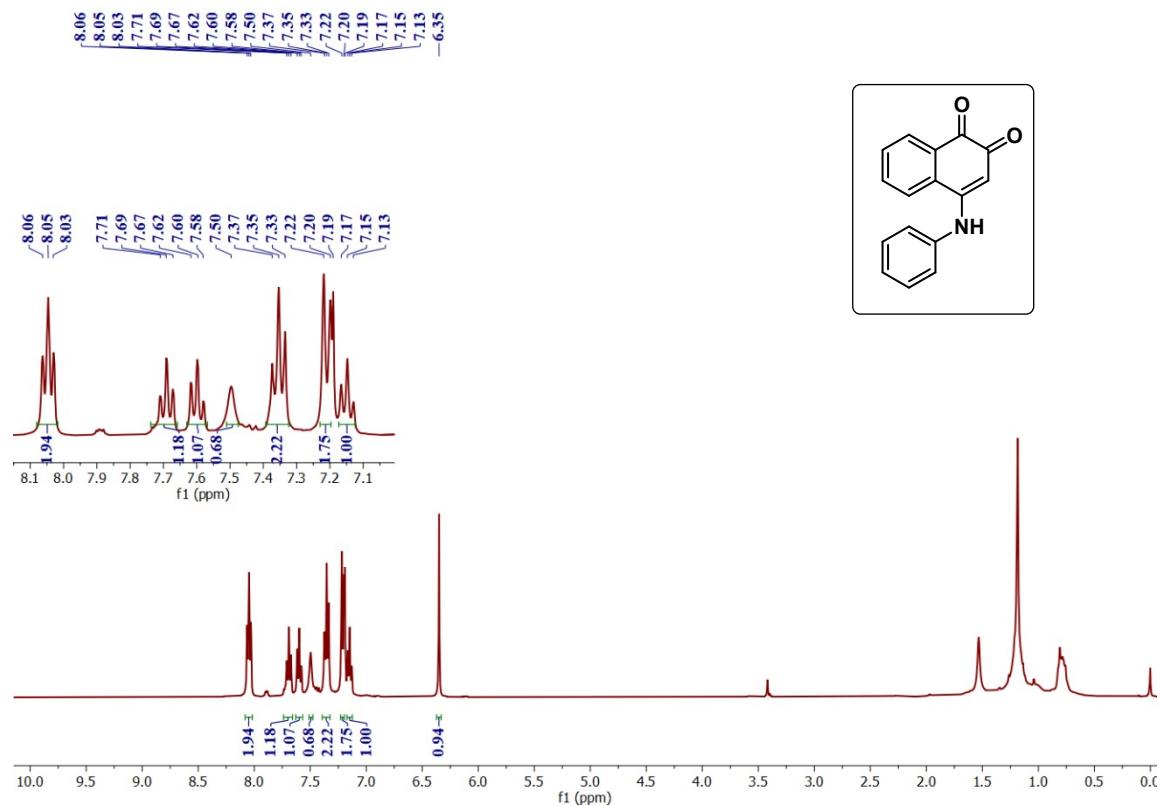


Figure S8: ^1H NMR (400MHz) Spectrum of **7a** in CDCl_3 at 28 °C.

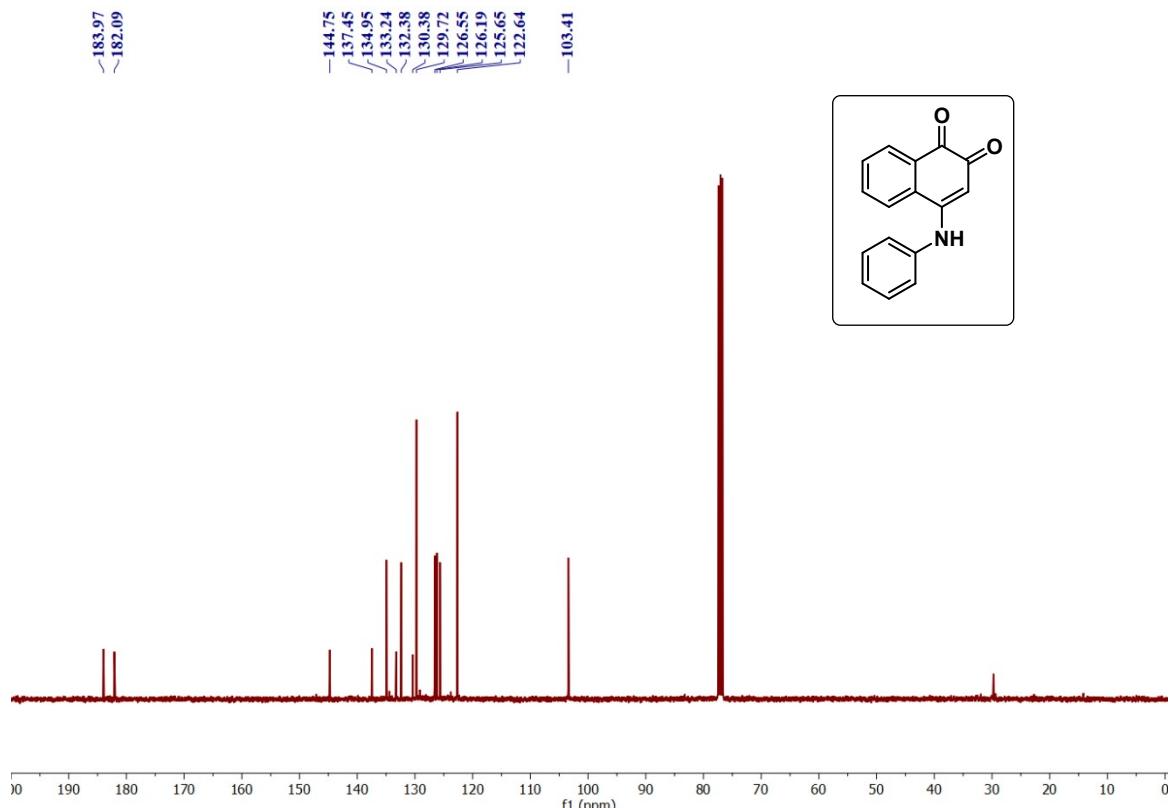


Figure S9: ^{13}C NMR (100 MHz) Spectrum of **7a** in CDCl_3 at 28 °C.

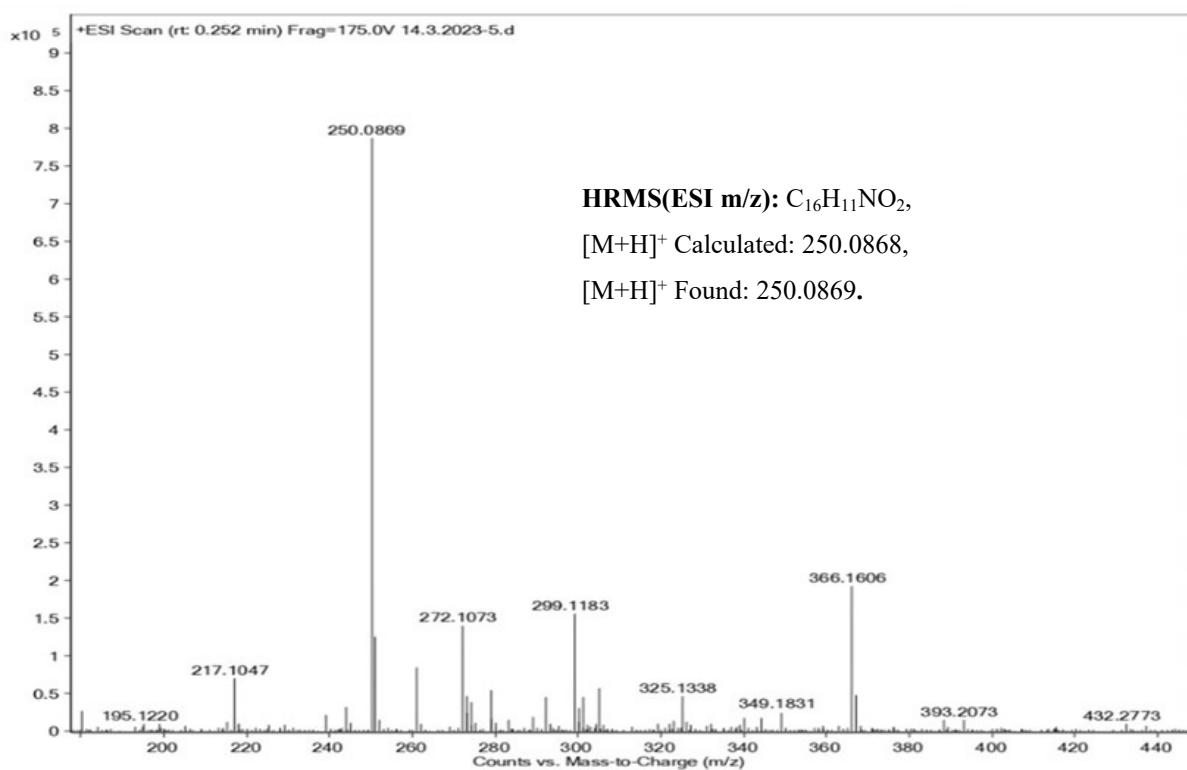


Figure S10: HRMS Spectrum of **7a**.

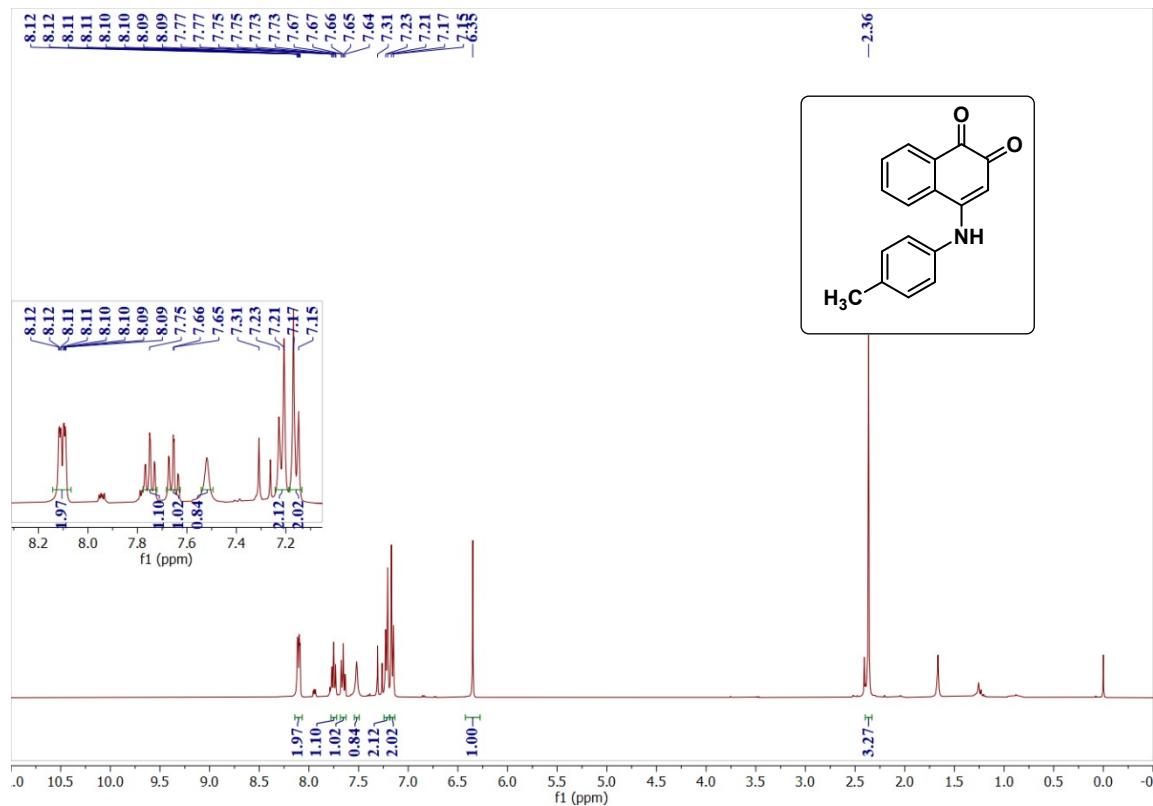


Figure S11: ^1H NMR (400MHz) Spectrum of **7b** in CDCl_3 at 28 °C.

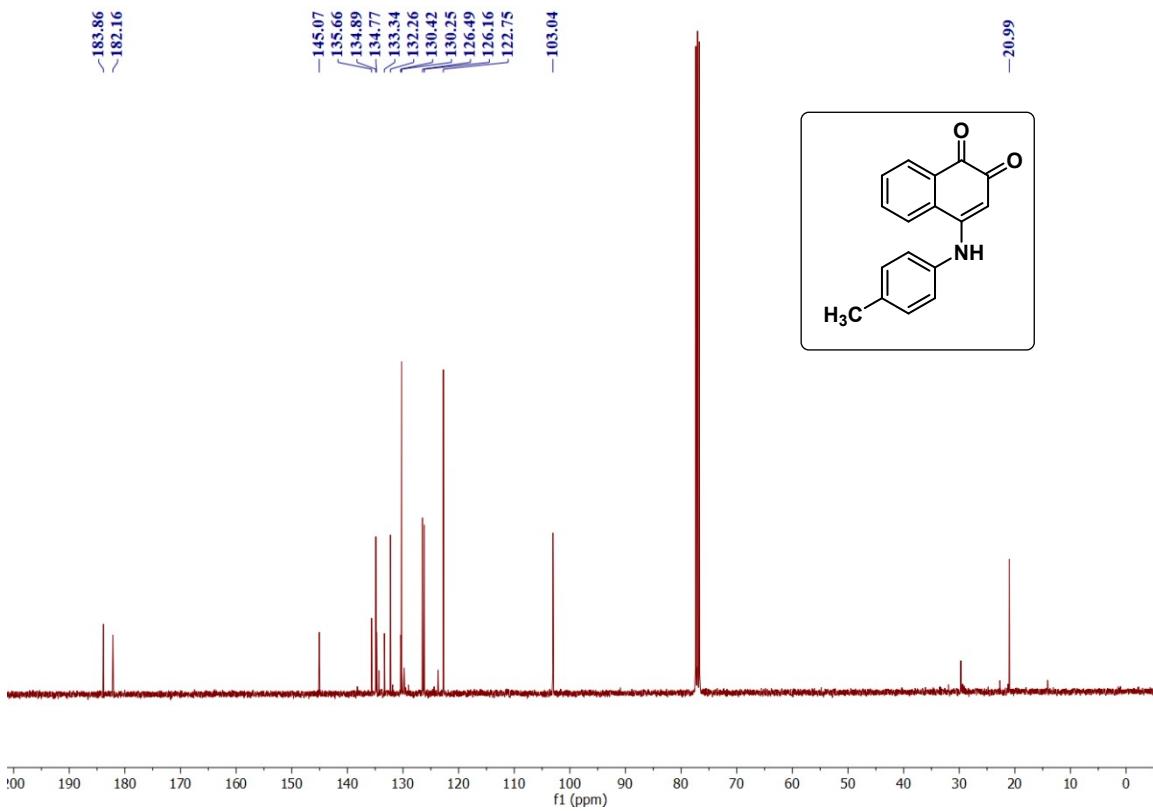


Figure S12: ^{13}C NMR (100 MHz) Spectrum of **7b** in CDCl_3 at 28 °C.

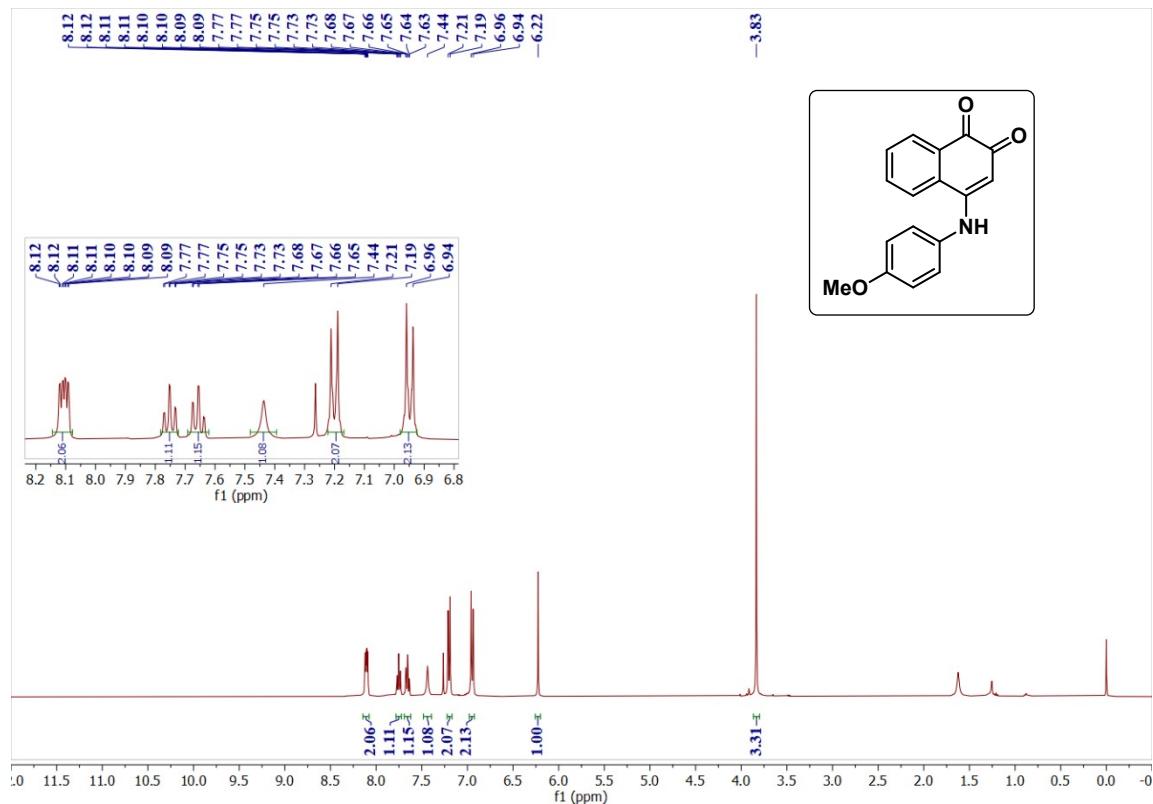


Figure S13: ^1H NMR (400MHz) Spectrum of **7c** in CDCl_3 at 28 °C.

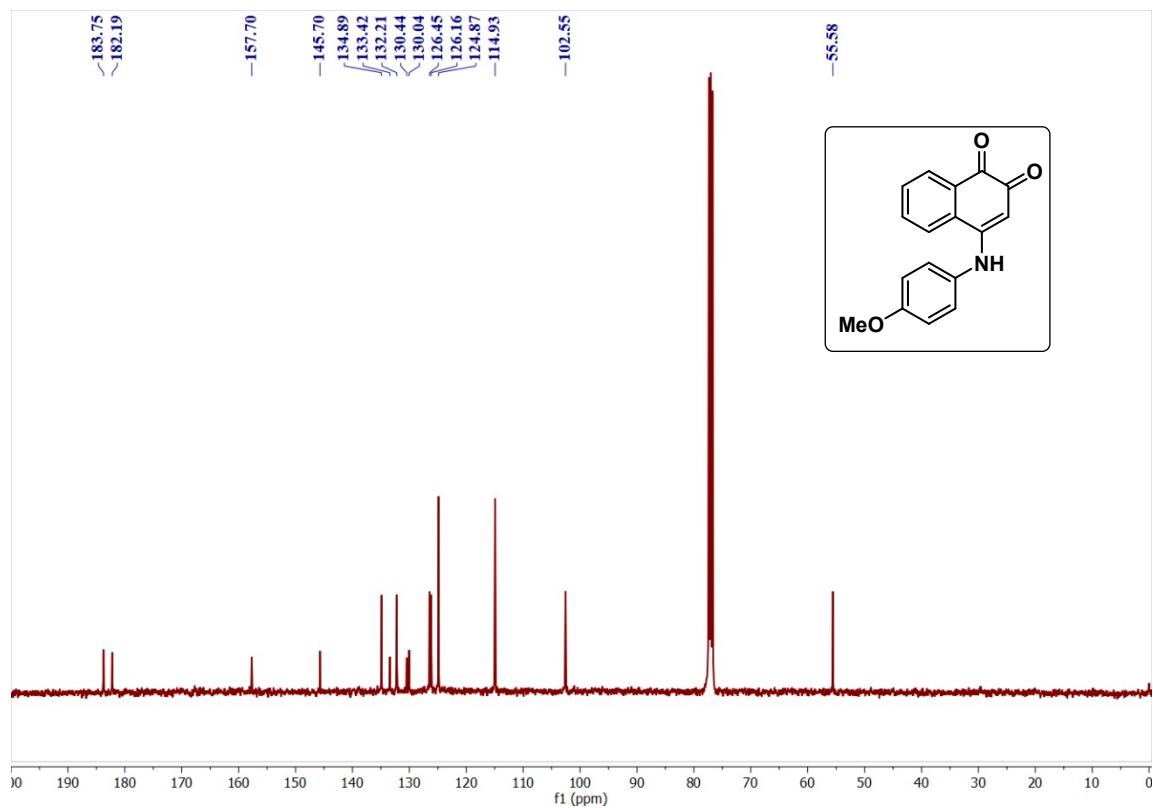


Figure S14: ^{13}C NMR (100 MHz) Spectrum of **7c** in CDCl_3 at 28 °C.

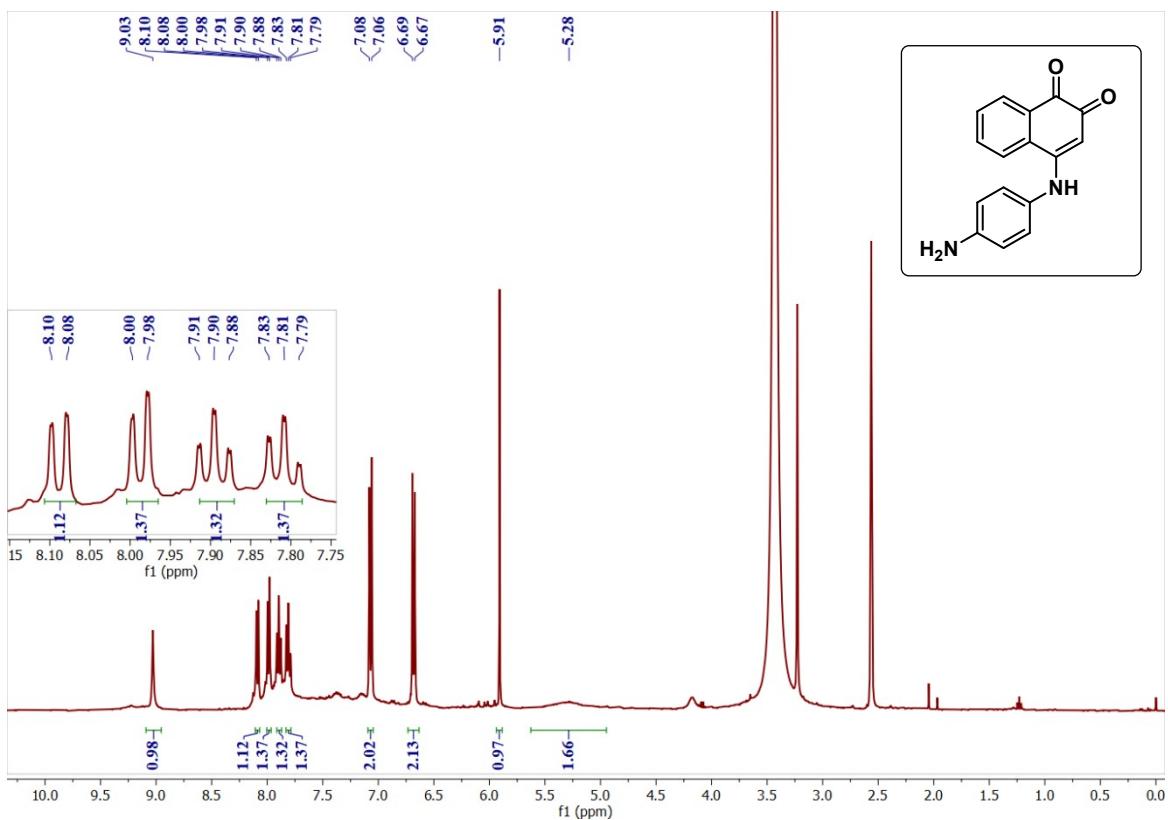


Figure S15: ^1H NMR (400MHz) Spectrum of **7d** in $\text{DMSO}-d_6$ at 80 °C.

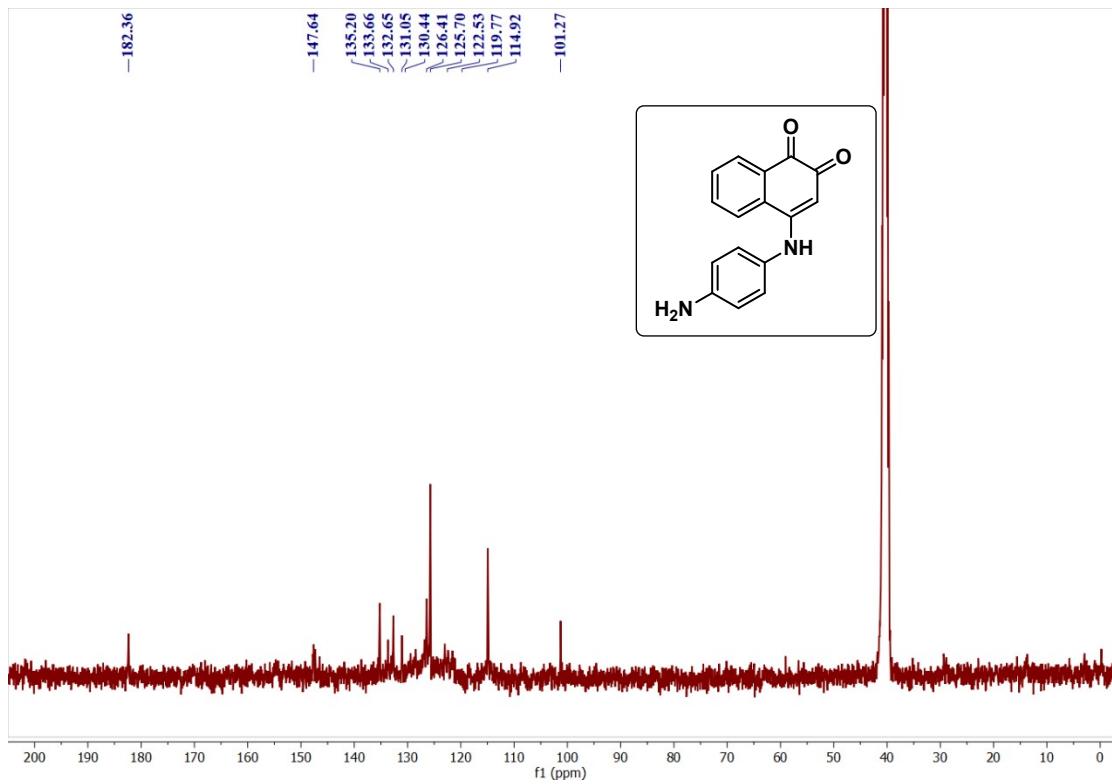


Figure S16: ^{13}C NMR (100 MHz) Spectrum of **7d** in $\text{DMSO}-d_6$ at 80 °C.

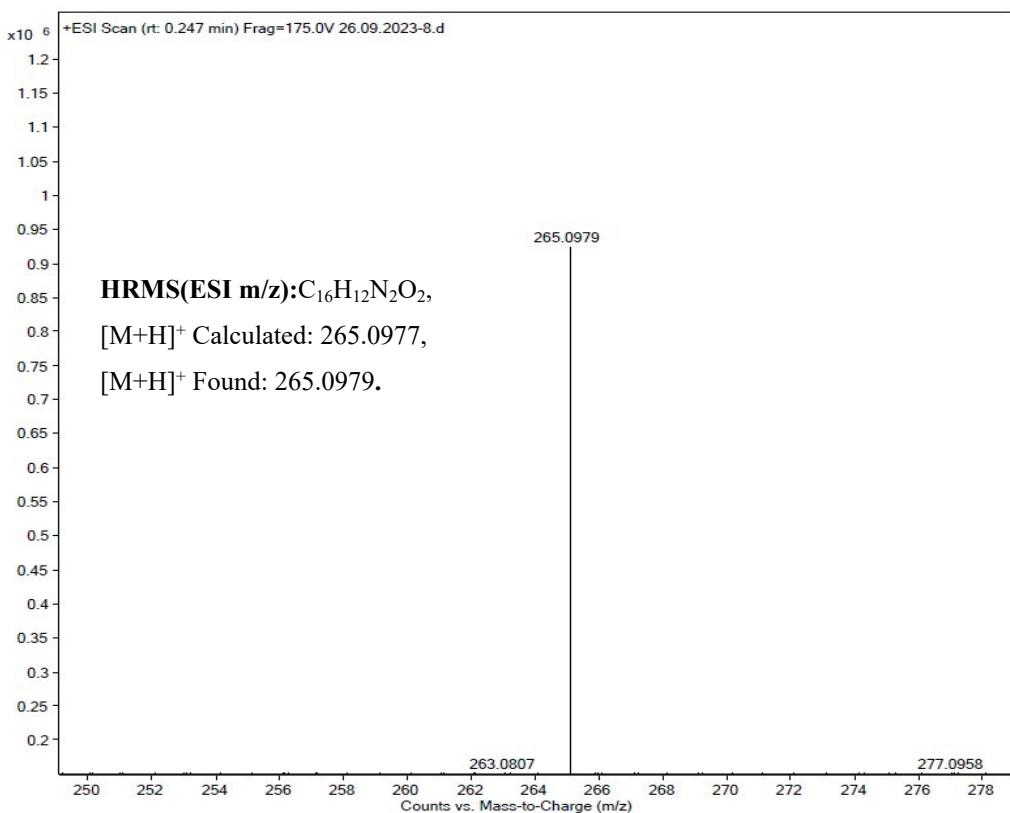


Figure S17: HRMS Spectrum of 7d.

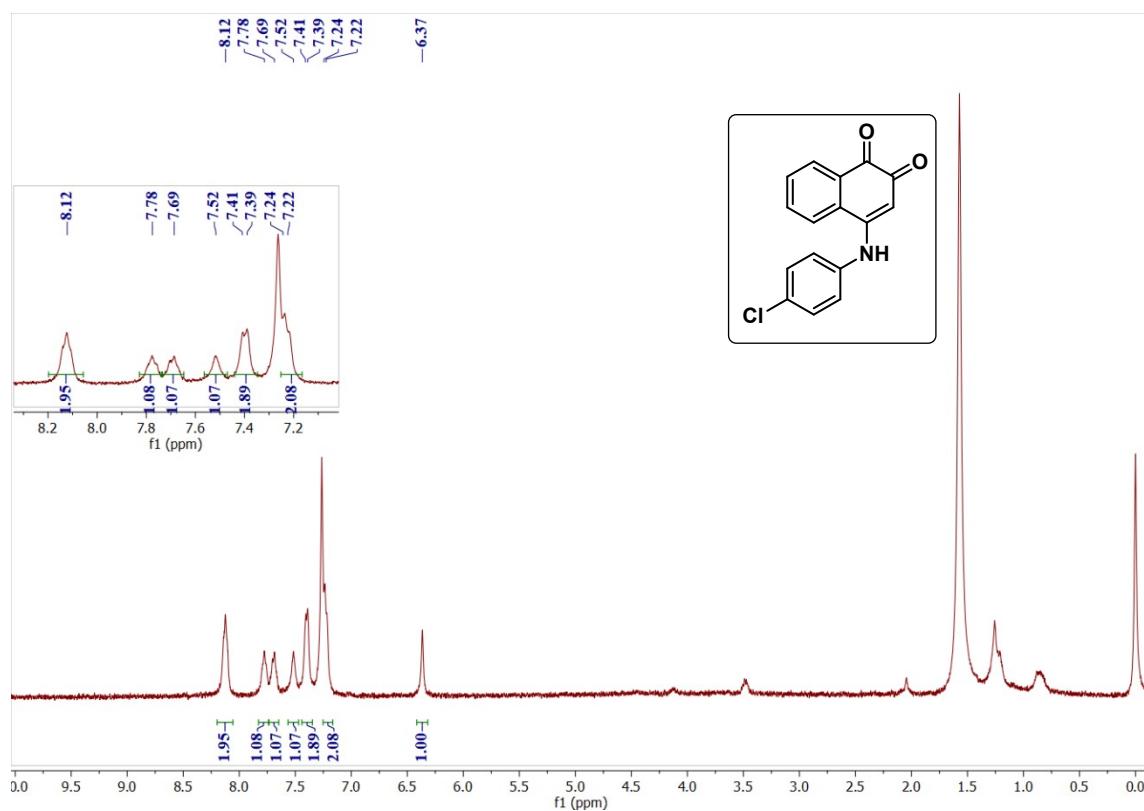


Figure S18: ¹H NMR (400MHz) Spectrum of 7e in CDCl₃ at 28 °C.

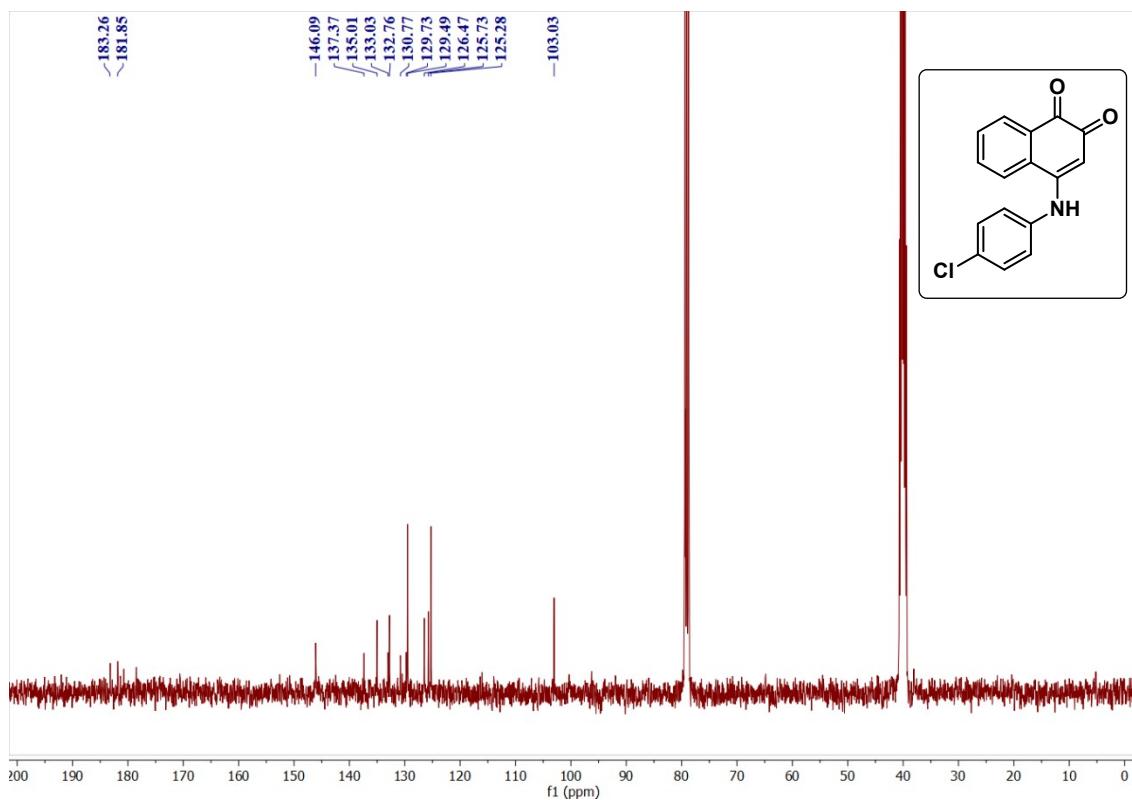


Figure S19: ^{13}C NMR (100 MHz) Spectrum of 7e in $\text{CDCl}_3+\text{DMSO}-d_6$ at 28 °C.

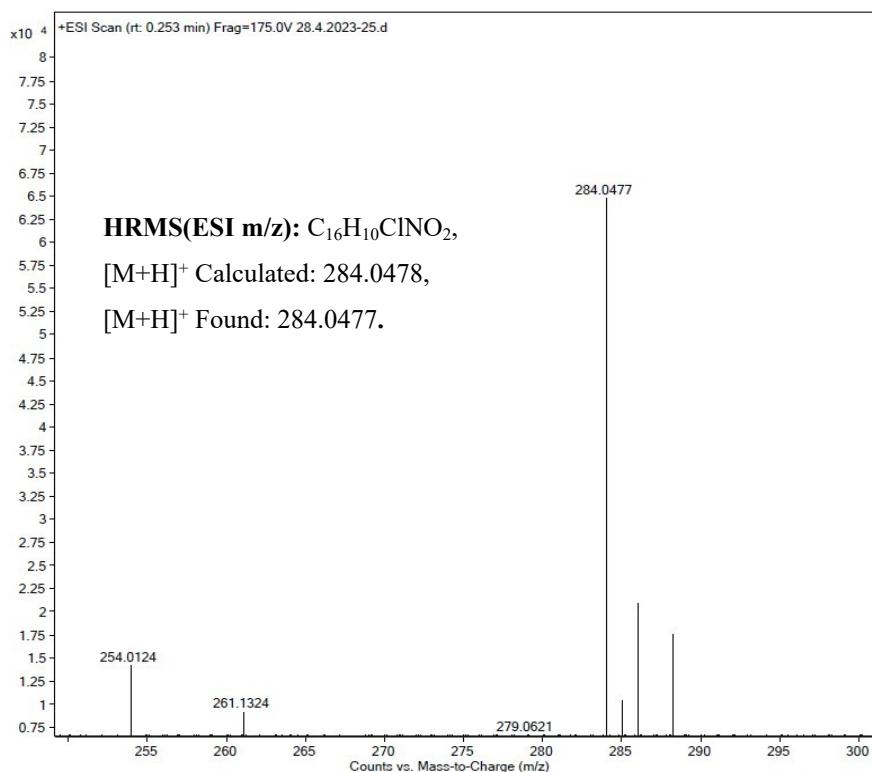


Figure S20: HRMS Spectrum of 7e.

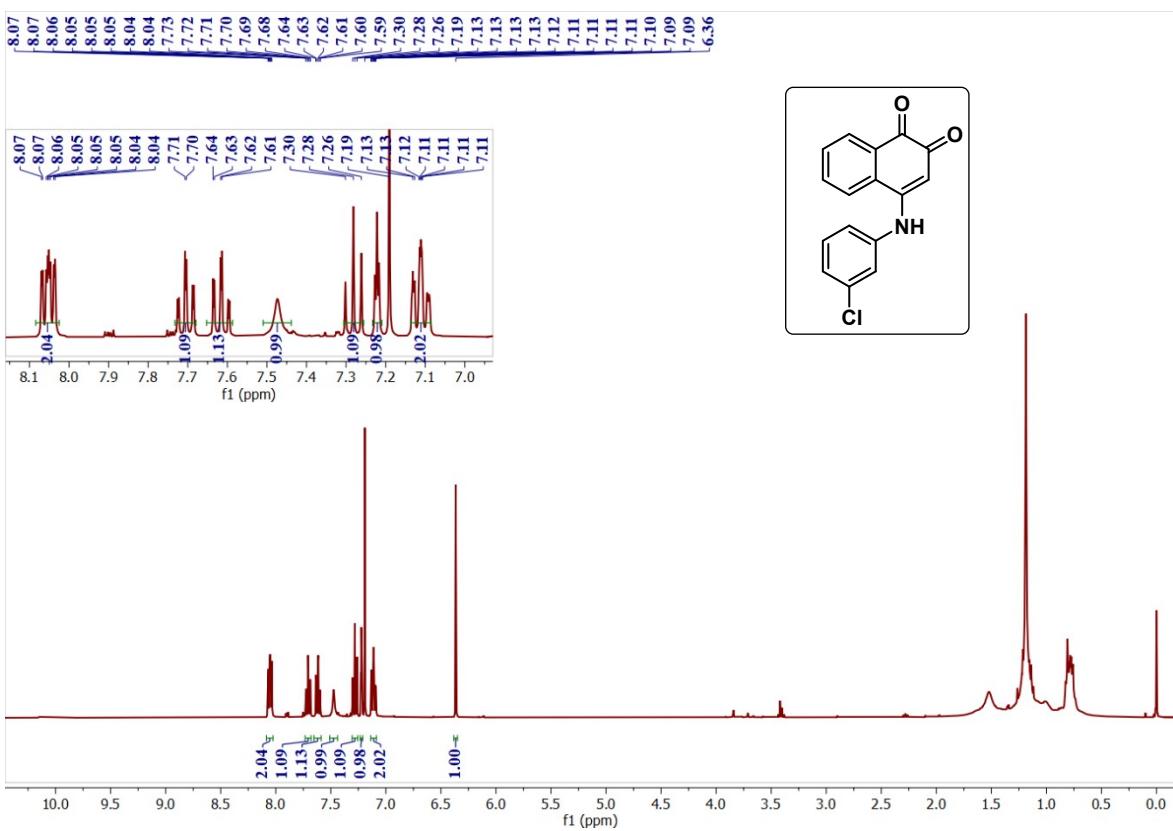


Figure S21: ^1H NMR (400MHz) Spectrum of **7f** in CDCl_3 at 28 °C.

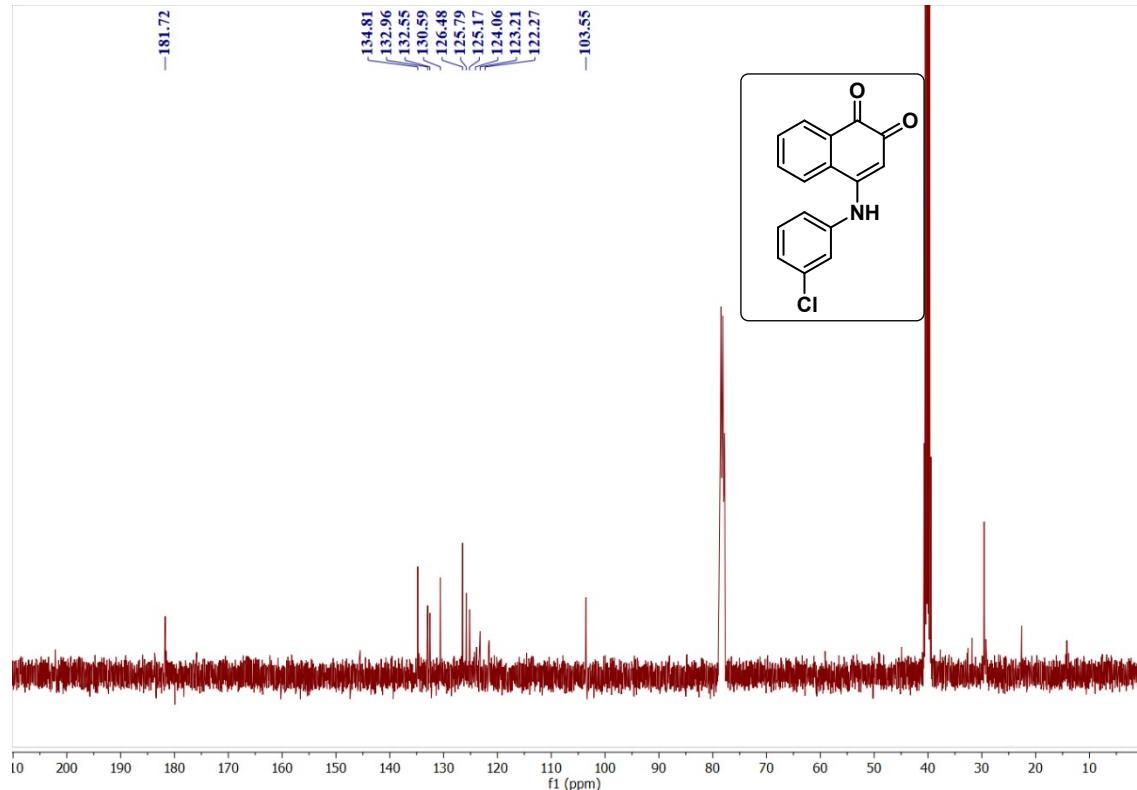


Figure S22: ^{13}C NMR (100 MHz) Spectrum of **7f** in $\text{CDCl}_3+\text{DMSO}-d_6$ at 28 °C.

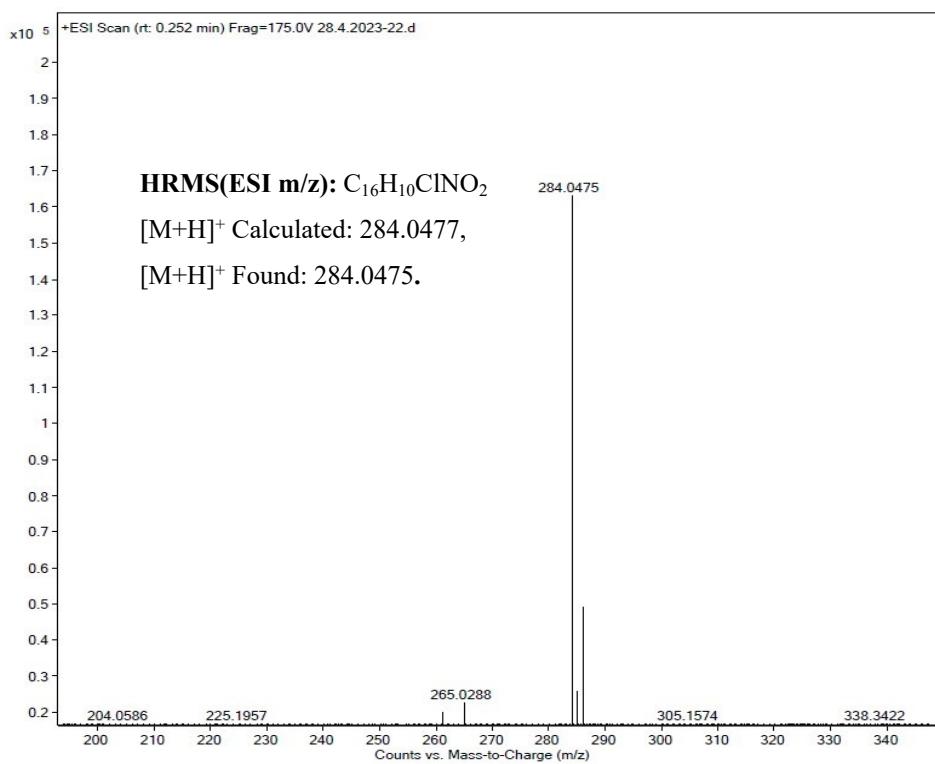


Figure S23: HRMS Spectrum of 7f.

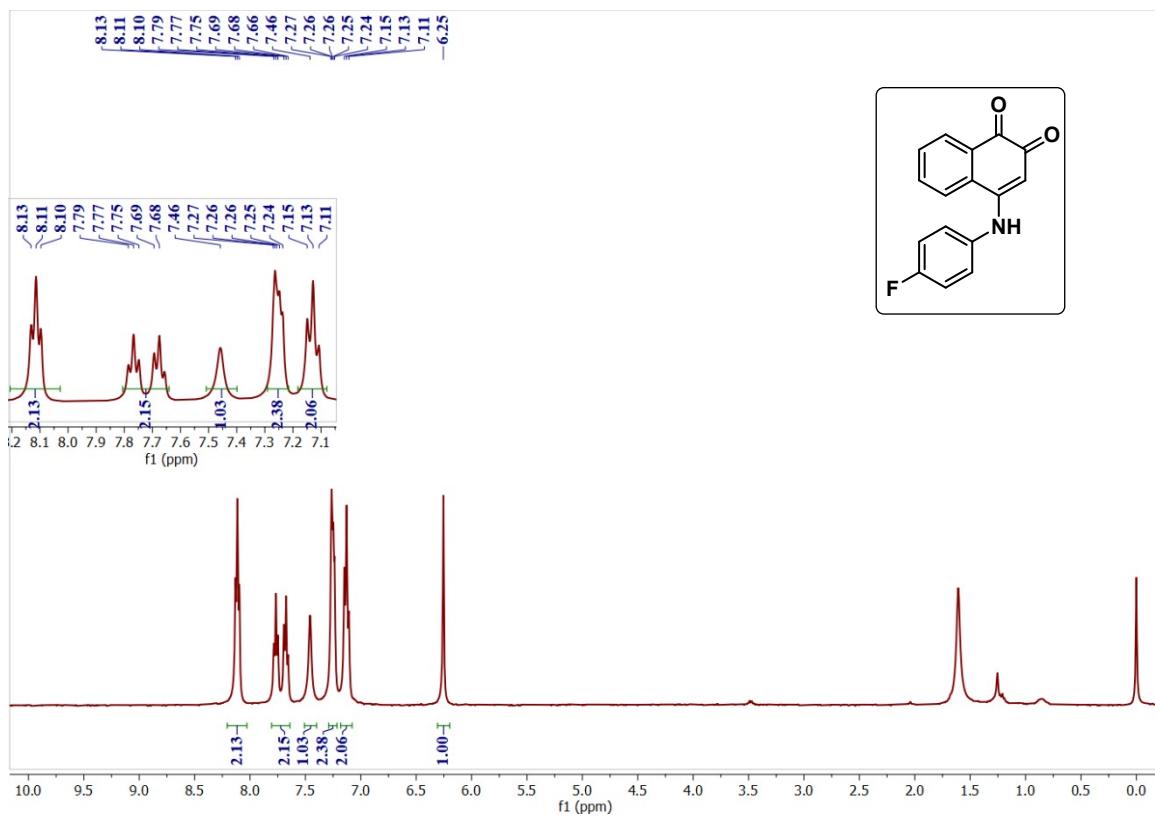


Figure S24: ¹H NMR (400MHz) Spectrum of 7g in CDCl₃ at 28 °C.

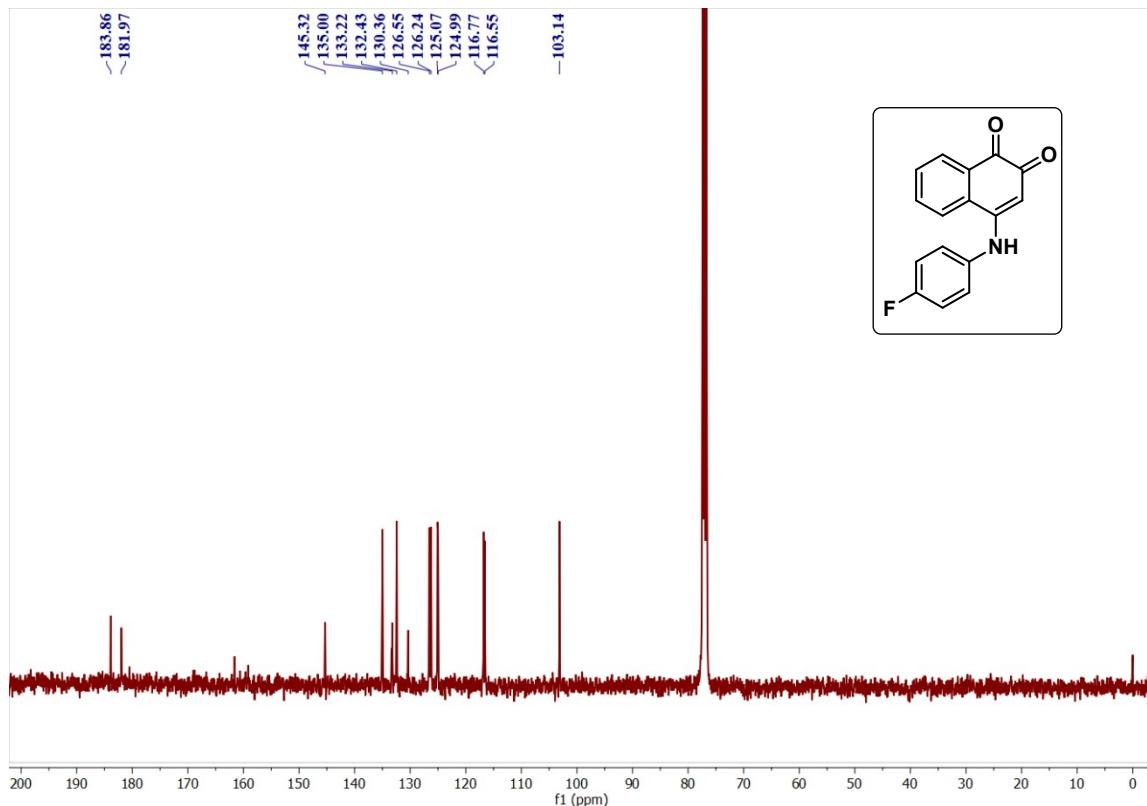


Figure S25: ^{13}C NMR (100 MHz) Spectrum of **7g** in CDCl_3 at 28 °C.

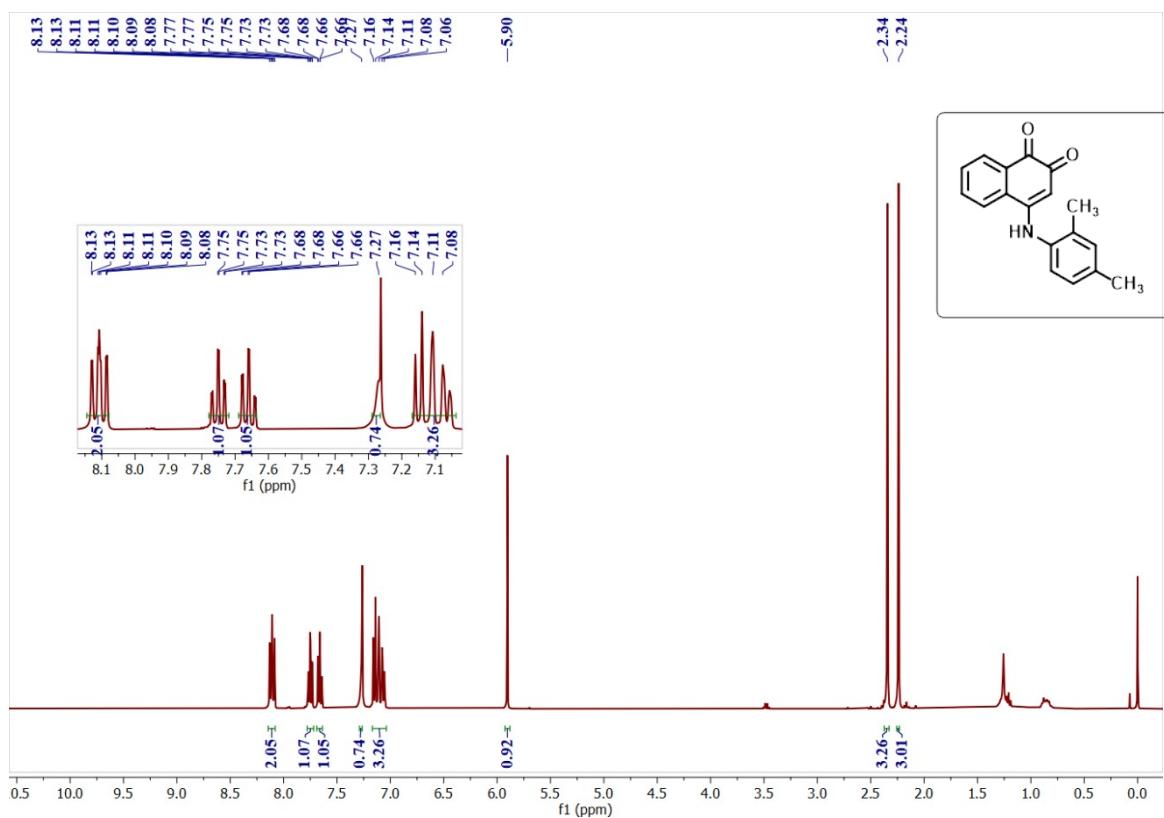


Figure S26: ^1H NMR (400MHz) Spectrum of 7h in CDCl_3 at 28 °C.

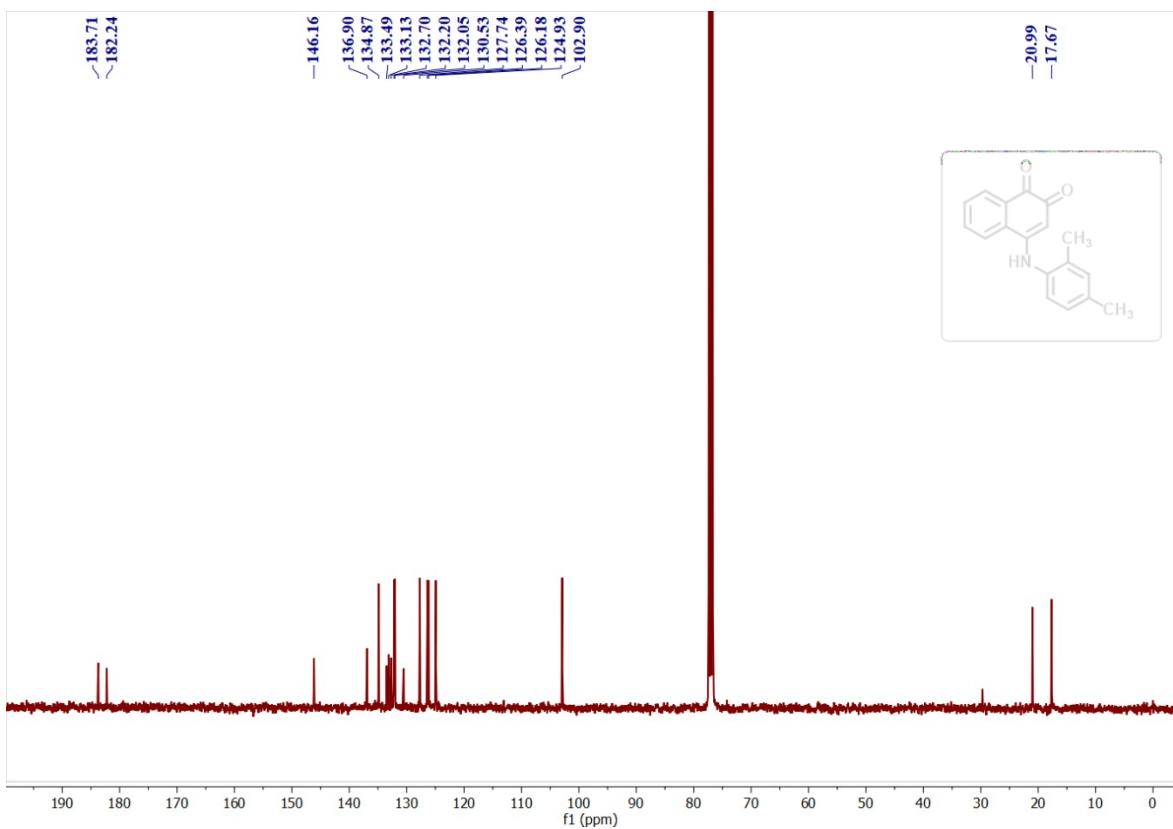
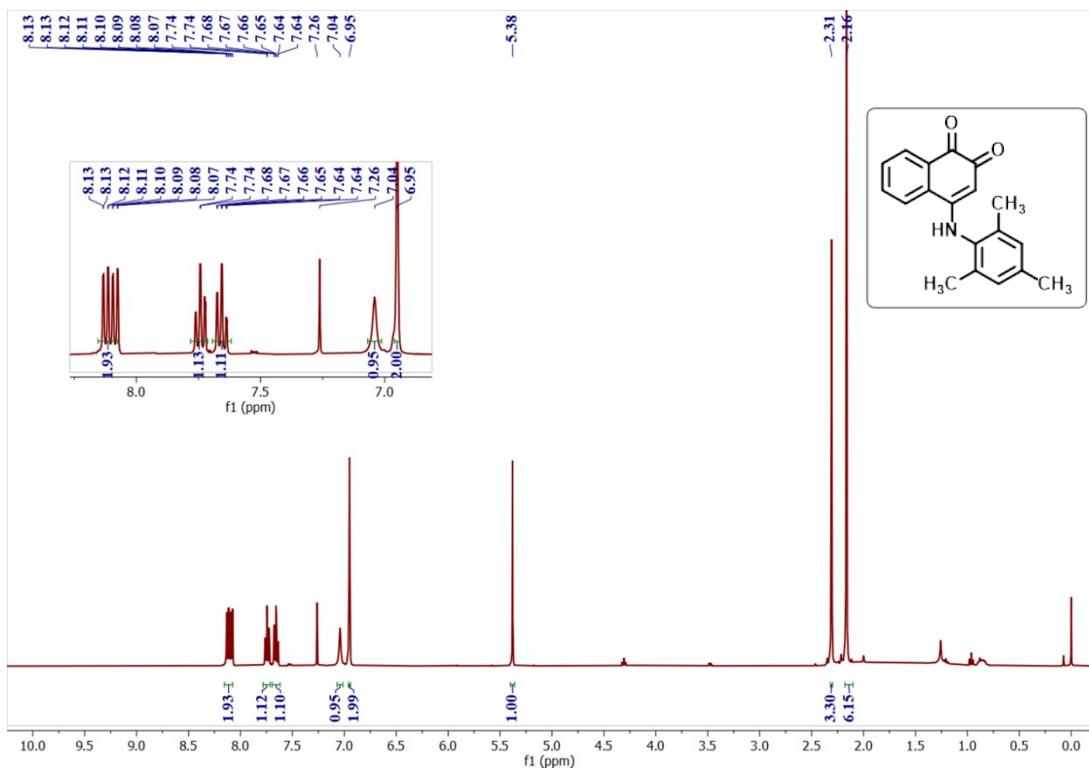


Figure S27: ^{13}C NMR (100 MHz) Spectrum 7h in CDCl_3 at 28 °C.



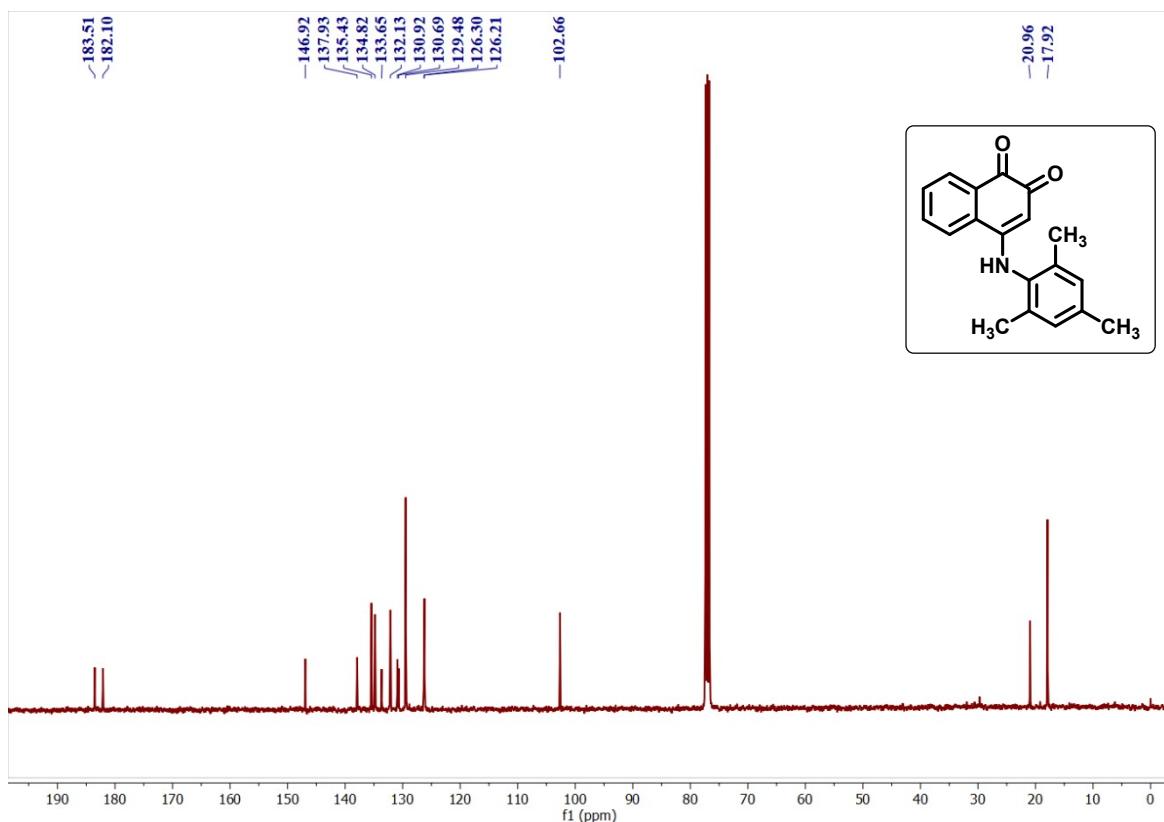


Figure S29: ^{13}C NMR (100 MHz) Spectrum 7i in CDCl_3 at 28 °C.

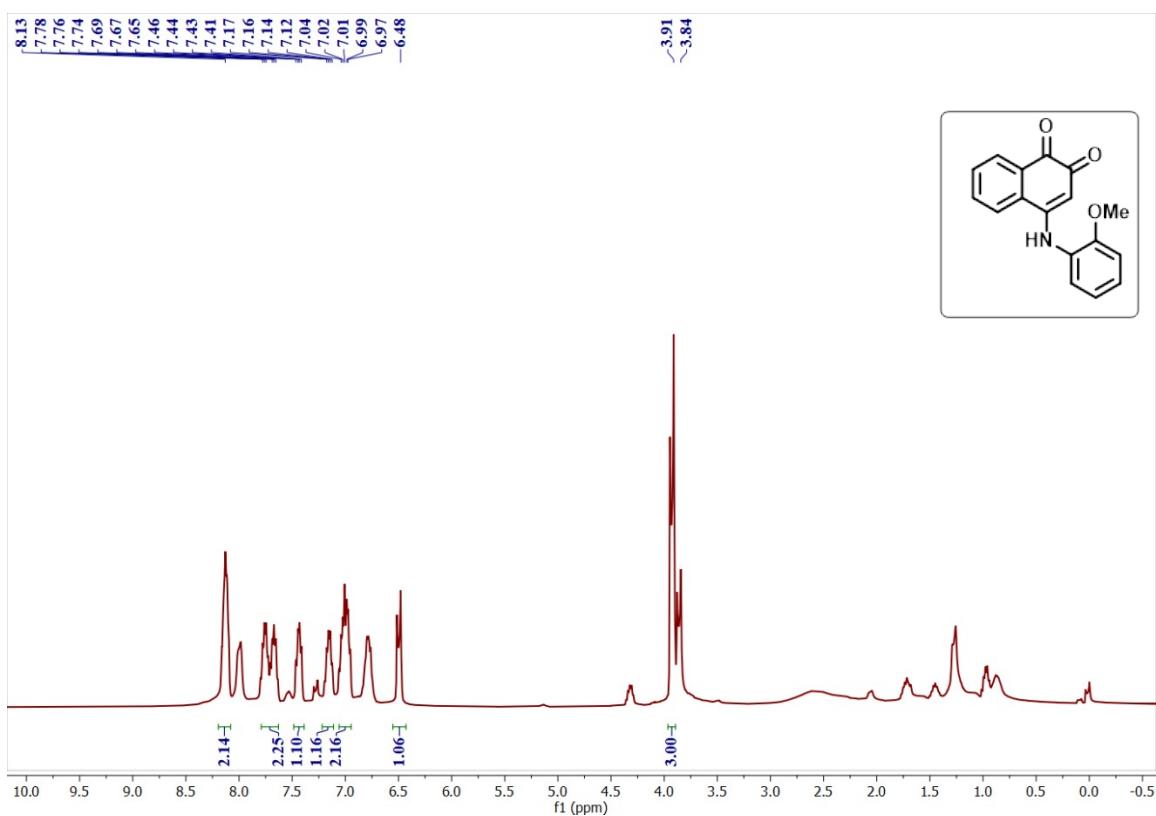


Figure S30: ^1H NMR (400MHz) Spectrum of 7j in CDCl_3 at 28 °C.

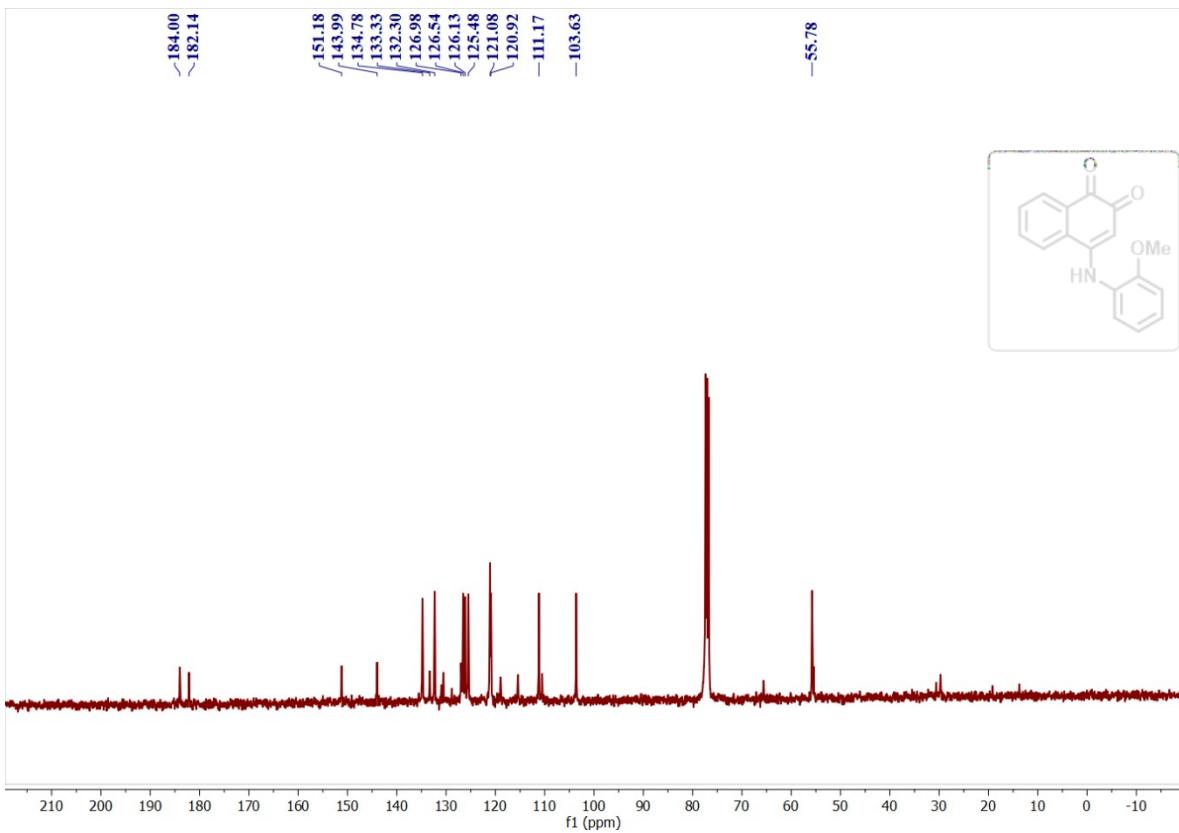


Figure S31: ^{13}C NMR (100 MHz) Spectrum 7j in CDCl_3 at 28 °C.

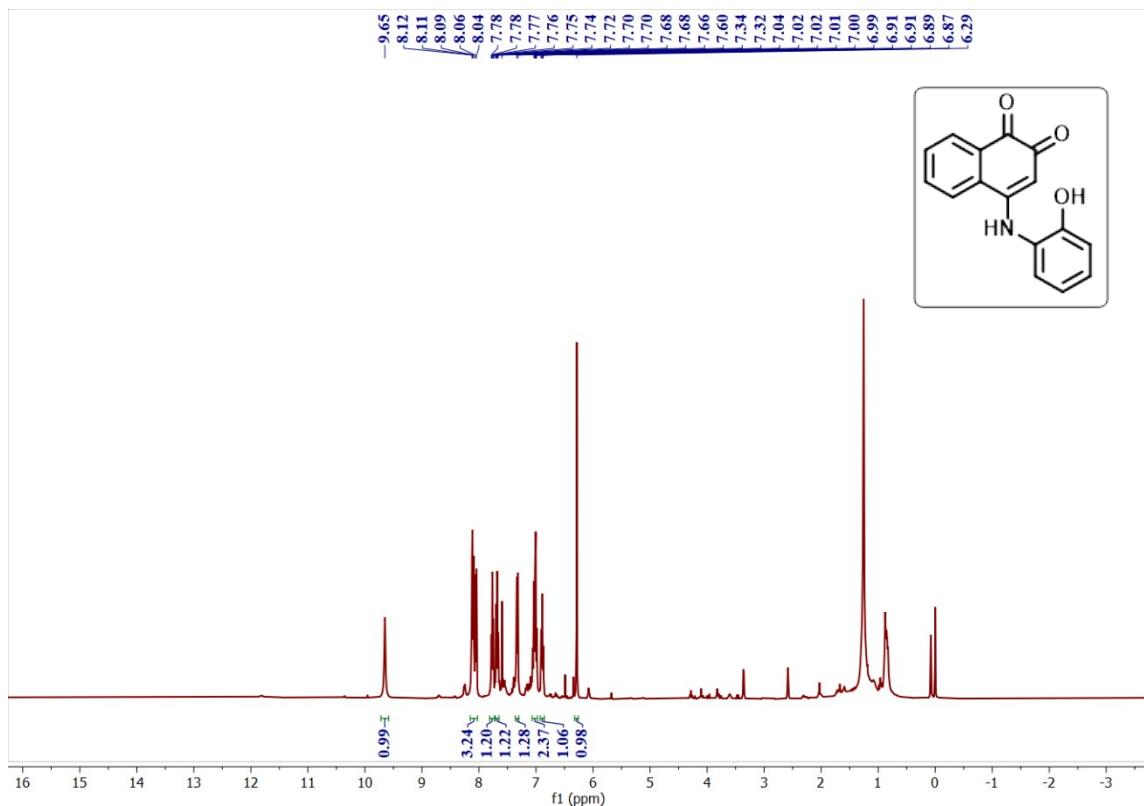


Figure S32: ^1H NMR (400MHz) Spectrum of 7k in $\text{CDCl}_3 + \text{DMSO}$ at 28 °C.

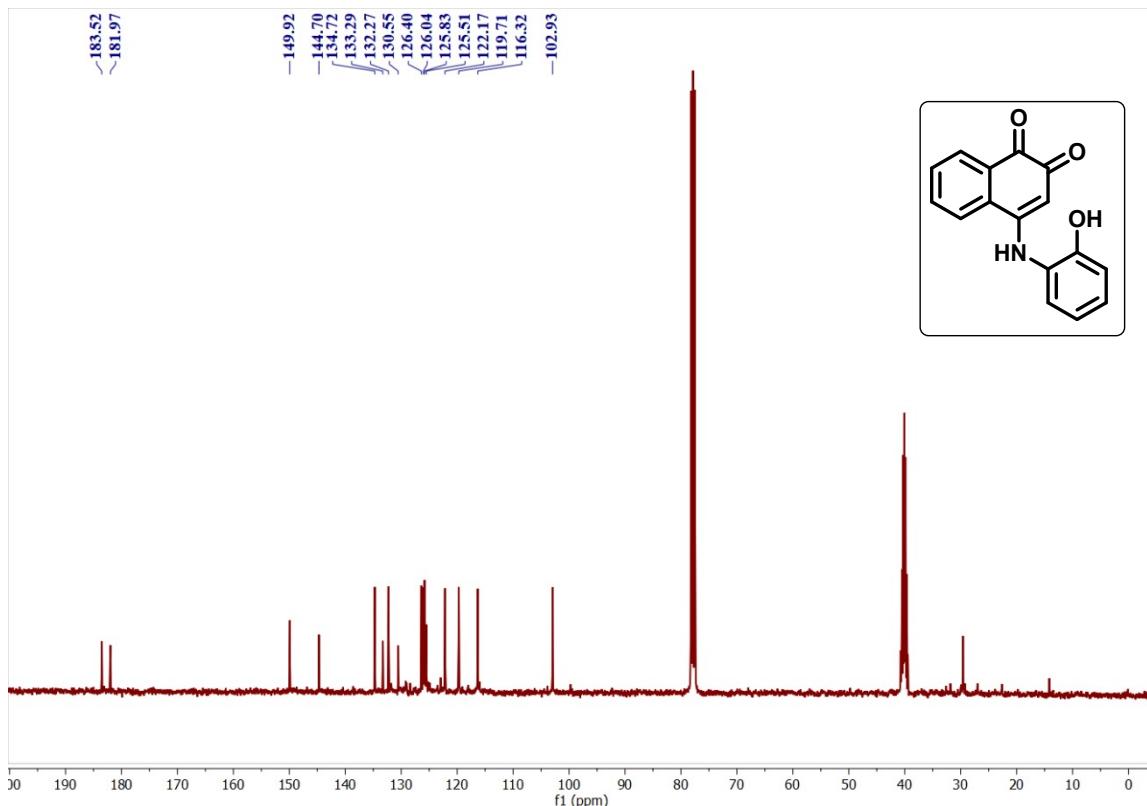


Figure S33: ^{13}C NMR (100 MHz) Spectrum 7k in $\text{CDCl}_3 + \text{DMSO}$ at 28 °C.

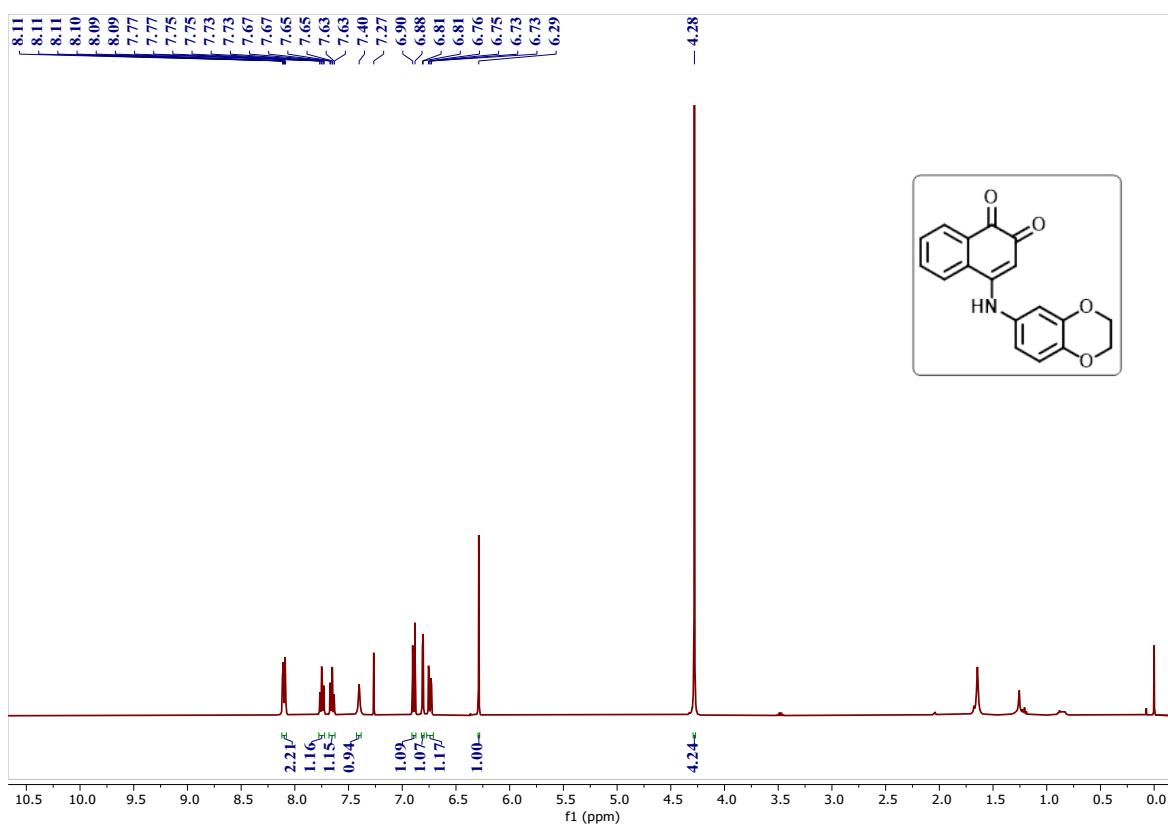


Figure S34: ^1H NMR (400MHz) Spectrum of 7l in CDCl_3 at 28 °C.

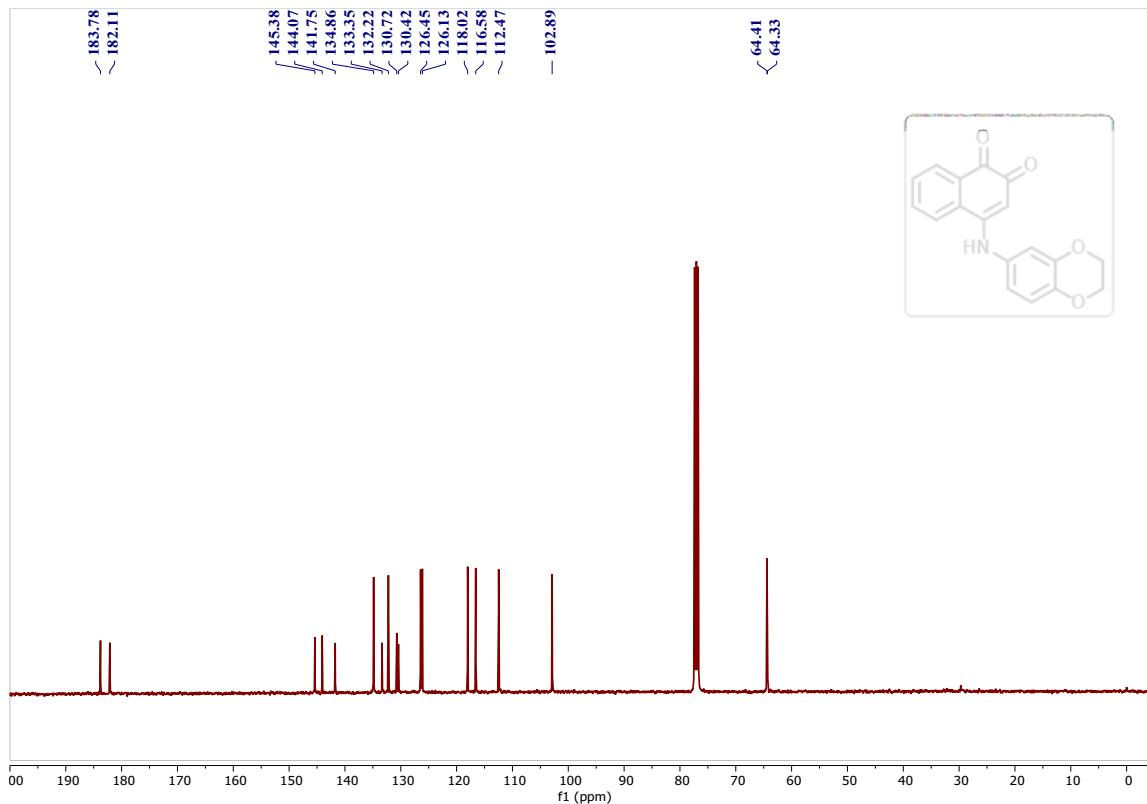


Figure S35: ^{13}C NMR (100 MHz) Spectrum 7l in CDCl_3 at 28 °C.

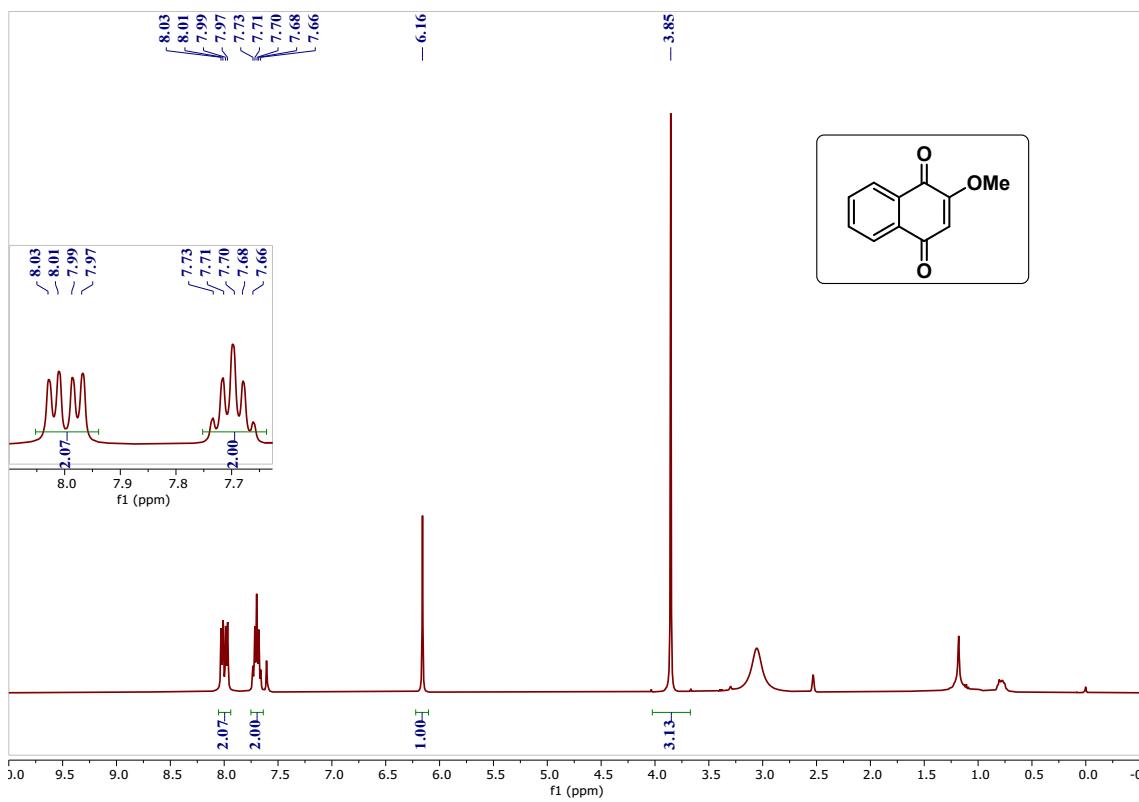


Figure S36: ^1H NMR (400MHz) Spectrum of **8a** in $\text{CDCl}_3+\text{DMSO}-d_6$ at 28 °C.

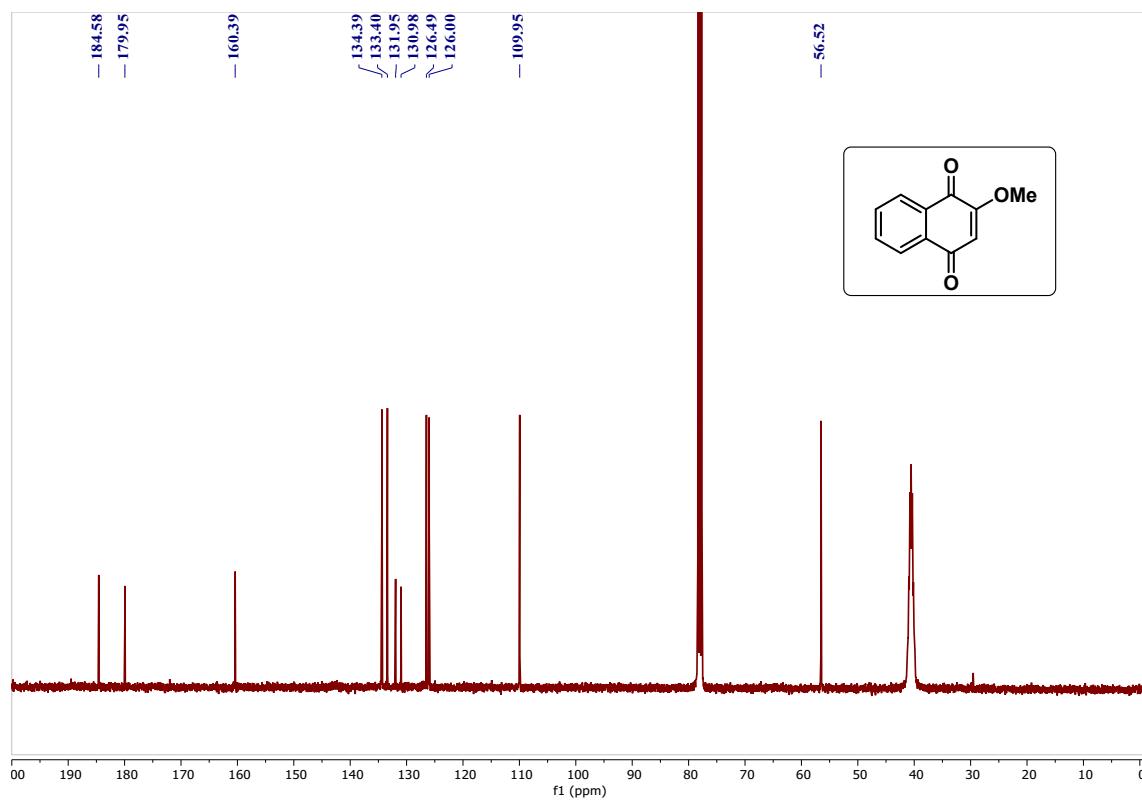


Figure S37: ^{13}C NMR (100 MHz) Spectrum of **8a** in $\text{CDCl}_3+\text{DMSO}-d_6$ at 28 °C.

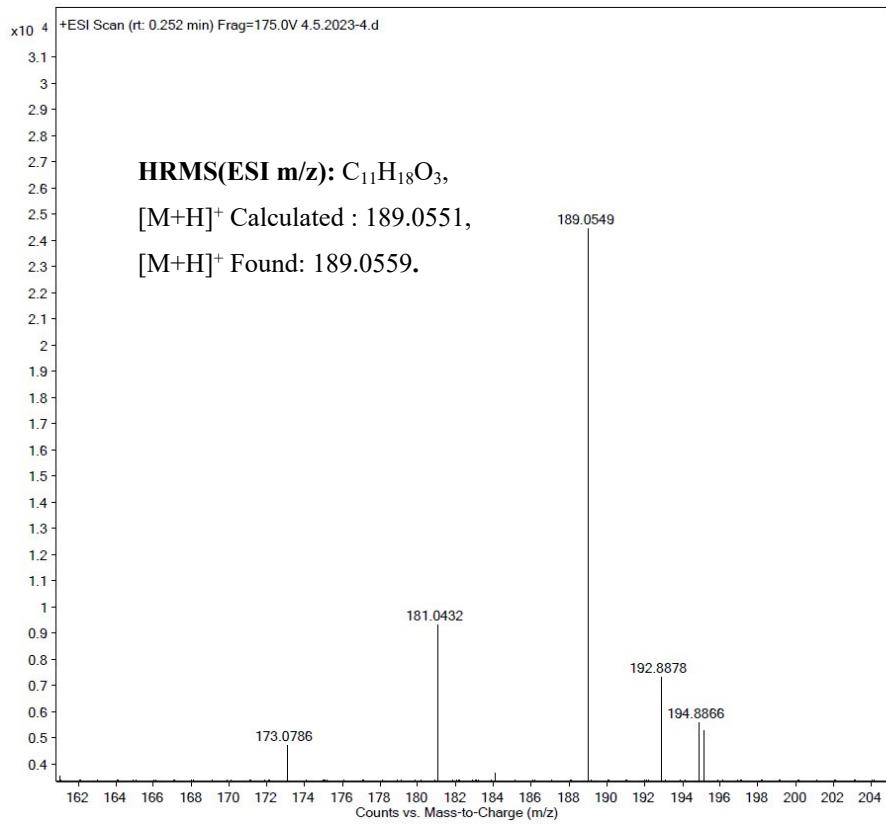


Figure S27: HRMS Spectrum of **8a**.

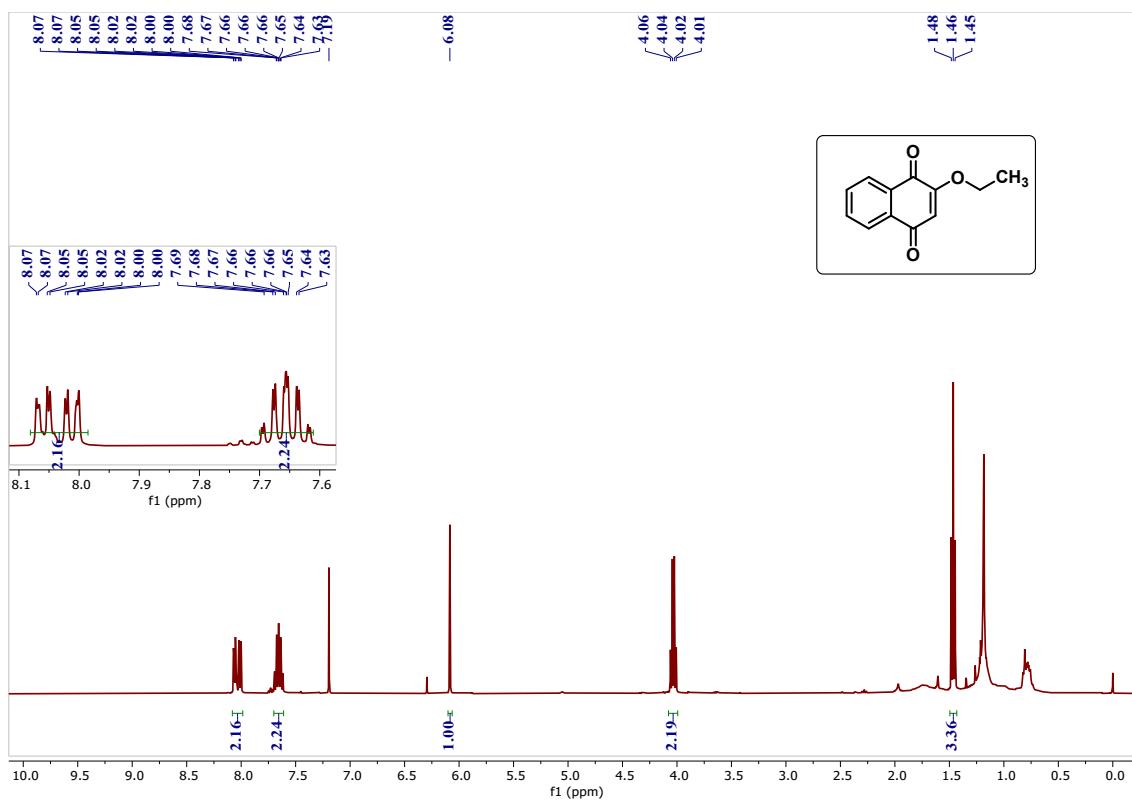


Figure S38: ^1H NMR (400MHz) Spectrum of **8b** in CDCl_3 at 28 °C.

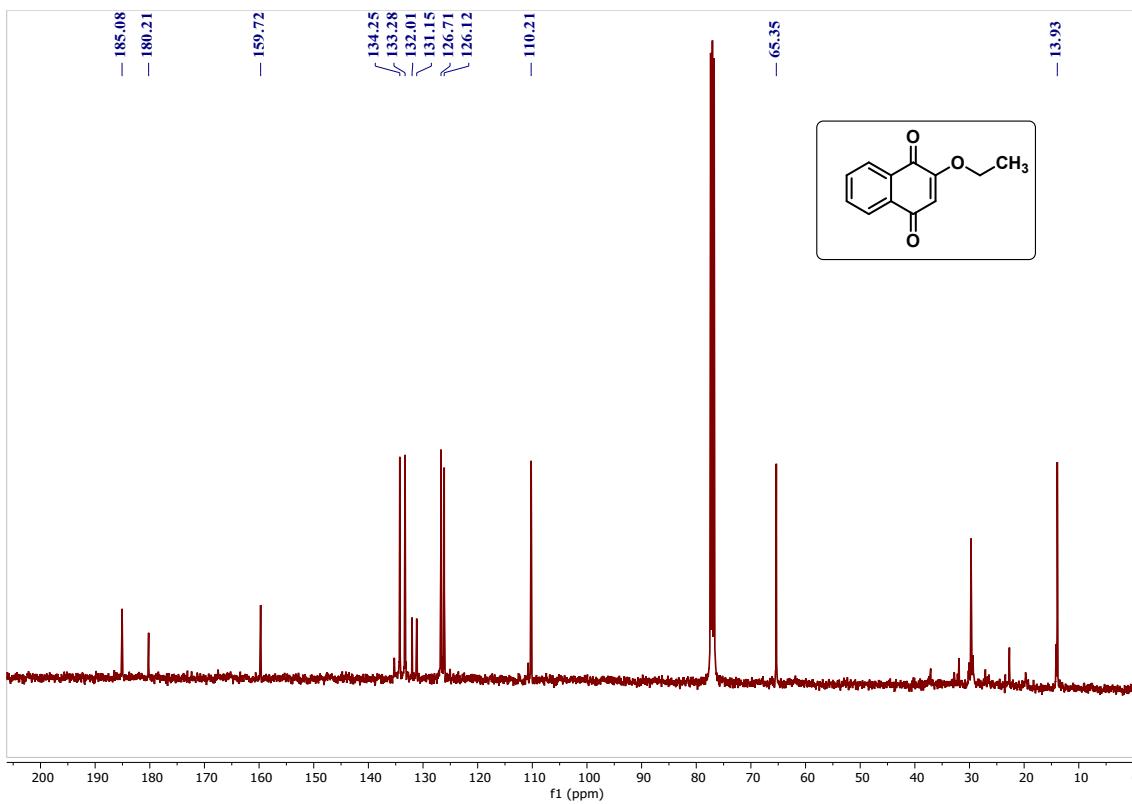


Figure S39: ^{13}C NMR (100 MHz) Spectrum of **8b** in CDCl_3 at 28 °C.

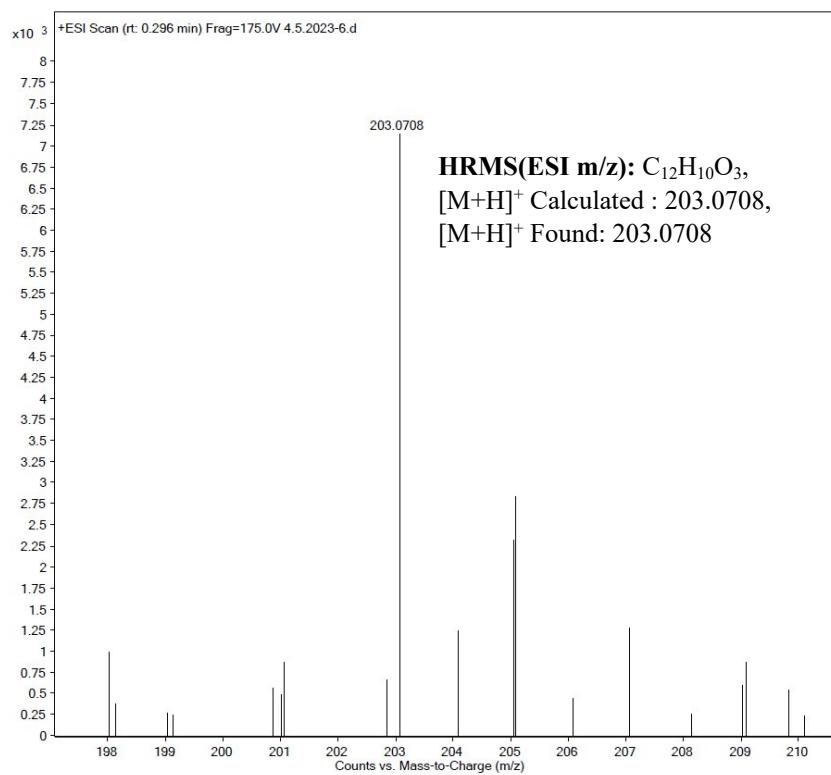


Figure S40: HRMS Spectrum of **8b**.

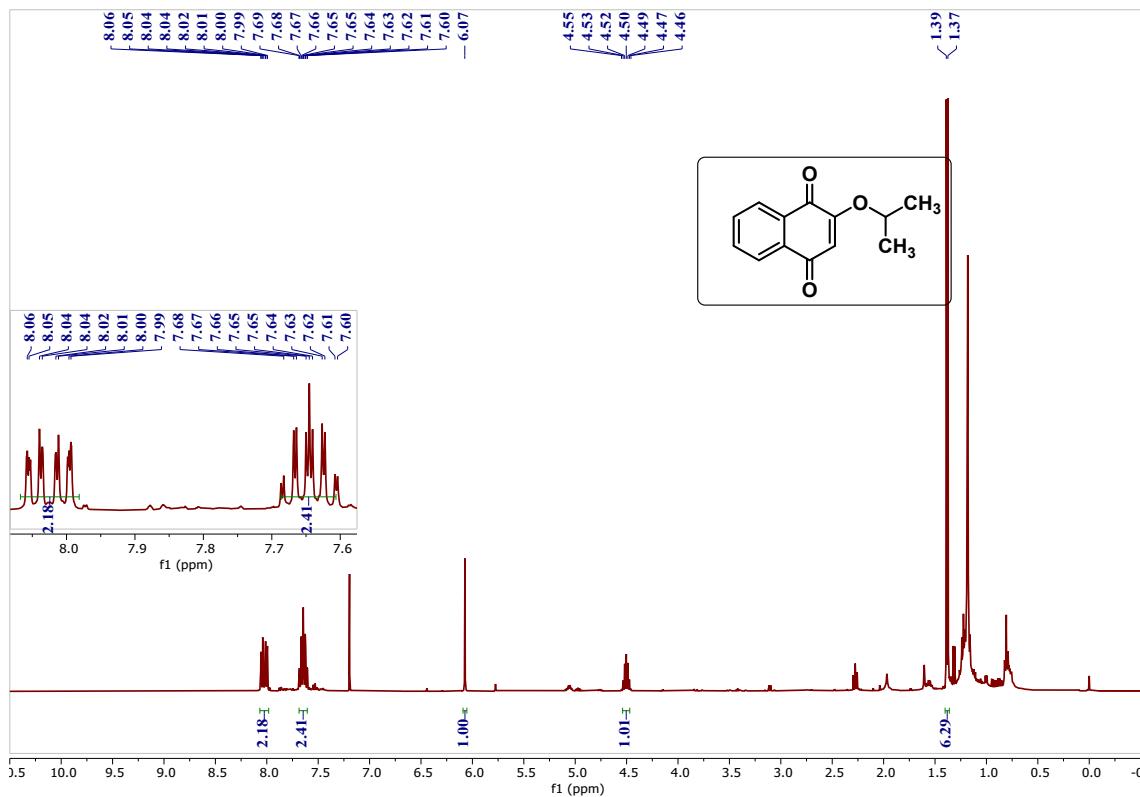


Figure S41: ¹H NMR (400MHz) Spectrum of **8c** in CDCl₃ at 28 °C.

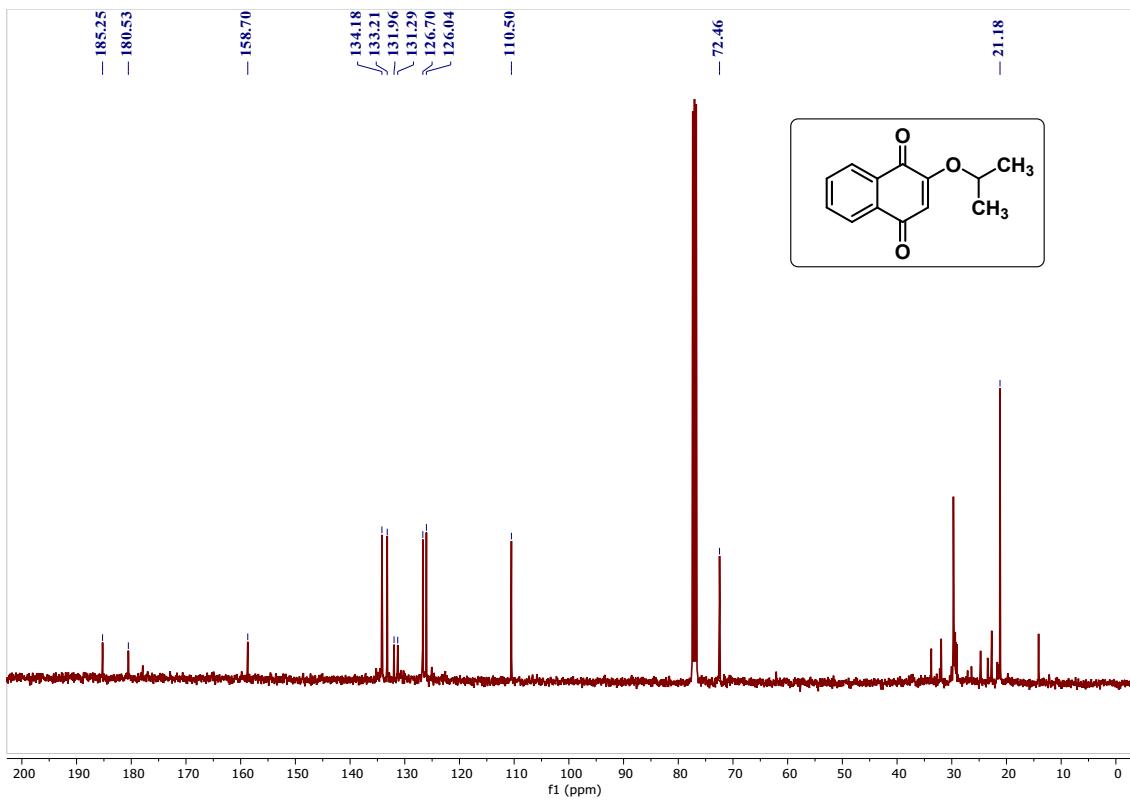


Figure S42: ^{13}C NMR (100 MHz) Spectrum of **8c** in CDCl_3 at 28 °C.

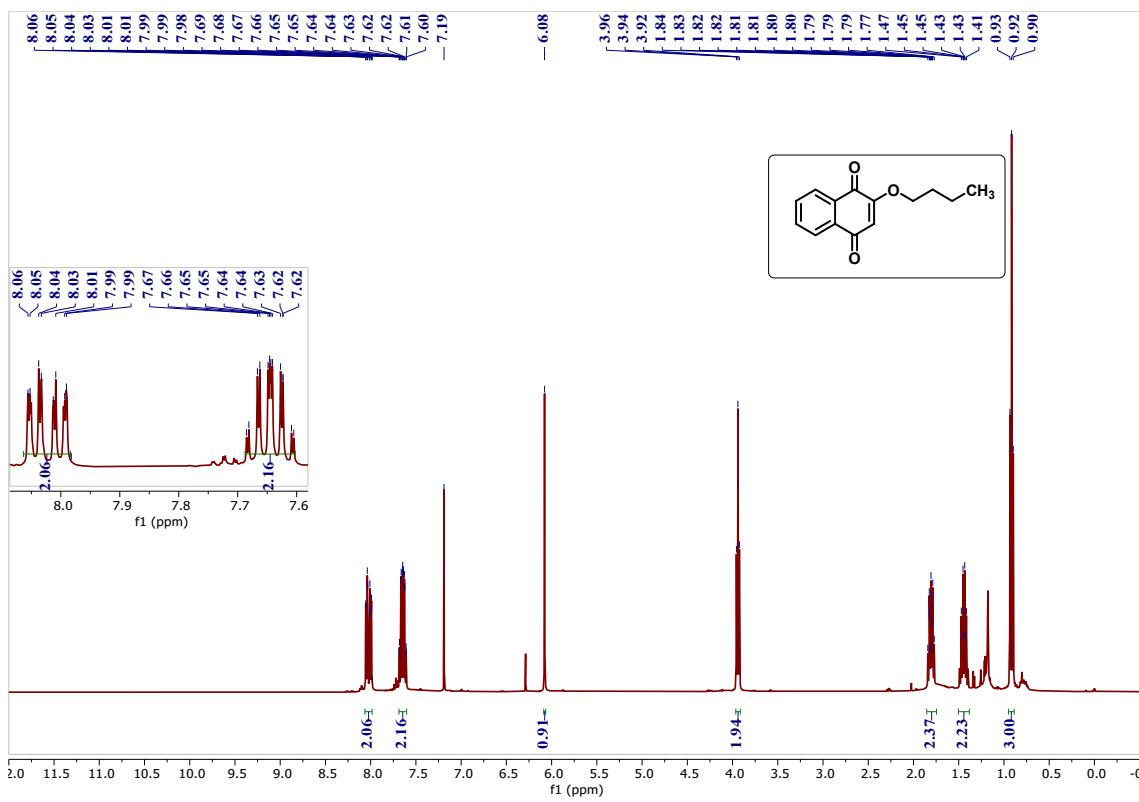


Figure S43: ^1H NMR (400MHz) Spectrum of **8d** in CDCl_3 at 28 °C.

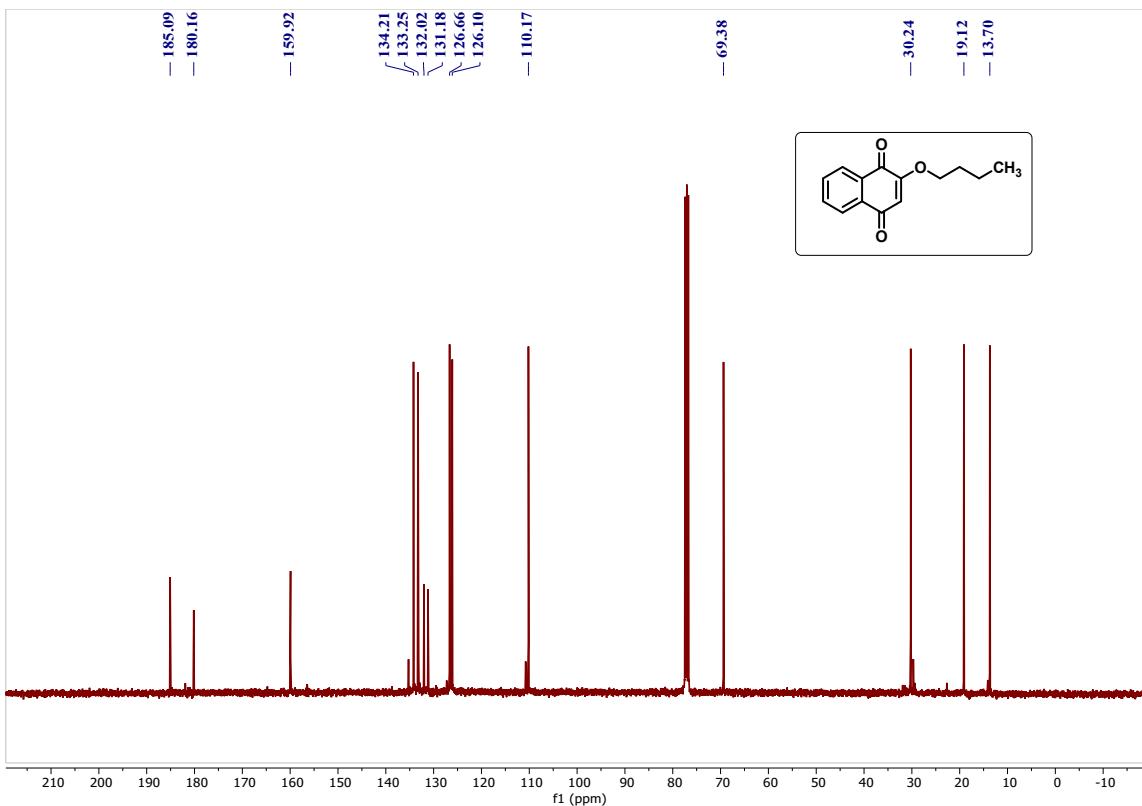


Figure S44: ^{13}C NMR (100 MHz) Spectrum of **8d** in CDCl_3 at 28 °C.

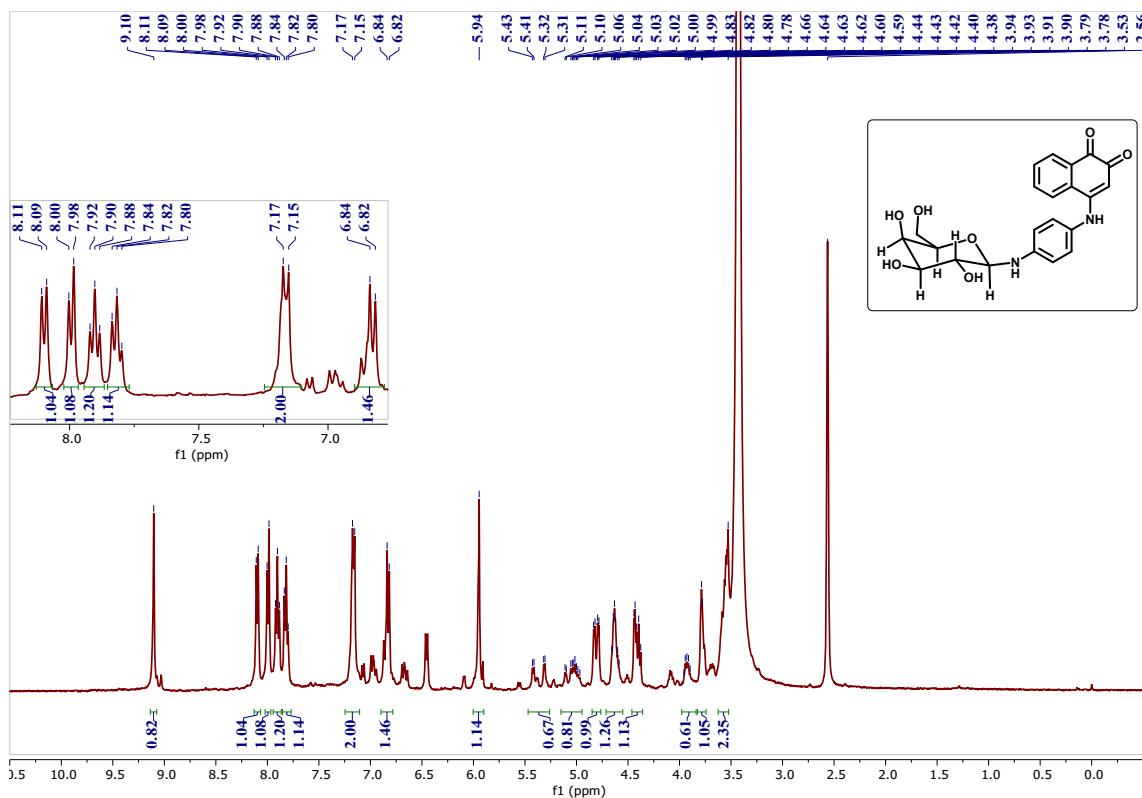


Figure S45: ^1H NMR (400MHz) Spectrum of **10** in $\text{DMSO}-d_6$ at 28 °C.

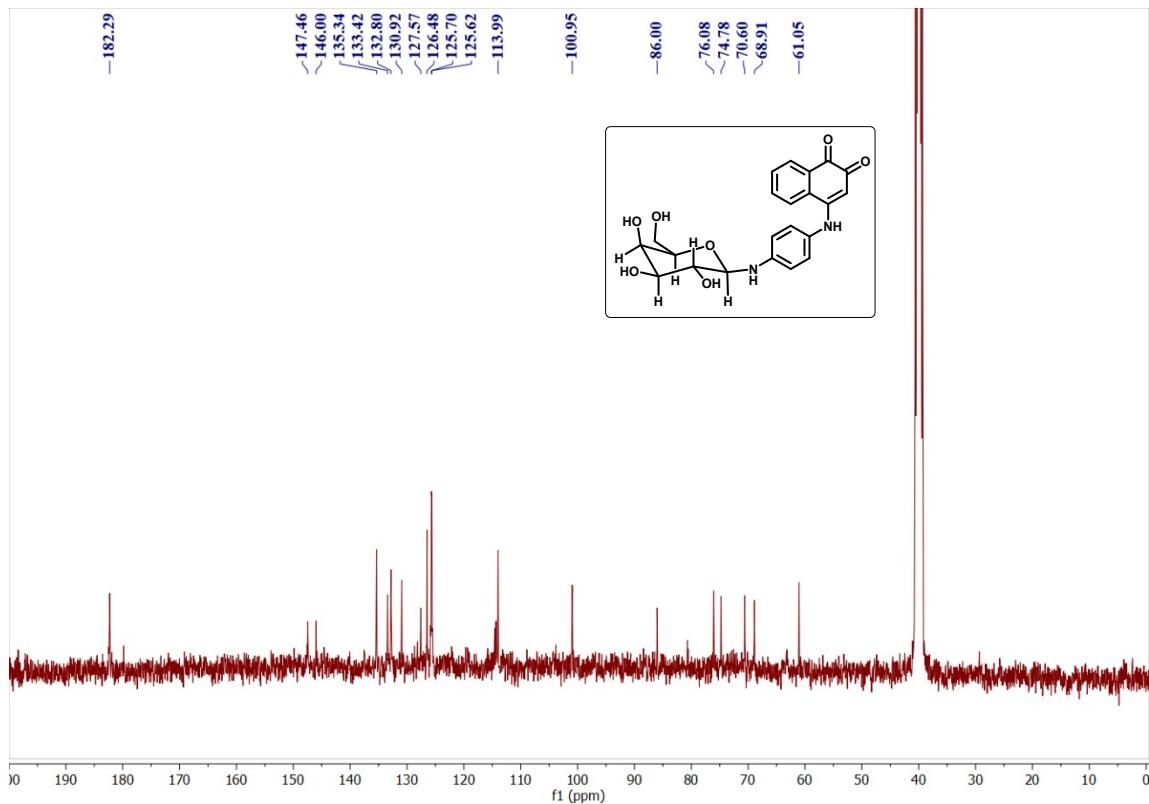


Figure S46: ^{13}C NMR (100 MHz) Spectrum of **10** in $\text{DMSO}-d_6$ at 28 °C.

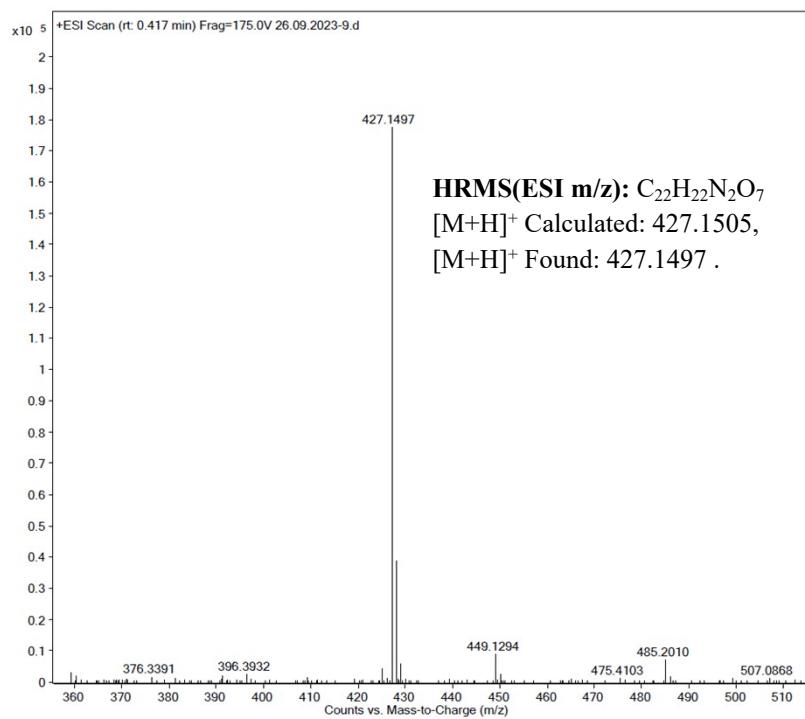


Figure S47: HRMS Spectrum of **10**.

7 IR

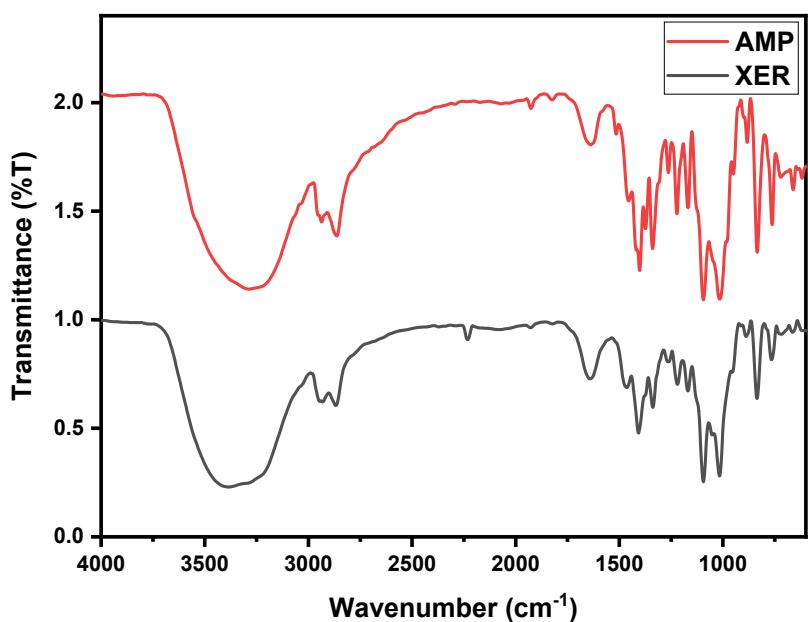


Figure S48: Comparison of IR spectrum of amorphous HMDBS and xerogel of HMDBS Eutectogels.

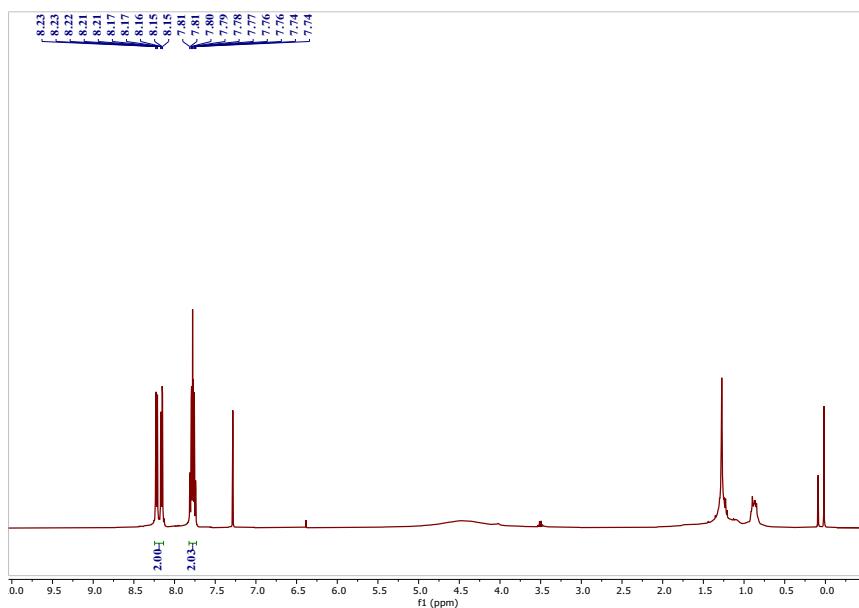


Figure S49: ¹H-NMR Spectra of compound 8a in CDCl₃ (400 MHz).⁸

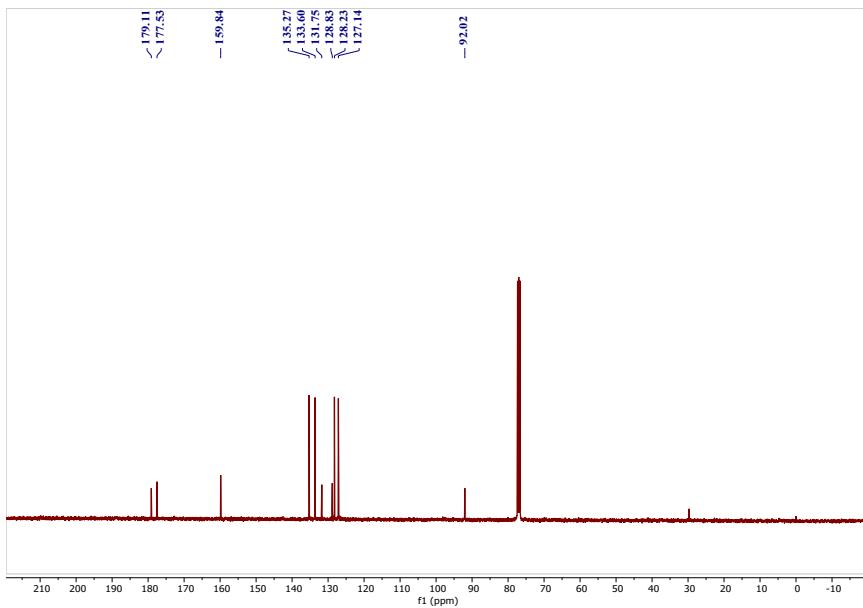


Figure S50. ¹³C-NMR Spectra of 8a in CDCl₃ (100 MHz)⁸

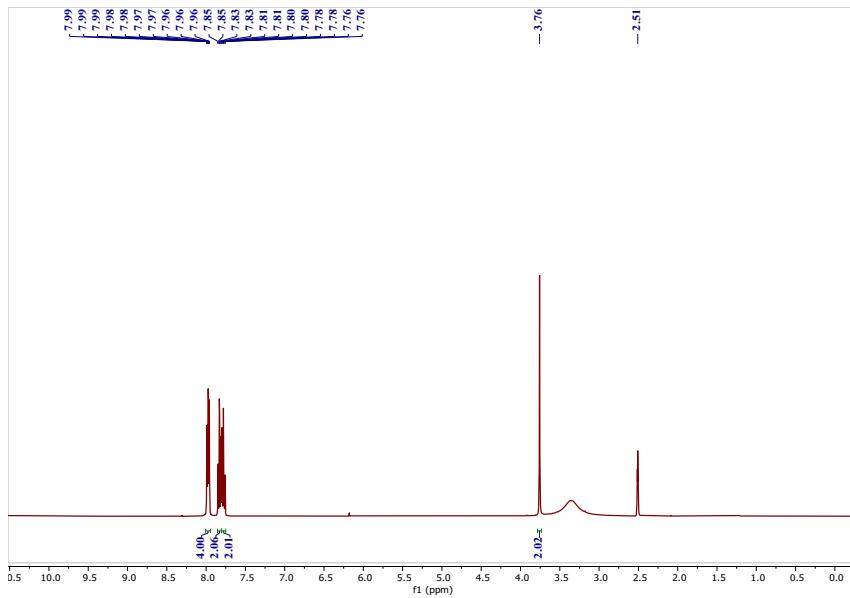


Figure S51: ¹H NMR spectra of Compound 8b in DMSO-d₆ (400 MHz).⁹

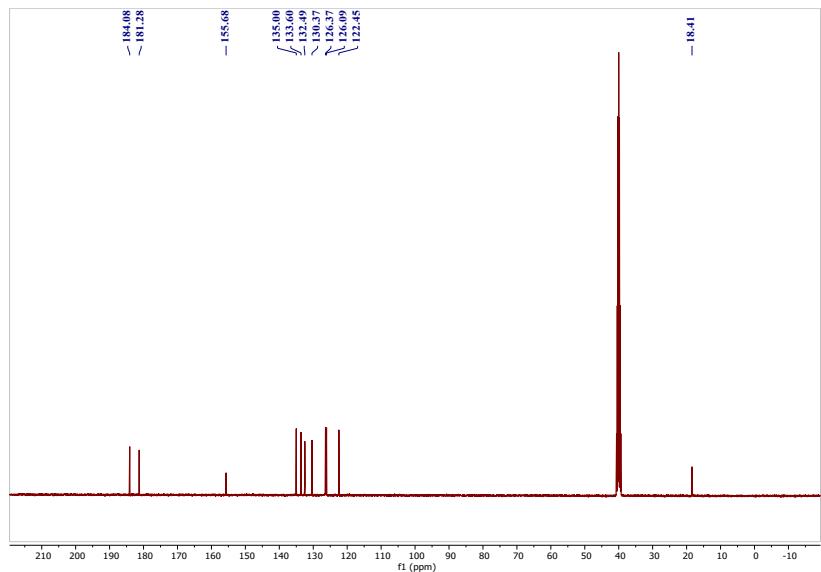


Figure S52. ^{13}C -NMR Spectra of **8b** in DMSO-d_6 (100 MHz).⁹

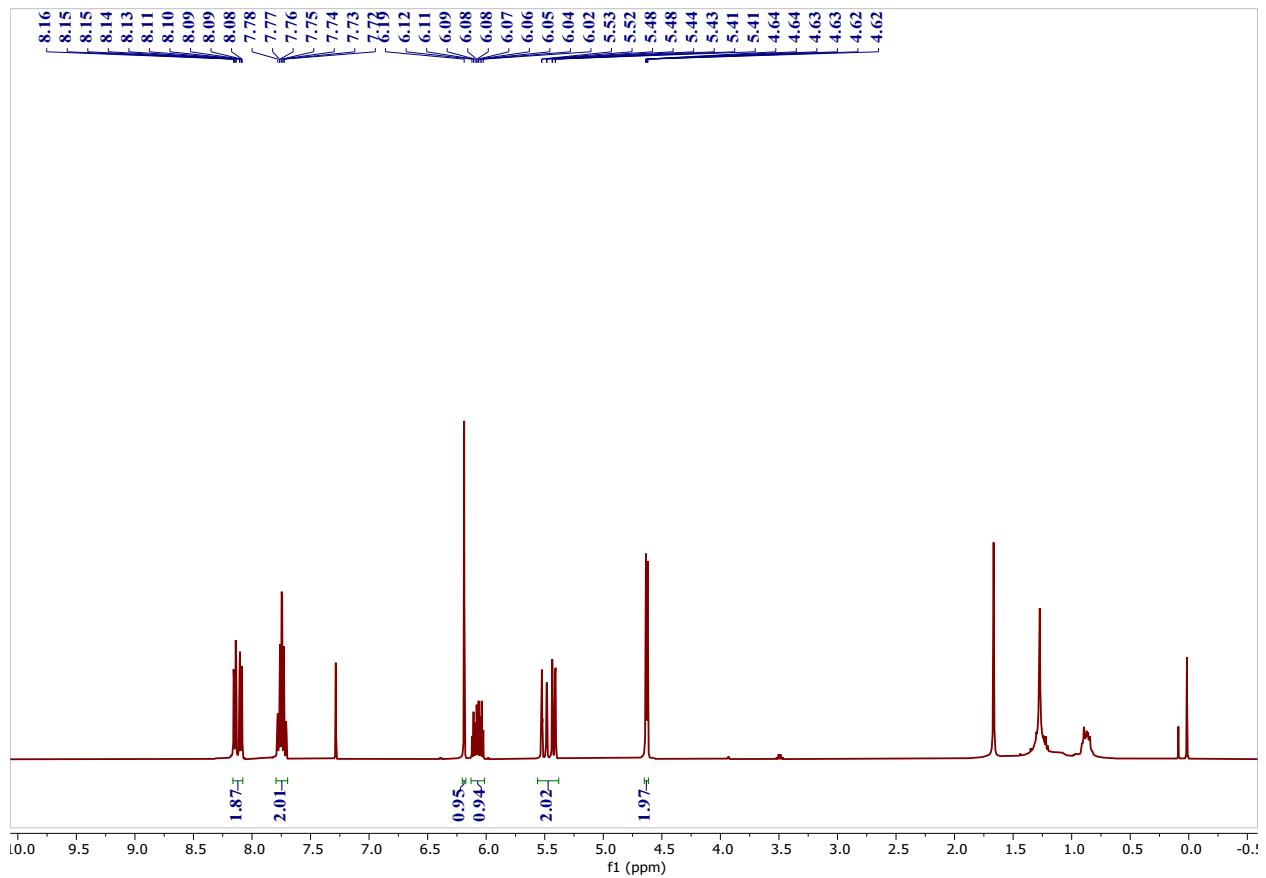


Figure S53 ^1H -NMR Spectra of **8c** in CDCl_3 (400 MHz).¹⁰

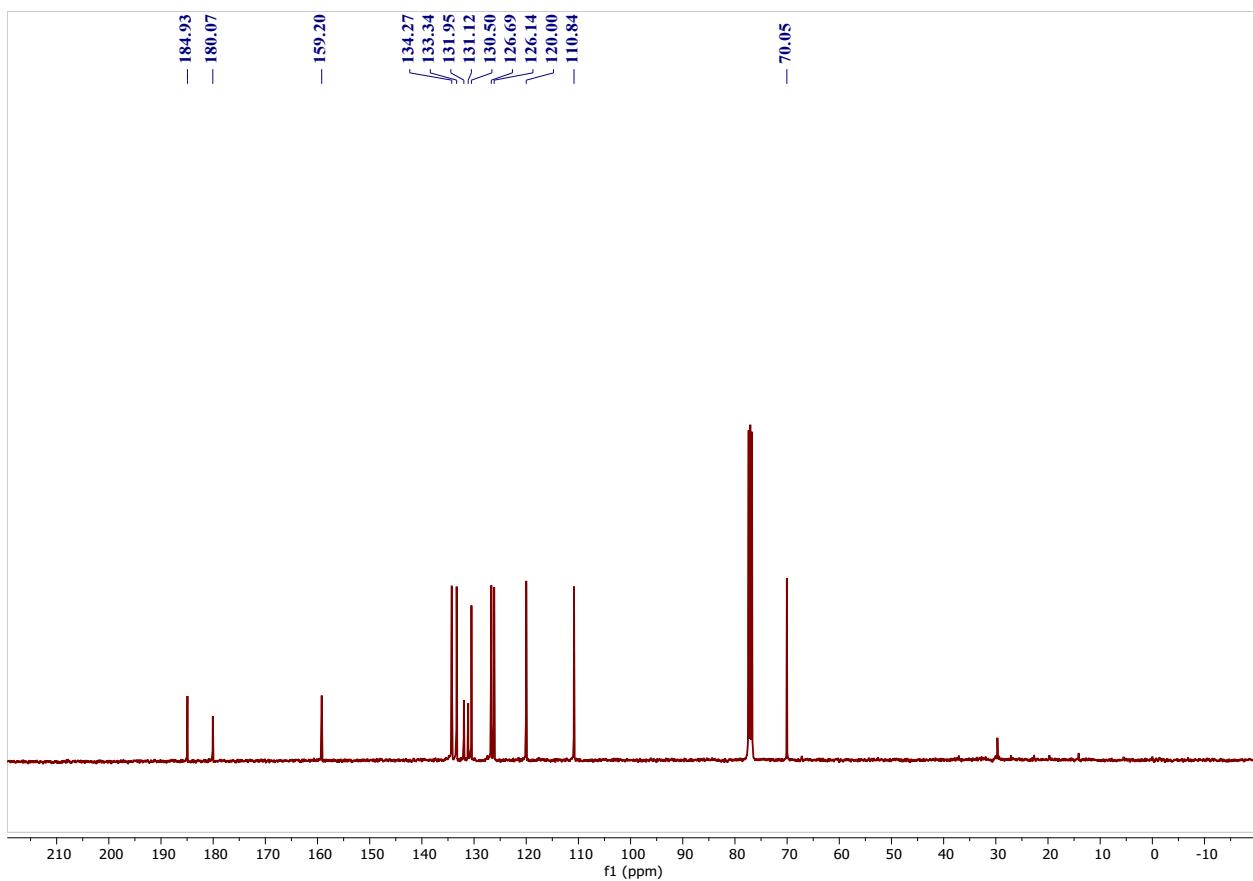


Figure S54. ^{13}C -NMR Spectra of 8c in CDCl_3 (100 MHz).¹⁰

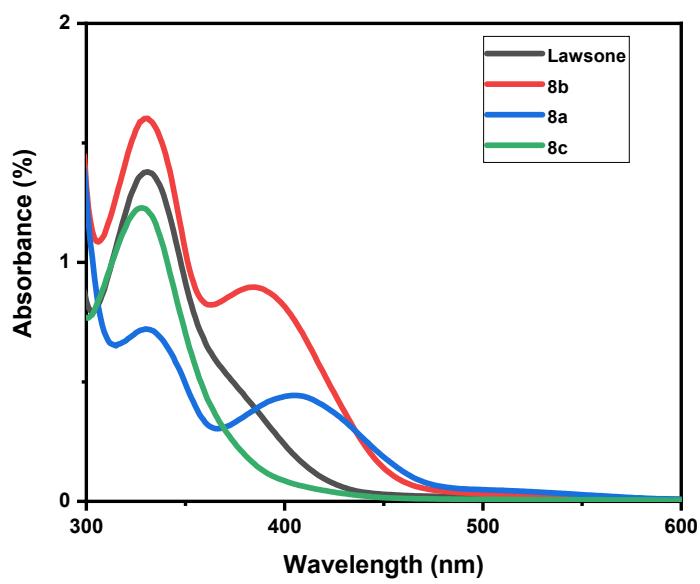


Figure S55. UV-vis spectra of lawsone, compound 8a, 8b, and 8c in THF.

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