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Supporting Information

Photoinduced metal-free trifluoro/perfluoroalkylation of heteroarenes

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1. General Information.

Nuclear magnetic resonance (NMR) spectra were recorded in deuterated solvents with residual protonated solvent signal as internal reference on Bruker-Ava-500. Chemical shifts are reported in parts per million using the solvent resonance internal standard (chloroform, 7.26 and 77.0 ppm). Data is reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constant, and integration. Electrospray and electron impact high resolution mass spectrometry was performed by Bruker mass spectrometer. The data is recorded as the ionization method followed by the calculated and measured masses. The Fluorescence emission intensities were measured on Agilent Cary Eclipse Fluorescence Spectrometer. Solvents for starting material preparation and coupling reactions were dried before use. Green LEDs (2.50 W, λ = 535 nm) Rebel LED, mounted on a 25 mm cool base was purchased from commercial supplier Luxeon Star LEDs Quadica Developments Inc.10-3447 30 Ave N. Lethbridge, Alberta T1H 7B5 Canada.

2. Experimental Section

2.1. Scope of Heteroarenes



11

l

1m

1n







1r

2.2: Preparation of Substrate

Materials: 1a-c, 1f, 1g, 1h, 1i, 1j were all prepared according to the previous reports.¹

1l², 1m³, 1o⁴, 1p⁵, 1s⁶ were all prepared according to the previous reports.

1d, 1e, 1k, 1n,1o, 1q were purchased from commercial sources and used without further purification and transformation.

2.2.1. General procedure for the Cbz protection of tryptamines



To a solution of tryptamine (1 equiv) in dichloromethane (0.1 M) was added a saturated aqueous solution of NaHCO₃ (5 equiv). The suspension was vigorously stirred and freshly distilled benzyl chloroformate (1.1 equiv) was added. The mixture was allowed to stir at rt for 2 h then the phases were separated and the aqueous phase extracted with dichloromethane. The combined organic extracts were washed with brine, dried over anhydrous sodium sulfate and concentrated in vacuo. The crude product was purified by flash column chromatography on silica gel using ethyl acetate and hexanes.

2.2.2 General procedure of synthesis of 3-substituted indole:



Indole-3-butyric acid for **1k** and Indole acetic acid for **1l** (1.0 equiv), respective alcohol (4.40 mmol, 1.1 equiv), N,N'-dicyclohexylcarbodiimide (1.2 equiv), and 4-dimethylaminopyridine (0.1 equiv) in a round-bottom flask. Dry THF was added, and the reaction mixture was stirred for 15 h at room temperature. After completion of reaction, the white precipitate was filtered off and the solution was concentrated by evaporation of the solvent. Purification by column chromatography on silica gel (EtOAc:hexane = 3:7) gave a solid **1h** and **1i**.

2.2.3 General procedure of synthesis of 3-substituted indole:



Into a 100 mL three-neck flask equipped with a magnetic stir bar, glass stopper and dropping funnel, dry toluene (3 mL), Et₃SiH (1.39 g, 12 mmol) and Cl₃CCOOH (1.23 g, 7.5 mmol) were added under N₂ atmosphere. The mixture was heated to 70 °C and treated with a solution of cyclododecanone (1.0 g, 5.5 mmol) and indole (586 mg, 5 mmol) in toluene (2 mL) dropwise. The reaction mixture was then stirred for 2 h at this temperature. After cooling to room temperature, the mixture was quenched with saturated Na₂CO₃, extracted with Et₂O. The combined organic layer was dried over anhydrous MgSO, filtrated, concentrated under vacuum, and the residue was purified by flash column chromatography (petroleum ether/EtOAc = 30:1) to give a white solid (740 mg, 52 %).

3-cyclododecyl-1H-indole



¹H NMR (500 MHz, CDCl₃) δ 7.93 – 7.83 (m, 1H), 7.66 (d, *J* = 7.9 Hz, 1H), 7.35 (d, *J* = 8.1 Hz, 1H), 7.18 (t, *J* = 7.5 Hz, 1H), 7.11 (t, *J* = 7.5 Hz, 1H), 6.97 (s, 1H), 3.13 (p, *J* = 6.4 Hz, 1H), 1.92 – 1.82 (m, 2H), 1.66 (ddt, *J* = 13.2, 7.4, 5.5 Hz, 2H), 1.56 (ddd, *J* = 17.9, 7.7, 3.3 Hz, 3H), 1.50 – 1.43 (m, 6H), 1.42 – 1.31 (m, 10H). ¹³C NMR (126 MHz, CDCl₃) δ 136.6, 127.4, 122.3, 121.9, 120.4, 119.5, 119.0, 111.2, 77.4, 77.2, 76.9, 31.1, 30.7, 24.0, 24.0, 23.9, 23.7,

22.9. HRMS (ESI/QTOF), m/z: [M-H] ⁻ Calcd. C₂₀H₂₈N 282.2222; Found 282.2224.

2.3 Preparation of perfluoroalkanesulfinates.



Perfluoroiodine compounds ($R_f = C_4F_9$, C_6F_{13} , C_8F_{17}) (10 mmol) was bubbled into CH₃CN (6 mL) in a round bottom flask at 0°C. NaHCO₃ (1.68 g, 20 mmol, 2 equiv.), Na₂S₂O₄ (1.58 g, 10 mmol, 2 equiv), and H₂O (10 mL) were added. The reaction mixture was stirred at room temperature overnight and then extracted with ethyl acetate (3 × 50 mL). The combined organic layers were dried over anhydrous MgSO₄ and concentrated to dryness. The residue was washed with diethyl ether (3 × 20 mL) and dried in vacuum to give 1c/1d as a white solid. The spectral data matched with those reported in the literature.⁹



3: Photo Reaction Setup





- (a) Whole Setup Setup
- (b) DC Regulator controlling Current and voltage
- (c) Box- cover with holes for RB and RB illuminated by Green LEDs
- (d) Uncovered box (6' * 6') with 4 LEDs

Each RB was at 5cm distance from LEDs.

4: General procedures for the synthesis of 3 and spectral data



Scheme 1: Preparation of trifluoromethylated/perfluoroalkylated heteroarenes

0.2 mmol of **1** (1.0 equiv.), trifluoro/perfluoro alkyl sulfinates (0.6 mmol, 3 equiv.) were taken in a long neck round bottom flask and 2.5 mol % of Eosin Y was added into it followed by ammonium peroxodisulfate (3.0 equiv), and the RB was capped with septum. The mixture was degassed and filled with N₂ (three times). Subsequently 0.1M MeCN solution was added into the reaction mixture via syringe. The reaction mixture was then irradiated with green LED for 24 h. After completion (monitored through TLC), reaction was quenched with saturated NaHCO₃ solution and extracted with ethyl-acetate (3 x 10 mL), washed with brine solution. After removal of solvent in vacuo, the product was purified by silica gel chromatography using EtOAc-hexane (3:7 to 5:5) as eluent to provide the desired product.

Analytical Data for Products

benzyl (2-(2-(trifluoromethyl)-1H-indol-3-yl)ethyl)carbamate (3a): Physical state: White



liquid; Yield: 52 mg (72 %). ¹H NMR (500 MHz, CDCl₃) δ 8.46 (s, 1H), 7.69 (d, J = 8.0 Hz, 1H), 7.41 – 7.31 (m, 7H), 7.17 (t, J = 7.5 Hz, 1H), 5.11 (s, 2H), 4.84 (d, J = 7.4 Hz, 1H), 3.51 (q, J = 6.7 Hz, 2H), 3.14 (t, J = 7.4 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 156.54, 136.71,

135.40, 134.17, 128.64, 128.22, 127.56, 125.14, 122.54(C-F, $1J_{C-F} = 268.38$ Hz), 121.02, 120.66, 120.41 (C-F, $1J_{C-F} = 268.38$ Hz), 111.94, 111.49, 66.80, 41.74, 24.59. ¹⁹F NMR (471 MHz, CDCl₃) δ -58.06. HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₁₉H₁₆N₂O₂F₃ 361.1164; Found 361.1172.

tert-butyl (2-(2-(trifluoromethyl)-1H-indol-3-yl)ethyl)carbamate (3b): Physical state: White liquid; Yield: 44 mg (67 %). ¹H NMR (400 MHz, CDCl₃) δ 8.59 (s, 1H), 7.69 (d, J = 8.3 Hz, 1H), 7.41 (dd, J = 8.3, 1.0 Hz, 1H), 7.34 – 7.30 (m, 1H), 7.20 – 7.16 (m, 1H), 4.63 (s, 1H), 3.41 (s, 2H), 3.10 (t, J = 6.9 Hz, 2H), 1.44 (s, 9H). ¹³C NMR (126 MHz, CDCl₃) δ 155.95, 135.29, 127.49, 125.17 (C-F, $1J_{C-F} = 269.6$ Hz), 124.93, 123.04 (C-F, $1J_{C-F} = 269.6$ Hz), 122.39, 122.10, 120.90 (C-F, $1J_{C-F} = 269.6$ Hz), 120.75, 120.73, 120.35, 118.76 (C-F, $1J_{C-F} = 269.6$ Hz), 115.36, 111.78, 41.15, 28.38, 28.27, 24.53. ¹⁹F NMR (376 MHz, CDCl₃) δ -57.91. HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₁₆H₁₈N₂O₂F₃ 327.1321; Found 327.1329.

2-(trifluoromethyl)-1H-indole-3-carbaldehyde (3c): Physical state: White solid; Yield: 21

CHO mg (50 %). ¹H NMR (500 MHz, DMSO-*d*₆) δ 13.41 (s, 1H), 10.23 (s, 1H), R_{H}° CF₃ 8.24 (d, *J* = 8.0 Hz, 1H), 7.61 – 7.59 (m, 1H), 7.43 – 7.41 (m, 1H), 7.37 – 7.34 (m, 1H). ¹³C NMR (126 MHz, DMSO-*d*₆) δ 184.24, 135.36, 131.35 (C-F, 2*J*_{C-F} = 39.0 Hz), 131.04 (C-F, 2*J*_{C-F} = 39.0 Hz), 130.72 (C-F, 2*J*_{C-F} = 39.0 Hz), 130.41 (C-F, 2*J*_{C-F} = 39.0 Hz), 125.82, 124.33, 124.02 (C-F, 1*J*_{C-F} = 270.9 Hz), 123.84, 121.99, 121.87 (C-F, 1*J*_{C-F} = 270.9 Hz), 119.71 (C-F, 1*J*_{C-F} = 270.9 Hz), 115.45, 113.19. ¹⁹F NMR (471 MHz, CDCl₃) δ -56.65. HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₁₀H₅NO₂F₃ 212.0323; Found 212.0333.

3-nitro-2-(trifluoromethyl)-1H-pyrrolo[2,3-β]pyridine (3d): Physical state: White solid; NO₂ Vield: 21 mg (45 %). ¹H NMR (500 MHz, DMSO-*d*₆) δ 8.69 (dd, J = 8.0, 1.6 Hz, 1H), 8.63 (dd, J = 5.0, 1.6 Hz, 1H), 7.59 (dd, J = 8.1, 5.0 Hz, 1H), 4.01 (s, 1H). ¹³C NMR (126 MHz, DMSO) δ 144.01, 143.98, 132.56, 124.15, 123.24 (C-F, $1J_{C-F} = 270.9$ Hz), 121.09 (C-F, $1J_{C-F} = 270.9$ Hz), 120.04, 118.94 (C-F, $1J_{C-F} = 270.9$ Hz), 116.78 (C-F, $1J_{C-F} = 270.9$ Hz), 116.36. ¹⁹F NMR (471 MHz, CDCl₃) δ -60.37. HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₁₁H₂N₃OF₂ 230.0166; Found 230.0166.

2-(trifluoromethyl)-1H-indole-3-carbonitrile (3e): Physical state: White liquid; Yield: 22 mg (52 %). ¹H NMR (500 MHz, CDCl₃) δ 9.40 (s, 1H), 7.84 (d, J = 8.1 Hz, 1H), 7.55 (d, J = 8.3 Hz, 1H), 7.48 (t, J = 7.7 Hz, 1H), 7.40 (t, J = 7.5 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 134.54,

126.98, 126.95, 123.89, 120.83 (C-F, $1J_{C-F} = 270.9 \text{ Hz}$), 120.75, 118.68 (C-F, $1J_{C-F} = 270.9 \text{ Hz}$), 112.88, 112.69. ¹⁹F NMR (471 MHz, CDCl₃) δ -60.31. HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd.C₁₀H₄N₂F₃ 209.0327; Found 209.0334.

3-cyclohexyl-2-(trifluoromethyl)-1H-indole (3f): Physical state: White liquid; **Yield:** 37 mg (69 %). ¹**H NMR (500 MHz, CDCl₃)** δ 8.20 (s, 1H), 7.88 (d, J = 8.2 Hz, 1H), 7.39 (d, J = 8.3 Hz, 1H), 7.29 (t, J = 7.4 Hz, 1H), 7.17 – 7.13 (m, 1H), 3.06 – 3.00 (m, 1H), 1.98 – 1.80 (m, 7H), 1.48 – 1.36 (m, 3H). ¹³**C NMR (126 MHz, CDCl₃)** δ 135.61, 126.51, 124.43, 124.06,

124.03, 123.37 (C-F, $1J_{C-F} = 269.64$ Hz), 122.39, 121.23 (C-F, $1J_{C-F} = 269.64$ Hz), 120.13, 36.39, 32.99, 27.13, 26.35. ¹⁹F NMR (471 MHz, CDCl₃) δ -57.47. HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₁₅H₁₅NF₃ 266.1157; Found 266.1162.

3-cyclododecyl-2-(trifluoromethyl)-1H-indole (3g): Physical state: White liquid; Yield: 45



mg (64%). ¹H NMR (500 MHz, CDCl₃) δ 8.19 (s, 1H), 7.80 (d, J = 8.2 Hz, 1H), 7.40 (d, J = 8.3 Hz, 1H), 7.29 (t, J = 7.7 Hz, 1H), 7.14 (t, J = 7.6 Hz, 1H), 3.30 (t, J = 6.9 Hz, 1H), 2.10 – 2.05 (m, 2H), 1.71 – 1.66 (m, 2H), 1.63 (d, J = 6.2 Hz, 1H), 1.46 (q, J = 6.7 Hz, 6H), 1.44 – 1.26 (m, 11H). ¹³C NMR (126 MHz, CDCl₃) δ 135.86, 126.49, 124.42, 123.74, 123.42 (C-F, $1J_{C-F} = 269.64$ Hz), 122.57, 121.26 (C-F, $1J_{C-F} = 269.64$ Hz), 120.05,

119.03, 112.08, 30.71, 30.26, 24.44, 23.77, 22.45. ¹⁹F NMR (471 MHz, CDCl₃) δ -57.38. HRMS (ESI/QTOF), m/z: [M-H] ⁻ Calcd. C₂₁H₂₇NF₃ 350.2096; Found 350.2105.

(1R,2S,5R)-2-isopropyl-5-methylcyclohexyl 2-(2-(trifluoromethyl)-1H-indol-3-yl)acetate



(3h): Physical state: White liquid; Yield: 65 mg (85 %). ¹H NMR (500 MHz, Chloroform-*d*) δ 8.43 (s, 1H), 7.66 (d, J = 8.1 Hz, 1H), 7.38 (dd, J = 8.3, 1.4 Hz, 1H), 7.32 (dd, J = 8.2, 7.0 Hz, 1H), 7.22 – 7.18 (m, 1H), 4.70 – 3.90 (m, 1H), 3.90 (s, 2H), 1.99 – 1.95 (m, 1H), 1.68 – 1.60 (m, 4H), 1.44 – 143 (m, 1H), 1.33 – 1.27 (m, 1H), 1.02 – 0.90 (m, 1H), 0.87 (d, J = 6.6 Hz, 3H), 0.84 – 0.81 (m, 1H), 0.78 (d,

J = 7.0 Hz, 3H), 0.64 (d, J = 7.0 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 170.29, 135.27, 127.49, 125.12, 122.93 (C-F, $2J_{C-F} = 35.3$ Hz), 122.65 (C-F, $2J_{C-F} = 35.3$ Hz), 121.06, 120.79 (C-F, $1J_{C-F} = 269.64$ Hz), 120.41, 118.65 (C-F, $1J_{C-F} = 269.64$ Hz), 111.82, 75.16, 47.11, 40.77, 34.33, 31.48, 30.33, 26.22, 23.50, 22.12, 20.75, 16.27. ¹⁹F NMR (471 MHz, CDCl₃) δ -58.49. HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₂₁H₂₅NO₂F₃ 380.1837; Found 380.1841.

1,3-dioxoisoindolin-2-yl 4-(2-(trifluoromethyl)-1Hindol-3-yl)butanoate (3i) : Physical state: White liquid; 10 $V = CF_3$ **Yield:** 46 mg (55 %). ¹**H NMR (500 MHz, CDCl₃)** δ 8.36 (s, 1H), 7.90 – 7.88 (m, 2H), 7.80 – 7.78 (m, 2H), 7.72 (d, J = 8.0 Hz, 1H), 7.41 (d, J = 8.3 Hz, 1H), 7.33 (t, J = 7.6 Hz, 1H), 7.21 (t, J = 7.5 Hz, 1H), 3.06 (t, J = 8.2 Hz, 2H), 2.73 (t, J = 7.5 Hz, 2H), 2.17 (p, J = 7.6 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 169.55, 162.11, 135.40, 135.01, 134.94, 134.90, 129.07, 127.41, 125.90 (C-F, $1J_{C-F}$ = 262.1 Hz), 125.09, 124.20, 124.16, 124.13, 123.83 (C-F, $1J_{C-F}$ = 262.1 Hz), 121.75 (C-F, $1J_{C-F}$ = 262.1 Hz), 120.95, 120.37, 119.65 (C-F, $1J_{C-F}$ = 262.1 Hz), 111.90, 30.59, 25.69, 22.94. ¹⁹F NMR (376 MHz, CDCl₃) δ -58.03. HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₂₁H₁₄N₂O₄F₃ 415.0906; Found 415.0899.

1,1,1,3,3,3-hexafluoropropan-2-yl 2-(2-(trifluoromethyl)-1H-indol-3-yl)acetate (3j):



Physical state: White liquid; Yield: 47 mg (60 %). ¹H NMR (500 MHz, CDCl₃) δ 8.53 (s, 1H), 7.64 (d, J = 8.1 Hz, 1H), 7.45 (d, J = 8.3 Hz, 1H), 7.40 (t, J = 7.6 Hz, 1H), 7.29 – 7.26 (m, 1H), 5.78 (hept, J = 6.0 Hz, 1H), 4.15 (d, J = 1.3 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 167.69, 135.23, 126.98, 125.56, 124.79 (C-F, $1J_{C-F}$ = 269.64 Hz),

123.91 (C-F, $2J_{C-F} = 37.3 \text{ Hz}$), 123.61 (C-F, $2J_{C-F} = 37.3 \text{ Hz}$), 123.32 (C-F, $2J_{C-F} = 37.3 \text{ Hz}$), 123.02 (C-F, $2J_{C-F} = 37.3 \text{ Hz}$), 122.66 (C-F, $1J_{C-F} = 269.64 \text{ Hz}$), 121.62, 120.52 (C-F, $1J_{C-F} = 269.64 \text{ Hz}$), 119.76, 112.11, 108.25 (C-F, $3J_{C-F} = 3.7 \text{ Hz}$), 108.23 (C-F, $3J_{C-F} = 3.7 \text{ Hz}$), 108.20 (C-F, $3J_{C-F} = 3.7 \text{ Hz}$), 108.18 (C-F, $3J_{C-F} = 3.7 \text{ Hz}$), 67.66 (C-F, q, J=34.9), 67.39 (C-F, q, J=34.9), 67.11 (C-F, q, J=34.9), 66.83 (C-F, q, J=34.9), 66.56 (C-F, q, J=34.9), 28.86. ¹⁹F **NMR (471 MHz, CDCl₃)** δ -58.82 (3F), -73.39 (6F). **HRMS (ESI/QTOF), m/z:** [M-H]⁻ Calcd. C₁₄H₇NO₂F₉ 392.0333; Found 392.0341.

5-(trifluoromethyl)-1H-imidazole-4-carbaldehyde (3k): Physical state: White solid; Yield: 30 mg (90 %). ¹H NMR (500 MHz, DMSO- d_6) δ 14.00 (s, 1H), 9.88 (s, 1H), 8.18 (s, 1H). ¹³C NMR (126 MHz, DMSO- d_6) δ 179.30, 140.65, 136.11 (C-F, 2 J_{C-F} = 39.06 Hz), 135.82 (C-F, 2 J_{C-F} = 39.06 Hz), 135.50 (C-F, 2 J_{C-F} = 39.06 Hz), 135.18 (C-F, 2 J_{C-F} = 39.06 Hz), 129.61, 124.56 (C-F, 1 J_{C-F} = 268.38 Hz), 122.43 (C-F, 1 J_{C-F} = 268.38 Hz), 120.30 (C-F, 1 J_{C-F} = 268.38 Hz), 118.17 (C-F, 1 J_{C-F} = 268.38 Hz). ¹⁹F NMR (376 MHz, CDCl₃) δ -55.88. HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₅H₂N₂OF₃ 163.0119; Found 163.0112.

2-phenyl-3-(trifluoromethyl)imidazo[1,2-α]pyridine (3l): Physical state: white solid; Yield: 46 mg (88 %). ¹H NMR (400 MHz, CDCl₃)

δ 8.31 (d, J = 7.0 Hz, 1H), 7.75 – 7.69 (m, 3H), 7.49 – 7.36 (m, 4H), 7.01 – 6.97 (m, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 148.07, 146.16, 132.90, 129.71 (C-F, $J_{C-F} = 1.5$ Hz), 129.69 (C-F, $J_{C-F} = 1.5$ Hz), 129.68 (C-F, $J_{C-F} = 1.5$ Hz), 129.66 (C-F, $J_{C-F} = 1.5$ Hz), 129.08, 128.48, 128.30, 128.29, 127.14, 125.66 (C-F, $J_{C-F} = 3.3$ Hz), 125.63 (C-F, $J_{C-F} = 3.3$ Hz), 125.59 (C-F, $J_{C-F} = 3.3$ Hz), 125.56 (C-F, $J_{C-F} = 3.3$ Hz), 118.14, 114.09. ¹⁹F NMR (376 MHz, CDCl₃) δ -57.56. HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₁₄H₈N₂F₃ 261.0640; Found 261.0645.

2-(trifluoromethyl)benzo[β]thiophene-3-carbaldehyde (3m): Physical state: White liquid;



Yield: 36 mg (79 %). ¹**H NMR (500 MHz, CDCl₃)** δ 10.43 (s, 1H), 8.11 (d, J = 8.3 Hz, 1H), 7.93 (d, J = 8.1 Hz, 1H), 7.60 – 7.53 (m, 2H). ¹³**C NMR (126 MHz, CDCl₃)** δ 183.83 (C-F, $J_{C-F} = 4.4$ Hz), 183.79 (C-F, J_{C-F}

= 4.4 Hz), 183.76 (C-F, J_{C-F} = 4.4 Hz), 183.72 (C-F, J_{C-F} = 4.4 Hz), 144.35, 141.43, 135.86, 128.78, 126.48, 125.84 (C-F, $I_{J_{C-F}}$ = 273.4 Hz), 125.24 (C-F, J_{C-F} = 2.4 Hz), 125.22 (C-F, J_{C-F} = 2.4 Hz), 125.20 (C-F, J_{C-F} = 2.4 Hz), 125.17 (C-F, J_{C-F} = 2.4 Hz), 123.67 (C-F, $I_{J_{C-F}}$ = 273.4 Hz), 123.30, 121.50 (C-F, $I_{J_{C-F}}$ = 273.4 Hz), 119.32 (C-F, $I_{J_{C-F}}$ = 273.4 Hz). ¹⁹F NMR (376 MHz, CDCl₃) δ -53.82. HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₁₀H₄OSF₃ 228.9935; Found 228.9941.

2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-4-(trifluoromethyl)-1H-indol-3-yl)acetic



acid (30): Physical state: Yellow Solid; Yield: 38 mg (45%). ¹H NMR (500 MHz, CDCl₃) δ 7.59 (d, J = 8.5 Hz, 2H), 7.41 (d, J = 8.5 Hz, 2H), 7.30 (d, J = 9.2 Hz, 1H), 6.77 (d, J = 9.2 Hz, 1H), 3.80 (s, 3H), 3.77 (q, J = 2.5 Hz, 2H), 2.20 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 177.24, 168.14, 154.91,

154.89, 140.21, 139.30, 133.29, 132.03, 131.58, 131.48, 129.40, 128.88, 127.87 (C-F, $I_{J_{C-F}} = 273.42 \text{ Hz}$), 127.71, 127.23 (C-F, $J_{C-F} = 2.52 \text{ Hz}$), 127.21 (C-F, $J_{C-F} = 2.52 \text{ Hz}$), 127.20 (C-F, $J_{C-F} = 2.52 \text{ Hz}$), 125.69 (C-F, $I_{J_{C-F}} = 273.42 \text{ Hz}$), 123.52 (C-F, $I_{J_{C-F}} = 273.42 \text{ Hz}$), 121.35 (C-F, $I_{J_{C-F}} = 273.42 \text{ Hz}$), 57.74, 29.71, 13.72. ¹⁹F NMR (376 MHz, CDCl₃) δ -51.59. HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₂₀H₁₄NO₄F₃Cl 424.0563; Found 424.0560.

ethyl (E)-2-benzylidene-4,4,4-trifluorobutanoate (3p): Physical state: White solid; Yield:



46 mg (90 %). ¹H NMR (500 MHz, CDCl₃) δ 8.23 (s, 1H), 7.51 (d, J = 5.8 Hz, 2H), 7.46 (d, J = 6.6 Hz, 3H), 4.59 (s, 2H), 4.37 (q, J = 7.2 Hz, 2H), 1.38 (t, J = 7.2 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 165.60, 148.75, 133.25, 130.32, 129.10 (J = 13.86), 128.99 (J = 13.86), 120.89, 118.28,

117.51, 62.21, 49.46. ¹⁹F NMR (**376** MHz, CDCl₃) δ -77.98.

3-(trifluoromethyl)-2H-chromen-2-one (3q): Physical state: White $figure CF_3$ solid; Yield: 32 mg (75 %). ¹H NMR (500 MHz, CDCl₃) δ 8.16 (s, 1H), 7.70 - 7.62 (m, 2H), 7.40 - 7.37 (m, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 155.92, 154.65, 143.46 (C-F, $3J_{C-F} = 4.9$ Hz), 143.42 (C-F, $3J_{C-F} = 4.9$ Hz), 143.38 (C-F, $3J_{C-F} = 4.9$ Hz), 143.34 (C-F, $3J_{C-F} = 4.9$ Hz), 143.48, 133.76, 129.53, 125.32, 124.62 (C-F, $1J_{C-F} = 273.42$ Hz), 122.46 (C-F, $1J_{C-F} = 273.42$ Hz), 121.32, 120.29 (C-F, $1J_{C-F} = 273.42$ Hz), 118.13 (C-F, $1J_{C-F} = 273.42$ Hz), 118.09 (C-F, $2J_{C-F} = 34.02$ Hz), 117.82 (C-F, $2J_{C-F} = 34.02$ Hz), 117.56 (C-F, $2J_{C-F} = 34.02$ Hz), 117.29 (C-F, $2J_{C-F} = 34.02$ Hz), 117.00, 116.80, 116.42. ¹⁹F NMR (376 MHz, CDCl₃) δ -66.08. HRMS (ESI/QTOF), m/z: [M+H]⁻ Calcd. C₁₀H₆O₂F₃ 215.0321; Found 215.0320.

1-benzyl-3-(trifluoromethyl)quinoxaline-2(1H)-one (3r): Physical state: White solid; **Vield:** 49 mg (80 %). ¹**H NMR (500 MHz, CDCl₃)** δ 7.96 (dd, J = 8.1, 1.5Hz, 1H), 7.58 – 7.55 (m, 1H), 7.38 – 7.22 (m, 7H), 5.50 (s, 2H). ¹³**C NMR (126 MHz, CDCl₃)** δ 151.90, 144.60 (C-F, 2 J_{C-F} = 34.02 Hz), 144.33 (C-F, 2 J_{C-F} = 34.02 Hz), 144.06 (C-F, 2 J_{C-F} = 34.02 Hz), 143.79 (C-F, 2 J_{C-F} =

34.02 Hz), 134.58, 134.14, 133.63, 132.02, 131.33, 129.23, 128.21, 127.19, 124.68, 123.36 (C-F, $1J_{C-F} = 277.2$ Hz), 121.16 (C-F, $1J_{C-F} = 277.2$ Hz), 118.96 (C-F, $1J_{C-F} = 277.2$ Hz), 116.77 (C-F, $1J_{C-F} = 277.2$ Hz), 114.95, 46.18. ¹⁹F NMR (376 MHz, CDCl₃) δ -69.80. HRMS (ESI/QTOF), m/z: [M+H]⁻ Calcd. C₁₆H₁₂N₂OF₃ 305.0902; Found 305.0904.

benzyl (2-(2-(nonafluoro-4l12-buta-1,3-diyn-1-yl)-1H-indol-3-yl)ethyl)carbamate (3s):



Physical state: White liquid; Yield: 68 mg (66 %). ¹H NMR (500NHCbzMHz, CDCl₃) δ 8.62 - 8.57 (m, 1H), 7.75 (d, J = 8.1 Hz, 1H), 7.42 -7.32 (m, 7H), 7.17 (t, J = 7.6 Hz, 1H), 5.13 (s, 2H), 4.93 (t, J = 6.2Hz, 1H), 3.53 (q, J = 6.8 Hz, 2H), 3.13 (t, J = 7.3 Hz, 2H). ¹³C NMR

(126 MHz, CDCl₃) δ 156.50, 136.55, 136.19, 128.53, 128.12 (J = 3.78), 128.09 (J = 3.78), 127.62, 125.17, 120.83, 120.34, 119.99 (J = 28.96), 119.76 (J = 28.96), 117.64, 111.81, 66.70, 41.81, 24.79. ¹⁹F NMR (376 MHz, CDCl₃) δ -80.80 – -80.85 (m, 3F), -107.52 (s, 2F), -122.66 (s, 2F), -125.67 – -125.75 (m, 2F). HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₂₂H₁₆N₂O₂F₉ 511.1068; Found 511.1059.

benzyl (2-(2-(tridecafluoro-6l16-hexa-1,3,5-triyn-1-yl)-1H-indol-3-yl)ethyl)carbamate NHCbz (3t): Physical state: White liquid; Yield: 73 mg (60 %). ¹H NMR (500 $_{13}$ **MHz, CDCl₃)** δ 8.48 (s, 1H), 7.75 (d, J = 8.1 Hz, 1H), 7.42 - 7.31 (m, 7H), 7.18 (t, J = 7.6 Hz, 1H), 5.12 (s, 2H), 4.90 (s, 1H), 3.52 (q, J = 6.8 Hz, 2H), 3.13 (t, J = 7.3 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 156.46, 136.56, 136.14, 128.52, 128.11 (J = 2.52), 128.09 (J = 2.52), 127.63, 125.21, 120.87, 120.37, 120.04 (J = 28.98), 119.81(J = 28.98), 117.71, 111.76, 66.69, 41.80, 24.79. ¹⁹F NMR (376 MHz, CDCl₃) δ -80.66 (t, J = 9.8 Hz, 3F), -107.32 - -107.51 (m, 2F), -121.57 - -121.84 (m, 4F), -122.60 - - 122.67 (m, 2H), -125.94 - -126.03 (m, 2H). HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₂₄H₁₆N₂O₂F₁₃ 611.1004; Found 611.1016.

benzyl



(2-(2-(heptadecafluoro-8-octa-1,3,5,7-tetrayn-1-yl)-1H-indol-3yl)ethyl)carbamate (3u): Physical state: White liquid; Yield: 75 mg (53 %). ¹H NMR (500 MHz, CDCl₃) δ 8.43 (s, 1H), 7.75 (d, J = 8.2Hz, 1H), 7.42 - 7.32 (m, 7H), 7.19 (d, J = 7.7 Hz, 1H), 5.12 (s, 2H), 4.88

(d, J = 6.4 Hz, 1H), 3.51 (t, J = 6.9 Hz, 2H), 3.12 (d, J = 7.5 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 155.41, 135.53, 135.09, 127.49, 127.08 (J = 3.78), 127.05 (J = 3.78), 126.60, 124.19, 119.85, 119.35, 119.02 (J = 28.98), 118.79 (J = 28.98), 116.70, 110.71, 65.65, 40.77, 23.76. ¹⁹F NMR (376 MHz, CDCl₃) δ -80.68 (t, J = 10.0 Hz, 3F), -107.37 (t, J = 14.2 Hz, 2F), -121.74 (s, 7F), -122.59(s, 3F), -125.95 - -126.04 (m, 2F). HRMS (ESI/QTOF), m/z: [M-H]⁻ Calcd. C₂₆H₁₆N₂O₂F₁₇711.0940; Found 711.0958.

GC- Analysis Data for 3n



Compound	Retention Time	Yield (Starting material) %	Yield (Conversion)
	7.564	31 (Unreacted); [M] = 120.1	
CF ₃	8.745		57.5; [M] = 188.1
3n	7.719		12; [M] = 256.1
F ₃ C CF ₃ 3n'			







5: Mechanistic Investigation

5.1: Radical trapping experiments

Effect of TEMPO









Figure 1: GC-MS Spectra for 5

5.2. UV- Visible Studies

UV-Visible spectra were taken on a Agilent Cary 60 UV-Vis Spectrophotometer of catalyst (Eosin -Y) solutions as well solutions of **1a**, oxidant and **2a** (1* 10⁻³ M). To investigate the possibility of a donor-acceptor complex between **1a**, **2a**, oxidant and Eosin Y along with other combination were studied and respective spectra are shown in Figure 2.



Figure 2: UV-Visible Spectra

Conclusion

The aforementioned studies clearly show that the addition of the 1a, 2a, oxidant doesn't affect the stability of Eosin Y - as well as no evidence of EDA is concluding from above all spectral graphs, indicating that no possibility of EDA complex in the reaction medium.

5.3. Fluorescence titration of photocatalyst

Fluorescence Quenching Experiments Test conditions for quenching reaction (I_0 and I are respective fluorescence intensities in the absence and presence of the indicated concentrations of the quenchers).

Eosin Y: 1.3 mg dissolved in 25 mL ACN (0.00008 M)

Quencher:

9.12 mg of (NH₄)₂S₂O₈ dissolved in 10 mL ACN (0.004 M)

15.6 mg of CF₃SO₂Na dissolved in 10 mL ACN (0.004 M)

11.8 mg 1a dissolved in 10 mL ACN (0.004 M)

General procedure: 1 mL of prepared solution containing Eosin Y was added to a cuvette, keep the total volume at 3 mL, Quenchers and ACN were adjusted according to the table shown in Florescence Graphs.

Entry	Eosin Y	Quenchers	ACN	Total Volume
1	1 mL	0 mL	2 mL	3 mL
2	1 mL	20 µL	1.98 mL	3 mL
3	1 mL	25 μL	1.75 mL	3 mL
4	1 mL	30 µL	1.70 mL	3 mL
5	1 mL	40 µL	1.60 mL	3 mL
6	1 mL	45 μL	1.55 mL	3 mL

Table 1: Preparation of Solution



Figure 3: Fluorescence quenching experiments with $(NH_4)_2S_2O_8$



Figure 4: Stern-Volmer plots of (NH₄)₂S₂O₈



Figure 5: Fluorescence quenching experiments with CF₃SO₂Na



Figure 6: Stern-Volmer plots of CF₃SO₂Na



Figure 7: Fluorescence quenching experiments with 1a



Figure 8: Stern-Volmer plots of 1a

Conclusion

The aforementioned studies clearly show that the strongest quenching of Eosin Y occurs in the presence of oxidant as compared to substrate **1a** i.e., $(NH_{4)2}S_2O_8$, No quenching was observed with CF_3SO_2Na indicating that quenching of the catalytic cycle is with $(NH_{4)2}S_2O_8$ favouring Oxidative quenching cycle initiated by single electron reduction of Eosin Y on irradiation with green light.

5.3: Electron Paramagnetic Resonance (EPR) Analysis

Continuous wave (CW) EPR spectra were obtained using a Bruker A300-9.5/12/S/W instrument with X-band of 8.75-9.65 GHz. The spectral data was collected at 77 K with the following spectrometer settings: microwave power = 0.48 mW, center field = 3350 G, sweep width = 100 G, sweep time = 30 s, modulation frequency = 9.6 GHz, modulation amplitude = 10 G, time constant = 0.01 ms.

For all the EPR measurements, the corresponding sample solution was transferred into the EPR tube and then placed in liquid Nitrogen to freeze the sample solution prior to recording of the spectra. After that, the sample tube was inserted in the EPR cavity which was kept frozen with continuous supply of liquid nitrogen for the recording of the spectra. For experiments in which the sample was irradiated, the sample tube was kept at 6 cm distance from the 535 nm Kessil lamp.



Scheme 4: General procedure for EPR study

Procedure: (Standard Condition) An oven dried 7 mL glass vial equipped with magnetic stirring bar was charged with 0.2 mmol of **1** (1.0 equiv.), trifluoro alkyl sulfinates (0.6 mmol, 3 equiv.) were taken in a long neck round bottom flask and 2.5 mol % of Eosin Y was added into it followed by ammonium peroxodisulfate (3.0 equiv. A screw cap equipped with a septum was then fitted tightly to the reaction vial. Next, the vial was placed into a magnetic stirrer and standard sample without irradiation with green light was recorded immediately by transferring into EPR tube and EPR spectra was recorded after freezing the solution by using liquid nitrogen (Figure 9).



Figure 9: EPR analysis of standard solution at 77 K

6: References

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7: NMR Spectral Data

















110 100 90 f1 (ppm)



























48 -49 -50 -51 -52 -53 -54 -55 -56 -57 -58 -59 -60 -61 -62 -63 -64 -65 -66 -67 -68 -69 -70 -71 -72 -73 -74 -75 -76 -77 -78 -79 -80 -81 -82 -83 -84 -85 -86 -87 -8 f1 (ppm)





-47 -48 -49 -50 -51 -52 -53 -54 -55 -56 -57 -58 -59 -60 -61 -62 -63 -64 -65 -66 -67 -68 -69 -70 -71 -72 -73 -74 -75 -76 -77 -78 -79 -80 -81 -82 -83 -84 -85 -86 f1 (ppm)











-48 -49 -50 -51 -52 -53 -54 -55 -56 -57 -58 -59 -60 -61 -62 -63 -64 -65 -66 -67 -68 -69 -70 -71 -72 -73 -74 -75 -76 -77 -78 -79 -80 -81 -8. f1 (ppm)

































