

**Exploring Metal/Base-Free Porphyrin Involving Carboxyl Functionalized Pyridine Moiety for Photocatalytic N-Arylation of Benzamide Validated by RSM**

Bhairav Chandraday Mataghare<sup>a</sup>, and Pundlik Rambhau Bhagat<sup>a\*</sup>

[a] Department of Chemistry, School of Advanced Sciences, Vellore Institute of Technology, Vellore-632014, India.

\*Corresponding author e-mail address: [drprbhagat111@gmail.com](mailto:drprbhagat111@gmail.com) (Pundlik Rambhau Bhagat)

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Signature SIF VIT VELLORE  
DCA-IL



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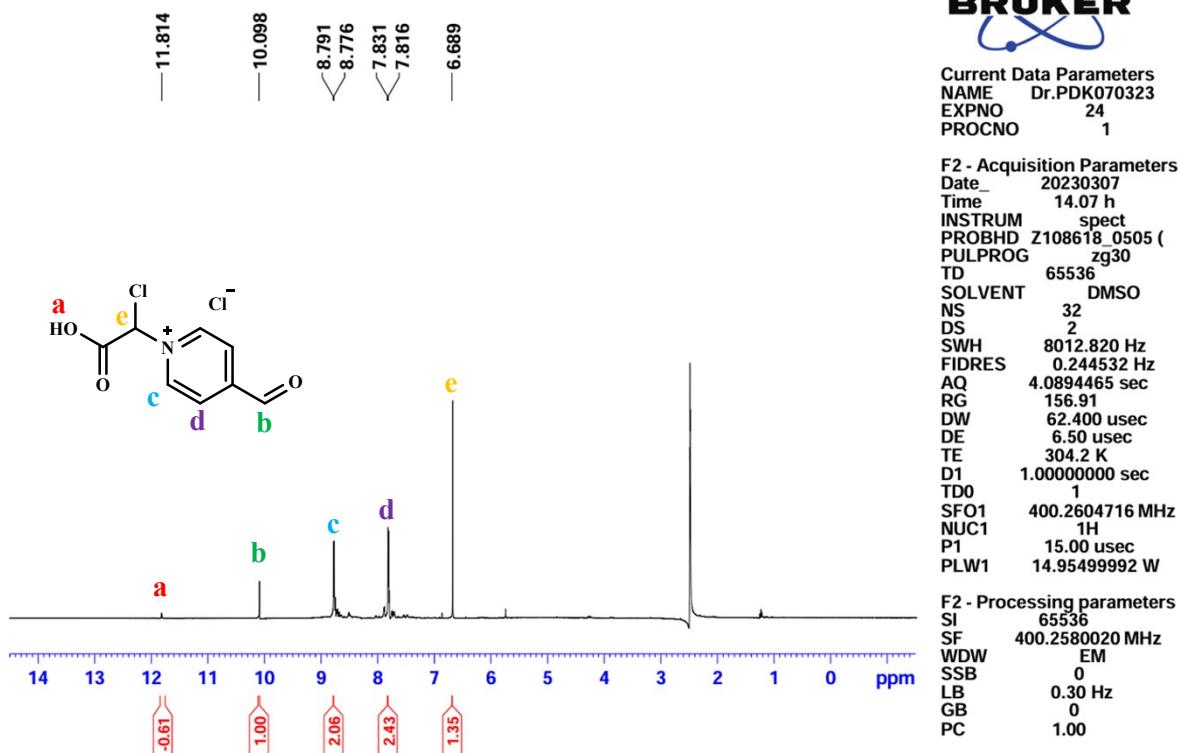


Fig S1.  $^1\text{H}$  NMR spectrum of 1-(carboxychloromethyl)-4-formyl-pyridin-1-ium chloride [1A]

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Current Data Parameters  
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PROCNO 1

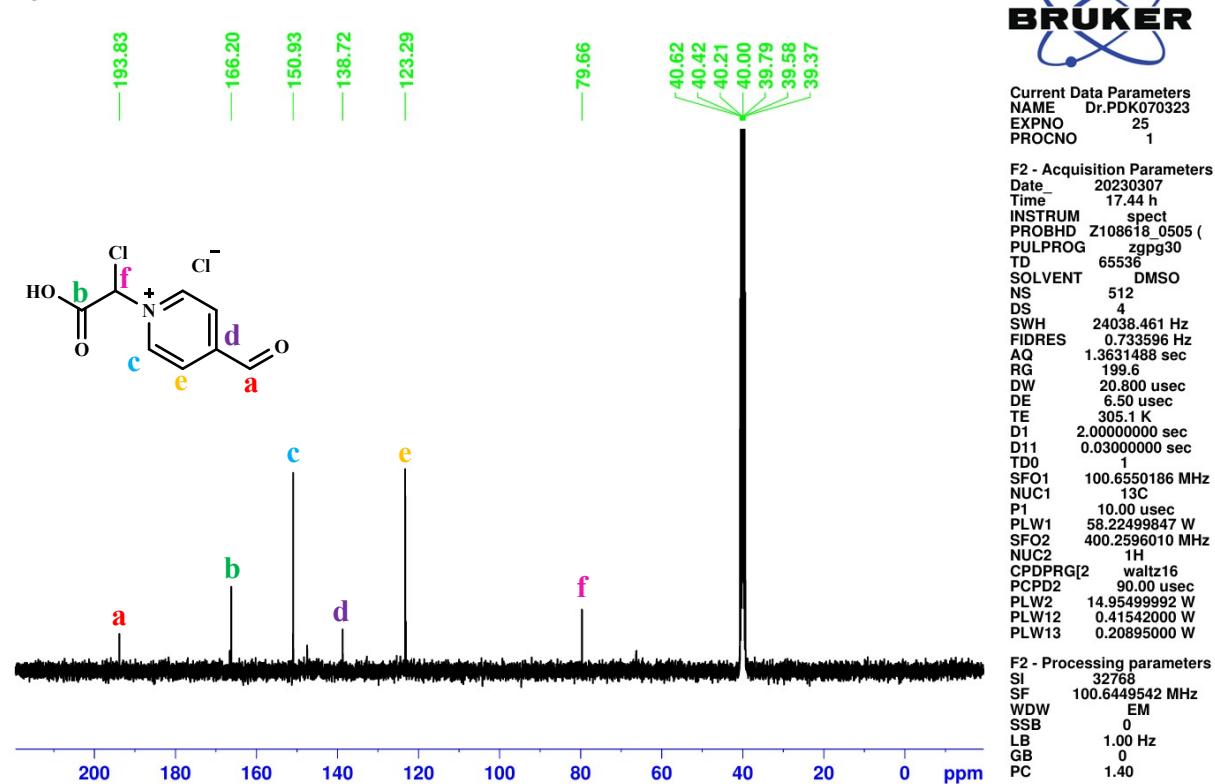


Fig S2.  $^{13}\text{C}$ NMR spectrum of 1-(carboxychloromethyl)-4-formyl-pyridin-1-ium chloride [1A]

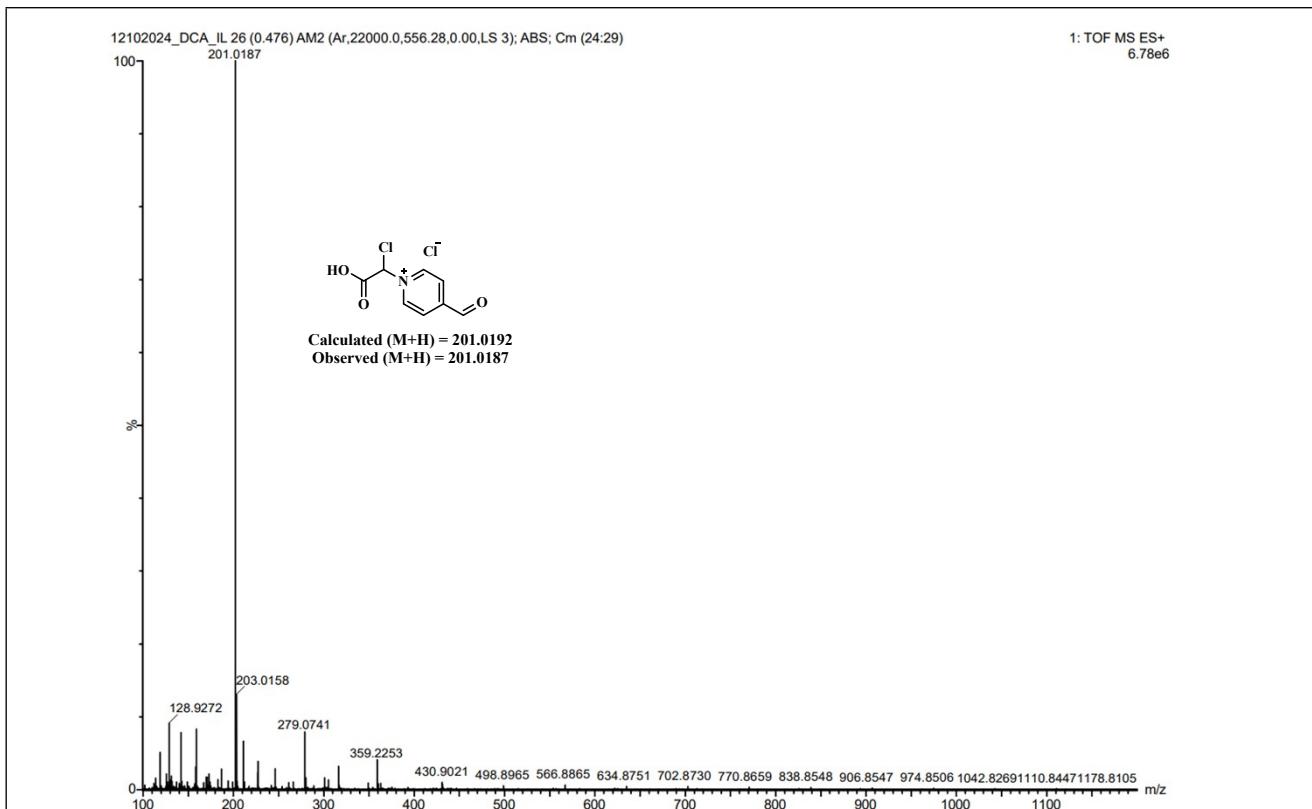


Fig S3. TOF MS ES +Ve mode of 1-(carboxychloromethyl)-4-formyl-pyridin-1-i um chloride [1A]

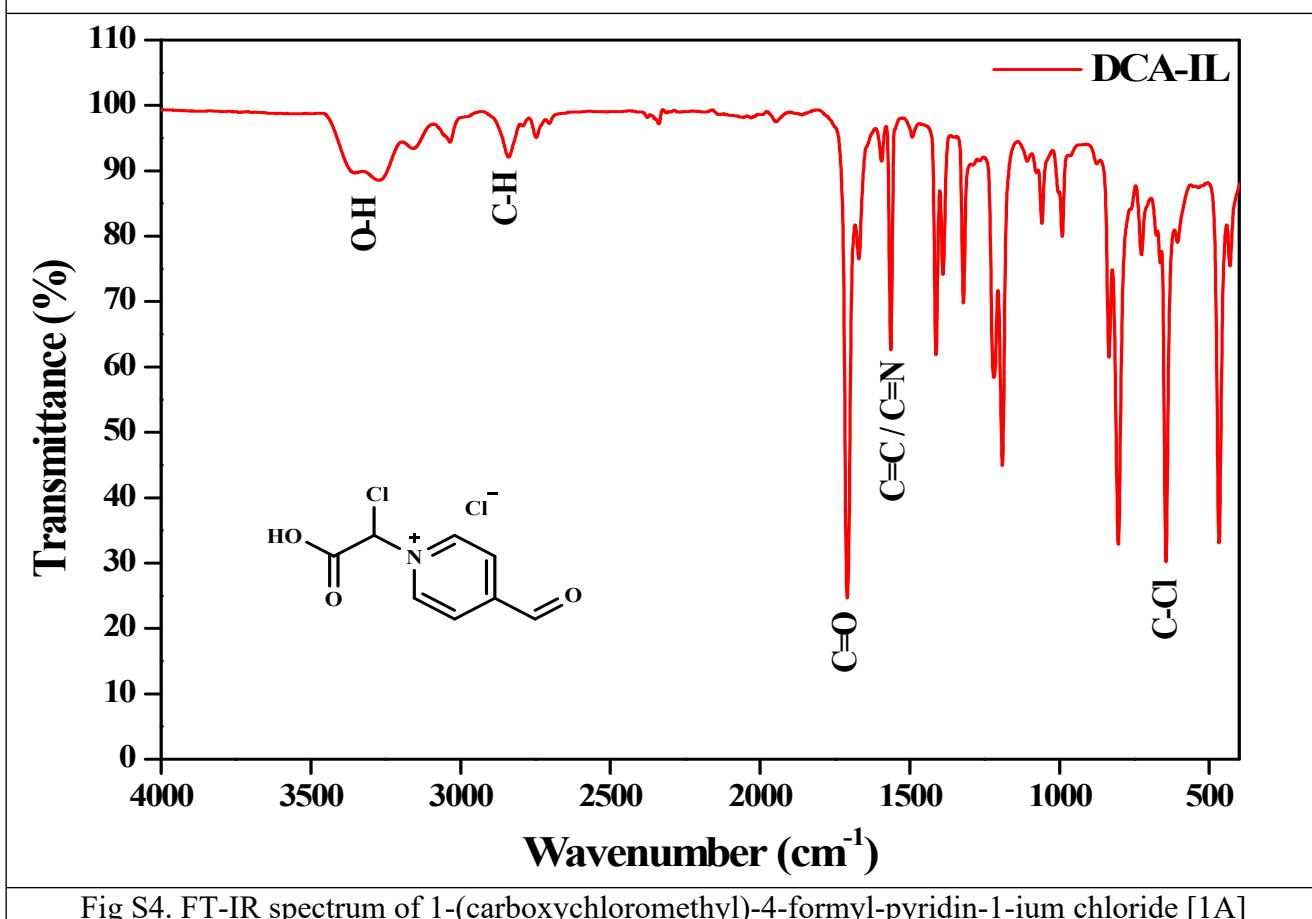


Fig S4. FT-IR spectrum of 1-(carboxychloromethyl)-4-formyl-pyridin-1-i um chloride [1A]

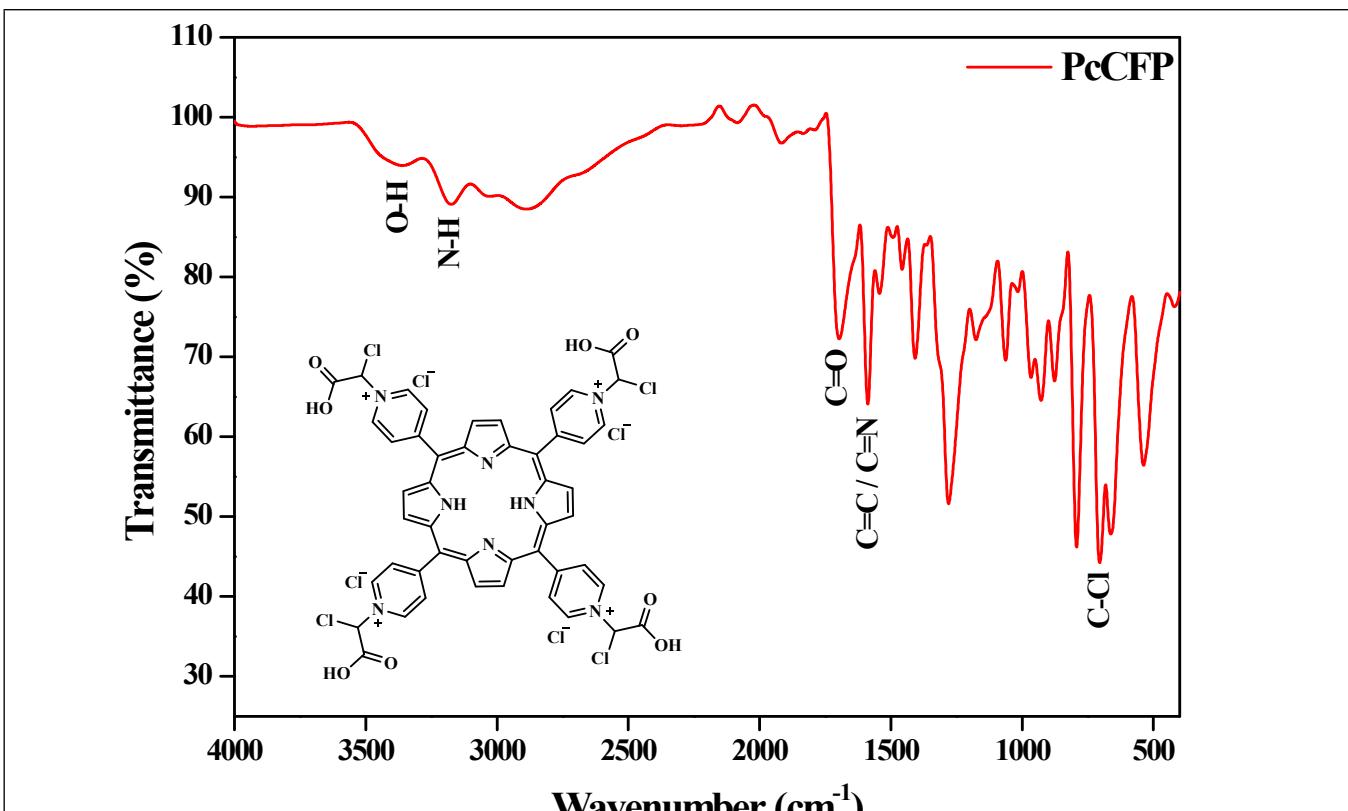


Fig S5. FT-IR Spectrum of PCCFP Photocatalyst

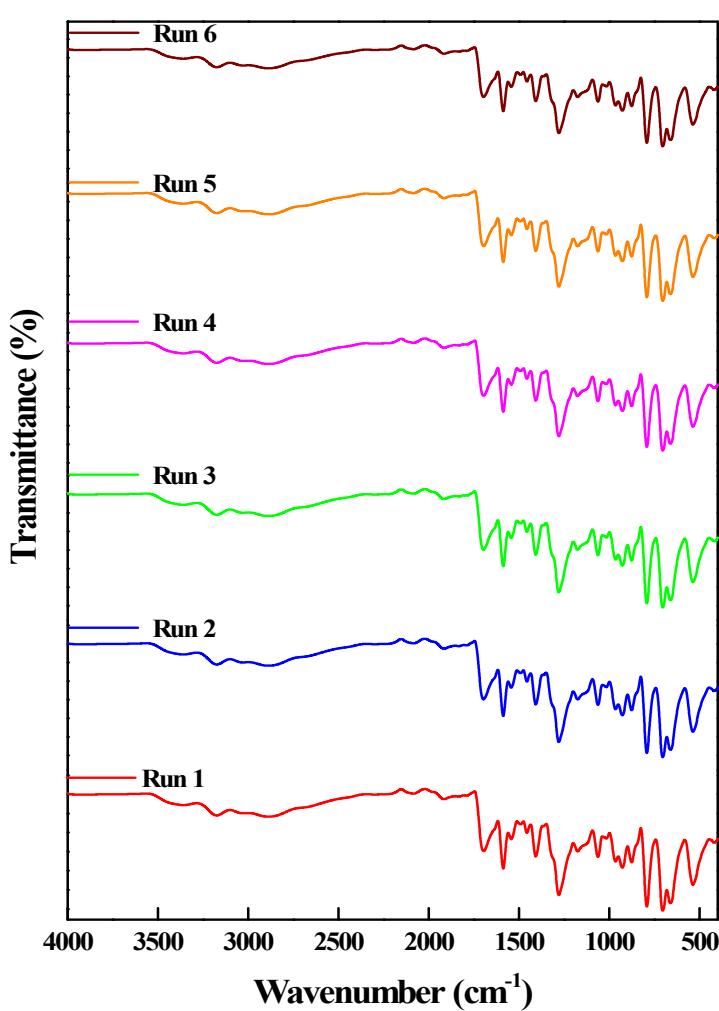


Fig S6. Comparative FT-IR Spectra of PCCFP Photocatalyst recycled upto 6<sup>th</sup> Run

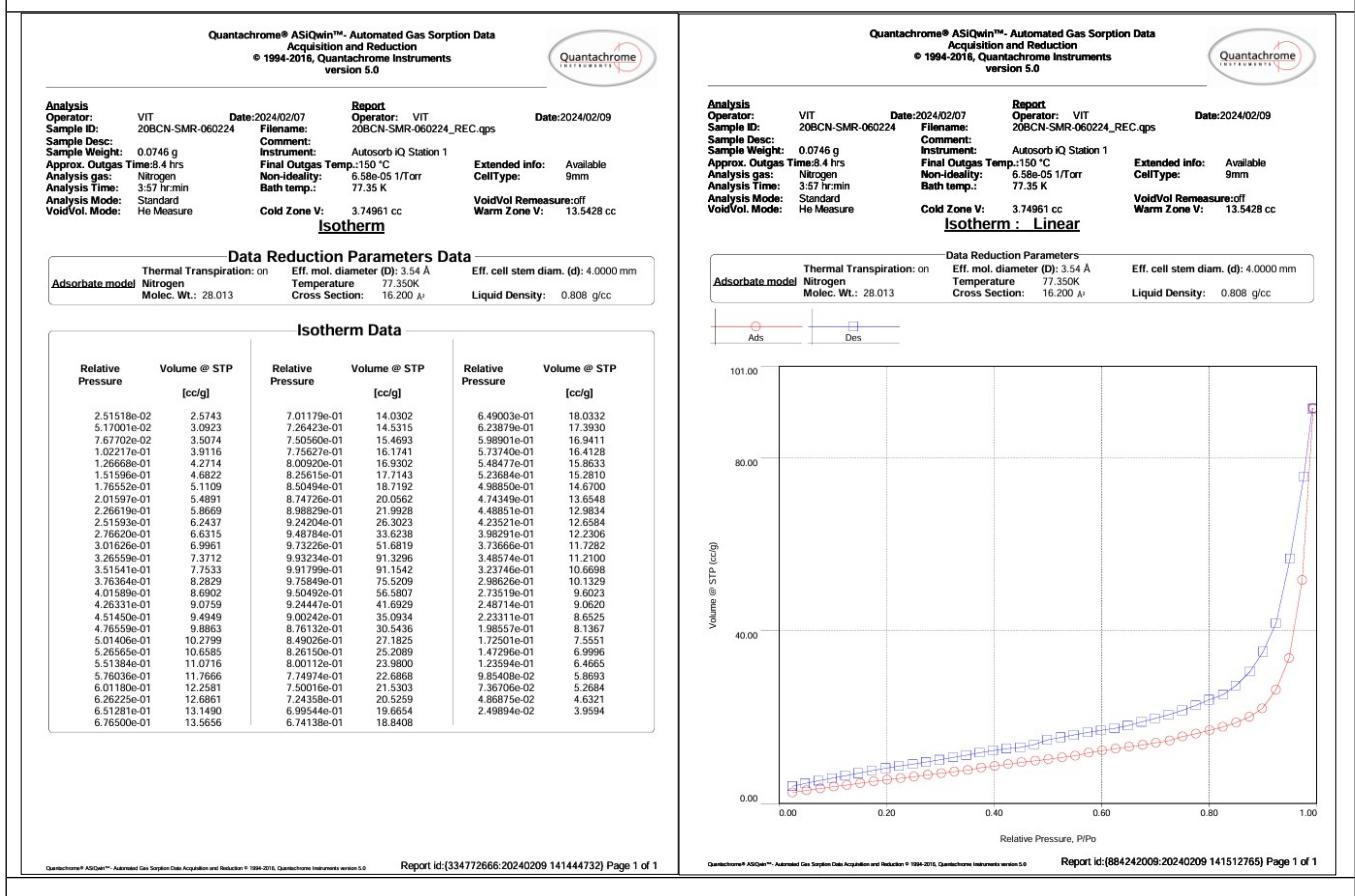
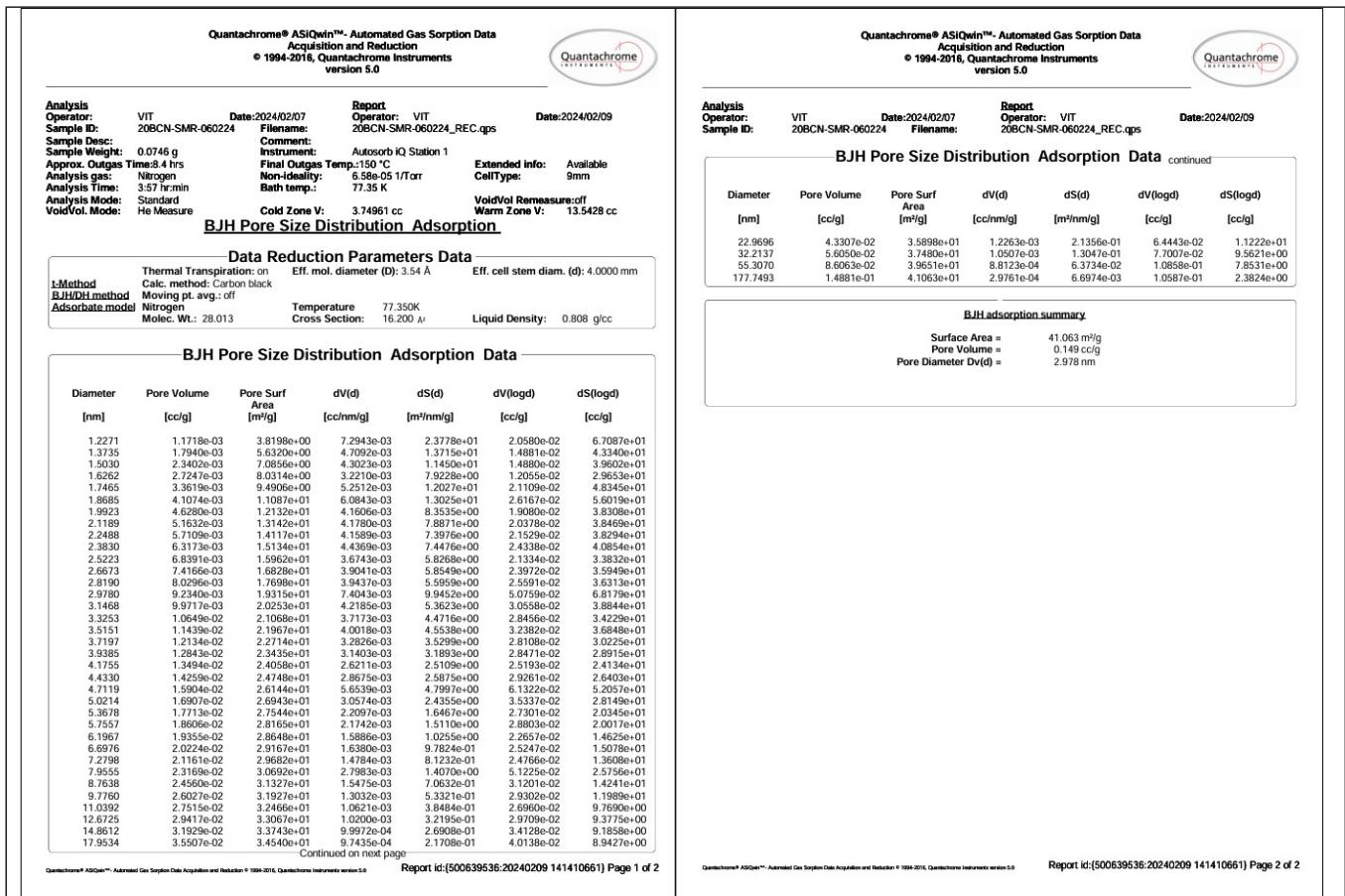
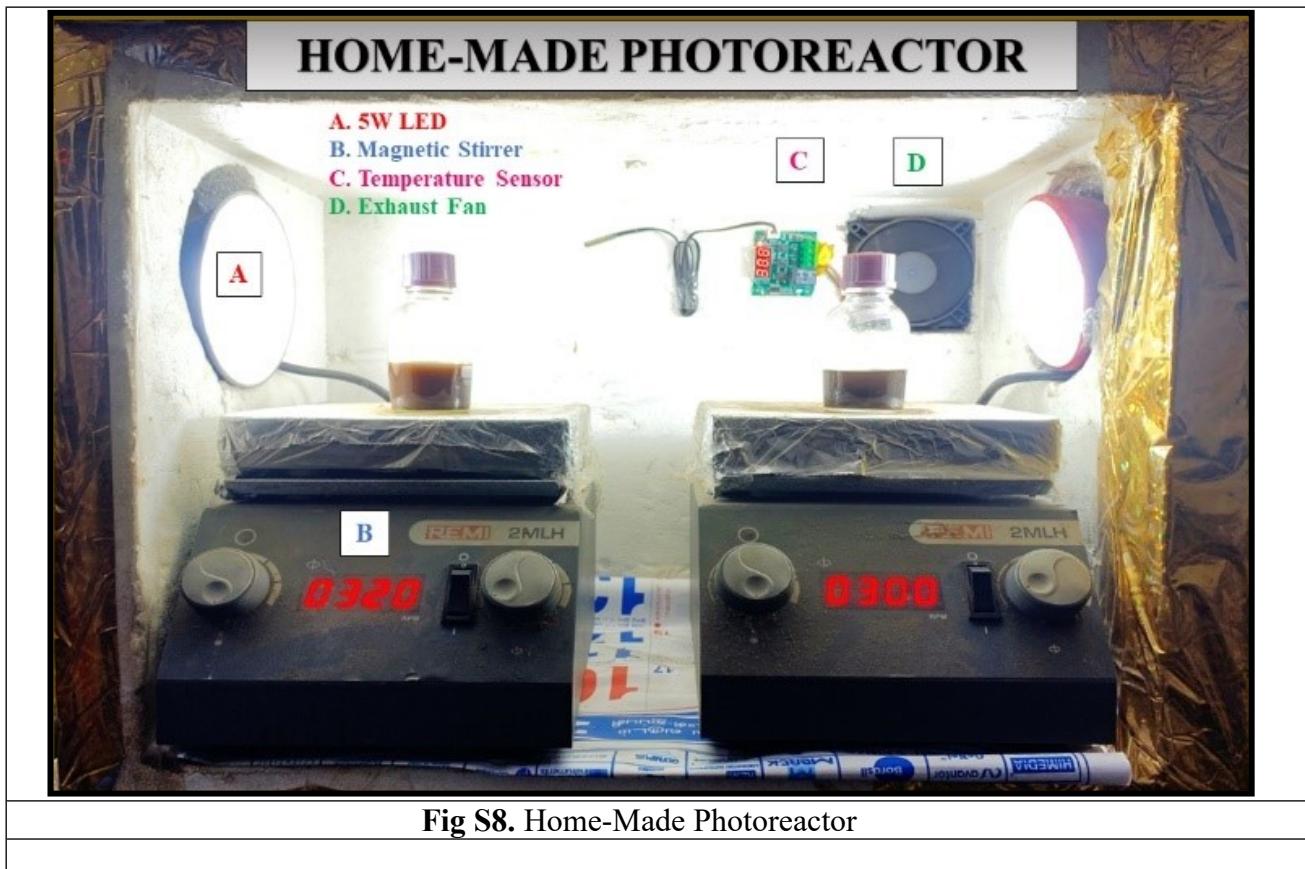


Fig S7. BET Data of PcfCFP Photocatalyst

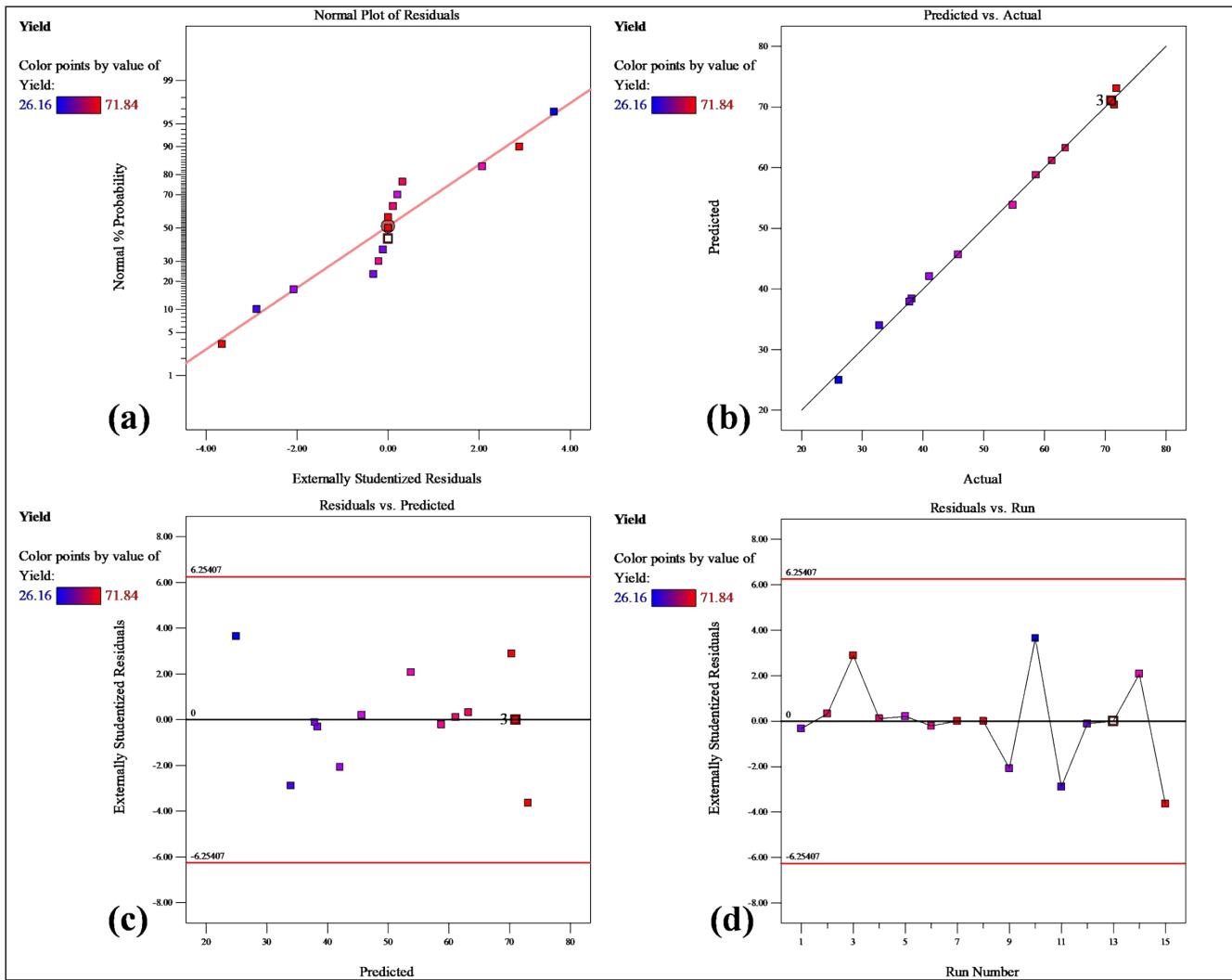


**Fig S8.** Home-Made Photoreactor

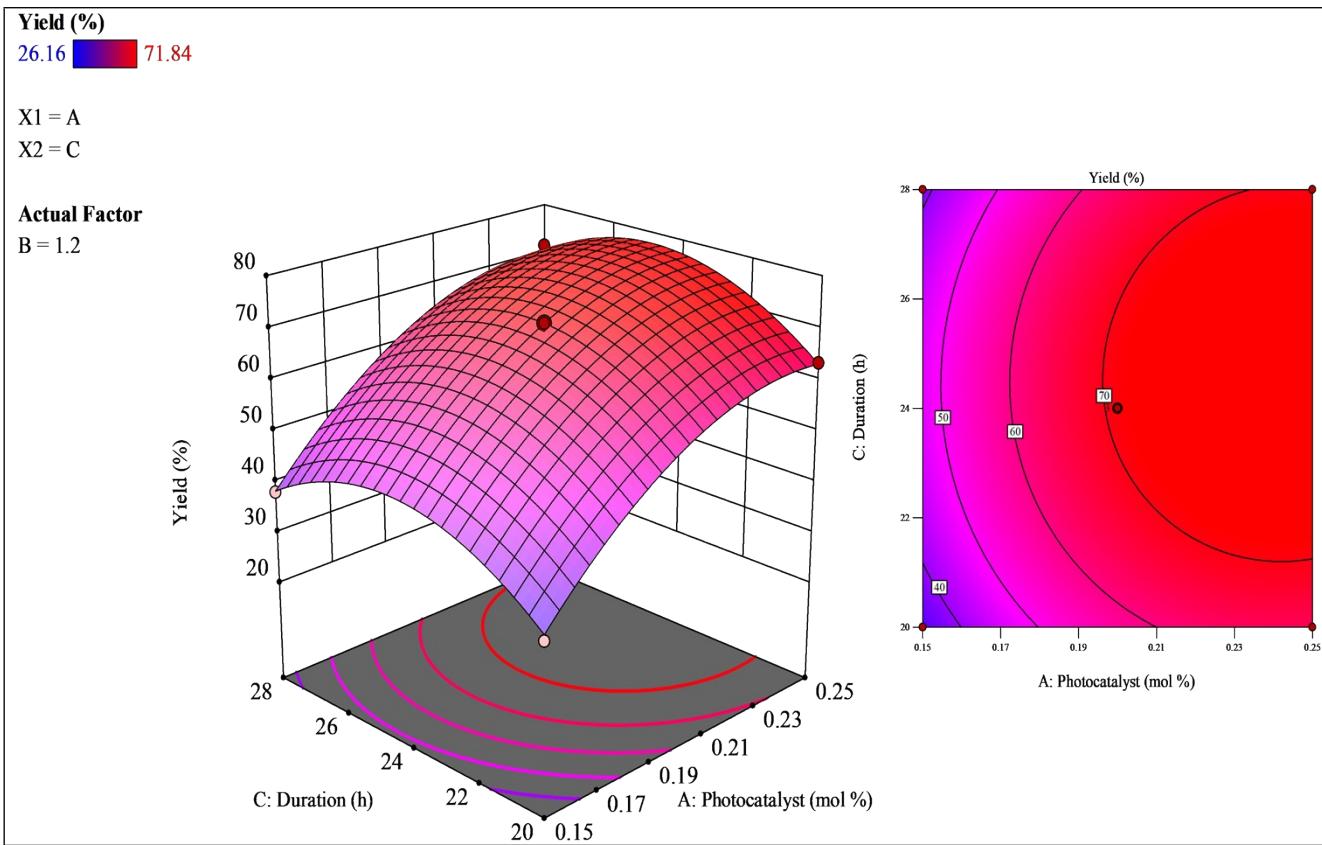
#### Explanation of Normal Distribution Plots:

The use of normal probability plots allows for a comparison between expected and residual responses, facilitating a visual assessment of the normality of these residuals, as depicted in Figure S9. Evaluating the model's adequacy necessitates a comparison between predicted values and experimental data. It is vital to ascertain whether the residuals exhibit a normal distribution to ensure an optimal distribution of data. This analysis plays a significant role in validating the reliability of the outputs generated by the predictive model. Additionally, the configuration of data points in relation to the fitted line can be employed to evaluate which model aligns most closely with the data. The plot of normal probability against externally studentized residuals (Fig S9. a) exhibited a linear association, though with some deviations, suggesting that data points conforming to the principles of normal distribution would yield superior model performance. Therefore, linearity in the data would be evident if the connecting line forms a 45° angle.

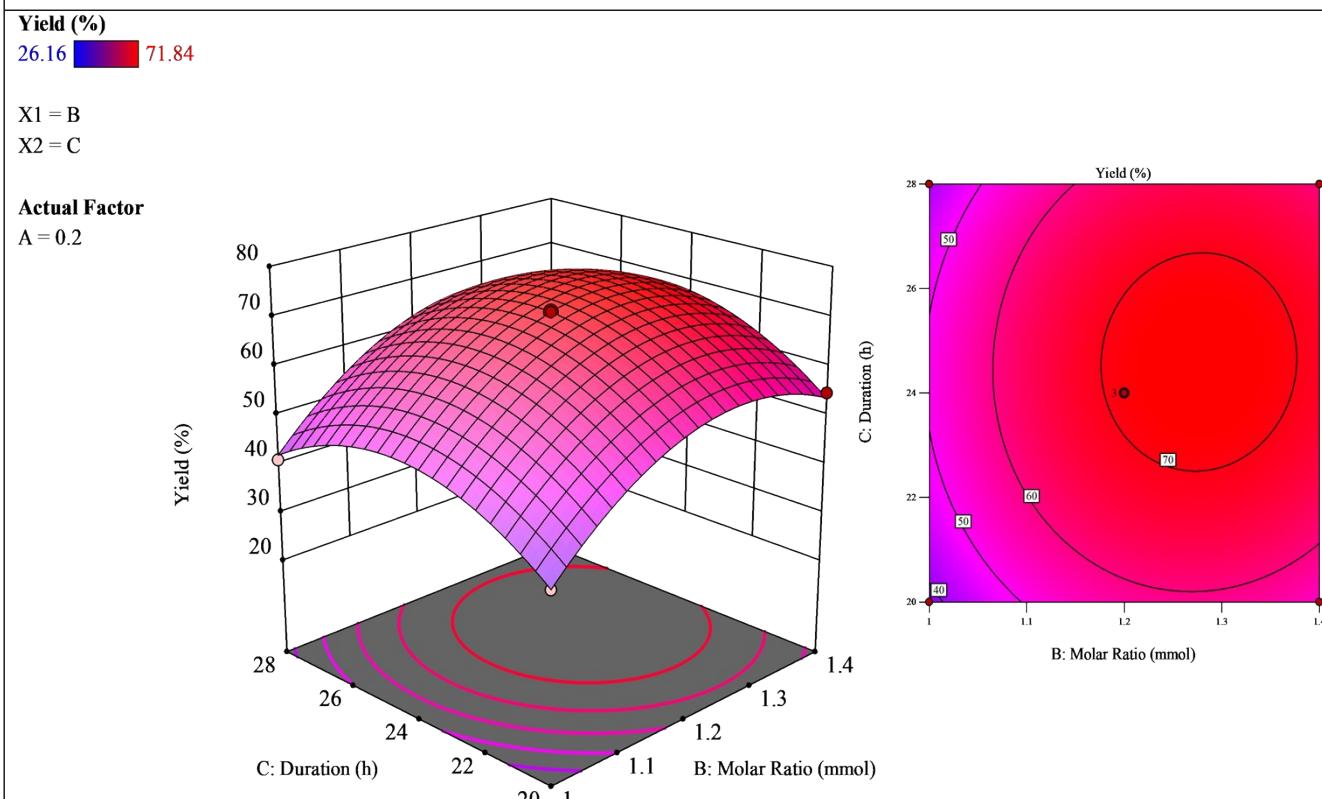
To determine the adequacy of the models used for the conversion of Benzamide to N-Aryl Amides through the photocatalytic process, we assessed both residuals and the distribution plot. The alignment of the plotted points with the theoretical normality line was analyzed to evaluate the model's suitability and significance (Fig S9. b). Likewise, Fig S9.c adeptly showcases the random characteristics of the residuals concerning the predicted response, highlighting the absence of any noticeable patterns or trends in the experimental data. This scatter plot confirms the lack of systematic variation, reinforcing the unpredictable nature of the residuals. Additionally, a plot of residuals plotted against the run number displays a random and scattered distribution (Fig S9. d). Such results are pivotal in affirming the adequacy and credibility of the model utilized.



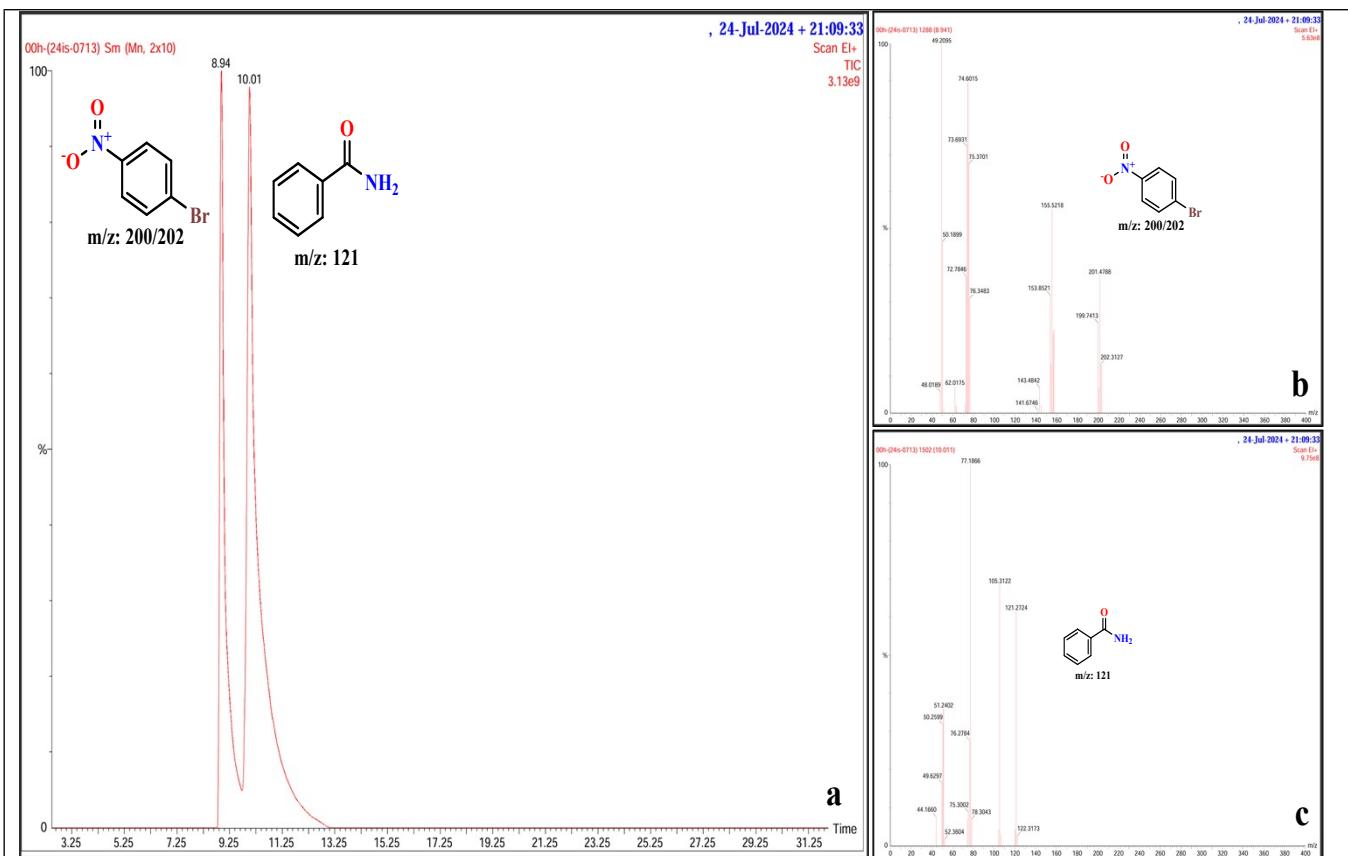
**Fig S9.** Plot illustrating the normal distribution of Linear models **a.** depicting the efficiency of PCCFP Photocatalyst for the N-Arylation of Benzamide. **b.** Comparison between the observed and predicted results for photocatalytic N-Arylation of Benzamide. Normal probability plots of residual for photo-transesterification of UFO to Biofuel as a function of **c.** Predicted Response and **d.** Run Number



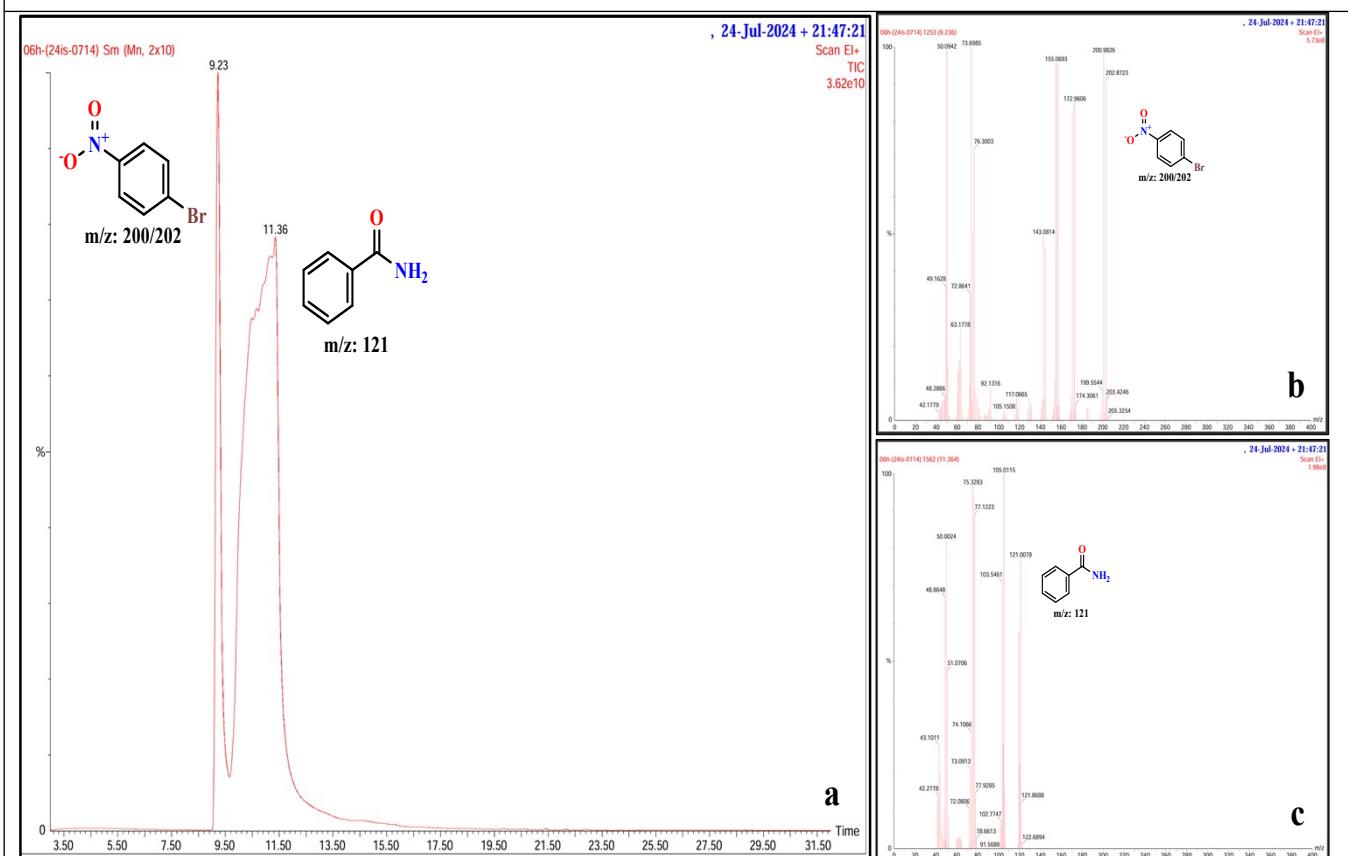
**Fig S10.** 3-D Surface and Contour Plot of N-arylation of Benzamide highlighting the relationship between the amount of PcCFP Photocatalyst (A) and duration of reaction (C) with constant molar ratio of reactants (B)



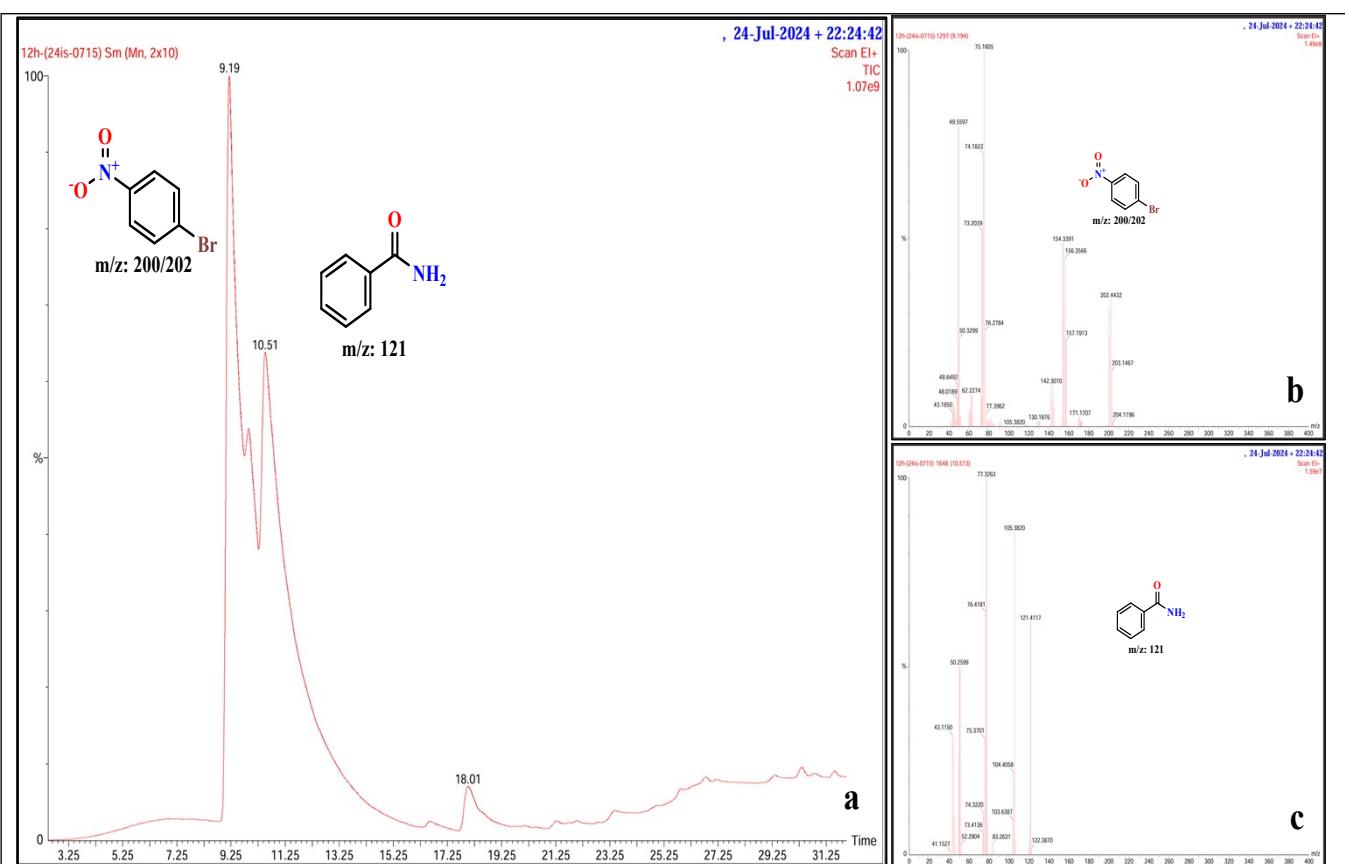
**Fig S11.** 3-D Surface and Contour Plot of N-arylation of Benzamide highlighting the relationship between the molar ratio of reactants (B) and duration of reaction (C) with constant amount of PcCFP Photocatalyst (A)



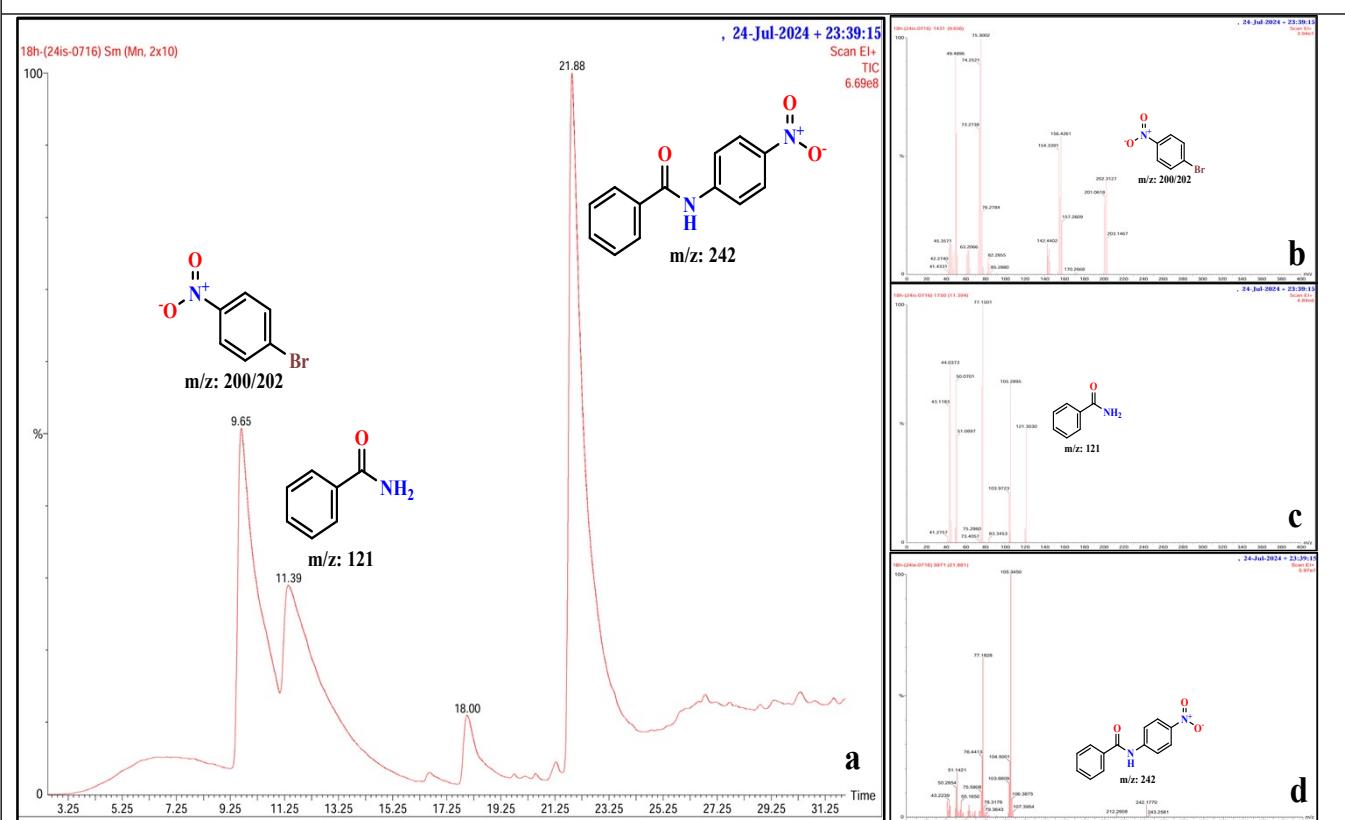
**Fig S12.** **a.** GC-MS Chromatogram of sample taken initially at 0 hour of reaction time. **b.** m/z of Ar-Br obtained at 8.94 RT in chromatogram. **c.** m/z of benzamide obtained at 10.01 RT in chromatogram.



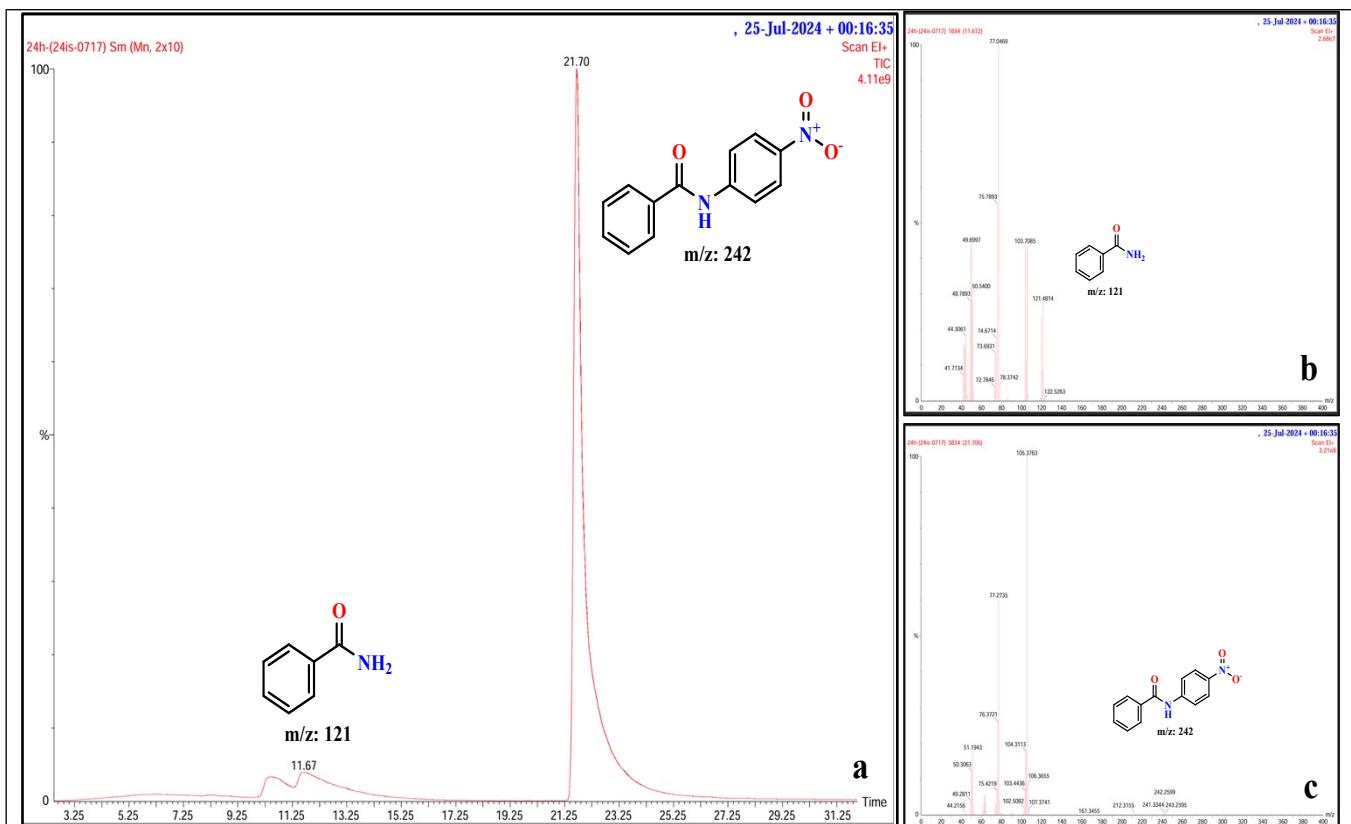
**Fig S13.** **a.** GC-MS Chromatogram of sample taken initially at 6 hour of reaction time. **b.** m/z of Ar-Br obtained at 9.23 RT in chromatogram. **c.** m/z of benzamide obtained at 11.36 RT in chromatogram.



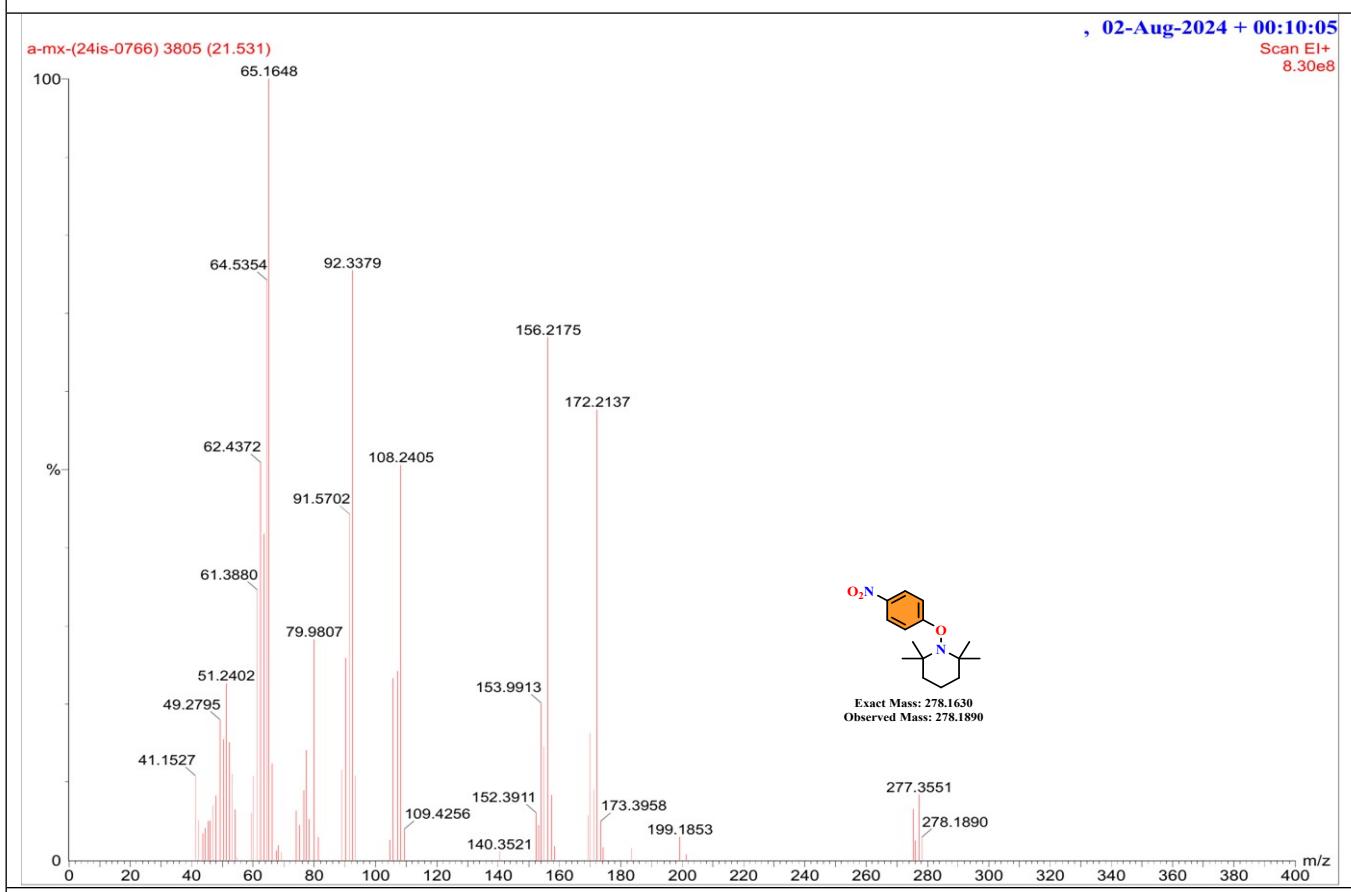
**Fig S14. a.** GC-MS Chromatogram of sample taken initially at 12 hour of reaction time. **b.** m/z of Ar-Br obtained at 9.19 RT in chromatogram. **c.** m/z of benzamide obtained at 10.51 RT in chromatogram.



**Fig S15. a.** GC-MS Chromatogram of sample taken initially at 18 hour of reaction time. **b.** m/z of Ar-Br obtained at 9.65 RT in chromatogram. **c.** m/z of benzamide obtained at 11.39 RT in chromatogram. **d.** m/z of product obtained at 21.88 RT in chromatogram.



**Fig S16.** a. GC-MS Chromatogram of sample taken initially at 24 hour of reaction time. b.  $m/z$  of benzamide traces obtained at 11.67 RT in chromatogram. c.  $m/z$  of product obtained at 21.70 RT in chromatogram.



**Fig S17.** GC-Mass Spectrum of TEMPO adduct formed from Aryl radical

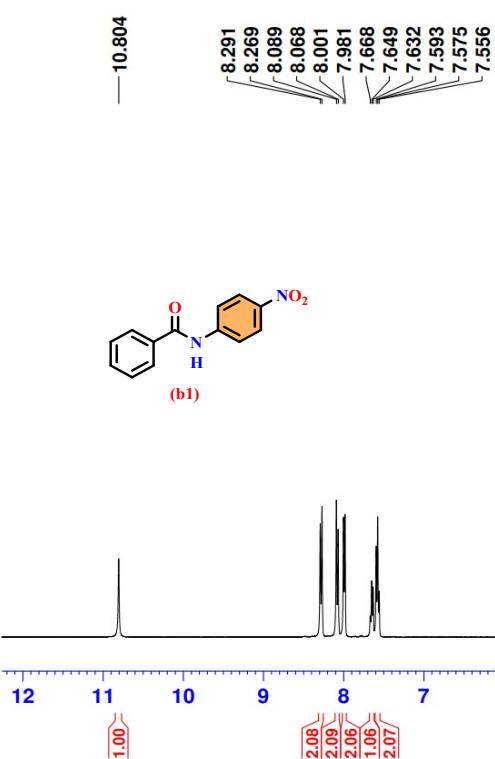
**Table S1.** Precursor with respective halogens and its Product obtained via N-arylation of Benzamide

Sr. No	Product Code	Reactant R1	Reactants R2	Product	Yield (%)
1.	b1 <sub>i</sub>			 <chem>CC(=O)Nc1ccccc1Brc2ccccc2[N+](=O)[O-]</chem>	71%
2.	b1 <sub>ii</sub>			 <chem>CC(=O)Nc1ccccc1Clc2ccccc2[N+](=O)[O-]</chem>	29%
3.	b2 <sub>i</sub>			 <chem>CC(=O)Nc1ccccc1Brc2ccccc2C(=O)Nc3ccccc3</chem>	64%
4.	b2 <sub>ii</sub>			 <chem>CC(=O)Nc1ccccc1Clc2ccccc2C(=O)Nc3ccccc3</chem>	27%
5.	b3 <sub>i</sub>			 <chem>CC(=O)Nc1ccccc1OCc2ccccc2[N+](=O)[O-]</chem>	61%
6.	b3 <sub>ii</sub>			 <chem>CC(=O)Nc1ccccc1OCc2ccccc2C(=O)Nc3ccccc3</chem>	22%
7.	b4 <sub>i</sub>			 <chem>CC(=O)Nc1ccccc1[N+](=O)[O-]c2ccccc2[N+](=O)[O-]</chem>	73%
8.	b4 <sub>ii</sub>			 <chem>CC(=O)Nc1ccccc1[N+](=O)[O-]c2ccccc2[N+](=O)[O-]</chem>	35%
9.	b5 <sub>i</sub>			 <chem>CC(=O)Nc1ccccc1Brc2ccccc2</chem>	63%
10.	b5 <sub>ii</sub>			 <chem>CC(=O)Nc1ccccc1Clc2ccccc2</chem>	30%
11.	b5 <sub>iii</sub>			 <chem>CC(=O)Nc1ccccc1Ic2ccccc2</chem>	72%
12.	b6 <sub>i</sub>			 <chem>CC(=O)Nc1ccccc1Brc2ccccc2</chem>	58%

13.	b6 <sub>ii</sub>				21%
14.	b7				54%
15.	b8				67%
16.	b9				60%
17.	b10				62%
18.	b11				61%
19.	b12				73%
20.	b13 <sub>i</sub>				59%
21.	b13 <sub>ii</sub>				22%
22.	b14 <sub>i</sub>				60%
23.	b14 <sub>ii</sub>				26%
24.	b15				68%
25.	b16				51%
26.	b17				60%

27.	b18				63%
28.	b19				53%
29.	b20				55%
30.	b21				61%
31.	b22				28%
32.	b23				69%
33.	b24 <sub>i</sub>				70%
34.	b24 <sub>ii</sub>				28%
35.	b25				31%

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Bz-01-Br



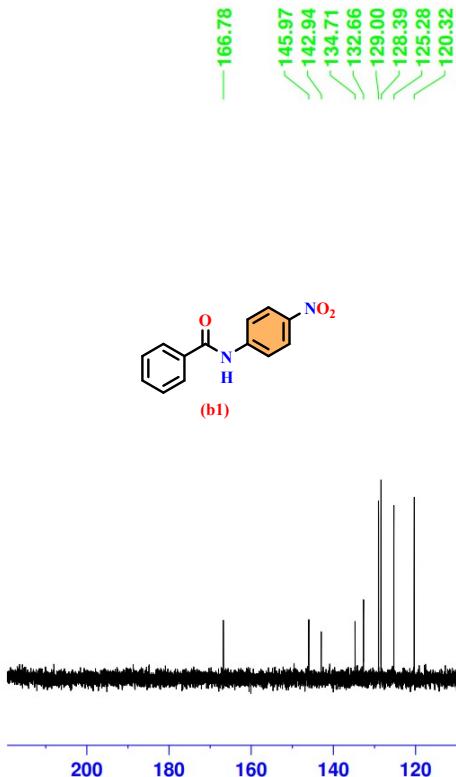
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PROCNO 1

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TD 65536  
SOLVENT DMSO  
NS 32  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 199.6  
DW 62.400 usec  
DE 6.50 usec  
TE 307.5 K  
D1 1.0000000 sec  
TD0 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

F2 - Processing parameters  
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SF 400.2580000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

Fig S18.  $^1\text{H}$  NMR spectrum of *N*-(4-nitrophenyl)benzamide (**b1**)

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Bz-01-Br



Current Data Parameters  
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PROCNO 1

F2 - Acquisition Parameters  
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DS 4  
SWH 24038.461 Hz  
FIDRES 0.733596 Hz  
AQ 1.3631488 sec  
RG 199.6  
DW 20.800 usec  
DE 6.50 usec  
TE 304.2 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TD0 1  
SFO1 100.6550186 MHz  
NUC1  $^{13}\text{C}$   
P1 10.00 usec  
PLW1 56.49300003 W  
SFO2 400.2596010 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 90.00 usec  
PLW2 15.21399975 W  
PLW12 0.42261001 W  
PLW13 0.21257000 W

F2 - Processing parameters  
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SF 100.6449542 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

Fig S19.  $^{13}\text{C}$  NMR spectrum of *N*-(4-nitrophenyl)benzamide (**b1**)

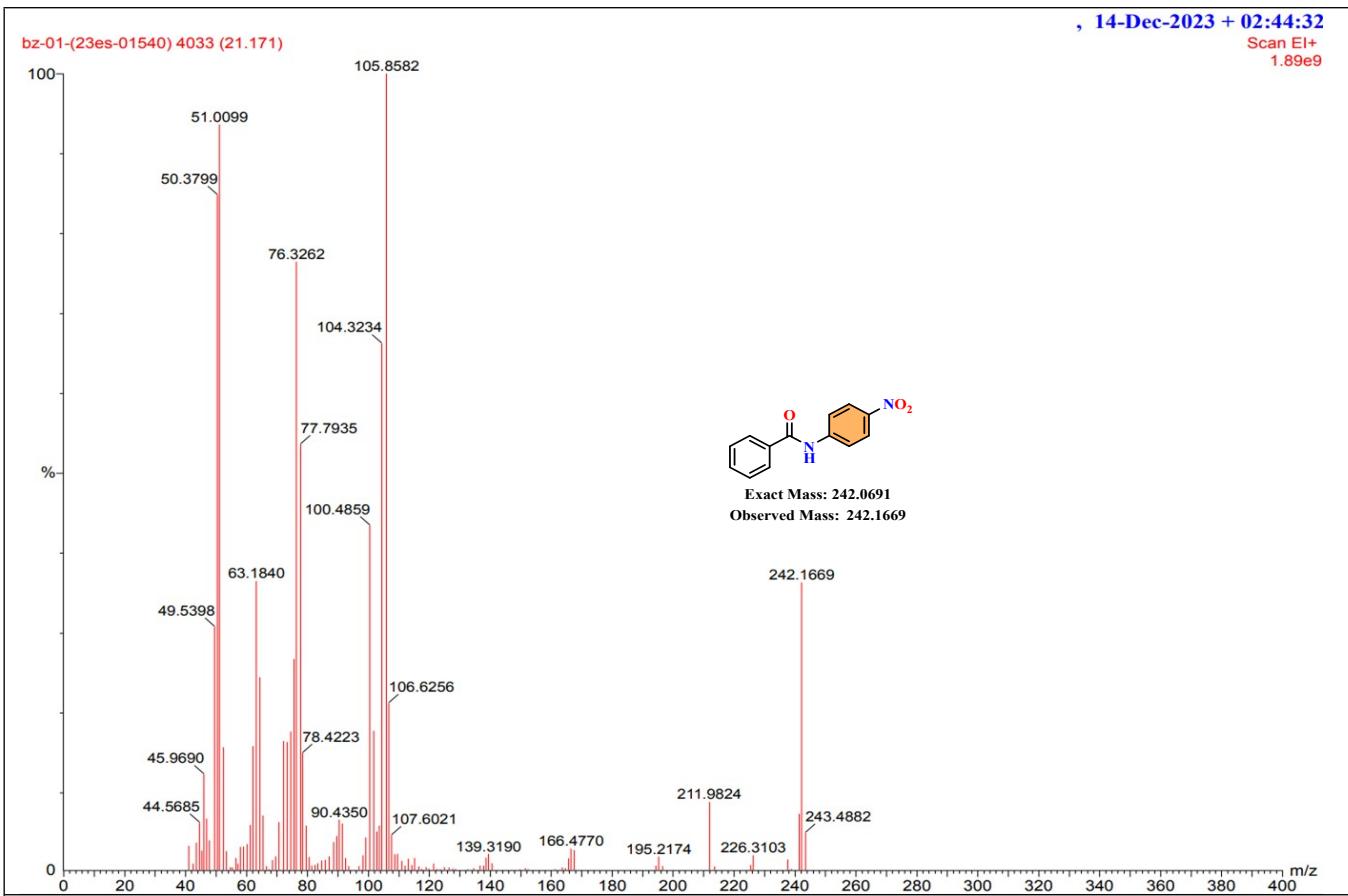
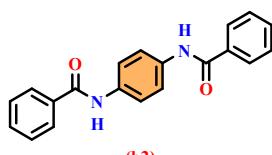


Fig S20. GC-Mass spectrum ( $m/z$ ) of *N*-(4-nitrophenyl)benzamide ( $b_{1j}$ )

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BZ-02-Br



10.159 ——————  
[ 7.906    7.887    7.686    7.540    7.522    7.505    7.481    7.464    7.445 ]



Current Data Parameters  
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EXPNO 24  
PROCNO 1

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DS 2  
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FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 127.79  
DW 62.400 usec  
DE 6.50 usec  
TE 308.5 K  
D1 1.0000000 sec  
TDO 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

F2 - Processing parameters  
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SF 400.2580303 MHz  
WDW EM  
SSB 0  
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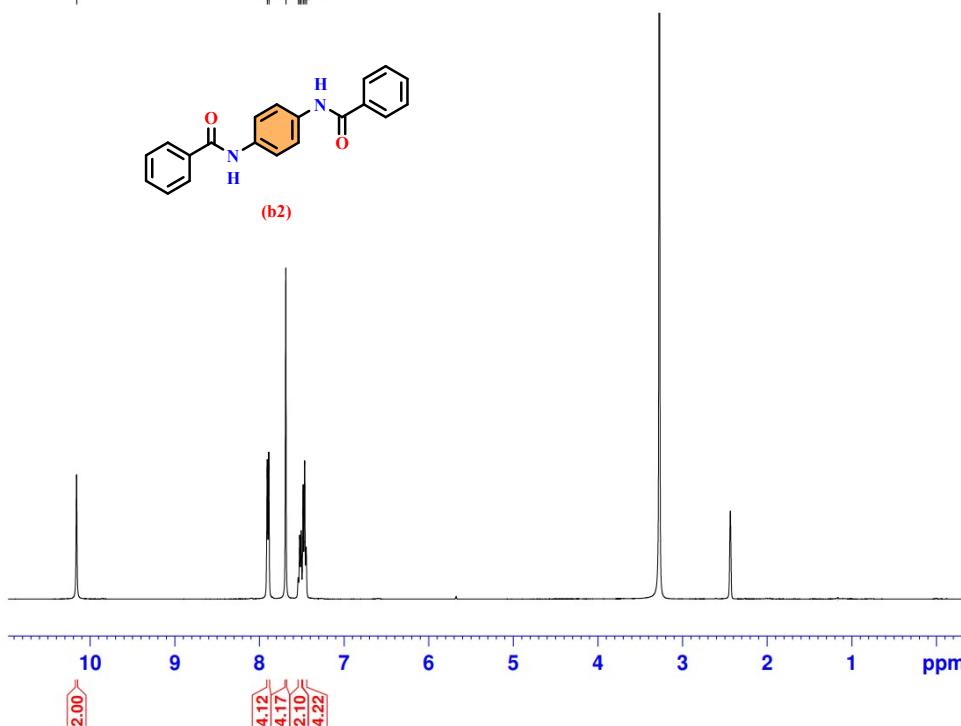


Fig S21.  $^1\text{H}$  NMR spectrum of *N,N'*-(1,4-phenylene)dibenzamide ( $b_{2j}$ )

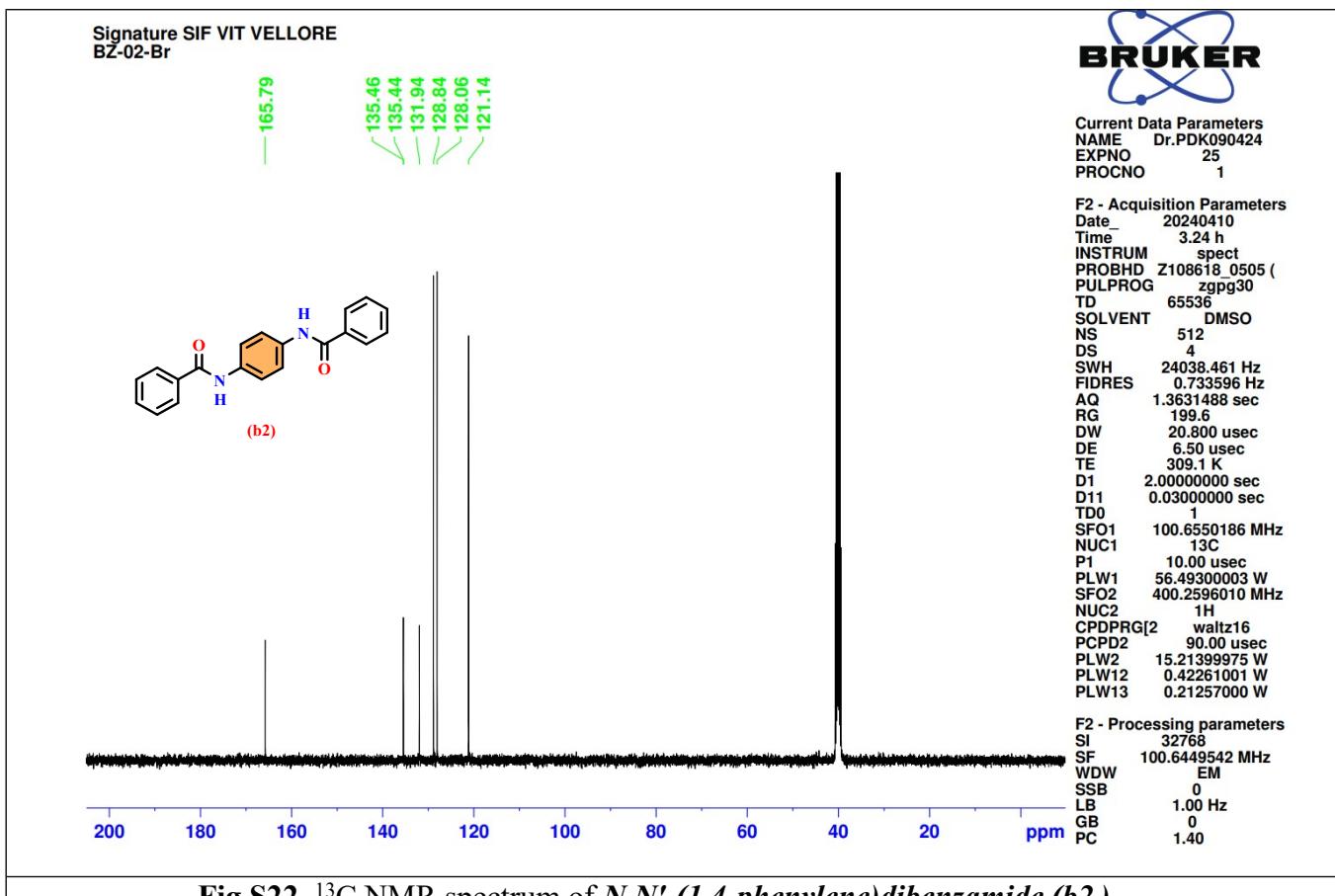


Fig S22. <sup>13</sup>C NMR spectrum of *N,N'*-(1,4-phenylene)dibenzamide (*b2<sub>i</sub>*)

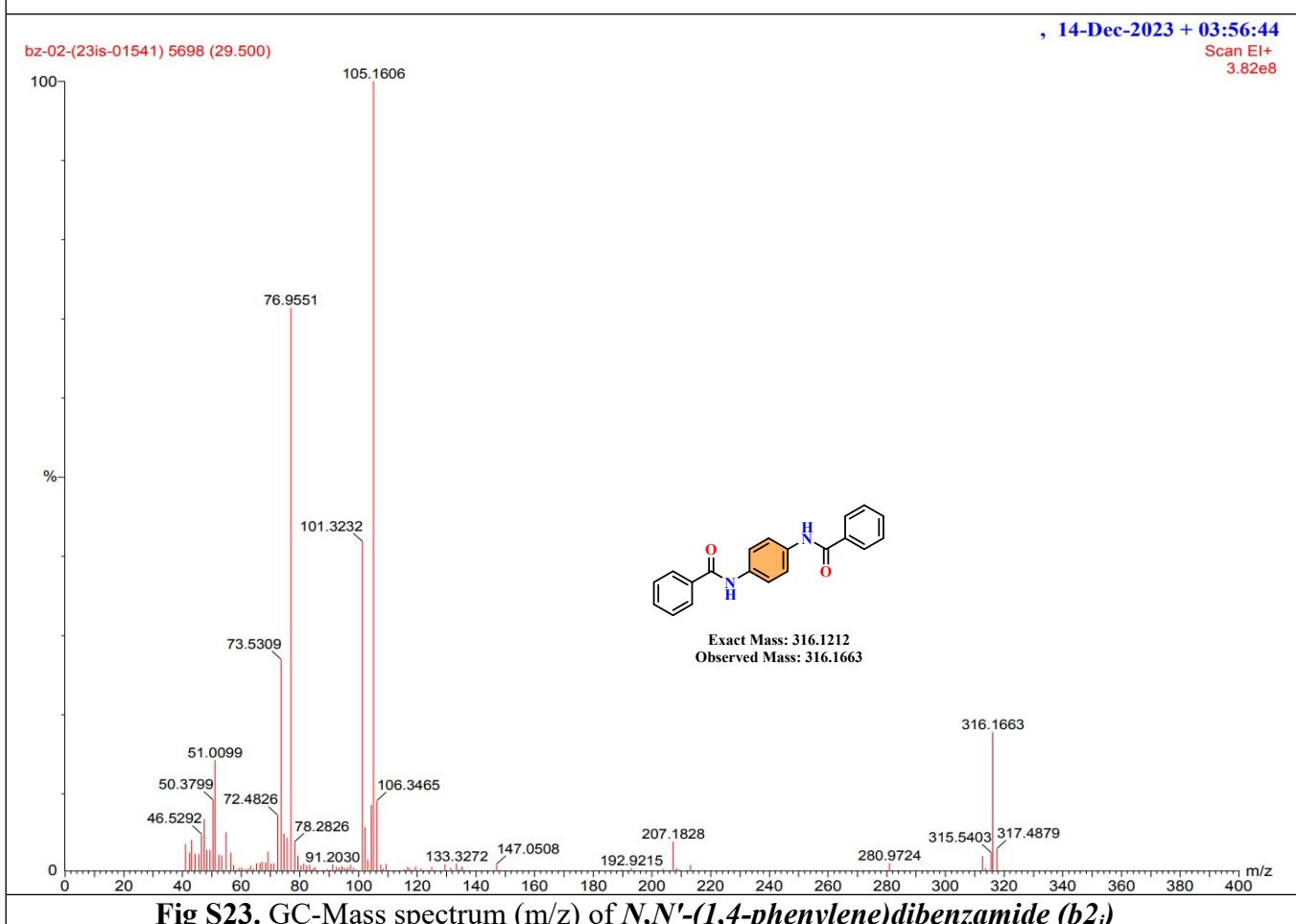


Fig S23. GC-Mass spectrum (m/z) of *N,N'*-(1,4-phenylene)dibenzamide (*b2<sub>i</sub>*)

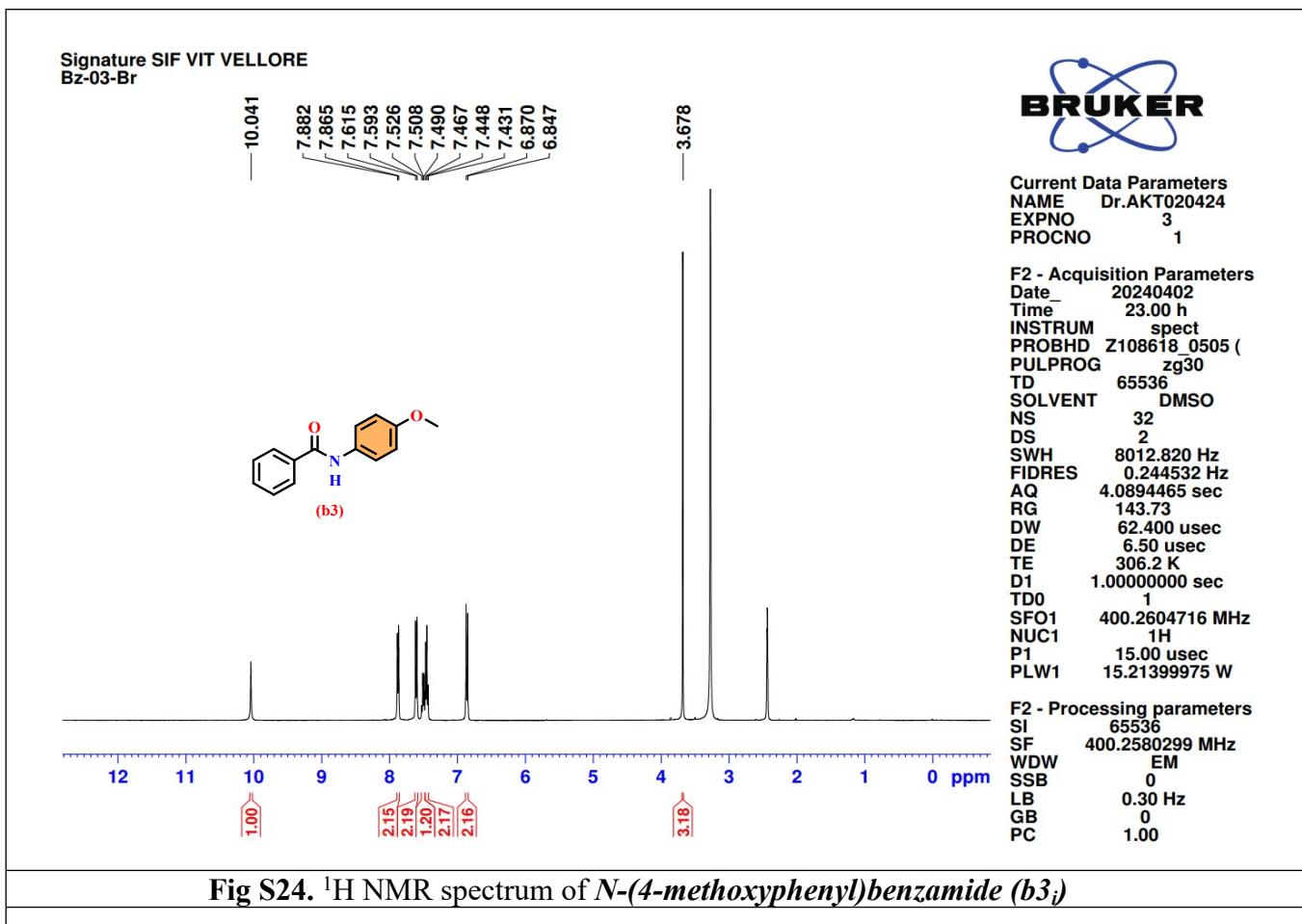


Fig S24.  $^1\text{H}$  NMR spectrum of *N*-(4-methoxyphenyl)benzamide (*b3*)

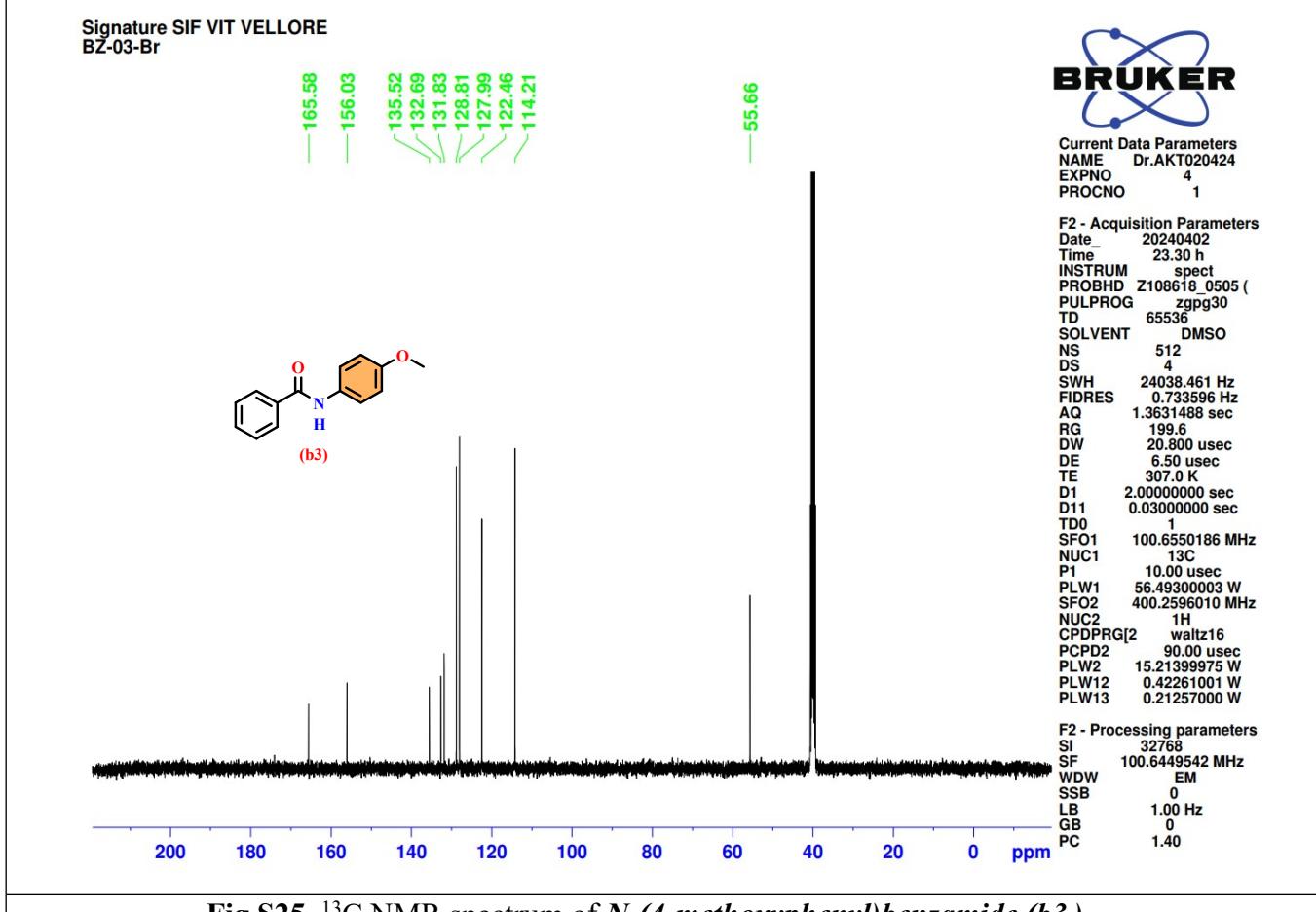
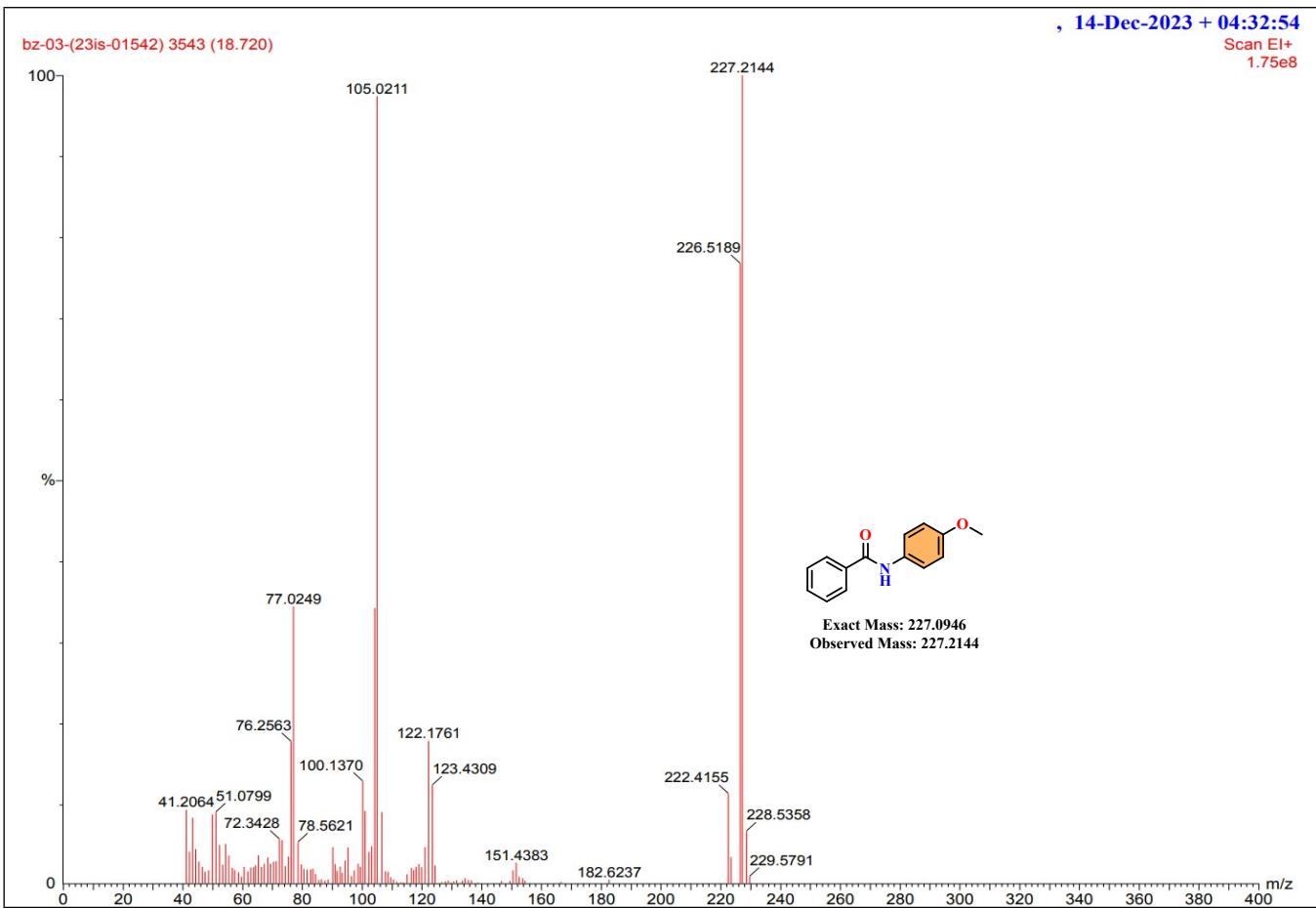


Fig S25.  $^{13}\text{C}$  NMR spectrum of *N*-(4-methoxyphenyl)benzamide (*b3*)



**Fig S26.** GC-Mass spectrum ( $m/z$ ) of *N*-(4-methoxyphenyl)benzamide ( $b3_i$ )

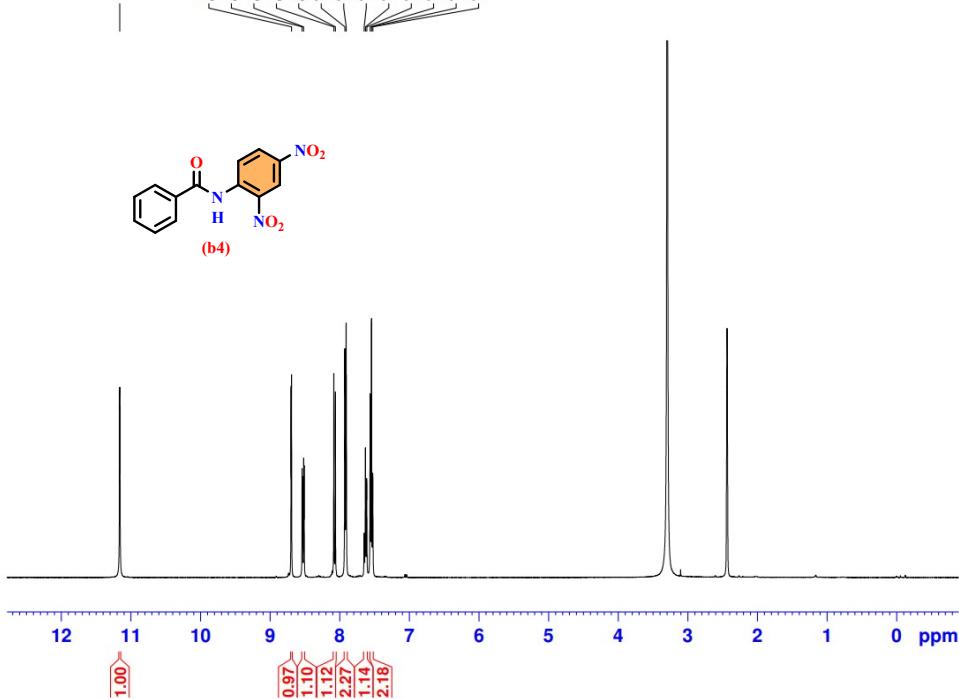
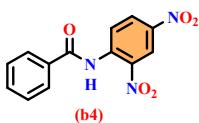
Signature SIF VIT VELLORE  
Bz-04-Br



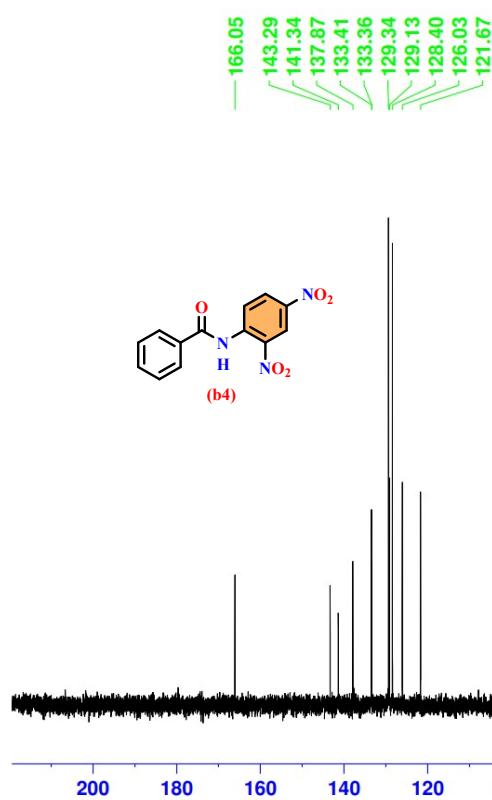
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EXPNO 34  
PROCNO 1

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PULPROG zg30  
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SOLVENT DMSO  
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DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 127.79  
DW 62.400 usec  
DE 6.50 usec  
TE 304.2 K  
D1 1.0000000 sec  
TD0 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

F2 - Processing parameters  
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SF 400.2580287 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



**Fig S27.**  $^1\text{H}$  NMR spectrum of *N*-(2,4-dinitrophenyl)benzamide ( $b4_i$ )



Current Data Parameters  
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EXPNO 26  
PROCNO 1  
  
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Time 9.05 h  
INSTRUM spect  
PROBHD Z108618\_0505 (   
PULPROG zgpg30  
TD 65536  
SOLVENT DMSO  
NS 512  
DS 4  
SWH 24038.461 Hz  
FIDRES 0.733596 Hz  
AQ 1.3631488 sec  
RG 199.6  
DW 20.800 usec  
DE 6.50 usec  
TE 310.0 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TDO 1  
SFO1 100.6550186 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 56.49300003 W  
SFO2 400.2596010 MHz  
NUC2 1H  
CPDPRG[2 waltz16  
PCPD2 90.00 usec  
PLW2 15.21399975 W  
PLW12 0.42261001 W  
PLW13 0.21257000 W  
  
F2 - Processing parameters  
SI 32768  
SF 100.6449542 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

Fig S28.  $^{13}\text{C}$  NMR spectrum of *N*-(2,4-dinitrophenyl)benzamide (*b4*)

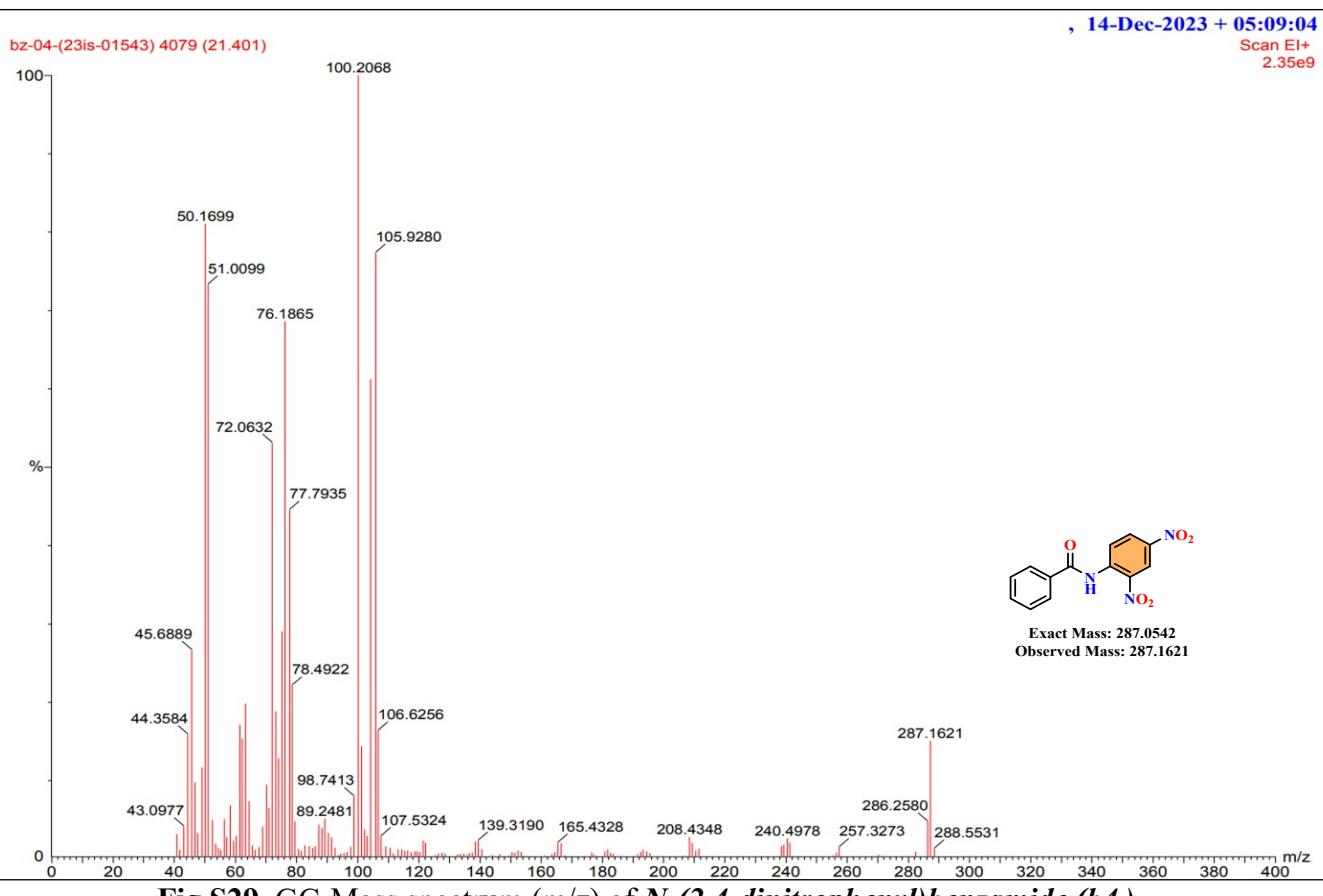


Fig S29. GC-Mass spectrum ( $m/z$ ) of *N*-(2,4-dinitrophenyl)benzamide (*b4*)

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BZ-05-Br

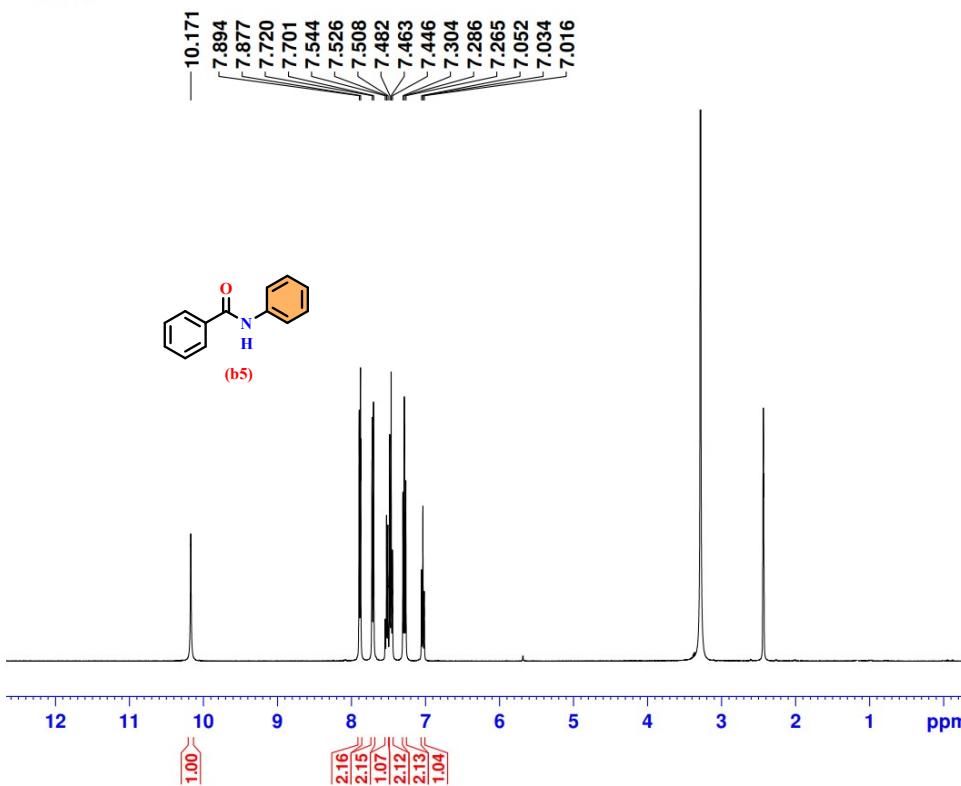


Fig S30.  $^1\text{H}$  NMR spectrum of *N*-phenylbenzamide (*b5*)

Signature SIF VIT VELLORE  
BZ-05-Br

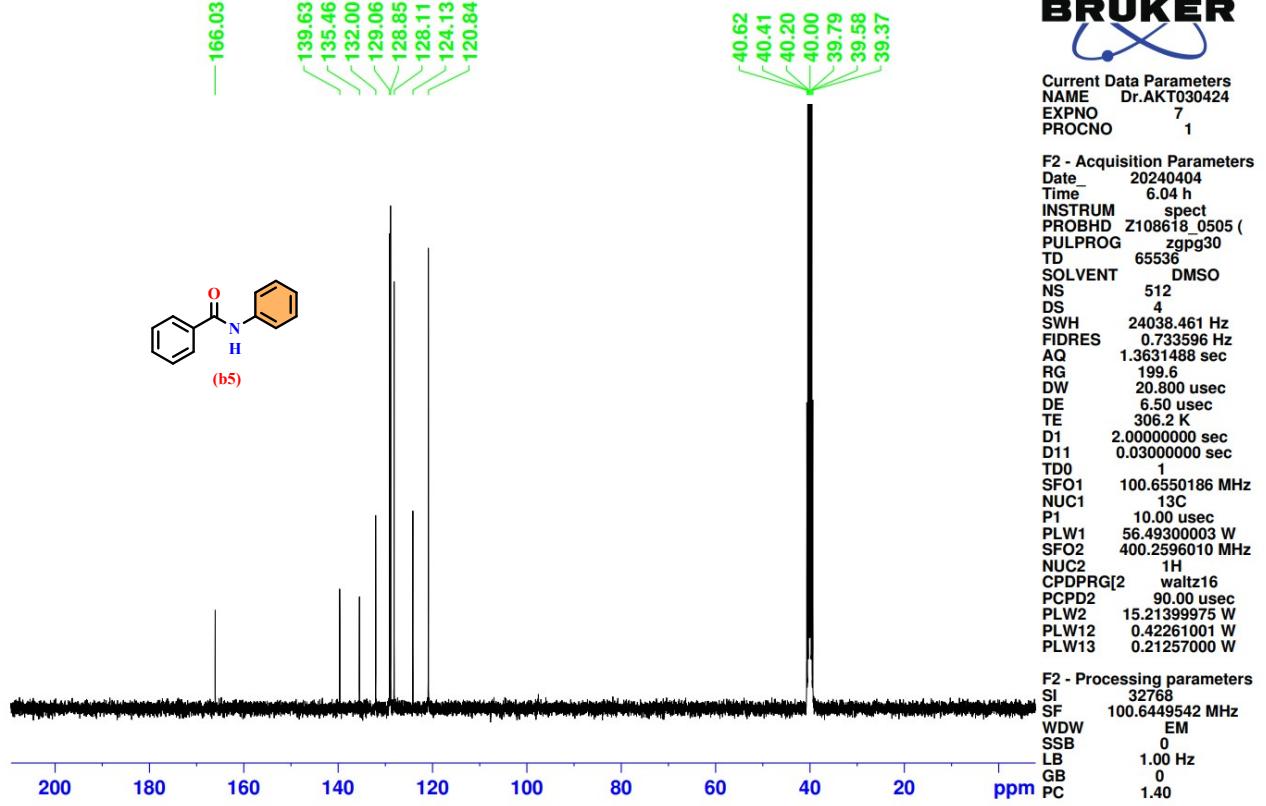
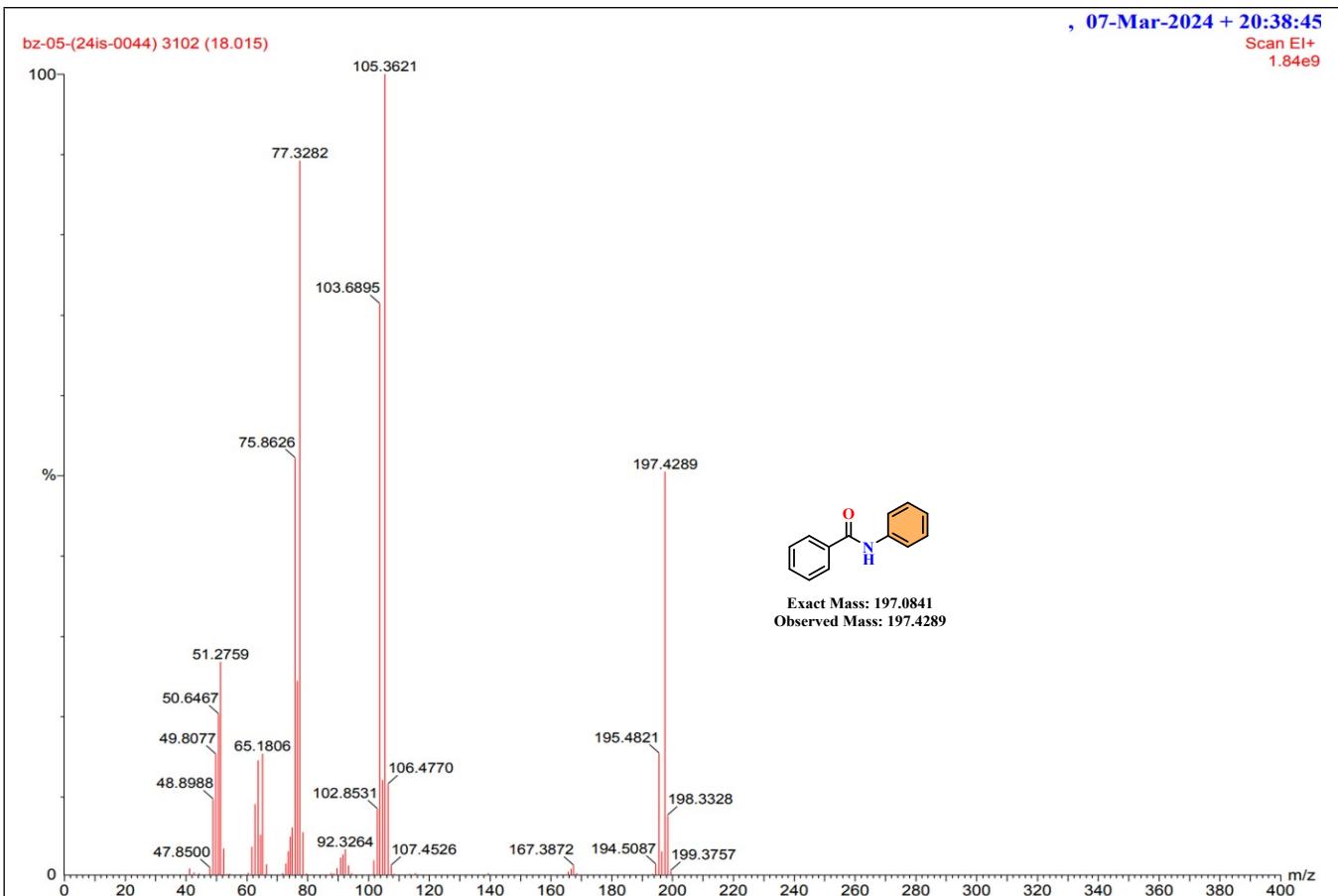
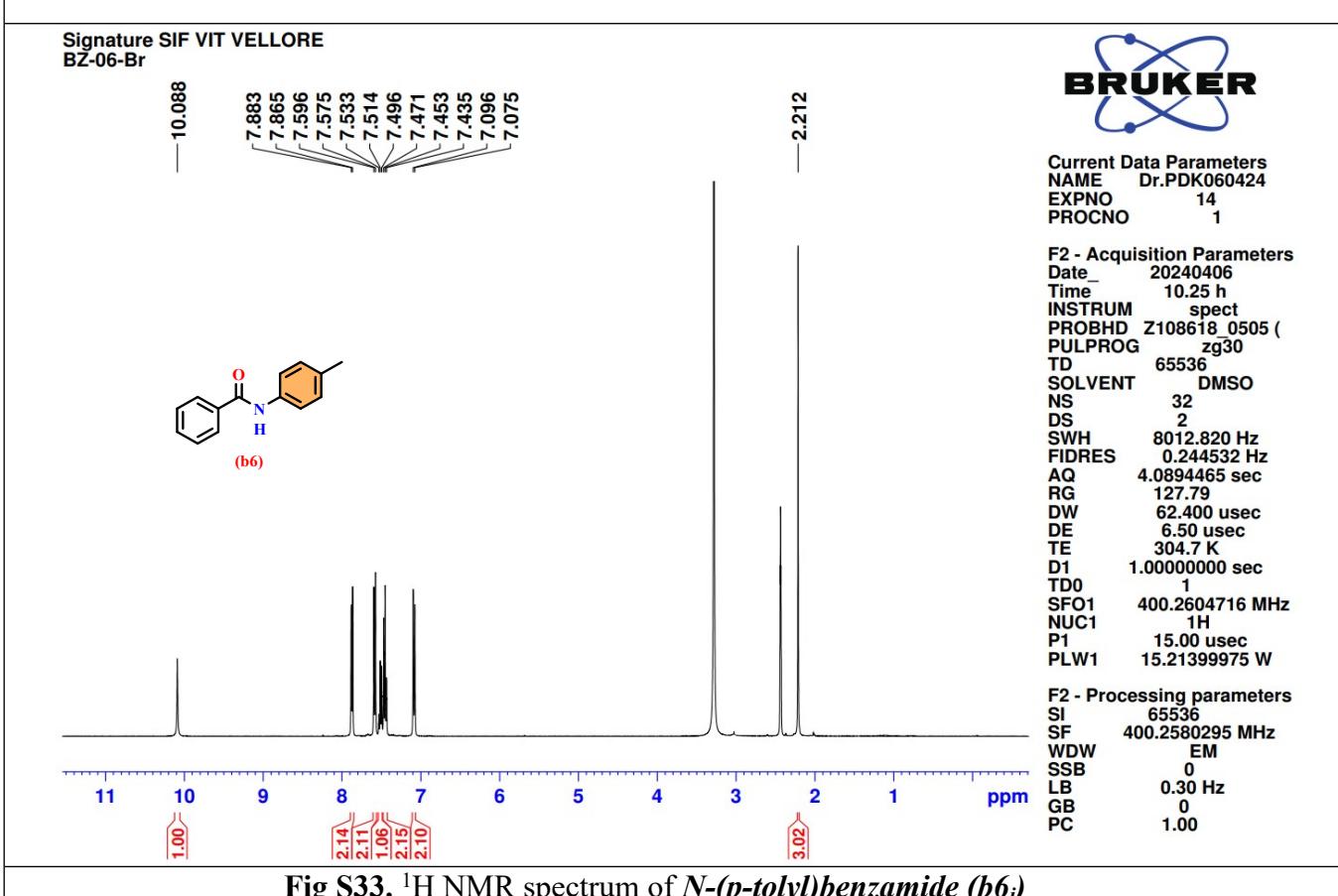


Fig S31.  $^{13}\text{C}$  NMR spectrum of *N*-phenylbenzamide (*b5*)



**Fig S32.** GC-Mass spectrum (m/z) of *N*-phenylbenzamide (*b5i*)



**Fig S33.**  $^1\text{H}$  NMR spectrum of *N*-(*p*-tolyl)benzamide (*b6i*)

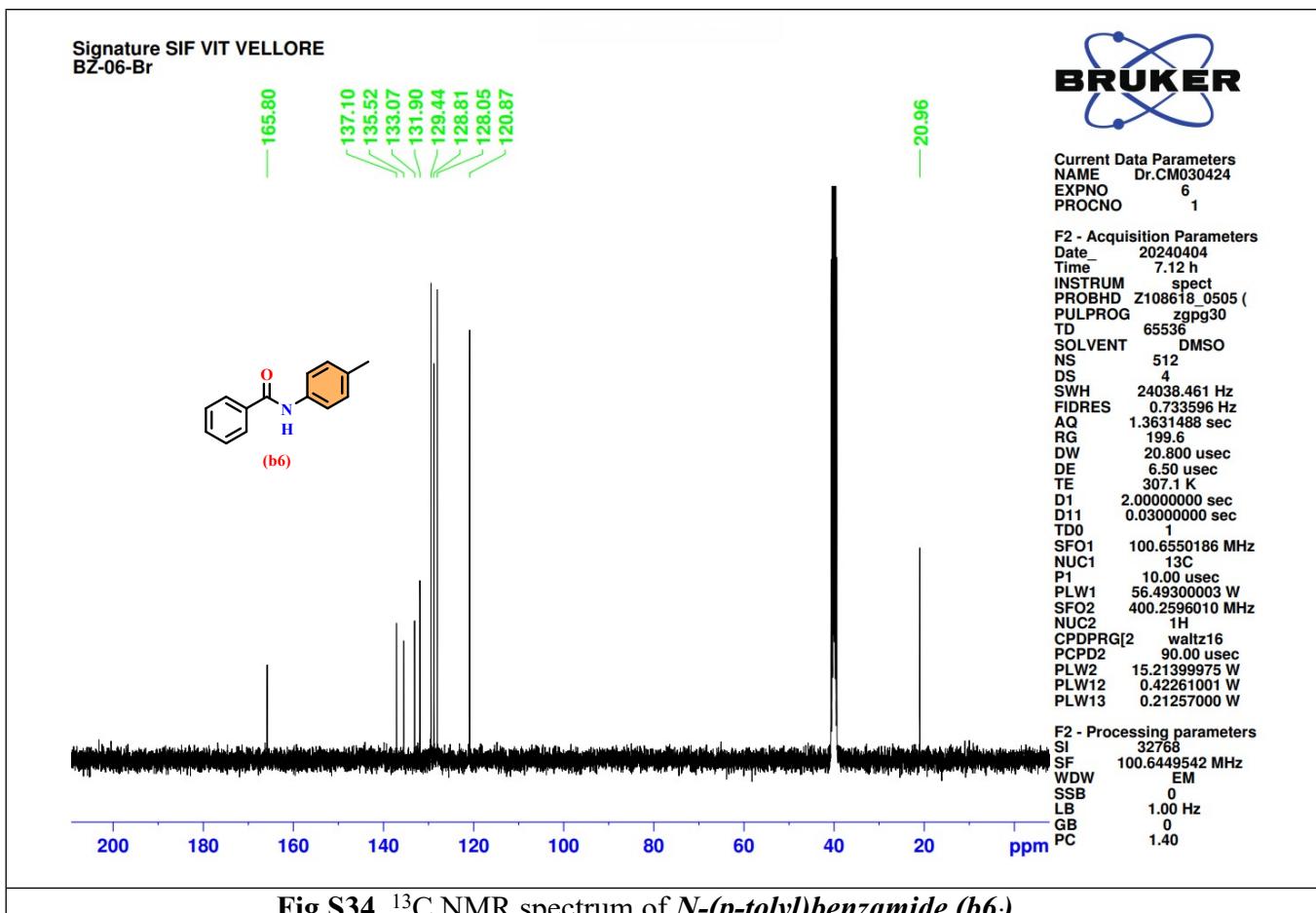


Fig S34. <sup>13</sup>C NMR spectrum of *N*-(*p*-tolyl)benzamide (*b6i*)

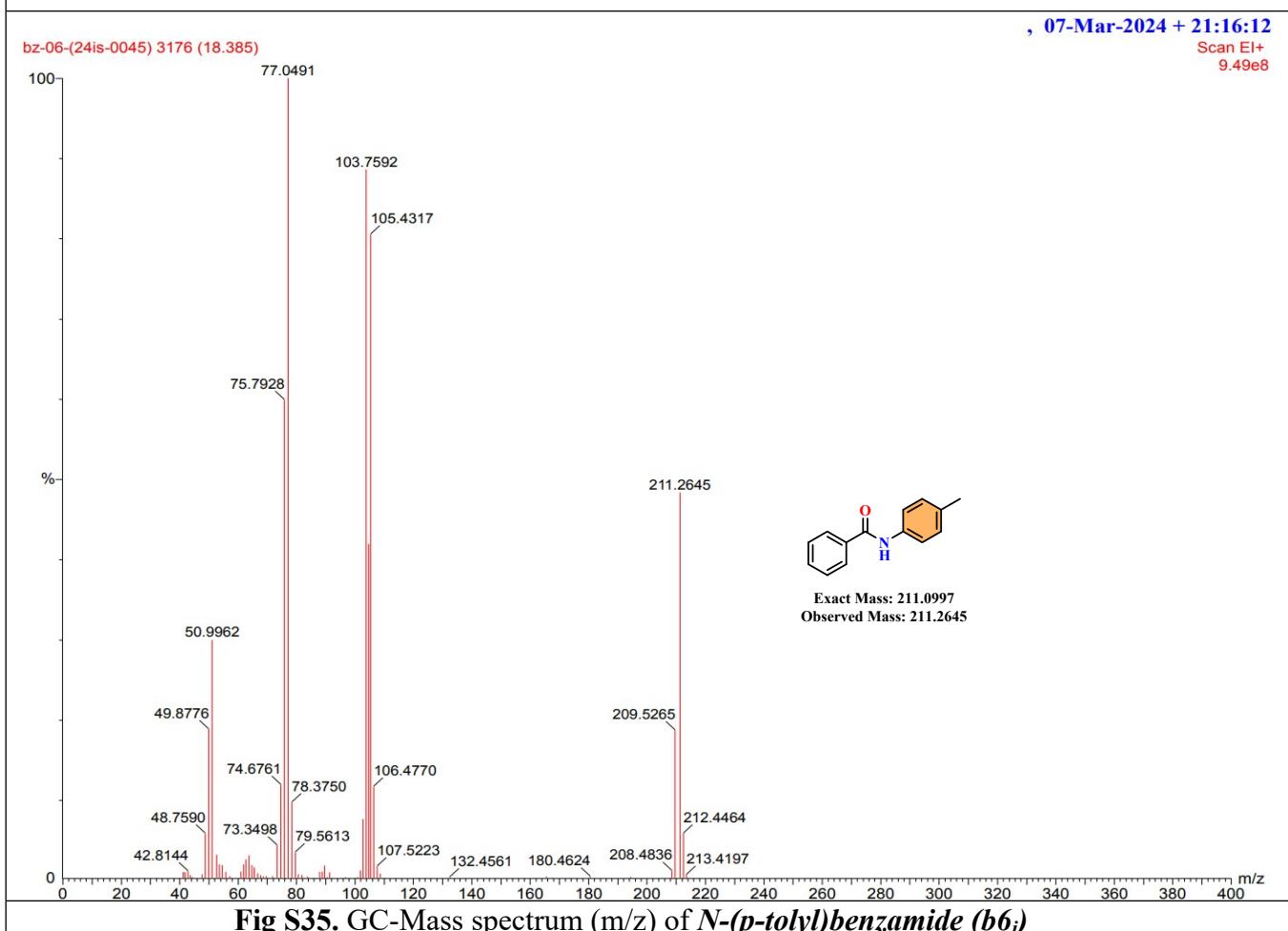


Fig S35. GC-Mass spectrum (m/z) of *N*-(*p*-tolyl)benzamide (*b6i*)

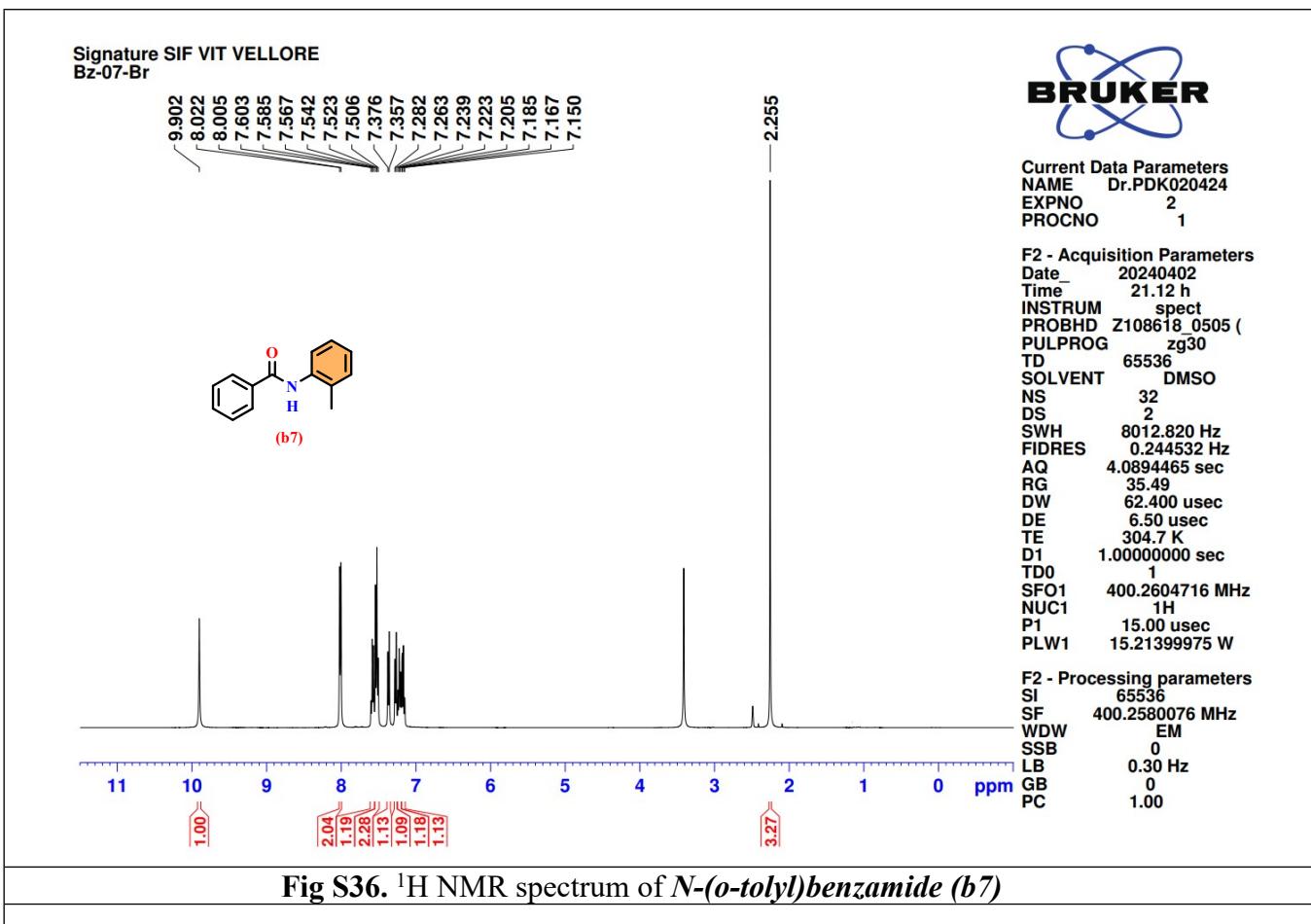


Fig S36.  $^1\text{H}$  NMR spectrum of *N*-(*o*-tolyl)benzamide (**b7**)

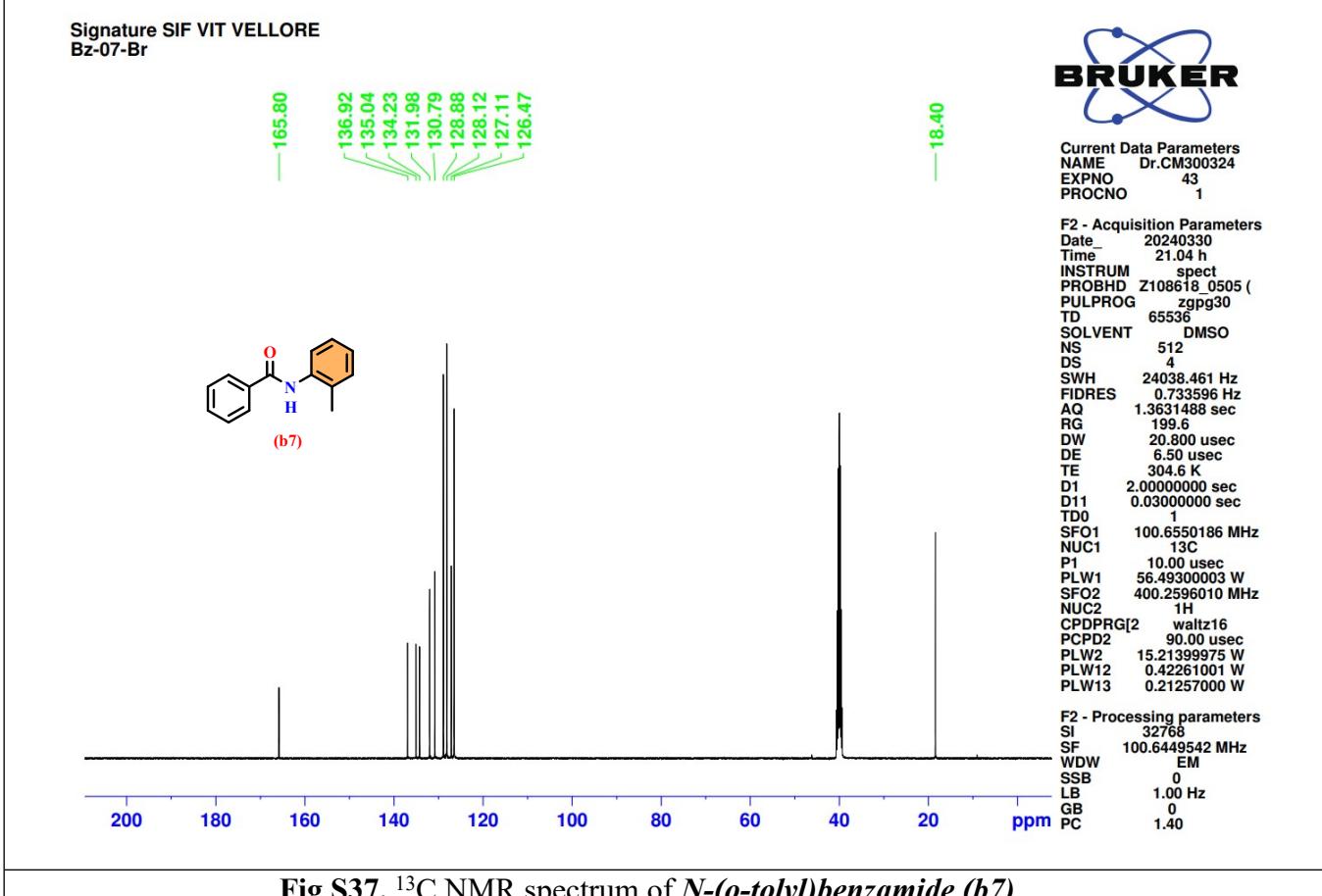


Fig S37.  $^{13}\text{C}$  NMR spectrum of *N*-(*o*-tolyl)benzamide (**b7**)

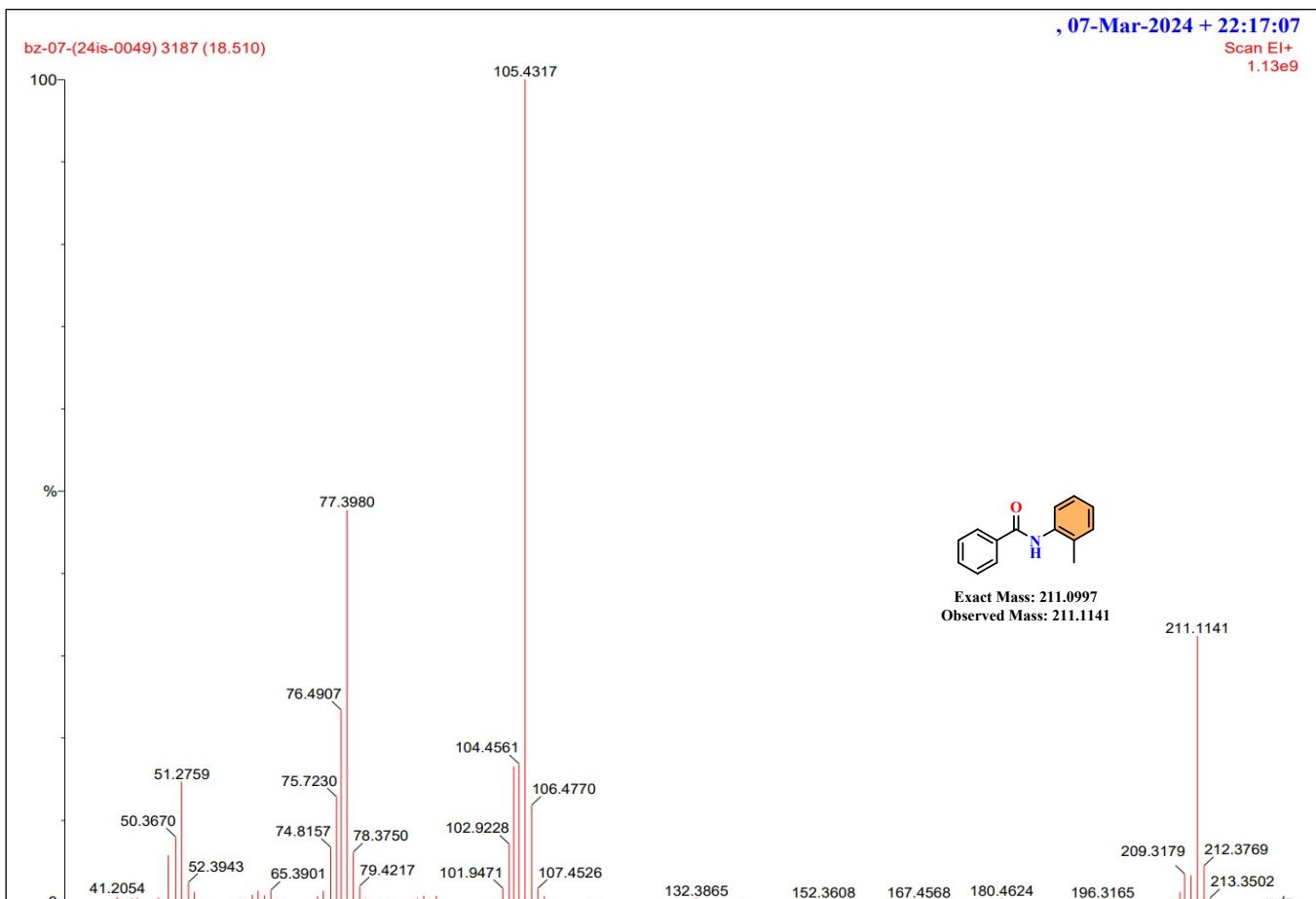


Fig S38. GC-Mass spectrum ( $m/z$ ) of *N*-(*o*-tolyl)benzamide (**b7**)

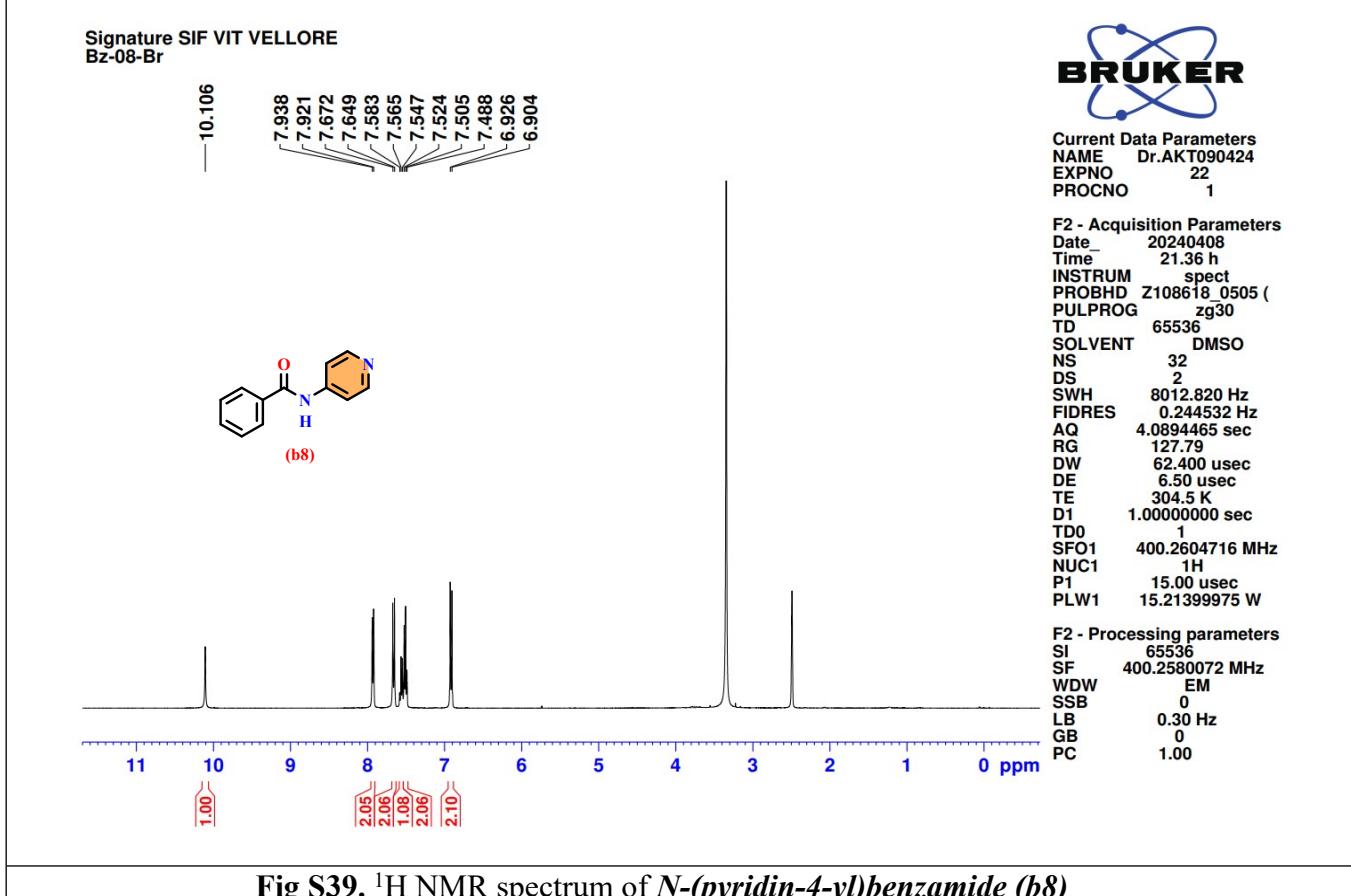


Fig S39.  $^1\text{H}$  NMR spectrum of *N*-(pyridin-4-yl)benzamide (**b8**)

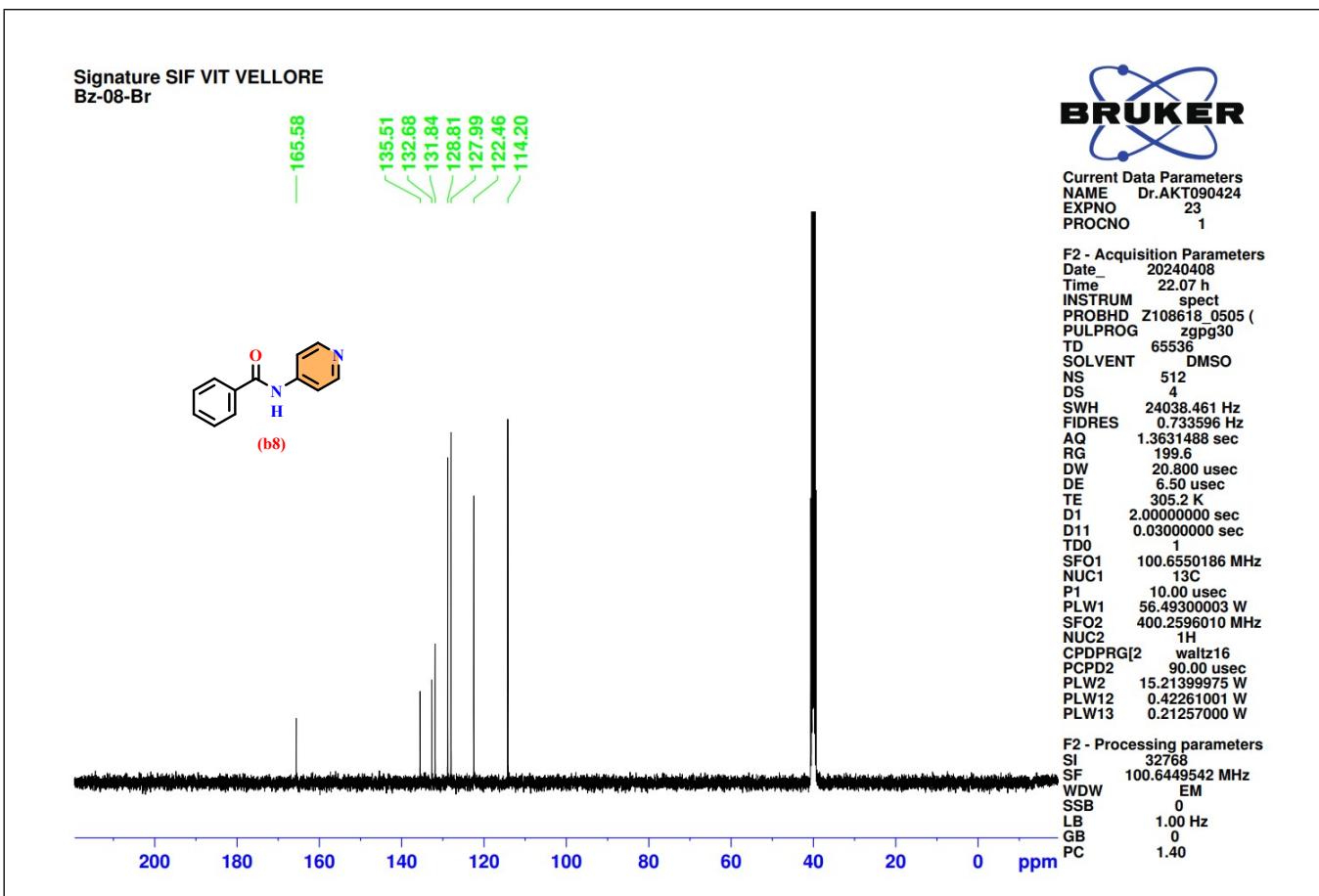


Fig S40.  $^{13}\text{C}$  NMR spectrum of *N*-(pyridin-4-yl)benzamide (*b8*)

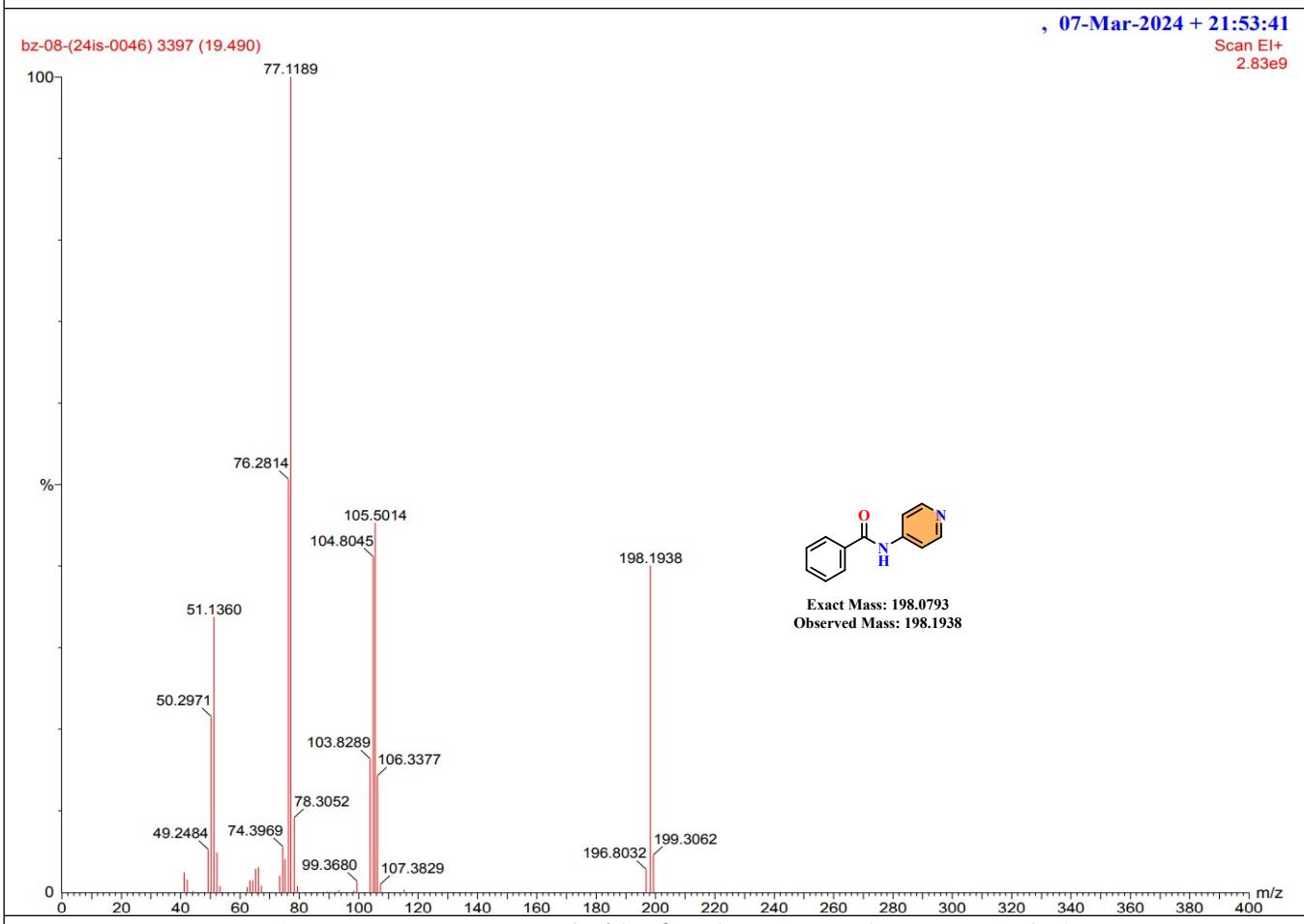


Fig S41. GC-Mass spectrum (m/z) of *N*-(pyridin-4-yl)benzamide (*b8*)

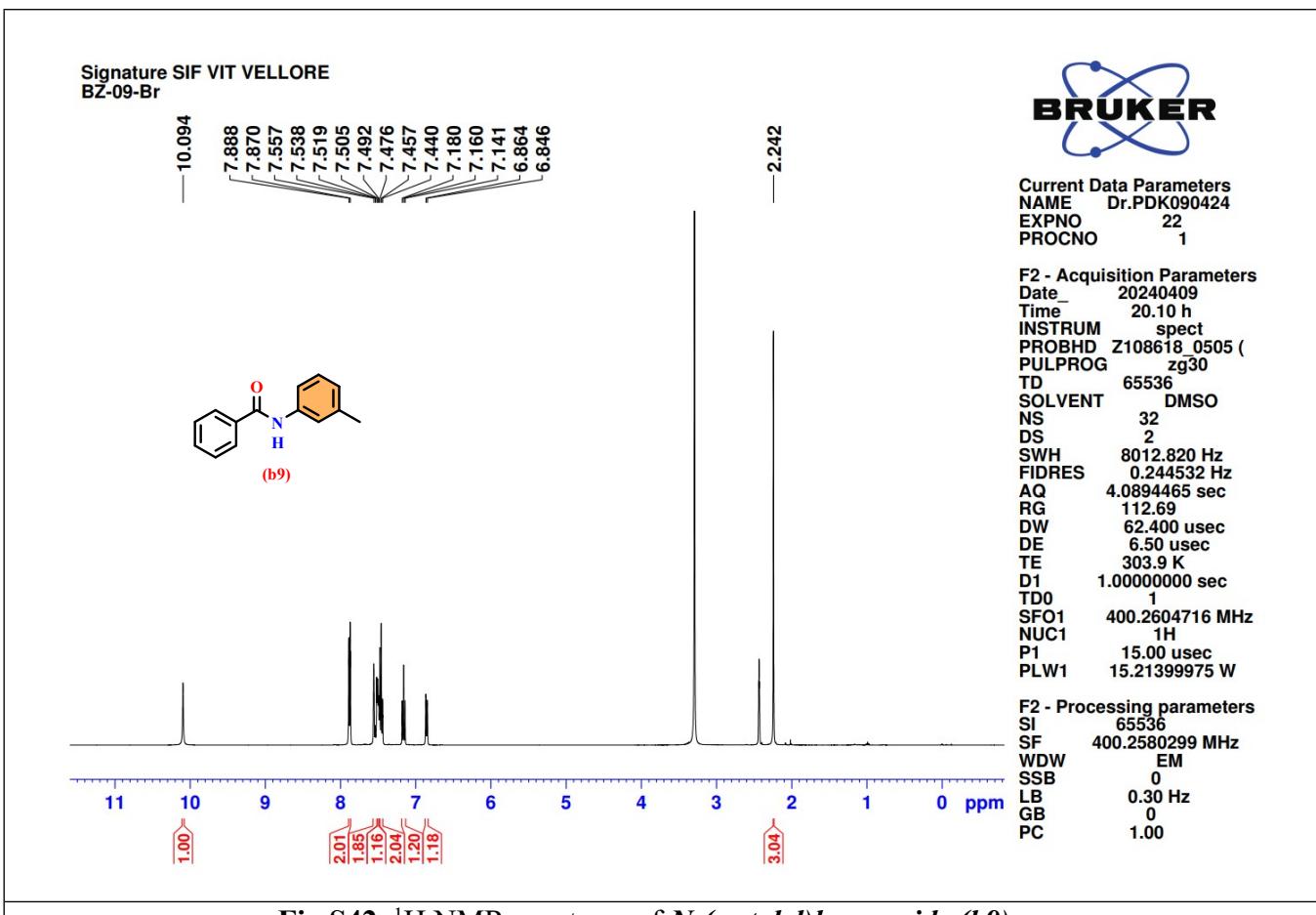


Fig S42.  $^1\text{H}$  NMR spectrum of *N*-(*m*-tolyl)benzamide (*b9*)

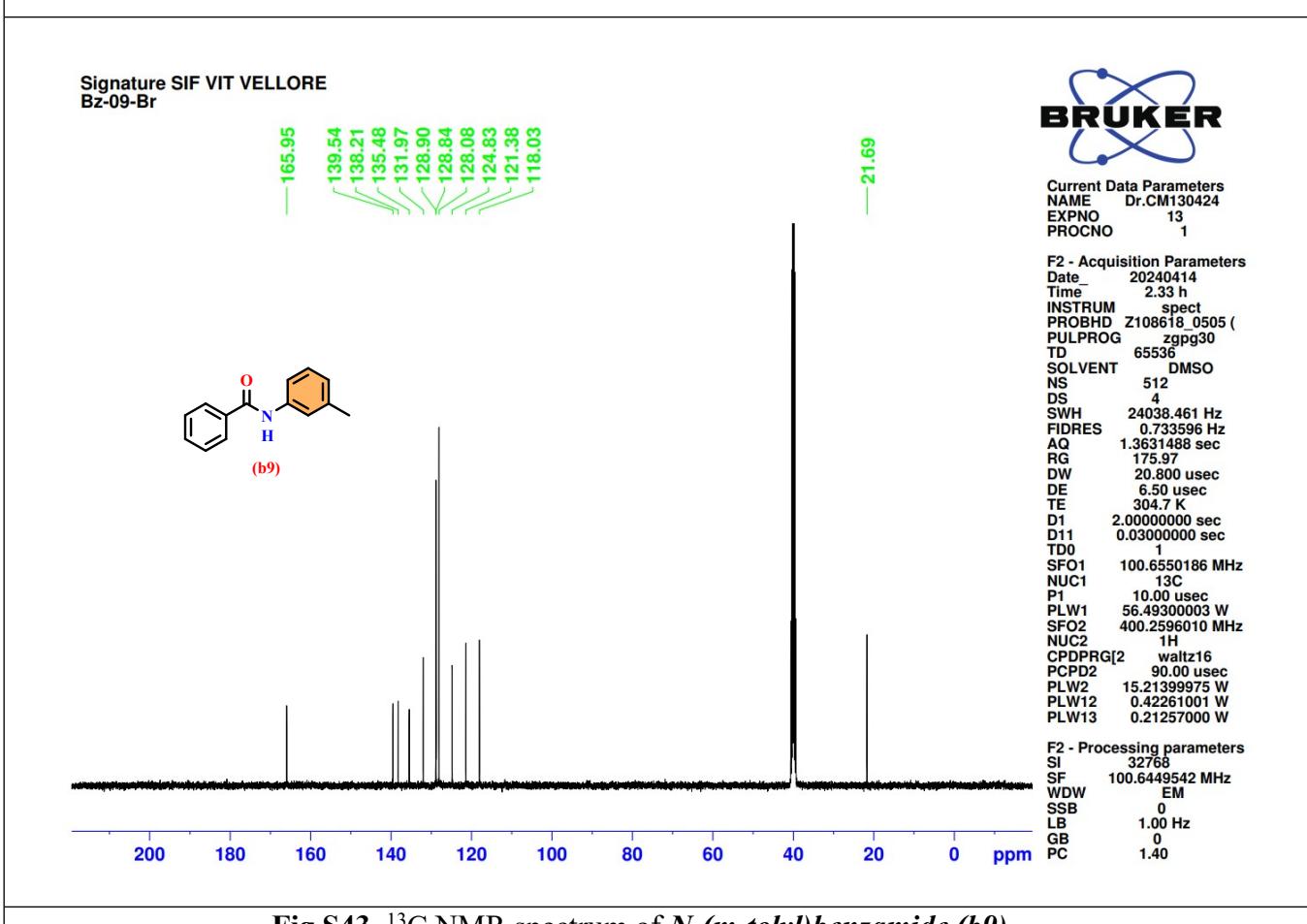
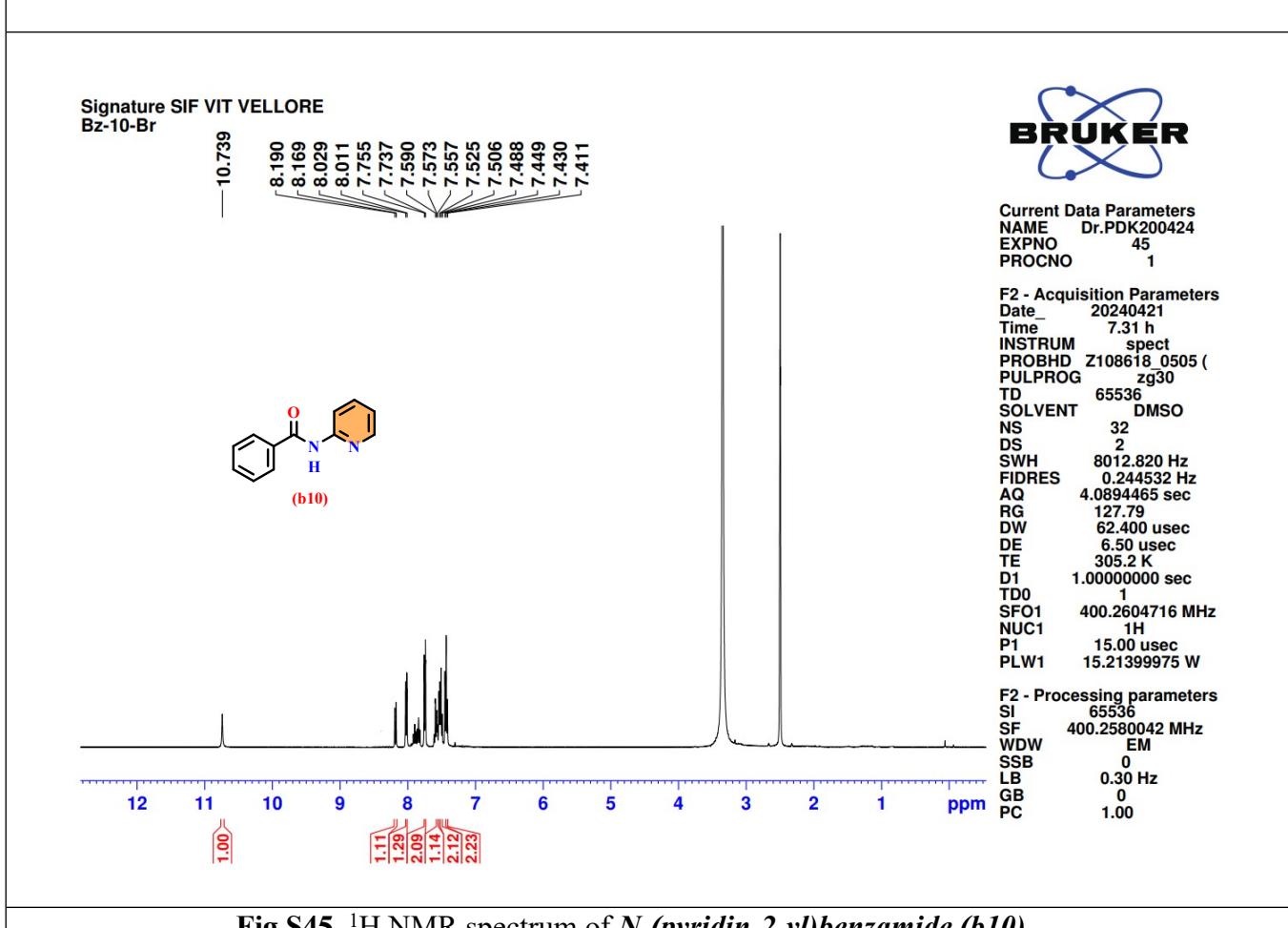
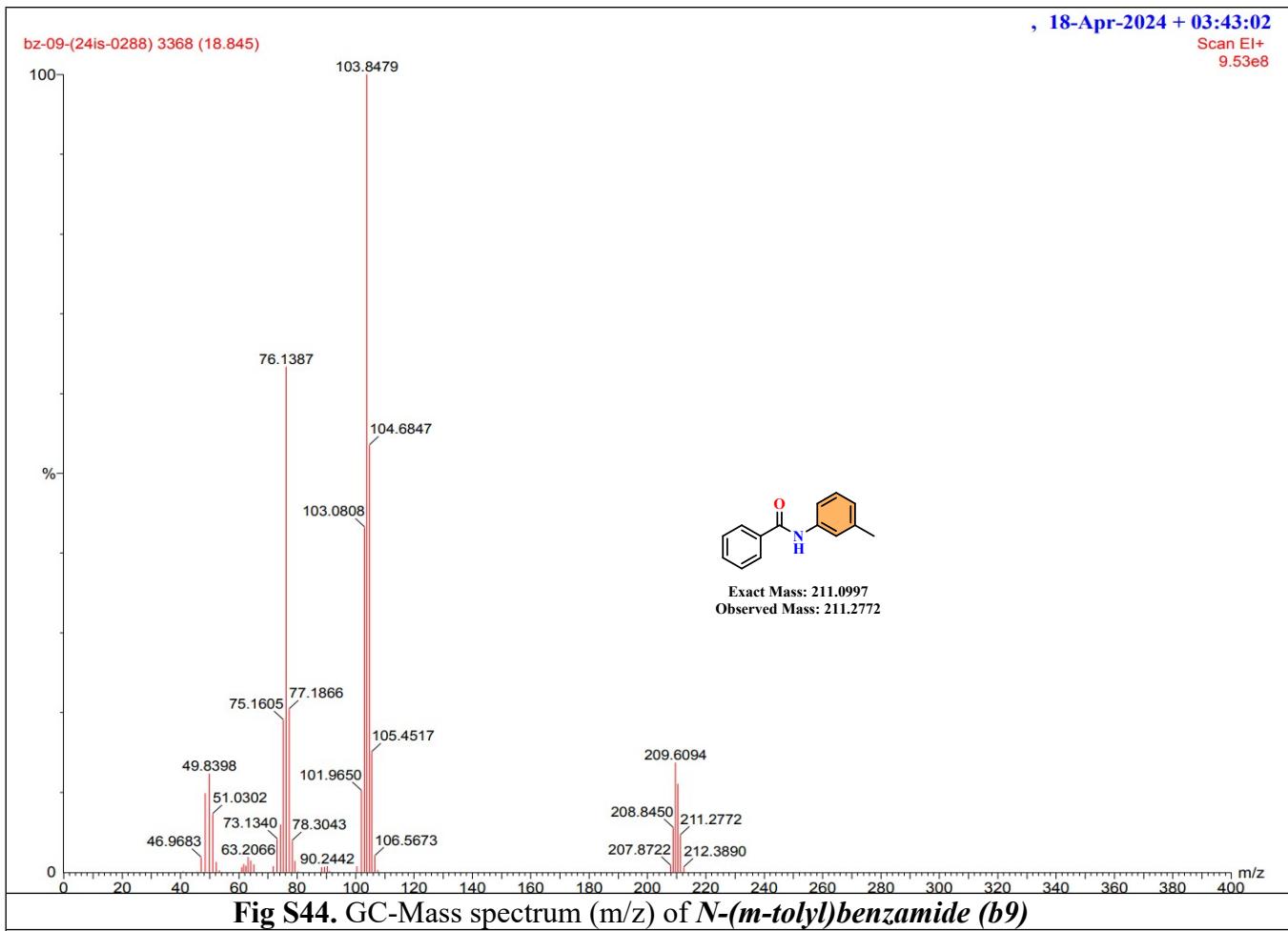


Fig S43.  $^{13}\text{C}$  NMR spectrum of *N*-(*m*-tolyl)benzamide (*b9*)



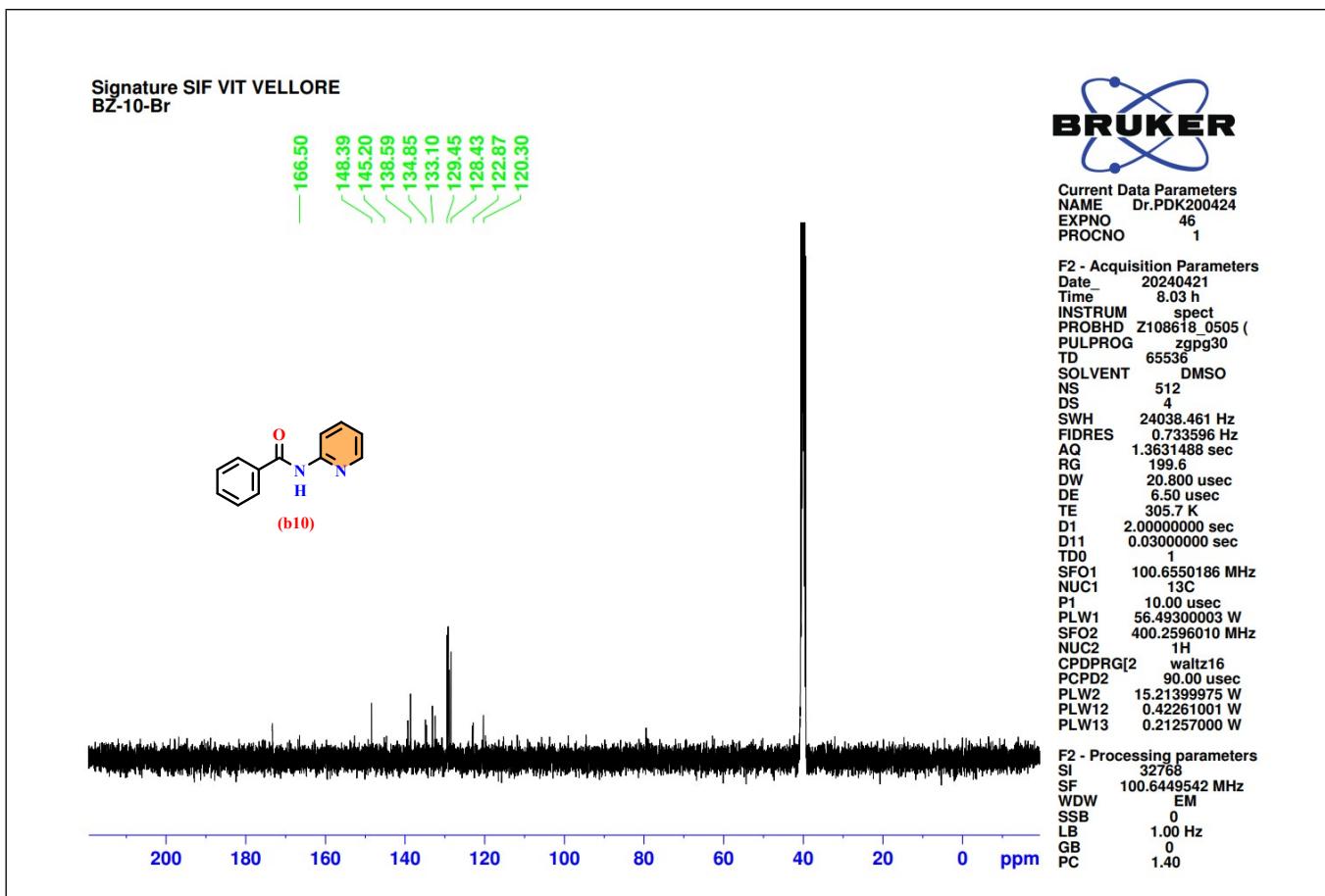


Fig S46.  $^{13}\text{C}$  NMR spectrum of *N*-(pyridin-2-yl)benzamide (*b10*)

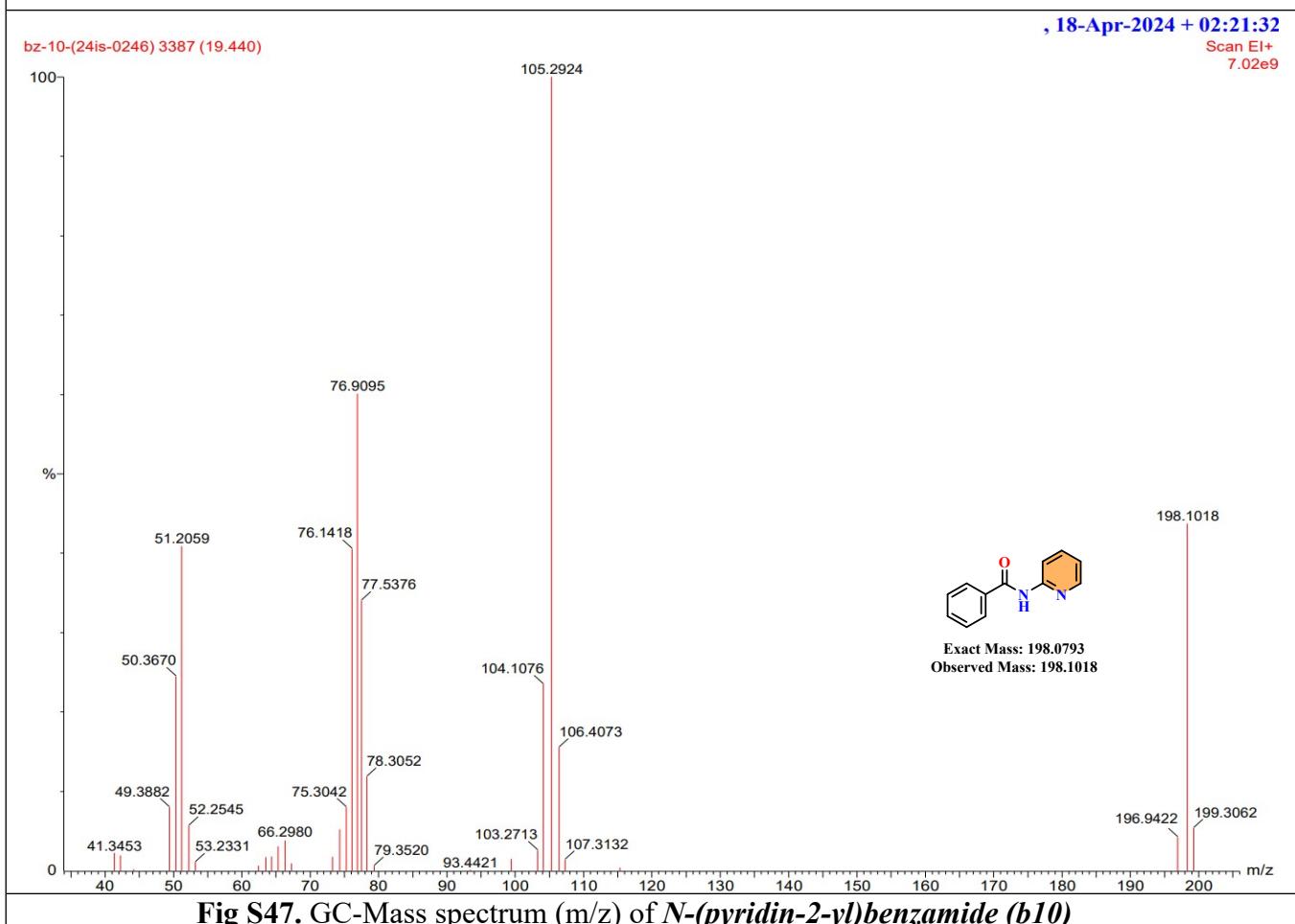


Fig S47. GC-Mass spectrum (m/z) of *N*-(pyridin-2-yl)benzamide (*b10*)

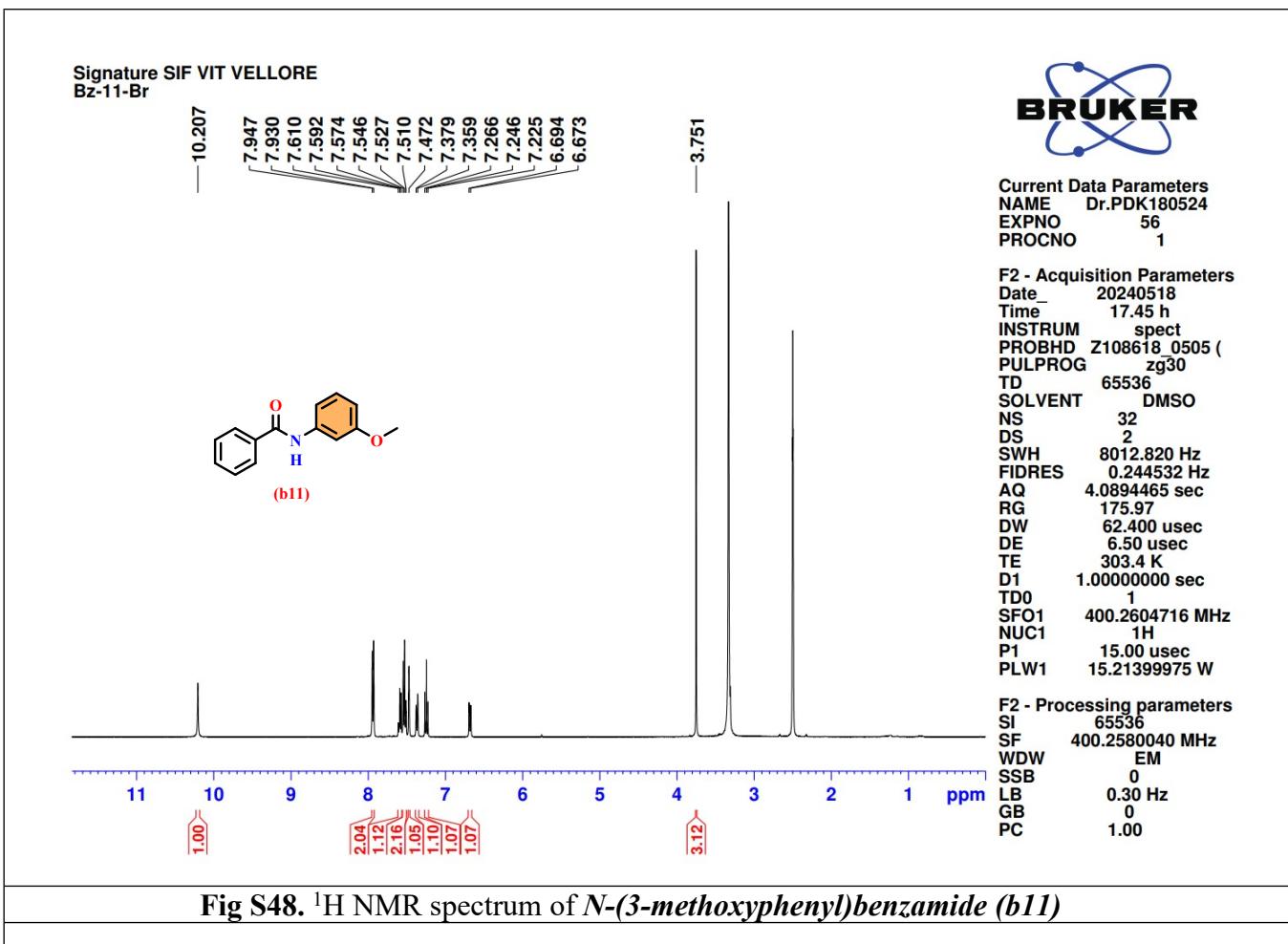


Fig S48.  $^1\text{H}$  NMR spectrum of *N*-(3-methoxyphenyl)benzamide (**b11**)

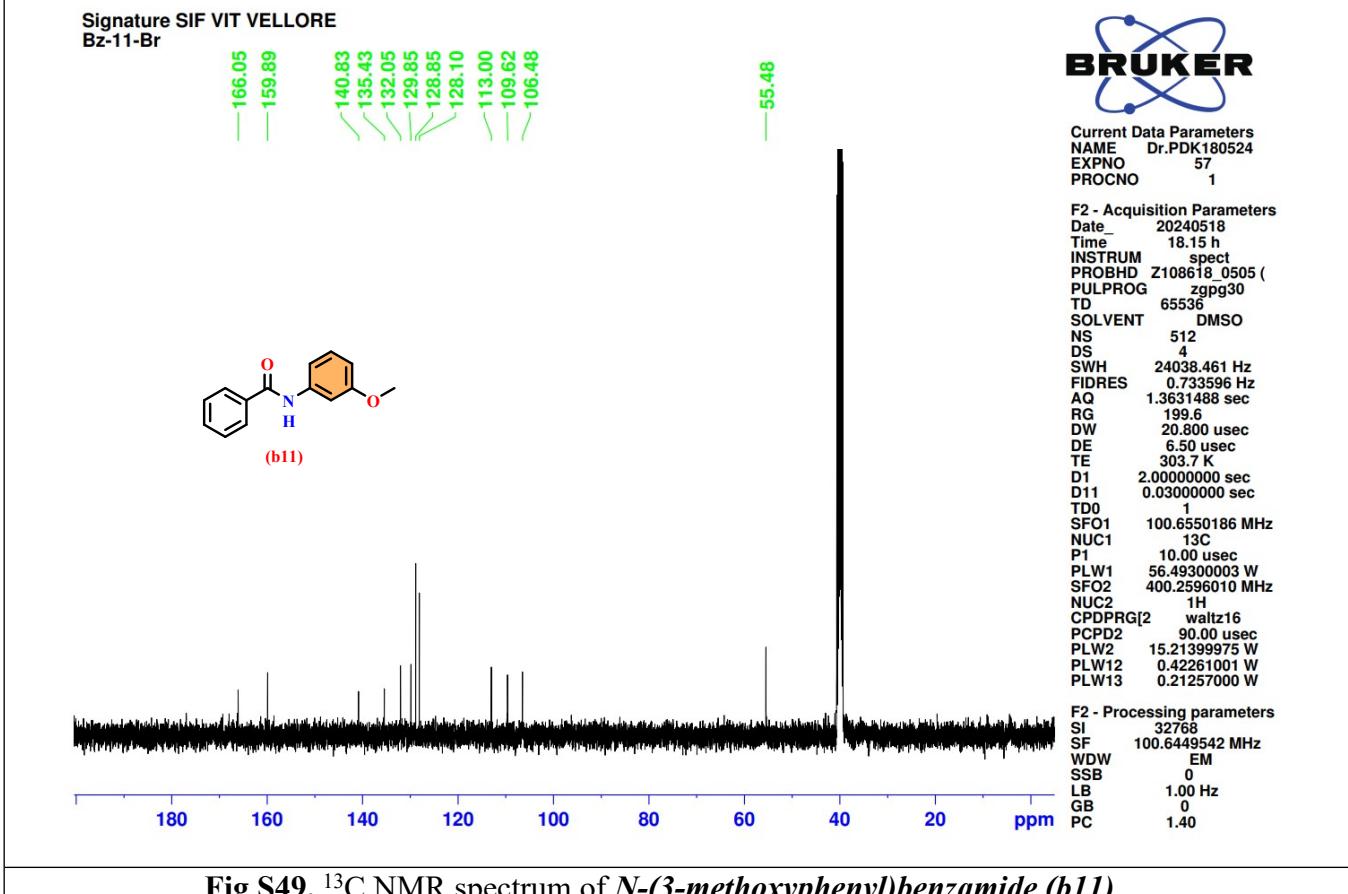
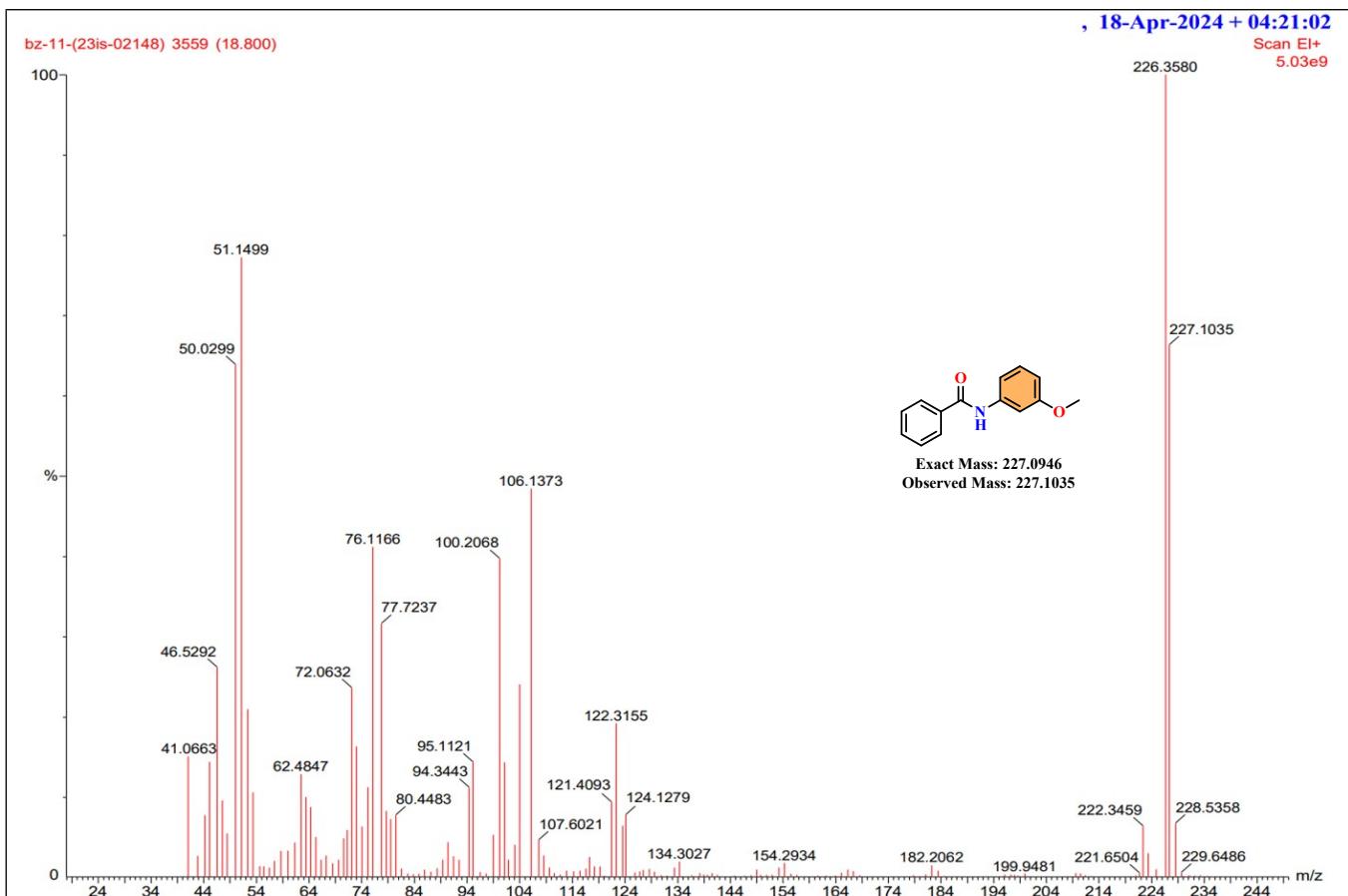


Fig S49.  $^{13}\text{C}$  NMR spectrum of *N*-(3-methoxyphenyl)benzamide (**b11**)



**Fig S50.** GC-Mass spectrum ( $m/z$ ) of *N*-(3-methoxyphenyl)benzamide (**b11**)

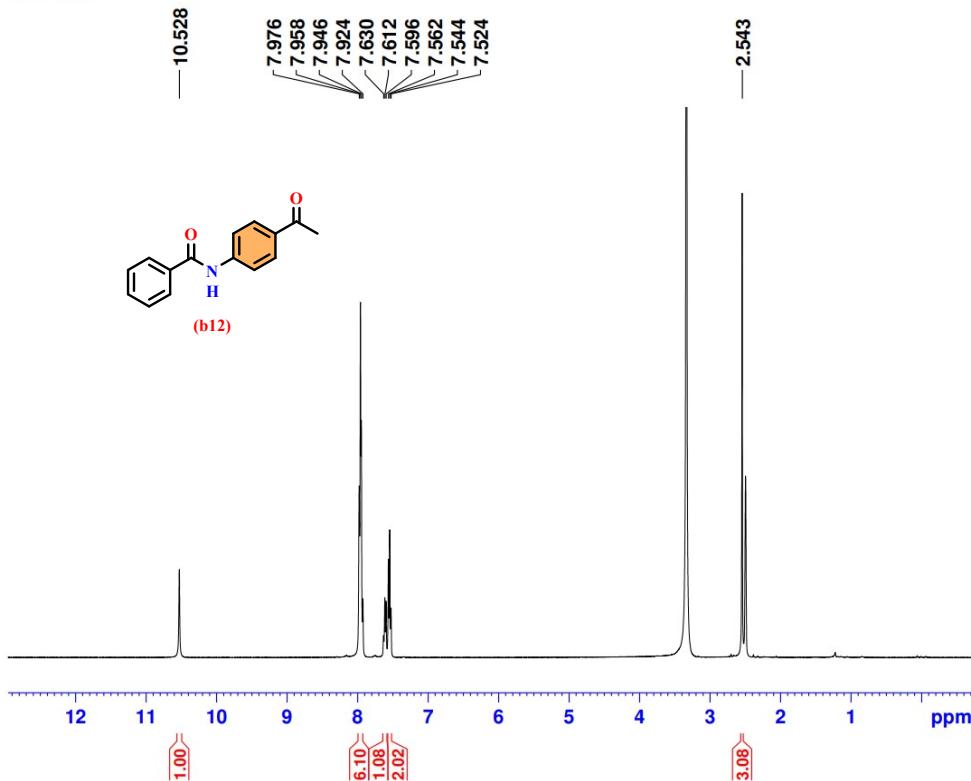
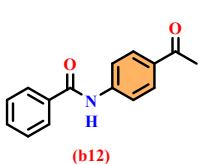
Signature SIF VIT VELLORE  
BZ-12-Br



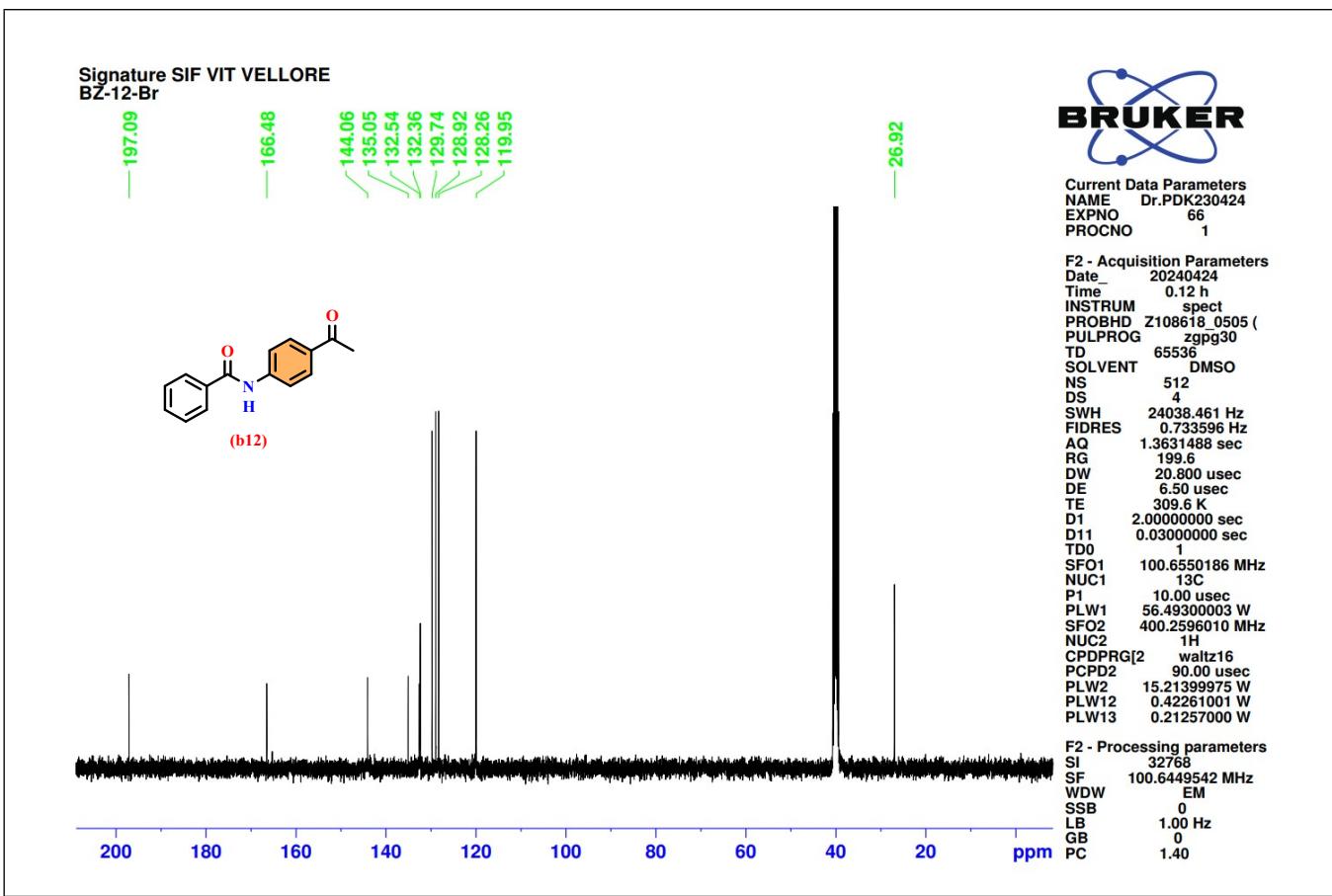
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AQ 4.0894465 sec  
RG 112.69  
DW 62.400 usec  
DE 6.50 usec  
TE 309.0 K  
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TD0 1  
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NUC1 1H  
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PLW1 15.21399975 W

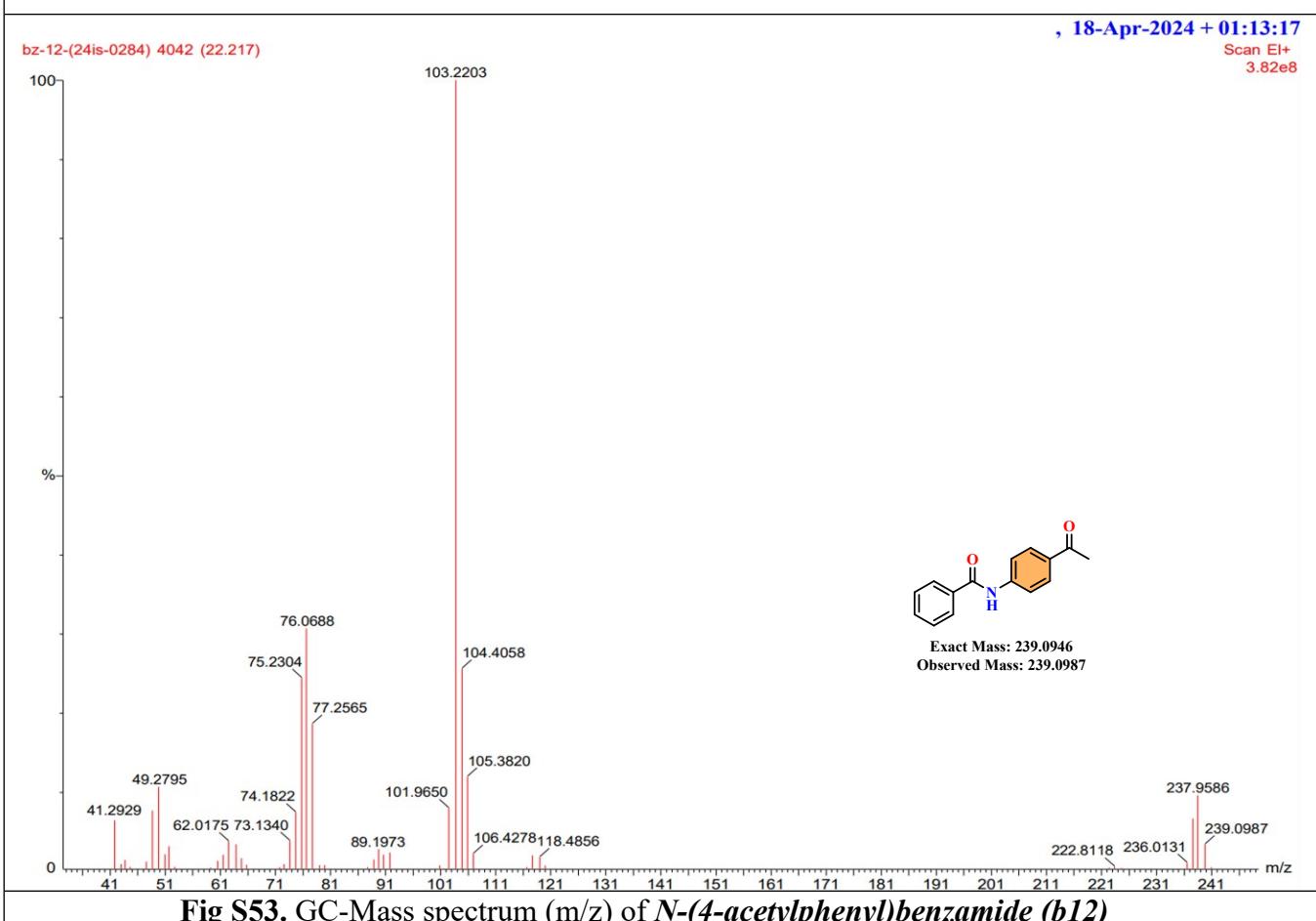
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LB 0.30 Hz  
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PC 1.00



**Fig S51.**  $^1\text{H}$  NMR spectrum of *N*-(4-acetylphenyl)benzamide (**b12**)



**Fig S52.**  $^{13}\text{C}$  NMR spectrum of *N*-(4-acetylphenyl)benzamide (**b12**)



**Fig S53.** GC-Mass spectrum (m/z) of *N*-(4-acetylphenyl)benzamide (**b12**)

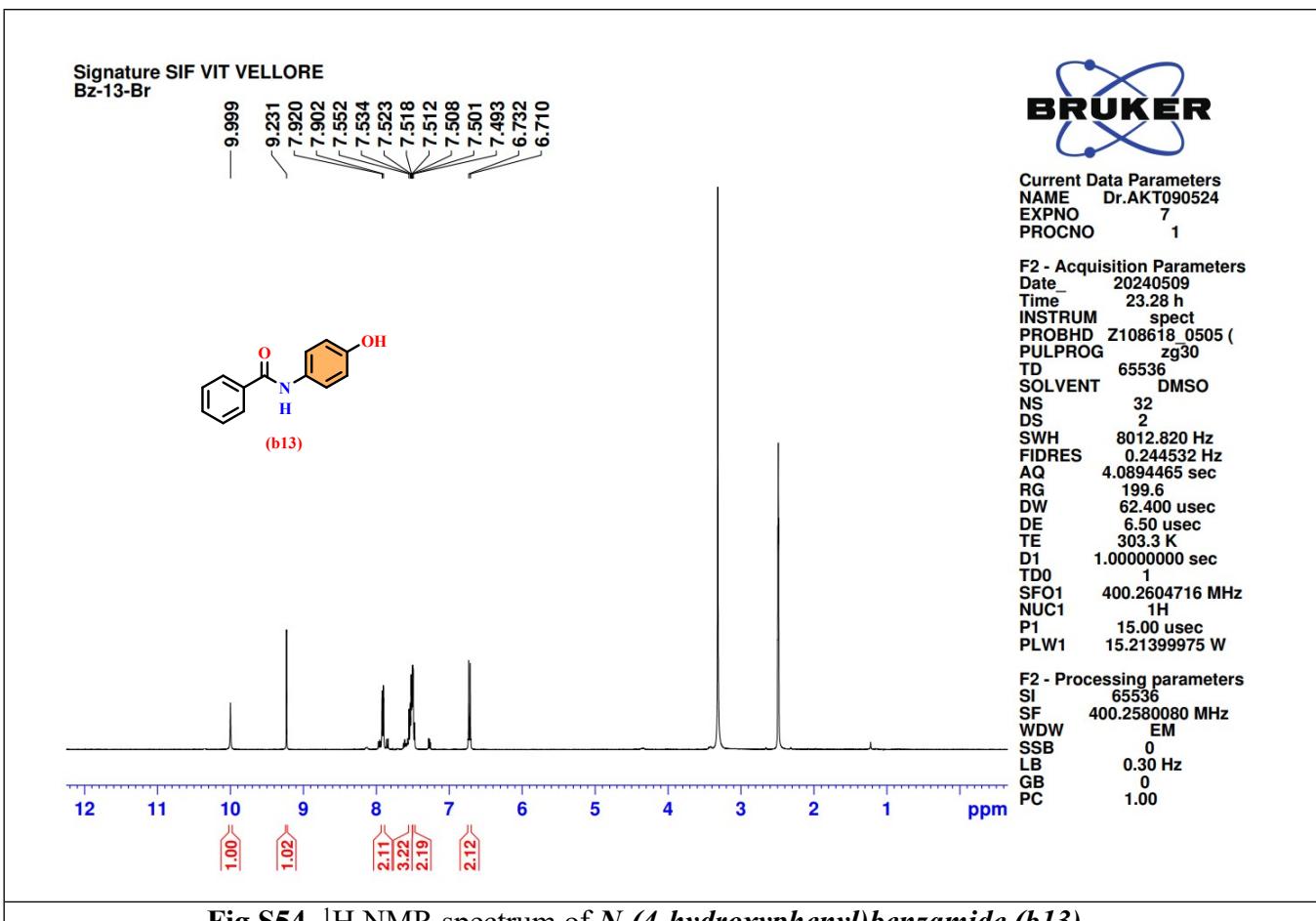


Fig S54.  $^1\text{H}$  NMR spectrum of *N*-(4-hydroxyphenyl)benzamide (**b13**)

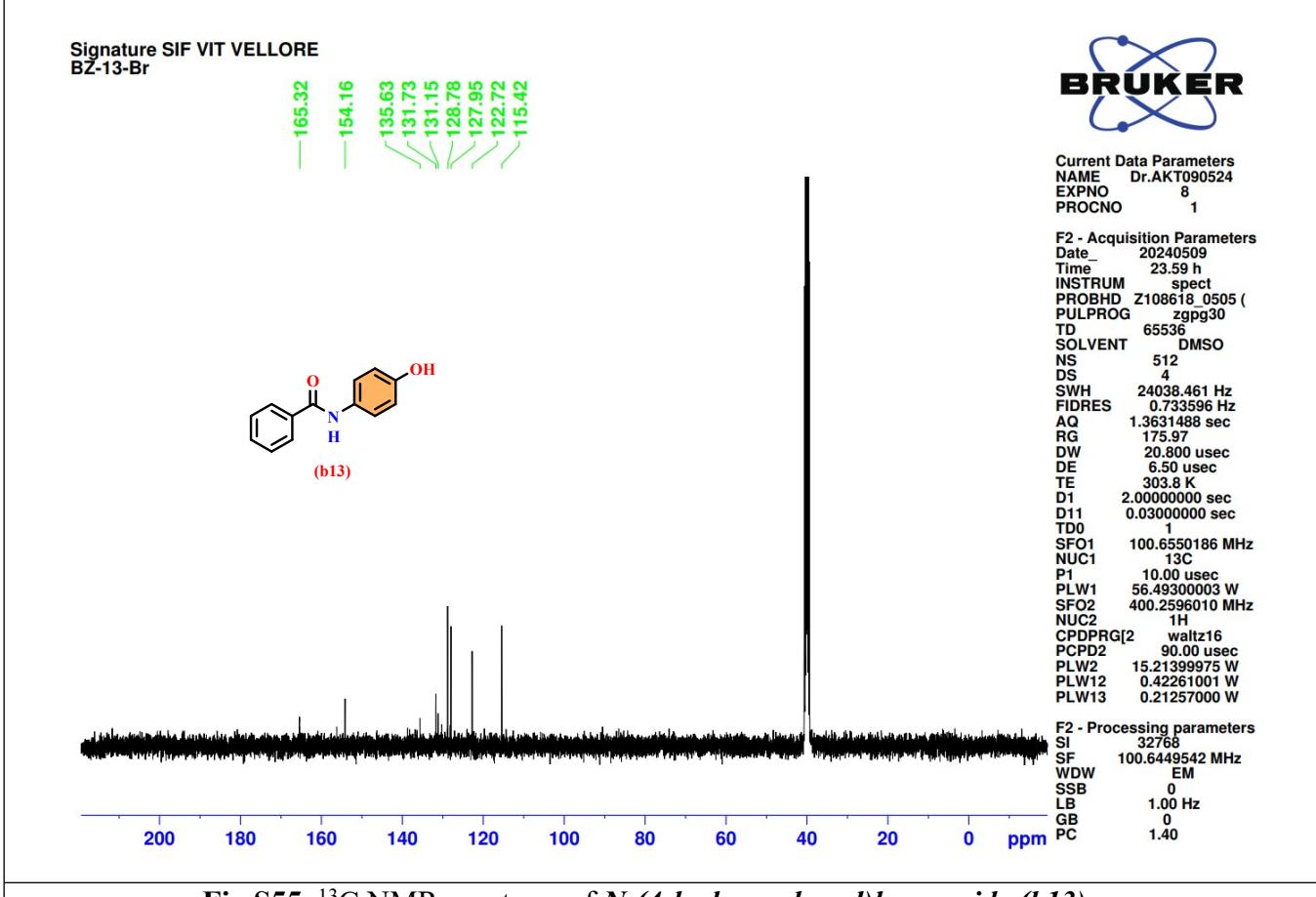


Fig S55.  $^{13}\text{C}$  NMR spectrum of *N*-(4-hydroxyphenyl)benzamide (**b13**)

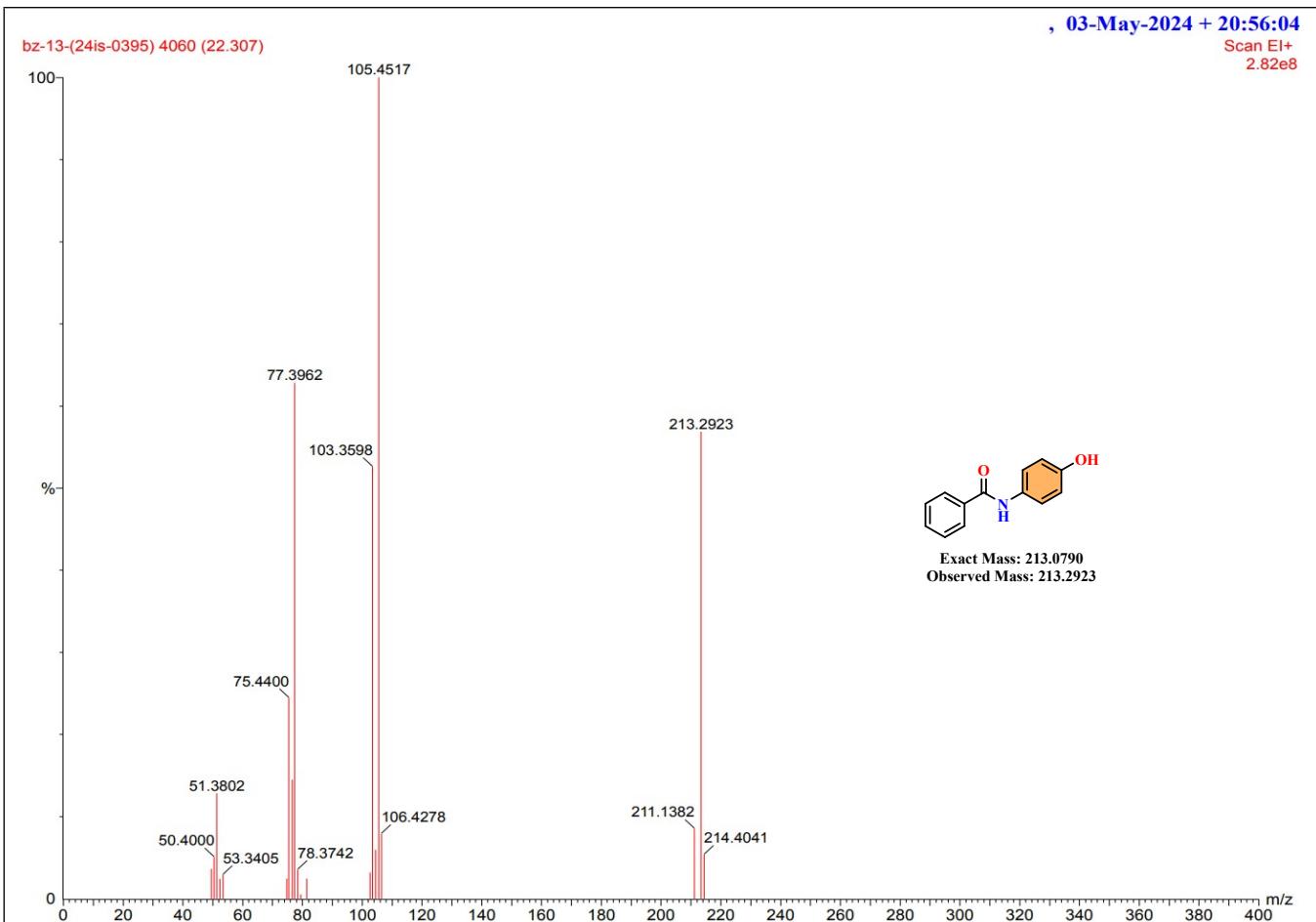


Fig S56. GC-Mass spectrum (m/z) of *N*-(4-hydroxyphenyl)benzamide (**b13**)

Signature SIF VIT VELLORE  
BZ-14-Br

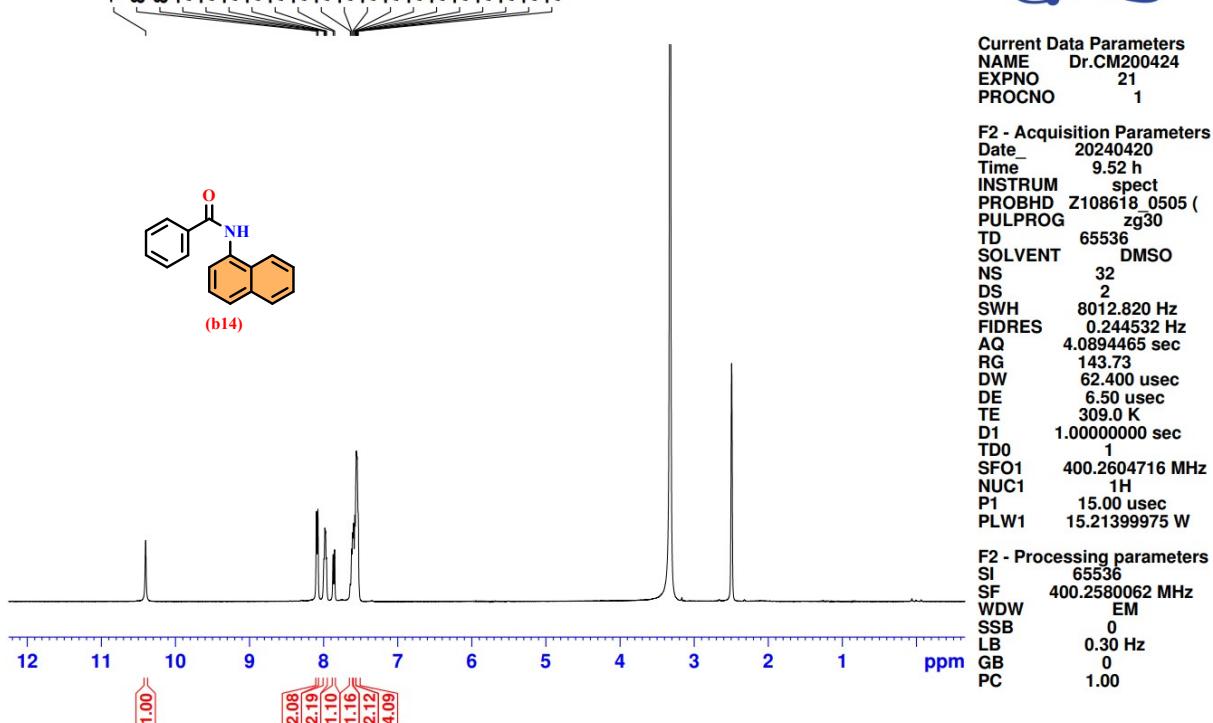


Fig S57.  $^1\text{H}$  NMR spectrum of *N*-(naphthalen-1-yl)benzamide (**b14**)

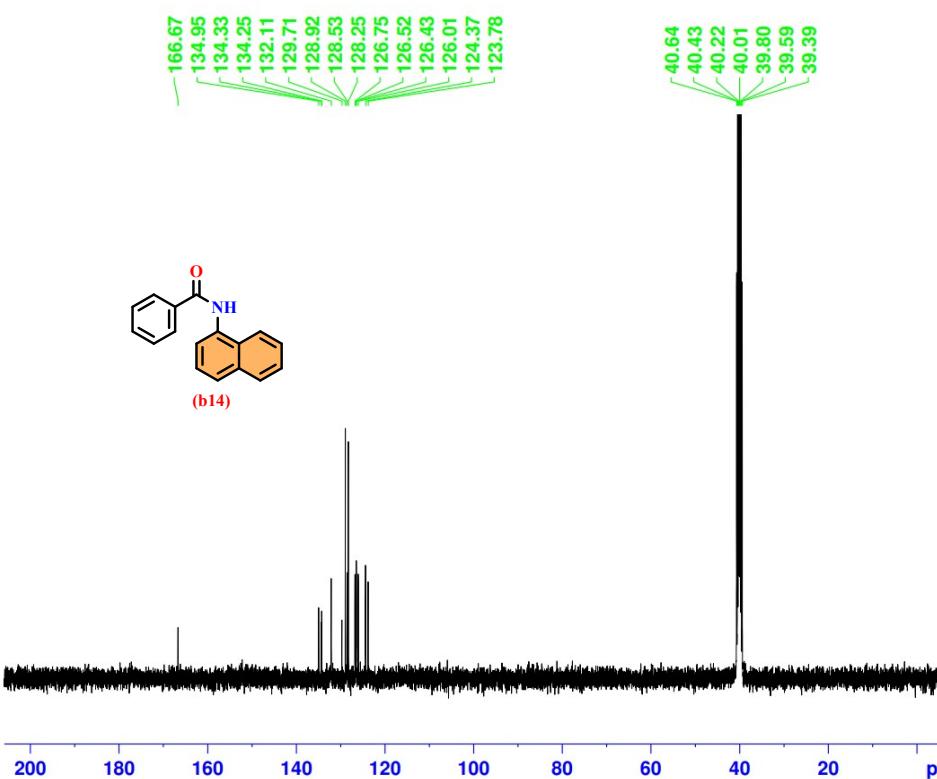
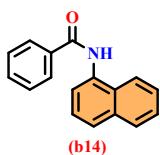
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BZ-14-Br



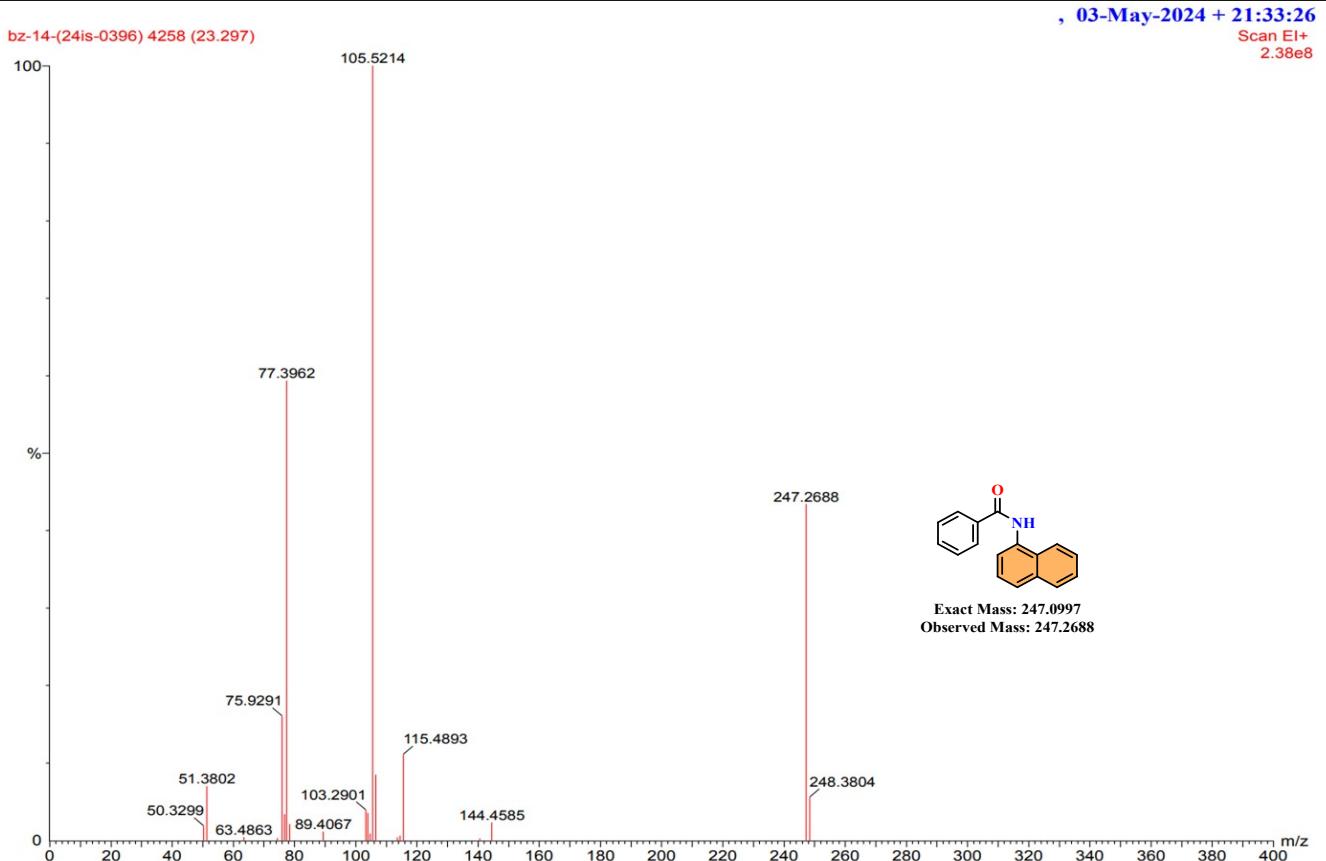
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EXPNO 39  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240424  
Time 9.05 h  
INSTRUM spect  
PROBHD Z108618\_0505 (   
PULPROG zpgpg30  
TD 65536  
SOLVENT DMSO  
NS 512  
DS 4  
SWH 24038.461 Hz  
FIDRES 0.733596 Hz  
AQ 1.3631488 sec  
RG 199.6  
DW 20.800 usec  
DE 6.50 usec  
TE 309.0 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TD0 1  
SFO1 100.6550186 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 56.49300003 W  
SFO2 400.2596010 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 90.00 usec  
PLW2 15.21399975 W  
PLW12 0.42261001 W  
PLW13 0.21257000 W

F2 - Processing parameters  
SI 32768  
SF 100.6449542 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40



**Fig S58.** <sup>13</sup>C NMR spectrum of *N*-(naphthalen-1-yl)benzamide (**b14**)



**Fig S59.** GC-Mass spectrum (m/z) of *N*-(naphthalen-1-yl)benzamide (**b14**)

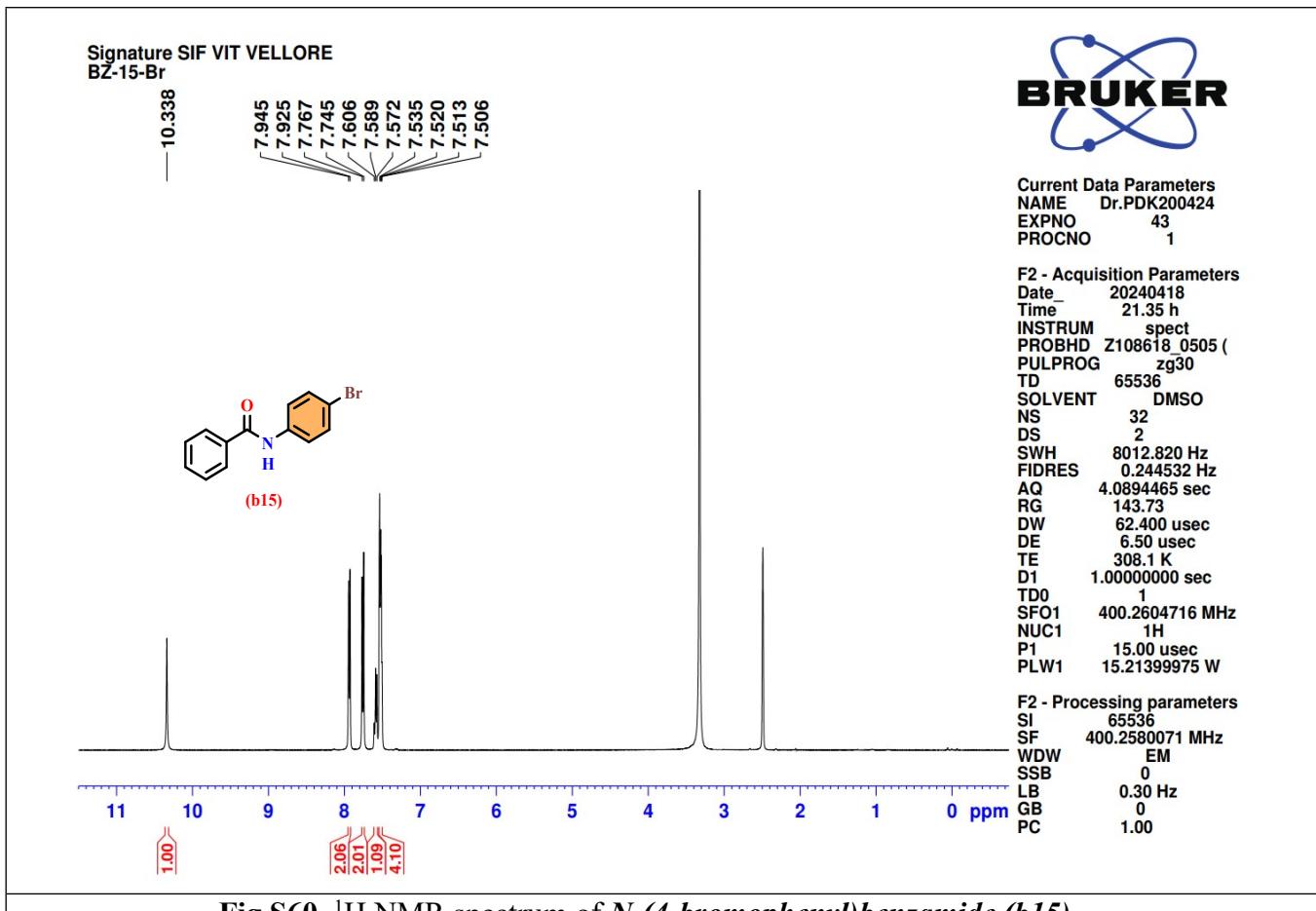


Fig S60.  $^1\text{H}$  NMR spectrum of *N*-(4-bromophenyl)benzamide (**b15**)

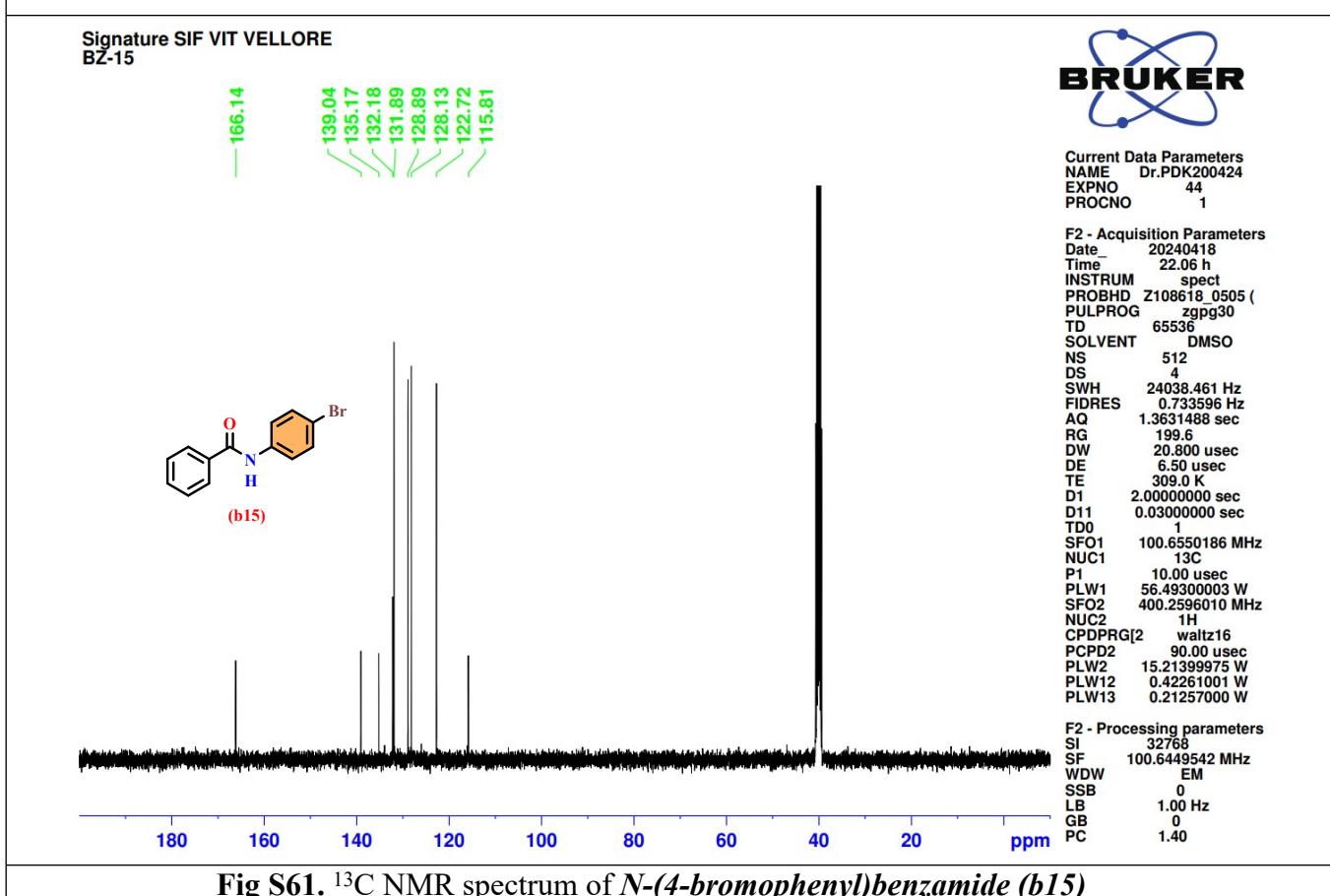
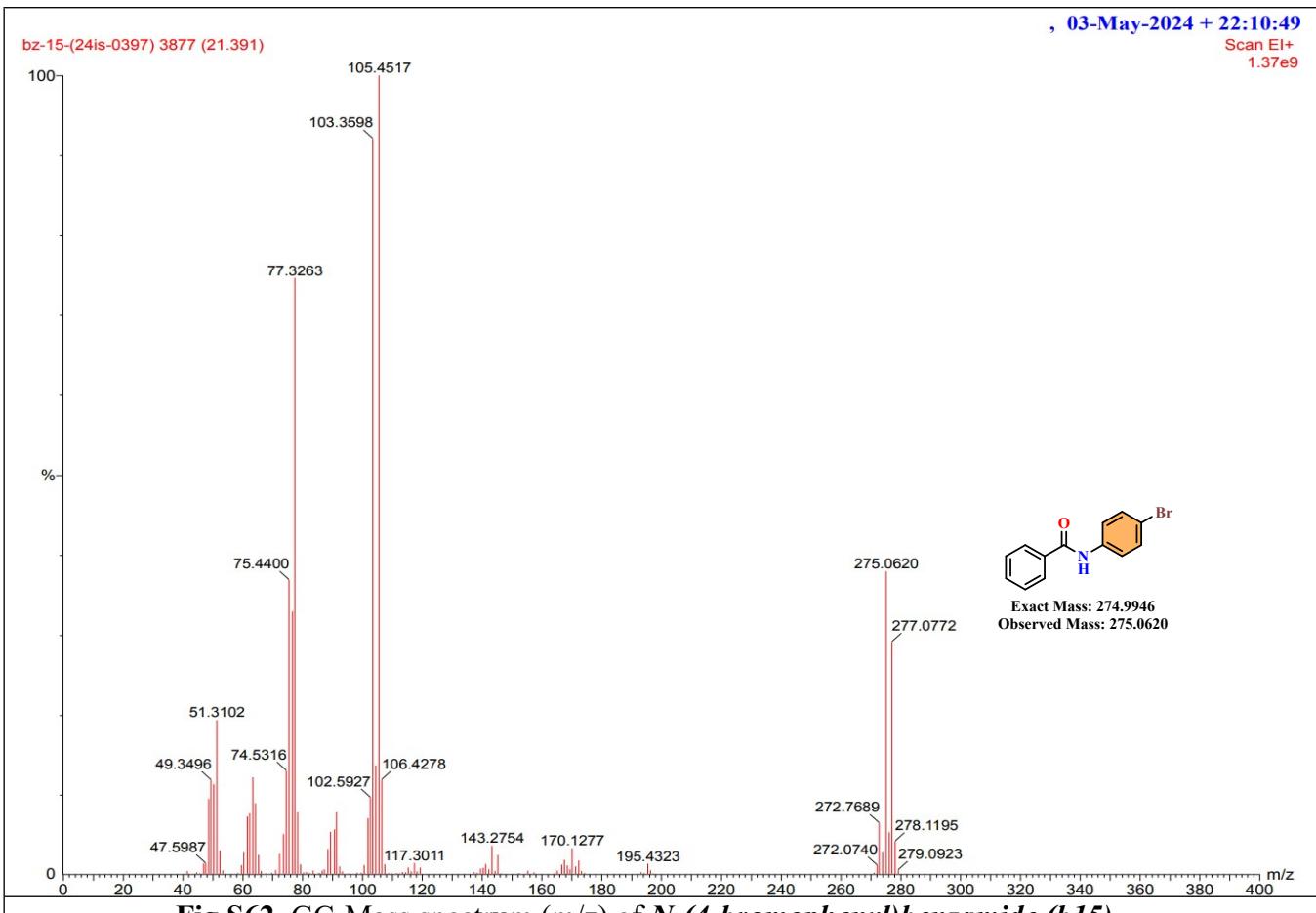
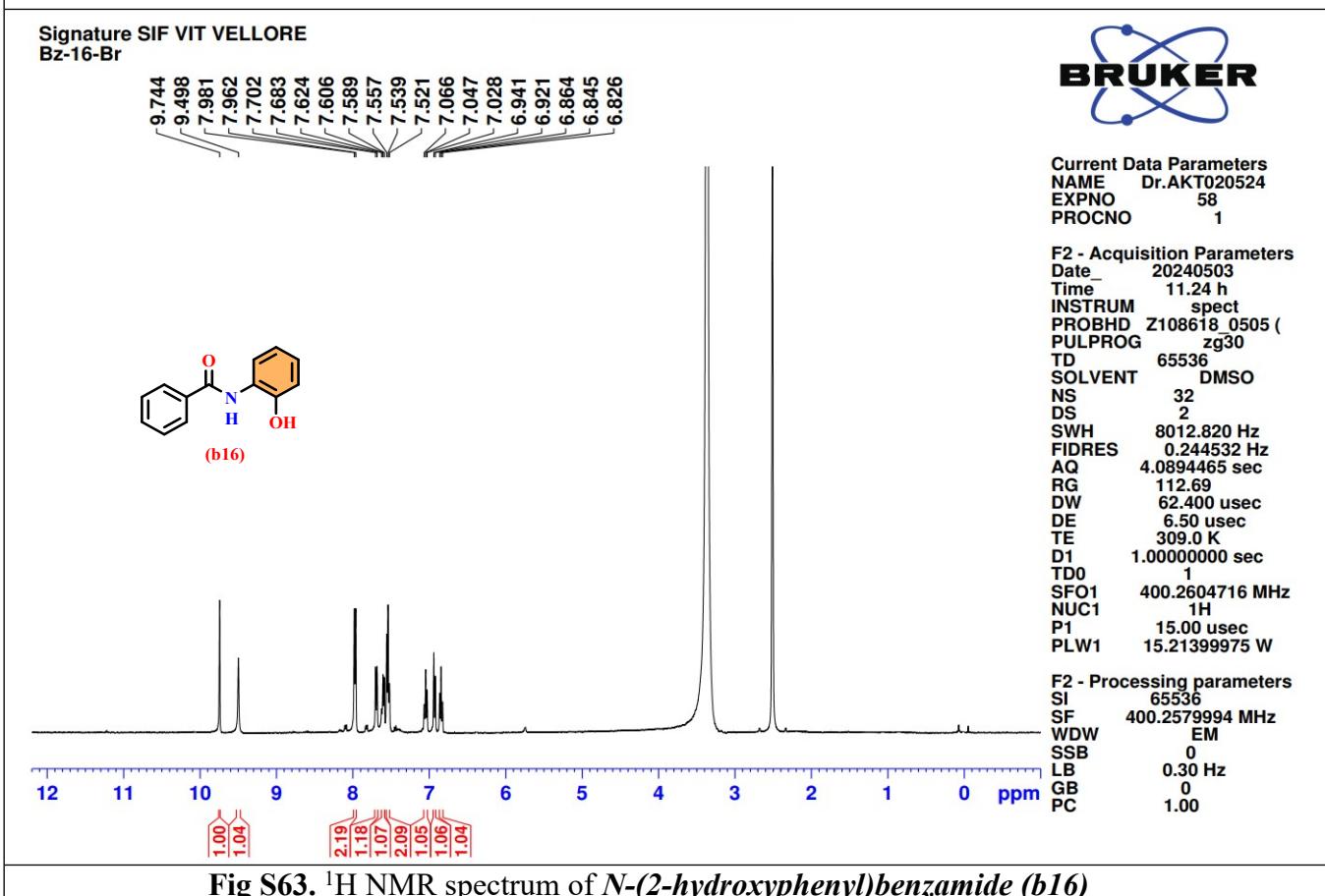


Fig S61.  $^{13}\text{C}$  NMR spectrum of *N*-(4-bromophenyl)benzamide (**b15**)



**Fig S62.** GC-Mass spectrum (m/z) of *N*-(4-bromophenyl)benzamide (**b15**)



Signature SIF VIT VELLORE  
Bz-16-Br



Current Data Parameters  
NAME Dr.AKT020524  
EXPNO 59  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240503  
Time 11.55 h  
INSTRUM spect  
PROBHD Z108618\_0505 (PULPROG zpgpg30  
TD 65536  
SOLVENT DMSO  
NS 512  
DS 4  
SWH 24038.461 Hz  
FIDRES 0.733596 Hz  
AQ 1.3631488 sec  
RG 199.6  
DW 20.800 usec  
DE 6.50 usec  
TE 309.2 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TDO 1  
SF01 100.6550186 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 56.49300000 W  
SF02 400.2596010 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 90.00 usec  
PLW2 15.21399975 W  
PLW12 0.42261001 W  
PLW13 0.21257000 W

F2 - Processing parameters  
SI 32768  
SF 100.6449542 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

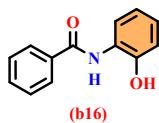


Fig S64.  $^{13}\text{C}$  NMR spectrum of *N*-(2-hydroxyphenyl)benzamide (**b16**)

bz-16-(24is-0399) 4097 (22.412)

, 03-May-2024 + 21:33:11  
Scan El+  
4.24e8

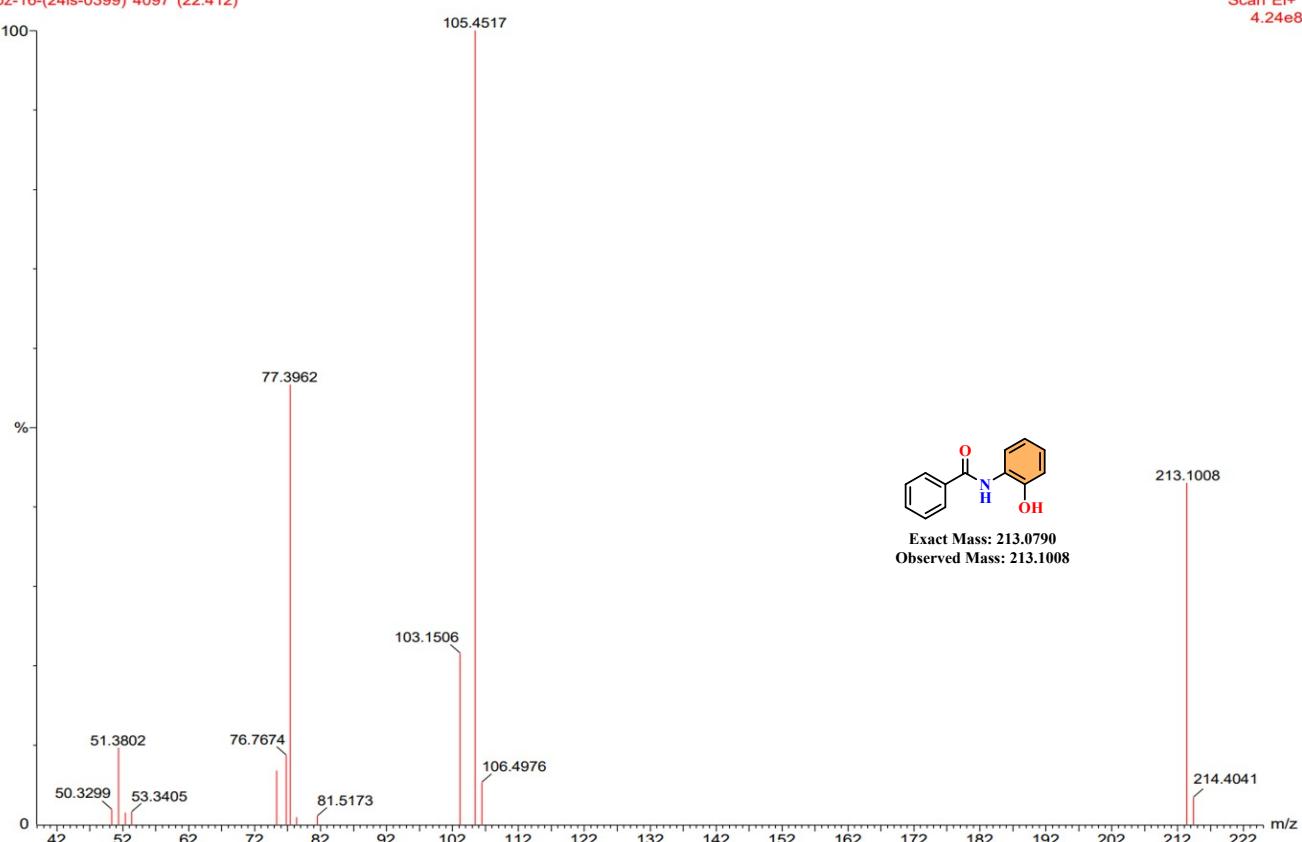
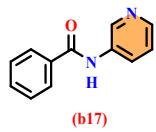


Fig S65. GC-Mass spectrum (m/z) of *N*-(2-hydroxyphenyl)benzamide (**b16**)

Signature SIF VIT VELLORE  
Bz-17-Br



10.440  
8.929  
8.313  
8.302  
8.200  
8.179  
8.176  
7.956  
7.953  
7.623  
7.606  
7.589  
7.557  
7.539  
7.519  
7.411  
7.395  
7.379



Current Data Parameters  
NAME Dr.PDK070524  
EXPNO 4  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240507  
Time 19.58 h  
INSTRUM spect  
PROBHD Z108618\_0505 (   
PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 32  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 98.85  
DW 62.400 usec  
DE 6.50 usec  
TE 308.7 K  
D1 1.0000000 sec  
TD0 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

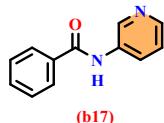
F2 - Processing parameters  
SI 65536  
SF 400.2580075 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

**Fig S66.**  $^1\text{H}$  NMR spectrum of *N*-(pyridin-3-yl)benzamide (b17)

Signature SIF VIT VELLORE  
Bz-17-Br



166.45  
144.91  
142.34  
136.33  
134.79  
132.37  
128.95  
128.19  
127.98  
124.06



Current Data Parameters  
NAME Dr.PDK070524  
EXPNO 11  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240507  
Time 20.28 h  
INSTRUM spect  
PROBHD Z108618\_0505 (   
PULPROG zgpg30  
TD 65536  
SOLVENT DMSO  
NS 512  
DS 4  
SWH 24038.461 Hz  
FIDRES 0.733596 Hz  
AQ 1.3631488 sec  
RG 199.6  
DW 20.800 usec  
DE 6.50 usec  
TE 308.7 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TD0 1  
SFO1 100.6550186 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 56.49300003 W  
SFO2 400.2596010 MHz  
NUC2 1H  
CPDPRG[2 waltz16  
PCPD2 90.00 usec  
PLW2 15.21399975 W  
PLW12 0.42261001 W  
PLW13 0.21257000 W

F2 - Processing parameters  
SI 32768  
SF 100.6449542 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

**Fig S67.**  $^{13}\text{C}$  NMR spectrum of *N*-(pyridin-3-yl)benzamide (b17)

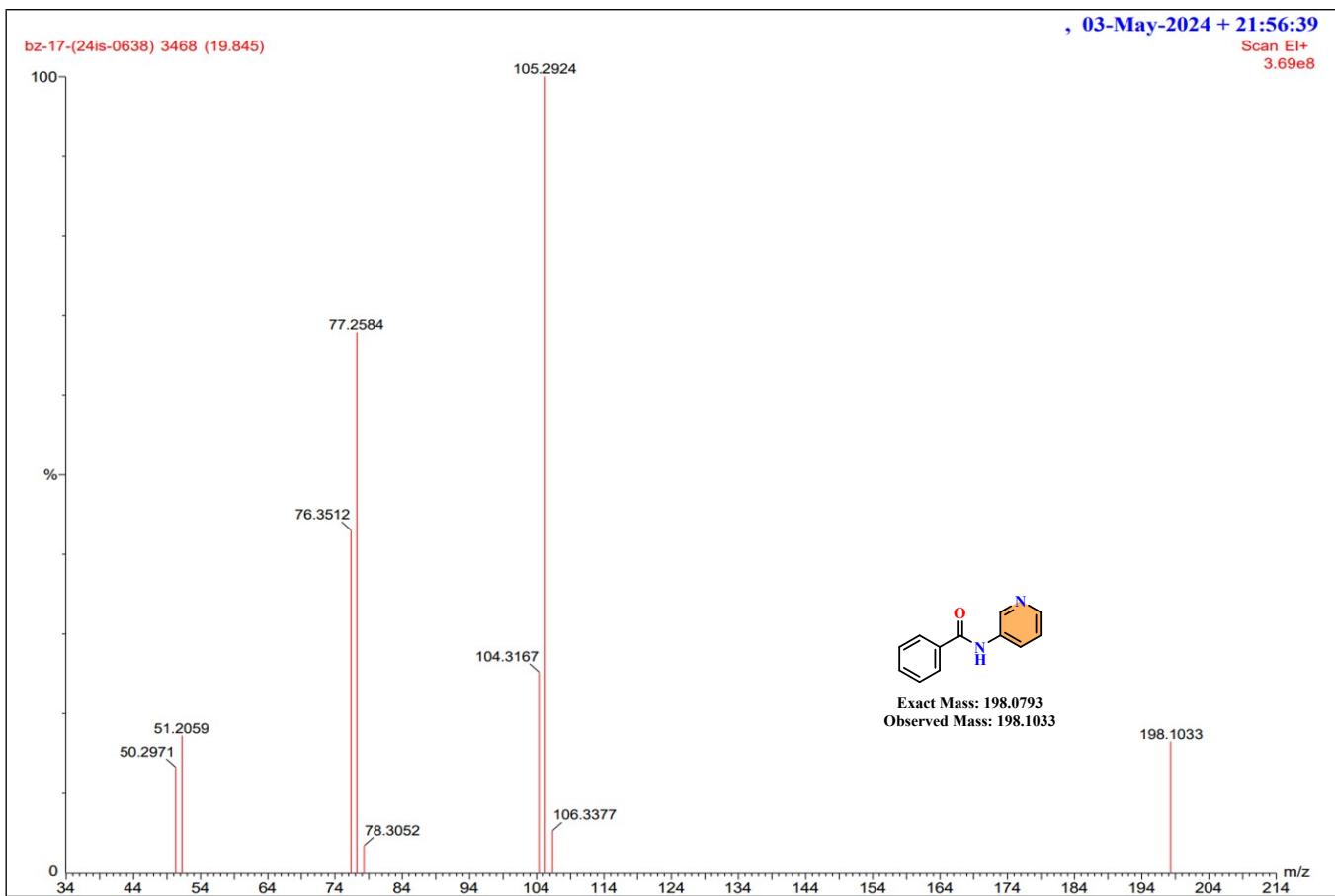


Fig S68. GC-Mass spectrum ( $m/z$ ) of *N*-(pyridin-3-yl)benzamide (b17)

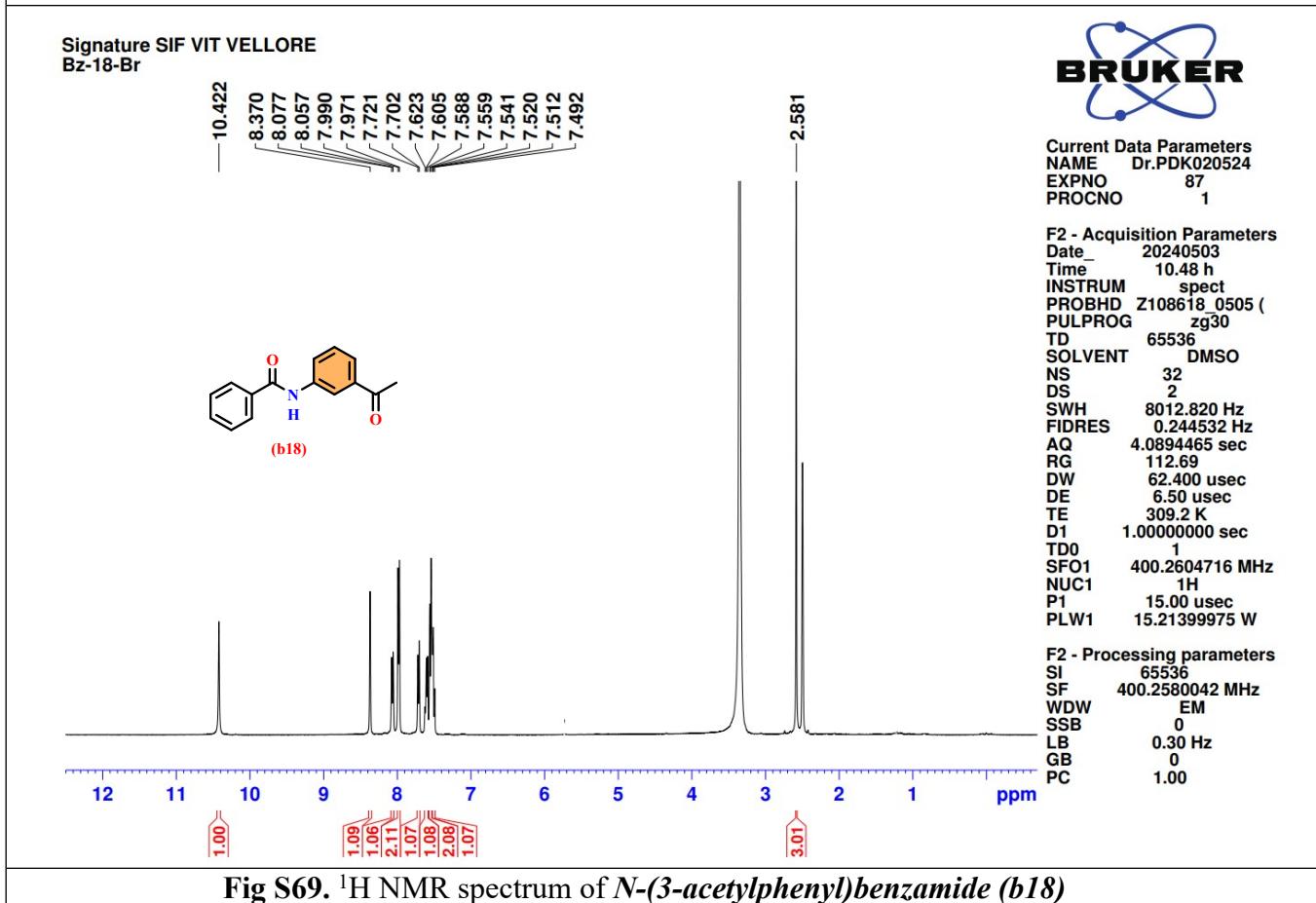


Fig S69.  $^1\text{H}$  NMR spectrum of *N*-(3-acetylphenyl)benzamide (b18)

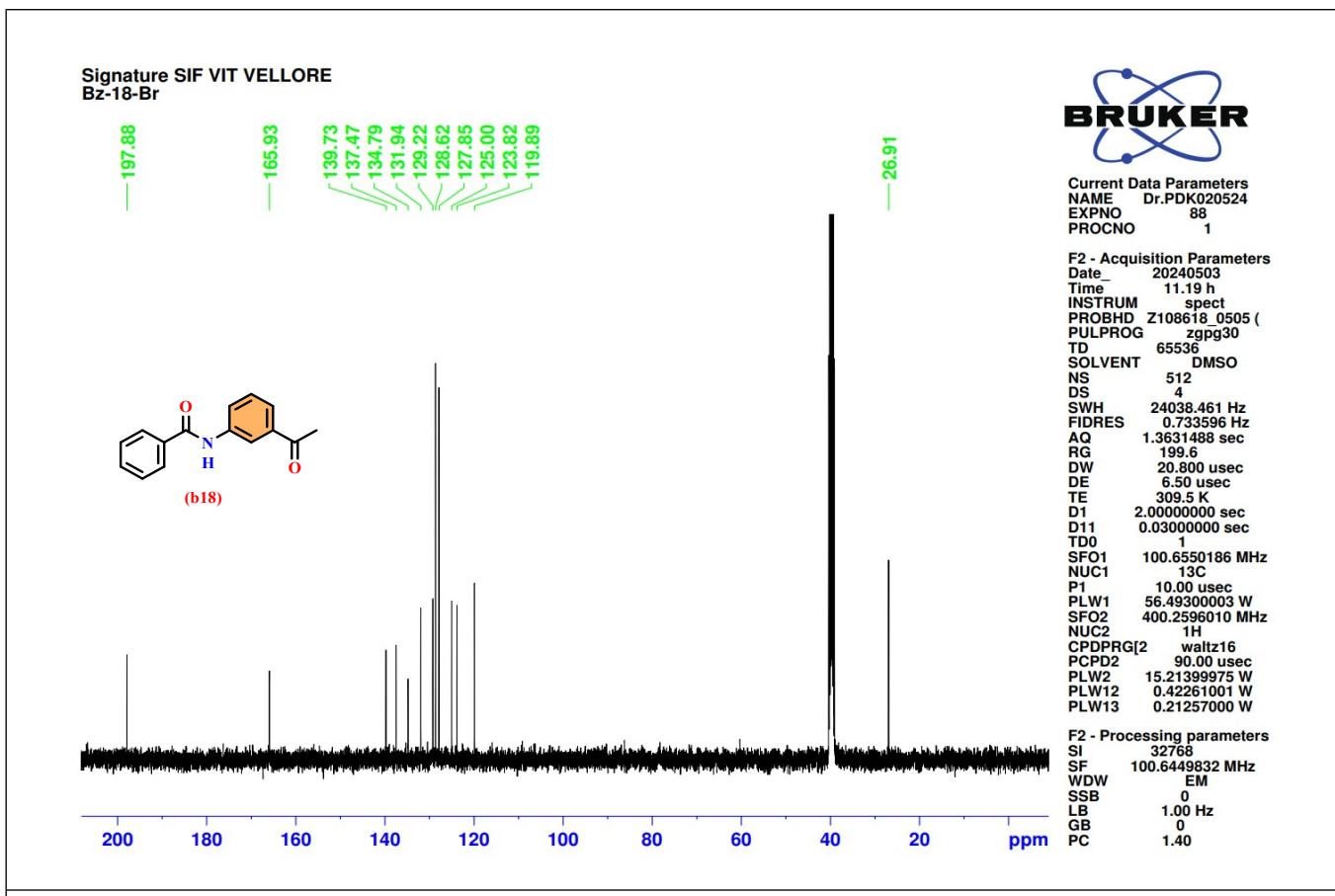


Fig S70. <sup>13</sup>C NMR spectrum of *N*-(3-acetylphenyl)benzamide (*b18*)

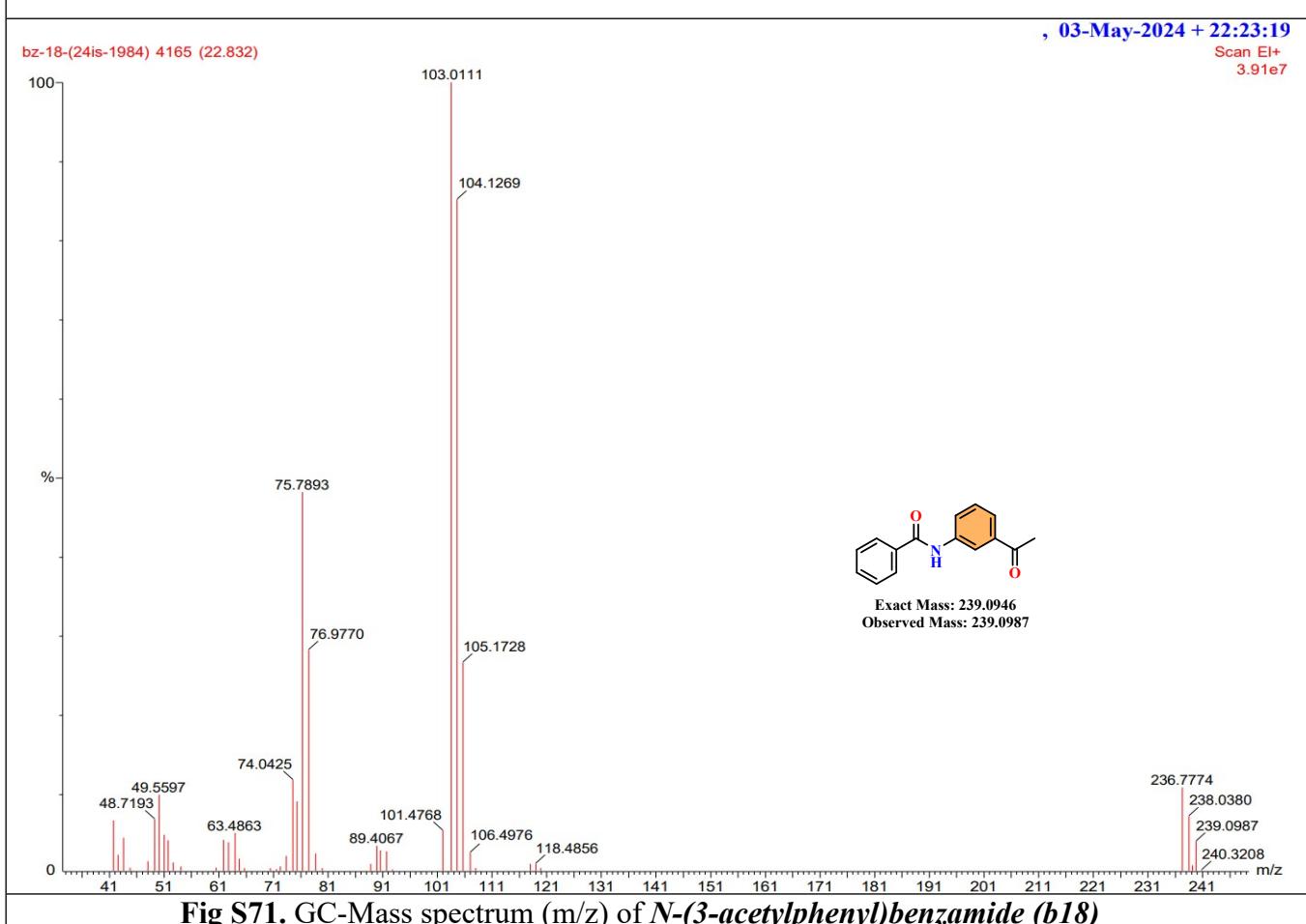


Fig S71. GC-Mass spectrum (m/z) of *N*-(3-acetylphenyl)benzamide (*b18*)

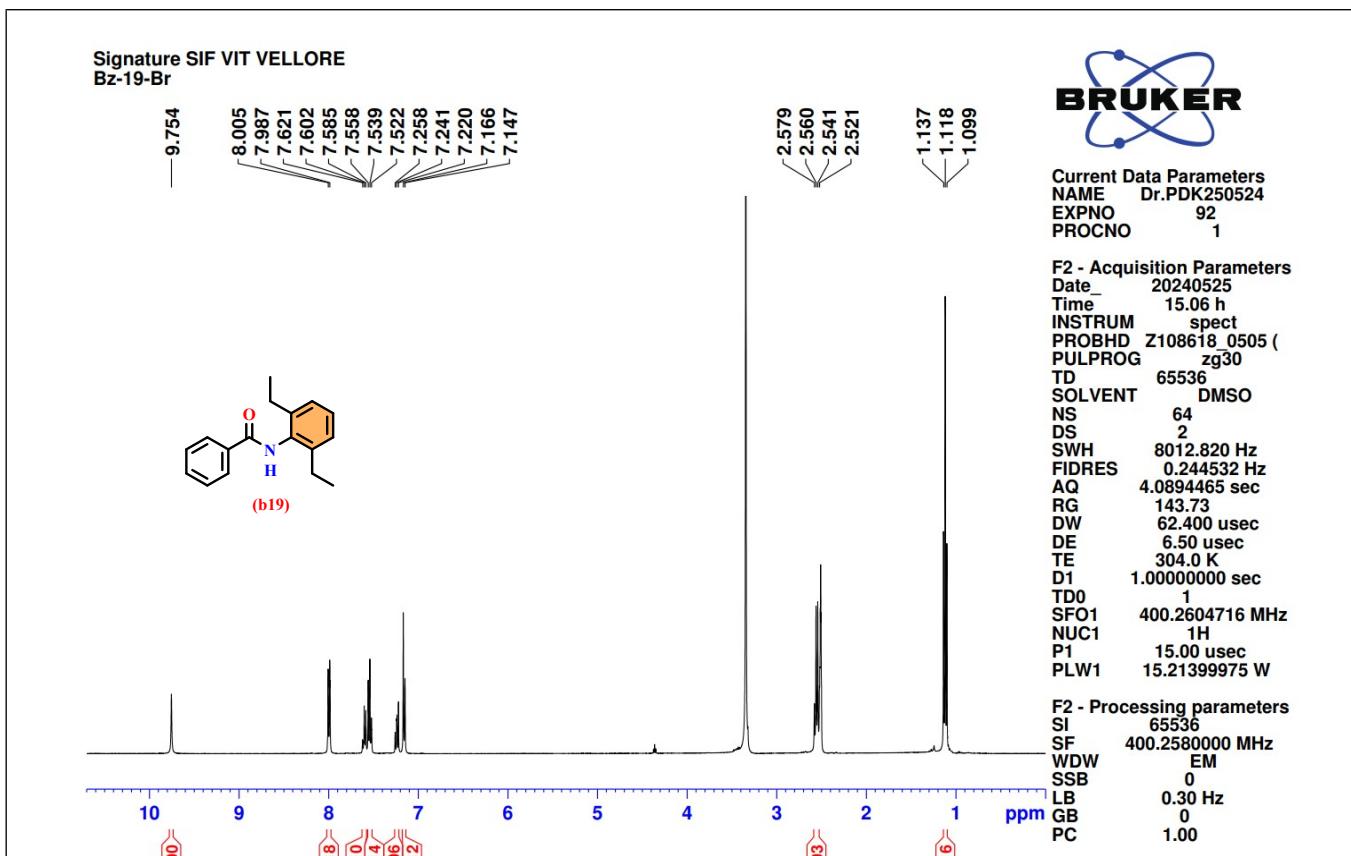


Fig S72.  $^1\text{H}$  NMR spectrum of *N*-(2,6-diethylphenyl)benzamide (**b19**)

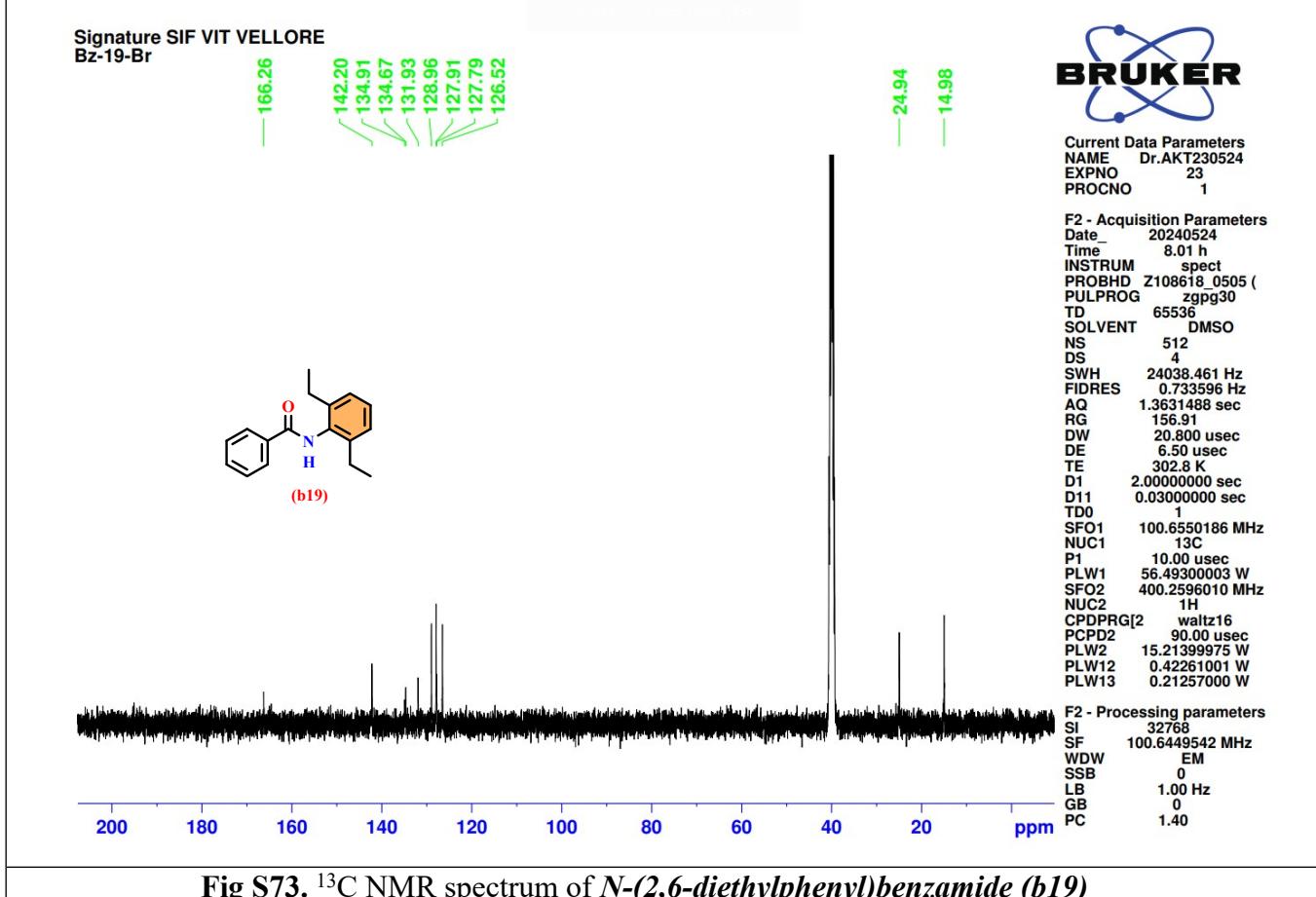
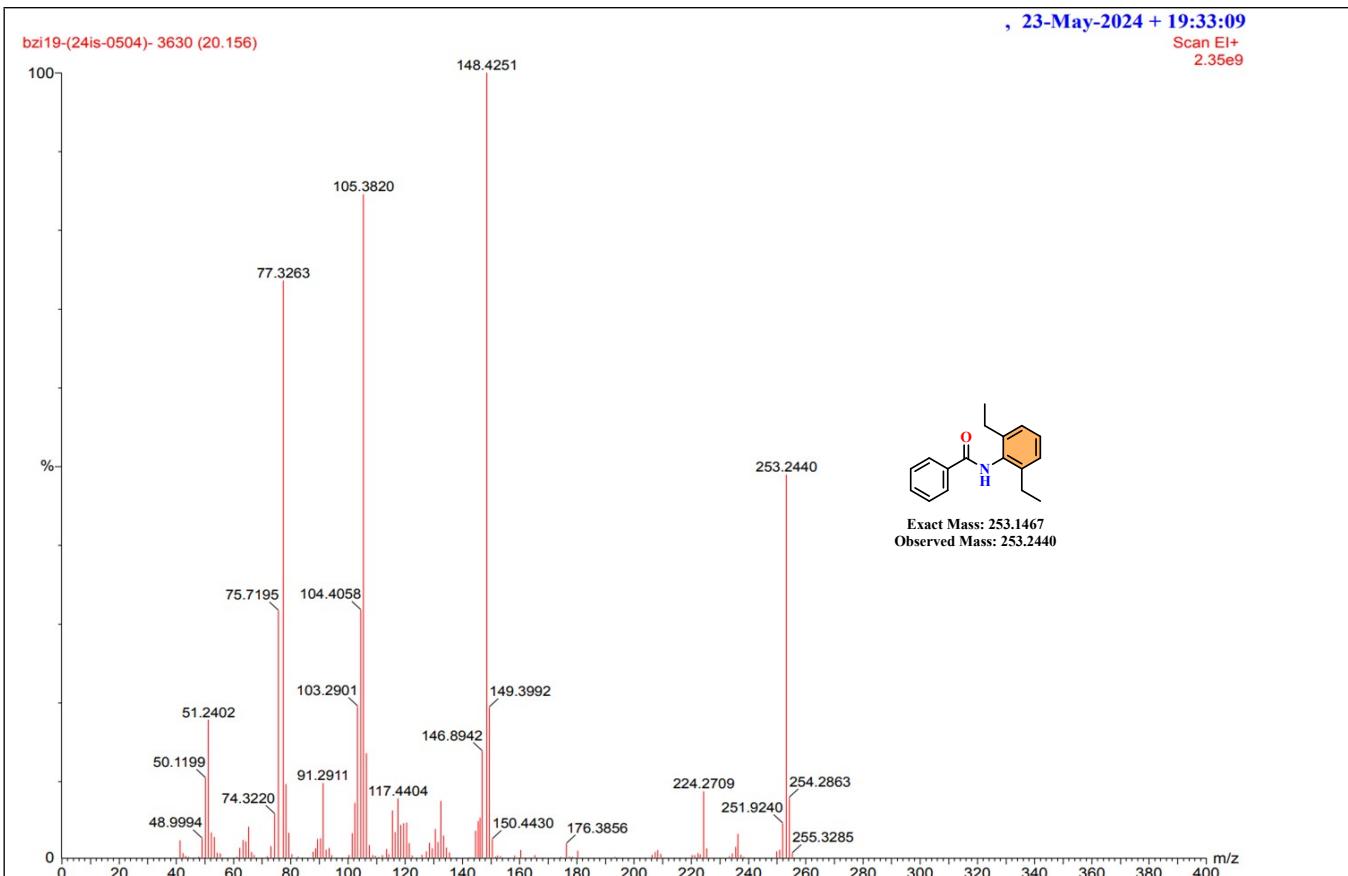
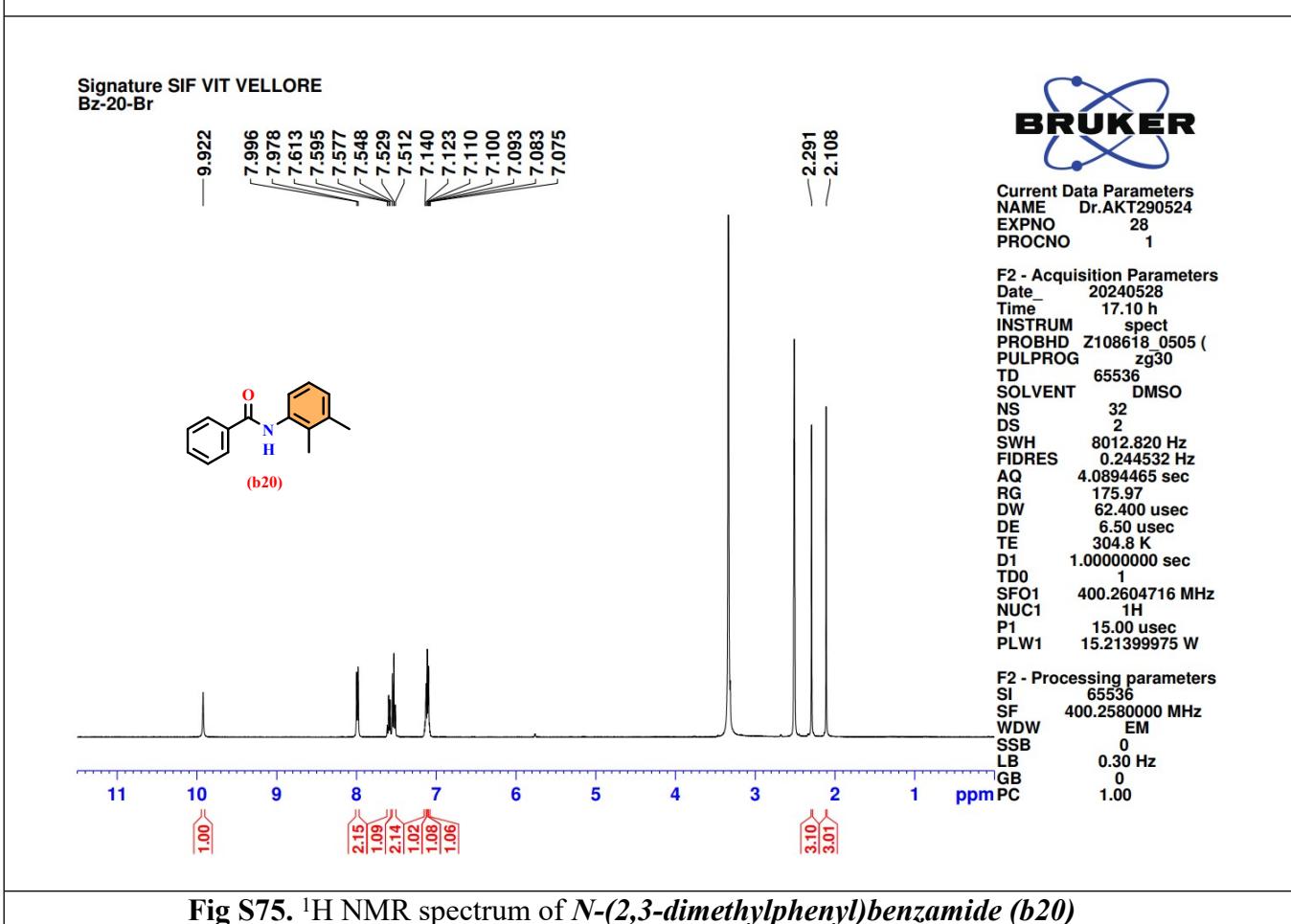


Fig S73.  $^{13}\text{C}$  NMR spectrum of *N*-(2,6-diethylphenyl)benzamide (**b19**)



**Fig S74.** GC-Mass spectrum (*m/z*) of *N*-(2,6-diethylphenyl)benzamide (**b19**)



**Fig S75.**  $^1\text{H}$  NMR spectrum of *N*-(2,3-dimethylphenyl)benzamide (**b20**)

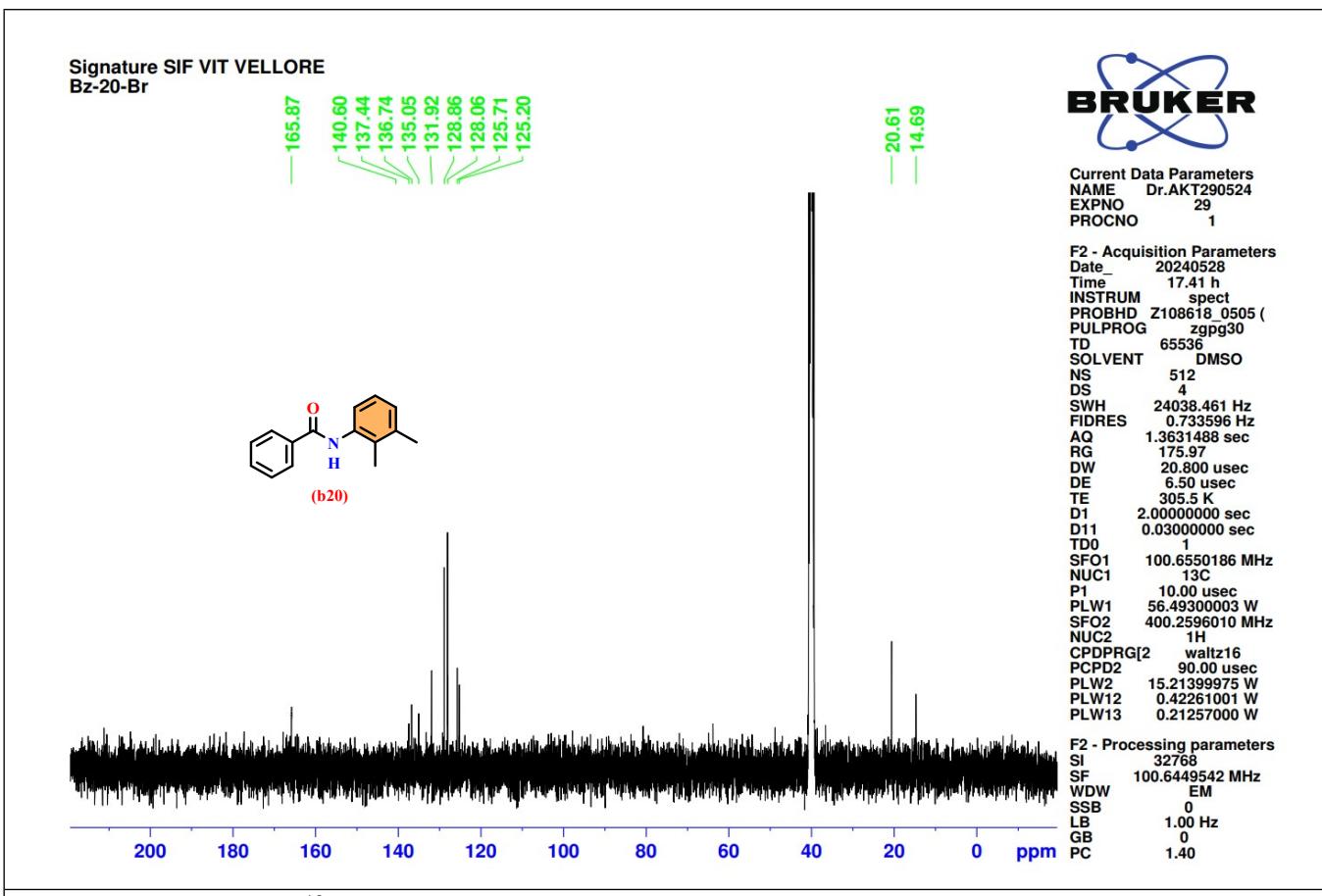


Fig S76.  $^{13}\text{C}$  NMR spectrum of *N*-(2,3-dimethylphenyl)benzamide (*b*20)

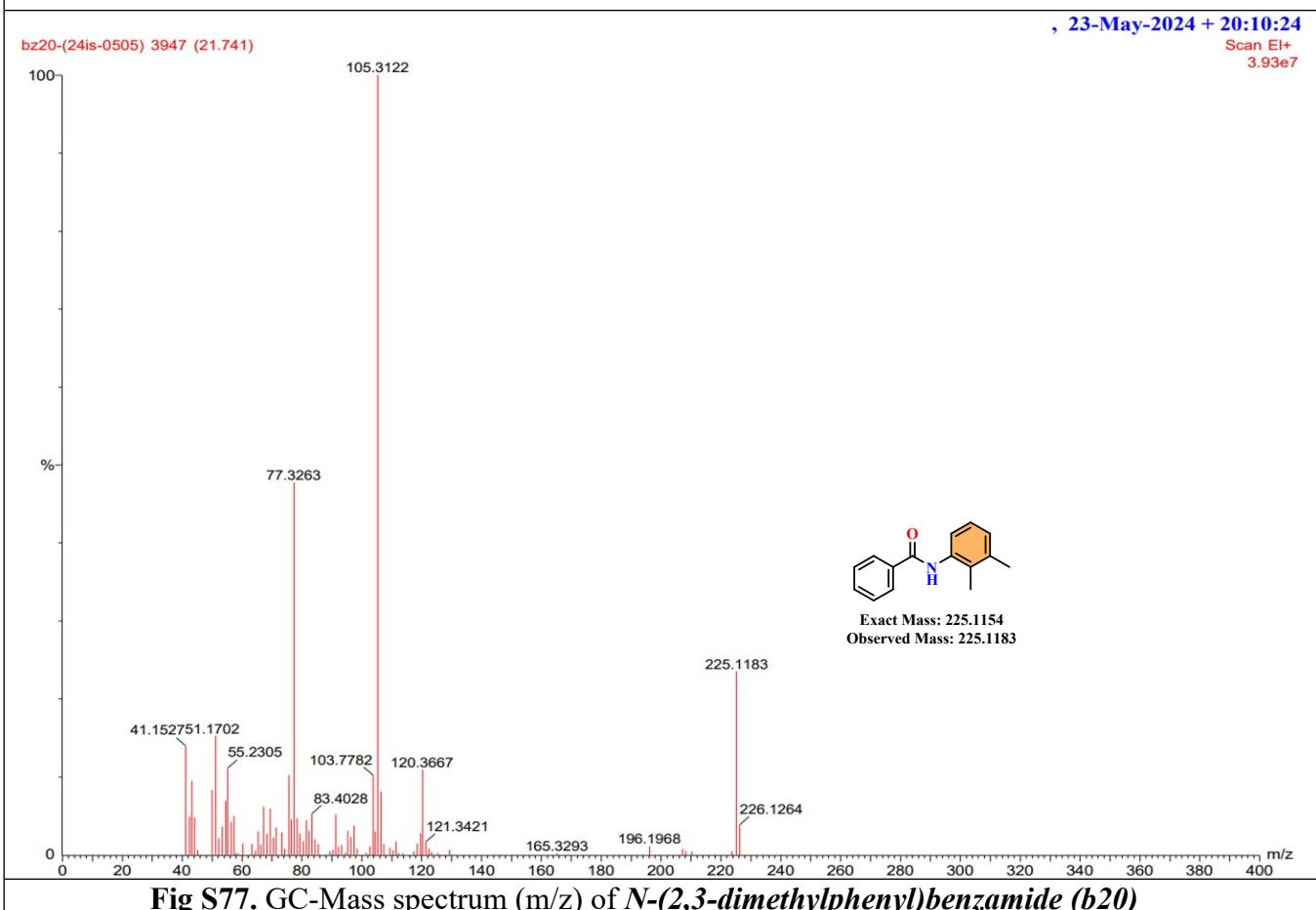


Fig S77. GC-Mass spectrum (m/z) of *N*-(2,3-dimethylphenyl)benzamide (*b*20)

Signature SIF VIT VELLORE  
Bz-21-Br



Current Data Parameters  
NAME Dr.AKT290524  
EXPNO 30  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240529  
Time 4.43 h  
INSTRUM spect  
PROBHD Z108618\_0505 (PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 32  
DS 2  
SWH 8012.80 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 175.97  
DW 62.400 usec  
DE 6.50 usec  
TE 302.0 K  
D1 1.0000000 sec  
TDO 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

F2 - Processing parameters  
SI 65536  
SF 400.2580000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

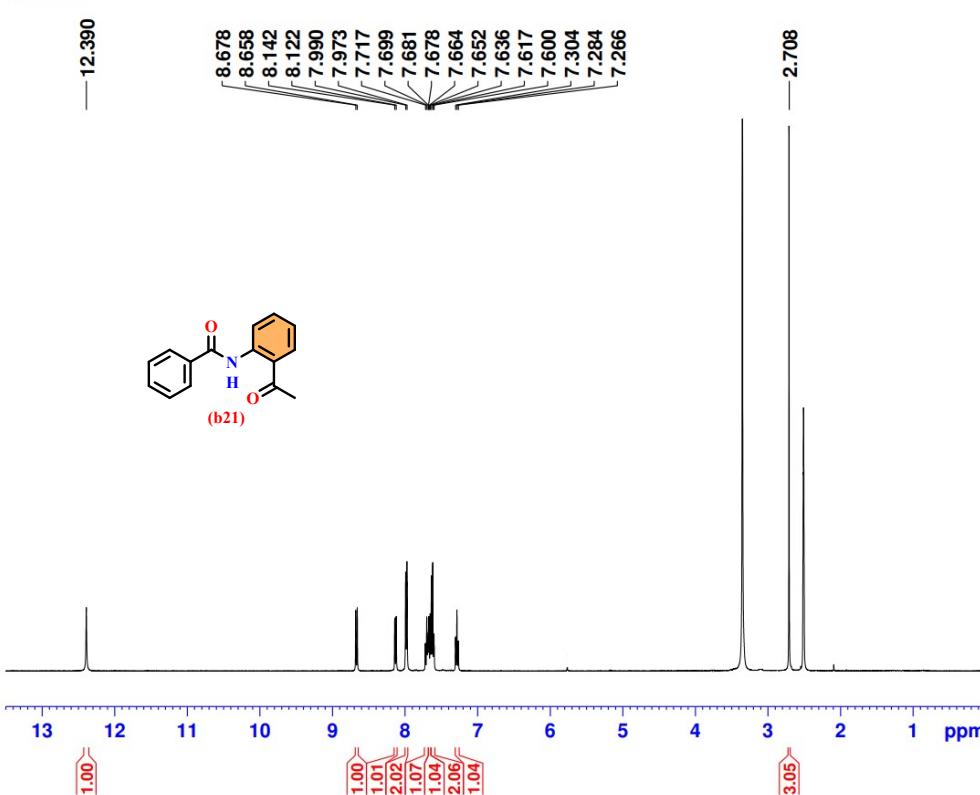


Fig S78.  $^1\text{H}$  NMR spectrum of *N*-(2-acetylphenyl)benzamide (b21)

Signature SIF VIT VELLORE  
Bz-21-Br



Current Data Parameters  
NAME Dr.AKT290524  
EXPNO 31  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240529  
Time 5.14 h  
INSTRUM spect  
PROBHD Z108618\_0505 (PULPROG zgpg30  
TD 65536  
SOLVENT DMSO  
NS 512  
DS 4  
SWH 24038.461 Hz  
FIDRES 0.733596 Hz  
AQ 1.3631488 sec  
RG 156.91  
DW 20.800 usec  
DE 6.50 usec  
TE 302.6 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TDO 1  
SFO1 100.6550186 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 56.49300003 W  
SF02 400.2596010 MHz  
NUC2 1H  
CPDPG[2 waltz16  
PCPD2 90.00 usec  
PLW2 15.21399975 W  
PLW12 0.42261001 W  
PLW13 0.21257000 W

F2 - Processing parameters  
SI 32768  
SF 100.6449542 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

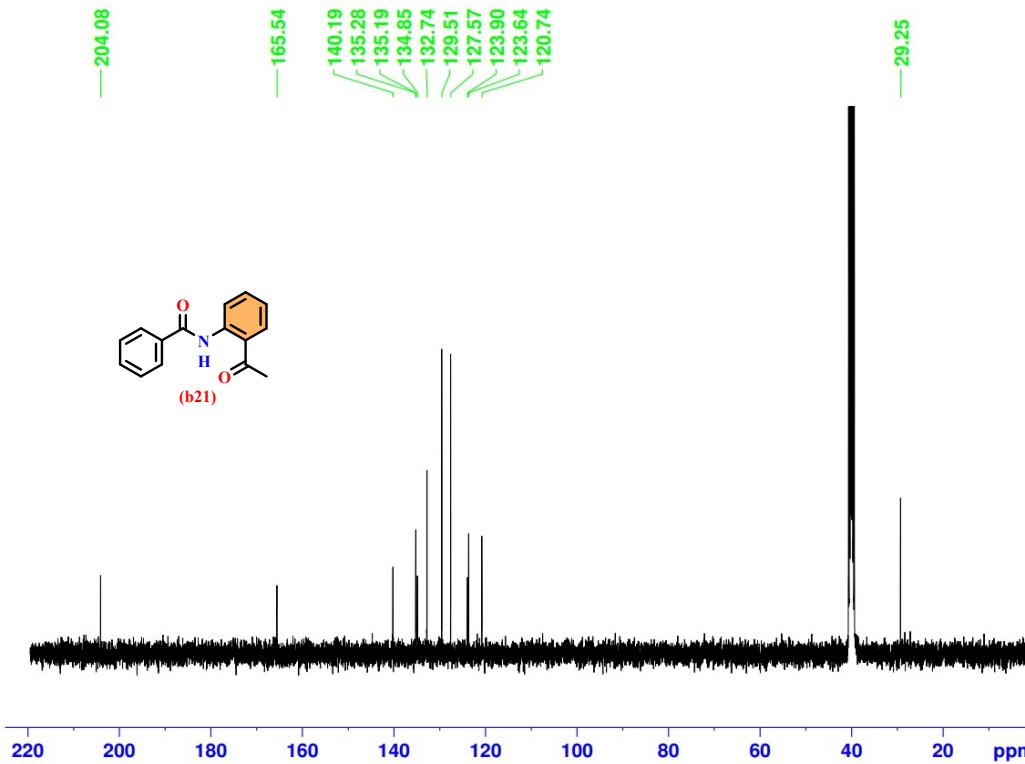
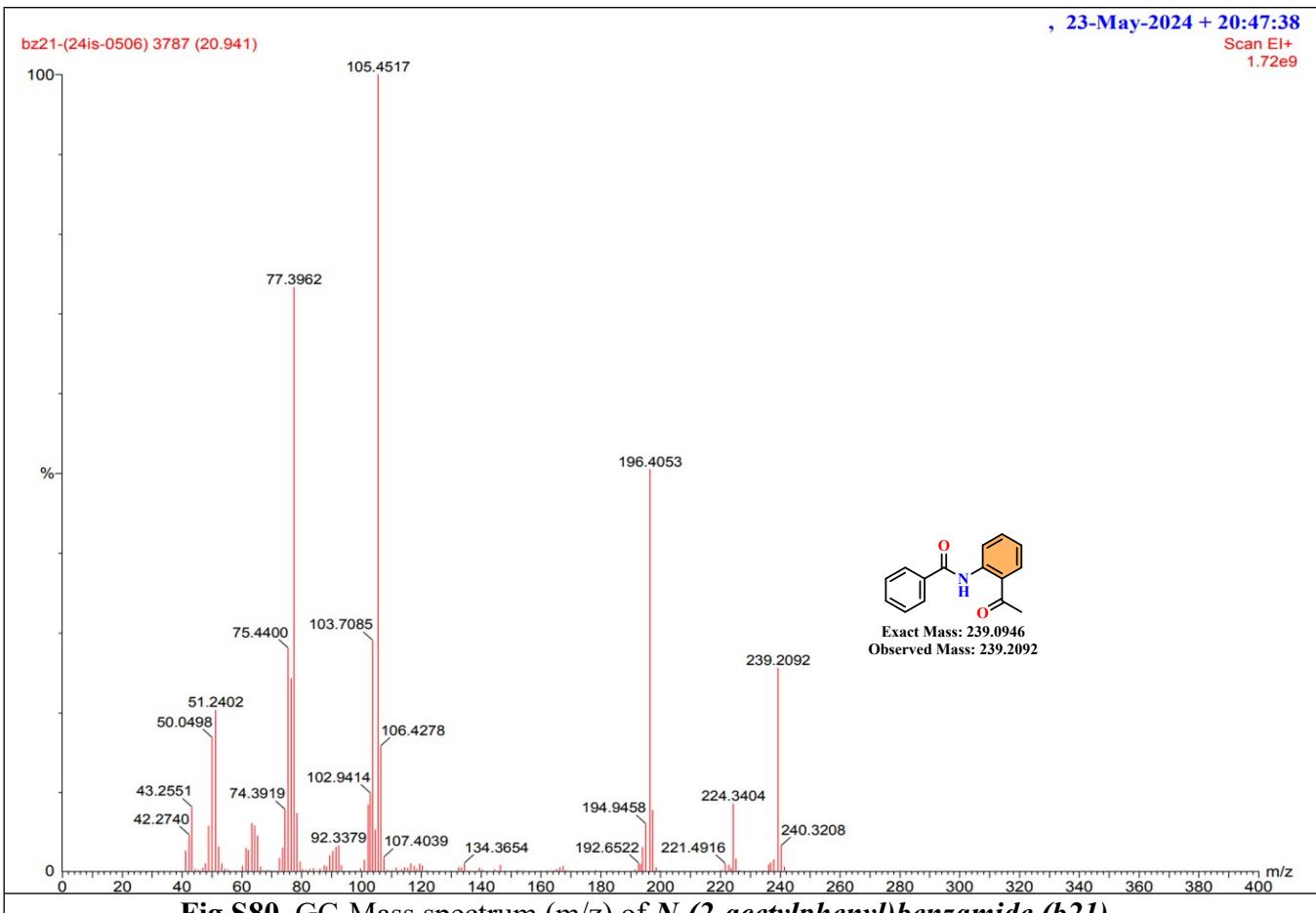
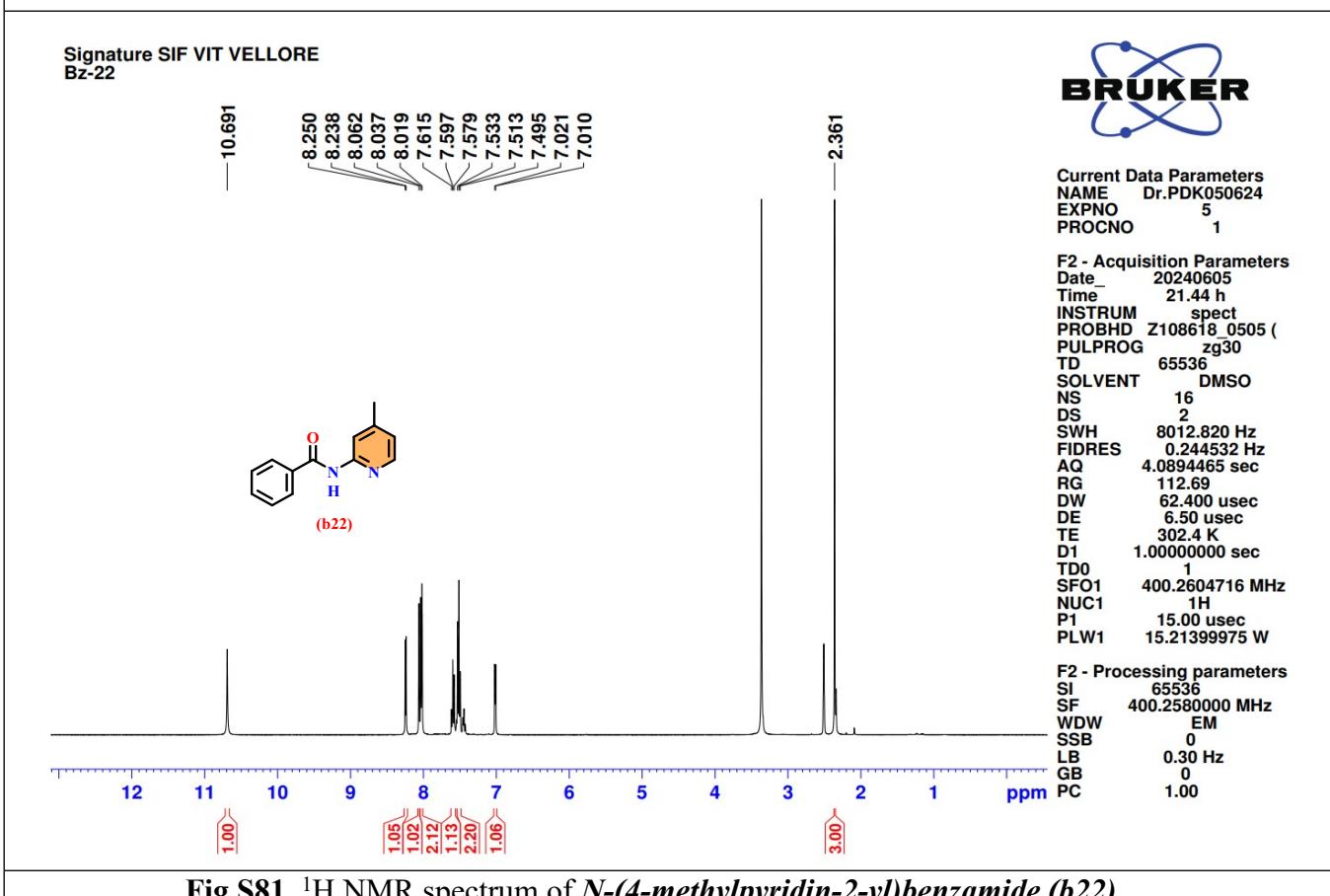


Fig S79.  $^{13}\text{C}$  NMR spectrum of *N*-(2-acetylphenyl)benzamide (b21)



**Fig S80.** GC-Mass spectrum ( $m/z$ ) of *N*-(2-acetylphenyl)benzamide (**b21**)



**Fig S81.**  $^1\text{H}$  NMR spectrum of *N*-(4-methylpyridin-2-yl)benzamide (**b22**)

Current Data Parameters  
NAME Dr.PDK050624  
EXPNO 6  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20240605  
Time 22.15 h  
INSTRUM spect  
PROBHD Z108618\_0505 (PULPROG zpg30  
TD 65536  
SOLVENT DMSO  
NS 512  
DS 4  
SWH 24038.461 Hz  
FIDRES 0.733596 Hz  
AQ 1.3631488 sec  
RG 156.91  
DW 20.800 usec  
DE 6.50 usec  
TE 302.8 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TD0 1  
SFO1 100.6550186 MHz  
NUC1 <sup>13</sup>C  
P1 10.00 usec  
PLW1 56.49300003 W  
SFO2 400.2596010 MHz  
NUC2 <sup>1</sup>H  
CPDPG[2 waltz16  
PCPD2 90.00 usec  
PLW2 15.21399975 W  
PLW12 0.42261001 W  
PLW13 0.21257000 W

F2 - Processing parameters  
SI 32768  
SF 100.6449542 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

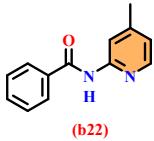


Fig S82. <sup>13</sup>C NMR spectrum of *N*-(4-methylpyridin-2-yl)benzamide (b22)

Bz-22-(24ls-0703) 3606 (20.036)

, 15-Jul-2024 + 22:16:00

Scan El+  
1.22e9

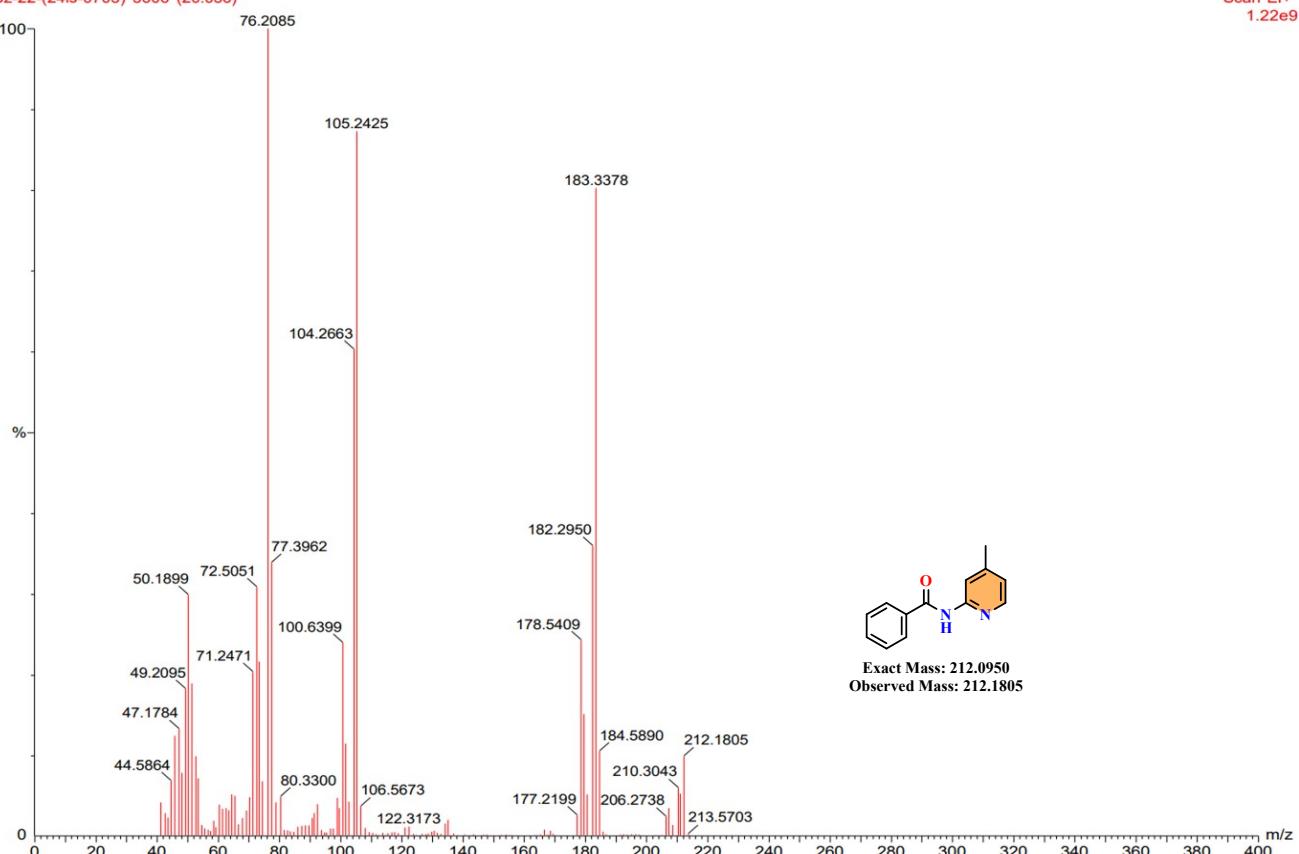


Fig S83. GC-Mass spectrum (m/z) of *N*-(4-methylpyridin-2-yl)benzamide (b22)

Signature SIF VIT VELLORE  
Bz-23



Current Data Parameters  
NAME Dr.PDK150724  
EXPNO 25  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240716  
Time 14.04 h  
INSTRUM spect  
PROBHD Z108618\_0505 (PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 32  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 127.79  
DW 62.400 usec  
DE 6.50 usec  
TE 303.1 K  
D1 1.0000000 sec  
TD0 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

F2 - Processing parameters  
SI 65536  
SF 400.2580073 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

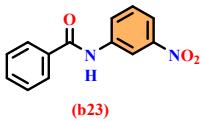


Fig S84.  $^1\text{H}$  NMR spectrum of *N*-(3-nitrophenyl)benzamide (**b23**)

Signature SIF VIT VELLORE  
Bz-23



Current Data Parameters  
NAME Dr.PDK150724  
EXPNO 26  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240716  
Time 14.36 h  
INSTRUM spect  
PROBHD Z108618\_0505 (PULPROG zgpg30  
TD 65536  
SOLVENT DMSO  
NS 512  
DS 4  
SWH 24038.461 Hz  
FIDRES 0.733596 Hz  
AQ 1.3631488 sec  
RG 175.97  
DW 20.800 usec  
DE 6.50 usec  
TE 303.8 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TD0 1  
SFO1 100.6550186 MHz  
NUC1  $^{13}\text{C}$   
P1 10.00 usec  
PLW1 56.49300003 W  
SFO2 400.2596010 MHz  
NUC2  $^1\text{H}$   
CPDPRG[2 waltz16  
PCPD2 90.00 usec  
PLW2 15.21399975 W  
PLW12 0.42261001 W  
PLW13 0.21257000 W

F2 - Processing parameters  
SI 32768  
SF 100.6449542 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

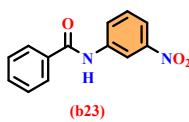
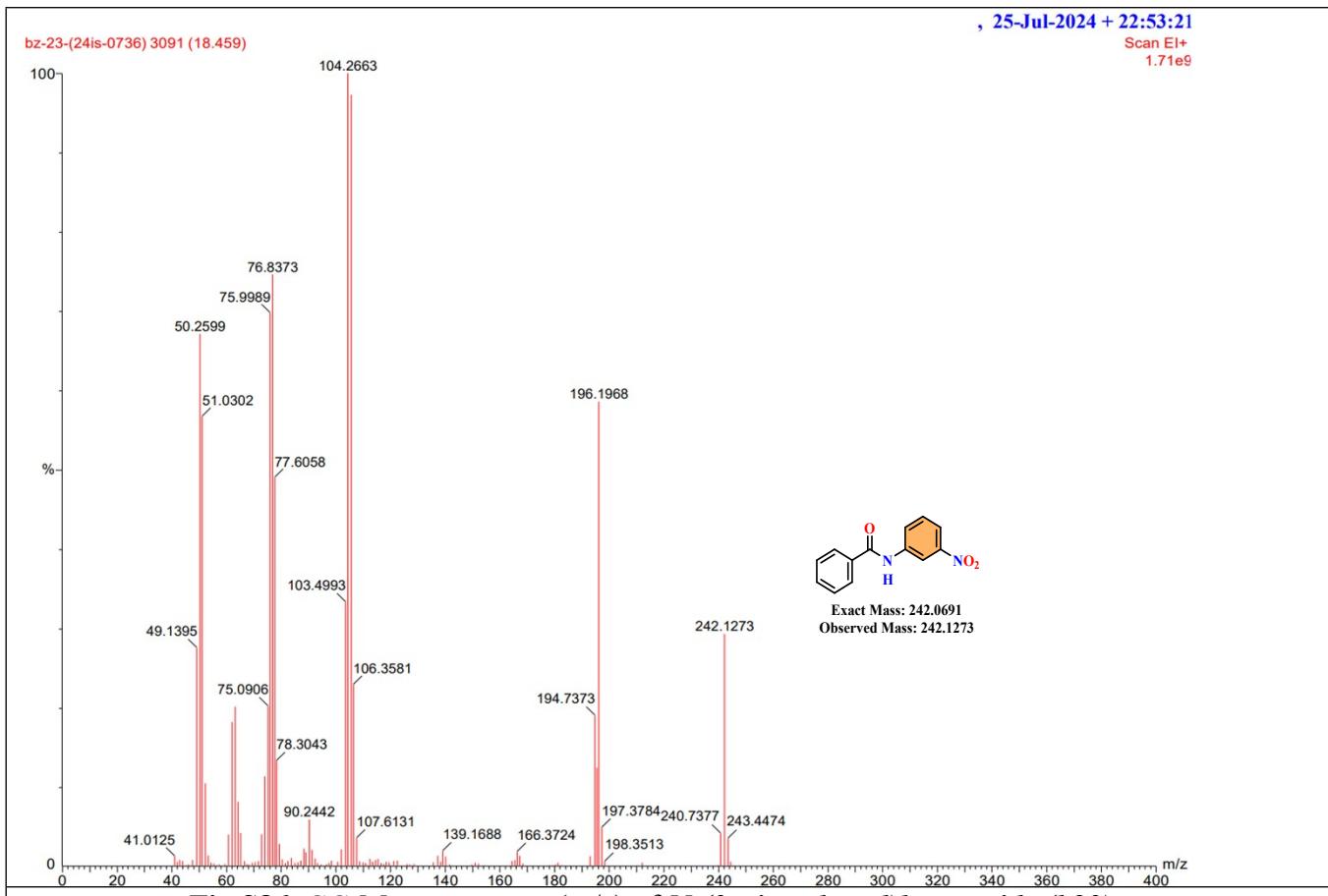
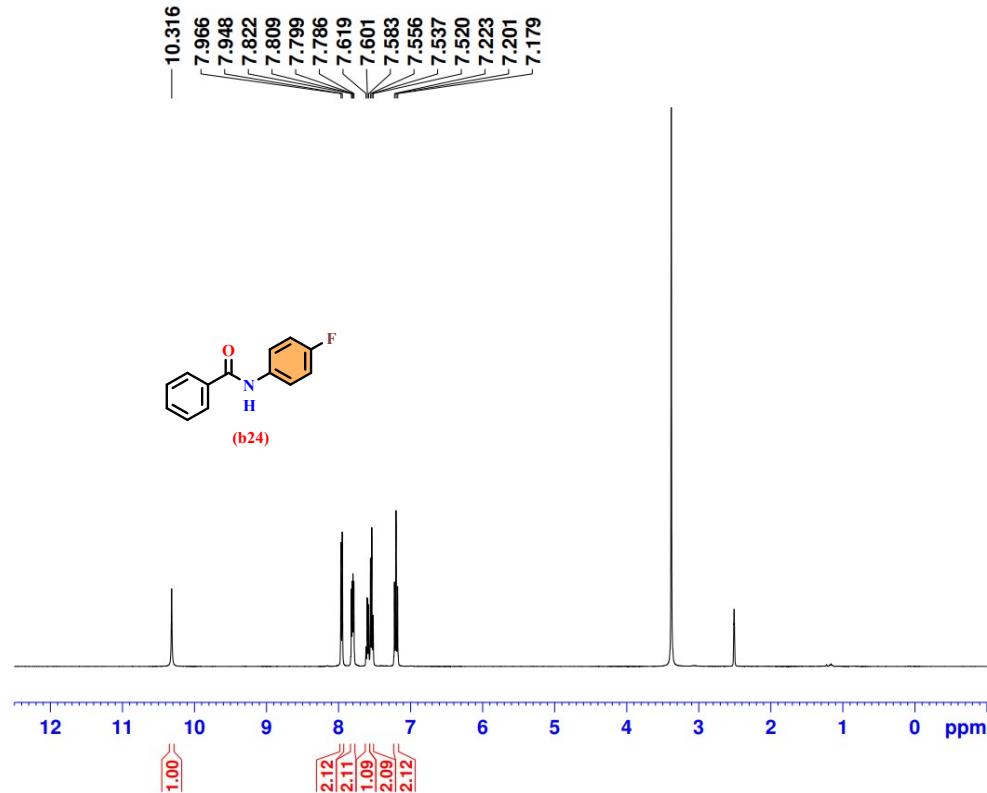


Fig S85.  $^{13}\text{C}$  NMR spectrum of *N*-(3-nitrophenyl)benzamide (**b23**)



**Fig S86.** GC-Mass spectrum (m/z) of *N*-(3-nitrophenyl)benzamide (b23)

Signature SIF VIT VELLORE  
Bz-24F



**Fig S87.**  $^1\text{H}$  NMR spectrum of *N*-(4-fluorophenyl)benzamide (b24)

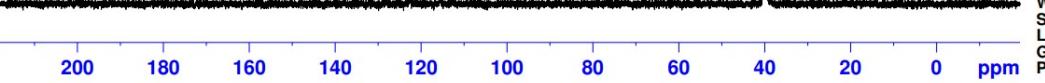
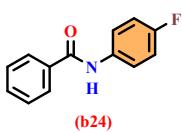
Signature SIF VIT VELLORE  
Bz-24



Current Data Parameters  
NAME Dr.PDK090724  
EXPNO 15  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240709  
Time 20.17 h  
INSTRUM spect  
PROBHD Z108618\_0505 (PULPROG zgpg30  
TD 65536  
SOLVENT DMSO  
NS 512  
DS 4  
SWH 24038.461 Hz  
FIDRES 0.733596 Hz  
AQ 1.3631488 sec  
RG 199.6  
DW 20.800 usec  
DE 6.50 usec  
TE 304.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TD0 1  
SFO1 100.6550186 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 56.49300003 W  
SFO2 400.2596010 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 90.00 usec  
PLW2 15.21399975 W  
PLW12 0.42261001 W  
PLW13 0.21257000 W

F2 - Processing parameters  
SI 32768  
SF 100.6449542 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40



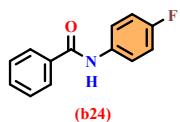
Signature SIF VIT VELLORE  
Bz-24F

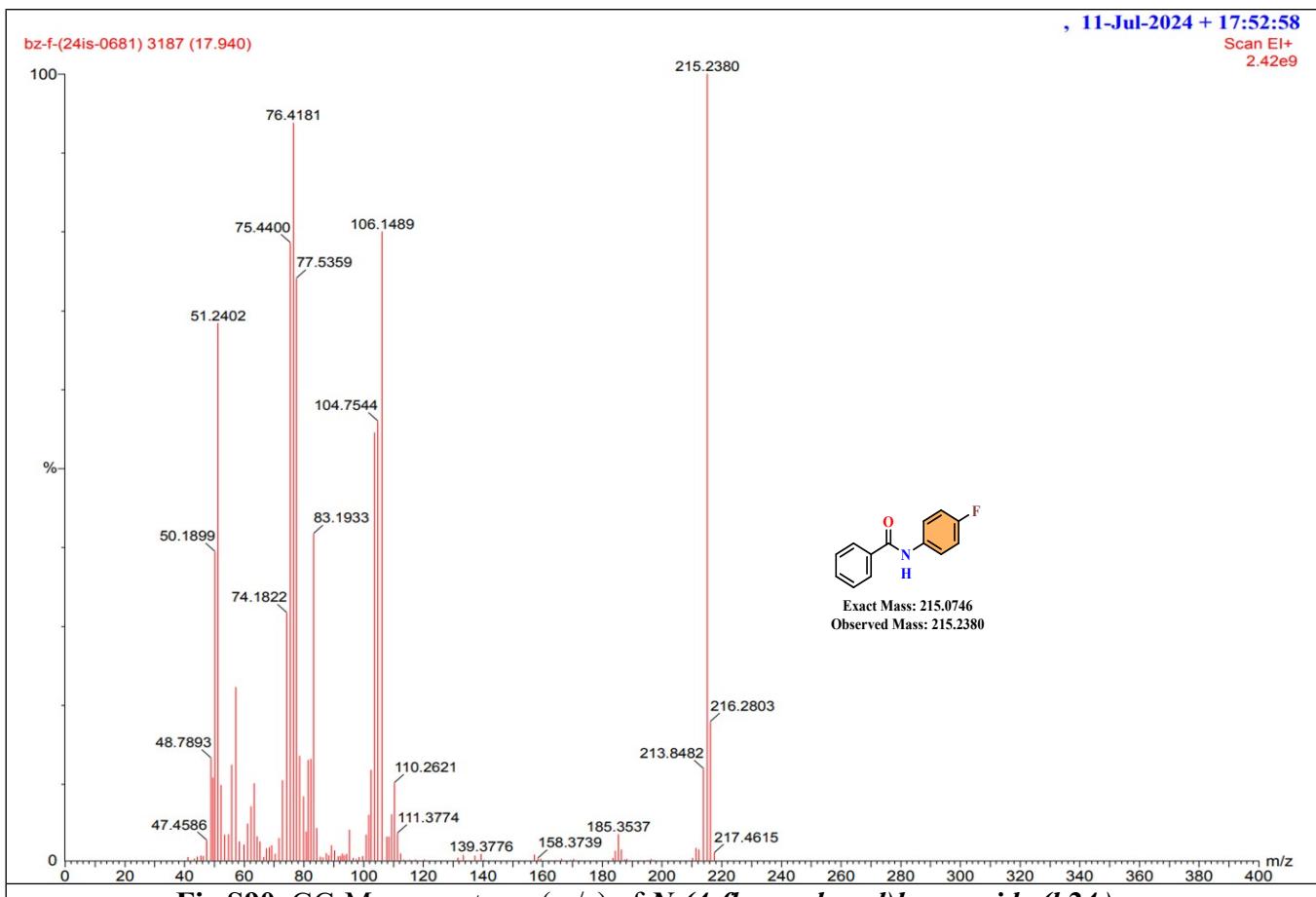


Current Data Parameters  
NAME Dr.PDK090724  
EXPNO 16  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240709  
Time 20.19 h  
INSTRUM spect  
PROBHD Z108618\_0505 (PULPROG zgflqpn  
TD 131072  
SOLVENT DMSO  
NS 16  
DS 4  
SWH 89285.711 Hz  
FIDRES 1.362392 Hz  
AQ 0.7340032 sec  
RG 199.6  
DW 5.600 usec  
DE 6.50 usec  
TE 303.5 K  
D1 1.00000000 sec  
TD0 1  
SFO1 376.5811447 MHz  
NUC1 19F  
P1 15.00 usec  
PLW1 20.11800003 W

F2 - Processing parameters  
SI 65536  
SF 376.6188065 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00





**Fig S90.** GC-Mass spectrum (m/z) of *N*-(4-fluorophenyl)benzamide (**b24**)

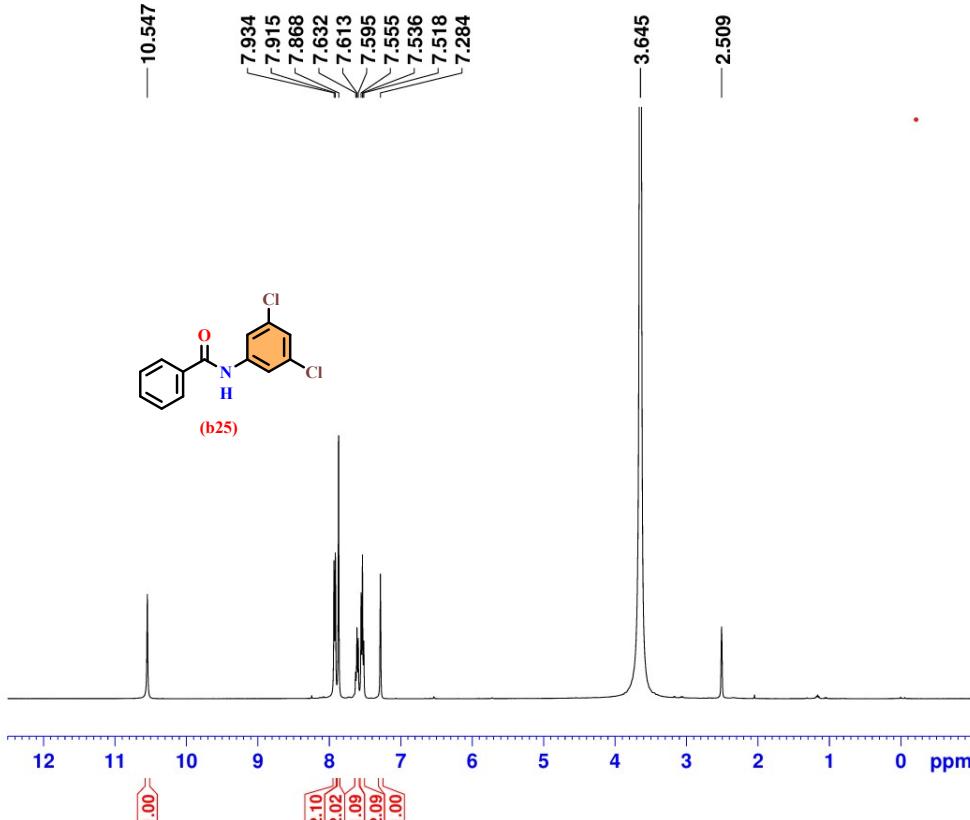
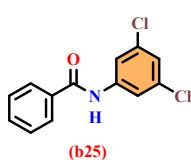
Signature SIF VIT VELLORE  
Bz-25



Current Data Parameters  
NAME Dr.PDK031024  
EXPNO 4  
PROCNO 1

F2 - Acquisition Parameters  
Date 20241003  
Time 20.42 h  
INSTRUM spect  
PROBHD Z108618\_0505 ( PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 32  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 35.49  
DW 62.400 usec  
DE 6.50 usec  
TE 302.7 K  
D1 1.000000000 sec  
TD0 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

F2 - Processing parameters  
SI 65536  
SF 400.2580000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



**Fig S91.**  $^1\text{H}$  NMR spectrum of *N*-(3,5-dichlorophenyl)benzamide (**b25**)

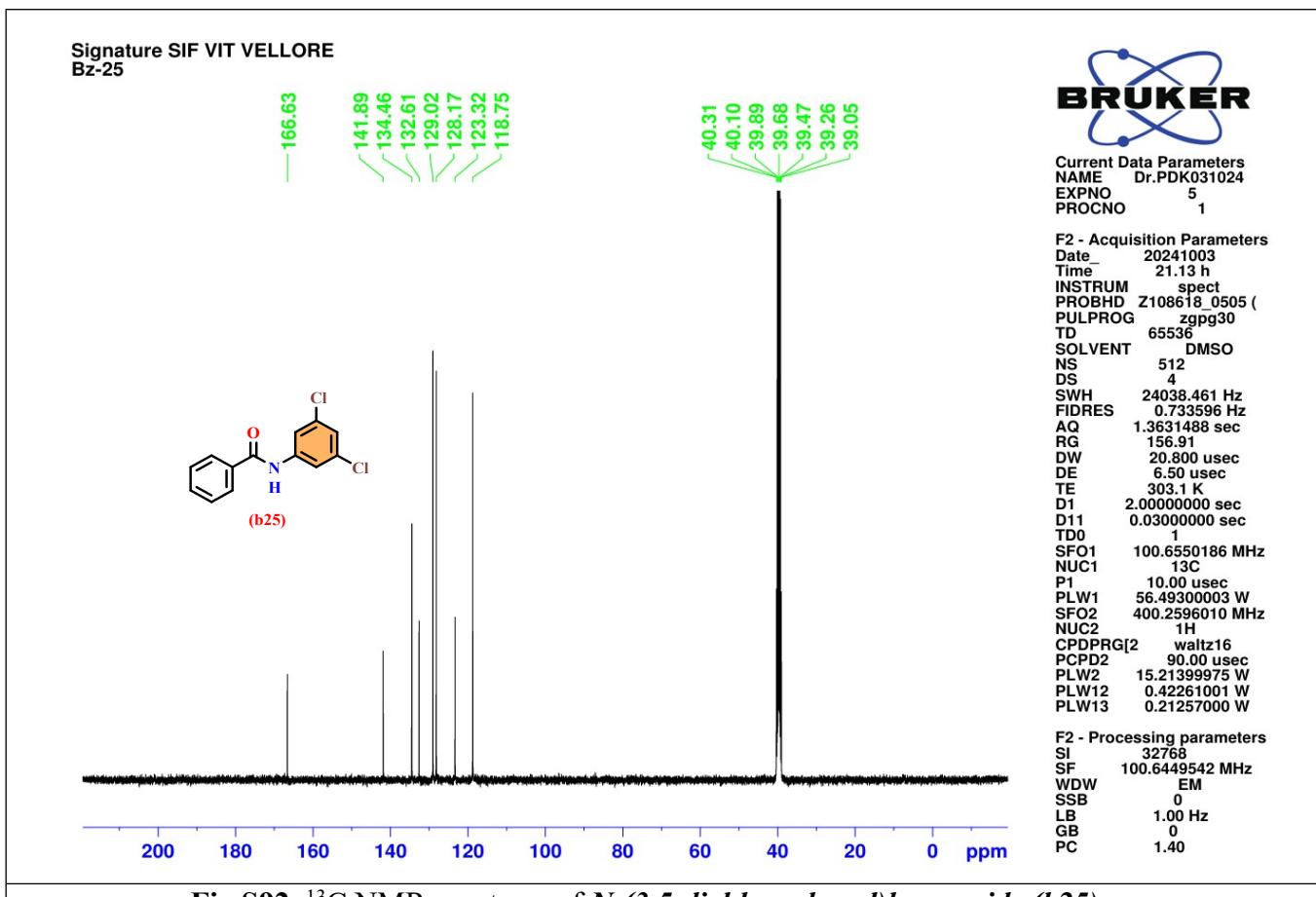


Fig S92. <sup>13</sup>C NMR spectrum of *N*-(3,5-dichlorophenyl)benzamide (b25)

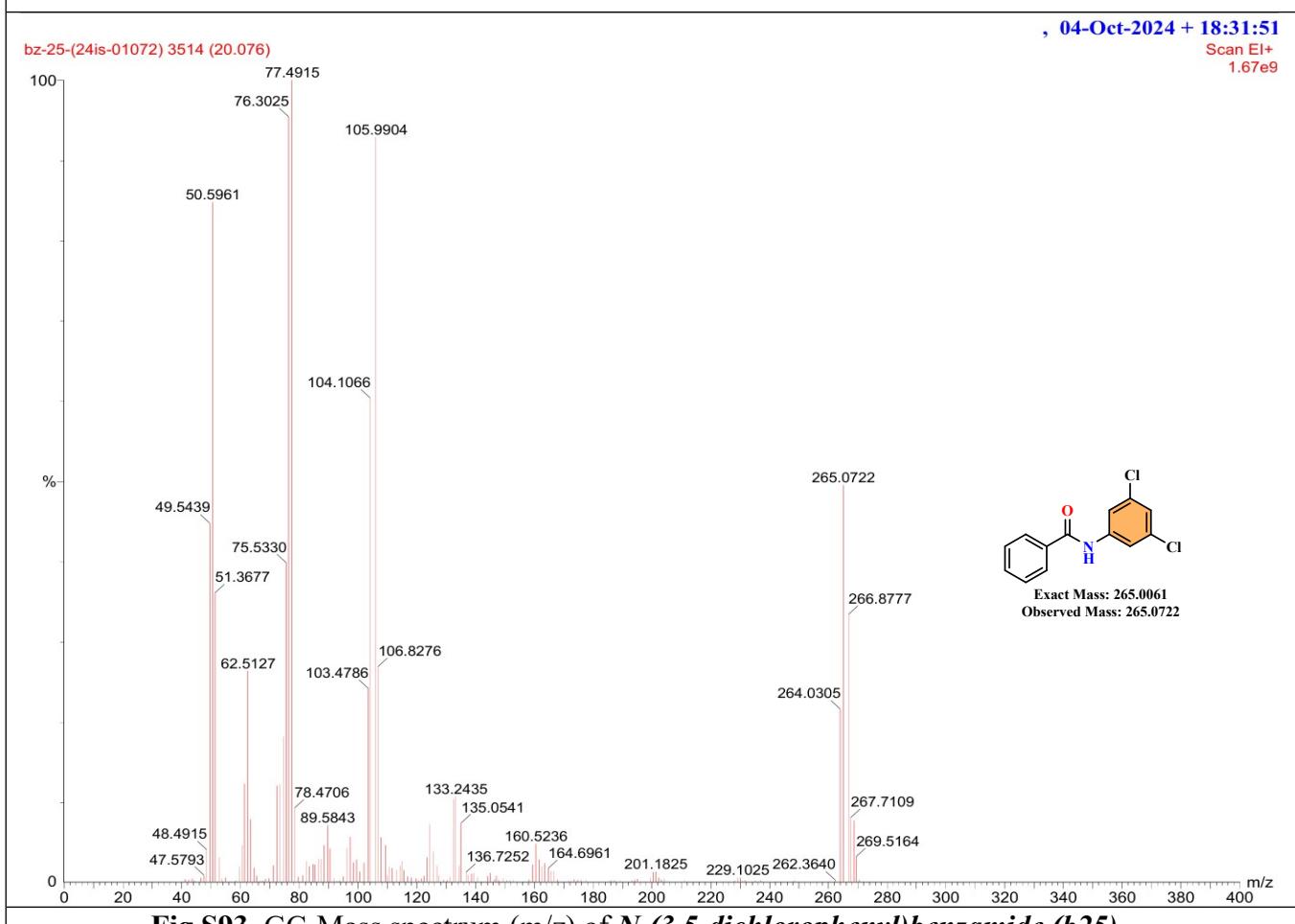


Fig S93. GC-Mass spectrum (m/z) of *N*-(3,5-dichlorophenyl)benzamide (b25)

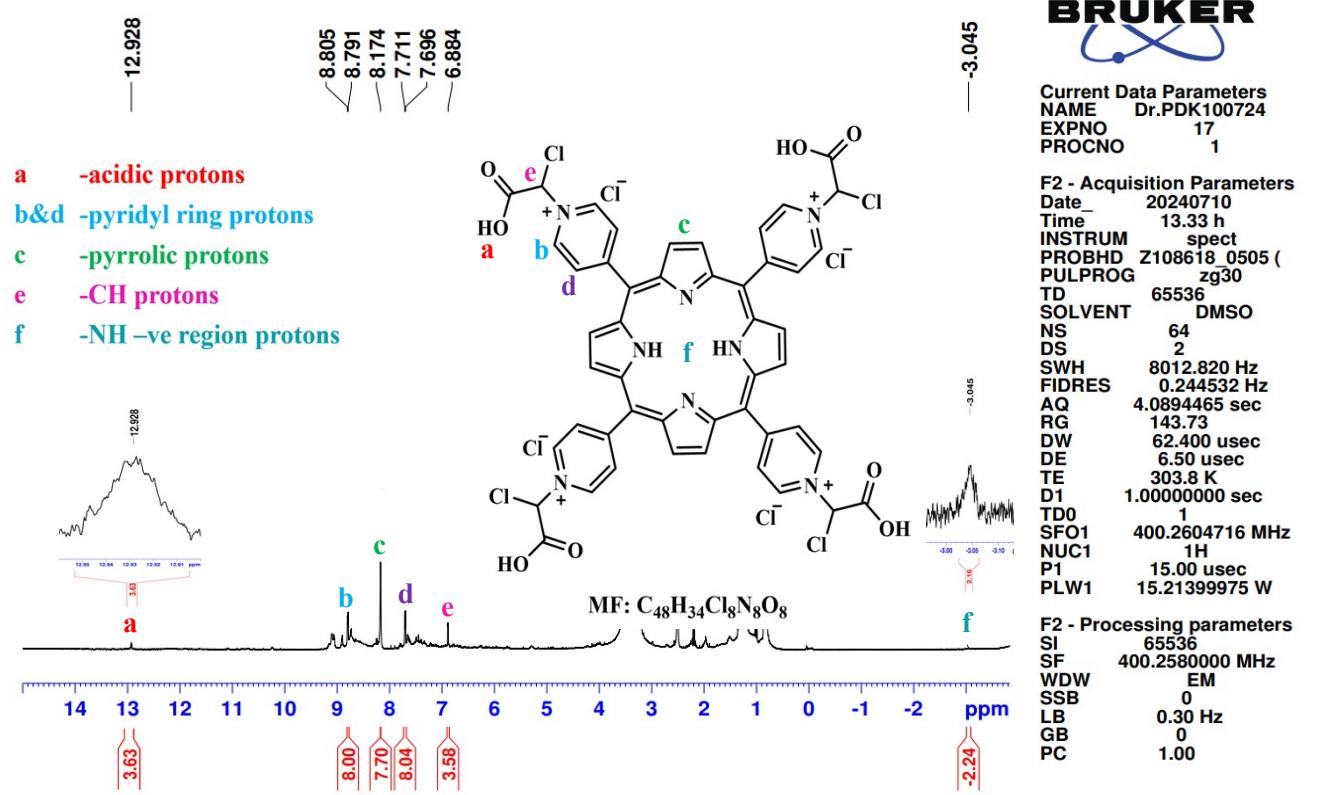


Fig S94. <sup>1</sup>H NMR spectrum of PcCFP Photocatalyst

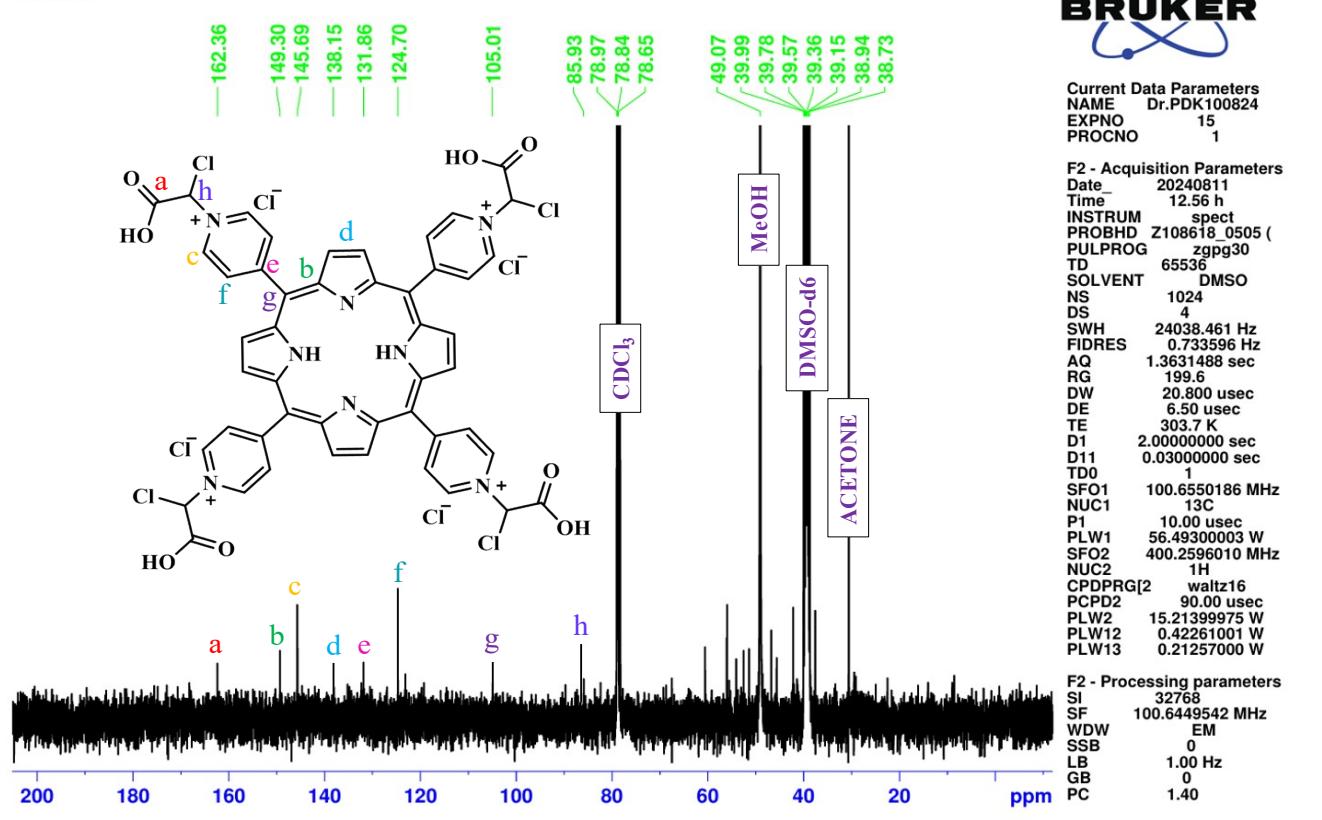


Fig S95. <sup>13</sup>C NMR spectrum of PcCFP Photocatalyst

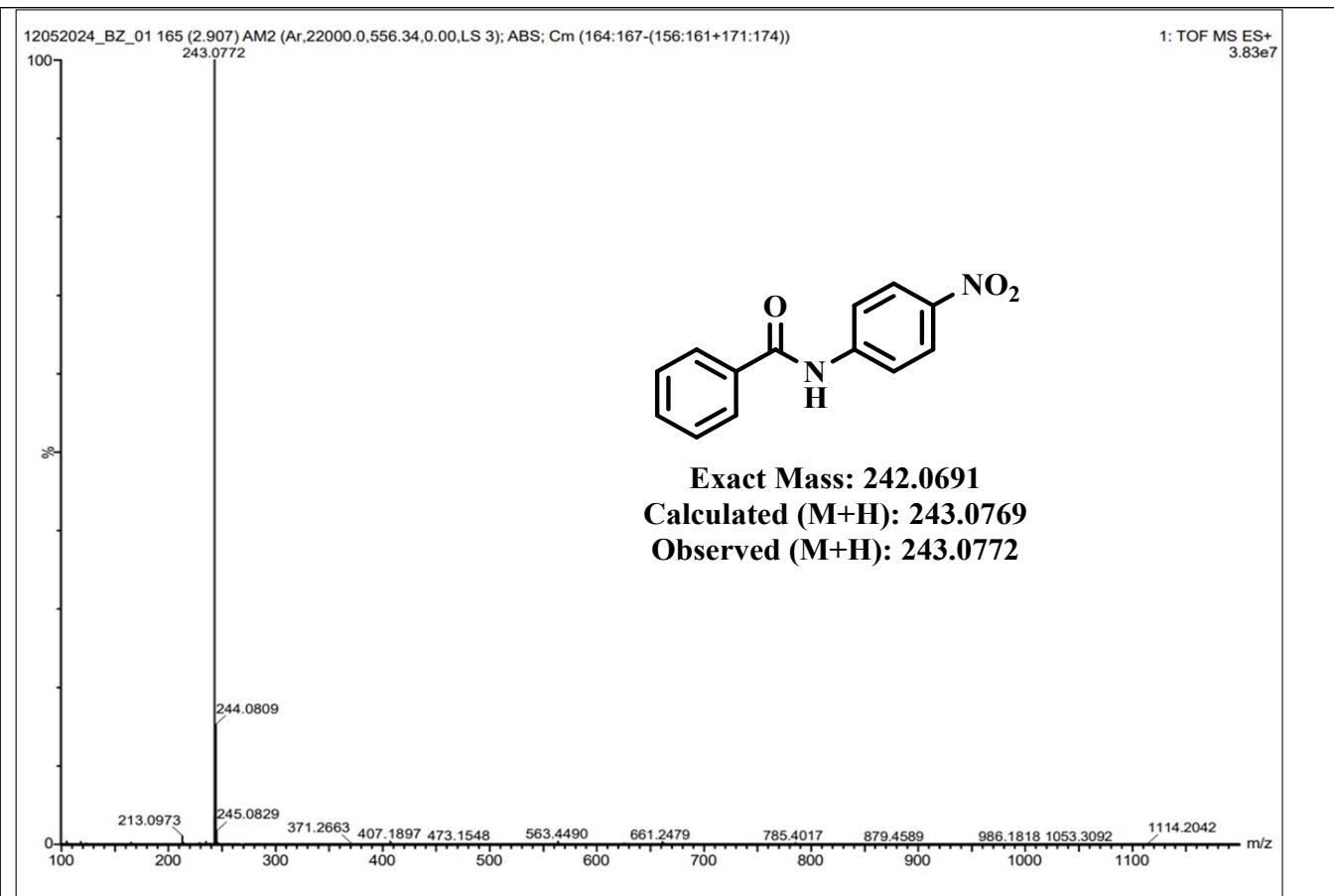


Fig S96. High Resolution Mass spectrum (m/z) of *N*-(4-nitrophenyl)benzamide (*b1<sub>i</sub>*)

Signature SIF VIT VELLORE  
BZ-01-CI

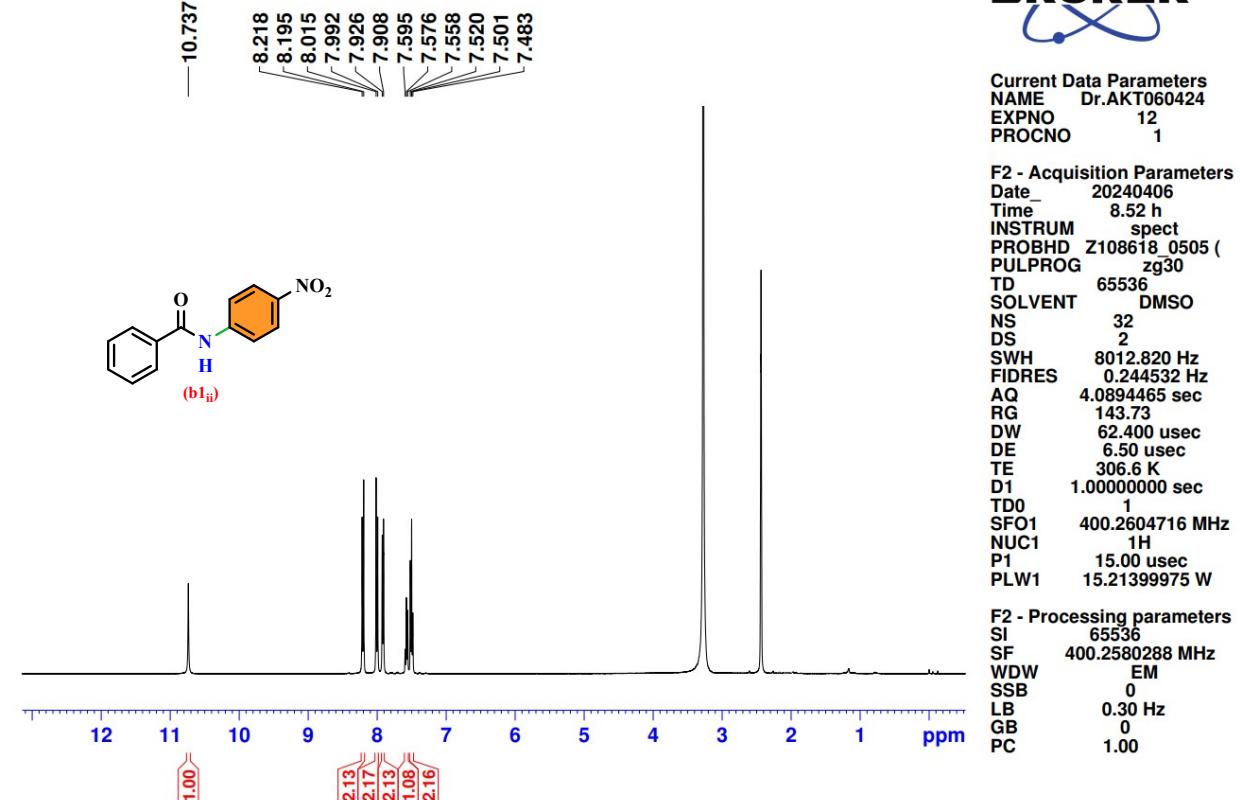


Fig S97. <sup>1</sup>H NMR spectrum of *N*-(4-nitrophenyl)benzamide (*b1<sub>ii</sub>*)

Signature SIF VIT VELLORE  
BZ-02-Cl

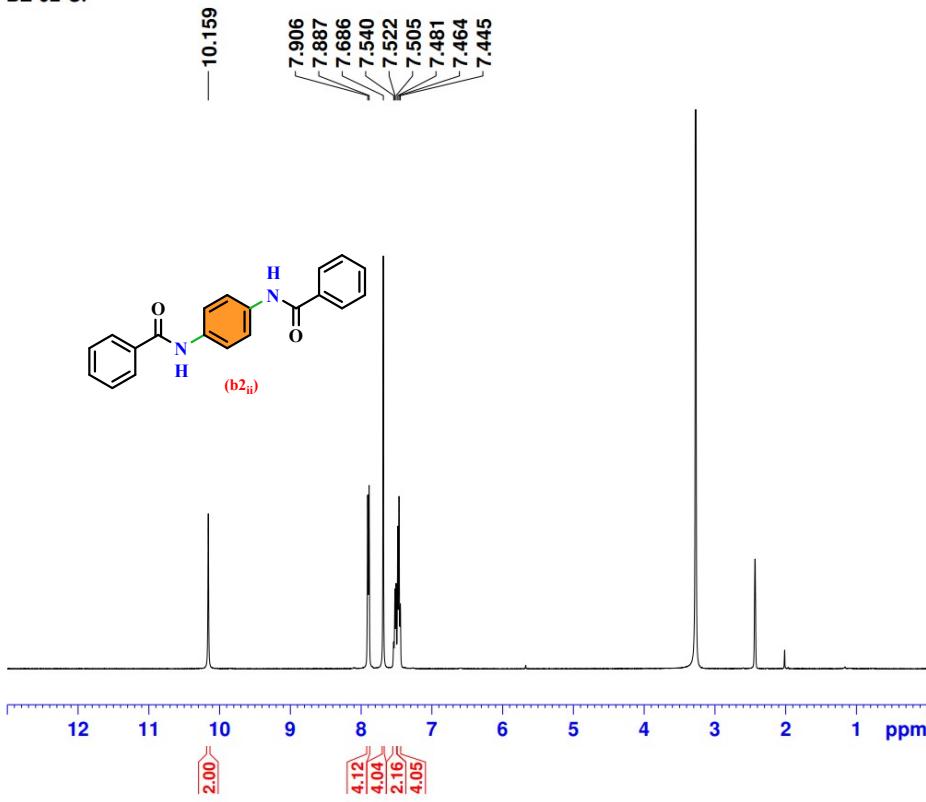


Fig S98.  $^1\text{H}$  NMR spectrum of  $N,N'$ -(1,4-phenylene)dibenzamide ( $b2_{ii}$ )

Signature SIF VIT VELLORE  
BZ-03-Cl

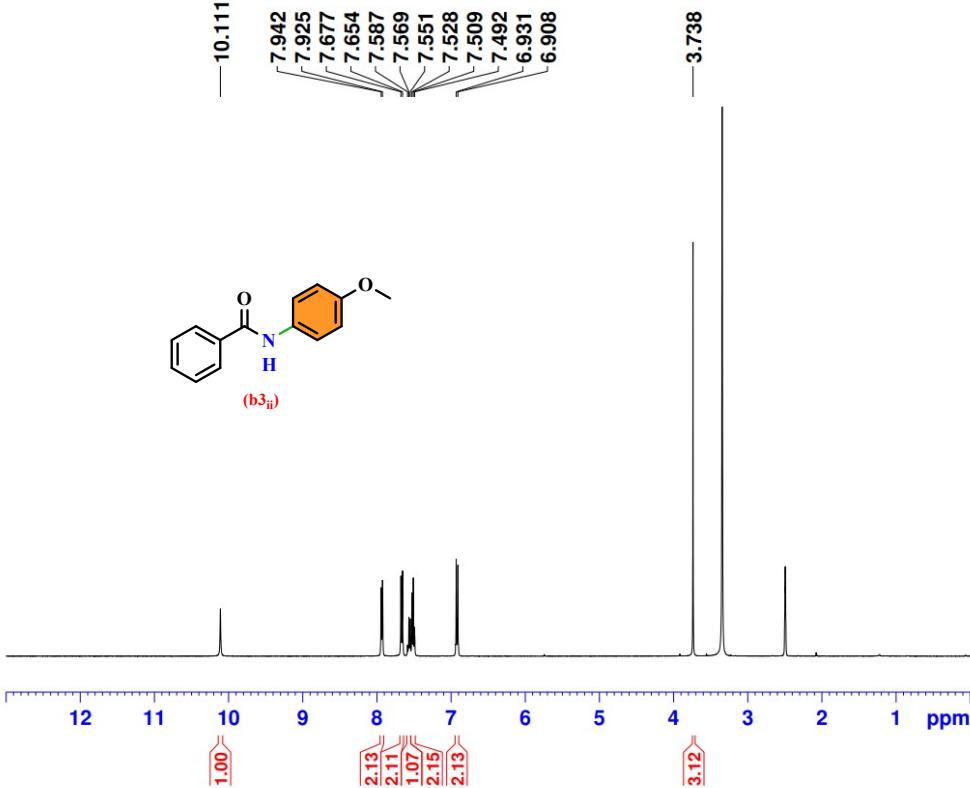


Fig S99.  $^1\text{H}$  NMR spectrum of  $N$ -(4-methoxyphenyl)benzamide ( $b3_{ii}$ )

Signature SIF VIT VELLORE  
BZ-04-Cl



Current Data Parameters  
NAME Dr.PDK200424  
EXPNO 52  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240420  
Time 12.30 h  
INSTRUM spect  
PROBHD Z108618\_0505 (PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 32  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 127.79  
DW 62.400 usec  
DE 6.50 usec  
TE 307.2 K  
D1 1.0000000 sec  
TD0 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

F2 - Processing parameters  
SI 65536  
SF 400.2580291 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

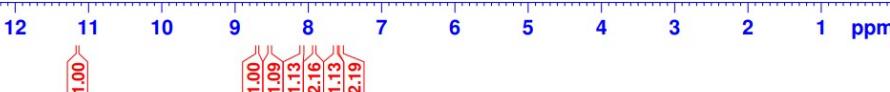
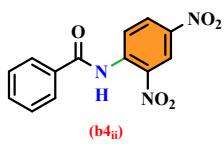


Fig S100. <sup>1</sup>H NMR spectrum of *N*-(2,4-dinitrophenyl)benzamide (*b4*<sub>ii</sub>)

Signature SIF VIT VELLORE  
BZ-05-Cl



Current Data Parameters  
NAME Dr.AKT080424  
EXPNO 21  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240408  
Time 16.15 h  
INSTRUM spect  
PROBHD Z108618\_0505 (PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 32  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 127.79  
DW 62.400 usec  
DE 6.50 usec  
TE 303.7 K  
D1 1.0000000 sec  
TD0 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

F2 - Processing parameters  
SI 65536  
SF 400.2580297 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

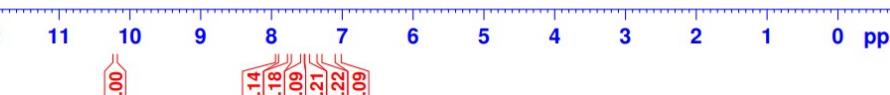
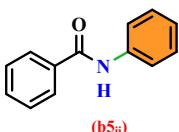


Fig S101. <sup>1</sup>H NMR spectrum of *N*-phenylbenzamide (*b5*<sub>ii</sub>)

Signature SIF VIT VELLORE  
BZ-05-I



Current Data Parameters  
NAME Dr.PDK200424  
EXPNO 52  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240420  
Time 12.30 h  
INSTRUM spect  
PROBHD Z108618\_0505 (zg30)  
TD 65536  
SOLVENT DMSO  
NS 32  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 127.79  
DW 62.400 usec  
DE 6.50 usec  
TE 307.2 K  
D1 1.0000000 sec  
TD0 1  
SF01 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

F2 - Processing parameters  
SI 65536  
SF 400.2580291 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

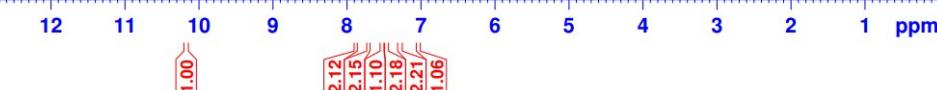
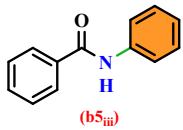


Fig S102. <sup>1</sup>H NMR spectrum of *N*-phenylbenzamide (*b5*<sub>iii</sub>)

Signature SIF VIT VELLORE  
BZ-06-Cl



Current Data Parameters  
NAME Dr.AKT080424  
EXPNO 18  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240408  
Time 16.00 h  
INSTRUM spect  
PROBHD Z108618\_0505 (zg30)  
TD 65536  
SOLVENT DMSO  
NS 32  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 127.79  
DW 62.400 usec  
DE 6.50 usec  
TE 303.7 K  
D1 1.0000000 sec  
TD0 1  
SF01 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

F2 - Processing parameters  
SI 65536  
SF 400.2580134 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

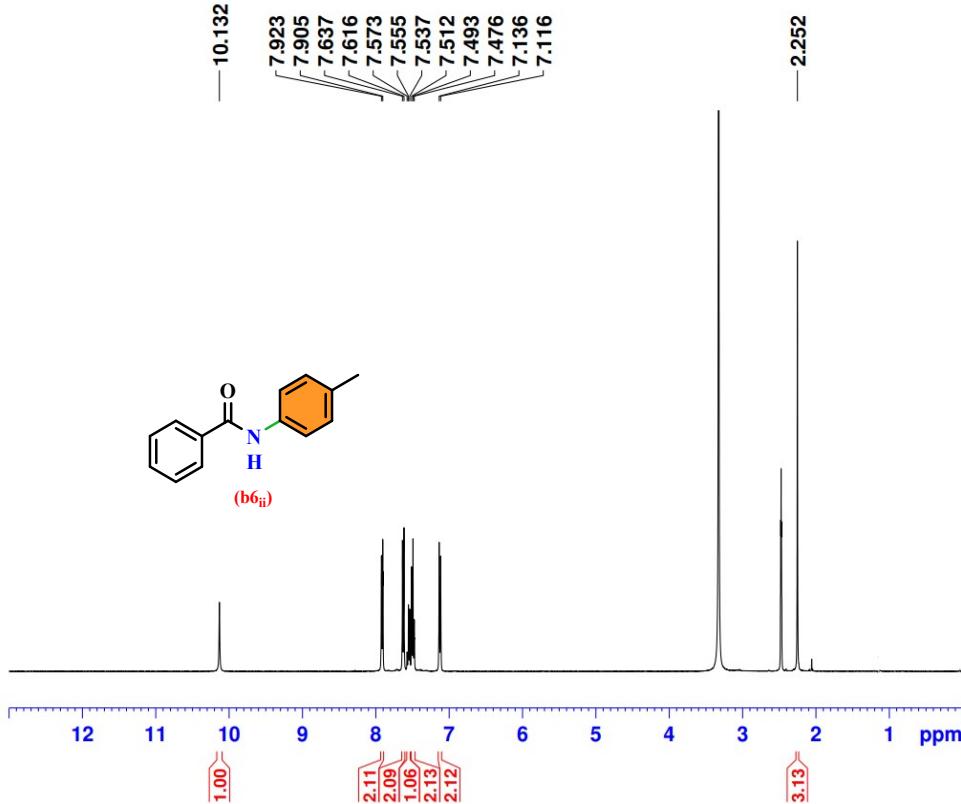
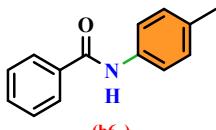


Fig S103. <sup>1</sup>H NMR spectrum of *N*-(*p*-tolyl)benzamide (*b6*)

Signature SIF VIT VELLORE  
Bz-13-Cl



Current Data Parameters  
NAME Dr.AKT090524  
EXPNO 7  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240509  
Time 23.28 h  
INSTRUM spect  
PROBHD Z108618\_0505 (PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 32  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 199.6  
DW 62.400 usec  
DE 6.50 usec  
TE 303.3 K  
D1 1.0000000 sec  
TD0 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

F2 - Processing parameters  
SI 65536  
SF 400.2580198 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

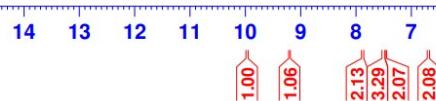
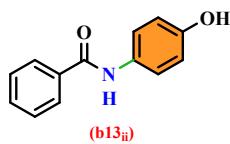


Fig S104. <sup>1</sup>H NMR spectrum of *N*-(4-hydroxyphenyl)benzamide (*b*13<sub>ii</sub>)

Signature SIF VIT VELLORE  
BZ-14-Cl



Current Data Parameters  
NAME Dr.ASK220424  
EXPNO 63  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240422  
Time 15.28 h  
INSTRUM spect  
PROBHD Z108618\_0505 (PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 32  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 127.79  
DW 62.400 usec  
DE 6.50 usec  
TE 305.7 K  
D1 1.0000000 sec  
TD0 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

F2 - Processing parameters  
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SF 400.2580060 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

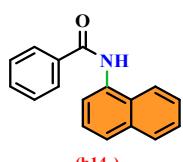


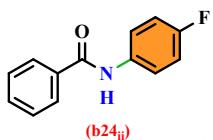
Fig S105. <sup>1</sup>H NMR spectrum of *N*-(naphthalen-1-yl)benzamide (*b*14<sub>ii</sub>)



Current Data Parameters  
NAME Dr.PDK250724  
EXPNO 46  
PROCNO 1

F2 - Acquisition Parameters  
Date 20240725  
Time 19.26 h  
INSTRUM spect  
PROBHD Z108618\_0505 (   
PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 32  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 98.85  
DW 62.400 usec  
DE 6.50 usec  
TE 303.5 K  
D1 1.0000000 sec  
TD0 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 15.21399975 W

F2 - Processing parameters  
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SF 400.2580282 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



(b24ii)

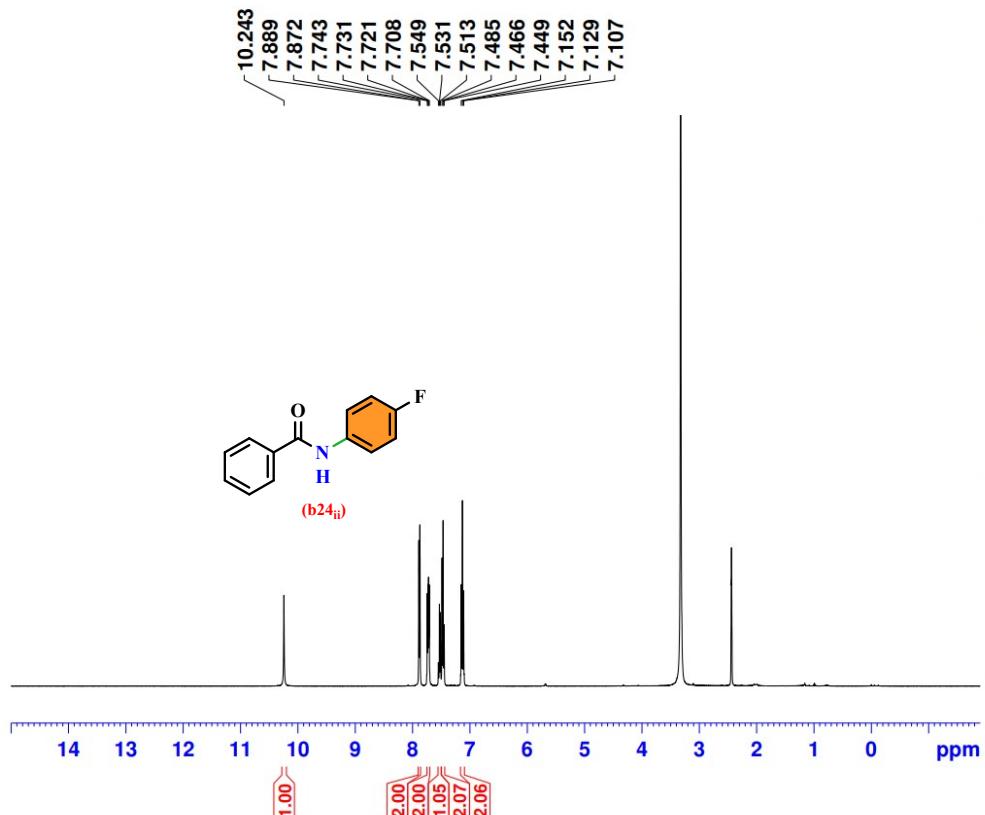
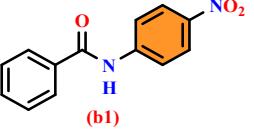
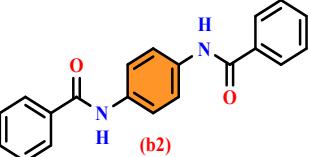
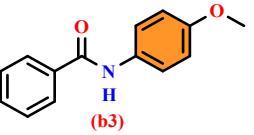
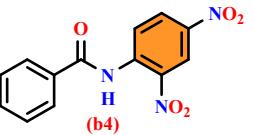
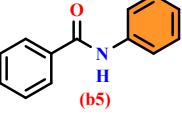
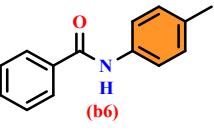
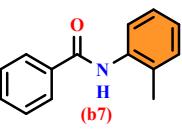
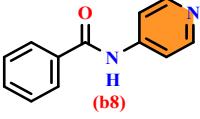
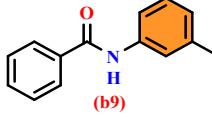
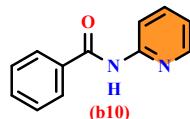
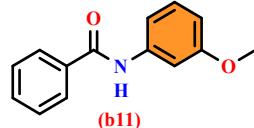
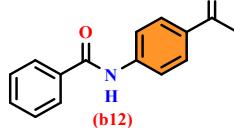
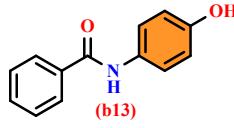
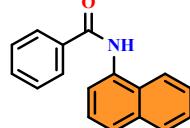
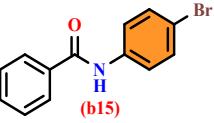
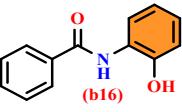
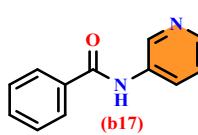
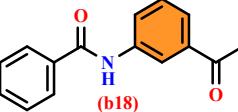
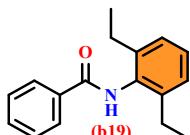
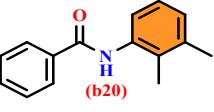


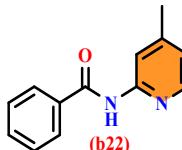
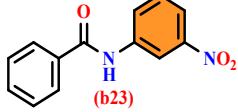
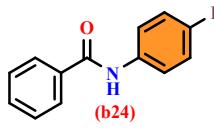
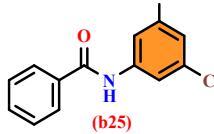
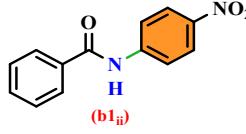
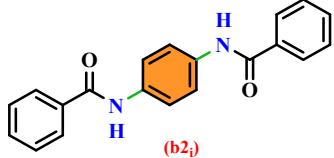
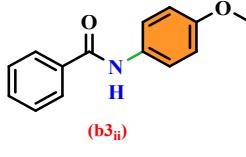
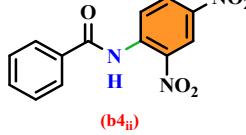
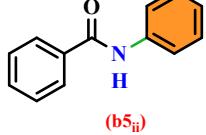
Fig S106.  $^1\text{H}$  NMR spectrum of *N*-(4-fluorophenyl)benzamide (*b24<sub>ii</sub>*)

**Table S2.** Spectral data of Products

Sr. No	CODE	Characterization Data	Derivative Structure
1.	b1 <sub>i</sub>	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.80 (s, 1H), 8.28 (d, J = 8.80 Hz, 2H), 8.08 (d, J = 8.40 Hz, 2H), 7.99 (d, J = 8.00 Hz, 2H), 7.65 (t, J = 6.80 Hz, 1H), 7.58 (t, J = 7.60 Hz, 2H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 166.78, 145.97, 142.94, 134.71, 132.66, 129, 128.39, 125.28, 120.32. GCMS exact m/z <sup>+</sup> 242.0691 & observed m/z <sup>+</sup> 242.1669. HRMS calculated (M+H) 243.0691 & observed (M+H) 243.0772.	
2.	b2 <sub>i</sub>	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.16 (s, 2H), 7.90 (d, J = 7.60 Hz, 4H), 7.69 (s, 4H), 7.52 (t, J = 6.80 Hz, 2H), 7.46 (t, J = 7.60 Hz, 4H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 165.79, 135.46, 135.44, 131.94, 128.84, 128.06, 121.14. GCMS exact m/z 316.1212 & observed m/z is 316.1663.	
3.	b3 <sub>i</sub>	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.04 (s, 1H), 7.87 (d, J = 6.80 Hz, 2H), 7.60 (d, J = 8.80 Hz, 2H), 7.51 (t, J = 7.20 Hz, 1H), 7.45 (t, J = 6.80 Hz, 2H), 6.86 (d, J = 9.20 Hz, 2H), 3.68 (s, 3H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 165.58, 156.03, 135.52, 132.69, 131.83, 128.81, 127.99, 122.46, 114.21, 55.66. GCMS exact m/z 227.0946 & observed m/z is 227.2144.	
4.	b4 <sub>i</sub>	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 11.16 (s, 1H), 8.69 (s, 1H), 8.53 (d, J = 9.20 Hz, 1H), 8.07 (d, J = 8.80 Hz, 1H), 7.92 (d, J = 7.20 Hz, 2H), 7.63 (t, J = 7.20 Hz, 1H), 7.54 (t, J = 7.20 Hz, 1H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 166.05, 143.29, 141.34, 137.87, 133.41, 133.36, 129.34, 129.13, 128.40, 126.03, 121.67. GCMS exact m/z 287.0542 & observed m/z is 287.1621.	
5.	b5 <sub>i</sub>	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.17 (s, 1H), 7.89 (d, J = 6.80 Hz, 2H), 7.71 (d, J = 7.60 Hz, 2H), 7.53 (t, J = 7.20 Hz, 1H), 7.46 (t, J = 6.80 Hz, 2H), 7.29 (t, J = 8.40 Hz, 2H), 7.03 (t, J = 7.20 Hz, 1H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 166.03, 139.63, 135.46, 132.00, 129.06, 128.85, 128.11, 124.13, 120.84. GCMS exact m/z 197.0841 & observed m/z is 197.4289.	
6.	b6 <sub>i</sub>	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.09 (s, 1H), 7.87 (d, J = 7.20 Hz, 2H), 7.59 (d, J = 8.40 Hz, 2H), 7.51 (t, J = 7.20 Hz, 1H), 7.45 (t, J = 7.20 Hz, 2H), 7.09 (d, J = 8.40 Hz, 2H), 2.21(s, 3H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 165.80, 137.10, 135.52, 133.07, 131.90, 129.44, 128.81, 128.05, 120.87, 20.96. GCMS exact m/z 211.0997 & observed m/z is 211.2645.	
7.	b7	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 9.90 (s, 1H), 8.01 (d, J = 6.80 Hz, 2H), 7.59 (t, J = 7.20 Hz, 1H), 7.52 (t, J = 6.80 Hz, 2H), 7.37 (d, J = 7.60 Hz, 1H), 7.27 (d, J = 7.60 Hz, 1H), 7.22 (t, J = 7.20 Hz, 1H), 7.17 (t, J = 6.80 Hz, 1H), 2.25 (s, 3H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 165.80, 136.92, 135.04, 134.23, 131.98, 130.79, 128.88, 128.12, 127.11, 126.47, 18.40. GCMS exact m/z 211.0997 &	

		observed m/z is 211.1141.	
8.	b8	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.11 (s, 1H), 7.93 (d, J = 6.80 Hz, 2H), 7.66 (d, J = 9.20 Hz, 2H), 7.57 (t, J = 7.20 Hz, 1H), 7.51 (t, J = 6.80 Hz, 2H), 6.92 (d, J = 8.80 Hz, 2H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 165.58, 135.51, 132.68, 131.84, 12881, 127.99, 122.46, 114.20. GCMS exact m/z 198.0793 & observed m/z is 198.1938.	
9.	b9	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.09 (s, 1H), 7.88 (d, J = 7.20 Hz, 2H), 7.55 (d, J = 7.60 Hz, 2H), 7.51 (t, J = 5.20 Hz, 1H), 7.46 (t, J = 6.80 Hz, 2H), 7.16 (t, J = 7.60 Hz, 1H), 6.86 (d, J = 7.20 Hz, 1H), 2.24 (s, 3H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 165.95, 139.54, 138.21, 135.48, 131.97, 128.90, 128.84, 128.08, 124.83, 121.38, 118.03, 21.69. GCMS exact m/z 211.0977 & observed m/z is 211.2772.	
10.	b10	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.74 (s, 1H), 8.18 (d, J = 8.40 Hz, 1H), 8.02 (d, J = 7.20 Hz, 1H), 7.75 (d, J = 7.20 Hz, 2H), 7.57 (t, J = 6.40 Hz, 1H), 7.51 (t, J = 7.20 Hz, 2H), 7.43 (t, J = 7.60 Hz, 2H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 166.50, 148.39, 145.20, 138.59, 134.85, 133.10, 129.45, 128.43, 122.87, 120.30. GCMS exact m/z 198.0793 & observed m/z is 198.1018.	
11.	b11	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.21 (s, 1H), 7.94 (d, J = 6.80 Hz, 2H), 7.59 (t, J = 6.40 Hz, 1H), 7.53 (t, J = 6.80 Hz, 2H), 7.47 (s, 1H), 7.37 (d, J = 8.00 Hz, 1H), 7.25 (t, J = 8.40 Hz, 1H), 6.68 (d, J = 8.40 Hz, 1H), 3.75 (s, 3H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 166.05, 159.89, 140.83, 135.43, 132.05, 129.85, 128.85, 128.10, 113.00, 109.62, 106.48, 55.48. GCMS exact m/z 227.0946 & observed m/z is 227.1035.	
12.	b12	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.53 (s, 1H), 7.92-7.95 (m, 6H), 7.61 (t, J = 6.40 Hz, 1H), 7.54 (t, J = 8.00 Hz, 2H), 2.54 (s, 3H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 197.09, 166.48, 144.06, 135.05, 132.54, 129.74, 128.92, 128.26, 119.95, 26.92. GCMS exact m/z 239.0946 & observed m/z is 239.0987.	
13.	b13 <sub>i</sub>	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.00 (s, 1H), 9.23 (s, 1H), 7.91 (d, J = 7.20 Hz, 2H), 7.54 (d, J = 7.20 Hz, 2H), 7.52 (t, J = 2.40 Hz, 1H), 7.51 (t, J = 2.80 Hz, 2H), 6.72 (d, J = 8.80 Hz, 2H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 165.32, 154.16, 135.63, 131.73, 131.15, 128.78, 127.95, 122.72, 115.42. GCMS exact m/z 213.0790 & observed m/z is 213.2923.	
14.	b14 <sub>i</sub>	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.40 (s, 1H), 8.09 (d, J = 7.60 Hz, 2H), 7.96-7.97 (m, 2H), 7.86 (d, J = 8.00 Hz, 1H), 7.53-7.64 (m, 7H). <sup>13</sup> C NMR 100 MHz, DMSO-d6: δ 166.67, 134.95, 134.33, 134.25, 132.11, 129.71, 128.92, 128.53, 128.25, 126.75, 126.52, 126.43, 126.01, 124.37, 123.78. GCMS exact m/z 247.0997 & observed m/z is 247.2688.	
15.	b15	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.34 (s, 1H), 7.94	

		(d, $J = 8.00$ Hz, 2H), 7.76 (d, $J = 8.80$ Hz, 2H), 7.59 (t, $J = 6.80$ Hz, 1H), 7.51-7.54 (m, 4H). $^{13}\text{C}$ NMR 100 MHz, DMSO-d6: $\delta$ 166.14, 139.04, 135.17, 132.18, 131.89, 128.89, 128.13, 122.72, 115.81. GCMS exact m/z 274.9946 & observed m/z is 275.0620.	
16.	b16	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 9.74 (s, 1H), 9.50 (s, 1H), 7.97 (d, $J = 7.60$ Hz, 2H), 7.69 (d, $J = 7.60$ Hz, 1H), 7.61 (t, $J = 6.80$ Hz, 1H), 7.54 (t, $J = 7.20$ Hz, 2H), 7.05 (t, $J = 7.60$ Hz, 1H), 6.93 (d, $J = 8.00$ Hz, 1H), 6.85 (t, $J = 7.60$ Hz, 1H). $^{13}\text{C}$ NMR 100 MHz, DMSO-d6: $\delta$ 165.93, 149.76, 134.81, 132.15, 128.99, 127.94, 126.36, 126.16, 124.52, 119.54, 116.48. GCMS exact m/z 213.0790 & observed m/z is 213.1008.	
17.	b17	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 10.44 (s, 1H), 8.93 (s, 1H), 8.31 (d, $J = 4.40$ Hz, 1H), 8.19 (d, $J = 8.40$ Hz, 1H), 7.97 (d, $J = 8.00$ Hz, 2H), 7.61 (t, $J = 6.80$ Hz, 1H), 7.54 (t, $J = 8.00$ Hz, 2H), 7.40 (t, $J = 6.40$ Hz, 1H). $^{13}\text{C}$ NMR 100 MHz, DMSO-d6: $\delta$ 166.45, 144.91, 142.34, 136.33, 134.79, 132.37, 128.95, 128.19, 127.98, 124.06. GCMS exact m/z 198.0793 & observed m/z is 198.1033.	
18.	b18	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 10.44 (s, 1H), 8.37 (s, 1H), 8.07 (d, $J = 8.00$ Hz, 1H), 7.98 (d, $J = 7.60$ Hz, 2H), 7.71 (d, $J = 7.60$ Hz, 1H), 7.61 (t, $J = 6.80$ Hz, 1H), 7.54 (t, $J = 8.40$ Hz, 2H), 7.50 (d, $J = 8.00$ Hz, 1H). $^{13}\text{C}$ NMR 100 MHz, DMSO-d6: $\delta$ 197.88, 165.93, 139.73, 137.47, 134.79, 131.94, 129.22, 128.62, 127.85, 125.00, 123.82, 119.89, 26.91. GCMS exact m/z 239.0946 & observed m/z is 239.0987.	
19.	b19	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 9.75 (s, 1H), 8.00 (s, 1H), 7.60 (t, $J = 6.80$ Hz, 1H), 7.54 (t, $J = 6.80$ Hz, 2H), 7.24 (t, $J = 8.40$ Hz, 1H), 7.16 (d, $J = 7.60$ Hz, 2H), 2.55 (q, $J = 8.00$ Hz, 4H), 1.12 (t, $J = 7.60$ Hz, 6H). $^{13}\text{C}$ NMR 100 MHz, DMSO-d6: $\delta$ 166.26, 142.20, 134.91, 134.67, 131.93, 128.96, 127.91, 127.79, 126.52, 24.94, 14.98. GCMS exact m/z 253.1467 & observed m/z 253.2440.	
20.	b20	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 9.92 (s, 1H), 7.99 (d, $J = 7.20$ Hz, 2H), 7.60 (t, $J = 7.20$ Hz, 1H), 7.53 (t, $J = 6.80$ Hz, 2H), 7.08-7.14 (m, 3H), 2.29 (s, 3H), 2.11 (s, 3H). $^{13}\text{C}$ NMR 100 MHz, DMSO-d6: $\delta$ 165.87, 140.60, 137.44, 136.74, 135.05, 131.92, 128.86, 128.06, 125.71, 125.20, 20.61, 14.69. GCMS exact m/z 225.1154 & observed m/z 225.1183.	
21.	b21	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 12.39 (s, 1H), 8.67 (d, $J = 8.00$ Hz, 1H), 8.13 (d, $J = 8.00$ Hz, 1H), 7.98 (d, $J = 6.80$ Hz, 2H), 7.70 (t, $J = 7.20$ Hz, 1H), 7.66 (t, $J = 4.80$ Hz, 1H), 7.62 (t, $J = 6.80$ Hz, 2H), 7.28 (t, $J = 7.20$ Hz, 1H), 2.71 (s, 3H). $^{13}\text{C}$ NMR 100 MHz, DMSO-d6: $\delta$ 204.08, 165.54, 140.19, 135.28, 135.19, 134.85, 132.74, 129.51, 127.57, 123.90, 123.64, 120.74, 29.25. GCMS exact m/z 239.0946 & observed m/z 239.2092.	
22.	b22	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 10.69 (s, 1H), 8.24	

		(d, $J = 4.80$ Hz, 1H), 8.06 (s, 1H), 8.03 (d, $J = 7.20$ Hz, 2H), 7.60 (t, $J = 7.20$ Hz, 1H), 7.51 (t, $J = 7.20$ Hz, 2H), 7.02 (d, $J = 4.40$ Hz, 1H), 2.36 (s, 3H). $^{13}\text{C}$ NMR 100 MHz, DMSO-d6: $\delta$ 166.35, 152.69, 149.25, 148.02, 134.60, 132.35, 128.82, 128.40, 121.31, 115.60, 21.41. GCMS exact m/z 212.0950 & observed m/z 212.1805.	 <b>(b22)</b>
23.	b23	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 10.70 (s, 1H), 8.80 (s, 1H), 8.18 (d, $J = 8.00$ Hz, 1H), 7.98 (d, $J = 7.20$ Hz, 2H), 7.95 (d, $J = 1.60$ Hz, 1H), 7.66 (t, $J = 3.20$ Hz, 1H), 7.62 (t, $J = 7.20$ Hz, 1H), 7.56 (t, $J = 7.20$ Hz, 2H). $^{13}\text{C}$ NMR 100 MHz, DMSO-d6: $\delta$ 166.55, 148.38, 140.83, 134.68, 132.54, 130.56, 129.01, 128.24, 126.66, 118.63, 114.81. GCMS exact m/z 242.0691 & observed m/z 242.1273.	 <b>(b23)</b>
24.	b24 <sub>i</sub>	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 10.32 (s, 1H), 7.96 (d, $J = 7.20$ Hz, 2H), 7.79-7.80 (m, 2H), 7.60 (t, $J = 7.20$ Hz, 1H), 7.54 (t, $J = 6.80$ Hz, 2H), 7.20 (t, $J = 8.80$ Hz, 2H). $^{13}\text{C}$ NMR 100 MHz, DMSO-d6: $\delta$ 165.95, 159.95, 157.56, 136.00, 135.97, 135.26, 132.07, 128.87, 128.09, 122.69, 122.62, 115.76, 115.54. $^{19}\text{F}$ NMR 377 MHz, DMSO-d6: $\delta$ – (118.93-118.86, m, 1F). GCMS exact m/z 215.0746 & observed m/z 215.2380.	 <b>(b24)</b>
25.	b25	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 7.92 (d, $J = 7.60$ Hz, 2H), 7.87 (s, 2H), 7.61 (t, $J = 7.20$ Hz, 1H), 7.54 (t, $J = 6.80$ Hz, 2H), 7.28 (s, 1H). $^{13}\text{C}$ NMR 100 MHz, DMSO-d6: $\delta$ 166.63, 141.89, 134.46, 132.61, 129.02, 128.17, 123.32, 118.75. GCMS exact m/z 265.0061 & observed m/z 265.0722.	 <b>(b25)</b>
26.	b1 <sub>ii</sub>	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 10.74 (s, 1H), 8.20 (d, $J = 8.80$ Hz, 2H), 8.00 (d, $J = 8.40$ Hz, 2H), 7.91 (d, $J = 8.00$ Hz, 2H), 7.57 (t, $J = 6.80$ Hz, 1H), 7.5 (t, $J = 7.60$ Hz, 2H).	 <b>(b1<sub>ii</sub>)</b>
27.	b2 <sub>ii</sub>	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 10.16 (s, 2H), 7.90 (d, $J = 7.60$ Hz, 4H), 7.69 (s, 4H), 7.52 (t, $J = 6.80$ Hz, 2H), 7.46 (t, $J = 7.60$ Hz, 4H).	 <b>(b2<sub>ii</sub>)</b>
28.	b3 <sub>ii</sub>	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 10.11 (s, 1H), 7.93 (d, $J = 6.80$ Hz, 2H), 7.66 (d, $J = 8.80$ Hz, 2H), 7.57 (t, $J = 7.20$ Hz, 1H), 7.5 (t, $J = 6.80$ Hz, 2H), 6.92 (d, $J = 9.20$ Hz, 2H), 3.73 (s, 3H).	 <b>(b3<sub>ii</sub>)</b>
29.	b4 <sub>ii</sub>	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 11.15 (s, 1H), 8.7 (s, 1H), 8.52 (d, $J = 9.20$ Hz, 1H), 8.08 (d, $J = 8.80$ Hz, 1H), 7.92 (d, $J = 7.20$ Hz, 2H), 7.63 (t, $J = 7.20$ Hz, 1H), 7.53 (t, $J = 7.20$ Hz, 1H).	 <b>(b4<sub>ii</sub>)</b>
30.	b5 <sub>ii</sub>	$^1\text{H}$ NMR 400 MHz, DMSO-d6: $\delta$ 10.21 (s, 1H), 7.92 (d, $J = 6.80$ Hz, 2H), 7.74 (d, $J = 7.60$ Hz, 2H), 7.56 (t, $J = 7.20$ Hz, 1H), 7.49 (t, $J = 6.80$ Hz, 2H), 7.32 (t, $J = 8.40$ Hz, 2H), 7.06 (t, $J = 7.20$ Hz, 1H).	 <b>(b5<sub>ii</sub>)</b>

31.	b5 <sub>iii</sub>	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.18 (s, 1H), 7.89 (d, J = 6.80 Hz, 2H), 7.71 (d, J = 7.60 Hz, 2H), 7.52 (t, J = 7.20 Hz, 1H), 7.46 (t, J = 6.80 Hz, 2H), 7.29 (t, J = 8.40 Hz, 2H), 7.02 (t, J = 7.20 Hz, 1H).	
32.	b6 <sub>ii</sub>	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.13 (s, 1H), 7.91 (d, J = 7.20 Hz, 2H), 7.62 (d, J = 8.40 Hz, 2H), 7.55 (t, J = 7.20 Hz, 1H), 7.49 (t, J = 7.20 Hz, 2H), 7.12 (d, J = 8.40 Hz, 2H), 2.25(s, 3H).	
33.	b13 <sub>ii</sub>	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 9.97 (s, 1H), 9.2 (s, 1H), 7.88 (d, J = 7.20 Hz, 2H), 7.52 (d, J = 7.20 Hz, 2H), 7.49 (t, J = 2.40 Hz, 1H), 7.46 (t, J = 2.80 Hz, 2H), 6.69 (d, J = 8.80 Hz, 2H).	
34.	b14 <sub>ii</sub>	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.42 (s, 1H), 8.09 (d, J = 7.60 Hz, 2H), 7.96-7.99 (m, 2H), 7.86 (d, J = 8.00 Hz, 1H), 7.53-7.64 (m, 7H).	
35.	b24 <sub>ii</sub>	<sup>1</sup> H NMR 400 MHz, DMSO-d6: δ 10.24 (s, 1H), 7.88 (d, J = 7.20 Hz, 2H), 7.71-7.74 (m, 2H), 7.53 (t, J = 7.20 Hz, 1H), 7.47 (t, J = 6.80 Hz, 2H), 7.13 (t, J = 8.80 Hz, 2H).	

### **ICP-MS of PcCFP Photocatalyst**

A quantity of 1 mg of PC was dissolved in 4 mL of aqua regia within a borosilicate glass beaker until complete dissolution was achieved. The resulting transparent solution was subsequently transferred to a 100 mL volumetric flask, where double distilled water was added to bring the total volume to 100 mL. This final volume corresponds to a 10 ppm solution of the photocatalyst. From this 100 mL solution, 0.5 mL was extracted using a pipette and added to a 10 mL volumetric flask, which was then filled with double distilled water to achieve a total volume of 10 mL. This preparation results in a concentration of 0.5 ppm, equivalent to 500 ppb, necessary for the sample preparation.

The ICP-MS analysis was carried out by using 500 ppb of solution and the metals are detected in Below Detection Limit (BDL)

**Table S3.** Metal detection in Photocatalyst by ICP-MS

<b>Sample ID</b>	<b>Cu 63 (ppb)</b>	<b>Fe 57 (ppb)</b>	<b>Ni 60 (ppb)</b>	<b>Zn 66 (ppb)</b>	<b>Pd 106 (ppb)</b>	<b>Ir 193 (ppb)</b>
Standard 1	2.000	2.000	2.000	2.000	2.000	2.000
Standard 2	9.960	9.270	9.992	9.555	9.999	9.924
Standard 3	49.949	47.653	49.998	49.613	50.056	52.032
Standard 4	100.069	95.294	100.367	99.412	109.008	110.906
Standard 5	215.366	192.887	200.236	199.040		
PcCFP	4.947	9.401	0.898	20.488	0.033	0.007