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Supporting information for

# Visible-Light-Induced Cascade Radical Cyclization to Access Sulfamoylated Indole[2, 1-a]isoquinoline Benzo[4,5]imidazo[2,1-*a*]isoquinolin-6(5*H*)-ones

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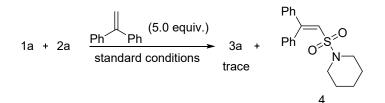
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# **Contents Page**

1. The radical trapping experiments	S3
2. Stern-Volmer Luminescence Quenching Analysis	S3-S5
3. Larger-Scale Experiments	S5
4. <sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>19</sup> F NMR spectra	S6-S52

#### **1.** The radical trapping experiments



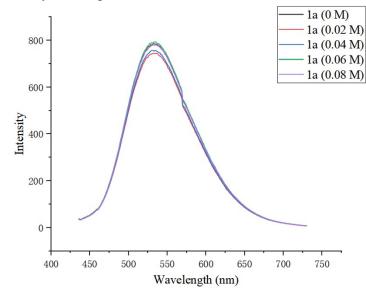
Add **1a** (0.20 mmol), **2a** (0.46 mmol), 1,1-stilbene (1.2 mmol, 4.0 equiv.) to the sealed tube and dry DMAc (1.0 mL), The reaction mixture was evacuated and with pure N<sub>2</sub> for three times and stirred under the irradiation of 36 W blue LEDs at room temperature for 1 h. After irradiation, Then, the mixture was diluted with water (10 mL) and extracted with ethyl acetate (5 mL × 3). The combined organic phase was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure to give a residue which was purified by silica gel column chromatography to afford the compound **3a** (trace) and compound **4** (27 mg, 8% yield). Characteristic data for compound **4**: : <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.41-7.30 (m, 8H), 7.24 (d, J = 7.6 Hz, 2H), 6.63 (s, 1H), 3.05 (t, J = 4.6 Hz, 4H), 1.54-1.49 (m, 6H).

#### 2. Stern-Volmer Luminescence Quenching Analysis

Emission intensities were recorded using an Agilent Cary Eclipse Fluorescence Spectrophotometer. First, the emission intensity of 1a solutions was observed at 532 nm. The solutions were irradiated at 426 nm (Maximum absorption wavelength of 1a) and fluorescence was measured from 436 nm to 730 nm. The solution of 4-CzIPN-Br (1 mM, 10 mL), 1a (1 M, 10 mL) and 2a (1 M, 10 mL) were prepared in air.

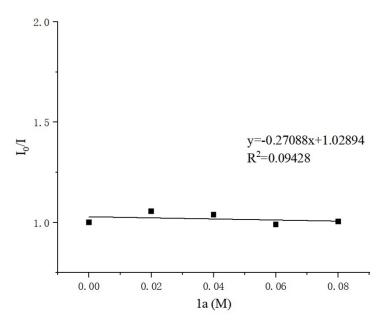
**For Experiment 1:** Constant photocatalyst; Varied 2-methyl-1-(2-phenyl-1H-benzo[d]imidazol-1-yl)prop-2-en-1-one.

Add 30  $\mu$ L 4-CzIPN-Br solution and 0  $\mu$ L, 60  $\mu$ L, 120  $\mu$ L, 180  $\mu$ L, 240  $\mu$ L 1a solution respectively in the quartz cuvette, then diluted the solution to 3 mL.



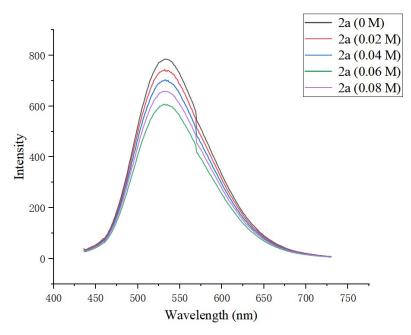
Supplementary figure 1: Fluorescence spectrum of PC + 1a.

Comment: 1a does not react with the excited photocatalyst.



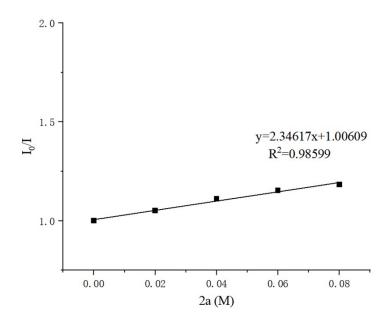
Supplementary figure 2: Stern-Volmer Luminescence Quenching Analysis of PC + 1a.

For Experiment 2: Constant photocatalyst; Varied piperidine-1-sulfonyl chloride. Add 30  $\mu$ L 4-CzIPN-Br solution and 0  $\mu$ L, 60  $\mu$ L, 120  $\mu$ L, 180  $\mu$ L, 240  $\mu$ L 1a solution respectively in the quartz cuvette, then diluted the solution to 3 mL.



Supplementary figure 3: Fluorescence spectrum of PC + 2a.

Comment: Only 2a could quench the photocatalyst.



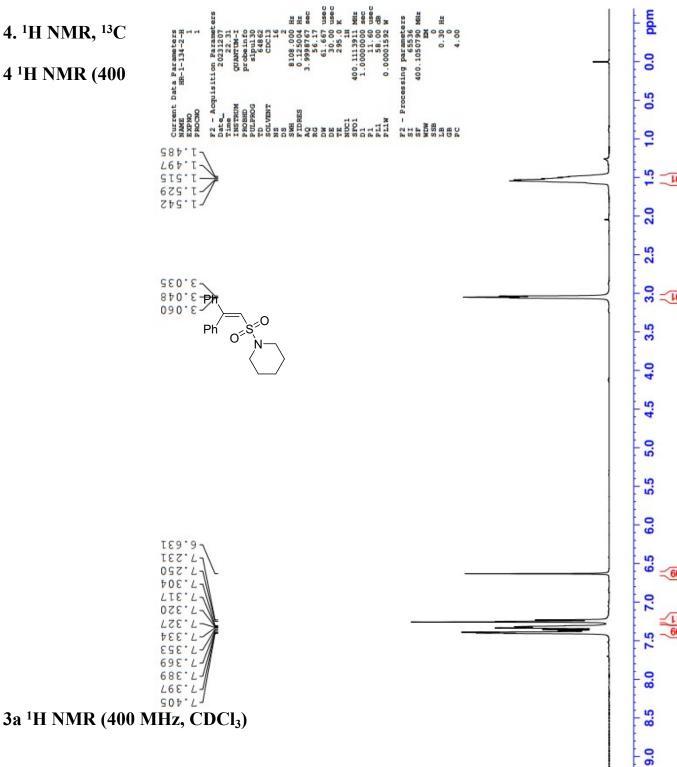
Supplementary figure 4: Stern-Volmer Luminescence Quenching Analysis of PC + 2a.

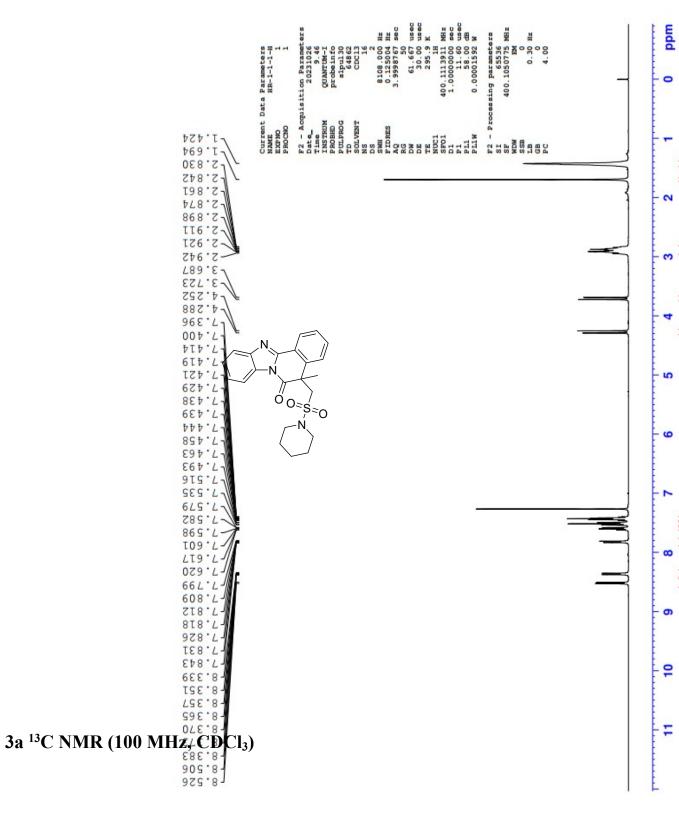
#### 3. Larger-Scale Experiments

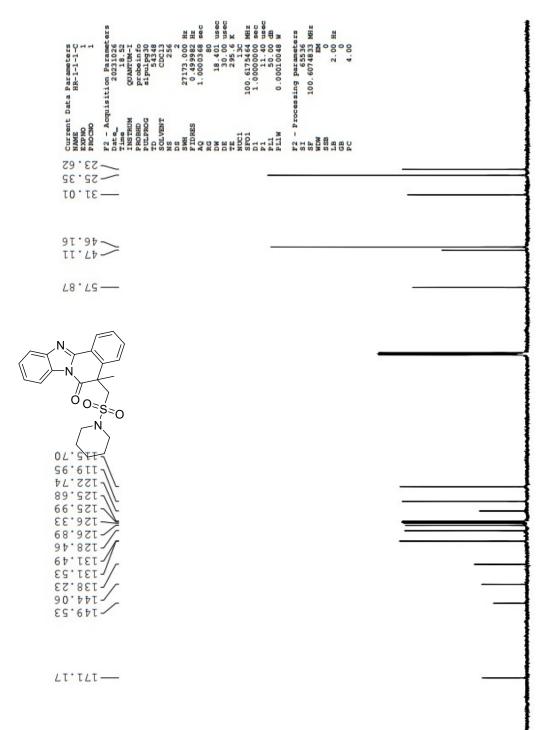
A frame-dried 100 mL Schlenk flask with a magnetic stirring bar was charged with 2methyl-1-(2-phenyl-1H-benzo[d]imidazol-1-yl)prop-2-en-1-one (1a) (1.05 g, 4.0 mmol), 4-CzIPN-Br (28.4 mg, 0.02 mmol) and dry DMA (20.0 mL) was added via syringe under nitrogen. Then piperidine-1-sulfonyl chloride (2a) (1.69 g, 9.2 mmol) was added and the reaction mixture was stirred under irradiation by a 36 W blue LEDs at 25 °C for 2.5 hours under nitrogen. The reaction mixture diluted with water (200 mL) and extracted with ethyl acetate (100 mL) for three times. The combined organic phase was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated to give a residue which was purified by silica gel chromatography, eluting with petroleum ether/ethyl acetate, to give compound 4a as a light yellow solid (1.60 g, 98%).



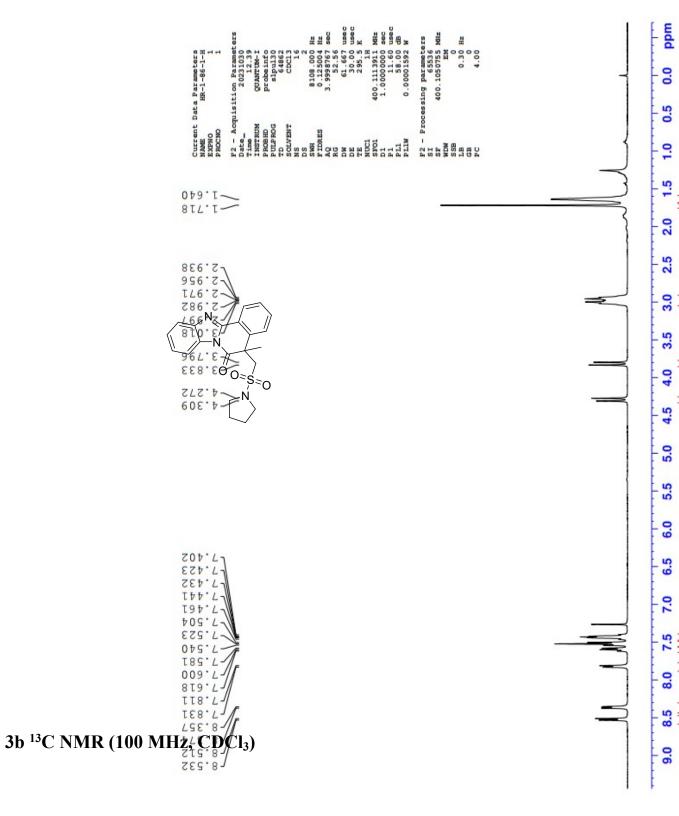
## 4<sup>1</sup>H NMR (400

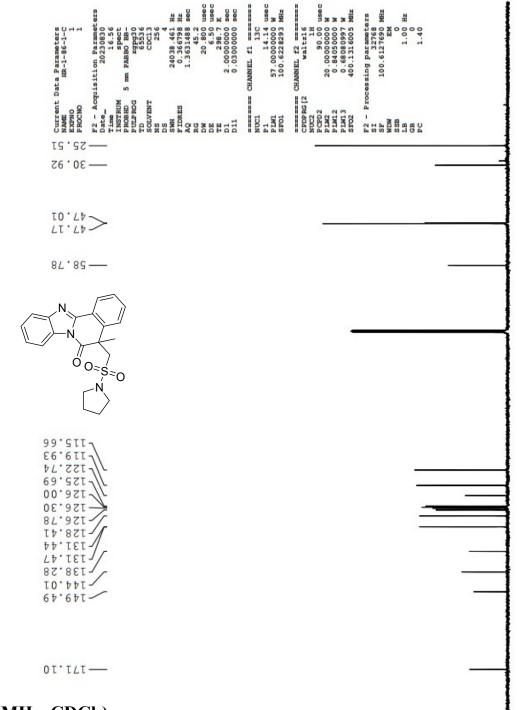




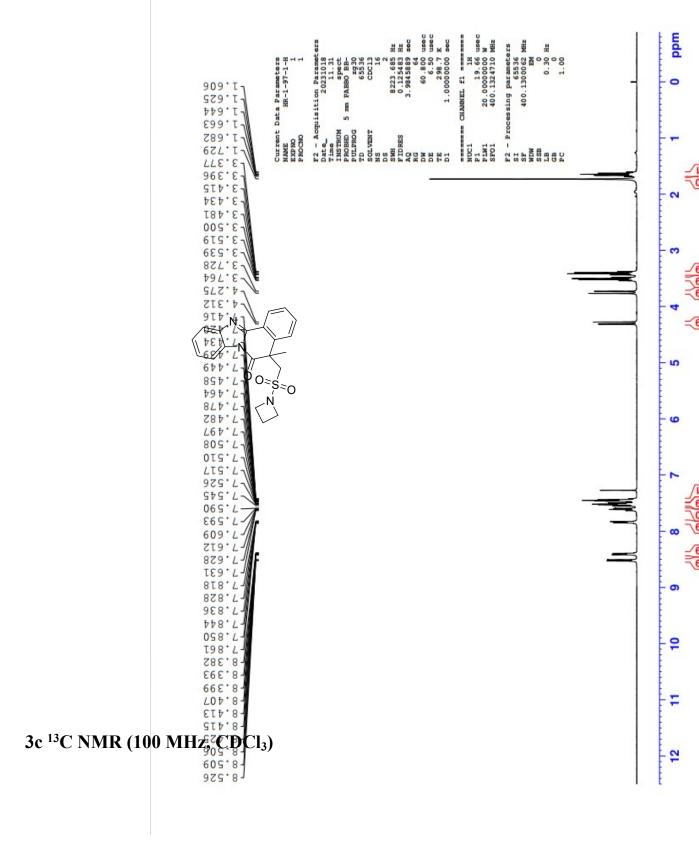


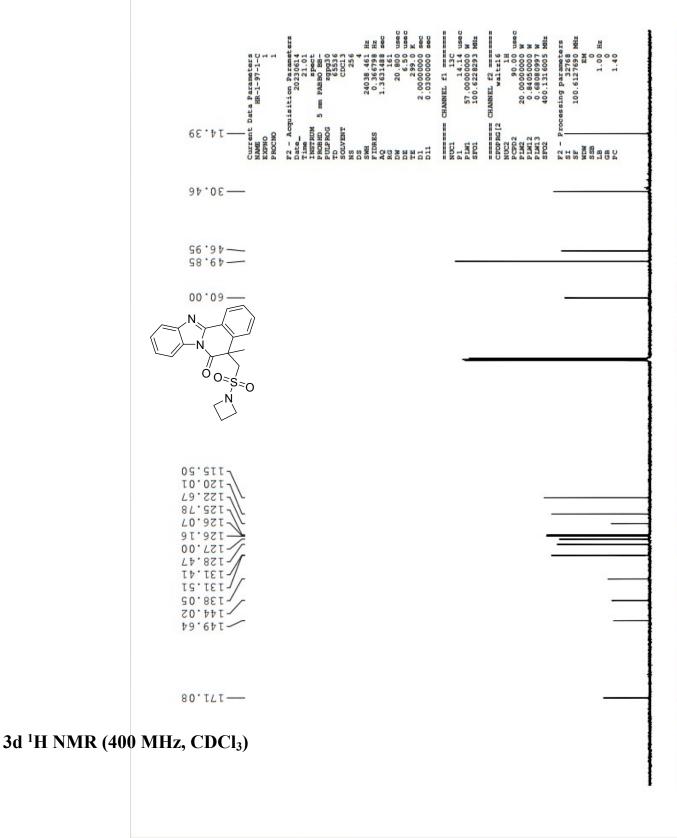
3b <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

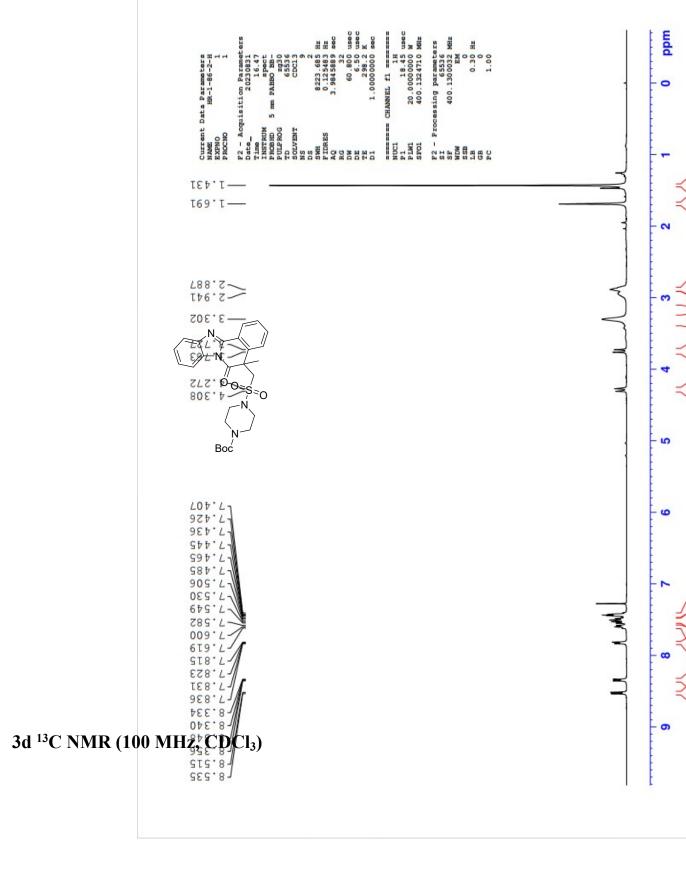


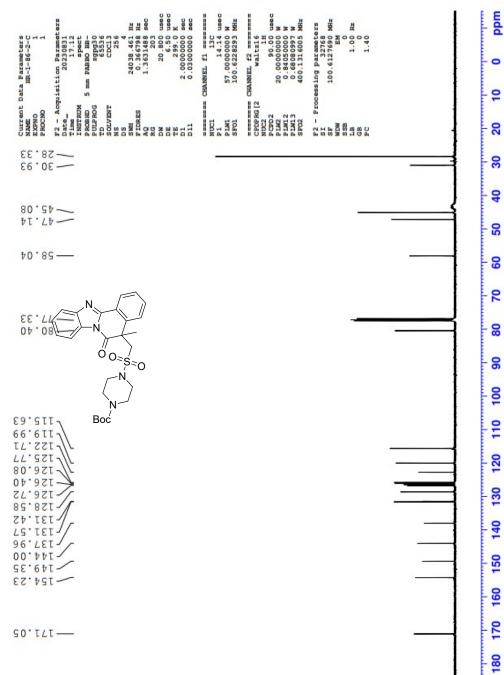


# 3c<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



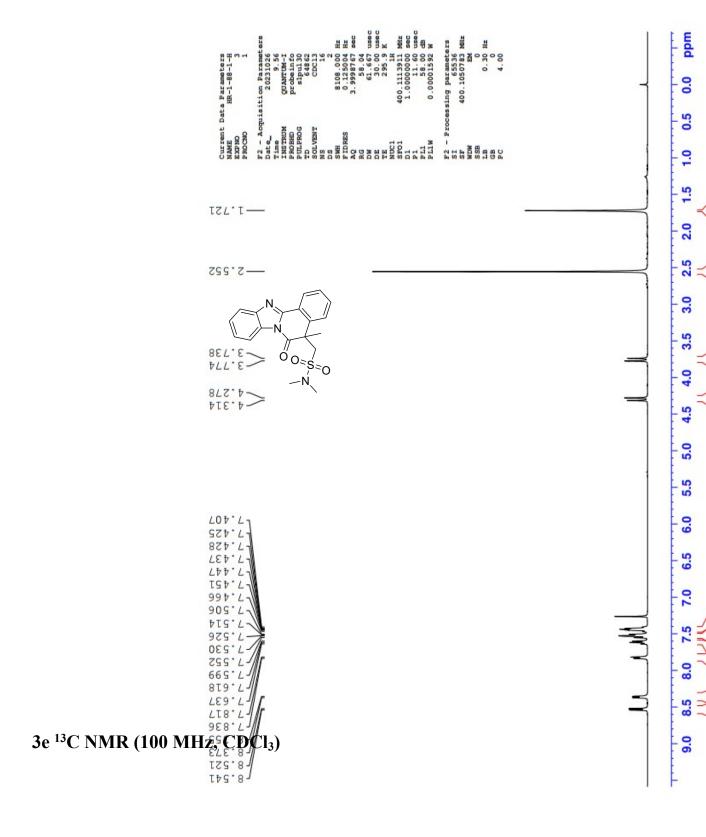


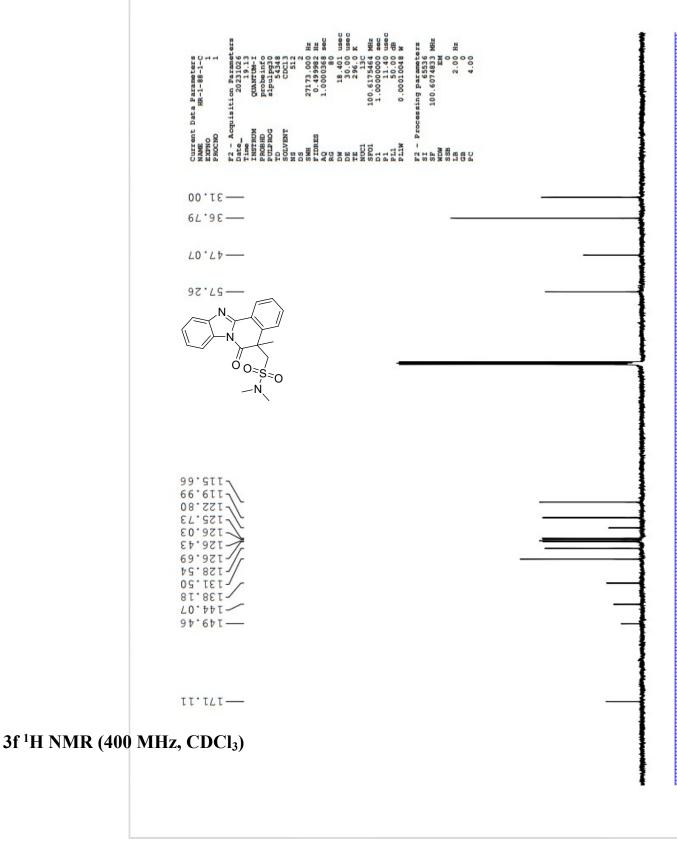


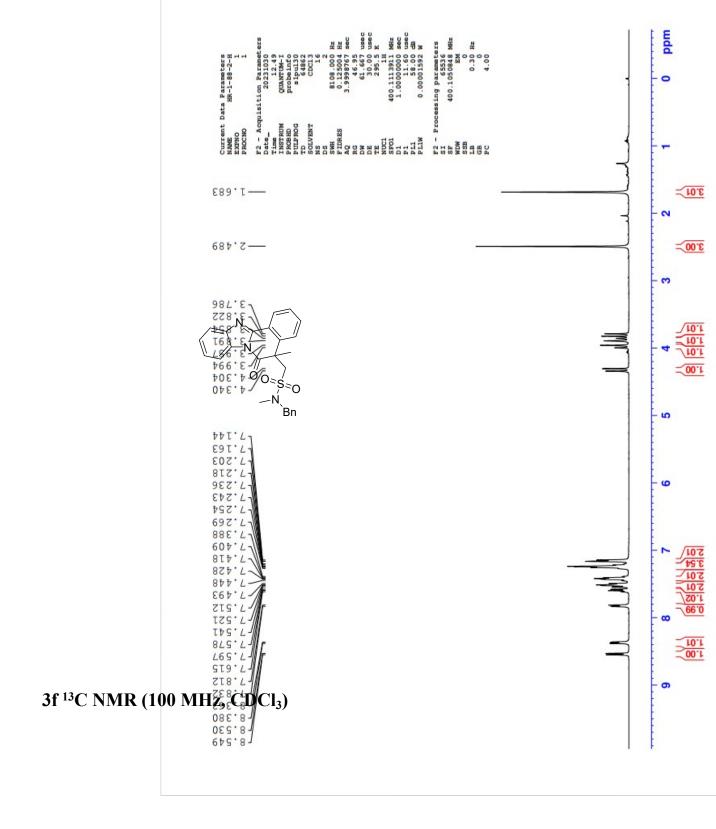


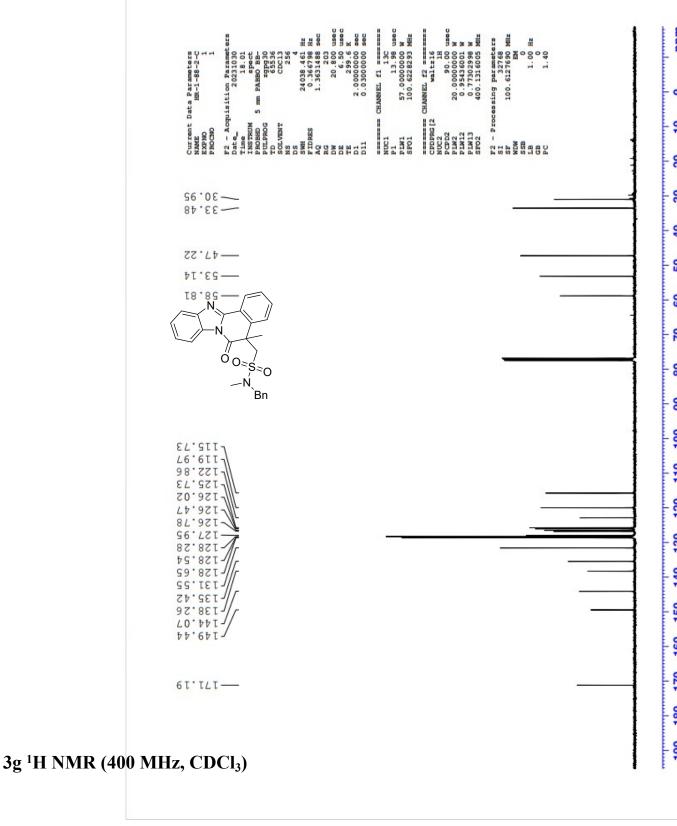
mdd

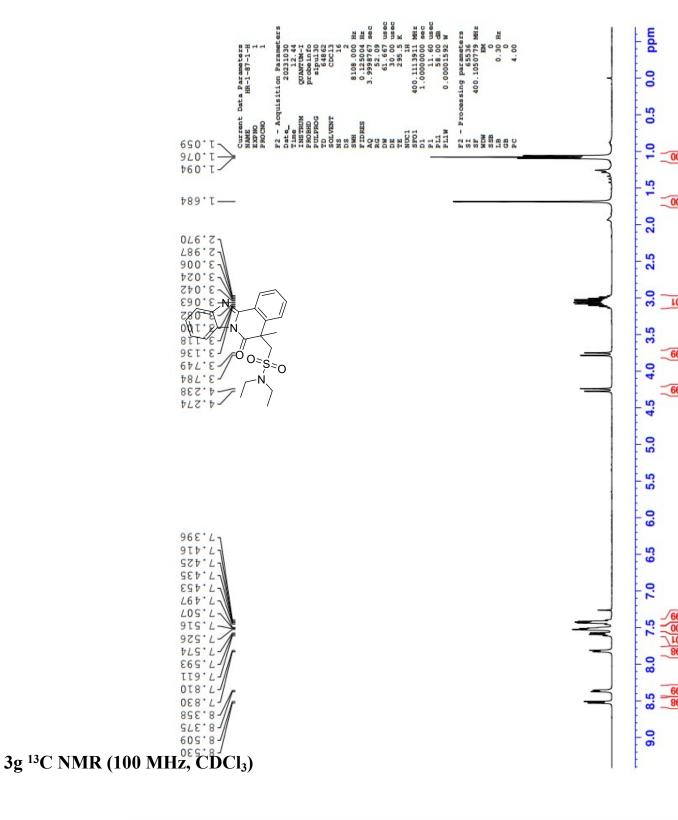
3e <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

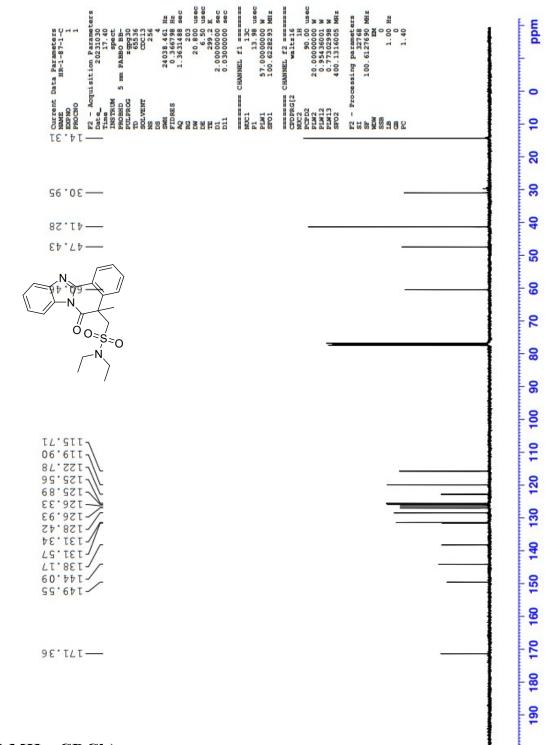




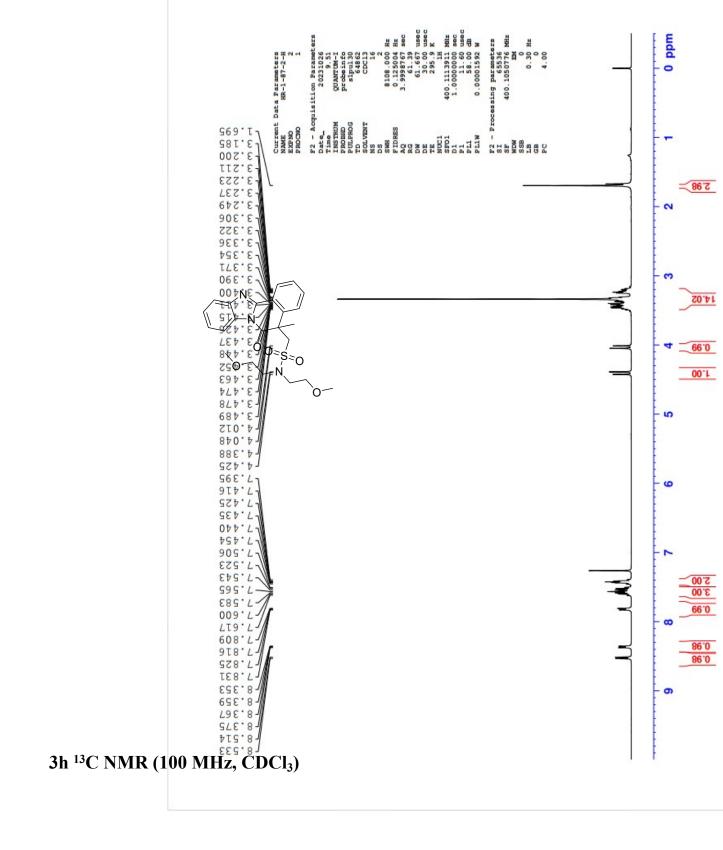


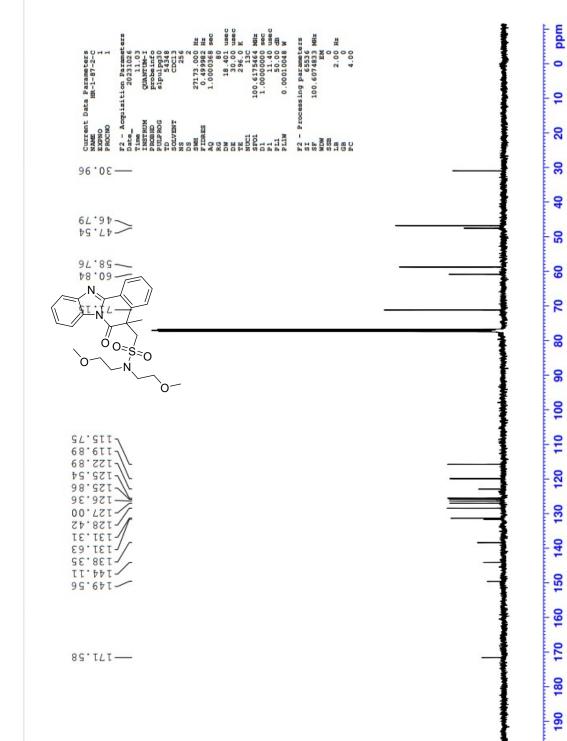




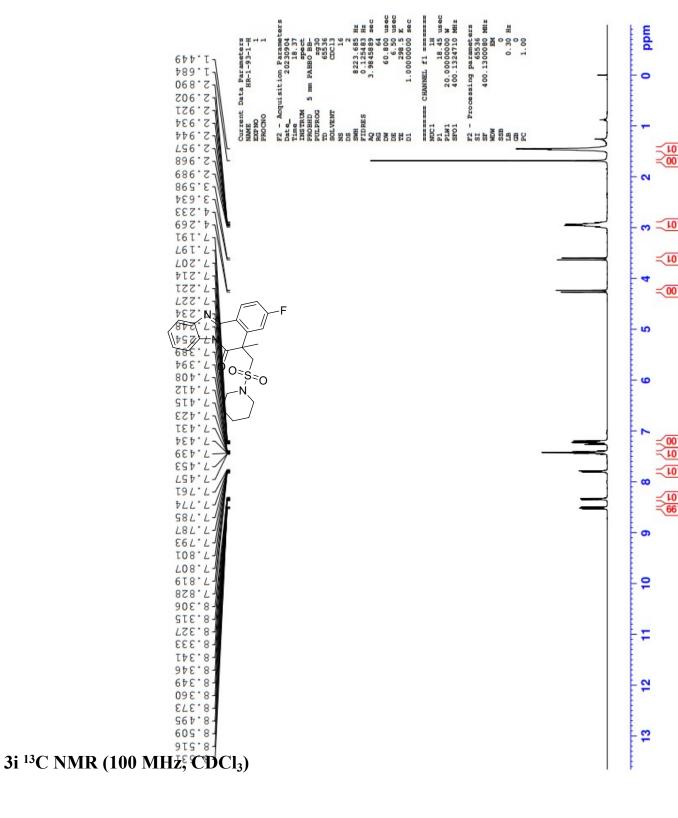


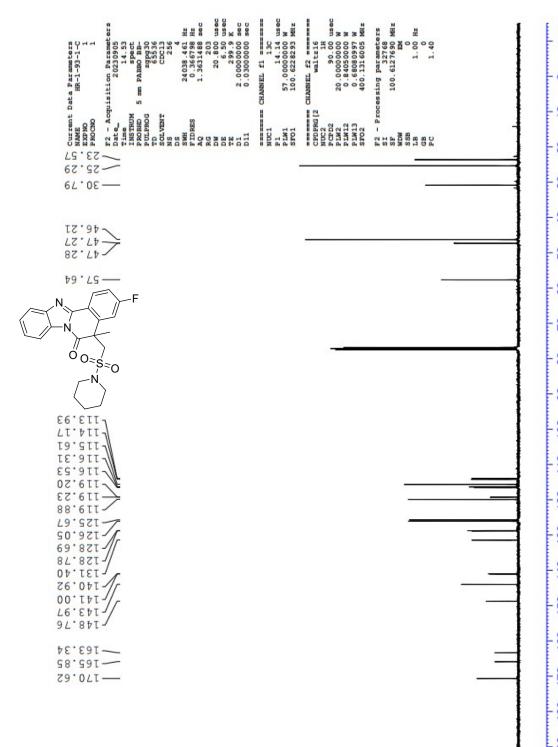
### 3h <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)





3i <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



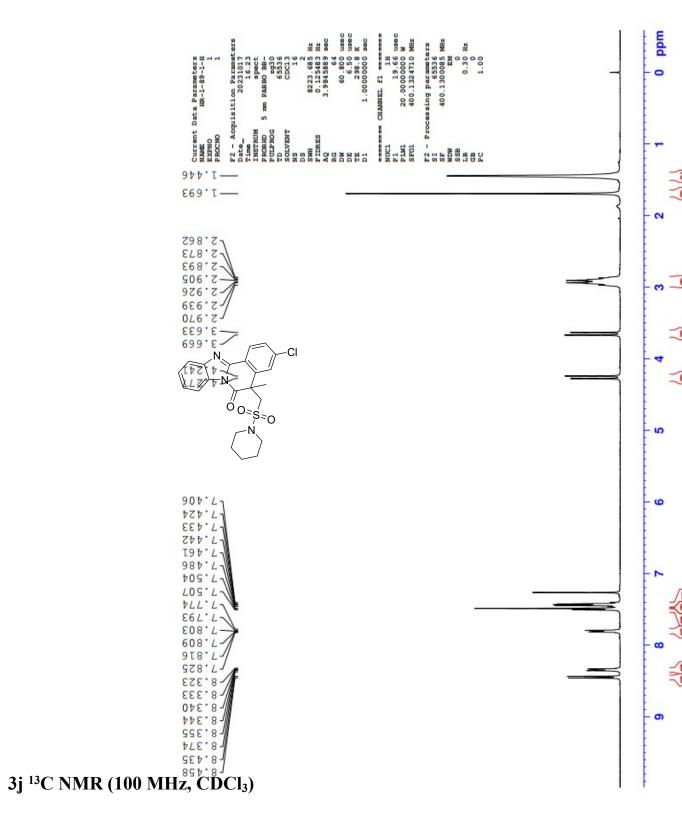


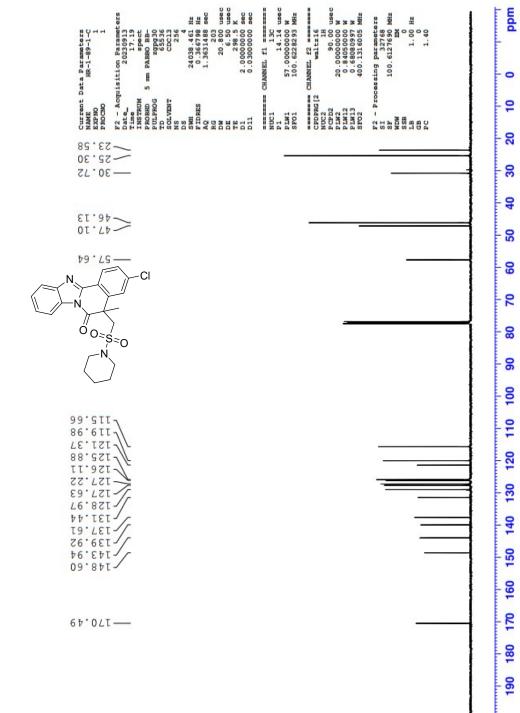
3i<sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)

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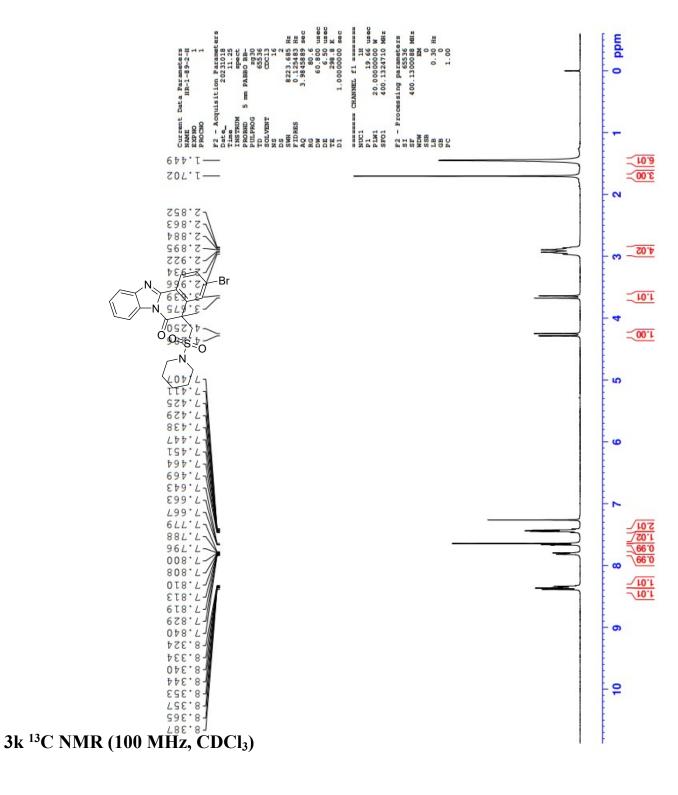
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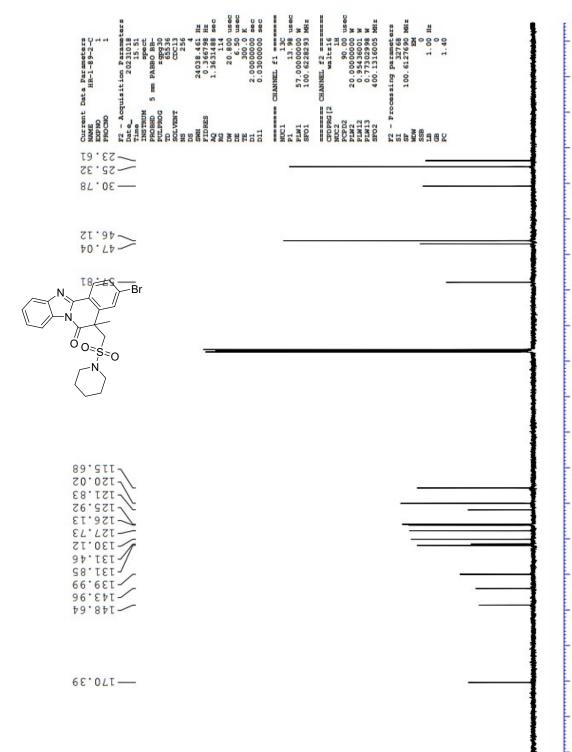
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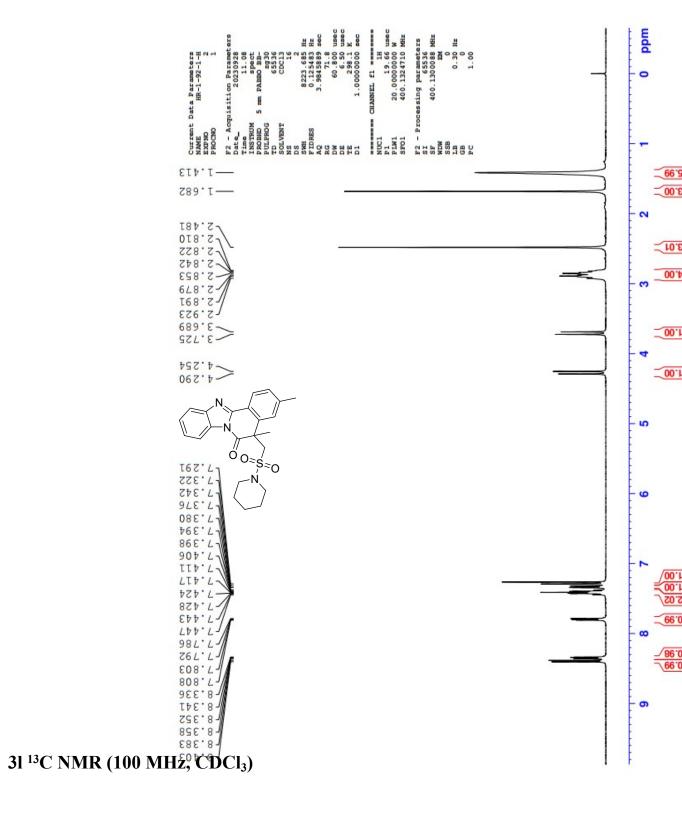


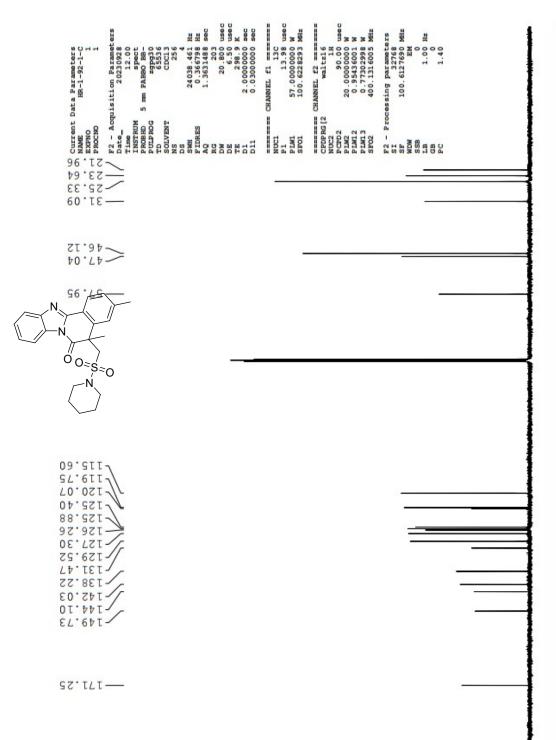
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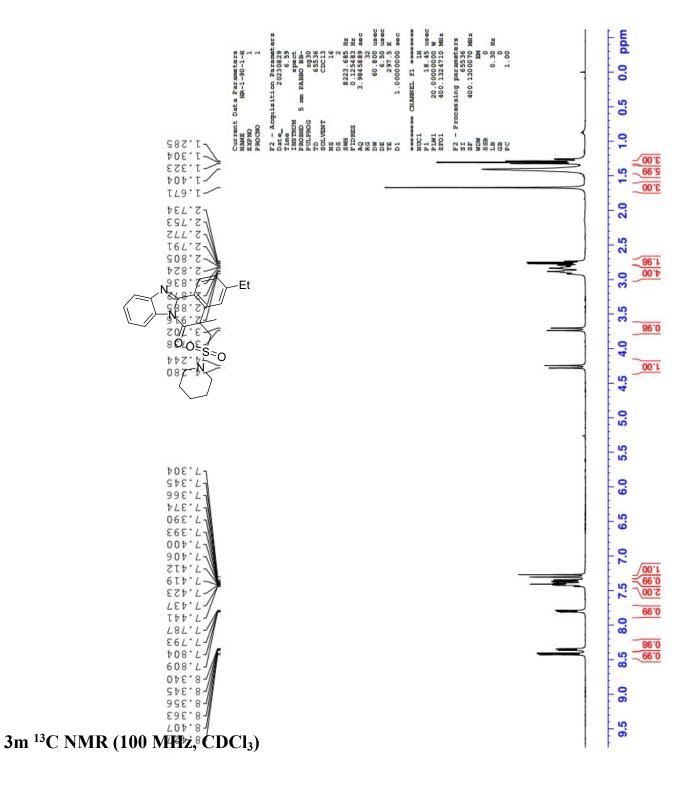


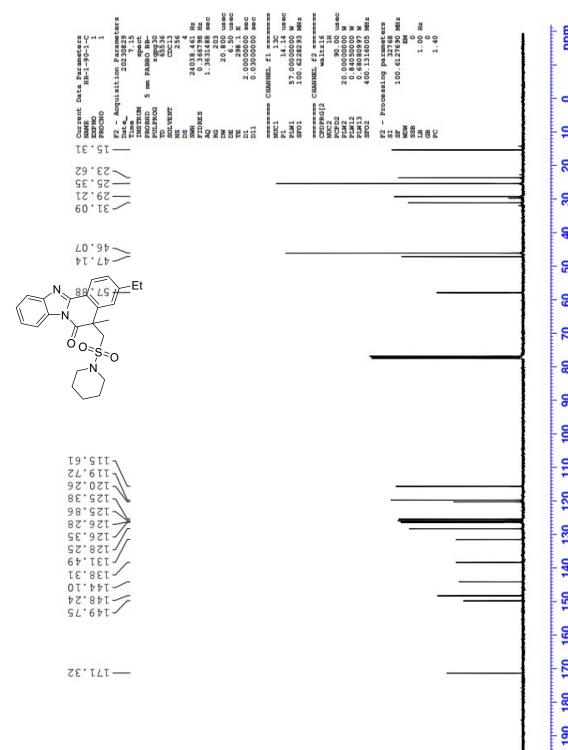
3l <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



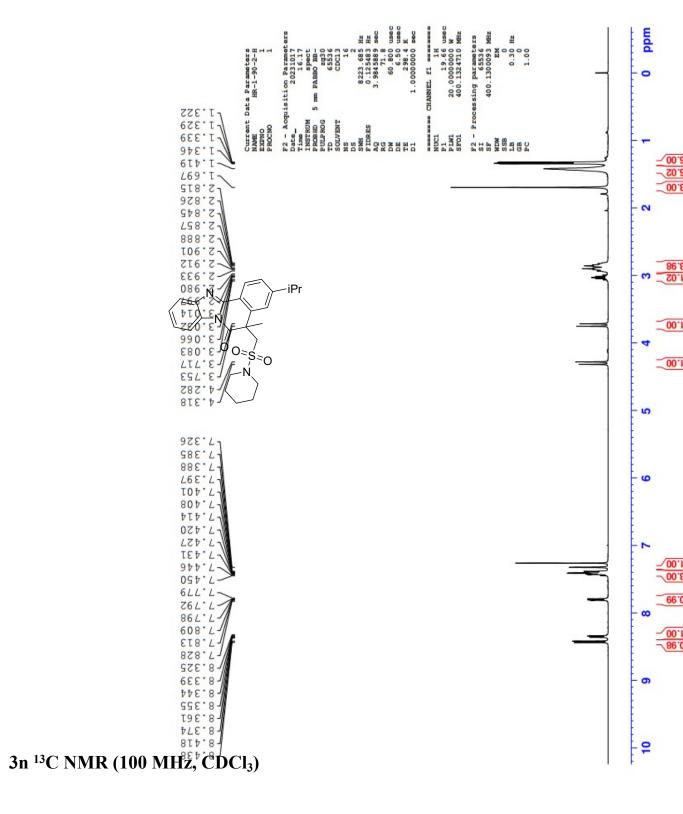


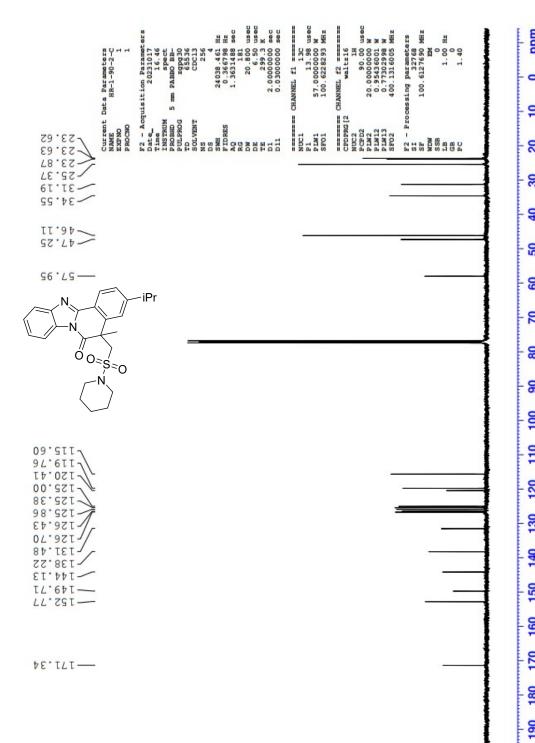
3m <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)





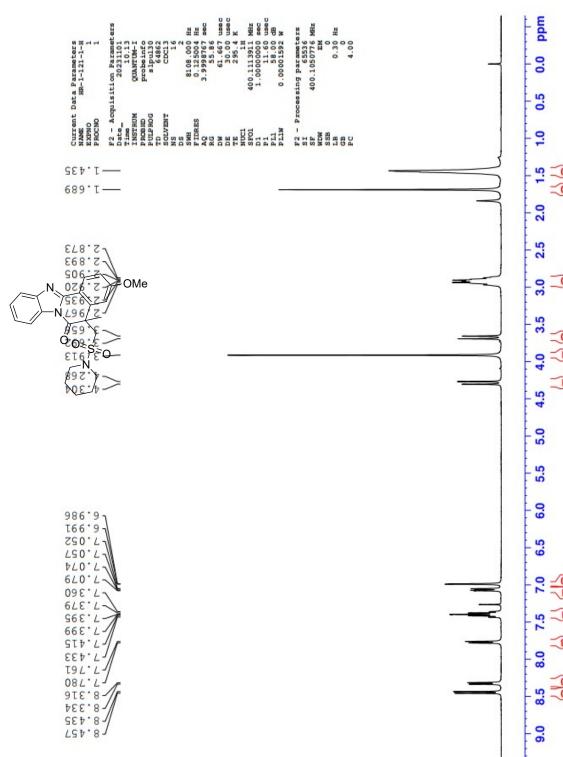
3n <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



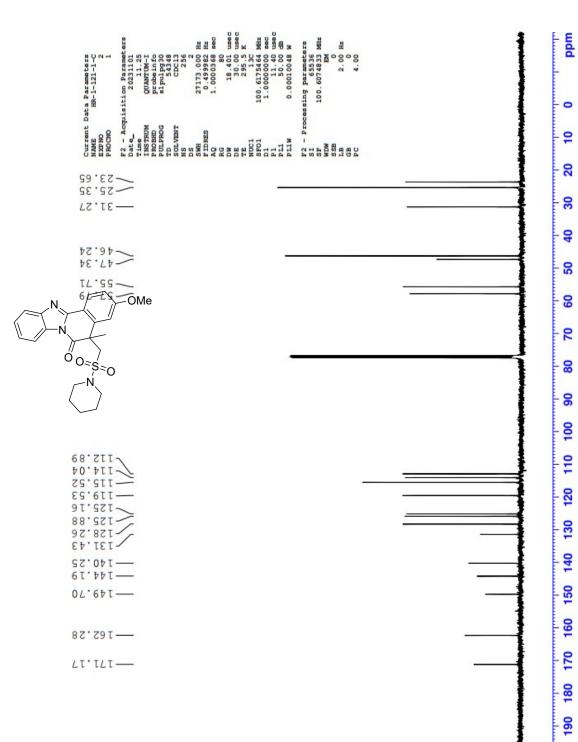


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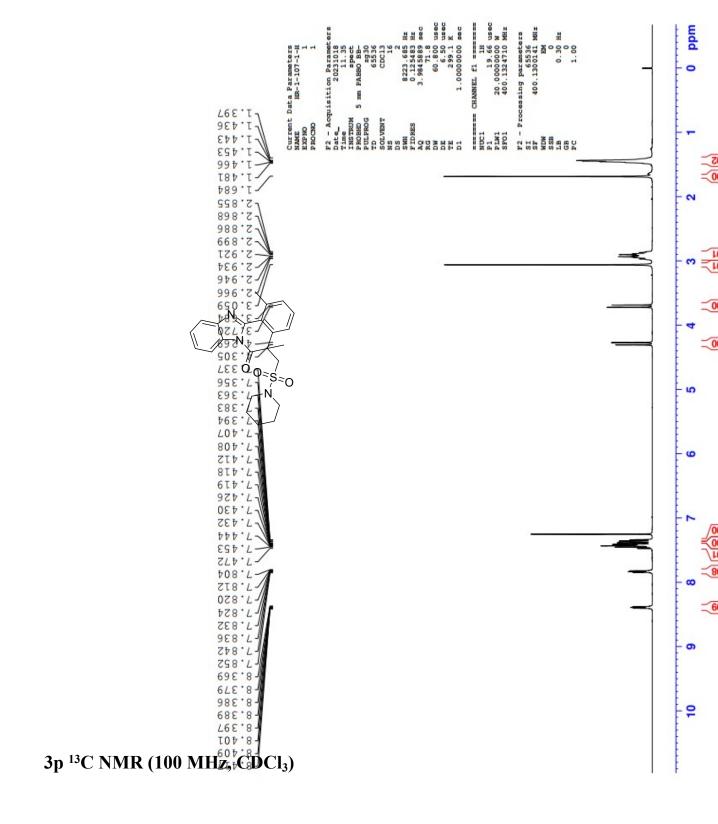
30 <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

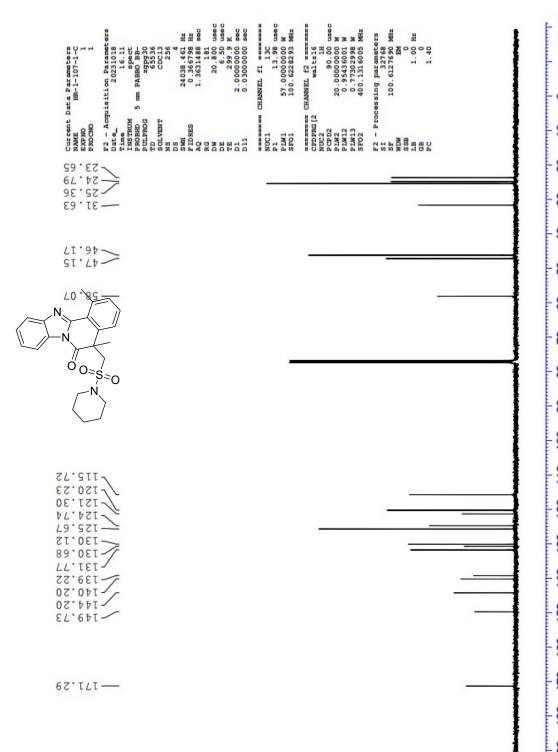


## 30 <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)

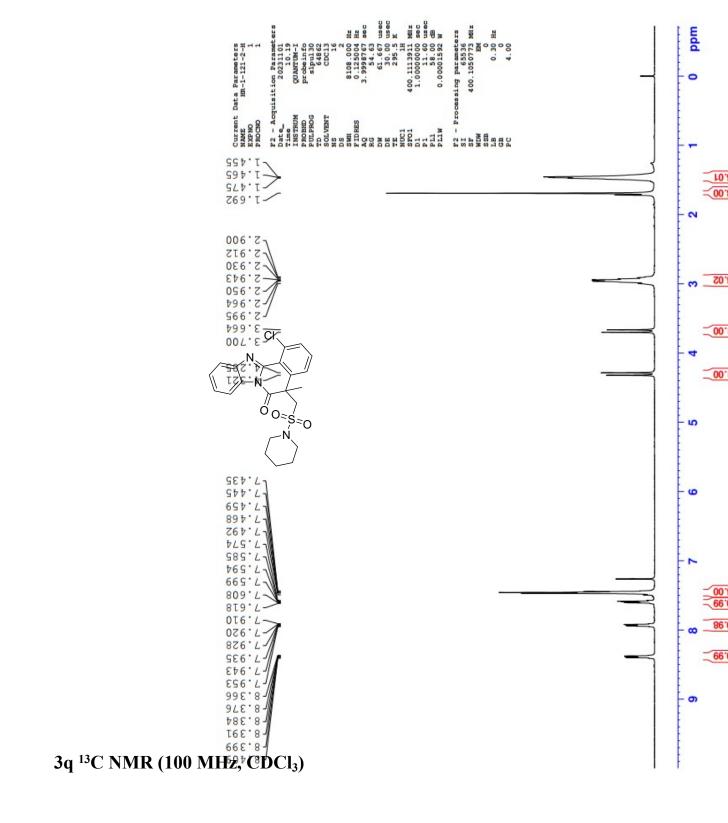


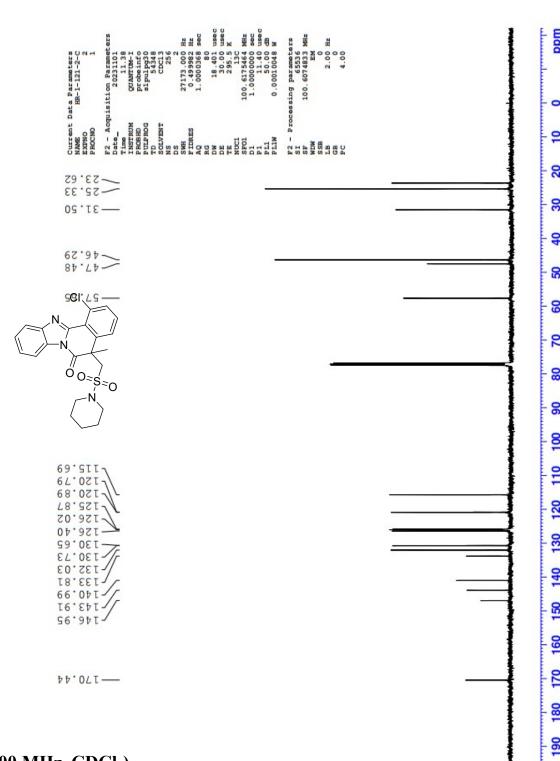
3p <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



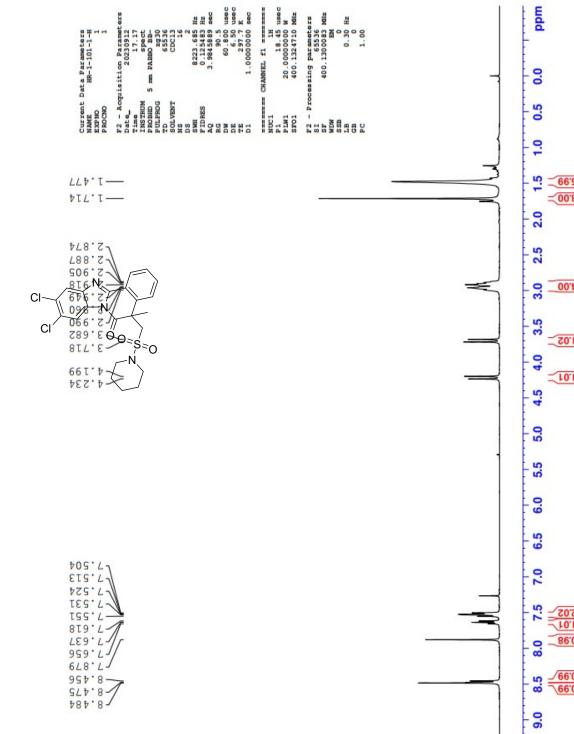


3q <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

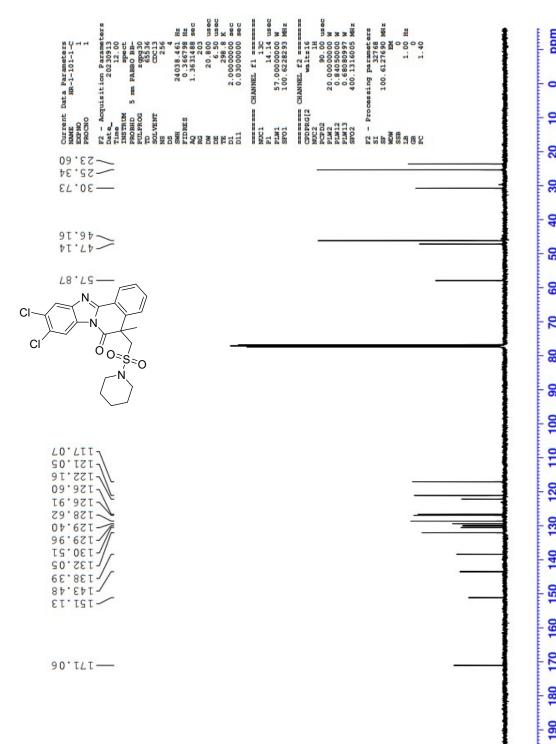




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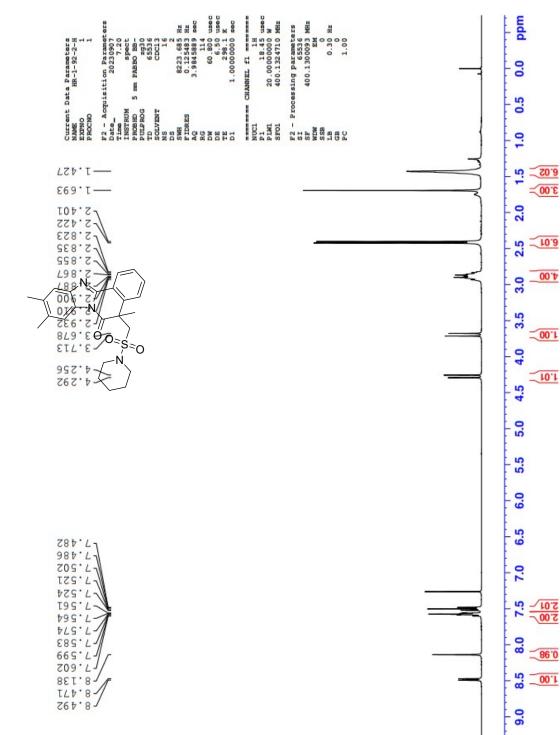


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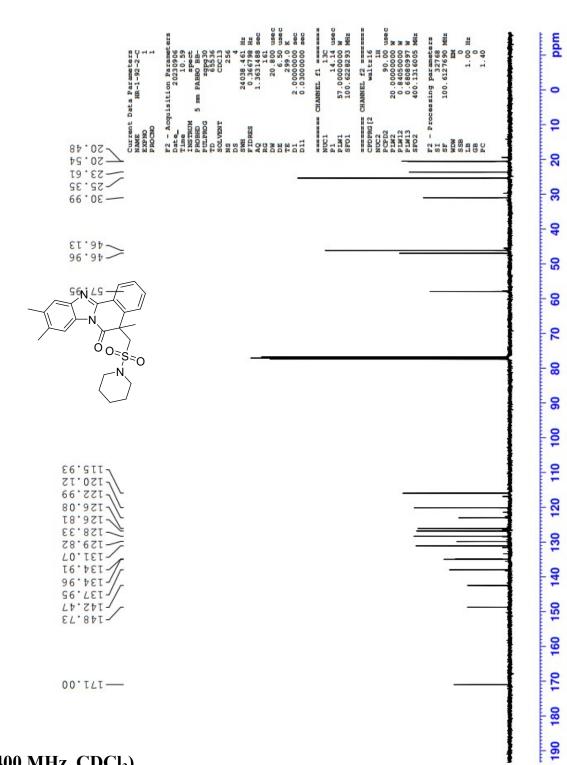


naa

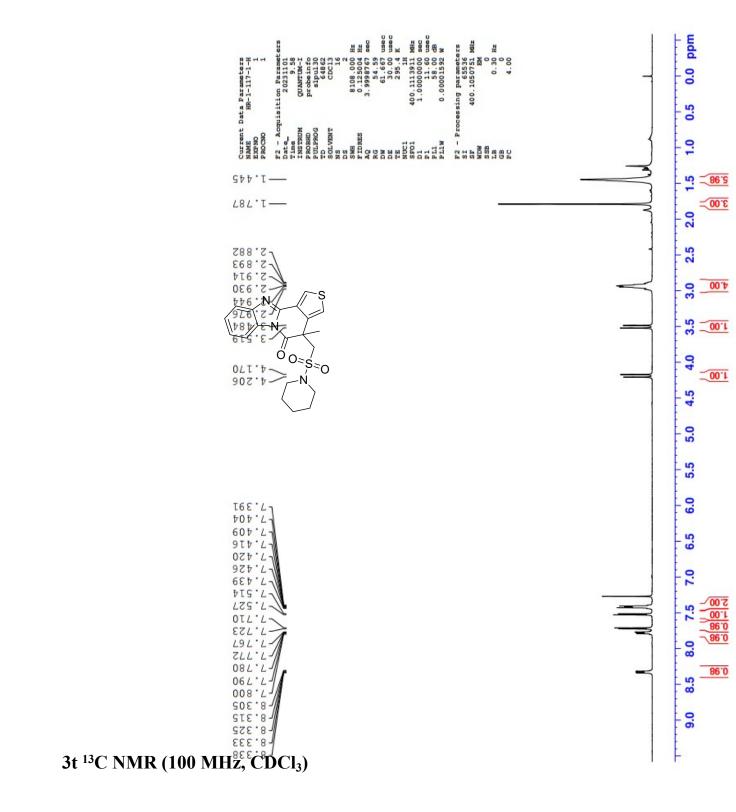
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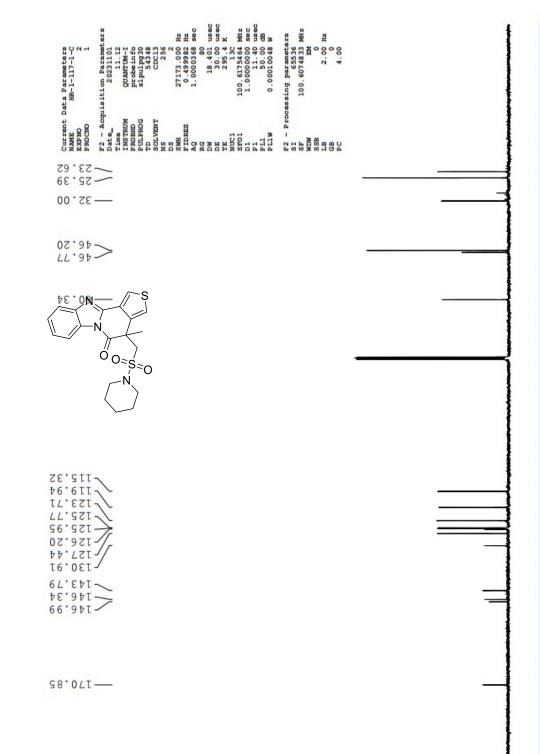


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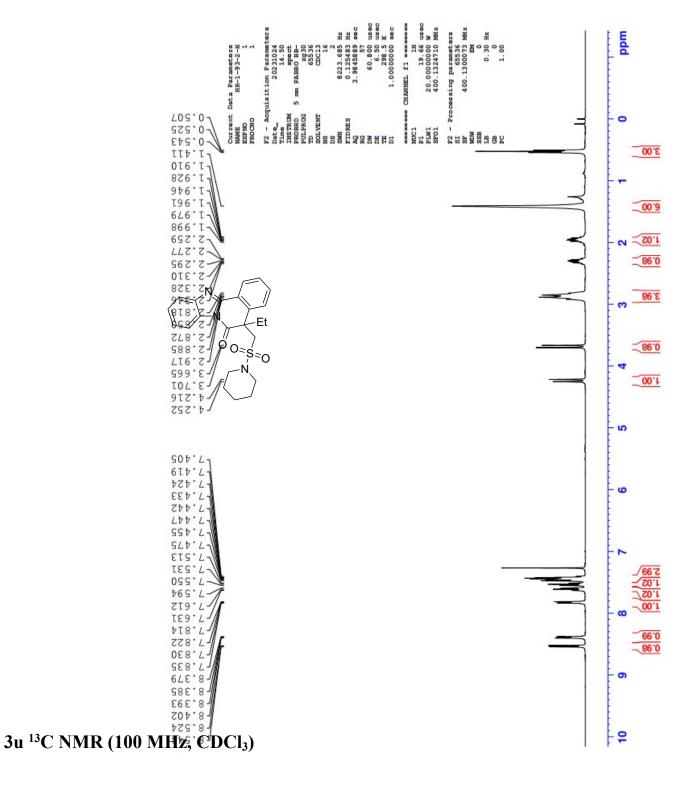


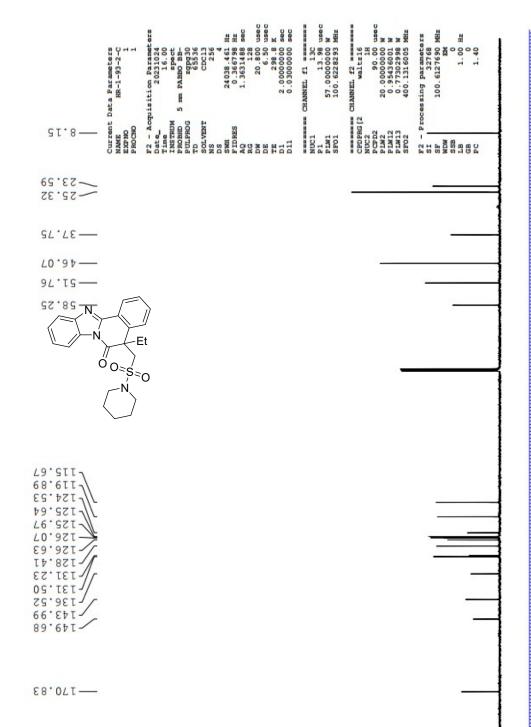
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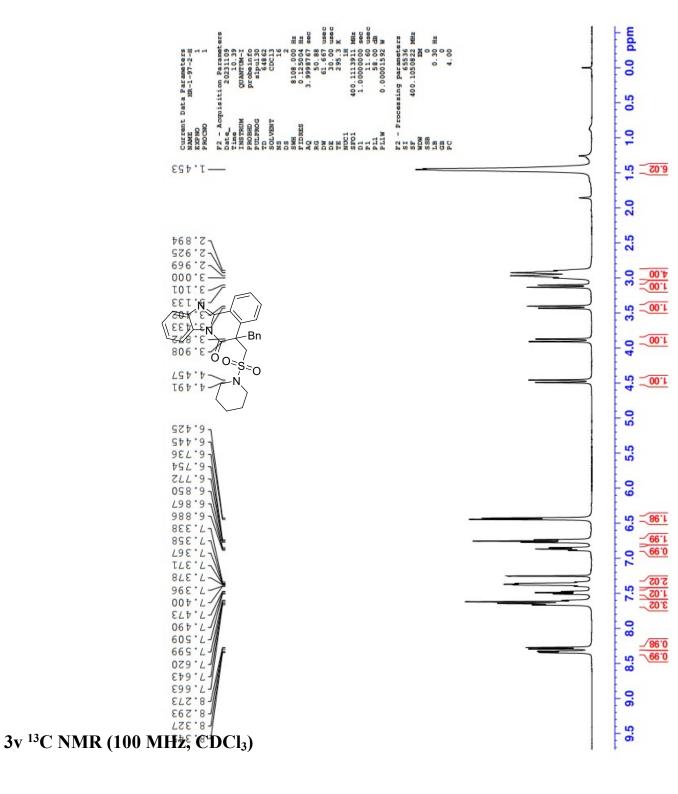


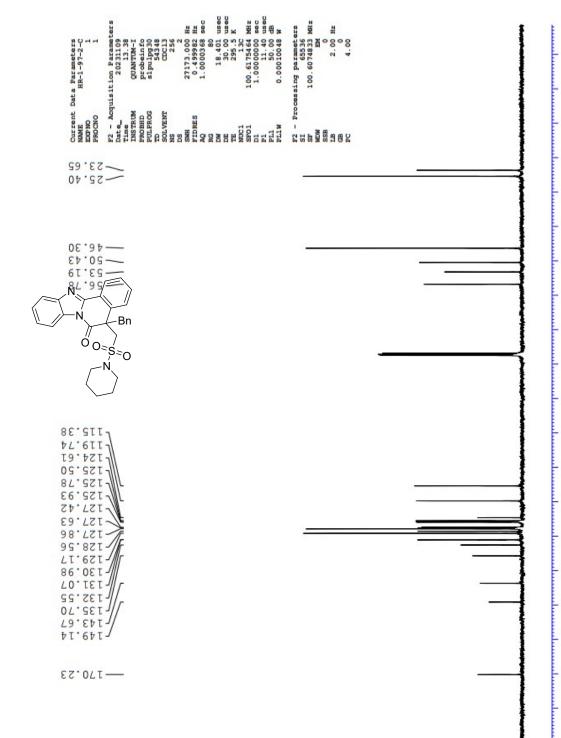
3u <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)





3v <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)





S51