

## Supplementary Information for Manuscript Entitled

### Stereoselective ring-opening polymerization of *rac*-lactide catalyzed by proton sponge bisphosphazene of 1,8- bis(hexamethyltriaminophosphazenylnaphthalene

Zhenqing Zhang,<sup>a</sup> Xiaoyu Guo,<sup>a</sup> Xinhui Kou,<sup>b</sup> Na Zhao,<sup>a,\*</sup> and Zhibo Li<sup>a,c,\*</sup>

<sup>a</sup> Key Laboratory of Biobased Polymer Materials, Shandong Provincial Education Department, College of Polymer Science and Engineering, Qingdao University of Science and Technology, Qingdao, Shandong 266042, China

<sup>b</sup> Analyses and Testing Center, Qingdao University of Science and Technology, Qingdao, Shandong 266042, China

<sup>c</sup> College of Chemical Engineering, Qingdao University of Science and Technology, Qingdao, Shandong 266042, China

\*Corresponding Author: E-mail: zhaona@qust.edu.cn; zbli@qust.edu.cn

#### Experimental Section

##### Materials.

Toluene and tetrahydrofuran (THF) were purified by purging with dry nitrogen, followed by passing through columns of activated alumina. *Rac*-LA was purchased from Macklin Inc. and recrystallized twice from toluene. Benzyl alcohol (BnOH), purchased from Shanghai Aladdin Biochemical Technology Co., Ltd, was stirred with CaH<sub>2</sub> at room temperature for 24 hours and distilled under reduced pressure prior to use. Other chemicals were purchased from commercial suppliers and used without further purification unless otherwise noted. HMPN,<sup>1</sup> U-1/U-2/U-3/U-4,<sup>2, 3</sup> and SQ-1/SQ-2<sup>4</sup> were synthesized according to the procedures reported before.

##### General Considerations.

All moisture/oxygen sensitive reactions/compounds were operated using standard

Schlenk technique or glovebox technique in an atmosphere of high-purity nitrogen. Nuclear magnetic resonance (NMR) spectra were recorded on a Bruker AVANCE NEO 400 MHz NMR spectrometer (400 MHz for  $^1\text{H}$  NMR) at 298 K. Chemical shifts were reported in  $\delta$  (ppm) and were referenced to tetramethylsilane (0.00 ppm). Gel permeation chromatography (GPC) experiments were performed on an Agilent HPLC system equipped with a model 1260 Hip degasser, a model 1260 Iso pump and a model 1260 differential refractometer detector with using THF as mobile phase at a flow rate of 1.0 mL/min at 40 °C. One PLgel5  $\mu\text{m}$  guard column and three Mz-Gel SD<sub>plus</sub> columns ( $10^3$  Å,  $10^4$  Å, and  $10^5$  Å, linear range of  $M_w = 1000 - 2 \cdot 10^6$  Da) were connected in series. The molecular weight and dispersity were calculated using polystyrene standards with narrow molecular weight distribution as references. The sample concentration used for GPC analysis was about 5 mg/mL. Matrix-assisted laser desorption/ionization time-of-flight mass spectroscopy (MALDI-TOF MS) analyses were conducted on a Bruker Microflex MALDI-TOF MS spectrometer equipped with a 337 nm nitrogen laser. The polymer was dissolved in THF, and *trans*-2-[3-(4-*tert*-butylphenyl)-2-methyl-2-propenylidene]-malononitrile (DCTB) was used as the matrix and  $\text{CF}_3\text{COONa}$  as the ionization agent. Differential scanning calorimetry (DSC) was performed using a TA differential scanning calorimeter DSC 25 that was calibrated using high purity indium. Measurements were performed under  $\text{N}_2$  atmosphere with a flow rate of 50 mL/min. Each sample with a mass of 5 - 10 mg was used for the measurement.

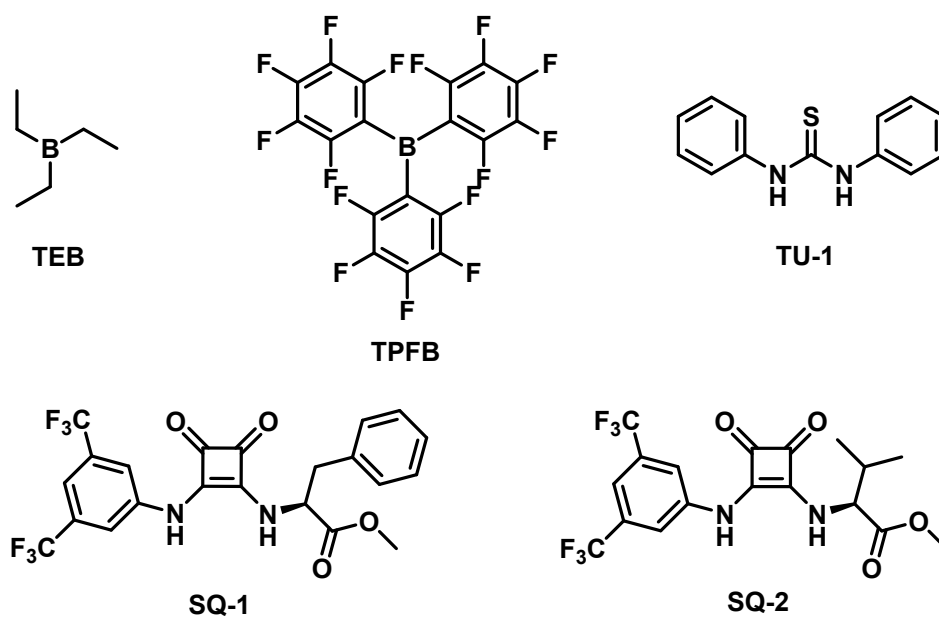
**General procedure for ROP of *rac*-LA catalyzed by HMPN (Table 1, run 2).**

In a glove-box, HMPN (4.2 mg, 0.01 mmol) and BnOH (1.0  $\mu\text{L}$ , 0.01 mmol) were dissolved in 1 mL of toluene and stirred for 10 min in a Schlenk tube. Then *rac*-LA (144 mg, 1.0 mmol) was added into the mixture to start the polymerization at room temperature (25 °C). After the desired polymerization time, acetic acid was added to quench the reaction and a small part of solution was taken and analyzed by  $^1\text{H}$  NMR to determine the conversion. The conversion of *rac*-LA monomer was calculated based on the integration (I) ratio  $I_{\text{PLA}}/[I_{\text{PLA}} + I_{\text{rac-LA}}]$  of the methine proton, which is at

5.18 ppm for PLA polymer and 4.87 ppm for *rac*-LA monomer. The other solution was poured into cold methanol (100 mL) to precipitate polymer, which was dried in a vacuum oven overnight.

**General procedure for ROP of *rac*-LA catalyzed by HMPN/cocatalyst (Table 2, run 2).**

In a glove-box, *rac*-LA (144 mg, 1.0 mmol) was dissolved in toluene (0.5 mL) in a Schlenk tube. In a second tube, HMPN (4.2 mg, 0.01 mmol), BnOH (1.0  $\mu$ L, 0.01 mmol), U-2 (6.5 mg, 0.03 mmol) were mixed in toluene (0.5 mL) and stirred for 2 min. The latter mixture was then added into *rac*-LA solution to start the polymerization at room temperature (25 °C). After the desired polymerization time, acetic acid was added to quench the reaction and a small part of solution was taken and analysed by  $^1\text{H}$  NMR to determine the conversion.

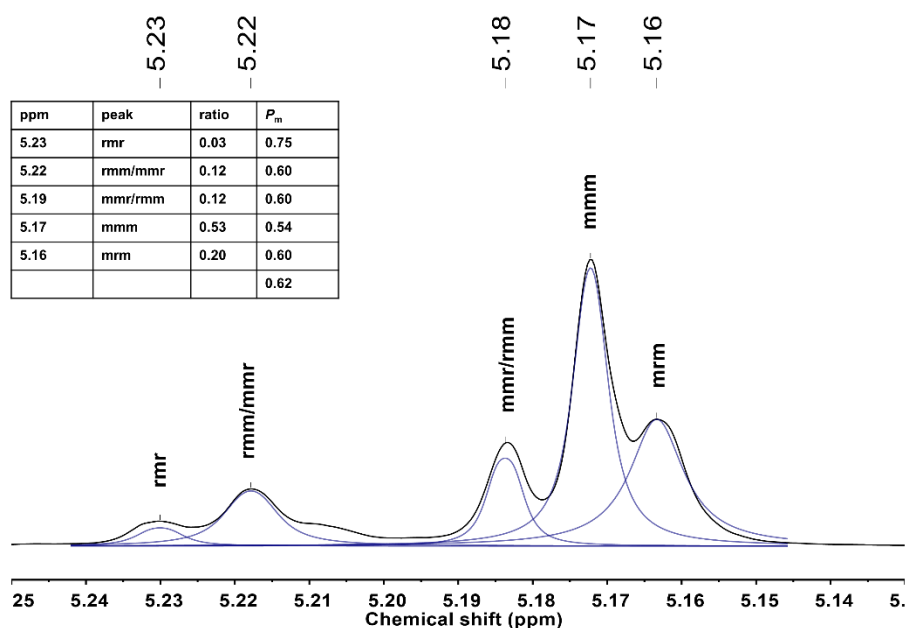


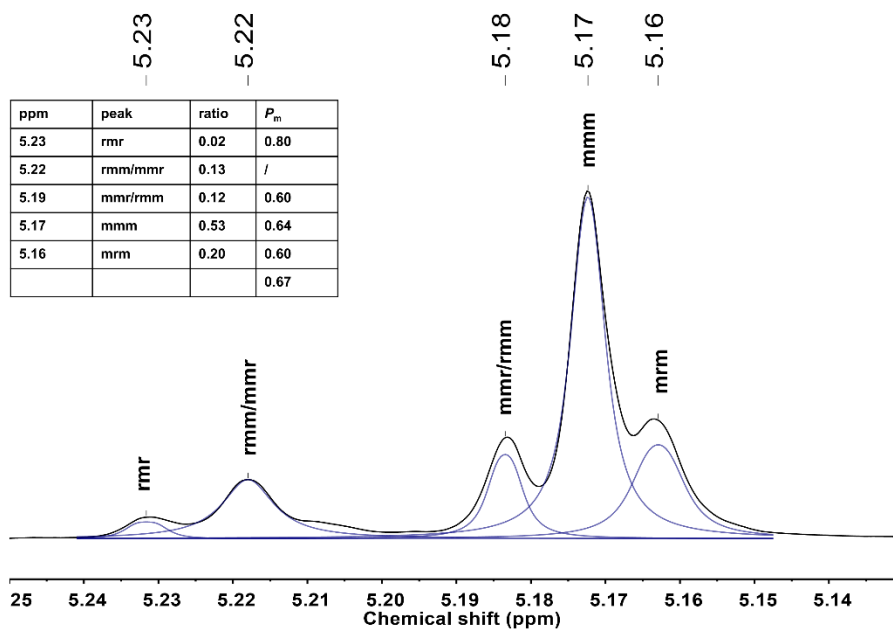
**Scheme S1.** Chemical structures of the cocatalysts

**Table S1.** ROP of *rac*-LA catalyzed by binary catalytic system<sup>a</sup>

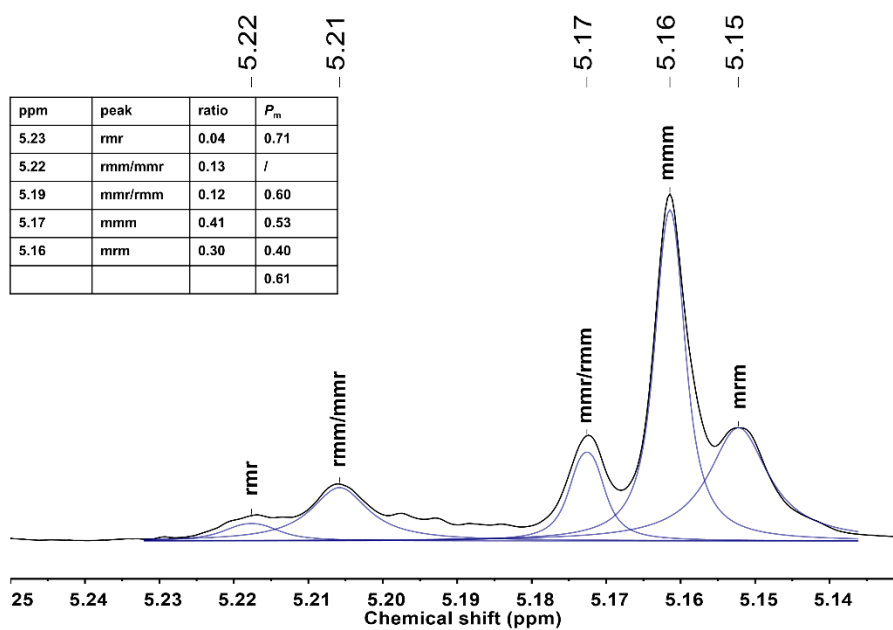
run	M/B/I/C <sup>b</sup>	Cocatalyst	Solvent	Time (min)	Conv. <sup>c</sup> (%)	$M_n^d$ (kg/mol)	$\bar{D}^d$	$P_m^e$
1	100/1/1/2	TEB	Toluene	60	95	3.1	1.26	0.66
2	100/1/1/1	TPFB	Toluene	720	0	n.d.	n.d.	n.d.
3	100/1/1/3	SQ-1	THF	20	8	21.6	1.36	0.67
4	100/1/1/3	SQ-2	THF	20	5	22.0	1.33	0.68
5	100/1/1/3	TU-1	toluene	10	88	21.2	1.09	0.76

<sup>a</sup>Conditions:  $[rac\text{-LA}]_0 = 1.0$  mol/L, the polymerizations were carried out at room temperature using HMPN as catalyst. <sup>b</sup>M/B/I/C indicates the mole ratio of  $[rac\text{-LA}]_0/[HMPN]_0/[BnOH]_0/[cocatalyst]_0$ . <sup>c</sup>Determined by <sup>1</sup>H-NMR. <sup>d</sup>Determined by GPC at 40 °C in THF using standard polystyrene as a reference. <sup>e</sup>Determined by homonuclear decoupled <sup>1</sup>H NMR spectroscopy of purified polymers.

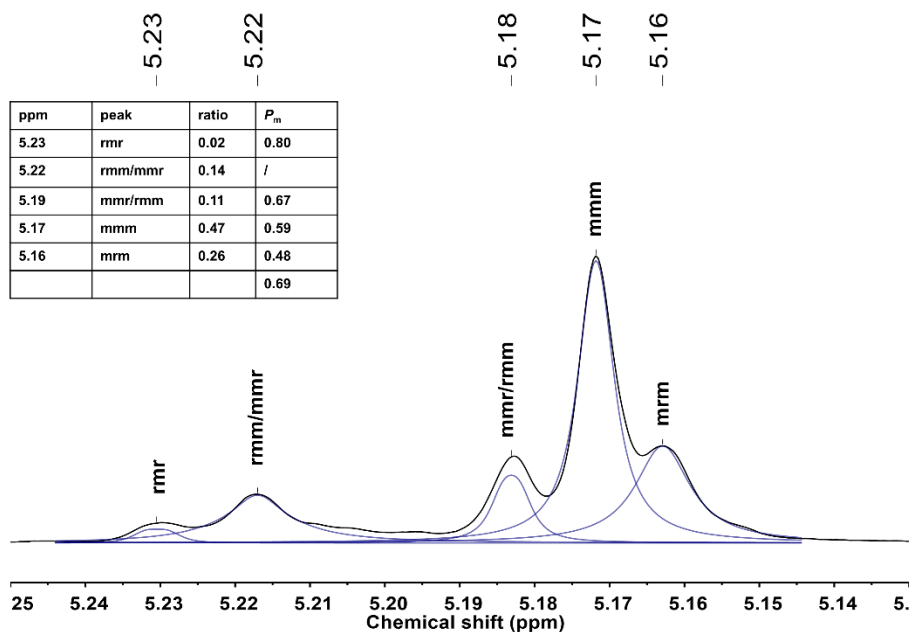
**Figure S1.** Homonuclear decoupled <sup>1</sup>H NMR spectrum of PLA sample (Table 1, run 1).



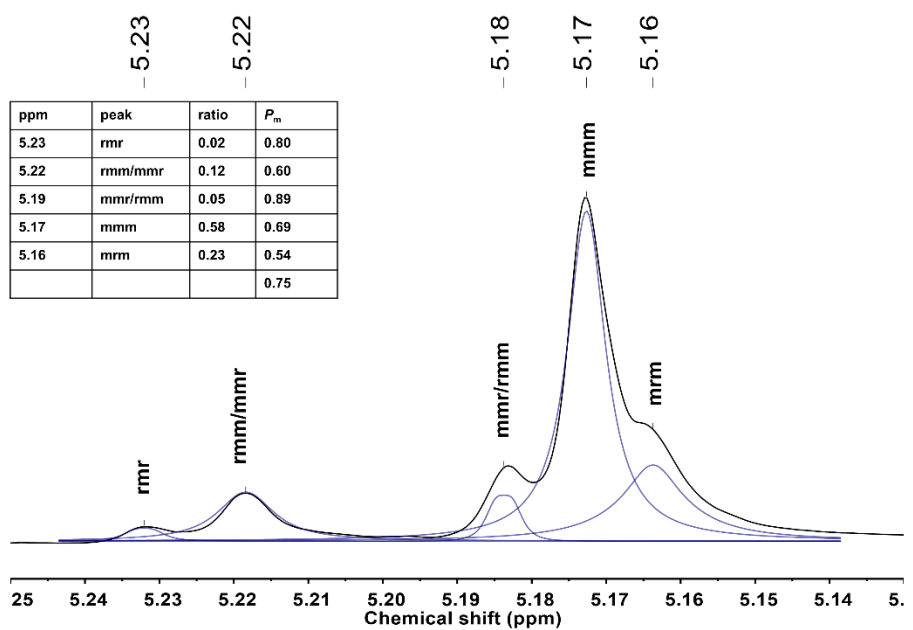
**Figure S2.** Homonuclear decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 1, run 2).



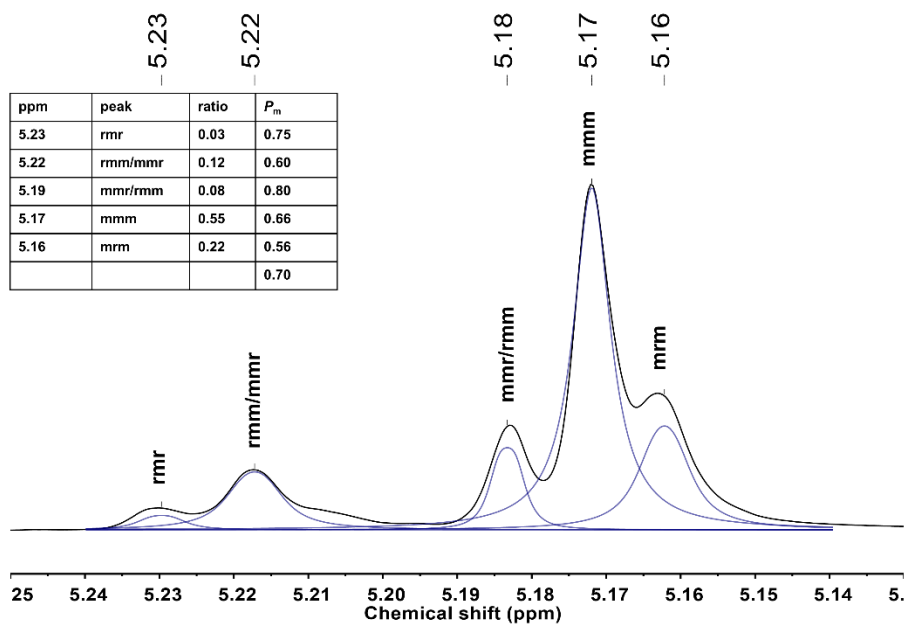
**Figure S3.** Homonuclear decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 1, run 3).



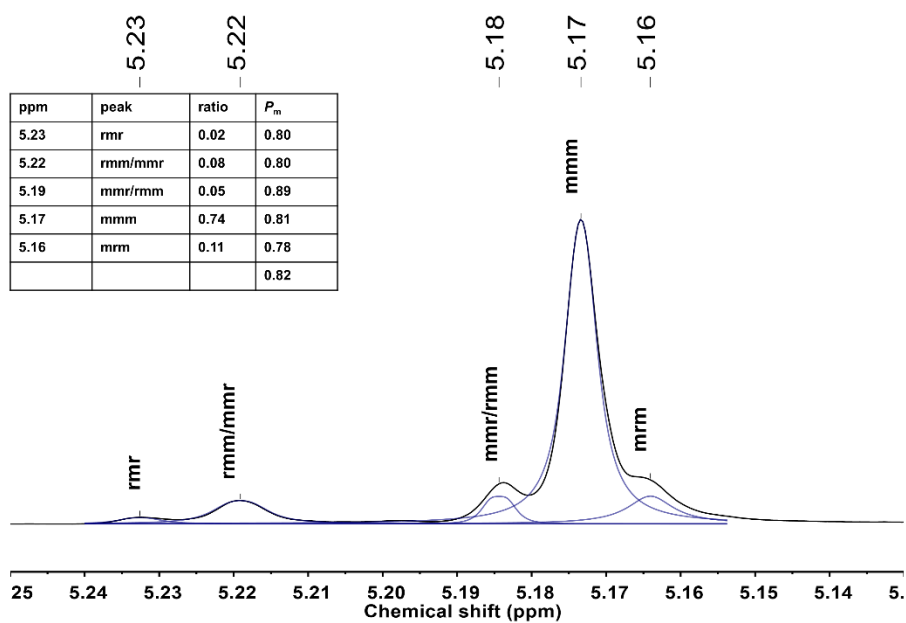
**Figure S4.** Homuncular decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 1, run 4).



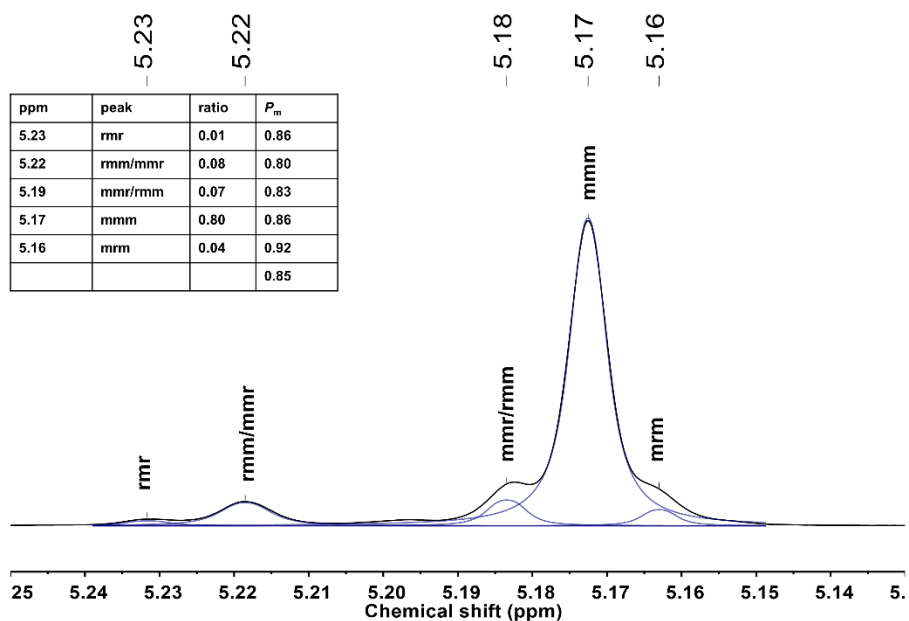
**Figure S5.** Homuncular decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 1, run 5).



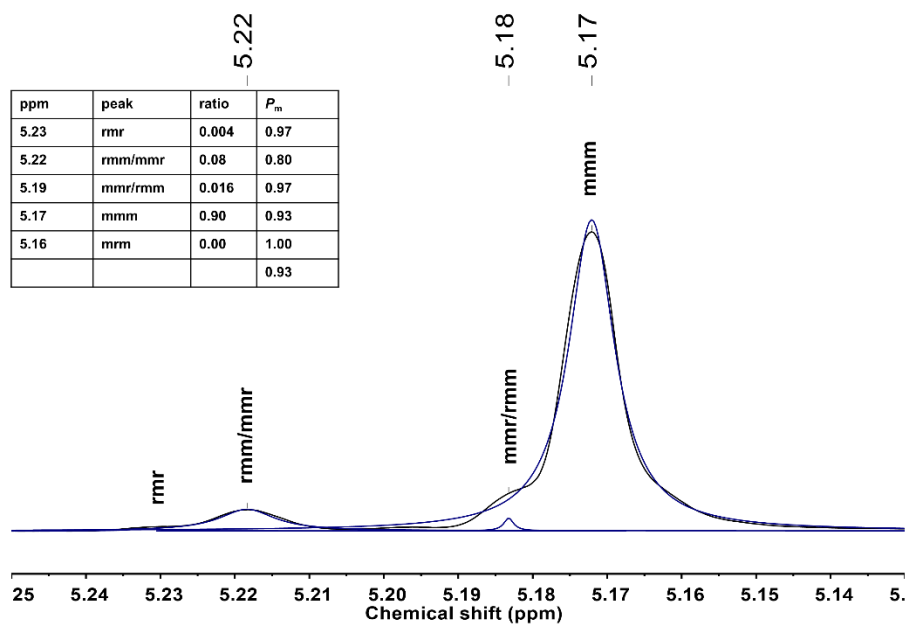
**Figure S6.** Homuncular decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 1, run 7).



**Figure S7.** Homuncular decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 1, run 8).

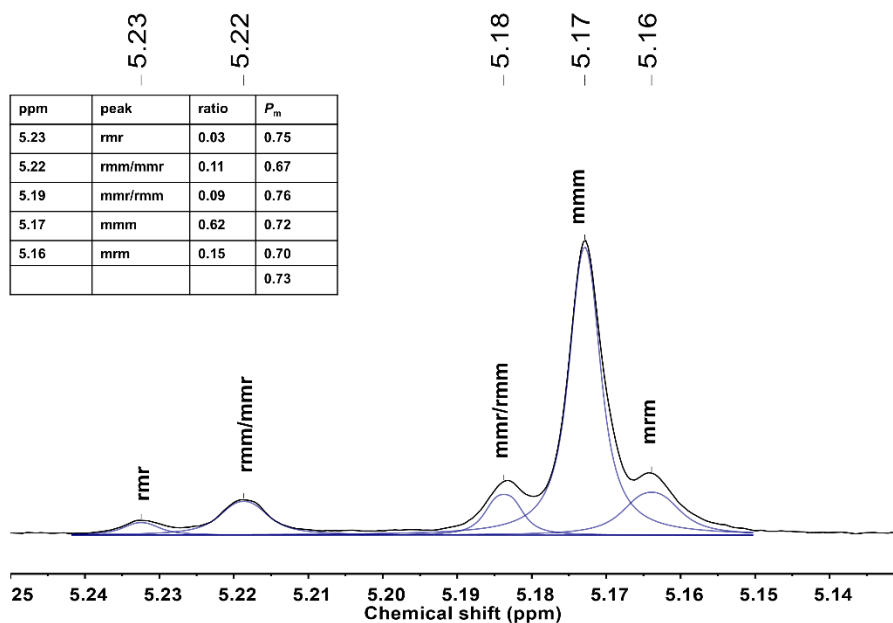


**Figure S8.** Homounuclear decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 1, run 9).

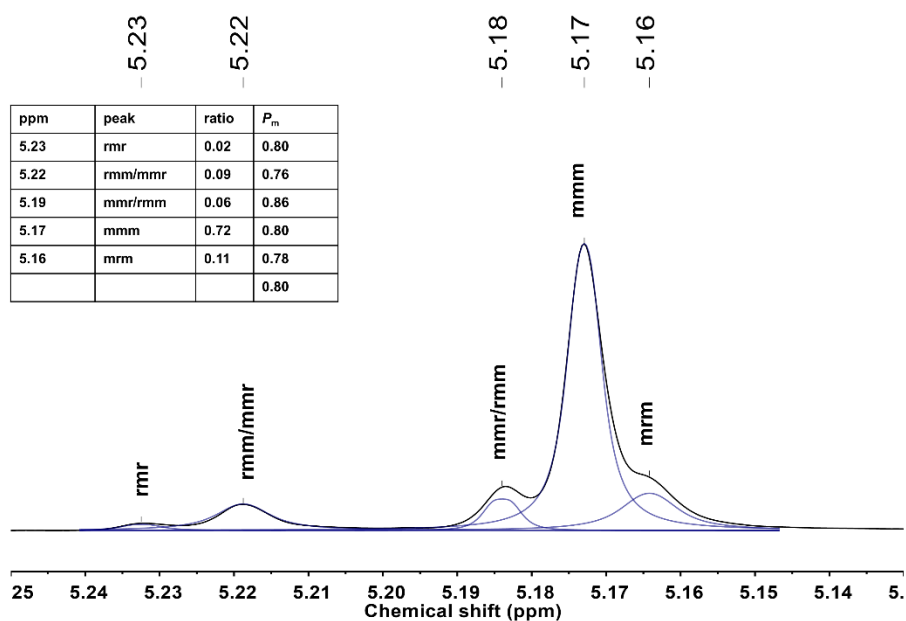


**Figure S9.** Homounuclear decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 1, run 10).

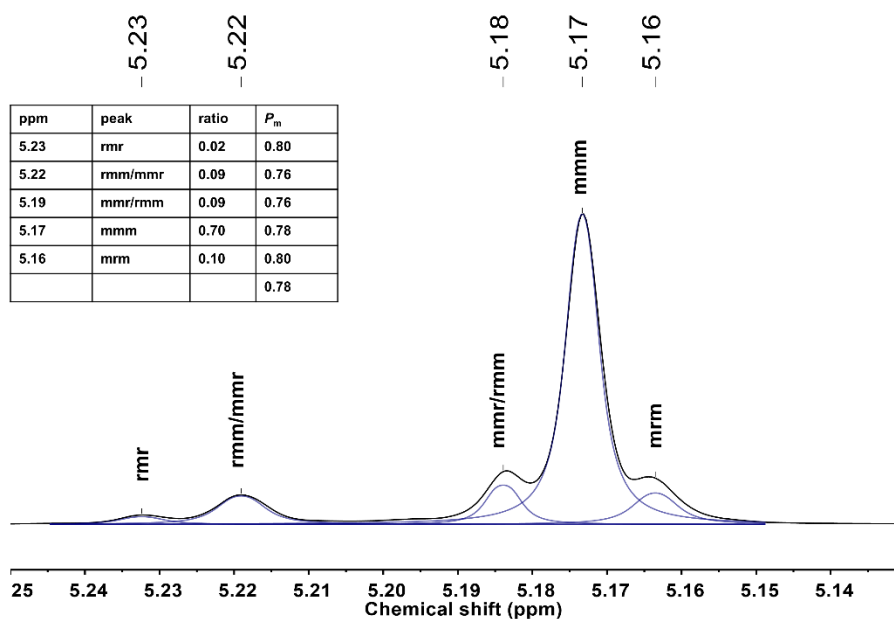




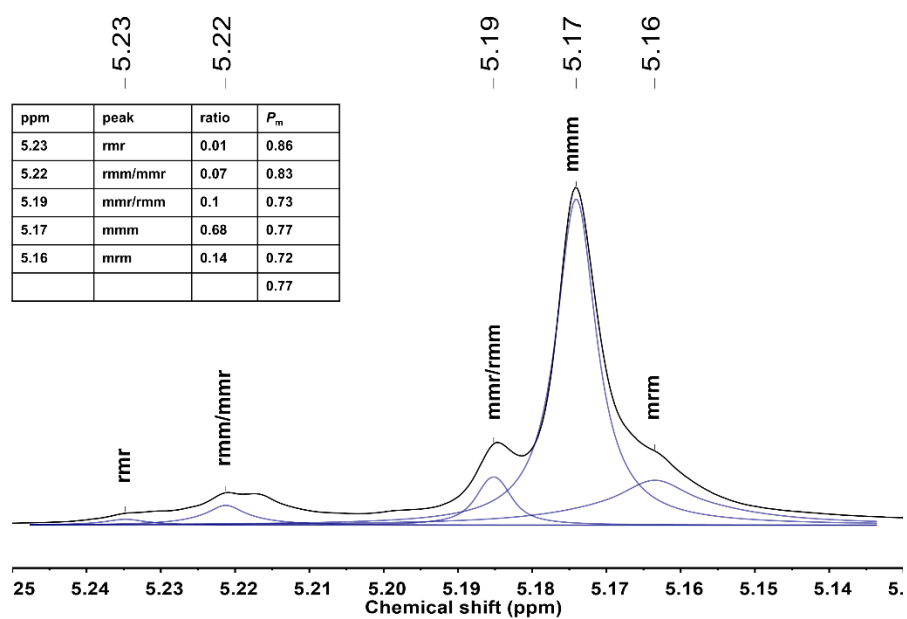
**Figure S10.** Homuncular decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 2, run 1).



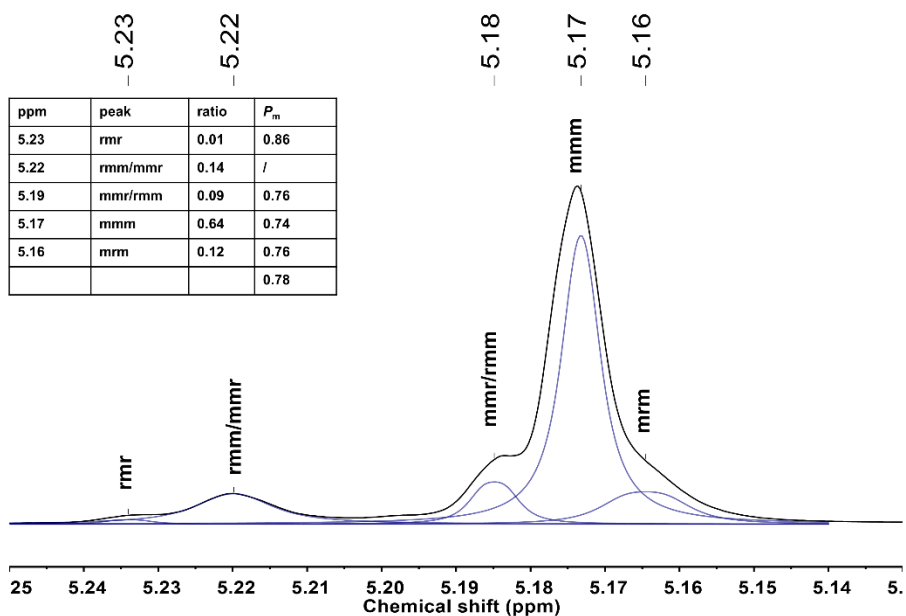
**Figure S11.** Homuncular decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 2, run 2).



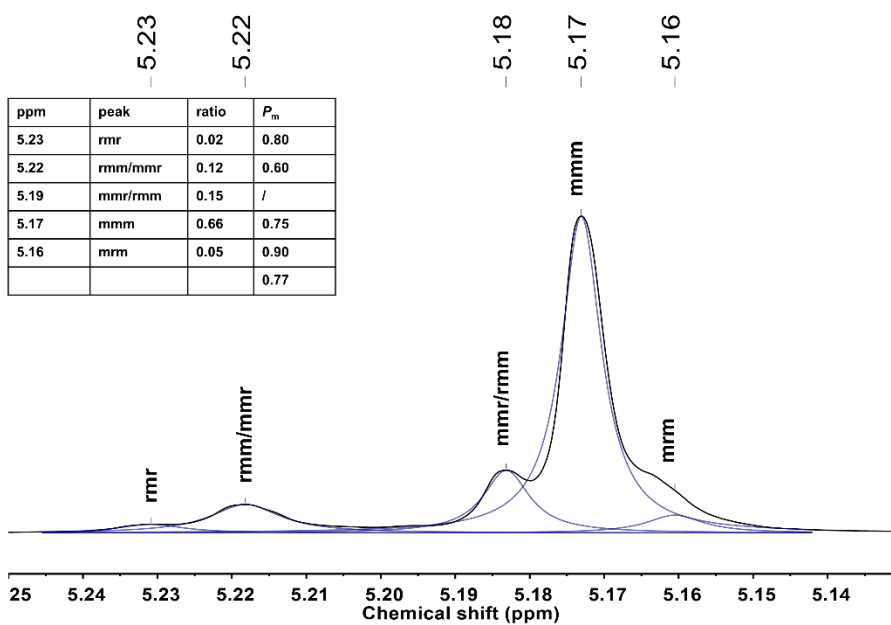
**Figure S12.** Homuncular decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 2, run 3).



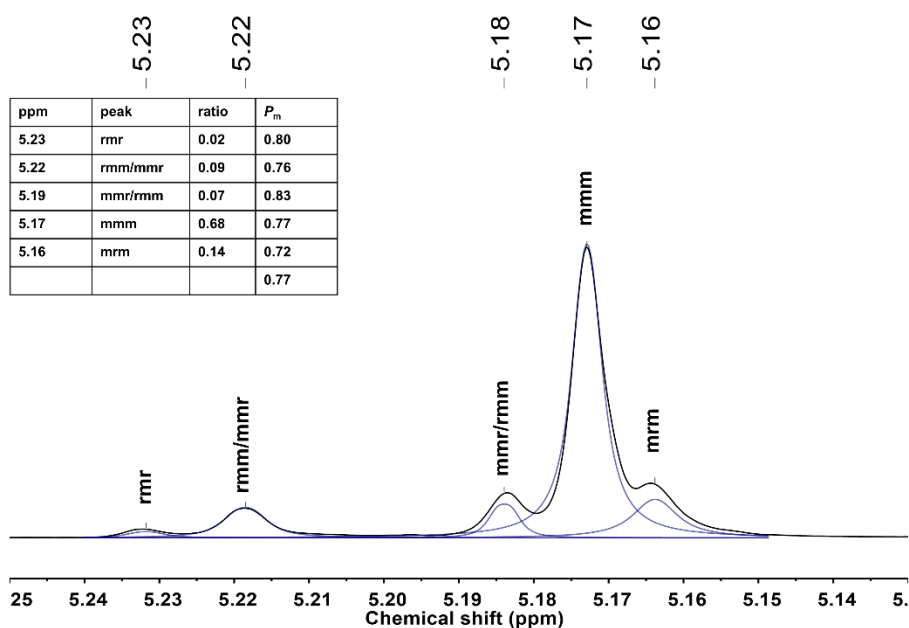
**Figure S13.** Homuncular decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 2, run 4).



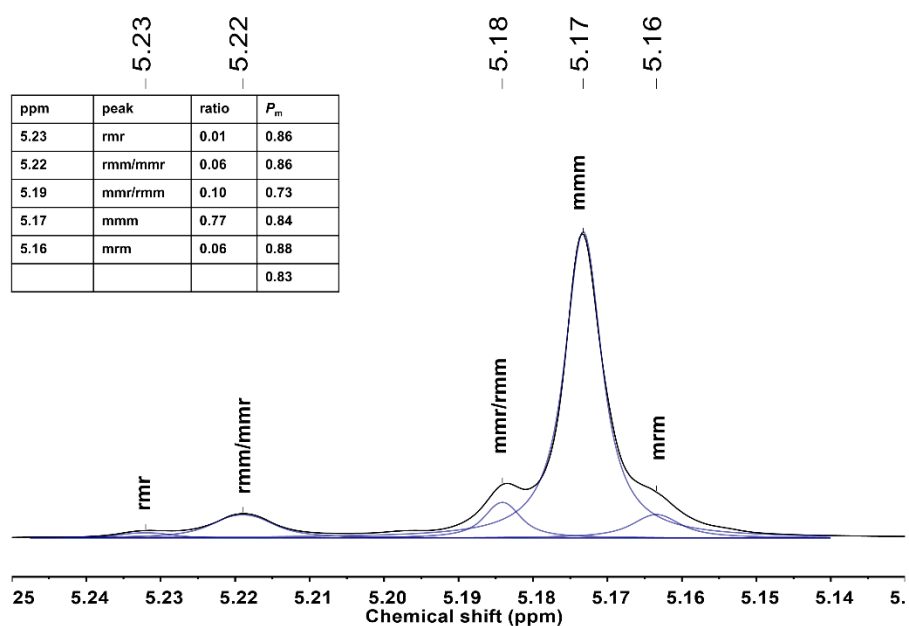
**Figure S14.** Homuncular decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 2, run 5).



**Figure S15.** Homuncular decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 2, run 6).



**Figure S16.** Homonuclear decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 2, run 7).



**Figure S17.** Homonuclear decoupled  $^1\text{H}$  NMR spectrum of PLA sample (Table 2, run 8).

## References

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