Supporting information

Exploiting Chemoselectivity for Discrete Oligomer Synthesis through Sequential IrAAC and CuAAC Reactions

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I. General Information

All air or moisture sensitive reactions were conducted in oven-dried glassware under nitrogen atmosphere using dry solvents. Flash column chromatography was performed over silica gel (200-300 mesh) purchased from Qingdao Puke Co., China. Alkynes and common organic chemicals were purchased from commercial suppliers, such as Sigma-Aldrich® and *J*&K® Scientific Ltd., and used as received. Iridium catalyst was purchased from Strem® Chemicals, Inc.

NMR. ¹H and ¹³C spectra were collected on a Bruker AV 400 MHz NMR spectrometer using residue solvent peaks as an internal standard (¹H NMR: CDCl₃ at 7.26 ppm, DMSO-*d6* at 2.54 ppm; ¹³C NMR: CDCl₃ at 77.0 ppm, DMSO-*d6* at 39.5 ppm).

MS. ESI-MS analyses were measured on Agilent 6540 UHD Accurate-Mass Q-TOF. FTMS were measured on Bruker Solarix XR Fourier transform ion cyclotron resonance mass spectrometer.

LC-MS. LC-MS analysis was measured on Xevo G2 Qtof with Waters Acquity SDS. The instrument was calibrated in the m/z range 50–1800 with an electrospray source (ESI-MS). The conditions of the reversed phase HPLC method are as follows: initial 5.0% acetonitrile - 95.0% water, 3 min 15.0% acetonitrile - 85.0% water, 9 min 50.0% acetonitrile - 50.0% water, 11 min 100.0% acetonitrile - 0.0% water, 13 min 5.0% acetonitrile - 95.0% water. Flow rate: 0.3 ml/min; Column temperature of 45 °C; Injection volume of 5 μ L. The signal was monitored at 214 nm.

SEC. Samples (5 mg) were dissolved in THF (1 mL) and filtered prior to injection. SEC analyses were performed on a Waters 1525 Gel chromatography with three mixed-bed GPC columns in series (three Waters Styragel HT3 THF (7.8*300mm Column)), and THF mobile phase run at 35 °C for 40 min. The differential refractive index of each compound was monitored using a WAT038040 (2414) detector. **Fluorescence.** Fluorescence performance was recorded on Hitachi F-7000 FL Spectrophotometer. The spectra were recorded with EX slit 5.0 nm, EM slit 5.0 nm, PMT voltage 300~800 V and three repetitions.

II. Preparation of Substrates

Diazido monomers A1, 1 A2 2 and A3 3 were prepared according to references.



Scheme S1. Synthetic route to T3.

Synthesis of 4-((4-((*tert*-butyldimethylsilyl)oxy)but-1-yn-1-yl)thio)phenol (T2) (steps i-ii in Scheme S1): (i) At -78 °C, to a solution of alkyne (30 mmol, 1.0 eq.) in dry THF (0.25 M) under N₂ atmosphere was slowly added *n*-BuLi (1.1 eq.). The mixture was stirred at the same temperature for 1 hour before disulfide (1.0 eq.) and EtI (1.0 eq.) were added. Then the mixture was allowed warming to room temperature and stirred for 2 hours before a saturated aqueous NH₄Cl solution was added. The aqueous phase was separated and extracted with ethyl acetate (EA) for three times. The combined organic phase was washed with brine, dried over Na₂SO₄ and evaporated under vacuum to give the crude product of thioalkyne, which was then purified by silica gel flash column chromatography to give pure thioalkyne. (ii) The TBS-protected thioalkyne was dissolved in THF (0.5 M), TBAF (1.0 eq.) was added at 0 °C and stirred another 5 minutes. Upon completion indicated by TLC, EA was added and the mixture was washed by water and brine. The organic layer was dried over MgSO₄, and evaporated under vacuum to give the crude product of thioalkyne, which was then purified by silica gel flash column chromatography to give pure product T2 as yellow oil in 71% overall yield (6.56 g). Rf = 0.5 (PE/EA = 5:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.30-7.26 (m, 2 H), 6.81-6.78 (m, 2 H), 5.89 (s, 1 H), 3.78 (t, *J* = 8.0 Hz, 2 H), 2.61 (t, *J* = 8.0 Hz, 2 H), 0.89 (s, 9 H), 0.08 (s, 6 H).

¹³C NMR (100 MHz, CDCl₃)δ 154.9, 128.8, 123.3, 116.4, 95.0, 67.7, 61.9, 25.9, 25.6, 24.5, 18.3, 5.3.



Synthesis of *tert*-butyldimethyl((4-((4-(prop-2-yn-1-yloxy)phenyl)thio)but-3-yn-1-yl)oxy)silane

(T3) (step iii in Scheme S1): T2 was dissolved in MeCN (0.5 M), K_2CO_3 (2.0 eq.) was added and stirred for 30 minutes, then 3-bromopropyne (1.2 eq.) was added at 0 °C and stirred over night at 60 °C. Upon completion indicated by TLC, the reaction solution was filtered under reduced pressure and purified by silica gel flash column chromatography to give pure product T3 as colorless oil in 83% yield (6.12 g).

Rf = 0.5 (PE/EA = 20:1).

¹**H** NMR (400 MHz, CDCl₃) δ 7.38-7.35 (m, 2 H), 6.97-6.94 (m, 2 H), 4.68 (d, *J* = 4.0 Hz, 2 H), 3.78 (t, *J* = 8.0 Hz, 2 H), 2.63 (t, *J* = 8.0 Hz, 2 H), 2.52 (t, *J* = 4.0 Hz, 1 H), 0.91 (s, 9 H), 0.08 (s, 6 H).

¹³C NMR (100 MHz, CDCl₃) δ 156.5, 128.2, 124.8, 115.9, 95.6, 78.2, 75.7, 67.2, 61.7, 56.0, 25.9, 24.6, 18.3, 5.3.

HRMS m/z (ESI) calcd. for C₁₉H₂₇O₂SSi (M+H)⁺ 347.1496, found 347.1490.

III. Azide Reactivity Study



Scheme S2. IrAAC reaction of bis(azides) with thioalkyne T1.

General procedure:

In a glove box, to an oven-dried vial was added the building unit involving bis(azides) (1.2 eq. or 2.0 eq.), **T1** (0.2 mmol, 1.0 eq.), [Ir(COD)Cl]₂ (2 mol %) and DCE (0.3 M). The vial was capped and removed from the glove box. The reaction mixture was stirred at room temperature for 3 h until the reaction completed (confirmed by TLC), and then concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography to give the mixed product.



Scheme S3. Selective reaction of T1 and A1.

The ratio of **T-A**, **A-T'** and **T-A-T'** is determined by ¹H NMR analysis. At the completion of the reaction, the reaction solution was concentrated by vacuum and added the internal standard. The ratio of **T1-A1** and **T1-A1-T1'** was determined to be 92:8, by comparing the relative valves of the peak observed at 5.45 ppm (s, 2 H) for **T1-A1** and 5.49 ppm (s, 2 H) for **T1-A1-T1'**. When the **A1** was 2.0 equivalents, the ratio of **T1-A1** and **T1-A1-T1'** was up to 99:1.



Figure S1. Comparison of ¹H NMR for reaction mixture, T1-A1 and T1-A1-T1'.



Scheme S4. Selective reaction of T1 and A2.

The ratio of **T1-A2** and **T1-A2-T1**' was determined to be 93:7, by comparing the relative valves of the peak observed at 5.45 ppm (s, 2 H) for **T1-A2** and 5.44 ppm (s, 2 H) for **T1-A2-T1**'. When the **A2** was 2.0 equivalents, the ratio of **T1-A2** and **T1-A2-T1**' was up to 99:1.

Reaction mixture and internal standard



Figure S2. Comparison of ¹H NMR for reaction mixture, T1-A2 and T1-A2-T1'.



Scheme S5. Selective reaction of T1 and A3.



1-(4-azidobenzyl)-4-(2-((tert-butyldimethylsilyl)oxy)ethyl)-5-(phenylthio)-1H-1,2,3-triazole (T1-

A1) was obtained as brown oil in 85% yield (79.2 mg).

Rf = 0.5 (PE/EA = 3:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.16-7.13 (m, 5 H), 6.8-6.80 (m, 4 H), 5.45 (s, 2 H), 3.90 (t, *J* = 8.0 Hz,

2 H), 2.9 (t, *J* = 8.0 Hz, 2 H), 0.81 (s, 9 H), 0.05 (s, 6 H).

¹³**C NMR** (100 MHz, CDCl₃) δ 150.8, 140.0, 133.8, 131.1, 129.6, 129.3, 126.7, 126.5, 124.3, 119.1, 62.0, 51.7, 29.1, 25.8, 18.2, 5.4.

HRMS m/z (ESI) calcd. for C₂₃H₃₁N₆OSSi (M+H)⁺ 467.2044, found 467.2046.



1-(3-azidobenzyl)-4-(2-((*tert*-butyldimethylsilyl)oxy)ethyl)-5-(phenylthio)-1*H*-1,2,3-triazole (T1-A2) was obtained as brown oil in 86% yield (80.2 mg).

Rf = 0.5 (PE/EA = 3:1).

¹H NMR (400 MHz, CDCl₃) δ 7.18-7.12 (m, 4 H), 6.96 (d, *J* = 8.0 Hz, 1 H), 6.85-6.81 (m, 3 H), 6.76 (s, 1 H), 5.45 (s, 2 H), 3.91 (t, *J* = 8.0 Hz, 2 H), 2.96 (t, *J* = 8.0 Hz, 2 H), 0.81 (s, 9 H), 0.05 (s, 6 H).
¹³C NMR (100 MHz, CDCl₃) δ 150.8, 140.3, 136.3, 133.7, 130.0, 129.2, 126.6, 126.5, 124.4, 118.7, 62.0, 51.8, 29.1, 25.8, 18.2, 5.4.

HRMS m/z (ESI) calcd. for C₂₃H₃₁N₆OSSi (M+H)⁺ 467.2044, found 467.2049.



1-(2-azidobenzyl)-4-(2-((*tert***-butyldimethylsilyl)oxy)ethyl)-5-(phenylthio)-1***H***-1,2,3-triazole (1) was obtained as brown oil in 94% yield (87.6 mg).**

Rf = 0.5 (PE/EA = 3:1).

¹H NMR (400 MHz, CDCl₃) δ 7.25-7.21 (m, 1 H), 7.15-7.08 (m, 3 H), 7.00-6.91 (m, 3 H), 6.88-6.86 (m, 2 H), 5.48 (s, 2 H), 3.93 (t, *J* = 8.0 Hz, 2 H), 2.98 (t, *J* = 8.0 Hz, 2 H), 0.83 (s, 9 H), 0.02 (s, 6 H).
¹³C NMR (100 MHz, CDCl₃) δ 150.5, 137.5, 133.7, 129.4, 129.4, 129.1, 126.9, 126.5, 125.8, 124.8, 118.0, 62.1, 47.1, 29.2, 25.9, 18.3, 5.4.

HRMS m/z (ESI) calc. for C₂₃H₃₁N₆OSSi (M+H)⁺ 467.2044, found 467.2046.



Scheme S6. Different reactivity of azido groups in IrAAC and CuAAC.



In a glove box, to an oven-dried vial was added **T1-A3**(0.2 mmol, 1.0 eq.), **T1** (1.2 eq.), [Ir(COD)Cl]₂ (2 mol %) and DCE (0.3 M). The vial was capped and removed from the glove box. The reaction mixture was stirred at room temperature for 24 h, then concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography to give the pure product **T1-A3-T1**' as brown oil in 33% yield (50.1 mg).

Rf = 0.5 (PE/EA = 2:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.21-7.09 (m, 8 H), 6.94-6.87 (m, 5 H), 6.79-6.76 (m, 1 H), 5.22 (s, 2 H), 4.01 (t, *J* = 8.0 Hz, 2 H), 3.92 (t, *J* = 8.0 Hz, 2 H), 3.06 (t, *J* = 8.0 Hz, 2 H), 2.97 (t, *J* = 8.0 Hz, 2 H), 0.86 (d, *J* = 12.0 Hz, 18 H), 0.03 (d, *J* = 12.0 Hz, 12 H).

¹³C NMR (100 MHz, CDCl₃) δ 150.2, 149.6, 133.7, 133.2, 132.2, 130.3, 129.3, 129.2, 128.8, 128.3, 128.2, 127.9, 127.5, 126.9, 125.7, 62.0, 47.5, 29.1, 25.9, 18.3, 5.4, 5.3.



T1-A3 (0.2 mmol, 1.0 eq.) and phenylacetylene (1.2 eq.) were added into the solution of DMF (0.5 M) involving CuBr (0.2 eq.), PMDETA (0.2 eq.), NaAsc (0.5 eq.). The reaction mixture was stirred for 10 min until the reaction completed (confirmed by TLC). Then diluted with ethyl acetate, washed with

brine (three times), dried over Na₂SO₄, filtered, and evaporated under vacuum. The residue was purified by column chromatography to give **T1-A3-T**" as white solid in 91% yield (103.4 mg). Rf = 0.5 (PE/EA = 2:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.94 (s, 1 H), 7.90-7.87 (m, 2 H), 7.50-7.32 (m, 6 H), 7.18 (d, *J* = 4.0 Hz, 1 H), 7.09-7.07 (m, 3 H), 6.78-6.76 (m, 2 H), 5.55 (s, 2 H), 3.88 (t, *J* = 8.0 Hz, 2 H), 2.93 (t, *J* = 8.0 Hz, 2 H), 0.81 (s, 9 H), 0.05 (s, 6 H).

¹³**C NMR** (100 MHz, CDCl₃) δ 150.2, 149.6, 133.7, 133.2, 132.2, 130.3, 129.3, 129.2, 128.8, 128.3, 128.2, 127.9, 127.5, 126.9, 125.7, 62.0, 47.5, 29.1, 25.9, 18.3, 5.4, 5.3.

IV. Construction of Oligomers via Sequential IrAAC and CuAAC reactions

General procedures:

IrAAC: In a glove box, to an oven-dried vial was added the thioalkyne building unit (first step was **T1**) (1.0 eq.), the building unit **A3** (1.2 eq.), $[Ir(COD)Cl]_2$ (2 mol %) and DCE (0.3 M). The vial was capped and removed from the glove box. The reaction mixture was stirred at room temperature for 2-6 h until the reaction completed (confirmed by TLC), and then concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography to give the desired product.

CuAAC: The purified unprotected azide building unit (1.0 eq.) and the thioalkyne building unit **T3** (1.1 eq.) were added into the solution of DMF (0.5 M) involving CuBr (0.2 eq.), PMDETA (0.2 eq.), NaAsc (0.5 eq.). The mixture was stirred at room temperature. The solution was then cooled to room temperature, diluted with ethyl acetate, washed with brine (three times), dried over Na₂SO₄, filtered, and evaporated under vacuum. The residue was purified by column chromatography on silica gel to afford the thioalkyne building unit for next cycle.



2 was prepared as yellow oil from **1** (4.4 mmol, 2.06 g, 1.0 eq.) and **T3** (4.8 mmol, 1.68 g, 1.1 eq.) in 90% yield (4.12 g).

Rf = 0.5 (PE/EA = 5:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.78 (s, 1 H), 7.39-7.33 (m, 4 H), 7.25 (d, *J* = 8.0 Hz, 1 H), 7.10 (m, 4 H), 7.02 (d, *J* = 8.0 Hz, 2 H), 6.76-6.73 (m, 2 H), 5.51 (s, 2 H), 5.25 (s, 2 H), 3.89 (t, *J* = 8.0 Hz, 2 H), 3.78 (t, *J* = 8.0 Hz, 2 H), 2.94 (t, *J* = 8.0 Hz, 2 H), 2.64 (t, *J* = 8.0 Hz, 2 H), 0.89 (s, 9 H), 0.81 (s, 9 H), 0.07 (s, 6 H), 0.04 (s, 6 H).

¹³C NMR (100 MHz, CDCl₃) δ 157.2, 150.5, 144.2, 135.1, 133.3, 130.5, 130.2, 129.6, 129.3, 129.1, 128.3, 126.9, 126.7, 125.5, 124.9, 124.5, 115.7, 62.0, 61.7, 47.3, 29.1, 25.8, 24.6, 18.2, 5.4.
HRMS *m/z* (ESI) calc. for C₄₂H₅₇N₆O₃S₂Si₂ (M+H)⁺ 813.3467, found 813.3459.



3 was prepared as yellow-brown oil from **2** (4.0 mmol, 3.26 g, 1.0 eq.) and **A3** (4.8 mmol, 0.85 g, 1.2 eq.) in 87% yield (3.43 g).

Rf = 0.4 (PE/EA = 3:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.82 (s, 1 H), 7.43-7.35 (m, 2 H), 7.30-7.26 (m, 2 H), 7.13 (t, *J* = 4.0 Hz, 4 H), 7.08 (d, *J* = 8.0 Hz, 1 H), 7.00-6.95 (m, 3 H), 6.85-6.78 (m, 5 H), 5.53 (d, *J* = 12.0 Hz, 4 H), 5.20 (s, 2 H), 3.97 (t, *J* = 8.0 Hz, 2 H), 3.92 (t, *J* = 8.0 Hz, 2 H), 3.03 (t, *J* = 8.0 Hz, 2 H), 2.97 (t, *J* = 8.0 Hz, 2 H), 0.87 (s, 9 H), 0.83 (s, 9 H), 0.02 (s, 6 H), 0.01 (s, 6 H).

¹³**C NMR** (100 MHz, CDCl₃) δ 157.6, 150.5, 149.8, 143.9, 137.3, 135.1, 133.3, 130.6, 130.3, 129.7, 129.3, 129.2, 129.1, 127.0, 126.8, 126.2, 126.0, 125.5, 124.9, 124.9, 124.8, 124.6, 118.0, 115.6, 62.1, 61.9, 47.3, 47.1, 29.2, 25.8, 18.3, 5.4.

HRMS m/z (ESI) calc. for C₄₉H₆₃N₁₂O₃S₂Si₂ (M+H)⁺ 987.4121, found 987.4124.



4 was prepared as light brown oil from 3 (3.1 mmol, 3.06 g, 1.0 eq.) and T3 (3.4 mmol, 1.18 g, 1.1 eq.) in 90% yield (3.72 g).

Rf = 0.2 (PE/EA = 2:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.90 (s, 1 H), 7.80 (s, 1 H), 7.41-7.24 (m, 8 H), 7.11-7.08 (m, 4 H), 7.02-6.96 (m, 3 H), 6.90 (d, *J* = 8.0 Hz, 2 H), 6.81-6.76 (m, 4 H), 5.53 (d, *J* = 8.0 Hz, 4 H), 5.28 (s, 2 H), 5.15 (s, 2 H), 3.95-3.87 (m, 4 H), 3.77 (t, *J* = 8.0 Hz, 2 H), 3.00-2.92 (m, 4 H), 2.62 (t, *J* = 8.0 Hz, 2 H), 0.89 (s, 9 H), 0.83 (d, *J* = 4.0 Hz, 18 H), 0.07 (s, 6 H), 0.01 (d, *J* = 16.0 Hz, 12 H).

¹³C NMR (100 MHz, CDCl₃) δ 157.8, 157.2, 150.5, 149.7, 144.2, 143.8, 135.1, 135.0, 133.3, 130.7, 130.6, 130.3, 130.2, 129.7, 129.4, 129.3, 129.2, 129.1, 128.3, 127.0, 126.8, 126.6, 125.5, 125.0, 124.7, 124.6, 124.3, 115.8, 62.0, 61.9, 61.7, 47.3, 29.1, 25.8, 24.6, 18.3, 18.3, 5.4, 5.3.

HRMS m/z (ESI) calc. for C₆₈H₈₉N₁₂O₅S₃Si₃ (M+H)⁺ 1333.5543, found 1333.5546.



5 was prepared as white solid from **4** (2.6 mmol, 3.47 g, 1.0 eq.) and **A3** (3.2 mmol, 0.55 g, 1.2 eq.) in 87% yield (3.41 g).

Rf = 0.1 (DCM/MeOH = 100:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.94 (s, 1 H), 7.83 (s, 1 H), 7.41-7.26 (m, 7 H), 7.12-7.04 (m, 5 H), 6.98-6.92 (m, 6 H), 6.84-6.80 (m, 7 H), 5.52 (t, *J* = 12.0 Hz, 6 H), 5.22 (d, *J* = 16.0 Hz, 4 H), 3.95-3.89 (m, 6 H), 3.01-2.94 (m, 6 H), 0.85-0.82 (m, 27 H), 0.03-0.00 (m, 18 H).

¹³**C NMR** (100 MHz, CDCl₃) δ 157.8, 157.6, 150.4, 149.7, 149.6, 143.9, 143.7, 137.2, 135.1, 135.0, 133.3, 130.6, 130.5, 130.2, 129.7, 129.3, 129.2, 129.0, 127.0, 126.8, 126.6, 126.2, 126.0, 125.5, 124.9, 124.8, 124.6, 124.2, 117.9, 115.8, 115.6, 62.1, 62.0, 61.9, 47.3, 47.3, 47.0, 29.1, 29.1, 25.8, 18.2, 5.5, 5.4.

HRMS m/z (ESI) calcd. for C₇₅H₉₅N₁₈O₅S₃Si₃ (M+H)⁺ 1507.6197, found 1507.6188.



6 was prepared as white solid from **5** (2.2 mmol, 3.31 g, 1.0 eq.) and **T3** (2.42 mmol, 0.84 g, 1.1 eq.) in 86% yield (3.51 g).

Rf = 0.2 (DCM/MeOH = 80:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.94-7.92 (d, *J* = 8.0 Hz, 2 H), 7.83 (s, 1 H), 7.40-7.26 (m, 11 H), 7.24-7.09 (m, 4 H), 7.03-6.88 (m, 8 H), 6.83-6.77 (m, 6 H), 5.55 (t, *J* = 4.0 Hz, 6 H), 5.28 (s, 2 H), 5.20 (d, *J* = 12.0 Hz, 4 H), 3.96-3.89 (m, 6 H), 3.78 (t, *J* = 8.0 Hz, 2 H), 3.01-2.93 (m, 6 H), 2.62 (t, *J* = 8.0 Hz, 2 H), 0.90 (s, 9 H), 0.84 (d, *J* = 8.0 Hz, 27 H), 0.08 (s, 6 H), 0.00 (d, *J* = 16.0 Hz, 18H).

¹³C NMR (100 MHz, CDCl₃) δ 157.8, 157.1, 150.4, 149.6, 144.1, 143.7, 143.7, 135.1, 134.9, 133.2, 130.6, 130.6, 130.4, 130.2, 129.6, 129.3, 129.2, 129.0, 128.2, 127.0, 126.8, 126.5, 125.5, 124.9, 124.6, 124.6, 124.5, 124.2, 115.8, 115.7, 62.0, 61.9, 61.6, 47.3, 47.3, 29.1, 25.8, 24.6, 18.2, 5.5, 5.4.
HRMS *m/z* (ESI) calcd. for C₉₄H₁₂₀N₁₈NaO₇S₄Si₄ (M+Na)⁺ 1875.7444, found 1875.7716.



7 was prepared as white solid from **6** (1.8 mmol, 3.33 g, 1.0 eq.) and **A3** (2.2 mmol, 0.38 g, 1.2 eq.) in 96% yield (3.43 g).

Rf = 0.3 (DCM/MeOH = 70:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.93 (d, *J* = 4.0 Hz, 2 H), 7.82 (s, 1 H), 7.41-7.23 (m, 10 H), 7.11-7.08 (m, 4 H), 7.04 (d, *J* = 8.0 Hz, 1 H), 6.97-6.90 (m, 9 H), 6.83-6.76 (m, 9 H), 5.53-5.50 (m, 8 H), 5.20-5.16 (m, 6 H), 3.96-3.87 (m, 8 H), 3.01-2.92 (m, 8 H), 0.83-0.80 (m, 36 H), 0.05-0.01 (m, 24 H).

¹³C NMR (100 MHz, CDCl₃) δ 157.8, 157.6, 150.4, 149.7, 143.7, 135.0, 133.3, 129.3, 129.1, 127.0, 126.8, 126.6, 126.3, 125.6, 125.0, 124.9, 124.7, 124.3, 118.0, 115.8, 115.6, 62.1, 62.0, 61.9, 47.4, 47.1, 29.1, 25.9, 18.3, 5.4.

HRMS m/z (ESI) calcd. for C₁₀₁H₁₂₇N₂₄O₇S₄Si₄ (M+H)⁺ 2027.8274, found 2027.8277.



8 was prepared as white solid from **7** (1.6 mmol, 3.23 g, 1.0 eq.) and **T3** (1.76 mmol, 0.61 g, 1.1 eq.) in 88% yield (3.34 g).

Rf = 0.5 (DCM/MeOH = 50:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.93-7.90 (m, 3 H), 7.81 (s, 1 H), 7.35-7.26 (m, 13 H), 7.21 (d, *J* = 8.0 Hz, 1 H) 7.08-7.06 (m, 4 H), 7.00 (d, *J* = 8.0 Hz, 2 H), 6.93-6.89 (m, 9 H), 6.85-6.75 (m, 8 H), 5.52-5.49 (m, 8 H), 5.25 (s, 2 H), 5.17 (d, *J* = 12 Hz, 6 H), 3.93-3.86 (m, 8 H), 3.75 (t, *J* = 8.0 Hz, 2 H), 2.98-2.90 (m, 8 H), 2.59 (t, *J* = 8.0 Hz, 2 H), 0.87 (s, 9 H), 0.87-0.79 (m, 36 H), 0.79 (s, 6 H), 0.07-0.01 (m, 24 H).

¹³C NMR (100 MHz, CDCl₃) δ 157.8, 157.1, 150.4, 149.6, 144.1, 143.7, 135.0, 134.9, 133.2, 130.6, 130.6, 130.4, 130.2, 129.6, 129.4, 129.2, 129.1, 129.0, 128.2, 127.0, 126.8, 126.6, 125.5, 124.9, 124.7, 124.5, 124.1, 115.8, 115.7, 61.9, 61.8, 61.6, 47.3, 47.2, 29.0, 25.8, 25.8, 24.5, 18.2, 5.4.
HRMS *m/z* (ESI) calcd. for C₁₂₀H₁₅₃N₂₄O₉S₅Si₅ (M+H)⁺ 2373.9697, found 2373.9707.



9 was prepared as white solid from **8** (1.4 mmol, 3.31 g, 1.0 eq.) and **A3** (1.68 mmol, 0.30 g, 1.2 eq.) in 90% yield (3.42 g).

Rf = 0.5 (DCM/MeOH = 50:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.93 (d, *J* = 4.0 Hz, 3 H), 7.81 (s, 1 H), 7.40-7.23 (m, 13 H), 7.10-7.07 (m, 4 H), 7.03 (d, *J* = 8.0 Hz, 1 H), 6.96-6.89 (m, 12 H), 6.82-6.76 (m, 11 H), 5.52-5.46 (m, 10 H), 5.19-5.15 (m, 8 H), 3.95-3.86 (m, 10 H), 3.01-2.91 (m, 10 H), 0.83-0.80 (m, 45 H), 0.06-0.02 (m, 30 H).

¹³C NMR (100 MHz, CDCl₃) δ 157.8, 149.6, 143.7, 135.0, 130.7, 130.2, 129.7, 129.4, 129.3, 129.1, 127.0, 126.8, 126.6, 126.0, 125.5, 124.7, 124.2, 117.9, 115.8, 115.6, 62.1, 62.0, 61.9, 47.4, 47.1, 29.6, 29.1, 25.9, 25.8, 18.2, 5.4.

HRMS m/z (ESI) calcd. for C₁₂₇H₁₅₉N₃₀O₉S₅Si₅ (M+H)⁺ 2548.0351, found 2548.0357.



10 was prepared as white solid from **9** (1.1 mmol, 2.80 g, 1.0 eq.) and **T3** (1.4 mmol, 0.49 g, 1.1 eq.) in 93% yield (2.89 g).

Rf = 0.4 (DCM/MeOH = 50:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.93-7.92 (m, 4 H), 7.90 (s, 1 H), 7.8-7.25 (m, 18 H), 7.10-7.09 (m, 4 H), 7.01 (s, 1 H), 6.99 (s, 1 H), 6.93-6.86 (m, 12 H), 6.82-6.76 (m, 10 H), 5.52-5.49 (m, 10 H), 5.26 (s, 2 H), 5.18-5.15 (m, 8 H), 3.93-3.86 (m, 10 H), 3.76 (t, *J* = 8.0 Hz, 2 H), 2.99-2.93 (m, 10 H), 2.60 (t, *J* = 8.0 Hz, 2 H), 0.88 (s, 9 H), 0.82-0.07 (d, *J* = 8.0 Hz, 45 H), 0.06 (s, 6 H), 0.04 (d, *J* = 12.0 Hz, 30 H).

¹³C NMR (100 MHz, CDCl₃) δ 157.8, 149.6, 143.7, 143.7, 135.0, 130.7, 130.6, 130.2, 129.4, 129.3, 129.2, 129.1, 128.3, 127.0, 126.8, 126.6, 125.5, 124.7, 124.2, 115.8, 62.0, 61.9, 47.4, 29.1, 25.9, 25.8, 18.2, 5.4.

HRMS *m/z* (ESI) calcd. for C₁₄₆H₁₈₄N₃₀NaO₁₁S₆Si₆ (M+Na)⁺ 2916.1593, found 2916.5479.



Scheme S7. Synthesis of 11.

Synthesis of 11: T4 (0.02 mmol, 4.8 mg, 1.0 eq.) and 9 (0.048 mmol, 122.4 mg, 2.2 eq.) were added into the solution of DMF (0.5 M) involving CuBr (0.4 eq.), PMDETA (0.4 eq.), NaAsc (1.0 eq.). The mixture was stirred for 25 min at room temperature. The solution was then cooled to room temperature, diluted with ethyl acetate, washed with brine (three times), dried over Na_2SO_4 , filtered, and evaporated under vacuum. The residue was purified by column chromatography on silica gel to get 11 as white solid in 81% yield (86.5 mg).

Rf = 0.2 (DCM/MeOH = 50:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.96-7.83 (d, 10 H), 7.39-7.23 (m, 35 H), 7.11-7.07 (m, 8 H), 6.91-6.89 (m, 22 H), 6.82-6.76 (m, 20 H), 5.53 (d, *J* = 12.0 Hz, 20 H), 5.31-5.16 (m, 20 H), 3.92 (s, 20 H), 3.87 (s, 3 H), 2.97 (s, 20 H), 0.82-0.80 (m, 90 H), 0.05-0.02 (m, 60 H).

¹³**C NMR** (100 MHz, CDCl₃) δ 157.9, 135.1, 135.1, 133.3, 130.8, 130.6, 130.3, 129.3, 129.1, 127.1, 126.8, 125.6, 124.1, 115.8, 61.9, 47.5, 29.1, 25.9, 18.3, 5.4.

FTMS m/z (ESI) calcd. for C₂₆₈H₃₂₈N₆₀O₂₂S₁₀Si₁₀ (M) 5338.13, found 1336.60 (M+4H)⁴⁺, 1782.18 (M+3H)³⁺.



Scheme S8. Synthesis of 12.

Synthesis of 12: In a glove box, to an oven-dried vial was added **A4** (0.02 mmol, 3.7 mg, 1.0 eq.), **10** (0.048 mmol, 139.1 mg, 2.2 eq.), [Ir(COD)Cl]₂ (4 mol %) and DCE (0.1 M). The vial was capped and removed from the glove box. The reaction mixture was stirred at room temperature for 2 h until the reaction completed (confirmed by TLC), and then concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography to give the desired product **12** as white solid in 70% yield (83.0 mg).

Rf = 0.3 (DCM/MeOH = 50:1).

¹**H NMR** (400 MHz, CDCl₃) δ 8.00 (s, 2 H), 7.94 (s, 6 H), 7.81 (s, 2 H), 7.38-7.35 (m, 10 H), 7.33-7.30 (m, 17 H), 7.28-7.23 (m, 3 H), 7.10-7.07 (m, 8 H), 7.01 (s, 4 H), 6.93-6.76 (m, 52 H), 5.53 (d, *J* = 12.0 Hz, 20 H), 5.43 (s, 4 H), 5.22-5.15 (m, 20 H), 3.90-3.86 (m, 24 H), 2.99-2.91 (m, 24 H), 0.82-0.80 (m, 108 H), 0.06-0.02 (m, 72 H).

¹³**C NMR** (100 MHz, CDCl₃) δ 157.8, 157.7, 153.2, 152.8, 151.9, 150.4, 149.8, 149.6, 143.7, 143.7, 135.1, 135.0, 133.3, 130.7, 130.6, 130.2, 130.1, 129.4, 129.3, 129.1, 128.0, 127.0, 126.7, 125.5, 124.9, 124.8, 124.7, 124.2, 115.8, 62.0, 61.9, 47.4, 47.3, 29.1, 25.9, 18.3, 5.4.

FTMS m/z calcd. for C₃₀₀H₃₇₆N₆₆O₂₂S₁₂Si₁₂ (M) 5974.42, found 998.68 (M+6H)⁶⁺, 1994.50 (M+3H)³⁺.



Scheme S9. Synthesis of 13.

Synthesis of M1: In a glove box, to an oven-dried vial was added 10 (0.6 mmol, 1.73 g, 1.0 eq.), BnN₃ (0.72 mmol, 0.09 g, 1.2 eq.), $[Ir(COD)Cl]_2$ (2 mol %) and DCE (0.1 M). The vial was capped and removed from the glove box. The reaction mixture was stirred at room temperature for 3 h until the reaction completed (confirmed by TLC), and then concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography to give the desired product M1 as white solid in 95% yield (1.58 g).

Rf = 0.4 (DCM/MeOH = 50:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.93 (d, *J* = 8.0 Hz, 4 H), 7.81 (s, 1 H), 7.40-7.27 (m, 14 H), 7.25-7.19 (m, 4 H), 7.15-7.13 (m, 2 H), 7.10-7.07 (m, 4 H), 6.93-6.89 (m, 14 H), 6.84-6.76 (m, 12 H), 5.52-5.47 (m, 12 H), 5.20-5.15 (m, 10 H), 3.93-3.86 (m, 12 H), 2.98-2.91 (m, 12 H), 0.82-0.80 (m, 54 H), 0.03 (d, *J* = 12.0 Hz, 36 H).

¹³**C NMR** (100 MHz, CDCl₃) δ 157.8, 157.6, 150.4, 149.6, 143.9, 143.7, 143.7, 135.0, 130.7, 130.6, 130.0, 129.4, 129.1, 128.6, 128.0, 127.8, 127.0, 126.8, 126.6, 125.5, 124.8, 124.7, 124.2, 115.8, 62.0, 61.9, 47.4, 47.3, 29.1, 25.8, 18.2, 5.4.



Synthesis of 13: the **M1** was dissolved in MeOH/DCM (2:1, v/v), added the HCl (12 M, 2.0 eq.) at 0 °C stirred 10 min. After the reaction completed, then concentrated under reduced pressure, diluted with DCM, washed with NaHCO₃ aqueous solution, dried over Na₂SO₄, filtered, and evaporated under vacuum. The residue was purified by column chromatography on silica gel to get **13** as white solid in 94% yield (1.17 g).

Rf = 0.5 (DCM/MeOH = 10:1).

¹**H NMR** (400 MHz, DMSO) δ 8.64 (s, 4 H), 8.54 (s, 1 H), 7.50-7.39 (m, 15 H), 7.26-7.24 (m, 3 H), 7.24-7.15 (m, 3 H), 7.11-7.08 (m, 5 H), 7.0-6.93 (m, 22 H), 6.88 (d, *J* = 8.0 Hz, 2 H), 5.56-5.53 (m, 12 H), 5.22-5.16 (m, 10 H), 4.75-4.70 (m, 6 H), 3.65-3.59 (m, 12 H), 2.82-2.74 (m, 12 H).

¹³**C NMR** (100 MHz, DMSO) δ 157.7, 149.7, 149.1, 149.0, 142.9, 142.8, 135.4, 134.9, 134.8, 132.9, 130.9, 130.7, 130.2, 130.1, 129.4, 129.3, 129.2, 129.0, 128.5, 127.8, 127.5, 127.1, 126.9, 126.3, 126.1, 126.0, 125.6, 124.3, 123.8, 123.3, 115.9, 61.1, 59.9, 59.8, 51.3, 47.2, 28.9.

ESI-MS m/z calcd. for C₁₁₇H₁₀₇N₃₃O₁₁S₆ (M) 2341.7152, found 2342.1785 (M+H)⁺, 2365.1450 (M+Na)⁺.



Scheme S10. Synthesis of 14.

Synthesis of M2: TsCl (9.0 eq.), Et_3N (12.0 eq.), and DMAP (0.12 eq.) were added in the solution of DCM and 13 (0.4 mmol, 0.94g, 1.0 eq.), stirred about 3 h, the reaction was completed, then concentrated under reduced pressure. The residue was purified by column chromatography on silica gel to get M2 as white solid in 89% yield.

Rf = 0.5(DCM/MeOH = 50:1)

¹**H NMR** (400 MHz, CDCl₃) δ 7.92 (s, 4 H), 7.80 (s, 1 H), 7.67-7.65 (m, 12 H), 7.40-7.22 (m, 30 H), 7.15-7.04 (m, 6 H), 6.95-6.82 (m, 24 H), 6.76-6.74 (m, 2 H), 5.53-5.48 (m, 12 H), 5.20 (t, *J* = 12.0 Hz, 10 H), 4.31-4.24 (m, 12 H), 3.07-3.01 (m, 12 H), 2.40 (s, 18 H).

¹³**C NMR** (100 MHz, CDCl₃) δ 158.1, 147.8, 146.9, 144.8, 143.8, 135.0, 132.6, 131.3, 130.8, 130.3, 129.8, 129.5, 129.3, 128.7, 127.8, 125.6, 124.8, 123.2, 116.0, 52.3, 47.7, 47.7, 25.4, 21.6.



Synthesis of 14: Added potassium azide (9.0 eq.) to the solution of M2 in DMF, heat 80 °C, stirred 3 h before a saturated aqueous NaCl solution was added. The aqueous phase was separated and extracted with ethyl acetate (EA) for three times. The combined organic phase was washed with brine, dried over Na₂SO₄ and evaporated under vacuum to give the which was then purified by silica gel flash column chromatography to give pure product 14 as white solid in 80% yield (642.7 mg).

Rf = 0.5 (DCM/MeOH = 50:1)

¹**H NMR** (400 MHz, CDCl₃) δ 7.90 (d, *J* = 8.0 Hz, 4 H), 7.79 (s, 1 H), 7.44-7.35 (m, 10 H), 7.31 (d, *J* = 4.0 Hz, 4 H), 7.25-7.23 (m, 3 H), 7.19-7.13 (m, 6 H), 7.04 (d, *J* = 8.0 Hz, 4 H), 6.93-6.83 (m, 21 H), 6.79-6.77 (m, 2 H), 5.56 (s, 8 H), 5.53 (d, *J* = 4.0 Hz, 4 H), 5.20 (t, *J* = 8.0 Hz, 10 H), 3.60-3.54 (m, 12 H), 2.99-2.94 (m, 12 H).

¹³**C NMR** (100 MHz, CDCl₃) δ 158.0, 148.5, 143.8, 135.1, 130.9, 130.4, 129.7, 129.5, 128.7, 128.3, 127.9, 127.1, 127.0, 125.7, 124.8, 124.7, 123.7, 116.0, 116.0, 61.9, 49.9, 47.7, 25.5.



Scheme S11. Synthesis of 15a.

Synthesis of 15a: M4 (0.29 mmol, 111.2 mg, 7.2 eq.) and **14** (0.04 mmol, 100 mg, 1.0 eq.) were added into the solution of DMF (0.1 M) involving CuBr (1.2 eq.), PMDETA (1.2 eq.), NaAsc (6.0 eq.). The

reaction mixture was stirred for 25 min at room temperature. The solution was then cooled to room temperature, diluted with ethyl acetate, washed with brine (three times), dried over Na₂SO₄, filtered, and evaporated under vacuum. The residue was purified by column chromatography on silica gel to get **15a** as white solid in 51% yield (96.2 mg).

Rf = 0.4 (DCM/MeOH = 40:1)

¹**H NMR** (400 MHz, CDCl₃) δ 7.88 (s, 4 H), 7.77 (s, 1 H), 7.52-7.51 (m, 5 H), 7.46 (s, 1 H), 7.34-7.30 (m, 11 H), 7.21-7.16 (m, 16 H), 7.09-6.91 (m, 103 H), 6.81-6.75 (m, 20 H), 6.67-6.65 (m, 14 H), 5.52 (t, *J* = 8.0 Hz, 12 H), 5.16 (t, *J* = 12.0 Hz, 10 H), 5.00 (t, *J* = 4.0 Hz, 12 H), 4.73-4.68 (m, 12 H), 3.29-3.23 (m, 12 H).

FTMS m/z calcd. for C₂₉₁H₂₃₃N₅₁O₁₁S₆ (M) 4808.76, found 1604.97 (M+3H)³⁺, 2407.45 (M+2H)²⁺.



Scheme S12. Synthesis of 15b.

Synthesis of 15b: M5 (0.29 mmol, 62.1 mg, 7.2 eq.) and **14** (0.04 mmol, 100 mg, 1.0 eq.) were added into the solution of DMF (0.1 M) involving CuBr (1.2 eq.), PMDETA (1.2 eq.), NaAsc (6.0 eq.). The reaction mixture was stirred at room temperature. The solution was then cooled to room temperature, diluted with ethyl acetate, washed with brine (three times), dried over Na₂SO₄, filtered, and evaporated under vacuum. The residue was purified by column chromatography on silica gel to get **15b** as white solid in 73% yield (105.7 mg).

Rf = 0.3 (DCM/MeOH = 40:1).

¹**H NMR** (400 MHz, CDCl₃) δ 7.96 (d, *J* = 4.0 Hz, 4 H), 7.82 (s, 1 H), 7.65 (t, *J* = 4.0 Hz, 5 H) 7.58 (s, 1 H), 7.48-7.45 (m, 6 H), 7.41-7.29 (m, 14 H), 7.24-7.10 (m, 3 H), 7.09-7.04 (m, 6 H), 6.93-6.71 (m, 37 H), 6.64-6.61 (m, 2 H), 6.11-6.08 (m, 6 H), 5.52-5.49 (m, 12 H), 5.20-5.13 (m, 22 H), 4.77-4.73

(m, 12 H), 3.32,-3.24 (m, 12 H), 2.37-2.35 (m, 18 H).

FTMS m/z calcd. for C₁₉₅H₁₆₁N₅₁O₂₃S₆ (M) 3776.13, found 1260.45 (M+3H)³⁺, 1891.31 (M+2H)²⁺.

V. SEC Traces



Figure S3. GPC traces of 1~12.

VI. NMR Spectra



Figure S4. Comparison of ¹H NMR for **1-12**.



Figure S6. ¹³C NMR spectra of T2 (CDCl₃, 100 MHz).



Figure S8. ¹³C NMR spectra of T3 (CDCl₃, 100 MHz).



Figure S10. ¹³C NMR spectra of T1-A1 (CDCl₃, 100 MHz).



Figure S12. ¹³C NMR spectra of T1-A2 (CDCl₃, 100 MHz).



Figure S14. ¹³C NMR spectra of 1 (CDCl₃, 100 MHz).



Figure S15. ¹H NMR spectra of T1-A3-T1' (CDCl₃, 400 MHz).



Figure S16. ¹³C NMR spectra of T1-A3-T1' (CDCl₃, 100 MHz).



Figure S17. ¹H NMR spectra of T1-A3-T" (CDCl₃, 400 MHz).



Figure S18. ¹³C NMR spectra of **T1-A3-T**" (CDCl₃, 100 MHz).



Figure S20. ¹³C NMR spectra of 2 (CDCl₃, 100 MHz).





Figure S22. ¹³C NMR spectra of 3 (CDCl₃, 100 MHz).





Figure S24. ¹³C NMR spectra of 4 (CDCl₃, 100 MHz).



Figure S26. ¹³C NMR spectra of 5 (CDCl₃, 100 MHz).





Figure S28. ¹³C NMR spectra of 6 (CDCl₃, 100 MHz).



Figure S30. ¹³C NMR spectra of 7 (CDCl₃, 100 MHz).



Figure S32. ¹³C NMR spectra of 8 (CDCl₃, 100 MHz).





Figure S34. ¹³C NMR spectra of 9 (CDCl₃, 100 MHz).



Figure S36. ¹³C NMR spectra of 10 (CDCl₃, 100 MHz).



Figure S38. ¹³C NMR spectra of 11 (CDCl₃, 100 MHz).



160 150 140 130 120 110 100 f1 (ppm)

Figure S40. ¹³C NMR spectra of 12 (CDCl₃, 100 MHz).



Figure S42. ¹³C NMR spectra of M1 (CDCl₃, 100 MHz).



Figure S43. ¹H NMR spectra of 13 (DMSO,400 MHz).





Figure S44. ¹³C NMR spectra of 13 (DMSO, 100 MHz).



Figure S46. ¹³C NMR spectra of M2 (CDCl₃, 100 MHz).



Figure S48. ¹³C NMR spectra of 14 (CDCl₃, 100 MHz).

180 170 160 150 140 130 120 110 100 f1 (ppm)

210

200

190

90

80 70

60

10

 $\begin{array}{c} 7.88\\ 7.75\\ 7.51\\ 7.51\\ 7.55\\ 7.75\\ 7.75\\ 7.75\\ 7.75\\ 7.75\\ 7.75\\ 7.75\\ 7.75\\ 7.75\\ 7.75\\ 7.72\\ 7.70\\$



Figure S49. ¹H NMR spectra of 15a (CDCl₃, 400 MHz).

$\begin{array}{c} 7.96\\ 6.77\\ 7.75\\$



Figure S50. ¹H NMR spectra of 15b (CDCl₃, 400 MHz).

VII. References

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