Supporting Information

Sustainable synthesis of long-acting local anesthetics ropivacaine and levobupivacaine under batch and continuous flow via asymmetric hydrogenation

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1. General Information

All commercially available reagents were used without further purification. Tetrahydrofuran and toluene were dried with sodium chips and indicated by benzophenone, other anhydrous solvents were purchased from Aladdin. Chromatography was conducted by using 300–400 mesh silica gel. All new compounds were characterized by NMR spectroscopy, high resolution mass spectrometry (HRMS), FT-IR spectroscopy and melting point (if solids). NMR spectra were recorded on a 400 MHz NMR spectrometer. Reference values for residual solvents were taken as $\delta = 7.26$ (CDCl₃) ppm, $\delta = 2.50$ (DMSO-*d*₆) ppm for ¹H NMR and $\delta = 77.16$ (CDCl₃) ppm, $\delta = 49.00$ (MeOH-*d*₄) ppm, $\delta = 39.52$ (DMSO-*d*₆) ppm for ¹³C NMR. Coupling constants (*J*) were given in Hz and multiplicities for coupled signals were denoted as: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad and dd = double doublet etc. Infrared (IR) spectra were recorded on a Perkin Elmer Spectrum Two FT-IR spectrometer. High-resolution mass spectra (HRMS) were recorded on a Bruker microTOF Q III by the ESI method. Melting points (m.p.) were recorded on an SRS-optic melting point apparatus. Chiral HPLC was performed using a Daicel Chiralcel IC column, Chiralcel OJ-H column, Chiralcel AD-H column and Chiralcel OD-H column.

2. Experimental Procedures

2.1 General Procedures for Picolinate esters

2-ester–substituted pyridines were synthesized from 2-carboxypyridines with the corresponding alcohols via esterification. Methyl picolinate **1a** was commercially available.



picolinic acid (500.0 mg, 4.1 mmol, 1.0 equiv.), 4-DMAP (248.1 mg, 2.0 mmol, 0.5 equiv.) and EDCI (1.2 g, 6.1 mmol, 1.5 equiv.) were dissolved in dichloromethane (15 mL) and stirred at 0 °C for 15 min. Alcohol (4.5 mmol, 1.1 equiv.) was added to the reaction mixture that was then stirred at 25 °C for 12 h. Water was added and the solution was extracted with dichloromethane. The combined organic layer was washed with brine, dried over anhydrous Na_2SO_4 , filtered and concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 100:1) to give the desired products.

Isopropyl picolinate (1b): 590.4 mg, 88% yield, colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.73 - 8.72 (m, 1H), 8.09 - 8.06 (m, 1H), 7.81 - 7.62 (m, 1H), 7.43 - 7.40 (m, 1H), 5.35 - 5.26 (m, 1H), 1.38 (d, *J* = 6.0 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 164.8, 149.9, 148.7, 136.9, 126.7, 125.1, 69.6, 21.9. HRMS (ESI) m/z calcd for C₉H₁₁NNaO₂ [M + Na]⁺: 188.0682, found: 188.0680.

tert-Butyl picolinate (1c): 727.9 mg, 93% yield, colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.75 \sim - 8.73 (m, 1H), 8.05 (dd, *J* = 7.6, 2.4 Hz, 1H), 7.83 - 7.78 (m, 1H), 7.44 - 7.41 (m, 1H), \sim CO_{2/Bu} 1.64 (s, 9H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 164.4, 149.9, 149.8, 136.9, 126.5, 124.9, 82.4, 28.3. HRMS (ESI) m/z calcd for C₁₀H₁₃NNaO₂ [M + Na]⁺: 202.0838, found: 202.0837.

Cyclopropylmethyl picolinate (1d): 618.9 mg, 86% yield, colorless oil. ¹H NMR (400 MHz,

Chloroform-*d*) δ 8.72 (t, *J* = 3.6 Hz, 1H), 8.10 (dd, *J* = 7.6, 2.4 Hz, 1H), 7.81 – 7.77 (m, 1H), 7.44 – 7.40 (m, 1H), 4.20 (d, *J* = 7.6 Hz, 2H), 1.31 – 1.26 (m, 1H), 058 – 0.56 (m, 2H), 0.39 – 0.29 (m, 2H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 165.4, 149.9,

148.4, 137.0, 126.8, 125.2, 70.8, 10.0, 3.6. HRMS (ESI) m/z calcd for $C_{10}H_{11}NNaO_2$ [M + Na]⁺: 200.0682, found: 200.0684.

Cyclopentylmethyl picolinate (1e): 758.6 mg, 91% yield, colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.78 – 8.68 (m, 1H), 8.07 (dd, *J* = 8.0, 2.4 Hz, 1H), 7.80 (t, *J* = 7.6 Hz, 1H), 7.47 – 7.38 (m, 1H), 4.27 (d, *J* = 7.6 Hz, 2H), 2.42 – 2.34 (m, 1H), 1.87 – 1.72 (m, 2H), 1.66 – 1.50 (m, 4H), 1.39 – 1.25 (m, 2H). ¹³C NMR (100 MHz,

Chloroform-*d*) δ 165.3, 150.0, 148.4, 137.0, 126.8, 125.1, 69.8, 38.7, 29.5, 25.3. HRMS (ESI) m/z calcd for C₁₂H₁₅NNaO₂ [M + Na]⁺: 228.0995, found: 228.0998.

 Cyclohexylmethyl picolinate (1f): 801.5 mg, 90% yield, colorless oil. ¹H NMR (400 MHz, Chloroform

 d) δ 8.73 – 8.71 (m, 1H), 8.07 (dd, J = 7.6, 2.4 Hz, 1H), 7.84 – 7.74 (m, 1H), 7.46 –

 7.38 (m, 1H), 4.18 (d, J = 4.0 Hz, 2H), 1.86 – 1.58 (m, 6H), 1.28 – 0.93 (m, 5H). ¹³C

 NMR (100 MHz, Chloroform-d) δ 165.3, 150.0, 148.4, 136.9, 126.8, 125.1, 70.9,

 $37.1,\,29.8,\,26.4,\,25.7.\ HRMS\ (ESI)\ m/z\ calcd\ for\ C_{13}H_{17}NNaO_2\ [M+Na]^+:\,242.1151,\ found:\ 242.1149.$

Benzyl picolinate (1g): 831.4 mg, 96% yield, colorless oil. ¹H NMR (400 MHz, Chloroform-d) δ 8.76
 - 8.74 (m, 1H), 8.17 - 8.06 (m, 1H), 7.87 - 7.72 (m, 1H), 7.53 - 7.24 (m, 6H), 5.45 (s, 2H). ¹³C NMR (100 MHz, Chloroform-d) δ 165.0, 149.9, 148.0, 137.0, 135.6, 128.6, 128.5, 128.4, 126.9, 125.2, 67.5. HRMS (ESI) m/z calcd for C₁₃H₁₁NNaO₂

[M + Na]⁺: 236.0682, found: 236.0681.

4-(Trifluoromethyl)benzyl picolinate (1h): 993.7 mg, 87% yield, white solid, m.p. = 70.2 - 72.4 °C. $\int_{0}^{CF_{3}} {}^{1}H NMR (400 MHz, Chloroform-d) \delta 8.63 - 8.61 (m, 1H), 8.00 (d, J = 8.0 Hz, 1H), 7.71 - 7.67 (m, 1H), 7.50 - 7.45 (m, 4H), 7.35 - 7.32 (m, 1H), 5.36 (s, 2H).$ ${}^{13}C NMR (100 MHz, Chloroform-d) \delta 164.7, 149.8, 147.5, 139.5, 136.9, 130.2$ (q, J = 32.2 Hz), 128.3, 127.0, 125.4 (q, J = 3.7 Hz), 125.2, 123.9 (q, J = 270.5 Hz), 66.2. HRMS (ESI) $m/z \text{ calcd for } C_{14}H_{10}F_{3}NNaO_{2} [M + Na]^{+}: 304.0556, \text{ found: } 304.0553.$

4-Cyanobenzyl picolinate (1i): 822.5 mg, 85% yield, white solid, m.p. = 94.0 – 95.4 °C. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.77 – 8.69 (m, 1H), 8.11 (d, *J* = 8.0 Hz, 1H), 7.85 – 7.80 (m, 1H), 7.64 – 7.61 (m, 2H), 7.57 – 7.52 (m, 2H), 7.49 – 7.46 (m, 1H), 5.45 (s, 2H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 164.8, 150.0, 147.5, 140.9, 137.2,

132.4, 128.6, 127.3, 125.4, 118.5, 112.1, 66.2. HRMS (ESI) m/z calcd for $C_{14}H_{10}N_2NaO_2$ [M + Na]⁺: 261.0634, found: 261.0634.

4-(Methoxycarbonyl)benzyl picolinate (1j): 980.6 mg, 89% yield, white solid, m.p. = 60.3 - 62.6 °C. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.74 - 8.72 (m, 1H), 8.10 (d, *J* = 8.0 Hz, 1H), 8.00 (d, *J* = 8.0 Hz, 2H), 7.80 (t, *J* = 8.0 Hz, 1H), 7.50 (d, *J* = 8.4 Hz, 2H), 7.47 - 7.41 (m, 1H), 5.46 (s, 2H), 3.86 (s, 3H). ¹³C NMR (100 MHz, 100 MHz Chloroform-*d*) δ 166.7, 164.9, 150.0, 147.7, 140.7, 137.1, 130.1, 129.9, 128.0, 127.1, 125.3, 66.7, 52.2. HRMS (ESI) m/z calcd for C₁₅H₁₃NNaO₄ [M + Na]⁺: 294.0737, found: 294.0740.

4-Nitrobenzyl picolinate (1k): 943.9 mg, 90% yield, white solid, m.p. = 126.1 - 127.8 °C. ¹H NMR



(400 MHz, Chloroform-*d*) δ 8.84 – 8.75 (m, 1H), 8.24 (d, *J* = 8.0 Hz, 2H), 8.17 (d, *J* = 8.0 Hz, 1H), 7.91 – 7.87 (m, 1H), 7.66 (d, *J* = 8.4 Hz, 2H), 7.58 – 7.49 (m, 1H), 5.56 (s, 2H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 164.8, 150.1, 147.8,

147.5, 142.9, 137.2, 128.7, 127.4, 125.5, 123.9, 65.9. HRMS (ESI) m/z calcd for $C_{13}H_{10}N_2NaO_4$ [M + Na]⁺: 281.0533, found: 281.0535.

4-Methylbenzyl picolinate (11): 784.6 mg, 85% yield, colorless oil. ¹H NMR (400 MHz, Chloroform *d*) δ 8.71 – 8.59 (m, 1H), 8.02 (dd, *J* = 7.6, 2.0 Hz, 1H), 7.72 – 7.68 (m, 1H), 7.41 – 7.24 (m, 3H), 7.08 (d, *J* = 8.0 Hz, 2H), 5.32 (s, 2H), 2.25 (s, 3H). ¹³C NMR

 $^{\circ}$ (100 MHz, Chloroform-*d*) δ 165.0, 149.9, 148.1, 138.3, 137.0, 132.7, 129.3, 128.8, 126.9, 125.2, 67.5, 21.2. HRMS (ESI) m/z calcd for C₁₄H₁₃NNaO₂ [M + Na]⁺: 250.0838, found: 250.0834.

3-Methylbenzyl picolinate (1m): 803.0 mg, 87% yield, colorless oil. ¹H NMR (400 MHz, Chloroformd) δ 8.68 – 8.66 (m, 1H), 8.03 (dd, J = 8.0, 2.4 Hz, 1H), 7.74 – 7.69 (m, 1H), 7.39 – 7.32 (m, 1H), 7.20 – 7.16 (m, 3H), 7.05 (d, J = 7.2 Hz, 1H), 5.33 (s, 2H), 2.26 (s, 3H). ¹³C NMR (100 MHz, Chloroform-d) δ 165.0, 150.0, 148.1, 138.3, 137.0,

135.5, 129.4, 129.2, 128.5, 126.9, 125.7, 125.3, 67.6, 21.4. HRMS (ESI) m/z calcd for $C_{14}H_{13}NNaO_2$ [M + Na]⁺: 250.0838, found: 250.0842.

3-Methoxybenzyl picolinate (1n): 879.3 mg, 89% yield, colorless oil. ¹H NMR (400 MHz, Chloroformd) δ 8.71 – 8.58 (m, 1H), 8.03 (d, J = 7.6 Hz, 1H), 7.77 – 7.67 (m, 1H), 7.44 – 7.31 (m, 1H), 7.21 – 7.17 (m, 1H), 7.02 – 6.91 (m, 2H), 6.77 (d, J = 8.8 Hz, 1H), 5.33 (s, 2H), 3.70 (s, 3H). ¹³C NMR (100 MHz, Chloroform-d) δ 165.0, 159.7,

149.9, 147.9, 137.1, 137.0, 129.7, 127.0, 125.3, 120.7, 114.0, 114.0, 67.3, 55.2. HRMS (ESI) m/z calcd for $C_{14}H_{13}NNaO_3$ [M + Na]⁺: 266.0788, found: 266.0784.

4-Bromobenzyl picolinate (10): 1.0 g, 87% yield, colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ Br 8.71 - 8.64 (m, 1H), 8.04 (d, *J* = 8.0 Hz, 1H), 7.77 - 7.21 (m, 1H), 7.42 - 7.38 (m, 3H), 7.27 (d, *J* = 8.0 Hz, 2H), 5.31 (s, 2H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 165.0, 150.0, 147.8, 137.1, 134.7, 131.8, 130.3, 127.1, 125.3, 122.5, 66.7.

HRMS (ESI) m/z calcd for $C_{13}H_{10}BrNNaO_2 [M + Na]^+: 313.9787$, found: 313.9786.

3-Bromobenzyl picolinate (1p): 1.0 g, 85% yield, colorless oil. ¹H NMR (400 MHz, Chloroform-d) δ



8.72 – 8.62 (m, 1H), 8.06 (d, J = 8.0 Hz, 1H), 7.78 – 7.74 (m, 1H), 7.55 (s, 1H), 7.43 – 7.30 (m, 3H), 7.18 – 7.14 (m, 1H), 5.33 (s, 2H). ¹³C NMR (100 MHz, Chloroform-d) δ 164.9, 150.0, 147.8, 137.9, 137.1, 131.6, 131.5, 130.3, 127.2,

127.2, 125.4, 122.7, 66.6. HRMS (ESI) m/z calcd for $C_{13}H_{10}BrNNaO_2$ [M + Na]⁺: 313.9787, found: 313.9792.

2-Bromobenzyl picolinate (1q): 996.6 mg, 84% yield, colorless oil. ¹H NMR (400 MHz, Chloroformd) δ 8.83 – 8.75 (m, 1H), 8.16 (d, J = 8.0 Hz, 1H), 7.87 – 7.83 (m, 1H), 7.59 (d, J = 8.0 Hz, 1H), 7.56 – 7.46 (m, 2H), 7.35 – 7.31 (m, 1H), 722 – 7.18 (m, 1H), 5.54 (s, 2H). ¹³C NMR (100 MHz, Chloroform-d) δ 164.8, 150.1, 147.9, 137.1, 135.0, 132.9, 130.1, 129.9, 127.7, 127.1, 125.5, 123.5, 67.0. HRMS (ESI) m/z calcd for C₁₃H₁₀BrNNaO₂ [M + Na]⁺: 313.9787, found: 313.9791.

4-Chlorobenzyl picolinate (1r): 865.1 mg, 86% yield, colorless oil. ¹H NMR (400 MHz, Chloroform-



d) δ 8.77 – 8.69 (m, 1H), 8.10 (d, *J* = 8.0Hz, 1H), 7.83 – 7.78 (m, 1H), 7.46 – 7.39 (m, 3H), 7.33 – 7.26 (m, 2H), 5.39 (s, 2H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 164.9, 149.9, 147.7, 137.0, 134.3, 134.1, 129.9, 128.7, 127.0, 125.2, 66.6.

HRMS (ESI) m/z calcd for $C_{13}H_{10}CINNaO_2 [M + Na]^+$: 270.0292, found: 270.0291.

4-Fluorobenzyl picolinate (1s): 854.6 mg, 91% yield, colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.75 – 8.73 (m, 1H), 8.12 – 8.09 (m, 1H), 7.83 – 7.79 (m, 1H), 7.51 – 7.41 (m, 3H), 7.10 – 6.98 (m, 2H), 5.40 (s, 2H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 165.1, 162.9 (d, *J* = 245.5 Hz), 150.1, 148.0, 137.1, 131.6 (d, *J* = 3.3 Hz), 130.8

(d, *J* = 8.3 Hz), 127.1, 125.4, 115.6 (d, *J* = 21.6 Hz), 66.9. HRMS (ESI) m/z calcd for C₁₃H₁₀FNNaO₂ [M + Na]⁺: 254.0588, found: 254.0592.

2.2 General Procedures for the synthesis of pyridinium salts

A mixture of 2-substituted pyridine (1.0 mmol), benzyl bromide (1.2 mmol) and 10.0 mL acetone or methanol was stirred at 0 °C for 12-96 h. Ether was added, the resulting precipitate was collected and rinsed with ethyl acetate to give the solid product which was directly used for the hydrogenation. If the desired product was not precipitated, the reaction mixture was purified by column chromatography on silica gel using DCM/MeOH (20:1) to give the desired products (35%–96% yield).

2.3 General Procedure for Hydrogenation of pyridinium salts



A mixture of $[Ir(COD)Cl]_2$ (1.3 mg, 2.0 µmol, 2.0 mol%) and (*R*,*R*)-BDPP (L6) (1.8 mg, 4.0 µmol, 4.0 mol%) were dissolved in degassed MeOAc (2.0 mL) at argon atmosphere, and the resulting solution was allowed to stirred for 20 min, followed by the addition of the substrate **2** (0.1 mmol, 1.0 equiv.) and KI (16.6 mg, 0.1 mmol, 1.0 equiv.). The resulting mixture was transferred to an autoclave, which was purged (3 × 5 atm) and charged with H₂ (600 psi), then the reaction mixtures were stirred at -20 °C for 72 h. After careful release of the hydrogen gas, the reaction mixture was filtrated and concentrated in vacuo. Flash chromatography on silica gel using petroleum ether/ethyl acetate as the eluent gave the products. The enantiomeric excesses were determined by chiral HPLC.



Asymmetric hydrogenation of *N*-benzyl-2-(methoxycarbonyl)pyridinium salts (**2a**) at gram scale: A mixture of $[Ir(COD)Cl]_2$ (5.5 mg, 8.0 µmol, 0.25 mol%) and (*R*,*R*)-BDPP (**L6**) (7.0 mg, 16.0 µmol, 0.5 mol%) were dissolved in degassed MeOAc (30.0 mL) at argon atmosphere, and the resulting solution

was allowed to stirred for 20 min, followed by the addition of the substrate **2a** (1.0 g, 3.2 mmol, 1.0 equiv.). The resulting mixture was transferred to an autoclave, which was purged (3×5 atm) and charged with H₂ (600 psi), then the reaction mixtures were stirred at -20 °C for 72 h. After careful release of the hydrogen gas, the reaction mixture was filtrated and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 150:1) to give the desired product **3a** (711.6 mg, 94% yield, 95% *ee*) as a pale-yellow oil.

Asymmetric hydrogenation of *N*-benzyl-2-((benzyloxy)carbonyl)pyridinium salts (**2g**) at gram scale: A mixture of $[Ir(COD)Cl]_2$ (4.2 mg, 6.5 µmol, 0.25 mol%) and (*R*,*R*)-BDPP (**L6**) (5.8 mg, 13.0 µmol, 0.5 mol%) were dissolved in degassed MeOAc (30.0 mL) at argon atmosphere, and the resulting solution was allowed to stirred for 20 min, followed by the addition of the substrate **2g** (1.0 g, 2.6 mmol, 1.0 equiv.). The resulting mixture was transferred to an autoclave, which was purged (3 × 5 atm) and charged with H₂ (600 psi), then the reaction mixtures were stirred at -20 °C for 72 h. After careful release of the hydrogen gas, the reaction mixture was filtrated and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 150:1) to give the desired product **3g** (773.0 mg, 96% yield, 93% *ee*) as a pale-yellow oil.

(*R*)-1-Benzylpiperidine-2-carboxylate methyl ester (3a): 21.7 mg, 93% yield, 95% *ee*, $[\alpha]_D^{20} = 80.7$ (c = 1.0, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.36 – 7.18 (m, 5H), 3.78 (d, *J* = 13.6 Hz, 1H), 3.73 (s, 3H), 3.41 (d, *J* = 13.6 Hz, 1H), 3.17 (dd, *J* = 7.6, 4.4 Hz, 1H), 2.96 – 2.91 (m, 1H), 2.19 – 2.13 (m, 1H), 1.90 – 1.77 (m, 2H), 1.66 – 1.50 (m, 3H), 1.42 – 1.32 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 174.5, 138.3, 129.3, 128.2, 127.1, 64.5, 60.7, 51.6, 50.2, 29.7, 25.4, 22.6. HRMS (ESI) m/z calcd for C₁₄H₂₀NO₂ [M + H]⁺: 234.1489, found: 234.1490. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 90/10, flow = 0.5 mL/min, retention time 8.8 min and 11.4 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate isopropyl ester (3b)¹: 23.0 mg, 88% yield, 91% *ee*, $[\alpha]_D^{20} = 80.1$ (c = 0.5, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.34 – 7.23 (m, 5H), 5.16 – 5.06 (m, 1H), 3.81 (d, *J* = 13.2 Hz, 1H), 3.39 (d, *J* = 13.2 Hz, 1H), 3.09 (t, *J* = 6.4 Hz, 1H), 2.96 – 2.90 (m, 1H), 2.16 – 2.10 (m, 1H), 1.90 – 1.46 (m, 6H), 1.30 – 1.26 (m, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.7, 138.5, 129.4, 128.2, 127.1, 67.8, 64.8, 60.6, 50.3, 29.7, 25.4, 22.7, 22.1, 22.0. HRMS (ESI) m/z calcd for C₁₆H₂₄NO₂ [M + H]⁺: 262.1802, found: 262.1805. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 8.6 min and 9.6 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate tert-butyl ester (3c)²: 25.6 mg, 93% yield, 96% *ee*, $[\alpha]_D^{20} = 76.1$ (c = 0.5, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.41 – 7.21 (m, 5H), 3.83 (d, *J* = 13.2 Hz, 1H), 3.39 (d, *J* = 13.2 Hz, 1H), 3.01 (dd, *J* = 7.6, 4.4 Hz, 1H), 3.03 – 2.89 (m, 1H), 2.15 – 2.09 (m, 1H), 1.86 – 1.76 (m, 2H), 1.66 – 1.53 (m, 3H), 1.49 (s, 9H), 1.40 – 1.32 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.4, 138.9, 129.3, 128.2, 127.0, 80.6, 65.1, 60.5, 50.2, 29.8, 28.3, 25.5, 22.7. HRMS (ESI) m/z calcd for C₁₇H₂₆NO₂ [M + H]⁺: 276.1958, found: 276.1960. HPLC: Chiralcel OJ-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 7.7 min and 8.2 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate cyclopropylmethyl ester (3d): 23.5 mg, 86% yield, 92% *ee*, $\begin{bmatrix} \alpha \end{bmatrix}_{D}^{20} = 69.4 (c = 0.5, CHCl_3), colorless oil. ¹H NMR (400 MHz, Chloroform-$ *d* $) \delta 7.35 - 7.24 (m, 5H), 4.09 - 3.92 (m, 2H), 3.82 (d,$ *J*= 13.2 Hz, 1H), 3.42 (d,*J*= 13.2 Hz, 1H), 3.15 (dd,*J*= 8.0, 4.4 Hz, 1H), 2.97 - 2.92 (m, 1H), 2.20 - 2.11 (m, 1H), 1.91 - 1.79 (m, 2H), 1.58 - 1.53 (m, 3H), 1.42 - 1.32 (m, 1H), 1.24 - 1.15 (m, 1H), 0.59 - 0.55 (m, 2H), 0.35 - 0.29 (m, 2H). ¹³C NMR (100 MHz, Chloroform-*d* $) \delta 174.3, 138.3, 129.5, 128.3, 127.2, 69.3, 64.8, 60.7, 50.4, 29.8, 25.4, 22.7, 10.1, 3.48, 3.46. HRMS (ESI) m/z calcd for C₁₇H₂₄NO₂ [M + H]⁺: 274.1802, found: 274.1802. HPLC: Chiralcel OJ-H column, 254 nm, 30 °C,$ *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 9.0 min and 10.5 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate cyclopentylmethyl ester (3e): 27.4 mg, 91% yield, 92% *ee*, $\left[\alpha\right]_{D}^{20} = 61.3 \text{ (c} = 0.5, \text{ CHCl}_3\text{), colorless oil. }^{1}\text{H NMR} (400 \text{ MHz, Chloroform-}d) \delta$ 7.40 - 7.26 (m, 4H), 7.25 - 7.21 (m, 1H), 4.04 (d, *J* = 7.2 Hz, 2H), 3.81 (d, *J* = 13.6 Hz, 1H), 3.44 (d, *J* = 13.6 Hz, 1H), 3.17 (dd, *J* = 7.2, 4.8 Hz, 1H), 2.96 - 2.91 (m,

1H), 2.30 - 2.11 (m, 2H), 1.85 - 1.72 (m, 4H), 1.61 - 1.55 (m, 8H), 1.46 - 1.32 (m, 2H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 174.3, 138.5, 129.4, 128.3, 127.1, 68.5, 64.5, 60.7, 50.2, 38.8, 29.9, 29.6, 25.51, 25.50, 22.6. HRMS (ESI) m/z calcd for C₁₉H₂₈NO₂ [M + H]⁺: 302.2115, found: 302.2119. HPLC: Chiralcel AD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 8.7 min and 9.3 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate cyclohexylmethyl ester (3f): 28.4 mg, 90% yield, 94% *ee*, $\begin{bmatrix} \alpha \end{bmatrix}_{D}^{20} = 78.1 (c = 0.5, CHCl_3), colorless oil. ¹H NMR (400 MHz, Chloroform-$ *d* $) \delta 7.35 - 7.27 (m, 4H), 7.25 - 7.24 (m, 1H), 3.95 (d,$ *J*= 6.4 Hz, 2H), 3.81 (d,*J*= 13.2 Hz, 1H), 3.44 (d,*J*= 13.2 Hz, 1H), 3.17 (t,*J*= 6.4 Hz, 1H), 2.96 - 2.93 (m, 1H), 2.20 - 2.15 (m, 1H), 1.85 - 1.80 (m, 2H), 1.80 - 1.64 (m, 6H), 1.57 - 1.52 (m, 3H), 1.42 - 1.34 (m, 1H), 1.25 - 1.11 (m, 3H), 1.03 - 0.94 (m, 2H). ¹³C NMR (100 MHz, Chloroform-*d* $) <math>\delta$ 174.3, 138.5, 129.3, 128.3, 127.1, 69.7, 64.5, 60.7, 50.2, 37.3, 29.92, 29.89, 26.5, 25.8, 25.5, 22.7. HRMS (ESI) m/z calcd for

C₂₀H₃₀NO₂ [M + H]⁺: 316.2271, found: 316.2272. HPLC: Chiralcel OJ-H column, 254 nm, 30 °C, n-

(*R*)-1-Benzylpiperidine-2-carboxylate benzyl ester $(3g)^3$: 29.7 mg, 96% yield, 93% *ee*, $[\alpha]_D^{20} = 64.9$ (c = 0.5, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.43 – 7.18 (m, 10H), 5.18 (s, 2H), 3.79 (dd, *J* = 13.2, 2.0 Hz, 1H), 3.42 (d, *J* = 13.2 Hz, 1H), 3.21 (t, *J* = 6.0 Hz, 1H), 2.98 – 2.92 (m, 1H), 2.25 – 2.11 (m, 1H), 1.86 – 1.82 (m, 2H), 1.69 – 1.47 (m, 3H), 1.42 – 1.33 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.8, 138.4, 136.1, 129.3, 128.7, 128.5, 128.4, 128.2, 127.1, 66.2, 64.2, 60.6, 50.2, 29.7, 25.4, 22.5. HRMS (ESI) m/z calcd for C₂₀H₂₄NO₂ [M + H]⁺: 310.1802, found: 310.1805. HPLC: Chiralcel OJ-H column, 254 nm, 30 °C, *n*-

hexane/i-propanol = 95/5, flow = 0.5 mL/min, retention time 19.3 min and 21.8 min (maj).

hexane/i-propanol = 95/5, flow = 0.5 mL/min, retention time 7.5 min and 8.0 min (maj).

(R)-1-Benzylpiperidine-2-carboxylate-4-(trifluoromethyl)benzyl ester (3h)⁴: 32.8 mg, 87% yield, 93%

ee, $[\alpha]_D^{20} = 86.3$ (c = 0.5, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform*d*) δ 7.61 (d, *J* = 8.0 Hz, 2H), 7.47 (d, *J* = 8.0 Hz, 2H), 7.30 – 7.24 (m, 5H), 5.21 (s, 2H), 3.78 (d, *J* = 13.2 Hz, 1H), 3.43 (d, *J* = 13.2 Hz, 1H), 3.25 (dd, *J* = 7.2,

4.8 Hz, 1H), 2.98 – 2.93 (m, 1H), 2.23 – 2.18 (m, 1H), 1.88 – 1.82 (m, 2H), 1.62 – 1.53 (m, 3H), 1.44 – 1.35 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.6, 140.1, 138.3, 130.6 (q, *J* = 32.1 Hz), 129.2, 128.4, 128.3, 127.2, 125.7 (q, *J* = 3.7 Hz), 124.1 (q, *J* = 270.4 Hz), 65.2, 64.2, 60.7, 50.2, 29.7, 25.4,

22.5. HRMS (ESI) m/z calcd for $C_{21}H_{23}F_3NO_2$ [M + H]⁺: 378.1675, found: 378.1673. HPLC: Chiralcel OJ-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 17.8 min and 22.0 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate 4-cyanobenzyl ester (3i): 28.4 mg, 85% yield, 92% *ee*, $[\alpha]_D^{20} = 93.1$ (c = 1.0, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.64 (d, J = 8.4 Hz, 2H), 7.46 (d, J = 8.0 Hz, 2H), 7.34 – 7.20 (m, 5H), 5.20 (s, 2H), 3.77 (d, J = 13.2 Hz, 1H), 3.43 (d, J = 13.6 Hz, 1H), 3.25 (dd, J = 7.2, 4.8 Hz, 1H), 2.98 – 2.93 (m, 1H), 2.24 – 2.18 (m, 1H), 1.87 – 1.84 (m, 2H), 1.62 – 1.53 (m, 3H), 1.48 – 1.36 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.5, 141.4, 138.3, 132.5, 129.1, 128.6, 128.3, 127.2, 118.6, 112.2, 65.0, 64.2, 60.7, 50.2, 29.7, 25.3, 22.5. HRMS (ESI) m/z calcd for C₂₁H₂₃N₂O₂ [M + H]⁺: 335.1754, found: 335.1750. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 23.7 min and 30.1 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate 4-(methoxycarbonyl)benzyl ester (3j)⁵: 32.7 mg, 89% yield, $92\% \ ee, \ [\alpha]_D^{20} = 79.8 \ (c = 0.5, CHCl_3), \text{ colorless oil. }^{1}H \ NMR \ (400 \ MHz, Chloroform-d) \delta 7.95 \ (d, J = 8.0 \ Hz, 2H), 7.36 \ (d, J = 8.0 \ Hz, 2H), 7.22 - 7.19 \ (m, 5H), 5.14 \ (s, 2H), 3.85 \ (s, 3H), 3.72 \ (d, J = 13.2 \ Hz, 1H), 3.37 \ (d, J = 13.6 \ Hz, 1H), 3.20 - 3.17 \ (m, 1H), 2.95 - 2.83 \ (m, 1H), 2.20 - 2.10 \ (m, 1H), 1.80 - 1.76 \ (m, 2H), 1.54 - 1.47 \ (m, 3H), 1.34 - 1.31 \ (m, 1H). \ ^{13}C \ NMR \ (100 \ MHz, Chloroform-d) \delta 173.6, 166.9, 141.2, 138.3, 130.1, 130.0, 129.3, 128.3, 128.0, 127.2, 65.5, 64.1, 60.6, 52.3, 50.2, 29.7, 25.4, 22.5. \ HRMS \ (ESI) \ m/z \ calcd for C_{22}H_{26}NO4 \ [M + H]^+: 368.1856, found: 368.1851. \ HPLC: Chiralcel AD-H \ column, 254 \ nm, 30 \ ^{\circ}C, n-hexane/i-propanol = 95/5, flow = 0.5 \ mL/min, retention \ time 17.7 \ min \ and 19.2 \ min \ (maj).$

(*R*)-1-Benzylpiperidine-2-carboxylate 4-nitrobenzyl ester (3k): 31.9 mg, 90% yield, 92% *ee*, $[\alpha]_D^{20} =$ NO₂
89.1 (c = 0.5, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.21 (d, *J* = 8.8 Hz, 2H), 7.52 (d, *J* = 8.8 Hz, 2H), 7.31 – 7.22 (m, 5H), 5.25 (s, 2H), 3.78 (d, *J* = 13.2 Hz, 1H), 3.44 (d, *J* = 13.6 Hz, 1H), 3.27 (dd, *J* = 7.2, 4.4 Hz,

1H), 2.99 – 2.93 (m, 1H), 2.25 – 2.19 (m, 1H), 1.88 – 1.83 (m, 2H), 1.59 – 1.55 (m, 3H), 1.46 – 1.34 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.5, 147.9, 143.4, 138.3, 129.1, 128.7, 128.3, 127.3, 124.0, 64.7, 64.1, 60.7, 50.2, 29.7, 25.4, 22.5. HRMS (ESI) m/z calcd for C₂₀H₂₃N₂O₄ [M + H]⁺: 355.1652, found: 355.1652. HPLC: Chiralcel AD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 22.7 min and 24.0 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate 4-methylbenzyl ester (31)⁵: 27.5 mg, 85% yield, 93% *ee*, $[\alpha]_D^{20}$ = 71.9 (c = 0.6, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.27 -7.14 (m, 9H), 5.13 (s, 2H), 3.78 (d, *J* = 13.2 Hz, 1H), 3.41 (d, *J* = 13.2 Hz, 1H), 3.20 - 3.17 (m, 1H), 2.97 - 2.91 (m, 1H), 2.34 (s, 3H), 2.19 - 2.14 (m, 1H), 1.85

-1.80 (m, 2H), 1.60 - 1.52 (m, 3H), 1.40 - 1.32 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.9, 138.4, 138.2, 133.2, 129.4, 129.3, 128.6, 128.2, 127.1, 66.2, 64.3, 60.6, 50.2, 29.7, 25.4, 22.6, 21.3. HRMS (ESI) m/z calcd for C₂₁H₂₆NO₂ [M + H]⁺: 324.1958, found: 324.1963. HPLC: Chiralcel OJ-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 16.7 min and 22.1 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate 3-methylbenzyl ester (3m): 28.1 mg, 87% yield, 92% *ee*,
$$[\alpha]_D^{20}$$

= 61.9 (c = 0.5, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.31
- 7.11 (m, 9H), 5.14 (s, 2H), 3.79 (d, *J* = 13.2 Hz, 1H), 3.42 (d, *J* = 13.2 Hz, 1H),
3.21 (dd, *J* = 7.2, 5.2 Hz, 1H), 2.98 – 2.93 (m, 1H), 2.34 (s, 3H), 2.25 – 2.13 (m,

1H), 1.89 - 1.81 (m, 2H), 1.63 - 1.54 (m, 3H), 1.43 - 1.34 (m, 1H). ¹³C NMR (100 MHz, Chloroformd) δ 173.9, 138.4, 136.0, 129.3, 129.2, 129.1, 128.6, 128.3, 128.3, 127.1, 125.5, 66.3, 64.3, 60.6, 50.2, 29.7, 25.4, 22.6, 21.5. HRMS (ESI) m/z calcd for C₂₁H₂₆NO₂ [M + H]⁺: 324.1958, found: 324.1958. HPLC: Chiralcel OJ-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 16.1 min and 17.9 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate 3-methoxybenzyl ester (3n): 30.2 mg, 89% yield, 92% *ee*, $\begin{bmatrix} \alpha \end{bmatrix}_{D}^{20} = 76.4 \text{ (c} = 0.6, \text{ CHCl}_3\text{), colorless oil. }^{1}\text{H NMR} (400 \text{ MHz, Chloroform-} d) \delta 7.30 - 7.23 \text{ (m, 6H)}, 6.97 - 6.84 \text{ (m, 3H)}, 5.15 \text{ (s, 2H)}, 3.82 - 3.80 \text{ (m, 1H)}, 3.79 \text{ (s, 3H)}, 3.43 \text{ (d, } J = 13.2 \text{ Hz, 1H}\text{)}, 3.24 - 3.21 \text{ (m, 1H)}, 2.99 - 2.93 \text{ (m, 2H)}$

1H), 2.24 – 2.16 (m, 1H), 1.89 – 1.82 (m, 2H), 1.60 – 1.53 (m, 3H), 1.42 – 1.34 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.8, 159.9, 138.3, 137.7, 129.8, 129.3, 128.3, 127.2, 120.6, 114.0, 113.8, 66.1, 64.3, 60.6, 55.4, 50.2, 29.7, 25.4, 22.5. HRMS (ESI) m/z calcd for C₂₁H₂₆NO₃ [M + H]⁺: 340.1907, found: 340.1904. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 12.6 min and 13.7 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate 4-bromobenzyl ester (30)⁵: 33.8 mg, 87% yield, 93% *ee*, $[\alpha]_D^{20}$ = 61.3 (c = 0.5, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.54 = 7.39 (m, 2H), 7.29 = 7.20 (m, 7H), 5.11 (s, 2H), 3.77 (d, J = 13.2 Hz, 1H), 3.41 (d, J = 13.6 Hz, 1H), 3.21 (dd, J = 7.2, 4.8 Hz, 1H), 2.97 = 2.92 (m, 1H), 2.21 = 2.16 (m, 1H), 1.85 = 1.81 (m, 2H), 1.57 = 1.53 (m, 3H), 1.39 = 1.34 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.7, 138.3, 135.2, 131.9, 130.2, 129.2, 128.3, 127.2, 122.5, 65.4, 64.2, 60.7, 50.2, 29.7, 25.4, 22.5. HRMS (ESI) m/z calcd for C₂₀H₂₃BrNO₂ [M + H]⁺: 388.0907, found: 388.0909. HPLC: Chiralcel OJ-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 12.1 min and 13.8min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate 3-bromobenzyl ester (3p): 33.0 mg, 85% yield, 92% *ee*, $[\alpha]_D^{20}$ = 67.7 (c = 0.5, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.53 (t, *J* = 1.9 Hz, 1H), 7.47 – 7.43 (m, 1H), 7.31 – 7.19 (m, 7H), 5.13 (s, 2H), 3.78 (d, *J* = 13.6 Hz, 1H), 3.43 (d, *J* = 13.2 Hz, 1H), 3.23 (dd, *J* = 7.2, 4.8 Hz, 1H),

2.98 – 2.93 (m, 1H), 2.28 – 2.14 (m, 1H), 1.86 – 1.81 (m, 2H), 1.63 – 1.54 (m, 3H), 1.44 – 1.35 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.7, 138.4, 138.3, 131.5, 131.3, 130.3, 129.3, 128.3, 127.2, 126.9, 122.7, 65.2, 64.1, 60.7, 50.2, 29.7, 25.4, 22.5. HRMS (ESI) m/z calcd for C₂₀H₂₃BrNO₂ [M + H]⁺: 388.0907, found: 388.0906. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 11.3 min and 12.8 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate 2-bromobenzyl ester (3q): 32.6 mg, 84% yield, 91% *ee*, $[\alpha]_D^{20}$ = 61.0 (c = 0.5, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.43 (dd, *J* = 8.0, 1.2 Hz, 1H), 7.27 (dd, *J* = 7.6, 1.6 Hz, 1H), 7.18 – 7.03 (m, 7H), 5.10 (s, 2H), 3.68 (d, *J* = 13.2 Hz, 1H), 3.32 (d, *J* = 13.6 Hz, 1H), 3.13 (t, *J* = 6.0 Hz, 1H), 2.85 – 2.79 (m, 1H), 2.10 – 2.04 (m, 1H), 1.76 – 1.72 (m, 2H), 1.48 – 1.39 (m, 3H), 1.28 – 1.22 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.6, 138.5, 135.5, 133.1, 130.3, 130.0, 129.3, 128.3, 127.7, 127.1, 123.8, 65.9, 64.0, 60.6, 50.0, 29.7, 25.4, 22.5. HRMS (ESI) m/z calcd for $C_{20}H_{23}BrNO_2$ [M + H]⁺: 388.0907, found: 388.0908. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 11.0 min and 11.7 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate 4-chlorobenzyl ester (3r): 29.6 mg, 86% yield, 88% *ee*, $[\alpha]_D^{20}$ $(\alpha)_{\text{Bn}}^{C_1} = 67.1 (c = 0.6, \text{CHCl}_3)$, colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.36 -7.19 (m, 9H), 5.13 (s, 2H), 3.77 (d, *J* = 13.6 Hz, 1H), 3.41 (d, *J* = 13.6 Hz, 1H), 3.21 (dd, J = 7.2, 4.8 Hz, 1H), 2.97 – 2.92 (m, 1H), 2.21 – 2.15 (m, 1H), 1.85 – 1.78 (m, 2H), 1.57 - 1.52 (m, 3H), 1.41 - 1.34 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.7, 138.3, 134.7, 134.3, 129.9, 129.3, 128.9, 128.3, 127.2, 65.4, 64.3, 60.6, 50.2, 29.7, 25.4, 22.5. HRMS(ESI) m/z calcd for C₂₀H₂₃ClNO₂ [M + H]⁺: 344.1412, found: 344.1415. HPLC: Chiralcel OD-H column,<math>254 nm, $30 \,^{\circ}$ C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 15.2 min and 15.9 min (maj).

(*R*)-1-Benzylpiperidine-2-carboxylate 4-fluorobenzyl ester (3s): 29.8 mg, 91% yield, 92% *ee*, $[\alpha]_D^{20}$ = 90.5 (c = 0.5, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.39 = 7.32 (m, 2H), 7.28 = 7.21 (m, 5H), 7.05 = 7.01 (m, 2H), 5.14 (s, 2H), 3.76 (d, J) = 13.2 Hz, 1H), 3.40 (d, J = 13.2 Hz, 1H), 3.19 (dd, J = 7.6, 4.8 Hz, 1H), 2.97 = 13.2 Hz, 1H) = 1

2.92 (m, 1H), 2.22 – 2.15 (m, 1H), 1.84 – 1.81 (m, 2H), 1.61 – 1.54 (m, 3H), 1.43 – 1.32 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.8, 162.8 (d, *J* = 245.4 Hz), 138.3, 132.0 (d, *J* = 3.1 Hz), 130.5 (d, *J* = 8.2 Hz), 129.3, 128.3, 127.2, 115.6 (d, *J* = 21.4 Hz), 65.5, 64.3, 60.6, 50.2, 29.7, 25.4, 22.6. HRMS (ESI) m/z calcd for C₂₀H₂₃FNO₂ [M + H]⁺: 328.1707, found: 328.1708. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 10.1 min and 11.4 min (maj).

(R)-1-(4-(Trifluoromethyl)benzyl)piperidine-2-carboxylate benzyl ester (3t): 31.7 mg, 84% yield, 90%



ee, $[\alpha]_D^{20} = 99.6$ (c = 1.0, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-_{CO2Bn} *d*) δ 7.53 (d, *J* = 8.0 Hz, 2H), 7.41 (d, *J* = 8.0 Hz, 2H), 7.38 - 7.30 (m, 5H), 5.23 -5.12 (m, 2H), 3.83 (d, *J* = 14.0 Hz, 1H), 3.47 (d, *J* = 14.0 Hz, 1H), 3.26 (t, *J* = 5.6 Hz, 1H), 2.96 - 2.90 (m, 1H), 2.23 - 2.18 (m, 1H), 1.90 - 1.85 (m, 2H), 1.60 - 1.54

(m, 3H), 1.45 - 1.38 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.6, 143.0, 136.0, 130.9 (q, J = 32.2 Hz), 129.2, 128.7, 128.5, 128.5, 127.1 (q, J = 3.8 Hz), 124.4 (q, J = 270.2 Hz), 66.3, 64.2, 60.1, 50.1, 29.6, 25.4, 22.4. HRMS (ESI) m/z calcd for C₂₁H₂₃F₃NO₂ [M + H]⁺: 378.1675, found: 378.1678. HPLC: Chiralcel AD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 9.5 and 10.1 min (maj).

(R)-1-(4-(Chlorobenzyl)benzyl)piperidine-2-carboxylate benzyl ester (3u): 30.6 mg, 89% yield, 91%

NCO2Br

ee, $[\alpha]_D^{20} = 61.2$ (c = 0.65, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform*d*) δ 7.40 – 7.29 (m, 5H), 7.25 – 7.17 (m, 4H), 5.20 – 5.13 (m, 2H), 3.73 (d, *J* = 13.2 Hz, 1H), 3.37 (d, *J* = 13.2 Hz, 1H), 3.21 (t, *J* = 5.6 Hz, 1H), 2.94 – 2.89 (m, 1H), 2.23 – 2.10 (m, 1H), 1.88 – 1.80 (m, 2H), 1.60 – 1.52 (m, 3H), 1.40 – 1.36 (m, 1H).

¹³C NMR (100 MHz, Chloroform-*d*) δ 173.7, 137.1, 136.1, 132.8, 130.5, 128.7, 128.5, 128.5, 128.4, 66.3, 64.2, 59.9, 50.1, 29.7, 25.4, 22.5. HRMS (ESI) m/z calcd for $C_{20}H_{23}CINO_2$ [M + H]⁺: 344.1412, found: 344.1407. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 10.8 and 11.7 min (maj).

(R)-1-(4-(Methoxycarbonyl)benzyl)piperidine-2-carboxylate benzyl ester (3v): 33.8 mg, 92% yield,



92% *ee*, $[\alpha]_D^{20} = 70.3$ (c = 0.65, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.95 (d, *J* = 8.0 Hz, 2H), 7.43 – 7.28 (m, 7H), 5.25 – 5.09 (m, 2H), 3.90 (s, 3H), 3.82 (d, *J* = 14.0 Hz, 1H), 3.46 (d, *J* = 13.6 Hz, 1H), 3.23 (t, *J* = 6.0 Hz, 1H), 2.97 – 2.86 (m, 1H), 2.25 – 2.12 (m, 1H), 1.88 – 1.84 (m, 2H),

1.62 - 1.53 (m, 3H), 1.43 - 1.35 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.6, 167.2, 144.2, 136.1, 129.6, 129.0, 129.0, 128.7, 128.5, 128.4, 66.3, 64.2, 60.3, 52.1, 50.2, 29.6, 25.4, 22.4. HRMS (ESI) m/z calcd for C₂₂H₂₆NO₄ [M + H]⁺: 368.1856, found: 368.1854 HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 15.4 and 16.1 min (maj).

(R)-1-(2,4-Difluorobenzyl)piperidine-2-carboxylate benzyl ester (3w): 30.7 mg, 89% yield, 98% ee,



 $[\alpha]_D^{20} = 59.1 \text{ (c} = 0.5, \text{ CHCl}_3\text{), colorless oil. }^{1}\text{H NMR (400 MHz, Chloroform-}d\text{)} \delta$ 7.43 - 7.25 (m, 6H), 6.86 - 6.69 (m, 2H), 5.27 - 5.12 (m, 2H), 3.70 (d, J = 13.6 Hz, 1H), 3.57 (d, J = 14.0 Hz, 1H), 3.22 (dd, J = 7.6, 4.4 Hz, 1H), 2.98 - 2.93 (m, 1H), 2.24 - 2.18 (m, 1H), 1.87 - 1.80 (m, 2H), 1.62 - 1.53 (m, 3H), 1.41 - 1.31 (m, 1H).

¹³C NMR (100 MHz, Chloroform-*d*) δ 173.7, 163.0 (dd, J = 78.4, 11.8 Hz), 160.6 (dd, J = 79.6, 12.0 Hz), 136.1, 132.4 (dd, J = 9.0, 6.3 Hz), 128.7, 128.5, 128.4, 121.0 (dd, J = 16.3 Hz), 111.1 (dd, J = 20.8, 3.7 Hz), 103.5 (dd, J = 26.0, 25.5Hz), 66.4, 64.2, 52.5, 50.2, 29.8, 25.4, 22.5. HRMS (ESI) m/z calcd for C₂₀H₂₂F₂NO₂ [M + H]⁺: 346.1613, found: 346.1609. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 95/5, flow = 0.5 mL/min, retention time 27.6 min and 29.0 min (maj).

(*R*)-1-benzyl-2-phenylpiperidine (3x): 14.4 mg, 76% yield, 33% *ee*, $[\alpha]_D^{20} = 13.4$ (c = 0.5, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.36 – 7.15 (m, 5H), 3.99 (d, *J* = 13.6 Hz, 1H), 3.18 (d, *J* = 13.6 Hz, 1H), 2.75 – 2.69 (m, 1H), 2.31 – 2.27 (m, 1H), 1.97 – 1.91 (m, 1H), 1.65 – 1.61 (m, 2H), 1.56 – 1.21 (m, 4H), 1.15 (d, *J* = 6.0 Hz, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 139.7, 129.1, 128.1, 126.7, 58.6, 56.5, 52.3, 34.9, 26.2, 24.1, 19.6. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 90/10, flow = 0.5 mL/min, retention time 10.1 min and 11.5 min (maj).

(*R*)-1-benzyl-2-phenylpiperidine (3y): 20.6 mg, 82% yield, 60% *ee*, $[\alpha]_D^{20} = 24.9$ (c = 1.0, CHCl₃), colorless oil. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.45 (d, *J* = 7.6 Hz, 2H), 7.32 (t, *J* = 7.6 Hz, 2H), 7.28 – 7.15 (m, 6H), 3.76 (d, *J* = 13.6 Hz, 1H), 3.10 (dd, *J* = 11.2, 2.8 Hz, 1H), 2.99 – 2.94 (m, 1H), 2.80 (d, *J* = 13.6 Hz, 1H), 1.96 – 1.90 (m, 1H), 1.80 – 1.74 (m, 2H), 1.66 – 1.55 (m, 3H), 1.42 – 1.33 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 145.9, 140.0, 128.8, 128.6, 128.1, 127.6, 127.0, 126.6, 69.4, 59.9, 53.5, 37.2, 26.2, 25.4. HRMS (ESI) m/z calcd for C₁₈H₂₂N [M + H]⁺: 252.1747, found: 252.1743. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 90/10, flow = 0.5 mL/min, retention time 13.6 min and 15.1 min (maj).

2.4 General procedure for asymmetric hydrogenation under continuous flow

All process parts, including fittings, tubes, valves and junctions that hold pressure were purchased from SHENZHEN INSFTECH CO,. Ltd. The specification of the reaction coil is 0.5 ml/m. The information of other main components is summarized in Table S1.

Table S1 Components details of reactor system

Name	Information
Pump	Sanotac high pressure HPLC pump AP0030 (0-10 mL/min; 20 MPa)
MFC	SHENZHEN INSFTECH CO,. Ltd. FCM-1050 (0-500sccm,10MPa)
BPR	SHENZHEN INSFTECH CO,. Ltd. FAV-1500B (0-500mL/min, 10MPa)
Mixer	SHENZHEN INSFTECH CO,, Ltd. MGL-2000 (200*250µm, 2000Psi)

A mixture of $[Ir(COD)Cl]_2$ (2.0 mol%) and (*R*,*R*)-BDPP (4.0 mol%) was dissolved in a degassed solvent EA/MeOH at argon atmosphere, and the resulting solution was allowed to be stirred at room temperature for 30.0 min. Then, *N*-benzyl-2-methylpyridinium salt **2a** (1.0 equiv.) and KI (1.0 equiv.) were added. The process was washed by EA/MeOH at a liquid flow rate of 5 mL/min and gas flow rate of 10.0 sccm (avoid back flow of liquid to gas flow meter) for 10.0 minutes and then pressurized the BPR. After the reactor was pressurized to 8.0 MPa, the aforehand reaction medium was pumped instead of solvent. Liquid flow rate was set at 0.2 mL/min and gas flow rate was keeping 20.0 sccm. The liquid holding capacity of the reaction coil can be adjusted according to the needs. When reaction finished, system was depressurized by releasing the gas slowly, and washed the whole system by pumping ethanol for 10.0 minutes. The reaction mixture was filtrated and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 150:1) to give the desired product **3a** (94% yield, 92% *ee*) as a pale-yellow oil.



Figure S1 AH of 2a under continuous flow.



Figure S2 Set-up for asymmetric hydrogenation under continuous flow.

2.5 Procedures for Products Transformations



A solution of the methyl ester **3a** (100.0 mg, 0.43 mmol, 1.0 equiv.) in THF (2.5 mL) was added dropwise to a stirred suspension of LiAlH₄ (24.4 mg, 0.64 mmol, 1.5 equiv.) in THF (2.5 mL) at 0 °C under an argon atmosphere. The resulting mixture was stirred at 25 °C for 8 h and then 2 M NaOH(aq) (1 µL per 1 mg of LiAlH₄), Et₂O (5 mL) and Na₂SO₄ were carefully added. The solids were removed by filtration through Celite and evaporated under reduced pressure to give the crude product. The residue was purified by silica gel flash column chromatography (DCM/MeOH = 100:1) to give compound **4** (84.5 mg, 96%, 97% *ee*) as a pale-yellow oil. $[\alpha]_D^{20} = 34.2$ (c = 0.5, CHCl₃). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.38 – 7.19 (m, 5H), 4.06 (d, *J* = 13.2 Hz, 1H), 3.86 (dd, *J* = 10.8, 4.4 Hz, 1H), 3.52 (dd, *J* = 10.4, 4.0 Hz, 1H), 3.32 (d, *J* = 13.2 Hz, 1H), 2.89 – 2.84 (m, 1H), 2.53 (s, 1H), 2.47 – 2.37 (m, 1H), 2.17 – 2.12 (m, 1H), 1.79 – 1.49 (m, 4H), 1.39 – 1.31 (m, 2H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 139.1, 129.0, 128.5, 127.1, 62.4, 61.0, 57.8, 50.9, 27.4, 24.2, 23.5. HRMS (ESI) m/z calcd for C₁₃H₂₀NO [M + H]⁺: 206.1539, found: 206.1537. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 90/10, flow = 0.5 mL/min, retention time 12.1 min and 12.8 min (maj).



To a solution of the methyl ester **3a** (100.0 mg, 0.43 mmol, 1.0 equiv.) in anhydrous THF (5.0 mL) was added methyl magnesium bromide (1.0 M in THF, 1.3 mL, 1.29 mmol, 3.0 equiv.) dropwise under argon atmosphere at 0 °C. The resulting mixture was stirred at room temperature for 4 h and then quenched with aqueous NH₄Cl solution, extracted with ethyl acetate. Organic phases were combined and dried over anhydrous Na₂SO₄, filtered and the solvent was evaporated under reduced pressure. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 50:1) to give compound **5** (91.0 mg, 91% yield, 93% *ee*) as a pale-yellow oil. $[\alpha]_D^{20} = -5.41$ (c = 0.4, CHCl₃). ¹H NMR (400 MHz, Chloroform-d) δ 7.39 – 7.21 (m, 5H), 3.96 – 3.84 (m, 2H), 2.87 – 2.80 (m, 1H), 2.59 – 2.53 (m, 2H), 1.87 – 1.59 (m, 4H), 1.59 – 1.49 (m, 2H), 1.43 – 1.36 (m, 1H), 1.26 (s, 3H), 1.21 (s, 3H). ¹³C NMR (100 MHz, Chloroform-d) δ 140.1, 128.8, 128.5, 127.2, 72.4, 68.5, 58.6, 46.0, 29.5, 26.7, 21.5, 21.1, 18.7. HRMS (ESI) m/z calcd for C₁₅H₂₄NO [M + H]⁺: 234.1852, found: 234.1847. HPLC: Chiralcel OJ-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 90/10, flow = 0.5 mL/min, retention time 11.9 min (maj) and 13.6 min.



An oven-dried vial equipped with a stir bar was charged with the methyl ester **3a** (100.0 mg, 0.43 mmol, 1.0 equiv.), aniline (46.9 µL, 0.51 mmol, 1.2 equiv.) placed under a positive pressure of argon, and subjected to three evacuation/backfilling cycles. Toluene (5.0 mL) and LiHMDS (1.0 M in THF, 0.9 mL, 0.86 mmol, 2.0 equiv.) were sequentially added with vigorous stirring at 25 °C, and the reaction mixture was stirred at room temperature overnight. The reaction mixture was quenched with aqueous NH₄Cl solution, diluted with ethyl acetate (5.0 mL), the organic layer was washed with water, brine, dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 20:1) to give compound **6** (112.3 mg, 89% yield, 96% *ee*) as a white solid, m.p. = 113.0 – 115.1 °C. $[\alpha]_D^{20} = 43.1$ (c = 1.0, CHCl₃). ¹H NMR

(400 MHz, Chloroform-d) δ 8.84 (s, 1H), 7.56 – 7.54 (m, 2H), 7.36 – 7.24 (m, 7H), 7.10 – 7.06 (m, 1H), 3.93 (d, J = 13.6 Hz, 1H), 3.28 (d, J = 13.6 Hz, 1H), 3.00 – 2.95 (m, 2H), 2.11 – 2.06 (m, 2H), 1.77 – 1.62 (m, 3H), 1.51 – 1.26 (m, 2H). ¹³C NMR (100 MHz, Chloroform-d) δ 172.7, 138.0, 137.9, 129.1, 128.63, 128.59, 127.4, 124.1, 119.4, 68.1, 61.0, 51.5, 29.9, 24.6, 23.4. HRMS (ESI) m/z calcd for C₁₉H₂₃N₂O [M + H]⁺: 295.1805, found: 295.1807. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 90/10, flow = 0.5 mL/min, retention time 15.7 min (maj) and 19.0 min.



To a stirred solution of the methyl ester **3a** (100.0 mg, 0.43 mmol, 1.0 equiv.) in MeOH (5.0 mL) was added 10% Pd/C (10.0 mg, 10.0 wt%). The resulting mixture was stirred overnight at 25 °C under H₂ (1 atm, balloon) and then filtered, washed with water and concentrated under reduced pressure.

The crude was dissolved in acetic acid (5.0 mL), potassium cyanate (69.8 mg, 0.86 mmol, 1.0 equiv.) was added and the reaction mixture was allowed to stir at 25 °C for 12 h. The reaction mixture was quenched with aqueous NaHCO₃ solution, extracted with ethyl acetate, the combined organic phase was washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 10:1) to give compound 7 (57.5 mg, 87% over 2 steps, 98% *ee*) as a white solid, m.p. = 131.3 – 133.3 °C. $[\alpha]_D^{20}$ = 77.1 (c = 1.0, CHCl₃). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.62 (s, 1H), 4.13 (dd, *J* = 13.6, 4.4 Hz, 1H), 3.82 – 3.78 (m, 1H), 2.83 – 2.76 (m, 1H), 2.21 – 2.17 (m, 1H), 2.01 – 1.98 (m, 1H), 1.75 – 1.72 (m, 1H), 1.54 – 1.31 (m, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.8, 154.4, 58.8, 39.3, 27.7, 25.0, 22.8. HRMS (ESI) m/z calcd for C₇H₁₁N₂O₂ [M + H]⁺: 155.0815, found: 155.0817. HPLC: Chiralcel IC column, 210 nm, 30 °C, *n*-hexane/*i*-propanol = 90/10, flow = 0.5 mL/min, retention time 29.2 min (maj) and 33.7 min.



To a stirred solution of the methyl ester **3a** (100.0 mg, 0.43 mmol, 1.0 equiv.) in MeOH (5.0 mL) was added 10% Pd/C (10.0 mg, 10.0 wt%). The resulting mixture was stirred overnight at 25 °C under H₂ (1 atm, balloon) and then filtered, washed with water and concentrated under reduced pressure. The crude was dissolved in acetonitrile (6.0 mL), (*R*)-propylene oxide (39.0 µL, 0.56 mmol, 1.3 equiv.) was added. Next magnesium trifluoromethanesulfonate (37.8 mg, 0.21 mmol, 0.5 equiv.) was added and the reaction was stirred and heated at 75 °C for 17 h. After this time, the reaction was cooled to room temperature and sodium carbonate solution (2 M) was added. The mixture was extracted with ethyl acetate, the combined organic phase was washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 50:1) to give compound **8** (65.3 mg, 90% yield, 97% *ee*) as a pale-yellow oil. $[\alpha]_D^{20} = 45.5$ (c = 0.5, CHCl₃). ¹H NMR (400 MHz, Chloroform-*d*) δ 4.58 – 4.51 (m, 1H), 2.88 – 2.79 (m, 1H), 2.70 – 2.57 (m, 3H), 2.24 – 2.19 (m, 1H), 2.15 – 2.08 (m, 1H), 1.92 – 1.81 (m, 1H), 1.65 – 1.49 (m, 3H), 1.46 (d, *J* = 6.8 Hz, 3H), 1.37 – 1.21 (m, 1H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 170.3, 74.4, 64.6, 55.73, 55.68, 27.6, 24.8, 24.6, 20.8. HRMS (ESI) m/z calcd for C₉H₁₆NO₂ [M + H]⁺: 170.1176, found:

170.1173. HPLC: Chiralcel IC column, 210 nm, 30 °C, *n*-hexane/*i*-propanol = 90/10, flow = 0.5 mL/min, retention time 44.3 min (maj) and 48.7 min.



To a stirred solution of the methyl ester **3a** (100.0 mg, 0.43 mmol, 1.0 equiv.) in MeOH (5.0 mL) were added 10% Pd/C (10.0 mg, 10.0 wt%). The resulting mixture was stirred overnight at 25 °C under H₂ (1 atm, balloon) and then filtered, washed with MeOH and concentrated under reduced pressure. The crude was dissolved in DMSO, 1-Fluoro-2-nitrobenzene (22.6 µL, 0.21 mmol, 0.5 equiv.) was added. The mixture was heated to 110 °C for 24 hours. After cooled to room temperature. The mixture was extracted with ethyl acetate, the combined organic phase was washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The resulting residue was next dissolved in methanol (5.0 mL), zinc powder (420.4 mg, 6.43 mmol, 15.0 equiv.) and ammonium chloride (343.9 mg, 6.43 mmol, 15.0 equiv.) was added to this solution, and the mixture was stirred at 25 °C overnight. The mixture solution was then filtered, concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 20:1) to give compound 9 (68.5 mg, 79% yield over 3 steps, 92% ee) as a white solid, m.p. = 181.2 - 183.1 °C. $[\alpha]_D^{20} = 4.8$ (c = 0.8, CHCl₃). ¹H NMR (400 MHz, Chloroform-d) δ 9.36 (s, 1H), 7.03 – 6.94 (m, 1H), 6.80 – 6.77 (m, 3H), 3.78 (d, J = 12.0 Hz, 1H), 3.57 (dd, J = 12.0, 3.2 Hz, 1H), 2.77 - 2.70 (m, 1H), 2.24 - 2.20 (m, 1H), 1.99 - 1.93 (m, 1H), 1.79 – 1.57 (m, 3H), 1.55 – 1.44 (m, 1H). ¹³C NMR (100 MHz, Chloroform-d) δ 169.8, 135.7, 126.5, 124.2, 119.5, 115.6, 112.2, 59.9, 46.7, 26.9, 23.9, 23.4. HRMS (ESI) m/z calcd for $C_{12}H_{15}N_2O$ [M + H]⁺: 203.1179, found: 203.1182. HPLC: Chiralcel IC column, 254 nm, 30 °C, n-hexane/i-propanol = 90/10, flow = 0.5 mL/min, retention time 36.7 min and 40.3 min (maj).

2.6 The synthesis of ropivacaine and levobupivacaine under batch and flow

Synthesis of (-)-3a

Batch Reaction



A mixture of $[Ir(COD)CI]_2$ (2.2 mg, 2.0 µmol, 0.1 mol%) and (*S*,*S*)-BDPP (*ent*-L6) (2.6 mg, 4.0 µmol, 0.2 mol%) were dissolved in degassed MeOAc (30.0 mL) at argon atmosphere, and the resulting solution was allowed to stirred for 20 min, followed by the addition of the substrate **2a** (1.0 g, 3.25 mmol, 1.0 equiv.) and KI (539.5 mg, 3.25 mmol, 1.0 equiv.). The resulting mixture was transferred to an autoclave, which was purged (3 × 5 atm) and charged with H₂ (600 psi), then the reaction mixtures were stirred at -20 °C for 72 h. After careful release of the hydrogen gas, the reaction mixture was filtrated and concentrated in vacuo. Flash chromatography on silica gel using petroleum ether/ethyl acetate as the eluent gave the products. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 100:1) to give compound (–)-**3a** (0.72 g, 95% yield, 95% *ee*) as a pale-yellow oil.

 $[\alpha]_D^{20} = -75.1$ (c = 1.0, CHCl₃). HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 90/10, flow = 0.5 mL/min, retention time 8.8 min (maj) and 11.5 min.



Continuous Flow Reaction

A mixture of $[Ir(COD)Cl]_2$ (2.0 mol%) and (*R*,*R*)-BDPP (4.0 mol%) was dissolved in a degassed solvent EtOAc/MeOH at argon atmosphere, and the resulting solution was allowed to be stirred at room temperature for 30 min. Then, *N*-benzyl-2-methylpyridinium salt **2a** (1.0 equiv.) was added. The process was washed by EA/MeOH at a liquid flow rate of 5 mL/min and gas flow rate of 10.0 sccm (avoid back flow of liquid to gas flow meter) for 10.0 minutes and then pressurized the BPR. After the reactor was pressurized to 8 MPa, the aforehand reaction medium was pumped instead of solvent. Liquid flow rate was set at 0.2 mL/min and gas flow rate was keeping 20 sccm. The liquid holding capacity of the reaction coil can be adjusted according to the needs. When reaction finished, system was depressurized by releasing the gas slowly, and washed the whole system by pumping ethanol for 10.0 minutes. the reaction mixture was filtrated and concentrated in vacuo. Flash chromatography on silica gel using petroleum ether/ethyl acetate as the eluent gave the products. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 100:1) to give compound (–)-**3a** (94% yield, 91% *ee*) as a pale-yellow oil. HPLC: Chiralcel OD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 90/10, flow = 0.5 mL/min, retention time 8.8 min (maj) and 11.5 min.

Synthesis of 14

Batch Reaction



To a stirred solution of the methyl ester (-)-3a (200.0 mg, 5.8 mmol, 1.0 equiv.) in MeOH (25.0 mL) were added 10% Pd/C (180.0 mg, 10.0 wt%). The resulting mixture was stirred overnight at 25 °C under H₂ (1 atm, balloon) and then filtered, washed with water and concentrated under reduced pressure to afford compound 13 (114.2 mg, 93% yield) as a yellow oil, which was directly used in the next step without further purification.

Continuous Flow Reaction



A solution of (–)-**3a** in MeOH (0.8 M) was pumped at a flow rate of 1.8 mL/min, which combined with hydrogen stream at 100 sccm in a T-shape mixer. The gas-liquid mixed solution was passed through a fixed bed reactor containing 10% Pd/C (2.0 g) and SiO₂ (18.0 g) (9.0 mL internal volume) at 25 °C with a 5.0 min residence time. The outlet of the fixed bed reactor was connected to a back-pressure regulator to control a stable system pressure at 1.0 MPa. The output of the reaction mixture was concentrated in vacuo to afford crude secondary amine **14** (97% yield) without further purification.

Synthesis of 16

Batch Reaction



An oven-dried vial equipped with a stir bar was charged with the methyl ester **14** (100.0 mg, 0.70 mmol, 1.0 equiv.), aniline (103.2 μ L, 0.84 mmol, 1.2 equiv.) placed under a positive pressure of argon, and subjected to three evacuation/backfilling cycles. Toluene (5.0 mL) and LiHMDS (1.0 M in THF, 1.4 mL, 1.40 mmol, 2.0 equiv.) were sequentially added with vigorous stirring at 25 °C, and the reaction mixture was stirred overnight. The reaction mixture was quenched with aqueous NH₄Cl solution, diluted with ethyl acetate (5.0 mL), the organic layer was washed with water, brine, dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography (DCM/MeOH = 100:1) to give compound **16** (144.4 mg, 89% yield, 92% *ee*). HPLC: Chiralcel OJ-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 90/10, flow = 0.5 mL/min, retention time 12.6 min and 14.6 min (maj).

Continuous Flow Reaction



The self-build flow reactor equipment consists of two Fusion 4000 type syringe pumps for reagent/solvent delivery and a 1.5 mL PTFE coil reactor (0.8 mm i.d.). Before the start of the actual experiment, the reactor was primed with toluene. The syringe pump A was used to introduce the solution of compound 14 (0.5 M, 1.0 equiv.) and 15 (1.2 equiv.) in toluene (0.5 mL/min), the syringe pump B was used to introduce the solution of LiHMDS (1.0 M in THF, 2.0 equiv., 0.5 mL /min). The two solutions were mixed through a T-shape mixer and pumped through the coil reactor (1.5 mL, internal volume, t_R = 1.5 min) at 25 °C. A 75 psi BPR was connected at the outlet of coil reactor. The system was allowed to come to steady state by waiting three residence times prior to collecting product. The reaction mixture was collected, quenched by addition of water. The mixture was extracted with ethyl acetate, the combined organic phase was washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 10:1) to give compound 16 (95% yield, 91% ee) as a white solid, m.p. = 121.1 - 1000123.2 °C. $[\alpha]_D^{20} = 38.6$ (c = 2.0, HCl 1.0 M) [Lit.⁶ $[\alpha]_D^{22} = 33.0$ (c = 2.0, HCl 1.0 M)]. ¹H NMR (400 MHz, Chloroform-d) & 8.23 (s, 1H), 7.13 – 6.95 (m, 3H), 3.41 (dd, J = 10.0, 3.2 Hz, 1H), 3.14 – 3.09 (m, 1H), 2.85 – 2.70 (m, 1H), 2.22 (s, 6H), 2.10 – 2.05 (m, 1H), 1.86 – 1.82 (m, 2H), 1.65 – 1.58 (m, 2H), 1.57 – 1.41 (m, 2H). ¹³C NMR (100 MHz, Chloroform-d) δ 172.5, 135.2, 133.8, 128.3, 127.1, 60.8, 46.0, 30.5, 26.1, 24.2, 18.6. HRMS (ESI) m/z calcd for $C_{14}H_{21}N_{20}$ [M + H]⁺: 233.1648, found: 233.1644. HPLC: Chiralcel OJ-H column, 254 nm, 30 °C, n-hexane/i-propanol = 90/10, flow = 0.5 mL/min, retention time 11.9 min and 13.9 min (maj).

Synthesis of 17

Batch Reaction



To a stirred solution of the amide **16** (50.0 mg, 0.22 mmol, 1.0 equiv.) in isopropyl alcohol (2.0 mL) were added potassium carbonate (89.3 mg, 0.65 mmol, 3.0 equiv.) and alkyl halide (0.65 mmol, 3.0 equiv.). Water (0.5 mL) was added and the mixture was stirred at 100 °C overnight, the solvent was evaporated and water was added. The mixture was extracted with ethyl acetate, the combined organic phase was washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 20:1) to

give desired compound **17a**: 56.7 mg, 96% yield, 93% *ee*. HPLC: Chiralcel AD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 90/10, flow = 0.5 mL/min, retention time 13.8 min and 14.6 min (maj). **17b**: 60.2 mg, 97% yield, 93% *ee*. HPLC: Chiralcel AD-H column, 254 nm, 30 °C, *n*-hexane/*i*-propanol = 90/10, flow = 0.5 mL/min, retention time 19.7 min and 22.4 min (maj).

Continuous Flow Reaction



The self-build flow reactor equipment consists of two Fusion 4000 type syringe pumps for reagent/solvent delivery and a 6.0 mL PTFE coil reactor (0.8 mm i.d.). Before the start of the actual experiment, the reactor was primed with iPrOH/H2O. The syringe pump A was used to introduce the solution of compound 16 (0.05 M, 1.0 equiv.) and RBr (3.0 equiv.) in iPrOH (50.0 µL/min), the syringe pump B was used to introduce the solution of K₂CO₃ (0.15 M in *i*PrOH/H₂O, 3.0 equiv., 50.0 μL/min). The two solutions were mixed through a T-shape mixer and pumped through the coil reactor (6.0 mL, internal volume, $t_{\rm R} = 60.0$ min) at 80 °C. A 75 psi BPR was connected at the outlet of coil reactor. The system was allowed to come to steady state by waiting three residence times prior to collecting product. The reaction mixture was collected, the solvent was evaporated and water was added. The mixture was extracted with ethyl acetate, the combined organic phase was washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 20:1) to give desired compound 17a: 97% yield, 92% ee, white solid, m.p. = 138.3 - 140.1 °C. $[\alpha]_D^{25} = -72.6$ (c = 2.0, MeOH) [Lit.⁶ $[\alpha]_D^{25} = -80.0$ (c = 2.0, MeOH), m.p. = 145.0 – 147.0 °C]. ¹H NMR (400 MHz, Chloroform-d) δ 8.16 (s, 1H), 7.10 – 7.04 (m, 3H), 3.22 – 3.17 (m, 1H), 2.87 (dd, J = 10.4, 3.6 Hz, 1H), 2.83 – 2.72 (m, 1H), 2.24 (s, 6H), 2.21 – 1.98 (m, 3H), 1.81 - 1.63 (m, 4H), 1.59 - 1.44 (m, 2H), 1.41 - 1.27 (m, 1H), 0.91 (t, J = 7.6 Hz, 3 H).¹³C NMR (100 MHz, Chloroform-d) δ 173.1, 135.4, 133.8, 128.4, 127.1, 68.7, 59.5, 51.7, 30.8, 25.0, 23.6, 20.8, 18.8, 11.7. HRMS (ESI) m/z calcd for $C_{17}H_{27}N_2O$ [M + H]⁺: 275.2118, found: 275.2117. HPLC: Chiralcel AD-H column, 254 nm, 30 °C, n-hexane/i-propanol = 90/10, flow = 0.5 mL/min, retention time 14.1 min and 14.9 min (maj). 17b: 97% yield, 91% ee, white solid, m.p. = 133.1 - 135.3 °C. $[\alpha]_D^{20} = -89.5$ (c = 1.0, MeOH) [Lit.⁷ $[\alpha]_D^{20} = -77.0$ (c = 1.0, MeOH), m.p. = 135.0 - 137.0 °C]. ¹H NMR (400 MHz, Chloroform-d) δ 8.16 (s, 1H), 7.10 – 7.05 (m, 3H), 3.23 – 3.18 (m, 1H), 2.94 – 2.76 (m, 2H), 2.30 - 2.26 (m, 1H), 2.25 (s, 6H), 2.16 - 2.00 (m, 2H), 1.83 - 1.42 (m, 6H), 1.41 - 1.22 (m, 3H), 0.92 (t, J = 7.2 Hz, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.1, 135.4, 133.9, 128.4, 127.2, 68.7, 57.7, 51.8, 30.8, 29.9, 25.0, 23.7, 20.8, 18.9, 14.3. HRMS (ESI) m/z calcd for C₁₈H₂₉N₂O [M + H]⁺: 289.2274, found: 289.2277. HPLC: Chiralcel AD-H column, 254 nm, 30 °C, n-hexane/i-propanol = 90/10, flow = 0.5 mL/min, retention time 20.4 min and 23.1 min (maj).

3. References

- X. Frogneux, N. von Wolff, P. Thuéry, G. Lefèvre and T. Cantat, *Chem. Eur. J.*, 2016, 22, 2930-2934.
- N. S. Mahajani, R. I. L. Meador, T. J. Smith, S. E. Canarelli, A. A. Adhikari, J. P. Shah, C. M. Russo, D. R. Wallach, K. T. Howard, A. M. Millimaci and J. D. Chisholm, *J. Org. Chem.*, 2019, 84 (12), 7871-7882.
- A. Jordan, K. D. Whymark, J. Sydenham and H. F. Sneddon, *Green Chem.*, 2021, 23 (17), 6405-6413.
- 4. Y.-S. Bao, C.-Y. Chen and Z.-Z. Huang, J. Org. Chem., 2012, 77 (18), 8344-8349.
- 5. R. R. Anugu, S. Munnuri and J. R. Falck, J. Am. Chem. Soc., 2020, 142 (11), 5266-5271.
- 6. T. K. Beng and R. E. Gawley, J. Am. Chem. Soc., 2010, 132 (35), 12216-12217.
- 7. N. Shankaraiah, R. A. Pilli and L. S. Santos, *Tetrahedron Lett.*, 2008, 49 (34), 5098-5100.

4. NMR Spectra



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1b



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 1b



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1c



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 1c



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1d



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 1d



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1e



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 1e



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1f



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 1f



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1g



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 1g



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1h



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 1h



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 2i



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 2i



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1j





¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1k



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 1k



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 11



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 11



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1m



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 1m



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1n



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 1n



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 10



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 10



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1p



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 1p


¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 1q



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 1q



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1r



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 1r



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 1s



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 1s



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3a



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 3a



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3b



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 3b



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3c



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 3c



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3d



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 3d



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3e



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 3e



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3f



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 3f



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3g



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 3g



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 3h



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 3h



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 3i



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 3i



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3j



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 3j



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 3k



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 3k



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 31



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 31



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3m



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 3m



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 3n



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 3n



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 30



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 30



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 3p



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 3p



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3q



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 3q



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3r



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 3r



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 3s



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 3s



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 3t



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 3t



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 3u



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 3u



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3v



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 3v



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3w



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 3w



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 3x



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 3x



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 3y



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 3y



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 4



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 4



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 5



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 5



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 6



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 6



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 7



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 7



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 8



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 8



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 9



¹³C-NMR Spectrum (100 MHz, Chloroform-*d*) of Compound 9



¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 16



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 16



¹H-NMR Spectrum (400 MHz, Chloroform-d) of Compound 17a



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 17a


¹H-NMR Spectrum (400 MHz, Chloroform-*d*) of Compound 17b



¹³C-NMR Spectrum (100 MHz, Chloroform-d) of Compound 17b

5. HPLC Spectra





Ret Time [min]	Туре	Width [min]	Area [mAU * s]	Height [mAU]	Area%
8.774	MM m	0.18	148.54	13.19	2.44
11.385	MM m	0.25	5938.85	361.54	97.56



Ret Time [min]	Туре	Width [min]	Area [mAU * s]	Height [mAU]	Area%
8.755	MM m	0.17	252.11	22.46	4.15
11.364	MM m	0.24	5815.70	355.56	95.85























Ret Time [min]	Туре	Width [min]	Area [mAU * s]	Height [mAU]	Area%
18.875	MM m	0.48	2574.91	80.44	49.53
21.402	MM m	0.49	2624.25	81.63	50.47









Ret Time [min]	Type	Width [min]	Area [mAU * s]	Height [mAU]	Area%
23.720	MM m	0.43	496.25	17.40	49.46
30.254	MM m	0.49	507.16	13.75	50.54









Ret Time [min]	Туре	Width [min]	Area [mAU * s]	Height [mAU]	Area%
22.794	MM m	0.51	10816.27	324.47	49.84
24.160	MM m	0.36	10886.89	458.81	50.16





























Ret Time [min]	Туре	Width [min]	Area [mAU * s]	Height [mAU]	Area%
15.113	MM m	0.32	8925.84	433.81	49.69
15.904	MM m	0.35	9036.70	397.26	50.31

















Ket Thile [hilli]	Type	width [mm]	Alca [IIIAO s]	fieight [hix0]	Alca/0	
15.447	MM m	0.28	491.91	28.19	4.00	
16.131	MM m	0.37	11797.57	491.05	96.00	
						7









































Ret Time [min]	Туре	Width [min]	Area [mAU * s]	Height [mAU]	Area%
8.841	MM m	0.19	4432.49	347.17	49.94
11.529	MM m	0.24	4443.38	276.29	50.06



102.37

8.37

2.53

11.495

MM m





9.5 10 10.5 11 11.5 12 12.5 13 13.5 14 14.5 15 15.5 16 16.5 17 17.5 18 18.5 19 19.5 20 Time [min]

Ret Time [min]	Туре	Width [min]	Area [mAU * s]	Height [mAU]	Area%
12.056	MM m	0.40	3249.94	118.71	50.11
14.170	MM m	0.70	3235.15	68.85	49.89



Ret Time [min]	Туре	Width [min]	Area [mAU * s]	Height [mAU]	Area%
12.626	MM m	0.34	66.52	2.42	3.82
14.586	MM m	0.56	1674.33	42.54	96.18





Ret Time [min]	Type	Width [min]	Area [mAU * s]	Height [mAU]	Area%
13.888	MM m	0.26	27128.86	1647.57	49.78
14.709	MM m	0.27	27374.08	1575.79	50.22







15 15 5 16 16 5 17 17 5 18 18 5 19 19 5 20 20 5 21 21 5 22 22 5 23 23 5 24 24 5 25 25 5 26 26 5 27 27 5 28 28 5 29 29 5 30 Time [min]

Ret Time [min]	Туре	Width [min]	Area [mAU * s]	Height [mAU]	Area%
19.972	MM m	0.48	1303.95	41.42	50.01
22.783	MM m	0.50	1303.39	39.27	49.99



Timo.	min
1 11 12	

Ret Time [min]	Туре	Width [min]	Area [mAU * s]	Height [mAU]	Area%
19.664	MM m	0.33	35.27	1.28	3.38
22.351	MM m	0.51	1007.21	29.67	96.62



Ret Time [min]	Туре	Width [min]	Area [mAU * s]	Height [mAU]	Area%
20.374	MM m	0.34	35.03	1.24	4.35
23.062	MM m	0.45	770.97	23.55	95.65
6. Crystallographic Data



Figure 1. ORTEP of the molecular structure of 7.

Diffraction-quality crystal of compound 7 was obtained in ethyl acetate and petroleum ether.

CCDC 2221385 contains the supplementary crystallographic data for compound 7.

Empirical formula	C7H10N2O2
Formula weight	154.17
Temperature/K	302.0
Crystal system	monoclinic
Space group	P21
a/Å	5.3586(2)
b/Å	6.1328(3)
c/Å	11.1471(5)
α/°	90
β/°	98.5290(10)
$\gamma/^{\circ}$	90
Volume/Å ³	362.28(3)
Ζ	2
$\rho_{calc}g/cm^3$	1.413
µ/mm ⁻¹	0.878
F(000)	164.0
Crystal size/mm ³	0.5 imes 0.26 imes 0.22
Radiation	$CuK\alpha (\lambda = 1.54178)$
2θ range for data collection/°	17.458 to 136.312
Index ranges	$-6 \le h \le 6, -7 \le k \le 7, -13 \le 1 \le 13$
Reflections collected	5724
Independent reflections	$1310 [R_{int} = 0.0405, R_{sigma} = 0.0332]$
Data/restraints/parameters	1310/1/100
Goodness-of-fit on F ²	1.099
Final R indexes [I>=2 σ (I)]	$R_1 = 0.0317, wR_2 = 0.0831$
Final R indexes [all data]	$R_1 = 0.0318, wR_2 = 0.0832$
Largest diff. peak/hole / e Å ⁻³	0.17/-0.16
Flack parameter	-0.04(7)