

SUPPLEMENTARY INFORMATION

FOR

Deoxygenation of N-heterocyclic *N*-oxides using isopropanol as the recyclable reductant

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TABLE OF CONTENTS

Table of Contents.....	S2
General Information.....	S7
General Methods.....	S7
Chromatography.....	S7
NMR & IR Spectroscopy.....	S7
Mass Spectrometry.....	S8
Starting Materials.....	S8
Experimental Data.....	S9
Experimental Procedures and Compounds Characterisation Data.....	S9
Preparation of N-Heterocyclic <i>N</i> -Oxides (1).....	S9
Preparation of Methyl Quinoline-3-carboxylate <i>N</i> -Oxide (1a).....	S9
Preparation of Methyl Quinoline-6-carboxylate <i>N</i> -Oxide (1b).....	S10
Preparation of Ethyl Quinoline-6-carboxylate <i>N</i> -Oxide (1c).....	S11
Preparation of Isopropyl Quinoline-6-carboxylate <i>N</i> -Oxide (1d).....	S12
Preparation of <i>tert</i> -Butyl Quinoline-6-carboxylate <i>N</i> -Oxide (1e).....	S13
Preparation of 3-Acetylquinoline <i>N</i> -Oxide (1f).....	S13
Preparation of <i>N,N</i> -Diethyl-3-quinolinecarboxamide <i>N</i> -Oxide (1g).....	S14
Preparation of <i>N,N</i> -Diphenyl-3-quinolinecarboxamide <i>N</i> -Oxide (1h).....	S15
Preparation of 6-Quinolinecarboxamide <i>N</i> -Oxide (1i).....	S16
Preparation of <i>N,N</i> -Dibenzyl-6-quinolinecarboxamide <i>N</i> -Oxide (1j).....	S17
Preparation of 2-Quinolinecarbonitrile <i>N</i> -Oxide (1k).....	S19
Preparation of 3-Quinolinecarbonitrile <i>N</i> -Oxide (1l).....	S19
Preparation of 4-Chloroquinoline <i>N</i> -Oxide (1m).....	S19
Preparation of 6-Fluoroquinoline <i>N</i> -Oxide (1n).....	S20
Preparation of 6-Chloroquinoline <i>N</i> -Oxide (1o).....	S20
Preparation of 6-Bromoquinoline <i>N</i> -Oxide (1p).....	S21
Preparation of 6-Iodoquinoline <i>N</i> -Oxide (1q).....	S21
Preparation of 3-Bromoquinoline <i>N</i> -Oxide (1r).....	S21

Preparation of 6-Phenylquinoline <i>N</i> -Oxide (1s).....	S22
Preparation of 2-Phenylquinoline <i>N</i> -Oxide (1t).....	S22
Preparation of 2,6-Dimethylquinoline <i>N</i> -Oxide (1u).....	S23
Preparation of 6-Methylquinoline <i>N</i> -Oxide (1v).....	S23
Preparation of 3-Methylquinoline <i>N</i> -Oxide (1w).....	S23
Preparation of 6-Methoxyquinoline <i>N</i> -Oxide (1x).....	S24
Preparation of 6-Benzoyloxyquinoline <i>N</i> -Oxide (1y).....	S24
Preparation of 3-[Bis(<i>tert</i> -butoxycarbonyl)amino]quinoline <i>N</i> -Oxide (1z).....	S25
Preparation of Benzo[<i>h</i>]quinoline <i>N</i> -Oxide (1aa).....	S26
Preparation of Benzo[<i>f</i>]quinoline <i>N</i> -Oxide (1ab).....	S27
Preparation of 1-Isoquinolinecarbonitrile <i>N</i> -Oxide (1ac).....	S28
Preparation of 1-Benzoylisoquinoline <i>N</i> -Oxide (1ad).....	S29
Preparation of 1-Acetylisquinoline <i>N</i> -Oxide (1ae).....	S29
Preparation of Methyl 1-Isoquinolinecarboxylate <i>N</i> -Oxide (1af).....	S30
Preparation of Methyl 2-quinoxalinecarboxylate <i>N</i> -Oxide (1ag).....	S31
Preparation of 4-Phenylpyridine <i>N</i> -Oxide (1ah).....	S32
Preparation of 3-Phenylpyridine <i>N</i> -Oxide (1ai).....	S33
Preparation of 2-Phenylpyridine <i>N</i> -Oxide (1aj).....	S33
Preparation of 4-Benzoylpyridine <i>N</i> -Oxide (1ak).....	S34
Preparation of 3-Benzoylpyridine <i>N</i> -Oxide (1al).....	S34
Preparation of 2-Benzoylpyridine <i>N</i> -Oxide (1am).....	S35
Preparation of 4-Acetylpyridine <i>N</i> -Oxide (1an).....	S35
A Comprehensive Optimisation Study	S36
Table S1. Catalyst Screening.....	S36
Table S2. Solvent Screening.....	S37
Table S3. Further Solvent Screening.....	S38
Table S4. Screening the Equivalent of IPA.....	S39
Table S5. Further Screening the Equivalent of IPA.....	S39
Table S6. Screening the Molarity of Acetone.....	S40
Table S7. Further Screening the Molarity of Acetone.....	S40

Table S8. Screening Mes-Acr ⁺ Derivatives as a Photocatalyst.....	S41
Table S9. Confirming the Optimised Conditions.....	S42
General Procedure for the Optimisation Studies (Table 1).....	S43
General Procedure for the Organic Photoredox-Catalysed Deoxygenation Reaction of N-Heterocyclic N-Oxide Using Isopropyl Alcohol as a Sustainable Reducing Agent (Table 2).....	S44
Characterisation Data for the Deoxygenated N-Heterocycles (2).....	S44
Preparation of Methyl Quinoline-3-carboxylate (2a).....	S44
Preparation of Methyl Quinoline-6-carboxylate (2b).....	S45
Preparation of Ethyl Quinoline-6-carboxylate (2c).....	S45
Preparation of Isopropyl Quinoline-6-carboxylate (2d).....	S46
Preparation of <i>tert</i> -Butyl Quinoline-6-carboxylate (2e).....	S46
Preparation of 3-Acetylquinoline (2f).....	S47
Preparation of <i>N,N</i> -Diethyl-3-quinolinecarboxamide (2g).....	S47
Preparation of <i>N,N</i> -Diphenyl-3-quinolinecarboxamide (2h).....	S48
Preparation of 6-Quinolinecarboxamide (2i).....	S48
Preparation of <i>N,N</i> -Dibenzyl-6-quinolinecarboxamide (2j).....	S49
Preparation of 2-Quinolinecarbonitrile (2k).....	S49
Preparation of 3-Quinolinecarbonitrile (2l).....	S50
Preparation of 4-Chloroquinoline (2m).....	S50
Preparation of 6-Fluoroquinoline (2n).....	S51
Preparation of 6-Chloroquinoline (2o).....	S51
Preparation of 6-Bromoquinoline (2p).....	S52
Preparation of 6-Iodoquinoline (2q).....	S52
Preparation of 3-Bromoquinoline (2r).....	S52
Preparation of 6-Phenylquinoline (2s).....	S53
Preparation of 2-Phenylquinoline (2t).....	S53
Preparation of 2,6-Dimethylquinoline (2u).....	S54
Preparation of 6-Methylquinoline (2v).....	S54
Preparation of 3-Methylquinoline (2w).....	S54
Preparation of 6-Methoxyquinoline (2x).....	S55

Preparation of 6-Benzoyloxyquinoline (2y).....	S55
Preparation of 3-[Bis(<i>tert</i> -butoxycarbonyl)amino]quinoline (2z).....	S56
Preparation of Benzo[<i>h</i>]quinoline (2aa).....	S56
Preparation of Benzo[<i>f</i>]quinoline (2ab).....	S57
Preparation of 1-Isoquinolinecarbonitrile (2ac).....	S57
Preparation of 1-Benzoylisoquinoline (2ad).....	S58
Preparation of 1-Acetylisquinoline (2ae).....	S58
Preparation of Methyl 1-Isoquinolinecarboxylate (2af).....	S59
Preparation of Methyl 2-quinoxalinecarboxylate (2ag).....	S59
Preparation of 4-Phenylpyridine (2ah).....	S60
Preparation of 3-Phenylpyridine (2ai).....	S60
Preparation of 2-Phenylpyridine (2aj).....	S60
Preparation of 4-Benzoylpyridine (2ak).....	S61
Preparation of 3-Benzoylpyridine (2al).....	S61
Preparation of 2-Benzoylpyridine (2am).....	S62
Preparation of 4-Acetylpyridine (2an).....	S62
Preparation of 2-(2-(<i>N</i> -Oxidopyridine-2-yl)ethyl)quinoline (1ao').....	S63
Preparation of 2-(2-(<i>N</i> -Oxidopyridine-2-yl)ethyl)quinoline <i>N</i> -Oxide (1ao).....	S63
Preparation of 2-(2-(2-Quinoliny)ethyl)pyridine <i>N</i> -Oxide (2ao).....	S64
Experimental Procedure of Mechanistic studies.....	S66
Experimental Procedure for Scheme 1a (In the Absence of a Photocatalyst)	S66
Experimental Procedure for Scheme 1b (In the Absence of Visible Light)	S66
Experimental Procedure for Quantum yield(Φ) measurement.....	S66
Fig. S1 UV-Vis spectra of ferrioxalate/1,10-phenanthroline solutions after light irradiation.....	S67
Fig. S2 Moles of Fe ²⁺ formed vs. irradiation time.....	S68
Experimental Procedure for Scheme 1c (Visible Light On/Off Experiment)	S69
Experimental Procedure for Scheme 1d (Employing Radical Inhibitor)	S70
Experimental Procedure for Scheme 1e (Radical Trapping Experiment)	S70
Experimental Procedure for Radical Trapping Experiment II (Footnote 46)	S71
Stern-Volmer Luminescence Quenching Experiments (Fig. 2)	S72

Fig. S3 Emission Quenching of 7 by 1x	S72
Fig. S4 Emission Quenching of 7 by quinoline <i>N</i> -oxide.....	S72
Fig. S5 Emission Quenching of 7 by 1b	S72
Fig. S6 Emission Quenching of 7 by IPA.....	S72
Cyclic Voltammetry Measurements (Fig.2).....	S73
Fig. S7 Cyclic Voltammetry Measurement of 1x	S73
Fig. S8 Cyclic Voltammetry Measurement of quinoline <i>N</i> -oxide.....	S74
Fig. S9 Cyclic Voltammetry Measurement of 1b	S74
NMR Spectroscopic Studies for Confirming the Generation of Acetone as a Byproduct.....	S75
Fig. S10 Acetone Confirm in ¹ H NMR.....	S75
Fig. S11 Acetone Confirm in ¹³ C NMR.....	S75
Experimental Procedure for Multigram-Scale Deoxygenation Reaction (Scheme 3).....	S76
Fig. S12 Gram-scale Photoreactor (light-off)	S76
Fig. S13 Gram-scale Photoreactor (light-on)	S76
Experimental Procedure for Recycling Process (Scheme 4)	S77
Table S10. Recycling Process.....	S78
Fig. S14 Recycling Process (simple distillation).....	S79
Fig. S15 Recycling Process (Kugelrohr distillation).....	S79
References.....	S80
NMR Spectra.....	S82

GENERAL INFORMATION

General Methods

Experiments involving moisture- and/or air-sensitive compounds were performed in oven- or flame-dried glassware with rubber septa under a positive pressure of nitrogen using standard Schlenk techniques. Nonaqueous reagents were transferred by hypodermic syringe. Heating was accomplished by silicon oil bath using a temperature controller. Brine is defined as a saturated aqueous solution of sodium chloride. Organic solutions were concentrated under reduced pressure at 30 °C (water bath temperature) using a Büchi rotary evaporator, unless otherwise noted. Reactions were magnetically stirred and monitored by thin-layer chromatography (TLC) or GC. Yields refer to chromatographically and spectroscopically (¹H NMR) homogeneous (>95%) materials, unless otherwise stated. Melting points were measured using a Stuart SMP50 apparatus and are uncorrected.

Chromatography

TLC was performed on E. Merck 60–F254 precoated plates (0.25 mm), and spots were visualised by UV fluorescence (254 nm) quenching, iodine vapor, and KMnO₄ staining. Flash chromatography was carried out with Merck silica gel 60 (200–400 mesh) according to the procedure of Still et al.¹

NMR & IR Spectroscopy

¹H and ¹³C NMR spectra were recorded with Bruker Avance III Ascend 500 (500 MHz and 126 MHz, respectively) and Avance III HD (400 MHz and 101 MHz, respectively) spectrometers. ¹⁹F NMR spectra were obtained on a Bruker Avance III Ascend 500 spectrometer (471 MHz). Chemical shifts of the ¹H NMR (CDCl₃: 7.26 ppm, MeOD: 4.78 ppm, DMSO-*d*₆: 2.50 ppm) and ¹³C NMR (CDCl₃: 77.00 ppm, MeOD: 49.15 ppm, DMSO-*d*₆: 39.50 ppm) spectra were referenced to residual solvent peaks or tetramethylsilane (0.00 ppm) as an internal standard. ¹H NMR spectra are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quarter, quin = quintet, sext = sextet, sept = septet, m = multiplet, br = broad), coupling constant (*J*) in Hz and integration. Infrared (IR) spectra were recorded on a Bruker AVATAR 370 DTGS spectrometer using thin film samples on KBr plates and are reported in terms of frequency of absorption (cm⁻¹).

Mass Spectrometry

Low-resolution mass spectra were acquired using a HP 6890 GC system coupled with a HP 5973 mass selective detector with electron impact (EI) mode or an Agilent 1260 infinity LC-MS system coupled with an Agilent 6120 quadrupole mass spectrometer with electrospray ionisation (ESI) mode. High-resolution mass spectra (HRMS) were obtained from the Organic Chemistry Research Center (OCRC) at Sogang University (Seoul, Korea) using a Bruker Compact ESI-TOF mass spectrometer.

Starting Materials

Unless otherwise stated, all reagents and solvents were purchased at the highest commercial quality from commercial suppliers (Sigma-Aldrich, TCI, Alpha Aesar, Strem, or Acros) and used as received without further purification.

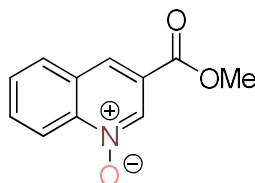
EXPERIMENTAL DATA

Experimental Procedures and Compounds Characterisation Data

Preparation of N-Heterocyclic N-Oxides (1)

Unless stated otherwise, N-heterocyclic N-oxides used in this study were prepared according to the literature procedures. They were further purified by recrystallisation from an appropriate mixture of solvents, as indicated, and then stored in a desiccator.

Preparation of Methyl Quinoline-3-carboxylate N-Oxide (1a)



A 250-mL Schlenk-type flask, equipped with a magnetic stir bar and a cold finger condenser, was charged with 3-quinolinecarboxylic acid (2.86 g, 16.5 mmol, 1.0 equivalent) and SOCl_2 (18 mL). The resulting mixture was heated at 60 °C for 4 h. To this mixture was added MeOH (60 mL), NEt_3 (24 mL), and the resulting mixture was stirred overnight at room temperature. The reaction mixture was poured into water (20 mL). The organic layer was separated, and the aqueous layer was extracted with EtOAc (15 mL \times 3). The combined organic layer was washed successively with aqueous solution of saturated NaHCO_3 , water, and brine, dried over anhydrous Na_2SO_4 , filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography ($\text{CH}_2\text{Cl}_2/\text{MeOH} = 10:1$) on silica gel to afford methyl quinoline-3-carboxylate (**2a**, 2.9 g, 96 %) as a white solid. A 100-mL round-bottomed flask was charged with methyl quinoline-3-carboxylate (748.8 mg, 4.00 mmol, 1.00 equivalent) and CH_2Cl_2 (13.3 mL), and the resulting mixture was cooled in an ice-water bath. A separate pear-shaped flask was charged with *m*-CPBA (77%, 1.34 g, 6.00 mmol, 1.5 equivalent) in CH_2Cl_2 (17 mL), and the resulting *m*-CPBA solution was added dropwise to the flask containing the pyridine at 0 °C via a syringe. The pear-shaped flask was rinsed with CH_2Cl_2 (1.0 mL \times 3), and the resulting solution was added via a syringe. After stirring for 10 minutes, the cold reaction mixture was allowed to slowly warm to room temperature. After stirring for 30 minutes at that temperature, the reaction mixture was poured

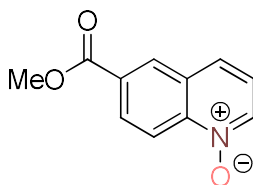
into water (20 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (15 mL × 3). The combined organic layer was washed successively with aqueous solution of saturated NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (EtOAc/MeOH = 9:1) on silica gel to afford an off-white solid, which was further purified by recrystallization from a hot mixture of EtOAc and hexanes (1:1, v/v) to yield the title compound as a white solid (565 mg, 71%). Spectroscopic data matched with those reported in the literature.^{4,6}

¹H NMR (500 MHz, CDCl₃) δ 9.0 (d, *J* = 1.4 Hz, 1H), 8.8–8.7 (m, 1H), 8.4 (s, 1H), 8.0 (dd, *J* = 8.2, 1.4 Hz, 1H), 7.8 (ddd, *J* = 8.5, 6.9, 1.4 Hz, 1H), 7.7 (ddd, *J* = 8.2, 6.9, 1.2 Hz, 1H), 4.0 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 163.7, 142.9, 134.8, 132.4, 129.4 (d, *J* = 10.0 Hz), 129.0, 127.5, 124.2, 119.8, 52.8.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₁H₁₀NO₃) 204.1, found 204.1.

Preparation of Methyl Quinoline-6-carboxylate *N*-Oxide (1b)



A 100-mL round-bottomed flask was charged with methyl quinoline-6-carboxylate (748.8 mg, 4.00 mmol, 1.00 equivalent) and CH₂Cl₂ (13.3 mL), and the resulting mixture was cooled in an ice-water bath. A separate pear-shaped flask was charged with *m*-CPBA (77%, 1.34 g, 6.00 mmol, 1.50 equivalent) in CH₂Cl₂ (17 mL), and the resulting *m*-CPBA solution was added dropwise to the flask containing the pyridine at 0 °C via a syringe. The pear-shaped flask was rinsed with CH₂Cl₂ (1.0 mL × 3), and the resulting solution was added via a syringe. After stirring for 10 minutes, the cold reaction mixture was allowed to slowly warm to room temperature. After stirring for 30 minutes at that temperature, the reaction mixture was poured into water (20 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (15 mL × 3). The combined organic layer was washed successively with aqueous solution of saturated NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (EtOAc/MeOH = 10:1) on silica gel to afford an off-white solid, which was further

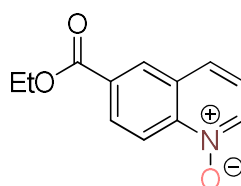
purified by recrystallization from a hot mixture of EtOAc and hexanes (10:1, v/v) to yield the title compound as a white solid (577 mg, 71%).

¹H NMR (300 MHz, CDCl₃) δ 8.7 (d, *J* = 10.9 Hz, 1H), 8.6–8.5 (m, 2H), 8.2 (dt, *J* = 9.2, 1.7 Hz, 1H), 7.8 (d, *J* = 8.3 Hz, 1H), 7.3 (dd, *J* = 8.5, 6.1 Hz, 1H), 3.9 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ 165.7, 143.2, 137.1, 131.0, 130.4, 129.9, 129.8, 126.6, 121.9, 120.3, 52.7.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₁H₁₀NO₃) 204.1, found 204.1.

Preparation of Ethyl Quinoline-6-carboxylate *N*-Oxide (**1c**)



A 100-mL schlenk flask, equipped with a magnetic stir bar and a reflux condenser, was charged with 6-quinolinecarboxylic acid (1.4 g, 8.00 mmol, 1.0 equivalent), ethanol (50 mL), and H₂SO₄ (1.0 mL). The resulting mixture was heated to reflux overnight. The mixture was allowed to cool to room temperature, diluted with CH₂Cl₂, and neutralized with an aqueous solution of saturated NaHCO₃. The mixture was then extracted with CH₂Cl₂ (15 mL × 3). The combined organic layer was washed successively with water and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (hexanes/EtOAc = 2:1) on silica gel to afford ethyl quinoline-6-carboxylate (**2c**, 1.2 g, 68 %) as a white solid. A 25-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with **2c** (503 mg, 2.50 mmol, 1.0 equivalent) and CH₂Cl₂ (7.0 mL). The resulting mixture was cooled in an ice-water bath. A separate pear-shaped flask was charged with *m*-CPBA (77%, 647 mg, 3.75 mmol, 1.5 equivalent) in CH₂Cl₂ (10 mL), and the resulting *m*-CPBA solution was added dropwise to the flask containing **2c** at 0 °C via a syringe. The pear-shaped flask was rinsed with CH₂Cl₂ (1.0 mL × 3), and the resulting solution was added via a syringe. After stirring for 10 minutes, the cold reaction mixture was allowed to slowly warm to room temperature. After stirring for 13 hours at that temperature, the reaction mixture was poured into water (20 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (15 mL × 3). The combined organic layer was washed successively with aqueous solution of saturated NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue

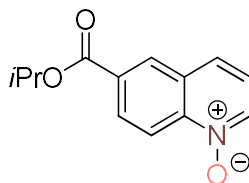
was purified by flash column chromatography (EtOAc/MeOH = 8:1) on silica gel to afford an off-white solid, which was further purified by recrystallization from a hot mixture of EtOAc and hexanes (1:1, v/v) to yield the title compound as a white solid (364 mg, 67%). Spectroscopic data matched with those reported in the literature.^{4,6}

¹H NMR (500 MHz, CDCl₃) δ 8.77 (d, *J* = 9.1 Hz, 1H), 8.59 (d, *J* = 1.8 Hz, 1H), 8.56 (dd, *J* = 6.0, 1.0 Hz, 1H), 8.31 (dd, *J* = 9.1, 1.8 Hz, 1H), 7.84–7.78 (m, 1H), 7.35 (dd, *J* = 8.4, 6.1 Hz, 1H), 4.44 (q, *J* = 7.1 Hz, 2H), 1.43 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 165.2, 143.2, 137.0, 130.8 (two carbons), 129.9, 129.8, 126.4, 121.8, 120.3, 61.7, 14.3.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₂H₁₂NO₃) 218.1, found 218.1

Preparation of Isopropyl Quinoline-6-carboxylate *N*-Oxide (**1d**)



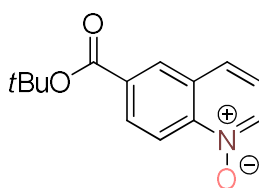
A 100-mL round-bottomed flask was charged with isopropyl quinoline-6-carboxylate (**1d**, 861 mg, 4.00 mmol, 1.00 equivalent) and CH₂Cl₂ (13.3 mL), and the resulting mixture was cooled in an ice-water bath. A separate pear-shaped flask was charged with *m*-CPBA (77%, 1.34 g, 6.00 mmol, 1.50 equivalent) in CH₂Cl₂ (17 mL), and the resulting *m*-CPBA solution was added dropwise to the flask containing the pyridine at 0 °C via a syringe. The pear-shaped flask was rinsed with CH₂Cl₂ (1.0 mL × 3), and the resulting solution was added via a syringe. After stirring for 10 minutes, the cold reaction mixture was allowed to slowly warm to room temperature. After stirring for 4 hours at that temperature, the reaction mixture was poured into water (20 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (15 mL × 3). The combined organic layer was washed successively with aqueous solution of saturated NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (EtOAc/MeOH = 8:1) on silica gel to afford an off-white solid, which was further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to yield the title compound as a white solid (604 mg, 65%).

¹H NMR (500 MHz, CDCl₃) δ 8.7 (d, *J* = 9.2 Hz, 1H), 8.5 (dd, *J* = 7.9, 1.7 Hz, 2H), 8.3 (dd, *J* = 9.1, 1.8 Hz, 1H), 7.8 (d, *J* = 8.5 Hz, 1H), 7.3 (dd, *J* = 8.4, 6.1 Hz, 1H), 5.3 (hept, *J* = 6.3 Hz, 1H), 1.4 (d, *J* = 6.4 Hz, 5H).

¹³C NMR (126 MHz, CDCl₃) δ 164.8, 143.3, 137.1, 131.2, 130.8, 130.0, 129.9, 126.6, 121.9, 120.3, 69.5, 22.0.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₃H₁₄NO₃) 232.1, found 232.1

Preparation of *tert*-Butyl Quinoline-6-carboxylate *N*-Oxide (1e)



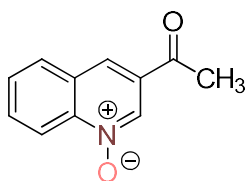
Prepared according to the literature procedure⁶ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, *v/v*) to afford the title compound as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.97 (dd, *J* = 4.2, 1.8 Hz, 1H), 8.50 (d, *J* = 1.9 Hz, 1H), 8.29–8.21 (m, 2H), 8.10 (d, *J* = 8.8 Hz, 1H), 7.43 (dd, *J* = 8.3, 4.2 Hz, 1H), 1.63 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 165.2, 152.2, 149.9, 137.2, 130.5, 129.9, 129.5, 129.0, 127.3, 121.6, 81.5, 28.2.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₄H₁₆NO₃) 246.1, found 246.1.

Preparation of 3-Acetylquinoline *N*-Oxide (1f)



A 100-mL round-bottomed flask was charged with methyl quinoline-3-carboxylate^{4,8} (748.8 mg, 4.00 mmol, 1.00 equivalent) and CH₂Cl₂ (13.3 mL), and the resulting mixture was cooled in an ice-water bath. A separate pear-shaped flask was charged with *m*-CPBA (77%, 1.34 g, 6.00 mmol, 1.50 equivalent) in CH₂Cl₂ (17 mL), and the resulting *m*-CPBA solution was added dropwise to the flask containing the pyridine at 0 °C via a syringe. The pear-shaped flask was rinsed with CH₂Cl₂ (1.0 mL × 3), and the

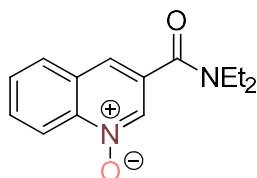
resulting solution was added via a syringe. After stirring for 10 minutes, the cold reaction mixture was allowed to slowly warm to room temperature. After stirring for 30 minutes at that temperature, the reaction mixture was poured into water (20 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (15 mL × 3). The combined organic layer was washed successively with aqueous solution of saturated NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (EtOAc/MeOH = 15:1) on silica gel to afford an off-white solid, which was further purified by recrystallization from a hot mixture of EtOAc and hexanes (10:1, v/v) to yield the title compound as a white solid (546 mg, 67%).

¹H NMR (500 MHz, CDCl₃) δ 8.9 (d, *J* = 1.1 Hz, 1H), 8.7 (d, *J* = 8.7 Hz, 1H), 8.2 (s, 1H), 8.0 (d, *J* = 8.1 Hz, 1H), 7.8 (ddd, *J* = 8.5, 6.9, 1.2 Hz, 1H), 7.7 (t, *J* = 7.5 Hz, 1H), 2.7 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 194.3, 143.0, 134.0, 132.7, 130.6, 129.9, 129.7, 129.2, 126.4, 120.0, 26.7.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₁H₁₀NO₂) 188.1, found 188.1.

Preparation of *N,N*-Diethyl-3-quinolinecarboxamide *N*-Oxide (1g)



A 25-mL round-bottomed flask was charged with *N,N*-diethyl-3-quinolinecarboxamide⁹ (114 mg, 0.500 mmol, 1.00 equivalent) and CH₂Cl₂ (1.7 mL), and the resulting mixture was cooled in an ice-water bath. A separate pear-shaped flask was charged with *m*-CPBA (77%, 168 mg, 0.750 mmol, 0.75 equivalent) in CH₂Cl₂ (1.5 mL), and the resulting *m*-CPBA solution was added dropwise to the flask containing the pyridine at 0 °C via a syringe. The pear-shaped flask was rinsed with CH₂Cl₂ (0.5 mL × 2), and the resulting solution was added via a syringe. After stirring for 10 minutes, the cold reaction mixture was allowed to slowly warm to room temperature. After stirring for 30 minutes at that temperature, the reaction mixture was poured into water (10 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (15 mL × 3). The combined organic layer was washed successively with aqueous solution of saturated NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, filtered,

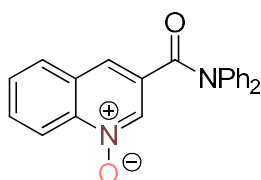
and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (EtOAc/MeOH = 9:1) on silica gel to afford the title compound as a yellow dense oil (64.4 mg, 53%).

¹H NMR (500 MHz, CDCl₃) δ 8.71 (d, *J* = 8.7 Hz, 1H), 8.51 (d, *J* = 1.4 Hz, 1H), 7.87 (dd, *J* = 8.1, 1.4 Hz, 1H), 7.78 (ddd, *J* = 8.6, 6.9, 1.4 Hz, 1H), 7.71 (s, 1H), 7.66 (ddd, *J* = 8.1, 6.8, 1.2 Hz, 1H), 3.45 (d, *J* = 111.4 Hz, 4H), 1.21 (d, *J* = 38.9 Hz, 6H).

¹³C NMR (126 MHz, CDCl₃) δ 166.2, 141.5, 133.5, 131.1, 130.9, 129.6, 129.4, 128.5, 123.0, 119.7, 77.3, 77.0, 76.7, 43.5, 39.7, 14.3, 12.8.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₄H₁₇N₂O₂) 245.1, found 245.1.

Preparation of *N,N*-Diphenyl-3-quinolinecarboxamide *N*-Oxide (**1h**)



A 25-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with 3-quinolinecarboxylic acid (520 mg, 3.00 mmol, 1.0 equivalent), 1,1'-carbodiimidazole (535 mg, 3.30 mmol, 1.1 equivalent), and dry DMF (4.4 mL, 0.69 M). After stirring in an oil bath (bath temperature = 40 °C) for 24 hours, the mixture was treated with diphenylamine (1.02 g, 6.00 mmol, 2.0 equivalent) followed by DBU (457 mg, 3.00 mmol, 1.0 equivalent). After stirring at 80 °C for 11 hours, the mixture was allowed to cool to room temperature and extracted with CH₂Cl₂ (10 mL × 3). The combined organic layer was washed with brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (hexanes/EtOAc = 3:2) on silica gel to afford *N,N*-diphenyl-3-quinolinecarboxamide (**2h**, 407 mg, 42 %) as an off-white solid. A 25-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with **2h** (407 mg, 1.26 mmol, 1.0 equivalent) and CH₂Cl₂ (2.1 mL). The resulting mixture was cooled in an ice-water bath. A separate pear-shaped flask was charged with *m*-CPBA (77%, 422 mg, 1.88 mmol, 1.5 equivalent) in CH₂Cl₂ (2.1 mL), and the resulting *m*-CPBA solution was added dropwise to the flask containing **2h** at 0 °C via a syringe. The pear-shaped flask was rinsed with CH₂Cl₂ (1.0 mL × 3), and the resulting solution was added via a syringe. After stirring for 10 minutes, the cold reaction mixture was allowed to slowly

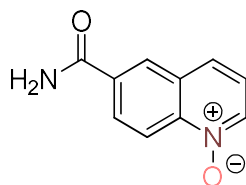
warm to room temperature. After stirring for 12 hours at that temperature, the reaction mixture was poured into water (20 mL). The organic layer was separated, and the aqueous layer was extracted with CH_2Cl_2 (10 mL \times 3). The combined organic layer was washed successively with an aqueous solution of saturated NaHCO_3 , water, and brine, dried over anhydrous Na_2SO_4 , filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (EtOAc as a sole eluent) on silica gel to afford an off-white solid, which was further purified by recrystallisation from a hot mixture of EtOAc and hexanes (5:1, *v/v*) to yield the title compound as a white solid (305 mg, 71%). All spectroscopic data matched with those reported in the literature.⁹

^1H NMR (500 MHz, CDCl_3) δ 8.66 (d, J = 8.7 Hz, 1H), 8.60 (s, 1H), 7.81–7.73 (m, 4H), 7.62 (t, J = 7.6 Hz, 1H), 7.35 (t, J = 7.6 Hz, 5H), 7.28–7.21 (m, 8H).

^{13}C NMR (126 MHz, CDCl_3) δ 165.9, 142.8, 141.6, 135.3, 131.7, 130.3, 129.6, 129.3, 129.1, 129.0, 127.3, 126.6, 119.8.

LC-MS (ESI) m/z calculated for $[\text{M}+1]^+$ ($\text{C}_{22}\text{H}_{17}\text{N}_2\text{O}_2$) 341.1, found 341.1.

Preparation of 6-Quinolinecarboxamide *N*-Oxide (1i)



A 50-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with methyl quinoline-6-carboxylate (**2b**, 740 mg, 3.95 mmol, 1.0 equivalent) and methanol (3 mL). The resulting mixture was heated in an oil bath at 45 °C and treated with aqueous ammonia (2.5 mL) via a syringe. After stirring for 10 minutes, the same volume of aqueous ammonia (2.5 mL) was added. The mixture was stirred for 10 minutes before being poured into water. The mixture was extracted with EtOAc (3 \times 30 mL). The combined organic layer was washed with brine, dried over Na_2SO_4 , filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (hexanes/EtOAc = 3:1 to 15:1) on silica gel to afford 6-quinolinecarboxamide (**2i**, 286 mg, 42%) as a white solid.

^1H NMR (300 MHz, MeOD) δ 8.87 (dd, J = 4.4, 1.7 Hz, 1H), 8.41 (dd, J = 17.2, 1.9 Hz, 2H), 8.16 (dd, J = 8.9, 2.1 Hz, 2H), 8.02 (d, J = 8.9 Hz, 1H), 7.54 (dd, J = 8.3, 4.3 Hz, 2H).

^{13}C NMR (75 MHz, MeOD) δ 171.3, 153.0, 149.9, 139.4, 133.4, 129.7, 129.5, 129.3, 129.0, 123.3.

GC-MS (EI, 70 eV) m/z calculated for $[M]^+$ ($C_{10}H_8N_2O$) 172.1, found 172.1.

A 50-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with **2i** (177 mg, 1.00 mmol, 1.0 equivalent) and CH_2Cl_2 (2 mL). The resulting mixture was cooled in an ice-water bath. In a pear-shaped flask, *m*-CPBA (77%, 690 mg, 4.00 mmol, 1.5 equivalent) was dissolved in CH_2Cl_2 (1.3 mL). The resulting *m*-CPBA solution was added dropwise to the flask containing **2i** at 0 °C via a syringe. After stirring for 5 minutes, the cold reaction mixture was allowed to slowly warm to room temperature and was then treated with CH_2Cl_2 (5 mL). After stirring overnight at room temperature, the reaction mixture was treated with K_2CO_3 (1.66 g, 12.0 mmol, 4.5 equivalent). The resulting mixture was stirred for 3 hours, filtered through a pad of Celite®, and washed with CH_2Cl_2 . The filtrate was concentrated using a rotary evaporator. After dissolving the reaction mixture in a 3:1 mixture of CH_2Cl_2 and MeOH, the resulting solution was directly loaded on a silica gel column and eluted with a 3:1 mixture of EtOAc and MeOH to afford 6-quinolinecarboxamide (**1i**, 124 mg, 66%) as a white solid.

¹H NMR (400 MHz, DMSO) δ 8.68–8.59 (m, 2H), 8.56 (d, J = 9.2 Hz, 1H), 8.33 (s, 1H), 8.22 (dd, J = 9.0, 2.0 Hz, 1H), 8.00 (d, J = 8.5 Hz, 1H), 7.70 (s, 1H), 7.53 (dd, J = 8.5, 6.1 Hz, 1H).

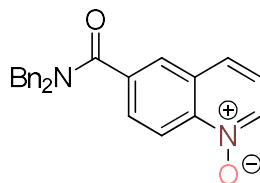
¹³C NMR (101 MHz, DMSO) δ 166.9, 141.7, 136.4, 134.3, 129.8, 128.7, 128.6, 125.9, 122.6, 119.2.

IR (KBr) 3378, 3199, 1656, 1625, 1611, 1588, 1461, 1407, 1316, 1268, 1025, 915, 857, 794 cm^{-1} .

HRMS (ESI, TOF) m/z calculated for $[M + Na]^+$ ($C_{10}H_8N_2NaO_2$) 211.0478, found 211.0478.

Melting point: 202–205 °C (dec.)

Preparation of *N,N*-Dibenzyl-6-quinolinecarboxamide *N*-Oxide (**1j**)



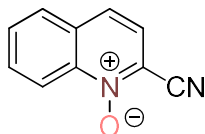
A 25-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with 6-quinolinecarboxylic acid (693 mg, 4.00 mmol, 1.0 equivalent), 1,1'-carbodiimidazole (685 mg, 4.00 mmol, 1.0 equivalent), and dry DMF (5.8 mL, 0.69 M). After stirring in an oil bath (bath temperature = 40 °C) for 2 hours, the mixture was treated with dibenzylamine (2.37 g, 12.0 mmol, 3.0 equivalent)

followed by DBU (609 mg, 4.00 mmol, 1.0 equivalent). After stirring at 80 °C for 15 hours, the mixture was allowed to cool to room temperature and extracted with CH₂Cl₂ (10 mL × 3). The combined organic layer was washed with brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (hexanes/EtOAc = 1:2) on silica gel to afford *N,N*-dibenzyl-6-quinolinecarboxamide (**2j**, 253 mg, 21 %) as an off-white solid. A 25-mL round-bottomed flask equipped with a magnetic stir bar was charged with **2j** (352 mg, 0.500 mmol, 1.0 equivalent) and CH₂Cl₂ (1.7 mL). The resulting mixture was cooled in an ice-water bath. A separate pear-shaped flask was charged with *m*-CPBA (77%, 168 mg, 0.750 mmol, 1.5 equivalent) in CH₂Cl₂ (1.7 mL), and the resulting *m*-CPBA solution was added dropwise to the flask containing **2j** at 0 °C via a syringe. The pear-shaped flask was rinsed with CH₂Cl₂ (1.0 mL × 3), and the resulting solution was added via a syringe. After stirring for 10 minutes, the cold reaction mixture was allowed to slowly warm to room temperature. After stirring for 12 hours at that temperature, the reaction mixture was poured into water (20 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (10 mL × 3). The combined organic layer was washed successively with an aqueous solution of saturated NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (EtOAc as a sole eluent) on silica gel to afford an off-white solid, which was further purified by recrystallisation from a hot mixture of EtOAc and hexanes (3:1, v/v) to yield the title compound as a white solid (135 mg, 73%). All spectroscopic data matched with those reported in the literature.⁸

¹H NMR (500 MHz, CDCl₃) δ 8.72 (d, *J* = 9.0 Hz, 1H), 8.48 (d, *J* = 6.0 Hz, 1H), 7.96 (s, 1H), 7.79 (d, *J* = 8.9 Hz, 1H), 7.64 (d, *J* = 8.5 Hz, 1H), 7.43–7.27 (m, 13H), 7.10 (s, 2H), 4.75 (s, 2H), 4.38 (s, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 170.4, 141.5, 136.6, 136.4, 136.1, 135.8, 130.1, 128.8, 128.7, 128.4, 128.2, 127.7, 127.7, 126.7, 126.5, 125.5, 121.8, 120.4, 51.5, 47.3.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₂₄H₂₁N₂O₂) 369.2, found 369.2

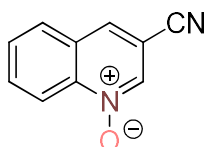
Preparation of 2-Quinolinecarbonitrile *N*-Oxide(1k)

Prepared according to the literature procedure⁸ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.66 (d, *J* = 8.9 Hz, 1H), 7.92 (dd, *J* = 8.1, 1.7 Hz, 1H), 7.83 (ddd, *J* = 8.7, 7.0, 1.5 Hz, 1H), 7.80–7.70 (m, 2H), 7.52 (d, *J* = 8.7 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 142.2, 131.6, 131.2, 131.1, 128.7, 124.7, 123.5, 121.5, 120.0, 113.0.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₀H₇N₂O) 171.1, found 171.1.

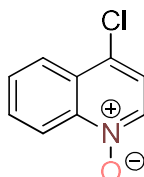
Preparation of 3-Quinolinecarbonitrile *N*-Oxide (1l)

Prepared according to the literature procedure^{4,8} and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.75 (d, *J* = 8.7 Hz, 1H), 8.63 (s, 1H), 8.09 (s, 1H), 8.01–7.91 (m, 2H), 7.81 (t, *J* = 7.6 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 143.3, 134.9, 133.5, 130.5, 130.0, 129.1, 129.0, 120.1, 114.9, 107.2.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₀H₇N₂O) 171.1, found 171.1.

Preparation of 4-Chloroquinoline *N*-Oxide (1m)

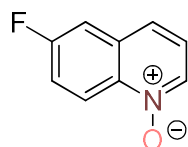
Prepared according to the literature procedure¹² and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.75 (dd, *J* = 8.8, 1.2 Hz, 1H), 8.42 (d, *J* = 6.6 Hz, 1H), 8.20 (dd, *J* = 8.4, 1.4 Hz, 1H), 7.82 (ddd, *J* = 8.6, 6.9, 1.4 Hz, 1H), 7.74 (ddd, *J* = 8.3, 6.9, 1.3 Hz, 1H), 7.36 (d, *J* = 6.6 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 142.2, 135.1, 131.1, 129.8, 129.7, 128.1, 125.2, 121.0, 120.4.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₉H₇ClNO) 180.0, found 180.0.

Preparation of 6-Fluoroquinoline *N*-Oxide (1n)



Prepared according to the literature procedure^{4,11} and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, *v/v*) to afford the title compound as a white solid.

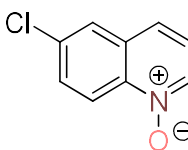
¹H NMR (500 MHz, CDCl₃) δ 8.76 (dd, *J* = 9.0, 5.2 Hz, 1H), 8.47 (d, *J* = 6.0 Hz, 1H), 7.66 (d, *J* = 8.5 Hz, 1H), 7.53–7.45 (m, 2H), 7.31 (dd, *J* = 8.5, 6.0 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 161.8 (d, ¹*J*_{C-F} = 251.8 Hz), 138.7, 135.0, 131.7 (d, *J* = 10.0 Hz), 124.9 (d, ⁴*J*_{C-F} = 5.8 Hz), 122.9 (d, ³*J*_{C-F} = 9.3 Hz), 122.2, 120.3 (d, ²*J*_{C-F} = 25.5 Hz), 111.5 (d, ²*J*_{C-F} = 22.7 Hz).

¹⁹F NMR (471 MHz, CDCl₃) δ -109.4.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₉H₇FNO) 164.1, found 164.1.

Preparation of 6-Chloroquinoline *N*-Oxide (1o)

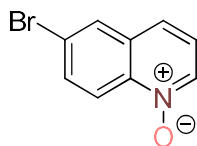


Prepared according to the literature procedure⁹ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, *v/v*) to afford the title compound as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.69 (d, *J* = 9.3 Hz, 1H), 8.49 (d, *J* = 7.1 Hz, 1H), 7.85 (d, *J* = 2.2 Hz, 1H), 7.71–7.60 (m, 2H), 7.32 (dd, *J* = 8.5, 6.1 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 140.3, 135.8, 135.2, 131.4, 131.3, 126.9, 124.8, 122.4, 121.9.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₉H₇ClNO) 180.0, found 180.0.

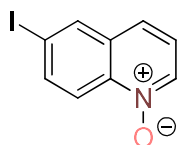
Preparation of 6-Bromoquinoline N-Oxide (1p)

Prepared according to the literature procedure¹⁹ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.62 (d, *J* = 9.2 Hz, 1H), 8.51 (d, *J* = 6.0 Hz, 1H), 8.04 (d, *J* = 2.0 Hz, 1H), 7.82 (dd, *J* = 9.2, 2.1 Hz, 1H), 7.63 (d, *J* = 8.5 Hz, 1H), 7.32 (dd, *J* = 8.5, 6.0 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 140.4, 135.8, 133.8, 131.6, 130.1, 124.6, 123.3, 122.2, 121.8.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₉H₇BrNO) 224.0, found 224.0.

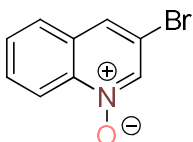
Preparation of 6-Iodoquinoline N-Oxide (1q)

Prepared according to the literature procedure⁸ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.49 (d, *J* = 6.1 Hz, 1H), 8.44 (d, *J* = 9.2 Hz, 1H), 8.25 (d, *J* = 1.9 Hz, 1H), 7.97 (dd, *J* = 9.1, 1.9 Hz, 1H), 7.59 (d, *J* = 8.5 Hz, 1H), 7.29 (dd, *J* = 8.5, 6.0 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 140.9, 139.0, 136.6, 135.8, 132.0, 124.2, 122.0, 121.5, 95.1.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₉H₇I_{NO}) 272.0, found 272.0.

Preparation of 3-Bromoquinoline N-Oxide (1r)

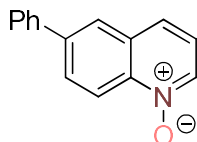
Prepared according to the literature procedure⁸ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.62 (dd, *J* = 8.7, 0.9 Hz, 1H), 8.58 (d, *J* = 1.6 Hz, 1H), 7.85 (s, 0H), 7.79–7.68 (m, 2H), 7.63 (ddd, *J* = 8.2, 6.9, 1.3 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 140.6, 137.1, 130.5, 130.3, 129.8, 127.6, 127.4, 119.9, 114.4.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₉H₇BrNO) 224.0, found 224.0.

Preparation of 6-Phenylquinoline N-Oxide (1s)



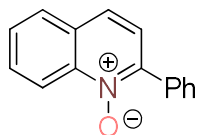
Prepared according to the literature procedure¹⁶ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.81 (d, *J* = 1.7 Hz, 1H), 8.72 (d, *J* = 8.9 Hz, 1H), 7.91–7.85 (m, 2H), 7.75–7.68 (m, 1H), 7.62 (dd, *J* = 7.2, 2.2 Hz, 3H), 7.52–7.46 (m, 2H), 7.43 (dd, *J* = 8.5, 6.2 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 140.4, 136.0, 135.1, 135.0, 130.4, 130.2, 129.4, 129.2, 129.0, 128.4, 127.1, 123.4, 119.8.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₅H₁₂NO) 222.1, found 222.1.

Preparation of 2-Phenylquinoline N-Oxide (1t)

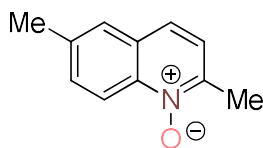


Prepared according to the literature procedure^{4,12,14} and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.84 (d, *J* = 8.7 Hz, 1H), 7.99–7.93 (m, 2H), 7.83 (dd, *J* = 8.2, 1.6 Hz, 1H), 7.79–7.69 (m, 2H), 7.61 (ddd, *J* = 8.2, 6.9, 1.3 Hz, 1H), 7.53–7.41 (m, 4H).

¹³C NMR (126 MHz, CDCl₃) δ 145.1, 142.3, 133.5, 130.6, 130.6, 129.6, 129.6, 128.5, 128.3, 128.0, 125.3, 123.4, 120.3.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₅H₁₂NO) 222.1, found 222.1.

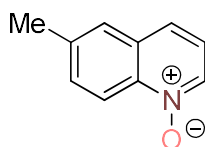
Preparation of 2,6-Dimethylquinoline N-Oxide (1u)

Prepared according to the literature procedure¹⁸ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, *v/v*) to afford the title compound as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.67 (d, *J* = 8.7 Hz, 1H), 7.60–7.53 (m, 3H), 7.28 (s, 1H), 2.71 (s, 3H), 2.53 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 144.9, 140.1, 137.7, 132.4, 129.3, 126.9, 124.6, 122.9, 119.3, 21.3, 18.6.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₁H₁₂NO) 174.1, found 174.1.

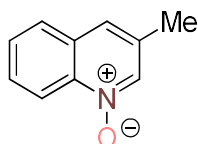
Preparation of 6-Methylquinoline N-Oxide (1v)

Prepared according to the literature procedure^{4,14} and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, *v/v*) to afford the title compound as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.63 (d, *J* = 8.9 Hz, 1H), 8.47 (d, *J* = 5.9 Hz, 1H), 7.65 (d, *J* = 8.5 Hz, 1H), 7.62 (s, 1H), 7.59 (dd, *J* = 8.9, 1.7 Hz, 1H), 7.27–7.23 (m, 1H), 2.54 (s, 3H)

¹³C NMR (101 MHz, CDCl₃) δ 140.1, 139.0, 135.0, 132.6, 130.7, 127.0, 125.5, 120.9, 119.6, 21.4

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₀H₁₀NO) 160.1, found 160.1.

Preparation of 3-Methylquinoline N-Oxide (1w)

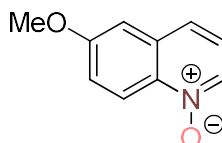
Prepared according to the literature procedure¹⁹ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, *v/v*) to afford the title compound as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.67 (d, *J* = 8.4 Hz, 1H), 8.40 (d, *J* = 1.5 Hz, 1H), 7.75 (dd, *J* = 8.1, 1.4 Hz, 1H), 7.66 (ddd, *J* = 8.6, 6.9, 1.4 Hz, 1H), 7.58 (ddd, *J* = 8.1, 6.9, 1.3 Hz, 1H), 7.50 (s, 1H), 2.43 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 139.7, 137.0, 131.2, 130.2, 129.3, 128.7, 127.4, 125.4, 119.6, 18.7.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₀H₁₀NO) 160.1, found 160.1.

Preparation of 6-Methoxyquinoline *N*-Oxide (**1x**)



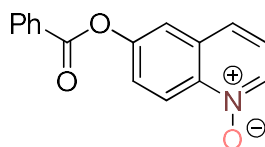
Prepared according to the literature procedure¹¹ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, *v/v*) to afford the title compound as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.65 (d, *J* = 9.6 Hz, 1H), 8.39 (dd, *J* = 6.0, 1.1 Hz, 1H), 7.63 (d, *J* = 8.5 Hz, 1H), 7.42–7.34 (m, 1H), 7.25–7.22 (m, 1H), 7.10 (d, *J* = 2.4 Hz, 1H), 3.94 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 159.5, 137.3, 133.8, 132.0, 125.0, 122.8, 121.5 (two carbons), 105.8, 55.7.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₀H₁₀NO₂) 176.1, found 176.1.

Preparation of 6-Benzoyloxyquinoline *N*-Oxide (**1y**)



A 100-ml round-bottomed flask, equipped with a magnetic stir bar, was charged with 6-hydroxyquinoline (155 mg, 1.07 mmol, 1.0 equivalent), 4-(*N,N*-dimethylamino)pyridine (DMAP, 6.6 mg, 0.054 mmol, 5 mol%), Et₃N (5 mL), and CH₂Cl₂ (25 mL). To this mixture was added dropwise benzoyl chloride (156 mg, 1.07 mmol, 1.0 equivalent) via a syringe at room temperature, and the resulting mixture was stirred for 1 hour. The reaction was quenched by H₂O (1 mL), and the mixture was extracted with CH₂Cl₂ (10 mL × 3). The combined organic layer was washed with brine, dried with Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (hexanes/EtOAc = 2:1) on silica gel to afford 6-benzoyloxyquinoline (**2y**, 188 mg, 71 %).

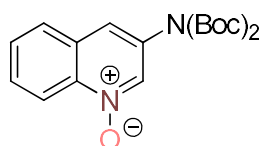
as an off-white solid. A 25-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with **2y** (188 mg, 0.760 mmol, 1.0 equivalent) and CH₂Cl₂ (1.2 mL). The resulting mixture was cooled in an ice-water bath. A separate pear-shaped flask was charged with *m*-CPBA (77%, 254 mg, 1.13 mmol, 1.5 equivalent) in CH₂Cl₂ (1.3 mL), and the resulting *m*-CPBA solution was added dropwise to the flask containing **2y** at 0 °C via a syringe. The pear-shaped flask was rinsed with CH₂Cl₂ (1 mL × 3), and the resulting solution was added via a syringe. After stirring for 10 minutes, the cold reaction mixture was allowed to slowly warm to room temperature. After stirring for 1 hour at that temperature, the reaction mixture was poured into water (20 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (10 mL × 3). The combined organic layer was washed successively with an aqueous solution of saturated NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (EtOAc/MeOH = 9:1) on silica gel to afford the title compound as an off-white solid, which was further purified by recrystallisation from a hot mixture of EtOAc and hexanes (5:1, *v/v*) to yield the title compound as a white solid (102 mg, 51 %). All spectroscopic data matched with those reported in the literature.⁸

¹H NMR (500 MHz, CDCl₃) δ 8.80 (d, *J* = 9.5 Hz, 1H), 8.49 (d, *J* = 6.0 Hz, 1H), 8.21 (d, *J* = 6.9 Hz, 2H), 7.76 (d, *J* = 2.6 Hz, 1H), 7.71–7.57 (m, 4H), 7.52 (t, *J* = 7.9 Hz, 2H), 7.29 (dd, *J* = 8.5, 6.0 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 164.8, 150.8, 139.7, 135.4, 134.1, 131.3, 130.3, 128.9, 128.8, 125.6, 125.5, 122.0, 121.9, 119.2.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₆H₁₂NO₃) 266.1, found 266.1.

Preparation of 3-[Bis(*tert*-butoxycarbonyl)amino]quinoline *N*-Oxide (**1z**)



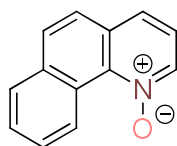
Prepared according to the literature procedure^{4,10} and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, *v/v*) to afford the title compound as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.68 (d, *J* = 8.8 Hz, 1H), 8.36 (d, *J* = 1.8 Hz, 1H), 7.82 (dd, *J* = 8.1, 1.3 Hz, 1H), 7.73 (ddd, *J* = 8.5, 6.9, 1.4 Hz, 1H), 7.62 (ddd, *J* = 8.1, 6.8, 1.2 Hz, 1H), 7.53 (d, *J* = 1.8 Hz, 1H), 1.39 (s, 18H).

^{13}C NMR (126 MHz, CDCl_3) δ 150.7, 140.6, 136.2, 133.2, 130.7, 129.3, 129.2, 128.4, 124.7, 119.8, 84.2, 27.9.

LC-MS (ESI) m/z calculated for $[\text{M}+1]^+$ ($\text{C}_{19}\text{H}_{25}\text{N}_2\text{O}_5$) 361.2, found 361.2.

Preparation of Benzo[*h*]quinoline *N*-Oxide (1aa)

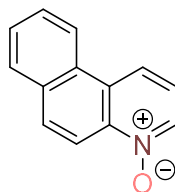


A 25-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with benzo[*h*]quinoline (**2aa**, 269 mg, 1.50 mmol, 1.0 equivalent) and CH_2Cl_2 (5 mL). The resulting mixture was cooled in an ice-water bath. In a pear-shaped flask, *m*-CPBA (77%, 388 mg, 2.25 mmol, 1.5 equivalent) was dissolved in CH_2Cl_2 (2.5 mL). The resulting *m*-CPBA solution was added dropwise to the flask containing **2aa** at 0 °C via a syringe. After stirring for 5 minutes, the cold reaction mixture was allowed to slowly warm to room temperature and stirred overnight. The reaction mixture was re-cooled to 0 °C and treated with an additional *m*-CPBA solution (77%, 129 mg, 0.75 mmol, 0.5 equivalent) in CH_2Cl_2 (2 mL) via a syringe. After stirring for 5 hours, the reaction mixture was poured into water (10 mL). The organic layer was separated, and the aqueous layer was extracted with CH_2Cl_2 (10 mL \times 3). The combined organic layer was washed successively with aqueous solution of saturated NaHCO_3 , water, and brine. It was then dried over anhydrous Na_2SO_4 , filtered, and concentrated on a rotary evaporator. The residue obtained was purified by flash column chromatography (EtOAc/MeOH = 7:1) on silica gel to afford the title compound (**1aa**, 204 mg, 70%) as a white solid. All spectroscopic data matched with those reported in the literature.²⁰

^1H NMR (400 MHz, CDCl_3) δ 10.84 (dd, J = 7.3, 2.6 Hz, 1H), 8.63 (dd, J = 6.3, 1.3 Hz, 1H), 7.88 (dd, J = 7.3, 2.2 Hz, 1H), 7.82–7.66 (m, 4H), 7.58 (d, J = 8.9 Hz, 1H), 7.34 (dd, J = 8.1, 6.2 Hz, 1H).

^{13}C NMR (101 MHz, CDCl_3) δ 139.3, 138.5, 134.1, 131.2, 130.6, 129.1, 128.3, 128.1, 127.7, 126.1, 125.7, 125.0, 121.2.

LC-MS (ESI) m/z calculated for $[\text{M}+1]^+$ ($\text{C}_{13}\text{H}_{10}\text{NO}$) 196.1, found 196.1.

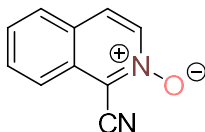
Preparation of Benzo[*f*]quinoline *N*-Oxide (1ab)

A 25-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with benzo[*f*]quinoline (**2ab**, 269 mg, 1.50 mmol, 1.0 equivalent) and CH₂Cl₂ (5 mL). The resulting mixture was cooled in an ice-water bath. In a pear-shaped flask, *m*-CPBA (77%, 388 mg, 2.25 mmol, 1.5 equivalent) was dissolved in CH₂Cl₂ (2.5 mL). The resulting *m*-CPBA solution was added dropwise to the flask containing **2ab** at 0 °C via a syringe. After stirring for 5 minutes, the cold reaction mixture was allowed to slowly warm to room temperature and stirred overnight. The reaction mixture was re-cooled to 0 °C and treated with an additional *m*-CPBA solution (77%, 129 mg, 0.75 mmol, 0.5 equivalent) in CH₂Cl₂ (2 mL) via a syringe. After stirring for 5 hours, the reaction mixture was poured into water (10 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (10 mL × 3). The combined organic layer was washed successively with aqueous solution of saturated NaHCO₃, water, and brine. It was then dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The residue obtained was purified by flash column chromatography (EtOAc/MeOH = 7:1) on silica gel to afford the title compound (**1ab**, 219 mg, 75%) as a white solid. All spectroscopic data matched with those reported in the literature.²⁵

¹H NMR (400 MHz, CDCl₃) δ 8.67 (d, *J* = 10.3 Hz, 1H), 8.59–8.46 (m, 2H), 8.41 (d, *J* = 8.7 Hz, 1H), 7.97 (d, *J* = 9.5 Hz, 1H), 7.93–7.86 (m, 1H), 7.72–7.61 (m, 2H), 7.39 (dd, *J* = 8.5, 6.1 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 141.1, 136.2, 132.1, 132.1, 129.1, 128.9, 128.6, 128.5, 128.0, 123.3, 121.4, 120.5, 117.2.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₃H₁₀NO) 196.1, found 196.1.

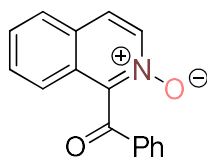
Preparation of 1-Isoquinolinecarbonitrile *N*-Oxide (1ac)

A 100-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with 1-isoquinolinecarbonitrile (**2ac**) (308 mg, 2.00 mmol, 1.00 equivalent) and CH₂Cl₂ (3.0 mL). The resulting mixture was cooled in an ice-water bath. A separate pear-shaped flask was charged with *m*-CPBA (77%, 672 mg, 3.00 mmol, 1.5 equivalent) in CH₂Cl₂ (4 mL), and the resulting *m*-CPBA solution was added dropwise to the flask containing **2ac** at 0 °C via a syringe. The pear-shaped flask was rinsed with CH₂Cl₂ (1.0 mL × 3), and the resulting solution was added via a syringe. After stirring for 10 minutes, the cold reaction mixture was allowed to slowly warm to room temperature. After stirring for 12 hours at that temperature, the reaction mixture was poured into water (20 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (15 mL × 3). The combined organic layer was washed successively with aqueous solution of saturated NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The residue thus obtained was purified by flash column chromatography (EtOAc/MeOH = 12:1) on silica gel to afford the title compound as a white solid, which was further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to yield the title compound as a white solid (186 mg, 55%). Spectroscopic data matched with those reported in the literature.^{9,24}

¹H NMR (500 MHz, CDCl₃) δ 8.16 (d, *J* = 7.2 Hz, 1H), 7.99 (dd, *J* = 8.4, 0.9 Hz, 1H), 7.87 (dt, *J* = 8.1, 0.8 Hz, 1H), 7.84 (d, *J* = 7.3 Hz, 1H), 7.80 (ddd, *J* = 8.4, 7.1, 1.2 Hz, 1H), 7.68 (ddd, *J* = 8.2, 7.0, 1.2 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 137.2, 131.9, 130.3, 129.7, 127.7, 127.6, 127.2, 123.2, 121.9, 111.3.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₃H₇N₂O) 171.1, found 171.1.

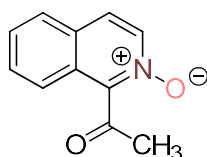
Preparation of 1-Benzoylisoquinoline *N*-Oxide (1ad)

Prepared according to the literature procedure²⁵ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.13 (d, *J* = 7.2 Hz, 1H), 7.84 (dt, *J* = 8.7, 1.7 Hz, 3H), 7.77 (d, *J* = 7.2 Hz, 1H), 7.59 (dddd, *J* = 8.2, 6.9, 3.5, 1.4 Hz, 2H), 7.54 (td, *J* = 7.6, 6.9, 1.4 Hz, 1H), 7.49 (d, *J* = 8.2 Hz, 1H), 7.47–7.42 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 189.7, 142.4, 136.7, 135.3, 134.6, 130.2, 129.3, 129.2, 129.1, 128.8, 127.5, 127.3, 125.0, 123.2.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₆H₁₂NO₂) 250.1, found 250.1.

Preparation of 1-Acetylisoquinoline *N*-Oxide (1ae)

A 100-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with isoquinoline (517 mg, 4.00 mmol, 1.0 equivalent), acetaldehyde (705 mg, 16.0 mmol, 4.0 equivalent), tetra-*n*-butylammonium bromide (387 mg, 1.20 mmol, 0.3 equivalent), K₂S₂O₈ (2.16 g, 8.00 mmol, 2.0 equivalent), and 1,2-dichloroethane (33 mL). The resulting mixture was heated in an oil bath (bath temperature = 100 °C) under reflux for 3 hours. The mixture was allowed to cool to room temperature, and the solvent was removed under vacuum. The crude mixture was diluted with CH₂Cl₂, washed successively with an aqueous solution of saturated NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography on silica gel (CH₂Cl₂ as a sole eluent) to afford 1-acetylisoquinoline (**2ae**, 437 mg, 64 %) as a white solid. A 25-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with **2ae** (437 mg, 2.55 mmol, 1.0 equivalent) and CH₂Cl₂ (4.0 mL). The resulting mixture was cooled in an ice-water bath. A separate pear-shaped flask was charged with *m*-CPBA (77%, 1.14 g,

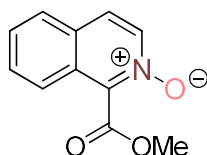
5.11 mmol, 2.0 equivalent) in CH₂Cl₂ (4.5 mL), and the resulting *m*-CPBA solution was added dropwise to the flask containing **2ae** at 0 °C via a syringe. The pear-shaped flask was rinsed with CH₂Cl₂ (1.0 mL × 3), and the resulting solution was added via a syringe. After stirring for 10 minutes, the cold reaction mixture was allowed to slowly warm to room temperature. After stirring for 17 hours at that temperature, the reaction mixture was poured into water (20 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (15 mL × 3). The combined organic layer was washed successively with an aqueous solution of saturated NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (EtOAc as a sole eluent) on silica gel to afford an off-white solid, which was further purified by recrystallisation from a hot mixture of hexanes and EtOAc (3:1, v/v) to yield the title compound as a white solid (209 mg, 44%). Spectroscopic data matched with those reported in the literature.⁸

¹H NMR (500 MHz, CDCl₃) δ 8.05 (d, *J* = 7.1 Hz, 1H), 7.82–7.74 (m, 1H), 7.67 (dd, *J* = 17.3, 7.6 Hz, 2H), 7.58 (pd, *J* = 6.9, 1.3 Hz, 4H), 2.73 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 197.0, 143.2, 136.4, 130.2, 128.9, 127.1, 126.0, 124.9, 123.0, 29.7.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₁H₁₀NO₂) 188.1, found 188.1.

Preparation of Methyl 1-Isoquinolinecarboxylate *N*-Oxide (**1af**)



Prepared according to the literature procedure²⁷. In a 50-mL Schlenk-type flask, equipped with a magnetic stir bar and a reflux condenser, 1-isoquinolinecarboxylic acid (346 mg, 2.00 mmol, 1.0 equivalent), MeOH (20 mL), and H₂SO₄ (1 mL) were combined. The resulting mixture was heated in an oil bath (bath temperature = 80 °C) under reflux overnight. The mixture was allowed to cool to room temperature, diluted with CH₂Cl₂, and neutralised with an aqueous solution of saturated NaHCO₃. The mixture was extracted with CH₂Cl₂ (10 mL × 3). The combined organic layer was washed successively with water and brine and dried over anhydrous Na₂SO₄. It was filtered and concentrated on a rotary evaporator. The residue obtained was purified by flash column chromatography (Hex/EtOAc = 2:1) on

silica gel to afford methyl 1-isoquinolinecarboxylate (**2af**, 329 mg, 88%) as a white solid. All spectroscopic data matched with those reported in the literature.

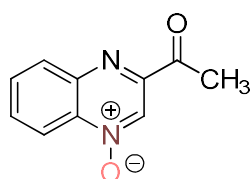
A 25-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with methyl 1-isoquinolinecarboxylate (**2af**, 281 mg, 1.50 mmol, 1.0 equivalent) and CH₂Cl₂ (5 mL). The resulting mixture was cooled in an ice-water bath. In a pear-shaped flask, *m*-CPBA (77%, 388 mg, 2.25 mmol, 1.5 equivalent) was dissolved in CH₂Cl₂ (2.5 mL). The resulting *m*-CPBA solution was added dropwise to the flask containing **2af** at 0 °C via a syringe. After stirring for 5 minutes, the cold reaction mixture was allowed to slowly warm to room temperature. After stirring overnight, the reaction mixture was poured into water (10 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (10 mL × 3). The combined organic layer was washed successively with aqueous solution of saturated NaHCO₃, water, and brine. It was dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (EtOAc/MeOH = 15:1) on silica gel to afford the title compound (**1af**, 238 mg, 78%) as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.07 (d, *J* = 7.2 Hz, 1H), 7.75 (dd, *J* = 8.4, 1.8 Hz, 1H), 7.66 (d, *J* = 7.2 Hz, 1H), 7.61–7.50 (m, 3H), 4.09 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 162.4, 138.6, 136.6, 130.2, 129.0, 128.4, 127.1, 126.5, 124.9, 123.0, 53.5.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₁H₁₀NO₃) 204.1, found 204.1

Preparation of Methyl 2-Quinoxalinecarboxylate *N*-Oxide (**1ag**)



A 150-mL round-bottomed flask, equipped with a magnetic stir bar and a reflux condenser, was charged with quinoxaline (651 mg, 5.00 mmol, 1.0 equivalent), acetaldehyde (1.10 g, 20.0 mmol, 4.0 equivalent), tetra-*n*-butylammonium bromide (484 mg, 1.50 mmol, 0.3 equivalent), K₂S₂O₈ (2.70 g, 10.0 mmol, 2.0 equivalent), and 1,2-dichloroethane (42 mL). The resulting mixture was heated in an oil bath (bath temperature = 100 °C) under reflux for 2 hours. The mixture was allowed to cool to room temperature, and the solvent was removed under vacuum. The crude mixture was diluted with CH₂Cl₂, washed

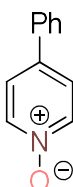
successively with an aqueous solution of saturated NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography on silica gel (hexanes/EtOAc = 7:1) to afford 2-acetylquinoxaline (**2ag**) as a white solid (437 mg, 64 %). A 50-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with **2ag** (528 mg, 3.08 mmol, 1.0 equivalent) and CH₂Cl₂ (5.0 mL). The resulting mixture was cooled in an ice-water bath. A separate pear-shaped flask was charged with *m*-CPBA (77%, 1.38 g, 6.16 mmol, 2.0 equivalent) in CH₂Cl₂ (5.3 mL), and the resulting *m*-CPBA solution was added dropwise to the flask containing **2ag** at 0 °C via a syringe. The pear-shaped flask was rinsed with CH₂Cl₂ (1.0 mL × 3), and the resulting solution was added via a syringe. After stirring for 10 minutes, the cold reaction mixture was allowed to slowly warm to room temperature. After stirring for 10 hours at that temperature, the reaction mixture was poured into water (20 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (15 mL × 3). The combined organic layer was washed successively with aqueous solution of saturated NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (hexene/EtOAc = 4:1) on silica gel to afford a yellow solid, which was further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to yield the title compound as a white solid (209 mg, 44%). Spectroscopic data matched with those reported in the literature.¹⁷

¹H NMR (500 MHz, CDCl₃) δ 8.86 (s, 1H), 8.55 (dd, *J* = 8.5, 1.7 Hz, 1H), 8.19 (dd, *J* = 8.3, 1.6 Hz, 1H), 7.84 (dddd, *J* = 25.8, 8.4, 7.0, 1.5 Hz, 2H), 2.79 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 197.9, 149.8, 144.4, 138.1, 132.3, 132.3, 132.1, 127.3, 119.2, 25.4.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₀H₉N₂O₂) 189.1, found 189.1.

Preparation of 4-Phenylpyridine *N*-Oxide (**1ah**)



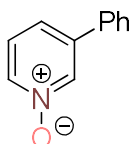
Prepared according to the literature procedure²¹ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.29–8.22 (m, 2H), 7.62–7.55 (m, 2H), 7.54–7.40 (m, 5H).

^{13}C NMR (101 MHz, CDCl_3) δ 139.3, 138.9, 136.1, 129.3, 129.2, 126.4, 123.7.

LC-MS (ESI) m/z calculated for $[\text{M}+1]^+$ ($\text{C}_{11}\text{H}_{10}\text{NO}$) 172.1, found 172.1.

Preparation of 3-Phenylpyridine *N*-Oxide (1ai)



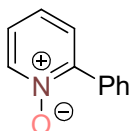
Prepared according to the literature procedure²² and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (2:1, v/v) to afford the title compound as a white solid.

^1H NMR (500 MHz, CDCl_3) δ 8.41 (t, J = 1.9 Hz, 1H), 8.15 (dt, J = 6.4, 1.2 Hz, 1H), 7.51–7.37 (m, 6H), 7.29 (dd, J = 8.1, 6.3 Hz, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 140.3, 137.6, 137.5, 135.1, 129.4 (two carbons), 126.9, 125.9, 124.6.

LC-MS (ESI) m/z calculated for $[\text{M}+1]^+$ ($\text{C}_{11}\text{H}_{10}\text{NO}$) 172.1, found 172.1.

Preparation of 2-Phenylpyridine *N*-Oxide (1aj)

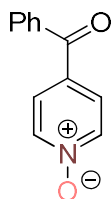


Prepared according to the literature procedure²³ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

^1H NMR (400 MHz, CDCl_3) δ 8.33 (dd, J = 6.5, 1.3 Hz, 1H), 7.82 (dd, J = 8.0, 1.7 Hz, 2H), 7.53–7.39 (m, 4H), 7.30 (td, J = 7.7, 1.3 Hz, 1H), 7.22 (ddd, J = 7.6, 6.5, 2.2 Hz, 1H).

^{13}C NMR (101 MHz, CDCl_3) δ 149.3, 140.5, 132.6, 129.6, 129.2, 128.3, 127.4, 125.5, 124.5.

LC-MS (ESI) m/z calculated for $[\text{M}+1]^+$ ($\text{C}_{11}\text{H}_{10}\text{NO}$) 172.1, found 172.1.

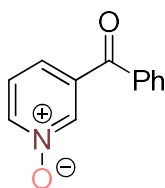
Preparation of 4-Benzoylpyridine N-Oxide (1ak)

Prepared according to the literature procedure²⁴ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.26–8.20 (m, 2H), 7.76–7.65 (m, 4H), 7.65–7.58 (m, 1H), 7.50 (tt, *J* = 7.5, 1.5 Hz, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 192.1, 139.4 (two carbons), 136.1, 133.3, 132.9, 129.6 (two carbons), 128.8 (two carbons), 127.0 (two carbons).

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₂H₁₀NO₂) 200.1, found 200.1.

Preparation of 3-Benzoylpyridine N-Oxide (1al)

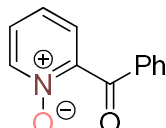
A 25-mL round-bottomed flask, equipped with a magnetic stir bar, was charged with 3-benzoylpyridine (**2al**, 275 mg, 1.50 mmol, 1.0 equivalent) and CH₂Cl₂ (5 mL). The resulting mixture was cooled in an ice-water bath. In a pear-shaped flask, *m*-CPBA (77%, 388 mg, 2.25 mmol, 1.5 equivalent) was dissolved in CH₂Cl₂ (2.5 mL). The resulting *m*-CPBA solution was added dropwise to the flask containing **2al** at 0 °C via a syringe. After stirring for 5 minutes, the cold reaction mixture was allowed to slowly warm to room temperature. After stirring overnight, the reaction mixture was poured into water (10 mL). The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (10 mL × 3). The combined organic layer was then washed successively with aqueous solution of saturated NaHCO₃, water, and brine. It was dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography (EtOAc/MeOH = 8:1) on silica gel to afford the title compound (**1al**, 63 mg, 21%) as a brown solid. All spectroscopic data matched with those reported in the literature.²⁶

¹H NMR (400 MHz, CDCl₃) δ 8.46 (s, 1H), 8.30 (d, *J* = 6.5 Hz, 1H), 7.73 (d, *J* = 7.9 Hz, 2H), 7.58 (dd, *J* = 10.2, 4.2 Hz, 2H), 7.50–7.42 (m, 2H), 7.42–7.32 (m, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 191.6, 141.7, 140.1, 136.6, 135.5, 133.8, 129.9, 128.8, 126.5, 125.9.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₂H₁₀NO₂) 200.1, found 200.1.

Preparation of 2-Benzoylpyridine *N*-Oxide (1am)



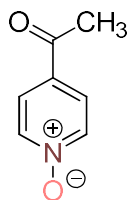
Prepared according to the literature procedure¹³ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.20 (ddd, *J* = 4.9, 2.6, 1.1 Hz, 1H), 7.86–7.77 (m, 2H), 7.62–7.53 (m, 1H), 7.44 (tt, *J* = 7.7, 1.8 Hz, 2H), 7.40–7.33 (m, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 189.4, 147.1, 140.0, 135.1, 134.2, 129.3 (two carbons), 128.9 (two carbons), 127.1, 125.7, 125.7.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₂H₁₀NO₂) 200.1, found 200.1.

Preparation of 4-Acetylpyridine *N*-Oxide (1an)



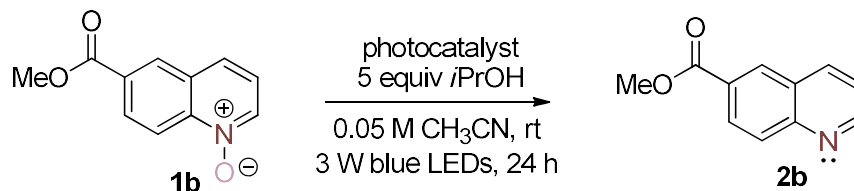
Prepared according to the literature procedure¹⁵ and further purified by recrystallisation from a hot mixture of hexanes and EtOAc (1:1, v/v) to afford the title compound as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.19 (d, *J* = 6.7 Hz, 2H), 7.76 (d, *J* = 7.3 Hz, 2H), 2.55 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 193.7, 139.6 (two carbons), 132.3, 125.1 (two carbons), 26.3.

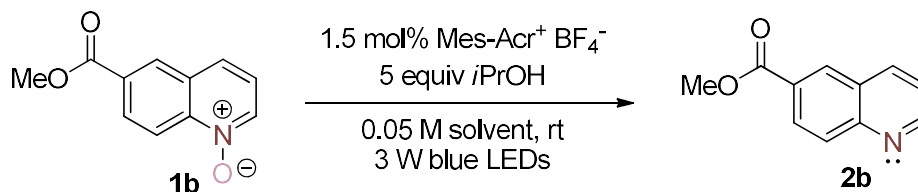
LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₇H₈NO₂) 138.1, found 138.1.

A Comprehensive Optimisation Study

Table S1. Catalyst Screening^a

Entry	Photocatalyst (mol%)	Yield (%)
1 ^b	eosin Y disodium salt (1.0)	11
2 ^b	eosin Y disodium salt (3.0)	21
3 ^b	eosin Y (1.0)	10
4 ^b	eosin Y (3.0)	22
5 ^b	rosebengal (1.0)	trace
6 ^b	rosebengal (3.0)	6
7 ^b	rosebengal (5.0)	10
8 ^b	rhodamine B (1.0)	trace
9 ^b	rhodamine 6G (1.0)	trace
10	9,10-dicyanoanthracene (1.0)	trace
11	fluorescein (1.0)	trace
12	4CzIPN (1.0) (3)	50
13	2,4,6-triphenylpyrylium BF ₄ ⁻ (1.0) (4)	30
14	Mes-Acr ⁺ BF ₄ ⁻ (1.0) (5)	63
15	Mes-Acr ⁺ ClO ₄ ⁻ (1.0) (6)	60
16	9-mesityl-10-phenylacridinium ⁺ BF ₄ ⁻ (1.0)	57

^aConditions: **1b** (0.20 mmol), IPA (0.08 mL), and a photocatalyst (1.0–5.0 mol%) in degassed acetonitrile (4 mL, 0.05 M). ^bUnder the irradiation of green LEDs.

Table S2. Solvent Screening^a

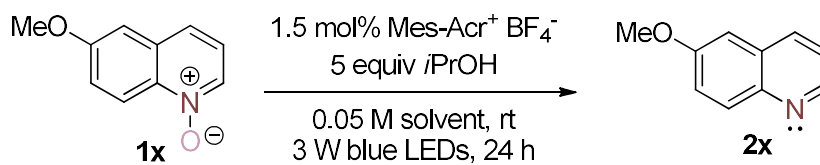
Entry	Solvent	Time (h)	Yield (%)
1	DCM	24	34
2	DCE	24	33
3	MeOH	20	45
4	EtOH	22	40
5	BuOH	23	41
6	toluene	18	82
7	xylene	20	63
8	THF	10	41
9	DMA	23	47
10	DMSO	14	53
11	DMF	24	49
12	acetonitrile	24	63
13	acetone	16	78
14	1,4-dioxane	4.5	58
15	2-butanone	22	56
16	toluene/acetonitrile (7/1)	13	88
17	toluene/DCM (40/1)	13	84
18	toluene/THF (3/1)	12	80
19	toluene/acetone (3/1)	14	83
20	toluene/DMSO (5/1)	24	76
21	toluene/anisole (20/1)	13	81
22	toluene/butanone (3/1)	12	86
23	toluene/acetonitrile (10/1)	13	79
24	toluene/acetonitrile (4/1)	19	84

25

toluene/acetonitrile (1/1)

24

80

^aConditions: **1b** (0.20 mmol), IPA (0.08 mL), and a photocatalyst (1.5 mol%) in degassed solvent (4 mL, 0.05 M).**Table S3. Further Solvent Screening^a**

Entry	solvent	Yield (%)
1	toluene/acetonitrile (7/1)	43
2	acetone	54
3	2-butanone	42
4	acetonitrile	33
5	1,4-dioxane	32
6	DCM	34
7	MeOH	39
8	toluene	53
9	THF	38
10	DMF	36

^aConditions: **1x** (0.20 mmol), IPA (0.08 mL), and a photocatalyst (1.5 mol%) in degassed solvent (4 mL, 0.05 M).

Table S4. Screening the Equivalent of IPA^a

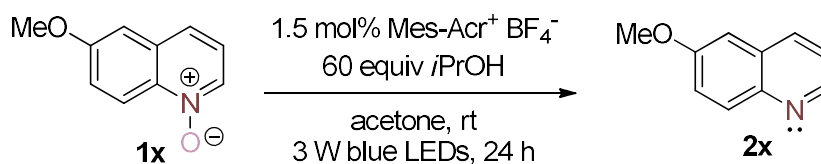
Entry	Equivalent of IPA	Time (h)	Yield (%)
1	1.1	24	43
2	20	10.5	72
3	40	9	74
4	60	8	77
5	75	7.5	76
6	120	6	66

^aConditions: **1b** (0.20 mmol), IPA (1.1–120 equivalent), and a photocatalyst (0.5–1.5 mol%) in degassed acetone (4 mL, 0.05 M).

Table S5. Further Screening the Equivalent of IPA^a

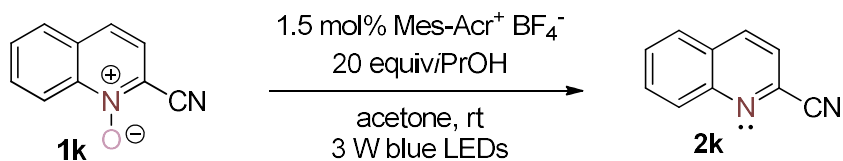
Entry	Equivalent of IPA	Yield (%)
1	1.1	14
2	10	31
3	30	44
4	40	50
5	50	57
6	60	64
7	75	55
8	90	52

^aConditions: **1x** (0.20 mmol), IPA (1.1–90 equivalent), and a photocatalyst (1.5 mol%) in degassed solvent (4 mL, 0.05 M).

Table S6. Screening the Molarity of Acetone^a

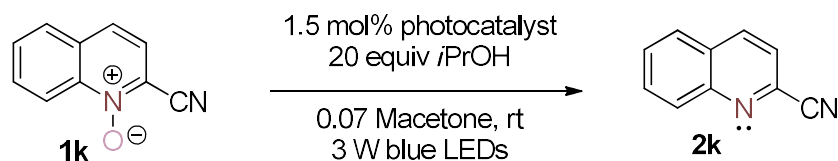
Entry	Molarity of Acetone	Yield (%)
1	0.3	7
2	0.2	18
3	0.1	33
4	0.07	64
5	0.05	63
6	0.03	58

^aConditions: **1x** (0.20 mmol), IPA (0.92 mL), and a photocatalyst (1.5 mol%) in degassed solvent (0.67–6.7 mL, 0.1–0.03 M).

Table S7. Further Screening the Molarity of Acetone^a

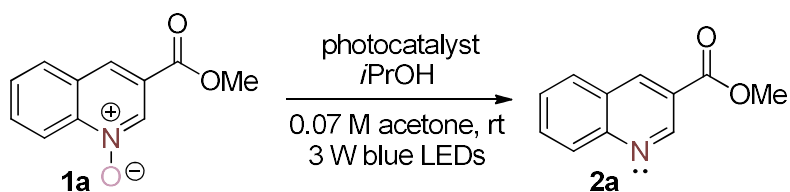
Entry	Molarity of Acetone	Time (h)	Yield (%)
1	0.1	5	73
2	0.07	5	83
3	0.05	6.5	77

^aConditions: **1k** (0.20 mmol), IPA (0.30 mL), and a photocatalyst (1.5 mol%) in degassed solvent (2–4 mL, 0.1–0.05 M).

Table S8. Screening Mes-Acr⁺ Derivatives as a Photocatalyst^a

Entry	Photocatalyst (mol%)	Time (h)	Yield (%)
1	Mes-Acr ⁺ BF ₄ ⁻ (1.5) (5)	6.5	83
2	Mes-Acr ⁺ ClO ₄ ⁻ (1.5) (6)	7	81
3	9-mesityl-10-phenylacridinium ⁺ BF ₄ ⁻ (1.5)	11	80
4	Mes- <i>t</i> Bu ₂ Acr ⁺ BF ₄ ⁻ (1.5) (7)	5	90
5	10,10'-dimethyl-9,9'-biacridinium ²⁺ 2 NO ₃ ⁻ (1.5)	24	48
6	9-phenylacridine (1.5)	24	51
7	10-methyl-9-phenylacridinium ⁺ ClO ₄ ⁻ (1.5)	22	73
8	2,7,10-trimethyl-9-(2,4,6-trimethylphenyl)acridinium ⁺ ClO ₄ ⁻ (1.5)	16	86
9	9-(2,6-dimethylphenyl)-10-methylacridinium ⁺ ClO ₄ ⁻ (1.5)	24	77
10	9-[1,1'-biphenyl]-2-yl-10-methylacridinium ⁺ ClO ₄ ⁻ (1.5)	22	80

^aConditions: **1k** (0.20 mmol), IPA (0.30 mL), and a photocatalyst (1.5 mol%) in degassed solvent (2.9 mL, 0.07 M).

Table S9. Confirming the Optimised Conditions^a

Entry	Photocatalyst (mol%)	Equivalent of IPA	Time (h)	Yield (%)
1 ^b	eosin Y disodium salt (1.0)	30	24	Trace
2 ^b	rosebengal (1.0)	30	24	Trace
3	4CzIPN (1.0) (3)	30	7	69
4	2,4,6-triphenylpyrylium BF ₄ ⁻ (1.0) (4)	30	24	42
5	Mes-Acr ⁺ BF ₄ ⁻ (1.0) (5)	30	8	78
6	Mes-Acr ⁺ ClO ₄ ⁻ (1.0) (6)	30	8.5	77
7	Mes- <i>t</i> Bu ₂ Acr ⁺ BF ₄ ⁻ (1.0) (7)	30	4	79
8	Mes- <i>t</i> Bu ₂ Acr ⁺ BF ₄ ⁻ (1.0) (7)	1.1	22	43
9	Mes- <i>t</i> Bu ₂ Acr ⁺ BF ₄ ⁻ (1.0) (7)	15	5	76
10	Mes- <i>t</i> Bu ₂ Acr ⁺ BF ₄ ⁻ (1.0) (7)	4	3	85
11	Mes- <i>t</i> Bu ₂ Acr ⁺ BF ₄ ⁻ (1.0) (7)	60	3	89
12	Mes- <i>t</i> Bu ₂ Acr ⁺ BF ₄ ⁻ (1.0) (7)	75	2.5	80
13	Mes- <i>t</i> Bu ₂ Acr ⁺ BF ₄ ⁻ (0.5) (7)	60	3.5	85
14	Mes- <i>t</i> Bu ₂ Acr ⁺ BF ₄ ⁻ (1.5) (7)	60	2.5	92

^aConditions: **1a** (0.20 mmol), IPA (1.1–75 equivalent), and a photocatalyst (0.5–1.5 mol%) in degassed solvent (2.9 mL, 0.07 M).

^bUnder the irradiation of green LEDs.

General Procedure for the Optimisation Studies (Table 1)

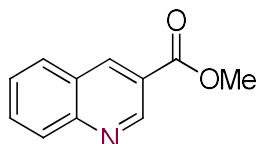
In a 10-mL test tube, equipped with a small magnetic stir bar, methyl quinoline-3-carboxylate *N*-oxide (**1a**, 40.6 mg, 0.200 mmol, 1.00 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 1.7 mg, 3.0 μ mol, 1.5 mol%), and isopropanol (IPA, 0.92 mL) were combined. The test tube was fitted with a rubber septum, and acetone (2.90 mL, 0.07 M) was added via a syringe under a nitrogen atmosphere. After degassing by N₂ bubbling at room temperature for 5 minutes, the reaction mixture was irradiated with two 3 W blue LEDs at room temperature for the indicated time (see Table 1). Upon completion of the reaction as determined by TLC, the reaction mixture was concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography on silica gel (hexanes/EtOAc = 3:1) to afford methyl quinoline-3-carboxylate (**2a**) as a white solid.

General Procedure for the Organic Photoredox-Catalysed Deoxygenation Reaction of N-Heterocyclic N-Oxide Using Isopropyl Alcohol as a Sustainable Reducing Agent (Table 2)

In a 10-mL test tube, equipped with a small magnetic stir bar, N-heterocyclic N-oxide (**1**, 0.200 mmol, 1.00 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 1.7 mg, 3.0 μ mol, 1.5 mol%), and IPA (0.92 mL) were combined. The test tube was fitted with a rubber septum, and acetone (2.9 mL, 0.07 M) was added via a syringe under a nitrogen atmosphere. After degassing by N₂ bubbling at room temperature for 5 minutes, the reaction mixture was irradiated with two 3 W blue LEDs at room temperature for the indicated time (Table 2). Upon completion of the reaction as determined by TLC, the reaction mixture was concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography on silica gel (hexanes and EtOAc) to afford the corresponding deoxygenated N-heterocycle (**2**). For N-heterocyclic N-oxides **1n**, **1o**, **1p**, **1t**, **1u**, **1v**, **1w**, **1x** and **1ao**, a photoreactor equipped with four 3 W blue LEDs were used under otherwise identical conditions.

Characterisation Data for the Deoxygenated N-Heterocycles (**2**)

Preparation of Methyl Quinoline-3-carboxylate (**2a**)

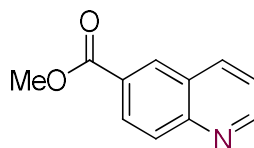


Prepared using methyl quinoline-3-carboxylate N-oxide (**1a**, 40.6 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). Methyl quinoline-3-carboxylate (**2a**, 34.5 mg, 92%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 9.4 (m, 1H), 8.8 (d, *J* = 2.3 Hz, 1H), 8.2 (d, *J* = 8.4 Hz, 1H), 7.9 (d, *J* = 8.2 Hz, 1H), 7.8 (t, *J* = 7.3 Hz, 1H), 7.6 (t, *J* = 7.5 Hz, 1H), 4.0 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 165.6, 149.6, 149.3, 139.0, 132.0, 129.1, 129.0, 127.5, 126.8, 122.9, 52.4.

GC-MS (EI, 70 eV) *m/z* calculated for [M]⁺ (C₁₁H₉NO₂) 187.1, found 187.1.

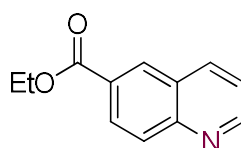
Preparation of Methyl Quinoline-6-carboxylate (2b)

Prepared using methyl quinoline-6-carboxylate *N*-oxide (**1b**, 40.6 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). Methyl quinoline-6-carboxylate (**2b**, 30.1 mg, 80%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.9 (dd, $J = 4.3, 1.8$ Hz, 1H), 8.5 (d, $J = 2.0$ Hz, 1H), 8.2 (dd, $J = 8.9, 2.0$ Hz, 1H), 8.2 (dd, $J = 8.2, 0.9$ Hz, 1H), 8.1 (d, $J = 8.7$ Hz, 1H), 7.4 (dd, $J = 8.2, 4.3$ Hz, 1H), 3.9 (s, 3H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 166.5, 152.5, 150.1, 137.3, 131.0, 129.8, 128.9, 128.1, 127.4, 121.8, 52.4.

GC-MS (EI, 70 eV) m/z calculated for $[\text{M}]^+$ ($\text{C}_{11}\text{H}_9\text{NO}_2$) 187.1, found 187.1.

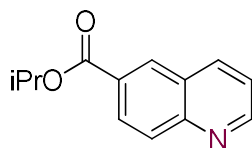
Preparation of Ethyl Quinoline-6-carboxylate (2c)

Prepared using ethyl quinoline-6-carboxylate *N*-oxide (**1c**, 43.4 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). Ethyl quinoline-6-carboxylate (**2c**, 32.7 mg, 81%) was isolated (hexanes/EtOAc = 1.5:1) as a pale-yellow oil.

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 9.0 (dd, $J = 4.3, 1.8$ Hz, 1H), 8.6 (d, $J = 2.1$ Hz, 1H), 8.3 (dd, $J = 8.9, 2.0$ Hz, 1H), 8.3 (dd, $J = 8.3, 1.9$ Hz, 1H), 8.1 (d, $J = 8.9$ Hz, 1H), 7.5 (dd, $J = 8.3, 4.2$ Hz, 1H), 4.5 (q, $J = 7.1$ Hz, 2H), 1.5 (t, $J = 7.1$ Hz, 3H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 166.1, 152.4, 150.0, 137.3, 130.9, 129.7, 129.0, 128.5, 127.4, 121.8, 61.4, 14.4.

GC-MS (EI, 70 eV) m/z calculated for $[\text{M}]^+$ ($\text{C}_{12}\text{H}_{11}\text{NO}_2$) 201.1, found 201.1.

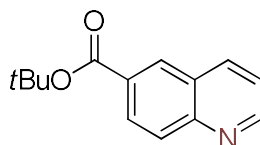
Preparation of Isopropyl Quinoline-6-carboxylate (2d)

Prepared using isopropyl quinoline-6-carboxylate *N*-oxide (**1d**, 46.2 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). Isopropyl quinoline-6-carboxylate (**2d**, 33.2 mg, 74%) was isolated (hexanes/EtOAc = 1.5:1) as a brown solid.

¹H NMR (500 MHz, CDCl₃) δ 9.0 (dd, J = 4.2, 1.8 Hz, 1H), 8.6 (d, J = 2.0 Hz, 1H), 8.3 (dd, J = 8.8, 1.9 Hz, 1H), 8.2 (dd, J = 8.2, 2.4 Hz, 1H), 8.1 (d, J = 8.9 Hz, 1H), 5.3 (hept, J = 6.3 Hz, 1H), 1.4 (d, J = 6.3 Hz, 6H).

¹³C NMR (126 MHz, CDCl₃) δ 165.7, 152.5, 150.1, 137.4, 130.9, 129.8, 129.1, 129.0, 127.5, 121.9, 69.0, 22.1.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₃H₁₃NO₂) 215.1, found 215.2.

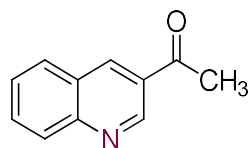
Preparation of *tert*-Butyl Quinoline-6-carboxylate (2e)

Prepared using *tert*-butyl quinoline-6-carboxylate (**1e**, 49.1 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). *tert*-Butyl quinoline-6-carboxylate (**2e**, 36.7 mg, 80%) was isolated (hexanes/EtOAc = 2:1) as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.97 (dd, J = 4.2, 1.8 Hz, 1H), 8.50 (d, J = 2.0 Hz, 1H), 8.24 (ddd, J = 9.2, 4.1, 2.2 Hz, 2H), 8.10 (d, J = 8.8 Hz, 1H), 7.43 (dd, J = 8.3, 4.3 Hz, 1H), 1.63 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 165.3, 152.3, 150.0, 137.4, 130.7, 130.1, 129.6, 129.2, 127.5, 121.8, 81.7, 28.3.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₄H₁₅NO₂) 229.1, found 229.2.

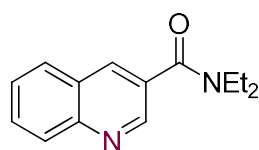
Preparation of 3-Acetylquinoline (2f)

Prepared using 3-acetylquinoline *N*-oxide (**1f**, 37.4 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 3-Acetylquinoline (**2f**, 29.9 mg, 87%) was isolated (hexanes/EtOAc = 1:1) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 9.4 (d, J = 2.0 Hz, 1H), 8.7 (d, J = 2.4 Hz, 1H), 8.1 (d, J = 8.4 Hz, 1H), 7.9 (d, J = 8.2 Hz, 1H), 7.8 (ddd, J = 8.4, 6.8, 1.4 Hz, 1H), 7.6 (t, J = 7.5 Hz, 1H), 2.7 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 196.8, 149.8, 149.2, 137.5, 132.2, 129.5, 129.5, 129.3, 127.7, 126.9, 26.9.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₁H₉NO) 171.1, found 171.1.

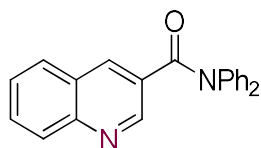
Preparation of *N,N*-Diethyl-3-quinolinecarboxamide (2g)

Prepared using *N,N*-diethyl-3-quinolinecarboxamide *N*-oxide (**1g**, 48.9 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). *N,N*-Diethyl-3-quinolinecarboxamide (**2g**, 41.1 mg, 89%) was isolated (hexanes/EtOAc = 1:3) as a yellow oil.

¹H NMR (500 MHz, CDCl₃) δ 8.90 (s, 1H), 8.16 (s, 1H), 8.09 (d, J = 8.5 Hz, 1H), 7.81 (d, J = 8.2 Hz, 1H), 7.76–7.69 (m, 1H), 7.59–7.51 (m, 1H), 3.57 (br s, 2H), 3.29 (br s, 2H), 1.24 (br s, J = 26.9 Hz, 3H), 1.12 (br s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 168.7, 148.1, 148.0, 134.1, 130.5, 130.2, 129.4, 128.2, 127.4, 127.1, 43.6, 39.7, 14.4, 13.0.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₄H₁₆N₂O) 228.1, found 228.1.

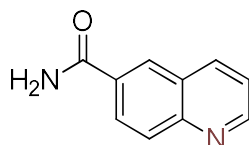
Preparation of *N,N*-Diphenyl-3-quinolinecarboxamide (2h)

Prepared using *N,N*-diphenyl-3-quinolinecarboxamide *N*-oxide (**1h**, 68.1 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). *N,N*-Diphenyl-3-quinolinecarboxamide (**2h**, 62.9 mg, 97%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

^1H NMR (500 MHz, CDCl_3) δ 8.89 (d, J = 2.2 Hz, 1H), 8.37 (d, J = 2.2 Hz, 1H), 8.04 (d, J = 8.4 Hz, 1H), 7.78–7.70 (m, 3H), 7.53 (t, J = 7.5 Hz, 1H), 7.32 (t, J = 7.7 Hz, 5H), 7.27–7.21 (m, 8H).

^{13}C NMR (126 MHz, CDCl_3) δ 168.2, 149.6, 148.1, 143.3, 137.8, 131.0, 129.4, 129.3, 129.2, 128.6, 127.5, 127.3, 127.0, 126.8.

GC-MS (EI, 70 eV) m/z calculated for $[\text{M}]^+$ ($\text{C}_{22}\text{H}_{16}\text{N}_2\text{O}$) 324.1, found 324.1.

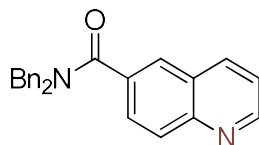
Preparation of 6-Quinolinecarboxamide (2i)

Prepared using 6-quinolinecarboxamide *N*-oxide (**1i**, 37.6 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 6-Quinolinecarboxamide (**2i**, 28.1 mg, 82%) was isolated (hexanes/acetone = 1:3) as a white solid.

^1H NMR (300 MHz, MeOD) δ 8.87 (dd, J = 4.4, 1.7 Hz, 1H), 8.41 (dd, J = 17.2, 1.9 Hz, 2H), 8.16 (dd, J = 8.9, 2.1 Hz, 2H), 8.02 (d, J = 8.9 Hz, 1H), 7.54 (dd, J = 8.3, 4.3 Hz, 2H).

^{13}C NMR (75 MHz, MeOD) δ 171.3, 153.0, 149.9, 139.4, 133.4, 129.7, 129.5, 129.3, 129.0, 123.3.

GC-MS (EI, 70 eV) m/z calculated for $[\text{M}]^+$ ($\text{C}_{10}\text{H}_8\text{N}_2\text{O}$) 172.1, found 172.1.

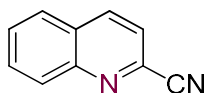
Preparation of *N,N*-Dibenzyl-6-quinolinecarboxamide (2j)

Prepared using *N,N*-dibenzyl-6-quinolinecarboxamide *N*-oxide (**1j**, 73.7 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). *N,N*-Dibenzyl-6-quinolinecarboxamide (**2j**, 63.5 mg, 90%) was isolated (hexanes/EtOAc = 1:3) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.93 (dd, J = 4.2, 1.8 Hz, 1H), 8.12 (d, J = 8.4 Hz, 2), 7.97 (d, J = 2.0 Hz, 1H), 7.82 (dd, J = 8.6, 1.9 Hz, 1H), 7.44–7.29 (m, 9H), 7.15 (d, J = 7.9 Hz, 2H), 4.77 (s, 2H), 4.45 (s, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 171.6, 151.6, 148.4, 136.8, 136.5, 136.3, 134.4, 130.1, 128.9, 128.6, 127.8, 127.7, 127.0, 126.6, 121.9, 51.7, 47.2.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₂₄H₂₀N₂O) 352.2, found 352.2.

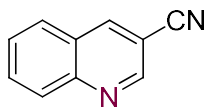
Preparation of 2-Quinolinecarbonitrile (2k)

Prepared using 2-quinolinecarbonitrile *N*-oxide (**1k**, 34.0 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.30 mL), and acetone (2.9 mL, 0.07 M). 2-Quinolinecarbonitrile (**2k**, 28.4 mg, 90%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.33 (d, J = 8.4 Hz, 1H), 8.18 (d, J = 8.5 Hz, 1H), 7.92 (dd, J = 8.2, 1.4 Hz, 1H), 7.86 (ddd, J = 8.5, 6.9, 1.5 Hz, 1H), 7.73 (t, J = 7.6 Hz, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 148.3, 137.6, 133.7, 131.4, 130.1, 129.6, 128.8, 127.9, 123.4, 117.7.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₀H₆N₂) 154.1, found 154.1.

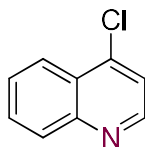
Preparation of 3-Quinolinecarbonitrile (2l)

Prepared using 3-quinolinecarbonitrile *N*-oxide (**1l**, 34.0 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.30 mL), and acetone (2.9 mL, 0.07 M). 3-Quinolinecarbonitrile (**2l**, 24.1 mg, 78%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 9.02 (d, $J = 2.1$ Hz, 1H), 8.52 (dd, $J = 2.2, 0.8$ Hz, 1H), 8.16 (d, $J = 9.0$ Hz, 1H), 7.93–7.84 (m, 2H), 7.68 (ddd, $J = 8.1, 6.9, 1.2$ Hz, 1H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 149.8, 148.8, 141.6, 132.9, 129.9, 128.6, 128.3, 126.3, 117.2, 106.6.

GC-MS (EI, 70 eV) m/z calculated for $[\text{M}]^+$ ($\text{C}_{10}\text{H}_6\text{N}_2$) 154.1, found 154.1.

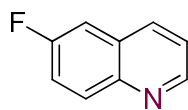
Preparation of 4-Chloroquinoline (2m)

Prepared using 4-chloroquinoline *N*-oxide (**1m**, 35.9 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 4-Chloroquinoline (**2m**, 24.9 mg, 76%) was isolated (hexanes/EtOAc = 10:1) as a white solid.

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.78 (d, $J = 4.7$ Hz, 1H), 8.22 (dd, $J = 8.4, 1.5$ Hz, 1H), 8.13 (dd, $J = 8.5, 0.9$ Hz, 1H), 7.77 (ddd, $J = 8.4, 6.8, 1.5$ Hz, 1H), 7.64 (ddd, $J = 8.4, 6.8, 1.3$ Hz, 1H), 7.48 (d, $J = 4.7$ Hz, 1H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 149.9, 149.1, 142.9, 130.6, 129.9, 127.8, 126.6, 124.3, 121.4.

GC-MS (EI, 70 eV) m/z calculated for $[\text{M}]^+$ ($\text{C}_9\text{H}_6\text{ClN}$) 163.0, found 163.1.

Preparation of 6-Fluoroquinoline (2n)

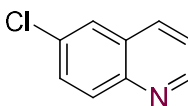
Prepared using 6-fluoroquinoline *N*-oxide (**1n**, 32.6 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 6-Fluoroquinoline (**2n**, 20.9 mg, 71%) was isolated (hexanes/EtOAc = 2:1) as a yellow oil.

^1H NMR (500 MHz, CDCl_3) δ 8.88 (dd, J = 4.4, 1.4 Hz, 1H), 8.11 (dd, J = 9.5, 5.3 Hz, 2H), 7.48 (td, J = 8.7, 2.7 Hz, 1H), 7.42 (ddd, J = 8.5, 5.3, 3.4 Hz, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 160.5 (d, $^1J_{\text{C-F}}$ = 248.0 Hz), 149.8 (d, $^4J_{\text{C-F}}$ = 2.9 Hz), 145.5, 135.5 (d, $^4J_{\text{C-F}}$ = 5.5 Hz), 132.1 (d, $^3J_{\text{C-F}}$ = 9.2 Hz), 129.0 (d, $^3J_{\text{C-F}}$ = 9.9 Hz), 121.9, 119.9 (d, $^2J_{\text{C-F}}$ = 25.7 Hz), 110.8 (d, $^2J_{\text{C-F}}$ = 21.6 Hz).

^{19}F NMR (376 MHz, CDCl_3) δ -113.3.

GC-MS (EI, 70 eV) m/z calculated for $[\text{M}]^+$ ($\text{C}_9\text{H}_6\text{FN}$) 147.0, found 147.1.

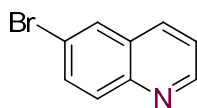
Preparation of 6-Chloroquinoline (2o)

Prepared using 6-chloroquinoline *N*-oxide (**1o**, 35.9 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 6-Chloroquinoline (**2o**, 24.5 mg, 75%) was isolated (hexanes/EtOAc = 3:1) as a yellow oil.

^1H NMR (500 MHz, CDCl_3) δ 8.90 (dd, J = 4.3, 1.8 Hz, 1H), 8.05 (t, J = 9.9 Hz, 1H), 7.79 (d, J = 2.3 Hz, 1H), 7.64 (dd, J = 8.9, 2.4 Hz, 1H), 7.41 (dd, J = 8.4, 4.3 Hz, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 150.6, 146.6, 135.4, 132.5, 131.1, 130.6, 128.9, 126.5, 122.0.

GC-MS (EI, 70 eV) m/z calculated for $[\text{M}]^+$ ($\text{C}_9\text{H}_6\text{ClN}$) 163.0, found 163.1.

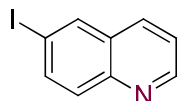
Preparation of 6-Bromoquinoline (2p)

Prepared using 6-bromoquinoline *N*-oxide (**1p**, 44.8 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 6-Bromoquinoline (**2p**, 31.2 mg, 75%) was isolated (hexanes/EtOAc = 3:1) as a yellow oil.

¹H NMR (500 MHz, CDCl₃) δ 8.91 (dd, J = 4.3, 1.7 Hz, 1H), 8.06 (dd, J = 8.3, 1.9 Hz, 1H), 8.00–7.95 (m, 2H), 7.77 (dd, J = 9.0, 2.3 Hz, 1H), 7.41 (dd, J = 8.3, 4.2 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 150.7, 146.8, 135.3, 133.1, 131.2, 129.9, 129.5, 122.0, 120.6.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₉H₆BrN) 207.0, found 207.1.

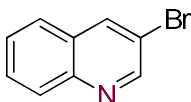
Preparation of 6-Iodoquinoline (2q)

Prepared using 6-iodoquinoline *N*-oxide (**1q**, 54.2 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 6-Iodoquinoline (**2q**, 38.1 mg, 75%) was isolated (hexanes/EtOAc = 10:1) as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.89 (d, J = 4.5 Hz, 1H), 8.17 (d, J = 2.3 Hz, 1H), 7.99 (d, J = 8.3 Hz, 1H), 7.95–7.88 (m, 3H), 7.81 (d, J = 8.9 Hz, 1H), 7.37 (dd, J = 8.3, 4.2 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 151.0, 147.3, 138.2, 136.6, 134.9, 131.3, 130.0, 121.8, 92.2.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₉H₆I₁N) 255.0, found 254.9.

Preparation of 3-Bromoquinoline (2r)

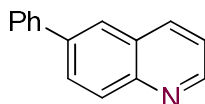
Prepared using 3-bromoquinoline *N*-oxide (**1r**, 44.8 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 3-Bromoquinoline (**2r**, 34.2 mg, 82%) was isolated (hexanes/EtOAc = 10:1) as a yellow oil.

¹H NMR (500 MHz, CDCl₃) δ 8.90 (d, *J* = 2.4 Hz, 1H), 8.30 (d, *J* = 2.4 Hz, 1H), 8.08 (d, *J* = 9.0 Hz, 1H), 7.72 (td, *J* = 6.1, 5.5, 2.9 Hz, 2H), 7.60–7.53 (m, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 151.4, 146.3, 137.4, 129.9, 129.5, 129.2, 127.7, 127.0, 117.2.

GC-MS (EI, 70 eV) *m/z* calculated for [M]⁺ (C₉H₆BrN) 207.0, found 207.1.

Preparation of 6-Phenylquinoline (2s)



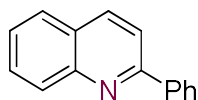
Prepared using 6-phenylquinoline *N*-oxide (**1s**, 44.2 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μmol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 6-Phenylquinoline (**2s**, 37.9 mg, 92%) was isolated (hexanes/EtOAc = 10:1) as a yellow oil.

¹H NMR (500 MHz, CDCl₃) δ 9.20 (s, 1H), 8.30 (s, 1H), 8.16 (d, *J* = 8.5 Hz, 1H), 7.87 (d, *J* = 8.1 Hz, 1H), 7.71 (d, *J* = 7.9 Hz, 3H), 7.61–7.48 (m, 3H), 7.44 (t, *J* = 7.4 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 149.9, 147.2, 137.8, 133.9, 133.3, 129.5, 129.2, 129.2, 128.2, 128.1, 128.0, 127.4, 127.1.

GC-MS (EI, 70 eV) *m/z* calculated for [M]⁺ (C₁₅H₁₁N) 205.1, found 205.1.

Preparation of 2-Phenylquinoline (2t)

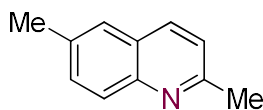


Prepared using 2-phenylquinoline *N*-oxide (**1t**, 44.2 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μmol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 2-Phenylquinoline (**2t**, 37.6 mg, 92%) was isolated (hexanes/DCM = 2.5:1) as a yellow oil.

¹H NMR (500 MHz, CDCl₃) δ 8.22 (s, 1H), 8.21–8.17 (m, 7H), 7.88 (d, *J* = 8.5 Hz, 2H), 7.83 (dd, *J* = 8.1, 1.5 Hz, 2H), 7.74 (ddd, *J* = 8.4, 6.9, 1.6 Hz, 2H), 7.58–7.51 (m, 6H), 7.51–7.44 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 157.5, 148.4, 139.8, 136.9, 129.9, 129.8, 129.4, 129.0, 127.7, 127.6, 127.3, 126.4, 119.1.

GC-MS (EI, 70 eV) *m/z* calculated for [M]⁺ (C₁₅H₁₁N) 205.1, found 205.1.

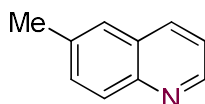
Preparation of 2,6-Dimethylquinoline (2u)

Prepared using 2,6-dimethylquinoline *N*-oxide (**1u**, 34.6 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 2,6-Dimethylquinoline (**2u**, 24.0 mg, 76%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 7.93 (dd, J = 12.8, 8.4 Hz, 2H), 7.54–7.46 (m, 2H), 7.23 (d, J = 8.4 Hz, 1H), 2.72 (s, 3H), 2.50 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 158.0, 146.4, 135.8, 135.6, 131.8, 128.3, 126.6, 126.5, 122.1, 25.3, 21.6.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₁H₁₁N) 157.1, found 157.2.

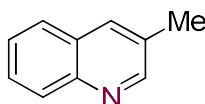
Preparation of 6-Methylquinoline (2v)

Prepared using 6-methylquinoline *N*-oxide (**1v**, 31.8 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 6-Methylquinoline (**2v**, 22.3 mg, 78%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.59 (d, J = 8.9 Hz, 1H), 8.41 (dd, J = 6.0, 1.1 Hz, 1H), 7.60 (d, J = 8.4 Hz, 1H), 7.57 (s, 1H), 7.53 (dd, J = 8.9, 2.0 Hz, 1H), 7.20 (dd, J = 8.5, 6.0 Hz, 1H), 2.49 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 140.1, 139.0, 134.9, 132.6, 130.6, 127.0, 125.4, 120.9, 119.5, 21.4.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₀H₉N) 143.1, found 143.2.

Preparation of 3-Methylquinoline (2w)

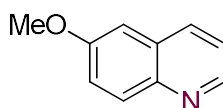
Prepared using 3-methylquinoline *N*-oxide (**1w**, 31.8 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 3-Methylquinoline (**2w**, 21.4 mg, 75%) was isolated (hexanes/EtOAc = 5:1) as a yellow oil.

¹H NMR (500 MHz, CDCl₃) δ 8.76 (d, *J* = 2.4 Hz, 1H), 8.07 (d, *J* = 8.5 Hz, 1H), 7.92–7.88 (m, 1H), 7.73 (dd, *J* = 8.2, 1.5 Hz, 1H), 7.64 (ddd, *J* = 8.4, 6.8, 1.6 Hz, 1H), 7.50 (ddd, *J* = 8.2, 6.8, 1.3 Hz, 1H), 2.50 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 152.4, 146.5, 134.9, 130.6, 129.2, 128.6, 128.2, 127.2, 126.7, 18.9.

GC-MS (EI, 70 eV) *m/z* calculated for [M]⁺ (C₁₀H₉N) 143.1, found 143.2.

Preparation of 6-Methoxyquinoline (2x)



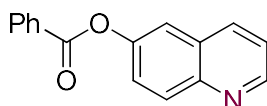
Prepared using 6-methoxyquinoline *N*-oxide (**1x**, 35.0 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μmol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 6-Methoxyquinoline (**2x**, 25.1 mg, 79%) was isolated (hexanes/EtOAc = 10:1) as a yellow oil.

¹H NMR (500 MHz, CDCl₃) δ 8.76 (dd, *J* = 4.4, 1.5 Hz, 1H), 8.09–7.98 (m, 2H), 7.36 (ddd, *J* = 11.3, 8.8, 3.6 Hz, 2H), 7.06 (d, *J* = 2.7 Hz, 1H), 3.92 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 157.9, 147.8, 144.3, 135.1, 130.8, 129.4, 122.5, 121.5, 105.2, 55.7.

GC-MS (EI, 70 eV) *m/z* calculated for [M]⁺ (C₁₀H₉NO) 159.1, found 159.1.

Preparation of 6-Benzoyloxyquinoline (2y)

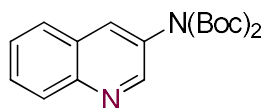


Prepared using 6-benzoyloxyquinoline *N*-oxide (**1y**, 53.0 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μmol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 6-Benzoyloxyquinoline (**2y**, 37.6 mg, 76%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.94 (dd, *J* = 4.3, 1.8 Hz, 1H), 8.27 (m, *J* = 7.0, 1.4 Hz, 2H), 8.20 (d, *J* = 9.0 Hz, 1H), 8.15 (dd, *J* = 8.3, 2.1 Hz, 1H), 7.71 (d, *J* = 2.6 Hz, 1H), 7.67 (tt, *J* = 7.0, 1.4 Hz, 1H), 7.61 (dd, *J* = 9.2, 2.6 Hz, 1H), 7.55 (t, *J* = 7.9 Hz, 2H), 7.43 (dd, *J* = 8.2, 4.3 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 165.2, 150.3, 148.9, 146.5, 135.9, 133.9, 131.2, 130.3, 129.4, 128.8, 128.7, 124.9, 121.7, 118.7.

GC-MS (EI, 70 eV) *m/z* calculated for [M]⁺ (C₁₆H₁₁NO₂) 249.1, found 249.1.

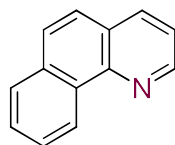
Preparation of 3-[Bis(*tert*-butoxycarbonyl)amino]quinoline (2z)

Prepared using 3-[bis(*tert*-butoxycarbonyl)amino]quinoline *N*-oxide (**1z**, 72.1 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 3-[Bis(*tert*-butoxycarbonyl)amino]quinoline (**2z**, 63.7 mg, 93%) was isolated (hexanes/EtOAc = 4:1) as a yellow solid.

¹H NMR (500 MHz, CDCl₃) δ 8.68 (d, J = 2.4 Hz, 1H), 8.09 (d, J = 8.4 Hz, 1H), 7.91 (d, J = 2.4 Hz, 1H), 7.79 (dd, J = 7.5, 1.9 Hz, 1H), 7.70 (tt, J = 6.9, 1.7 Hz, 1H), 7.54 (tt, J = 6.9, 1.7 Hz, 1H), 1.39 (s, 18H).

¹³C NMR (126 MHz, CDCl₃) δ 151.3, 150.6, 146.8, 133.6, 132.9, 129.8, 129.3, 127.9, 127.8, 127.1, 83.6, 28.0.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₉H₂₄N₂O₄) 344.2, found 344.1.

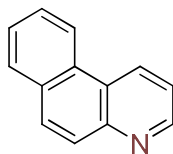
Preparation of Benzo[*h*]quinoline (2aa)

Prepared using benzo[*h*]quinoline *N*-oxide (**1aa**, 39.0 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). Benzo[*h*]quinoline (**2aa**, 29.1 mg, 81%) was isolated (hexanes/EtOAc = 5:1) as a pale yellow solid.

¹H NMR (500 MHz, CDCl₃) δ 9.32 (dd, J = 8.3, 1.1 Hz, 1H), 9.01 (dd, J = 4.4, 1.8 Hz, 1H), 8.15 (dd, J = 8.1, 1.8 Hz, 1H), 7.91 (dd, J = 7.9, 1.6 Hz, 1H), 7.85–7.64 (m, 4H), 7.51 (dd, J = 8.1, 4.4 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 148.8, 146.6, 135.8, 133.6, 131.5, 128.2, 127.8, 127.8, 127.1, 126.4, 125.4, 124.4, 121.8.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₃H₉N) 179.1, found 179.2.

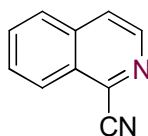
Preparation of Benzo[*f*]quinoline (2ab)

Prepared using benzo[*f*]quinoline *N*-oxide (**1ab**, 39.0 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). Benzo[*f*]quinoline (**2ab**, 25.9 mg, 72%) was isolated (hexanes/EtOAc = 5:1) as a pale yellow solid.

¹H NMR (400 MHz, CDCl₃) δ 8.96 (dd, J = 4.4, 1.6 Hz, 1H), 8.92 (dd, J = 8.4, 1.9 Hz, 1H), 8.60 (d, J = 7.9 Hz, 1H), 8.03–7.95 (m, 2H), 7.93 (dd, J = 7.4, 2.0 Hz, 1H), 7.73–7.60 (m, 2H), 7.54 (dd, J = 8.4, 4.4 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 149.8, 148.3, 131.8, 131.0, 130.8, 129.7, 128.8, 128.3, 127.4, 127.2, 125.5, 122.7, 121.4.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₃H₉N) 179.1, found 179.2.

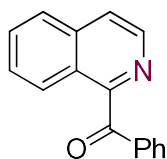
Preparation of 1-Isoquinolinecarbonitrile (2ac)

Prepared using 1-isoquinolinecarbonitrile *N*-oxide (**1ac**, 34.0 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 1-Isoquinolinecarbonitrile (**2ac**, 26.0 mg, 84%) was isolated (hexanes/acetone = 3:1) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.63 (d, J = 5.5 Hz, 1H), 8.33–8.28 (m, 1H), 7.94 (dd, J = 7.6, 1.5 Hz, 1H), 7.89 (d, J = 5.5 Hz, 1H), 7.80 (dddd, J = 18.6, 8.2, 6.9, 1.4 Hz, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 143.3, 135.9, 134.8, 131.8, 129.9, 129.3, 127.3, 125.3, 124.5, 115.8.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₀H₆N₂) 154.1, found 154.1.

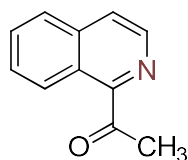
Preparation of 1-Benzoylisoquinoline (2ad)

Prepared using 1-benzoylisoquinoline *N*-oxide (**1ad**, 49.9 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 1-Benzoylisoquinoline (**2ad**, 44.4 mg, 84%) was isolated (hexanes/EtOAc = 5:1) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.60 (d, J = 5.6 Hz, 1H), 8.22 (dd, J = 8.5, 1.0 Hz, 1H), 8.00–7.95 (m, 2H), 7.91 (d, J = 8.2 Hz, 1H), 7.80 (dd, J = 5.6, 0.8 Hz, 1H), 7.73 (ddd, J = 8.2, 6.9, 1.4 Hz, 1H), 7.60 (dddd, J = 8.9, 5.6, 3.1, 1.4 Hz, 2H), 7.50–7.43 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 194.8, 156.5, 141.2, 136.8, 136.7, 133.7, 130.8, 130.8, 128.5, 128.4, 127.2, 126.5, 126.2, 122.7.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₆H₁₁NO) 233.1, found 233.1.

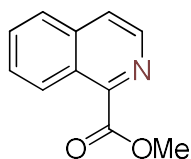
Preparation of 1-Acetylisquinoline (2ae)

Prepared using 1-acetylisquinoline *N*-oxide (**1ae**, 37.4 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 1-Acetylisquinoline (**2ae**, 27.2 mg, 80%) was isolated (hexanes/acetone = 3:1) as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.95 (d, J = 7.9 Hz, 1H), 8.57 (dd, J = 5.5, 1.9 Hz, 1H), 7.88–7.77 (m, 2H), 7.75–7.62 (m, 2H), 2.86 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 202.8, 152.9, 141.2, 137.2, 130.4, 129.2, 127.1, 127.0, 125.9, 124.7, 28.7.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₁H₉NO) 171.1, found 171.1.

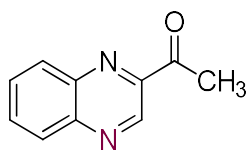
Preparation of Methyl 1-Isoquinolinecarboxylate (2af)

Prepared using Methyl 1-isoquinolinecarboxylate *N*-oxide (**1af**, 40.6 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). Methyl 1-isoquinolinecarboxylate (**2af**, 33.9 mg, 91%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.82 (d, *J* = 8.5 Hz, 1H), 8.61 (dd, *J* = 5.6, 1.6 Hz, 1H), 7.86 (d, *J* = 7.9 Hz, 1H), 7.80 (d, *J* = 5.8 Hz, 1H), 7.75–7.63 (m, 2H), 4.08 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 166.4, 148.3, 141.7, 137.0, 130.7, 128.9, 127.2, 127.0, 126.5, 124.4, 53.1.

GC-MS (EI, 70 eV) *m/z* calculated for [M]⁺ (C₁₁H₉NO₂) 187.1, found 187.1.

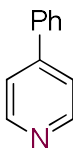
Preparation of Methyl 2-quinoxalinecarboxylate (2ag)

Prepared using methyl 2-quinoxalinecarboxylate *N*-oxide (**1ag**, 37.6 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). Methyl 2-quinoxalinecarboxylate (**2ag**, 25.0 mg, 73%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 9.47 (s, 1H), 8.16 (ddd, *J* = 13.1, 8.2, 1.8 Hz, 2H), 7.92–7.80 (m, 2H), 2.84 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 199.8, 146.6, 143.9, 143.0, 141.1, 132.2, 130.7, 130.5, 129.4, 25.5.

GC-MS (EI, 70 eV) *m/z* calculated for [M]⁺ (C₁₀H₈N₂O) 172.1, found 172.1.

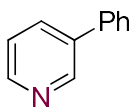
Preparation of 4-Phenylpyridine (2ah)

Prepared using 4-phenylpyridine *N*-oxide (**1ah**, 34.2 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 4-Phenylpyridine (**2ah**, 28.2 mg, 90%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.65 (dd, J = 4.4, 1.8 Hz, 2H), 7.67–7.59 (m, 2H), 7.55–7.38 (m, 5H).

¹³C NMR (126 MHz, CDCl₃) δ 150.3, 148.3, 138.1, 129.1, 129.1, 127.0, 121.6.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₁H₉N) 155.1, found 155.2.

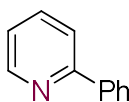
Preparation of 3-Phenylpyridine (2ai)

Prepared using 3-phenylpyridine *N*-oxide (**1ai**, 34.2 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 3-Phenylpyridine (**2ai**, 18.8 mg, 61%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 8.85 (dd, J = 2.4, 0.9 Hz, 1H), 8.59 (dd, J = 4.8, 1.6 Hz, 1H), 7.86 (ddd, J = 7.9, 2.4, 1.6 Hz, 1H), 7.61–7.53 (m, 2H), 7.47 (ddd, J = 7.5, 6.7, 1.3 Hz, 2H), 7.43–7.37 (m, 1H), 7.35 (ddd, J = 7.9, 4.8, 0.9 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 148.5, 148.4, 137.8, 136.6, 134.3, 129.1, 128.1, 127.2, 123.5.

GC-MS (EI, 70 eV) m/z calculated for [M]⁺ (C₁₁H₉N) 155.1, found 155.2.

Preparation of 2-Phenylpyridine (2aj)

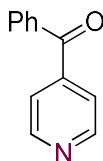
Prepared using 2-phenylpyridine *N*-oxide (**1aj**, 34.2 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 2-Phenylpyridine (**2aj**, 23.0 mg, 74%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.70 (dt, *J* = 4.9, 1.5 Hz, 1H), 8.02–7.96 (m, 2H), 7.79–7.70 (m, 2H), 7.48 (td, *J* = 6.6, 6.1, 1.7 Hz, 1H), 7.45–7.39 (m, 1H), 7.23 (td, *J* = 4.7, 2.3 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 157.6, 149.8, 139.5, 136.9, 129.1, 128.9, 127.0, 122.2, 120.7.

GC-MS (EI, 70 eV) *m/z* calculated for [M]⁺ (C₁₁H₉N) 155.1, found 155.2.

Preparation of 4-Benzoylpyridine (2ak)



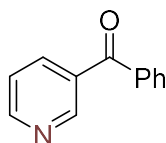
Prepared using 4-benzoylpyridine *N*-oxide (**1ak**, 39.8 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μmol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 4-Benzoylpyridine (**2ak**, 33.0 mg, 74%) was isolated (hexanes/EtOAc = 3:1) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.83–8.75 (m, 2H), 7.83–7.75 (m, 2H), 7.62 (tt, *J* = 7.0, 1.4 Hz, 1H), 7.59–7.53 (m, 2H), 7.53–7.45 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 195.2, 150.4 (two carbons), 144.4, 136.0, 133.6, 130.2 (two carbons), 128.7 (two carbons), 122.9 (two carbons).

GC-MS (EI, 70 eV) *m/z* calculated for [M]⁺ (C₁₂H₉NO) 183.1, found 183.1.

Preparation of 3-Benzoylpyridine (2al)



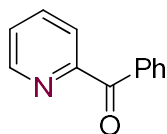
Prepared using 3-benzoylpyridine *N*-oxide (**1al**, 39.8 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μmol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 3-Benzoylpyridine (**2al**, 19.9 mg, 54 %) was isolated (hexanes/EtOAc = 3:1) as a brown solid.

¹H NMR (500 MHz, CDCl₃) δ 8.98 (dd, *J* = 2.2, 1.0 Hz, 1H), 8.80 (dd, *J* = 4.9, 1.8 Hz, 1H), 8.11 (dt, *J* = 7.8, 1.9 Hz, 1H), 7.85–7.78 (m, 2H), 7.68–7.59 (m, 1H), 7.56–7.48 (m, 2H), 7.44 (ddd, *J* = 7.9, 4.9, 1.0 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 195.0, 152.9, 151.1, 137.3, 136.8, 133.3, 130.1, 128.7, 123.5.

GC-MS (EI, 70 eV) m/z calculated for $[M]^+$ ($C_{12}H_9NO$) 183.1, found 183.1.

Preparation of 2-Benzoylpyridine (**2am**)



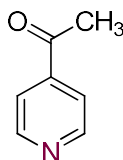
Prepared using 2-benzoylpyridine *N*-oxide (**1am**, 39.8 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 2-Benzoylpyridine (**2am**, 30.3 mg, 74%) was isolated (hexanes/DCM = 5:4) as a yellow solid.

1H NMR (500 MHz, $CDCl_3$) δ 8.72 (d, J = 4.9 Hz, 1H), 8.10–8.01 (m, 3H), 7.89 (tt, J = 7.8, 2.4 Hz, 1H), 7.59 (td, J = 7.3, 1.2 Hz, 1H), 7.53–7.42 (m, 3H).

^{13}C NMR (126 MHz, $CDCl_3$) δ 193.9, 155.1, 148.6, 137.1, 136.3, 132.9, 131.0 (two carbons), 128.2 (two carbons), 126.2, 124.6.

GC-MS (EI, 70 eV) m/z calculated for $[M]^+$ ($C_{12}H_9NO$) 183.1, found 183.1.

Preparation of 4-Acetylpyridine (**2an**)

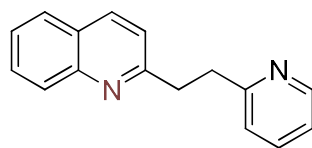


Prepared using 4-acetylpyridine *N*-oxide (**1an**, 27.4 mg, 0.200 mmol, 1.0 equivalent), **7** (1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.92 mL), and acetone (2.9 mL, 0.07 M). 4-Acetylpyridine (**2an**, 19.3 mg, 80%) was isolated (hexanes/EtOAc = 2:1) as a brown oil.

1H NMR (500 MHz, $CDCl_3$) δ 8.78 (d, J = 6.3 Hz, 2H), 7.70 (d, J = 6.3 Hz, 2H), 2.60 (s, 3H).

^{13}C NMR (126 MHz, $CDCl_3$) δ 197.4, 151.0 (two carbons), 142.8, 121.3 (two carbons), 26.7.

GC-MS (EI, 70 eV) m/z calculated for $[M]^+$ (C_7H_7NO) 121.1, found 121.1.

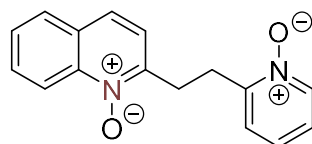
Preparation of 2-{2-(*N*-Oxidopyridine-2-yl)ethyl}quinoline (1ao')

This compound was prepared following a literature procedure.¹³ In a 60-mL thick-walled pressure vessel containing a magnetic stir bar, quinaldine (430 mg, 3.00 mmol, 1.0 equivalent), 2-pyridinemethanol (655 mg, 6.00 mmol, 2.0 equivalent), cesium hydroxide monohydrate (1.51 g, 9.00 mmol, 3.0 equivalent), and xylenes (3.0 mL) was combined in the air. The vessel was then fitted with a PTFE cap and immersed into a pre-heated oil bath (bath temperature = 160 °C). After stirring for 3 days, while heating the reaction mixture, the vessel was occasionally open to the air. The reaction mixture was then allowed to cool to room temperature. The reaction mixture was dissolved in a mixture of MeOH and CH₂Cl₂ (1:1, v/v) and poured into water. The mixture was extracted with CH₂Cl₂ (3 × 10 mL), and the combined organic layer was washed with brine (15 mL), dried over anhydrous sodium sulfate, filtered and concentrated on a rotary evaporator. Repetitive purification of the resulting residue by flash column chromatography (hexanes/EtOAc = 3:1 to neat EtOAc to hexanes/acetone = 9:1 to 15:1) on silica gel gave the title compound **1ao'** (91 mg, 13%) as a brown solid.

¹H NMR (400 MHz, CDCl₃) δ 8.61–8.52 (m, 1H), 8.06 (dd, *J* = 12.2, 8.5 Hz, 2H), 7.78 (dd, *J* = 8.1, 1.7 Hz, 1H), 7.70 (ddd, *J* = 8.4, 6.9, 1.6 Hz, 1H), 7.60–7.45 (m, 2H), 7.34–7.26 (m, 1H), 7.21–7.08 (m, 2H), 3.50–3.41 (m, 2H), 3.41–3.31 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 161.8, 161.2, 149.4, 148.1, 136.4, 136.3, 129.5, 129.0, 127.6, 126.9, 125.9, 123.2, 121.7, 121.3, 38.9, 38.1.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₆H₁₅N₂) 235.1, found 235.1.

Preparation of 2-{2-(*N*-Oxidopyridine-2-yl)ethyl}quinoline *N*-oxide (1ao)

This compound was prepared following a literature procedure.¹³ To a stirred solution of diheteroarylethane **1ao'** (342 mg, 0.86 mmol, 1.0 equivalent) in CH₂Cl₂ (2.3 mL) at 0 °C (ice-water bath),

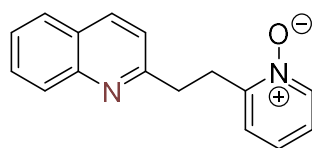
a solution of *m*-CPBA (>99%, 504 mg, 2.92 mmol, 2.0 equivalent) in CH₂Cl₂ (3.5 mL) was added dropwise via cannula over 30 minutes. The reaction mixture was then allowed to slowly warm to room temperature over 30 minutes. After stirring for 2 hours at the same temperature, another solution of *m*-CPBA (>99%, 252 mg, 1.46 mmol, 1.0 equivalent) in CH₂Cl₂ (1.5 mL) was added dropwise to the reaction mixture via cannula. The reaction mixture was then heated under reflux for 24 hours. After cooling to room temperature, the reaction mixture was poured into water and extracted with CH₂Cl₂ (3 × 10 mL). The combined organic layer was successively washed with a saturated solution of NaHCO₃ (15 mL), water (15 mL), and brine (15 mL). It was dried over anhydrous Na₂SO₄, filtered, and concentrated on a rotary evaporator. The crude mixture obtained was purified by flash column chromatography (EtOAc/MeOH = 4:1 to 2:1) on silica gel, yielding the title compound (**1ao**, 275 mg, 70%) as a brown solid.

¹H NMR (400 MHz, CDCl₃) δ 8.74 (d, *J* = 8.8 Hz, 1H), 8.24 (dd, *J* = 6.0, 1.8 Hz, 1H), 7.82–7.67 (m, 2H), 7.57 (ddd, *J* = 8.2, 6.6, 5.3 Hz, 2H), 7.33–7.20 (m, 2H), 7.17–7.05 (m, 2H), 3.61–3.51 (m, 2H), 3.51–3.40 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 150.9, 147.4, 141.6, 139.6, 130.4, 129.3, 128.1, 128.0, 126.3, 125.8, 125.3, 124.0, 122.4, 119.5, 28.3, 27.2.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₆H₁₅N₂O₂) 267.1, found 267.1.

Preparation of 2-{2-(2-Quinoliny)ethyl}pyridine *N*-oxide (**2ao**)



In a 10-mL test tube, equipped with a small magnetic stir bar, 2-{2-(2-quinoliny)ethyl}pyridine *N*-oxide (**1ao**, 53.3 mg, 0.200 mmol, 1.0 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 1.7 mg, 3.0 μmol, 1.5 mol%), and IPA (0.92 mL) were combined. The test tube was fitted with a rubber septum, and acetone (2.9 mL, 0.07 M) was added via a syringe under a nitrogen atmosphere. After degassing by N₂ bubbling at room temperature for 5 minutes, the reaction mixture was irradiated with two 3 W blue LEDs at room temperature for 51 hours. Afterward, the reaction mixture was concentrated on a rotary evaporator. The analysis of the crude reaction mixture by ¹H-NMR revealed the presence of a 5.5:1 mixture of mono-*N*-oxide **2ao** and diheteroarylethane **1ao'**. The crude

mixture was purified by flash column chromatography on silica gel (hexanes/EtOAc = 2:1), resulting in the isolation of 2-(2-(2-quinolinyl)ethyl)pyridine (**2ao**, 43 mg, 86%) as a brown oil.

¹H NMR (400 MHz, CDCl₃) δ 8.26 (dd, *J* = 5.5, 2.3 Hz, 1H), 8.02 (d, *J* = 8.3 Hz, 2H), 7.75 (dd, *J* = 8.1, 1.7 Hz, 1H), 7.66 (ddd, *J* = 8.4, 6.9, 1.6 Hz, 1H), 7.47 (ddd, *J* = 8.0, 6.8, 1.3 Hz, 1H), 7.33–7.27 (m, 1H), 7.24 (dd, *J* = 6.4, 3.5 Hz, 1H), 7.15–7.04 (m, 2H), 3.45 (tt, *J* = 7.8, 3.8 Hz, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 160.8, 151.5, 147.8, 139.6, 136.4, 129.4, 128.8, 127.6, 126.8, 126.3, 125.9, 125.5, 123.7, 121.5, 34.7, 30.5.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₆H₁₅N₂O) 251.1, found 251.1.

Experimental Procedure for Mechanistic studies

Experimental Procedure for Scheme 1a (In the absence of a photocatalyst)

A 10-mL test tube, equipped with a small magnetic stir bar, was charged with 3-quinolinecarbonitrile *N*-oxide (**2I**, 34.0 mg, 0.200 mmol, 1.0 equivalent) and IPA (0.30 mL). The test tube was fitted with a rubber septum, and acetone (2.9 mL, 0.07 M) was added via a syringe under an atmosphere of nitrogen. After degassing by N₂ bubbling at room temperature for 5 minutes, the reaction mixture was irradiated with two 3 W blue LEDs at room temperature for 4 hours. TLC analysis indicated that no conversion of **1I** into the deoxygenated product **2I** occurred.

Experimental Procedure for Scheme 1b (In the absence of visible light)

In a 10-mL test tube, equipped with a small magnetic stir bar, 6-iodoquinoline *N*-oxide (**1q**, 54.2 mg, 0.200 mmol, 1.0 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 1.7 mg, 3.0 μmol, 1.5 mol%), and IPA (0.92 mL) were combined. The test tube was fitted with a rubber septum and covered with aluminum foil, and acetone (2.9 mL, 0.07 M) was added via a syringe under a nitrogen atmosphere. After degassing by N₂ bubbling at room temperature for 5 minutes, the reaction mixture was stirred in the dark for 3 hours. TLC analysis indicated that no conversion of **1q** into the deoxygenated product **2q** occurred.

Experimental Procedure for Quantum yield(Φ) measurement

(1) Photon-flux measurement:

The photon flux of the photoreactor and quantum yield of this reaction were determined by standard ferrioxalate actinometry² following a modified literature procedure.³ A 0.15 M ferrioxalate solution was prepared by dissolving potassium ferrioxalate trihydrate (736.8 mg, 1.5 mmol) in 10 mL of 0.05 M H₂SO₄. A buffer solution was prepared by dissolving sodium acetate (2.25 g, 27.4 mmol) in 10 mL of 0.5 M H₂SO₄. Then, 4.0 mL of the 0.15 M ferrioxalate solution was added to 4.0 mL vial. The vial was irradiated for 15, 30, and 60 s. After irradiation, 100 μL of the solution was transferred to an aluminum foil covered 10 mL volumetric flask containing 1,10-phenanthroline (15 mg, 0.08 mmol) dissolved in 3.0 mL of the buffer solution. To the flask, water was added to make a total volume of 10 mL. After mixing the reagent

completely, the solution was stored for 1 h in the dark. 400 μL of the solution was transferred to a 1 cm path quartz cuvette and the absorbance at $\lambda = 452 \text{ nm}$ was measured by using UV-visible spectrophotometer (Fig. S1). This process was repeated for 15, 30, and 60 s and the absorbance of a non-irradiated sample was also measured.

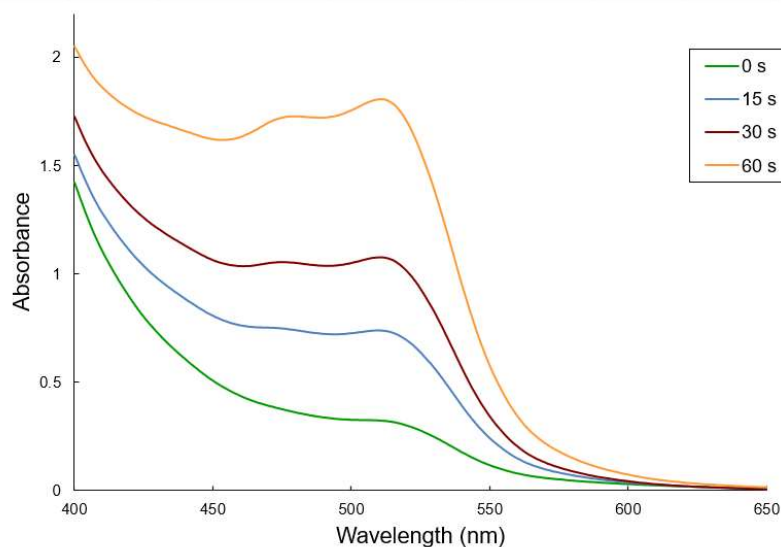


Fig. S1 UV-Vis spectra of ferrioxalate/1,10-phenanthroline solutions after light irradiation

The number of newly generated Fe^{2+} ions (mole) was calculated by following equation:

$$\text{mol Fe}^{2+} = \frac{V_1 V_3 \Delta A(510 \text{ nm})}{V_2 l \epsilon(510 \text{ nm})}$$

Where V_1 is the volume of 0.15 M ferrioxalate solution irradiate ($4.0 \times 10^{-3} \text{ L}$), V_2 is the volume of the aliquot for measurement of the concentration of Fe^{2+} ions ($1.0 \times 10^{-4} \text{ L}$), V_3 is the final volume after complexation with 1,10- phenanthroline ($1.0 \times 10^{-2} \text{ L}$), $\Delta A(510 \text{ nm})$ is the difference in absorbance at $\lambda = 510 \text{ nm}$ between the irradiated and non-irradiated ferrioxalate/1,10 phenanthroline solutions, l is the optical path length of the irradiation (1 cm), and $\epsilon(510 \text{ nm})$ is the molar absorptivity of the $\text{Fe}(\text{phen})_3^{2+}$ complex at $\lambda = 510 \text{ nm}$ ($11100 \text{ L mol}^{-1} \text{ cm}^{-1}$). The moles of Fe^{2+} were plotted as a function of time (Fig. S2).

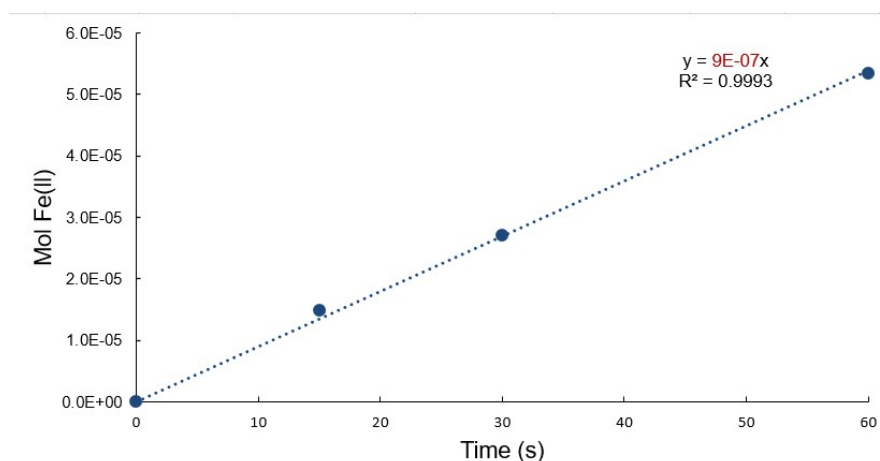


Fig. S2 Moles of Fe²⁺ formed vs. irradiation time

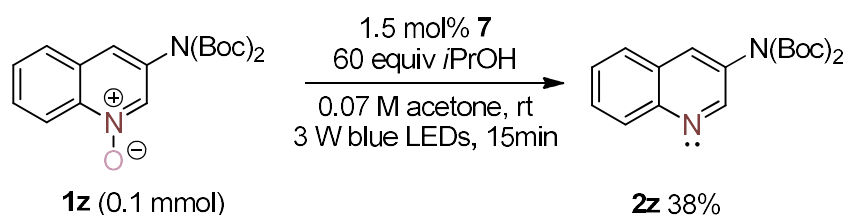
The photon flux was calculated by following equation:

$$\text{photon flux} = \frac{\text{mol Fe}^{2+}}{\Phi \times t \times f}$$

Where Φ is the quantum yield of the ferrioxalate actinometer (0.845 for a 0.15 M solution at 457 nm), t is the time, and f is the fraction of absorbed light at $\lambda = 452$ nm ($f = 1 - 10^{-A}$, $A(452 \text{ nm})$ of 0.15 M ferrioxalate solution measured in a 1 cm path quartz cuvette by UV-Vis spectroscopy to be >5 , therefore $f = 1.00$).

$$\text{photon flux} = \frac{9.0 \times 10^{-7} \text{ mol s}^{-1}}{0.845 \times 1.00} = 1.07 \times 10^{-6} \text{ einstein s}^{-1}$$

(2) Quantum yield(Φ) measurement:



In a 10-mL test tube, equipped with a small magnetic stir bar, 3-[bis(*tert*-butoxycarbonyl)amino]quinoline *N*-oxide (**1z**, 36.0 mg, 0.100 mmol, 1.0 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 0.9 mg, 1.5 μmol , 1.5 mol%), and IPA (0.46 mL) were combined. The test tube was fitted with a rubber septum, and acetone (1.45 mL, 0.07 M) was added via a syringe under a nitrogen atmosphere. After degassing by N₂ bubbling at room temperature for 5 minutes, the reaction mixture

was irradiated with two 3 W blue LEDs ($\lambda = 452$ nm) at room temperature for 900 s (15 minutes). Product yield of **2z** (38%, 3.8×10^{-5} mol) was measured by $^1\text{H-NMR}$ analysis of the crude mixture in the presence of CH_2Br_2 as an internal standard. The quantum yield (Φ) was calculated using the following equation.

$$\Phi = \frac{n(\text{product}, \text{mol})}{\text{photon flux} \times t \times f}$$

Where t represents the time (15 minutes = 900 seconds) and f denotes the fraction of light absorbed by the reaction mixture at $\lambda = 452$ nm, where $f = 1 - 10^{-A}$ (with A being the absorbance of the reaction mixture at $\lambda = 452$ nm measured in a 1 cm path quartz cuvette by a UV-Vis spectrometer; the measured $A = 3.333$, thus $f \approx 1$). The quantum yield was then calculated to be 0.03 by the following equation.

$$\Phi = \frac{3.8 \times 10^{-5} \text{ mol}}{1.07 \times 10^{-6} \text{ einstein s}^{-1} \times 900 \text{ s} \times 1.00} = 0.03$$

The quantum yield studies indicate that this is not a radical-chain propagation mechanism as the $\Phi < 1$.

Experimental Procedure for Scheme 1c (Visible Light On/Off Experiment)

Five 10-mL test tubes, each fitted with a magnetic stir bar, were prepared. Methyl 3-quinolinecarboxylate *N*-oxide (**1a**, 40.6 mg, 0.200 mmol, 1.0 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 1.7 mg, 3.0 μmol , 1.5 mol%), and IPA (0.92 mL) were added to the tubes. The tubes were sealed with rubber septa, and acetone (2.9 mL, 0.07 M) was injected using a syringe under a nitrogen atmosphere. After degassing by bubbling N_2 at room temperature for 5 minutes, all tubes were exposed to room temperature irradiation with two 3 W blue LEDs for 50 minutes. Subsequently, one tube was chosen, and its contents were concentrated using a rotary evaporator. The resulting residue was purified via flash column chromatography on silica gel (hexanes/EtOAc = 3:1) to obtain methyl 3-quinolinecarboxylate (**2a**, 19.5 mg, 52%) as a white solid. The remaining four tubes were covered with aluminum foil and kept under stirring with 3 W blue LEDs. After 30 minutes, one tube was selected, and its contents were concentrated using a rotary evaporator. The obtained residue was purified similarly, yielding methyl 3-quinolinecarboxylate (**2a**, 19 mg, 51%) as a white solid. Following the removal of the aluminum foil, the three remaining tubes were stirred under irradiation for 50 minutes. One tube was chosen, and its contents were concentrated and purified as before, resulting in methyl 3-

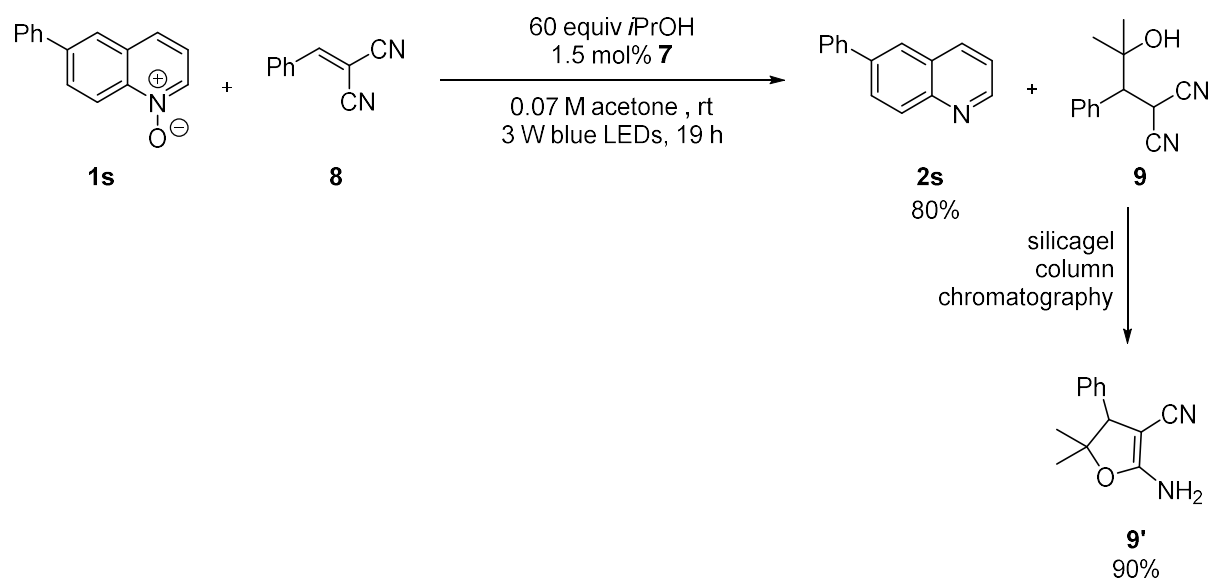
quinolinecarboxylate (**2a**, 25 mg, 68%) as a white solid. Additionally, two tubes were covered with aluminum foil and irradiated for 30 minutes. The contents of one tube were purified as previously described, yielding methyl 3-quinolinecarboxylate (**2a**, 26 mg, 70%) as a white solid. The other tube, after removing the aluminum foil, was stirred under irradiation for 50 minutes. Similar purification of its contents produced 3-quinolinecarboxylate (**2a**, 34 mg, 92%) as a white solid.

Experimental Procedure for Scheme 1d (employing radical inhibitor)

In a couple of 10-mL test tubes, each equipped with a small magnetic stir bar, 3-acetylquinoline *N*-oxide (**1f**, 37.4 mg, 0.20 mmol, 1.0 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 1.7 mg, 3.0 μ mol, 1.5 mol%), and IPA (0.92 mL) were combined. One tube was charged with (2,2,6,6-tetramethylpiperidin-1-yl)oxyl (TEMPO, 188 mg, 1.2 mmol, 6.0 equivalent), and the other tube was charged with 2,2-diphenyl-1-picrylhydrazyl (DPPH, 87 mg, 0.22 mmol, 1.1 equivalent). These test tubes were fitted with rubber septa, and acetone (2.9 mL, 0.07 M) was added via a syringe under a nitrogen atmosphere. After degassing by N₂ bubbling at room temperature for 5 minutes, the reaction mixtures were irradiated with two 3 W blue LEDs at room temperature for 7 hours. In the reaction with TEMPO, the resulting residue was directly purified by flash column chromatography on silica gel (hexanes/EtOAc = 1:1) to afford 3-acetylquinoline (**2f**, 8.9 mg, 26%) as a white solid. For the reaction with DPPH, TLC analysis indicated that no conversion of **1f** into the deoxygenated product **2f** occurred.

Experimental Procedure for Scheme 1e (radical trapping experiment)

In a 10-mL test tube, equipped with a small magnetic stir bar, methyl quinoline-3-carboxylate *N*-oxide (**1a**, 40.6 mg, 0.200 mmol, 1.0 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 1.7 mg, 3.0 μ mol, 1.5 mol%), and benzylidenemalononitrile (**8**, 30.8 mg, 0.200 mmol, 1.0 equivalent), IPA (0.92 mL) were combined. The test tube was fitted with a rubber septum, and CD₃CN (2.9 mL, 0.07 M) was added via a syringe under a nitrogen atmosphere. After degassing by N₂ bubbling at room temperature for 5 minutes, the reaction mixture was irradiated with two 3 W blue LEDs at room temperature for 1 hours. Methyl quinoline-3-carboxylate (**2a**, 60%) and 2-(2-hydroxy-2-methyl-1-phenylpropyl)propanedinitrile (**9**, 50%) were determined by ¹H NMR analysis, utilising dibromomethane as an internal standard.

Experimental Procedure for radical trapping experiment II (footnote 46)

In a 10-mL test tube, equipped with a small magnetic stir bar, 6-phenylquinoline *N*-oxide (**1s**, 44.3 mg, 0.20 mmol, 1.0 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 1.7 mg, 3.0 μ mol, 1.5 mol%), and benzylidenemalononitrile (**8**, 30.8 mg, 0.20 mmol, 1.00 equivalent), IPA (0.92 mL) were combined. The test tube was fitted with a rubber septum, and acetone (2.9 mL, 0.07 M) was added via a syringe under a nitrogen atmosphere. After degassing by N₂ bubbling at room temperature for 5 minutes, the reaction mixture was irradiated with two 3 W blue LEDs at room temperature for 19 hours. Upon completion of the reaction as determined by TLC, the reaction mixture was concentrated on a rotary evaporator. The resulting residue was purified by flash column chromatography on silica gel (hexanes/EtOAc = 10:1 to 3:1) to afford 6-phenylquinoline (**2s**, 33.0 mg, 80%) and 2-amino-5,5-dimethyl-4-phenyl-4,5-dihydrofuran-3-carbonitrile (**9'**, 38.4 mg, 90%). The latter compound resulted from the cyclisation of 2-(2-hydroxy-2-methyl-1-phenylpropyl)propanedinitrile (**9**) during the course of column chromatography.

2-Amino-5,5-dimethyl-4-phenyl-4,5-dihydrofuran-3-carbonitrile (9')

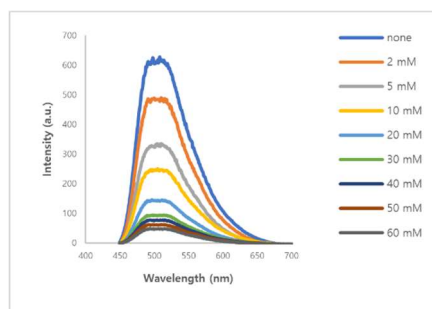
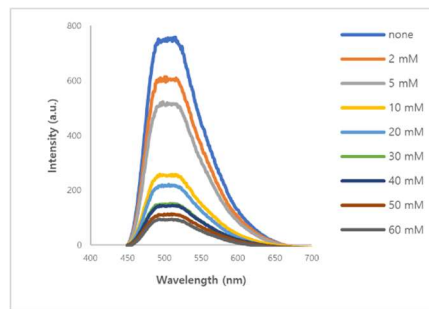
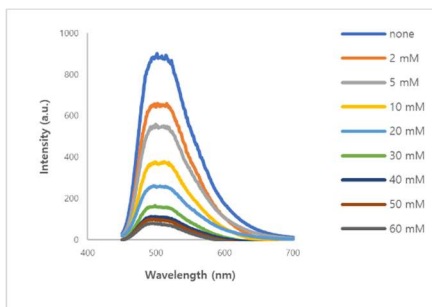
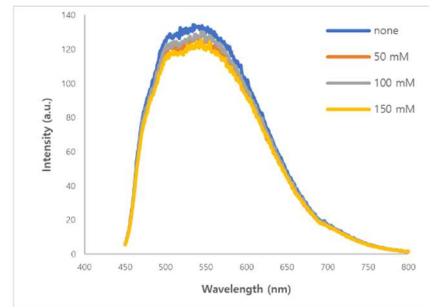
¹H NMR (400 MHz, CDCl₃) δ 7.35 (t, *J* = 7.4 Hz, 2H), 7.29 (d, *J* = 7.4 Hz, 1H), 7.20 (d, *J* = 7.7 Hz, 2H), 4.95 (s, 2H), 4.04 (s, 1H), 1.55 (s, 3H), 0.88 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 167.0, 138.8, 128.6, 128.4, 127.6, 119.8, 91.4, 57.6, 55.8, 29.2, 24.3.

LC-MS (ESI) *m/z* calculated for [M+1]⁺ (C₁₃H₁₅N₂O) 215.1, found 215.1.

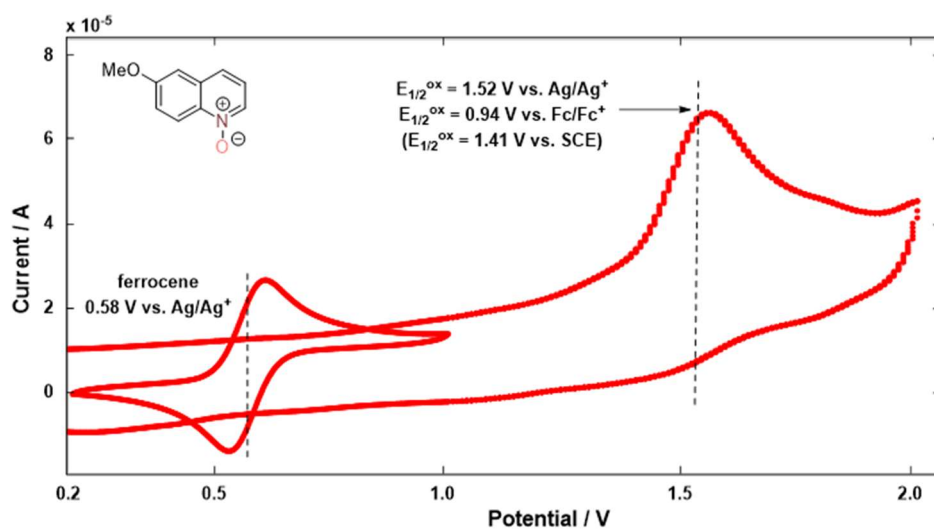
Stern-Volmer Luminescence Quenching Experiments (Fig. 2)**Emission Quenching of Mes-*t*Bu₂Acr⁺ (7)**

A 150-mL volumetric flask containing the photocatalyst, 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 2.8 mg, 5 μmol) was fitted with a rubber septum and charged with acetone (ca. 101 mL) via a syringe. The resulting homogeneous solution was degassed by sparging with nitrogen until its volume reached to 100 mL. The solutions of IPA (100 mM), **1b**, **1x**, and quinoline *N*-oxide (60 mM) in acetone were also prepared similarly. Stern-Volmer luminescence quenching experiments were performed using a Varian Cary eclipse fluorescence spectrophotometer. In each experimentation, the solution of **7** and varying concentrations of quencher (IPA, **1b**, **1x**, or quinoline *N*-oxide) were combined in degassed acetone in 1 cm screw-top quartz cuvettes. For the emission quenching of **7**, the photocatalyst concentration was 20 μM, the solution was excited at 450 nm, and the emission intensity was observed at 510 nm. Plots were constructed according to the Stern-Volmer equation $I_0/I = 1 + k_q\tau_0[Q]$. I_0 and I represent the intensities of the emission in the absence and presence of a quencher at 510 nm, respectively.

**Fig. S3** Emission Quenching of **7** by **1x****Fig. S5** Emission Quenching of **7** by **1b****Fig. S4** Emission Quenching of **7** by quinoline *N*-oxide**Fig. S6** Emission Quenching of **7** by IPA

Cyclic Voltammetry Measurements (Fig. 2)

Cyclic voltammograms were collected with a Gamry Potentiostat. Samples were prepared with 0.004 mmol of substrate in 4 mL of 0.1 M tetra-*n*-butylammonium hexafluorophosphate in anhydrous, degassed acetonitrile. Measurements employed a glassy carbon working electrode, platinum wire counter electrode, Ag/Ag⁺ reference electrode, and a scan rate of 100 mV/s. After adding 0.004 mmol ferrocene as an internal standard, the potential of ferrocene was measured under the same conditions. All potentials reported here vs. SCE and been converted by adding + 0.47 V and V to the potentials measured vs. Fc/Fc⁺.

**Fig.S7** Cyclic Voltammetry Measurement of **1x**

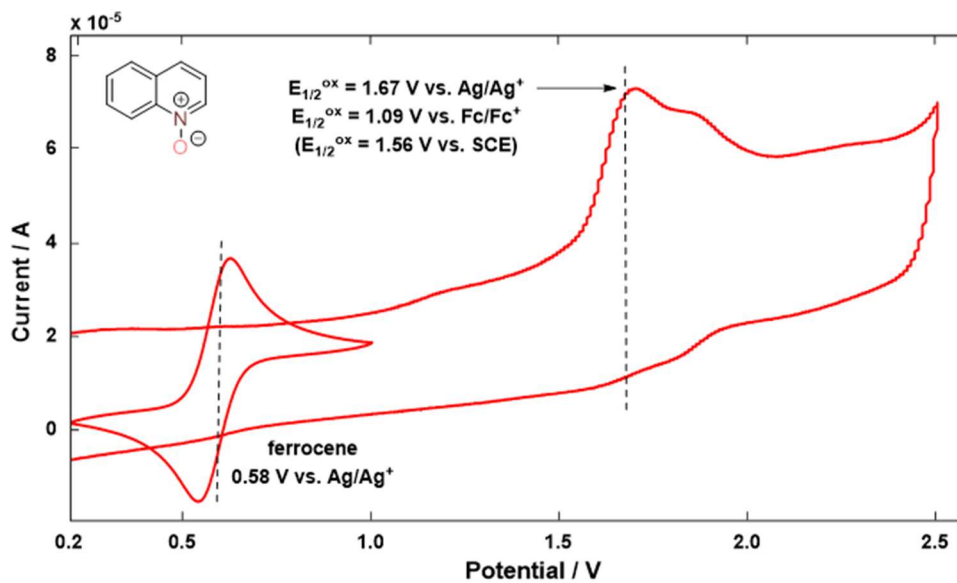


Fig.S8 Cyclic Voltammery Measurement of quinoline *N*-oxide

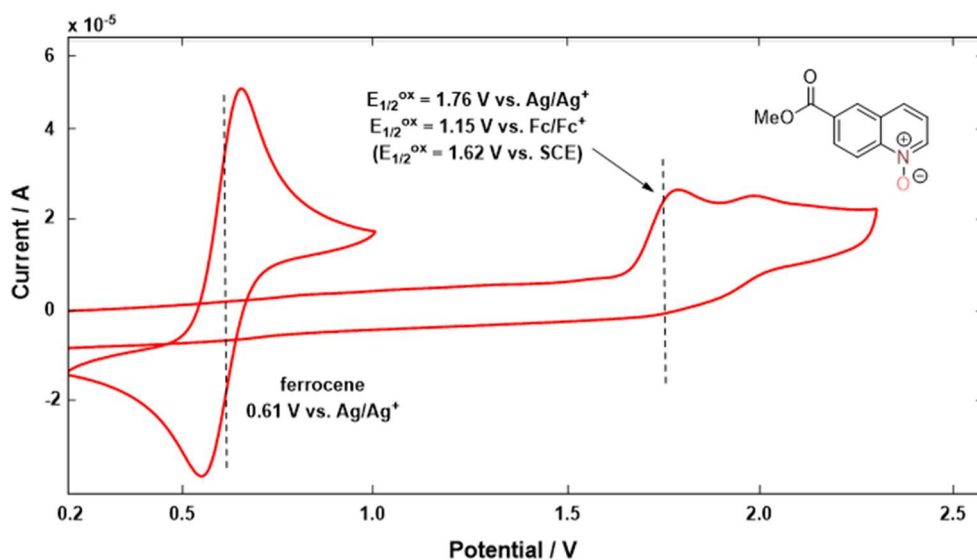
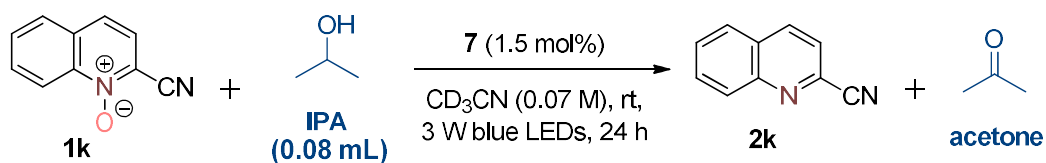
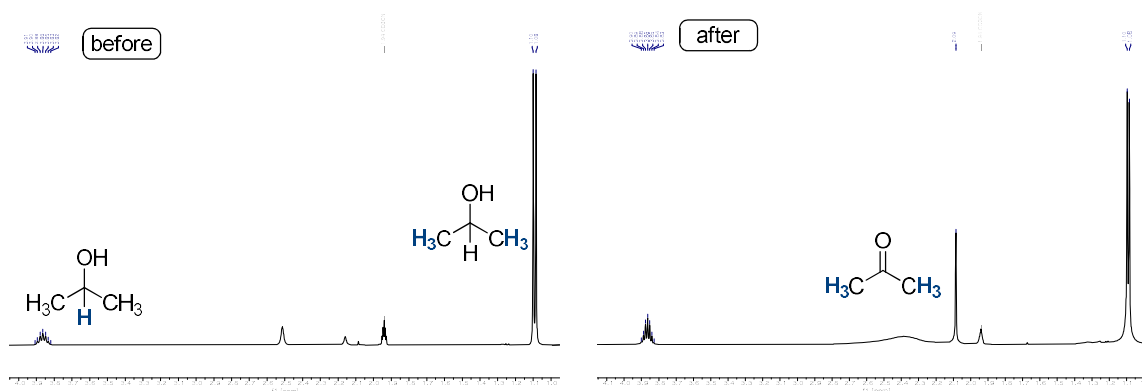
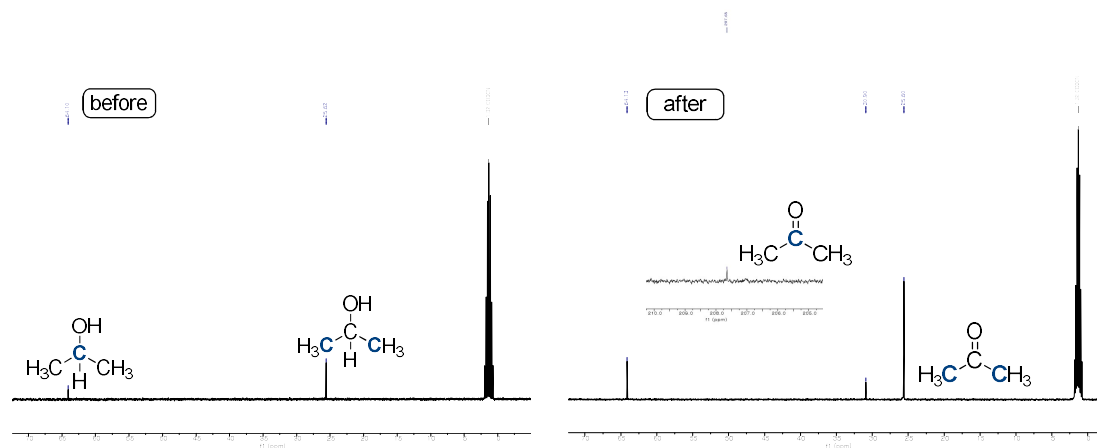


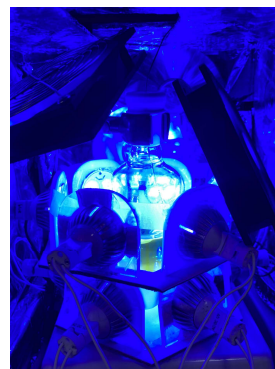
Fig.S9 Cyclic Voltammery Measurement of 1b

NMR Spectroscopic Studies for Confirming the Generation of Acetone as a Byproduct



In a 10-mL test tube, equipped with a small magnetic stir bar, 2-quinolinecarbonitrile *N*-oxide (**1k**, 34.0 mg, 0.200 mmol, 1.0 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 1.7 mg, 3.0 μ mol, 1.5 mol%), IPA (0.08 mL), and acetonitrile-*d*₃ (2.9 mL, 0.07 M) were combined. A small portion (ca. 0.6 mL) of the resulting mixture was transferred into a J-Young tube via a syringe, and the homogeneous mixture was thoroughly degassed using the freeze-pump-melting method (3 cycles). The reaction mixture was irradiated with two 3 W blue LEDs at room temperature for 24 hours. The disappearance of IPA and the simultaneous generation of acetone were validated by both ¹H and ¹³C NMR analyses, as shown in Fig. S10 and Fig. S11.

Fig. S10 Acetone Confirm in ¹H NMRFig. S11 Acetone Confirm in ¹³C NMR

Experimental Procedure for Multigram-Scale Deoxygenation Reaction (Scheme 3)**Fig. S12** Gram-scale Photoreactor (light-off)**Fig. S13** Gram-scale Photoreactor (light-on)**(a) Recrystallisation**

In a 350-mL test tube, equipped with a magnetic stir bar, 2-quinolinecarbonitrile *N*-oxide (**1k**, 1.3 g, 7.5 mmol, 1.0 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 65 mg, 0.11 mmol, 1.5 mol%), and IPA (11.5 mL) were combined. The flask was fitted with a rubber septum, and acetone (107 mL, 0.07 M) was added via a syringe under a nitrogen atmosphere. After degassing by N₂ bubbling at room temperature for 30 minutes, the reaction mixture was irradiated with twelve 3 W blue LEDs at room temperature for 11 hours. Afterward, the reaction mixture was concentrated on a rotary evaporator. The resulting residue was purified by recrystallisation from a hot mixture of hexanes and IPA (1:1, *v/v*) to afford 2-quinolinecarbonitrile (**2k**, 0.97 g, 84%) as a pale yellow solid.

In a 350-mL test tube, equipped with a magnetic stir bar, 3-[bis(*tert*-butoxycarbonyl)amino]quinoline *N*-oxide (**1z**, 1.0 g, 3.0 mmol, 1.0 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 25.6 mg, 0.045 mmol, 1.5 mol%), and IPA (14 mL) were combined. The flask was fitted with a rubber septum, and acetone (43 mL, 0.07 M) was added via a syringe under a nitrogen atmosphere. After degassing by N₂ bubbling at room temperature for 30 minutes, the reaction mixture was irradiated with twelve 3 W blue LEDs at room temperature for 23 hours. Afterward, the reaction mixture was concentrated on a rotary evaporator. The resulting residue was purified by recrystallisation from a hot mixture of hexanes and IPA (3:1, *v/v*) to afford 3-[bis(*tert*-butoxycarbonyl)amino]quinoline (**2z**, 0.95 g, 92%) as a white solid.

(b) Kugelrohr distillation

In a 350-mL test tube, equipped with a magnetic stir bar, 4-benzoylpyridine *N*-oxide (**1ak**, 1.2 g, 6.0 mmol, 1.0 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 51 mg, 0.09 mmol, 1.5 mol%), and IPA (27.5 mL) were combined. The flask was fitted with a rubber septum, and acetone (86 mL, 0.07 M) was added via a syringe under a nitrogen atmosphere. After degassing by N₂ bubbling at room temperature for 30 minutes, the reaction mixture was irradiated with twelve 3 W blue LEDs at room temperature for 15 hours. Afterward, the reaction mixture was concentrated on a rotary evaporator. The resulting residue was purified by Kugelrohr distillation to afford 4-benzoylpyridine (**2ak**, 1.1 g, 97%) as a brown oil.

In a 350-mL test tube, equipped with a magnetic stir bar, 4-acetylpyridine *N*-oxide (**1an**, 1.1 g, 8.0 mmol, 1.0 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 69 mg, 0.11 mmol, 1.5 mol%), and IPA (37 mL) were combined. The flask was fitted with a rubber septum, and acetone (114 mL, 0.07 M) was added via a syringe under a nitrogen atmosphere. After degassing by N₂ bubbling at room temperature for 30 minutes, the reaction mixture was irradiated with twelve 3 W blue LEDs at room temperature for 15 hours. Afterward, the reaction mixture was concentrated on a rotary evaporator. The resulting residue was purified by Kugelrohr distillation to afford 4-acetylpyridine (**2an**, 0.84 g, 87%) as a white solid.

Experimental Procedure for Recycling Process (Scheme 4)

In a 250-mL round-bottomed flask, equipped with a magnetic stir bar, 4-benzoylpyridine *N*-oxide (**1ak**, 1.02 g, 5.1 mmol, 1.0 equivalent), 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 44 mg, 0.077 mmol, 1.5 mol%), and IPA (25 mL) were combined. The flask was fitted with a rubber septum, and acetone (73 mL, 0.07 M) was added via a syringe under a nitrogen atmosphere. After degassing by N₂ bubbling at room temperature for 30 minutes, the reaction mixture was irradiated with twelve 3 W blue LEDs at room temperature for 17 hours. Afterward, the round-bottomed flask was connected to a distillation still and heated in an oil bath to collect acetone and IPA together. The residue in the round-bottomed flask was then transferred to a small bulb, which was assembled with a Büchi Kugelrohr distillation apparatus to collect a small amount of acetone and IPA together. The resulting

mixture in the bulb was purified by Kugelrohr distillation to afford 4-benzoylpyridine (**2ak**, 0.86 g, 92%) as a white solid. The combined mixture of acetone and IPA (a total of 93 mL) was determined to be approximately 2.7:1 by ¹H-NMR spectroscopy. In another 250-mL round-bottomed flask, equipped with a magnetic stir bar, 4-benzoylpyridine *N*-oxide (**1ak**, 1.02 g, 5.1 mmol, 1.0 equivalent) and 9-mesityl-3,6-di-*tert*-butyl-10-phenylacridinium tetrafluoroborate (**7**, 44 mg, 0.076 mmol, 1.5 mol%) were combined. The flask was fitted with a rubber septum, and the recovered mixture of acetone and IPA was added via a syringe under a nitrogen atmosphere. The resulting mixture was treated with a small amount of acetone (3–7 mL) via a syringe. After degassing by N₂ bubbling at room temperature for 30 minutes, the reaction mixture was irradiated with twelve 3 W blue LEDs at room temperature for 15–19 hours. This recycling process was repeated 9 times more, and the results are summarised in Table S10.

Table S10. Recycling Process^a

Runs	Recovered IPA and acetone (mL)	Amount of acetone added (mL)	Time (h)	Yield (%)
1	91	7	15	92
2	92	6	15	94
3	92	6	15.5	97
4	94	4	16	95
5	94	4	17	95
6	93	5	18	96
7	94	4	18.5	97
8	94	4	18.5	96
9	95	3	19	96
10	94	-	19	96

^aConditions: **1ak** (5.1 mmol), IPA (25 mL), and **7**(1.5 mol%) in degassed acetone (73 mL, 0.07 M).

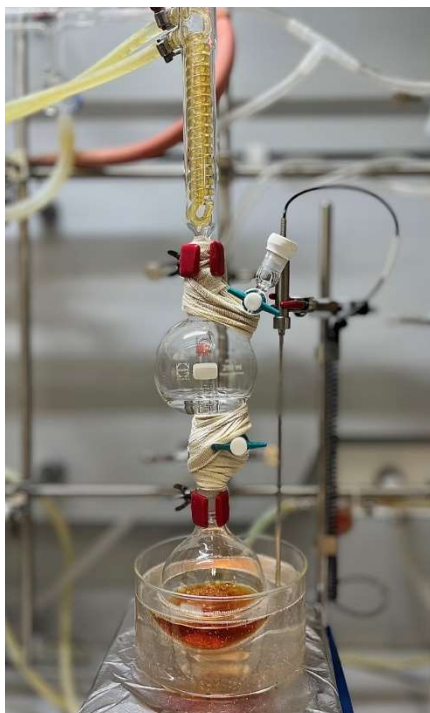


Fig. S14 Recycling Process (simple distillation)



Fig. S15 Recycling Process (Kugelrohr distillation)

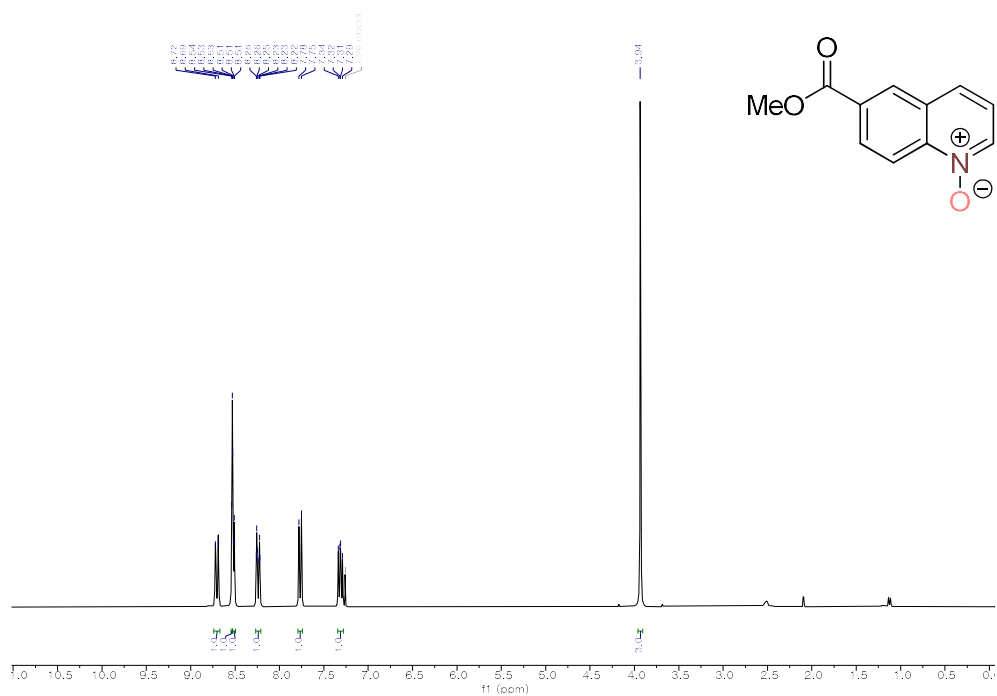
References

1. W.C. Still, M. Kahn and A. Mitra, *J. Org. Chem.*, 1978, **43**, 2923–2925.
2. S. R. Atriardi, J.-Y. Kim, Y. Anita and S. K. Woo, *Bull. Korean Chem. Soc.*, 2023, **44**, 50–54.
3. Z. Zhou, J. Kweon, H. Jung, D. Kim, S. Seo and S. Chang, *J. Am. Chem. Soc.*, 2022, **144**, 9161–9171.
4. J. H. An, K. D. Kim and J. H. Lee, *J. Org. Chem.*, 2021, **86**, 2876–2894.
5. H. Ishii, K. Minegishi, K. Nagatsu and M. R. Zhang, *Tetrahedron*, 2015, **71**, 1588–1596.
6. *PCT Int. Appl.*, WO2008059370 A2, 2008.
7. J. Jeong, D. Lee and S. Chang, *Chem. Commun.*, 2015, **51**, 7035–7038.
8. J. H. An, S. H. Kim and J. H. Lee, *Org. Biomol. Chem.*, 2021, **19**, 3735–3742.
9. J. Jeong, P. Patel, H. Hwang and S. Chang, *Org. Lett.*, 2014, **16**, 4598–4601.
10. A. Biswas, U. Karmakar, S. Nandi and R. Samanta, *J. Org. Chem.*, 2017, **82**, 8933–8942.
11. H. Hwang, J. Kim, J. Jeong and S. Chang, *J. Am. Chem. Soc.*, 2014, **136**, 10770–10776.
12. M. Holmquist, G. Blay, M. C. Muñoz, J. R. Pedro, *Org. Lett.*, 2014, **16**, 1204–1207.
13. K. D. Kim and J. H. Lee, *Org. Lett.*, 2018, **20**, 7712–7716.
14. X. Ma, H. Dang, J. A. Rose, P. R. Rablen and S. Herzon, *J. Am. Chem. Soc.*, 2017, **139**, 5998–6007.
15. A. Biswas, S. Sarkar and R. Samanta, *Chem. Eur. J.*, 2019, **25**, 3000–3004.
16. Z. Y. Liu, L. L. Huang, D. M. Chen and Z. H. Yuan, *Rapid Commun. Mass Spectrom.*, 2010, **24**, 909–918.
17. K. Hanaoka, K. Kikuchi, S. Kobayashi and T. Nagano, *J. Am. Chem. Soc.*, 2007, **129**, 13502–13509.
18. G. Li, C. Jia and K. Sun, *Org. Lett.*, 2013, **15**, 5198–5201.
19. A. Biswas, U. Karmakar, A. Pal and R. Samanta, *Chem. Eur. J.*, 2016, **22**, 13826–13830.
20. A. K. Jha and N. Jain, *Chem. Commun.*, 2016, **52**, 1831–1834.
21. P. Hu, X. Wang, B. Zhang, S. Zhang, Q. Wang and Z. Wang, *ChemMedChem*, 2014, **9**, 928–931.
22. E. Hayashi and A. Miyashita, *Yakugaku Zasshi*, 1977, **97**, 1334–1344.
23. L. Qixing, W. Chunqin, Z. Haifeng, W. Baigui, L. Jinliang, C. Lu and F. Yigang, *Org. Lett.*, 2018, **20**, 971–974.

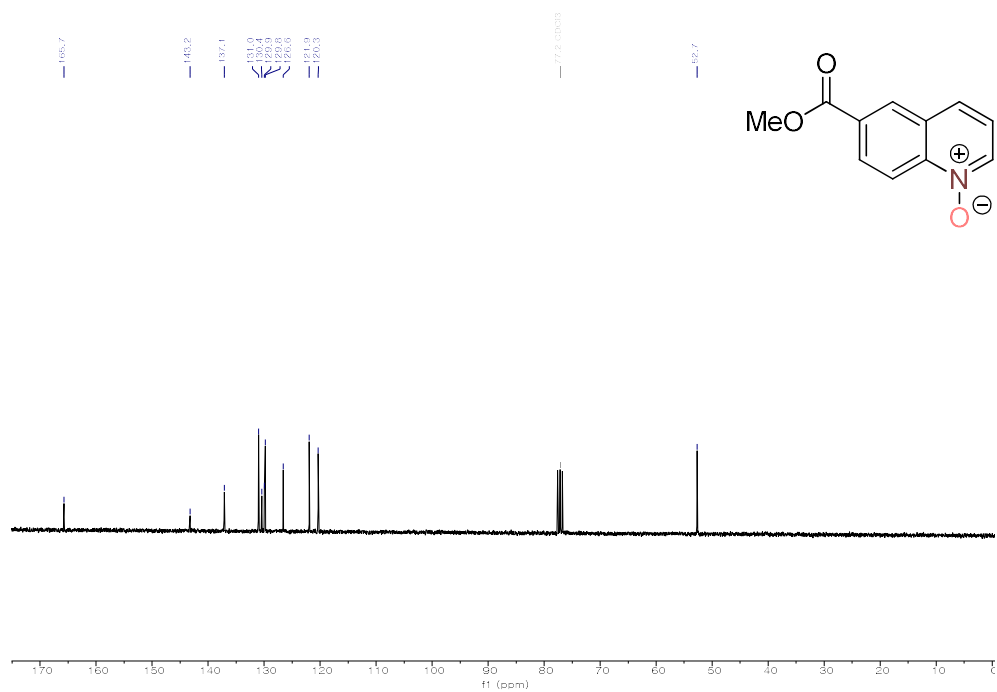
24. D. I. Bugaenko, M. A. Yurovskaya and A. V. *Org. Lett.*, 2021, **23**, 6099–6104.
25. D. Kim, P. Ghosh, N. Y. Kwon, S. H. Han, S. Han, N. K. Mishra, S. Kim and I. S. Kim, *J. Org. Chem.*, 2020, **85**, 2476–2485.
26. W. J. Lominac, M. L. D'Angelo, M. D. Smith, D. A. Ollison and J. M. Hanna Jr., *Tetrahedron Lett.*, 2012, **53**, 906–909.
27. *US Pat.*, US2014/022801, 2014.

¹H & ¹³C NMR Spectra of Methyl Quinoline-6-carboxylate *N*-Oxide (1b)

¹H NMR (300 MHz, CDCl₃)

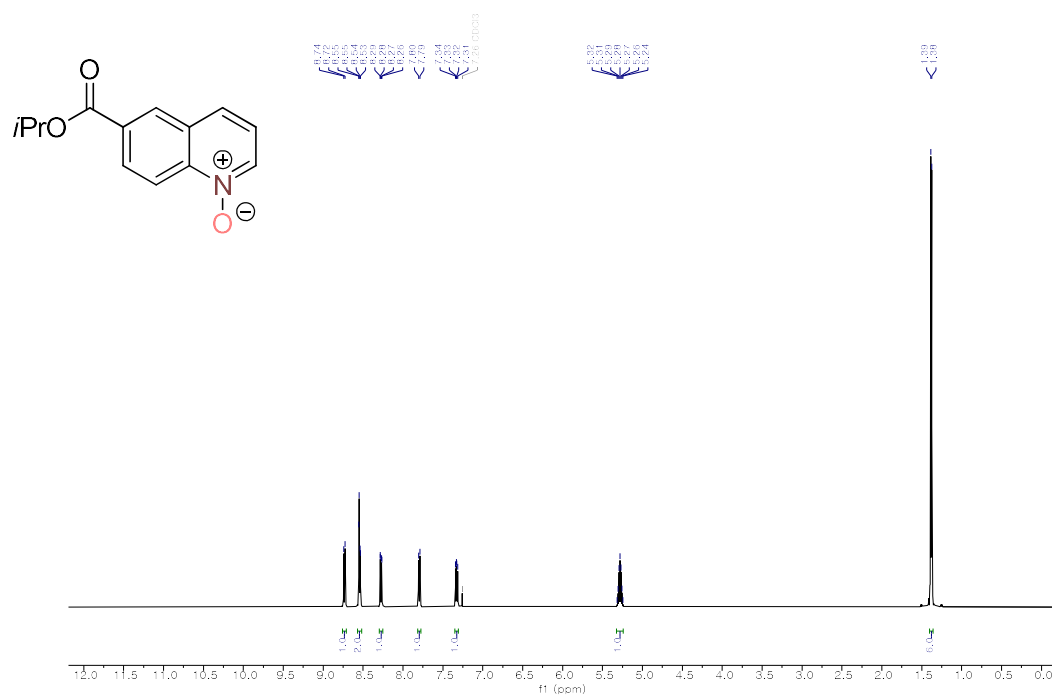


¹³C NMR (75 MHz, CDCl₃)

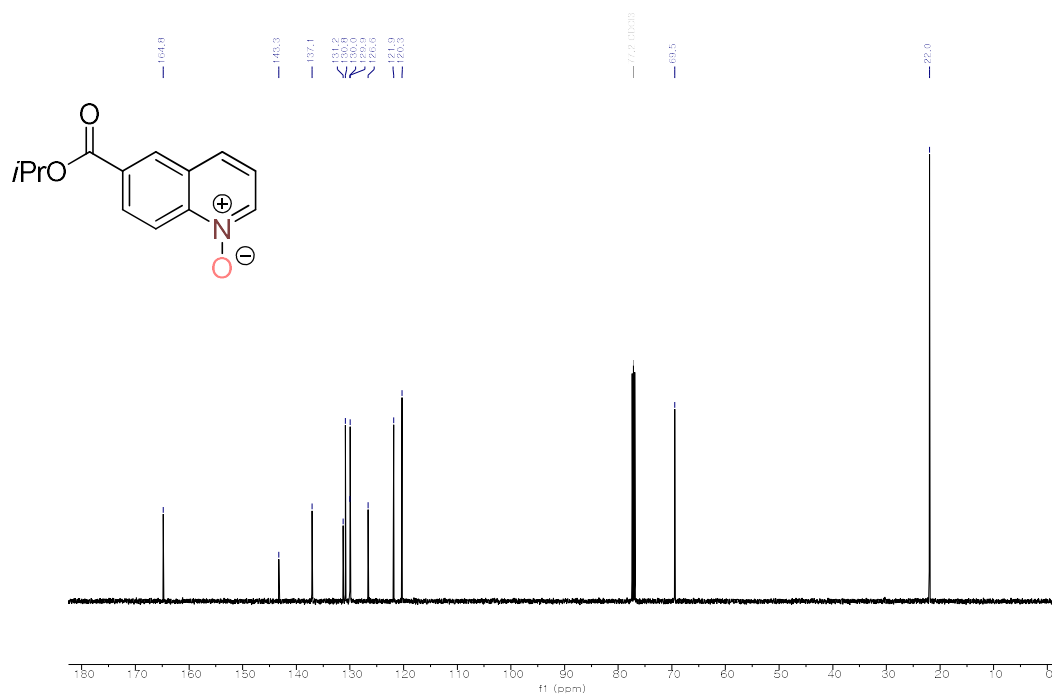


¹H & ¹³C NMR Spectra of Isopropyl Quinoline-6-carboxylate *N*-Oxide (1d)

¹H NMR (500 MHz, CDCl₃)

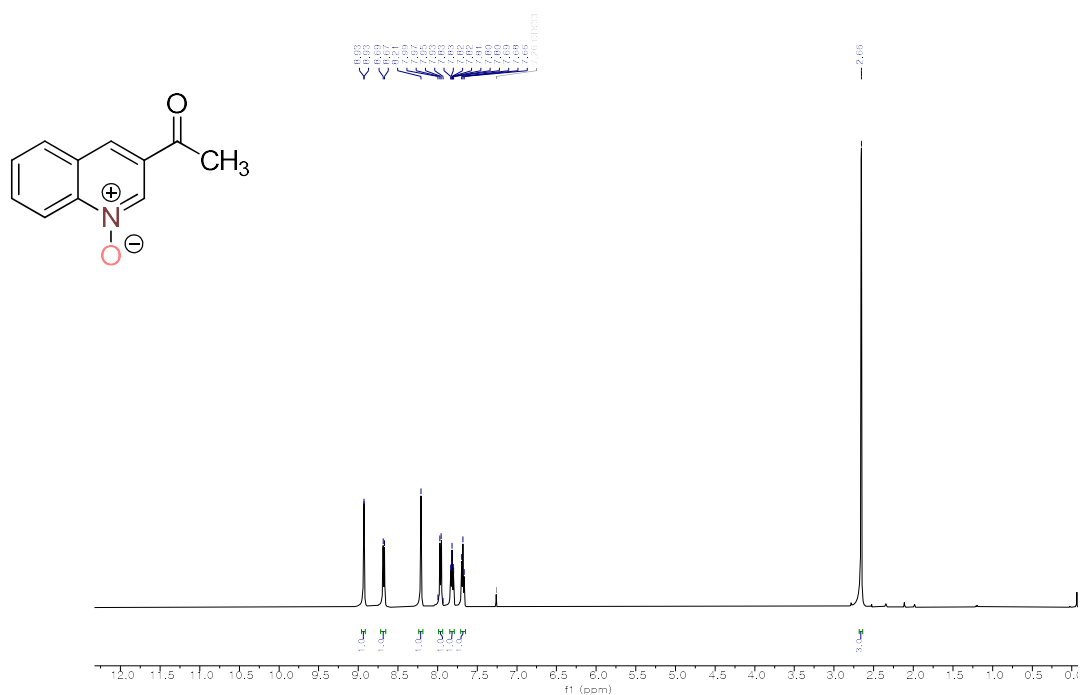


¹³C NMR (126 MHz, CDCl₃)

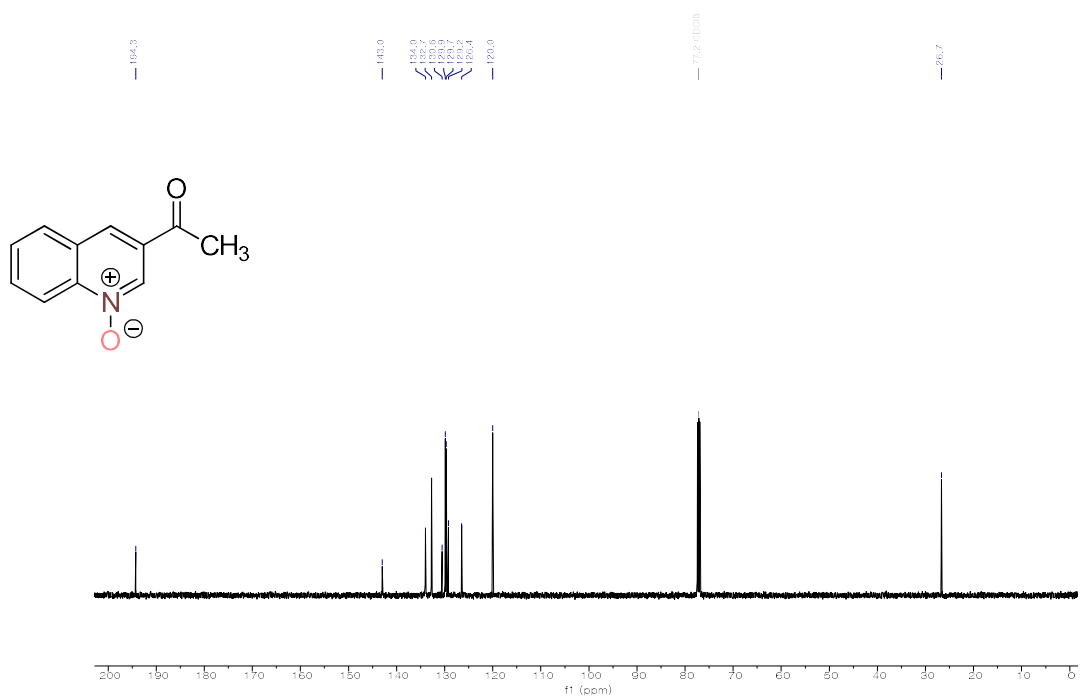


¹H & ¹³C NMR Spectra of 3-Acetylquinoline N-Oxide (1f)

¹H NMR (500 MHz, CDCl₃)

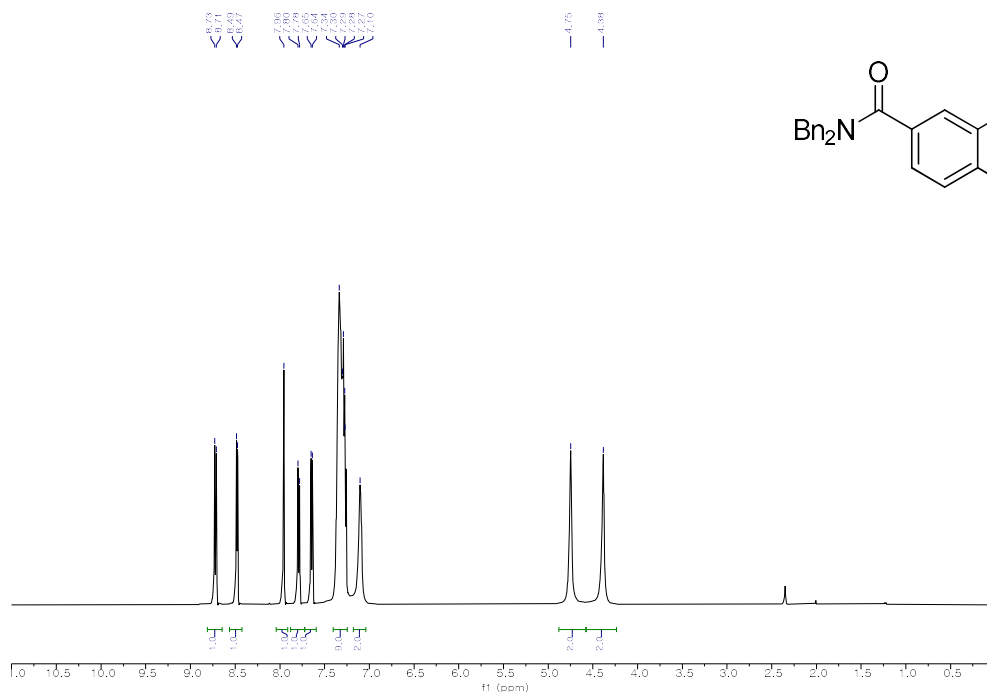


¹³C NMR (126 MHz, CDCl₃)

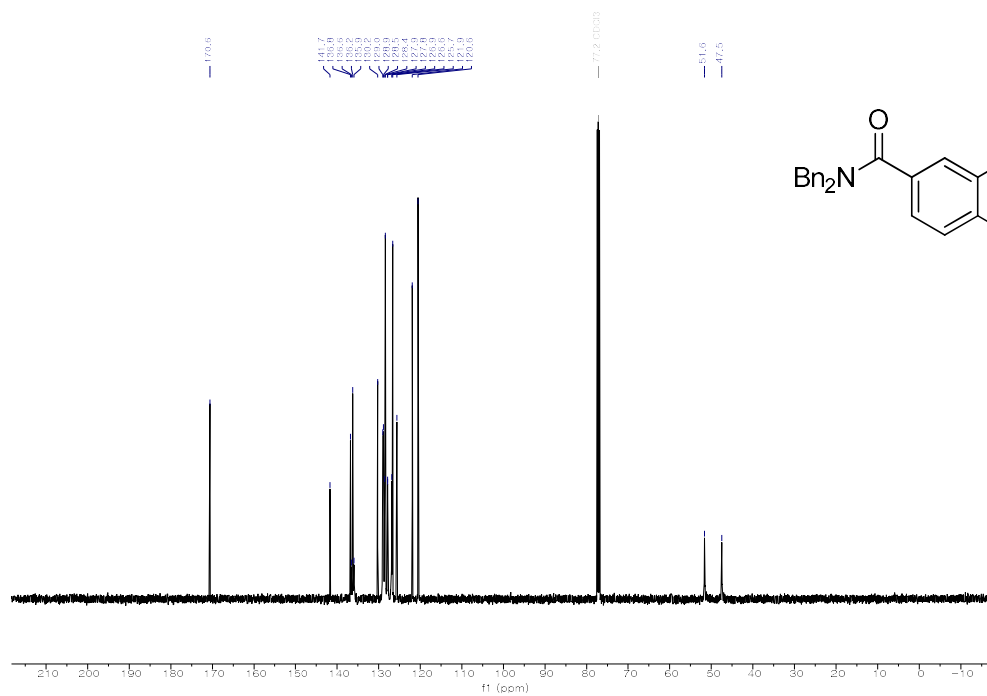


¹H & ¹³C NMR Spectra of *N,N*-Dibenzyl-6-quinolinecarboxamide *N*-Oxide (1j)

¹H NMR (500 MHz, CDCl₃)

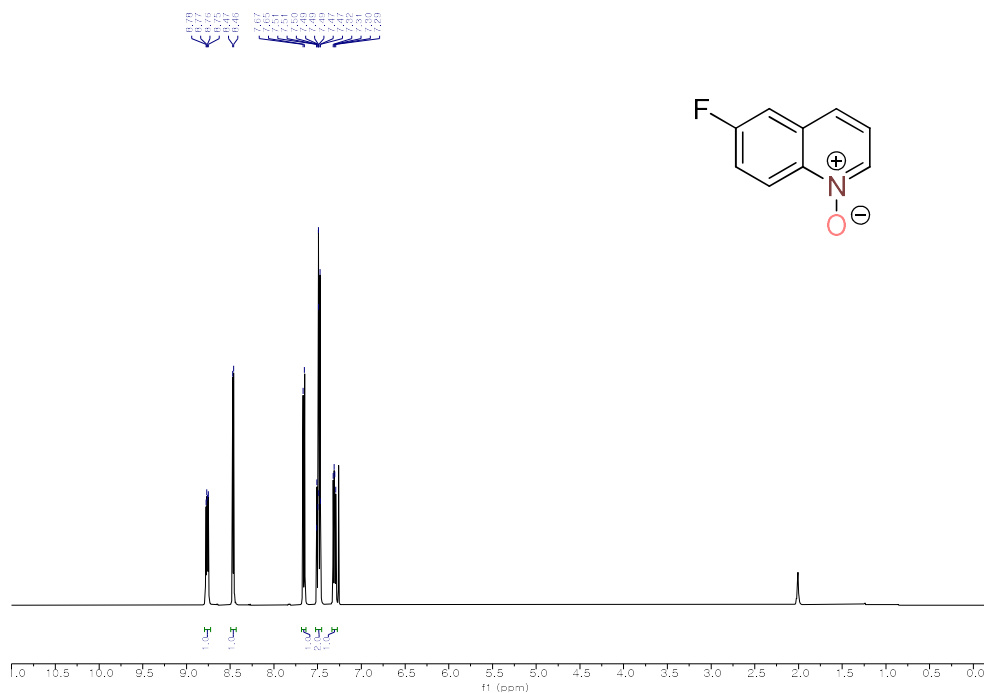


¹³C NMR (126 MHz, CDCl₃)

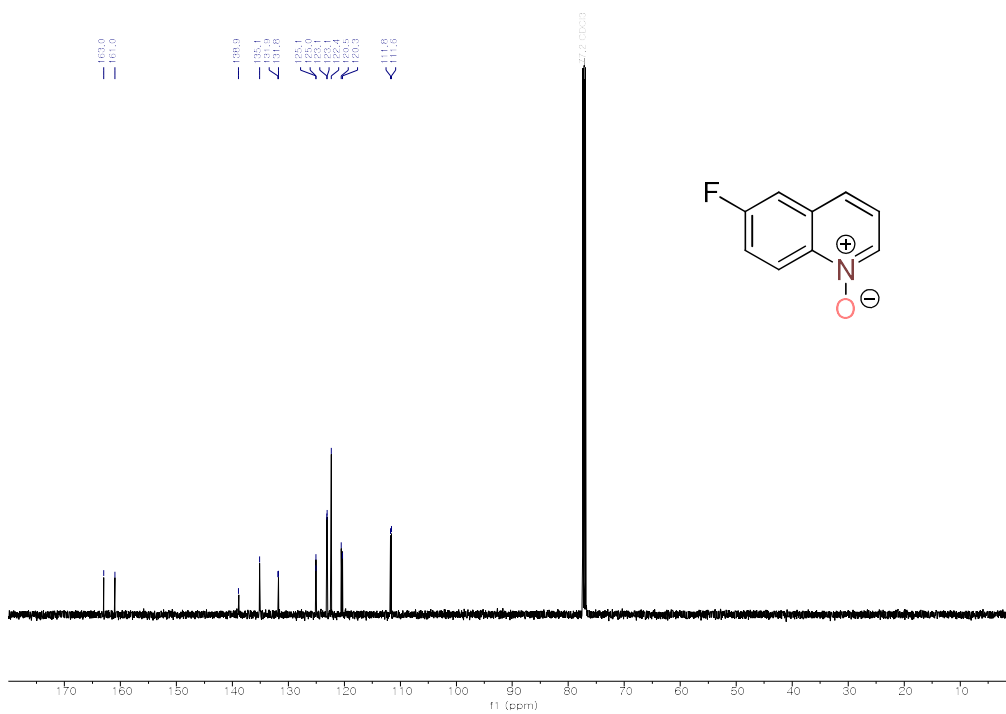


¹H & ¹³C & ¹⁹F NMR Spectra of 6-Fluoroquinoline N-Oxide (1n)

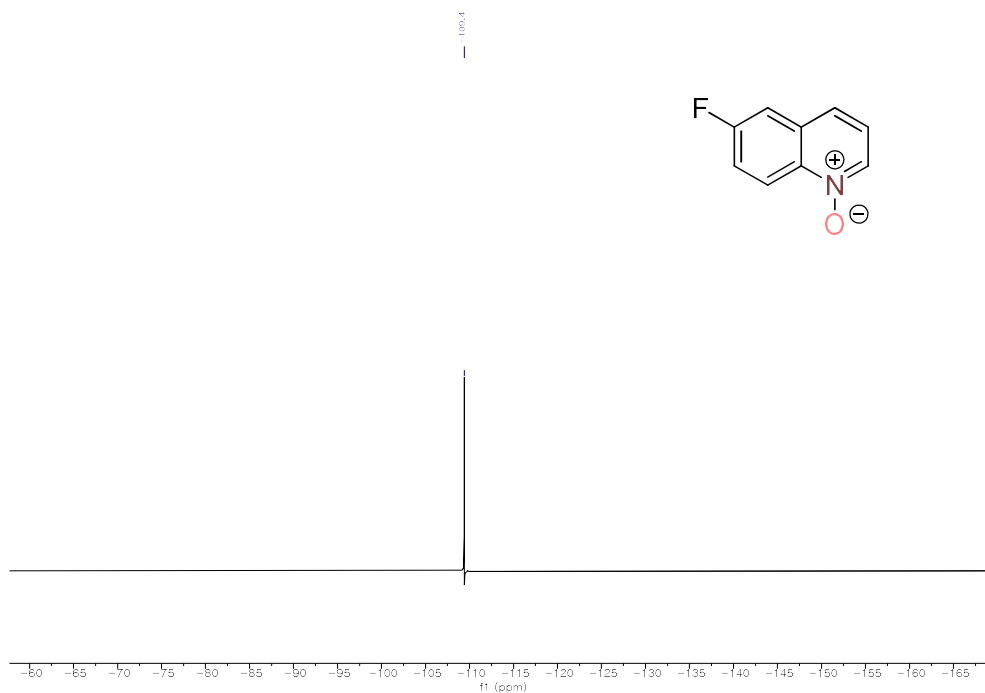
¹H NMR (500 MHz, CDCl₃)



¹³C NMR (126 MHz, CDCl₃)

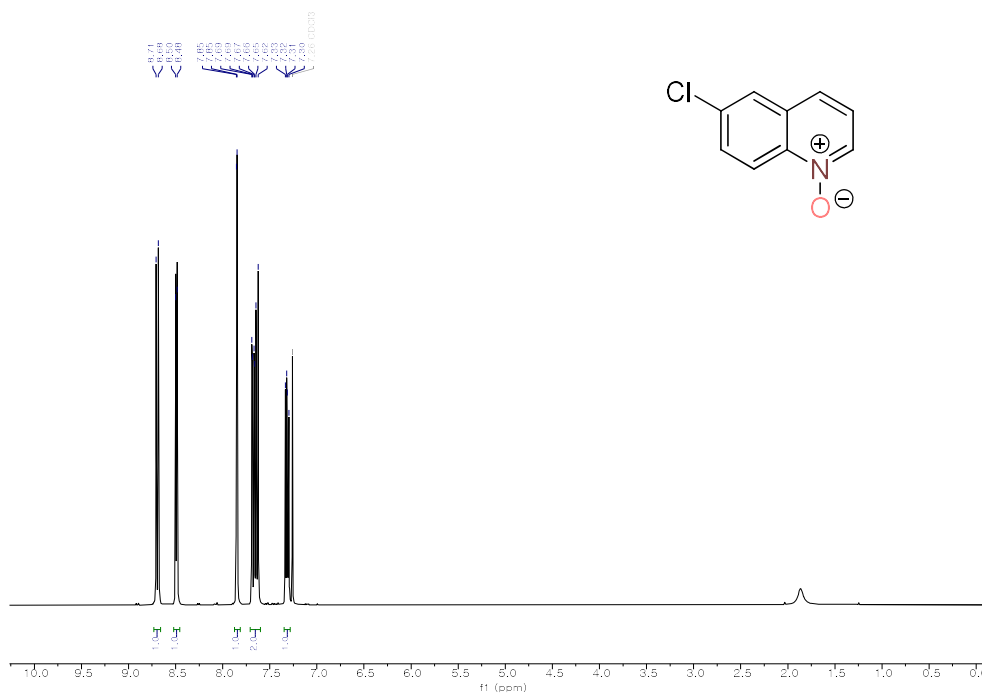


¹⁹F NMR (471 MHz, CDCl₃)



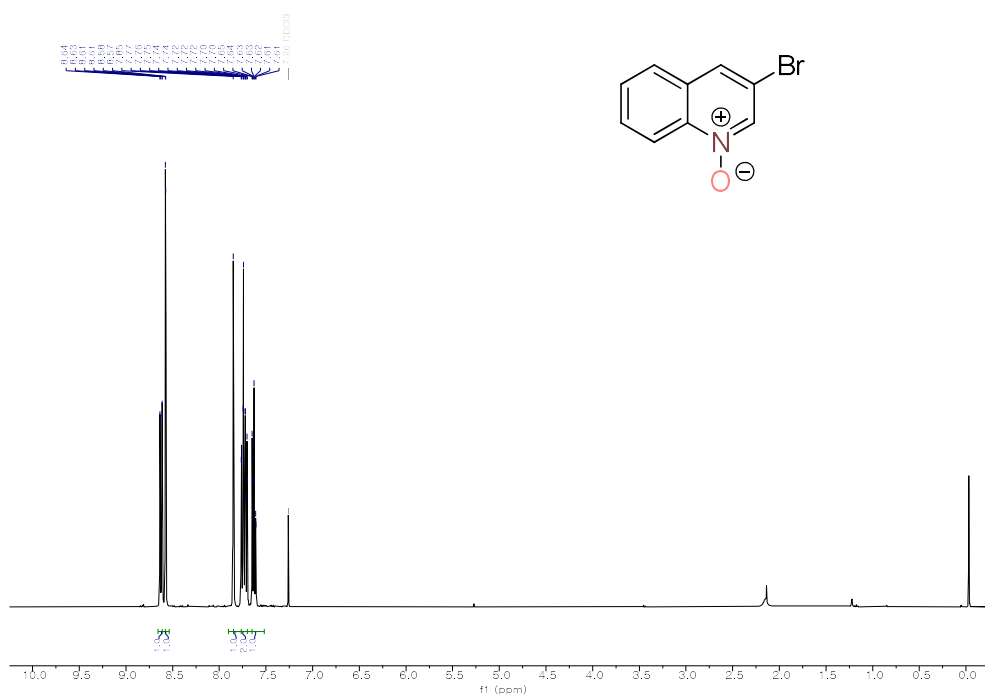
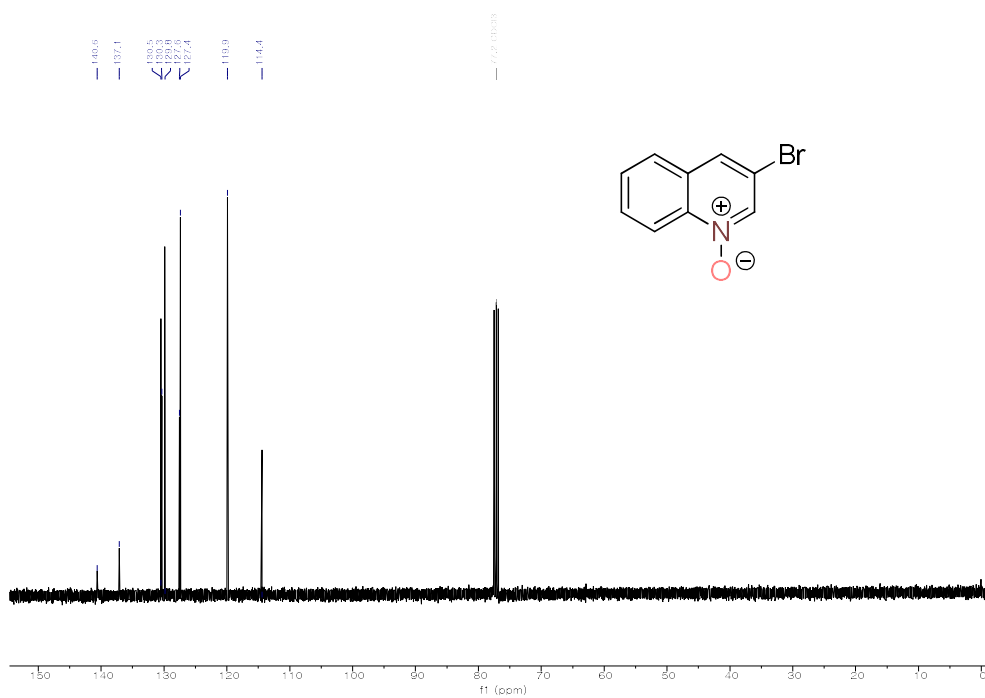
¹H & ¹³C NMR Spectra of 6-Chloroquinoline *N*-Oxide (1o)

¹H NMR (400 MHz, CDCl₃)



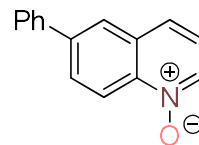
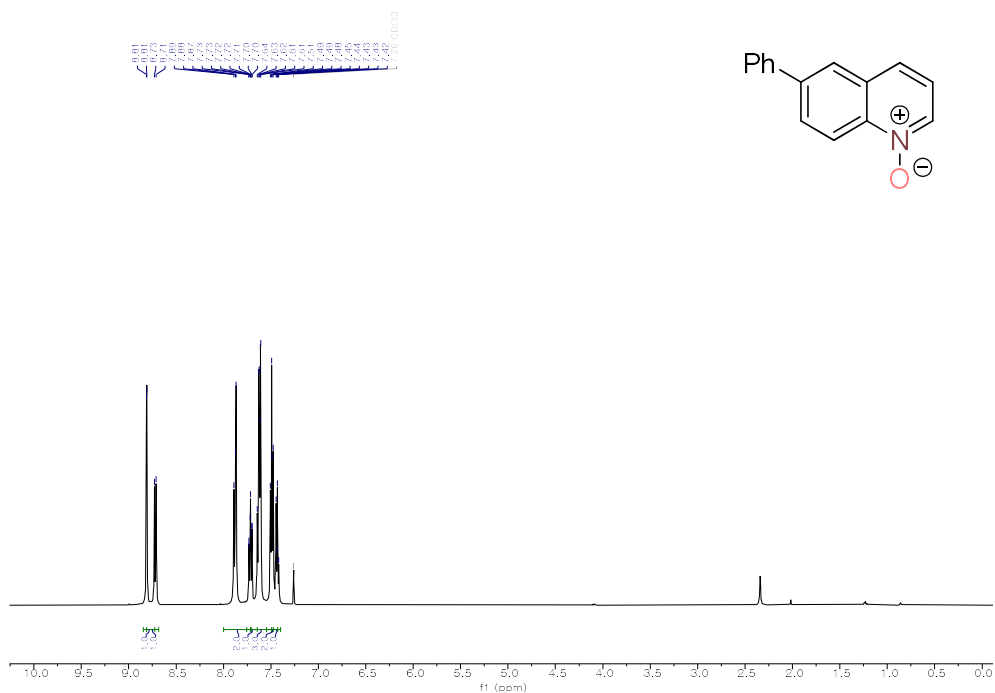
¹³C NMR (101 MHz, CDCl₃)



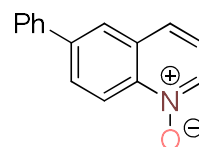
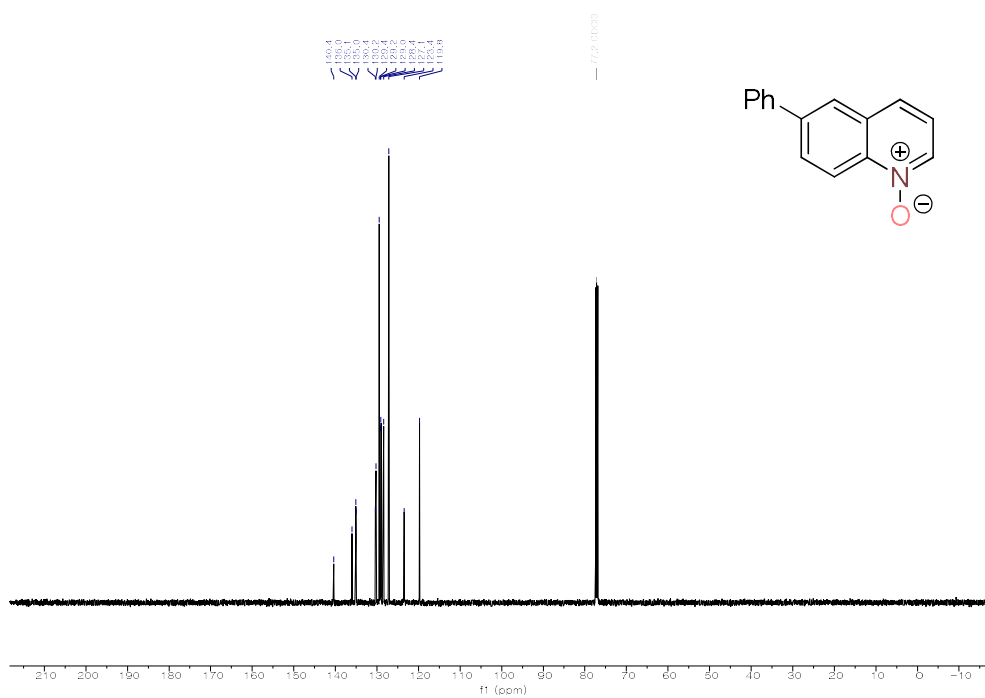
¹H & ¹³C NMR Spectra of 3-Bromoquinoline *N*-Oxide (1r)**¹H NMR (400 MHz, CDCl₃)****¹³C NMR (101 MHz, CDCl₃)**

¹H & ¹³C NMR Spectra of 6-Phenylquinoline *N*-Oxide (1s)

¹H NMR (500 MHz, CDCl₃)

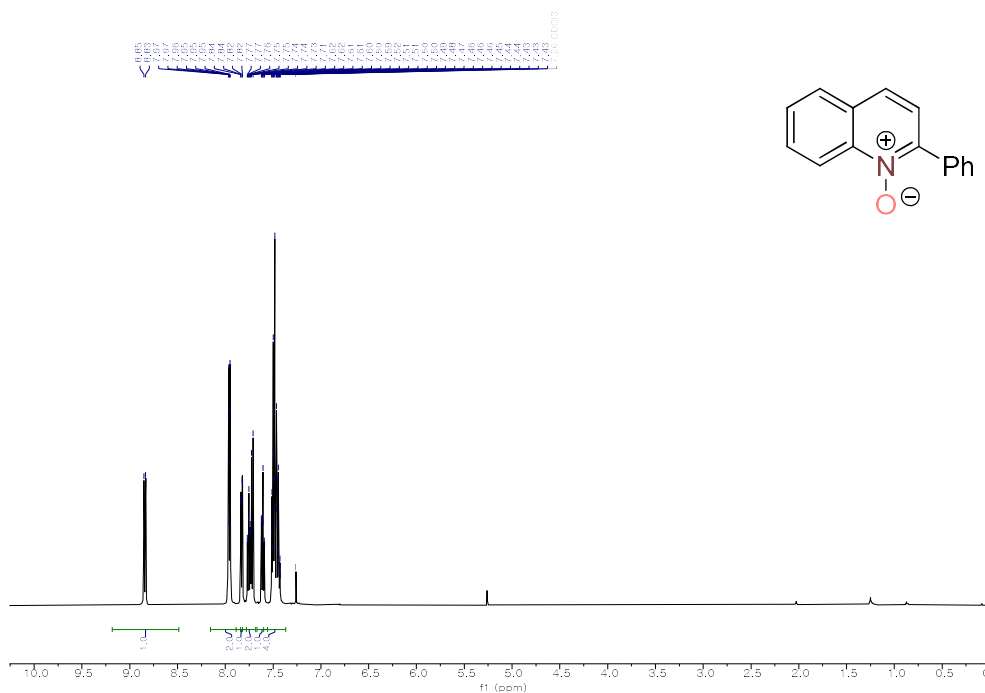


¹³C NMR (126 MHz, CDCl₃)

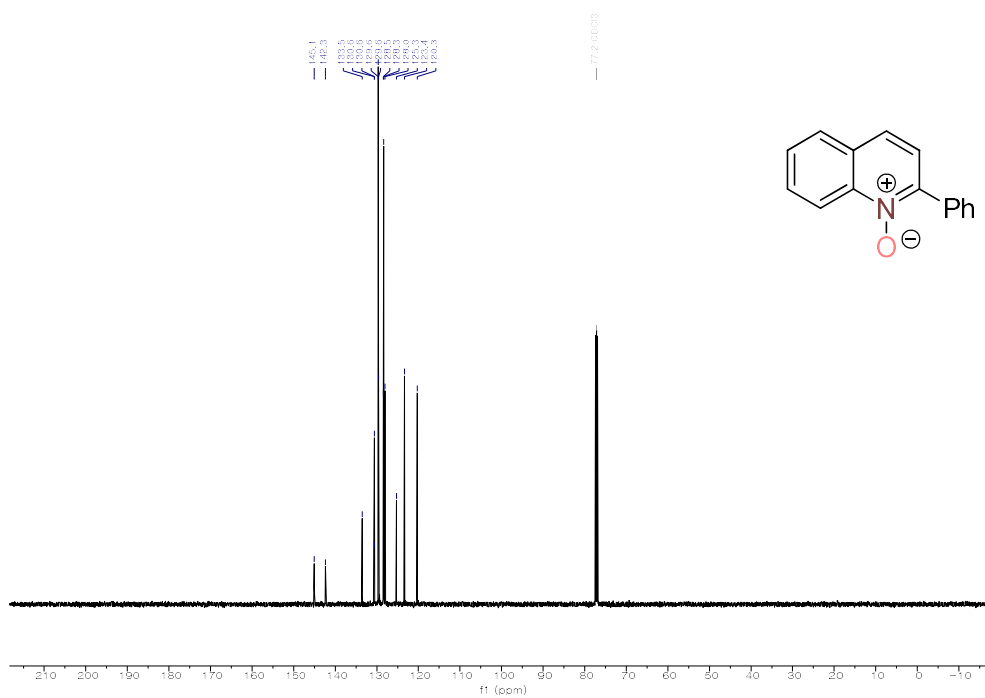


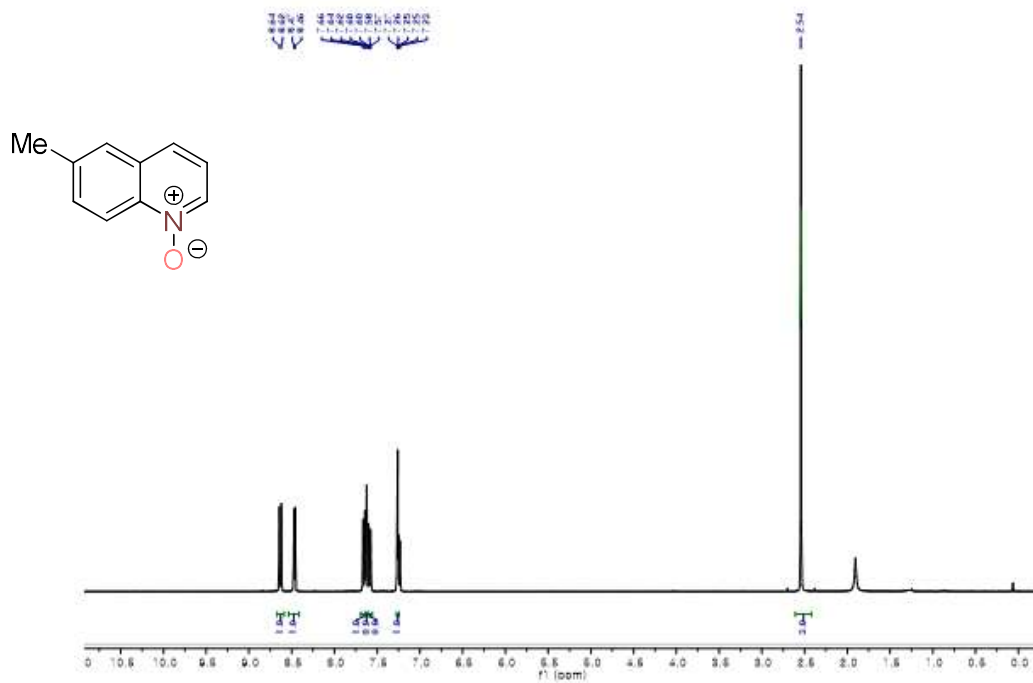
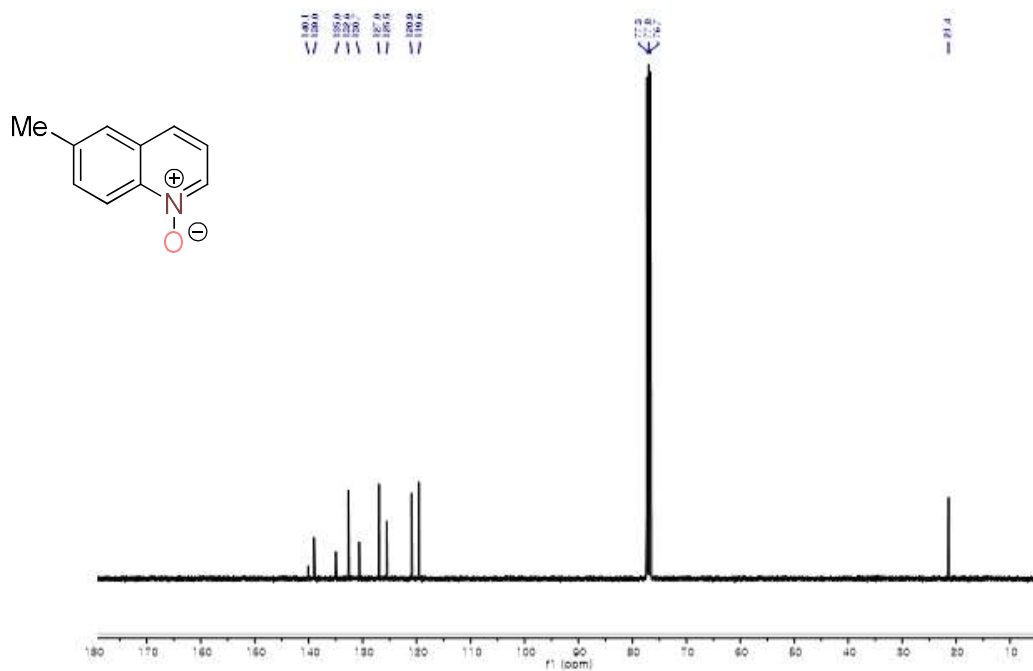
¹H & ¹³C NMR Spectra of 2-Phenylquinoline *N*-Oxide (1t)

¹H NMR (500 MHz, CDCl₃)



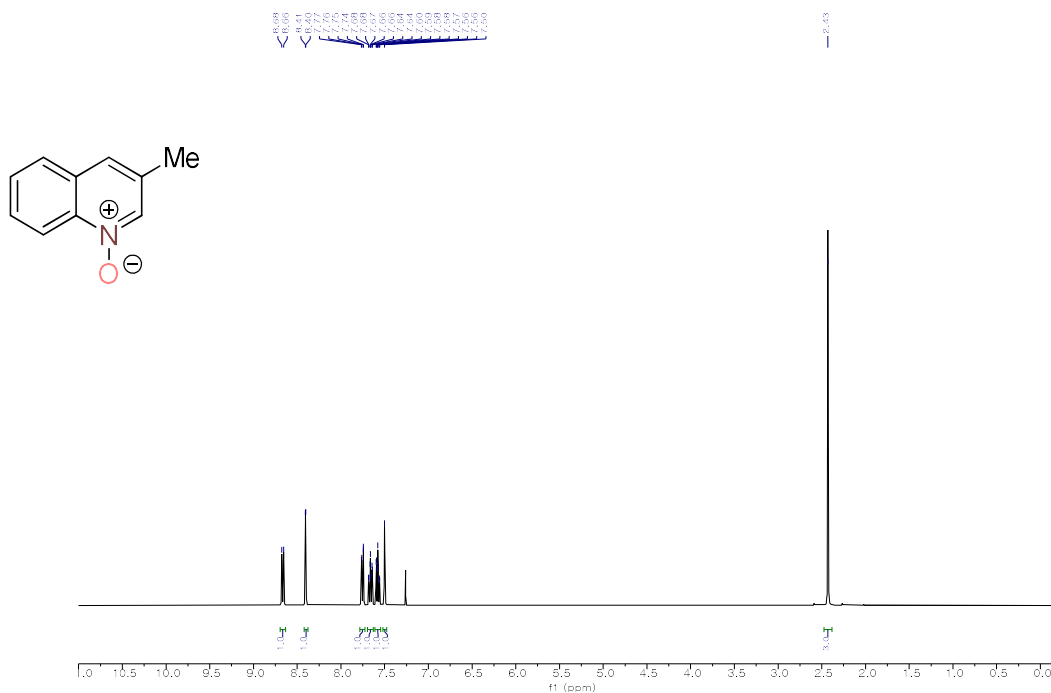
¹³C NMR (126 MHz, CDCl₃)



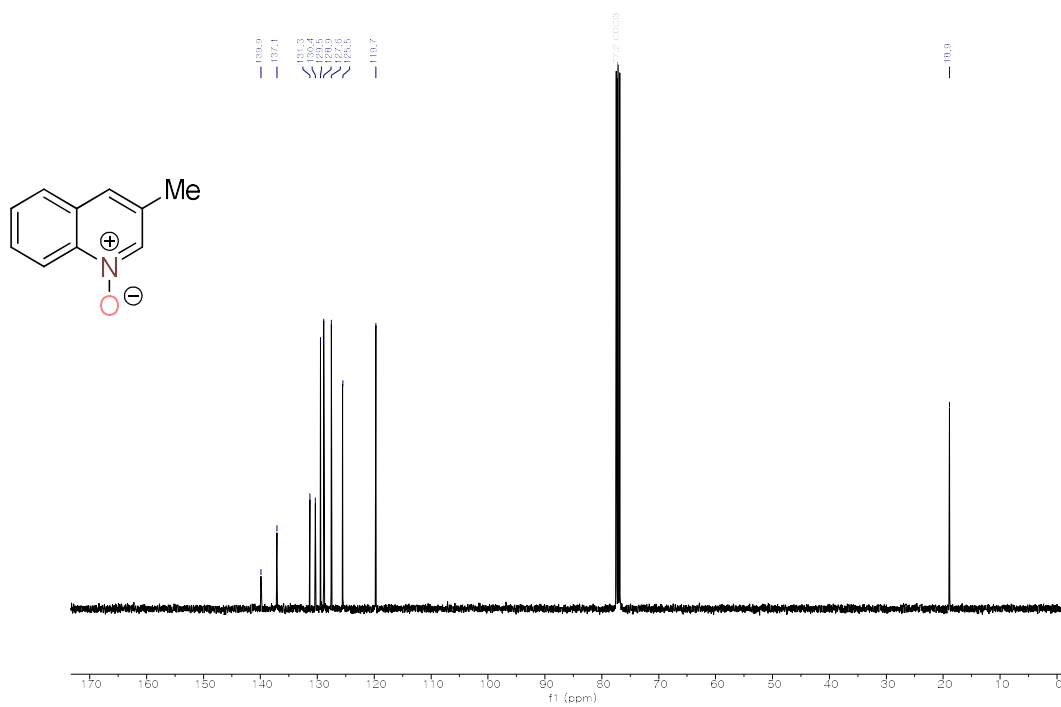
¹H & ¹³C NMR Spectra of 6-Methylquinoline *N*-Oxide (1v)**¹H NMR (400 MHz, CDCl₃)****¹³C NMR (101 MHz, CDCl₃)**

¹H & ¹³C NMR Spectra of 3-Methylquinoline *N*-Oxide (1w)

¹H NMR (400 MHz, CDCl₃)

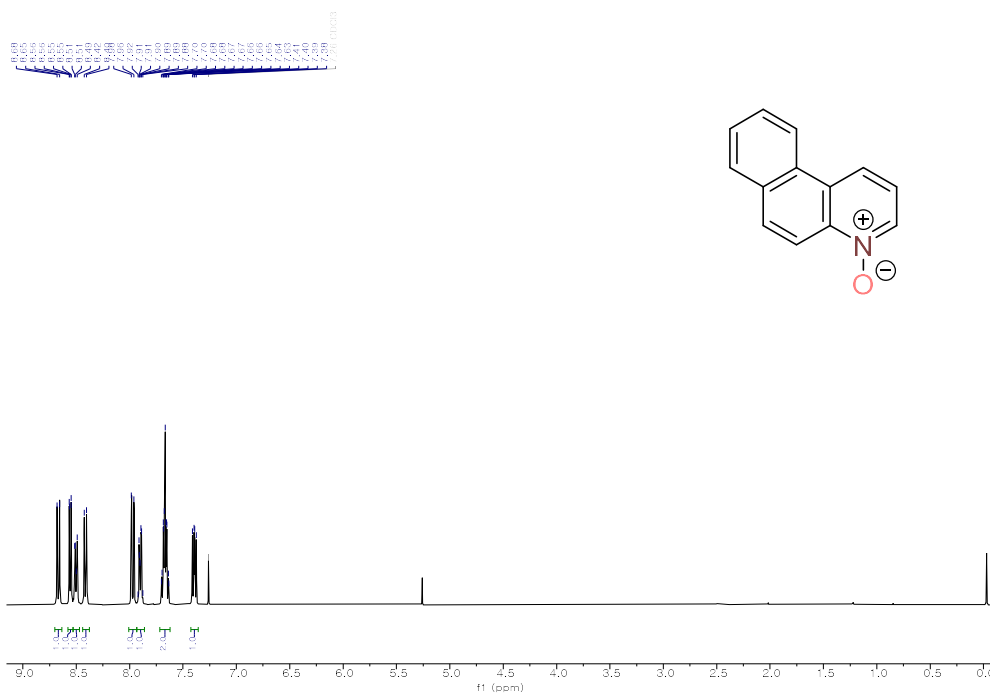


¹³C NMR (101 MHz, CDCl₃)

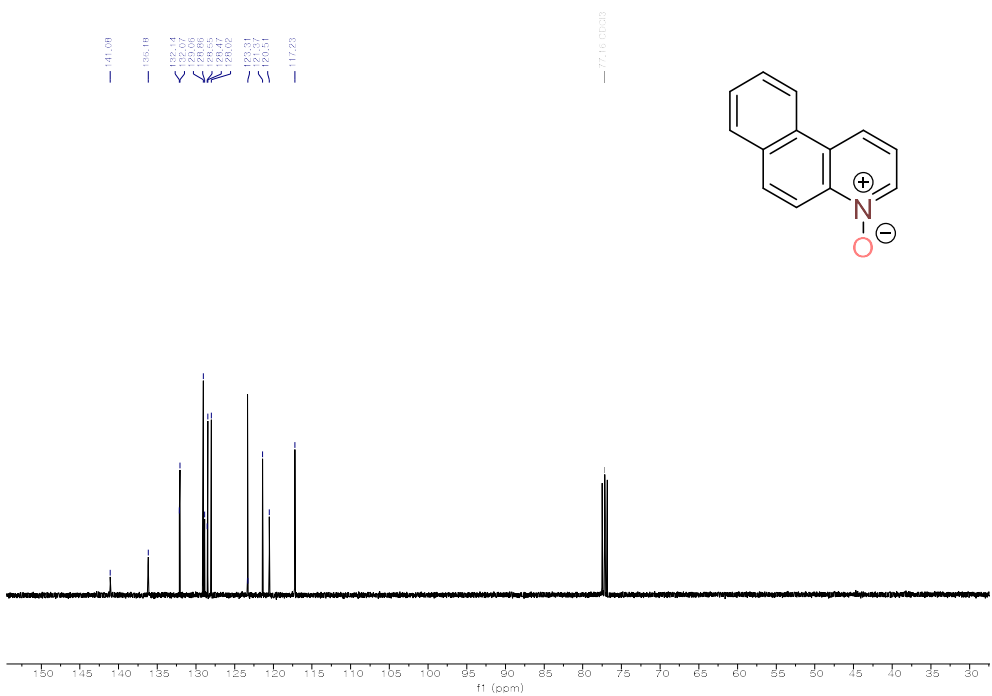


¹H & ¹³C NMR Spectra of Benzo[*f*]quinoline *N*-Oxide (1ab)

¹H NMR (400 MHz, CDCl₃)

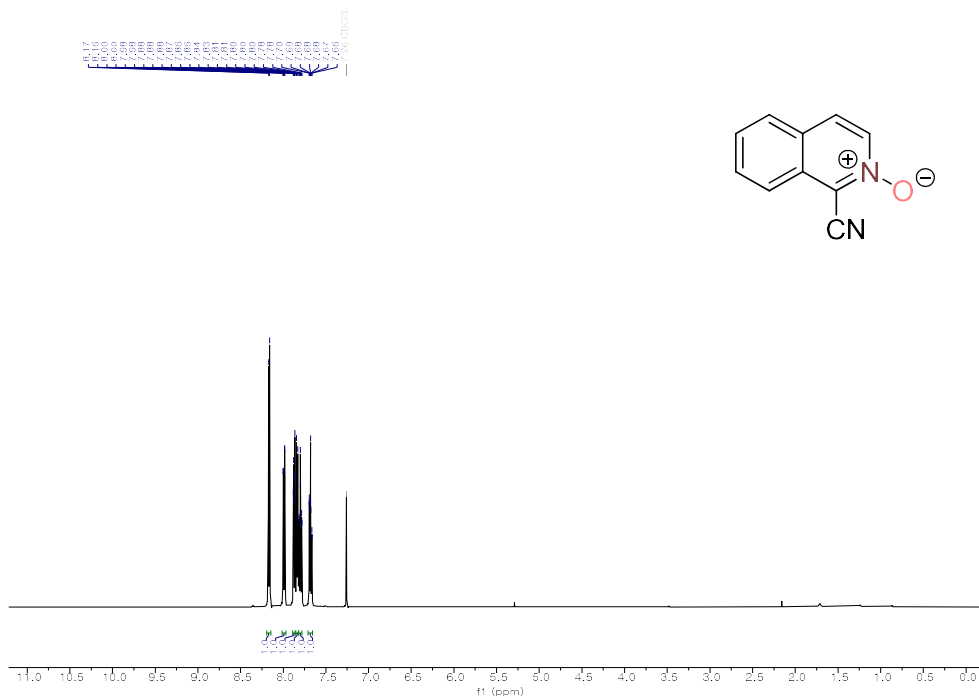


¹³C NMR (101 MHz, CDCl₃)

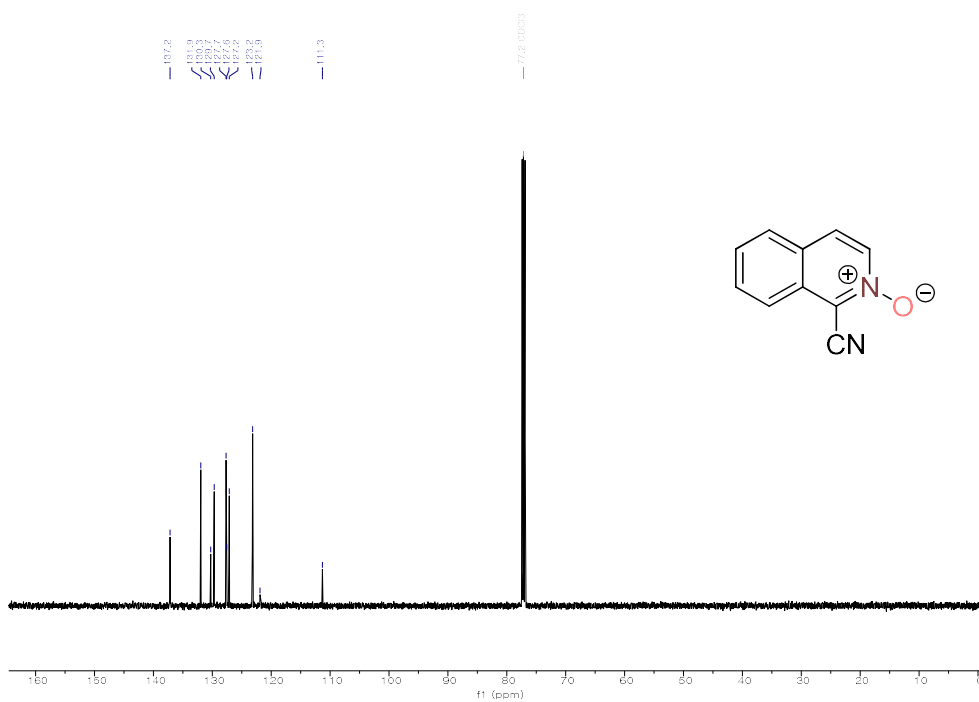


¹H & ¹³C NMR Spectra of 1-Isoquinolinecarbonitrile N-Oxide (1ac)

¹H NMR (500 MHz, CDCl₃)

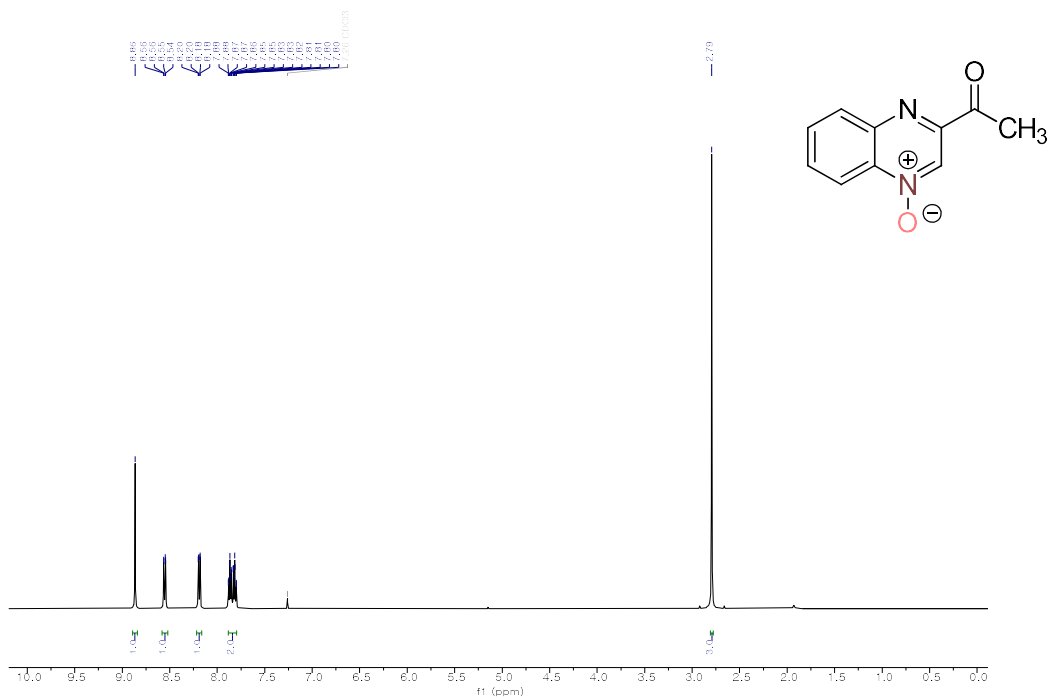


¹³C NMR (126 MHz, CDCl₃)

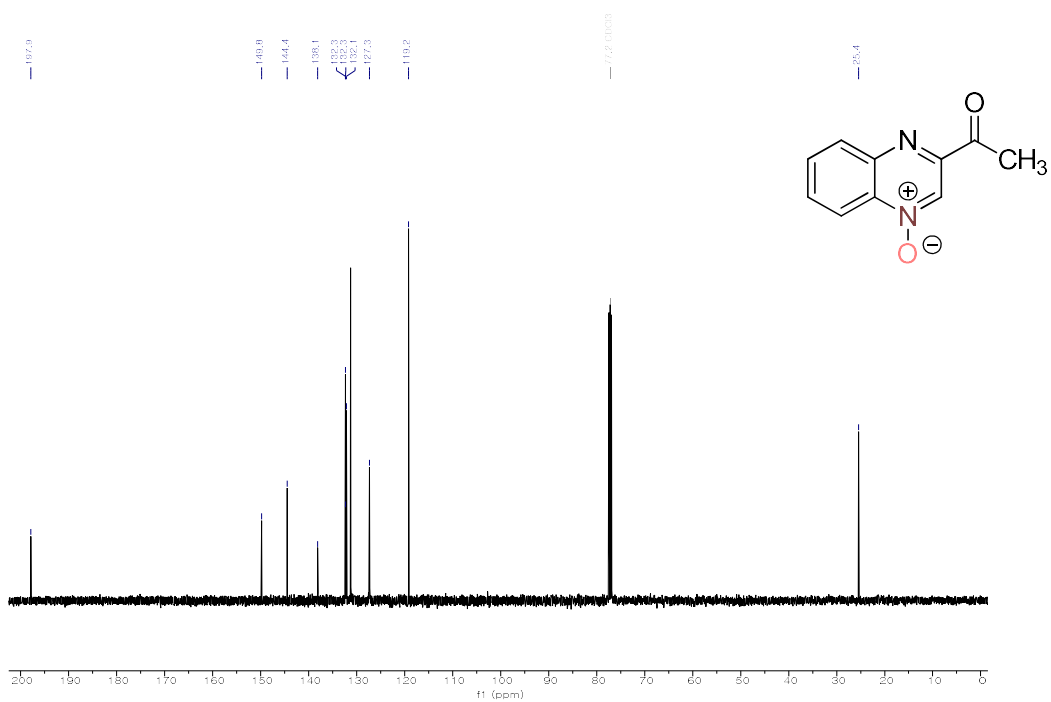


¹H & ¹³C NMR Spectra of Methyl 2-Quinoxalinecarboxylate *N*-Oxide (1ag)

¹H NMR (500 MHz, CDCl₃)

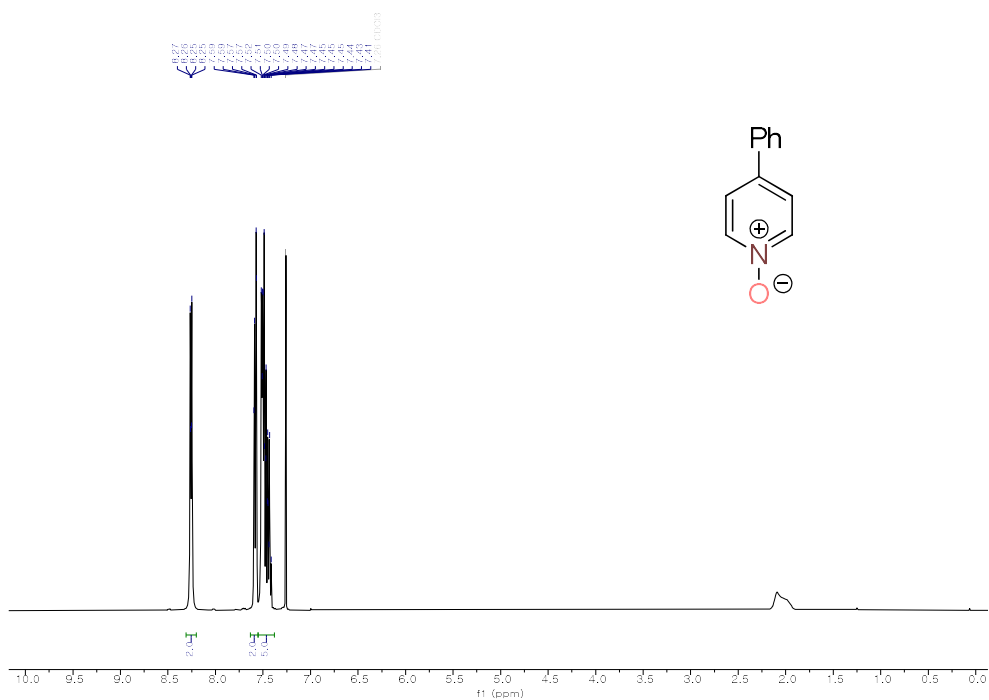


¹³C NMR (126 MHz, CDCl₃)

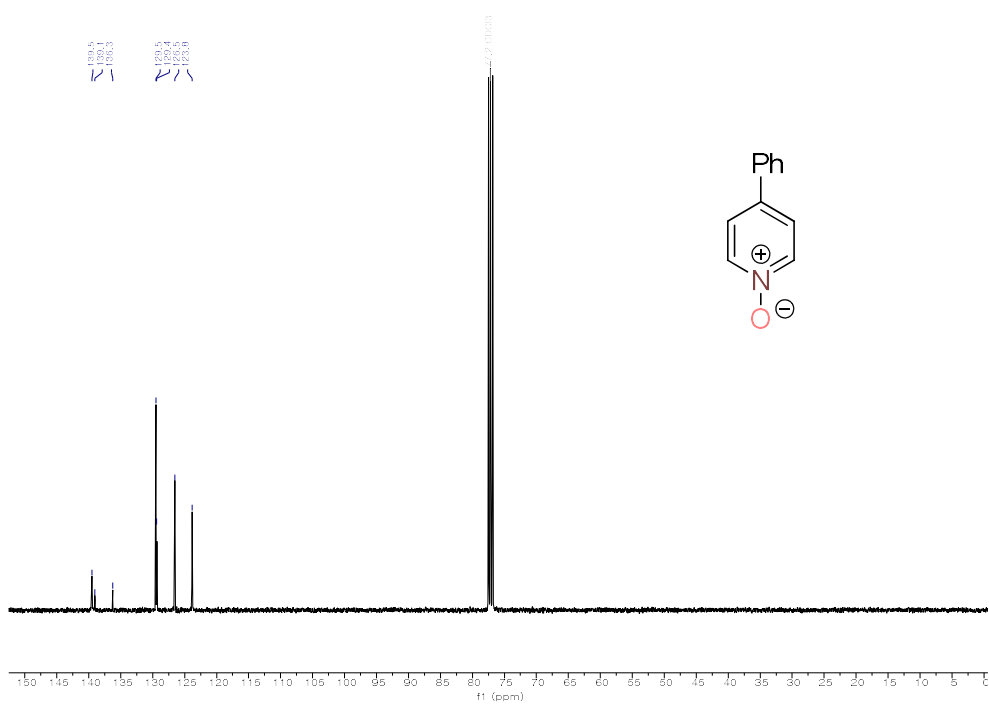


¹H & ¹³C NMR Spectra of 4-Phenylpyridine *N*-Oxide (1ah)

¹H NMR (500 MHz, CDCl₃)

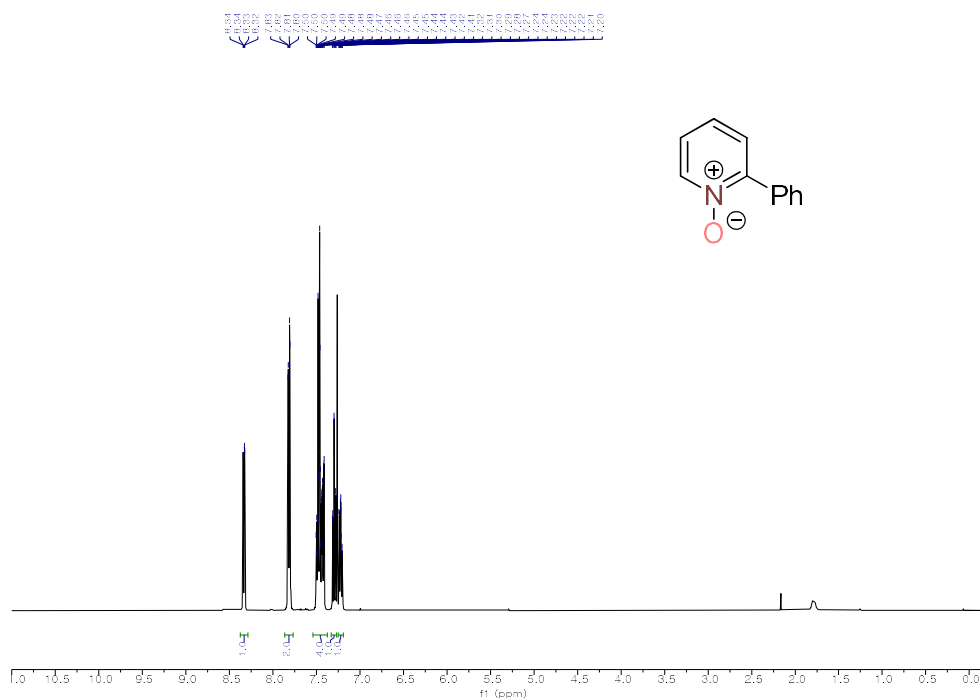


¹³C NMR (101 MHz, CDCl₃)

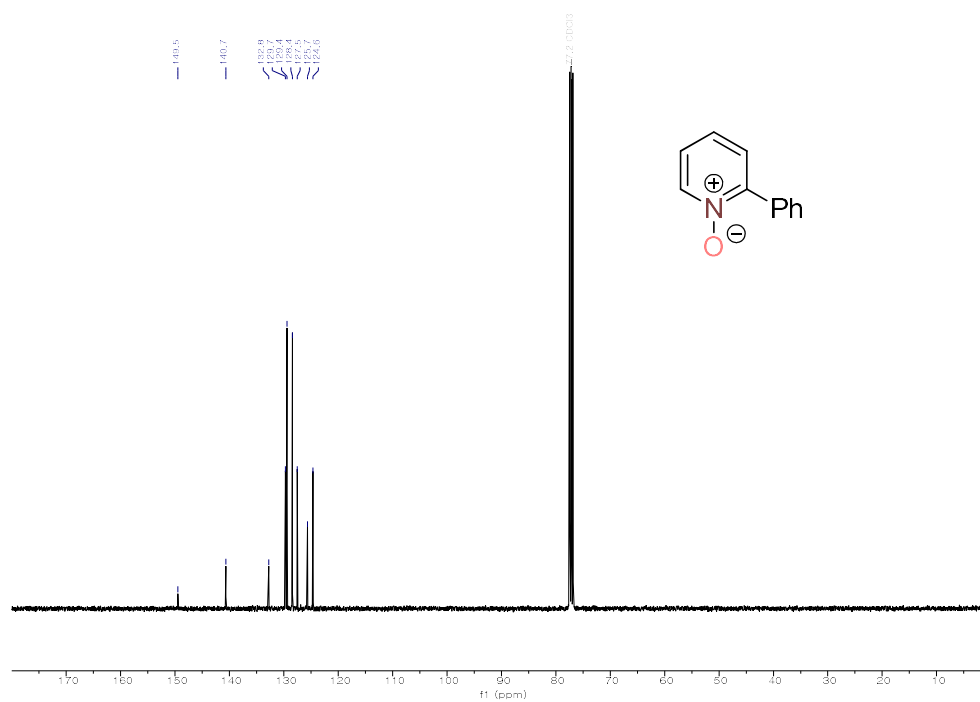


¹H & ¹³C NMR Spectra of 2-Phenylpyridine N-Oxide (1aj)

¹H NMR (400 MHz, CDCl₃)

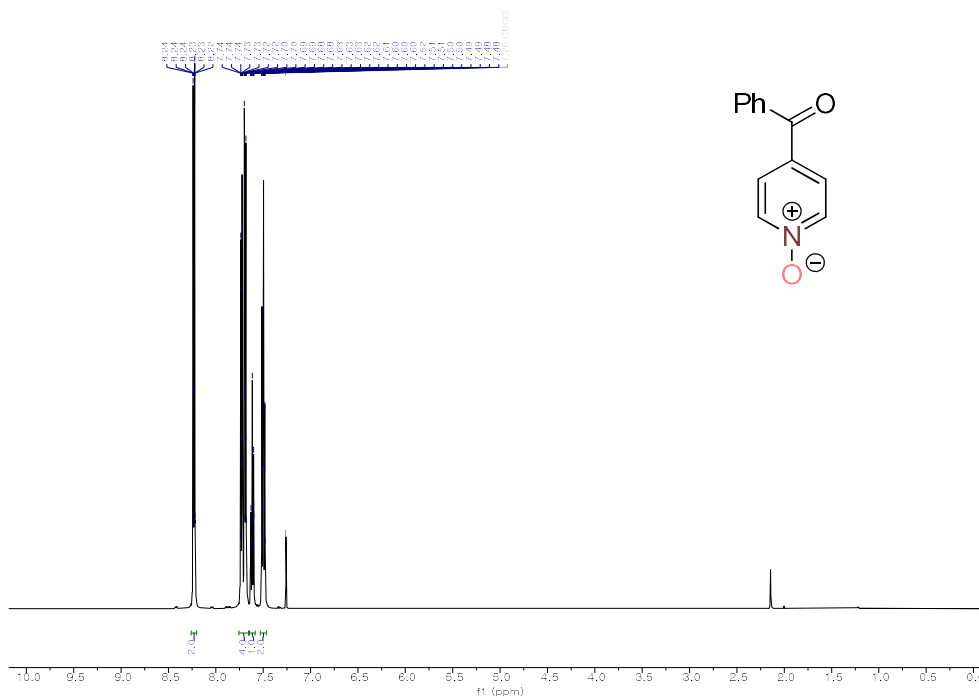


¹³C NMR (101 MHz, CDCl₃)

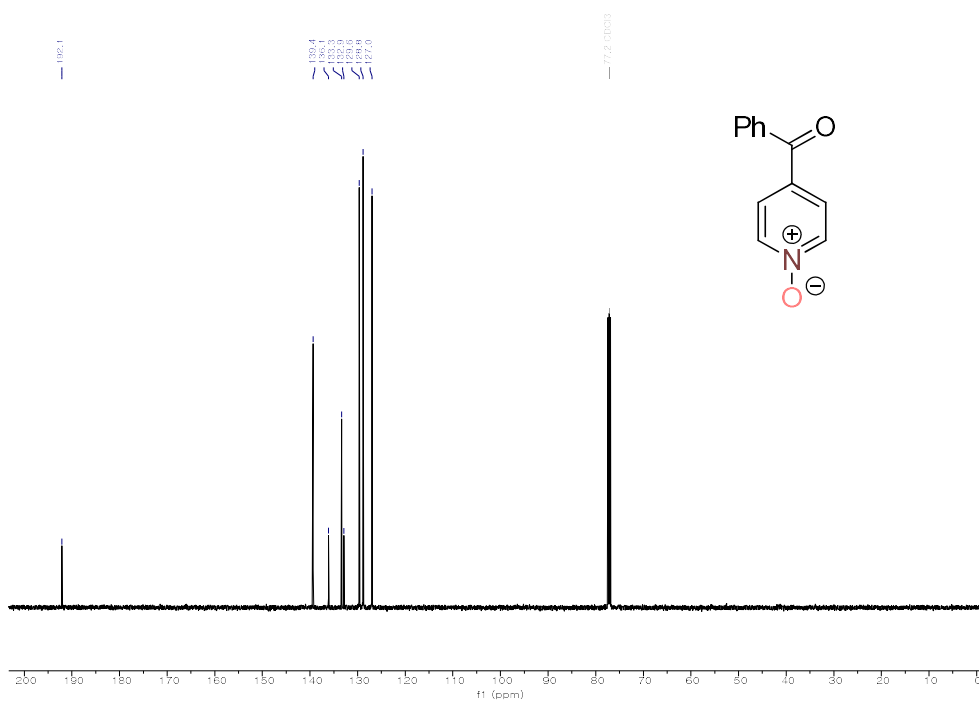


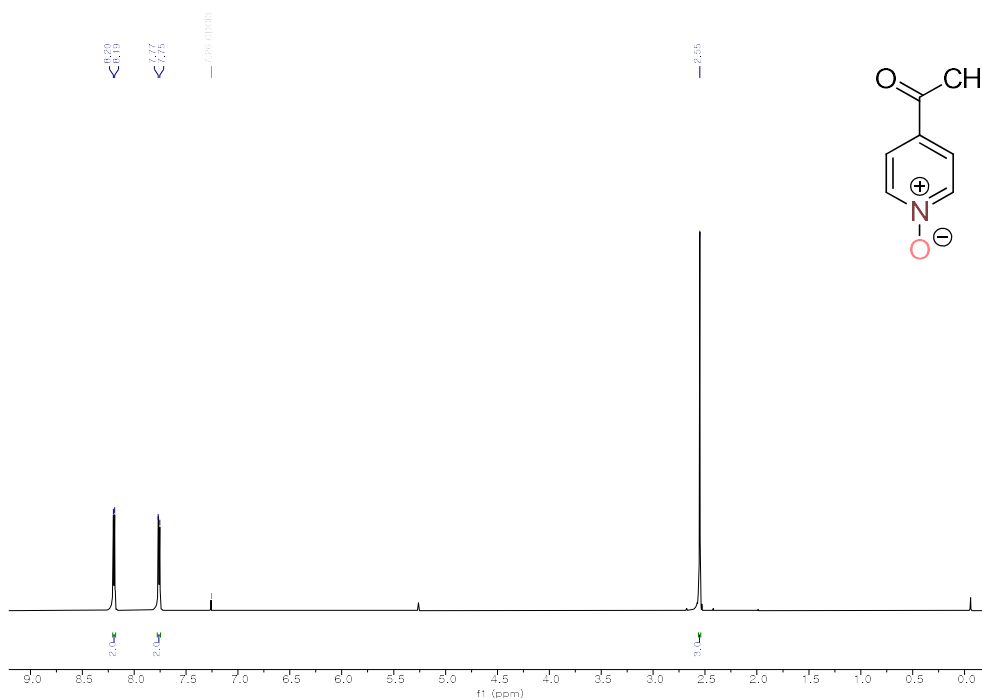
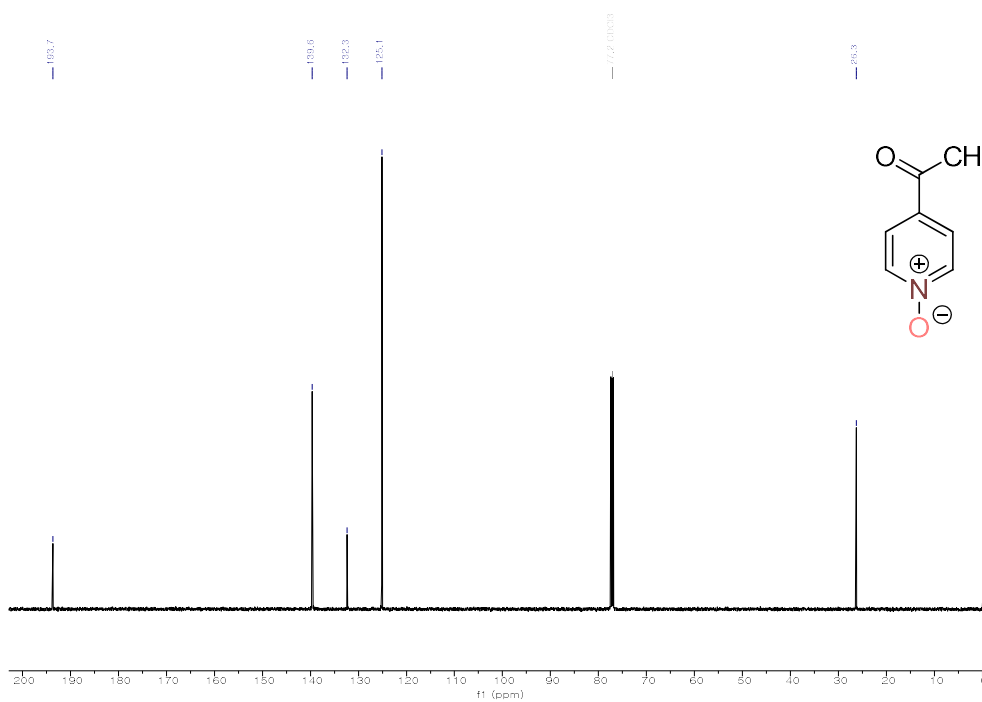
¹H & ¹³C NMR Spectra of 4-Benzoylpyridine *N*-Oxide (1ak)

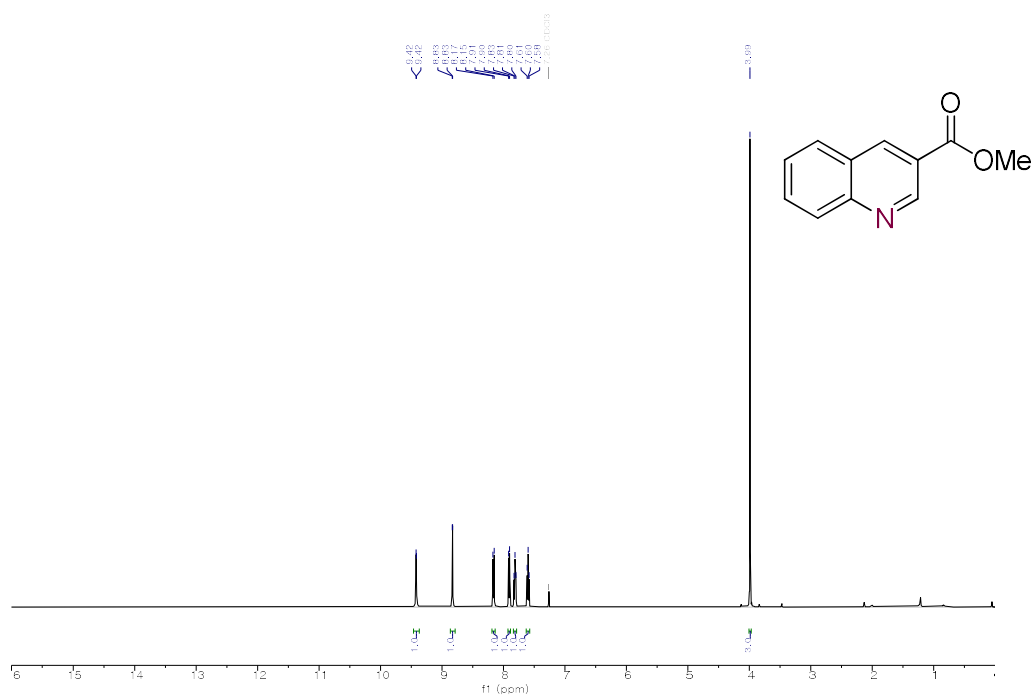
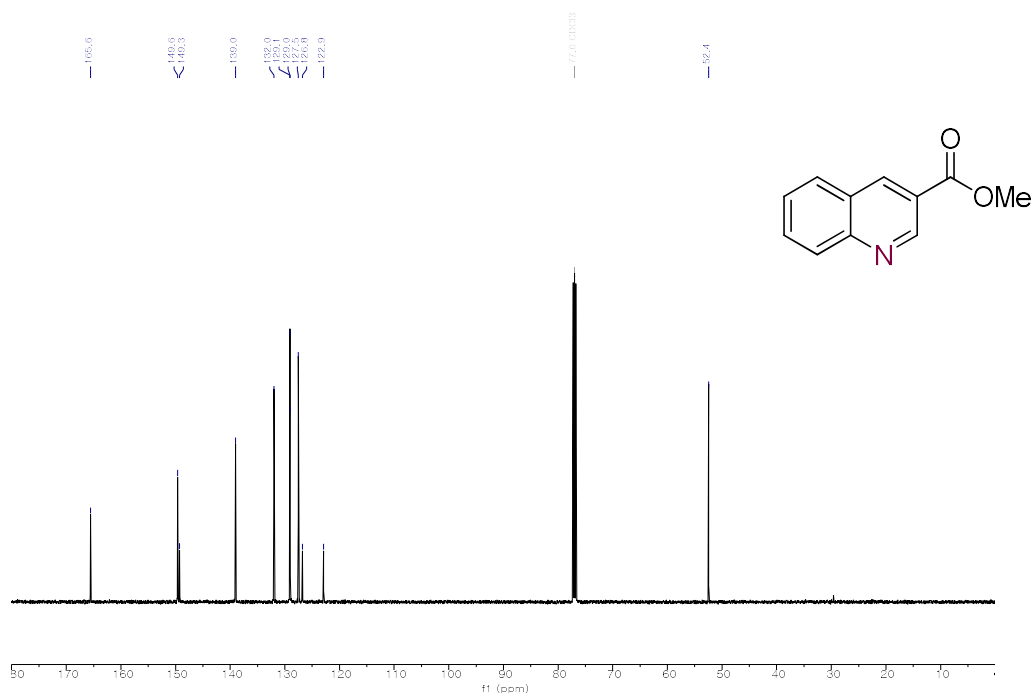
¹H NMR (500 MHz, CDCl₃)



¹³C NMR (126 MHz, CDCl₃)

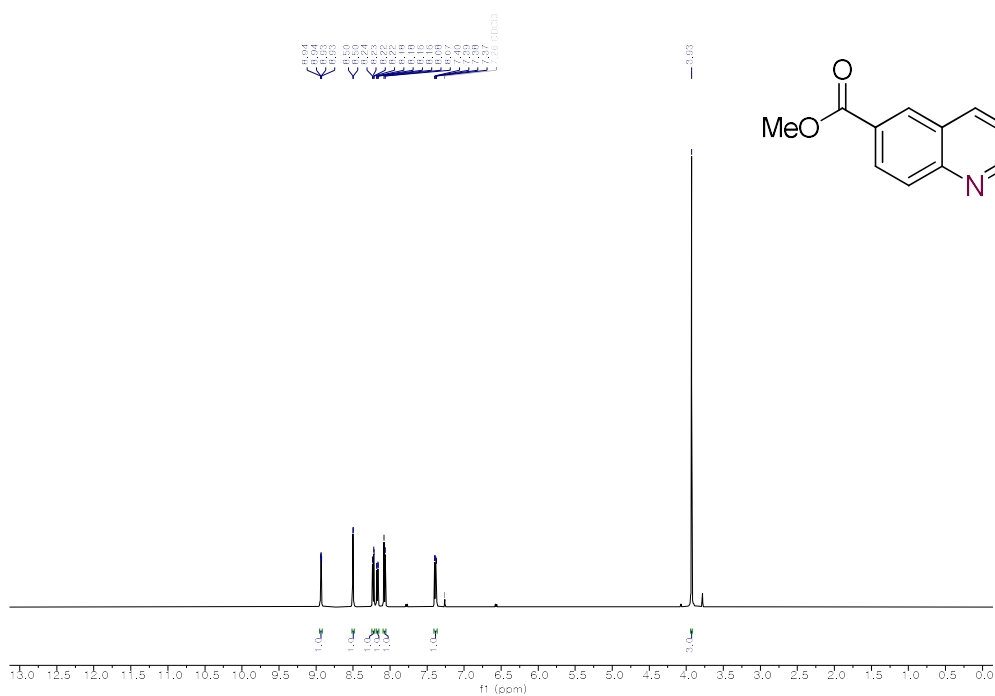


¹H & ¹³C NMR Spectra of 4-Acetylpyridine *N*-Oxide (1an)**¹H NMR (500 MHz, CDCl₃)****¹³C NMR (126 MHz, CDCl₃)**

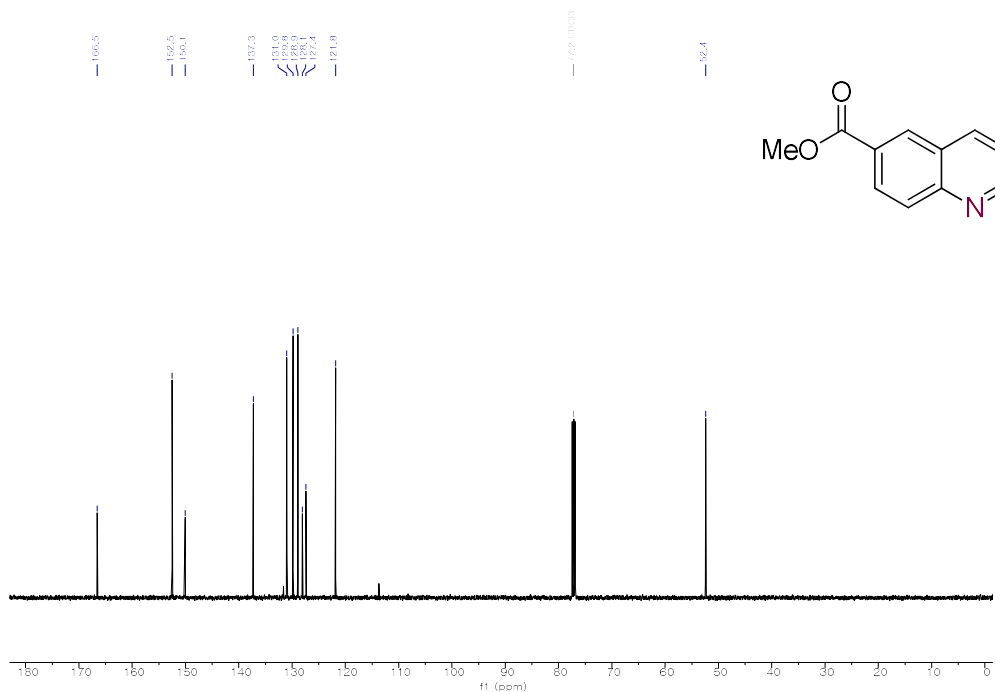
¹H & ¹³C NMR Spectra of Methyl Quinoline-3-carboxylate (2a)**¹H NMR (500 MHz, CDCl₃)****¹³C NMR (126 MHz, CDCl₃)**

¹H & ¹³C NMR Spectra of Methyl Quinoline-6-carboxylate (2b)

¹H NMR (500 MHz, CDCl₃)

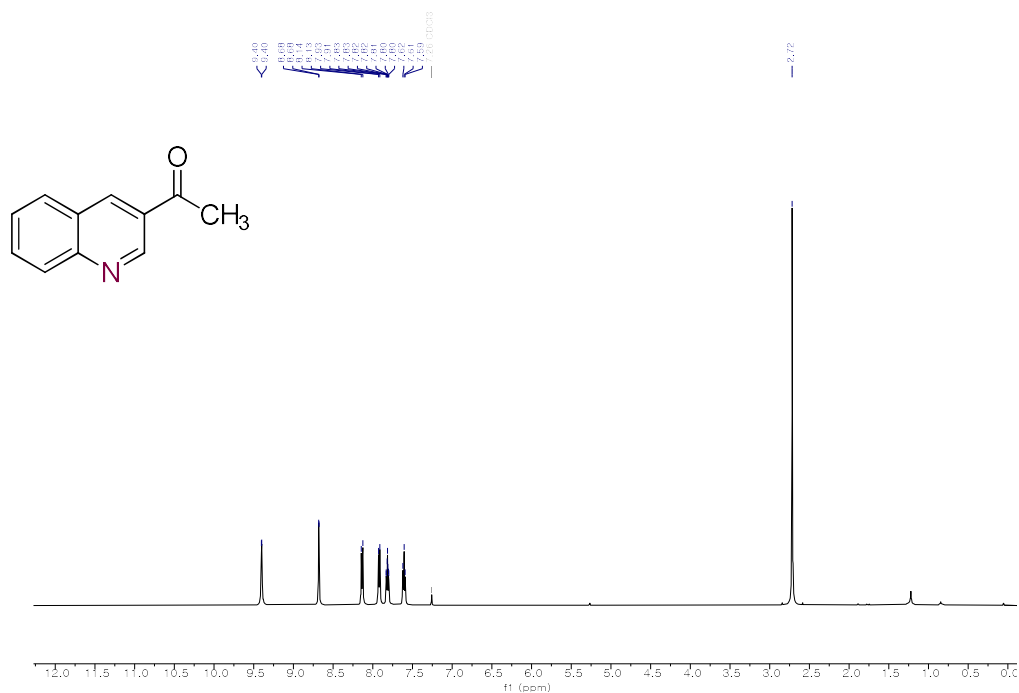


¹³C NMR (126 MHz, CDCl₃) δ

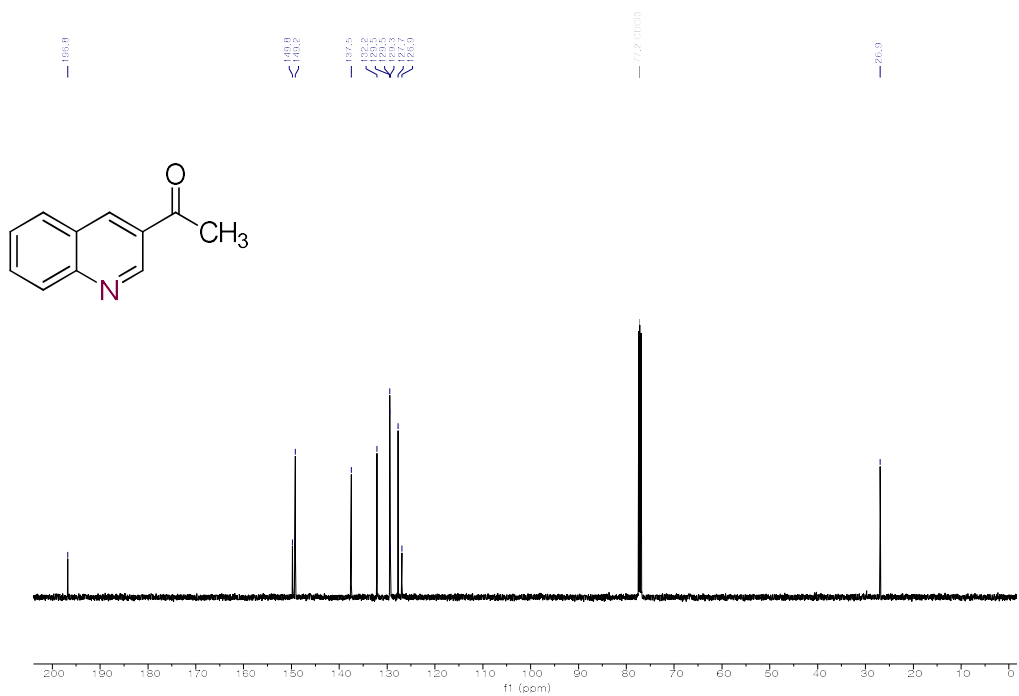


¹H & ¹³C NMR Spectra of 3-Acetylquinoline (2f)

¹H NMR (500 MHz, CDCl₃)

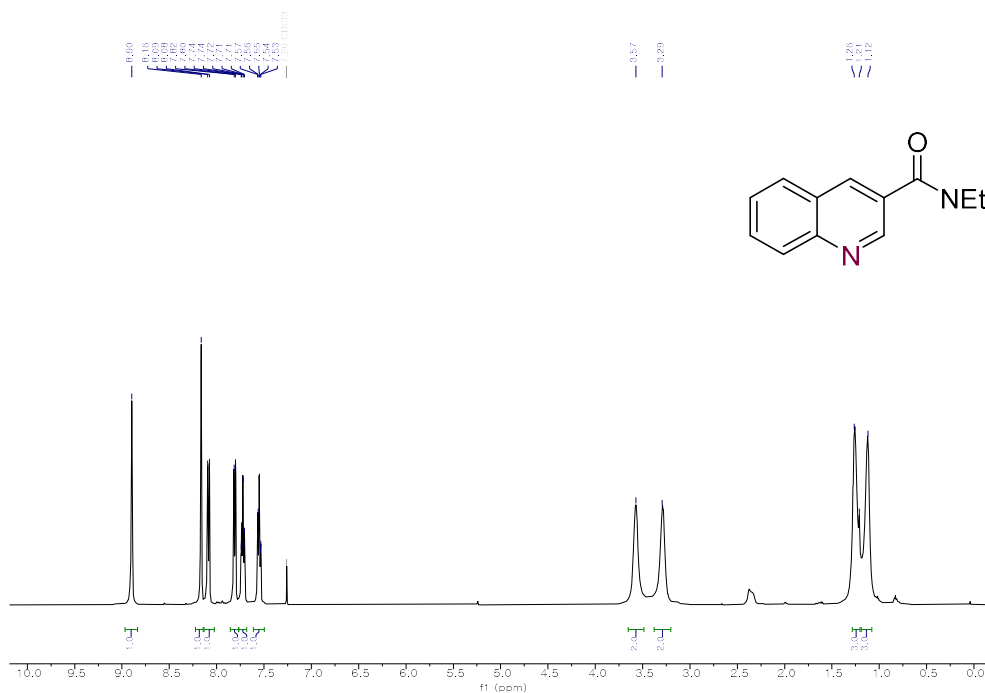


¹³C NMR (126 MHz, CDCl₃)

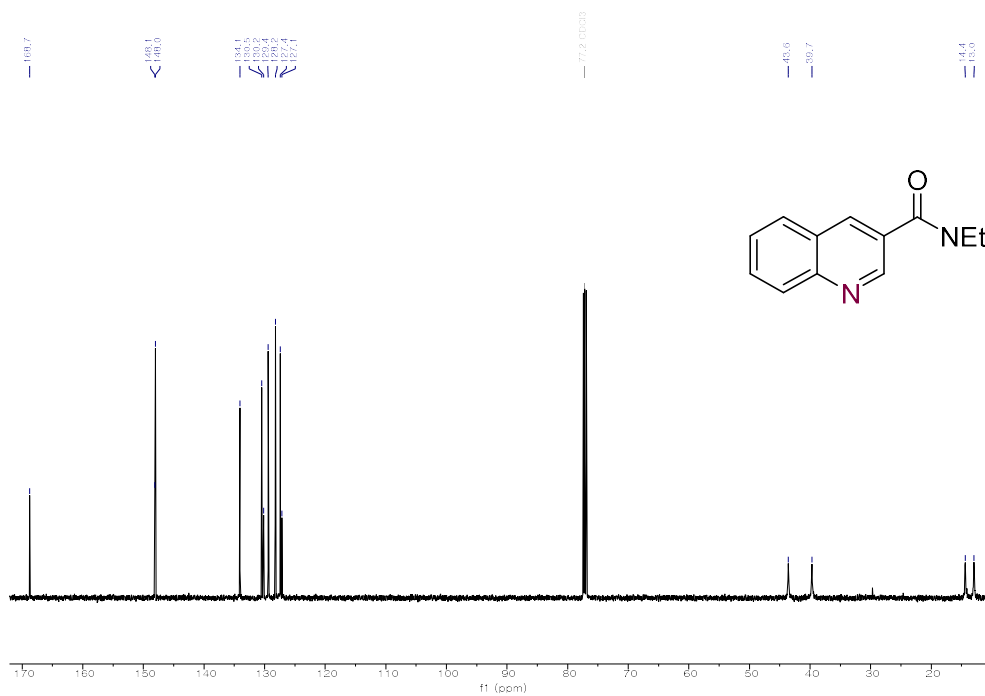


¹H & ¹³C NMR Spectra of *N,N*-Diethyl-3-quinolinecarboxamide (2g)

¹H NMR (500 MHz, CDCl₃)

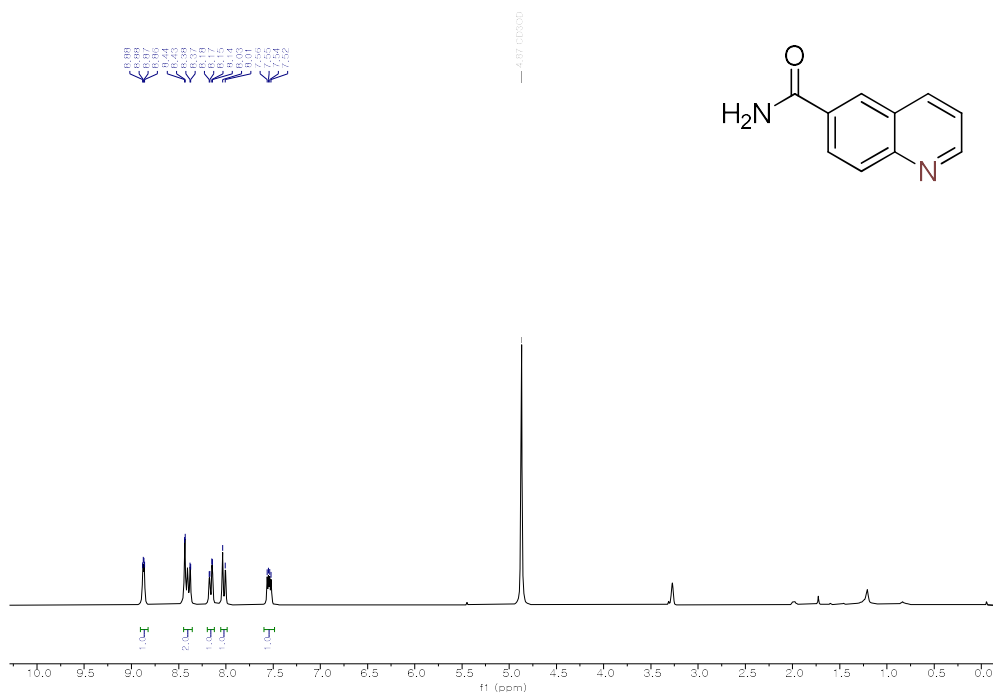


¹³C NMR (126 MHz, CDCl₃)

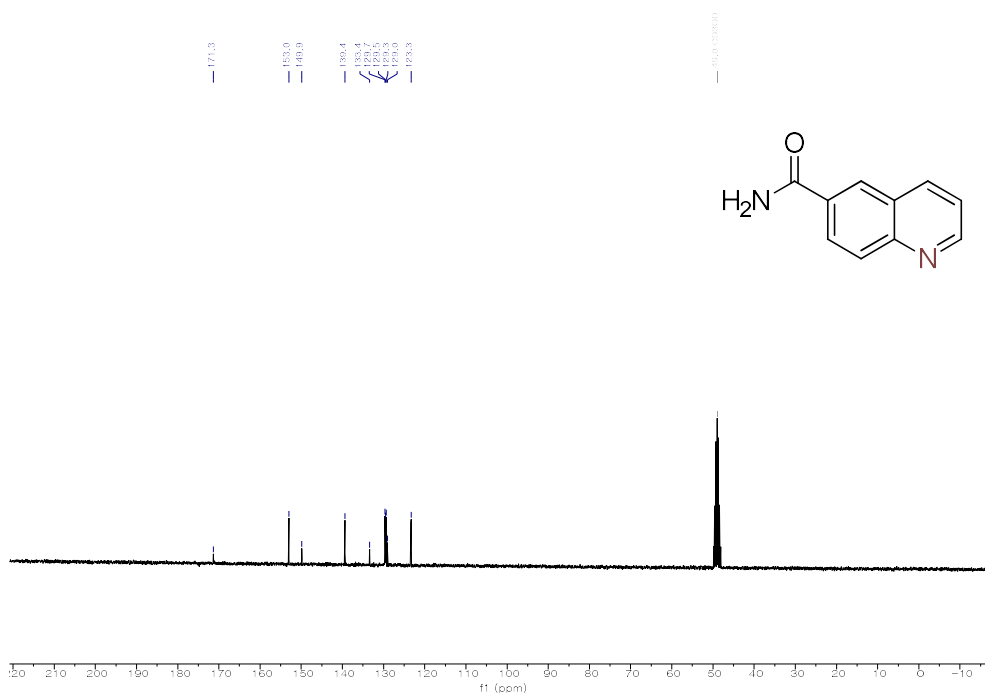


¹H & ¹³C NMR Spectra of 6-Quinolinecarboxamide (2i)

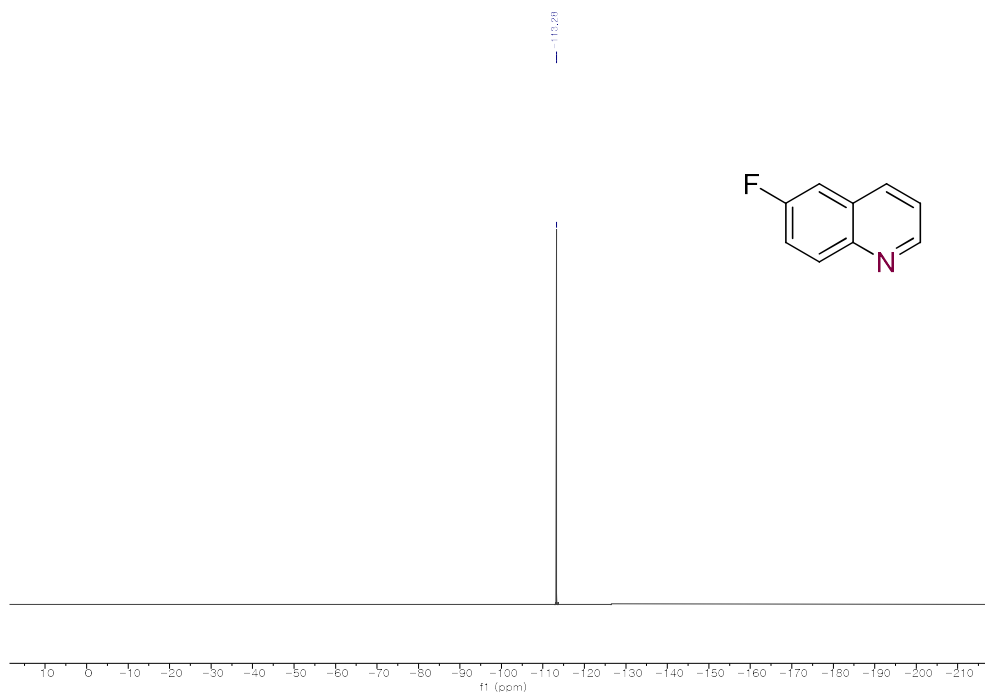
¹H NMR (300 MHz, MeOD)



¹³C NMR (75 MHz, MeOD)

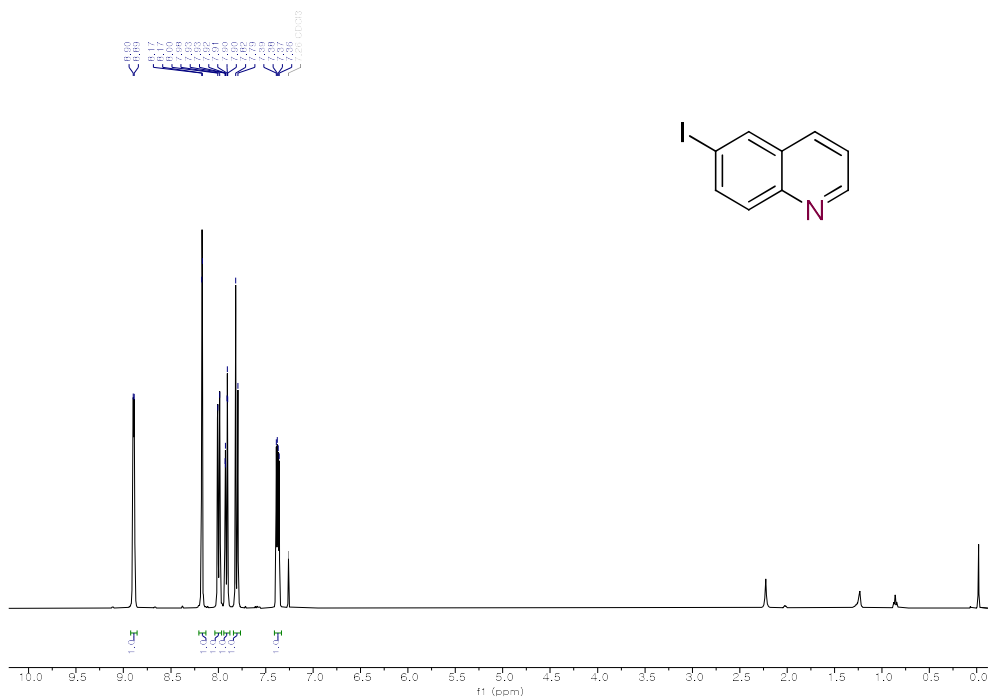


¹⁹F NMR (376 MHz, CDCl₃)

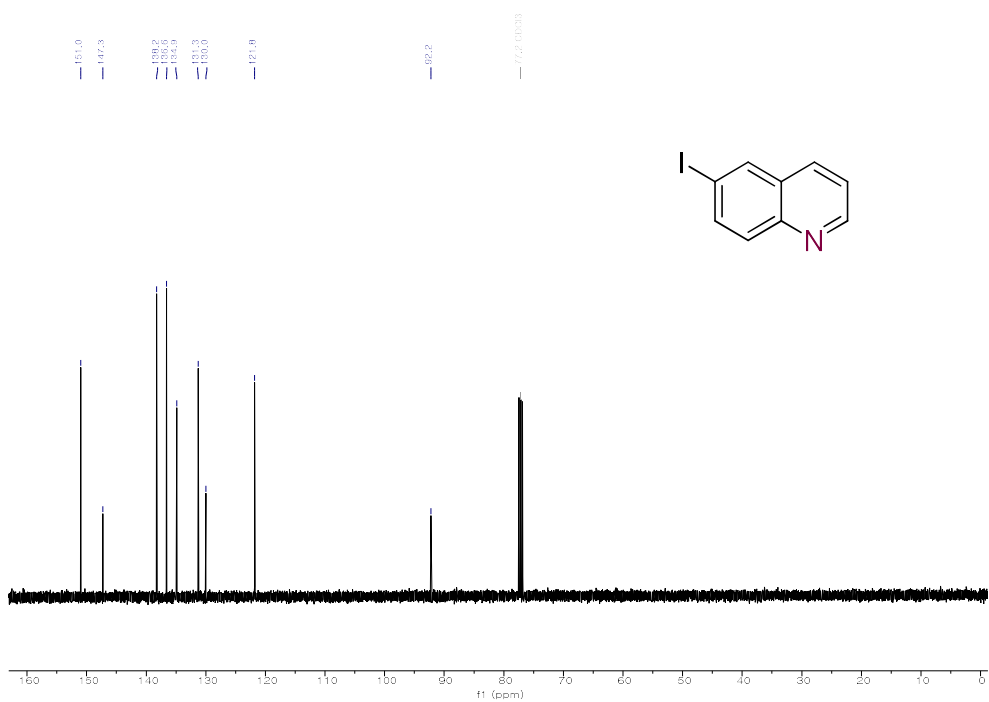


¹H & ¹³C NMR Spectra of 6-Iodoquinoline (2q)

¹H NMR (400 MHz, CDCl₃)

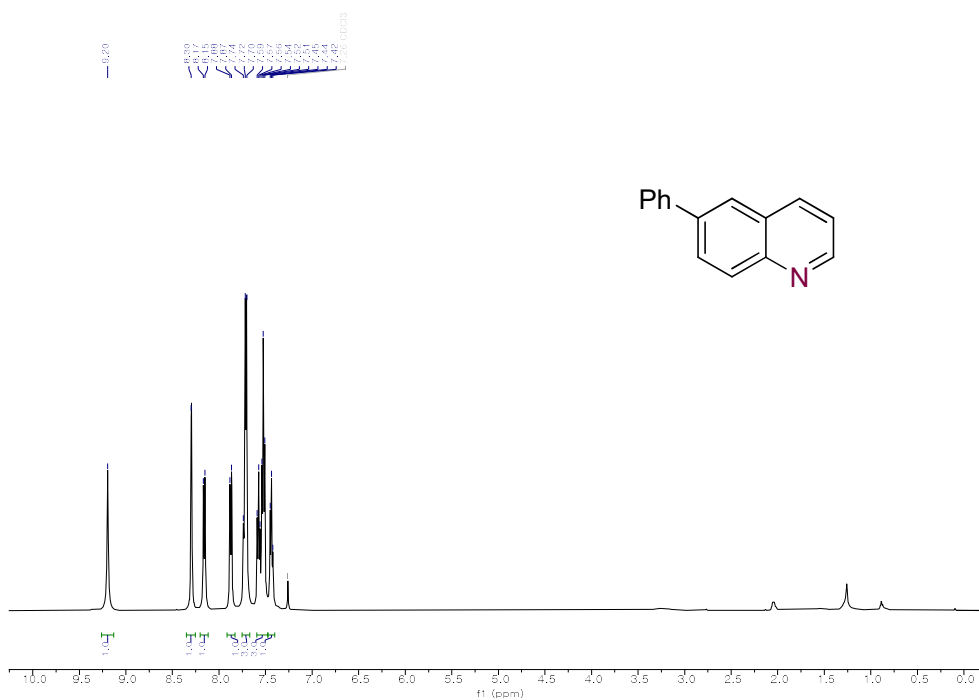


¹³C NMR (101 MHz, CDCl₃)



¹H & ¹³C NMR Spectra of 6-Phenylquinoline (2s)

¹H NMR (500 MHz, CDCl₃)

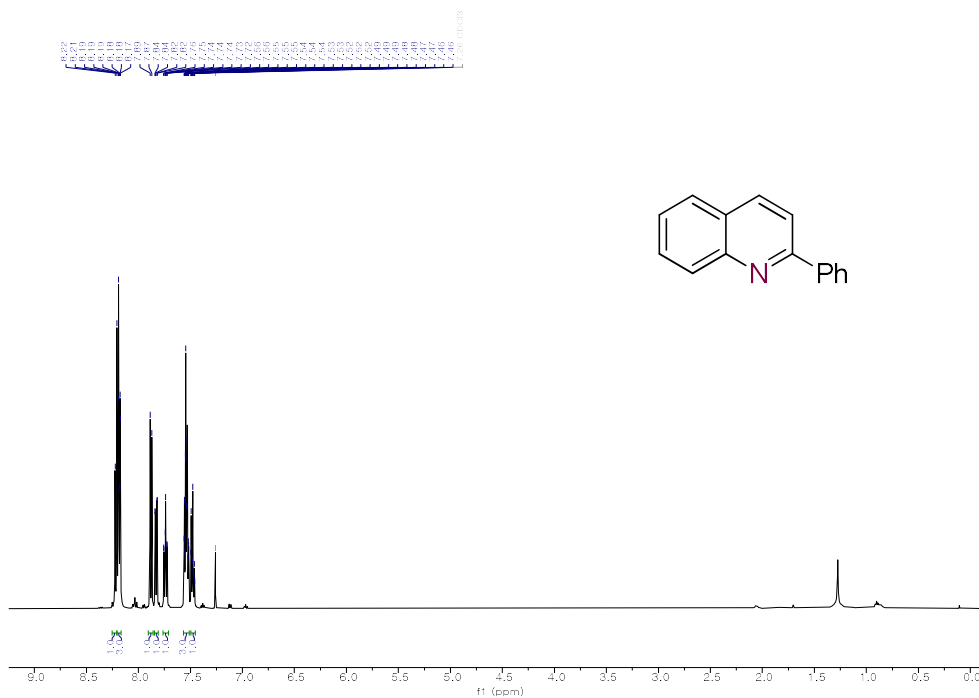


¹³C NMR (126 MHz, CDCl₃)



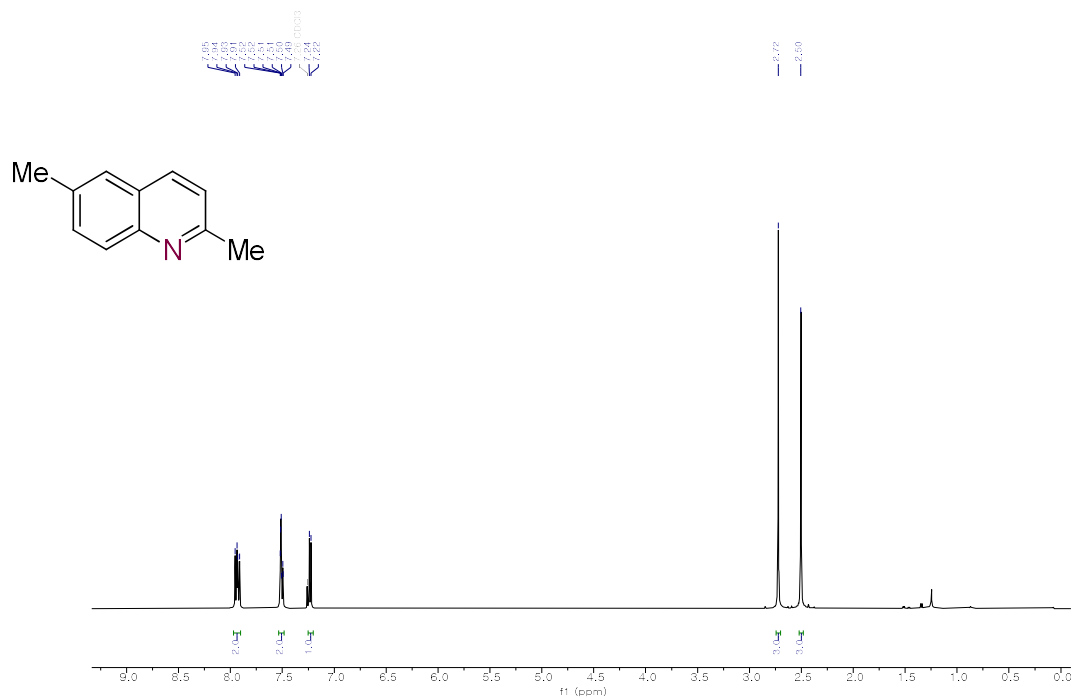
¹H & ¹³C NMR Spectra of 2-Phenylquinoline (2t)

¹H NMR (500 MHz, CDCl₃)

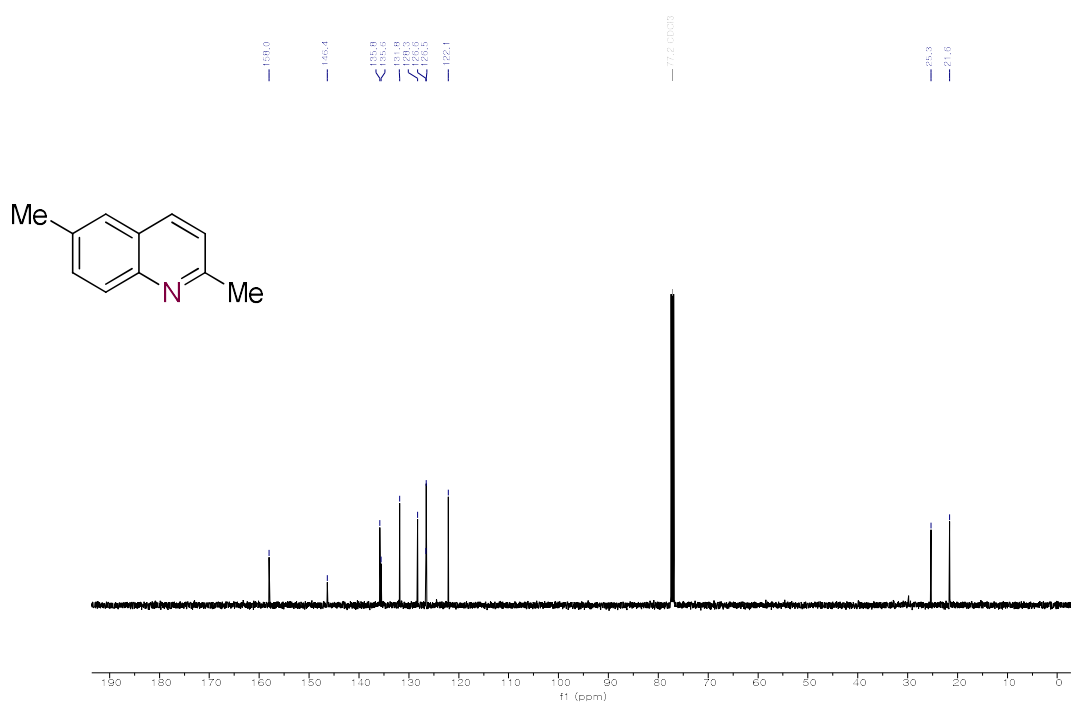


¹H & ¹³C NMR Spectra of 2,6-Dimethylquinoline (2u)

¹H NMR (500 MHz, CDCl₃)

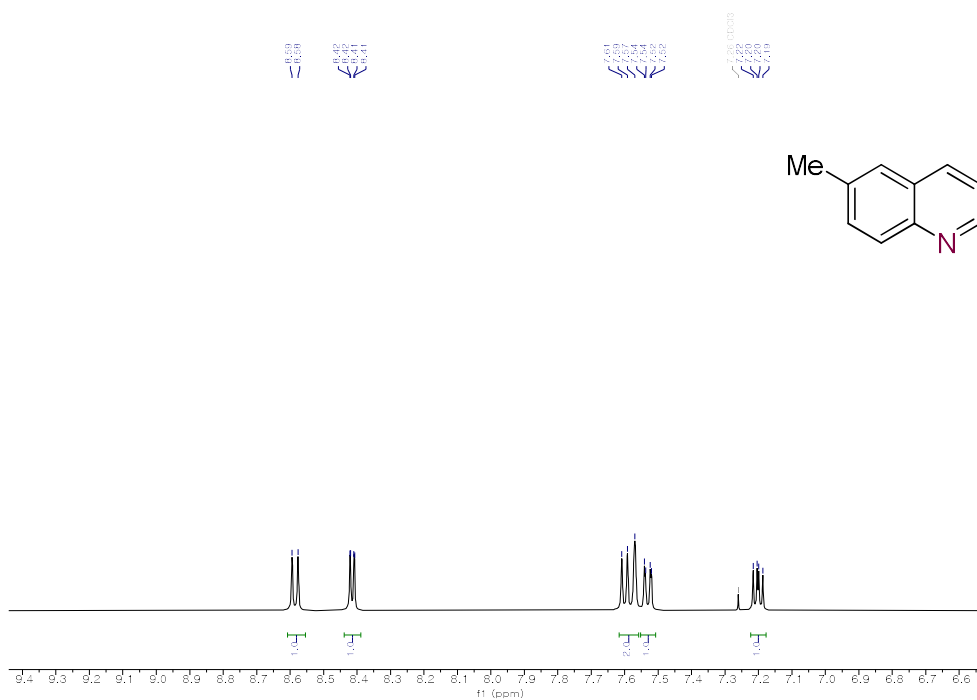


¹³C NMR (126 MHz, CDCl₃)

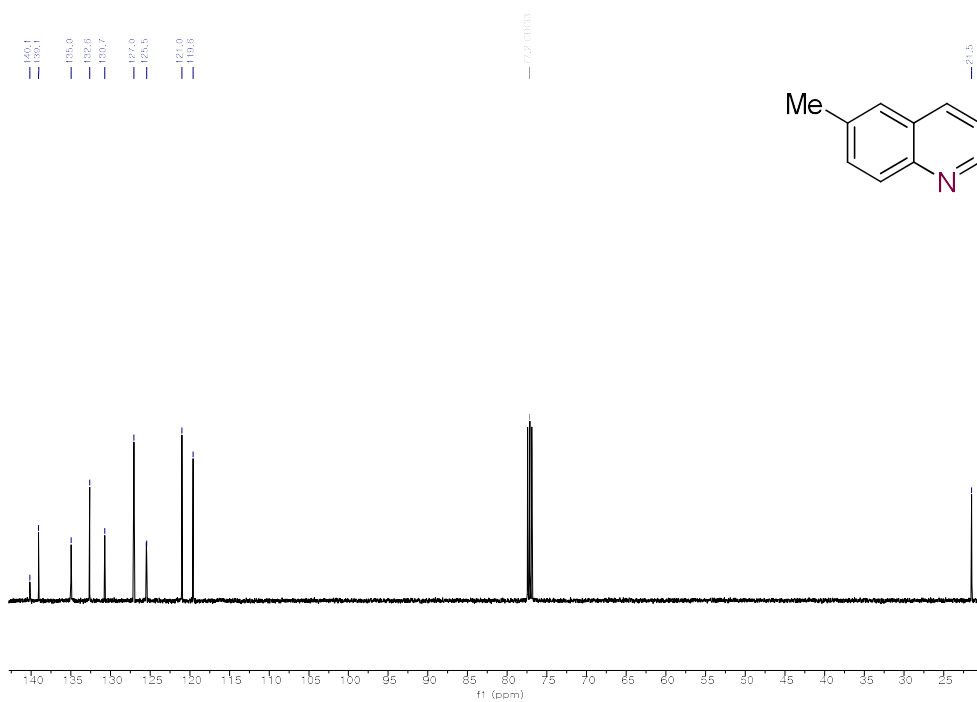


¹H & ¹³C NMR Spectra of 6-Methylquinoline (2v)

¹H NMR (500 MHz, CDCl₃)

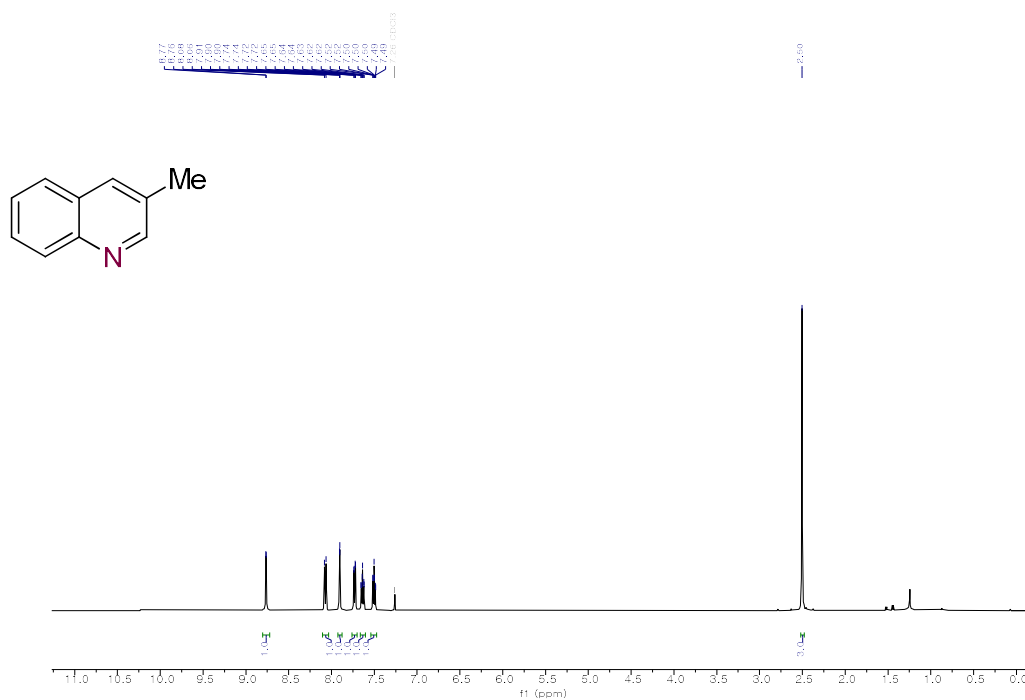


¹³C NMR (126 MHz, CDCl₃)

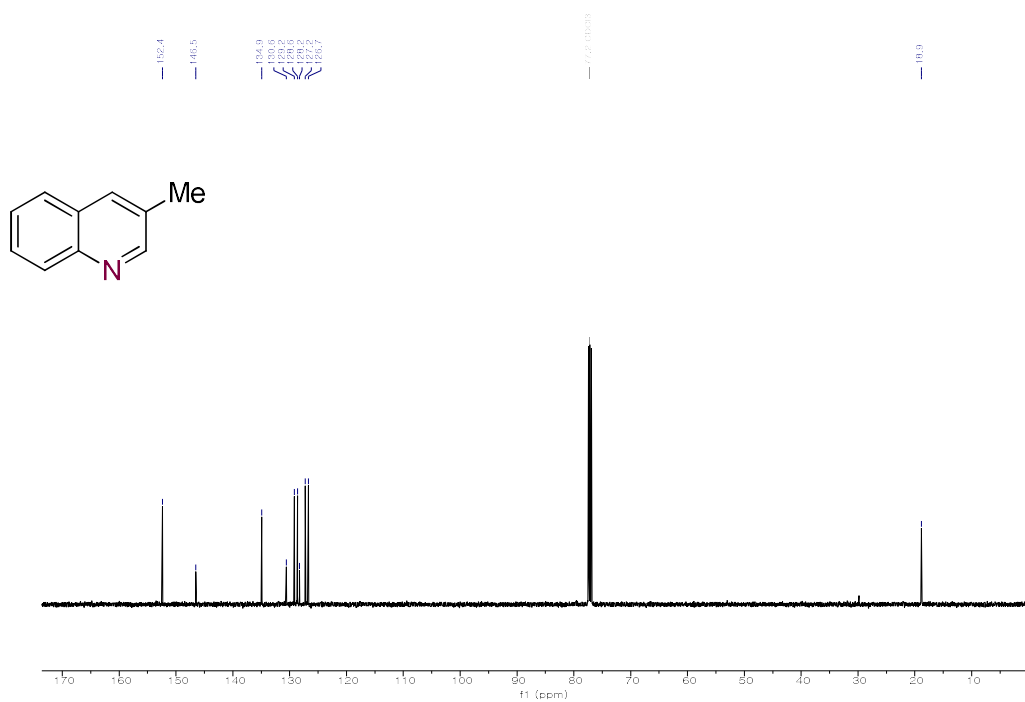


¹H & ¹³C NMR Spectra of 3-Methylquinoline (2w)

¹H NMR (500 MHz, CDCl₃)

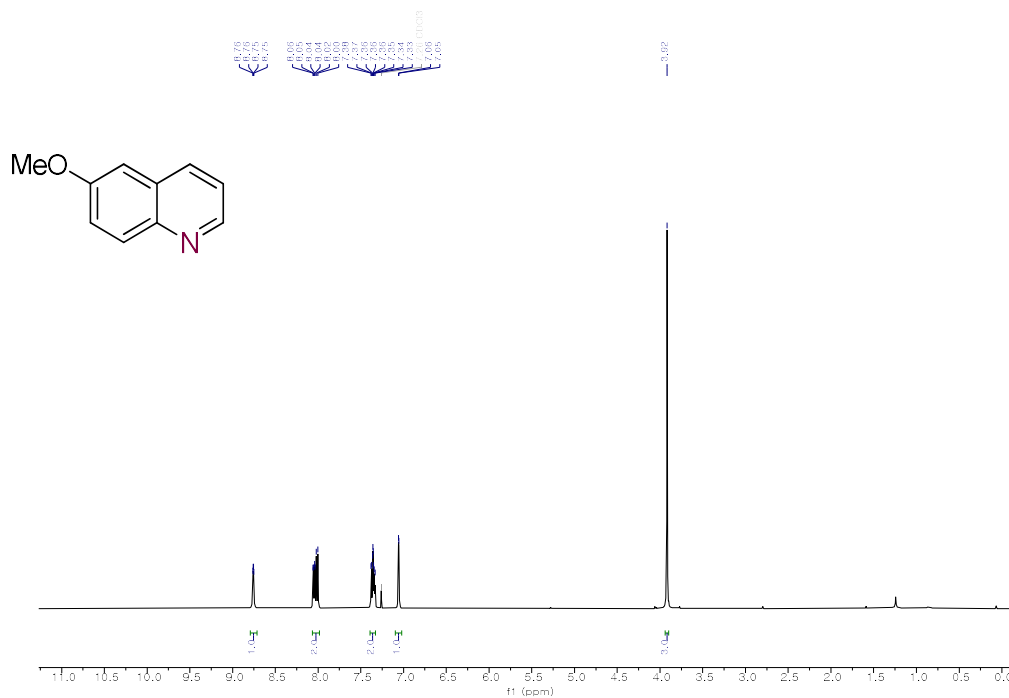


¹³C NMR (126 MHz, CDCl₃)

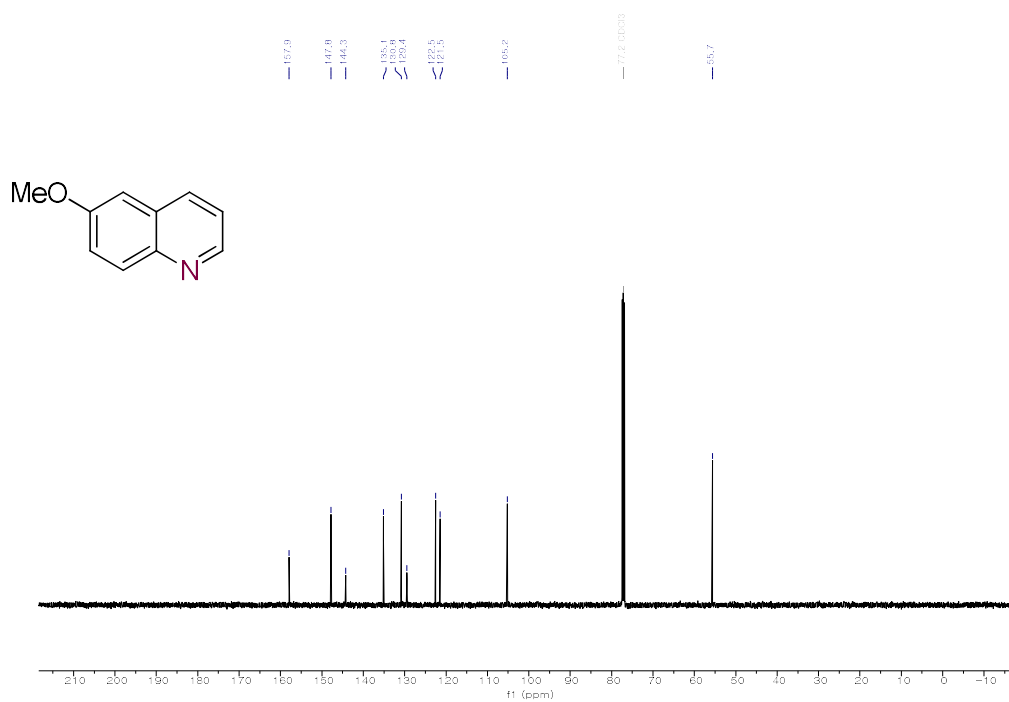


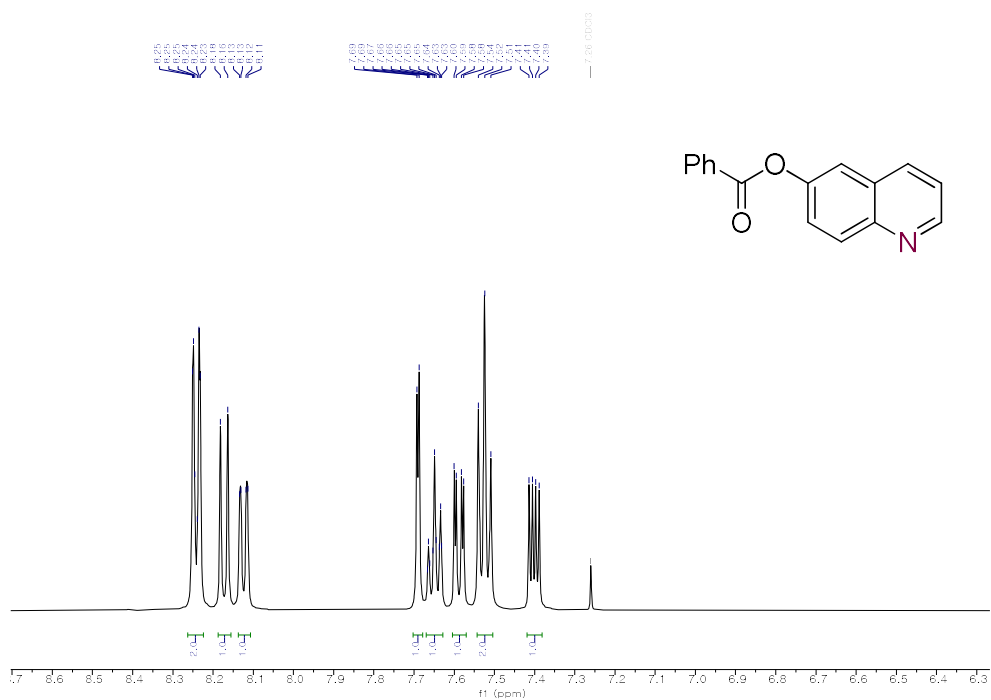
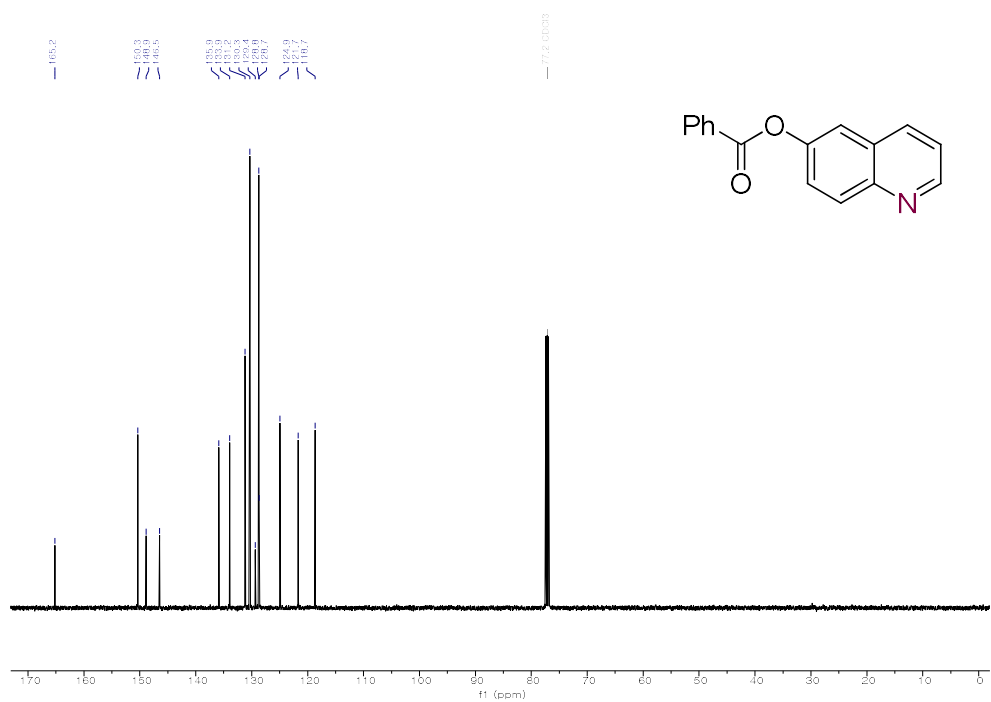
¹H & ¹³C NMR Spectra of 6-Methoxyquinoline (2x)

¹H NMR (500 MHz, CDCl₃)



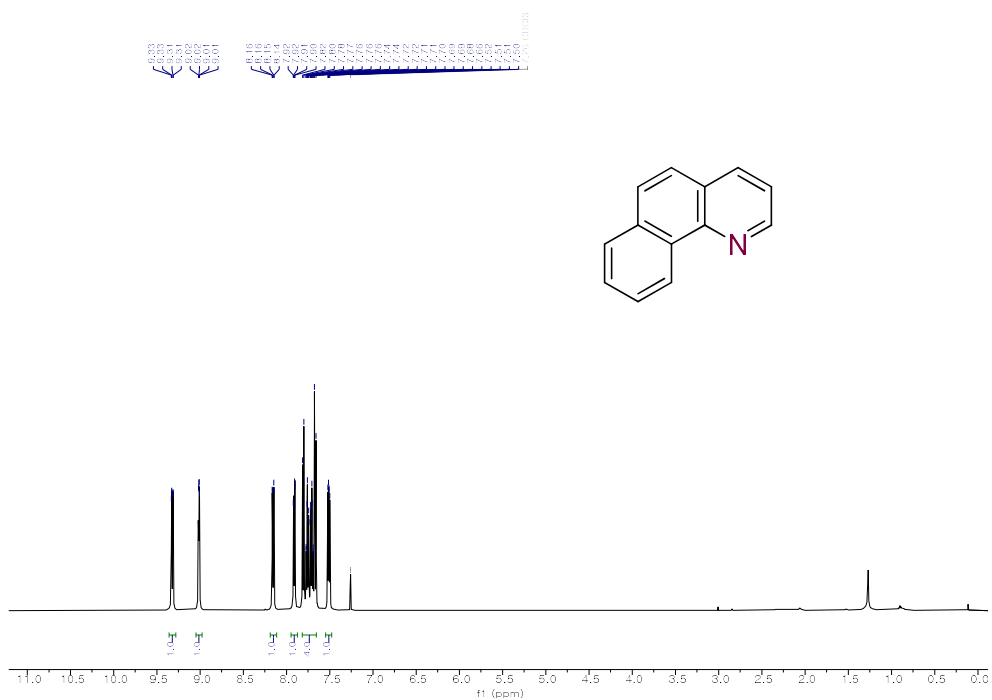
¹³C NMR (126 MHz, CDCl₃)



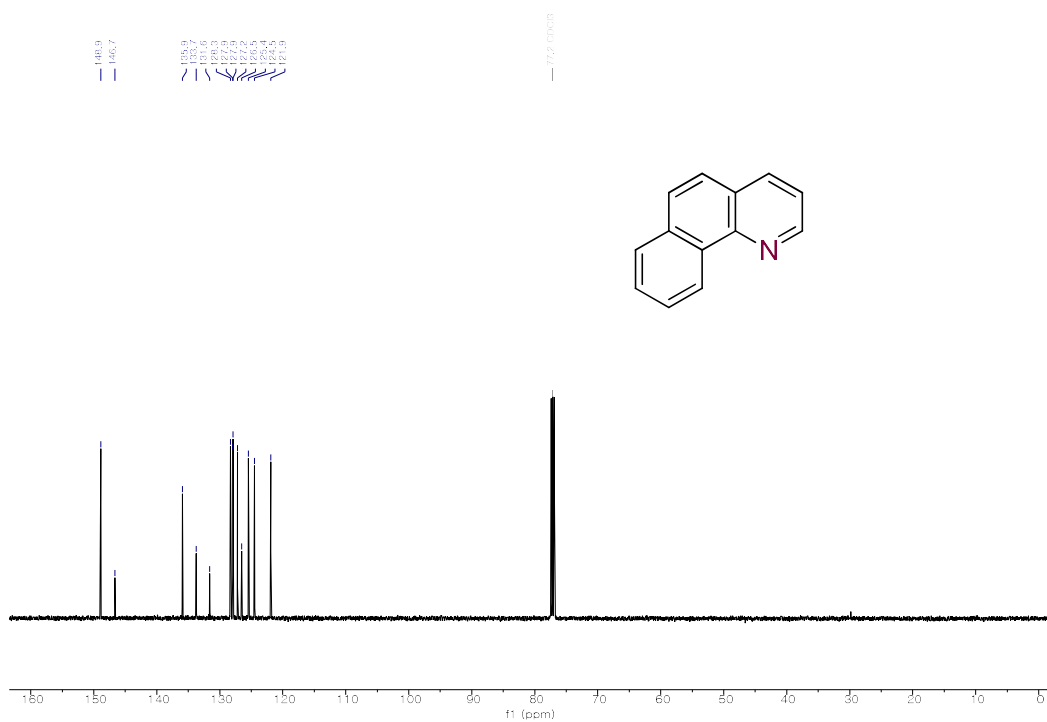
¹H & ¹³C NMR Spectra of 6-Benzoyloxyquinoline (2y)**¹H NMR (500 MHz, CDCl₃)****¹³C NMR (126 MHz, CDCl₃)**

¹H & ¹³C NMR Spectra of Benzo[*h*]quinoline (2aa)

¹H NMR (500 MHz, CDCl₃)

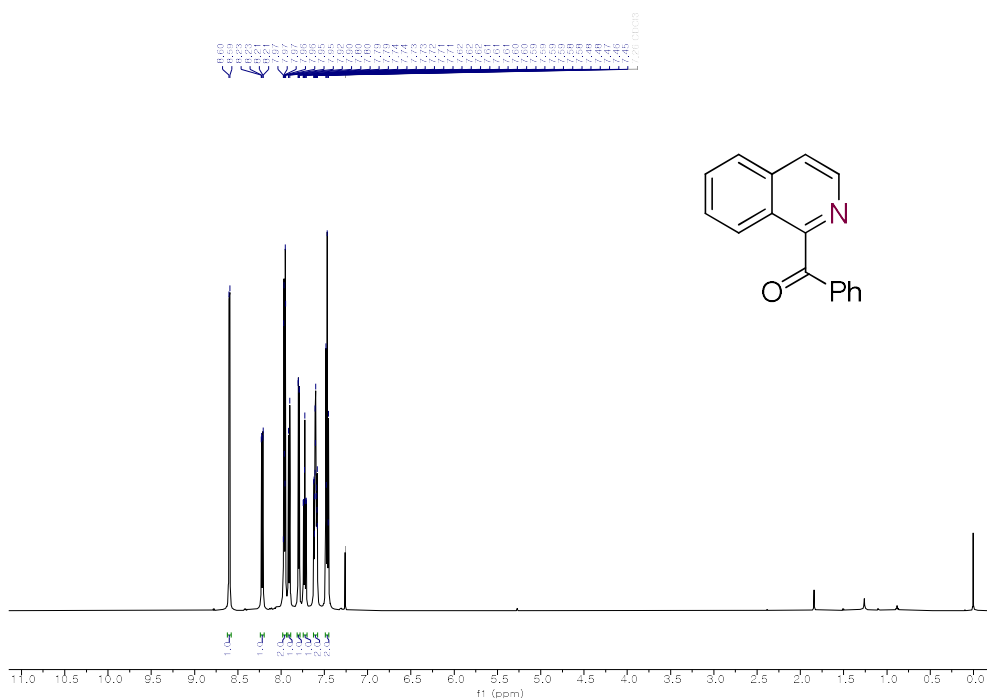


¹³C NMR (126 MHz, CDCl₃)

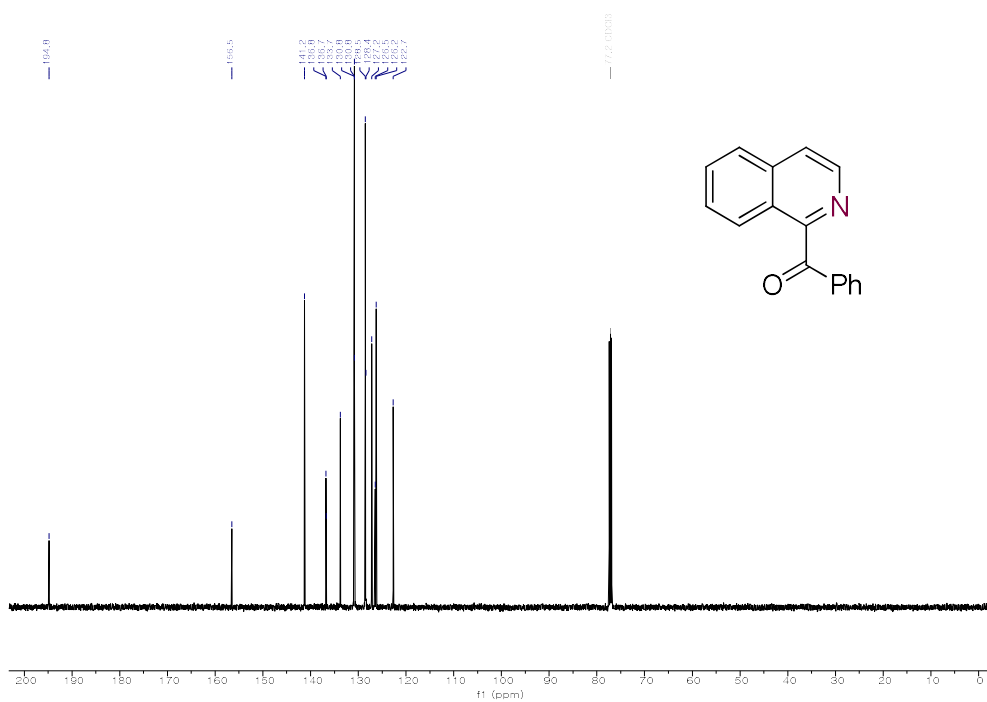


¹H & ¹³C NMR Spectra of 1-Benzoylisoquinoline (2ad)

¹H NMR (500 MHz, CDCl₃)

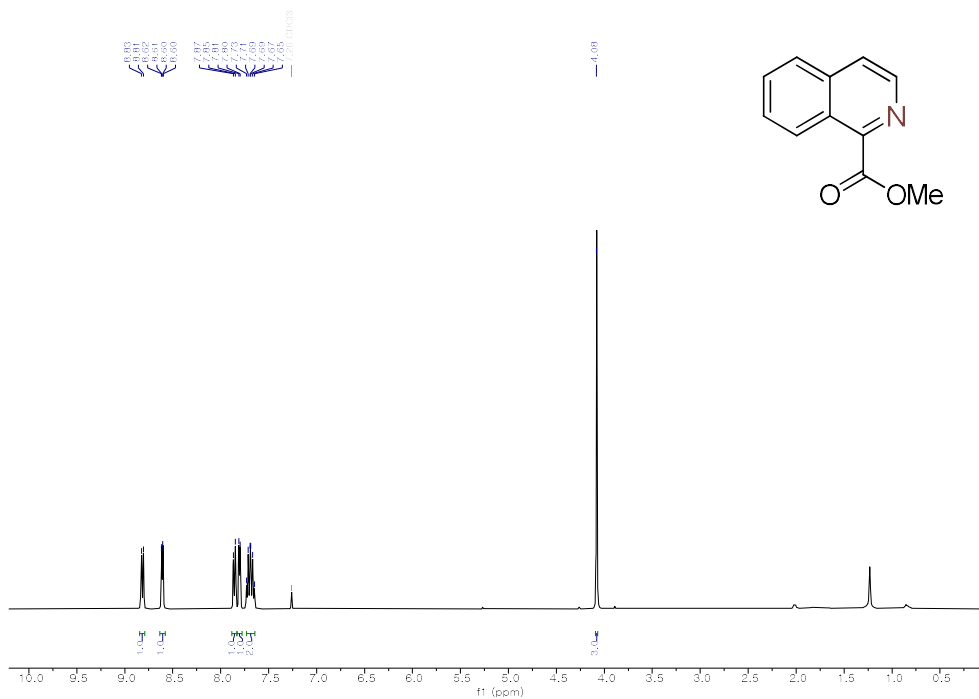


¹³C NMR (126 MHz, CDCl₃)

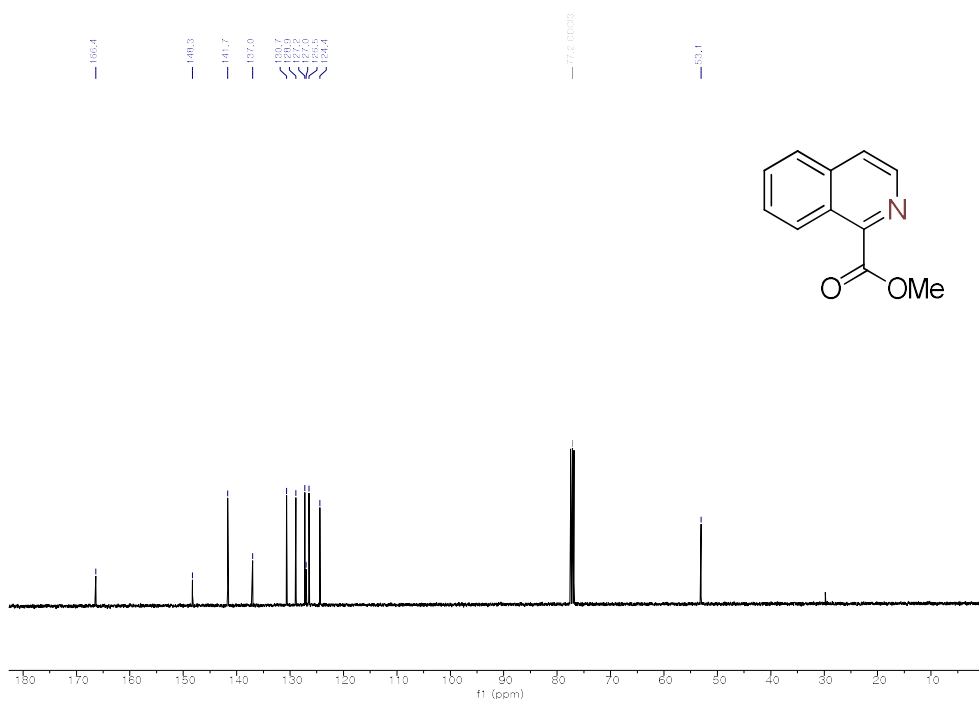


Preparation of Methyl 1-Isoquinolinecarboxylate (2af)

¹H NMR (400 MHz, CDCl₃)

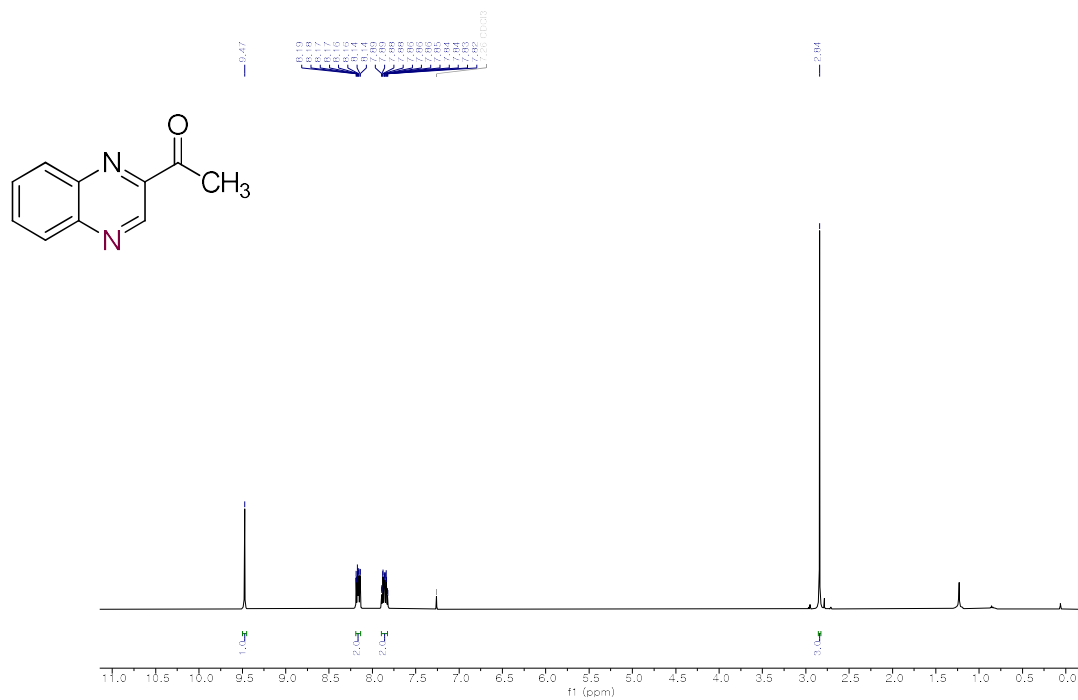


¹³C NMR (101 MHz, CDCl₃)

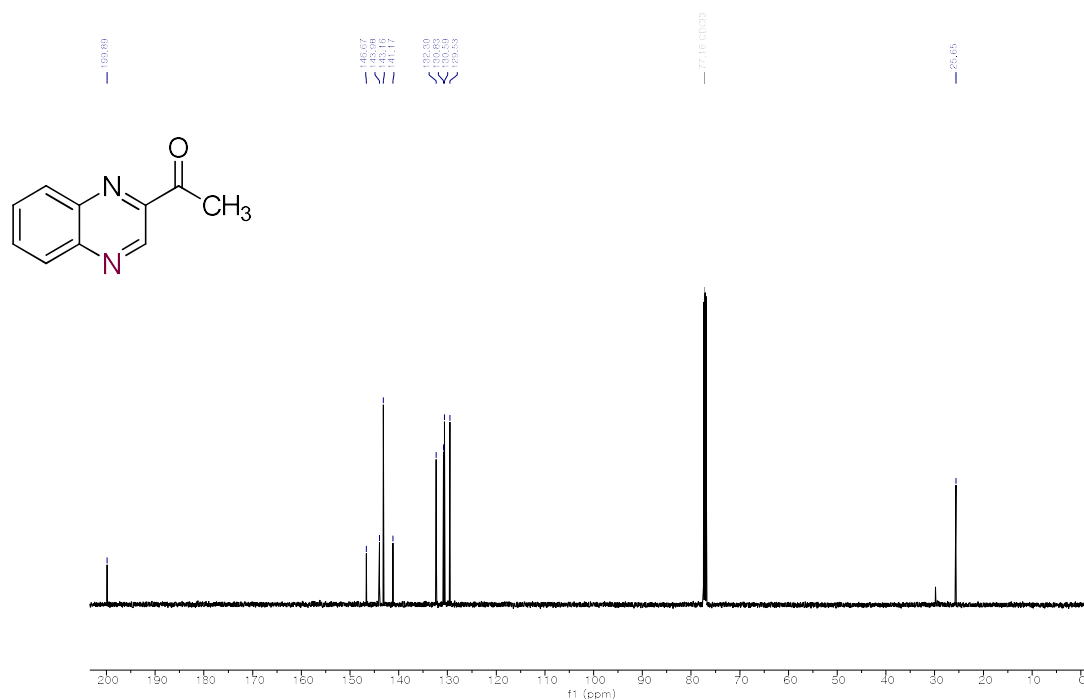


¹H & ¹³C NMR Spectra of Methyl 2-Quinoxalinecarboxylate (2ag)

¹H NMR (500 MHz, CDCl₃)

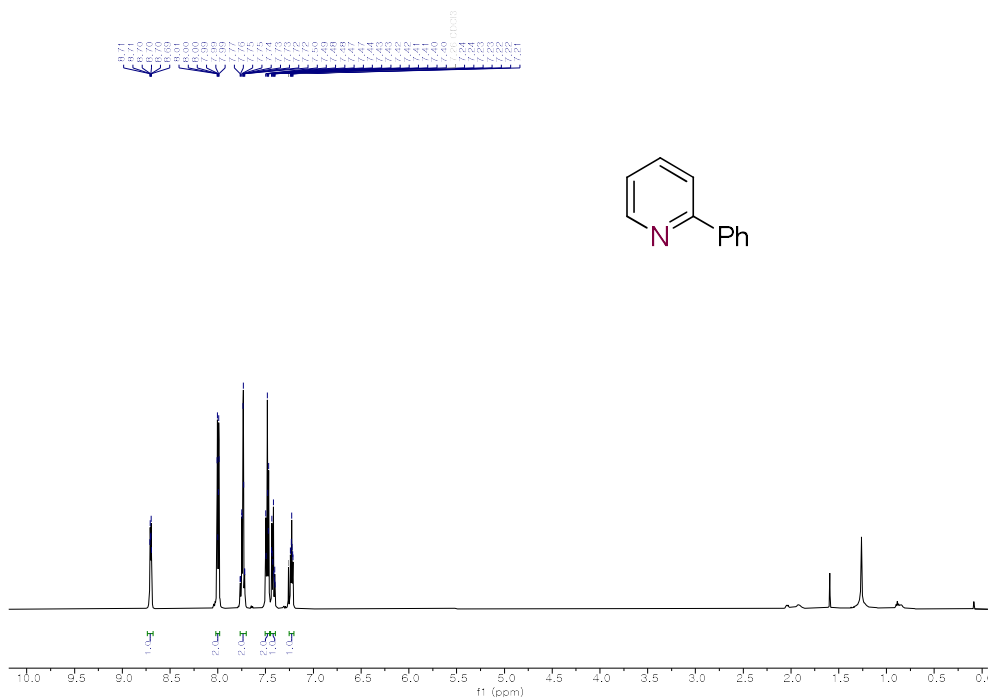


¹³C NMR (126 MHz, CDCl₃)

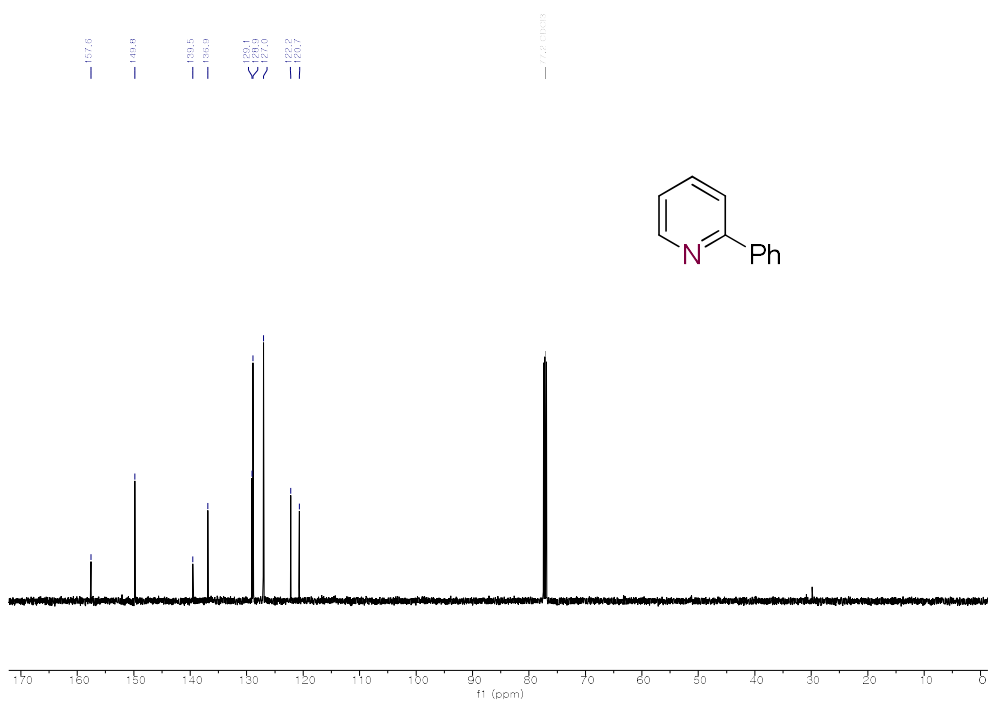


¹H & ¹³C NMR Spectra of 2-Phenylpyridine (2aj)

¹H NMR (500 MHz, CDCl₃)

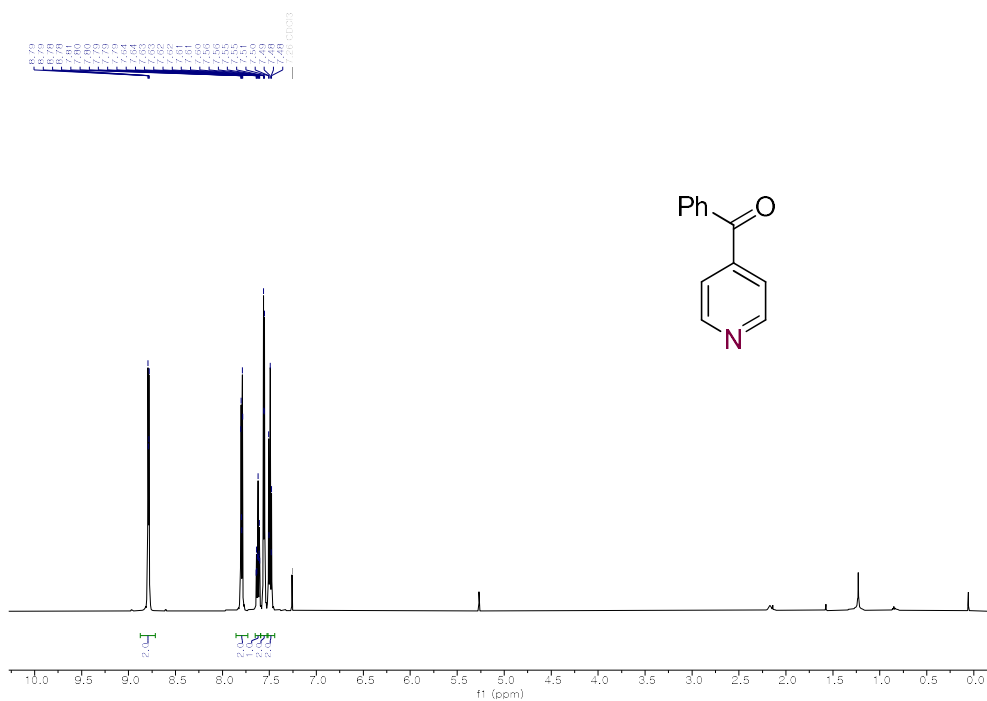


¹³C NMR (126 MHz, CDCl₃)

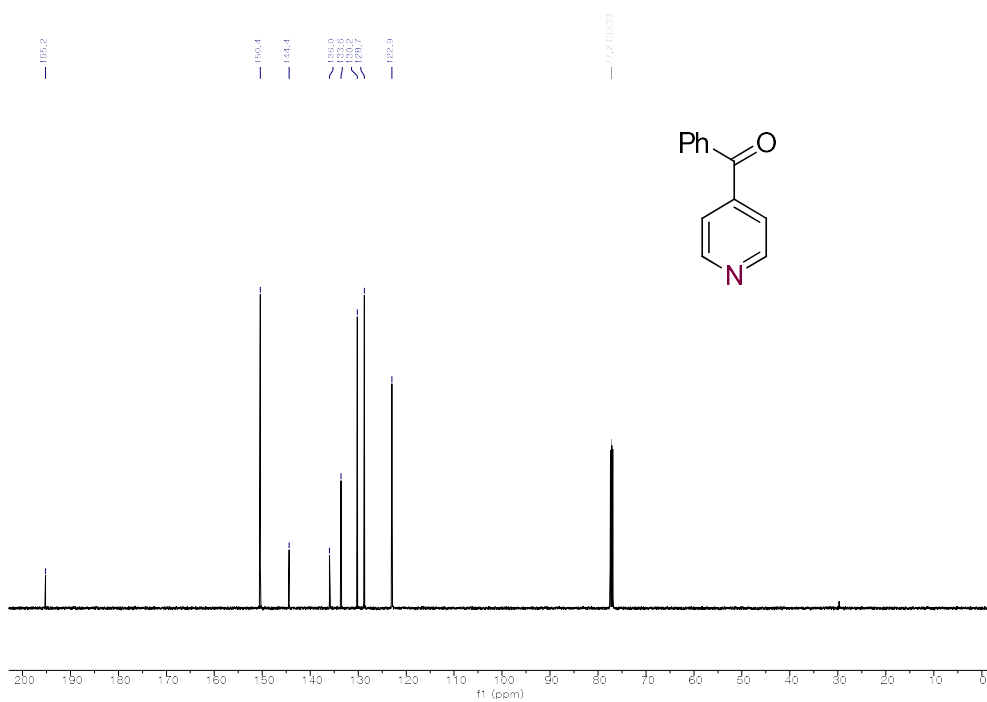


¹H & ¹³C NMR Spectra of 4-Benzoylpyridine (2ak)

¹H NMR (500 MHz, CDCl₃)

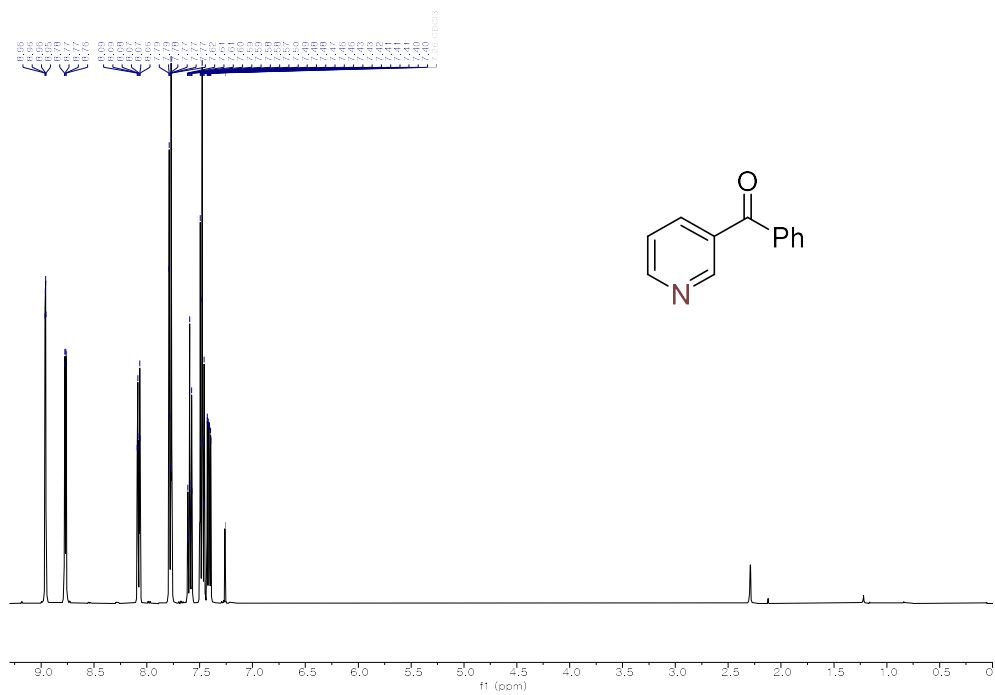


¹³C NMR (126 MHz, CDCl₃)

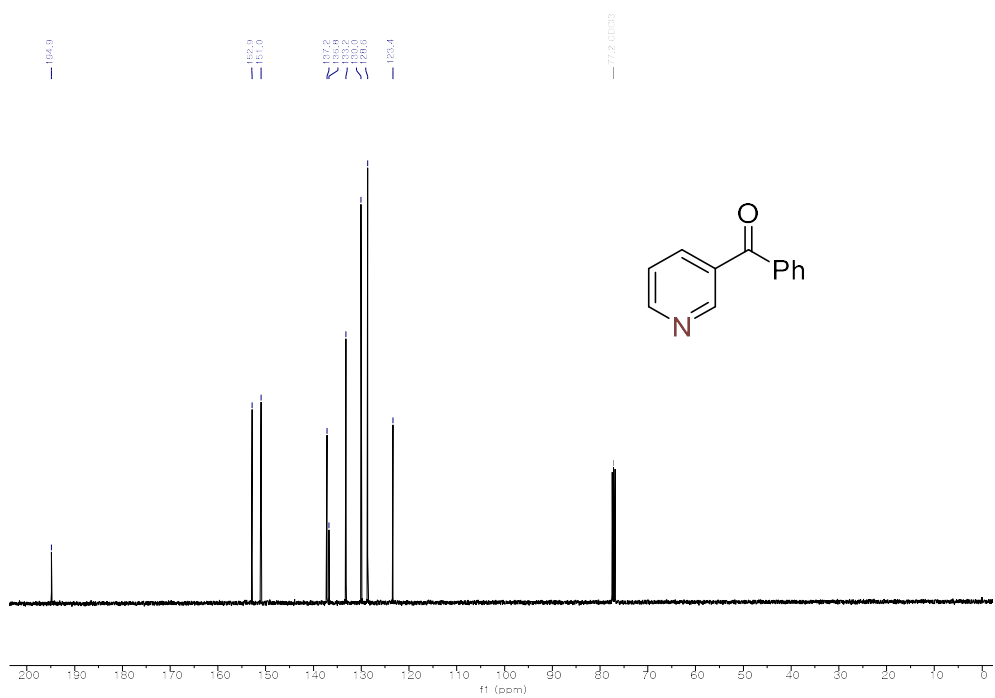


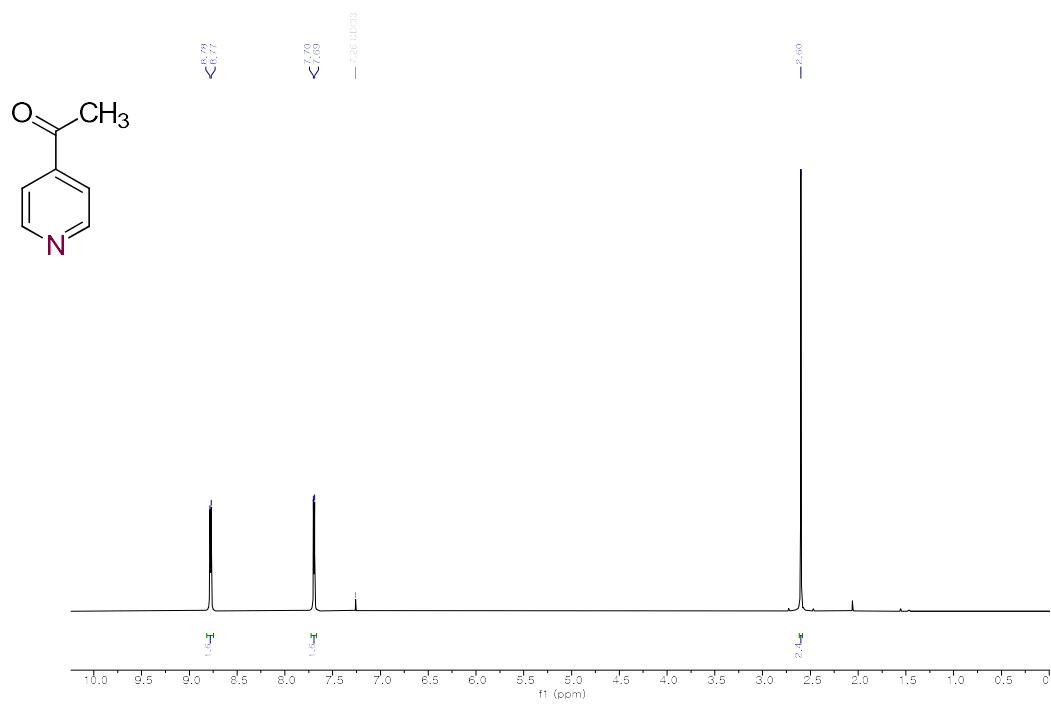
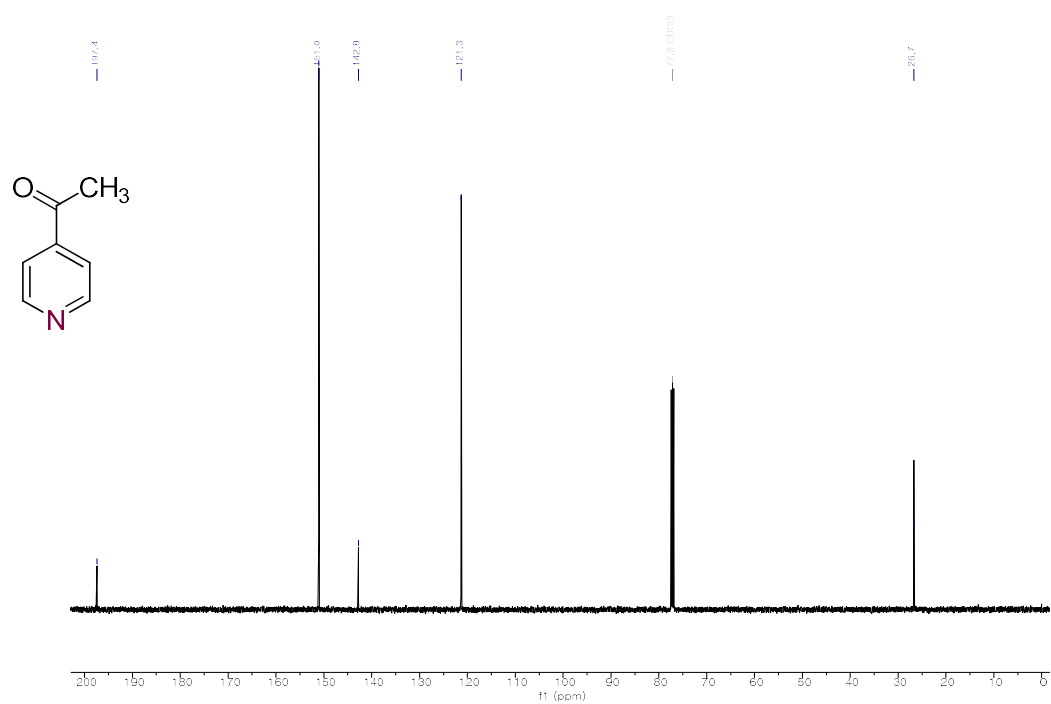
¹H & ¹³C NMR Spectra of 3-Benzoylpyridine (2aI)

¹H NMR (500 MHz, CDCl₃)



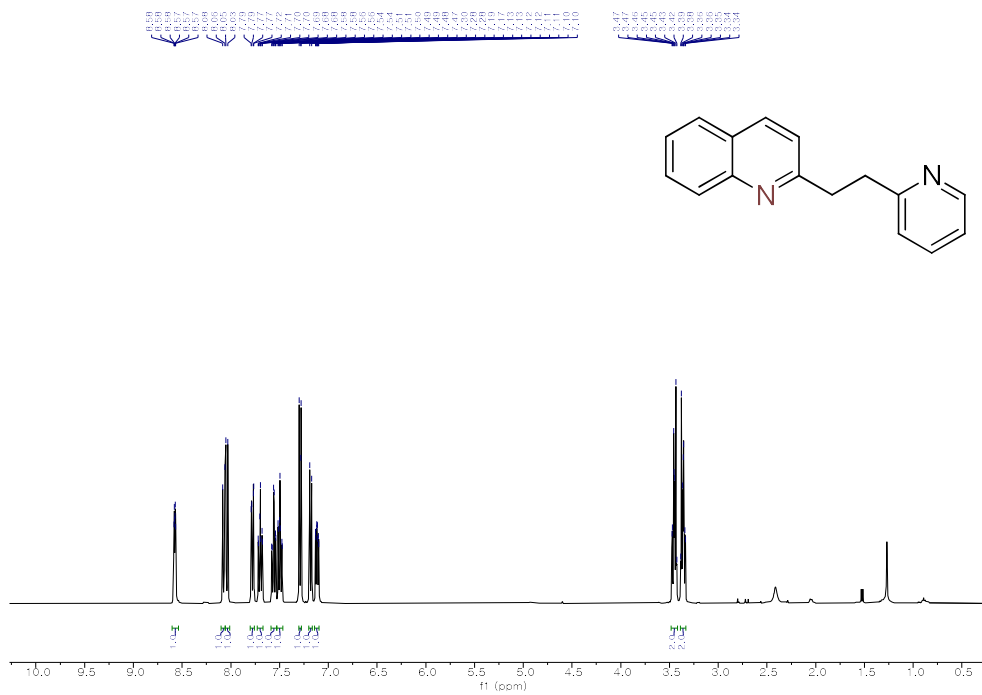
¹³C NMR (126 MHz, CDCl₃)



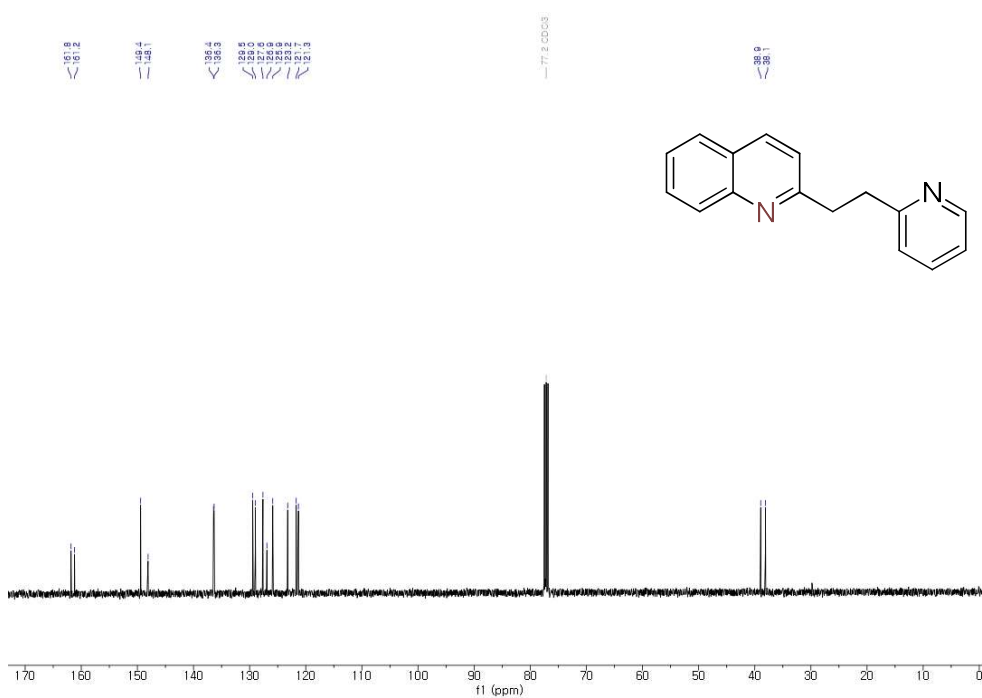
¹H & ¹³C NMR Spectra of 4-Acetylpyridine (2an)**¹H NMR (500 MHz, CDCl₃)****¹³C NMR (126 MHz, CDCl₃)**

¹H & ¹³C NMR Spectra of 2-{2-(*N*-Oxidopyridine-2-yl)ethyl}quinoline (1ao')

¹H NMR (400 MHz, CDCl₃)

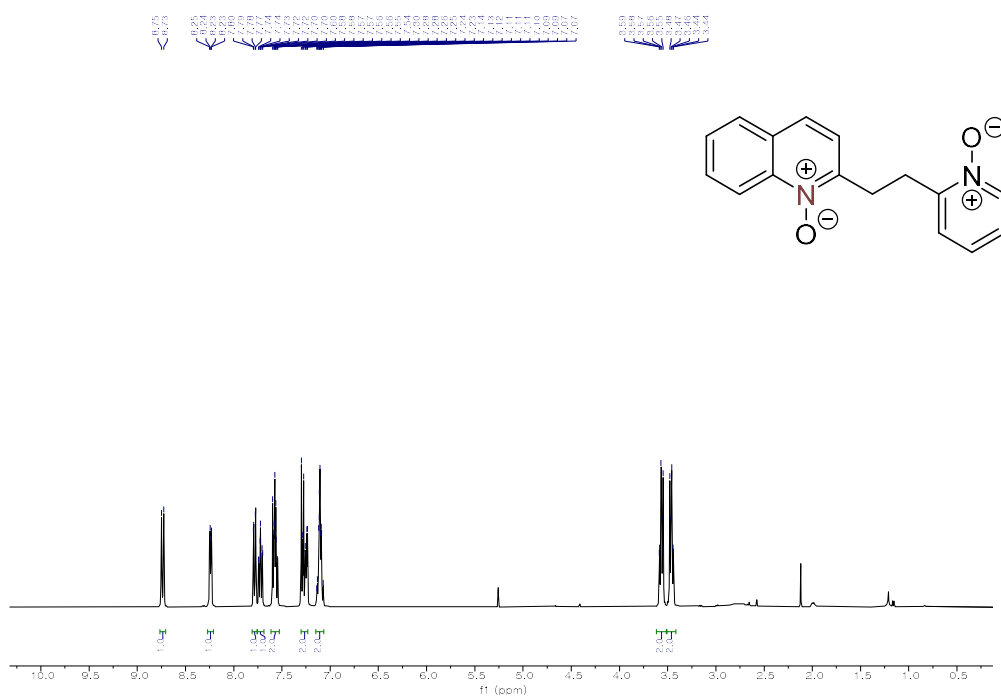


¹³C NMR (101 MHz, CDCl₃)

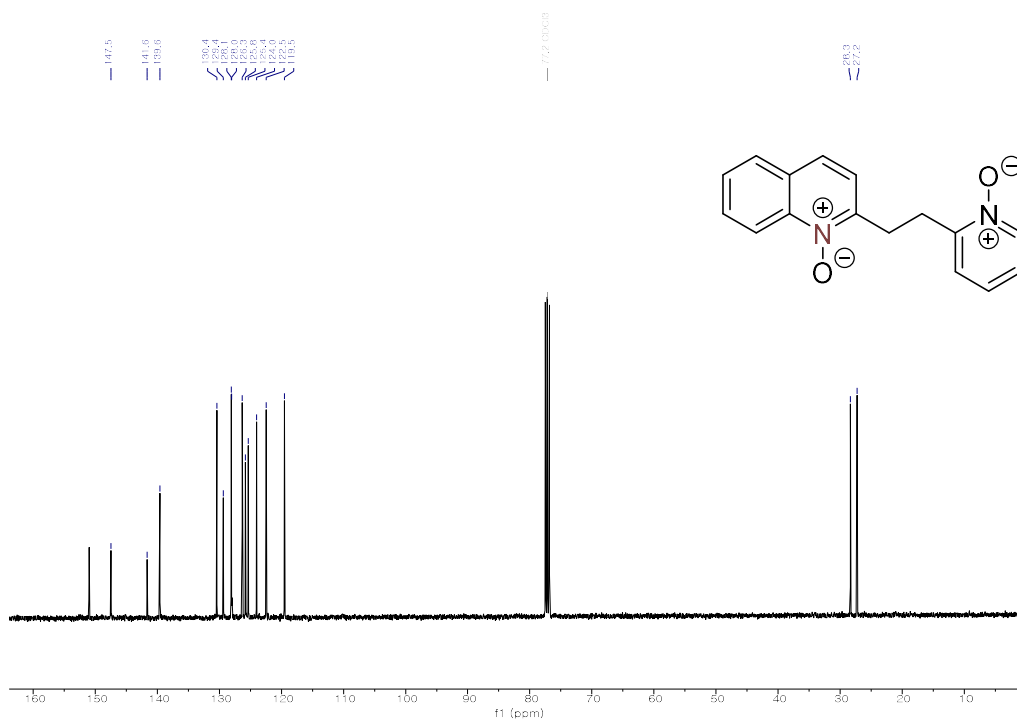


¹H & ¹³C NMR Spectra of 2-{2-(*N*-Oxidopyridine-2-yl)ethyl}quinoline *N*-oxide (1ao)

¹H NMR (400 MHz, CDCl₃)

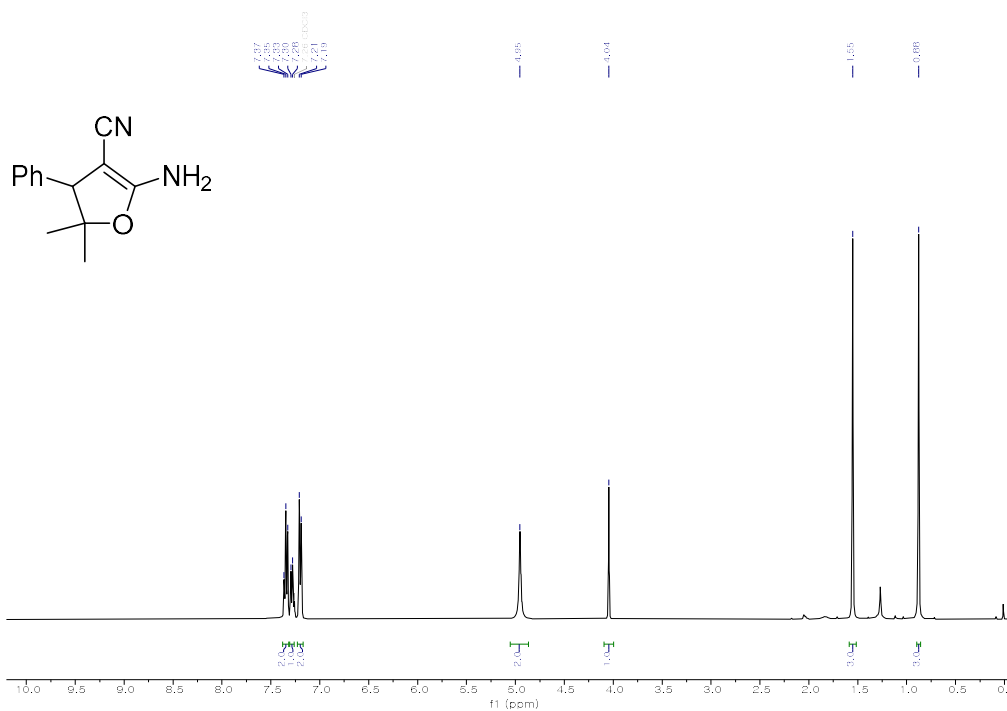


¹³C NMR (101 MHz, CDCl₃)



¹H & ¹³C NMR Spectra of 2-Amino-5,5-dimethyl-4-phenyl-4,5-dihydrofuran-3-carbonitrile (9')

¹H NMR (400 MHz, CDCl₃)



¹³C NMR (101 MHz, CDCl₃)

