

Biological Activity of Lyophilized Chitosan Scaffolds with Inclusion of Chitosan and Zinc Oxide Nanoparticles

Received 00th January 20xx,
Accepted 00th January 20xx

DOI: 10.1039/x0xx00000x

Jorge Eliecer Viloria Angarita^a, Daniel Insuasty^b, Juan David Rodríguez M.^c, Jorge Iván Castro^d, Carlos Humberto Valencia-Llano^e, Paula A. Zapata^f, Johannes Delgado-Ospina^g, Diana Paola Navia-Porras^g, Alberto Albis^h, Carlos David Grande-Tovar^{a*}

The constant demand for biocompatible and non-invasive materials for regenerative medicine in accidents and various diseases has driven the development of innovative biomaterials that promote biomedical applications. In this context, using sol-gel and ionotropic gelation methods, zinc oxide nanoparticles (NPs-ZnO) and chitosan nanoparticles (NPs-CS) were synthesized with sizes of 20.0 nm and 11.98 nm, respectively. These nanoparticles were incorporated into chitosan scaffolds through the freeze-drying method, generating a porous morphology with small (<100 μm), medium (100–200 μm), and large (200–450 μm) pore sizes. Moreover, the four formulations showed preliminary bioactivity after hydrolytic degradation, facilitating the formation of a hydroxyapatite (HA) layer on the scaffold surface, as evidenced by the presence of Ca (4%) and P (5.1%) during hydrolytic degradation. The scaffolds exhibited average antibacterial activity of F1=92.93%, F2=99.90%, F3=74.10%, and F4=88.72% against four bacterial strains: *K. pneumoniae*, *E. cloacae*, *S. enterica*, and *S. aureus*. In vivo evaluation confirmed the biocompatibility of the functionalized scaffolds, where F2 showed accelerated resorption attributed to the NPs-ZnO. At the same time, F3 exhibited controlled degradation with NPs-CS acting as initiation points for degradation. On the other hand, F4 combined NPs-CS and NPs-ZnO, resulting in progressive degradation, reduced inflammation, and an organized extracellular matrix. All the results presented expand the boundaries in tissue engineering and regenerative medicine by highlighting the crucial role of nanoparticles in optimizing scaffold properties.

^a Grupo de Investigación de Fotoquímica y Fotobiología, Universidad del Atlántico, Carrera 30 Número 8-49, Puerto Colombia 081008, Colombia

^b Departamento de Química y Biología, División de Ciencias Básicas, Universidad del Norte, Km 5 Vía Puerto Colombia, Barranquilla 081007, Colombia

^c Programa de Medicina, Facultad de Ciencias de la Salud, Universidad Libre. Km 5 Vía Puerto Colombia, Barranquilla 081007, Colombia.

^d Tribology, Polymers, Powder Metallurgy and Solid Waste Transformations Research Group, Universidad del Valle, Calle 13 No. 100-00, Cali 76001, Colombia; Jorge.castro@correounivalle.edu.co (J.I.C)

^e Grupo Biomateriales Dentales, Escuela de Odontología, Universidad del Valle, Calle 4B # 36-00, Cali 76001, Colombia

^f Grupo de Polímeros, Facultad de Química y Biología, Universidad de Santiago de Chile, Santiago 9170020, Chile

^g Grupo de Investigación Biotecnología, Facultad de Ingeniería, Universidad de San Buenaventura Cali, Carrera 122 # 6-65, Cali 76001

^h Grupo de Investigación en Bioprocesos, Universidad del Atlántico, Facultad de Ingeniería, Alberto Albis; albertoalbis@uniatlantico.edu.co. Carrera 30 Número 8-49, Puerto Colombia 081008, Colombia

*Correspondence: carlosgrande@mail.uniatlantico.edu.co; Tel.: +57-5-3599-484

Supplementary information

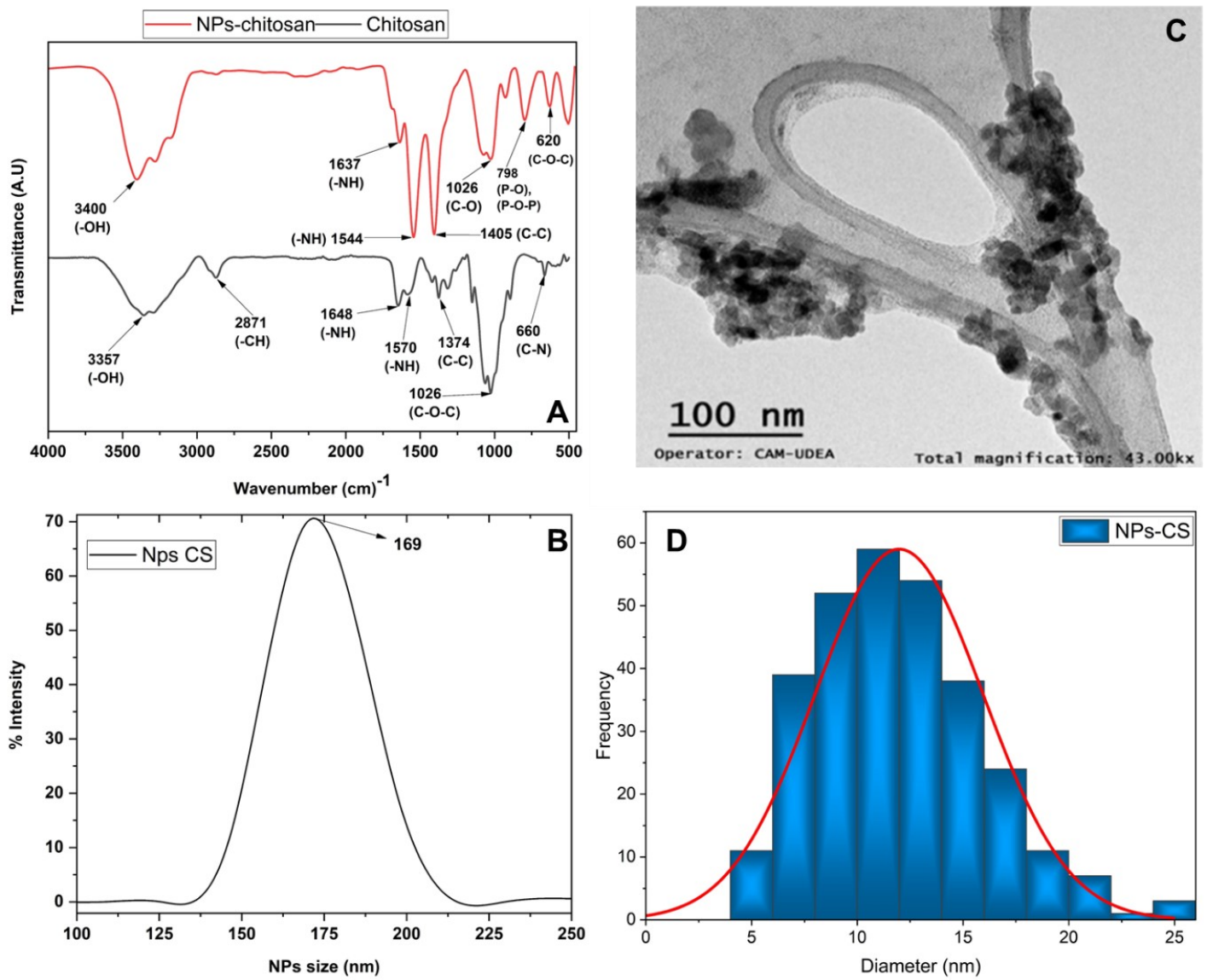


Figure S1. A) FTIR Spectrum of chitosan and NPs-CS. B) Size distribution profile of NPs-CS. C) TEM images of NPs-CS. D) Size histogram of NPs-CS.

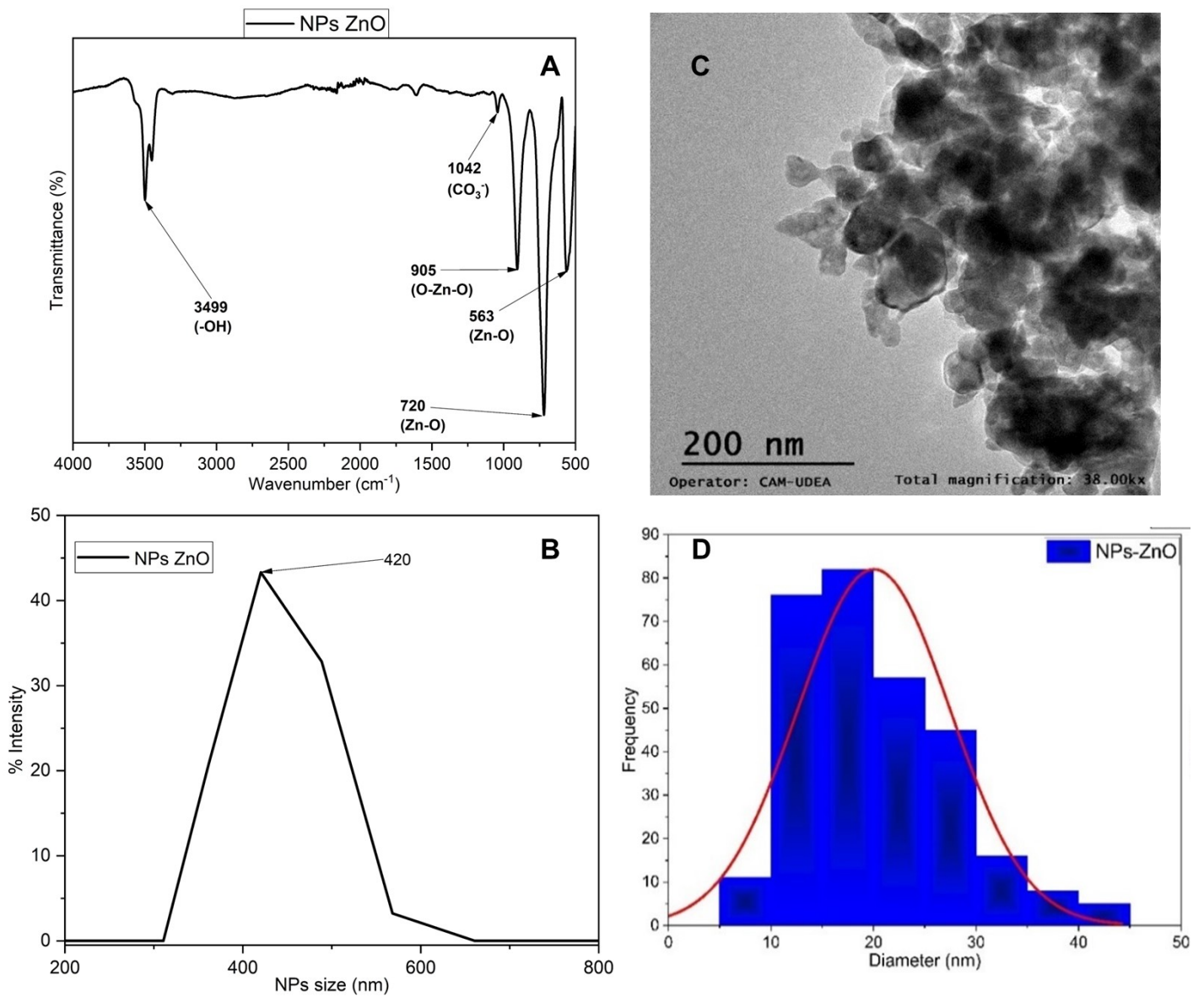


Figure S2. A) FTIR Spectrum of NPs-ZnO. B) Size distribution profile of NPs-ZnO. C) TEM images of NPs-ZnO. D) Size histogram of NPs-ZnO.