Supporting Information

Cu-doped and 2-propylimidazole-modified nanoceria (CeO₂@Cu-PrIm) oxidase-like nanozyme for total antioxidant capacity assay of fruits

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Fig. S1. XPS spectra of survey scan for CeO₂@Cu-PrIm.



Fig. S2. The stability of the oxidase-like catalytic activity of CeO₂@Cu-PrIm. (a) Batch stability of CeO₂@Cu-PrIm. (b) Storage stability of CeO₂@Cu-PrIm.



Fig. S3. The oxidase-like activity of $CeO_2@Cu-PrIm$ after the cycle of centrifugation. The recycled $CeO_2@Cu-PrIm$ was obtained by centrifuging the reaction solution at 8000 g for 10 min. The above cycle was repeated five times.



Fig. S4. The UV-vis absorption spectra describing the relationship between $CeO_2@Cu$ -PrIm catalytic activity and concentration.



Fig. S5. The detection of AA using the DPPH \cdot free radical scavenging method.



Fig. S6. The anti-interference capacity of the reaction system of $CeO_2@Cu-PrIm/ox-TMB$ assay.



Fig. S7. The detection of TAC of three vitamin C tablets using the DPPH \cdot free radical scavenging method.

Supplementary Tables

Catalyst	<i>K_m</i> [mM]	V _{max} [10 ⁻⁸ M/s]	Reference
CeO ₂ @Cu-PrIm	6.521	77.45	This work
CeO ₂ NPs	3.8	70	1
Nano-CeO ₂	0.42	10.04	2
Ce-BPyDC	0.16	26.8	3
Ce-MOF (MVCM)	0.00037	550	4
Dex-FeMnzyme	0.33	13.29	5
MIL-53(Fe)	1.08	8.78	6
Mn ₃ O ₄ NPs	0.025	5.07	7

Table S1. Comparison of kinetic parameters between $CeO_2@Cu-PrIm$ and reported oxidase mimics.

Nanomaterials	Linear range (µM)	LOD (µM)	Reference
CeO ₂ @Cu-PrIm	1-70	1.26	This work
Ce-BPyDC	1-20	0.28	3
Dex-FeMnzyme	1-30	1.17	5
MIL-53(Fe)	28.6-190.5	15	6
CuNCs	0.5-10	0.11	8
CP ₆₀₀₋₆	0.8-80	35	9
SNC	100-5000	80	10
Fe-NC NTs	0.2-20	0.131	11

 Table S2. Comparison of reported oxidase mimics for the detection of AA.



Fig. S8. The EDS Mapping of CeO₂@Cu-PrIm NPs (a) HAADF of CeO₂@Cu-PrIm NPs

(b) C atoms (c) N atoms (d) O atoms (e) Ce atoms (f) Cu atoms

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