

## Supplementary Information

### Ultrasonic-assisted Europium Decorated Cuprous Oxide Nanoparticles: Exploring their Photothermal Capabilities and Antioxidant Properties for Biomedical Applications

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### Estimation of Photothermal Conversion Efficiency

The photothermal conversion efficiencies of the Eu-Cu<sub>2</sub>O NPs and Cu<sub>2</sub>O NPs were determined according to previously reported methods. Detailed calculation is as follows.

$$\eta = \frac{hS(T_{\max} - T_0) - Q_{Dis}}{I(1 - 10^{-A_{808}})} \quad \dots (1)$$

$h$  is the heat transfer coefficient;  $S$  is the surface area of the container; the value of  $hS$  is obtained from Eq. 4 and Fig. 4d and 4f.

The maximum steady temperature ( $T_{\max}$ ) of the solution of Eu-Cu<sub>2</sub>O NPs and Cu<sub>2</sub>O NPs was 55.3 °C and 46.6 °C respectively.

The ambient temperature ( $T_0$ ) of the solution of Eu-Cu<sub>2</sub>O NPs and Cu<sub>2</sub>O NPs was 28.4 °C and 24 °C, respectively.

Therefore, the temperature change ( $T_{\max} - T_0$ ) of the solution of Eu-Cu<sub>2</sub>O NPs and Cu<sub>2</sub>O NPs was 26.9 °C and 22.6 °C, respectively.

The laser power  $I$  is 1W. The absorbances of Eu-Cu<sub>2</sub>O NPs and Cu<sub>2</sub>O NPs at 808 nm  $A_{808}$  were 0.282 and 0.228, respectively.  $Q_{Dis}$  denotes the heat dissipated from the light absorbed by the solvent and container.

To calculate  $hS$ , a dimensionless parameter  $\theta$  was introduced as follows:

$$\theta = \frac{T - T_0}{T_{\max} - T_0} \quad \dots (2)$$

A sample system time constant  $t_s$  was calculated as Eq. 3

$$t = -t_s \ln(\theta) \quad \dots (3)$$

According to figures 4d and 4f,  $t_s$  was calculated to be 489.67 s and 591.161 s for Eu-Cu<sub>2</sub>O NPs and Cu<sub>2</sub>O NPS, respectively.

$$hS = \frac{m_D C_D}{\tau_s} \dots\dots\dots (4)$$

Additionally,  $m$  is 1g, and  $C$  is 4.2 J/g °C. Based on Eq . 4,  $hS$  is measured to be 8.5 mW/ °C and 7.1 mW/ °C for Eu-Cu<sub>2</sub>O NPs and Cu<sub>2</sub>O NPs respectively.

$Q_{Dis}$  measured heat dissipated from the light absorbed by the quartz sample cell that was measured using a quartz cuvette containing pure water and found to be 2.1 W

Substituting the values of all parameters in Eq. 1, the photothermal conversion efficiency ( $\eta$ ) at 808 nm of Eu-Cu<sub>2</sub>O NPs and Cu<sub>2</sub>O NPs was calculated to be 44 % and 36 %.

### Estimation of Crystallite Size

Estimation of Crystallite size from XRD data using the Scherrer equation, given by:

$$D = (K.\lambda)/(\beta \cos \theta) \dots\dots\dots (5)$$

$D$ =crystallite size (nm),  $K$  = Shape factor (0.9 for spherical nanoparticles)  $\lambda$ : for Cu K-alpha, 0.15418 nm),  $\beta$ : Full Width at Half Maximum (FWHM) of the peak (in radians).

For Eu-Cu<sub>2</sub>O NPs, FWHM = 0.57203° = .00998 radians

For Cu<sub>2</sub>O NPs, FWHM = 0.38207° = .00667 radians

$\theta$ : Bragg angle (in radians),  $2\theta$ = 26.4°,  $\theta$ =.2304 radians

For Eu-Cu<sub>2</sub>O NPs,  $D = (0.9) (0.15418) / (0.00998) \cos (.2304) = 14.28$  nm

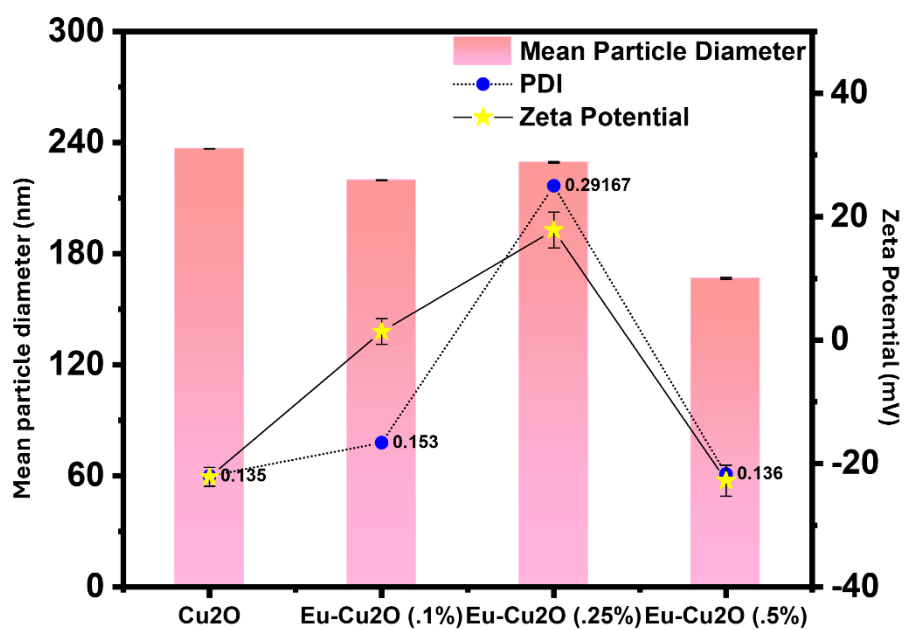
For Cu<sub>2</sub>O NPs,  $D = (0.9) (0.15418) / (0.00667) \cos (.2304) = 21.37$  nm

Entry	Mean particle diameter	PDI	Zeta Potential
Cu <sub>2</sub> O NPs	236.67± 1.52 nm	0.135	-22.1± 0.12 mV
Eu-Cu <sub>2</sub> O NPs (.1%)	219.67±2.08 nm	0.153	1.4 ± 0.20 mV
Eu-Cu <sub>2</sub> O NPs (.25%)	229.33±17.89 nm	0.291	17.8 ± 0.38 mV
Eu-Cu <sub>2</sub> O NPs (.5%)	166.67 ± 2.52 nm	0.136	-22.7 ± 0.55 mV

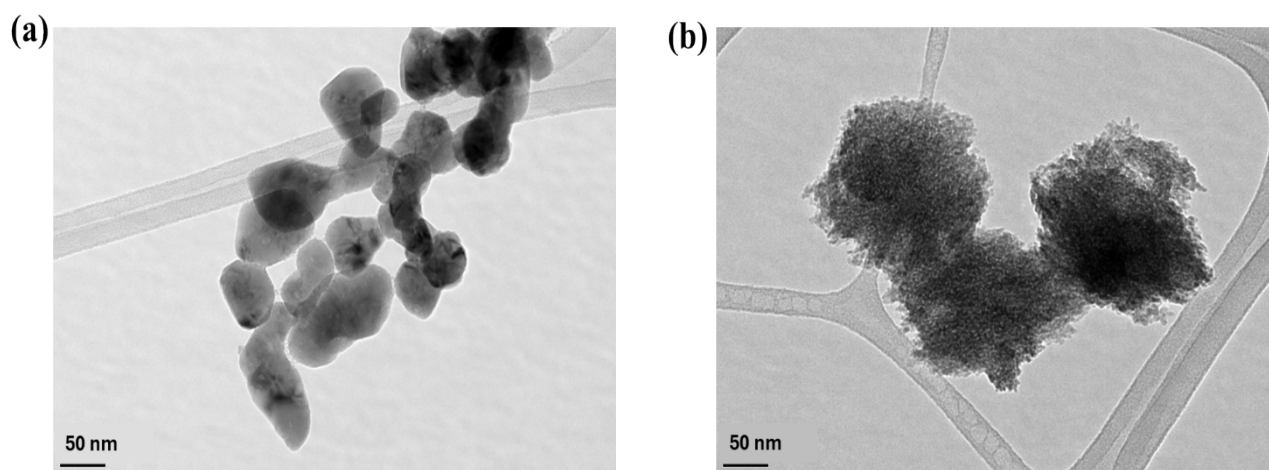
**Table S1.** Table representing the mean particle diameters, PDI and zeta-potential values measured using DLS for various nanoparticles.

Entry	Cell Line	Test Method	Incubation period	Cell Viability	Ref.
Cuprous oxide (Cu <sub>2</sub> O) nanoparticles	B16-F10 (Melanoma)	MTT	48 h	50% at 1.992 mg/ml	[37]
ZnO@polymer core-shell nanoparticles	U251 (Glioblastoma)	MTT	48 h	~80% at 10 mg/ml	[S1]
“? Dextran-coated CeO <sub>2</sub> Nanoparticles	MG-63 (Osteosarcoma)	MTS	24 h	50% at >250 mg/ml	[S2]
TiO <sub>2</sub> -DOX nanoparticles	MCF-7/ADM (Breast cancer)	MTT	24 h	~84% at 10 mg/ml DOX	[S3]
Iron doped Copper oxide nanoparticles	C6 (Glioma)	LDH	5 h	~45% at 1000 mM	[16]
Europium-doped CeO <sub>2</sub> nanoparticles	BV2 (Glial Cells)	MTT	4 h	60% at 1 mg/ml	[45]
Europium decorated Cu <sub>2</sub> O nanoparticles	KB (Epidermal Carcinoma)	PI cell death assay using flow cytometry	4 h	~80% at 50 mg/ml	Present work

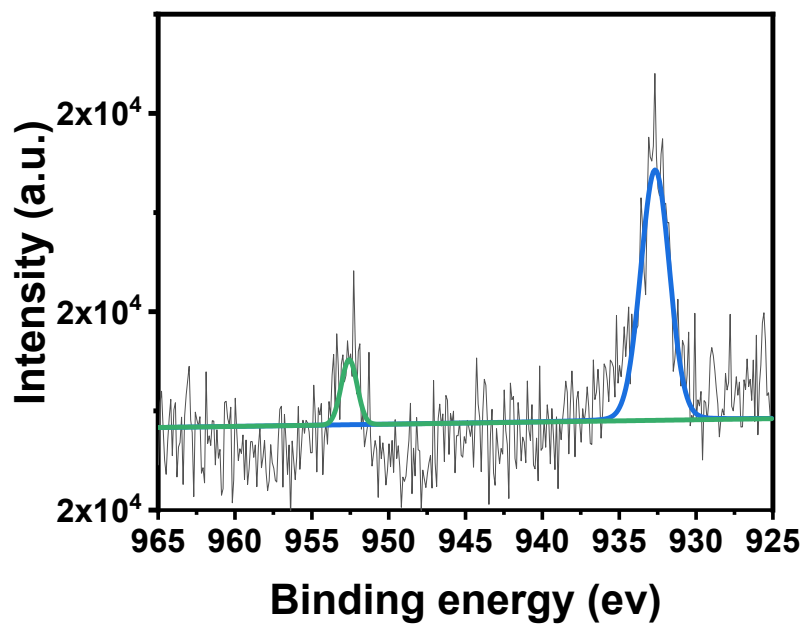
**Table S2.** Comparative analysis of cellular viabilities of different metal oxide nanoparticles.



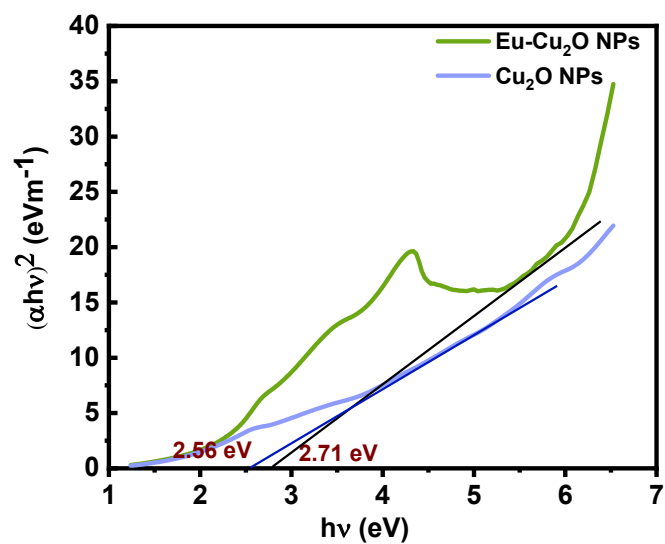
**Fig. S1.** DLS analysis of undecorated Cu<sub>2</sub>O NPs and decorated Eu-Cu<sub>2</sub>O NPs with variable europium content.



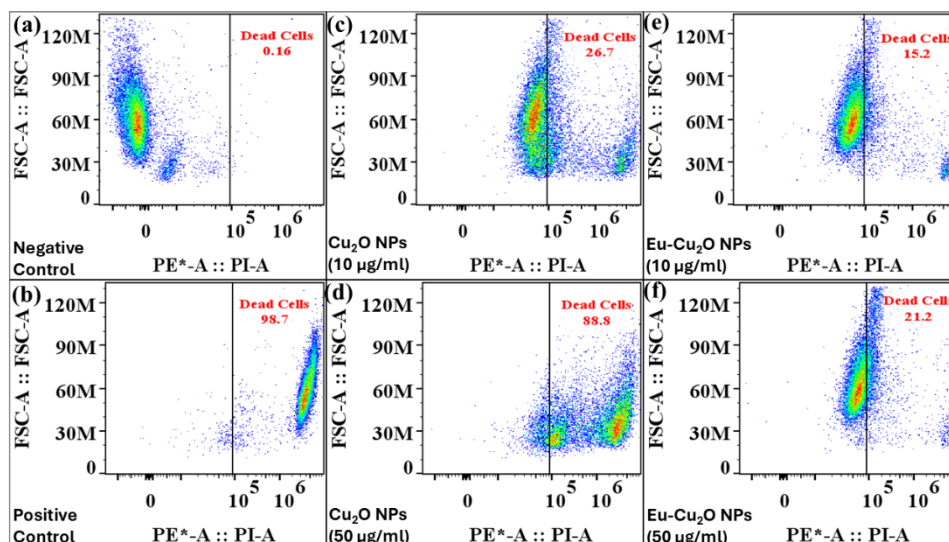
**Fig. S2.** TEM images of (a) Cu<sub>2</sub>O NPs and (b) Eu-Cu<sub>2</sub>O NPs.



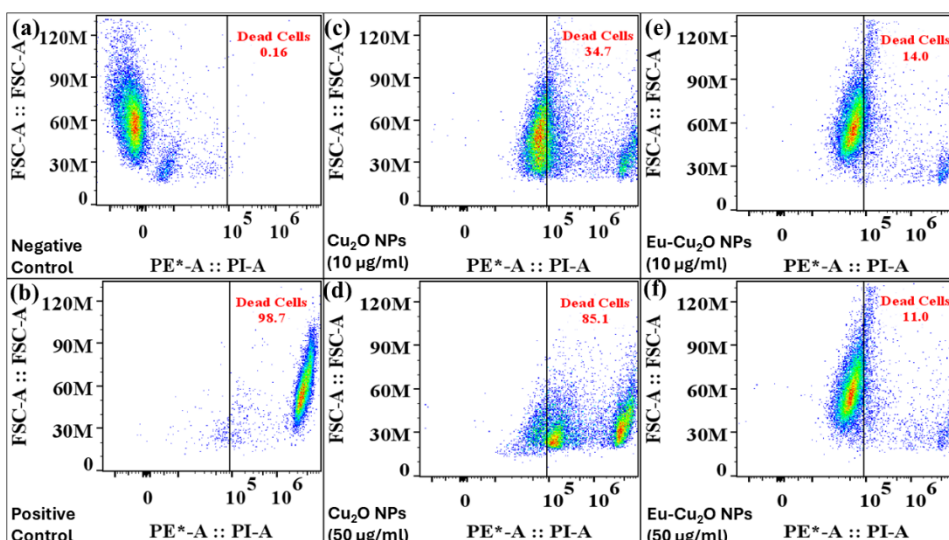
**Fig. S3.** XPS Elemental spectrum (Cu 2p) of pristine Cu<sub>2</sub>O NPs



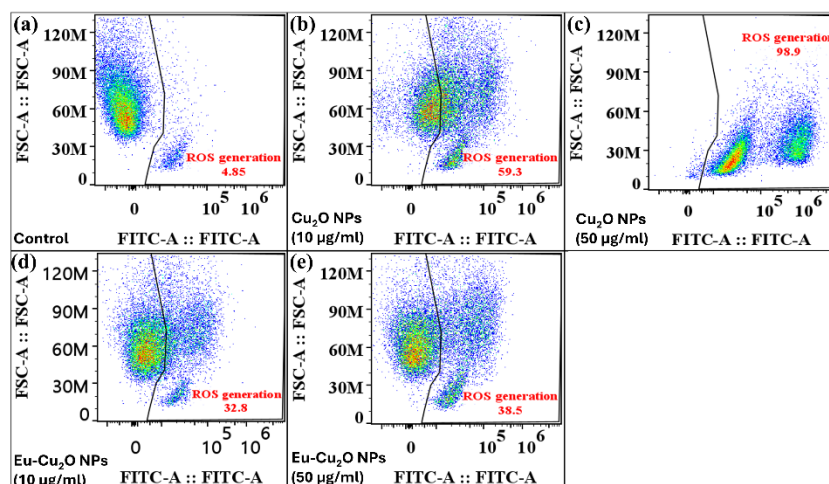
**Fig. S4.** Tauc Plot for direct band gap energy analysis of Eu-Cu<sub>2</sub>O NPs and Cu<sub>2</sub>O NPs.



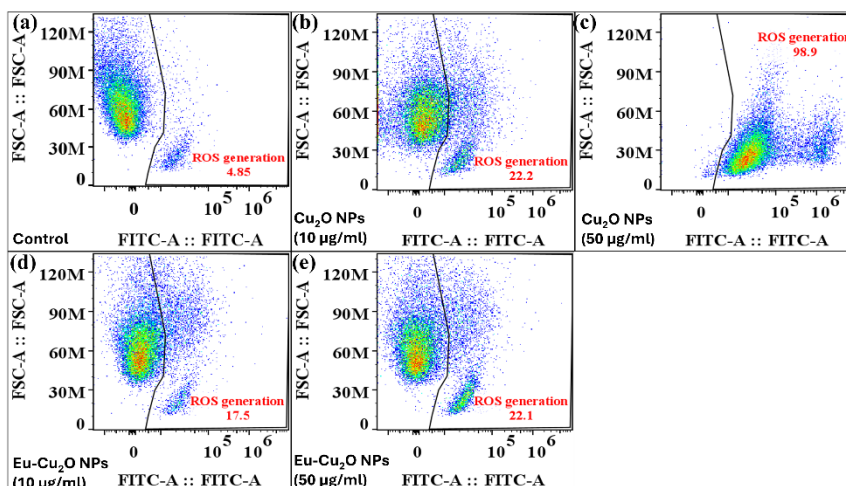
**Fig. S5.** Cell viability assay (dark) using flow cytometry with propidium iodide staining. (a) Untreated cells (b) induced dead cells (c) Cells treated with Cu<sub>2</sub>O NPs (10 µg/mL) (d) Cells treated with Cu<sub>2</sub>O NPs (50 µg/mL) (e) Cells treated with Eu-Cu<sub>2</sub>O NPs (10 µg/mL) (f) Cells treated with Eu-Cu<sub>2</sub>O NPs (50 µg/mL).



**Fig. S6.** Cell viability assay (under NIR irradiation) using flow cytometry with propidium iodide staining. (a) Untreated cells (b) induced dead cells (c) Cells treated with Cu<sub>2</sub>O NPs (10 µg/mL) (d) Cells treated with Cu<sub>2</sub>O NPs (50 µg/mL) (e) Cells treated with Eu-Cu<sub>2</sub>O NPs (10 µg/mL) (f) Cells treated with Eu-Cu<sub>2</sub>O NPs (50 µg/mL).

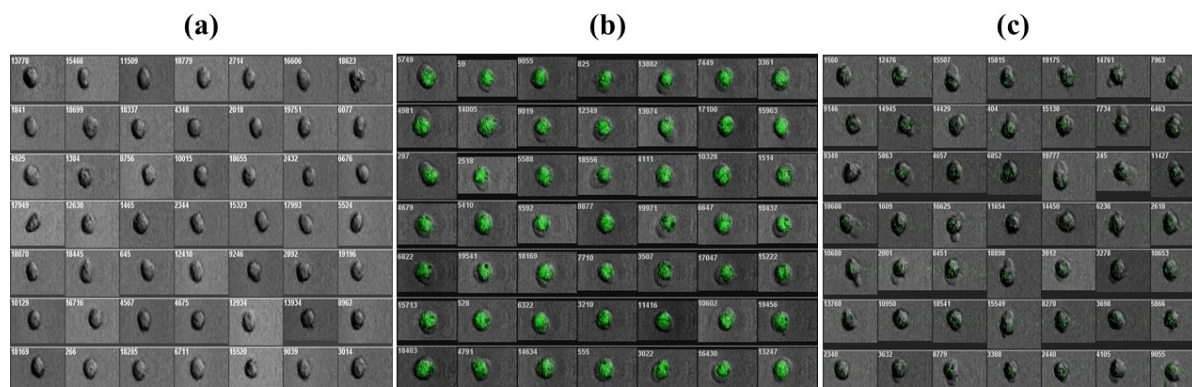


**Fig. S7.** DCFH-DA assay (dark) for ROS detection using flow cytometry in (a) Untreated cells (b) Cells treated with Cu<sub>2</sub>O NPs (10 µg/mL) (c) Cells treated with Cu<sub>2</sub>O NPs (50 µg/mL) (d) Cells treated with Eu-Cu<sub>2</sub>O NPs (10 µg/mL) (e) Cells treated with Eu-Cu<sub>2</sub>O NPs (50 µg/mL)

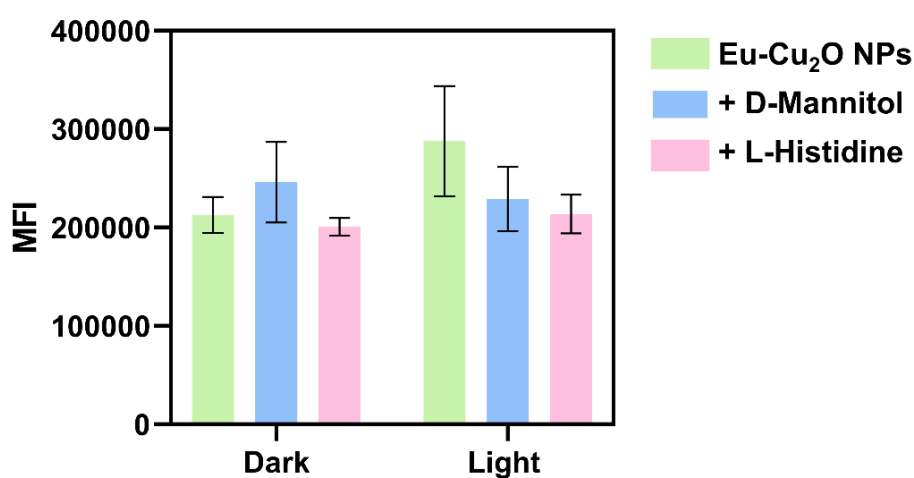


**Fig. S8.** DCFH-DA assay (NIR irradiated) for ROS detection using flow cytometry in (a) Untreated cells (b) Cells treated with Cu<sub>2</sub>O NPs (10 µg/mL) (c) Cells treated with Cu<sub>2</sub>O NPs (50 µg/mL) (d) Cells treated with Eu-Cu<sub>2</sub>O NPs (10 µg/mL) (e) Cells treated with Eu-Cu<sub>2</sub>O NPs (50 µg/ml)





**Fig. S9.** Representative images of cells as obtained from Image enabled Flow Cytometry (BD FACS Discover S8) for (a) untreated (b) Cu<sub>2</sub>O NPs (50 µg/mL) and (c) Eu-Cu<sub>2</sub>O NPs (50 µg/mL) treated cells for estimation of ROS generation on light irradiation using DCFH-DA Assay. Green fluorescence indicates total ROS generation. The images have been processed



using BD CellView.

**Fig. S10.** ROS quenching studies to study ROS generation by Eu-Cu<sub>2</sub>O NPs through the flow cytometric approach, using DCFH-DA as the fluorescent probe and ROS quenchers (D-mannitol and L-histidine). (n=3)

#### Supplementary References:

[S1] Z.Y Zhang, Y. D. Xu, Y. Y Ma, L. L Qiu, Y. Wang, J. L. Kong, H.M Xiong, *Angew. Chem.*, 2013, **52(15)**,4127-4131.

- [S2] E. Alpaslan, H. Yazici, N. H. Golshan, K. S. Ziemer and T. J. Webster, 2015, *ACS Biomater Sci Eng*, **1**, 1096–1103.
- [S3] W Ren, L Zeng, Z Shen, L Xiang, A Gong, J Zhang, C Mao, A Li, T Paunesku, G E. Woloschak, N S. Hosmaned, A Wu, 2013, *RSC Adv.*, **3**, 20855.