

Supporting Information

Palladium-Catalyzed Regio- and Stereoselective Phosphination of Cyclic Biarylsulfonium Salts to Access Atropoisomeric Phosphines

Jinghui Sun,^{a+} Yifei Yan,^{a+} Xuanxuan Chen,^a Zhiwei Huang^a and Yinhua Huang^{a*}

^a College of Materials, Chemistry and Chemical Engineering, Hangzhou Normal University,
Hangzhou 311121, China

⁺These authors contributed equally

Email: yhhuang@hznu.edu.cn

Table of Contents:

1. General Information	2
2. Experimental Details	2
2.1 Preparation of Substrates.....	2
2.2 Optimization of Conditions.....	14
2.3 Synthesis of the Atropoisomeric Phosphine Products.....	16
3. Scale-Up Synthesis of Enantiomerically Pure 3aa and 3ae	16
4. Mechanistic Considerations	18
4.1 Radical Trapping Experiments.....	18
4.2 The reaction of 1a' with 2a	19
4.3 Proposed Catalytic Cycle	19
5. Application of 3aa/3ae as a Chiral Ligand	20
6. Characterization of the Products	28
7. References	46
8. X-Ray Crystallographic Data of 1a , 3aa , 3ae	48
9. NMR Spectra and Chiral HPLC Charts	54

1. General Information

All air-sensitive manipulations were carried out with standard Schlenk techniques under nitrogen or argon. Solvents were degassed prior to use when necessary. NMR spectra were recorded on Bruker AMX 500 spectrophotometer (500 MHz for ^1H , 126 MHz for ^{13}C , 202 MHz for ^{31}P and 471 Hz for ^{19}F). Chemical shifts are reported in δ (ppm) referenced to an internal SiMe_4 standard ($\delta = 0$ ppm) for ^1H NMR, chloroform-*d* ($\delta = 77.0$ ppm) for ^{13}C NMR. The following abbreviations were used; s: singlet, d: doublet, t: triplet, q: quartet, m: multiplet, br: broad. Optical rotations were measured on an Anton Paar MCP 200 polarimeter. HRMS (ESI-TOF) were recorded on a time-of-flight (TOF) LC/MS instrument. Flash column chromatography was performed with Silica gel 60 (Merck) or Al_2O_3 (activated 200) (Merck). X-ray diffraction studies were performed on a 'Bruker D8 Venture' diffractometer. Enantiomeric excesses (ee%) were determined by HPLC analysis on Shimadzu HPLC with Daicel or Chiralcel chiral columns. All chemicals and solvents were purchased from commercial companies (Macklin, Energy Chemical, Sigma-Aldrich) and used as received. Solvents were degassed before use if necessary.

2. Experimental Details

2.1 Preparation of Substrates

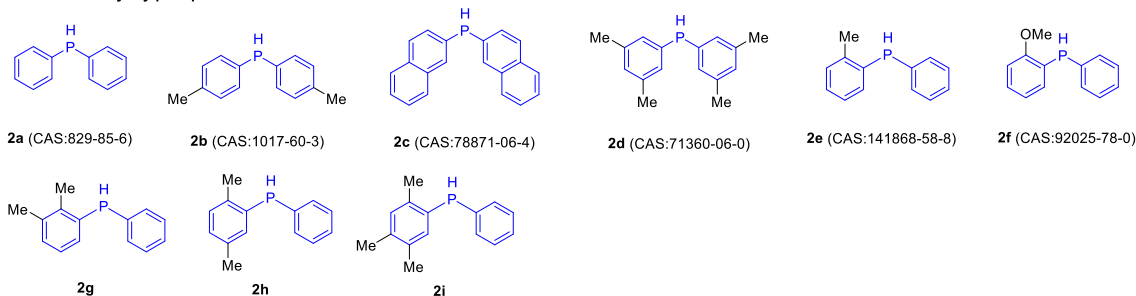
The cyclic biarylsulfonium salt substrates **1a-1c**, **1e-1x** are new compounds and they were prepared by the method shown below.^{1,2} **1d** (CAS: 2764681-19-6)³ is a known compound and was prepared by the same method as shown below. The good cyclic biarylsulfonium salt substrates and the limitation of the cyclic biarylsulfonium salt substrates which we tested are shown below (Scheme S1).

The secondary arylphosphine substrates **2a** (CAS: 829-85-6), **2d** (CAS: 71360-06-0) was purchased from commercial company and used as received. **2b** (CAS: 1017-60-3),⁴ **2c** (CAS: 78871-06-4),⁴ **2e** (CAS: 141868-58-8),⁵ **2f** (CAS: 92025-78-0),⁶ **2h** (CAS: 2764725-33-7)⁷ are known compounds which were prepared according to the reported literature procedure. **2g** and **2i** are new compounds which were prepared according to the reported method.⁵ The good secondary arylphosphine substrates and the limitation of the secondary phosphine substrates which we tested are shown below (Scheme S2).

The figure displays 17 chemical structures, labeled 1a through 1v. Structures 1a-1c, 1e, and 1f are 1,1'-bis(phenylthio)anthracene derivatives with various substituents on the phenyl rings: 1a (4-methyl), 1b (4-methyl, 2-methyl), 1c (4-methyl, 2-methyl, 3-methyl), 1e (4-ethyl), and 1f (4-methoxy). Structure 1d is the precursor 1,1'-bis(phenylthio)anthracene (CAS: 2764681-19-6). Structures 1g-1l are 1,1'-bis(phenylthio)anthracene derivatives with various substituents on the phenyl rings: 1g (4-phenyl), 1h (4-phenyl, 2-phenyl), 1i (4-fluoro), 1j (4-chloro), 1k (4-bromo), and 1l (4-(trifluoromethyl)). Structures 1m-1q are 1,1'-bis(phenylthio)anthracene derivatives with various substituents on the phenyl rings: 1m (4-phenyl), 1n (4-methyl, 2-methyl, 3-methyl), 1o (4-methyl, 2-methyl, 3-methyl, 5-methyl), 1p (4-methyl, 2-bromo, 3-bromo), and 1q (4-methyl, 2-phenyl, 3-phenyl). Structures 1r-1v are 1,1'-bis(phenylthio)anthracene derivatives with various substituents on the phenyl rings: 1r (4-methyl, 2-phenyl, 3-phenyl), 1s (4-methyl, 2-tolyl, 3-tolyl), 1t (4-methyl, 2-adamantyl, 3-adamantyl), 1u (4-methyl, 2-bromo, 3-bromo), and 1v (4-methyl, 2-(4-thienylphenyl), 3-(4-thienylphenyl)).

1a' (no reaction, at 25 °C)
 1a'' (no reaction, at 25 °C)
 1w (no reaction, at 25 °C)
 1x (no reaction, at 25 °C)
 (58%, 5% ee) (55 °C)

Good secondary arylphosphine substrates



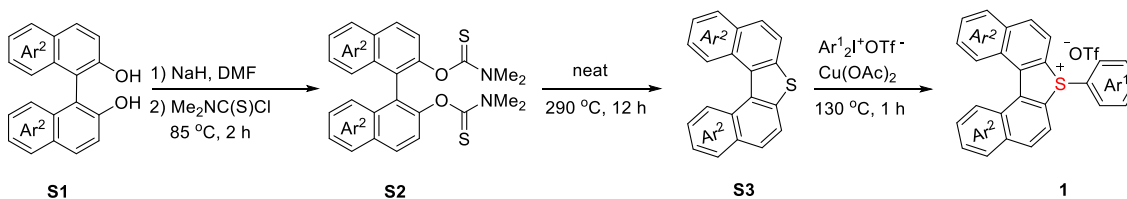
2j
 (no reaction, at 25 °C)
 36%, 73% ee (45 °C)

2k
 (no reaction, at 25 °C)

2l
 (no reaction, at 25 °C)

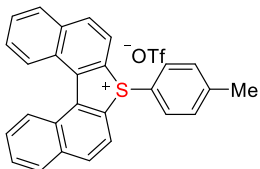
2m (CAS:829-84-5)

2.1.1 General Procedure A (GPA) for the Synthesis of Substrates **1**¹⁻³



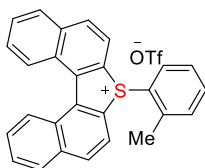
To a solution of binols **S1**⁸ (10 mmol) in dry DMF (20 mL) was added NaH (60% oil dispersion, 0.88 g, 22 mmol) in portions at 0 °C. The resulting yellow mixture was added dimethylthiocarbamoyl chloride (2.8 g, 22 mmol), and the solution was heated to 85 °C and kept stirring for 2 h. After completion, the reaction mixture was cooled to room temperature and quenched with 1 N KOH (80 mL) at 0 °C. The precipitate was filtered, washed thoroughly with water, and dissolved in CH₂Cl₂. The CH₂Cl₂ solution was dried over Na₂SO₄. After evaporation of the solvent, the crude solid was recrystallized from CH₂Cl₂/petroleum ether to afford the intermediate **S2** which was heated in an autoclave at 290 °C for 12 h. The reaction mixture was cooled to room temperature and then purified by column chromatography on silica gel (hexane/ethyl acetate = 95/5) to afford the intermediate **S3** which was directed used for next step.^{1,2}

Under an inert atmosphere, an oven dried flask was charged with the above obtained **S3** (0.50 mmol, 1.0 equiv), diaryliodonium salt Ar₁₂I⁺OTf⁻ (0.50 mmol, 1.0 equiv), copper (II) acetate (9.1 mg, 0.050 mmol, 10 mol%). The mixture was then heated to 130 °C and kept vigorous stirring for 1 h. After completion, the mixture was cooled to room temperature and concentrated under reduced pressure. The residue was purified by chromatography on silica gel (eluting with PE/EA 90/10, then DCM/MeOH 92/8) to afford the crude product. The crude product was the further purified by recrystallization (with ether), and the solid was collected by filtration and dried under vacuum to give the substrate **1**.³

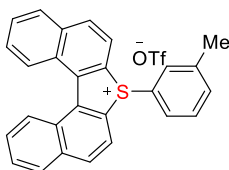


7-(*p*-Tolyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1a**).** Following General Procedure A (GPA), **1a** was prepared as a brown solid (139.4 mg, 53% yield). ¹H NMR (500 MHz, CDCl₃) δ 8.57 (d,

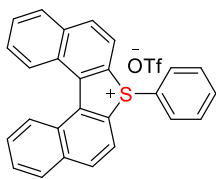
$J = 8.5$ Hz, 2H), 8.19 (d, $J = 8.7$ Hz, 2H), 8.16 (d, $J = 8.7$ Hz, 2H), 8.11 (d, $J = 7.8$ Hz, 2H), 7.80 (t, $J = 7.7$ Hz, 2H), 7.75 (t, $J = 7.7$ Hz, 2H), 7.60 (d, $J = 8.4$ Hz, 2H), 7.29 (d, $J = 8.4$ Hz, 2H), 2.36 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 147.0, 137.8, 136.6, 133.2, 132.5, 131.1, 130.9, 129.5, 129.4, 129.1, 127.6, 127.4, 121.9, 121.0 (q, $J = 320.8$ Hz), 120.8, 21.7. ^{19}F NMR (471 MHz, CDCl_3) δ -78.1. HRMS (ESI-TOF) m/z : $[\text{M}]^+$ Calcd for $\text{C}_{27}\text{H}_{19}\text{S}^+$: 375.1202; Found: 375.1207.



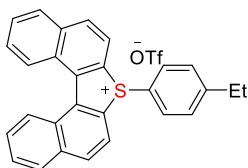
7-(*o*-Tolyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1b). Following General Procedure A (GPA), **1b** was prepared as a brown solid (89.4 mg, 34% yield). ^1H NMR (500 MHz, CDCl_3) δ 8.56 (d, $J = 8.5$ Hz, 2H), 8.22 (d, $J = 8.5$ Hz, 2H), 8.13-8.10 (m, 4H), 7.81 (t, $J = 7.6$ Hz, 2H), 7.75 (t, $J = 7.6$ Hz, 2H), 7.52-7.51 (m, 2H), 7.12 (brs, 1H), 6.70 (brs, 1H), 3.00 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 143.9, 138.1, 136.6, 135.2, 133.8, 133.3, 130.5, 129.5, 129.4, 129.3, 129.2, 127.6, 127.5, 121.9, 120.8 (q, $J = 321.3$ Hz), 20.3. ^{19}F NMR (471 MHz, CDCl_3) δ -78.2. HRMS (ESI-TOF) m/z : $[\text{M}]^+$ Calcd for $\text{C}_{27}\text{H}_{19}\text{S}^+$: 375.1202; Found: 375.1202.



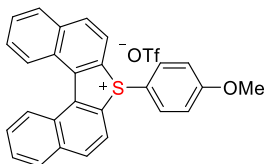
7-(*m*-Tolyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1c). Following General Procedure A (GPA), **1c** was prepared as a brown solid (105.2 mg, 40% yield). ^1H NMR (500 MHz, CDCl_3) δ 8.52 (d, $J = 8.5$ Hz, 2H), 8.13 (d, $J = 8.7$ Hz, 2H), 8.10 (d, $J = 8.7$ Hz, 2H), 8.05 (d, $J = 8.0$ Hz, 2H), 7.74 (t, $J = 7.5$ Hz, 2H), 7.69 (t, $J = 7.5$ Hz, 2H), 7.63 (s, 1H), 7.36 (t, $J = 4.6$ Hz, 1H), 7.27 (d, $J = 4.6$ Hz, 2H), 2.28 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 142.8, 137.9, 136.7, 136.3, 133.3, 131.6, 130.6, 129.5, 129.4, 129.1, 127.7, 127.4, 124.4, 122.0, 120.9 (q, $J = 321.3$ Hz), 21.3. ^{19}F NMR (471 MHz, CDCl_3) δ -78.2. HRMS (ESI-TOF) m/z : $[\text{M}]^+$ Calcd for $\text{C}_{27}\text{H}_{19}\text{S}^+$: 375.1202; Found: 375.1205.



7-Phenyl-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1d) (CAS: 2764681-19-6).³ Following General Procedure A (GPA), **1d** was prepared as a brown solid (161.3 mg, 63% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.59 (d, *J* = 8.5 Hz, 2H), 8.24-8.20 (m, 4H), 8.14 (d, *J* = 8.1 Hz, 2H), 7.84 (t, *J* = 7.4 Hz, 2H), 7.79-7.75 (m, 4H), 7.66 (t, *J* = 7.5 Hz, 1H), 7.52 (t, *J* = 7.5 Hz, 2H). **¹³C NMR** (126 MHz, CDCl₃) δ 137.9, 136.6, 135.2, 133.3, 131.8, 131.0, 130.8, 129.5, 129.1, 127.7, 127.4, 125.0, 122.1, 121.3 (q, *J* = 321.3 Hz). **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.1. **HRMS** (ESI-TOF) *m/z*: [M]⁺ Calcd for C₂₆H₁₇S⁺: 361.1045; Found: 361.1048.

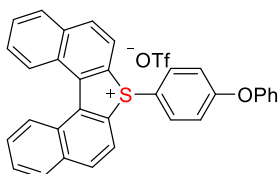


7-(4-Ethylphenyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1e). Following General Procedure A (GPA), **1e** was prepared as a brown solid (183.6 mg, 68% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.53 (d, *J* = 8.5 Hz, 2H), 8.18 (d, *J* = 8.4 Hz, 2H), 8.13 (d, *J* = 8.4 Hz, 2H), 8.09 (d, *J* = 8.1 Hz, 2H), 7.79 (t, *J* = 7.6 Hz, 2H), 7.73 (t, *J* = 7.6 Hz, 2H), 7.59 (d, *J* = 8.0 Hz, 2H), 7.30 (d, *J* = 8.0 Hz, 2H), 2.63 (q, *J* = 7.6 Hz, 2H), 1.16 (t, *J* = 7.6 Hz, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 152.9, 137.7, 136.6, 133.1, 131.4, 131.2, 131.1, 129.5, 129.4, 129.1, 127.6, 127.4, 122.0, 121.0, 121.7 (q, *J* = 321.0 Hz), 28.9, 14.7. **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.1. **HRMS** (ESI-TOF) *m/z*: [M]⁺ Calcd for C₂₈H₂₁S⁺: 389.1358; Found: 389.1353.

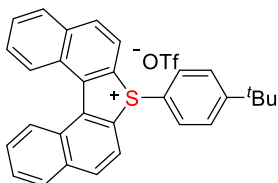


7-(4-Methoxyphenyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1f). Following General Procedure A (GPA), **1f** was prepared as a brown solid (62.3 mg, 23% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.52 (d, *J* = 8.2 Hz, 2H), 8.14 (d, *J* = 8.2 Hz, 2H), 8.07 (d, *J* = 8.1 Hz, 2H), 8.06 (d, *J* = 8.1 Hz, 2H), 7.70-7.69 (m, 4H), 7.46 (d, *J* = 8.1 Hz, 2H), 6.88 (d, *J* = 8.1 Hz, 2H), 3.72 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 165.3, 137.5, 136.5, 133.3, 133.1, 131.5, 129.5, 129.4,

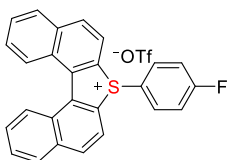
129.0, 127.6, 127.4, 121.6, 117.5, 112.4, 56.1. **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.1. **HRMS** (ESI-TOF) m/z: [M]⁺ Calcd for C₂₇H₁₉OS⁺: 391.1151; Found: 391.1156.



7-(4-Phenoxyphenyl)-7H-dinaphtho[2,1-b:1',2'-d]thiophen-7-ium trifluoromethanesulfonate (1g). Following General Procedure A (GPA), **1g** was prepared as a brown solid (166.1 mg, 55% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.41 (d, *J* = 8.6 Hz, 2H), 8.11 (d, *J* = 8.6 Hz, 2H), 8.07 (d, *J* = 8.6 Hz, 2H), 8.01 (d, *J* = 8.2 Hz, 2H), 7.70 (t, *J* = 7.6 Hz, 2H), 7.63 (t, *J* = 7.6 Hz, 2H), 7.53 (d, *J* = 8.8 Hz, 2H), 7.27 (t, *J* = 7.8 Hz, 2H), 7.11 (t, *J* = 7.8 Hz, 1H), 6.90 (d, *J* = 8.2 Hz, 2H), 6.88 (d, *J* = 8.8 Hz, 2H). **¹³C NMR** (126 MHz, CDCl₃) δ 164.2, 153.9, 137.6, 136.6, 133.5, 133.2, 131.5, 130.4, 129.5, 129.4, 129.1, 127.6, 127.4, 125.8, 122.0, 120.9 (d, *J* = 321.0 Hz), 120.7, 119.7, 115.0. **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.0. **HRMS** (ESI-TOF) m/z: [M+H]⁺ Calcd for C₃₂H₂₁SOH⁺: 454.1386; Found: 454.1387.

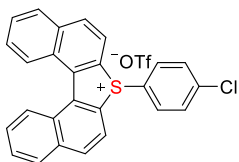


7-(4-(Tert-butyl)phenyl)-7H-dinaphtho[2,1-b:1',2'-d]thiophen-7-ium trifluoromethanesulfonate (1h). Following General Procedure A (GPA), **1h** was prepared as a brown solid (247.1 mg, 87% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.46 (d, *J* = 8.5 Hz, 2H), 8.11 (d, *J* = 8.5 Hz, 2H), 8.07 (d, *J* = 8.5 Hz, 2H), 8.02 (d, *J* = 8.1 Hz, 2H), 7.71 (t, *J* = 7.5 Hz, 2H), 7.65 (t, *J* = 7.5 Hz, 2H), 7.53 (d, *J* = 8.1 Hz, 2H), 7.42 (d, *J* = 8.1 Hz, 2H), 1.16 (s, 9H). **¹³C NMR** (126 MHz, CDCl₃) δ 159.7, 137.8, 136.6, 133.2, 131.2, 130.8, 129.5, 129.4, 129.1, 129.0, 127.6, 127.4, 122.1, 120.8 (q, *J* = 320.0 Hz), 120.7, 35.5, 30.8. **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.1. **HRMS** (ESI-TOF) m/z: [M]⁺ Calcd for C₃₀H₂₅S⁺: 417.1671; Found: 417.1672.

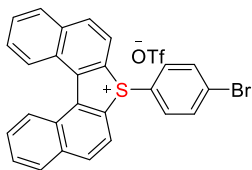


7-(4-Fluorophenyl)-7H-dinaphtho[2,1-b:1',2'-d]thiophen-7-ium trifluoromethanesulfonate (1i). Following General Procedure A (GPA), **1i** was prepared

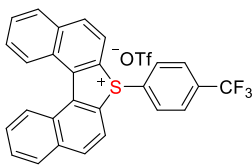
as a brown solid (185.5 mg, 70% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.59 (d, *J* = 8.5 Hz, 2H), 8.21 (brs, 4H), 8.12 (d, *J* = 8.1 Hz, 2H), 7.84-7.75 (m, 6H), 7.12 (t, *J* = 7.7 Hz, 2H). **¹³C NMR** (126 MHz, CDCl₃) δ 166.6 (d, *J* = 260.6 Hz), 137.8, 136.7, 134.1 (d, *J* = 10.0 Hz), 133.3, 30.9, 129.6, 129.5, 129.1, 127.7, 127.4, 122.0, 120.9 (q, *J* = 314.8 Hz), 120.0, 119.3 (d, *J* = 23.5 Hz). **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.2, -99.5. **HRMS** (ESI-TOF) *m/z*: [M]⁺ Calcd for C₂₆H₁₆SF⁺: 379.0951; Found: 379.0956.



7-(4-Chlorophenyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1j). Following General Procedure A (GPA), **1j** was prepared as a brown solid (226.2 mg, 83% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.52 (d, *J* = 8.5 Hz, 2H), 8.14 (brs, 4H), 8.06 (d, *J* = 8.1 Hz, 2H), 7.76 (t, *J* = 7.6 Hz, 2H), 7.70 (t, *J* = 7.6 Hz, 2H), 7.64 (d, *J* = 8.7 Hz, 2H), 7.39 (d, *J* = 8.7 Hz, 2H). **¹³C NMR** (126 MHz, CDCl₃) δ 142.3, 138.0, 136.7, 133.4, 132.4, 132.0, 130.8, 129.6, 129.5, 129.2, 127.7, 127.5, 123.4, 122.2, 120.7 (q, *J* = 320.8 Hz). **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.2. **HRMS** (ESI-TOF) *m/z*: [M]⁺ Calcd for C₂₆H₁₆SCl⁺: 395.0656; Found: 395.0652.

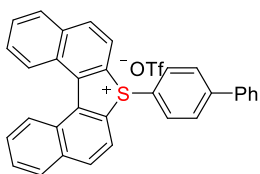


7-(4-Bromophenyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1k). Following General Procedure A (GPA), **1k** was prepared as a brown solid (205.8 mg, 70% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.59 (d, *J* = 8.5 Hz, 2H), 8.23 (brs, 4H), 8.14 (d, *J* = 7.5 Hz, 2H), 7.84 (t, *J* = 7.5 Hz, 2H), 7.78 (t, *J* = 7.5 Hz, 2H), 7.64 (brs, 4H). **¹³C NMR** (126 MHz, CDCl₃) δ 138.0, 136.7, 135.0, 133.4, 132.4, 130.8, 130.6, 129.6, 129.5, 129.2, 127.7, 127.5, 124.1, 122.2, 120.8 (q, *J* = 320.1 Hz). **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.2. **HRMS** (ESI-TOF) *m/z*: [M]⁺ Calcd for C₂₆H₁₆SBr⁺: 439.0151; Found: 439.0147.

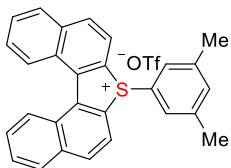


7-(4-(Trifluoromethyl)phenyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1l). Following General Procedure A

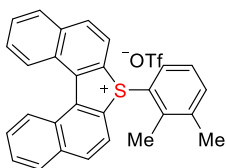
(GPA), **1l** was prepared as a brown solid (95.7 mg, 33% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.48 (d, *J* = 8.6 Hz, 2H), 8.19 (d, *J* = 8.6 Hz, 2H), 8.14 (d, *J* = 8.6 Hz, 2H), 8.04 (d, *J* = 8.1 Hz, 2H), 7.84 (d, *J* = 8.4 Hz, 2H), 7.75 (t, *J* = 7.4 Hz, 2H), 7.70-7.64 (m, 4H). **¹³C NMR** (126 MHz, CDCl₃) δ 138.3, 136.8, 136.4 (q, *J* = 34.0 Hz), 133.5, 131.4, 130.3, 130.0, 129.6, 129.5, 129.1, 128.5 (q, *J* = 3.8 Hz), 127.8, 127.4, 122.5 (q, *J* = 274.1 Hz), 122.2, 120.5 (q, *J* = 320.8 Hz). **¹⁹F NMR** (471 MHz, CDCl₃) δ -63.6, -78.2. **HRMS** (ESI-TOF) *m/z*: [M]⁺ Calcd for C₂₇H₁₆SF₃⁺: 429.0919; Found: 429.0914.



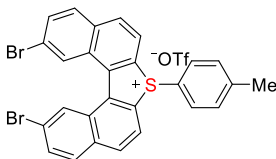
7-([1,1'-biphenyl]-4-yl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1m**)**. Following General Procedure A (GPA), **1m** was prepared as a brown solid (61.7 mg, 21% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.58 (d, *J* = 8.5 Hz, 2H), 8.20 (brs, 4H), 8.12 (d, *J* = 8.1 Hz, 2H), 7.83-7.74 (m, 6H), 7.67 (d, *J* = 8.1 Hz, 2H), 7.47 (d, *J* = 6.9 Hz, 2H), 7.43-7.37 (m, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 148.2, 138.0, 137.9, 136.6, 133.3, 131.5, 130.9, 130.2, 129.6, 129.5, 129.2, 129.1, 127.7, 127.4, 127.3, 122.7, 122.0, 121.2 (q, *J* = 321.3 Hz). **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.13. **HRMS** (ESI-TOF) *m/z*: [M]⁺ Calcd for C₃₂H₂₁S⁺: 437.1358; Found: 437.1356.



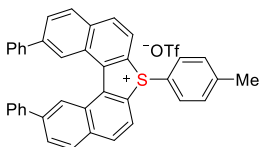
7-(3,5-Dimethylphenyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1n**)**. Following General Procedure A (GPA), **1n** was prepared as a brown solid (54.0 mg, 20% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.51 (d, *J* = 8.5 Hz, 2H), 8.14 (d, *J* = 8.5 Hz, 2H), 8.09 (d, *J* = 8.5 Hz, 2H), 8.05 (d, *J* = 7.8 Hz, 2H), 7.74 (t, *J* = 7.6 Hz, 2H), 7.68 (t, *J* = 7.6 Hz, 2H), 7.23 (s, 2H), 7.17 (s, 1H), 2.20 (s, 6H). **¹³C NMR** (126 MHz, CDCl₃) δ 142.4, 137.7, 137.3, 136.6, 133.2, 130.6, 129.5, 129.4, 129.1, 128.1, 127.6, 127.4, 123.7, 121.9, 121.0 (q, *J* = 321.3 Hz), 21.2. **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.13. **HRMS** (ESI-TOF) *m/z*: [M]⁺ Calcd for C₂₈H₂₁S⁺: 389.1358; Found: 389.1357.



7-(2,3-Dimethylphenyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1o). Following General Procedure A (GPA), **1o** was prepared as a brown solid (110.7 mg, 41% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.56 (d, *J* = 8.5 Hz, 2H), 8.12 (d, *J* = 8.7 Hz, 2H), 8.10 (d, *J* = 8.7 Hz, 2H), 7.80 (t, *J* = 7.5 Hz, 2H), 7.74 (t, *J* = 7.5 Hz, 2H), 7.35 (d, *J* = 7.3 Hz, 1H), 6.89-6.97 (m, 1H), 6.36 (brs, 1H), 3.06 (brs, 3H), 2.40 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 142.6, 141.8, 137.8, 136.5, 136.4, 133.4, 130.8, 129.6, 129.5, 129.2, 129.0, 127.7, 127.3, 121.8, 121.2 (q, *J* = 318.6 Hz), 20.8, 15.3. **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.2. **HRMS** (ESI-TOF) *m/z*: [M]⁺ Calcd for C₂₈H₂₁S⁺: 389.1358; Found: 389.1359.

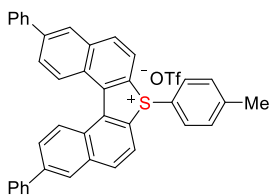


2,12-Dibromo-7-(*p*-tolyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1p). Following General Procedure A (GPA), **1p** was prepared as a brown solid (242.5 mg, 71% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.57 (s, 2H), 8.11 (s, 4H), 7.90 (d, *J* = 8.7 Hz, 2H), 7.75 (d, *J* = 8.7 Hz, 2H), 7.47 (d, *J* = 7.8 Hz, 2H), 7.21 (d, *J* = 7.8 Hz, 2H), 2.28 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 147.3, 135.8, 134.8, 133.3, 132.8, 132.6, 132.3, 131.1, 131.0, 129.4, 129.3, 122.6, 122.5, 120.6 (q, *J* = 320.0 Hz), 120.2, 21.7. **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.1. **HRMS** (ESI-TOF) *m/z*: [M]⁺ Calcd for C₂₇H₁₇SBBr₂⁺: 530.9412; Found: 530.9414.

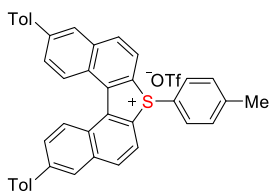


2,12-Diphenyl-7-(*p*-tolyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1q). Following General Procedure A (GPA), **1q** was prepared as a brown solid (169.0 mg, 50% yield). **¹H NMR** (400 MHz, CDCl₃) δ 8.53 (brs, 2H), 8.16 (d, *J* = 8.1 Hz, 2H), 8.06 (d, *J* = 8.5 Hz, 4H), 7.82 (d, *J* = 6.8 Hz, 2H), 7.51 (d, *J* = 8.0 Hz, 2H), 7.22-7.14 (m, 12H), 2.27 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 147.0, 140.5, 139.3, 137.6, 135.6, 133.0, 132.5, 131.0, 130.1, 129.4, 129.2, 129.1, 128.4,

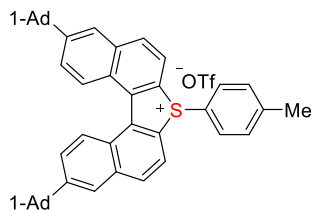
127.3, 125.3, 122.0, 121.2 (d, $J = 321.3$ Hz), 121.0, 21.7. ^{19}F NMR (471 MHz, CDCl_3) δ -78.0. **HRMS** (ESI-TOF) m/z : $[\text{M}]^+$ Calcd for $\text{C}_{39}\text{H}_{27}\text{S}^+$: 527.1828; Found: 527.1823.



3,11-Diphenyl-7-(*p*-tolyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1r). Following General Procedure A (GPA), **1r** was prepared as a brown solid (145.8 mg, 43% yield). ^1H NMR (500 MHz, CDCl_3) δ 8.58 (d, $J = 8.8$ Hz, 2H), 8.21-8.13 (m, 6H), 7.97 (d, $J = 8.8$ Hz, 2H), 7.77 (d, $J = 7.5$ Hz, 4H), 7.59-7.53 (m, 6H), 7.50-7.45 (m, 2H), 7.30 (d, $J = 8.4$ Hz, 2H), 2.36 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 146.9, 141.8, 138.9, 137.5, 137.0, 133.3, 132.5, 130.9, 130.8, 129.3, 128.7, 128.0, 127.9, 127.4, 127.0, 126.7, 122.5, 121.1, 121.0 (q, $J = 323.1$ Hz), 21.7. ^{19}F NMR (471 MHz, CDCl_3) δ -78.0. **HRMS** (ESI-TOF) m/z : $[\text{M}]^+$ Calcd for $\text{C}_{39}\text{H}_{27}\text{S}^+$: 527.1828; Found: 527.1828.

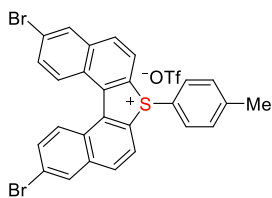


3,7,11-Tri-*p*-tolyl-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1s). Following General Procedure A (GPA), **1s** was prepared as a brown solid (91.8 mg, 26% yield). ^1H NMR (400 MHz, CDCl_3) δ 8.59 (d, $J = 8.9$ Hz, 2H), 8.20-8.10 (m, 6H), 7.96 (d, $J = 8.9$ Hz, 2H), 7.67 (d, $J = 8.0$ Hz, 4H), 7.56 (t, $J = 7.6$ Hz, 2H), 7.34 (d, $J = 7.9$ Hz, 4H), 7.28 (d, $J = 8.8$ Hz, 2H), 2.45 (s, 6H), 2.35 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 146.9, 141.6, 138.7, 137.5, 137.0, 135.9, 133.2, 132.5, 130.9, 130.5, 130.0, 127.8, 127.2, 126.8, 126.2, 122.3, 121.1 (d, $J = 321.9$ Hz), 21.7, 21.2. ^{19}F NMR (471 MHz, CDCl_3) δ -78.08. **HRMS** (ESI-TOF) m/z : $[\text{M}]^+$ Calcd for $\text{C}_{41}\text{H}_{31}\text{S}^+$: 555.2141; Found: 555.2144.

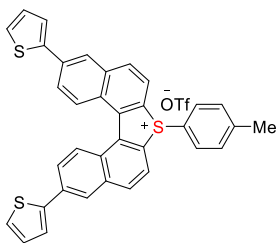


3,11-Di((3*r*,5*r*,7*r*)-adamantan-1-yl)-7-(*p*-tolyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1t). Following

General Procedure A (GPA), **1t** was prepared as a brown solid (119.1 mg, 30% yield). **¹H NMR** (400 MHz, CDCl₃) δ 8.55 (d, *J* = 9.0 Hz, 2H), 8.07 (d, *J* = 8.6 Hz, 2H), 8.01 (d, *J* = 8.6 Hz, 2H), 7.94 (s, 2H), 7.85 (d, *J* = 9.0 Hz, 2H), 7.56 (d, *J* = 8.1 Hz, 2H), 7.27 (d, *J* = 8.1 Hz, 2H), 2.35 (s, 3H), 2.20 (s, 6H), 2.07 (s, 12H), 1.89-1.80 (m, 12H). **¹³C NMR** (126 MHz, CDCl₃) δ 152.8, 146.7, 137.7, 133.1, 132.4, 130.9, 130.3, 127.5, 127.4, 125.9, 124.5, 122.0, 120.6 (d, *J* = 321.0 Hz), 42.9, 36.8, 36.7, 28.8, 21.7. **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.1. **HRMS** (ESI-TOF) *m/z*: [M]⁺ Calcd for C₄₇H₄₇S⁺: 643.3393; Found: 643.3388.

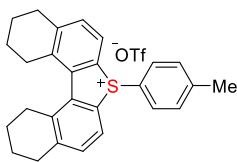


3,11-Dibromo-7-(*p*-tolyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1u**)**. Following General Procedure A (GPA), **1u** was prepared as a brown solid (47.8 mg, 14% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.68-8.65 (m, 2H), 8.16 (d, *J* = 2.1 Hz, 1H), 7.88 (d, *J* = 8.6 Hz, 2H), 7.84 (d, *J* = 8.6 Hz, 2H), 7.80 (d, *J* = 1.8 Hz, 1H), 7.82-7.78 (m, 2H), 7.53 (dd, *J* = 2.1, 9.1 Hz, 1H), 7.35 (d, *J* = 8.2 Hz, 2H), 7.19 (d, *J* = 8.7 Hz, 1H), 7.13 (d, *J* = 8.2 Hz, 2H), 2.31 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 142.6, 137.3, 132.6, 132.3, 131.6, 131.5, 131.1, 130.9, 128.3, 127.3, 124.0, 123.2, 120.5 (d, *J* = 308.8 Hz), 119.5, 21.7. **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.0. **HRMS** (ESI-TOF) *m/z*: [M]⁺ Calcd for C₂₇H₁₇SBr₂⁺: 530.9412; Found: 530.9417.



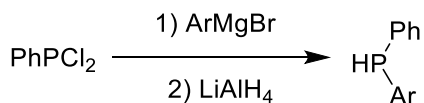
3,11-Di(thiophen-2-yl)-7-(*p*-tolyl)-7*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1v**)**. Following General Procedure A (GPA), **1v** was prepared as a brown solid (324.3 mg, 47% yield). **¹H NMR** (500 MHz, CDCl₃) δ 8.27 (d, *J* = 8.9 Hz, 2H), 8.03-7.91 (m, 6H), 7.80 (d, *J* = 8.6 Hz, 2H), 7.66 (s, 2H), 7.49-7.41 (m, 6H), 7.21 (d, *J* = 7.9 Hz, 2H), 2.29 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 146.9, 140.0, 137.2, 137.0, 136.1, 133.1, 132.4, 130.8, 130.1, 127.7, 127.6, 127.2, 126.3, 126.0, 125.3, 122.8, 122.2, 121.1 (d, *J* = 321.3 Hz), 121.0, 21.7. **¹⁹F NMR**

(471 MHz, CDCl₃) δ -78.0. **HRMS** (ESI-TOF) m/z : [M]⁺ Calcd for C₃₅H₂₃S₃⁺: 539.0956; Found: 539.0958.

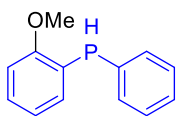


7-(*p*-Tolyl)-2,3,4,7,10,11,12,13-octahydro-1*H*-dinaphtho[2,1-*b*:1',2'-*d*]thiophen-7-ium trifluoromethanesulfonate (1x). Following General Procedure A (GPA), **1x** was prepared as a brown solid (189.7 mg, 71% yield). **¹H NMR** (500 MHz, CDCl₃) δ 7.83 (d, J = 8.0 Hz, 2H), 7.48 (d, J = 8.0 Hz, 2H), 7.41 (d, J = 8.0 Hz, 2H), 7.32 (d, J = 8.0 Hz, 2H), 3.00-2.95 (m, 8H), 2.38 (s, 3H), 1.96-1.91 (m, 4H), 1.73-1.68 (m, 4H). **¹³C NMR** (126 MHz, CDCl₃) δ 146.4, 146.3, 139.9, 138.5, 132.3, 132.0, 130.2, 128.4, 125.7, 124.7, 120.9 (q, J = 321.3 Hz), 32.0, 28.9, 22.1, 21.6, 21.1. **¹⁹F NMR** (471 MHz, CDCl₃) δ -78.1. **HRMS** (ESI-TOF) m/z : [M]⁺ Calcd for C₂₇H₂₇S⁺: 383.1828; Found: 383.1828.

2.1.2 General Procedure B (GPB) for the Synthesis of Substrates 2⁵



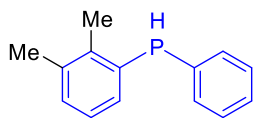
To a solution of PhPCl₂ (4.49 g, 25.1 mmol) in THF (50 mL) was added slowly ArMgBr (25.1 mmol in 50 mL of THF, prepared from ArBr (5.00 g, 25.1 mmol) and Mg (0.72 g, 30.1 mmol) in 50 mL of THF) at -80 °C under N₂, and the mixture was stirred at -80 °C for 2 h before it was warmed to room temperature and stirred for another 2 h. The mixture was cooled to 0 °C and LiAlH₄ (1.14 g, 30.1 mmol) was added slowly. The mixture was kept stirring at room temperature for 2 h. Degassed water (10 mL) was added dropwise at 0 °C and the mixture was extracted with ether (40 mL × 3). The organic layer was combined and dried over MgSO₄. The solvent was removed by distillation, the product was prepared and purified by high vacuum microdistillation as a colorless liquid.



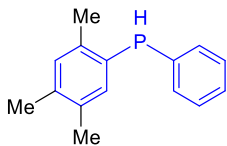
(2-Methoxyphenyl)(phenyl)phosphane (2f) (CAS: 92025-78-0).⁶

Following General Procedure B (GPB), **2f** was prepared as a colorless liquid (1.2 g, 23%

yield). **¹H NMR** (500 MHz, CDCl₃) δ 7.42-7.38 (m, 2H), 7.17-7.16 (m, 2H), 7.14-7.09 (m, 1H), 7.04-7.01 (m, 1H), 6.77-6.67 (m, 3H), 5.60 (brs, 1H), 3.63 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 159.4 (d, *J* = 10.1 Hz), 155.1, 133.7 (d, *J* = 7.6 Hz), 133.5 (d, *J* = 17.6 Hz), 129.4, 128.4, 127.6, 127.4 (d, *J* = 6.3 Hz), 120.0 (d, *J* = 2.5 Hz), 119.1, 114.5, 109.2 (d, *J* = 1.1 Hz), 54.5. **³¹P NMR** (202 MHz, CDCl₃) δ -54.4 (s, 1P). **HRMS** (ESI-TOF) *m/z*: [M+H]⁺ Calcd for C₁₃H₁₃OPH⁺ 217.0777; Found 217.0778.



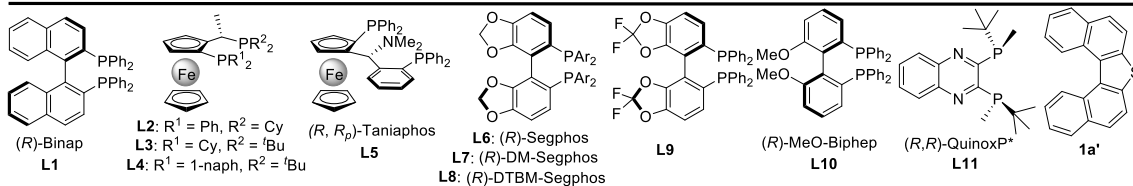
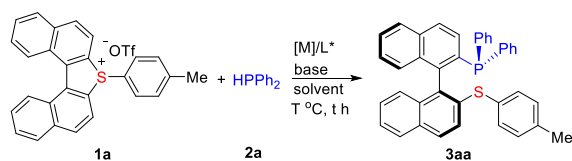
(2,3-Dimethylphenyl)(phenyl)phosphane (2g). Following General Procedure B (GPB), **2g** was prepared as a colorless liquid (2.8 g, 52% yield). **¹H NMR** (500 MHz, CDCl₃) δ 7.41-7.38 (m, 2H), 7.23-7.19 (m, 4H), 7.08-7.07 (m, 1H), 7.00-6.97 (m, 1H), 5.35-4.94 (brs, 1H), 2.29 (s, 3H), 2.21 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 140.1 (d, *J* = 17.6 Hz), 137.2 (d, *J* = 3.8 Hz), 134.3 (d, *J* = 17.4 Hz), 134.2 (d, *J* = 20.6 Hz), 133.2 (d, *J* = 6.3 Hz), 130.9, 128.8, 128.7, 128.6, 126.0 (d, *J* = 2.5 Hz), 20.8, 18.4 (d, *J* = 18.9 Hz). **³¹P NMR** (202 MHz, CDCl₃) δ -45.2 (s, 1P). **HRMS** (ESI-TOF) *m/z*: [M+K]⁺ Calcd for C₁₄H₁₅PK⁺: 253.0543; Found: 253.0539.



Phenyl(2,4,5-trimethylphenyl)phosphane (2i). Following General Procedure B (GPB), **2h** was prepared as a colorless liquid (4.5 g, 79% yield). **¹H NMR** (500 MHz, CDCl₃) δ 7.31-7.30 (m, 2H), 7.20-7.18 (m, 3H), 7.11 (d, *J* = 6.7 Hz, 1H), 6.89 (s, 1H), 4.71 (brs, 1H), 2.22 (s, 3H), 2.13 (s, 3H), 2.10 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 139.1 (d, *J* = 3.8 Hz), 137.8 (d, *J* = 5.4 Hz), 136.8 (d, *J* = 11.3 Hz), 136.7, 134.2, 133.8 (d, *J* = 17.6 Hz), 133.6 (d, *J* = 17.6 Hz), 131.7 (d, *J* = 3.8 Hz), 128.5 (d, *J* = 6.3 Hz), 128.2, 21.0 (d, *J* = 10.1 Hz), 19.5, 19.2. **³¹P NMR** (202 MHz, CDCl₃) δ -49.8 (s, 1P). **HRMS** (ESI-TOF) *m/z*: [M+K]⁺ Calcd for C₁₅H₁₇PK⁺: 267.0699; Found: 267.0695.

2.2 Optimization of Conditions

Table S1. Optimization of conditions^[a]



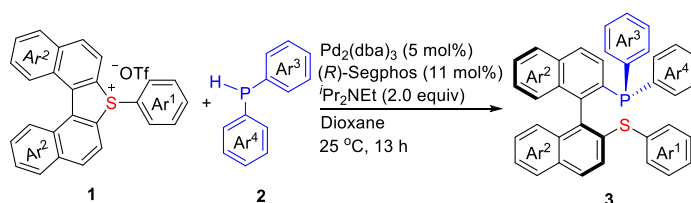
entry	1	L*	solvent	base	yield ^[b] (%)	ee ^[c] (%)
1	1a	L1	THF	Et ₃ N	96	62
2	1a	L2	THF	Et ₃ N	42	53
3	1a	L3	THF	Et ₃ N	58	5
4 ^[d]	1a	L4	THF	Et ₃ N	53	0
5	1a	L5	THF	Et ₃ N	93	65
6	1a	L6	THF	Et ₃ N	58	79
7	1a	L7	THF	Et ₃ N	72	52
8	1a	L8	THF	Et ₃ N	87	0
9	1a	L9	THF	Et ₃ N	39	58
10	1a	L10	THF	Et ₃ N	80	63
11	1a	L11	THF	Et ₃ N	10	-64
12	1a	L6	THF	Barton'base	41	<5
13	1a	L6	THF	Cs ₂ CO ₃	69	40
14	1a	L6	THF	K ₃ PO ₄	82	77
15	1a	L6	dioxane	K ₃ PO ₄	62	87
16	1a	L6	toluene	K ₃ PO ₄	85	64
17	1a	L6	MTBE	K ₃ PO ₄	91	60
18	1a	L6	DCE	K ₃ PO ₄	81	70
19 ^[e]	1a	L6	dioxane	K ₂ HPO ₄	64	90
20 ^[e]	1a	L6	dioxane	^t Pr ₂ NEt	74	90
21 ^{[e][f]}	1a	L6	dioxane	^t Pr ₂ NEt	95	93
22 ^[e]	1a'	L6	dioxane	^t Pr ₂ NEt	0	n.a.
23 ^g	1a	none	THF	Et ₃ N	35	-
24 ^h	1a	none	THF	Et ₃ N	42	-
25	1a	none	THF	Et ₃ N	46	-
26 ^h	1a	L3	THF	Et ₃ N	37.5	5

[a] Reaction conditions: **1** (0.120 mmol), **2a** (0.120 mmol), base (1.5 equiv), and Pd₂(dba)₃ (2.5 mol%) in solvent (4.0 mL) at 25 °C for 13 h, unless otherwise noted. THF = tetrahydrofuran, MTBE = methyl *tert*-butyl ether. [b] The yields were obtained by ¹H NMR analysis of the crude reaction mixture with the aid of Cl₂CHCHCl₂ as an internal standard. [c] The ee was determined by HPLC on a chiral stationary phase column. [d] The reaction was performed at 60 °C (no reaction at 25 °C). [e] Pd₂(dba)₃ (5 mol%). [f] **1a** (0.132 mmol), base (2.0 equiv). [g] Pd₂(dba)₃ instead of [Pd(Allyl)Cl]₂ [h] Pd₂(dba)₃ instead of Pd(OAc)₂

A typical procedure for entry 21:

To a solution of $\text{Pd}_2(\text{dba})_3$ (5.5 mg, 0.006 mmol, 5 mol%) in degassed dioxane (4 mL) in a Schlenk tube under nitrogen was added (*R*)-Segphos (8.1 mg, 0.0132 mmol, 11 mol%). The solution was stirred at room temperature (25 °C) for 30 min, and then sulfonium salt **1a** (69.4 mg, 0.132 mmol, 1.10 equiv), $^i\text{Pr}_2\text{NEt}$ (31.0 mg, 0.240 mmol, 2.00 equiv) and HPPPh_2 **2a** (22.3 mg, 0.120 mmol, 1.00 equiv) were added. The mixture was stirred at 25 °C for 13 h. After completion, the reaction mixture was passed through a short column of silica-gel with EtOAc as eluent. The solvent was removed on a rotary evaporator and the residue was subjected to silica gel chromatography (EtOAc/petroleum ether = 1/10) to give **3aa** as a white solid (63.9 mg, 95% yield, 93% ee).

2.3 Synthesis of the Atropoisomeric Phosphine Products

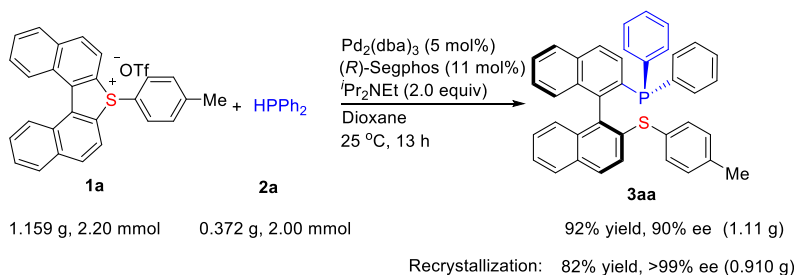


General Procedure C (GPC):

To a solution of $\text{Pd}_2(\text{dba})_3$ (5.5 mg, 0.006 mmol, 5 mol%) in degassed dioxane (4 mL) in a Schlenk tube under nitrogen was added (*R*)-Segphos (8.1 mg, 0.0132 mmol, 11 mol%). The solution was stirred at room temperature for 30 min, and then sulfonium salt **1** (0.132 mmol, 1.10 equiv), $^i\text{Pr}_2\text{NEt}$ (31.0 mg, 0.240 mmol, 2.00 equiv) and diarylphosphine **2** (0.120 mmol, 1.00 equiv) were added. The mixture was stirred at 25 °C for 13 h. After completion, the reaction mixture was passed through a short column of silica-gel with EtOAc as eluent. The solvent was removed on a rotary evaporator and the residue was subjected to silica gel chromatography (EtOAc/petroleum ether = 1/10) to give **3** as a white solid.

3. Scale-Up Synthesis of Enantiomerically Pure 3aa and 3ae

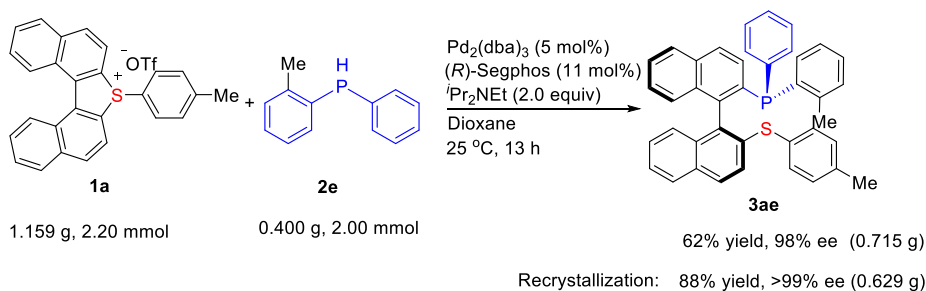
(1) Scaled-up synthesis of enantiomerically pure 3aa (2 mmol scale)



To a solution of $\text{Pd}_2(\text{dba})_3$ (91.9 mg, 0.100 mmol, 5 mol%) in degassed dioxane (70 mL) in a Schlenk tube under nitrogen was added $(R)\text{-Segphos}$ (135.3 mg, 0.220 mmol, 11 mol%). The solution was stirred at room temperature (25 °C) for 30 min, and then sulfonium **1a** (1.159 g, 2.20 mmol, 1.10 equiv), $i\text{Pr}_2\text{NEt}$ (0.157 g, 4.00 mmol, 2.00 equiv), HPPH₂ **2a** (372.0 mg, 2.00 mmol, 1.00 equiv) were added. The mixture was stirred at 25 °C for 13 h. After completion, the reaction mixture was passed through a short column of silica-gel with EtOAc as eluent. The solvent was removed on a rotary evaporator and the residue was subjected to silica gel chromatography (EtOAc/petroleum ether = 1/10) to give **3aa** as a white solid (1.11 g, 92% yield, 90% ee).

The above obtained **3aa** was recrystallized from CHCl_3 /acetone (via evaporation) to afford enantiomerically pure **3aa** (0.91 g, 82% yield, >99% ee) as colorless crystals.

(2) Scaled-up synthesis of enantiomerically pure **3ae** (2 mmol scale)



To a solution of $\text{Pd}_2(\text{dba})_3$ (91.9 mg, 0.100 mmol, 5 mol%) in degassed dioxane (70 mL) in a Schlenk tube under nitrogen was added $(R)\text{-Segphos}$ (135.3 mg, 0.220 mmol, 11 mol%). The solution was stirred at room temperature (25 °C) for 30 min, and then sulfonium **1a** (1.159 g, 2.20 mmol, 1.10 equiv), $i\text{Pr}_2\text{NEt}$ (0.157 g, 4.00 mmol, 2.00 equiv), **2e** (0.400 g, 2.00 mmol, 1.00 equiv) were added. The mixture was stirred at 25 °C for 13 h. After completion, the reaction mixture was passed through a short column of silica-gel with EtOAc as eluent. The solvent was removed on a rotary evaporator and the residue

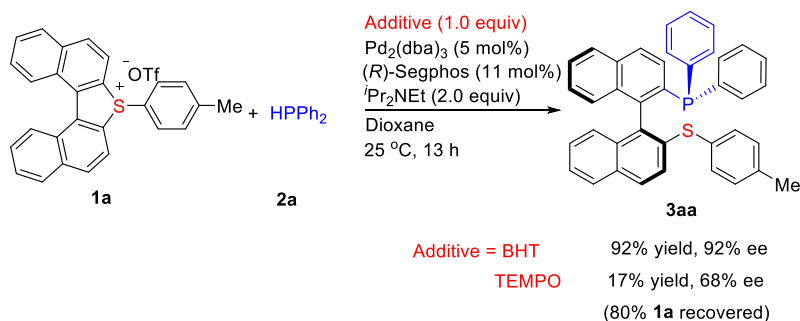
was subjected to silica gel chromatography (EtOAc/petroleum ether = 1/10) to give **3ae** as a white solid (0.715 g, 68% yield, 98% ee).

The above obtained **3ae** was recrystallized from CHCl₃/acetone (via evaporation) to afford enantiomerically pure **3ae** (0.629 g, 88% yield, >99% ee) as colorless crystals.

4. Mechanistic Considerations

4.1 Radical Trapping Experiments

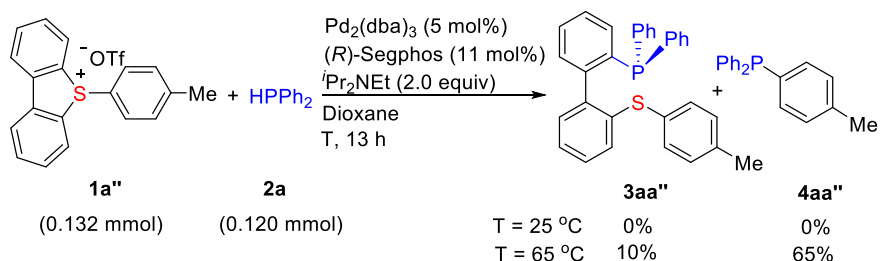
Radical trapping experiments using BHT (2,6-di-tert-butyl-4-methylphenol)/TEMPO (2,2,6,6-tetramethyl-1-piperidinyloxy) as a radical scavenger have been conducted as shown below and the results excluded the radical pathway.



To a solution of $\text{Pd}_2(\text{dba})_3$ (5.5 mg, 0.006 mmol, 5 mol%) in degassed dioxane (4 mL) in a Schlenk tube under nitrogen was added (*R*)-Segphos (8.1 mg, 0.0132 mmol, 11 mol%). The solution was stirred at room temperature (25 °C) for 30 min, and then sulfonium **1a** (69.4 mg, 0.132 mmol, 1.10 equiv), $i\text{Pr}_2\text{NEt}$ (31.0 mg, 0.240 mmol, 2.00 equiv), HPPH_2 **2a** (22.3 mg, 0.120 mmol, 1.00 equiv), BHT (2,6-di-tert-butyl-4-methylphenol, 26.4 mg, 0.120 mmol, 1.00 equiv) were added. The mixture was stirred at 25 °C for 13 h. After completion, the reaction mixture was passed through a short column of silica-gel with EtOAc as eluent. The solvent was removed on a rotary evaporator and the residue was subjected to silica gel chromatography (EtOAc/petroleum ether = 1/10) to give **3aa** as a white solid (61.9 mg, 92% yield, 92% ee).

When TEMPO (2,2,6,6-tetramethyl-1-piperidinyloxy) (18.7 mg, 0.120 mmol, 1.00 equiv)] was used instead of BHT as a radical scavenger under otherwise identical conditions, the yield and enantioselectivity of **3aa** were low (17% yield, 68% ee), **1a** (80%) being recovered.

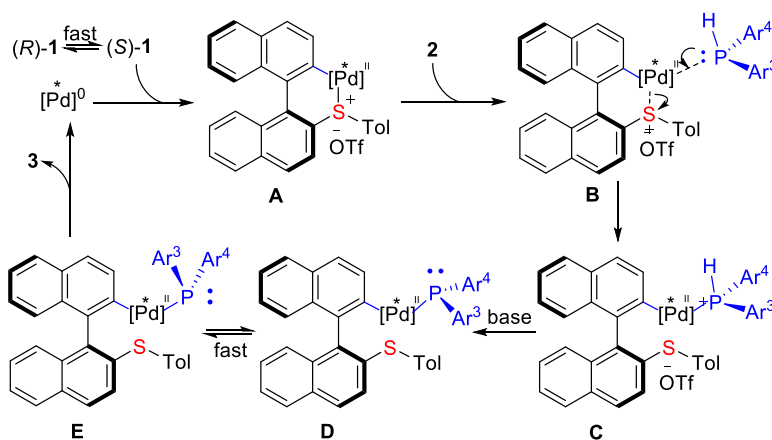
4.2 The reaction of 1a'' with 2a



To a solution of $\text{Pd}_2(\text{dba})_3$ (5.5 mg, 0.006 mmol, 5 mol%) in degassed dioxane (4 mL) in a Schlenk tube under nitrogen was added $(R)\text{-Segphos}$ (8.1 mg, 0.0132 mmol, 11 mol%). The solution was stirred at room temperature (25 °C) for 30 min, and then sulfonium salt **1a'** (51.2 mg, 0.132 mmol, 1.10 equiv), $i\text{Pr}_2\text{NEt}$ (31.0 mg, 0.240 mmol, 2.00 equiv) and HPPH_2 **2a** (22.3 mg, 0.120 mmol, 1.00 equiv) were added. The mixture was stirred at 25 °C for 13 h. No reaction was observed.

When the reaction was carried out at 65 °C instead of 25 °C under otherwise identical conditions, **4aa'** (21.6 mg, 65% yield) and **3aa'** (5.5 mg, 10% yield) were isolated (EtOAc/petroleum ether = 1/10).

4.3 Proposed Catalytic Cycle



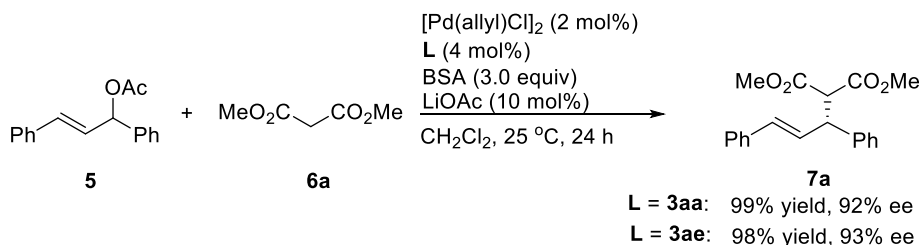
Scheme S3. Proposed catalytic cycle

A plausible catalytic cycle was proposed as depicted in Scheme S3. $(R_a)\text{-1}$ and $(S_a)\text{-1}$ are in rapid equilibrium^{3,9} with each other via the rotation of the $\text{C}_{\text{aryl}}\text{-C}_{\text{aryl}}$ single bond.

The (*R*)-Segphos-ligated Pd⁽⁰⁾ complex undergoes oxidative addition preferentially to (S)-**1** cleaving an endo C-S bond to form biaryl [Pd]^{II} species **A**. The following coordination of **2** (**B**) leads to the formation of complex **C** followed by the abstraction of a proton to give the [Pd]^{II}-phosphido species^{10,11} **D** and **E** which undergo a rapid equilibrium (when Ar³ ≠ Ar⁴). Subsequent reductive elimination of the more thermodynamic stable **D** furnishes the chiral phosphine product **3** and regenerates the [Pd]⁰ species, closing a catalytic cycle. The key point is that the equilibrium between **D** and **E** must be much faster than the reductive elimination in this dynamic kinetic resolution process in order to achieve high enantio-control of P-stereogenic centre. Notably, the exist of P,S-ligands which have high affinity to metals does not interrupt the catalytic cycle, presumably due to the stronger chelating ability of Segphos coordinated with palladium.

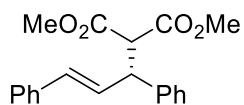
5. Application of 3aa/3ae as a Chiral Ligand

(1) Pd-catalyzed asymmetric allylic alkylation reaction¹²



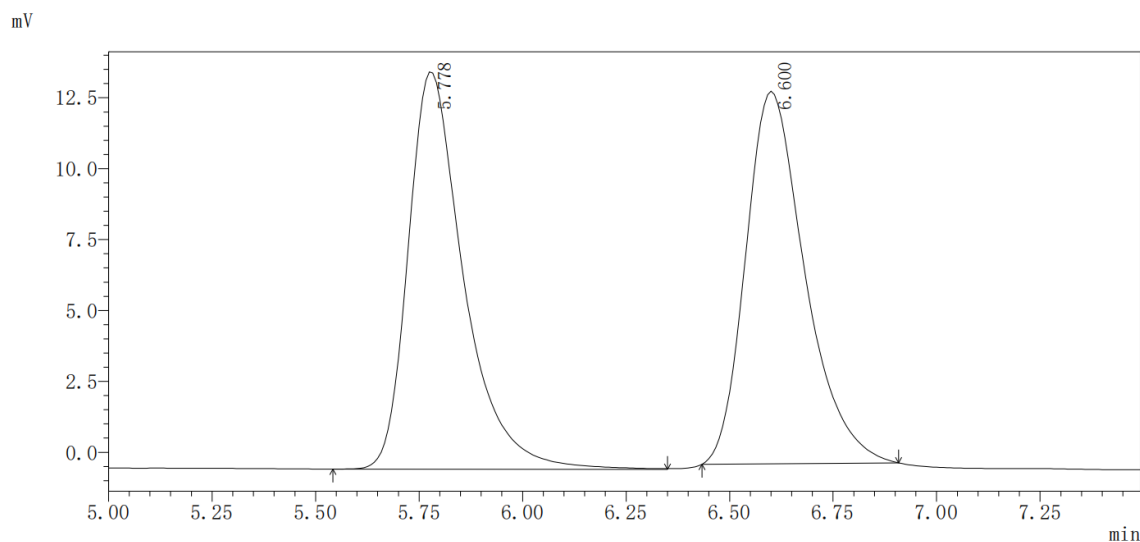
[PdCl(C₃H₅)₂] (1.4 mg, 2 mol%), LiOAc (1.3 mg, 10 mol%), **3aa** (4.5 mg, 4 mol%) and CH₂Cl₂ (1 mL) were placed in a Schlenk tube under argon. The resulting mixture was stirred at room temperature (25 °C) for 30 min. Racemic 1,3-diphenyl-2-propenyl acetate **5** (50.4 mg, 0.200 mmol) in CH₂Cl₂ (1 mL), dimethyl malonate **6a** (79.8 mg, 0.600 mmol) in CH₂Cl₂ (1 mL) and N,O-bis(trimethylsilyl)-acetamide (BSA) (122.0 mg, 0.600 mmol) in CH₂Cl₂ (1 mL) were added subsequently, and the reaction mixture was stirred at room temperature (25 °C) for 24 h. After completion, the reaction mixture was quenched with saturated NH₄Cl aqueous solution (10.0 mL) and extracted with Et₂O (3 x 10 mL). The combined organic layer was washed with brine (10 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash chromatography on silica gel (hexanes/EtOAc = 20/1) to afford product **7a** (64.2 mg, 99% yield, 92% ee) as a colorless oil.

When **3ae** (4.7 mg, 4 mol%) was used instead of **3aa** under otherwise identical conditions, product **7a** was obtained (63.6 mg, 98% yield, 93% ee) as a colorless oil.



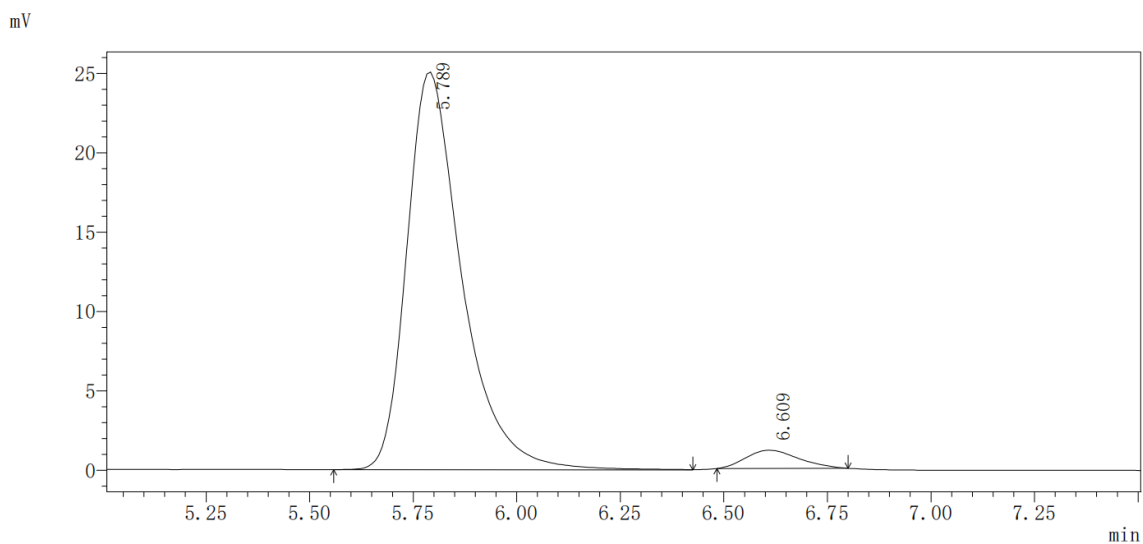
Dimethyl (R,E)-2-(1,3-diphenylallyl)malonate (7a). **7a** was prepared according to above method (64.2 mg, 99% yield, 93% ee) and purified by flash column chromatography (hexanes/EtOAc = 20/1) as a colorless oil. $[\alpha]_D^{23} +17.2$ (*c* 0.61, CHCl₃) for 93% ee. ¹H NMR (500 MHz, CDCl₃) δ 7.25-7.10 (m, 10H), 6.40 (d, *J* = 15.8 Hz, 1H), 6.26 (dd, *J* = 15.7, 8.6 Hz, 1H), 4.19 (dd, *J* = 10.7, 8.8 Hz, 1H), 3.88 (d, *J* = 10.9 Hz, 1H), 3.62 (s, 3H), 3.43 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 168.2, 167.8, 140.2, 136.8, 131.9, 129.1, 128.7, 128.5, 127.9, 127.6, 127.2, 126.4, 57.7, 52.7, 52.6, 52.5, 49.2, 41.1. The ¹H NMR matched the literature reported data.⁶ The ee was determined on a Chiralcel IA column with 15% i-PrOH/hexane, flow = 1.0 mL/min, λ = 254 nm, λ = 254 nm, t (major) = 5.8 min, t (minor) = 6.7 min.

HPLC charts (with 3aa as the ligand):



Detector A channel 254 nm

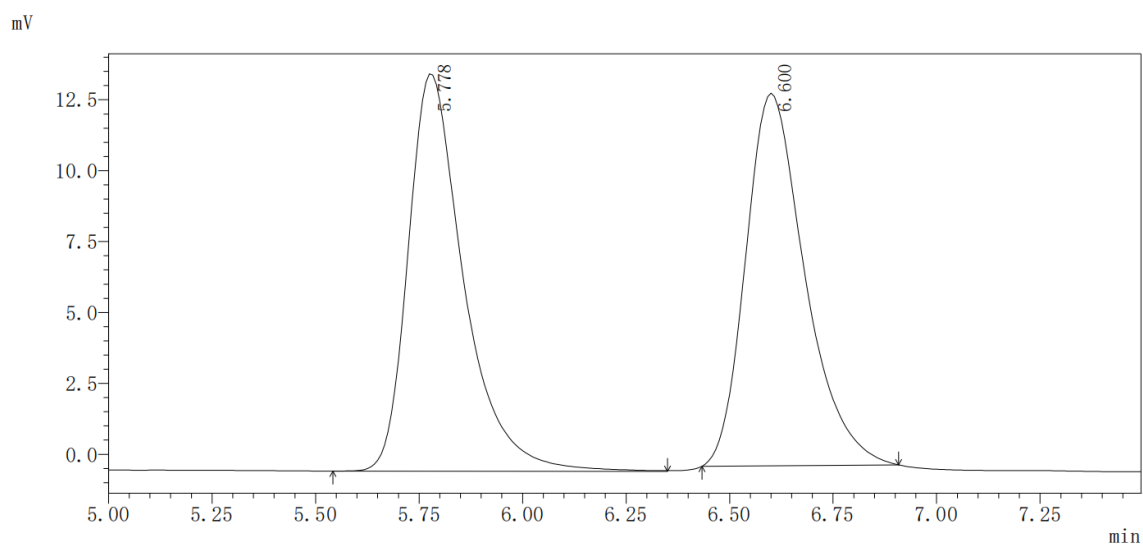
Peak	Ret. Time	Area	Height	Conc.
1	5.778	130985	14008	50.033
2	6.600	130811	13136	49.967
Total		261796	27145	



Detector A channel 254 nm

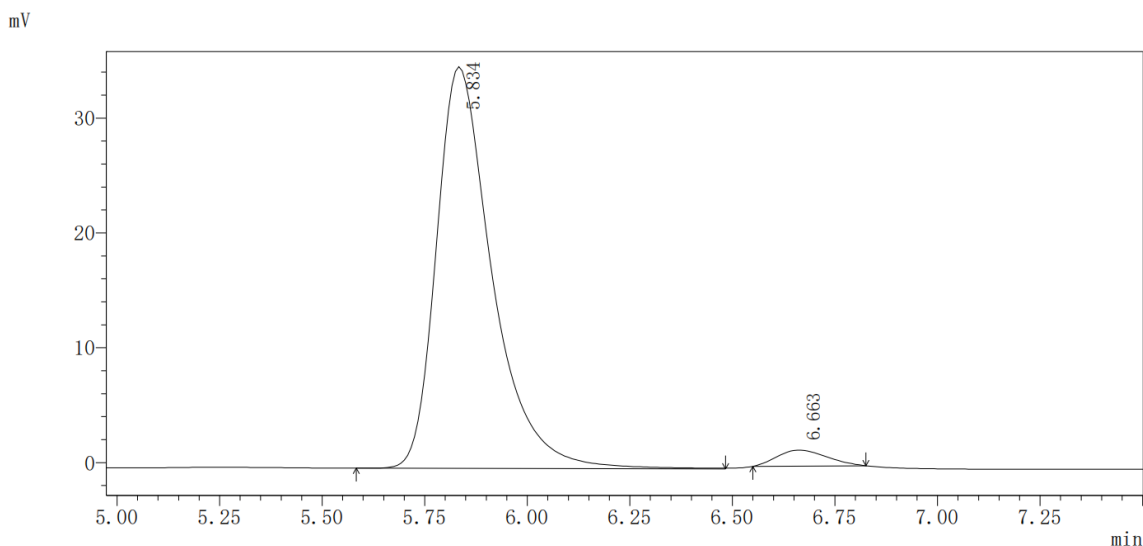
Peak	Ret. Time	Area	Height	Conc.
1	5.789	232267	25061	95.810
2	6.609	10158	1150	4.190
Total		242424	26211	

HPLC charts (with 3ae as the ligand):



Detector A channel 254 nm

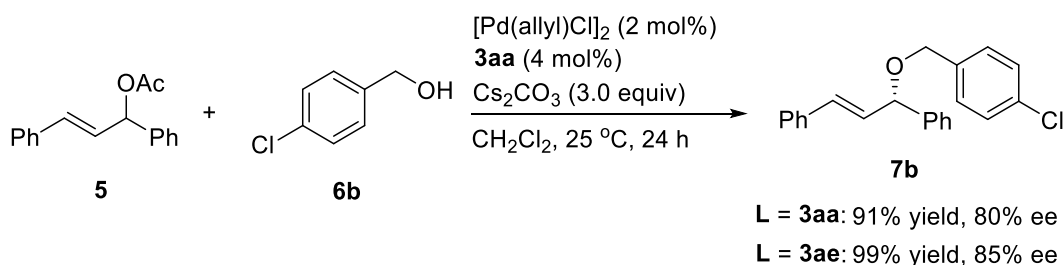
Peak	Ret. Time	Area	Height	Conc.
1	5.778	130985	14008	50.033
2	6.600	130811	13136	49.967
Total		261796	27145	



Detector A channel 254 nm

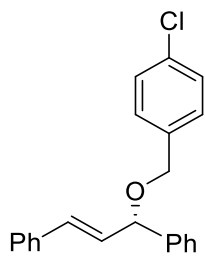
Peak	Ret. Time	Area	Height	Conc.
1	5.834	327704	34961	96.414
2	6.663	12190	1420	3.586
Total		339894	36380	

(2) Pd-catalyzed asymmetric allylic etherification reaction¹³



[PdCl(C₃H₅)]₂ (1.4 mg, 2 mol%), **3aa** (4.5 mg, 4 mol%) and CH₂Cl₂ (1 mL) were placed in a Schlenk tube under argon. The resulting mixture was stirred at room temperature (25 °C) for 30 min. Racemic 1,3-diphenyl-2-propenyl acetate **5** (50.4 mg, 0.200 mmol) was added and the mixture was stirred for another 10 min. (4-Chlorophenyl) methanol **6b** (85.8 mg, 0.600 mmol) and Cs₂CO₃ (195.0 mg, 0.600 mmol) were added. After stirring at room temperature for 24 h, the solvent was evaporated in vacuo and the residue was purified by flash column chromatography on silica gel to give product **7b** as a colorless oil (60.8 mg, 91% yield, 80% ee).

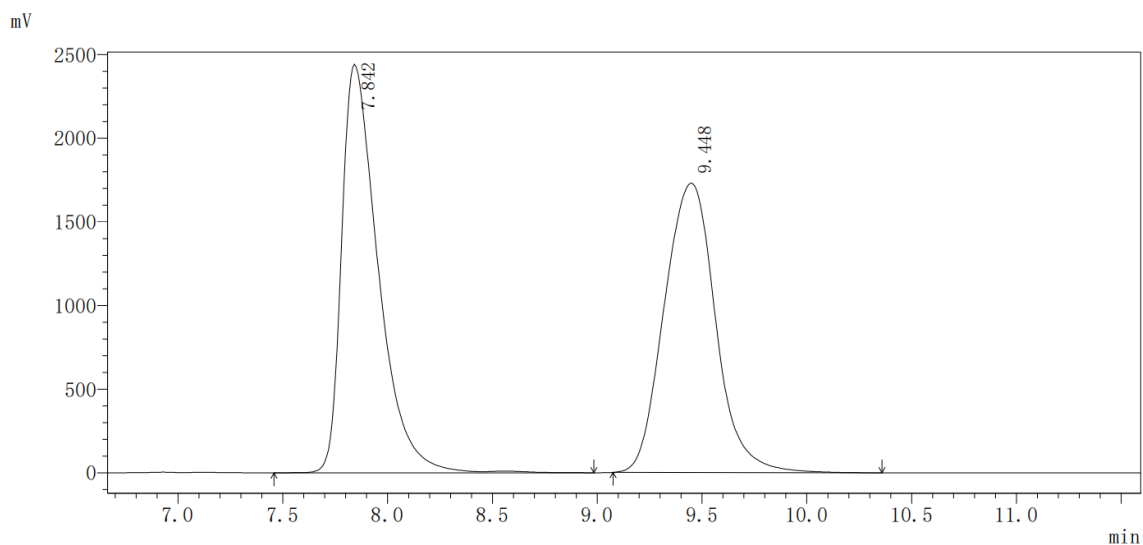
When **3ae** (4.7 mg, 4 mol%) was used instead of **3aa** under otherwise identical conditions, product **7b** was obtained (66.3 mg, 99% yield, 85% ee) as a colorless oil.



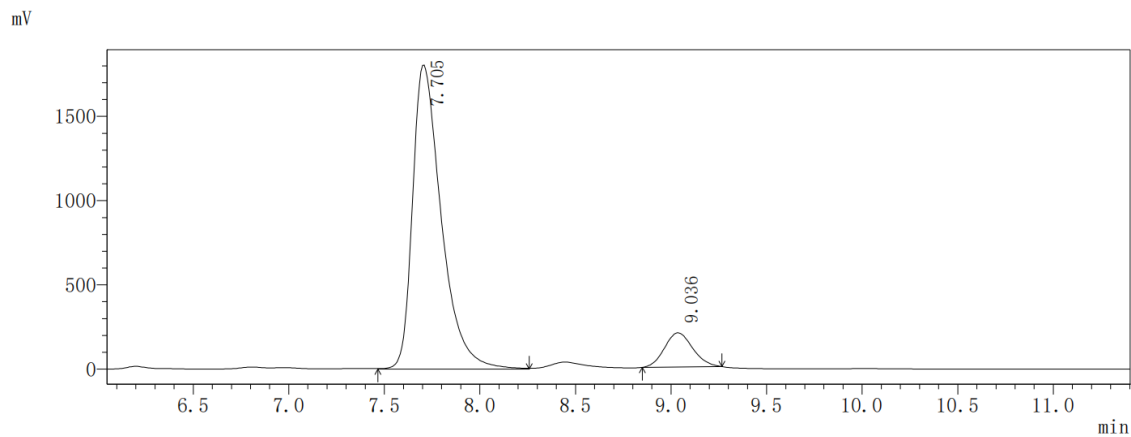
(S,E)-3-((4-chlorobenzyl)oxy)prop-1-ene-1,3-diyl dibenzene (7b).

7b was prepared according to above method and purified by flash column chromatography (hexanes/EtOAc = 10/1) as a colorless oil (66.3 mg, 99% yield, 85% ee). $[\alpha]_D^{23} +20.0$ (c 0.5, CHCl₃) for 85% ee. **¹H NMR** (500 MHz, CDCl₃) δ 7.32-7.08 (m, 14H), 6.52 (d, J = 15.9 Hz, 1H), 6.22 (dd, J = 15.9, 7.1 Hz, 1H), 4.88 (d, J = 7.1 Hz, 1H), 4.24 (dd, J = 12.2, 18.6 Hz, 2H). **¹³C NMR** (126 MHz, CDCl₃) δ 139.9, 135.9, 135.4, 132.2, 130.7, 129.0, 128.0, 127.6, 127.5, 127.4, 126.8, 125.9, 125.6, 80.8, 68.3. The ee was determined on a Chiralcel IB column with 0.1% i-PrOH/hexane, flow = 0.8 mL/min, λ = 254 nm, t (major) = 8.3 min, t (minor) = 9.6 min.

HPLC charts (with 3aa as the ligand):



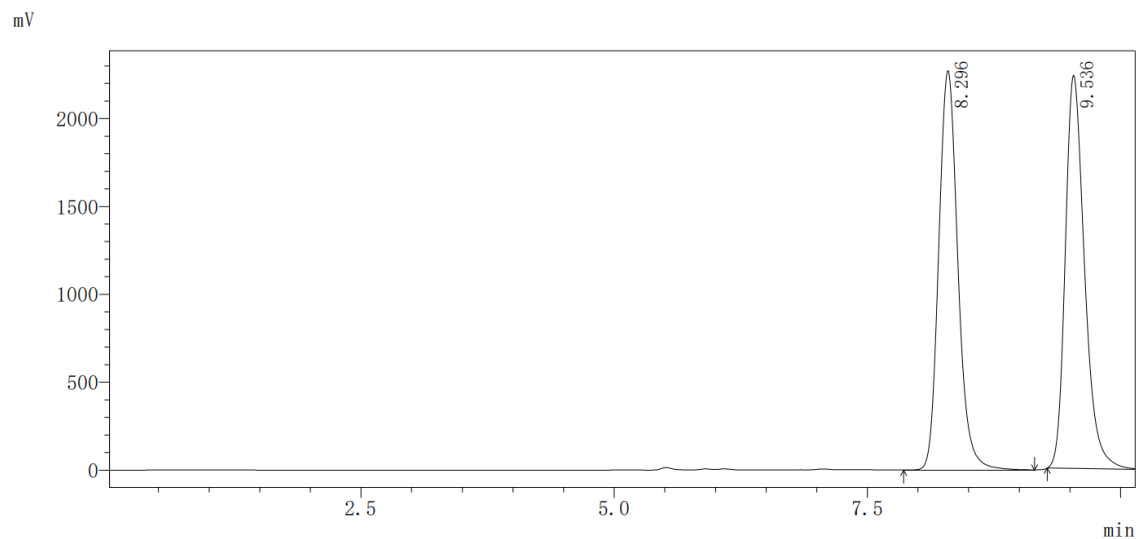
Peak	Ret. Time	Area	Height	Conc.
1	7.842	29789108	2442504	50.267
2	9.448	29472236	1729292	49.733
Total		59261343	4171796	



Detector A channel 254 nm

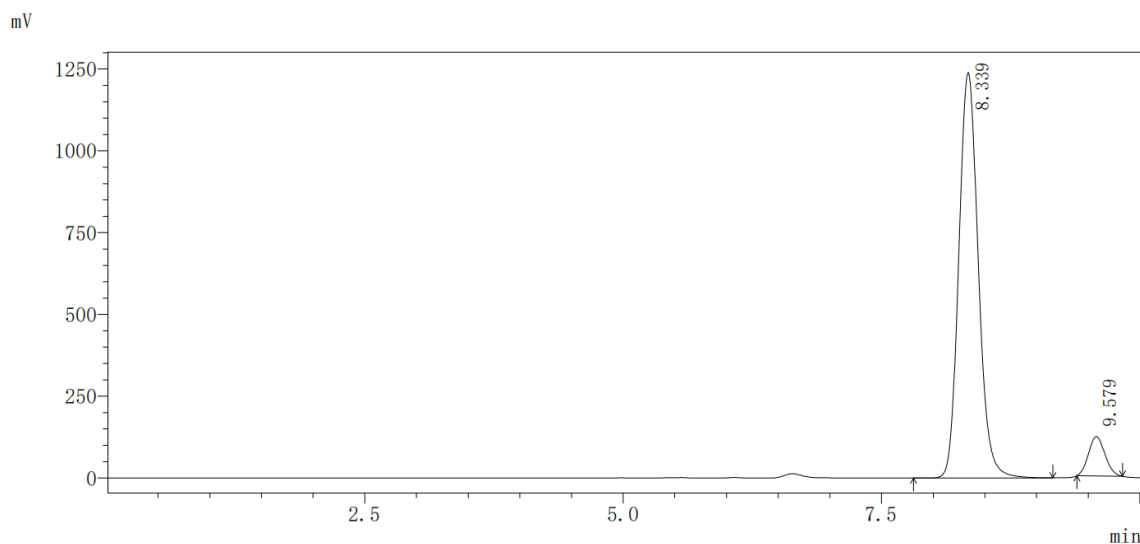
Peak	Ret. Time	Area	Height	Conc.
1	7.705	18909207	1804991	90.397
2	9.036	2008701	199244	9.603
Total		20917908	2004235	

HPLC charts (with 3ae as the ligand):



Detector A channel 254 nm

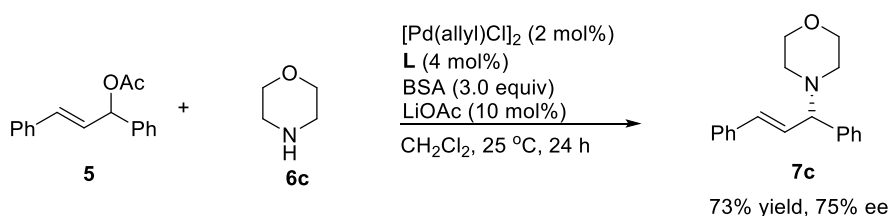
Peak	Ret. Time	Area	Height	Conc.
1	8.296	29998950	2271585	50.326
2	9.536	29610120	2237289	49.674
Total		59609070	4508874	



Detector A channel 254 nm

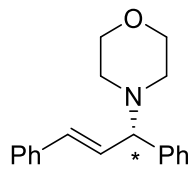
Peak	Ret. Time	Area	Height	Conc.
1	8.339	16263381	1239419	92.401
2	9.579	1337577	120581	7.599
Total		17600958	1360000	

(3) Pd-catalyzed asymmetric allylic amination reaction^{14,15}



[PdCl(C₃H₅)]₂ (1.4 mg, 2 mol%), LiOAc (1.3 mg, 10 mol%), **3aa** (4.5 mg, 4 mol%) and CH₂Cl₂ (1 mL) were placed in a Schlenk tube under argon. The resulting mixture was stirred at room temperature (25 °C) for 30 min. Racemic 1,3-diphenyl-2-propenyl acetate **5** (50.4 mg, 0.200 mmol) in CH₂Cl₂ (0.4 mL), morpholine **6c** (52.3 mg, 0.600 mmol) in CH₂Cl₂ (1 mL), and N,O-bis(trimethylsilyl)-acetamide (BSA) (122.0 mg, 0.600 mmol) in CH₂Cl₂ (1 mL) were added subsequently. The reaction mixture was stirred at room temperature (25 °C) for 24 h. After completion, the reaction mixture was quenched with saturated NH₄Cl aqueous solution (10.0 mL) and extracted with Et₂O (3 x 10 mL). The combined organic layer was washed with brine (10 mL), dried over Na₂SO₄, filtered, and

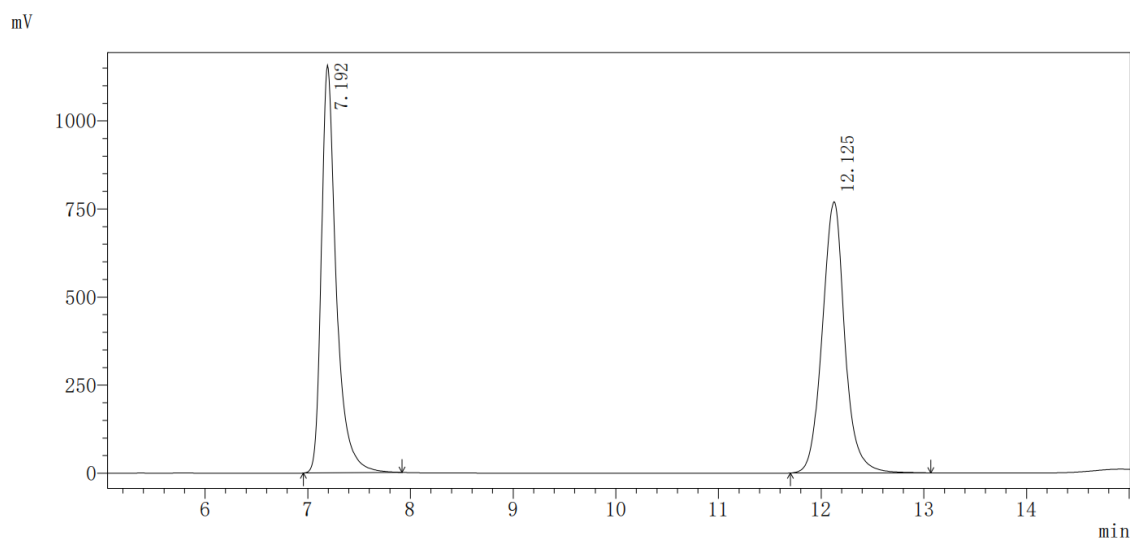
concentrated. The residue was purified by flash chromatography on silica gel (hexanes/EtOAc = 10/1) to afford product **7c** (40.8 mg, 73% yield, 75% ee) as a colorless oil.



(*S,E*)-4-(1,3-diphenylallyl)morpholine (7c). **7c** was prepared according

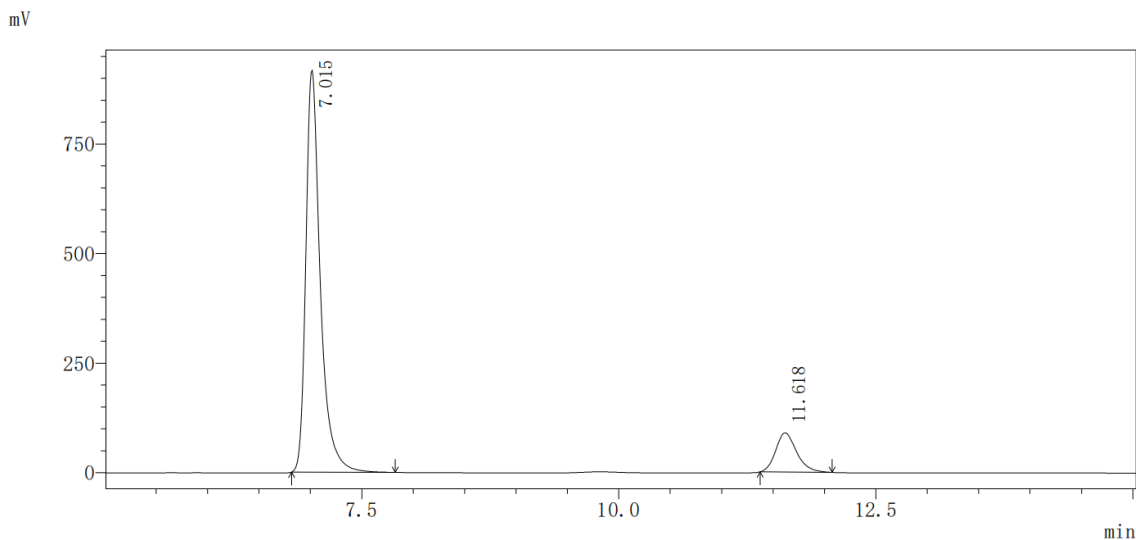
to above method and purified by flash column chromatography (hexanes/EtOAc = 10/1) as a colorless oil (40.8 mg, 73% yield, 75% ee). $[\alpha]_D^{23} + 13.7$ (*c* 0.73, CHCl₃) for 75% ee.

¹H NMR (500 MHz, CDCl₃) δ 7.33-7.11 (m, 10H), 6.49 (d, *J* = 15.8 Hz, 1H), 6.20 (dd, *J* = 15.8, 8.9 Hz, 1H), 3.71 (d, *J* = 8.9 Hz, 1H), 3.63 (t, *J* = 4.6 Hz, 4H), 2.42-2.52 (m, 2H), 2.34-2.29 (m, 2H). **¹³C NMR** (126 MHz, CDCl₃) δ 141.6, 136.8, 131.6, 131.4, 128.7, 128.6, 128.1, 127.6, 127.4, 126.4, 74.9, 67.2, 52.4. The ¹H NMR matched the literature reported data.^[8] The ee was determined on a Chiralcel IB column with 1% i-PrOH/hexane, flow = 1.0 mL/min, λ = 254 nm, *t* (major) = 7.02 min, *t* (minor) = 11.62 min.



Detector A channel 254 nm

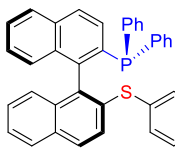
Peak	Ret. Time	Area	Height	Conc.
1	7.192	11663217	1156499	49.871
2	12.125	11723581	769899	50.129
Total		23386797	1926398	



Detector A channel 254 nm

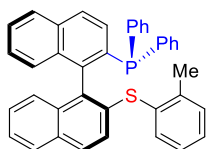
Peak	Ret. Time	Area	Height	Conc.
1	7.015	8911796	916980	87.574
2	11.618	1264537	89360	12.426
Total		10176334	1006340	

6. Characterization of the Products



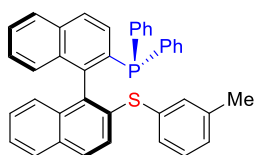
(S)-Diphenyl(2'-(*p*-tolylthio)-[1,1'-binaphthalen]-2-yl) phosphane (3aa). Following General Procedure C (GPC), **3aa** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (63.9 mg, 95% yield, 93% ee). $[\alpha]_D^{23}$ -134.0 (*c* 1.0, CHCl₃) for 93% ee. ^1H NMR (500 MHz, CDCl₃) δ 7.96-7.94 (m, 2H), 7.82 (d, *J* = 8.5 Hz, 2H), 7.55-7.52 (m, 2H), 7.38-7.28 (m, 9H), 7.24-7.21 (m, 3H), 7.19-7.12 (m, 4H), 7.07 (d, *J* = 7.8 Hz, 2H), 7.00-6.97 (m, 1H), 6.81 (d, *J* = 8.4 Hz, 1H), 2.35 (s, 3H). ^{13}C NMR (126 MHz, CDCl₃) δ 144.0, 143.7, 138.1 (d, *J* = 12.6 Hz), 137.8, 137.2 (d, *J* = 13.6 Hz), 136.7 (d, *J* = 2.9 Hz), 135.9 (d, *J* = 10.8 Hz), 134.9 (d, *J* = 8.6 Hz), 134.1 (d, *J* = 21.0 Hz), 133.8, 133.7 (d, *J* = 2.6 Hz), 133.6, 133.1 (d, *J* = 18.2 Hz), 132.7 (d, *J* = 7.7 Hz), 131.4, 130.8, 130.7 (d, *J* = 2.0 Hz), 129.9, 128.6, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.7, 126.8 (d, *J* = 29.2 Hz), 126.5 (d, *J* = 2.5 Hz), 126.3, 126.2, 126.1, 125.2, 21.2. ^{31}P NMR (202 MHz, CDCl₃) δ -14.5 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol =

99.5/0.5, 0.5 mL/min, λ = 254 nm, t (major) = 10.4 min, t (minor) = 11.0 min. **HRMS** (ESI-TOF) m/z: $[M+H]^+$ Calcd for $C_{39}H_{29}PSH^+$: 561.1800; Found: 561.1803.



(S)-Diphenyl(2'-(*o*-tolylthio)-[1,1'-binaphthalen]-2-yl)phosphane

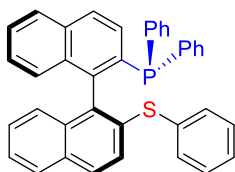
(3ba). Following General Procedure C (GPC), **3ba** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (57.8 mg, 86% yield, 96% ee). $[\alpha]_D^{23}$ -84.4 (*c* 0.64, $CHCl_3$) for 96% ee. **1H NMR** (500 MHz, $CDCl_3$) δ 7.85-7.82 (m, 2H), 7.69 (t, J = 8.9 Hz, 2H), 7.44-7.39 (m, 2H), 7.28-7.24 (m, 3H), 7.22-7.17 (m, 6H), 7.12-7.10 (m, 3H), 7.10-7.03 (m, 2H), 7.02-6.98 (m, 3H), 6.95 (d, J = 8.8 Hz, 1H), 6.87-6.83 (m, 1H), 6.65 (d, J = 8.5 Hz, 1H), 2.10 (s, 3H). **^{13}C NMR** (126 MHz, $CDCl_3$) δ 143.7 (d, J = 34.9 Hz), 141.8, 138.3 (d, J = 12.8 Hz), 137.2 (d, J = 13.8 Hz), 136.4 (d, J = 11.3 Hz), 136.1 (d, J = 2.9 Hz), 135.3, 134.3 (d, J = 8.6 Hz), 134.1, 134.0, 133.9, 133.8 (d, J = 2.8 Hz), 133.1, 133.0, 132.6, 132.5 (d, J = 7.6 Hz), 131.2, 130.8 (d, J = 2.3 Hz), 130.5, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.8, 127.0, 126.7, 126.6, 126.5, 126.2, 126.0, 125.0, 124.9, 20.8. **^{31}P NMR** (202 MHz, $CDCl_3$) δ -14.4 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IB, n-hexane/isopropanol = 99.9/0.1, 0.5 mL/min, λ = 254 nm, t (major) = 11.0 min, t (minor) = 12.3 min. **HRMS** (ESI-TOF) m/z: $[M+H]^+$ Calcd for $C_{39}H_{29}PSH^+$: 561.1800; Found: 561.1795.



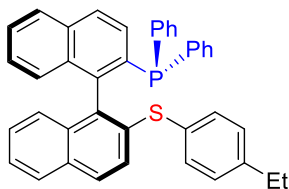
(S)-Diphenyl(2'-(*m*-tolylthio)-[1,1'-binaphthalen]-2-yl)phosphane

(3ca). Following General Procedure C (GPC), **3ca** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (65.4 mg, 97% yield, 85% ee). $[\alpha]_D^{23}$ -161.5 (*c* 0.52, $CHCl_3$) for 85% ee. **1H NMR** (500 MHz, $CDCl_3$) δ 7.83-7.81 (m, 2H), 7.72 (d, J = 8.6 Hz, 2H), 7.42-7.38 (m, 2H), 7.24-7.17 (m, 6H), 7.15-7.10 (m, 4H), 7.07-6.98 (m, 6H), 6.95-6.88 (m, 3H), 6.72 (d, J = 8.1 Hz, 1H), 2.11 (s, 3H). **^{13}C NMR** (126 MHz, $CDCl_3$) δ 143.9 (d, J = 34.9 Hz), 138.8, 137.9 (d, J = 12.4 Hz), 137.4 (d, J = 13.7 Hz), 136.2 (d, J = 2.4 Hz), 135.8 (d, J = 6.9 Hz), 135.5 (d, J = 26.1 Hz), 134.4, 134.0, 133.9, 133.8, 133.7, 133.6, 133.5, 133.1, 133.0, 132.8 (d, J =

7.7 Hz), 131.6, 130.8, 130.7, 130.1, 128.8, 128.7, 128.4, 128.2, 128.1, 128.0, 127.9, 127.8, 127.0, 126.8, 126.7, 126.5 (d, $J = 2.6$ Hz), 126.3, 126.2, 125.3, 21.2. ^{31}P NMR (202 MHz, CDCl_3) δ -14.7 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, $\lambda = 230$ nm, t (major) = 16.4 min, t (minor) = 17.3 min. HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{39}\text{H}_{29}\text{PSH}^+$: 561.1800; Found: 561.1798.

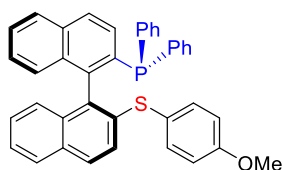


(S)-Diphenyl(2'-(phenylthio)-[1,1'-binaphthalen]-2-yl)phosphane (3da). Following General Procedure C (GPC), **3da** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (57.2 mg, 87% yield, 92% ee). $[\alpha]_{\text{D}}^{23}$ -151.8 (c 0.54, CHCl_3) for 92% ee. ^1H NMR (500 MHz, CDCl_3) δ 7.84-7.82 (m, 2H), 7.73-7.71 (m, 2H), 7.43-7.39 (m, 2H), 7.23-7.17 (m, 7H), 7.16-7.10 (m, 8H), 7.07-7.00 (m, 4H), 6.88 (t, $J = 7.7$ Hz, 1H), 6.70 (d, $J = 8.5$ Hz, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 143.8 (d, $J = 34.7$ Hz), 138.0 (d, $J = 12.7$ Hz), 137.2 (d, $J = 13.6$ Hz), 135.9 (d, $J = 2.9$ Hz), 135.8 (d, $J = 2.0$ Hz), 135.8, 134.9, 134.1, 133.9, 133.8, 33.7 (d, $J = 2.9$ Hz), 133.1, 133.0, 132.9, 132.8, 132.7 (d, $J = 7.7$ Hz), 131.6, 130.7 (d, $J = 2.2$ Hz), 129.0, 128.7, 128.4, 128.2 (d, $J = 12.8$ Hz), 128.1, 127.8 (d, $J = 1.8$ Hz), 127.4, 127.0, 126.9, 126.7, 126.5 (d, $J = 2.6$ Hz), 126.3, 126.2, 125.4. ^{31}P NMR (202 MHz, CDCl_3) δ -14.6 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, $\lambda = 230$ nm, t (major) = 17.2 min, t (minor) = 18.0 min. HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{38}\text{H}_{27}\text{PSH}^+$: 547.1644; Found: 547.1640.

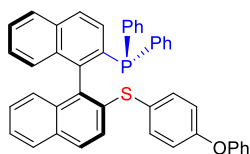


(S)-(2'-((4-Ethylphenyl)thio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3ea). Following General Procedure C (GPC), **3ea** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (58.7 mg, 85% yield, 92% ee). $[\alpha]_{\text{D}}^{23}$ -155.0 (c 1.0, CHCl_3) for 92% ee. ^1H NMR (500 MHz, CDCl_3) δ 7.81-7.78 (m, 2H), 7.68-7.66 (m, 2H), 7.40-7.36 (m, 2H), 7.24-7.12

(m, 10H), 7.09-7.07 (m, 2H), 7.04-6.98 (m, 4H), 6.94 (d, $J = 8.5$ Hz, 2H), 6.85-6.82 (m, 1H), 6.67 (d, $J = 8.4$ Hz, 1H), 2.49 (q, $J = 7.6$ Hz, 2H), 1.10 (t, $J = 7.6$ Hz, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 144.1, 143.9 (d, $J = 34.6$ Hz), 138.2 (d, $J = 12.8$ Hz), 137.2 (d, $J = 13.7$ Hz), 136.7 (d, $J = 2.9$ Hz), 135.9 (d, $J = 10.8$ Hz), 135.1 (d, $J = 8.6$ Hz), 134.2, 134.0, 133.8, 133.6, 133.2, 133.0, 132.7 (d, $J = 7.6$ Hz), 131.5, 131.2, 130.8, 128.7, 128.6, 128.4, 128.3, 128.2, 128.1, 128.0, 127.8, 127.0, 126.7, 126.5 (d, $J = 2.6$ Hz), 126.4, 126.3, 125.3, 28.6, 15.4. ^{31}P NMR (202 MHz, CDCl_3) δ -14.5 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, $\lambda = 230$ nm, t (major) = 9.5 min, t (minor) = 10.0 min. HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{40}\text{H}_{31}\text{PSH}^+$: 575.1957; Found: 575.1956.

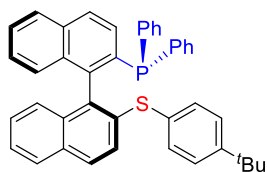


(S)-(2'-((4-Methoxyphenyl)thio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3fa). Following General Procedure C (GPC), **3fa** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/5) as a white solid (35.5 mg, 51% yield, 94% ee). $[\alpha]_{\text{D}}^{23}$ -84.4 (c 0.64, CHCl_3) for 94% ee. ^1H NMR (500 MHz, CDCl_3) δ 7.84-7.82 (m, 2H), 7.70-7.67 (m, 2H), 7.43-7.39 (m, 2H), 7.27-6.98 (m, 16H), 6.86 (t, $J = 7.7$ Hz, 1H), 6.71-6.68 (m, 2H), 6.67 (d, $J = 8.5$ Hz, 1H), 3.70 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 159.8, 143.8 (d, $J = 35.3$ Hz), 138.1 (d, $J = 12.9$ Hz), 137.5 (d, $J = 3.0$ Hz), 137.2 (d, $J = 13.6$ Hz), 136.0, 135.9 (d, $J = 10.8$ Hz), 134.1, 134.0, 133.9, 133.8, 133.6 (d, $J = 2.7$ Hz), 133.2, 133.0, 132.7 (d, $J = 7.7$ Hz), 131.3, 130.7 (d, $J = 2.2$ Hz), 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.7, 126.9, 126.7, 126.5 (d, $J = 2.6$ Hz), 126.2, 126.0, 125.5, 125.0, 124.5, 114.8, 55.3. ^{31}P NMR (202 MHz, CDCl_3) δ -14.4 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, $\lambda = 230$ nm, t (major) = 22.2 min, t (minor) = 25.0 min. HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{39}\text{H}_{29}\text{POSH}^+$: 577.1749; Found: 577.1747.



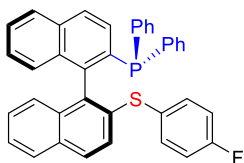
(S)-(2'-((4-Phenoxyphenyl)thio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3ga). Following General Procedure C (GPC), **3ga** was prepared

and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (74.2 mg, 97% yield, 93% ee). $[\alpha]_D^{23}$ -120.0 (*c* 1.0, CHCl₃) for 93% ee. **¹H NMR** (500 MHz, CDCl₃) δ 7.83-7.80 (m, 2H), 7.73-7.69 (m, 2H), 7.41-7.38 (m, 2H), 7.27-7.14 (m, 12H), 7.13-7.08 (m, 2H), 7.05-6.98 (m, 5H), 6.92-6.85 (m, 3H), 6.75-6.72 (m, 2H), 6.68 (d, *J* = 8.4 Hz, 1H). **¹³C NMR** (126 MHz, CDCl₃) δ 157.5, 156.6, 143.8 (d, *J* = 34.6 Hz), 138.0 (d, *J* = 12.7 Hz), 137.2 (d, *J* = 13.6 Hz), 136.5 (d, *J* = 3.0 Hz), 135.9 (d, *J* = 10.8 Hz), 135.3, 135.1 (d, *J* = 8.6 Hz), 134.1, 133.9, 133.8, 133.7 (d, *J* = 2.6 Hz), 133.2, 133.0, 132.8 (d, *J* = 7.7 Hz), 131.5, 130.8, 129.9, 128.7, 128.5, 128.3, 128.2, 128.1, 128.0, 127.9, 127.0, 126.8, 126.5 (d, *J* = 2.6 Hz), 126.3, 126.2, 125.3, 123.8, 119.4, 119.1. **³¹P NMR** (202 MHz, CDCl₃) δ -14.5 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 99.9/0.1, 0.5 mL/min, λ = 254 nm, *t* (minor) = 9.1 min, *t* (major) = 11.2 min. **HRMS** (ESI-TOF) *m/z*: [M+H]⁺ Calcd for C₄₄H₃₁POSH⁺: 639.1906; Found: 639.1906.

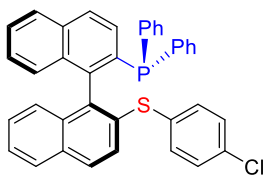


(S)-(2'-((4-(*Tert*-butyl)phenyl)thio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3ha).

Following General Procedure C (GPC), **3ha** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (55.1 mg, 76% yield, 94% ee). $[\alpha]_D^{23}$ -107.0 (*c* 1.0, CHCl₃) for 94% ee. **¹H NMR** (500 MHz, CDCl₃) δ 7.81-7.79 (m, 2H), 7.70-7.67 (m, 2H), 7.40-7.37 (m, 2H), 7.23-6.99 (m, 14H), 7.05-6.98 (m, 4H), 6.84 (t, *J* = 7.6 Hz, 1H), 6.67 (d, *J* = 6.1 Hz, 1H), 1.19 (s, 9H). **¹³C NMR** (126 MHz, CDCl₃) δ 150.8, 143.9 (d, *J* = 34.7 Hz), 138.1 (d, *J* = 12.8 Hz), 137.2 (d, *J* = 13.7 Hz), 136.4 (d, *J* = 3.2 Hz), 153.9 (d, *J* = 10.8 Hz), 135.4 (d, *J* = 8.6 Hz), 134.2, 134.0, 133.8, 133.7 (d, *J* = 2.7 Hz), 133.2, 133.0, 132.9, 132.8 (d, *J* = 7.6 Hz), 131.5, 131.1, 130.7, 130.6, 128.6, 128.5, 128.4, 128.2, 128.1, 128.0, 127.8, 126.9, 126.7, 126.5, 126.4, 126.3, 126.2, 126.1, 125.3, 34.6, 31.3. **³¹P NMR** (202 MHz, CDCl₃) δ -14.5 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IB, n-hexane/isopropanol = 99.9/0.1, 0.5 mL/min, λ = 254 nm, *t* (minor) = 11.5 min, *t* (major) = 12.5 min. **HRMS** (ESI-TOF) *m/z*: [M+H]⁺ Calcd for C₄₂H₃₅PSH⁺: 603.2270; Found: 603.2273.

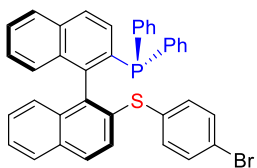


(S)-(2'-((4-Fluorophenyl)thio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3ia). Following General Procedure C (GPC), **3ia** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (61.8 mg, 91% yield, 90% ee). $[\alpha]_D^{23}$ -148.5 (*c* 1.0, CHCl₃) for 90% ee. ¹H NMR (500 MHz, CDCl₃) δ 7.82-7.80 (m, 2H), 7.71-7.69 (m, 2H), 7.40-7.37 (m, 2H), 7.22-7.08 (m, 12H), 7.05-6.97 (m, 4H), 6.90-6.86 (m, 1H), 6.80-6.75 (m, 2H), 6.70 (d, *J* = 2.1 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 162.5 (d, *J* = 248.4 Hz), 143.7 (d, *J* = 34.5 Hz), 137.9 (d, *J* = 12.6 Hz), 137.2 (d, *J* = 13.4 Hz), 136.0 (d, *J* = 2.5 Hz), 135.8 (d, *J* = 10.7 Hz), 135.6 (d, *J* = 8.6 Hz), 135.3, 135.2, 134.1, 133.9, 133.8, 133.7 (d, *J* = 2.7 Hz), 133.2, 133.0, 132.8 (d, *J* = 7.7 Hz), 131.6, 130.7, 129.8 (d, *J* = 3.3 Hz), 128.9, 128.5, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.0, 126.8, 126.5, 126.4, 126.3, 125.5, 116.3, 116.1. ³¹P NMR (202 MHz, CDCl₃) δ -14.6 (s, 1P). ¹⁹F NMR (471 MHz, CDCl₃) δ -133.7. The enantiomeric excess was determined by Daicel Chiralpak IB, n-hexane/isopropanol = 99.9/0.1, 0.5 mL/min, λ = 254 nm, t (minor) = 13.6 min, t (major) = 14.4 min. HRMS (ESI-TOF) *m/z*: [M+H]⁺ Calcd for C₃₈H₂₆PFSh⁺: 565.1550; Found: 565.1551.

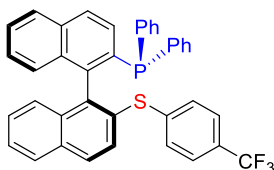


(S)-(2'-((4-Chlorophenyl)thio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3ja). Following General Procedure C (GPC), **3ja** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (67.7 mg, 97% yield, 81% ee). $[\alpha]_D^{23}$ -102.1 (*c* 1.0, CHCl₃) for 81% ee. ¹H NMR (500 MHz, CDCl₃) δ 7.83-7.81 (m, 2H), 7.75-7.72 (m, 2H), 7.42-7.37 (m, 2H), 7.26-7.10 (m, 10H), 7.07-6.98 (m, 8H), 6.93-6.90 (m, 1H), 6.73 (d, *J* = 8.4 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 143.7 (d, *J* = 34.4 Hz), 137.7 (d, *J* = 12.3 Hz), 137.2 (d, *J* = 13.4 Hz), 136.7 (d, *J* = 8.6 Hz), 135.6 (d, *J* = 10.8 Hz), 135.0 (d, *J* = 3.0 Hz), 134.0, 133.9, 133.8, 133.7 (d, *J* = 2.7 Hz), 133.6, 133.5, 133.3, 133.1, 133.0, 132.8 (d, *J* = 7.6 Hz), 131.8, 130.7, 129.1, 128.9, 128.5, 128.3, 128.2, 128.1, 127.9, 127.8, 127.1, 127.0, 126.8, 126.5, 126.4 (d, *J* = 2.6 Hz), 125.7. ³¹P NMR (202 MHz, CDCl₃) δ -14.8 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5

mL/min, λ = 230 nm, t (major) = 21.8 min, t (minor) = 23.9 min. **HRMS** (ESI-TOF) m/z: $[M+H]^+$ Calcd for $C_{38}H_{26}PClSH^+$: 581.1254; Found: 581.1251.

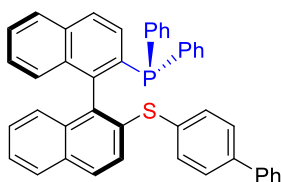


(S)-(2'-((4-Bromophenyl)thio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3ka). Following General Procedure C (GPC), **3ka** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (70.5 mg, 94% yield, 80% ee). $[\alpha]_D^{23}$ -136.0 (*c* 1.0, $CHCl_3$) for 80% ee. **1H NMR** (500 MHz, $CDCl_3$) δ 7.83-7.81 (m, 2H), 7.75 (t, *J* = 7.8 Hz, 2H), 7.43-7.37 (m, 2H), 7.27-7.19 (m, 4H), 7.17-7.10 (m, 8H), 7.07-7.04 (m, 2H), 7.01-6.96 (m, 4H), 6.92 (t, *J* = 7.6 Hz, 1H), 6.74 (d, *J* = 8.5 Hz, 1H). **^{13}C NMR** (126 MHz, $CDCl_3$) δ 143.7 (d, *J* = 34.5 Hz), 137.6 (d, *J* = 12.2 Hz), 137.2 (d, *J* = 13.4 Hz), 136.8 (d, *J* = 8.6 Hz), 135.6 (d, *J* = 10.7 Hz), 134.7 (d, *J* = 3.0 Hz), 134.6, 134.0, 133.9, 133.7 (d, *J* = 2.7 Hz), 133.6, 133.5, 133.1, 132.9, 132.8 (d, *J* = 7.6 Hz), 132.0, 131.9, 130.7 (d, *J* = 2.0 Hz), 129.0, 128.5, 128.3, 128.2, 128.1, 128.0, 127.9 (d, *J* = 4.2 Hz), 127.3, 127.0, 126.8, 126.5 (d, *J* = 34.5 Hz), 126.4 (d, *J* = 2.5 Hz), 125.8, 121.3. **^{31}P NMR** (202 MHz, $CDCl_3$) δ -14.8. (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, λ = 230 nm, t (major) = 22.0 min, t (minor) = 23.9 min. **HRMS** (ESI-TOF) m/z: $[M+H]^+$ Calcd for $C_{38}H_{26}PBrSH^+$: 625.0749; Found: 625.0749.



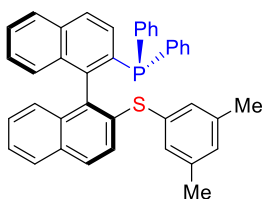
(S)-Diphenyl(2'-((4-(trifluoromethyl)phenyl)thio)-[1,1'-binaphthalen]-2-yl)phosphane (3la). Following General Procedure C (GPC), **3la** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (54.7 mg, 74% yield, 48% ee). $[\alpha]_D^{23}$ +134.0 (*c* 1.0, $CHCl_3$) for 48% ee. **1H NMR** (500 MHz, $CDCl_3$) δ 7.81-7.75 (m, 4H), 7.39-7.35 (m, 3H), 7.29 (t, *J* = 7.4 Hz, 1H), 7.18-7.15 (m, 3H), 7.12-6.85 (m, 14H), 6.81 (d, *J* = 8.5 Hz, 1H). **^{13}C NMR** (126 MHz, $CDCl_3$) δ 143.8 (d, *J* = 34.6 Hz), 141.6 (d, *J* = 0.9 Hz), 139.1 (d, *J* = 8.7 Hz), 137.4 (d, *J* = 2.7 Hz), 137.3 (d, *J* = 0.7 Hz), 135.4 (d, *J* = 10.8 Hz), 134.0, 133.9 (d, *J* = 2.6 Hz), 133.8, 133.6, 133.0, 132.9 (d, *J* = 7.9 Hz), 132.8, 132.7 (d, *J* = 2.9 Hz), 132.4, 130.7, 129.9,

129.3, 128.7, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.1, 126.8, 126.6, 126.4 (d, $J = 2.6$ Hz), 126.3, 125.5, 125.4 (d, $J = 11.2$ Hz), 125.3, 124.1 (q, $J = 272.3$ Hz). ^{31}P NMR (202 MHz, CDCl_3) δ -15.1 (s, 1P). ^{19}F NMR (471 MHz, CDCl_3) δ -62.5. The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, $\lambda = 230$ nm, t (major) = 18.7 min, t (minor) = 20.0 min. **HRMS** (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{39}\text{H}_{26}\text{PF}_3\text{SH}^+$: 615.1518; Found: 615.1518.



(S)-(2'-([1,1'-Biphenyl]-4-ylthio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3ma).

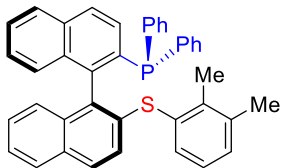
Following General Procedure C (GPC), **3ma** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (71.1 mg, 95% yield, 89% ee). $[\alpha]_{\text{D}}^{23}$ -144.6 (c 1.0, CHCl_3) for 89% ee. ^1H NMR (500 MHz, CDCl_3) δ 7.84-7.81 (m, 2H), 7.76-7.72 (m, 2H), 7.46-7.44 (m, 2H), 7.42-7.39 (m, 2H), 7.35-7.29 (m, 5H), 7.26-7.19 (m, 6H), 7.18-7.10 (m, 6H), 7.07-7.00 (m, 4H), 6.92-6.89 (m, 1H), 6.73 (d, $J = 8.5$ Hz, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 143.9 (d, $J = 34.5$ Hz), 140.3 (2C), 137.9 (d, $J = 12.4$ Hz), 137.2 (d, $J = 13.6$ Hz), 136.2 (d, $J = 8.5$ Hz), 135.7 (d, $J = 11.4$ Hz), 135.6 (d, $J = 2.9$ Hz), 134.2, 134.1, 134.0, 133.8, 133.7 (d, $J = 1.2$ Hz), 133.1, 133.0, 132.9, 132.8, 132.7, 131.7, 130.7, 128.8, 128.7, 128.4, 128.2, 128.1, 127.9, 127.6, 127.5, 127.2, 127.0, 126.7, 126.5 (d, $J = 2.6$ Hz), 126.4, 126.3, 125.5. ^{31}P NMR (202 MHz, CDCl_3) δ -14.7 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, $\lambda = 230$ nm, t (major) = 12.8 min, t (minor) = 14.3 min. **HRMS** (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{44}\text{H}_{31}\text{PSH}^+$: 623.1957; Found: 623.1955.



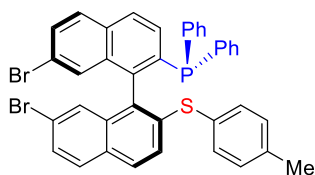
(S)-(2'-((3,5-Dimethylphenyl)thio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3na).

Following General Procedure C (GPC), **3na** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (67.0 mg, 97% yield, 89% ee). $[\alpha]_{\text{D}}^{23}$ -247.8 (c 0.57, CHCl_3) for 89% ee. ^1H NMR

(500 MHz, CDCl₃) δ 7.82-7.80 (m, 2H), 7.72-7.70 (m, 2H), 7.42-7.38 (m, 2H), 7.24-7.21 (m, 3H), 7.20-7.13 (m, 6H), 7.12-7.10 (m, 1H), 7.07-7.00 (m, 4H), 6.93-6.89 (m, 1H), 6.77 (s, 2H), 6.74-6.72 (m, 2H), 2.07 (s, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 144.0 (d, J = 34.8 Hz), 138.6, 137.9 (d, J = 12.5 Hz), 137.5 (d, J = 13.9 Hz), 136.4 (d, J = 2.8 Hz), 135.6 (d, J = 10.4 Hz), 135.4 (d, J = 8.4 Hz), 134.0, 133.9, 133.8, 133.7 (d, J = 2.7 Hz), 133.2, 133.0, 132.8 (d, J = 7.7 Hz), 131.5, 131.0, 130.8, 129.5, 128.6, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.7, 127.0, 126.7, 126.6, 126.5 (d, J = 2.6 Hz), 126.3, 126.2, 125.3, 125.2, 21.1. ³¹P NMR (202 MHz, CDCl₃) δ -14.7 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, λ = 254 nm, t (major) = 18.2 min, t (minor) = 19.1 min. HRMS (ESI-TOF) m/z: [M+H]⁺ Calcd for C₄₀H₃₁PSH⁺: 575.1957; Found: 575.1960.

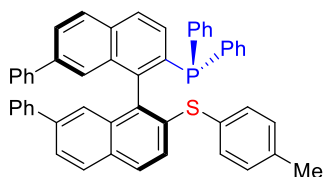


(S)-(2'-((2,3-Dimethylphenyl)thio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (30a). Following General Procedure C (GPC), **30a** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (63.1 mg, 91% yield, 84% ee). [α]_D²³ -54.0 (c 0.5, CHCl₃) for 84% ee. ¹H NMR (500 MHz, CDCl₃) δ 7.85-7.81 (m, 2H), 7.68 (t, J = 19.1 Hz, 2H), 7.44-7.39 (m, 2H), 7.29-7.26 (m, 2H), 7.21-7.16 (m, 7H), 7.13-7.10 (m, 1H), 7.06-6.99 (m, 5H), 6.93-6.88 (m, 2H), 6.86-6.82 (m, 1H), 6.64 (d, J = 2.1 Hz, 1H), 2.18 (s, 3H), 2.07 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 143.7 (d, J = 35.2 Hz), 140.5, 138.4 (d, J = 13.0 Hz), 137.7, 137.4 (d, J = 13.8 Hz), 136.7 (d, J = 2.7 Hz), 136.5 (d, J = 10.9 Hz), 134.1, 134.0, 133.8, 133.7 (d, J = 2.6 Hz), 133.5, 133.1, 133.0, 132.4 (d, J = 7.7 Hz), 131.1, 130.8 (d, J = 2.2 Hz), 130.3, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.7, 126.9, 126.6, 126.2, 126.1, 125.9, 124.9, 124.8, 21.0, 17.2. ³¹P NMR (202 MHz, CDCl₃) δ -14.41 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 99.9/0.1, 0.5 mL/min, λ = 254 nm, t (minor) = 8.7 min, t (major) = 9.0 min. HRMS (ESI-TOF) m/z: [M+H]⁺ Calcd for C₄₀H₃₁PSH⁺: 575.1957; Found: 575.1961.



(S)-(7,7'-Dibromo-2'-(p-tolylthio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3pa).

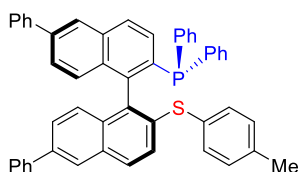
Following General Procedure C (GPC), **3pa** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (69.6 mg, 81% yield, 87% ee). $[\alpha]_D^{23}$ -120.0 (*c* 1.0, CHCl₃) for 87% ee. **¹H NMR** (500 MHz, CDCl₃) δ 7.79 (d, *J* = 8.5 Hz, 1H), 7.68 (d, *J* = 8.7 Hz, 1H), 7.64 (d, *J* = 8.8 Hz, 1H), 7.54 (d, *J* = 8.7 Hz, 1H), 7.48 (dd, *J* = 2.1, 8.6 Hz, 1H), 7.39 (dd, *J* = 2.7, 8.5 Hz, 1H), 7.26-7.24 (m, 2H), 7.20-7.09 (m, 11H), 7.01-6.97 (m, 4H), 6.71 (d, *J* = 1.6 Hz, 1H), 2.23 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 141.7 (d, *J* = 34.4 Hz), 138.7 (d, *J* = 2.9 Hz), 138.4, 138.1 (d, *J* = 12.5 Hz), 137.6 (d, *J* = 12.6 Hz), 136.0 (d, *J* = 12.9 Hz), 134.6 (d, *J* = 2.9 Hz), 134.2, 134.1, 134.0, 133.9, 133.6 (d, *J* = 7.4 Hz), 133.1, 133.0 (d, *J* = 8.2 Hz), 132.9, 132.3, 131.0 (d, *J* = 1.8 Hz), 130.5, 130.1, 130.0, 129.9, 129.8, 129.6, 129.0, 128.7, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1 (d, *J* = 2.7 Hz), 128.0, 127.9, 126.6, 121.2, 121.1, 21.2. **³¹P NMR** (202 MHz, CDCl₃) δ -13.8 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, λ = 230 nm, *t* (minor) = 10.7 min, *t* (major) = 11.7 min. **HRMS** (ESI-TOF) *m/z*: [M+H]⁺ Calcd for C₃₉H₂₇PBr₂SH⁺: 717.0011; Found: 717.0015.



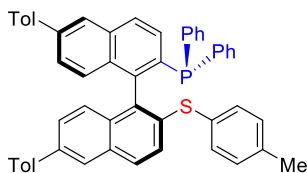
(S)-(7,7'-Diphenyl-2'-(p-tolylthio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3qa).

Following General Procedure C (GPC), **3qa** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (55.3 mg, 65% yield, 75% ee). $[\alpha]_D^{23}$ -113.4 (*c* 1.0, CHCl₃) for 75% ee. **¹H NMR** (500 MHz, CDCl₃) δ 7.88 (d, *J* = 8.5 Hz, 1H), 7.85 (d, *J* = 8.5 Hz, 1H), 7.78 (d, *J* = 8.5 Hz, 1H), 7.73 (d, *J* = 8.7 Hz, 1H), 7.65 (dd, *J* = 1.7, 8.5 Hz, 1H), 7.48-7.45 (m, 2H), 7.37 (s, 1H), 7.30 (d, *J* = 7.5 Hz, 2H), 7.23-7.07 (m, 16H), 7.02-6.91 (m, 8H), 2.20 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 144.2 (d, *J* = 35.0 Hz), 141.2, 141.0, 139.4, 139.1, 138.7 (d, *J* = 12.9 Hz), 137.8, 137.1 (d, *J* = 13.9 Hz), 137.0 (d, *J* = 2.8 Hz), 136.5 (d, *J* = 10.7 Hz), 135.5 (d, *J* = 8.8 Hz), 134.2, 134.1, 134.0, 133.5, 133.1, 133.0 (d, *J* = 7.7 Hz), 132.9, 132.8,

131.0, 130.9 (d, $J = 2.2$ Hz), 130.7, 129.9, 128.8, 128.7, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.7, 127.5, 127.4, 127.2, 127.0, 126.9, 126.5, 125.1, 124.5 (d, $J = 2.6$ Hz), 124.3, 21.2. **^{31}P NMR** (202 MHz, CDCl_3) δ -14.8 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, $\lambda = 254$ nm, t (minor) = 12.5 min, t (major) = 13.7 min. **HRMS** (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{51}\text{H}_{37}\text{PSH}^+$: 713.2426; Found: 713.2425.

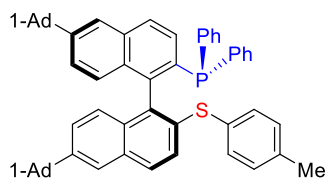


(S)-(6,6'-Diphenyl-2'-(*p*-tolylthio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3ra). Following General Procedure C (GPC), **3ra** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (82.2 mg, 96% yield, 91% ee). $[\alpha]_{\text{D}}^{23}$ -59.0 (c 1.0, CHCl_3) for 91% ee. **^1H NMR** (500 MHz, CDCl_3) δ 8.00 (d, $J = 1.5$ Hz, 1H), 7.88-7.85(m, 2H), 7.73 (d, $J = 8.8$ Hz, 1H), 7.57 (d, $J = 7.4$ Hz, 2H), 7.50 (d, $J = 7.4$ Hz, 2H), 7.46 (dd, $J = 1.9, 8.9$ Hz, 1H), 7.42 (dd, $J = 2.8, 8.5$ Hz, 1H), 7.34-7.21 (m, 10H), 7.19-7.08 (m, 7H), 7.02-7.01 (m, 4H), 6.92 (d, $J = 8.0$ Hz, 2H), 6.73 (d, $J = 8.8$ Hz, 1H), 2.18 (s, 3H). **^{13}C NMR** (126 MHz, CDCl_3) δ 143.5 (d, $J = 34.5$ Hz), 140.9, 141.0, 139.7, 138.0 (d, $J = 12.9$ Hz), 137.9, 137.8, 137.1 (d, $J = 13.4$ Hz), 136.9 (d, $J = 2.8$ Hz), 136.2 (d, $J = 11.0$ Hz), 134.8 (d, $J = 8.5$ Hz), 134.4, 134.2, 134.1, 133.7, 133.2, 133.1, 132.9 (d, $J = 2.6$ Hz), 132.0 (d, $J = 7.6$ Hz), 131.8, 131.2, 130.8, 130.0, 129.0, 128.9, 128.8, 128.7, 128.5, 128.4, 128.3, 128.2, 127.9, 127.6, 127.5, 127.3, 127.1 (d, $J = 2.4$ Hz), 126.8, 126.7, 126.5, 126.1, 125.9, 125.8, 21.3. **^{31}P NMR** (202 MHz, CDCl_3) δ -14.3 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, $\lambda = 254$ nm, t (minor) = 17.2 min, t (major) = 18.7 min. **HRMS** (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{51}\text{H}_{37}\text{PSH}^+$: 713.2426; Found: 713.2422.



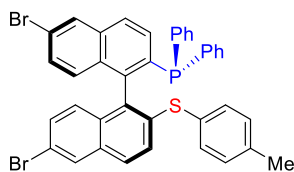
(S)-(6,6'-Di-*p*-tolyl-2'-(*p*-tolylthio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3sa). Following General Procedure C (GPC), **3sa** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white

solid (81.0 mg, 91% yield, 94% ee). $[\alpha]_D^{23}$ -27 (*c* 1.0, CHCl₃) for 94% ee. **¹H NMR** (500 MHz, CDCl₃) δ 8.00 (d, *J* = 1.7 Hz, 1H), 7.88-7.86 (m, 2H), 7.74 (d, *J* = 8.8 Hz, 1H), 7.50-7.41 (m, 6H), 7.27-7.23 (m, 4H), 7.19-7.02 (m, 12H), 7.04-7.02 (m, 3H), 6.94 (d, *J* = 7.9 Hz, 2H), 6.73 (d, *J* = 8.8 Hz, 1H), 2.30 (s, 6H), 2.21 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 143.5 (d, *J* = 34.6 Hz), 139.5, 138.1, 138.0 (d, *J* = 12.8 Hz), 137.9, 137.8, 137.7, 137.4, 137.1 (d, *J* = 13.4 Hz), 137.0, 136.6 (d, *J* = 2.8 Hz), 135.9 (d, *J* = 10.8 Hz), 134.9 (d, *J* = 8.5 Hz), 134.3, 134.1, 133.6, 133.2, 133.0, 132.8 (d, *J* = 2.6 Hz), 131.8 (d, *J* = 7.4 Hz), 131.7, 131.1 (d, *J* = 2.1 Hz), 130.9, 129.9, 129.7, 129.6, 129.5, 128.8, 128.7, 128.4, 128.3, 128.2, 128.1, 127.9, 127.3, 127.2, 127.1, 127.0 (d, *J* = 2.6 Hz), 126.8, 126.7, 126.4, 125.8, 125.7, 125.4, 121.3, 21.2, 21.1, 21.0. **³¹P NMR** (202 MHz, CDCl₃) δ -14.3 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, λ = 254 nm, *t* (minor) = 18.9 min, *t* (major) = 21.9 min. **HRMS** (ESI-TOF) *m/z*: [M+H]⁺ Calcd for C₅₃H₄₁PSH⁺: 741.2739; Found: 741.2741.

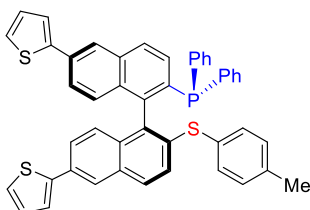


((S)-6,6'-Di((3S,5S,7S)-adamantan-1-yl)-2'-(*p*-tolylthio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3ta). Following General Procedure C (GPC), **3ta** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (85.6 mg, 86% yield, 97% ee). $[\alpha]_D^{23}$ +75.0 (*c* 1.0, CHCl₃) for 97% ee. **¹H NMR** (500 MHz, CDCl₃) δ 7.89-7.84 (m, 2H), 7.69 (d, *J* = 1.8 Hz, 1H), 7.66-7.62 (m, 2H), 7.40-7.36 (m, 2H), 7.34 (d, *J* = 1.7 Hz, 1H), 7.23-7.15 (m, 7H), 7.06-7.02 (m, 2H), 6.99-6.97 (m, 2H), 6.94-6.92 (m, 2H), 6.87-6.83 (m, 2H), 6.74 (d, *J* = 8.7 Hz, 1H), 2.24 (s, 3H), 2.05-2.03 (s, 6H), 1.90-1.87 (m, 12H), 1.75-1.66 (m, 12H). **¹³C NMR** (126 MHz, CDCl₃) δ 151.0, 148.3, 139.9 (d, *J* = 7.7 Hz), 137.9, 136.0, 134.6 (d, *J* = 2.3 Hz), 133.8, 132.9 (d, *J* = 11.3 Hz), 132.4, 132.3, 132.0, 131.9, 131.6 (d, *J* = 4.7 Hz), 131.3, 130.8 (d, *J* = 11.8 Hz), 130.6, 130.5, 130.4, 130.3, 129.8, 129.7, 129.3, 128.4, 128.3, 127.5, 127.4, 126.9, 126.8, 126.6, 125.9, 125.5, 125.2, 124.7, 123.1, 122.6, 43.3, 42.9, 36.8, 36.7, 36.4, 36.1, 28.9, 28.8, 21.2. **³¹P NMR** (202 MHz, CDCl₃) δ -14.5 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 95.0/5.0, 1.0 mL/min, λ = 254 nm, *t* (major) = 6.7 min, *t* (minor) =

14.8 min. **HRMS** (ESI-TOF) m/z : $[M+H]^+$ Calcd for $C_{59}H_{57}PSH^+$: 829.3991; Found: 829.3993.

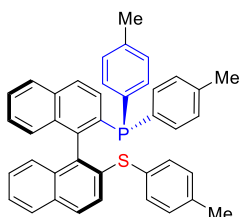


(S)-(6,6'-Dibromo-2'-(p-tolylthio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3ua). Following General Procedure C (GPC), **3ua** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (58.5 mg, 68% yield, 81% ee). $[\alpha]_D^{23} +41.0$ (c 1.0, $CHCl_3$) for 81% ee. **1H NMR** (500 MHz, $CDCl_3$) δ 7.99 (d, J = 1.9 Hz, 1H), 7.84 (d, J = 1.9 Hz, 1H), 7.74 (d, J = 8.5 Hz, 1H), 7.60 (d, J = 8.8 Hz, 1H), 7.41 (dd, J = 2.1, 8.5 Hz, 1H), 7.28 (dd, J = 2.0, 9.0 Hz, 1H), 7.23-7.10 (m, 9H), 7.07 (t, J = 7.5 Hz, 2H), 6.99-6.93 (m, 5H), 6.84 (dd, J = 2.0, 9.0 Hz, 1H), 6.38 (d, J = 9.0 Hz, 1H), 2.25 (s, 3H). **^{13}C NMR** (126 MHz, $CDCl_3$) δ 142.9 (d, J = 34.3 Hz), 138.7 (d, J = 22.7 Hz), 138.4, 138.0, 134.9, 134.3, 134.2, 134.1, 134.0, 133.9, 133.8 (d, J = 3.4 Hz), 133.5, 133.1, 133.0, 132.7, 132.6 (d, J = 35.1 Hz), 130.6, 130.3, 130.2 (d, J = 10.8 Hz), 130.1, 129.8 (d, J = 13.6 Hz), 129.5, 128.68 (d, J = 14.4 Hz), 128.4, 128.3, 128.2, 128.1, 128.0 (d, J = 3.1 Hz), 127.7 (d, J = 19.2 Hz), 127.5, 127.1, 126.5, 126.4 (d, J = 2.9 Hz), 122.0, 121.5, 121.3, 119.3, 118.8, 21.2. **^{31}P NMR** (202 MHz, $CDCl_3$) δ -14.1 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 99.9/0.1, 0.5 mL/min, λ = 254 nm, t (minor) = 11.1 min, t (major) = 11.7 min. **HRMS** (ESI-TOF) m/z : $[M+H]^+$ Calcd for $C_{39}H_{27}PBr_2SH^+$: 717.0011; Found: 717.0010.

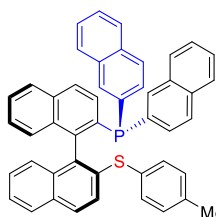


(S)-(6,6'-Di(thiophen-2-yl)-2'-(p-tolylthio)-[1,1'-binaphthalen]-2-yl)diphenylphosphane (3va). Following General Procedure C (GPC), **3va** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (45.9 mg, 53% yield, 89% ee). $[\alpha]_D^{23} -71.4$ (c 0.64, $CHCl_3$) for 89% ee. **1H NMR** (500 MHz, $CDCl_3$) δ 8.02 (d, J = 1.7 Hz, 1H), 7.90 (d, J = 1.7 Hz, 1H), 7.86 (d, J = 8.5 Hz, 1H), 7.72 (d, J = 8.8 Hz, 1H), 7.48 (dd, J = 1.8, 8.8 Hz, 1H), 7.45 (dd, J = 1.3, 2.9 Hz, 1H), 7.43-7.40 (m, 2H), 7.37-7.34 (m, 2H), 7.33-7.30 (m, 2H), 7.26-7.20

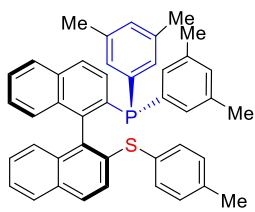
(m, 4H), 7.18-7.17 (m, 2H), 7.14-7.10 (m, 4H), 7.08-7.02 (m, 5H), 6.95 (d, $J = 8.0$ Hz, 2H), 6.69 (d, $J = 8.8$ Hz, 1H), 2.23 (s, 3H). **^{13}C NMR** (126 MHz, CDCl_3) δ 143.4, 142.2, 142.0, 137.9, 136.7 (d, $J = 2.9$ Hz), 135.9 (d, $J = 10.8$ Hz), 134.8, 134.3, 134.2, 134.1, 133.7, 133.2, 133.0, 132.8 (d, $J = 2.5$ Hz), 132.6 (d, $J = 2.4$ Hz), 132.5, 131.7, 131.2 (d, $J = 2.2$ Hz), 130.7, 130.5, 130.0, 139.0, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1, 127.9, 127.0 (d, $J = 2.7$ Hz), 126.8, 126.6, 126.5, 126.4, 126.3, 126.2, 125.9, 125.4, 125.3, 125.0, 124.8, 121.0, 120.5, 21.2. **^{31}P NMR** (202 MHz, CDCl_3) δ -14.4 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, $\lambda = 254$ nm, t (major) = 14.0 min, t (minor) = 16.7 min. **HRMS** (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{47}\text{H}_{33}\text{PS}_3\text{H}^+$: 725.1555; Found: 725.1552.



(S)-Di-*p*-tolyl(2'-(*p*-tolylthio)-[1,1'-binaphthalen]-2-yl)phosphane (3ab). Following General Procedure C (GPC), **3ab** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (65.8 mg, 93% yield, 91% ee). $[\alpha]_{\text{D}}^{23}$ -92.0 (c 1.0, CHCl_3) for 91% ee. **^1H NMR** (500 MHz, CDCl_3) δ 7.81 (d, $J = 8.4$ Hz, 2H), 7.70-7.68 (m, 2H), 7.42-7.38 (m, 2H), 7.16-7.18 (m, 2H), 7.15-7.10 (m, 4H), 7.07 (d, $J = 8.0$ Hz, 2H), 6.96-6.86 (m, 9H), 6.69 (d, $J = 8.5$ Hz, 1H), 2.21 (s, 3H), 2.20 (s, 3H), 2.18 (s, 3H). **^{13}C NMR** (126 MHz, CDCl_3) δ 143.5 (d, $J = 34.2$ Hz), 138.2, 137.6, 137.5, 136.7 (d, $J = 2.9$ Hz), 126.4 (d, $J = 11.3$ Hz), 135.2 (d, $J = 8.5$ Hz), 134.5 (d, $J = 11.2$ Hz), 134.1, 133.9, 133.8 (d, $J = 17.3$ Hz), 133.7, 133.4, 133.2, 133.1, 132.7 (d, $J = 7.5$ Hz), 131.5, 131.0, 130.6 (d, $J = 1.9$ Hz), 129.8, 129.1, 129.0, 128.9, 128.8, 128.7, 128.5, 128.3, 128.1, 127.8, 126.8, 126.6, 126.4 (d, $J = 2.5$ Hz), 126.3, 126.2, 126.1, 125.1, 21.3, 21.2, 21.1. **^{31}P NMR** (202 MHz, CDCl_3) δ -16.0 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IB, n-hexane/isopropanol = 99.9/0.1, 0.5 mL/min, $\lambda = 254$ nm, t (minor) = 11.3 min, t (major) = 12.6 min. **HRMS** (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{41}\text{H}_{33}\text{PSH}^+$: 589.2113; Found: 589.2109.

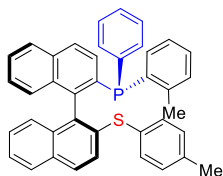


(S)-Di(naphthalen-2-yl) (2'-(p-tolylthio)-[1,1'-binaphthalen]-2-yl)phosphane (3ac). Following General Procedure C (GPC), **3ac** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (65.9 mg, 83% yield, 96% ee). $[\alpha]_D^{23}$ -159.1 (*c* 0.45, CHCl₃) for 96% ee. ¹H NMR (500 MHz, CDCl₃) δ 7.84-7.81 (m, 2H), 7.73 (d, *J* = 8.8 Hz, 1H), 7.70-7.63 (m, 5H), 7.59 (d, *J* = 8.4 Hz, 1H), 7.53-7.49 (m, 2H), 7.44-7.41 (m, 3H), 7.38-7.19 (m, 9H), 7.10-7.07 (m, 1H), 7.05 (d, *J* = 8.1 Hz, 2H), 6.83 (d, *J* = 8.0 Hz, 2H), 6.67 (d, *J* = 3.8 Hz, 2H), 2.15 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 144.0 (d, *J* = 34.3 Hz), 137.6, 136.5 (d, *J* = 3.1 Hz), 135.2, 135.0, 134.4, 134.3, 133.8, 133.7 (d, *J* = 2.5 Hz), 133.3, 133.2 (d, *J* = 2.1 Hz), 133.1 (2C), 133.0, 132.9, 132.8 (d, *J* = 7.6 Hz), 131.5, 130.8, 130.6 (d, *J* = 2.2 Hz), 130.3, 130.1, 129.9, 129.8, 129.7, 128.7, 128.5, 128.2, 128.1, 128.0, 127.7 (3C), 127.6 (2C), 127.5, 127.0, 126.7, 126.5 (3C), 126.1 (3C), 126.0 (2C), 125.2, 21.1. ³¹P NMR (202 MHz, CDCl₃) δ -13.1 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IB, n-hexane/isopropanol = 99.9/0.1, 0.5 mL/min, λ = 254 nm, *t* (minor) = 10.6 min, *t* (major) = 15.6 min. HRMS (ESI-TOF) *m/z*: [M+H]⁺ Calcd for C₄₇H₃₃PSH⁺: 661.2113; Found: 661.2115.

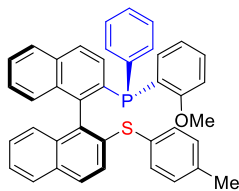


(S)-Bis(3,5-dimethylphenyl)(2'-(p-tolylthio)-[1,1'-binaphthalen]-2-yl)phosphane (3ad). Following General Procedure C (GPC), **3ad** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (68.8 mg, 93% yield, 81% ee). $[\alpha]_D^{23}$ -160.0 (*c* 1.0, CHCl₃) for 81% ee. ¹H NMR (500 MHz, CDCl₃) δ 7.80 (d, *J* = 8.5 Hz, 2H), 7.68 (dd, *J* = 3.2, 8.4 Hz, 2H), 7.46 (dd, *J* = 2.8, 8.6 Hz, 1H), 7.39-7.36 (m, 1H), 7.21-7.16 (m, 4H), 7.11 (d, *J* = 8.1 Hz, 2H), 6.91 (d, *J* = 8.0 Hz, 2H), 6.86 (d, *J* = 7.7 Hz, 3H), 6.72 (d, *J* = 21.6 Hz, 2H), 6.67 (d, *J* = 8.5 Hz, 1H), 6.61 (d, *J* = 8.1 Hz, 2H), 2.20 (s, 3H), 2.06 (s, 6H), 2.02 (s, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 143.5 (d, *J* = 34.2 Hz), 137.8 (d, *J* = 12.0 Hz), 137.6, 137.5, 137.4, 137.3, 137.2,

136.8, 136.7, 136.6, 136.4 (d, $J = 3.3$ Hz), 135.6 (d, $J = 8.4$ Hz), 133.8 (d, $J = 2.5$ Hz), 133.7, 133.2, 133.1, 132.8 (d, $J = 7.5$ Hz), 132.1, 131.9, 131.6, 131.3, 130.9, 130.8 (d, $J = 1.8$ Hz), 130.7, 130.3, 129.8, 129.6, 128.6, 128.4, 128.2, 127.7, 126.8, 126.7, 126.6, 126.5, 126.4, 126.0, 125.2, 21.4, 21.3, 21.2. ^{31}P NMR (202 MHz, CDCl_3) δ -13.7 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, $\lambda = 230$ nm, t (major) = 15.5 min, t (minor) = 19.4 min. HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{43}\text{H}_{37}\text{PSH}^+$: 617.2426; Found: 617.2424.

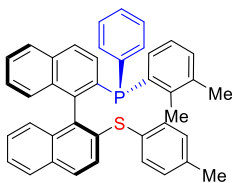


(S)-Phenyl(*o*-tolyl)((*S*)-2'-(*p*-tolylthio)-[1,1'-binaphthalen]-2-yl)phosphane (3ae). Following General Procedure C (GPC), **3ae** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (45.7 mg, 66% yield, 97% ee, 13:1 dr). $[\alpha]_{\text{D}}^{23}$ -78.0 (c 1.0, CHCl_3) for 97% ee. ^1H NMR (500 MHz, CDCl_3) δ 7.83-7.81 (m, 2H), 7.66 (d, $J = 8.6$ Hz, 2H), 7.40 (t, $J = 7.4$ Hz, 1H), 7.32 (dd, $J = 2.8, 8.5$ Hz, 1H), 7.21-7.10 (m, 12H), 6.96 (d, $J = 8.1$ Hz, 2H), 6.94-6.92 (m, 2H), 6.80-6.73 (m, 2H), 6.61 (d, $J = 8.5$ Hz, 1H), 2.23 (s, 3H), 1.75 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 143.4 (d, $J = 21.9$ Hz), 143.1 (d, $J = 16.3$ Hz), 137.8, 137.5 (d, $J = 12.8$ Hz), 136.8 (d, $J = 2.8$ Hz), 136.0 (d, $J = 11.6$ Hz), 135.3 (d, $J = 14.2$ Hz), 134.6 (d, $J = 8.1$ Hz), 134.4, 133.7, 133.6, 133.5, 133.4 (d, $J = 2.6$ Hz), 132.8 (d, $J = 7.2$ Hz), 131.5, 131.0, 130.5 (d, $J = 1.3$ Hz), 139.9, 129.7 (d, $J = 5.0$ Hz), 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.6, 126.7, 126.6, 126.3 (d, $J = 2.7$ Hz), 126.2, 125.9, 125.7, 125.1, 21.3 (d, $J = 23.4$ Hz), 21.2. ^{31}P NMR (202 MHz, CDCl_3) δ -24.1 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, $\lambda = 280$ nm, t (major) = 9.3 min, t (minor) = 11.0 min. HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{40}\text{H}_{31}\text{PSH}^+$: 575.1957; Found: 575.1958.



(*R*)-(2-Methoxyphenyl)(phenyl)((*S*)-2'-(*p*-tolylthio)-[1,1'-binaphthalen]-2-yl)phosphane (3af). Following General Procedure C (GPC), **3af** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10)

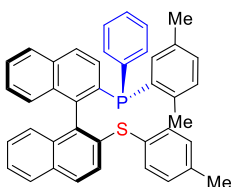
as a white solid (45.4 mg, 64% yield, 50% ee, 6:1 dr). $[\alpha]_D^{23}$ -68.0 (*c* 0.64, CHCl₃) for 50% ee. **¹H NMR** (500 MHz, CDCl₃) δ 7.82 (d, *J* = 8.2 Hz, 1H), 7.80 (d, *J* = 8.5 Hz, 1H), 7.70 (d, *J* = 8.2 Hz, 1H), 7.64 (d, *J* = 8.8 Hz, 1H), 7.41-7.38 (m, 1H), 7.35-7.32 (m, 1H), 7.27-7.11 (m, 10H), 7.05-6.98 (m, 3H), 6.91-6.87 (m, 3H), 6.70 (t, *J* = 7.4 Hz, 1H), 6.66-6.61 (m, 2H), 3.41 (s, 3H), 2.21 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 161.1 (d, *J* = 15.3 Hz), 143.2 (d, *J* = 35.0 Hz), 137.6, 137.1 (d, *J* = 2.9 Hz), 136.7 (d, *J* = 12.7 Hz), 135.4 (d, *J* = 11.0 Hz), 134.6 (d, *J* = 33.2 Hz), 134.3 (d, *J* = 3.5 Hz), 134.2, 134.1, 133.7, 133.6, 132.8 (d, *J* = 7.8 Hz), 131.5, 130.8 (d, *J* = 2.1 Hz), 130.7, 129.9, 129.8, 129.0, 128.4, 128.2, 128.1, 128.0, 127.8, 126.7, 126.6, 126.5, 126.4, 126.3 (d, *J* = 2.5 Hz), 126.0, 125.2, 120.7, 110.1, 55.2, 21.2. **³¹P NMR** (202 MHz, CDCl₃) δ -25.3 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, λ = 280 nm, *t* (major) = 11.8 min, *t* (minor) = 13.8 min. **HRMS** (ESI-TOF) *m/z*: [M+H]⁺ Calcd for C₄₀H₃₁POSH⁺: 591.1906; Found: 591.1903.



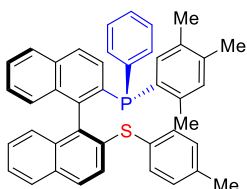
(S)-(2,3-Dimethylphenyl)(phenyl)((S)-2'-(p-tolylthio)-[1,1'-

binaphthalen]-2-yl)phosphane (3ag). Following General Procedure C (GPC), **3ag** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (43.2 mg, 61% yield, 92% ee, 10:1 dr). $[\alpha]_D^{23}$ -120.6 (*c* 1.0, CHCl₃) for 92% ee. **¹H NMR** (500 MHz, CDCl₃) δ 7.80 (d, *J* = 8.4 Hz, 2H), 7.66-7.63 (m, 2H), 7.38-7.35 (m, 1H), 7.33 (dd, *J* = 2.8, 8.5 Hz, 1H), 7.23-7.10 (m, 12H), 6.96 (d, *J* = 8.1 Hz, 2H), 6.86-6.83 (m, 2H), 6.70-6.66 (m, 1H), 6.51 (d, *J* = 8.4 Hz, 1H), 2.22 (s, 3H), 1.92 (s, 3H), 1.62 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 143.0 (d, *J* = 33.9 Hz), 142.1 (d, *J* = 27.8 Hz), 138.0 (d, *J* = 12.9 Hz), 137.8, 136.9 (d, *J* = 12.1 Hz), 136.8 (d, *J* = 2.8 Hz), 136.4 (d, *J* = 4.8 Hz), 135.1 (d, *J* = 11.7 Hz), 134.7 (d, *J* = 8.1 Hz), 133.7, 133.6, 133.5 (d, *J* = 2.6 Hz), 133.4, 132.7, 131.6, 131.1, 130.4, 130.1, 129.9, 129.8, 128.4, 128.3, 128.2, 128.1, 127.9, 127.4, 126.7, 126.6, 126.5, 126.4, 126.3 (d, *J* = 2.6 Hz), 126.1, 125.8, 125.4, 125.0, 21.2, 20.5 (d, *J* = 2.9 Hz), 17.6 (d, *J* = 29.4 Hz). **³¹P NMR** (202 MHz, CDCl₃) δ -23.9 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, λ = 280 nm, *t* (major) = 8.6 min, *t* (minor) =

9.8 min. **HRMS** (ESI-TOF) m/z : $[M+H]^+$ Calcd for $C_{41}H_{33}PSH^+$: 589.2113; Found: 589.2117.



(S)-(2,5-Dimethylphenyl)(phenyl)((S)-2'-(p-tolylthio)-[1,1'-binaphthalen]-2-yl)phosphane (3ah). Following General Procedure C (GPC), **3ah** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (50.1 mg, 71% yield, 91% ee, 16:1 dr). $[\alpha]_D^{23}$ -37.0 (c 1.0, $CHCl_3$) for 91% ee. **1H NMR** (500 MHz, $CDCl_3$) δ 7.83-7.80 (m, 2H), 7.64 (d, J = 8.6 Hz, 2H), 7.41-7.37 (m, 1H), 7.31 (dd, J = 2.8, 8.5 Hz, 1H), 7.21-7.11 (m, 11H), 6.95 (d, J = 7.9 Hz, 1H), 6.80-6.74 (m, 2H), 6.69-6.62 (m, 2H), 6.61 (d, J = 8.4 Hz, 1H), 2.22 (s, 3H), 2.05 (s, 3H), 1.70 (s, 3H). **^{13}C NMR** (126 MHz, $CDCl_3$) δ 143.2 (d, J = 33.7 Hz), 140.0 (d, J = 28.0 Hz), 137.8, 137.5 (d, J = 12.9 Hz), 126.8 (d, J = 2.8 Hz), 126.2 (d, J = 11.9 Hz), 134.9, 134.8, 134.7 (d, J = 7.8 Hz), 133.8, 133.7, 133.6, 133.4 (d, J = 2.6 Hz), 133.2 (d, J = 1.6 Hz), 133.1 (d, J = 1.0 Hz), 132.8 (d, J = 7.2 Hz), 131.5, 131.1, 130.6, 129.9, 129.8 (d, J = 5.9 Hz), 129.6 (d, J = 4.2 Hz), 129.2, 128.5, 128.4, 128.3, 128.2, 128.1 (d, J = 8.0 Hz), 128.0, 127.6, 126.7, 126.6, 126.5, 126.3, 125.8, 125.1, 21.2, 21.1, 20.8 (d, J = 23.0 Hz). **^{31}P NMR** (202 MHz, $CDCl_3$) δ -23.2 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, λ = 230 nm, t (major) = 16.3 min, t (minor) = 20.0 min. **HRMS** (ESI-TOF) m/z : $[M+H]^+$ Calcd for $C_{41}H_{33}PSH^+$: 589.2113; Found: 589.2116.



(S)-Phenyl((S)-2'-(p-tolylthio)-[1,1'-binaphthalen]-2-yl)(2,4,5-trimethylphenyl)phosphane (3ai). Following General Procedure C (GPC), **3ai** was prepared and purified by flash column chromatography (EtOAc/petroleum ether = 1/10) as a white solid (47.0 mg, 65% yield, 89% ee, 10:1 dr). $[\alpha]_D^{23}$ -92.0 (c 1.0, $CHCl_3$) for 89% ee. **1H NMR** (500 MHz, $CDCl_3$) δ 7.82-7.79 (m, 2H), 7.64-7.63 (m, 2H), 7.40-7.34 (m, 2H), 7.23-7.11 (m, 10H), 7.00-6.98 (m, 1H), 6.95 (d, J = 7.9 Hz, 2H), 6.75-6.73 (m, 1H), 6.63 (d, J = 4.4 Hz, 1H), 6.58 (d, J = 8.6 Hz, 1H), 6.53 (d, J = 5.0 Hz, 1H), 2.22 (s,

3H), 2.03 (s, 3H), 1.96 (s, 3H), 1.67 (s, 3H). **¹³C NMR** (126 MHz, CDCl₃) δ 143.0 (d, *J* = 33.5 Hz), 140.6 (d, *J* = 28.2 Hz), 137.8 (d, *J* = 12.6 Hz), 137.7, 136.9, 135.7, 134.7, 134.5, 133.7, 133.6, 133.5, 133.4 (d, *J* = 1.2 Hz), 133.3 (d, *J* = 2.5 Hz), 132.8 (d, *J* = 7.2 Hz), 132.7, 131.6 (d, *J* = 12.4 Hz), 131.5, 131.2, 131.1, 130.5, 129.9, 129.7, 128.4, 128.3, 128.2, 128.1, 127.9, 127.6, 126.6, 126.5, 126.4, 136.3, 126.2, 125.7, 125.0, 21.2, 20.7 (d, *J* = 22.7 Hz), 19.4, 19.3. **³¹P NMR** (202 MHz, CDCl₃) δ -24.1 (s, 1P). The enantiomeric excess was determined by Daicel Chiralpak IA, n-hexane/isopropanol = 98.6/1.4, 0.5 mL/min, λ = 254 nm, t (major) = 8.2 min, t (minor) = 9.7 min. **HRMS** (ESI-TOF) m/z: [M+H]⁺ Calcd for C₄₂H₃₅PSH⁺: 603.2270; Found: 603.2274.

7. References

- (1) Shimada, T.; Cho, Y.-H.; Hayashi, T. Nickel-Catalyzed Asymmetric Grignard Cross-Coupling of Dinaphthothiophene Giving Axially Chiral 1,1'-Binaphthyls. *J. Am. Chem. Soc.* **2002**, *124*, 13396– 13397.
- (2) Cho, Y.-H.; Kina, A.; Shimada, T.; Hayashi, T. Asymmetric synthesis of axially chiral biaryls by nickel-catalyzed Grignard cross-coupling dibenzothiophenes. *J. Org. Chem.* **2004**, *69*, 3811– 3823.
- (3) Zhang, Q.; Xue, X.; Hong, B.; Gu, Z. Torsional strain inversed chemoselectivity in a Pd-catalyzed atroposelective carbonylation reaction of dibenzothiophenium. *Chem. Sci.* **2022**, *13*, 3761– 3765.
- (4) Busacca, C. A.; Lorenz, J. C.; Grinberg, N.; Haddad, N.; Hrapchak, M.; Latli, B.; Senanayake, C. H. A Superior Method for the Reduction of Secondary Phosphine Oxides. *Org. Lett.* **2005**, *7* (19), 4277– 4280.
- (5) Huang, Y.; Li, Y.; Leung, P. H.; Hayashi, T. Asymmetric Synthesis of P-Stereogenic Diarylphosphinites by Palladium-Catalyzed Enantioselective Addition of Diarylphosphines to Benzoquinones. *J. Am. Chem. Soc.* **2014**, *136*, 4865–4868.
- (6) Vedejs, E.; Donde, Y. Stereogenic P-Trisubstituted Phosphorus by Crystallization-Induced Asymmetric Transformation: A Practical Synthesis of Phenyl(o-anisyl)methylphosphine Borane. *J. Am. Chem. Soc.* **1997**, *119*, 9293– 9294.

-
- (7) Zhu, J.; Ye, Y.; Huang, Y. Palladacycle-Catalyzed Olefinic C–P Cross-Coupling of Alkenylsulfonium Salts with Diarylphosphines to Access Alkenylphosphines. *Organometallics* **2022**, *41*, 2342–2348.
- (8) C. A. Sojda, M. C. Kozlowski. HAT Catalysts to Facilitate Acid Mediated Cleavage of Biaryl C–C Bonds in Binaphthols. *Org. Lett.* **2022**, *24*, 8326–8330.
- (9) Shimada, T.; Cho, Y.-H.; Hayashi, T. Nickel-catalyzed asymmetric Grignard cross-coupling of dinaphthothiophene giving axially chiral 1,1'-Binaphthyls. *J. Am. Chem. Soc.* **2002**, *124*, 13396–13397.
- (10) Blank, N. F.; Moncarz, J. R.; Brunner, T. J.; Scriban, C.; Anderson, B. J.; Amir, O.; Glueck, D. S.; Zakharov, L. N.; Golen, J. A.; Incarvito, C. D.; Rheingold, A. L. Palladium-Catalyzed Asymmetric Phosphination. Scope, Mechanism, and Origin of Enantioselectivity. *J. Am. Chem. Soc.* **2007**, *129*, 6847–6858.
- (11) Chan, V. S.; Bergman, R. G.; Toste, F. D. Pd-Catalyzed Dynamic Kinetic Enantioselective Arylation of Silylphosphines. *J. Am. Chem. Soc.* **2007**, *129*, 15122–15123.
- (12) Shintani, R.; Duan, W.-L.; Okamoto, K.; Hayashi, T. Palladium/chiral phosphine-olefin complexes: X-ray crystallographic analysis and the use in catalytic asymmetric allylic alkylation. *Tetrahedron: Asymmetry* **2005**, *16*, 3400–3405.
- (13) Lam, F. L.; Au-Yeung, T. T.-L.; Kwong, F. Y.; Zhou, Z.; Wong, K. Y.; Chan, A. S. C. Palladium-(S_pR)-FerroNPS-Catalyzed Asymmetric Allylic Etherification: Electronic Effect of Nonconjugated Substituents on Benzylic Alcohols on Enantioselectivity. *Angew. Chem., Int. Ed.* **2008**, *47*, 1280–1283.
- (14) Chen, J. M.; Lang, F.; Li, D.; Cun, L. F.; Zhu, J.; Deng, J. G.; Liao, J. Palladium-catalyzed asymmetric allylic nucleophilic substitution reactions using chiral tert-butanefulfinylphosphine ligands. *Tetrahedron: Asymmetry* **2009**, *20*, 1953–1956.
- (15) J.-H. Wu, S. Fang, X. Zheng, J. He, Y. Ma, Z. Su, T. Wang. Organocatalytic Dynamic Kinetic Resolution Enabled Asymmetric Synthesis of Phosphorus-Containing Chiral Helicenes. *Angew. Chem. Int. Ed.* **2023**, *62*, e202309515.

8. X-Ray Crystallographic Data of 1a, 3aa, 3ae

(1) Crystal data and structure refinement for compound 1a

Colorless crystals of product **1a** (CCDC-2303113) suitable for X-ray crystallographic analysis were obtained by recrystallization from CHCl₃/acetone (via evaporation). The ORTEP drawing of compound **1a** is shown in Figure S1.

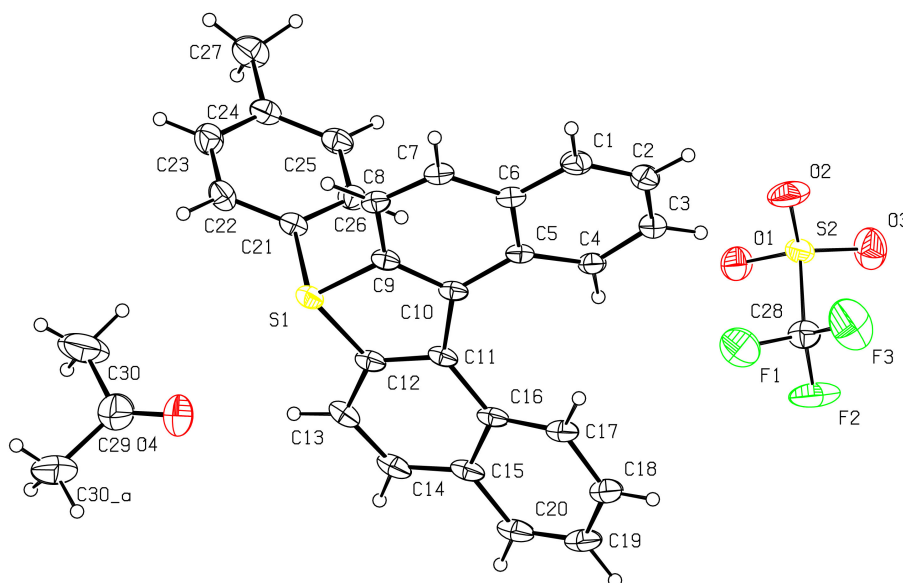


Figure S1. ORTEP illustration of compound 1a with thermal ellipsoids drawn at 50% probability level.

Table S2. Crystal data and structure refinement for compound 1a

Identification code	230914_HYH_S1
Empirical formula	C ₂₉ H ₄₄ F ₆ O ₄ S ₁
Formula weight	1107.18
Temperature/K	170.00
Crystal system	orthorhombic
Space group	Pbcn
a/Å	23.6038 (16)
b/Å	12.3774 (9)
c/Å	17.1095 (11)

$\alpha / ^\circ$	90
$\beta / ^\circ$	90
$\gamma / ^\circ$	90
Volume/ $\text{\AA}^3$	4998.6(6)
Z	4
$\rho_{\text{calc}}/\text{g}/\text{cm}^3$	1.471
μ / mm^{-1}	1.587
F(000)	2288.0
Crystal size/ mm^3	$0.15 \times 0.09 \times 0.04$
Radiation	GaK α ($\lambda = 1.34139$)
2Θ range for data collection/ $^\circ$	6.516 to 114.454
Index ranges	$-29 \leq h \leq 29$, $-15 \leq k \leq 15$, $-16 \leq l \leq 21$
Reflections collected	71816
Independent reflections	5159 [$R_{\text{int}} = 0.1416$, $R_{\text{sigma}} = 0.0759$]
Data/restraints/parameters	5159/241/419
Goodness-of-fit on F^2	1.042
Final R indexes [$I \geq 2\sigma(I)$]	$R_1 = 0.0667$, $wR_2 = 0.1613$
Final R indexes [all data]	$R_1 = 0.1018$, $wR_2 = 0.1835$
Largest diff. peak/hole / $e \text{ \AA}^{-3}$	0.70/-0.44

(2) Crystal data and structure refinement for compound 3aa

Colorless crystals of product **3aa** (CCDC-2303114) suitable for X-ray crystallographic analysis were obtained by recrystallization from CHCl₃/acetone (via evaporation). The ORTEP drawing of compound **3aa** is shown in Figure S2.

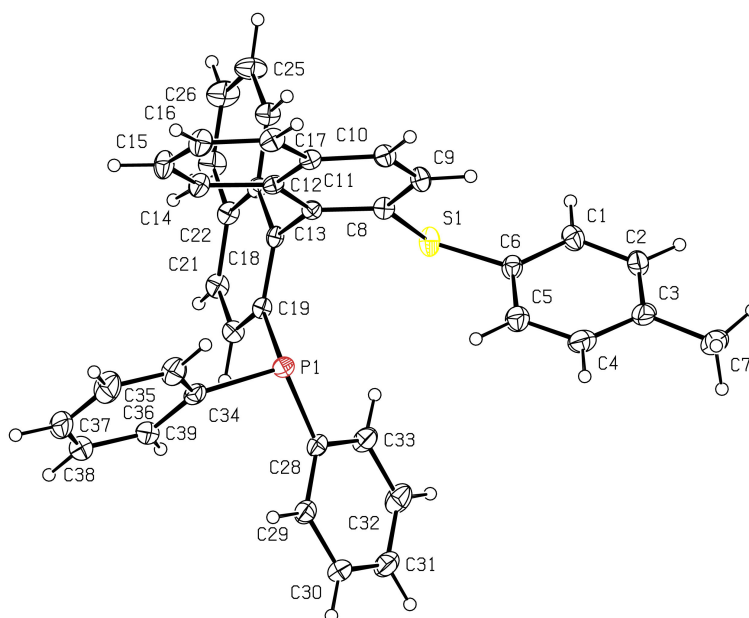


Figure S2. ORTEP illustration of compound **3aa** with thermal ellipsoids drawn at 50% probability level.

Table S3. Crystal data and structure refinement for compound **3aa**

Identification code	mo_230915_HYH_P1_0m
Empirical formula	C ₃₉ H ₂₉ PS
Formula weight	560.65
Temperature/K	170.00
Crystal system	triclinic
Space group	P1
a/Å	11.0390 (6)
b/Å	15.1879 (8)
c/Å	16.0708 (7)
α /°	112.211 (2)
β /°	104.135 (2)

$\gamma / ^\circ$	103.663(2)
Volume/ $\text{\AA}^3$	2251.2(2)
Z	3
$\rho_{\text{calc}}/\text{g}/\text{cm}^3$	1.241
μ / mm^{-1}	0.188
F(000)	882.0
Crystal size/ mm^3	$0.18 \times 0.16 \times 0.07$
Radiation	MoK α ($\lambda = 0.71073$)
2Θ range for data collection/ $^\circ$	4.088 to 55.106
Index ranges	$-14 \leq h \leq 14$, $-19 \leq k \leq 19$, $-20 \leq l \leq 20$
Reflections collected	109565
Independent reflections	20597 [$R_{\text{int}} = 0.0626$, $R_{\text{sigma}} = 0.0453$]
Data/restraints/parameters	20597/3/1111
Goodness-of-fit on F^2	1.080
Final R indexes [$I \geq 2\sigma(I)$]	$R_1 = 0.0470$, $wR_2 = 0.1012$
Final R indexes [all data]	$R_1 = 0.0591$, $wR_2 = 0.1094$
Largest diff. peak/hole / $\text{e \AA}^{-3}$	0.56/-0.24
Flack parameter	0.010(18)

(3) Crystal data and structure refinement for compound 3ae

Colourless crystals of product **3ae** (CCDC-2306477) suitable for X-ray crystallographic analysis were obtained by recrystallization from CHCl₃/acetone (via evaporation). The ORTEP drawing of compound **3ae** is shown in Figure S3.

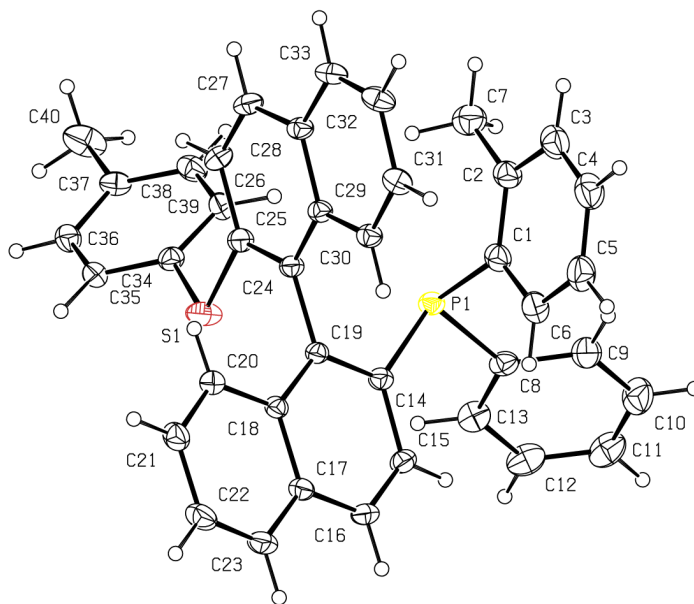


Figure S3. ORTEP illustration of compound 3ae with thermal ellipsoids drawn at 50% probability level.

Table S4. Crystal data and structure refinement for compound 3ae

Table 1 Crystal data and structure refinement for mo_231106_SJH_HP1_0m.	
Identification code	mo_231106_SJH_HP1_0m
Empirical formula	C ₄₀ H ₃₁ PS
Formula weight	574.68
Temperature/K	170.00
Crystal system	monoclinic
Space group	P2 ₁
a/Å	8.1621(3)
b/Å	11.2980(4)
c/Å	16.5498(6)

$\alpha / ^\circ$	90
$\beta / ^\circ$	94.610(2)
$\gamma / ^\circ$	90
Volume/ $\text{\AA}^3$	1521.21(10)
Z	2
$\rho_{\text{calc}}/\text{cm}^3$	1.255
μ/mm^{-1}	0.187
F(000)	604.0
Crystal size/ mm^3	$0.42 \times 0.19 \times 0.08$
Radiation	MoK α ($\lambda = 0.71073$)
2Θ range for data collection/ $^\circ$	4.37 to 55.08
Index ranges	$-10 \leq h \leq 10$, $-14 \leq k \leq 14$, $-21 \leq l \leq 21$
Reflections collected	48812
Independent reflections	7005 [$R_{\text{int}} = 0.0388$, $R_{\text{sigma}} = 0.0260$]
Data/restraints/parameters	7005/1/381
Goodness-of-fit on F^2	1.074
Final R indexes [$I \geq 2\sigma(I)$]	$R_1 = 0.0371$, $wR_2 = 0.0925$
Final R indexes [all data]	$R_1 = 0.0401$, $wR_2 = 0.0951$
Largest diff. peak/hole / $e \text{\AA}^{-3}$	0.34/-0.19
Flack parameter	0.001(17)

9. NMR Spectra and Chiral HPLC Charts

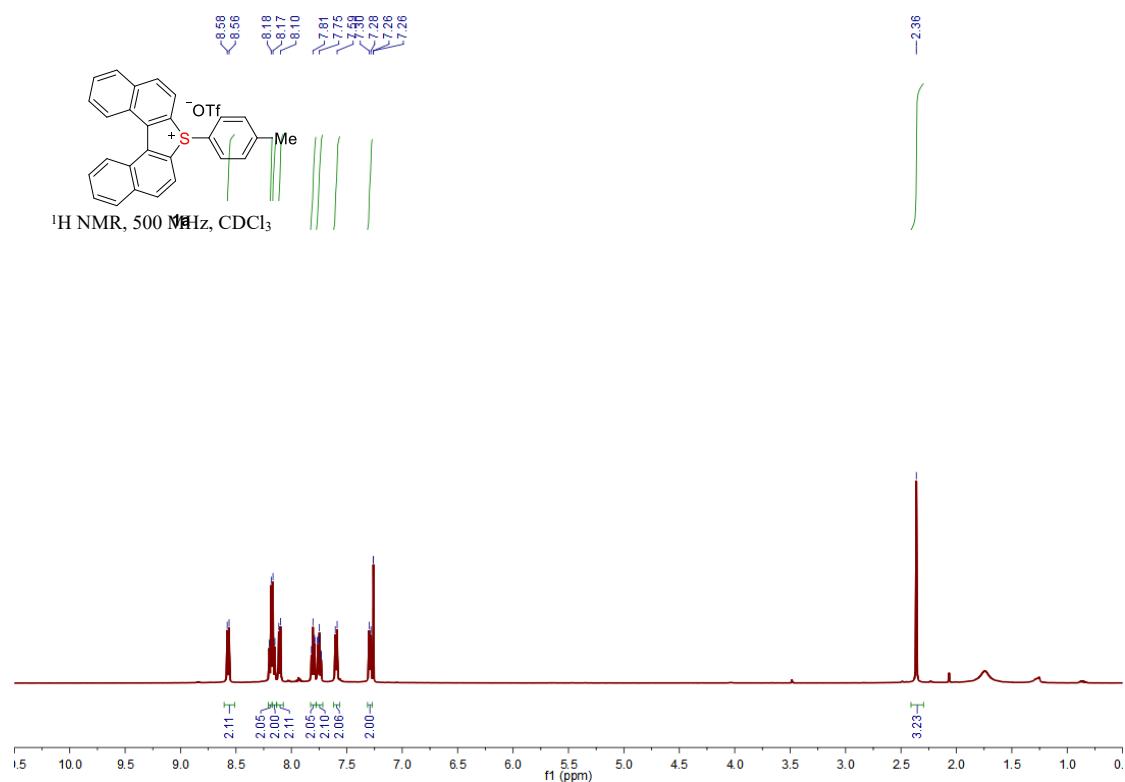


Figure S4. ¹H NMR spectrum of 1a (500 MHz, CDCl₃)

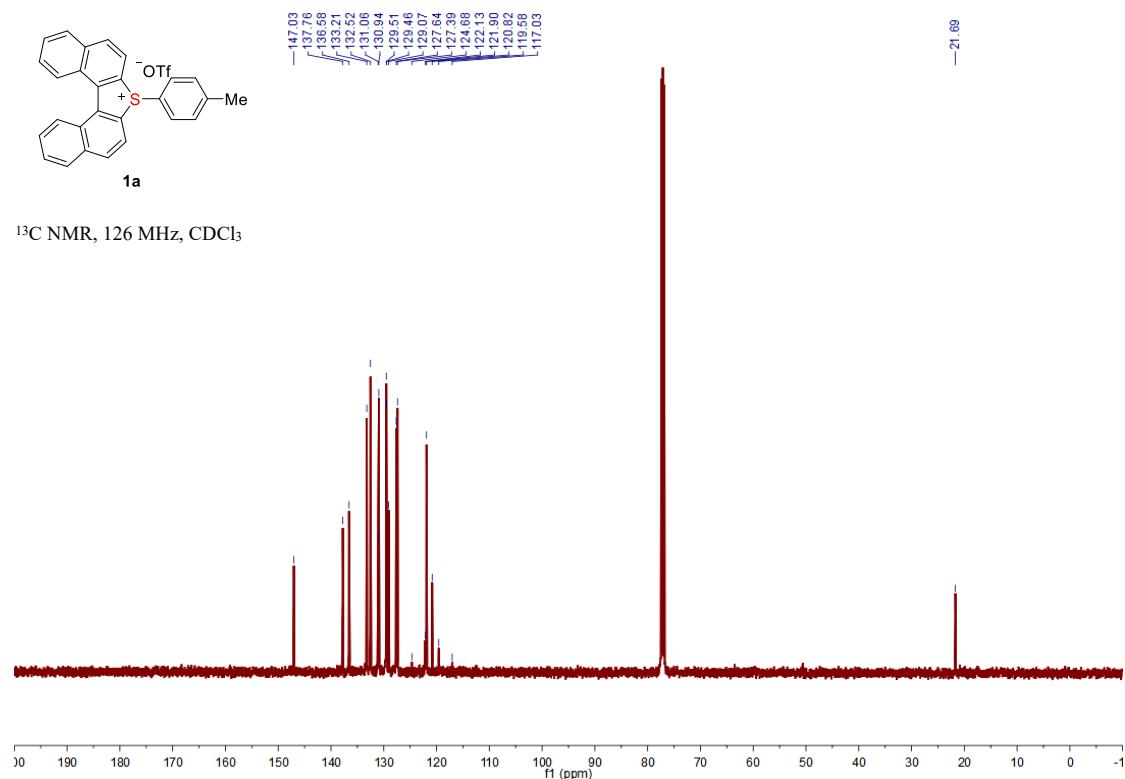


Figure S5. ¹³C NMR spectrum of 1a (126 MHz, CDCl₃)

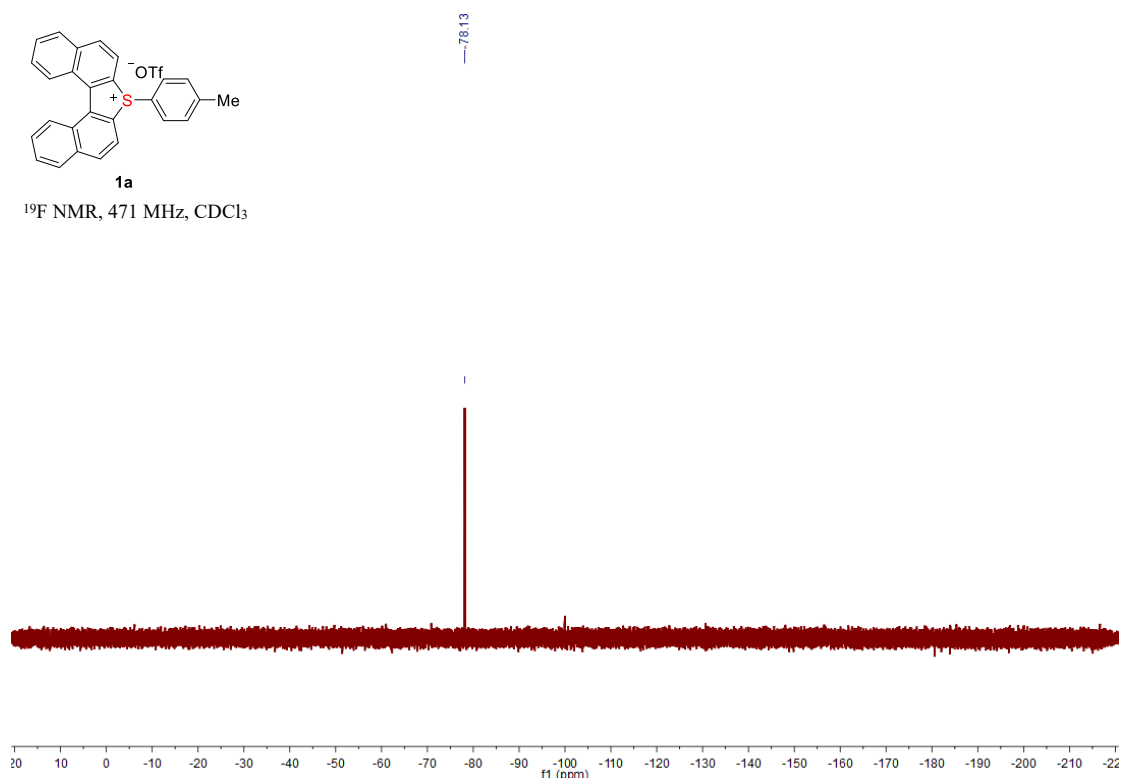


Figure S6. ^{19}F NMR spectrum of 1a (471 MHz, CDCl_3)

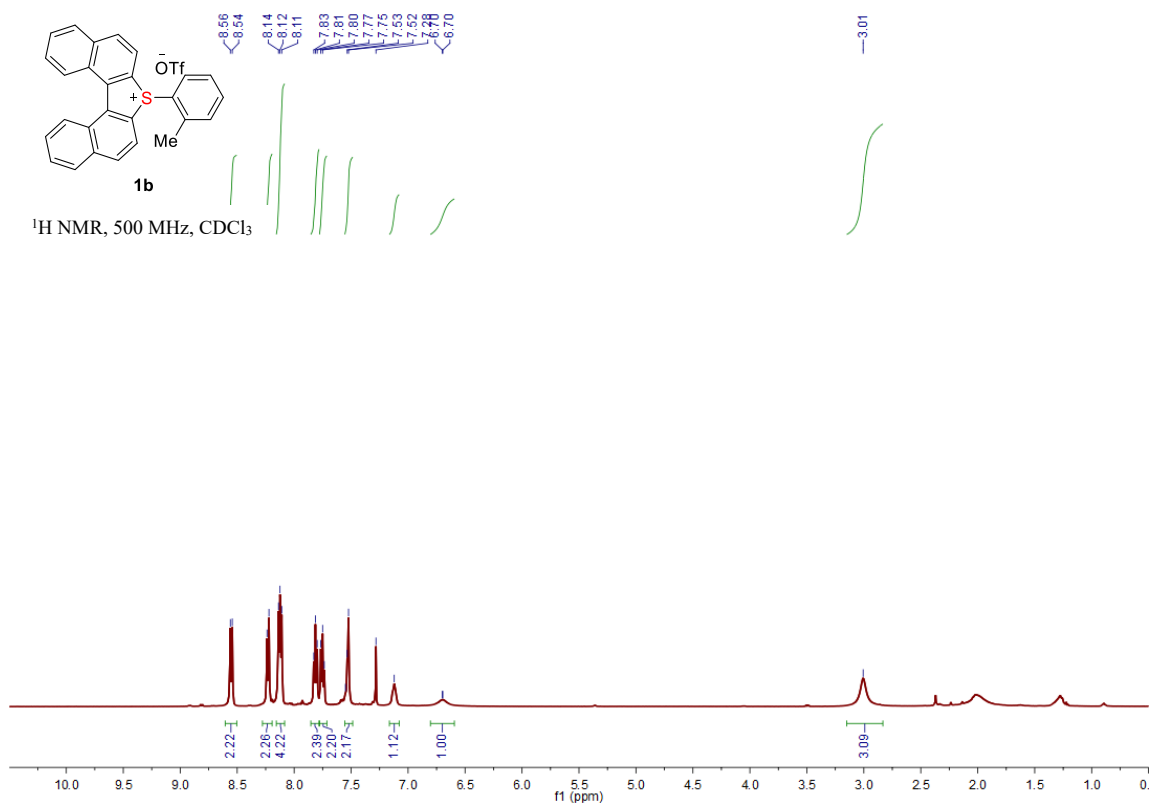


Figure S7. ^1H NMR spectrum of 1b (500 MHz, CDCl_3)

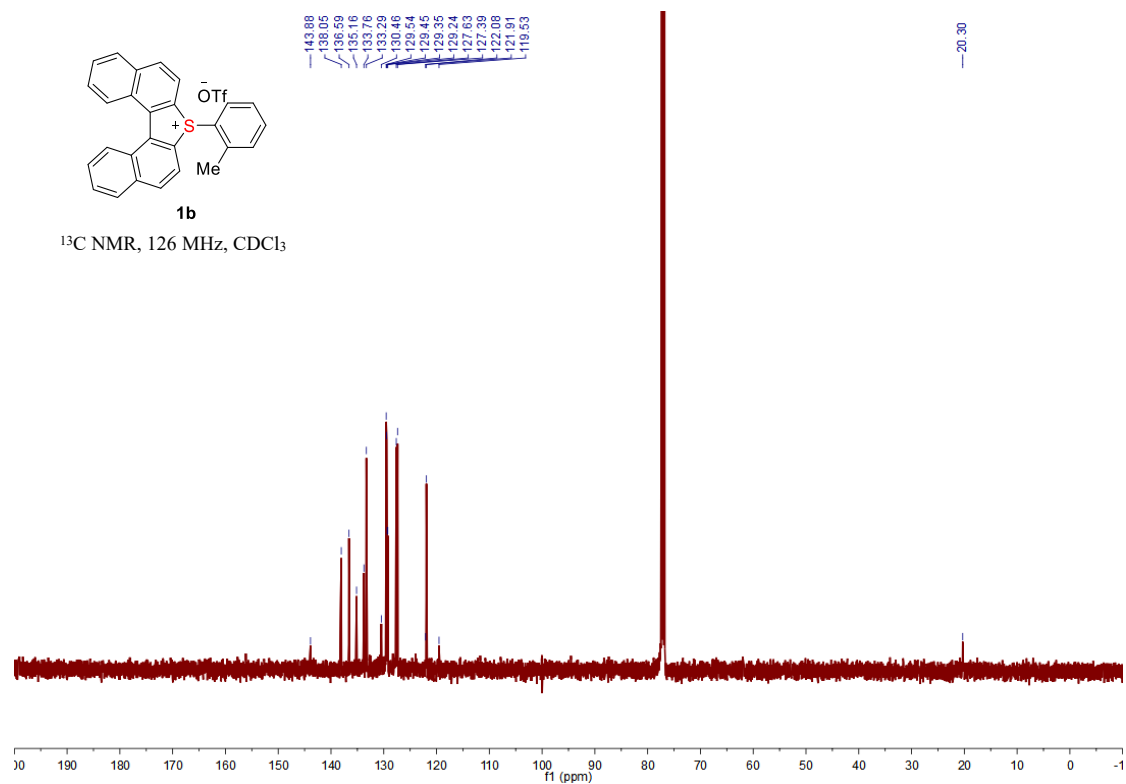


Figure S8. ^{13}C NMR spectrum of **1b** (126 MHz, CDCl_3)

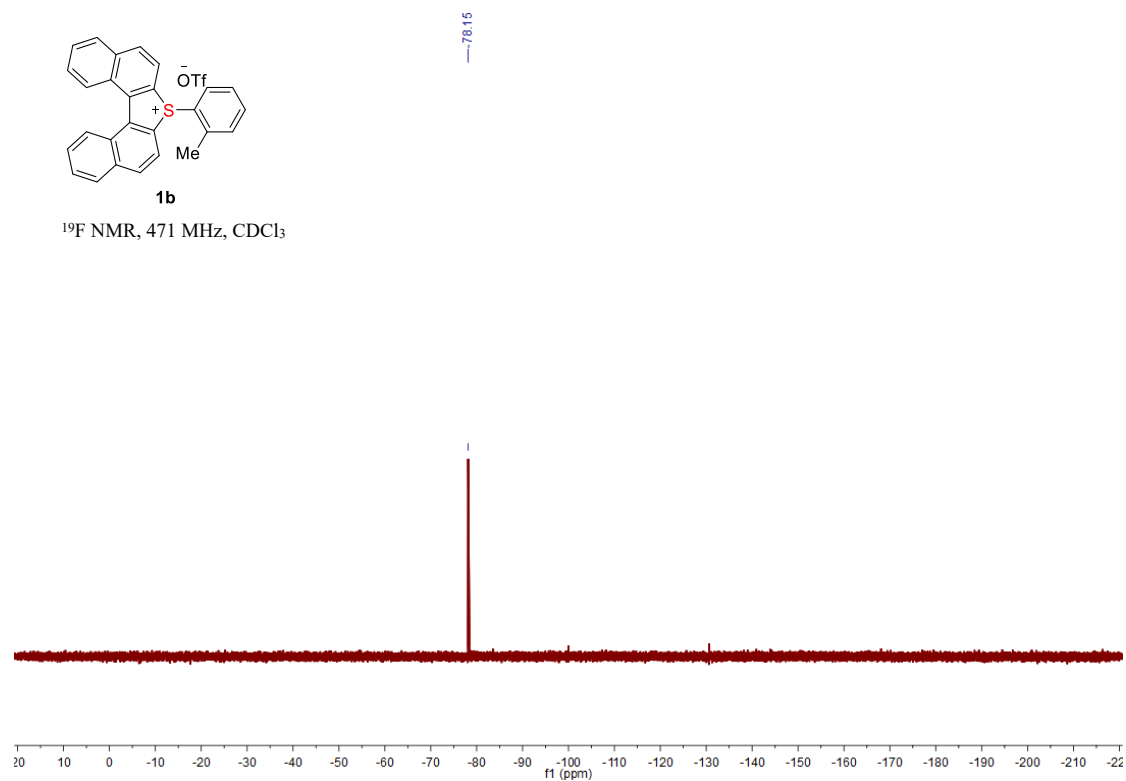


Figure S9. ^{19}F NMR spectrum of **1b** (471 MHz, CDCl_3)

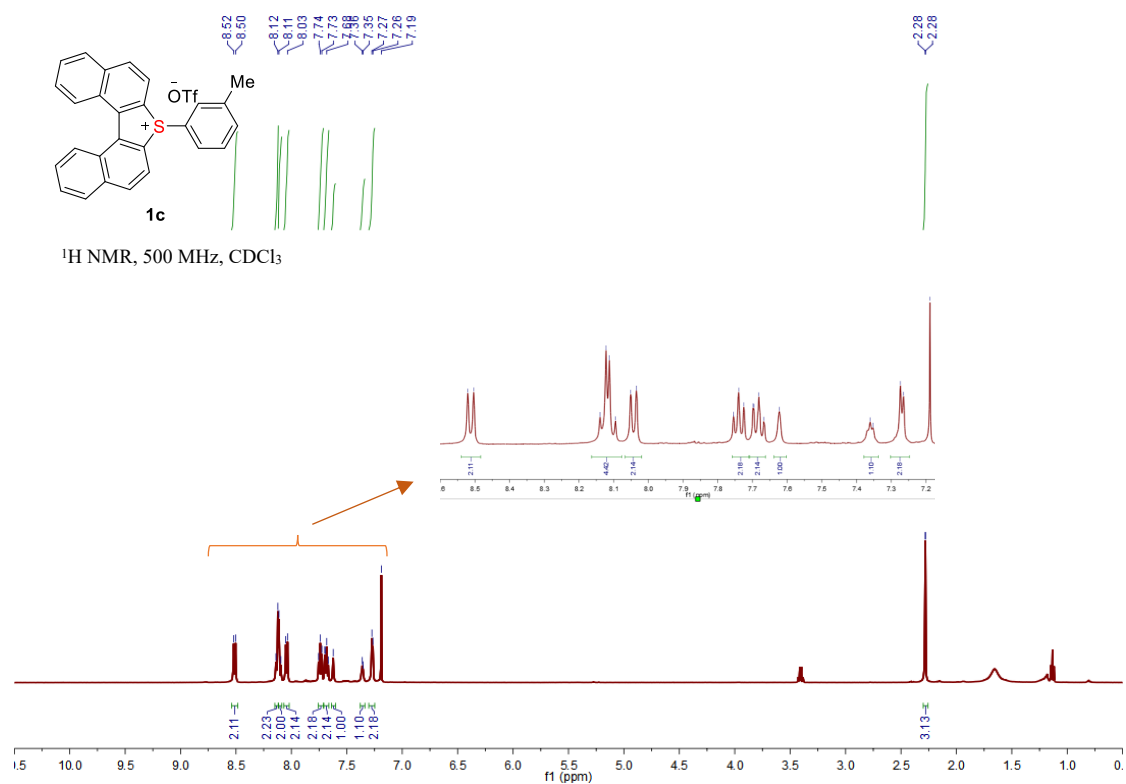


Figure S10. ^1H NMR spectrum of **1c** (500 MHz, CDCl_3)

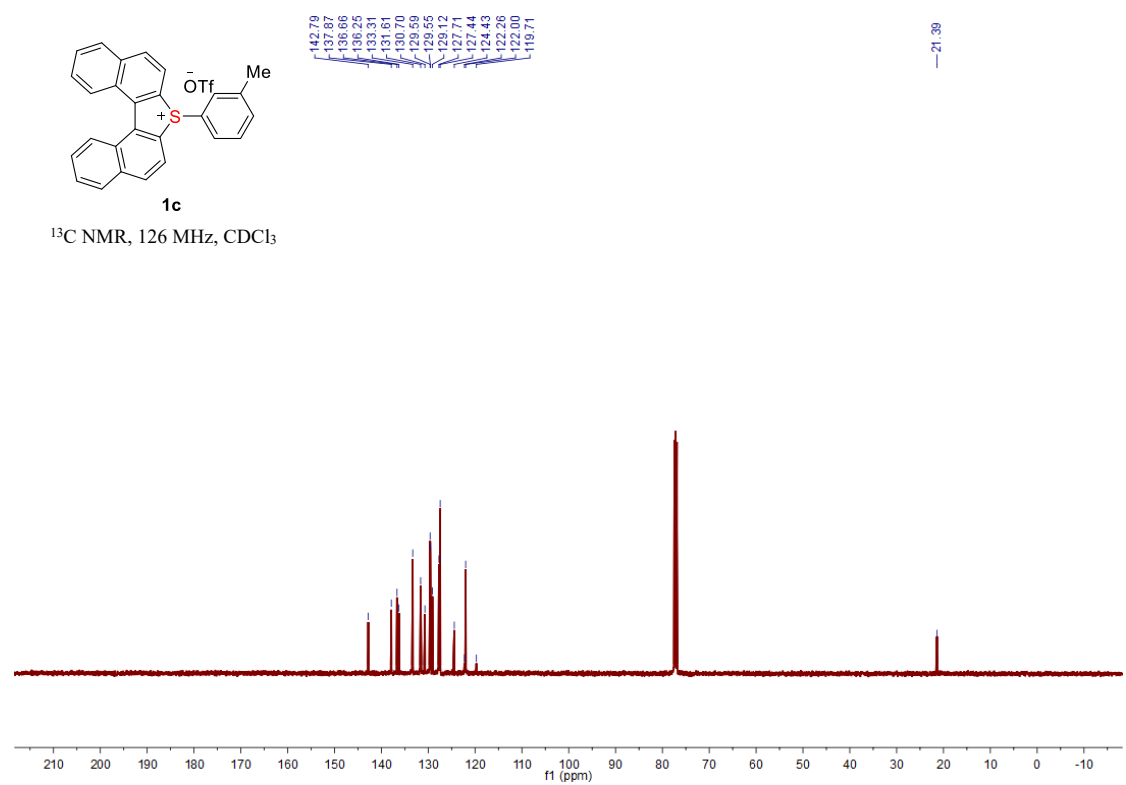


Figure S11. ^{13}C NMR spectrum of **1c** (126 MHz, CDCl_3)

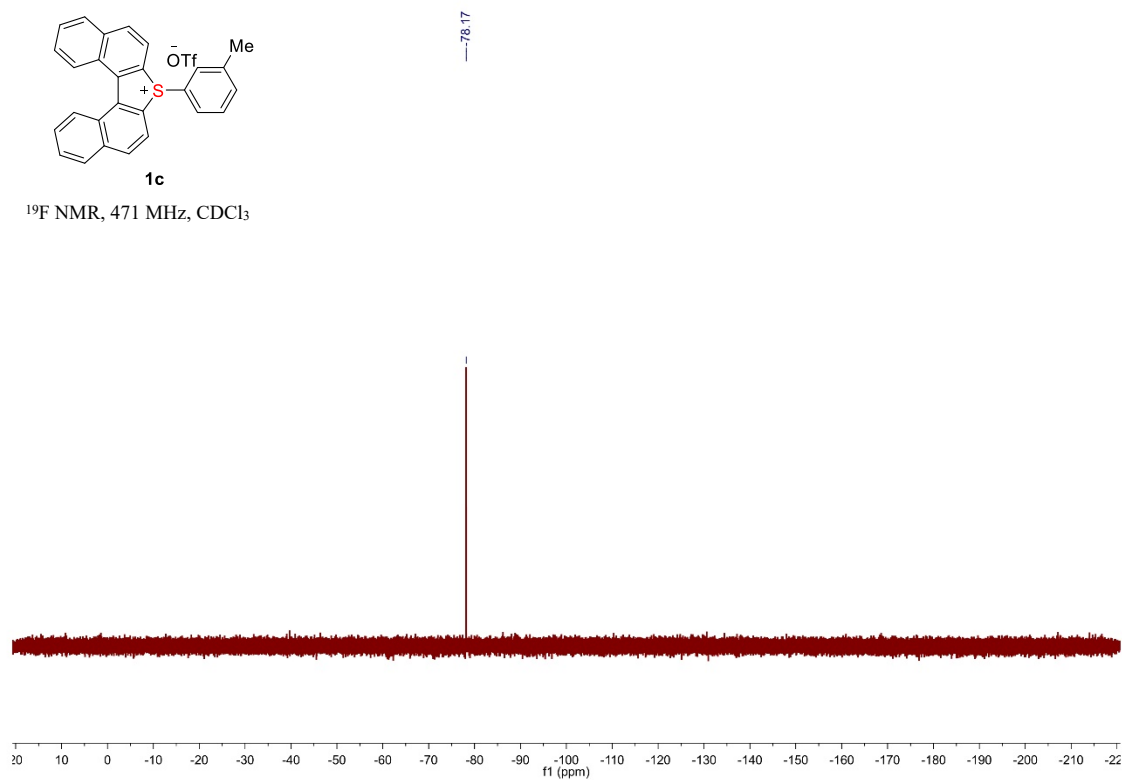


Figure S12. ^{19}F NMR spectrum of **1c** (471 MHz, CDCl_3)

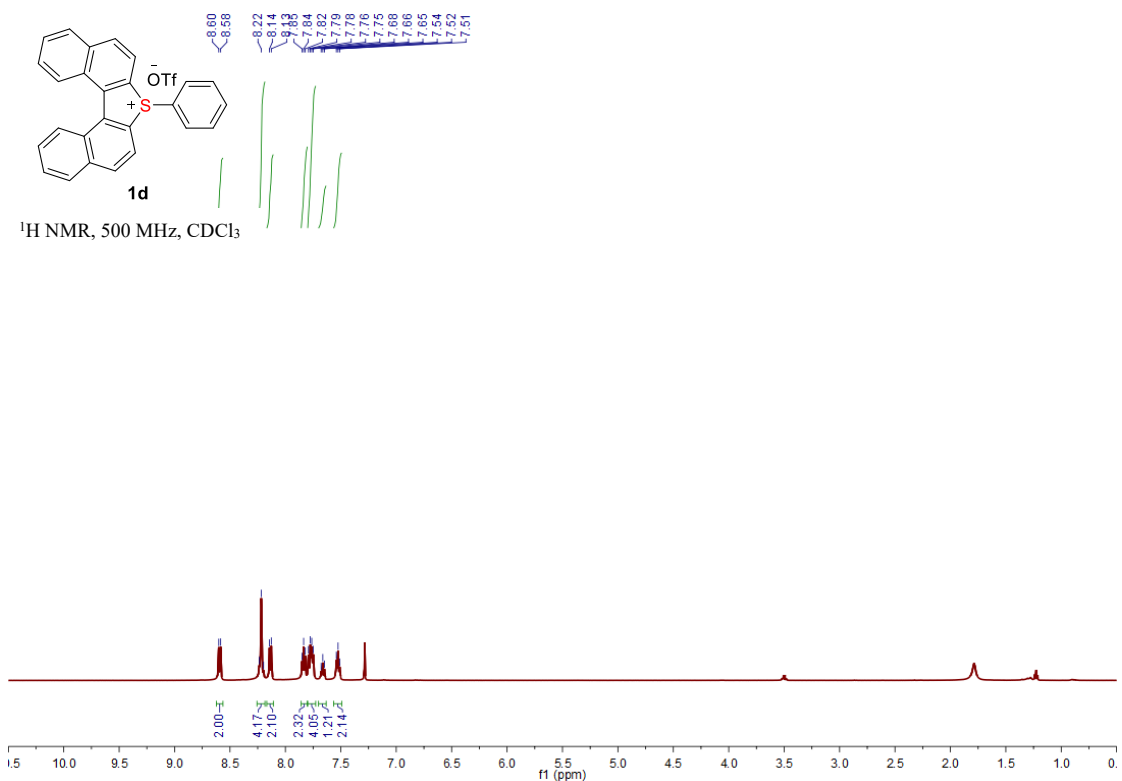


Figure S13. ^1H NMR spectrum of **1d** (500 MHz, CDCl_3)

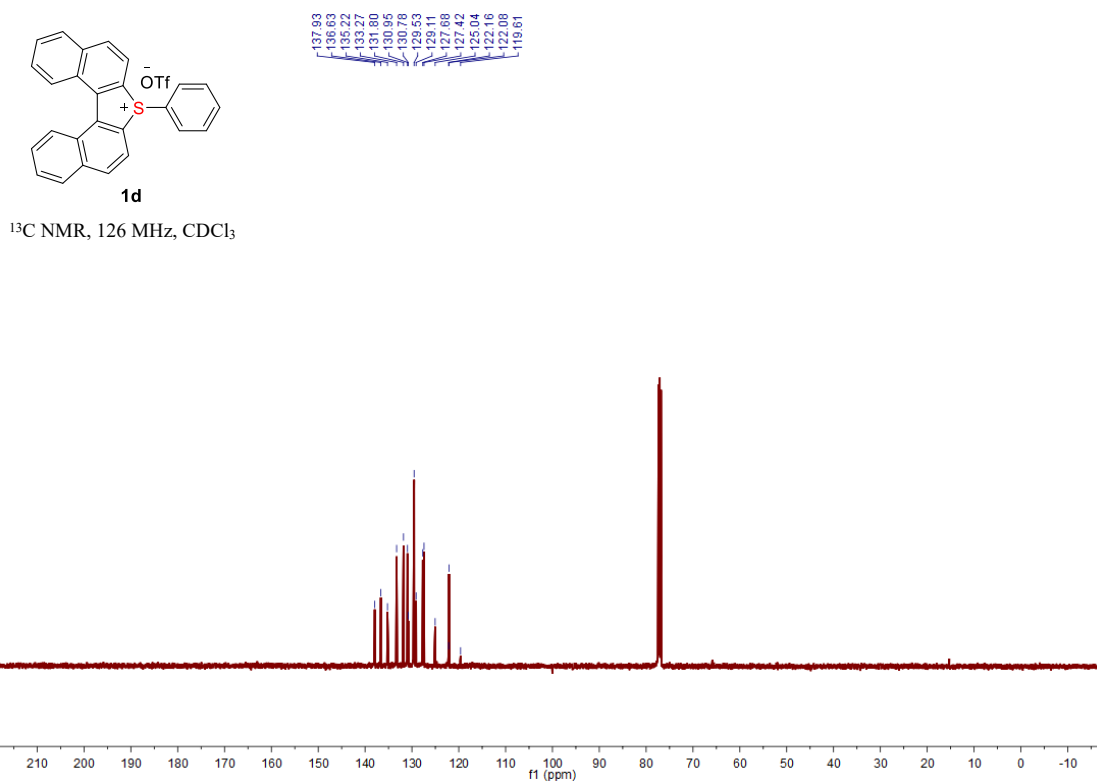


Figure S14. ¹³C NMR spectrum of **1d** (126 MHz, CDCl₃)

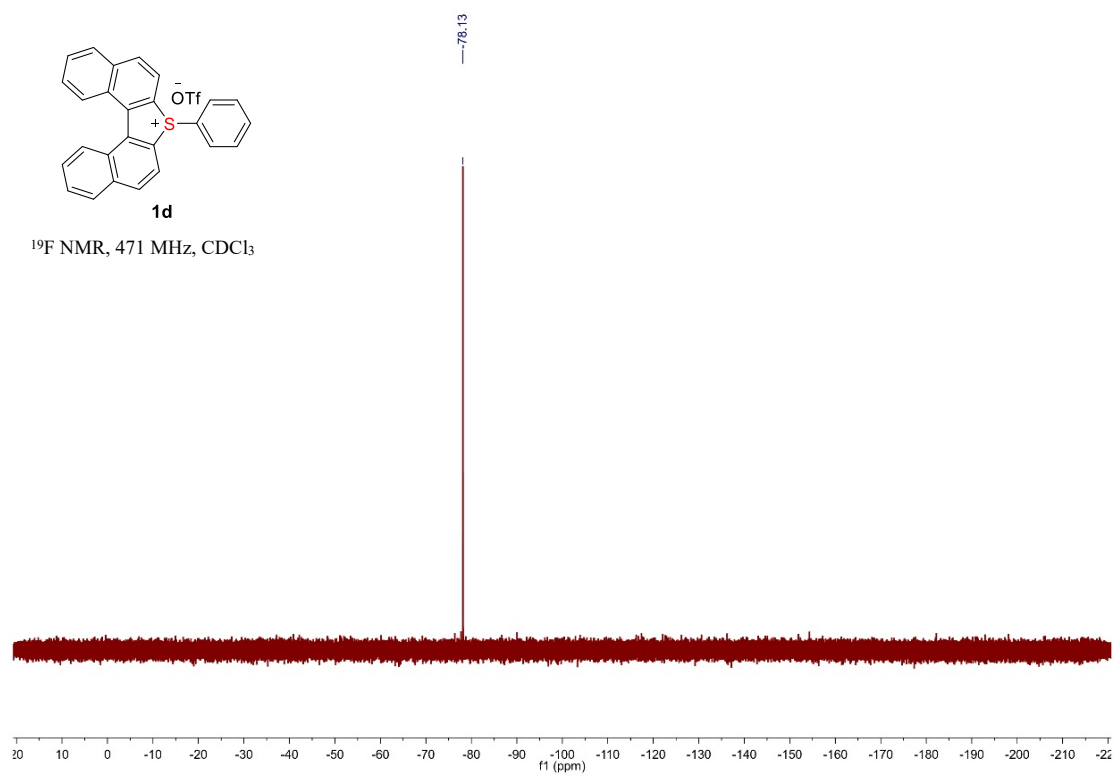


Figure S15. ¹⁹F NMR spectrum of **1d** (471 MHz, CDCl₃)

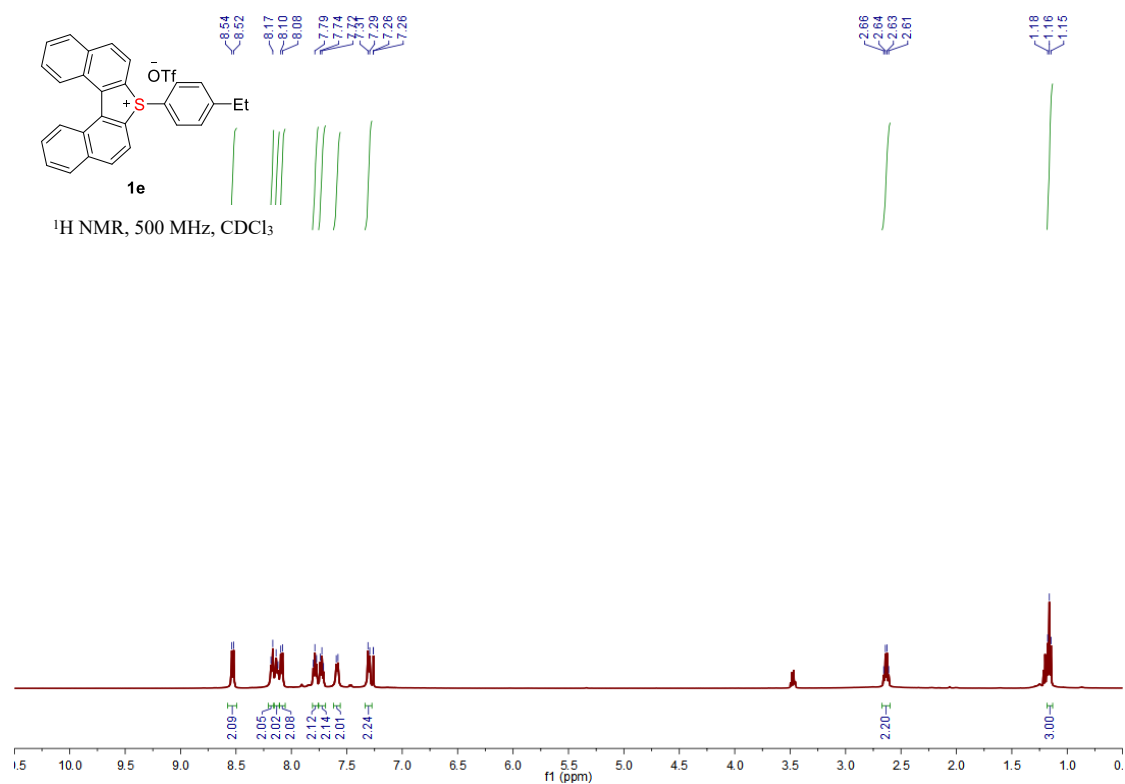


Figure S16. ¹H NMR spectrum of 1e (500 MHz, CDCl₃)

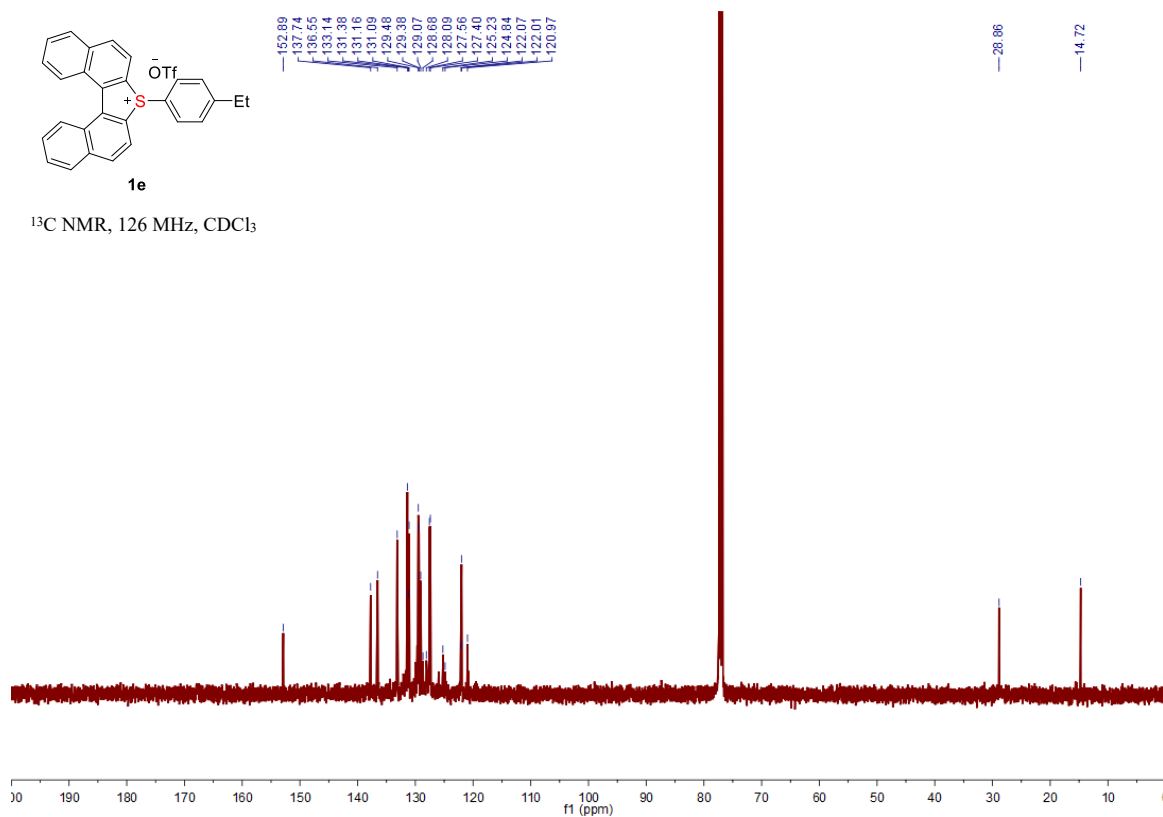


Figure S17. ¹³C NMR spectrum of 1e (126 MHz, CDCl₃)

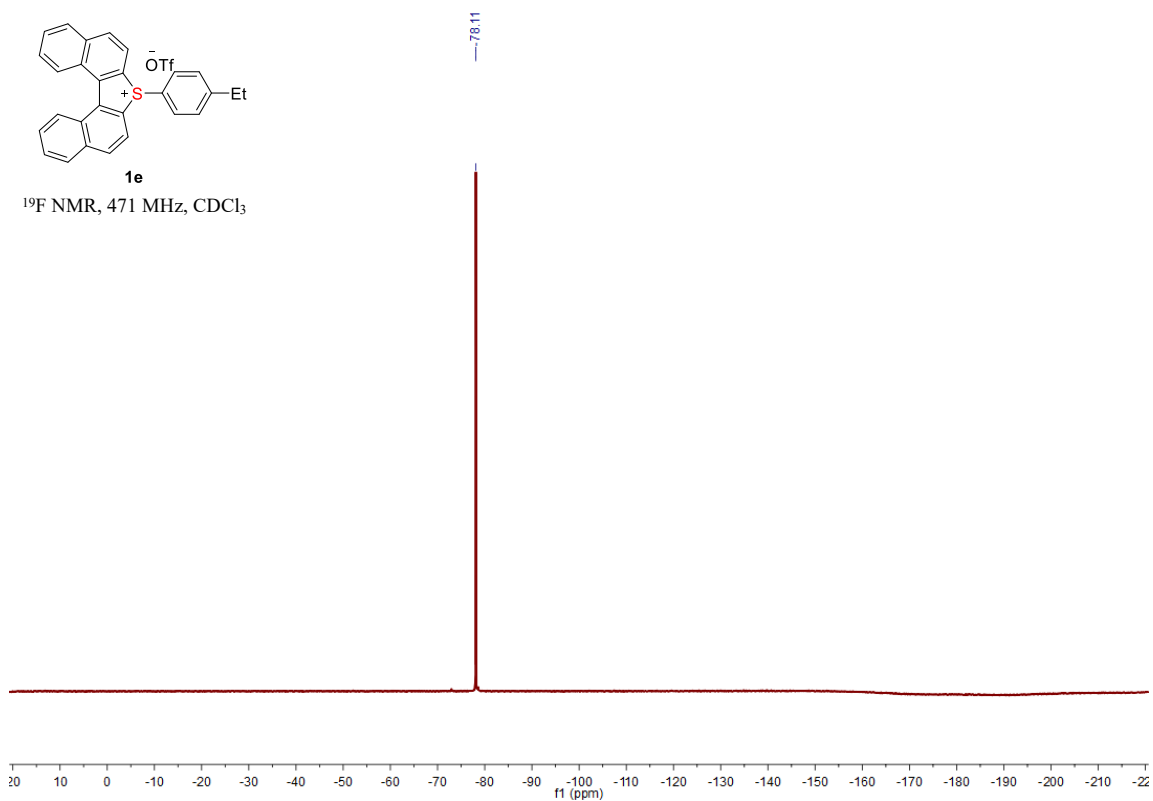


Figure S18. ¹⁹F NMR spectrum of **1e** (471 MHz, CDCl₃)

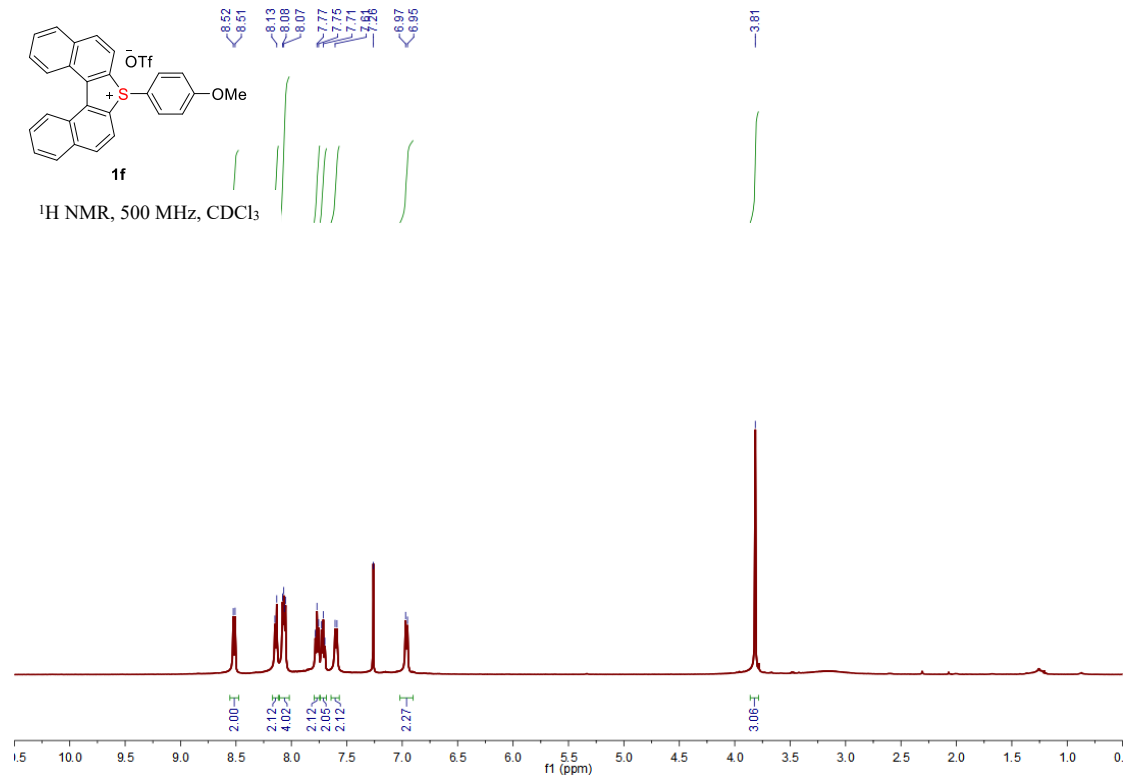


Figure S19. ¹H NMR spectrum of **1f** (500 MHz, CDCl₃)

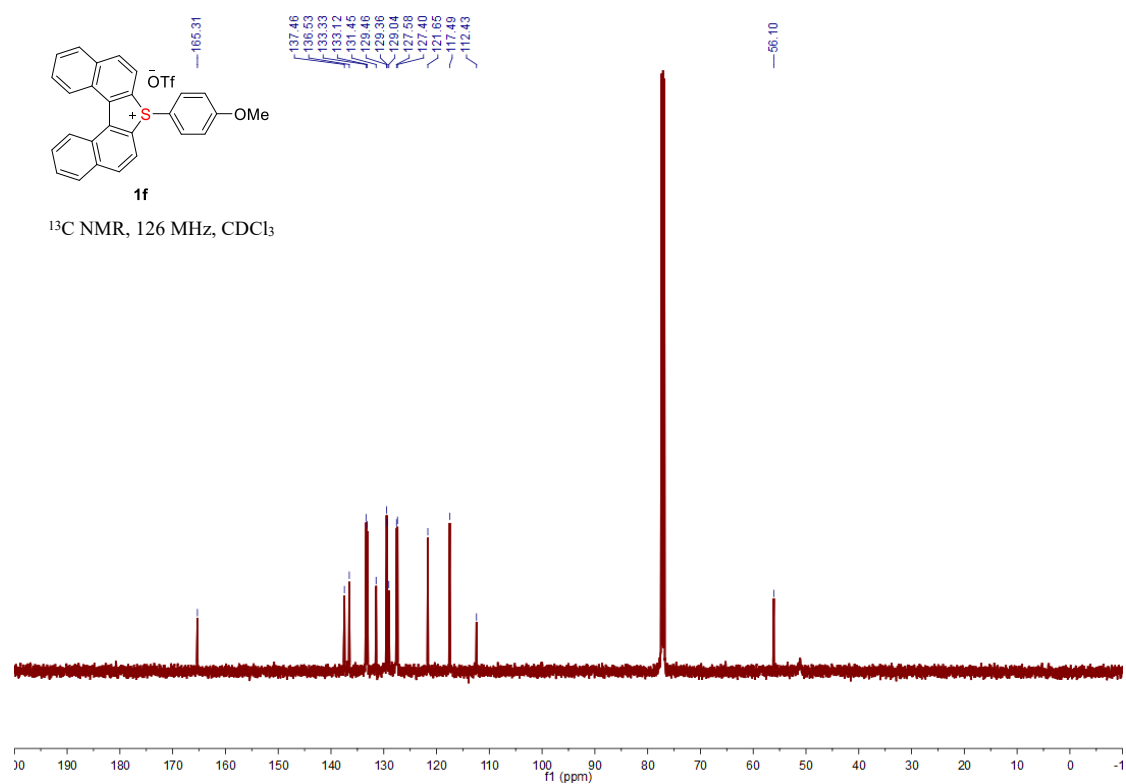


Figure S20. ^{13}C NMR spectrum of **1f** (126 MHz, CDCl_3)

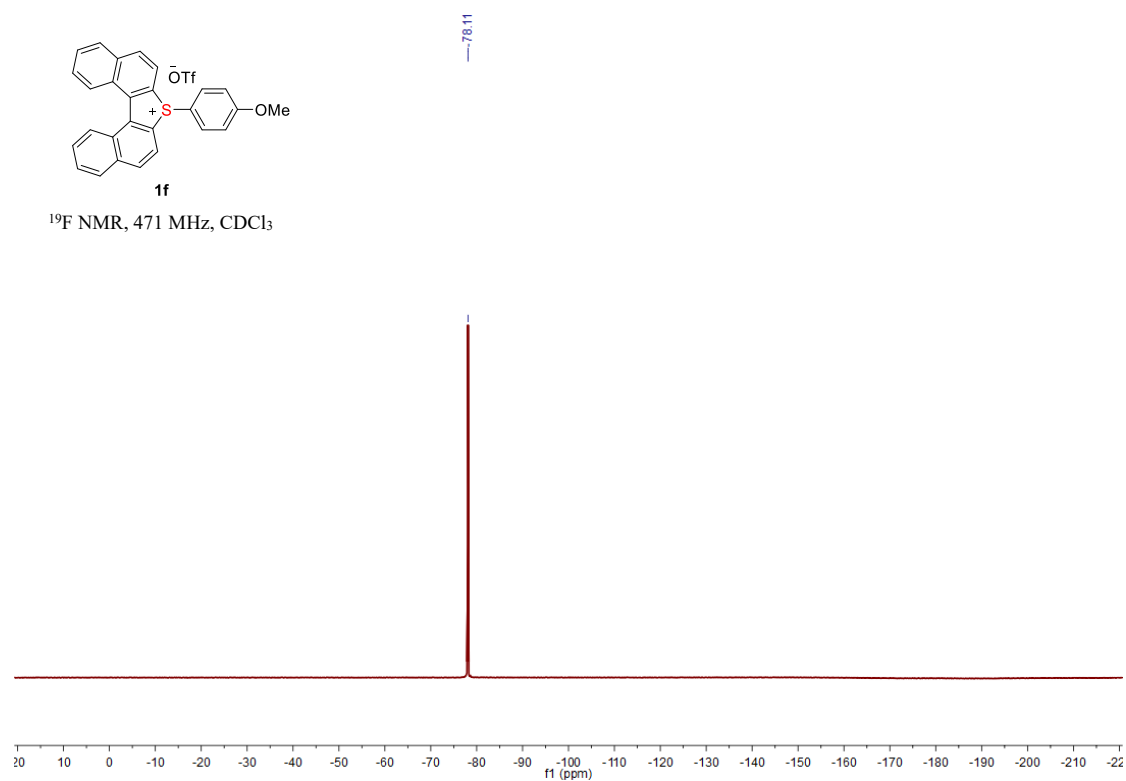


Figure S21. ^{19}F NMR spectrum of **1f** (471 MHz, CDCl_3)



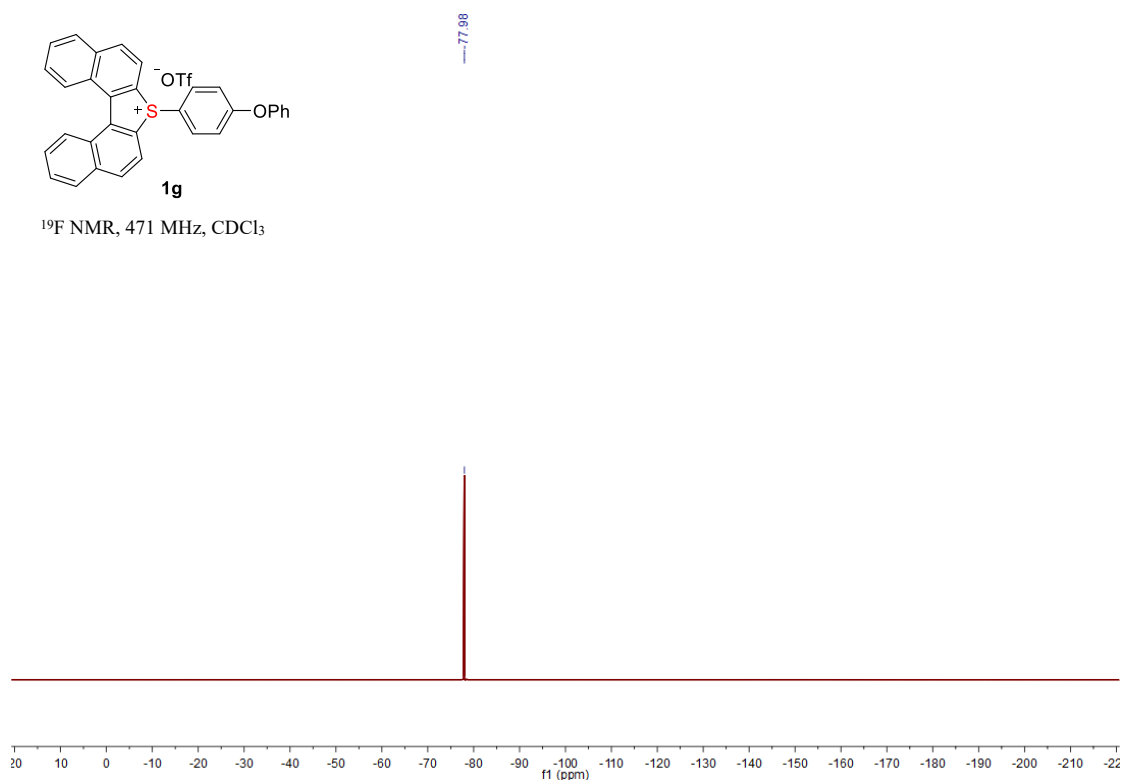


Figure S24. ^{19}F NMR spectrum of 1g (471 MHz, CDCl_3)

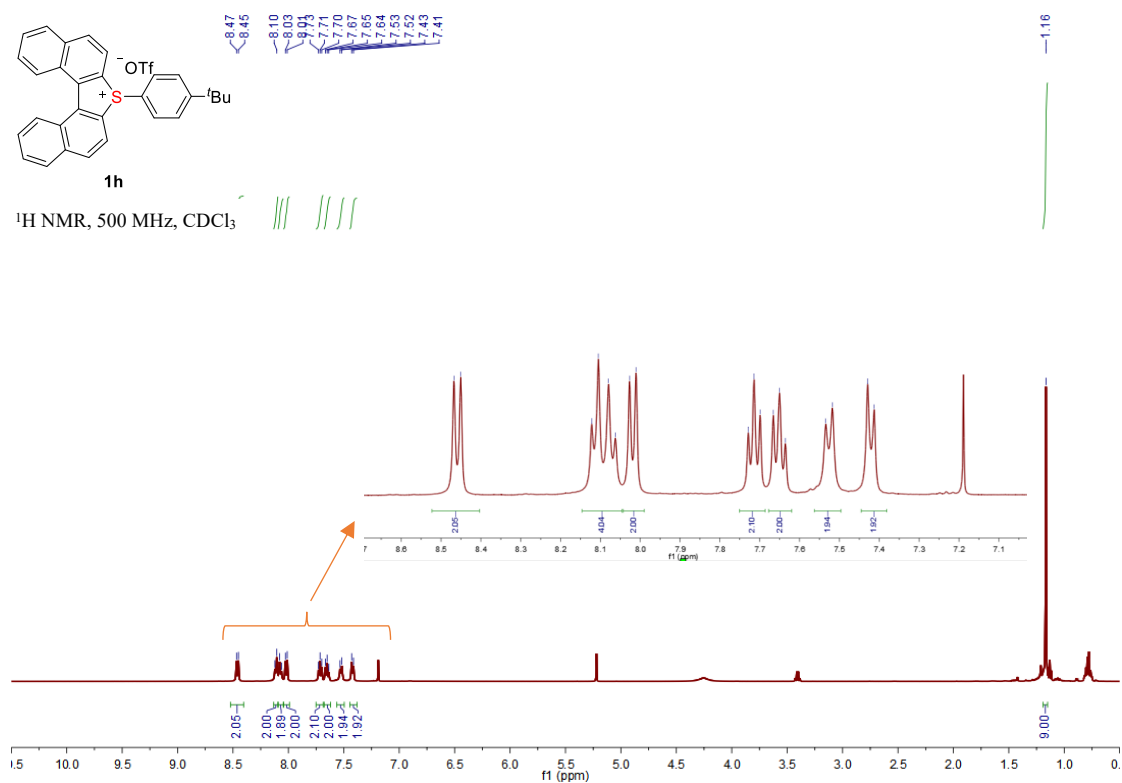


Figure S25. ^1H NMR spectrum of 1h (500 MHz, CDCl_3)

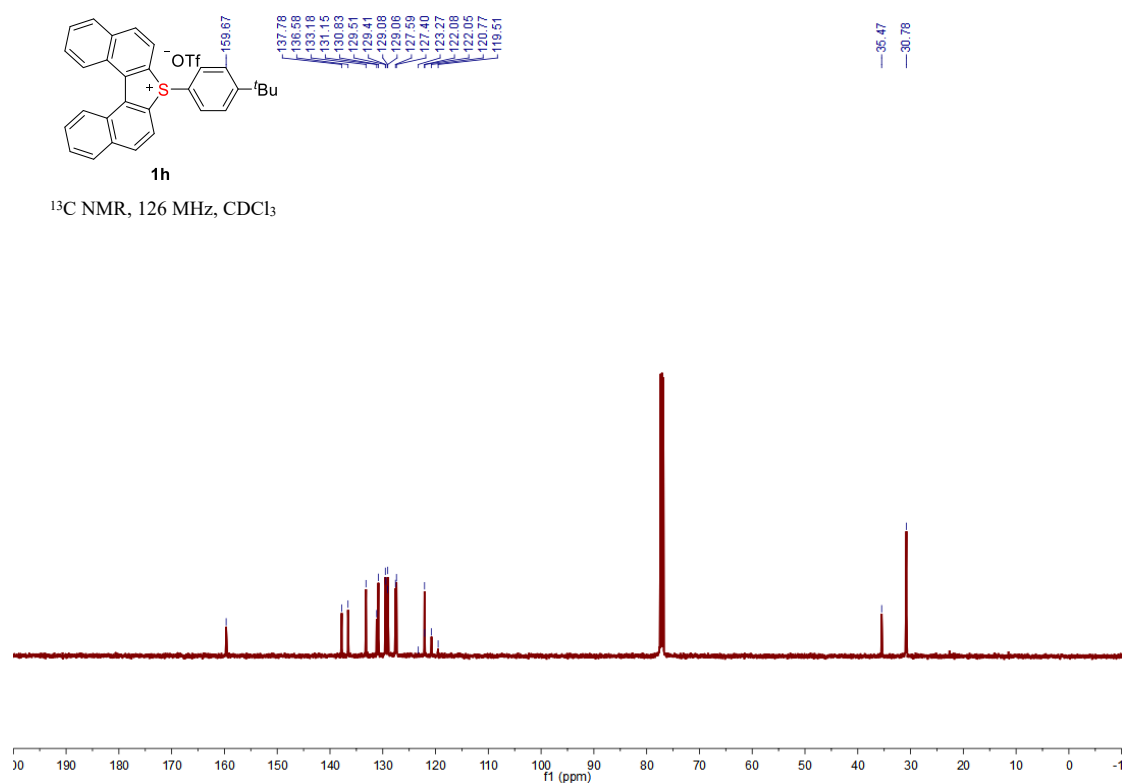


Figure S26. ^{13}C NMR spectrum of **1h** (126 MHz, CDCl_3)

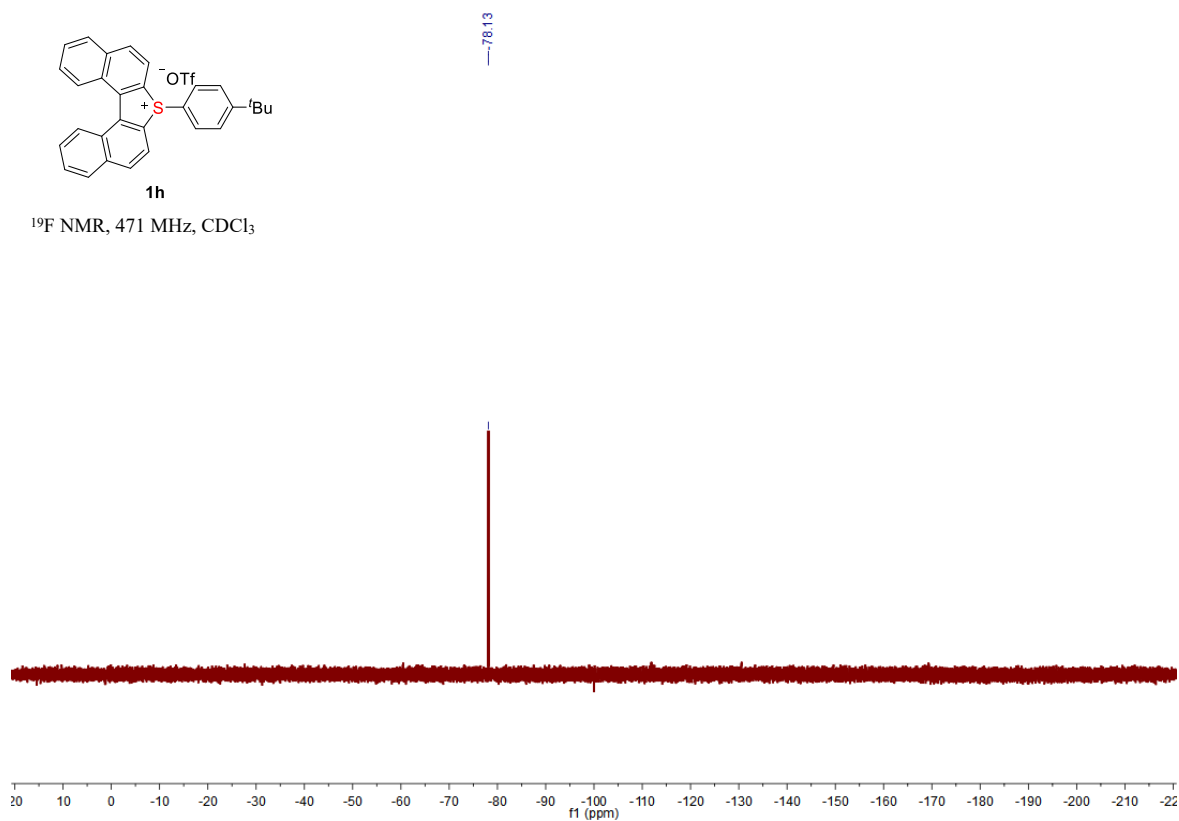


Figure S27. ^{19}F NMR spectrum of **1h** (471 MHz, CDCl_3)

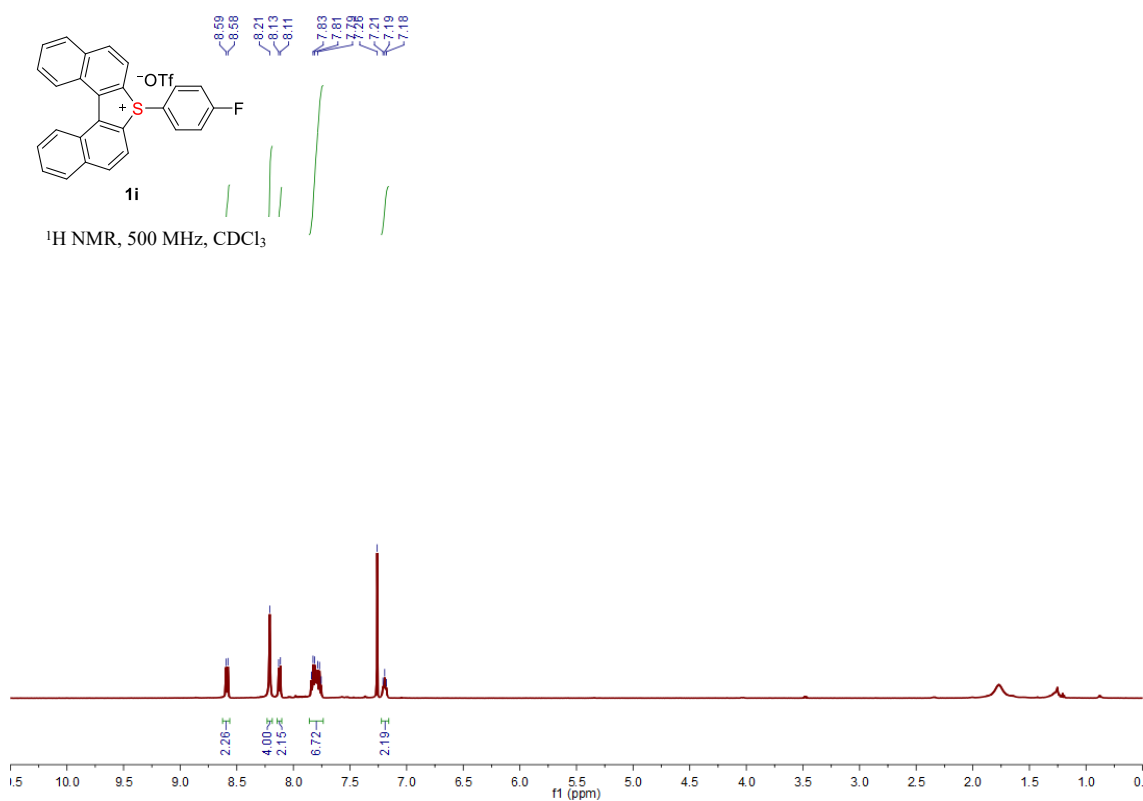


Figure S28. ^1H NMR spectrum of **1i (500 MHz, CDCl_3)**

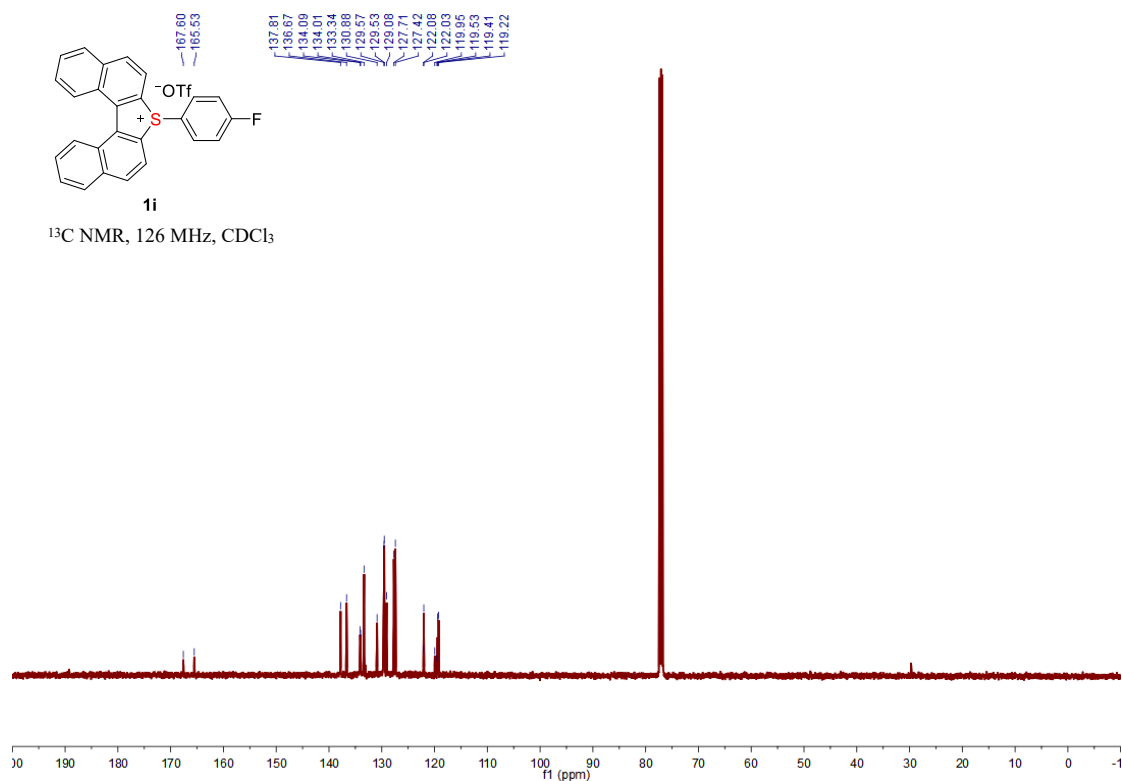


Figure S29. ^{13}C NMR spectrum of **1i (126 MHz, CDCl_3)**

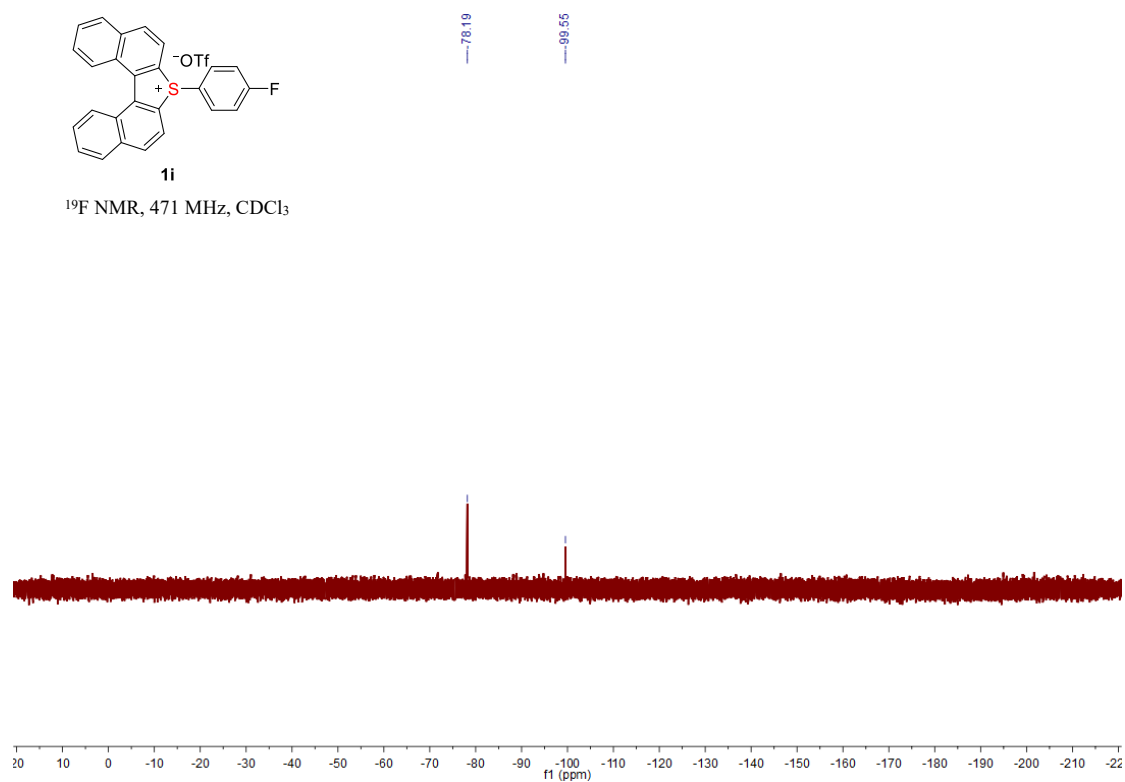


Figure S30. ^{19}F NMR spectrum of **1i** (471 MHz, CDCl_3)

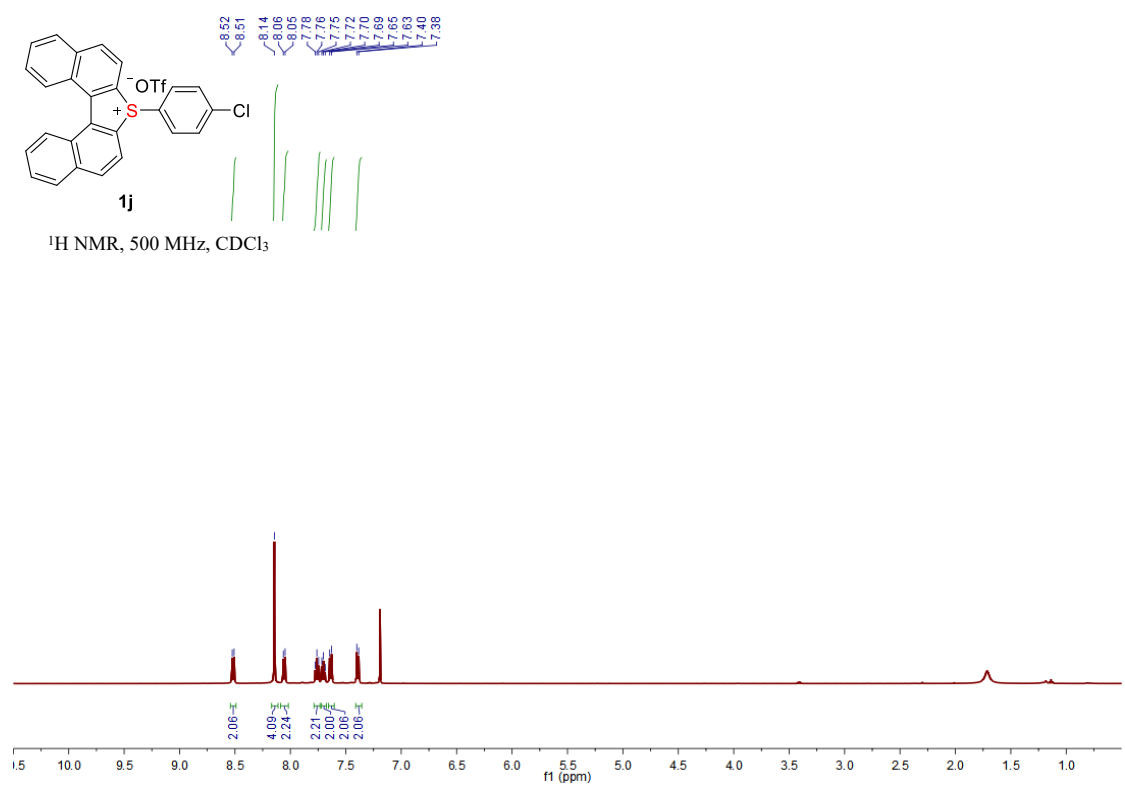


Figure S31. ^1H NMR spectrum of **1j** (500 MHz, CDCl_3)

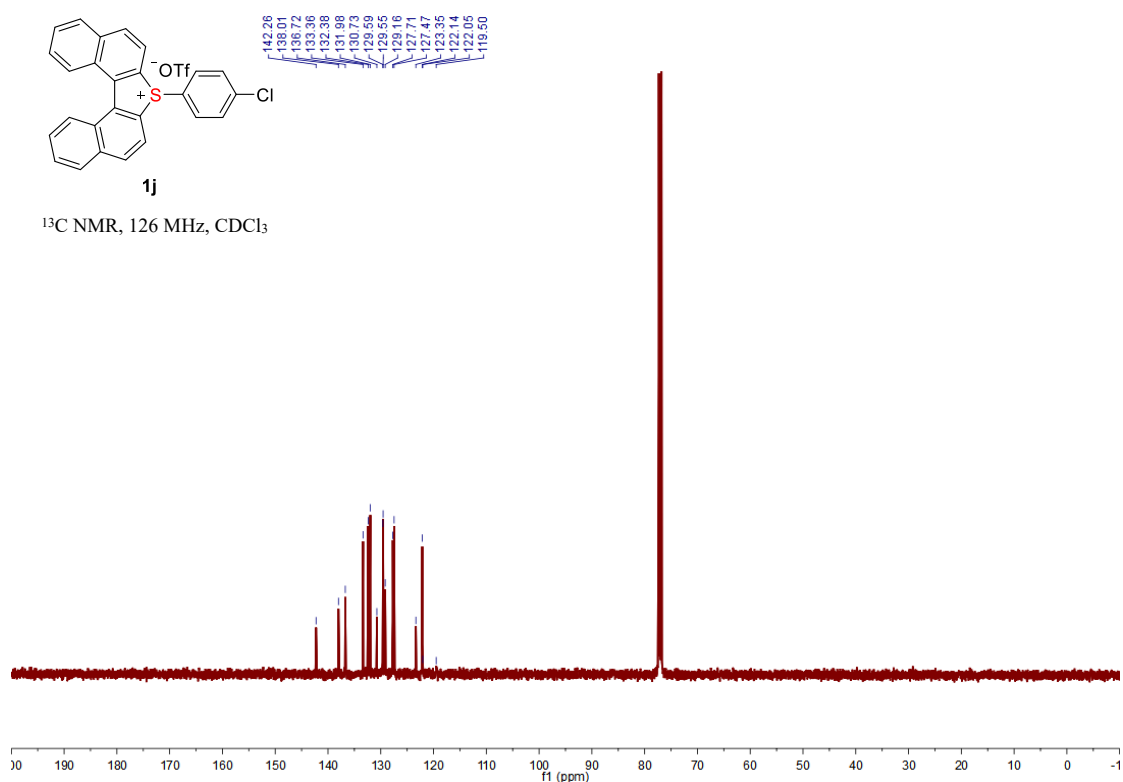


Figure S32. ^{13}C NMR spectrum of **1j** (126 MHz, CDCl_3)

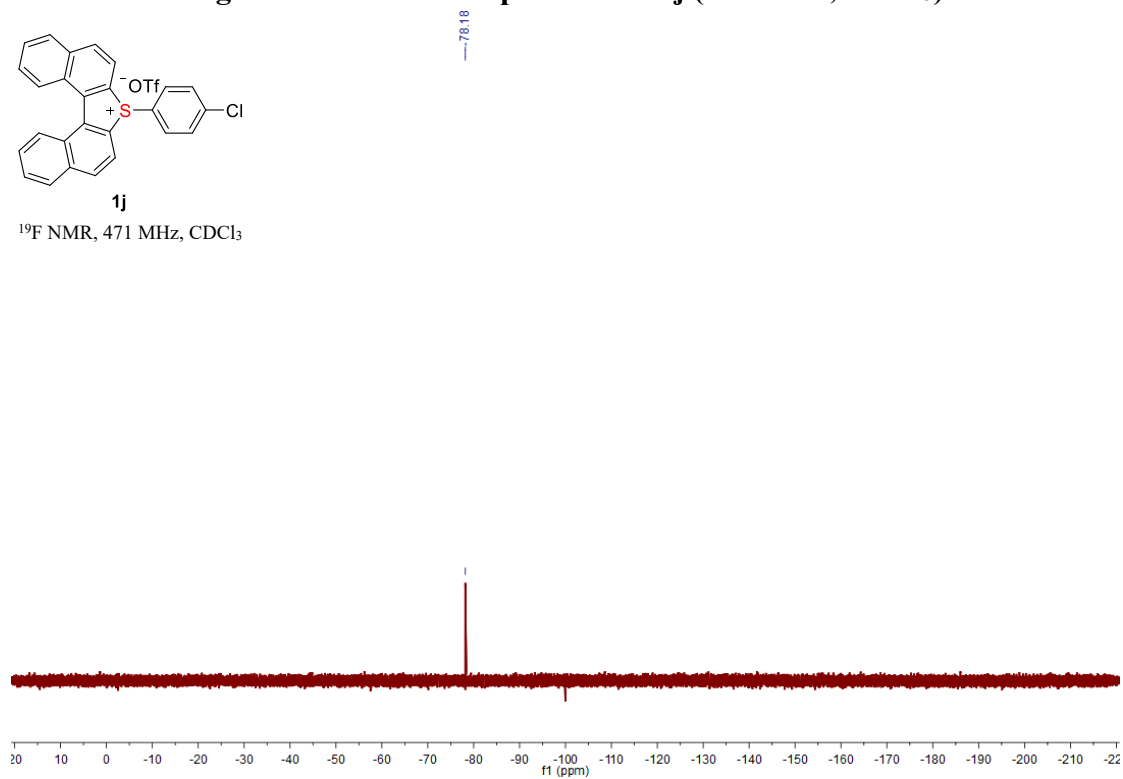


Figure S33. ^{19}F NMR spectrum of **1j** (471 MHz, CDCl_3)

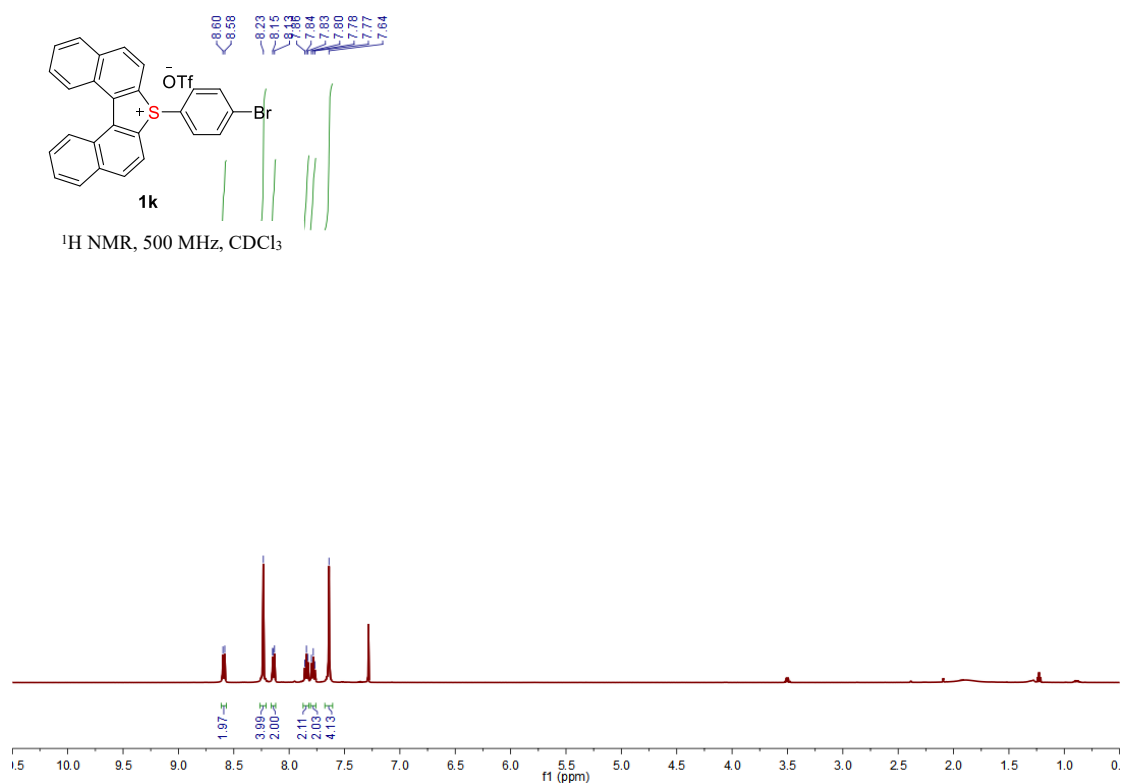


Figure S34. ^1H NMR spectrum of **1k** (500 MHz, CDCl_3)

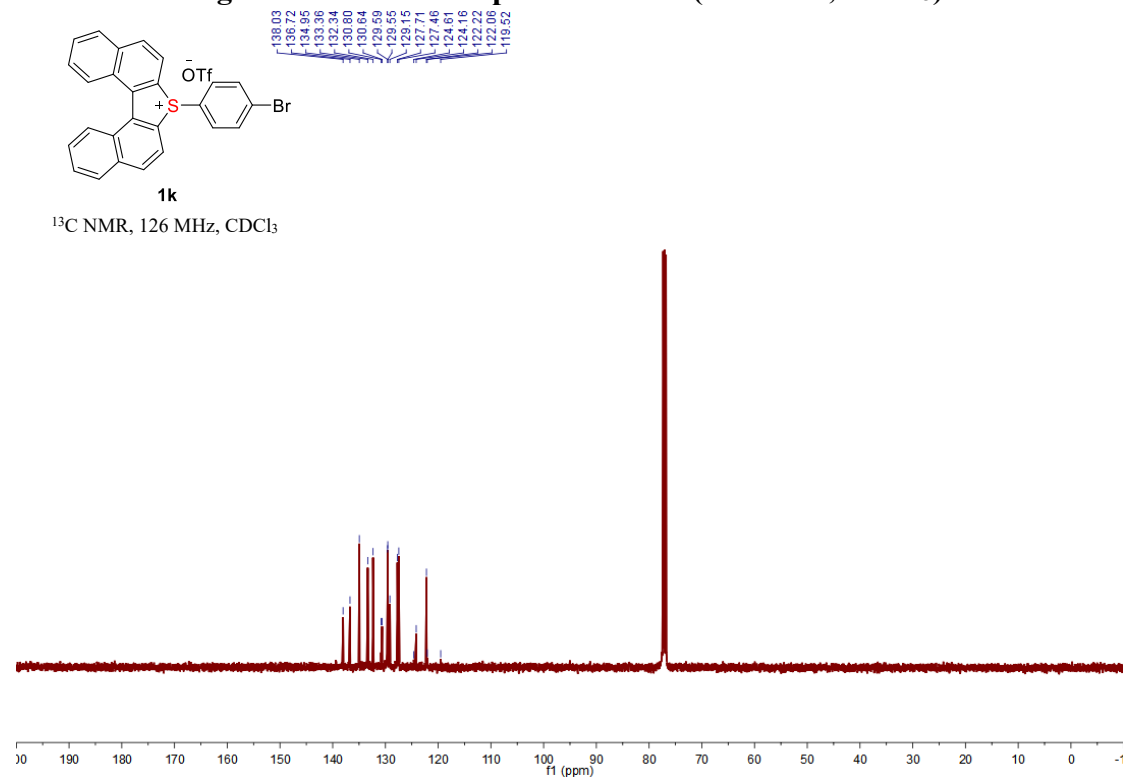


Figure S35. ^{13}C NMR spectrum of **1k** (126 MHz, CDCl_3)

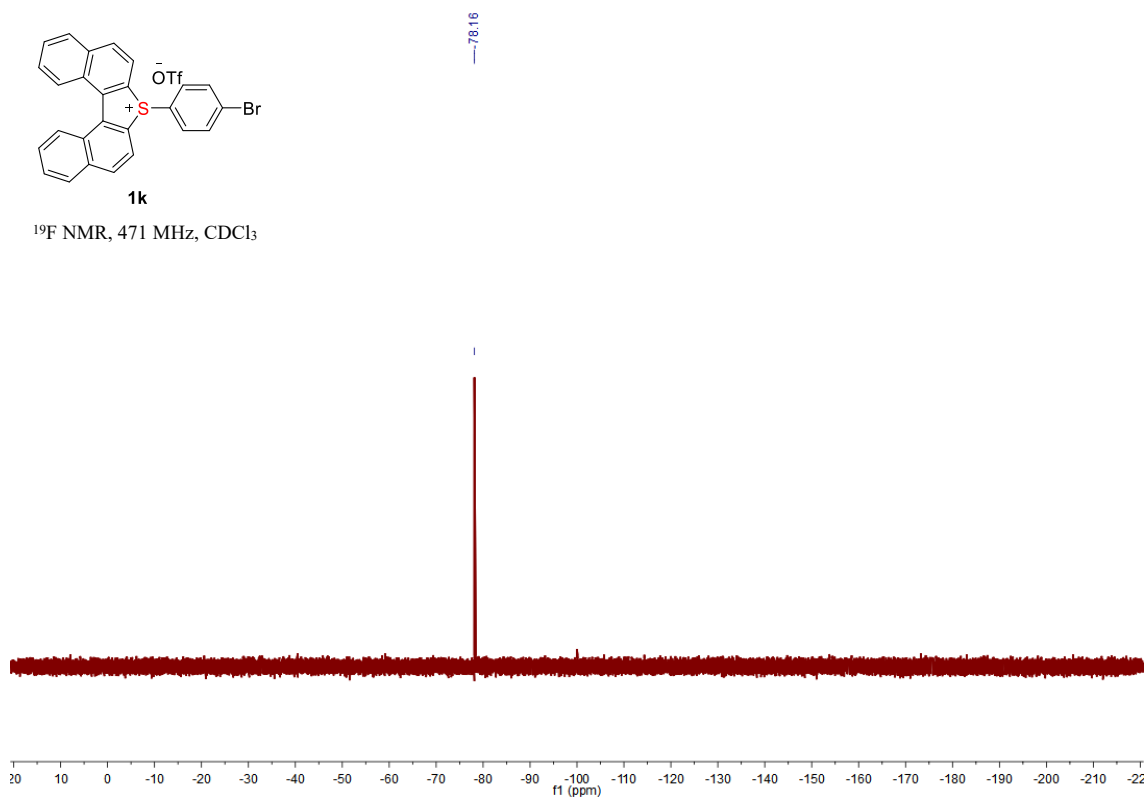


Figure S36. ^{19}F NMR spectrum of **1k** (471 MHz, CDCl_3)

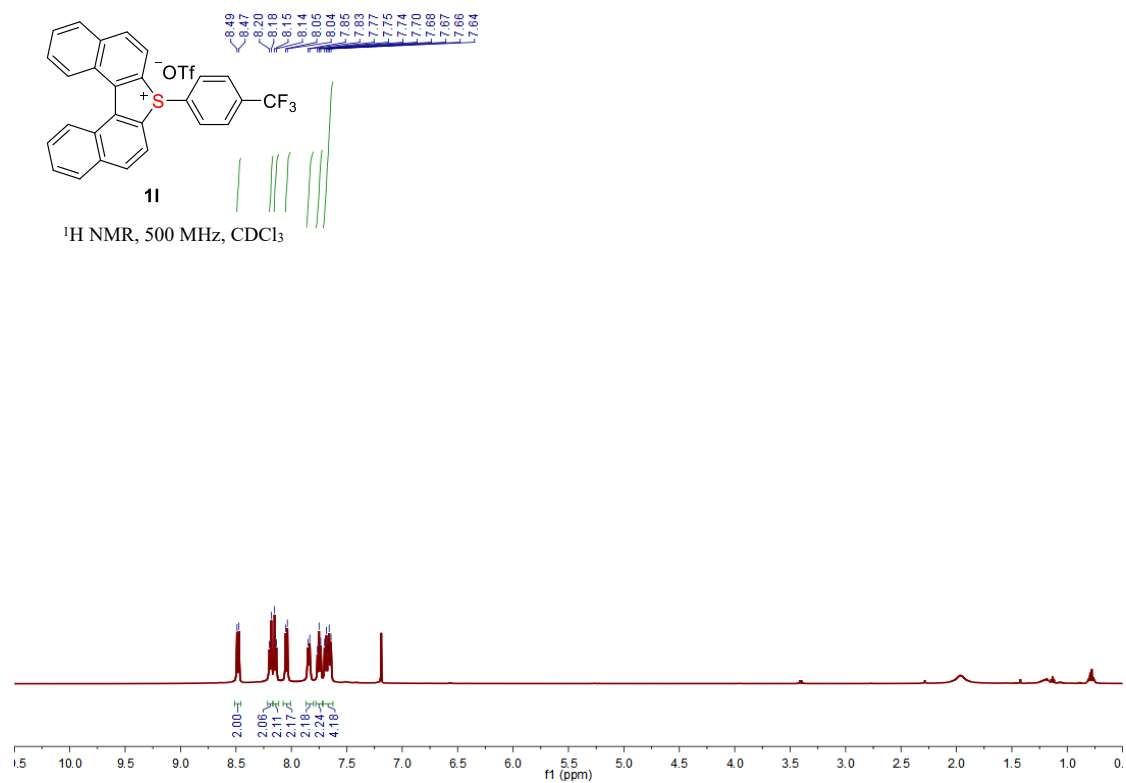


Figure S37. ^1H NMR spectrum of **1l** (500 MHz, CDCl_3)

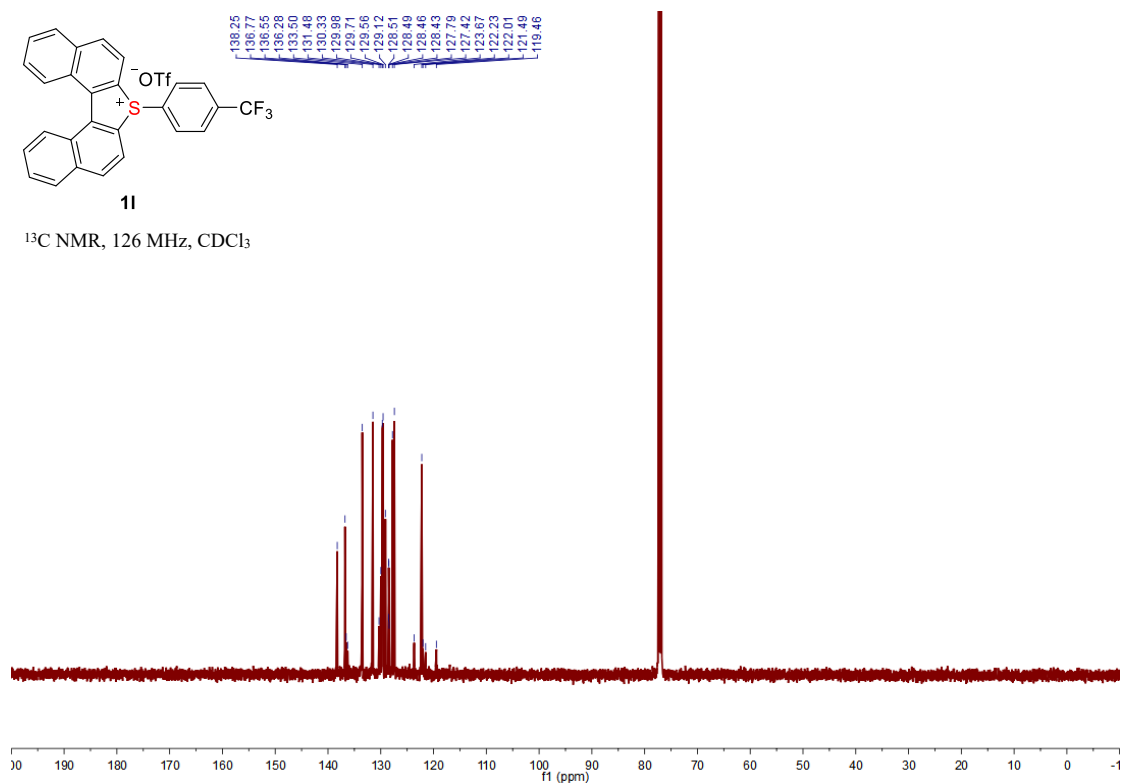


Figure S38. ^{13}C NMR spectrum of **1l** (126 MHz, CDCl_3)

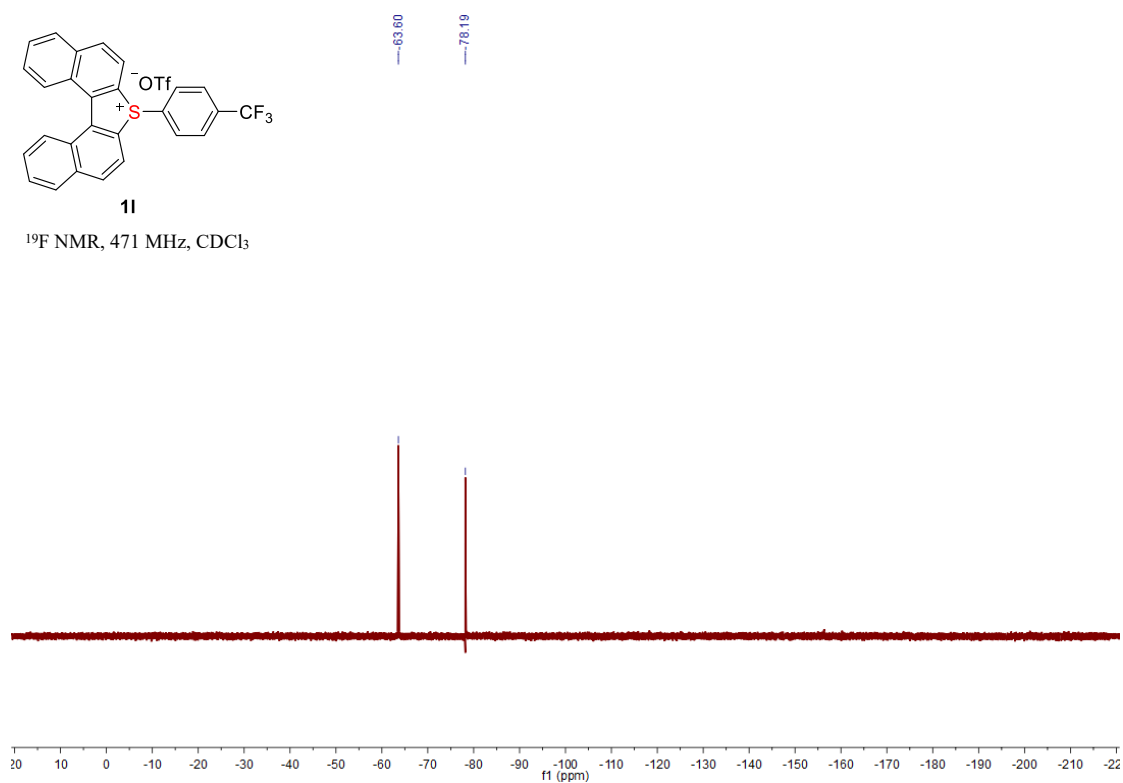


Figure S39. ^{19}F NMR spectrum of **1l** (471 MHz, CDCl_3)

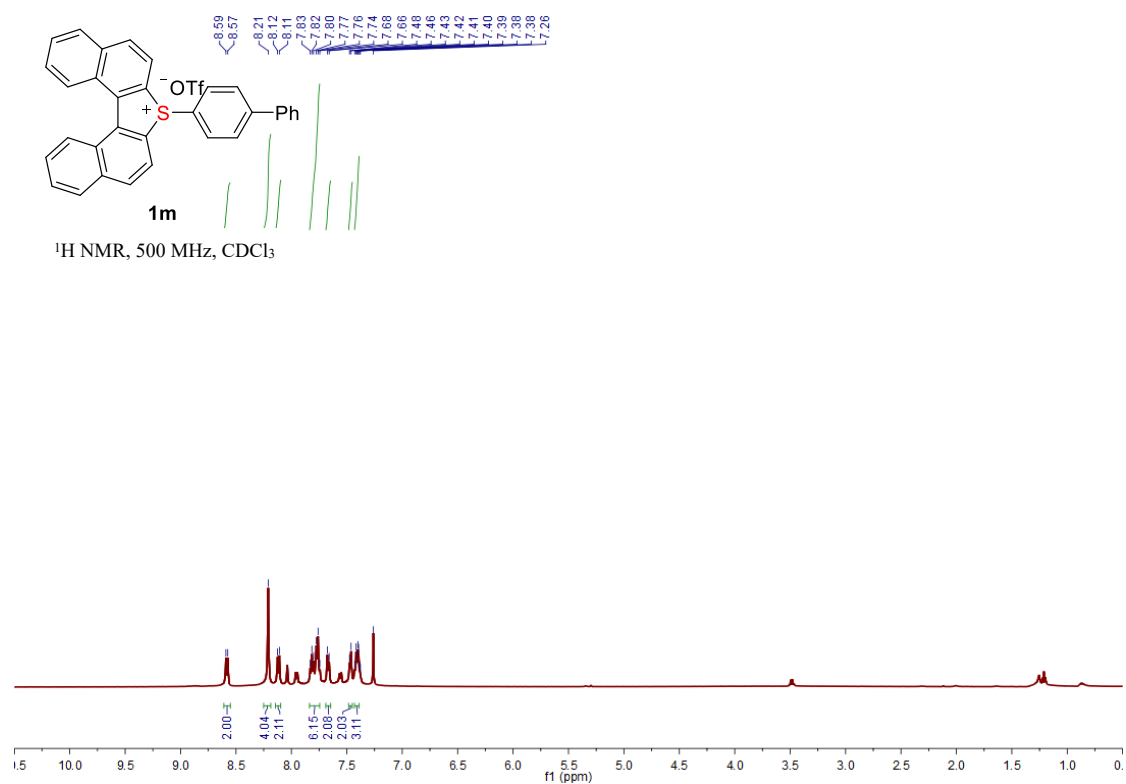


Figure S40. ^1H NMR spectrum of **1m (500 MHz, CDCl_3)**

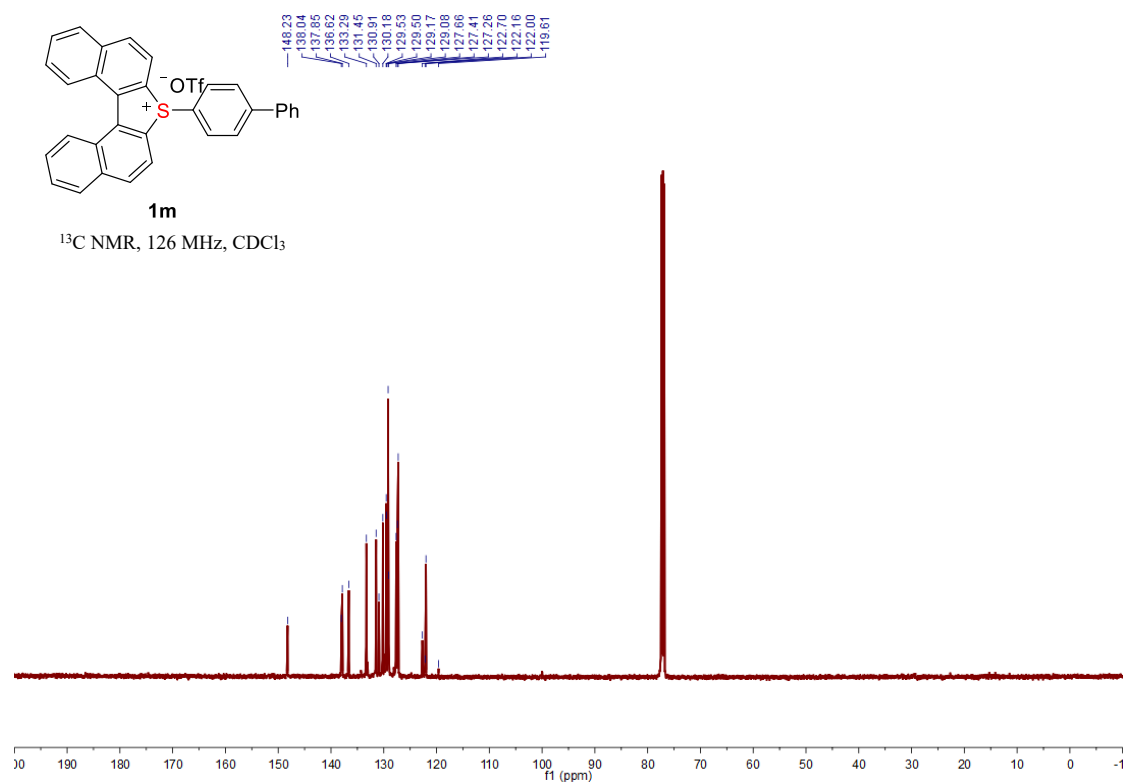


Figure S41. ^{13}C NMR spectrum of **1m (126 MHz, CDCl_3)**

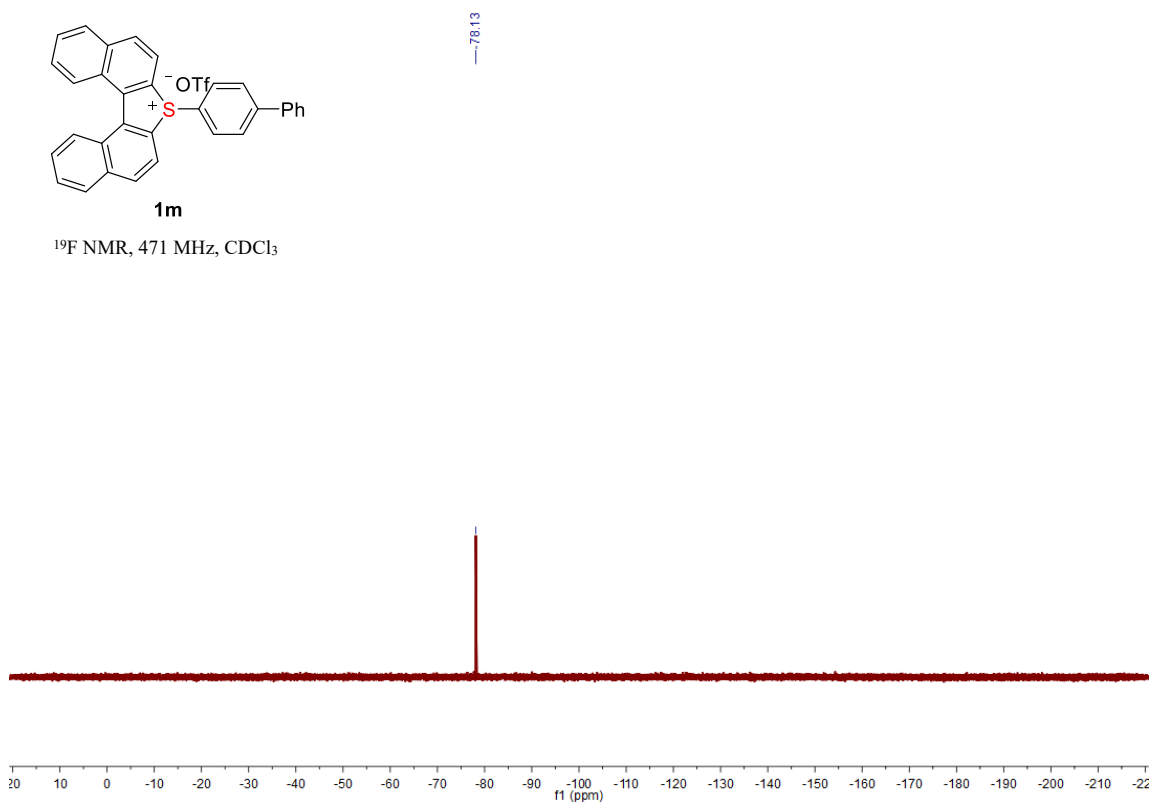


Figure S42. ^{19}F NMR spectrum of **1m** (471 MHz, CDCl_3)

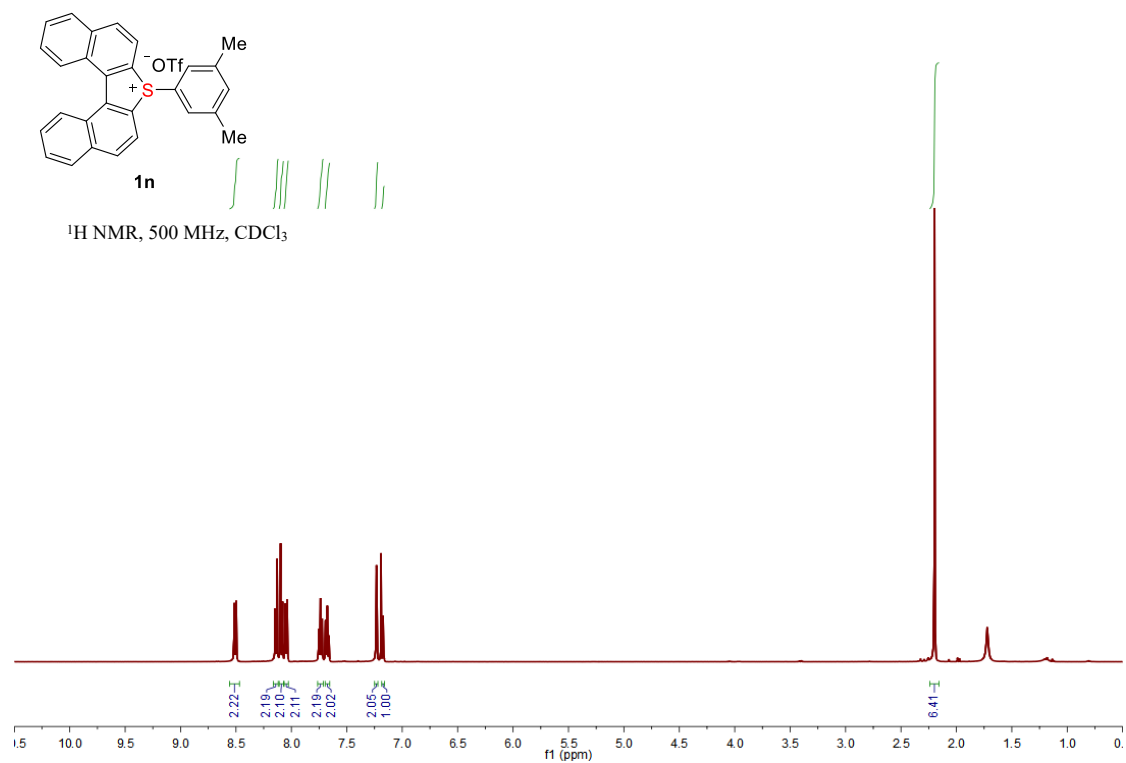


Figure S43. ^1H NMR spectrum of **1n** (500 MHz, CDCl_3)

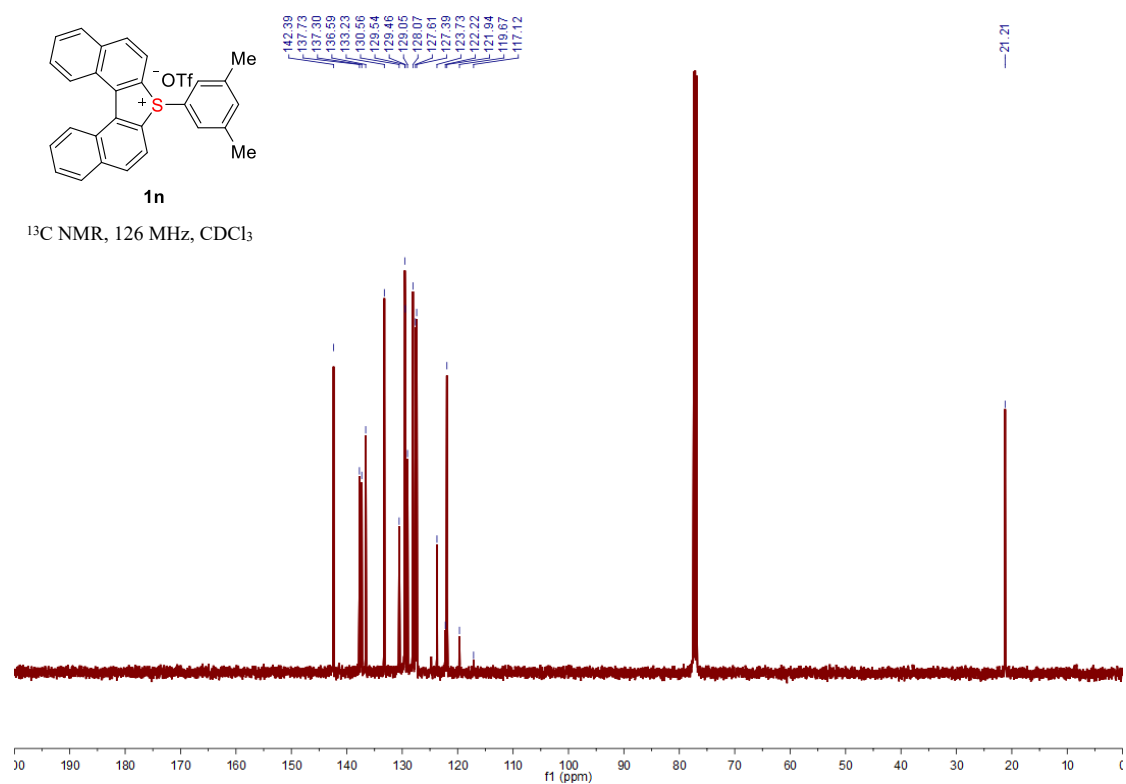


Figure S44. ^{13}C NMR spectrum of **1n** (126 MHz, CDCl_3)

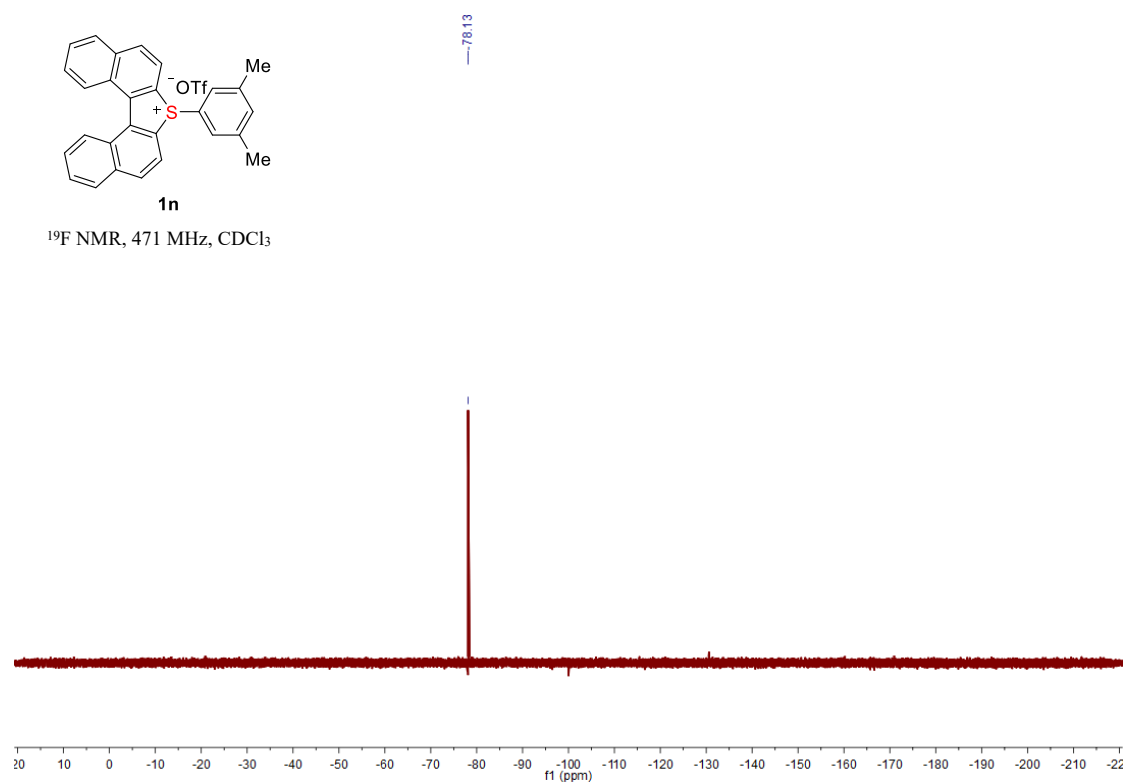


Figure S45. ^{19}F NMR spectrum of **1n** (471 MHz, CDCl_3)

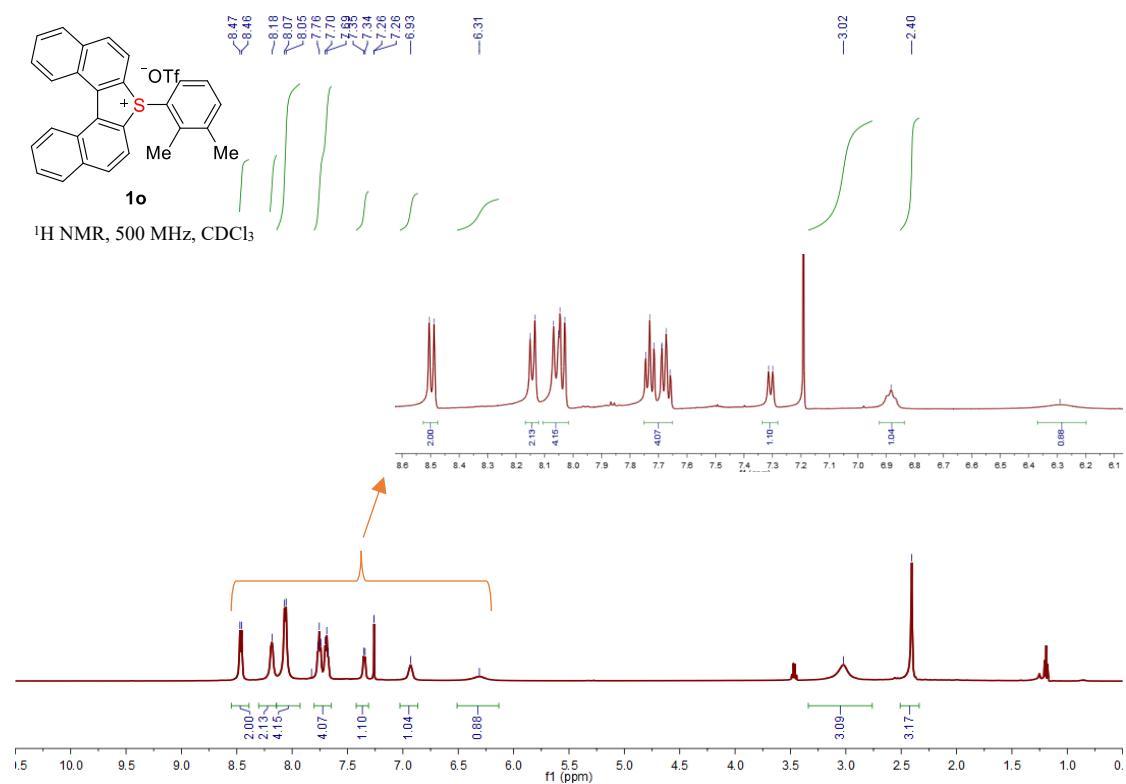


Figure S46. ¹H NMR spectrum of **1o** (500 MHz, CDCl₃)

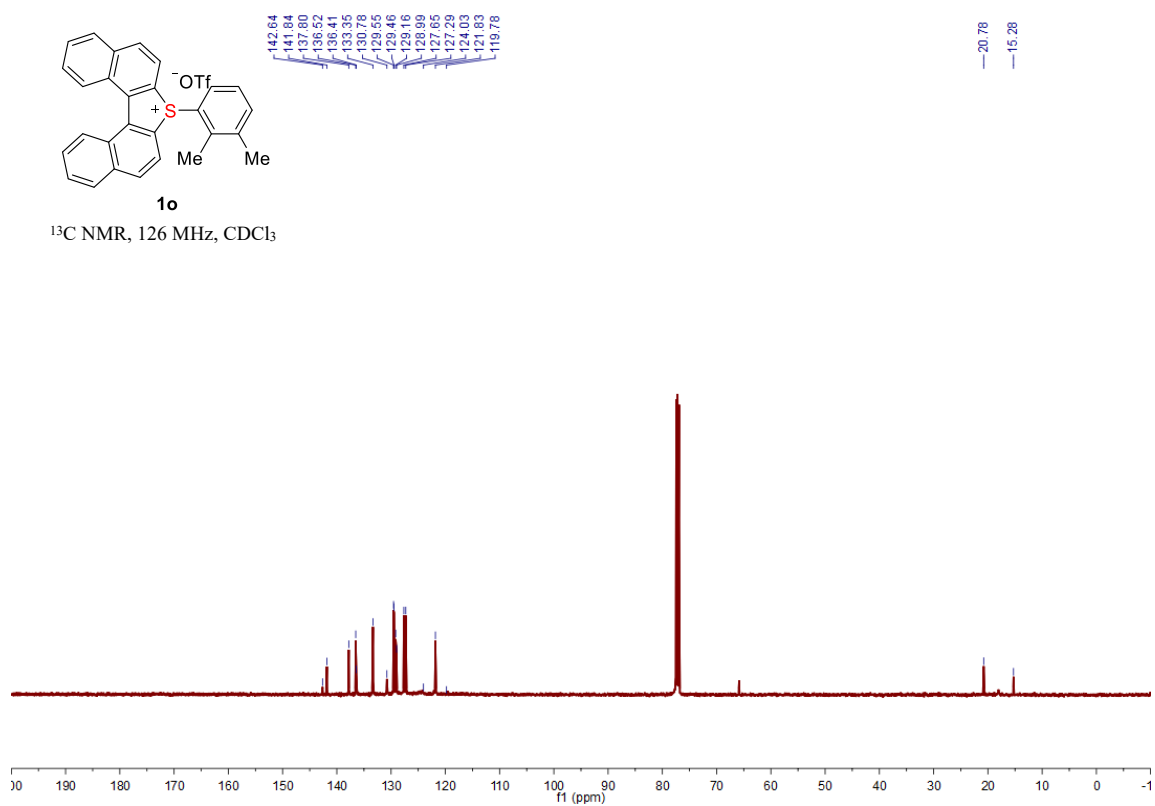


Figure S47. ¹³C NMR spectrum of **1o** (126 MHz, CDCl₃)

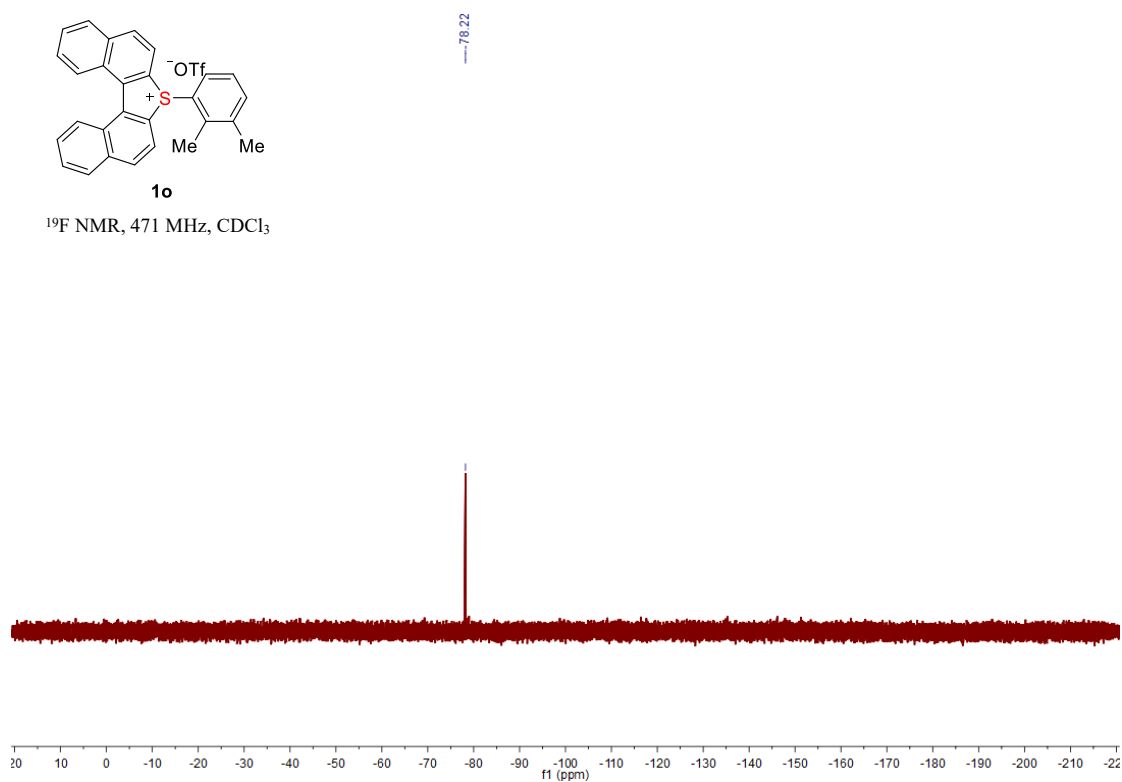


Figure S48. ^{19}F NMR spectrum of 1o (471 MHz, CDCl_3)

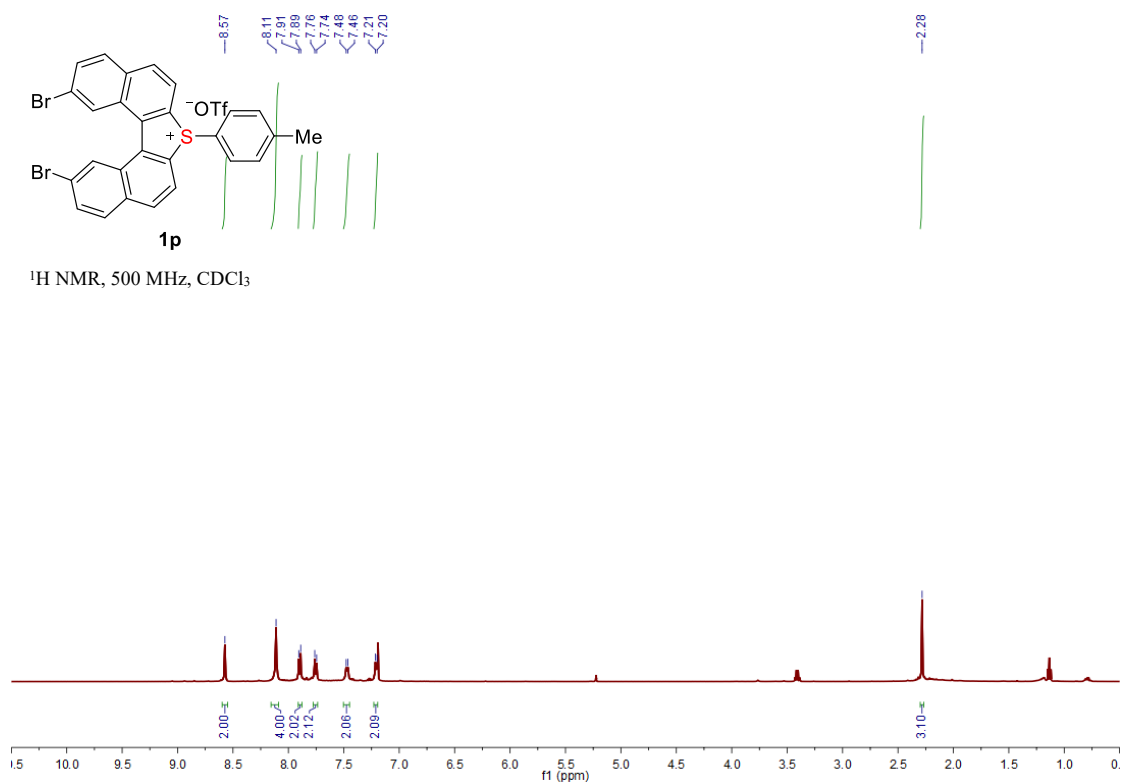


Figure S49. ^1H NMR spectrum of 1p (500 MHz, CDCl_3)

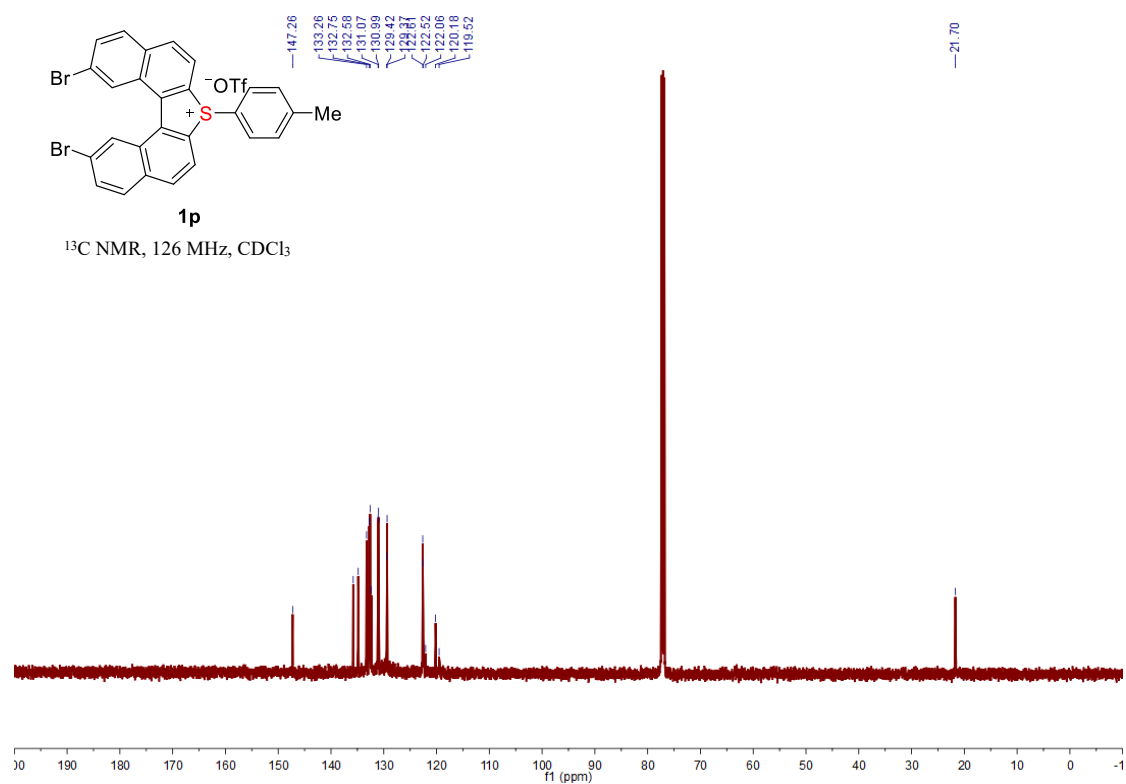


Figure S50. ¹³C NMR spectrum of 1p (126 MHz, CDCl₃)

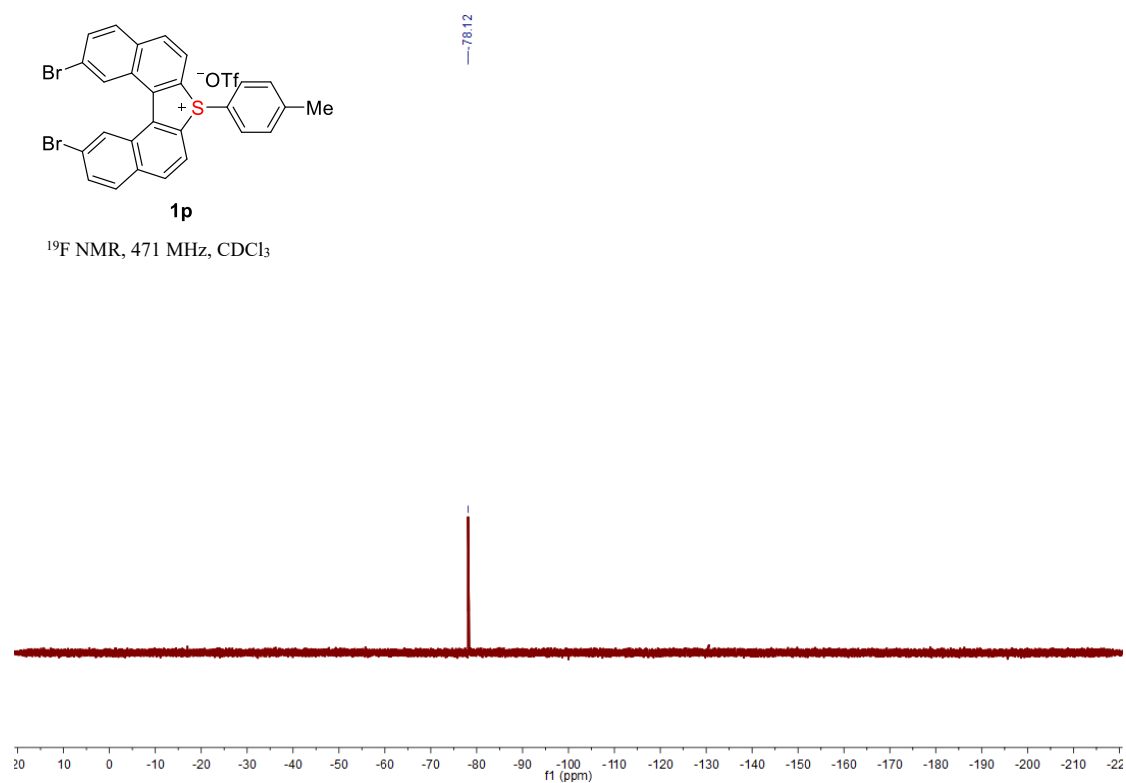


Figure S51. ¹⁹F NMR spectrum of 1p (471 MHz, CDCl₃)

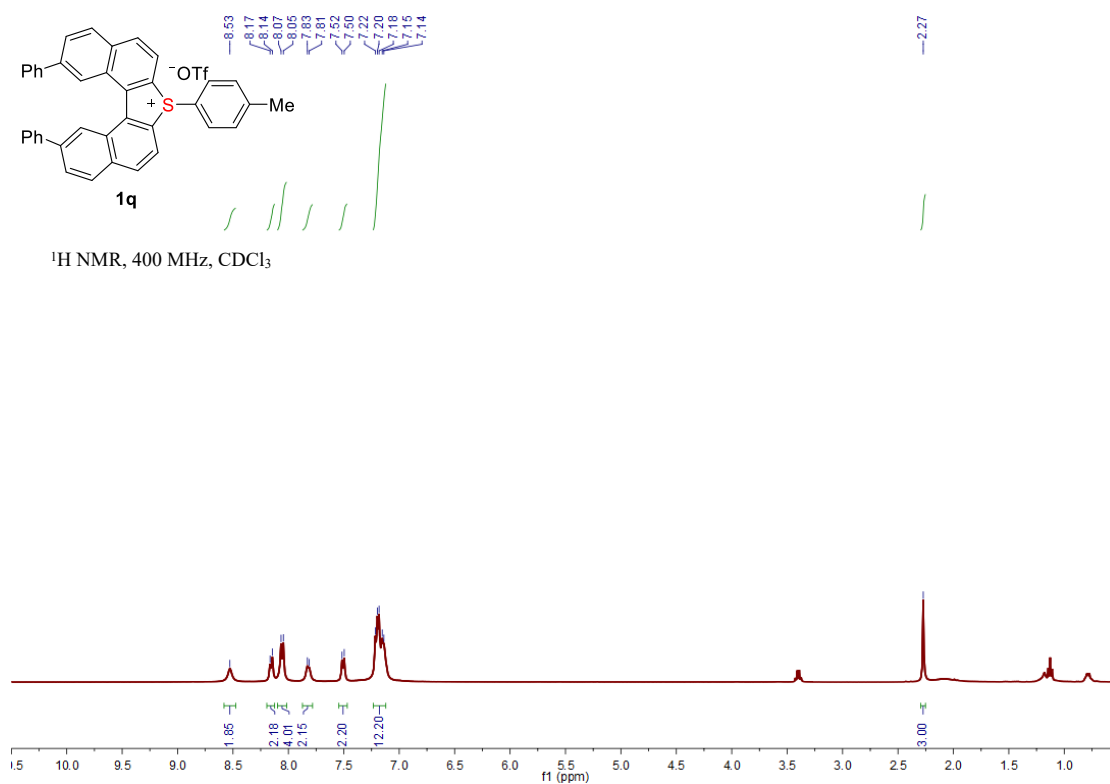


Figure S52. ^1H NMR spectrum of **1q** (400 MHz, CDCl_3)

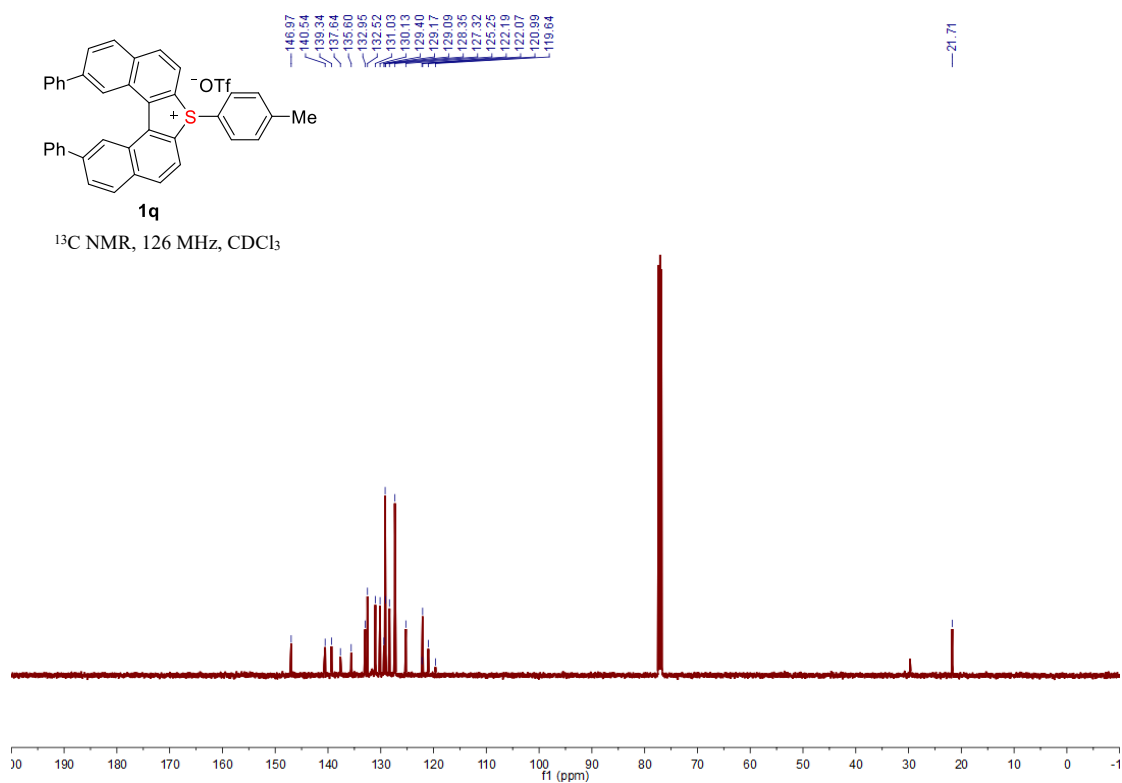


Figure S53. ^{13}C NMR spectrum of **1q** (126 MHz, CDCl_3)

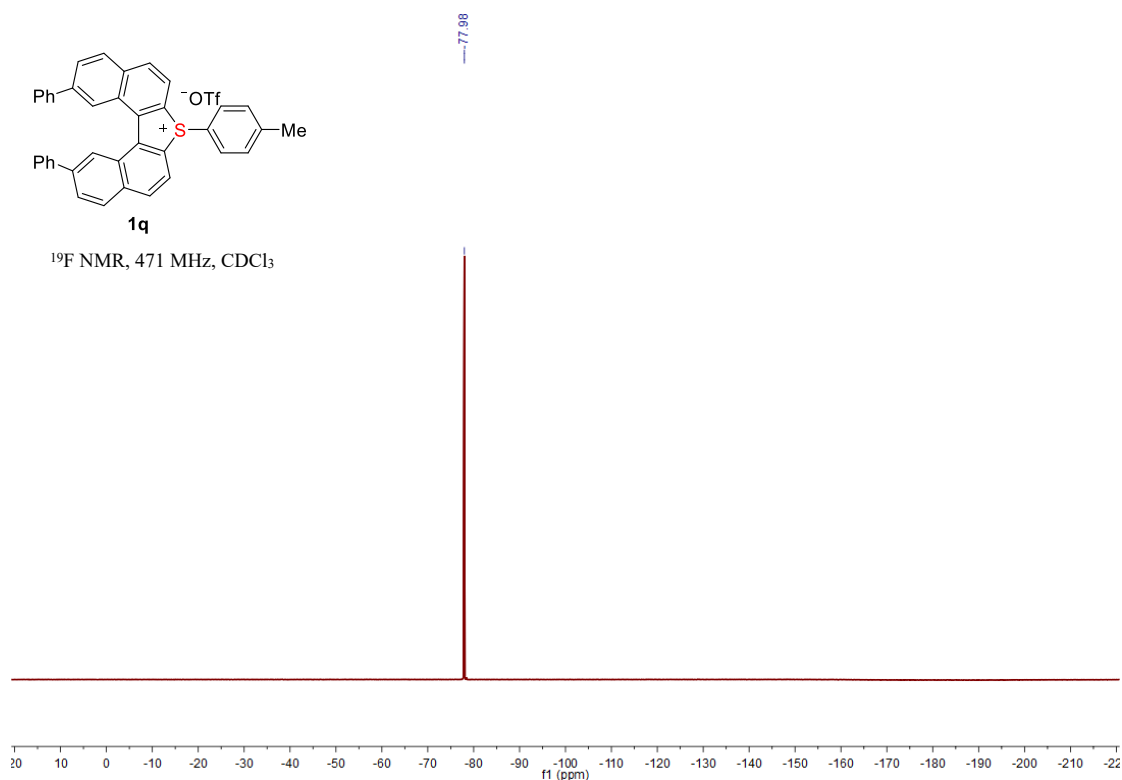


Figure S54. ^{19}F NMR spectrum of 1q (471 MHz, CDCl_3)

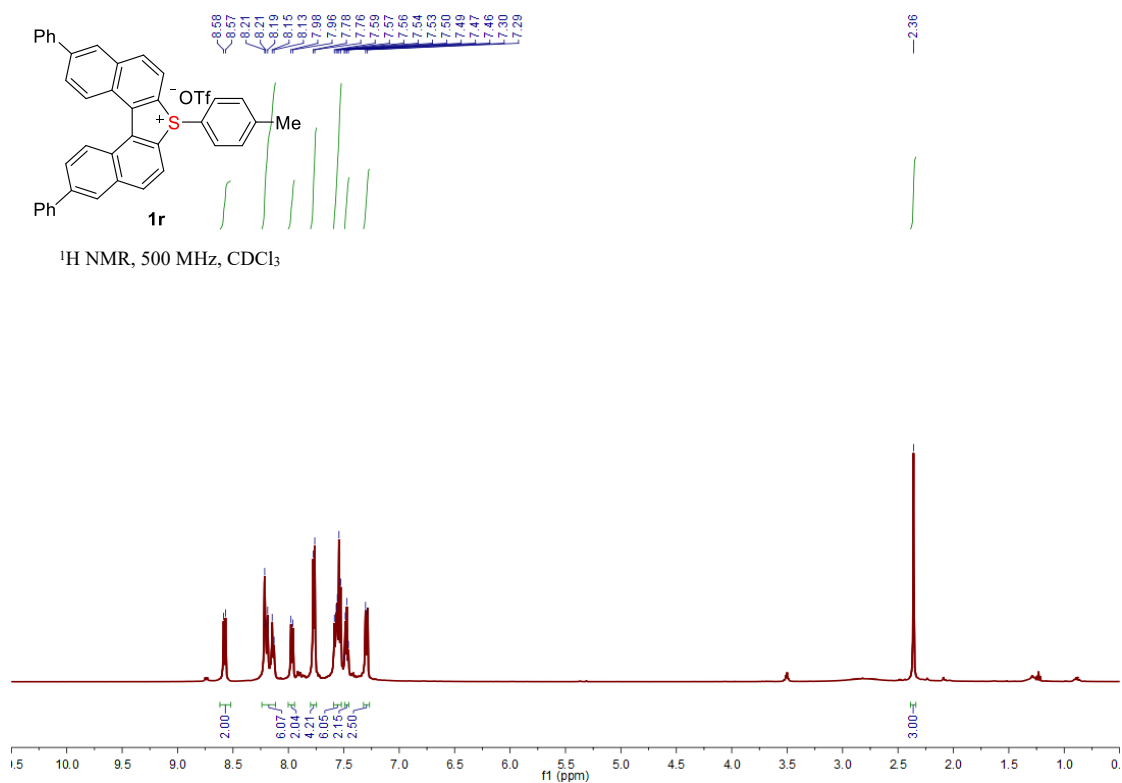


Figure S55. ^1H NMR spectrum of 1r (500 MHz, CDCl_3)

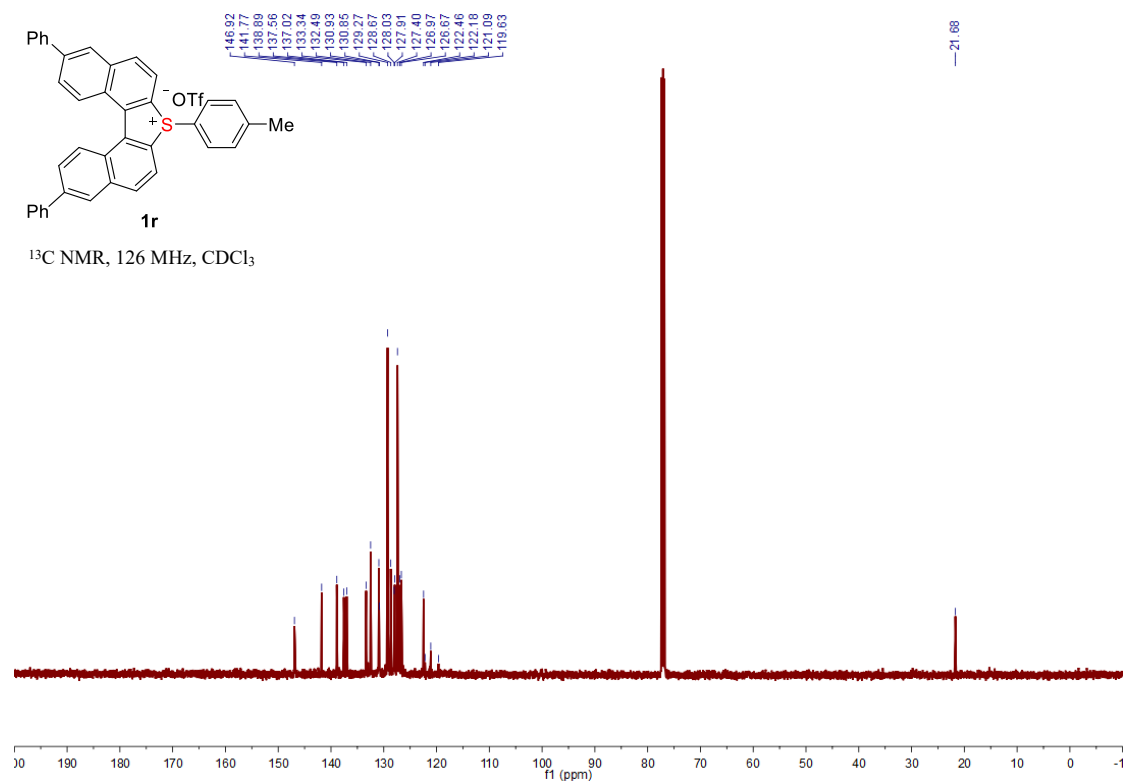


Figure S56. ^{13}C NMR spectrum of **1r** (126 MHz, CDCl_3)

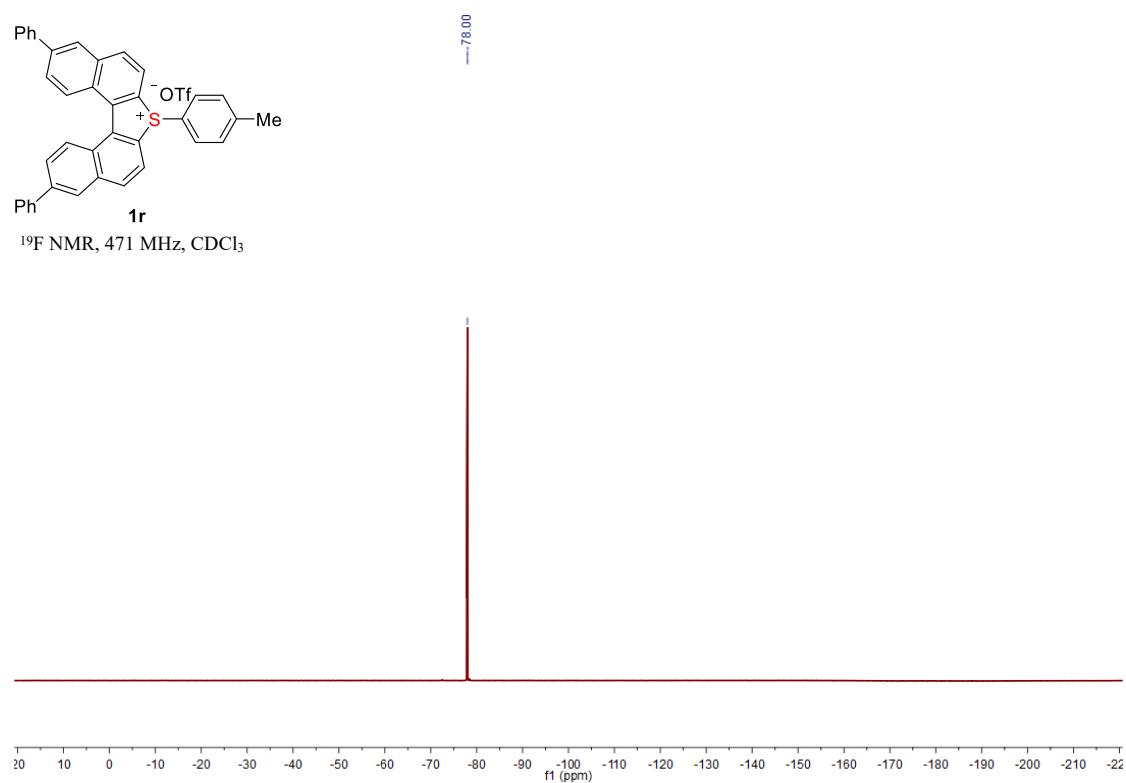


Figure S57. ^{19}F NMR spectrum of **1r** (471 MHz, CDCl_3)

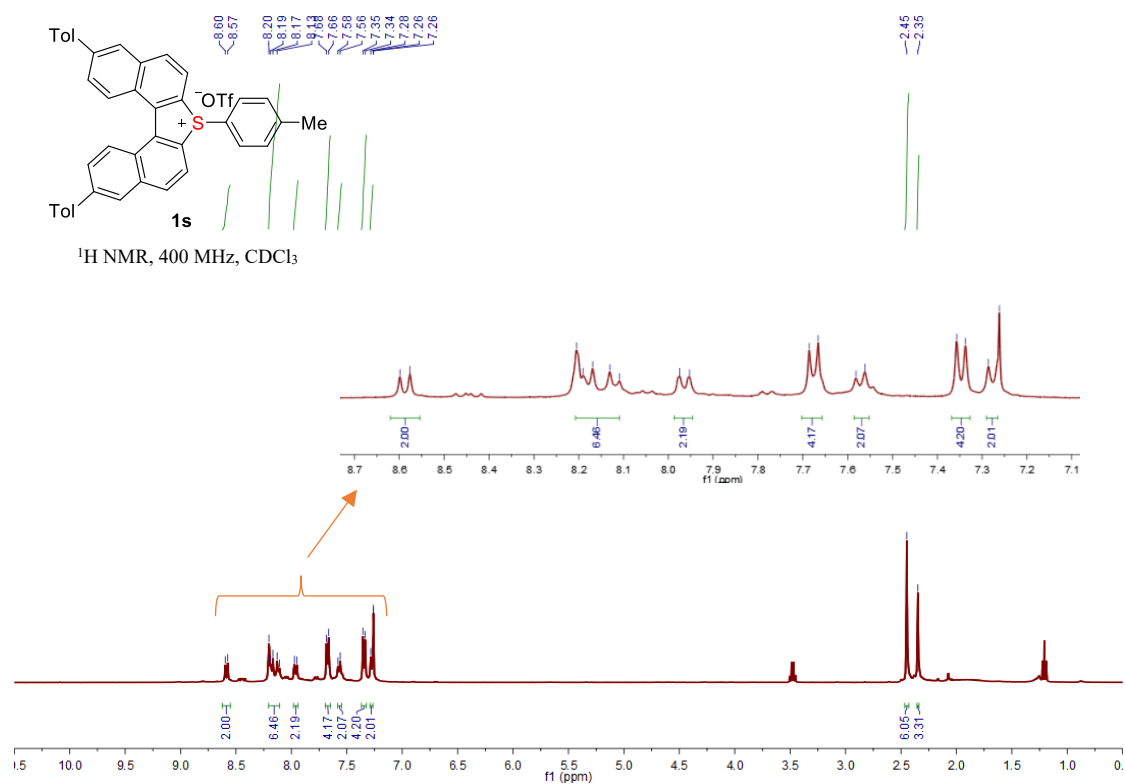


Figure S58. ¹H NMR spectrum of 1s (400 MHz, CDCl₃)

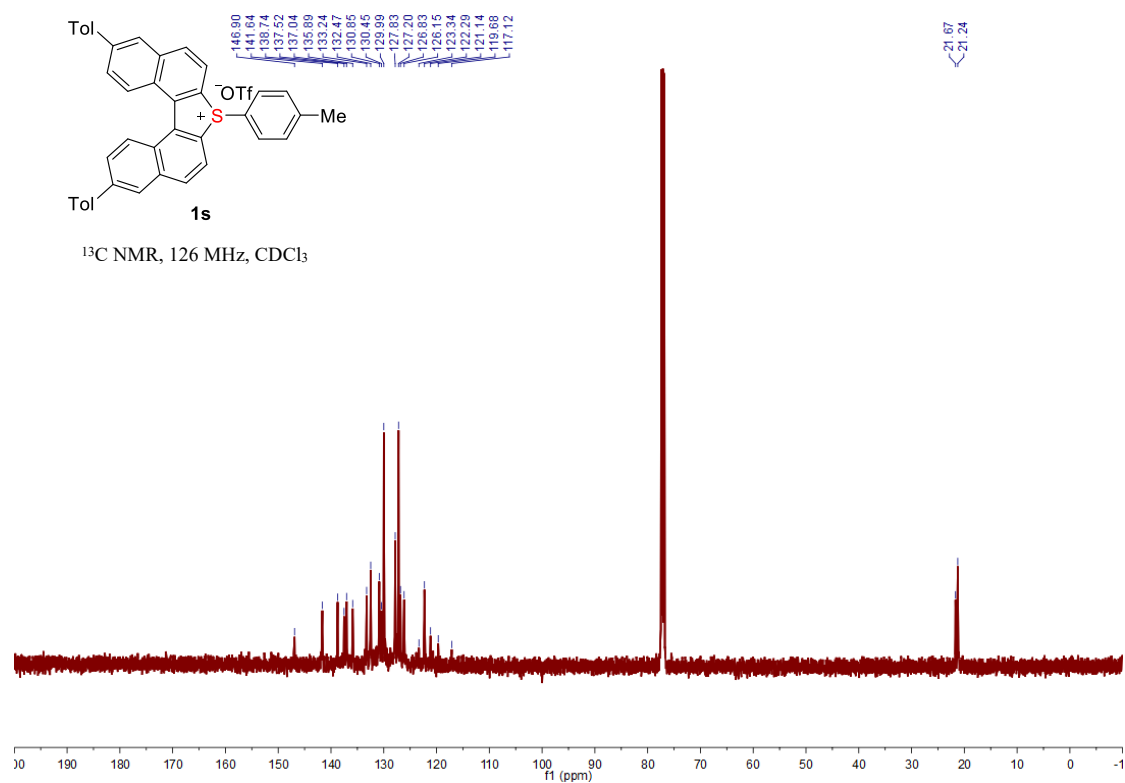


Figure S59. ¹³C NMR spectrum of 1s (126 MHz, CDCl₃)

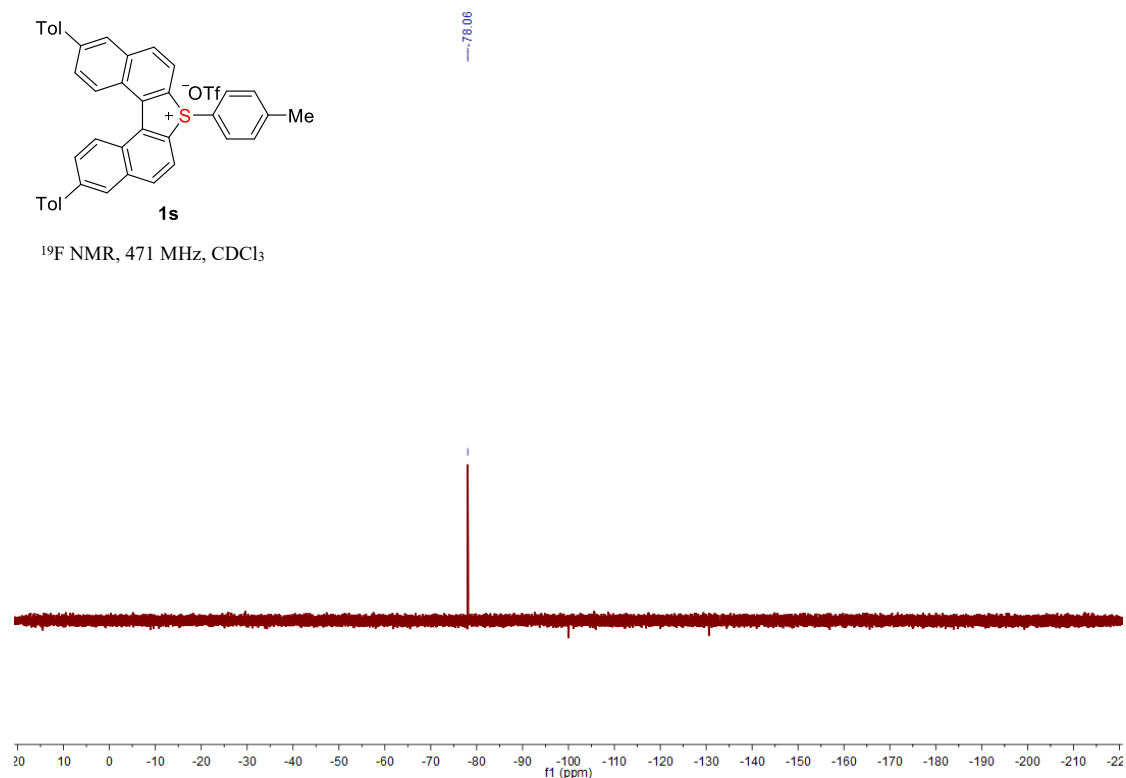


Figure S60. ^{19}F NMR spectrum of 1s (471 MHz, CDCl_3)

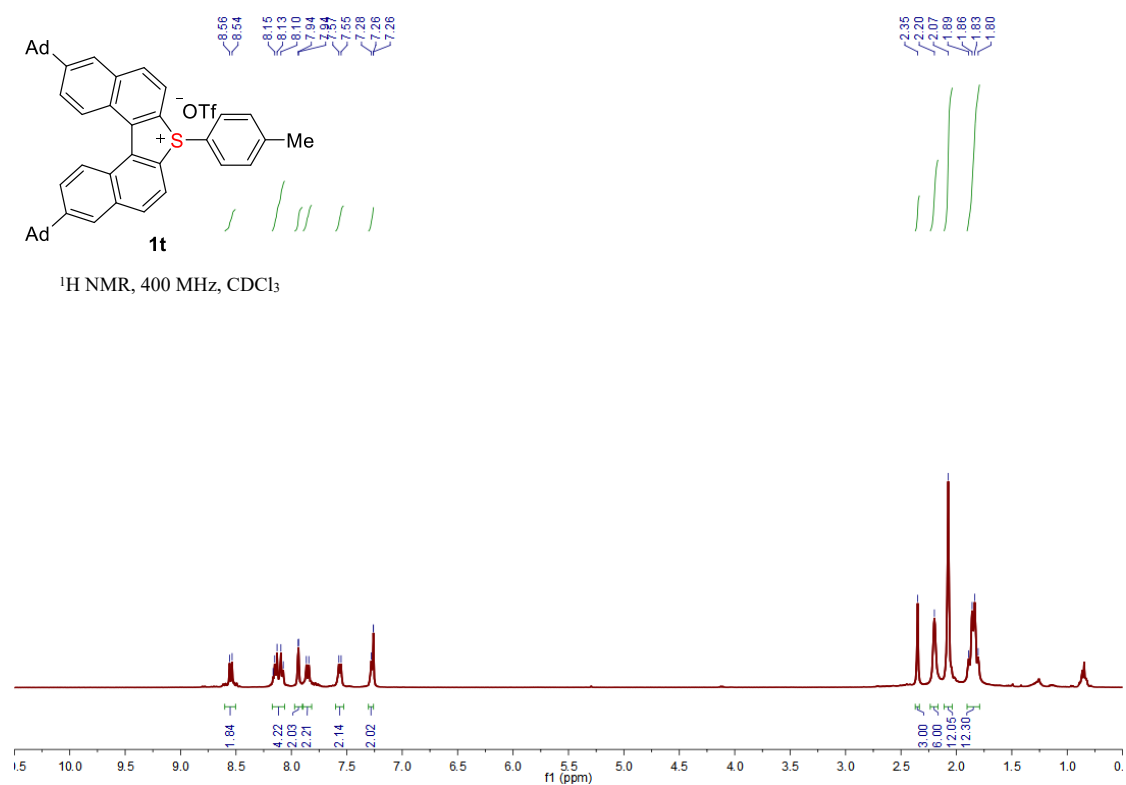


Figure S61. ^1H NMR spectrum of 1t (400 MHz, CDCl_3)

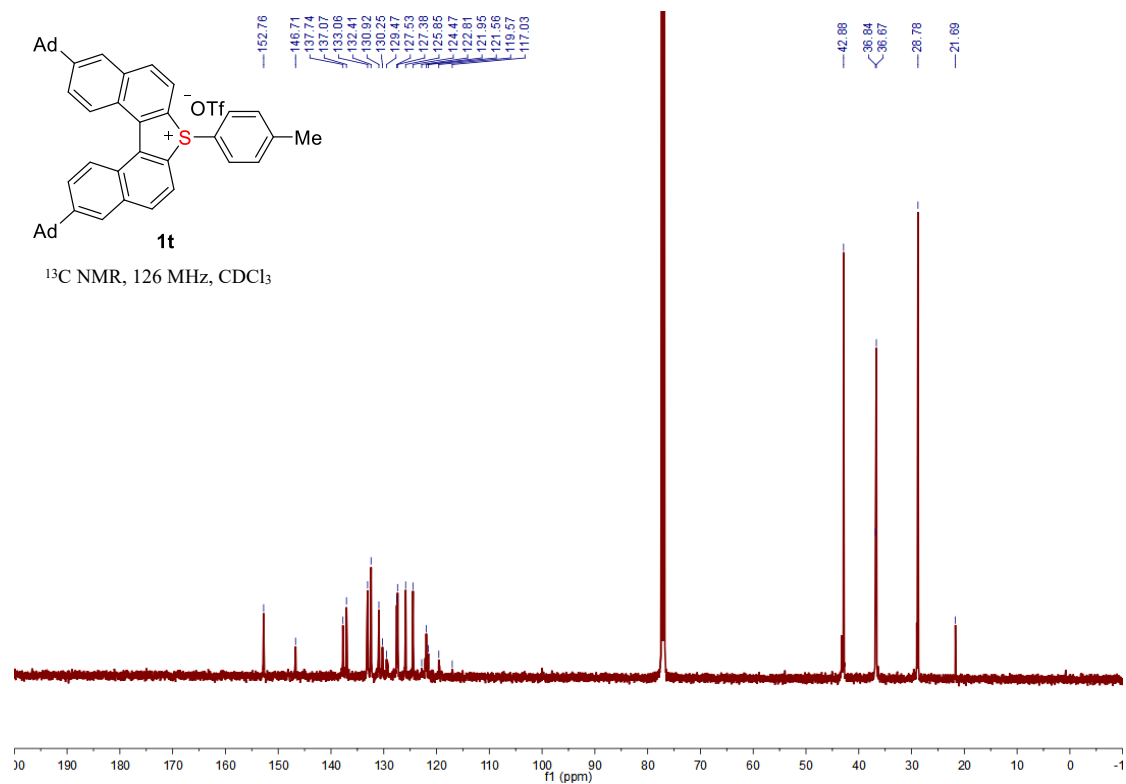


Figure S62. ¹³C NMR spectrum of **1t** (126 MHz, CDCl₃)

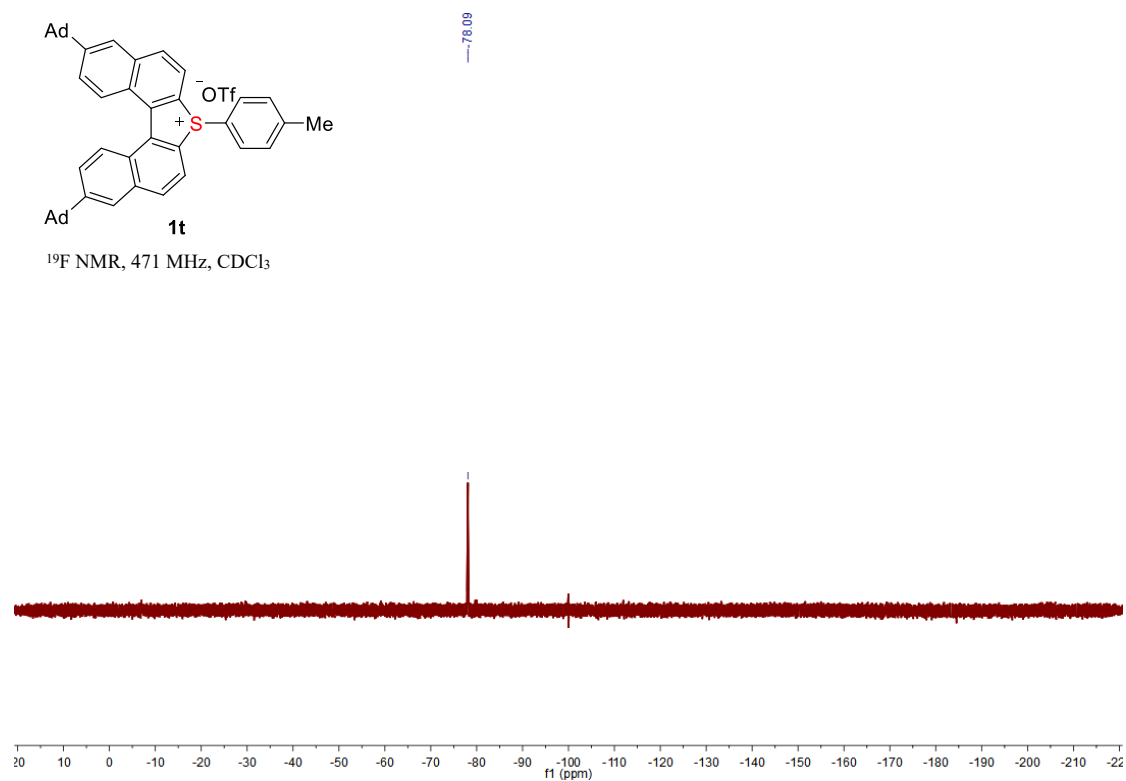


Figure S63. ¹⁹F NMR spectrum of **1t** (471 MHz, CDCl₃)

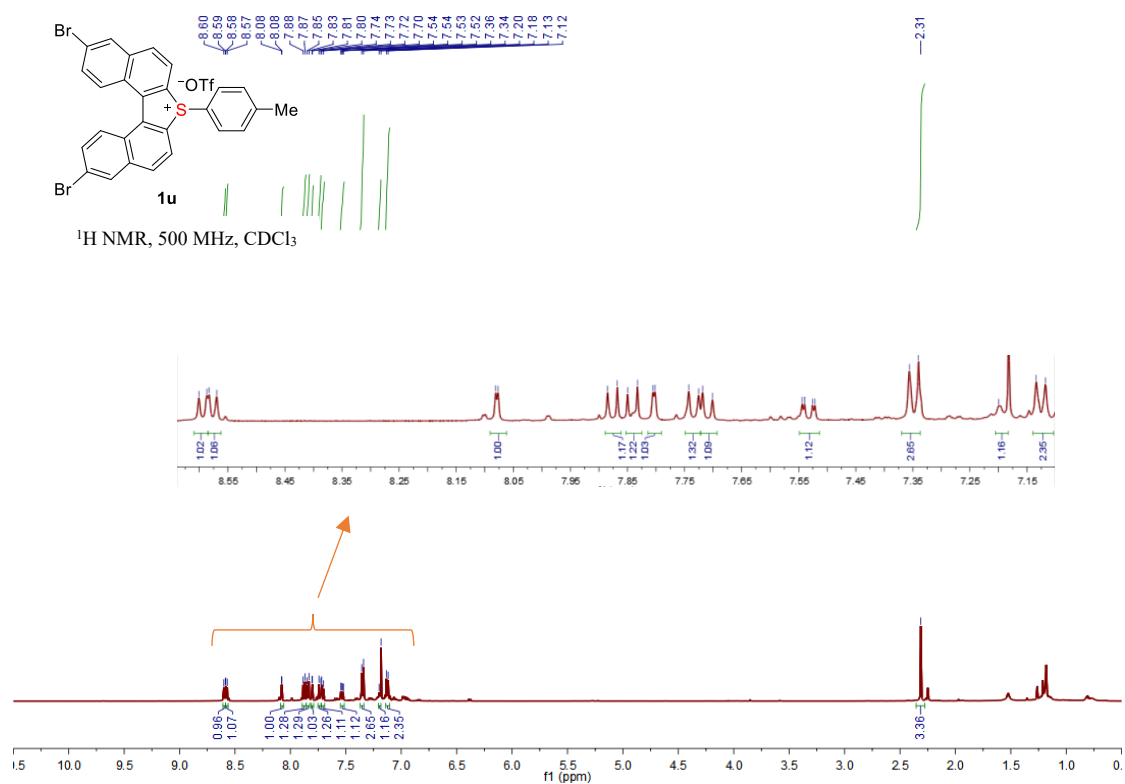


Figure S64. ^1H NMR spectrum of **1u** (500 MHz, CDCl_3)

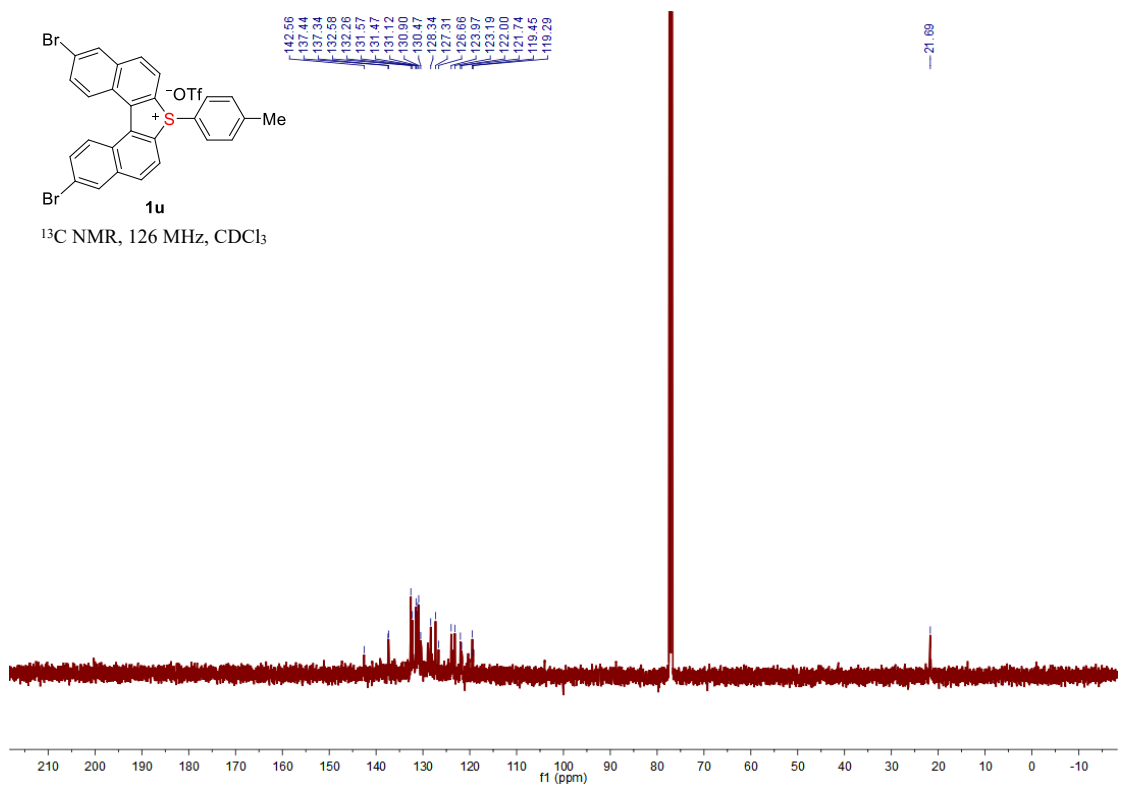


Figure S65. ^{13}C NMR spectrum of **1u** (126 MHz, CDCl_3)

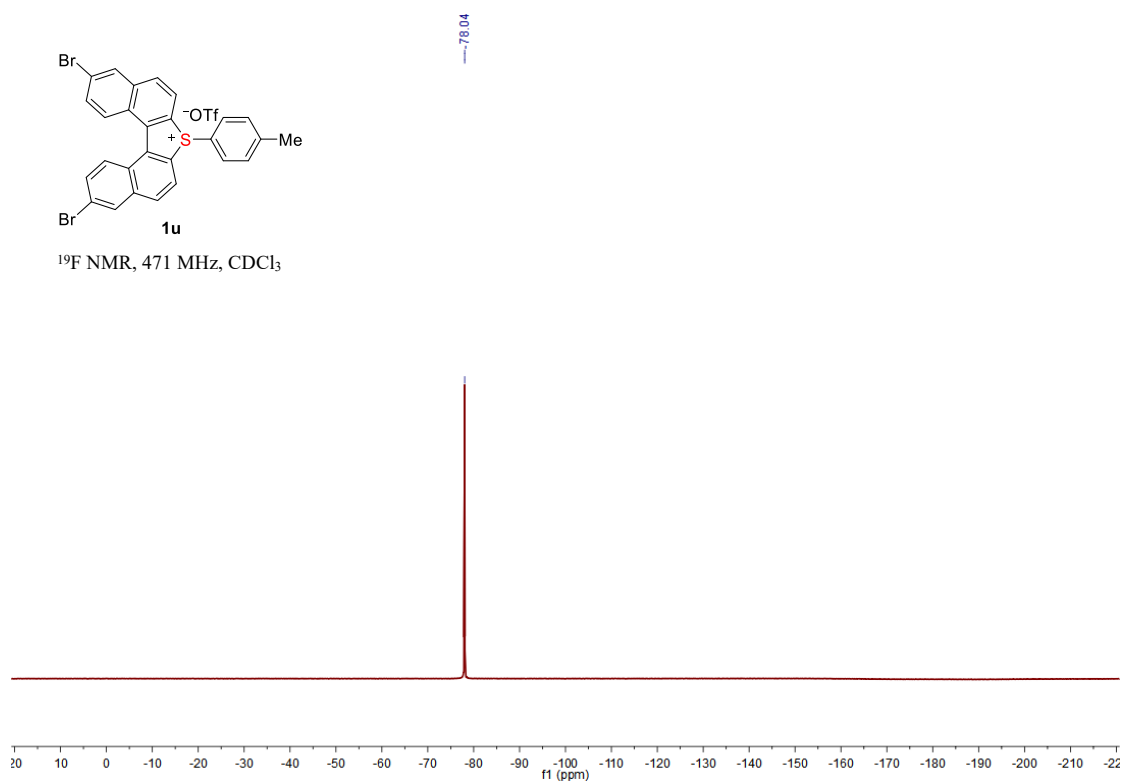


Figure S66. ¹⁹F NMR spectrum of 1u (471 MHz, CDCl₃)

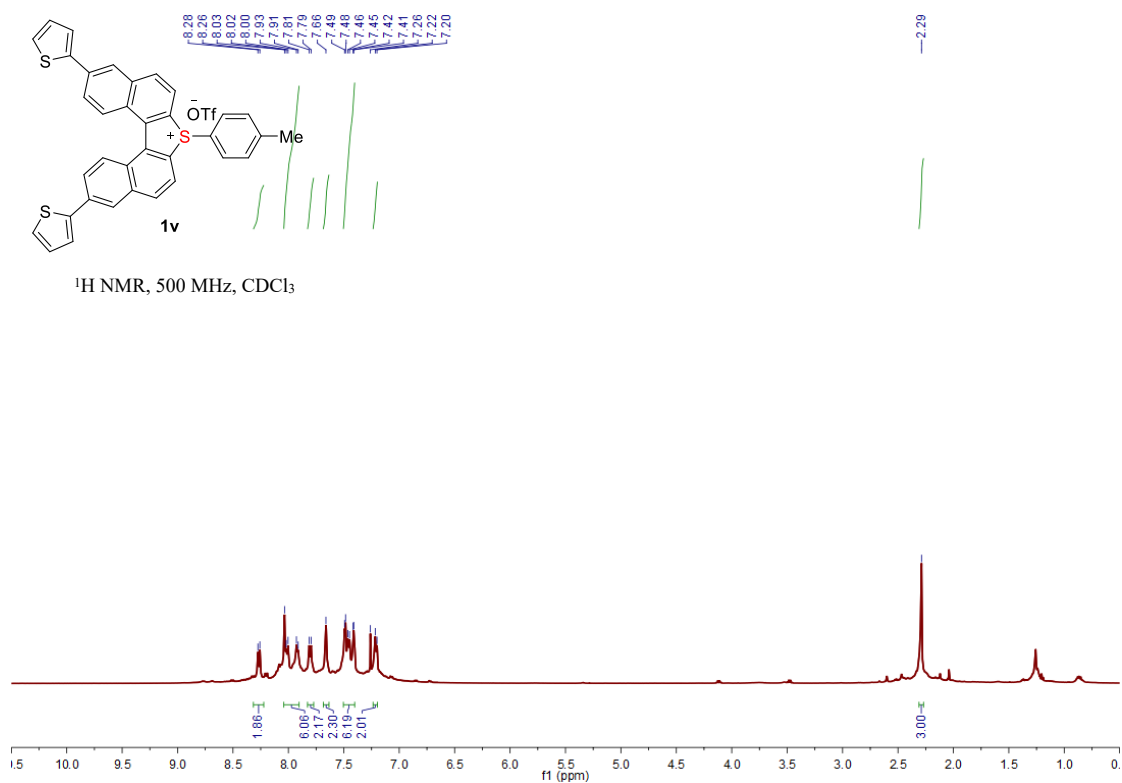


Figure S67. ¹H NMR spectrum of 1v (500 MHz, CDCl₃)

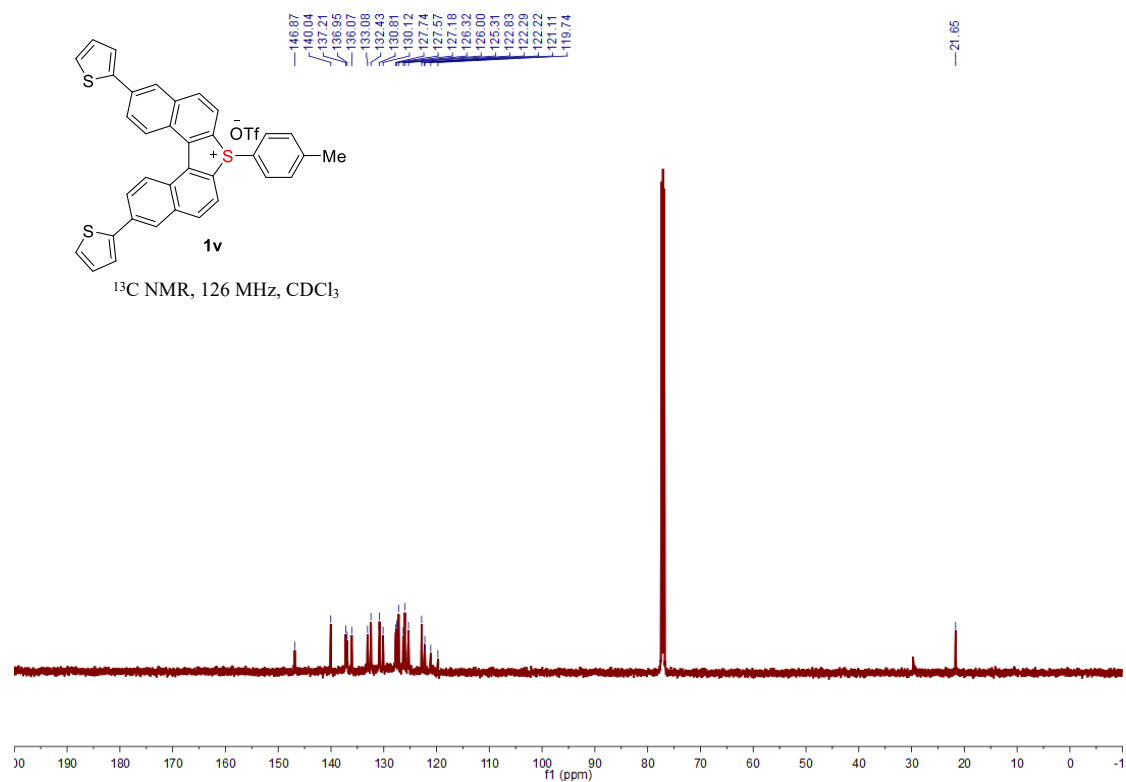


Figure S68. ^{13}C NMR spectrum of **1v** (126 MHz, CDCl_3)

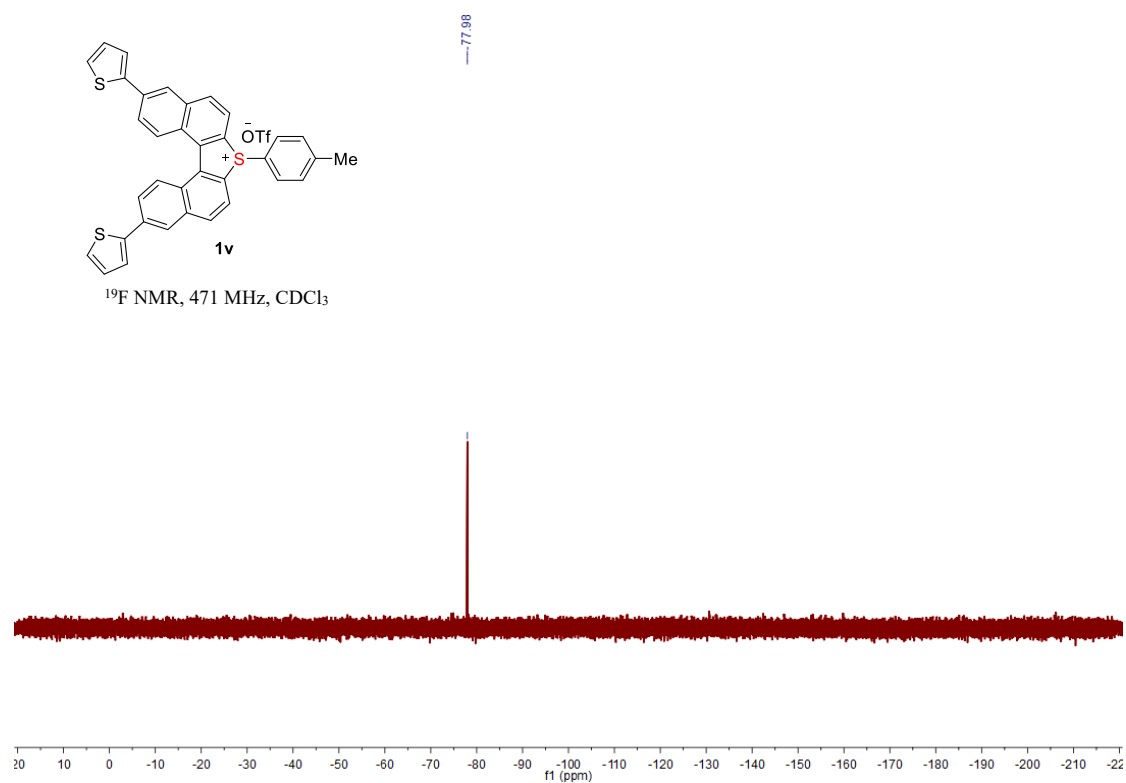


Figure S69. ^{19}F NMR spectrum of **1v** (471 MHz, CDCl_3)

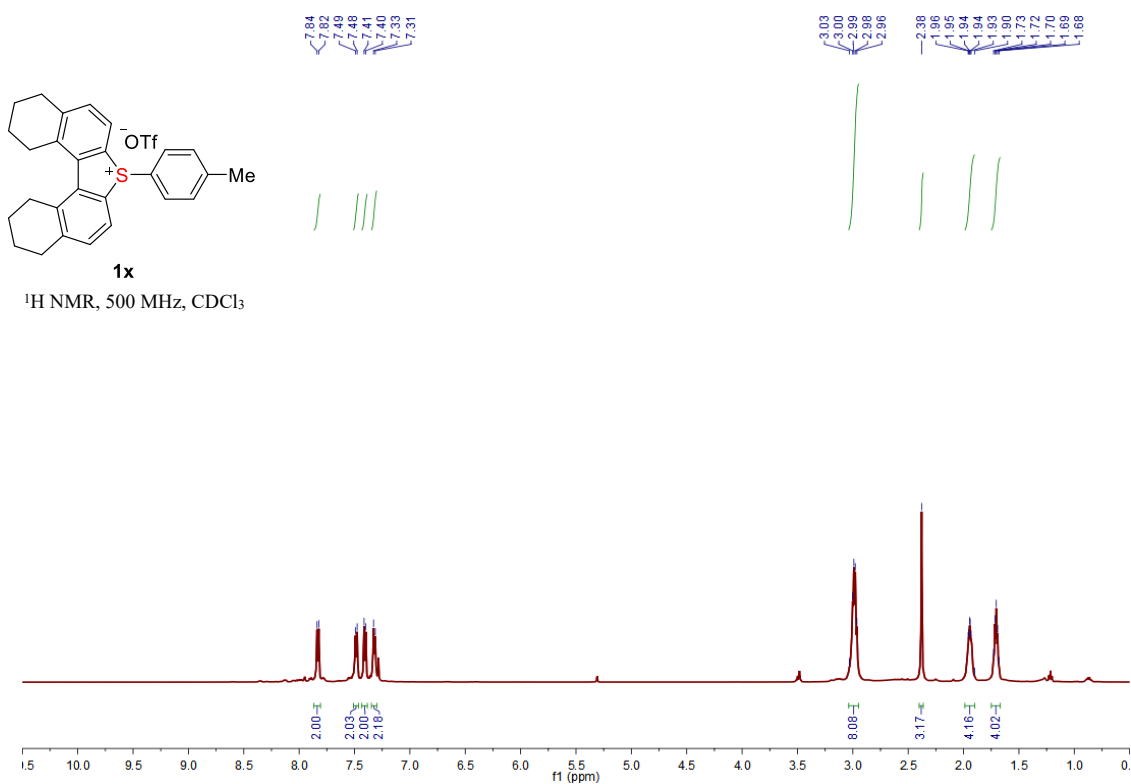


Figure S70. ^1H NMR spectrum of **1x** (500 MHz, CDCl_3)

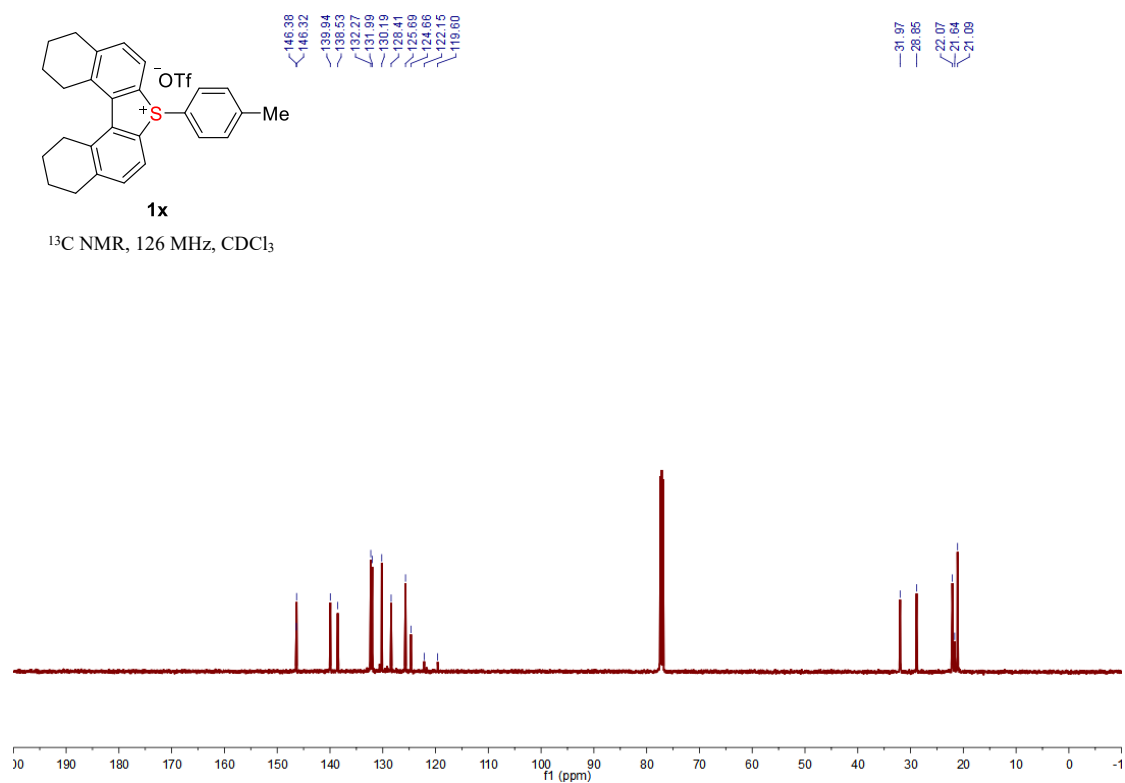


Figure S71. ^{13}C NMR spectrum of **1x** (126 MHz, CDCl_3)

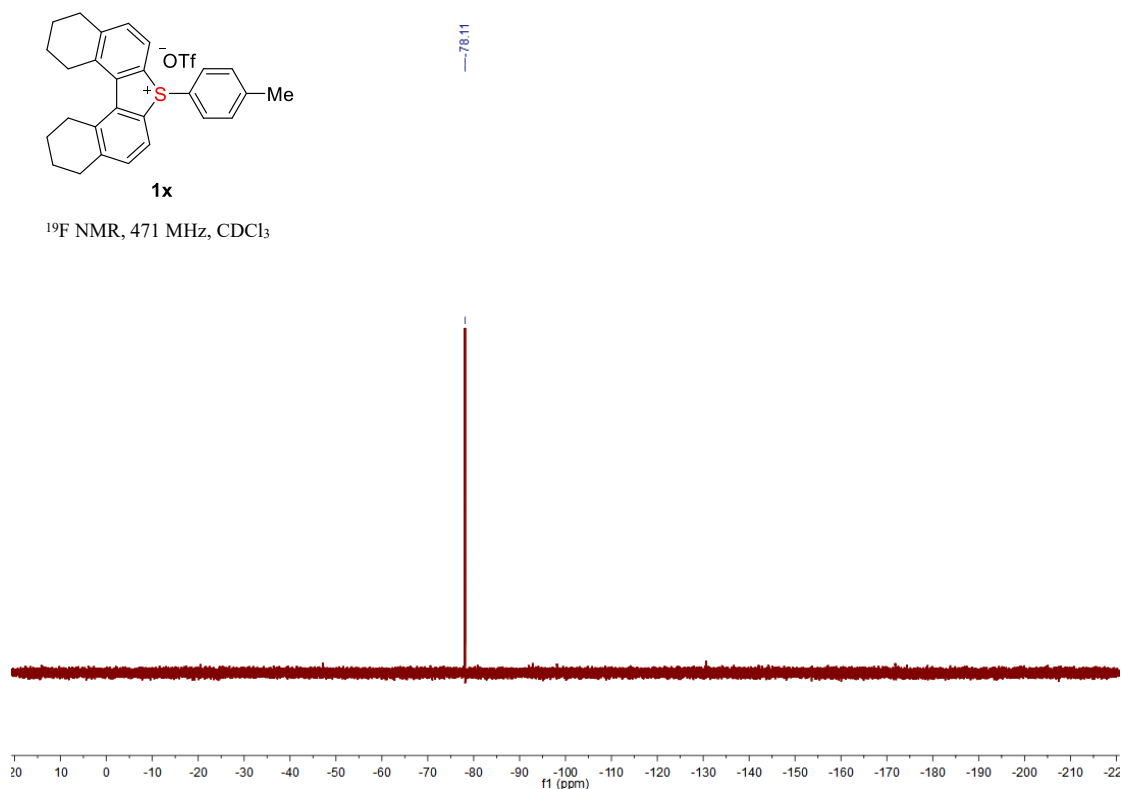


Figure S72. ^{19}F NMR spectrum of 1x (471 MHz, CDCl_3)

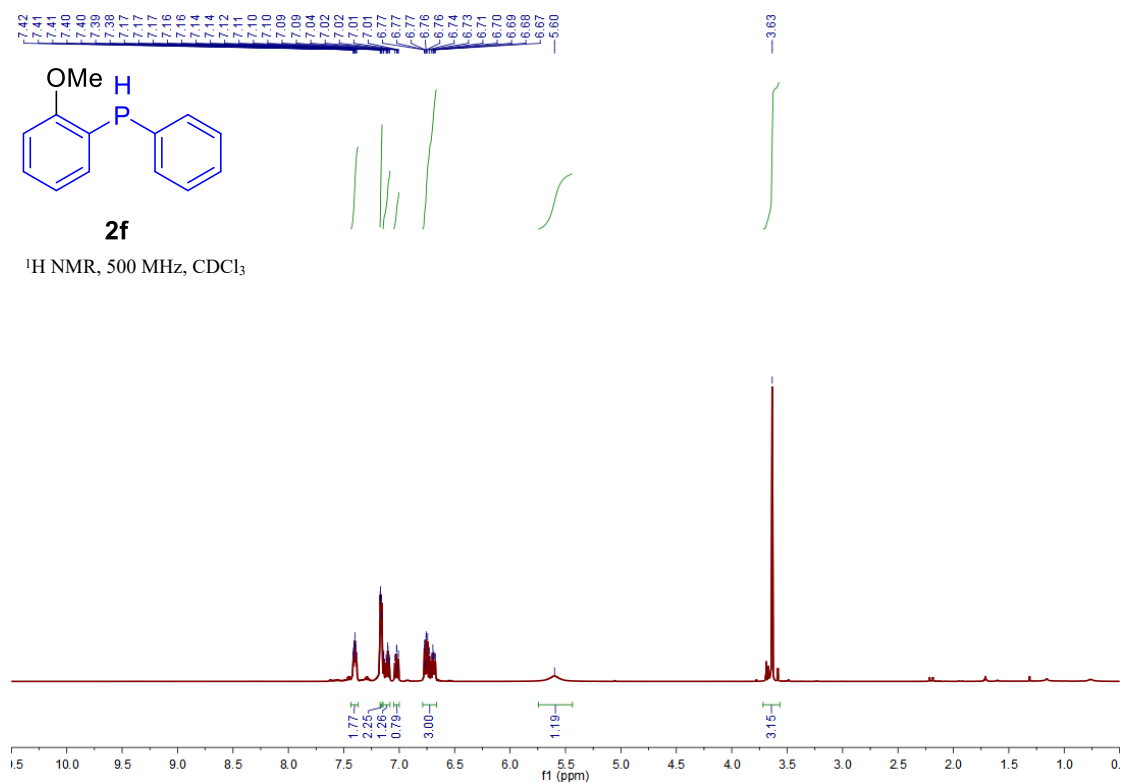


Figure S73. ^1H NMR spectrum of 2f (500 MHz, CDCl_3)

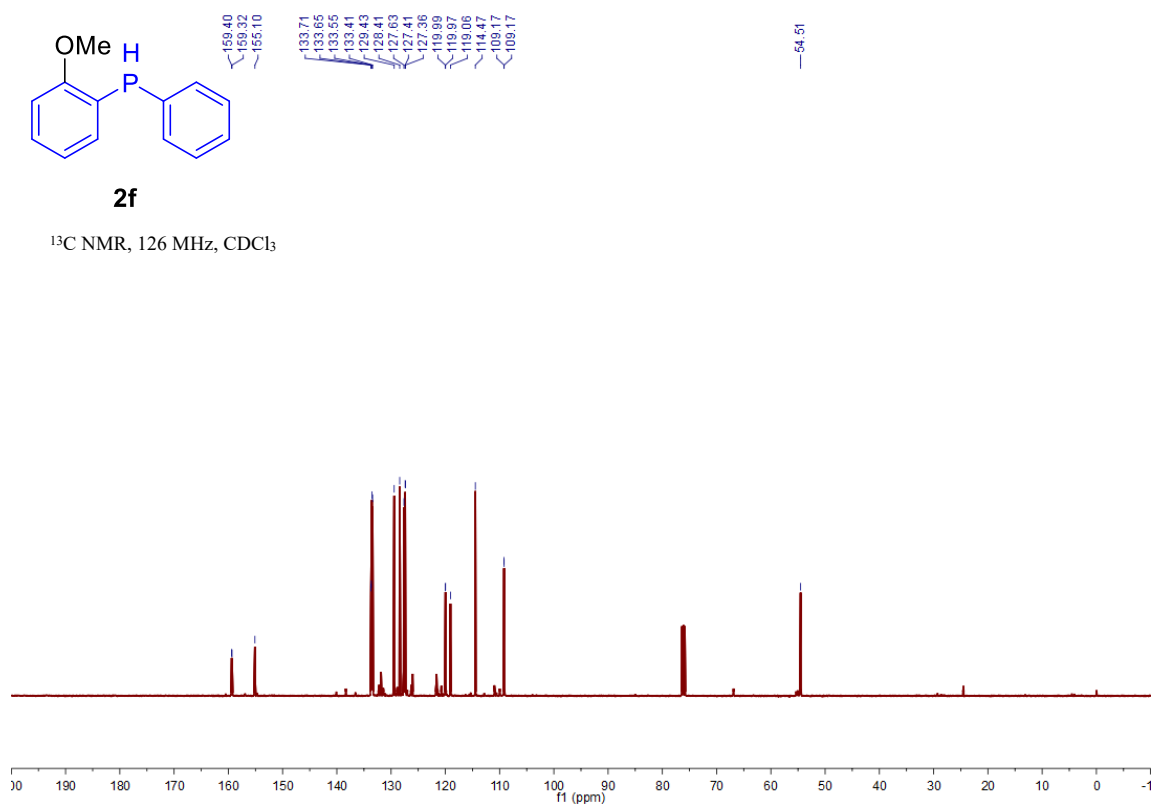


Figure S74. ^{13}C NMR spectrum of **2f** (126 MHz, CDCl_3)

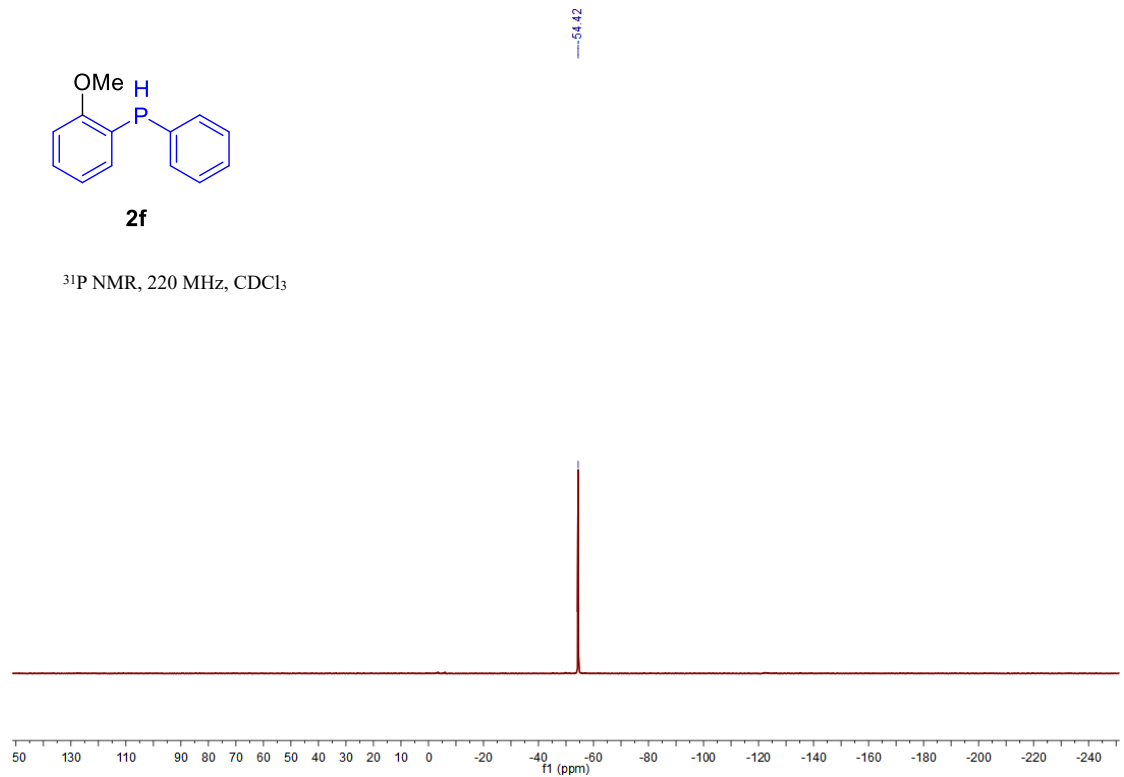


Figure S75. ^{31}P NMR spectrum of **2f** (220 MHz, CDCl_3)

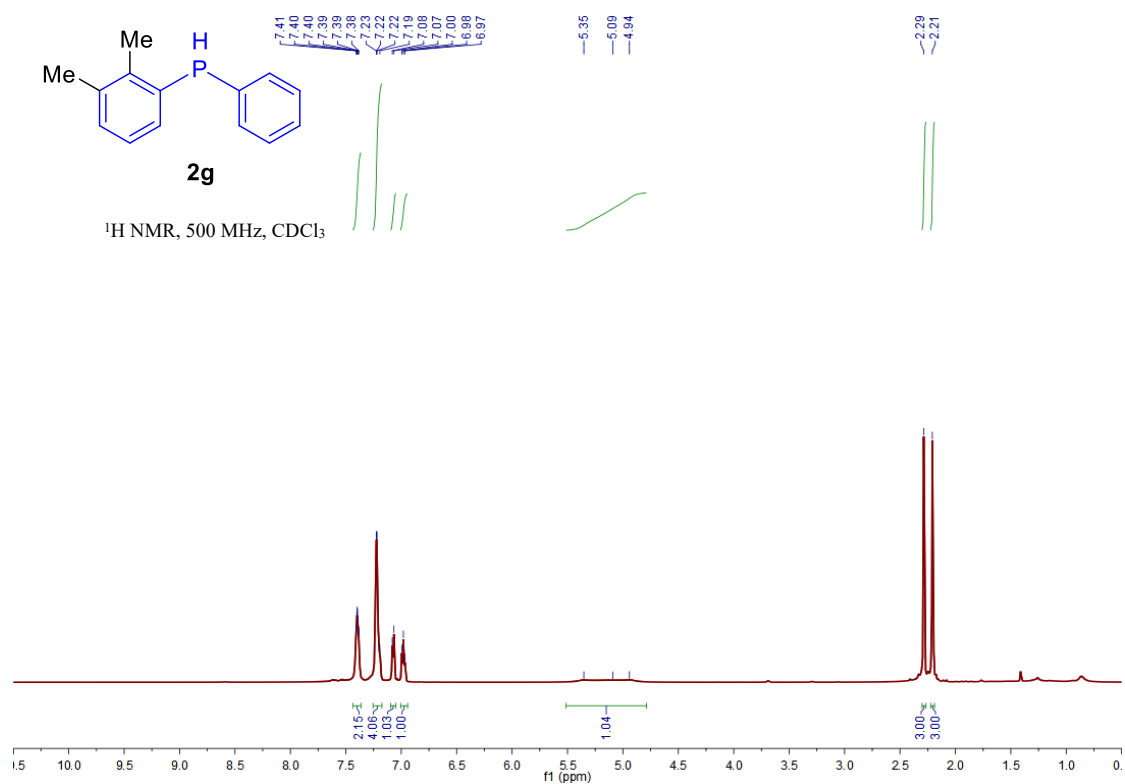


Figure S76. ^1H NMR spectrum of 2g (500 MHz, CDCl_3)

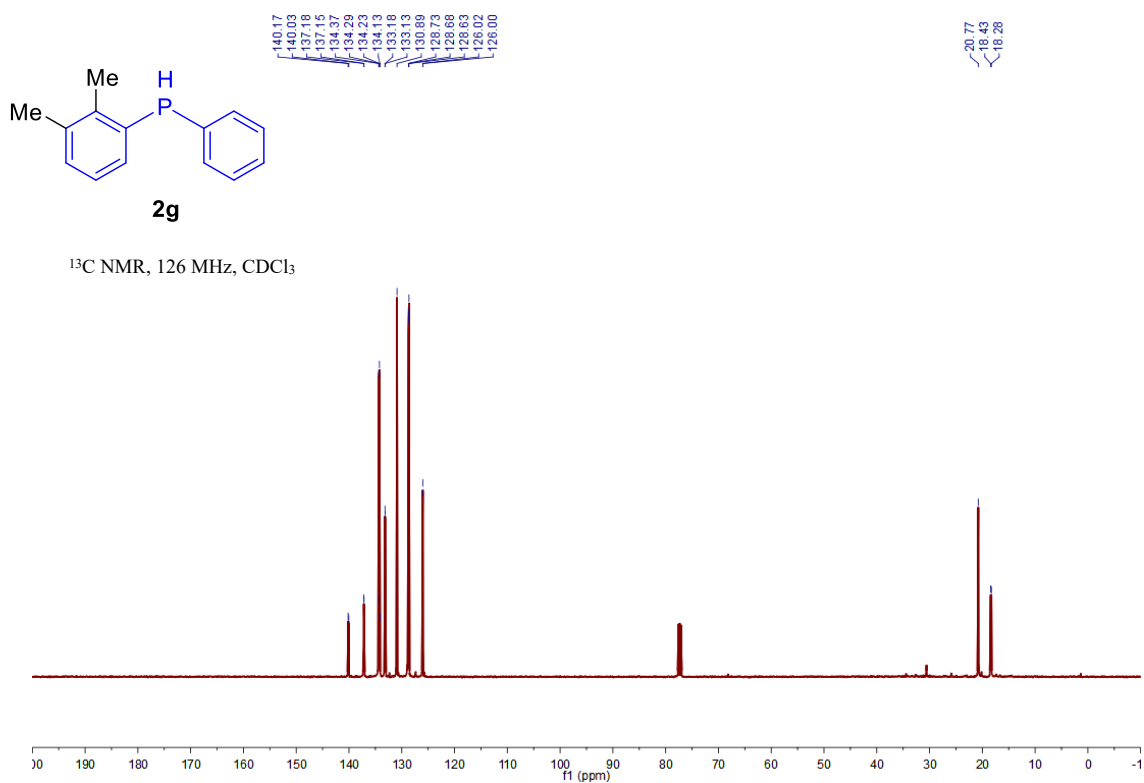


Figure S77. ^{13}C NMR spectrum of 2g (126 MHz, CDCl_3)

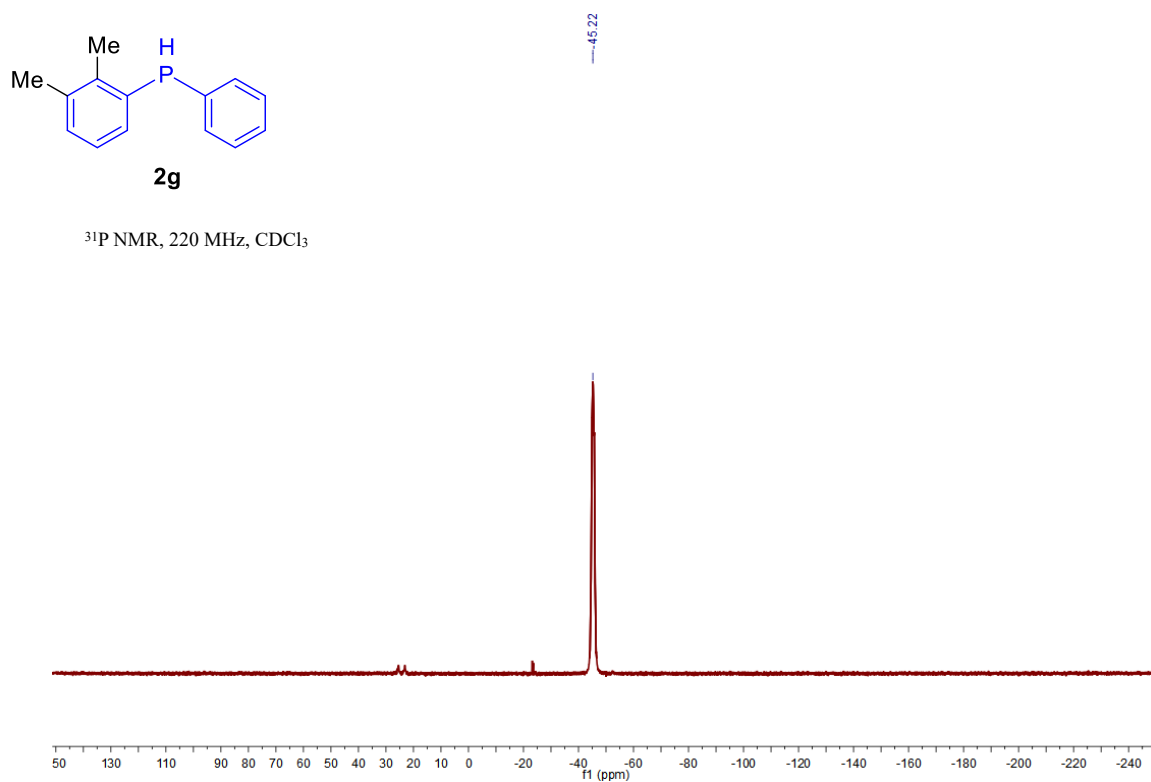


Figure S78. ^{31}P NMR spectrum of **2g** (202 MHz, CDCl_3)

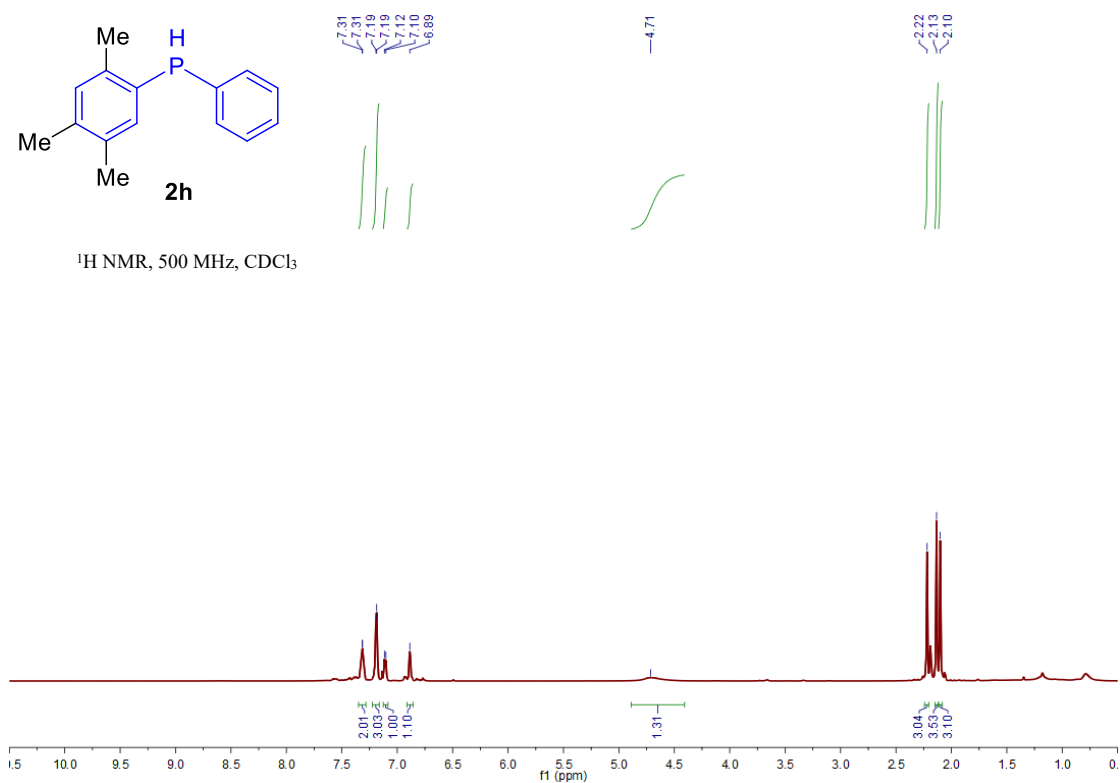


Figure S79. ^1H NMR spectrum of **2h** (500 MHz, CDCl_3)

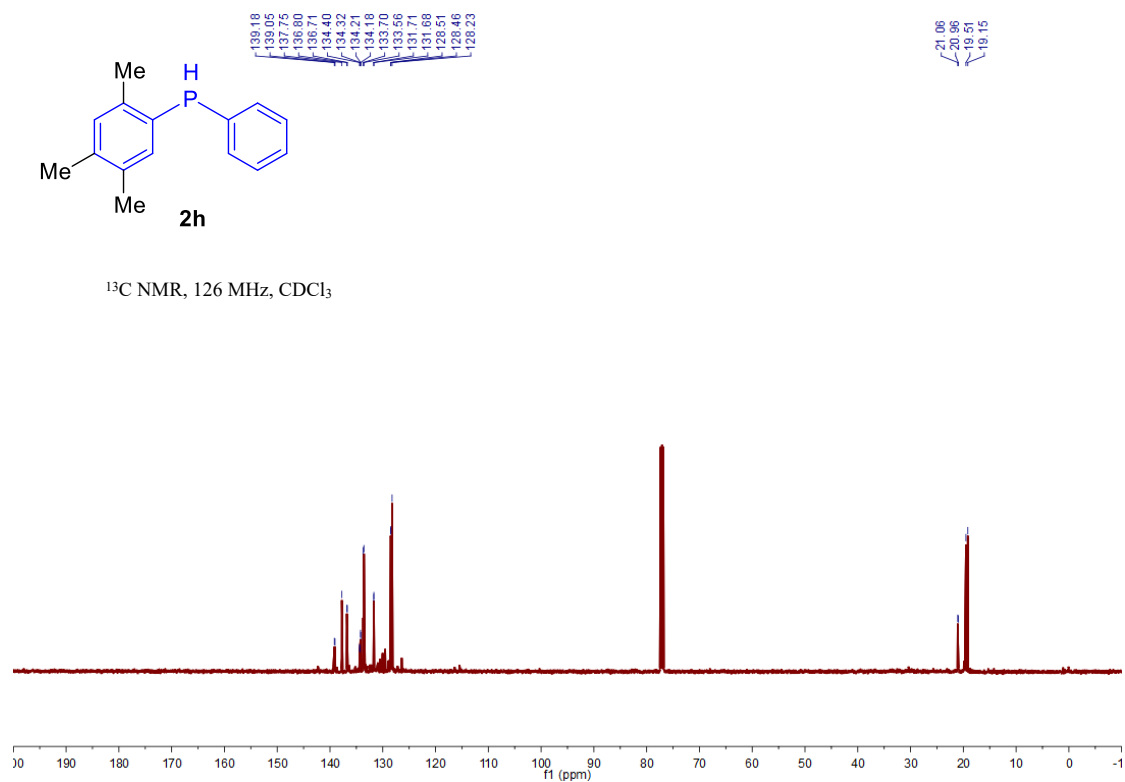


Figure S80. ^{13}C NMR spectrum of 2h (126 MHz, CDCl_3)

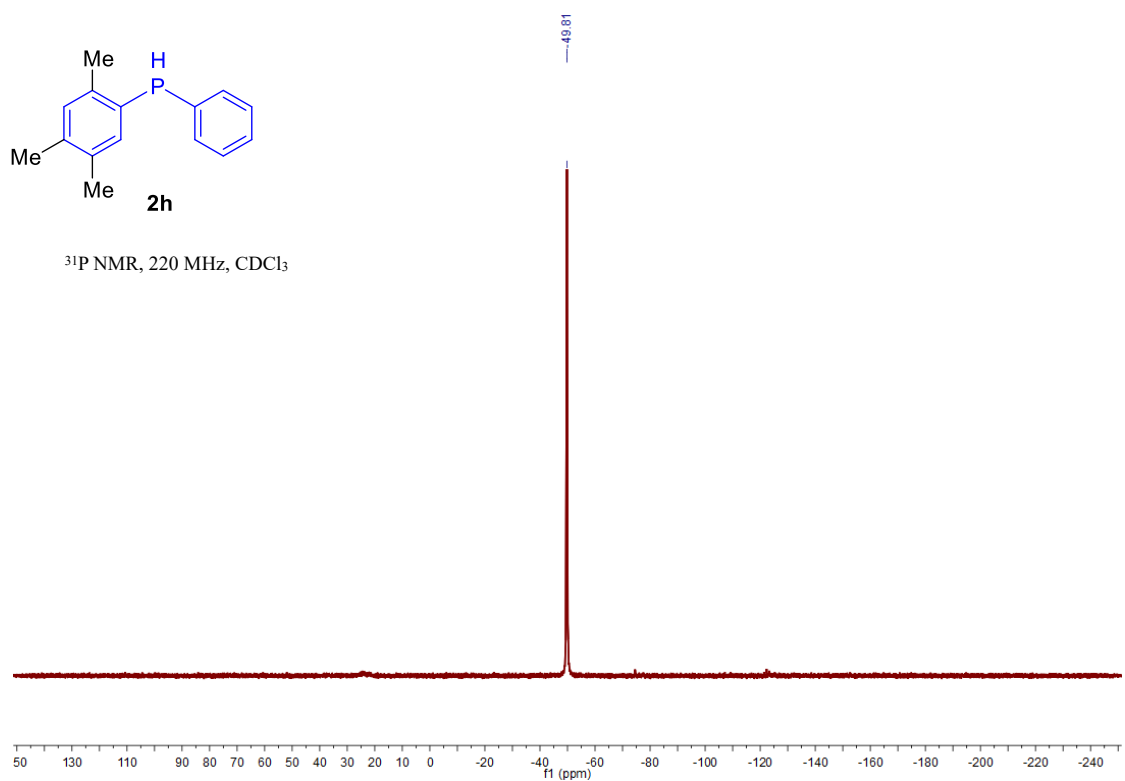


Figure S81. ^{31}P NMR spectrum of 2h (202 MHz, CDCl_3)

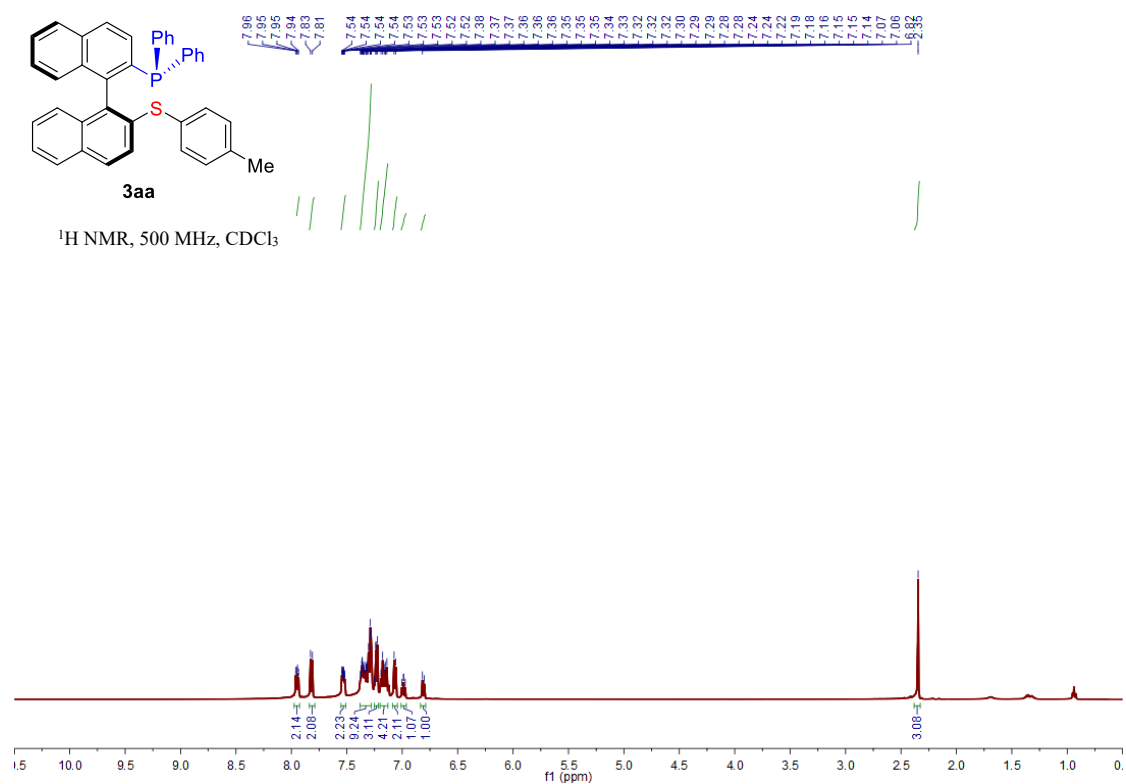


Figure S82. ¹H NMR spectrum of **3aa** (500 MHz, CDCl₃)

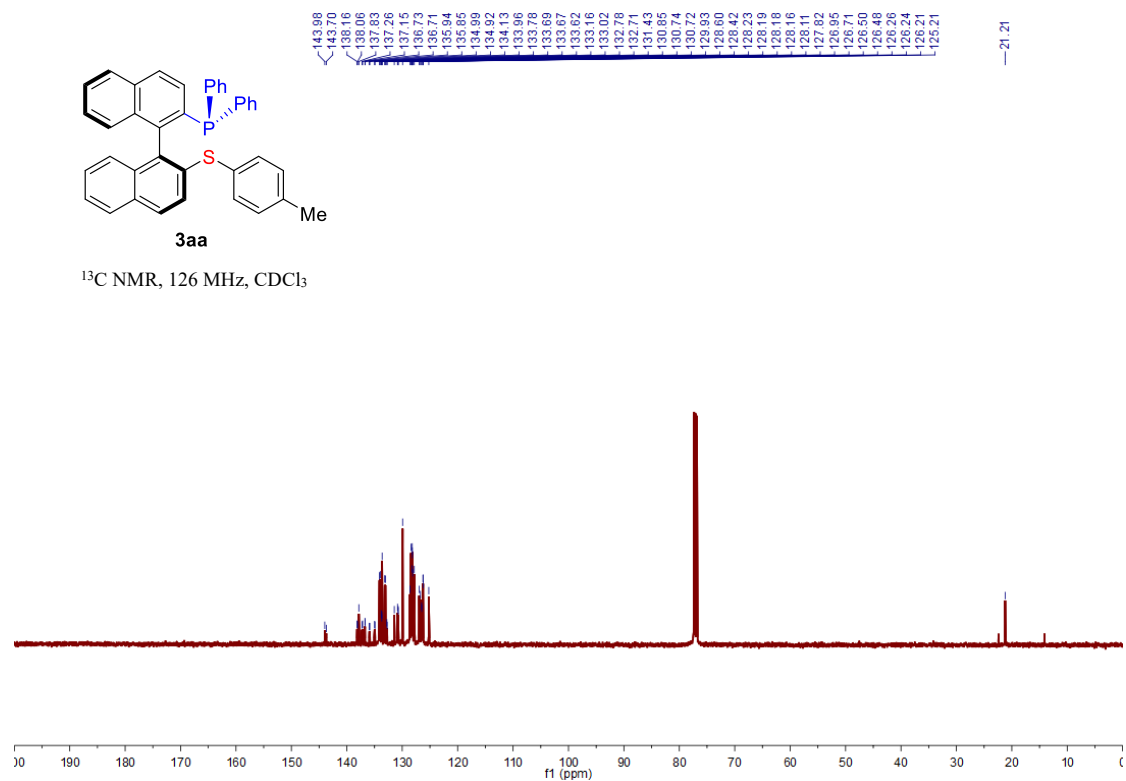


Figure S83. ¹³C NMR spectrum of **3aa** (126 MHz, CDCl₃)

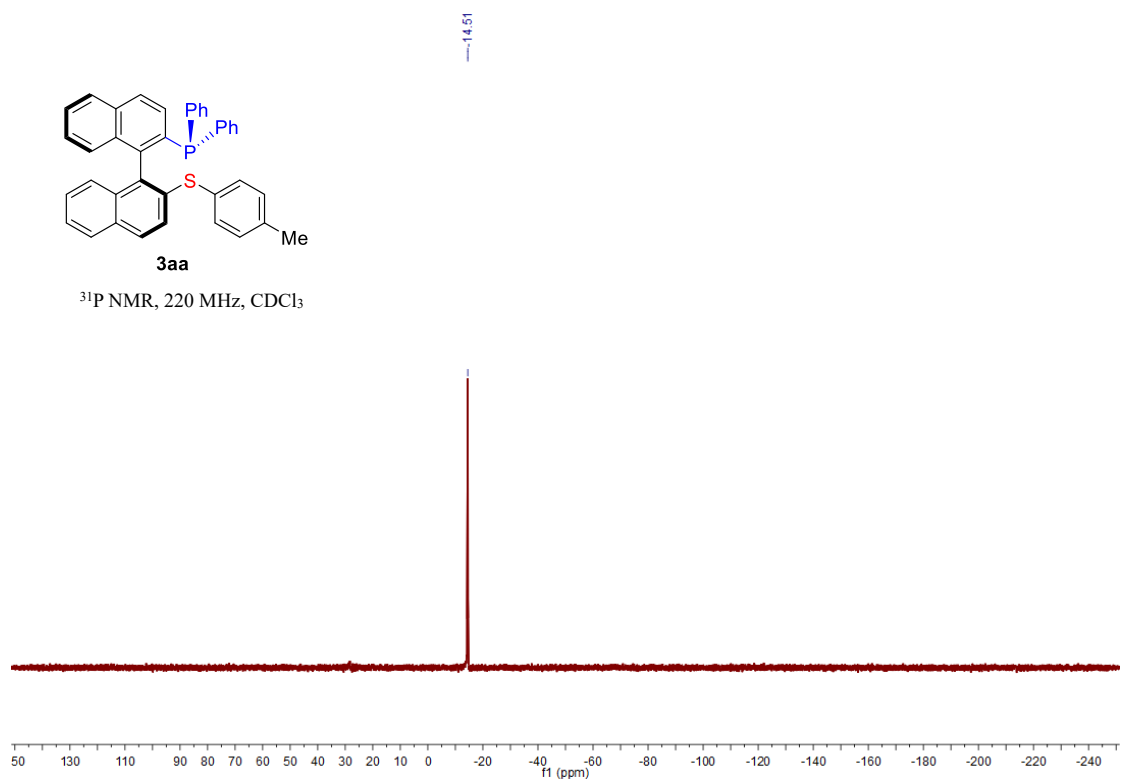


Figure S84. ³¹P NMR spectrum of 3aa (202 MHz, CDCl₃)

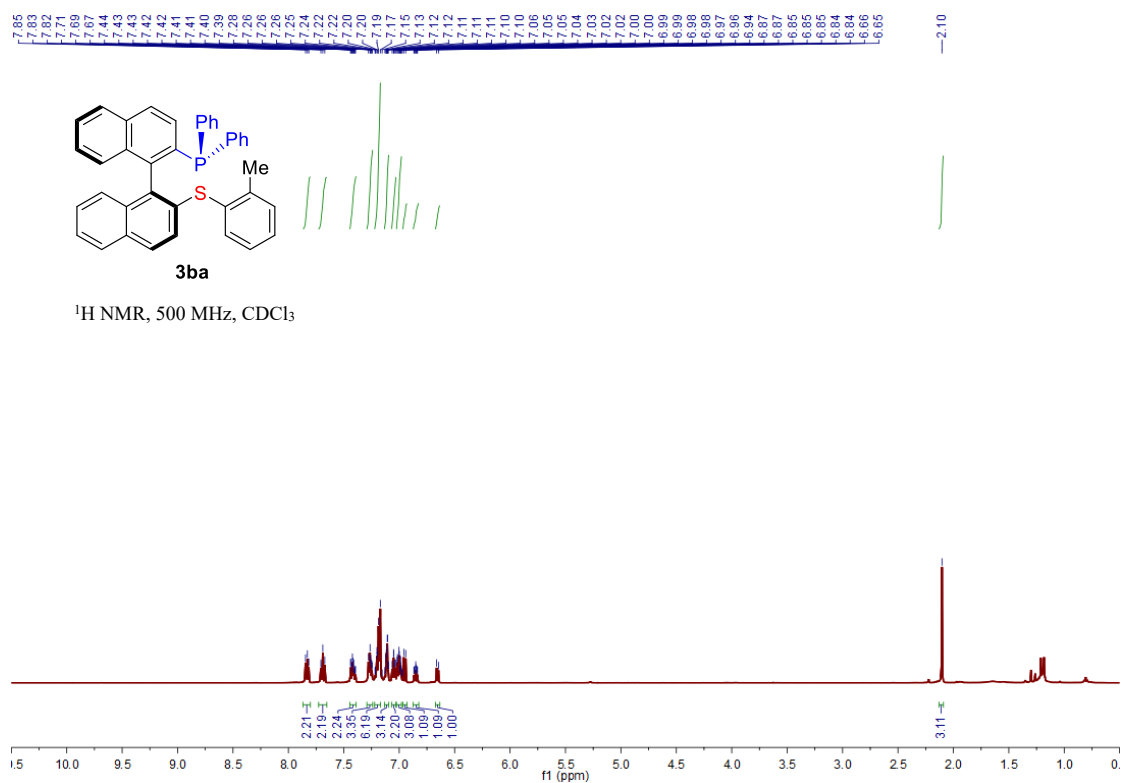
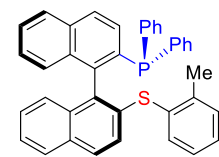
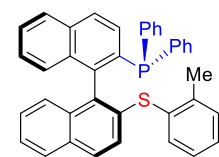


Figure S85. ¹H NMR spectrum of 3ba (500 MHz, CDCl₃)



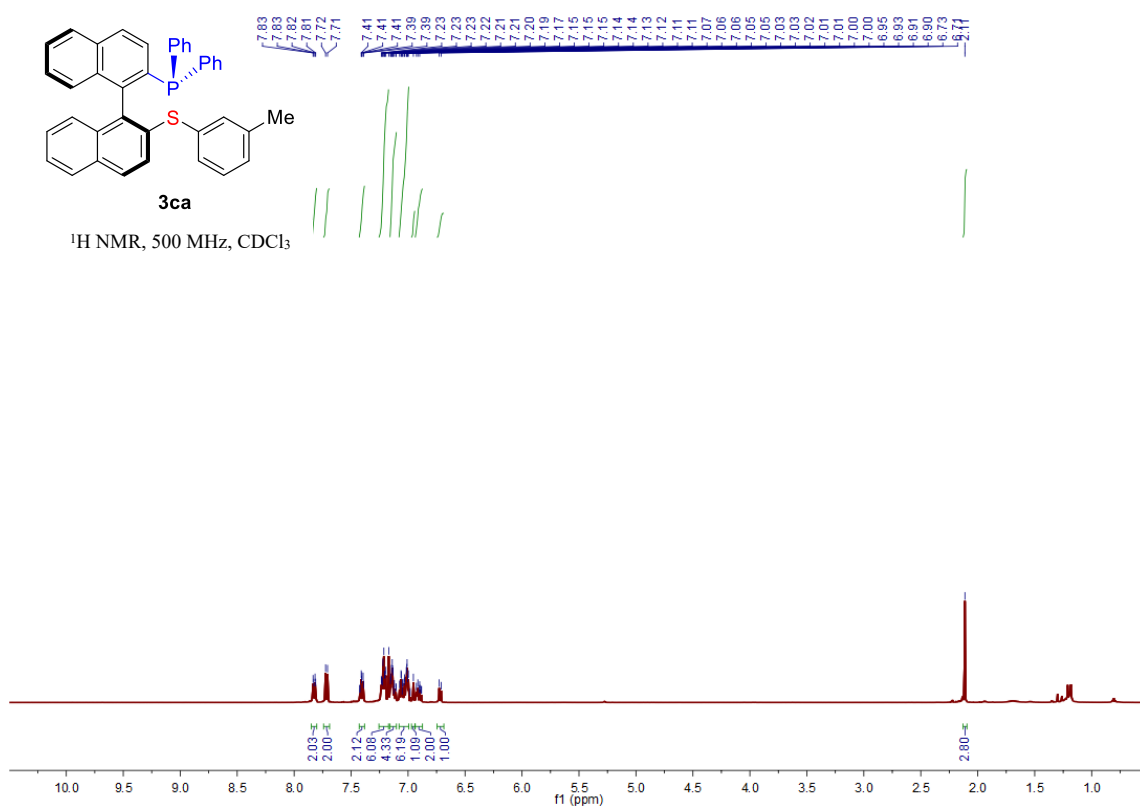


Figure S88. ¹H NMR spectrum of **3ca** (500 MHz, CDCl₃)

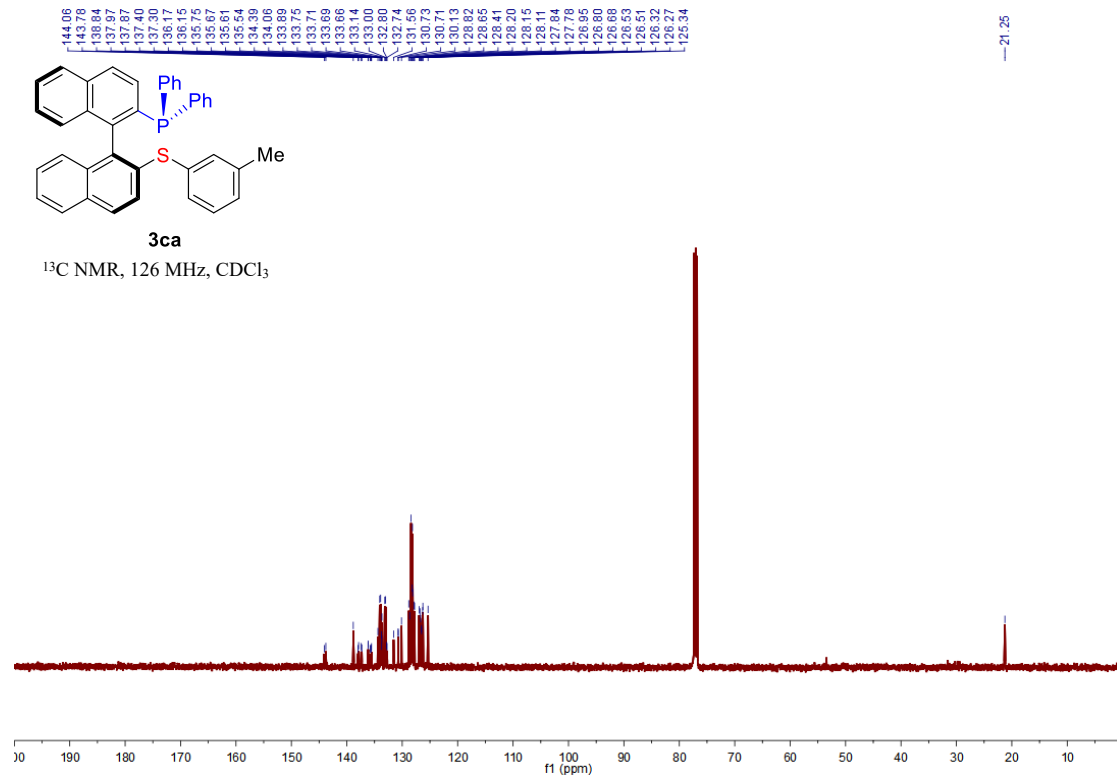


Figure S89. ¹³C NMR spectrum of **3ca** (126 MHz, CDCl₃)

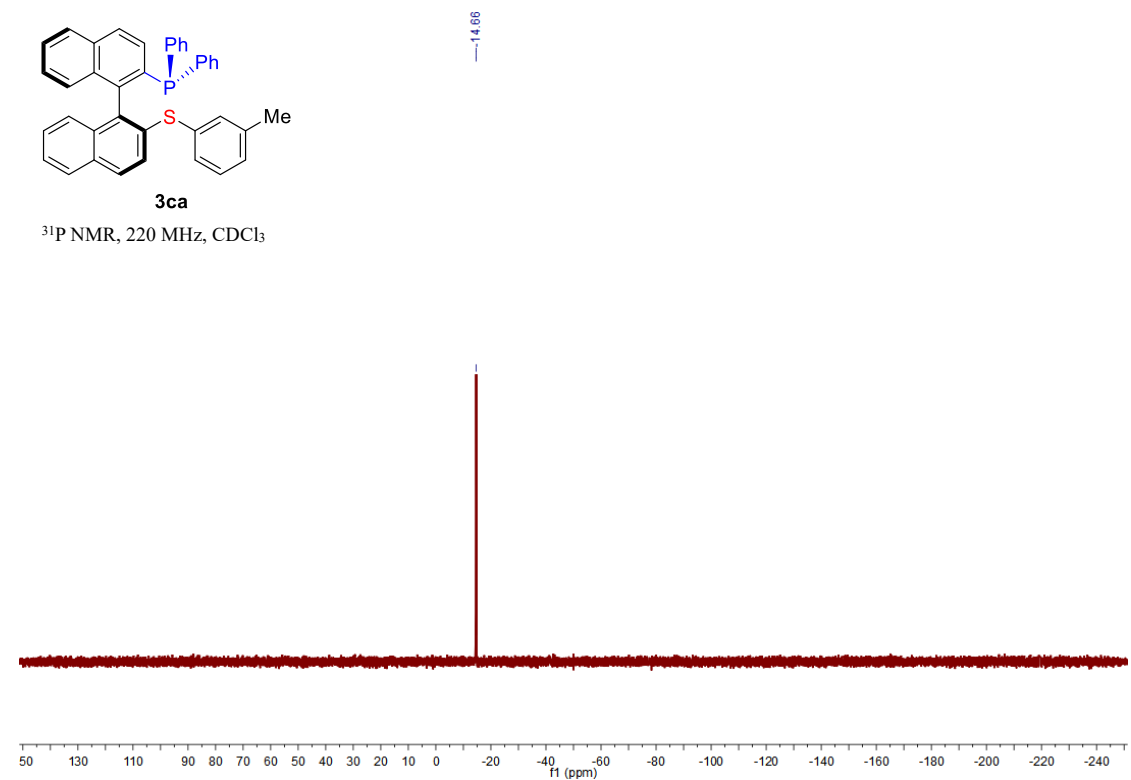


Figure S90. ^{31}P NMR spectrum of 3ca (202 MHz, CDCl_3)

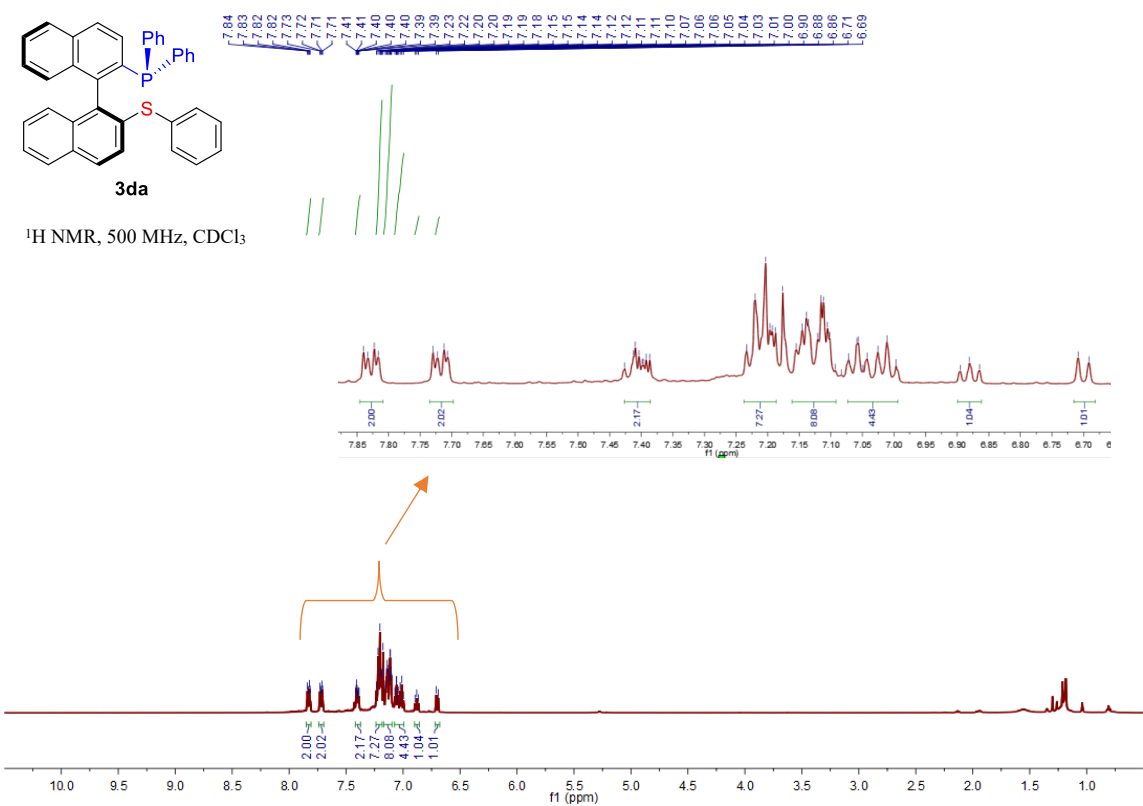


Figure S91. ^1H NMR spectrum of 3da (500 MHz, CDCl_3)

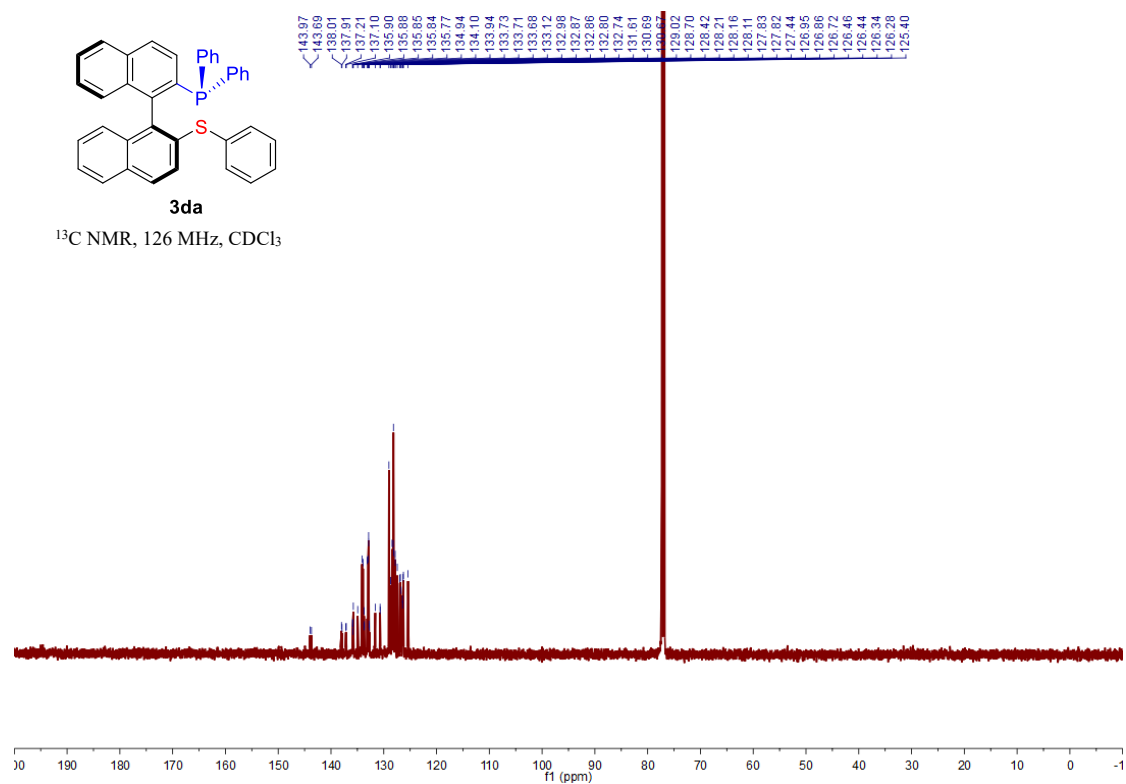


Figure S92. ^{13}C NMR spectrum of 3da (126 MHz, CDCl_3)

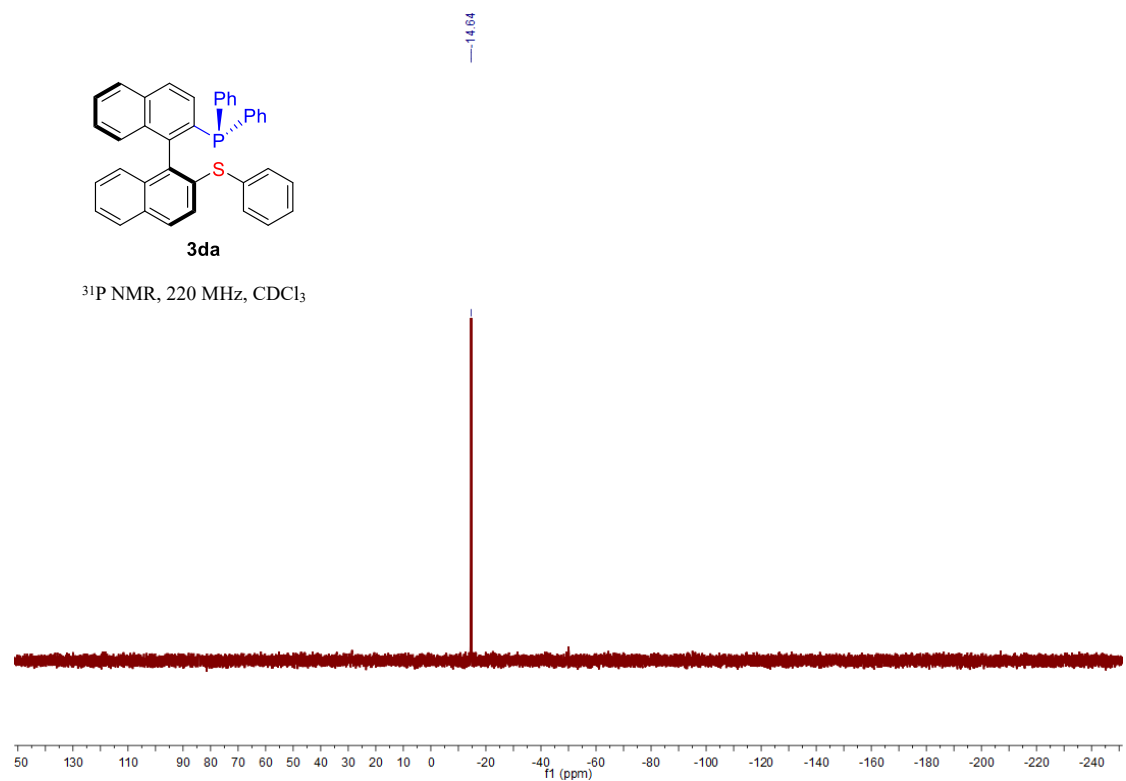


Figure S93. ^{31}P NMR spectrum of 3da (202 MHz, CDCl_3)

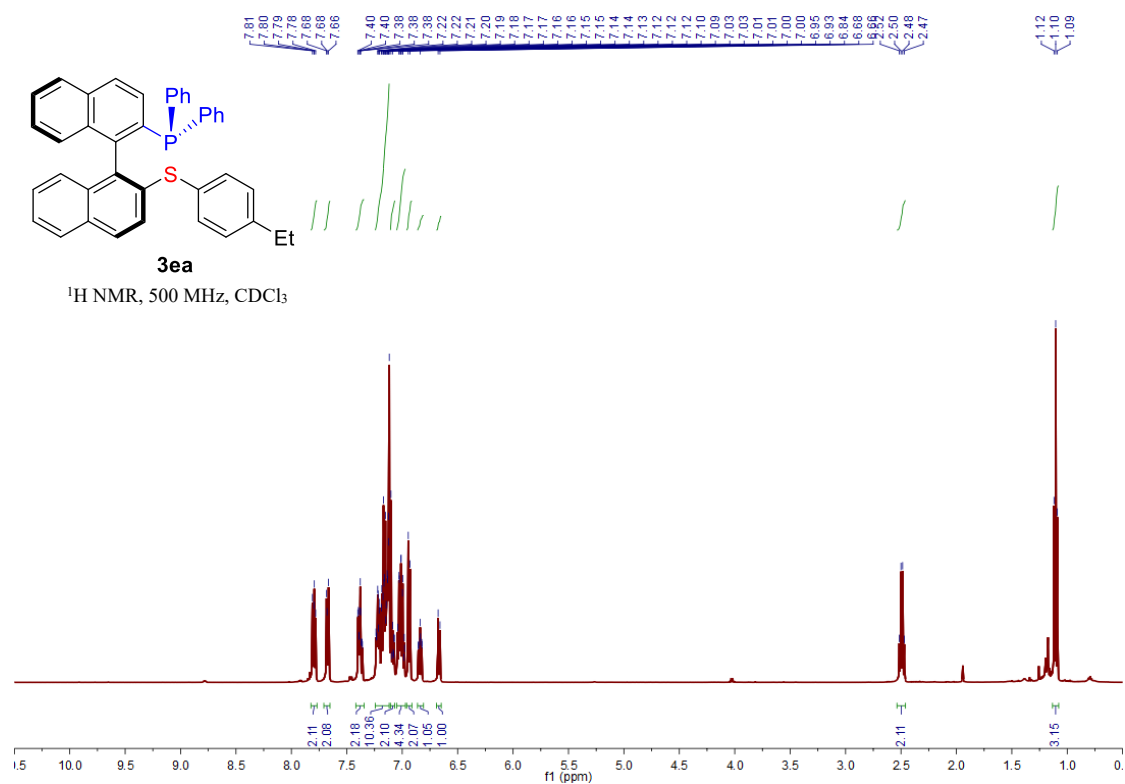


Figure S94. ¹H NMR spectrum of **3ea** (500 MHz, CDCl₃)

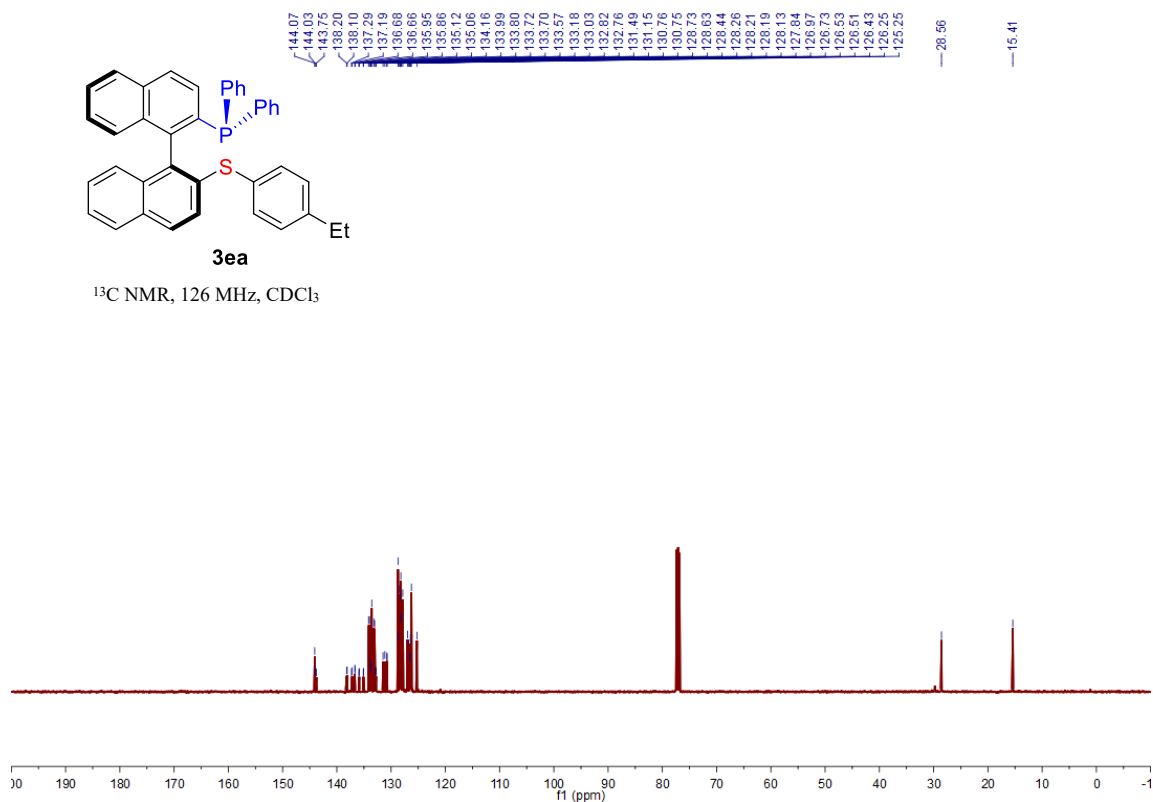


Figure S95. ¹³C NMR spectrum of **3ea** (126 MHz, CDCl₃)

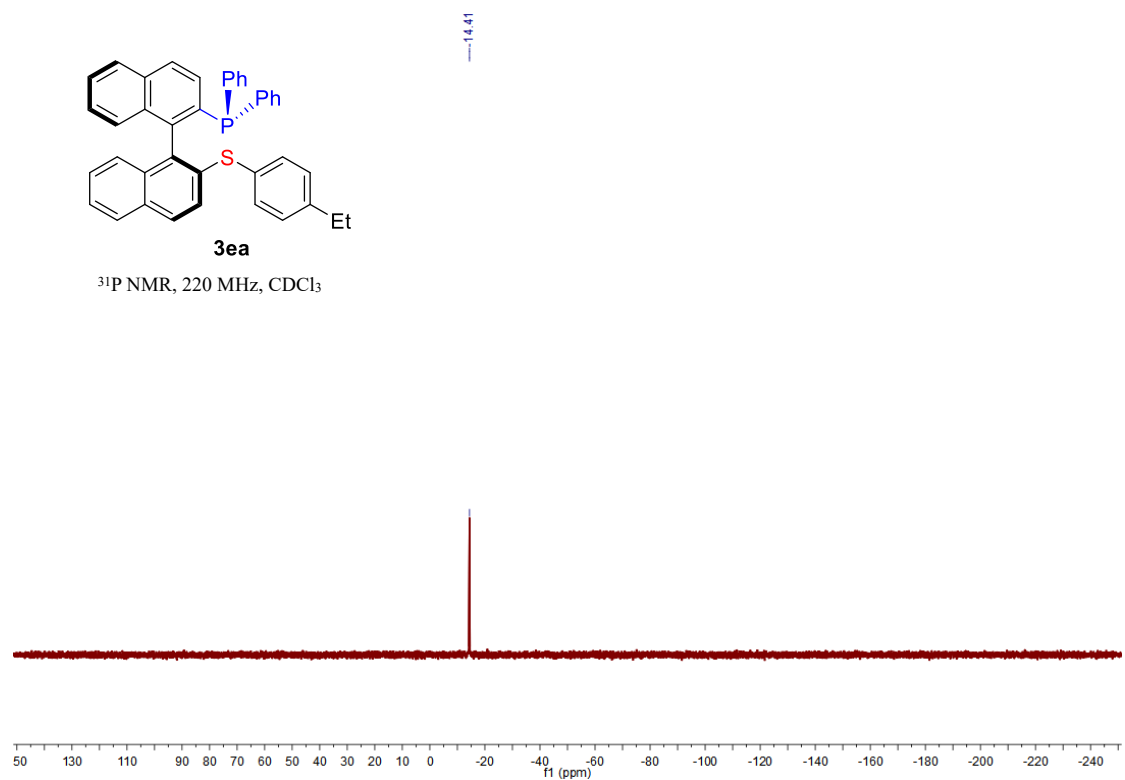


Figure S96. ^{31}P NMR spectrum of **3ea** (202 MHz, CDCl_3)

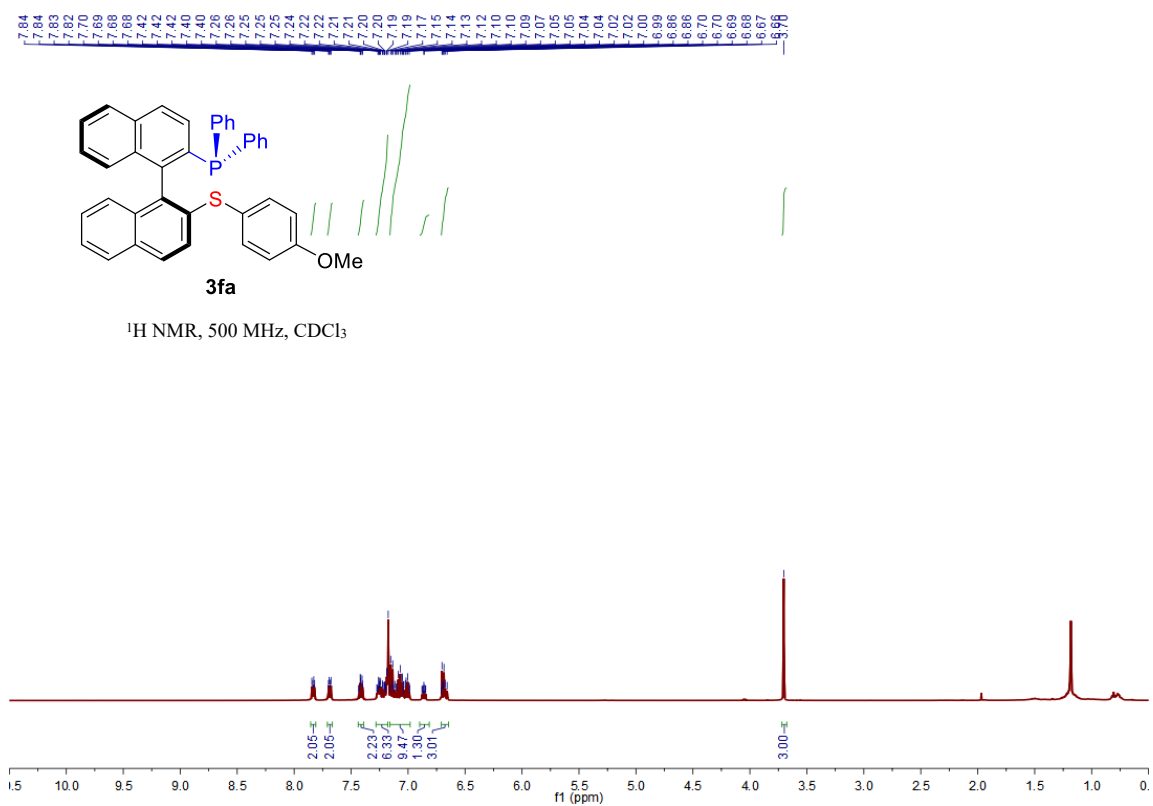


Figure S97. ^1H NMR spectrum of **3fa** (500 MHz, CDCl_3)

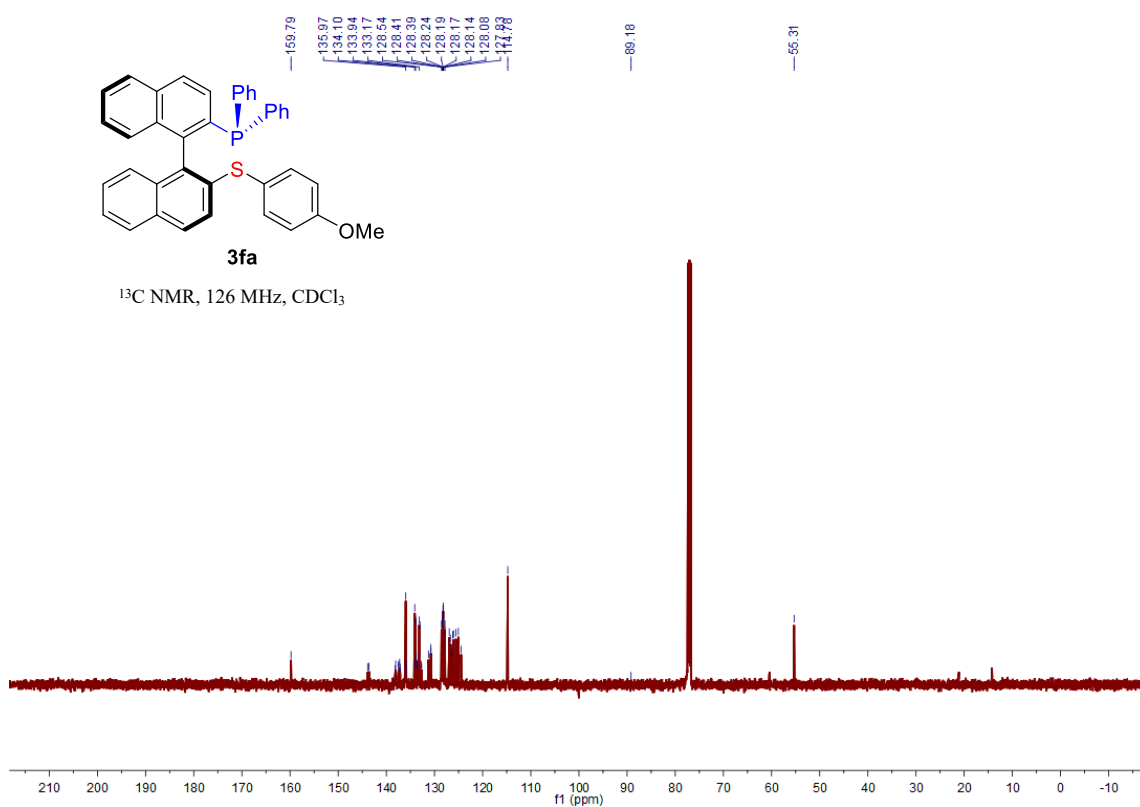


Figure S98. ¹³C NMR spectrum of **3fa** (126 MHz, CDCl₃)

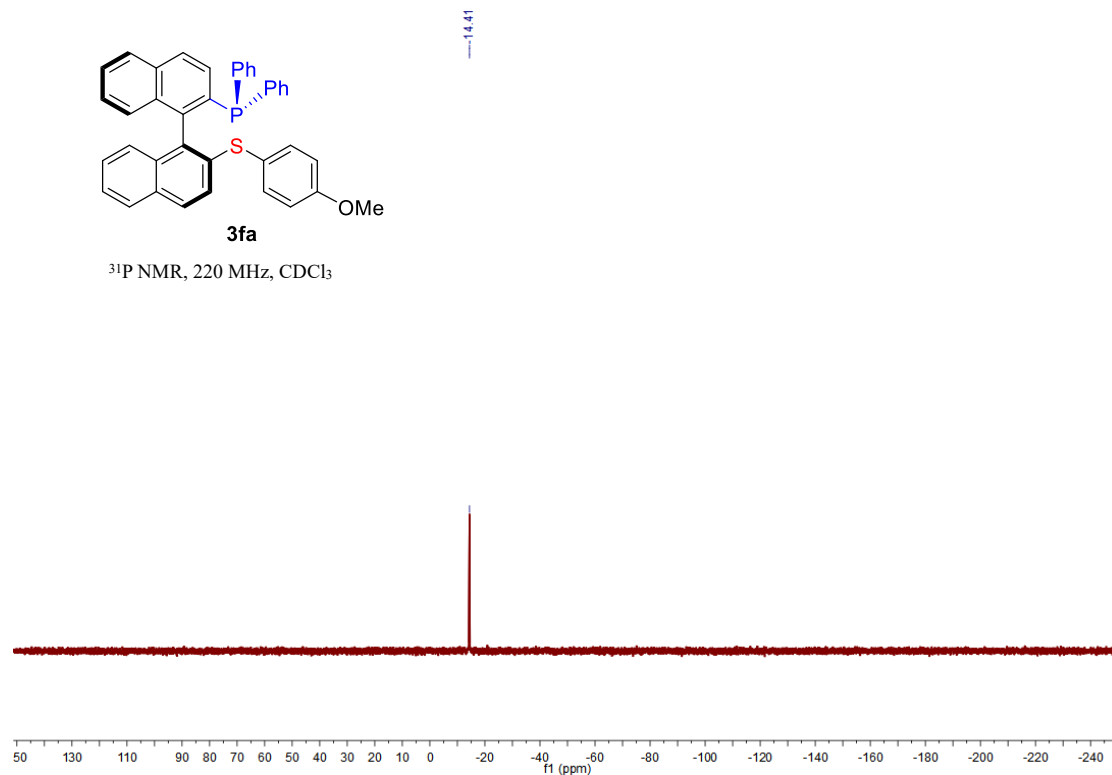


Figure S99. ³¹P NMR spectrum of **3fa** (202 MHz, CDCl₃)

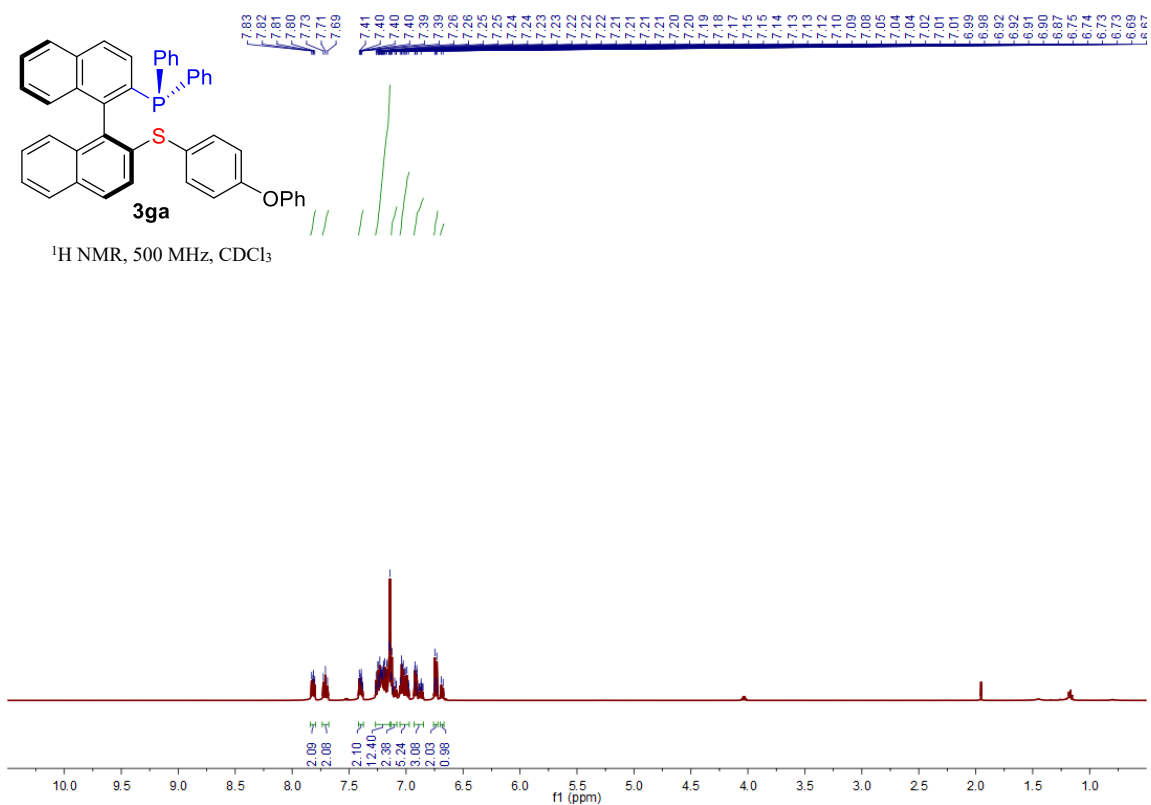


Figure S100. ¹H NMR spectrum of **3ga** (500 MHz, CDCl₃)

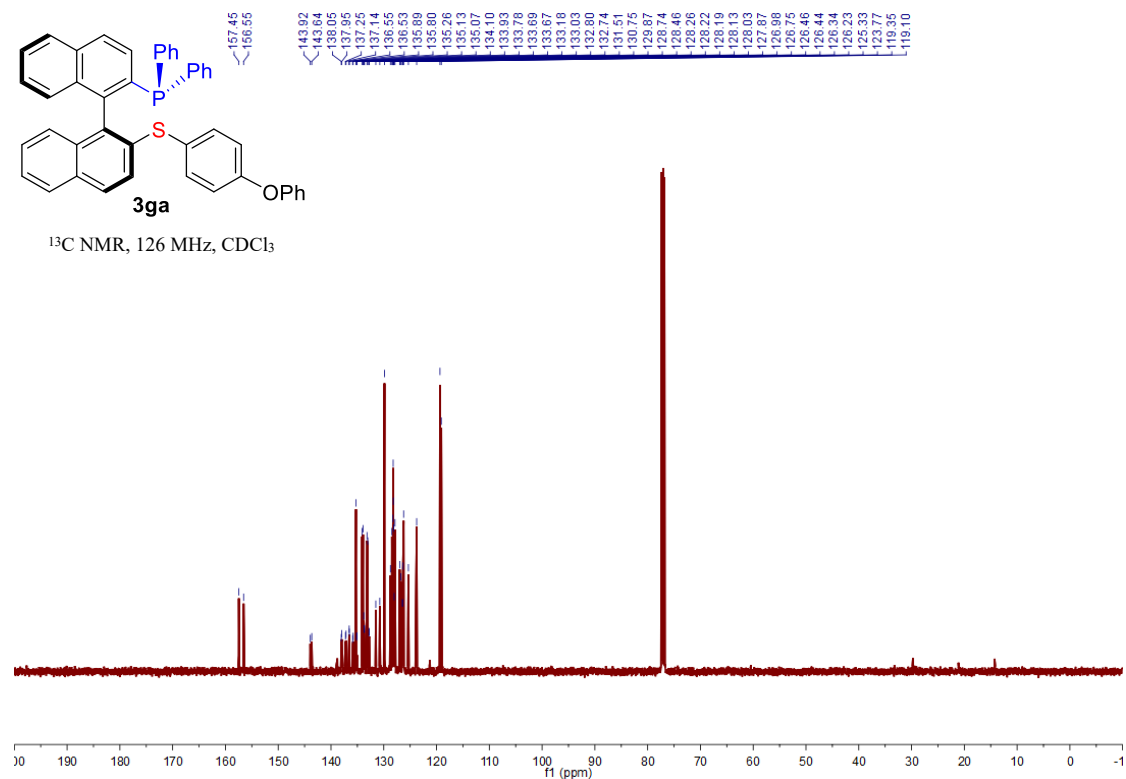


Figure S101. ¹³C NMR spectrum of **3ga** (126 MHz, CDCl₃)

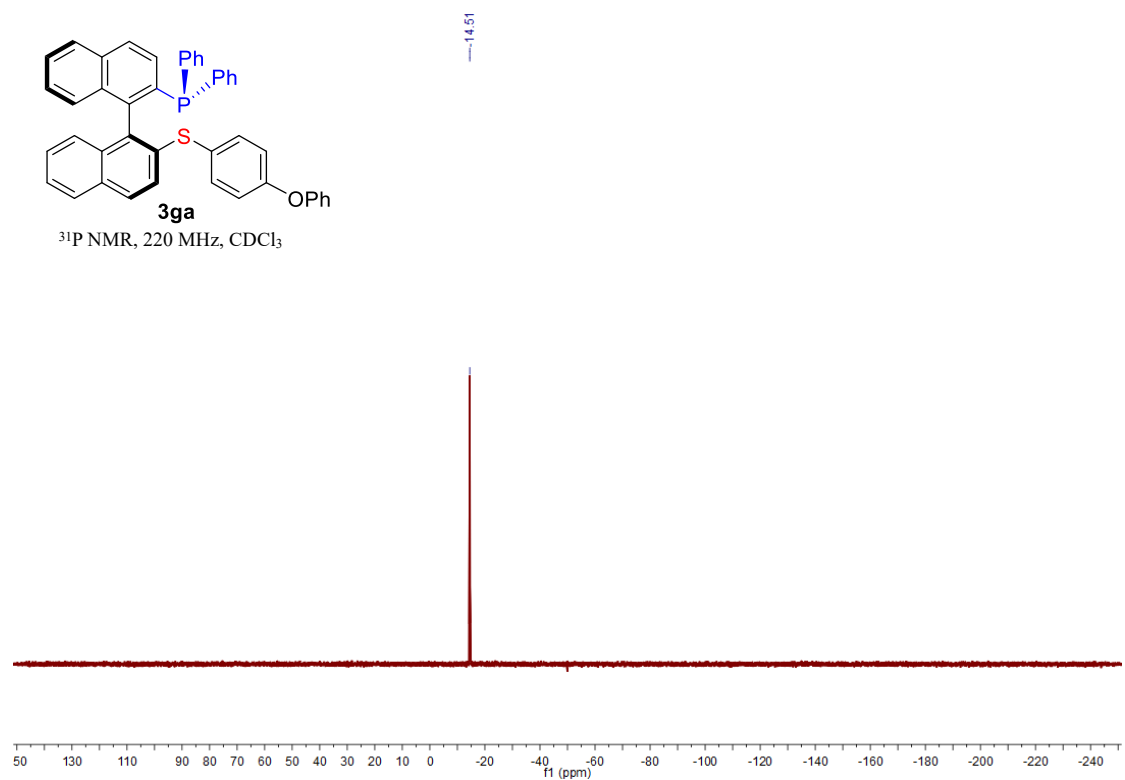


Figure S102. ^{31}P NMR spectrum of **3ga** (202 MHz, CDCl_3)

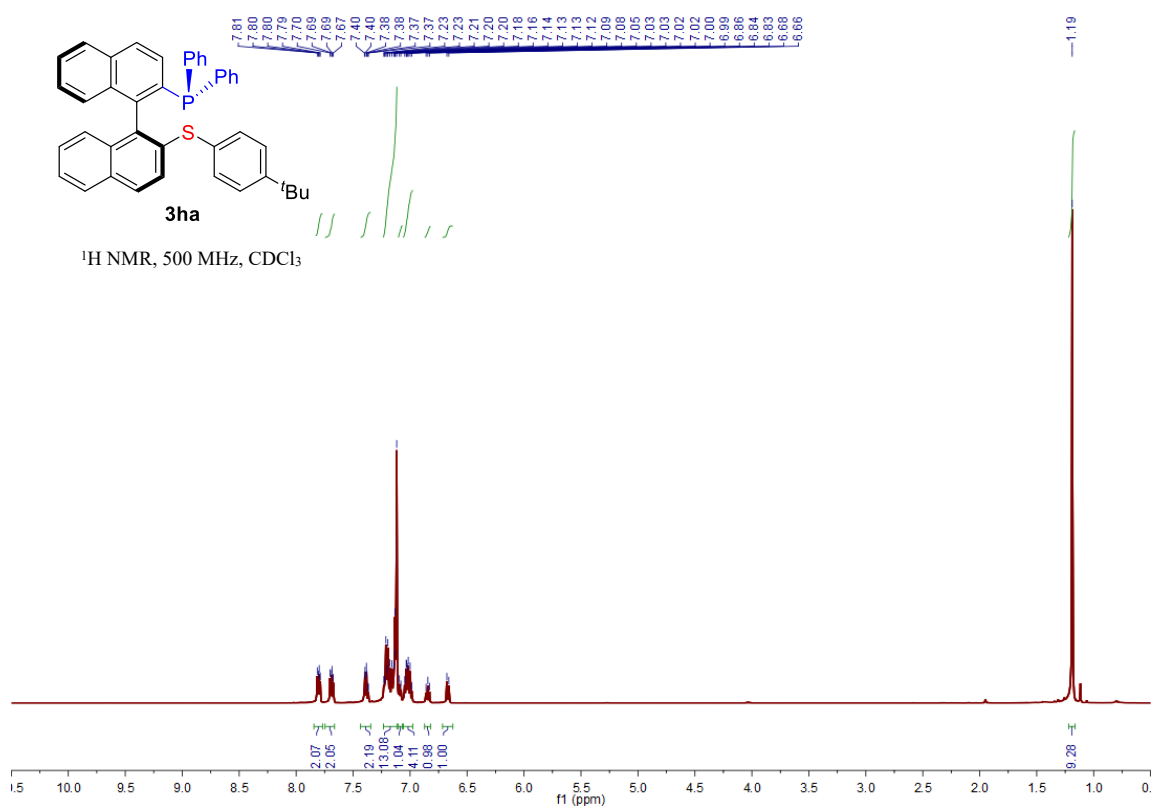


Figure S103. ^1H NMR spectrum of **3ha** (500 MHz, CDCl_3)

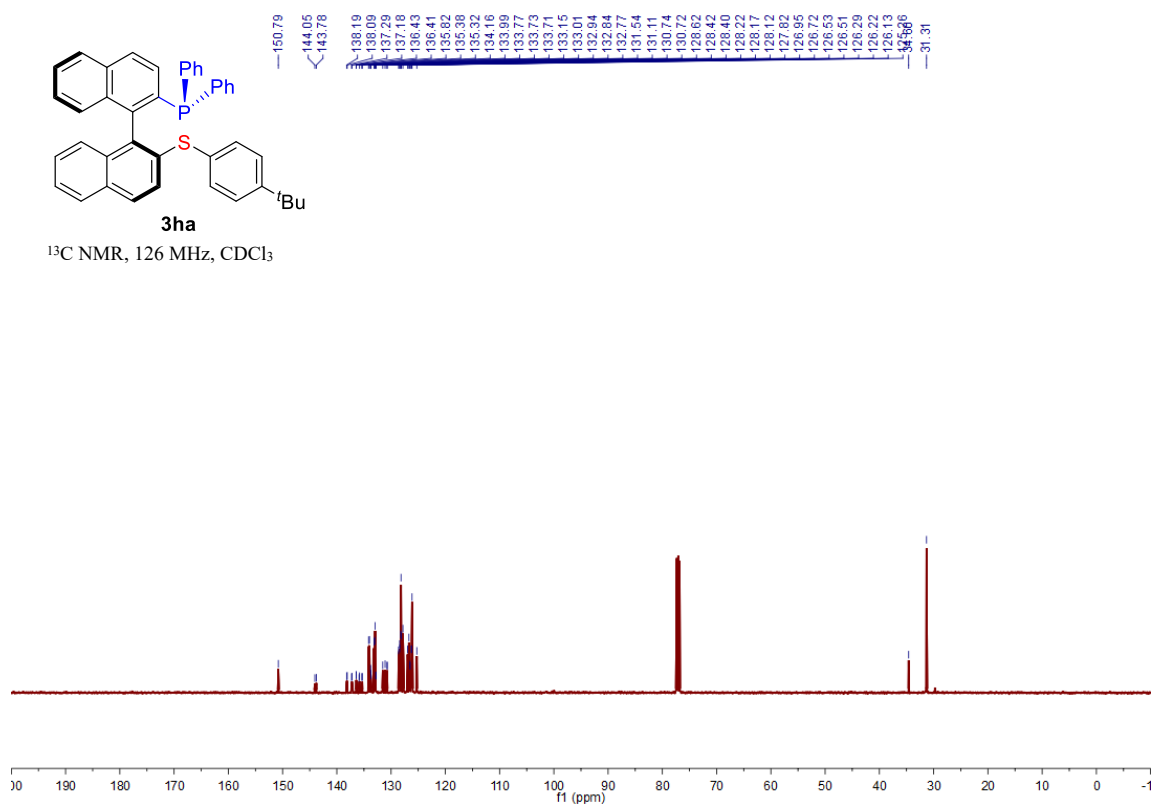


Figure S104. ^{13}C NMR spectrum of **3ha** (126 MHz, CDCl_3)

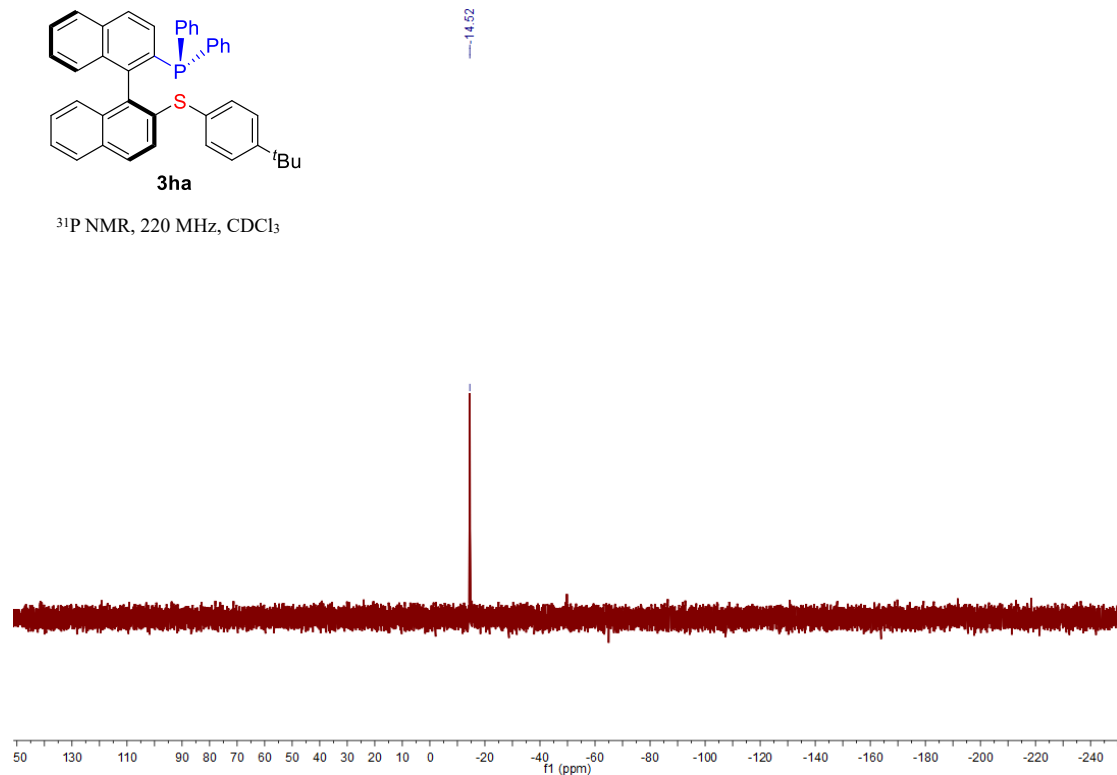


Figure S105. ^{31}P NMR spectrum of **3ha** (202 MHz, CDCl_3)

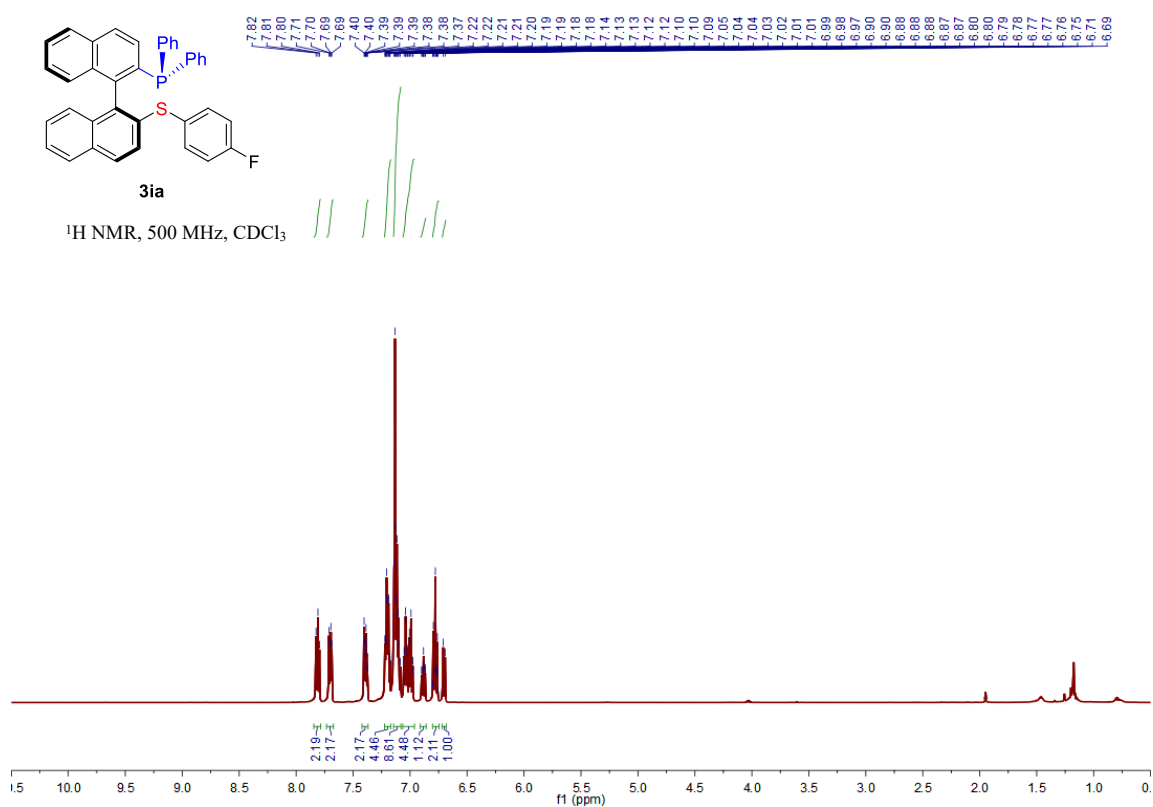


Figure S106. ¹H NMR spectrum of 3ia (500 MHz, CDCl₃)

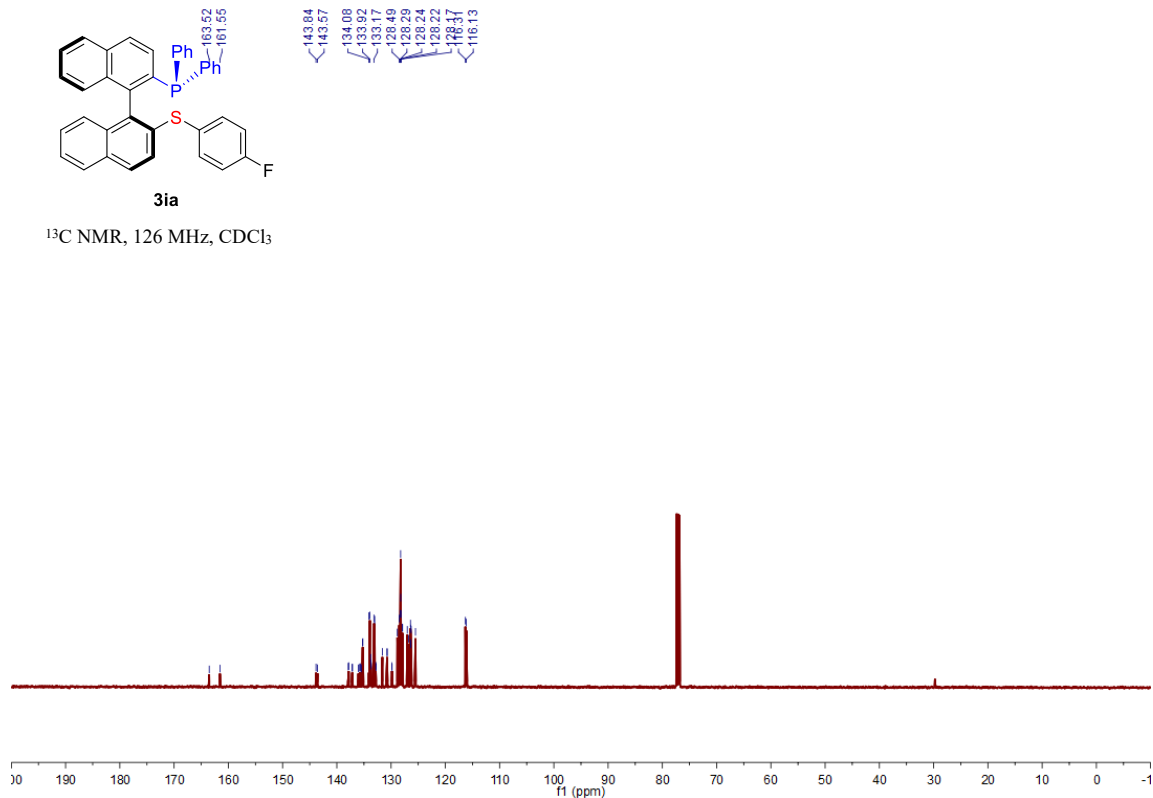


Figure S107. ¹³C NMR spectrum of 3ia (126 MHz, CDCl₃)

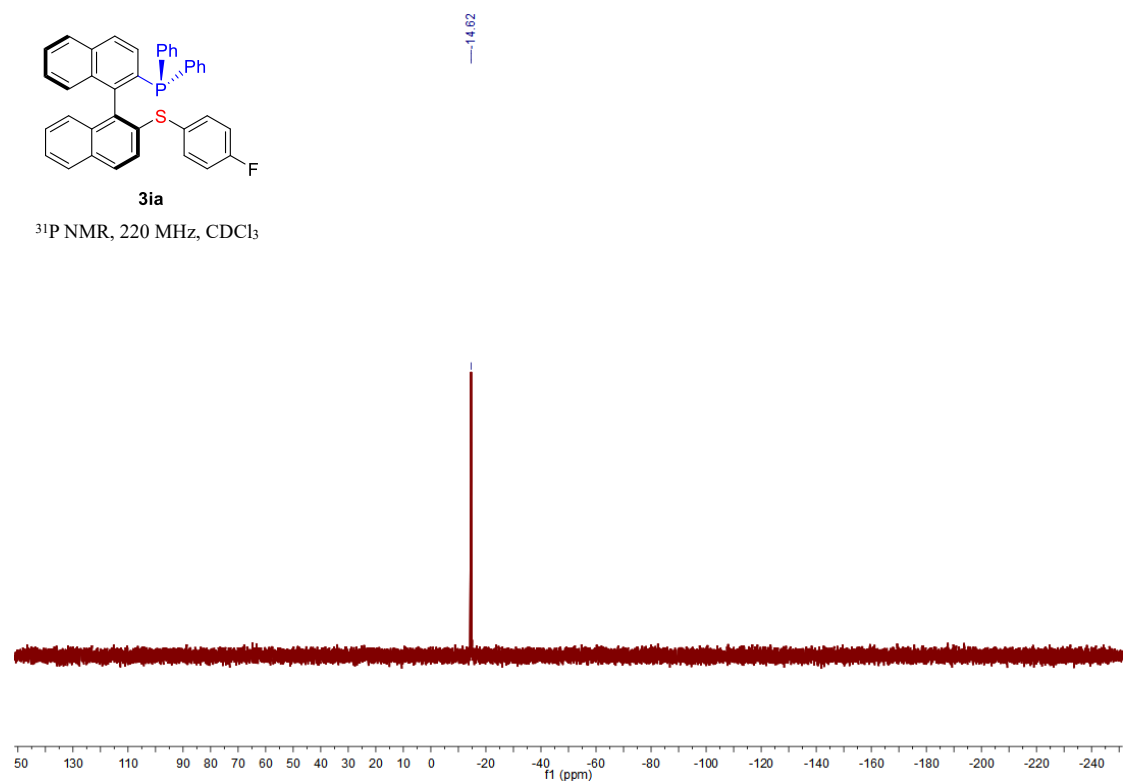


Figure S108. ^{31}P NMR spectrum of 3ia (202 MHz, CDCl_3)

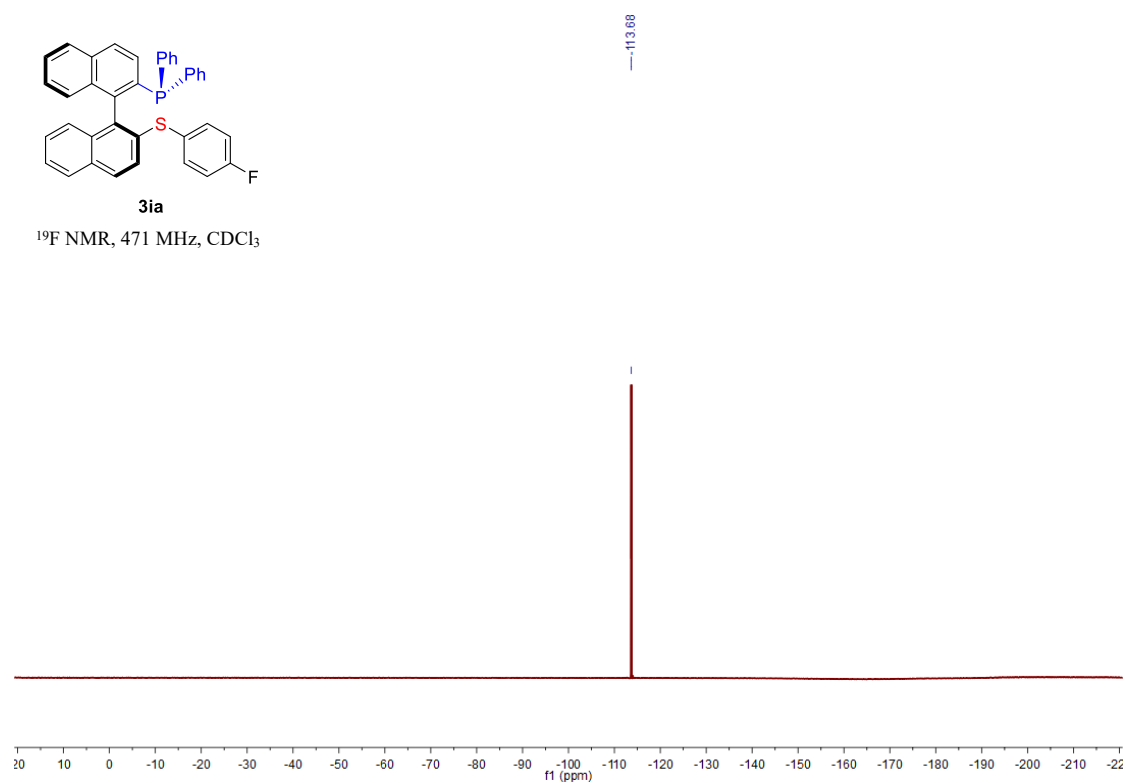


Figure S109. ^{19}F NMR spectrum of 3ia (471 MHz, CDCl_3)

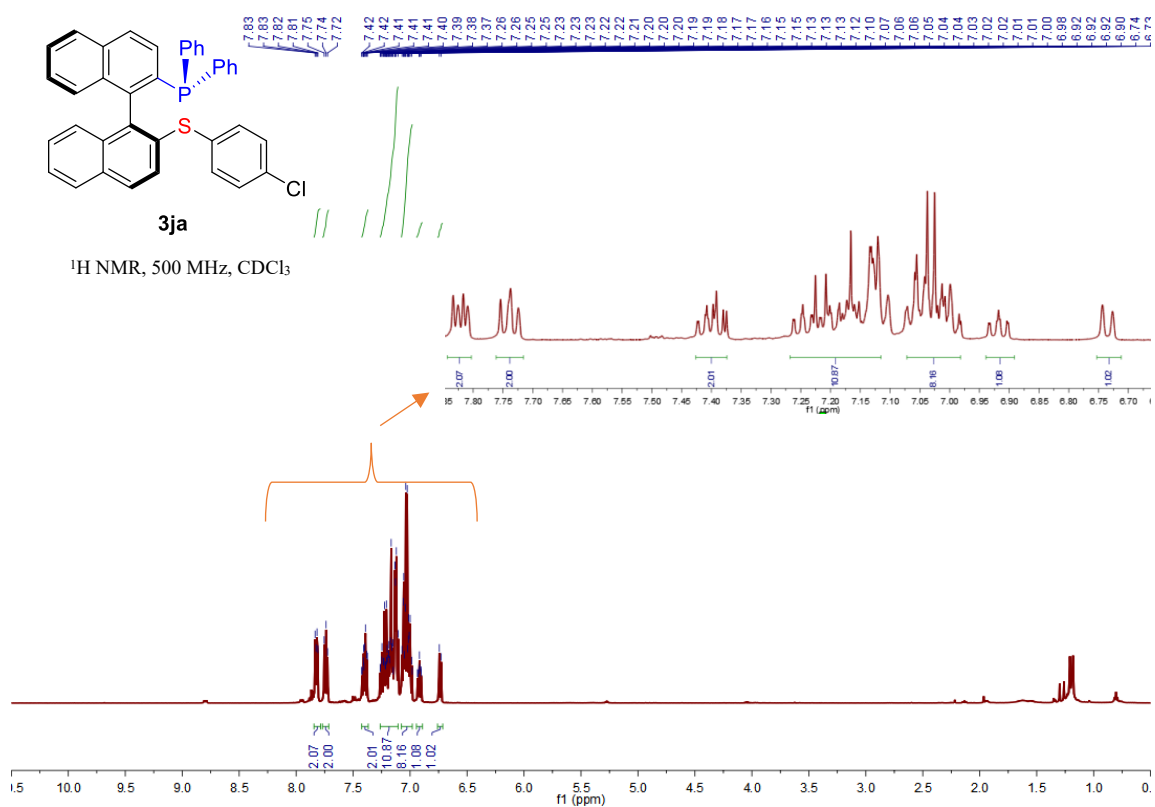


Figure S110. ¹H NMR spectrum of **3ja** (500 MHz, CDCl₃)

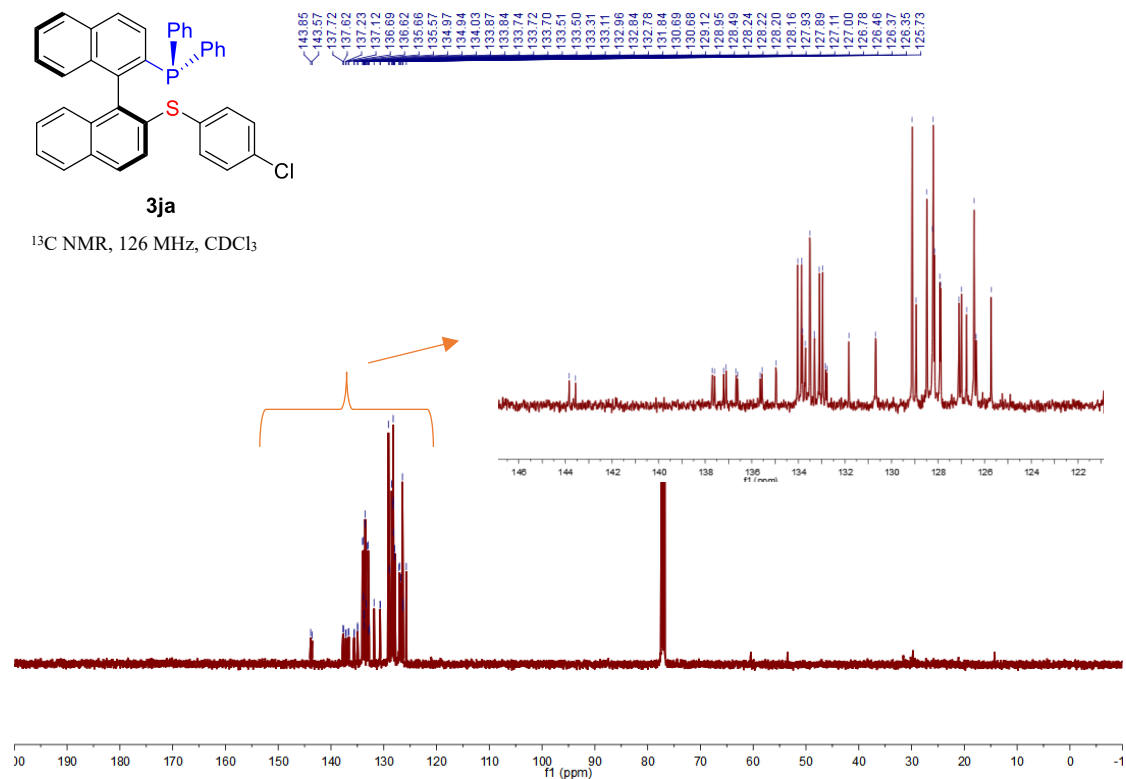


Figure S111. ¹³C NMR spectrum of **3ja** (126 MHz, CDCl₃)

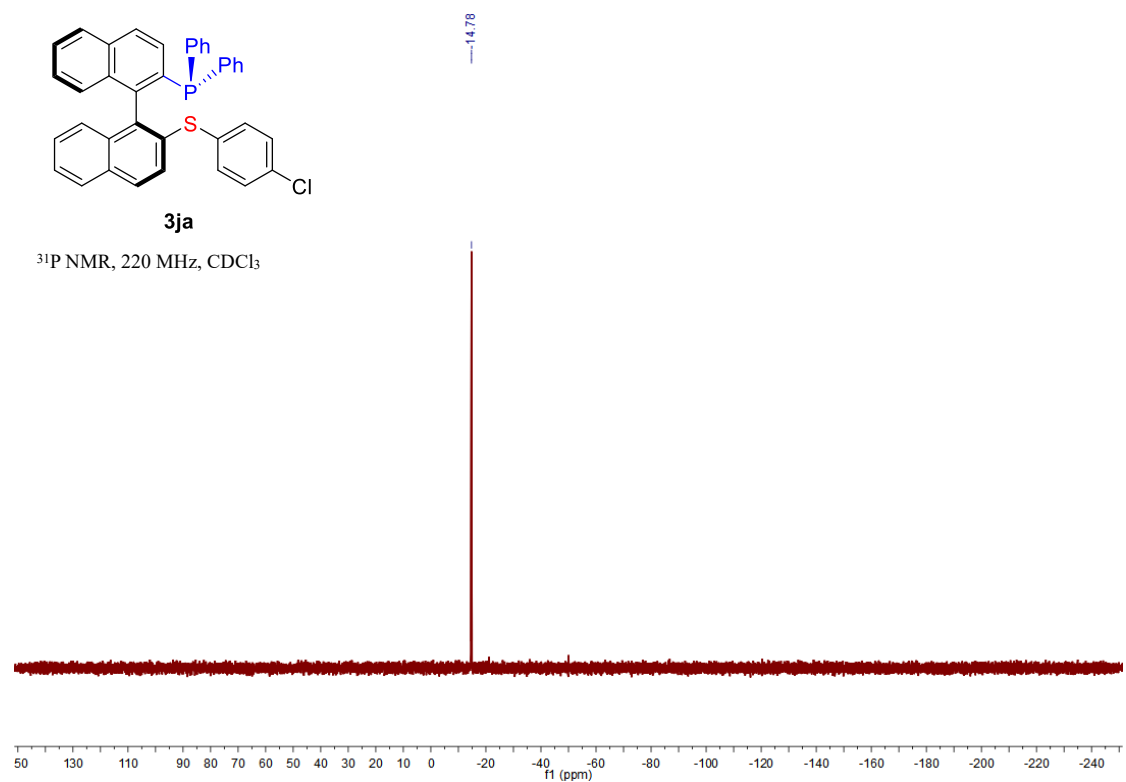


Figure S112. ^{31}P NMR spectrum of **3ja** (202 MHz, CDCl_3)

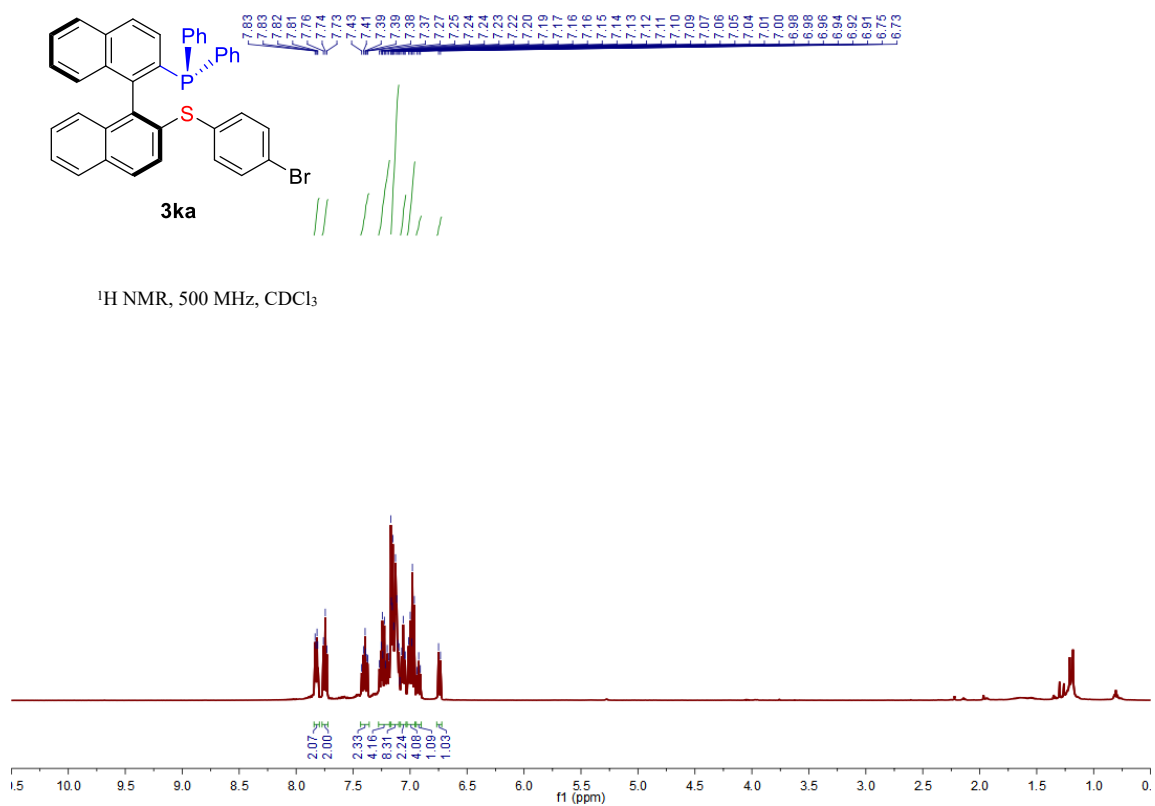


Figure S113. ^1H NMR spectrum of **3ka** (500 MHz, CDCl_3)

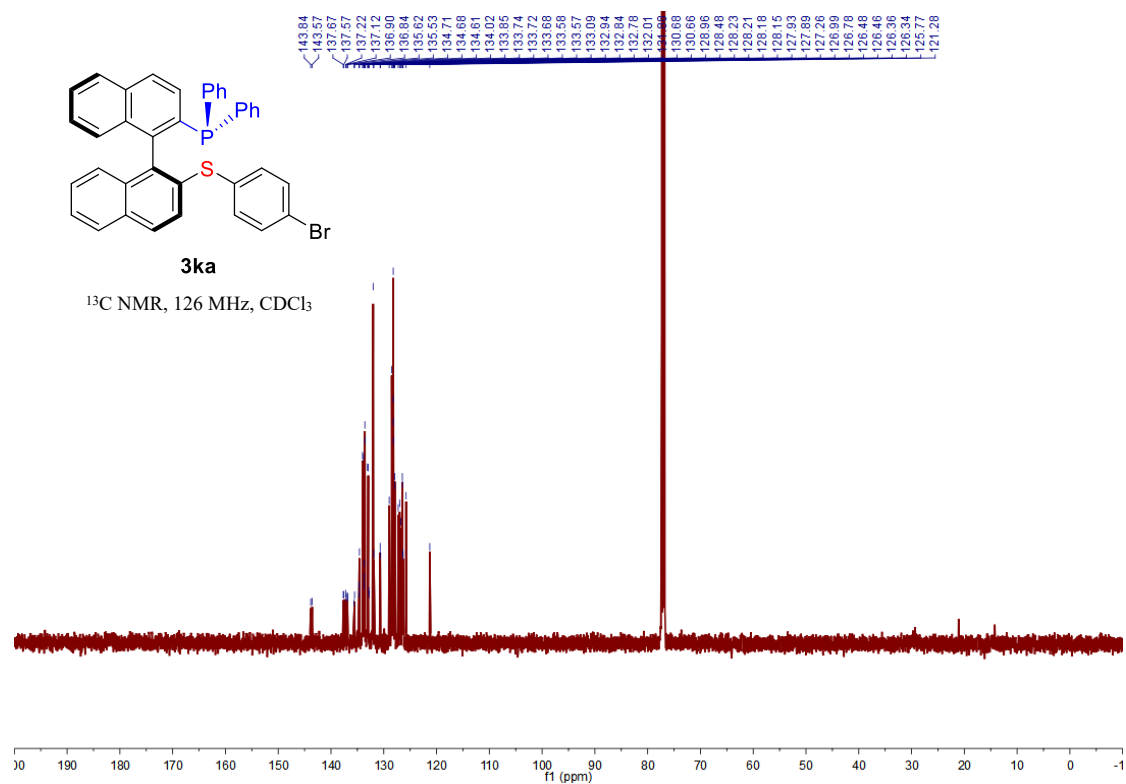


Figure S114. ^{13}C NMR spectrum of **3ka** (126 MHz, CDCl_3)

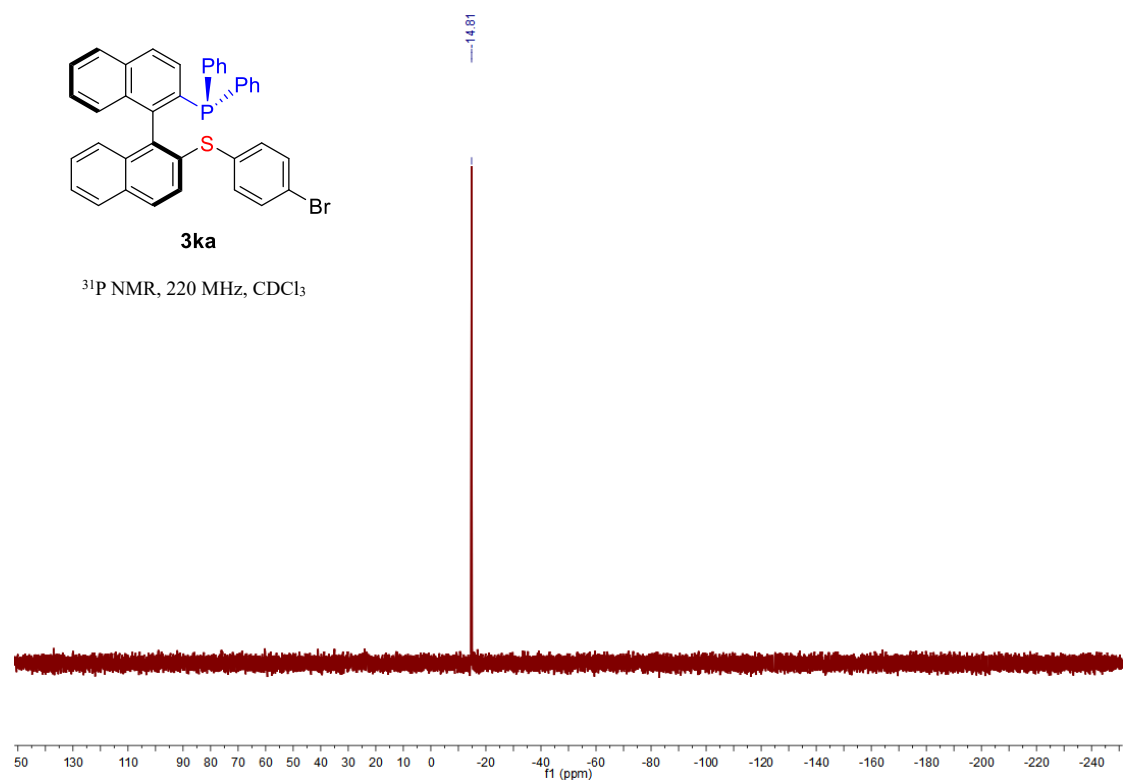


Figure S115. ^{31}P NMR spectrum of **3ka** (202 MHz, CDCl_3)

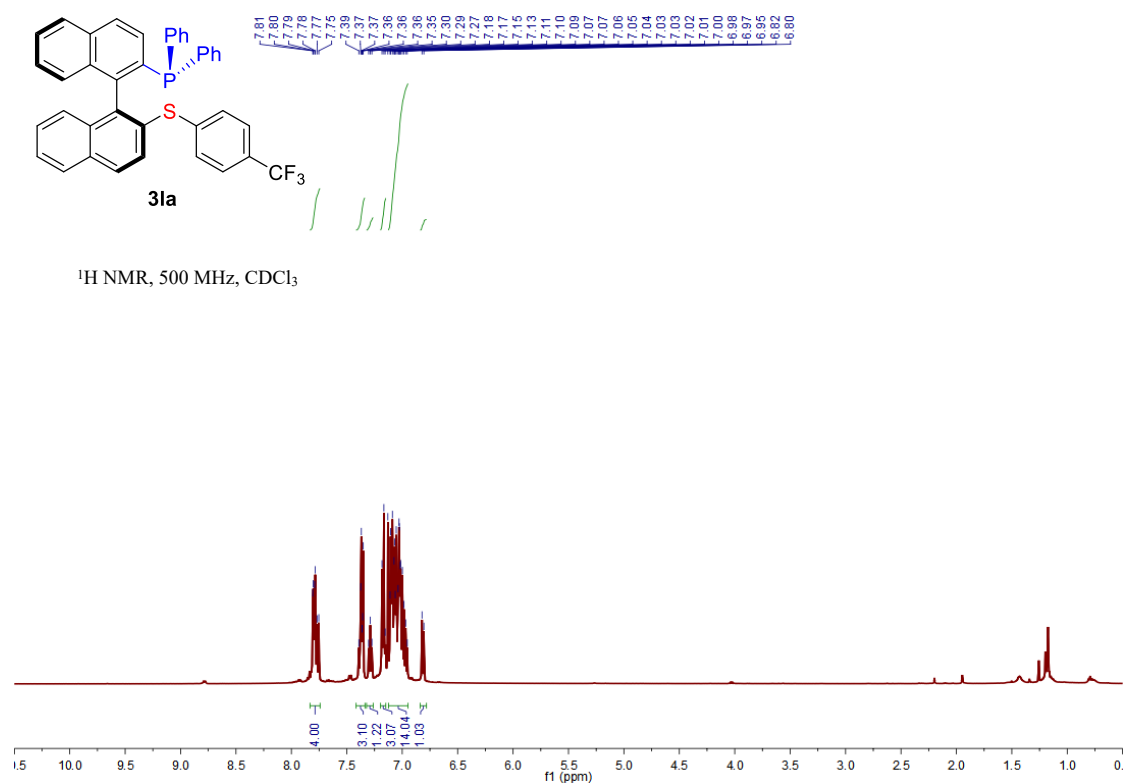


Figure S116. ¹H NMR spectrum of 3la (500 MHz, CDCl₃)

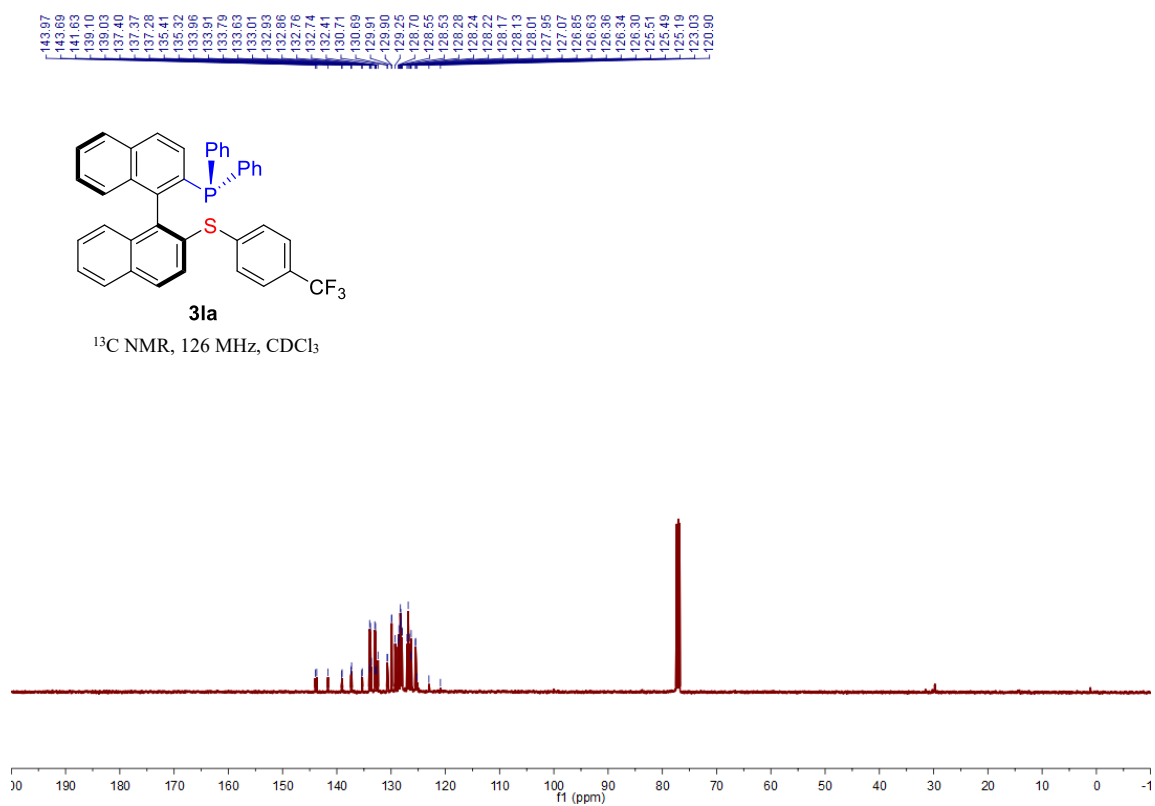


Figure S117. ¹³C NMR spectrum of 3la (126 MHz, CDCl₃)

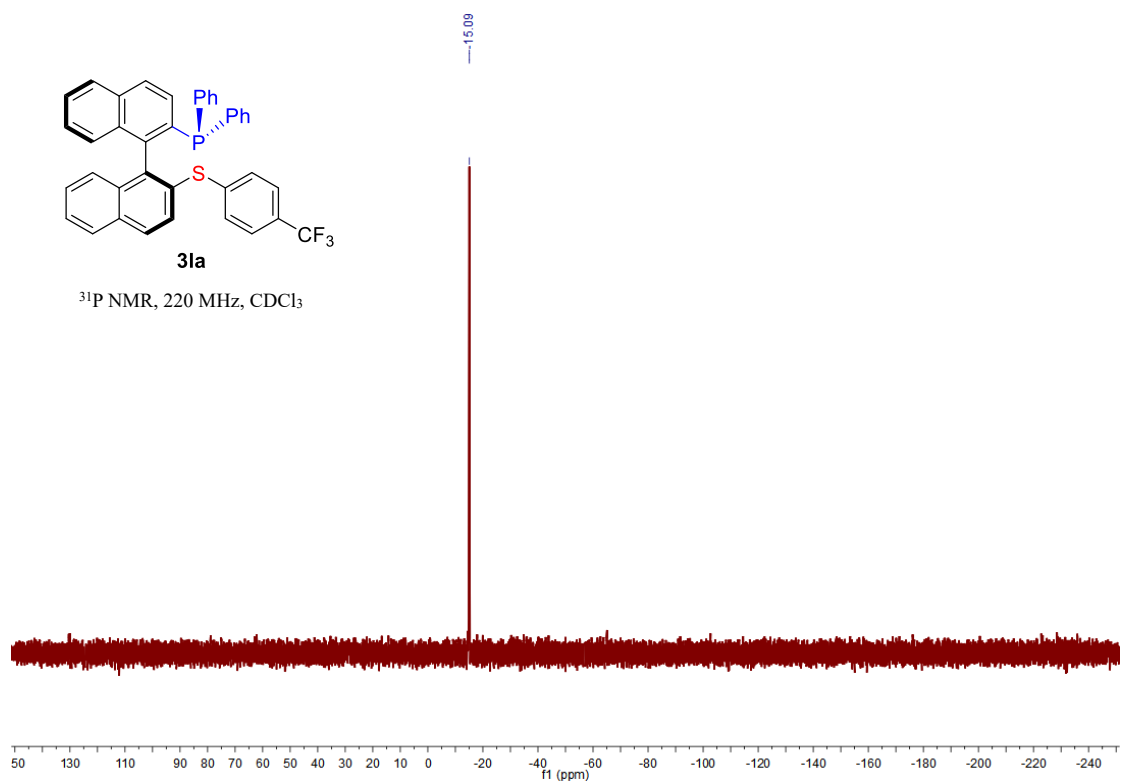


Figure S118. ³¹P NMR spectrum of 3la (202 MHz, CDCl₃)

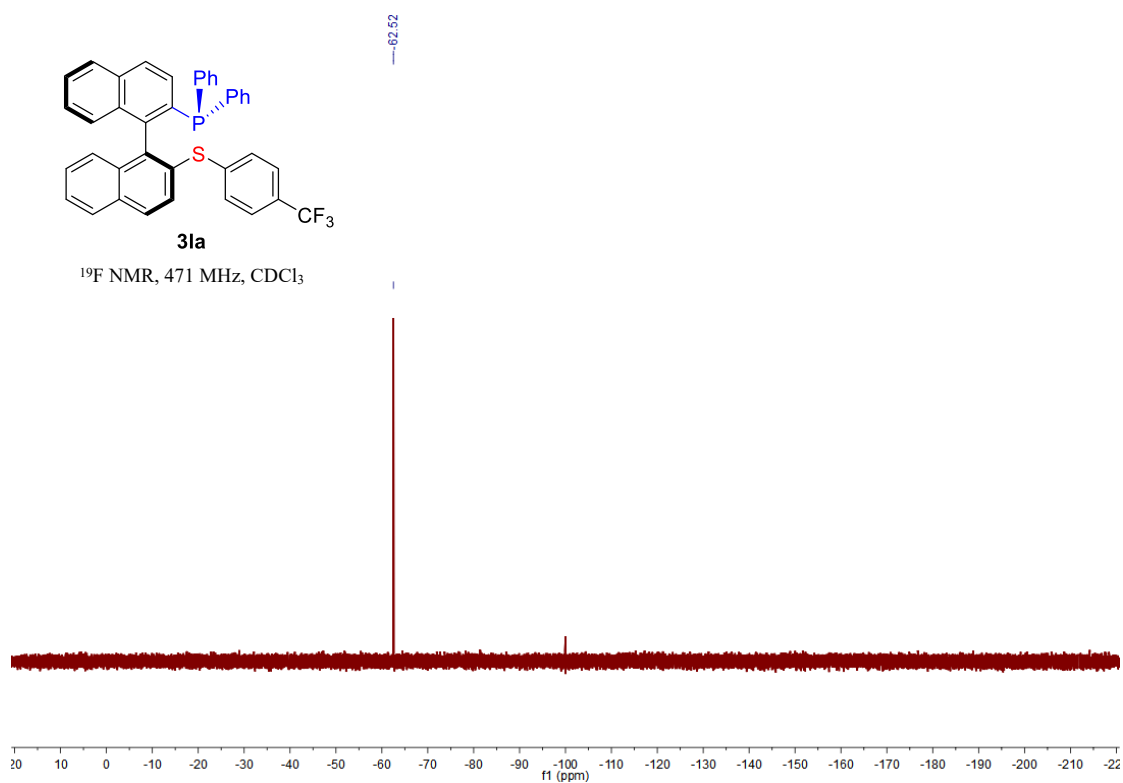


Figure S119. ¹⁹F NMR spectrum of 3la (471 MHz, CDCl₃)

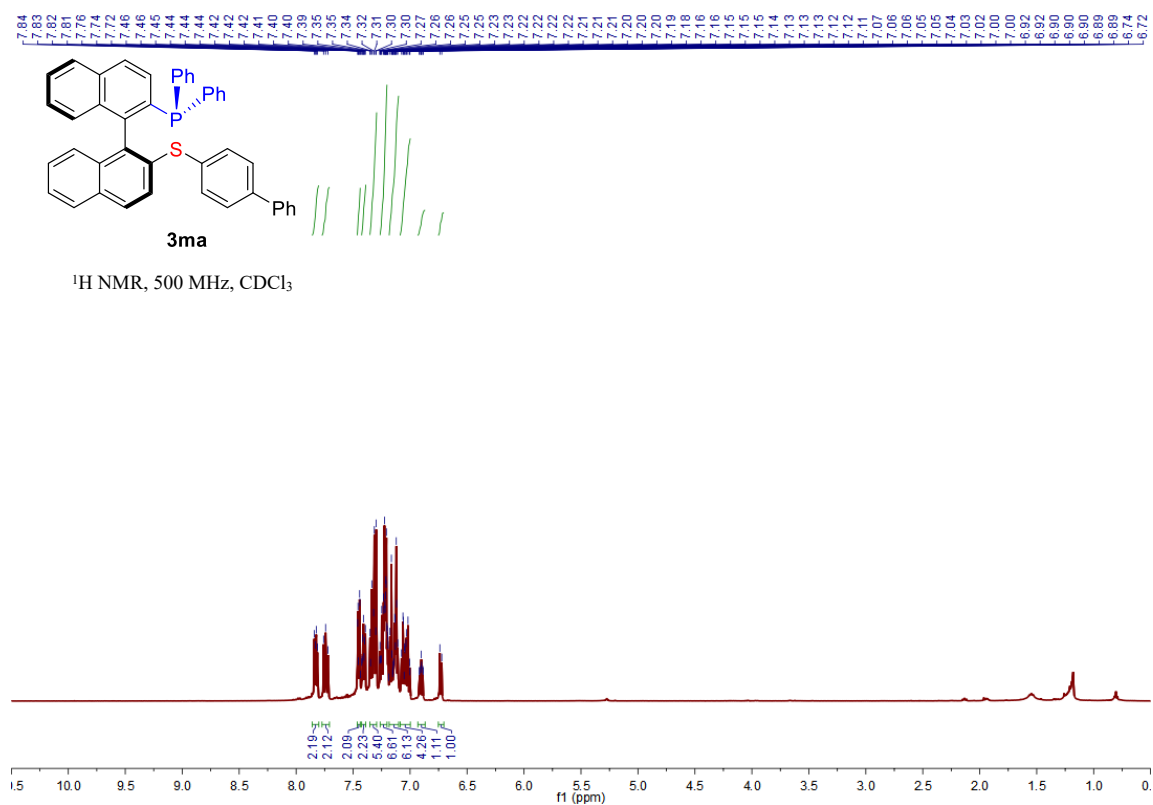


Figure S120. ^1H NMR spectrum of 3ma (500 MHz, CDCl_3)

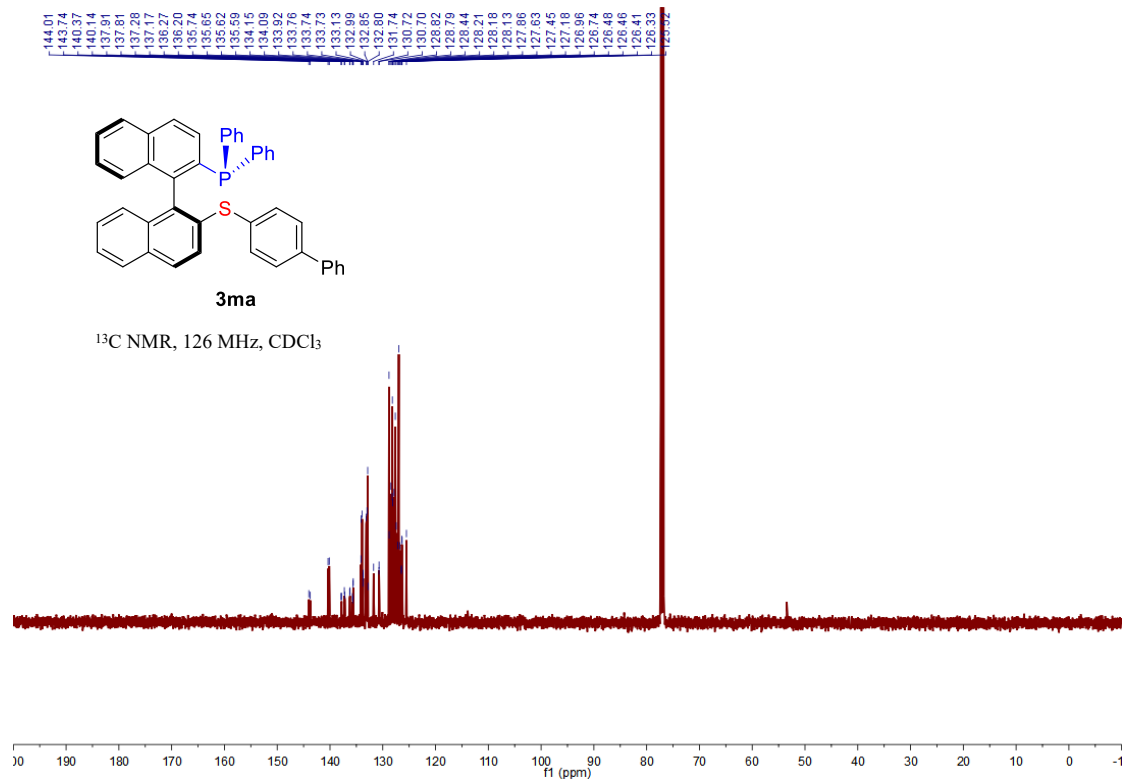


Figure S121. ^{13}C NMR spectrum of 3ma (126 MHz, CDCl_3)

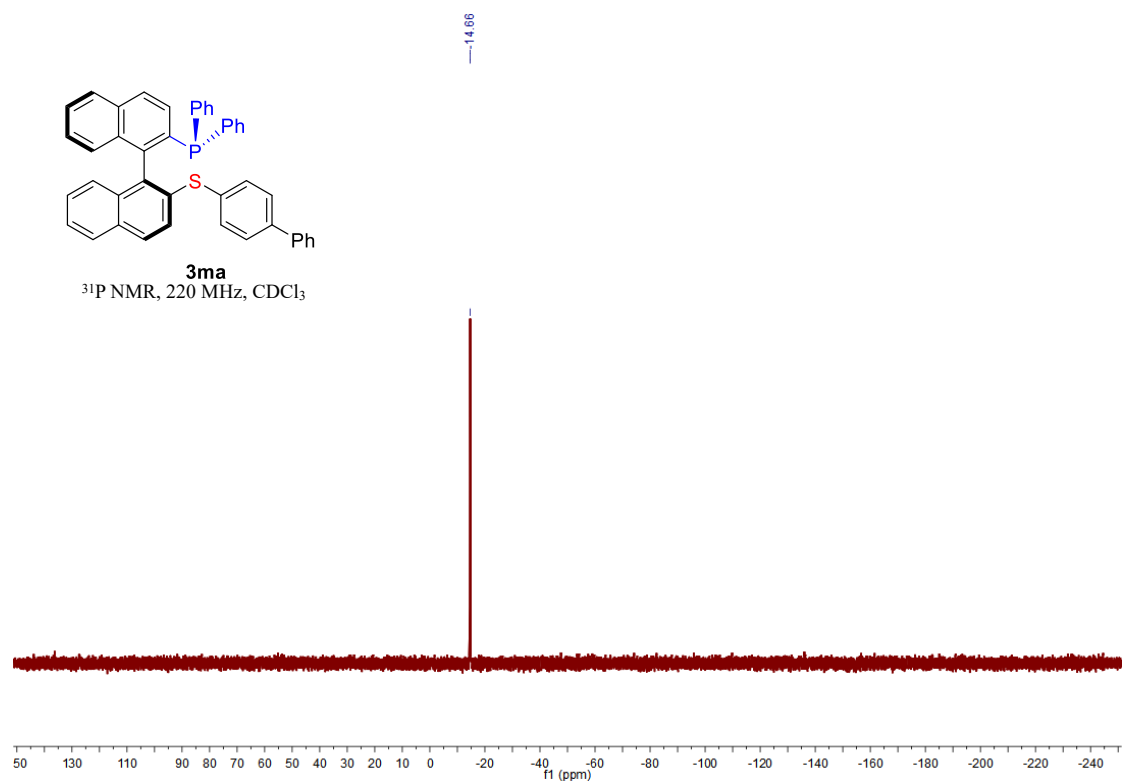


Figure S122. ^{31}P NMR spectrum of **3ma** (202 MHz, CDCl_3)

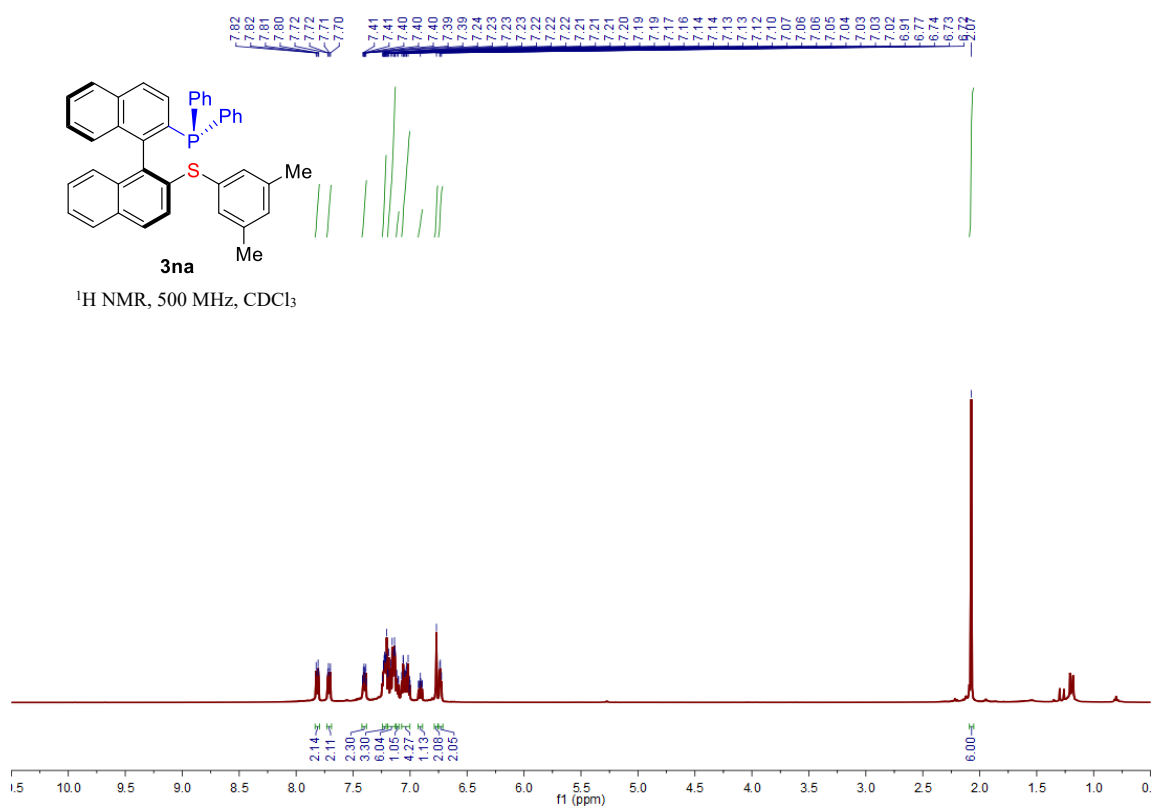


Figure S123. ^1H NMR spectrum of **3na** (500 MHz, CDCl_3)

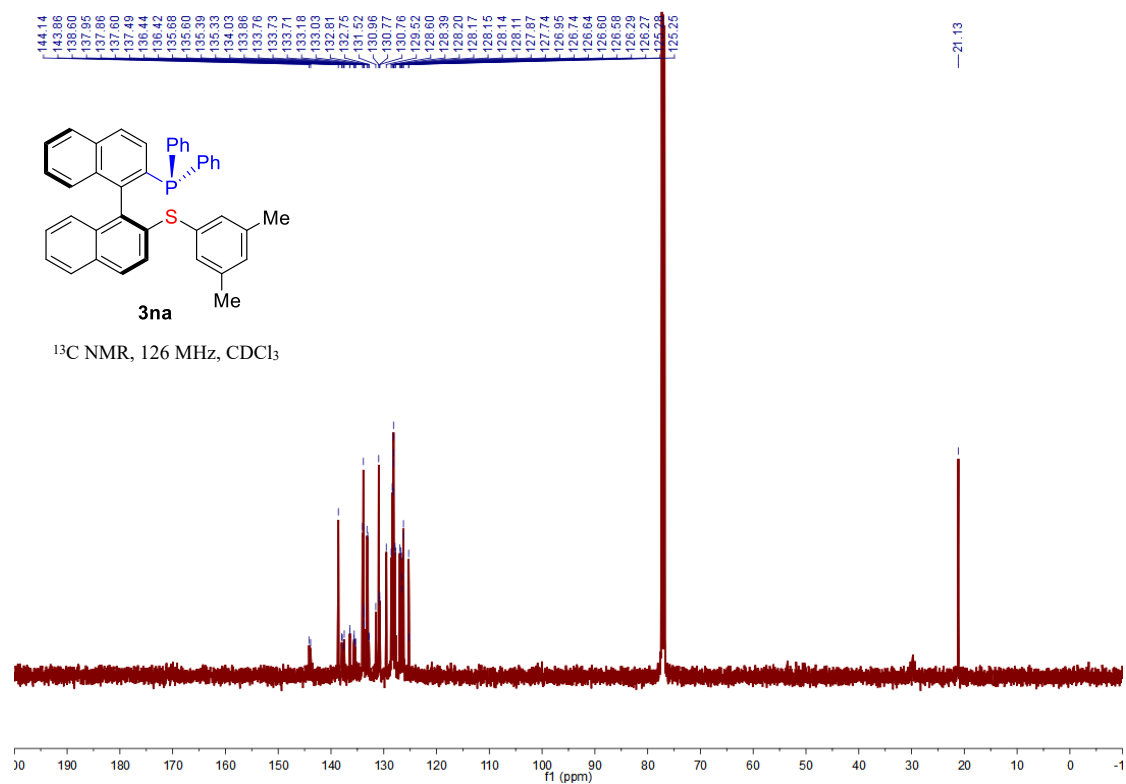


Figure S124. ^{13}C NMR spectrum of **3na** (126 MHz, CDCl_3)

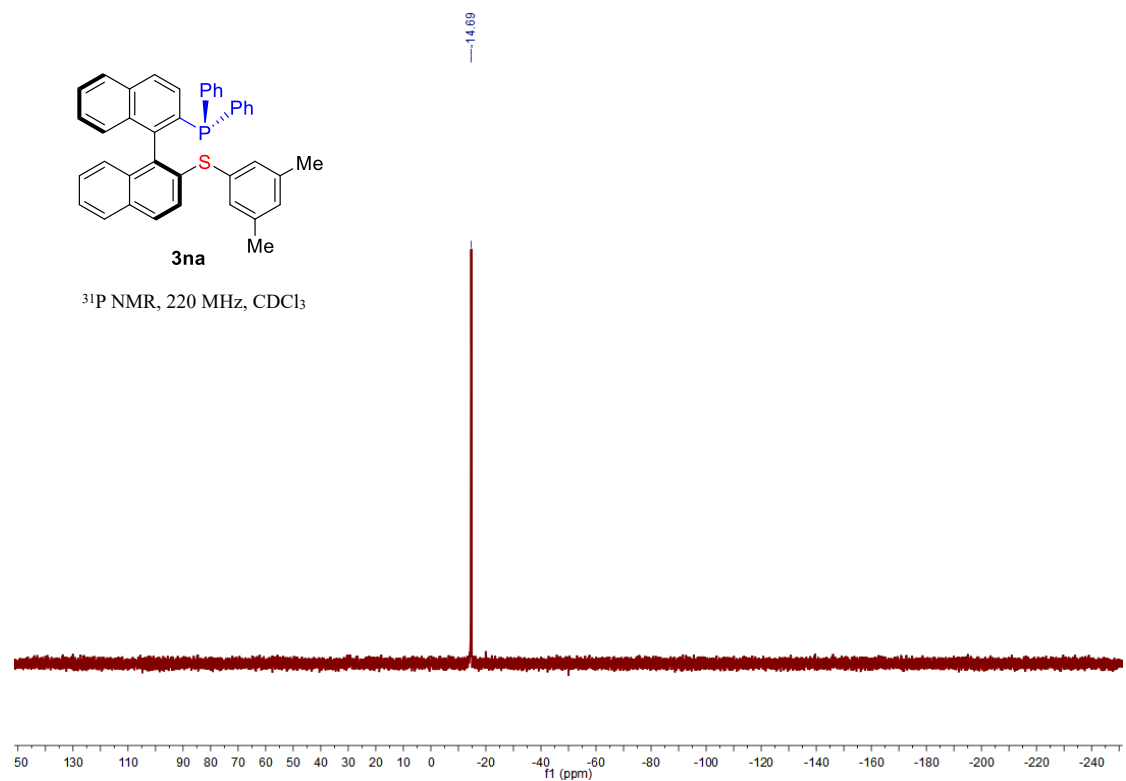


Figure S125. ^{31}P NMR spectrum of **3na** (202 MHz, CDCl_3)

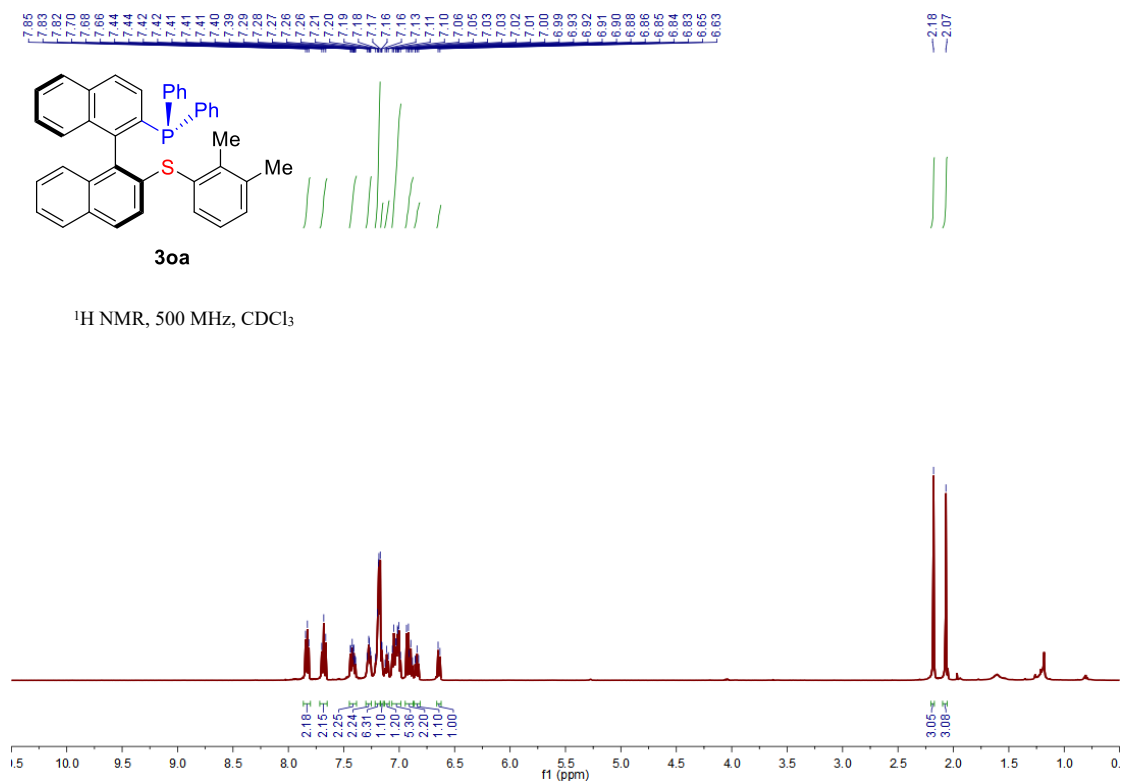


Figure S126. ^1H NMR spectrum of 30a (500 MHz, CDCl_3)

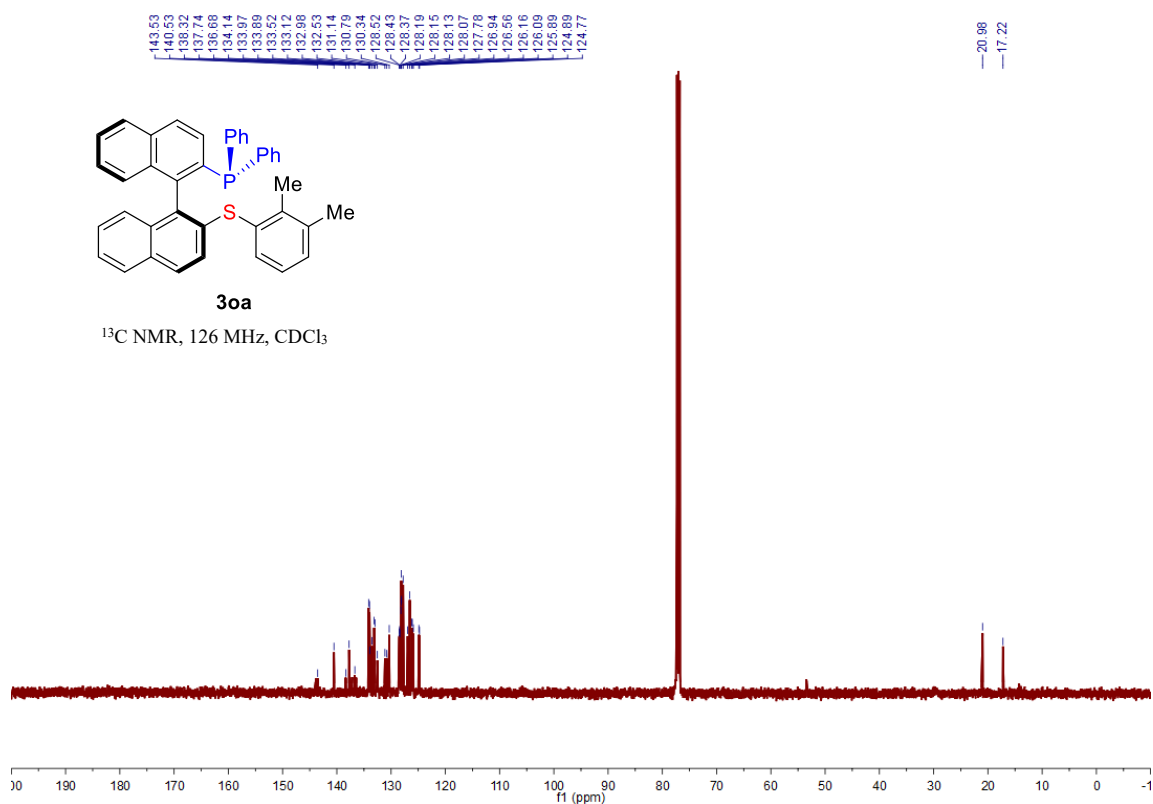


Figure S127. ^{13}C NMR spectrum of 30a (126 MHz, CDCl_3)

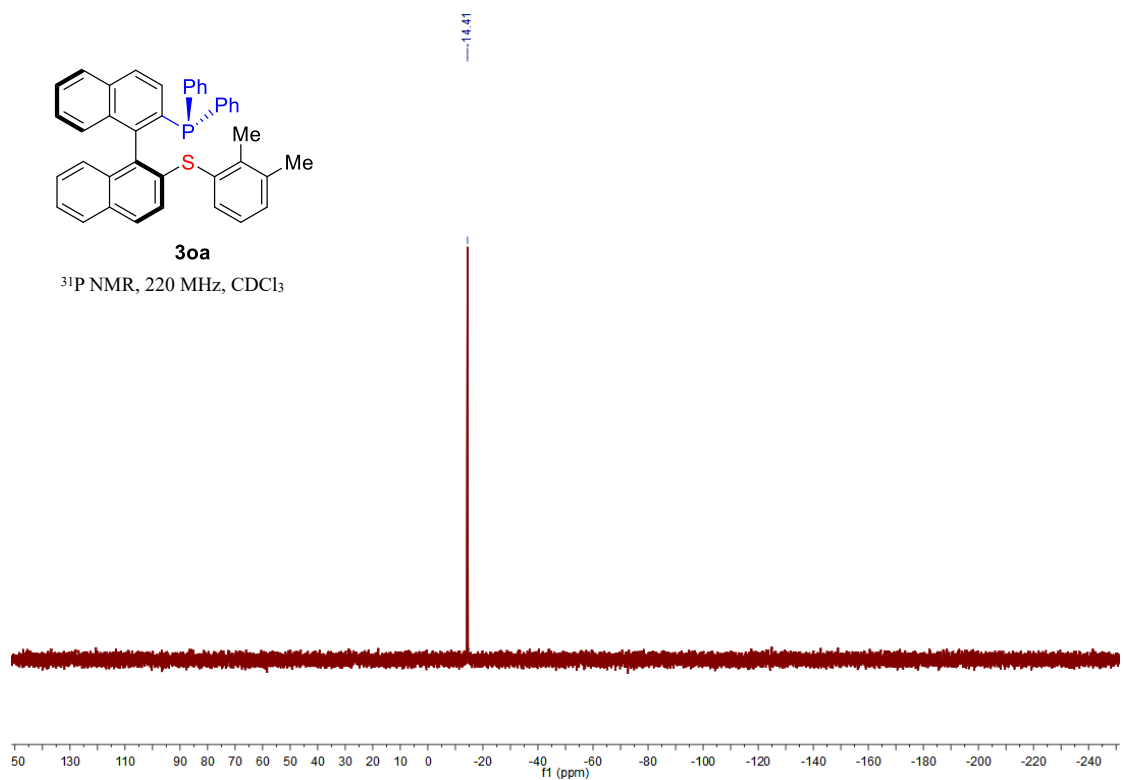


Figure S128. ^{31}P NMR spectrum of **3oa** (202 MHz, CDCl_3)

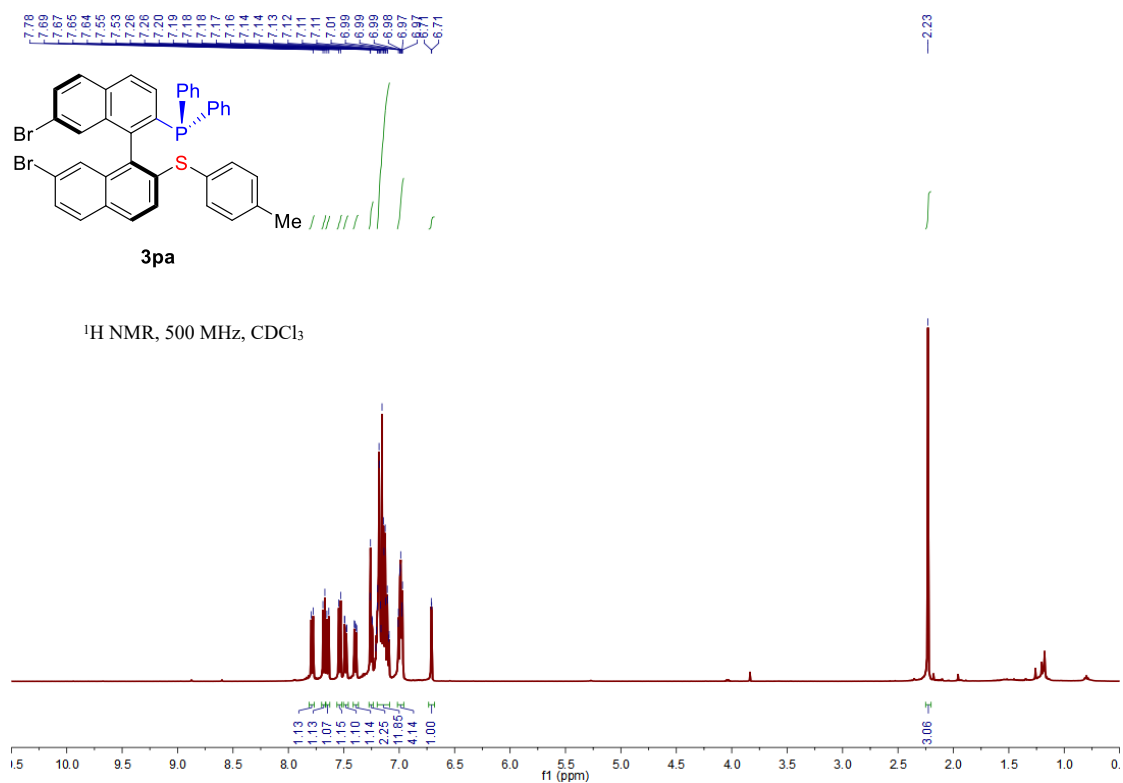


Figure S129. ^1H NMR spectrum of **3pa** (500 MHz, CDCl_3)

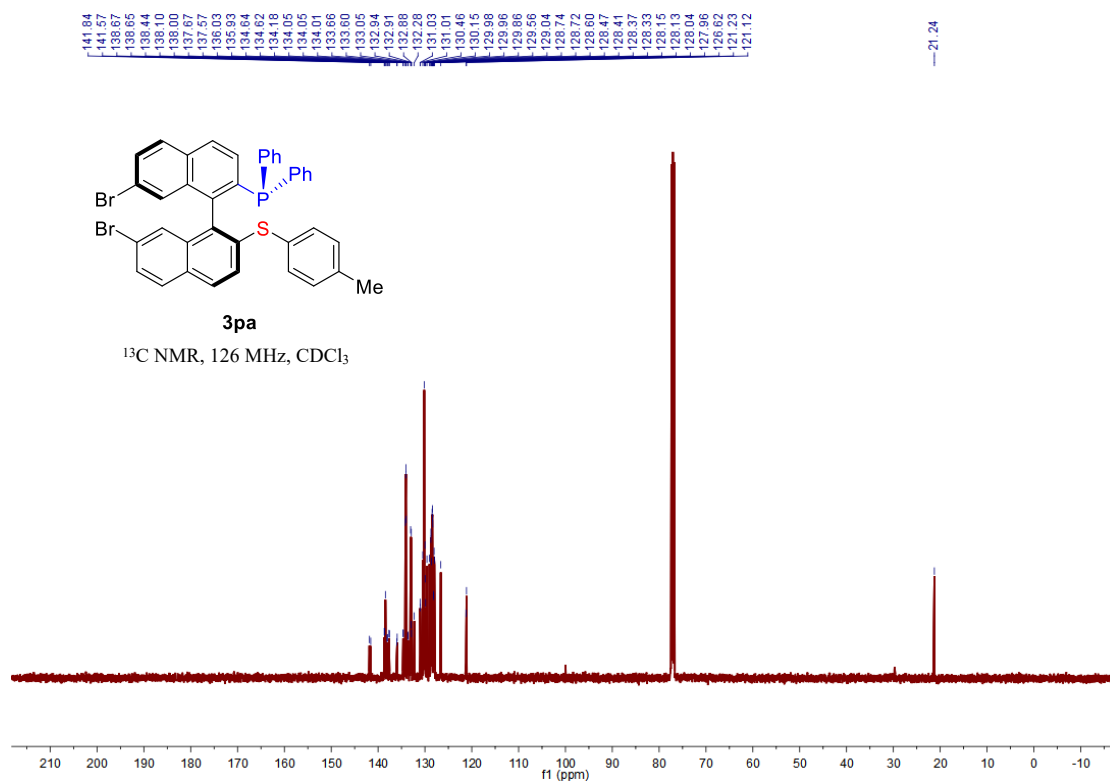


Figure S130. ^{13}C NMR spectrum of **3pa** (126 MHz, CDCl_3)

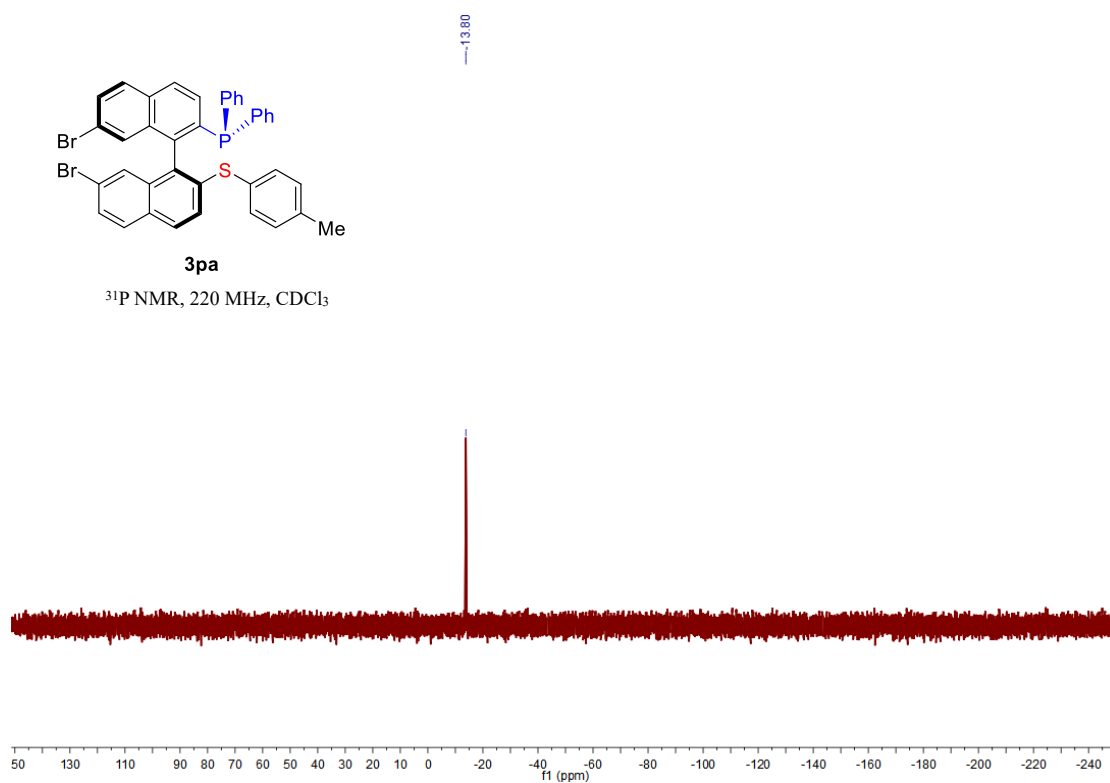
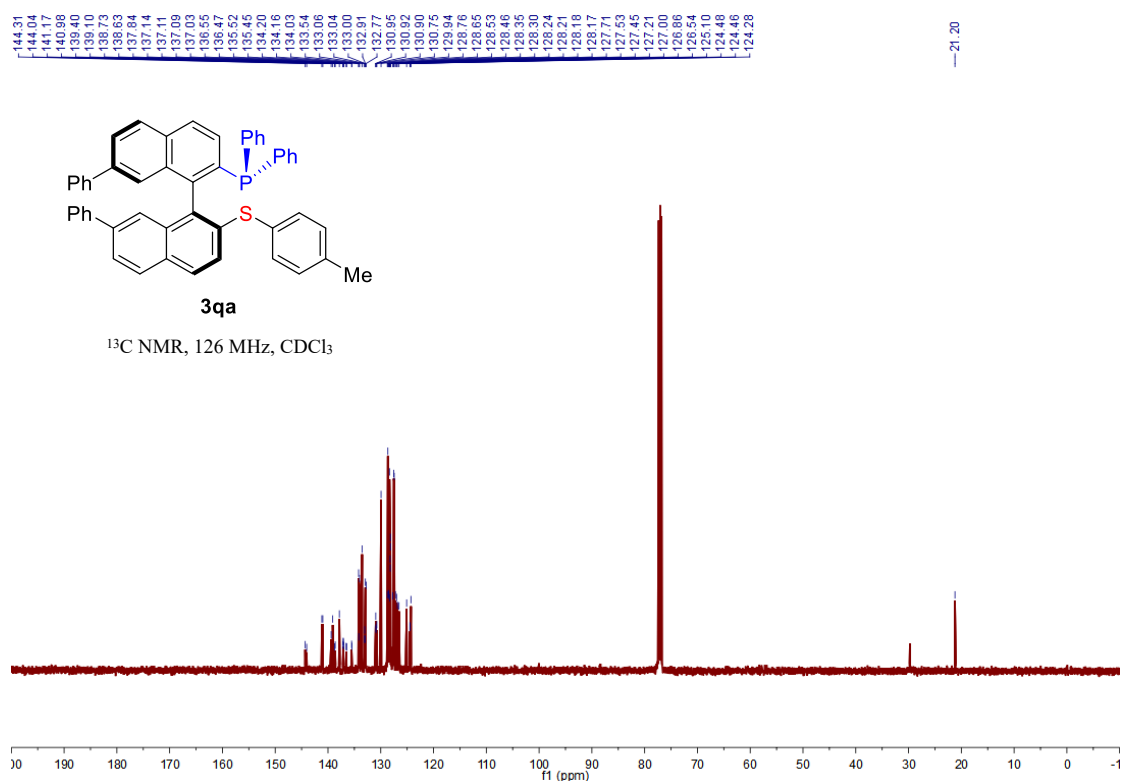
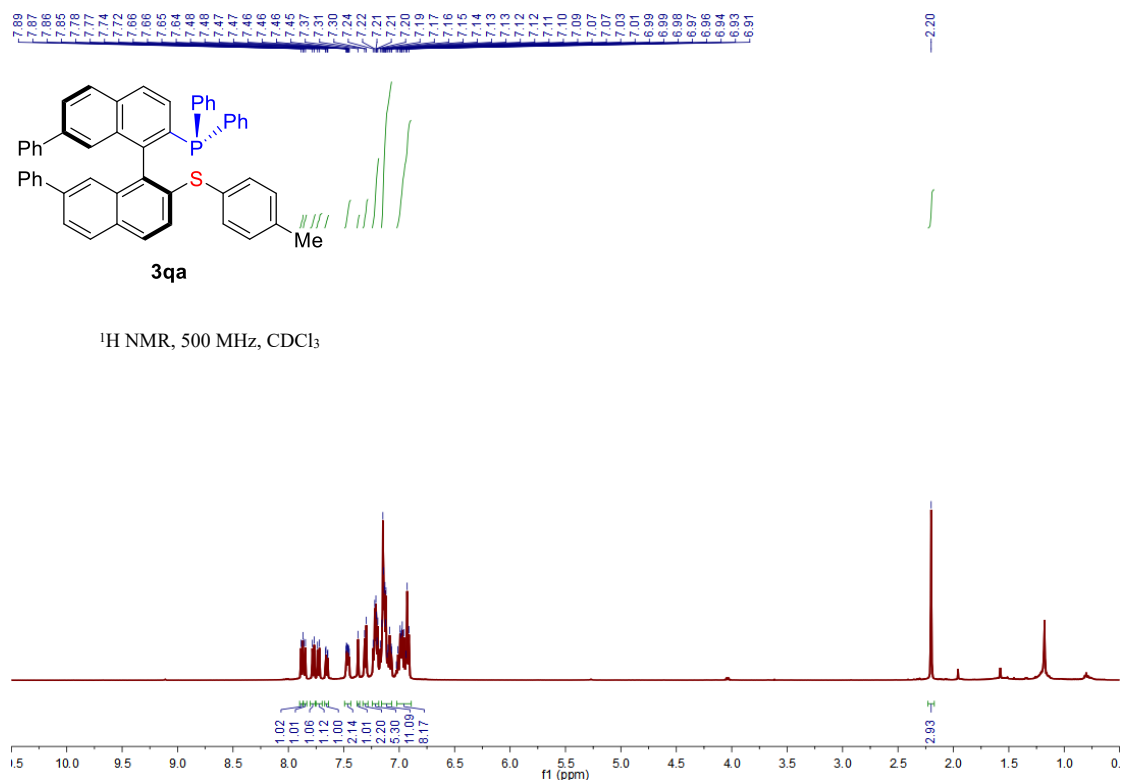


Figure S131. ^{31}P NMR spectrum of **3pa** (202 MHz, CDCl_3)



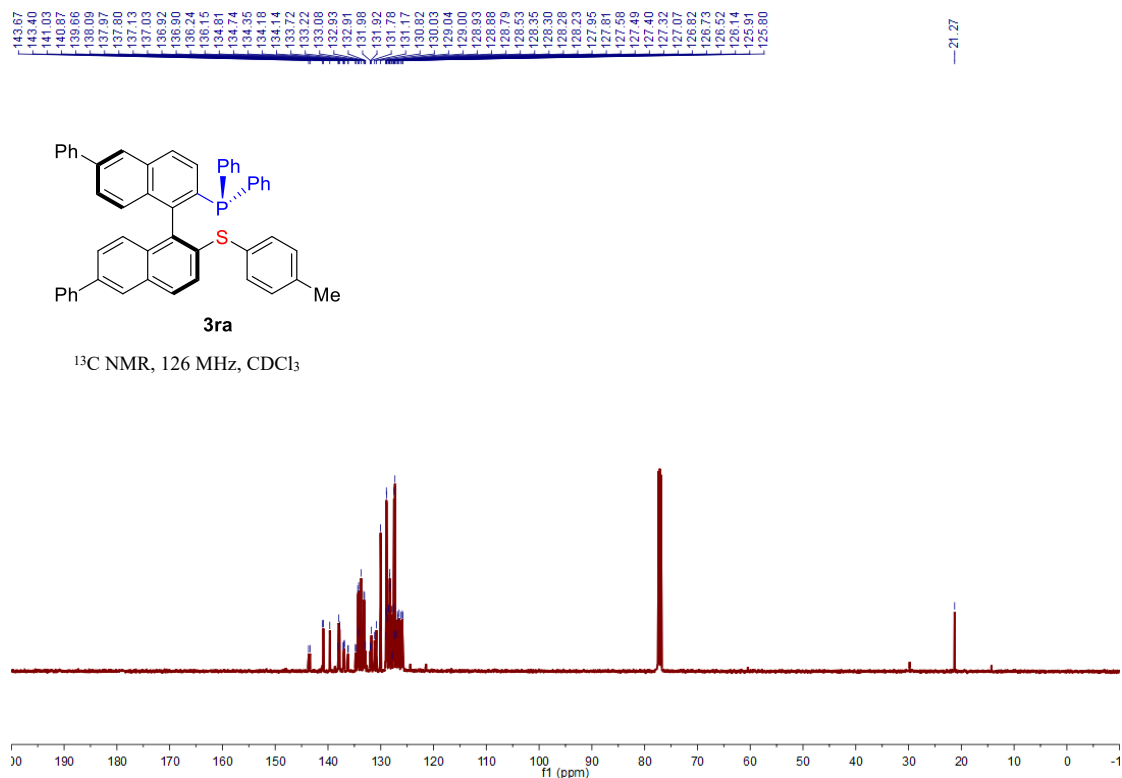


Figure S136. ^{13}C NMR spectrum of 3ra (126 MHz, CDCl_3)

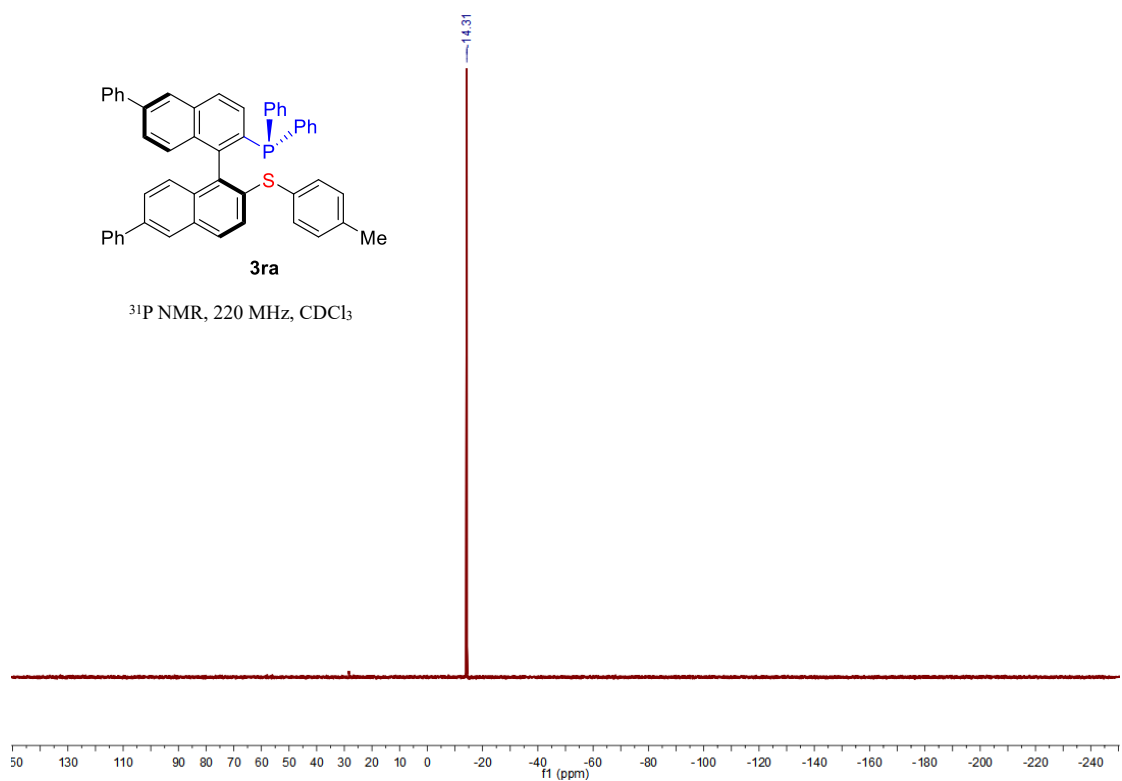


Figure S137. ^{31}P NMR spectrum of 3ra (202 MHz, CDCl_3)

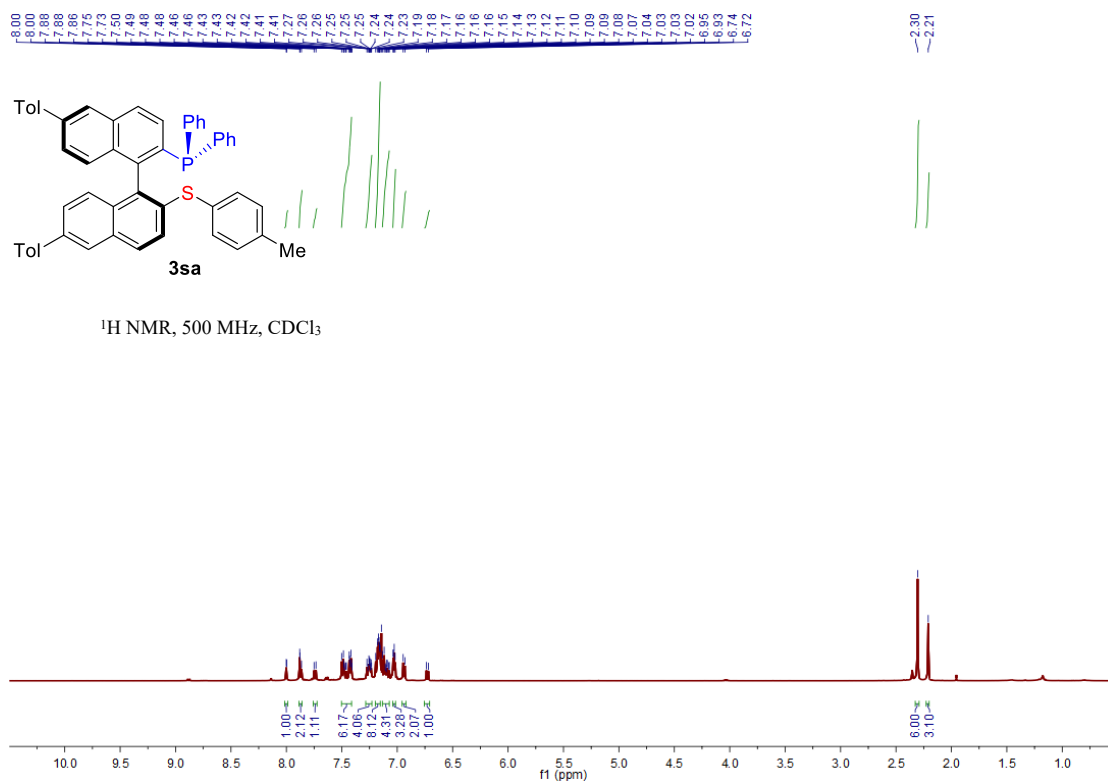


Figure S138. ¹H NMR spectrum of 3sa (500 MHz, CDCl₃)

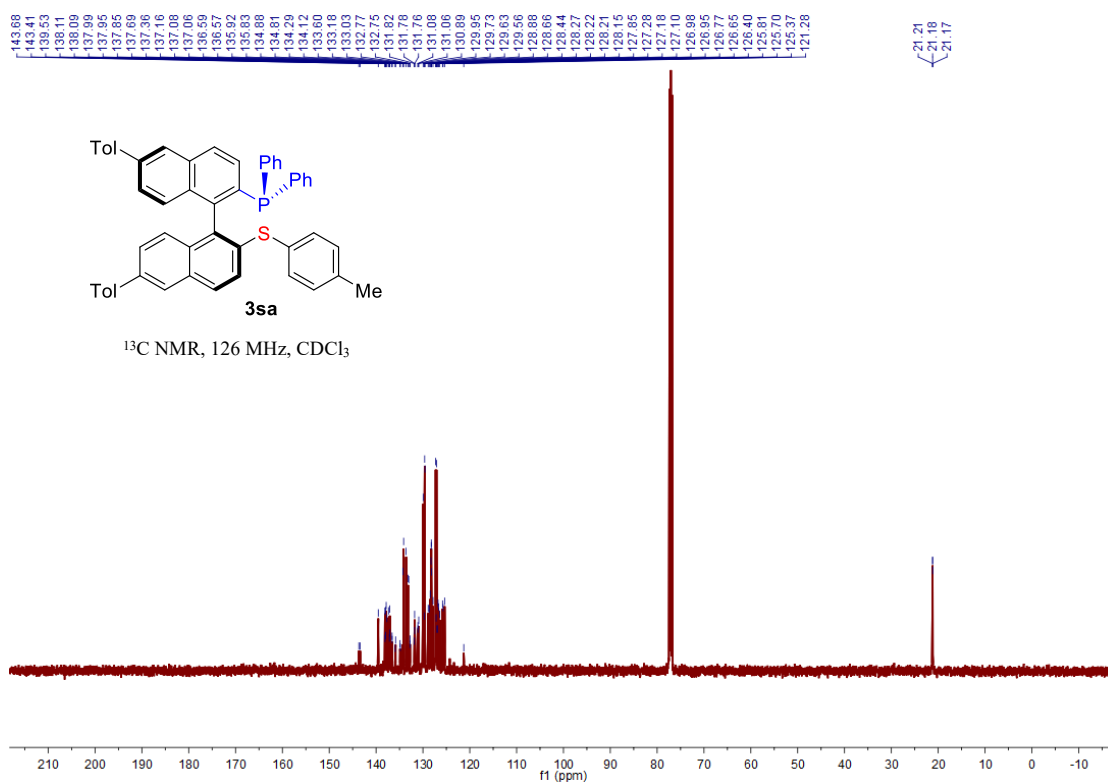


Figure S139. ¹³C NMR spectrum of 3sa (126 MHz, CDCl₃)

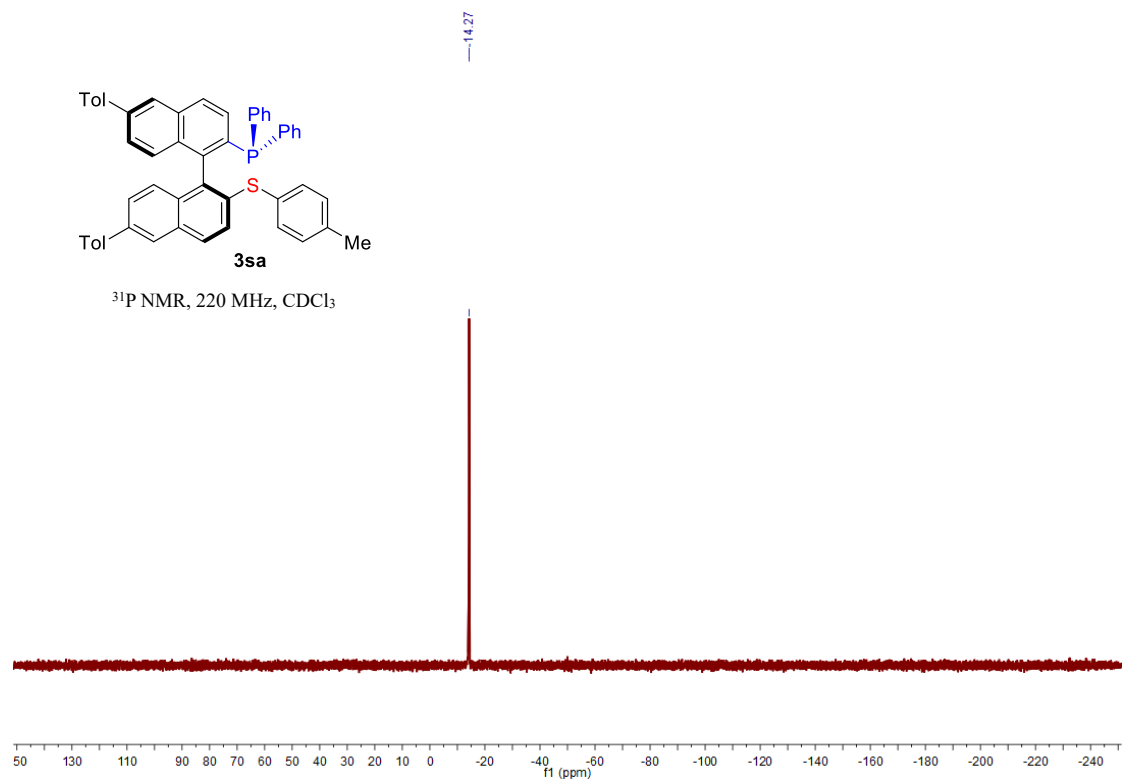


Figure S140. ³¹P NMR spectrum of 3sa (202 MHz, CDCl₃)

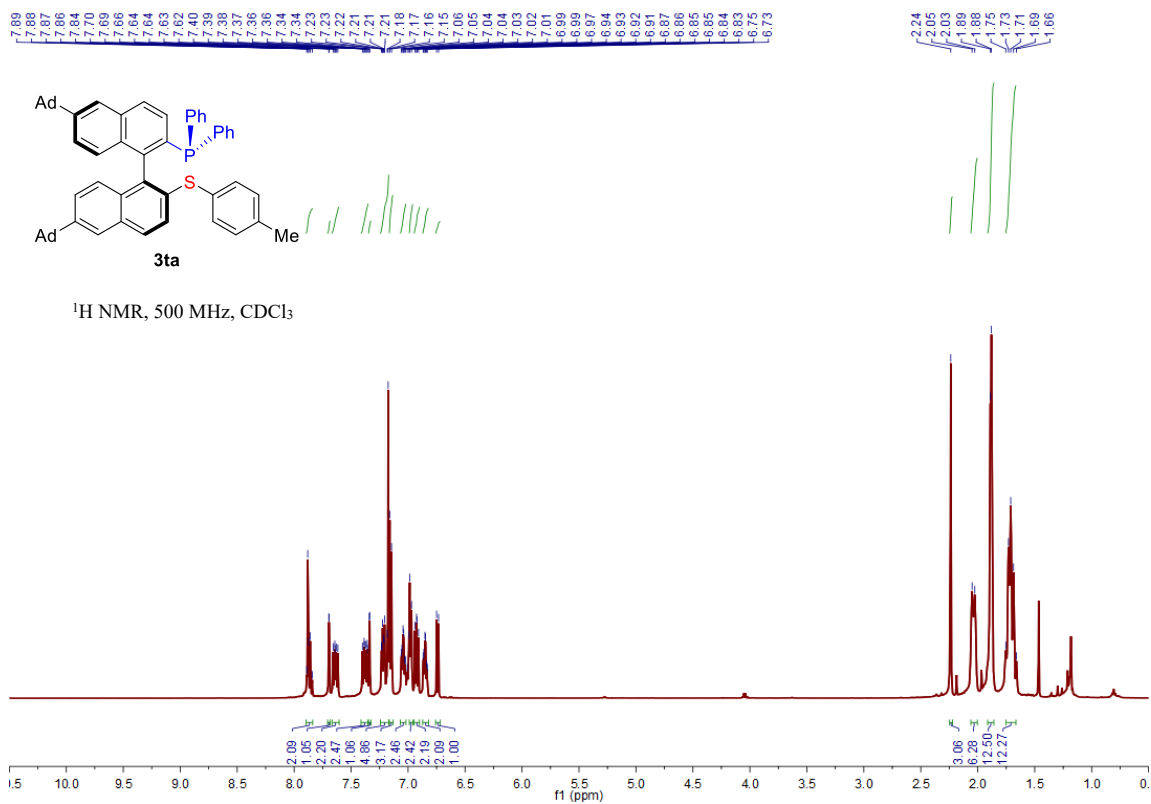


Figure S141. ¹H NMR spectrum of 3ta (500 MHz, CDCl₃)

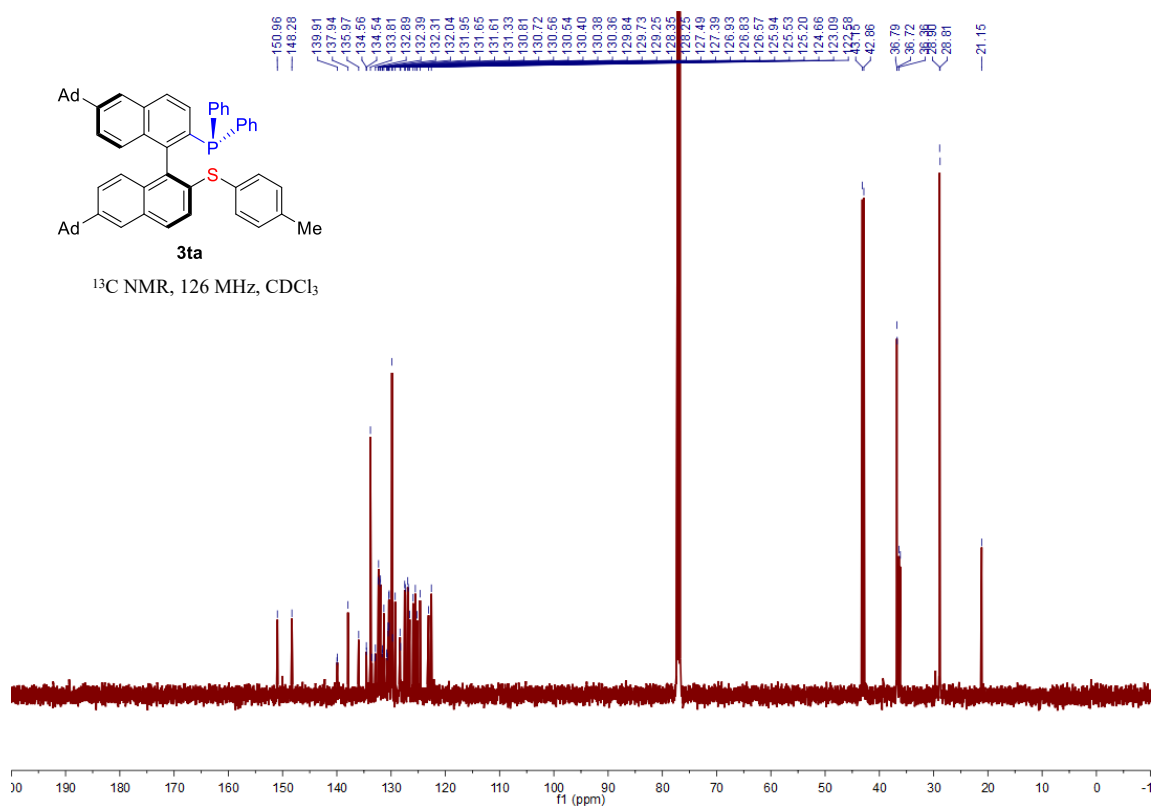


Figure S142. ^{13}C NMR spectrum of **3ta** (126 MHz, CDCl_3)

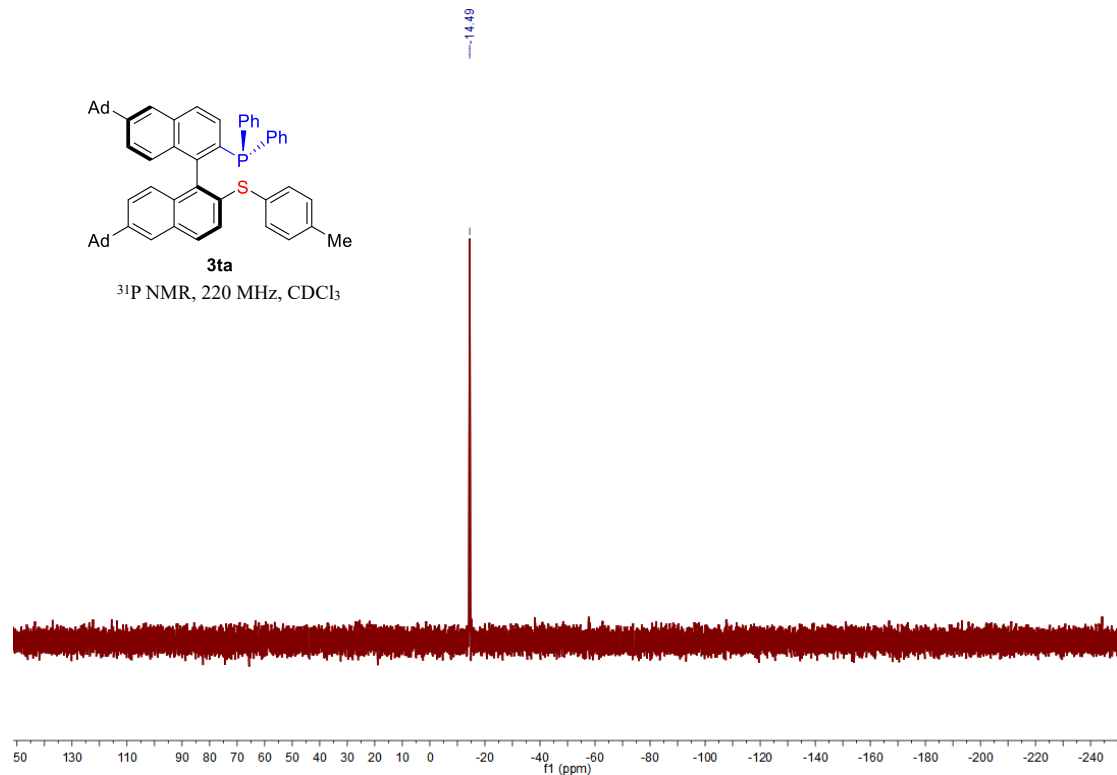


Figure S143. ^{31}P NMR spectrum of **3ta** (202 MHz, CDCl_3)

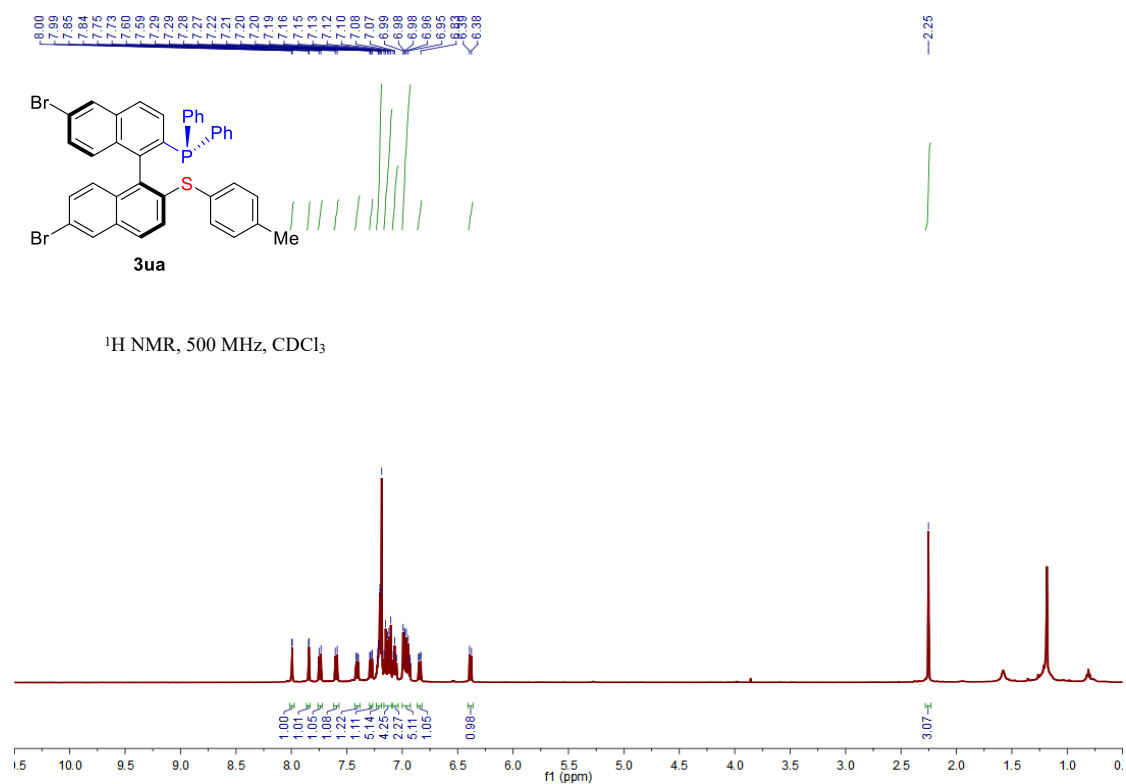


Figure S144. ^1H NMR spectrum of 3ua (500 MHz, CDCl_3)

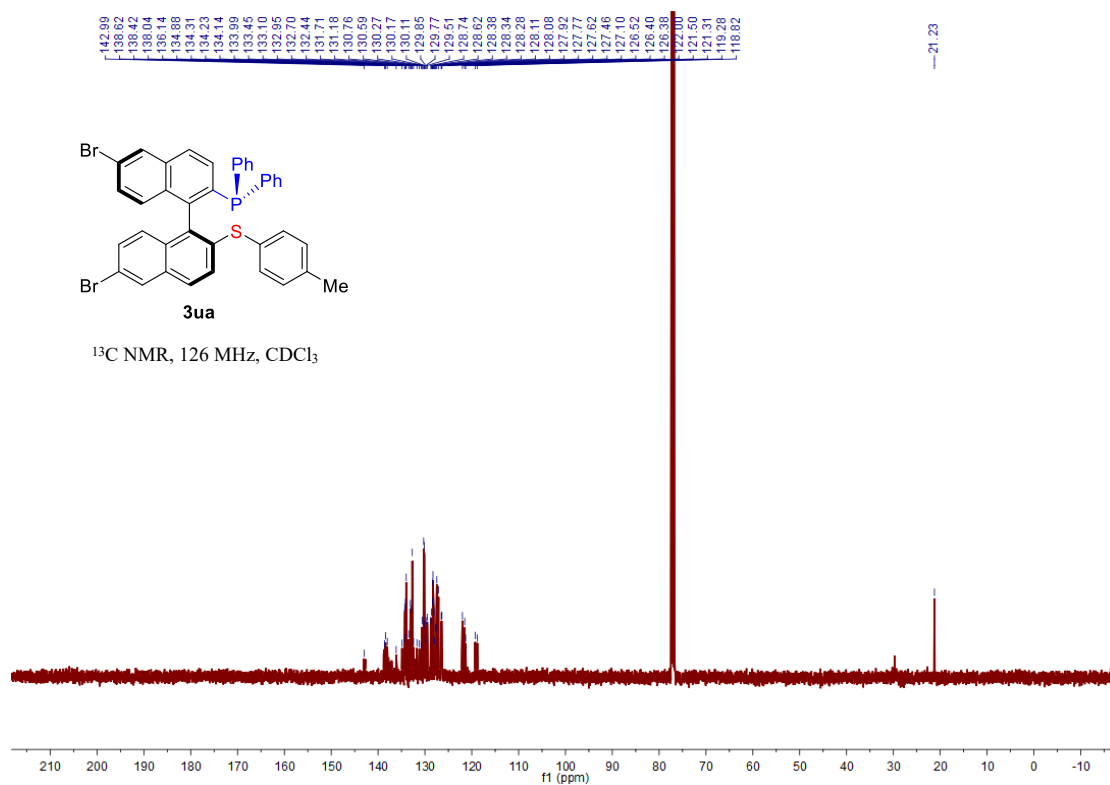


Figure S145. ^{13}C NMR spectrum of 3ua (126 MHz, CDCl_3)

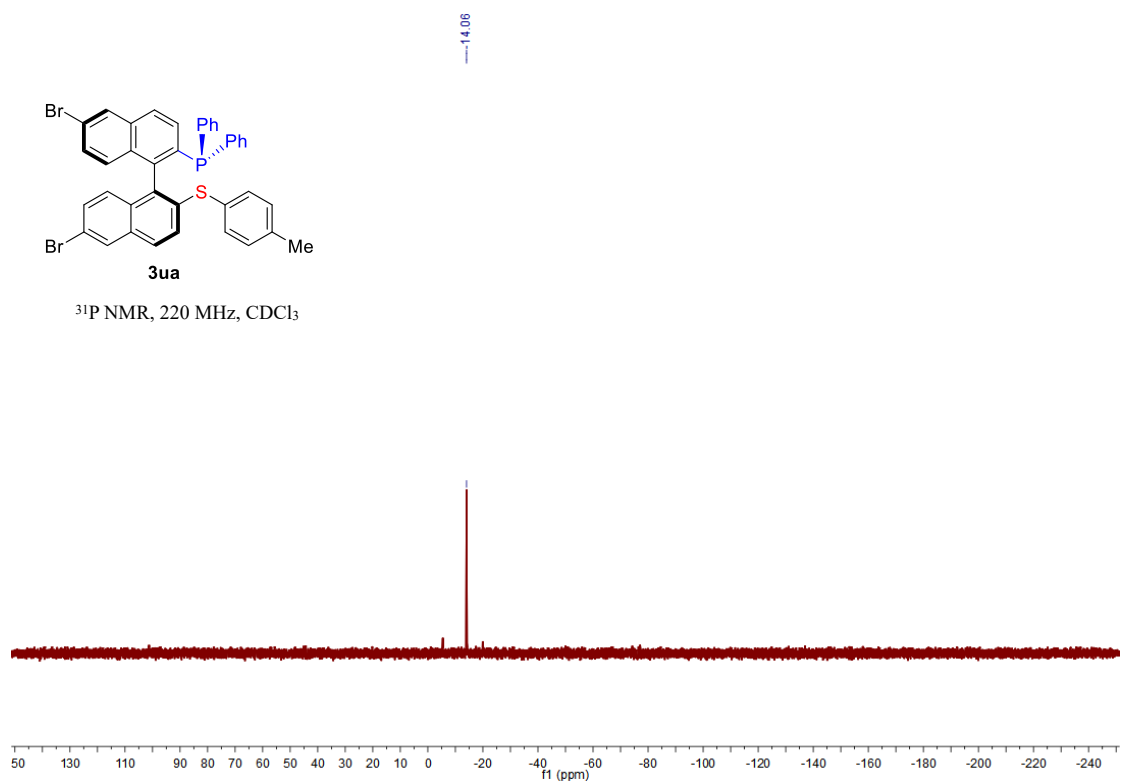


Figure S146. ^{31}P NMR spectrum of **3ua** (202 MHz, CDCl_3)

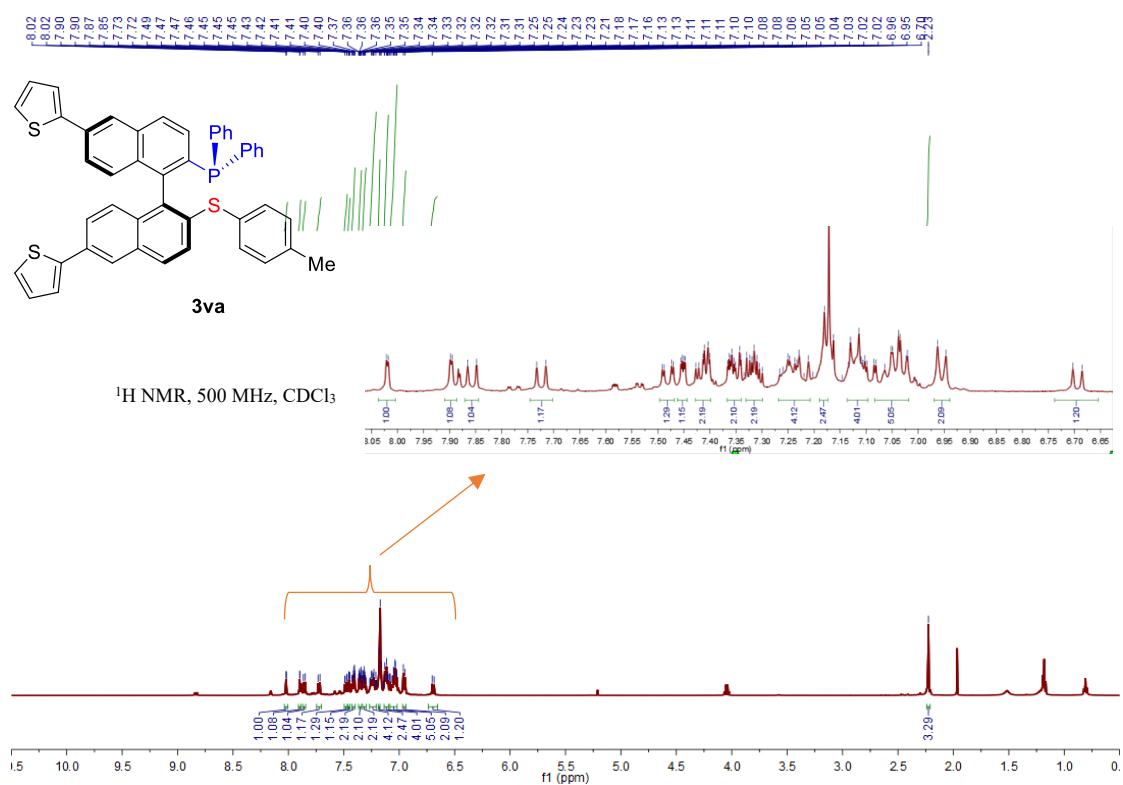


Figure S147. ^1H NMR spectrum of **3va** (500 MHz, CDCl_3)

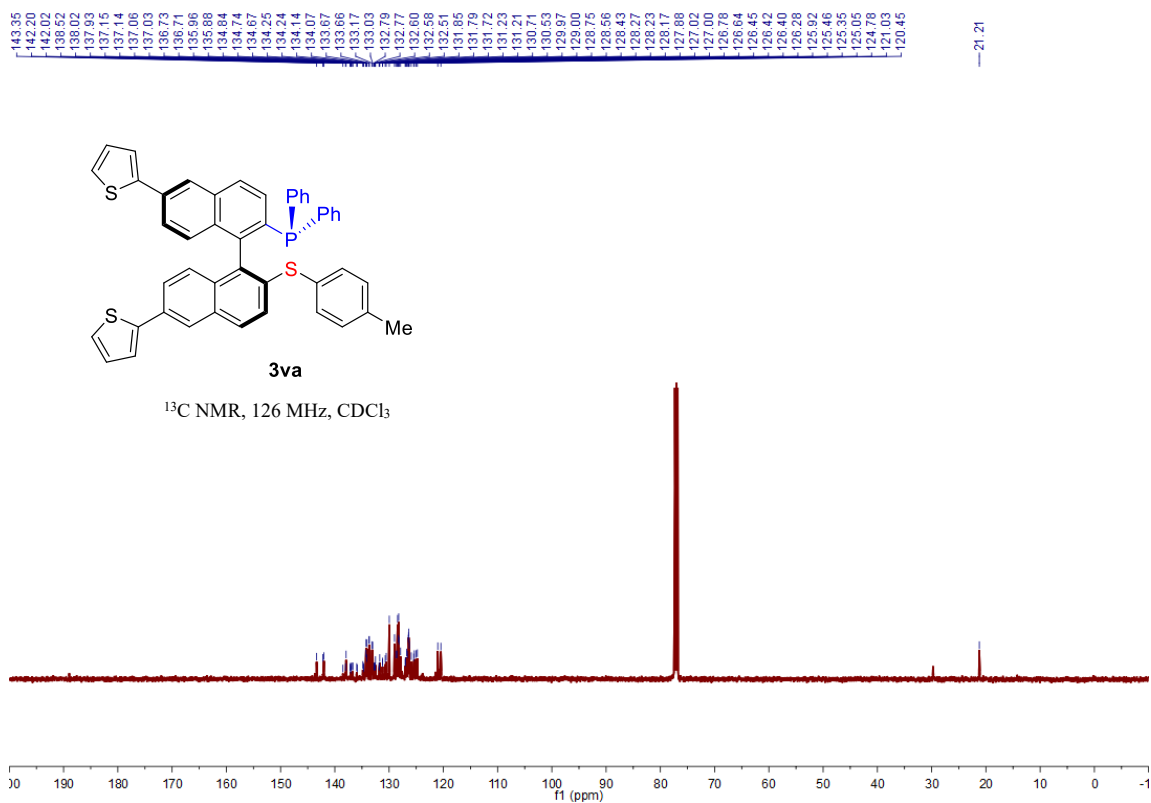


Figure S148. ^{13}C NMR spectrum of 3va (126 MHz, CDCl_3)

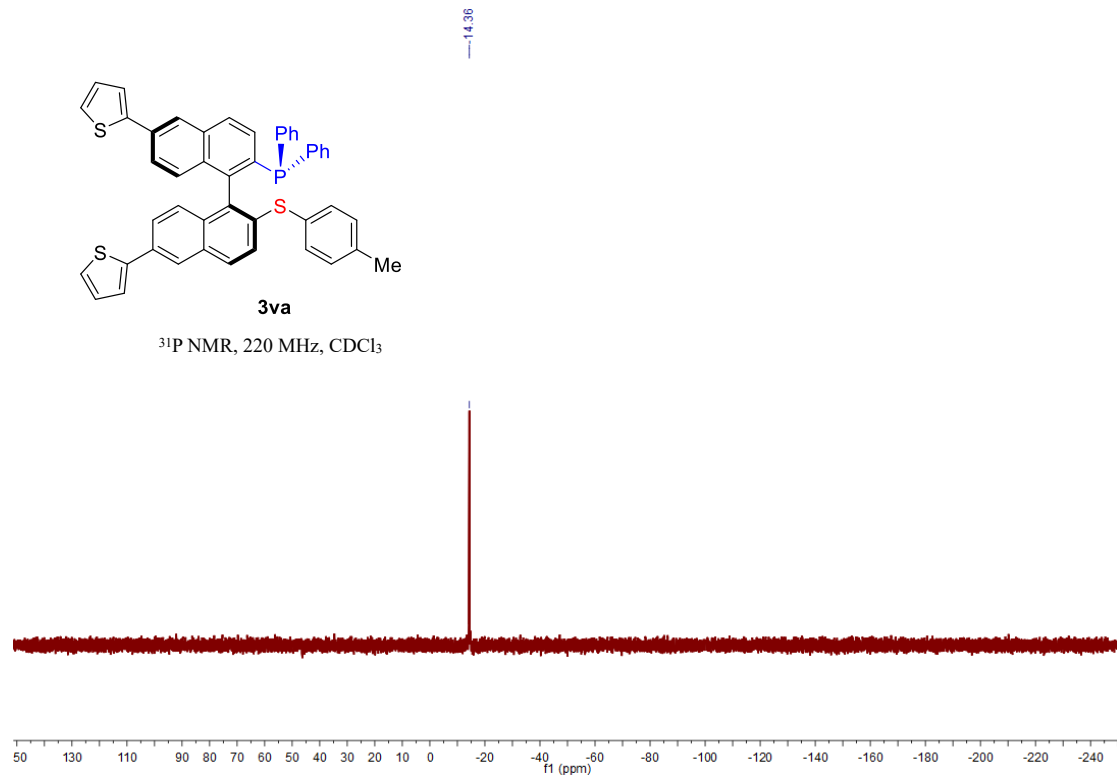


Figure S149. ^{31}P NMR spectrum of 3va (202 MHz, CDCl_3)

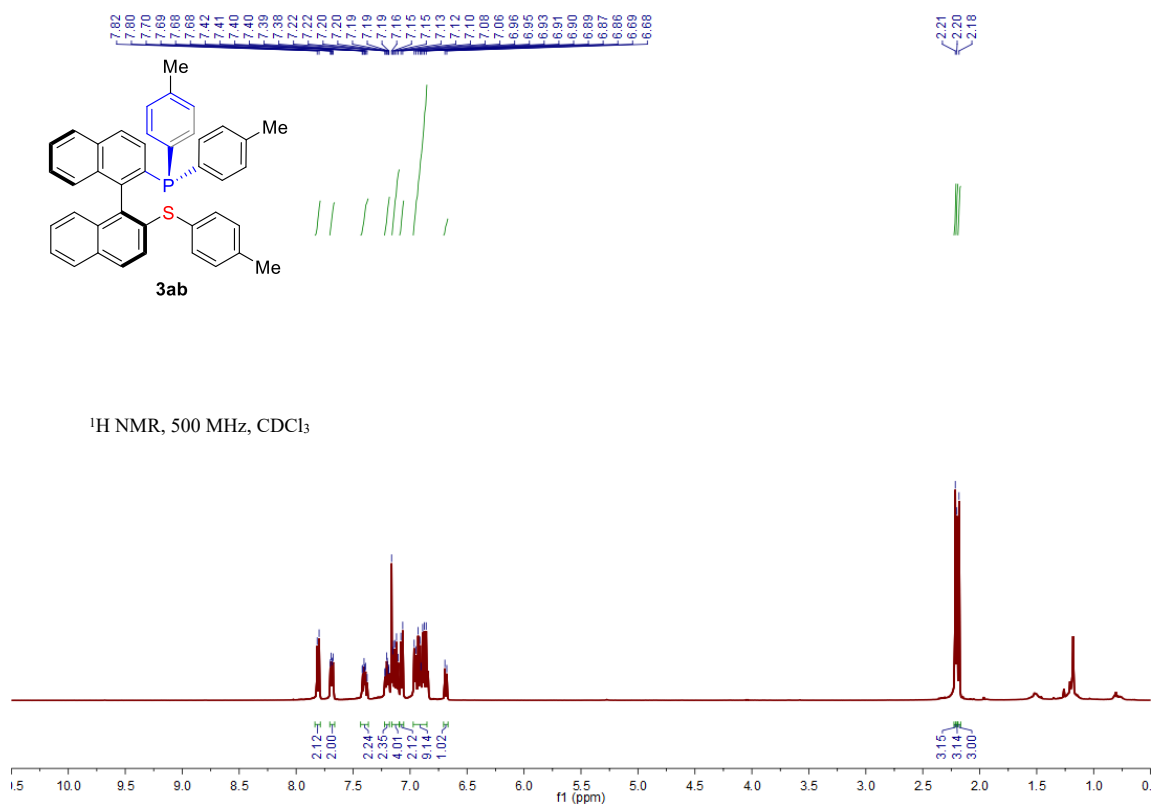


Figure S150. ¹H NMR spectrum of 3ab (500 MHz, CDCl₃)

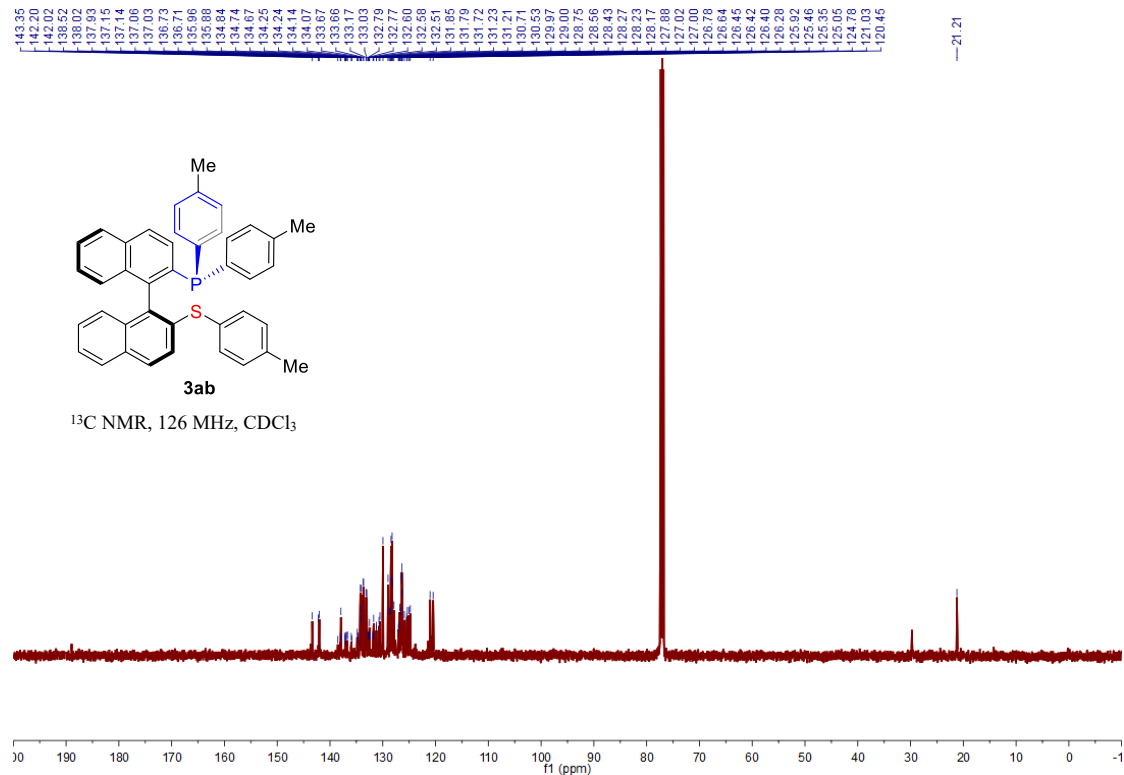


Figure S151. ¹³C NMR spectrum of 3ab (126 MHz, CDCl₃)

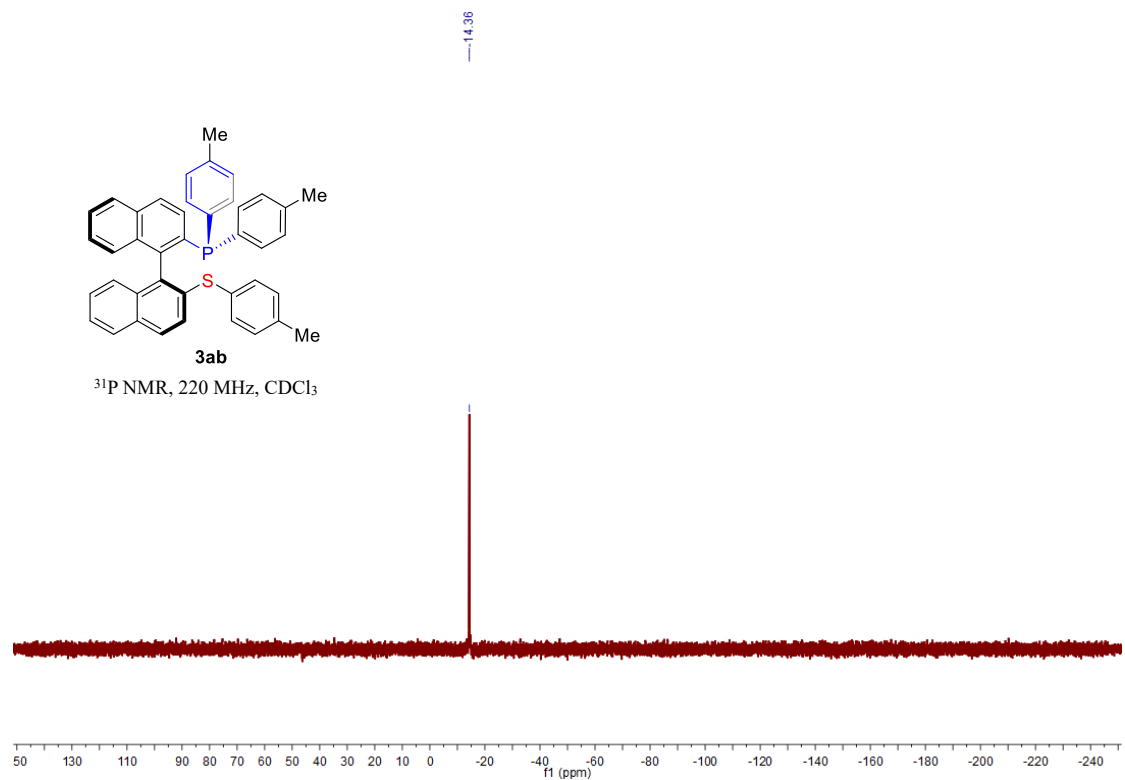


Figure S152. ^{31}P NMR spectrum of **3ab** (202 MHz, CDCl_3)

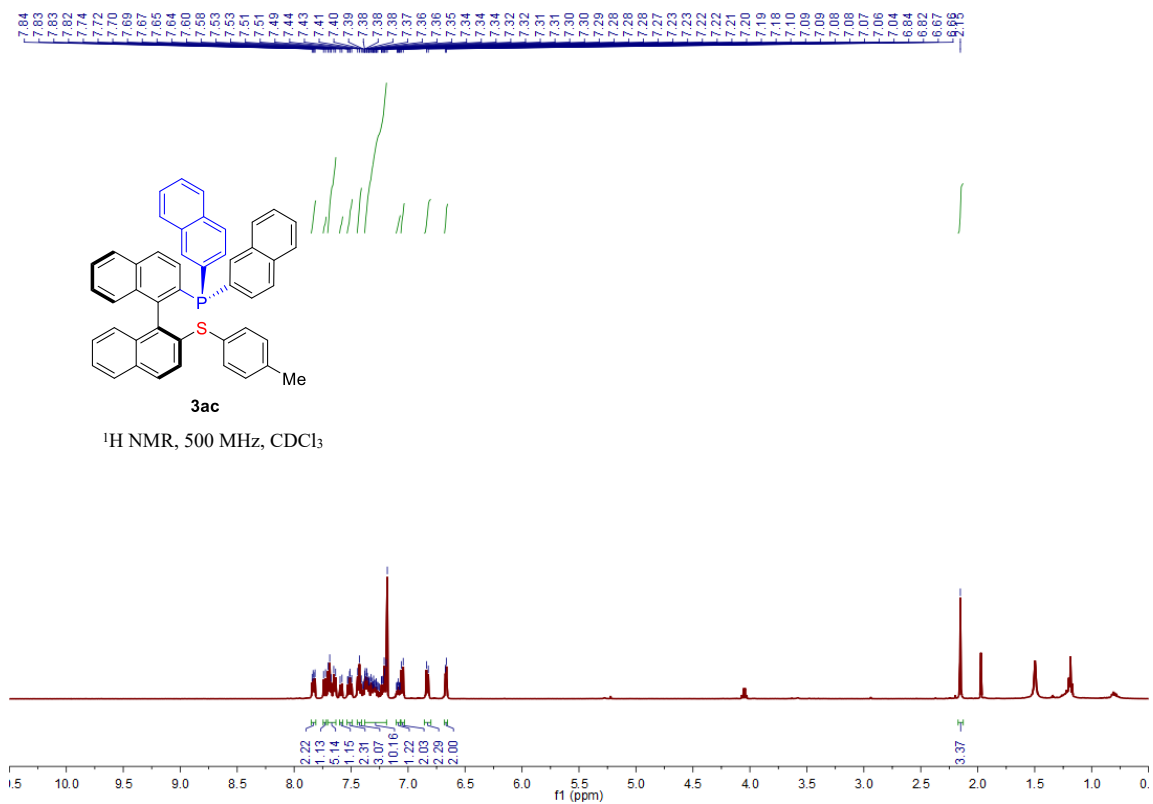


Figure S153. ^1H NMR spectrum of **3ac** (500 MHz, CDCl_3)

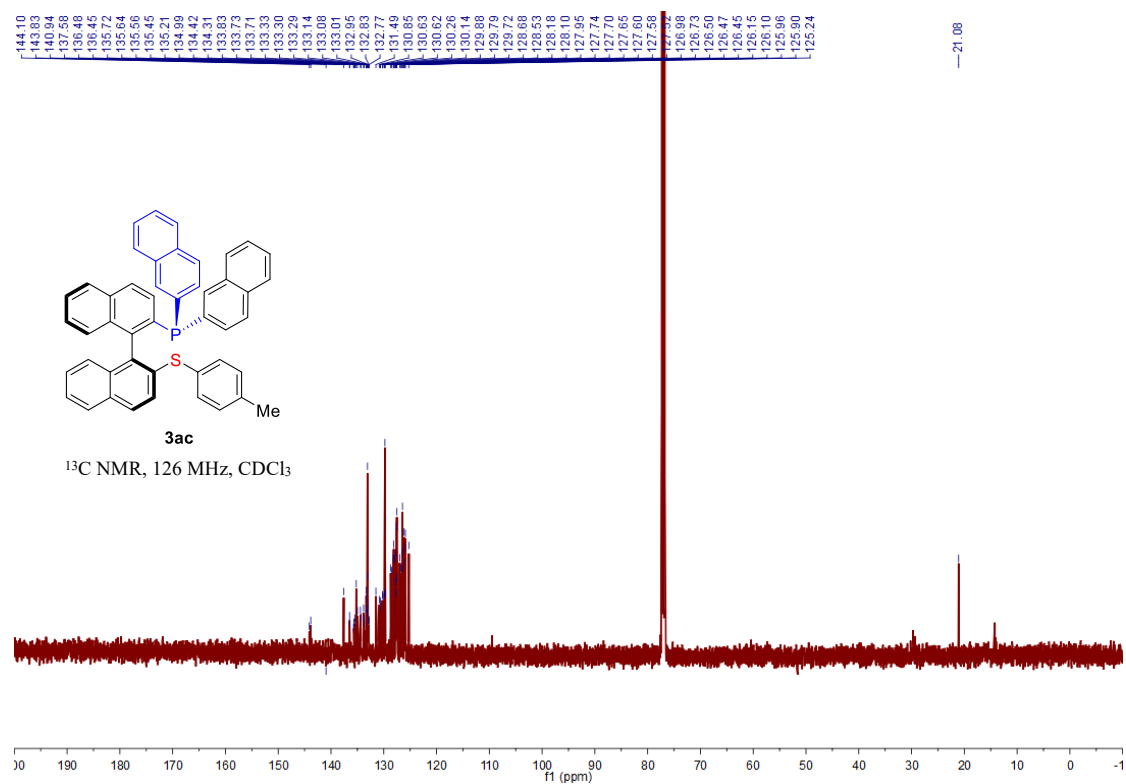


Figure S154. ^{13}C NMR spectrum of 3ac (126 MHz, CDCl_3)

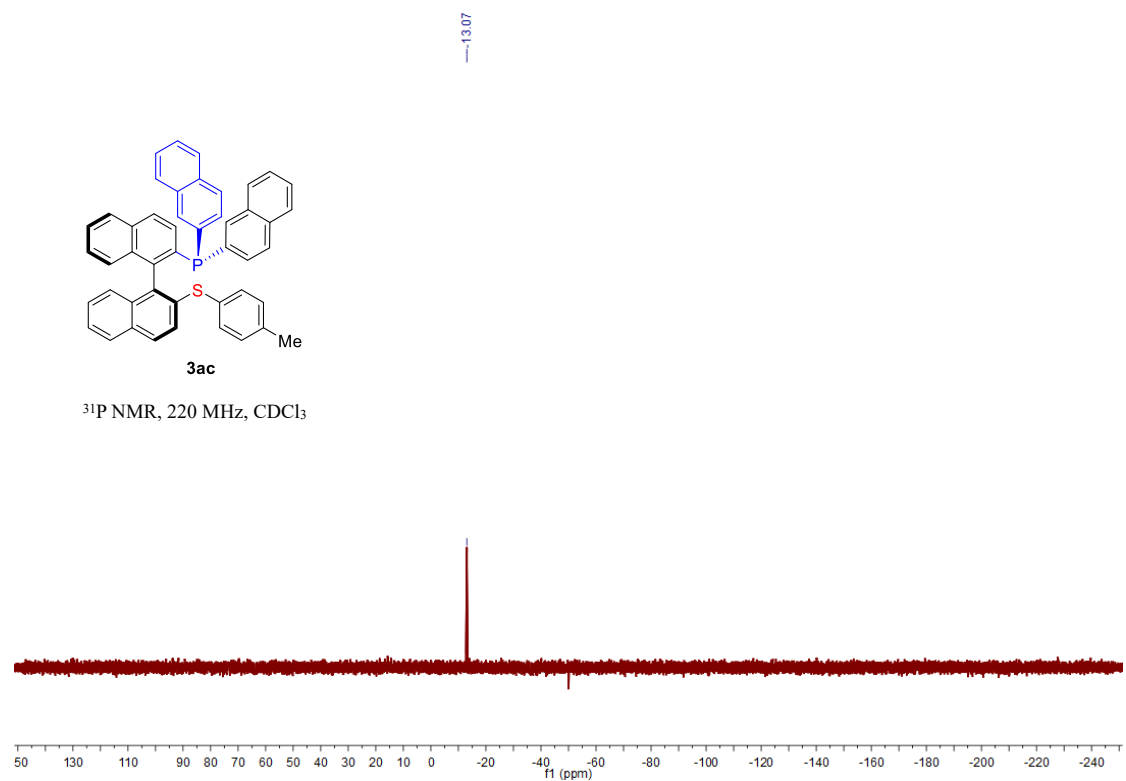


Figure S155. ^{31}P NMR spectrum of 3ac (202 MHz, CDCl_3)

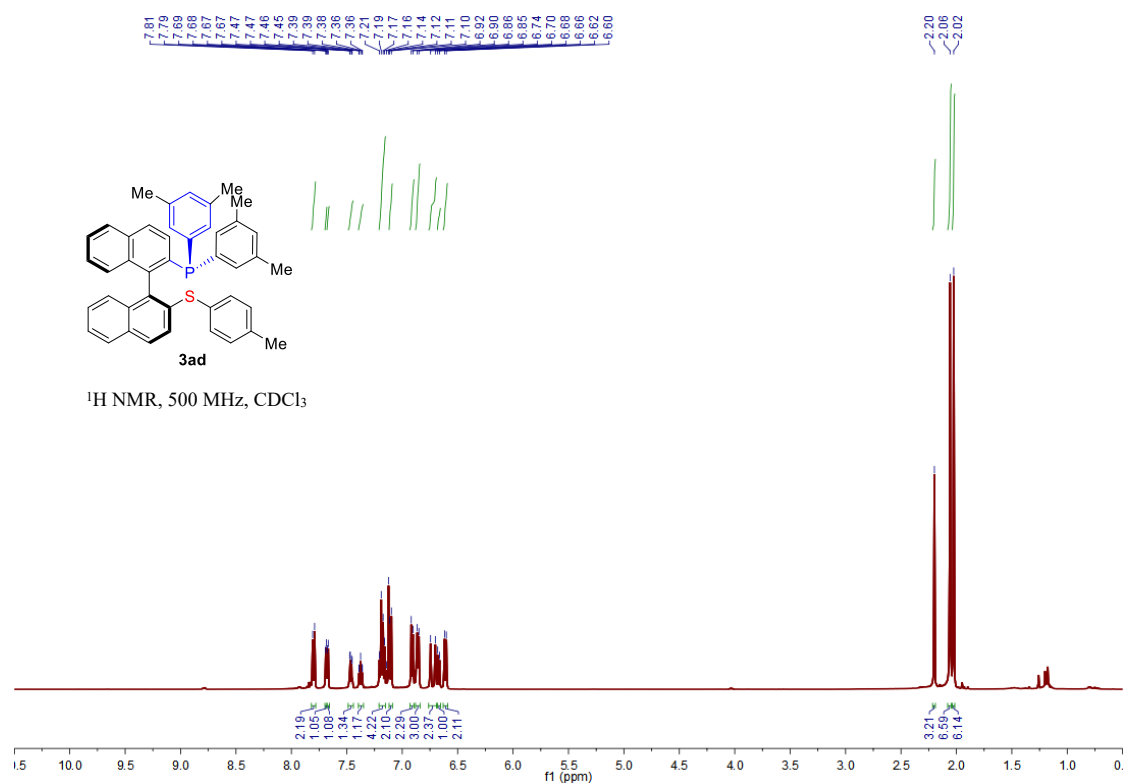


Figure S156. ¹H NMR spectrum of 3ad (500 MHz, CDCl₃)

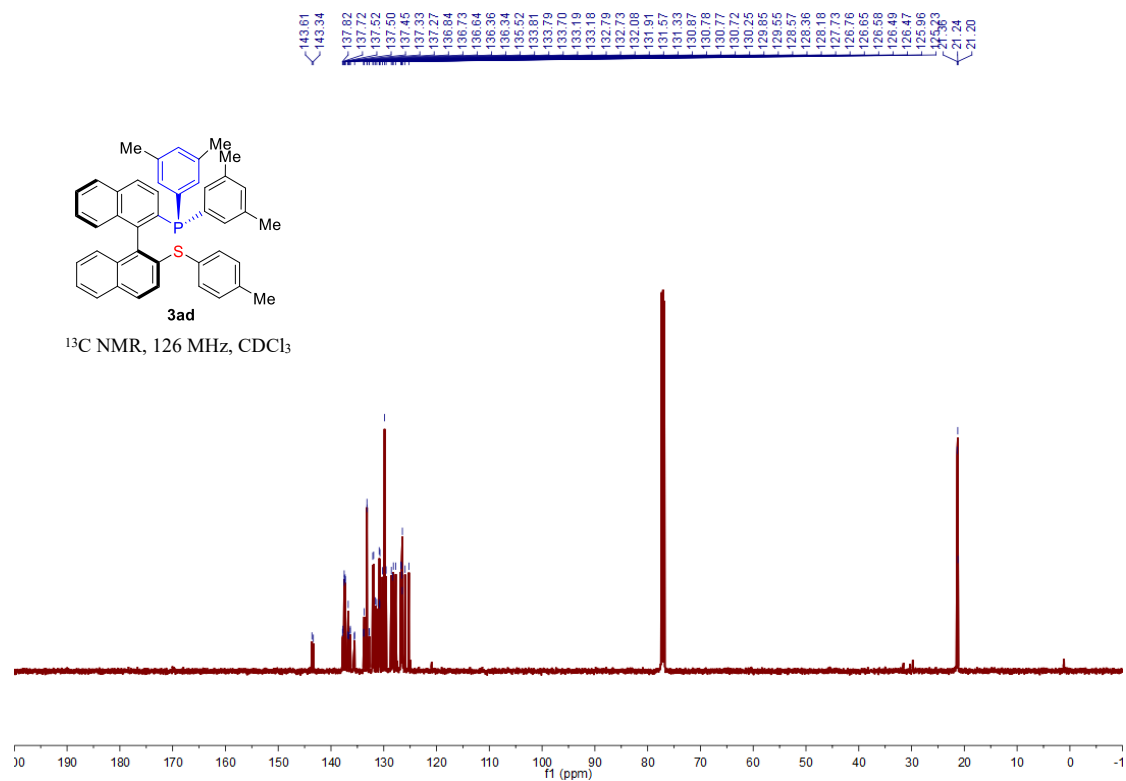


Figure S157. ¹³C NMR spectrum of 3ad (126 MHz, CDCl₃)

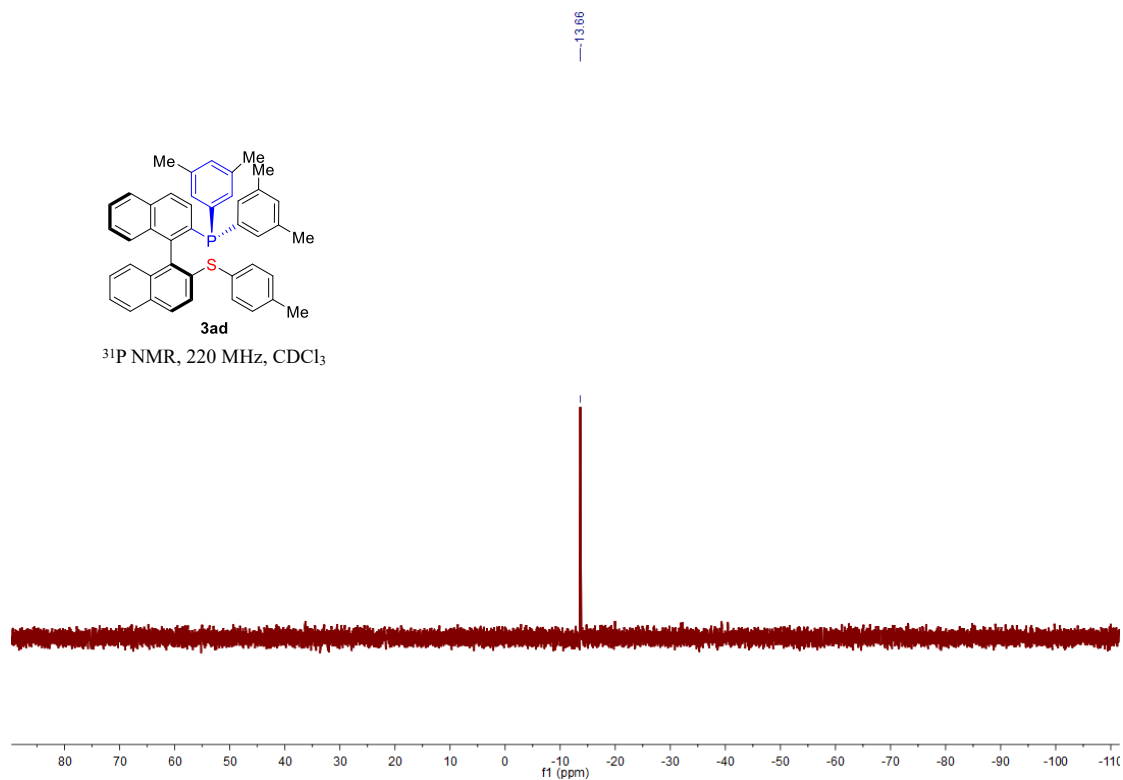


Figure S158. ^{31}P NMR spectrum of 3ad (202 MHz, CDCl_3)

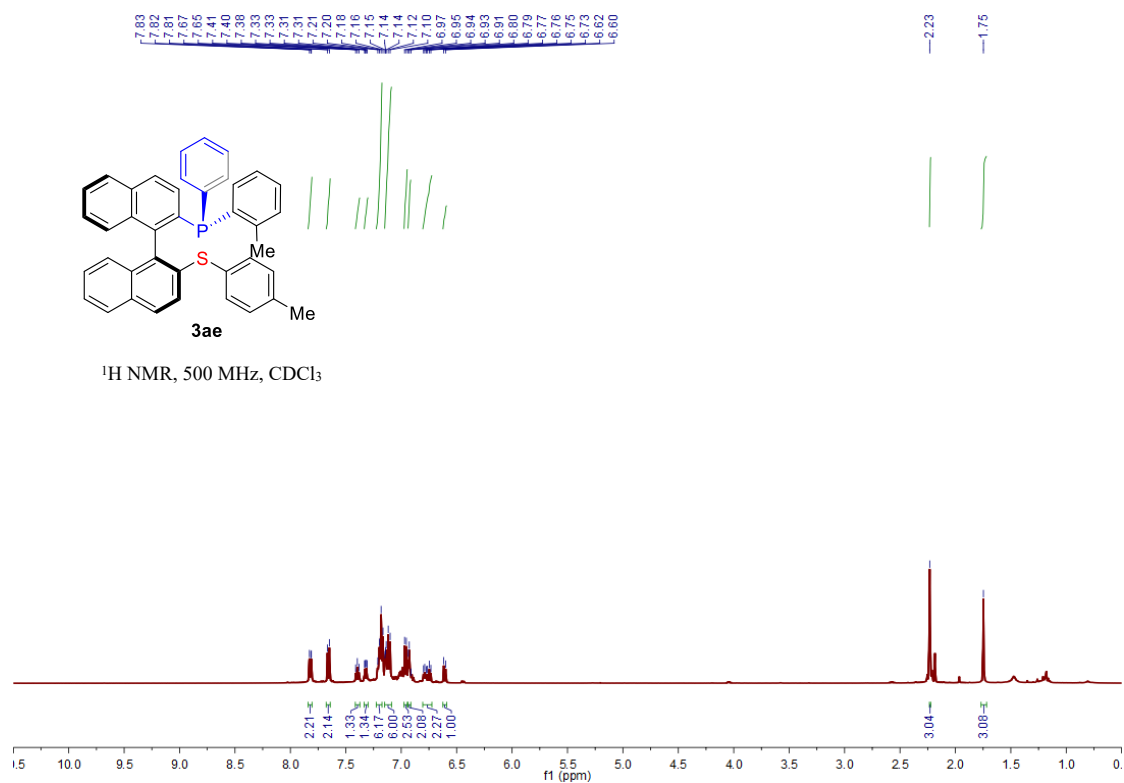


Figure S159. ^1H NMR spectrum of 3ae (500 MHz, CDCl_3)

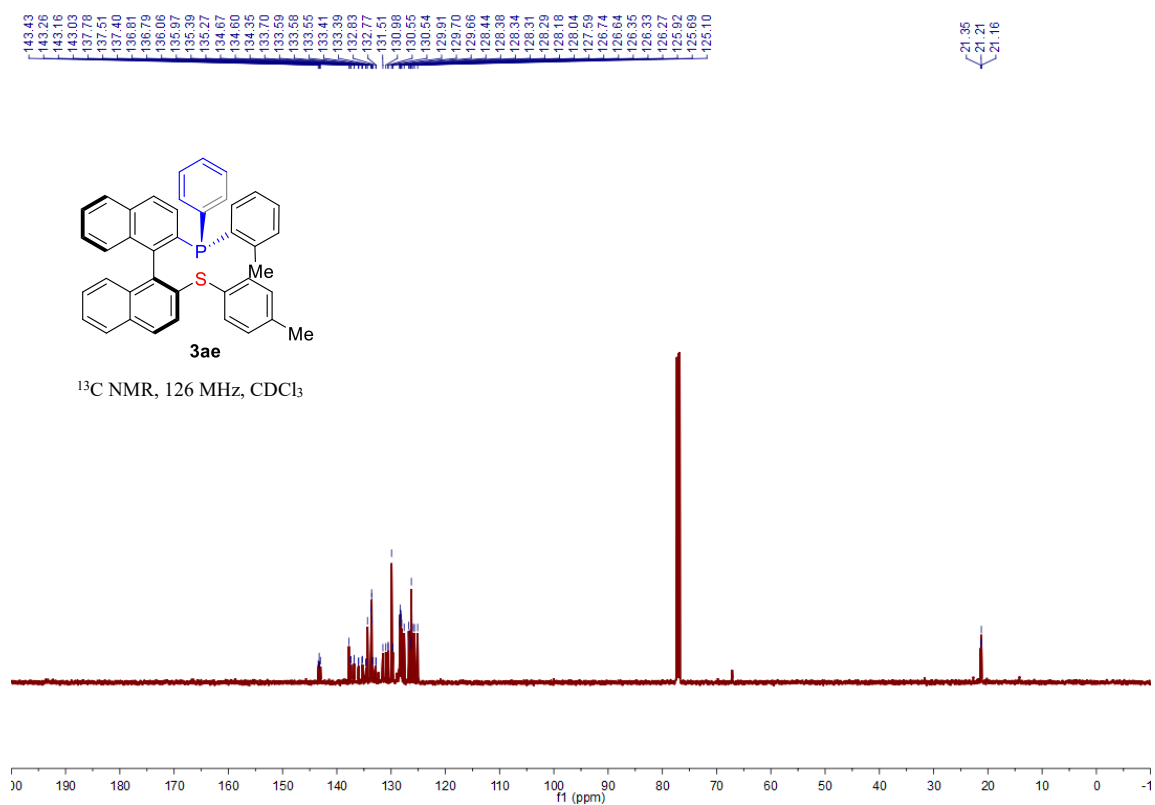


Figure S160. ^{13}C NMR spectrum of **3ae** (126 MHz, CDCl_3)

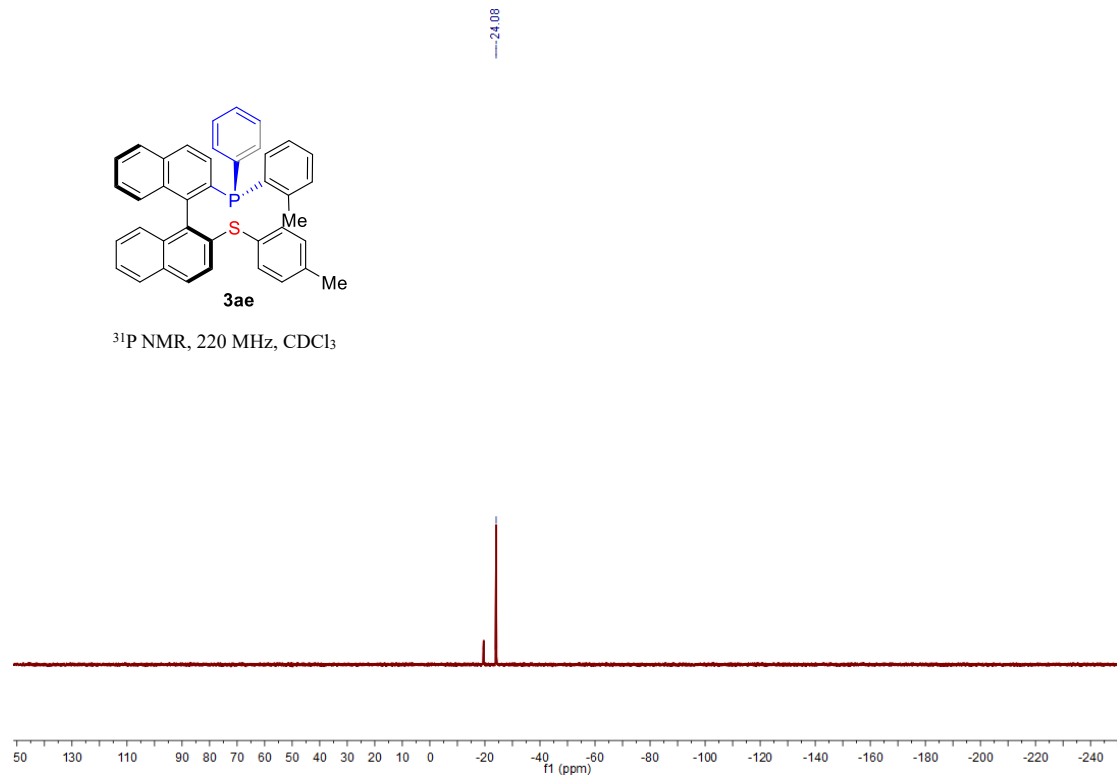


Figure S161. ^{31}P NMR spectrum of **3ae** (202 MHz, CDCl_3)

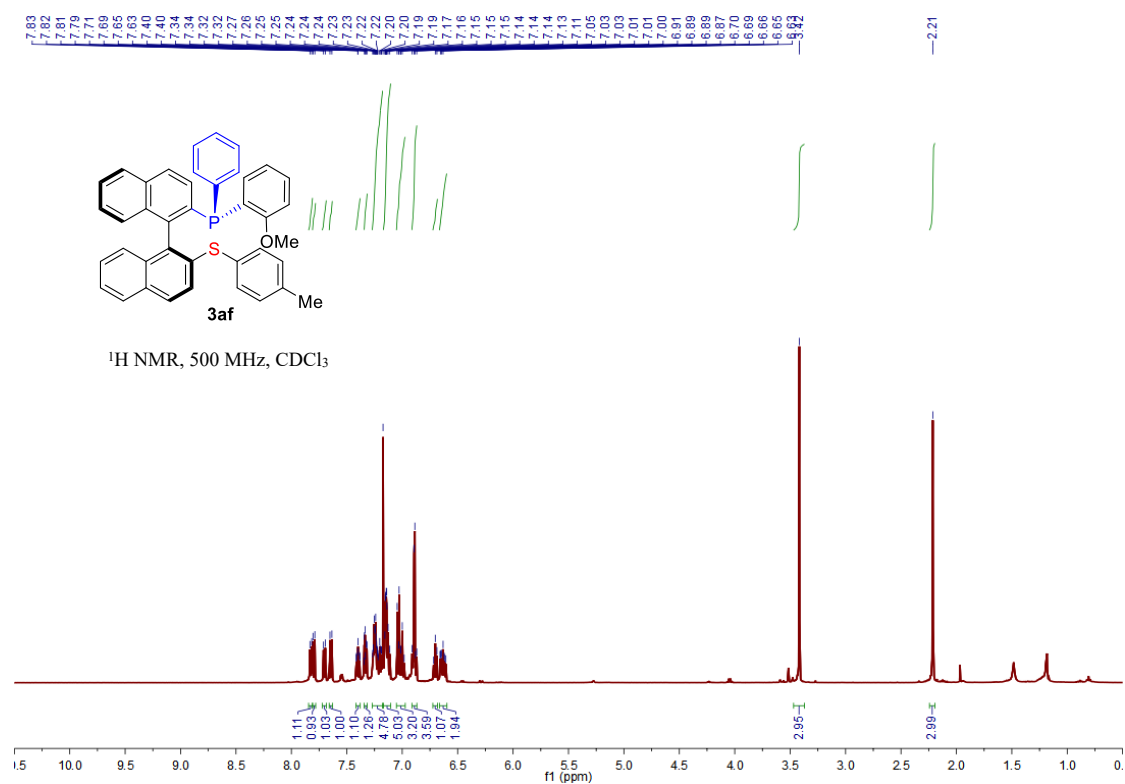


Figure S162. ¹H NMR spectrum of 3af (500 MHz, CDCl₃)

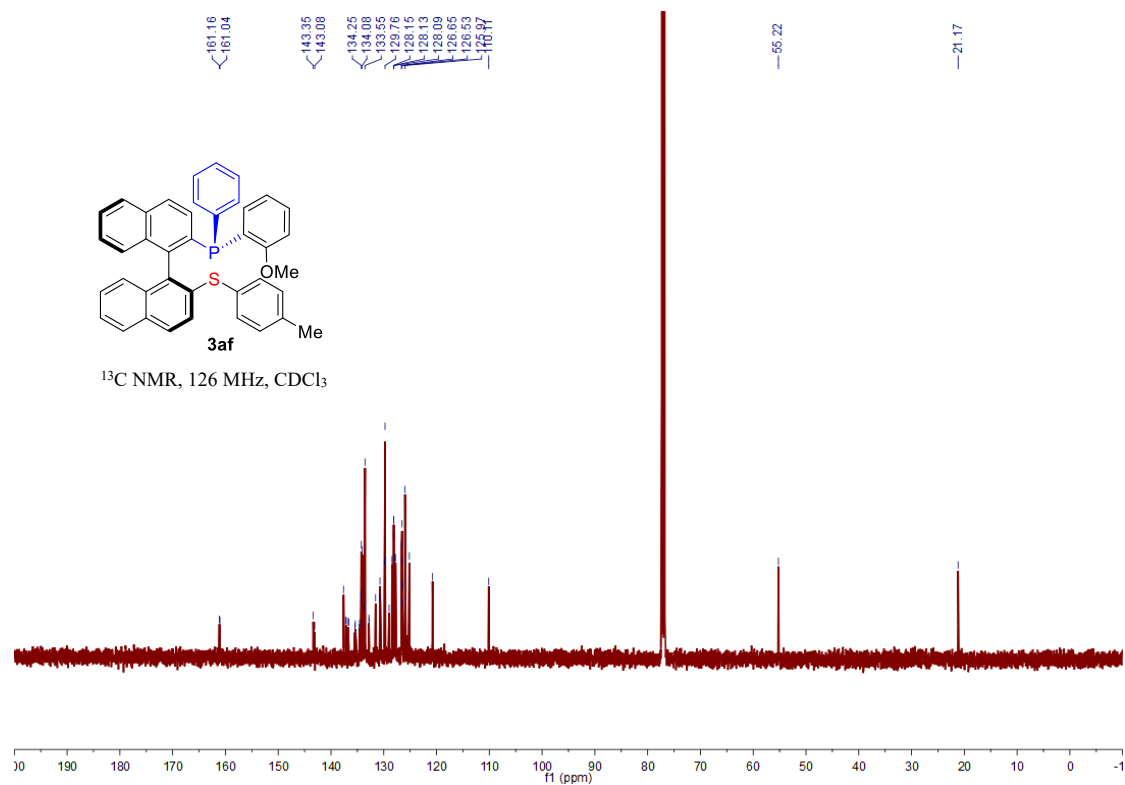


Figure S163. ¹³C NMR spectrum of 3af (126 MHz, CDCl₃)

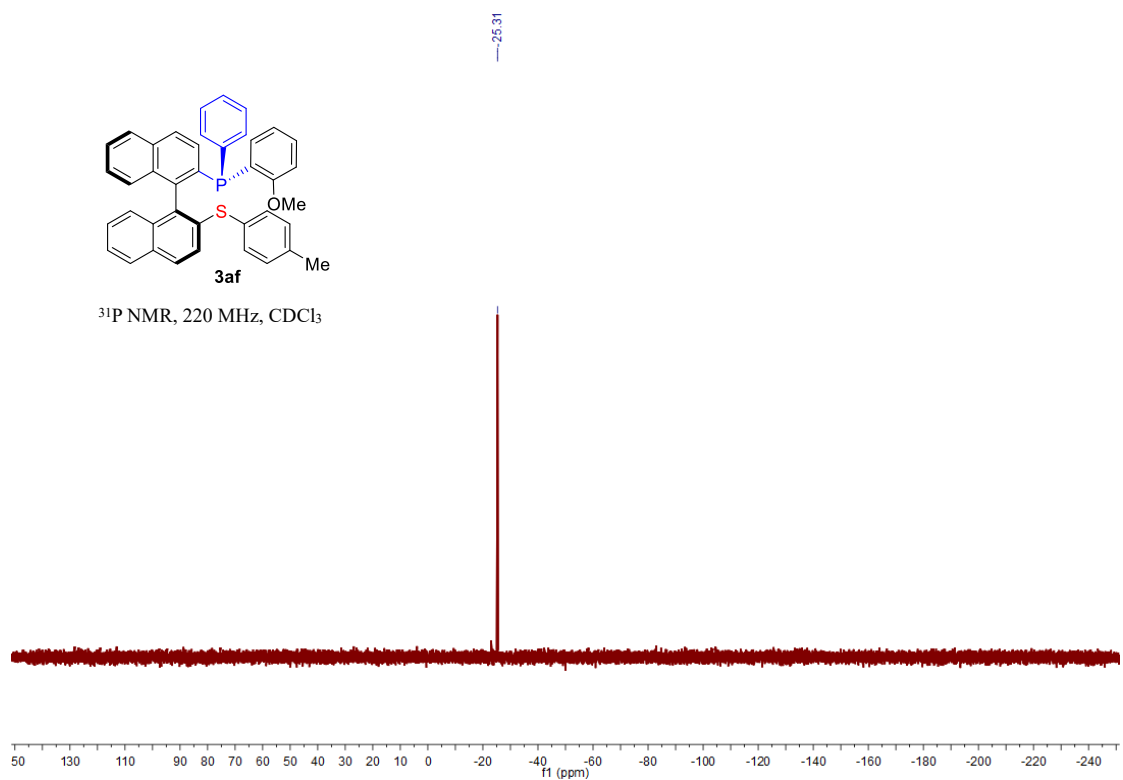


Figure S164. ^{31}P NMR spectrum of **3af** (202 MHz, CDCl_3)

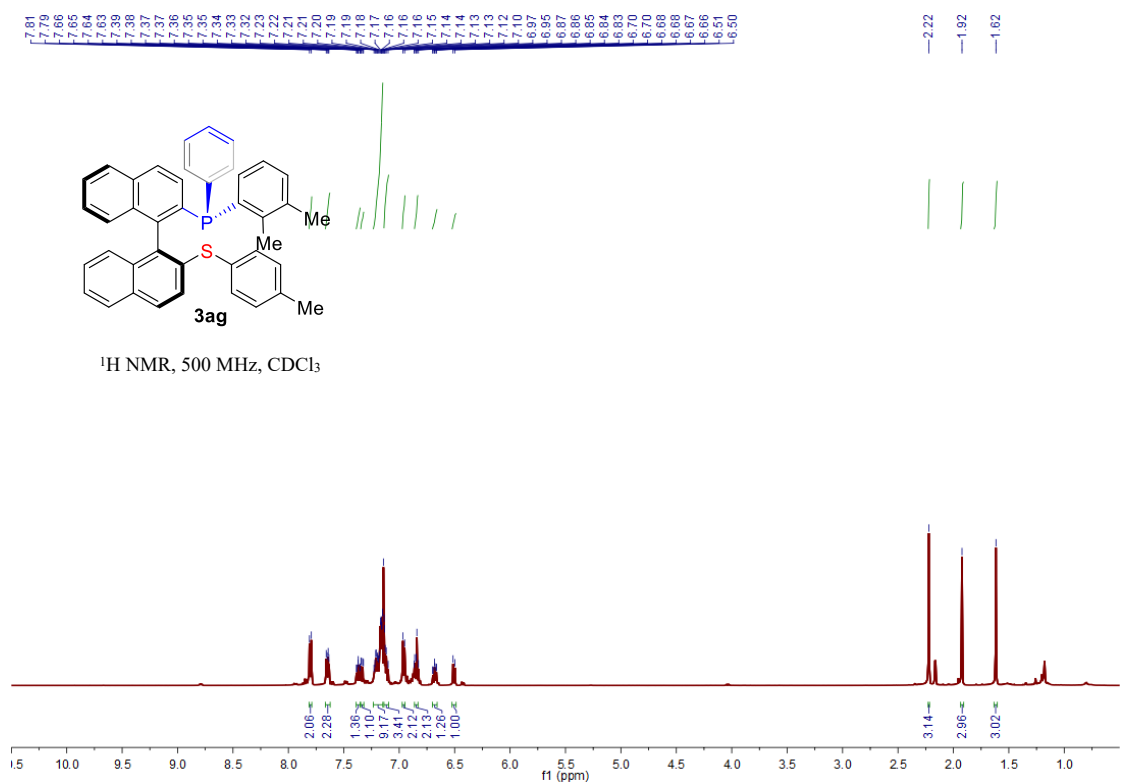


Figure S165. ^1H NMR spectrum of **3ag** (500 MHz, CDCl_3)

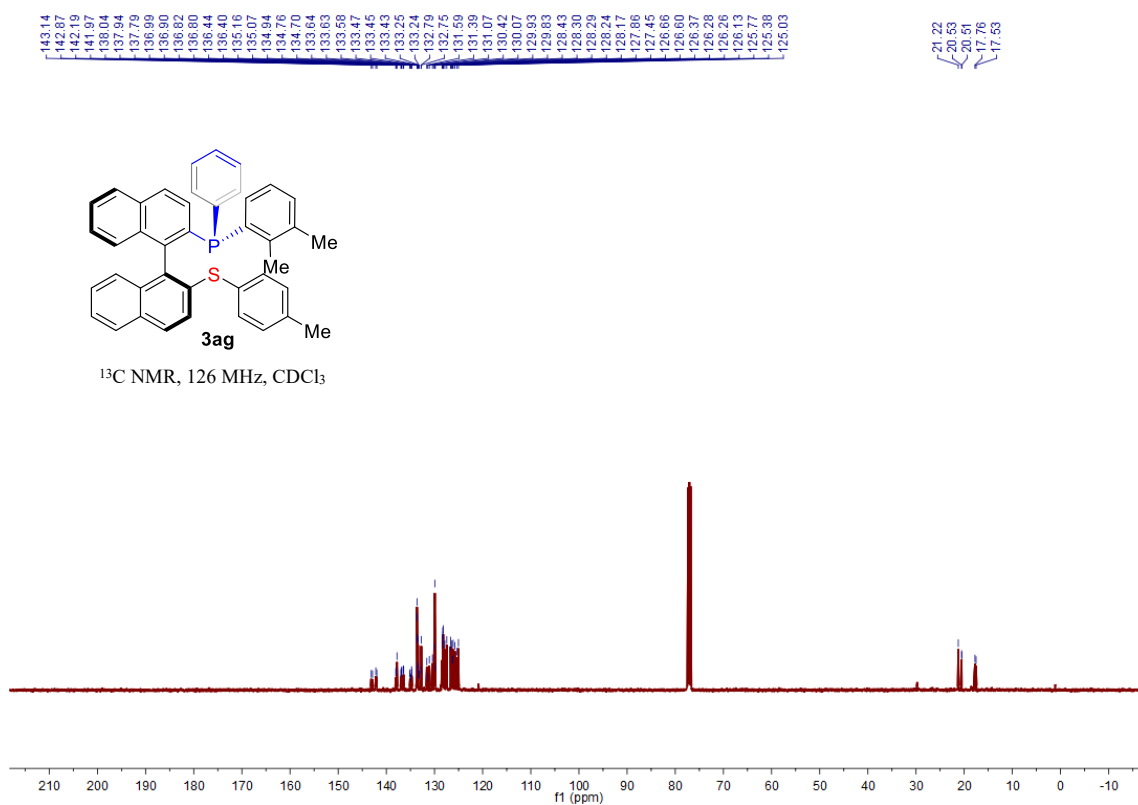


Figure S166. ¹³C NMR spectrum of **3ag** (126 MHz, CDCl₃)

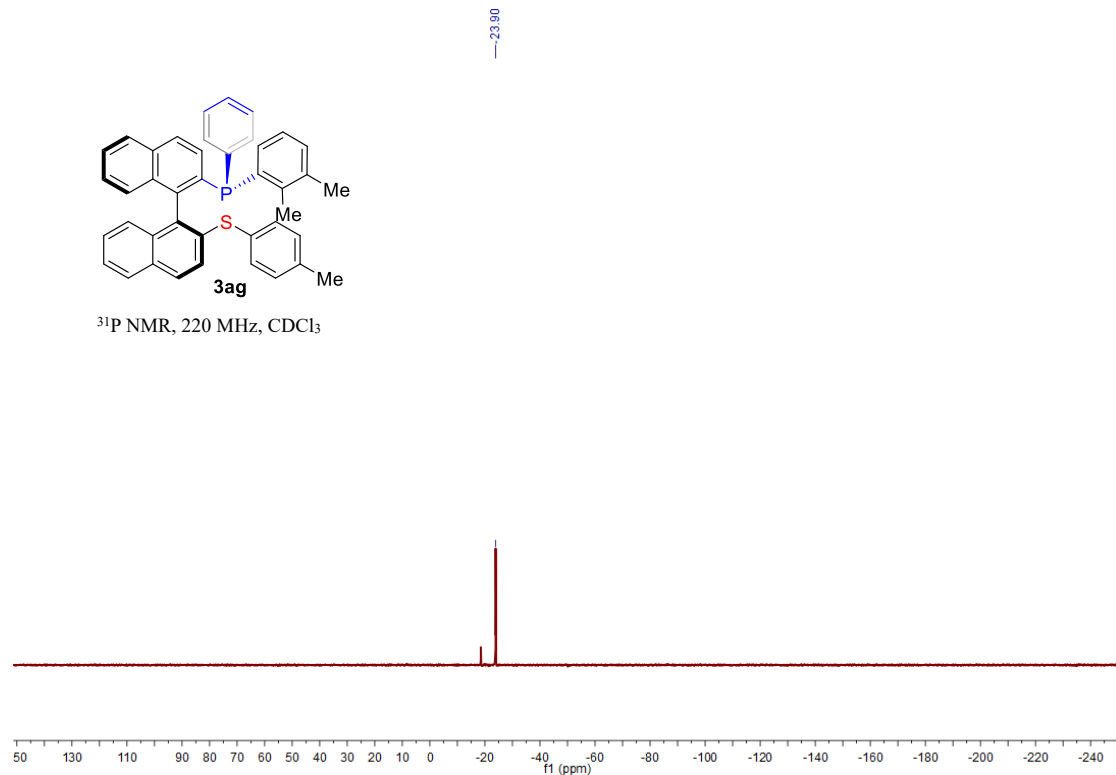


Figure S167. ³¹P NMR spectrum of **3ag** (202 MHz, CDCl₃)

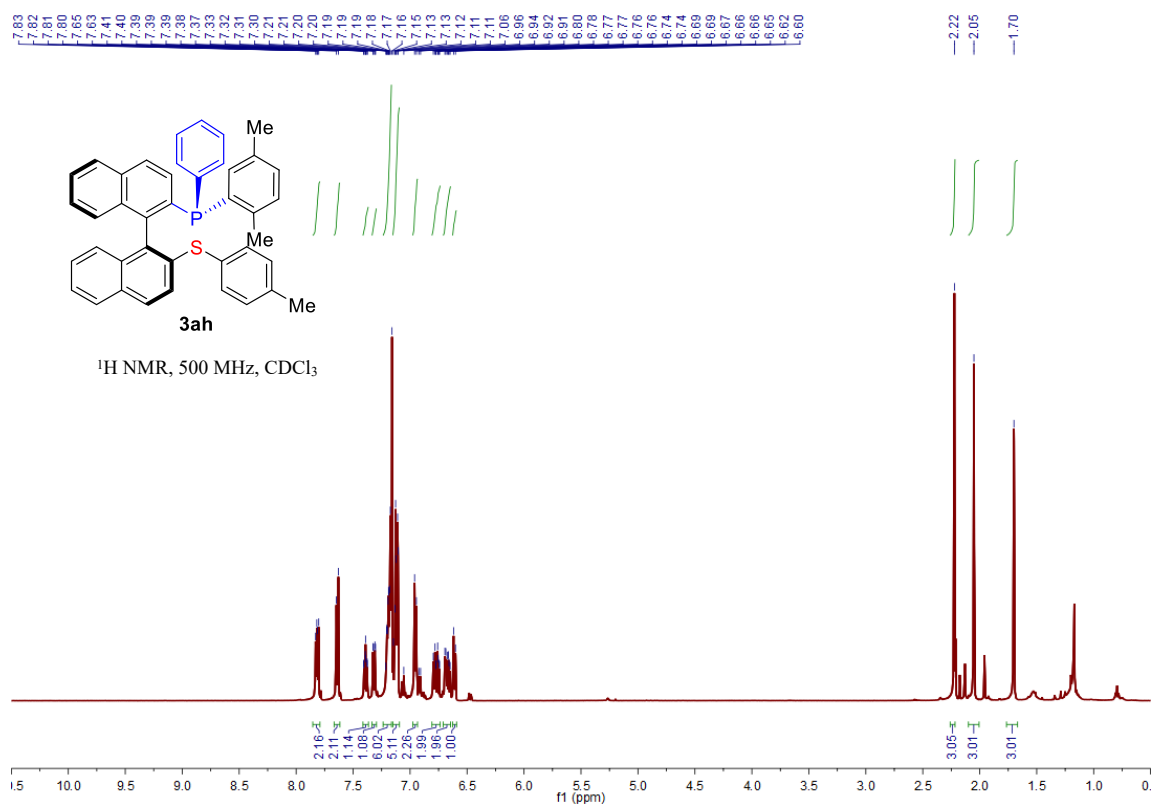


Figure S168. ^1H NMR spectrum of 3ah (500 MHz, CDCl_3)

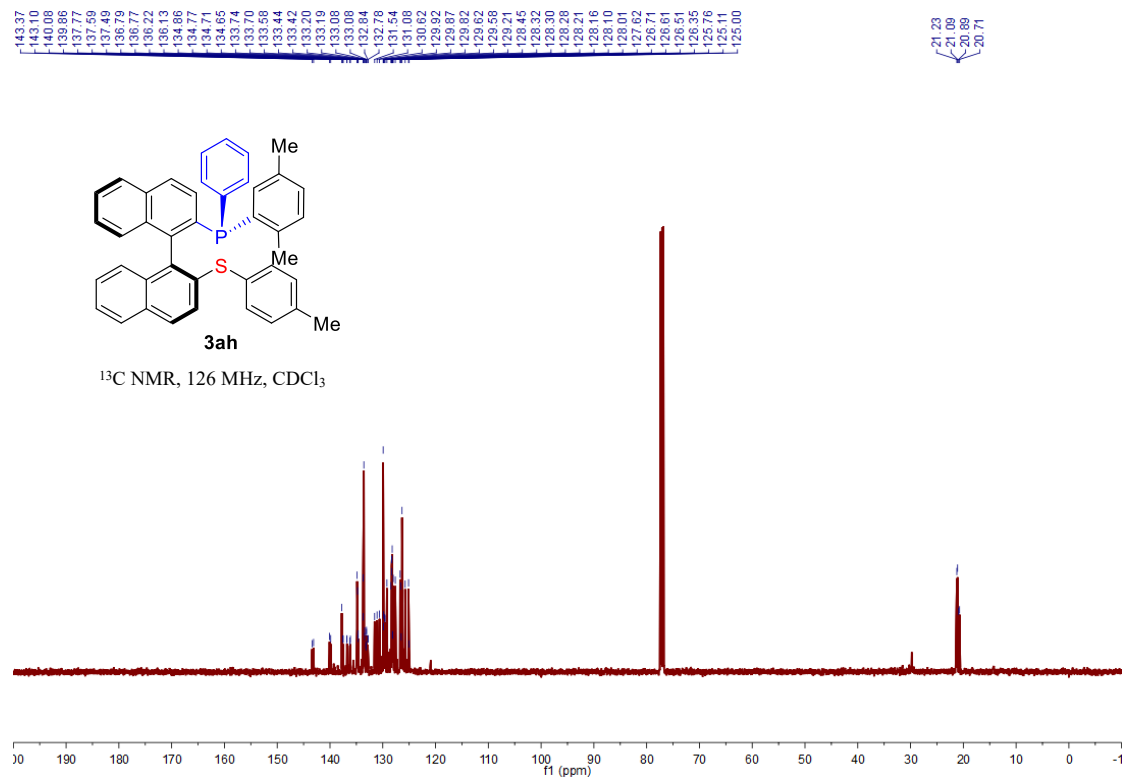


Figure S169. ^{13}C NMR spectrum of 3ah (126 MHz, CDCl_3)

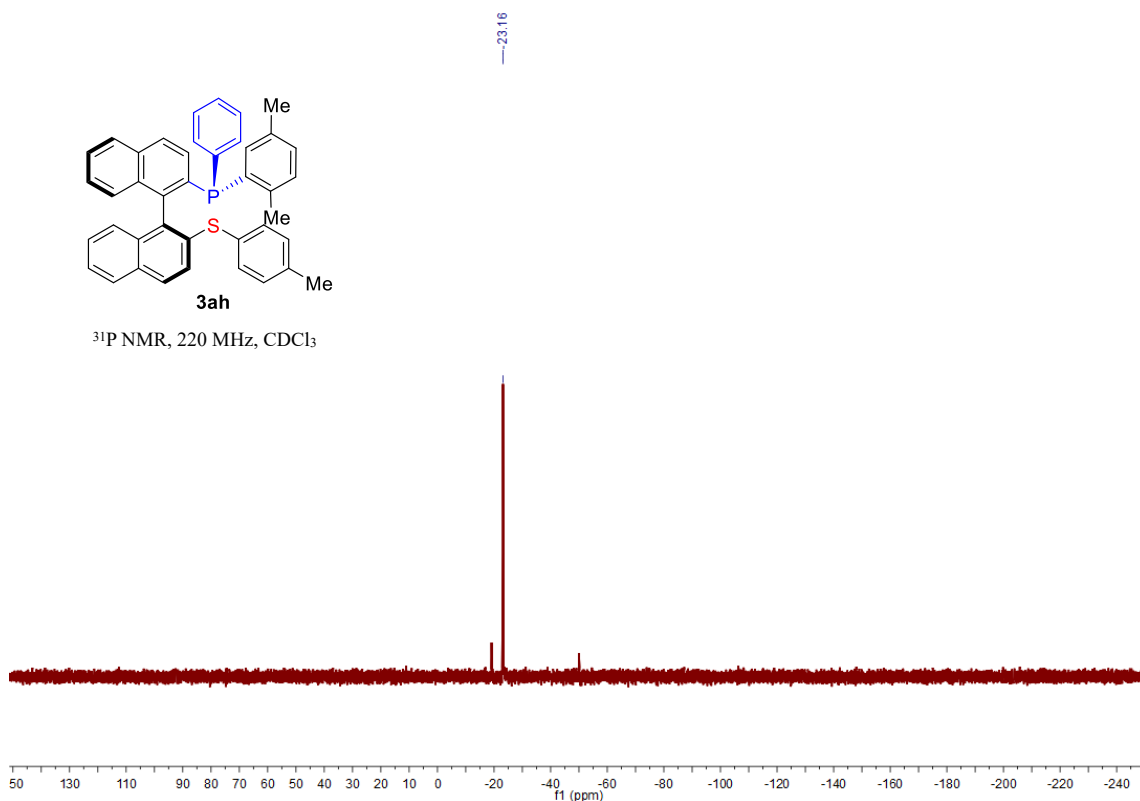


Figure S170. ^{31}P NMR spectrum of **3ah** (202 MHz, CDCl_3)

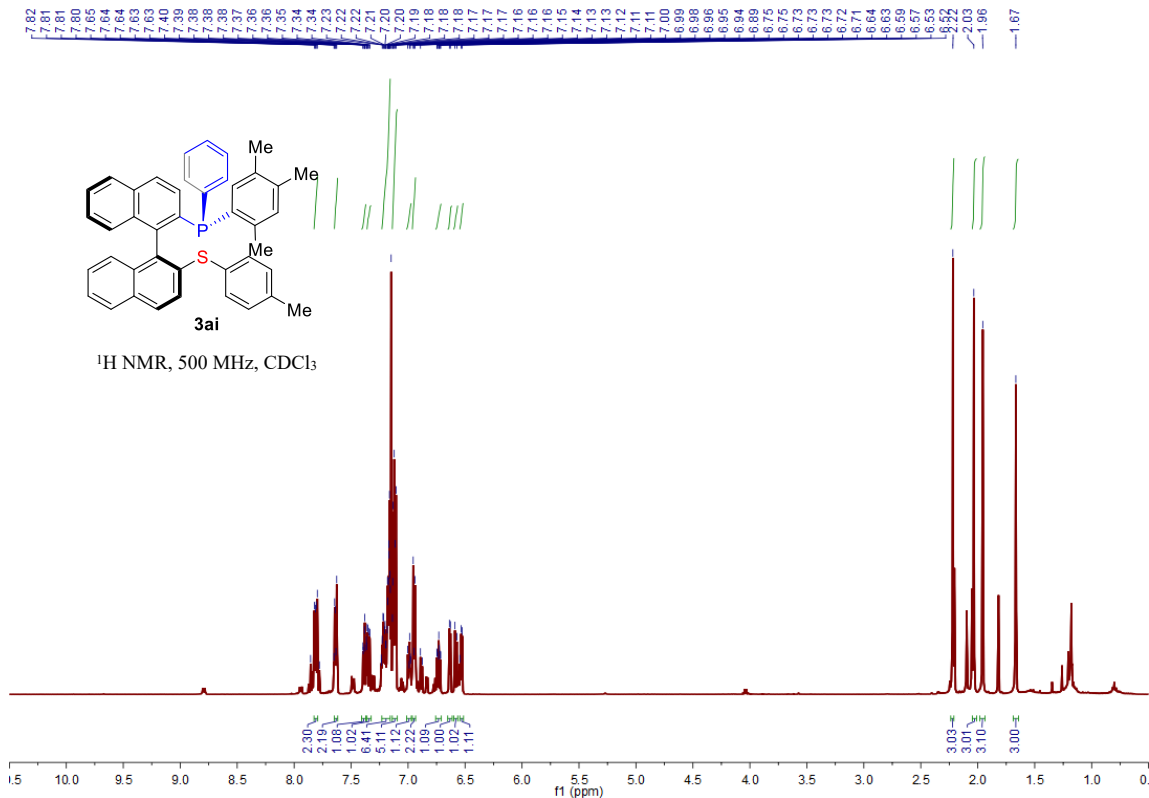


Figure S171. ^1H NMR spectrum of **3ai** (500 MHz, CDCl_3)

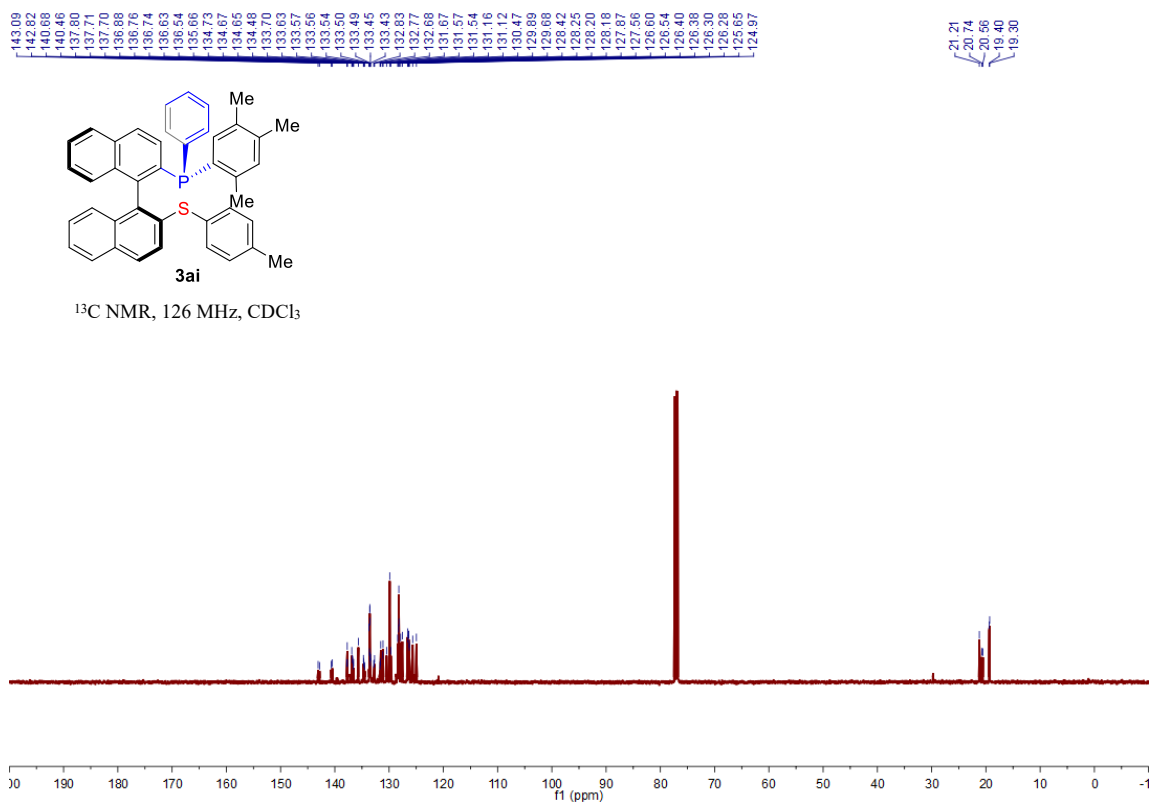


Figure S172. ^{13}C NMR spectrum of **3ai** (126 MHz, CDCl_3)

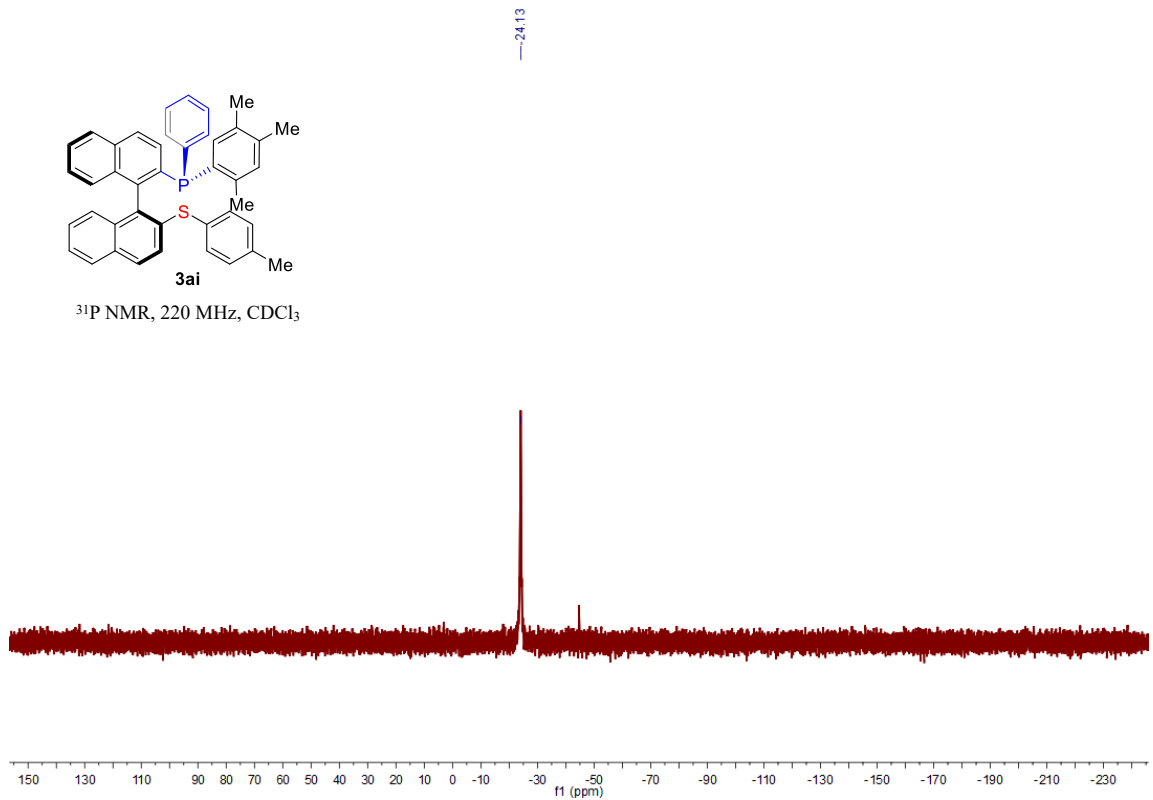


Figure S173. ^{31}P NMR spectrum of **3ai** (202 MHz, CDCl_3)

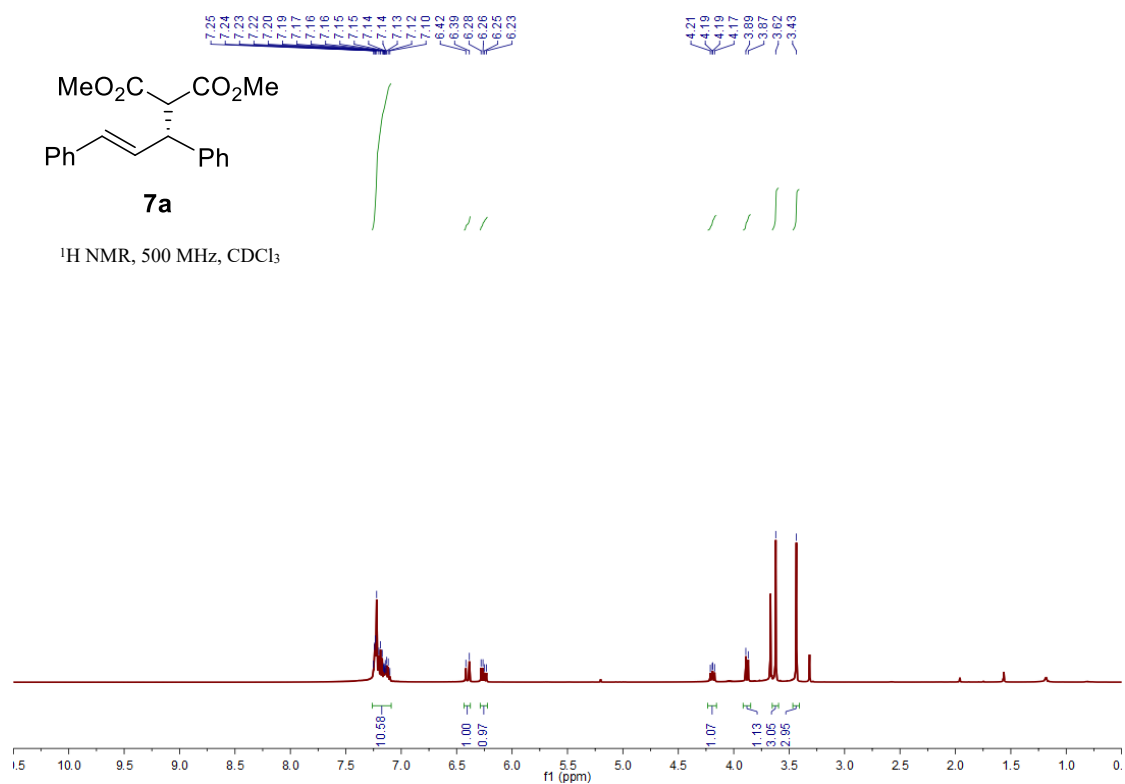


Figure S174. ^1H NMR spectrum of **7a** (500 MHz, CDCl_3)

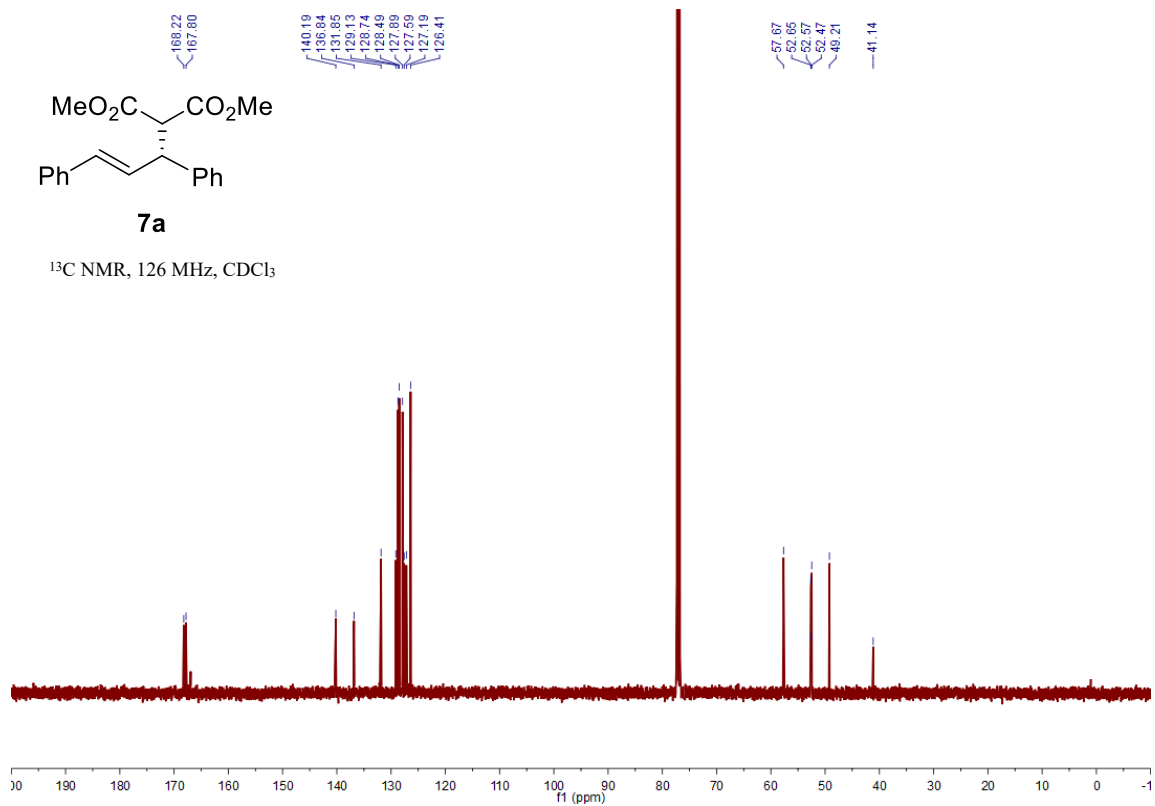


Figure S175. ^{13}C NMR spectrum of **7a** (126 MHz, CDCl_3)

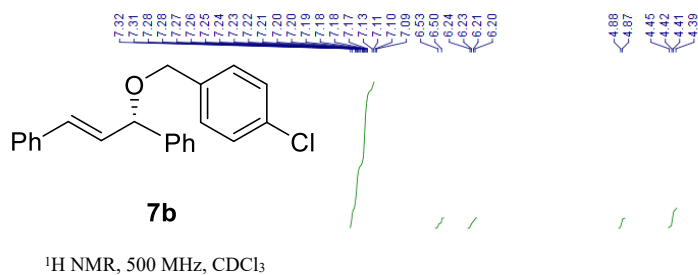


Figure S176. ^1H NMR spectrum of **7b** (500 MHz, CDCl_3)

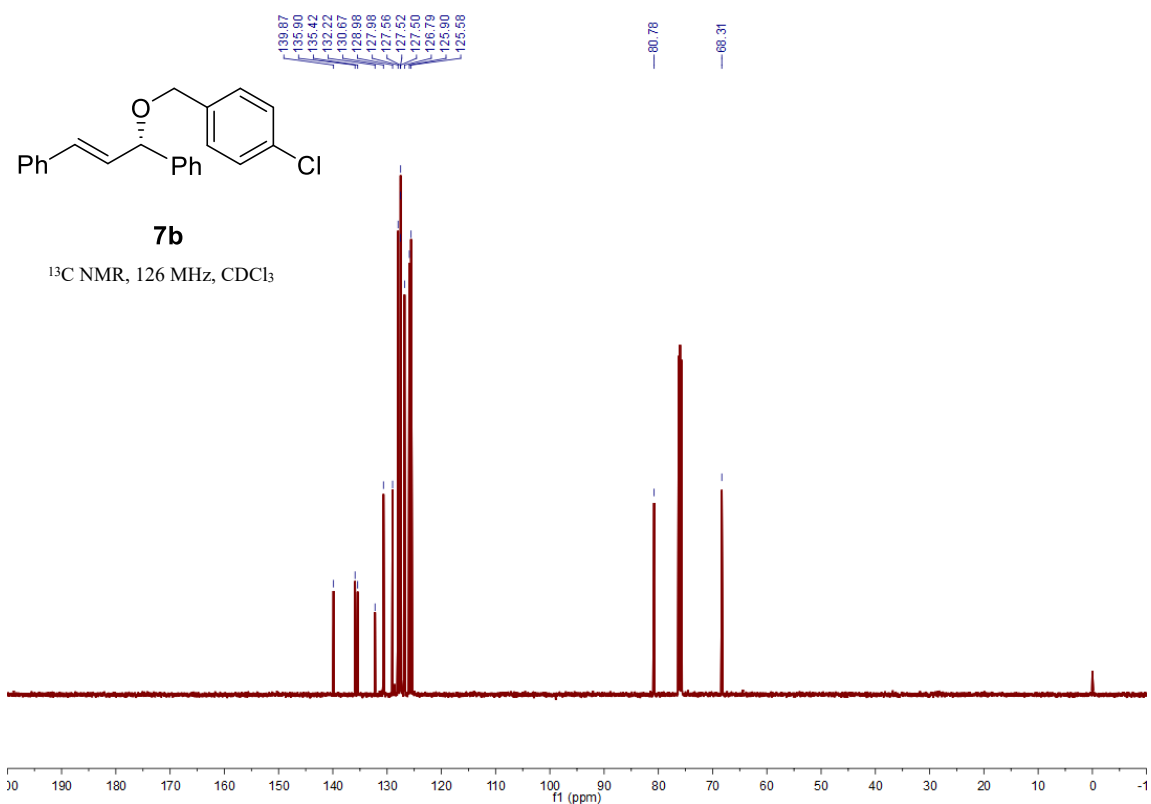


Figure S177. ^{13}C NMR spectrum of **7b** (126 MHz, CDCl_3)

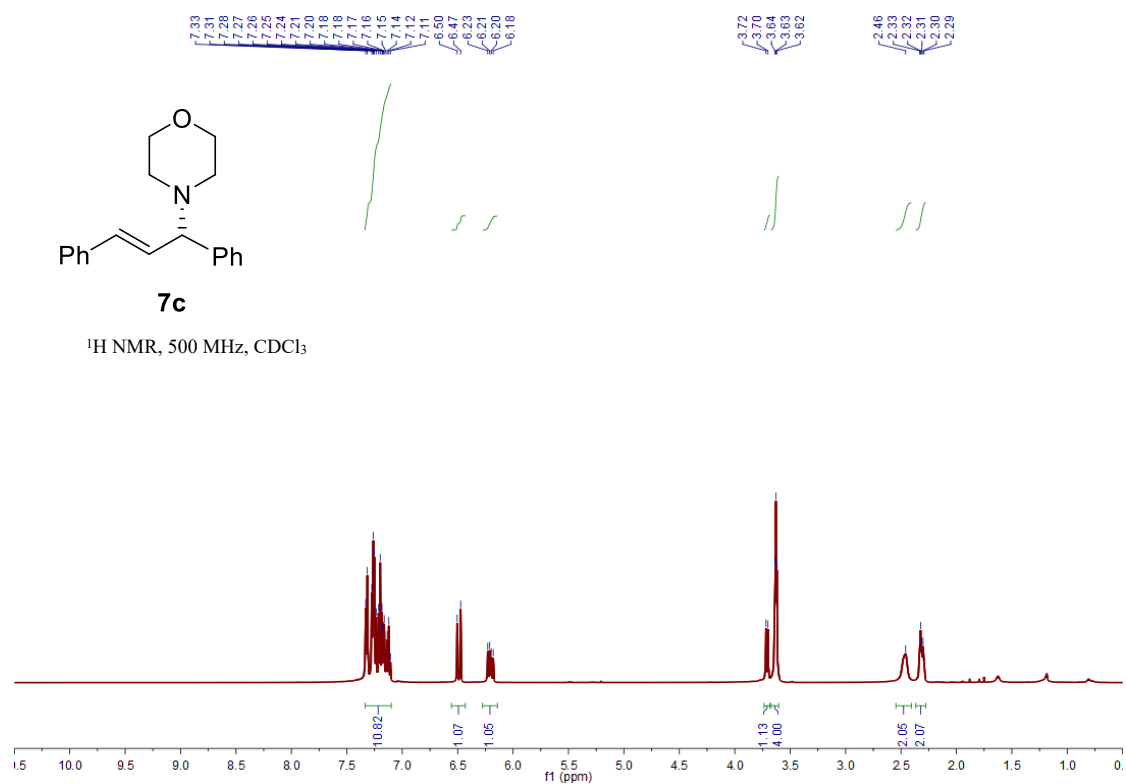


Figure S178. ^1H NMR spectrum of **7c** (500 MHz, CDCl_3)

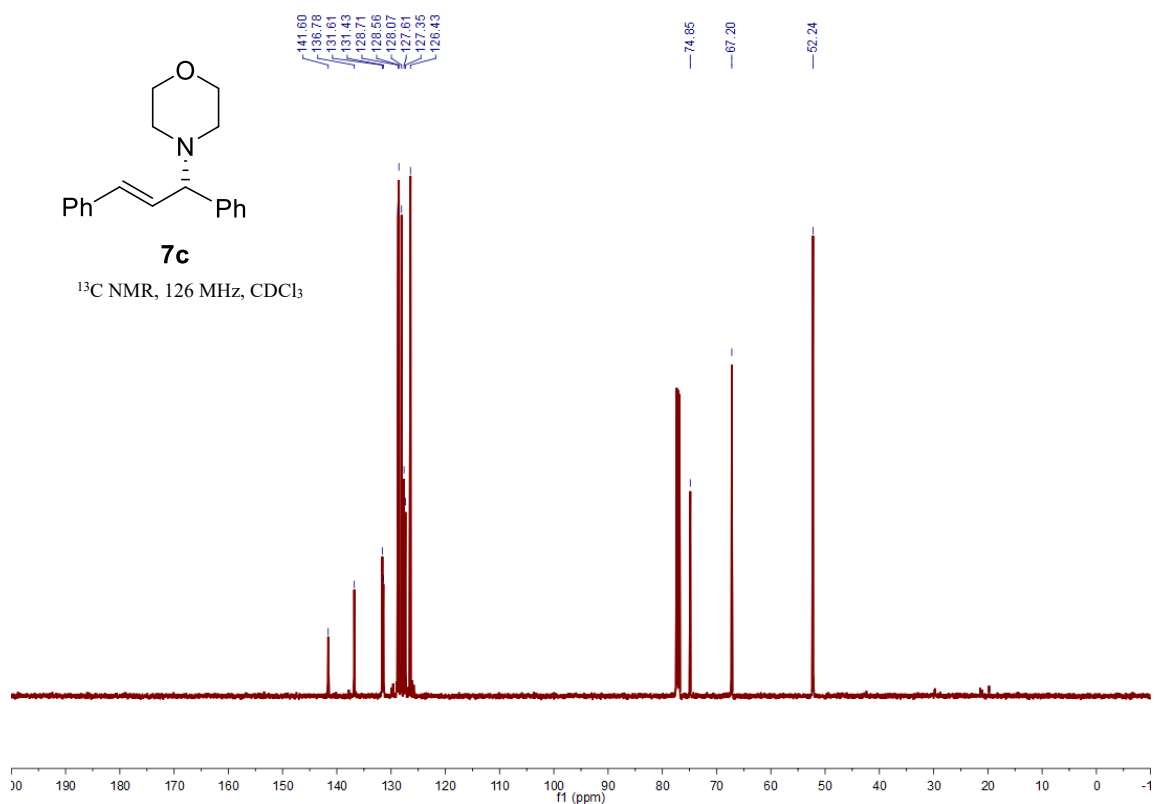
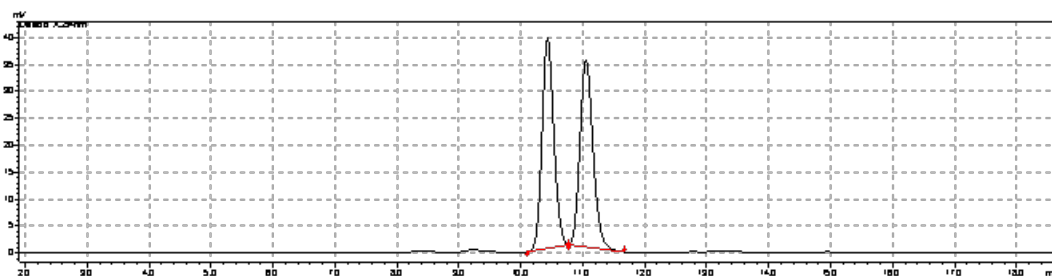
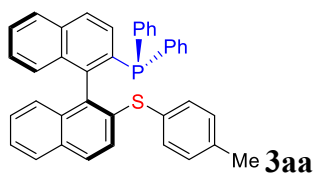


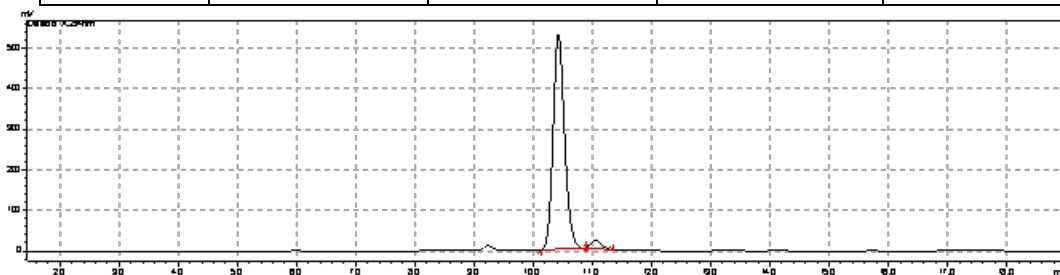
Figure S178. ^{13}C NMR spectrum of **7c** (126 MHz, CDCl_3)

HPLC Charts



Detector A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	10.426	515491	38951	50.243
2	11.046	510509	34763	49.757
Total		1026000	73714	100.000

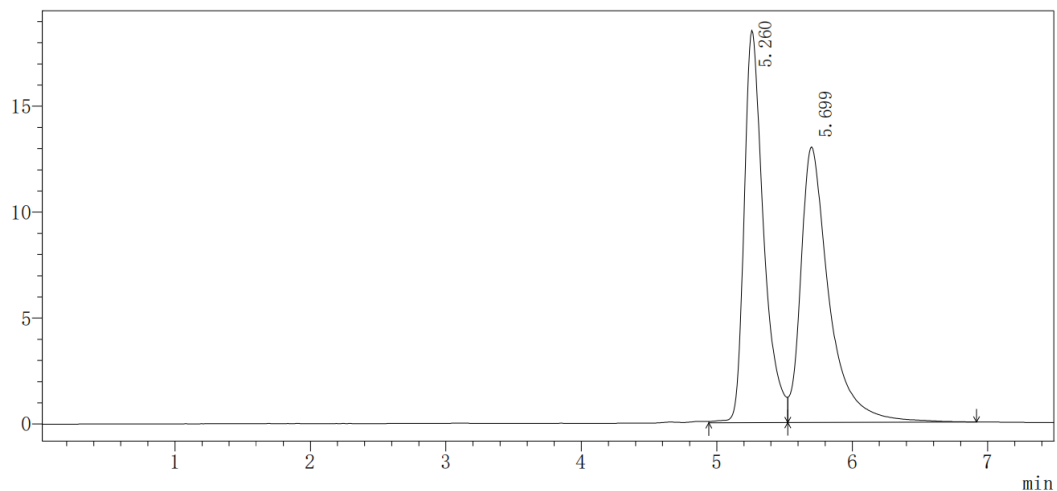


Detector A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	10.420	7085790	528525	96.572
2	11.048	251555	20128	3.428
Total		7337345	548653	100.000

After Recrystallization:

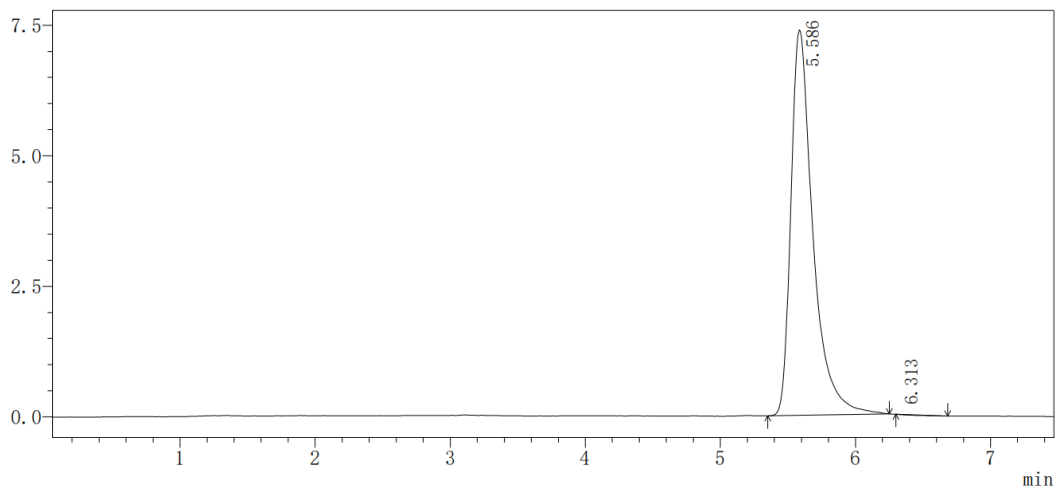
mV



channel 254 nm

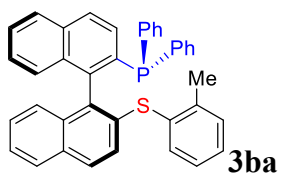
Peak	Ret. Time	Area	Height	Conc.
1	5.260	185738	18514	49.569
2	5.699	188964	13014	50.431
Total		374702	31527	

mV

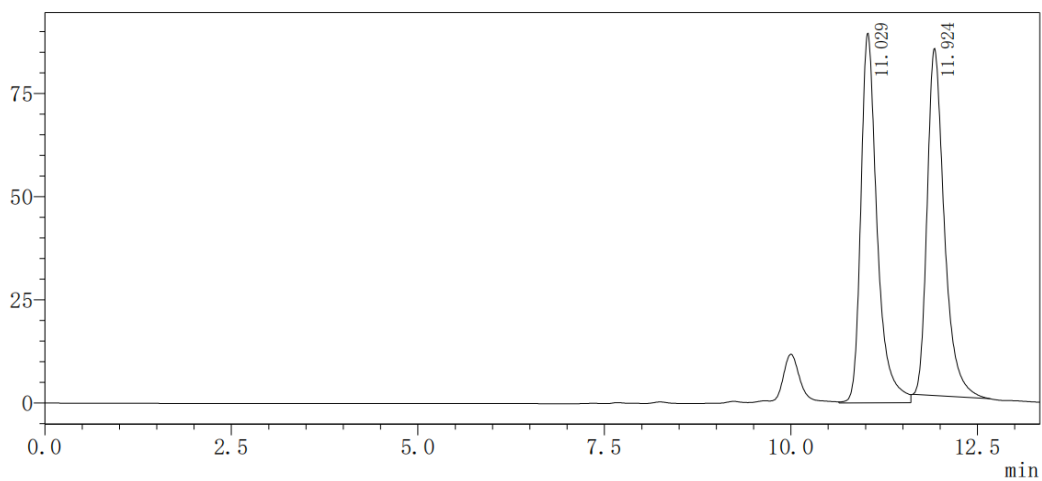


channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	5.586	83682	7376	99.771
2	6.271	192	24	0.229
Total		83874	7400	



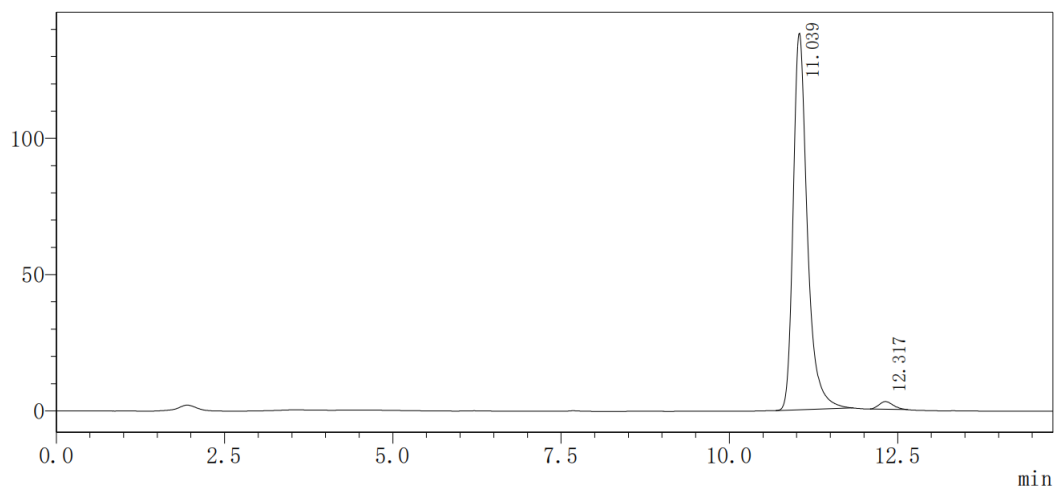
mV



Detector A channel 254 nm

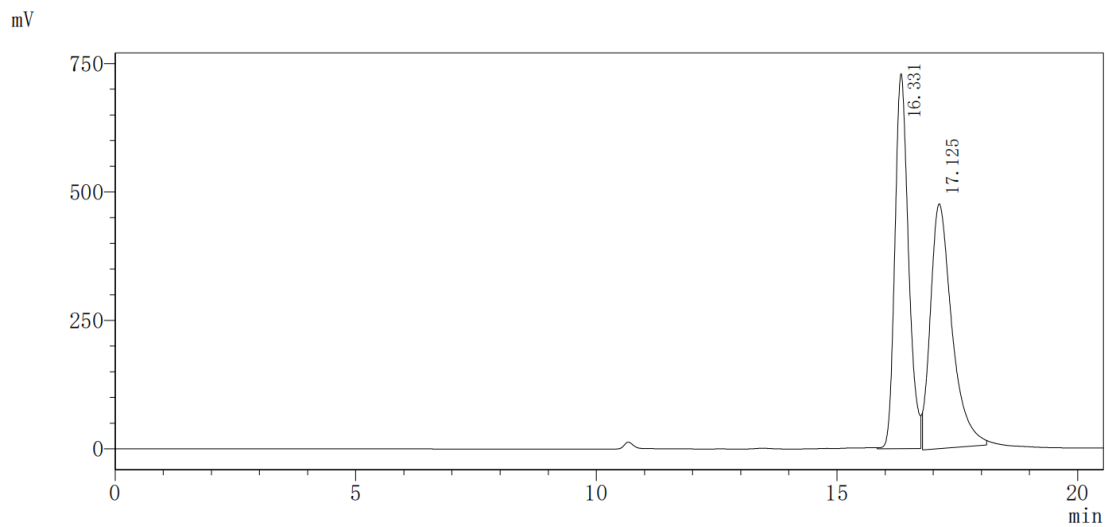
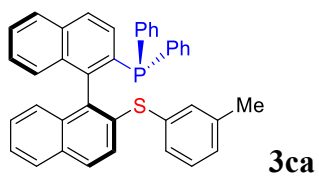
Peak	Ret. Time	Area	Height	Conc.
1	11.029	1366805	89528	50.364
2	11.924	1347036	84083	49.636
Total		2713841	173610	

mV



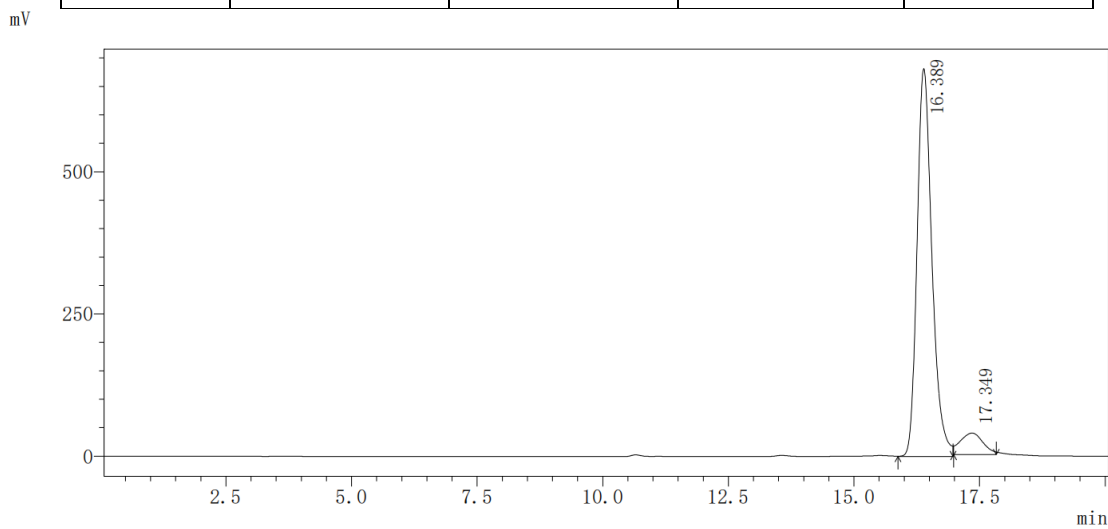
Detector A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	11.039	2011711	138131	97.971
2	12.317	41671	2834	2.029
Total		2053382	140965	



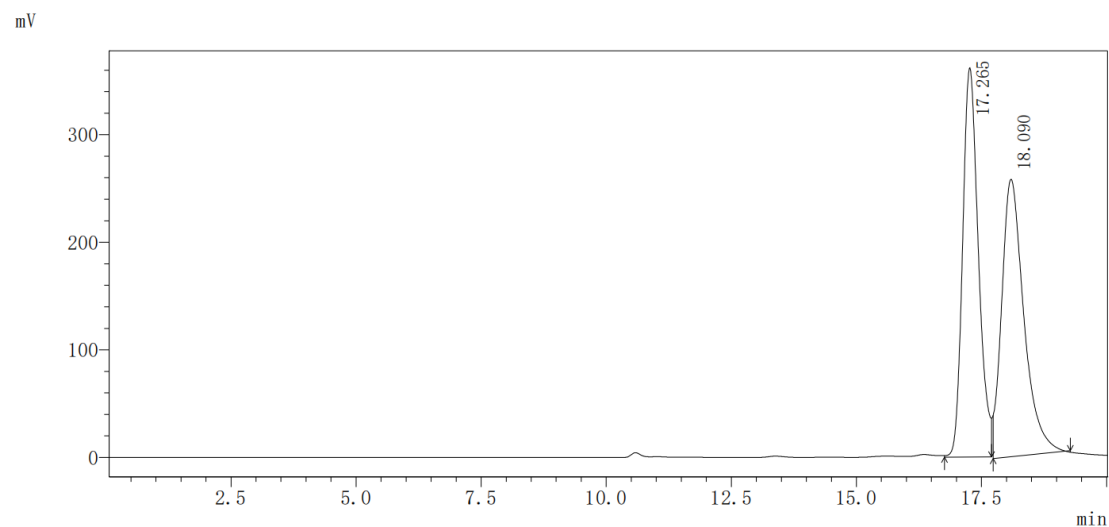
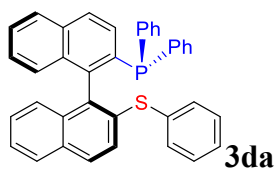
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	16.331	14342519	729557	49.162
2	17.125	14831466	476772	50.838
Total		29173985	1206329	



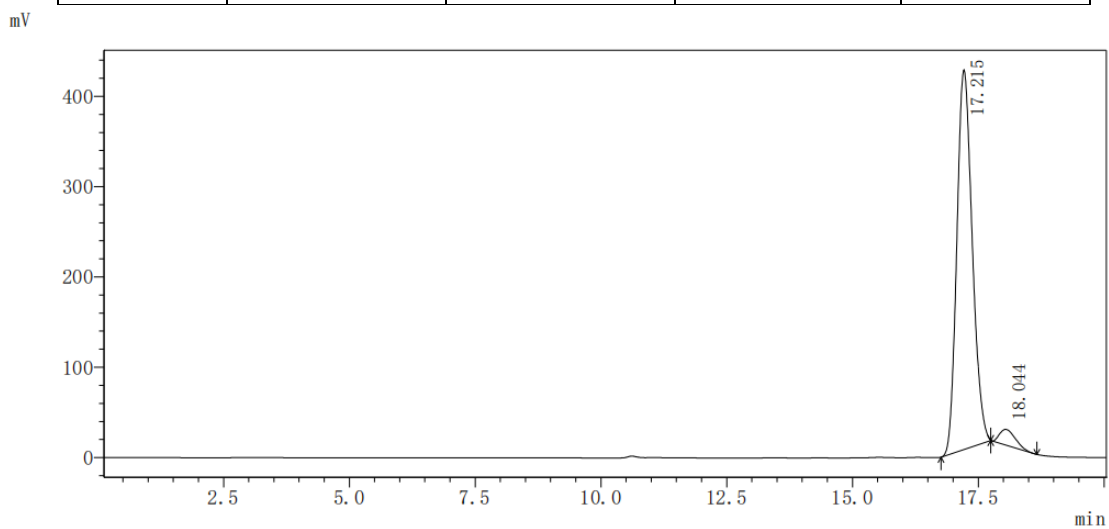
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	16.389	14076368	682075	92.249
2	17.349	1182727	37754	7.751
Total		15259095	719830	



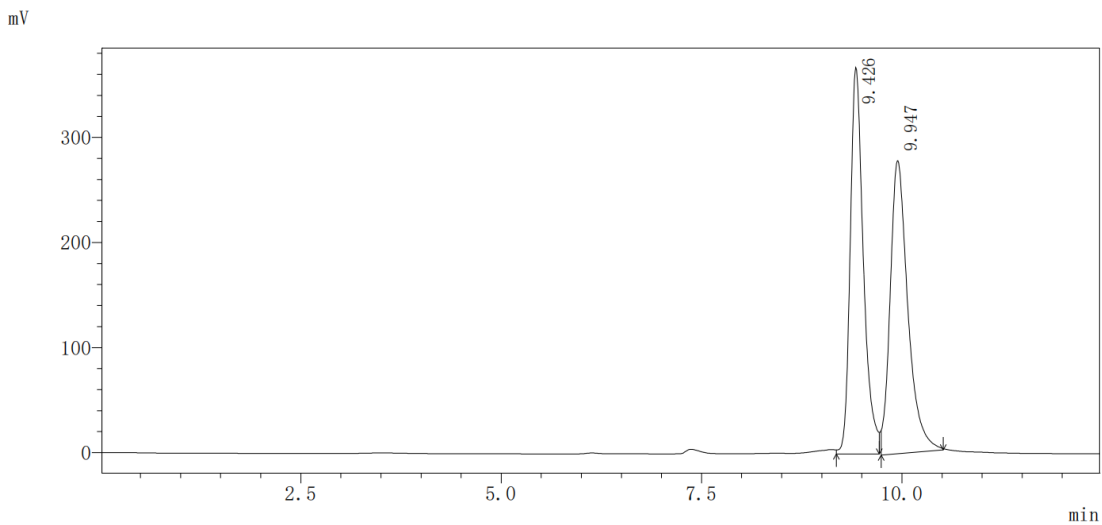
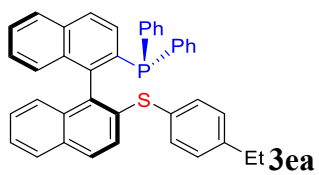
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	17.265	7907065	361841	49.923
2	18.090	7931431	257962	50.077
Total		15838496	619803	



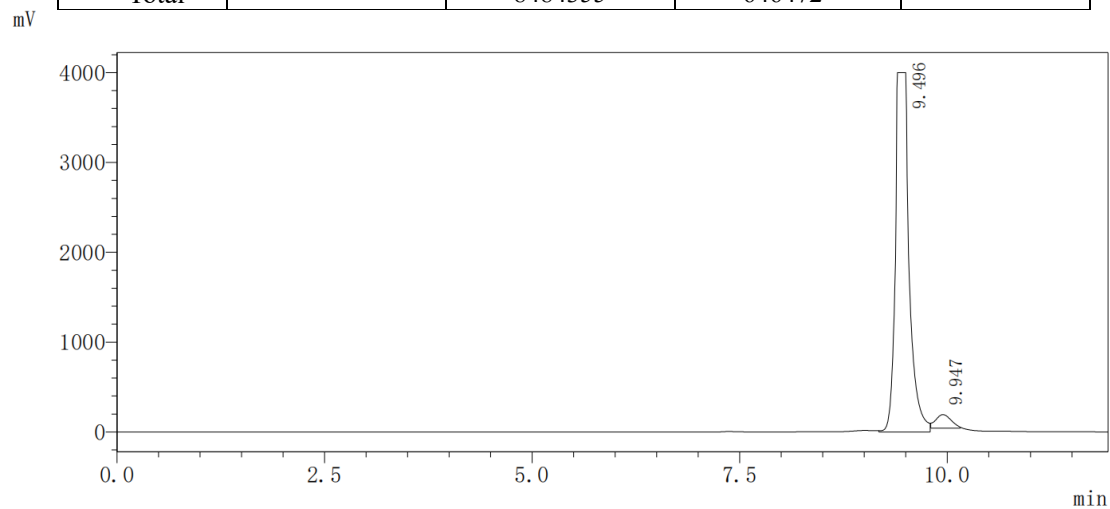
Detector A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	17.215	8945100	420426	95.971
2	18.044	375524	17237	4.029
Total		9320624	437663	



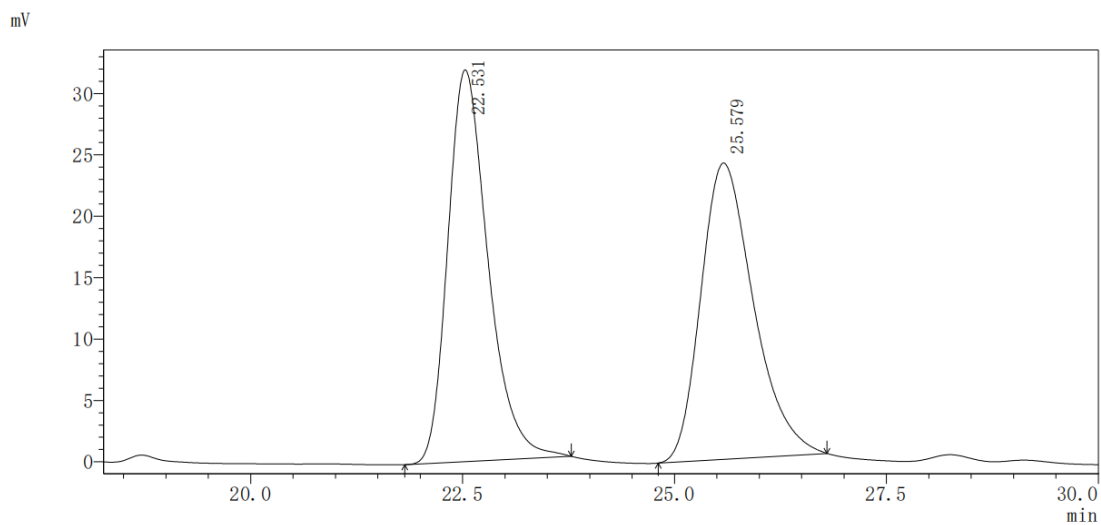
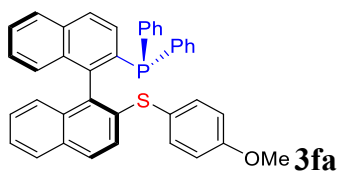
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	9.426	4205311	367695	49.565
2	9.947	4279044	278777	50.435
Total		8484355	646472	



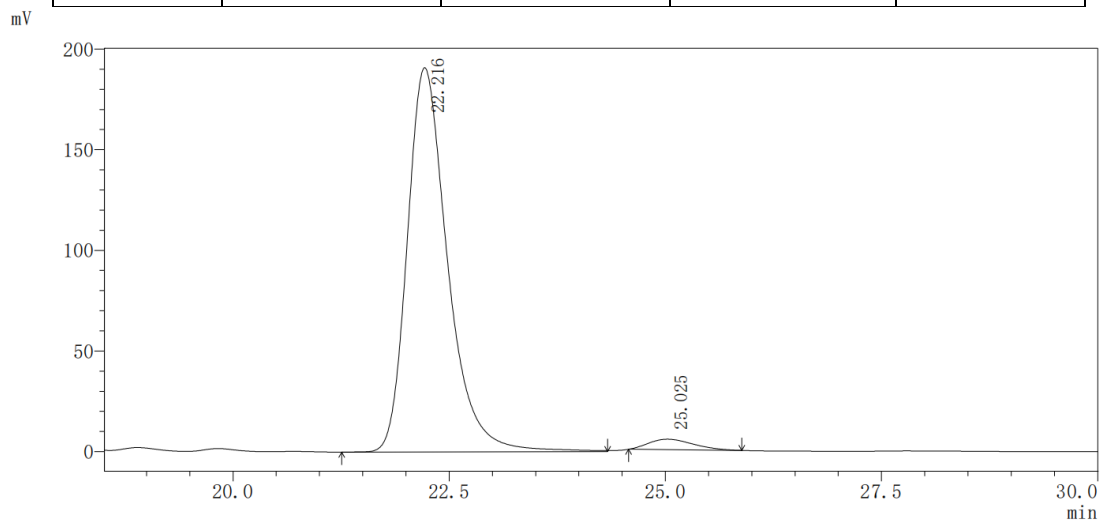
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	9.496	46577763	3999403	95.962
2	9.947	1959851	151525	4.038
Total		48537614	4150928	



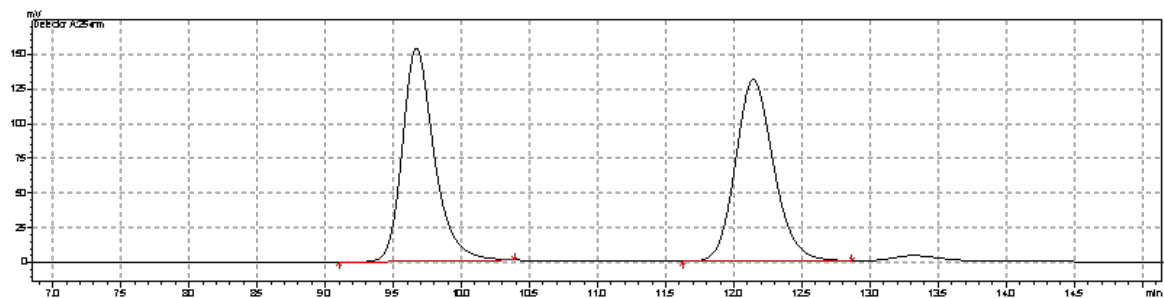
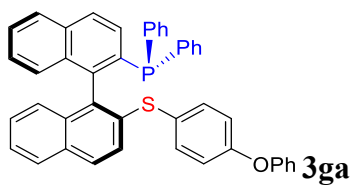
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	22.531	1057075	31917	50.350
2	25.579	1042363	24126	49.650
Total		2099438	56042	



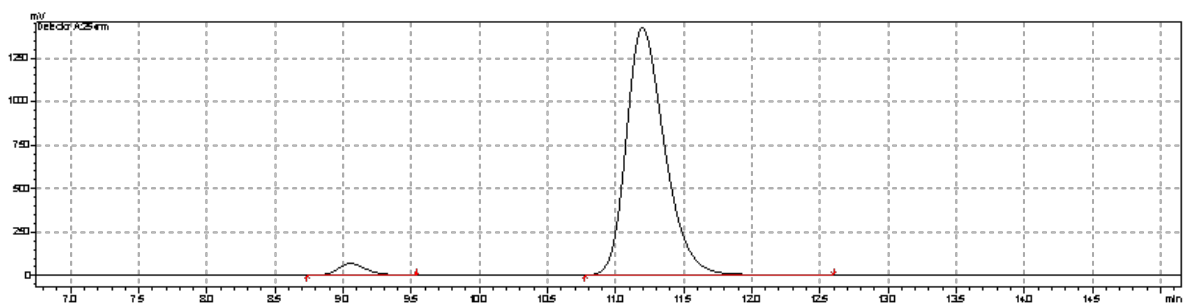
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	22.216	6208063	190972	97.074
2	25.025	187139	5205	2.926
Total		6395202	196177	



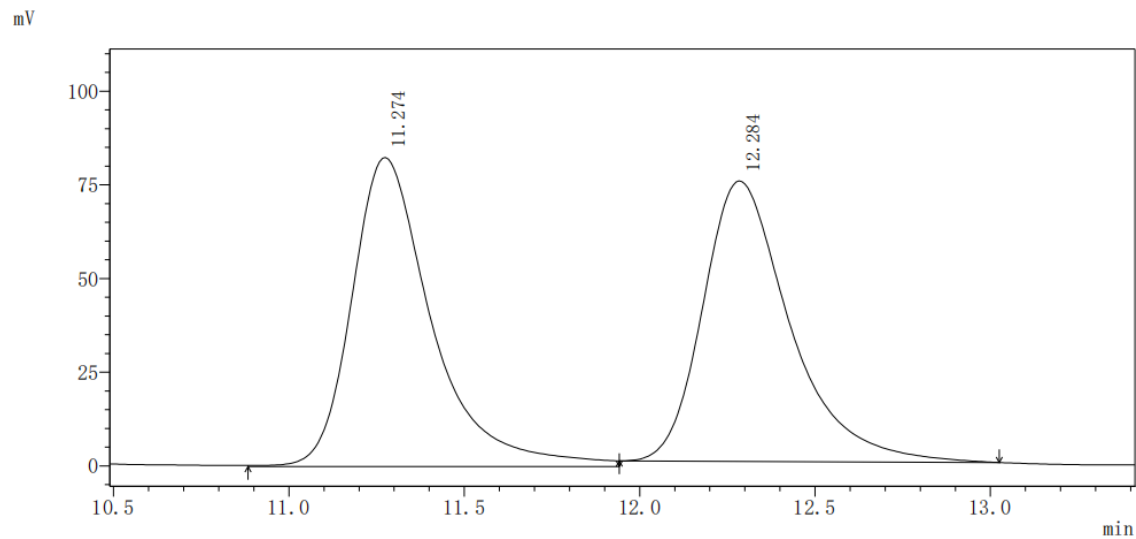
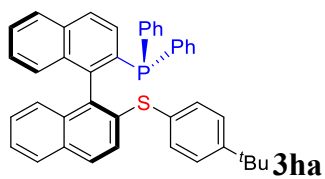
Detector A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	9.664	2493696	153561	49.445
2	12.138	2549694	130844	50.555
Total		5043390	284405	



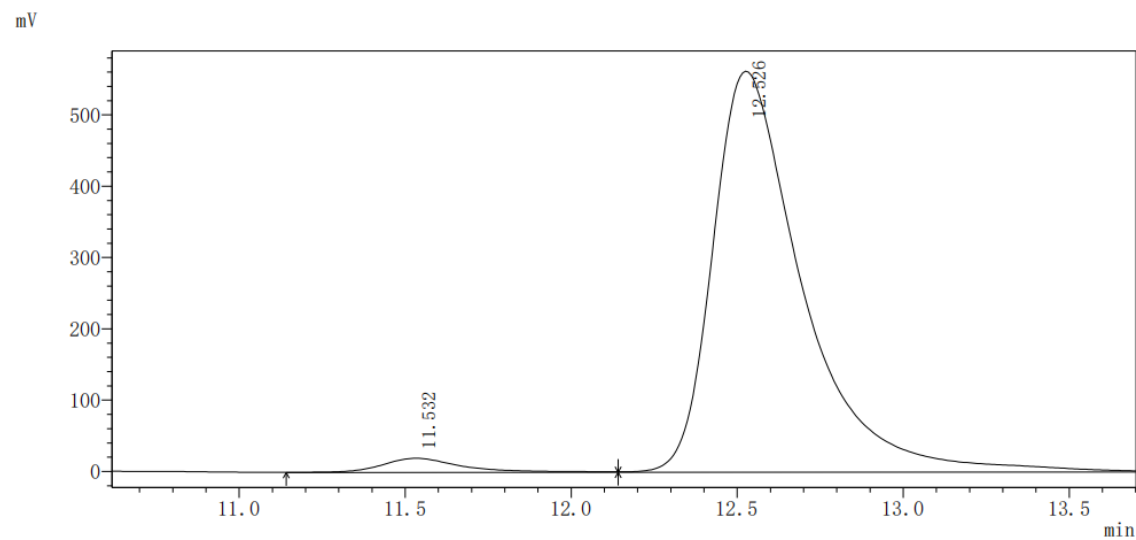
Detector A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	9.050	1027705	68490	3.573
2	11.193	27737484	1422924	96.427
Total		28765189	1491415	



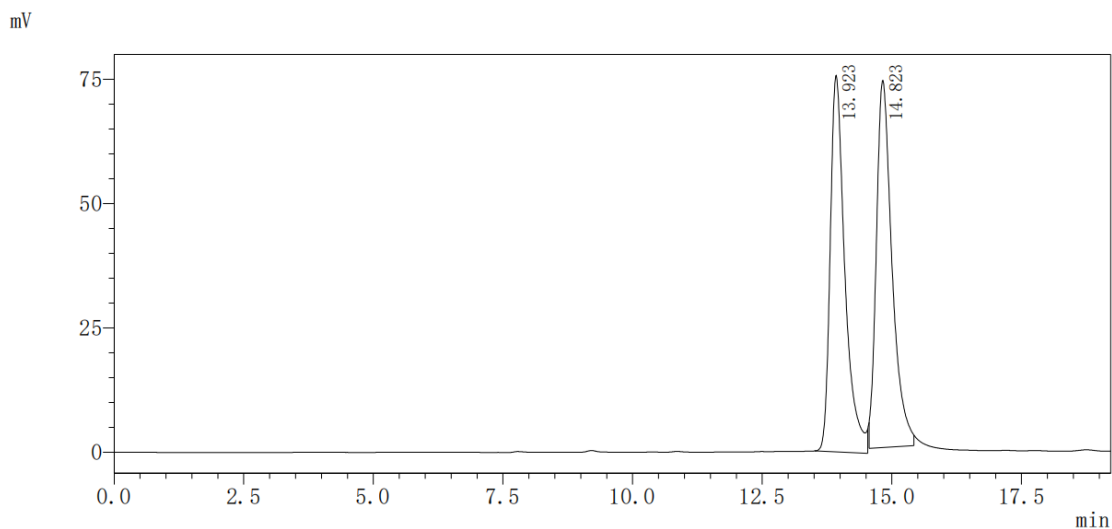
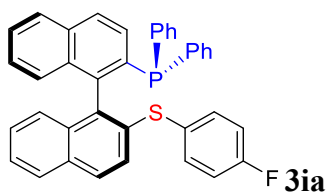
Detector A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	11.274	1314358	82520	50.384
2	12.284	1294302	74877	49.616
Total		2608659	157397	



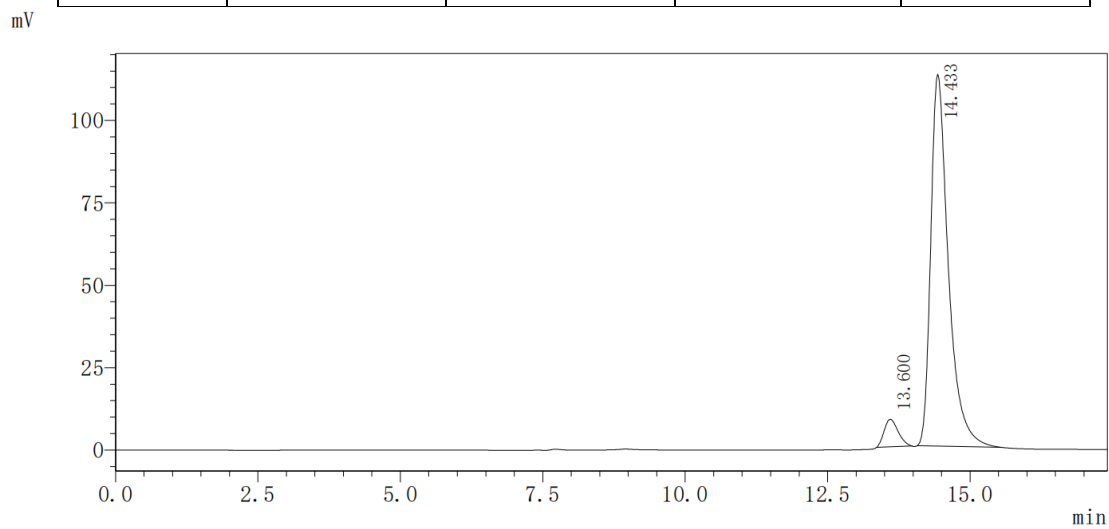
Detector A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	11.532	345067	19950	3.143
2	12.526	10634208	562637	96.857
Total		10979274	582588	



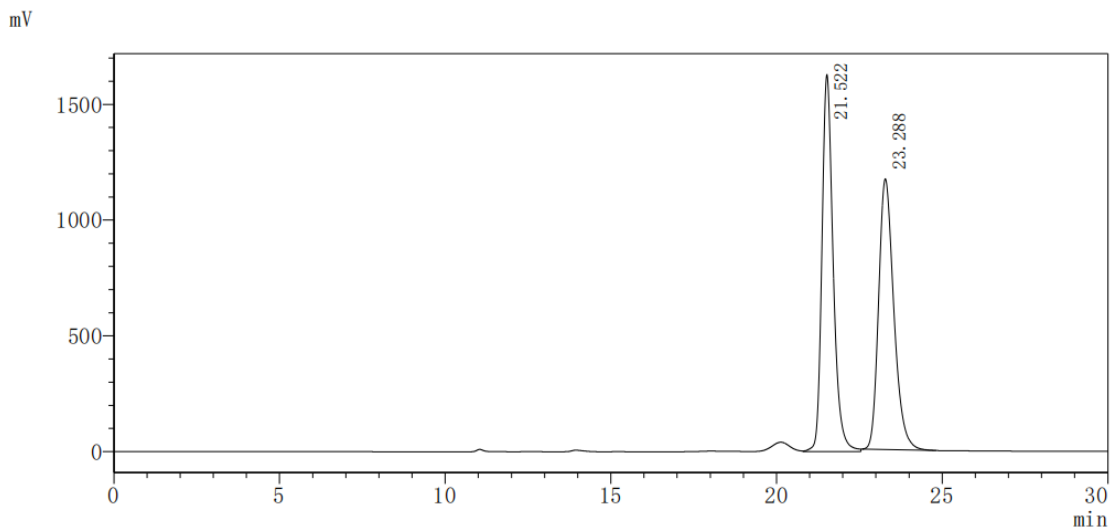
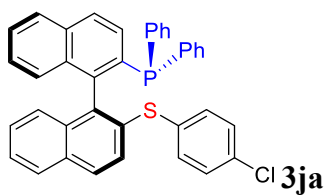
Detector A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	13.923	1469358	75533	46.993
2	14.823	1657433	74524	53.007
Total		3126791	150057	



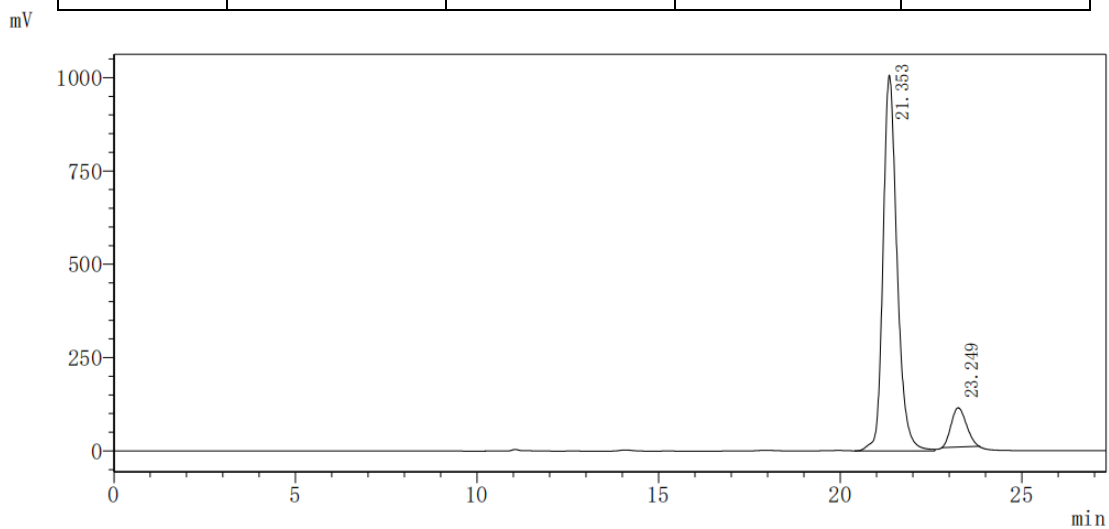
Detector A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	13.600	132050	8064	5.133
2	14.433	2440535	112942	94.867
Total		2572585	121006	



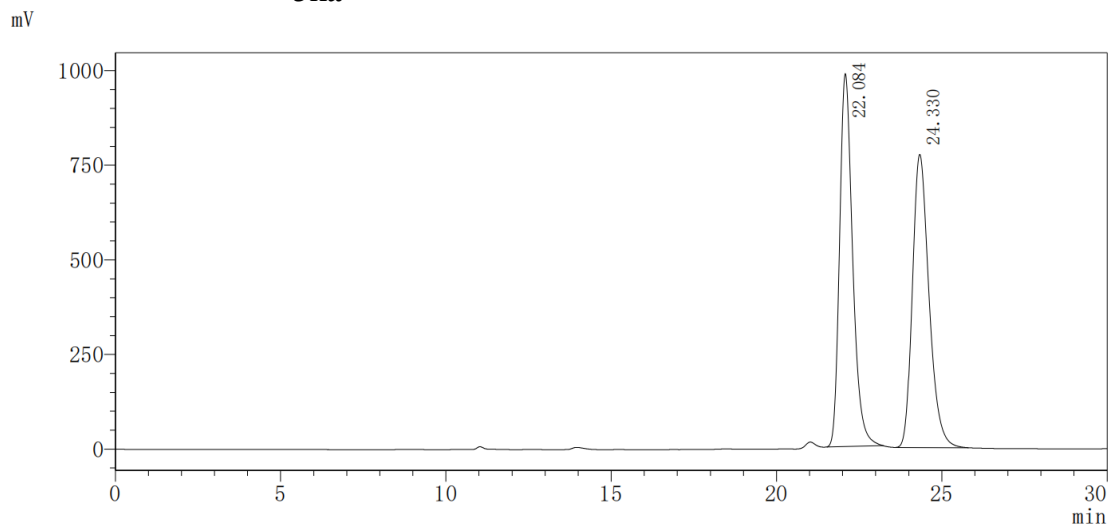
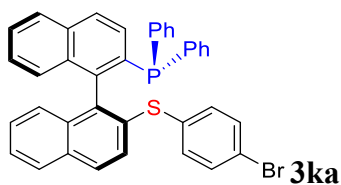
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	21.522	38483085	1628009	50.739
2	23.288	37361593	1168771	49.261
Total		75844678	2796780	



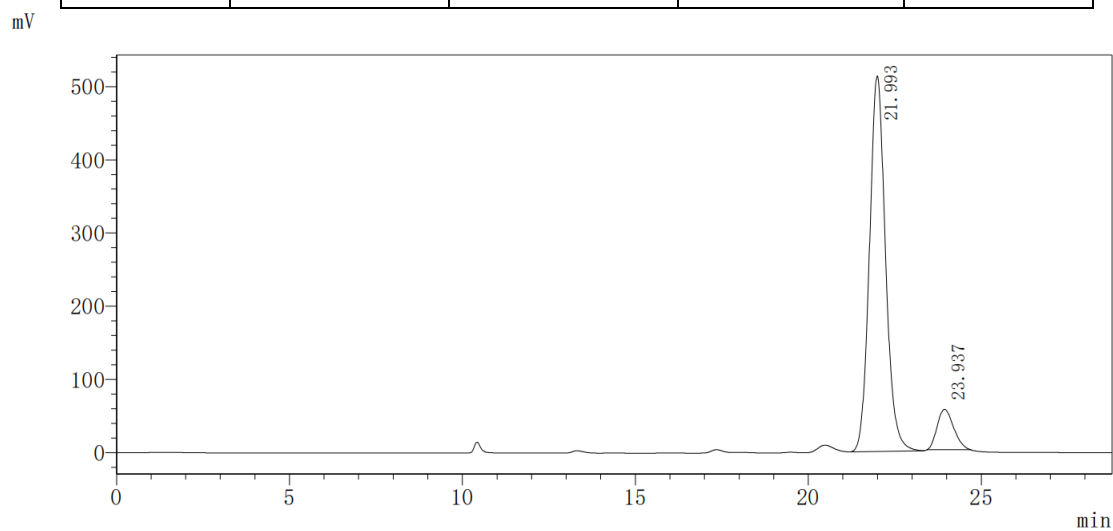
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	21.820	2093391	63840	90.780
2	23.944	212611	6711	9.220
Total		2306001	70551	



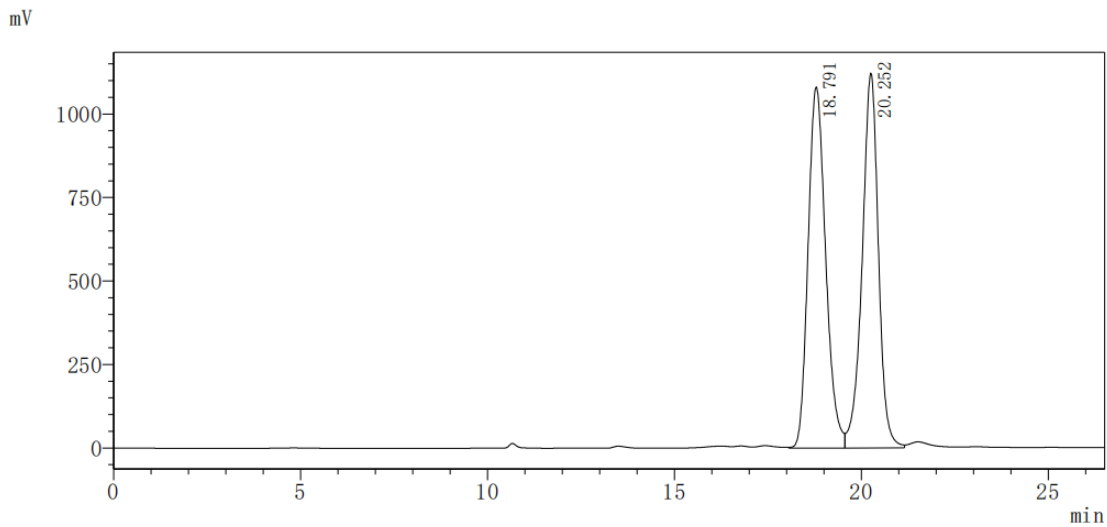
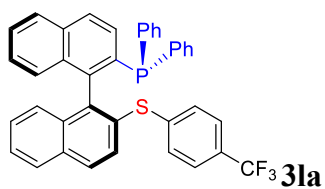
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	22.084	28079549	992408	50.293
2	24.330	27737294	779191	49.680
Total		55832224	1772027	



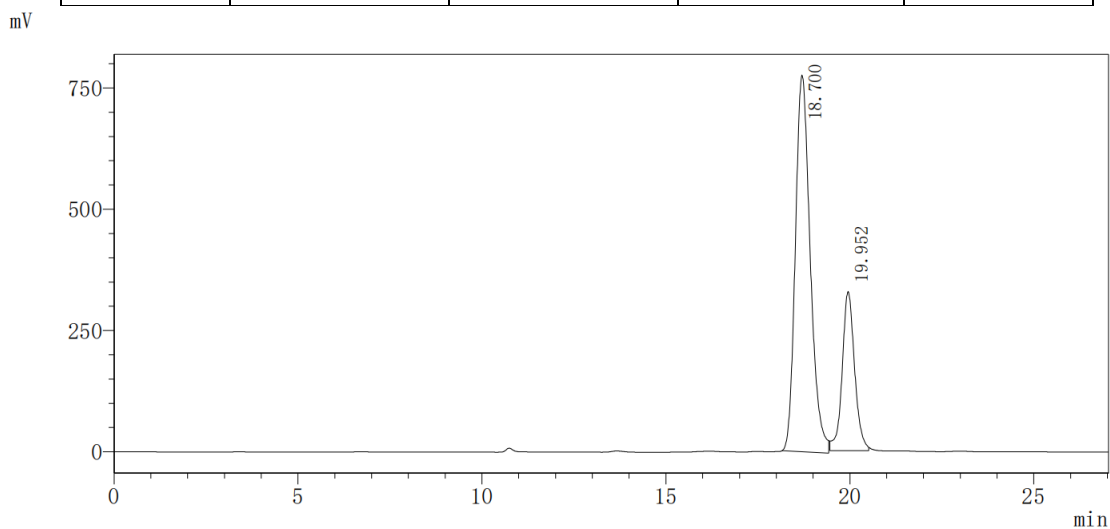
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	21.993	16580572	512733	90.040
2	23.937	1834118	55265	9.960
Total		18414690	567998	



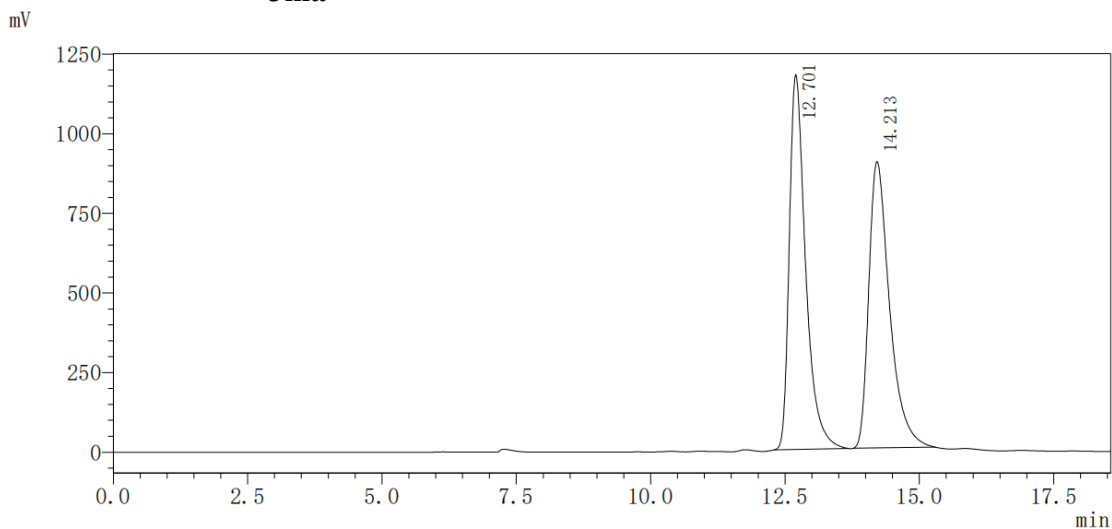
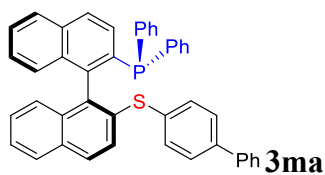
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	18.791	35361704	1081141	51.089
2	20.252	33853805	1121921	48.911
Total		69215509	2203063	



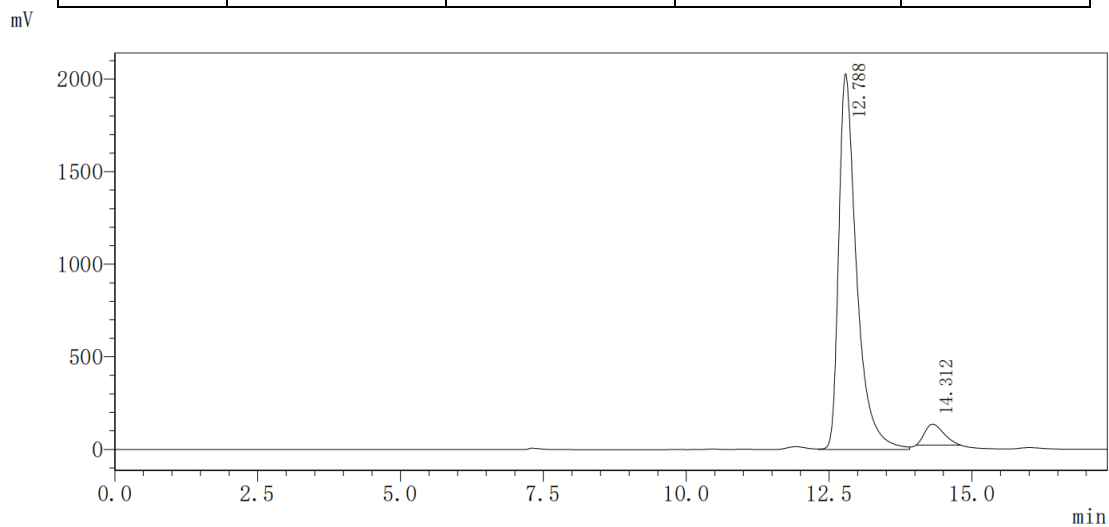
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	18.700	21920315	776657	73.694
2	19.952	7824858	330649	26.306
Total		29745172	1107307	



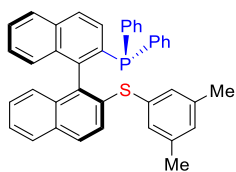
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	12.701	24807405	1176326	50.618
2	14.213	24201275	899144	49.382
Total		49008680	2075470	

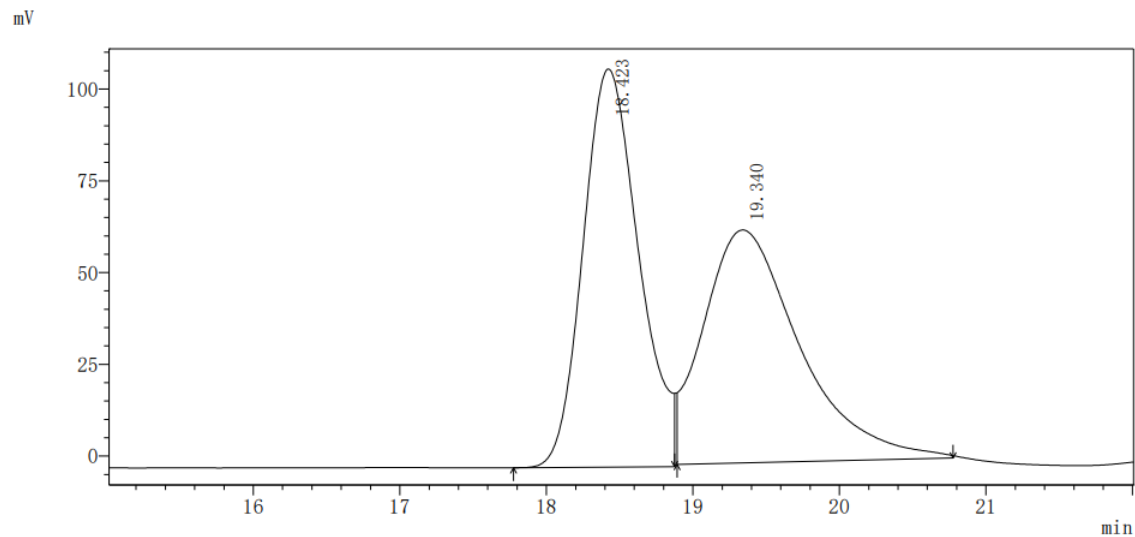


Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	12.788	44222438	2028831	94.439
2	14.312	2603904	112887	5.561
Total		46826342	2141717	

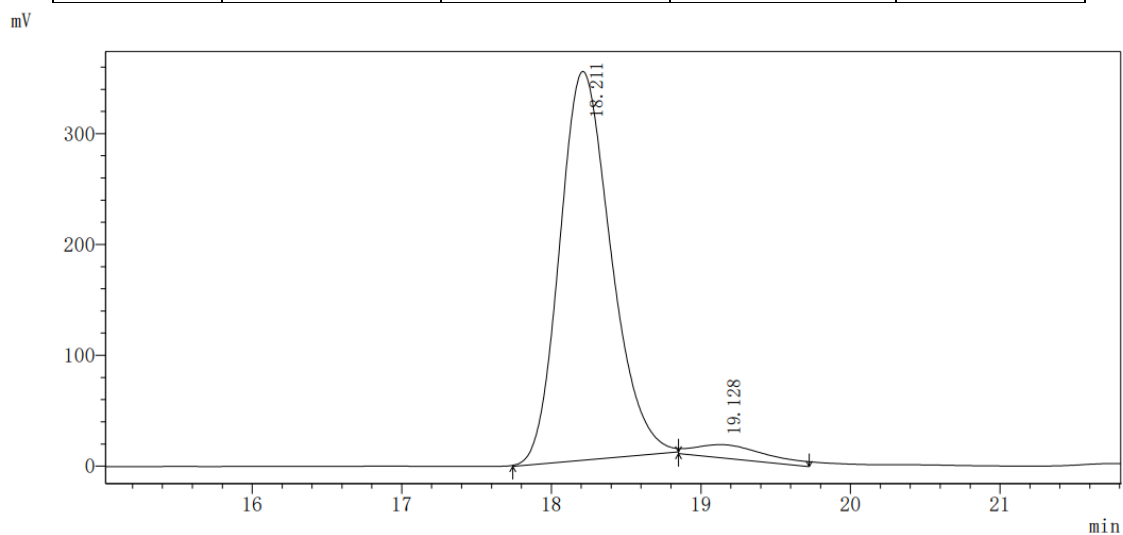


3na



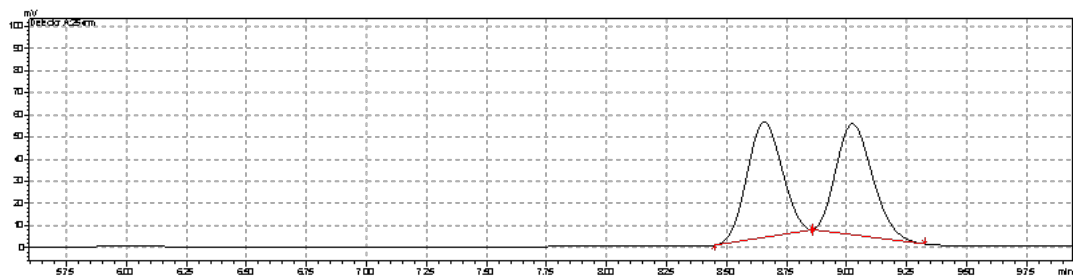
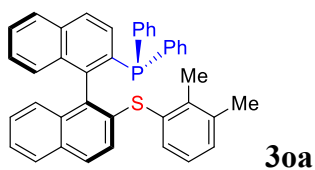
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	18.423	2867401	108487	49.705
2	19.340	2901420	63469	50.295
Total		5768821	171956	



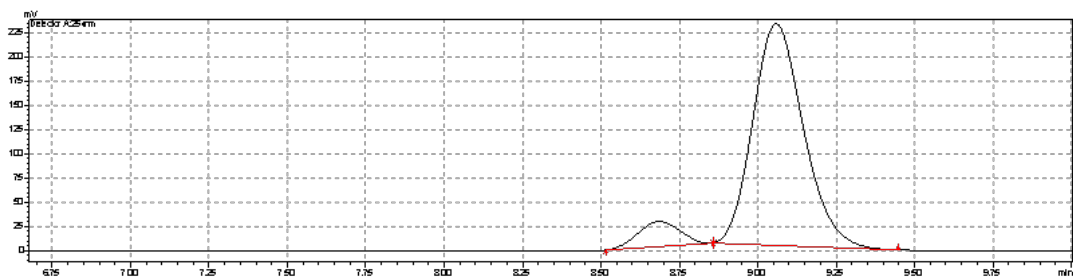
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	18.211	8823734	356128	94.593
2	19.128	504374	17109	5.407
Total		9328108	373237	



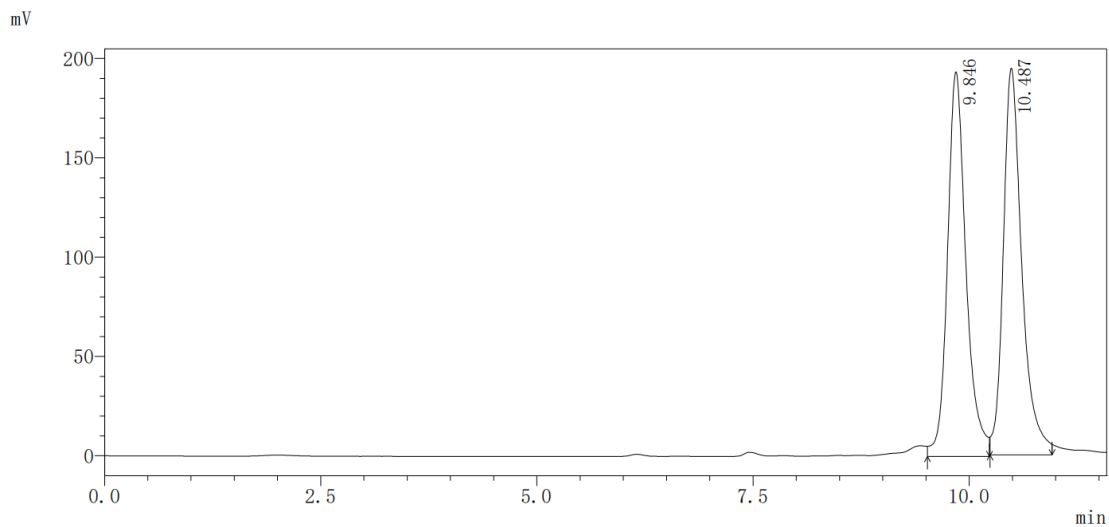
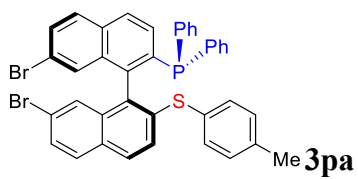
Detector A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	8.652	526852	52115	49.960
2	9.019	527698	50045	50.040
Total		1054549	102160	



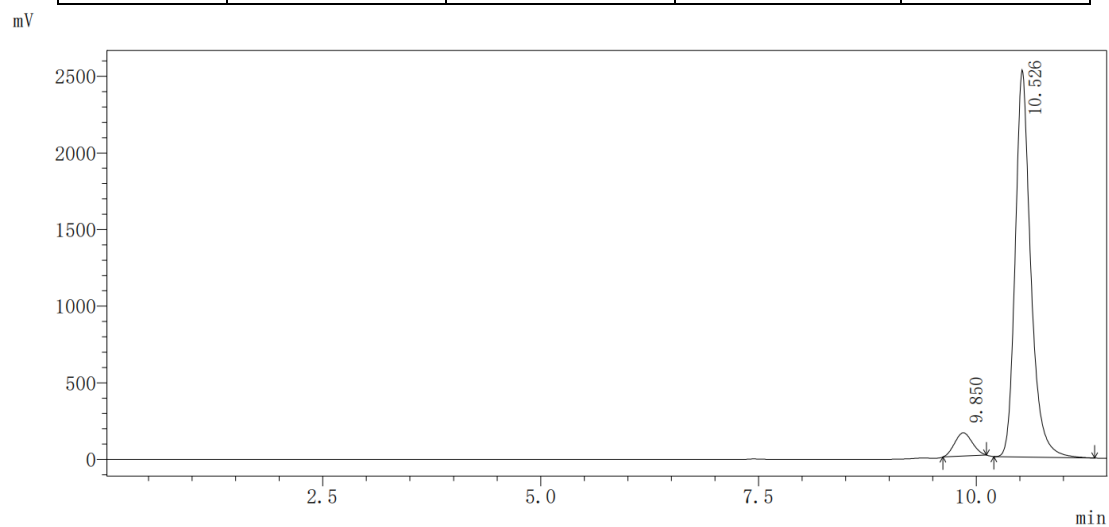
Detector A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	8.648	61465	6520	8.192
2	9.029	688828	58566	91.808
Total		750293	65085	



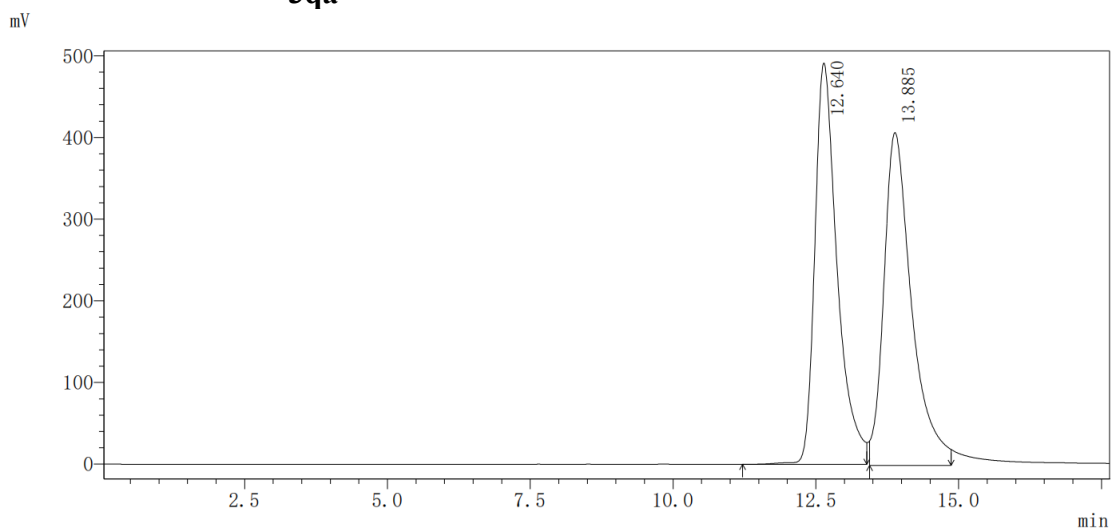
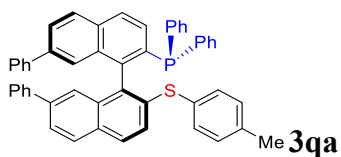
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	9.846	2882126	193541	49.799
2	10.487	2905338	194566	50.201
Total		5787464	388107	



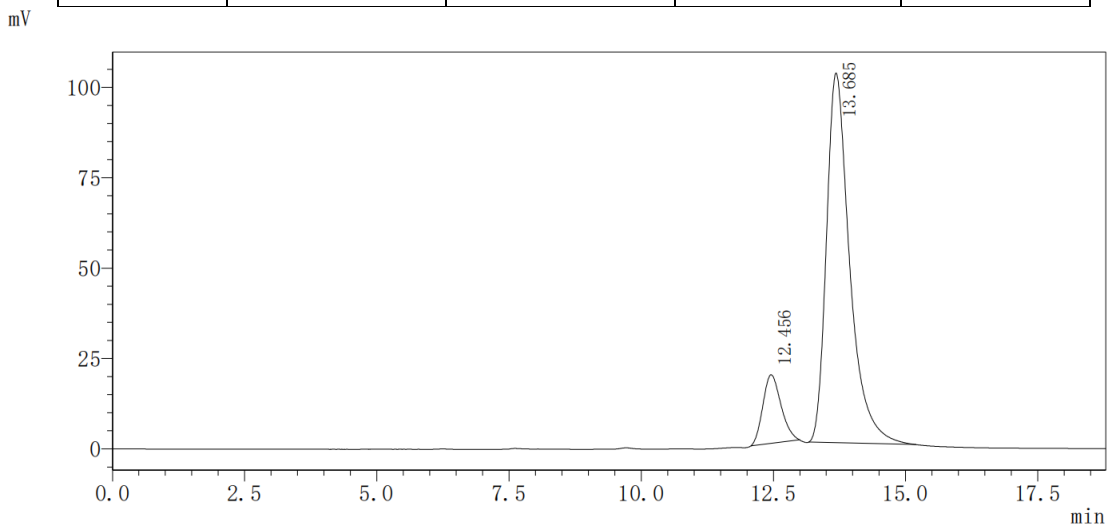
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	9.850	2148147	152642	6.413
2	10.526	31350153	2525115	93.587
Total		33498299	2677757	



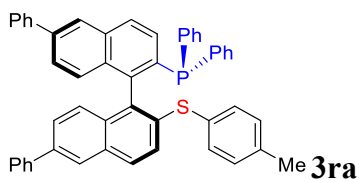
Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	12.640	13384161	491437	49.267
2	13.885	13782649	407551	50.733
Total		27166810	898988	

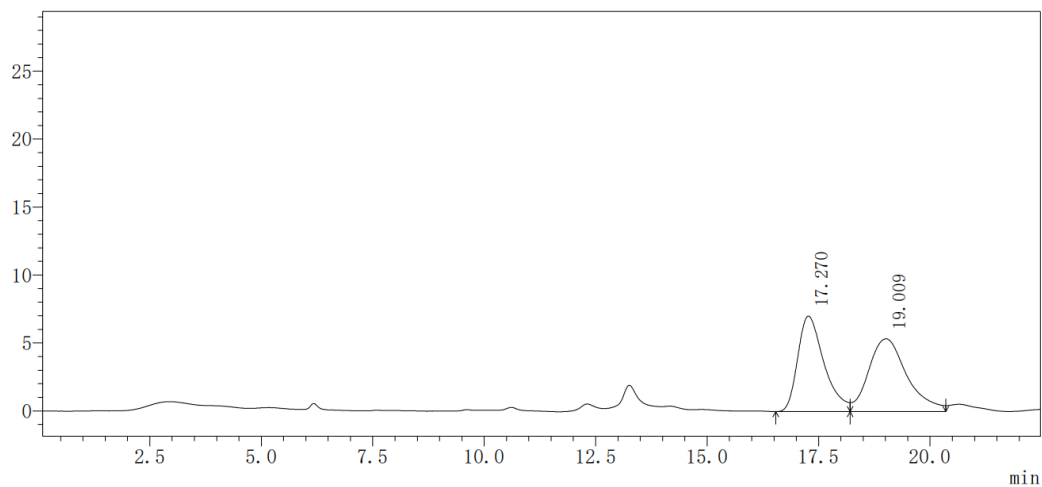


Detector A channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	12.456	452906	18980	12.638
2	13.685	3130802	102248	87.362
Total		3583708	121228	



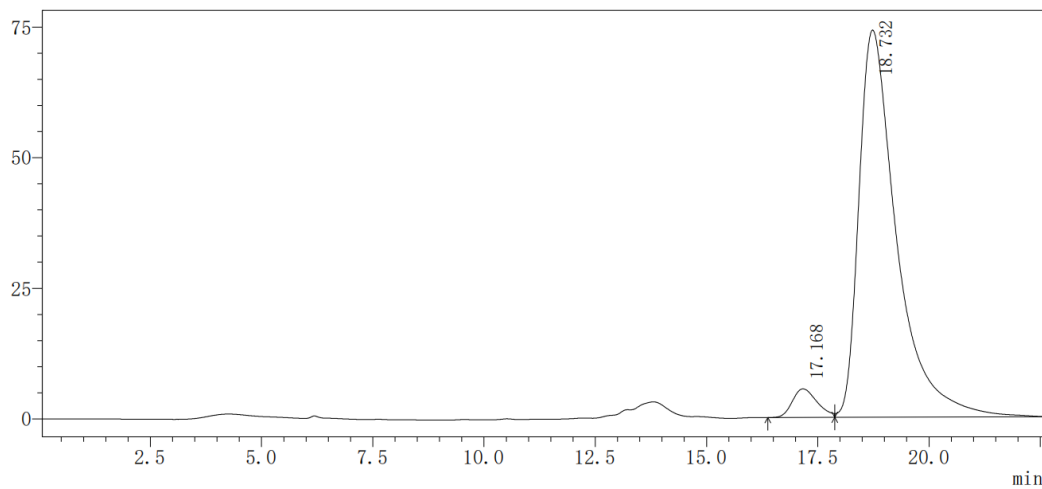
mV



Detector A channel 254 nm

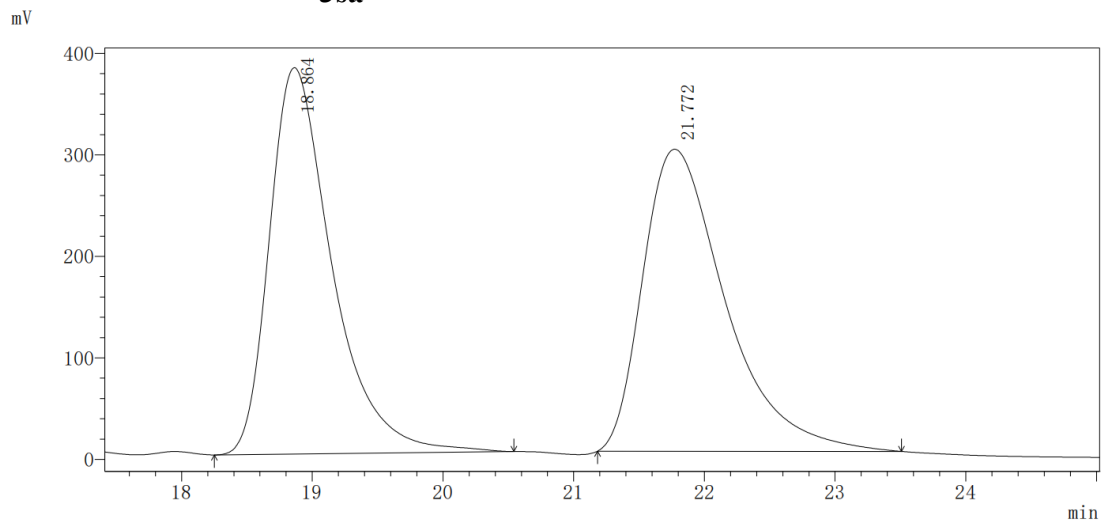
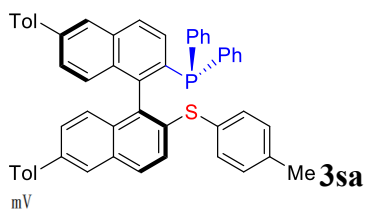
Peak	Ret. Time	Area	Height	Conc.
1	17.270	289144	7041	49.573
2	19.009	294130	5268	50.427
Total		583274	12308	

mV



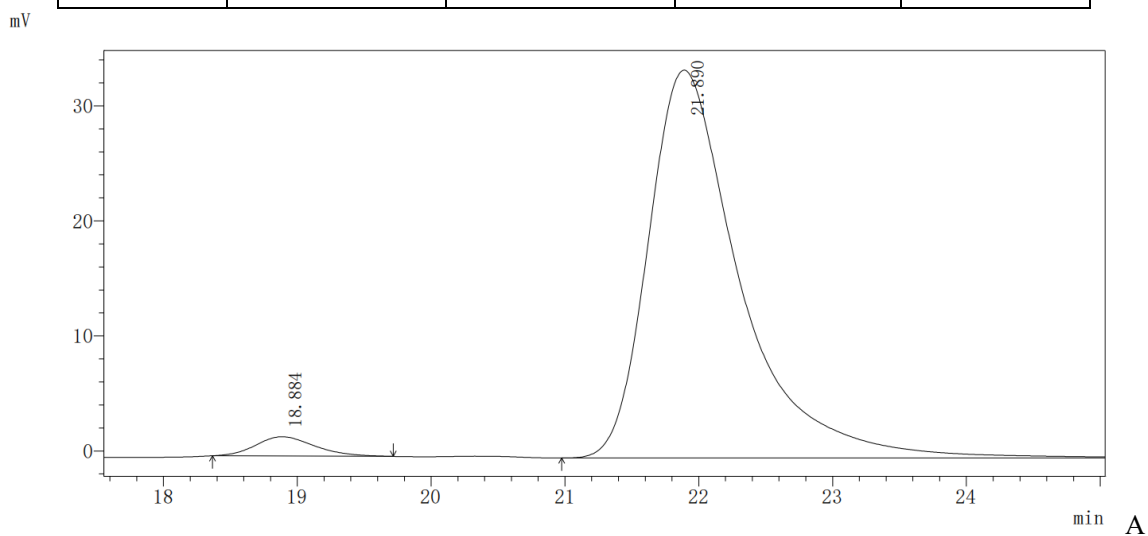
channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	17.168	212565	5489	4.630
2	18.732	4378801	74107	95.370
Total		4591366	79596	



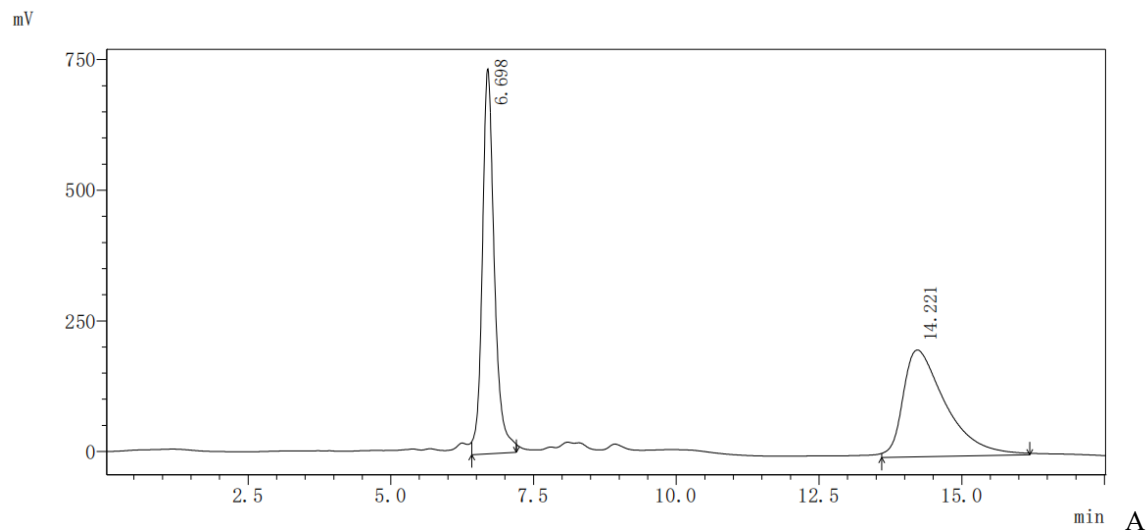
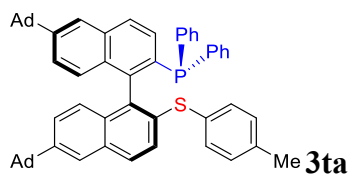
A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	18.864	12682222	380682	50.015
2	21.772	12674691	294658	49.985
Total		25356913	675340	



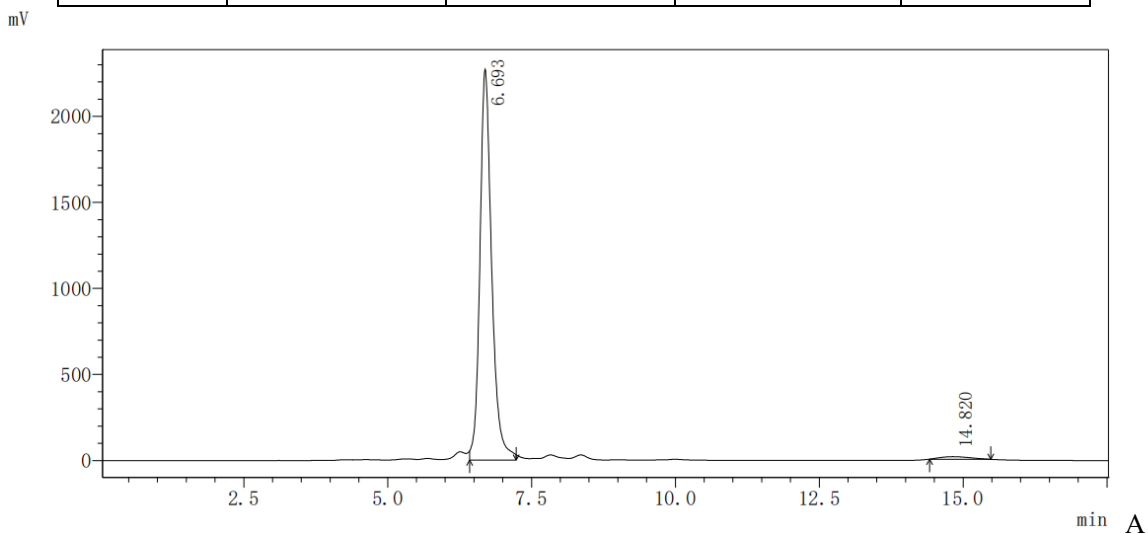
channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	18.884	51219	1673	3.081
2	21.890	1611103	33723	96.919
Total		1662322	35397	



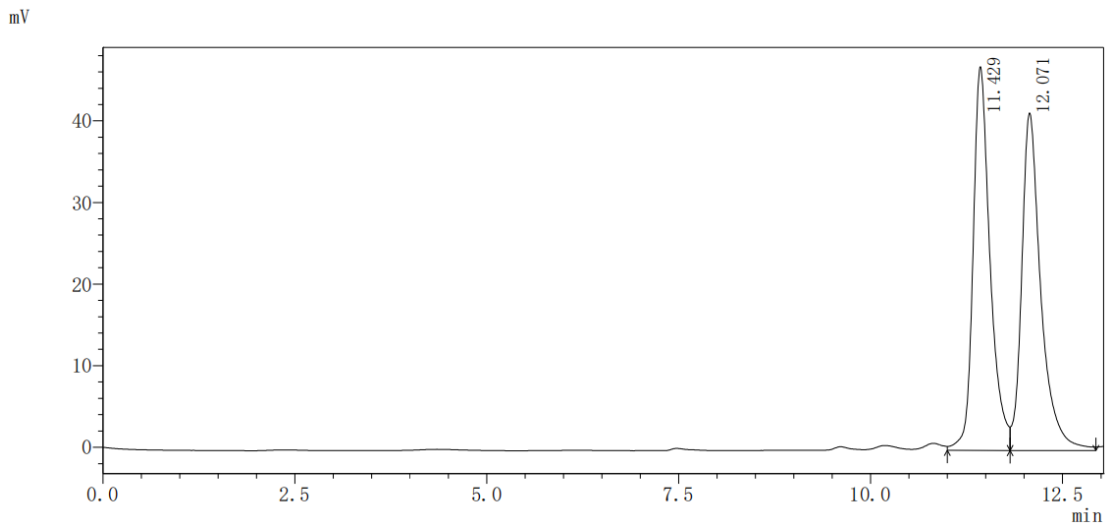
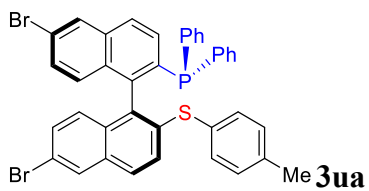
channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	6.698	10812421	736933	50.325
2	14.221	10672856	204493	49.675
Total		21485277	941426	



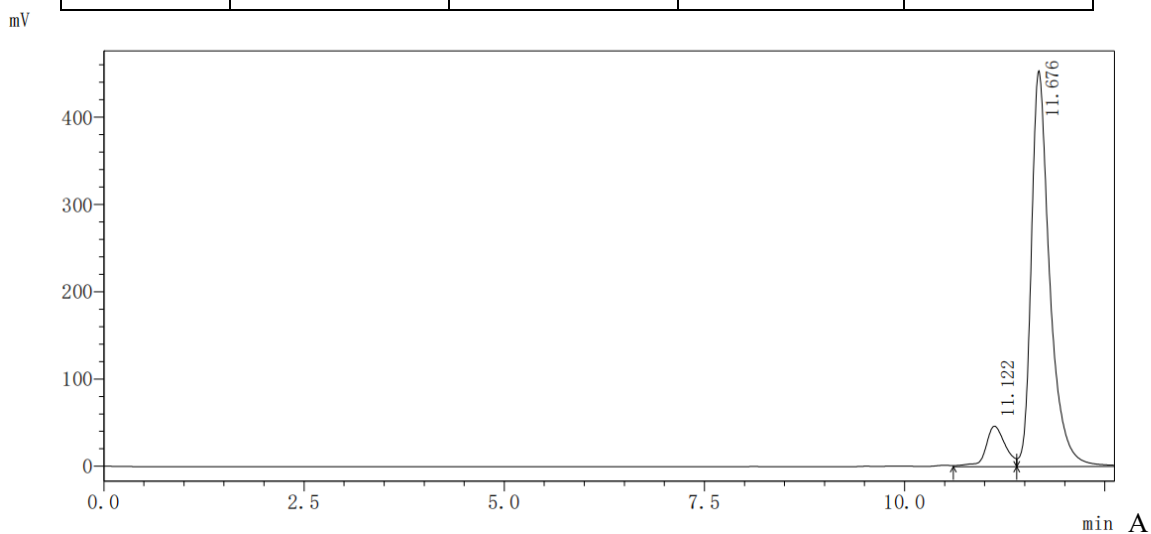
channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	6.693	32227823	2270379	98.341
2	14.820	543519	14943	1.659
Total		32771342	2285323	



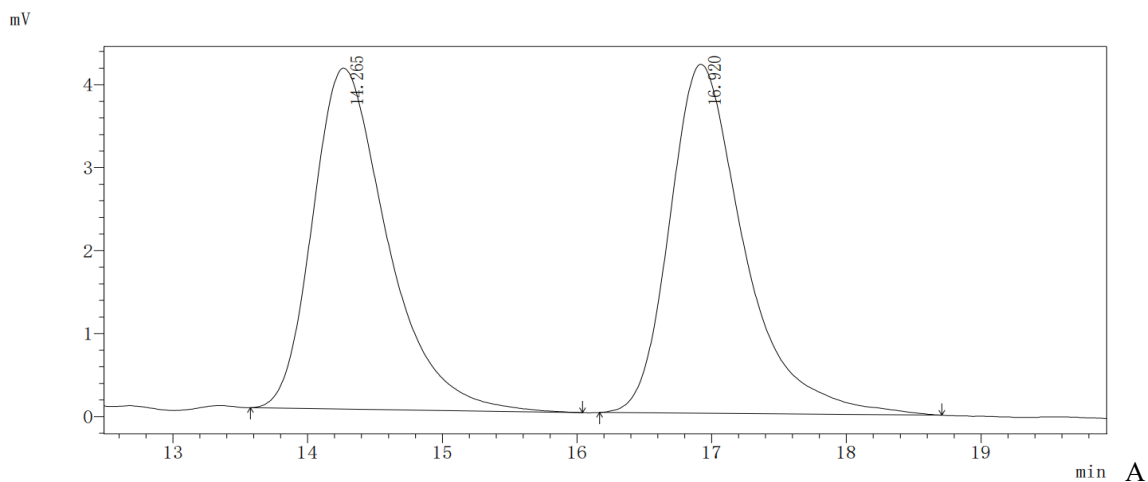
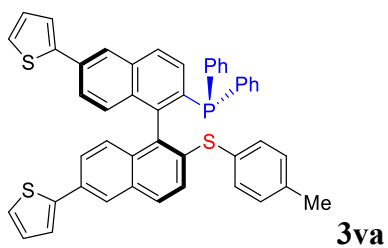
A channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	11.429	727190	46993	50.245
2	12.071	720101	41355	49.755
Total		1447292	88348	



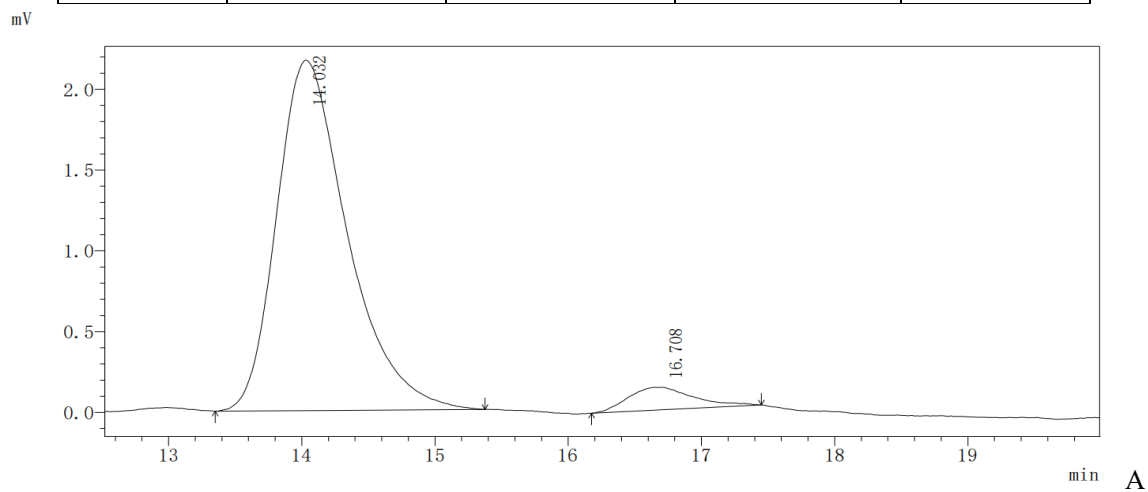
channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	11.122	783729	46371	9.746
2	11.676	7257643	453235	90.254
Total		8041372	499607	



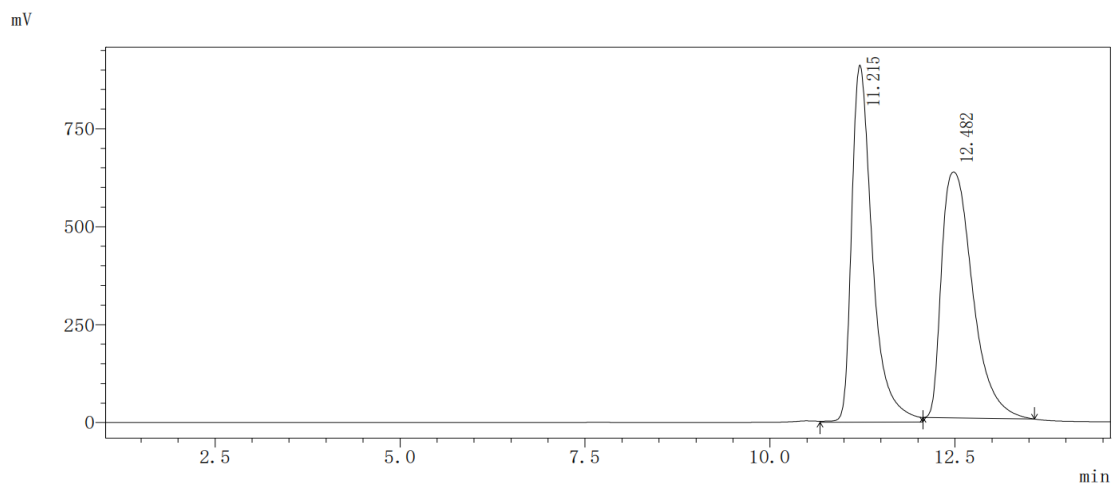
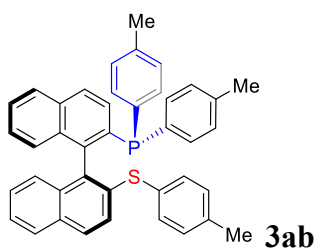
channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	14.265	158720	4110	49.543
2	16.920	161650	4206	50.457
Total		320370	8316	



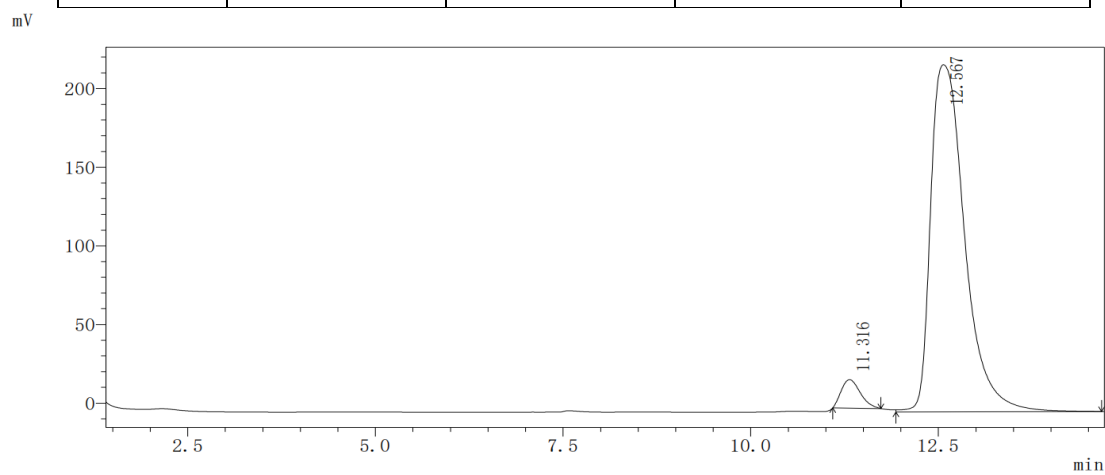
channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	14.032	80235	2170	94.403
2	16.708	4757	139	5.597
Total		84992	2308	



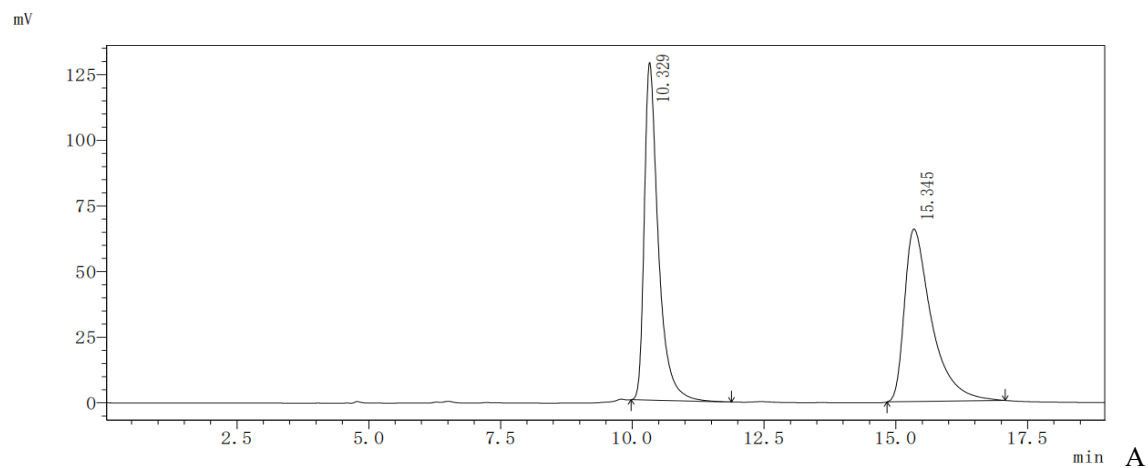
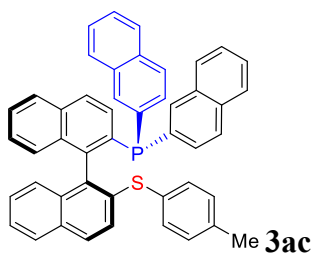
channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	11.215	17870355	911391	49.294
2	12.482	18381924	627899	50.706
Total		36252278	1539289	



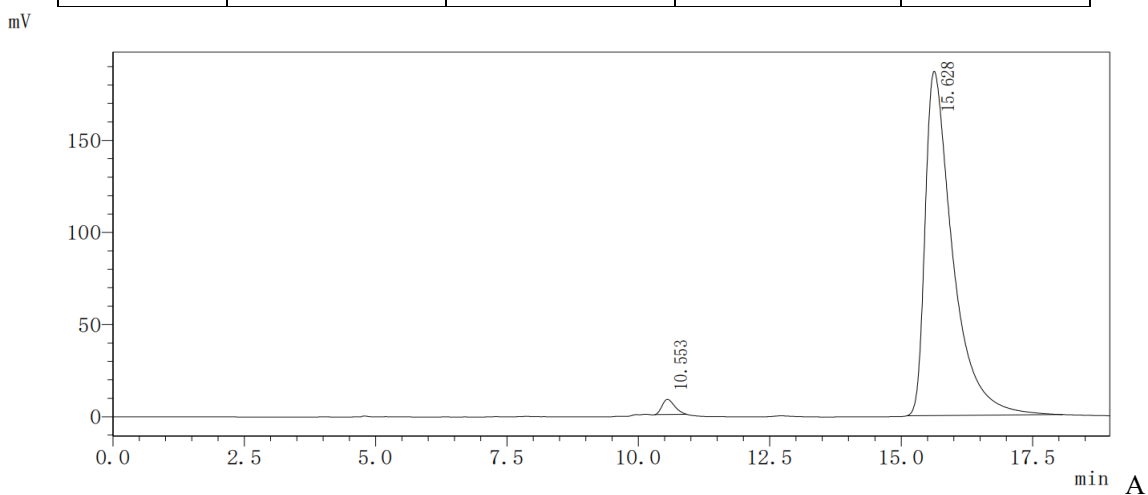
channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	11.316	318894	18051	4.374
2	12.567	6970982	220774	95.626
Total		7289876	238826	



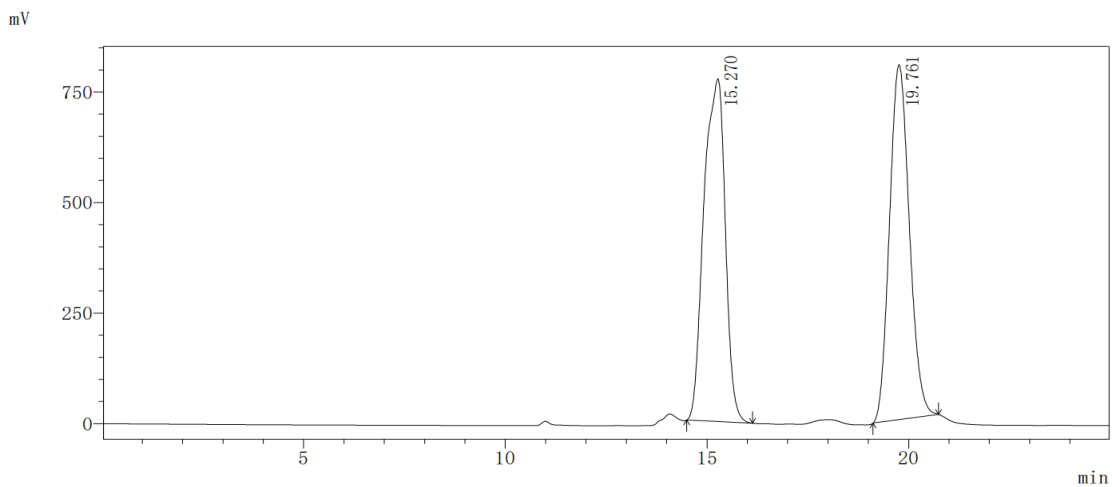
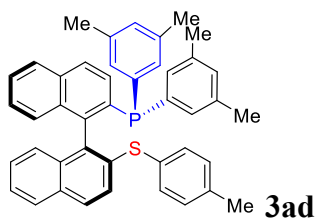
channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	10.329	2346624	128159	50.052
2	15.345	2341720	65648	49.948
Total		4688343	193806	



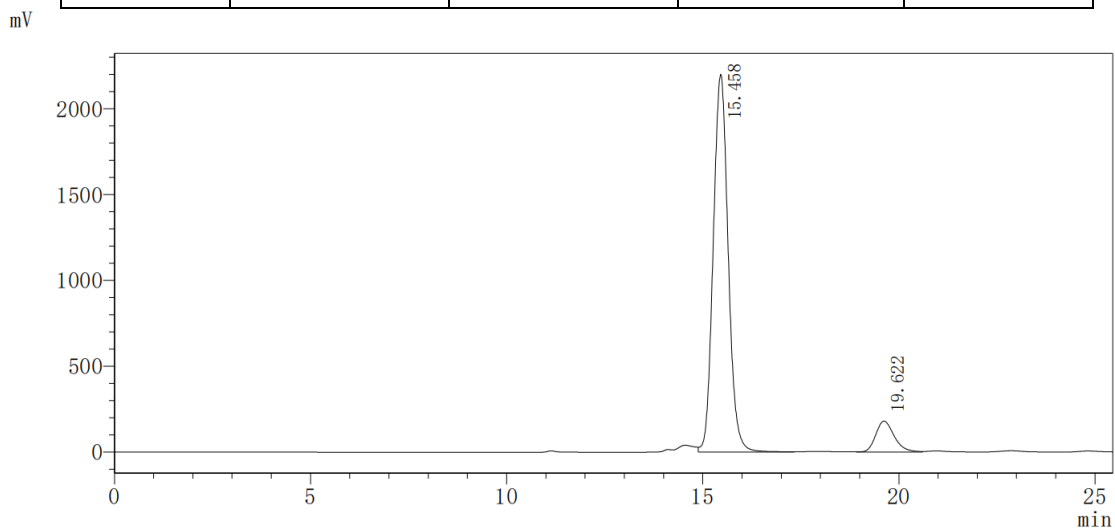
channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	10.553	135889	8193	1.950
2	15.628	6834009	186820	98.050
Total		6969898	195013	



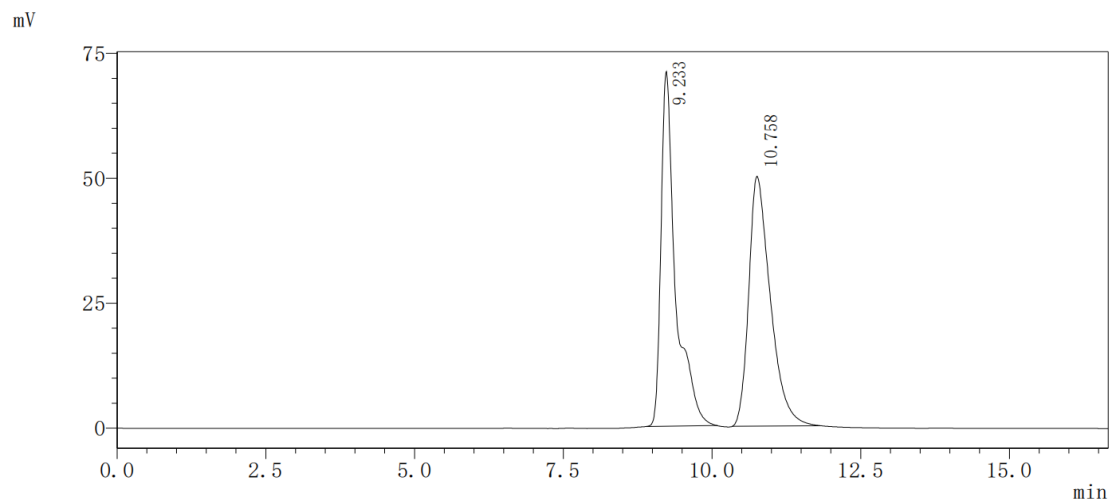
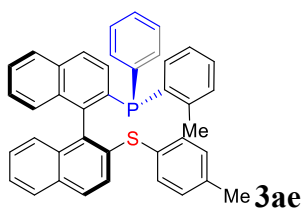
channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	15.270	28460037	774810	50.890
2	19.761	27464875	802525	49.110
Total		55924912	1577335	



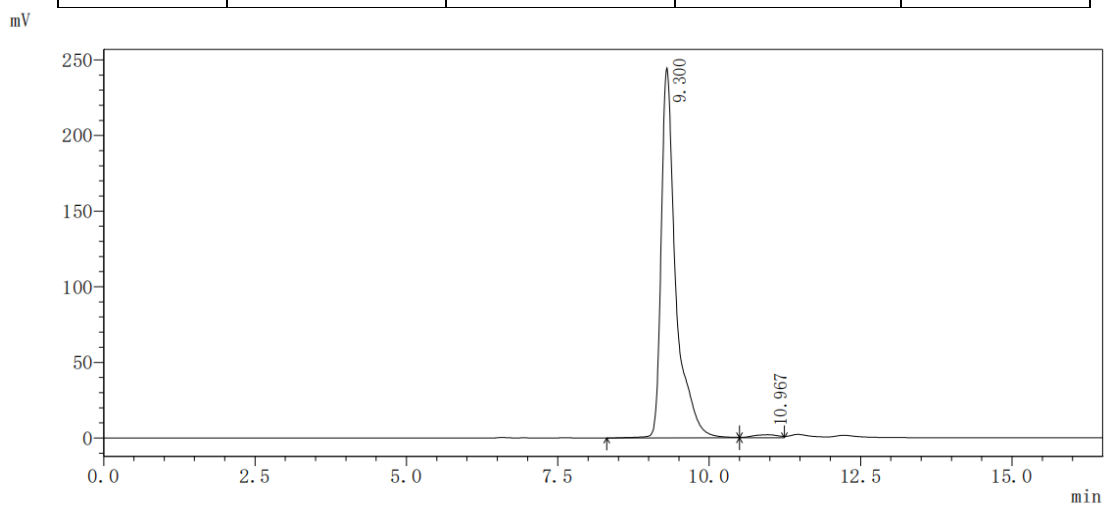
channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	15.458	58360888	2200430	90.716
2	19.622	5972676	180840	9.284
Total		64333564	2381270	



channel 280 nm

Peak	Ret. Time	Area	Height	Conc.
1	9.233	1206934	70941	48.884
2	10.758	1262034	49993	51.116
Total		2468968	120933	

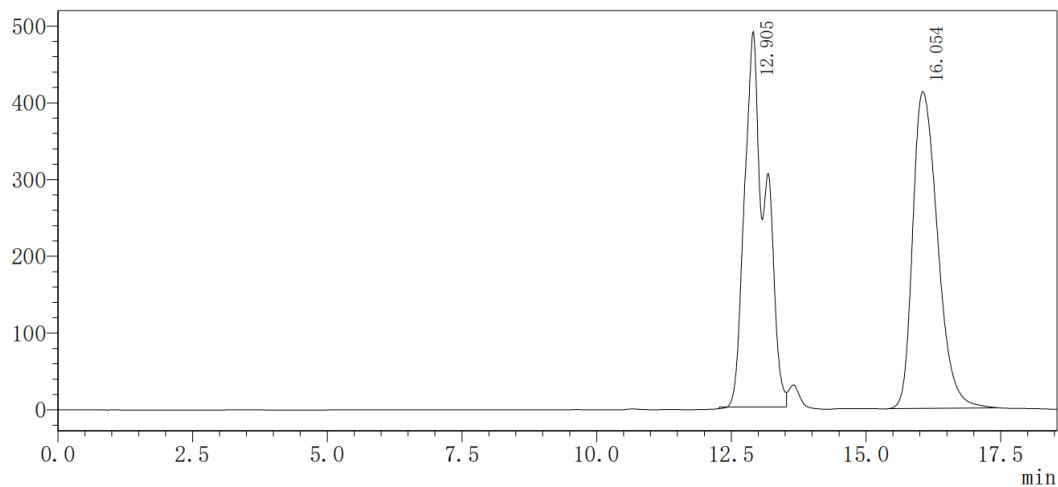


channel 280 nm

Peak	Ret. Time	Area	Height	Conc.
1	9.300	4086665	244530	98.565
2	10.967	59495	2034	1.435
Total		4146160	246564	

After recrystallization

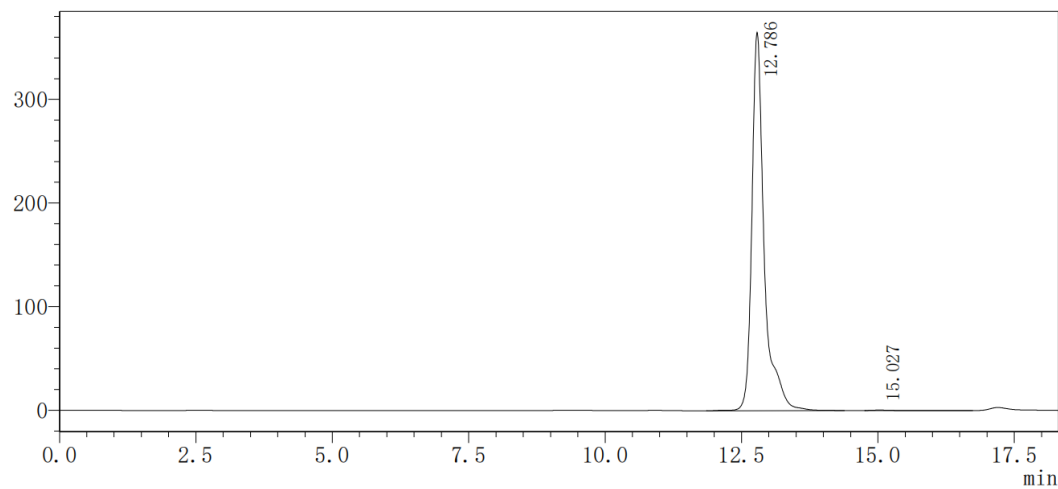
mV



channel 254 nm

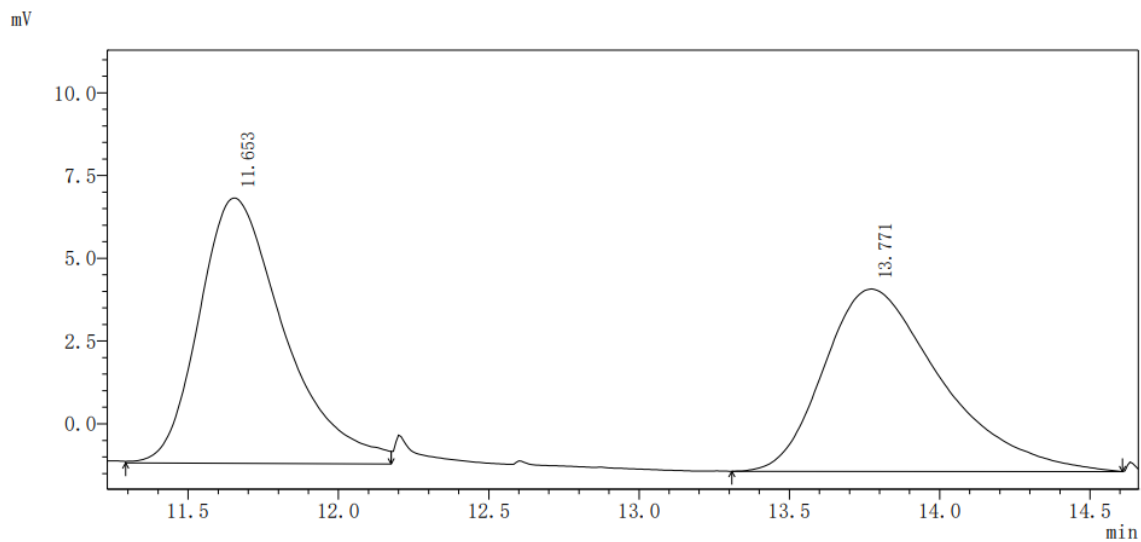
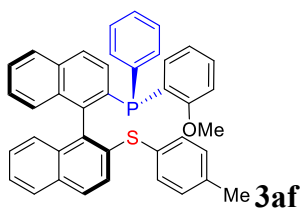
Peak	Ret. Time	Area	Height	Conc.
1	12.905	13336701	488730	50.064
2	16.054	13302745	412692	49.936
Total		26639446	901422	

mV



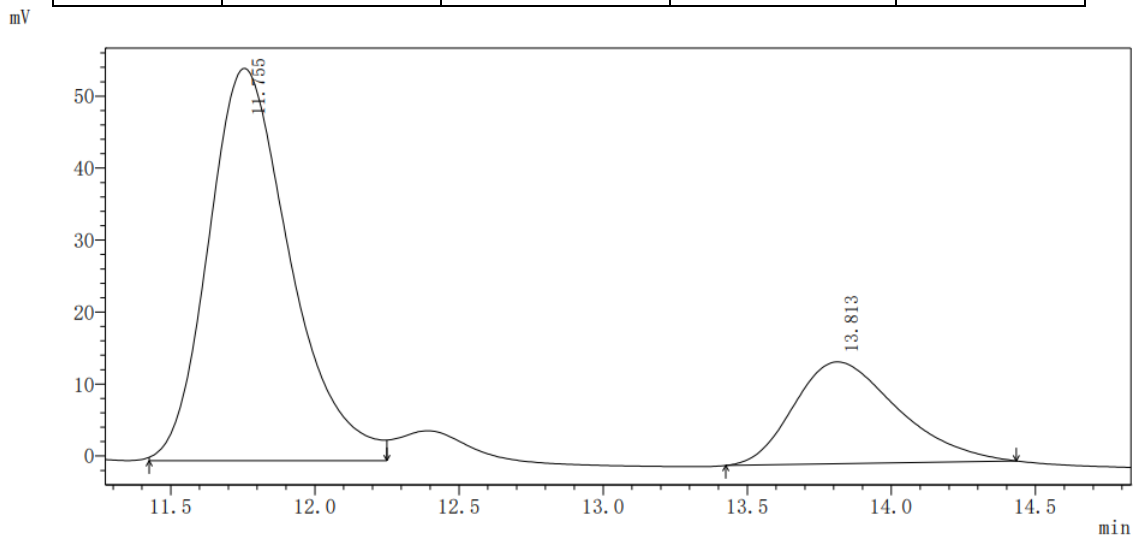
channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	12.786	5763209	364836	99.903
2	15.027	5575	402	0.097
Total		5768784	365237	



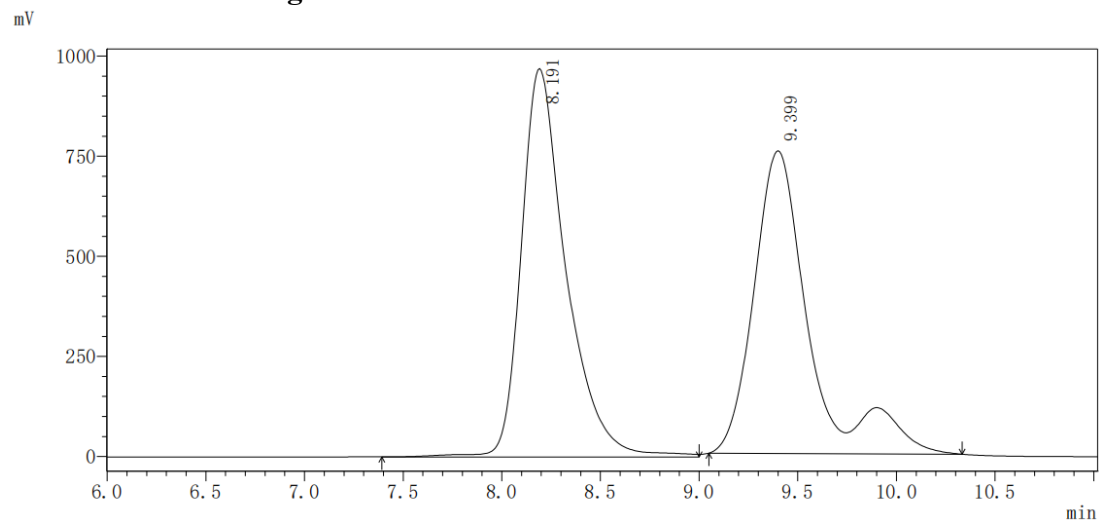
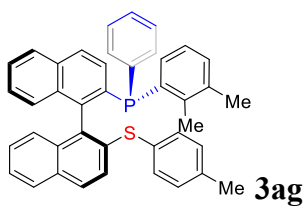
channel 280 nm

Peak	Ret. Time	Area	Height	Conc.
1	11.653	157805	8015	51.358
2	13.771	149461	5509	48.642
Total		307266	13524	



channel 280 nm

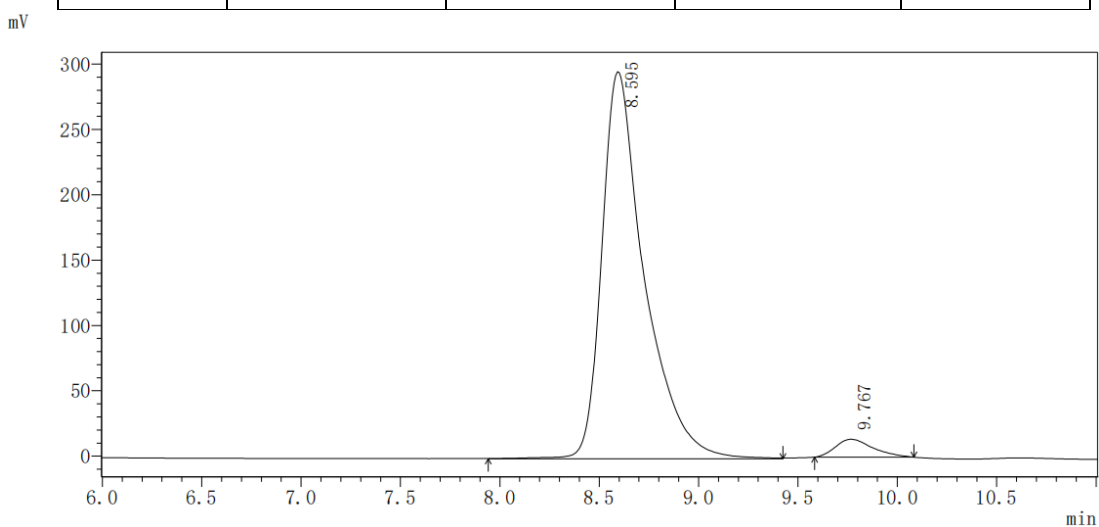
Peak	Ret. Time	Area	Height	Conc.
1	11.755	1078708	54492	75.204
2	13.813	355677	14143	24.796
Total		1434385	68635	



(mixed with diastereomers)

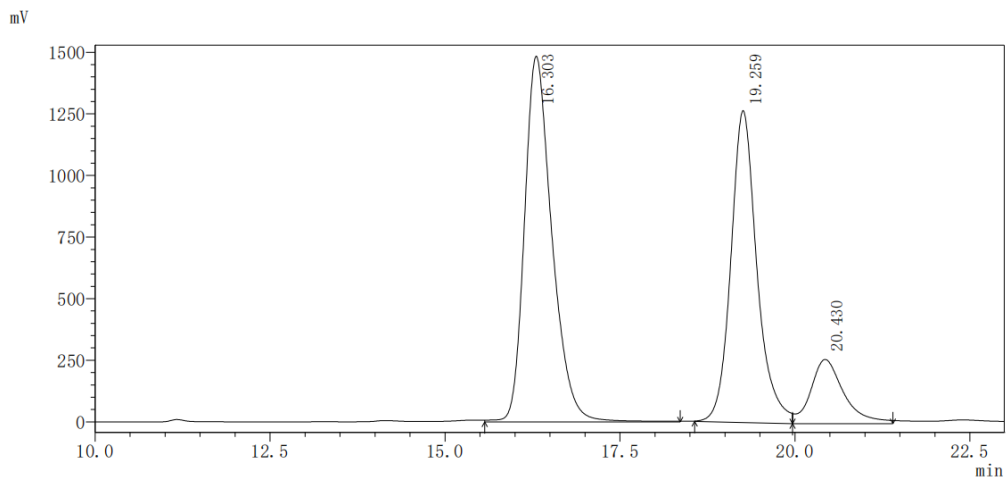
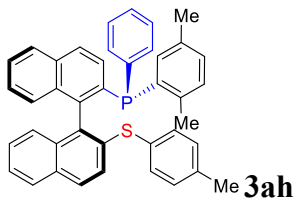
channel 280 nm

Peak	Ret. Time	Area	Height	Conc.
1	8.191	15343105	970125	50.863
2	9.399	14822576	756019	49.137
Total		30165680	1726145	



channel 280 nm

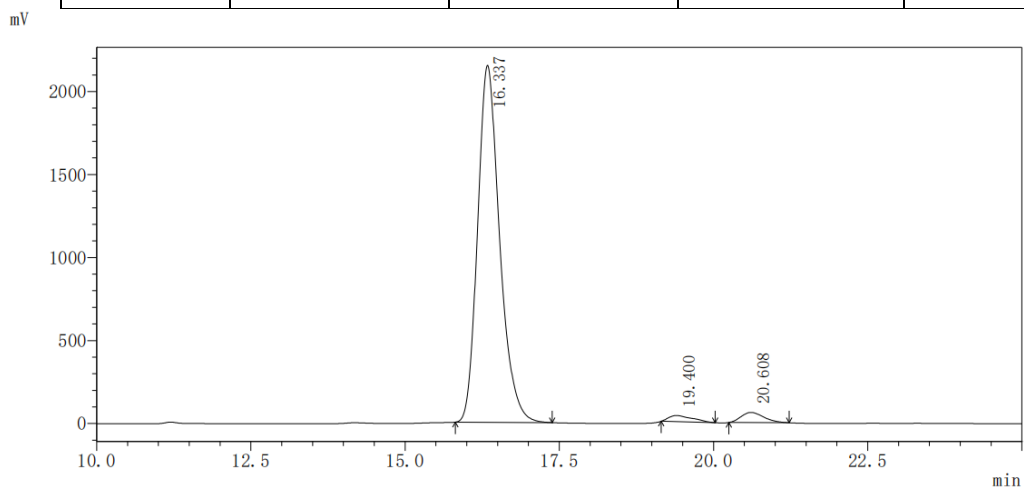
Peak	Ret. Time	Area	Height	Conc.
1	8.595	4411947	296094	96.119
2	9.767	178132	13671	3.881
Total		4590079	309765	



(mixed with diastereomers)

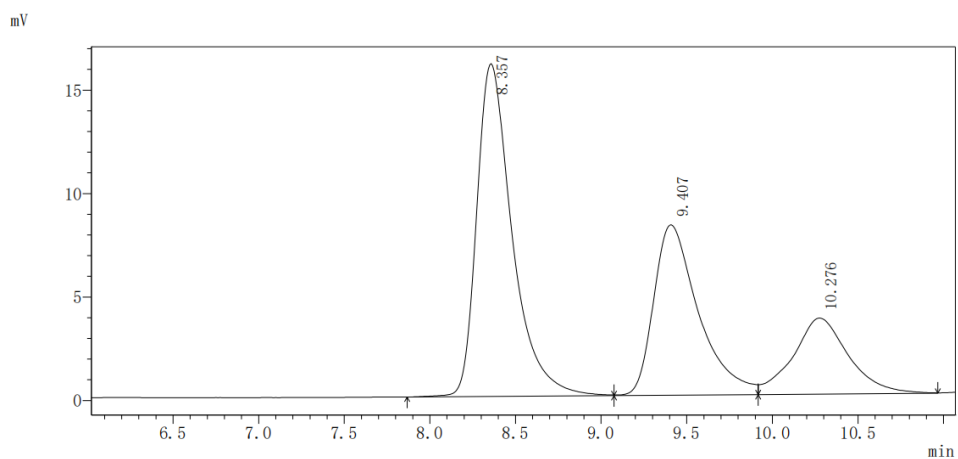
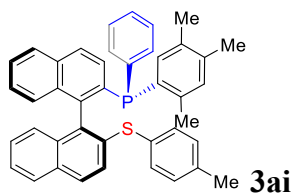
channel 230 nm

Peak	Ret. Time	Area	Height	Conc.
1	16.303	40550639	1483927	49.440
2	19.259	32848704	1266227	40.050
3	20.430	8620300	260844	10.510
Total		82019644	3010999	



channel 230 nm

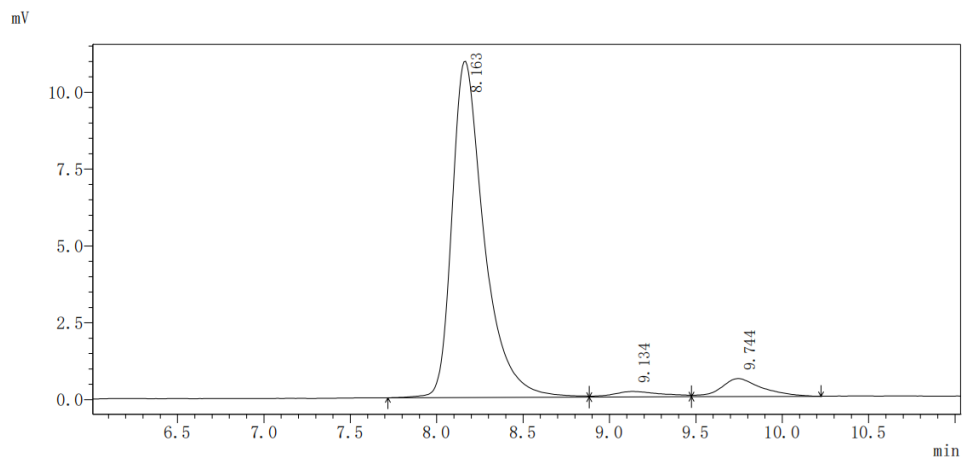
Peak	Ret. Time	Area	Height	Conc.
1	16.337	53459749	2149621	95.443
2	19.400	996853	37245	1.780
3	20.608	1555660	60929	2.777
Total		56012261	2247795	



(mixed with diastereomers)

channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	8.357	231668	16078	49.908
2	9.407	149720	8235	32.254
3	10.276	82803	3679	17.838
Total		464191	27992	



channel 254 nm

Peak	Ret. Time	Area	Height	Conc.
1	8.163	141650	10930	93.626
2	9.134	3534	178	2.336
3	9.744	6109	486	4.038
Total		151293	11594	