# **Electronic Supplementary information (ESI)**

# Sulphur-Atom Positional Engineering in Perylenimide: Structure-Property

# **Relationships and H-aggregation Directed Type-I Photodynamic Therapy**

*Mst Nasima Khatun*<sup>[a]</sup>, *Satyendu Nandy*<sup>[b]</sup>, *Hirakjyoti Roy*<sup>[b]</sup>, *Siddhartha Sankar Ghosh*\*<sup>[b,c]</sup>, *Sachin Kumar*\*<sup>[b]</sup>, *and Parameswar Krishnan Iyer*\*<sup>[a,c]</sup>

<sup>a</sup>Department of Chemistry, Indian Institute of Technology Guwahati, Guwahati-781039, Assam, India.

<sup>b</sup>Department of Bioscience and Bioengineering, Indian Institute of Technology Guwahati, Guwahati-781039, Assam, India

<sup>c</sup>Centre for Nanotechnology, Indian Institute of Technology Guwahati, Guwahati-781039, Assam, India.

FAX: +913612582349; E-mail: pki@iitg.ac.in.

# Contents

1. Experiment Section	2
1.1. Materials and Instrumentation	2
1.2. Preparation of the Test Solution	3
1.3. Preparation of FESEM samples	3
1.4. Theoretical Studies	3
1.5. Photoluminescence Quantumn Yield Calculations	3
1.6. Synthesis of <b>RPNI-O</b> and <b>RPNI-S</b> derivatives	3
1.7. Computational Studies 7	
1.8. <sup>1</sup> O <sub>2</sub> generation quantum yield Estimation	8
1.9. Reactive oxygen species (ROS) Generation/Measurement	8
1.10. The Point of Zero Zeta Potential Evaluation	10
1.11. The Energy of the Valence Band and Conduction Band Calculations	10
1.12. Cell studies	11
1.12.1. Cell culture	11
1.12.2. Dark & Light Cytotoxicity Evaluated by MTT assay	11
1.12.3. Cellular Uptake	11
1.12.4. Live/dead cell co-staining assay	12
2. Supporting figures	63
3. References	83

# 1. Experimental Section

1.1. Materials and Instrumentations: All reagents and starting materials, including Perylene-3,4,9,10-tetracarboxylic acid anhydride (PDA), imidazole, Zinc acetate [Zn (OAc)<sub>2</sub>], and various boronic acid derivatives such as phenylboronic acid, thianthrenyl boronic acid, and 2,2'-bithiophene-5-boronic acid pinacol ester, and other reactive oxygen species detector such as 2',7'-Dichlorodihydrofluorescein diacetate were (DCFDA), 2,2,6,6-Tetramethylpiperidin (TEMP), 9,10-Anthracenediyl-bis(methylene)dimalonic acid (ABDA), Terephthalic acid (TA), Dihydrorhodamine 123 (DHR 123), 2,3-Bis-(2-methoxy-4-nitro-5-sulfophenyl)-2Htetrazolium-5-carboxanilide (XTT) and 5,5-dimethyl-1-pyrroline-N-oxide (DMPO) were purchased from were purchased from Sigma Aldrich (INDIA) in reagent-grade quality. HPLCgrade solvents were obtained from Fisher Scientific Ltd. and RANKEM. Nuclear magnetic resonance (NMR) spectra, including <sup>1</sup>H and <sup>13</sup>C, were recorded using a Bruker Avance 400 MHz spectrometer. The residual solvent signal was used as an internal reference for all solutions used in the NMR experiments. Mass spectra were obtained using MALDI-TOF spectrometry. UV/visible (UV/vis) and photoluminescence (PL) spectra were recorded using a Perkin-Elmer Model Lambda-750 spectrophotometer and a Horiba Fluoromax-4 spectrofluorometer, respectively. Measurements were performed at 298 K using 4 mm quartz cuvettes. A laser diode at 375 nm (DeltaDiode-375) was the excitation source for both UV/vis and PL experiments. The compounds' hydrodynamic diameter and Zeta potential were measured using a Malvern Zetasizer instrument. Field emission scanning electron microscopy (FESEM) images were obtained using a Sigma Carl ZEISS field emission scanning electron microscope. Single crystal data were collected using a Bruker SMART APEX diffractometer with a CCD area detector. The electron spin resonance (ESR) technique was recorded using Bruker EMXplus-10/12 Bruker. For photodynamic therapy studies, a white light source with an intensity of 50 mW cm<sup>-2</sup>, specifically the "Pick Ur Needs 120W Plastic Lithium Battery Search Light Long 1 Km Range with Multi-Functional + Blinker Rechargeable Handheld Torch" in black, was utilized. DMEM, Propidium iodide (PI), and Calcein-AM (AM) were purchased from High Media.

**1.2. Preparation of the Test Solution**: Stock solutions of **RPNI-O** materials, including **PPI**, **THPI**, and **API**, as well as derivatives of **RPNI-S** such as **PPIS**, **THPIS**, and **APIS**, were prepared at a concentration of 20 mM in dimethyl sulfoxide (DMSO). To study the aggregation behavior, test solutions of the materials were prepared at a concentration of 100  $\mu$ M. These test

solutions were prepared by varying the water fraction in DMSO. Before recording the spectra, the resulting solutions were thoroughly shaken at room temperature to ensure proper mixing and dispersion of the compounds.

#### **1.3. Preparation of FESEM samples:**

The morphological analysis of the supramolecular self-assembly was conducted using field emission scanning electron microscopy (FESEM). A simple methodology was employed to prepare the samples for imaging, as depicted in Fig. S8b. A dilute suspension of the perylene derivatives at a concentration of 100  $\mu$ M was prepared using a mixture of 99.9% water and -0.1% DMSO. Subsequently, this suspension was drop-casted onto a glass surface coated with aluminum foil. The samples were then allowed to air dry overnight at room temperature before analysis.

**1.4. Theoretical Studies**: Density functional theory (DFT) calculations were employed to evaluate the electronic properties of **RPNI-O** and **RPNI-S** derivatives. The Gaussian 16 package, incorporating the B3LYP hybrid functional, was utilized to perform these calculations. For each derivative, the ground state optimized geometries were obtained, and the electron density and energy of the highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbital (LUMO) were determined. The calculations employed the 6-31G basis set, which is known to provide accurate results within a reasonable computational time. <sup>1,2</sup>

**1.5. Photoluminescence quantum Yield Calculations**: The fluorescence quantum yields  $(\Phi_{PL})$  of **RPNI-O** and **RPNI-S** derivatives were determined by comparing them to the fluorescence quantum yield of Rhodamine 6G ( $\Phi_r = 0.95$  in ethanol) using a specific equation. The equation used is as follows:

$$\Phi_{PL} = \Phi_r \left( A_r F_s / A_s F_r \right) \left( \eta_s^2 / \eta_r^2 \right)$$
(S1)

In this equation, s and r refer to the sample and reference, respectively.  $\Phi$  represents the quantum yield, A denotes the absorbance, F signifies the relative integrated fluorescence intensity, and  $\eta$  represents the refractive index.

## 1.6. Synthesis of RPNI-O and RPNI-S derivatives

**Synthetic procedures for peryleneimide (PNI-O): PNI-O** was synthesized through a simple condensation reaction employing hydrothermal reaction procedures. Perylene-3,4,9,10-

tetracarboxylic acid anhydride (PDA) (2.54 mmol), amine (1.35 mmol), Zn (OAC)<sub>2</sub> (1.72 mmol), imidazole (5g), and water (1ml) were combined in a Teflon sleeve and thoroughly mixed until a homogeneous mixture was obtained. The homogeneous mixture was transferred to a hydrothermal autoclave reactor vessel and sealed. The reactor was then subjected to a pressure of 15 bar and placed in an oven. The temperature inside the autoclave reactor was gradually increased to 190 °C over 20 hours. The reaction was allowed to proceed under these conditions. After 20 hours, the heat was turned off, and the reaction mixture was allowed to cool down to room temperature. The resulting dark reddish crude product was extracted with chloroform (200 ml) and water (50 ml) under acidic PH. The extraction process was repeated four times to ensure adequate separation. The extracted product was washed four times with water, dried over Na<sub>2</sub>SO<sub>4</sub>, and subsequently filtered. The filtered solution was concentrated using a rotary evaporator to remove the solvent, resulting in a concentrated residue. The concentrated residue was purified using column chromatography with chloroform as the eluent. This purification step yielded the desired dark reddish compound (400 mg) with a 40% yield.

**Characterization data for PNI-O:** <sup>1</sup>H NMR (400 MHz, CDCl3) 0.84 (t, 3H),1.28-1.44 (m, 6H), 1.70 (dd, 2H), 4.09-4.14 (t, 2H), 7.52 (t, 2H), 7.80 (d, 2H), 8.24 (d, 2H), 8.29 (d, 2H), 8.44 (d, 2H)). <sup>13</sup>C NMR (101 MHz, CDCl3)  $\delta$  14.09, 22.58, 26.92, 28.19, 31.64, 40.49, 120.12, 120.83, 123.67, 126.73, 126.96, 127.75, 129.10, 130.90, 131.29, 134.35, 136.94, 163.86. MALDI-TOF: calculated for C<sub>28</sub>H<sub>23</sub>NO<sub>2</sub>: 405.17 [M]<sup>+</sup>, Found: 404.95 [M-H]<sup>+</sup> **Synthetic Route** 



Scheme S1. The synthetic method employed to prepare PNI-O.

**Synthetic procedures for BrPNI-O: PNI-O** (1 mmol) was dissolved in 5 mL of chlorobenzene with gentle heating. Subsequently, bromine (4.5 mmol) was added to the solution, and the resulting mixture was stirred for 4.5 hours at 50 °C. After completion of the reaction, chlorobenzene and any unreacted bromine were removed under vacuum. The resulting solid, which appeared red, was subjected to crystallization using methanol. This process yielded a 95% yield of the desired compound.

Characterization data for 8-bromo-2-hexyl-1H-benzo[5,10]anthra[2,1,9-def]isoquinoline-1,3(2H)-dione (BrPNI-O): <sup>1</sup>H NMR (400 MHz, CDCl3)  $\delta$  0.93 (t, 3H), 1.3-1.6 (m, 6H), 1.88 (dd, 2H), 3.77 (t, 2H), 7.69 (t, 1H),7.86 (d, 1H), 8.13 (d, 1H), 8.27 (d, 2H), 8.33 (d, 1H). 8.38 (d, 1) 8.55 (dd, 2H). <sup>13</sup>C NMR (101 MHz, CDCl3)  $\delta$ .14.09, 22.58, 26.92, 28.19, 31.64, 40.49, 120.12, 123.67, 126.96, 127.75, 129.15, 130.80, 131.32, 172.56 MALDI-TOF: calculated for C<sub>28</sub>H<sub>22</sub>BrNO<sub>2</sub>: 484.38 [M]<sup>+</sup>, Found: 483.39 [M-H]<sup>+</sup>

**Synthetic Route** 



Scheme S2. The synthetic method employed to prepare BrPNI-O.

**Synthetic procedures for RPNI-O:** In a 50 mL round-bottom flask, Compound **BrPNI-O** (0.5 mmol) and the corresponding boronic acid (1 mmol) were combined with 5 mg of Pd (PPh<sub>3</sub>)<sub>4</sub> catalyst. The mixture was then subjected to nitrogen gas for 30 min to remove any oxygen. Then, 6 mL of THF (tetrahydrofuran) was added to the flask, followed by 2 mL of a 2.0 M potassium carbonate solution. The resulting mixture was stirred at 85 °C under a nitrogen atmosphere, and the progression of the reactions was monitored using thin-layer chromatography (TLC). After 24 hours, the solutions were cooled and extracted using CHCl<sub>3</sub> (chloroform). The organic layers were subsequently dried using anhydrous sodium sulfate. The solvent was evaporated, and the resulting product was purified using column chromatography, yielding a 60% yield of the desired Red solid for **PPI** and **API** and **THPI** (brownish solid) compound.

Characterization data for 2-hexyl-8-phenyl-1H-benzo[5,10]anthra[2,1,9-def]isoquinoline-1,3(2H)-dione (PPI): <sup>1</sup>H NMR (400 MHz, CDC13)  $\delta$  0.89 (t, 3H), 1.41 (m, 6H), 1.76 (dt, 2H), 4.14 (t, 2H), 7.53 (m, 7H),7.98 (d, 1H) 8.37 (d, 2H), 8.42 (t, 2H), 8.55 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDC13)  $\delta$  14.07, 22.62, 26.77, 28.07, 29.02, 31.56, 40.39, 114.10, 120.13, 119.69, 123.56, 126.05, 128.52, 129.20, 130.01, 131.99, 132.66, 136.66, 139.74, 143.11, 163.73. MALDI-TOF: calculated for C<sub>34</sub>H<sub>29</sub>NO<sub>2</sub>: 481.20 [M]<sup>+</sup>, Found: 481.321[M]<sup>+</sup> Characterization data for 8-([2,2'-bithiophen]-5-yl)-2-hexyl-1H-benzo[5,10]anthra[2,1,9-def]isoquinoline-1,3(2H)-dione (THPI): <sup>1</sup>H NMR (400 MHz, CDCl3)  $\delta$  0.88 (t, 3 H), 1.39 (m,5H), 1.75 (dt,3H), 4.19 (t,2H), 7.08 (t, 1H), 7.29 (m, 4H), 7.64 (t, 1H), 7.70 (d, 1H), 8.35 (m, 3H), 8.41 (t, 2H), 8.54 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl3)  $\delta$  12.98, 22.92, 26.64, 27.25, 28.02, 30.90, 32.50, 39.56, 113.69, 118.96, 119.56, 122.50, 126.83, 127.54, 128.52, 131.29, 135.54, 138.75, 142.01, 162.55. MALDI-TOF: calculated for C<sub>36</sub>H<sub>27</sub>NO<sub>2</sub>S<sub>2</sub>: 569.14 [M]<sup>+</sup>, Found:569.43 [M]<sup>+</sup>

Characterizationdatafor2-hexyl-8-(thianthren-1-yl)-1H-benzo[5,10]anthra[2,1,9-def]isoquinoline-1,3(2H)-dione (API):  ${}^{1}$ H NMR (400 MHz, CDCl3)  $\delta$  0.89 (t, 3H), 1.39 (m, 6H),1.77 (m, 2H), 4.22 (t, 2H), 7.15 (t, 3H), 7.34 (d, 1H), 7.40 (t, 3H), 7.54 (dd, 3H), 7.65 (d, 3H).8.50 (m, 1) 8.64 (dd, 2H).  ${}^{13}$ C NMR (101 MHz, CDCl3)  $\delta$  14.09, 27.09, 28, 59, 31.57, 40.49,120.36, 121.25, 126.28, 127.84, 128.67, 129.12, 132.4, 135.44, 136.43, 137.39, 139.66, 164.12.MALDI-TOF: calculated for C40H29NO2S2: 619.79 [M]<sup>+</sup>, Found: 620.79 [M+H]<sup>+</sup>

## **Synthetic Route**



Scheme S3. The synthetic method employed to prepare RPNI-O.

**Synthetic procedures for RPNI-S:** In a 20 mL flask, **RPNI-O** (0.5 mmol) and Lawesson's reagent (1.5 mmol) were dissolved in dry toluene. The resulting solution was refluxed under a nitrogen atmosphere for 24 hours. After completion of the reaction, the mixture was allowed to cool down to room temperature, and the solvent was evaporated under reduced pressure. The residue was purified using column chromatography on silica gel, employing a hexane/dichloromethane (DCM) mixture as the eluent. Subsequent drying under vacuum conditions yielded a bluish powder (53-58% yield).

Characterization data for 2-hexyl-8-phenyl-1H-benzo[5,10]anthra[2,1,9-def]isoquinoline-1,3(2H)-dithione (PPIS): <sup>1</sup>H NMR (400 MHz, CDC13)  $\delta$  0.90 (t, 3H), 1.40 (m,6H), 1.78 (dt, 2H) ), 4.21 (m, 2H), 7.16 (q, 3H), 7.34 (d, 3H), 7.41 (t, 1H), 7.53 (d, 1H), 7.56 (d, 1H), 7.65 (d, 1H), 8.48 (dd, 1H), 8.54 (d, 1H) 8.63 (dd, 2H). <sup>13</sup>C NMR (101 MHz, CDCl3)  $\delta$  13.07, 21.66, 25.87, 27.03, 28.67, 30.57, 49.54, 113.04, 118.31, 119.47, 121.45, 122.33, 125.63, 126.94, 127.56, 128.09, 129.95, 131.95, 132.25, 138.75, 142.09, 165.3. MALDI-TOF: calculated for C<sub>34</sub>H<sub>27</sub>NS<sub>2</sub>: 513.15 [M]<sup>+</sup>, Found: 513.29 [M]<sup>+</sup>

Characterization data for 8-([2,2'-bithiophen]-5-yl)-2-hexyl-1H-benzo[5,10]anthra[2,1,9-def]isoquinoline-1,3(2H)-dithione (THPIS): <sup>1</sup>H NMR (400 MHz, CDCl3)  $\delta$  0.89 (t, 3H), 1.41 (m, 6H), 1.76 (dt, 2H) 4.19 (t,2H), 7.08 (t, 1H), 7.29 (m, 6H), 7.64 (t, 1H), 7.71 (d, 1H), 8.35 (m, 3H), 8.40 (dd, 1H), 8.54 (t, 2H). <sup>13</sup>C NMR (101 MHz, CDCl3)  $\delta$  13.09, 21.59, 25.67, 27.04, 28.77, 30.67, 48.54, 114.04, 117.31, 118.57, 120.55, 121.43, 123.73, 125.74, 125.56, 127.01, 128.95, 130.05, 131.25, 141.45, 142.09, 169.3. MALDI-TOF: calculated for C<sub>36</sub>H<sub>27</sub>NS<sub>4</sub>: 601.10 [M]<sup>+</sup>, Found: 601.19 [M]<sup>+</sup>

Characterizationdatafor2-hexyl-8-(thianthren-1-yl)-1H-benzo[5,10]anthra[2,1,9-def]isoquinoline-1,3(2H)-dithione (APIS): $^{1}$ H NMR (400 MHz, CDCl3)  $\delta$  0.91 (t, 3H), 1.42 (m,6H),1.78 (dt, 2H),4.21 (t, 2H),7.16 (t, 3H),7.34 (t, 1H),7.40 (t, 1H),7.51 (d, 4H),7.65 (d, 2H),8.47 (m, 1H),8.54 (d, 1H),8.63 (dd, 2H).14.10,22.61,26.89,28.12,31.62,42.4,120.36,121.25,123.40,124.03,128.67,129.21,132.43,135.44,136.01,137.89,139.66,170.30.MALDI-TOF: calculated forC40H29NS4:651.11 [M]<sup>+</sup>,Found:650.40 [M-H]<sup>+</sup>

**Synthetic Route** 



Scheme S4. The synthetic method employed to prepare RPNI-S.

# **1.7.** Computational studies

This study examined the optimization and frequencies of triplet excited states, excitation energies, and transition properties of these peryleneimide materials RPNI-O and RPNI-S

derivatives. These calculations were performed using the time-dependent B3LYP/6-31G (d, p) method within the Tamm-Dancoff approximation (TDA). The computational software used for these calculations was the Gaussian 16 package, with the ORCA 5.0 module utilized at the B3LYP DEF2-SVP level. Furthermore, the spin-orbit coupling (SOC) constants for **PPI**, **THPI**, **API**, **PPIS**, **THPIS**, and **APIS** were also computed at the abovementioned level, employing the ORCA 5.0 software.<sup>3,4</sup>

**1.8.** <sup>1</sup>O<sub>2</sub> generation quantum yield Estimation: In this study, dilute solutions of Rose Bengal were used as a reference ( $\Phi_{\Delta} = 0.76$  in ethanol). To eliminate the inner filter effect, the absorbance maxima of Rose Bengal and Photosensitizer were maintained at 0.2. Additionally, various solutions of **RPNI-O** and **RPNI-S** derivatives were prepared at a concentration of 100  $\mu$ M. Then, 100 uM ABDA was added to these solutions. The absorbance of ABDA at 378 nm was measured at various irradiation times using white light (400-700 nm, 50 mW cm<sup>-2</sup>). This measurement allowed the determination of the decay rate of the photosensitizing process. The  $\Phi_{\Delta}$  of **RPNI-O** and **RPNI-S** derivatives in 99% PBS in DMSO was then calculated using the following equations:

 $\Phi_{\text{AIEgen}} = \Phi_{\text{RB}} (K_{\text{AIEgen}} A_{\text{RB}}/K_{\text{RB}} A_{\text{AIEgen}}) \dots (S2)$ 

Where  $\Phi_{AIE}$  is the <sup>1</sup>O<sub>2</sub> generation quantum yield of AIEgen in 99% PBS in DMSO. A<sub>RB</sub> and A<sub>AIEgen</sub> represent the light absorbed by Rose Bengal and AIEgen, respectively, at an absorbance 0.2. The absorption bands were integrated into the 400-700 nm wavelength range to determine these values. K<sub>RB</sub> and K<sub>AIEgen</sub> are the decomposition rate constants of ABDA by Rose Bengal and AIEgen, respectively. These constants were determined by plotting Ln (A<sub>0</sub>/A) against time, where A<sub>0</sub> and A are the absorbances of ABDA at 378 nm by photosensitizers under white light irradiation at different time points.<sup>5,6</sup>

## 1.9. Reactive oxygen species (ROS) Generation/Measurement

Singlet Oxygen ( $^{1}O_{2}$ ) Test: This experiment was conducted to quantify the generation  $^{1}O_{2}$ . Singlet oxygen, produced through the photoirradiation of PSs at 99% PBS in DMSO, was detected using ESR spectroscopy in the presence of TEMP. The ESR signals from the free-radical TEMPO, formed from the reaction of singlet oxygen with TEMP, were recorded after irradiation with white light. Solutions containing 100  $\mu$ M TEMP and 100  $\mu$ M **RPNI-O** or **RPNI-S** PSs were inserted quantitatively into quartz capillaries at 99% PBS in DMSO. These solutions were then subjected to white light irradiation at a power density of 50 mW cm<sup>-2</sup> for 30 min. ESR spectra of the spin-trapped radicals were recorded in the range of 3000-3600 G after irradiation. Background interference was corrected using a sample before irradiation.<sup>7</sup>

Further, a mixed solution containing 100  $\mu$ M ABDA (a singlet oxygen sensor) and 100  $\mu$ M of various **RPNI-O** and **RPNI-S** derivatives solutions was prepared in 99% PBS in DMSO. The solution was irradiated with white light at a power density of 50 mW cm<sup>-2</sup> for different durations: 0, 2, 4, 6, 8, 10, and 20 min. The degradation of ABDA's absorbance at 378 nm was measured promptly after each interval.

<u>Total ROS Test:</u> The total ROS test measures the overall reactive oxygen species (ROS) levels. DCFDA, a chemically reduced form of fluorescein, was used as a fluorescent indicator for ROS.\_A solution of 10  $\mu$ M DCFDA and 100  $\mu$ M of various solutions of **RPNI-O** and **RPNI-S** derivatives was prepared at 99% PBS in DMSO.\_The solution was irradiated with white light at a power density of 50 mW cm<sup>-2</sup> for different durations: 0, 5, 10, 15, 20, 25, and 30 min.\_The fluorescence spectra of the mixed solution were rapidly measured.

# Typ-I ROS :

# Superoxide Anion Radical (O2 • ) Test:

1) The feasibility of the reaction producing  $O_2^{\bullet-}$  was checked through theoretical calculations using the ORCA 5.0 software.

2) ESR analysis was conducted to verify the generation of  $O_2$ <sup>--</sup> utilizing DMPO as the spintrapping agent. The experimental samples were quantitatively inserted into quartz capillaries, comprising 100 µM DMPO at 99% PBS in DMSO and 100 µM of **PPIS**, **THPIS**, or **APIS**. Subsequently, the spin spectra were monitored after subjecting the solutions to 10 min white light irradiation (50 mW cm<sup>-2</sup>).

3) XTT was employed as the indicator for  $O_2^{\bullet-}$ , which undergoes conversion to XTT formazan in the presence of  $O_2^{\bullet-}$ . A mixed solution of XTT (100  $\mu$ M) and **PPIS**, **THPIS**, or **APIS** PSs (100  $\mu$ M) was prepared. Subsequently, the solution was irradiated with white light for durations of 0, 5, 10, 20, and 30 min, and the UV-vis spectra of the mixed solution were promptly measured. Additionally, UV-vis spectra of XTT irradiated solely by white light were recorded as the control. 4) DHR123 was utilized as the indicator for superoxide anion radicals, which undergo conversion to Rhodamine 123 in the presence of  $O_2^{\bullet-}$ . Solutions containing 100  $\mu$ M **PPIS**, **THPIS**, or **APIS** at 99% PBS in DMSO were prepared. Subsequently, the cuvette containing the solution was exposed to white light (400-700 nm, 50 mW cm<sup>-2</sup>) for varying durations (5-30 min), and the fluorescence spectra were promptly recorded after each irradiation. In the control group, a DHR123 solution without photosensitizers was irradiated. ( $\lambda_{ex} = 500$  nm,  $\lambda_{em} = 527$  nm).

<u>Hydroxyl Radical (HO') Test:</u> This test aimed to measure the generation of hydroxyl radicals (HO').\_A mixed solution containing 100  $\mu$ M TA and 100  $\mu$ M of various solutions of **RPNI-O** and **RPNI-S** derivatives compounds was prepared in 99% PBS in DMSO.\_The subsequent testing process was the same as determining total ROS using DCFDA as the probe.\_Upon trapping with HO', TA was converted to hydroxyl terephthalic acid, resulting in an enhanced fluorescent intensity ( $\lambda_{ex} = 315$  nm).

## 1.10. The Point of Zero Zeta Potential Evaluation

According to the literature,<sup>8</sup> the points of zero zeta potential (PZZP) for **RPNI-O** and **RPNI-S** derivatives were calculated. The aqueous solutions of these Perylenimide materials were initially at a neutral pH of about 7. However, by introducing a dilute solution of sodium hydroxide or dilute hydrochloric acid, the pH of the solutions could be adjusted to either alkaline or acidic conditions. When the pH values of the **RPNI-O** and **RPNI-S** materials were altered within 9, 7.4, 5.6, to 3 and 1, a noticeable change in the zeta potential was observed. Specifically, the zeta potential transitioned from negative to positive as the pH values changed from alkaline to acidic.

#### 1.11. The Energy of the Valence Band and Conduction Band Calculations

The valence band energy  $(E_V)$  was determined by calculating the energy of the

band ( $E_V$ ) from X-ray photoelectron spectroscopy (XPS) data. To calculate the conduction band energy ( $E_C$ ), the energy gap was first estimated from the onset absorption data from UVvisible spectroscopy. Subsequently,  $E_C$  was calculated concerning the normal hydrogen electrode (NHE) using Equations S3 and S4, as shown below:

$$E_{\rm V} = -E_{\rm HOMO} - 4.5 \tag{S3}$$

$$E_{\rm C} = -E_{\rm LUMO} - 4.5 \tag{S4}$$

The calculated  $E_V$  and  $E_C$  values for the **RPNI-O** and **RPNI-S** derivatives were recorded in Table S17. Additionally,  $E_C$  and  $E_V$  at a point of zero zeta potential (PZZP) with a pH of 5.6 were computed based on literature data using Equations S5 and S6, respectively:

$$E_{\rm C,pH} = E_{\rm C} + 0.059 \times (PZZP - pH) \tag{S5}$$

$$E_{\rm V,pH} = E_{\rm V} + 0.059 \times (PZZP - pH) \tag{S6}$$

Where PZZP represents the points of zero zeta potential for the **RPNI-O** and **RPNI-S** materials at pH 5.6. The estimated  $E_C$  and  $E_V$  values at a pH of 5.6 for **PPI, THPI, API, PPIS, THPIS**, and **APIS** were also included in Table S17.

## 1.12. Cell studies

**1.12.1. Cell culture:** HeLa (cervical cancer cells) and MCF 7 (breast cancer cells) were procured from the National Centre for Cell Sciences (NCCS) in Pune, India. These cells were cultured in Dulbecco's Modified Eagle Medium (DMEM), supplemented with 10% fetal bovine serum and 100 U/ml each of penicillin and streptomycin. The cell cultures were maintained in a controlled environment with 5% CO<sub>2</sub> at 37 °C.

**1.12.2.** Dark & Light Cytotoxicity Evaluated by MTT assay: In the cell viability assay conducted on cancer cells (HeLa and MCF7 cell lines), the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) method was employed. Initially, 5000 cells were seeded into individual wells of a 96-well plate and incubated in DMEM media inside a 5% CO<sub>2</sub> humidified incubator at 37 °C for 24 hours. Subsequently, the cells were treated with different concentrations of **RPNI-O** and **RPNI-S** compounds for 4 hours. After the 4-hour treatment, specific cell samples were exposed to white light using a handheld torch, 50 mW cm<sup>-2</sup>, for 20 min. Following irradiation, the cells were further incubated with other treatment groups for 24 hours at 37 °C in the same 5% CO<sub>2</sub> humidified incubator. Upon completion of the incubation period, the MTT assay was performed. The MTT assay measures cell viability based on the reduction of MTT by mitochondrial enzymes in viable cells, producing a formazan product. This formazan product is detected spectrophotometrically at 570 nm, with a reference filter set at 655 nm. The resulting absorbance of the formazan product is directly proportional to the number of viable cells present in the samples.

**1.12.3. Cellular Uptake:** After an overnight incubation period, Hela cells were seeded into 96-well plates with a cell density of  $1 \times 10^4$  cells per well. Twenty-four hours later, the Hela cells

were treated with varying concentrations of **RPNI-S** derivatives such as **PPIS**, **THPIS**, and **APIS**, respectively, at 20  $\mu$ M, while HEK 293T cells were treated with 100  $\mu$ M of the same compounds. Additionally, Hela cells were exposed to **RPNI-O** derivative at a concentration of 20  $\mu$ M and 100  $\mu$ M, and HEK 293T cells were treated with 100  $\mu$ M of the corresponding compound. These treatments were performed at different intervals under normal oxygen conditions ranging from 0, 1, 2, 3, 4, 5, and 6 hours. Subsequently, the cells were washed three times with DPBS to remove any excess **RPNI-O** and **RPNI-S** compounds. Each well's optical density (OD) values were then measured using a multifunction microplate reader (BioTek, Synergy H1). (BioTek, Synergy H1,  $\lambda_{abs.max} = 500$  nm for **PPIS**, THPIS,  $\lambda_{abs.max} = 580$  nm for **THPIS**, and 636 nm for **APIS**).

**1.12.4.** Live/dead cell co-staining assay: In the study, HeLa cells were initially seeded in 35mm glass-bottomed dishes at a density of 3 x 10<sup>5</sup> cells per dish and cultured overnight in standard culture media. Subsequently, the cells were treated with **RPNI-S** derivatives, such as **PPIS**, **THPIS**, and **APIS**, at a concentration of 50  $\mu$ M for 4 hours. After the 4-hour treatment, the cells were subjected to white light irradiation using a handheld torch with a specific wavelength (50 mW cm<sup>-2</sup>) for 20 min. The cells were co-stained with 2  $\mu$ M Calcein AM and 4  $\mu$ M propidium iodide (PI) for I h. After washing with DPBS (Dulbecco's Phosphate-Buffered Saline), fluorescence images of the cells were acquired using a fluorescence microscope. Additionally, another set of HeLa cells was treated with the same **RPNI-S** derivatives at a concentration of 50  $\mu$ M for 4 hours. These cells were treated with 2',7'dichlorodihydrofluorescein diacetate (DCFDA) at 10  $\mu$ M for I h. After washing with DPBS, the cells were irradiated with the same handheld torch (50 mW cm<sup>-2</sup>) for 20 min, and their fluorescence images were acquired. Moreover, HeLa cells were cultured in a normoxic environment (21% O<sub>2</sub>) for 24 hours.

**Table S1** Below is a comparison table of recently published heavy-atom-free materials, highlighting their design strategies, structure-property relationships, and photosensitizing behaviors for PDT efficacy.

		J. Am. Chem. Soc.
References	<b>Present work</b>	<b>2019</b> , <i>141</i> , 16243-
		16248

Materials		THPIS	PPIS	API	MANI-S
AIE		Non- fluorescent (a)	Non- fluorescent (a)	Red AIEE <sub>gen</sub> (a)	Non-fluorescent (b)
Stokes- shift (nm)		g	g	146	g
$\Phi_{ m PL}$		g	g	0.85 (a)	g
$\Phi_{\Delta}$		0.29 (c)	0.11 (c)	g	1.00 (d)
Light used		50mWcm <sup>-2</sup> (e)	50 mWcm <sup>-2</sup> (e)	50 mWcm <sup>-2</sup> (e)	100 mW cm <sup>-2</sup> (e)
ROS	Types	Type-I, Type-II ( <sup>1</sup> O <sub>2</sub> , O <sub>2</sub> )	Туре-I (О2 <sup></sup> )	Type-I (O₂ <sup></sup> )	Туре-I, Туре-II ( <sup>1</sup> О <sub>2</sub> , О <sub>2</sub> -)
	Specificity	Specific	Specific	Specific	Not very specific
Total ROS	DCFDA (µM)	10	10	10	10
	T (min)	5	5	5	3

	I	1x10 <sup>6</sup>	2.5x10 <sup>5</sup>	5x10 <sup>4</sup>	7.5x10 <sup>4</sup>
ROS (mechanism)		DCFDA, ABDA, TA, ESR, Theoretical (ΔG, SOC), Band energy/NHE)	DCFDA, ABDA, TA, ESR, Theoretical (ΔG, SOC), Band energy/NHE)	DCFDA, ABDA, TA, ESR, Theoretical (ΔG, SOC), Band energy/NHE)	DCFDA, DPBF, DHE, Theoretical (SOC)
PDT (Cancer cell)	In vitro	Normoxia (Hela, MCF7, normal cell)	Normoxia (Hela, MCF7, normal cell)	g	Hypoxia (Hela cell)
	Selectivity	Selective	Selective	g	g
Liposome		Х	Х	g	Albumin
Design strategy		Structure-property relationship and application (H-aggregation unveiled PDT efficacy)			Application-based
		S-atom at the carbonyl and functional position			S-atom at the carbonyl position

a, b, c, and d, respectively, are the different solvent systems (a = 99%  $f_w$ , b = toluene, c = 99% PBS, and d = ethanol, respectively). In contrast, e is the white light used for the photosensitization [g = not observed, x = not used (normal cell non-toxic)].

**Table S2** Comparison of the blue-shifted  $\lambda_{abs.max}$  with the reported data has demonstrated enhanced H-aggregation, which is crucial for PDT efficacy by boosting the SOC value.

	References	Materials	Blue-shifted λ <sub>abs.max</sub> (nm)	H-aggregation
		PPIS	89	Highly pronounced
	<b>Present work</b>	THPIS	76	Highly pronounced
		APIS	23	Moderate
		РТТе	74	Pronounced
а	Adv. Mater. 2022, 34, 210814	PTSe	71	Pronounced
		PTS	30	Moderate
b	Nature Materials 2015, 14, 685-690	DPhCzT	14	Moderate
с	Nanoscale <b>2016</b> , 8, 17422-17426	(OB4) <sub>4</sub>	10	Moderate

a, b, and c are the references 41, 28, and 29 present in the main manuscript.

Table S3 CIE coordinates for the RPNI-O and RPNI-S derivatives in solution, aggregated, and solid state.

	Solu	ition	Aggre	egated	So	lid
Materials						
	X	Y	X	Y	X	Y
PPI	0.562	0.437	0.638	0.36	0.689	0.31
THPI	0.635	0.364	0.644	0.355	0.725	0.274
API	0.504	0.493	0.617	0.382	0.684	0.315
PPIS	0.73	0.269	А	А	А	A
THPIS	0.724	0.275	A	A	А	A
APIS	0.734	0.265	A	А	А	A

A represents not observed due to non-emissive characteristics.



Fig. S1 Photophysical characteristics of all the **RPNI-O** (**PPI**, **THPI**, and **API**) and **RPNI-S** (**PPIS**, **THPIS**, and **APIS**) derivatives. (a) The PL spectra in their solution state represent monomeric emission ( $\lambda_{ex} = 500$  nm). (b) PL spectra in their aggregated-state (at 99%  $f_w$  in DMSO) ( $\lambda_{ex} = 500$  nm). (c) Solid state PL spectra ( $\lambda_{ex} = 500$  nm). (d) Fluorescence intensity plot at various  $f_w$  in DMSO at their corresponding  $\lambda_{em,max}$ .



**Fig. S2** (a and b) The absorbance and emission peak area of rhodamine 6G. (c and d) The absorbance and emission peak area of **API** at 40%  $f_w$  in DMSO. IF: integrated fluorescence intensity.



**Fig. S3** (a and b) The absorbance and emission peak area of rhodamine 6G. (c, d), (e, f) and (g, h) The absorbance and emission peak area of **PPI**, **THPI**, and **API** at 99%  $f_w$  in DMSO. IF: integrated fluorescence intensity. Rh6G represent rhodamine 6G.



**Fig. S4** Normalized UV-vis spectra (a-c) of **RPNI-O** derivatives (**PPI, THP**I, and **API**), and (d-f) **RPNI-S** derivatives (**PPIS, THPIS,** and **APIS**) at various water fractions ( $f_w$ ) in DMSO. [**RPNI-O**] or [**RPNI-S**] = 100  $\mu$ M. Inset **Fig. S4** (c') Altered normalized UV-vis spectra of 0 and 99 %  $f_w$ .

Materials	Water fraction ( <i>f</i> <sub>w</sub> )	$\lambda_{abs.max}(nm)$	Α	$\Delta\lambda$ (nm)
	0%	506	0.394105	
PPI				60
	99%	446	0.159005	
	0%	517	0.059899	
THPI				46
	99%	471	0.021479	
	0%	510	0.248319	
API				10
	99%	500	0.151884	
	0%	590	0.053059	
PPIS				89
	99%	501	0.006136	

Table S4 UV-vis absorption data for the RPNI-O and RPNI-S derivatives.

	0%	580	0.030367	
THPIS				76
	99%	514	0.012369	
	0%	636	0.37716	
APIS				23
	99%	613	0.220526	

A,  $\Delta\lambda$ , are the absorbance and wavelength maxima shifts in their absorption.



**Fig. S5** Solvatochromic effect of the **RPNI-O** derivatives. (a-c) Normalized absorbance, (d-f) Normalized fluorescence spectra of **PPI**, **THPI**, and **API** in various solvents from nonpolar to polar (100  $\mu$ M,  $\lambda_{ex} = 500$  nm).

**Table S5** The solvatochromic UV-vis data for the **RPNI-O** derivatives in various solvents from nonpolar to polar (100  $\mu$ M,  $\lambda_{ex} = 500$  nm).

Materials	Solvents	λabs.max (nm)	$\Delta\lambda$ (nm)
	Toluene	488	
PPI	Dioxane	488	16
	CHCl <sub>3</sub>	494	-

	CH <sub>3</sub> CN	497	
	МеОН	498	
	DMF	502	
	<b>D</b> 1/(20	201	_
	DMSO	504	
	Toluene	504	
	Toluelle	504	
	Dioxane	508	_
	2 1011011	200	
	CHCl <sub>3</sub>	514	-
THPI	CH <sub>3</sub> CN	514	16
	MeOH	518	
			_
	DMF	522	
	DMSO	522	_
	DWISO	522	
	Toluene	504	
	Dioxane	507	
	CHCl <sub>3</sub>	507	
API	CH <sub>3</sub> CN	507	13
	МеОН	512	_
	IVICOII	512	
	DMF	513	-
	DMSO	517	

 $\Delta\lambda$  is the wavelength maxima shift in their absorption

**Table S6** The solvatochromic PL data for the **RPNI-O** derivatives in various solvents from nonpolar to polar (100  $\mu$ M,  $\lambda_{ex} = 500$  nm).

Materials	Solvents	λ <sub>em.max</sub> (nm)	Δλ (nm)
PPI	Toluene	539	45

	Dioxane	547	
	CHCl <sub>3</sub>	562	
	CH <sub>3</sub> CN	566	
	MeOH	573	
	DMF	578	
	DMSO	584	
	Toluene	602	
	Dioxane	609	•
	CHCl <sub>3</sub>	611	•
THPI	CH <sub>3</sub> CN	622	51
	МеОН	623	
	DMF	631	
	DMSO	653	•
	Toluene	538	
	Dioxane	539	
	CHCl <sub>3</sub>	554	
API	CH <sub>3</sub> CN	554	34
	МеОН	559	
	DMF	559	
	DMSO	572	

 $\Delta\lambda$ , is the wavelength maxima shift in their absorption.

#### S-atom engineering at the functional positions



Fig. S6 Capturing daylight photographs of **RPNI-O** derivatives (**PPI, THPI**, and **API**) and **RPNI-S** derivatives (**PPIS, THPIS, and APIS**), respectively at 0%, 40%, and 99%  $f_w$  in DMSO.



S-atom engineering at the carbonyl positions: non-emissive characteristics

**Fig. S7** (a1-c4) 2D Excitation-emission (EEM) contour projections of the **RPNI-S** derivatives at different concentrations (left: 100  $\mu$ M; middle: 300  $\mu$ M and 500  $\mu$ M; right: 1 mM) in DMSO. [Inset: arrow represents the fluorescence intensity].



**Fig. S8** (a) FESEM images of the micro-assembly of the **THPIS**, spontaneously formed at 99.9%  $f_w$  in DMSO (100  $\mu$ M) by a simple drop-casting technique on aluminum foil coated on a glass slide, followed by drying at room temperature, as graphically represented in b. (b) Schematic illustration of the highly reproducible, simple, cost-effective, and feasible strategy for preparing micro- and nano-supramolecular self-assembly from **RPNI-O** and **RPNI-S** derivatives.



**Fig. S9** Single crystal structures of (a) **PPI**, (b) **THPI**, and (c) **API**, respectively, show evidence of Hand J- aggregate formation in these molecules, corresponding to their schematic showing the  $\pi$ - $\pi$ -stacking interaction among the **PNI-O** cores. The angle between the transition dipoles and the interconnected axis is indicated by  $\theta$ . (d-f) single crystal unit cell of (d) **PPI**, (e) **THPI**, and (f) **API**, respectively.

Name	PPI	THPI	API
Formula	$C_{34}H_{29}NO_2$	$C_{36}H_{27}NO_2S_2$	$C_{40}H_{29}NO_2S_2$
Formula weight	108.37	70.71	607.75
Wavelength (Å)	0.71073	0.71073	0.71073
Space Group	P -1	P -1	P -1
Crystal System	triclinic	triclinic	triclinic
Temperature	293 (K)	293 (K)	293 (K)
	a 17.6473(9)	a 7.3852(5)	a 8.766(6)

Table S7 SCXRD data of the RPNI-O derivatives.

Cell Lengths	b 7.5383(4)	b 11.7473(8)	b 11.916(8)
(Å)	c 18.6063(9)	c 16.5975(11)	c 15.464(11)
	α 89.999(2)	α 83.044(2)	α 101.43(2)
Cell Angles (o)	β 94.196(2)	β 84.880(2)	β 92.94(2)
	γ 90.014(2)	γ 77.501(2)	γ 111.20(2)
Cell Volume (Å3)	2468.6(2)	1392.51(16)	1462.8(18)
Z	1	2	2
Density (g/cm3)	0.418	0.361	1.380
F(000)	78	98	636
hmax, kmax, lmax	2,8,2	8,1,3	9,13,16
CCDC	2325822	2325824	2325826



**Fig. S10** The ROS quantum yield of **RPNI-S** was determined through chemical trapping measurements. ABDA is undergoing photodegradation with (a) Rose Bengal, (d) **PPIS**, (g) **THPIS**, and (j) **APIS**, respectively. Rate constants for the decomposition of ABDA in the presence of (b) Rose Bengal, (e) **PPIS**, (h) **THPIS**, and (k) **APIS**, respectively. The integrated area of the absorption peak for (c) Rose Bengal. (f) **PPIS**, (i) **THPIS**, and (l) **APIS**, respectively.



**Fig. S11** The ROS quantum yield of **RPNI-O** was determined through chemical trapping measurements. ABDA is undergoing photodegradation with (a) **PPI**, (d) **THPI**, and (g) **API**, respectively. Rate constants for the decomposition of ABDA in the presence of (b) **PPI**, (e) **THPI**, and (h) **API**, respectively. The integrated area of the absorption peak for (c) **PPI**, (f) **THPI**, and (i) **API**, respectively.

**Table S8** Spin-orbit coupling matrix elements (SOCMEs) of **PPIS** compound in optimized structure.  $\langle S_m | H_{SO} | T_n \rangle$ , cm<sup>-1</sup> and  $\Delta E_{ST}$ , eV calculated using ORCA 5.0 at the B3LYP/6-31G (d, p) and B3LYP/DEF2SVP level.

Excite state electronic transition	$\Delta E_{\rm ST} ({ m eV})$	SOC (cm <sup>-1</sup> )
$S_1 \leftrightarrow T_1$	0.402	8.33
$S_1 \leftrightarrow T_2$	0.205	3.00
$S_2 \leftrightarrow T_1$	0.794	6.53

$S_2 {\leftrightarrow} T_2$	0.597	4.16
$S_2{\leftrightarrow}T_3$	0.251	7.24
$S_2{\leftrightarrow}T_4$	0.096	10.80
$S_3 {\leftrightarrow} T_2$	0.99	7.88
S₃↔T₃	0.637	5.46
S₃↔T₄	0.482	4.25
$S_4 {\leftrightarrow} T_4$	0.99	6.00
$S_4 {\leftrightarrow} T_5$	0.637	1.34
S₄⇔T <sub>6</sub>	0.482	0.75
S₅⇔T₅	0.681	3.01
S₅⇔T <sub>6</sub>	0.36	3.69
S₅⇔T7	0.16	2.63
S₅⇔T∗	0.092	1.25
S₅⇔T₂	0.046	5.01
S₅⇔T10	0.02	6.57
S <sub>6</sub> ↔T₅	0.72	5.09
S <sub>6</sub> ↔T <sub>6</sub>	0.3990	6.16
$S_6 \leftrightarrow T_7$	0.199	5.20
$S_6 \leftrightarrow T_8$	0.131	3.08
S <sub>6</sub> ↔T9	0.085	2.43
$S_6 \leftrightarrow T_{10}$	0.059	1.76
$S_7 \leftrightarrow T_5$	0.754	6.54

S <sub>7</sub> ↔T <sub>6</sub>	0.44	6.50
$S_7 \leftrightarrow T_7$	0.233	9.16
$S_7 \leftrightarrow T_8$	0.165	3.02
S <sub>7</sub> ↔T <sub>9</sub>	0.119	1.32
$S_7 \leftrightarrow T_{10}$	0.093	3.01
$S_8 \leftrightarrow T_5$	0.864	1.18
S <sub>8</sub> ↔T <sub>6</sub>	0.543	2.05
S <sub>8</sub> ↔T <sub>7</sub>	0.343	2.62
S <sub>8</sub> ↔T <sub>8</sub>	0.275	0.88
S <sub>8</sub> ↔T <sub>9</sub>	0.229	5.74
$S_8 \leftrightarrow T_{10}$	0.203	4.75
S₂↔T₅	0.905	1.38
S₂⇔T6	0.584	1.99
S₂↔T7	0.384	1.57
S₂↔T8	0.316	1.09
S₂↔T₂	0.27	2.65
S <sub>9</sub> ↔T <sub>10</sub>	0.244	4.32
$S_{10} {\leftrightarrow} T_7$	0.46	2.79
$S_{10} \leftrightarrow T_8$	0.392	1.34
S10↔T9	0.346	4.79
S10↔T10	0.32	3.83

**Table S9** SOCMEs of **THPIS** compound in optimized structure.  $\langle S_m | H_{SO} | T_n \rangle$ , cm<sup>-1</sup> and  $\Delta E_{ST}$ , eV calculated using ORCA 5.0 at the B3LYP/6-31G (d, p) and B3LYP/DEF2SVP level.

Excite state electronic transition	$\Delta E_{\rm ST} ({\rm eV})$	SOC (cm <sup>-1</sup> )
$S_1 \leftrightarrow T_1$	0.418	6.90
$S_1 \leftrightarrow T_2$	0.202	2.32
$S_2 \leftrightarrow T_1$	0.81	5.49
$S_2 \leftrightarrow T_2$	0.594	5.15
S₂↔T₃	0.247	6.14
$S_2 \leftrightarrow T_4$	0.094	9.20
S₃↔T₂	0.798	5.47
S₃↔T₃	0.451	3.75
S₃↔T₄	0.298	2.70
S₃↔T₅	0.017	0.60
S₃↔T₃	0.857	4.90
S₄↔T₄	0.704	3.61
S₄↔T₅	0.423	0.65
S₄↔T₀	0.043	0.44
S₅⇔T₄	0.892	4.91
S₅⇔T₅	0.611	1.65
S₅⇔T <sub>6</sub>	0.231	0.87
S₅⇔T7	0.054	0.52
S <sub>6</sub> ↔T <sub>5</sub>	0.921	3.56

S <sub>6</sub> ↔T <sub>6</sub>	0.541	5.97
S <sub>6</sub> ↔T <sub>7</sub>	0.364	4.40
$S_6 \leftrightarrow T_8$	0.227	3.92
S <sub>6</sub> ↔T9	0.075	3.02
S <sub>6</sub> ↔T <sub>10</sub>	0.046	1.91
S <sub>7</sub> ↔T₅	0.98	2.14
S <sub>7</sub> ↔T <sub>6</sub>	0.60	3.32
S <sub>7</sub> ↔T <sub>7</sub>	0.423	3.29
S <sub>7</sub> ↔T <sub>8</sub>	0.286	3.00
S <sub>7</sub> ↔T <sub>9</sub>	0.134	5.45
S <sub>7</sub> ↔T <sub>10</sub>	0.105	0.80
S <sub>8</sub> ↔T <sub>6</sub>	0.649	2.74
$S_8 {\leftrightarrow} T_7$	0.472	4.14
$S_8 \leftrightarrow T_8$	0.3350	4.50
S <sub>8</sub> ↔T <sub>9</sub>	0.183	0.57
$S_8 \leftrightarrow T_{10}$	0.154	0.75
S9↔T6	0.724	2.13
S9↔T7	0.547	1.57
S9↔T8	0.41	2.00
S9↔T9	0.258	4.21
S <sub>9</sub> ↔T <sub>10</sub>	0.229	1.41
S10↔T6	0.826	1.14

S10↔T7	0.649	0.80
S10↔T8	0.512	0.88
S10↔T9	0.36	2.33
S10↔T10	0.331	0.44

**Table S10** SOCMEs of **APIS** compound in optimized structure.  $\langle S_m | H_{SO} | T_n \rangle$ , cm<sup>-1</sup> and  $\Delta E_{ST}$ , eV calculated using ORCA 5.0 at the B3LYP/6-31G (d, p) and B3LYP/DEF2SVP level.

Excite state electronic transition	$\Delta E_{\rm ST} ({ m eV})$	SOC (cm <sup>-1</sup> )
$S_1 \leftrightarrow T_1$	0.386	4.44
$S_1 \leftrightarrow T_2$	0.207	1.62
$S_2 \leftrightarrow T_1$	0.778	6.37
$S_2 \leftrightarrow T_2$	0.599	4.47
$S_2 \leftrightarrow T_3$	0.253	3.83
$S_2 \leftrightarrow T_4$	0.098	5.73
S₃↔T₂	0.907	2.36
S₃⇔T₃	0.561	2.58
$S_3 \leftrightarrow T_4$	0.406	2.16
S₄↔T₃	0.682	5.63
S₄↔T₄	0.527	4.34
S₄↔T₅	0.121	2.16
S₅⇔T₄	0.916	3.63
S₅⇔T₅	0.51	1.34
S₅↔T <sub>6</sub>	0.311	1.46

S₅⇔T7	0.034	0.42
S <sub>6</sub> ↔T₄	0.958	1.61
S <sub>6</sub> ⇔T₅	0.552	2.18
S <sub>6</sub> ⇔T <sub>6</sub>	0.353	1.00
S <sub>6</sub> ⇔T <sub>7</sub>	0.076	0.52
S <sub>7</sub> ⇔T₅	0.897	1.47
S <sub>7</sub> ⇔T <sub>6</sub>	0.698	3.49
S <sub>7</sub> ↔T <sub>7</sub>	0.421	2.96
S <sub>7</sub> ↔T <sub>8</sub>	0.325	2.54
S <sub>7</sub> ↔T <sub>9</sub>	0.161	1.60
S <sub>7</sub> ↔T <sub>7</sub>	0.128	0.20
S <sub>8</sub> ⇔T₅	0.911	1.11
S <sub>8</sub> ⇔T <sub>6</sub>	0.712	4.25
$S_8 \leftrightarrow T_7$	0.435	3.15
$S_8 \leftrightarrow T_8$	0.339	2.94
$S_8 \leftrightarrow T_9$	0.175	1.71
$S_8 \leftrightarrow T_{10}$	0.142	3.28
S9↔T5	0.956	1.89
S9↔T6	0.757	2.57
S9↔T7	0.48	1.85
S9↔T8	0.384	1.88
S9↔T9	0.22	1.36

S <sub>9</sub> ↔T <sub>10</sub>	0.187	3.04
S10↔T6	0.853	1.16
S10↔T7	0.576	0.77
S10↔T8	0.48	0.97
S10↔T9	0.316	0.69
S10↔T10	0.283	2.04

**Table S11** SOCMEs of **PPI** compound in optimized structure.  $\langle S_m | H_{SO} | T_n \rangle$ , cm<sup>-1</sup> and  $\Delta E_{ST}$ , eV calculated using ORCA 5.0 at the B3LYP/6-31G (d, p) and B3LYP/DEF2SVP level.

Excite state electronic transition	$\Delta E_{\rm ST} \left( { m eV}  ight)$	SOC (cm <sup>-1</sup> )
S₁↔T₁	1.196	0.3
$S_1 \leftrightarrow T_2$	-0.17	0.77
$S_2 \leftrightarrow T_1$	1.678	3.70
$S_2 \leftrightarrow T_2$	0.305	2.14
S₂↔T₃	0.207	2.39
S₂↔T₄	0.19	3.53
S₂↔T₅	0.048	3.72
$S_3 \leftrightarrow T_2$	0.609	0.39
S₃↔T₃	0.511	1.93
S₃↔T₄	0.352	0.95
S₃↔T₅	0.21	1.56
S₄↔T₃	0.576	3.81
S₄⇔T₄	0.559	1.51
---------------------------------	-------	------
S₄↔T₅	0.417	3.22
S₄↔T6	0.275	2.14
$S_4 \leftrightarrow T_7$	0.143	1.89
$S_4 \leftrightarrow T_8$	0.174	3.69
S₄↔T9	0.083	3.13
$S_4 \leftrightarrow T_{10}$	0.024	0.78
S₅⇔T₄	0.594	2.45
S₅⇔T₅	0.452	3.54
S₅⇔T₀	0.31	3.00
S₅↔T7	0.209	0.88
S₅⇔T8	0.178	2.49
S₅⇔T9	0.118	2.77
S₅⇔T10	0.059	0.95
S <sub>6</sub> ↔T₅	0.53	3.17
S <sub>6</sub> ↔T <sub>6</sub>	0.388	3.12
$S_6 \leftrightarrow T_7$	0.287	1.67
$S_6 \leftrightarrow T_8$	0.256	1.23
S <sub>6</sub> ↔T9	0.196	1.74
S <sub>6</sub> ↔T <sub>10</sub>	0.137	0.79
S <sub>7</sub> ↔T₅	0.585	2.62
S7↔T6	0.443	2.64

$S_7 \leftrightarrow T_7$	0.342	1.49
S <sub>7</sub> ↔T <sub>8</sub>	0.311	1.54
S <sub>7</sub> ↔T <sub>9</sub>	0.251	1.32
$S_7 \leftrightarrow T_{10}$	0.192	0.59
S <sub>8</sub> ⇔T <sub>6</sub>	0.522	2.35
$S_8 \leftrightarrow T_7$	0.421	1.92
$S_8 \leftrightarrow T_8$	0.39	2.75
S <sub>8</sub> ↔T <sub>9</sub>	0.33	1.58
S <sub>8</sub> ↔T <sub>10</sub>	0.271	0.35
S9↔T6	0.614	2.07
S₂↔T7	0.513	3.03
S₂↔T8	0.482	2.49
S₂↔T₂	0.422	1.32
S₂↔T₁0	0.363	0.26
S10↔T6	0.642	2.67
S <sub>10</sub> ↔T <sub>7</sub>	0.541	2.15
S10↔T8	0.51	3.07
S10↔T9	0.45	1.58
S10↔T10	0.391	0.35

**Table S12** SOCMEs of **THPI** compound in optimized structure.  $\langle S_m | H_{SO} | T_n \rangle$ , cm<sup>-1</sup> and  $\Delta E_{ST}$ , eV calculated using ORCA 5.0 at the B3LYP/6-31G (d, p) and B3LYP/DEF2SVP level.

Excite state electronic transition	$\Delta E_{\rm ST} ({ m eV})$	SOC (cm <sup>-1</sup> )

S₁↔T₁	0.981	0.65
$S_1 \leftrightarrow T_2$	0.036	0.85
$S_2{\leftrightarrow}T_1$	1.442	0.91
$S_2 \leftrightarrow T_2$	0.497	0.74
$S_3{\leftrightarrow}T_2$	0.761	2.33
S₃↔T₃	0.257	1.33
S₃↔T₄	0.203	1.07
S₃↔T₅	0.176	0.3
S₃↔T₀	0.153	1.85
$S_3 \leftrightarrow T_7$	0.04	3.37
$S_4 \leftrightarrow T_3$	0.554	0.32
S₄⇔T₄	0.50	1.47
S₄↔T₅	0.473	0.92
S₄↔T₀	0.45	0.47
S₄↔T7	0.337	1
S₄↔T <sub>8</sub>	0.184	0.4
S₄↔Tջ	0.091	0.97
S₄↔T10	0.05	1.66
S₅⇔T₄	0.568	2.26
S₅⇔T₅	0.541	1.55
S₅⇔T₀	0.518	1.07
S₅⇔T7	0.405	1.36

S₅⇔T8	0.252	0.51
S₅⇔T9	0.159	1.63
S₅↔T₁0	0.118	2.25
S <sub>6</sub> ↔T₅	0.56	2.67
S <sub>6</sub> ↔T <sub>6</sub>	0.537	2.01
$S_6 \leftrightarrow T_7$	0.424	3.44
$S_6 \leftrightarrow T_8$	0.271	2.55
S <sub>6</sub> ↔T <sub>9</sub>	0.178	1.81
S <sub>6</sub> ⇔T <sub>10</sub>	0.137	3.50
S <sub>7</sub> ↔T₅	0.683	1.90
S <sub>7</sub> ↔T <sub>6</sub>	0.66	1.58
S <sub>7</sub> ↔T <sub>7</sub>	0.547	2.98
S <sub>7</sub> ↔T <sub>8</sub>	0.394	3.06
S <sub>7</sub> ↔T <sub>9</sub>	0.301	1.54
S <sub>7</sub> ↔T <sub>10</sub>	0.26	0.51
$S_8 \leftrightarrow T_6$	0.687	0.37
$S_8 {\leftrightarrow} T_7$	0.574	0.24
$S_8 \leftrightarrow T_8$	0.421	0
S <sub>8</sub> ↔T9	0.328	0.32
S <sub>8</sub> ↔T <sub>10</sub>	0.287	0.2
S9↔T6	0.728	1.59
S9↔T7	0.651	1.68

S₂↔T8	0.462	1.55
S₂↔Tջ	0.369	1.82
S <sub>9</sub> ↔T <sub>10</sub>	0.328	1.73
S10↔T6	0.783	0.87
S10↔T7	0.67	1.08
S10↔T8	0.517	1.84
$S_{10} \leftrightarrow T_9$	0.424	1.65
S10↔T10	0.383	2.07

**Table S13** SOCMEs of **API** compound in optimized structure.  $\langle S_m | H_{SO} | T_n \rangle$ , cm<sup>-1</sup> and  $\Delta E_{ST}$ , eV calculated using ORCA 5.0 at the B3LYP/6-31G (d, p) and B3LYP/DEF2SVP level.

Excite state electronic transition	$\Delta E_{\rm ST} \left( { m eV}  ight)$	SOC (cm <sup>-1</sup> )
$S_1 \leftrightarrow T_1$	1.015	1.920
$S_1 \leftrightarrow T_2$	0.003	0.57
$S_2 \leftrightarrow T_1$	1.238	0.87
$S_2 \leftrightarrow T_2$	0.226	2.36
$S_3 \leftrightarrow T_2$	0.551	2.18
S₃⇔T₃	0.141	0.69
S₃⇔T₄	0.116	0.223
S₃⇔T₅	0.096	0.47
S₄⇔T₃	0.234	2.04
S₄↔T₄	0.209	1.87
S₄↔T₅	0.189	2.43

S₄↔T₀	0.087	3.13
S₄↔T7	0.041	2.97
S₅⇔T₄	0.571	1.95
S₅⇔T₅	0.551	0.28
S₅⇔T₀	0.449	0.48
S₅⇔T7	0.403	0.77
S₅⇔T <sub>8</sub>	0.265	0.39
S₅⇔Tջ	0.167	0.96
S₅⇔T10	0.122	1.43
$S_6 \leftrightarrow T_5$	0.571	2.87
$S_6 \leftrightarrow T_6$	0.469	1.84
$S_6 \leftrightarrow T_7$	0.423	1.76
$S_6 \leftrightarrow T_8$	0.285	1.67
S <sub>6</sub> ↔T <sub>9</sub>	0.187	0.97
$S_6 \leftrightarrow T_{10}$	0.142	2.23
S <sub>7</sub> ↔T₅	0.694	2.93
S <sub>7</sub> ↔T <sub>6</sub>	0.592	1.42
S <sub>7</sub> ↔T <sub>7</sub>	0.546	1.61
S <sub>7</sub> ⇔T <sub>8</sub>	0.408	1.89
S <sub>7</sub> ↔T <sub>9</sub>	0.31	1.23
S <sub>7</sub> ↔T <sub>10</sub>	0.265	0.74
S <sub>8</sub> ↔T <sub>6</sub>	0.619	0.93

$S_8 \leftrightarrow T_7$	0.573	0.85
$S_8 \leftrightarrow T_8$	0.435	1.22
S <sub>8</sub> ↔T9	0.337	0.84
$S_8 \leftrightarrow T_{10}$	0.292	0.81
S9↔T6	0.692	0.56
S9↔T7	0.646	0.75
S9↔T8	0.508	1.36
S₂↔T₂	0.41	1.47
S₂↔T₁0	0.365	1.72
S <sub>10</sub> ↔T <sub>6</sub>	0.735	0.49
$S_{10}{\leftrightarrow}T_7$	0.689	0.56
$S_{10} {\leftrightarrow} T_8$	0.551	1.09
S <sub>10</sub> ↔T9	0.453	1.13
S10↔T10	0.408	1.45

Table S14a Comparision of the SOCMEs with the most probable lowest excited singlet and triplet state transitions is conducted using the optimized isolated structures of previously reported PSs and conventional organic materials.

	References	Materials	SOC (cm <sup>-1</sup> )
		PPIS	10.88
	Present Work	THPIS	9.20
		APIS	5.73
a	Adv. Mater. 2022, 34, 2108146.	PTTe	0.28
9	Angew. Chem. Int. Ed. 2018, 57, 7997-8001.	o-MCBA	0.624

10	ACS Nano <b>2019</b> , 13, 11283-11293.	TPAPyPF6	0.648015
b	ACS Nano 2020, 14, 16840-16853.	TPE-PTB	4.53
11	ACS Nano <b>2021</b> , 15, 7328-7339.	MeTIN	1.242
12	Chem. Sci. <b>2020</b> , 11, 10921-10927.	РТР	0.04
с	J. Mater. Chem. B. <b>2022</b> , 10, 6228-6236.	gem-OMe-TPE-1CUM	0.340
d	Nanoscale <b>2016</b> , 8, 17422-17426	(OB4)1	0.42
13	Phys. Chem. Chem. Phys. 2023, 25, 24386-24394.	BODIPY heterodimers	0.74
14	J. Phys. Chem. A 2007, 111, 10490-10499.	monoaza[5]helicenes	0.1-3.0
15	J. Phys. Chem. C. 2023, 127, 2694-2704.	BT2O3Cz	0.462
e	Biomaterials <b>2021</b> , 275, 120934.	TPAIN	0.7250

a, b, c, d, and e correspond to references 41, 40, 50, 29, and 49, respectively, present in the main manuscript.

**Table S14b** Calculated different excite state singlet (S) and triplet (T) state energy of the **RPNI-O** and **RPNI-S** derivatives via ORCA 5.0 at the B3LYP/6-31G (d, p) and B3LYP/DEF2SVP level.

Materials		S (eV)									T (eV)									
	S <sub>1</sub>	$S_2$	$S_3$	$S_4$	$S_5$	<b>S</b> <sub>6</sub>	$\mathbf{S}_7$	<b>S</b> <sub>8</sub>	S9	S <sub>10</sub>	T <sub>1</sub>	<b>T</b> <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	<b>T</b> <sub>7</sub>	T <sub>8</sub>	T9	T <sub>10</sub>
PPIS	1.61	2.004	2.390	2.811	3.147	3.186	3.220	3.330	3.371	3.447	1.210	1.407	1.753	1.908	2.466	2.787	2.987	3.055	3.101	3.127
THPIS	1.595	1.987	2.191	2.597	2.785	3.095	3.154	3.203	3.278	3.380	1.177	1.393	1.740	1.893	2.174	2.554	2.731	2.868	3.020	3.049
APIS	1.613	2.005	2.313	2.434	2.823	2.865	3.210	3.224	3.269	3.365	1.227	1.406	1.752	1.907	2.313	2.512	2.789	2.885	3.049	3.082
PPI	2.677	3.152	3.456	3.521	3.556	3.634	3.689	3.768	3.860	3.888	1.474	2.847	2.945	2.962	3.104	3.246	3.347	3.378	3.438	3.497
THPI	2.403	2.864	3.128	3.425	3.493	3.512	3.635	3.662	3.703	3.758	1.422	2.367	2.871	2.925	2.952	2.975	3.088	3.241	3.334	3.375
API	2.513	2.736	3.061	3.154	3.516	3.536	3.659	3.686	3.759	3.802	1.498	2.510	2.920	2.945	2.965	3.067	3.113	3.251	3.349	3.394



**Fig. S12** ROS generation by the **RPNI-S** derivatives. (a-c) Plots of ABDA (100  $\mu$ M) degradation rates at  $\lambda_{abs.max}$  of 378 nm by the various PSs (100  $\mu$ M) (a) **PPIS**, (b) **THPIS**, and (c) **APIS**, respectively and (d-f) Evaluation of the general ROS generation using DCFDA (10  $\mu$ M) indicator by PSs (100  $\mu$ M) (d) **PPIS**, (e) **THPIS** and (f) **APIS**, respectively upon white light excitation for different time periods at 99% PBS fraction in DMSO ( $\lambda_{ex}$ =500 nm). Inset: Chemical structure of the **RPNI-S** derivatives.



**Fig. S13** (a) Plot of fluorescence intensities of DCFDA indicator in the presence of the **RPNI-O** and **RPNI-S** derivatives, respectively. ROS generation by the **RPNI-O** derivatives. (b-d) Plots of ABDA (100  $\mu$ M) degradation rates at  $\lambda_{abs.max}$  of 378 nm by the various PSs (100  $\mu$ M) (b) **PPI**, (c) **THPI**, and (d) **API**, respectively and (e-g) Evaluation of the general ROS generation using DCFDA (10  $\mu$ M) indicator by PSs (100  $\mu$ M) (e) **PPI**, (f) **THPI** 

and (g) **API**, respectively upon white light excitation for different time periods at 99% PBS fraction in DMSO ( $\lambda_{ex}$ =500 nm). Inset: Chemical structure of the **RPNI-O** derivatives.



**Fig. S14** (a) PL spectra of TA at 528 nm. (b) Zeta potential versus pH values of **RPNI-O** derivatives. (c)  $E_C$  and  $E_V$  of **RPNI-O** derivatives at pH 5.6. The energy scale is expressed in relation to NHE.  $E_H$  for H<sub>2</sub>O/HO<sup>•</sup>, <sup>1</sup>O<sub>2</sub>/O<sub>2</sub>, and O<sub>2</sub>/O<sub>2</sub> <sup>-</sup> are 2.2, 1.88, and -0.2 eV, respectively.

Table S15 Zeta potential data of RPNI-O and RPNI-S derivatives at different PH.

Materials	РН							
	1	3	5.6	7.4	9			
ΡΡΙζ	1.49	-3.99	-11.90	-31.30	-32.50			

ΤΗΡΙζ	-2.18	-11.90	-19.30	-20.10	-22.70
ΑΡΙζ	3.35	-16.10	-16.30	-30.90	-35.80
ΡΡΙSζ	2.68	-1.89	-10.79	-21.13	-22.45
ΤΗΡΙSζ	2.18	-7.90	-18.20	-19.10	-20.70
ΑΡΙδζ	4.35	-13.4	-14.30	-31.90	-38.80

 $\zeta$  is the zeta potential of the respective materials.



**Fig. S15** XPS spectra displaying valence band energy (**E**<sub>v</sub>): (a) **PPIS**, (b) **THPIS**, (c) **APIS**, (d) **PPI**, (e) **THPI**, and (f) **API**, respectively.



**Fig. S16** UV-vis spectroscopy of exhibiting onset absorption (a) **PPIS**, (b) **THPIS**, (c) **APIS**, (d) **PPI**, (e) **THPI**, and (f) **API**, respectively.

 Table S16 Optical band gap data of RPNI-O and RPNI-S derivatives calculated from the onset absorption of the UV-vis spectroscopy.

Materials	λonset	E <sub>g</sub> (eV)
PPIS	742	1.67
THPIS	766	1.61
APIS	764	1.62
PPI	595	2.08
THPI	627	1.98
API	597	2.07

 $\lambda_{onset}$  onset UV-vis absorption

Table S17  $E_C$  and  $E_V$  at a pH of 5.6 of **RPNI-O** and **RPNI-S** derivatives.

Materials	Eg	Еномо	Elumo	Ev	Ec	PZZP	Ev,5.6	Ec,5.6

PPIS	1.67	-5.60	-3.93	1.10	-0.57	2.68	0.92772	-0.74228
THPIS	1.61	-5.48	-3.87	0.98	-0.63	2.18	0.77822	-0.83178
APIS	1.62	-5.23	-3.61	0.73	-0.89	4.35	0.65625	-0.96375
PPI	2.08	-5.62	-3.54	1.12	-0.96	1.49	0.87751	-1.20249
THPI	1.98	-5.61	-3.63	1.11	-0.87	-2.18	0.65098	-1.32902
API	2.07	-5.17	-3.10	0.67	-1.40	3.35	0.53725	-1.53275

PZZP, the point of zero zeta potential

**Table S18** Calculated Gibbs free energy of  $O_2$  using ORCA 5.0 at the B3LYP/6-31G (d, p) andB3LYP/DEF2SVP level using quantum mechanism package.

Materials		ΔG (Kcal mol <sup>-1</sup> )	
	O <sub>2</sub>	-150.09346315	
PPIS	PPIS+O <sub>2</sub>	-2310.53867284	-0.548
	PPIS	-2160.44608328	
	O <sub>2</sub>	-150.09346315	
THPIS	THPIS+O <sub>2</sub>	-3182.63436769	-40.83
	THPIS	-3032.55916191	
	O2	-150.09346315	
APIS	APIS+O <sub>2</sub>	-3336.04242916	-10.26
	APIS	-3185.96532129	

**Table S19** Calculated Gibbs free energy of **O**<sub>2</sub> using ORCA 5.0 at the B3LYP/6-31G (d, p) and B3LYP/DEF2SVP level using a quantum mechanism package.

#### -----

# **O**<sub>2</sub>

GIBBS FREE ENERGY

The Gibbs free energy is G = H - T\*S Total enthalpy ... -150.07131287 Eh Total entropy correction ... -0.02215028 Eh -13.90 kcal/mol

Final Gibbs free energy ... -150.09346315 Eh

For completeness - the Gibbs free energy minus the electronic energy

G-E(el) ... -0.01441918 Eh -9.05 kcal/mol

Timings for individual modules:

Sum of individual times  $\dots$  12.524 sec (= 0.209 min)

GTO integral calculation ... 2.186 sec (= 0.036 min) 17.5 %

SCF iterations ... 4.342 sec (= 0.072 min) 34.7 %

Analytical frequency calculation... 5.995 sec (= 0.100 min) 47.9 %

**Table S20** Calculated Gibbs free energy of **PPIS** using ORCA 5.0 at the B3LYP/6-31G (d, p) and B3LYP/DEF2SVP level using quantum mechanism package.

#### -----

\_\_\_\_\_

# PPIS

## GIBBS FREE ENERGY

**Table S21** Calculated Gibbs free energy of **PPIS** and **O**<sub>2</sub> using ORCA 5.0 at the B3LYP/6-31G (d,p) and B3LYP/DEF2SVP level using a quantum mechanism package.

-----

\_\_\_\_\_

GIBBS FREE ENERGY

### PPIS +O<sub>2</sub>

The Gibbs free energy is G = H - T\*STotal enthalpy ... -2310.44123598 Eh Total entropy correction ... -0.09743686 Eh -61.14 kcal/mol ------Final Gibbs free energy ... -2310.53867284 Eh For completeness - the Gibbs free energy minus the electronic energy G-E(el) ... 0.45873772 Eh 287.86 kcal/mol Timings for individual modules: Sum of individual times ... 19281.091 sec (= 321.352 min) GTO integral calculation ... 103.385 sec (= 1.723 min) 0.5 % SCF iterations ... 12827.456 sec (= 213.791 min) 66.5 % SCF Gradient evaluation ... 2722.936 sec (= 45.382 min) 14.1 % Geometry relaxation ... 24.973 sec (= 0.416 min) 0.1 % Analytical frequency calculation... 3602.340 sec (= 60.039 min) 18.7 % **Table S22** Calculated Gibbs free energy of **THPIS** using ORCA 5.0 at the B3LYP/6-31G (d,p) and B3LYP/DEF2SVP level using quantum mechanism package.

#### THPIS \_\_\_\_\_ GIBBS FREE ENERGY \_\_\_\_\_ The Gibbs free energy is $G = H - T^*S$ Total enthalpy ... -3032.47582297 Eh Total entropy correction ... -0.08333893 Eh -52.30 kcal/mol Final Gibbs free energy ... -3032.55916191 Eh For completeness - the Gibbs free energy minus the electronic energy G-E(el) ... 0.47671745 Eh 299.14 kcal/mol Timings for individual modules: Sum of individual times $\dots$ 3337.858 sec (= 55.631 min) GTO integral calculation ... 4.858 sec (= 0.081 min) 0.1 % SCF iterations ... 251.657 sec (= 4.194 min) 7.5 % Analytical frequency calculation... 3081.344 sec (= 51.356 min) 92.3 %

**Table S23** Calculated Gibbs free energy of **THPIS** and **O**<sub>2</sub> using ORCA 5.0 at the B3LYP/6-31G (d, p) and B3LYP/DEF2SVP level using a quantum mechanism package.

\_\_\_\_\_ THPIS+O<sub>2</sub> GIBBS FREE ENERGY \_\_\_\_\_ The Gibbs free energy is  $G = H - T^*S$ Total enthalpy ... -3182.55043554 Eh Total entropy correction ... -0.08393215 Eh -52.67 kcal/mol Final Gibbs free energy ... -3182.63436769 Eh For completeness - the Gibbs free energy minus the electronic energy G-E(el) ... 0.48055338 Eh 301.55 kcal/mol Timings for individual modules: Sum of individual times  $\dots$  5309.271 sec (= 88.488 min) GTO integral calculation ... 3.579 sec (= 0.060 min) 0.1 % SCF iterations ... 2394.155 sec (= 39.903 min) 45.1 % Analytical frequency calculation... 2911.537 sec (= 48.526 min) 54.8 %

**Table S24** Calculated Gibbs free energy of **APIS** using ORCA 5.0 at the B3LYP/6-31G (d,p) and B3LYP/DEF2SVP level using quantum mechanism package.

#### -----

## APIS

## GIBBS FREE ENERGY

\_\_\_\_\_

The Gibbs free energy is G = H - T\*S Total enthalpy ... -3185.87795057 Eh Total entropy correction ... -0.08737072 Eh -54.83 kcal/mol

Final Gibbs free energy ... -3185.96532129 Eh

For completeness - the Gibbs free energy minus the electronic energy

G-E(el) ... 0.52321755 Eh 328.32 kcal/mol

Timings for individual modules:

Sum of individual times  $\dots$  4288.483 sec (= 71.475 min)

GTO integral calculation ... 2.835 sec (= 0.047 min) 0.1 %

SCF iterations ... 288.080 sec (= 4.801 min) 6.7 %

Analytical frequency calculation... 3997.568 sec (= 66.626 min) 93.2 %

**Table S25** Calculated Gibbs free energy of **APIS** and **O**<sub>2</sub> using ORCA 5.0 at the B3LYP/6-31G (d, p) and B3LYP/DEF2SVP level using a quantum mechanism package.

APIS+O<sub>2</sub>

## GIBBS FREE ENERGY

\_\_\_\_\_

The Gibbs free energy is G = H - T\*STotal enthalpy ... -3335.95158265 Eh Total entropy correction ... -0.09084651 Eh -57.01 kcal/mol -------Final Gibbs free energy ... -3336.04242916 Eh For completeness - the Gibbs free energy minus the electronic energy G-E(el) ... 0.52511206 Eh 329.51 kcal/mol Timings for individual modules: Sum of individual times ... 7444.600 sec (= 124.077 min) GTO integral calculation ... 73.874 sec (= 1.231 min) 1.0 % SCF iterations ... 2387.693 sec (= 39.795 min) 32.1 % Analytical frequency calculation... 4983.033 sec (= 83.051 min) 66.9 %



**Fig. S17**  $O_2$ <sup>--</sup> production of three **RPNI-S** using the reduction product of XTT as the indicator. (a) UV-vis spectra changes of XTT alone, and (b-d) UV-vis spectra changes of XTT in the presence of (b) **PPIS**, (c) **THPIS**, and (d) **APIS** PSs, respectively, under the white light irradiation with different time. [Inset: (a') represents the partial magnified parts of Fig. a].



**Fig. S18** O<sub>2</sub><sup>•-</sup> generation of three **RPNI-S** PSs using the DHR 123 indicator. PL spectra of DHR 123 in 99% PBS in DMSO,  $f_{PBS}$  in the presence of (a) **PPIS**, (b) **THPIS**, and (c) **APIS**, respectively, after exposure to white light irradiation at different times. [**RPNI-S**] =100  $\mu$ M, [DHR 123] = 10  $\mu$ M].



**Fig. S19** (a-c) Cellular uptake data of the (a) **PPIS**, (b) **THPIS**, and (c) **APIS** in Hela cell. [concentration of **PPIS**, **THPIS**, and **APIS**: 20  $\mu$ M]. (d-f) Cellular uptake data of the (d) **PPIS**, (e) **THPIS**, and (f) **APIS** in HEK 293T. [concentration of **PPIS**, **THPIS**, and **APIS**: 100  $\mu$ M].



**Fig. S20** (a-c) Cellular uptake data of the (a) **PPI**, (b) **THPI**, and (c) **API** in Hela cell. [concentration of **PPI**, **THPI**, and **API**: 20  $\mu$ M]. (d-f) Cellular uptake data of the (d) **PPI**, (e) **THPI**, and (f) **API** in Hela cell. [concentration of **PPI**, **THPI**, and **API**: 100  $\mu$ M]. (g-i) Cellular uptake data of the (g) **PPI**, (h) **THPI**, and (i) **API** in HEK293. [concentration of **PPI**, **THPI**, and **API**: 100  $\mu$ M].



**Fig. S21** (a-c) Cell viability of HeLa cells after treatment with different concentrations of (a) **PPI**, (b) **THPI**, and (c) **API**, respectively, along with white light irradiation. (d-f) Cell cytotoxicity of HEK293Tcells after treatment with different concentrations of (d) **PPI**, (e) **THPI**, and (f) **API**, respectively, along with white light irradiation.



**Fig. S22** Fluorescence microscope images of HeLa cells after treatment with **APIS** under normoxia and loaded with calcein-AM (2  $\mu$ M, live cell marker). [Concentration of **APIS**: (50  $\mu$ M, scale bar: 100  $\mu$ m].



**Fig. S23** Fluorescence microscope images of HeLa cells after treatment with **APIS** under normoxia and loaded with PI (4  $\mu$ M, dead cell marker). [Concentration of **APIS**: (50  $\mu$ M, scale bar: 100  $\mu$ m].



**Fig. S24** Fluorescence microscope images of HeLa cells after treatment with **APIS** under normoxia and loaded with DCFDA (10  $\mu$ M, ROS detection probe). [Concentration of **APIS**: (50  $\mu$ M, scale bar: 100  $\mu$ m].

#### 2. Supporting figures



Fig. S25 <sup>1</sup>H NMR spectra of PNI-O.



Fig. S26<sup>13</sup>C NMR spectra of PNI-O.



Fig. S27 Maldi-TOF spectra of PNI-O.



Fig. S28 <sup>1</sup>H NMR spectra of BrPNI-O.



Fig. S29 <sup>13</sup>C NMR spectra of BrPNI-O.



Fig. S30 Maldi-TOF spectra of BrPNI-O.



Fig. S31 <sup>1</sup>H NMR spectra of PPI.



Fig. S32 <sup>13</sup>C NMR spectra of PPI.



Fig. S33 Maldi-TOF spectra of PPI.



Fig. S34 <sup>1</sup>H NMR spectra of THPI.



Fig. S35 <sup>13</sup>C NMR spectra of THPI.



Fig. S36 Maldi-TOF spectra of THPI.



Fig. S37 <sup>1</sup>H NMR spectra of API.



Fig. S38 <sup>13</sup>C NMR spectra of API.


Fig. S39 Maldi-TOF spectra of API.



Fig. S40 <sup>1</sup>H NMR spectra of PPIS.



Fig. S41 <sup>13</sup>C NMR spectra of PPIS.



Fig. S42 Maldi-TOF spectra of PPIS.



Fig. S43 <sup>1</sup>H NMR spectra of THPIS.



Fig. S44 <sup>13</sup>C NMR spectra of THPIS.



Fig. S45 Maldi-TOF spectra of THPIS.



Fig. S46 <sup>1</sup>H NMR spectra of APIS.



Fig. S47 <sup>13</sup>C NMR spectra of APIS.



Fig. S48 Maldi-TOF spectra of APIS.

## 3. References

- 1 A. D. Becke, J. Chem. Phys. 1993, 98, 5648-5652.
- 2 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman, and D. J. Fox, Gaussian, Inc., Wallingford CT, 2016.
- 3 F. Neese, WIREs Comp. Mol. Sci., 2018, 8, 1327.
- 4 F. Neese, WIREs Comp. Mol. Sci., 2022, 12, 1606.
- 5 D. Wang, M. M. S. Lee, G. Shan, R. T. K. Kwok, J. W. Y. Lam, H. Su, Y. Cai and B. Z. Tang, Adv. Mater., 2018, 30, 1802105.

- 6 W. Wu, D. Mao, S. Xu, M. Panahandeh-Fard, Y. Duan, F. Hu, D. Kong and B. Liu, *Adv. Funct. Mater.*, 2019, **29**, 1901791.
- 7 X. Shi, S. H. P. Sung, J. H. C. Chau, Y. Li, Z. Liu, R. T. K. Kwok, J. Liu, P. Xiao, J. Zhang, B. Liu, J. W. Y. Lam and B. Z. Tang, *Small Methods*, 2020, **4**, 2000046.
- 8 Y. Li, W. Zhang, J. Niu and Y. Chen, ACS Nano, 2012, 6, 5164-5173.
- 9 Y. Xiong, Z. Zhao, W. Zhao, H. Ma, Q. Peng, Z. He, X. Zhang, Y. Chen, X. He, J. W. Y. Lam and B. Z. Tang, *Angew. Chem., Int. Ed.*, 2018, **57**, 7997-8001.
- 10 Z. Liu, H. Zou, Z. Zhao, P. Zhang, G.-G. Shan, R. T. K. Kwok, J. W. Y. Lam, L. Zheng and B. Z. Tang, *ACS Nano*, 2019, **13**, 11283-11293.
- 11 Z. Yang, Z. Zhang, Z. Lei, D. Wang, H. Ma and B. Z. Tang, ACS Nano, 2021, 15, 7328-7339.
- 12 X. Wang, Y. Song, G. Pan, W. Han, B. Wang, L. Cui, H. Ma, Z. An, Z. Xie, B. Xu and W. Tian, *Chem. Sci.*, 2020, **11**, 10921-10927.
- 13 Z. Wang, L. Ma, H. Zhao, Y. Wan, X.-F. Zhang, Y. Li, Z. Kuang and A. Xia, *Phys. Chem. Chem. Phys.*, 2023, **25**, 24386-24394.
- 14 K. Schmidt, S. Brovelli, V. Coropceanu, D. Beljonne, J. Cornil, C. Bazzini, T. Caronna, R. Tubino, F. Meinardi, Z. Shuai and J.-L. Brédas, J. Phys. Chem. A, 2007, **111**, 10490-10499.
- 15 D. Barman and P. K. Iyer, J. Phys. Chem. C, 2023, 127, 2694-2704.