

SUPPORTING INFORMATION

Synergistically Activating Nucleophile Strategy Enabled Organocatalytic Asymmetric P-Addition of Cyclic Imines

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1. General information

All the starting materials were obtained from commercial sources and used without further purification unless otherwise stated. ^1H and ^{13}C NMR spectra were recorded on a Bruker AVANCE III HD (400 MHz) spectrometer in CDCl_3 . Chemical shifts (δ) are reported in ppm, and the residual solvent peak was used as an internal reference CDCl_3 [$\delta(^1\text{H}) = 7.26$ ppm, $\delta(^{13}\text{C}) = 77.0$ ppm]. Multiplicity was indicated as follows: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), dd (doublet of doublet), br s (broad singlet). Coupling constants (J) were reported in Hertz (Hz). CD spectra were acquired using a J-1500 CD spectrometer and an Applied Photophysics Chirascan spectrometer. All high resolution mass spectra were obtained on a Thermo LTQ mass spectrometer. For thin layer chromatography (TLC), Merck pre-coated TLC plates (Merck 60 F254) were used, and compounds were visualized with a UV light at 254 nm. Flash chromatographic separations were performed on Merck 60 (0.040-0.063 mm) mesh silica gel. Enantiomeric excesses were determined by HPLC analysis using chiral column described below in detail. Optical rotations were measured with polarimeter.

All the cyclic imines **1/1'**, **6** listed in Figure S2 were synthesized following general procedures D,^[4] E,^[5] and F.^[6] Phosphine oxides **2a**, **2g**, **3a**, **3b**, **3c** and **3d** were obtained from commercial sources, and other phosphine oxides **2** were synthesized following the previous reported literature and listed in Figure S3.^[3] All the phosphonium salt catalysts used in this study were prepared via a **P**-alkylation reaction of our previously reported organophosphines according to the known procedures.^[1] The structure and absolute configurations of products were assigned by optical rotation analysis of **5'b** and **5'j** (Figure S4), and CD analysis of compound **4a**, **5a**, **7a** and **7g** were tested and calculated (Figure S5-8).

2. Optimizations of reaction conditions

2.1 Optimization for asymmetric P-nucleophile addition to cyclic N-sulfonyl imines **1a** with secondary phosphine oxides **2a**

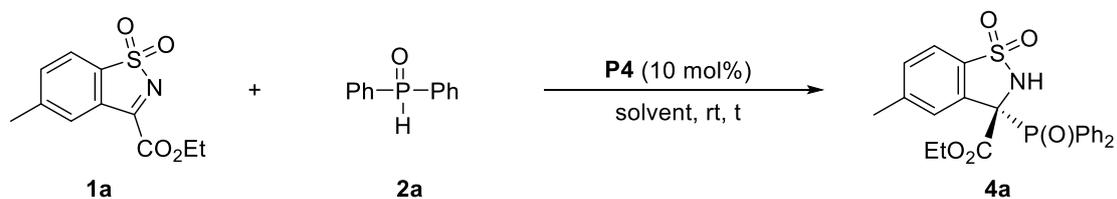
Table S1. The asymmetric P-nucleophile addition to cyclic ketimine **1a** with diphenylphosphine oxide **2a** catalyzed by different chiral phosphonium salts in DCE.^a

| entry | cat. | base(x equiv.) | t (h) | yield (%) ^b | ee (%) ^c |
|----------|-----------|--------------------------------------|----------|------------------------|---------------------|
| 1 | P0 | K ₂ CO ₃ (1.0) | 1 | 98 | 0 |
| 2 | - | - | 2 | 83 | 0 |
| 3 | P1 | - | 2 | 90 | 34 |
| 4 | P2 | - | 2 | 92 | 53 |
| 5 | P3 | - | 2 | 90 | 71 |
| 6 | P4 | - | 2 | 93 | 90 |
| 7 | P5 | - | 2 | 86 | 68 |
| 8 | P6 | - | 2 | 92 | 86 |
| 9 | P7 | - | 2 | 87 | 83 |

P4: R = ⁱPr; **P5:** R = Me
P6: R = ^tBu

^aReactions were performed with **1a** (0.1 mmol), **2a** (0.12 mmol), catalyst (10 mol%) in solvent (1.0 mL) at room temperature. ^bIsolated yield. ^cThe ee value was determined by HPLC analysis on a chiral stationary phase. Ts = 4-toluenesulfonyl. DCE = 1,2-dichloroethane.

Table S2. The asymmetric P-nucleophile addition to cyclic ketimine **1a** with diphenylphosphine oxide **2a** catalyzed by **P4**: screening solvents and other conditions.^a

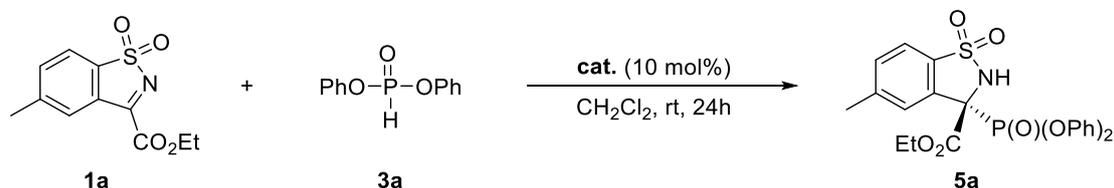


| entry | solvent | base(x equiv.) | t (h) | yield (%) ^b | ee (%) ^c |
|----------------|---------------------------------|--------------------------------------|----------|------------------------|---------------------|
| 1 | CH ₂ Cl ₂ | - | 2 | 93 | 90 |
| 2 | CHCl ₃ | - | 2 | 94 | 73 |
| 3 | Et ₂ O | - | 2 | 89 | 88 |
| 4 | toluene | - | 2 | 88 | 93 |
| 5 | hexane | - | 4 | 90 | 75 |
| 6 | DCE | - | 2 | 95 | >99 |
| 7 ^d | DCE | K ₂ CO ₃ (1.0) | 0.5 | 92 | <5 |
| 8 ^e | DCE | K ₂ CO ₃ (0.5) | 1 | 96 | 75 |
| 9 ^f | DCE | - | 2.5 | 92 | 89 |

^aReactions were performed with **1a** (0.1 mmol), **2a** (0.12 mmol), **P4** (10 mol%) in solvent (1.0 mL) at room temperature. ^bIsolated yield. ^cThe ee value was determined by HPLC analysis on a chiral stationary phase. ^d1.0 equivalent of K₂CO₃ was added. ^e0.5 equivalent of K₂CO₃ was added. ^fThe catalyst loading was 5 mol%. Ts = 4-toluenesulfonyl. c

2.2 Optimization for asymmetric P-nucleophile addition to cyclic N-sulfonyl imines **1a** with secondary phosphine oxides **3a**

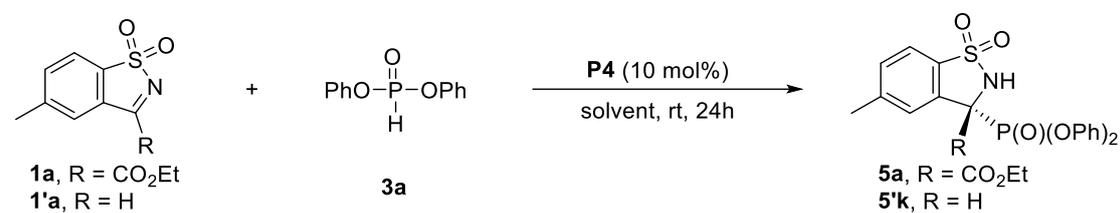
Table S3. The asymmetric P-nucleophile addition to cyclic ketimine **1a** with (PhO)₂P(O)H **3a** catalyzed by different chiral phosphonium salts in CH₂Cl₂. ^a



| entry | cat. | yield (%) ^b | ee (%) ^c |
|----------|-----------|------------------------|---------------------|
| 1 | P1 | 90 | 2 |
| 2 | P2 | 88 | 85 |
| 3 | P3 | 91 | 62 |
| 4 | P4 | 92 | 88 |
| 5 | P5 | 90 | 70 |
| 6 | P6 | 89 | 80 |
| 7 | P7 | 92 | 38 |

^aReactions were performed with **1a** (0.1 mmol), **3a** (0.12 mmol), catalyst (10 mol%) in CH₂Cl₂ (2.0 mL) at room temperature. ^bIsolated yield. ^cDetermined by HPLC analysis on a chiral stationary phase.

Table S4. The asymmetric **P**-nucleophile addition to cyclic ketimine **1a** with (PhO)₂P(O)H **3a** catalyzed by **P4**: screening of the solvents, and catalyst Loading. ^a

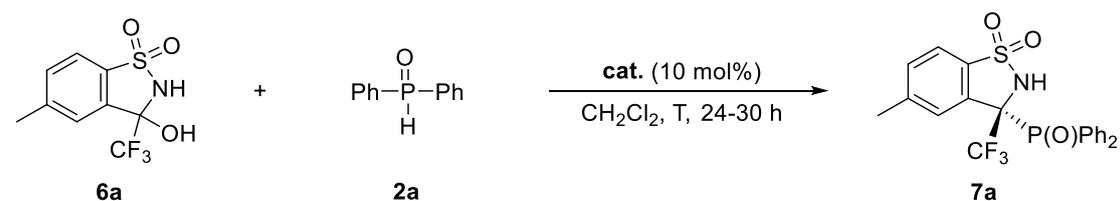


| entry | ketimine | solvent | t (h) | yield (%) ^b | ee (%) ^c |
|-------------------|------------|---------------------------------|-----------|------------------------|---------------------|
| 1 | 1a | CH ₂ Cl ₂ | 24 | 92 | 88 |
| 2 | 1a | CHCl ₃ | 24 | 92 | 73 |
| 3 | 1a | DCE | 24 | 95 | 91 |
| 4 | 1a | hexane | 48 | 91 | 95 |
| 5 | 1a | toluene | 24 | 96 | >99 |
| 6 ^d | 1a | toluene | 24 | 93 | 90 |
| 7 | 1'a | toluene | - | - | - |
| 8 ^e | 1'a | toluene | 54 | 91 | 88 |
| 9 ^e | 1'a | DCE | 48 | 92 | 90 |
| 10 ^e | 1'a | CH ₂ Cl ₂ | 48 | 95 | 98 |
| 11 ^{d,e} | 1'a | CH ₂ Cl ₂ | 48 | 94 | 89 |

^aReactions were performed with **1a** (0.1 mmol), **3a** (0.12 mmol), **P4** (10 mol%) in solvent (2.0 mL) at room temperature. ^bIsolated yield. ^cDetermined by HPLC analysis on a chiral stationary phase. ^dWith 5 mol% catalyst **P4**. ^e2.0 equiv. of Cs₂CO₃ was added.

3.1 Optimization of reaction conditions for the asymmetric **P**-nucleophile addition to CF₃-substituted cyclic N-sulfonyl amines **6a** with secondary phosphine oxides **2a**

Table S5. The asymmetric **P**-nucleophile addition to cyclic ketimine **6a** with diphenylphosphine oxide **2a**: screening different chiral phosphonium salts and other conditions. ^a



| entry | cat. | T (°C) | t (h) | yield (%) ^b | ee (%) ^c |
|-------|------|--------|-------|------------------------|---------------------|
|-------|------|--------|-------|------------------------|---------------------|

| | | | | | |
|----------------------|-----------|------------|-----------|-----------|-----------|
| 1 | P1 | -30 | 24 | 90 | 4 |
| 2 | P2 | -30 | 24 | 93 | 4 |
| 3 | P3 | -30 | 24 | 89 | 24 |
| 4 | P4 | -30 | 24 | 92 | 71 |
| 5 | P5 | -30 | 24 | 92 | 56 |
| 6 | P6 | -30 | 24 | 90 | 0 |
| 7 | P7 | -30 | 24 | 92 | 23 |
| 8 | P4 | -50 | 24 | 90 | 63 |
| 9^d | P4 | -30 | 30 | 93 | 94 |
| 10 ^d | P4 | -78 | 48 | 92 | 92 |
| 11 ^{d,e} | P4 | -30 | 48 | 92 | 28 |

^aReactions were performed with **6a** (0.1 mmol), **2a** (0.12 mmol), catalyst (10 mol%) in CH₂Cl₂ (1.0 mL). ^bIsolated yield. ^cDetermined by HPLC analysis on a chiral stationary phase. ^d3.0 mL CH₂Cl₂ was used. ^eWith 5 mol% catalyst **P4**.

3. Preparation of catalysts.

All the catalysts listed in Figure S1 were synthesized following general procedures A^[1], B and C^[2]. The catalysts **P1-P4** are known compounds, and their characterization data were in agreement with those reported in the literature.^[1,2] Unknown compounds **P5- P7**, **P4-1**, **P4-2**, **P4-3** and **P4-4** were fully characterized.

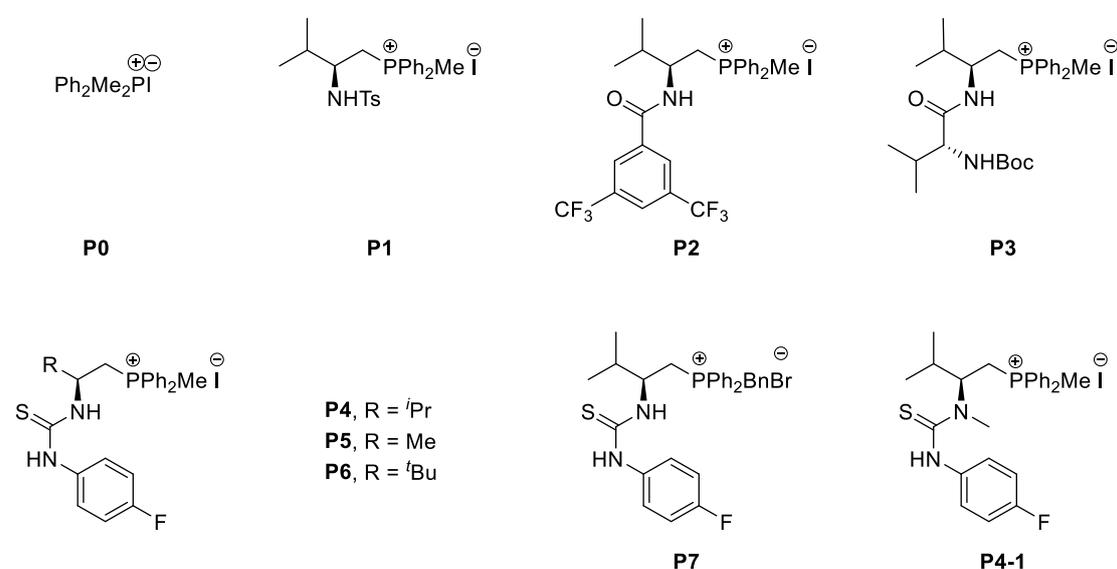
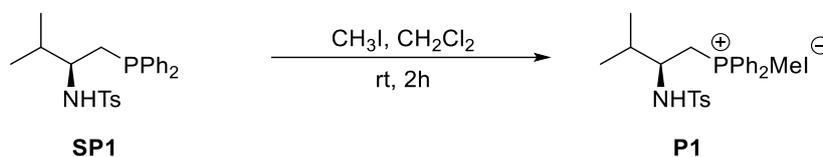


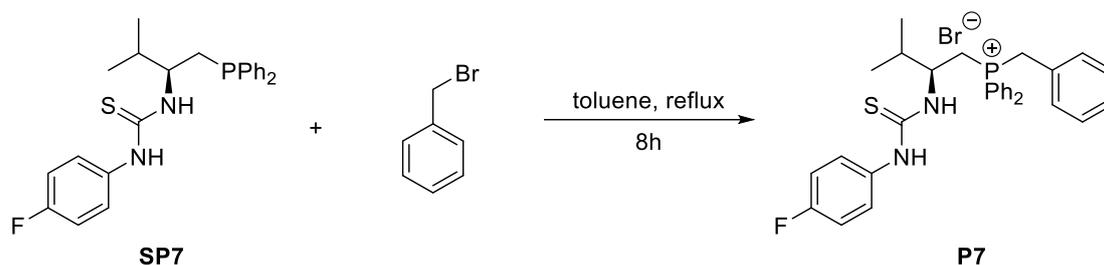
Figure S1. Phosphonium salt catalysts in this study.

(1) General procedure A: preparation of phosphonium salt P1



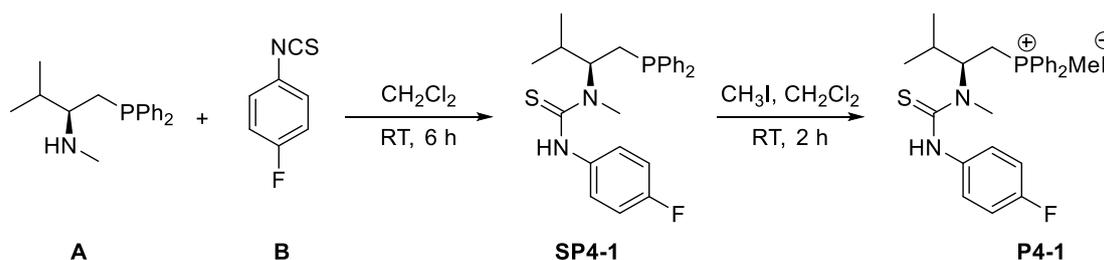
General procedure A: To a solution of phosphine **SP1** (1 mmol, 425 mg) in CH_2Cl_2 (10 mL) was slowly added the methyl iodide solution (4 mmol, 568 mg, 2.0 M in CH_2Cl_2). Then the mixture was allowed to stir at room temperature for 2 h. The reaction crude mixture was directly purified by flash chromatography dichloromethane/methanol = 20/1 to afford the desired chiral phosphonium salt **P1** as a yellow solid (93% yield). The phosphonium salts **P2-P6** were prepared from the above procedure by using the corresponding phosphines as reactants.^[1]

(2) General procedure B: preparation of phosphonium salt P7



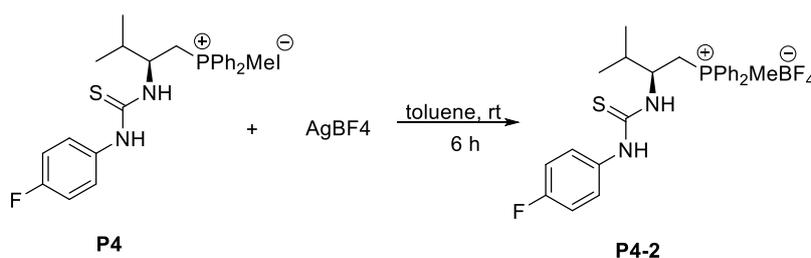
General procedure B: To a solution of phosphine **SP7** (1 mmol, 424 mg) in toluene was slowly added the benzyl bromide solution (1.2 mmol, 204 mg, 2.0 M in toluene). Then, the mixture was refluxed in toluene for 12 h. The reaction crude mixture was directly purified by flash chromatography dichloromethane/methanol = 20/1 to afford the desired chiral phosphonium salt **P7** as a white solid (95% yield).

(3) General procedure C: preparation of phosphonium salts P4-1



General procedure C: To a solution of compound **A** (0.4 mmol, 114 mg) in anhydrous CH₂Cl₂ was cooled to 0 °C, and a solution of **B** (0.48 mmol, 73.4 mg) in CH₂Cl₂ was added dropwise.^[2] Then, the reaction mixture was stirred at room temperature for 6 hours. The reaction crude mixture was directly purified by flash chromatography (petroleum ether/ethyl acetate = 5/1) to afford **SP4-1** (95% yield) as a colorless solid. To a solution of phosphine **SP4-1** (1.0 mmol, 438mg) in CH₂Cl₂ was slowly added the methyl iodide solution (4.0 mmol, 568 mg, 2.0 M in CH₂Cl₂). Then, the mixture was allowed to stir at room temperature for 24 h. The reaction crude mixture was directly purified by flash chromatography dichloromethane/methanol = 20/1 to afford the desired chiral phosphonium salt **P4-1** as a yellow solid (70% yield) was fully characterized.

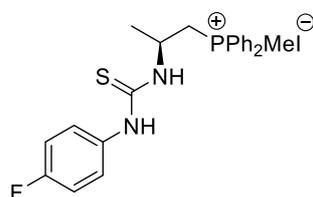
(4) General procedure C: preparation of phosphonium salts **P4-2**



To a solution of chiral phosphonium salt **P4** (0.1 mmol) in anhydrous toluene was quickly added AgBF₄ (0.3 mmol). Then, the mixture was allowed to stir at room temperature for 3 h. The reaction crude mixture was directly purified by flash chromatography dichloromethane/methanol = 20/1 to afford the desired chiral phosphonium salt **P4-2** as a yellow solid (85% yield). chiral phosphonium salt **P4-2**, **P4-3** and **P4-4** were fully characterized.

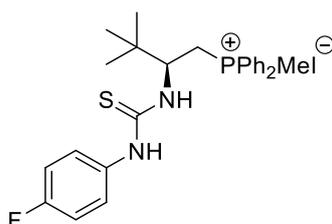
Characterization of the unknown phosphonium salts

(S)-2-(3-(4-fluorophenyl)thioureido)propyl(methyl)diphenylphosphonium iodide (P5)



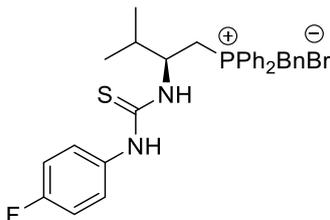
A yellow solid; ^1H NMR (400 MHz, CDCl_3) δ 8.79 (s, 1H), 8.71 (d, $J = 8.8$ Hz, 1H), 7.89 - 7.81 (m, 2H), 7.81 - 7.69 (m, 4H), 7.70 - 7.61 (m, 4H), 7.44 - 7.31 (m, 2H), 7.01 - 6.87 (m, 2H), 5.33 - 5.19 (m, 1H), 3.79 (dt, $J = 15.5, 11.0$ Hz, 1H), 3.11 - 2.97 (m, 1H), 2.85 (d, $J = 13.9$ Hz, 3H), 1.48 (dd, $J = 6.6, 2.1$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 180.34, 160.12 (d, $J = 242.9$ Hz), 135.14 (d, $J = 2.9$ Hz), 134.88 (d, $J = 2.9$ Hz), 134.59 (d, $J = 2.5$ Hz), 132.39 (d, $J = 10.2$ Hz), 132.25 (d, $J = 10.0$ Hz), 130.52 (d, $J = 6.4$ Hz), 130.40 (d, $J = 6.6$ Hz), 126.11 (d, $J = 8.0$ Hz), 119.50 (d, $J = 8.4$ Hz), 118.65 (d, $J = 8.7$ Hz), 115.15 (d, $J = 22.4$ Hz), 45.22 (d, $J = 4.1$ Hz), 30.94 (d, $J = 51.5$ Hz), 23.64 (d, $J = 14.0$ Hz), 8.62 (d, $J = 53.7$ Hz); ^{31}P NMR (162 MHz, CDCl_3) δ 20.94; HRMS (ESI) m/z calcd for $\text{C}_{23}\text{H}_{25}\text{N}_2\text{PSFI}$ $[\text{M}-\text{I}]^+ = 411.1460$, found = 411.1466.

(S)-(2-(3-(4-fluorophenyl)thioureido)-3,3-dimethylbutyl)(methyl)diphenylphosphonium iodide (P6)



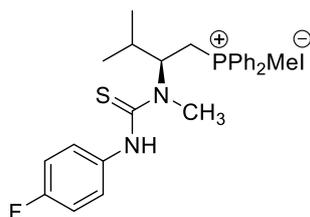
A yellow solid; ^1H NMR (400 MHz, CDCl_3) δ 9.21 (s, 1H), 8.88 (d, $J = 9.8$ Hz, 1H), 7.90 - 7.79 (m, 3H), 7.80 - 7.67 (m, 5H), 7.67 - 7.59 (m, 2H), 7.56 - 7.48 (m, 2H), 7.03 - 6.90 (m, 2H), 4.95 - 4.85 (m, 1H), 3.95 - 3.77 (m, 1H), 2.89 (d, $J = 14.1$ Hz, 3H), 2.75 (t, $J = 14.8$ Hz, 1H), 1.01 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 181.84, 160.21 (d, $J = 242.7$ Hz), 135.52 (d, $J = 3.0$ Hz), 135.04 (d, $J = 2.6$ Hz), 134.88 (d, $J = 3.0$ Hz), 132.55 (d, $J = 9.5$ Hz), 132.24 (d, $J = 10.3$ Hz), 130.61 (d, $J = 7.7$ Hz), 130.48 (d, $J = 8.0$ Hz), 126.18 (d, $J = 8.1$ Hz), 121.20, 120.35, 118.14, 117.28, 115.18 (d, $J = 22.4$ Hz), 56.29 (d, $J = 4.5$ Hz), 37.36 (d, $J = 11.7$ Hz), 26.32, 26.15 (d, $J = 53.4$ Hz), 8.09 (d, $J = 53.0$ Hz); ^{31}P NMR (162 MHz, CDCl_3) δ 24.13; HRMS (ESI) m/z calcd for $\text{C}_{26}\text{H}_{31}\text{N}_2\text{PSFI}$ $[\text{M}-\text{I}]^+ = 453.1930$, found = 453.1924.

(S)-benzyl(2-(3-(4-fluorophenyl)thioureido)-3-methylbutyl)diphenylphosphonium bromide (P7)



A white solid; ^1H NMR (400 MHz, CDCl_3) δ 9.29 (s, 1H), 9.20 (d, $J = 9.4$ Hz, 1H), 7.78 - 7.68 (m, 3H), 7.65 - 7.49 (m, 7H), 7.42 - 7.34 (m, 2H), 7.22 - 7.18 (m, 1H), 7.13 (t, $J = 7.6$ Hz, 2H), 6.93 - 6.82 (m, 4H), 5.25 - 5.08 (m, 1H), 4.88 - 4.66 (m, 2H), 3.55 - 3.42 (m, 1H), 2.69 - 2.56 (m, 1H), 2.04 - 1.92 (m, 1H), 0.94 (t, $J = 6.3$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 181.06, 159.71 (d, $J = 242.3$ Hz), 135.13 (d, $J = 2.7$ Hz), 135.03 (d, $J = 2.7$ Hz), 134.62 (d, $J = 2.7$ Hz), 133.36 (d, $J = 2.7$ Hz), 133.27 (d, $J = 2.5$ Hz), 130.38 (d, $J = 5.7$ Hz), 130.11 (d, $J = 9.8$ Hz), 129.99 (d, $J = 10.1$ Hz), 129.23 (d, $J = 3.1$ Hz), 129.02, 128.73 (d, $J = 3.6$ Hz), 128.21, 126.89 (d, $J = 8.5$ Hz), 125.48 (d, $J = 8.0$ Hz), 118.68, 117.86, 117.52, 116.69, 114.76 (d, $J = 22.3$ Hz), 52.66 (d, $J = 4.3$ Hz), 34.78 (d, $J = 12.6$ Hz), 28.91 (d, $J = 44.8$ Hz), 25.38 (d, $J = 51.5$ Hz), 18.41 (d, $J = 18.4$ Hz); ^{31}P NMR (162 MHz, CDCl_3) δ 24.29; HRMS (ESI) m/z calcd for $\text{C}_{31}\text{H}_{33}\text{N}_2\text{PSFBr}$ $[\text{M}-\text{Br}]^+ = 515.2081$, found = 515.2080.

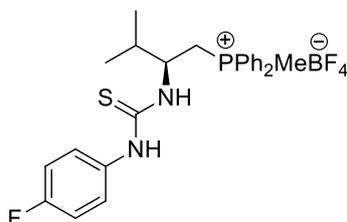
(S)-2-(2-(3-(4-fluorophenyl)-1-methylthioureido)propyl)(methyl)diphenylphosphonium iodide (P4-1)



A yellow solid; ^1H NMR (400 MHz, CDCl_3) δ 8.11 - 7.88 (m, 4H), 7.78 - 7.59 (m, 6H), 7.33 - 7.29 (m, 1H), 7.08 - 6.88 (m, 2H), 6.75 - 6.61 (m, 1H), 5.85 (dd, $J = 22.0, 6.8$ Hz, 1H), 4.49 - 4.25 (m, 1H), 3.22 (s, 1H), 2.92 (dd, $J = 22.0, 10.8$ Hz, 3H), 2.80 (s, 1H), 1.63 (s, 1H), 1.31 - 1.20 (m, 3H), 1.05 - 0.92 (m, 3H), 0.91 - 0.85 (m, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 183.05, 159.61, 135.44, 134.91, 132.67 (dd, $J = 13.4, 10.0$ Hz), 130.39, 130.26 (d, $J = 1.5$ Hz), 130.12, 128.60 (d, $J = 8.2$ Hz), 115.89, 115.26 (d, $J = 22.5$ Hz), 100.10, 60.51, 34.43, 32.79 (d, $J = 12.8$ Hz), 29.80, 19.67 (d, $J = 145.9$

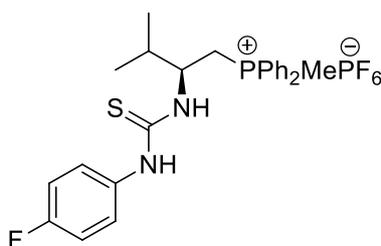
Hz), 15.95, 8.93 (d, $J = 53.5$ Hz); ^{31}P NMR (162 MHz, CDCl_3) δ 21.55; HRMS (ESI) m/z calcd for $\text{C}_{26}\text{H}_{31}\text{N}_2\text{PSFI}$ $[\text{M-I}]^+ = 453.1924$, found = 453.1920.

(S)-(2-(3-(4-fluorophenyl)thioureido)propyl)(methyl)diphenylphosphonium
tetrafluoroborate (P4-2)



A white solid; ^1H NMR (400 MHz, CD_3OD) δ 7.87 - 7.57 (m, 10 H), 7.56 - 7.51 (m, 1H), 7.29 (d, $J = 8.0$ Hz, 1H), 7.11 - 7.04 (m, 1H), 6.95 - 6.88 (m, 1H), 5.67 (d, $J = 10.0$ Hz, 1H), 3.30 - 3.21 (m, 2H), 3.11 - 3.00 (m, 1H), 2.72 - 2.65 (m, 3H), 2.36 (s, 1H), 1.03 (dd, $J = 6.8, 2.8$ Hz, 1H), 0.95 (d, $J = 6.8$ Hz, 3H), 0.65 (dd, $J = 11.6, 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CD_3OD) δ 160.97, 158.59, 156.81, 145.13, 140.16, 136.38 (d, $J = 2.8$ Hz), 135.90 (d, $J = 3.0$ Hz), 135.52 (dd, $J = 10.6, 3.1$ Hz), 135.30 (d, $J = 3.0$ Hz), 133.63 (d, $J = 9.8$ Hz), 133.40 - 132.91 (m), 127.70, 126.29 (d, $J = 8.5$ Hz), 123.10, 122.46 (d, $J = 45.6$ Hz), 121.80 (d, $J = 7.5$ Hz), 120.97 (dd, $J = 85.7, 45.0$ Hz), 117.16 (d, $J = 23.3$ Hz), 116.00 (d, $J = 22.5$ Hz), 55.60 (d, $J = 4.9$ Hz), 35.59 (d, $J = 13.2$ Hz), 33.98 (d, $J = 11.0$ Hz), 28.77 (d, $J = 53.3$ Hz), 25.90 (d, $J = 53.7$ Hz), 21.40, 17.94 (dd, $J = 223.9, 87.6$ Hz), 6.70 (dd, $J = 101.3, 54.8$ Hz); ^{31}P NMR (162 MHz, CDCl_3) δ 23.18; HRMS (ESI) m/z calcd for $\text{C}_{25}\text{H}_{29}\text{BN}_2\text{PSF}_5$ $[\text{M-BF}_4]^+ = 439.1768$, found = 439.1766.

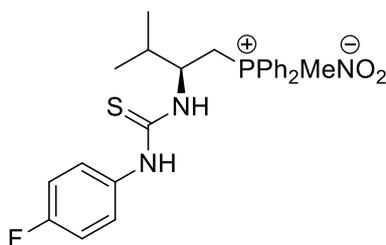
(S)-(2-(3-(4-fluorophenyl)thioureido)propyl)(methyl)diphenylphosphonium
hexafluorophosphate(V) (P4-3)



A white solid; ^1H NMR (400 MHz, CD_3OD) δ 7.89 - 7.75 (m, 6H), 7.74 - 7.64 (m, 4H), 7.62 - 7.57 (m, 1H), 7.36 - 7.30 (m, 1H), 7.14 - 6.93 (m, 2H), 4.82 (s, 3H), 3.34 -

3.29 (m, 1H), 3.10 - 2.97 (m, 1H), 2.83 (d, $J = 14.0$ Hz, 1H), 2.72 (dd, $J = 14.4, 3.6$ Hz, 2H), 2.40 (s, 1H), 1.05 - 0.97 (m, 3H), 0.69 (dd, $J = 10.8, 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CD_3OD) δ 145.18, 140.04, 135.92 (d, $J = 2.9$ Hz), 135.63 (dd, $J = 7.2, 4.0$ Hz), 133.55 (d, $J = 9.8$ Hz), 133.23 (dd, $J = 10.1, 6.3$ Hz), 131.47 - 130.79 (m), 128.10 (d, $J = 8.3$ Hz), 127.71, 122.10 (d, $J = 86.2$ Hz), 120.73 (d, $J = 86.1$ Hz), 116.68 (d, $J = 23.1$ Hz), 36.22 (d, $J = 13.2$ Hz), 33.96 (d, $J = 10.9$ Hz), 26.04 (d, $J = 53.3$ Hz), 21.42, 18.89 (d, $J = 78.3$ Hz), 17.15 (d, $J = 128.6$ Hz), 7.17 (dd, $J = 54.9, 9.8$ Hz); ^{31}P NMR (162 MHz, CDCl_3) δ 23.13; HRMS (ESI) m/z calcd for $\text{C}_{25}\text{H}_{29}\text{N}_2\text{P}_2\text{SF}_7$ $[\text{M}-\text{BF}_6]^+$ = 439.1768, found = 439.1768.

(S)-1-(4-fluorophenyl)-3-(1-(methyl(nitro)diphenylphosphoranylethyl)thiourea (P4-4)



A white solid; ^1H NMR (400 MHz, CDCl_3) δ 7.90 - 7.75 (m, 6H), 7.70 - 7.60 (m, 6H), 7.49 - 7.46 (m, 1H), 6.95 - 6.91 (m, 1H), 5.08 - 4.87 (m, 1H), 3.83 - 3.57 (m, 1H), 3.01 (d, $J = 14.0$ Hz, 1H), 2.91 - 2.81 (m, 3H), 2.37 (s, 1H), 2.09 - 2.00 (m, 1H), 1.249 - 1.19 (m, 1H), 0.99 (dd, $J = 10.8, 6.8$ Hz, 6H), 0.78 (d, $J = 6.8$ Hz, 1H), 0.65 (d, $J = 6.8$ Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 181.34, 160.06 (d, $J = 242.5$ Hz), 143.33, 139.27, 135.27 (dd, $J = 40.1, 3.0$ Hz), 135.07 (dd, $J = 47.9, 2.9$ Hz), 132.71 - 132.08 (m), 130.87 - 130.07 (m), 129.72, 126.47, 126.00 (d, $J = 8.0$ Hz), 121.51 (d, $J = 85.7$ Hz), 119.22 (dd, $J = 166.6, 85.5$ Hz), 117.36 (d, $J = 84.5$ Hz), 115.07 (d, $J = 22.4$ Hz), 53.15 (d, $J = 4.5$ Hz), 34.69 (d, $J = 12.8$ Hz), 32.90 (d, $J = 12.2$ Hz), 26.92 (d, $J = 53.1$ Hz), 20.51 (d, $J = 224.6$ Hz), 18.42, 8.86 (dd, $J = 117.1, 53.6$ Hz); ^{31}P NMR (162 MHz, CDCl_3) δ 23.16; HRMS (ESI) m/z calcd for $\text{C}_{25}\text{H}_{29}\text{N}_3\text{O}_2\text{PSF}$ $[\text{M}-\text{NO}_2]^+$ = 439.1768, found = 439.1760.

4. Preparation of cyclic imines **1/1'** and **6** and phosphine oxides **2**

All cyclic ketimines **1/1'** and **6** listed in Figure S2 were prepared following general procedures A^[4], B^[5] and C^[6]. All the phosphine oxides **2** were listed in Figure S3. Phosphine oxides **2a**, **2f**, **2g**, **3a**, **3b**, **3c** and **3d** were obtained from commercial sources, other phosphine oxides were synthesized following the previous reported literature. All the cyclic imines **1/1'** and **6** and phosphine oxides **2** were known compounds.

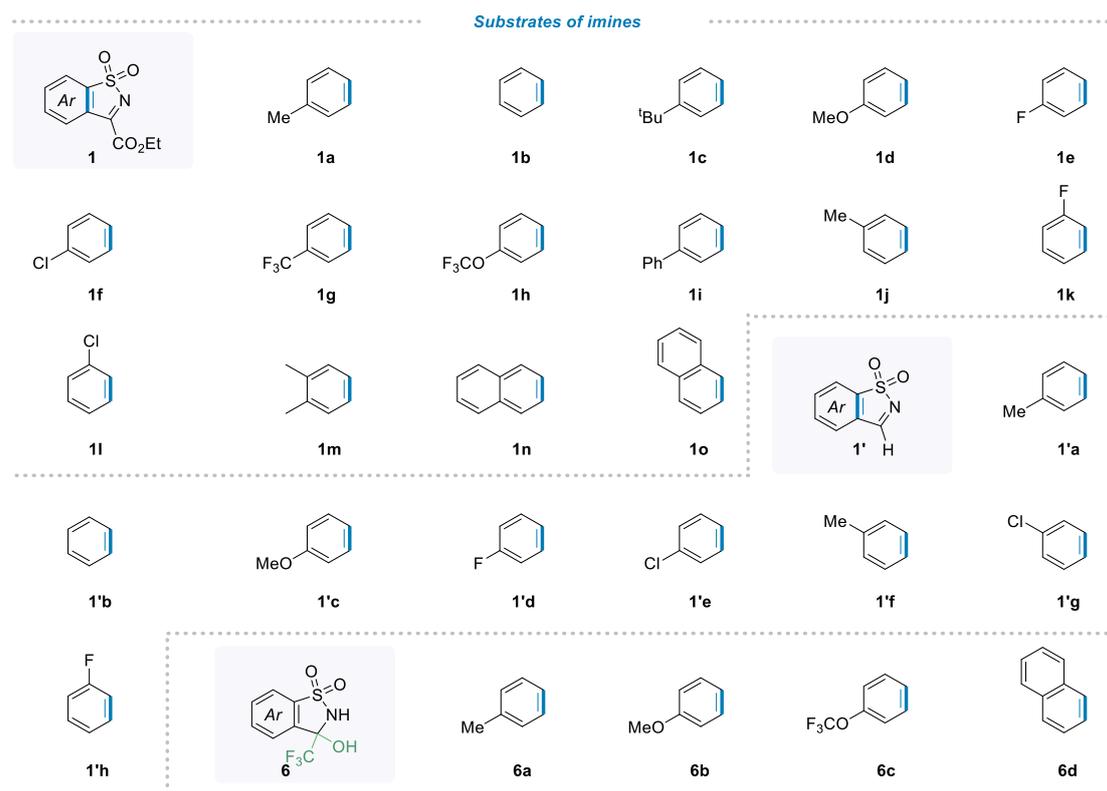


Figure S2. Substrates of cyclic imines **1/1'** and **6**

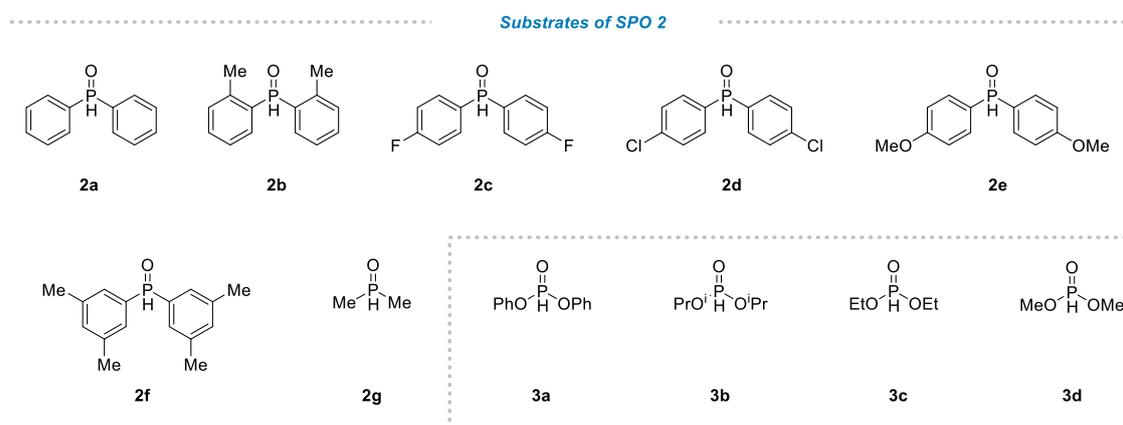
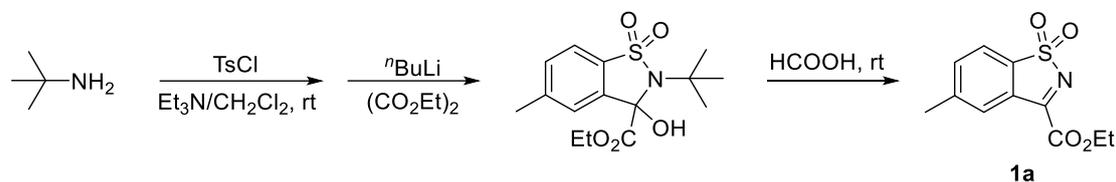


Figure S3. Substrates of phosphine oxides **2**.

4.1 Preparation of cyclic imines **1/1'** and **6**

(1) General procedure D: preparation of cyclic imines **1**

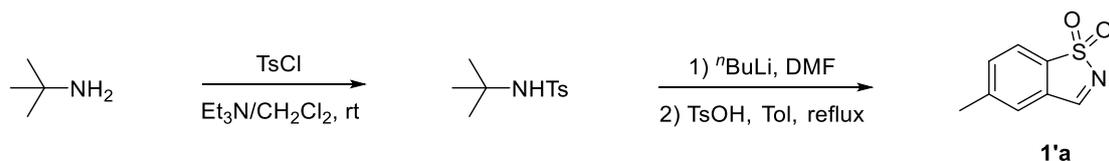


General procedure D: To a solution of tert-butylamine (30 mmol, 2.9 g) and triethylamine (40 mmol, 4.1 g) in DCM in an ice bath was added arylsulfonyl chloride (20 mmol, 3.8 g) dropwise. The mixture was stirred at room temperature overnight. It was washed with saturated sodium carbonate and brine. The organic layer was separated, and the aqueous layer was extracted with DCM. The combined organic extracts were dried over anhydrous sodium sulfate. The solvent was evaporated in vacuo to give the aryl sulfonamide as a solid without further purification.

n-butyllithium (30.75 mmol, 12.3 mL, 2.5 M in hexane) was added dropwise over a 20 minute period to a cold (0 °C), mechanically stirred solution of the aryl sulfonamide (15 mmol) in anhydrous tetrahydrofuran (100 mL) under a dry nitrogen atmosphere. After stirring an additional 25 min at 0 °C a precipitate formed. The suspension was cooled further to -78 °C and diethyl oxalate (45 mmol, 6.6 g) was added. The cooling bath was removed and the suspension was stirred at ambient temperature for 2 h. The reaction was quenched with 5% HCl (40 mL) and added to water (200 mL). The organics were extracted with diethyl ether (200 mL). The diethyl ether phase was washed with brine (200 mL). The solvent was removed and the crude product was obtained used directly in the next step.

To the crude product obtained above, formic acid (25 mL) was added and the suspension was stirred at room temperature under a dry nitrogen atmosphere. After 5 min dissolution occurred. After 20 h the solution was concentrated and the resultant solid was dissolved in DCM and concentrated (three times) to remove traces of formic acid. This afforded the title compound as a white solid which was further purified by flash chromatography.

(2) General procedure E: preparation of cyclic imines **1'**

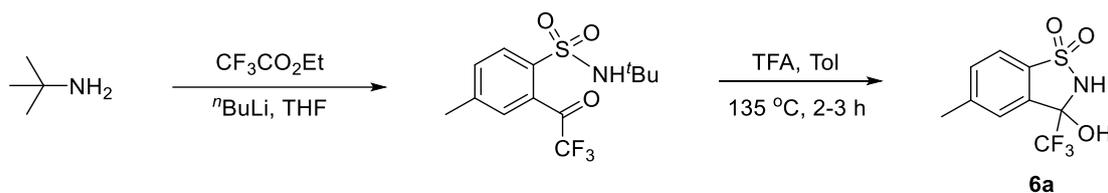


General procedure E: To a solution of tert-butylamine (30 mmol, 2.9 g) and triethylamine (40 mmol, 4.1 g) in CH_2Cl_2 in an ice bath was added arylsulfonyl chloride (20 mmol, 3.4 g) dropwise. The mixture was stirred at room temperature overnight. It was washed with saturated sodium carbonate and brine. The organic layer was separated, and the aqueous layer was extracted with CH_2Cl_2 . The combined organic extracts were dried over anhydrous sodium sulfate. The solvent was evaporated in vacuo to give the aryl sulfonamide as a solid without further purification.

The N-tert-butylbenzenesulfonamide (20 mmol, 4.5 g) in tetrahydrofuran (100 mL) held at $-78\text{ }^\circ\text{C}$ under nitrogen atmosphere was added dropwise a 1.6 M solution of n-butyllithium in tetrahydrofuran (44 mmol, 27 mL). After stirred at $-20\text{ }^\circ\text{C}$ for 0.5 h, the yellow mixture was placed at $-78\text{ }^\circ\text{C}$ and dimethyl formamide (30 mmol, 15 mL) was added. The solution was allowed to stir for 4 h, then warm slowly to room temperature. Saturated aqueous ammonium chloride (100 mL) was added, the mixture was transferred to a separatory funnel with EA, and the organic phase was separated. The aqueous phase was extracted with EA ($3 \times 50\text{ mL}$) and the organic extracts were combined, washed with brine (80 mL), dried over anhydrous sodium sulfate. The resulting mixture was concentrated in vacuo and further purification was performed by a short silica gel column.

Subsequently, to a solution of the above crude product in toluene, p-toluenesulfonic acid (0.2 mmol, 40 mg) was added. The mixture was stirred at $110\text{ }^\circ\text{C}$ for 5 h. The yellow oil was purified by a silica gel column eluted with PE/EA = 2/1 to give pure product as a yellow solid (58% yield).

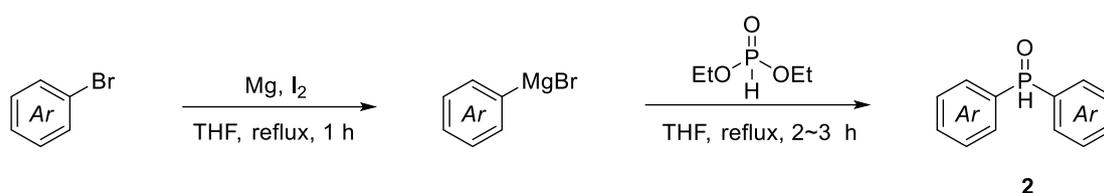
(3) General procedure F: Preparation of *in situ* cyclic imine **6**



General procedure F: n-Butyllithium (32 mmol, 13 mL, 2.5 M in hexance) was added dropwise over 20 minutes period to a cold (0 °C), mechanically stirred solution of the aryl sulfonamide (15 mmol, 3.4 g) in anhydrous tetrahydrofuran (100 ml) under a dry nitrogen atmosphere. After stirring an additional 25 min at 0 °C a precipitate formed. The suspension was cooled further to -78 °C and ethyl trifluoroacetate (45 mmol, 6.4 g) was added dropwise over 10 minutes. The resulting mixture was allowed to stir for 4 h at -30 °C and 2 h at ambient temperature. The reaction was quenched with 5% HCl (40 ml) and extracted with ether (50×3 mL). The combined ether phase was washed with brine (200 mL), dried over anhydrous Na₂SO₄. The solvent was removed and the crude product was obtained which can be used directly in the next step or purified by column chromatography (DCM/PE = 1/3-1/1).

To the crude product obtained above, TFA (75 mol) in toluene (20 mL) was added and the resulting mixture was stirred at 135 °C in sealed tube. After 2-3 h the solution was concentrated and the resultant solid was dissolved in CH₂Cl₂ (150 mL) and washed with saturated NaHCO₃ (50×2 mL) to remove traces of TFA. Then the organic phase was washed with brine and dried over Na₂SO₄. The solvent was removed and the obtained crude product was further purified by flash chromatography (PE/EA = 8/1-3/1) to give pure product as a white solid (68% yield).

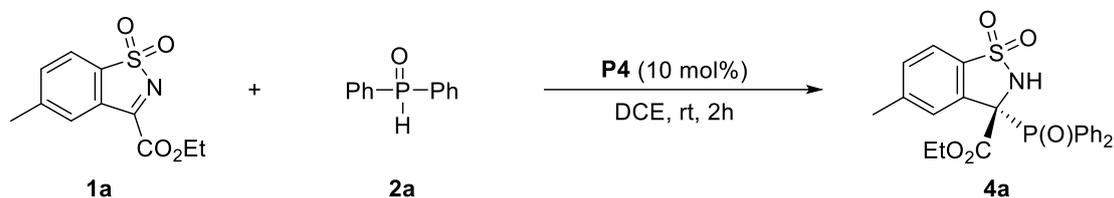
4.2 General procedure G: Preparation of phosphine oxides 2



General procedure G: Aryl bromide (10 mol, 1.6 g) in THF was added slowly to a stirred THF solution of I₂ (0.1 mol, 0.25 g), and heated under reflux for 1 hour. Then diethyl phosphate (3 mol, 0.42 g) in THF was added slowly under the cooling of an ice-water bath. The mixture thus obtained was heated under reflux for 1 hour. The resulting reaction mixture was cooled to 0 °C, and hydrochloric acid (50 mL, 6 M) was added slowly upon stirring. The solution was evaporated under reduced pressure at 40 °C. The residue was extracted with EA (150 mL). The organic layer was dried over anhydrous sodium sulfate and concentrated in vacuo, The crude product was purified by column chromatography on silica gel using PE/EA (1:1) as eluents.

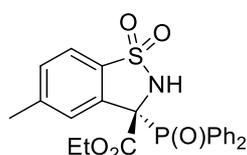
5. Representative procedure for asymmetric P-nucleophile addition

5.1 Representative procedure for asymmetric P-nucleophile addition to cyclic N-sulfonyl imines **1** with secondary phosphine oxides **2**

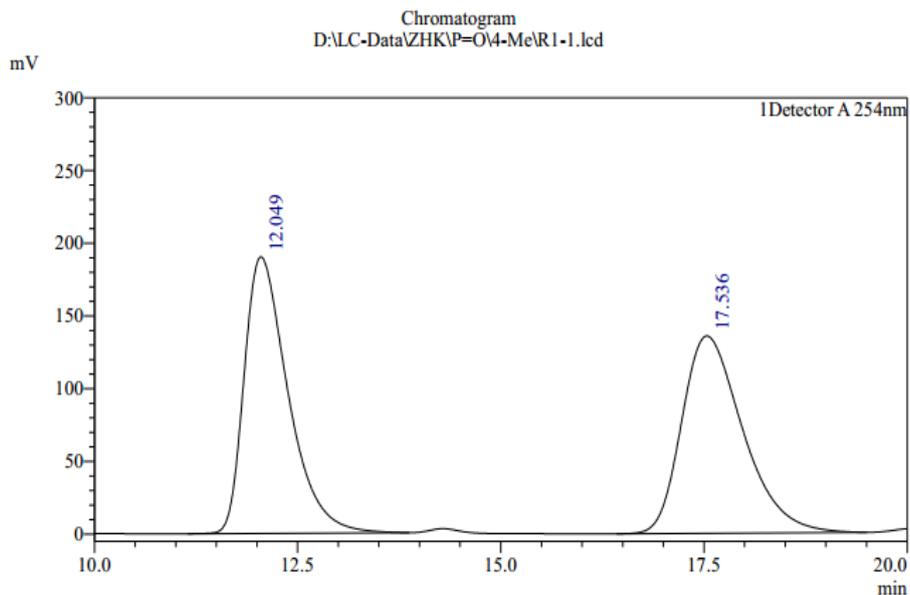


Representative procedure for 4a: To a flame-dried round bottle flask with a magnetic stirring bar were added cyclic ketimine **1a** (0.10 mmol, 25.3 mg), phosphine oxide **2a** (0.12 mmol, 24.2 mg), phosphonium salt **P4** (0.01 mmol, 5.7 mg), and DCE (1.0 mL). The reaction mixture was stirred at room temperature for 2 h. The solvent was removed under reduced pressure, and the residue was purified by column chromatography on silica gel (CH₂Cl₂/ethyl acetate = 3:1) to afford **4a** (96% yield) as a white solid.

(S)-ethyl 3-(diphenylphosphoryl)-5-methyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate (**4a**)



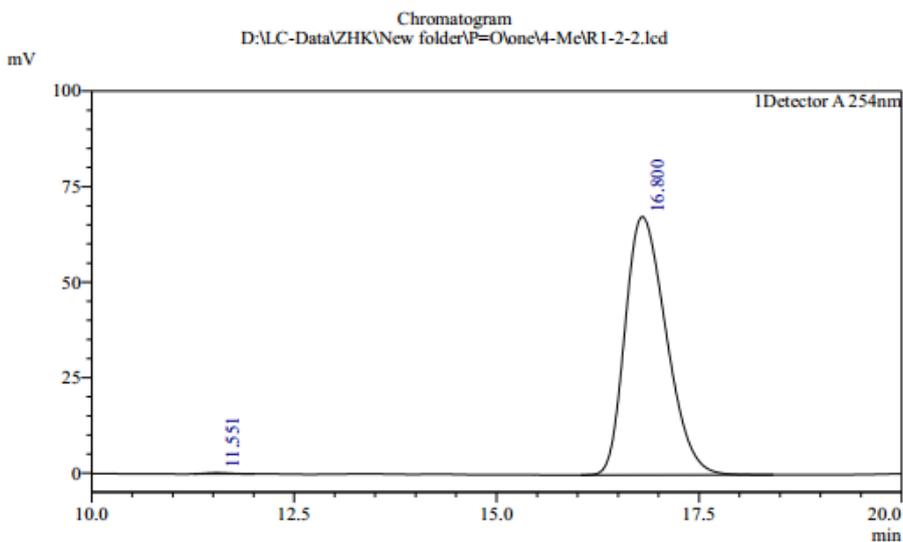
A white solid; m.p. = 185 - 187 °C; 43.7 mg, 96% yield; $[\alpha]_{\text{D}}^{25} = -70.67$ (*c* 1.1, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.17 - 8.13 (m, 1H), 8.12 - 8.05 (m, 2H), 7.64 - 7.56 (m, 3H), 7.55 - 7.49 (m, 2H), 7.46 (d, *J* = 8.0 Hz, 1H), 7.43 - 4.38 (m, 1H), 7.34 (d, *J* = 8.0 Hz, 1H), 7.30 - 7.23 (m, 3H), 6.00 (d, *J* = 4.0 Hz, 1H), 4.29 - 4.06 (m, 2H), 2.49 (s, 3H), 1.11 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.56 (d, *J* = 2.7 Hz), 144.64 (d, *J* = 1.9 Hz), 133.09 (d, *J* = 2.8 Hz), 132.82 (d, *J* = 2.8 Hz), 132.56 (dd, *J* = 16.7, 8.7 Hz), 131.85 (d, *J* = 1.6 Hz), 131.72 (d, *J* = 3.1 Hz), 131.43 (d, *J* = 2.4 Hz), 128.79 (d, *J* = 11.7 Hz), 128.42 (d, *J* = 2.4 Hz), 128.36, 128.18 (d, *J* = 12.3 Hz), 127.45 (d, *J* = 13.8 Hz), 126.48, 120.90 (d, *J* = 1.3 Hz), 70.02 (d, *J* = 71.4 Hz), 64.46, 21.97, 13.83; ³¹P NMR (162 MHz, CDCl₃) δ 28.18; HRMS (ESI) *m/z* calcd for C₂₃H₂₂NO₅PS [M+H]⁺ = 456.1035, found = 456.1038; The ee value was >99%, *t_R* (minor) = 12.1 min, *t_R* (major) = 16.9 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|--------|---------|---------|
| 1 | 12.049 | 7140364 | 190166 | 58.353 | 50.112 |
| 2 | 17.536 | 7108546 | 135720 | 41.647 | 49.888 |
| Total | | 14248910 | 325886 | 100.000 | 100.000 |

Racemic **4a**

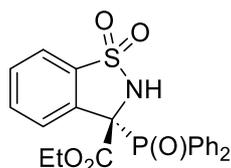


Peak Table

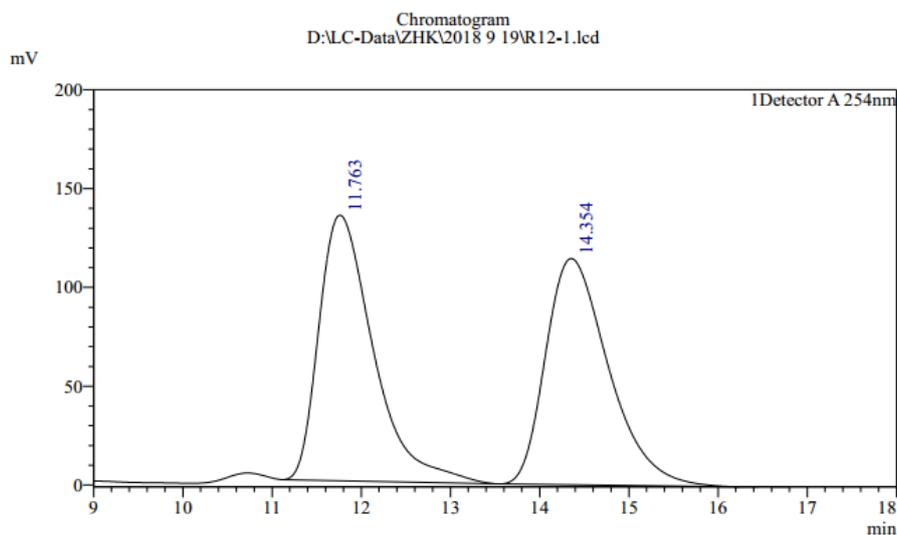
| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 11.551 | 5567 | 272 | 0.401 | 0.233 |
| 2 | 16.800 | 2386681 | 67601 | 99.599 | 99.767 |
| Total | | 2392247 | 67873 | 100.000 | 100.000 |

Enantiomerically enriched **4a**

(S)-ethyl 3-(diphenylphosphoryl)-2,3-dihydrobenzo[d]isothiazole
-3-carboxylate 1,1-dioxide (4b)



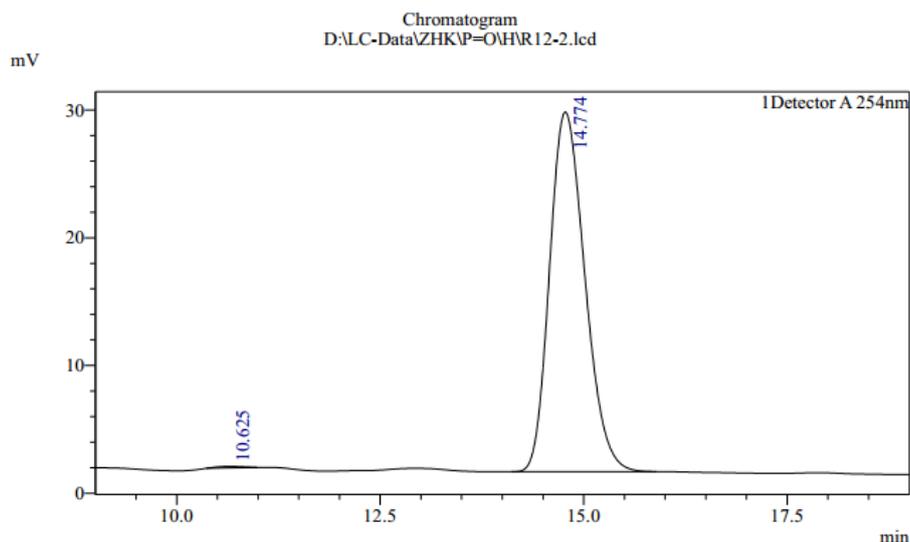
A white solid; m.p. = 178 - 180 °C; 42.3 mg, 96% yield; $[\alpha]_D^{25} = -13.32$ (*c* 0.9, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.38 (d, *J* = 7.6 Hz, 1H), 8.14 - 8.06 (m, 2H), 7.73 - 7.68 (m, 1H), 7.67 - 7.53 (m, 7H), 7.47 - 7.40 (m, 1H), 7.32 - 7.25 (m, 2H), 5.94 (d, *J* = 3.6 Hz, 1H), 4.29 - 4.10 (m, 2H), 1.16 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.63 (d, *J* = 2.6 Hz), 134.37 (d, *J* = 3.2 Hz), 133.51 (d, *J* = 1.7 Hz), 133.27 (d, *J* = 2.9 Hz), 132.99 (d, *J* = 2.8 Hz), 132.65 (dd, *J* = 18.7, 8.6 Hz), 131.26 (d, *J* = 2.3 Hz), 130.92 (d, *J* = 1.8 Hz), 128.93 (d, *J* = 11.7 Hz), 128.56 (d, *J* = 2.5 Hz), 128.33 (d, *J* = 12.3 Hz), 127.41 (d, *J* = 2.1 Hz), 126.38, 121.32 (d, *J* = 1.5 Hz), 70.29 (d, *J* = 70.8 Hz), 64.67, 13.94; ³¹P NMR (162 MHz, CDCl₃) δ 28.13; HRMS (ESI) *m/z* calcd for C₂₂H₂₁NO₅PS [M+H]⁺ = 478.0878, found = 478.0870; The ee value was >99%, *t_R* (minor) = 10.6 min, *t_R* (major) = 14.8 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|--------|---------|---------|
| 1 | 11.763 | 5698761 | 134415 | 53.996 | 50.374 |
| 2 | 14.354 | 5614092 | 114518 | 46.004 | 49.626 |
| Total | | 11312853 | 248933 | 100.000 | 100.000 |

Racemic **4b**

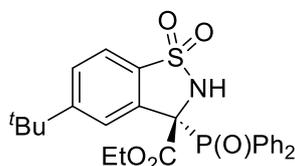


Peak Table

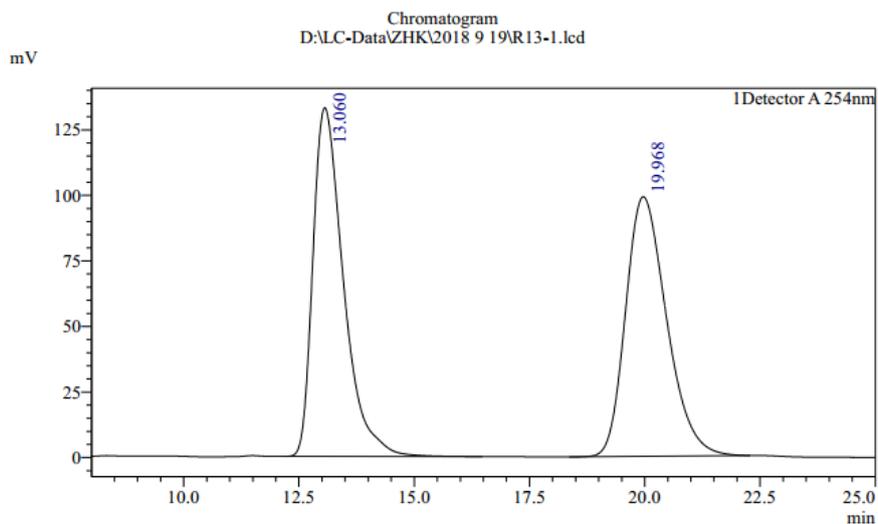
| Detector A 254nm | | | | | |
|------------------|-----------|--------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 10.625 | 2822 | 123 | 0.437 | 0.331 |
| 2 | 14.774 | 849237 | 28167 | 99.563 | 99.669 |
| Total | | 852059 | 28291 | 100.000 | 100.000 |

Enantiomerically enriched **4b**

(S)-ethyl 5-(tert-butyl)-3-(diphenylphosphoryl)-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (4c)



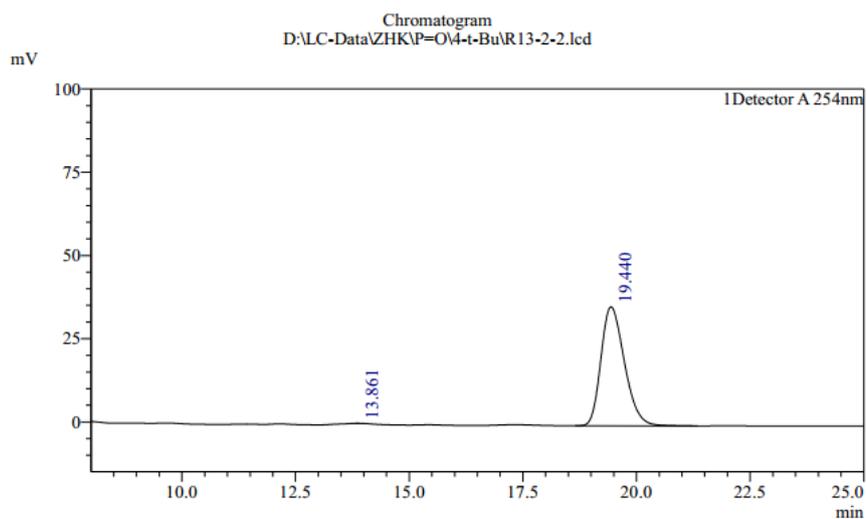
A white solid; m.p. = 117 - 119 °C; 46.2 mg, 93% yield; $[\alpha]_D^{25} = -63.67$ (c 1.1, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.34 - 8.30 (m, 1H), 8.13 - 8.04 (m, 2H), 7.70 - 7.61 (m, 3H), 7.60 - 7.53 (m, 2H), 7.54 (s, 1H), 7.49 (d, $J = 8.4$ Hz, 1H), 7.44 - 7.38 (m, 1H), 7.31 - 7.24 (m, 2H), 5.83 (s, 1H), 4.29 - 4.21 (m, 2H), 1.37 (s, 9H), 1.20 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 166.72 (d, $J = 2.8$ Hz), 157.80 (d, $J = 1.8$ Hz), 133.20 (d, $J = 2.8$ Hz), 132.87 (d, $J = 2.8$ Hz), 132.63 (dd, $J = 14.5, 8.5$ Hz), 131.44 (d, $J = 3.2$ Hz), 131.38 (d, $J = 2.4$ Hz), 128.90 (d, $J = 11.8$ Hz), 128.59, 128.26 (d, $J = 12.2$ Hz), 128.01 (d, $J = 1.5$ Hz), 127.59 (d, $J = 4.5$ Hz), 126.53, 125.65 (d, $J = 2.7$ Hz), 120.82 (d, $J = 1.3$ Hz), 70.18 (d, $J = 70.9$ Hz), 64.43, 35.70, 31.20, 13.97; ^{31}P NMR (162 MHz, CDCl_3) δ 28.13; HRMS (ESI) m/z calcd for $\text{C}_{26}\text{H}_{28}\text{NO}_5\text{PS}$ $[\text{M}+\text{Na}]^+ = 498.1504$, found = 498.1505; The ee value was >99%, t_R (minor) = 13.8 min, t_R (major) = 19.4 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Detector A 254nm | | | | | |
|------------------|-----------|----------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 13.060 | 6128099 | 133008 | 57.320 | 50.020 |
| 2 | 19.968 | 6123221 | 99036 | 42.680 | 49.980 |
| Total | | 12251320 | 232045 | 100.000 | 100.000 |

Racemic **4c**



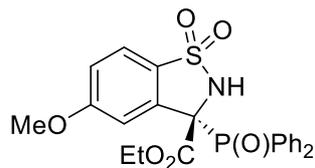
Peak Table

| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 13.861 | 2616 | 150 | 0.417 | 0.203 |
| 2 | 19.440 | 1286874 | 35768 | 99.583 | 99.797 |
| Total | | 1289490 | 35917 | 100.000 | 100.000 |

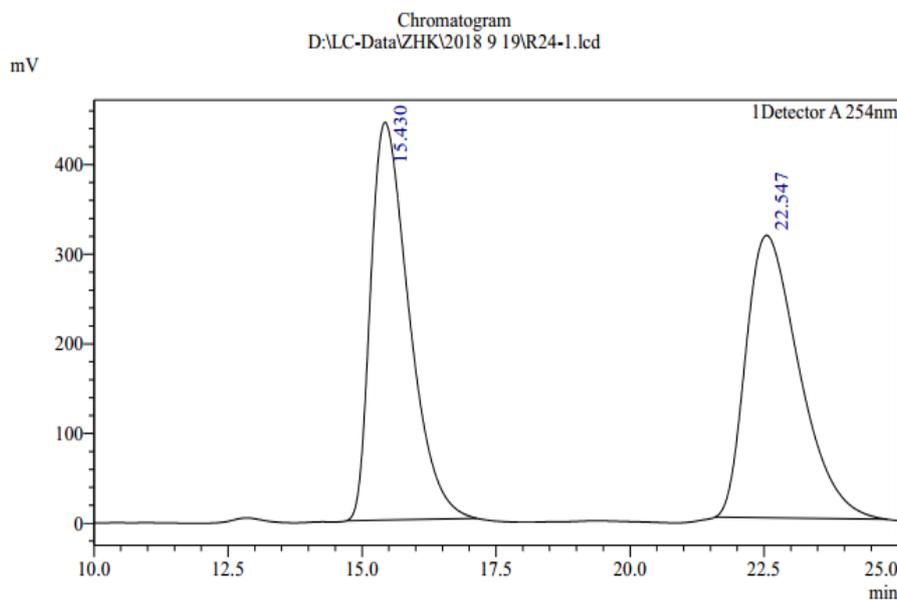
Enantiomerically enriched **4c**

(S)-ethyl 3-(diphenylphosphoryl)-5-methoxy-2,3-dihydrobenzo[d]

isothiazole-3-carboxylate 1,1-dioxide (4d)



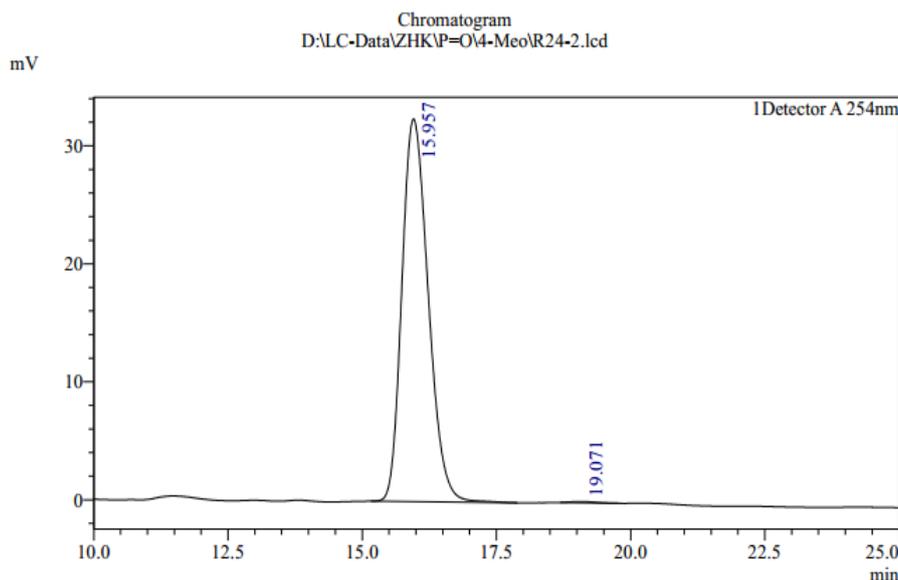
A white solid; m.p. = 125 - 126 °C; 42.9 mg, 91% yield; $[\alpha]_D^{25} = -52.67$ (c 1.8, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.14 - 8.05 (m, 2H), 7.82 - 7.78 (m, 1H), 7.68 - 7.59 (m, 3H), 7.58 - 7.51 (m, 2H), 7.49 - 7.41 (m, 2H), 7.34 - 7.27 (m, 2H), 7.05 (dd, $J = 8.8, 1.6$ Hz, 1H), 5.94 (d, $J = 4.0$ Hz, 1H), 4.28 - 4.14 (m, 2H), 3.93 (s, 3H), 1.14 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 166.43 (d, $J = 2.5$ Hz), 163.62 (d, $J = 1.6$ Hz), 133.87 (d, $J = 2.2$ Hz), 133.20 (d, $J = 2.6$ Hz), 132.96 (d, $J = 2.7$ Hz), 132.61 (dd, $J = 18.4, 8.6$ Hz), 128.87 (d, $J = 11.7$ Hz), 128.32, 128.31 (d, $J = 12.3$ Hz), 127.42 (d, $J = 15.3$ Hz), 126.45, 126.37 (d, $J = 3.0$ Hz), 122.45, 118.95 (d, $J = 1.1$ Hz), 111.38 (d, $J = 2.1$ Hz), 69.86 (d, $J = 70.8$ Hz), 64.54, 56.26, 13.91; ^{31}P NMR (162 MHz, CDCl_3) δ 28.18; HRMS (ESI) m/z calcd for $\text{C}_{23}\text{H}_{22}\text{NO}_6\text{PS}$ $[\text{M}+\text{H}]^+ = 472.0984$, found = 472.0981; The ee value was >99%, t_R (minor) = 16.0 min, t_R (major) = 19.1 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|--------|---------|---------|
| 1 | 15.430 | 22048437 | 443406 | 58.456 | 50.233 |
| 2 | 22.547 | 21843527 | 315127 | 41.544 | 49.767 |
| Total | | 43891964 | 758533 | 100.000 | 100.000 |

Racemic **4d**

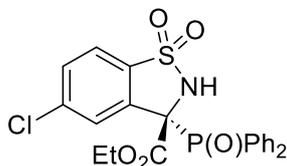


Peak Table

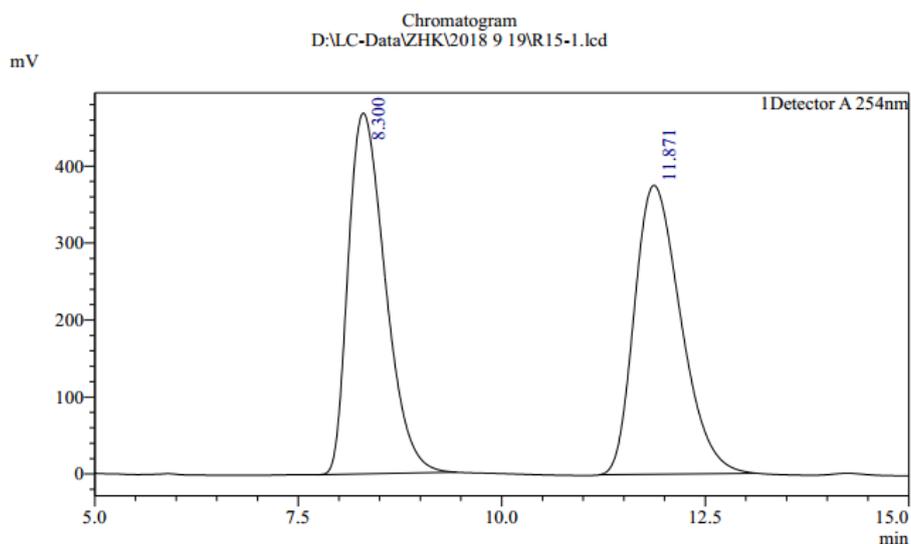
| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 15.957 | 1105712 | 32439 | 99.682 | 99.718 |
| 2 | 19.071 | 3129 | 103 | 0.318 | 0.282 |
| Total | | 1108841 | 32542 | 100.000 | 100.000 |

Enantiomerically enriched **4d**

(S)-ethyl 5-chloro-3-(diphenylphosphoryl)-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (4e)



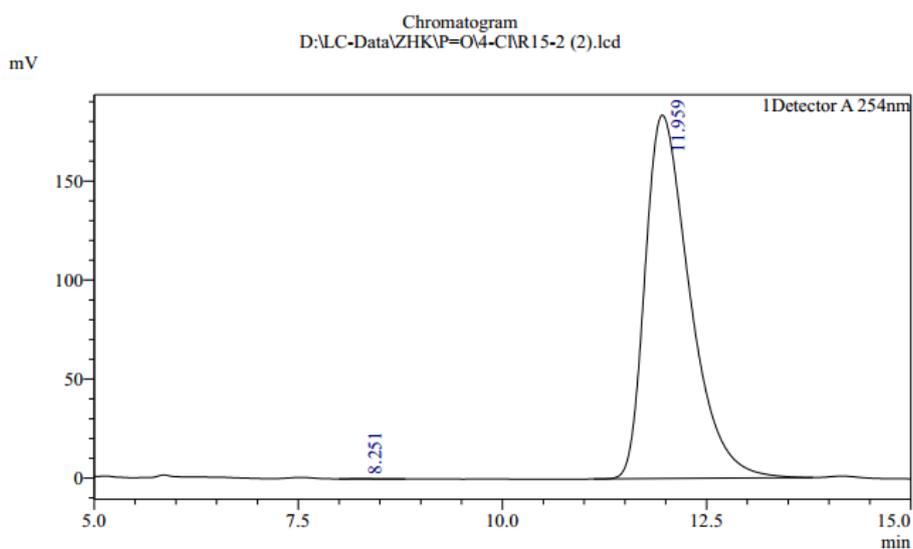
A white solid; m.p. = 139 - 141 °C; 44.7 mg, 94% yield; $[\alpha]_D^{25} = -34.22$ (*c* 0.2, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.40 - 8.36 (m, 1H), 8.14 - 8.05 (m, 2H), 7.70 - 7.61 (m, 3H), 7.61 - 7.54 (m, 2H), 7.53 - 7.43 (m, 3H), 7.36 - 7.28 (m, 2H), 6.01 (d, *J* = 4.0 Hz, 1H), 4.30 - 4.14 (m, 2H), 1.15 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.15 (d, *J* = 2.2 Hz), 140.09 (d, *J* = 2.1 Hz), 133.40 (d, *J* = 2.7 Hz), 133.27 (d, *J* = 2.4 Hz), 133.17 (d, *J* = 2.8 Hz), 132.85 (d, *J* = 3.1 Hz), 132.63 (dd, *J* = 25.8, 8.6 Hz), 131.47 (d, *J* = 1.6 Hz), 128.98 (d, *J* = 11.9 Hz), 128.54, 128.45 (d, *J* = 12.4 Hz), 128.06, 127.10 (d, *J* = 4.9 Hz), 126.08, 122.34 (d, *J* = 1.3 Hz), 69.90 (d, *J* = 69.3 Hz), 64.95, 13.91; ³¹P NMR (162 MHz, CDCl₃) δ 28.10; HRMS (ESI) *m/z* calcd for C₂₂H₁₉NO₅PSCl [M+H]⁺ = 476.0488, found = 476.0493; The ee value was >99%, *t_R* (minor) = 8.3 min, *t_R* (major) = 12.0 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Detector A 254nm | | | | | |
|------------------|-----------|----------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 8.300 | 14820184 | 468775 | 55.525 | 49.892 |
| 2 | 11.871 | 14884317 | 375491 | 44.475 | 50.108 |
| Total | | 29704501 | 844266 | 100.000 | 100.000 |

Racemic **4e**

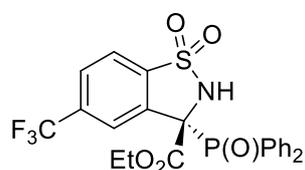


Peak Table

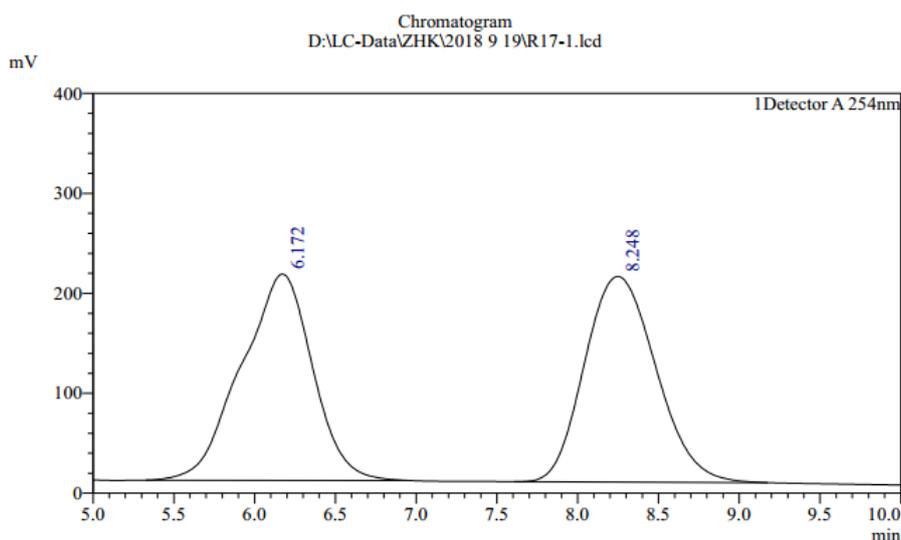
| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 8.251 | 2673 | 153 | 0.083 | 0.038 |
| 2 | 11.959 | 6977098 | 183521 | 99.917 | 99.962 |
| Total | | 6979771 | 183674 | 100.000 | 100.000 |

Enantiomerically enriched **4e**

(S)-ethyl 3-(diphenylphosphoryl)-5-(trifluoromethyl)-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (4f)



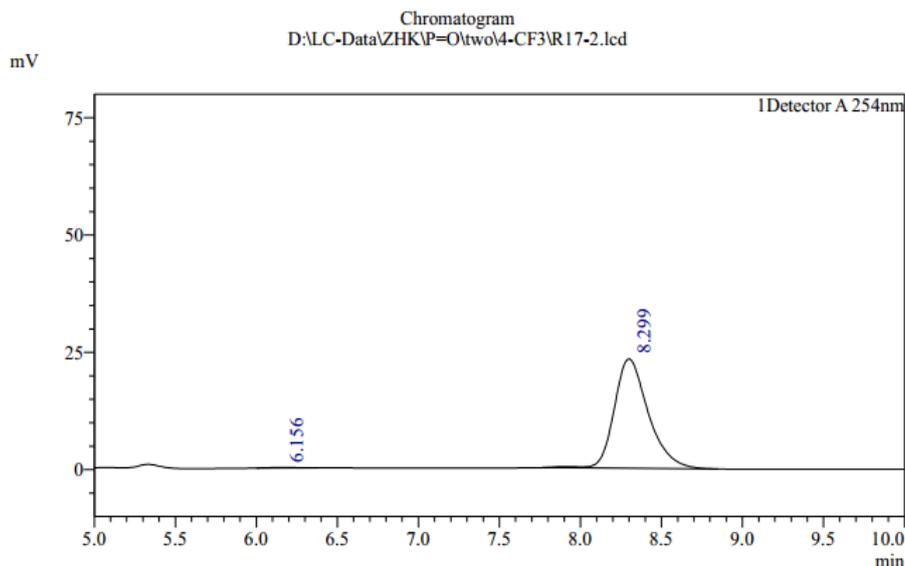
A white solid; m.p. = 112 - 113 °C; 45.8 mg, 90% yield; $[\alpha]_D^{25} = -42.37$ (c 0.9, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.63 (s, 1H), 8.14 - 8.06 (m, 2H), 7.81 (d, $J = 8.0$ Hz, 1H), 7.73 - 7.65 (m, 2H), 7.65 - 7.55 (m, 4H), 7.48 - 7.42 (m, 1H), 7.31 (m, 2H), 6.09 (d, $J = 3.6$ Hz, 1H), 4.30 - 4.15 (m, 2H), 1.16 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 166.03 (d, $J = 2.2$ Hz), 137.41, 135.61 (d, $J = 1.9$ Hz), 135.28 (d, $J = 1.8$ Hz), 133.50 (d, $J = 2.8$ Hz), 133.25 (d, $J = 2.8$ Hz), 132.61 (dd, $J = 23.4, 8.6$ Hz), 132.57, 129.05 (d, $J = 12.0$ Hz), 128.50 (d, $J = 12.4$ Hz), 126.96 (d, $J = 3.8$ Hz), 126.15 (dd, $J = 3.6, 2.6$ Hz), 125.91, 124.37, 122.08 (d, $J = 1.5$ Hz), 121.66, 118.94, 70.23 (d, $J = 68.9$ Hz), 65.03, 13.84; ^{31}P NMR (162 MHz, CDCl_3) δ 28.21; ^{19}F NMR (376 MHz, CDCl_3) δ -62.73; HRMS (ESI) m/z calcd for $\text{C}_{23}\text{H}_{19}\text{NO}_5\text{PSF}_3$ $[\text{M}+\text{Na}]^+ = 510.0752$, found = 510.0744; The ee value was >99%, t_R (minor) = 6.2 min, t_R (major) = 8.3 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|--------|---------|---------|
| 1 | 6.172 | 6335493 | 206418 | 50.086 | 50.025 |
| 2 | 8.248 | 6329172 | 205708 | 49.914 | 49.975 |
| Total | | 12664665 | 412126 | 100.000 | 100.000 |

Racemic **4f**

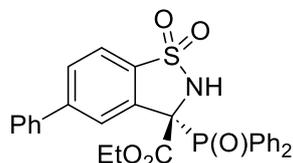


Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|--------|--------|---------|---------|
| 1 | 6.156 | 1322 | 105 | 0.448 | 0.385 |
| 2 | 8.299 | 342242 | 23332 | 99.552 | 99.615 |
| Total | | 343564 | 23437 | 100.000 | 100.000 |

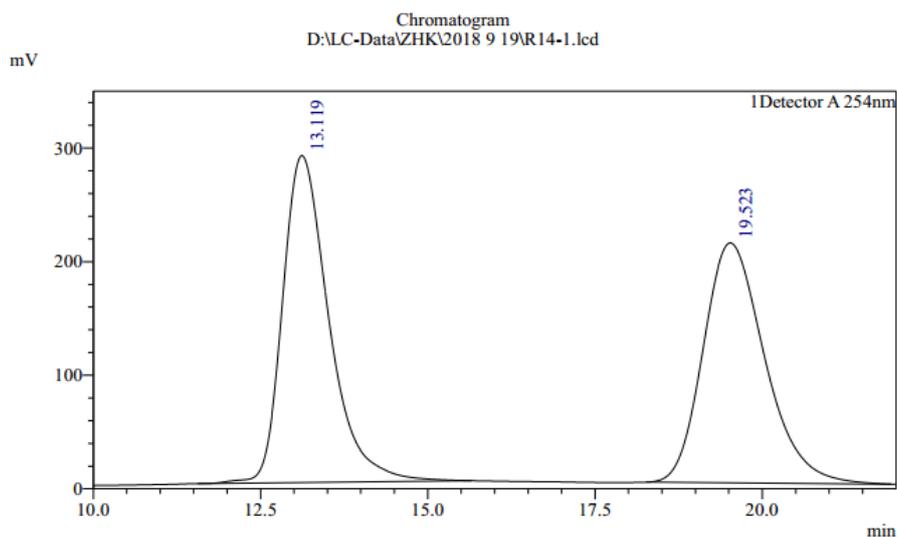
Enantiomerically enriched **4f**

(S)-ethyl 3-(diphenylphosphoryl)-5-phenyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (4g)



A white solid; m.p. = 165 - 167 °C; 46.5 mg, 90% yield; $[\alpha]_D^{25} = -78.33$ (*c* 1.2, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.61 - 8.55 (m, 1H), 8.18 - 8.09 (m, 2H), 7.81 - 7.76 (m, 1H), 7.70 - 7.63 (m, 6H), 7.62 - 7.56 (m, 2H), 7.55 - 7.49 (m, 2H), 7.48 - 7.41 (m, 2H), 7.30 - 7.27 (m, 2H), 5.97 (d, *J* = 3.2 Hz, 1H), 4.37 - 4.11 (m, 2H), 1.15 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.63 (d, *J* = 2.6 Hz), 146.77, 138.88, 133.29 (d, *J* = 2.7 Hz), 133.02 (d, *J* = 2.8 Hz), 132.91 (d, *J* = 3.2 Hz), 132.66 (dd, *J* = 19.4, 8.6 Hz), 132.10 (d, *J* = 2.2 Hz), 129.90 (d, *J* = 1.6 Hz), 129.28, 129.01, 128.90, 128.41 (d, *J* = 12.3 Hz), 128.34, 127.79, 127.45 (d, *J* = 9.2 Hz), 126.74 (d, *J* = 2.6 Hz), 126.45, 121.63, 70.27 (d, *J* = 70.6 Hz), 64.71, 13.98; ³¹P NMR (162 MHz, CDCl₃) δ 28.25; HRMS (ESI) *m/z* calcd for C₂₈H₂₄NO₅PS [M+H]⁺ = 518.1191, found = 518.1186; The ee value was >99%, *t_R* (minor) = 13.8 min, *t_R* (major) = 19.4 min (Chiralcel IC, λ = 254

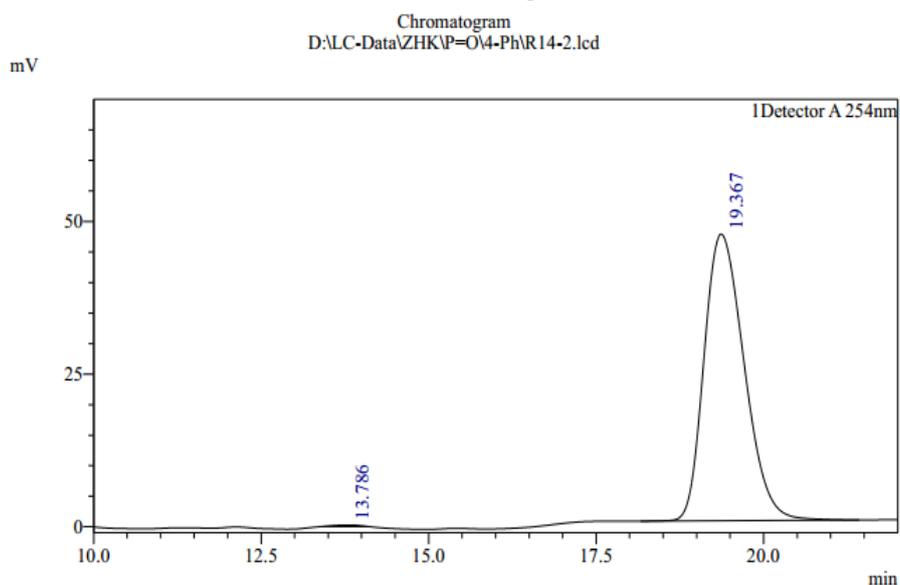
nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|--------|---------|---------|
| 1 | 13.119 | 13831731 | 287817 | 57.673 | 50.603 |
| 2 | 19.523 | 13502138 | 211233 | 42.327 | 49.397 |
| Total | | 27333869 | 499050 | 100.000 | 100.000 |

Racemic **4g**

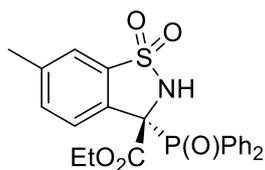


Peak Table

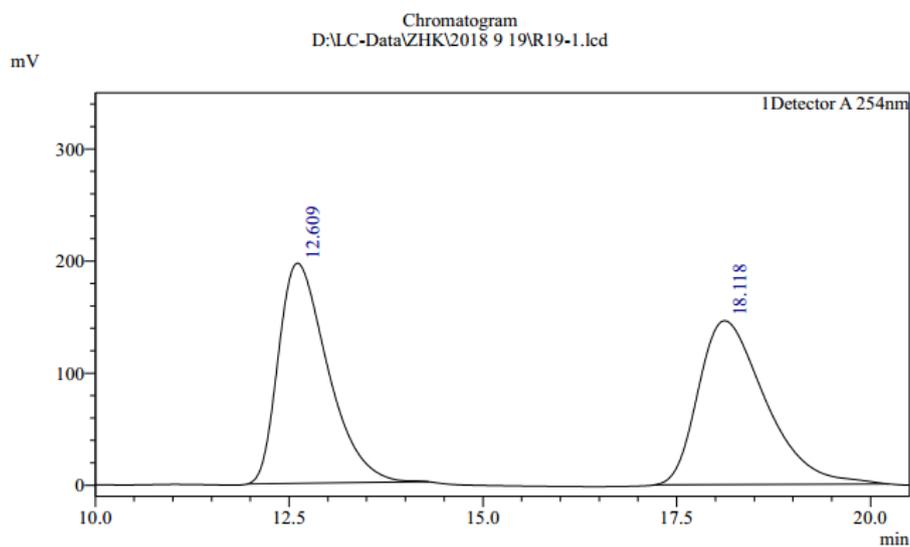
| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 13.786 | 8451 | 291 | 0.616 | 0.429 |
| 2 | 19.367 | 1960423 | 46971 | 99.384 | 99.571 |
| Total | | 1968874 | 47262 | 100.000 | 100.000 |

Enantiomerically enriched **4g**

**(S)-ethyl 3-(diphenylphosphoryl)-6-methyl-2,3-dihydrobenzo[d]
isothiazole-3-carboxylate 1,1-dioxide (4h)**



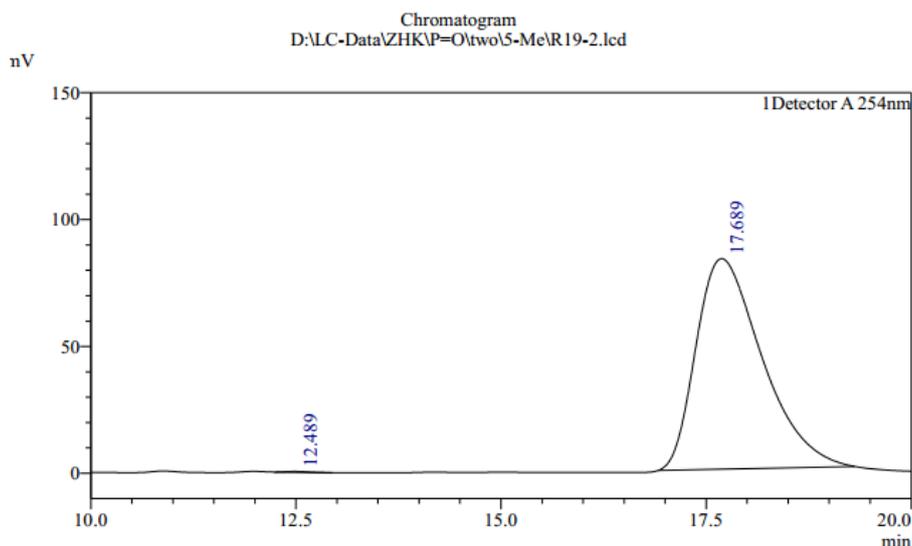
A white solid; m.p. = 147 - 149 °C; 42.3 mg, 93% yield; $[\alpha]_D^{25} = -34.72$ (*c* 0.9, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.24 (dd, *J* = 8.0, 1.2 Hz, 1H), 8.13 - 8.05 (m, 2H), 7.66 - 7.59 (m, 3H), 7.58 - 7.48 (m, 3H), 7.46 - 7.41 (m, 1H), 7.40 - 7.36 (m, 1H), 7.32 - 7.26 (m, 2H), 5.94 (d, *J* = 4.0 Hz, 1H), 4.28 - 4.10 (m, 2H), 2.42 (s, 3H), 1.13 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.80 (d, *J* = 3.0 Hz), 141.81 (d, *J* = 1.9 Hz), 134.68 (d, *J* = 1.7 Hz), 134.39 (d, *J* = 3.2 Hz), 133.16 (d, *J* = 2.7 Hz), 132.90 (d, *J* = 2.8 Hz), 132.62 (dd, *J* = 28.7, 8.7 Hz), 128.86 (d, *J* = 11.7 Hz), 128.52, 128.42 (d, *J* = 2.5 Hz), 128.28 (d, *J* = 12.3 Hz), 128.14 (d, *J* = 2.5 Hz), 127.56 (d, *J* = 2.8 Hz), 126.53, 121.17 (d, *J* = 1.4 Hz), 70.05 (d, *J* = 71.7 Hz), 64.52, 21.41, 13.91; ³¹P NMR (162 MHz, CDCl₃) δ 27.96 ; HRMS (ESI) *m/z* calcd for C₂₃H₂₂NO₅PS [M+H]⁺ = 456.1035, found = 456.1038; The ee value was >99%, *t_R* (minor) = 12.5 min, *t_R* (major) = 17.7 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|--------|---------|---------|
| 1 | 12.609 | 8717887 | 196415 | 57.348 | 49.774 |
| 2 | 18.118 | 8797097 | 146083 | 42.652 | 50.226 |
| Total | | 17514984 | 342497 | 100.000 | 100.000 |

Racemic **4h**

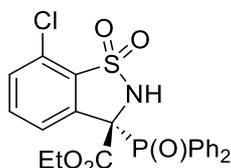


Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 12.489 | 6125 | 332 | 0.399 | 0.132 |
| 2 | 17.689 | 4648277 | 82985 | 99.601 | 99.868 |
| Total | | 4654403 | 83317 | 100.000 | 100.000 |

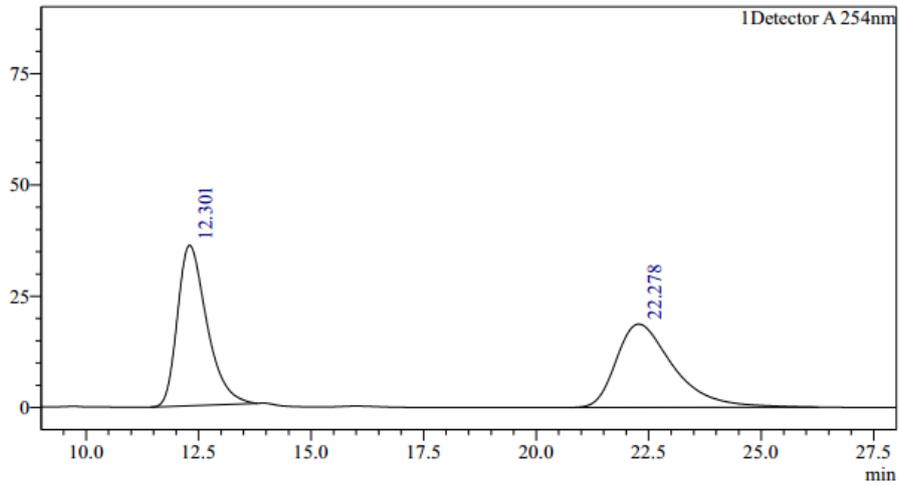
Enantiomerically enriched **4h**

(S)-ethyl 7-chloro-3-(diphenylphosphoryl)-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (4i)



A white solid; m.p. = 131 - 133 °C; 44.7 mg, 94% yield; $[\alpha]_D^{25} = -42.32$ (c 0.8, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.34 - 8.29 (m, 1H), 8.16 - 8.05 (m, 2H), 7.70 - 7.54 (m, 6H), 7.53 - 7.44 (m, 2H), 7.36 - 7.29 (m, 2H), 6.05 (d, $J = 4.4$ Hz, 1H), 4.28 - 4.07 (m, 2H), 1.14 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 166.27 (d, $J = 2.4$ Hz), 134.51 (d, $J = 1.8$ Hz), 133.91 (d, $J = 2.0$ Hz), 133.39 (d, $J = 2.8$ Hz), 133.29 (d, $J = 2.8$ Hz), 132.62 (dd, $J = 19.5, 8.7$ Hz), 132.30 (d, $J = 3.2$ Hz), 131.65 (d, $J = 1.7$ Hz), 128.98 (d, $J = 11.8$ Hz), 128.79 (d, $J = 1.7$ Hz), 128.48 (d, $J = 12.3$ Hz), 128.15, 127.16 (d, $J = 3.0$ Hz), 126.89 (d, $J = 2.5$ Hz), 126.10, 69.35 (d, $J = 69.7$ Hz), 64.88, 13.92; ^{31}P NMR (162 MHz, CDCl_3) δ 21.58 ; HRMS (ESI) m/z calcd for $\text{C}_{22}\text{H}_{19}\text{NO}_5\text{PSCl}$ $[\text{M}+\text{H}]^+ = 476.0488$, found = 476.0481; The ee value was 98%, t_R (minor) = 14.6 min, t_R (major) = 23.0 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).

Chromatogram
D:\LC-Data\ZHK\IP=O\two\6-C\IR20-1.lcd



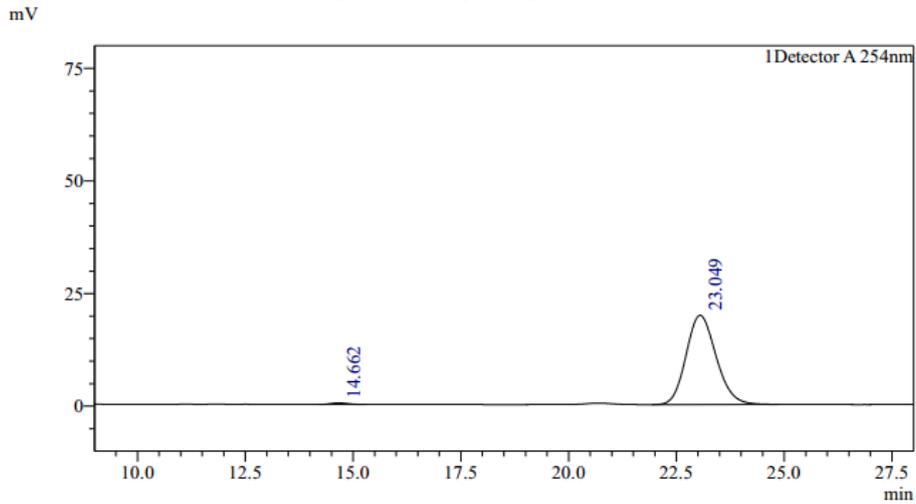
Detector A 254nm

Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 12.301 | 1638270 | 36081 | 65.856 | 49.872 |
| 2 | 22.278 | 1646665 | 18707 | 34.144 | 50.128 |
| Total | | 3284935 | 54787 | 100.000 | 100.000 |

Racemic 4i

Chromatogram
D:\LC-Data\ZHK\IP=O\two\6-C\IR20-2.lcd



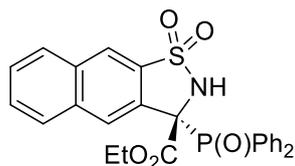
Detector A 254nm

Peak Table

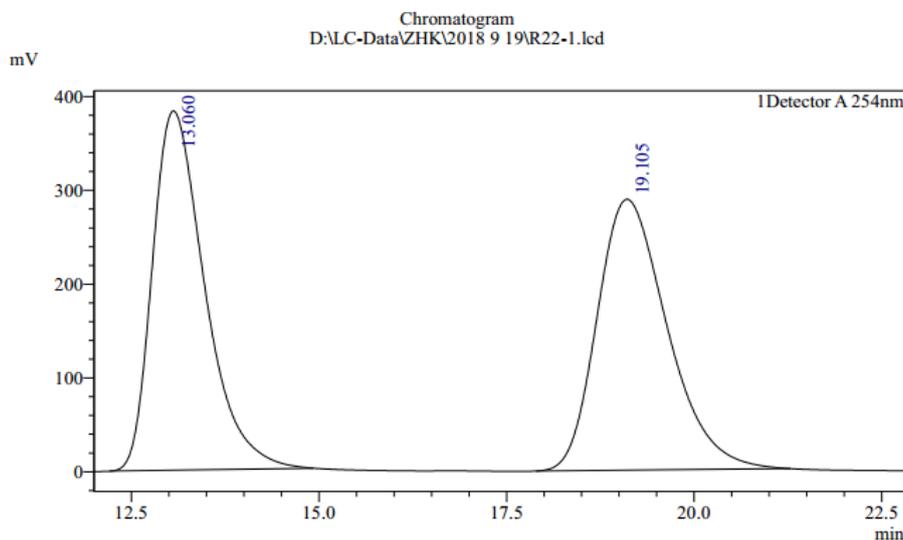
| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|--------|--------|---------|---------|
| 1 | 14.662 | 8846 | 334 | 1.656 | 0.912 |
| 2 | 23.049 | 960788 | 19812 | 98.344 | 99.088 |
| Total | | 969633 | 20146 | 100.000 | 100.000 |

Enantiomerically enriched 4i

**(S)-ethyl 3-(diphenylphosphoryl)-2,3-dihydronaphtho[2,3-d]
isothiazole-3-carboxylate 1,1-dioxide (4j)**



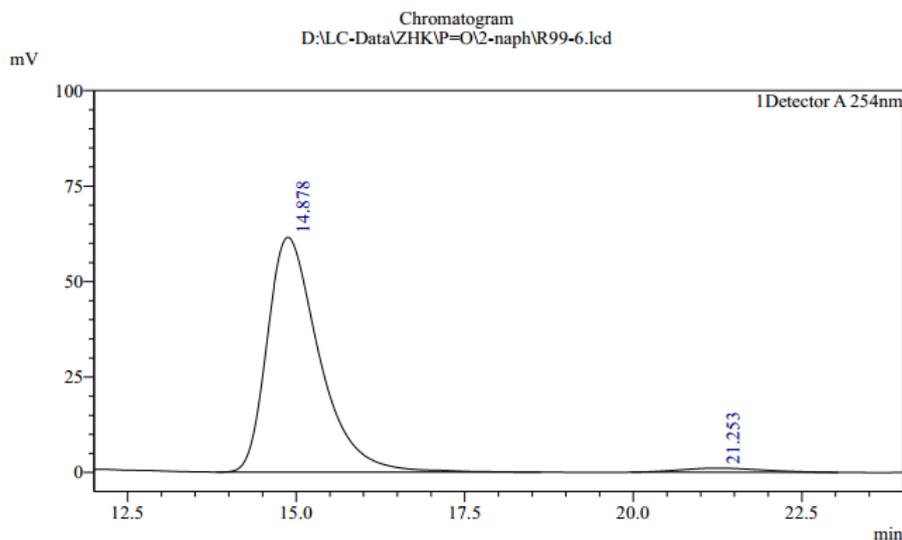
A white solid; m.p. = 116 - 118 °C; 45.2 mg, 92% yield; $[\alpha]_D^{25} = -37.43$ (*c* 0.6, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.88 - 8.83 (m, 1H), 8.21 - 8.06 (m, 4H), 7.95 (d, *J* = 8.4 Hz, 1H), 7.74 - 7.53 (m, 7H), 7.41 - 7.34 (m, 1H), 7.25 - 7.18 (m, 2H), 6.06 (d, *J* = 4.0 Hz, 1H), 4.33 - 4.13 (m, 2H), 1.16 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.98 (d, *J* = 3.1 Hz), 135.38 (d, *J* = 2.1 Hz), 133.23 (d, *J* = 2.6 Hz), 132.89 (d, *J* = 2.8 Hz), 132.67 (dd, *J* = 19.3, 8.6 Hz), 131.88 (d, *J* = 2.6 Hz), 129.42, 129.21 (d, *J* = 5.3 Hz), 128.93 (d, *J* = 11.7 Hz), 128.53 (d, *J* = 3.3 Hz), 128.47, 128.31 (d, *J* = 12.3 Hz), 127.63 (d, *J* = 12.5 Hz), 126.64, 126.20 (d, *J* = 2.6 Hz), 122.10 (d, *J* = 1.0 Hz), 70.05 (d, *J* = 71.5 Hz), 64.70, 13.98; ³¹P NMR (162 MHz, CDCl₃) δ 27.95; HRMS (ESI) *m/z* calcd for C₂₆H₂₂NO₅PS [M+H]⁺ = 492.1035, found = 492.1028; The ee value was 95%, *t_R* (minor) = 14.9 min, *t_R* (major) = 21.3 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Detector A 254nm

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|--------|---------|---------|
| 1 | 13.060 | 18565269 | 383108 | 57.020 | 49.737 |
| 2 | 19.105 | 18761516 | 288774 | 42.980 | 50.263 |
| Total | | 37326785 | 671882 | 100.000 | 100.000 |

Racemic **4j**



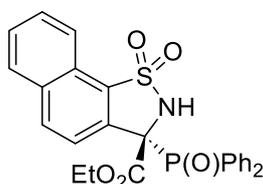
Peak Table

| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 14.878 | 3310077 | 61484 | 98.252 | 97.255 |
| 2 | 21.253 | 93432 | 1094 | 1.748 | 2.745 |
| Total | | 3403509 | 62578 | 100.000 | 100.000 |

Enantiomerically enriched **4j**

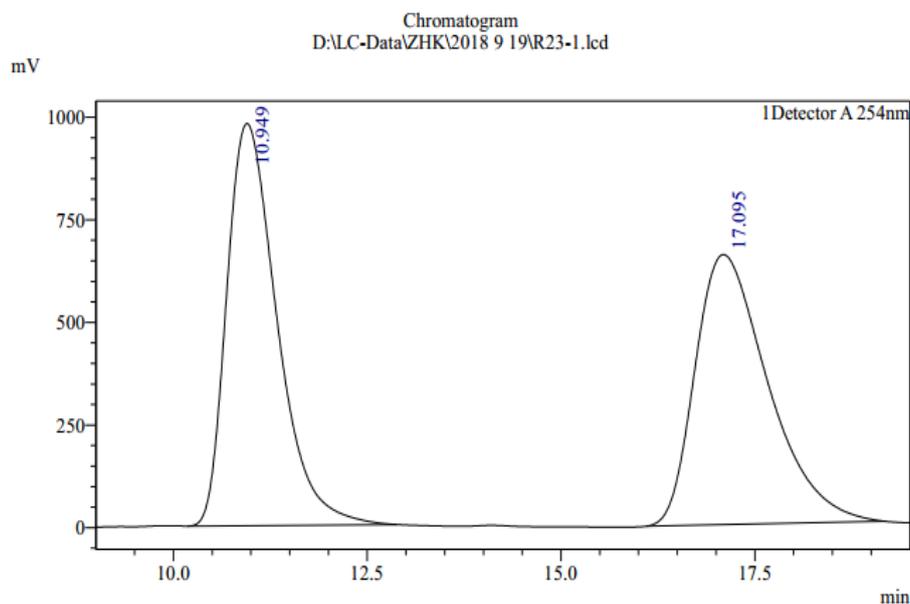
(S)-ethyl 3-(diphenylphosphoryl)-2,3-dihydronaphtho[2,1-d]

isothiazole-3-carboxylate 1,1-dioxide (4k)



A white solid; m.p. = 175 - 177 °C; 44.2 mg, 90% yield; $[\alpha]_D^{25} = -102.33$ (c 1.9, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.44 - 8.39 (m, 1H), 8.24 - 8.18 (m, 1H), 8.18 - 8.09 (m, 3H), 7.98 - 7.93 (m, 1H), 7.69 - 7.61 (m, 5H), 7.60 - 7.54 (m, 2H), 7.39 - 7.32 (m, 1H), 7.24 - 7.28 (m, 2H), 6.13 (d, $J = 4.0$ Hz, 1H), 4.30 - 4.14 (m, 2H), 1.17 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 166.66 (d, $J = 2.5$ Hz), 134.14 (d, $J = 1.3$ Hz), 133.99 (d, $J = 1.1$ Hz), 133.23 (d, $J = 2.8$ Hz), 132.99 (d, $J = 2.8$ Hz), 132.61 (dd, $J = 15.6, 8.7$ Hz), 131.02 (d, $J = 2.3$ Hz), 130.00 (d, $J = 4.0$ Hz), 129.17, 128.90 (d, $J = 11.8$ Hz), 128.60 (d, $J = 19.2$ Hz), 128.27 (d, $J = 12.3$ Hz), 127.45 (d, $J = 17.3$ Hz), 126.32, 124.98 (d, $J = 1.5$ Hz), 123.73 (d, $J = 1.7$ Hz), 122.99, 70.72 (d, $J = 70.8$ Hz), 70.01, 64.69, 13.94; ^{31}P NMR (162 MHz, CDCl_3) δ 28.32; HRMS (ESI) m/z calcd for $\text{C}_{26}\text{H}_{22}\text{NO}_5\text{PS}$ $[\text{M}+\text{H}]^+ = 492.1035$, found = 492.1036; The ee value was >99%, t_R (minor) = 11.4 min, t_R (major) = 16.7 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-

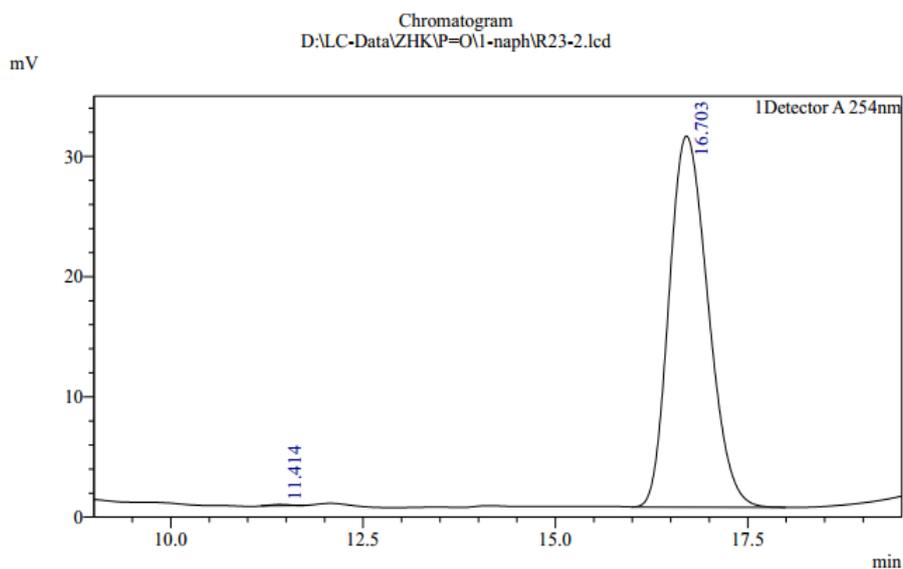
PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|---------|---------|---------|
| 1 | 10.949 | 44066617 | 979623 | 59.823 | 50.576 |
| 2 | 17.095 | 43063403 | 657919 | 40.177 | 49.424 |
| Total | | 87130020 | 1637542 | 100.000 | 100.000 |

Racemic 4k



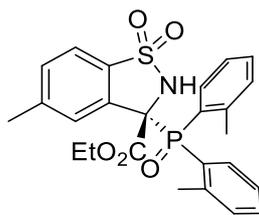
Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 11.414 | 1567 | 101 | 0.326 | 0.144 |
| 2 | 16.703 | 1084544 | 30855 | 99.674 | 99.856 |
| Total | | 1086111 | 30956 | 100.000 | 100.000 |

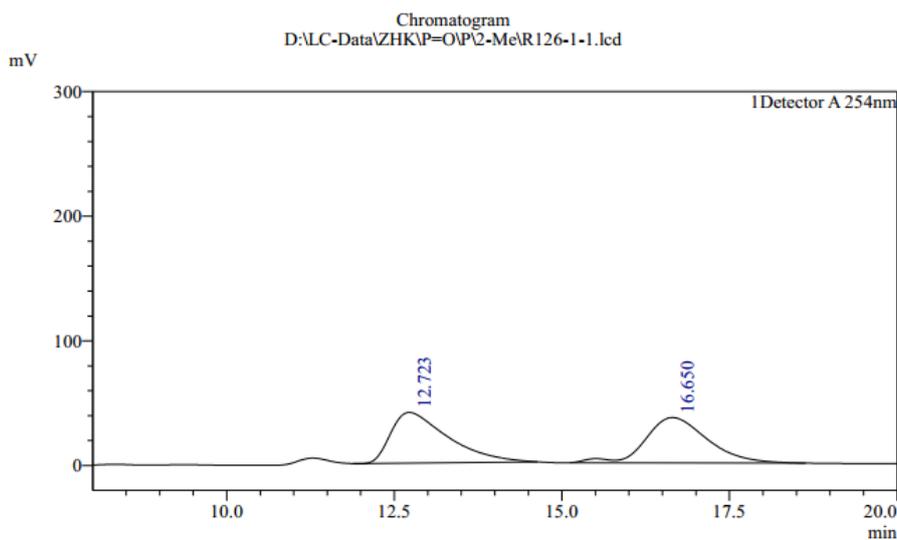
Enantiomerically enriched 4k

(S)-ethyl 3-(di-o-tolylphosphoryl)-5-methyl-2,3-dihydrobenzo[d]

isothiazole-3-carboxylate 1,1-dioxide (4l)



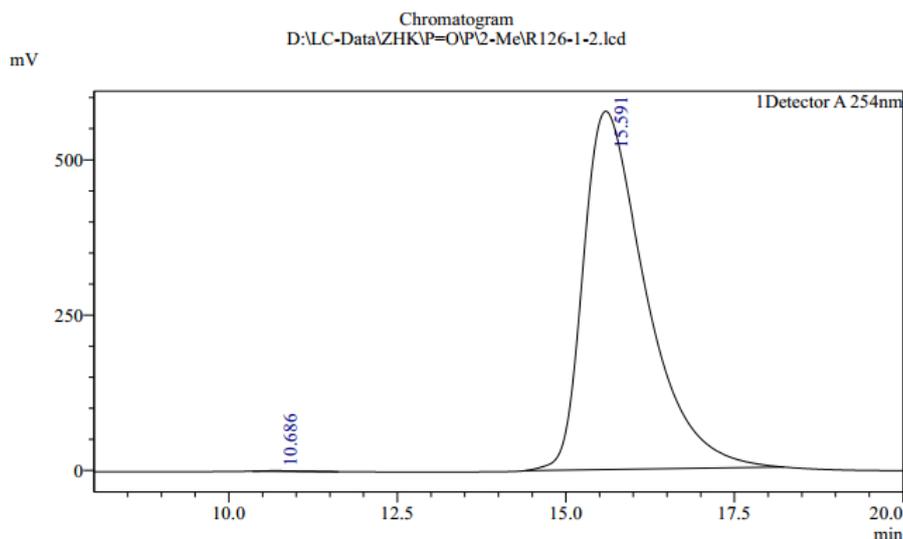
A white solid; m.p. = 171 - 173 °C; 44.4 mg, 92% yield; $[\alpha]_D^{25} = -41.00$ (*c* 1.1, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.24 (s, 1H), 7.67 - 7.7.58 (m, 1H), 7.54 - 7.38 (m, 3H), 7.33 - 7.24 (m, 3H), 7.23 - 7.16 (m, 1H), 7.07 - 6.94 (m, 2H), 5.97 (d, *J* = 5.6 Hz, 1H), 4.36 - 4.23 (m, 2H), 2.51 (s, 3H), 2.42 (s, 3H), 1.98 (s, 3H), 1.16 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.12 (d, *J* = 3.1 Hz), 144.46 (d, *J* = 1.9 Hz), 144.05 (dd, *J* = 20.4, 9.0 Hz), 134.32 (d, *J* = 11.0 Hz), 132.84 (d, *J* = 11.2 Hz), 132.50 (dd, *J* = 7.3, 2.7 Hz), 132.27 (d, *J* = 1.3 Hz), 132.12 (d, *J* = 11.9 Hz), 131.98 (d, *J* = 1.6 Hz), 131.88 (d, *J* = 3.1 Hz), 130.43 (d, *J* = 10.4 Hz), 129.69, 129.44 (d, *J* = 2.3 Hz), 128.72, 126.70, 125.71, 125.49 (d, *J* = 12.2 Hz), 125.01 (d, *J* = 12.7 Hz), 121.10 (d, *J* = 1.3 Hz), 70.01 (d, *J* = 71.4 Hz), 64.73, 22.03, 21.71 (d, *J* = 3.8 Hz), 21.11 (d, *J* = 2.9 Hz), 13.84; ³¹P NMR (162 MHz, CDCl₃) δ 36.35; HRMS (ESI) *m/z* calcd for C₂₅H₂₆NO₅PS [M+H]⁺ = 484.1348, found = 484.1347; The ee value was >99%, *t_R* (minor) = 10.7 min, *t_R* (major) = 15.6min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 12.723 | 2400607 | 40620 | 52.767 | 50.685 |
| 2 | 16.650 | 2335744 | 36359 | 47.233 | 49.315 |
| Total | | 4736351 | 76979 | 100.000 | 100.000 |

Racemic **4I**

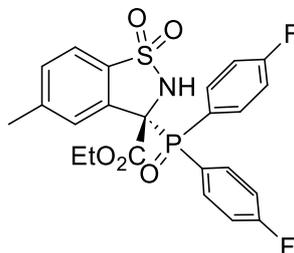


Peak Table

| Detector A 254nm | | | | | |
|------------------|-----------|----------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 10.686 | 38261 | 1022 | 0.177 | 0.103 |
| 2 | 15.591 | 37154116 | 576604 | 99.823 | 99.897 |
| Total | | 37192377 | 577626 | 100.000 | 100.000 |

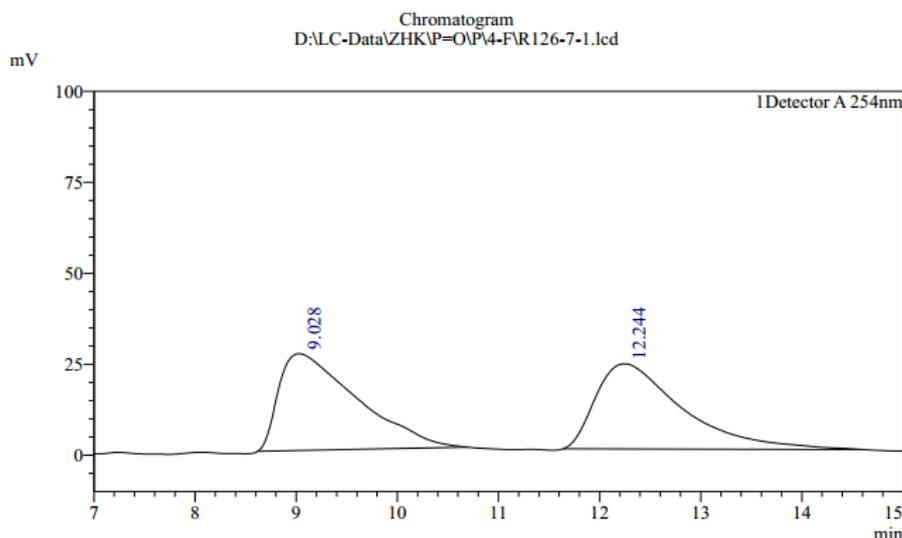
Enantiomerically enriched **4l**

(S)-ethyl -(bis(4-fluorophenyl)phosphoryl)-5-methyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (4m)



A white solid; m.p. = 163 - 167 °C; 46.2 mg, 94% yield; $[\alpha]_D^{25} = -47.67$ (*c* 1.3, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.17 - 8.05 (m, 3H), 7.67 - 7.55 (m, 2H), 7.49 (d, *J* = 8.0 Hz, 1H), 7.39 (d, *J* = 8.0 Hz, 1H), 7.31 - 7.22 (m, 2H), 7.02 - 6.93 (m, 2H), 5.94 (d, *J* = 4.4 Hz, 1H), 4.32 - 4.15 (m, 2H), 2.53 (s, 3H), 1.17 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 167.17 (dd, *J* = 13.3, 3.3 Hz), 166.53 (d, *J* = 2.7 Hz), 164.63 (dd, *J* = 12.7, 3.5 Hz), 144.97 (d, *J* = 2.0 Hz), 135.27 (q, *J* = 9.8 Hz), 132.17 (d, *J* = 1.9 Hz), 131.82 (d, *J* = 3.2 Hz), 131.24 (d, *J* = 2.3 Hz), 128.38 (d, *J* = 2.6 Hz), 124.21 (d, *J* = 3.3 Hz), 123.23 (dd, *J* = 6.9, 3.6 Hz), 122.14 (d, *J* = 3.5 Hz), 121.18 (d, *J* = 1.6 Hz), 116.53 (dd, *J* = 21.3, 12.8 Hz), 115.90 (dd, *J* = 21.4, 13.5 Hz), 70.14 (d, *J* = 73.6 Hz), 64.78, 22.09, 14.01; ³¹P NMR (162 MHz, CDCl₃) δ 26.92; ¹⁹F NMR (376 MHz, CDCl₃) δ -

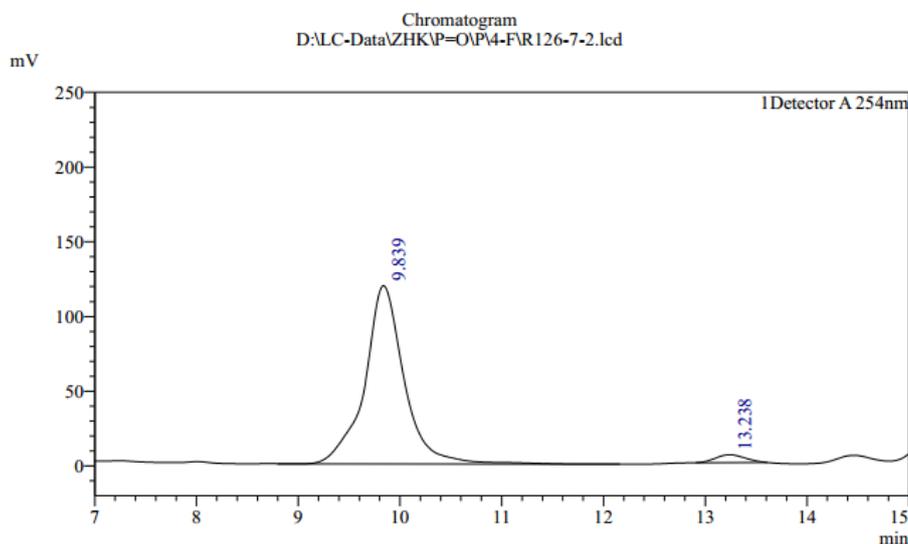
103.99; HRMS (ESI) m/z calcd for $C_{23}H_{20}NO_5PSF_2$ $[M+H]^+$ = 492.0846, found = 492.0849; The ee value was 94%, t_R (minor) = 9.8 min, t_R (major) = 13.2 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 9.028 | 1426741 | 26615 | 53.224 | 50.596 |
| 2 | 12.244 | 1393144 | 23391 | 46.776 | 49.404 |
| Total | | 2819885 | 50006 | 100.000 | 100.000 |

Racemic **4m**

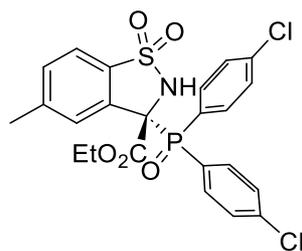


Peak Table

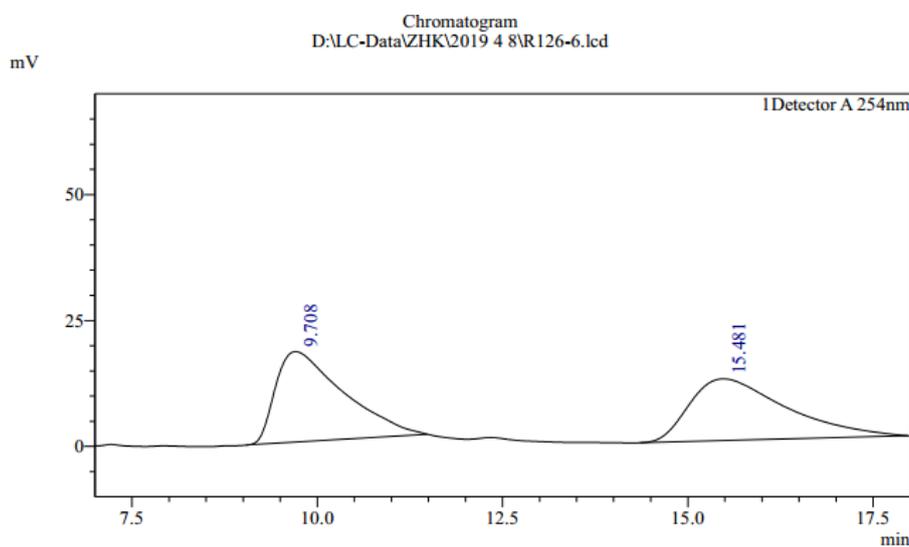
| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 9.839 | 3219598 | 119272 | 95.810 | 96.836 |
| 2 | 13.238 | 105200 | 5216 | 4.190 | 3.164 |
| Total | | 3324798 | 124488 | 100.000 | 100.000 |

Enantiomerically enriched **4m**

(S)-ethyl 3-(bis(4-chlorophenyl)phosphoryl)-5-methyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (4n)



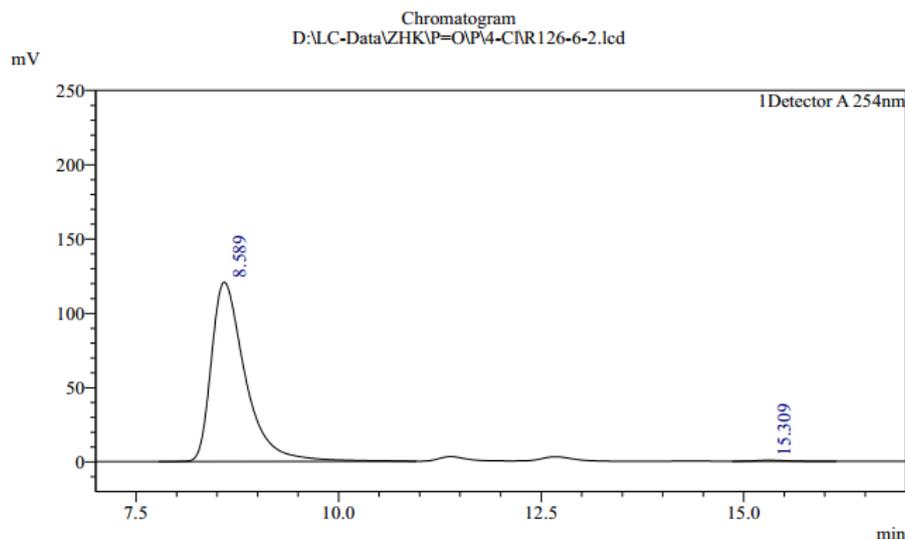
A white solid; m.p. = 148 - 150 °C; 48.6 mg, 93% yield; $[\alpha]_D^{25} = -44.80$ (*c* 1.4, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.11 (s, 1H), 8.06 - 7.97 (m, 2H), 7.59 - 7.47 (m, 5H), 7.38 (d, *J* = 8.0 Hz, 1H), 7.30 - 7.22 (m, 2H), 6.05 (d, *J* = 4.0 Hz, 1H), 4.30 - 4.4.13 (m, 2H), 2.51 (s, 3H), 1.17 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.34 (d, *J* = 2.8 Hz), 144.97 (d, *J* = 1.9 Hz), 140.24 (d, *J* = 3.6 Hz), 139.94 (d, *J* = 3.6 Hz), 133.94 (dd, *J* = 19.6, 9.4 Hz), 132.18 (d, *J* = 1.7 Hz), 131.76 (d, *J* = 3.3 Hz), 130.99 (d, *J* = 2.4 Hz), 129.36 (d, *J* = 12.4 Hz), 128.76 (d, *J* = 12.9 Hz), 128.32 (d, *J* = 2.5 Hz), 126.64, 125.66 (d, *J* = 2.2 Hz), 124.60, 121.18 (d, *J* = 1.6 Hz), 69.93 (d, *J* = 73.9 Hz), 64.75, 22.05, 13.97; ³¹P NMR (162 MHz, CDCl₃) δ 26.89; HRMS (ESI) *m/z* calcd for C₂₃H₂₀NO₅PSCl₂ [M+H]⁺ = 524.0255, found = 524.0253; The ee value was 99%, *t_R* (minor) = 8.6 min, *t_R* (major) = 15.3 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 9.708 | 1137330 | 17981 | 59.456 | 50.661 |
| 2 | 15.481 | 1107634 | 12262 | 40.544 | 49.339 |
| Total | | 2244964 | 30243 | 100.000 | 100.000 |

Racemic **4n**

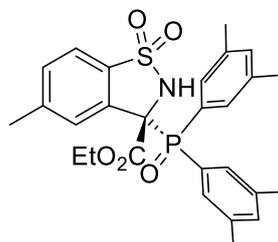


Peak Table

| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 8.589 | 3527411 | 120798 | 99.410 | 99.437 |
| 2 | 15.309 | 19958 | 717 | 0.590 | 0.563 |
| Total | | 3547369 | 121515 | 100.000 | 100.000 |

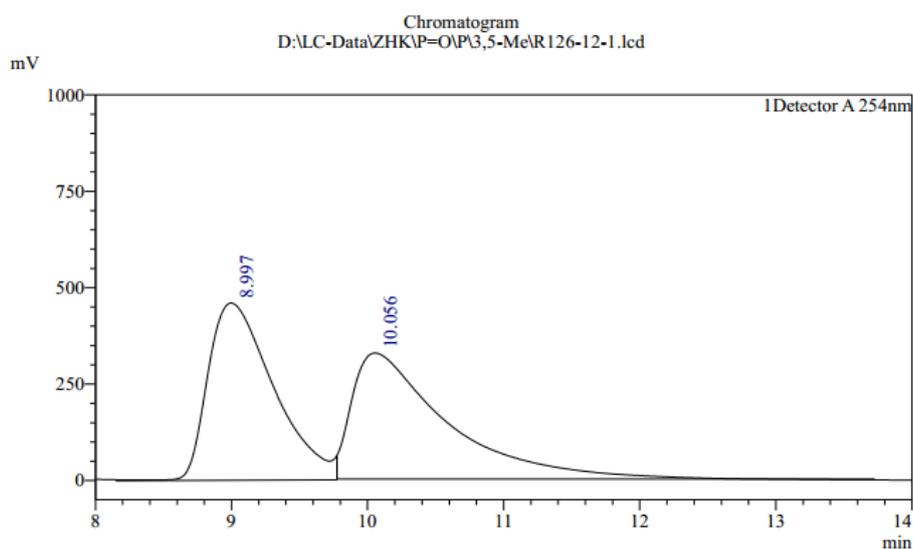
Enantiomerically enriched **4n**

(S)-ethyl 3-(bis(3,5-dimethylphenyl)phosphoryl)-5-methyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (4o)



A white solid; m.p. = 155 - 157 °C; 47.0 mg, 92% yield; $[\alpha]_D^{25} = -35.00$ (*c* 1.3, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.12 (s, 1H), 7.63 (d, *J* = 11.2 Hz, 2H), 7.48 (d, *J* = 8.0 Hz, 1H), 7.34 (d, *J* = 8.0 Hz, 1H), 7.25 - 7.16 (m, 3H), 7.03 (s, 1H), 5.79 (d, *J* = 3.2 Hz, 1H), 4.31 - 4.13 (m, 2H), 2.50 (s, 3H), 2.39 (s, 6H), 2.18 (s, 6H), 1.16 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.73 (d, *J* = 2.9 Hz), 144.60 (d, *J* = 1.8 Hz), 138.53 (d, *J* = 12.5 Hz), 137.87 (d, *J* = 13.0 Hz), 134.82 (d, *J* = 3.0 Hz), 134.45 (d, *J* = 2.9 Hz), 131.84 (dd, *J* = 8.1, 2.5 Hz), 131.69 (d, *J* = 1.7 Hz), 130.25 (d, *J* = 8.6 Hz), 130.00 (d, *J* = 8.4 Hz), 128.52 (d, *J* = 2.5 Hz), 128.36, 127.34 (d, *J* = 11.3 Hz), 126.25, 120.77 (d, *J* = 1.4 Hz), 70.11 (d, *J* = 70.0 Hz), 64.38, 22.02, 21.52, 21.24, 13.89; ³¹P NMR (162 MHz, CDCl₃) δ 29.24; HRMS (ESI) *m/z* calcd for C₂₇H₃₀NO₅PS [M+H]⁺ =

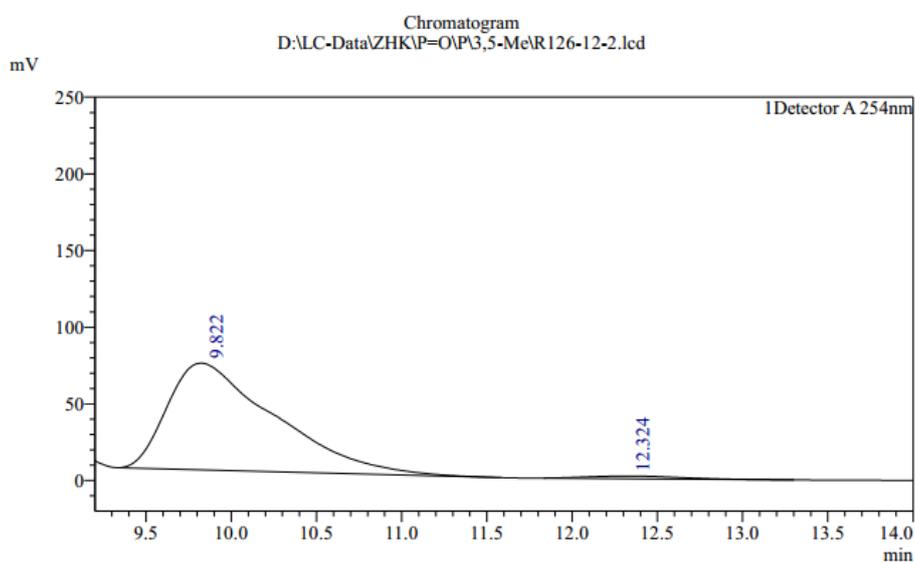
512.1661, found = 512.1162; The ee value was 96%, t_R (minor) = 9.8 min, t_R (major) = 12.3 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|--------|---------|---------|
| 1 | 8.997 | 15426703 | 459541 | 58.455 | 49.477 |
| 2 | 10.056 | 15752627 | 326607 | 41.545 | 50.523 |
| Total | | 31179330 | 786149 | 100.000 | 100.000 |

Racemic **4o**

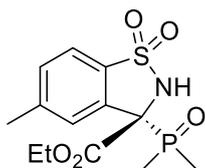


Peak Table

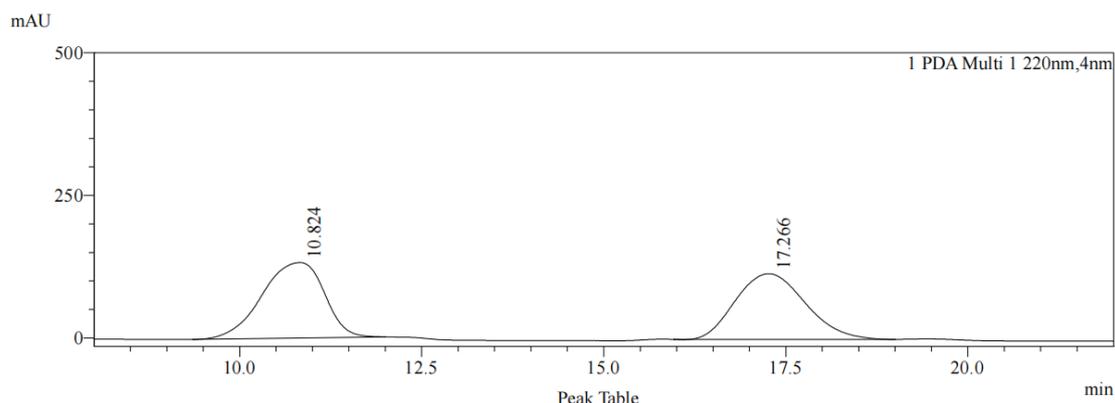
| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 9.822 | 3052516 | 69569 | 97.583 | 97.925 |
| 2 | 12.324 | 64683 | 1723 | 2.417 | 2.075 |
| Total | | 3117199 | 71292 | 100.000 | 100.000 |

Enantiomerically enriched **4o**

(S)-ethyl 3-(dimethylphosphoryl)-5-methyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (4p)

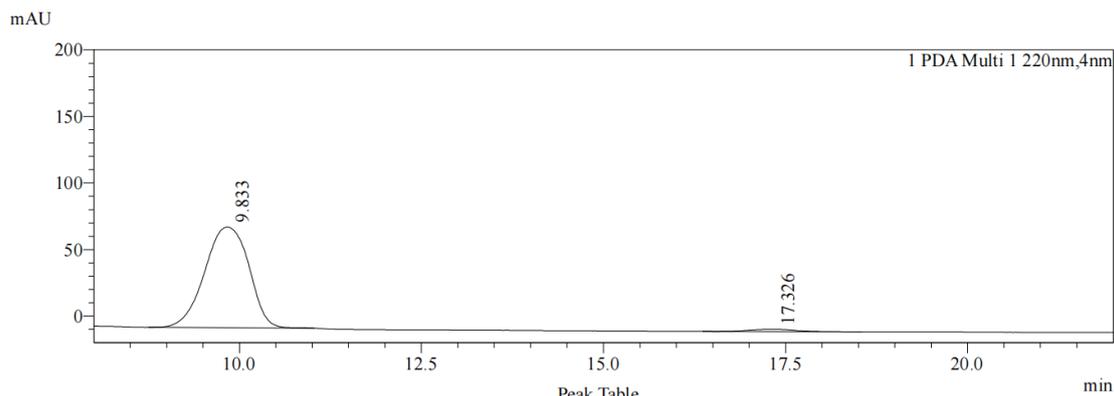


A white solid; m.p. = 94 - 97 °C; 31.4 mg, 95% yield; $[\alpha]_D^{25} = -10.68$ (c 0.6, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.96 (s, 1H), 7.66 (d, $J = 8.0$ Hz, 1H), 7.46 (d, $J = 8.0$ Hz, 1H), 4.49 - 4.36 (m, 2H), 2.52 (s, 3H), 1.63 (d, $J = 12.8$ Hz, 3H), 1.41 - 1.35 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 166.71 (d, $J = 1.5$ Hz), 145.35 (d, $J = 1.6$ Hz), 132.26 (d, $J = 1.8$ Hz), 131.74 (d, $J = 3.0$ Hz), 131.19 (d, $J = 1.8$ Hz), 127.97 (d, $J = 2.2$ Hz), 121.41 (d, $J = 1.2$ Hz), 68.74 (d, $J = 67.8$ Hz), 64.85, 22.12, 14.18, 13.25 (d, $J = 68.4$ Hz), 12.32 (d, $J = 71.6$ Hz); ^{31}P NMR (162 MHz, CDCl_3) δ 51.96; HRMS (ESI) m/z calcd for $\text{C}_{13}\text{H}_{18}\text{NO}_5\text{PS}$ $[\text{M}+\text{Na}]^+ = 354.0541$, found = 354.0546; The ee value was 96%, t_R (major) = 9.8 min, t_R (minor) = 17.3 min (Chiralcel IC, $\lambda = 220$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min)



| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|----------|---------|
| 1 | 10.824 | 132325 | 53.501 | 7895435 | 50.692 |
| 2 | 17.266 | 115009 | 46.499 | 7679829 | 49.308 |
| Total | | 247334 | 100.000 | 15575264 | 100.000 |

Racemic 4p



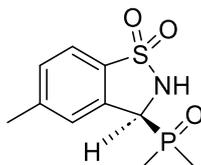
Peak Table

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|---------|---------|
| 1 | 9.833 | 75626 | 97.827 | 3194965 | 97.795 |
| 2 | 17.326 | 1680 | 2.173 | 72039 | 2.205 |
| Total | | 77306 | 100.000 | 3267005 | 100.000 |

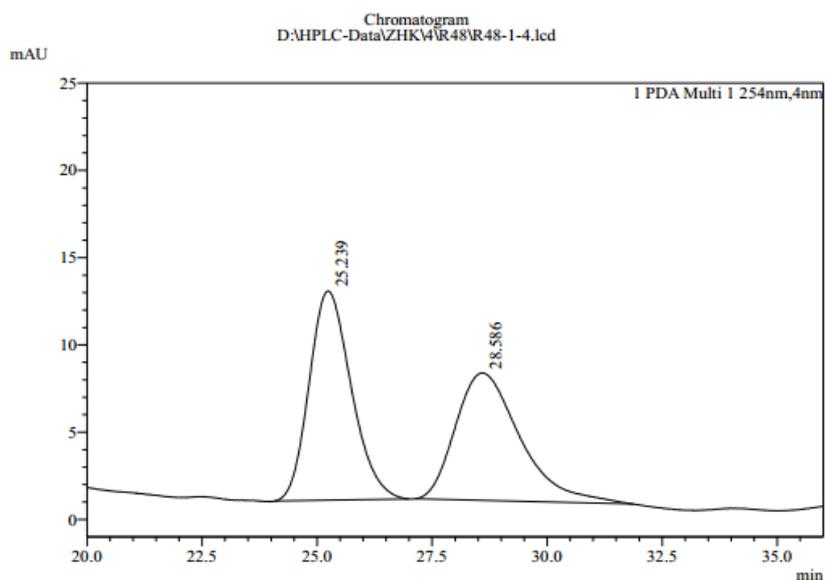
Enantiomerically enriched **4p**

(R)-3-(dimethylphosphoryl)-5-methyl-2,3-dihydrobenzo[d]isothiazole 1,1-dioxide

(4q)



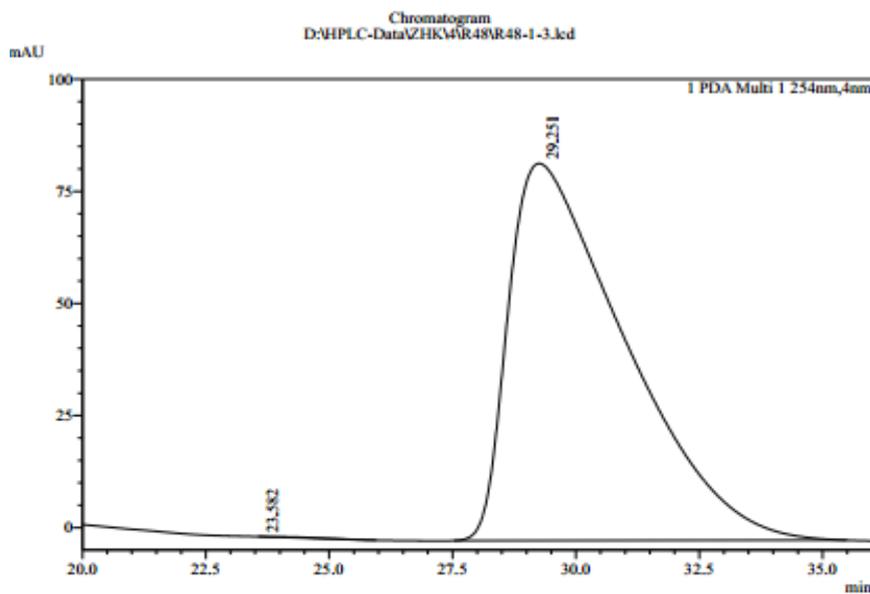
A white solid; m.p. = 85 - 87 °C; 23.6 mg, 91% yield; $[\alpha]_D^{25} = -15.00$ (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, DMSO) δ 7.75 (d, *J* = 8.0 Hz, 1H), 7.54 (s, 1H), 7.46 (d, *J* = 8.0 Hz, 1H), 5.11 (d, *J* = 9.2 Hz, 1H), 2.44 (s, 3H), 1.58 (d, *J* = 13.2 Hz, 3H), 1.09 (d, *J* = 12.8 Hz, 3H); ¹³C NMR (100 MHz, DMSO) δ 143.32 (d, *J* = 2.0 Hz), 134.05 (d, *J* = 2.3 Hz), 132.59 (d, *J* = 9.8 Hz), 130.64 (d, *J* = 1.4 Hz), 125.89 (d, *J* = 2.2 Hz), 120.79 (d, *J* = 1.3 Hz), 55.91 (d, *J* = 75.5 Hz), 21.28, 14.63 (d, *J* = 67.9 Hz), 10.65 (d, *J* = 67.4 Hz); ³¹P NMR (162 MHz, DMSO) δ 46.20; HRMS (ESI) *m/z* calcd for C₁₀H₁₄NO₃PS [M+Na]⁺ = 282.0330, found = 282.0327; The ee value was 99%, *t_R* (minor) = 23.6 min, *t_R* (major) = 29.3 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| PDA Ch1 254nm | | | | | |
|---------------|-----------|--------|---------|---------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 25.239 | 11983 | 62.126 | 762490 | 51.730 |
| 2 | 28.586 | 7305 | 37.874 | 711479 | 48.270 |
| Total | | 19288 | 100.000 | 1473969 | 100.000 |

Racemic **4q**

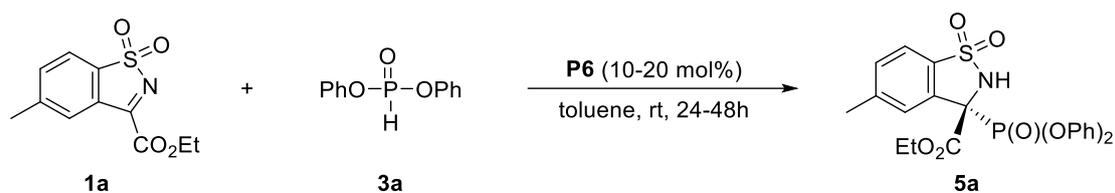


Peak Table

| PDA Ch1 254nm | | | | | |
|---------------|-----------|--------|---------|----------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 23.582 | 9 | 0.010 | 7928 | 0.057 |
| 2 | 29.251 | 84078 | 99.990 | 13823393 | 99.943 |
| Total | | 84087 | 100.000 | 13831321 | 100.000 |

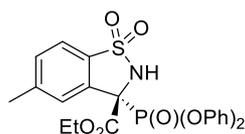
Enantiomerically enriched **4q**

5.2 Representative procedure for asymmetric P-nucleophile addition to cyclic N-sulfonyl imines **1/1'** with secondary phosphine oxides **3**

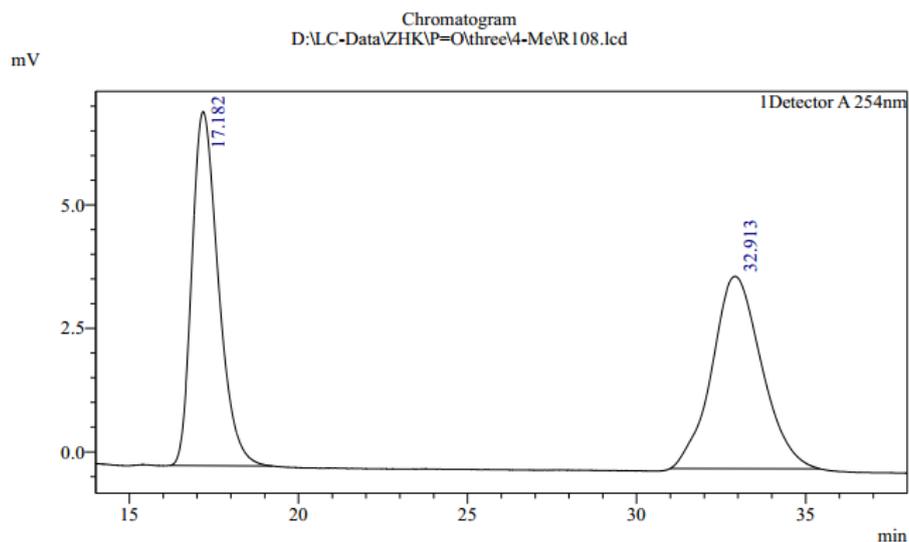


Representative procedure 5a: To a flame-dried round bottle flask with a magnetic stirring bar were added cyclic ketimine **1a** (0.10 mmol, 25.3mg), Phosphine Oxide **3a** (0.12 mmol, 28.1 mg), phosphonium salt **P4** (0.01 mmol, 5.7 mg), and toluene (2.0 mL). The reaction mixture was stirred at room temperature for 24 h. The solvent was removed under reduced pressure, and the residue was purified by column chromatography on silica gel (CH₂Cl₂/ethyl acetate = 3:1) to afford **5a** (93% yield) as a white solid.

(S)-ethyl 3-(diphenoxyphosphoryl)-5-methyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5a)



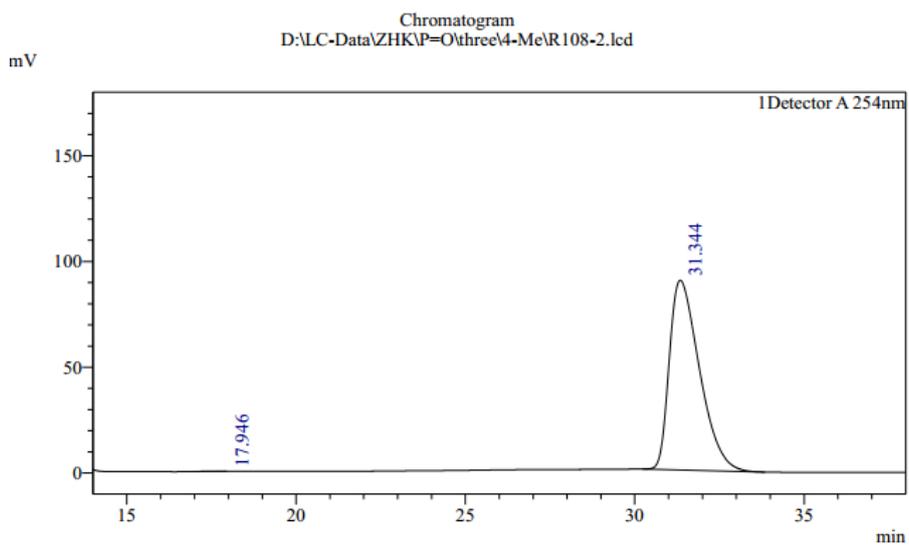
A white solid; m.p. = 135 - 137 °C; 45.3 mg, 93% yield; $[\alpha]^{25}_D = -14.37$ (*c* 0.3, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.95 - 7.87 (m, 1H), 7.70 (d, *J* = 8.0 Hz, 1H), 7.43 (d, *J* = 8.0 Hz, 1H), 7.35 - 7.27 (m, 2H), 7.24 - 7.14 (m, 5H), 7.11 - 7.05 (m, 1H), 7.01 - 6.94 (m, 2H), 6.09 (d, *J* = 5.2 Hz, 1H), 4.46 - 4.24 (m, 2H), 2.44 (s, 3H), 1.29 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.36 (d, *J* = 4.5 Hz), 150.50 (d, *J* = 9.6 Hz), 150.18 (d, *J* = 10.0 Hz), 145.02 (d, *J* = 2.7 Hz), 132.46 (d, *J* = 2.6 Hz), 132.14 (d, *J* = 5.1 Hz), 130.25 (d, *J* = 4.3 Hz), 129.91 (d, *J* = 0.6 Hz), 129.71, 127.86 (d, *J* = 3.0 Hz), 125.89 (d, *J* = 1.0 Hz), 125.57 (d, *J* = 0.7 Hz), 121.59 (d, *J* = 2.2 Hz), 120.58 (d, *J* = 4.4 Hz), 120.17 (d, *J* = 4.4 Hz), 67.62 (d, *J* = 166.5 Hz), 65.04, 22.01, 13.97; ³¹P NMR (162 MHz, CDCl₃) δ 4.32; HRMS (ESI) *m/z* calcd for C₂₃H₂₂NO₇PS [M+Na]⁺ = 510.0747, found = 510.0740; The ee value was >99%, *t_R* (minor) = 17.9 min, *t_R* (major) = 31.3 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Detector A 254nm | | | | | |
|------------------|-----------|--------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 17.182 | 389504 | 7168 | 64.810 | 49.411 |
| 2 | 32.913 | 398787 | 3892 | 35.190 | 50.589 |
| Total | | 788291 | 11060 | 100.000 | 100.000 |

Racemic **5a**

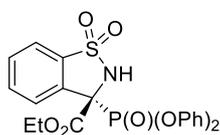


Peak Table

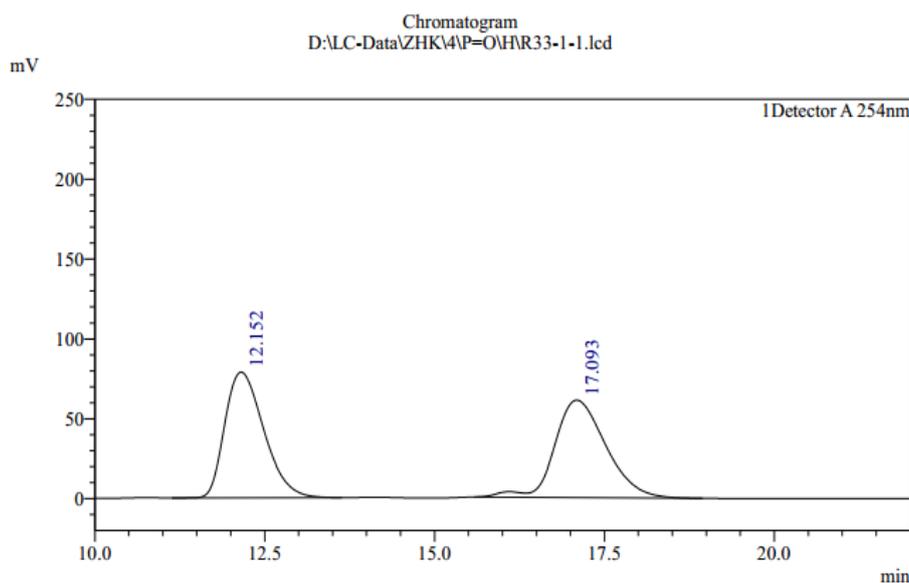
| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 17.946 | 1867 | 0 | 0.000 | 0.034 |
| 2 | 31.344 | 5541532 | 89720 | 100.000 | 99.966 |
| Total | | 5543398 | 89720 | 100.000 | 100.000 |

Enantiomerically enriched **5a**

(S)-ethyl 3-(diphenoxyphosphoryl)-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5b)



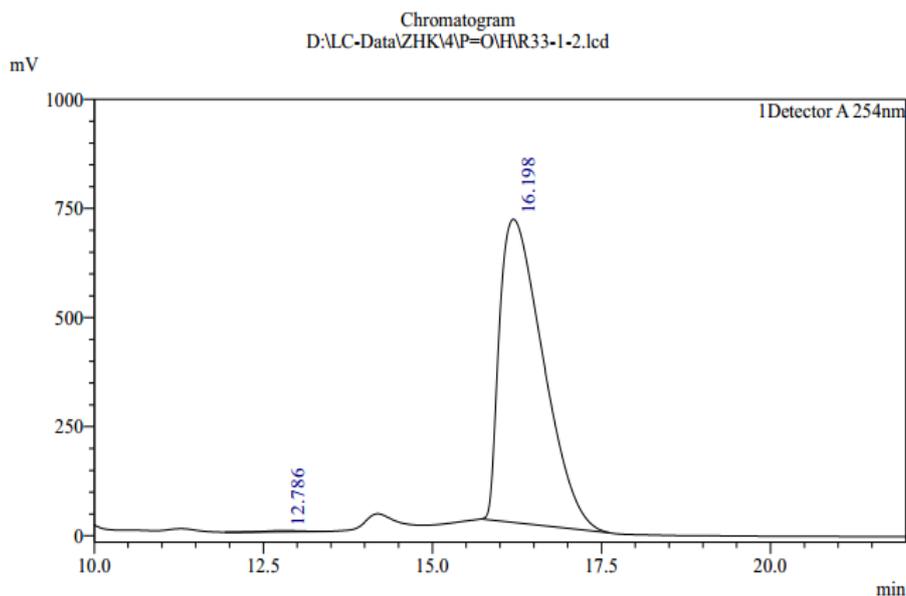
A white solid; m.p. = 98 - 100 °C; 45.4 mg, 96% yield; $[\alpha]_D^{25} = -32.18$ (c 0.6, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.23 - 8.11 (m, 1H), 7.88 - 7.79 (m, 1H), 7.72 - 7.59 (m, 2H), 7.34 - 7.26 (m, 2H), 7.22 - 7.12 (m, 5H), 7.11 - 7.05 (m, 1H), 7.01 - 6.92 (m, 2H), 6.22 (d, $J = 4.8$ Hz, 1H), 4.40 - 4.27 (m, 2H), 1.28 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 165.27 (d, $J = 4.5$ Hz), 150.24 (dd, $J = 26.7, 9.7$ Hz), 134.79 (d, $J = 5.0$ Hz), 134.37 (d, $J = 18.5$ Hz), 133.71 (d, $J = 2.7$ Hz), 131.49 (d, $J = 2.7$ Hz), 130.00 (d, $J = 4.5$ Hz), 129.89 (d, $J = 0.7$ Hz), 129.74, 127.76 (d, $J = 2.9$ Hz), 125.89 (d, $J = 1.0$ Hz), 125.61 (d, $J = 0.7$ Hz), 121.84 (d, $J = 2.2$ Hz), 120.56 (d, $J = 4.4$ Hz), 120.15 (d, $J = 4.4$ Hz), 67.77 (d, $J = 167.0$ Hz), 65.10, 13.92; ^{31}P NMR (162 MHz, CDCl_3) δ 4.33; HRMS (ESI) m/z calcd for $\text{C}_{22}\text{H}_{20}\text{NO}_7\text{PS}$ $[\text{M}+\text{Na}]^+ = 496.0590$, found = 496.0588; The ee value was >99%, t_R (minor) = 12.8 min, t_R (major) = 16.2 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 12.152 | 3144566 | 78698 | 56.327 | 49.474 |
| 2 | 17.093 | 3211375 | 61019 | 43.673 | 50.526 |
| Total | | 6355941 | 139717 | 100.000 | 100.000 |

Racemic **5b**

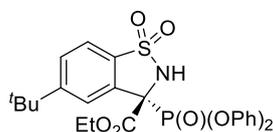


Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|--------|---------|---------|
| 1 | 12.786 | 75411 | 2330 | 0.334 | 0.242 |
| 2 | 16.198 | 31108919 | 694780 | 99.666 | 99.758 |
| Total | | 31184329 | 697111 | 100.000 | 100.000 |

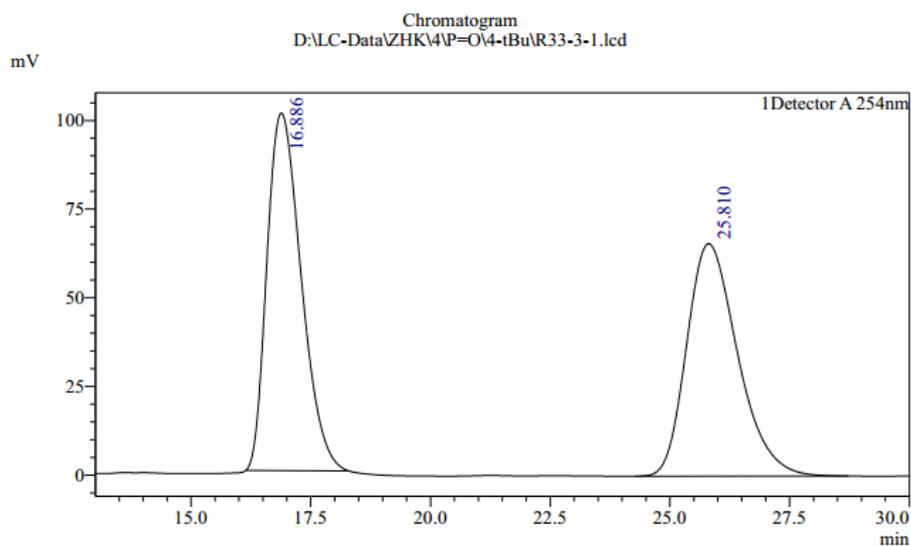
Enantiomerically enriched **5b**

(S)-ethyl 5-(tert-butyl)-3-(diphenoxyphosphoryl)-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5c)



A white solid; m.p. = 106 - 107 °C; 48.7 mg, 92% yield; $[\alpha]_D^{25} = -45.17$ (*c* 0.6, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.18 - 8.11 (m, 1H), 7.75 (dd, *J* = 8.4, 0.4 Hz, 1H), 7.68 - 7.62 (m, 1H), 7.35 - 7.26 (m, 2H), 7.22 - 7.13 (m, 5H), 7.10 - 7.03 (m, 1H), 6.98 - 6.90 (m, 2H), 6.08 (s, 1H), 4.46 - 4.25 (m, 2H), 1.32 - 1.26 (m, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 165.43 (d, *J* = 4.9 Hz), 158.11 (d, *J* = 2.4 Hz), 150.59 (d, *J* = 9.4 Hz), 150.14 (d, *J* = 10.0 Hz), 131.95 (d, *J* = 5.0 Hz), 129.94, 129.88, 129.67, 128.92 (d, *J* = 2.3 Hz), 125.93 (d, *J* = 0.9 Hz), 125.52, 124.85 (d, *J* = 3.0 Hz), 121.43 (d, *J* = 2.1 Hz), 120.63 (d, *J* = 4.4 Hz), 120.15 (d, *J* = 4.4 Hz), 67.73 (d, *J* = 166.1 Hz), 64.92, 35.70, 31.17, 14.01; ³¹P NMR (162 MHz, CDCl₃) δ 4.41; HRMS (ESI) *m/z* calcd for C₂₆H₂₈NO₇PS [M+Na]⁺ = 552.1222, found = 552.1226; The ee value was >99%, *t_R*

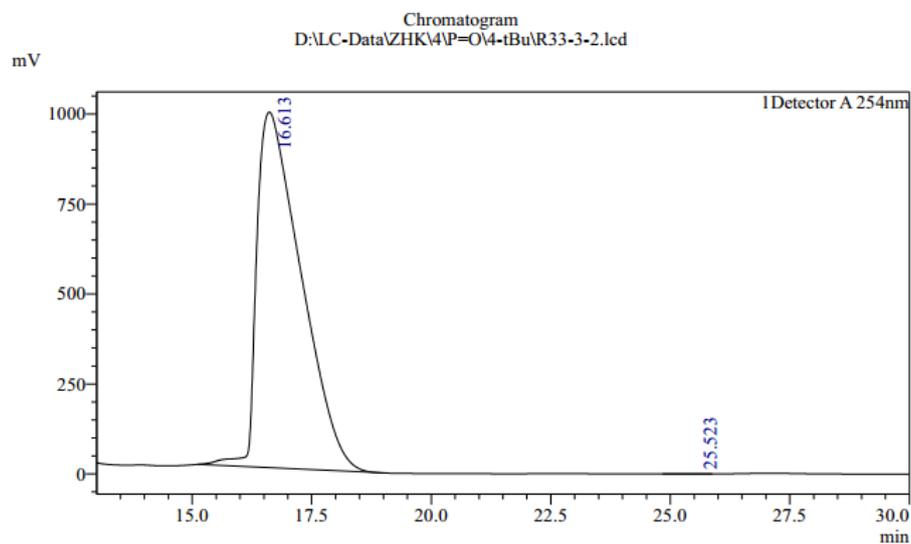
(major) = 16.6 min, t_R (minor) = 25.5 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 16.886 | 5003271 | 100797 | 60.600 | 50.801 |
| 2 | 25.810 | 4845440 | 65535 | 39.400 | 49.199 |
| Total | | 9848712 | 166332 | 100.000 | 100.000 |

Racemic 5c

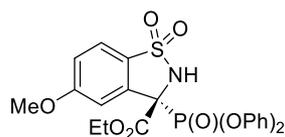


Peak Table

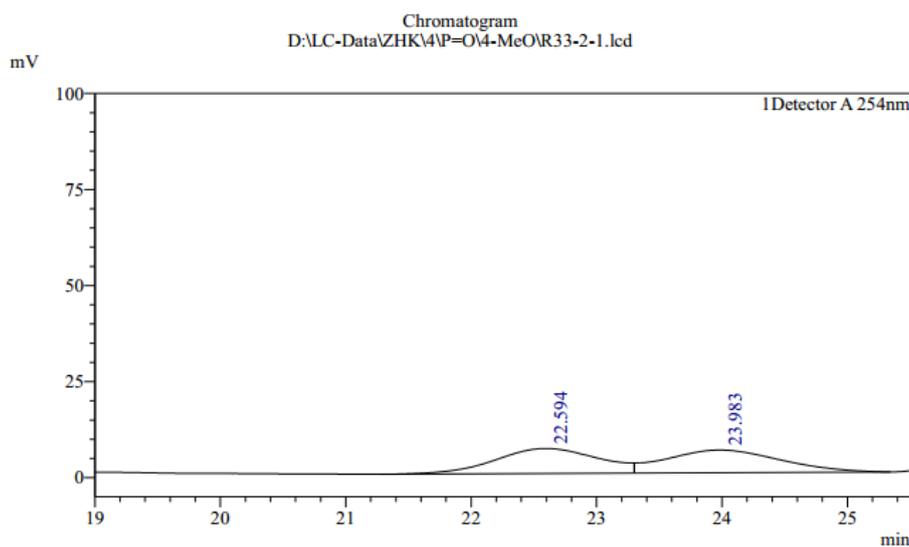
| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|--------|---------|---------|
| 1 | 16.613 | 63677188 | 988092 | 99.988 | 99.993 |
| 2 | 25.523 | 4254 | 123 | 0.012 | 0.007 |
| Total | | 63681442 | 988215 | 100.000 | 100.000 |

Enantiomerically enriched 5c

(S)-ethyl 3-(diphenoxyphosphoryl)-5-methoxy-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5d)



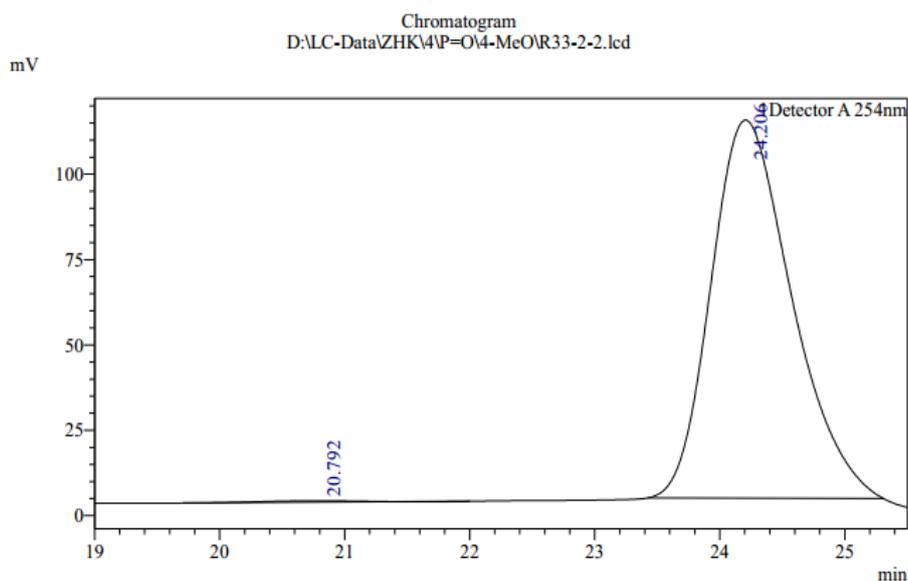
A white solid; m.p. = 107 - 110 °C; 48.8 mg, 97% yield; $[\alpha]_D^{25} = -16.67$ (c 0.6, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.71 (dd, $J = 8.4, 0.8$ Hz, 1H), 7.58 - 7.52(m, 1H), 7.34 - 7.29 (m, 2H), 7.23 - 7.16 (m, 5H), 7.14 - 7.08 (m, 2H), 7.04 - 6.98 (m, 2H), 6.05 (s, 1H), 4.41 - 4.28 (m, 2H), 3.84 (s, 3H), 1.30 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 165.26 (d, $J = 4.3$ Hz), 163.87 (d, $J = 2.5$ Hz), 150.55 (d, $J = 9.8$ Hz), 150.19 (d, $J = 9.6$ Hz), 132.54 (d, $J = 4.4$ Hz), 129.86 (d, $J = 19.5\text{Hz}$), 126.80 (d, $J = 4.9$ Hz), 125.95, 125.63, 123.16 (d, $J = 2.1$ Hz), 120.96, 120.61 (d, $J = 4.4$ Hz), 120.22 (d, $J = 4.4$ Hz), 118.78 (d, $J = 2.3$ Hz), 111.55 (d, $J = 2.7$ Hz), 67.46 (d, $J = 166.3$ Hz), 65.08, 56.16, 14.02; ^{31}P NMR (162 MHz, CDCl_3) δ 4.22; HRMS (ESI) m/z calcd for $\text{C}_{23}\text{H}_{22}\text{NO}_8\text{PS}$ $[\text{M}+\text{Na}]^+ = 526.0696$, found = 526.0691; The ee value was >99%, t_R (minor) = 20.8 min, t_R (major) = 24.2 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|--------|--------|---------|---------|
| 1 | 22.594 | 373113 | 6514 | 52.509 | 49.991 |
| 2 | 23.983 | 373254 | 5891 | 47.491 | 50.009 |
| Total | | 746367 | 12405 | 100.000 | 100.000 |

Racemic **5d**

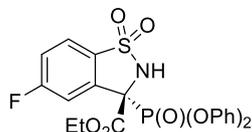


Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 20.792 | 21146 | 417 | 0.375 | 0.424 |
| 2 | 24.206 | 4961653 | 110722 | 99.625 | 99.576 |
| Total | | 4982799 | 111139 | 100.000 | 100.000 |

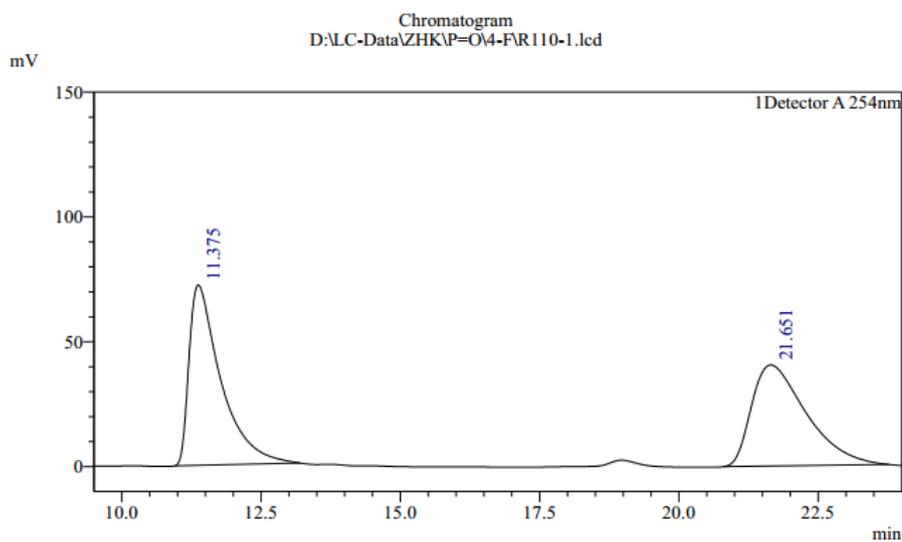
Enantiomerically enriched **5d**

(S)-ethyl 3-(diphenoxyphosphoryl)-5-fluoro-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5e)



A white solid; m.p. = 105 - 107 °C; 46.2 mg, 94% yield; $[\alpha]_D^{25} = -42.37$ (c 0.9, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.88 - 7.78 (m, 2H), 7.37 - 7.27 (m, 3H), 7.25 - 7.09 (m, 6H), 7.07 - 7.01 (m, 2H), 6.35 (d, *J* = 4.4 Hz, 1H), 4.39 - 4.30 (m, 2H), 1.29 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.72 (d, *J* = 3.2 Hz), 164.75 (d, *J* = 3.9 Hz), 164.17 (d, *J* = 3.1 Hz), 150.14 (dd, *J* = 25.6, 9.6 Hz), 133.14 (dd, *J* = 10.3, 4.6 Hz), 130.86 (dd, *J* = 4.9, 2.7 Hz), 129.93 (d, *J* = 0.6 Hz), 129.85, 125.89 (dd, *J* = 19.5, 1.0 Hz), 124.04 (dd, *J* = 10.0, 2.3 Hz), 120.48 (d, *J* = 4.4 Hz), 120.08 (d, *J* = 4.4 Hz), 119.72 (d, *J* = 2.4 Hz), 119.48 (d, *J* = 2.4 Hz), 115.22 (d, *J* = 2.7 Hz), 114.96 (d, *J* = 2.7 Hz), 67.26 (dd, *J* = 167.0, 2.2 Hz), 65.37, 13.89; ³¹P NMR (162 MHz, CDCl₃) δ 3.64; ¹⁹F NMR (376 MHz, CDCl₃) δ -102.22; HRMS (ESI) *m/z* calcd for C₂₂H₁₉NO₇PSF [M+Na]⁺ = 514.0491, found = 514.0487; The ee value was >99%, *t_R* (minor) = 12.8

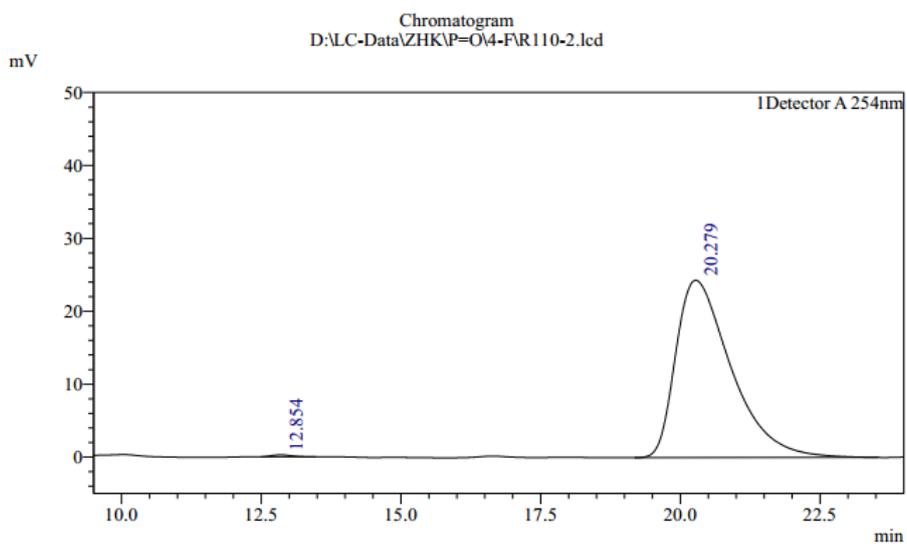
min, t_R (major) = 20.3 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 11.375 | 2866751 | 72190 | 64.055 | 50.875 |
| 2 | 21.651 | 2768088 | 40510 | 35.945 | 49.125 |
| Total | | 5634839 | 112700 | 100.000 | 100.000 |

Racemic 5e

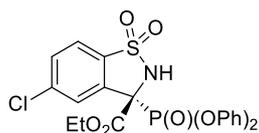


Peak Table

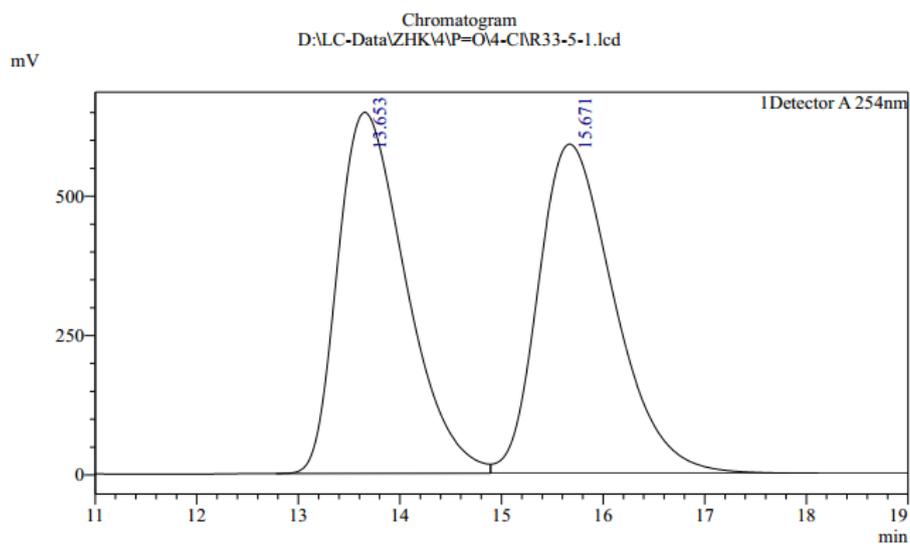
| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 12.854 | 6244 | 257 | 1.044 | 0.371 |
| 2 | 20.279 | 1678689 | 24328 | 98.956 | 99.629 |
| Total | | 1684933 | 24584 | 100.000 | 100.000 |

Enantiomerically enriched 5e

(S)-ethyl 5-chloro-3-(diphenoxyphosphoryl)-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5f)



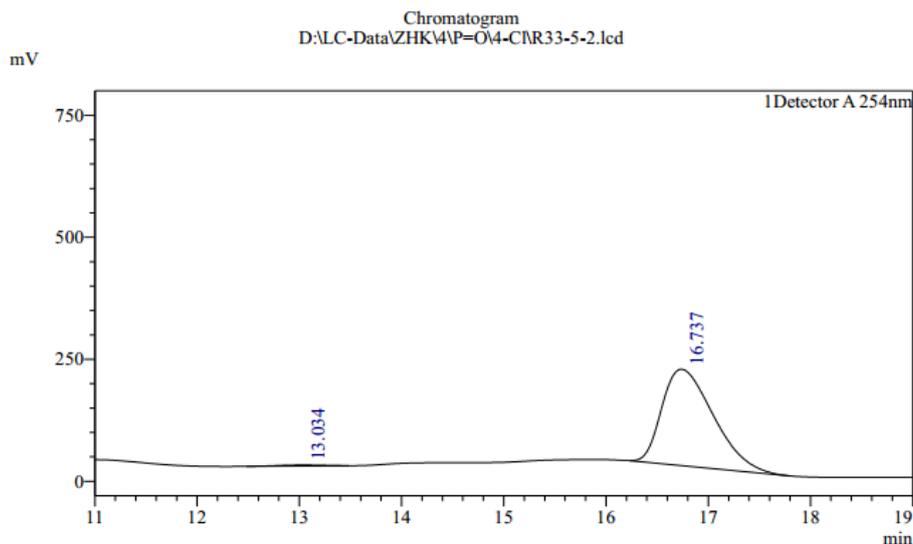
A white solid; m.p. = 92 - 94 °C; 48.2 mg, 95% yield; $[\alpha]_D^{25} = -31.83$ (c 0.6, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.17 - 8.09 (m, 1H), 7.80 - 7.72 (m, 1H), 7.61 - 7.58 (m, 1H), 7.31 - 7.28 (m, 2H), 7.27 - 7.20 (m, 2H), 7.20 - 7.10 (m, 4H), 7.08 - 7.02 (m, 2H), 4.42 - 4.30 (m, 2H), 1.30 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 164.70 (d, $J = 3.8$ Hz), 150.28 (d, $J = 9.5$ Hz), 150.02 (d, $J = 10.1$ Hz), 140.24 (d, $J = 3.2$ Hz), 133.23 (d, $J = 4.9$ Hz), 132.02 (d, $J = 4.5$ Hz), 131.93 (d, $J = 2.7$ Hz), 129.83 (d, $J = 7.7$ Hz), 127.86 (d, $J = 2.8$ Hz), 125.92, 125.72, 122.87 (d, $J = 2.2$ Hz), 120.39 (d, $J = 4.4$ Hz), 119.98 (d, $J = 4.4$ Hz), 67.27 (d, $J = 165.8$ Hz), 65.34, 13.85; ^{31}P NMR (162 MHz, CDCl_3) δ 3.44; HRMS (ESI) m/z calcd for $\text{C}_{22}\text{H}_{19}\text{NO}_7\text{PSCl}$ $[\text{M}+\text{Na}]^+ = 530.0192$, found = 530.0195; The ee value was 98%, t_R (minor) = 13.0 min, t_R (major) = 16.8 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|---------|---------|---------|
| 1 | 13.653 | 30154226 | 647798 | 52.306 | 49.565 |
| 2 | 15.671 | 30683470 | 590679 | 47.694 | 50.435 |
| Total | | 60837695 | 1238477 | 100.000 | 100.000 |

Racemic **5f**

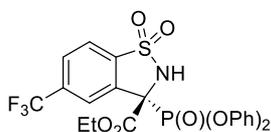


Peak Table

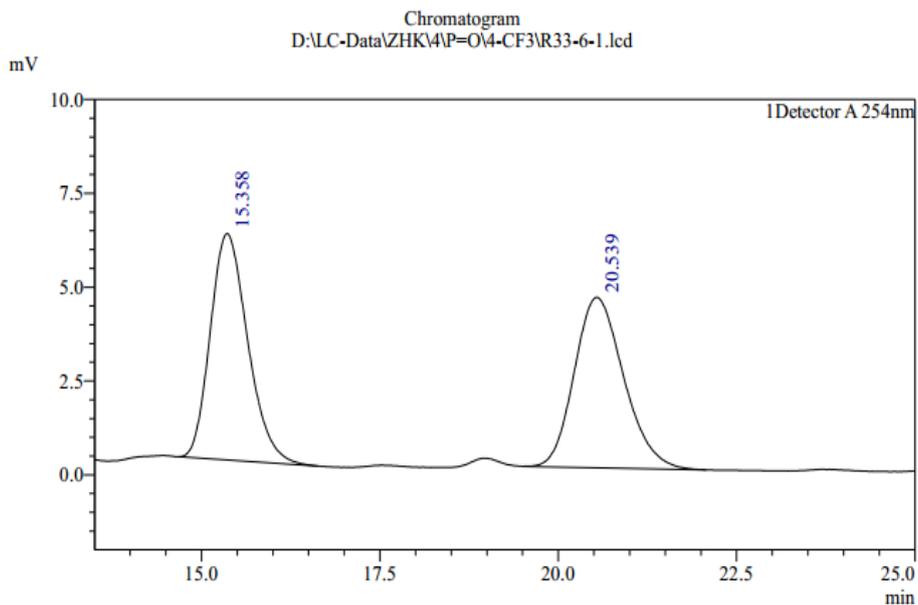
| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 13.034 | 63764 | 2000 | 1.003 | 0.916 |
| 2 | 16.737 | 6895016 | 197399 | 98.997 | 99.084 |
| Total | | 6958779 | 199399 | 100.000 | 100.000 |

Enantiomerically enriched **5f**

(S)-ethyl 3-(diphenoxyphosphoryl)-5-(trifluoromethyl)-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5g)



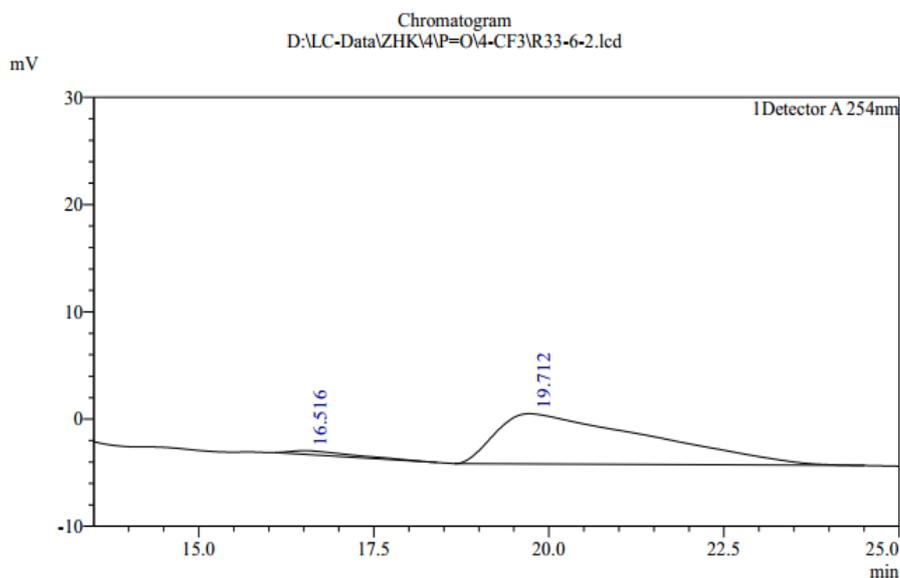
A white solid; m.p. = 96 - 98 °C; 50.9 mg, 94% yield; $[\alpha]_D^{25} = -45.00$ (*c* 0.6, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.43 (s, 1H), 7.99 - 7.86 (m 2H), 7.36 - 7.28 (m, 2H), 7.26 - 7.10 (m, 6H), 7.08 - 7.02 (m, 2H), 6.17 (s, 1H), 4.53 - 4.20 (m, 2H), 1.29 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.64 (d, *J* = 3.4 Hz), 150.41 (d, *J* = 9.6 Hz), 150.09 (d, *J* = 10.1 Hz), 137.97 (d, *J* = 4.8 Hz), 135.96 (d, *J* = 2.5 Hz), 135.63 (d, *J* = 2.5 Hz), 131.36 (d, *J* = 4.5 Hz), 129.99 (d, *J* = 6.2 Hz), 128.72 (dd, *J* = 6.4, 3.0 Hz), 126.11 (d, *J* = 0.8 Hz), 125.89, 125.44 (dd, *J* = 7.2, 3.2 Hz), 124.25 (d, *J* = 2.8 Hz), 122.81 (d, *J* = 2.1 Hz), 120.45 (d, *J* = 4.4 Hz), 119.93 (d, *J* = 4.6 Hz), 67.65 (d, *J* = 165.2 Hz), 65.58, 13.90; ³¹P NMR (162 MHz, CDCl₃) δ 3.05; HRMS (ESI) *m/z* calcd for C₂₃H₁₉NO₇PSF₃ [M+Na]⁺ = 564.0530, found = 564.0536; The ee value was 94%, *t_R* (minor) = 16.5 min, *t_R* (major) = 19.7 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|--------|--------|---------|---------|
| 1 | 15.358 | 220482 | 6031 | 57.051 | 50.182 |
| 2 | 20.539 | 218887 | 4540 | 42.949 | 49.818 |
| Total | | 439369 | 10571 | 100.000 | 100.000 |

Racemic **5g**

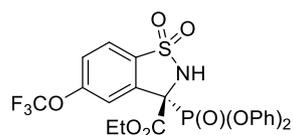


Peak Table

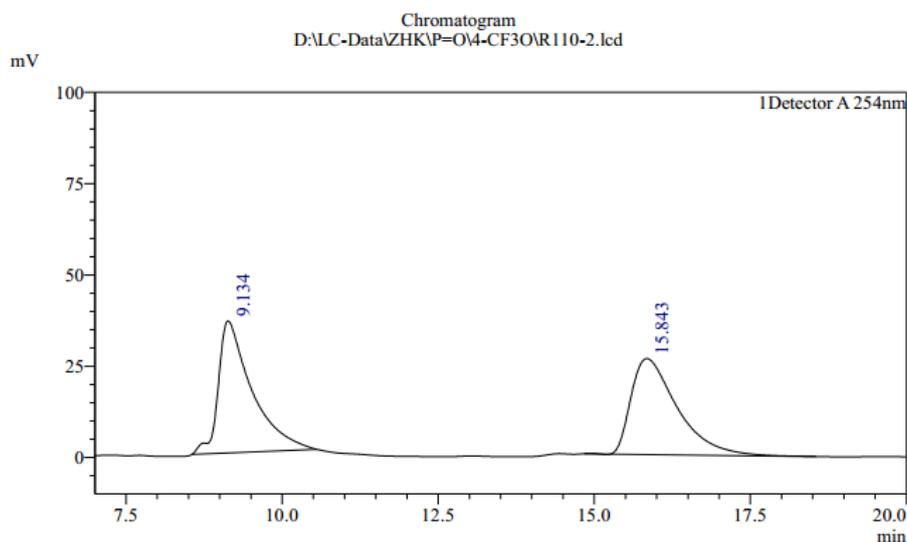
| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|--------|--------|---------|---------|
| 1 | 16.516 | 23749 | 345 | 6.855 | 3.205 |
| 2 | 19.712 | 717134 | 4682 | 93.145 | 96.795 |
| Total | | 740883 | 5027 | 100.000 | 100.000 |

Enantiomerically enriched **5g**

(S)-ethyl 3-(diphenoxyphosphoryl)-5-(trifluoromethoxy)-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5h)



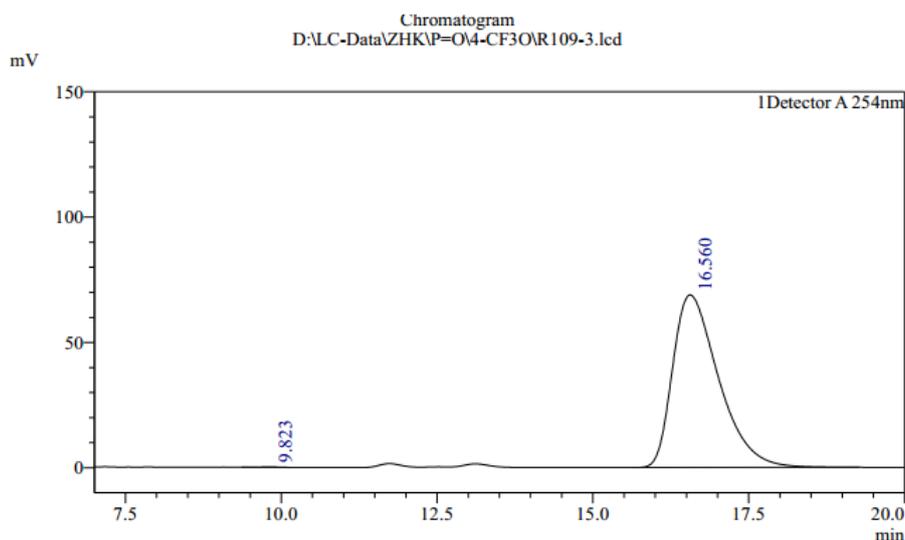
A white solid; m.p. = 85 - 88 °C; 50.1 mg, 90% yield; $[\alpha]_D^{25} = -31.67$ (c 0.7, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.05 - 8.01 (m, 1H), 7.89 - 8.85 (m, 1H), 7.47 (d, $J = 8.4$ Hz, 1H), 7.38 - 7.01 (m, 10H), 6.43 (d, $J = 4.8$ Hz, 1H), 4.43 - 4.27 (m, 2H), 1.27 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 164.61 (d, $J = 3.8$ Hz), 152.63 (dd, $J = 3.0, 1.9$ Hz), 150.13 (dd, $J = 28.7, 9.6$ Hz), 132.99 (d, $J = 4.9$ Hz), 132.71 (d, $J = 4.6$ Hz), 130.12 (d, $J = 1.0$ Hz), 129.92 (d, $J = 0.6$ Hz), 129.83, 129.46, 125.99 (d, $J = 0.9$ Hz), 125.79, 124.13, 123.69 (d, $J = 2.2$ Hz), 121.49, 120.61 (d, $J = 4.9$ Hz), 120.45 (d, $J = 4.5$ Hz), 119.97 (d, $J = 4.5$ Hz), 119.90, 118.90, 116.32, 115.39, 68.11 (dd, $J = 165.4$ Hz), 65.38, 13.75; ^{31}P NMR (162 MHz, CDCl_3) δ 3.39; ^{19}F NMR (376 MHz, CDCl_3) δ -57.87; HRMS (ESI) m/z calcd for $\text{C}_{23}\text{H}_{19}\text{NO}_8\text{PSF}_3$ $[\text{M}+\text{Na}]^+ = 580.0408$, found = 580.0400; The ee value was >99%, t_R (minor) = 9.8 min, t_R (major) = 16.7 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 9.134 | 1353193 | 36130 | 57.923 | 50.099 |
| 2 | 15.843 | 1347839 | 26246 | 42.077 | 49.901 |
| Total | | 2701032 | 62376 | 100.000 | 100.000 |

Racemic **5h**

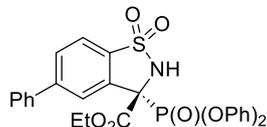


Peak Table

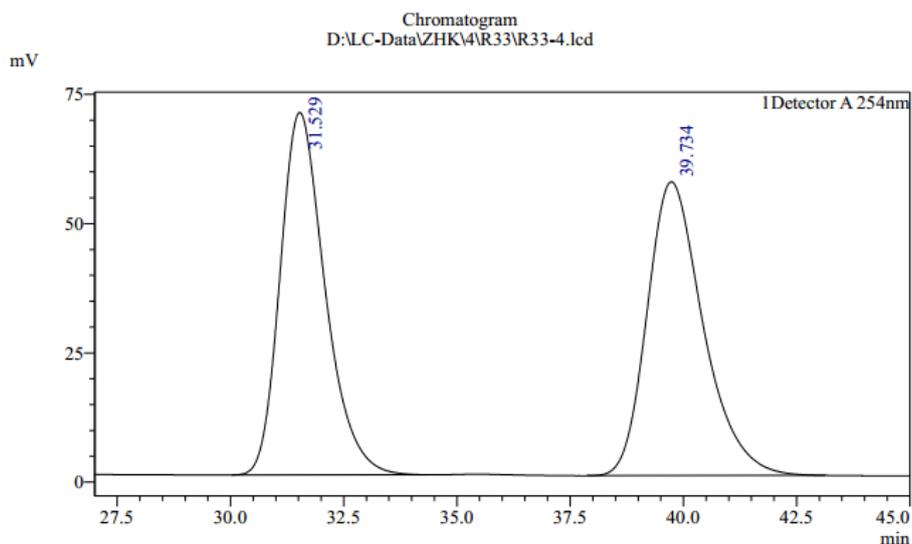
| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 9.823 | 3082 | 136 | 0.197 | 0.087 |
| 2 | 16.560 | 3536166 | 68881 | 99.803 | 99.913 |
| Total | | 3539248 | 69017 | 100.000 | 100.000 |

Enantiomerically enriched **5h**

(S)-ethyl 3-(diphenoxyphosphoryl)-5-phenyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5i)



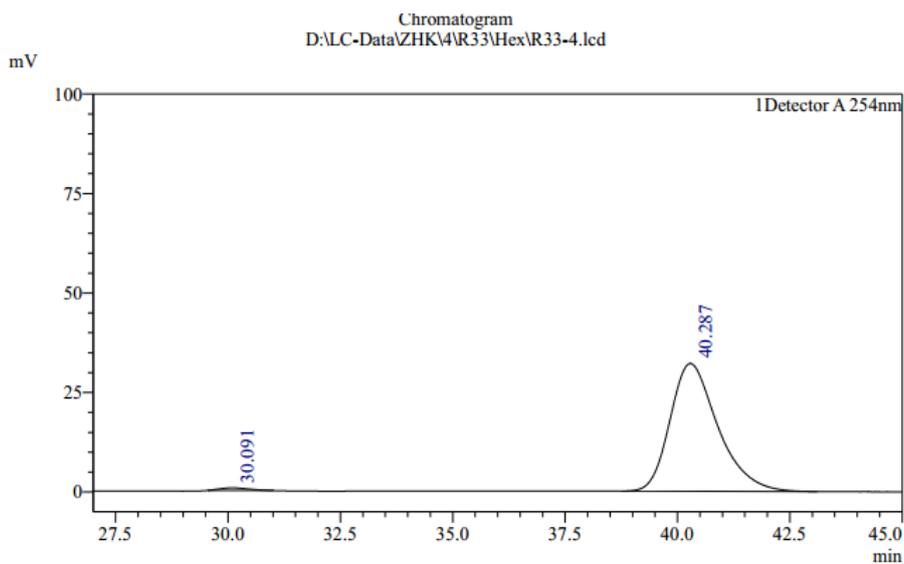
A white solid; m.p. = 106 - 107 °C; 51.6 mg, 94% yield; $[\alpha]_D^{25} = -38.17$ (*c* 0.6, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.34 - 8.28 (m, 1H), 7.92 - 7.79 (m, 2H), 7.55 - 7.40 (m, 5H), 7.37 - 7.29 (m, 2H), 7.23 - 7.12 (m, 5H), 7.069 - 7.03 (m, 1H), 7.02 - 6.97 (m, 2H), 6.14 (d, *J* = 3.6 Hz, 1H), 4.48 - 4.21 (m, 2H), 1.30 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.30 (d, *J* = 4.3 Hz), 150.53 (d, *J* = 9.6 Hz), 150.20 (d, *J* = 9.9 Hz), 147.29 (d, *J* = 2.5 Hz), 138.83, 133.31 (d, *J* = 4.9 Hz), 130.75 (d, *J* = 4.3 Hz), 130.59 (d, *J* = 2.6 Hz), 129.99, 129.75, 129.30, 129.05, 127.67, 126.27 (d, *J* = 2.9 Hz), 125.99, 125.66, 122.20 (d, *J* = 2.0 Hz), 120.61 (d, *J* = 4.4 Hz), 120.17 (d, *J* = 4.5 Hz), 67.77 (d, *J* = 165.6 Hz), 65.18, 14.07; ³¹P NMR (162 MHz, CDCl₃) δ 4.19; HRMS (ESI) *m/z* calcd for C₂₈H₂₄NO₇PS [M+Na]⁺ = 572.0898, found = 572.0899; The ee value was 98%, *t_R* (minor) = 30.1 min, *t_R* (major) = 40.3 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 31.529 | 4832525 | 70114 | 55.230 | 49.665 |
| 2 | 39.734 | 4897674 | 56835 | 44.770 | 50.335 |
| Total | | 9730199 | 126949 | 100.000 | 100.000 |

Racemic **5i**

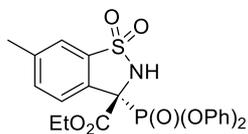


Peak Table

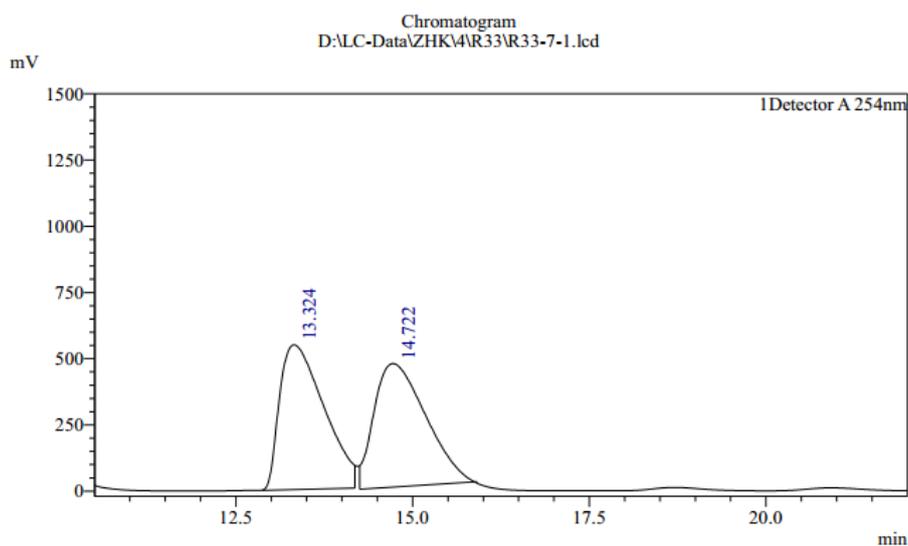
| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 30.091 | 29579 | 662 | 2.021 | 1.230 |
| 2 | 40.287 | 2374886 | 32107 | 97.979 | 98.770 |
| Total | | 2404466 | 32769 | 100.000 | 100.000 |

Enantiomerically enriched **5i**

(S)-ethyl 3-(diphenoxyphosphoryl)-6-methyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (**5j**)



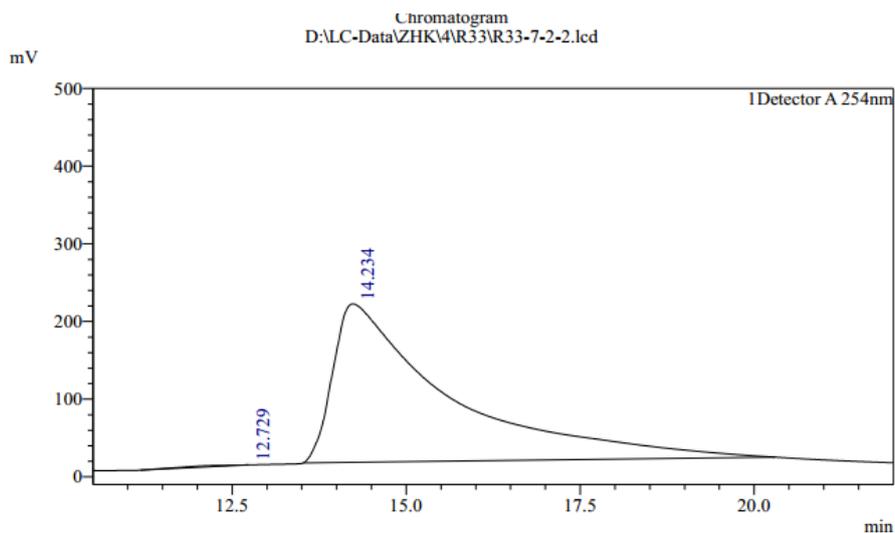
A white solid; m.p. = 98 - 99 °C; 45.8 mg, 94% yield; $[\alpha]_D^{25} = -33.47$ (c 0.7, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.02 (dd, $J = 8.4, 2.0$ Hz, 1H), 7.62 (s, 1H), 7.48 (d, $J = 8.0$ Hz, 1H), 7.33 - 7.27 (m, 2H), 7.24 - 7.07 (m, 6H), 7.02 - 6.96 (m, 2H), 6.07 (d, $J = 4.8$ Hz, 1H), 4.40- 4.25 (m, 2H), 2.47 (d, $J = 1.6$ Hz, 3H), 1.29 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 165.50 (d, $J = 4.9$ Hz), 150.47 (d, $J = 9.5$ Hz), 150.22 (d, $J = 10.1$ Hz), 142.53 (d, $J = 2.8$ Hz), 134.90 (d, $J = 5.0$ Hz), 134.86 (d, $J = 2.6$ Hz), 129.91, 129.77, 127.43 (d, $J = 2.8$ Hz), 127.31 (d, $J = 4.8$ Hz), 125.88, 125.58, 121.82 (d, $J = 2.1$ Hz), 120.63 (d, $J = 4.4$ Hz), 120.26 (d, $J = 4.7$ Hz), 67.61 (d, $J = 167.6$ Hz), 65.03, 21.46, 13.99; ^{31}P NMR (162 MHz, CDCl_3) δ 4.51; HRMS (ESI) m/z calcd for $\text{C}_{23}\text{H}_{22}\text{NO}_7\text{PS}$ $[\text{M}+\text{Na}]^+ = 510.0741$, found = 510.0746; The ee value was >99%, t_R (minor) = 12.7 min, t_R (major) = 14.3 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Detector A 254nm | | | | | |
|------------------|-----------|----------|---------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 13.324 | 23895413 | 547220 | 53.967 | 50.255 |
| 2 | 14.722 | 23652744 | 466778 | 46.033 | 49.745 |
| Total | | 47548157 | 1013997 | 100.000 | 100.000 |

Racemic **5j**

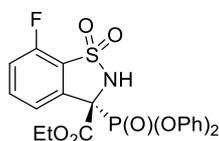


Peak Table

| Detector A 254nm | | | | | | |
|------------------|-----------|----------|--------|---------|---------|--|
| Peak# | Ret. Time | Area | Height | Height% | Area% | |
| 1 | 12.729 | 84291 | 2 | 0.001 | 0.358 | |
| 2 | 14.234 | 23453293 | 204022 | 99.999 | 99.642 | |
| Total | | 23537583 | 204024 | 100.000 | 100.000 | |

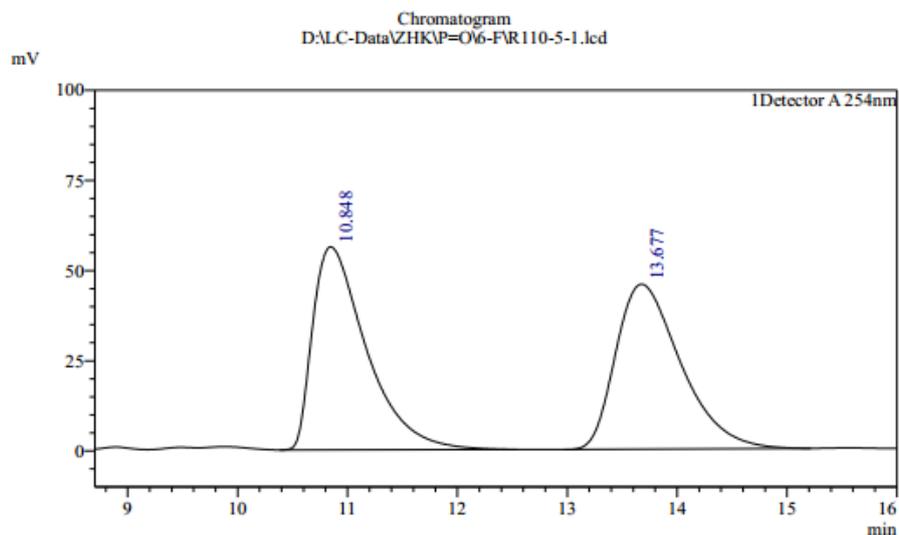
Enantiomerically enriched **5j**

(S)-ethyl 3-(diphenoxyphosphoryl)-7-fluoro-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5k)



A white solid; m.p. = 85 - 86 °C; 45.7 mg, 93% yield; $[\alpha]_D^{25} = -12.33$ (*c* 0.3, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.99 - 7.92 (m, 1H), 7.70 - 7.63 (m, 1H), 7.34 - 7.08 (m, 9H), 7.06 - 7.00 (m, 2H), 6.28 (d, *J* = 5.2 Hz, 1H), 4.38 - 4.29 (m, 2H), 1.28 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.91 (d, *J* = 3.8 Hz), 157.71 (d, *J* = 2.3 Hz), 155.13 (d, *J* = 2.2 Hz), 150.21 (dd, *J* = 29.6, 9.6 Hz), 136.19 (dd, *J* = 7.2, 2.7 Hz), 133.03 (dd, *J* = 3.9, 1.5 Hz), 129.96 (d, *J* = 0.6 Hz), 129.85, 126.01 (d, *J* = 1.0 Hz), 125.75 (d, *J* = 0.8 Hz), 123.70 (d, *J* = 3.3 Hz), 123.63 - 123.40 (q, *J* = 2.5 Hz), 123.02 (dd, *J* = 20.1, 4.8 Hz), 120.53 (d, *J* = 4.4 Hz), 120.13 (d, *J* = 4.5 Hz), 118.35 (dd, *J* = 18.1, 2.4 Hz), 68.58, 67.75 (d, *J* = 167.0 Hz), 13.94; ³¹P NMR (162 MHz, CDCl₃) δ 3.73; ¹⁹F NMR (376 MHz, CDCl₃) δ -115.25; HRMS (ESI) *m/z* calcd for C₂₂H₁₉NO₇PSF [M+Na]⁺ = 514.0491, found = 514.0498; The ee value was 99%, *t_R*

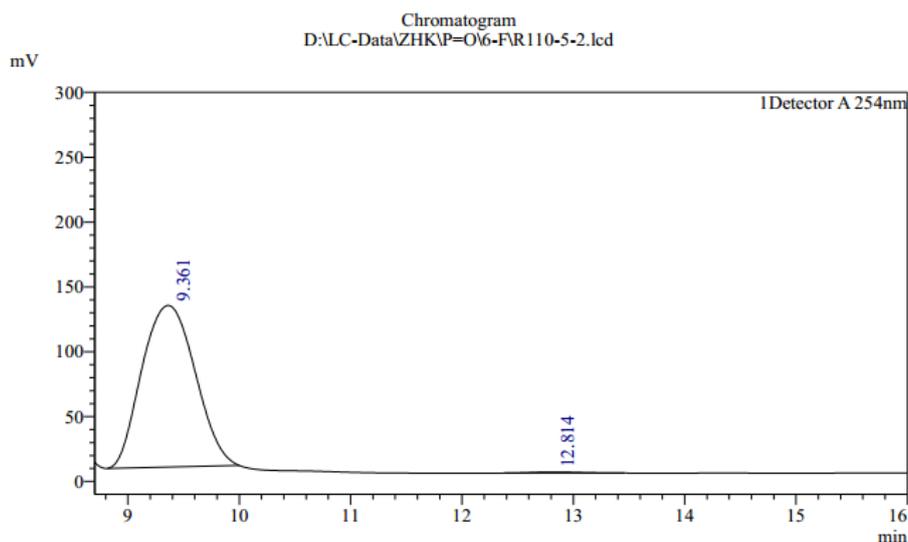
(major) = 9.4 min, t_R (minor) = 12.8 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 10.848 | 1919680 | 56314 | 55.168 | 50.440 |
| 2 | 13.677 | 1886184 | 45764 | 44.832 | 49.560 |
| Total | | 3805863 | 102077 | 100.000 | 100.000 |

Racemic **5k**

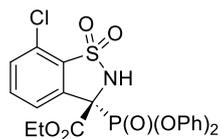


Peak Table

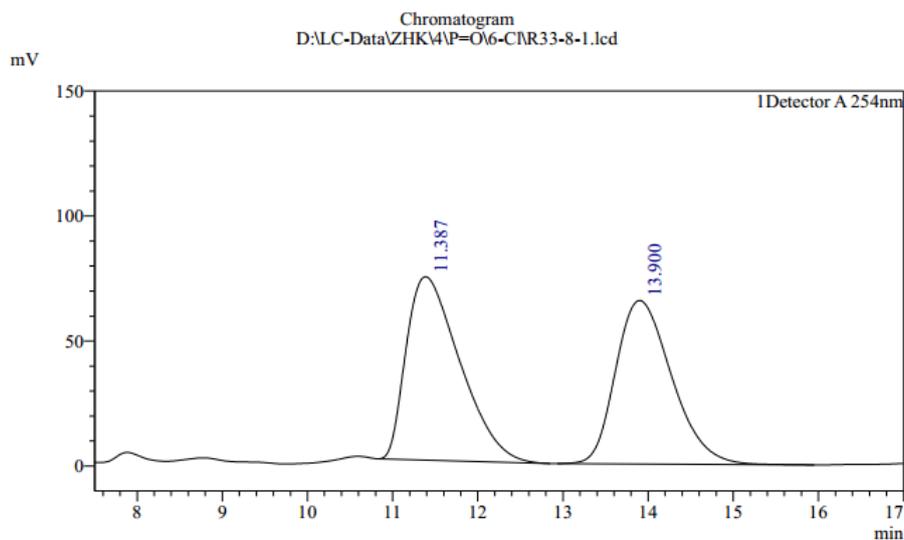
| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 9.361 | 4151361 | 124584 | 99.324 | 99.398 |
| 2 | 12.814 | 25152 | 848 | 0.676 | 0.602 |
| Total | | 4176513 | 125432 | 100.000 | 100.000 |

Enantiomerically enriched **5k**

(S)-ethyl 7-chloro-3-(diphenoxyphosphoryl)-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5l)



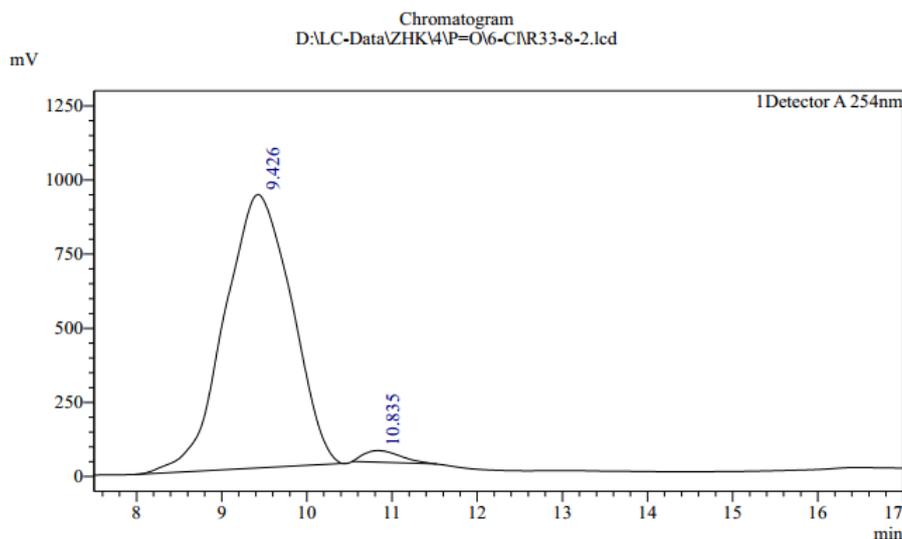
A white solid; m.p. = 81 - 83 °C; 47.2 mg, 93% yield; $[\alpha]_D^{25} = -31.83$ (*c* 0.6, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.64 - 7.50 (m, 3H), 7.47 - 7.40 (m, 2H), 7.39 - 7.28 (m, 6H), 7.24 - 7.14 (m, 2H), 5.53 (d, *J* = 3.6 Hz, 1H), 4.18 - 3.98 (m, 2H), 1.10 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.65, 150.22 (dd, *J* = 31.9, 7.3 Hz), 135.89, 135.05, 132.95 (d, *J* = 8.9 Hz), 132.53 (d, *J* = 5.7 Hz), 131.73, 130.00, 129.82, 129.41, 126.05, 123.38, 120.92 (d, *J* = 4.7 Hz), 120.71 (d, *J* = 4.9 Hz), 63.26, 61.87 (d, *J* = 5.1 Hz), 13.87; ³¹P NMR (162 MHz, CDCl₃) δ 3.81; HRMS (ESI) *m/z* calcd for C₂₂H₁₉NO₇PS [M+Na]⁺ = 530.0195, found = 530.0191; The ee value was 95%, *t_R* (major) = 9.4 min, *t_R* (minor) = 10.8 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 11.387 | 3216960 | 73343 | 52.858 | 51.827 |
| 2 | 13.900 | 2990155 | 65412 | 47.142 | 48.173 |
| Total | | 6207115 | 138754 | 100.000 | 100.000 |

Racemic 51

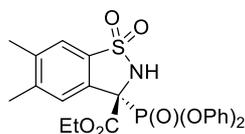


Peak Table

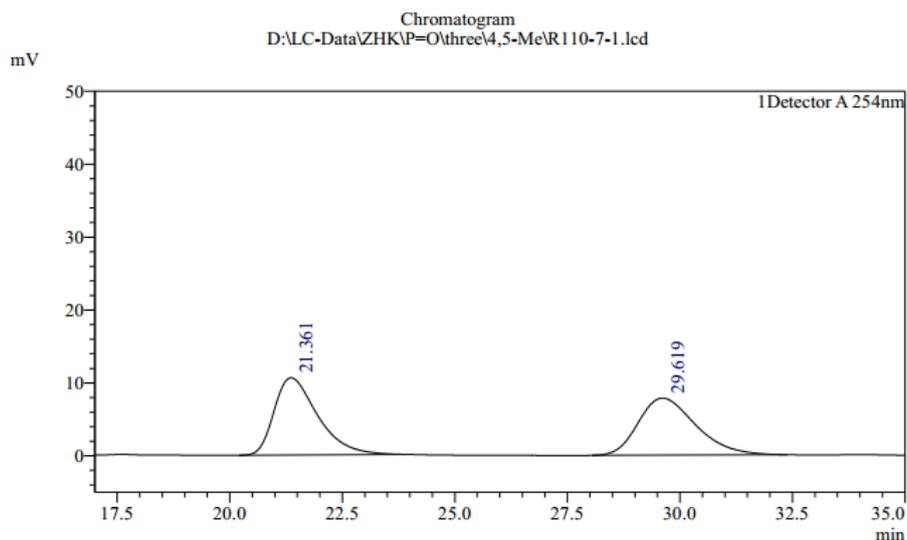
| Detector A 254nm | | | | | |
|------------------|-----------|----------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 9.426 | 50565136 | 921394 | 95.927 | 97.690 |
| 2 | 10.835 | 1195769 | 39126 | 4.073 | 2.310 |
| Total | | 51760906 | 960520 | 100.000 | 100.000 |

Enantiomerically enriched **5l**

(S)-ethyl 3-(diphenoxyphosphoryl)-5,6-dimethyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5m)



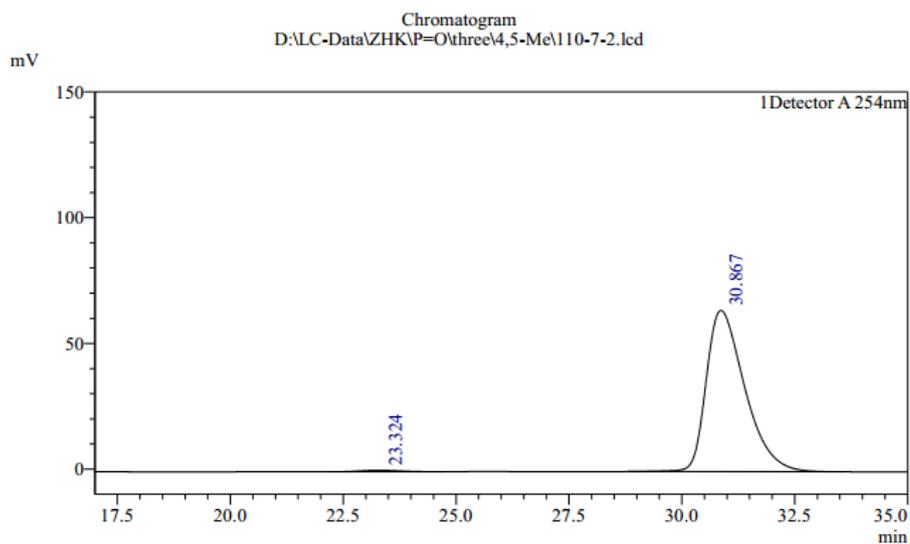
A white solid; m.p. = 147 - 149 °C; 46.1 mg, 92% yield; $[\alpha]_D^{25} = -31.67$ (*c* 0.7, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.87 - 7.81 (m, 1H), 7.57 (s, 1H), 7.33 - 7.27 (m, 2H), 7.23 - 7.13 (m, 5H), 7.13 - 7.06 (m, 1H), 7.04 - 6.98 (m, 2H), 6.08 (d, *J* = 5.2 Hz, 1H), 4.37 - 4.28 (m, 2H), 2.39 - 2.28 (m, 6H), 1.28 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.51 (d, *J* = 5.2 Hz), 150.36 (dd, *J* = 29.7, 9.7 Hz), 143.88 (d, *J* = 2.7 Hz), 141.28 (d, *J* = 2.8 Hz), 132.30 (d, *J* = 5.0 Hz), 129.85 (d, *J* = 0.6 Hz), 129.65, 127.94 (d, *J* = 2.9 Hz), 127.57 (d, *J* = 4.5 Hz), 125.80 (d, *J* = 1.0 Hz), 125.44 (d, *J* = 0.6 Hz), 121.98 (d, *J* = 2.2 Hz), 120.58 (d, *J* = 4.4 Hz), 120.19 (d, *J* = 4.5 Hz), 67.49 (d, *J* = 167.3 Hz), 64.90, 20.71, 20.14, 13.95; ³¹P NMR (162 MHz, CDCl₃) δ 4.58; HRMS (ESI) *m/z* calcd for C₂₄H₂₄NO₇PS [M+Na]⁺ = 524.0898, found = 524.0892; The ee value was 98%, *t_R* (minor) = 23.3 min, *t_R* (major) = 30.9 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 21.361 | 714649 | 10592 | 57.503 | 50.684 |
| 2 | 29.619 | 695353 | 7828 | 42.497 | 49.316 |
| Total | | 1410002 | 18419 | 100.000 | 100.000 |

Racemic **5m**

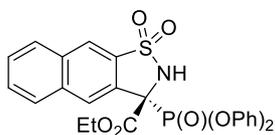


Peak Table

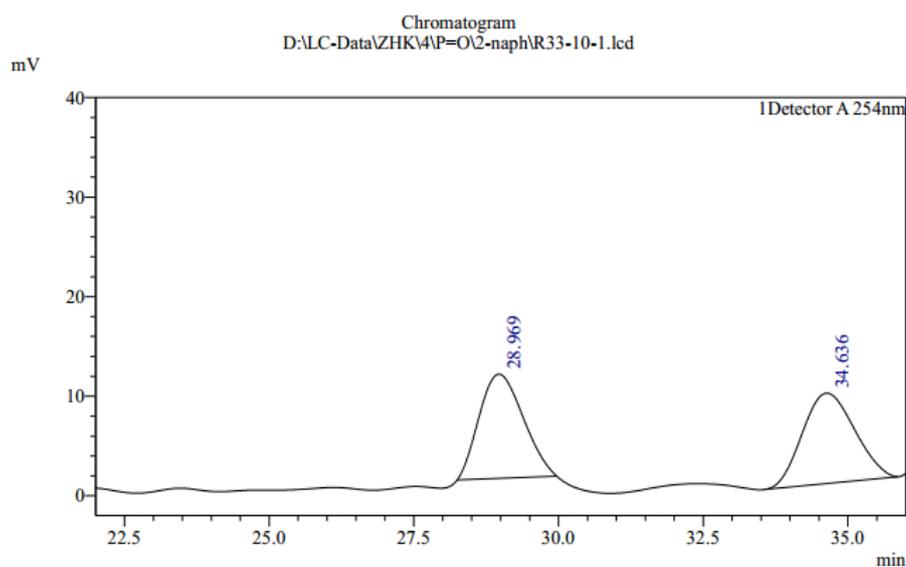
| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 23.324 | 38442 | 613 | 0.947 | 0.998 |
| 2 | 30.867 | 3814681 | 64120 | 99.053 | 99.002 |
| Total | | 3853123 | 64733 | 100.000 | 100.000 |

Enantiomerically enriched **5m**

(S)-ethyl 3-(diphenoxyphosphoryl)-2,3-dihydronaphtho[2,3-d]isothiazole-3-carboxylate 1,1-dioxide (5n)



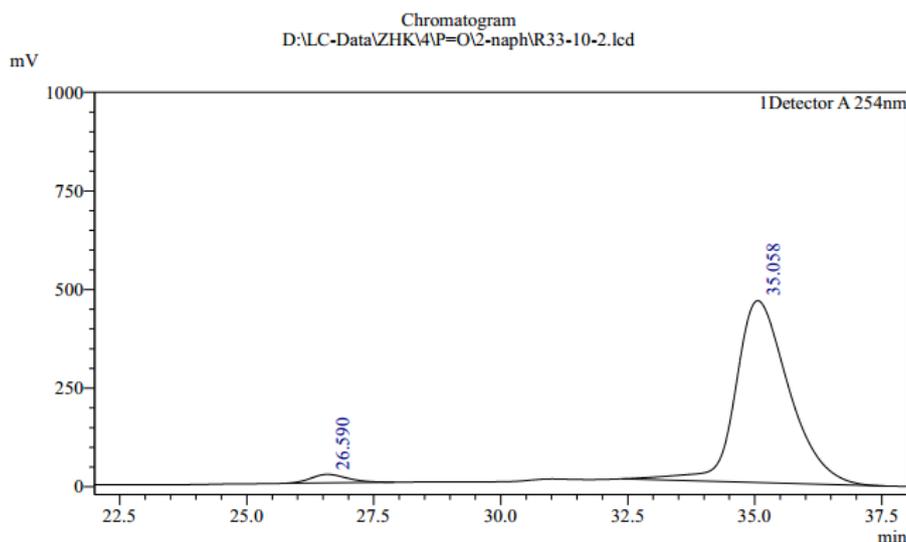
A white solid; m.p. = 107 - 108 °C; 48.1 mg, 92% yield; $[\alpha]_D^{25} = -57.67$ (c 0.7, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.44 (d, $J = 8.0$ Hz, 1H), 8.20 - 8.07(m, 2H), 7.99 (d, $J = 8.0$ Hz, 1H), 7.81 - 7.67 (m, 2H), 7.33 - 7.28 (m, 2H), 7.22 - 7.10 (m, 5H), 7.09 - 6.97 (m, 3H), 5.30 (s, 1H), 4.45 - 4.29 (m, 2H), 1.31 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 165.36 (d, $J = 4.2$ Hz), 150.36 (d, $J = 10.0$ Hz), 150.23 (d, $J = 10.0$ Hz), 134.58 (d, $J = 2.0$ Hz), 134.33 (d, $J = 1.5$ Hz), 130.65 (d, $J = 5.9$ Hz), 129.91, 129.74, 129.57, 129.43 (d, $J = 4.5$ Hz), 128.82 (d, $J = 16.1$ Hz), 125.76 (d, $J = 28.9$ Hz), 125.33 (d, $J = 2.2$ Hz), 123.30, 122.70 (d, $J = 1.6$ Hz), 120.64 (d, $J = 4.4$ Hz), 120.28 (d, $J = 4.5$ Hz), 67.96 (d, $J = 167.1$ Hz), 65.21, 14.01; ^{31}P NMR (162 MHz, CDCl_3) δ 4.20; HRMS (ESI) m/z calcd for $\text{C}_{26}\text{H}_{22}\text{NO}_7\text{PS}$ $[\text{M}+\text{Na}]^+ = 546.0741$, found $[\text{M}+\text{Na}]^+ = 546.0743$; The ee value was 94%, t_R (minor) = 26.6 min, t_R (major) = 35.1 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Detector A 254nm | | | | | |
|------------------|-----------|---------|--------|---------|---------|
| Peak# | Ret. Time | Area | Height | Height% | Area% |
| 1 | 28.969 | 552454 | 10452 | 53.505 | 49.315 |
| 2 | 34.636 | 567801 | 9082 | 46.495 | 50.685 |
| Total | | 1120254 | 19534 | 100.000 | 100.000 |

Racemic **5n**

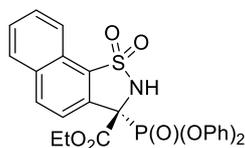


Peak Table

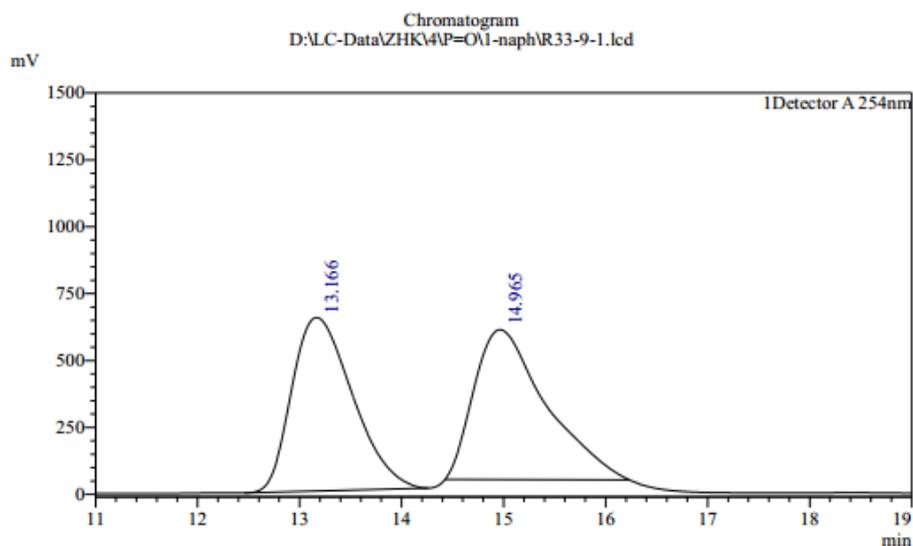
| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|--------|---------|---------|
| 1 | 26.590 | 1084290 | 21519 | 4.459 | 3.115 |
| 2 | 35.058 | 33728690 | 461061 | 95.541 | 96.885 |
| Total | | 34812981 | 482579 | 100.000 | 100.000 |

Enantiomerically enriched **5n**

(S)-ethyl 3-(diphenoxyphosphoryl)-2,3-dihydronaphtho[2,1-d]isothiazole-3-carboxylate 1,1-dioxide (5o)



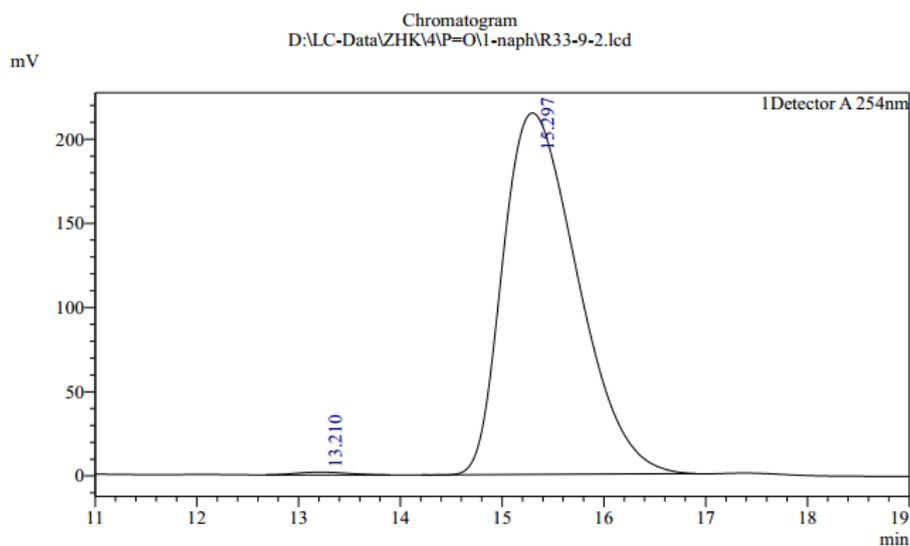
A white solid; m.p. = 108 - 110 °C; 48.1 mg, 92% yield; $[\alpha]_D^{25} = -15.87$ (*c* 0.3, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.67 - 7.57 (m, 1H), 8.37 (s, 1H), 8.08 - 7.92 (m, 2H), 7.73 - 7.56 (m, 2H), 7.37 - 7.28 (m, 2H), 7.21 - 7.14 (m, 3H), 7.14 - 7.06 (m, 2H), 7.02 - 6.90 (m, 3H), 5.30 (s, 1H), 4.47 - 4.29 (m, 2H), 1.32 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.65 (d, *J* = 4.9 Hz), 150.47 (d, *J* = 9.6 Hz), 150.24 (d, *J* = 10.0 Hz), 135.33 (d, *J* = 2.8 Hz), 133.57 (d, *J* = 2.0 Hz), 132.00 (d, *J* = 4.2 Hz), 129.93, 129.67, 129.40, 129.22 (d, *J* = 11.6 Hz), 128.77, 128.06 (d, *J* = 4.1 Hz), 125.92, 125.51, 125.18, 122.71 (d, *J* = 1.6 Hz), 120.61 (d, *J* = 4.4 Hz), 120.14 (d, *J* = 4.5 Hz), 115.46, 67.52 (d, *J* = 167.1 Hz), 65.14, 14.03; ³¹P NMR (162 MHz, CDCl₃) δ 4.57; HRMS (ESI) *m/z* calcd for C₂₆H₂₂NO₇PS [M+Na]⁺ = 546.0741, found = 546.0738; The ee value was 99%, *t_R* (minor) = 13.2 min, *t_R* (major) = 15.3 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|---------|---------|---------|
| 1 | 13.166 | 26827315 | 648949 | 53.671 | 49.420 |
| 2 | 14.965 | 27457531 | 560167 | 46.329 | 50.580 |
| Total | | 54284847 | 1209116 | 100.000 | 100.000 |

Racemic **5o**

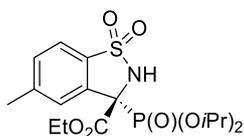


Peak Table

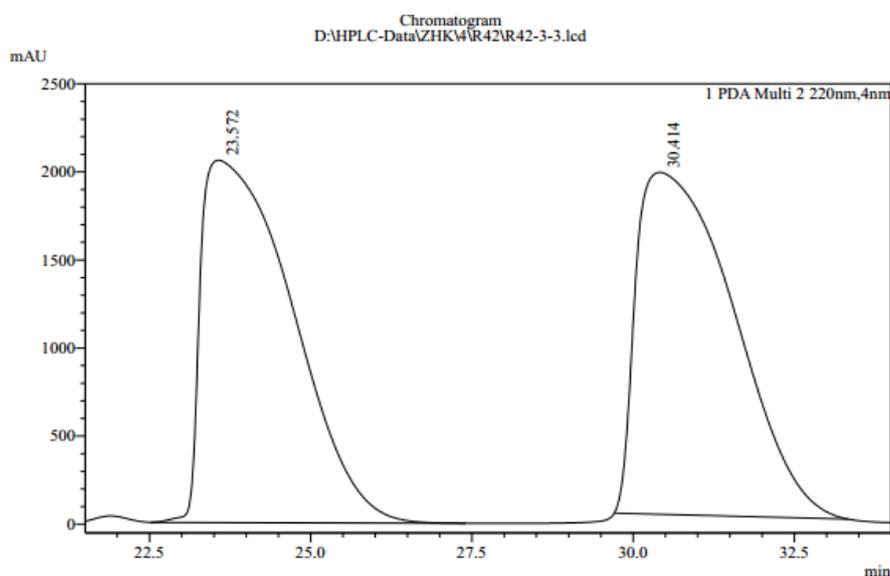
| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|----------|--------|---------|---------|
| 1 | 13.210 | 56820 | 1623 | 0.751 | 0.516 |
| 2 | 15.297 | 10947201 | 214629 | 99.249 | 99.484 |
| Total | | 11004021 | 216253 | 100.000 | 100.000 |

Enantiomerically enriched **5o**

(S)-ethyl 3-(diisopropoxyphosphoryl)-5-methyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5p)



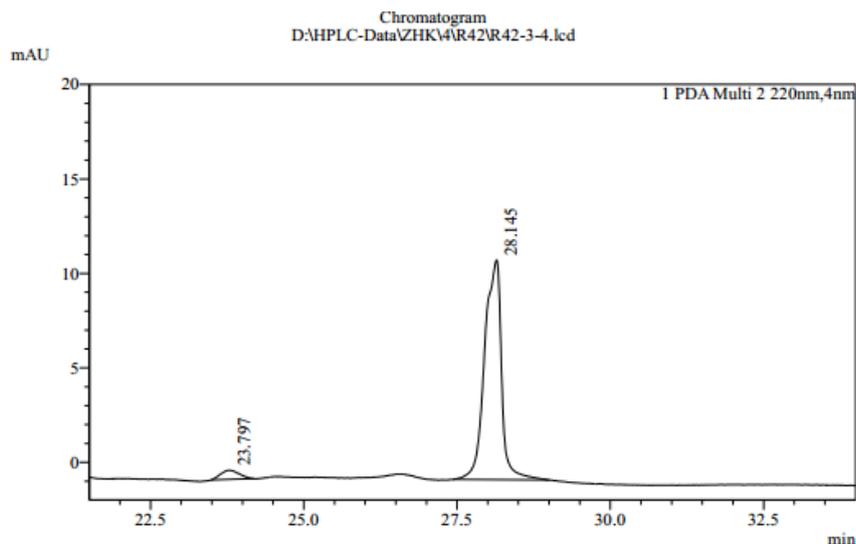
A white solid; m.p. = 91 - 93 °C; 39.0 mg, 93% yield; $[\alpha]_D^{25} = -22.37$ (*c* 0.6, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.89 (s, 1H), 7.64 (d, *J* = 8.0 Hz, 1H), 7.45 - 7.36 (m, 1H), 4.88 - 4.74 (m, 1H), 4.65 - 4.51 (m, 1H), 4.44 - 4.27 (m, 2H), 2.49 (s, 3H), 1.34 (dd, *J* = 7.2, 4.4 Hz, 6H), 1.28 (dd, *J* = 6.0, 2.8 Hz, 6H), 1.00 (d, *J* = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.28 (d, *J* = 3.0 Hz), 144.52 (d, *J* = 2.5 Hz), 131.99 (dd, *J* = 17.5, 4.8 Hz), 131.47 (d, *J* = 4.7 Hz), 127.92 (d, *J* = 2.8 Hz), 121.21 (d, *J* = 1.9 Hz), 74.53 (dd, *J* = 17.0, 7.9 Hz), 67.96 (d, *J* = 160.3 Hz), 64.35, 24.22 (dd, *J* = 3.1, 0.2 Hz), 23.74 (d, *J* = 5.8 Hz), 23.16 (d, *J* = 5.6 Hz), 22.08, 14.08; ³¹P NMR (162 MHz, CDCl₃) δ 10.29; HRMS (ESI) *m/z* calcd for C₁₇H₂₆NO₇PS [M+Na]⁺ = 442.1054, found = 442.1048; The ee value was 91%, *t_R* (minor) = 23.8 min, *t_R* (major) = 28.1 min (Chiralcel IC, λ = 220 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| PDA Ch2 220nm | | | | | |
|---------------|-----------|---------|---------|-----------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 23.572 | 2056900 | 51.443 | 202623371 | 49.724 |
| 2 | 30.414 | 1941504 | 48.557 | 204872008 | 50.276 |
| Total | | 3998404 | 100.000 | 407495379 | 100.000 |

Racemic **5p**

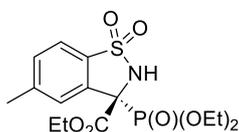


Peak Table

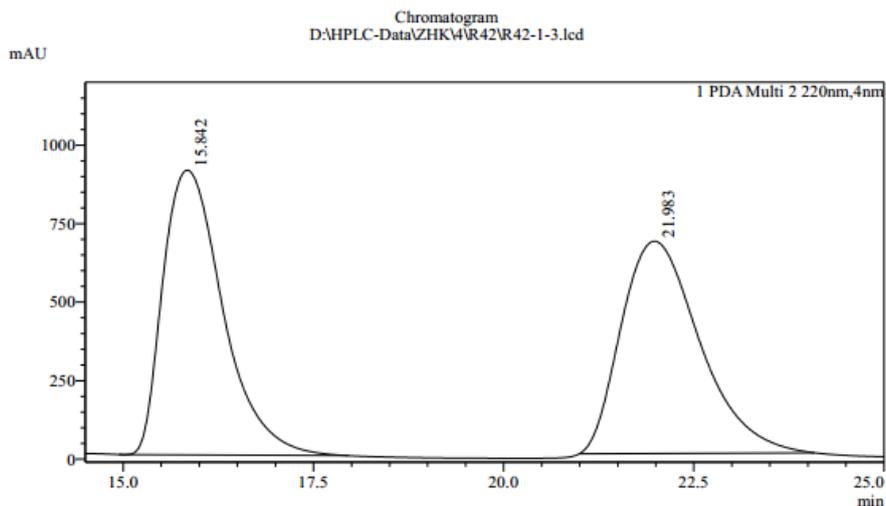
| PDA Ch2 220nm | | | | | |
|---------------|-----------|--------|---------|--------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 23.797 | 481 | 3.977 | 10305 | 4.407 |
| 2 | 28.145 | 11626 | 96.023 | 223556 | 95.593 |
| Total | | 12107 | 100.000 | 233861 | 100.000 |

Enantiomerically enriched **5p**

(S)-ethyl 3-(diethoxyphosphoryl)-5-methyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5q)



A white solid; m.p. = 85 - 87 °C; 36.8 mg, 94% yield; $[\alpha]_D^{25} = -22.37$ (c 0.6, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.87 - 7.82 (m, 1H), 7.66 (d, $J = 8.0$ Hz, 1H), 7.46 - 7.40 (m, 1H), 5.84 (s, 1H), 4.46 - 4.34 (m, 2H), 4.34 - 4.24 (m, 2H), 4.17 - 3.98 (m, 2H), 2.50 (s, 3H), 1.37 (t, $J = 7.2$ Hz, 3H), 1.30 (td, $J = 7.2, 0.8$ Hz, 3H), 1.23 (td, $J = 7.2, 0.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 166.10 (d, $J = 2.9$ Hz), 144.67 (d, $J = 2.5$ Hz), 132.10 (d, $J = 2.8$ Hz), 132.06 (d, $J = 5.0$ Hz), 131.51 (d, $J = 4.4$ Hz), 127.80 (d, $J = 2.7$ Hz), 121.33 (d, $J = 2.0$ Hz), 67.95 (d, $J = 160.7$ Hz), 65.96 (d, $J = 7.3$ Hz), 65.20 (d, $J = 7.3$ Hz), 64.60, 22.13, 16.55 (d, $J = 5.4$ Hz), 16.35 (d, $J = 5.4$ Hz), 14.12; ^{31}P NMR (162 MHz, CDCl_3) δ 12.05; HRMS (ESI) m/z calcd for $\text{C}_{15}\text{H}_{22}\text{NO}_7\text{PS}$ $[\text{M}+\text{Na}]^+$ = 414.0752, found = 414.0750; The ee value was 94%, t_R (major) = 16.5 min, t_R (minor) = 21.1 min (Chiralcel AS-H, $\lambda = 220$ nm, 30% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).

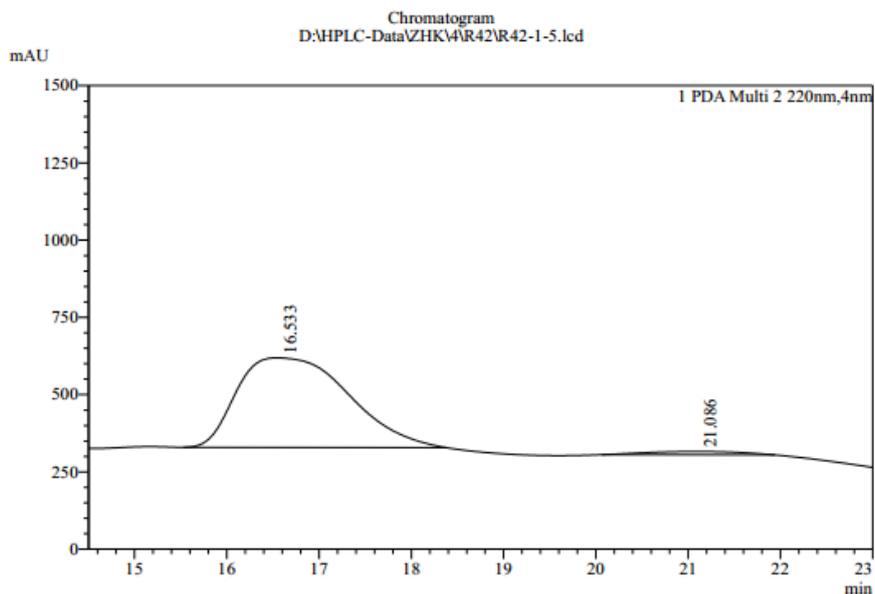


Peak Table

PDA Ch2 220nm

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|---------|---------|----------|---------|
| 1 | 15.842 | 906673 | 57.278 | 49801409 | 50.544 |
| 2 | 21.983 | 676253 | 42.722 | 48728520 | 49.456 |
| Total | | 1582926 | 100.000 | 98529929 | 100.000 |

Racemic **5q**



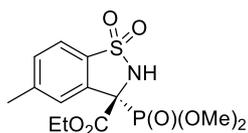
Peak Table

PDA Ch2 220nm

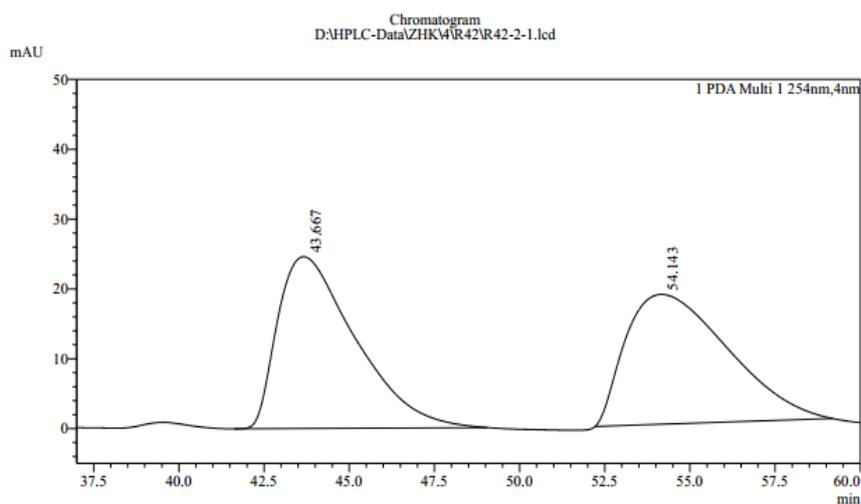
| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|----------|---------|
| 1 | 16.533 | 289891 | 96.426 | 24386843 | 96.924 |
| 2 | 21.086 | 10745 | 3.574 | 773992 | 3.076 |
| Total | | 300636 | 100.000 | 25160835 | 100.000 |

Enantiomerically enriched **5q**

(S)-ethyl 3-(dimethoxyphosphoryl)-5-methyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (5r)



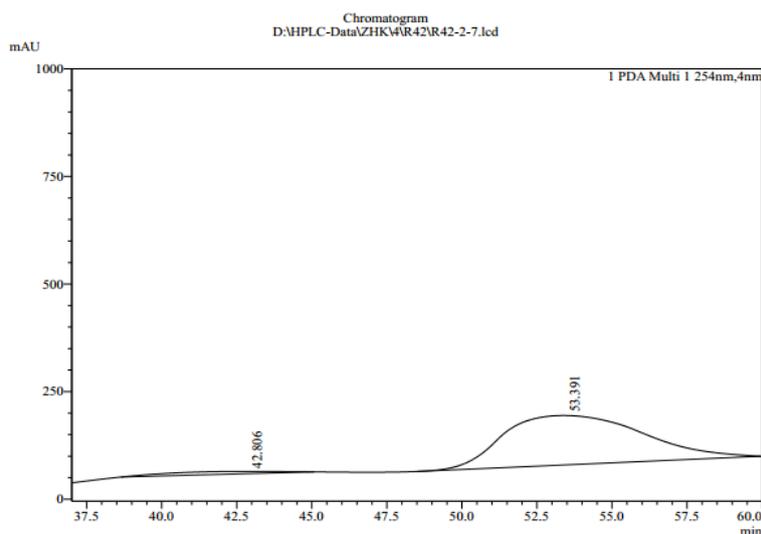
A white solid; m.p. = 101 - 103 °C; 34.1 mg, 94% yield; $[\alpha]_D^{25} = -22.37$ (c 0.6, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.79 (s, 1H), 7.67 (d, $J = 8.0$ Hz, 1H), 7.47 - 7.40 (m, 1H), 5.90 (s, 1H), 4.47 - 4.34 (m, 2H), 3.91 (d, $J = 10.4$ Hz, 3H), 3.76 (d, $J = 10.8$ Hz, 3H), 2.51 (s, 3H), 1.37 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 165.92 (d, $J = 3.0$ Hz), 144.85 (d, $J = 2.6$ Hz), 132.24 (d, $J = 2.4$ Hz), 131.97 (d, $J = 4.6$ Hz), 131.31 (d, $J = 4.8$ Hz), 127.55 (d, $J = 2.6$ Hz), 121.43 (d, $J = 2.0$ Hz), 67.89 (d, $J = 162.5$ Hz), 64.75, 56.40 (d, $J = 7.5$ Hz), 55.44 (d, $J = 7.3$ Hz), 22.13, 14.11; ^{31}P NMR (162 MHz, CDCl_3) δ 14.46; HRMS (ESI) m/z calcd for $\text{C}_{13}\text{H}_{18}\text{NO}_7\text{PS}$ $[\text{M}+\text{Na}]^+ = 386.0439$, found = 386.0447; The ee value was 91%, t_R (minor) = 42.8 min, t_R (major) = 53.4 min (Chiralcel AS-H, $\lambda = 254$ nm, 10% *i*-PrOH/hexanes, flow rate = 0.8 mL/min).



Peak Table

| PDA Ch1 254nm | | | | | |
|---------------|-----------|--------|---------|---------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 43.667 | 24599 | 56.924 | 3913970 | 50.174 |
| 2 | 54.143 | 18615 | 43.076 | 3886809 | 49.826 |
| Total | | 43215 | 100.000 | 7800779 | 100.000 |

Racemic **5r**



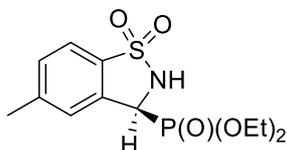
Peak Table

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|----------|---------|
| 1 | 42.806 | 5312 | 4.413 | 1678774 | 4.315 |
| 2 | 53.391 | 115041 | 95.587 | 37226302 | 95.685 |
| Total | | 120352 | 100.000 | 38905076 | 100.000 |

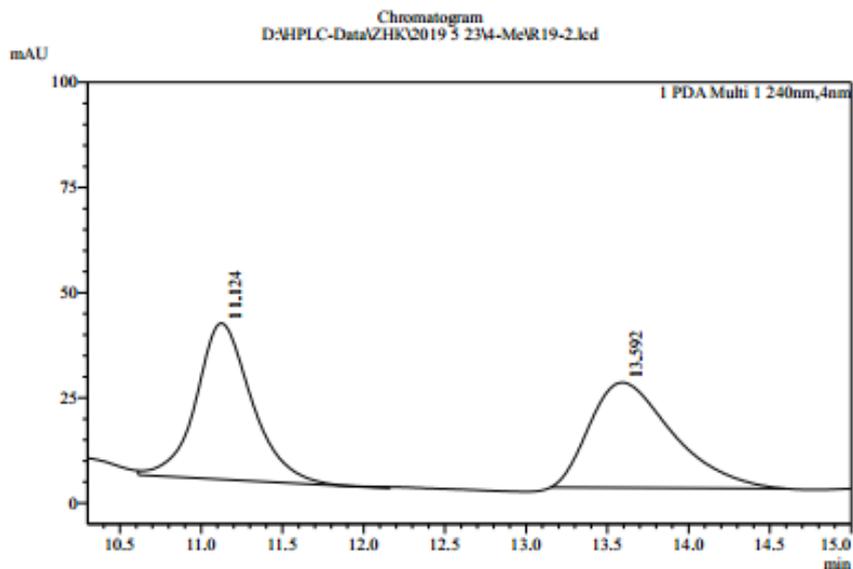
Enantiomerically enriched **5r**

(S)-diethyl (5-methyl-1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl)

Phosphonate (**5'a**)



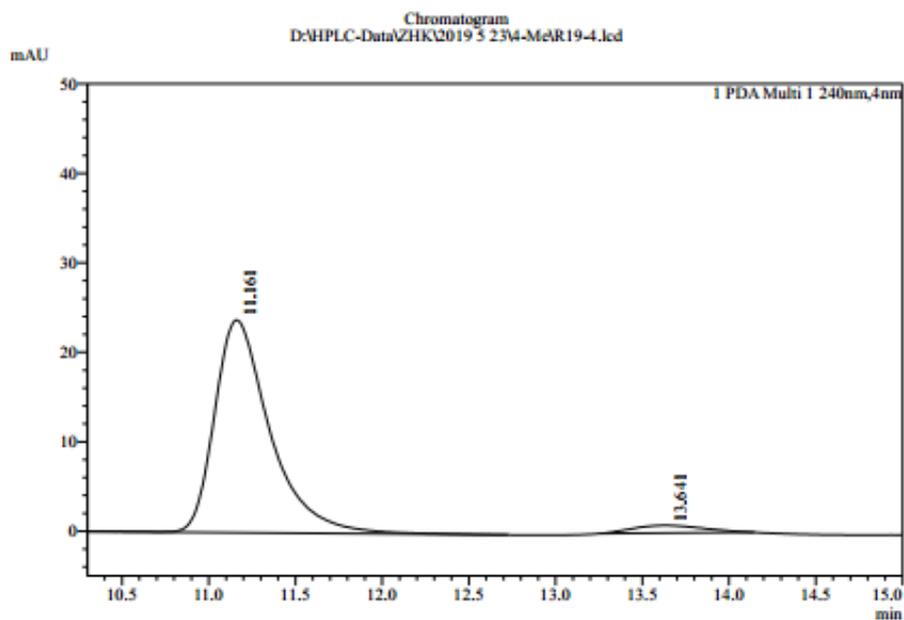
A white solid; m.p. = 90 - 92 °C; 29.3 mg, 92% yield; $[\alpha]_D^{25} = -36.33$ (*c* 0.5, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.68 (d, *J* = 8.0 Hz, 1H), 7.52 (s, 1H), 7.37 (d, *J* = 8.0 Hz, 1H), 6.00 - 5.87 (m, 1H), 5.00 (dd, *J* = 11.2, 4.8 Hz, 1H), 4.32 - 4.18 (m, 2H), 4.15 - 3.93 (m, 2H), 2.47 (s, 3H), 1.30 (t, *J* = 7.2 Hz, 3H), 1.22 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 144.39 (d, *J* = 2.7Hz), 132.67 (d, *J* = 5.2 Hz), 132.45 (d, *J* = 5.6 Hz), 131.08 (d, *J* = 2.3 Hz), 126.17 (d, *J* = 2.8 Hz), 121.39 (d, *J* = 2.7Hz), 64.58 (d, *J* = 7.0 Hz), 64.17 (d, *J* = 7.2 Hz), 54.09 (d, *J* = 160.9 Hz), 21.98, 16.54 (d, *J* = 5.6 Hz), 16.38 (d, *J* = 5.6 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 16.85; HRMS (ESI) *m/z* calcd for C₁₂H₁₈NO₅PS [M+Na]⁺ = 342.0530, found = 342.0526; The ee value was 91%, *t_R* (major) = 11.2 min, *t_R* (minor) = 13.6 min (Chiralcel AS-H, λ = 240 nm, 40% *i*-PrOH/hexanes, flow rate = 0.6 mL/min).



Peak Table

| PDA Ch1 240nm | | | | | |
|---------------|-----------|--------|---------|---------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 11.124 | 37126 | 59.778 | 893132 | 50.101 |
| 2 | 13.592 | 24981 | 40.222 | 889533 | 49.899 |
| Total | | 62107 | 100.000 | 1782665 | 100.000 |

Racemic **5'a**

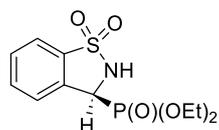


Peak Table

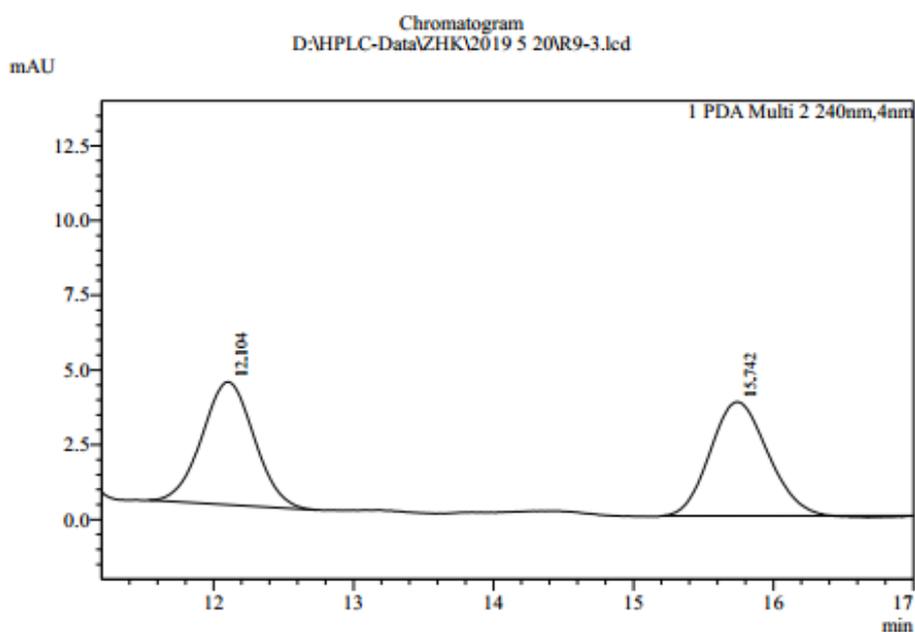
| PDA Ch1 240nm | | | | | |
|---------------|-----------|--------|---------|--------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 11.161 | 23755 | 96.570 | 529415 | 95.690 |
| 2 | 13.641 | 844 | 3.430 | 23843 | 4.310 |
| Total | | 24599 | 100.000 | 553258 | 100.000 |

Enantiomerically enriched **5'a**

(S)-diethyl (1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl)phosphonate (5'b)



A white solid; m.p. = 92 - 94 °C; 28.4 mg, 93% yield; $[\alpha]_D^{25} = -33.33$ (*c* 0.6, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.81 (d, *J* = 7.6 Hz, 1H), 7.74 (d, *J* = 7.6 Hz, 1H), 7.668 - 7.62(m, 1H), 7.61 - 7.54 (m, 1H), 6.27 - 6.14 (m, 1H), 5.07 (dd, *J* = 11.2, 4.4 Hz, 1H), 4.34 - 4.17 (m, 2H), 4.13 - 3.94 (m, 2H), 1.28 (t, *J* = 7.2 Hz, 3H), 1.22 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 135.12 (d, *J* = 5.6 Hz), 133.26 (d, *J* = 2.4 Hz), 132.37 (d, *J* = 5.2 Hz), 130.05 (d, *J* = 2.6 Hz), 126.04 (d, *J* = 2.8 Hz), 121.66 (d, *J* = 1.9 Hz), 64.76 (d, *J* = 6.8 Hz), 64.11 (d, *J* = 7.1 Hz), 54.26 (d, *J* = 161.4 Hz), 16.52 (d, *J* = 5.4 Hz), 16.38 (d, *J* = 5.5 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 16.83; HRMS (ESI) *m/z* calcd for C₁₁H₁₆NO₅PS [M+Na]⁺ = 328.0379, found = 328.0369; The ee value was 90%, *t_R* (major) = 11.2 min, *t_R* (minor) = 16.3 min (Chiralcel AS-H, λ = 240 nm, 40% *i*-PrOH/hexanes, flow rate = 0.6 mL/min).

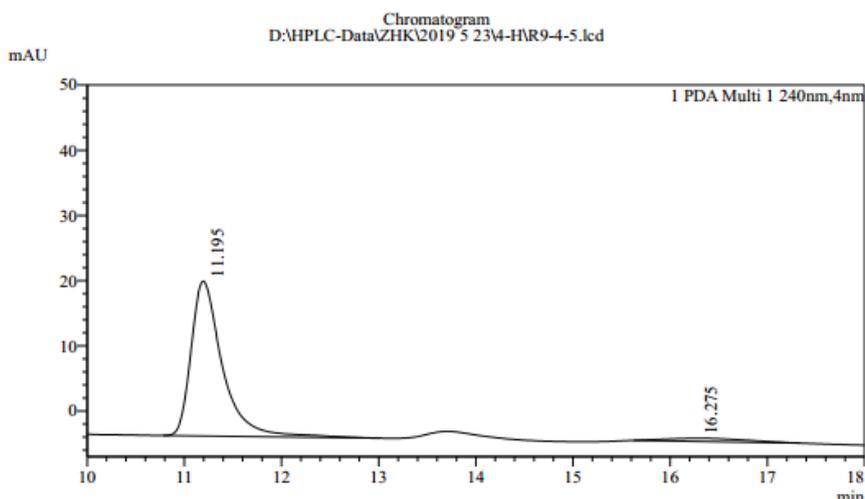


Peak Table

PDA Ch2 240nm

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|--------|---------|
| 1 | 12.104 | 4112 | 51.888 | 106896 | 49.680 |
| 2 | 15.742 | 3813 | 48.112 | 108272 | 50.320 |
| Total | | 7925 | 100.000 | 215168 | 100.000 |

Racemic **5'b**



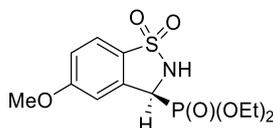
Peak Table

| PDA Ch1 240nm | | | | | |
|---------------|-----------|--------|---------|--------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 11.195 | 23720 | 98.028 | 533284 | 95.064 |
| 2 | 16.275 | 477 | 1.972 | 27687 | 4.936 |
| Total | | 24197 | 100.000 | 560971 | 100.000 |

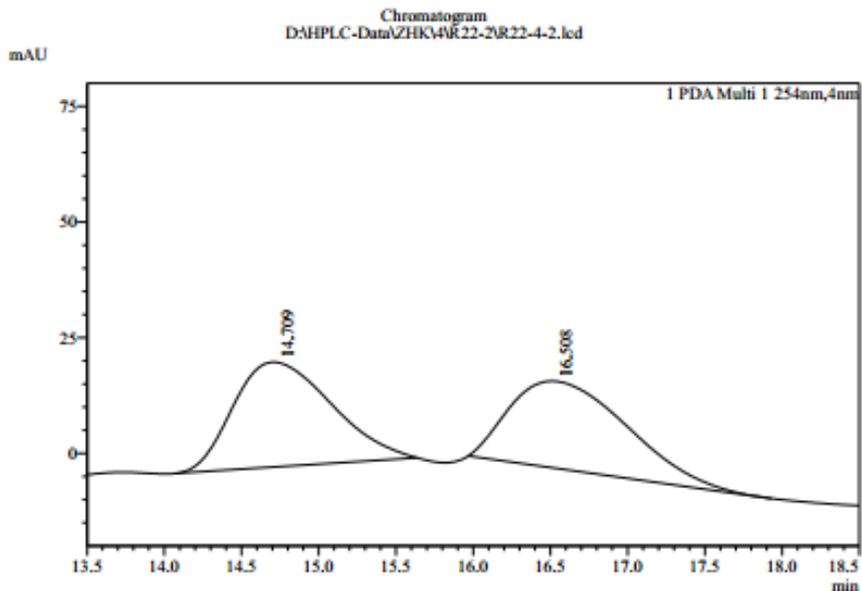
Enantiomerically enriched **5'b**

(S)-diethyl (5-methoxy-1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl)

Phosphonate (5'c)



A white solid; m.p. = 99 - 101 °C; 31.8 mg, 95% yield; $[\alpha]_D^{25} = -18.17$ (*c* 0.3, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.70 (dd, *J* = 8.4, 0.8 Hz, 1H), 7.19 (s, 1H), 7.12 - 7.7.0 (m, 1H), 4.98 (d, *J* = 11.6 Hz, 1H), 4.32 - 4.20(m, 2H), 4.15 - 3.95 (m, 2H), 3.89 (s, 3H), 1.33 (t, *J* = 7.2 Hz, 3H), 1.22 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 163.69 (d, *J* = 2.8 Hz), 135.01 (d, *J* = 5.4 Hz), 127.12 (d, *J* = 5.6 Hz), 123.03 (d, *J* = 1.8 Hz), 117.52 (d, *J* = 2.4 Hz), 109.63 (d, *J* = 2.6 Hz), 64.55 (d, *J* = 6.9 Hz), 64.25 (d, *J* = 7.1 Hz), 56.07, 54.00 (d, *J* = 159.9 Hz), 16.57 (d, *J* = 5.5 Hz), 16.41 (d, *J* = 5.7 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 16.62; HRMS (ESI) *m/z* calcd for C₁₂H₁₈NO₆PS [M+Na]⁺ = 358.0485, found = 358.0479; The ee value was 94%, *t_R* (minor) = 14.7 min, *t_R* (major) = 16.8 min (Chiralcel AS-H, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 0.6 mL/min).

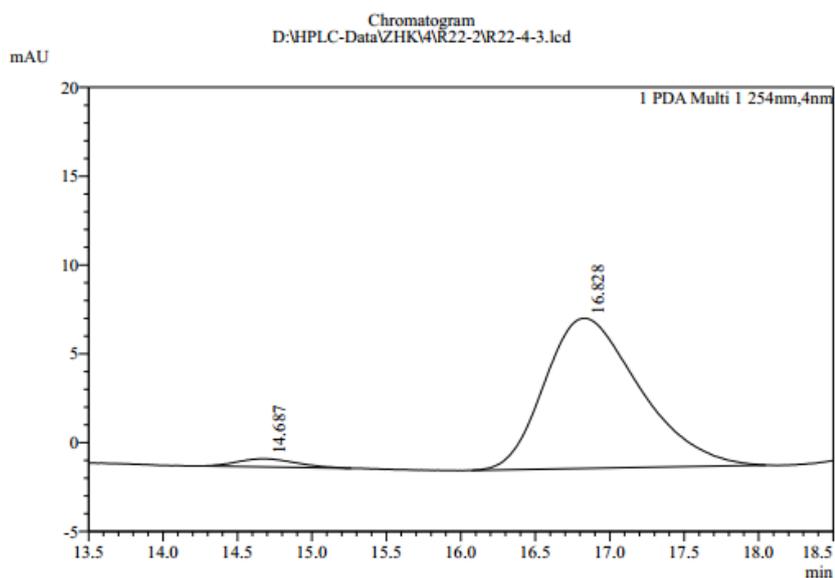


Peak Table

PDA Ch1 254nm

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|---------|---------|
| 1 | 14.709 | 22690 | 54.823 | 1003933 | 50.067 |
| 2 | 16.508 | 18698 | 45.177 | 1001233 | 49.933 |
| Total | | 41388 | 100.000 | 2005167 | 100.000 |

Racemic 5'**c**



Peak Table

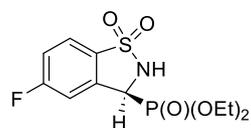
PDA Ch1 254nm

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|--------|---------|
| 1 | 14.687 | 455 | 5.106 | 12135 | 3.134 |
| 2 | 16.828 | 8449 | 94.894 | 375044 | 96.866 |
| Total | | 8904 | 100.000 | 387178 | 100.000 |

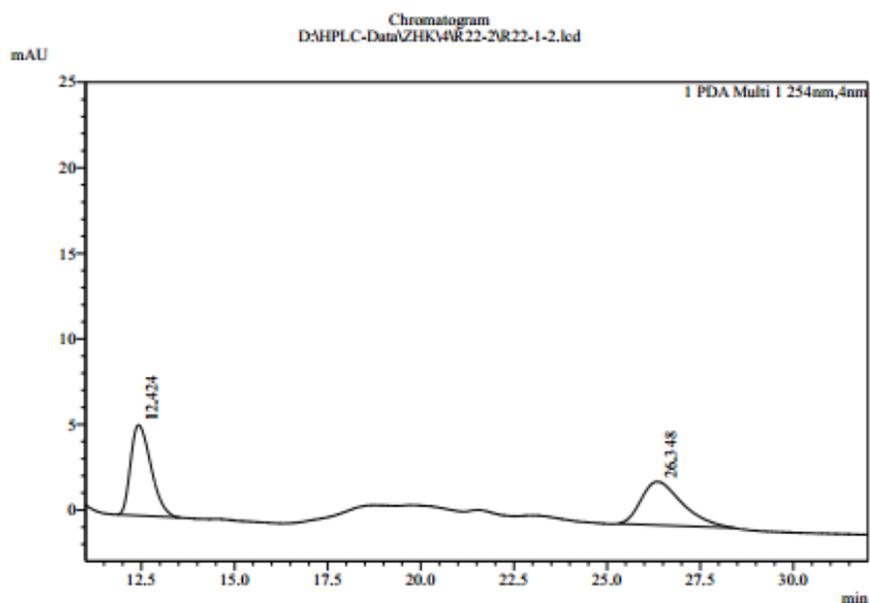
Enantiomerically enriched 5'**c**

(S)-diethyl (5-fluoro-1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl)phosphonate

(5'd)



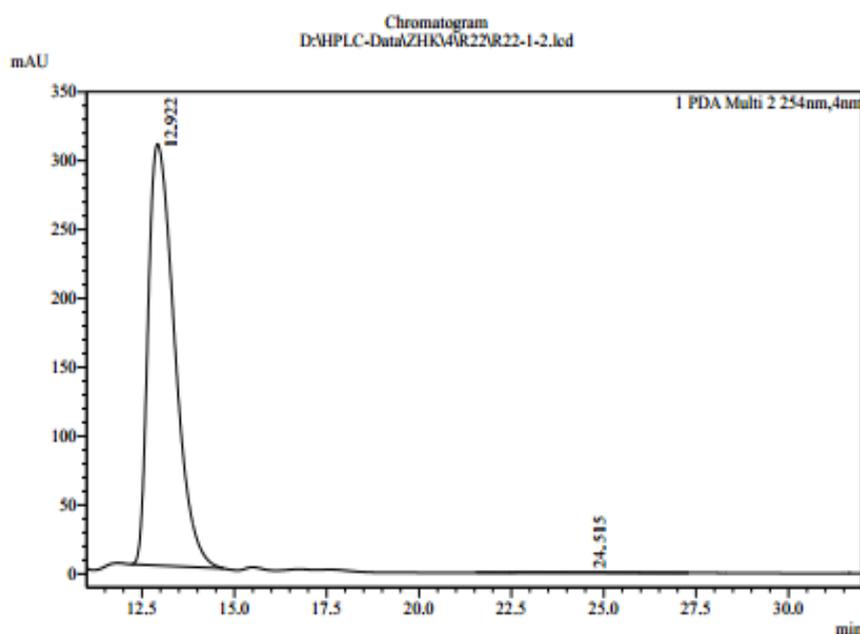
A white solid; m.p. = 95 - 97 °C; 29.4 mg, 91% yield; $[\alpha]_D^{25} = -15.47$ (c 0.3, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.70 - 7.61 (m, 1H), 7.52 (d, $J = 7.6$ Hz, 1H), 7.28 - 7.19 (m, 1H), 6.36 - 5.85 (m, 1H), 5.05 (d, $J = 12.0$ Hz, 1H), 4.34 - 4.20 (m, 2H), 4.18 - 4.00 (m, 2H), 1.32 (t, $J = 7.2$ Hz, 3H), 1.26 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 135.81 (d, $J = 2.5$ Hz), 135.75 (d, $J = 1.2$ Hz), 121.79 (d, $J = 3.1$ Hz), 121.75 (d, $J = 3.0$ Hz), 116.97 (d, $J = 2.4$ Hz), 116.79 (d, $J = 2.4$ Hz), 65.00 (d, $J = 7.0$ Hz), 64.27 (d, $J = 7.3$ Hz), 54.29 (d, $J = 162.7$ Hz), 16.55 (d, $J = 5.4$ Hz), 16.42 (d, $J = 5.6$ Hz); ^{31}P NMR (162 MHz, CDCl_3) δ 16.11; ^{19}F NMR (376 MHz, CDCl_3) δ -114.73; HRMS (ESI) m/z calcd for $\text{C}_{11}\text{H}_{15}\text{NO}_5\text{PSF}$ $[\text{M}+\text{Na}]^+ = 346.0279$, found = 346.0272; The ee value was >99%, t_R (major) = 12.9 min, t_R (minor) = 24.5 min (Chiralcel AS-H, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 0.6 mL/min).



Peak Table

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|--------|---------|
| 1 | 12.424 | 5292 | 67.588 | 201772 | 50.841 |
| 2 | 26.348 | 2538 | 32.412 | 195093 | 49.159 |
| Total | | 7830 | 100.000 | 396866 | 100.000 |

Racemic 5'd



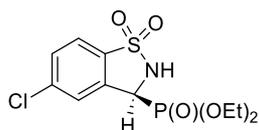
Peak Table

| PDA Ch2 254nm | | | | | |
|---------------|-----------|--------|---------|----------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 12.922 | 305938 | 99.886 | 15174901 | 99.583 |
| 2 | 24.515 | 349 | 0.114 | 63557 | 0.417 |
| Total | | 306287 | 100.000 | 15238457 | 100.000 |

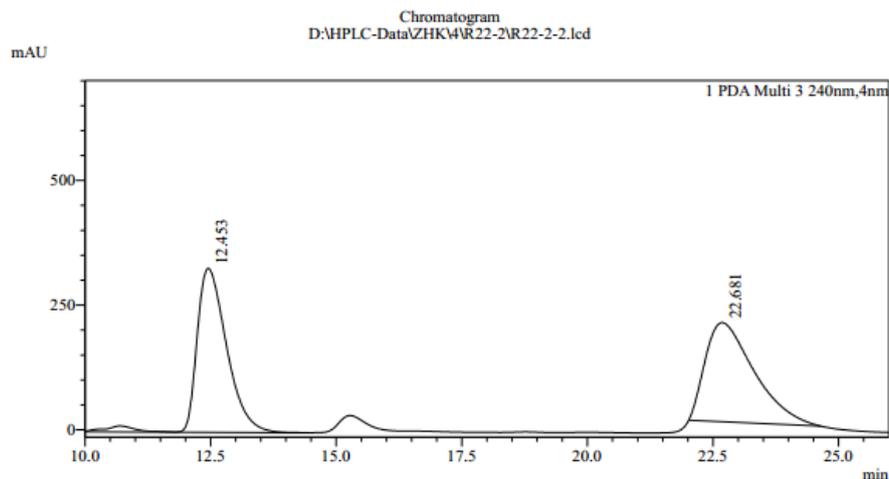
Enantiomerically enriched 5'd

(S)-diethyl (5-chloro-1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl)phosphonate

(5'e)



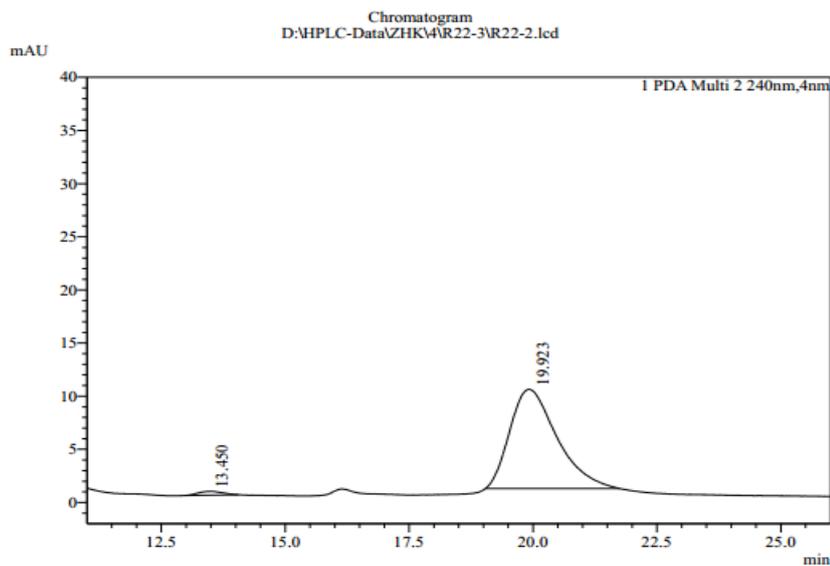
A white solid; m.p. = 91 - 93 °C; 30.8 mg, 91% yield; $[\alpha]_D^{25} = -12.67$ (c 0.3, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.75 - 7.65 (m, 2H), 7.57 - 7.51 (m, 1H), 5.02 (d, $J = 12.0$ Hz, 1H), 4.30 - 4.20 (m, 2H), 4.20 - 4.09 (m, 3H), 1.29 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 139.74 (d, $J = 3.0$ Hz), 134.62 (d, $J = 5.4$ Hz), 133.75 (d, $J = 5.3$ Hz), 130.60 (d, $J = 2.5$ Hz), 126.17 (d, $J = 2.7$ Hz), 122.84 (d, $J = 1.9$ Hz), 64.69 (dd, $J = 99.7, 7.9$ Hz), 54.71, 53.08, 16.52 (d, $J = 5.5$ Hz), 16.43 (d, $J = 5.4$ Hz); ^{31}P NMR (162 MHz, CDCl_3) δ 16.34; HRM (ESI) m/z calcd for $\text{C}_{11}\text{H}_{15}\text{NO}_5\text{PSCl}$ $[\text{M}+\text{Na}]^+ = 361.9984$, found $[\text{M}+\text{Na}]^+ = 361.9980$; The ee value was 96%, t_R (minor) = 13.5 min, t_R (major) = 19.9 min (Chiralcel IC, $\lambda = 240$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|----------|---------|
| 1 | 12.453 | 328357 | 62.316 | 13837779 | 50.729 |
| 2 | 22.681 | 198563 | 37.684 | 13440273 | 49.271 |
| Total | | 526921 | 100.000 | 27278052 | 100.000 |

Racemic 5'e



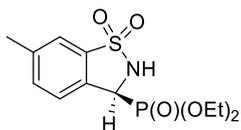
Peak Table

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|--------|---------|
| 1 | 13.450 | 355 | 3.666 | 11913 | 1.886 |
| 2 | 19.923 | 9339 | 96.334 | 619648 | 98.114 |
| Total | | 9694 | 100.000 | 631561 | 100.000 |

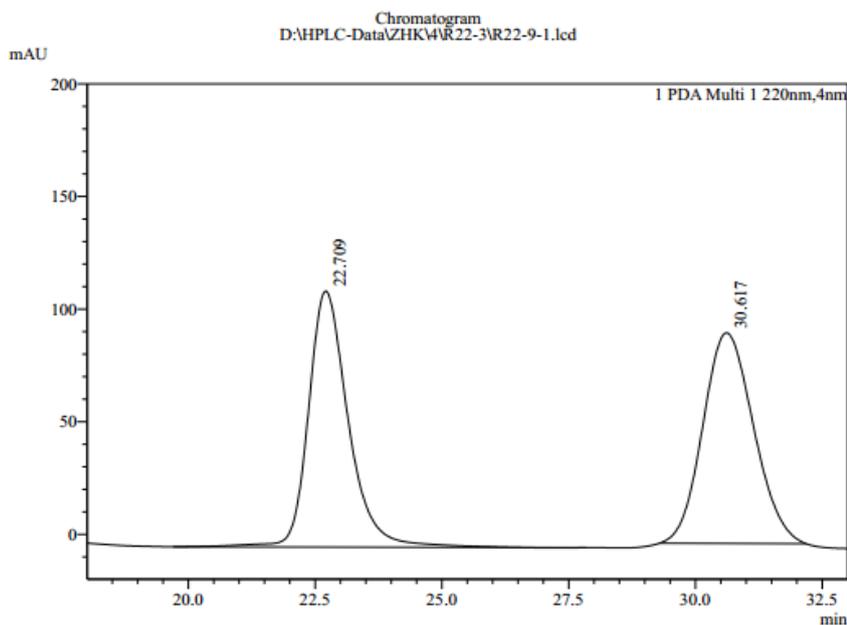
Enantiomerically enriched 5'e

(S)-diethyl (6-methyl-1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl)

Phosphonate (5'f)



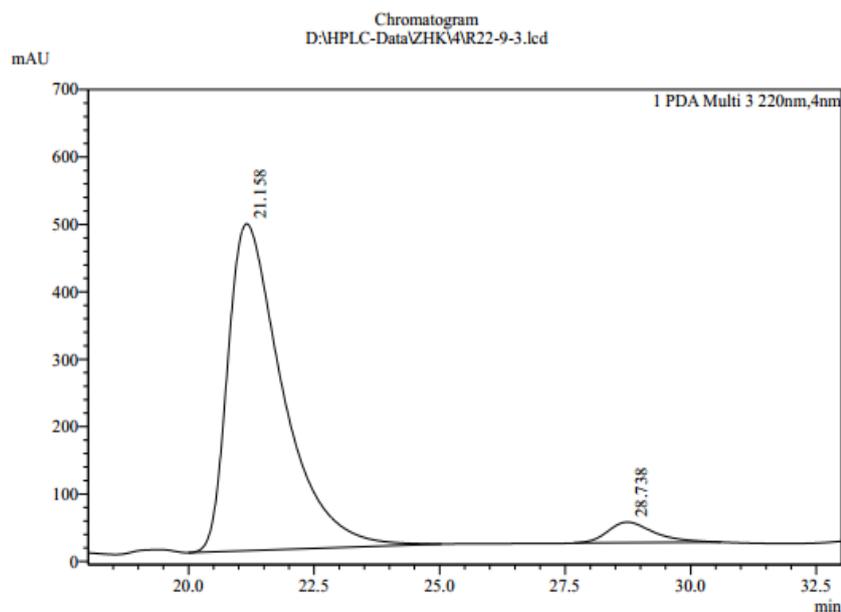
A white solid; m.p. = 92- 94 °C; 29.0 mg, 91% yield; $[\alpha]_D^{25} = -30.47$ (*c* 0.5, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.67 - 7.58 (m, 2H), 7.46 (d, *J* = 8.0 Hz, 1H), 5.59 - 5.41 (m, 1H), 5.12 - 4.88 (m, 1H), 4.34 - 4.15 (m, 2H), 4.13 - 3.92 (m, 2H), 2.47 (s, 3H), 1.31 (t, *J* = 7.2 Hz, 3H), 1.21 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 140.86 (d, *J* = 2.8 Hz), 135.10 (d, *J* = 5.7 Hz), 134.52 (d, *J* = 2.8 Hz), 129.44 (d, *J* = 5.3 Hz), 125.73 (d, *J* = 2.9 Hz), 121.63 (d, *J* = 1.8 Hz), 64.50 (d, *J* = 6.8 Hz), 64.15 (d, *J* = 7.2 Hz), 54.01 (d, *J* = 160.5 Hz), 21.45, 16.56 (d, *J* = 5.7 Hz), 16.41 (d, *J* = 5.5 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 16.86; HRMS (ESI) *m/z* calcd for C₁₂H₁₈NO₅PS [M+Na]⁺ = 342.0530, found = 342.0526; The ee value was 90%, *t_R* (major) = 21.2 min, *t_R* (minor) = 28.7 min (Chiralcel AS-H, λ = 220 nm, 40% *i*-PrOH/hexanes, flow rate = 0.6 mL/min).



Peak Table

| PDA Ch1 220nm | | | | | |
|---------------|-----------|--------|---------|----------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 22.709 | 113726 | 54.868 | 6237783 | 48.944 |
| 2 | 30.617 | 93545 | 45.132 | 6506981 | 51.056 |
| Total | | 207271 | 100.000 | 12744764 | 100.000 |

Racemic **5f**

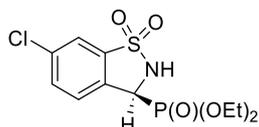


Peak Table

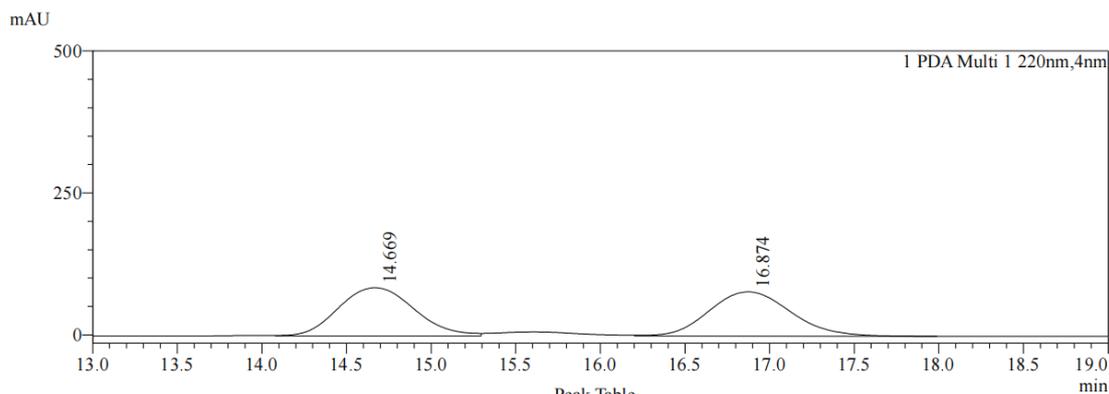
| PDA Ch3 220nm | | | | | |
|---------------|-----------|--------|---------|----------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 21.158 | 485099 | 94.087 | 36936207 | 94.950 |
| 2 | 28.738 | 30486 | 5.913 | 1964357 | 5.050 |
| Total | | 515586 | 100.000 | 38900565 | 100.000 |

Enantiomerically enriched **5'f**

(S)-diethyl (6-chloro-1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl) phosphonate (5'g)



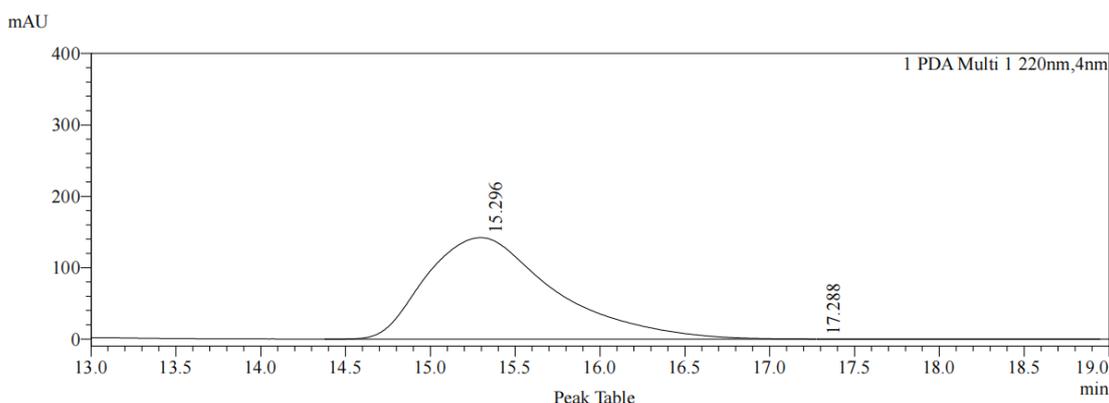
A white solid; m.p. = 87 - 89 °C; 30.5 mg, 90% yield; $[\alpha]_D^{25} = -34.18$ (*c* 0.5, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.72 (d, *J* = 7.6 Hz, 1H), 7.64 (d, *J* = 8.0 Hz, 1H), 7.56 - 7.47 (m, 1H), 6.29 (s, 1H), 5.10 (d, *J* = 10.0 Hz, 1H), 4.26 - 4.07 (m, 4H), 1.31 (t, *J* = 7.2 Hz, 3H), 1.21 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 137.31 (d, *J* = 4.5 Hz), 134.06 (d, *J* = 2.6 Hz), 131.83 (d, *J* = 3.9 Hz), 131.52 (d, *J* = 2.9 Hz), 131.46 (d, *J* = 6.5 Hz), 120.30 (d, *J* = 2.3 Hz), 64.80 (d, *J* = 7.1 Hz), 64.00 (d, *J* = 7.2 Hz), 54.69 (d, *J* = 154.5 Hz), 16.42 (d, *J* = 2.4 Hz), 16.36 (d, *J* = 2.1 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 17.03; HRMS (ESI) *m/z* calcd for C₁₁H₁₅NO₅PSCl [M+Na]⁺ = 361.9984, found = 361.9980; The ee value was 99%, *t_R* (major) = 15.3 min, *t_R* (minor) = 17.3 min (Chiralcel AS-H, λ = 220 nm, 40% *i*-PrOH/hexanes, flow rate = 0.6 mL/min).



Peak Table

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|---------|---------|
| 1 | 14.669 | 85255 | 52.202 | 2667639 | 50.330 |
| 2 | 16.874 | 78064 | 47.798 | 2632654 | 49.670 |
| Total | | 163318 | 100.000 | 5300293 | 100.000 |

Racemic 5'g



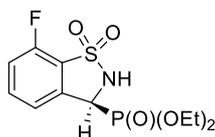
Peak Table

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|---------|---------|
| 1 | 15.296 | 142188 | 99.799 | 7501820 | 99.799 |
| 2 | 17.288 | 286 | 0.201 | 15086 | 0.201 |
| Total | | 142474 | 100.000 | 7516906 | 100.000 |

Enantiomerically enriched 5'g

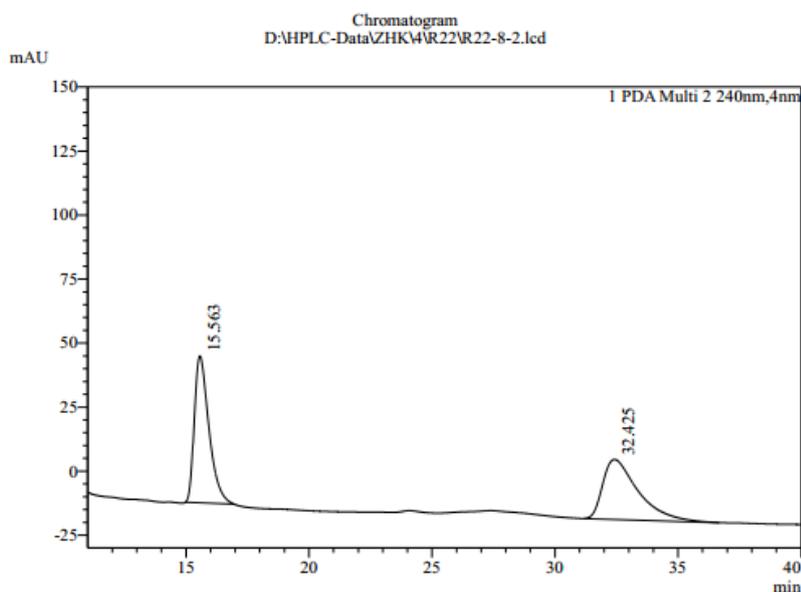
(S)-diethyl (7-fluoro-1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl)

phosphonate (5'h)



A white solid; m.p. = 85 - 87 °C; 29.1 mg, 90% yield; $[\alpha]_D^{25} = -40.00$ (c 0.7, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.79 (dd, $J = 8.4, 4.4$ Hz, 1H), 7.39 (d, $J = 8.4$ Hz, 1H), 7.32 - 7.23 (m, 1H), 6.60 (s, 1H), 5.04 (d, $J = 12.0$ Hz, 1H), 4.33 - 4.20 (m, 2H), 4.19 - 4.07 (m, 2H), 1.32 - 1.29 (t, $J = 7.2$ Hz, 3H), 1.29 - 1.25 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR

(100 MHz, CDCl₃) δ 165.39 (dd, $J = 253.5, 2.9$ Hz), 135.72 (dd, $J = 10.1, 5.7$ Hz), 131.27 (dd, $J = 5.2, 2.4$ Hz), 123.91 (dd, $J = 9.9, 1.8$ Hz), 118.16 (d, $J = 2.1$ Hz), 118.04 (d, $J = 2.2$ Hz), 113.19 (dd, $J = 24.9, 2.5$ Hz), 65.19 (d, $J = 7.1$ Hz), 64.15 (dd, $J = 24, 2.1$ Hz), 53.93 (d, $J = 162.6$ Hz), 16.51 (d, $J = 5.4$ Hz), 16.43 (d, $J = 5.7$ Hz); ³¹P NMR (162 MHz, CDCl₃) δ 16.24; ¹⁹F NMR (376 MHz, CDCl₃) δ -103.79; HRMS (ESI) m/z calcd for C₁₁H₁₅NO₅PSF [M+Na]⁺ = 346.0279, found = 346.0276; The ee value was 90%, t_R (major) = 15.7 min, t_R (minor) = 31.3 min (Chiralcel AS-H, $\lambda = 240$ nm, 40% *i*-PrOH/hexanes, flow rate = 0.6 mL/min).

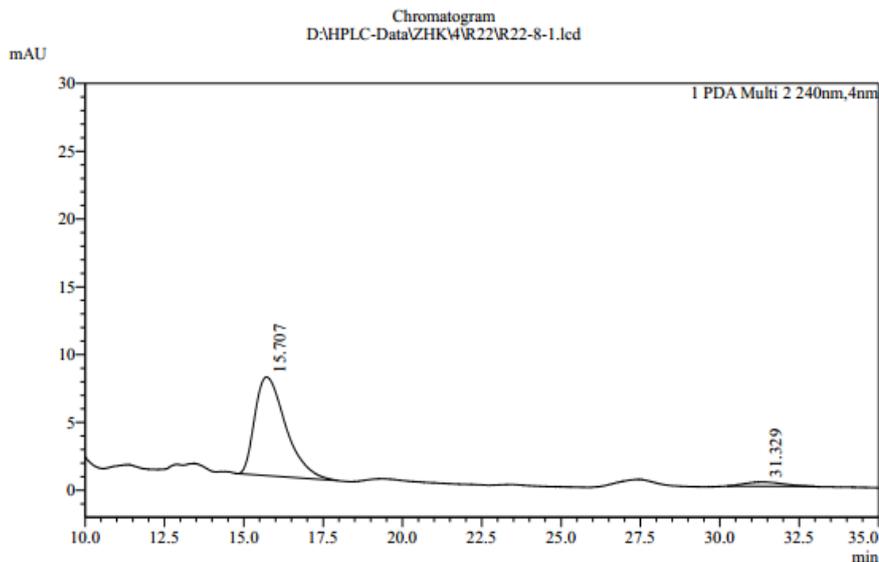


Peak Table

PDA Ch2 240nm

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|---------|---------|
| 1 | 15.563 | 57312 | 70.995 | 2364546 | 50.289 |
| 2 | 32.425 | 23415 | 29.005 | 2337344 | 49.711 |
| Total | | 80726 | 100.000 | 4701891 | 100.000 |

Racemic **5'h**



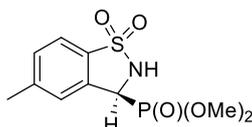
Peak Table

| PDA Ch2 240nm | | | | | |
|---------------|-----------|--------|---------|--------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 15.707 | 7269 | 95.871 | 493607 | 94.956 |
| 2 | 31.329 | 313 | 4.129 | 26219 | 5.044 |
| Total | | 7582 | 100.000 | 519826 | 100.000 |

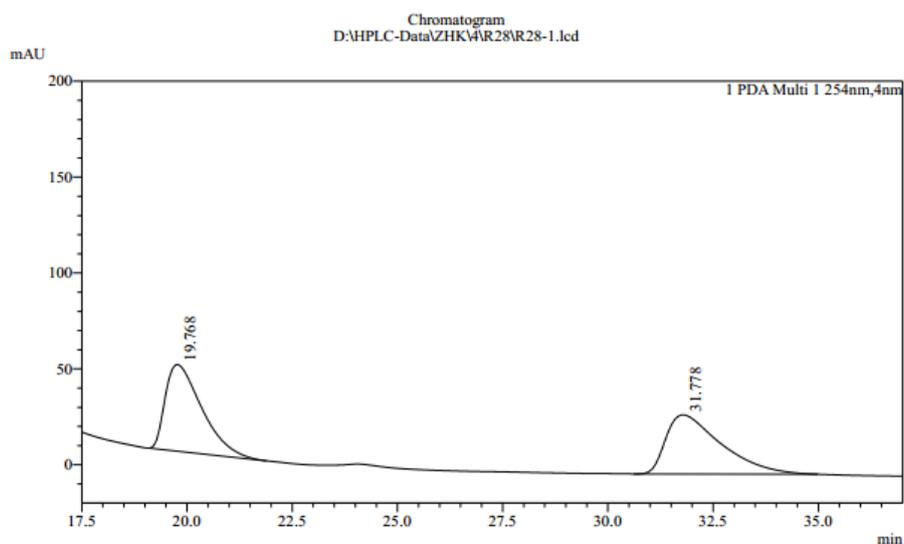
Enantiomerically enriched **5'h**

(S)-dimethyl (5-methyl-1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl)

Phosphonate (5'i)



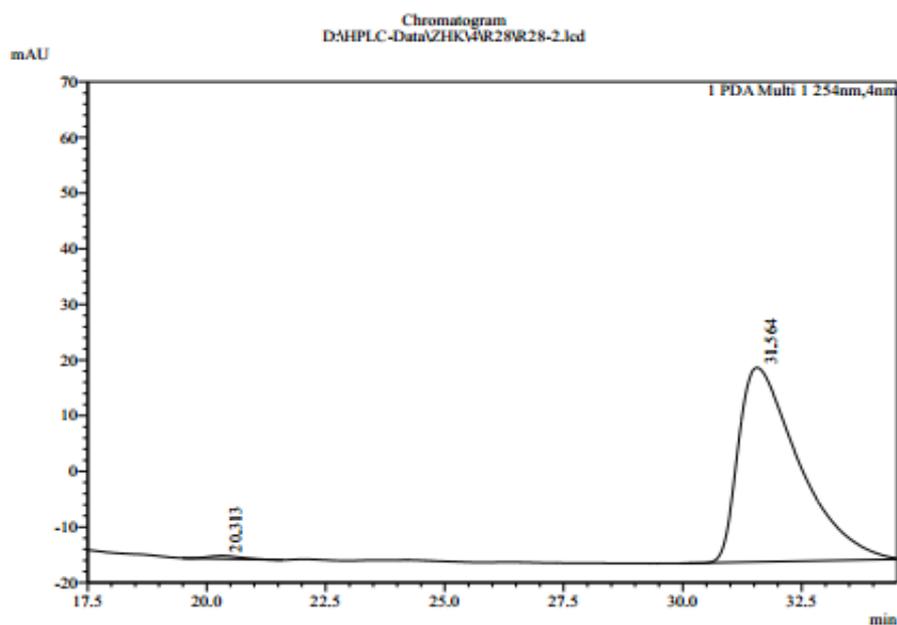
A white solid; m.p. = 89 - 91 °C; 27.1 mg, 93% yield; $[\alpha]_{D}^{25} = -20.50$ (c 0.5, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.68 (d, $J = 8.0$ Hz, 1H), 7.49 (s, 1H), 7.38 (d, $J = 8.0$ Hz, 1H), 6.14 (s, 1H), 5.05 (d, $J = 11.2$ Hz, 1H), 3.87 (d, $J = 10.4$ Hz, 3H), 3.72 (d, $J = 10.8$ Hz, 3H), 2.48 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 144.56 (d, $J = 4.8$ Hz), 132.44 (d, $J = 1.8$ Hz), 132.38 (d, $J = 2.4$ Hz), 131.21 (d, $J = 2.3$ Hz), 126.14 (d, $J = 2.8$ Hz), 121.46 (d, $J = 1.9$ Hz), 55.07 (d, $J = 7.0$ Hz), 54.38 (d, $J = 7.3$ Hz), 53.77 (d, $J = 162.6$ Hz), 21.97; ^{31}P NMR (162 MHz, CDCl_3) δ 19.22; HRMS (ESI) m/z calcd for $\text{C}_{10}\text{H}_{14}\text{NO}_5\text{PS}$ $[\text{M}+\text{Na}]^+ = 314.0217$, found = 314.0211; The ee value was 98%, t_R (minor) = 20.3 min, t_R (major) = 31.6 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| PDA Ch1 254nm | | | | | |
|---------------|-----------|--------|---------|---------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 19.768 | 45179 | 59.430 | 2732101 | 49.258 |
| 2 | 31.778 | 30842 | 40.570 | 2814446 | 50.742 |
| Total | | 76020 | 100.000 | 5546547 | 100.000 |

Racemic 5'i



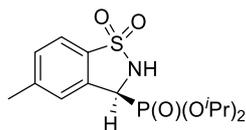
Peak Table

| PDA Ch1 254nm | | | | | |
|---------------|-----------|--------|---------|---------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 20.313 | 561 | 1.581 | 26587 | 0.852 |
| 2 | 31.564 | 34913 | 98.419 | 3093789 | 99.148 |
| Total | | 35474 | 100.000 | 3120376 | 100.000 |

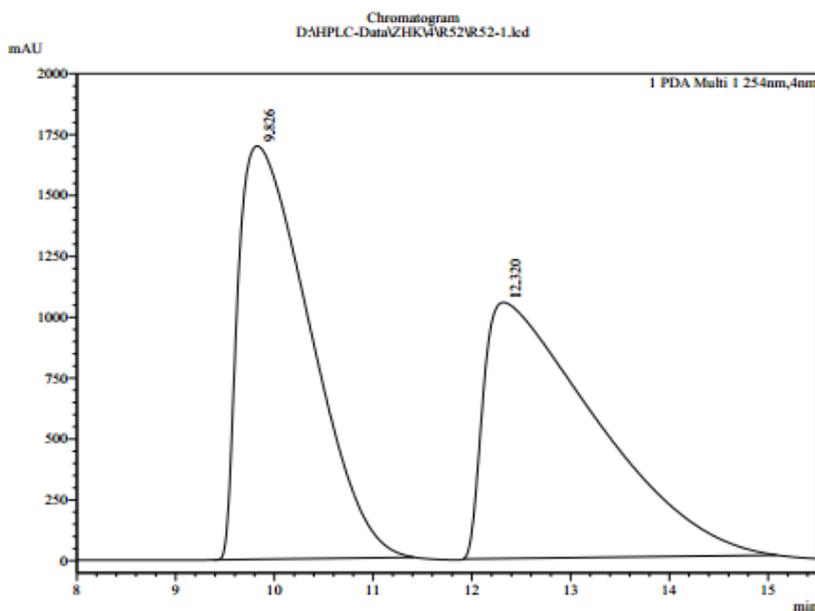
Enantiomerically enriched 5'i

(S)-diisopropyl (5-methyl-1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl)

Phosphonate (5'i)



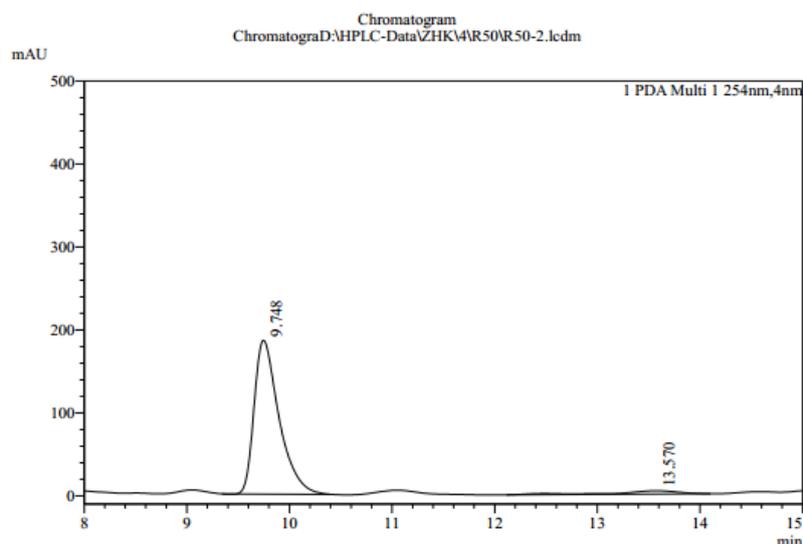
A white solid; m.p. = 88 - 90 °C; 31.9 mg, 92% yield; $[\alpha]_D^{25} = -11.00$ (c 0.3, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.67 (d, $J = 8.0$ Hz, 1H), 7.57 (s, 1H), 7.37 (d, $J = 8.0$ Hz, 1H), 5.46 (s, 1H), 4.93 (dd, $J = 11.6, 2.8$ Hz, 1H), 4.86 - 4.75 (m, 1H), 4.65 - 4.48 (m, 1H), 2.47 (s, 3H), 1.35 (d, $J = 6.0$ Hz, 3H), 1.30 (d, $J = 2.8$ Hz, 3H), 1.29 (d, $J = 2.8$ Hz, 3H), 1.03 (d, $J = 6.4$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 144.35 (d, $J = 2.6$ Hz), 132.93 (d, $J = 5.8$ Hz), 132.52 (d, $J = 5.7$ Hz), 130.98 (d, $J = 2.4$ Hz), 126.11 (d, $J = 2.8$ Hz), 121.35 (d, $J = 1.9$ Hz), 73.38 (d, $J = 14.5$ Hz), 73.38, 54.47 (d, $J = 159.5$ Hz), 24.25 (d, $J = 1.4$ Hz), 24.22 (d, $J = 1.6$ Hz), 23.94 (d, $J = 5.2$ Hz), 23.51 (d, $J = 5.1$ Hz), 21.97; ^{31}P NMR (162 MHz, CDCl_3) δ 14.85; HRMS (ESI) m/z calcd for $\text{C}_{14}\text{H}_{22}\text{NO}_5\text{PS}$ $[\text{M}+\text{Na}]^+ = 370.0843$, found = 370.0838; The ee value was 90%, t_R (minor) = 9.8 min, t_R (major) = 12.3 min (Chiralcel IC, $\lambda = 254$ nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|---------|---------|-----------|---------|
| 1 | 9.826 | 1696845 | 61.745 | 85262324 | 50.557 |
| 2 | 12.320 | 1051289 | 38.255 | 83382437 | 49.443 |
| Total | | 2748133 | 100.000 | 168644761 | 100.000 |

Racemic **5j**



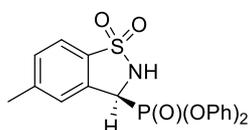
Peak Table

| PDA Ch1 254nm | | | | | |
|---------------|-----------|--------|---------|---------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 9.748 | 185462 | 97.939 | 3128663 | 95.027 |
| 2 | 13.570 | 3902 | 2.061 | 163715 | 4.973 |
| Total | | 189364 | 100.000 | 3292378 | 100.000 |

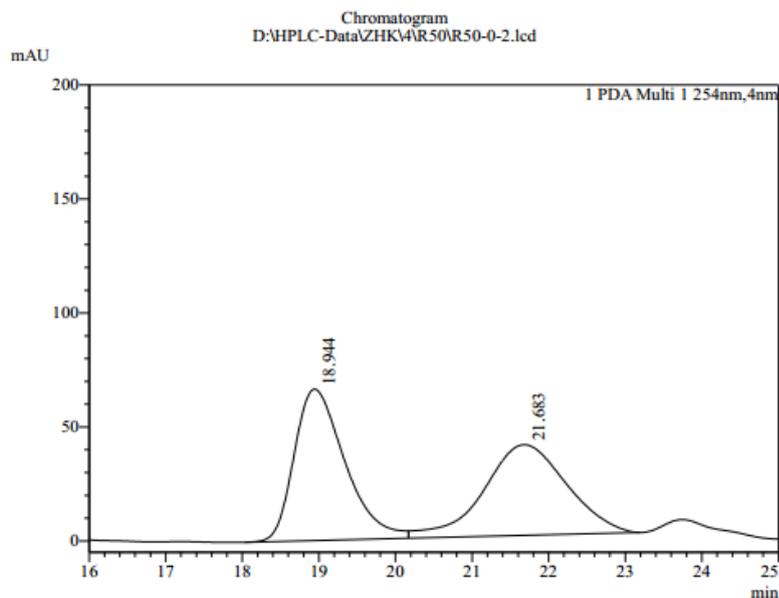
Enantiomerically enriched **5'j**

(S)-diphenyl (5-methyl-1,1-dioxido-2,3-dihydrobenzo[d]isothiazol-3-yl)

Phosphonate (5'k)



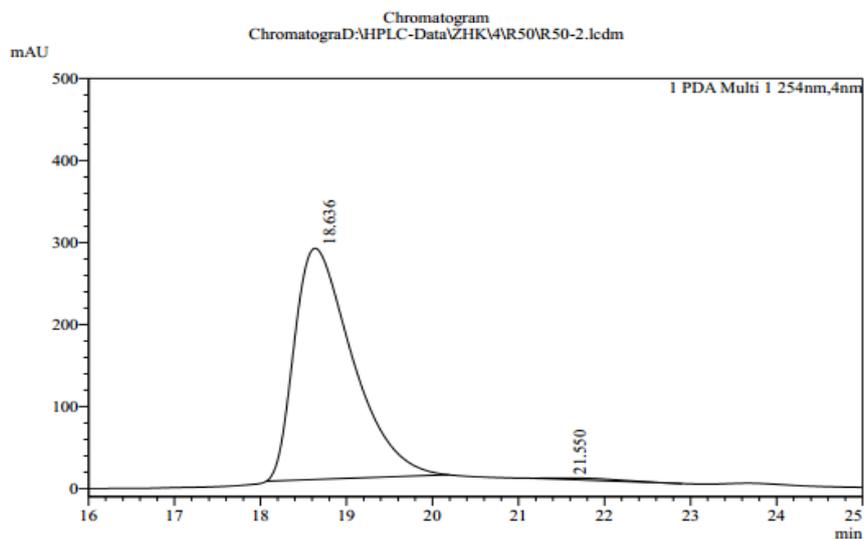
A white solid; m.p. = 91 - 93 °C; 39.4 mg, 95% yield; $[\alpha]_D^{25} = -17.43$ (*c* 0.3, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.72 (d, *J* = 8.0 Hz, 1H), 7.57 (s, 1H), 7.37 (d, *J* = 8.0 Hz, 1H), 7.33- 7.27 (m, 2H), 7.23 - 7.13 (m 5H), 7.12 - 7.05 (m, 1H), 7.01 - 6.93 (m, 2H), 5.73 (s, 1H), 5.30 (d, *J* = 9.2 Hz, 1H), 2.39 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 150.11 (d, *J* = 9.3 Hz), 149.85 (d, *J* = 9.3 Hz), 144.78 (d, *J* = 2.6 Hz), 132.45 (d, *J* = 6.2 Hz), 131.47 (d, *J* = 2.5 Hz), 131.30 (d, *J* = 5.1 Hz), 130.08, 129.77, 126.37 (d, *J* = 2.8 Hz), 125.98 (d, *J* = 0.9 Hz), 125.69, 121.61 (d, *J* = 2.0 Hz), 120.79 (d, *J* = 4.2 Hz), 120.46 (d, *J* = 4.3 Hz), 53.76 (d, *J* = 164.1 Hz), 21.86; ³¹P NMR (162 MHz, CDCl₃) δ 9.37; HRMS (ESI) *m/z* calcd for C₂₂H₁₈NO₅PS [M+Na]⁺ = 438.0530, found = 438.0526; The ee value was 98%, *t_R* (minor) = 18.6 min, *t_R* (major) = 21.6 min (Chiralcel IC, λ = 254 nm, 40% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|---------|---------|
| 1 | 18.944 | 66385 | 62.533 | 3058001 | 50.489 |
| 2 | 21.683 | 39774 | 37.467 | 2998769 | 49.511 |
| Total | | 106159 | 100.000 | 6056770 | 100.000 |

Racemic 5'k

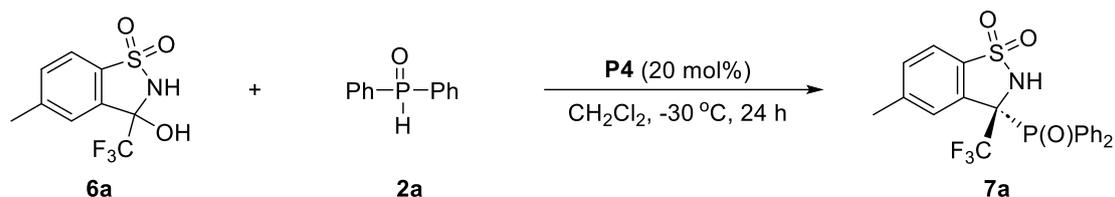


Peak Table

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|----------|---------|
| 1 | 18.636 | 281943 | 99.422 | 13254549 | 99.146 |
| 2 | 21.550 | 1640 | 0.578 | 114177 | 0.854 |
| Total | | 283583 | 100.000 | 13368726 | 100.000 |

Enantiomerically enriched 5'k

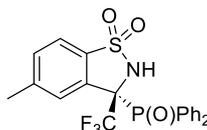
5.3 Representative procedure for the asymmetric P-nucleophile addition to CF₃-substituted cyclic N-sulfonyl amines 6 and phosphine oxide 2 and 3



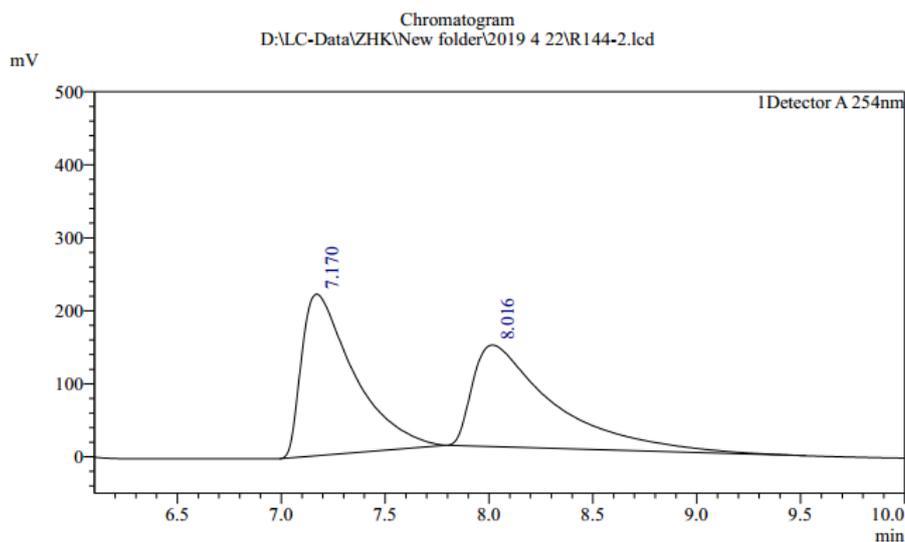
Representative procedure: To a flame-dried round bottle flask with a magnetic stirring bar were added the cyclic imine **6a** (0.1 mmol, 26.7 mg), Phosphine Oxide **2a** (0.12 mmol, 24.2 mg), and phosphonium salt **P4** (0.02 mmol, 11.3mg). followed by the addition of CH₂Cl₂ (1.0 ml). The reaction mixture was stirred at -30 °C for 24 h. The solvent was removed under reduced pressure, and the residue was purified by column chromatography on silica gel (CH₂Cl₂/ethyl acetate = 3:1) to afford **7a** (92% yield) as a white solid.

Condition b: carried out room temperature.

(S)-3-(diphenylphosphoryl)-5-methyl-3-(trifluoromethyl)-2,3-dihydrobenzo[d]isothiazole 1,1-dioxide (7a)



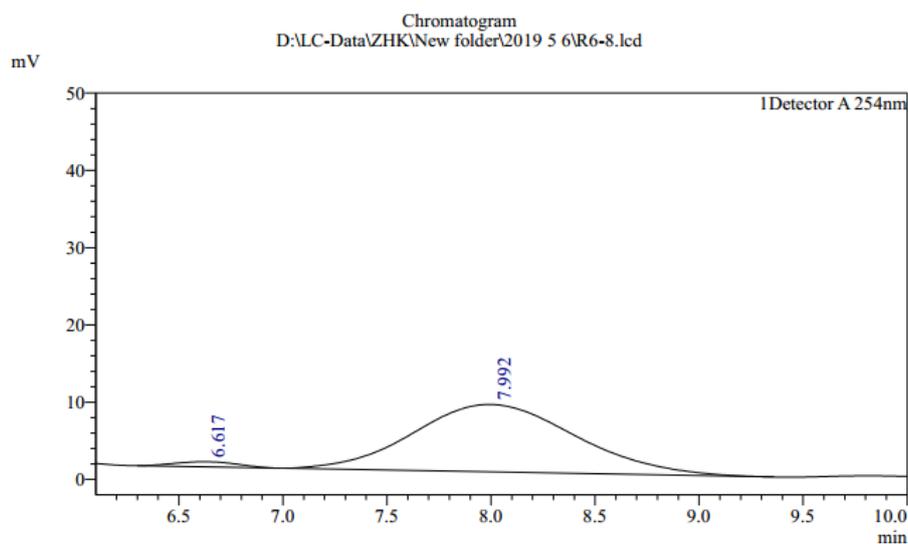
A white solid; m.p. = 85 - 87 °C; 41.5 mg, 92% yield; $[\alpha]_D^{25} = -12.32$ (*c* 0.6, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.13 - 8.04 (m, 2H), 7.98 - 7.86 (m, 3H), 7.55 - 7.47 (m, 2H), 7.47 - 7.27 (m, 6H), 7.00 (s, 1H), 2.43 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 145.26, 133.07 (d, *J* = 2.6 Hz), 132.94 (d, *J* = 8.8 Hz), 132.83 (d, *J* = 2.6 Hz), 132.48 (d, *J* = 8.1 Hz), 129.07, 128.88 (d, *J* = 12.2 Hz), 128.66, 128.31 (d, *J* = 12.1 Hz), 127.62 (d, *J* = 8.9 Hz), 126.54 (d, *J* = 2.8 Hz), 125.33 (d, *J* = 7.3 Hz), 122.51 (dd, *J* = 6.7, 1.3 Hz), 121.27, 21.79; ³¹P NMR (162 MHz, CDCl₃) δ 24.65; HRMS (ESI) *m/z* calcd for C₂₁H₁₇NO₃PSF₃ [M+Na]⁺ = 474.0517, found = 474.0525; The ee value was 94%, *t_R* (minor) = 6.6 min, *t_R* (major) = 8.0 min (Chiralcel IC, λ = 254 nm, 30% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 7.170 | 3881802 | 221096 | 61.397 | 50.879 |
| 2 | 8.016 | 3747666 | 139014 | 38.603 | 49.121 |
| Total | | 7629469 | 360110 | 100.000 | 100.000 |

Racemic **7a**

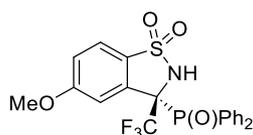


Peak Table

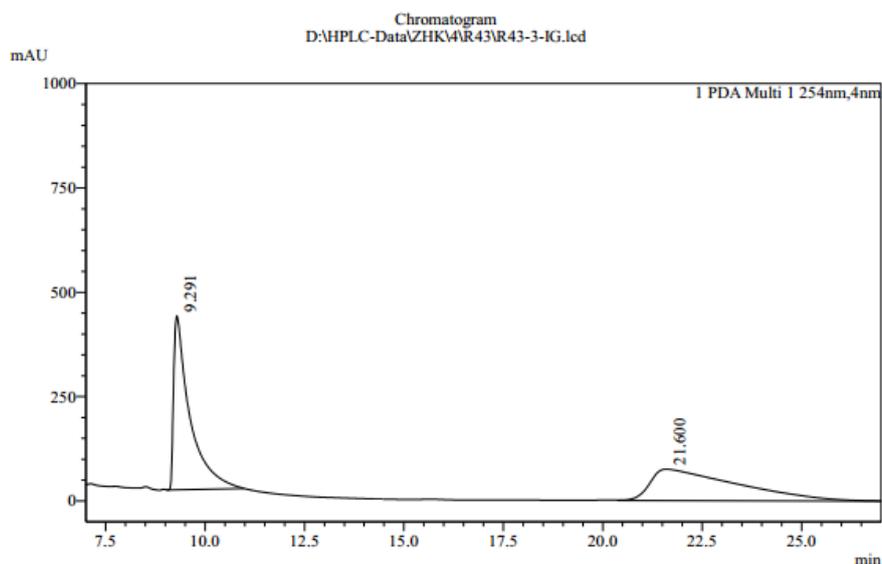
| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|--------|--------|---------|---------|
| 1 | 6.617 | 13975 | 669 | 7.114 | 2.857 |
| 2 | 7.992 | 475187 | 8728 | 92.886 | 97.143 |
| Total | | 489163 | 9397 | 100.000 | 100.000 |

Enantiomerically enriched **7a**

(S)-3-(diphenylphosphoryl)-5-methoxy-3-(trifluoromethyl)-2,3-dihydrobenzo[d]isothiazole 1,1-dioxide (7b)



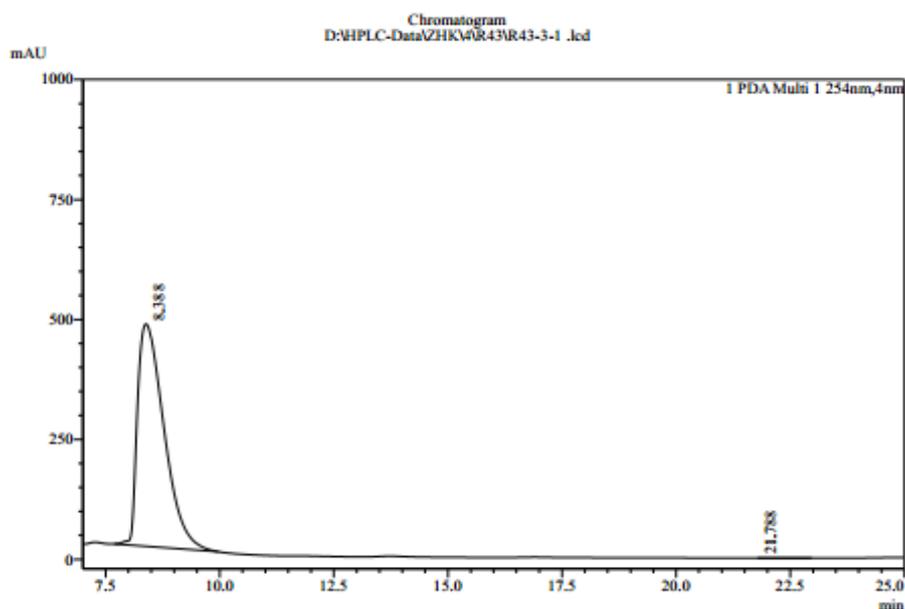
A white solid; m.p. = 88 - 90 °C; 44.4 mg, 95% yield; $[\alpha]_D^{25} = -14.17$ (c 0.6, CHCl_3); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.16 - 8.04 (m, 2H), 7.99 - 7.88 (m, 2H), 7.65 - 7.37 (m, 6H), 7.36 - 7.28 (m, 2H), 7.07 - 6.98 (m, 1H), 6.03 (s, 1H), 3.86 (s, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 163.94 (d, $J = 1.5$ Hz), 133.46 (d, $J = 2.8$ Hz), 133.15 (d, $J = 2.8$ Hz), 132.99 (d, $J = 8.9$ Hz), 132.48 (d, $J = 8.1$ Hz), 131.56 (d, $J = 1.5$ Hz), 129.11 (d, $J = 12.2$ Hz), 128.66 (d, $J = 12.1$ Hz), 127.64 (d, $J = 9.3$ Hz), 127.05 (d, $J = 2.7$ Hz), 126.63, 123.09, 119.75 (d, $J = 1.1$ Hz), 110.35, 56.50; $^{31}\text{P NMR}$ (162 MHz, CDCl_3) δ 24.93; HRMS (ESI) m/z calcd for $\text{C}_{21}\text{H}_{17}\text{NO}_4\text{PSF}_3$ $[\text{M}+\text{Na}]^+ = 490.0466$, found = 490.0466; The ee value was >99%, t_R (major) = 8.4 min, t_R (minor) = 21.8 min (Chiralcel IC, $\lambda = 254$ nm, 10% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| PDA Ch1 254nm | | | | | |
|---------------|-----------|--------|---------|----------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 9.291 | 415708 | 84.680 | 11820782 | 50.992 |
| 2 | 21.600 | 75209 | 15.320 | 11360881 | 49.008 |
| Total | | 490917 | 100.000 | 23181663 | 100.000 |

Racemic **7b**

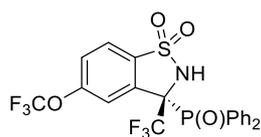


Peak Table

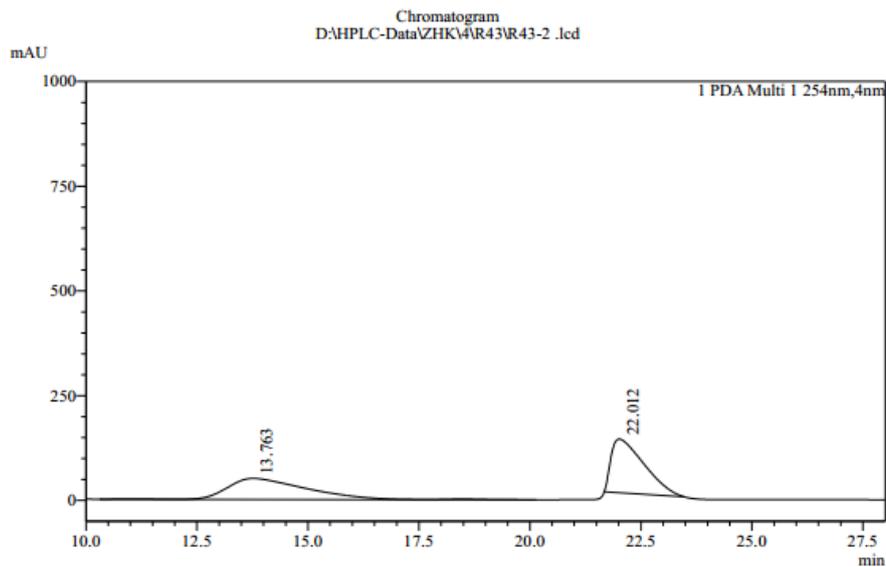
| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|----------|---------|
| 1 | 8.388 | 463398 | 100.000 | 18721825 | 99.997 |
| 2 | 21.788 | 1 | 0.000 | 576 | 0.003 |
| Total | | 463400 | 100.000 | 18722402 | 100.000 |

Enantiomerically enriched **7b**

(S)-3-(diphenylphosphoryl)-5-(trifluoromethoxy)-3-(trifluoromethyl)-2,3-dihydrobenzo[d]isothiazole 1,1-dioxide (7c)



A white solid; m.p. = 80 - 82 °C; 47.9 mg, 92% yield; $[\alpha]_D^{25} = -17.17$ (c 0.6, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.66 (s, 1H), 7.73 - 7.63 (m, 5H), 7.58 - 7.43 (m, 8H); ^{13}C NMR (100 MHz, CDCl_3) δ 132.67 (d, $J = 2.8$ Hz), 131.89, 131.62 (d, $J = 9.3$ Hz), 130.880, 130.77 (d, $J = 11.3$ Hz), 128.98 (d, $J = 12.7$ Hz), 120.75 (d, $J = 6.9$ Hz); ^{31}P NMR (162 MHz, CDCl_3) δ 21.59; HRMS (ESI) m/z calcd for $\text{C}_{21}\text{H}_{14}\text{NO}_4\text{PSF}_6$ $[\text{M}+\text{Na}]^+$ = 544.0183, found = 544.0194; The ee value was 98%, t_R (minor) = 13.7 min, t_R (major) = 21.5 min (Chiralcel IC, $\lambda = 254$ nm, 10% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).

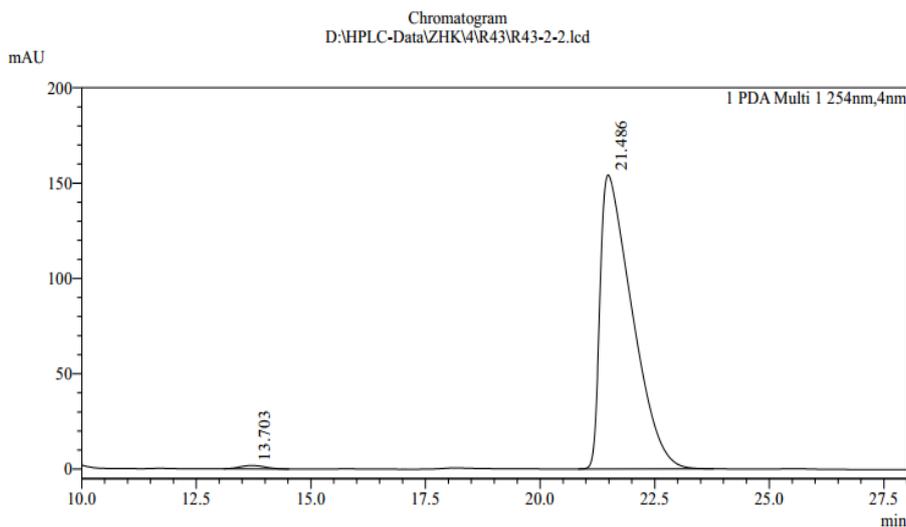


Peak Table

PDA Ch1 254nm

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|----------|---------|
| 1 | 13.763 | 50252 | 28.190 | 6194062 | 48.282 |
| 2 | 22.012 | 128006 | 71.810 | 6634778 | 51.718 |
| Total | | 178258 | 100.000 | 12828840 | 100.000 |

Racemic 7c



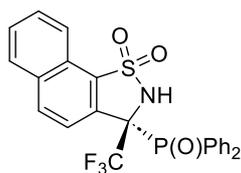
Peak Table

PDA Ch1 254nm

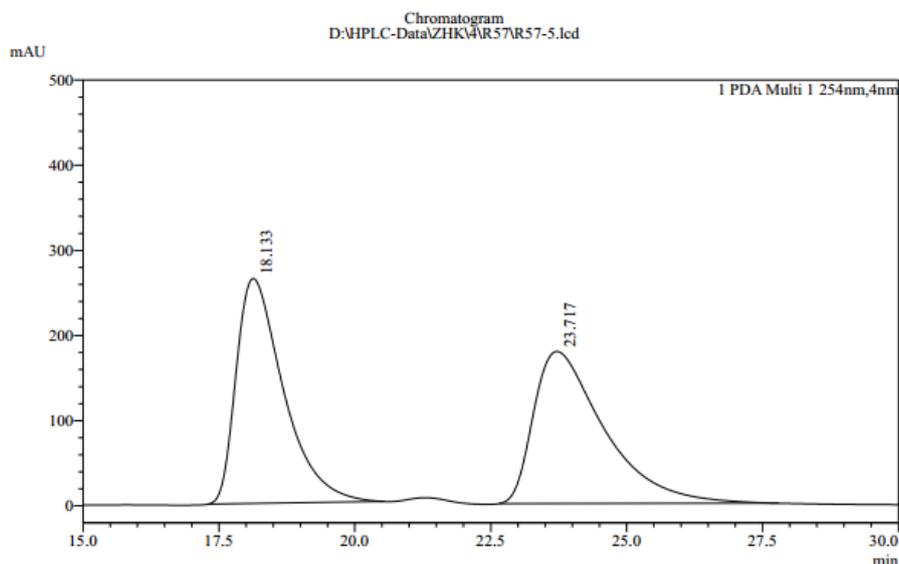
| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|---------|---------|
| 1 | 13.703 | 1756 | 1.126 | 65259 | 0.864 |
| 2 | 21.486 | 154115 | 98.874 | 7487308 | 99.136 |
| Total | | 155870 | 100.000 | 7552567 | 100.000 |

Enantiomerically enriched 7c

(S)-3-(diphenylphosphoryl)-3-(trifluoromethoxy)-2,3-dihydronaphtho[2,1-d]isothiazole 1,1-dioxide (7d)



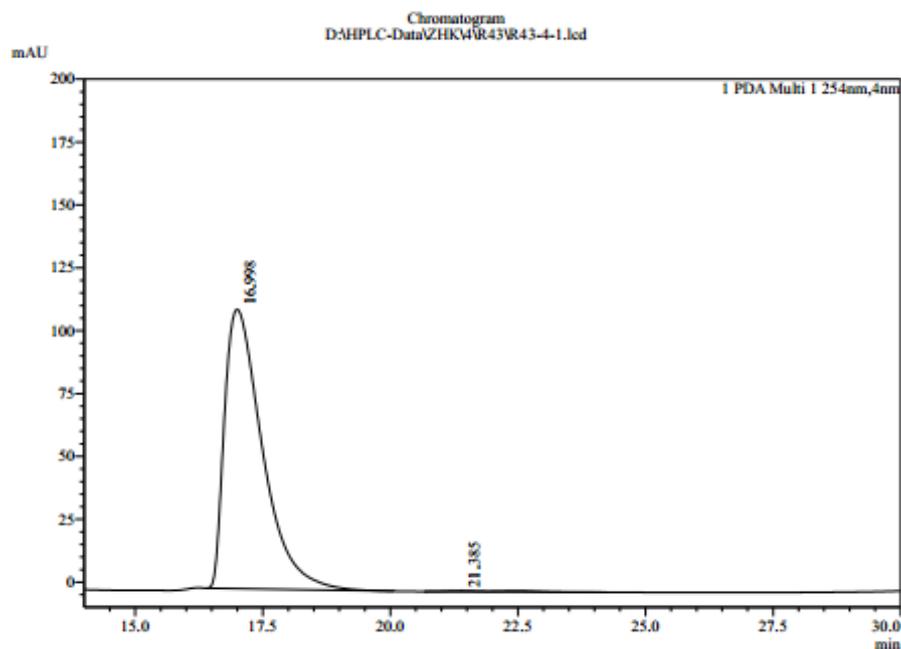
A white solid; m.p. = 95 - 97 °C; 44.8 mg, 92% yield; $[\alpha]_D^{25} = -20.83$ (c 0.6, CHCl_3); ^1H NMR (400 MHz, DMSO) δ 10.20 (s, 1H), 8.33 (d, $J = 8.7$ Hz, 1H), 8.21 - 8.05 (m, 5H), 7.84 - 7.63 (m, 7H), 7.35 - 7.18 (m, 3H); ^{13}C NMR (100 MHz, DMSO) δ 134.29, 133.65, 133.07 (d, $J = 2.5$ Hz), 132.72 (d, $J = 2.3$ Hz), 132.50 (d, $J = 8.7$ Hz), 131.77 (d, $J = 8.0$ Hz), 130.99 (d, $J = 3.6$ Hz), 129.63, 129.13, 128.95, 128.83, 128.16 (d, $J = 11.7$ Hz), 127.90, 127.75 (d, $J = 1.2$ Hz), 126.87, 123.81, 121.83 (d, $J = 4.8$ Hz); ^{31}P NMR (162 MHz, DMSO) δ 23.73; HRMS (ESI) m/z calcd for $\text{C}_{24}\text{H}_{17}\text{NO}_3\text{PSF}_3$ $[\text{M}+\text{Na}]^+ = 510.0517$, found = 510.0525; The ee value was 99%, t_R (minor) = 17.0 min, t_R (major) = 21.4min (Chiralcel IG, $\lambda = 254$ nm, 10% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| PDA Ch1 254nm | | | | | |
|---------------|-----------|--------|---------|----------|---------|
| Peak# | Ret. Time | Height | Height% | Area | Area% |
| 1 | 18.133 | 264142 | 59.673 | 15994126 | 49.778 |
| 2 | 23.717 | 178509 | 40.327 | 16136647 | 50.222 |
| Total | | 442651 | 100.000 | 32130774 | 100.000 |

Racemic **7d**



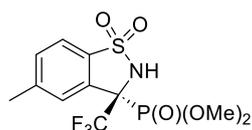
Peak Table

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|---------|---------|
| 1 | 16.998 | 111067 | 99.678 | 5742991 | 99.339 |
| 2 | 21.385 | 358 | 0.322 | 38195 | 0.661 |
| Total | | 111425 | 100.000 | 5781185 | 100.000 |

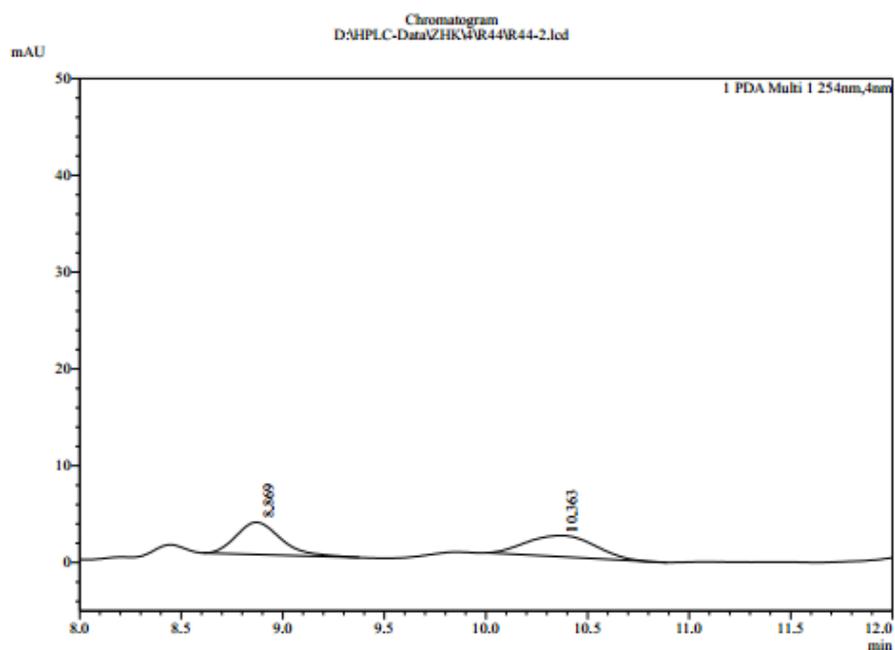
Enantiomerically enriched **7d**

(S)-dimethyl (5-methyl-1,1-dioxido-3-(trifluoromethyl)-2,3-

dihydrobenzo[d]isothiazol-3-yl)phosphonate (7e)



A white solid; m.p. = 85 - 87 °C; 33.0 mg, 92% yield; $[\alpha]_D^{25} = -9.67$ (*c* 0.7, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.76 (d, *J* = 8.0 Hz, 1H), 7.67 (s, 1H), 7.53 (d, *J* = 8.0 Hz, 1H), 5.88 (d, *J* = 6.8 Hz, 1H), 3.90 (d, *J* = 10.8 Hz, 3H), 3.67 (d, *J* = 10.8 Hz, 3H), 2.54 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 145.36 (d, *J* = 2.1 Hz), 141.47, 133.94, 132.98 (d, *J* = 1.8 Hz), 126.83 (d, *J* = 2.1 Hz), 121.96 (d, *J* = 1.3 Hz), 55.65 (dd, *J* = 23.5, 6.8 Hz), 29.85, 22.11; ³¹P NMR (162 MHz, CDCl₃) δ 12.99; HRMS (ESI) *m/z* calcd for C₁₁H₁₃NO₅PSF₃ [M+Na]⁺ = 382.0102, found = 382.0998; The ee value was 99%, *t_R* (minor) = 8.4 min, *t_R* (major) = 11.1 min (Chiralcel IC, λ = 254 nm, 10% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).

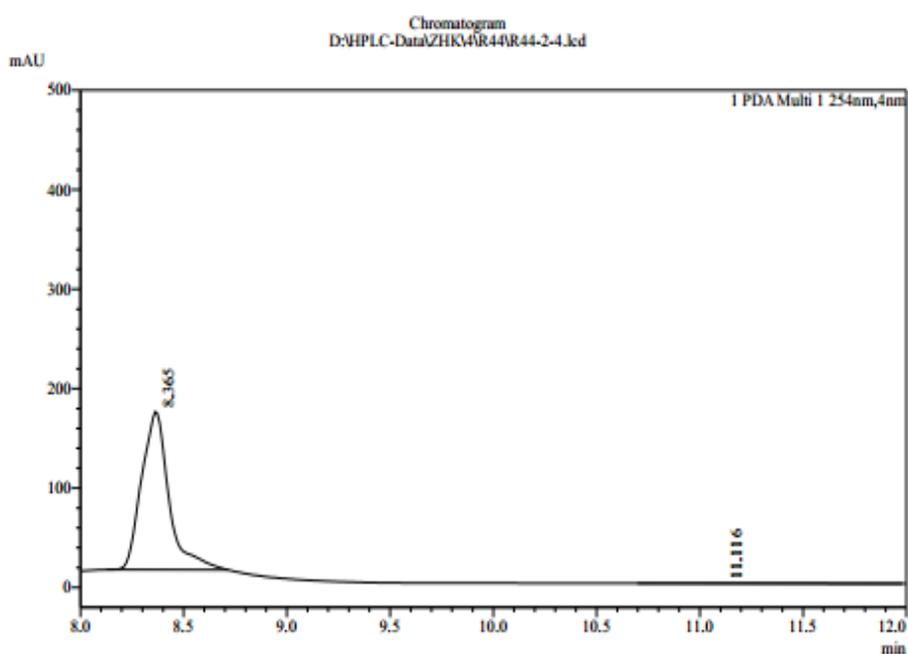


Peak Table

PDA Ch1 254nm

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|--------|---------|
| 1 | 8.869 | 3334 | 60.532 | 50969 | 50.043 |
| 2 | 10.363 | 2174 | 39.468 | 50881 | 49.957 |
| Total | | 5508 | 100.000 | 101850 | 100.000 |

Racemic 7e



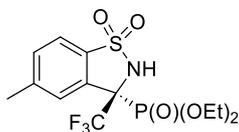
Peak Table

PDA Ch1 254nm

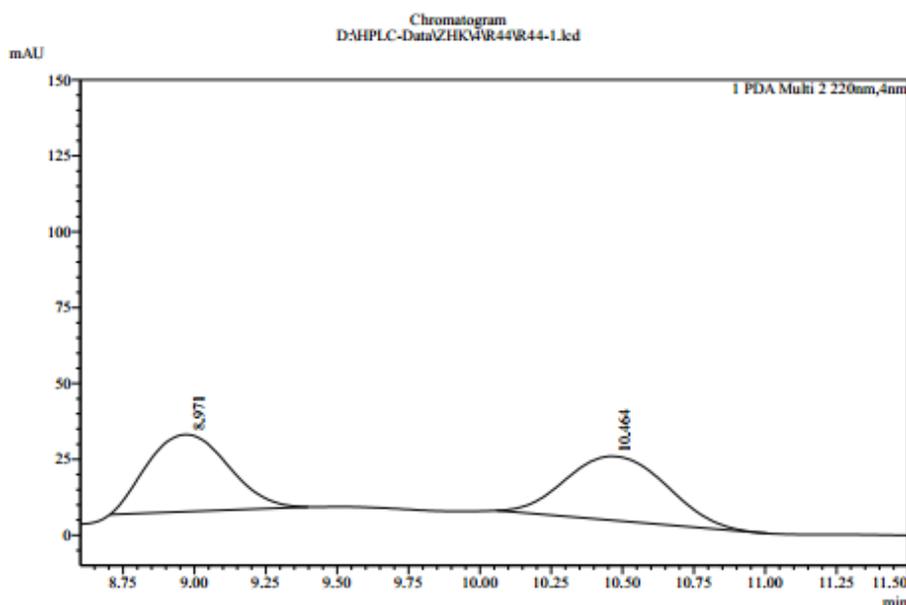
| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|---------|---------|
| 1 | 8.365 | 158972 | 99.908 | 1470203 | 99.494 |
| 2 | 11.116 | 147 | 0.092 | 7484 | 0.506 |
| Total | | 159119 | 100.000 | 1477687 | 100.000 |

Enantiomerically enriched 7e

(S)-diethyl (5-methyl-1,1-dioxido-3-(trifluoromethyl)-2,3-dihydrobenzo[d]isothiazol-3-yl)phosphonate (7f)



A white solid; m.p. = 79 - 91 °C; 35.2 mg, 91% yield; $[\alpha]_D^{25} = -10.50$ (*c* 0.6, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.00 - 7.78 (m, 1H), 7.67 - 7.64 (m, 1H), 7.30 (d, *J* = 8.0 Hz, 1H), 5.93 (s, 1H), 4.19 - 4.09 (m, 4H), 2.43 (d, *J* = 7.6 Hz, 3H), 1.36 (t, *J* = 7.2 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 144.20, 138.13, 130.33 (d, *J* = 4.2 Hz), 129.83, 128.92 (d, *J* = 10.0 Hz), 126.57, 67.20 (d, *J* = 32.0 Hz), 65.74 (d, *J* = 1.8 Hz), 62.19 (d, *J* = 5.7 Hz), 30.69, 29.83, 21.67, 16.41 (d, *J* = 6.2 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 10.55; HRMS (ESI) *m/z* calcd for C₁₃H₁₇NO₅PSF₃ [M+Na]⁺ = 410.0415, found = 410.0410; The ee value was 90%, *t_R* (minor) = 9.4 min, *t_R* (major) = 10.4 min (Chiralcel IC, λ = 220 nm, 10% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).

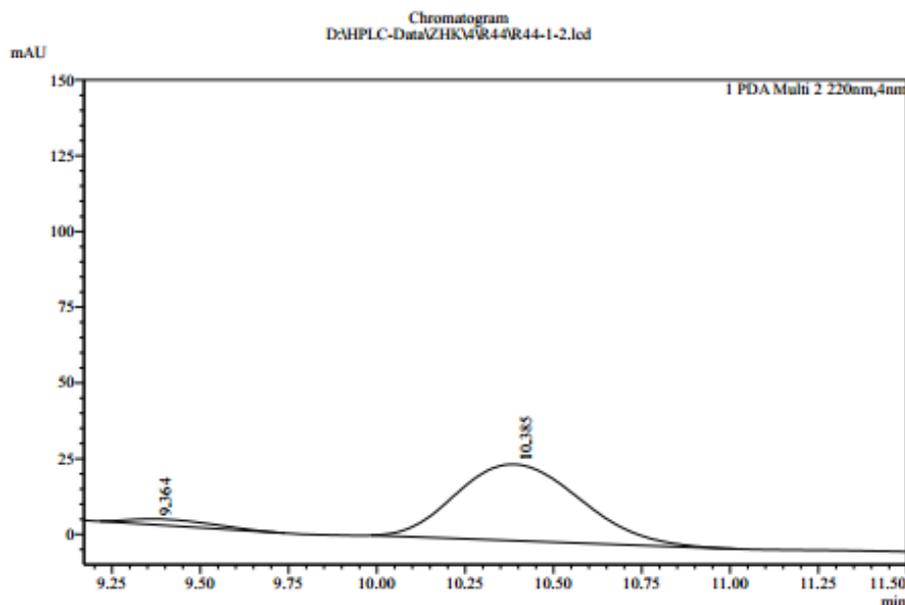


Peak Table

PDA Ch2 220nm

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|---------|---------|
| 1 | 8.971 | 25322 | 54.547 | 503207 | 49.119 |
| 2 | 10.464 | 21101 | 45.453 | 521255 | 50.881 |
| Total | | 46423 | 100.000 | 1024462 | 100.000 |

Racemic **7f**

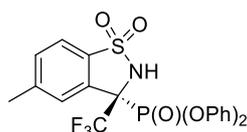


Peak Table

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|--------|---------|
| 1 | 9.364 | 1852 | 6.854 | 32798 | 5.047 |
| 2 | 10.385 | 25168 | 93.146 | 617099 | 94.953 |
| Total | | 27020 | 100.000 | 649897 | 100.000 |

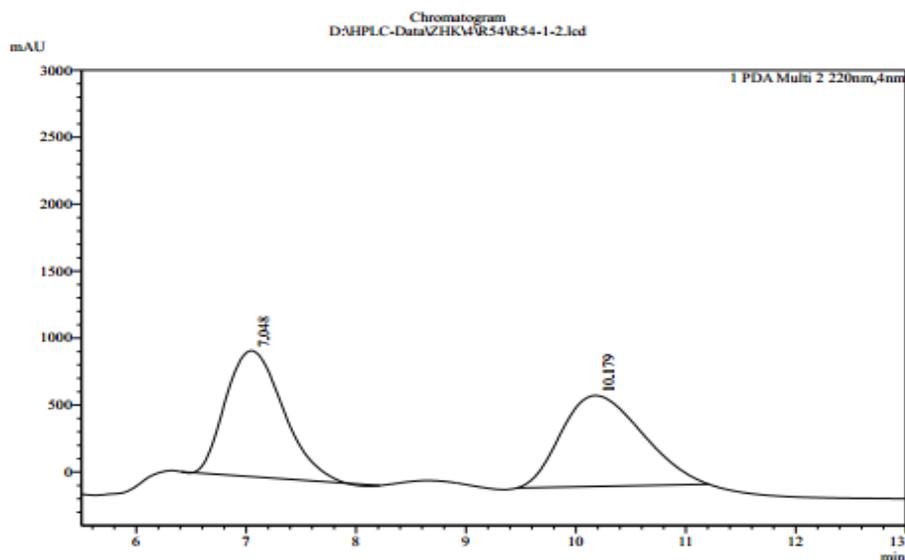
Enantiomerically enriched **7f**

(S)-diphenyl (5-methyl-1,1-dioxido-3-(trifluoromethyl)-2,3-dihydrobenzo[d]isothiazol-3-yl)phosphonate (7g)



A white solid; m.p. = 77 - 79 °C; 44.0 mg, 91% yield; $[\alpha]_D^{25} = -21.68$ (*c* 0.6, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.75 (d, *J* = 8.0 Hz, 1H), 7.66 (s, 1H), 7.43 (dd, *J* = 8.1, 0.5 Hz, 1H), 7.38 - 7.30 (m, 2H), 7.24 - 7.20 (m, 1H), 7.18 - 7.11 (m, 3H), 7.09 - 7.03 (m, 1H), 6.93 - 6.81 (m, 3H), 6.04 (d, *J* = 6.8 Hz, 1H), 2.34 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 155.75, 149.98 (dd, *J* = 84.6, 9.1 Hz), 145.48 (d, *J* = 2.3 Hz), 133.16 (d, *J* = 2.1 Hz), 132.78 (d, *J* = 5.4 Hz), 130.21 (d, *J* = 0.8 Hz), 129.72 (d, *J* = 8.2 Hz), 127.32, 126.41 (d, *J* = 1.1 Hz), 125.85, 122.02 (d, *J* = 1.4 Hz), 120.79, 120.49 (d, *J* = 4.3 Hz), 119.72 (d, *J* = 4.5 Hz), 115.43, 29.84, 21.83; ³¹P NMR (162 MHz, CDCl₃) δ 1.60; HRMS (ESI) *m/z* calcd for C₂₁H₁₇NO₅PSF₃ [M+Na]⁺ = 506.0415, found =

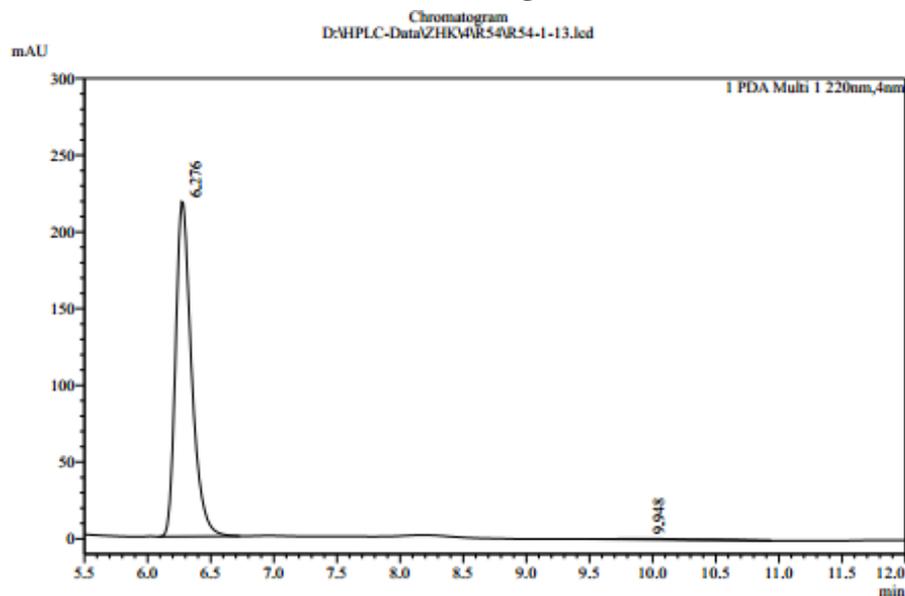
506.0420; The ee value was 97%, t_R (major) = 6.3 min, t_R (minor) = 9.9 min (Chiralcel IC, $\lambda = 220$ nm, 5% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|---------|---------|----------|---------|
| 1 | 7.048 | 940511 | 58.063 | 34207078 | 49.448 |
| 2 | 10.179 | 679294 | 41.937 | 34971334 | 50.552 |
| Total | | 1619805 | 100.000 | 69178412 | 100.000 |

Racemic **7g**

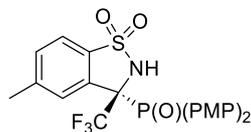


Peak Table

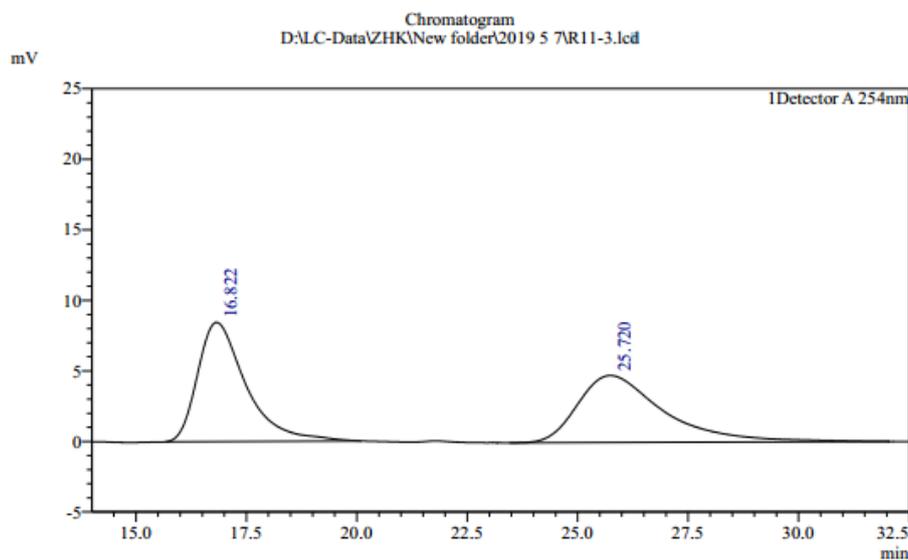
| Peak# | Ret. Time | Height | Height% | Area | Area% |
|-------|-----------|--------|---------|---------|---------|
| 1 | 6.276 | 218164 | 99.736 | 1921702 | 98.549 |
| 2 | 9.948 | 577 | 0.264 | 28292 | 1.451 |
| Total | | 218741 | 100.000 | 1949995 | 100.000 |

Enantiomerically enriched **7g**

(S)-3-(bis(4-methoxyphenyl)phosphoryl)-5-methyl-3-(trifluoromethyl)-2,3-dihydrobenzo[d]isothiazole 1,1-dioxide (7h)



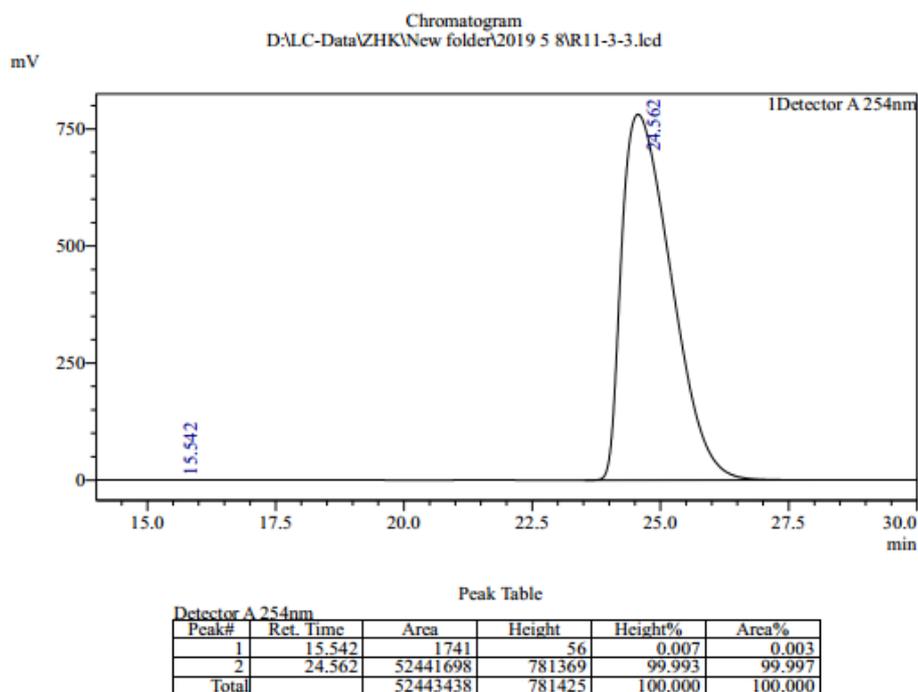
A white solid; m.p. = 81 - 83 °C; 38.6 mg, 92% yield; $[\alpha]_D^{25} = -10.43$ (*c* 0.7, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.99 - 7.91 (m, 2H), 7.82 - 7.75 (m, 3H), 7.54 (d, *J* = 8.0 Hz, 1H), 7.35 (d, *J* = 8.0 Hz, 1H), 7.05 - 6.98 (m, 2H), 6.84 - 6.75 (m, 2H), 5.37 (s, 1H), 3.87 (d, *J* = 1.6 Hz, 3H), 3.75 (d, *J* = 1.6 Hz, 3H), 2.43 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 163.49 (d, *J* = 2.8 Hz), 163.09 (d, *J* = 2.9 Hz), 145.16 (d, *J* = 0.9 Hz), 134.77 (d, *J* = 10.0 Hz), 134.31 (d, *J* = 9.6 Hz), 132.49, 132.37 (d, *J* = 2.7 Hz), 129.60, 127.36, 121.43, 118.68 (d, *J* = 24.7 Hz), 117.68, 114.59 (d, *J* = 13.1 Hz), 114.06 (d, *J* = 13.1 Hz), 55.55, 55.41, 21.90; ³¹P NMR (162 MHz, CDCl₃) δ 26.26; HRMS (ESI) *m/z* calcd for C₂₃H₂₁NO₅PSF₃ [M+Na]⁺ = 534.0728, found = 534.0720; The ee value was 99%, *t_R* (minor) = 15.5 min, *t_R* (major) = 24.6 min (Chiralcel IC, λ = 254 nm, 20% *i*-PrOH/hexanes, flow rate = 1.0 mL/min).



Peak Table

| Peak# | Ret. Time | Area | Height | Height% | Area% |
|-------|-----------|---------|--------|---------|---------|
| 1 | 16.822 | 642031 | 8446 | 64.040 | 50.108 |
| 2 | 25.720 | 639275 | 4743 | 35.960 | 49.892 |
| Total | | 1281305 | 13189 | 100.000 | 100.000 |

Racemic **7h**



Enantiomerically enriched **7h**

6. Investigation of the absolute stereochemistry

6.1. Determination of absolute configuration of **5'**

The absolute configuration of **5'b** and **5'j** were assigned to be *S* by comparing optical rotation of the same compounds reported in the literature (Scheme S4),^[6] and the configuration of **5'a**, **5'c** - **5'i** and **5'k** were assigned by analogy.

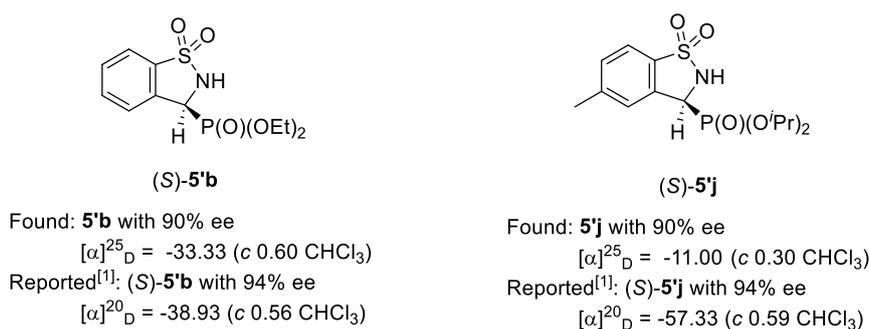


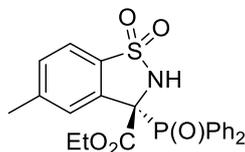
Figure S4. Optical rotation of **5'b** and **5'j** compared to reported

6.2. Determination of absolute configuration of **4**, **5** and **7**^[7].

In order to confirm the absolute configuration of compounds **4**, **5** and **7**. For four compounds, geometry optimization and electronic circular dichroism (ECD) spectrum calculations were performed at the PBE0-D3(BJ)/ 6-311+G(d,p) / SMD

(dichloromethane) level of theory,^[7b-7e] using Gaussian 09 program package.^[7f] The calculated ECD curves were generated using Multiwfn 3.8 software.^[7g]

A. Determination of absolute configuration of **4a**



The absolute configuration (AC) of **4a** was confirmed as *S* and detailed method was elaborated as following: the CD spectrum experiment of **4a** from the catalytic reaction was obtained, at the same time, the ECD spectra of (*S*)-**4a** were calculated. As shown in Figure S5, the simulated spectra are in good agreement with the experimental spectral data, and the *S* configuration could be reliably assigned to compound **4a**.

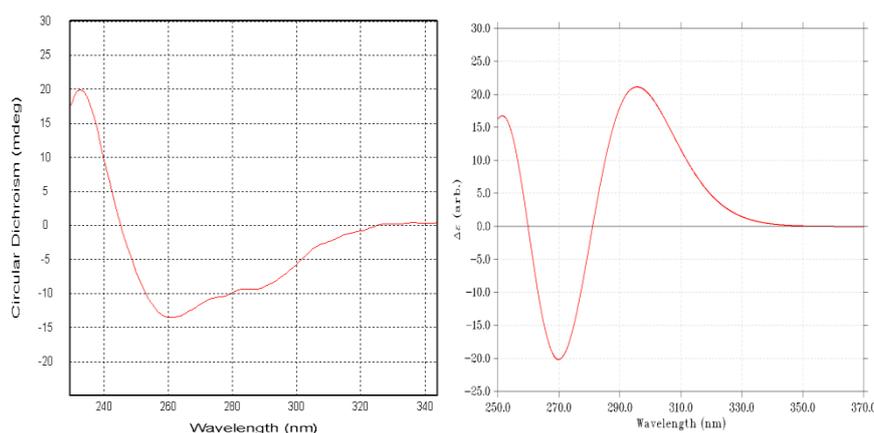
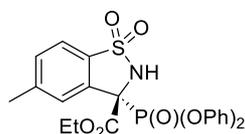


Figure S5. Experimental ECD spectra (left) and simulated spectra (right) proving *S*-conformer absolute configuration of **4a**.

B. Determination of absolute configuration of **5a**



As shown in Scheme S6, the simulated spectra of (*S*)-**5a** are in good agreement with the experimental spectral data, and the *S* configuration could be reliably assigned to compound **5a**.

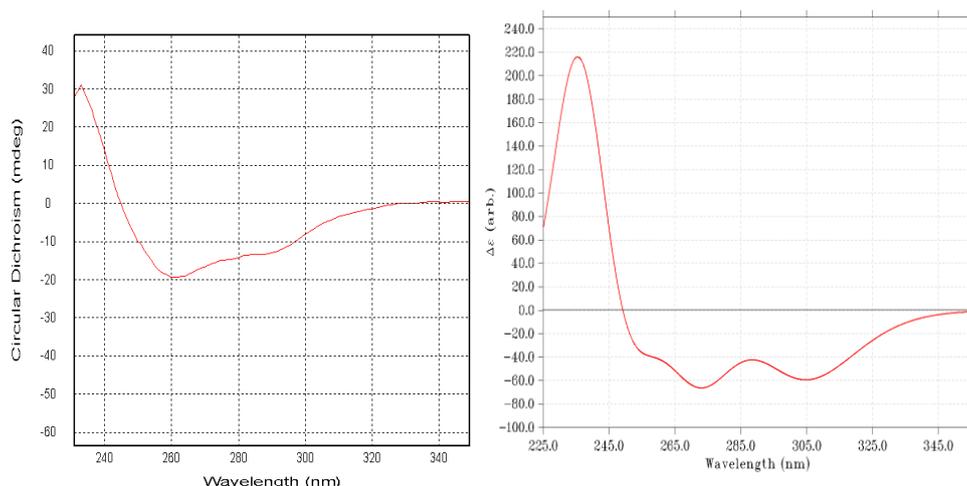


Figure S6. Experimental CD spectra (left) and simulated spectra (right) proving *S*-conformer **5a** absolute configuration

C. Determination of absolute configuration of **7a**

As shown in Scheme S7, the simulated spectra of (*S*)-**7a** are in good agreement with the experimental spectral data, and the *S* configuration could be reliably assigned to compound **7a**.

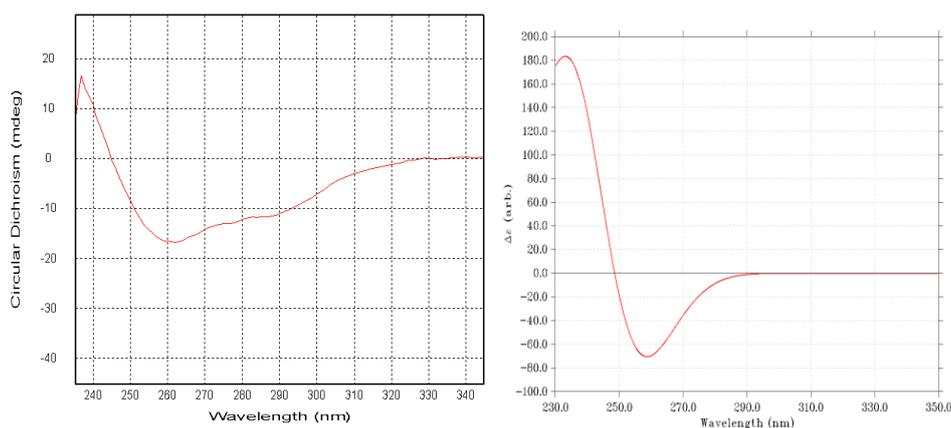
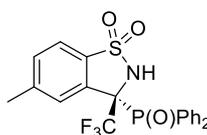
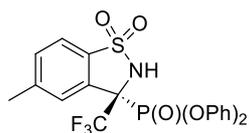


Figure S7. Experimental ECD spectra (left) and simulated spectra (right) proving *S*-conformer absolute configuration of **7a**.

D. Determination of absolute configuration of **7g**



As shown in Scheme S8, the simulated spectra of (*S*)-**7g** are in good agreement with the experimental spectral data, and the *S* configuration could be reliably assigned to compound **7g**.

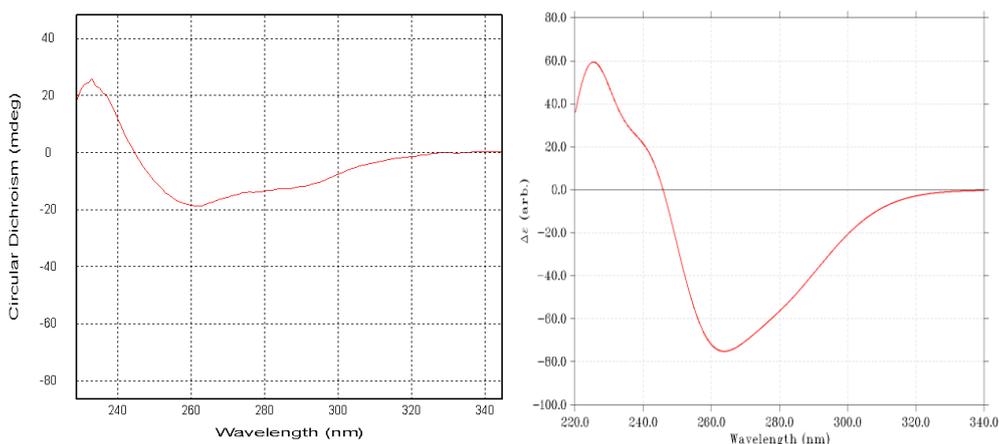


Figure S8. Experimental ECD spectra (left) and simulated spectra (right) proving *S*-conformer absolute configuration of **7g**.

6.3 Single crystal structure of catalyst **P4**

The absolute configuration of the catalyst **P4** was assigned as (*S*) by X-ray crystallographic analysis of a single crystal of **14** (Figure S9). CCDC 1935757 contains the supplementary crystallographic data of the chiral catalyst **P4** for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

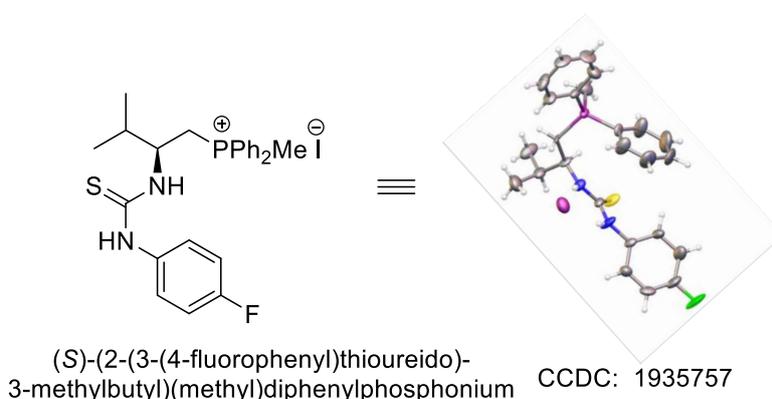


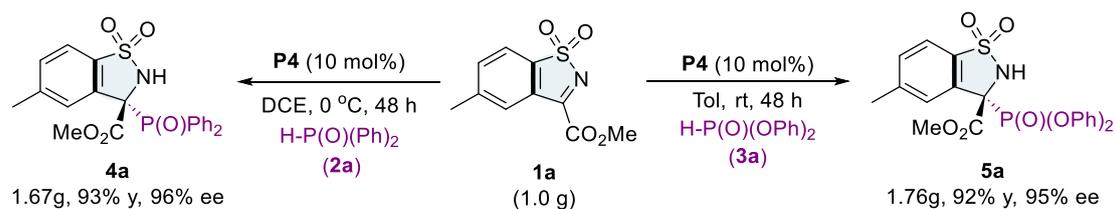
Figure S9. X-ray structure of **P4**

Table S6. Crystal data and structure refinement for **P4**

| | |
|---|---|
| Identification code | catalyst P4 |
| Empirical formula | C ₂₅ H ₂₉ FIN ₂ PS |
| Formula weight | 566.43 |
| Temperature/K | 295.4(6) |
| Crystal system | monoclinic |
| Space group | P2 ₁ |
| a/Å | 9.4752(4) |
| b/Å | 11.6241(3) |
| c/Å | 12.9271(6) |
| α/° | 90 |
| β/° | 111.550(5) |
| γ/° | 90 |
| Volume/Å ³ | 1324.26(10) |
| Z | 2 |
| ρ _{calc} /cm ³ | 1.421 |
| μ/mm ⁻¹ | 10.974 |
| F(000) | 572.0 |
| Crystal size/mm ³ | 0.7 × 0.6 × 0.2 |
| Radiation | CuKα (λ = 1.54184) |
| 2θ range for data collection/° | 7.352 to 131.016 |
| Index ranges | -11 ≤ h ≤ 11, -13 ≤ k ≤ 9, -12 ≤ l ≤ 15 |
| Reflections collected | 6569 |
| Independent reflections | 3343 [R _{int} = 0.1119, R _{sigma} = 0.0835] |
| Data/restraints/parameters | 3343/1/283 |
| Goodness-of-fit on F ² | 1.131 |
| Final R indexes [I ≥ 2σ (I)] | R ₁ = 0.0957, wR ₂ = 0.2193 |
| Final R indexes [all data] | R ₁ = 0.0978, wR ₂ = 0.2230 |
| Largest diff. peak/hole / e Å ⁻³ | 2.88/-2.26 |
| Flack parameter | 0.009(16) |

7. Gram-scale preparations and transformations

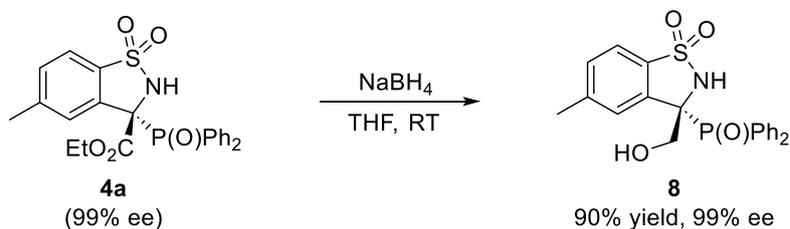
A. Procedure for the gram-scale synthesis of **4a** and **5a**



Gram-scale synthesis of 4a: To a flame-dried round bottle flask with a magnetic stirring bar were added the cyclic imine **1a** (4.0 mmol, 1012 mg), Phosphine Oxide **2a** (4.8 mmol, 969.6 mg), and phosphonium salt **P4** (0.4 mmol, 476 mg). followed by the addition of dry DCE (60.0 ml). The reaction mixture was stirred at 0 °C for 48 h. The solvent was removed under reduced pressure, and the residue was purified by column chromatography on silica gel (CH₂Cl₂/ethyl acetate = 3:1) to afford **4a** (93% yield) as a white solid.

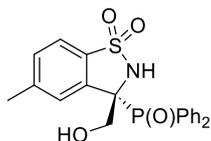
Gram-scale synthesis of 5a: To a flame-dried round bottle flask with a magnetic stirring bar were added the cyclic imine **1a** (4.0 mmol, 1012 mg), Phosphine Oxide **3a** (4.8 mmol, 1123.2 mg), and phosphonium salt **P6** (0.4 mmol, 226.4 mg). followed by the addition of toluene (40.0 mL). The reaction mixture was stirred at room temperature for 48 h. The solvent was removed under reduced pressure, and the residue was purified by column chromatography on silica gel (CH₂Cl₂/ethyl acetate = 3:1) to afford **5a** (92% yield) as a white solid.

B. Preparation of compound **8**^[8]



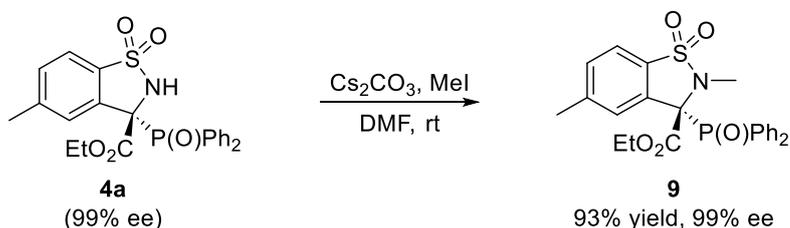
General procedure: To the solution of **4a** (0.2 mmol, 91.0 mg) in THF (5 mL) was added NaBH₄ (0.6 mmol, 22.8 mg) in three portions at 0 °C. The resulting mixture was stirred for 0.5 hour at room temperature. Then, the reaction was quenched with saturated aqueous chloride ammonium (2 mL). The organic solvent was removed in vacuo and the aqueous layer was extracted with CH₂Cl₂ (5 mL x 3). The combined organic layer was dried over anhydrous sodium sulfate, filtered and evaporated under vacuum. The residue was purified by flash chromatography over silica gel (CH₂Cl₂/ethyl acetate = 3:1) to afford the desired product **8**.

(S)-3-(diphenylphosphoryl)-3-(hydroxymethyl)-5-methyl-2,3-dihydrobenzo[d]isothiazole 1,1-dioxide (8)



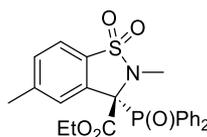
A white solide; m.p. = 142 - 144 °C; 74.3 mg, 90% yield, 99% ee; ^1H NMR (400 MHz, CDCl_3) δ 7.66 - 7.47 (m, 5H), 7.47 - 7.30(m 3H), 7.16 - 6.95 (m, 5H), 5.86 (s, 1H), 5.32 (s, 1H), 3.92 - 3.54 (m, 1H), 2.21 (s, 1H), 2.13 (s, 3H); ^{13}C NMR (100 MHz, $\text{CDCl}_3:\text{CD}_3\text{OD} = 1:1$) δ 143.81 (d, $J = 2.1$ Hz), 143.65, 137.72, 132.88, 132.69 (d, $J = 2.6$ Hz), 132.55 (d, $J = 3.0$ Hz), 132.40 (dd, $J = 12.9, 2.7$ Hz), 132.01 (d, $J = 3.7$ Hz), 131.58 (dd, $J = 56.4, 9.0$ Hz), 130.39 (d, $J = 1.9$ Hz), 130.06 (d, $J = 11.8$ Hz), 129.90, 129.53, 128.57 (d, $J = 13.0$ Hz), 128.12 (dd, $J = 66.8, 12.0$ Hz), 126.18, 125.42 (d, $J = 2.5$ Hz), 125.18, 124.64, 120.42 (d, $J = 1.6$ Hz), 120.10, 64.03, 58.53, 56.23 (d, $J = 84.4$ Hz); ^{31}P NMR (162 MHz, $\text{CDCl}_3:\text{CD}_3\text{OD} = 1:1$) δ 35.78; HRMS (ESI) m/z calcd for $\text{C}_{21}\text{H}_{20}\text{NO}_4\text{PS}$ $[\text{M}+\text{Na}]^+ = 436.0748$, found = 436.0740.

C. Preparation of compound 9^[9]



General procedure: To the solution of **4a** (0.2 mmol, 91.0 mg) in DMF (5 mL) was added CH_3I (0.6 mmol), The resulting mixture was stirred for 0.5 hour at room temperature. Then, the reaction was added Cs_2CO_3 (0.2 mmol). The resulting mixture was stirred for 4 hours at room temperature. The reaction was quenched with saturated aqueous NaCl (2 mL). The mixture layer was extracted with CH_2Cl_2 (10 mL x 3). The combined organic layer was dried over anhydrous sodium sulfate, filtered and evaporated under vacuum. The residue was purified by flash chromatography over silica gel ($\text{CH}_2\text{Cl}_2/\text{ethyl acetate} = 3:1$) to afford the desired product **9**.

(S)-ethyl 3-(diphenylphosphoryl)-2,5-dimethyl-2,3-dihydrobenzo[d]isothiazole-3-carboxylate 1,1-dioxide (9)



A white solide, m.p. = 184 - 186 °C; 87.2 mg, 93% yield, 99% ee; ^1H NMR (400 MHz, CDCl_3) δ 7.88 - 7.77 (m, 2H), 7.63 (d, J = 8.0 Hz, 1H), 7.59 - 7.48 (m, 4H), 7.46 - 7.37 (m, 4H), 7.33 (d, J = 8.0 Hz, 1H), 6.88 (s, 1H), 4.42 - 4.17 (m, 2H), 2.90 (s, 3H), 2.21 (s, 3H), 1.27 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 166.15 (d, J = 6.8 Hz), 143.54 (d, J = 1.7 Hz), 133.40 (d, J = 8.7 Hz), 132.92 (d, J = 2.7 Hz), 132.78 (d, J = 2.9 Hz), 131.57 (d, J = 1.6 Hz), 130.88 (d, J = 3.4 Hz), 130.56 (d, J = 2.6 Hz), 129.40 (d, J = 16.4 Hz), 128.65 (d, J = 11.9 Hz), 128.41 (d, J = 19.6 Hz), 127.96 (d, J = 12.0 Hz), 127.21 (d, J = 3.0 Hz), 121.24 (d, J = 0.8 Hz), 73.93 (d, J = 73.5 Hz), 63.38, 27.18, 21.78, 14.08; ^{31}P NMR (162 MHz, CDCl_3) δ 31.85; HRMS (ESI) m/z calcd for $\text{C}_{24}\text{H}_{24}\text{NO}_5\text{PS}$ $[\text{M}+\text{Na}]^+$ = 492.1010, found = 492.1002.

8. Mechanistic studies

8.1. Model reaction promoted by different catalysts

According to previous mechanistic results, we prepared the methylated phosphonium salt catalyst **P4-1**. As shown, employment of N-methylated catalyst **P4-1** led to the formation of the desired product **4a** in low yield with sharply decreased enantioselectivities (entry 2). Additionally, when methanol was used as solvent (entry 3), erosion of enantioselectivity was observed due to that the hydrogen-bond network was disrupted under polar system, Besides, when changed the optimal phosphonium salt catalyst P4 to its corresponding trivalent. phosphine (**P4-0**) under the standard conditions for the reaction, the yield and selectivity of the reaction decreased remarkably, giving nearly racemic product only with 35% isolated yield (entry 4). All these results indicated the importance of both hydrogen-bonding and ion-pair interactions in this phase-transfer system. Futhermore, we have also tested different negative ions with a neutral H-bonding trivalent phosphine catalyst (P4-0) under the standard conditions for the reaction. Not surprisingly, the yield and selectivity of the reaction decreased remarkably, giving nearly racemic product with moderate isolated

yield (entries 5-7), which indicated the synergistic effect between the hydrogen-bonding and ion pair of the bifunctional phosphonium salts. Additionally, the anions effect of the bifunctional phosphonium salt were investigated, the other types of bifunctional phosphonium salt catalysts with BF_4^- (**P4-2**)、 PF_6^- (**P4-3**) and NO_2^- (**P4-4**) were synthesized and applied in the model reaction under standard conditions, and the results shown that the enantioselectivities of this asymmetric P-addition reaction were dropped sharply (Table S7, entries 8-10). We speculated that mainly due to the steric hindrance and the tightness of ion pairs moiety in the catalyst.

Table S7. Asymmetric P-nucleophile addition to cyclic ketimine **1a with **2a** promoted by different phosphonium salts^a**

Catalysts examined

P4

P4-0

P4-1

P4-2: X = BF_4
P4-3: X = PF_6
P4-4: X = NO_2

| entry | cat. | sol. | T (°C) | t (h) | yield (%) ^b | ee (%) ^c |
|----------------|-------------|------|--------|-------|------------------------|---------------------|
| 1 | P4 | DCE | rt | 2 | 95 | >99 |
| 2 | P4-1 | DCE | rt | 2 | 94 | 8 |
| 3 | P4 | MeOH | rt | 0.5 | 92 | 0 |
| 4 | P4-0 | DCE | rt | 24 | 35 | <5 |
| 5 ^d | P4-0 | DCE | rt | 24 | 83 | <5 |
| 6 ^e | P4-0 | DCE | rt | 24 | 72 | <5 |
| 7 ^f | P4-0 | DCE | rt | 24 | 67 | 0 |
| 8 | P4-2 | DCE | rt | 2 | 80 | <5 |
| 9 | P4-3 | DCE | rt | 2 | 85 | <5 |
| 10 | P4-4 | DCE | rt | 2 | 90 | 12 |

^aReactions were performed with **1a** (0.1 mmol), **2a** (0.12 mmol), catalyst (0.01 mmol) in solvent (1.0

mL). ^bIsolated yield. ^cDetermined by HPLC analysis on a chiral stationary phase. ^dThe NaI (0.1 mmol) was used. ^eThe NaBr (0.1 mmol) was used. ^fThe NaCl (0.1 mmol) was used.

8.2. ¹H NMR titration of **1a** or **2a** with catalyst **P4**

¹H NMR titration experiments were conducted on a Bruker spectrometer by recording the changes of ¹H NMR spectra of the catalyst **P4** after the corresponding targets addition. In a typical experiment, the catalyst **P4** (28.3 mg, 0.05 mmol) was dissolved with CD₃Cl (0.5 mL) in nuclear magnetic tube. Then, the ¹H NMR spectrum of catalyst **P4** was recorded. After that, the corresponding equivalent of substrate (**1a** or **2a**) was stepwise added to the NMR-tube and chemical shifts of the protons on the catalyst **P4** were recorded after each addition.

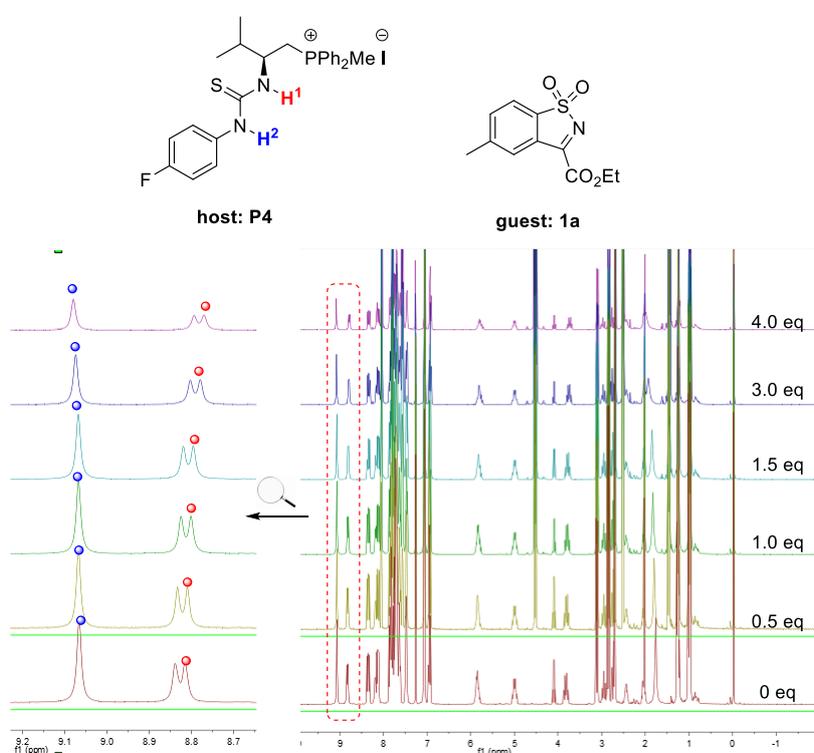


Figure S10. ¹H NMR spectrum of **P4** with **1a**

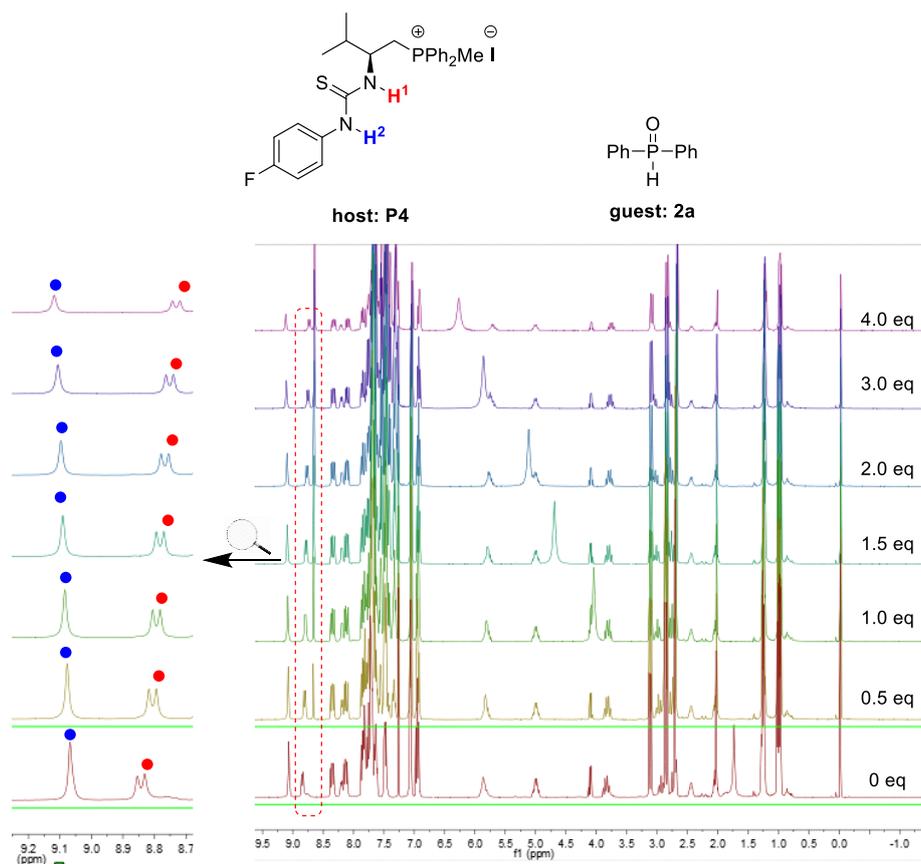


Figure S11. ^1H NMR spectrum of **P4** with **2a**

^1H NMR titration was conducted in chloroform-d, which was dried with potassium carbonate and degassed. The total concentration of the host (**P4**) and the guest (**1a/2a**) was 0.04 M. The proportion of the concentration of the host vs the total concentration varied from 0.2 equivalent to 0.8. The chemical shift of N-H¹ of **P4** were recorded. 1:1 binding pattern could also be observed in the Job plot.

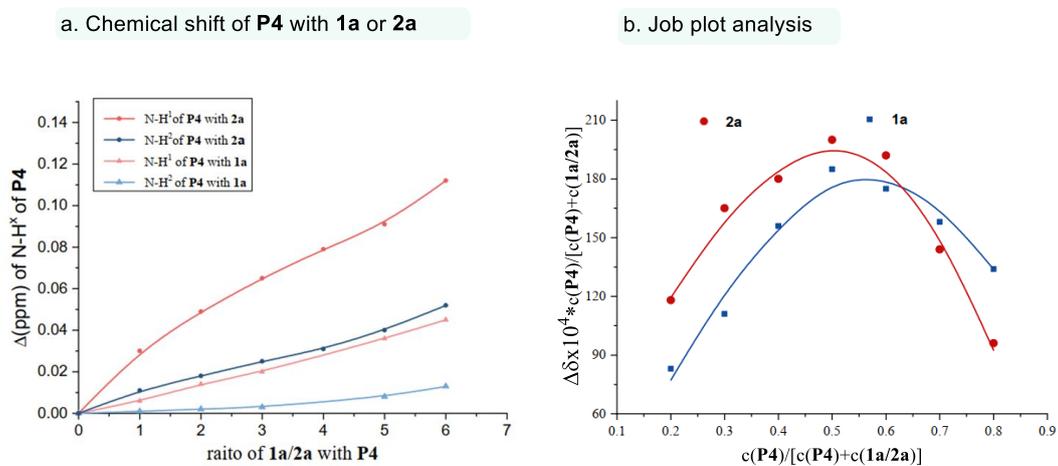


Figure S12. ^1H NMR titration of substrate **1a** or **2a** with **P4**

B. Proposed reaction catalytic cycle

It was plausible to propose a reaction catalytic cycle presented as Figure S13. At first the diphenylphosphine oxide **2a** was deprotonated by iodide ion and converted into diphenylphosphinite anion, which was stabilized and simultaneously activated by the catalyst **P4** via H-bonding and ion-pair interaction. Subsequently, the nucleophilic addition of the anionic **P**-species towards cyclic imine substrate **1a** preferentially occurred via the transition state **TS-1**, which quickly was protonated experiencing the transition state **TS-2** to produce product (*R*)-**5a** and release catalyst **P4** that would enter the next reaction cycle.

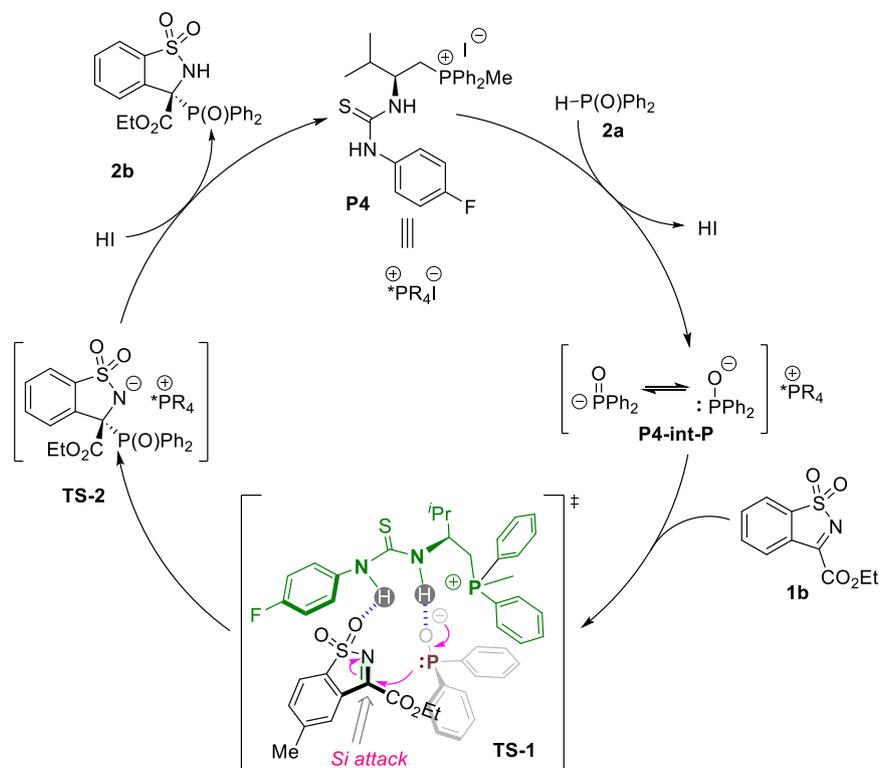


Figure S13. Proposed reaction mechanism

C. Structure characteristics of bifunctional phosphonium salt catalyst P4 by-DFT calculation

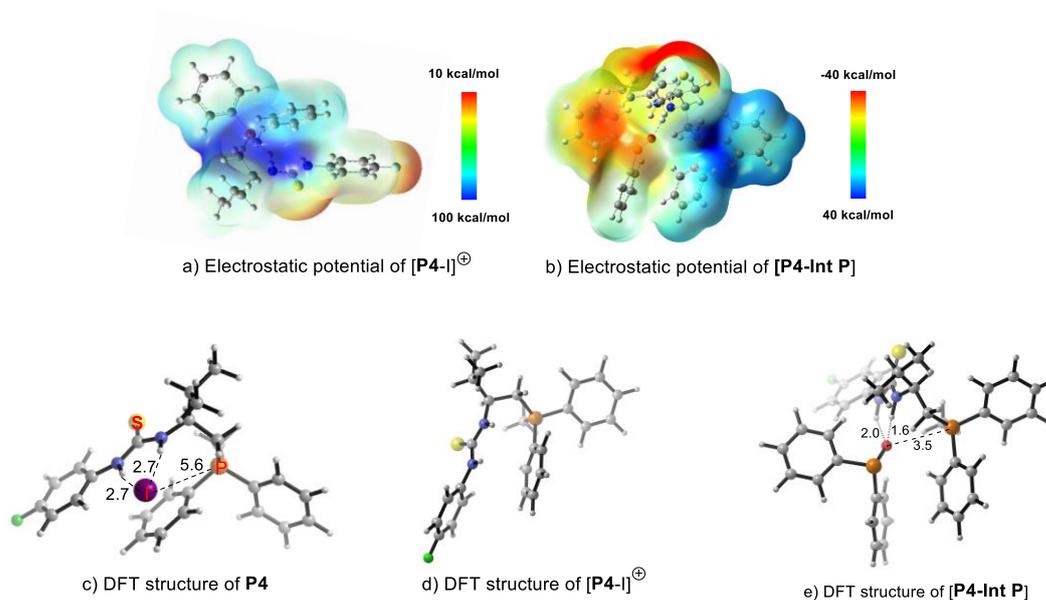


Figure S14. The electrostatic potential and DFT structure of catalysts, catalyst cations, and catalyst complexes. (The optimized structures were calculated at the M062X-D3/6-311+G(d,p)-SMD(CH₂Cl₂)/M062X-D3/6-31G(d,p)-SMD(CH₂Cl₂) level of theory).

D. Cartesian coordinates of DFT-computed structures

P4

| | | | |
|---|-------------|-------------|-------------|
| I | 0.88830600 | 2.79315600 | -1.90715500 |
| P | -2.35508800 | -1.02153100 | 0.62610700 |
| S | 1.02321100 | -0.62164700 | 3.03876100 |
| N | 0.00763600 | 1.12046800 | 1.27936200 |
| C | -1.26298100 | 1.21988000 | 1.96772600 |
| C | -3.78507200 | -1.40015600 | -0.41938400 |
| C | -0.87182300 | -1.49290100 | -0.28645000 |
| C | 3.95910400 | -0.09725800 | -0.64318100 |
| N | 2.02838800 | 0.32201800 | 0.68458100 |

| | | | |
|---|-------------|-------------|-------------|
| C | 3.24196500 | -0.36773000 | 0.54082400 |
| C | -2.39138400 | 0.75208100 | 1.03057400 |
| C | -4.83697600 | -0.49017500 | -0.60323600 |
| C | 4.97102000 | -1.95354700 | 1.17268200 |
| C | 0.53683800 | -1.19450800 | -2.23113000 |
| C | 3.76563800 | -1.30237600 | 1.44959400 |
| C | -0.59664500 | -0.84198500 | -1.50013500 |
| C | -1.50254800 | 2.64235800 | 2.53268800 |
| C | -1.69895800 | 3.70554600 | 1.45018200 |
| C | -0.01569300 | -2.49867800 | 0.18763800 |
| C | 1.38919400 | -2.19961700 | -1.76198700 |
| C | 1.03848900 | 0.30713300 | 1.61829600 |
| C | -2.65231000 | 2.65233100 | 3.54300800 |
| C | 1.11209200 | -2.85013800 | -0.55742100 |
| C | -5.92656300 | -0.84559600 | -1.40299300 |
| C | -2.46800700 | -1.97967000 | 2.15019500 |
| C | -3.82773000 | -2.66110600 | -1.03964700 |
| C | 5.16048200 | -0.74369500 | -0.91955900 |
| C | -4.92167900 | -3.00873700 | -1.83164500 |
| C | -5.97097300 | -2.10099200 | -2.01461200 |
| F | 6.81439800 | -2.30554800 | -0.26849300 |
| C | 5.65235100 | -1.67114900 | -0.00468300 |
| H | 0.09788700 | 1.65533200 | 0.40886800 |

| | | | |
|---|-------------|-------------|-------------|
| H | -1.19550600 | 0.54370000 | 2.83002700 |
| H | 3.55869700 | 0.62570400 | -1.35776300 |
| H | 1.83806200 | 0.94132900 | -0.11010700 |
| H | -2.37344400 | 1.30096600 | 0.07651500 |
| H | -3.37187500 | 0.92490400 | 1.49840500 |
| H | -4.81766500 | 0.49438800 | -0.13513500 |
| H | 5.38341200 | -2.68257200 | 1.87353100 |
| H | 0.76084400 | -0.67041700 | -3.16187000 |
| H | 3.22244400 | -1.52009600 | 2.36512200 |
| H | -1.25467800 | -0.05280200 | -1.86933500 |
| H | -0.57230800 | 2.87562800 | 3.07893800 |
| H | -0.87892300 | 3.70766300 | 0.71565900 |
| H | -1.74081100 | 4.70891400 | 1.90263200 |
| H | -2.64337100 | 3.55516800 | 0.90138000 |
| H | -0.21131300 | -3.00020800 | 1.13476900 |
| H | 2.28090400 | -2.46699300 | -2.33283100 |
| H | -3.62570700 | 2.46307600 | 3.05998000 |
| H | -2.72328900 | 3.63445200 | 4.03648700 |
| H | -2.50985100 | 1.89172000 | 4.32796200 |
| H | 1.78509100 | -3.62435600 | -0.18397400 |
| H | -6.74199300 | -0.13378900 | -1.54879000 |
| H | -3.35625200 | -1.63714700 | 2.70081700 |
| H | -1.56120100 | -1.81406400 | 2.75205100 |

| | | | |
|---|-------------|-------------|-------------|
| H | -2.57649800 | -3.04617000 | 1.90911600 |
| H | -3.00446800 | -3.36773700 | -0.91062600 |
| H | 5.71471100 | -0.53632800 | -1.83701000 |
| H | -4.95208600 | -3.98866800 | -2.31303300 |
| H | -6.82484100 | -2.37291200 | -2.63959300 |

[P4-I]⁺

| | | | |
|---|-------------|-------------|-------------|
| P | 2.16630900 | -0.17781400 | -0.56429500 |
| S | -1.64578000 | 1.70023800 | -1.76452500 |
| N | -0.22208600 | 1.65465000 | 0.50630800 |
| C | 0.93633500 | 2.31659000 | -0.07213300 |
| C | 3.73817000 | -1.00375900 | -0.23073700 |
| C | 0.84825900 | -1.19646700 | 0.12607800 |
| C | -4.50194500 | 0.27864100 | 1.27856100 |
| N | -2.20568600 | 0.58660000 | 0.63073500 |
| C | -3.50327200 | 0.10655600 | 0.31812500 |
| C | 2.19759500 | 1.47749800 | 0.19412800 |
| C | 4.87504000 | -0.28490500 | 0.15039400 |
| C | -5.05938700 | -1.08911300 | -1.08515300 |
| C | -0.20426900 | -2.16986100 | 2.06527300 |
| C | -3.78046900 | -0.59247000 | -0.85928300 |
| C | 0.80444900 | -1.38909700 | 1.51306700 |
| C | 1.07048300 | 3.76482600 | 0.44435500 |

| | | | |
|---|-------------|-------------|-------------|
| C | 1.27987900 | 3.84550400 | 1.95555000 |
| C | -0.10340600 | -1.79341400 | -0.70360400 |
| C | -1.15515800 | -2.77066300 | 1.23780700 |
| C | -1.35073800 | 1.29811200 | -0.14542600 |
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| C | 1.93606400 | -0.00634400 | -2.34310300 |
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| F | -7.26922200 | -1.38602800 | -0.33587500 |
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| H | 2.38101600 | 1.33581200 | 1.26612700 |
| H | 3.06927200 | 1.98753300 | -0.22960900 |
| H | 4.83856800 | 0.79064800 | 0.28707600 |
| H | -5.30234600 | -1.63107000 | -1.99251000 |
| H | -0.24223200 | -2.32074900 | 3.13909400 |

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| H | 1.55620900 | -0.93846800 | 2.15741900 |
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| H | 0.48861200 | 3.33020300 | 2.50897500 |
| H | 1.27506900 | 4.89082300 | 2.27691000 |
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| H | -6.56827200 | -0.10358300 | 1.80094900 |
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[P4-Int-P]

| | | | |
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| P | -2.67541600 | -0.31084900 | -0.30568300 |
|---|-------------|-------------|-------------|

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| N | 0.01416000 | -1.55543000 | 0.98676500 |
| C | -1.25404000 | -1.79916200 | 1.64415200 |
| C | -4.23863900 | -1.18448600 | -0.55114700 |
| C | -2.96278100 | 1.44626100 | -0.58964300 |
| C | 4.05801800 | -1.76733900 | -0.97986100 |
| N | 1.73444900 | -1.88984800 | -0.40470300 |
| C | 2.84888900 | -2.46946900 | -1.04784700 |
| C | -2.10486200 | -0.53687900 | 1.40934900 |
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| C | 3.91154100 | -4.13086400 | -2.44436800 |
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| C | -4.13732700 | 2.04112300 | -0.11390700 |
| C | -1.07723400 | -2.10589600 | 3.14537700 |
| C | -0.53695500 | -0.90684500 | 3.92175000 |
| C | -1.98481300 | 2.21487500 | -1.22701900 |
| C | -3.35703100 | 4.17606500 | -0.92489200 |
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| C | -5.86282400 | -2.84832100 | 0.08775400 |
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| C | 5.19033800 | -2.23623900 | -1.63497800 |
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| F | 6.19245700 | -3.89070900 | -2.97765800 |
| C | 5.09655800 | -3.41987000 | -2.34869900 |
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| H | 1.74255800 | -0.86441200 | -0.36287800 |
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| H | -3.00048400 | -0.51063300 | 2.03859900 |
| H | -4.10302200 | -2.39846700 | 1.23651100 |
| H | 3.87911700 | -5.04356200 | -3.02938000 |
| H | -5.23800000 | 3.87235800 | 0.08390500 |
| H | 1.83741000 | -4.17505100 | -1.88218100 |
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| H | -0.26205600 | -1.20396900 | 4.93807100 |
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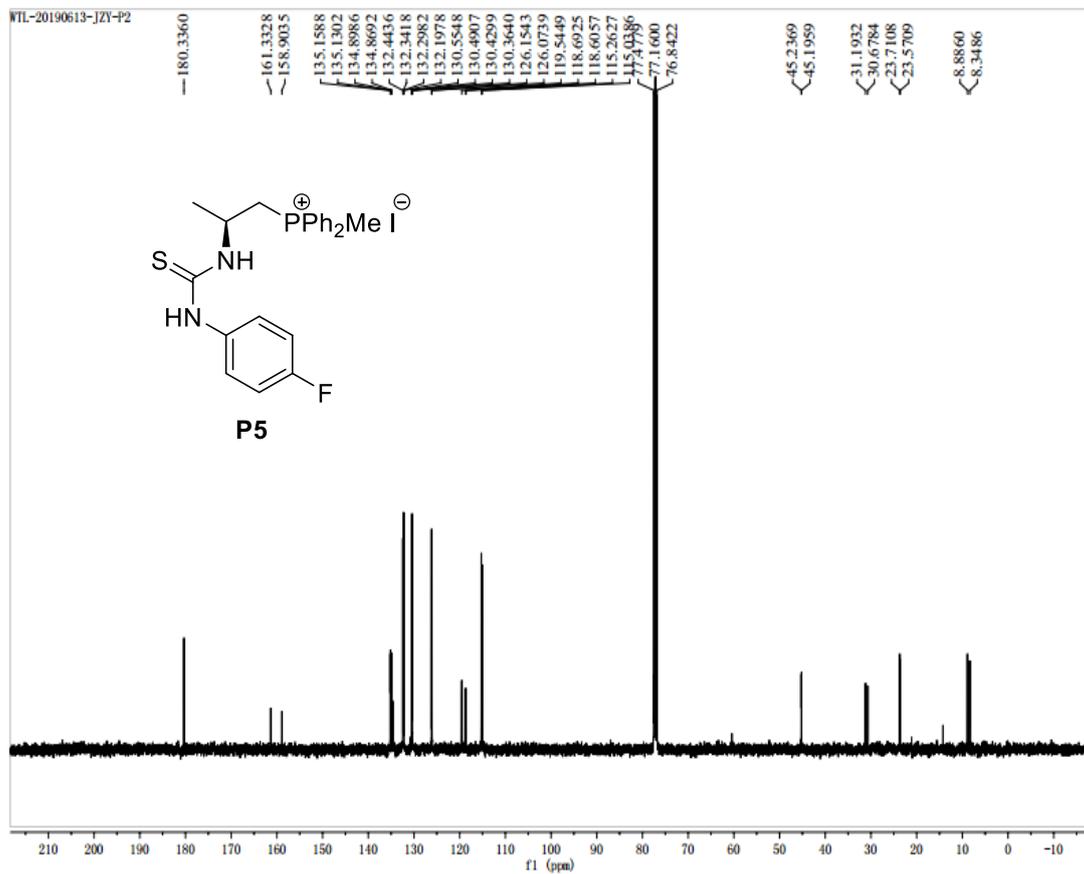
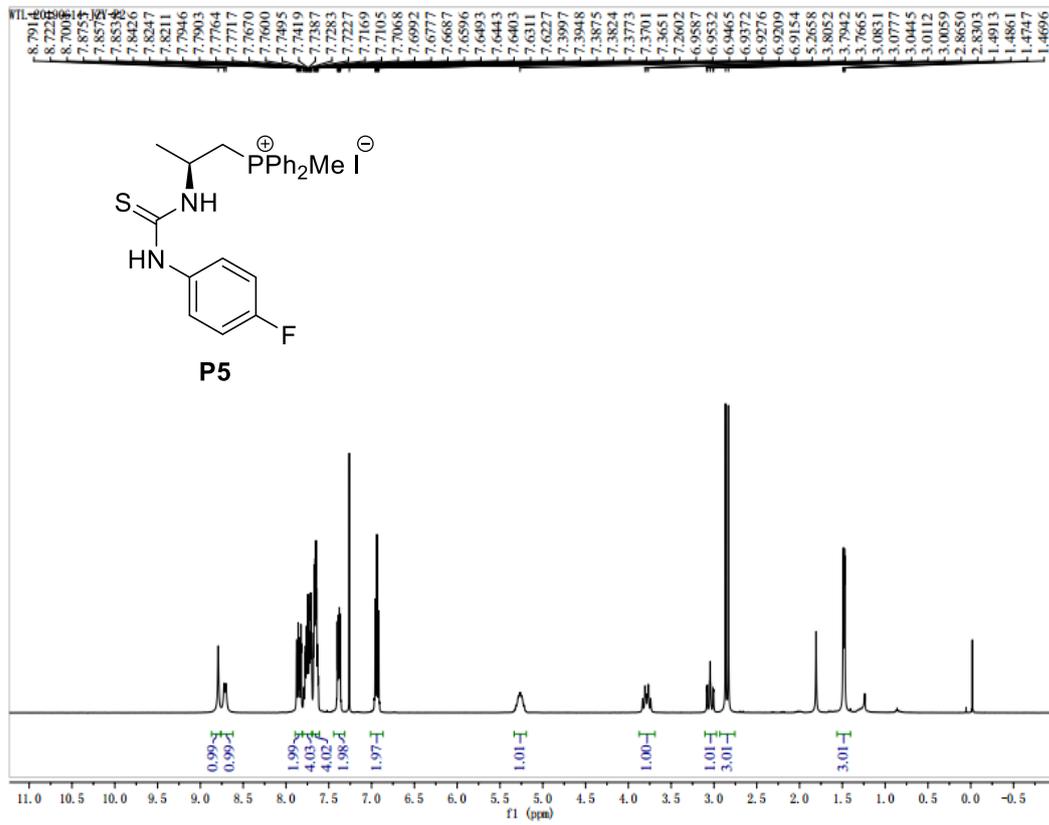
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| H | -6.74935800 | -1.34646300 | -2.83210600 |
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| O | 0.68928400 | 0.76417100 | 0.03506000 |
| C | 1.09275700 | 3.41193100 | -0.36938000 |
| C | 0.92358500 | 4.76328500 | -0.05149800 |
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| H | 0.64413500 | 5.04831000 | 0.96142900 |
| C | 1.55565400 | 4.03918100 | -2.66554500 |
| H | 1.51203100 | 2.00434900 | -1.92683000 |
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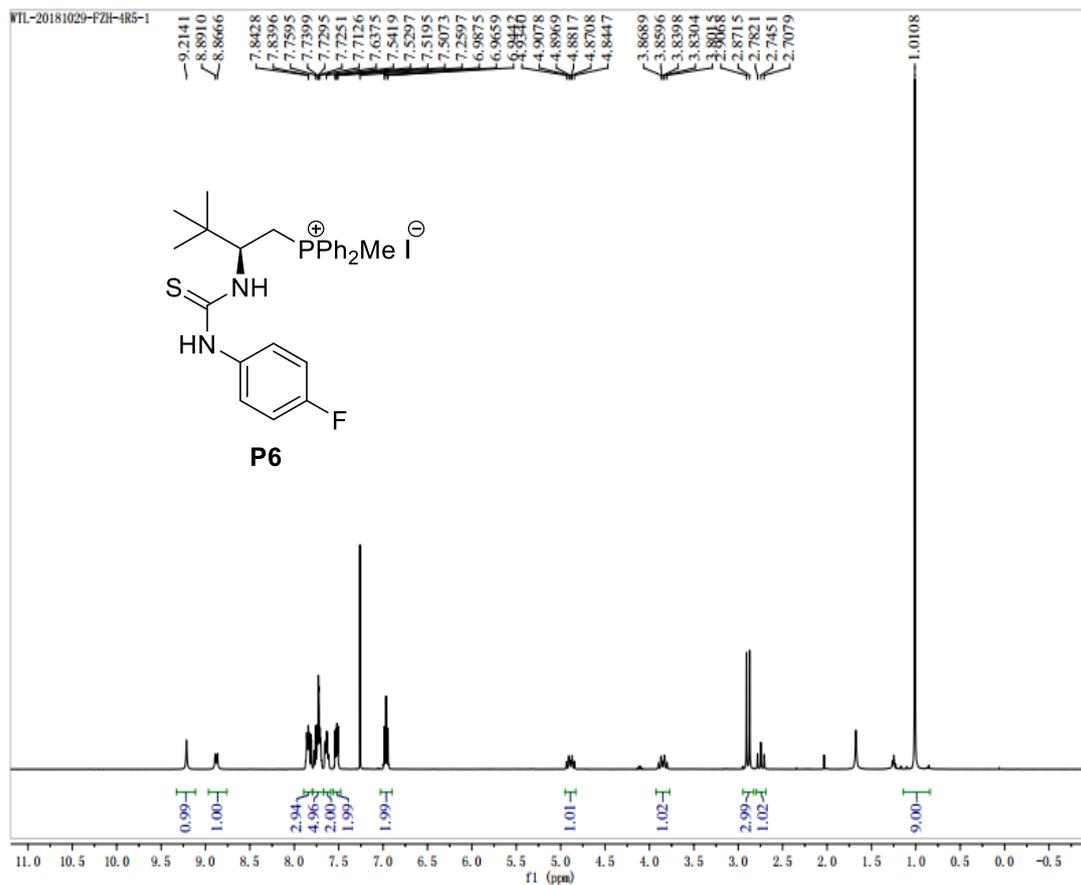
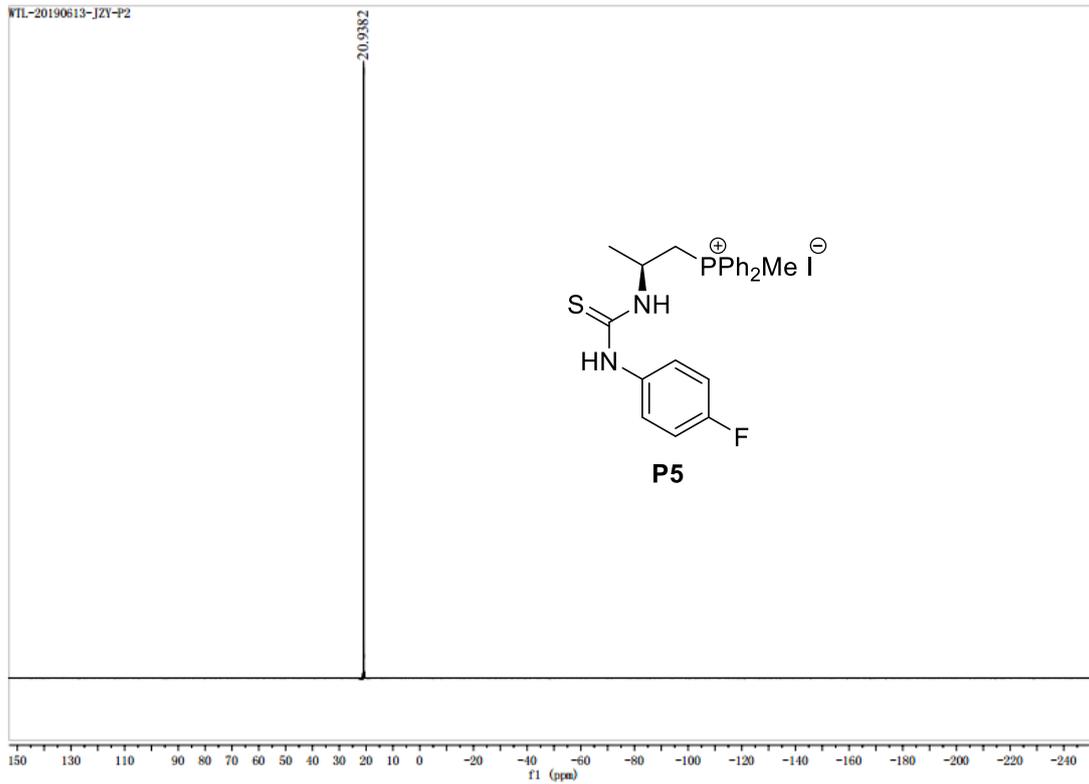
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| H | 2.51200200 | -0.14045100 | 1.59077800 |
| C | 4.48459800 | 3.02747600 | 2.59887600 |
| H | 2.82752300 | 4.13816900 | 1.80808300 |
| C | 5.03227200 | 1.76673400 | 2.83807200 |
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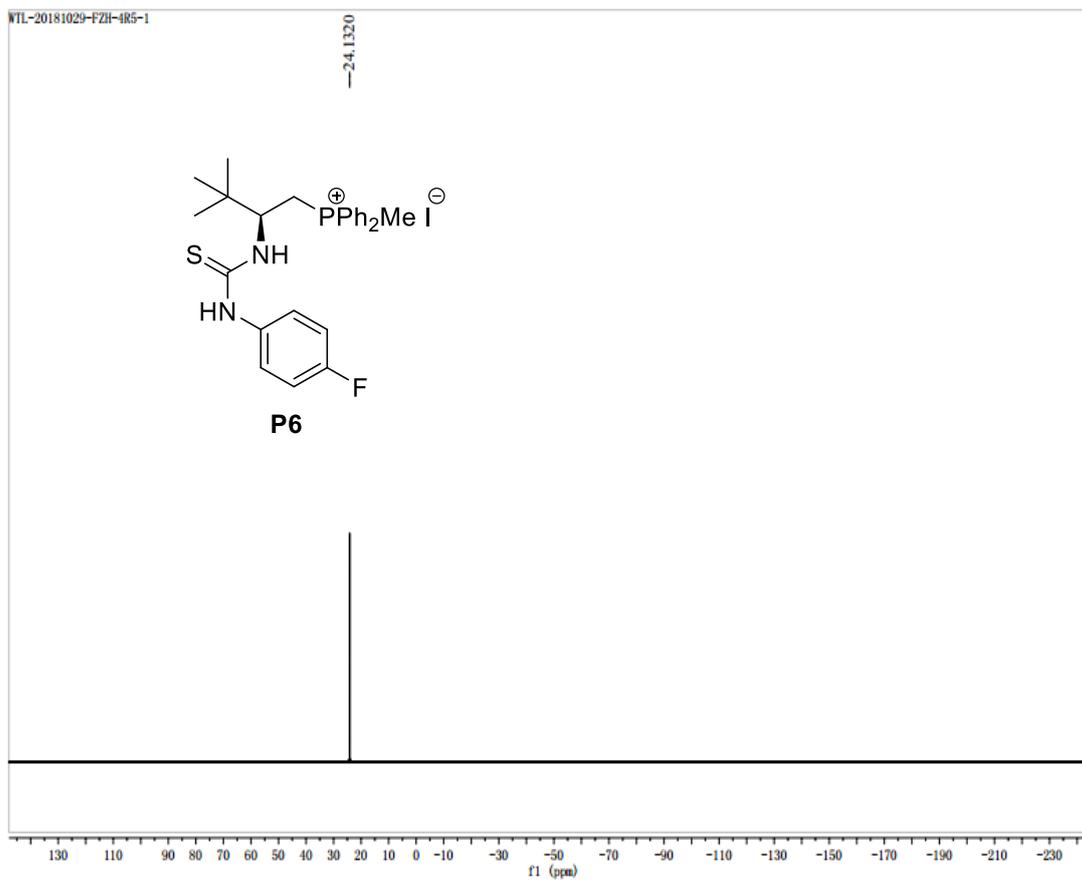
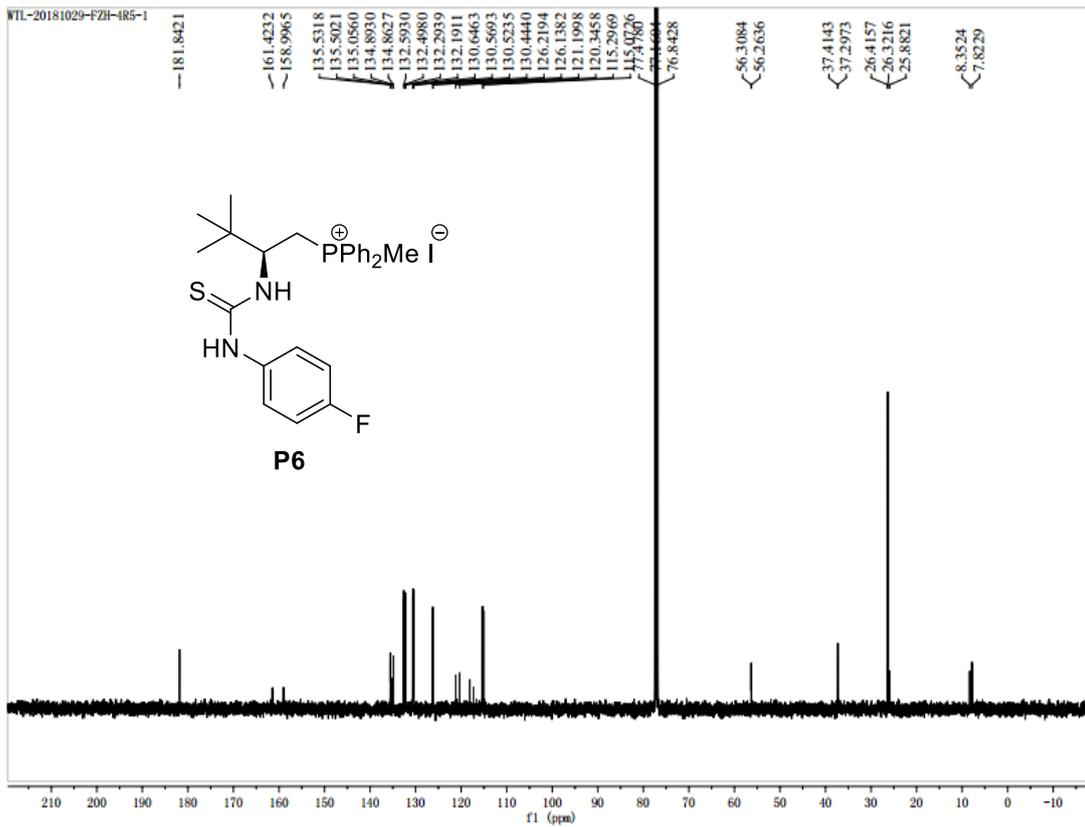
9. References

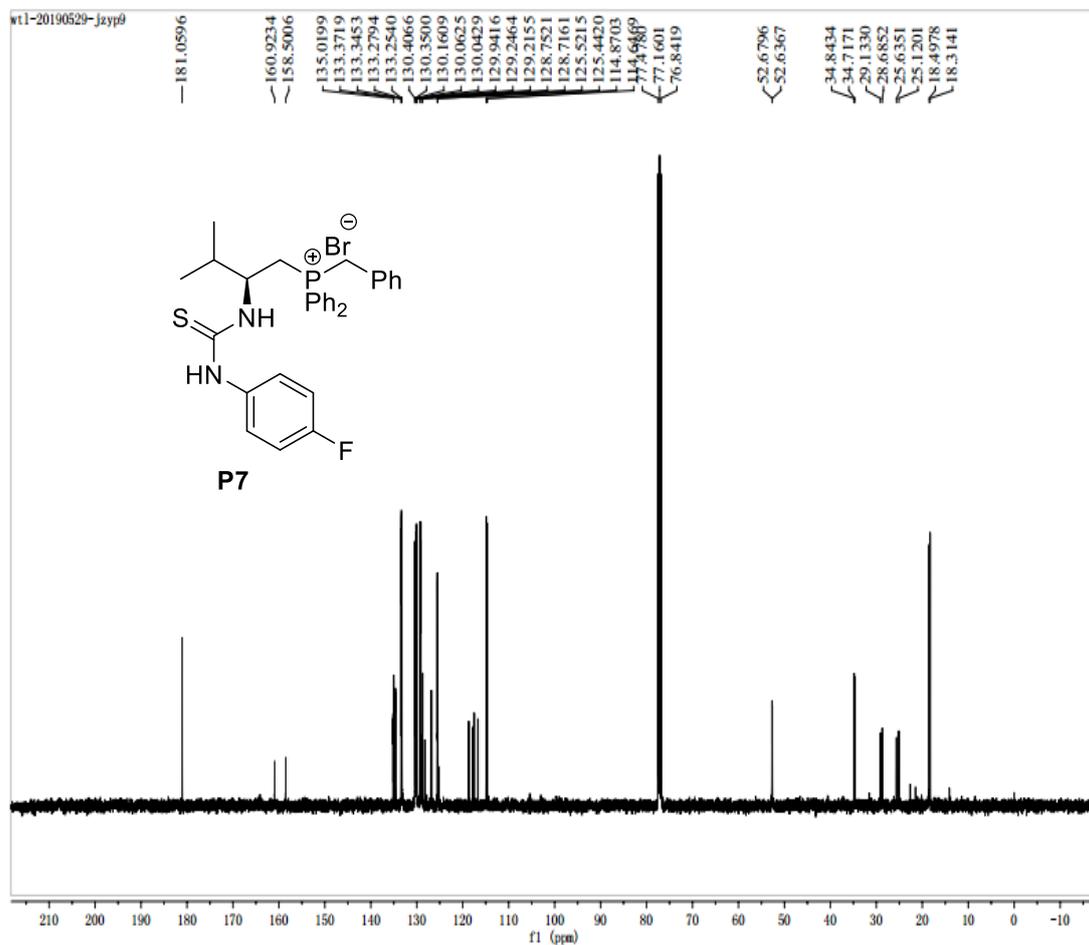
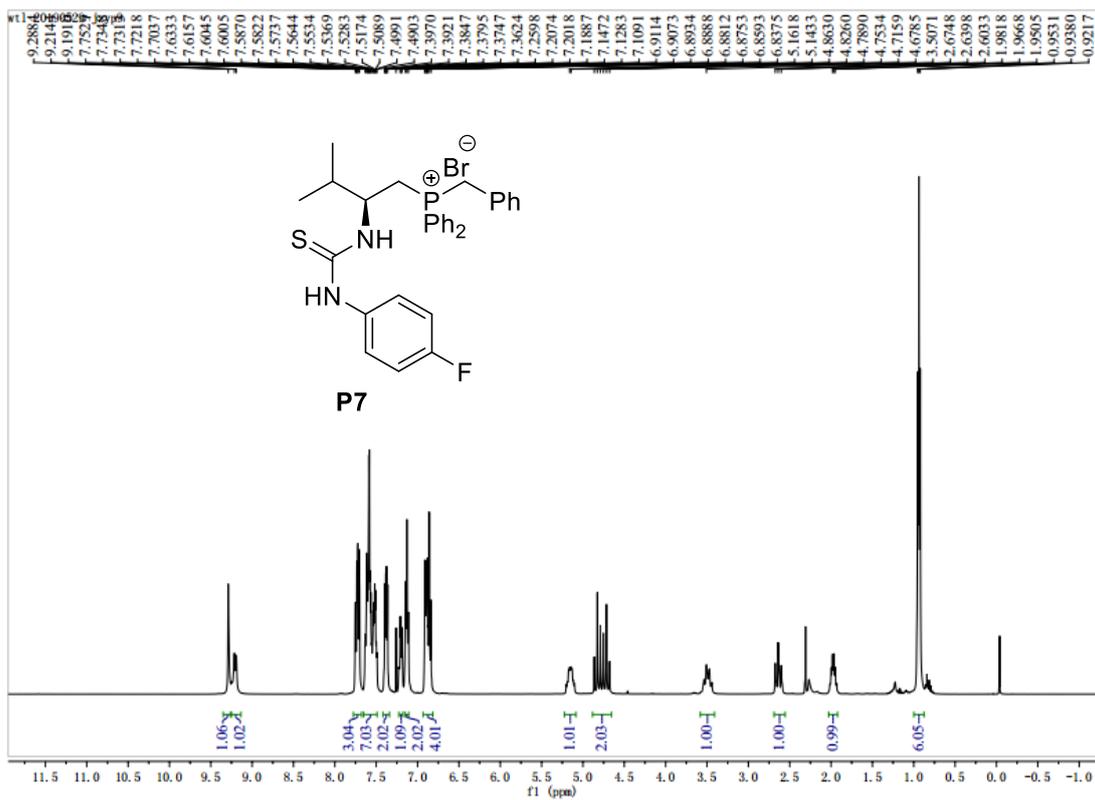
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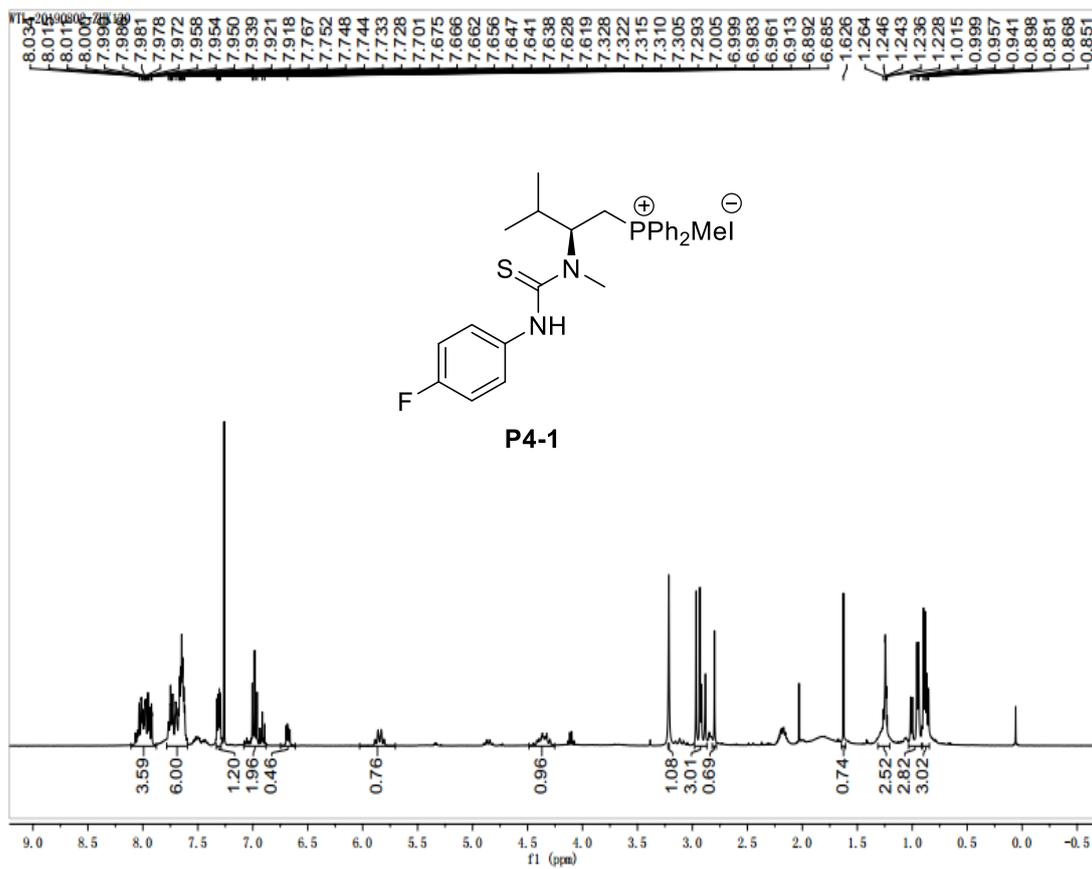
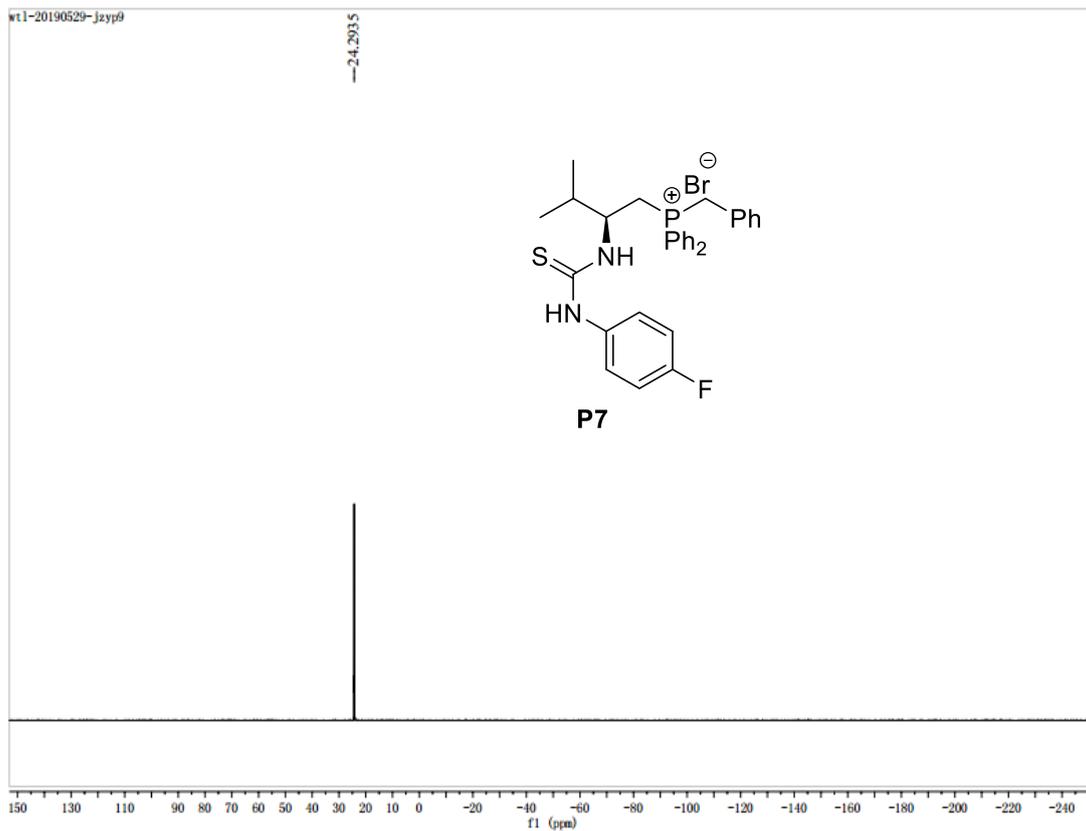
10. NMR spectra

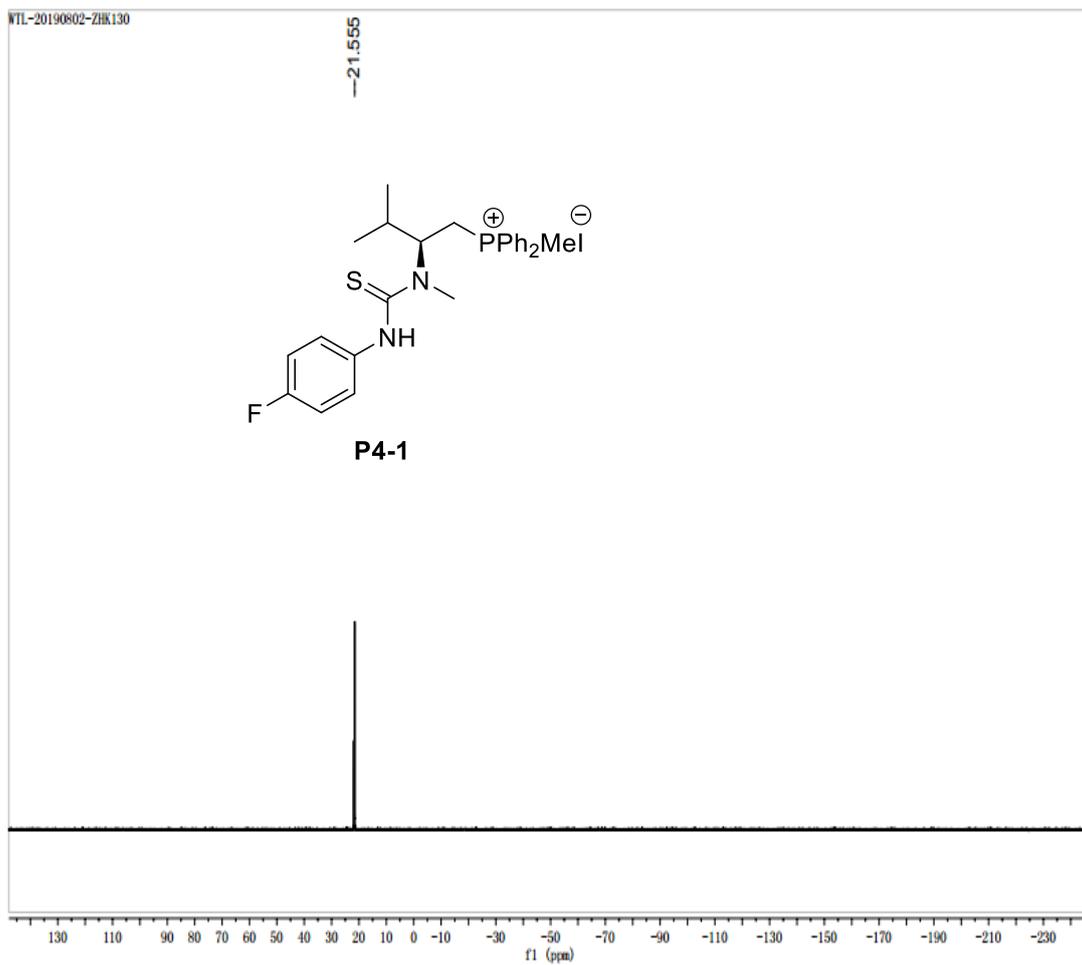
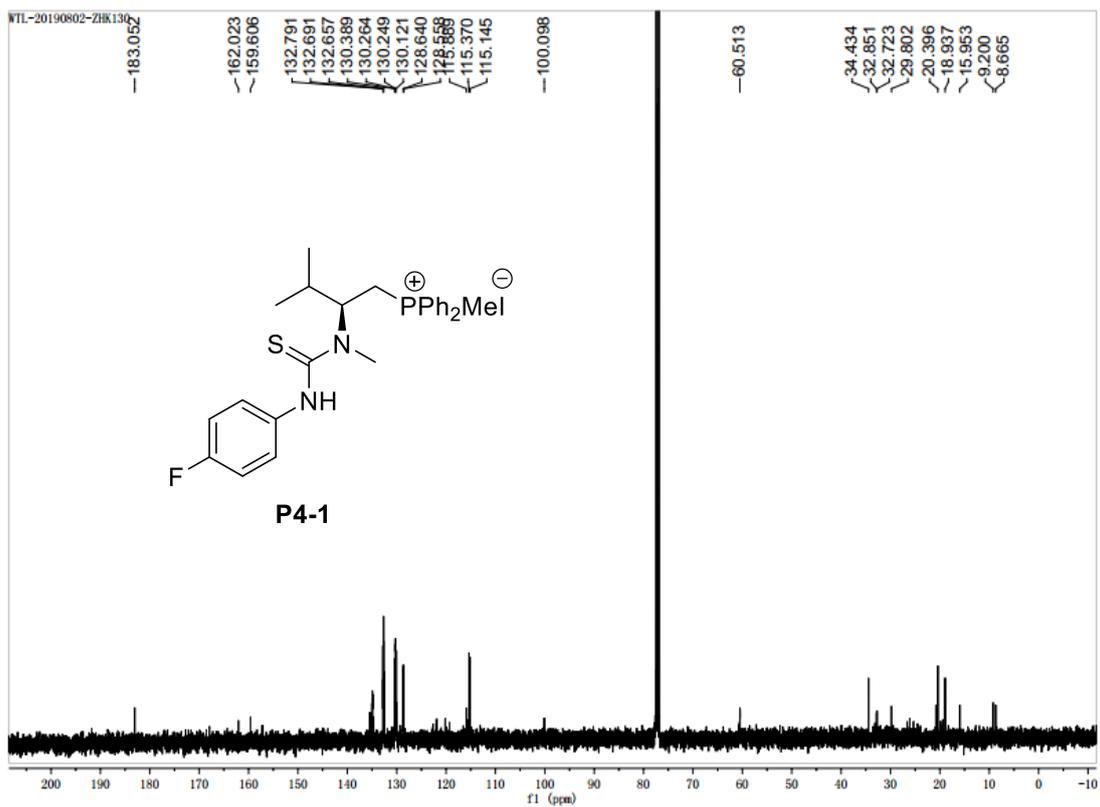


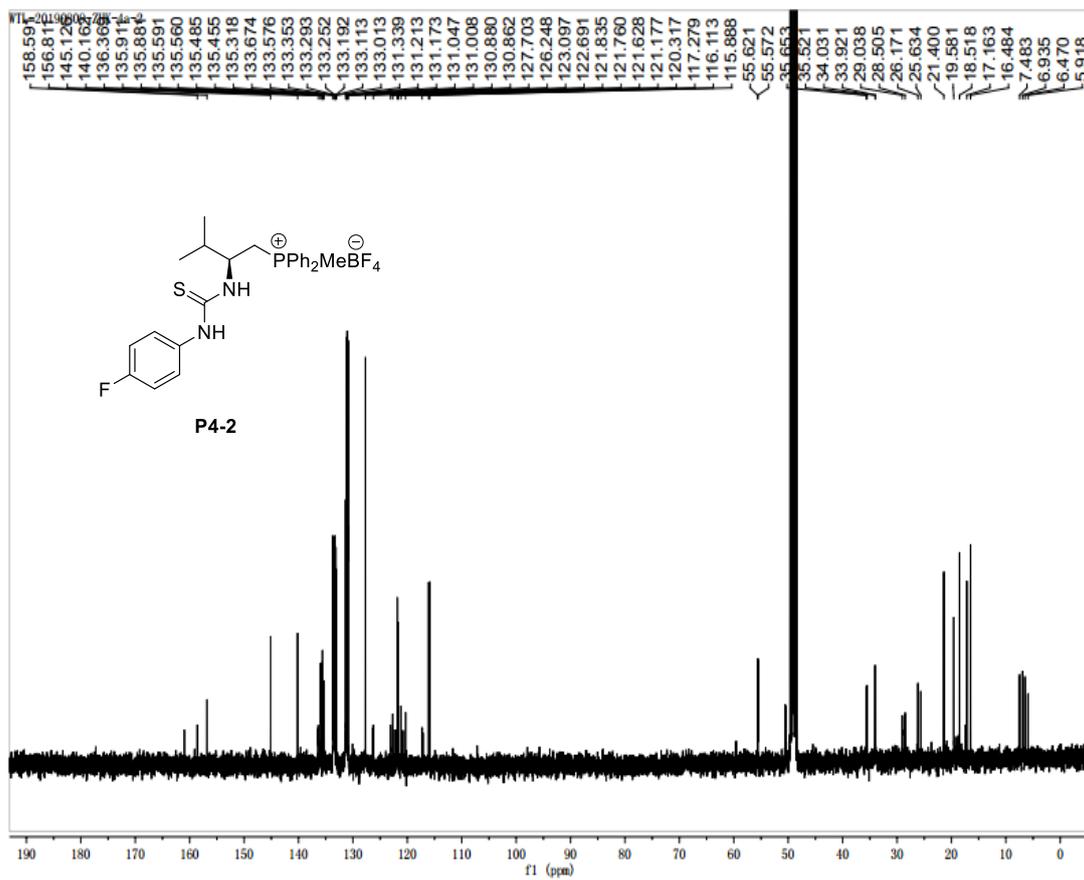
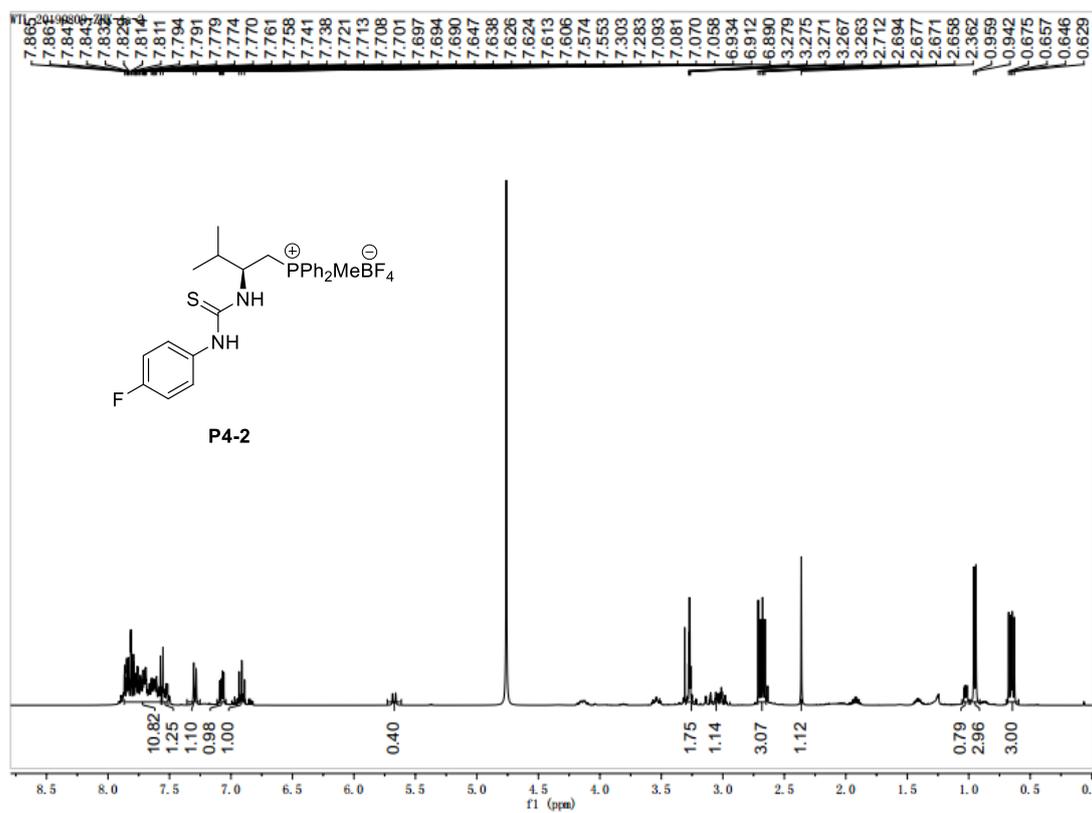


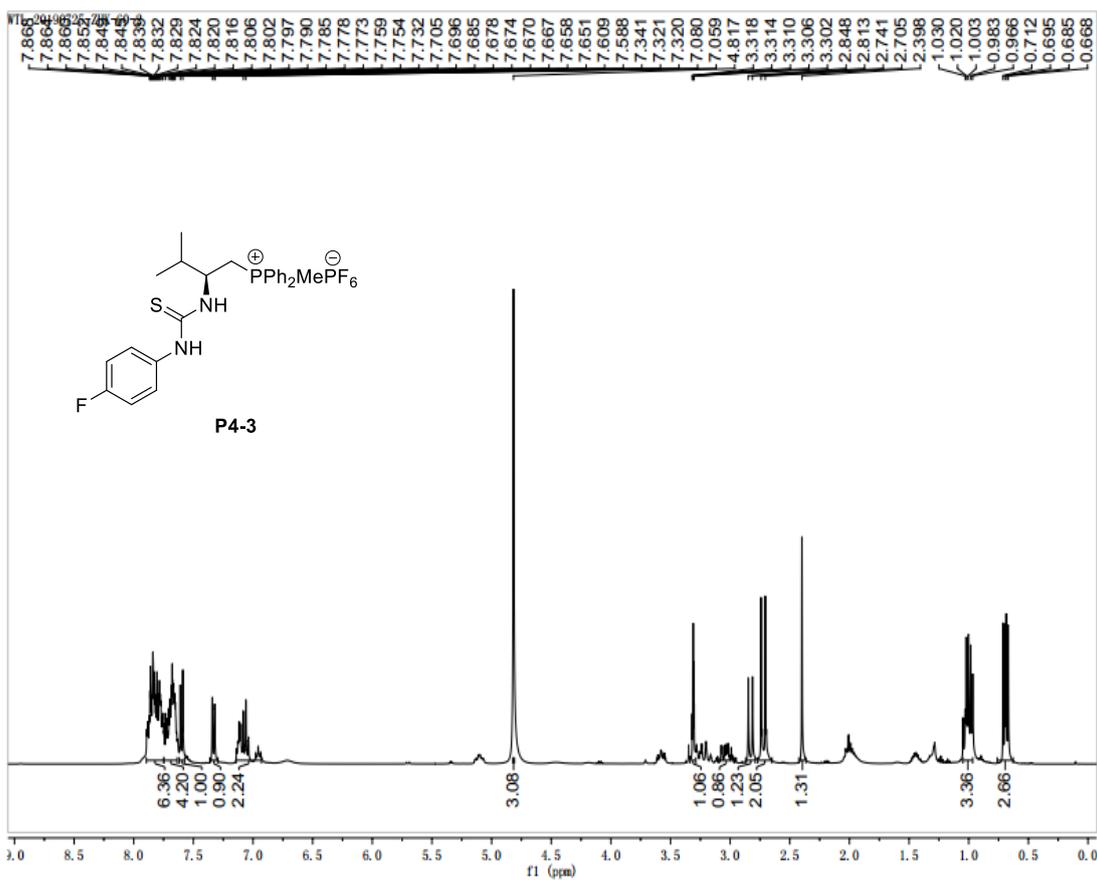
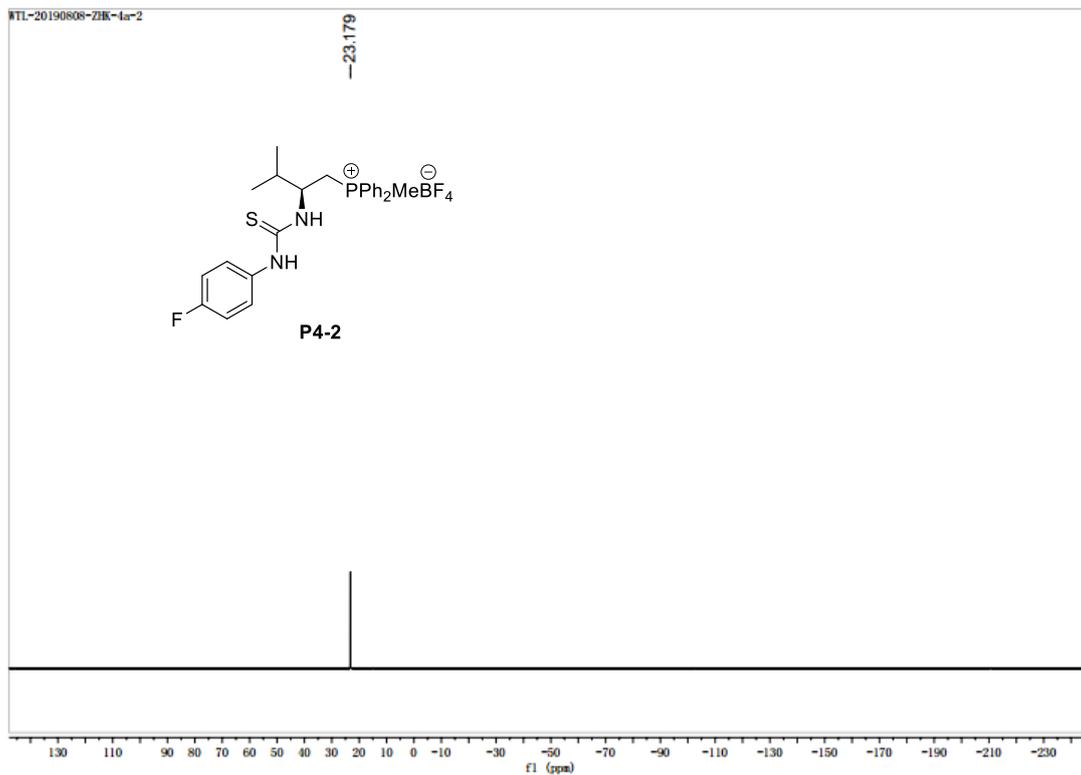


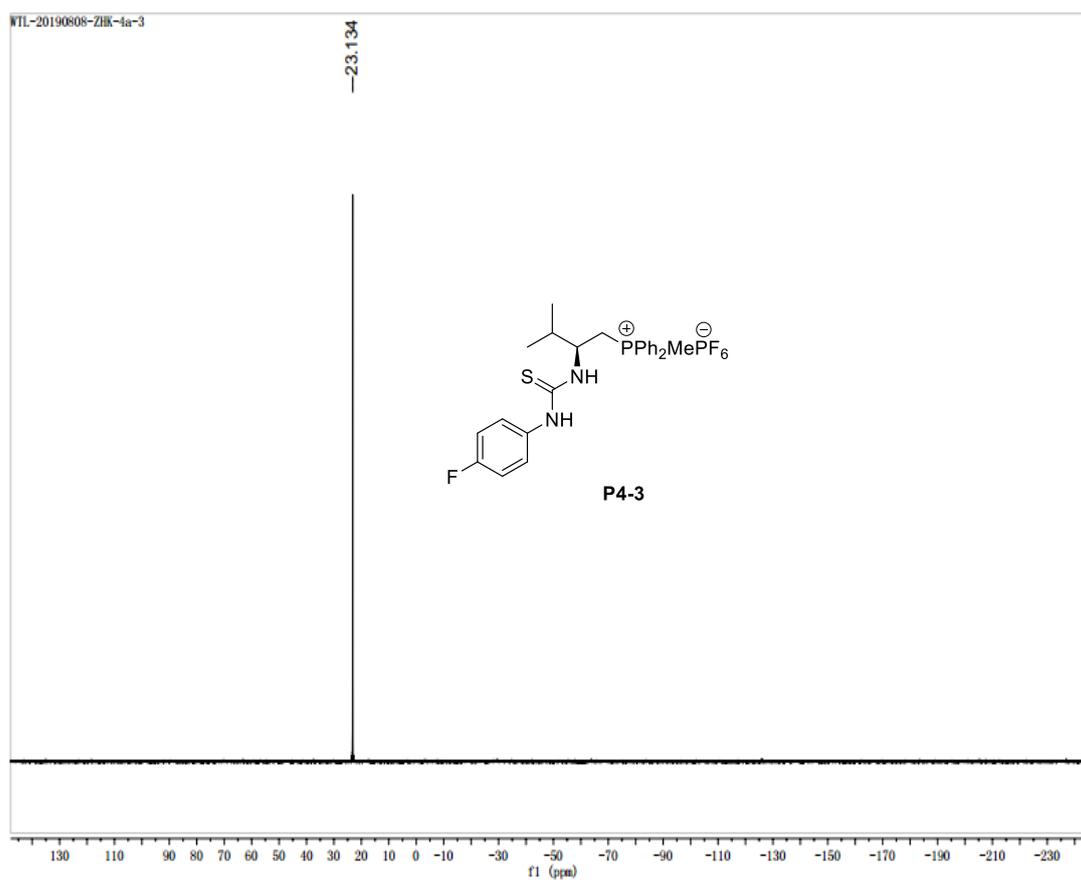
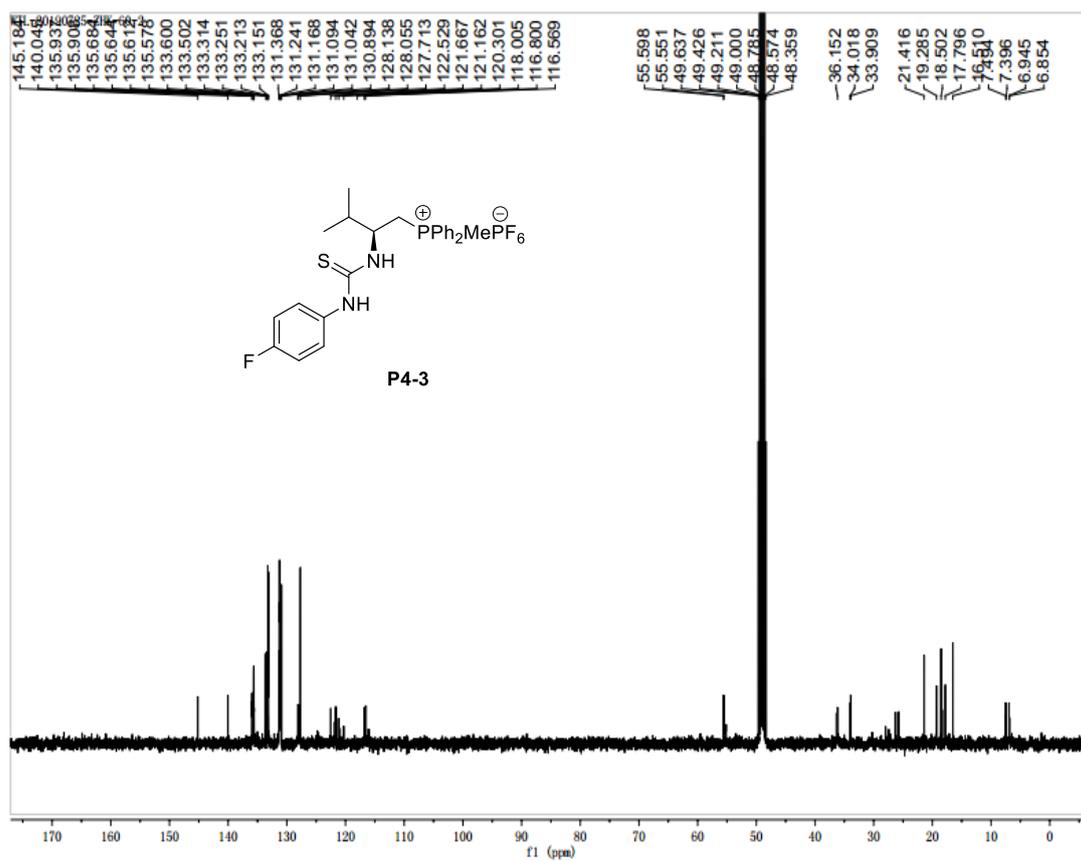


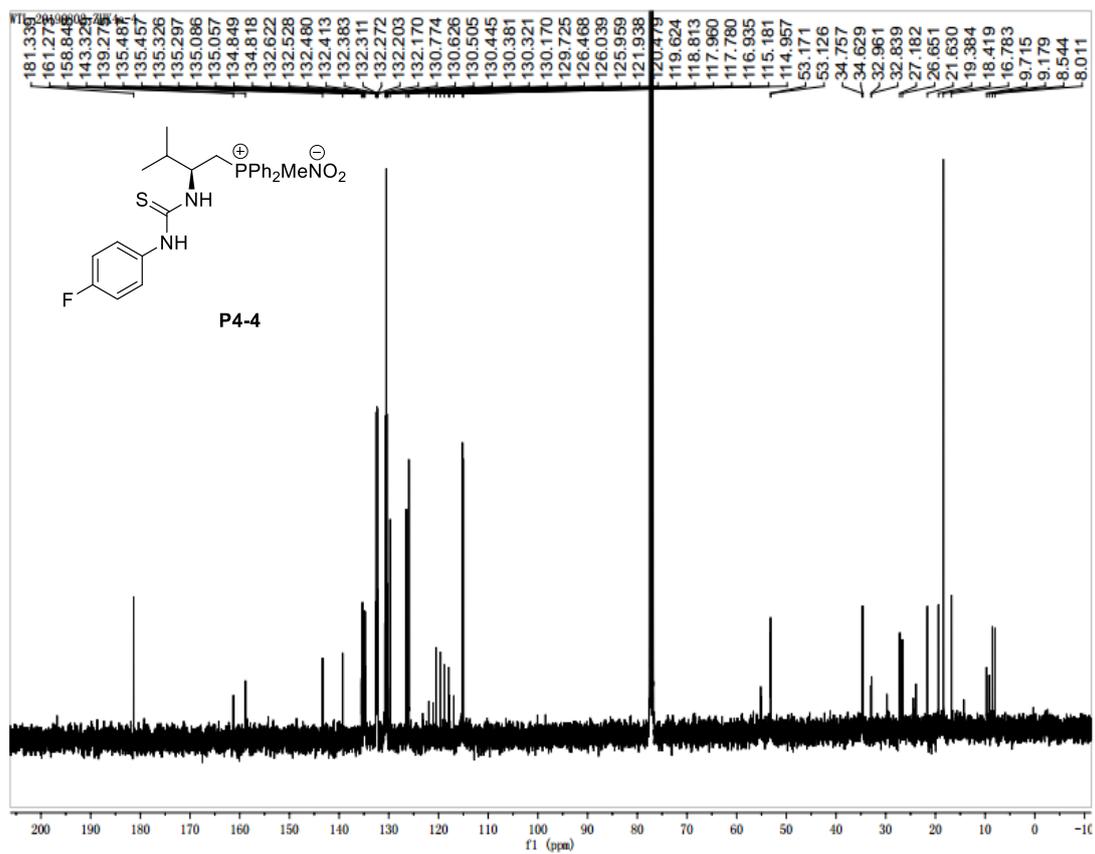
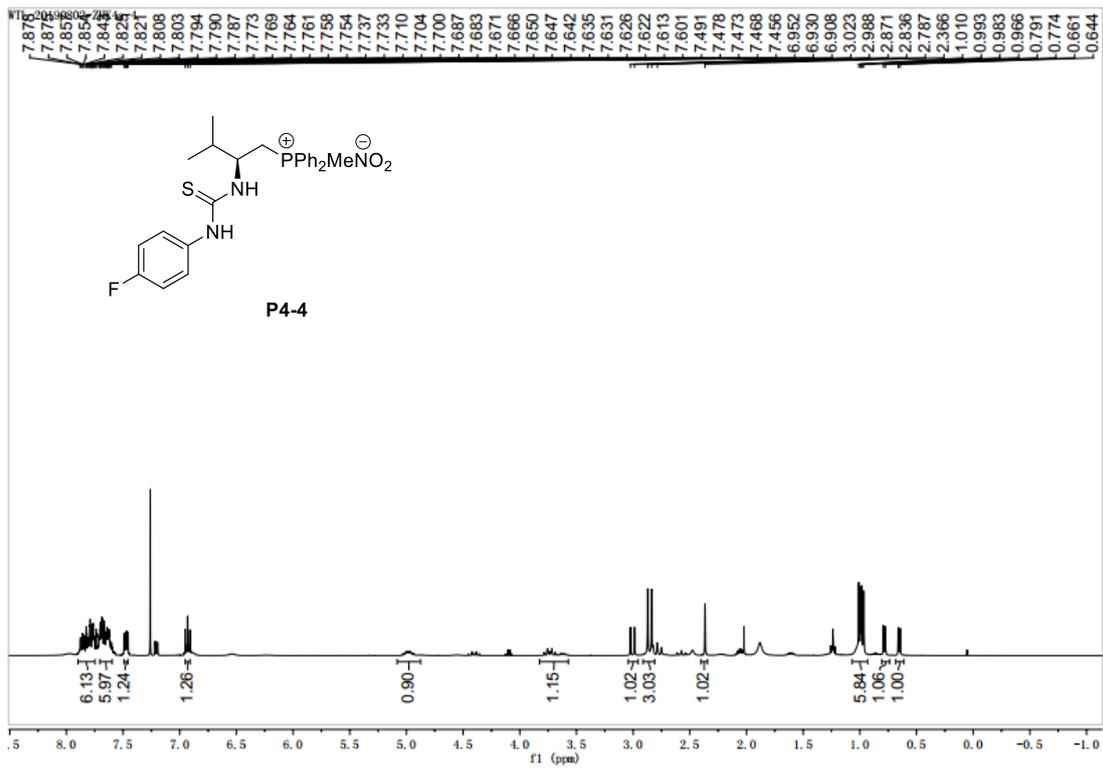


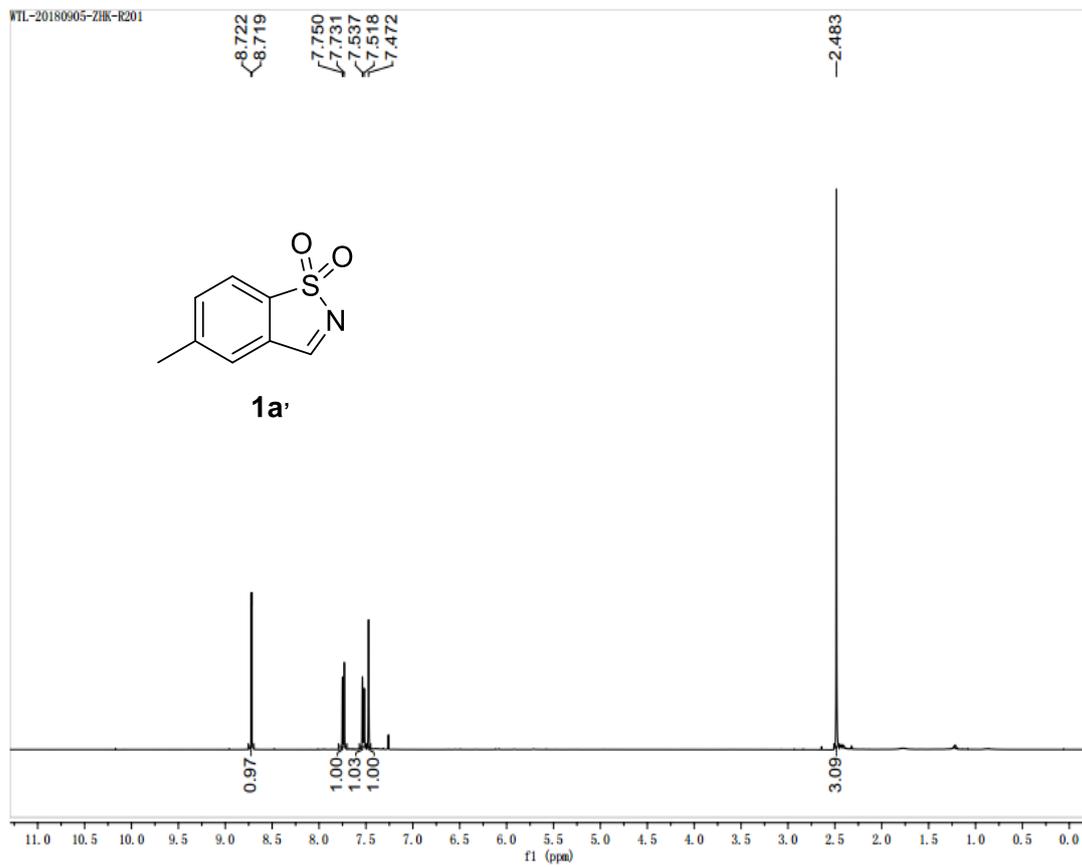
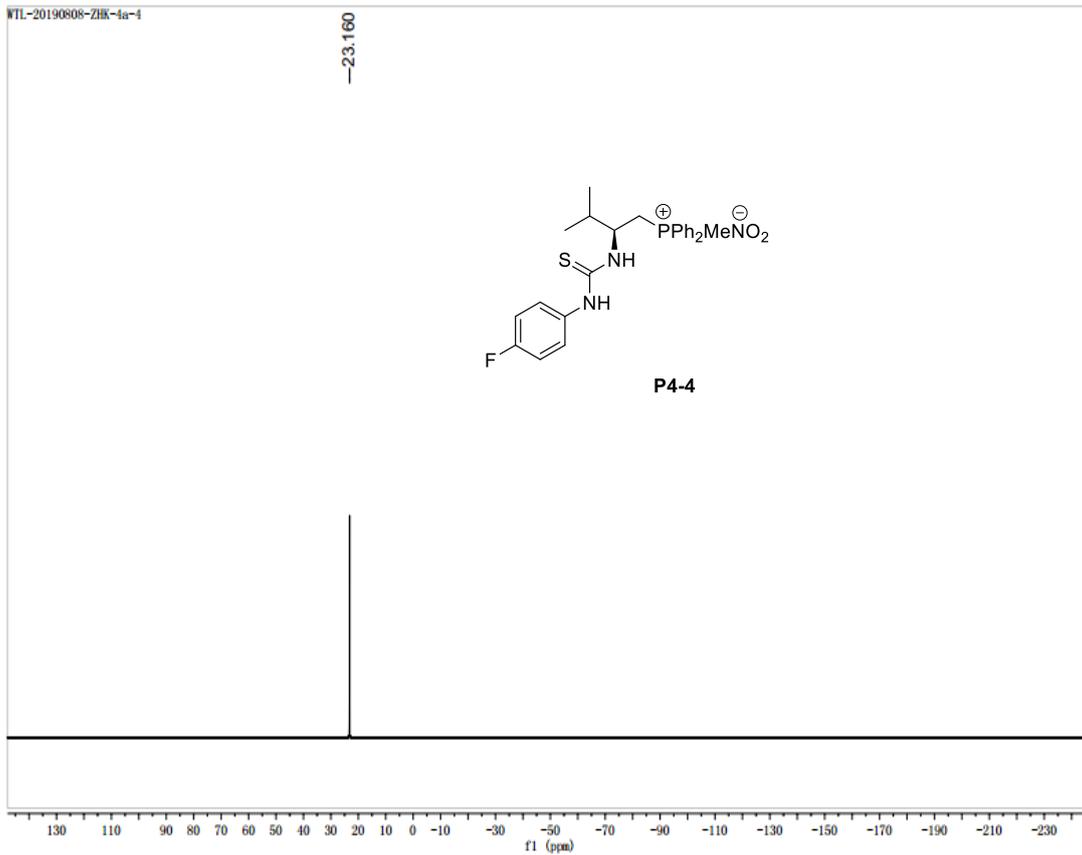


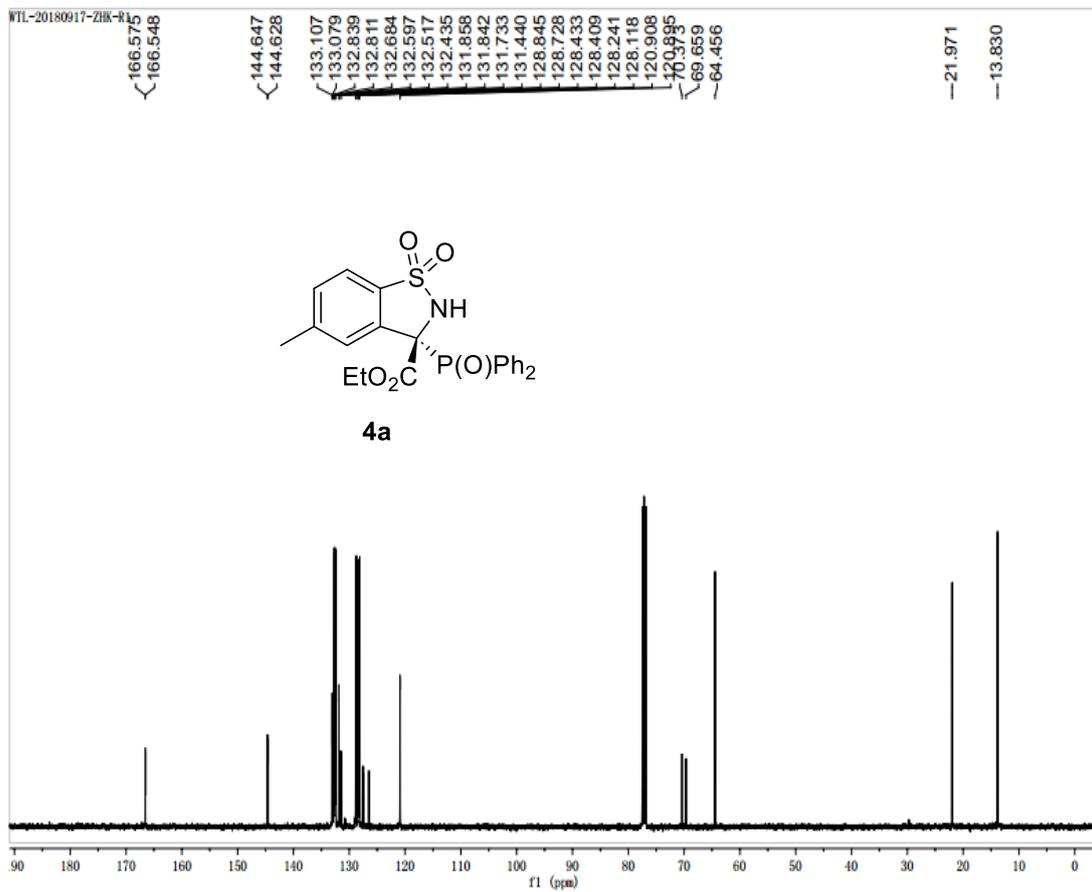
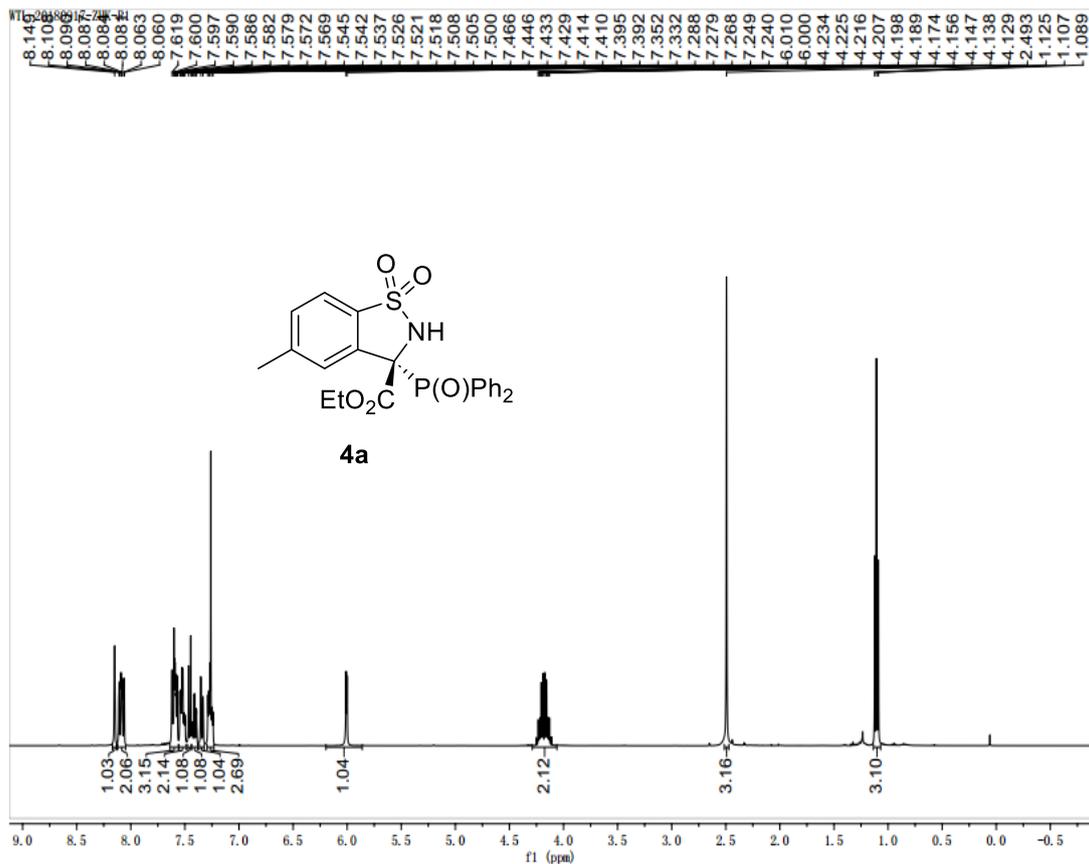




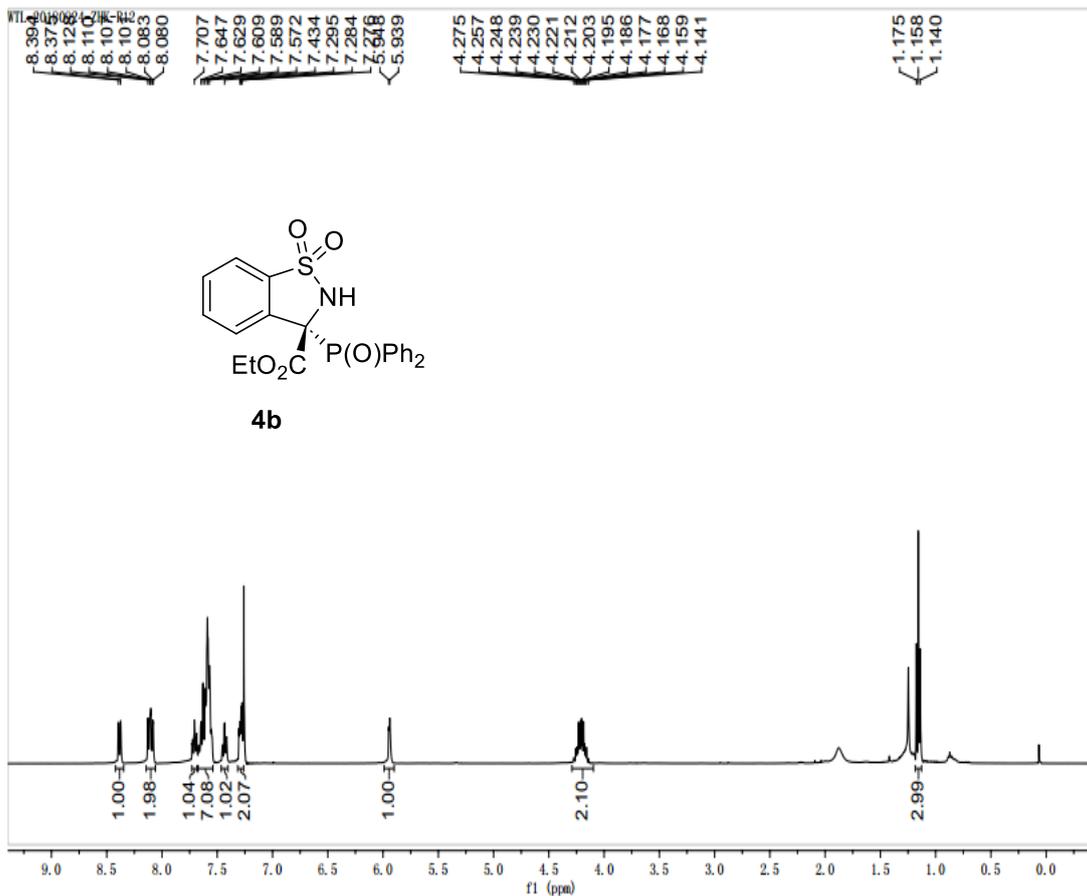
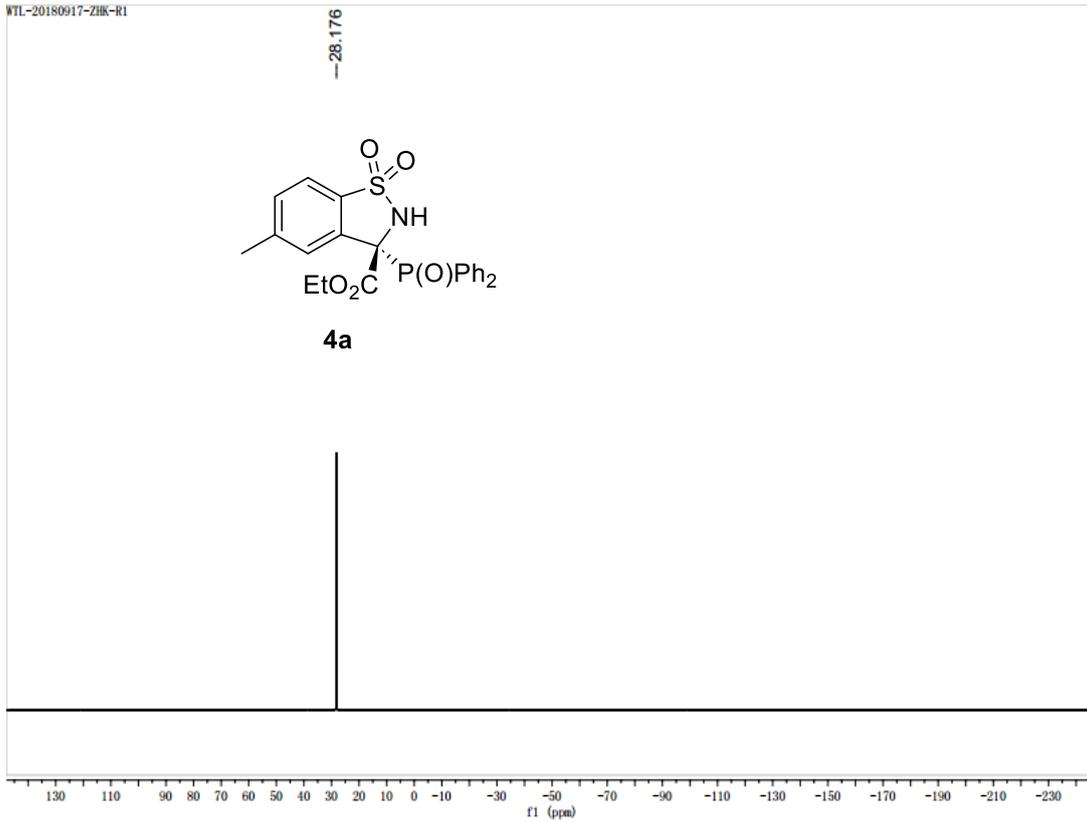


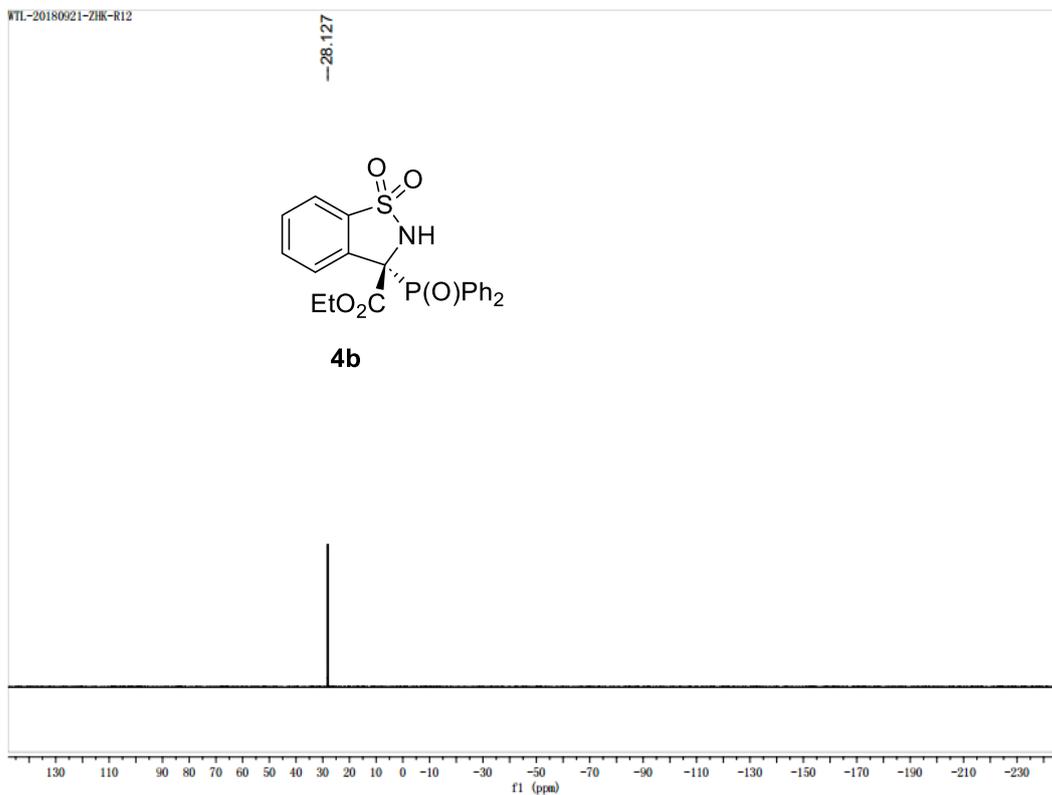
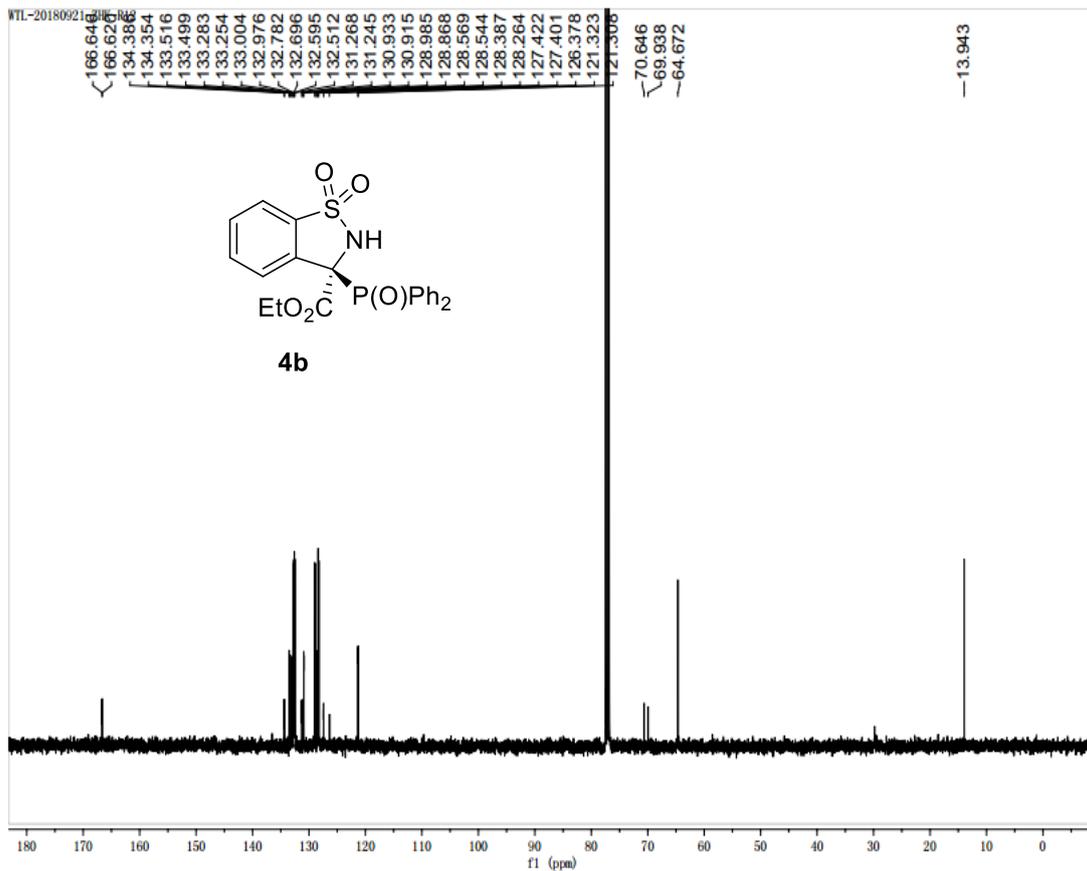


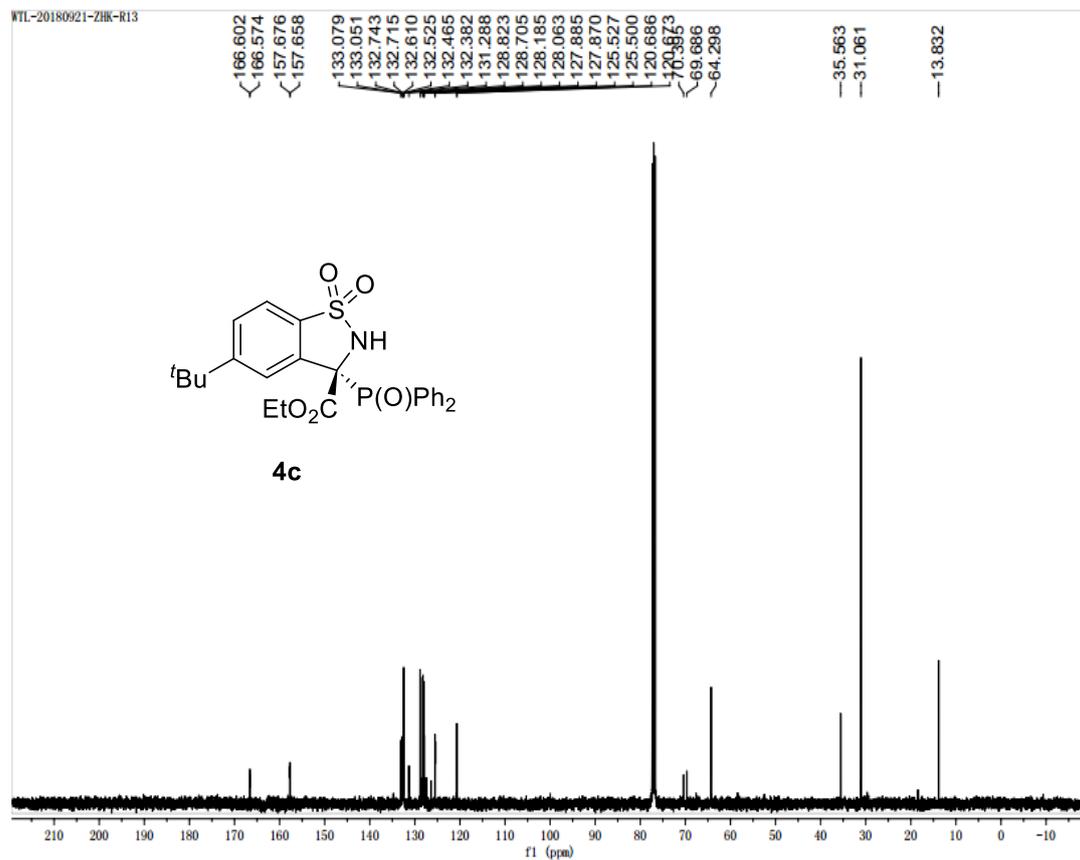
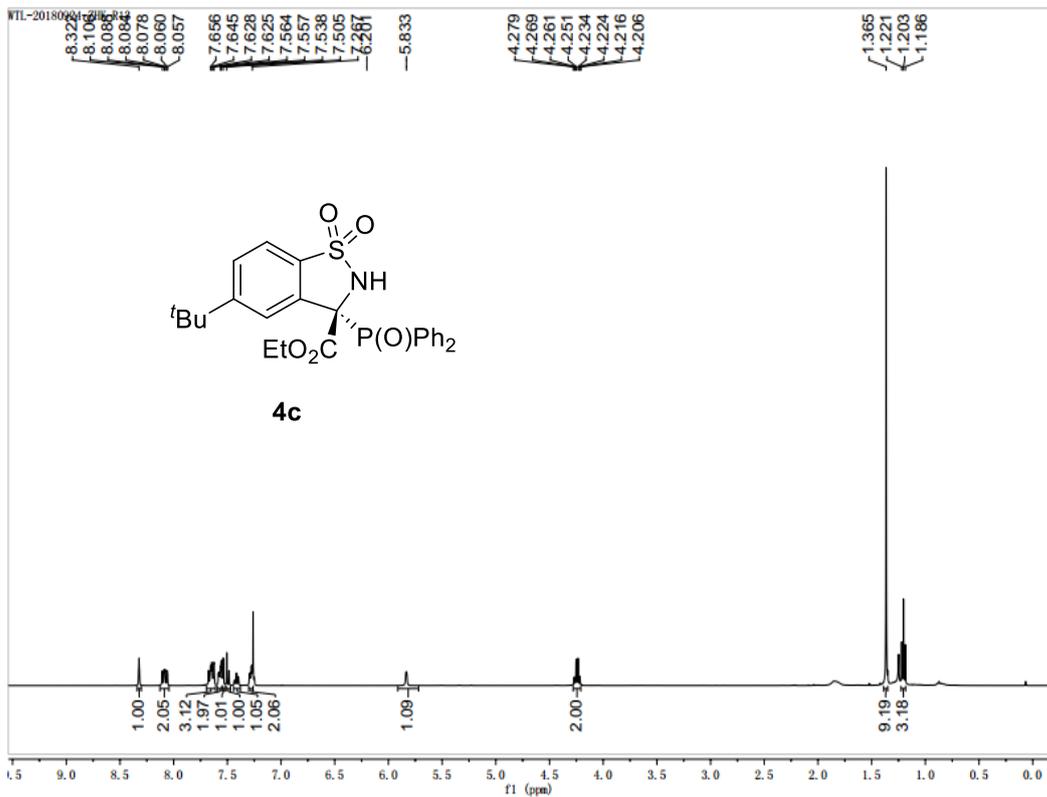


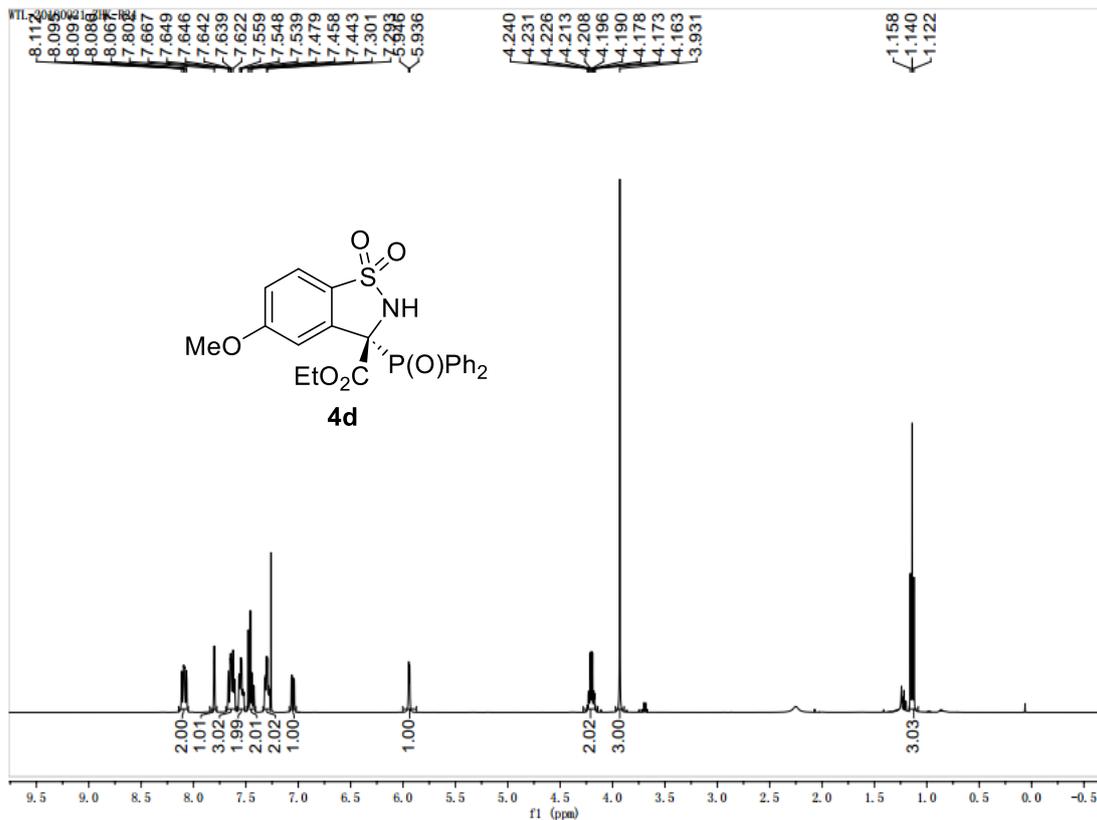
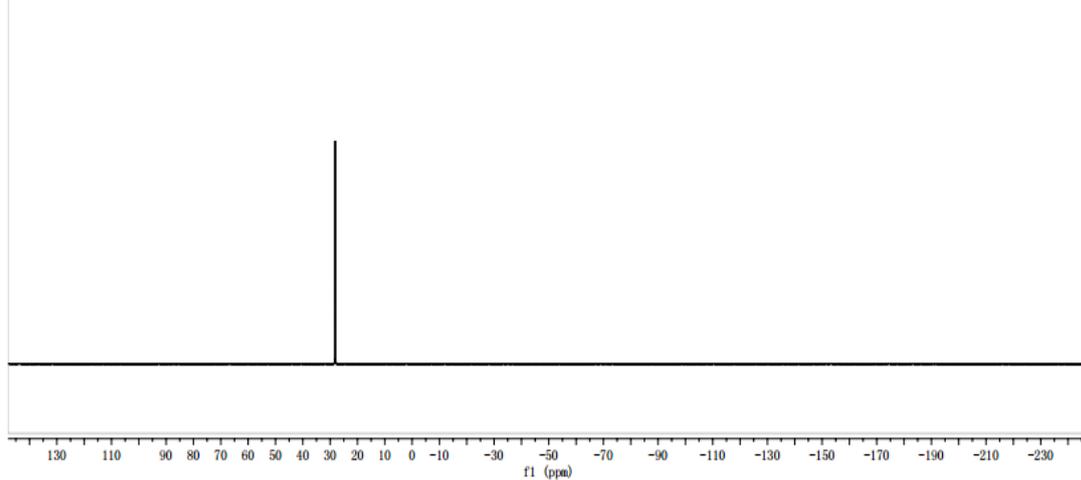
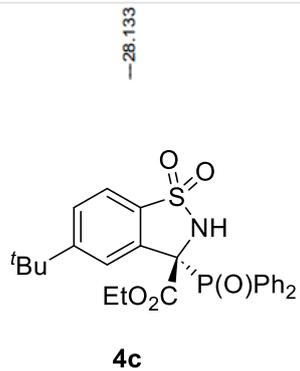


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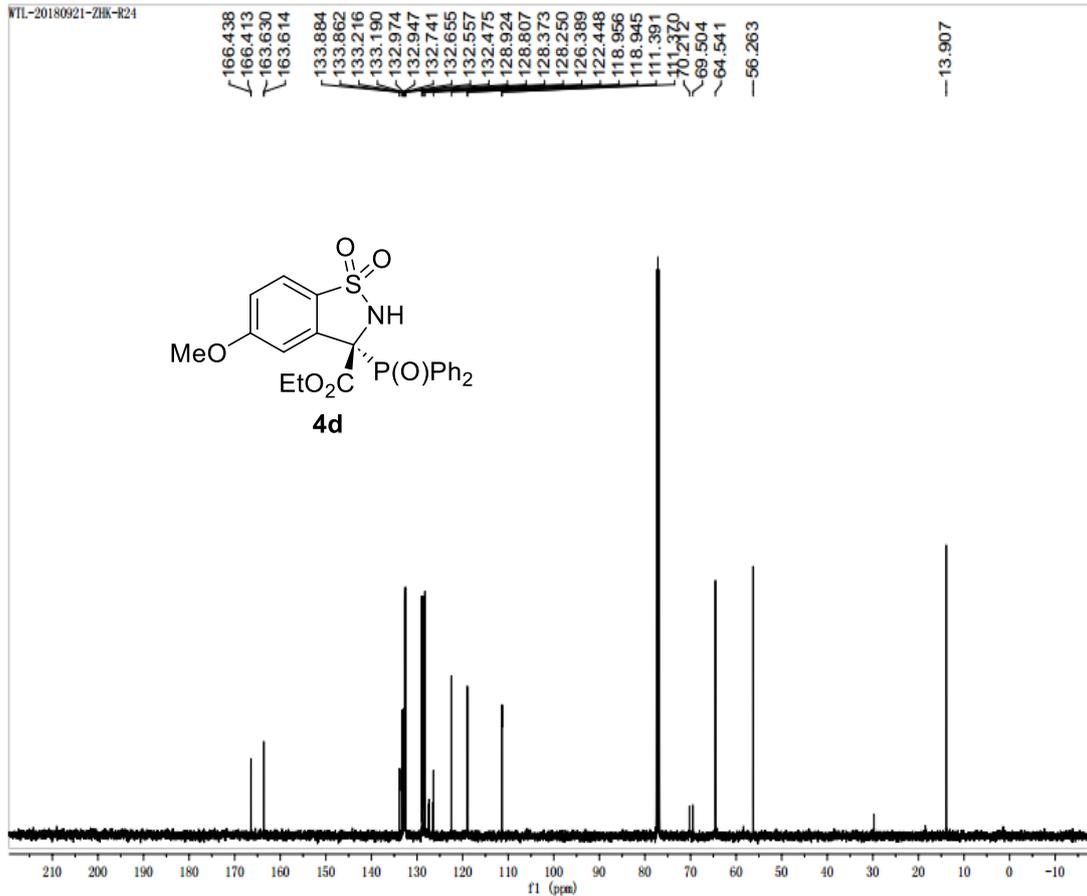




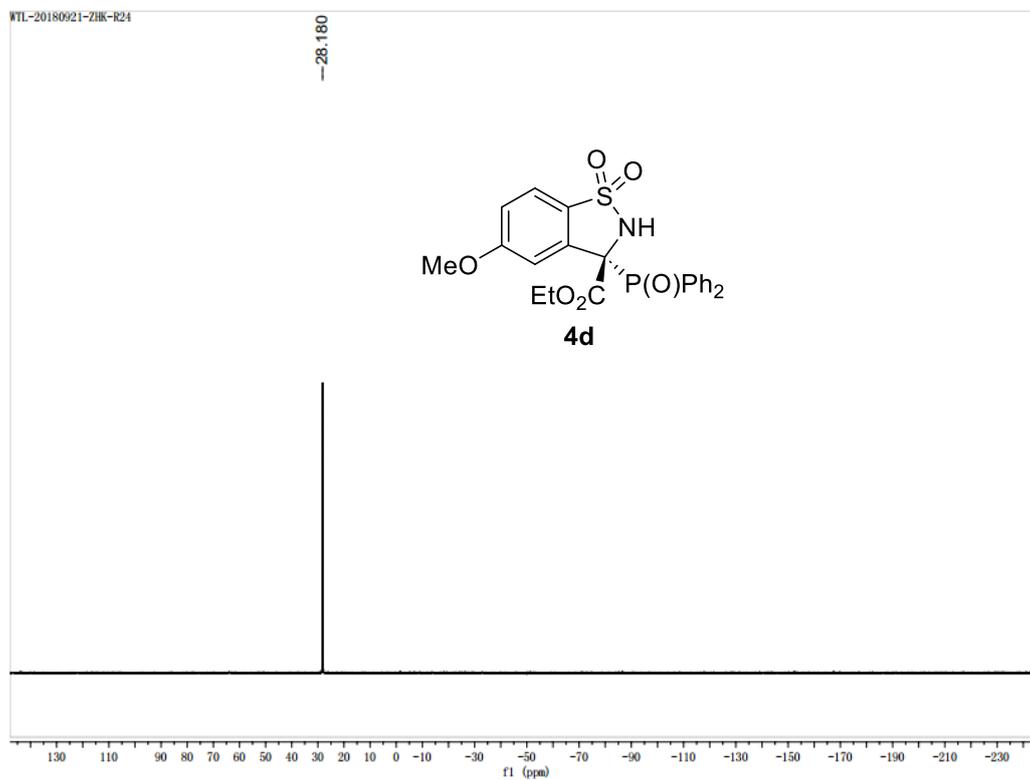


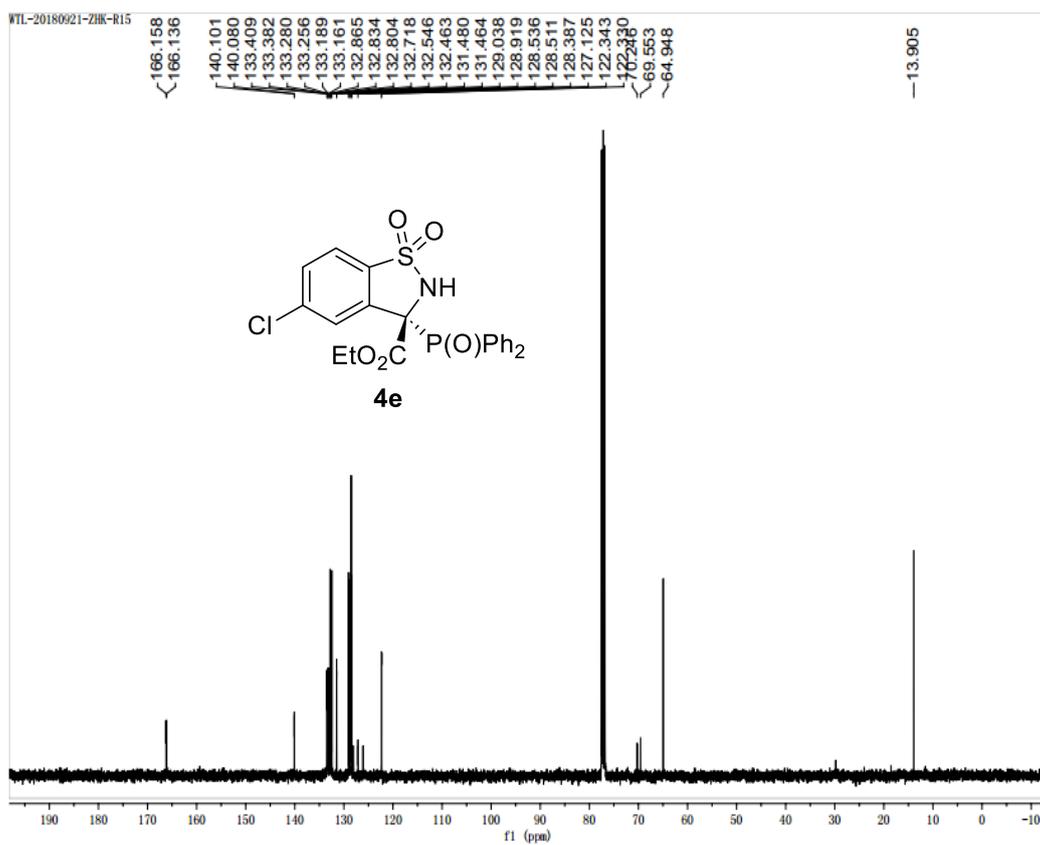
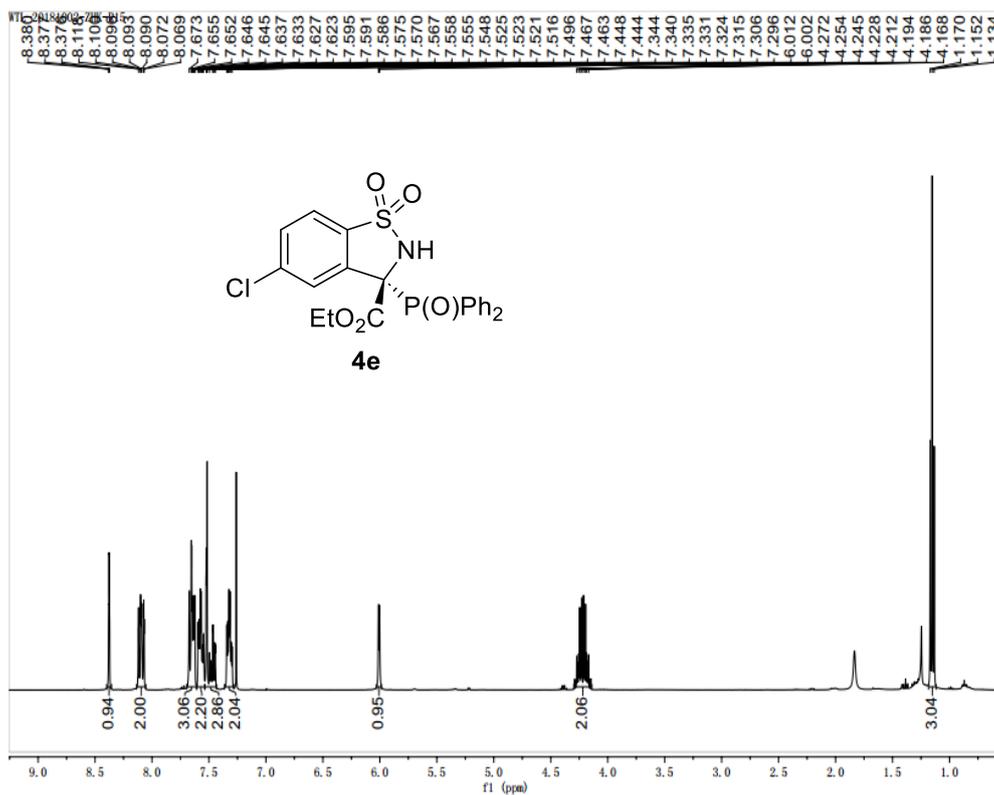


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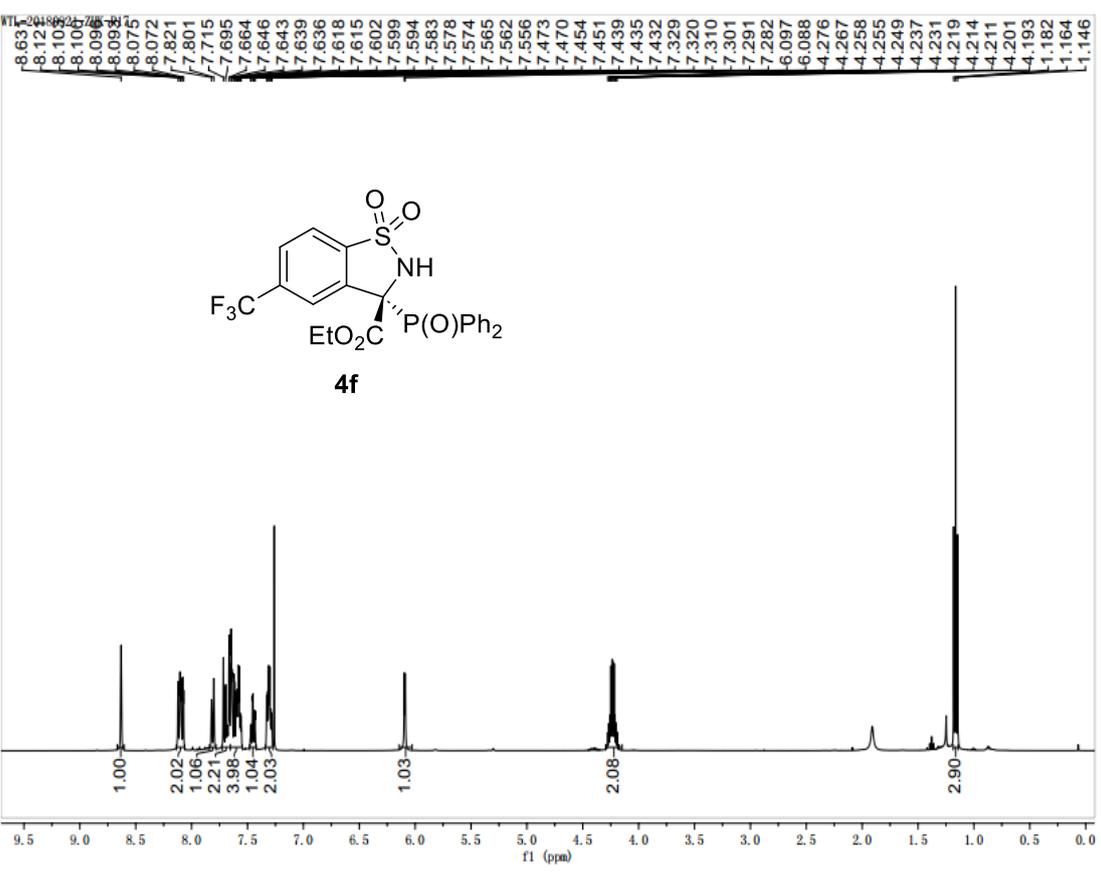
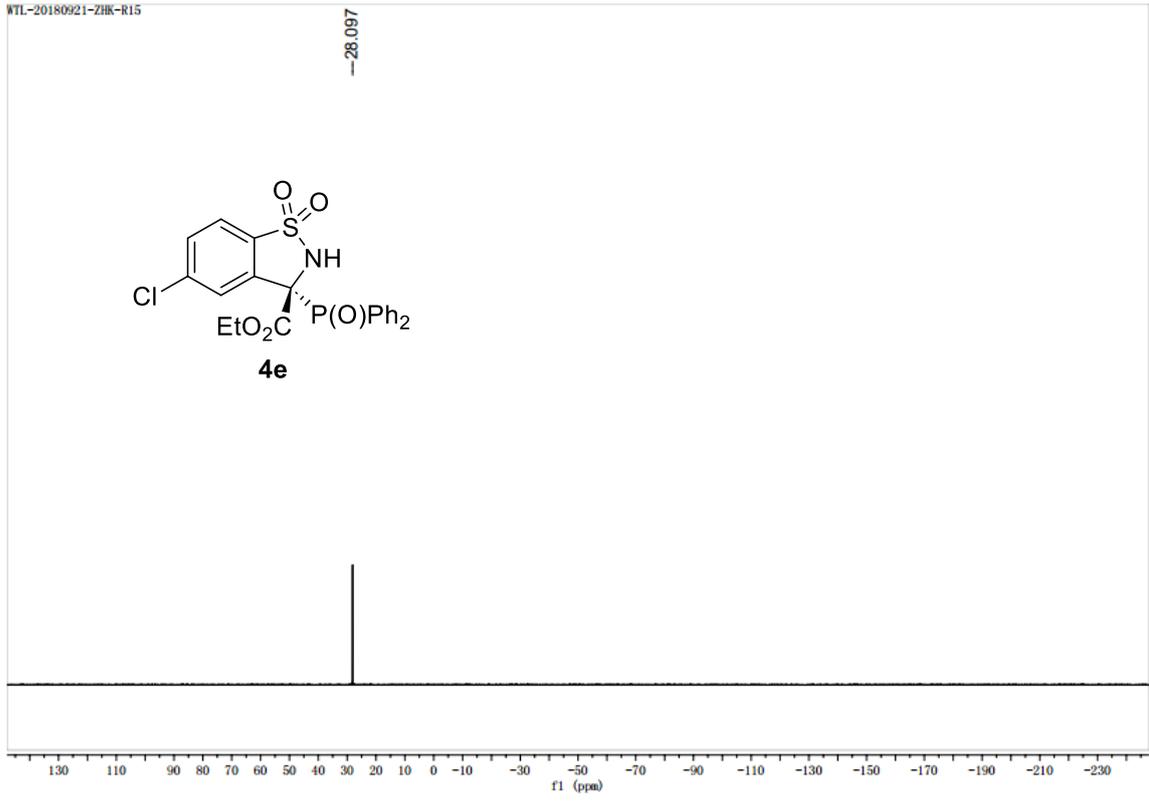


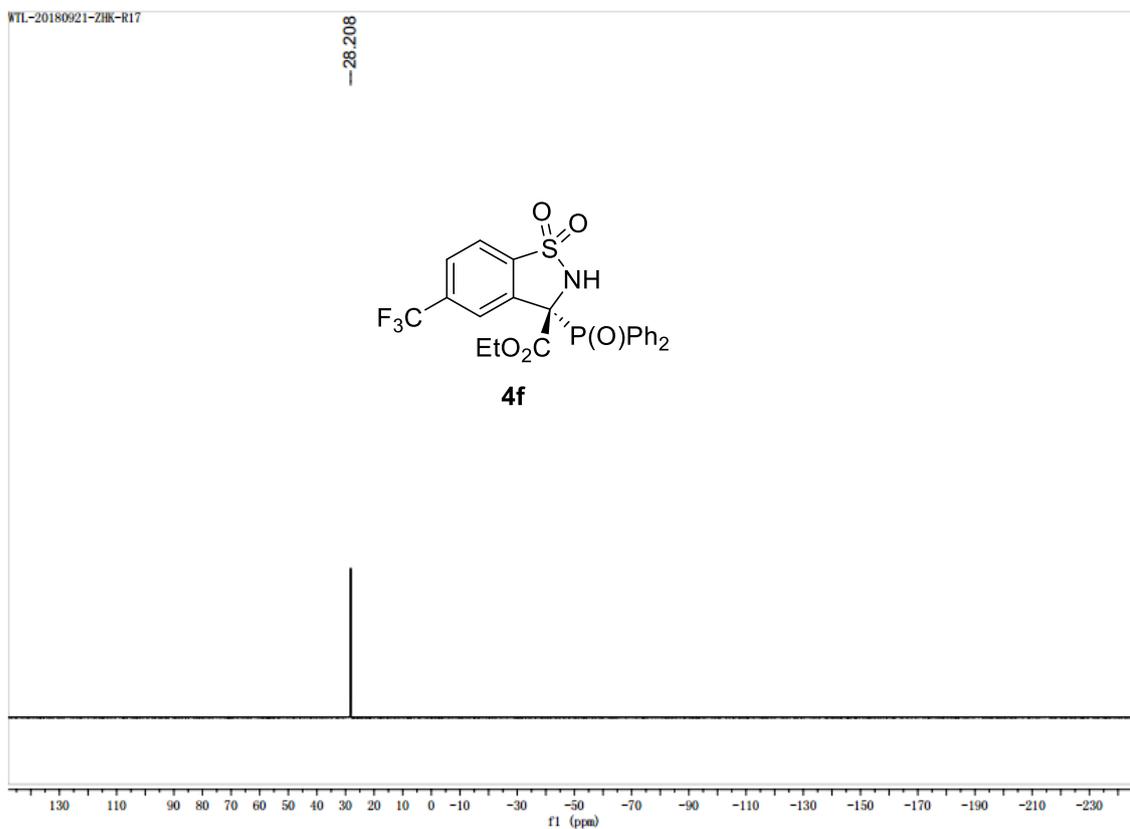
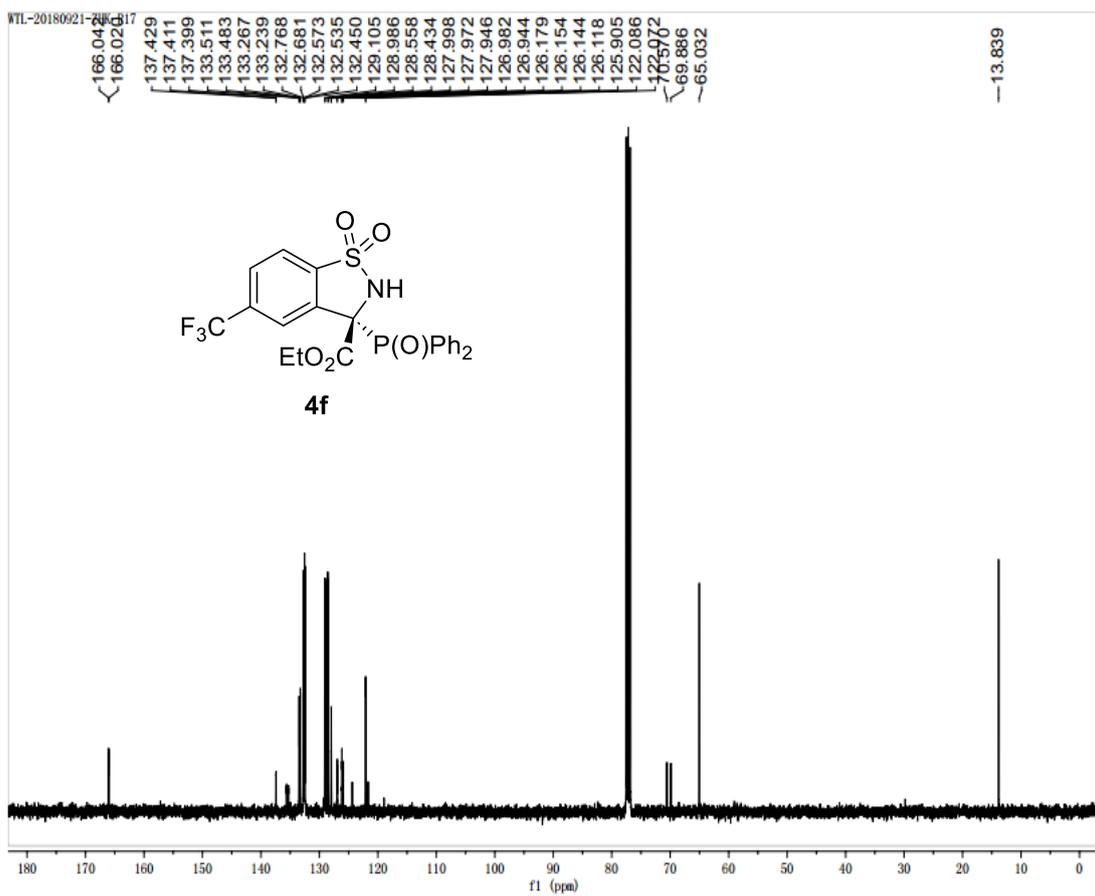
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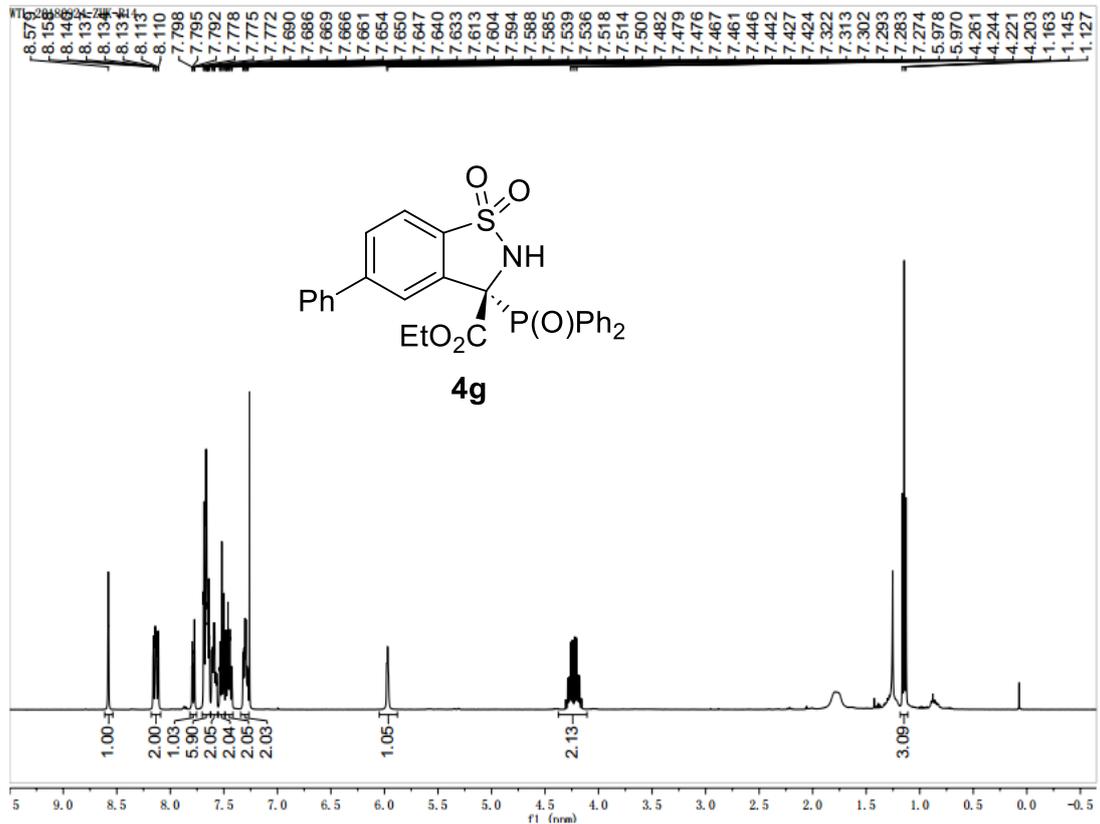
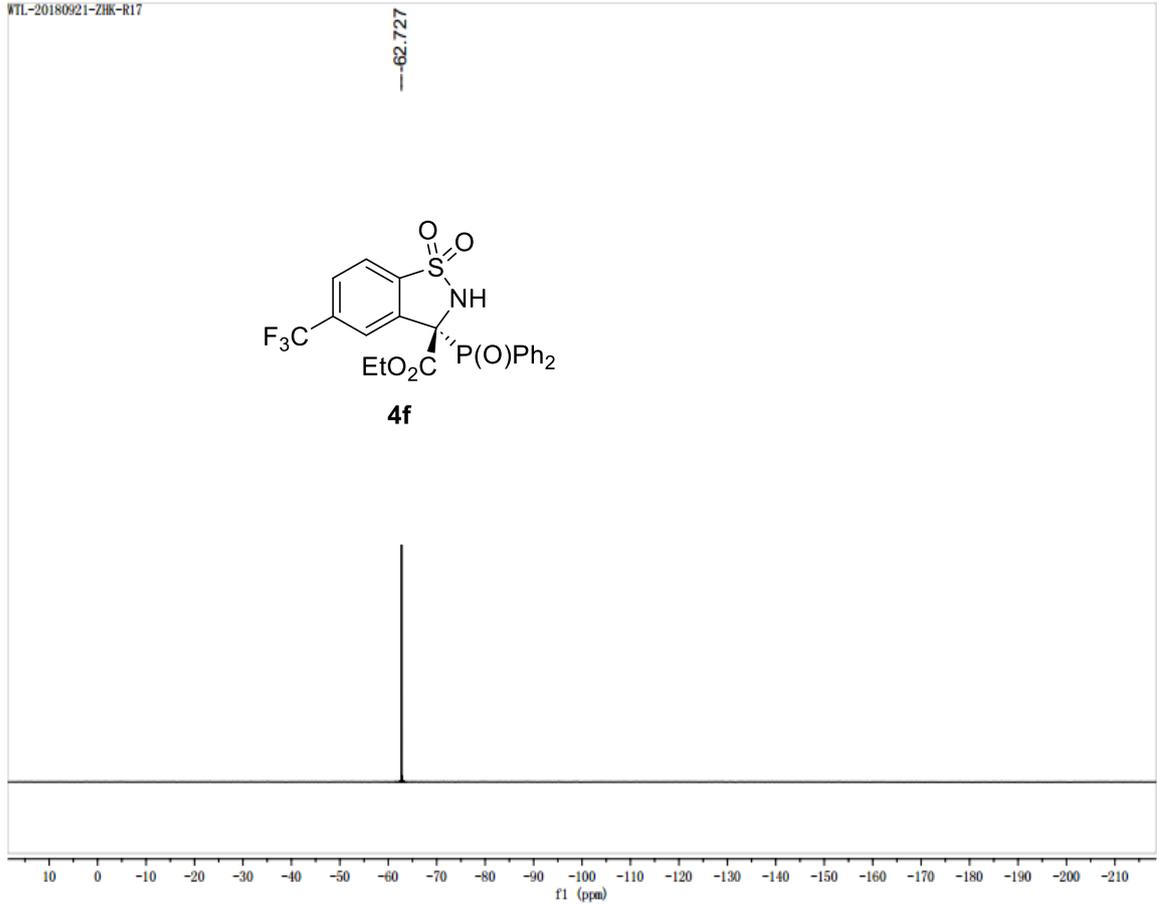


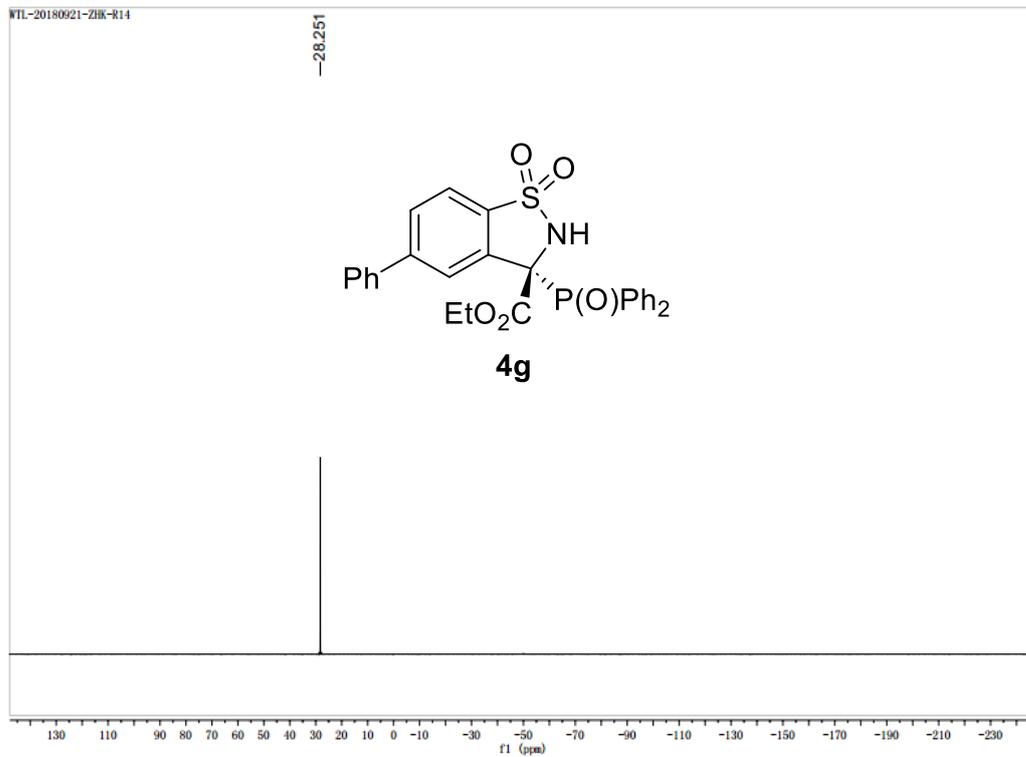
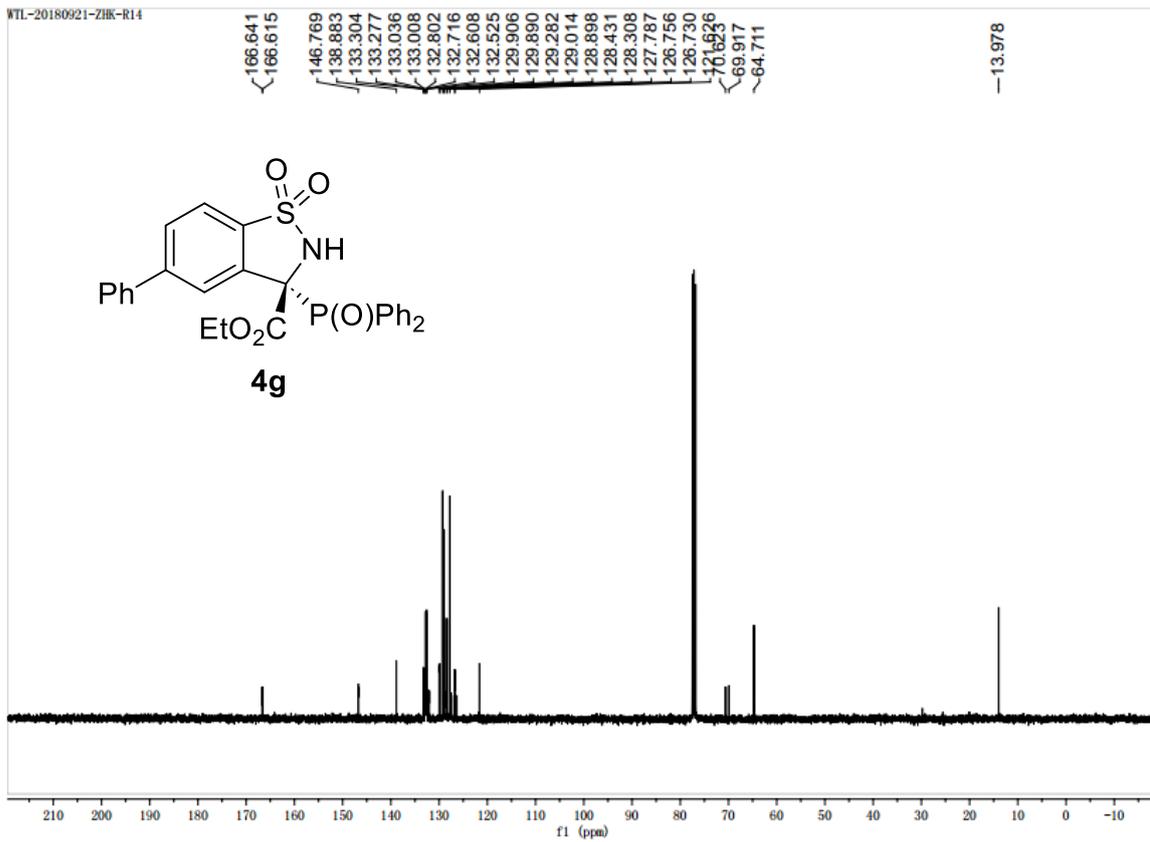
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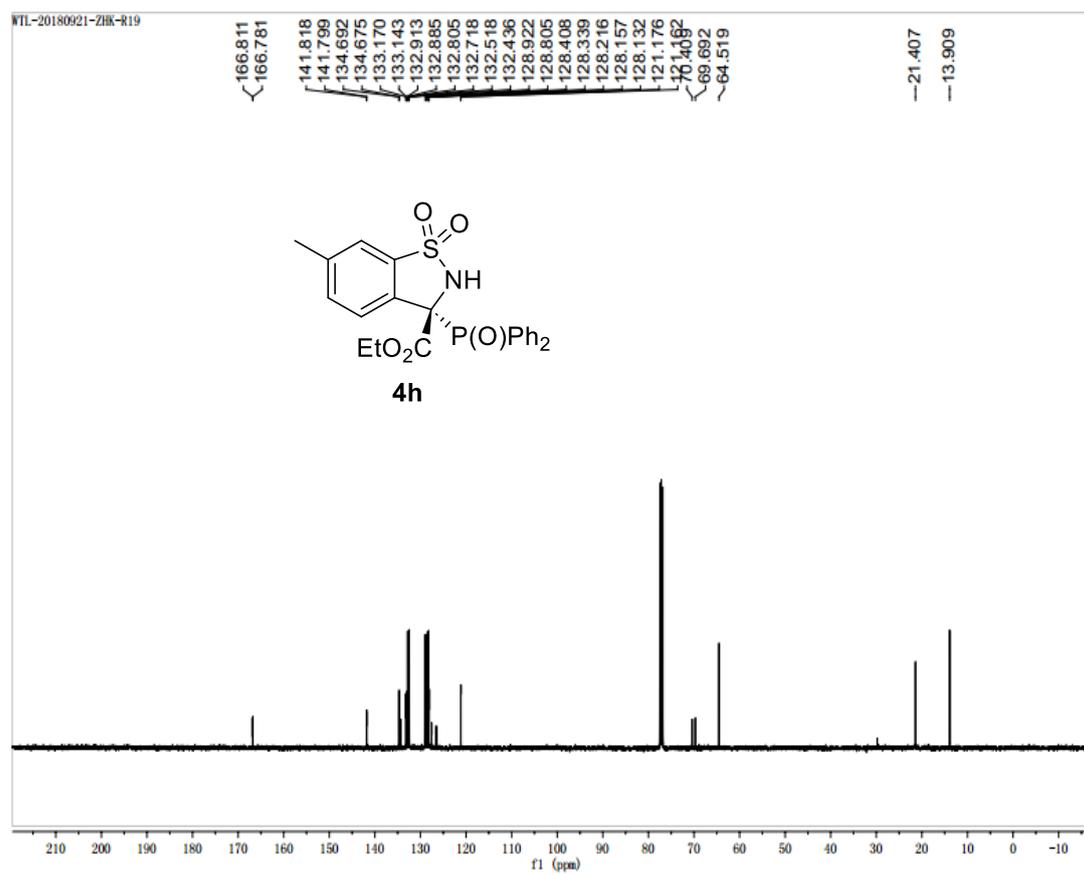
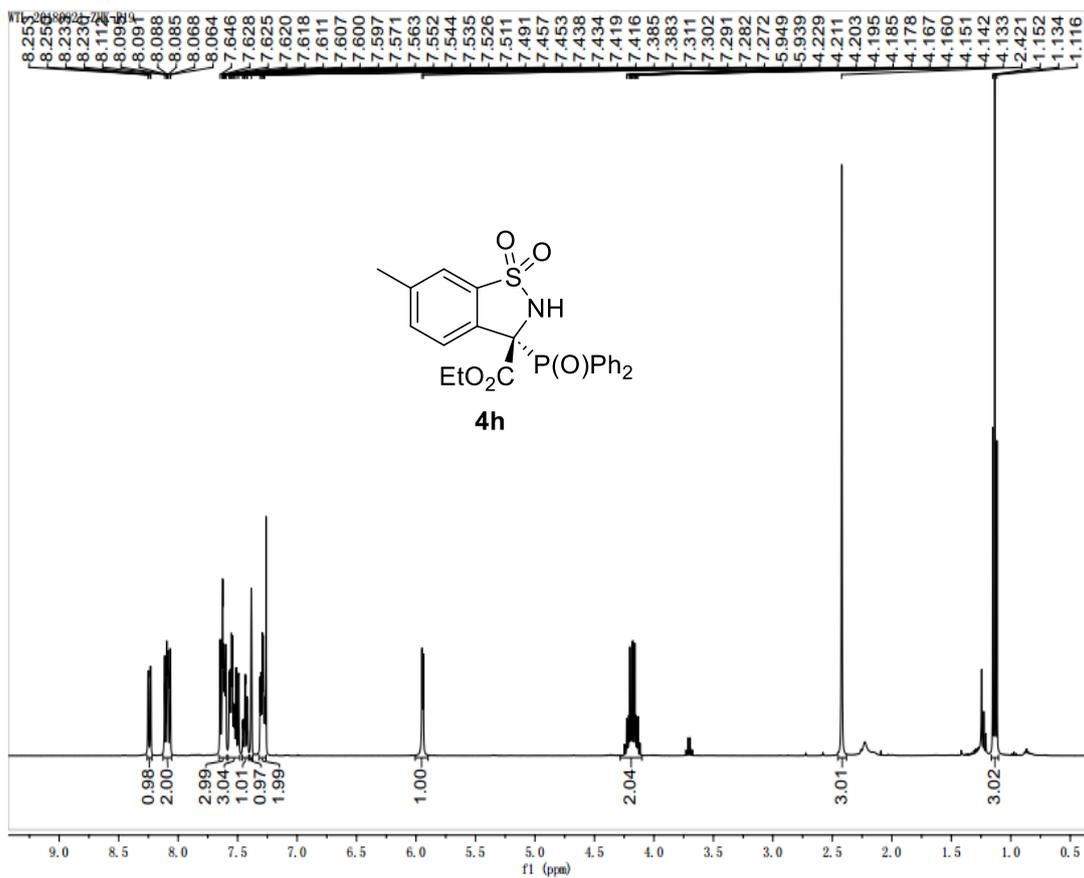


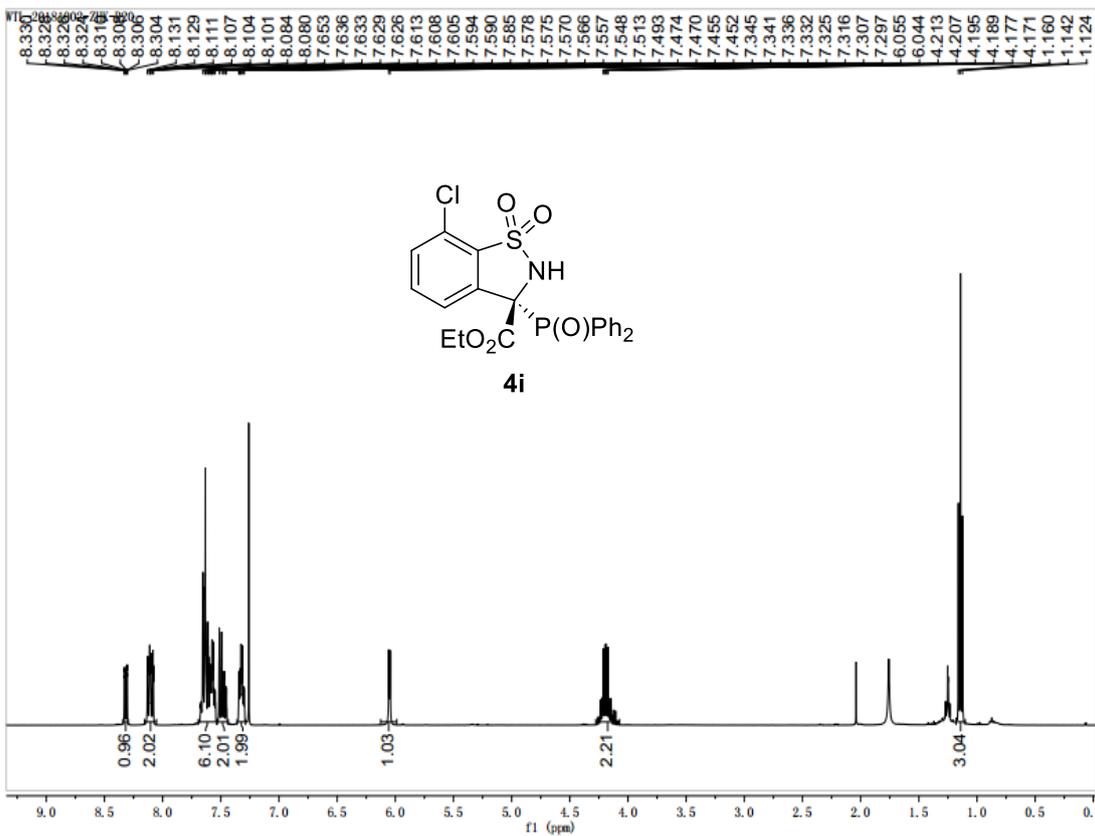
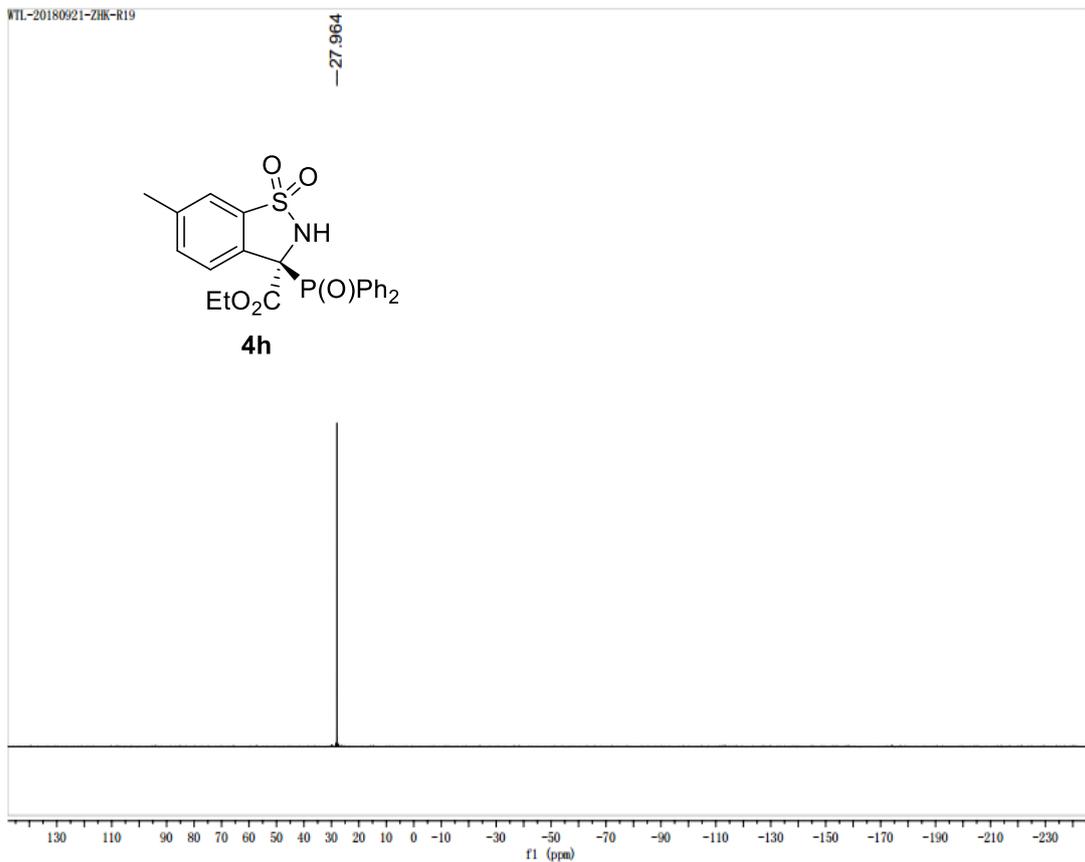


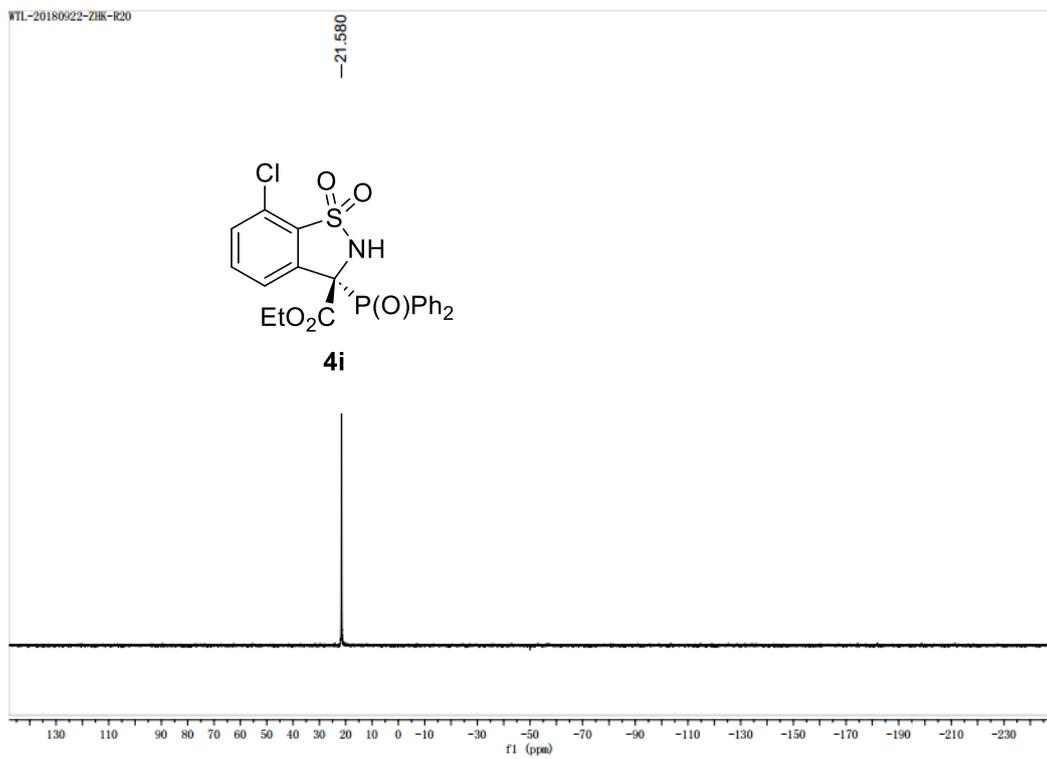
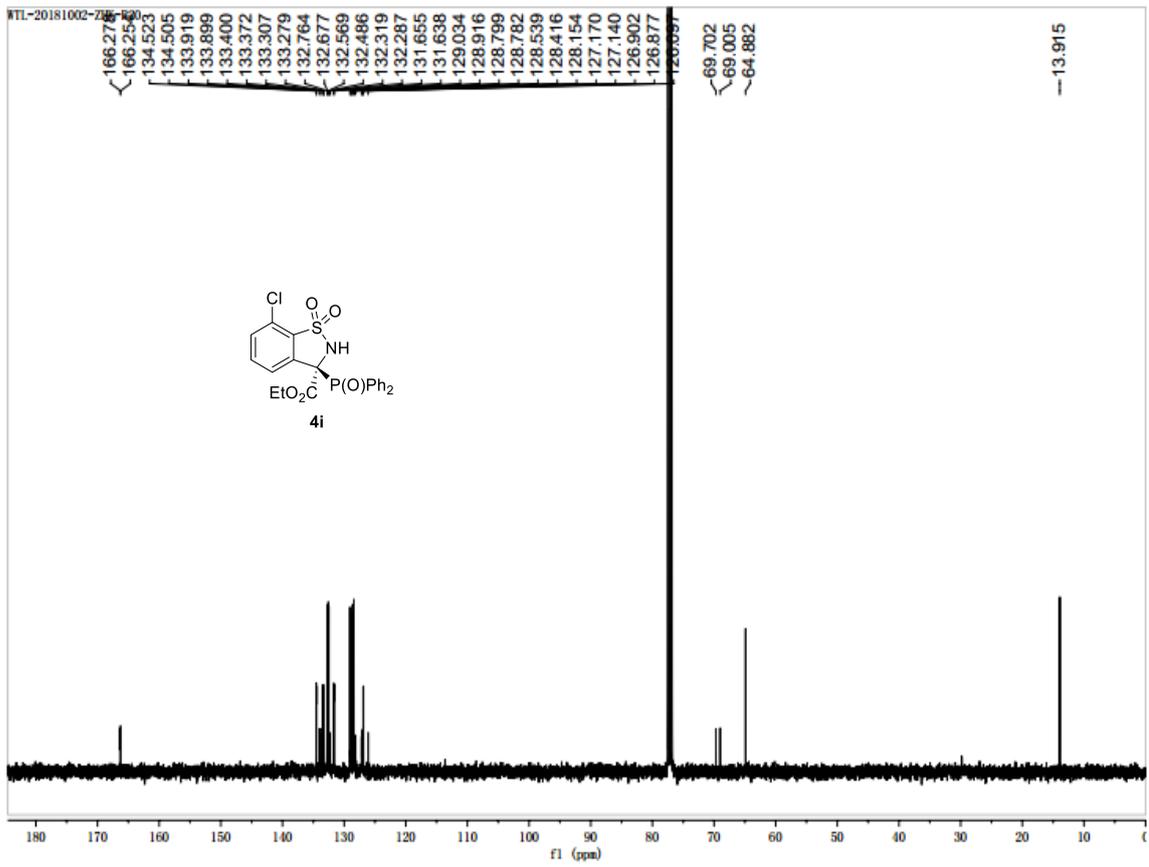
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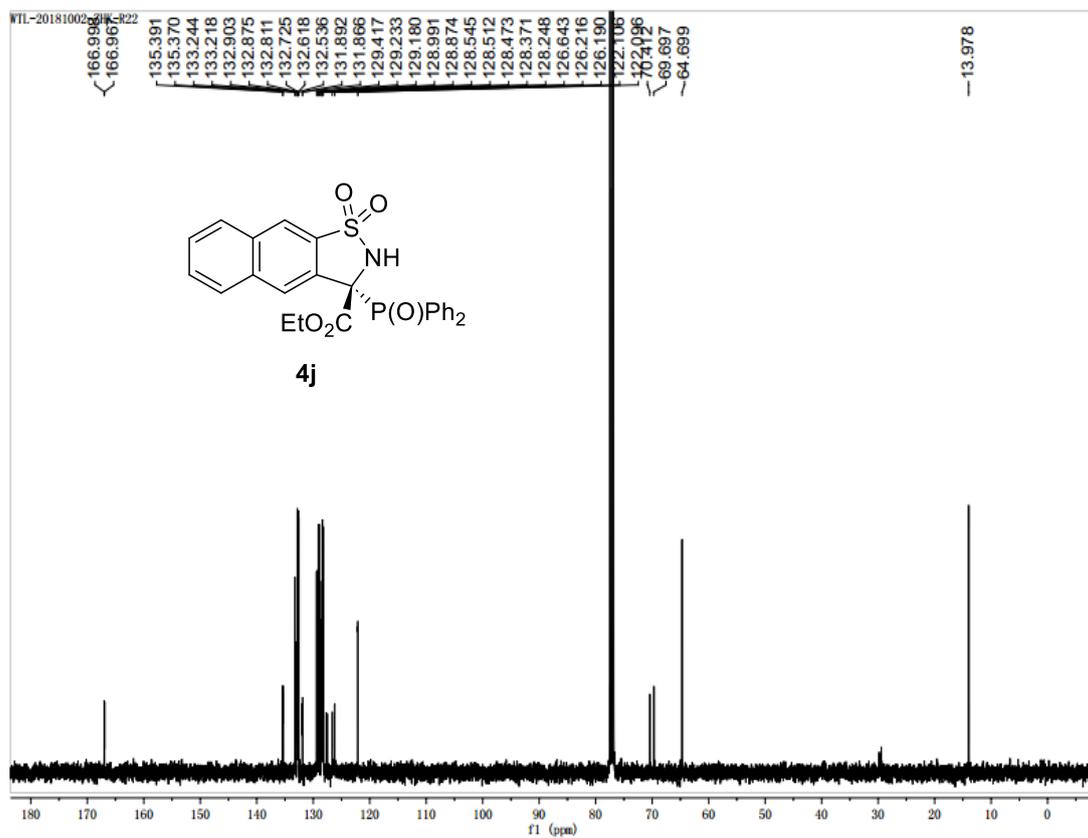
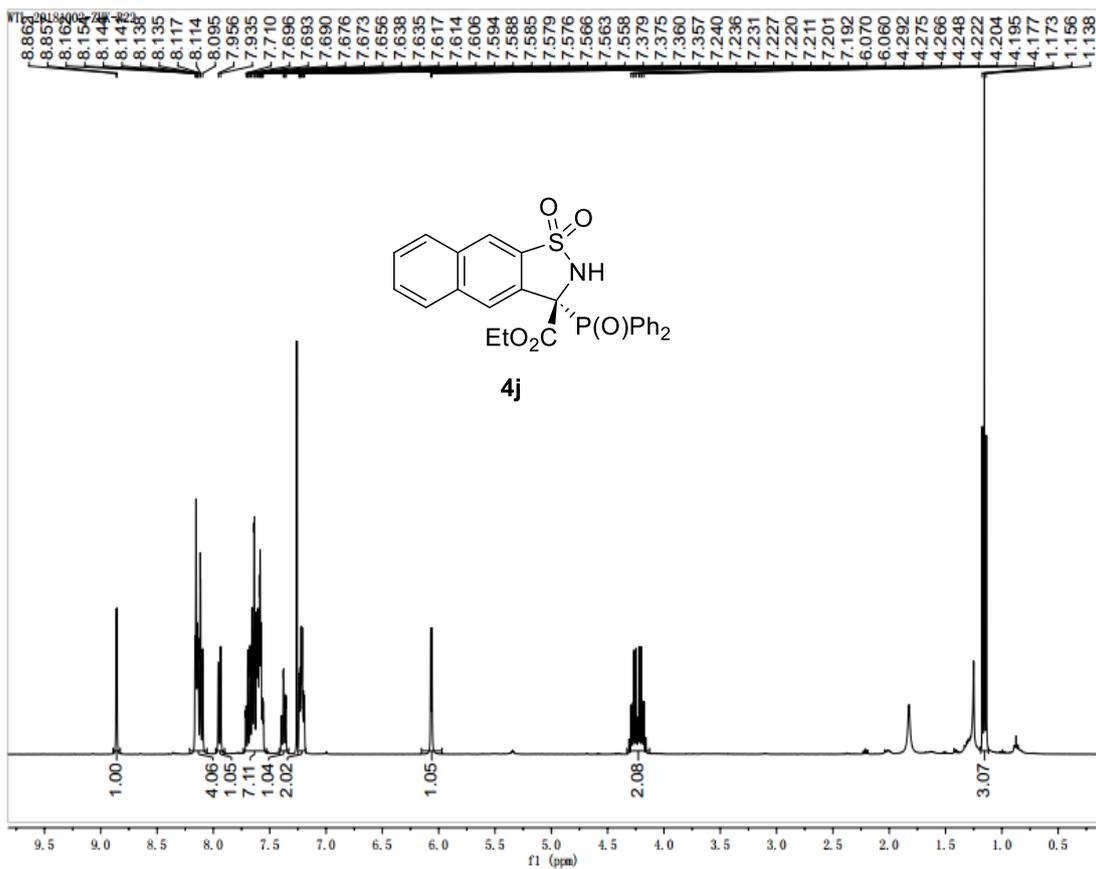




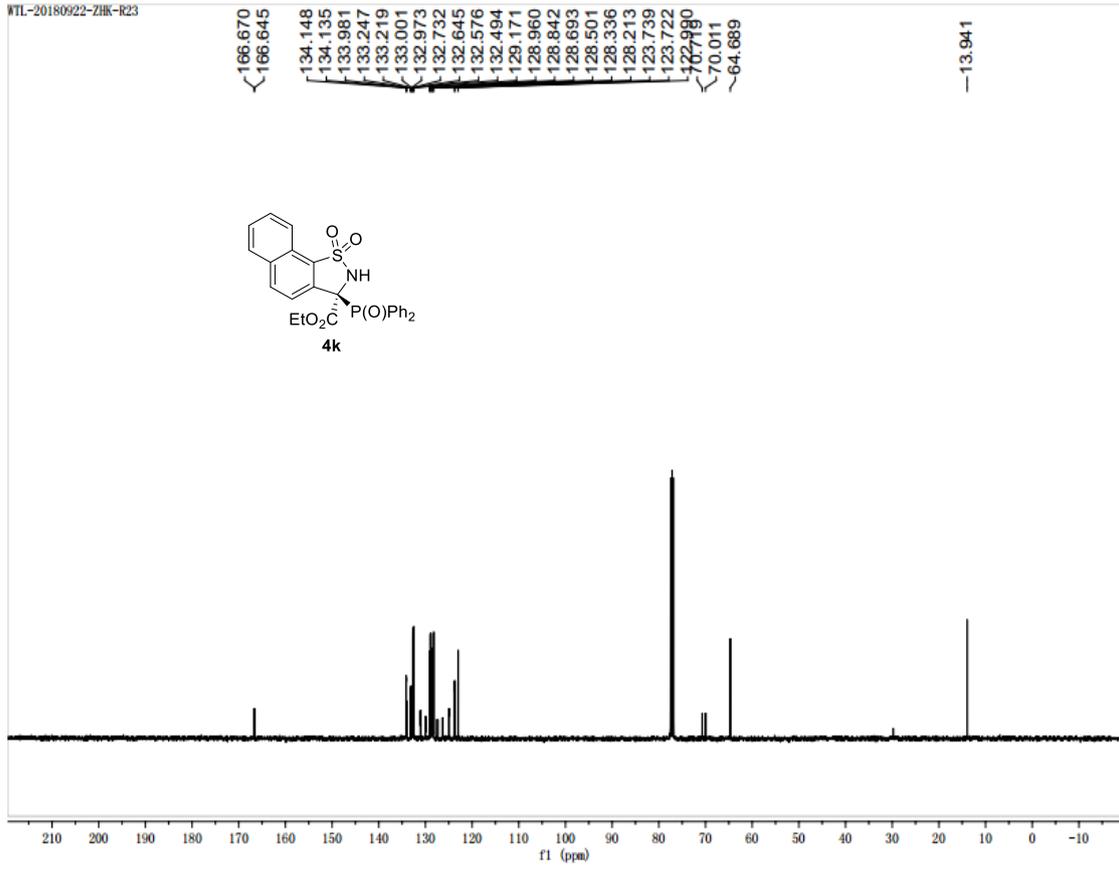




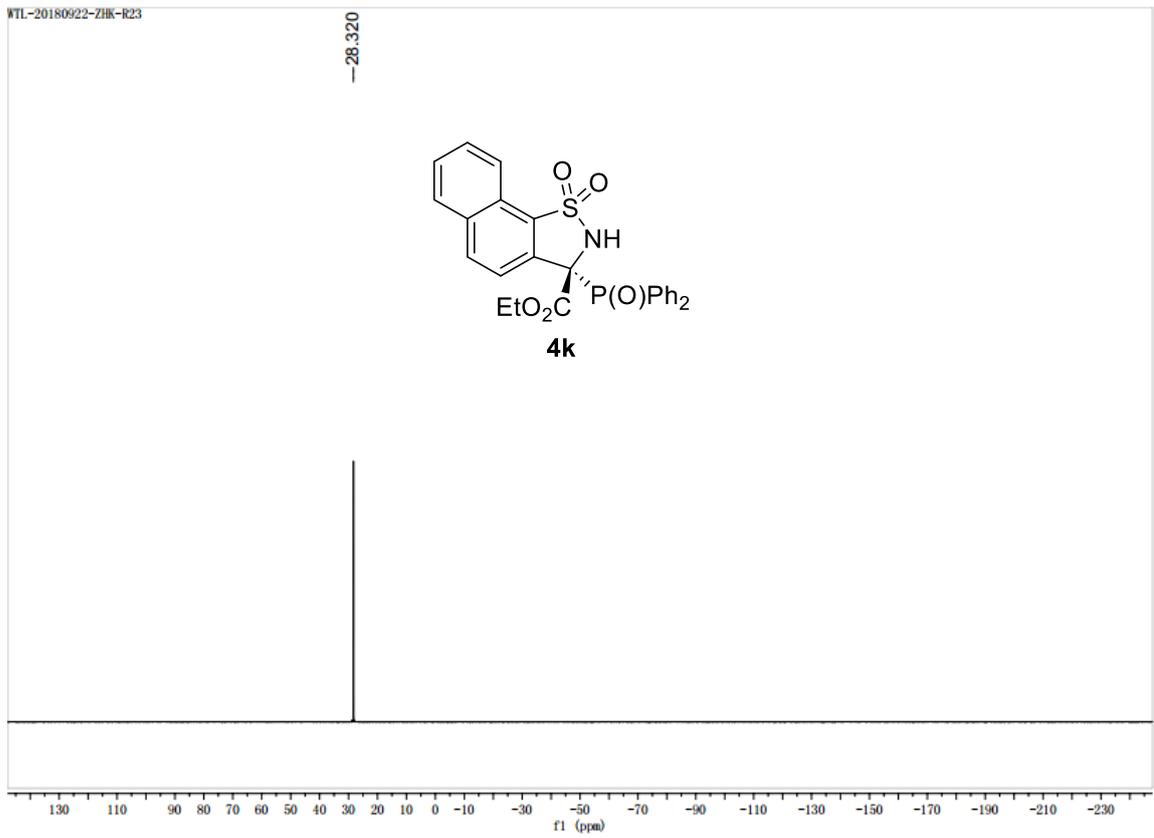


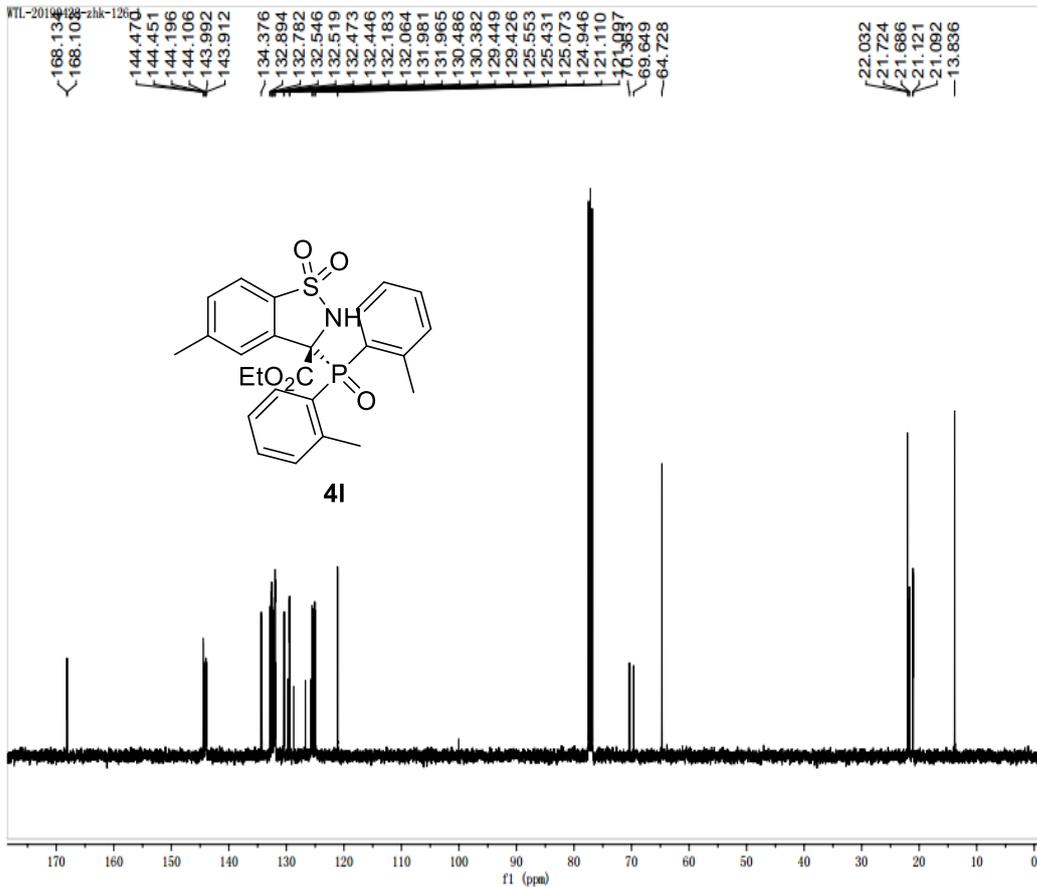
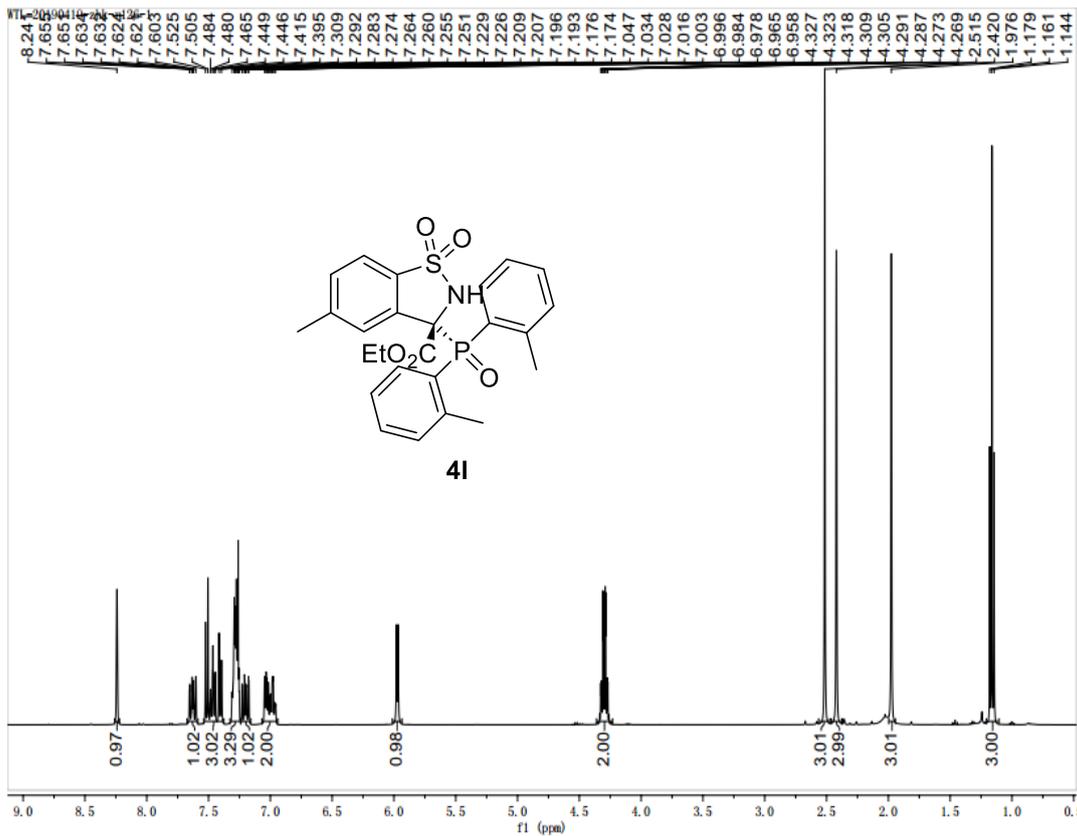


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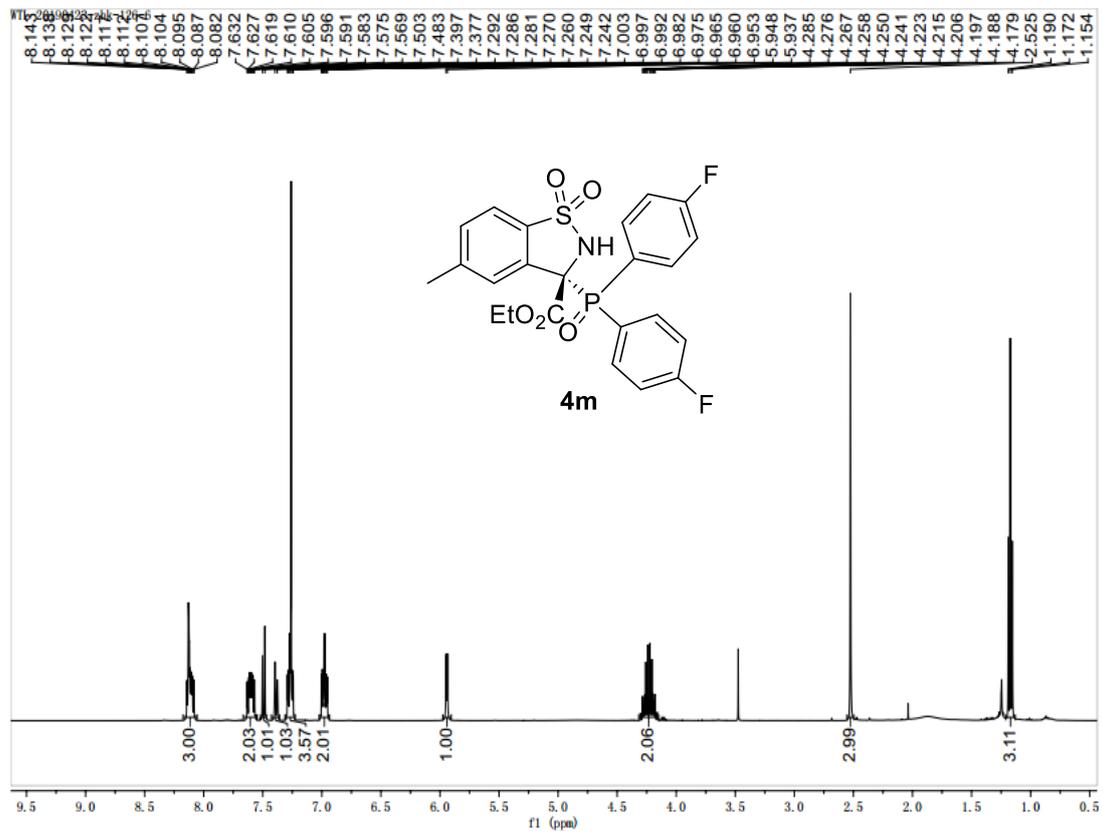
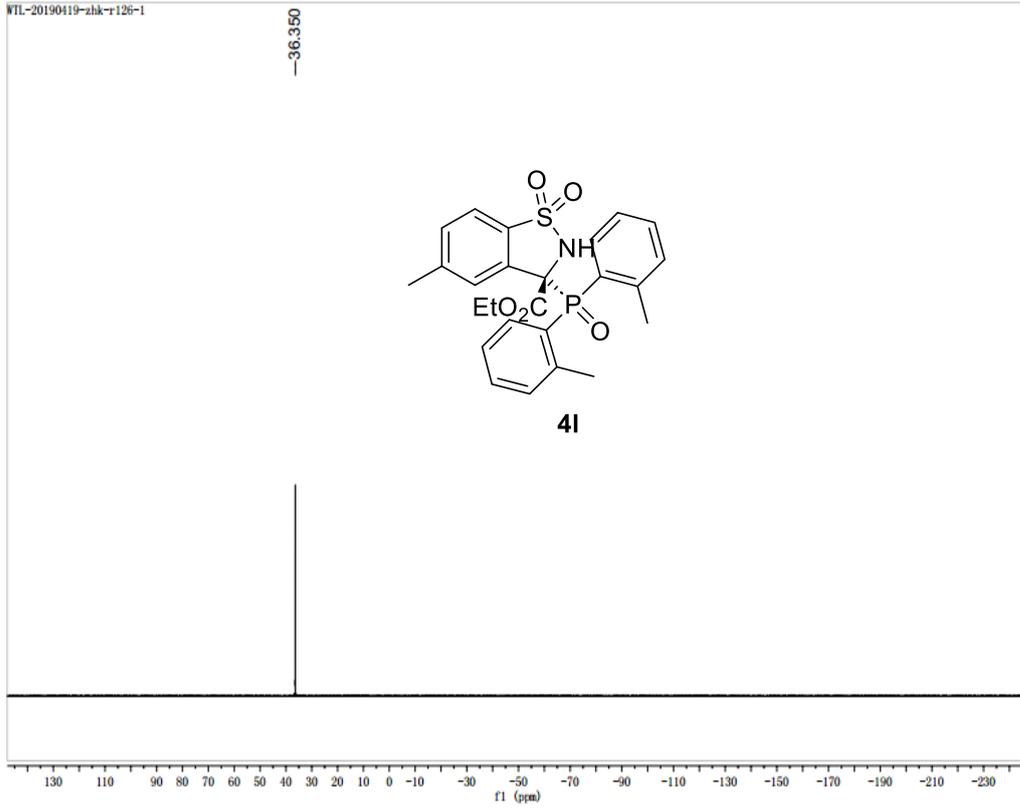


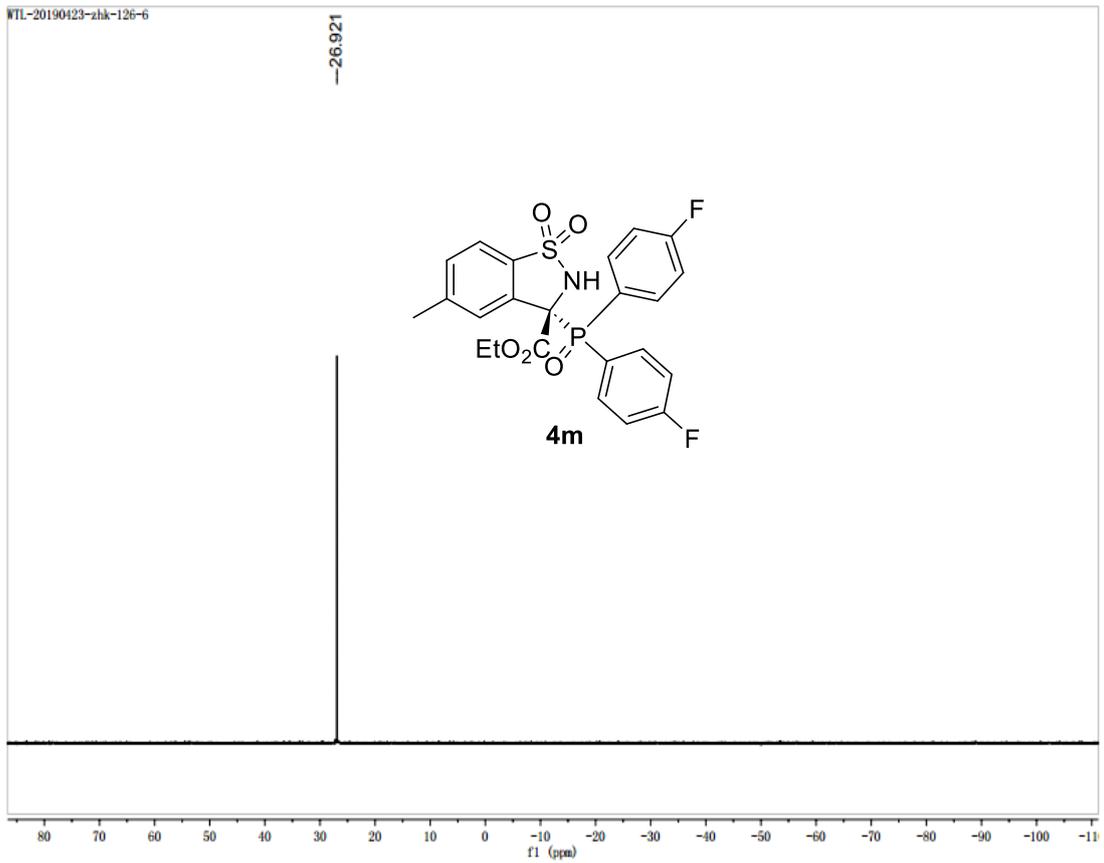
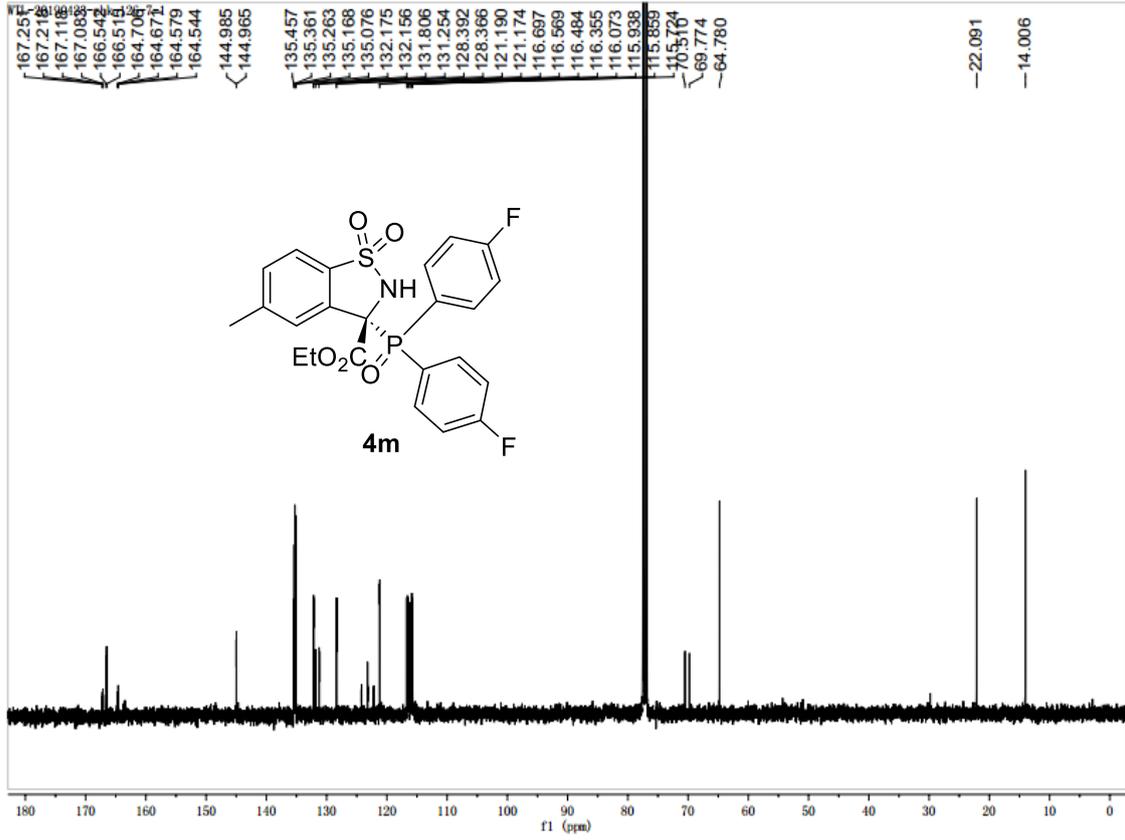
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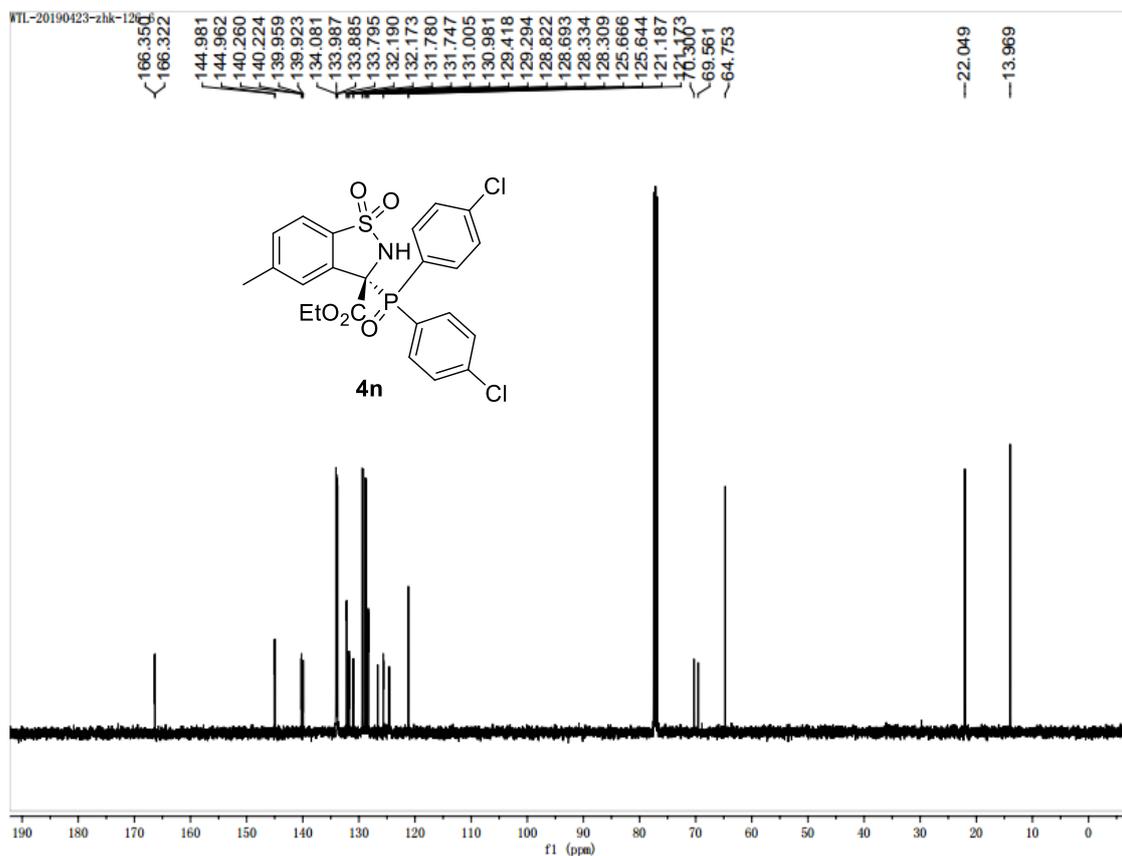
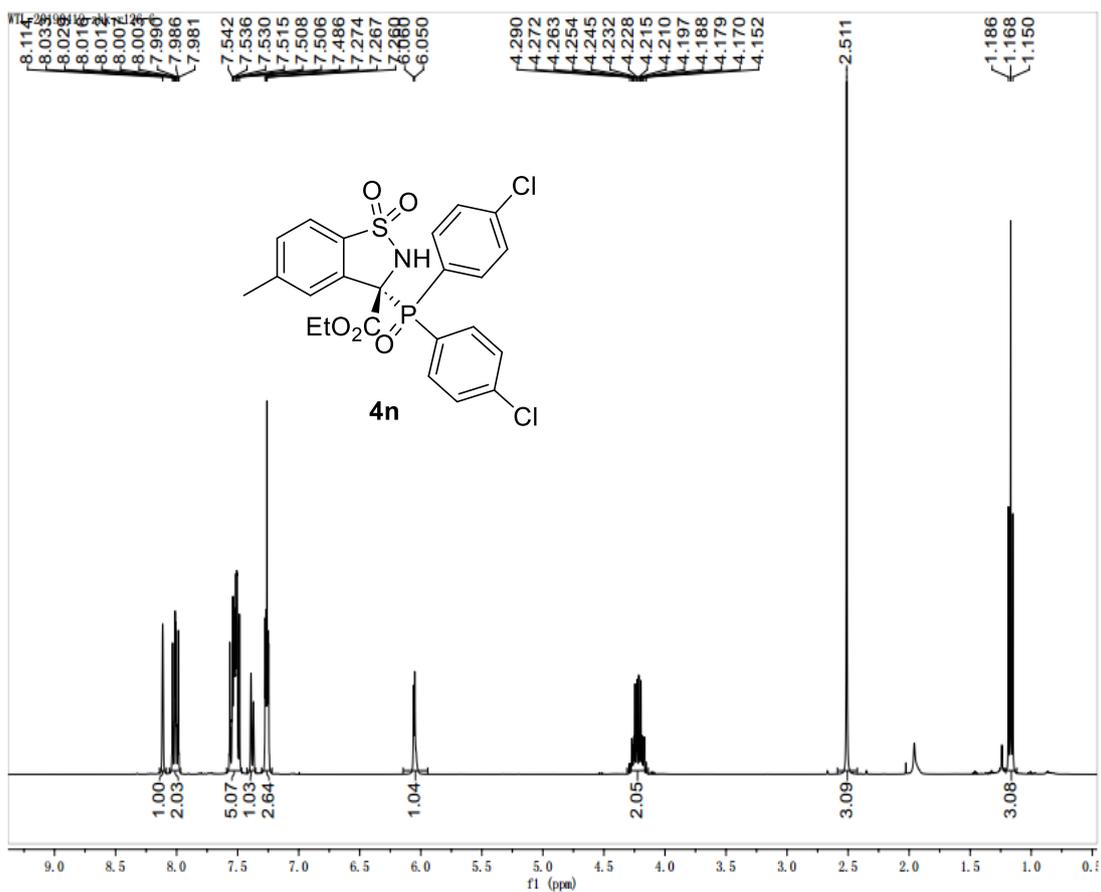


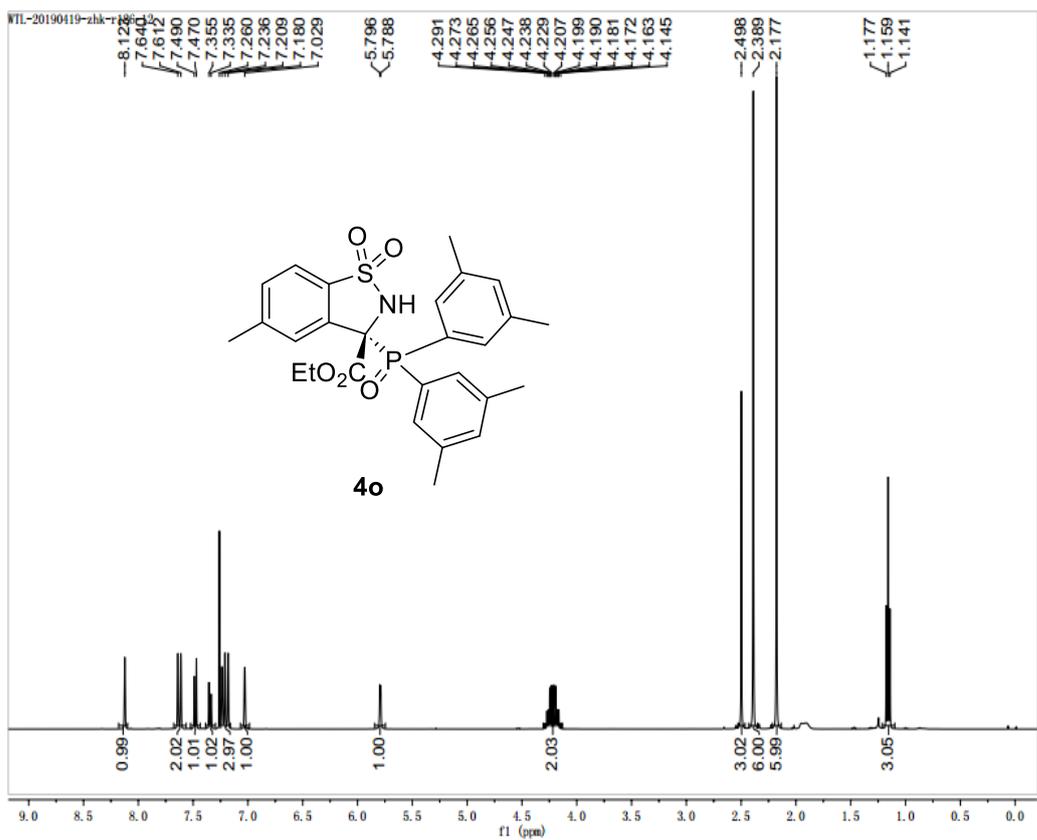
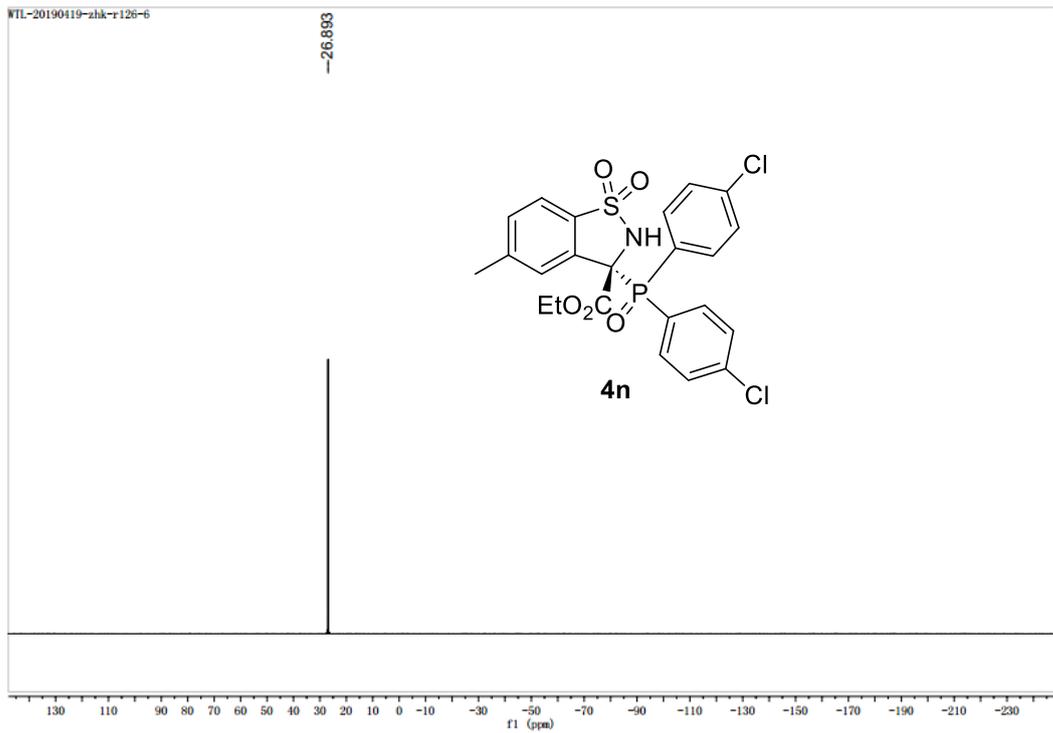


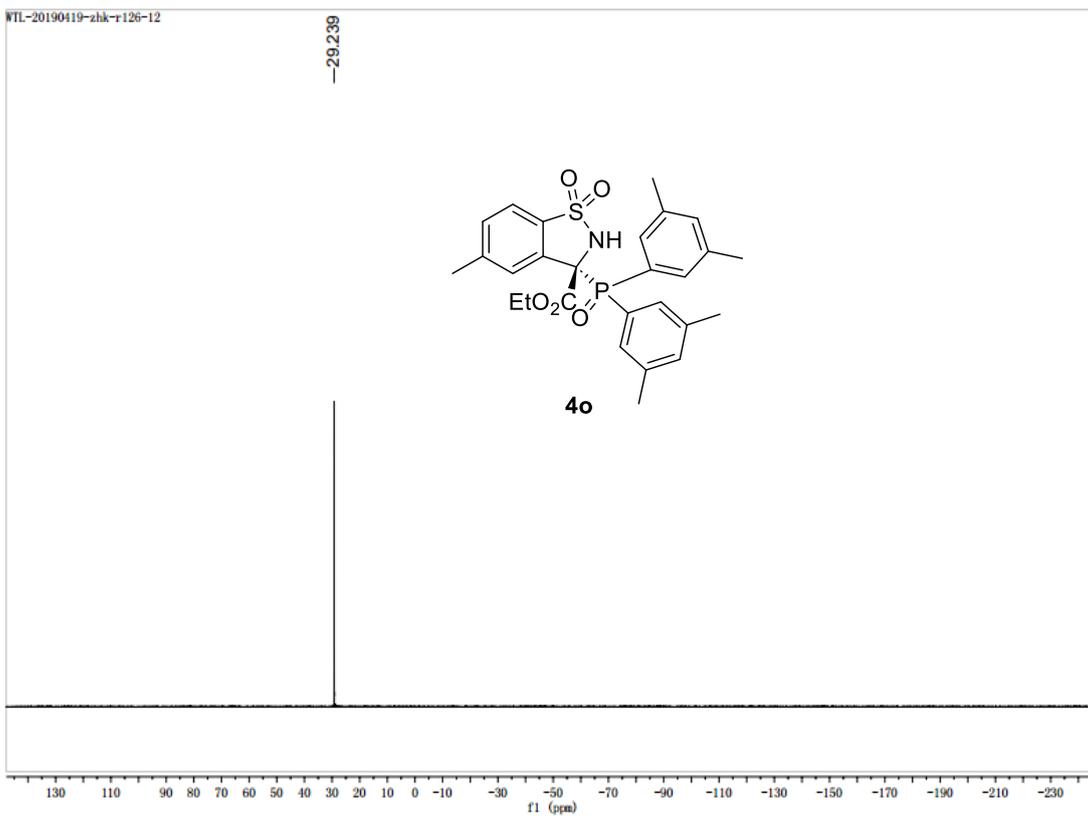
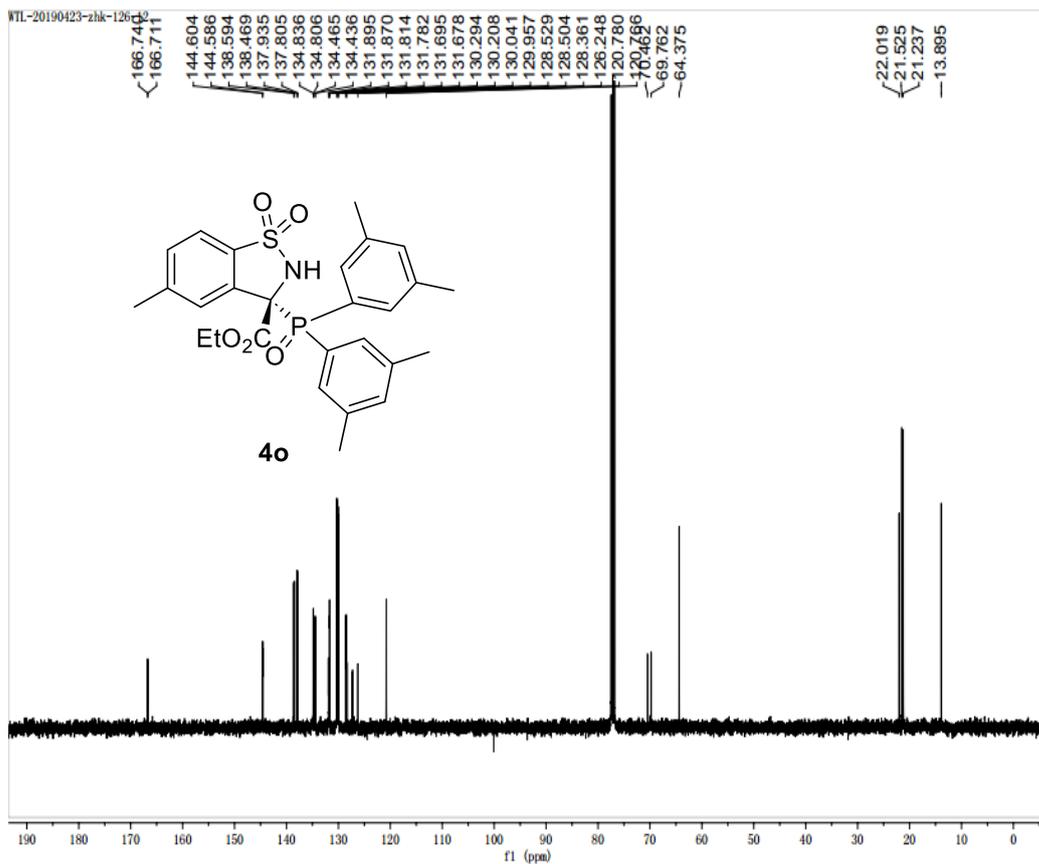
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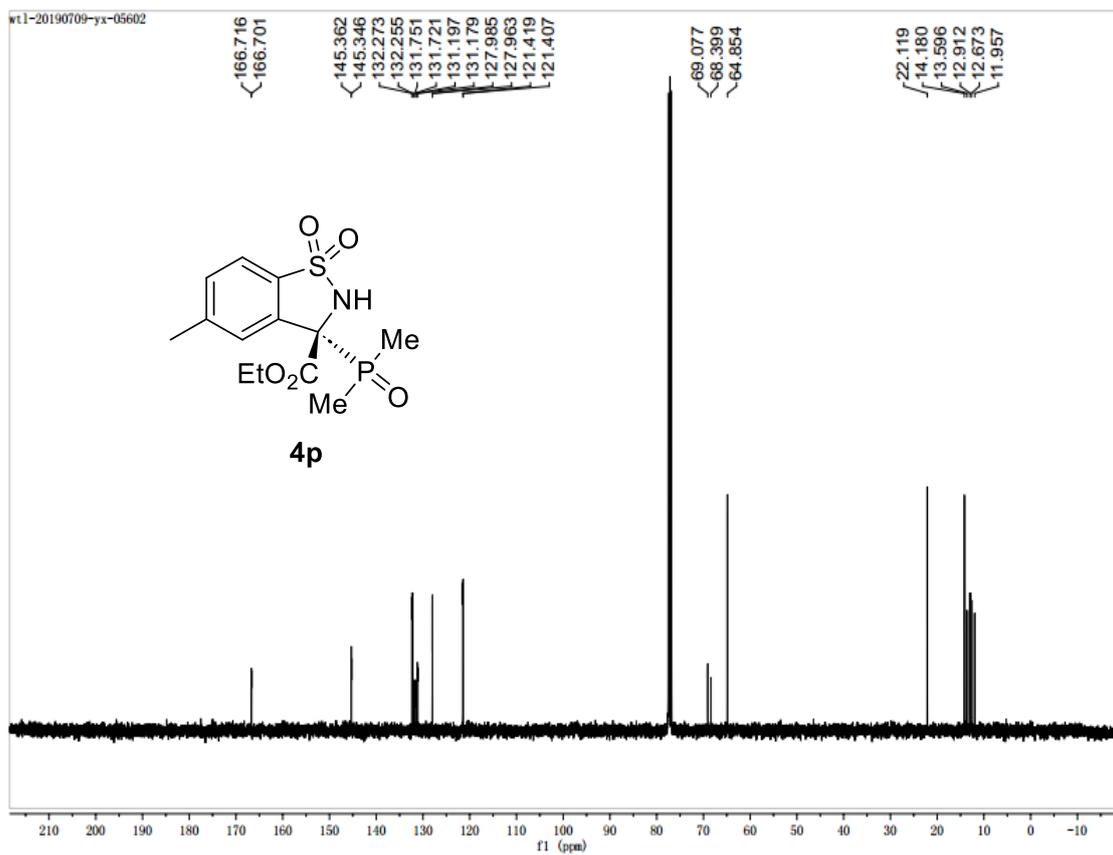
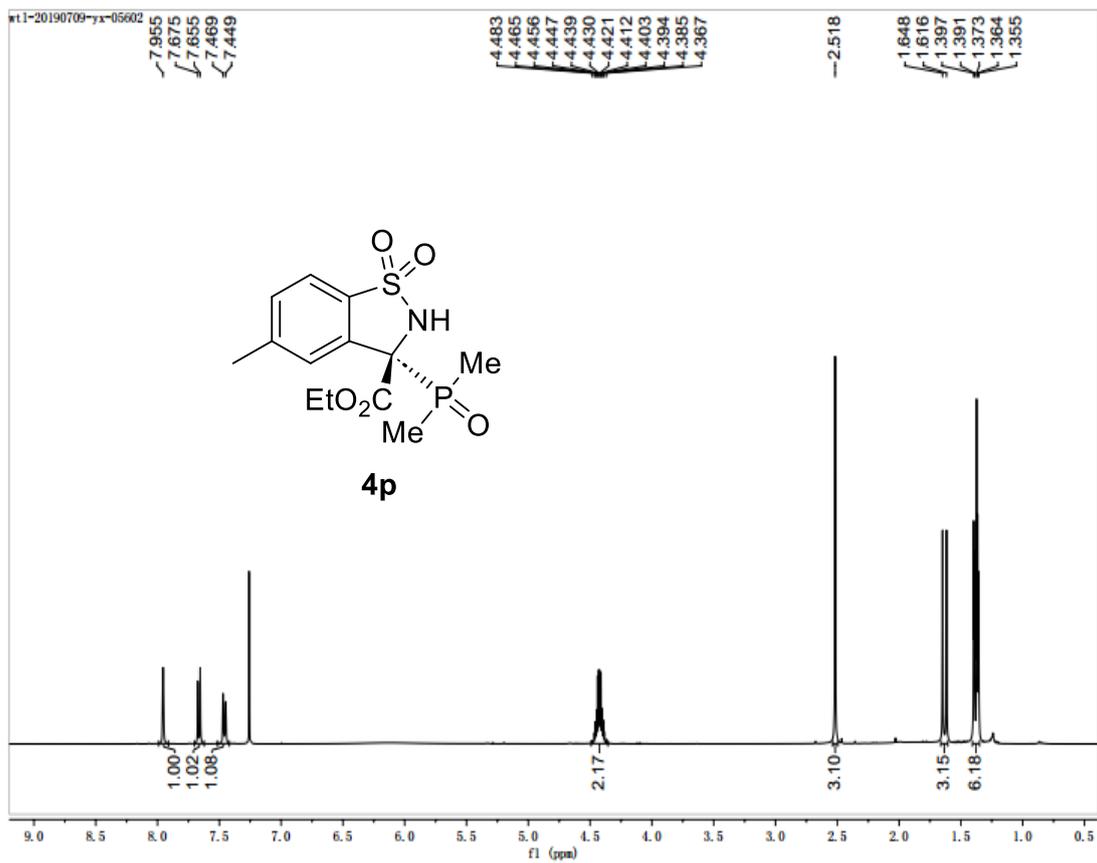


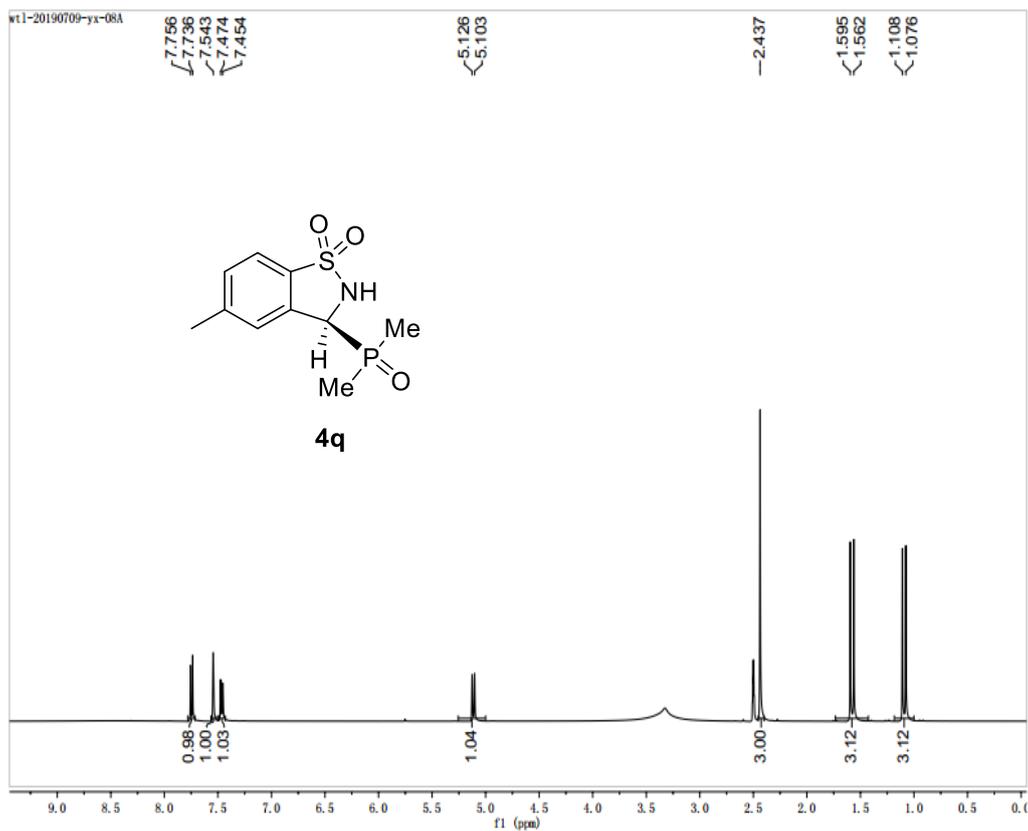
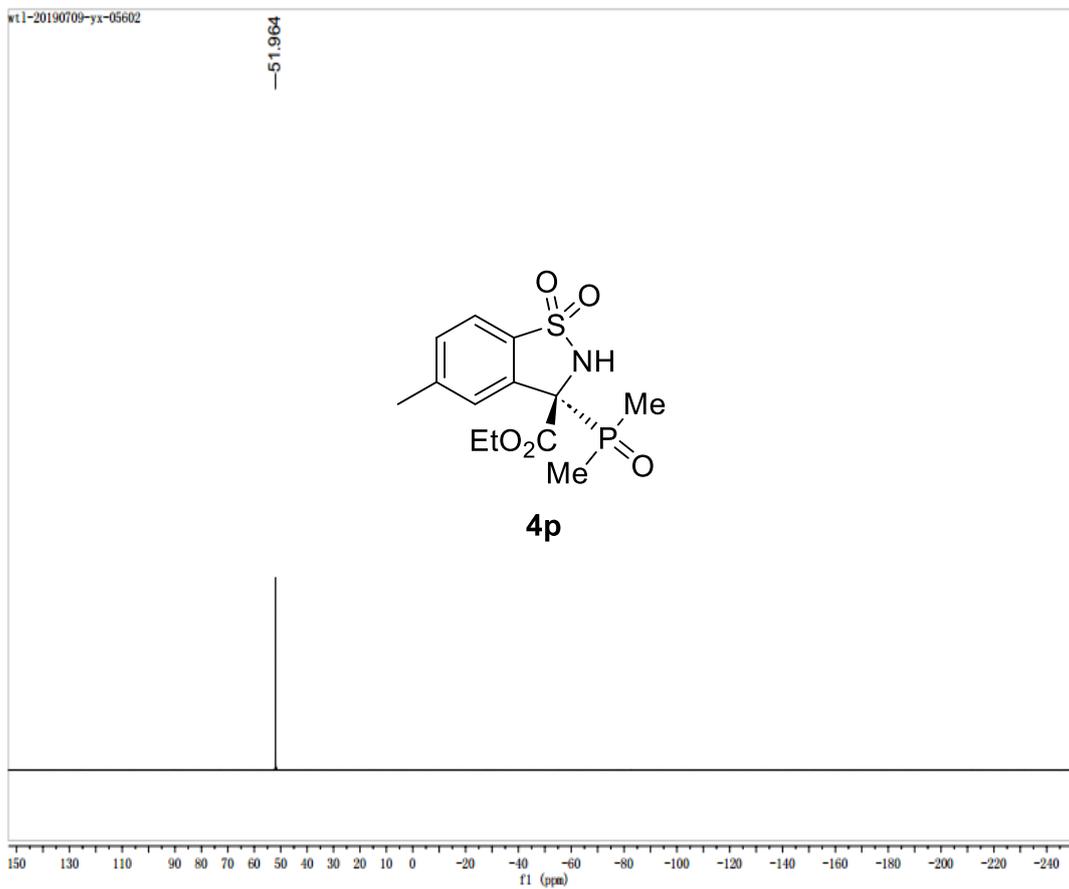


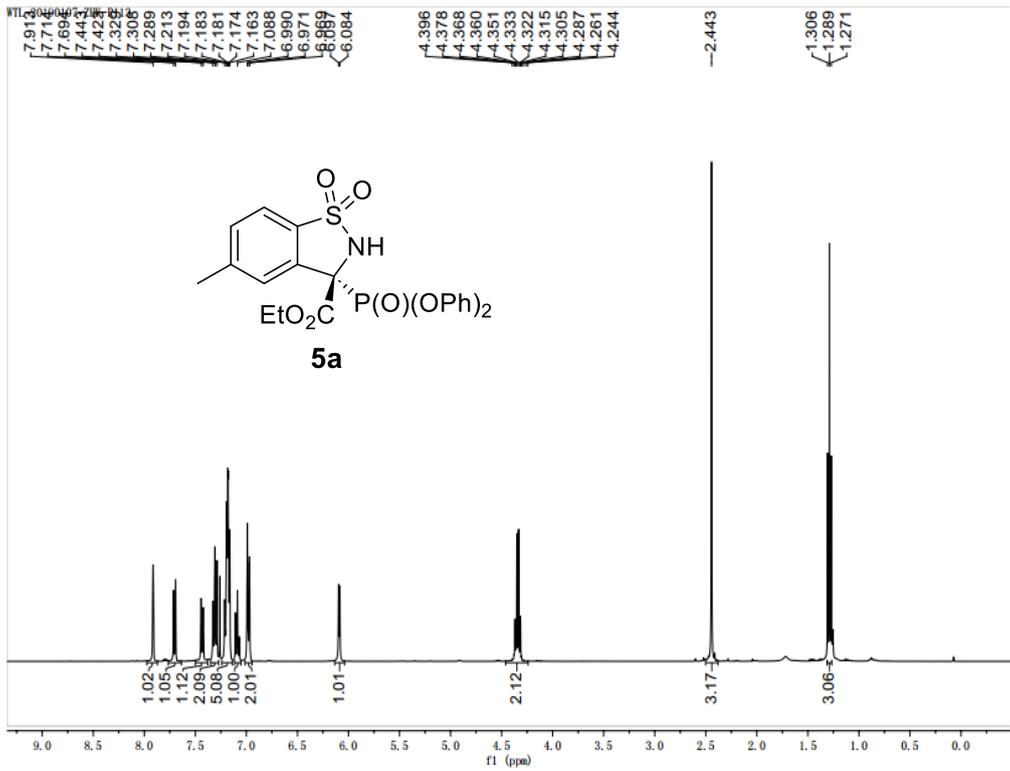
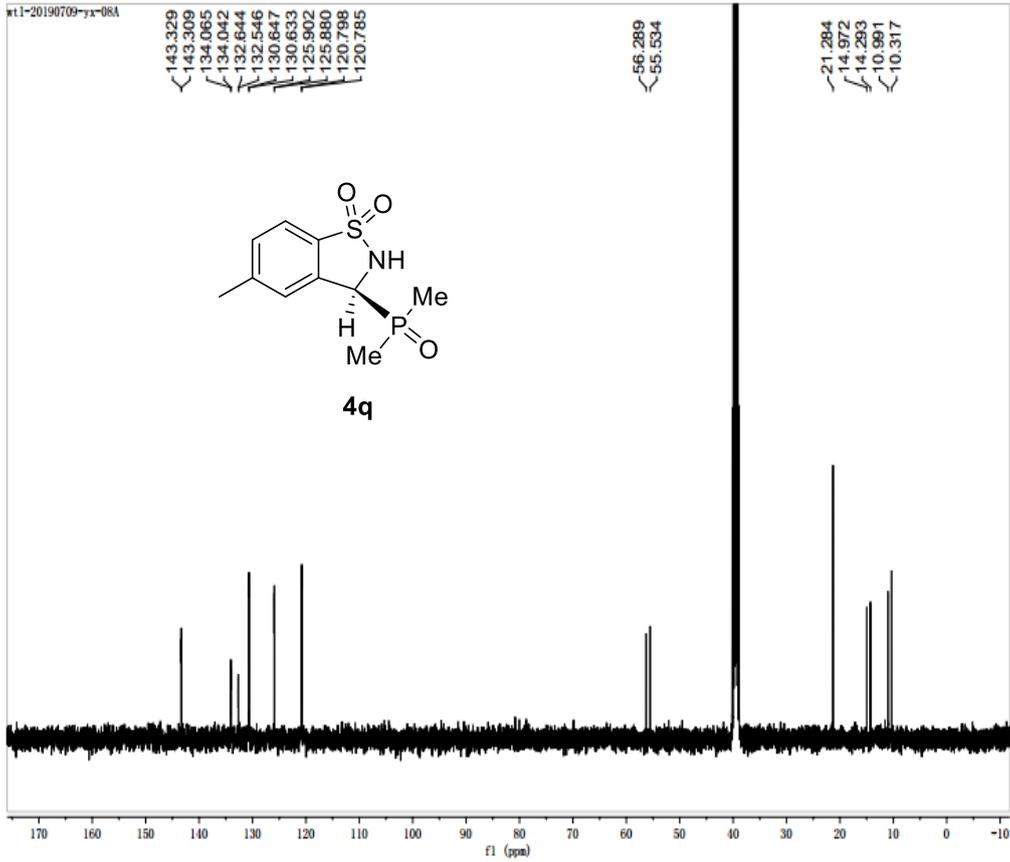


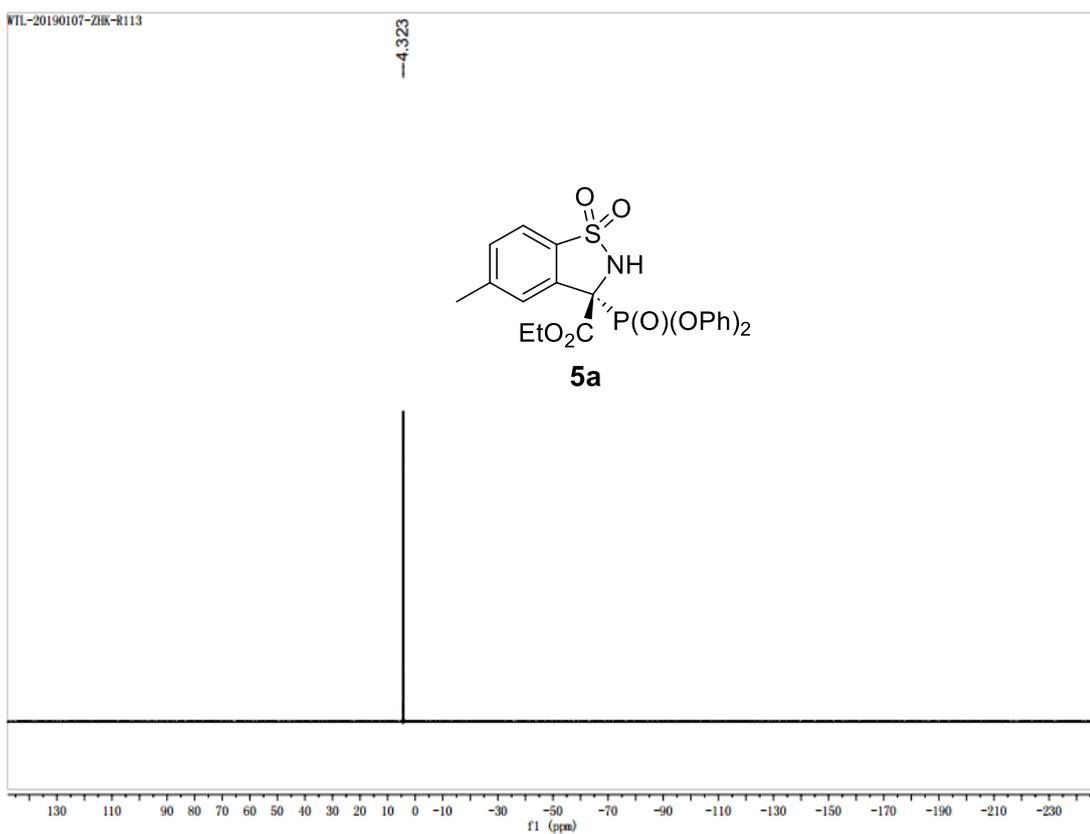
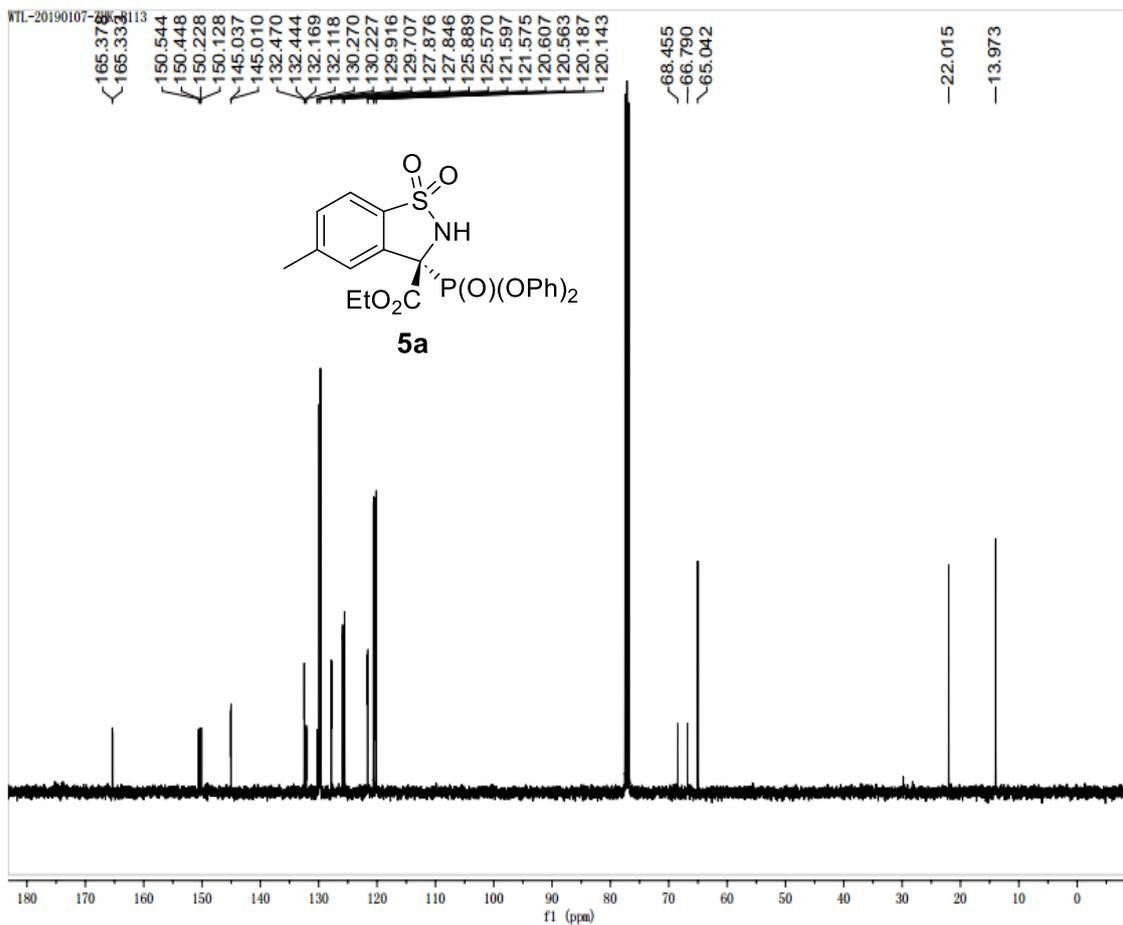


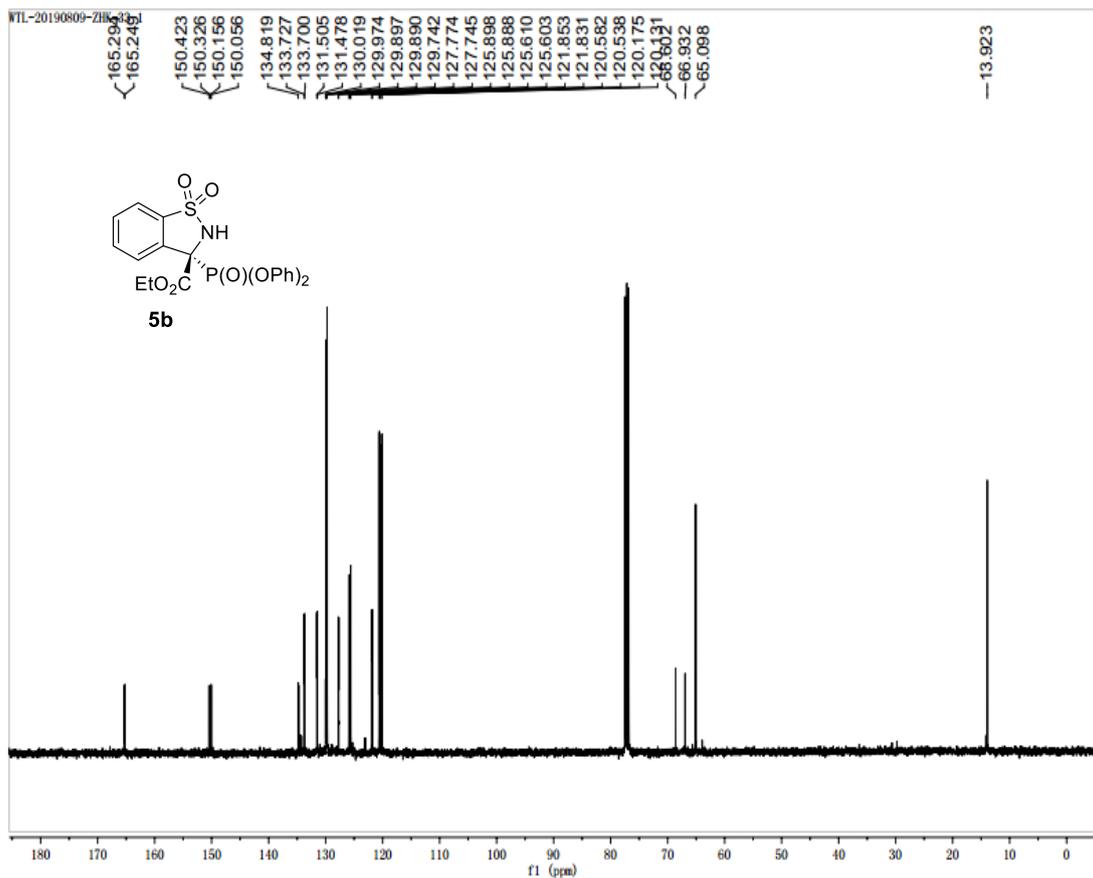
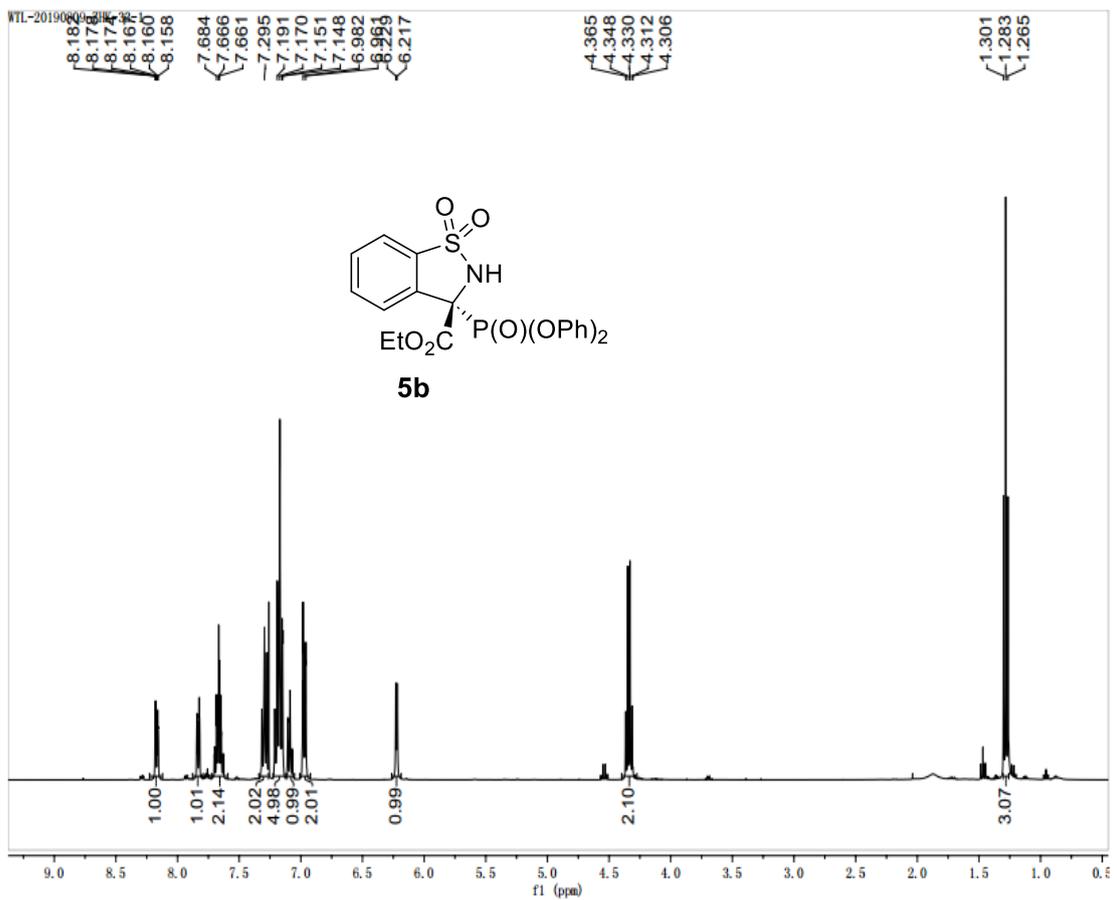




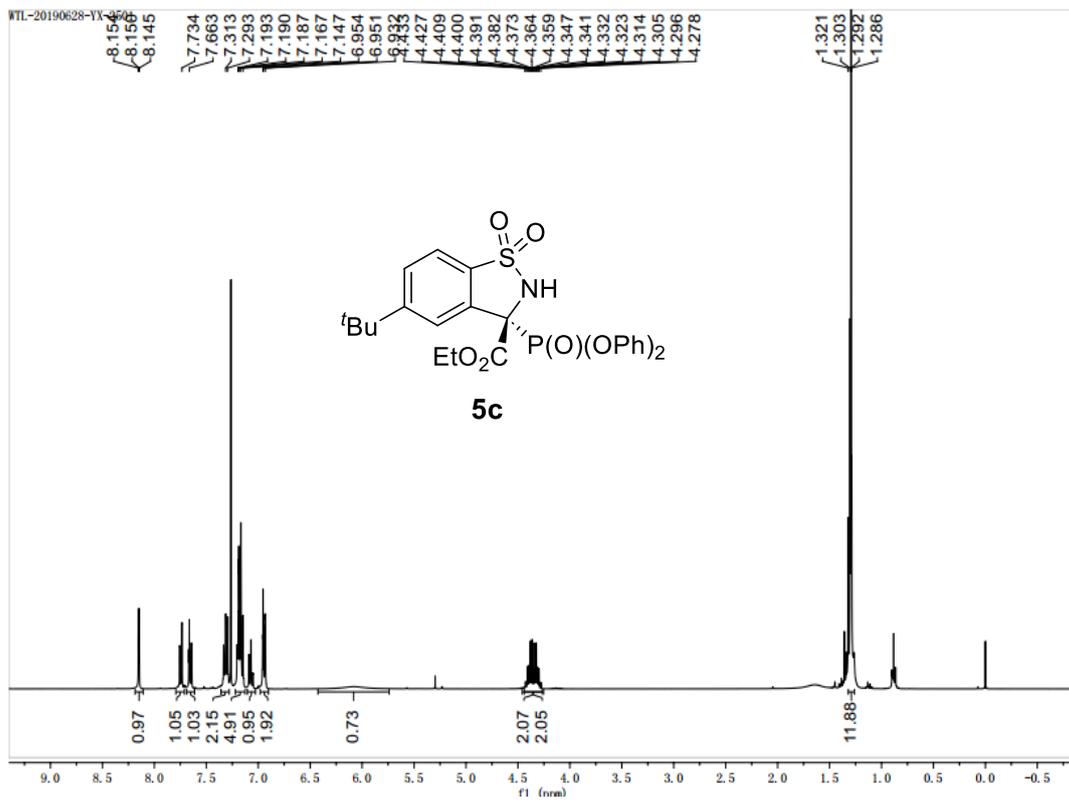
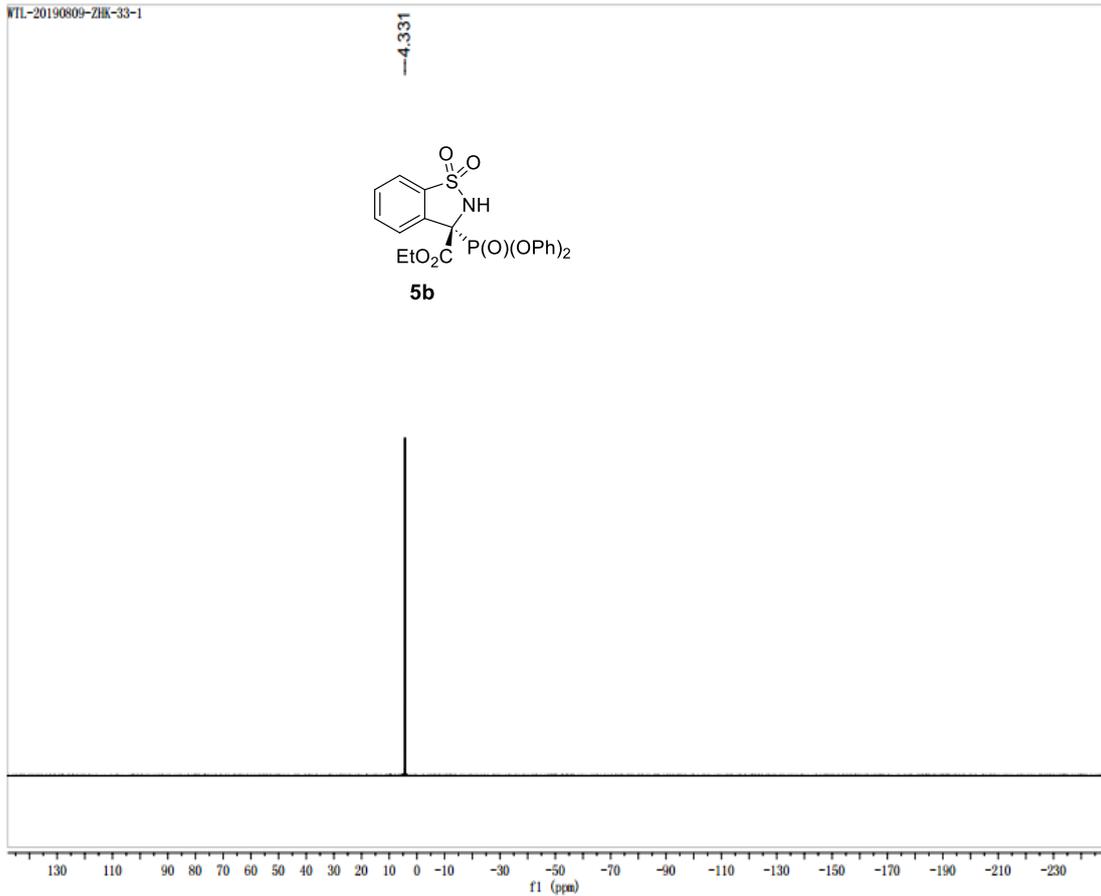


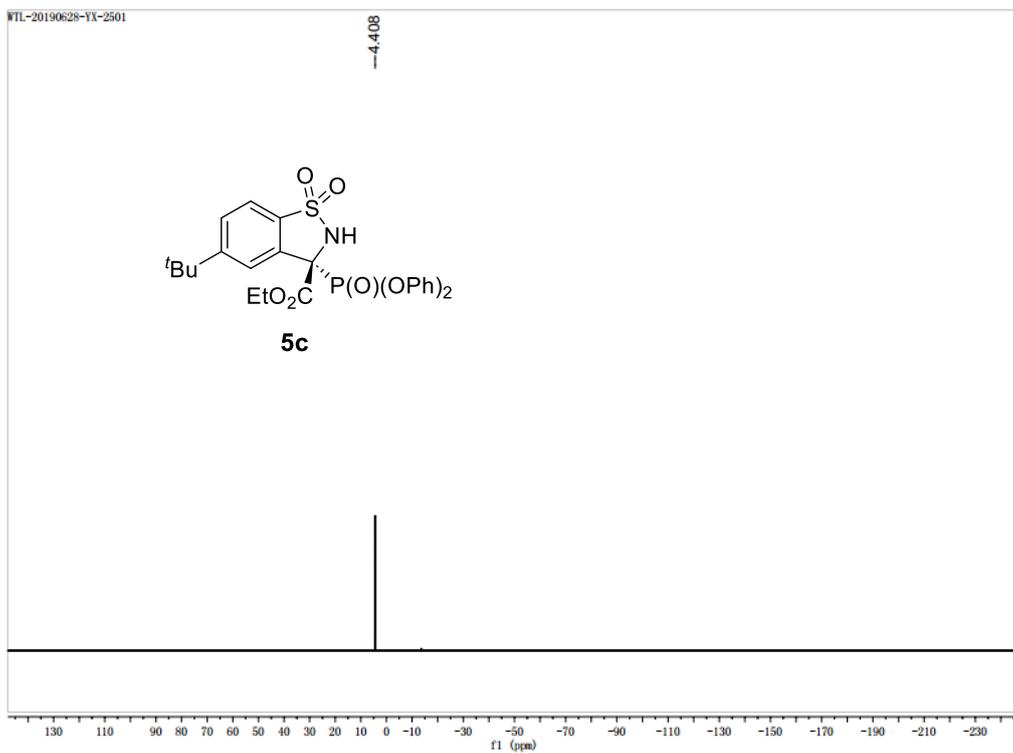
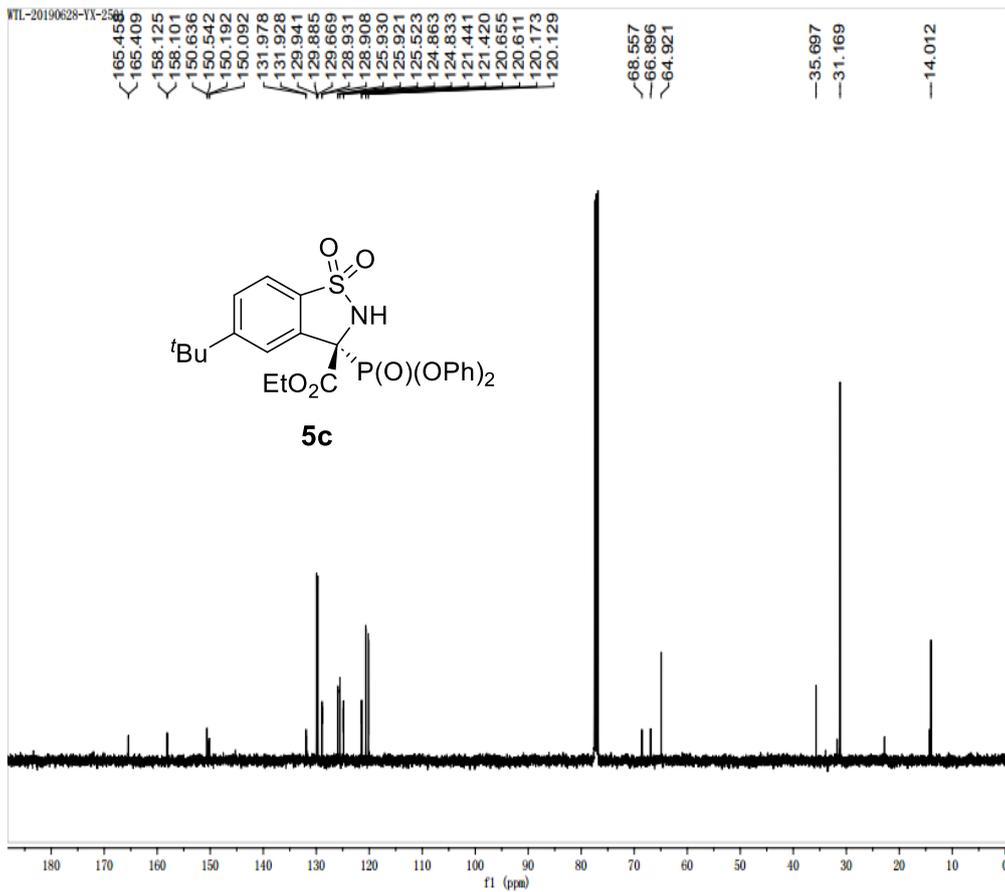


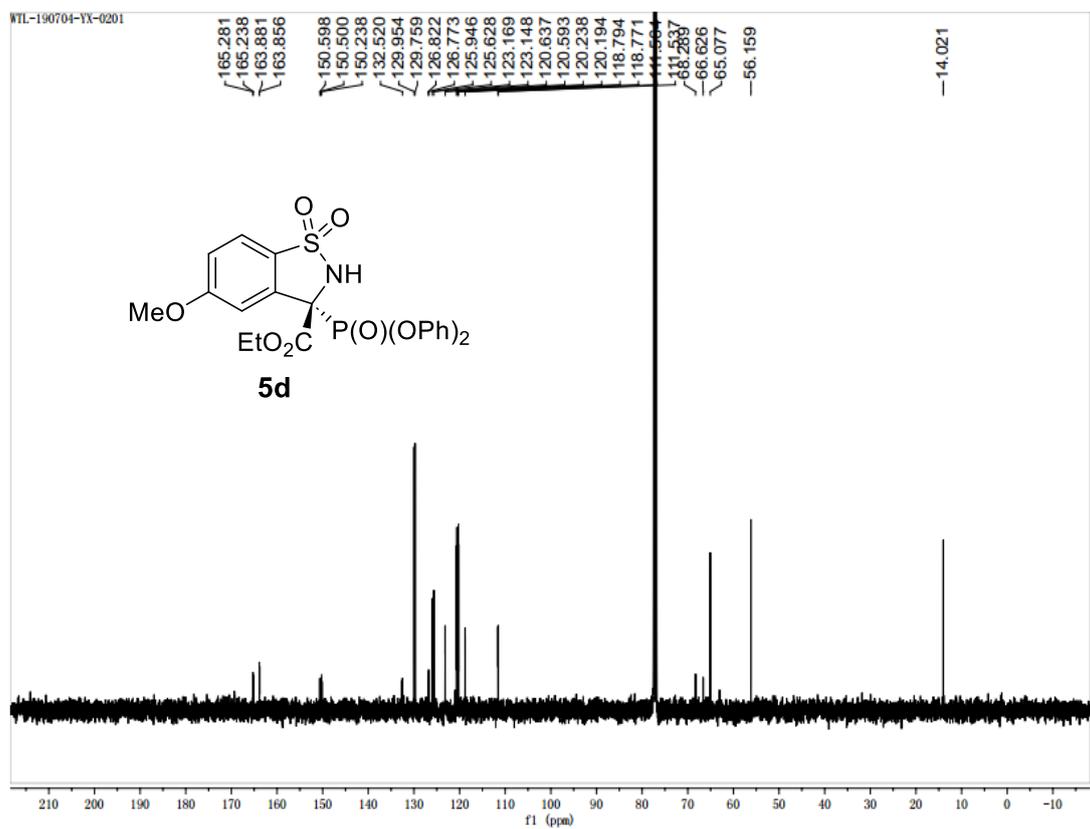
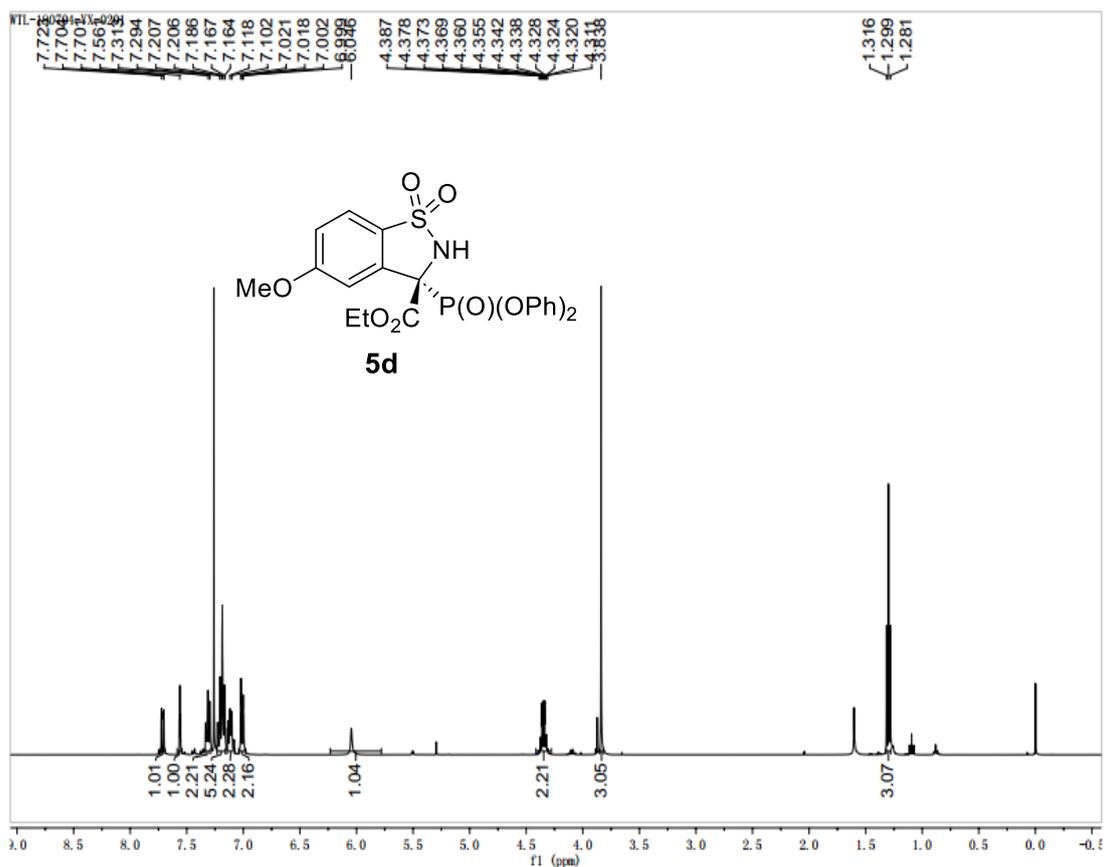


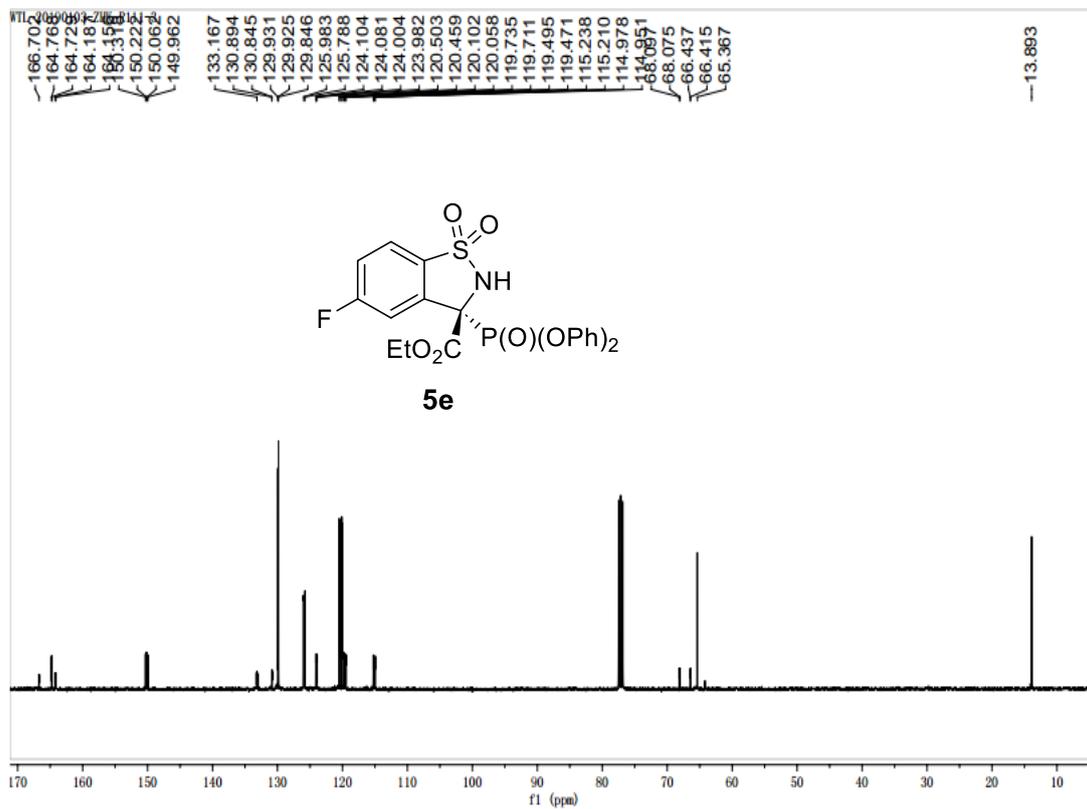
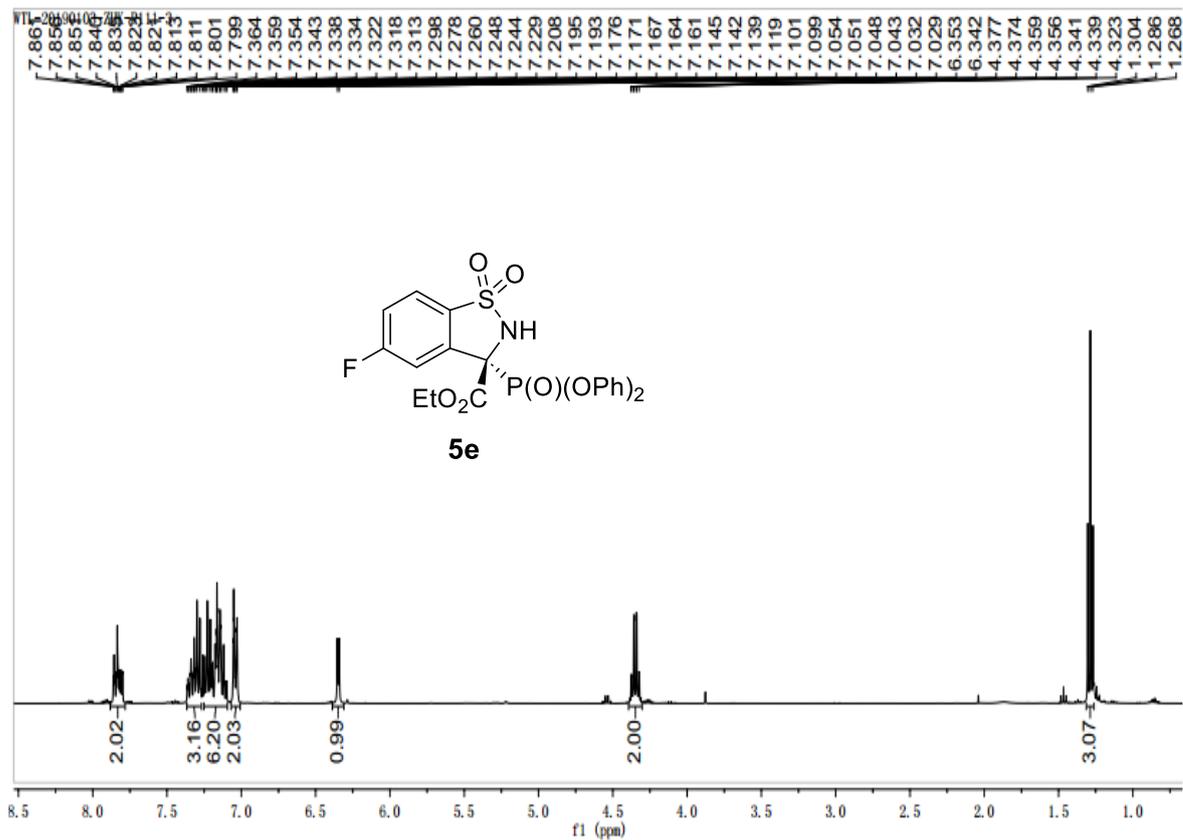


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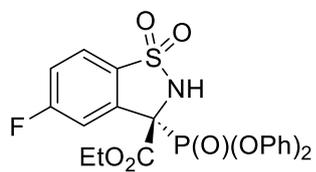




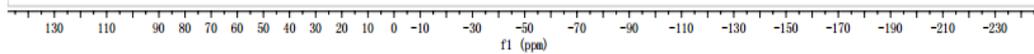


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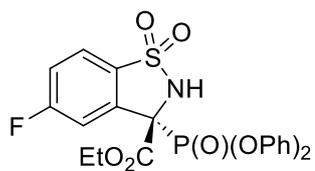


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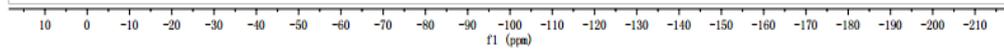


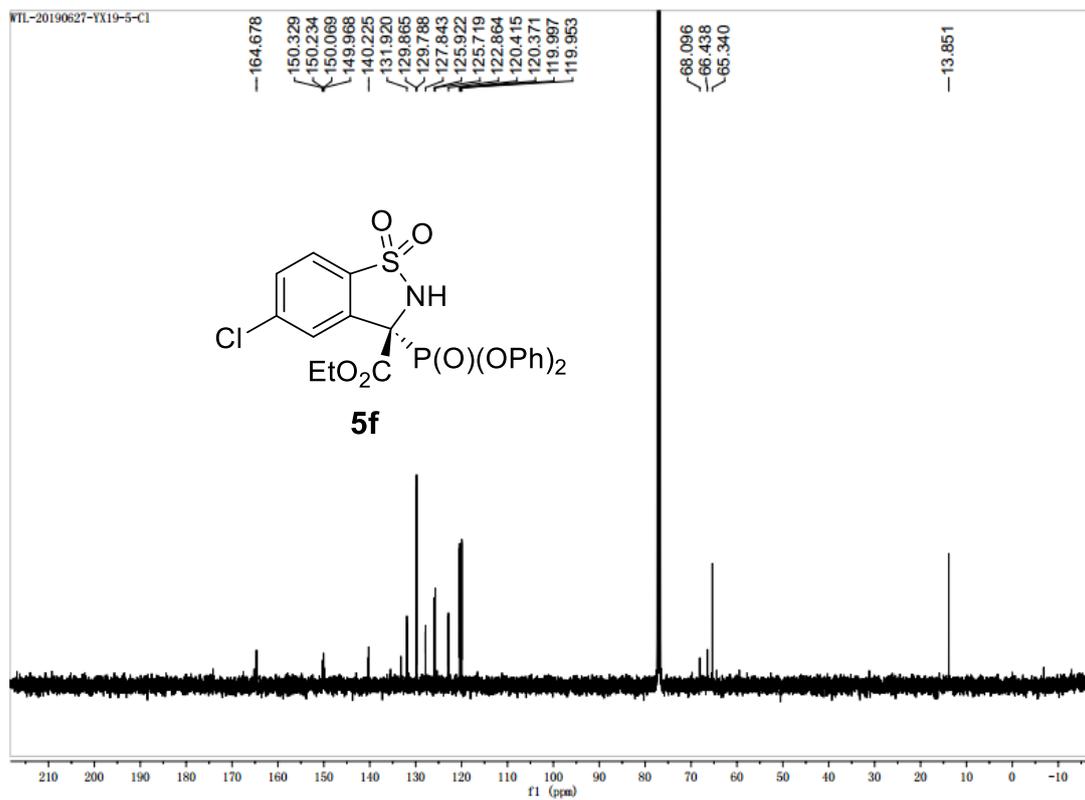
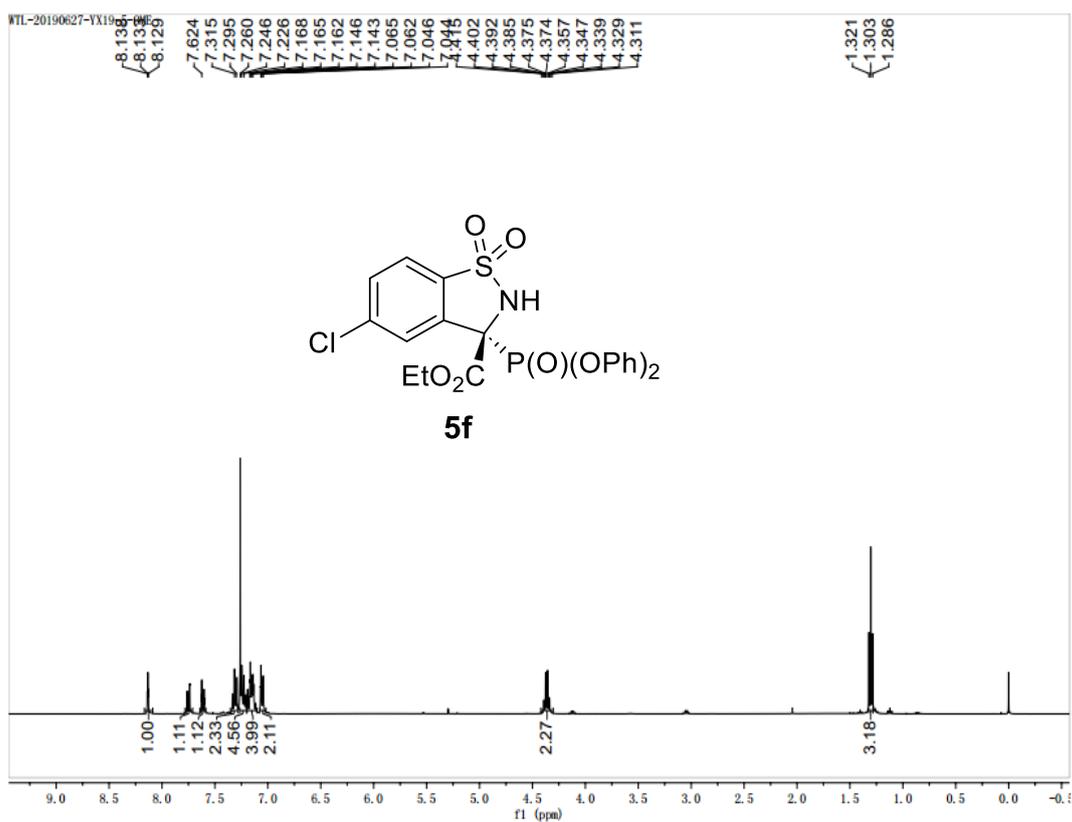
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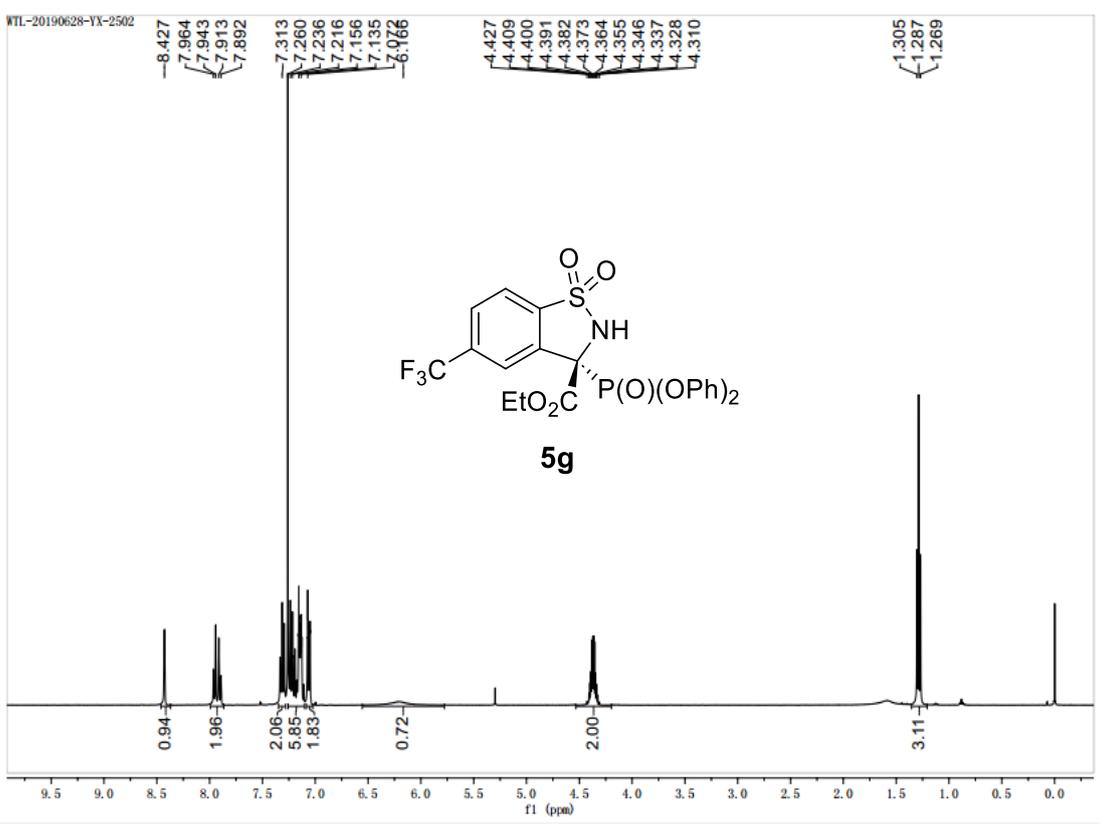
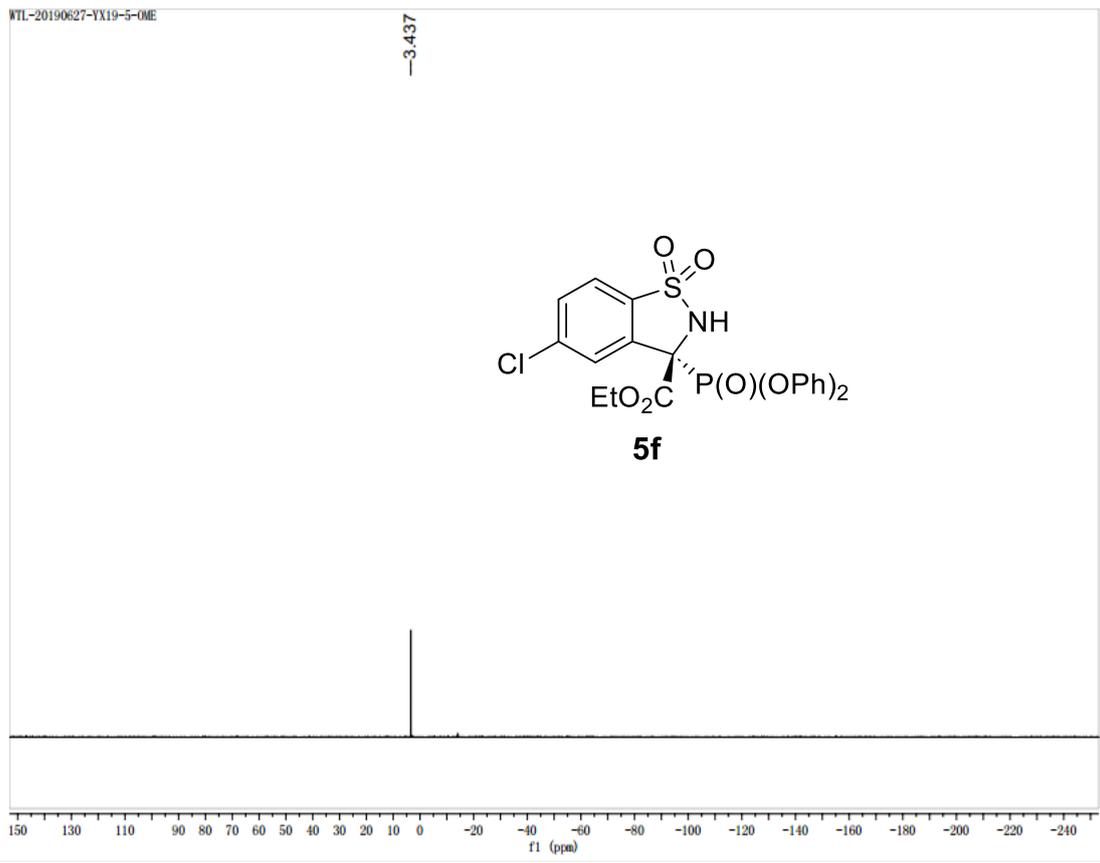
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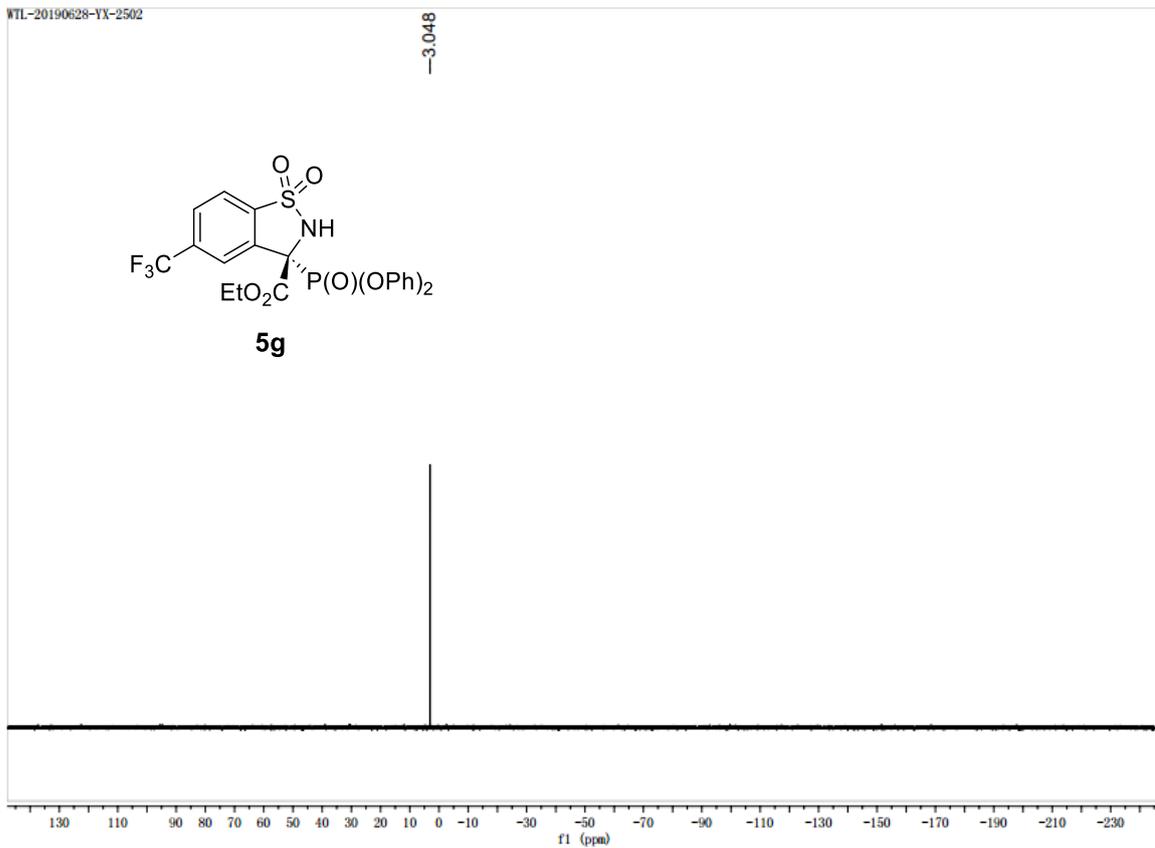
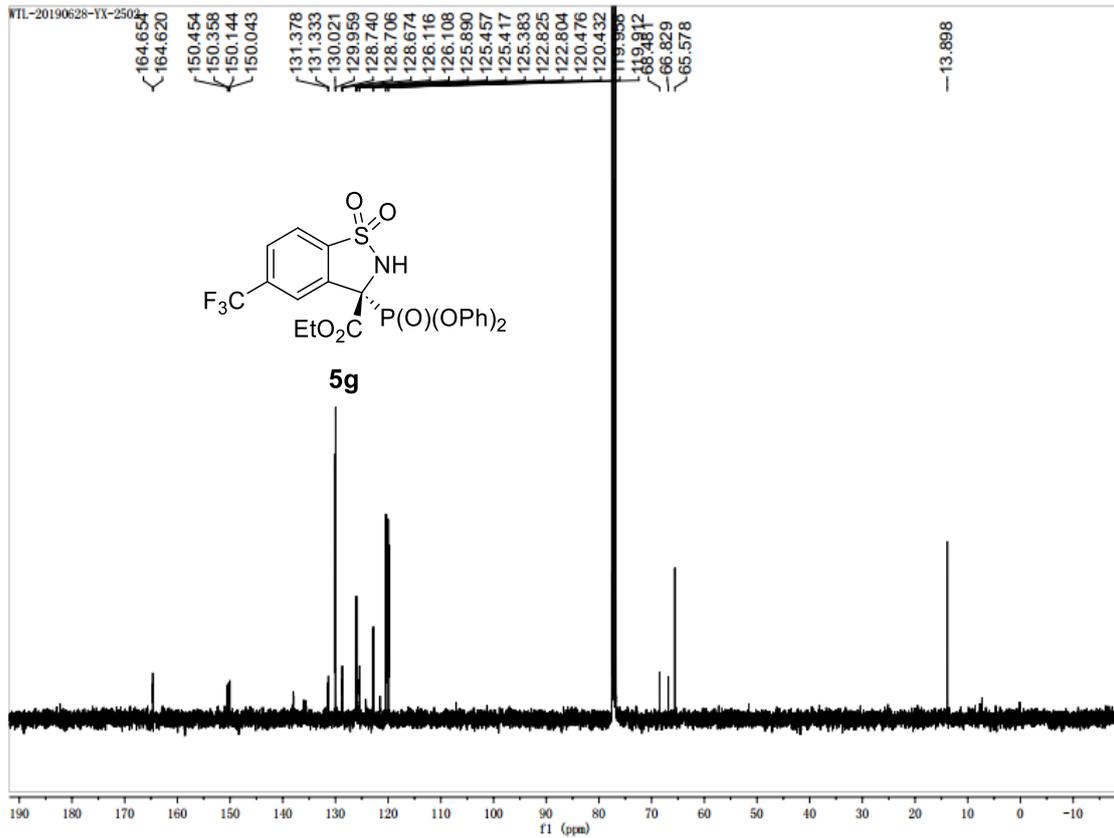


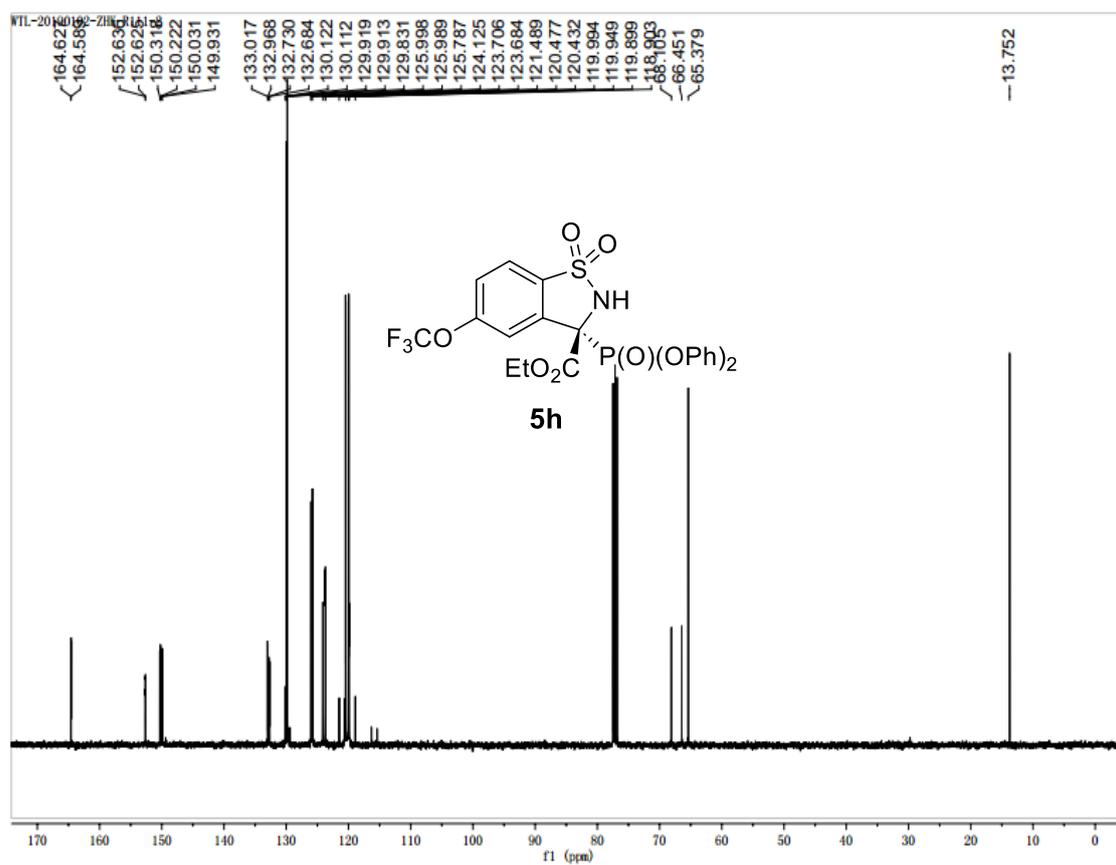
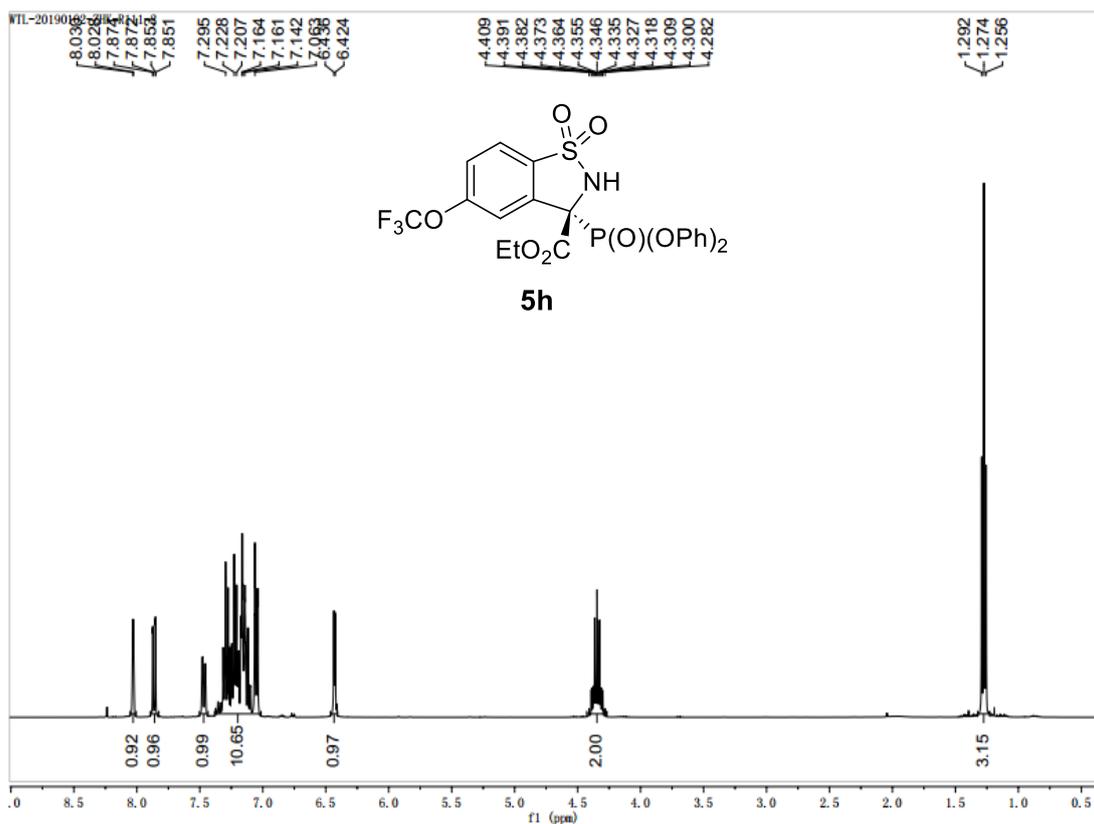
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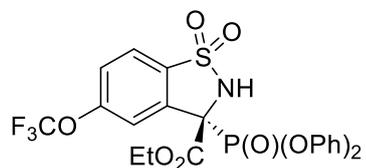




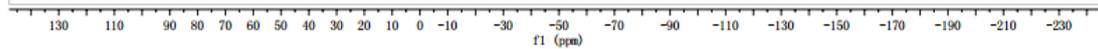


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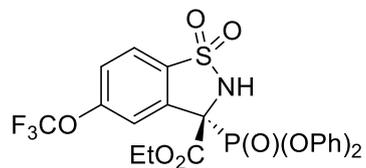


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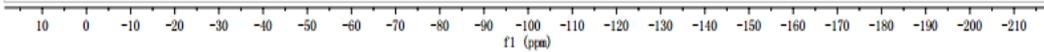


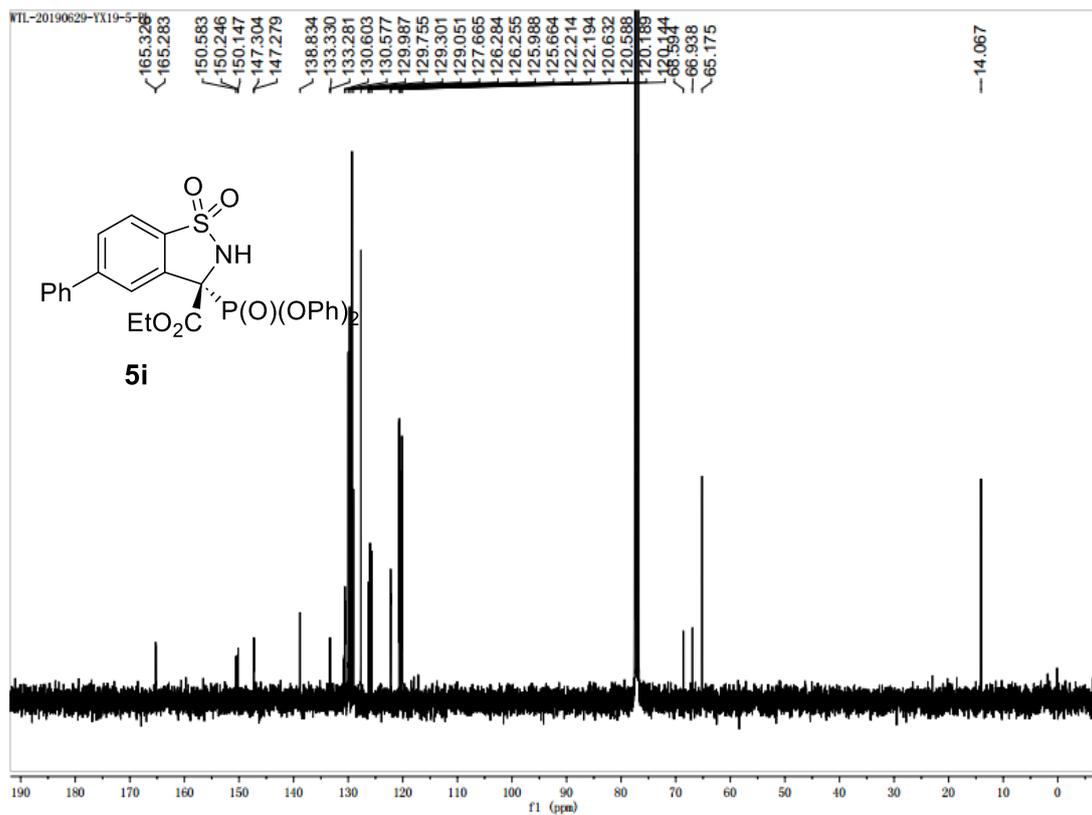
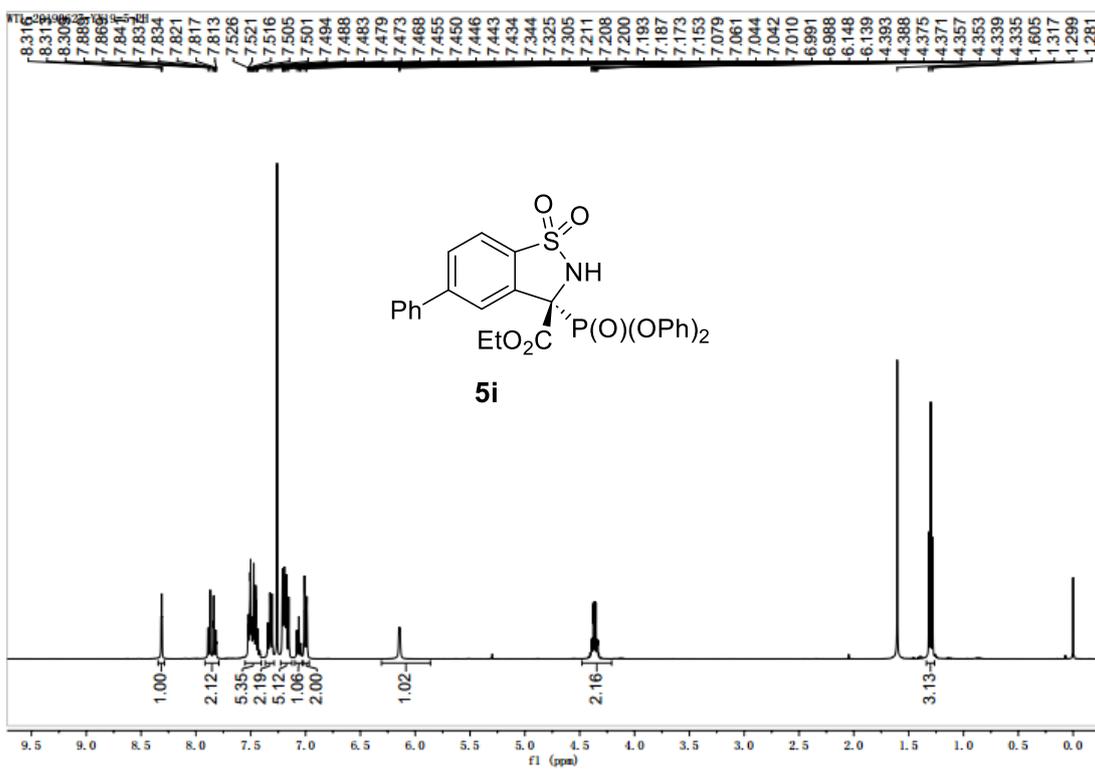
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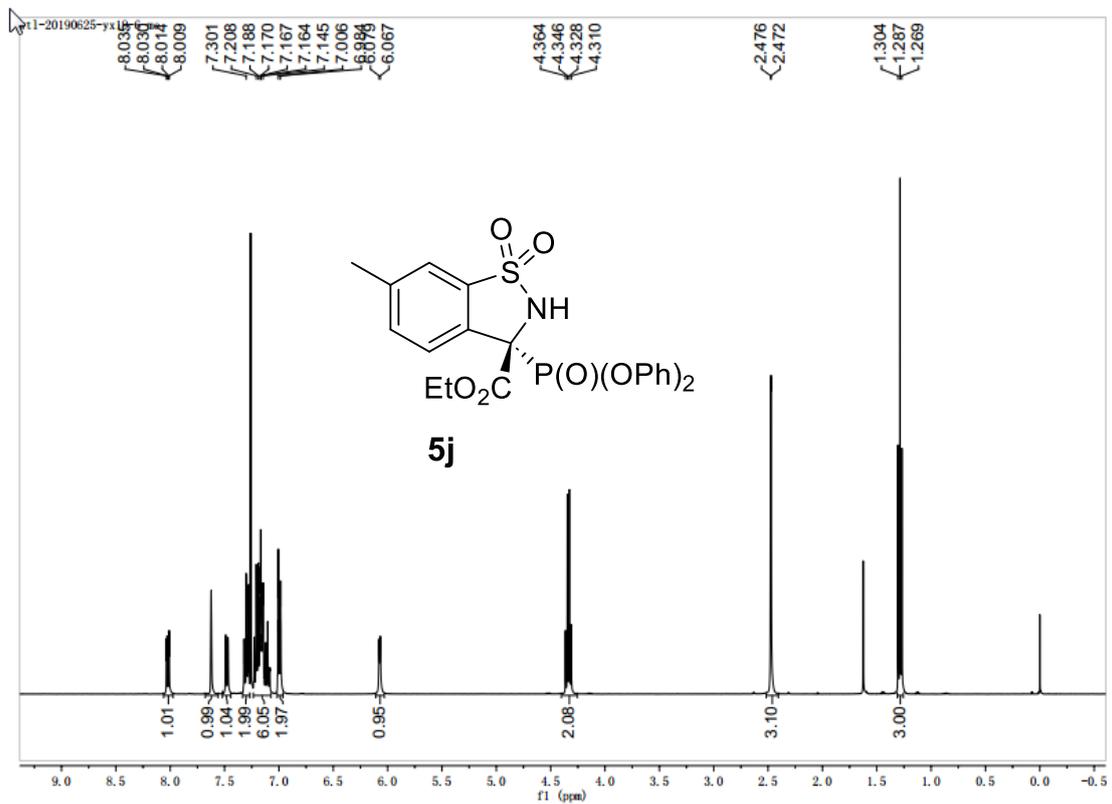
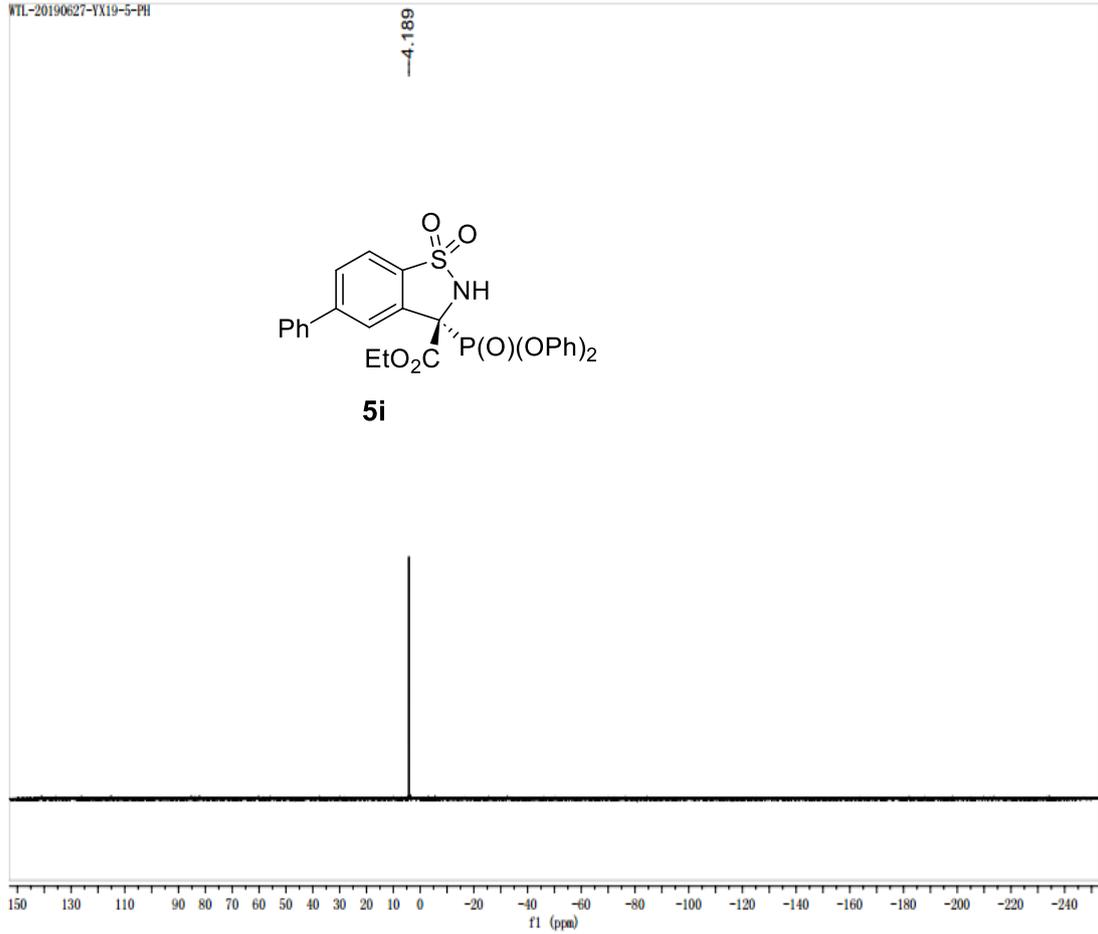


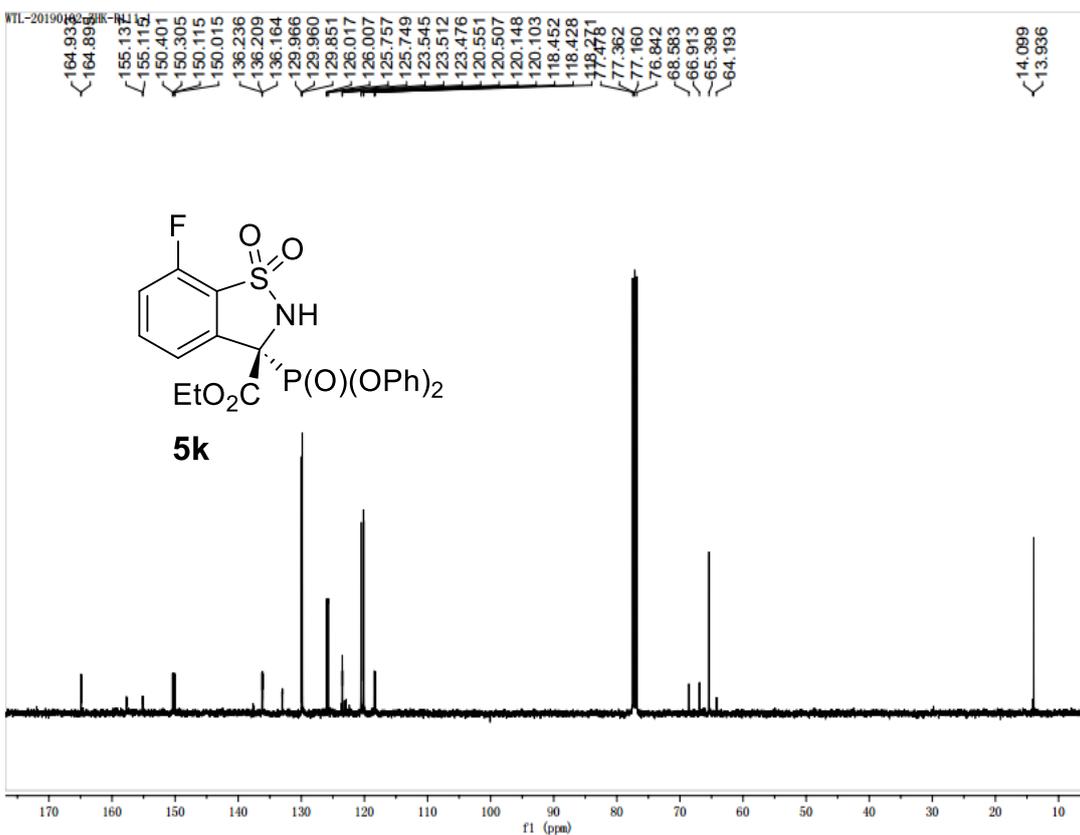
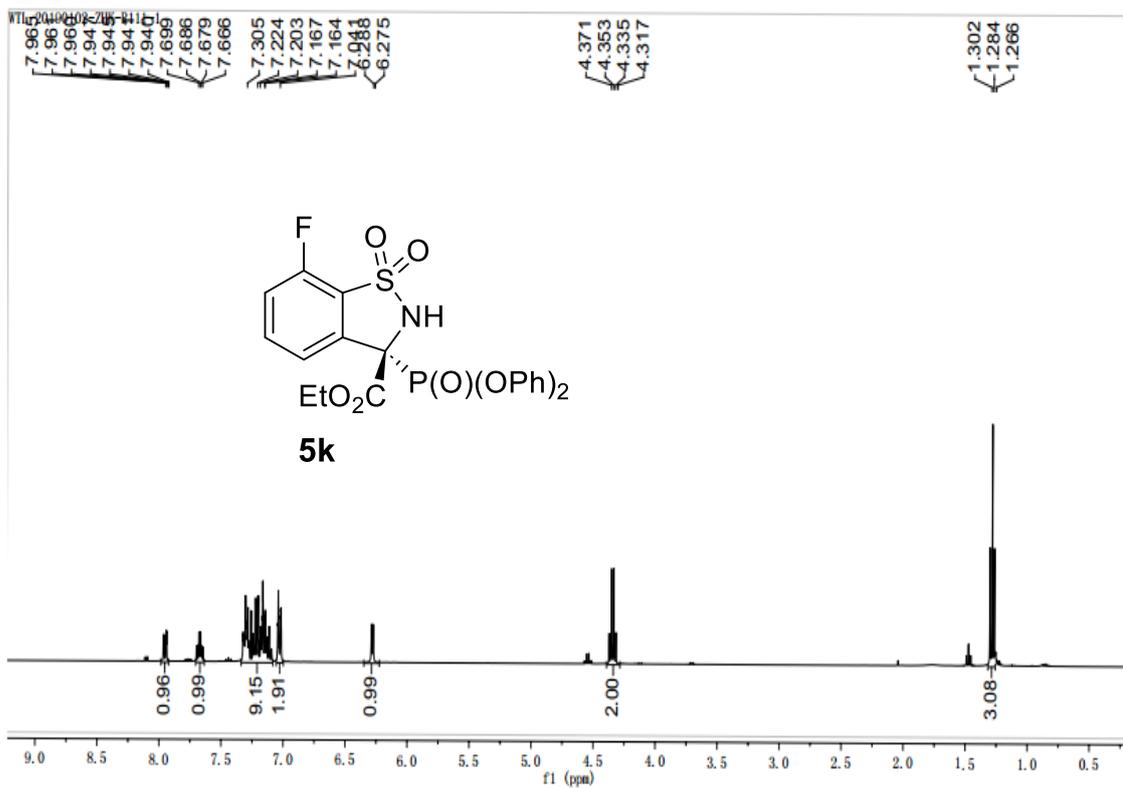
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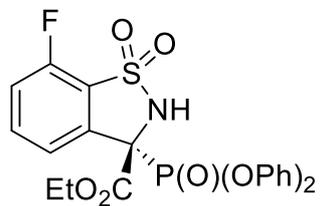
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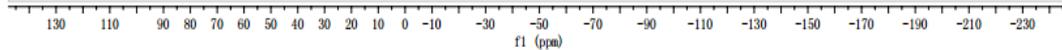


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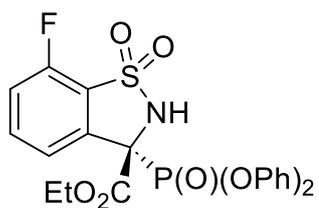
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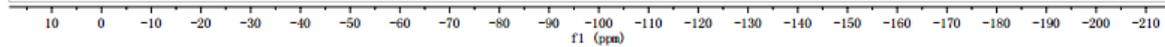
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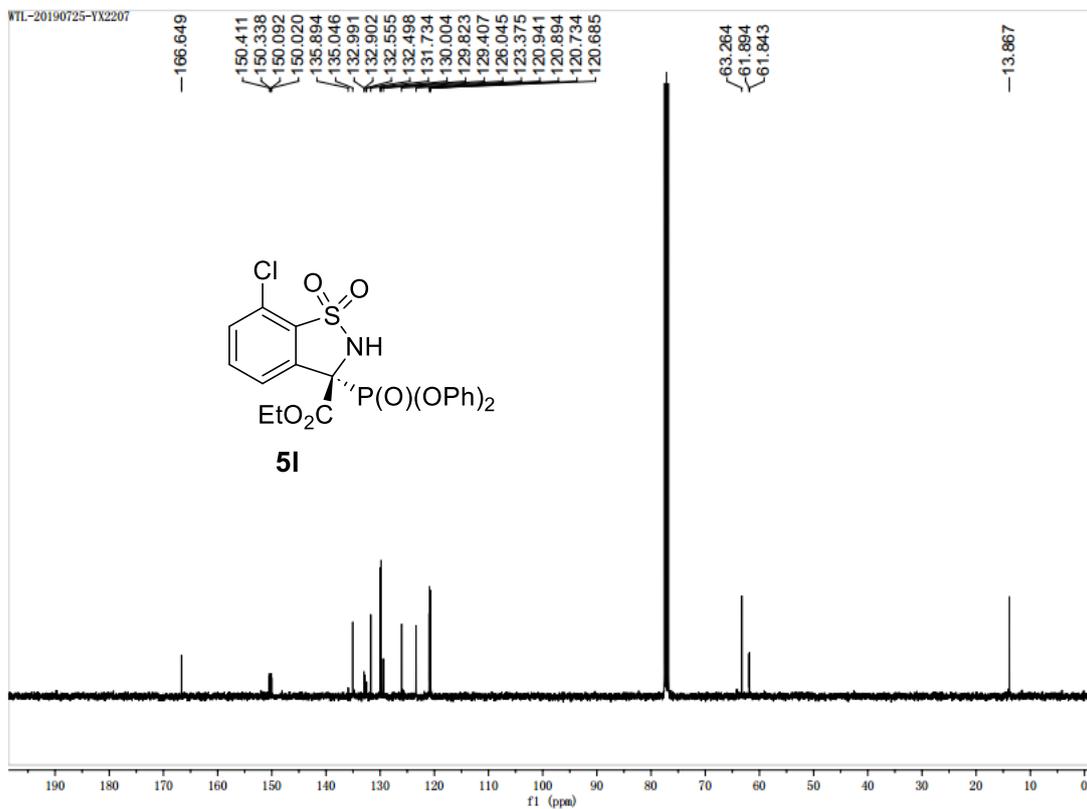
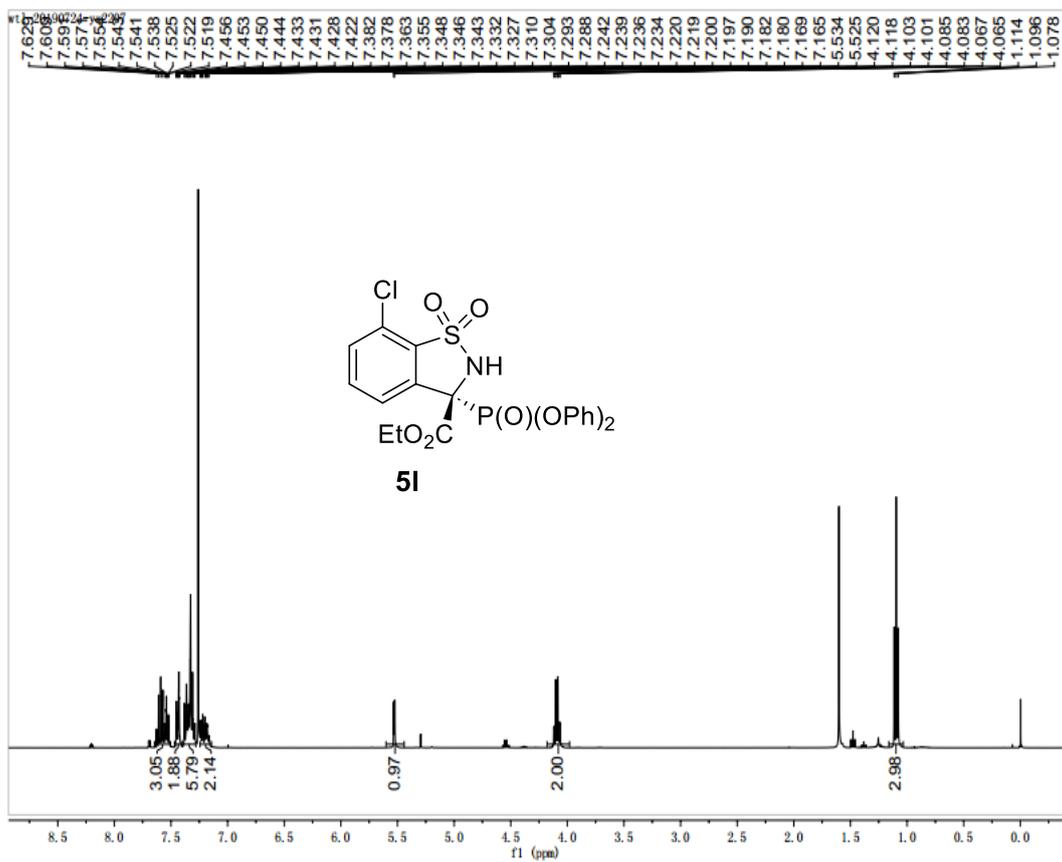


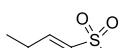
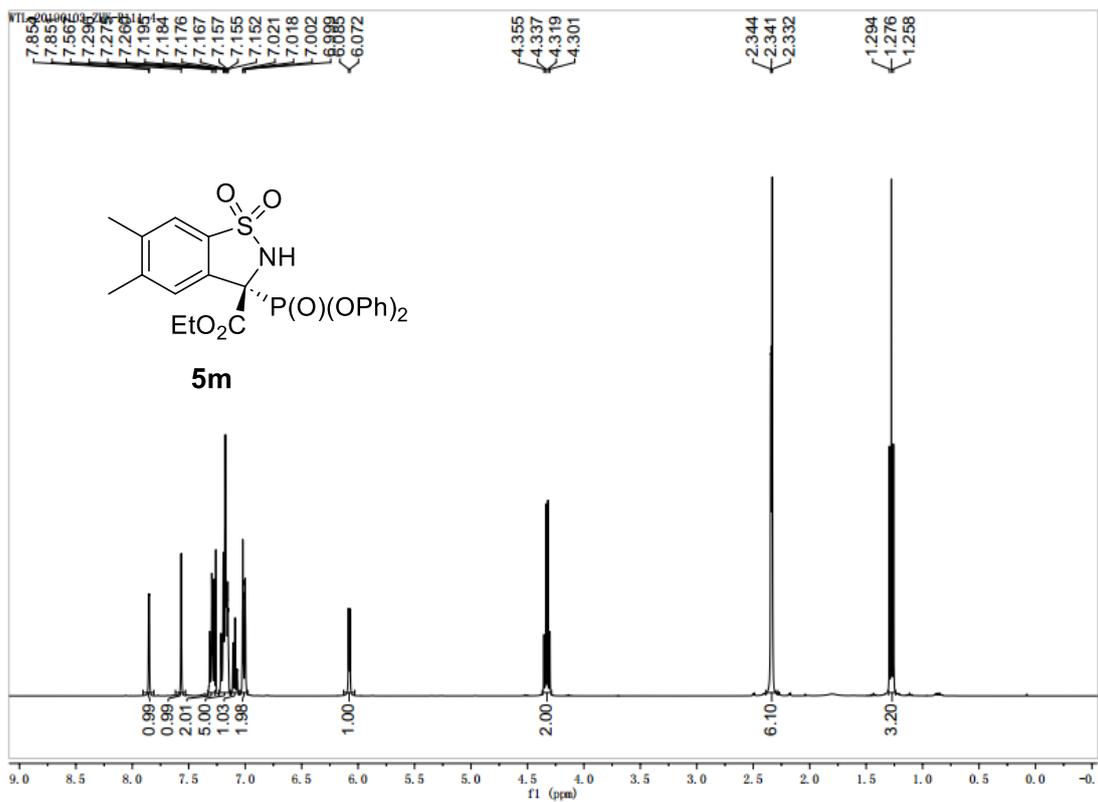
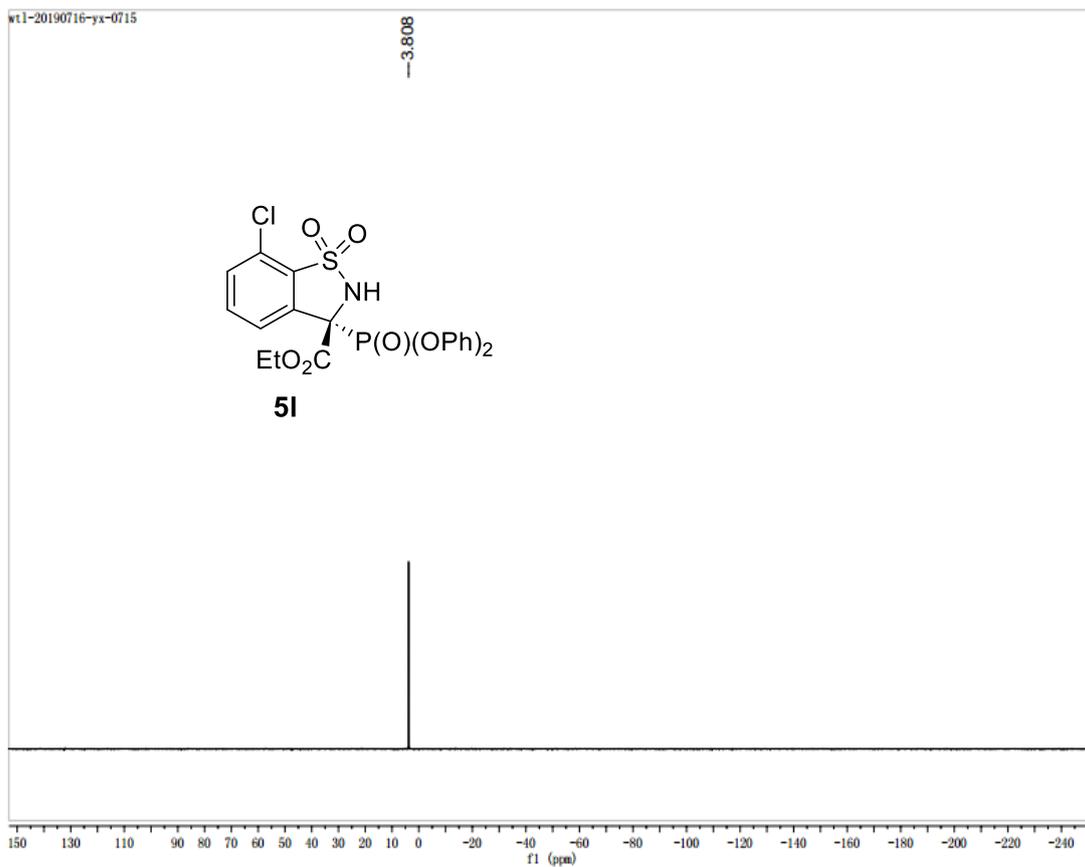
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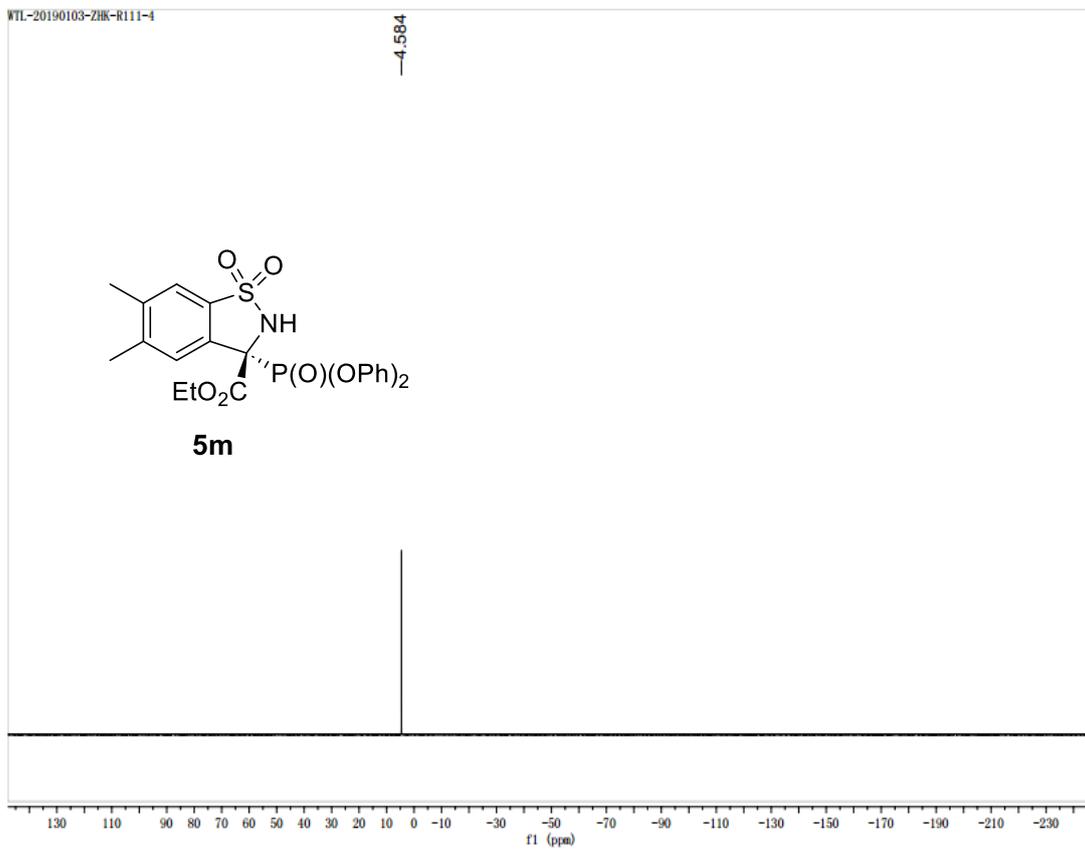
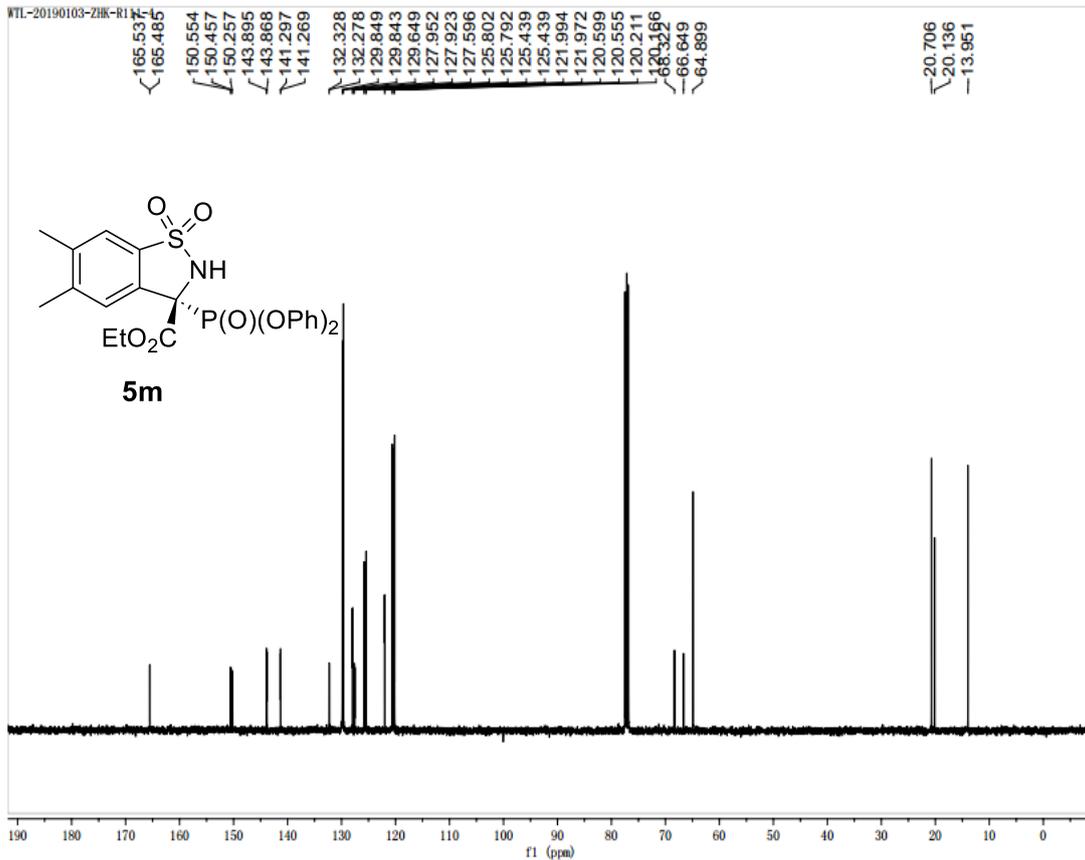


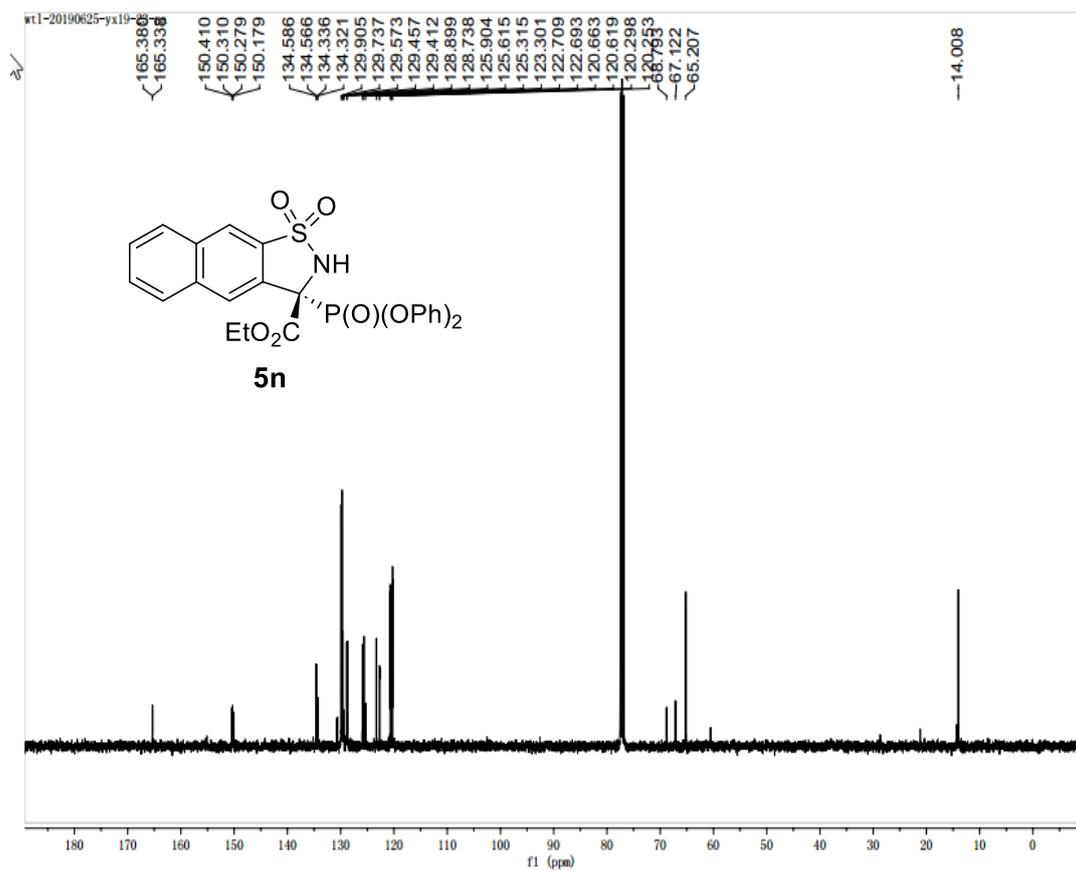
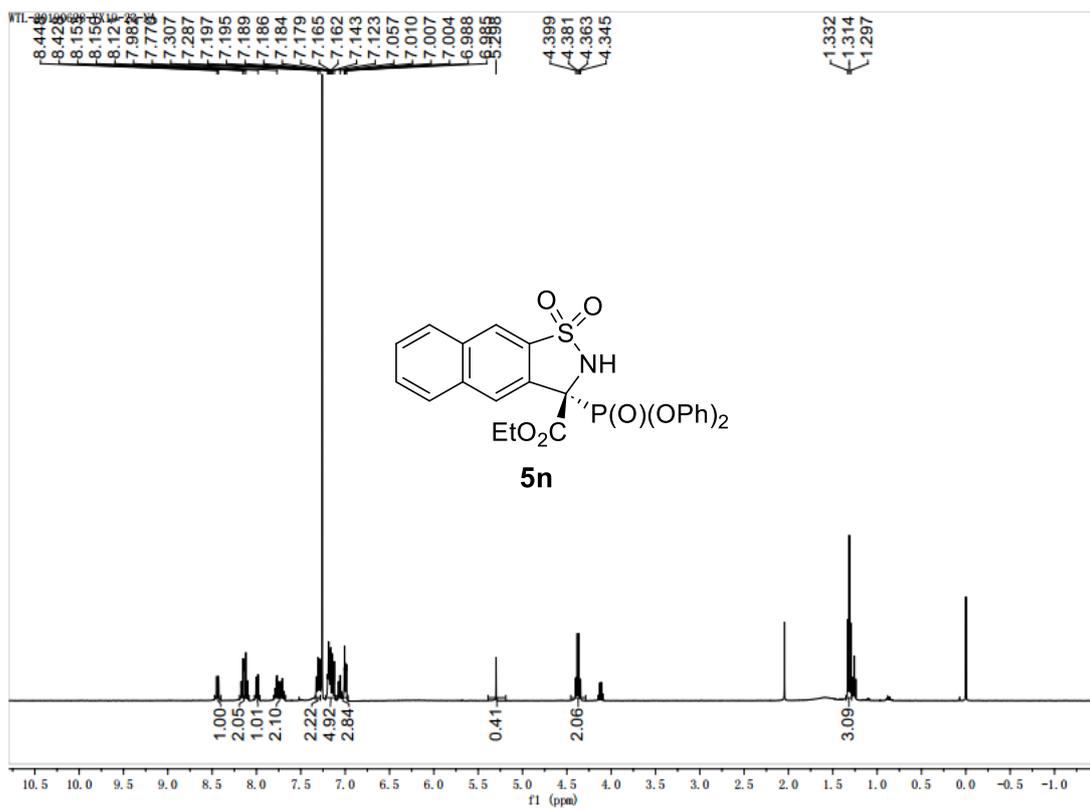
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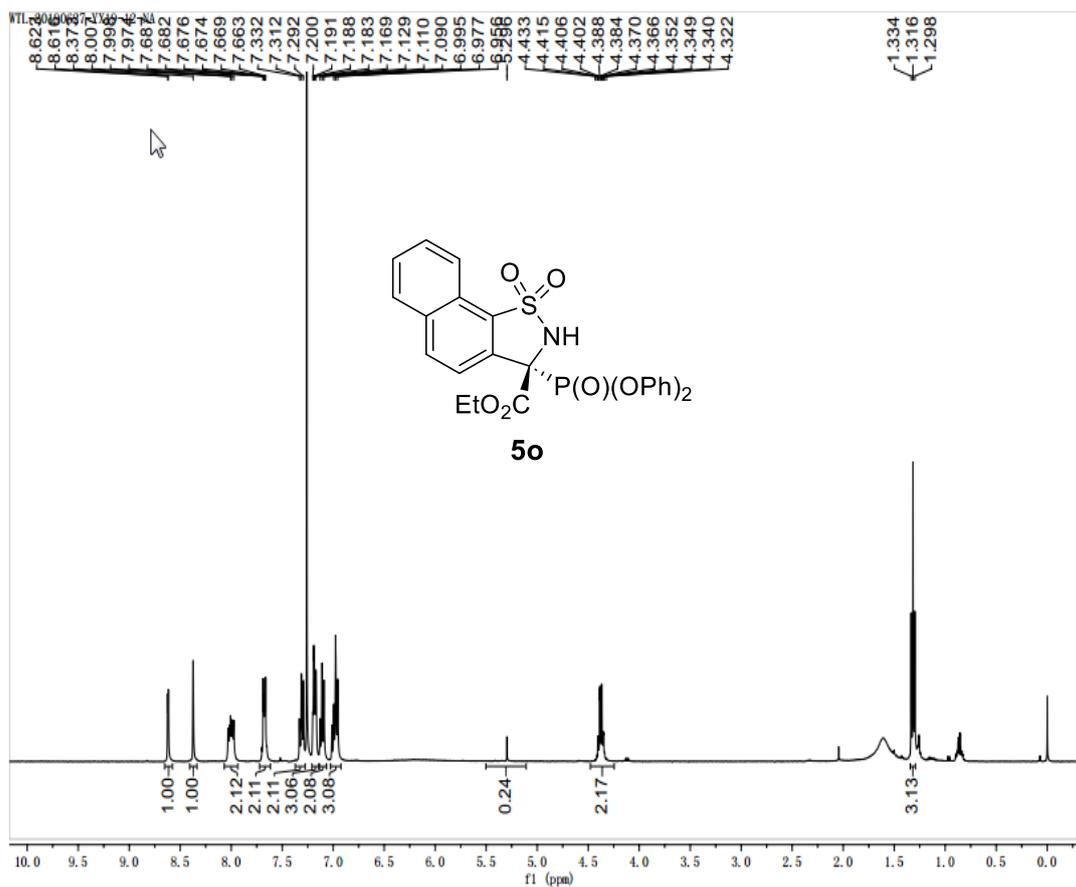
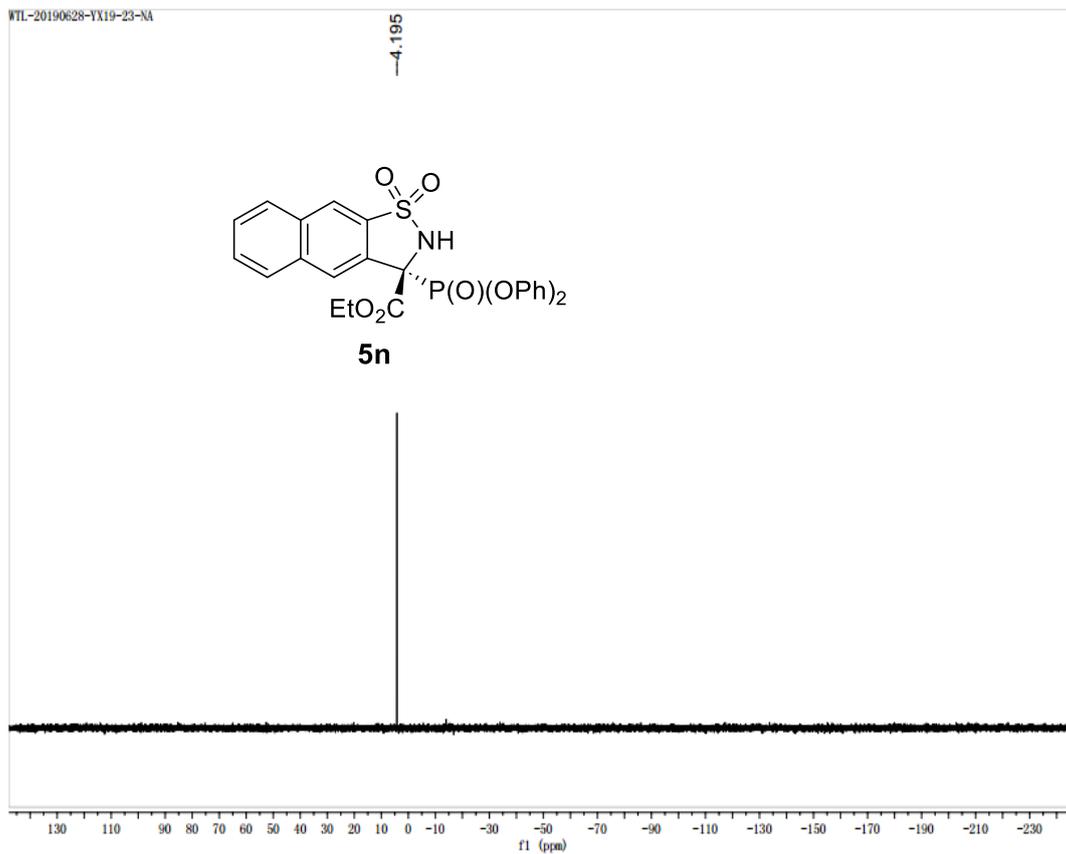


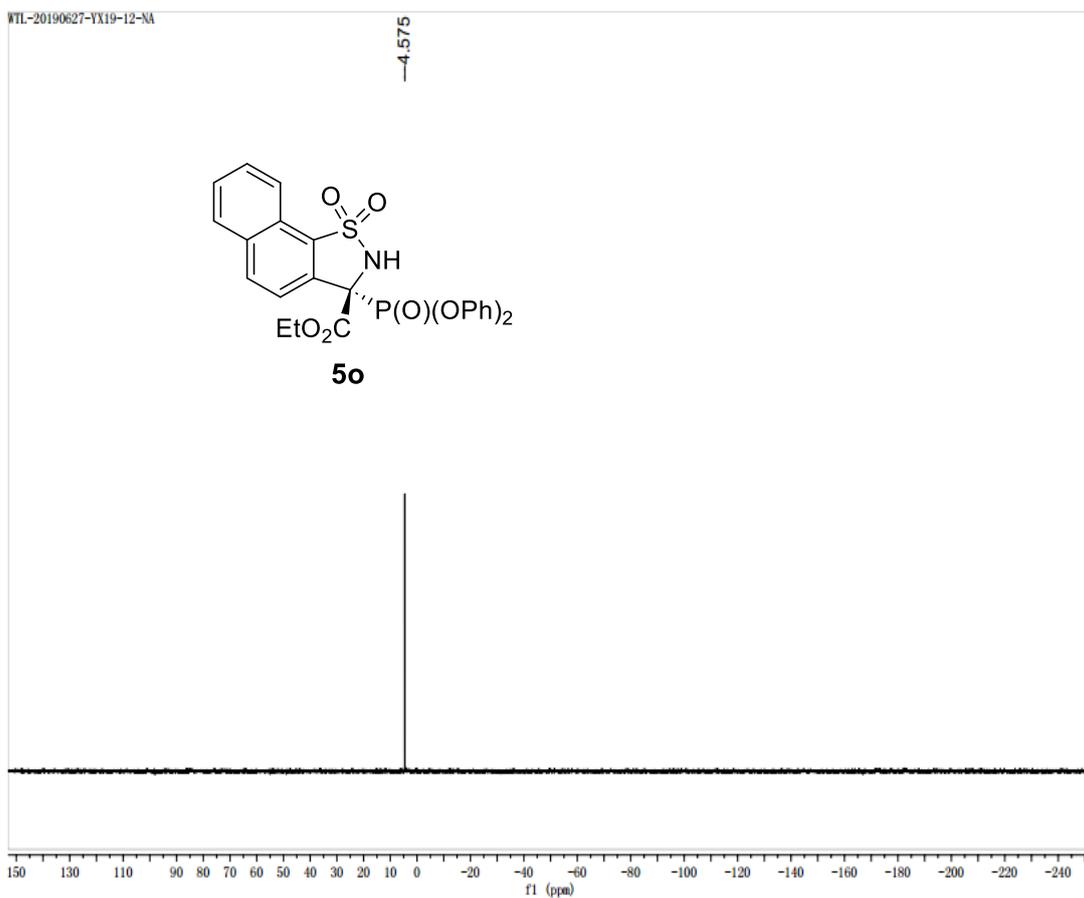
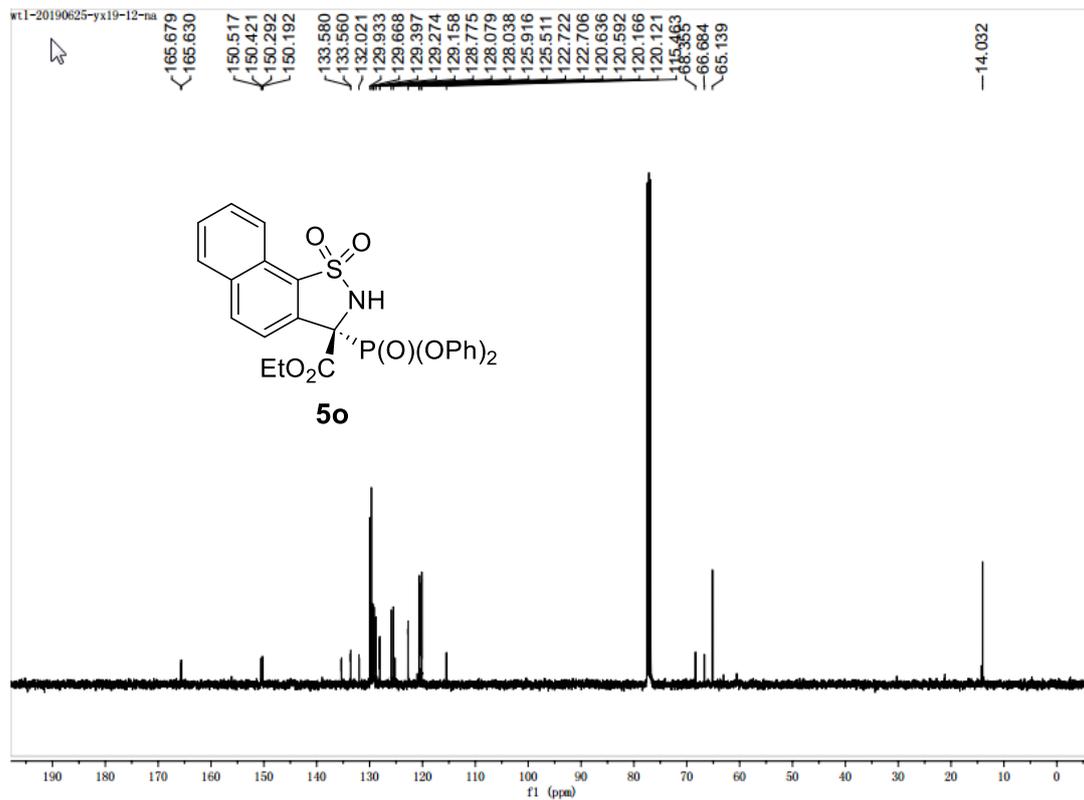


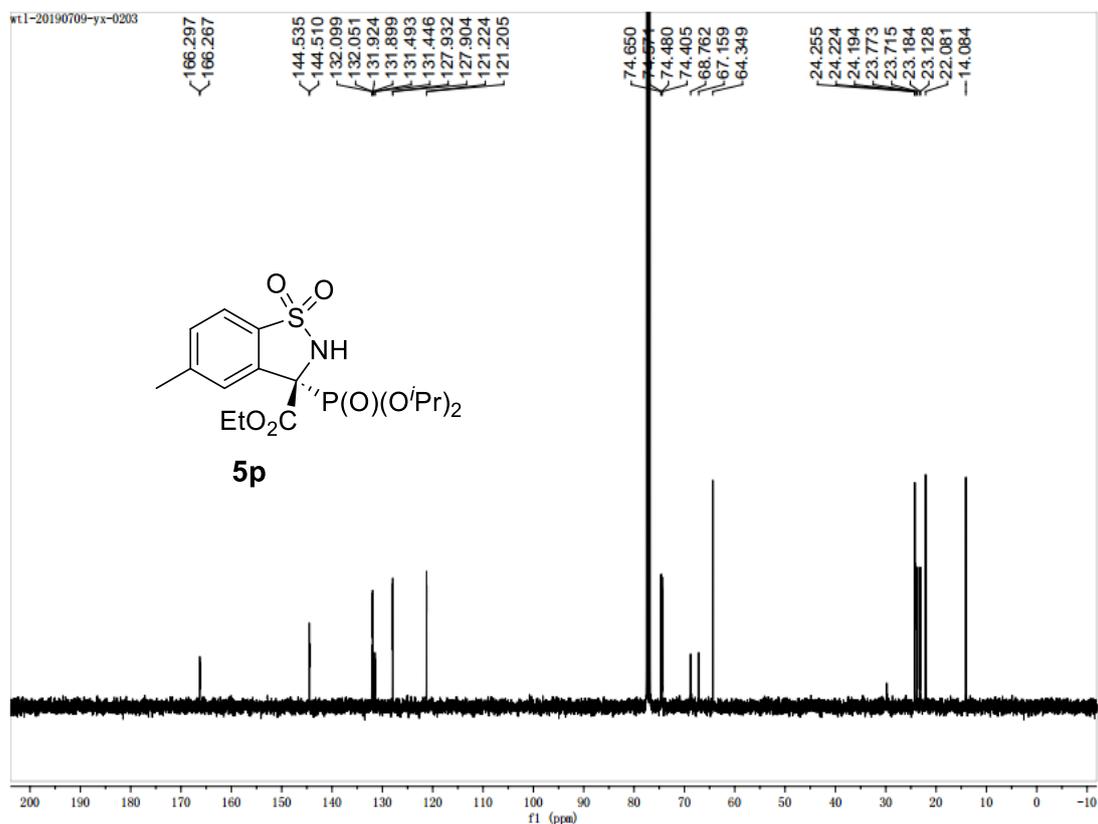
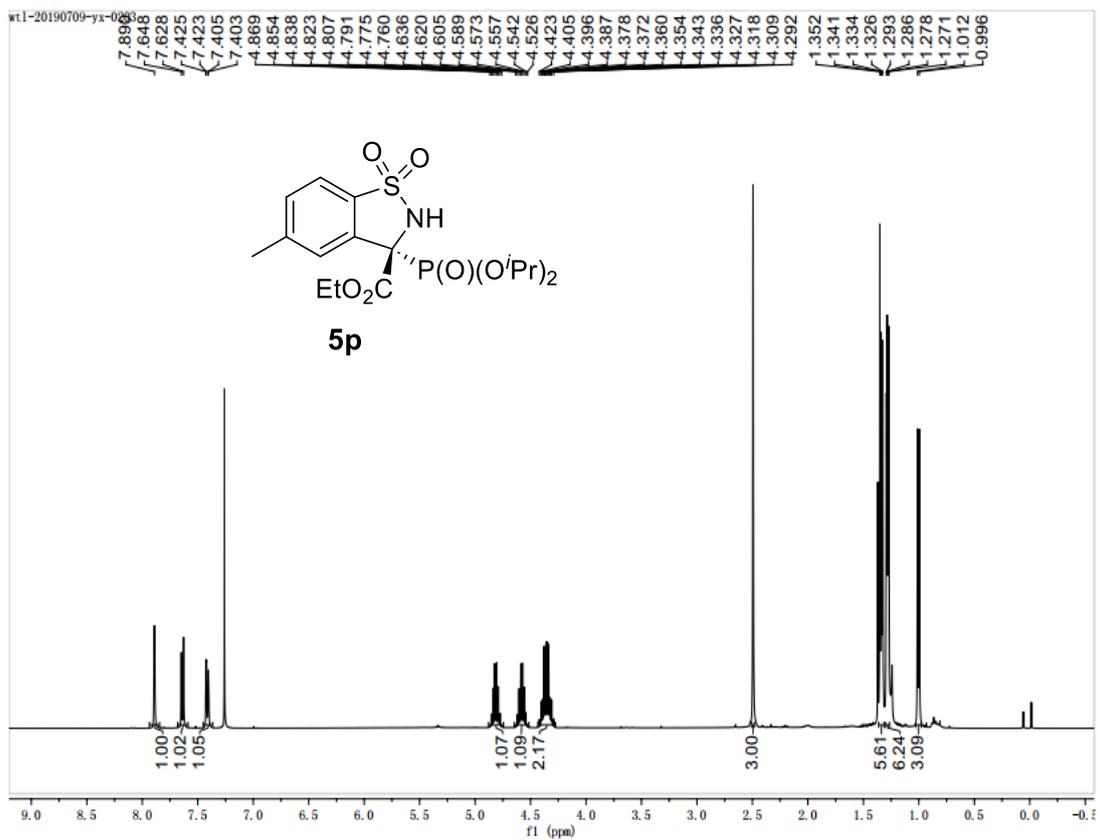


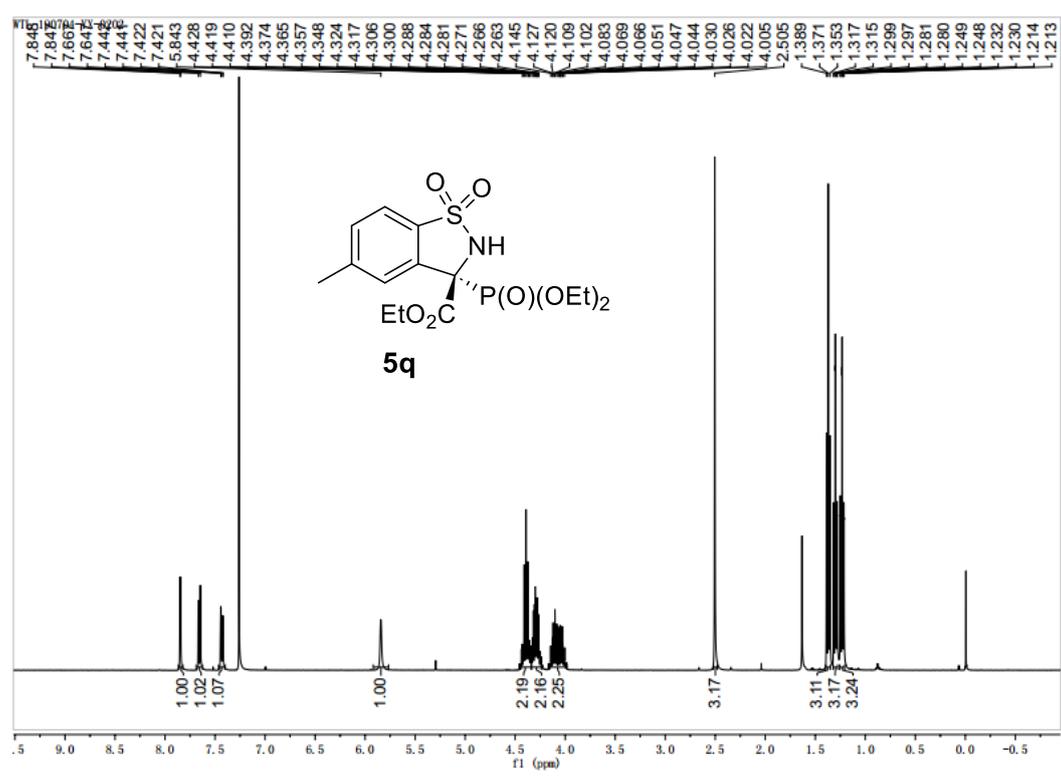
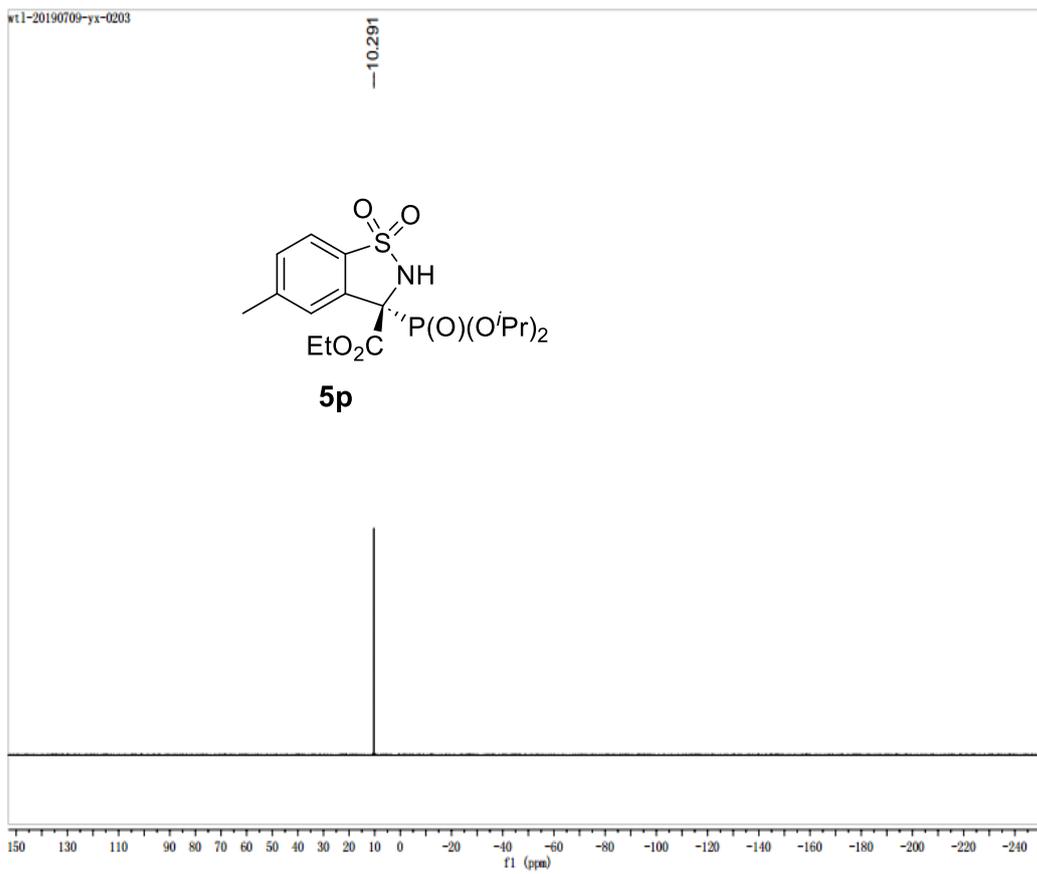


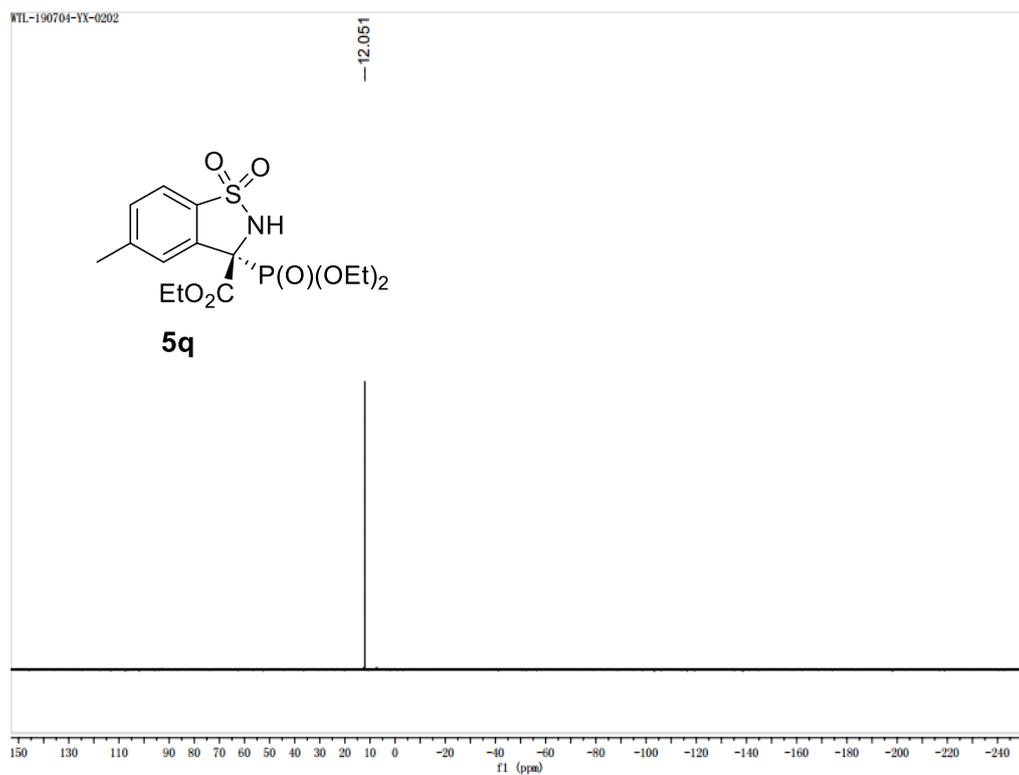
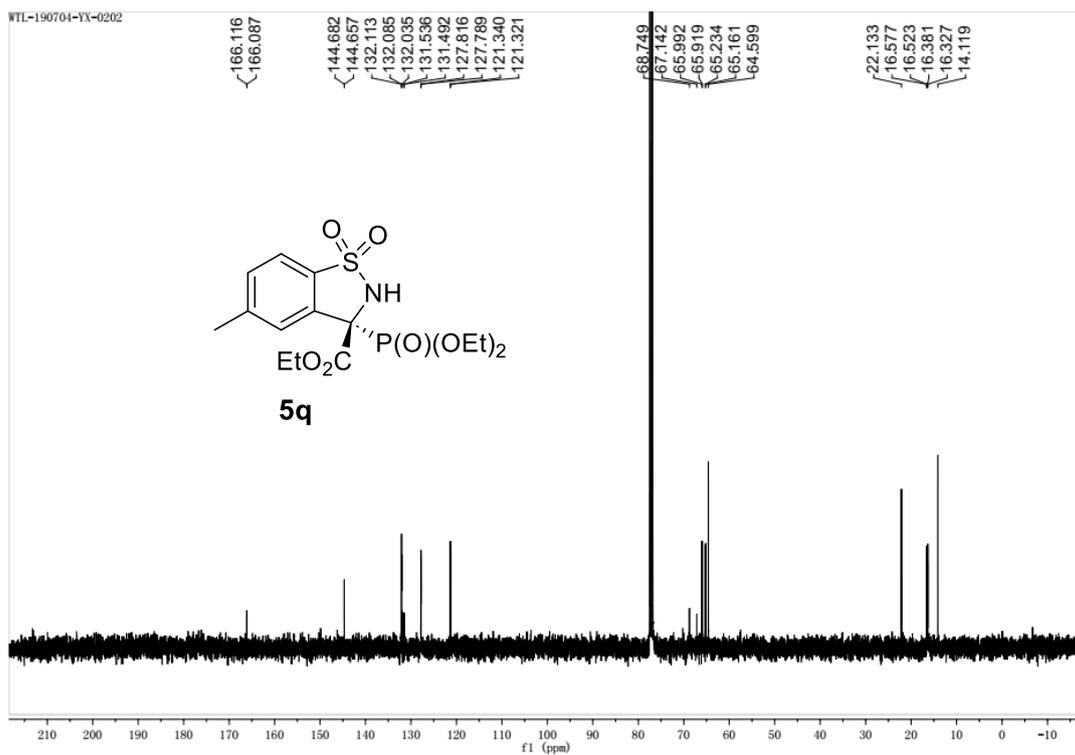


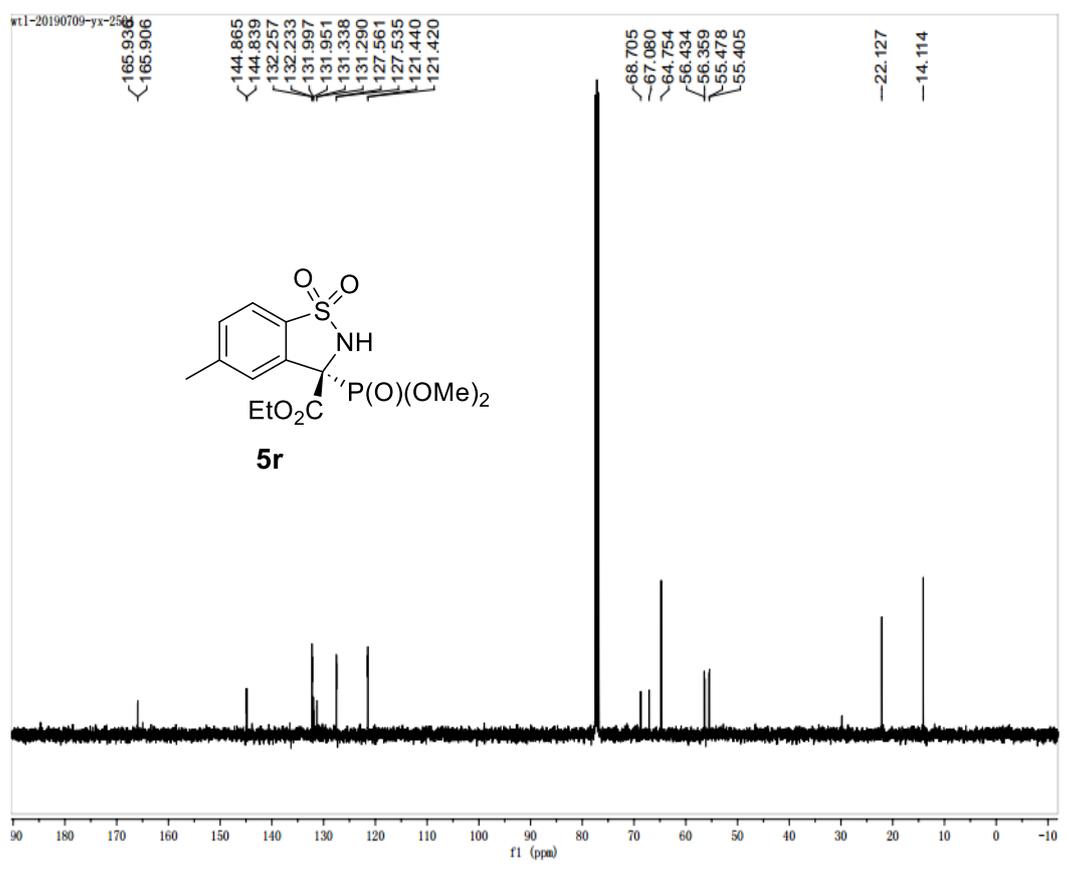
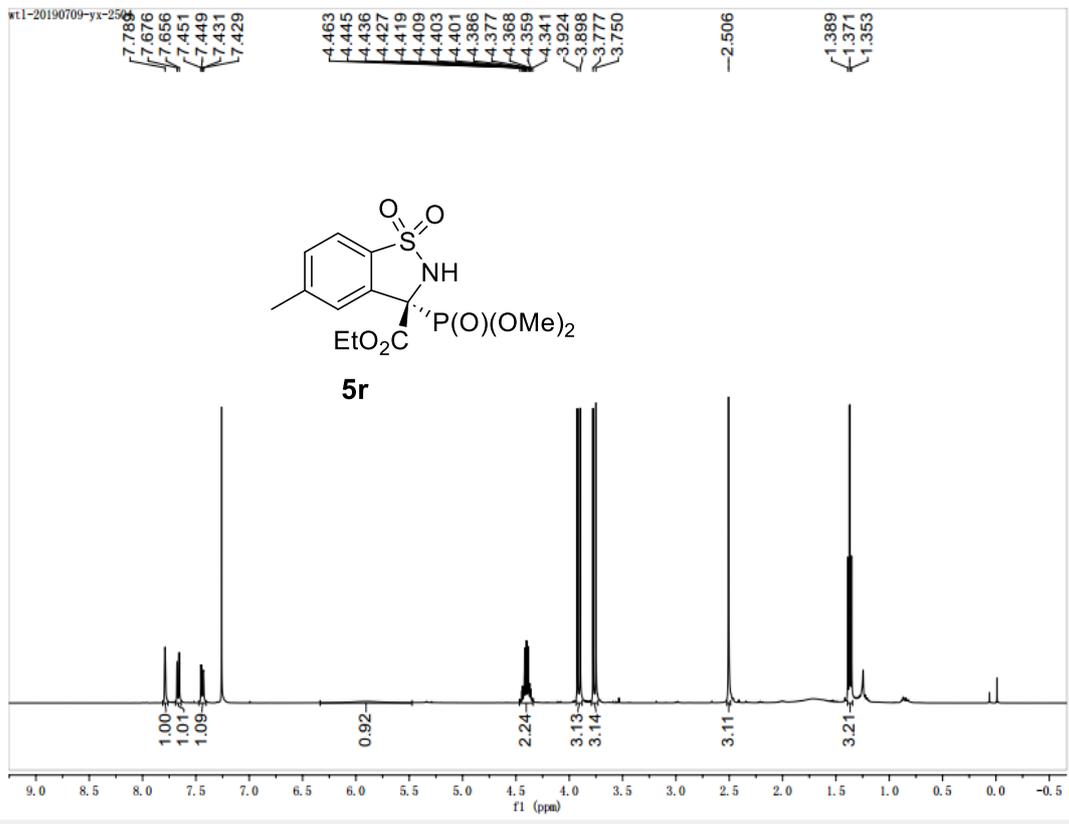


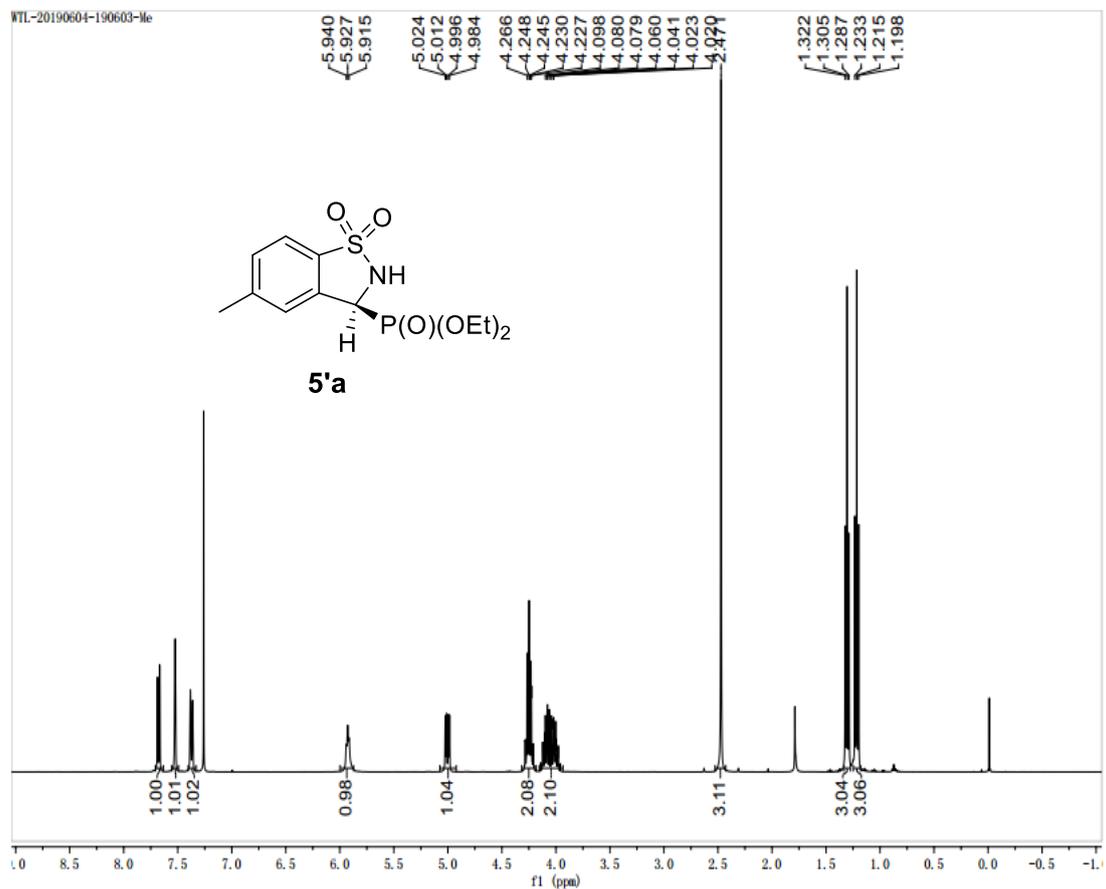
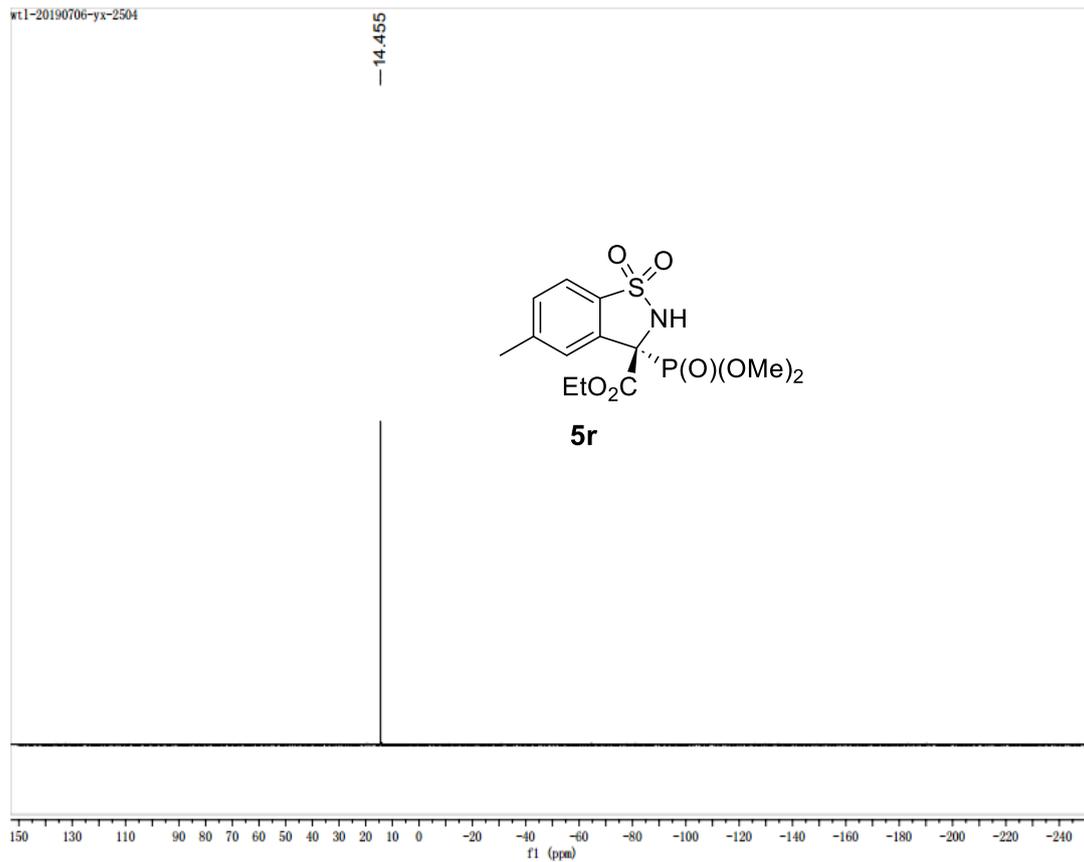


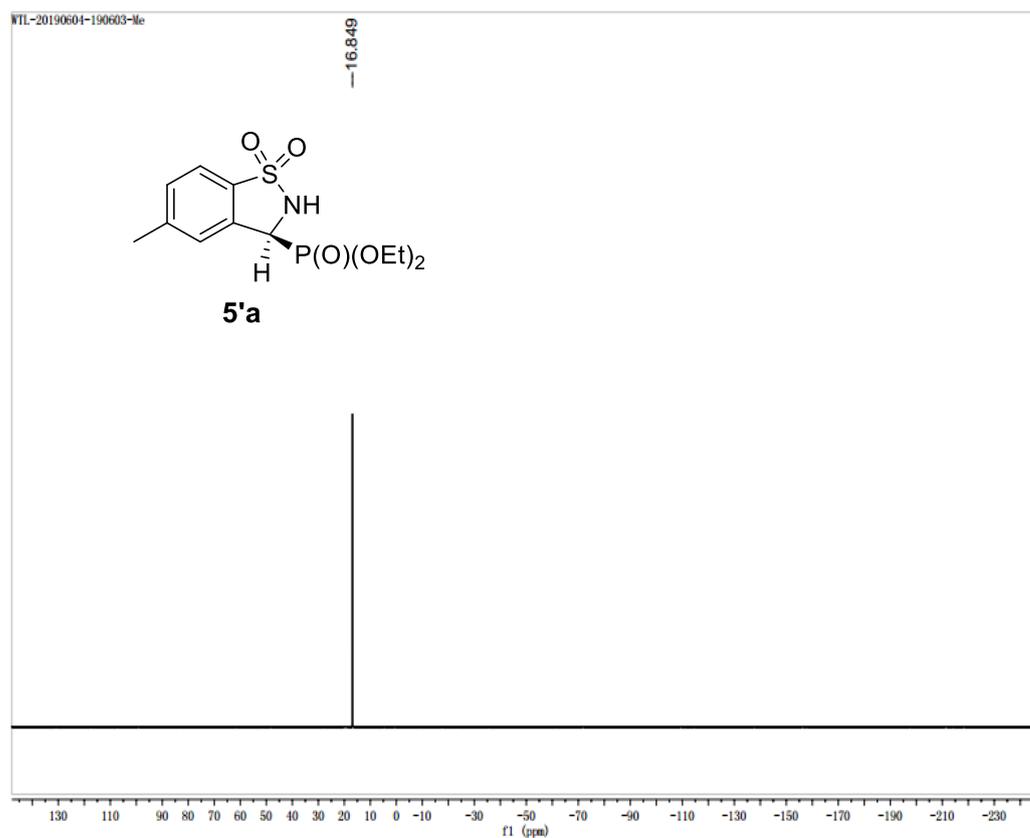
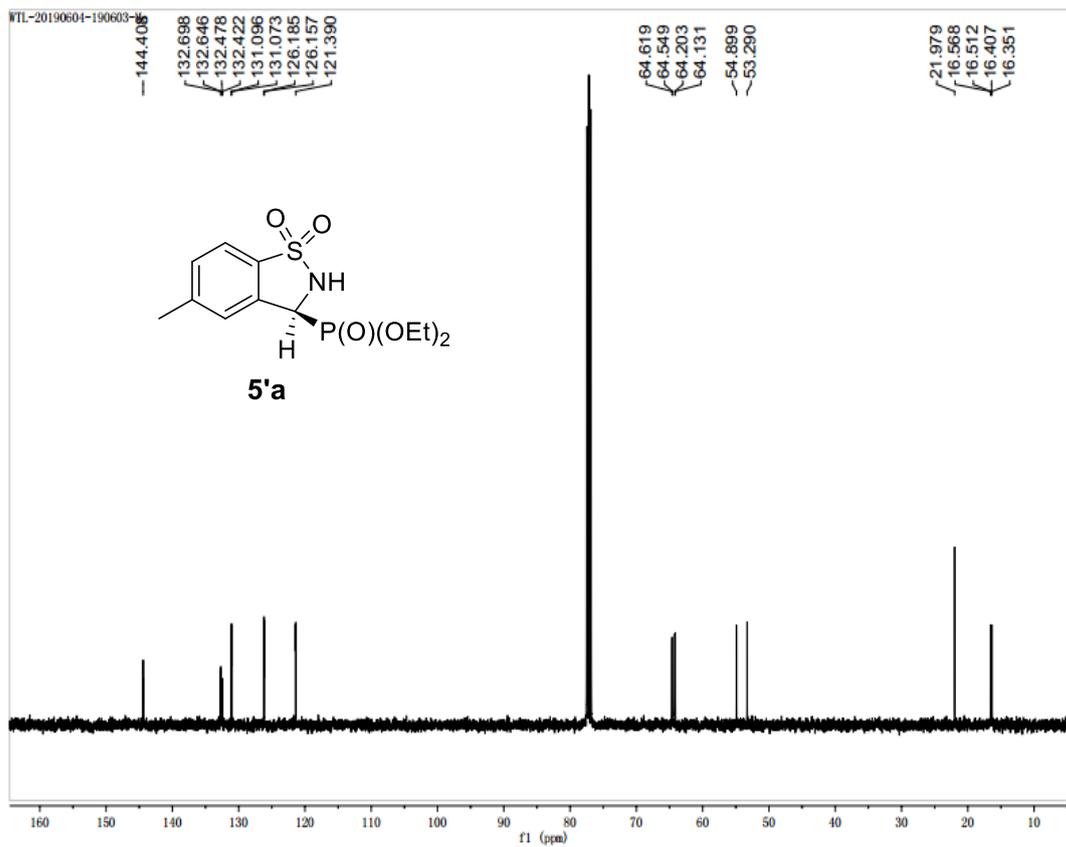


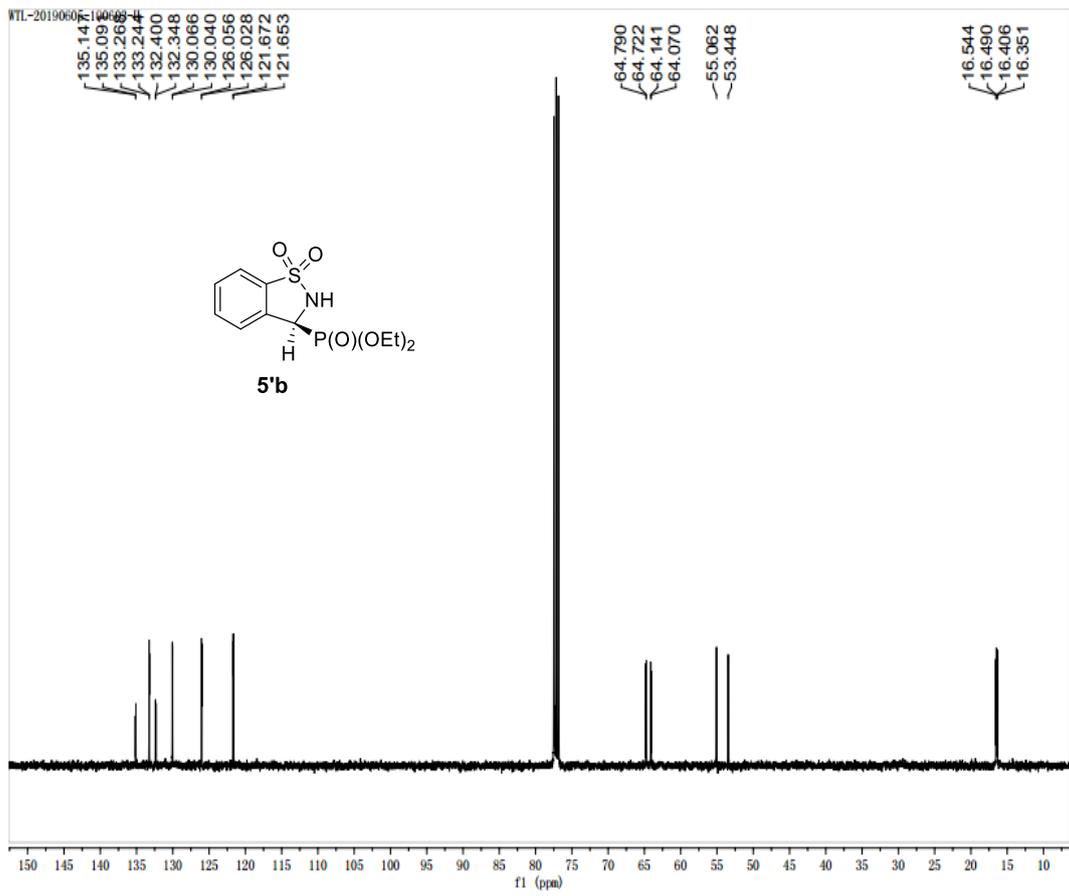
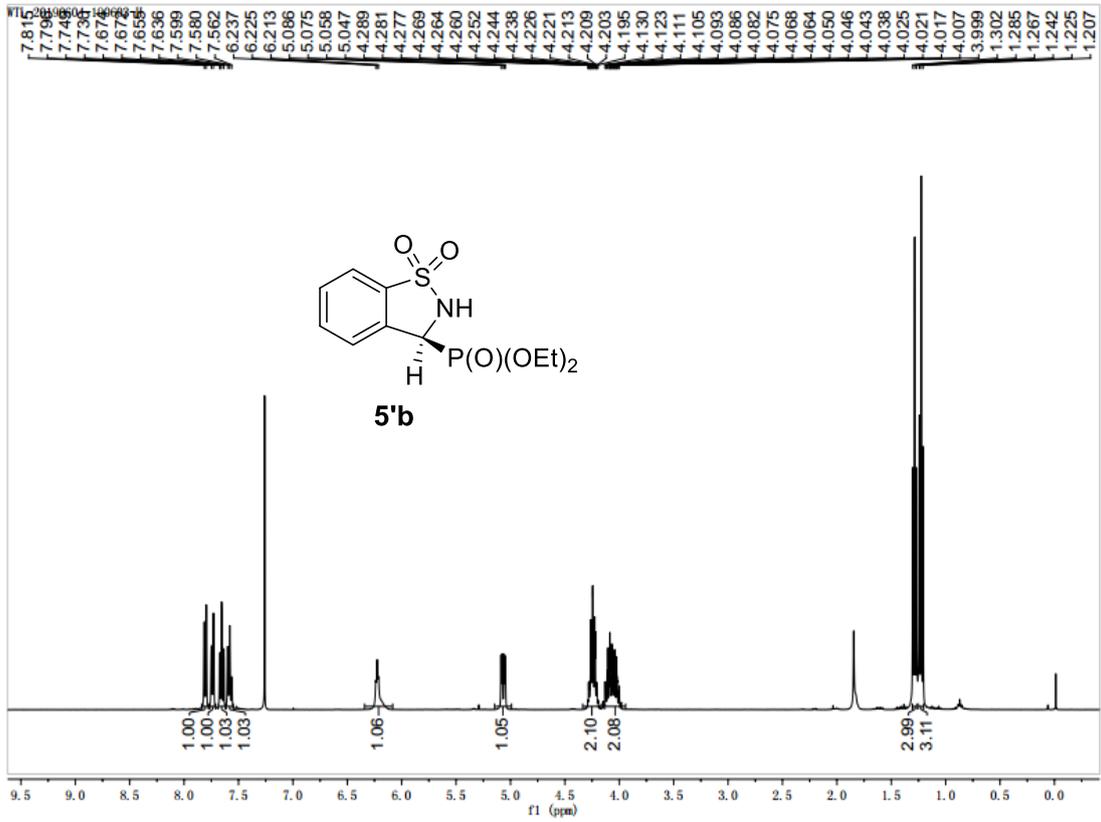


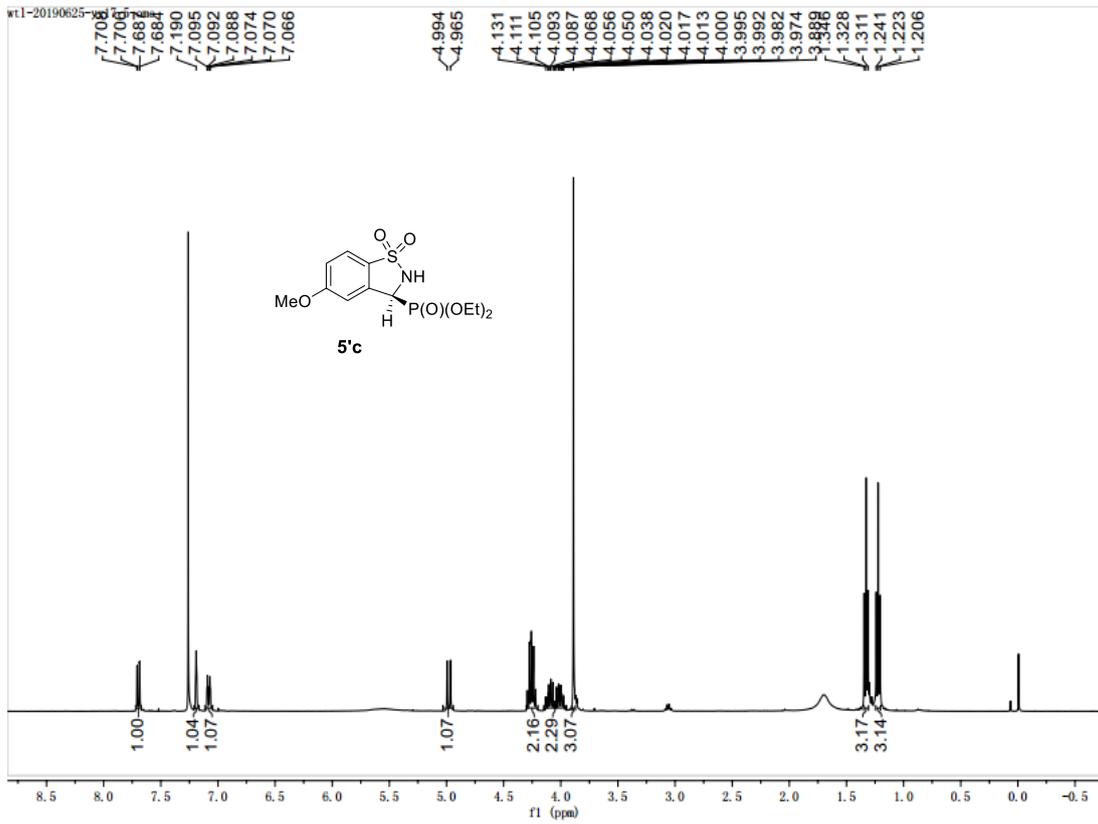
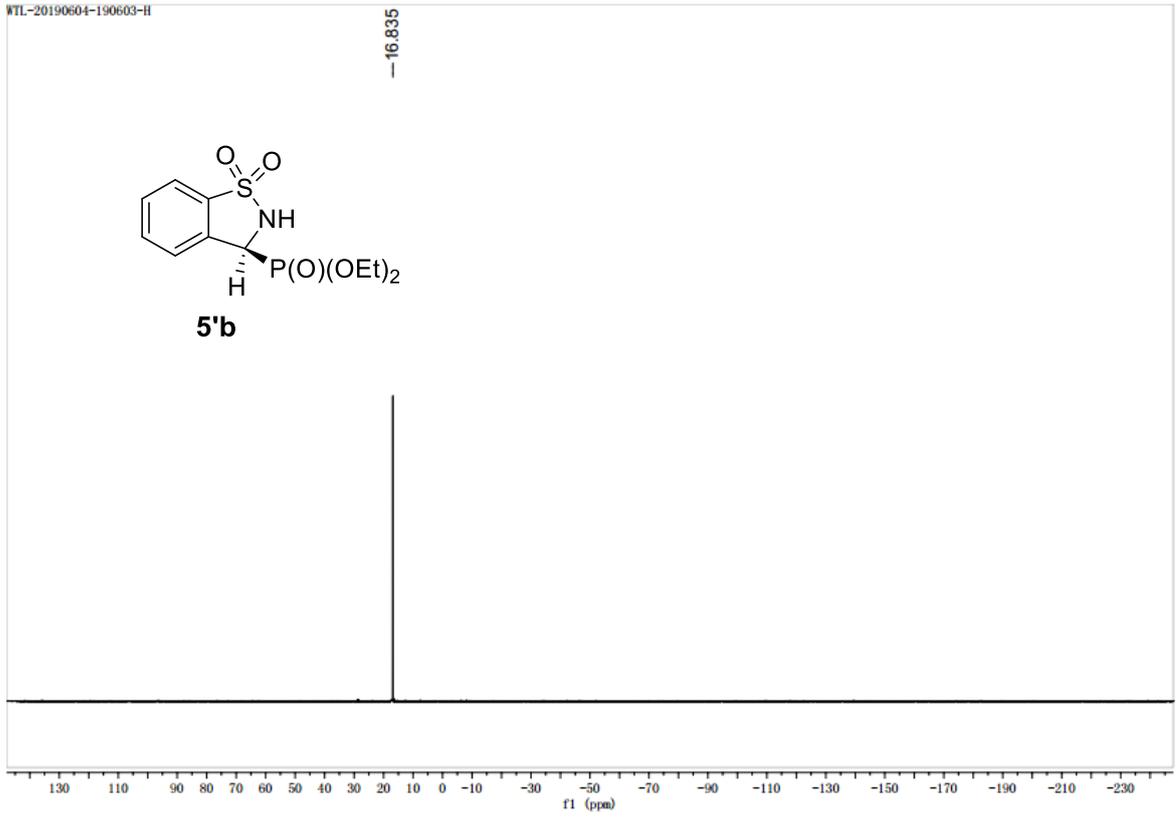


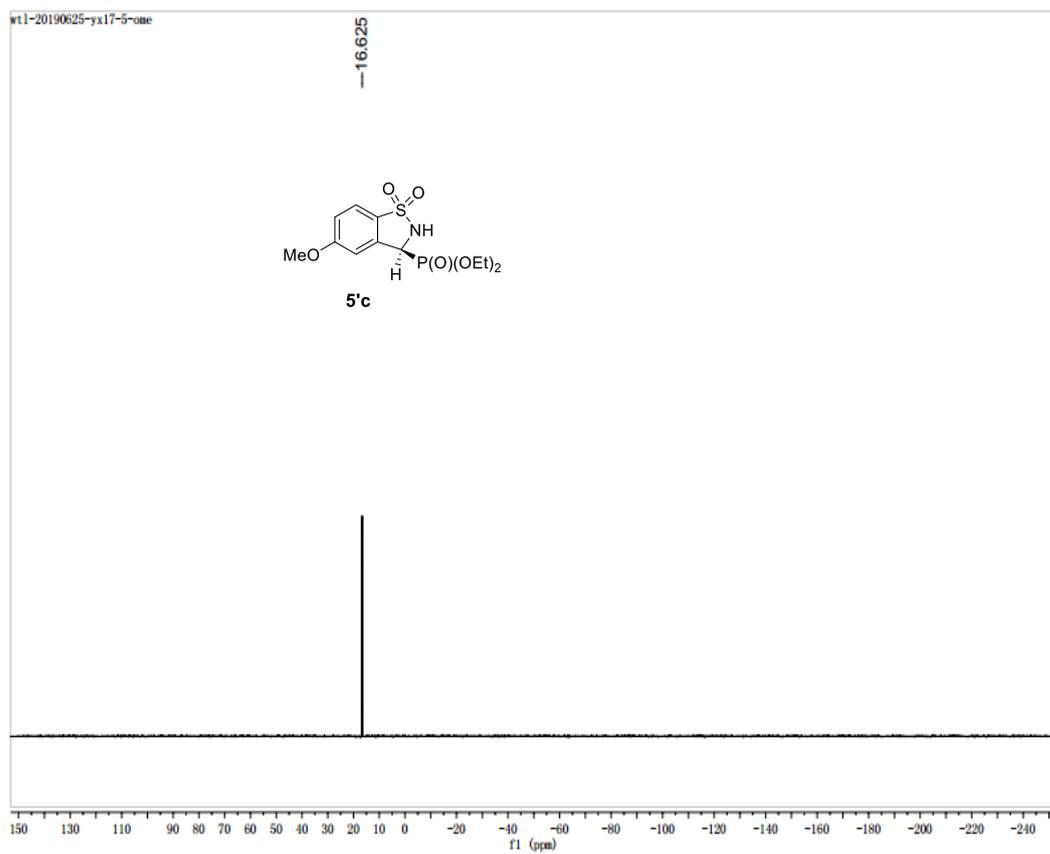
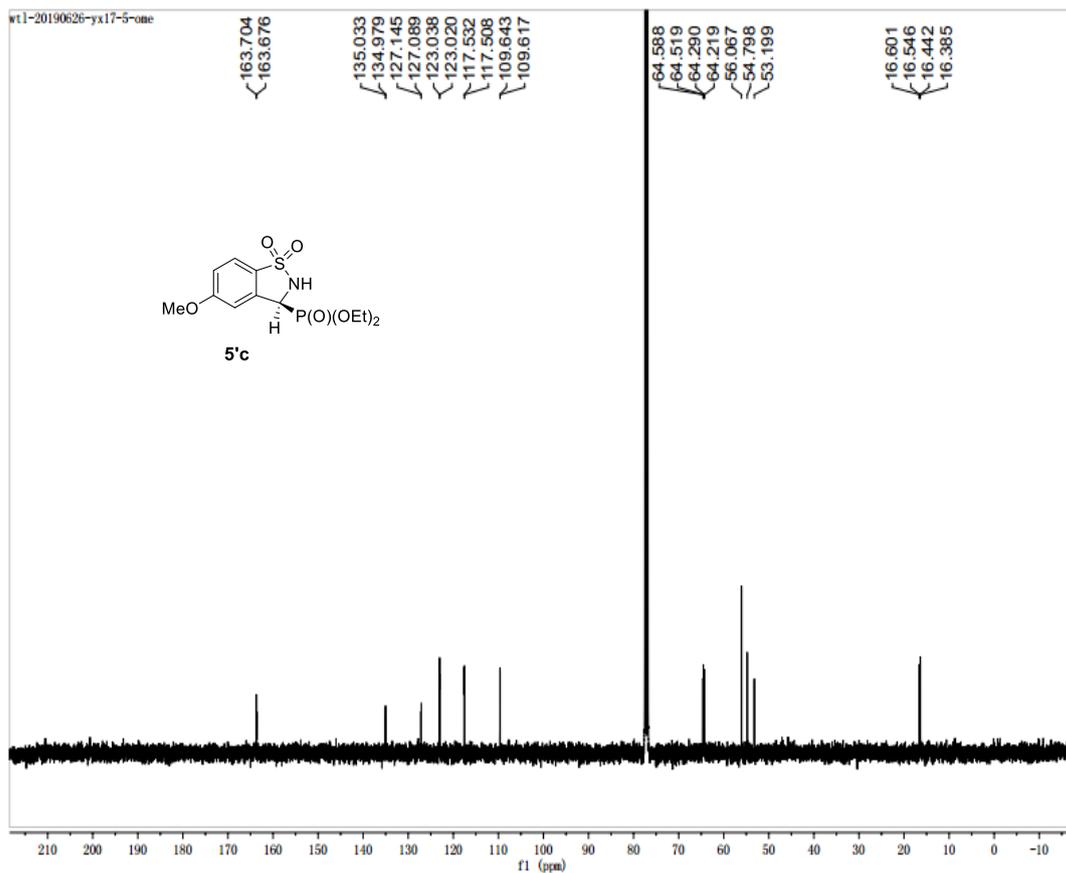


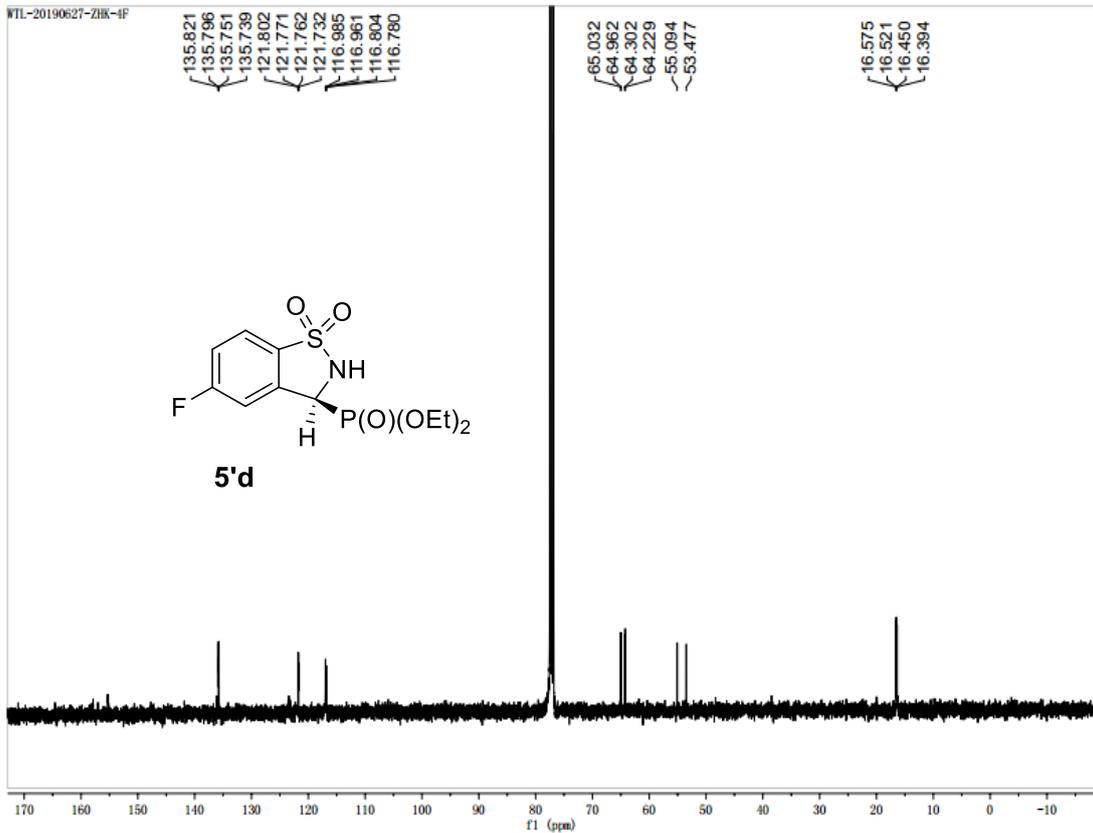
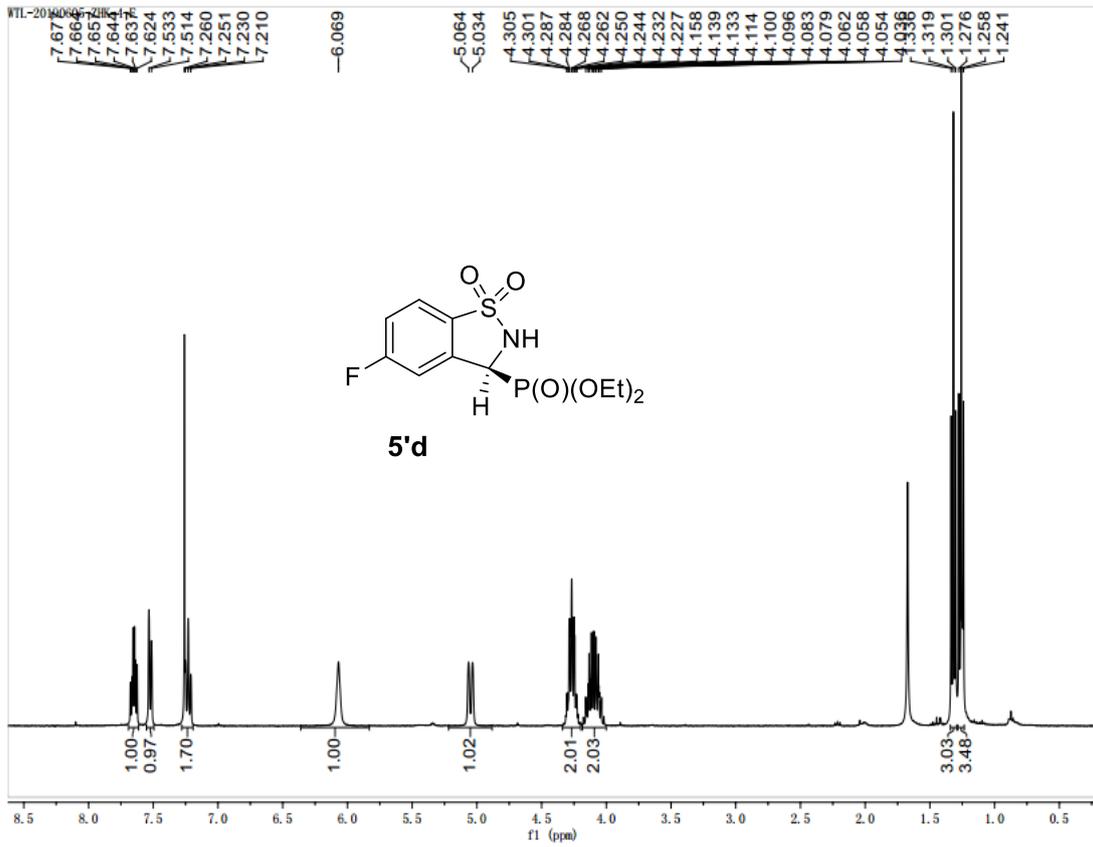






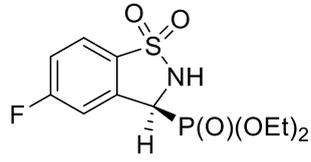




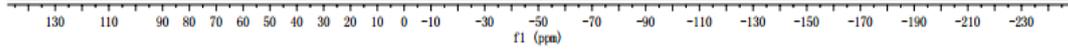


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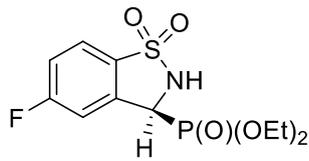


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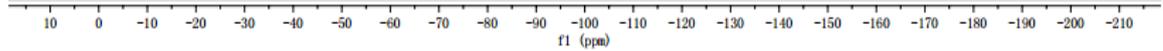


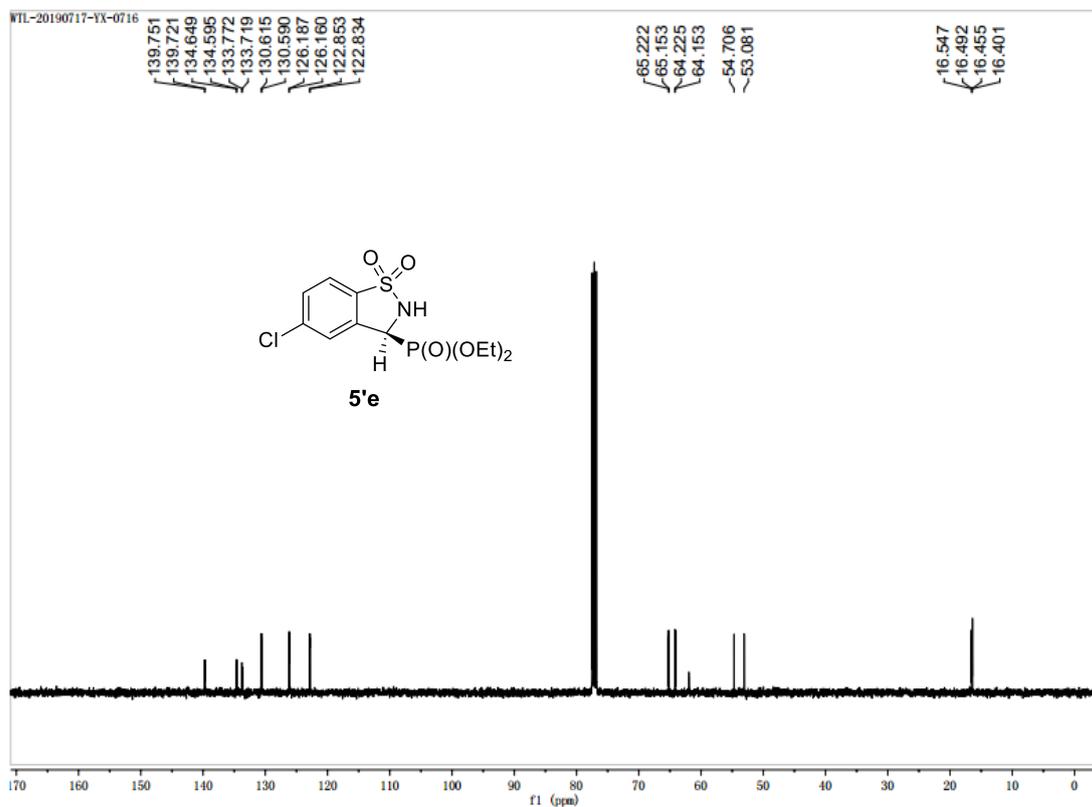
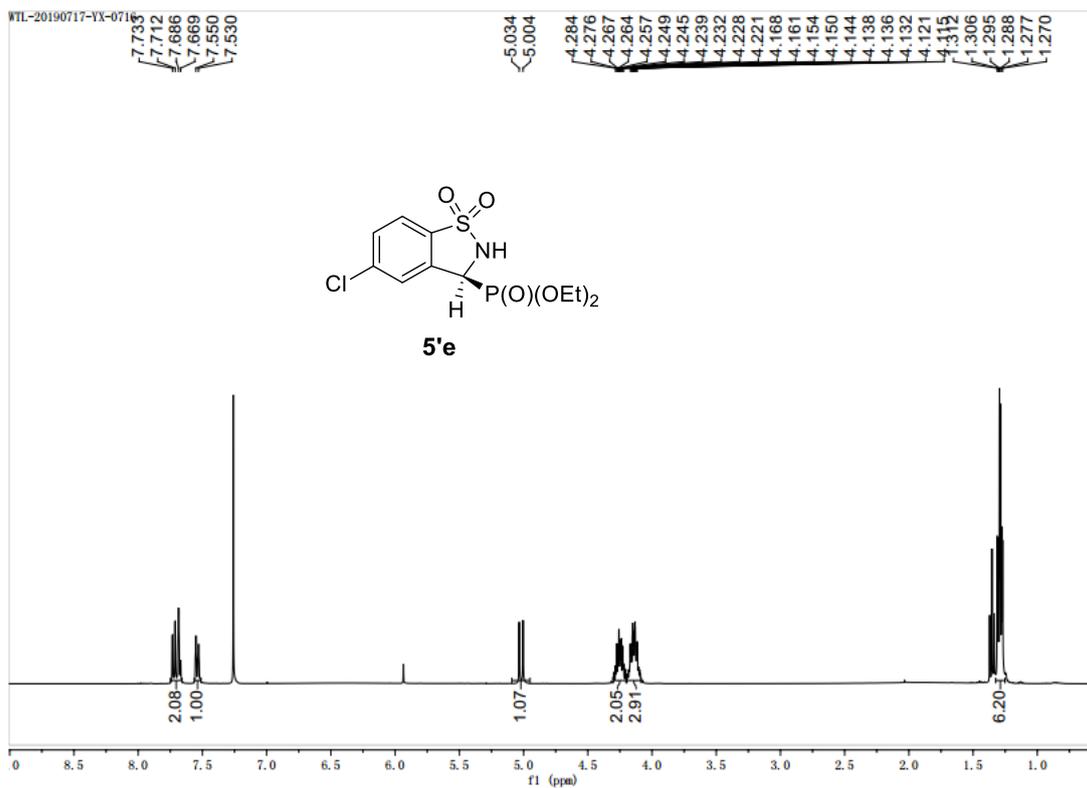
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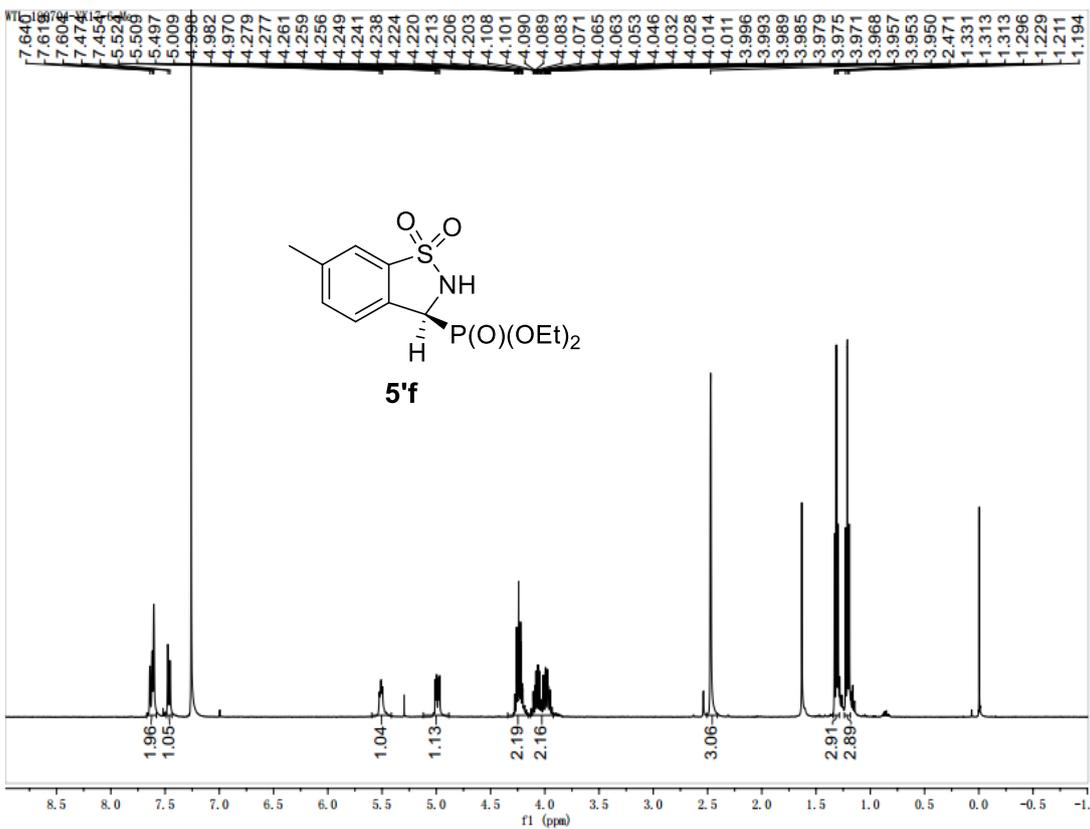
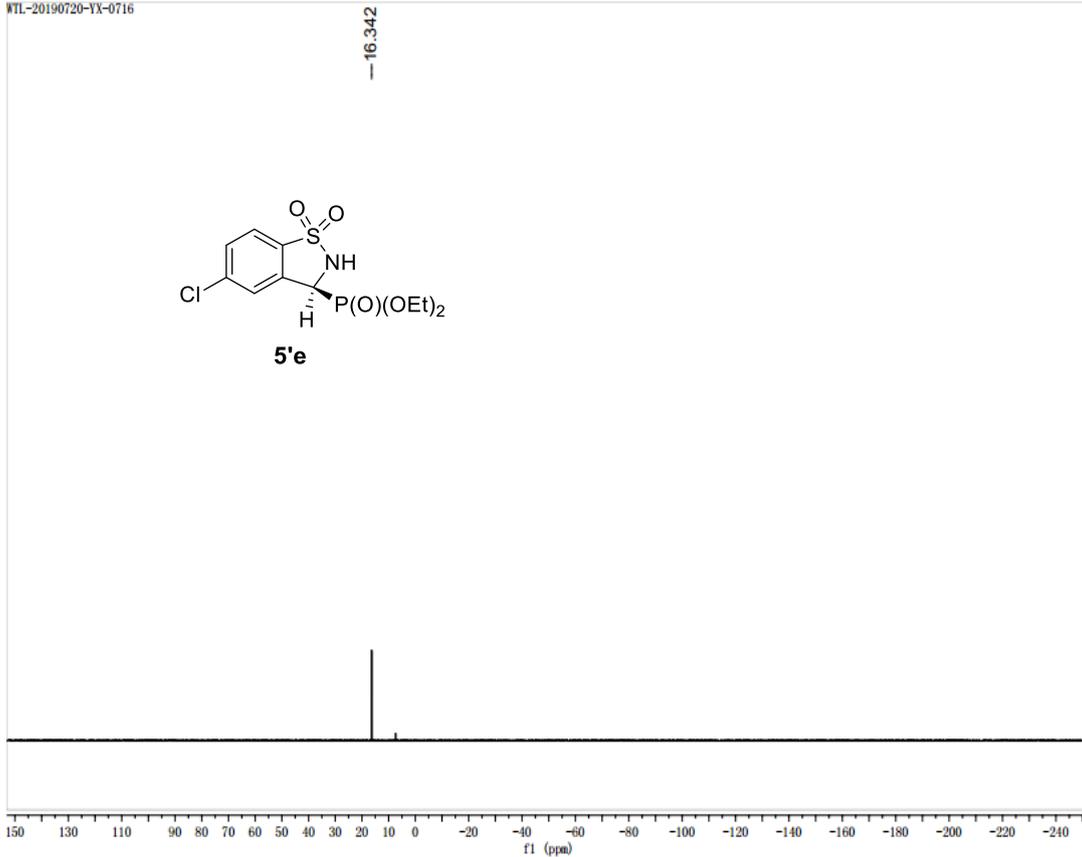
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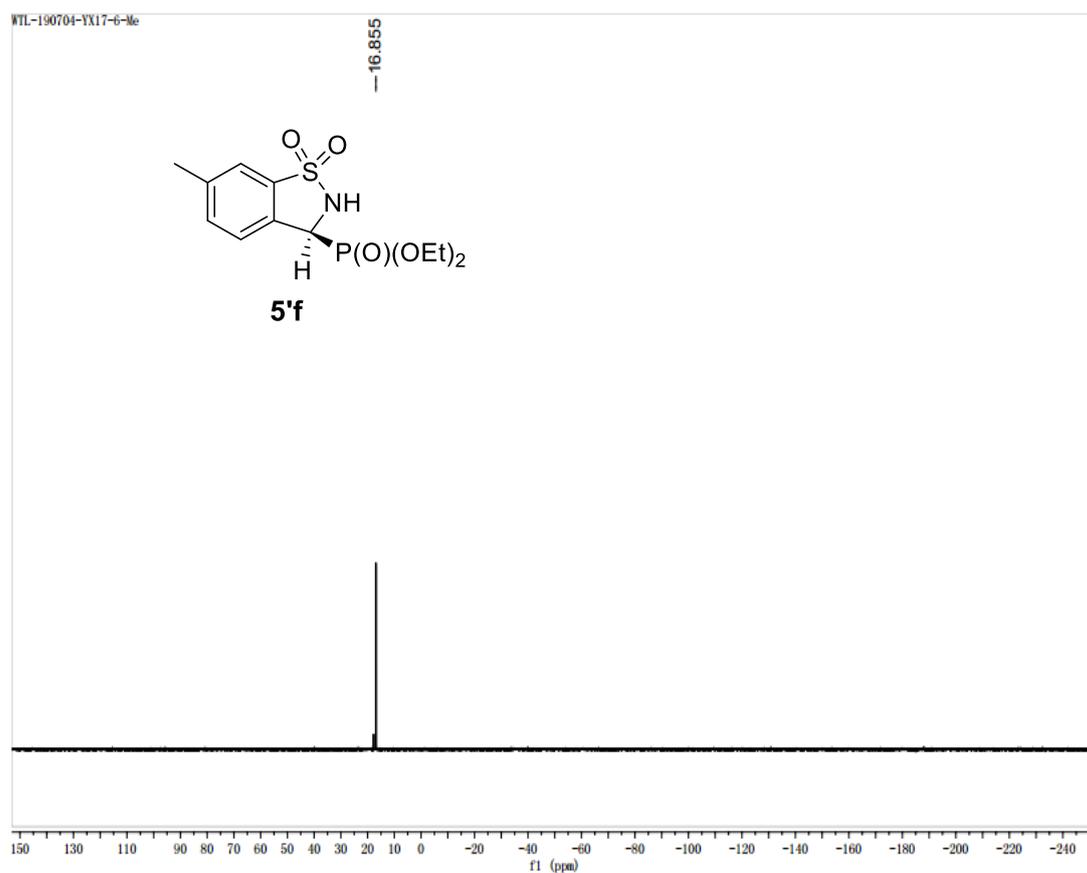
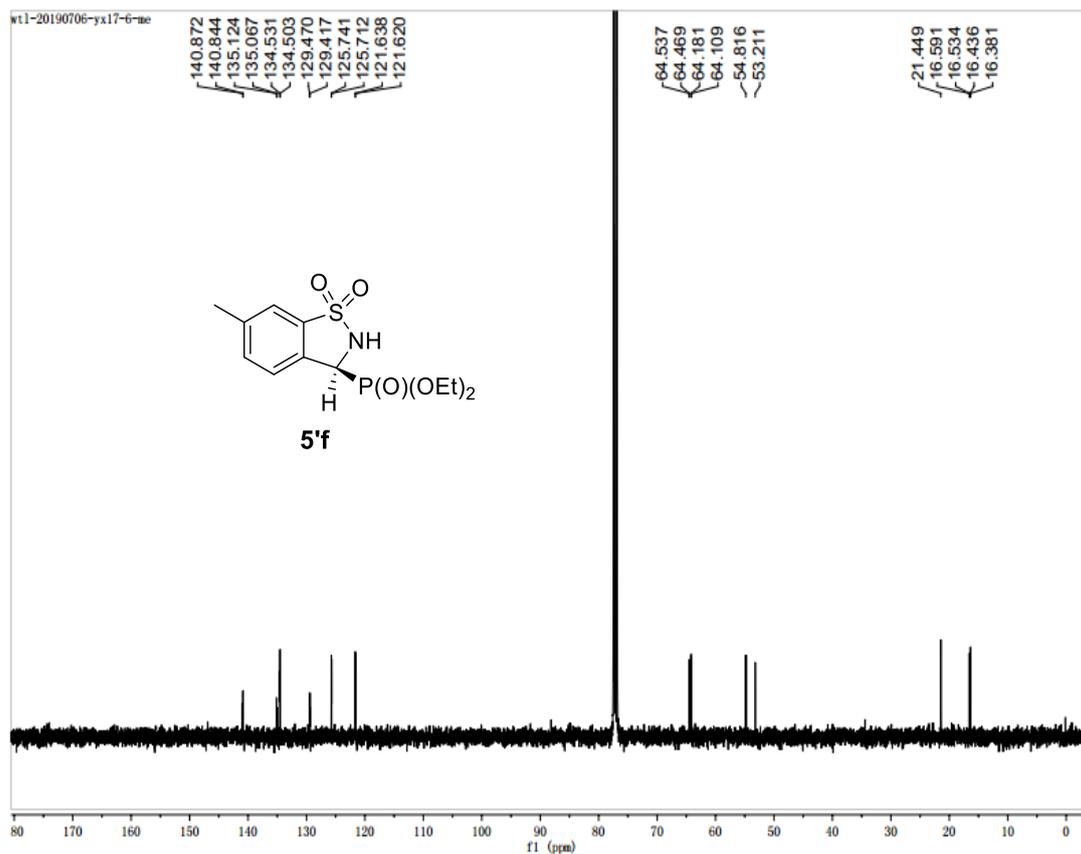


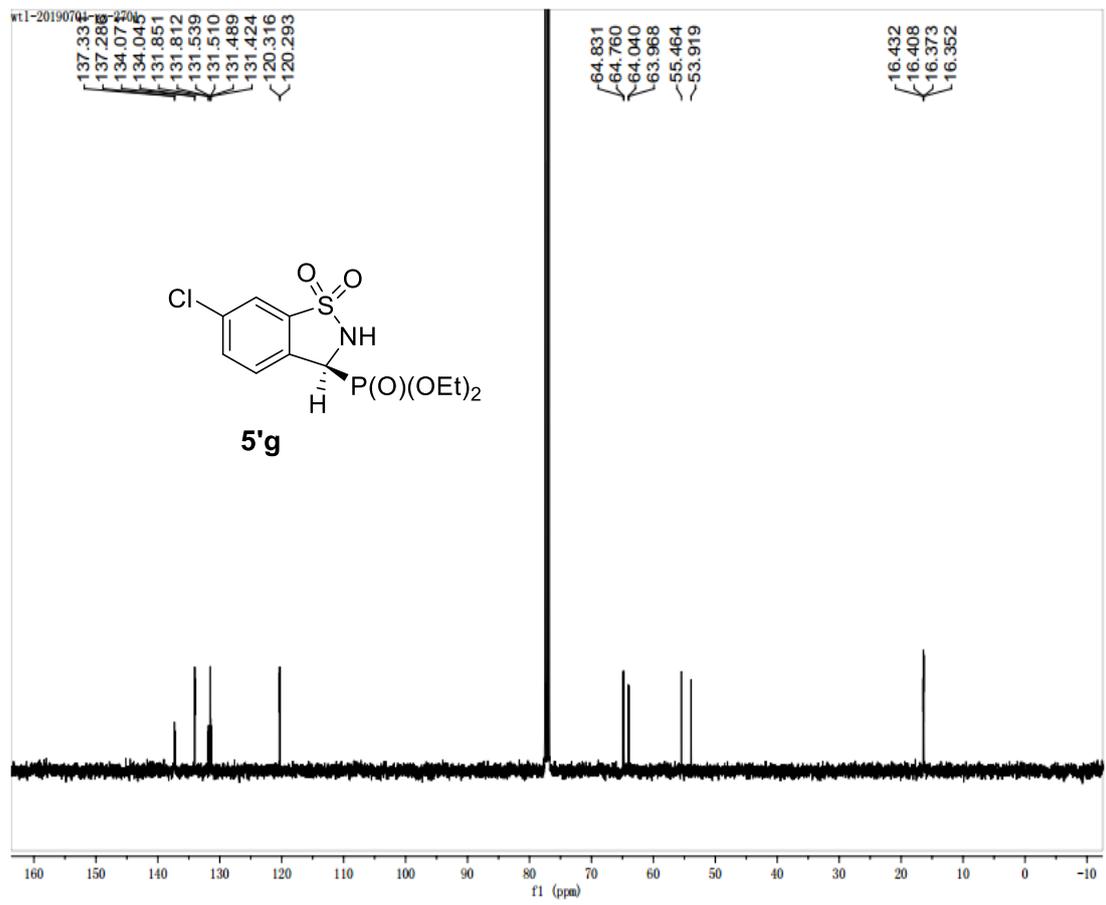
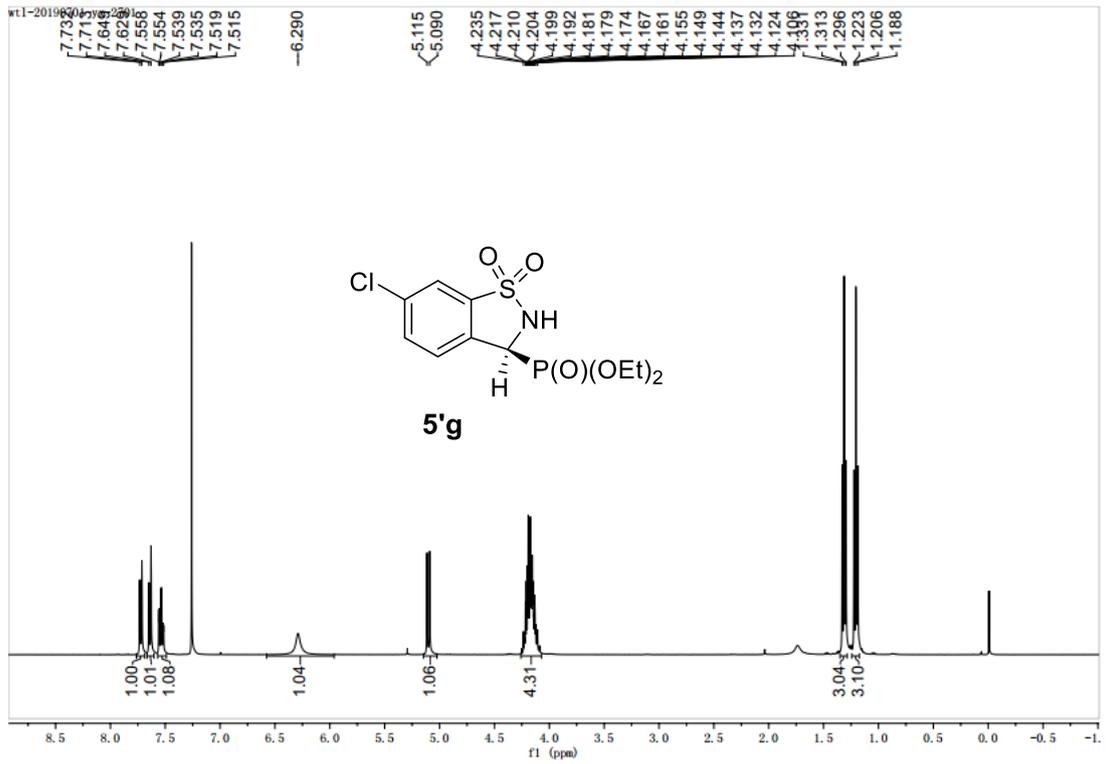
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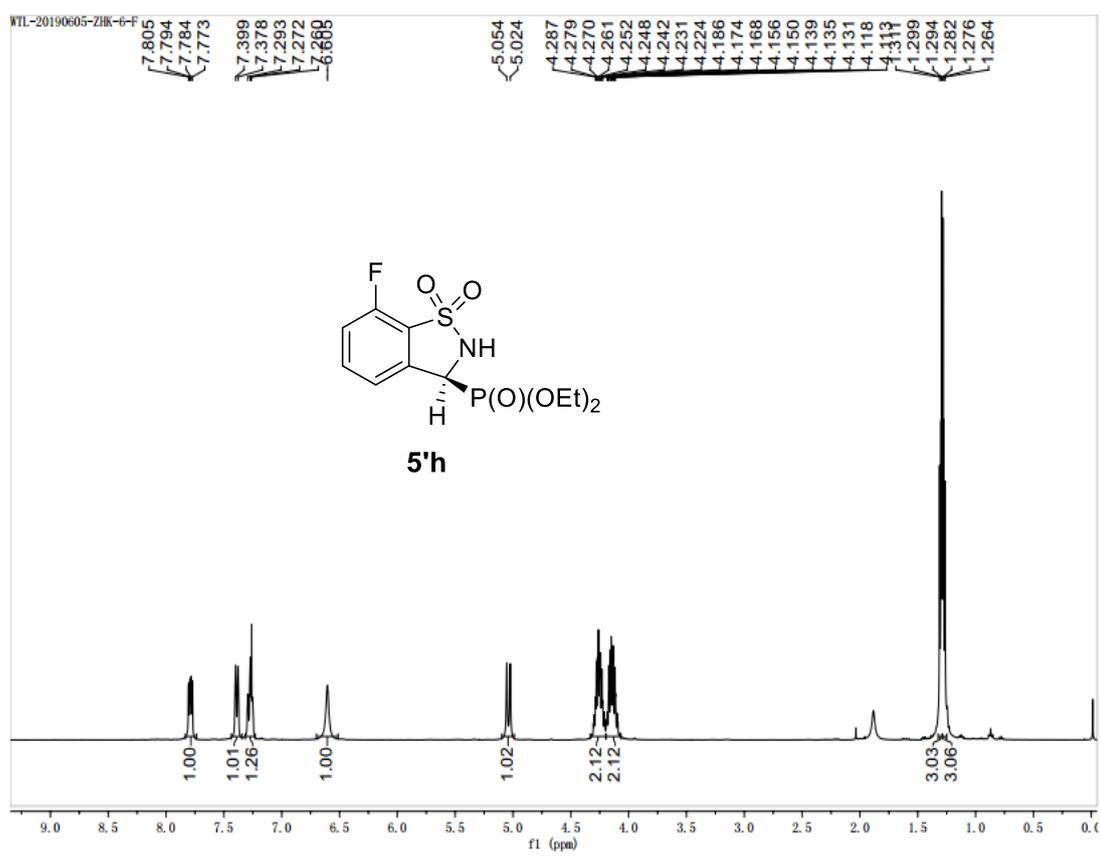
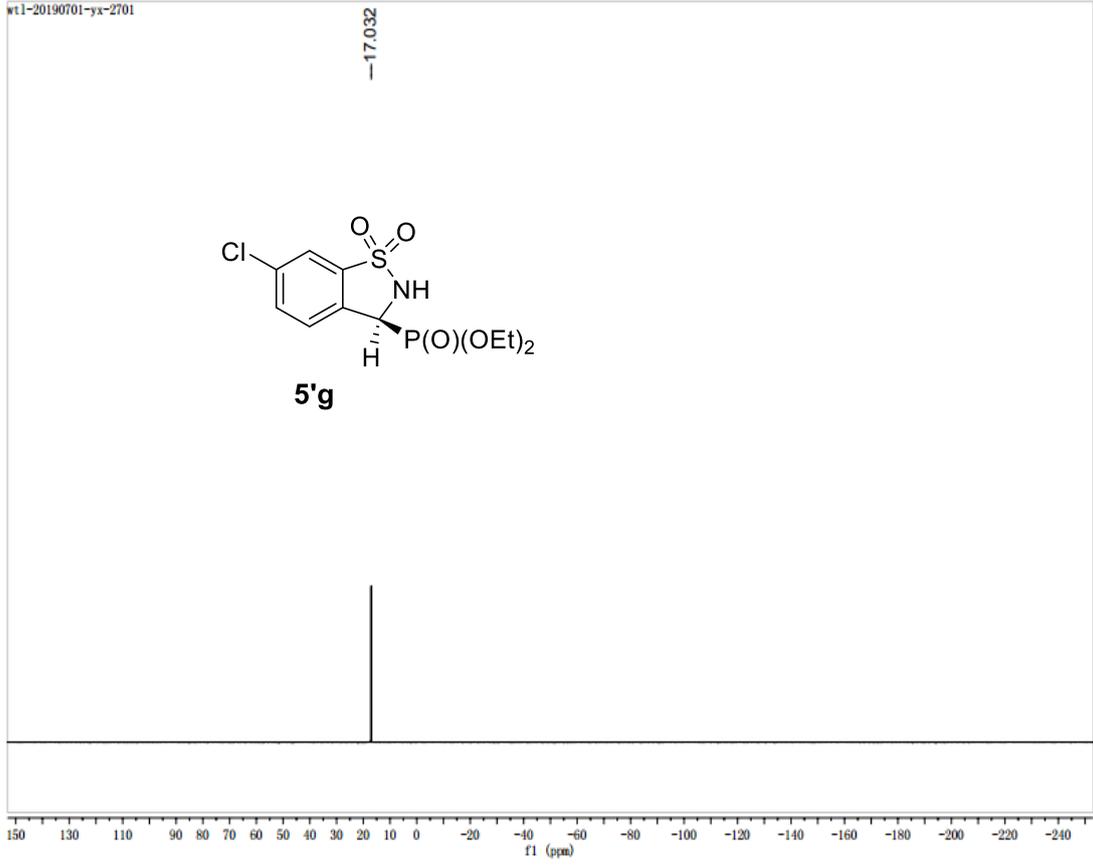


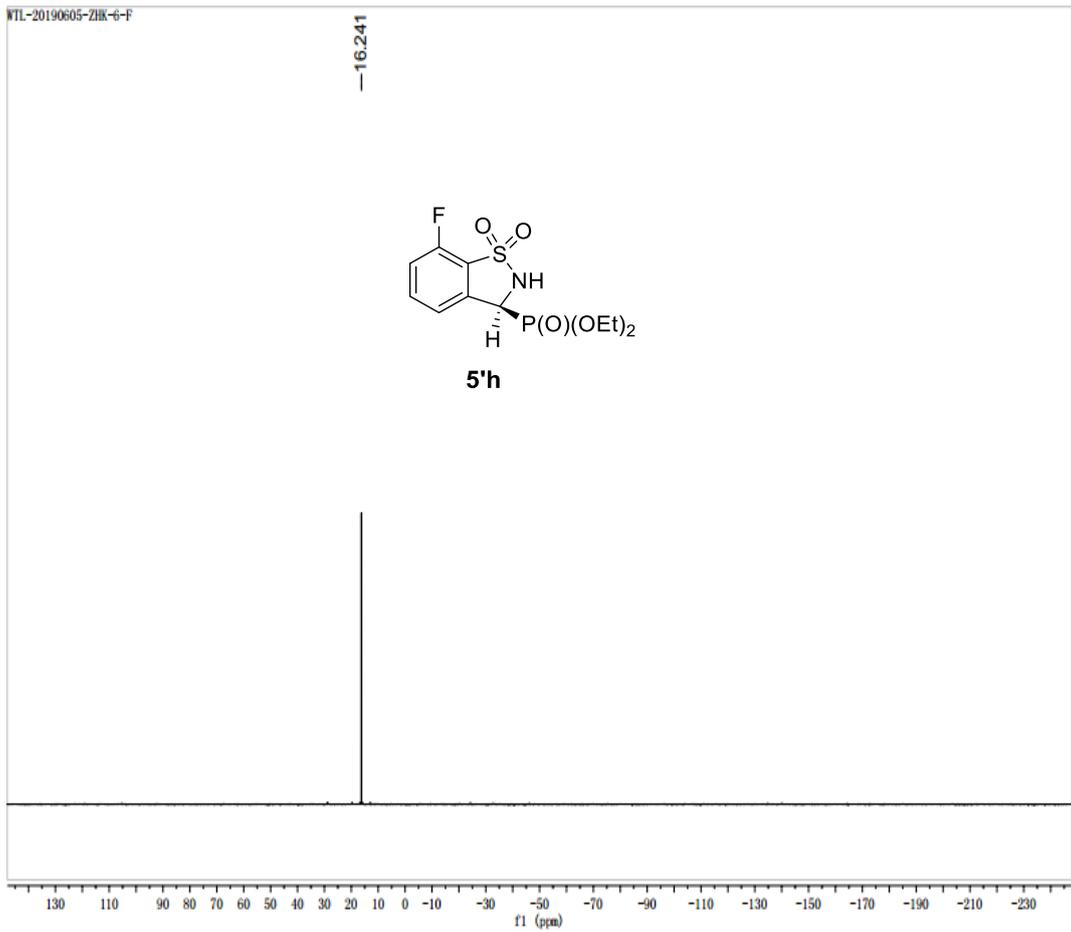
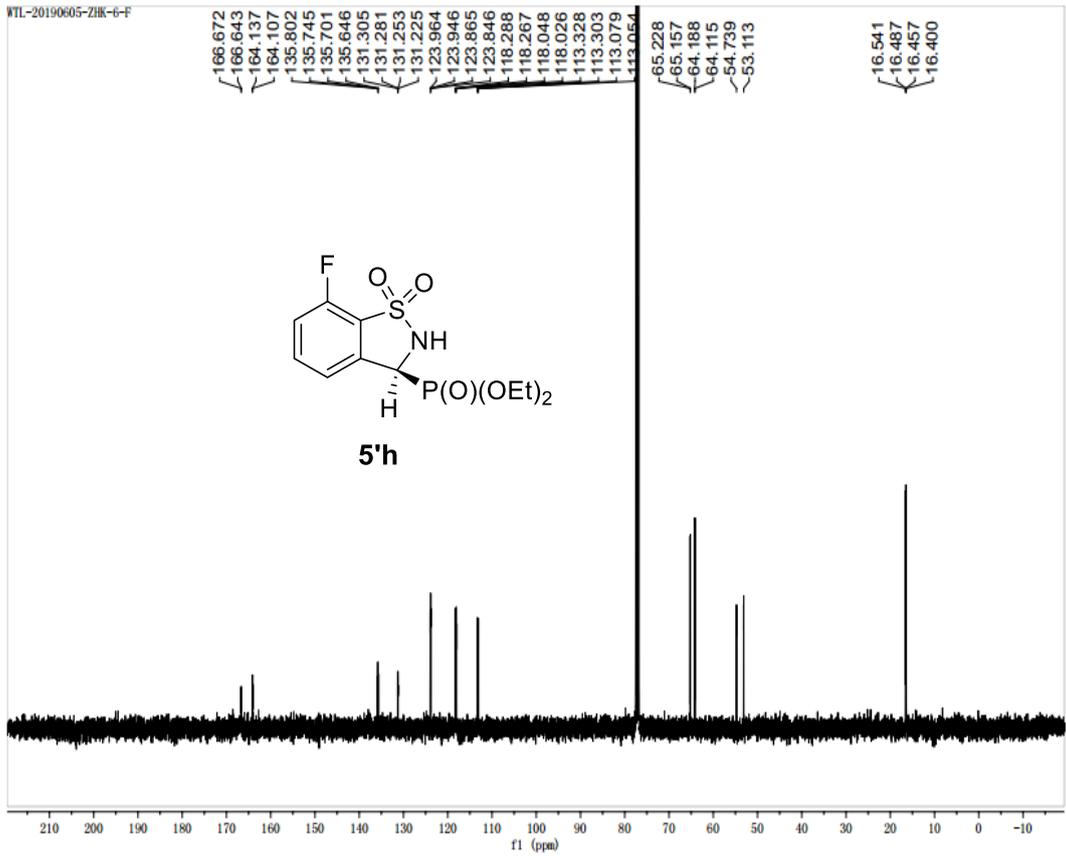




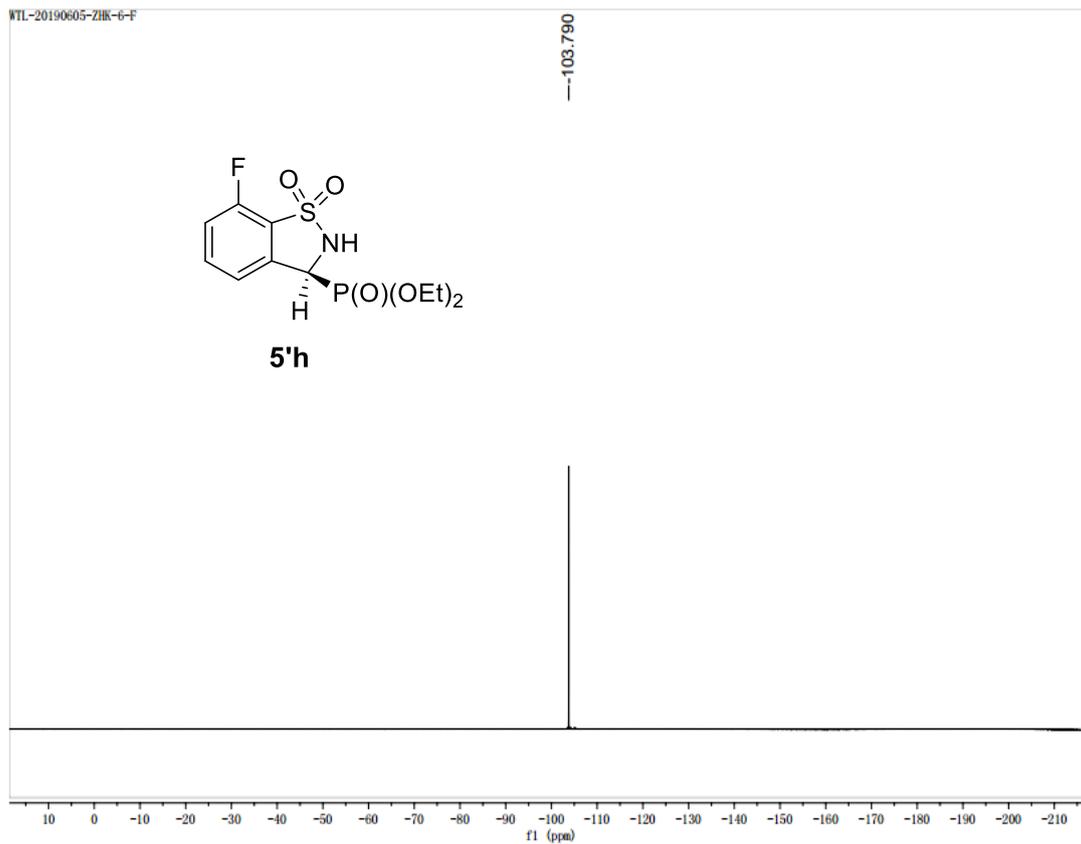




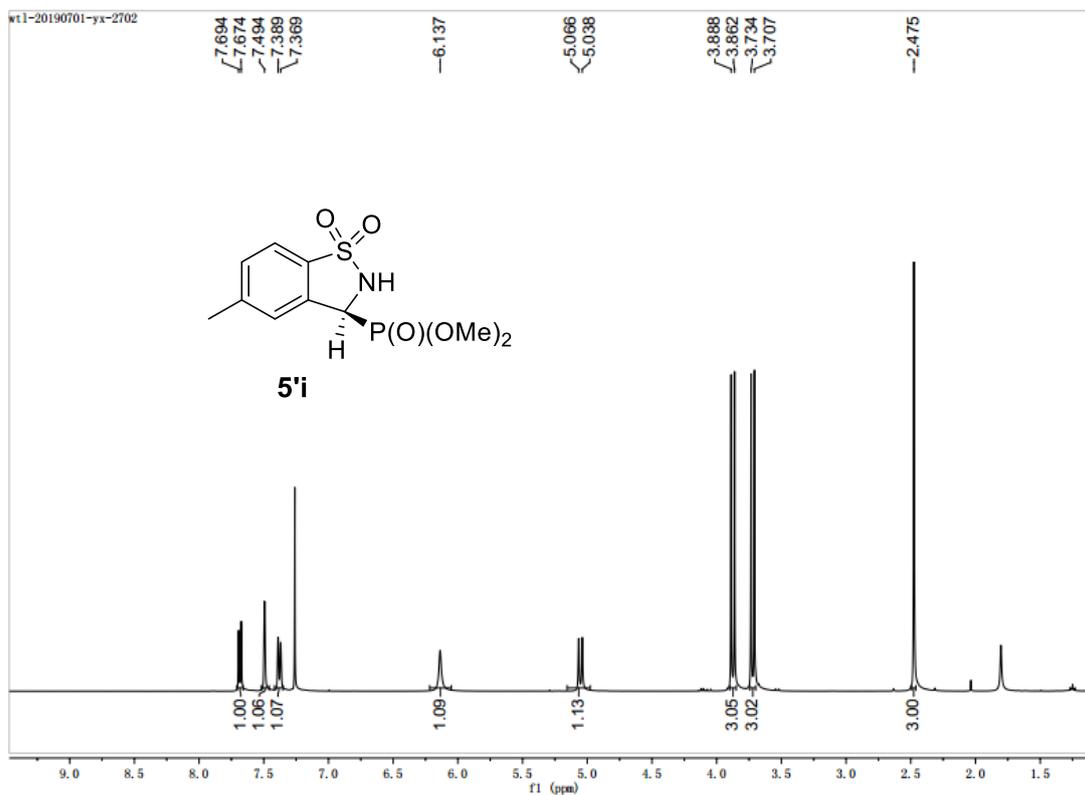


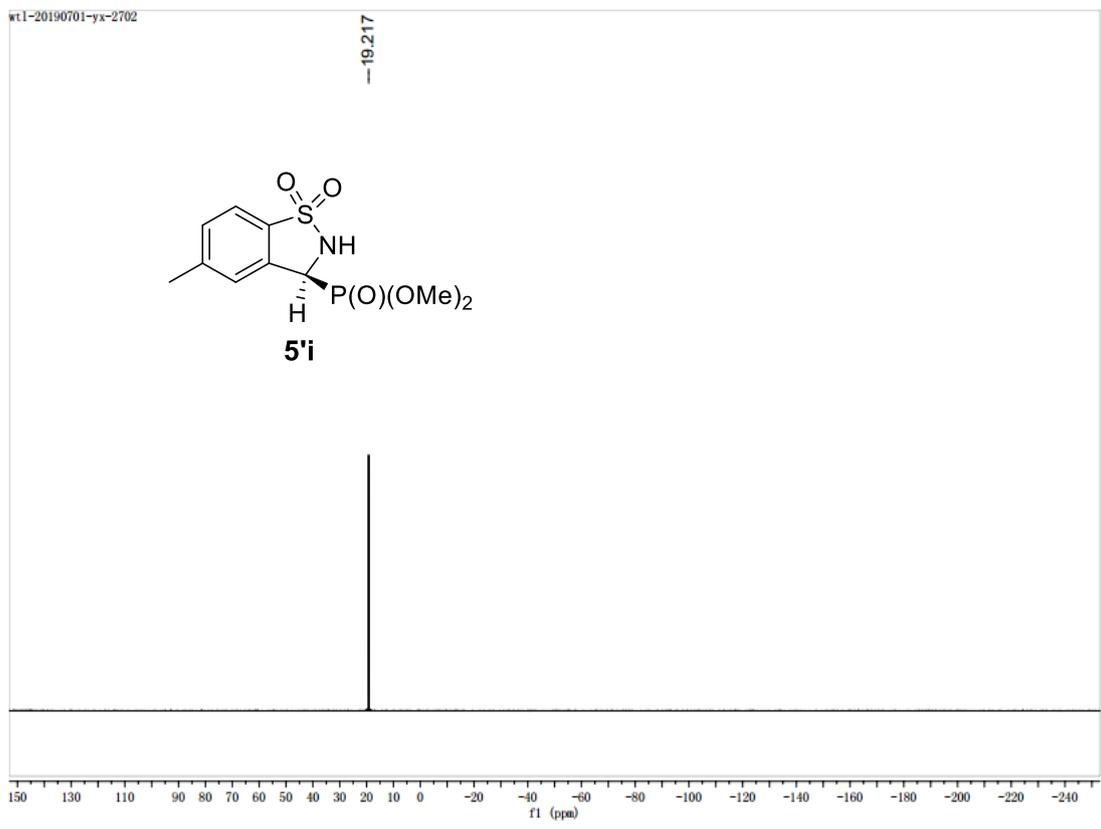
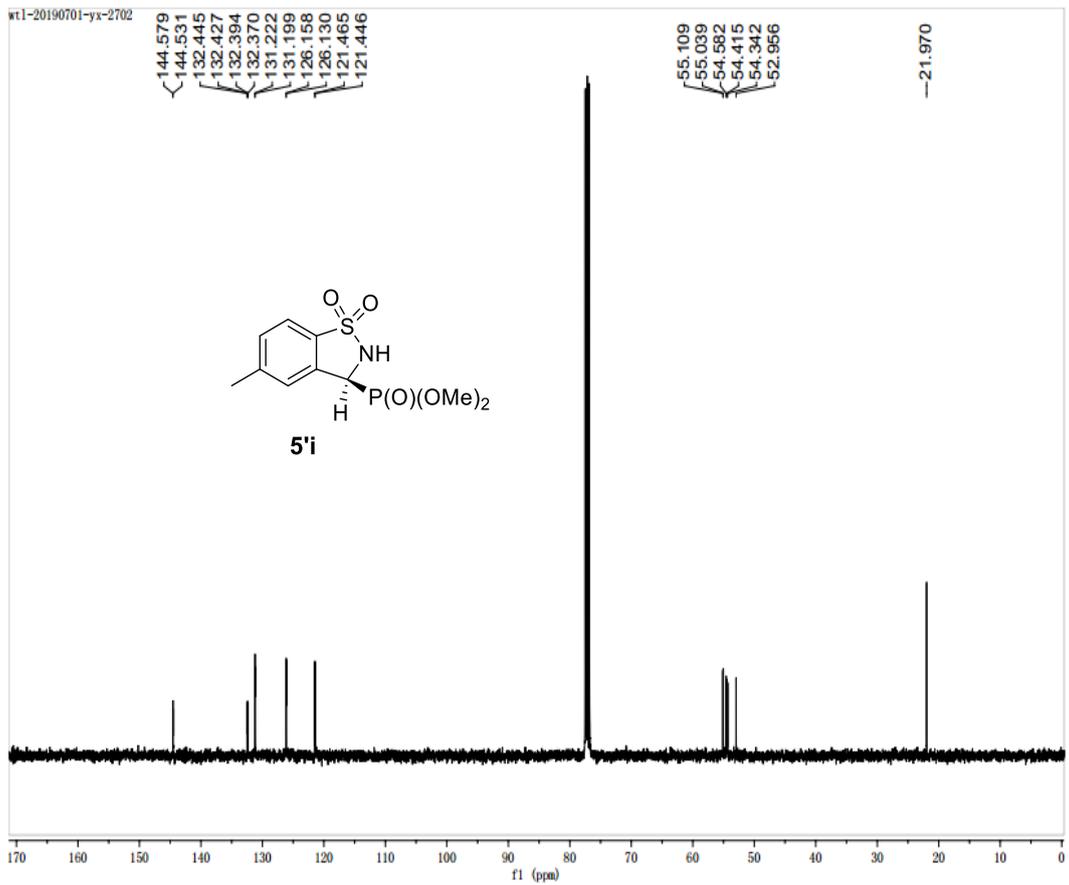


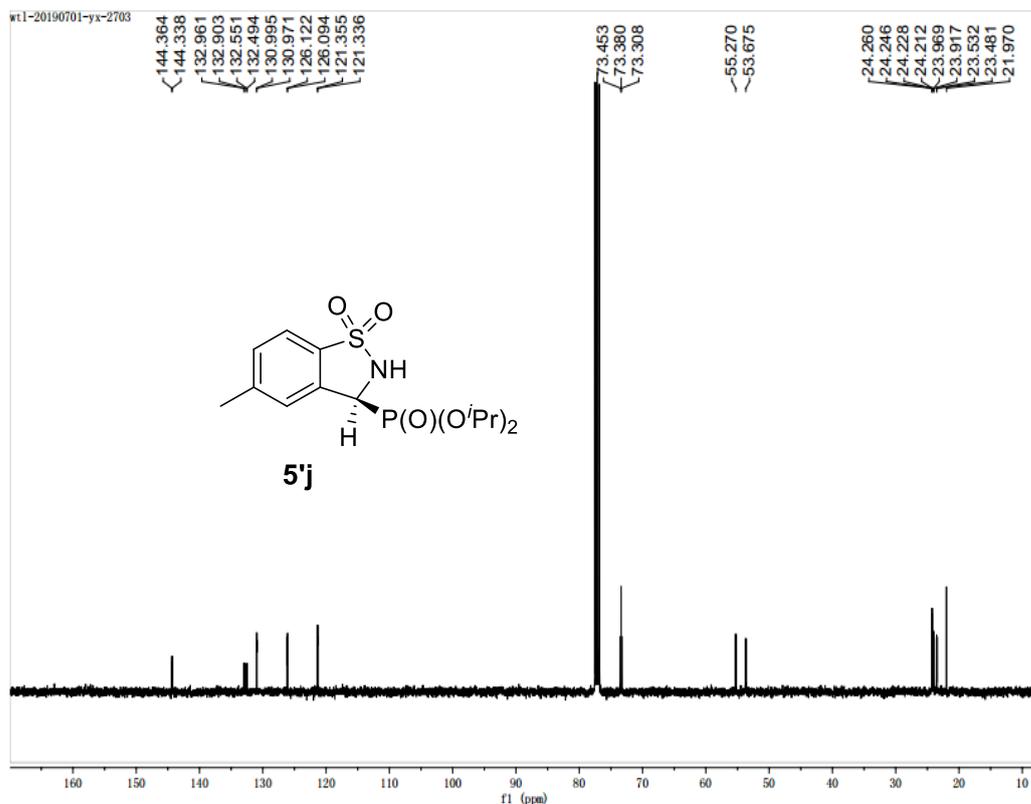
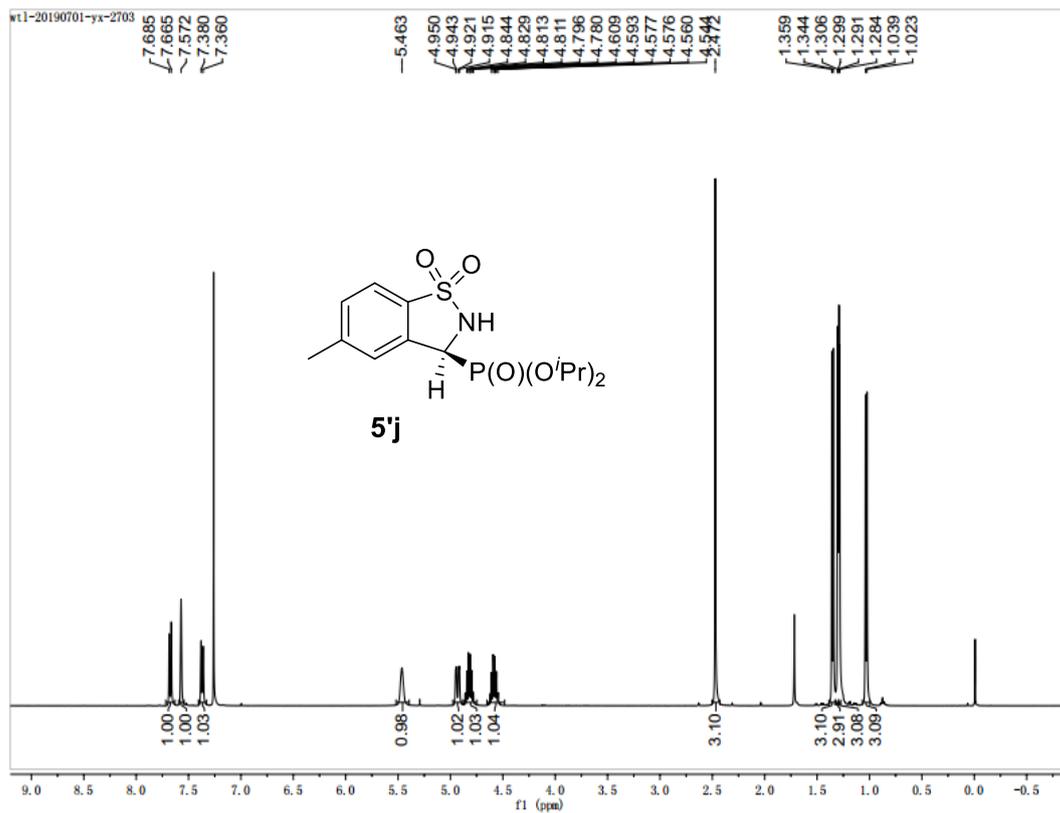
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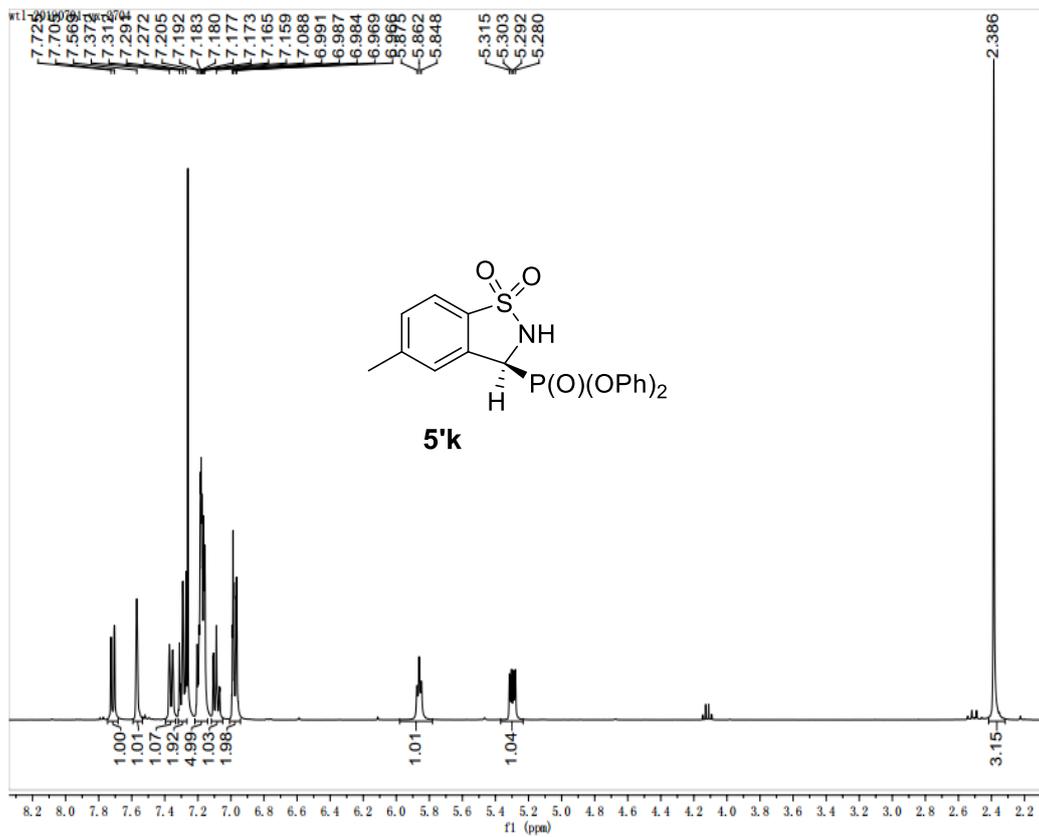
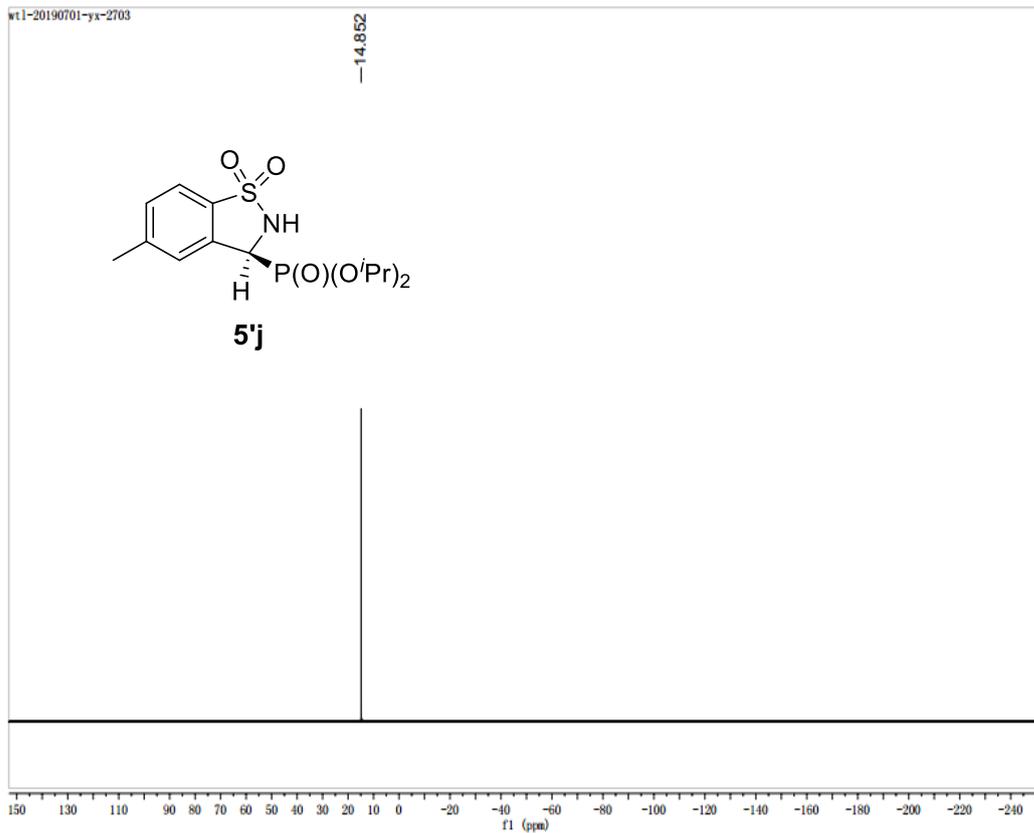


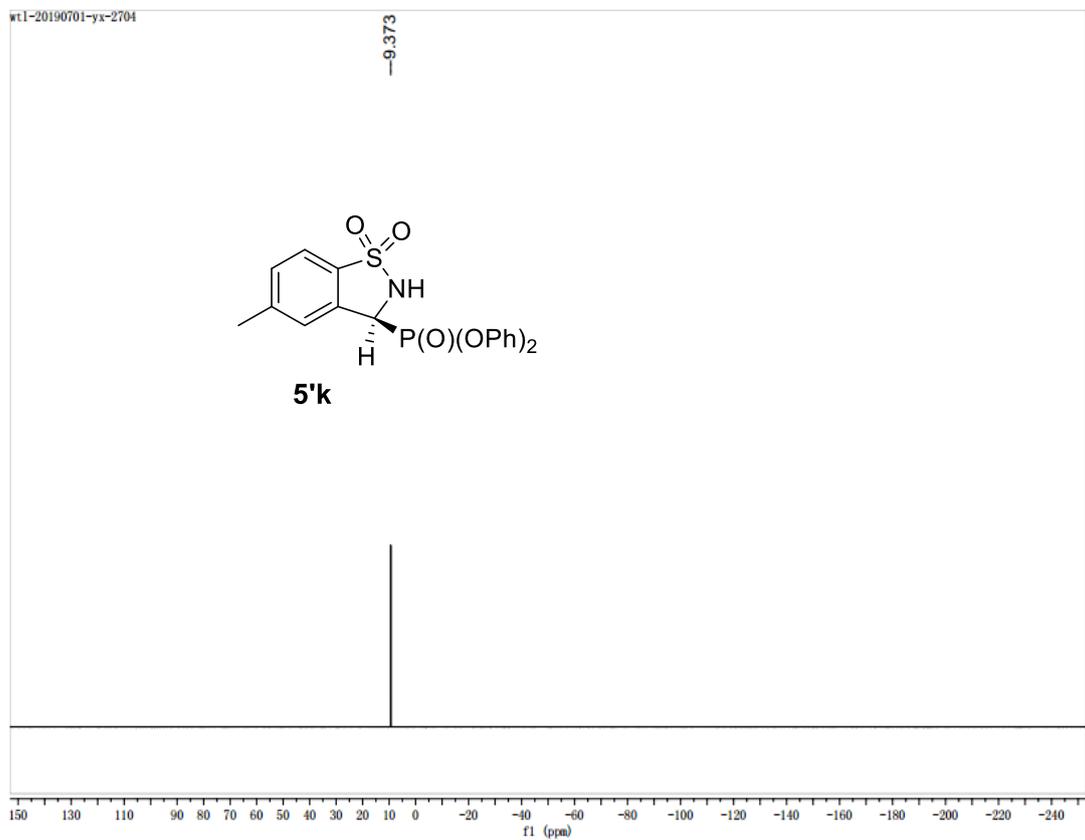
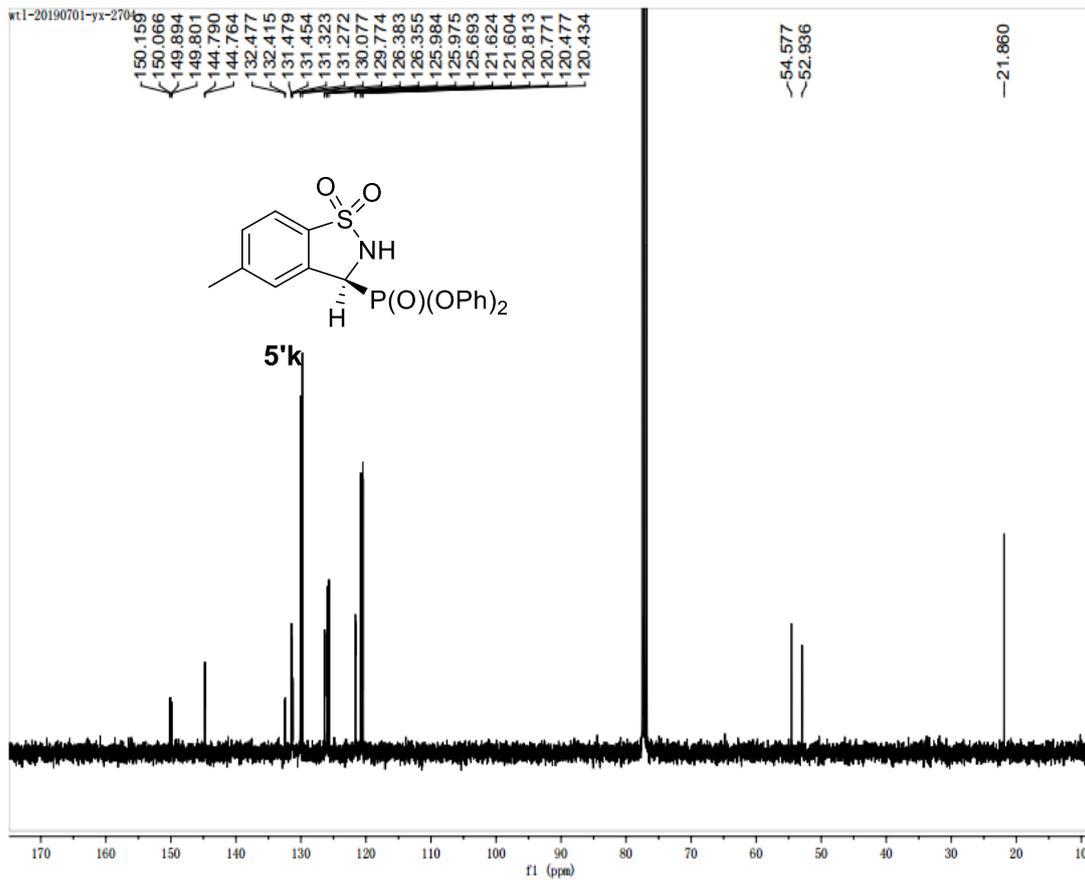
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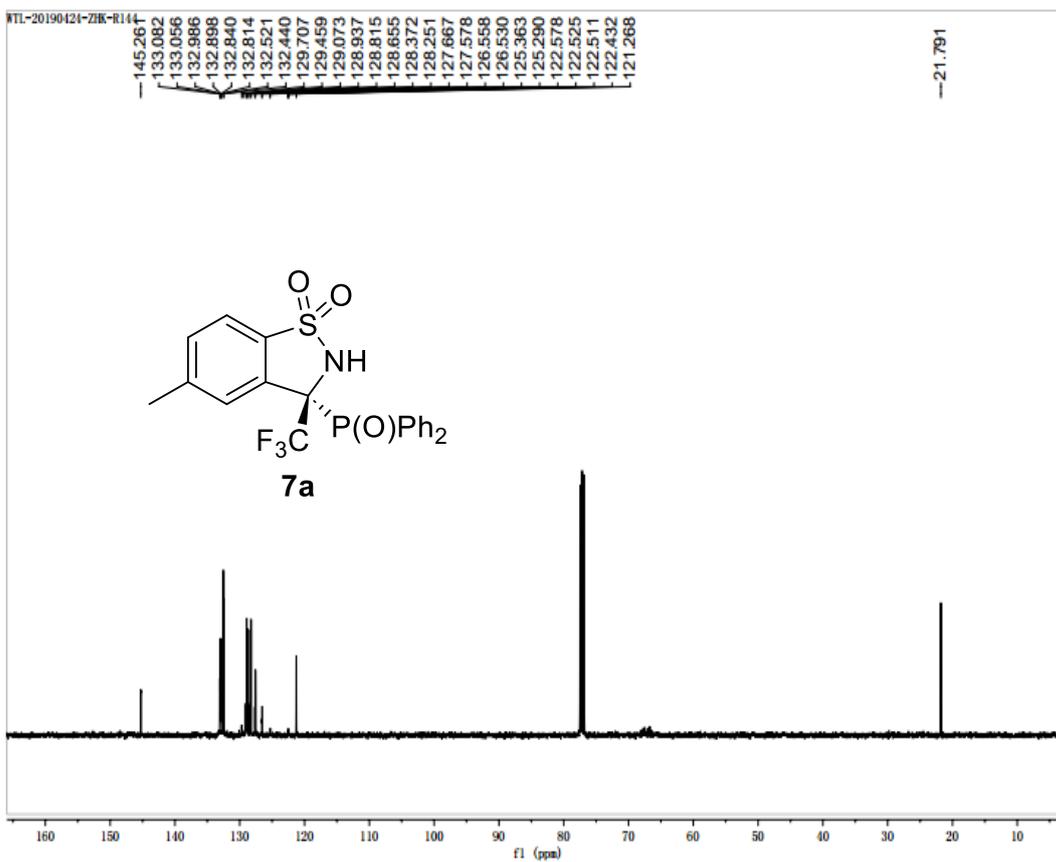
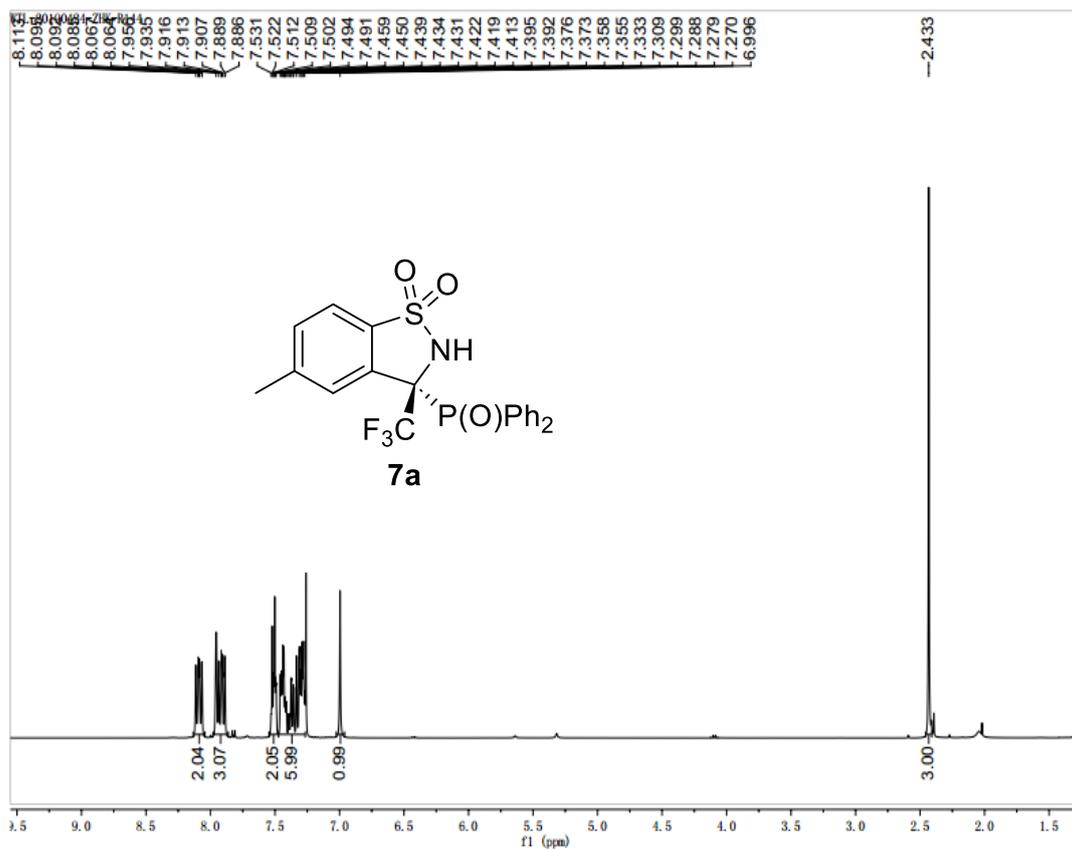




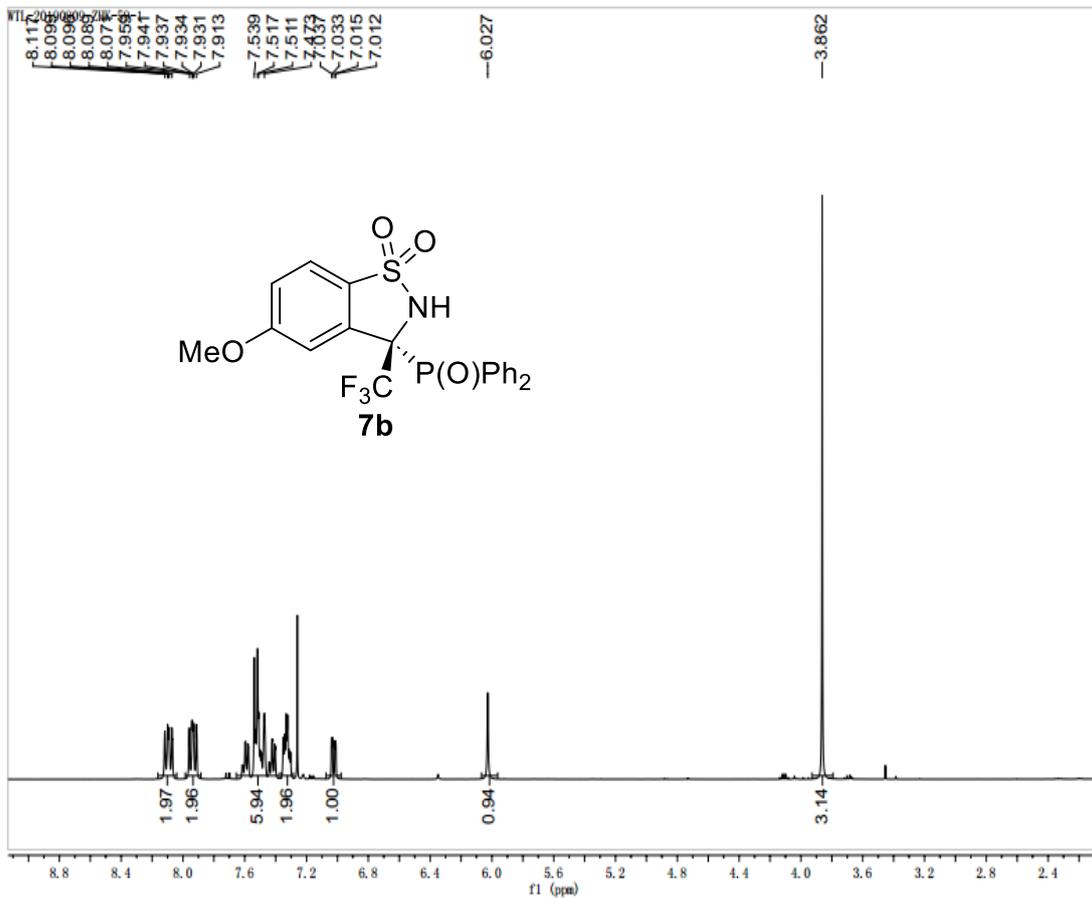
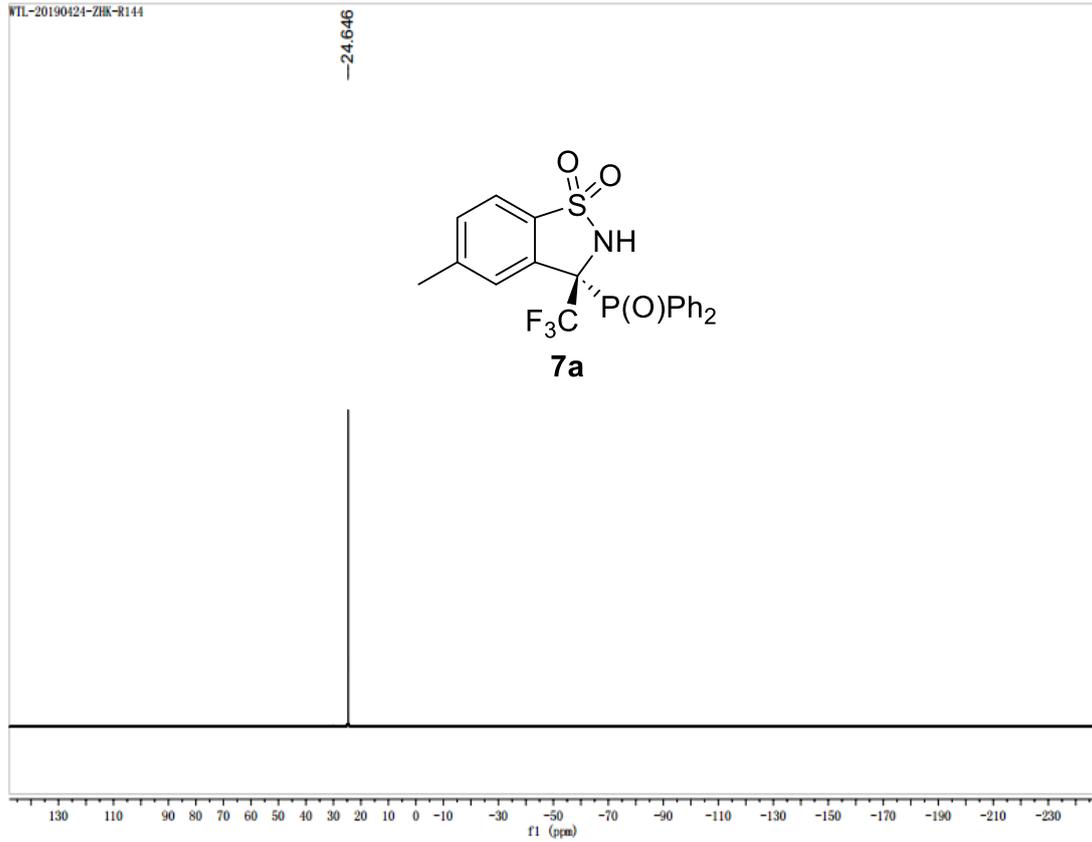


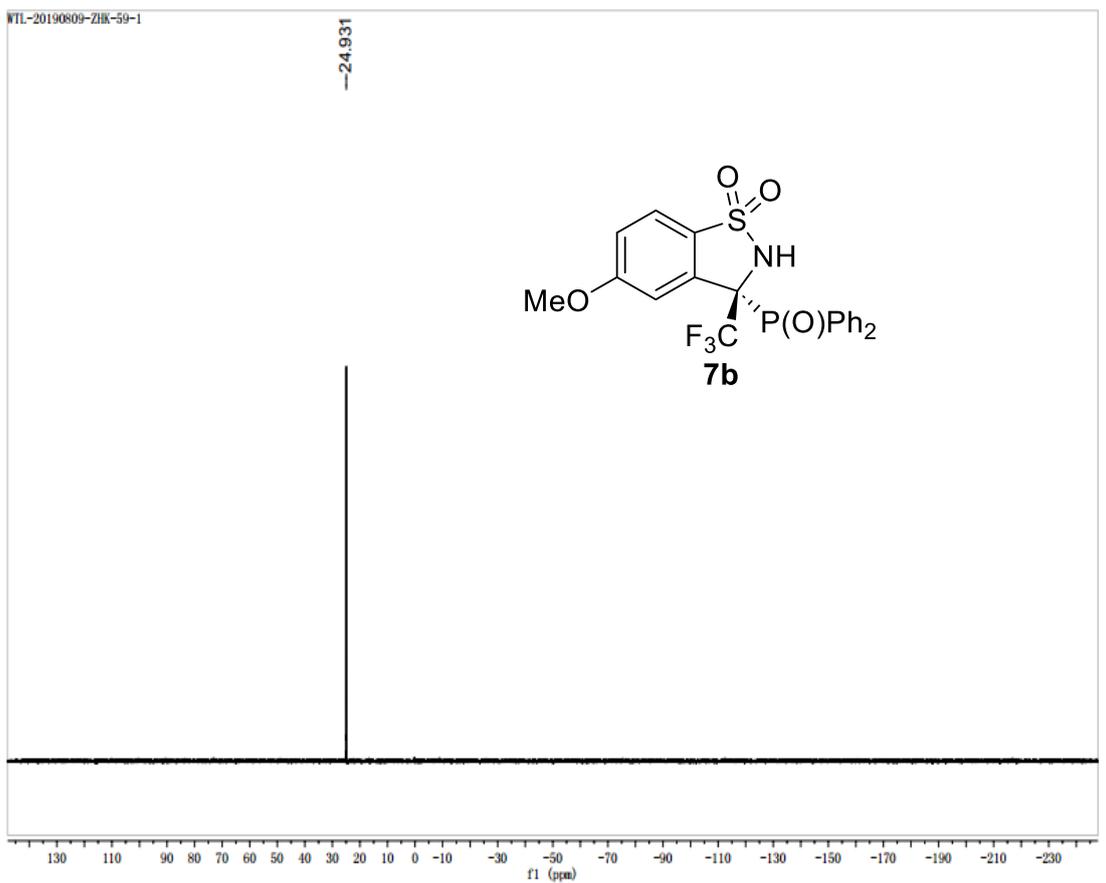
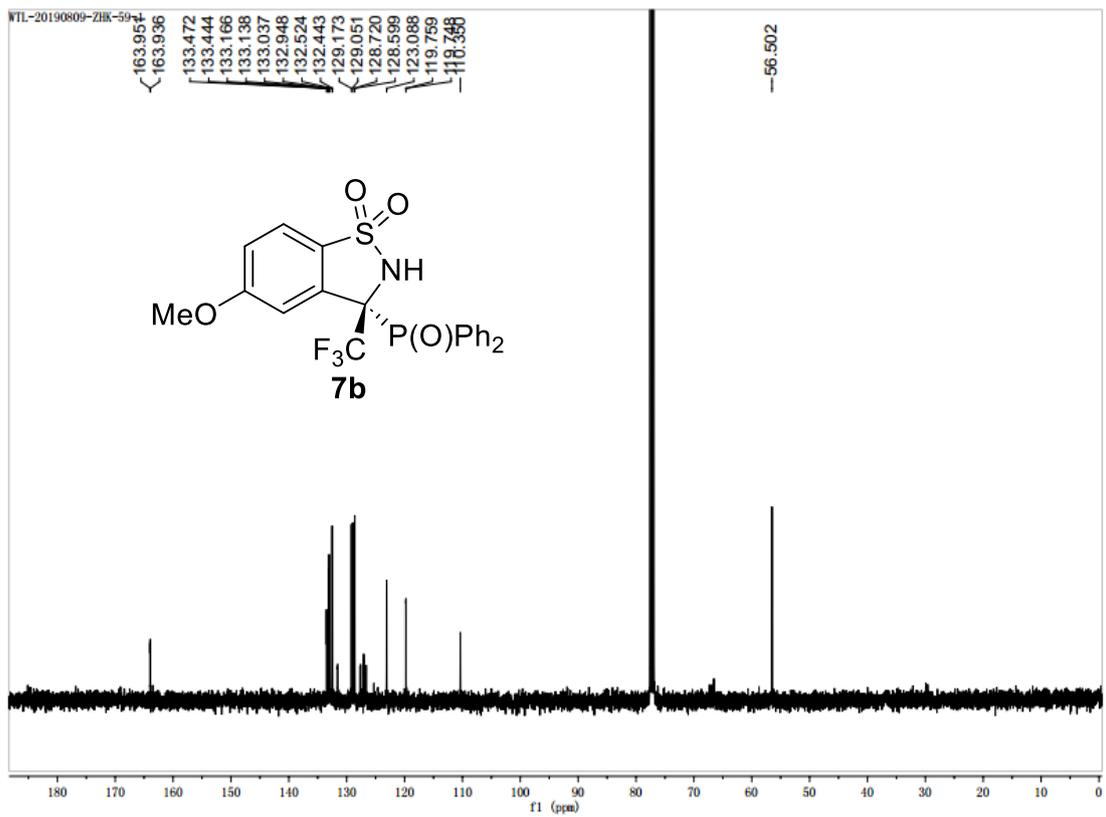


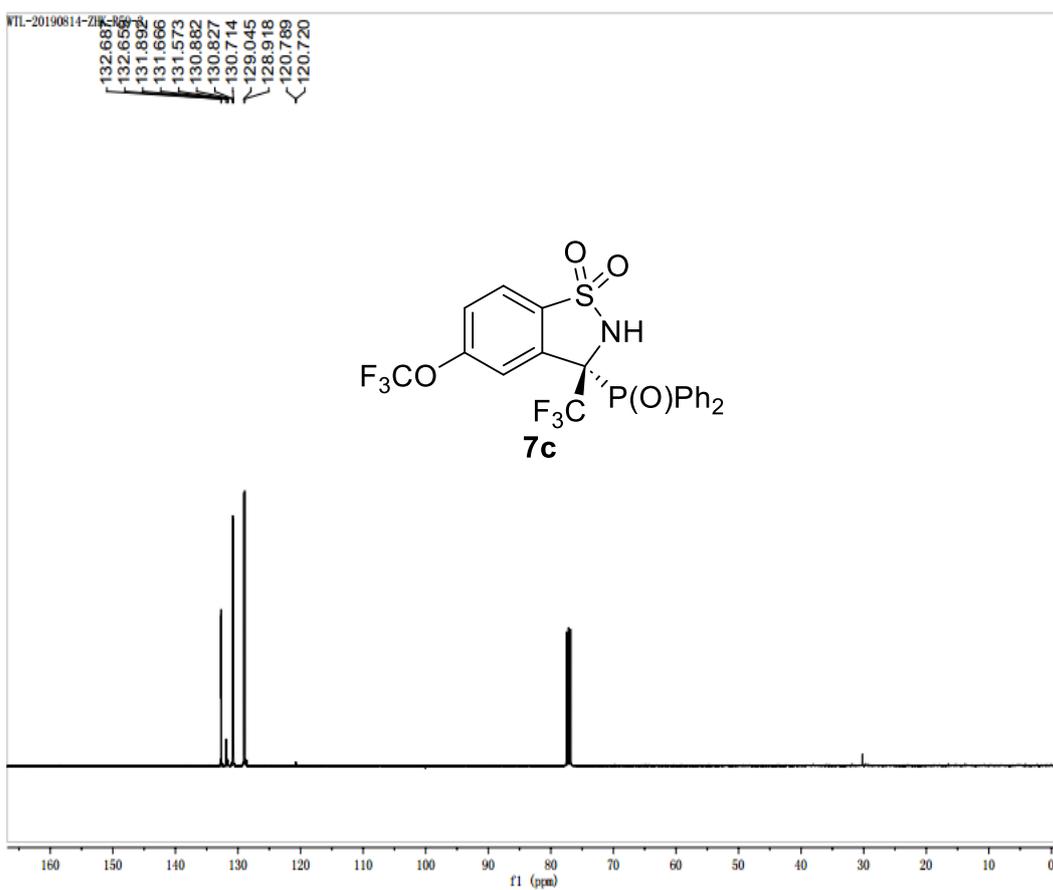
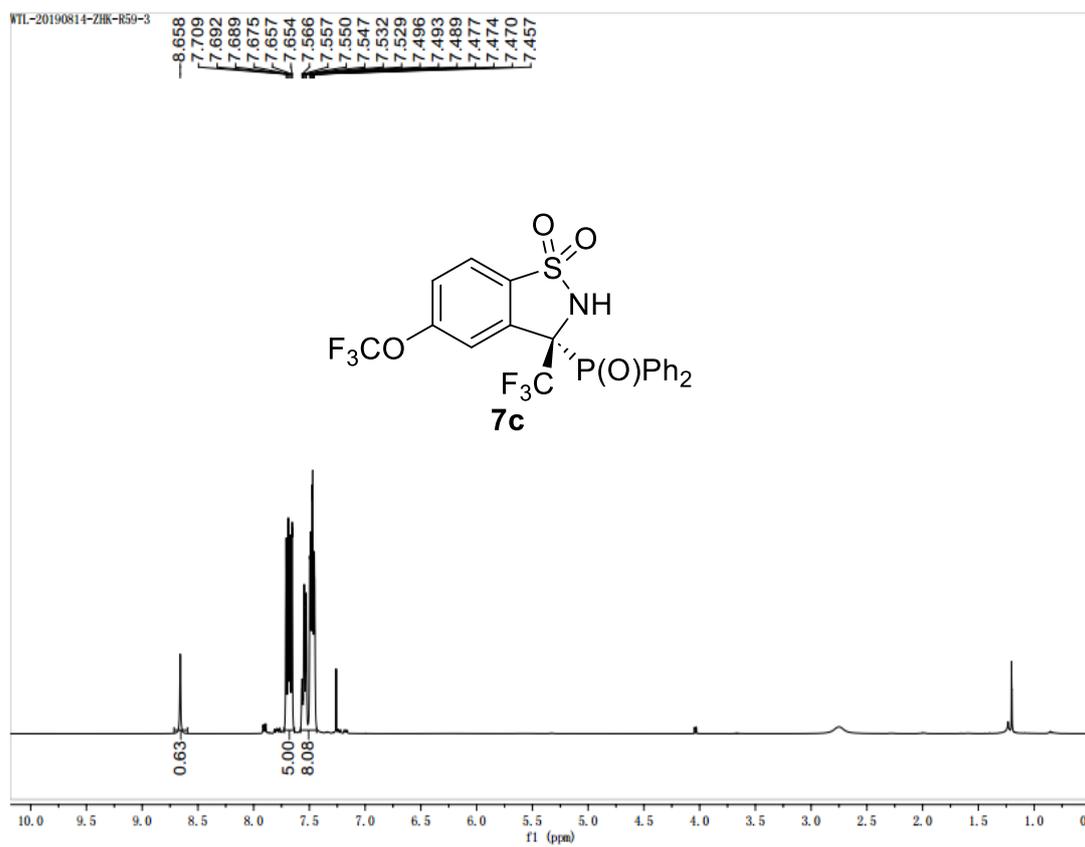




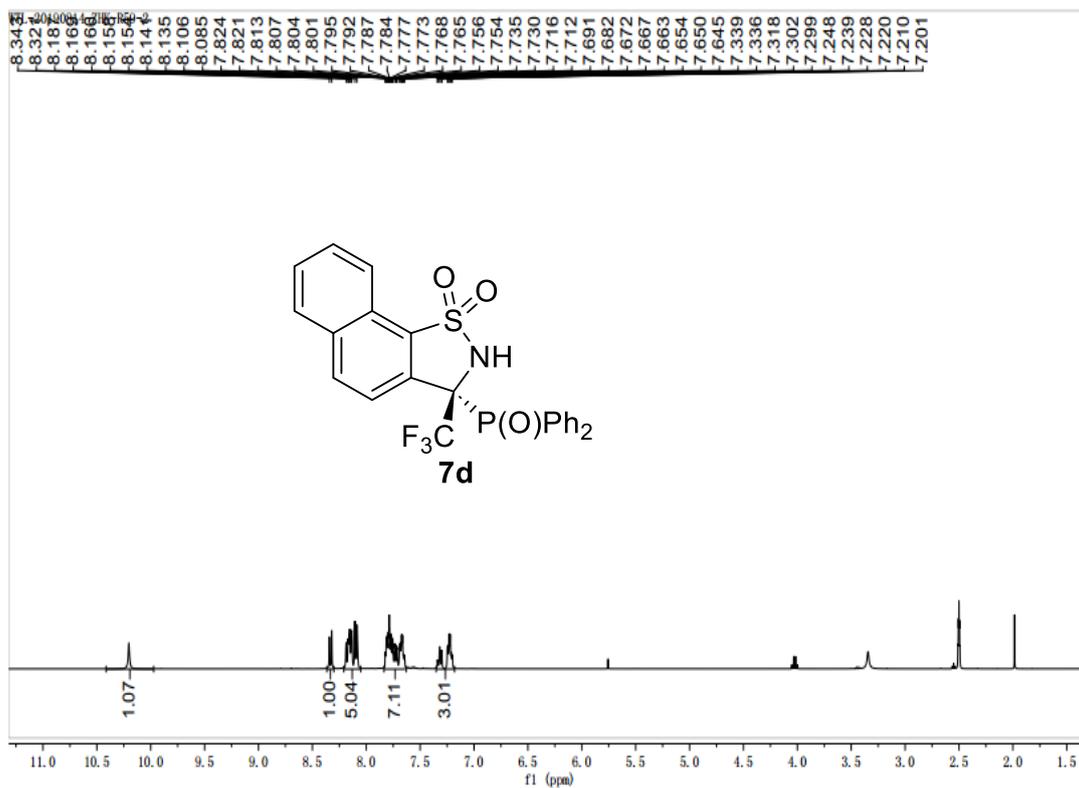
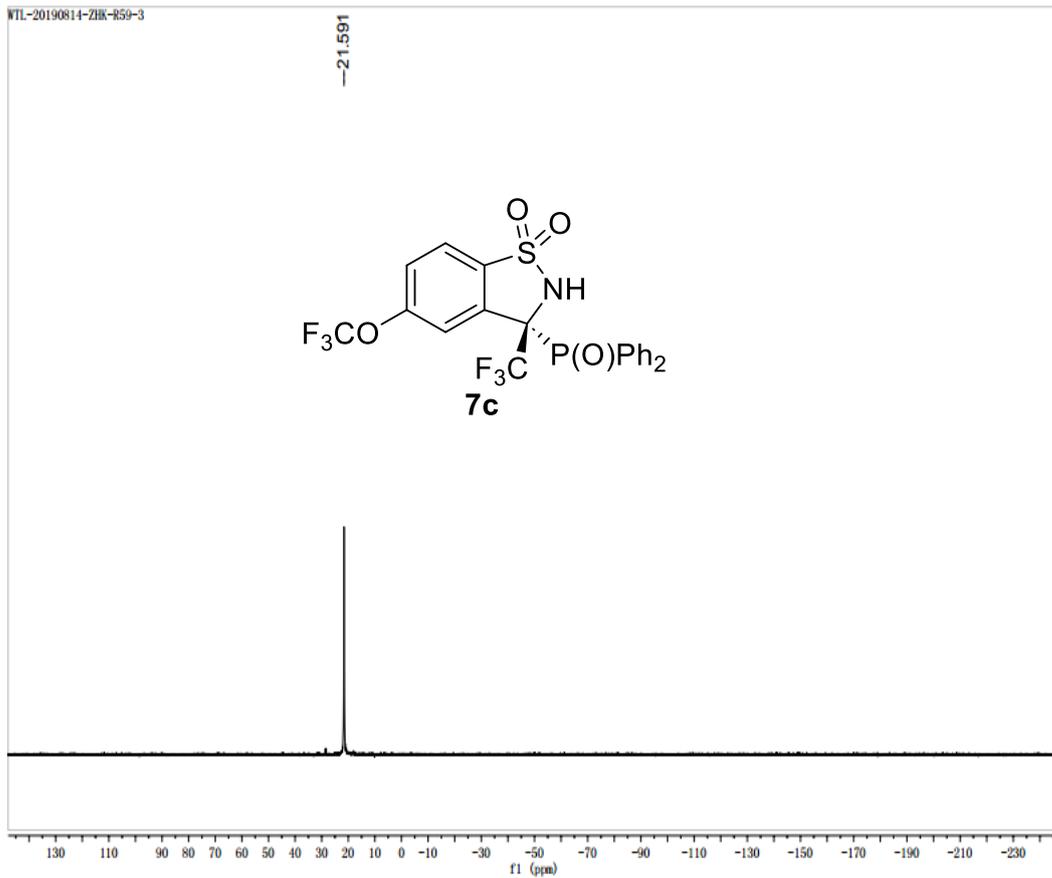
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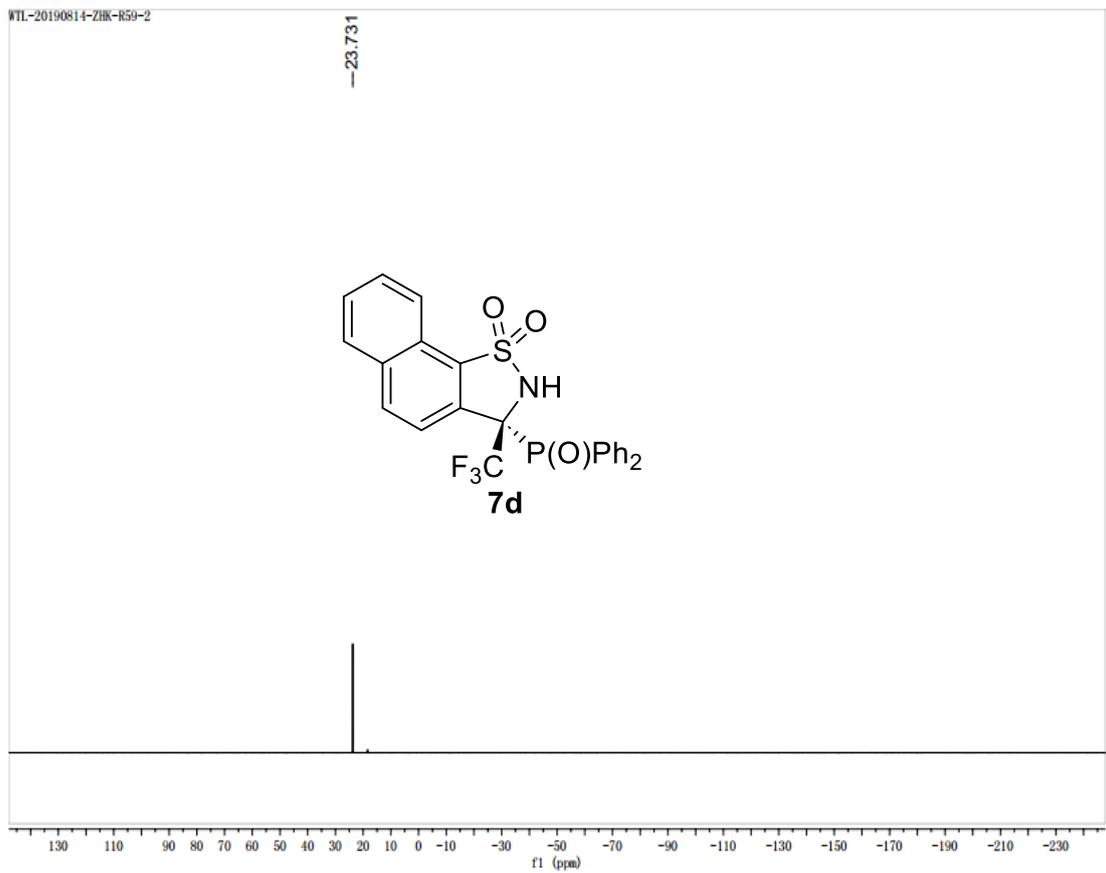
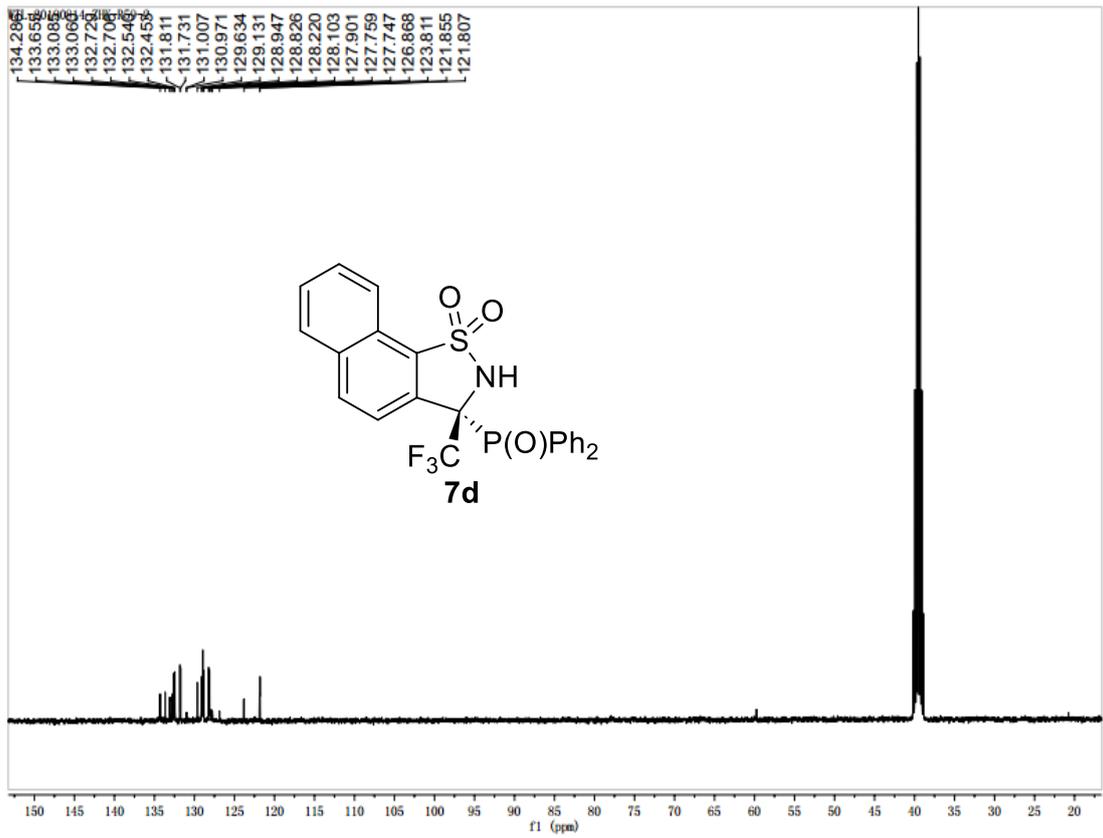


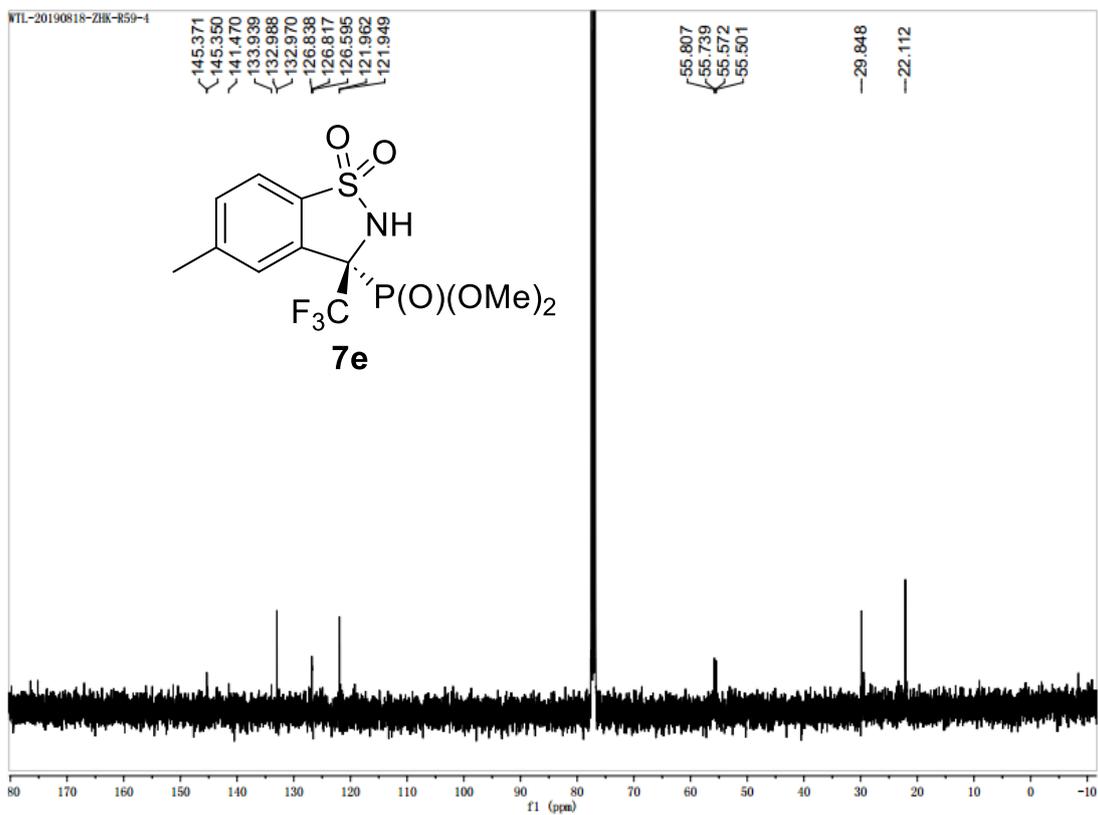
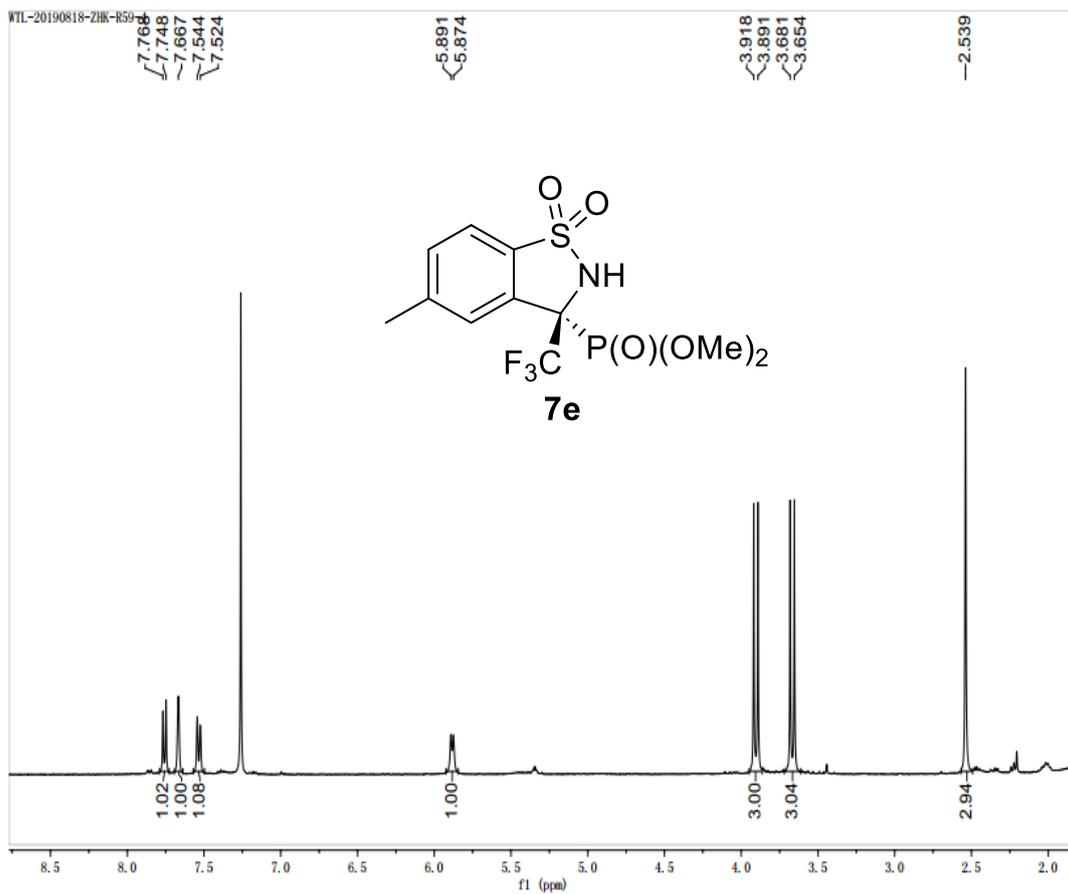


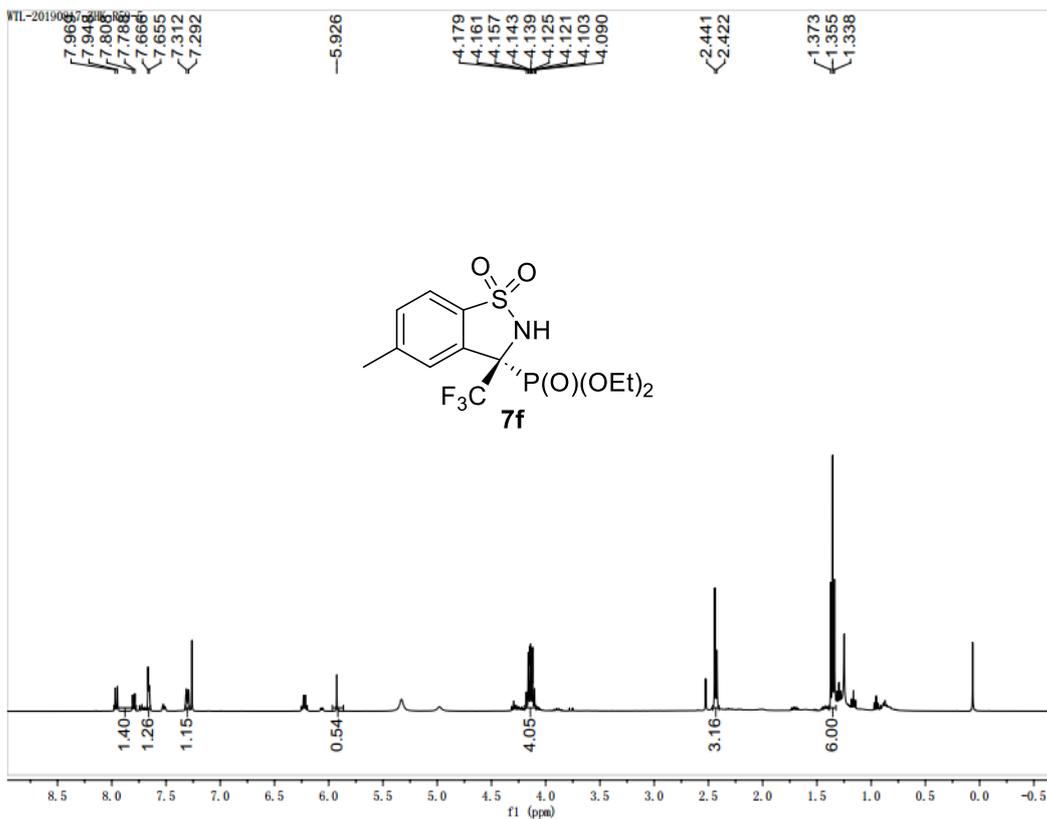
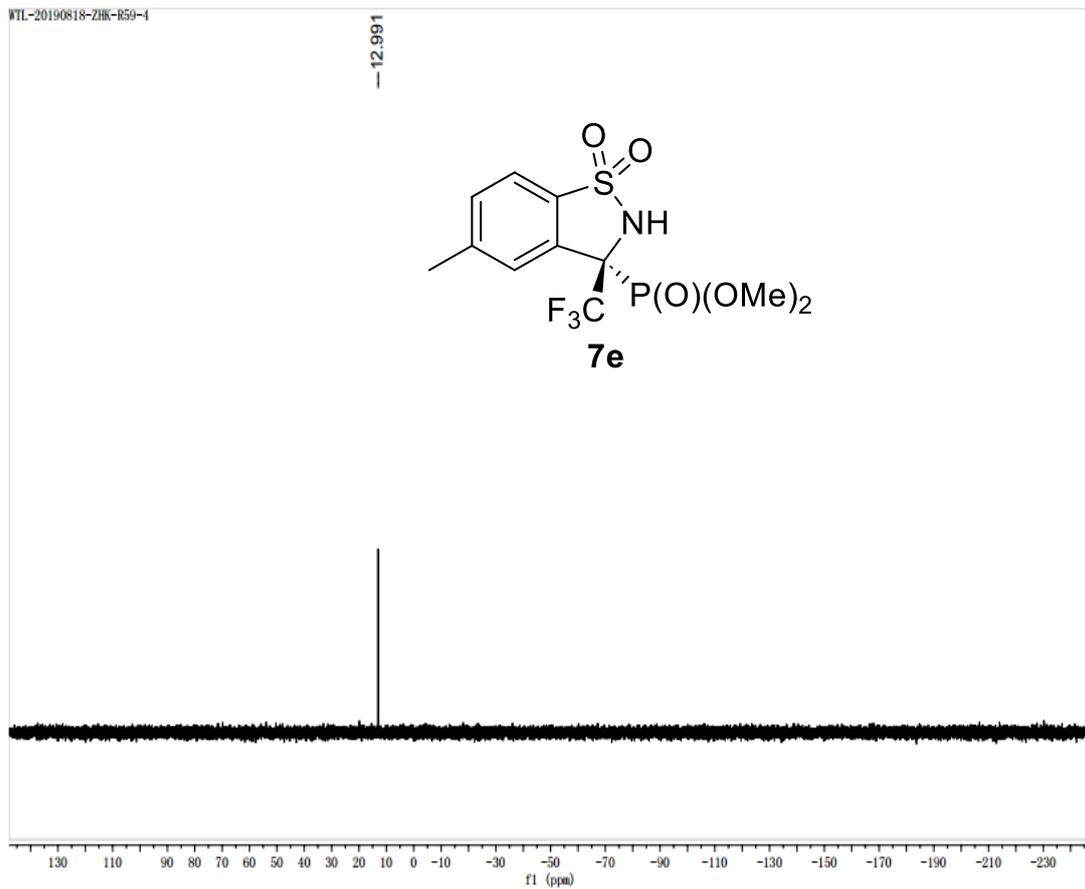


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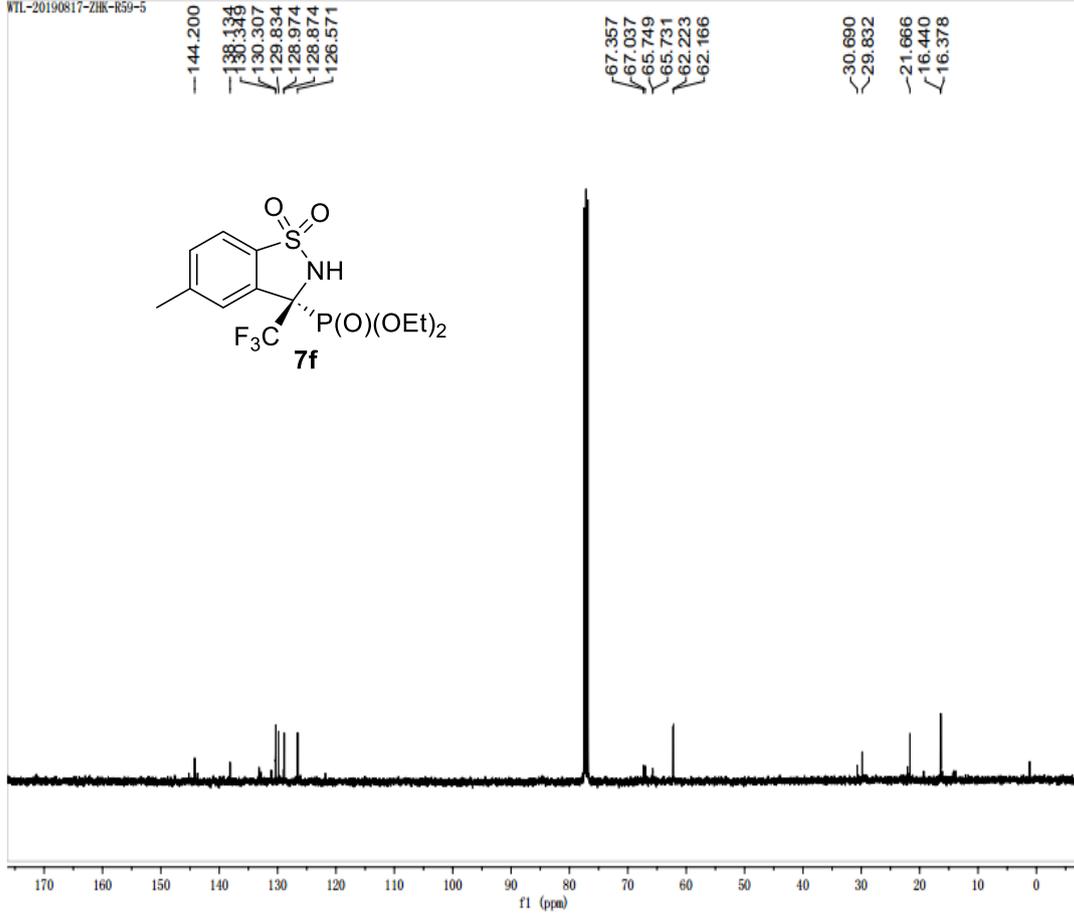




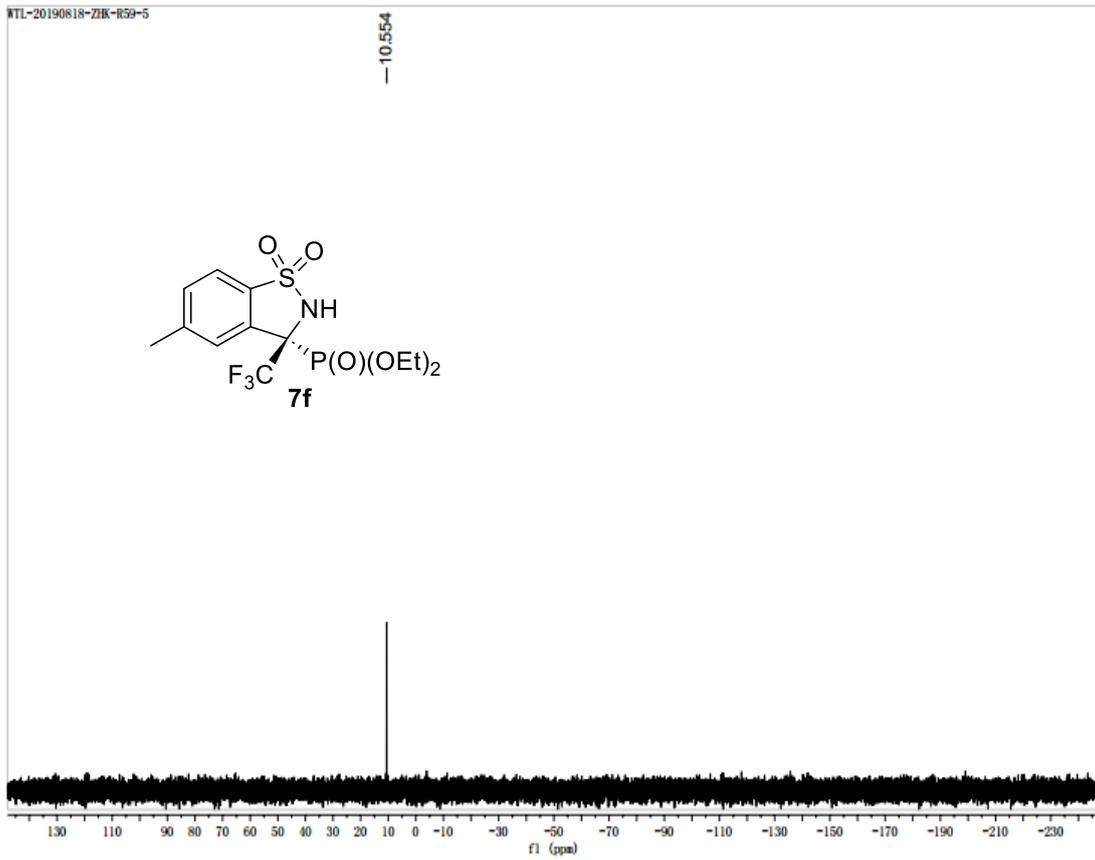


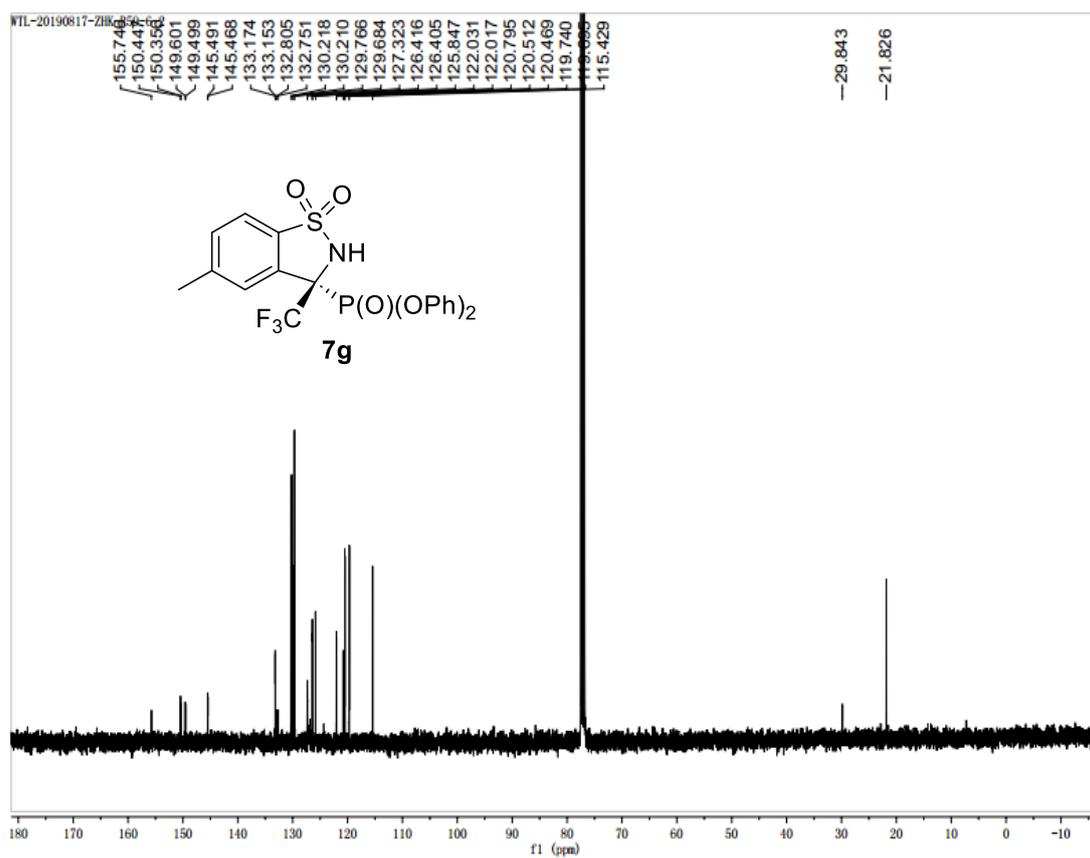
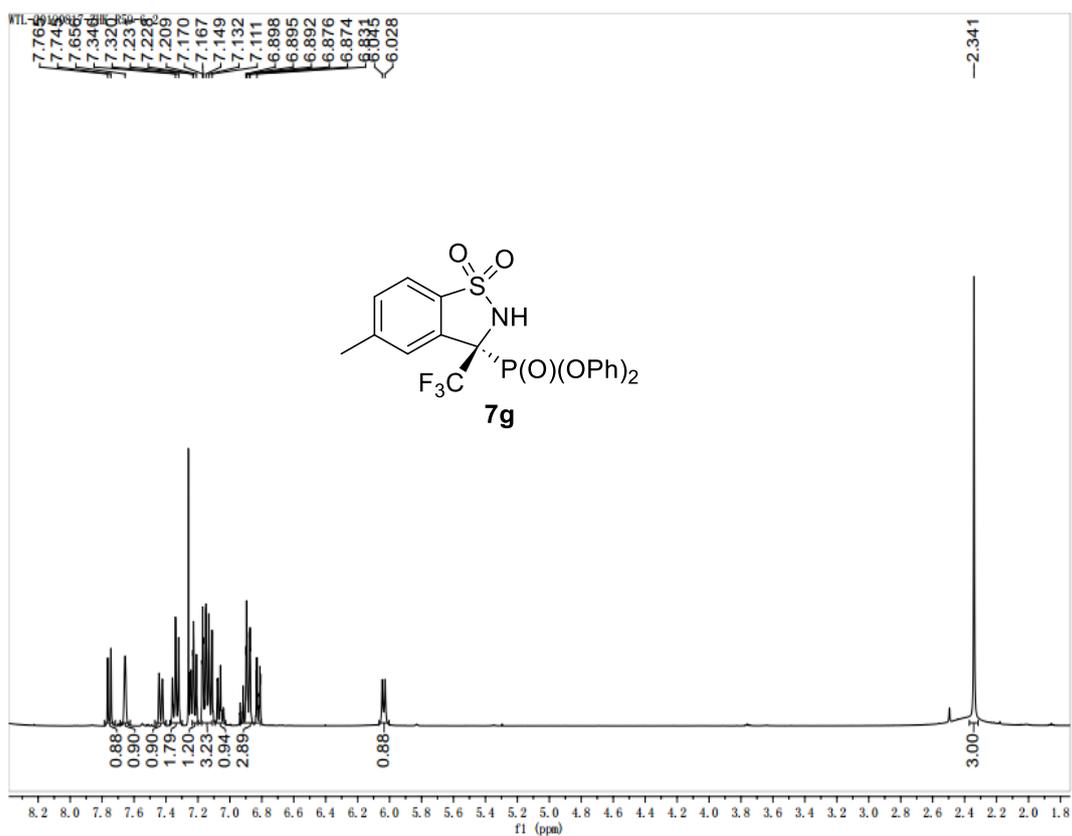


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WTL-20190818-ZHK-R59-5





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