

Supporting Information for

Integrated “all-in-one” strategy to construct highly efficient Pd catalyst for CO₂ transformation

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1. Experimental Section

1.1 General

All nuclear magnetic resonance (NMR) spectra were acquired using a Bruker Advance III 400 NMR spectrometer, with chemical shifts (δ) reported in ppm. Fourier-transform infrared (FT-IR) spectra were recorded on an Avatar Nicolet FT-IR spectrometer either neat with smart OMNI-transmission accessories or with KBr pellets using standard methods, with frequencies reported in cm^{-1} . Liquid chromatography–high–resolution mass spectrometry (LC-HR/MS) spectra were obtained using a Waters G2-XS QTOF mass spectrometer, with samples dissolved in methanol. X-ray photoelectron Spectrometer (XPS) measurements were conducted on an ESCALAB 250Xi X-ray photoelectron spectrometer (Thermo Fisher) using an Al Ka source (15 kV, 10 mA) with Ar etching for 30 min (the charge of C-C carbon species here was corrected to 284.8 eV).

All reagents were commercially sourced from Sinopharm Chemical Reagent Limited Corporation or Shanghai Aladdin Biochemical Technology Co., Ltd., and used without further purification unless specified otherwise. CO_2 (CO_2 , 99.99%) and simulated flue gas (CO_2/N_2 , Vol/Vol = 15:85) were procured from Ganzhou Shengda Gas Co., Ltd. Flash column chromatography was carried out using silica gel (300-400 mesh).

1.2 Synthesis and Characterization of Imidazolium Salts

Synthesis of Im-Br1-4

2-Bromoethyl glucoside (2.5 equivalent) and *N*-alkylated imidazole (1.0 equivalent) were dissolved in anhydrous acetonitrile and the solution was heated at 120°C for 48 hours. Upon cooling, the acetonitrile was evaporated

using a rotary evaporator. Purification was achieved by column chromatography on silica gel using a gradient elution of dichloromethane/methanol (V:V = 20:1 to 10:1), resulting in the isolation of products Im-Br**1-4** (84-96%) as pale yellow solids.

Synthesis of Im-Br5

Im-Br**1** (0.131 g, 1.0 mmol) was dissolved in anhydrous methanol (10 mL) and reacted with a catalytic quantity of freshly prepared sodium methoxide (NaOMe) to achieve a basic environment, maintaining a pH of about 9.0. The solution was stirred at room temperature (RT) overnight. Subsequent neutralization was carried out using dry Amberlite IR 120 (H^+) resin to achieve a pH of around 7.0. The crude mixture underwent simple washing successively with NaHCO_3 , NaCl, and H_2O . Solvent evaporation gave the product Im-Br**5** (95%) as a pale yellow viscous liquid.

Synthesis of Cat1-5

In a 100 mL round-bottom flask, Im-Br**1** (0.131 g, 0.10 mmol) were dissolved in 50.0 mL of dichloromethane, and aqueous solution of sodium tetrachloropalladate ($0.01 \text{ g}\cdot\text{mL}^{-1}$, 4.4 mL) was added in the flask and the mixture was stirred at RT overnight. The solution was washed twice with ultrapure water, then removed by rotary evaporation and dried under vacuum to get the product Cat**1** (0.123 g, 92%) as a reddish brown solid. Cat**2-4** were prepared in a similar method of that for Cat**1**. Cat**2-4** absorb moisture easily and followed the trend of the other palladium salts, being reddish brown.

Im-Br**1**, 75%, ^1H NMR (400 MHz, CDCl_3) δ 10.41 (s, 2H), 7.72 (dd, $J = 12.6, 7.6 \text{ Hz}$, 4H), 7.53 (s, 4H), 7.42 (t, $J = 12.4 \text{ Hz}$, 4H), 5.70 (s, 2H), 5.18 (s, 2H), 4.99 (t, $J = 9.4 \text{ Hz}$, 2H), 4.83 (t, $J = 9.6 \text{ Hz}$, 4H), 4.72 – 4.66 (m, 4H), 4.55 (d, $J = 7.9 \text{ Hz}$, 2H), 4.23 (s, 2H), 4.10 (d, $J = 5.0 \text{ Hz}$, 1H), 4.03 (s, 1H), 3.94 (d, $J = 11.2 \text{ Hz}$, 2H), 3.67 (d, $J = 9.5 \text{ Hz}$, 2H), 3.17 (s, 2H), 1.90 – 1.80 (m, 20H), 1.53 (s, 4H) ppm; ^{13}C NMR (101 MHz, CDCl_3) δ 170.6, 169.9, 169.4, 169.3,

142.4, 133.4, 132.1, 130.8, 130.1, 127.2, 127.0, 114.3, 113.5, 100.0, 72.5, 71.6, 70.8, 68.0, 61.6, 53.6, 50.6, 49.9, 47.8, 20.8, 20.5, 20.5, 20.4 ppm. FT-IR (cm^{-1}): 3422.78, 3029.74, 2959.65, 1749.14, 1629.30, 1564.71, 1431.76, 1374.37, 1227.92, 1038.52, 909.18, 756.04, 604.81, 544.35, 433.80. HR-MS m/z: [M-Br] $^{2+}$ calcd for $\text{C}_{54}\text{H}_{64}\text{N}_4\text{O}_{20}$, 544.20515; found 544.20352.

Im-Br2, 80%, ^1H NMR (400 MHz, CDCl_3) δ 9.78 (s, 2H), 7.74 (s, 2H), 7.54 (s, 4H), 7.49 (s, 2H), 5.57 (s, 4H), 5.16 (t, $J = 9.5$ Hz, 2H), 4.99 (t, $J = 9.7$ Hz, 2H), 4.88 (dd, $J = 9.6, 8.1$ Hz, 2H), 4.64 (dd, $J = 18.1, 5.8$ Hz, 4H), 4.58 – 4.51 (m, 2H), 4.19 (dd, $J = 12.5, 4.5$ Hz, 3H), 4.12 (dd, $J = 16.9, 6.5$ Hz, 3H), 4.00 (dd, $J = 9.6, 6.2$ Hz, 2H), 3.81 – 3.76 (m, 2H), 2.04 (s, 4H), 1.97 (d, $J = 8.5$ Hz, 20H) ppm; ^{13}C NMR (101 MHz, CDCl_3) δ 170.8, 170.2, 169.9, 169.7, 137.0, 134.4, 130.5, 123.6, 122.5, 100.5, 72.6, 72.1, 71.3, 68.5, 67.9, 62.0, 52.9, 50.0, 21.1, 20.8 ppm. FT-IR (cm^{-1}): 3422.68, 3142.06, 3094.39, 2959.81, 1749.10, 1633.94, 1563.48, 1436.24, 1373.71, 1228.29, 1163.63, 1039.94, 909.73, 840.19, 729.27, 603.28, 419.63. HR-MS m/z: [M-Br] $^{2+}$ calcd for $\text{C}_{48}\text{H}_{64}\text{N}_4\text{O}_{20}$, 494.18950; found, 494.18950.

Im-Br3, 85%, ^1H NMR (400 MHz, CDCl_3) δ 7.70 (d, $J = 3.2$ Hz, 4H), 7.44 (s, 4H), 5.67 – 5.42 (m, 4H), 5.19 – 5.12 (m, 2H), 5.05 (s, 1H), 4.98 (t, $J = 8.6$ Hz, 2H), 4.83 – 4.77 (m, 2H), 4.67 (s, 3H), 4.57 (d, $J = 7.8$ Hz, 3H), 4.22 (s, 1H), 4.14 – 4.06 (m, 4H), 3.83 (d, $J = 6.0$ Hz, 2H), 3.49 – 3.29 (m, 2H), 2.70 (s, 6H), 2.05 (d, $J = 8.4$ Hz, 6H), 2.01 – 1.96 (m, 14H), 1.94 (s, 4H) ppm; ^{13}C NMR (101 MHz, CDCl_3) δ 170.9, 170.2, 169.8, 145.4, 134.4, 129.5, 122.4, 100.5, 72.7, 72.1, 71.3, 68.5, 68.2, 62.0, 53.7, 51.7, 48.9, 21.1, 20.8, 11.4 ppm. FT-IR (cm^{-1}): 3432.60, 2958.18, 1748.75, 1632.64, 1531.70, 1429.29, 1375.24, 1229.56, 1038.05, 910.28, 749.72, 599.75, 550.80, 408.41. HR-MS m/z: [M-Br] $^{2+}$ calcd for $\text{C}_{46}\text{H}_{60}\text{N}_4\text{O}_{20}$, 508.20515; found, 508.20855.

Im-Br4, 88%, ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 10.02 (s, 2H), 8.11 (d, $J = 8.0$ Hz, 2H), 7.93 (d, $J = 7.8$ Hz, 2H), 7.66 (d, $J = 7.7$ Hz, 2H), 7.63 (d, $J = 8.2$ Hz, 2H), 7.56 (s, 4H), 5.78 (s, 4H), 4.51 (t, $J = 7.2$ Hz, 4H), 1.34 – 1.20 (m,

24H), 0.83 (dd, J = 8.0, 5.5 Hz, 6H) ppm. ^{13}C NMR (101 MHz, DMSO- d_6) δ 143.5, 135.7, 132.4, 131.8, 130.0, 127.7, 115.0, 115.0, 50.4, 49.6, 48.0, 32.2, 29.5, 26.8, 23.1, 14.9 ppm. FT-IR (cm^{-1}): 3129.71, 3043.10, 2926.50, 1748.60, 1661.68, 1612.57, 1561.02, 1518.02, 1463.46, 1422.94, 1379.67, 1345.07, 1263.28, 1215.10, 1194.00, 1131.38, 1019.60, 860.72, 765.03, 684.52, 568.91. HR-MS m/z: [M-Br] $^{2+}$ calcd for $\text{C}_{38}\text{H}_{52}\text{N}_4$, 282.20905; found, 282.21105.

Im-Br5, 94%, ^1H NMR (400 MHz, DMSO- d_6) δ 10.08 (s, 2H), 8.15 (d, J = 7.4 Hz, 2H), 7.96 (d, J = 7.4 Hz, 2H), 7.63 (t, J = 7.0 Hz, 4H), 7.57 (s, 4H), 5.82 (s, 4H), 5.24 (s, 2H), 5.09 (d, J = 30.9 Hz, 2H), 4.77 (s, 3H), 4.68 (s, 1H), 4.24 (d, J = 7.8 Hz, 2H), 4.15 – 4.09 (m, 2H), 4.04 – 3.97 (m, 2H), 3.18 – 3.08 (m, 4H), 3.03 (d, J = 8.1 Hz, 2H), 2.92 (d, J = 7.8 Hz, 2H) ppm; ^{13}C NMR (101 MHz, DMSO- d_6) δ 144.5, 135.7, 132.3, 131.7, 123.0, 127.7, 115.2, 114.9, 104.1, 77.9, 77.5, 74.4, 71.0, 67.0, 62.0, 50.4, 48.2 ppm. FT-IR (cm^{-1}): 3388.07, 3130.84, 3060.75, 2965.42, 2922.13, 2853.27, 1742.99, 1632.36, 1557.80, 1515.89, 1452.93, 1372.23, 1344.86, 1287.08, 1218.65, 1197.41, 1162.93, 1076.70, 1034.22, 895.34, 863.37, 758.29, 666.36, 566.14. HR-MS m/z: [M-Br] $^{2+}$ calcd for $\text{C}_{38}\text{H}_{48}\text{N}_4\text{O}_{12}$, 376.16289; found, 376.16494.

Cat1, 92%, ^1H NMR (400 MHz, CDCl_3) δ 9.88 (s, 2H), 8.06 (s, 4H), 7.87 (s, 4H), 7.60 (s, 4H), 5.89 (s, 3H), 5.30 (s, 1H), 5.12 (d, J = 9.1 Hz, 2H), 4.96 (d, J = 9.7 Hz, 4H), 4.90 – 4.78 (m, 4H), 4.70 (d, J = 7.1 Hz, 2H), 4.48 (s, 2H), 4.34 (s, 2H), 4.21 (d, J = 9.6 Hz, 2H), 4.09 (d, J = 10.8 Hz, 2H), 3.84 (s, 2H), 1.97 (d, J = 15.9 Hz, 20H), 1.70 (s, 4H) ppm; ^{13}C NMR (101 MHz, CDCl_3) δ 170.9, 170.2, 169.8, 142.2, 133.9, 132.6, 131.3, 130.9, 127.8, 114.8, 114.1, 100.5, 73.0, 72.0, 71.3, 68.5, 68.4, 62.0, 53.7, 51.4, 48.5, 21.2, 20.9, 20.9, 20.8 ppm. FT-IR (cm^{-1}): 3441.45, 2960.63, 1749.56, 1628.39, 1565.79, 1427.51, 1372.95, 1227.05, 1035.94, 749.83, 700.59, 599.07, 538.82, 430.84. HR-MS m/z: [M-PdCl₄] $^{2+}$ calcd for $\text{C}_{54}\text{H}_{64}\text{N}_4\text{O}_{20}$, 544.20515; found 544.20901.

Cat2, 98%, ^1H NMR (400 MHz, CDCl_3) δ 9.35 (s, 2H), 7.93 (s, 4H), 7.62 (s, 2H), 7.51 (s, 2H), 5.68 (s, 3H), 5.29 (s, 1H), 5.18 (t, J = 9.2 Hz, 2H), 5.02 (t,

J = 9.2 Hz, 2H), 4.91 (t, *J* = 8.1 Hz, 2H), 4.74 (d, *J* = 6.8 Hz, 4H), 4.59 (s, 2H), 4.31 (s, 2H), 4.23 (s, 4H), 4.16 (d, *J* = 11.5 Hz, 2H), 3.88 (d, *J* = 7.8 Hz, 2H), 2.01 (t, *J* = 15.8 Hz, 24H) ppm; ^{13}C NMR (101 MHz, CDCl_3) δ 170.9, 170.2, 170.0, 169.8, 136.6, 134.5, 130.9, 124.1, 122.8, 100.5, 72.7, 72.0, 71.4, 68.41, 68.2, 62.0, 53.7, 53.5, 50.5, 21.4, 21.3, 20.9 ppm. FT-IR (cm^{-1}): 3494, 3146, 2968, 1956, 1629, 1582, 1533, 1431, 1376, 1226, 1171, 1035, 912, 755, 604. HR-MS m/z: [M-PdCl₄]²⁺ calcd for C₄₈H₆₄N₄O₂₀, 494.18950; found, 494.19304.

Cat3, 98%, ^1H NMR (400 MHz, CDCl_3) δ 7.71 (s, 4H), 7.65 (s, 2H), 7.60 (s, 2H), 5.65 (t, *J* = 17.8 Hz, 4H), 5.20 (t, *J* = 9.4 Hz, 2H), 5.04 (t, *J* = 9.6 Hz, 2H), 4.90 (t, *J* = 8.7 Hz, 2H), 4.75 (d, *J* = 7.7 Hz, 2H), 4.64 (s, 4H), 4.36 (s, 2H), 4.31 – 4.19 (m, 4H), 4.15 (d, *J* = 11.4 Hz, 2H), 3.89 (d, *J* = 9.3 Hz, 2H), 2.90 (s, 6H), 2.10 (s, 6H), 2.02 (d, *J* = 9.5 Hz, 18H) ppm; ^{13}C NMR (101 MHz, CDCl_3) δ 171.0, 170.3, 170.0, 169.9, 145.4, 134.2, 130.1, 122.9, 122.5, 100.5, 72.9, 72.0, 71.4, 68.5, 68.2, 62.0, 52.5, 49.5, 21.4, 21.3, 21.0, 12.0 ppm. FT-IR (cm^{-1}): 3480, 3146, 3105, 2968, 2934, 2860, 1752, 1629, 1561, 1431, 1377, 1226, 1165, 1042, 987, 912, 803, 727, 604. HR-MS m/z: [M-PdCl₄]²⁺ calcd for C₄₆H₆₀N₄O₂₀, 508.20515; found, 508.20871.

Cat4, 95%, ^1H NMR (400 MHz, DMSO-*d*₆) δ 10.01 (s, 2H), 8.10 (d, *J* = 7.9 Hz, 2H), 7.92 (d, *J* = 7.8 Hz, 2H), 7.67 (t, *J* = 5.4 Hz, 2H), 7.63 (d, *J* = 8.2 Hz, 2H), 7.55 (s, 4H), 5.78 (s, 4H), 4.50 (t, *J* = 7.3 Hz, 4H), 1.30 (d, *J* = 3.1 Hz, 8H), 1.23 (d, *J* = 11.0 Hz, 16H), 0.85 – 0.81 (m, 6H) ppm; ^{13}C NMR (101 MHz, DMSO-*d*₆) δ 143.5, 135.7, 132.4, 131.9, 129.9, 127.8, 115.0, 114.9, 50.5, 48.0, 32.20, 29.5, 29.5, 29.5, 26.8, 23.1, 15.0 ppm. FT-IR (cm^{-1}): 3124.38, 3027.67, 2955.04, 2923.84, 2852.83, 1717.73, 1633.64, 1613.73, 1456.48, 1422.46, 1340.72, 1261.30, 1261.30, 1215.89, 1179.74, 1094.89, 1018.06, 868.22, 802.31, 752.41, 689.51, 602.02, 570.65. HR-MS m/z: [M-PdCl₄]²⁺ calcd for C₃₈H₅₂N₄, 282.20905; found, 282.21053.

Cat5, 97%, ^1H NMR (400 MHz, DMSO-*d*₆) δ 10.10 (s, 2H), 8.14 (d, *J* = 7.1 Hz, 2H), 7.95 (d, *J* = 7.4 Hz, 2H), 7.67 – 7.59 (m, 4H), 7.57 (s, 4H), 5.82 (s, 4H),

5.27 (d, J = 18.8 Hz, 2H), 5.07 (s, 2H), 4.76 (s, 4H), 4.23 (d, J = 7.7 Hz, 2H), 4.10 (s, 2H), 4.01 (s, 2H), 3.15 (t, J = 8.7 Hz, 2H), 3.12 – 3.07 (m, 2H), 3.03 (d, J = 9.0 Hz, 2H), 2.90 (t, J = 8.2 Hz, 2H) ppm; ^{13}C NMR (101 MHz, DMSO- d_6) δ 144.6, 135.7, 132.3, 131.7, 130.0, 127.8, 115.3, 114.9, 104.1, 77.9, 77.5, 74.4, 71.0, 67.1, 61.9, 50.4, 48.3 ppm. FT-IR (cm^{-1}): 2973.83, 2923.74, 2856.07, 1745.01, 1633.75, 1556.04, 1518.69, 1487.36, 1455.67, 1421.58, 1373.17, 1339.25, 1218.69, 1186.56, 1070.35, 1024.42, 949.53, 856.57, 758.90, 652.34, 599.07, 566.15, 534.58. HR-MS m/z: [M-PdCl₄]²⁺ calcd for C₃₈H₄₈N₄O₁₂, 376.16289; found, 376.16457.

1.3 Synthesis and Characterization of Propargylic Amines

For the synthesis of propargylic amine **6**, propargyl halide (1.0 mmol) was added dropwise to the corresponding amine (3.0 mmol) at 0°C in the dark, and the mixture was slowly warmed to RT. The reaction was then stirred for 12-24 hours. Subsequently, NaOH and dichloromethane were introduced. After extracting the aqueous layer with dichloromethane, the organic phase was washed with saturated brine, dried over MgSO₄, and the solvent was evaporated. The crude product was further purified by flash column chromatography on silica gel using a gradient elution of V_{PE}/V_{EA} = 20:1.

1.4 General Procedure for the Synthesis of Propargylic Amines with CO₂

In a standard procedure, Cat**1** (0.25-0.5 mol%), propargylic amine **6** (0.05 mmol), and NaOAc (0.075 mmol) were combined in DMSO (1.0 mL) within a reaction vessel. A simulated flue gas (CO₂/N₂, Vol/Vol = 15:85, bubbling) was introduced into the preheated reaction mixture using a long hollow needle (φ 0.7 × 200 mm) immersed in a 70°C oil bath. The reaction mixture was then stirred for a specified duration. The yield was quantified using ¹H NMR

spectroscopy. To determine the isolated yields of purified products, the crude material was concentrated and subjected to purification via column chromatography on silica gel using a gradient elution of petroleum ether/ethyl acetate (V:V = 20:1 to 10:1). The isolated yields were calculated based on the initial reactants.

2. Catalytic Section

2.1 Table S1. Comparison of TOFs with Other Reported Catalysts

Entry	Catalyst	T (°C)/t (h)	P (atm)	TOF (h ⁻¹)	TON	Cycles	Ref.
1	Cat1	70/0.083	0.15	3456	400	10	Herein
2	CuI/DBU	50/4	1.0	2	8	/	1
3	AgOAc	25/7	1.0	7	49	/	2
4	AgNO ₃ /DBU	60/2	1.0	94	188	/	3
5	CoBr ₂ /TBD	80/9	1.0	1	9	/	4
6	ZnCl ₂ (TBD) ₂	60/12	1.0	2	24	/	5
7	Ag ₂₇ -MOF	25/6	1.0	16	96	4	6
8	TOS-Ag ₄	25/24	1.0	4	96	5	7
9	Zn ₁₁₆	70/12	1.0	31	372	10	8
10	PdSCS	50/0.33	1.0	60	19.8	/	9
11	Ag@TpPa-1	60/18	1.0	17	306	5	10
12	Ag@2,6-FPP-TAPT	50/2	1.0	964	1928	4	11
13	Cu ₂ O@ZIF-8	40/6	1.0	3.3	19.8	5	12
14	Cu ^I /Cu ^{II} -MOF	30/0.17	1.0	230	39.1	5	13
15	Pd@BBA-2	40/2	1.0	43.68	87.36	5	14
15	AuCl(IPr)	40/15	1.0	3.0	45	/	15
16	AgCl(IPr)	40/15	1.0	3.0	45	/	16
17	[Au]	RT/24	air	5.33	127.9	/	17
18	Ag-HMP-2	60/20	1.0	10.6	212	/	18

Entry	Catalyst	T (°C)/t (h)	P (atm)	TOF (h ⁻¹)	TON	Cycles	Ref.
19	Ag-MOF-1	RT/24	1.0	0.56	13.44	/	19
20	2Gn[TEG][Au]	RT/24	1.0	1.77	42.48	/	20
21	AcGlu-Im-PdCl ₄	70/0.5	0.15	1440	720	/	21
22	Ag ₄ NC	25/2	1.0	2873	5746	5	22
23	Cu ₆ -NH ₂	30/1.5	1.0	387	580.5	5	23
24	AcGlu-Im-Br 3 /CuBr ₂	80/0.33	1.0	400	132	/	24
25	Cu ^I -TpBD-COF	80/5	1.0	1058	5290	9	25
26	Cu ^I Cu ^{II} DA/WN	40/5	1.0	38.4	192	5	26

3. Characterizations

3.1 Comparative NMR Spectra of Imidazolium Salts

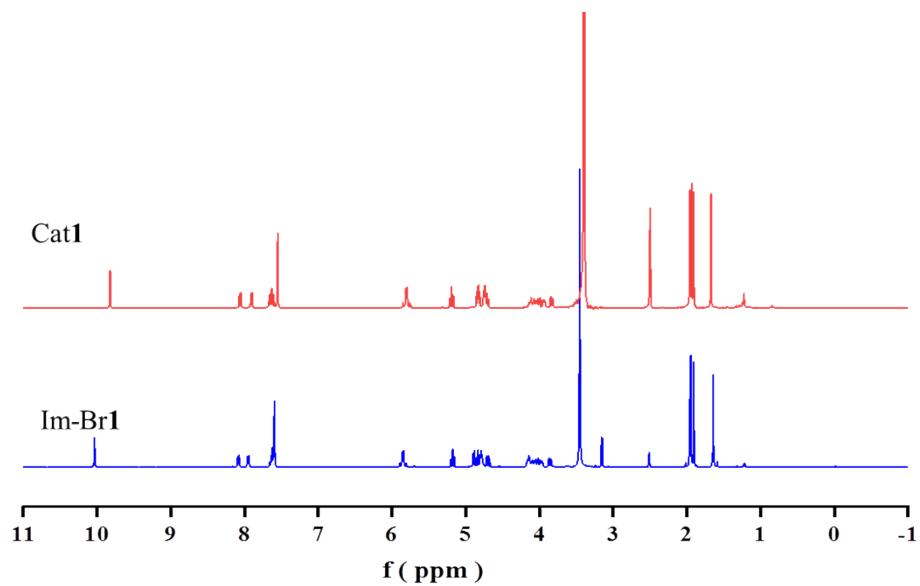


Fig. S1 ¹H NMR of Cat1/Im-Br1 in DMSO-*d*₆.

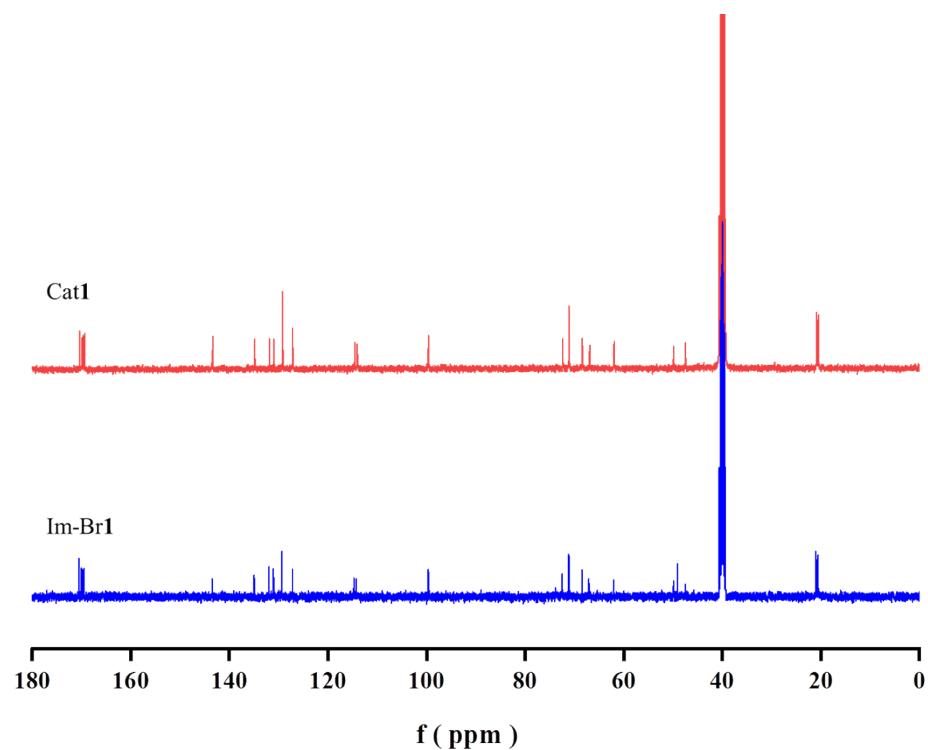


Fig. S2 ¹³C NMR of Cat1/Im-Br1 in DMSO-*d*₆.

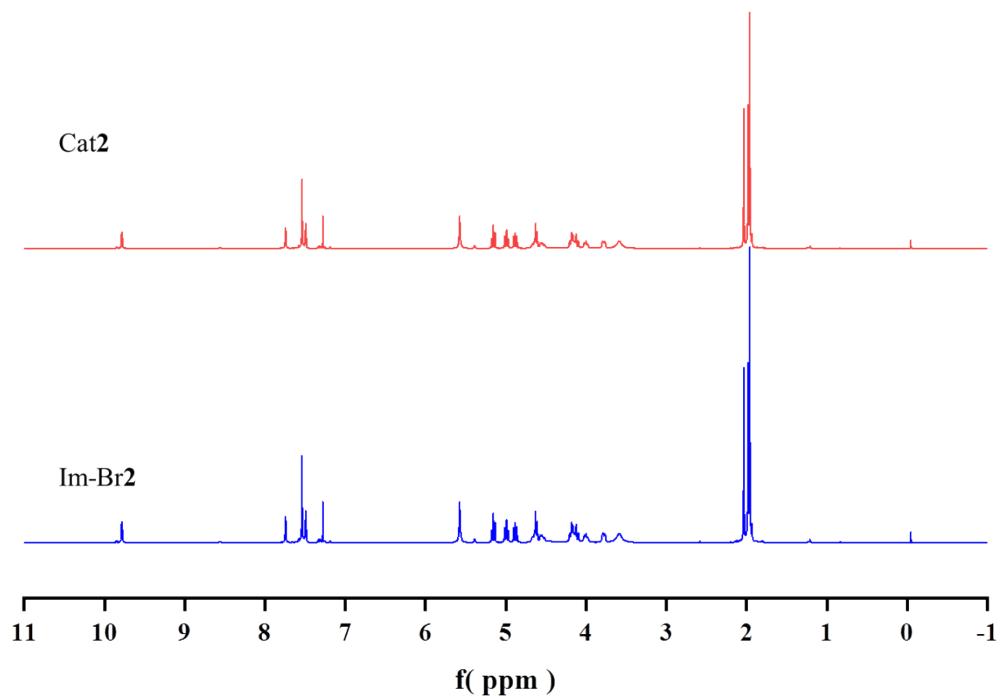


Fig. S3 ¹H NMR of Cat2/Im-Br2 in CDCl₃.

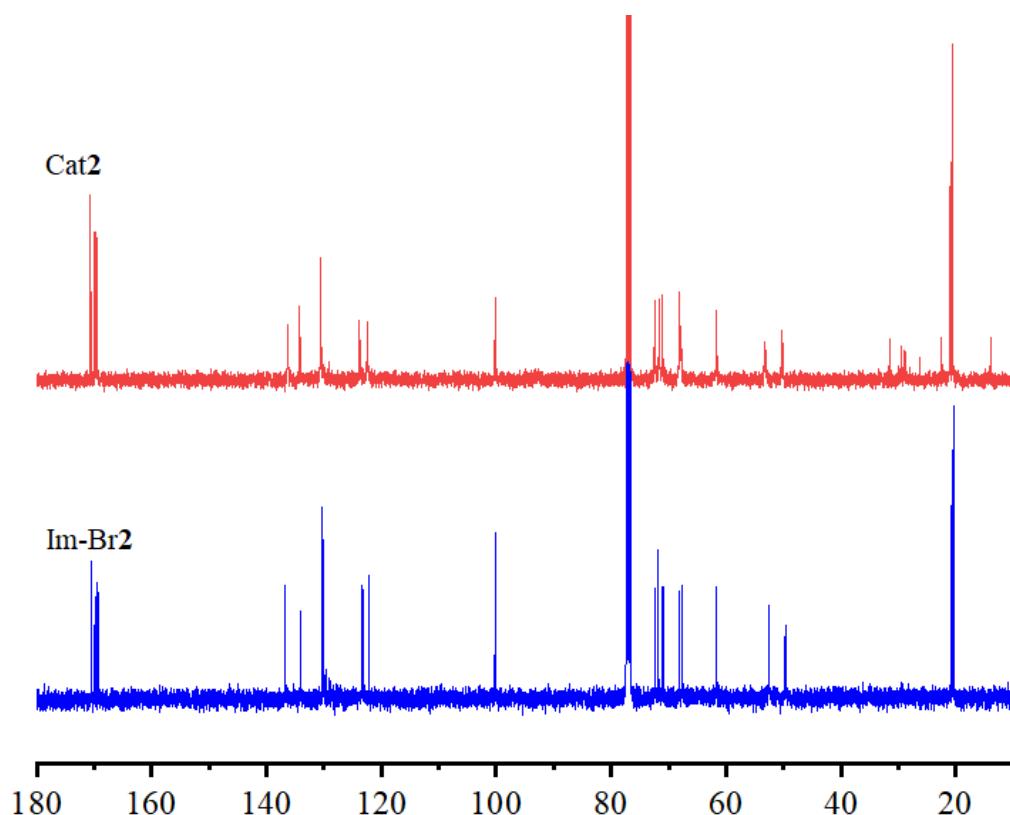


Fig. S4 ¹³C NMR of Cat2/Im-Br2 in CDCl₃.

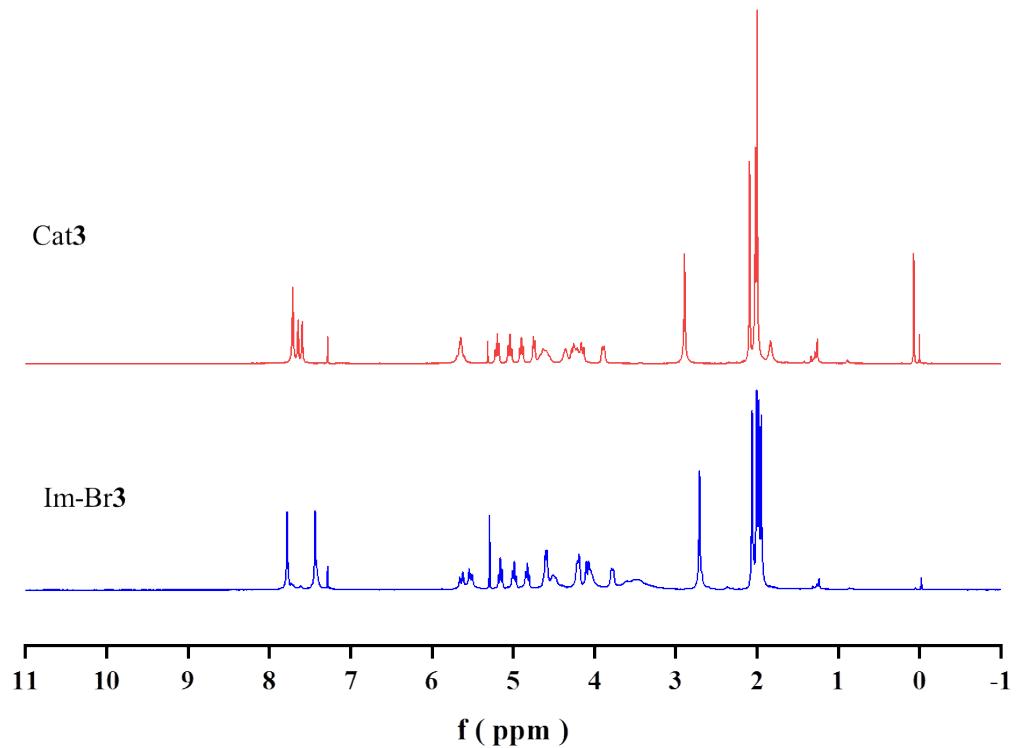


Fig. S5 ¹H NMR of Cat3/Im-Br3 in CDCl₃.

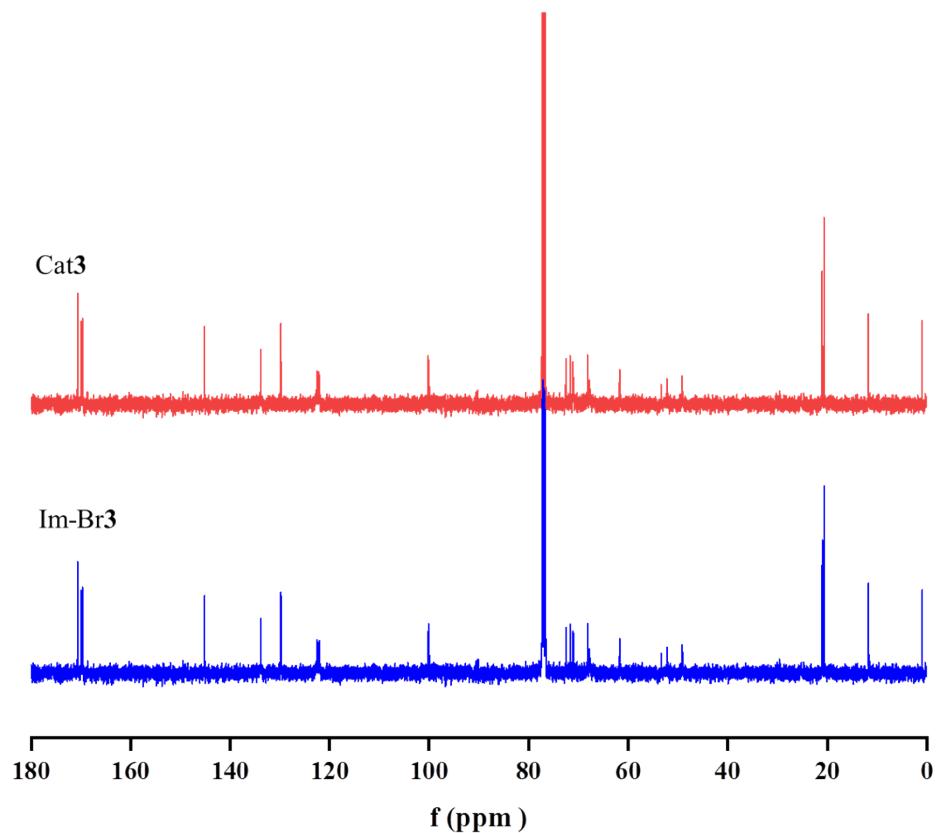


Fig. S6 ¹³C NMR of Cat3/Im-Br3 in CDCl₃.

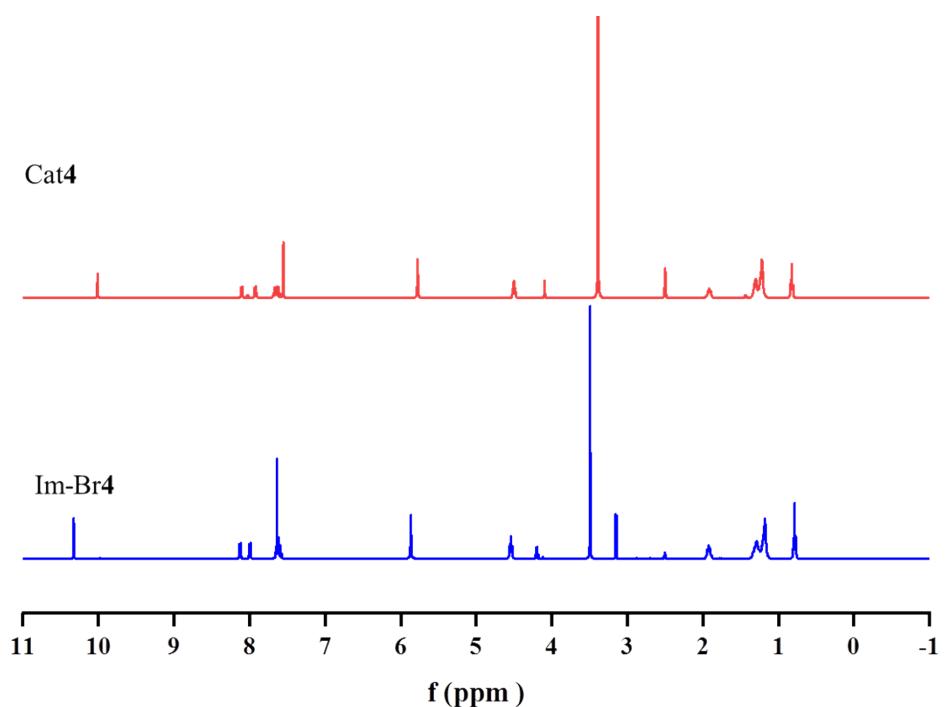


Fig. S7 ¹H NMR of Cat4/Im-Br4 in DMSO-*d*₆.

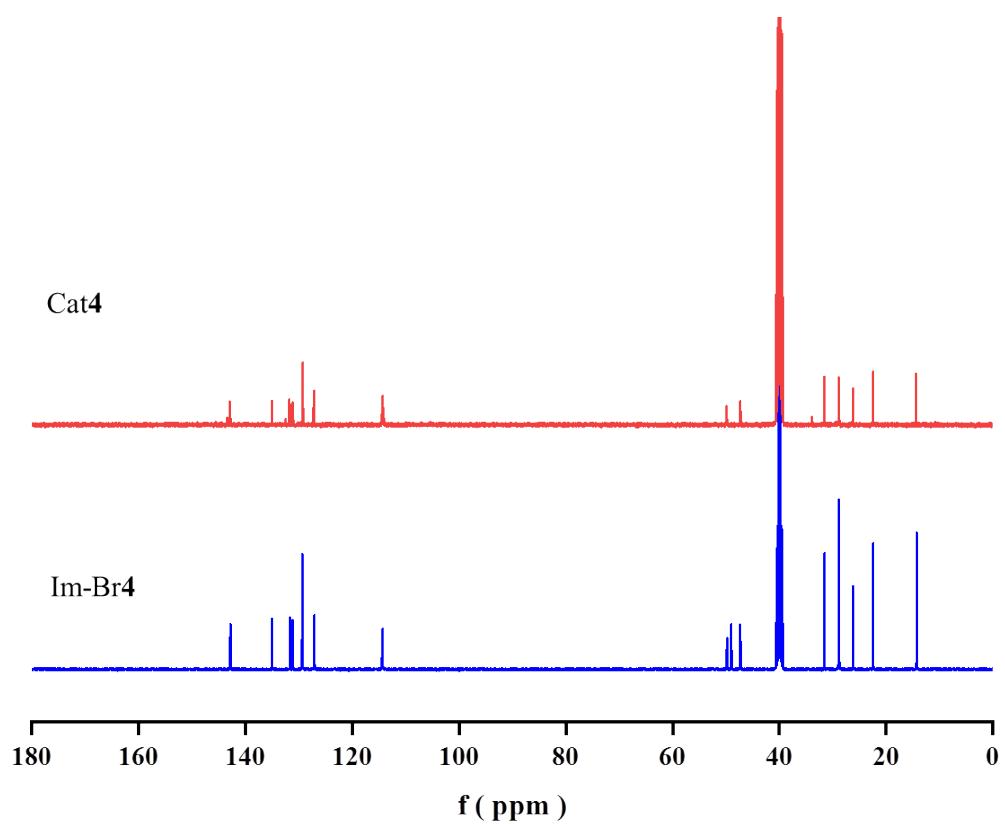


Fig. S8 ¹³C NMR of Cat4/Im-Br4 in DMSO-*d*₆.

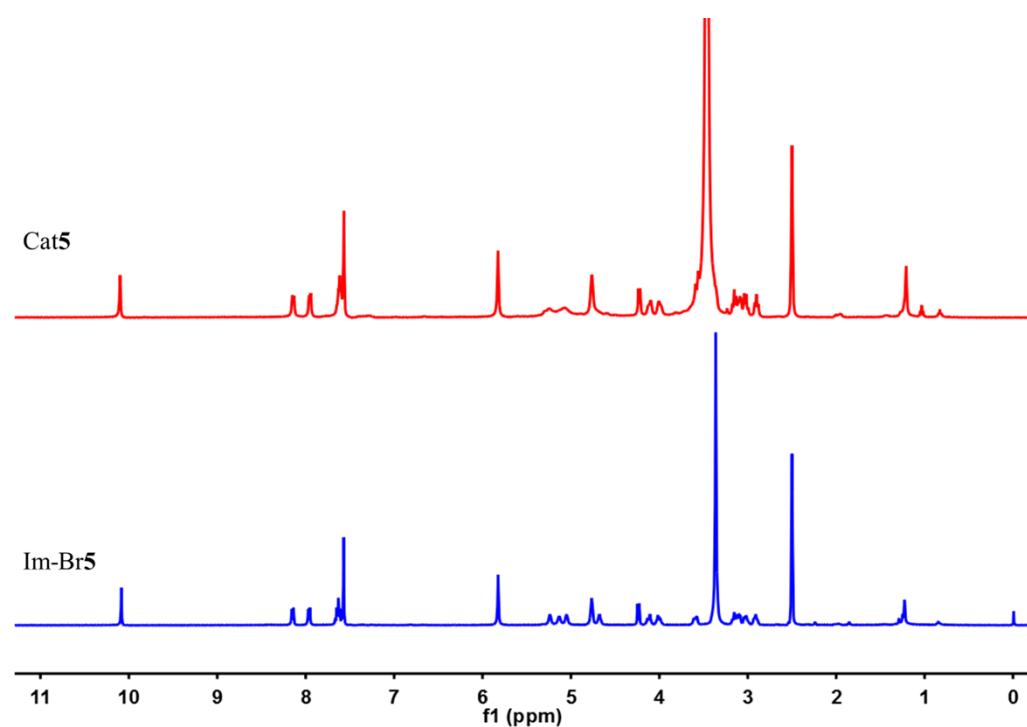


Fig. S9 ¹H NMR of Cat5/Im-Br5 in DMSO-*d*₆.

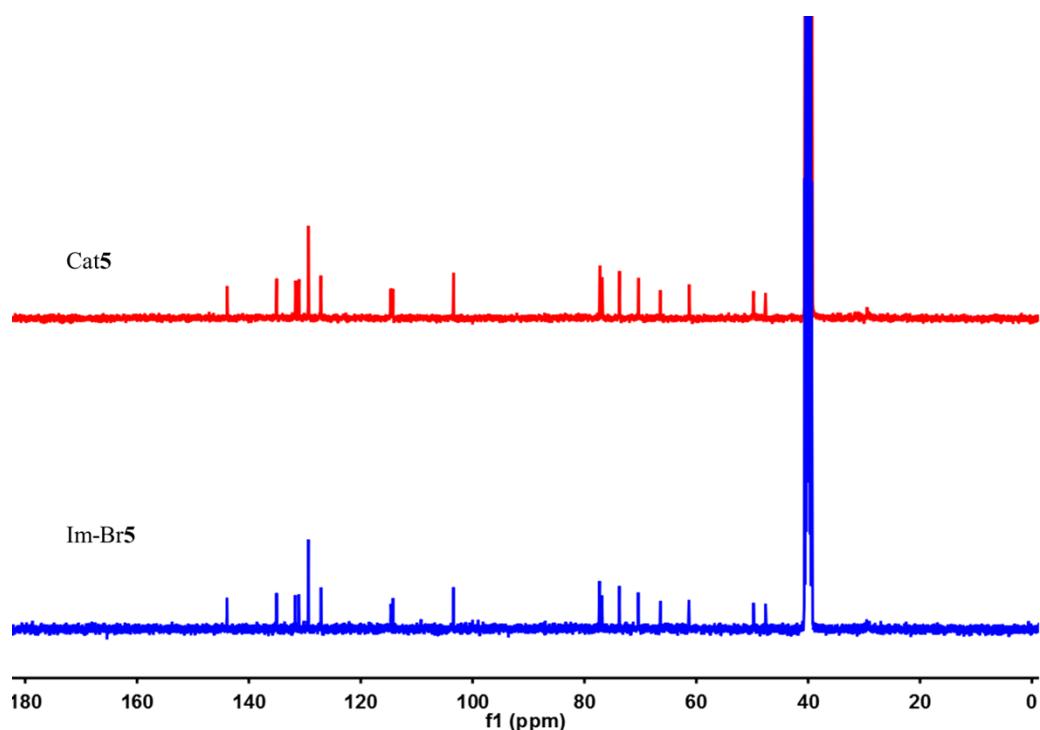


Fig. S10 ¹³C NMR of Cat5/Im-Br5 in DMSO-*d*₆.

3.2 Comparative FT-IR Spectra of Imidazolium Salts

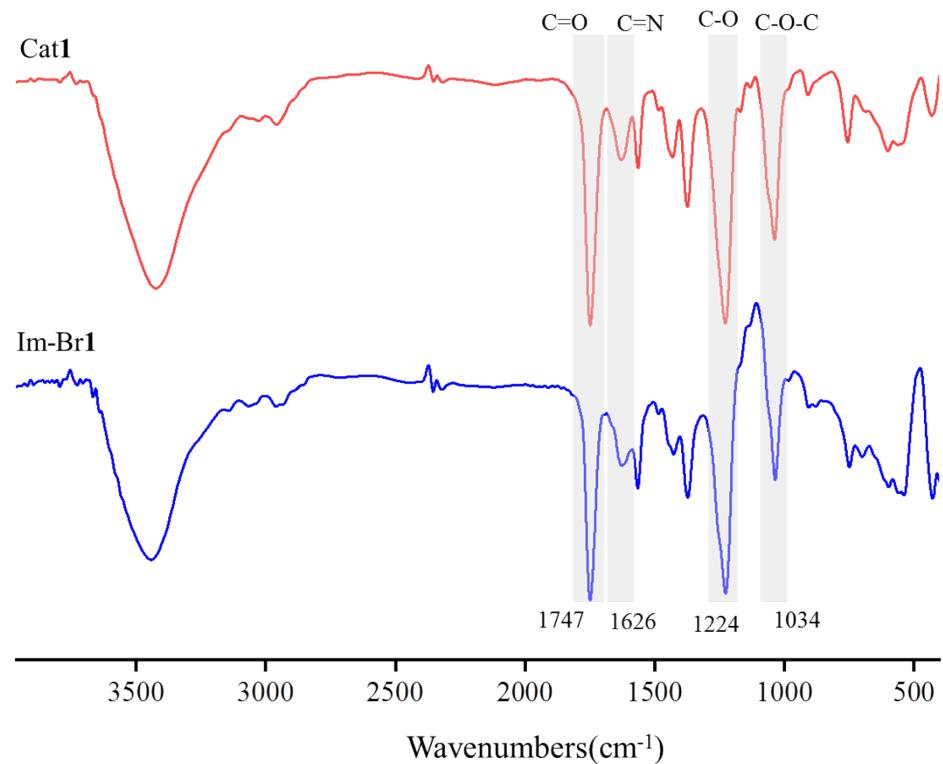


Fig. S11 FT-IR of **Cat1/Im-Br1**.

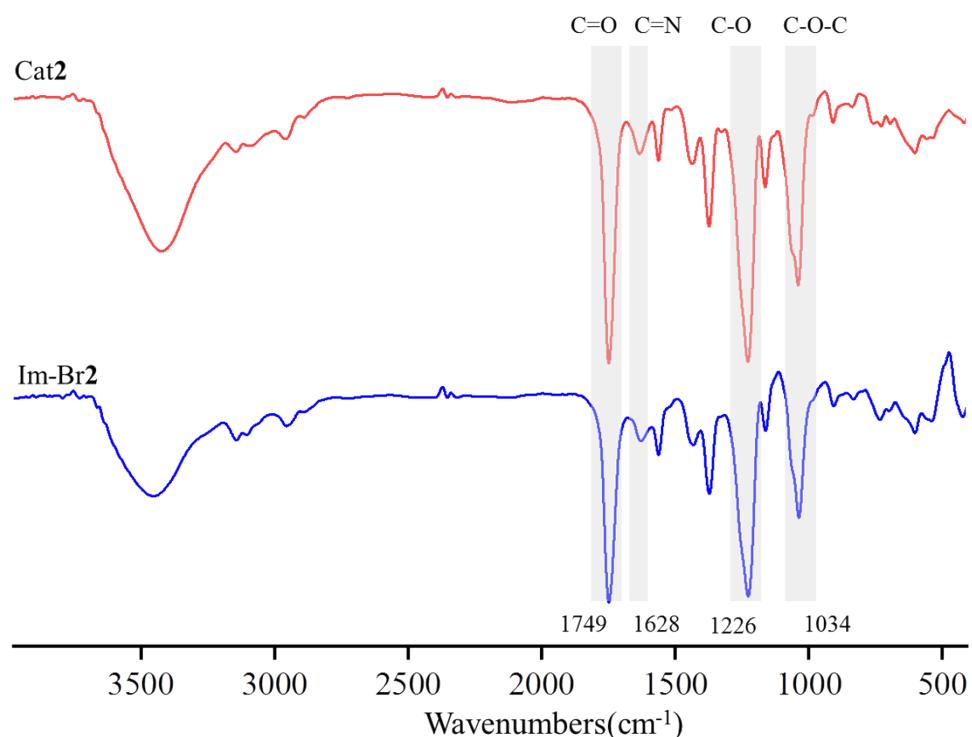


Fig. S12 FT-IR of **Cat2/Im-Br2**.

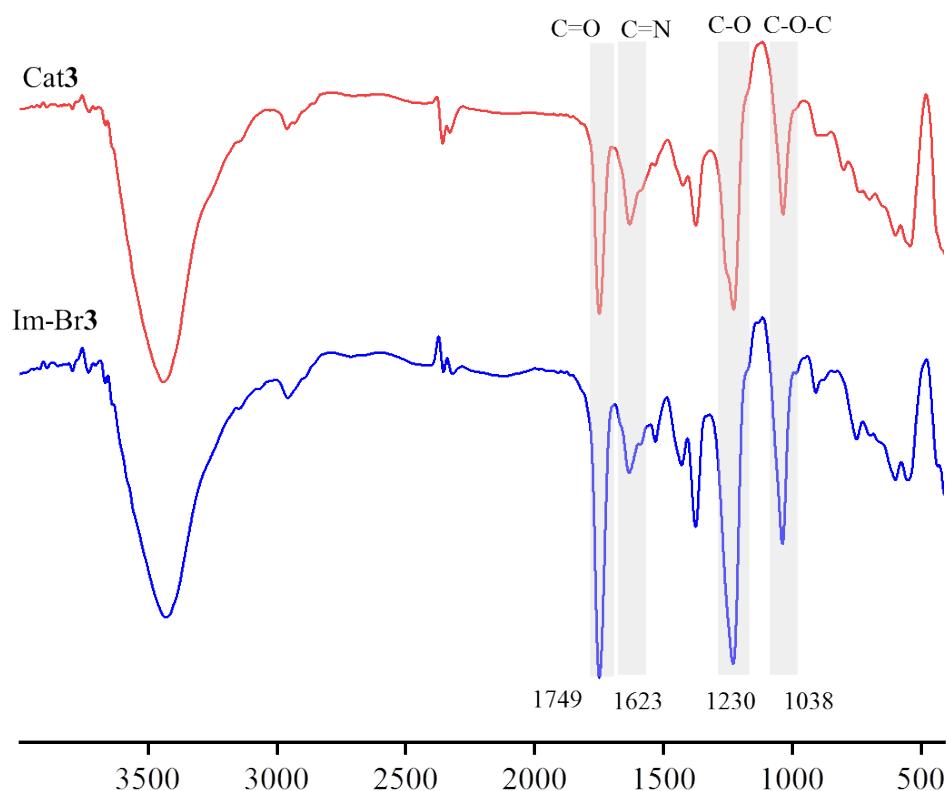


Fig. S13 FT-IR of **Cat3/Im-Br3**.

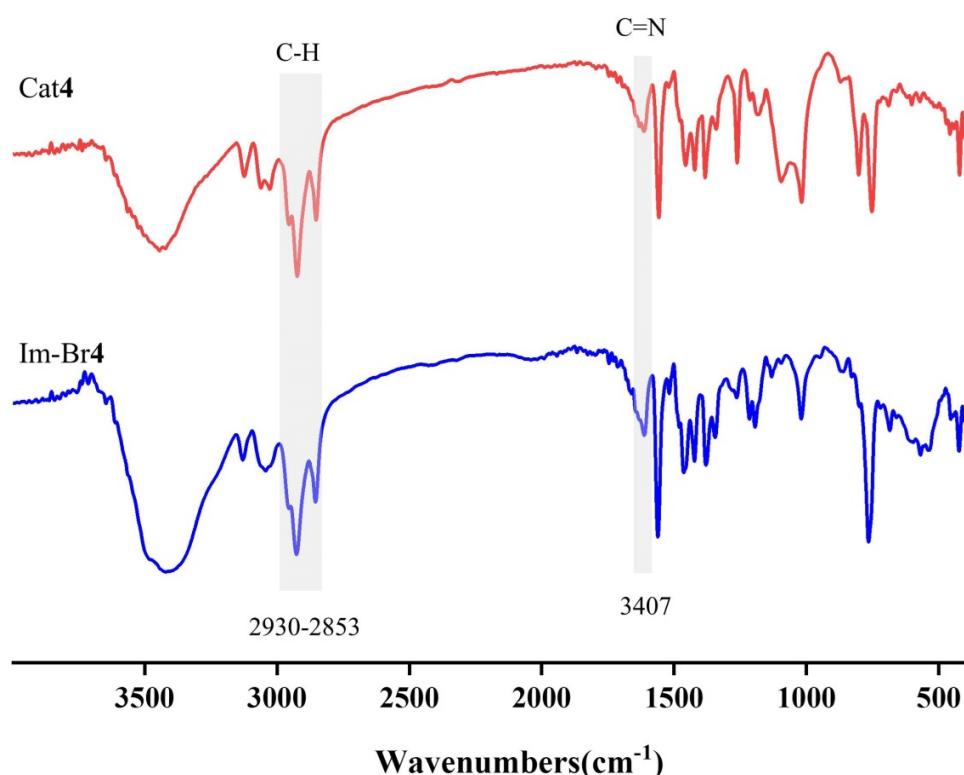


Fig. S14 FT-IR of **Cat4/Im-Br4**.

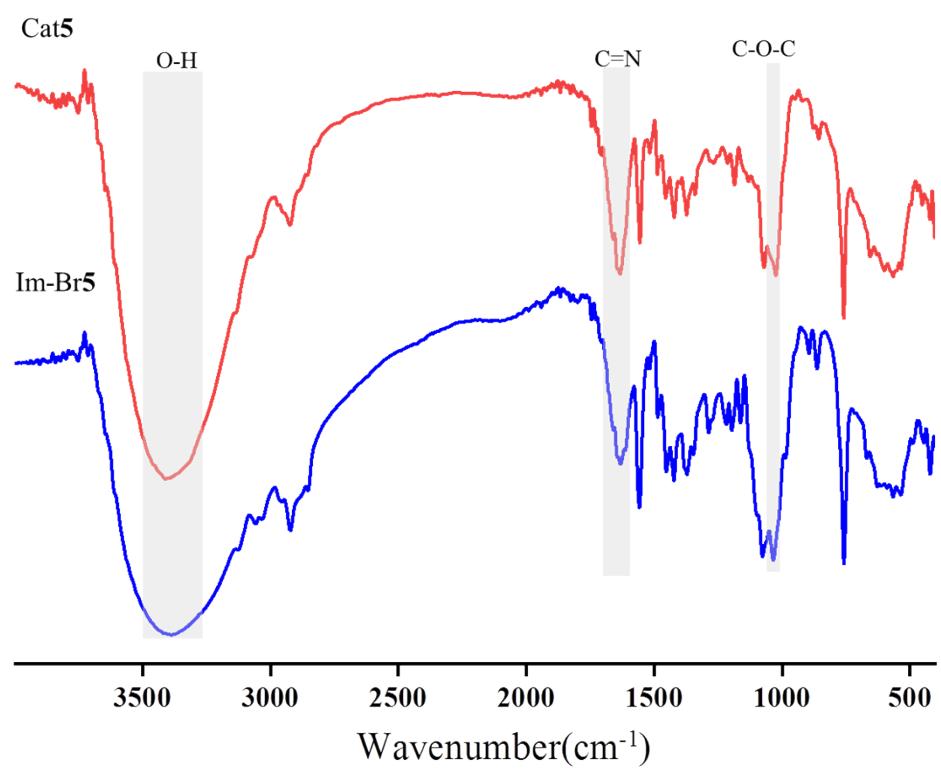
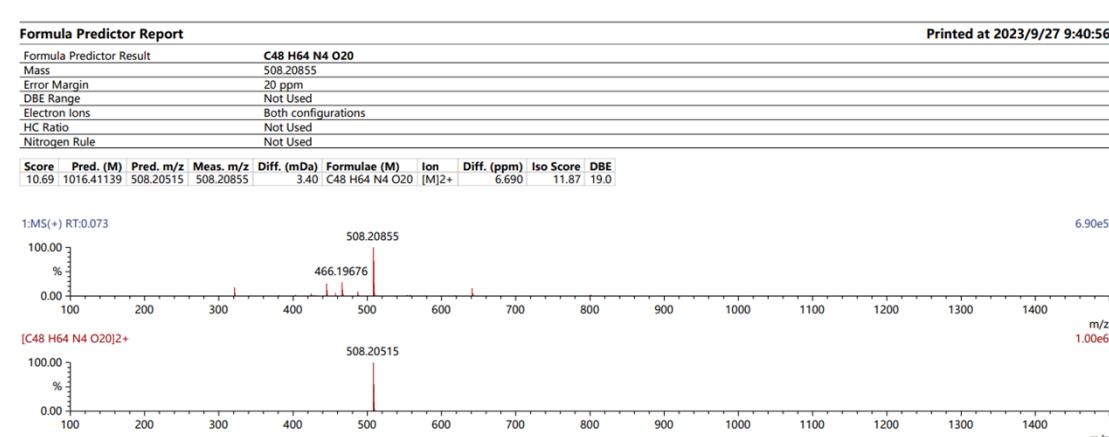
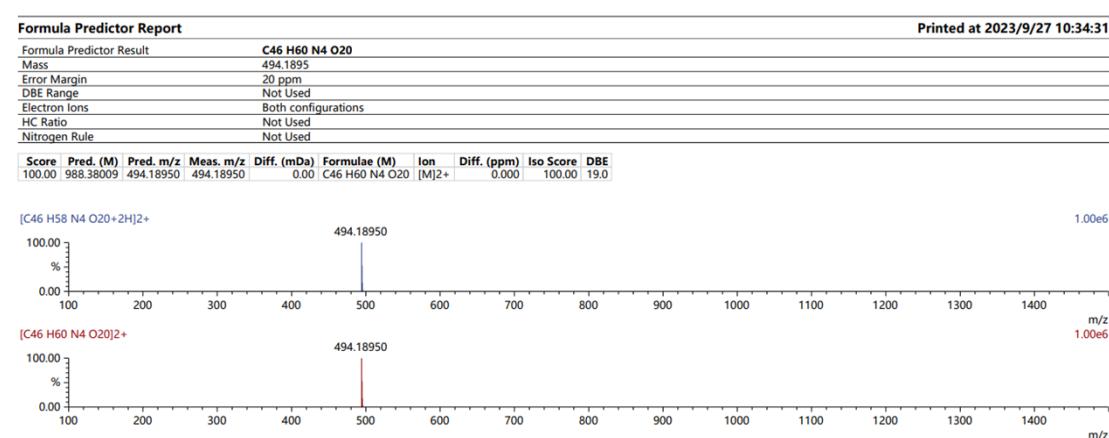
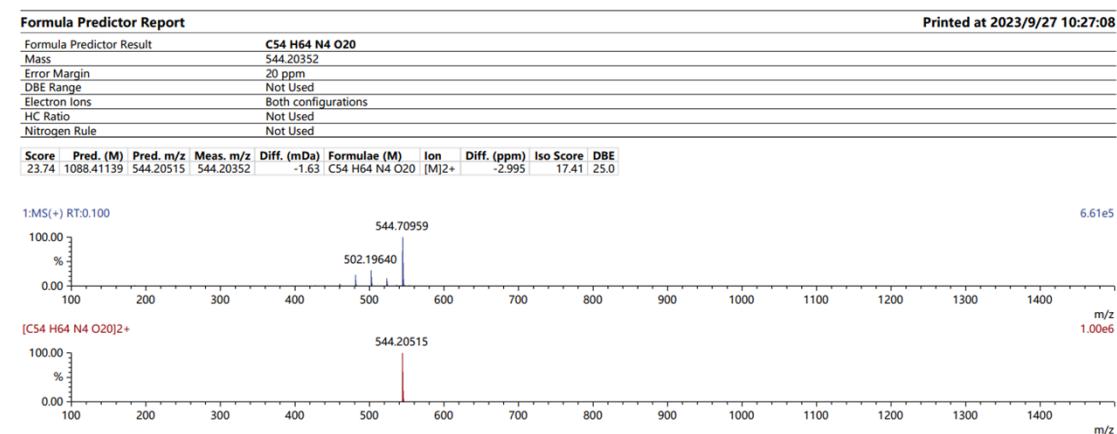


Fig. S15 FT-IR of **Cat5**/Im-Br5.

3.3 HR-MS Spectra of Imidazolium Salts



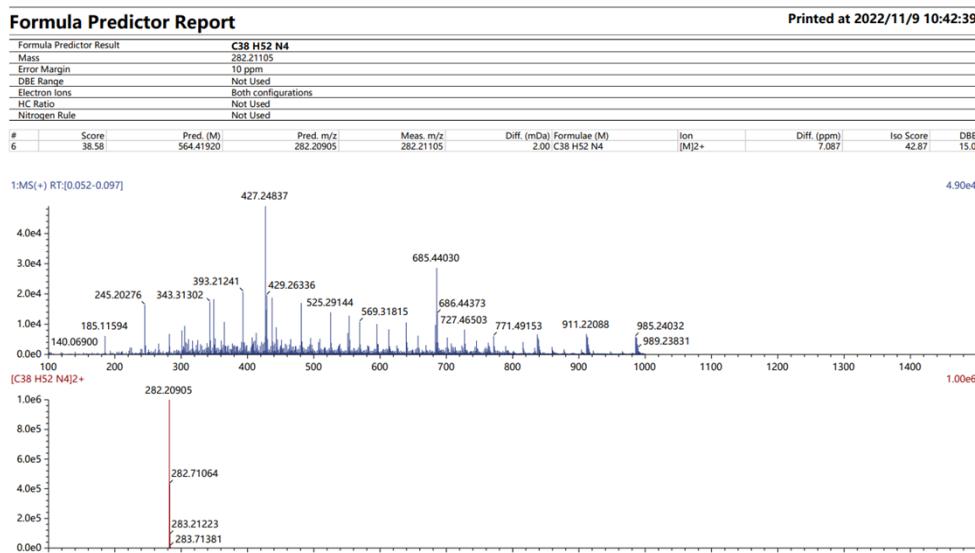


Fig. S19 HR/LC-MS of Im-Br4.

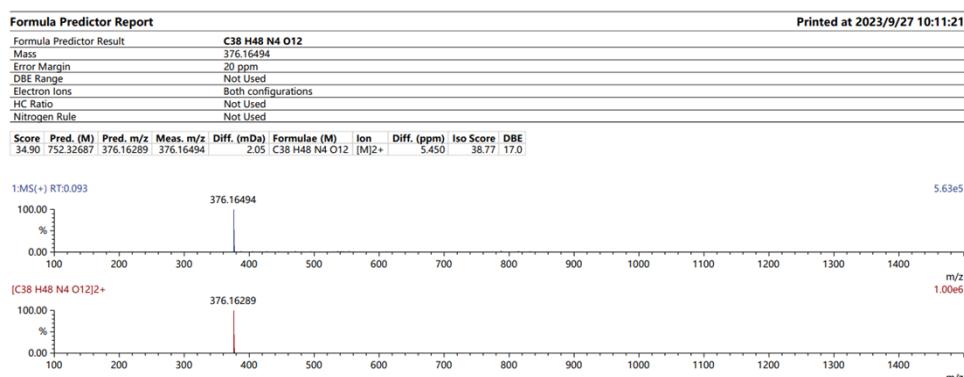


Fig. S20 HR/LC-MS of Im-Br⁵.

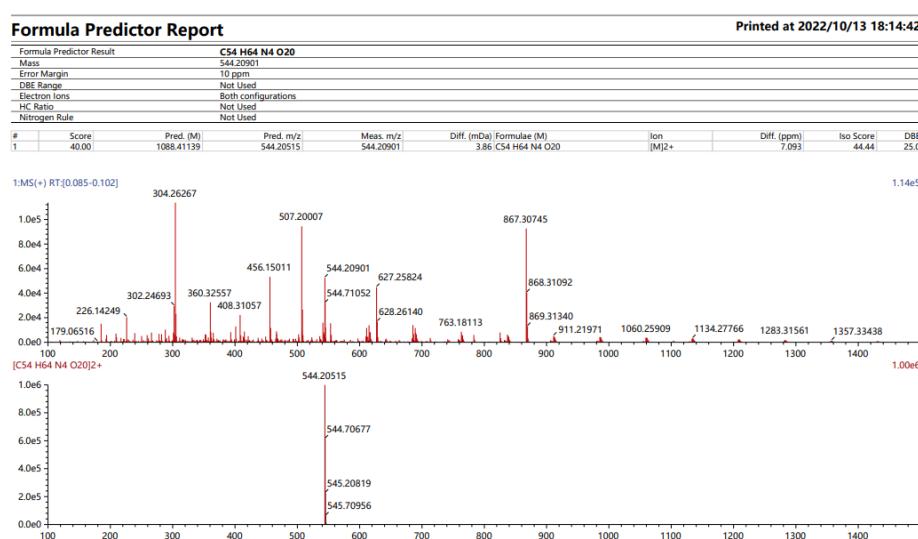


Fig. S21 HR/LC-MS of Cat1.

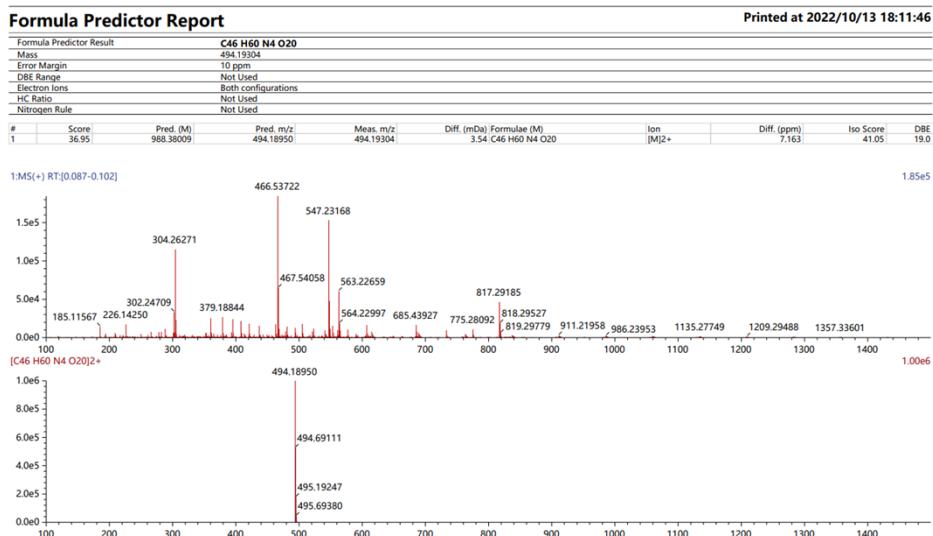


Fig. S22 HR/LC-MS of Cat2.

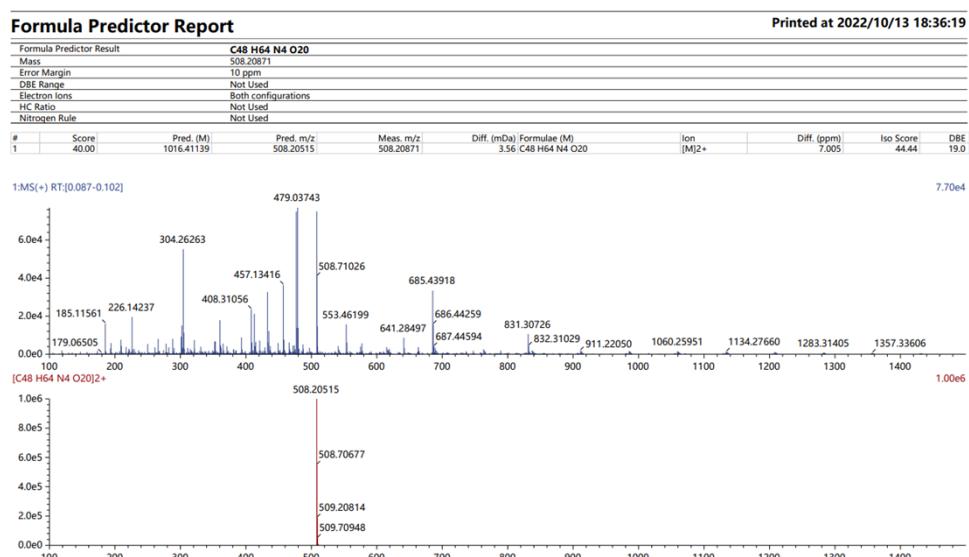


Fig. S23 HR/LC-MS of Cat3.

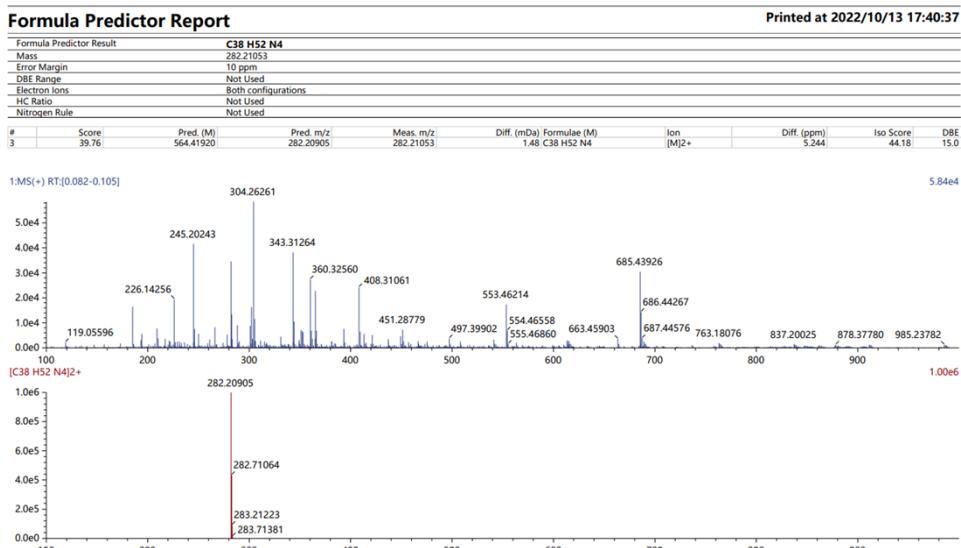


Fig. S24 HR/LC-MS of Cat4.

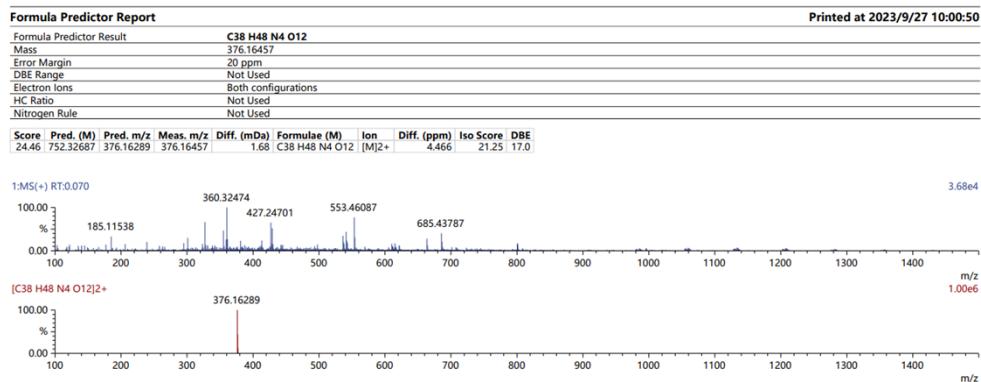


Fig. S25 HR/LC-MS of Cat5.

3.4 XPS Spectra of Cat1 and Cat5

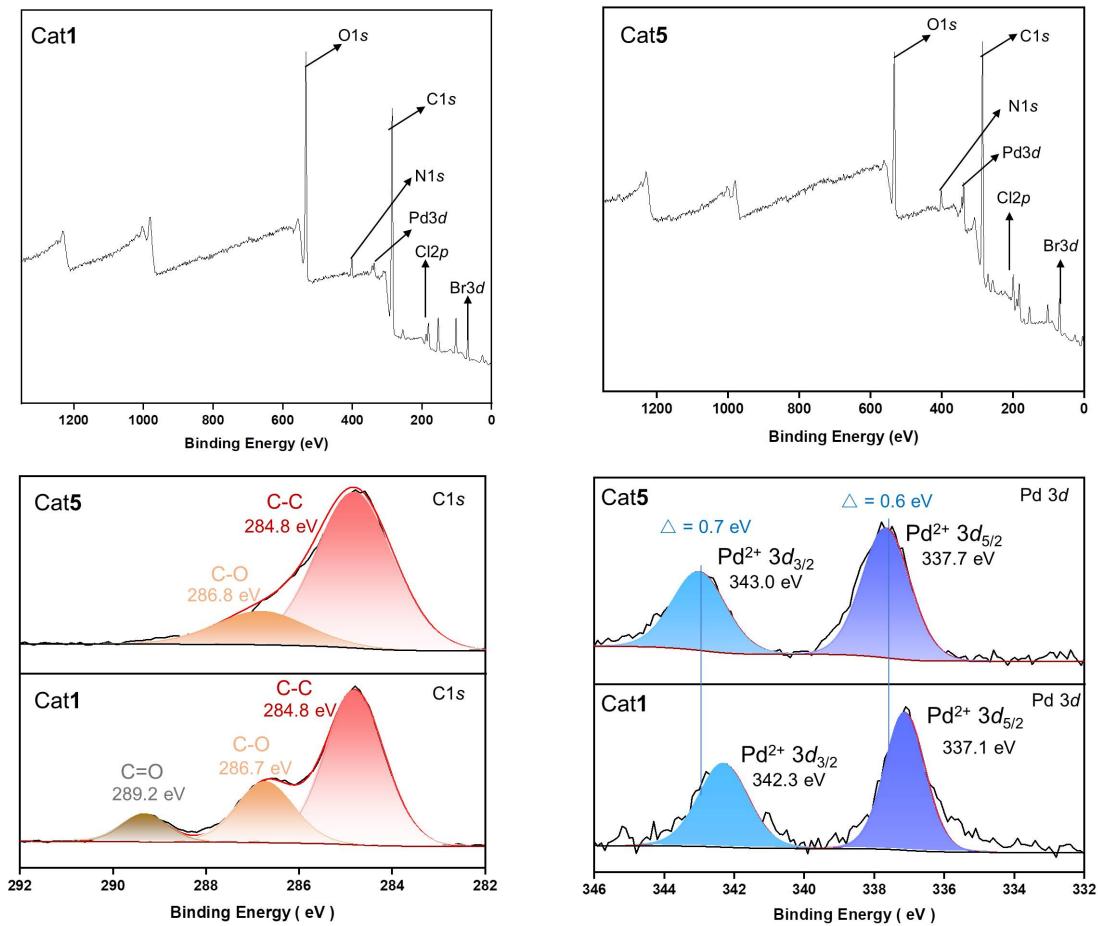


Fig. S26 XPS spectra of Cat1 and Cat5.

3.5 TGA Spectra of Cat1-5

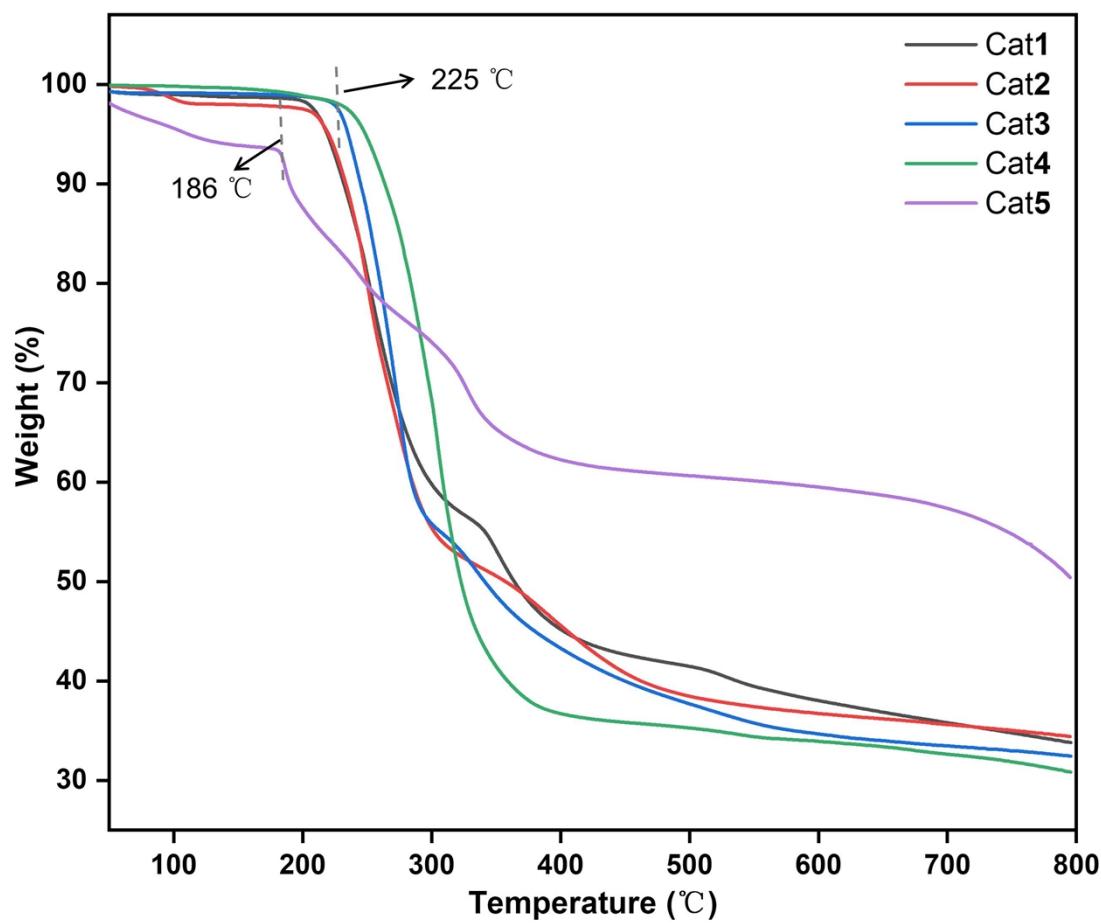


Fig. S27 TGA spectra of Cat1-5.

3.6 NMR Spectra and Color of Cat1 at Different Temperatures

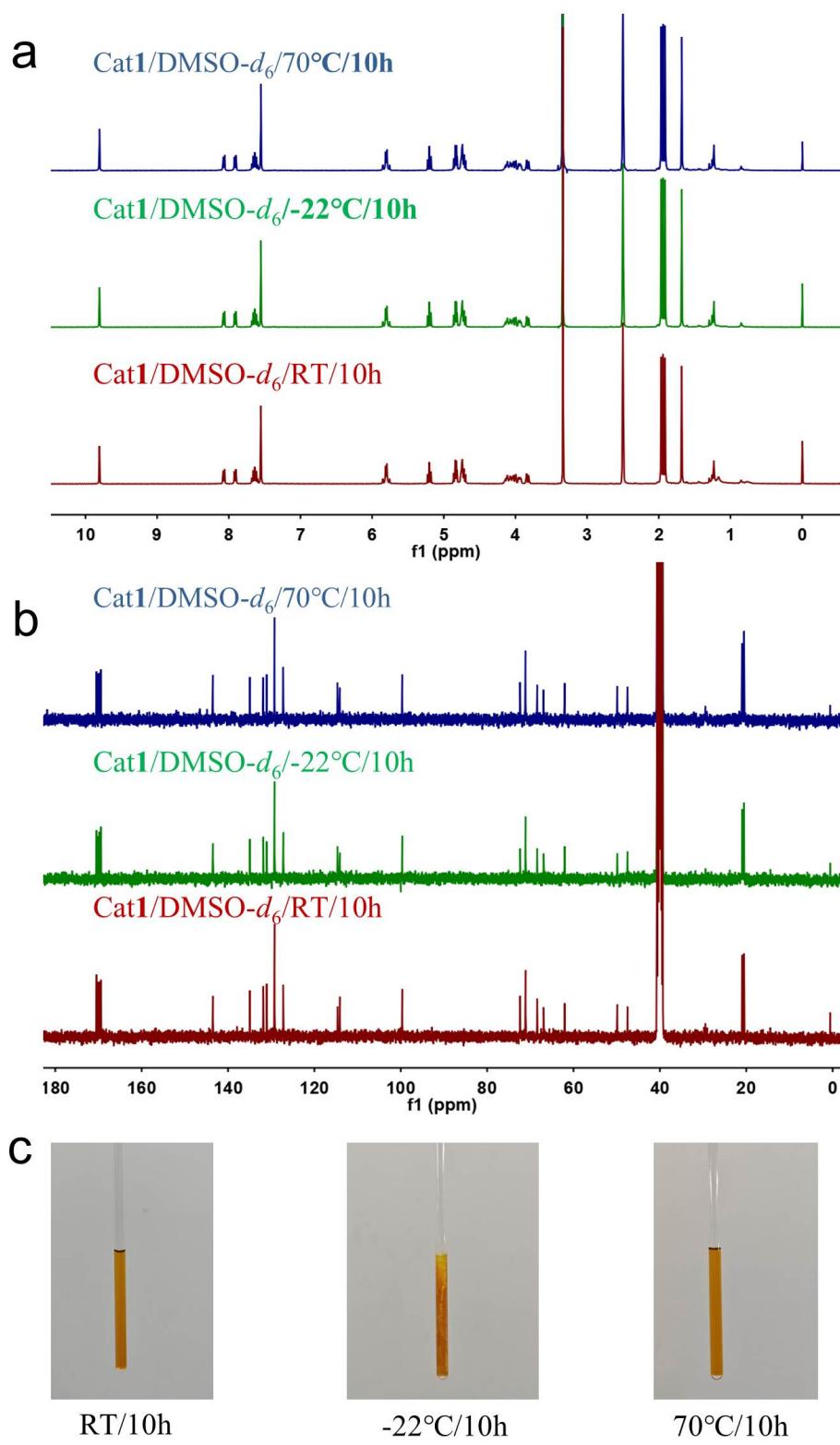


Fig. S28 NMR spectra (a and b) and color (c) of Cat1 in DMSO- d_6 at -22°C, 25-30°C, and 70°C.

4. Kinetic Experiments

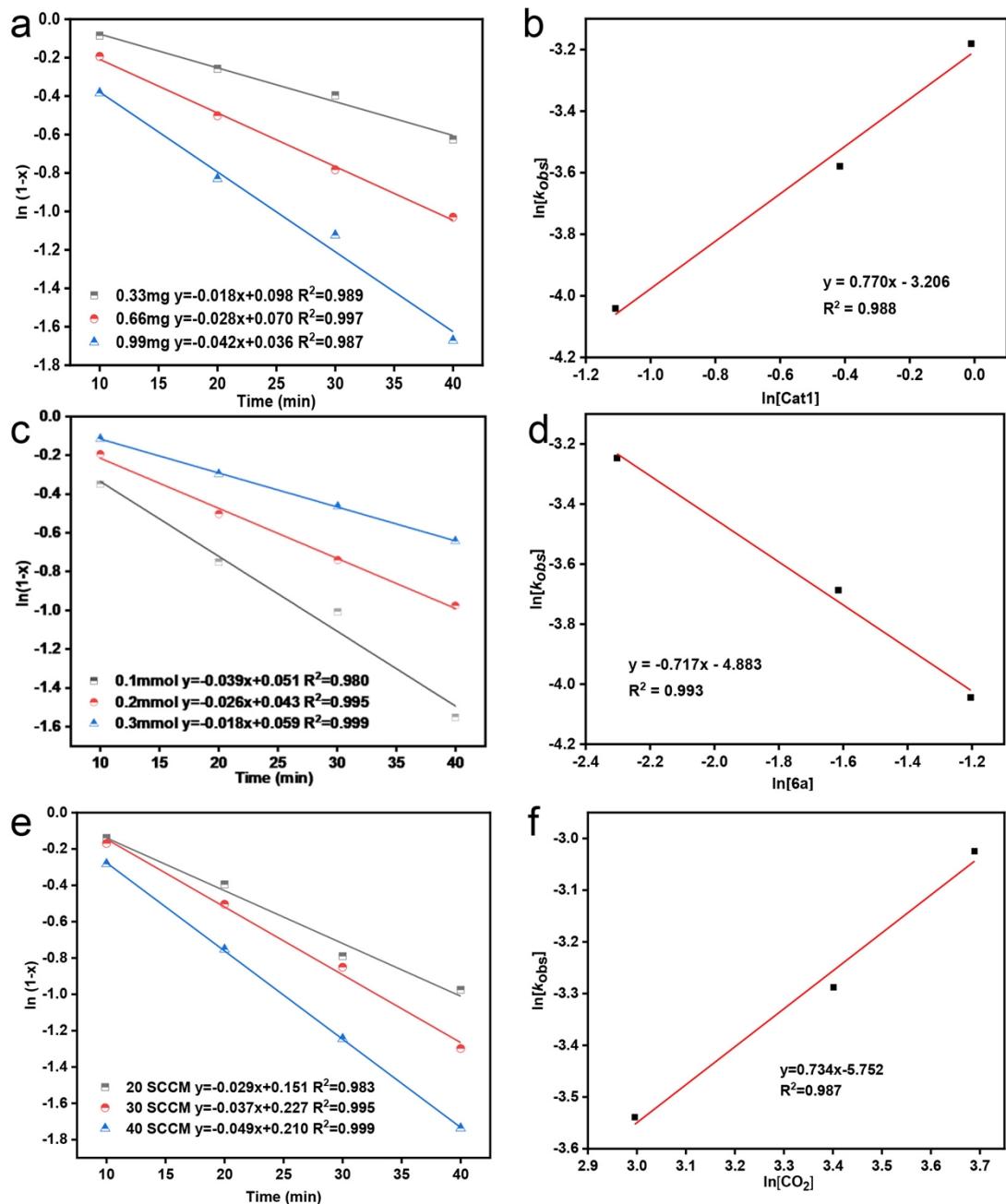


Fig. S29 a Pseudo-first order kinetic plots ($\ln(1-x)$ versus time) of three different dosages of Cat1. b Curve fitting of $\ln[k_{obs}]$ against $\ln[\text{Cat1}]$. c Pseudo-first order kinetic plots $\ln(1-x)$ versus time of three different concentrations of **6a**. d Curve fitting of $\ln[k_{obs}]$ against $\ln[6a]$; e Pseudo-first order kinetic plots ($\ln(1-x)$ versus time) of three different flow rates of CO_2 . f curve fitting of $\ln[k_{obs}]$ against $\ln[\text{CO}_2]$.

5. Gram-scale Reaction, Product Properties, and ^1H NMR

Spectra

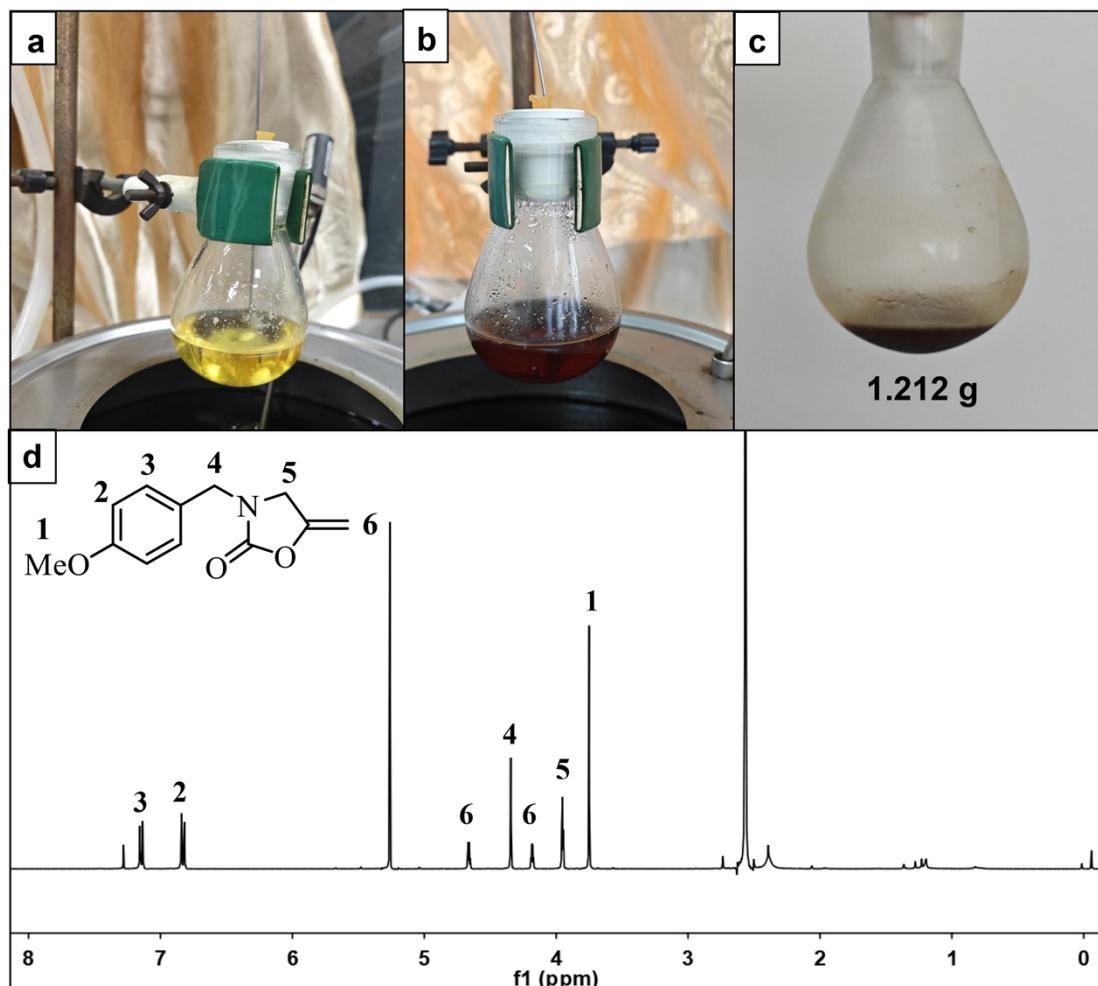


Fig. S30 Gram-scale synthesis. The reactions were carried out with **6a** (6.0 mmol, 1.05 g), Cat**1** (5.0 mol%, 6.6 mg), simulated flue gas (bubbling), NaOAc (3.0 mmol, 0.246 g), DMSO (20.0 mL) and stirred at 70 °C for 3.0 h. a System color before reaction; b System color after reaction. c Properties of purified product **7a**. d ^1H NMR of purified product **7a**.

6. Catalyst Stability Evaluation

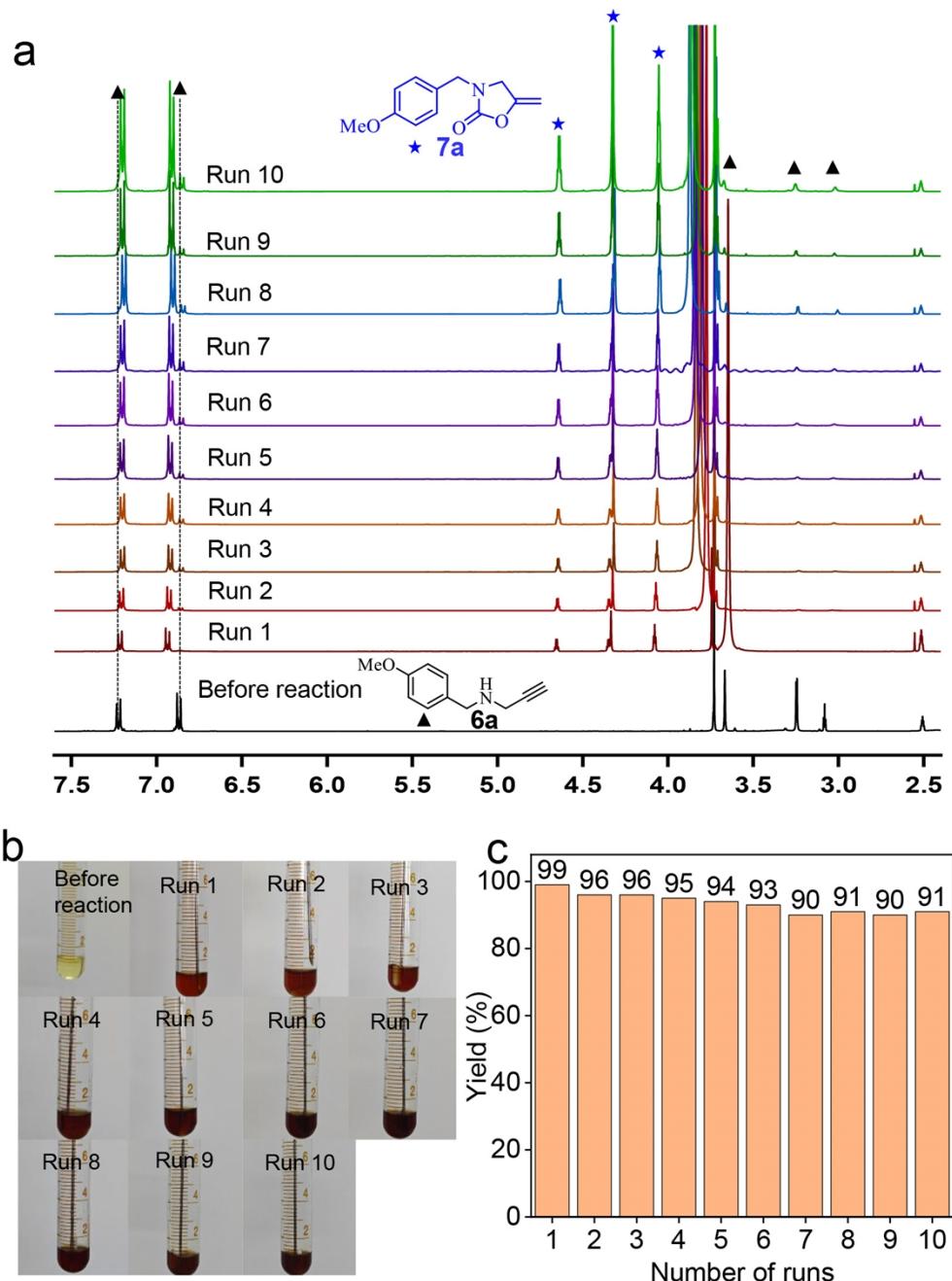


Fig. S31 The cycle catalytic performance of catalyst Cat1. Reaction conditions: **6a** (0.1 mmol, 17.5 mg), Cat1 (0.5 mol%, 0.66 mg), simulated flue gas (bubbling), NaOAc (0.3 mmol, 24.6 mg), DMSO (1.0 mL) and stirred at 70°C for 1.0 h. After each reaction, fresh **6a** was added to the catalytic system. Yields were determined by ¹H NMR. a Catalyst recycling study for Cat1. b Photos of catalytic system. c ¹H NMR tracked catalytic system.

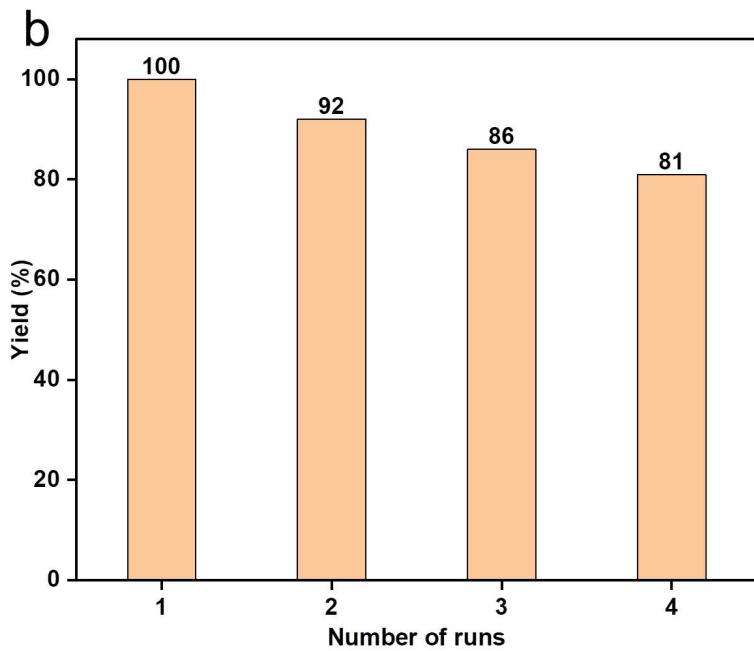
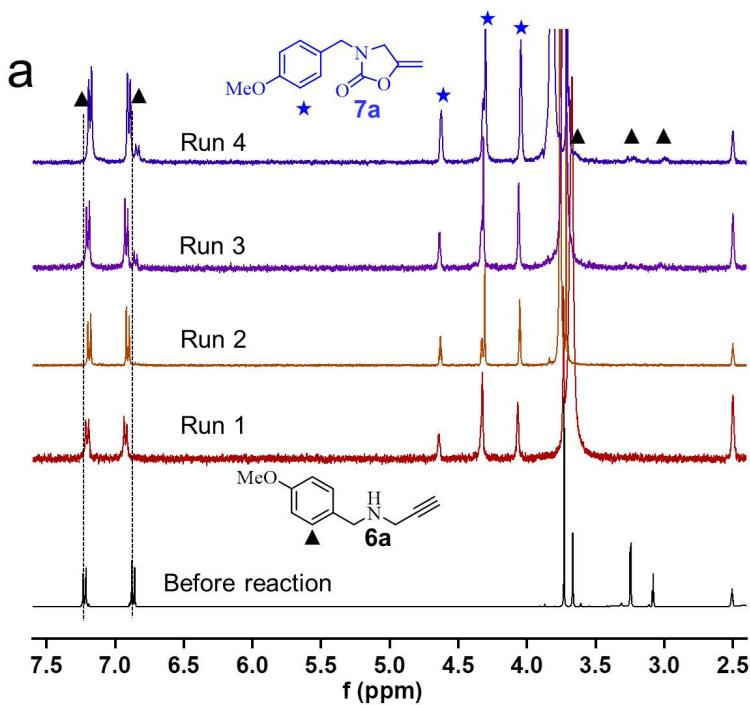


Fig. S32 The cycle catalytic performance of catalyst Cat5. Reaction conditions: **6a** (0.1 mmol, 17.5 mg), Cat5 (0.75 mol%, 0.78 mg), simulated flue gas (bubbling), NaOAc (0.3 mmol, 24.6 mg), DMSO-*d*₆ (1.0 mL) and stirred at 70°C for 1.0 h. After each reaction, fresh **6a** was added to the catalytic system. Yields were determined by ¹H NMR. a Catalyst recycling study for Cat5. b ¹H NMR tracked catalytic system.

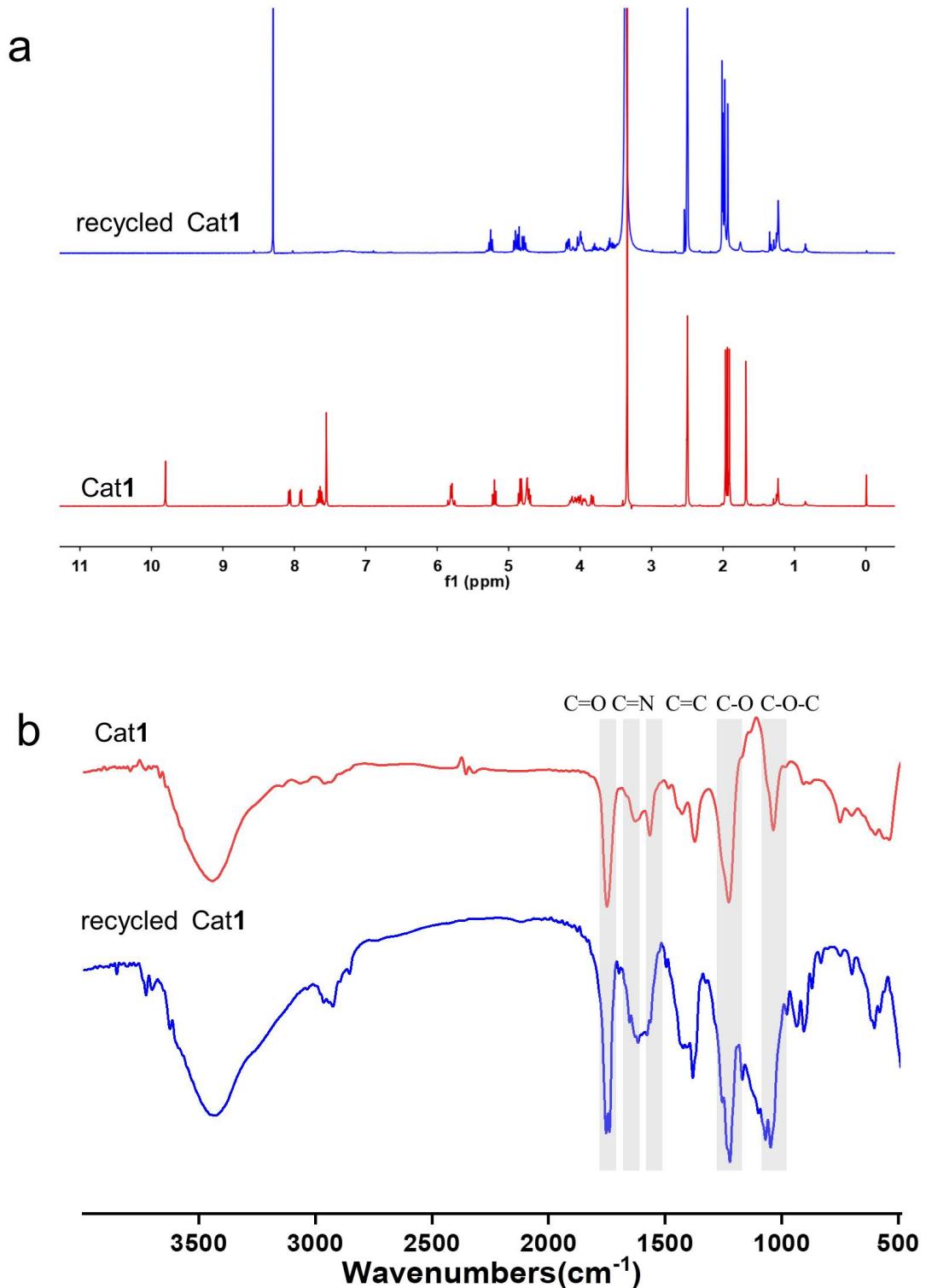


Fig. S33 a ^1H NMR spectra of Cat1 and recycled Cat1. b FT-IR spectra of Cat1 and recycled Cat1.

7. Mechanism Exploration

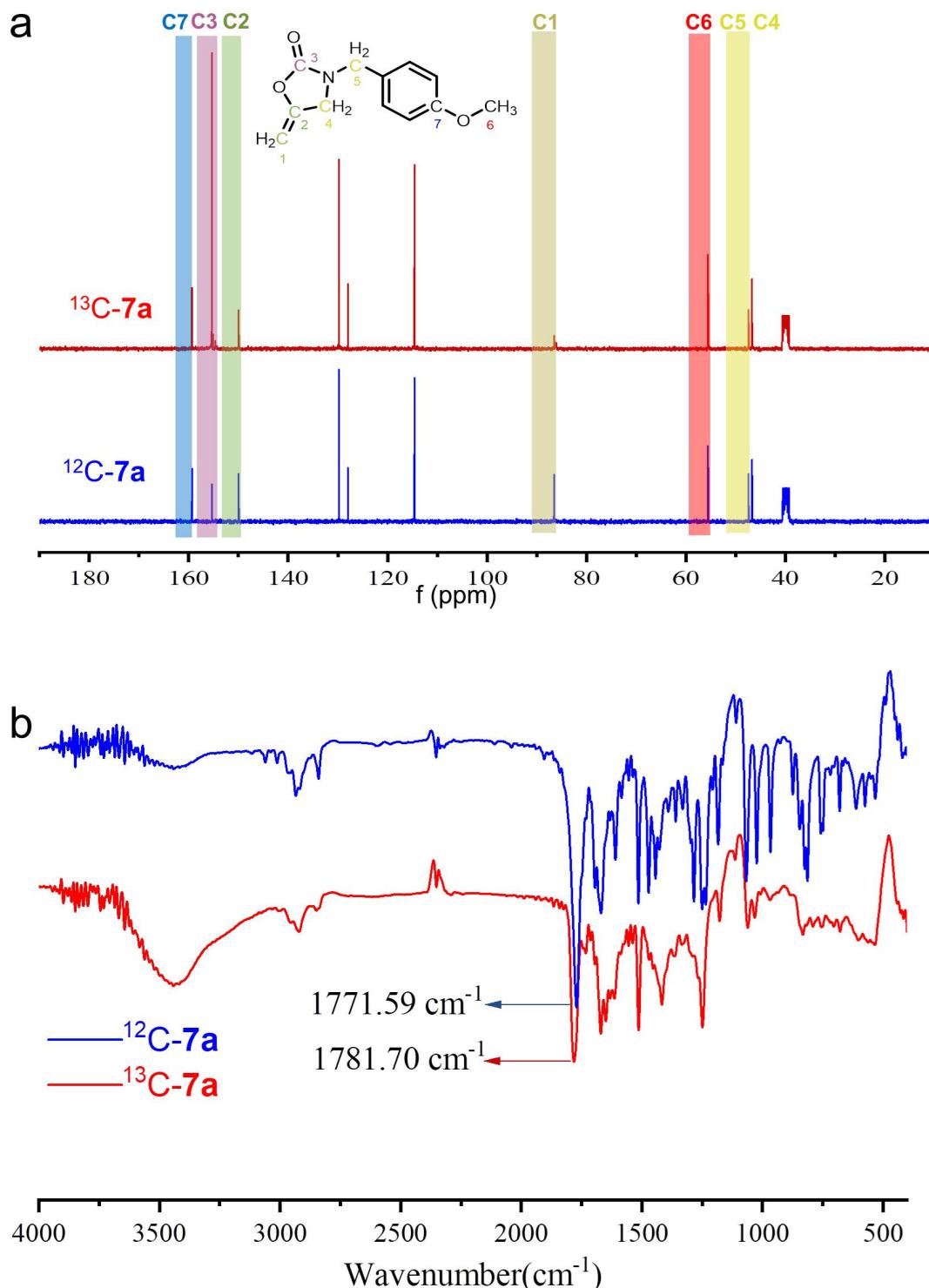


Fig. S34 a ^{13}C NMR comparison spectra of ^{13}C -isotope-labeling experiments for **7a**. b FT-IR spectra of ^{13}C -isotope-labeling experiments for **7a**.

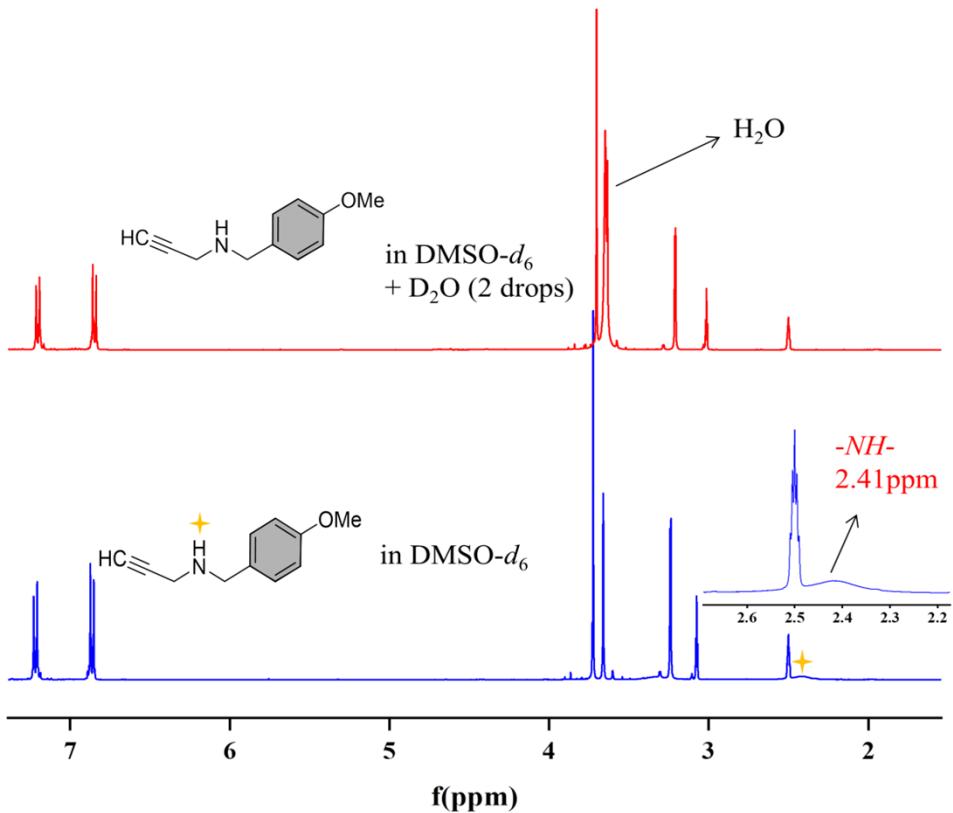


Fig. S35 Hydrogen–deuterium exchange experiment in $\text{D}_2\text{O}/\text{DMSO}-d_6$.

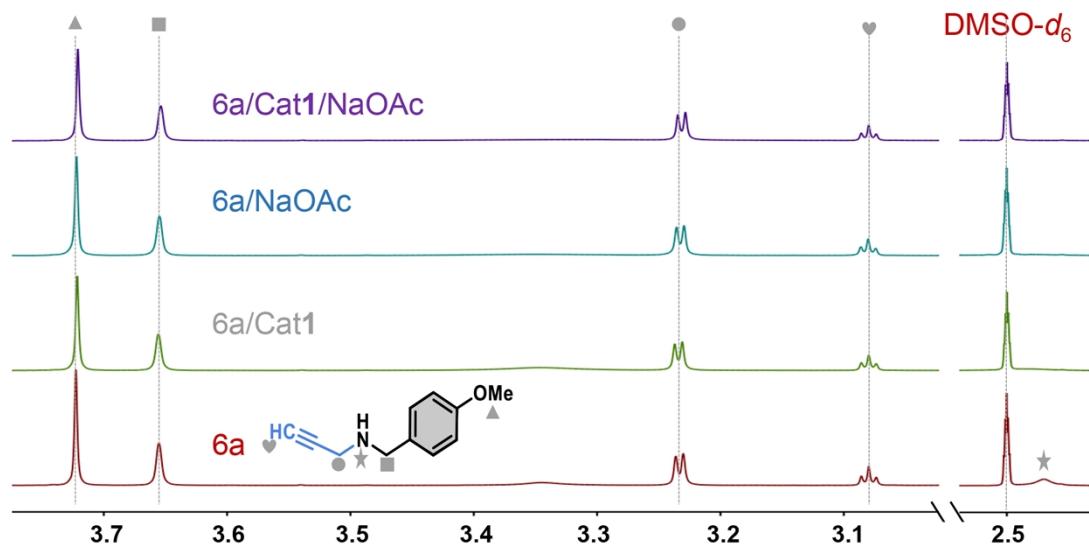


Fig. S36 Partially ^1H NMR spectra monitoring of substrate **6a**, **6a/Cat1**, **6a/NaOAc**, and **6a/Cat1/NaOAc** in $\text{DMSO}-d_6$.

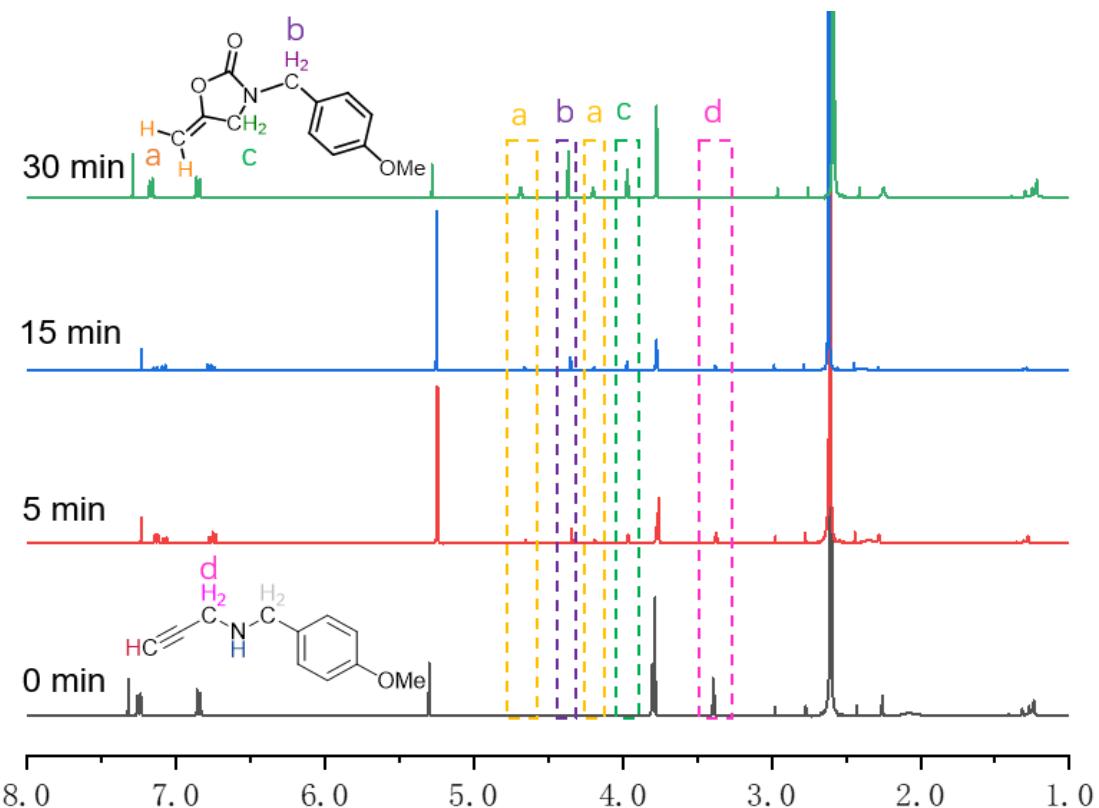


Fig. S37 Partially ^1H NMR spectra monitoring of Cat1-catalyzed carboxylative cyclization of **6a** and CO_2 at 70°C .

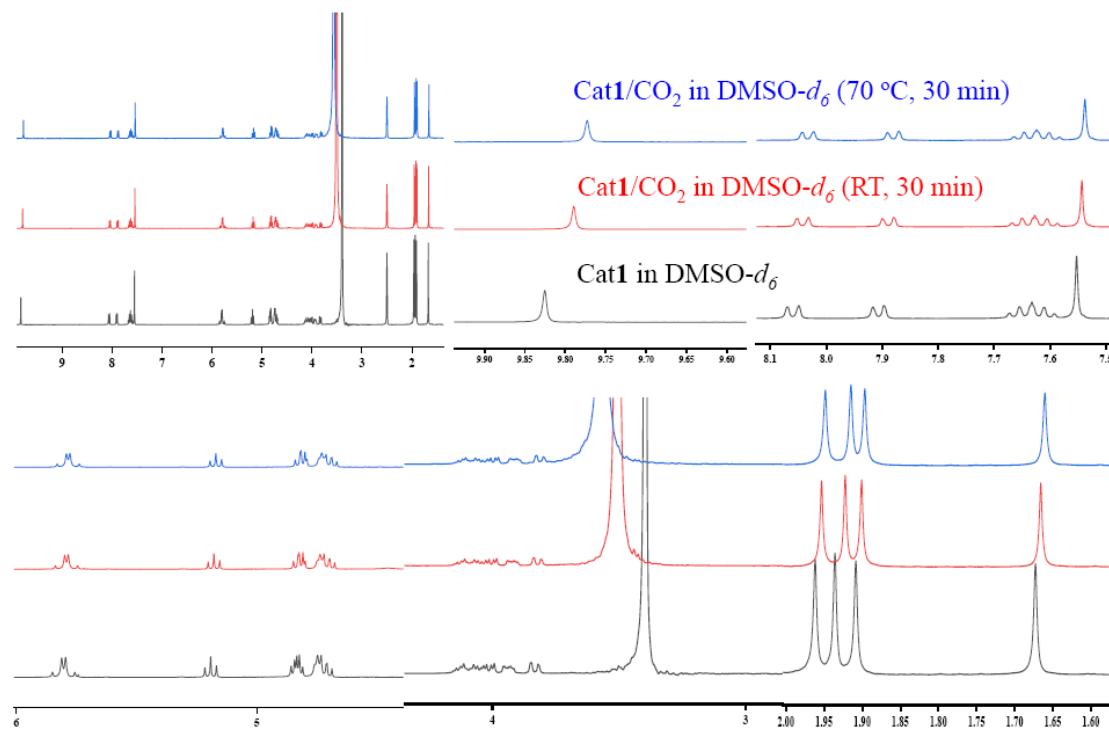


Fig. S38 The interactions between Cat1 and CO_2 were investigated by ^1H NMR.

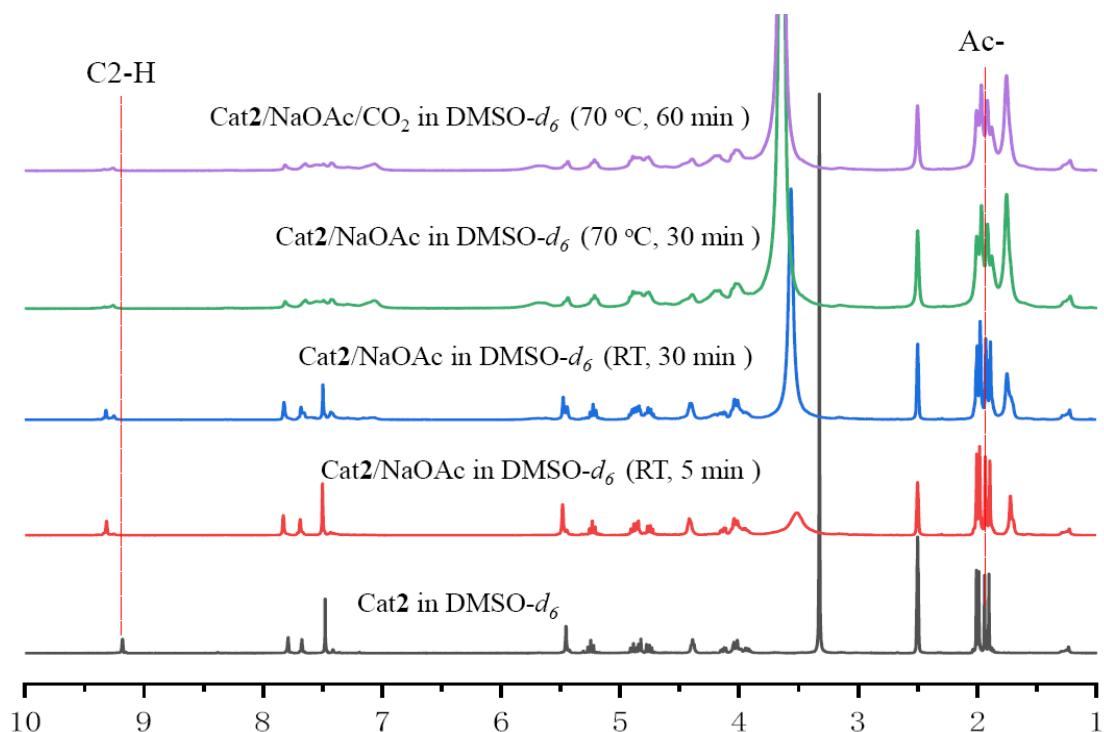


Fig. S39 The interactions between **Cat2** and NaOAc were investigated by ¹H NMR.

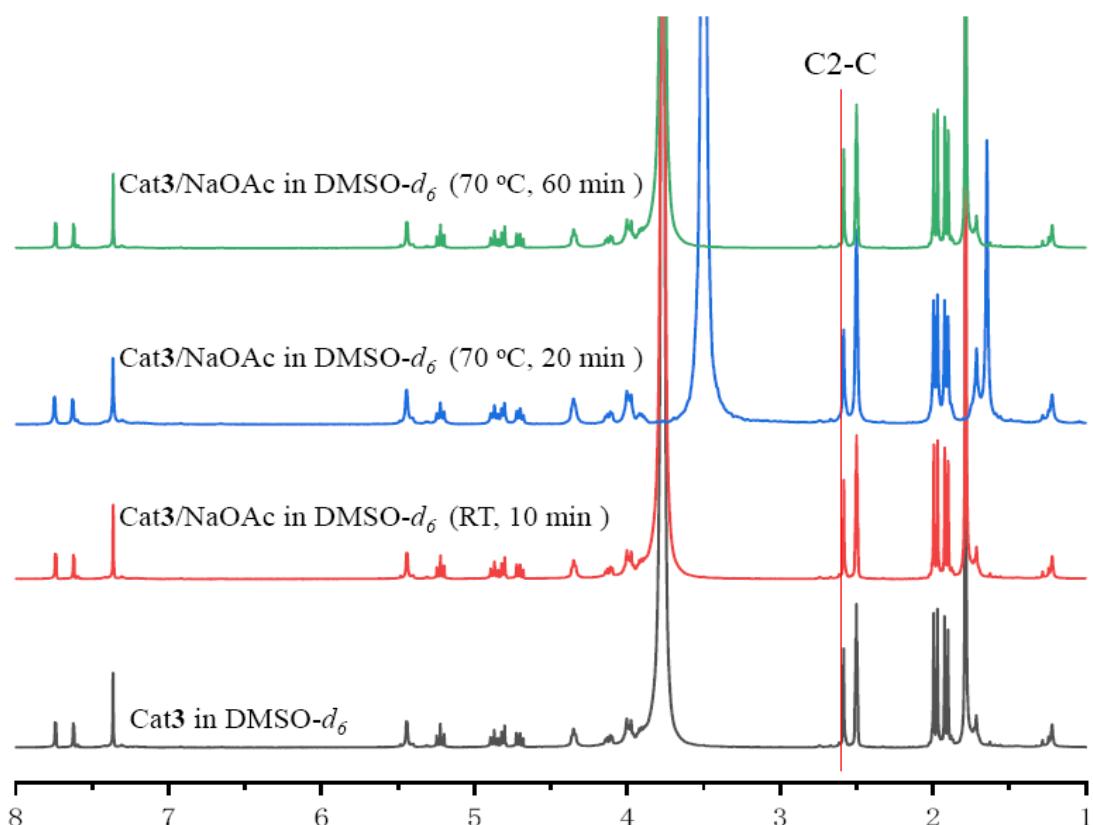


Fig. S40 The interactions between **Cat3** and NaOAc were investigated by ¹H NMR.

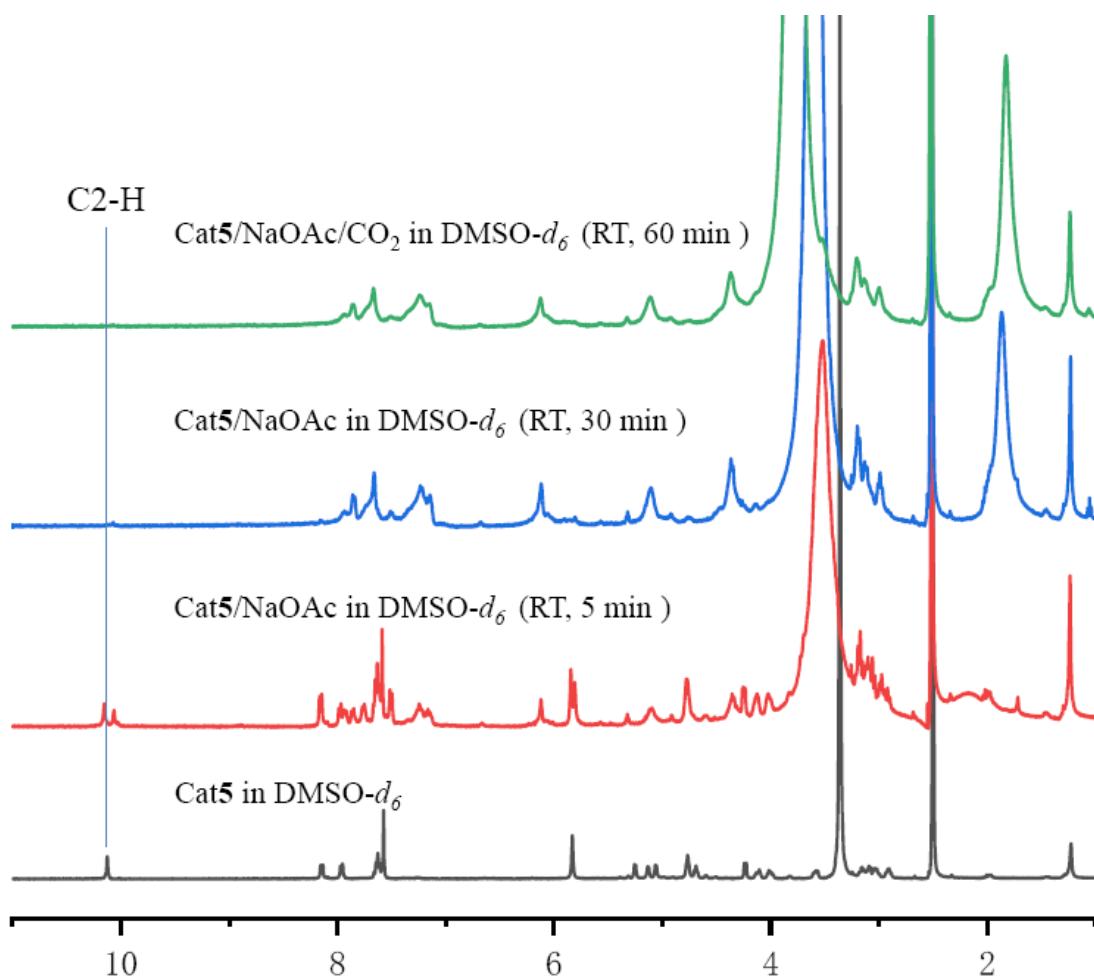


Fig. S41 The interactions between **Cat5**, NaOAc, and CO₂ were investigated by ¹H NMR.

7. NMR Spectra of Imidazolium Salts

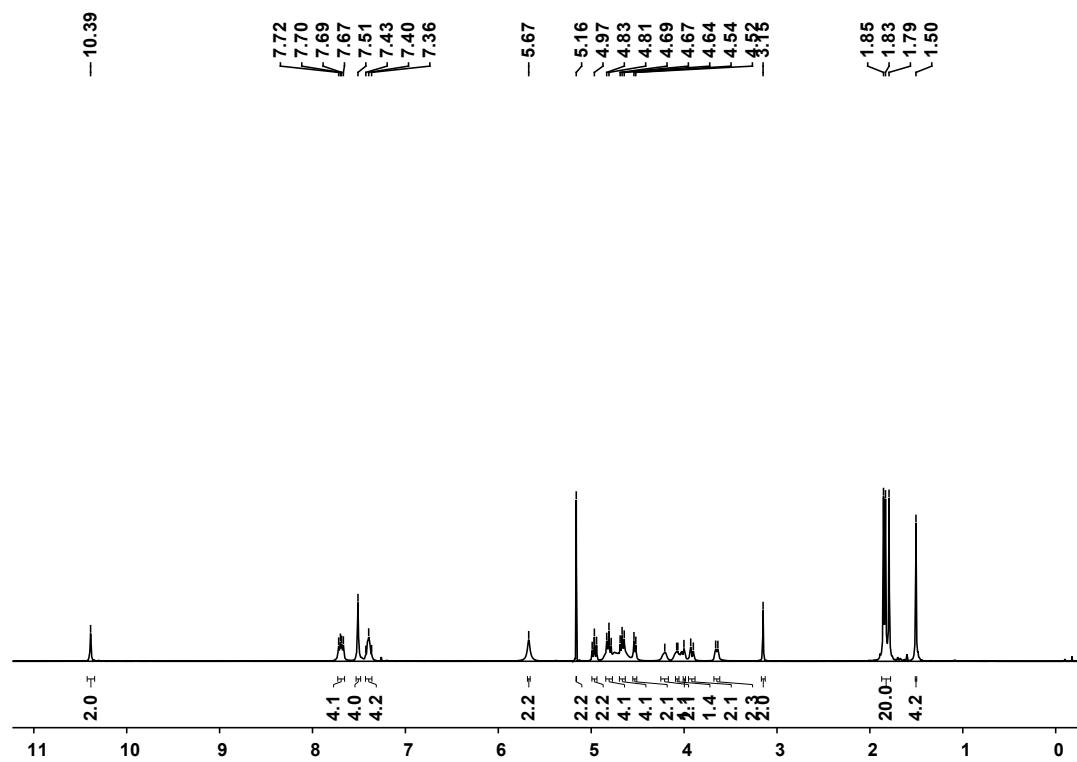


Fig. S42 ¹H NMR of Im-Br1 in CDCl₃.

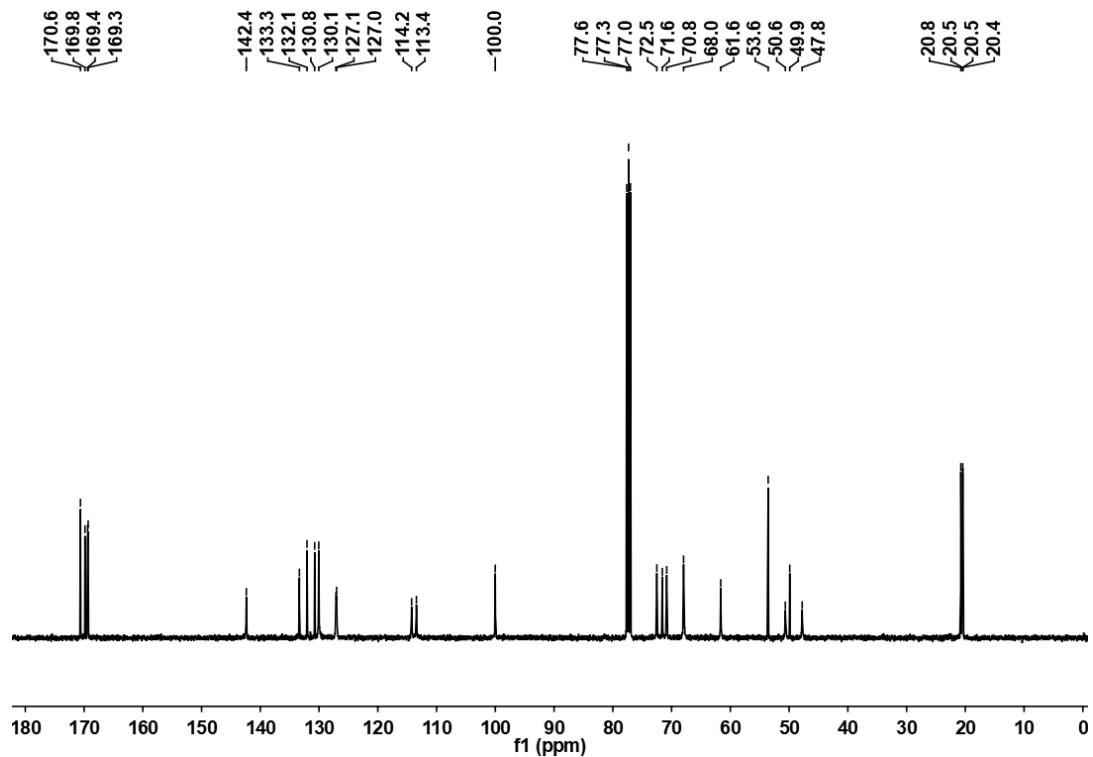


Fig. S43 ¹³C NMR of Im-Br1 in CDCl₃.

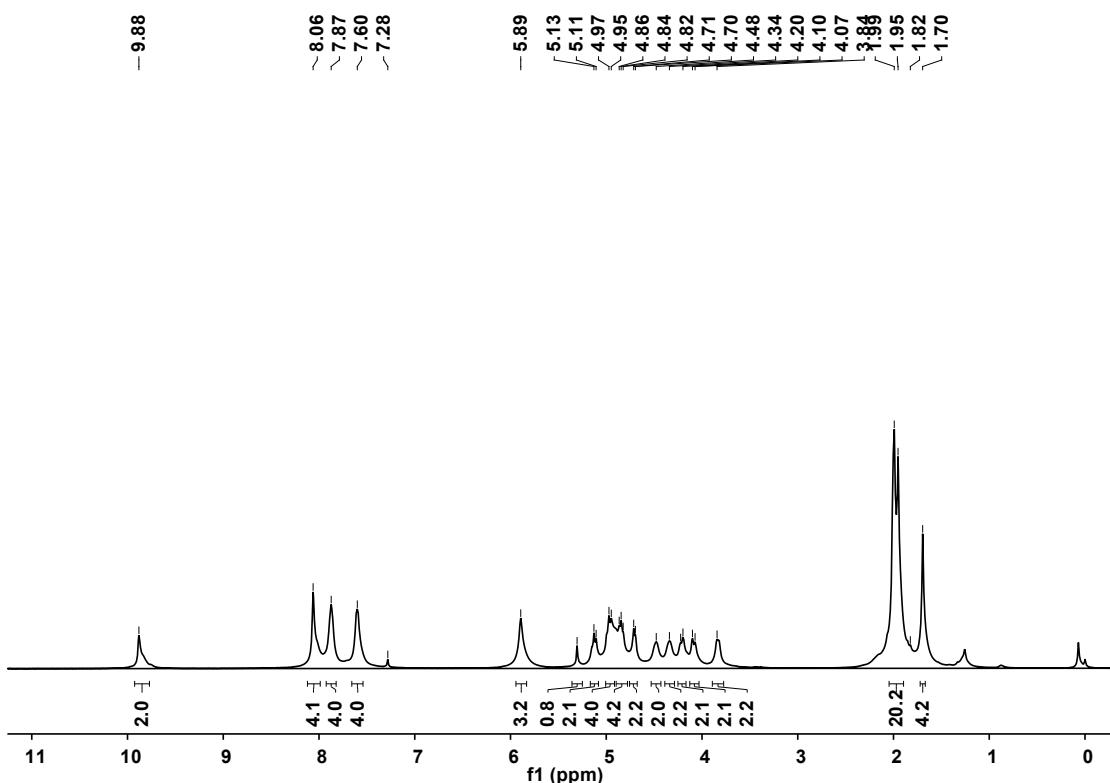


Fig. S44 ^1H NMR of Cat1 in CDCl_3 .

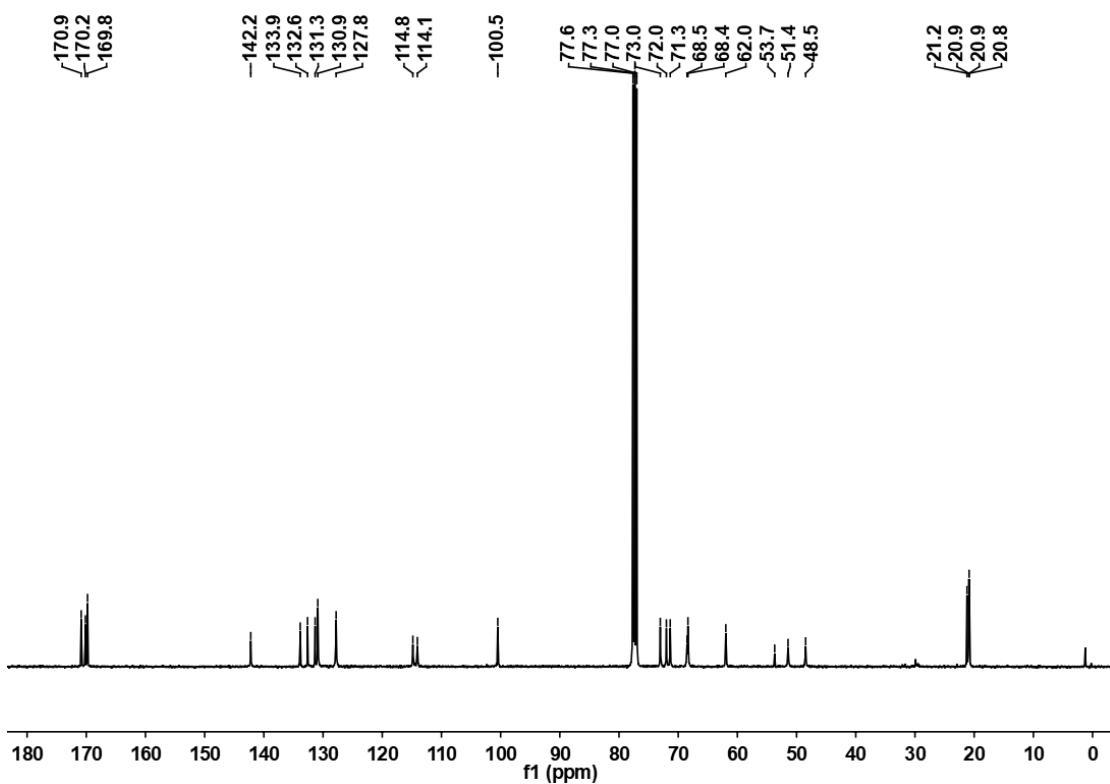


Fig. S45 ^{13}C NMR of Cat1 in CDCl_3 .

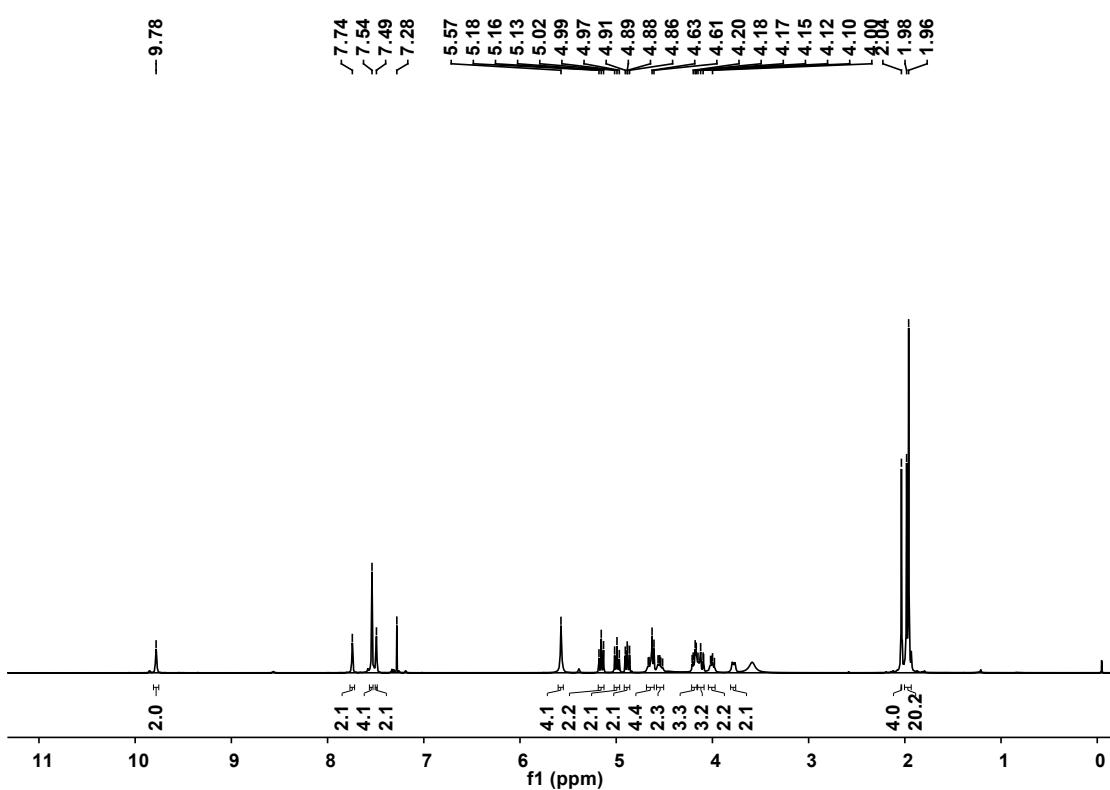


Fig. S46 ^1H NMR of Im-Br2 in CDCl_3 .

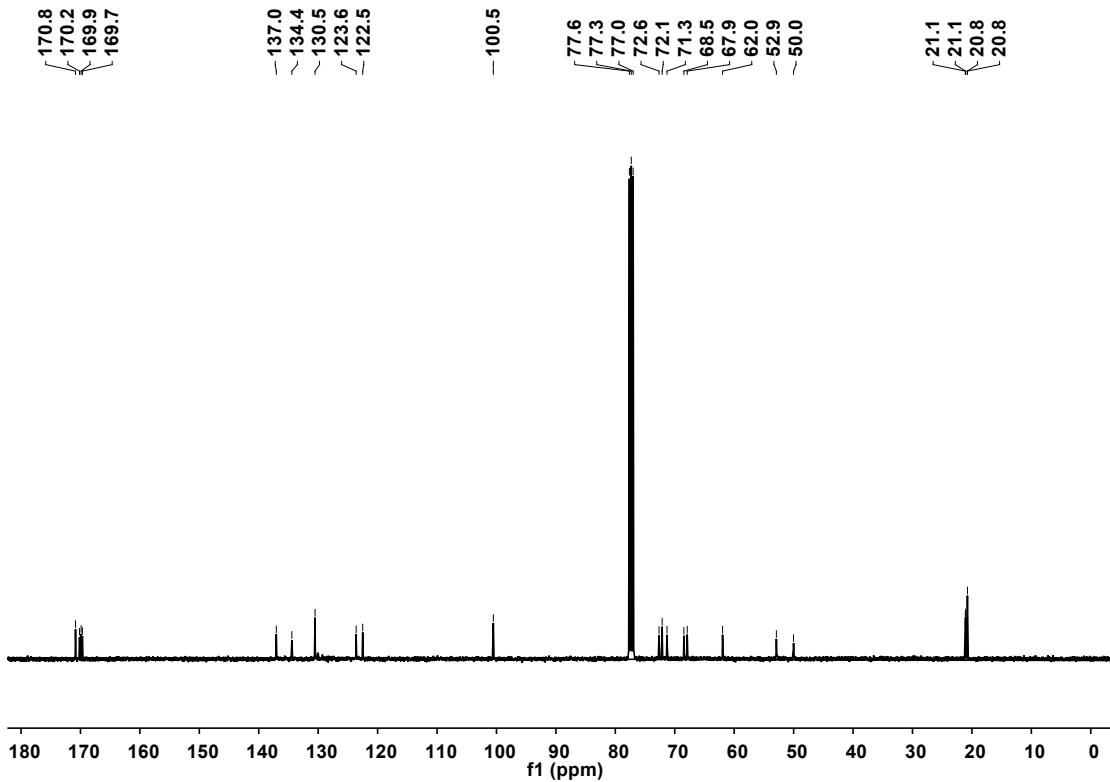


Fig. S47 ^{13}C NMR of Im-Br2 in CDCl_3 .

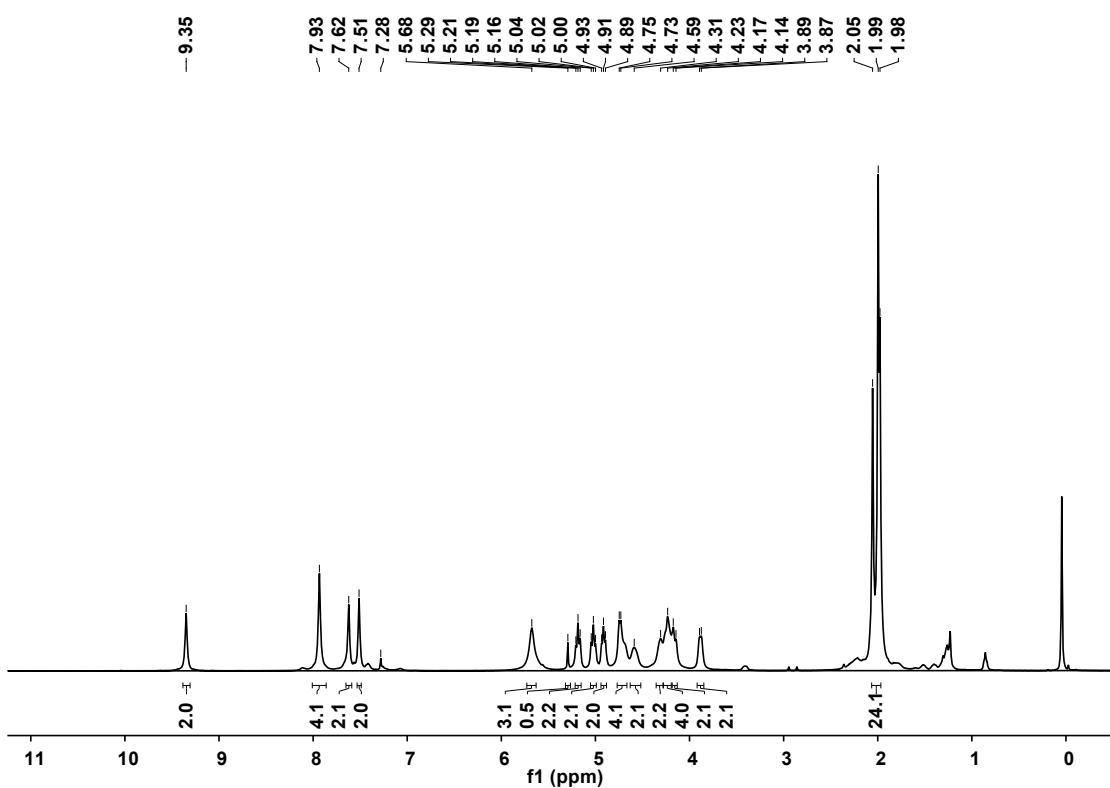


Fig. S48 ^1H NMR of Cat2 in CDCl_3 .

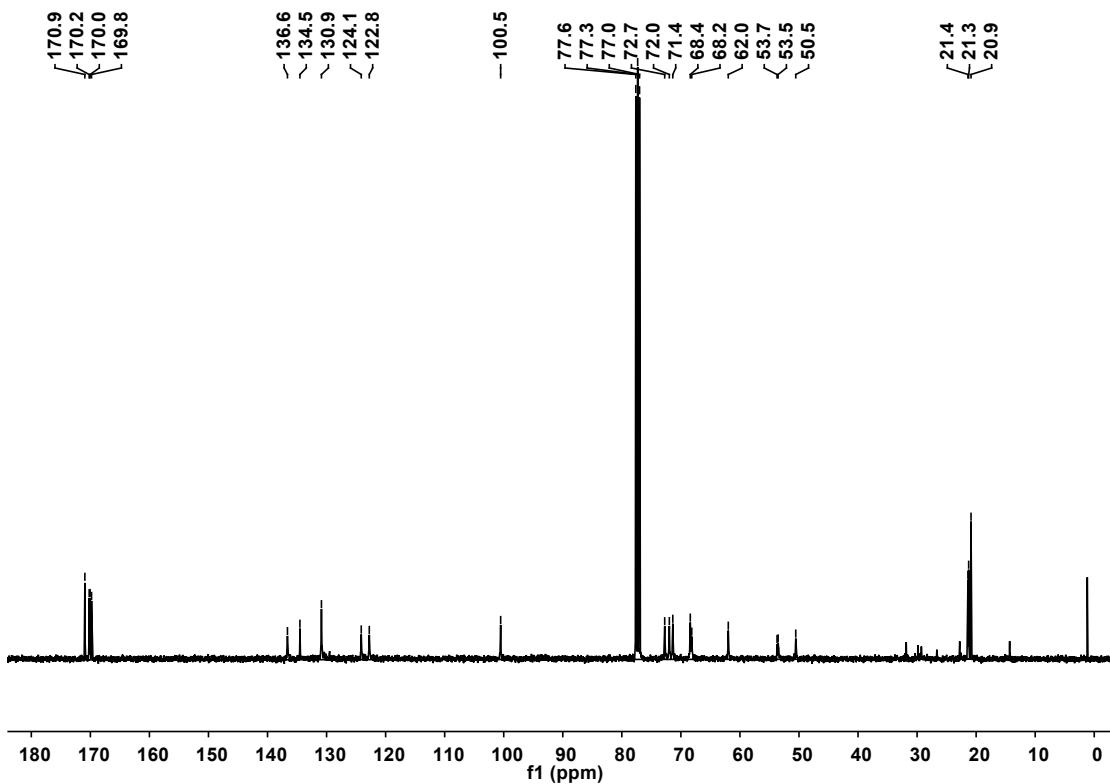


Fig. S49 ^{13}C NMR of Cat2 in CDCl_3 .

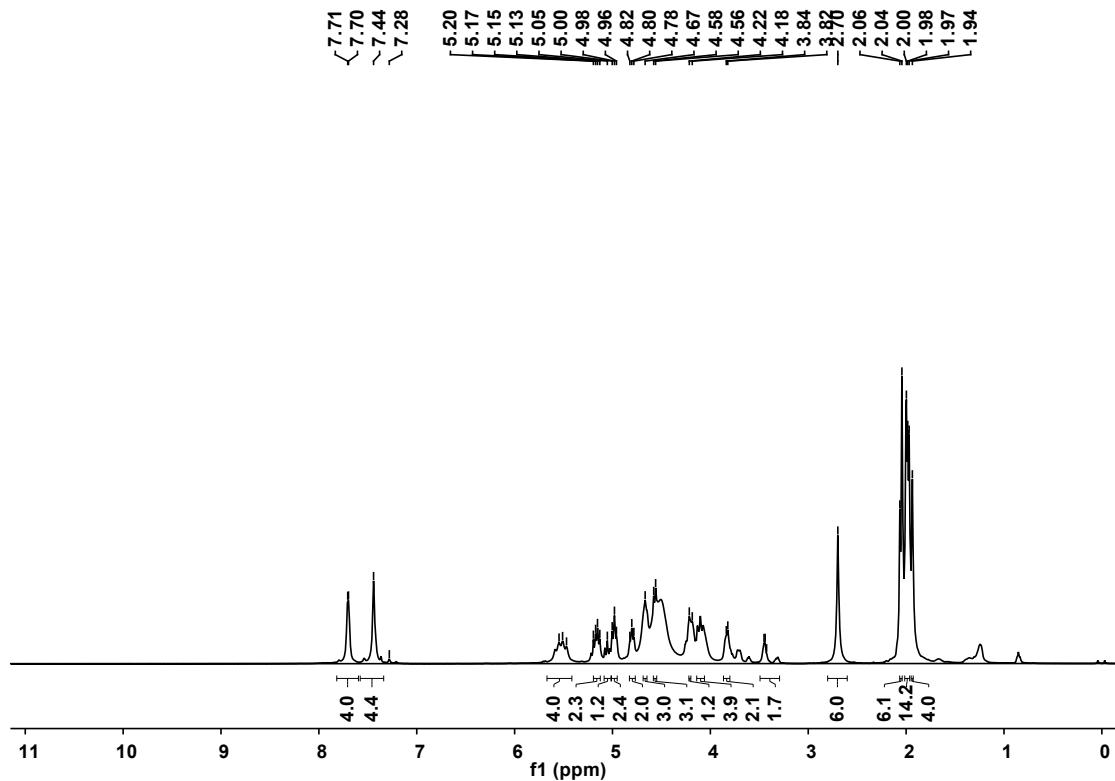


Fig. S50 ^1H NMR of Im-Br3 in CDCl_3 .

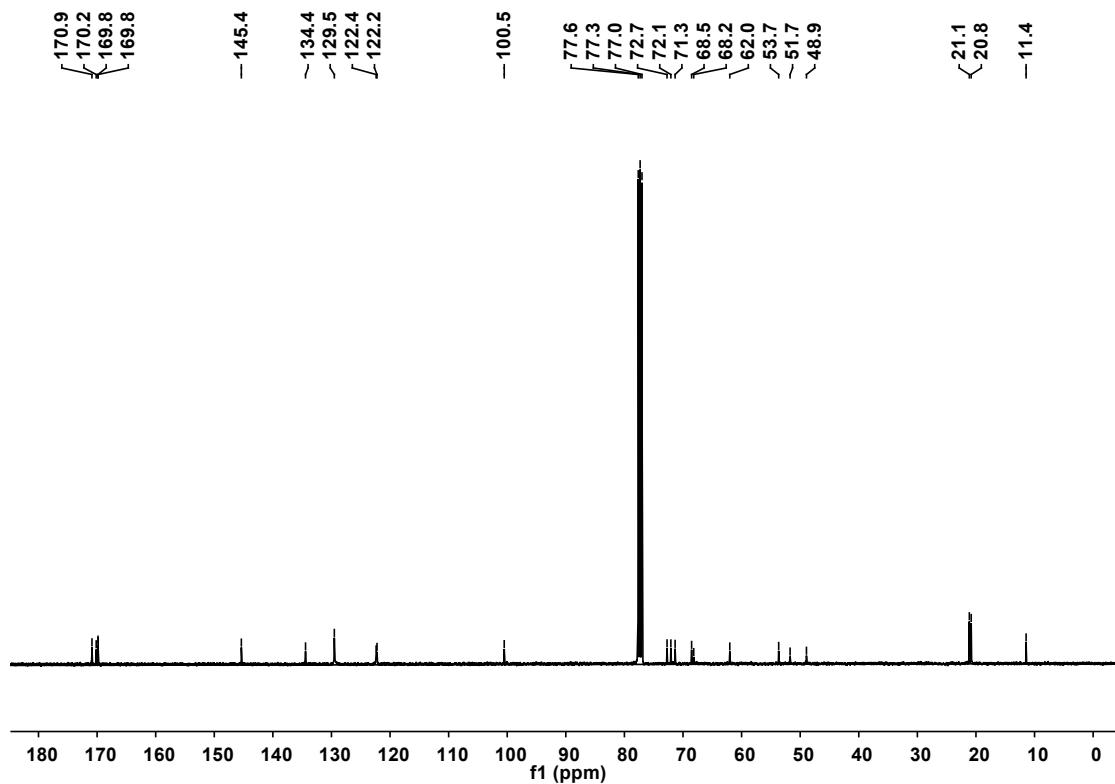


Fig. S51 ^{13}C NMR of Im-Br3 in CDCl_3 .

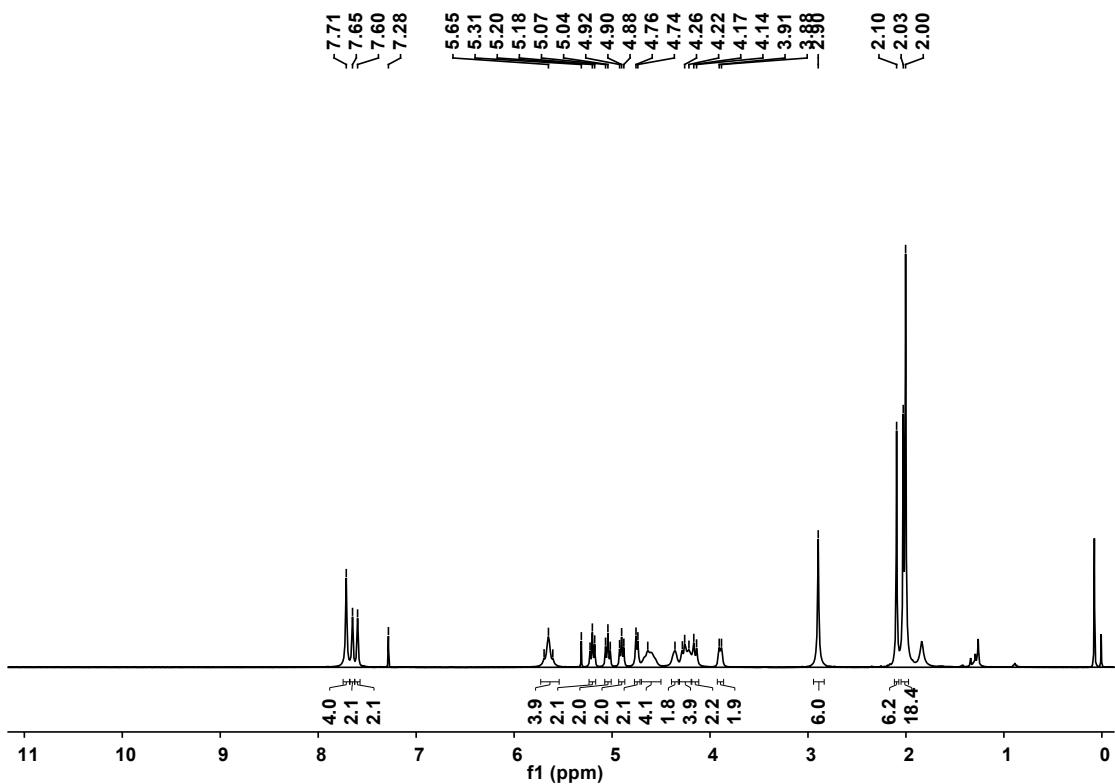


Fig. S52 ^1H NMR of Cat3 in CDCl_3 .

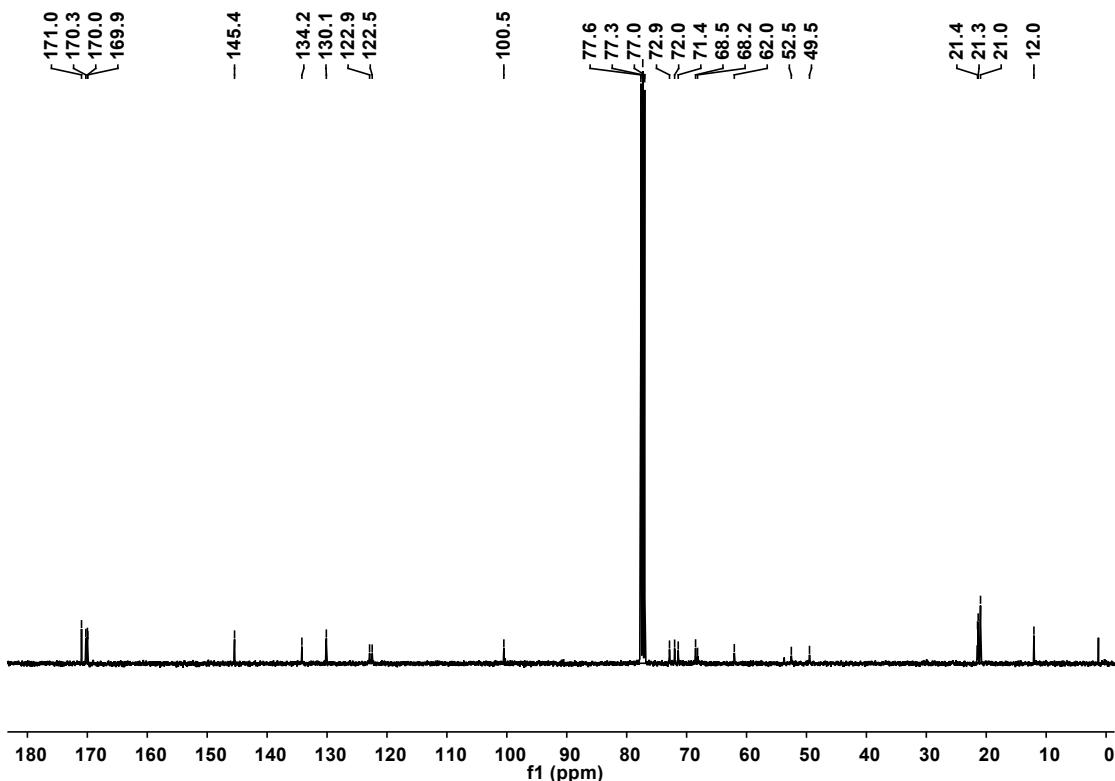


Fig. S53 ^{13}C NMR of Cat3 in CDCl_3 .

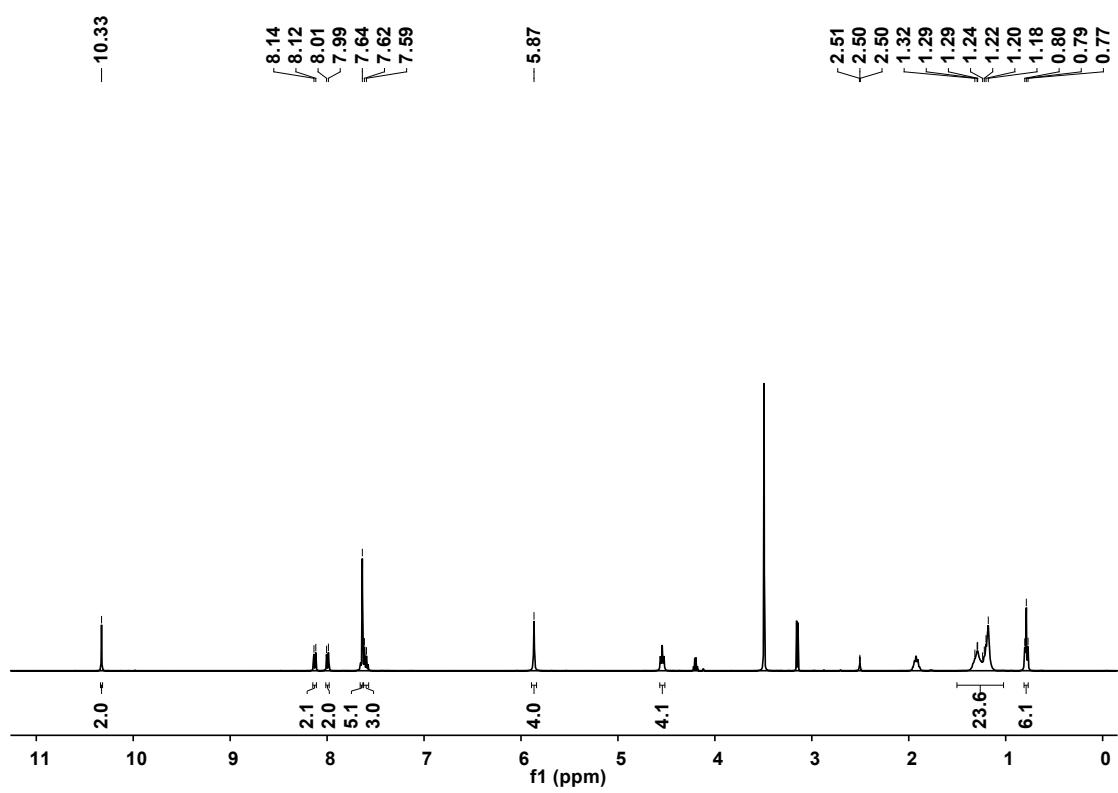


Fig. S54 ^1H NMR of Im-Br4 in $\text{DMSO}-d_6$.

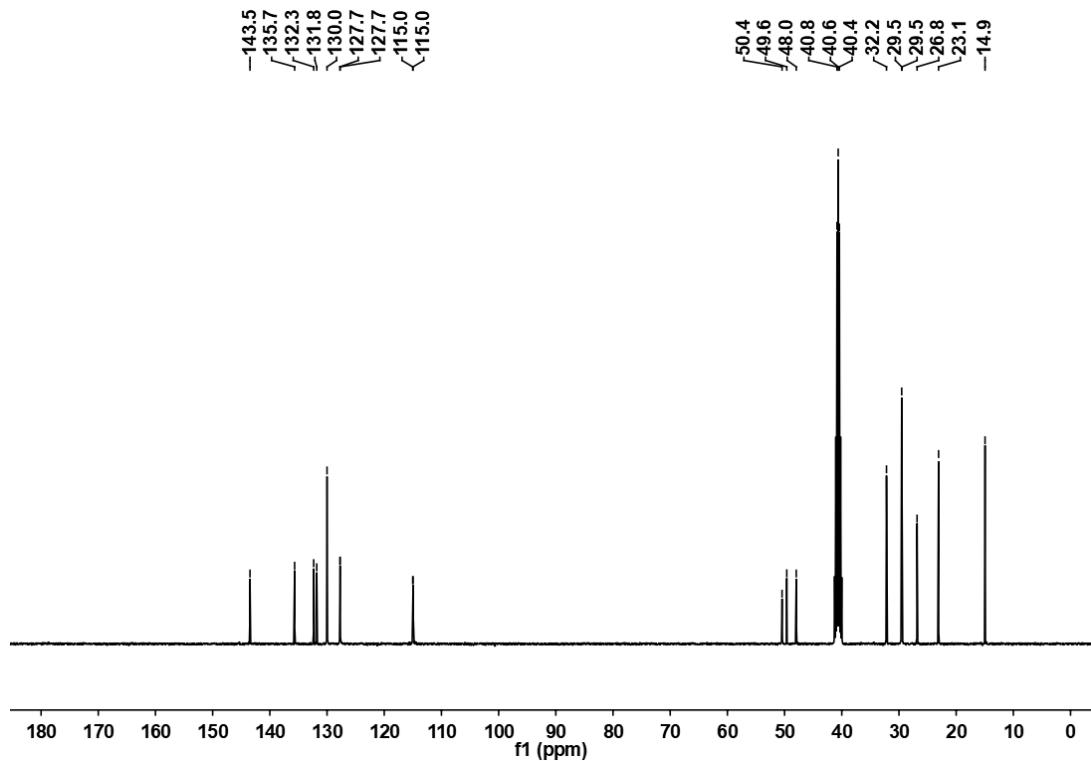


Fig. S55 ^{13}C NMR of Im-Br4 in $\text{DMSO}-d_6$.

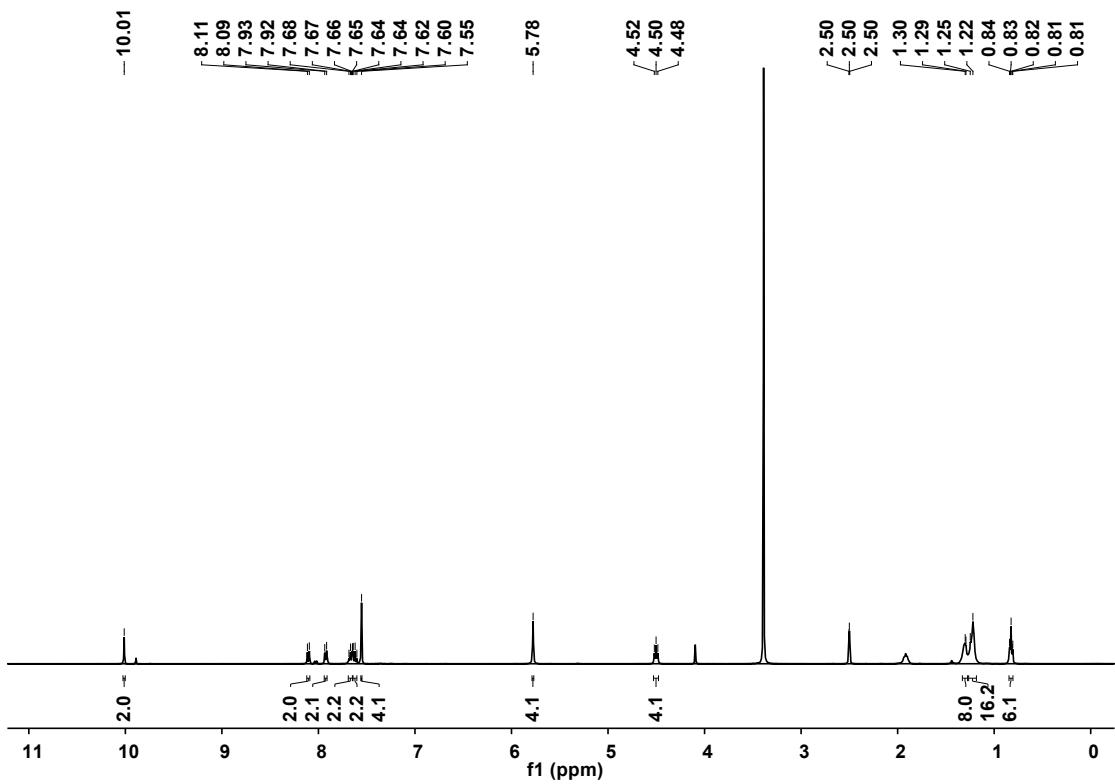


Fig. S56 ^1H NMR of Cat4 in $\text{DMSO}-d_6$.

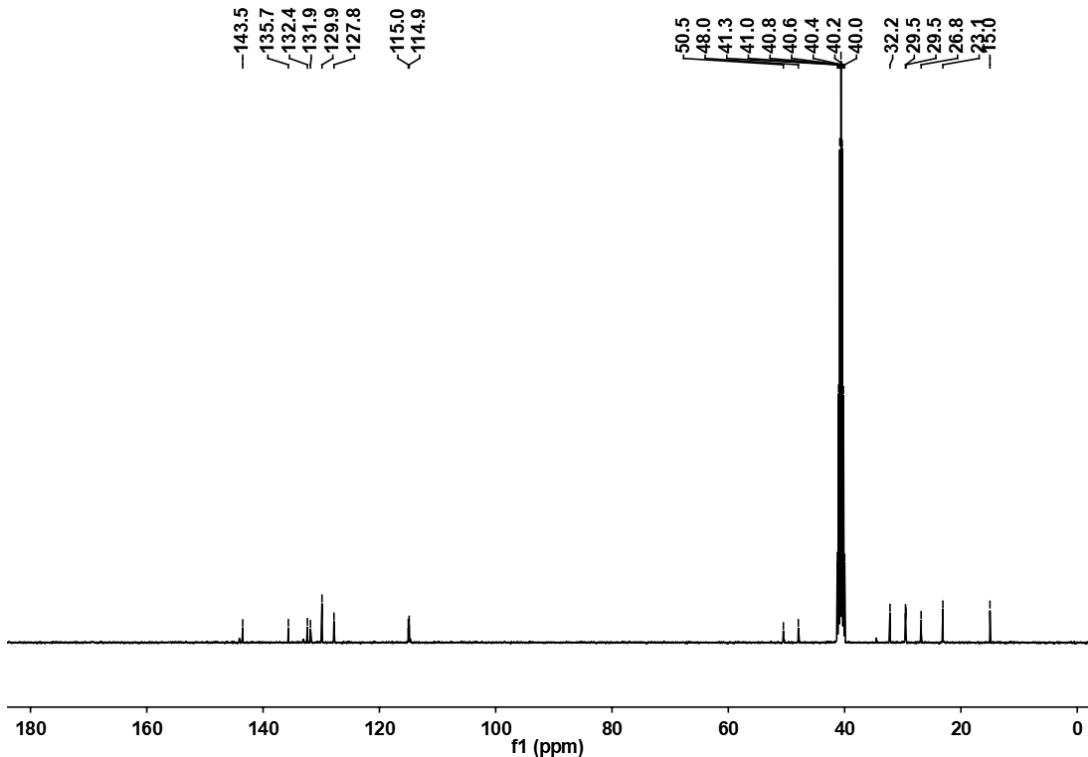


Fig. S57 ^{13}C NMR of Cat4 in $\text{DMSO}-d_6$.

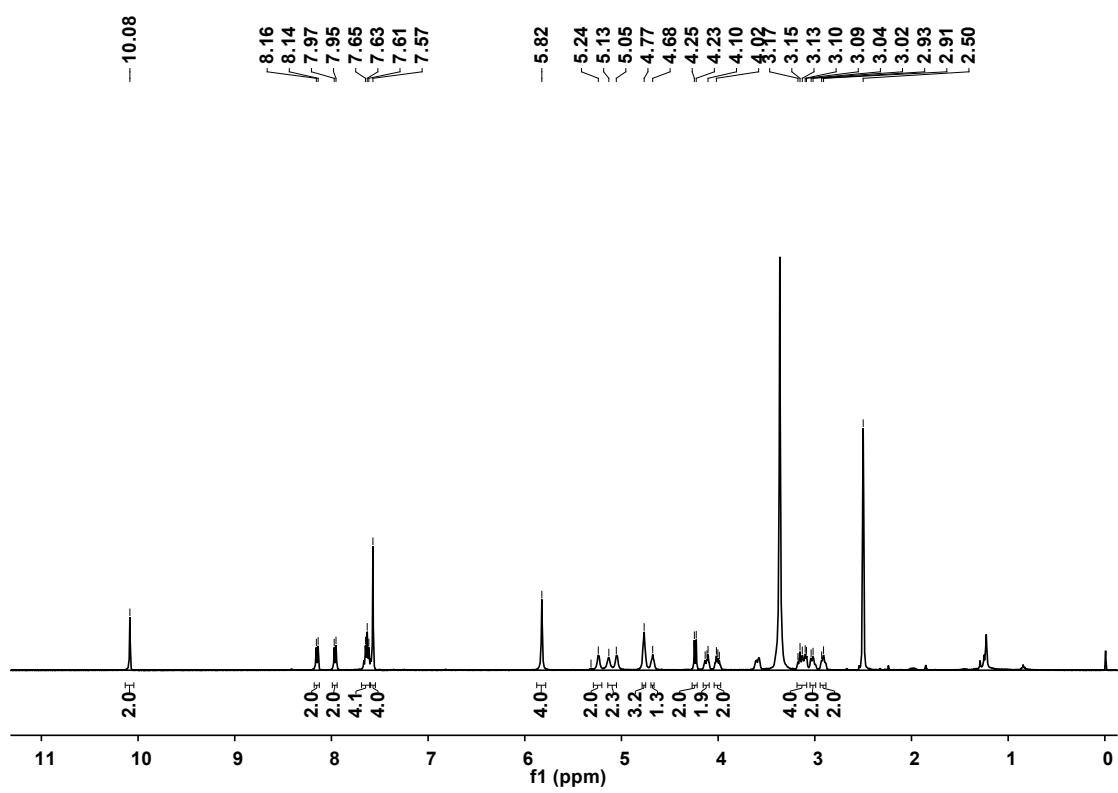


Fig. S58 ^1H NMR of Im-Br**5** in $\text{DMSO}-d_6$.

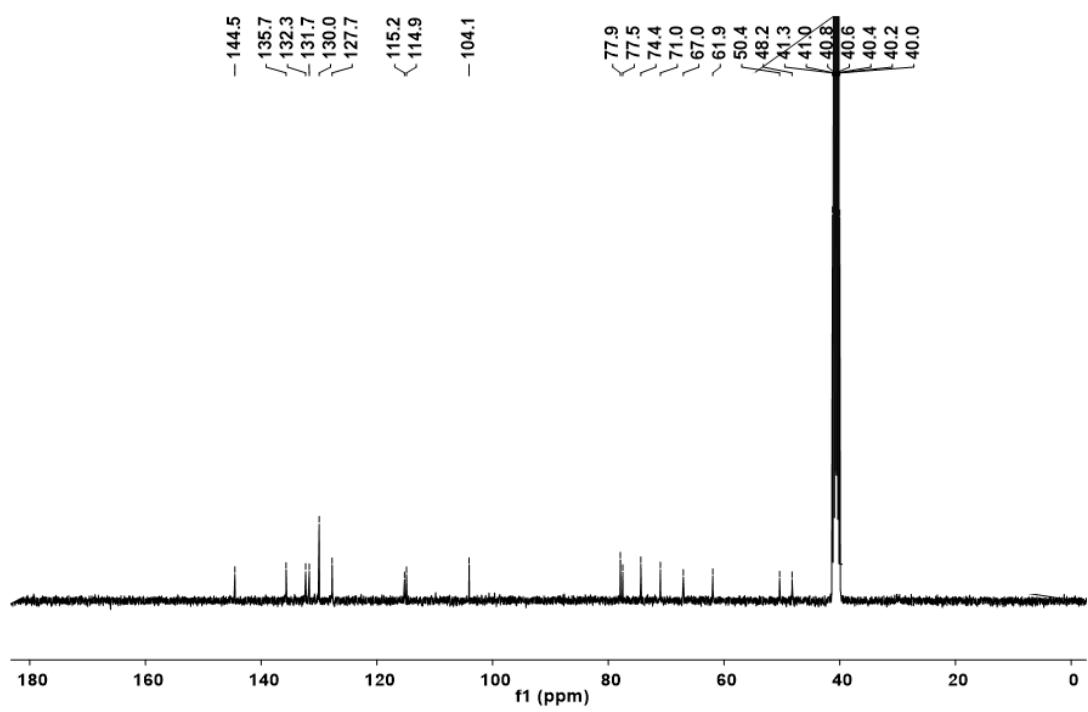


Fig. S59 ^{13}C NMR of Im-Br5 in $\text{DMSO}-d_6$.

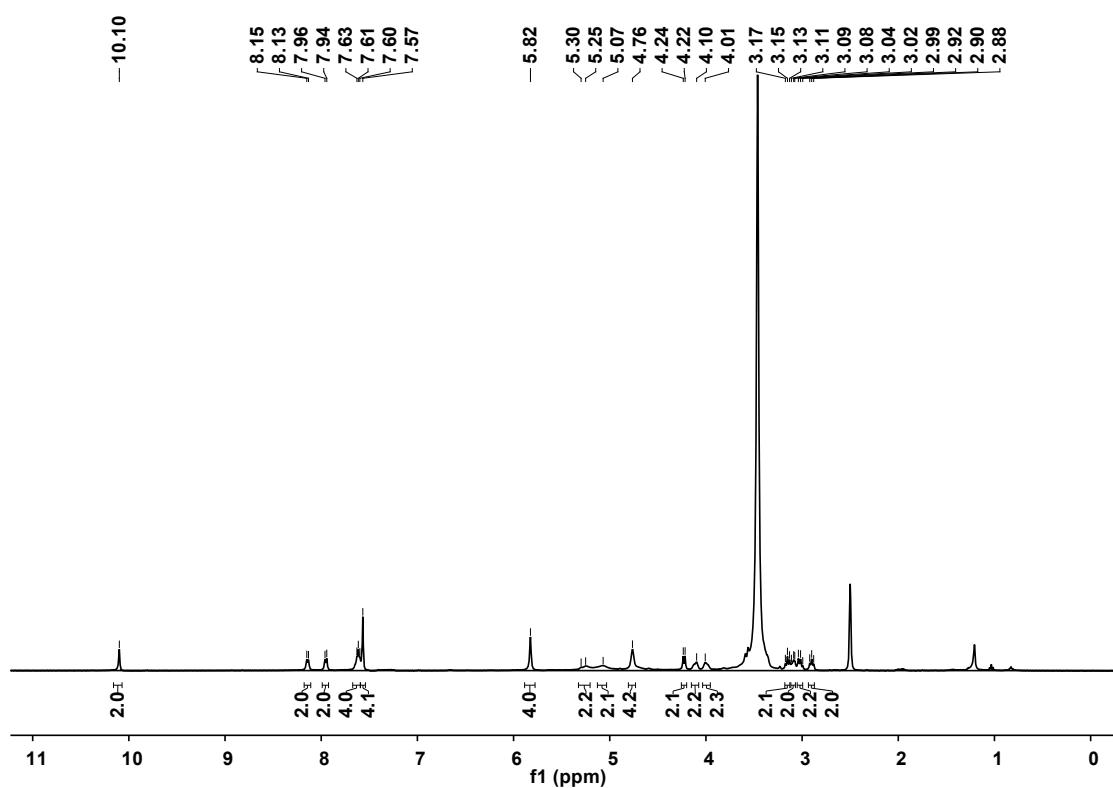


Fig. S60 ^1H NMR of Cat5 in $\text{DMSO}-d_6$.

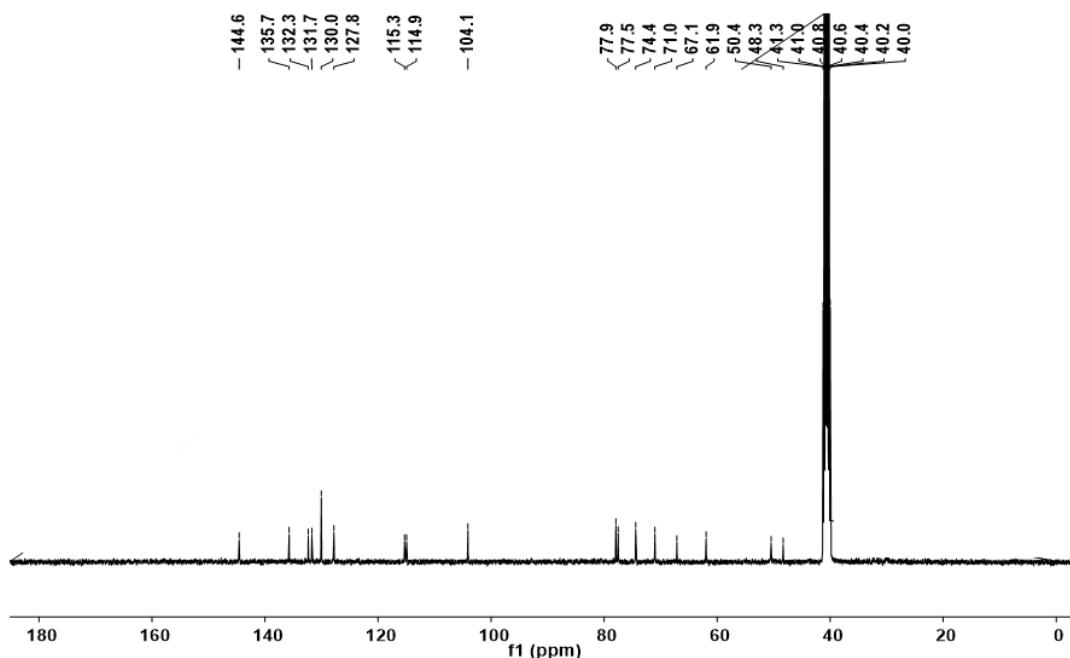


Fig. S61 ^{13}C NMR of Cat5 in $\text{DMSO}-d_6$.

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