

**Enhancing Biofouling Resistance in Microfiltration Membranes through Capsaicin-Derivative
Functionalization – Supporting Information**

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Text S1 Synthesis of P(VDF-CTFE)-g-PMAA

The synthesis of P(VDF-CTFE)-g-PMAA was carried out in two stages, as previously described in our study [1]. In the first stage, the carbon chloride bond on the P(VDF-CTFE) backbone was grafted with tert-butyl methacrylate (tBMA) through atom transfer radical polymerization (ATRP), followed by hydrolysis to obtain P(VDF-CTFE)-g-PMAA with carboxyl functional groups. To initiate this process, P(VDF-CTFE) with a chlorine content of 2.4 mmol (1.5 g) and CuCl (238 mg, 2.4 mmol) were placed in a 100 mL Schlenk flask under a nitrogen atmosphere. The flask was subjected to three cycles of vacuum extraction and nitrogen refilling, followed by the addition of 25 mL of N-methyl-2-pyrrolidone (NMP) which was dissolved by magnetic stirring. A mixture of tert-butyl methacrylate (96 mmol, 15.27 mL) and NMP (5 mL) containing PMDETA (0.24 mmol, 0.5 mL) was then added. The system was heated to 80 °C in an oil bath under a nitrogen atmosphere and allowed to proceed for 20 h before being quenched by exposing the system to air at room temperature. The resulting product was diluted with 5 mL of acetone and purified by adding it dropwise into a mixed solution of water and methanol (volume ratio of 1:2, 1.2 L). The (P(VDF-CTFE)-g-PtBMA) product was then washed with pure water until the filtrate was colorless and subsequently freeze-dried for further use.

In the second stage, the tert-butyl methacrylate on the backbone was hydrolyzed. Specifically, 2.5 g of P(VDF-CTFE)-g-PtBMA was added to a 250 mL round-bottomed flask containing 50 mL of toluene, followed by the addition of p-toluenesulfonic acid (TSA, 4.0 g). The mixture was heated to 85 °C for 8 h to hydrolyze the tert-butyl ester groups in the PtBMA units. The resulting P(VDF-CTFE)-g-PMAA product was washed by a mixed solution of water and ethanol (volume ratio of 1:2) and recovered by vacuum filtration. After drying under vacuum, P(VDF-CTFE)-g-PMAA was stored

for further use.

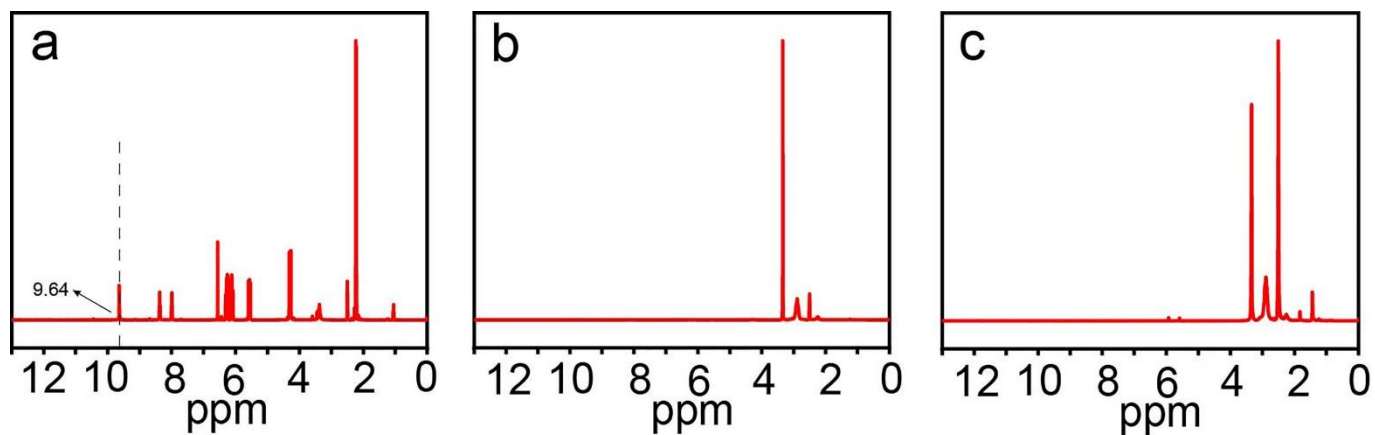


Figure S1. ^1H NMR spectra of (a) CD, (b) P (VDF-CTFE) and (c) P (VDF-CTFE)-g-PtBMA.

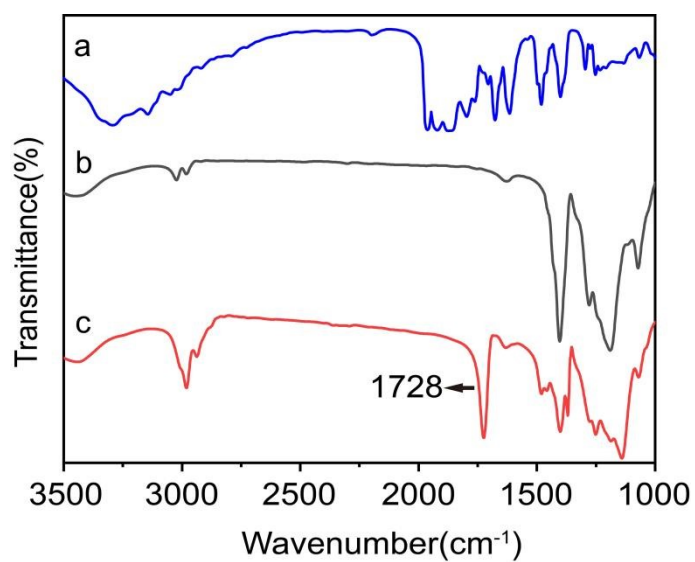


Figure S2. FTIR spectra of the (a) CD, (b) P (VDF-CTFE) and (c) P (VDF-CTFE)-g-PtBMA.

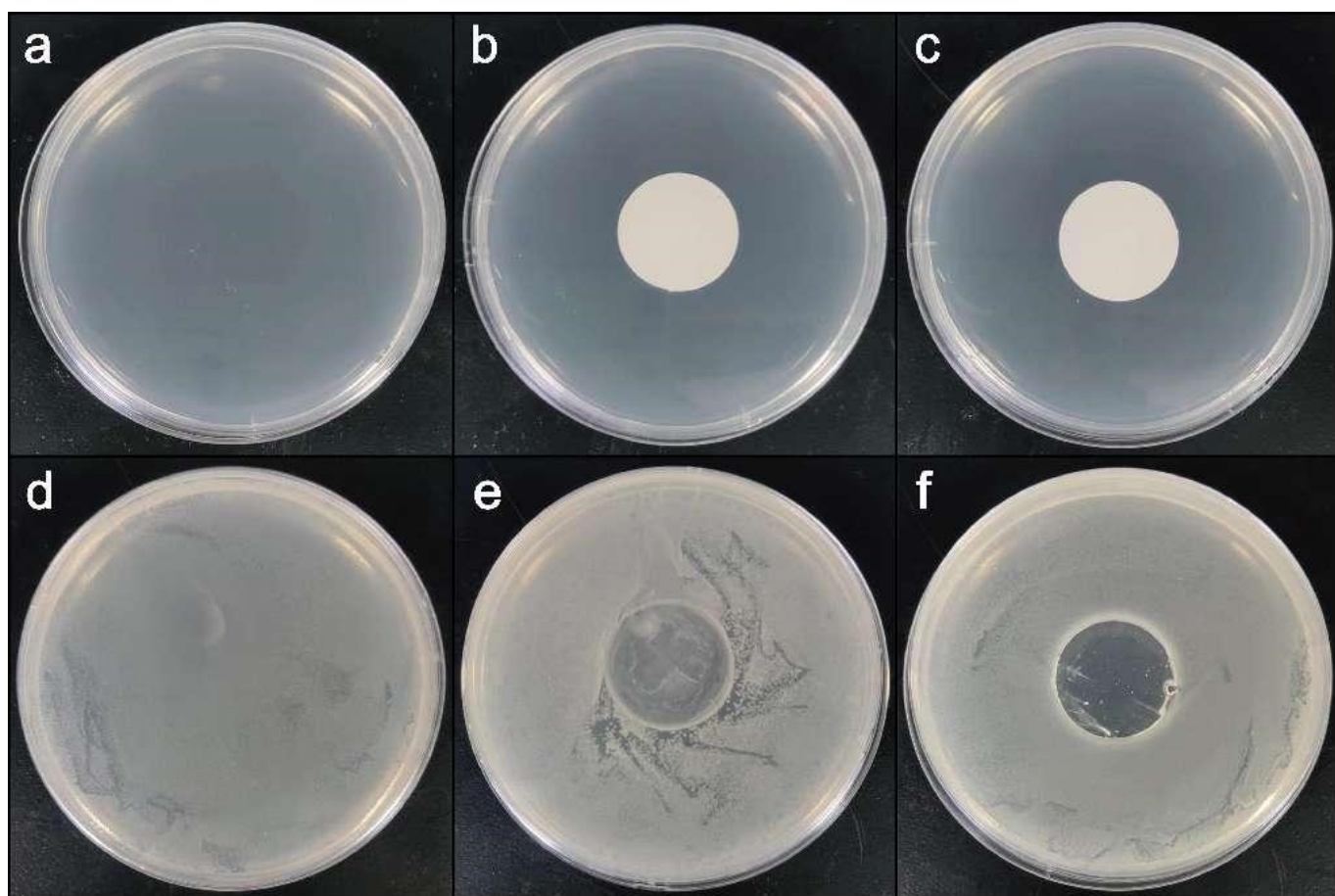


Figure S3. Zone of inhibition experiment without (b,e) and with (c,f) capsaicin derivatives CD.

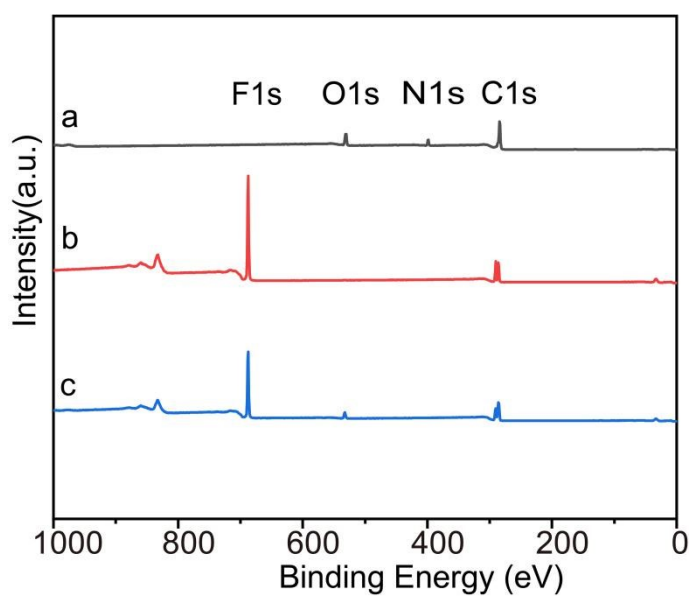


Figure S4. XPS of (a) CD, (b) P (VDF-CTFE) and (c) P (VDF-CTFE)-g-PtBMA.

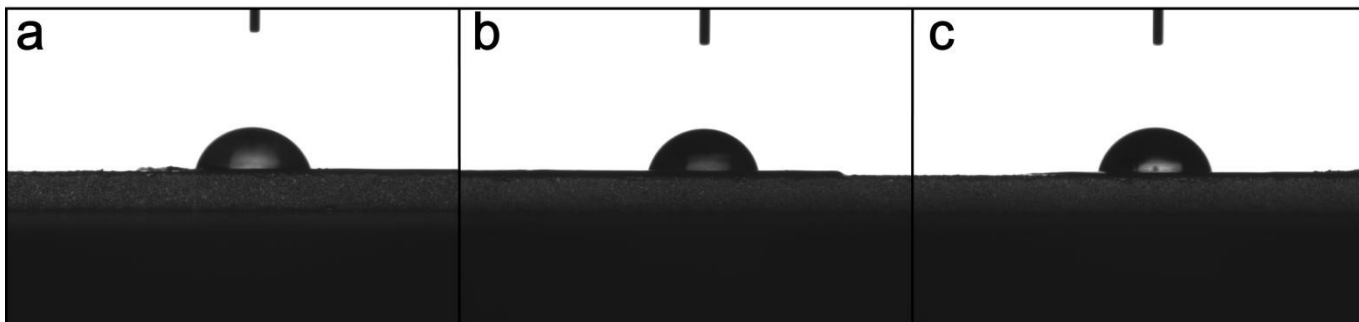


Figure S5. Water contact angle of the membranes. (a) M0, (b) M1, and (c) MA.

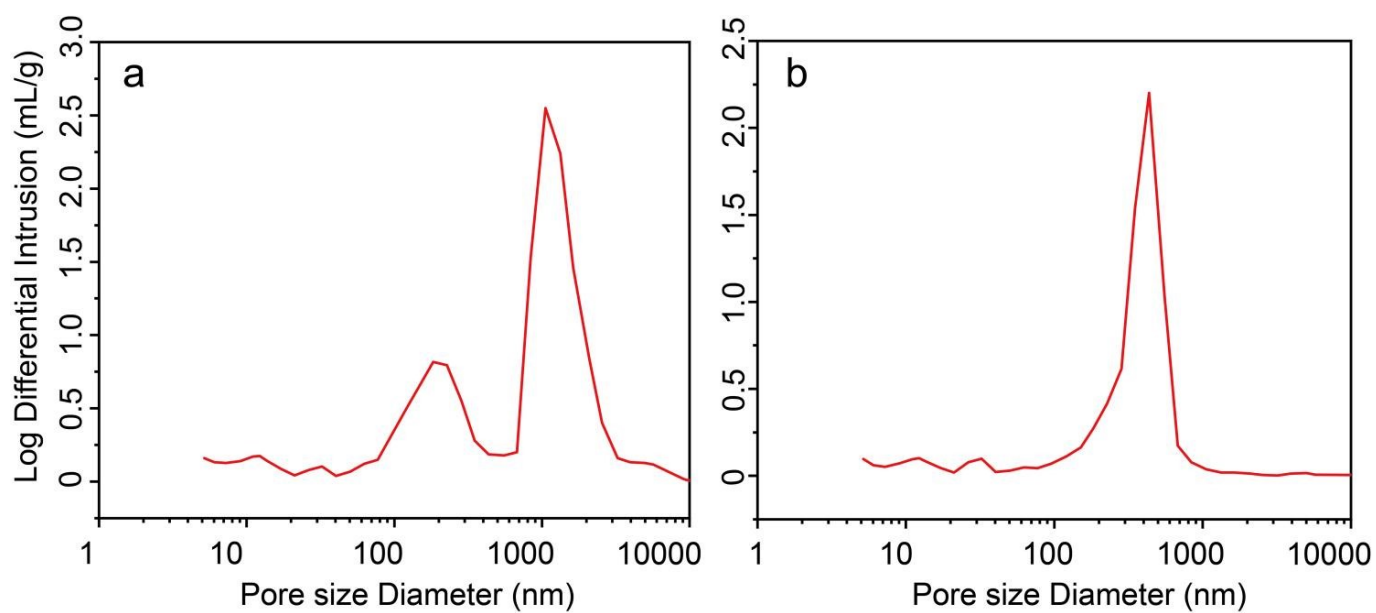


Figure S6. Pore size distribution of the M1 (a) and MA (b) membranes. (Note: determined by mercury intrusion porosimeter.)

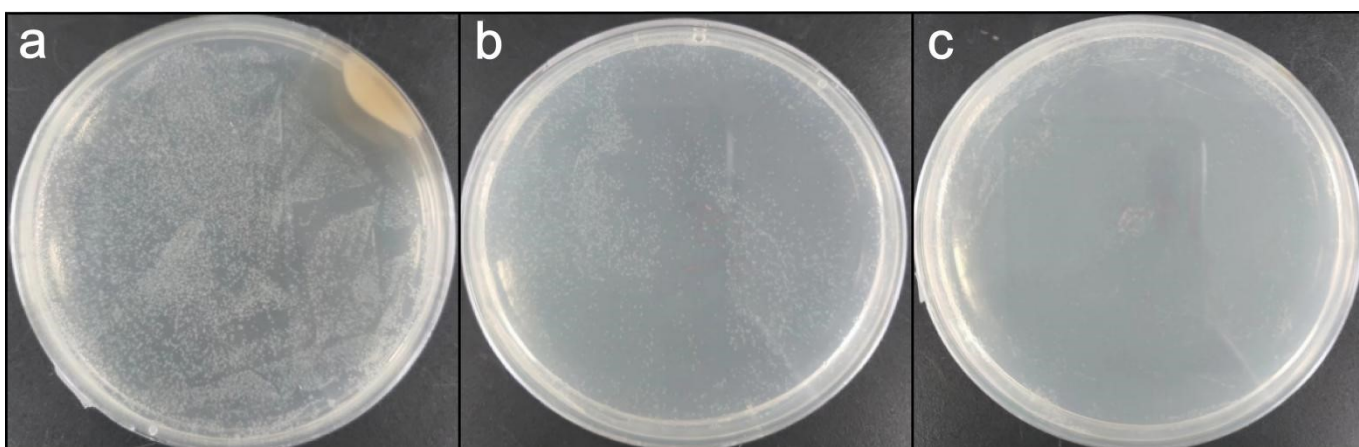


Figure S7. Growth of *E. coli* detached from M0 (a), M1 (b), and MA (c) on LB plates.

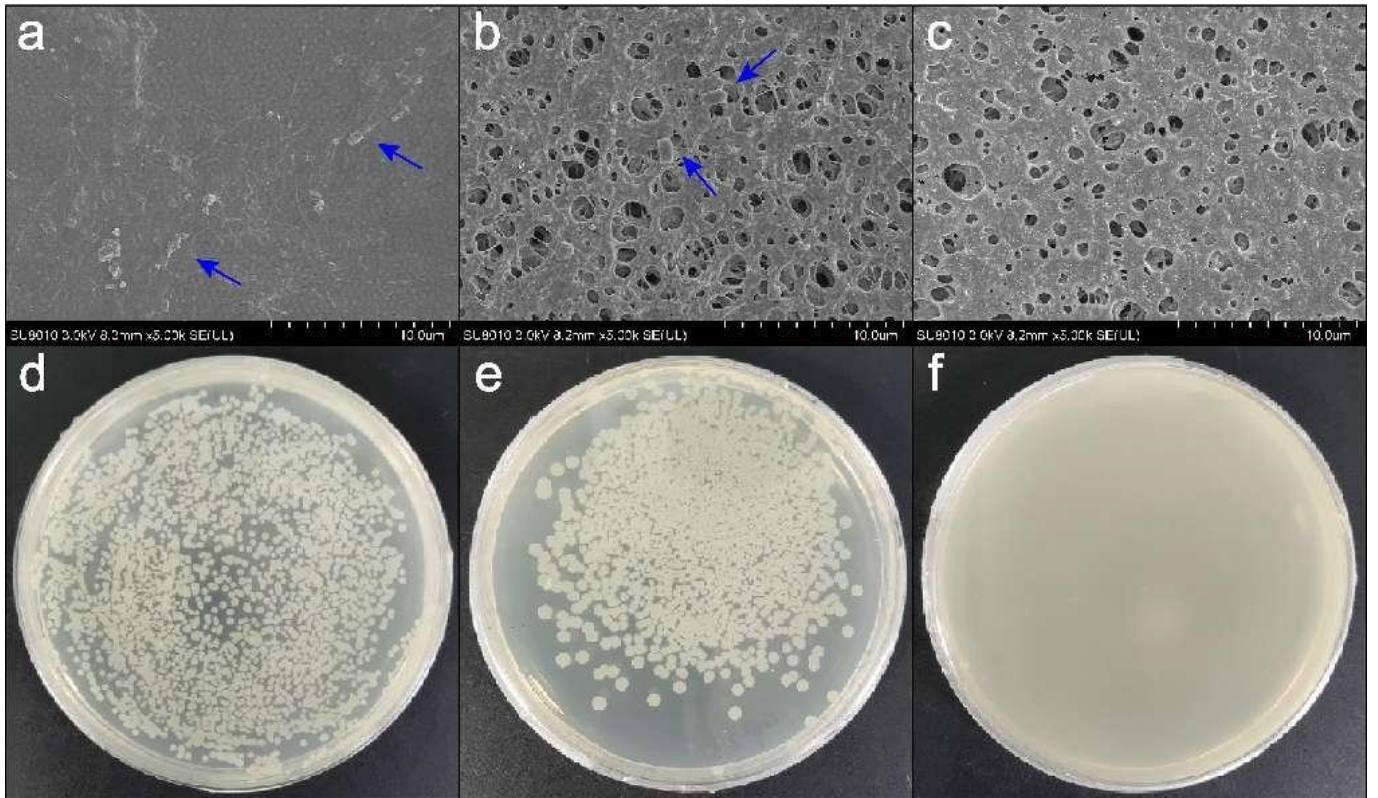


Figure S8. SEM image of M0 (a), M1 (b), and MA (c) fixed with glutaraldehyde after soaking in the *B. subtilis* suspension; the growth of *B. subtilis* detached from M0 (d), M1 (e), and MA (f) on LB plates.

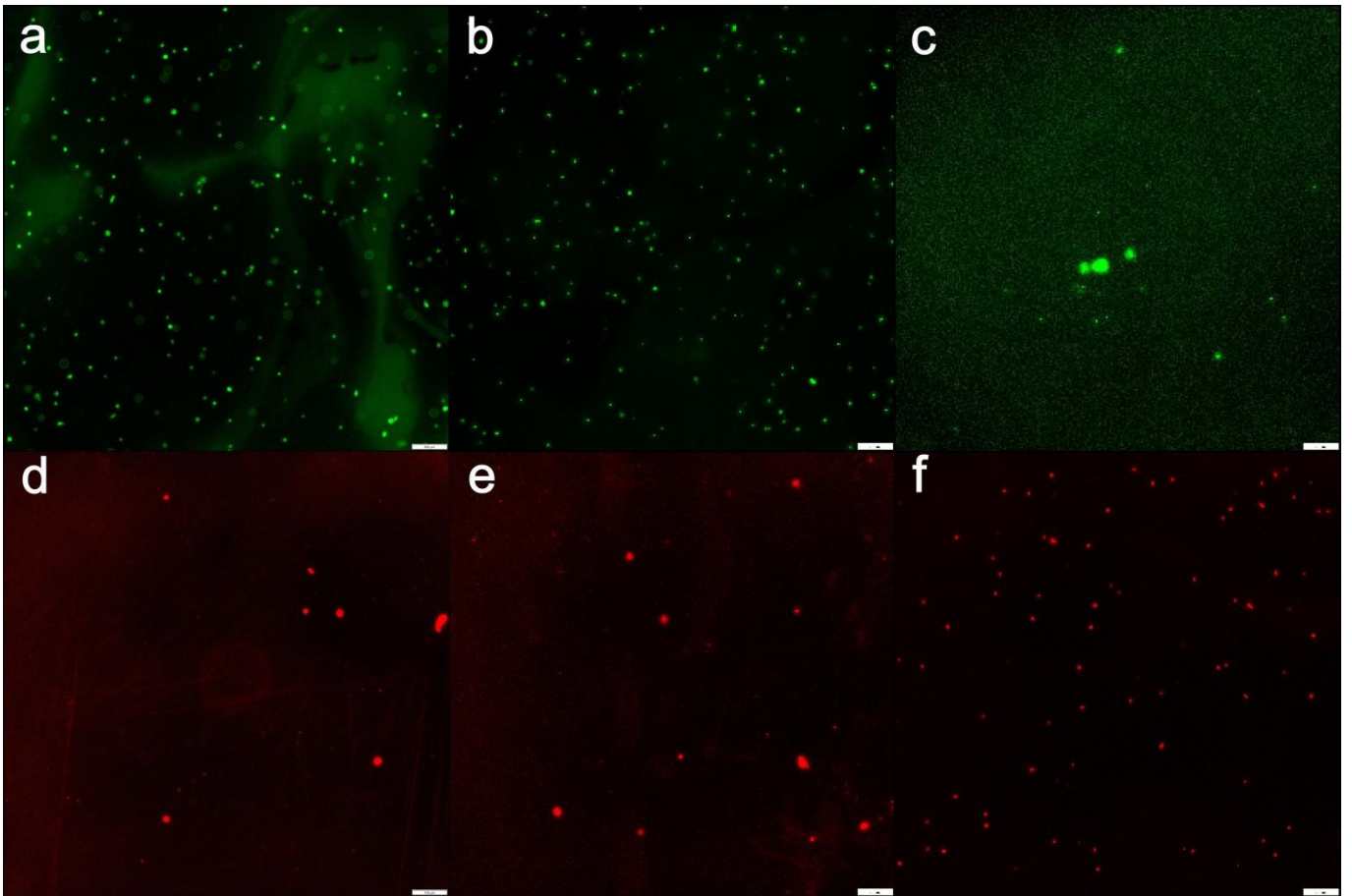


Figure S9. Fluorescence microscope images of *B. subtilis* on the M0 (a and d), M1(b and e) and MA (c and f) stained with SYTO 9 and PI, respectively. Note: the scale bar is 100 μm .

Table S1. Composition of elements (C, F, O, N) in the prepared materials.

| Materials | C% | F% | O% | N% |
|----------------------|-------|-------|-------|------|
| CD | 75.34 | 0 | 14.93 | 9.15 |
| P (VDF-CTFE) | 48.13 | 50.97 | 0 | 0 |
| P (VDF-CTFE)-g-PtBMA | 53.93 | 41.04 | 4.51 | 0 |
| P (VDF-CTFE)-g-PMAA | 49.71 | 47.99 | 1.77 | 0 |
| MA | 64.05 | 22.51 | 8.31 | 5.01 |

Note: Estimated from the XPS spectra.

Table S2. Relative flux decay (RFD) and relative flux recovery (RFR) of M1 and MA during filtration (transmembrane pressure: 0.01 MPa).

| Cycle number | | J0 | J1 | J2 | RFD | RFR |
|--------------|----|----------------------|----------------------|----------------------|-------|-------|
| | | (L/m ² h) | (L/m ² h) | (L/m ² h) | (%) | (%) |
| 1 | M1 | 41.38 | 21.04 | 28.66 | 49.15 | 69.26 |
| | MA | 62.07 | 43.62 | 57.52 | 29.72 | 92.67 |
| 2 | M1 | | 16.79 | 22.54 | 59.42 | 54.47 |
| | MA | | 39.56 | 56.24 | 36.27 | 90.61 |
| 3 | M1 | | 9.92 | 19.26 | 76.03 | 46.54 |
| | MA | | 36.62 | 52.73 | 41.00 | 84.95 |

Table S3. Comparison the performance of MA with the other antibacterial membranes.

| Membrane matrix | Antibacterial material | Modification reaction | Permeability ($Lm^{-2}h^{-1} / Bar$) | Relative Flux recovery (%) | Reference |
|----------------------------------------------------|------------------------|-----------------------|----------------------------------------|----------------------------|------------------|
| PES | Capsaicin derivative | UV-grafting | 120.0 | 92.3 | 1 |
| PSF | Capsaicin derivative | UV- grafting | 700.0 | 45.0 | 2 |
| CA | Quaternary ammonium | Etherification | 308.0 | 80.3 | 3 |
| PVDF | Ag^{+} | Blend | 671.0 | 61.6 | 4 |
| P(VDF-CTFE) grafted with capsaicin derivative (PD) | Capsaicin derivative | Esterification | 620.7 | 85.0 | This Work |

References:

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2. G. Xueli, W. Haizeng, W. Jian, H. Xing and G. Congjie, *Journal of Membrane Science*, 2013, **445**.
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