SUPPORTING INFORMATION

Dry and Wet Wrinkling of a Silk Fibroin Biopolymer by a Shape-Memory Material with Insight into Mechanical Effects on Secondary Structures in Silk Network

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TABLE OF CONTENTS

Index	Page
1. Supporting Methods	S4
Supporting Method I - SMP substrates for shape memory biaxial contraction.	S4
Supporting Method II - SMP substrates for shape memory bending actuated tension.	S4
Supporting Method III - SF Thin Film Application for SMP substrates undergoing shape	S5
memory biaxial contraction or shape memory bending actuated tension.	
2. Additional Supporting Figures	S 6
Effect of silk spin-coating speeds on wrinkle characteristics.	S 6
Figure S1 – Detailed characterization on the effects of silk spin-coating speeds on wrinkle	S6
characteristics at 70 °C.	
Figure S2 – Detailed characterization on the effects of silk spin-coating speeds on wrinkle	S 6
characteristics at 40 °C.	
Effect of varying methanol treatment time on secondary structures in silk network.	S7 - S22
Figure S3 – Background subtraction and deconvolution results of FTIR spectra within the	S7 - S9
Amide I region for SF-SMP with no methanol treatment (three independent replicates).	
Figure S4 - Background subtraction and deconvolution results of FTIR spectra within the	S10 - S12
Amide I region for SF-SMP with 30 min methanol treatment (three independent replicates).	
Figure S5 - Background subtraction and deconvolution results of FTIR spectra within the	S13 - S15
Amide I region for SF-SMP with 40 min methanol treatment (three independent replicates).	
Figure S6 - Background subtraction and deconvolution results of FTIR spectra within the	S16-S18
Amide I region for SF-SMP with 50 min methanol treatment (three independent replicates).	
Figure S7 - Background subtraction and deconvolution results of FTIR spectra within the	S19-S21
Amide I region for SF-SMP with 60 min methanol treatment (three independent replicates).	
Figure S8 - Detailed comparison of the secondary structure content with no methanol	S22
treatment and with varying methanol treatment times.	
Effect of shape memory uniaxial contraction on secondary structures in silk network	S23 - S40
Figure S9. Background subtraction and deconvolution of FTIR spectra within the Amide I	S23-S25
region following 40 °C heating for unstrained SF-SMP with no methanol treatment (three	
independent replicates).	
Figure S10. Background subtraction and deconvolution of FTIR spectra within the Amide I	S26-S28
region following 70 °C heating for unstrained SF-SMP with no methanol treatment (three	
independent replicates).	
Figure S11. Background subtraction and deconvolution of FTIR spectra within the Amide I	S29 - S31
region following 40 °C heating for strained SF-SMP with no methanol treatment (three	
independent replicates).	
Figure S12. Background subtraction and deconvolution of FTIR spectra within the Amide I	S32 - S34
region following 70 °C heating for strained SF-SMP with no methanol treatment (three	
independent replicates).	
Figure S13. Background subtraction and deconvolution of FTIR spectra within the Amide I	S35 - S37
region following 40 °C heating for strained SF-SMP with methanol treatment (three	
independent replicates).	~~~~
Figure S14. Background subtraction and deconvolution of FTIR spectra within the Amide I	S38 - S40
region following 70 °C heating for strained SF-SMP with methanol treatment (three	
independent replicates).	G 41 G 50
Effect of shape memory biaxial compression on secondary structures in silk network.	S41 - S58

Figure S15. Background subtraction and deconvolution of FTIR spectra within the Amide I	S41 - S43
region following 40 °C heating for strained SF-SMP with no methanol treatment (three	
independent replicates).	
Figure S16. Background subtraction and deconvolution of FTIR spectra within the Amide I	S44 - S46
region following 70 °C heating for strained SF-SMP with no methanol treatment (three	
independent replicates).	
Figure S17. Background subtraction and deconvolution of FTIR spectra within the Amide I	S47 - S49
region following 40 °C heating for strained SF-SMP with methanol treatment (three	
independent replicates).	
Figure S18. Background subtraction and deconvolution of FTIR spectra within the Amide I	S50 - S52
region following 70 °C heating for strained SF-SMP with methanol treatment (three	
independent replicates).	
Figure S19. Background subtraction and deconvolution of FTIR spectra within the Amide I	S53 - S55
region following 40 °C heating for unstrained SF-SMP with no methanol treatment (three	
independent replicates).	
Figure S20. Background subtraction and deconvolution of FTIR spectra within the Amide I	S56 - S58
region following 70 °C heating for unstrained SF-SMP with no methanol treatment (three	
independent replicates).	
Effect of shape memory bending actuated tension on secondary structures in silk	S59 - S83
network.	
Figure S21. Background subtraction and deconvolution of FTIR spectra within the Amide I	S59 - S61
region following 40 °C heating for strained SF-SMP with no methanol treatment (three	
independent replicates).	
Figure S22. Background subtraction and deconvolution of FTIR spectra within the Amide I	S62 - S64
region following 70 °C heating for strained SF-SMP with no methanol treatment (three	
independent replicates).	
Figure S23. Background subtraction and deconvolution of FTIR spectra within the Amide I	S65 - S67
region following 40 °C heating for strained SF-SMP with methanol treatment (three	
independent replicates).	
Figure S24. Background subtraction and deconvolution of FTIR spectra within the Amide I	S68 - S70
region following 70 °C heating for strained SF-SMP with methanol treatment (three	
independent replicates).	
Figure S25. Background subtraction and deconvolution of FTIR spectra within the Amide I	S71 - S73
region following 40 °C heating for unstrained SF-SMP with no methanol treatment (three	
independent replicates).	
Figure S26. Background subtraction and deconvolution of FTIR spectra within the Amide I	S74 - S76
region following 70 °C heating for unstrained SF-SMP with no methanol treatment (three	
independent replicates).	
Figure S27. Detailed comparison of the secondary structure content between strained and	S77
unstrained SF-SMP with exposure to methanol treatment following 40 °C and 70 °C	
heating.	
Figure S28. Background subtraction and deconvolution of FTIR spectra within the Amide I	S78 - S80
region following 40 °C heating for unstrained SF-SMP with methanol treatment (three	
independent replicates).	
Figure S29. Background subtraction and deconvolution of FTIR spectra within the Amide I	S81 - S83
region following 70 °C heating for unstrained SF-SMP with methanol treatment (three	
independent replicates).	

1. SUPPORTING METHODS

Supporting Method I - SMP substrates for shape memory biaxial contraction. To prepare SMP samples in which primarily compressive stresses are present in the substrate during shape recovery, we utilized a 4D-printing technique recently developed in our lab, known as programming via printing (PvP).⁵⁸ PvP involves using fused deposition modeling (FDM) to 3D-print an SMP substrate while simultaneously programming strains in the substrate at the fiber level during the printing process. This eliminates the need for a separate programming step following fabrication, and the SMP substrate can immediately undergo shape transformation after printing once exposed to an external stimulus (e.g., heat). The TPU pellets were dried for a minimum of 24 h in a vacuum oven at 50 °C prior to undergoing extrusion into filaments (Composer 450, 3Devo., The Netherlands). To print the SMP filament into desired substrates, STL files were converted to G-code using a slicing software (Ultimaker Cura., The Netherlands). Next, the SMP filament was loaded into a 3D printer (Ender 3 Pro., Creality) with a 0.4 mm nozzle diameter for depositing the semi-molten material onto the build plate in a layer-by-layer process. To ensure proper sample adhesion to the build plate, the plate was covered with Kapton tape and maintained at 25 °C. Rectangular samples (dimensions: 20 mm by 10 mm, 1.5 mm thick) were printed with fiber orientations of 0° and 90° alternating layer-by-layer for biaxial contraction. The samples were printed with a nozzle temperature of 200 °C, print speed of 30 mm/s, and print flow rate of 125%. Once printed, samples were stored in a desiccator until use.

Supporting Method II - *SMP substrates for shape memory bending actuated tension*. To prepare SMP samples in which primarily tensile stresses are present in the substrate during shape recovery, 3D printing was conducted but samples had to undergo conventional programming. The TPU filament was used to 3D print a "U-shaped" SMP substrate. To print the filament into the desired SMP substrate, the "U-shape" design was created in computer aided design software and STL files were converted to g-code using Ultimaker Cura. Next, the TPU filament was loaded into the Ender 3 Pro with 0.4 mm nozzle diameter. To optimize bed adhesion, the build plate was covered with Kapton tape and maintained at 50 °C. The U-shaped SMP substrates were printed with a nozzle temperature of 220 °C, print speed of 30 mm/s, and print

flow rate of 125%. To set the secondary shape of the SMP, the ends of the substrate were clamped, then the U-shaped SMP was preheated in an isothermal oven at 50 °C for 5 min, then tension was applied to the clamps to straighten the substrate, and then cooled in a freezer at -20 °C to fix the straightened, temporary shape. Once fixed, samples were removed from the clamps and stored in a desiccator until use.

Supporting Method III - *SF Thin Film Application for SMP substrates undergoing shape memory biaxial contraction or shape memory bending actuated tension.* Since one of the major limitations of spin coating is that it cannot create films on architecturally complex and/or curved surfaces, dip coating was used for application of a SF film to the surface of these SMP substrates. The stock concentration of SF was adjusted to 3% w/v by adding ultrapure water. To coat the SMP substrates with SF, samples were dip coated in a 20 ml scintillation vial for 2 min at room temperature ensuring that we prevent pre-recovery of the SMP. Then, by adapting a previously published fabrication procedure,⁵⁹ the silk coated SMP was immediately post-treated in a solution of 70% methanol for 5 sec. The treated samples were stored in a desiccator to dry overnight.

2. ADDITIONAL SUPPORTING FIGURES



FIGURE S1. Detailed characterization of the effects of SF spin-coating speeds on wrinkle (A) wavelength and (B) amplitude at 70 °C. (n=3, Student's t-test for two group comparisons, *p< 0.05, **p< 0.01, ***p<0.001, ***p<0.001).



FIGURE S2. Detailed characterization of the effects of SF spin-coating speeds on wrinkle (A) wavelength and (B) amplitude at 40 °C. (n=3, Student's t-test for two group comparisons, *p < 0.05, **p < 0.01, ***p < 0.001, ***p < 0.001).

No Methanol Trial 1



FTIR Deconvolution



No Methanol Trial 2



No Methanol Trial 3



FIGURE S3. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹). (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + SMP) with no methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with no methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.



S10





FIGURE S4. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹). (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + SMP) with 30 min methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with 30 min methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.







FIGURE S5. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹). (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + SMP) with 40 min methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with 40 min methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.







FIGURE S6. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹). (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + SMP) with 50 min methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with 50 min methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.







FIGURE S7. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹). (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + SMP) with 60 min methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with 60 min methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.



FIGURE S8. Methanol treatment provides secondary structural changes in the silk network and induces β -sheet formation. Detailed comparison of the secondary structure content with no methanol treatment and with varying methanol treatment times of 30 min, 40 min, 50 min, and 60 min.

Unstrained SF-SMP, No MeOH, 40 °C Trial



Unstrained SF-SMP, No MeOH, 40 °C Trial



Unstrained SF-SMP, No MeOH, 40 °C Trial

FTIR Background Subtraction



FIGURE S9. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 40 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + unstrained SMP) with no methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with no methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.



Unstrained SF-SMP, No MeOH, 70 °C Trial

Unstrained SF-SMP, No MeOH, 70 °C Trial



Unstrained SF-SMP, No MeOH, 70 °C Trial



FIGURE S10. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 70 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + unstrained SMP) with no methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with no methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.

Strained SF-SMP, No MeOH, 40 °C Trial



Strained SF-SMP, No MeOH, 40 °C Trial



Strained SF-SMP, No MeOH, 40 °C Trial



FIGURE S11. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 40 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + strained SMP) with no methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with no methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.

Strained SF-SMP, No MeOH, 70 °C Trial 1



Strained SF-SMP, No MeOH, 70 °C Trial



Strained SF-SMP, No MeOH, 70 °C Trial



FIGURE S12. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 70 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + strained SMP) with no methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with no methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.

Strained SF-SMP, With MeOH, 40 °C Trial



Strained SF-SMP, With MeOH, 40 °C Trial




FIGURE S13. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 40 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + strained SMP) with methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.



S38





FIGURE S14. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 70 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + strained SMP) with methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.









FIGURE S15. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 40 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + strained SMP) with no methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with no methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.







FIGURE S16. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 70 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + strained SMP) with no methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with no methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.







FIGURE S17. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 40 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + strained SMP) with methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.







FIGURE S18. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 70 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + strained SMP) with methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.











FIGURE S19. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 40 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + unstrained SMP) with no methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with no methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.







FIGURE S20. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 70 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + unstrained SMP) with no methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with no methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.







FIGURE S21. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 40 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + strained SMP) with no methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with no methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.





S63



FIGURE S22. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 70 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + strained SMP) with no methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with no methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.







S66





FIGURE S23. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 40 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + strained SMP) with methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.





S68





FIGURE S24. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 70 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + strained SMP) with methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.






Unstrained SF-SMP, No MeOH, 40 °C Trial



FIGURE S25. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 40 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + unstrained SMP) with no methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with no methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.

Unstrained SF-SMP, No MeOH, 70 °C Trial



FTIR Background Subtraction



Unstrained SF-SMP, No MeOH, 70 °C Trial

Unstrained SF-SMP, No MeOH, 70 °C Trial



FIGURE S26. Deconvolution of FTIR spectra within the Amide I region $(1600-1700 \text{ cm}^{-1})$ following 70 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + unstrained SMP) with no methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with no methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.



FIGURE S27. No significant differences were observed for secondary structural changes in the silk network between the strained and unstrained SF-SMP with exposure to methanol treatment following 40 $^{\circ}$ C and 70 $^{\circ}$ C heating.

Unstrained SF-SMP, With MeOH, 40 °C Trial



FTIR Background Subtraction

Unstrained SF-SMP, With MeOH, 40 °C Trial



Unstrained SF-SMP, With MeOH, 40 °C Trial 3



FIGURE S28. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 40 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + unstrained SMP) with methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.









Unstrained SF-SMP, With MeOH, 70 °C Trial



FIGURE S29. Deconvolution of FTIR spectra within the Amide I region (1600-1700 cm⁻¹) following 70 °C heating. (Top panels) Multiple linear regression is used to subtract the contribution of the SMP substrate (blue) from the total signal (SF + unstrained SMP) with methanol treatment (orange) through decomposition. Three independent replicates were measured for each condition (n=3). (Bottom panels) Deconvolution results of FTIR spectra for SF-SMP with methanol treatment. In the middle panel, raw data (red) and reproduced spectra (blue) are depicted, with the blue line representing the summation of the deconvoluted peaks. Residual fit errors are depicted in the top panels. The bottom panel shows the results from the deconvolution process; secondary structures are assigned based on peak locations within this panel. The normalized peak areas were used to determine the secondary structures found in the film.