

Supporting Information for

A Dual-Color ESIPT-based Probe for Simultaneous Detection of Hydrogen Sulfide and Hydrazine

Qian Gong, Youbo Lai, Weiyang Lin,*

Institute of Optical Materials and Chemical Biology, Guangxi Key Laboratory of
Electrochemical Energy Materials, School of Chemistry and Chemical Engineering,
Guangxi University, Nanning, Guangxi 530004, P. R. China.

E-mail: weiyanglin2013@163.com (W. Lin)

Table of contents

1. Materials and Instrument	3
2. Test solution configuration of the BDM-DNP	3
3. Cytotoxicity assay	4
4. Cells fluorescence imaging	4
5. Supporting figures and tables	5
Fig. S1	5
Fig. S2	6
Fig. S3	6
Fig. S4	7
Fig. S5	7
Fig. S6	8
Fig. S7	8
Fig. S8	9
Fig. S9	9
Fig. S10	10
Fig. S11	10
Fig. S12	11
Fig. S13	11
Fig. S14	12
Fig. S15	12

1. Materials and instrument

Common reagents or materials were obtained from commercial suppliers without further purification except as otherwise noted. All experiments used ultra-pure water. The pH measurements were performed with a PHS-3E pH meter. UV-vis absorption spectra were obtained on a Shimadzu UV-2700 spectrophotometer, and fluorescence spectra were measured on a HITACHI F4700 fluorescence spectrophotometer. The fluorescence imaging of cells was performed with a Leica TCS SP8 CARS confocal microscope. ^1H and ^{13}C NMR spectra were measured on an AVANCE III HD600 digital NMR spectrometer, using tetramethylsilane (TMS) as the internal reference. High-resolution mass spectrometric (HRMS) analyses were measured on Brooke solanX 70 FT-MS, Agilent 6540T.

2. Test solution configuration of the BDM-DNP

Except for special circumstances, all probe solutions, ion solutions, and related solutions required in this article were prepared and used by the following methods. (a) Preparation of **BDM-DNP** reserve solution: 5.9 mg of **BDM-DNP** solid dissolved in 1 mL DMSO solution to prepare reserve solution; (b) Manufacture of **BDM-DNP** test solution: 1.2 μL of the stock solution diluted with DMSO to 1.2 mL, then the diluted solution diluted with PBS buffer to 3 mL to prepare the final test solution required for testing. The final probe was tested at the concentration of 10 μM , and the solution was 60% DMSO and 40% PBS buffer. The spectral properties of the solutions were measured using a Shimadzu UV-2700 spectrophotometer and a HITACHI F4700 fluorescence spectrophotometer.

3. Cytotoxicity assay

In vitro, cytotoxicity was measured using the colorimetric methyl thiazolyl tetrazolium (MTT) assay on HeLa cells. Cells were seeded into the 96-well tissue

culture plate in the presence of 100 μ L Dulbecco's Modified Eagle Medium (DMEM) supplemented with 10% fetal bovine serum (FBS) and 1% penicillin/streptomycin at 37 °C and 5% CO₂ atmosphere for 24h. The cell DMEM medium from each well was replaced with fresh medium containing increasing concentrations of **BDM-DNP**, i.e., 0, 1, 2, 5, 10, 20, 30, and 40 μ M. Then the cell culture medium was removed and rinsed twice with PBS. Subsequently, 90 μ L of fresh DMEM medium and 10 μ L of MTT (5 mg/mL) were added to each well and incubated for 4 h. Absorbance of the solution was measured using a microplate reader. The cell viability was determined by assuming 100% cell viability for cells without **BDM-DNP**.

4. Cells fluorescence imaging

HeLa cells were cultured in Dulbecco's Modified Eagle Medium (DMEM) supplemented with 10% fetal bovine serum (FBS) and 1% streptomycin sulfate and penicillin, incubated at 37°C in a 5% CO₂ environment. For the fluorescence imaging study of exogenous H₂S and N₂H₄ in living HeLa cells, we conducted a control experiment, in which the HeLa cells were incubated with 10 μ M **BDM-DNP** in the culture medium.

To investigate exogenous H₂S and N₂H₄ in live cells, three HeLa cell lines were treated with NaHS and N₂H₄ for imaging studies, respectively, then cultured with **BDM-DNP** (10 μ M) for 15 minutes. Firstly, 10 mM stock solutions of **BDM-DNP** in DMSO and 100 mM hydrogen sulfide and hydrazine in water were prepared. The first HeLa cell lines were only cultured with **BDM-DNP** (10 μ M) for 15 minutes. The second cell lines were pretreated with H₂S and N₂H₄ (100 μ M) for 15 minutes and then incubated with **BDM-DNP** (10 μ M) for 15 minutes. The third cell lines were pretreated with H₂S and N₂H₄ (100 μ M) for 30 minutes before being incubated with **BDM-DNP** (10 μ M) for 15 minutes.

5. Supporting figures and tables

$^1\text{H NMR}$ (600 MHz, $\text{DMSO-}d_6$) δ 11.14 (s, 1H), 10.28 (s, 1H), 7.97 (d, $J=2.3$ Hz, 1H), 7.88 (dd, $J=8.7, 2.3$ Hz, 1H), 7.30 (d, $J=4.7$ Hz, 2H), 7.04 (d, $J=8.6$ Hz, 1H), 6.86 (s, 1H), 2.60 (s, 2H), 2.53 (s, 2H), 1.01 (s, 6H).

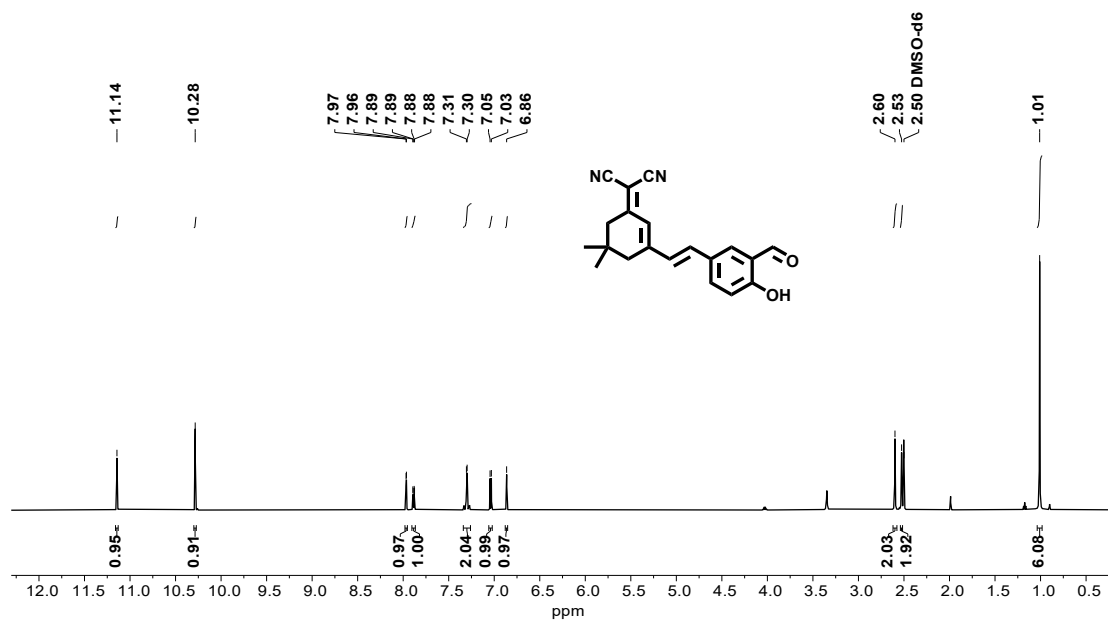


Fig. S1. $^1\text{H NMR}$ (600 MHz, $\text{DMSO-}d_6$) spectrum of compound 2.

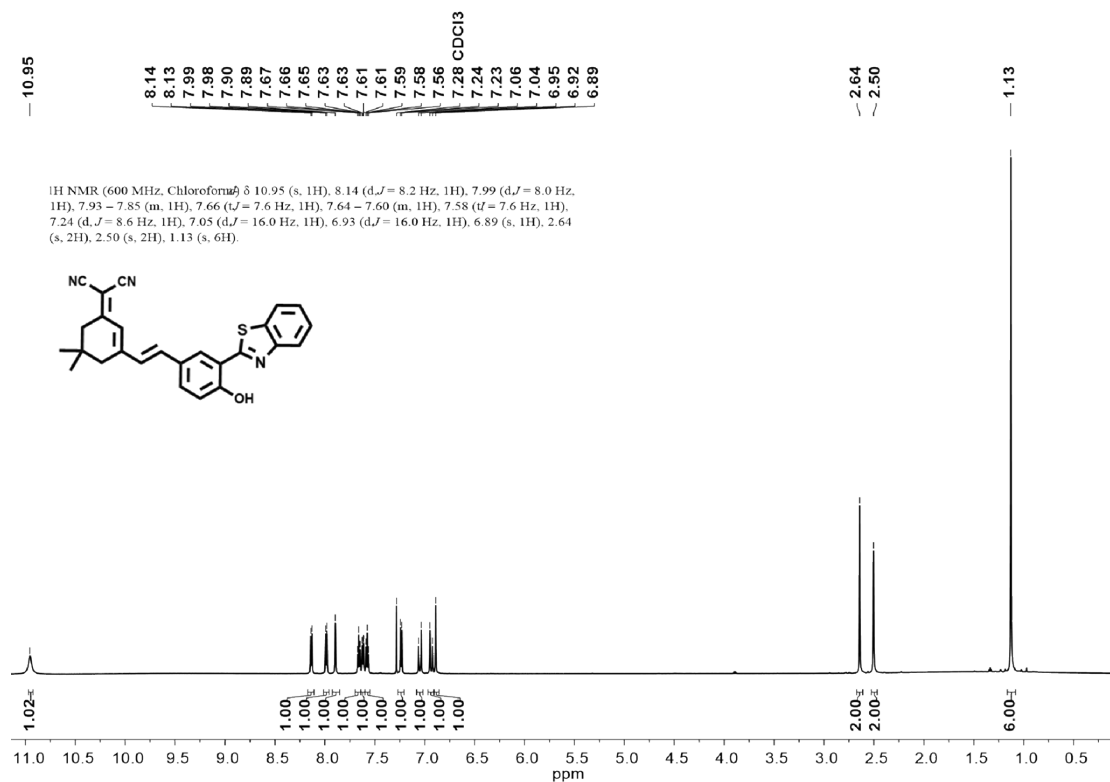


Fig. S2. ¹H NMR (600 MHz, Chloroform-*d*) spectrum of **BDM-OH**.

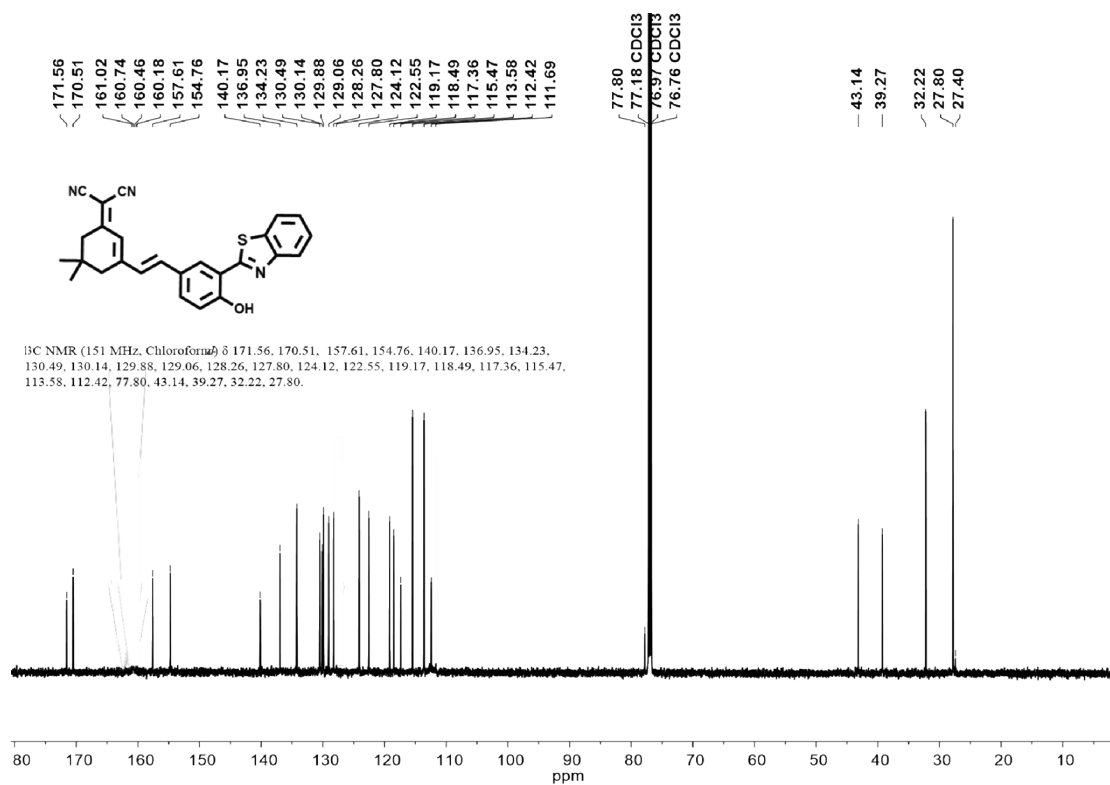


Fig. S3. ¹³C NMR (151 MHz, Chloroform-*d*) spectrum of **BDM-OH**.

^1H NMR (600 MHz, Chloroform-*d*) δ 8.93 (d, J = 2.6 Hz, 1H), 8.67 (s, 1H), 8.30 (dd, J = 9.1, 2.7 Hz, 1H), 8.07 (d, J = 8.2 Hz, 1H), 7.87 (d, J = 7.5 Hz, 1H), 7.71 (d, J = 10.5 Hz, 1H), 7.51 (t, J = 7.9 Hz, 1H), 7.41 (t, J = 7.6 Hz, 1H), 7.22 (d, J = 8.4 Hz, 1H), 7.12 (d, J = 12.8 Hz, 2H), 7.04 (d, J = 9.2 Hz, 1H), 6.91 (s, 1H), 2.62 (s, 2H), 2.49 (s, 2H), 1.11 (s, 6H).

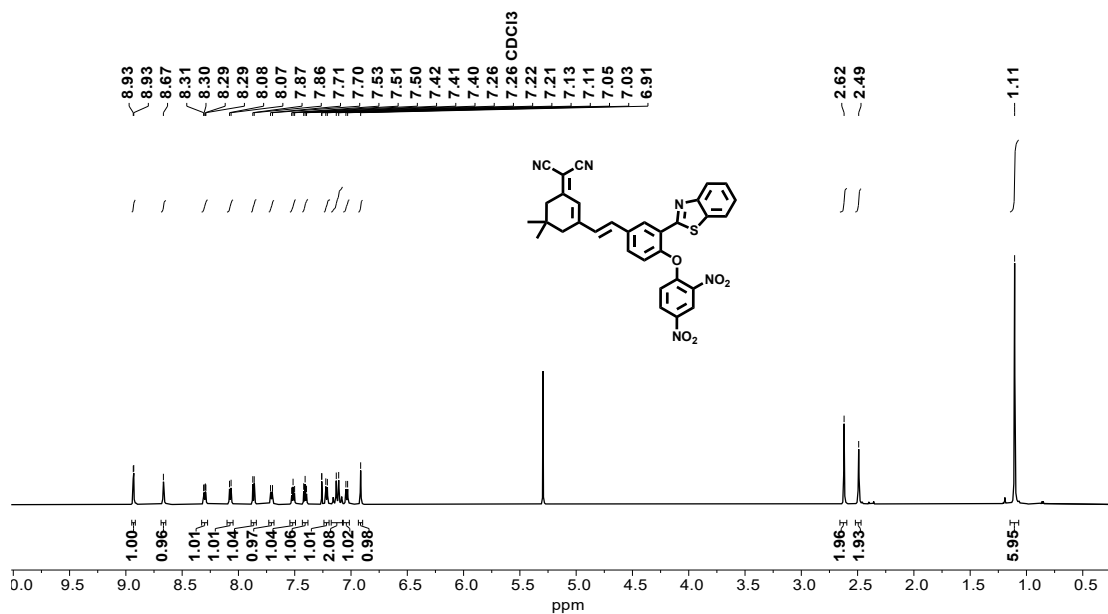


Fig. S4. ^1H NMR (600 MHz, Chloroform-*d*) spectrum of **BDM-DNP**.

^{13}C NMR (151 MHz, Chloroform-*d*) δ 160.14, 154.88, 152.85, 152.38, 151.35, 142.31, 134.92, 134.19, 131.15, 130.42, 130.21, 129.14, 126.83, 125.99, 122.14, 121.60, 118.59, 113.23, 112.54, 32.08.

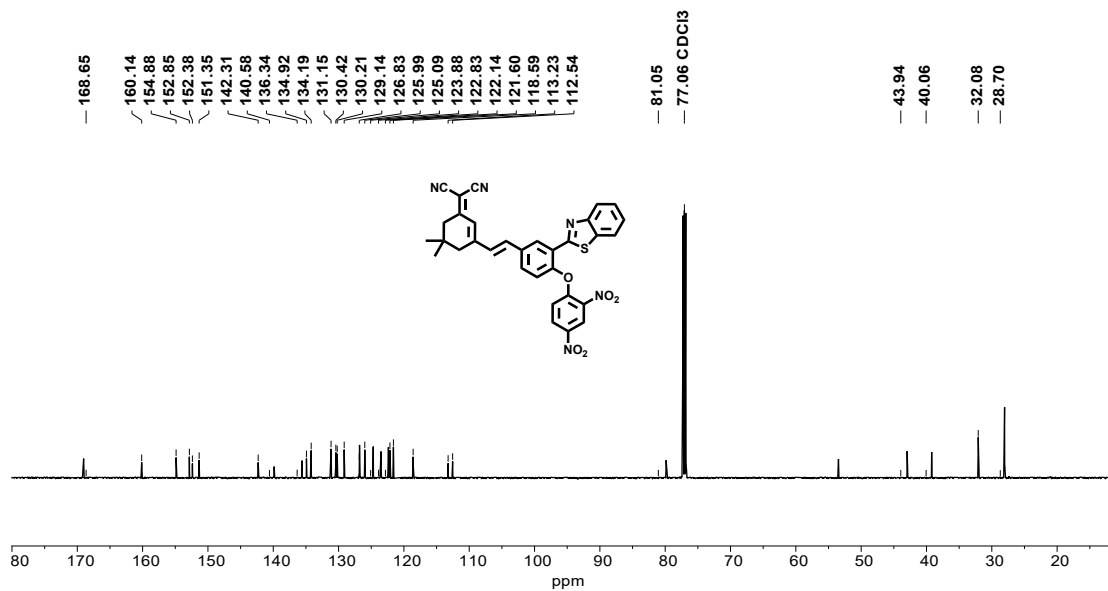


Fig. S5. ^{13}C NMR (151 MHz, Chloroform-*d*) spectrum of **BDM-DNP**.

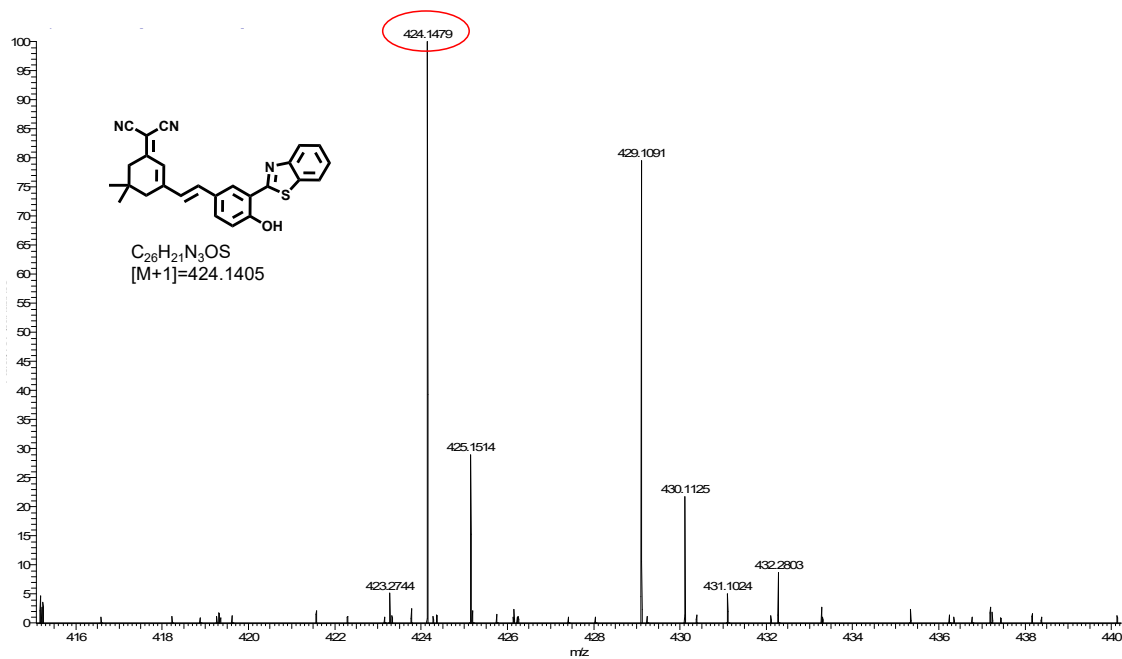


Fig. S6. HRMS (ESI) spectrum of **BDM-OH**

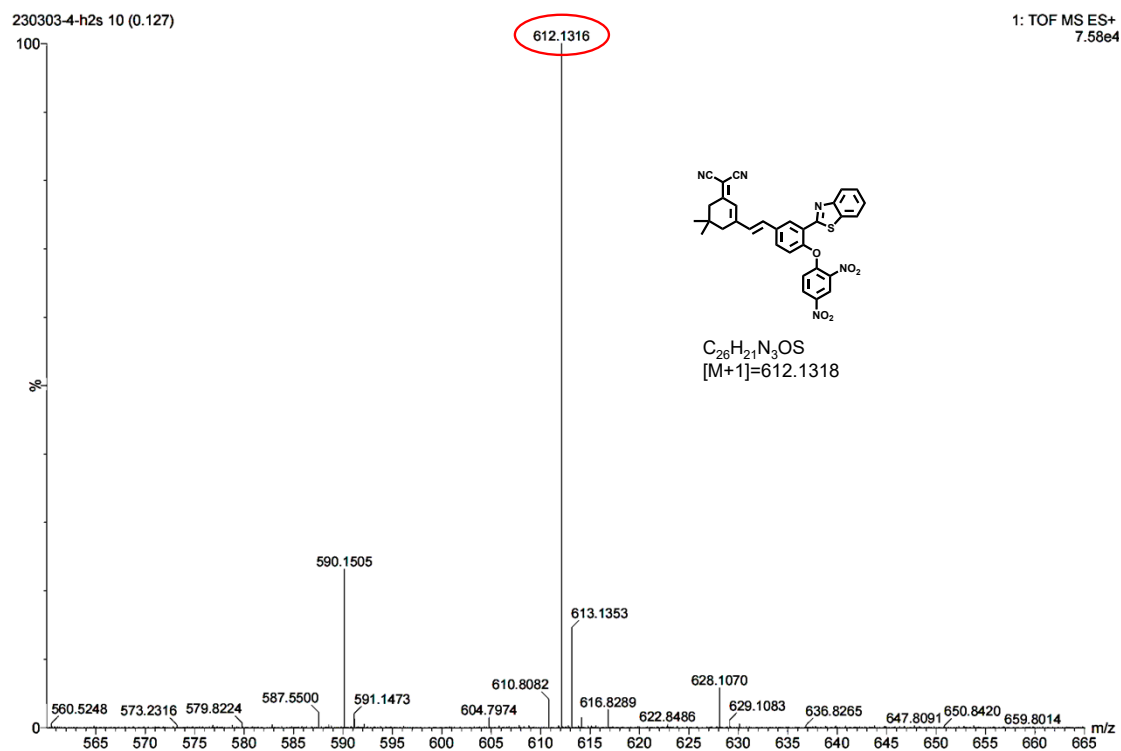


Fig. S7. HRMS (ESI) spectrum of **BDM-DNP**.

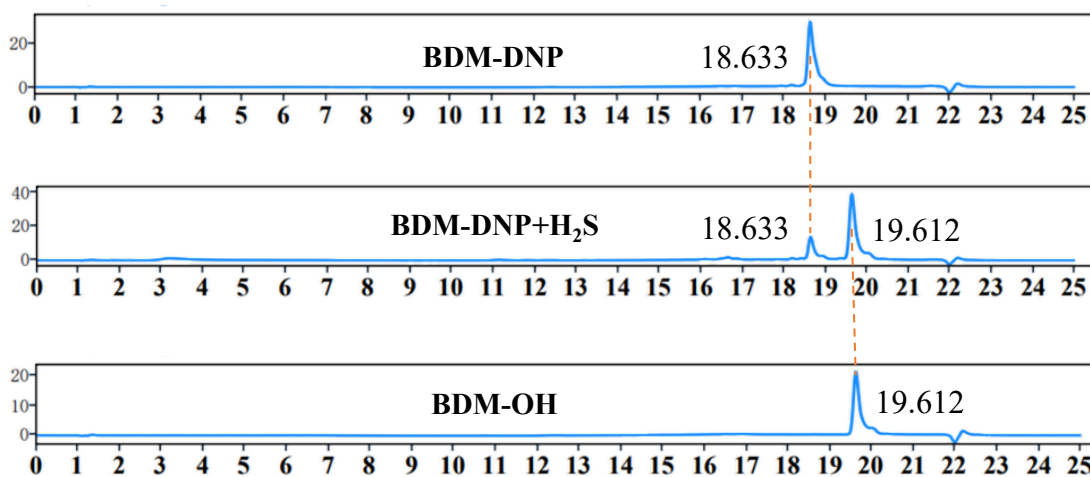


Fig. S8. The high-performance liquid chromatography (HPLC) of **BDM-DNP** and **BDM-OH** were tested in 10 mM PBS buffer solution (methyl alcohol: water = 1:9, pH = 7.4). (A) **BDM-DNP**; (B) **BDM-DNP** + H₂S; (C) **BDM-OH**.

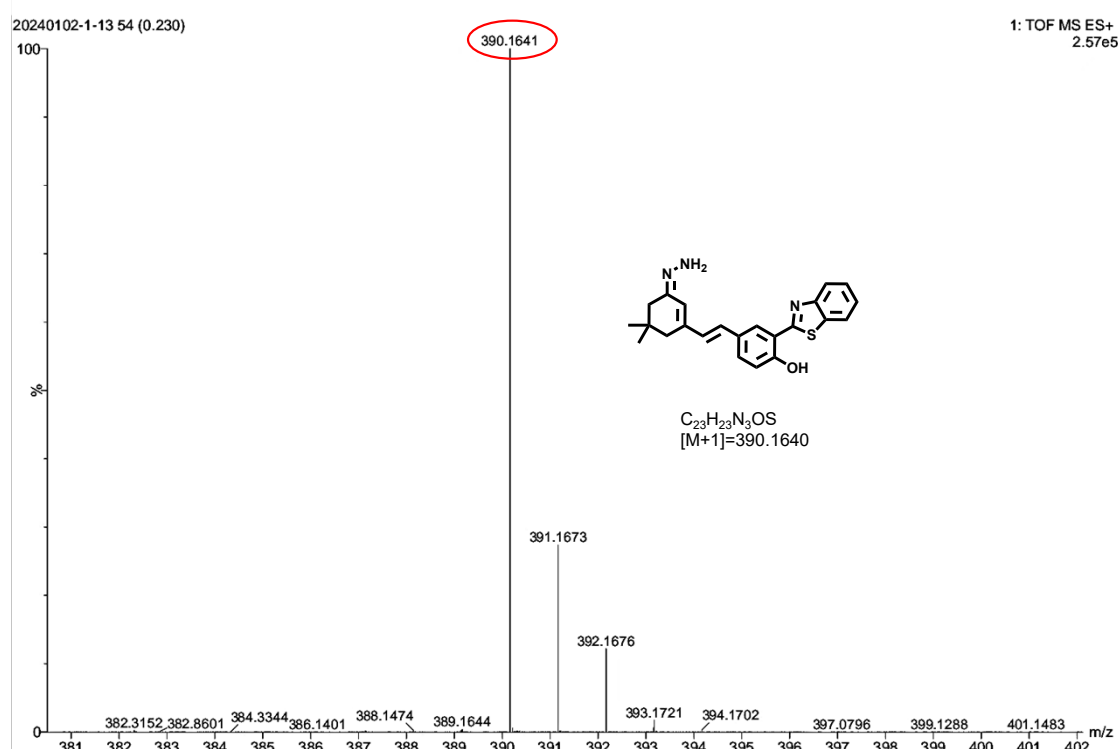


Fig. S9. HRMS (ESI) spectra of the reaction of **BDM-DNP** with N₂H₄.

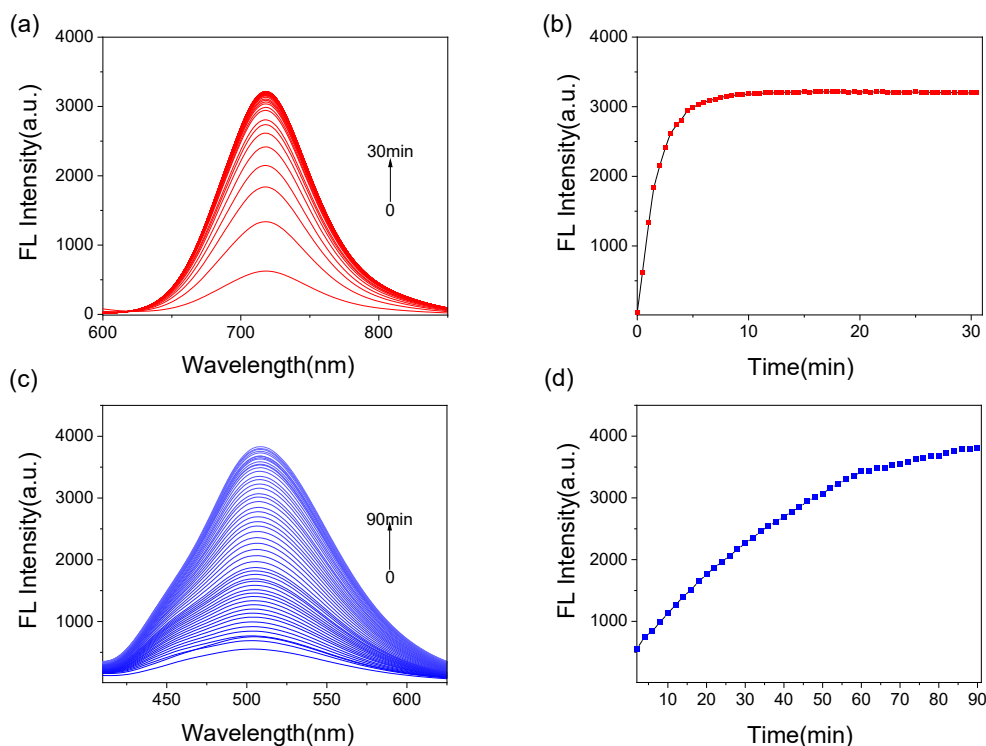


Fig. S10. (a) Fluorescence spectra of **BDM-DNP** (10 μM) with 1 mM H₂S at different times; (b) The response time of **BDM-DNP** with H₂S at 713 nm; (c) Fluorescence spectra of **BDM-DNP** (10 μM) with 1 mM N₂H₄ at different time; (d) The response time of **BDM-DNP** with N₂H₄ at 713 nm.

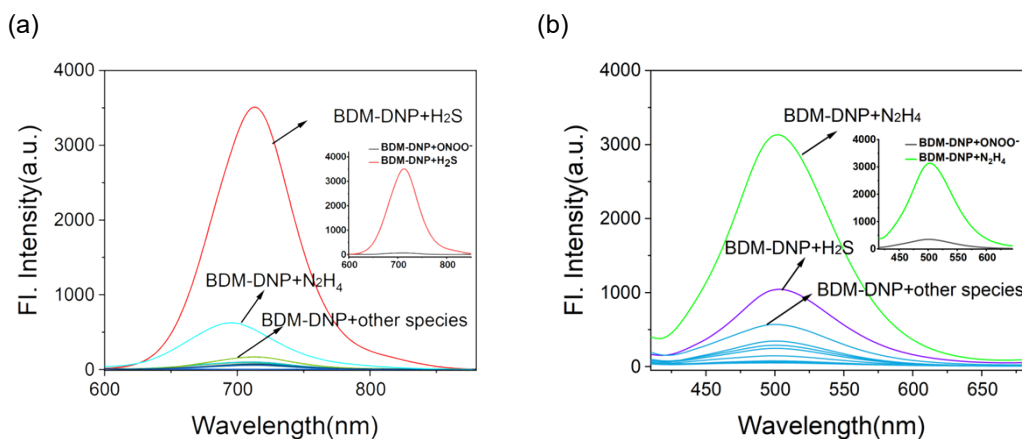


Fig. S11. The selectivity of **BDM-DNP** (10 μM) towards diverse relevant analytes (1 mM) against H₂S (1 mM) (a) and N₂H₄ (1 mM) (b) in the presence of coexisting analytes (1 mM). Insert: 1.H₂S; 2. N₂H₄ 3. Cys; 4. Hcy; 5. GSH; 6. Na₂S; 7. NaS₂O₃; 8. NaSCN; 9. NaCl; 10. NaH₂PO₄; 11. NaHCO₃; 12. NaNO₃; 13.CH₃COONa; 14. H₂O₂; 15. Gly; 16. Asp; 17.ONOO⁻.

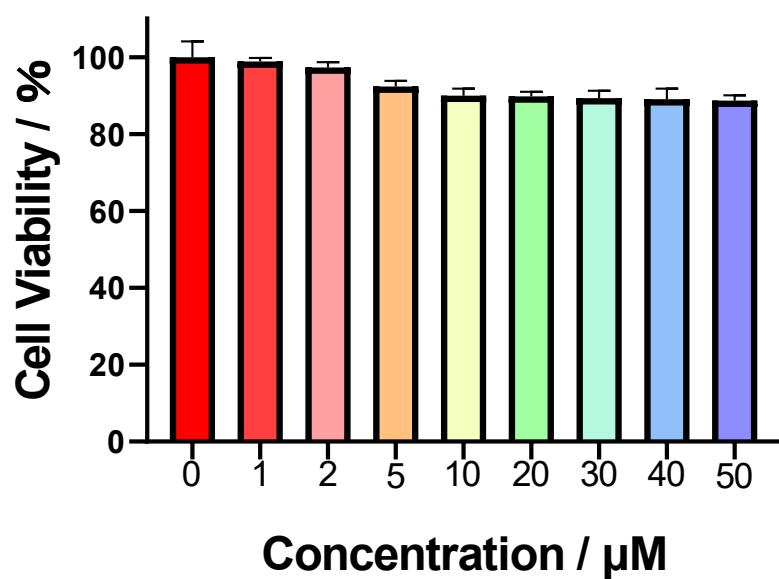


Fig. S12. The cytotoxicity of **BDM-DNP** in HeLa cells.

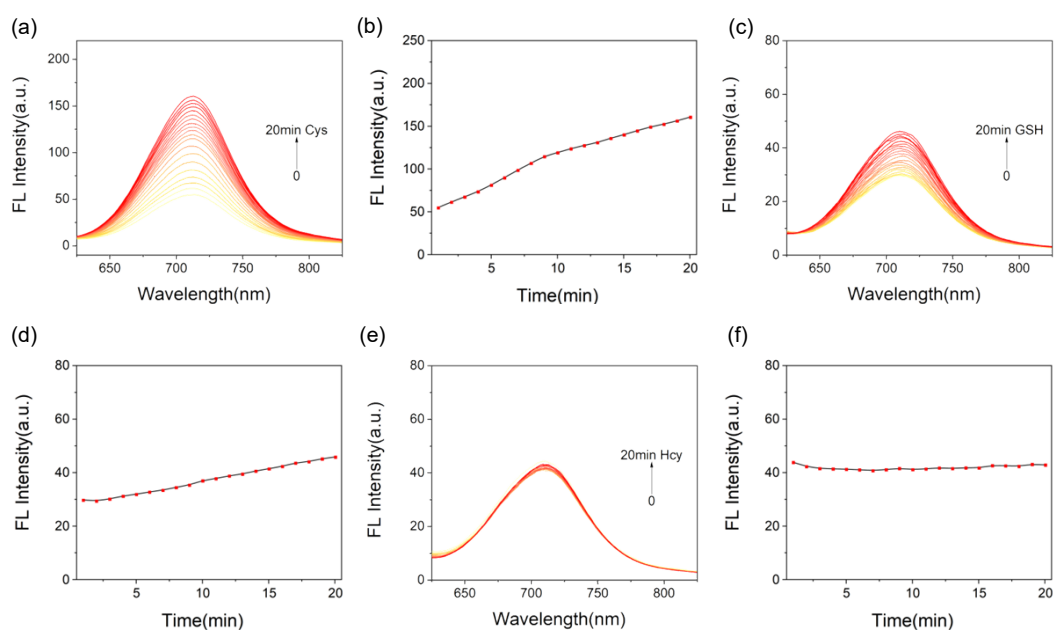


Fig. S13. Fluorescence spectra of **BDM-DNP** ($10 \mu\text{M}$) with $500 \mu\text{M}$ Cys (a), $500 \mu\text{M}$ GSH (c), and $500 \mu\text{M}$ Hcy (e) at different times; The response time of **BDM-DNP** with Cys (b), GSH (d) and Hcy (f) at 713 nm .

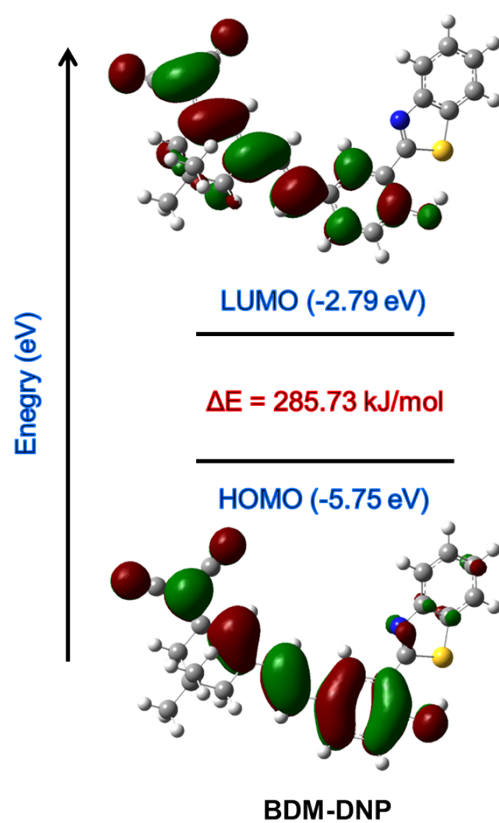


Fig. S14. Theoretical calculation of HOMO/LUMO energy for **BDM-DNP**.

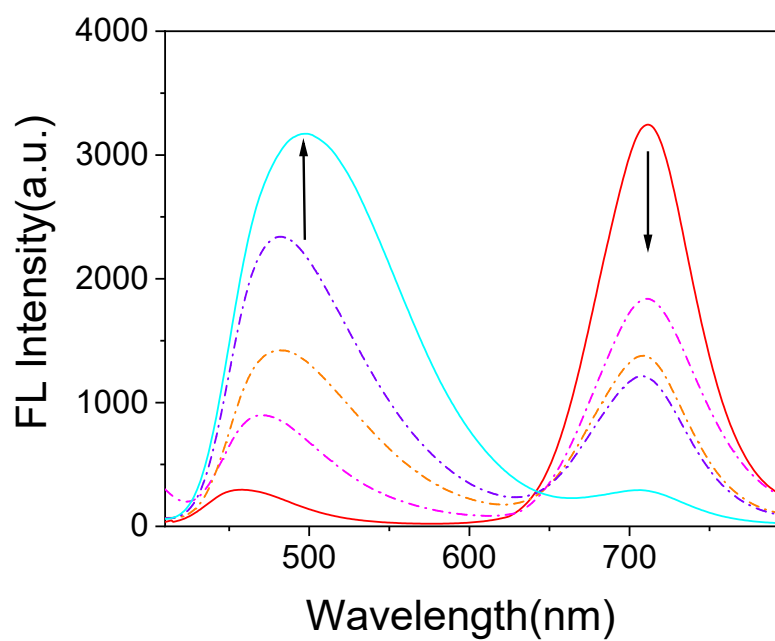


Fig. S15. Fluorescence spectra of **BDM-DNP** (10 μM) coexisting with 1 mM H_2S and 1 mM N_2H_4 .