

## Electronic Supplementary Material

### Bio-Ionic Liquid based Self Healable and Adhesive Ionic Hydrogel for the On-demand Transdermal Delivery of Chemotherapeutic Drug

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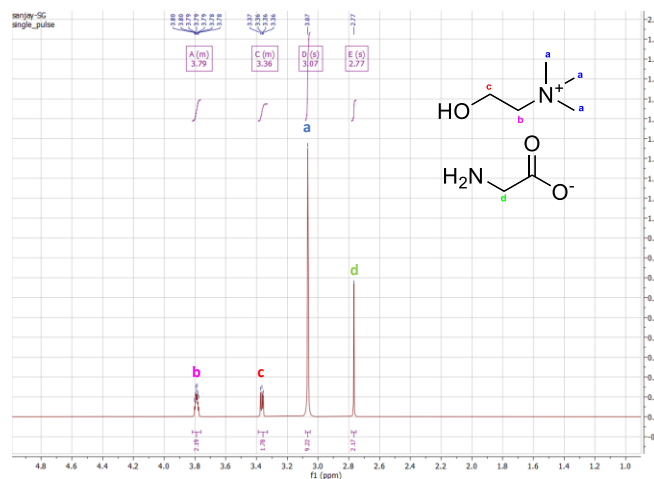
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### Mechanism for the Formation of Ionic Hydrogel

Ionic hydrogel was prepared by simple, drop wise addition of OA in aqueous solution of [Cho][Gly]. Here, [Cho][Gly] contain positively charged -NMe<sub>3</sub> whereas OA carries a negatively charged carboxylic -COO group that contributed to the hydrogel formation through electrostatic interaction. From the computational simulations we have confirmed that there is a significant change in the electronic charge distribution around the individual moieties before and after interacting with each other (**Figure 4**). In detail, there is a significant change in the negative charge distribution around the acid group of standalone OA<sup>1</sup> in regard with the OA interacting with the amine head group of the [Cho][Gly] (**Figure 4a**). Also, we can observe the changes in the positive charge distribution around the amine head of the [Cho][Gly]. This significant change in the charge distribution leads to a change in the behaviour of OA and [Cho][Gly] that in turn leads to changes in the shape of the aggregates from ellipsoidal to cylindrical to ionic hydrogel, SANS data. Further, the hydrogen bonding between the [Cho][Gly] and OA was also responsible for the formation of ionic hydrogel, FTIR results. These non-covalent interactions are confirmed through the change in the frequencies of the respective functional groups through FTIR and supported by the NCI analysis as well as the MESP calculations. The non-covalent interactions present between the molecules leads to the transition of the aggregates structure from ellipsoidal micelles to

34 cylindrical micelles to worm-like micelles to ionic hydrogel, as shown by SANS  
 35 analysis. The process of hydrogel formation is influenced by various factors, such  
 36 as the concentration of OA and pH of the system, and is important for  
 37 understanding the behavior of the molecules in different contexts.

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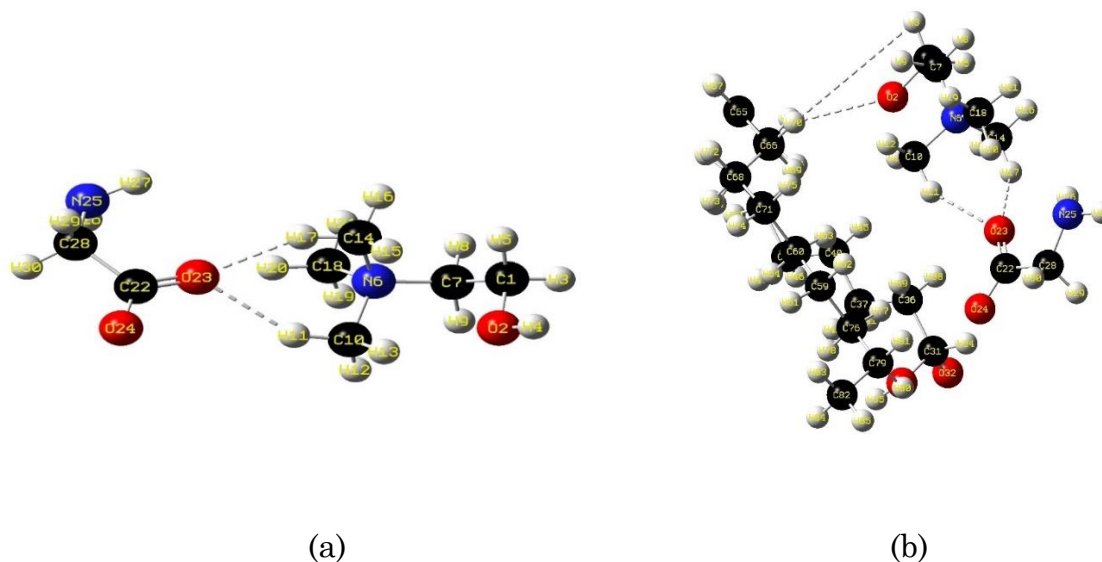
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**Figure S1:**  $^1\text{H-NMR}$  spectra of  $[\text{Cho}][\text{Gly}]$ .

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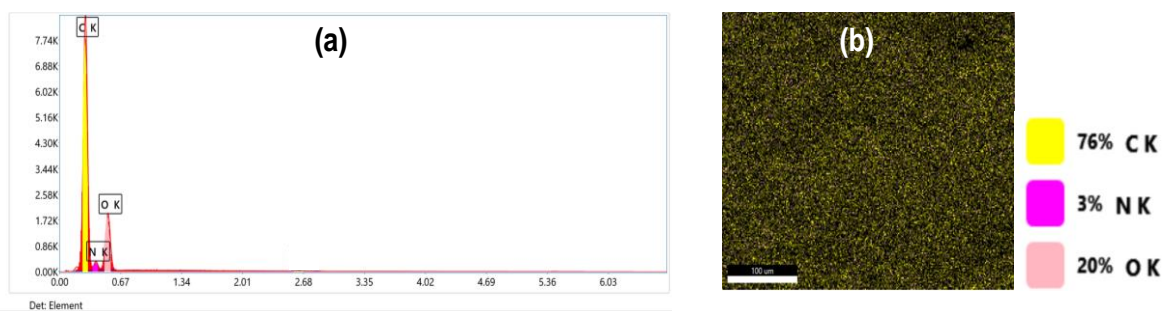
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**Figure S2:** Optimization structure of (a)  $[\text{Cho}][\text{Gly}]$  and (b)  $[\text{Cho}][\text{Gly}]/\text{Oleic acid}$   
 47 complex.

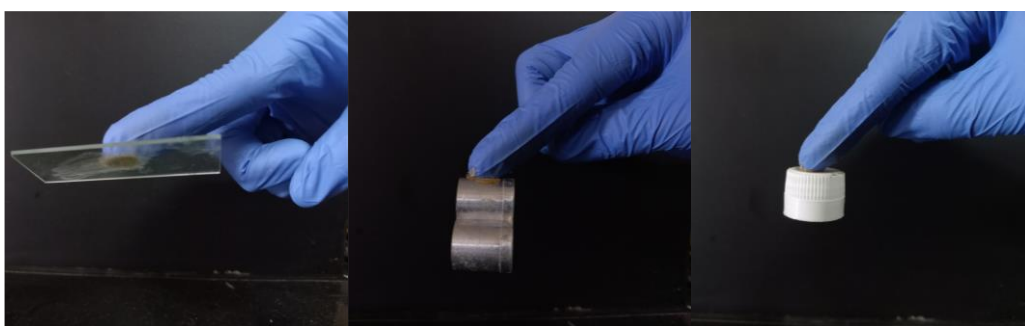
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50 **Figure S3:** EDX analysis of [Cho][Gly] and OA base ionic hydrogel (a) EDX  
 51 mapping and (b) EDX image.

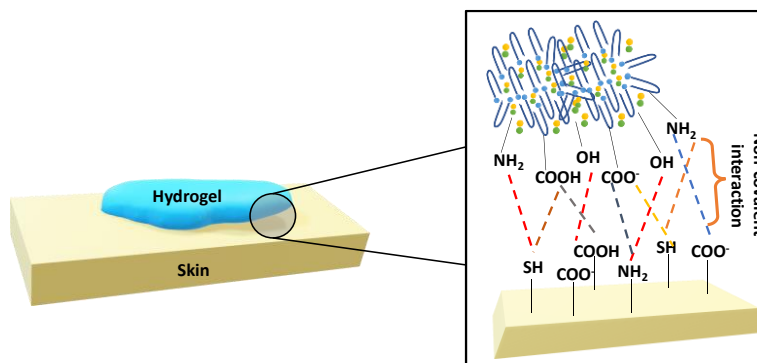
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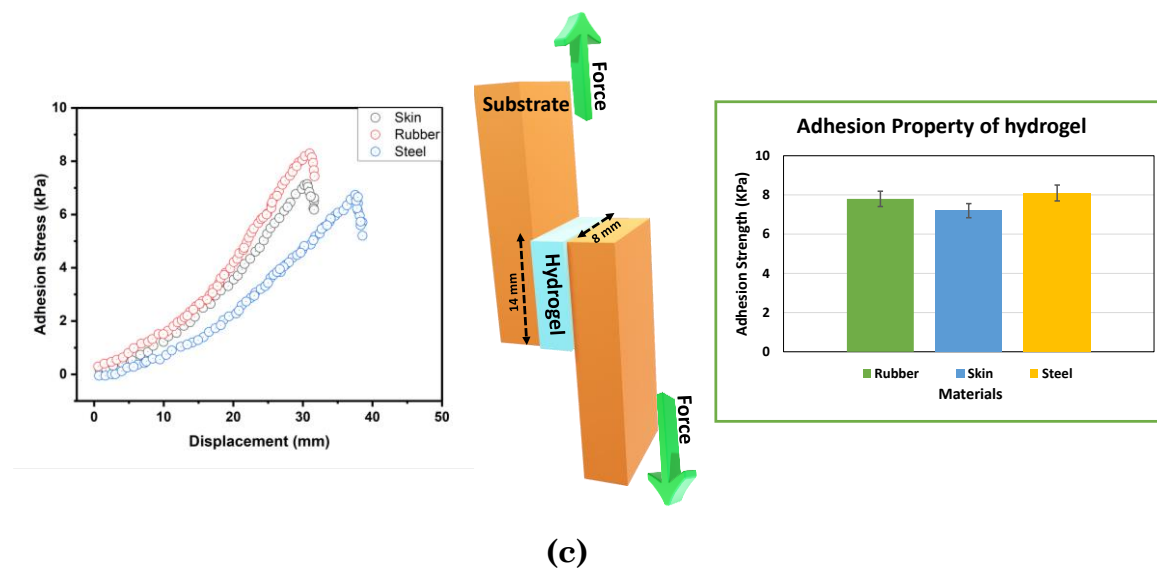
(a)



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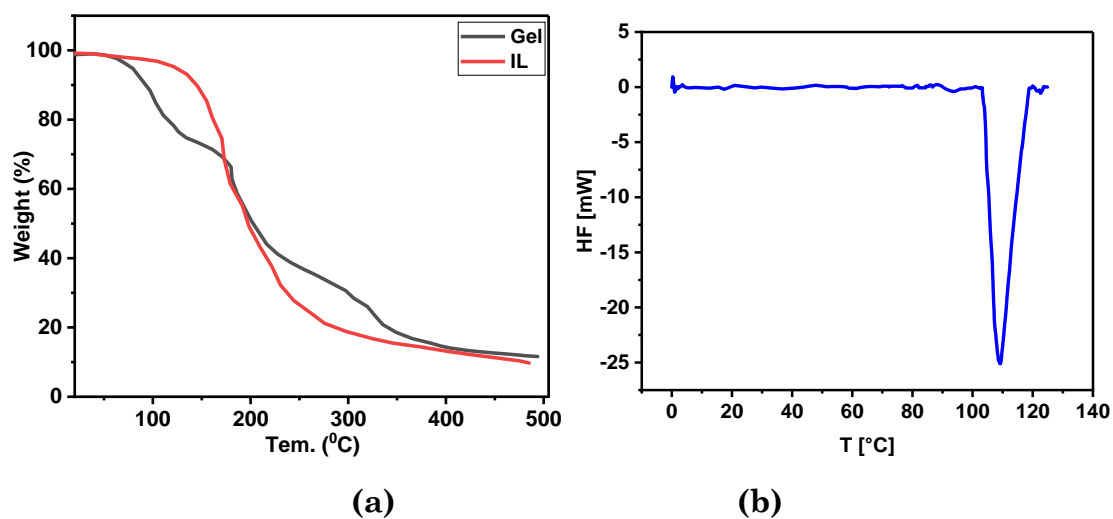
(b)



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59 **Figure S4:** (a) Visual image of adhesiveness of ionic hydrogel with glass, steel  
 60 and plastic materials, (b) Graphical representation of interaction  
 61 between ionic hydrogel and other material (c) Result of lap shear  
 62 test.



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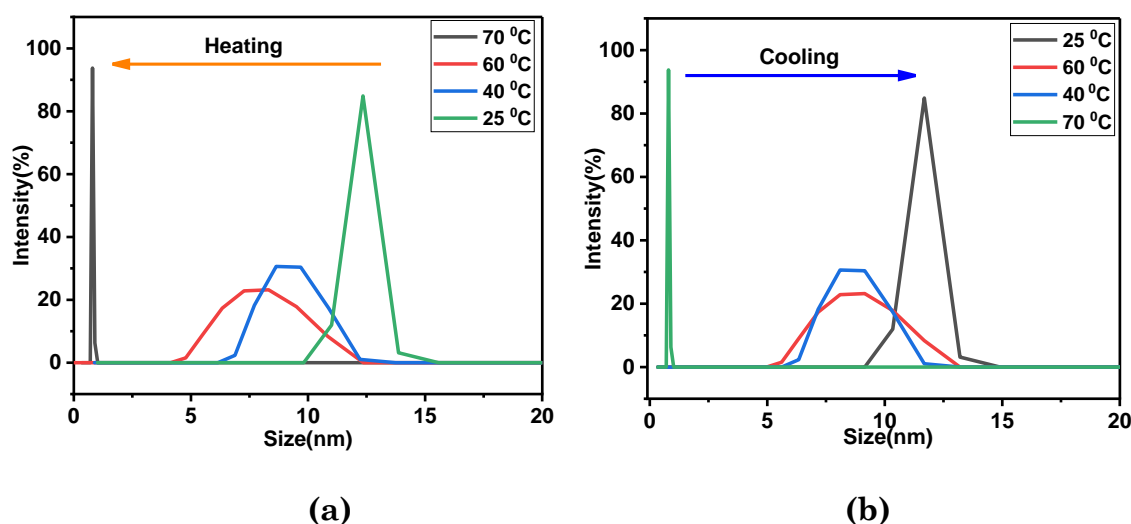
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66 **Figure S5:** (a) TGA graph of hydrogel, [Cho][Gly]-OA Gel (**Black Line**),  
 67 [Cho][Gly] (**Red Line**), (b) DSC analysis of hydrogel.

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## 69 Effect of temperature on the ionic hydrogel

70 The temperature is the most important stimulus for stimuli-responsive  
 71 hydrogels, which significantly impacts aggregate size and drug release. We used  
 72 DLS as a function of temperature to investigate the temperature-responsive  
 73 behaviour of ionic hydrogel and the effect of temperature on aggregate size. As  
 74 the temperature of the hydrogel increased from 25 to 75 °C, the average size and  
 75 PDI value within the hydrogel decreased (**Table S3** and **Figure S6**). The non-  
 76 covalent interactions within the hydrogel are weakened when the temperature  
 77 increases, but upon decreasing the temperature, these interactions reform,  
 78 leading to the gel, transitioning back into its original form (**Figure S6** and **Table**  
 79 **S4**).



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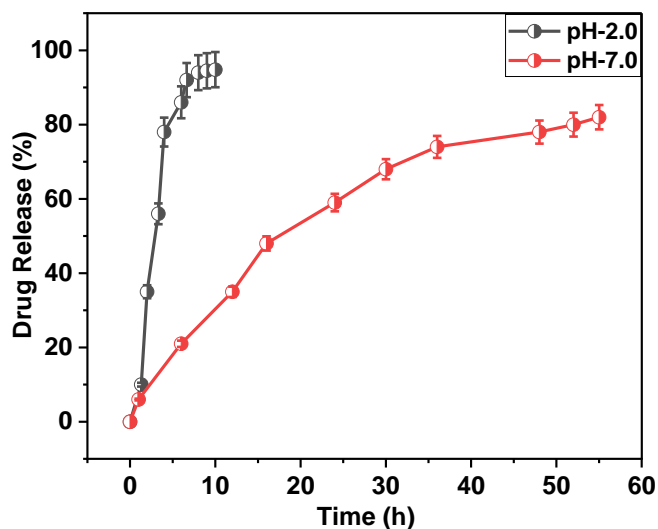
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82 **Figure S6:** Aggregates size of ionic hydrogel at different temperature measure  
 83 by DLS analysis, (a) Heating of gel and (b) Cooling of gel.

## 84 *In vitro* drug release

85 As illustrated in **Figure S7**, 94% of the burst release occurs within 8 hours,  
 86 observed at pH 2 (gastric condition), attributed to the deformation of the three-  
 87 dimensional network below pH 5, leading to the release of encapsulated drug  
 88 molecules. In the case of neutral pH (pH-7.0) condition, 78 % drug was released  
 89 within 50 h. These findings imply that when utilizing this system orally, adverse  
 90 effects may be observed in the human body.<sup>2,3</sup> In TDD, drug release is sustained  
 91 and directed towards the targeted site through the skin, thereby minimizing the

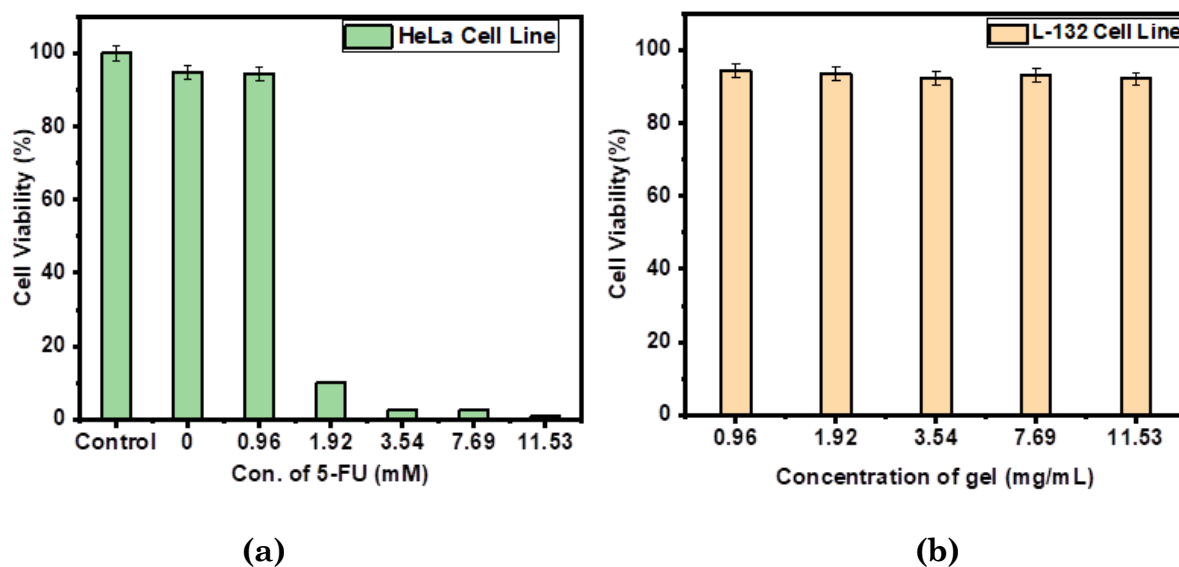
92 potential side effects associated with TDD-based systems as compared to other  
 93 ways.



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95 **Figure S7:** *In vitro* drug release at different pH values.

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99 **Figure S8:** (a) *In vitro* cytotoxicity of drug loaded ionic hydrogel on cervical  
 100 cancer cell line HeLa cells, (b) *in vitro* biocompatibility of ionic  
 101 hydrogel on normal cell line L-132 cell.

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**Tables**

105

106 **Table S1:** pH of system when addition of OA in aqueous solution of [Cho][Gly].

<b>[Cho][Gly] (M)</b>	<b>[OA] (M)</b>	<b>pH of system</b>	<b>Nature of system</b>
0.84	0.0	11.12	Solution
0.84	0.15	10.2	Solution
0.84	0.28	8.1	Viscus solution
0.84	0.63	7.5	Loose gel
0.84	0.75	7.0	Ionic Hydrogel

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108 **Table S2:** Compression of theoretical and practical elemental analysis of gel.

<b>Element present in gel</b>	<b>Theoretical % of element</b>	<b>% of element through EDX</b>
C	72.75	76.00
O	22.64	20.00
N	4.60	3.00

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110 **Table S3:** Average aggregates size of ionic hydrogel with increases  
111 temperature measured by DLS analysis.

<b>Temperature (°C)</b>	<b>Aggregate Size (nm)</b>	<b>PDI</b>
25	11.68	0.21
40	9.15	0.26
60	8.15	0.24
70	0.8	0.12

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113 **Table S4:** Average aggregates size of ionic hydrogel with decreases  
 114 temperature measured by DLS analysis.

Temperature (°C)	Aggregate Size (nm)	PDI
70	0.8	0.12
60	7.11	0.21
40	8.50	0.25
25	12.11	0.22

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116 **Table S5:** Cell viability data of ionic hydrogel with MCF-07 and M-132 Cell  
 117 lines.

Concentration of gel (mg/mL)	[Cho][Gly]-Gel	
	HaCaT cell line	L-132 cell line
0.96	98.2	94.23
1.92	98.0	93.54
3.54	97.5	92.35
7.69	95.1	93.21
11.53	93.2	92.10

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120 **Table S6:** FTIR peak shift of SC, after treated with various formulations.

Formulations	Lipid components of SC				Protein components of SC			
	Absorption CH <sub>2</sub> Asymmetric	Shift	Abs. CH <sub>2</sub> symm	Shift	Abs. -C=O Amide	Shift	Abs. HN- C=O amide	shift
[Cho][Gly]	2920.32	0.09	2850.8 1	0.02	1644.1 0	0.8	1539.2 5	0.05
[Cho][Gly]/OA hydrogel	2926.50	6.27	2857.2 5	6.46	1648.3 3	5.03	1545.5 4	6.3
PBS (Control)	2920.23	-	2850.7 9	-	1643.3 5	-	1539.2 0	-

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124 **References**

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