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Supporting Information

2 **Conditional Regulation of HCR for Rapid Visualization of Endogenous miR-21**

3 **in Cancer Cells**

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CONTENT

12

13	1. Materials and methods	3
14	1.1 Reagents and materials.....	3
15	1.2 Oligonucleotide labeling	3
16	1.3 Nucleic acids PAGE electrophoresis	3
17	1.4 HCR reaction under different buffers	4
18	1.5 Live-cell fluorescence imaging	4
19	2. Supplementary Figures and Tables	5
20	Fig. S1 Different types of nucleic acid assemblies	5
21	Table S1 The oligonucleotide sequences of HCR.....	6

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24 **1. Materials and methods**

25 **1.1 Reagents and materials**

26 The oligonucleotide was chemically synthesized by General Biosystems Co., Ltd (Anhui,
27 China). Acrylamide, Acryl/Bis 30% Solution (29:1), Water-DEPC Treated Water, RNase-free
28 ddH₂O, Ethidium bromide bought by Sangon Biotech Co., Ltd (Shanghai, China). Dulbecco's
29 modified essential media (DMEM), Hyclone fetal bovine serum (FBS), Hoechst 33342
30 (62249), AF568-NHS Ester, Polyethylenimine, Lipofectamine™ 3000 Transfection Reagent
31 were purchased from ThermoFisher Scientific. Hela cells (ATCC): CCL-2, human, female;
32 HEK293T(ATCC): CRL3216, human, sex unknown; 3T3(ATCC): L1, mouse, sex unknown.
33 All other chemicals, including solvents, were purchased from Sigma-Aldrich, Aladdin, and
34 Adamas and used without further purification.

35 **1.2 Oligonucleotide labeling**

36 The oligonucleotide was synthesized and purified by PAGE. FAM or active amino group
37 were modified on oligonucleotide chains H₁ and H₂, respectively. The oligonucleotide was
38 dissolved in ddH₂O and used directly for transcription in vitro. The purified H₂ oligonucleotide
39 and AF568-NHS Ester (molar ratio of 3:1) were incubated in PBS buffer (20 mM, pH 7.4) at
40 room temperature for 2 h and then purified by G-25 prepacked column. The modified H₁-FAM
41 and H₂-AF568 oligonucleotide chains were dissolved in the hairpin folding solution (20 mM
42 Tris-HCl, 137 mM NaCl, pH 7.4) to produce a mother solution with a final concentration of 10
43 μM.

44 **1.3 Nucleic acids PAGE electrophoresis**

45 All appliances were treated with ddH₂O water to remove bacteria and RNase. Each 10 mL
46 contains 6.66 mL 30% acrylamide, 1 mL 50×TAE, 50 μL 10% AP, and 5 μL TEMED, the
47 final concentration of electrophoresis gel was 15%. In vitro experiments, the uracil in the RNA

48 sequence of miR-21 was replaced with thymine called "I" (Table S1) to become a single-
49 stranded DNA (ssDNA) sequence to resist RNA degradation, which did not affect the
50 complementary base pairing between nucleic acid chains. The initiator chain "I" and the folded
51 hairpin chains "H₁" and "H₂" are mixed at different concentrations and react at room
52 temperature for 4 h or 37 °C 2 h. Then the 9 μL reacted sample is mixed with 1 μL 10×loading
53 buffer and added directly into the polyacrylamide gel without heating. After 135 V for 50 min,
54 it was stained with EB in ddH₂O for 10 min. The gel is photographed under ultraviolet light.
55 The extraction of image signal is realized through ImageJ.

56 **1.4 HCR reaction under different buffers**

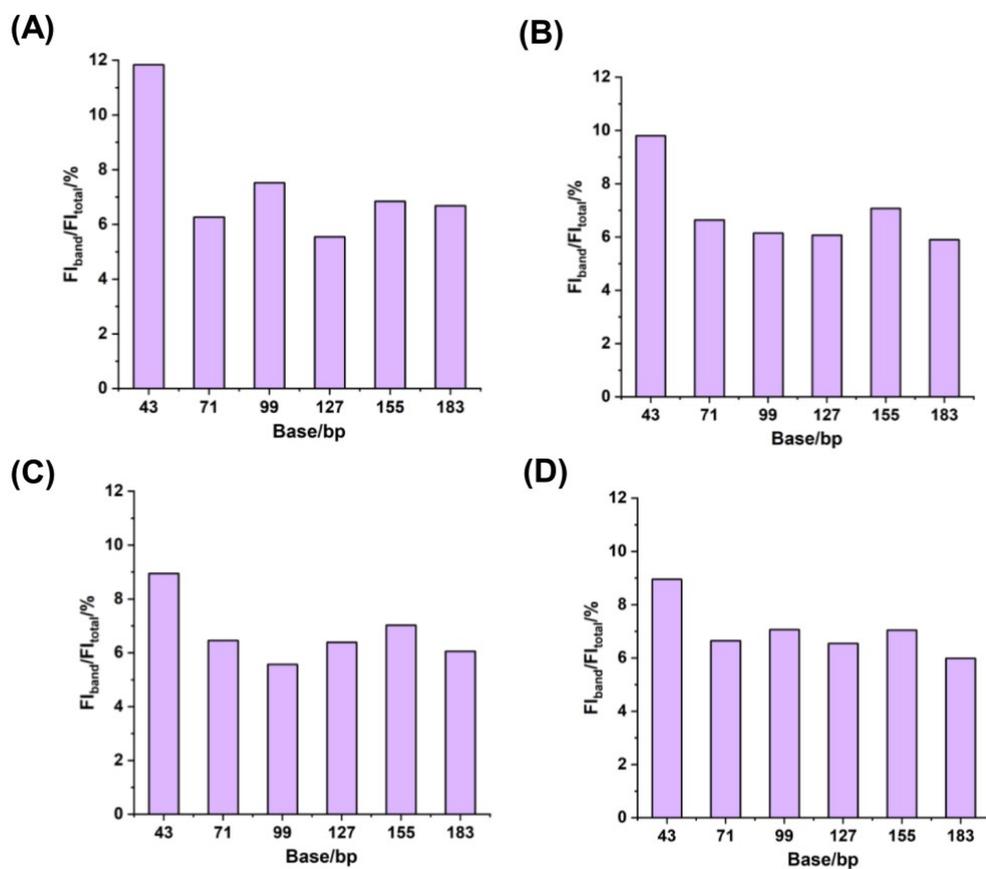
57 After the oligonucleotide chains "H₁" and "H₂" were folded in the nucleic acids hairpin
58 folding solution, they were added to six different assembly buffers respectively with the
59 simulated nucleic acid chain "I" of miR-21, and reacted at 37 °C 2 h to vary the influence of
60 different ion concentrations on the HCR assembly reaction. The six different buffer components
61 are as follows: buffer 1, 20 mM PBS, 10 mM MgCl₂; buffer 2, 20 mM PBS, 5 mM MgCl₂, 50
62 mM NaCl; buffer 3, 20 mM PBS, 10 mM MgCl₂, 50 mM NaCl; buffer 4, 20 mM PBS, 5 mM
63 MgCl₂, 50 mM NaCl, 20 mM HEPES; buffer 5, 20 mM PBS, 10 mM MgCl₂, 50 mM NaCl, 20
64 mM HEPES; buffer 6, 20 mM PBS, 5 mM MgCl₂. Then after polyacrylamide gel
65 electrophoresis, the gel is photographed under ultraviolet light. The extraction of image signal
66 is realized through ImageJ.

67 **1.5 Live-cell fluorescence imaging**

68 HeLa, 293T, 3T3 cells were seeded into a confocal imaging plate with 70-80% confluency.
69 The cells were transfected with 300 ng H₁-FAM and 300 ng H₂-AF568 in sequence per plate
70 (200 μL) using Lipofectamine 3000 (2 μL) and incubated for 2h. The cells were then incubated
71 with the fresh medium containing 500 nM Hoechst 33342. After incubation at 37 °C for 1 h
72 with 5% CO₂, the fluorescence imaging was performed with a 100 × oil-dripping objective lens
73 by using an ANDORTM living cell laser scanning confocal microscope (Revolution WD). For
74 H₁-FAM: λ_{ex} = 488 nm, λ_{em} = 500-570 nm, For H₂-AF549, λ_{ex} = 561 nm, λ_{em} = 580-650 nm.

75 The extraction of image signal is realized through ImageJ.

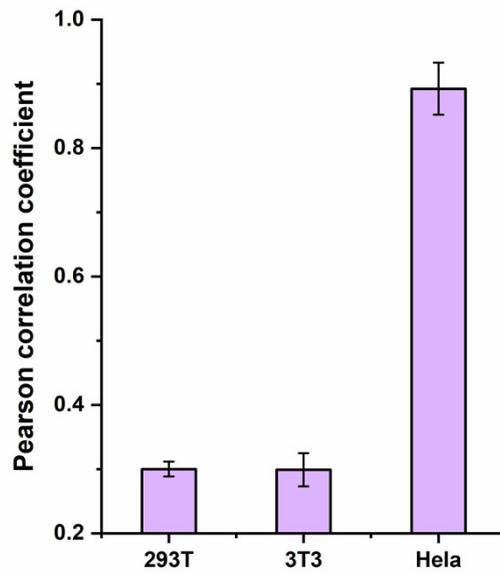
76 2. Supplementary Figures and Tables



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78 **Fig. S1(A) - (D)** The proportion of different types of nucleic acid assemblies, and the

79 concentration of miR-21 is 1 μM , 0.8 μM , 0.5 μM , 0.3 μM , respectively.



81

82 **Fig. S2** Colocalization of H1-FAM and H2-AF568 in three different cell lines in
 83 Figure 6A-C;

84 **Table S1** The oligonucleotide sequences of HCR.

Name	Sequence 5'-3'
miR-21	UAGCUUAUCAGACUGAUGUUGA
I	TAGCTTATCAGACTGATGTTGA
H₁	TCAACATCAGTCTGATAAGCTACTAAGTTAGCTTATC AGACTG
H₂	TAGCTTATCAGACTGATGTTGACAGTCTGATAAGCTA ACTTAG
H₁-FAM	TCAACATCAGTCTGATAAGCTACTAAGTTAGCTTATC AGACTG-FAM
H₂-AF568	AF568- TAGCTTATCAGACTGATGTTGACAGTCTGATAAGCTA ACTTAG

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