Supporting Information

Lipid structure-dependent CD1d functional stabilization and immunomodulation of endogenous glucosyl ceramides

Kazunari Ueki,¹ Risa Nozawa,¹ Takanori Matsumaru,¹ Sho Yamasaki^{2,3}

and Yukari Fujimoto¹

¹Faculty of Science and Technology, Keio University. Hiyoshi 3-14-1, Yokohama, Kanagawa 223-

8522, Japan

² Department of Molecular Immunology, Research Institute for Microbial Diseases, Osaka

University, Suita 565-0871, Japan

³ Laboratory of Molecular Immunology, Immunology Frontier Research Center (WPI-IFReC),

Osaka University, Suita 565-0871, Japan

E-mail: fujimotoy@keio.jp

Table of Contents

Supplemental Figures and Data:	
Synthesis	2
Biological assays	3
Experimental Section	
NMR Spectra	20

Ethics approval

Ethics approval for the genetic recombination experiments and the animal experiments was obtained from the Institutional Review Board of Keio University, in accordance with national and institutional guidelines for the safe and ethical conduct of genetic research.

Supplemental Figures and Data









Scheme S2. Synthesis of compounds 6



Figure S1. Antigen presenting cell (APC)-free assay for lipid binding to mCD1d-Fc fusion protein using the indicated a) GlcCer **1a–d**, **2a–d**. α -GalCer (KRN7000) was used as a reference. The graphs show the mean \pm SD of triplicate measurements, and the results shown are representative of at least three independent experiments. b) Overview of antigen presenting cell (APC)-free assay for lipid binding to mCD1d-Fc fusion protein.



Figure S2. Cytokines, IL-4, IL-17A and IL-10, secretion by mouse splenocytes following stimulation by ligands **1a–d** and **2a–d**. The graphs show the mean \pm SD of triplicate measurements, and the results shown are representative of at least three independent experiments. a) IL-4, b) IL-17A and c) IL-10 secretion stimulated with GlcCer **1a–d** and **2a–d**.



Figure S3. Inhibitory activities of GlcCer (1c, 2c) along with KRN7000 100 nM against IFN- γ induction in mouse splenocytes.

Experimental Section

Synthesis: General procedures

Nuclear magnetic resonance (¹H NMR, ¹³C NMR) spectra were measured in an indicated solvent with either JEOL AL400, ECX 400 or ECS 400. The proton chemical shifts in CDCl₃ are reported in parts per million (δ) using tetramethylsilane as an internal standard and coupling constants are in Heltz (Hz). The chemical shifts in other solvents are reported in ppm from the residual proton signal of the solvent. The chemical shifts for ¹³C NMR are reported in ppm from the internal solvent signal (CDCl₃, δ 77.16). High-resolution mass spectra (HRMS) of synthetic compounds were obtained on an electron spray ionization quadrupole time of flight (ESI-QTOF) mass spectrometer (micrOTOF-QII-HC; BRUKER). Analytical thin layer chromatography (TLC) was performed on Silica gel 60 F₂₅₄ Plates (Merck, 0.25 mm thickness). Silica gel column chromatography was performed using Silica gel 60 N [spherical neutral (Kanto Chemical Co., Inc., 40–50 mm)] at medium pressure (2–4 kgcm⁻² using indicated solvent systems. Reagents were purchased from commercial suppliers (TCI, nacalai tesque, FUJIFILM Wako Pure Chemical Corporation, Merck, Kanto Chemical Co., Inc., Watanabe Chemical Industries, ltd.) and were used without further purification. Unless otherwise noted, non-aqueous reactions were carried out under an argon atmosphere. Anhydrous dichloromethane, tetrahydrofuran, N, N-dimethylformamide, methanol, and toluene were purchased from Kanto Chemical Co., Inc.



Compound <u>S3</u>: To a stirred solution of S1¹ (1.00 g, 3.49 mmol) in anhydrous DMF (15.2 mL) were added NaH 60% suspension in mineral oil (616 mg, 15.4 mmol) and AllylBr (1.28 mL, 14.7 mmol) at 0 °C, and the mixture was stirred at room temperature for 18 h. The reaction was quenched with ice water, and the whole was extracted with Et₂O and washed with H₂O and brine, dried over anhydrous Na₂SO₄, filtered, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (*n*-hexane/EtOAc = 7/1) to afford **S3** (1.23 g, 79%) as clear oil; ¹H NMR (CDCl₃) δ 7.06–6.97 (m, 2H), 6.81 (d, *J* = 8.5 Hz, 2H), 6.06–5.84 (m, 4H), 5.34–5.23 (m, 4H), 5.18–5.15 (m, 4H), 4.47–4.39 (m, 1H), 4.30 (ttt, *J* = 21.3, 7.4, 2.3 Hz, 3H), 4.21–4.11 (m, 2H), 4.08–3.80 (m, 3H), 3.76 (t, *J* = 3.1 Hz, 3H), 3.63 (dt, *J* = 11.9, 4.1 Hz, 1H), 3.58–3.51 (m, 2H), 3.45 (t, *J* = 4.6 Hz, 2H); ¹³C NMR (CDCl₃) δ 155.2, 154.9, 150.7, 135.3, 134.9, 134.7, 134.4, 118.4, 118.1, 117.6, 117.3, 117.1, 116.9, 116.8, 116.5, 102.7, 96.5, 84.1, 81.4, 79.3, 75.0, 74.5, 73.9, 72.4, 70.6, 68.2, 55.6; HRMS (ESI-QTOF) calcd for C₂₅H₃₄O₇ [M+Na]⁺ 469.2197, found 469.2203.

Compound <u>S4</u>: To a stirred solution of S2¹ (4.00 g, 14.0 mmol) in anhydrous DMF (60.7 mL) were added NaH 60% suspension in mineral oil (2.46 g, 61.5 mmol) and AllylBr (5.32 mL, 61.5 mmol) at 0 °C, and the mixture was stirred at room temperature for 24 h. The reaction was quenched with ice water, and the whole was extracted with Et₂O and washed with H₂O and brine, dried over anhydrous Na₂SO₄, filtered, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (*n*-hexane/EtOAc = 6/1) to afford **S4** (5.16 g, 83%) as clear oil; ¹H NMR (CDCl₃) δ 7.05–6.92 (m, 2H), 6.84–6.68 (m, 2H), 6.03–5.73 (m, 4H), 5.46–5.43 (m, 1H), 5.36-5.09 (m, 8H), 4.77–4.72 (m, 1H), 4.44–3.87 (m, 13H), 3.84–3.79 (m, 1H), 3.70–3.54 (m, 1H), 3.49–3.30 (m, 1H); ¹³C NMR (CDCl₃) δ 155.2, 151.8, 135.5, 135.4, 135.3, 135.2, 135.1, 135.0, 134.5, 118.7, 117.4, 117.2, 117.1, 116.8, 116.7, 116.5, 114.5, 114.4, 103.2, 97.6, 81.5, 79.0, 78.3, 76.1, 74.5, 74.2, 74.0, 73.6, 72.5, 72.3, 72.0, 71.8, 69.7, 68.5, 55.7; HRMS (ESI-QTOF) calcd for C₂₅H₃₄O₇ [M+Na]⁺ 469.2197, found 469.2203.

Compound <u>S5</u>: To a stirred solution of **S3** (30.4 mg, 0.068 mmol) in MeCN (419 µL) and H₂O (419 µL) was added CAN (73.5 mg, 0.134 mmol), and the mixture was stirred at 0 °C for 15 min. The mixture was diluted with brine, and the whole was extracted with EtOAc, dried over anhydrous Na₂SO₄, filtered, and then concentrated under reduced pressure to affrd the residue, which was purified by silica gel column chromatography (*n*-hexane/EtOAc = 2/1) to afford **S5** (19.9 mg, 86%) as a brown solid; ¹H NMR (CDCl₃) δ : 6.02–5.86 (m, 4H), 5.27 (dt, *J* = 17.2, 5.1 Hz, 4H), 5.14 (m, 4H), 4.39–3.93 (m, 9H), 3.74–3.58 (m, 3H), 3.38 (m, 2H), 3.24–3.14 (1H, m); ¹³C NMR (CDCl₃) δ 135.7, 135.4, 115.8, 115.6, 115.4, 115.2, 89.6, 80.4, 79.6, 77.5, 73.5, 72.9, 72.8, 72.5, 72.4, 71.1, 70.3, 69.1; HRMS (ESI-QTOF) calcd for C₁₈H₂₈O₆ [M+Na]⁺ 363.1778, found 363.1787.

Compound <u>S6</u>: To a stirred solution of S4 (2.42 g, 5.42 mmol) in MeCN (33.9 mL) and H₂O (33.9 mL) was added CAN (5.94 g, 10.8 mmol), and the mixture was stirred at 0 °C for 40 min. The mixture was diluted with brine, and the whole was extracted with EtOAc, dried over anhydrous Na₂SO₄, filtered, and then concentrated under reduced pressure to afford the residue, which was purified by silica gel column chromatography (*n*-hexane/EtOAc = 2/1 to 1/1) to afford S6 (1.55 g, 84%) as a brown solid; ¹H NMR (CDCl₃) δ : 6.00–5.86 (m, 4H), 5.42–5.10 (m, 9H), 4.97–4.76 (m, 1H), 4.60 (dd,

J = 7.5, 1.5 Hz, 1H), 4.42–3.94 (m, 9H), 3.84–3.81 (m, 1H), 3.78–3.67 (m, 2H), 3.67–3.54 (m, 1H); ¹³C NMR (CDCl₃) δ 135.3, 134.9, 134.7, 134.3, 117.8, 117.5, 117.4, 116.9, 116.7, 97.6, 92.0, 74.5, 74.0, 72.7, 72.5, 71.6, 69.4, 68.9; HRMS (ESI-QTOF) calcd for C₁₈H₂₈O₆ [M+Na]⁺ 363.1778, found 363.1787.



Compound <u>5</u>: To a stirred solution of **S5** (261 mg, 0.767 mmol) in CH₂Cl₂ (15.3 mL) were added 2,2,2-trifluoro-*N*-phenylacetimidoyl chloride (145 μ L, 0.922 mmol), DIPEA (261 μ L, 1.53 mmol), and DMAP (81.5 mg, 0.667 mmol). The mixture was stirred at room temperature for 4 h, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (*n*-hexane/EtOAc = 9/1) to afford **5** (373 mg, 95%) as yellow oil; ¹H NMR (CDCl₃) δ : 7.28 (dd, *J* = 13.2, 4.9 Hz, 2H), 7.09 (t, *J* = 7.3 Hz, 1H), 6.83 (d, *J* = 7.3 Hz, 2H), 6.03–5.84 (m, 4H), 5.32–5.26 (m, 4H), 5.19 (tt, *J* = 9.0, 3.6 Hz, 4H), 4.41–3.98 (m, 9H), 3.85 (s, 1H), 3.74 (q, *J* = 7.8 Hz, 1H), 3.66 (dd, *J* = 12.7, 8.3 Hz, 2H), 3.52 (dd, *J* = 21.5, 11.7 Hz, 2H); ¹³C NMR (CDCl₃) δ 135.0, 134.7, 134.5, 134.4, 129.1, 128.7 (3C), 127.4, 120.6, 117.6 (2C), 117.5, 117.2, 117.1 (2C), 116.8 (2C), 116.7, 80.3, 74.4, 74.1, 73.9, 73.0, 72.5, 72.4; HRMS (ESI-QTOF) calcd for C₂₆H₃₂F₃O₆ [M+Na]⁺ 534.2174, found 534.2074.

Compound <u>6</u>: To a stirred solution of **S6** (1.10 g, 3.23 mmol) in CH₂Cl₂ (64.6 mL) were added 2,2,2-trifluoro-*N*-phenylacetimidoyl chloride (558 μ L, 3.55 mmol), DIPEA (1.10 mL, 6.46 mmol), and DMAP (276 mg, 2.26 mmol). The mixture was stirred at room temperature for 14 h, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (*n*-hexane/EtOAc = 9/1) to afford **6** (1.40 g, 85%) as yellow oil; ¹H NMR (CDCl₃) δ 7.34–7.03 (m, 4H), 7.10–6.98 (m, 1H), 6.80 (d, *J* = 7.2 Hz, 2H), 5.98–5.82 (m, 4H), 5.42–5.14 (m, 8H), 4.41–4.34 (m, 1H), 4.27–3.91 (m, 8H), 3.80–3.74 (m, 2H), 3.67–3.53 (m, 2H); ¹³C NMR (CDCl₃) δ 135.3, 135.2, 134.9, 134.8, 134.7 (2C), 134.4 (3C), 129.5, 128.8, 128.7, 126.5, 124.2, 120.5, 119.6, 119.4, 117.6 (2C), 117.3, 117.2, 117.0, 116.7 (2C), 81.4, 75.5, 74.3, 74.3, 74.2, 74.1, 72.9, 72.5, 72.0, 71.9, 68.3, 67.9; HRMS (ESI-QTOF) calcd for C₂₆H₃₂F₃O₆ [M+Na]⁺ 534.2174, found 534.2074.



Compound <u>S8</u>: To a stirred solution of S7² (877 mg, 1.55 mmol) in DMF (15.5 mL) were added NaH 60% suspension in mineral oil (99.2 mg, 2.48 mmol) and AllylBr (201 µL, 2.32 mmol) at 0 °C, and the mixture was stirred at room temperature for 23 h. The mixture was diluted with ice water. The whole was extracted with Et₂O and washed with H₂O and brine, dried over anhydrous Na₂SO₄, filtrated, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (*n*-hexane/EtOAc = 15/1) to afford **S8** (891 mg, 95%) as a white solid; ¹H NMR (CDCl₃) δ 7.51–7.41 (m, 5H), 7.35–7.22 (m, 10H), 5.80 (dq, *J* = 11.1, 5.4 Hz, 1H), 5.68–5.61 (m, 1H), 5.30–5.11 (m, 2H), 4.01 (dd, *J* = 13.1, 5.0 Hz, 1H), 3.90 (dd, *J* = 8.4, 5.7 Hz, 1H), 3.76 (dd, *J* = 12.9, 5.7 Hz, 1H), 3.63–3.56 (m, 1H), 3.30 (dd, *J* = 9.7, 6.6 Hz, 1H), 3.21 (q, *J* = 4.7 Hz, 1H), 2.05–1.98 (m, 2H), 1.44–1.16 (m, 20H), 0.93–0.84 (m, 3H); ¹³C NMR (CDCl₃) δ 143.8 (3C), 137.7, 134.7 (2C), 128.8, 127.9, 127.2, 127.1, 126.0, 116.7, 87.1, 79.6, 77.5, 77.1, 76.8, 69.0, 65.0, 63.1, 53.5, 32.4, 32.0, 29.8 (2C), 29.7, 29.6 (2C), 29.5, 29.3, 29.1, 22.8, 14.3; HRMS (ESI-QTOF) calcd for C₄₀H₅₃O₂ [M+Na]⁺ 630.4030, found 630.4022.



Compound 7: To a stirred solution of compound **S8** (810 mg, 1.33 mmol) in CH₂Cl₂ (6.8 mL) and MeOH (3.4 mL) was added *p*-TsOH·H₂O (278 mg, 1.46 mmol), and the mixture was stirred at room temperature for 2.5 h. The mixture was quenched with saturated aqueous NaHCO₃, and the whole was extracted with EtOAc and washed with saturated aqueous NaHCO₃ and brine, dried over anhydrous Na₂SO₄, filtrated, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (*n*-hexane/EtOAc = 5/1) to afford 7 (443 mg, 91%) as colorless oil; ¹H NMR (CDCl₃) δ 5.94–5.84 (m, 1H), 5.77 (dt, *J* = 15.1, 6.8 Hz, 1H), 5.39 (q, *J* = 8.0 Hz, 1H), 5.29–5.18 (m, 2H), 4.16–4.07 (m, 1H), 3.92–3.80 (m, 2H), 3.80–3.70 (m, 2H), 3.50 (q, *J* = 5.4 Hz, 1H), 2.17 (t, *J* = 6.3 Hz, 1H), 2.10 (q, *J* = 6.8 Hz, 2H), 1.41 (t, *J* = 6.8 Hz, 2H), 1.27–1.20 (m, 20H), 0.89 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (CDCl₃) δ 138.2, 134.3, 126.1, 117.3, 80.9, 77.4, 77.1, 76.8, 69.1, 66.0, 62.7, 32.4, 32.0, 29.7 (3C), 29.5 (2C), 29.5 (2C), 29.2, 29.0, 22.8, 14.2; HRMS (ESI-QTOF) calcd for C₂₁H₃₉O₂ [M+Na]⁺ 388.2934, found 388.2930.



Compound S10\alpha and S10\beta: To a solution of compound 5 (1284 mg, 2.52 mmol), and compound S9³ (860 mg, 1.93 mmol) in CH₂Cl₂ (29.7 mL) was stirred for 30 min in the presence of MS 4Å. The mixture was cooled to -50 °C then TMSOTf (90.8 μ L, 0.50 mmol). After stirring for 19 h, the reaction was quenched with Et₃N, the mixture was filtered, and the solvent was concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (nhexane/EtOAc = 6/1) to afford S10 α (597 mg, 33%) as yellow solid, and S10 β (732 mg, 40%) as yellow oil; **S10a**: ¹H NMR (CDCl₃) δ : 7.22 (t, J = 7.3 Hz, 2H), 6.83 (t, J = 13.5 Hz, 2H), 6.01–5.82 (m, 4H), 5.79-5.71 (m, 1H), 5.41 (dd, J = 15.5, 8.5 Hz, 1H), 5.27 (d, J = 17.1 Hz, 4H), 5.16 (dd, J = 17.1 Hz, 516.3, 10.0 Hz, 4H), 4.87 (d, J = 3.1 Hz, 1H), 4.52 (d, J = 11.4 Hz, 1H), 4.29 (m, 4H), 4.14–4.07 (m, 4H), 4.00 (m, 2H), 3.89 (t, J = 6.8 Hz, 1H), 3.78 (s, 3H), 3.71 (dd, J = 17.7, 8.8 Hz, 2H), 3.62 (t, J = 1.005.3 Hz, 3H), 3.53 (t, J = 8.9 Hz, 1H), 3.43 (t, J = 9.4 Hz, 1H), 3.35 (dd, J = 9.5, 3.3 Hz, 1H), 2.08 (dd, J = 9.5, 3.3 Hz, 1H), 3.53 (dd, J = 9.5, 3.5 Hz, 1H), 3.5 (dd, JJ = 15.9, 9.0 Hz, 2H), 1.41 (s, 2H), 1.26 (s, 20H), 0.88 (t, J = 6.5 Hz, 3H); ¹³C NMR (CDCl₃) δ 159.0, 138.0, 135.3, 135.1, 134.9, 134.8, 134.4, 130.1, 129.2, 129.1, 126.3, 125.9, 120.6, 120.1, 117.3, 117.1, 117.0, 116.7, 116.3, 114.2, 113.7, 97.9, 81.2, 79.3, 79.1, 74.1, 73.8, 72.4, 71.9, 70.4, 69.6, 68.3, 67.5, 64.3, 55.2, 32.3, 31.9, 29.6 (2C), 29.4, 29.3, 29.2, 29.0, 22.6; HRMS (ESI-QTOF) calcd for $C_{44}H_{69}N_{3}O_{8}$ [M+Na]⁺ 790.4977, found 790.4991; **<u>S10</u>**: ¹H NMR (CDCl₃) δ : 7.25 (dd, J = 16.6, 7.3) Hz, 2H), 6.85 (d, J = 8.8 Hz, 2H), 6.00–5.83 (m, 4H), 5.77–5.69 (m, 1H), 5.41 (dd, J = 15.6, 8.8 Hz, 1H), 5.26–5.22 (m, 3H), 5.15 (d, J = 10.2 Hz, 4H), 4.54 (d, J = 11.7 Hz, 1H), 4.29 (tt, J = 21.0, 6.6 Hz, 6H), 4.11 (dd, J = 12.7, 5.9 Hz, 3H), 4.02 (td, J = 12.6, 7.2 Hz, 2H), 3.94 (dd, J = 10.2, 6.8 Hz, 1H), 3.87 (dd, J = 8.3, 5.4 Hz, 1H), 3.78 (s, 3H), 3.68 (t, J = 5.6 Hz, 2H), 3.59 (dt, J = 17.4, 5.9 Hz, 2H), 3.37–3.32 (m, 3H), 3.18 (t, J = 8.3 Hz, 1H), 2.10 (q, J = 6.8 Hz, 2H), 1.40 (d, J = 6.8 Hz, 2H), 1.25 (d, J = 10.7 Hz, 20H), 0.88 (t, J = 6.6 Hz, 3H); ¹³C NMR (CDCl₃) δ 159.0, 138.0, 135.1, 135.0, 134.9, 134.7, 134.6, 134.5, 134.3, 130.0, 129.1, 125.7, 117.0, 116.9, 116.8 (2C), 116.5, 113.6, 103.2, 84.0, 81.3, 78.9, 76.7, 74.8, 74.3, 73.7, 73.5, 72.4, 69.4, 68.6, 68.4, 64.3, 55.1, 32.3, 31.8, 29.6 (2C), 29.4, 29.3, 29.1, 29.0, 22.6, 14.1, 14.0; HRMS (ESI-QTOF) calcd for C44H69N3O8 [M+Na]⁺ 790.4977, found 790.4991.



Compound 8\alpha and 8\beta: To a solution of compound 5 (294 mg, 0.57 mmol), and compound 7 (268 mg, 0.44 mmol) in CH₂Cl₂ (8.8 mL) was stirred for 30 min in the presence of MS 4Å. The mixture was cooled to -30 °C then TMSOTf (27 µL, 0.15mmol). After stirring for 7 h, the reaction was quenched with Et₃N, the mixture was filtered, and the solvent was concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (*n*-hexane/EtOAc = 6/1) to afford 8α (160 mg, 53%) as yellow solid, and 8β (121 mg, 40%) as yellow oil; 8α : ¹H NMR (CDCl₃) δ 5.96–5.78 (m, 5H), 5.73–5.65 (m, 1H), 5.37–5.17 (m, 5H), 5.16–5.05 (m, 5H), 4.34–4.13 (m, 6H), 4.12-3.96 (m, 5H), 3.91-3.73 (m, 3H), 3.70-3.50 (m, 4H), 3.39-3.31 (m, 3H), 3.20-3.09 (m, 1H), 2.08-1.95 (m, 2H), 1.34 (d, J = 5.0 Hz, 2H), 1.31-1.23 (m, 20H), 0.85 (t, J = 6.9 Hz, 3H); ${}^{13}C$ NMR (CDCl₃) δ 135.3, 135.1 (2C), 134.9 (2C), 134.8 (2C), 129.3, 126.2, 120.7, 117.1, 117.0, 116.9, 116.8, 116.7, 103.4, 84.2, 81.5, 77.5, 77.3, 77.1, 76.8, 75.0, 74.9, 74.5, 73.9, 72.5, 69.0, 68.9, 32.4, 32.0, 29.7 (3C), 29.5 (2C), 29.4 (2Cs), 29.3, 29.2, 29.1, 22.8, 14.2; HRMS (ESI-QTOF) calcd for C₃₉H₆₅N₃O₈₇ [M+Na]⁺ 710.4715, found 710.4711; **8β**: ¹H NMR (CDCl₃) δ 5.99–5.82 (m, 5H), 5.75–5.66 (m, 1H), 5.38–5.30 (m, 1H), 5.28–5.22 (m, 5H), 5.18–5.13 (m, 5H), 4.39–4.21 (m, 5H), 4.19–3.97 (m, 5H), 3.93–3.77 (m, 3H), 3.69–3.51 (m, 4H), 3.36–3.31 (m, 3H), 3.22–3.17 (m, 1H), 2.09–2.03 (m, 2H), 1.37 (t, J = 7.0 Hz, 2H), 1.26–1.21 (m, 20H), 0.87 (t, J = 6.8 Hz, 3H); ¹³C NMR (CDCl₃) δ 138.0, 137.8, 135.3, 135.2, 134.9, 134.8, 134.4, 117.1, 117.0, 116.9, 116.8, 116.7, 84.2, 84.1, 81.5, 79.7, 79.5, 77.5, 77.2, 76.8, 75.0, 74.9, 74.4, 73.9, 73.7, 72.5, 69.0, 67.5, 64.4, 58.5, 53.5, 49.6, 32.4, 32.0, 29.7, 29.5, 29.4, 29.3, 29.2, 29.1, 22.7, 14.2; HRMS (ESI-QTOF) calcd for C₃₉H₆₅N₃O_{8¥7} [M+Na]⁺ 710.4715, found 710.4720.

Compound <u>9a</u> and <u>9b</u>: To a solution of compound **6** (1.63 g, 3.19 mmol), and compound **7** (1.06 g, 2.90 mmol) in CH₂Cl₂ (44.6 mL) was stirred for 30 min in the presence of MS 4Å. The mixture was cooled to -30 °C then TMSOTf (136 µL, 0.75 mmol). After stirring for 15 h, the reaction was quenched with Et₃N, the mixture was filtered, and the solvent was concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (*n*-hexane/EtOAc = 6/1) to afford **9a** (734 mg, 37%) as yellow solid, and **9β** (861 mg, 43%) as yellow oil; **14β**: ¹H NMR (CDCl₃) δ 5.98–5.81 (m, 5H), 5.75–5.68 (m, 1H), 5.38–4.97 (m, 14H), 4.37–4.29 (m, 2H), 4.24–4.10 (m, 4H), 4.06–3.95 (m, 2H), 3.95–3.68 (m, 4H), 3.66–3.47 (m, 5H), 3.37–3.29 (m, 1H), 2.20–2.04 (m, 2H), 1.55–1.11 (m, 22H), 0.87 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (CDCl₃) δ 138.1, 135.5, 135.4, 135.1, 135.0, 134.8, 134.4, 129.5, 128.0, 126.5, 125.6, 120.6, 117.6, 117.1, 116.8, 116.7, 116.6, 103.8, 81.6,

79.6, 79.0, 74.1, 74.0, 73.4, 73.2, 72.5, 71.9, 69.0, 68.6, 64.4, 32.5, 32.0, 29.8, 29.6, 29.5, 29.3, 29.1, 22.8, 14.2; HRMS (ESI-QTOF) calcd for C₄₄H₆₉N₃O₈ [M+Na]⁺ 710.4715, found 710.4519.



Compound 10a: To a stirred solution of **8a** (148 mg, 0.22 mmol) in CHCl₃ (6.5 mL), MeOH (0.65 mL) and TFA (72 µL) was added [CpRu(C₃H₅)(C₉H₆NCOO)]PF₆ (9.0 mg, 17.2 µmol), and the mixture was stirred at room temperature for 15 h, and the reaction was quenched with SilicaMet[®] DMT, and the mixture was stirred at room temperature for 30 min, and filtered, and the solvent was concentrated to afford a residue, which was purified by silica gel column chromatography (CHCl₃/MeOH= 7/1) to afford **10a** (97.4 mg, 93%) as a brown solid; ¹H NMR (CD₃OD) δ : 5.72–5.64 (m, 1H), 5.43 (dd, *J* = 15.4, 7.5 Hz, 1H), 4.72 (dd, *J* = 9.5, 5.7 Hz, 1H), 4.09 (t, *J* = 6.1 Hz, 1H), 3.80–3.75 (m, 1H), 3.70 (dd, *J* = 11.8, 2.4 Hz, 1H), 3.58 (dt, *J* = 15.1, 4.5 Hz, 1H), 3.54–3.42 (m, 2H), 3.32–3.28 (m, 1H), 3.20 (dq, *J* = 9.7, 2.7 Hz, 3H), 1.98 (q, *J* = 6.7 Hz, 2H), 1.31 (d, *J* = 6.3 Hz, 2H), 1.19 (s, 20H), 0.82–0.77 (m, 3H); ¹³C NMR (CDCl₃) δ 135.8, 129.8, 101.0, 74.9, 74.0 (2C), 73.5, 73.4 (2C), 71.6 (2C), 68.8, 67.2, 62.5 (2C), 33.4, 33.1, 30.8 (2C), 30.6, 30.5, 30.3, 30.2, 23.7; HRMS (ESI-QTOF) calcd for C₂₄H₄₅N₃O₇ [M+Na]⁺ 510.3150, found: 510.3164.

Compound 106: To a stirred solution of **86** (244.3 mg, 0.36 mmol) in CHCl₃ (10.8 mL), MeOH (1.1 mL) and TFA (118 μ L) was added [CpRu(C₃H₅)(C₉H₆NCOO)]PF₆ (14.9 mg, 28.4 μ mol), and the mixture was stirred at room temperature for 16 h, and the reaction was quenched with SilicaMet[®] DMT, and the mixture was stirred at room temperature for 30 min, and filtered, and the solvent was concentrated to afford a residue, which was purified by silica gel column chromatography (CHCl₃/MeOH = 7/1) to afford**106** (150 mg, 87%) as a brown solid; ¹H NMR (CD₃OD) δ : 5.70–5.63 (m, 1H), 5.41 (dd, *J* = 15.4, 7.5 Hz, 1H), 4.19 (t, *J* = 9.6 Hz, 1H), 4.08 (dd, *J* = 7.3, 5.5 Hz, 1H), 3.83–3.75 (m, 2H), 3.61–3.51 (m, 3H), 3.28–3.14 (m, 3H), 3.10 (dd, *J* = 8.8, 7.9 Hz, 1H), 1.97 (q, *J* = 7.0 Hz, 2H), 1.30 (d, *J* = 6.7 Hz, 2H), 1.14 (d, *J* = 36.8 Hz, 20H), 0.80 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (CDCl₃) δ 135.9, 129.6, 104.5, 78.0 (2C), 75.0, 73.5 (2C), 71.5 (2C), 70.1, 67.3, 62.7 (2C), 33.4, 33.1, 30.8 (2C), 30.6, 30.5, 30.3, 30.2, 23.7, 14.5; HRMS (ESI-QTOF) calcd for C₂₄H₄₅N₃O₇ [M+Na]⁺ 510.3150, found 510.3163.

Compound 11a: To a stirred solution of **9a** (448 mg, 0.65 mmol) in CHCl₃ (19.8 mL), MeOH (1.98 mL) and TFA (217 µL) was added [CpRu(C₃H₅)(C₉H₆NCOO)]PF₆ (27.3 mg, 52.0 µmol), and the mixture was stirred at room temperature for 18 h, and the reaction was quenched with SilicaMet[®] DMT, and the mixture was stirred at room temperature for 30 min, and filtered, and the solvent was concentrated to afford a residue, which was purified by silica gel column chromatography (CHCl₃/MeOH= 7/1) to afford **11a** (259 mg, 82%) as a brown solid; ¹H NMR (CD₃OD) δ 5.78–5.70 (m, 1H), 5.51–5.46 (m, 1H), 4.22 (d, *J* = 7.7 Hz, 1H), 4.17 (dd, *J* = 7.0, 5.7 Hz, 1H), 3.90–3.83 (m, 2H), 3.78–3.71 (m, 2H), 3.70–3.59 (m, 2H), 3.55–3.44 (m, 2H), 3.29–3.28 (m, 1H), 2.07–1.99 (m, 2H), 1.38 (q, *J* = 6.8 Hz, 2H), 1.32–1.21 (m, 20H), 0.87 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (CDCl₃) δ 134.5, 128.3, 103.8, 75.2 (3C), 73.6, 72.2, 71.1(2C), 69.1, 68.7, 66.0, 61.2 (2C), 48.3, 48.1, 47.9, 47.7, 47.5, 47.3, 47.0, 32.1, 31.8, 29.5, 29.3, 29.2, 28.9, 28.9, 22.4, 13.2; HRMS (ESI-QTOF) calcd for C₂₄. H₄₅N₃O₇ [M+Na]⁺ 510.3150, found: 510.3164.



Compound 12a: To a compound **10a** (68.1 mg, 0.14 mmol) in THF (7.4 mL), MeOH (12.9 mL) and AcOH (5.5 mL), zinc dust (1.84 g, 28.3 mmol) was added. The mixture was subjected to sonication (bath sonication) for 20 min then filtered, and the filtrate was concentrated under the reduced pressure. The residue was dissolved in MeOH and treated with aqueous NH₄OH at room temperature for 1 h. The solvent was removed under reduced pressure to afford a residue, which was purified by silica gel column chromatography (CH₂Cl₂/MeOH/H₂O = 60/25/4) to afford **10a** (59.5 mg, 92%) as purple oil; ¹H NMR (CD₃OD) δ : 5.81–5.74 (m, 1H), 5.39 (dd, *J* = 15.2, 6.6 Hz, 1H), 4.73 (d, *J* = 3.6 Hz, 1H), 4.23 (dd, *J* = 12.2, 6.8 Hz, 1H), 3.94–3.84 (m, 1H), 3.71 (dd, *J* = 11.8, 1.8 Hz, 1H), 3.55 (dt, *J* = 16.9, 6.0 Hz, 2H), 3.42 (m, 3H), 3.29 (t, *J* = 5.0 Hz, 1H), 3.19 (dt, *J* = 14.3, 6.7 Hz, 1H), 2.00 (q, *J* = 6.9 Hz, 2H), 1.30 (s, 2H), 1.19 (s, 20H), 0.80 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (CD₃OD) δ 136.7, 126.1, 100.6, 74.9, 74.2, 73.3, 71.4 (2C), 62.5, 57.0, 33.4, 33.1, 30.9, 30.8 (3C), 30.7 (2C), 30.5, 30.4 (2C), 30.2, 23.7, 14.5; HRMS (ESI-QTOF) calcd for C₂4H₄₇NO₇ [M+Na]⁺ 484.3245, found: 484.3241.

Compound 12 β : To a compound **10** β (10.7 mg, 22.0 µmol) in THF (1.2 mL), MeOH (2.0 mL) and AcOH (870 µL), zinc dust (290 mg, 4.43 mmol) was added. The mixture was subjected to sonication (bath sonication) for 30 min then filtered, and the filtrate was concentrated under the reduced pressure. The residue was dissolved in MeOH and treated with aqueous NH₄OH at room temperature for 1 h.

The solvent was removed under reduced pressure to afford a residue, which was purified by silica gel column chromatography (CH₂Cl₂/MeOH/H₂O = 60/25/4) to afford **12**β (6.3 mg, 62%) as purple oil; ¹H NMR (CD₃OD) δ : 5.80–5.72 (m, 1H), 5.39 (dd, *J* = 15.4, 6.8 Hz, 1H), 4.21 (t, *J* = 6.3 Hz, 1H), 3.83 (ddd, *J* = 19.1, 13.5, 4.9 Hz, 3H), 3.56 (dd, *J* = 11.6, 5.7 Hz, 1H), 3.30–3.11 (m, 6H), 2.00 (q, *J* = 6.9 Hz, 2H), 1.32 (t, *J* = 7.5 Hz, 2H), 1.19 (s, 20H), 0.80 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (CD₃OD) δ 136.5, 128.5, 104.1, 78.1, 77.8, 74.8, 71.5, 71.1, 62.5, 56.7, 35.4, 33.4, 33.1, 30.9, 30.8 (3C), 30.7, 30.5 (2C), 30.4, 30.2, 23.7, 14.5; HRMS (ESI-QTOF) calcd for C₂₄H₄₇NO₇ [M+Na]⁺ 484.3245, found 484.3243.

Compound 13a: To a compound **11a** (148 mg, 0.30 mmol) in THF (11.1 mL), MeOH (19.4 mL) and AcOH (1.0 mL), zinc dust (4.0 g, 60.8 mmol) was added. The mixture was subjected to sonication (bath sonication) for 30 min then filtered, and the filtrate was concentrated under the reduced pressure. The residue was dissolved in MeOH and treated with aqueous NH₄OH at room temperature for 1 h. The solvent was removed under reduced pressure to afford a residue, which was purified by silica gel column chromatography (CH₂Cl₂/MeOH/H₂O = 60/25/4) to afford **13a** (67.9 mg, 48%) as purple oil; ¹H NMR (CD₃OD) δ 5.83 (dd, *J* = 15.0, 7.0 Hz, 1H), 5.47 (dd, *J* = 15.4, 6.6 Hz, 1H), 4.84 (d, *J* = 3.6 Hz, 1H), 4.30 (t, *J* = 5.5 Hz, 1H), 4.02–3.97 (m, 1H), 3.88 (t, *J* = 3.1 Hz, 1H), 3.82 (dd, *J* = 10.1, 3.6 Hz, 1H), 3.76–3.64 (m, 3H), 3.50–3.43 (m, 1H), 3.38 (dd, *J* = 7.5, 4.4 Hz, 1H), 3.30–3.27 (m, 1H), 2.07 (q, *J* = 6.7 Hz, 2H), 1.91 (d, *J* = 7.2 Hz, 2H), 1.39 (d, *J* = 6.1 Hz, 2H), 1.27 (s, 22H), 0.88 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (CD₃OD) δ 135.3, 127.0, 99.6, 71.6 (2C), 69.9, 69.6, 68.8, 64.5, 61.4, 55.5, 48.4, 48.1, 47.9, 47.7, 47.5, 47.3, 47.1, 32.1, 31.8, 29.5 (3C), 29.4 (2C), 29.2, 29.1, 28.9, 22.4, 21.8, 13.2; HRMS (ESI-QTOF) calcd for C₂₄H₄7NO₇ [M+Na]⁺ 484.3245, found: 484.3241.

12 or 13	RCO ₂ H (a–b) DMT-MM (OTf) EtOH/CH ₂ Cl ₂ rt	 α-GlcCer: 1a (25%), 1c (22%) β-GlcCer: 2a (40%), 2c (35%) α-GalCer: 3a (16%), 3c (17%) 	12 or 13	RCO ₂ H (c–d) HATU Et ₃ N, NMM THF rt	α-GlcCer: 1b (23%), 1d (23%) β-GlcCer: 2b (40%), 2d (44%) α-GalCer: 3b (73%), 3d (55%)
	R= /	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	b / (13	24:1 c	λ ₂₂ 24:0 d

Compound 1a: To a stirring solution of compound 12α (12.1 mg, 26.2 µmol) and oleic acid (8.14 mg, 28.8 µmol) in EtOH (250 µL) and CH₂Cl₂ (80 µL) at room temperature was added DMT-MM(OTf) (12.3 mg, 31.4 µmol). The mixture was stirred for 13 h at room temperature. The reaction was quenched with saturated aqueous NaHCO₃, the whole was extracted with EtOAc and washed with saturated aqueous NaHCO₃ and brine, dried over anhydrous Na₂SO₄, filtrated, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography

(CHCl₃/MeOH= 9/1 to 6/1) to afford **1a** (4.0 mg, 21%) as a white solid. ¹H NMR (CDCl₃/CD₃OD = 3/1) δ 5.67–5.61 (m, 1H), 5.36 (m, *J* = 15.5, 7.2 Hz, 1H), 5.26 (dq, *J* = 15.6, 5.1 Hz, 2H), 4.74 (d, *J* = 3.7 Hz, 1H), 4.01 (t, *J* = 7.0 Hz, 1H), 3.89–3.86 (m, 1H), 3.67 (m, 4H), 3.58 (t, *J* = 9.3 Hz, 1H), 3.48–3.45 (m, 1H), 3.37 (dd, *J* = 9.7, 3.7 Hz, 1H), 3.30 (t, *J* = 4.9 Hz, 1H), 3.27–3.26 (m, 2H), 2.16–2.03 (m, 2H), 1.93 (d, *J* = 3.7 Hz, 6H), 1.51 (s, 2H), 1.20 (s, 44H), 0.80 (t, *J* = 6.9 Hz, 6H); ¹³C–NMR (CDCl₃/CD₃OD = 3/1) δ 174.3, 133.8, 129.5, 129.3, 128.8, 99.0, 73.4, 71.7, 71.6, 69.8, 66.8, 61.0, 55.4, 53.2, 36.0, 32.0, 31.5 (2C), 29.4, 29.3 (5C), 29.2 (2C), 29.1 (2C), 29.0 (4C), 28.9 (4C), 28.8 (2C), 25.5, 22.3, 13.5 (2C); HRMS (ESI-QTOF) calcd for C₄₂H₇₉NO₈: [M+Na]⁺, 748.5698; found: 748.5698.

Compound 2a: To a stirring solution of compound **12** β (14.9 mg, 32.3 µmol) and oleic acid (10.0 mg, 35.5 µmol) in EtOH (300 µL) and CH₂Cl₂ (100 µL) at room temperature was added DMT-MM(OTf) (11.0 mg, 38.8 µmol). The mixture was stirred for 21 h at room temperature. The reaction was quenched with saturated aqueous NaHCO₃, the whole was extracted with EtOAc and washed with saturated aqueous NaHCO₃ and brine, dried over anhydrous Na₂SO₄, filtrated, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (CHCl₃/MeOH= 9/1 to 6/1) to afford **2a** (4.8 mg, 20%) as a white solid. ¹H NMR (CDCl₃/CD₃OD = 3/1) δ 5.61 (m, 1H), 5.38 (m, 1H), 5.26 (m, 2H), 4.19 (m, 1H), 4.09 (m, 1H), 4.01 (m, 1H), 3.95 (t, *J* = 5.3 Hz, 2H), 3.79 (m, 1H), 3.64 (m, 1H), 3.49 (m, 1H), 3.26 (m, 2H), 3.20 (m, 1H), 2.09 (m, 2H), 1.93 (m, 4H), 1.50 (m, 2H), 1.18 (m, 44H), 0.80 (m, 6H); ¹³C–NMR (CDCl₃/CD₃OD = 3/1) δ 173.6, 133.3, 128.8, 128.6, 128.2,102.0, 75.3, 72.5, 71.0, 69.0, 67.5, 60.3, 54.6, 52.2, 35.3, 31.3, 30.8, 28.6 (6C), 28.4 (4C), 28.3 (4C), 28.2 (3C), 28.1, 26.1, 24.9, 21.6, 12.8 (2C); HRMS (ESI-QTOF) calcd for C₄₂H₇₉NO₈: [M+Na]⁺, 748.5698; found: 748.5698

Compound 3a: To a stirring solution of compound **13α** (16.9 mg, 34.7 µmol) and oleic acid (14.7 mg, 52.0 µmol) in EtOH (325 µL) and CH₂Cl₂ (110 µL) at room temperature was added DMT-MM(OTf) (30.4 mg, 78.0 µmol). The mixture was stirred for 13 h at room temperature. The reaction was quenched with saturated aqueous NaHCO₃, the whole was extracted with EtOAc and washed with saturated aqueous NaHCO₃ and brine, dried over anhydrous Na₂SO₄, filtrated, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (CHCl₃/MeOH= 7/1 to 5/1) to afford **3a** (11.9 mg, 47%) as a white solid. ¹H NMR (CDCl₃/CD₃OD = 10/1) δ 5.54–5.47 (m, 1H), 5.23 (dd, *J* = 15.4, 6.8 Hz, 1H), 5.15–5.11 (m, 2H), 4.66 (d, *J* = 3.6 Hz, 1H), 4.29–4.19 (m, 3H), 3.89–3.73 (3, 11H), 3.62–3.40 (m, 6H), 2.05–1.96 (m, 2H), 1.79 (d, *J* = 5.4 Hz, 6H), 1.38 (s, 2H), 1.09–0.97 (m, 44H), 0.68–0.64 (m, 6H); ¹³C–NMR (CDCl₃/CD₃OD = 10/1) δ 174.6, 133.9, 129.8, 129.5, 129.0, 99.8, 77.6, 77.2, 76.9, 72.1, 70.1, 69.6, 68.9, 67.4, 61.6, 55.6, 53.6, 49.2, 48.9, 48.7, 48.5, 48.3, 48.1, 47.9, 36.3, 32.2, 31.7, 29.5, 29.4 (5C), 29.3 (2C), 29.2 (4C), 29.1 (4C), 27.0 (4C), 25.8, 22.5, 13.7 (2C); HRMS (ESI-QTOF) calcd for C₄₂H₇₉NO₈: [M+Na]⁺, 748.5698; found: 748.5698.

Compound 1b: To a stirring solution of compound **12** β (11.4 mg, 24.7 µmol) and stearic acid (7.0 mg, 24.7 µmol) in THF (310 µL) at room temperature was added HATU (9.4 mg, 24.7 µmol), Et₃N (10.9 µL, 49.4 µmol) and NMM (5.4 µL, 49.4 µmol). The mixture was stirred at room temperature for 21 h, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (CHCl₃/MeOH= 7/1 to 5/1) to afford **1b** (4.2 mg, 23%) as a white solid. ¹H NMR (CDCl₃/CD₃OD = 3/1) δ 5.68–5.61 (m, 1H), 5.36 (dd, *J* = 15.4, 7.2 Hz, 1H), 4.74 (d, *J* = 3.6 Hz, 1H), 4.01 (t, *J* = 7.0 Hz, 1H), 3.88 (dd, *J* = 6.8, 3.6 Hz, 1H), 3.72–3.63 (m, 4H), 3.58 (dd, *J* = 12.7, 5.9 Hz, 1H), 3.46 (t, *J* = 4.3 Hz, 1H), 3.38–3.35 (m, 1H), 3.29 (t, *J* = 9.3 Hz, 1H), 2.12–2.07 (m, 2H), 1.94 (tt, *J* = 29.2, 8.9 Hz, 2H), 1.51 (m, 2H), 1.18 (m, 50H), 0.81–0.78 (m, 6H); ¹³C–NMR (CDCl₃/CD₃OD = 3/1) δ 174.2, 133.6, 128.7, 98.8, 73.2, 71.6, 71.5, 71.2, 69.6, 66.5, 60.8, 53.1, 35.8 (2C), 31.8 (2C), 31.3 (2C), 29.1 (5C), 29.0 (3C), 28.9 (3C), 28.8 (3C), 28.7 (4C), 25.4 (2C), 22.0 (2C), 13.2 (2C); HRMS (ESI-QTOF) calcd for C₄2H₈₁NO₈: [M+Na]⁺, 750.5854; found: 750.5862.

Compound 2b: To a stirring solution of compound **12β** (10.4 mg, 22.5 µmol) and stearic acid (6.4 mg, 22.5 µmol) in THF (280 µL) at room temperature was added HATU (8.56 mg, 22.5 µmol), Et₃N (10.0 µL, 45.1 µmol) and NMM (5.0 µL, 45.1 µmol). The mixture was stirred at room temperature for 21 h, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (CHCl₃/MeOH= 7/1 to 5/1) to afford **2b** (6.5 mg, 40%) as a white solid. ¹H NMR (CDCl₃/CD₃OD = 3/1) δ 5.61 (t, *J* = 7.8 Hz, 1H), 5.37 (dd, *J* = 15.1, 7.3 Hz, 1H), 4.18 (d, *J* = 7.8 Hz, 1H), 4.08 (t, *J* = 5.0 Hz, 1H), 4.01 (t, *J* = 7.5 Hz, 1H), 3.92 (m, 1H), 3.78 (d, *J* = 9.6 Hz, 1H), 3.63 (dd, *J* = 11.9, 5.0 Hz, 1H), 3.49 (d, *J* = 7.3 Hz, 1H), 3.35–3.26 (m, 2H), 3.18 (t, *J* = 8.5 Hz, 2H), 2.09 (t, *J* = 7.5 Hz, 2H), 1.92 (t, *J* = 11.0 Hz, 2H), 1.50 (m, 2H), 1.18 (m, 50H), 0.80 (t, *J* = 6.6 Hz, 6H); ¹³C–NMR (CDCl₃/CD₃OD = 3/1) δ 174.3, 134.0, 128.8, 102.7, 76.0, 75.8, 73.2, 71.6, 69.7, 68.2, 61.0, 52.9, 36.0 (2C), 31.9 (2C), 31.5 (2C), 29.3 (2C), 29.2 (3C), 29.1 (6C), 29.0 (3C), 28.9 (2C), 28.8 (2C), 25.5 (2C), 22.2 (2C), 13.4 (2C); HRMS (ESI-QTOF) calcd for C₄₂H₈₁NO₈: [M+Na]⁺, 750.5854; found: 750.5861.

Compound 3b: To a stirring solution of compound **13α** (15.6 mg, 32.0 µmol) and stearic acid (13.7 mg, 48.0 µmol) in THF (400 µL) at room temperature was added HATU (18.2 mg, 48.0 µmol), Et₃N (14.1 µL, 64.0 µmol) and NMM (7.0 µL, 64.0 µmol). The mixture was stirred at room temperature for 20 h, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (CHCl₃/MeOH= 7/1 to 5/1) to afford **3b** (10.1 mg, 43%) as a white solid. ¹H NMR (CDCl₃/CD₃OD = 10/1) δ 5.71–5.62 (m, 1H), 5.38 (dd, *J* = 15.4, 6.8 Hz, 1H), 4.86–4.80 (m, 1H), 4.09–4.00 (m, 2H), 3.95–3.80 (m, 3H), 3.77–3.57 (m, 5H), 2.14–2.07 (m, 2H), 2.02–1.90 (m, 2H), 1.53 (s, 2H), 1.29–1.08 (m, 50H), 0.87–0.80 (m, 6H); ¹³C NMR (CDCl₃/CD₃OD = 10/1) δ 174.7, 134.0, 128.9, 99.8, 77.5, 77.2, 76.9, 72.2, 70.5, 70.1, 69.6, 68.9, 67.5, 61.6, 53.6, 49.3, 49.1, 48.8, 48.6, 48.4, 48.2, 48.0, 36.4, 32.3 (3C), 31.8 (3C), 29.6 (3C), 29.4 (3C), 29.3 (2C), 29.2 (2C), 29.1 (2C), 25.8

(3C), 22.5 (4C), 13.8 (2C); HRMS (ESI-QTOF) calcd for $C_{42}H_{81}NO_8$: $[M+Na]^+$, 750.5854; found: 750.5862.

Compound 1c: To a stirring solution of compound **12** β (10.8 mg, 23.5 µmol) and nervonic acid (12.9 mg, 35.2 µmol) in EtOH (220 µL) and CH₂Cl₂ (70 µL) at room temperature was added DMT-MM (OTf) (14.4 mg, 37.6 µmol). The mixture was stirred for 16 h at room temperature. The reaction was quenched with saturated aqueous NaHCO₃, the whole was extracted with EtOAc and washed with saturated aqueous NaHCO₃ and brine, dried over anhydrous Na₂SO₄, filtrated, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (CHCl₃/MeOH= 7/1 to 5/1) to afford **1c** (4.2 mg, 22%) as a white solid. ¹H NMR (CDCl₃/CD₃OD = 3/1) δ 5.74–5.67 (m, 1H), 5.43 (dd, *J* = 15.4, 7.5 Hz, 1H), 5.33 (t, *J* = 4.5 Hz, 2H), 4.82 (d, *J* = 3.6 Hz, 1H), 4.08 (t, *J* = 8.1 Hz, 1H), 3.95–3.92 (m, 1H), 3.77 (t, *J* = 5.4 Hz, 1H), 3.67–3.62 (m, 2H), 3.53 (t, *J* = 2.6 Hz, 1H), 3.40 (dd, *J* = 9.6, 3.8 Hz, 1H), 3.29 (dt, *J* = 8.3, 4.3 Hz, 3H), 2.19–2.15 (m, 2H), 2.02 (d, *J* = 5.2 Hz, 6H), 1.57 (m, 2H), 1.28 (m, 54H), 0.89 (dd, *J* = 8.5, 4.7 Hz, 6H); ¹³C NMR (CDCl₃/CD₃OD = 3/1) δ 174.3, 133.8 (2C), 129.4, 128.8, 99.0, 73.4, 71.7, 71.6, 71.5, 69.8, 66.8, 61.0, 53.2, 36.0, 32.0, 31.5 (4C), 29.3 (6C), 29.2 (3C), 29.1 (6C), 29.0 (2C), 28.9 (6C), 26.7, 25.5, 22.2, 13.5 (2C); HRMS (ESI-QTOF) calcd for C4₈H9₁NO₈: [M+Na]⁺, 832.6637; found: 832.6642.

Compound 2c: To a stirring solution of compound **12β** (10.6 mg, 23.0 µmol) and nervonic acid (12.7 mg, 34.6 µmol) in EtOH (220 µL) and CH₂Cl₂ (70 µL) at room temperature was added DMT-MM (OTf) (14.4 mg, 36.9 µmol). The mixture was stirred for 16 h at room temperature. The reaction was quenched with saturated aqueous NaHCO₃, the whole was extracted with EtOAc and washed with saturated aqueous NaHCO₃ and brine, dried over anhydrous Na₂SO₄, filtrated, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (CHCl₃/MeOH= 7/1 to 5/1) to afford **2c** (6.6 mg, 35%) as a white solid. ¹H NMR (CDCl₃/CD₃OD = 3/1) δ 5.65–5.57 (m, 1H), 5.36 (dd, *J* = 15.3, 7.6 Hz, 1H), 5.30–5.22 (m, 2H), 4.18 (d, *J* = 7.9 Hz, 1H), 4.09 (dd, *J* = 10.1, 4.5 Hz, 1H), 4.01 (t, *J* = 7.6 Hz, 1H), 3.91 (dd, *J* = 8.9, 4.6 Hz, 1H), 3.78 (d, *J* = 12.1 Hz, 1H), 3.63 (dd, *J* = 12.0, 5.3 Hz, 1H), 3.49 (t, *J* = 5.0 Hz, 1H), 3.31 (dq, *J* = 22.0, 4.5 Hz, 2H), 3.17 (t, *J* = 8.4 Hz, 2H), 2.08 (t, *J* = 7.6 Hz, 2H), 1.93 (t, *J* = 6.2 Hz, 6H), 1.50 (m, 2H), 0.81 (dd, *J* = 15.6, 9.1 Hz, 6H); ¹³C NMR (CDCl₃/CD₃OD = 3/1) δ 174.3, 133.9, 129.4 (2C), 128.8, 102.6, 76.0, 75.8, 73.2, 71.6, 69.7, 68.1, 61.0, 52.9, 36.0, 31.9, 31.5, 31.4, 29.3 (6C), 29.2 (3C), 29.1 (4C), 29.0 (4C), 28.9 (2C), 28.8 (4C), 26.7, 25.5, 25.5, 22.2, 13.4 (2C); HRMS (ESI-QTOF) calcd for C₄₈H₉₁NO₈: [M+Na]⁺, 832.6637; found: 832.6645.

Compound 3c: To a stirring solution of compound 13α (15.0, 30.8 µmol) and nervonic acid (16.9 mg, 46.1 µmol) in EtOH (290 µL) and CH₂Cl₂(100 µL) at room temperature was added DMT-MM (OTf) (27.0 mg, 69.2 µmol). The mixture was stirred for 24 h at room temperature. The reaction was

quenched with saturated aqueous NaHCO₃, the whole was extracted with EtOAc and washed with saturated aqueous NaHCO₃ and brine, dried over anhydrous Na₂SO₄, filtrated, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (CHCl₃/MeOH= 7/1 to 5/1) to afford **3c** (11.2 mg, 45%) as a white solid. ¹H NMR (CDCl₃/CD₃OD = 10/1) δ 5.68–5.61 (m, 1H), 5.37 (dd, *J* = 15.3, 7.0 Hz, 1H), 5.28–5.23 (m, 2H), 4.80 (d, *J* = 3.6 Hz, 1H), 4.43–4.32 (m, 3H), 3.89–3.86 (m, 1H), 3.76–3.63 (m, 7H), 2.11 (t, *J* = 7.6 Hz, 2H), 1.96–1.92 (m, 6H), 1.51 (d, *J* = 6.7 Hz, 2H), 1.35–1.19 (m, 50H), 0.80 (t, *J* = 6.7 Hz, 6H); ¹³C NMR (CDCl₃/CD₃OD = 10/1) δ 174.6, 134.0 (2C), 129.7, 129.0, 99.7, 77.6, 77.2, 76.9, 72.0, 70.5, 70.1, 69.6, 68.9, 67.4, 61.6, 53.6, 49.1, 48.9, 48.7, 48.5, 48.3, 48.1, 47.8, 36.3 (3C), 32.2 (3C), 31.7 (3C), 29.5 (3C), 29.4 (3C), 29.3 (3C), 29.2 (3C), 29.1 (3C), 27.0 (3C), 25.8 (3C), 22.5 (3C), 13.7 (2C); HRMS (ESI-QTOF) calcd for C₄₈H₉₁NO₈: [M+Na]⁺, 832.6637; found: 832.6642.

Compound 1d: To a stirring solution of compound **12** α (15.5 mg, 33.6 µmol) and lignoceric acid (12.4 mg, 33.6 µmol) in THF (420 µL) at room temperature was added HATU (12.8 mg, 33.6 µmol), Et₃N (14.8 µL, 67.2 µmol) and NMM (7.4 µL, 67.2 µmol). The mixture was stirred at room temperature for 21 h, and then concentrated under reduced pressure to afford **1d** (6.3 mg, 23%) as a white solid. ¹H NMR (CDCl₃/CD₃OD = 3/1) δ 5.64 (dd, *J* = 14.1, 7.3 Hz, 1H), 5.36 (dd, *J* = 15.4, 7.1 Hz, 1H), 4.74 (d, *J* = 3.9 Hz, 1H), 4.03 (dd, *J* = 13.9, 7.1 Hz, 1H), 3.89 (m, 1H), 3.70 (t, *J* = 5.1 Hz, 2H), 3.64 (d, *J* = 8.8 Hz, 2H), 3.58 (t, *J* = 9.3 Hz, 1H), 3.46 (m, 1H), 3.37 (dd, *J* = 9.5, 3.7 Hz, 1H), 3.30 (t, *J* = 9.3 Hz, 1H), 2.09 (dt, *J* = 18.7, 7.0 Hz, 2H), 1.95 (d, *J* = 7.3 Hz, 2H), 1.51 (m, 2H), 1.30–1.04 (m, 62H), 0.80 (t, *J* = 6.8 Hz, 6H); ¹³C NMR (CDCl₃/CD₃OD = 3/1) δ 174.3, 133.6, 128.8, 98.9, 73.3, 71.7, 71.6, 71.3, 69.7, 66.7, 60.9, 53.2, 35.9 (2C), 31.9 (2C), 31.4 (2C), 29.2 (6C), 29.1 (3C), 29.0 (6C), 28.9 (3C), 28.8 (6C), 25.5 (2C), 22.2 (2C), 13.4 (2C); HRMS (ESI-QTOF) calcd for C₄₂H₈₁NO₈: [M+Na]⁺, 834.6793; found: 834.6802.

Compound 2d: To a stirring solution of compound **12** β (10.2 mg, 22.1 µmol) and lignoceric acid (8.10 mg, 22.1 µmol) in THF (280 µL) at room temperature was added HATU (8.4 mg, 22.1 µmol), Et₃N (9.7 µL, 44.2 µmol) and NMM (4.8 µL, 44.2 µmol). The mixture was stirred at room temperature for 20 h, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (CHCl₃/MeOH= 7/1 to 5/1) to afford **2d** (7.9 mg, 44%) as a white solid. ¹H NMR (CDCl₃/CD₃OD = 3/1) δ 5.60 (d, *J* = 15.4 Hz, 1H), 5.36 (dd, *J* = 15.4, 7.7 Hz, 1H), 4.18 (d, *J* = 7.7 Hz, 1H), 4.10 (dd, *J* = 10.0, 4.5 Hz, 1H), 4.00 (t, *J* = 7.7 Hz, 1H), 3.91 (m, 1H), 3.78 (t, *J* = 5.9 Hz, 1H), 3.64–3.57 (m, 1H), 3.55–3.47 (m, 1H), 3.29 (t, *J* = 3.2 Hz, 1H), 3.25–3.24 (m, 2H), 3.17 (dd, *J* = 10.2, 6.6 Hz, 1H), 2.21 (dd, *J* = 16.5, 9.3 Hz, 2H), 2.08 (t, *J* = 7.7 Hz, 2H), 1.93 (d, *J* = 6.8 Hz, 2H), 1.51 (d, *J* = 7.2 Hz, 2H), 1.33–1.09 (m, 58H), 0.79 (t, *J* = 6.8 Hz, 6H); ¹³C NMR (CDCl₃/CD₃OD = 3/1) δ 174.3, 134.0, 128.8, 102.6, 76.0, 75.8, 73.2, 71.7, 69.7, 68.2, 61.0, 52.0, 36.1, 34.2 (2C), 32.0

(2C), 31.5 (2C), 29.3 (4C), 29.2 (6C), 29.1 (2C), 29.0 (2C), 28.9 (4C), 28.8 (4C), 25.6 (2C), 24.7 (2C), 22.2, 13.5 (2C); HRMS (ESI-QTOF) calcd for C₄₂H₈₁NO₈: [M+Na]⁺, 834.6793; found: 834.6787.

Compound 3d: To a stirring solution of compound **13** α (15.1 mg, 31.0 µmol) and lignoceric acid (17 .1mg, 46.4 µmol) in THF (390 µL) at room temperature was added HATU (17.7 mg, 46.4 µmol), Et₃N (13.7 µL, 61.9 µmol) and NMM (6.8 µL, 61.9 µmol). The mixture was stirred at room temperature for 24 h, and then concentrated under reduced pressure to afford a residue, which was purified by silica gel column chromatography (CHCl₃/MeOH= 7/1 to 5/1) to afford **3d** (4.8 mg, 19%) as a white solid. ¹H NMR (CDCl₃/CD₃OD = 10/1) δ 5.67–5.60 (m, 1H), 5.35 (dd, *J* = 15.3, 7.0 Hz, 1H), 4.78 (d, *J* = 3.8 Hz, 1H), 4.04–3.97 (m, 1H), 3.92–3.79 (m, 3H), 3.73–3.61 (m, 6H), 2.12–2.04 (m, 2H), 1.99–1.87 (m, 2H), 1.50 (s, 2H), 1.25 (m, 65H), 0.79 (t, *J* = 6.8 Hz, 6H); ¹³C NMR (CDCl₃/CD₃OD = 10/1) δ 174.3, 133.6, 128.8, 98.9, 73.3, 71.7, 71.6, 71.3, 69.7, 66.7, 60.9, 53.2, 35.9 (2C), 31.9 (2C), 31.4 (2C), 29.2 (6C), 29.1 (3C), 29.0 (6C), 28.9 (3C), 28.8 (6C), 25.5 (2C), 22.2 (2C), 13.4 (2C); HRMS (ESI-QTOF) calcd for C₄₂H₈₁NO₈: [M+Na]⁺, 834.6793; found: 834.6802.

AlphaScreen[®] assay for CD1d-Ligand Binding

The Mouse IgG Detection Kit (Perkin-Elmer Life Sciences) was used to determine CD1dligand interactions. Mouse CD1d:Ig fusion protein (BD Biosciences) in PBS containing 0.005% Tween 20 (final concentration: 5 nM, 10 μ L/well) was mixed with anti-mouse IgG acceptor beads in PBS containing 0.005% Tween 20 (final concentration: 10 μ g/mL, 10 μ L/well). After 60 min, Biotinyl-PE (Avanti) in PBS containing 0.005% Tween 20, 1% DMSO and 1% EtOH (final concentration; 2 mM, 10 μ L/well), and ligands in PBS containing 0.005% Tween 20 and 3% DMSO (final concentration range: 3-10000 nM, 10 mL/well) were added to the plate. After incubation at 37 °C for 23 h, Streptavidin Donor beads in PBS containing 0.005% Tween 20 (final concentration: 10 μ g/mL, 10 μ L/well) was added, and incubated for another 60 min. Samples were measured at 520-620 nm in a Spark[®] 10M microplate reader (TECAN).

APC (Antigen Presenting Cell)-free Assay for CD1d-Ligand Binding

Initially, 96-well microplates (multiwell plate 96F, Sumitomo Bakelite Co., Ltd.) were coated with mouse CD1d:Ig fusion protein (0.25 μ g/well, BD Biosciences) in PBS (100 μ L) at 37 °C for 24 h. After washing with PBS, various concentrations of ligand solution were added and incubated at 37 °C for 24 h. The above ligand solution was prepared by diluting the ligand stock solution in DMSO with PBS containing DMSO and Triton[®]X-100 (the final concentration: 1% DMSO and 0.005% Triton[®]X-100). After washing with PBS, 2E10 NKT hybridoma cells (5.0 × 10⁴ cells/well, Ref.: Nyambayar, D.; Iwabuchi, K.; Hedlund, E.; Murakawa, S.; Shirai, K.; Iwabuchi, C.; Kon, Y.; Miyazaki, Y.; Yanagawa, Y.; Onoe, K. *J. Clin. Exp. Hematop.* **2007**, *47*, 1-8.) were added and cultured

at 37 °C for 48 h. IL-2 in the supernatant was measured by ELISA kit (Biolegend).

Cytokine Secretion Assay Using Mouse Splenocyte

Spleen cell suspension (from C57BL/6*J* mice) was prepared in complete medium (RPMI 1640 media supplemented with 5% fetal bovine serum (FBS), and 1% penicillin-streptomycin) and seeded into 96 well plates (6.0×10^5 cells/well). Ligands were added and incubated at 37 °C for 50 h. IFN- γ , IL-4, IL-10 and IL-17A in the supernatant were measured by ELISA kit (Biolegend).

Cytokine Secretion Assay Using Mouse Splenocyte (inhibition of α-GalCer)

Spleen cell suspension (from C57BL/6*J* mice) was prepared in complete medium (RPMI 1640 media supplemented with 5% fetal bovine serum (FBS), and 1% penicillin-streptomycin) and seeded into 96 well plates (6.0×10^5 cells/well). The mixtures of a-GalCer (100 nM) and ligand were added and incubated at 37 °C for 50 h. IFN- γ in the supernatant were measured by ELISA kit (Biolegend).

Mincle Reporter Assay⁴

The ligands were dissolved in CH₂Cl₂/MeOH (2/1) at 1 mM, were diluted in isopropanol and added to 96-well plates with the indicated concentrations, followed by evaporation of the solvent. The 2B4 NFAT-GFP reporter cells expressing human Mincle/FcR γ were seeded before incubation at 37 °C for 18 h. The activation of NFAT-GFP reporter cells was monitored by flow cytometry.

References

- (a) Meng, X. B.; Yang, L. D.; Li, H.; Li, Q.; Cheng, T. M.; Cai, M. S.; Li, Z. J. *Carbohydr. Res.* 2002, 337, 977-981. (b) Dasgupta, S.; Rajput, V. K.; Roy, B.; Mukhopadhyay, B. J. Carbohydr. *Chem.* 2007, 26, 91-106.
- 2) Strelkov, I. S. et al. J. Org. Chem. 2009, 74, 8669.
- 3) Cairo, C. W.; Sandbhor, M. S.; Key, J. A.; Strelkov, I. S. J. Org. Chem. 2009, 74, 8669.
- 4) Yamasaki, S.; Ishikawa, E.; Sakuma, M.; Hara, H.; Ogata, K.; Saito, T. *Nat. Immunol.* **2008**, *9*, 1179-1188.



















Ы

Allylo OAllyl Allylo Allylo



















∕C₁₃H₂₇









Compound S10 α


Compound $S10\alpha$



Compound S10B







Compound 7α



Compound 7α



Compound 7β



∠C₁₃H₂₇

Compound 7β



∠C₁₃H₂₇

Compound 8α



Compound 8α



C₁₃H₂₇

Compound 9α



Compound 9α



Compound 9β



Compound 9β



Compound 10α



Compound 10α



Compound 10β



Compound 10β



C13H27

Compound 11α



Compound 11α



Compound 11β





Compound 12α



Compound 12α



C13H27

Compound 1a





Compound 1a



Compound 2a



Compound 2a



13H₂₇

Compound 3a



Compound 3a



Compound 1b



C₁₃H₂₇

Compound 1b



Compound 2b



Compound 2b



Compound 3b



Compound 3b



Compound 1c



C₁₃H₂₇
Compound 1c



Compound 2c



Compound 2c



Compound 3c



Compound 3c



13H₂₇

77

Compound 1d





78

Compound 1d



Compound 2d



Compound 2d



Compound 3d



Compound 3d

