

Supporting Information

Carbene-organocatalyzed enantioselective [5+5] annulation of dienols and dienals towards tetrahydroisochromen-1-ones

*Ke Xu[†], Jiayi Zhao[†], Kejun Lin, Tingshun Zhu**

[†]These authors contributed equally to this work.

Table of Contents

I.	General information	3
II.	General procedure for the preparation of substrates.....	3
III.	General procedure for the cycloaddition reactions of penta-substituted cyclohexene	4
IV.	Experimental details for product transformations	4
V.	Details of the Optimization of Reaction Conditions	6
VI.	Some ineffective results	9
VII.	Calculation details.....	10
VIII.	Characterization of the substrates.....	13
IX.	Characterization and SFC Chromatograms of Products.....	15
X.	Absolute configuration determination.....	47
XI.	Copies of ^1H NMR spectra of 1	52
XII.	Copies of ^1H NMR, ^{13}C NMR, ^{19}F NMR spectra of products.....	57
XIII.	References cited in the SI.....	105

I. General information

Commercial reagents were purchased from TCI, J&K, 3A Chemicals, Accela, Macklin, Bidepharm or Adamas and used without further purification. The solvents used in the experiments were all purchased anhydrous solvents and used directly. All reactions were carried out with oven-dried glassware. Analytical thin layer chromatography was performed on 0.20 mm silica gel HSGF-254 plates (Huanghai, China), and visualized under 254 nm UV light. Column chromatography was performed on 200-300 mesh silica gel (General-Reagent, China).

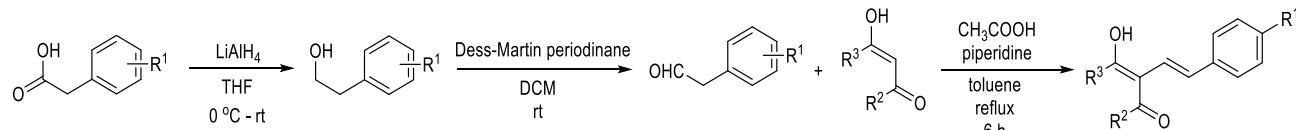
^1H , ^{19}F , ^{31}P and ^{13}C NMR spectra were recorded on a Bruker Ascend 400MHz spectrometer and Bruker Ultrashield 300MHz at ambient temperature. Chemical shifts were recorded in parts per million (ppm, δ) relative to chloroform (for ^1H NMR, $\delta = 7.26$ ppm, singlet; for ^{13}C NMR, $\delta = 77.16$ ppm, triplet). ^1H NMR splitting patterns are designated as singlet (s), doublet (d), triplet (t), quartet (q), dd (doublet of doublets); m (multiplets). All first-order splitting patterns were assigned based on the appearance of the multiplet.

Splitting patterns that could not be easily interpreted are designated as multiplet (m) or broad (br).

High resolution mass spectra of new compounds were recorded on Thermo scientific QExactive MS (ESI). Infrared (IR) spectra were recorded on PerkinElmer Frontier spectrometer and reported in wave numbers (cm^{-1}). The determination of enantiomeric excesses was performed via chiral stationary phases analysis using Supercritical Fluid Chromatography (SFC) by Waters Acuity UltraPerformance Convergence Chromatography (UPCC) (Chiral stationary phases column: Chiralpak OX-3, OD-3 column from Daicel Chiral Technologies and Trefoil CEL-1, CEL-2, AMY-1 column from Waters). Optical rotations were recorded on an Anton Paar MCP-500 polarimeter. Its X-ray diffraction data was collected on Rigaku Oxford diffraction SuperNova using the CuKa radiation at 150 K or 100 K. Electronic circular dichroism (ECD) spectra were recorded on a Circular Dichroism Spectrometer J1700.

II. General procedure for the preparation of substrates

(1) enol substrates **1**.^[1]

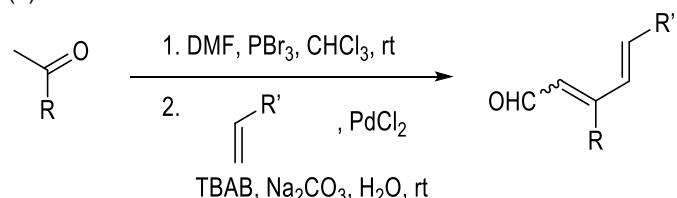


In a 50 mL Schlenk flask were placed substituted phenylacetic acid (5 mmol) and anhydrous THF (20 ml) under N₂ atmosphere. The solution was cooled down to 0 °C. LiAlH₄ (7.5 mmol) was added by portion and the mixture was stirred at 0 °C for 1.5 h. The mixture was quenched saturated aq. NH₄Cl, and the suspension was filtered through a celite pad. Then the filtrate was extracted with EtOAc (20 ml×3). The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. Crude corresponding phenethyl alcohol was used to the next step without further purification.

To a solution of phenethyl alcohol in 20 ml CH₂Cl₂, Dess-Martin periodinane (5 mmol) was added. The mixture was stirred for 2 h at room temperature. After the reaction, the solution was diluted by H₂O and extracted with CH₂Cl₂ (20 ml×3). The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by column chromatography with PE/ethyl acetate to give the corresponding substituted phenylacetaldehyde.

Acetic acid (190 μl , 3.3 mmol) and piperidine (54 μl , 0.55 mmol) were added to solution of ethyl acetoacetate (704 μl , 5.5 mmol) or other substituted diketones in toluene (20 ml) in two-neck round bottom flask. Equipped with DeaneStark apparatus and reflux condenser, corresponding phenylacetaldehyde (5 mmol) was added and the mixture was refluxed for 6 h until no more aldehyde was observed by TLC analysis. After completion of the reaction, the reaction mixture was diluted with by H₂O and extracted with ethyl acetate (30 ml×3). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by column chromatography with PE/ethyl acetate to give the corresponding product **1** as yellow oil or solids.

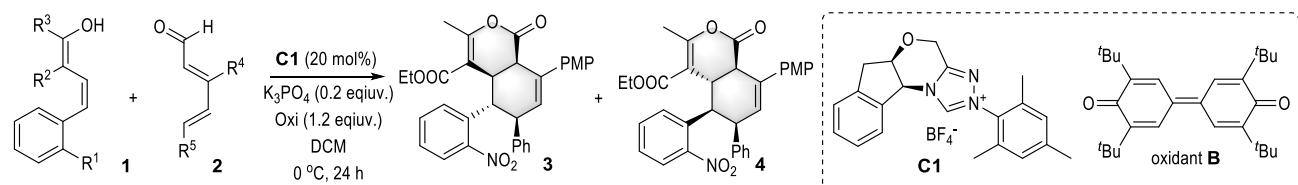
(2) unsaturated enals **2**.^[2]



DMF (2 ml, 26 mmol) in 25 mL dry CHCl₃ was added PBr₃ (2.2 ml, 23 mmol) dropwise at 0 °C, and a white solid precipitated during the addition process. The suspension was stirred at rt for a further 30 min, and acetophenone (10 mmol) in 5 mL dry CHCl₃ was added dropwise at 0 °C. The mixture was allowed to warm to rt and stirred for about 8 h. After complete consumption of acetophenone determined by TLC, the mixture was poured into ice water, and NaHCO₃ was added to neutralize to pH ≈ 6. After extraction with CH₂Cl₂, the organic layer was washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced

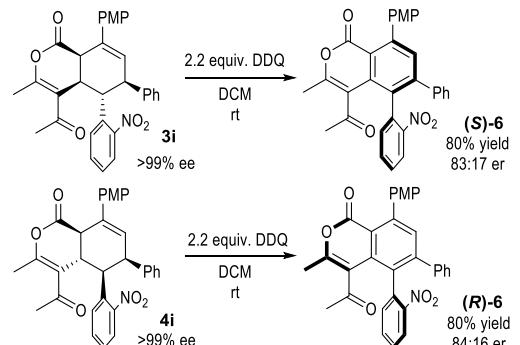
pressure and afford the crude product of β -bromo-enal. To the β -bromo-enal was added alkene (40 mmol), $PdCl_2$ (177 mg, 1 mmol), nBu_4NBr (3.2 g, 10 mmol), Na_2CO_3 (4.2 g, 40 mmol) and H_2O (50 ml). The mixture was stirred for 3 h, and then quenched with saturated NH_4Cl (aq.), extracted with $EtOAc$. The organic layer was washed with brine, dried over anhydrous Na_2SO_4 , concentrated under reduced pressure, and purified by silica gel column chromatography to afford the corresponding $\alpha, \beta, \gamma, \delta$ -diunsaturated enal **2**.

III. General procedure for the cycloaddition reactions of penta-substituted cyclohexene

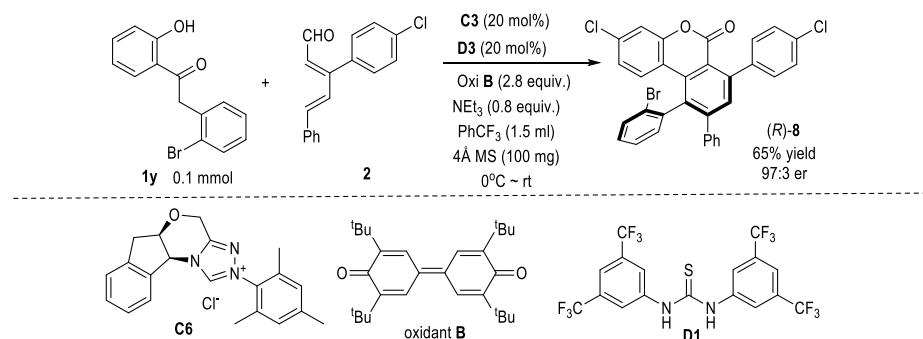


A solution of enol **1** (0.24 mmol), enal **2** (0.2 mmol), **C1** (16.4 mg, 20 mol%), K_3PO_4 (8.4 mg, 20 mol%) and oxidant **B** (82 mg, 0.24 mmol) in 6 mL DCM was stirred at 0 °C for 24 hours. The mixture was concentrated under reduced pressure and purified by silica gel column chromatography to afford the corresponding product **3** and **4**.

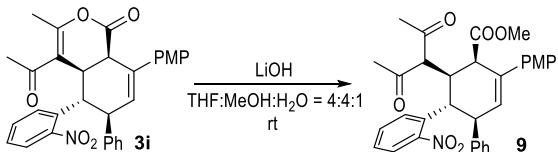
IV. Experimental details for product transformations



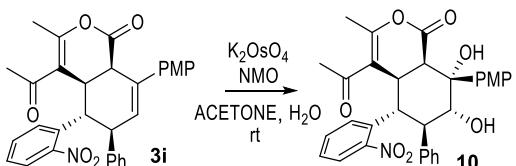
To a solution of **3i/4i** (50.9 mg, 0.1 mmol) in dichloromethane (2 ml) was added 2,3-dichloro-5,6-dicyanobenzoquinone (DDQ, 50 mg, 0.22 mmol). The mixture was stirred at rt for 2 h before being quenched with aqueous saturated $NaHCO_3$. Products were extracted with $EtOAc$ (10 ml \times 3), and extracts were dried ($MgSO_4$) and concentrated in vacuo. The crude product was purified by column chromatography (SiO_2 , hexane/ethyl acetate, by increasing the gradient from 10:1 to 3:1 v/v) to afford 38 mg (80% yield) of **6** as a yellow solid.^[3]



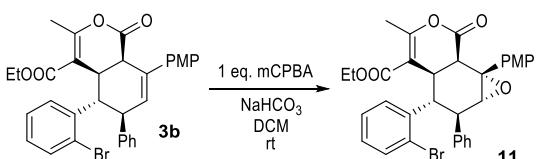
A solution of enol **1y** (0.1 mmol), enal **2** (0.2 mmol), **C3** (7.3 mg, 0.02 mmol), additive **D1** (10 mg, 0.02 mmol), NEt_3 (11 μ L, 0.08 mmol), oxidant **B** (123 mg, 0.3 mmol) and 100 mg 4 Å MS in 1.5 mL $PhCF_3$ was stirred at 0 °C and slowly warm (about 3 h) to rt for 24–72 hours. The mixture was concentrated under reduced pressure and purified by silica gel column chromatography to afford the corresponding product **(R)-8**.



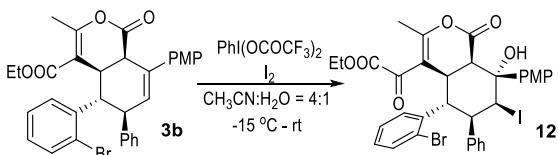
The lactone compound **3i** (0.1 mmol, 50.9mg) and LiOH monohydrate (24 mg, 10 equiv.) were charged to a 25 ml flask. To this mixture was then added MeOH (4 ml), THF (2 mL), and H₂O (1 ml). The reaction was then stirred for 2 h at room temperature and followed the course of the reaction by TLC until completion. The MeOH and THF were then removed in vacuo, and the resulting residue was diluted with H₂O (15 ml), ice, and EtOAc (20 ml). After acidification with 1M HCl, the solution was extracted with EtOAc for three times. The combined organic extract was washed with brine, dried over anhydrous Na₂SO₄, and concentrated in vacuo. The crude product was purified by silica gel column chromatography to afford **9** as a white solid (32 mg, 60% yield).^[4]



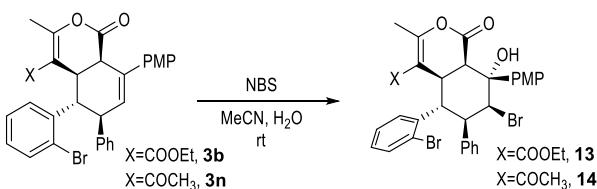
A suspension of **3i** (50.9 mg, 0.1 mmol), potassium osmate(VI) dihydrate (6 mg, 20 mmol) and N-methylmorpholine N-oxide (23 mg, 0.2 mmol) in acetone (0.8 ml) and water (0.2 ml) was vigorously stirred for 24 h at room temperature. The reaction mixture was quenched by the dropwise addition of sat. sodium sulfite (0.5 ml) and stirring continued for 5 mins before the black suspension was extracted with EtOAc (5 ml×3). The combined organic extracts were washed with H₂O and brine, dried over anhydrous Na₂SO₄ and concentrated in vacuo. The crude product was purified by silica gel column chromatography to afford **10** as a white solid (42 mg, 78% yield).^[5]



A solution of **3b** (57.2 mg, 0.1 mmol) in DCM (2 ml) was stirred at room temperature and Na₂CO₃ (10% w/v aq., 0.9 mL, 0.1 mmol) added in one portion. mCPBA (77% w/w, 23 mg, 0.1 mmol) was then added cautiously. Vigorous stirring was continued for 24 h and the reaction mixture subsequently diluted with DCM (10 ml). The aqueous and organic phases were separated and the organic layer washed with 2 ml sat. sodium thiosulfate, H₂O and brine before being dried over anhydrous Na₂SO₄ and concentrated in vacuo. The crude product was purified by **11** as a white solid (35 mg, 60% yield).^[5]



BTI (1.0 equiv.) was added at -15°C to a solution of **3b** (57.2 mg, 0.1 mmol) and I₂ (0.6 equiv.) in a 4:1 mixture of CH₃CN and H₂O and the vessel was allowed to reach room temperature. The red-brown colour of the solution disappears in a few minutes, and continually stirred for 2 h at room temperature. The reaction mixture was quenched by the dropwise addition of sat. sodium sulfite (0.5 ml) and stirring continued for 5 mins and then extracted with EtOAc (5 ml×3). The combined organic extracts were washed with H₂O and brine, dried over anhydrous Na₂SO₄ and concentrated in vacuo. The crude product was purified by silica gel column chromatography to afford **12** as a white solid (65 mg, 88% yield).^[6]



A solution of **3b/3n** (0.1 mmol), N-bromosuccinimide (0.1 mmol, 1 equiv), H₂O (5 mmol, 50 equiv) in MeCN (2 ml) was stirred at room temperature for 12 h and then extracted with EtOAc (5 ml×3). The combined organic extracts were washed with H₂O and brine, dried over anhydrous Na₂SO₄ and concentrated in vacuo. The crude product was purified silica gel column chromatography to afford **13** as a yellow solid (38 mg, 57% yield) or **14** as a yellow solid (54 mg, 86% yield).^[7]

V. Details of the Optimization of Reaction Conditions

Table S1. Screening of catalysts. [a][b][c]

The reaction scheme shows the condensation of aldehyde **2a** (bearing a PMP group) and alcohol **1a** (bearing an O₂N group) in THF at room temperature for 24 h. The reaction is catalyzed by NHC (20 mol%), Oxi B (1.5 equiv.), and Cs₂CO₃ (0.5 equiv.). Three products are formed: **3a**, **4a**, and **5a**. **3a** is a near-racemic mixture, while **4a** and **5a** show significant enantioselectivity.

Below the reaction scheme, the structures of the catalysts are shown:

- C1**: A tricyclic NHC catalyst with a BF₄⁻ counterion.
- C2**: A tricyclic NHC catalyst with a Cl⁻ counterion.
- C3**: A tricyclic NHC catalyst with a BF₄⁻ counterion.
- C4**: A tricyclic NHC catalyst with a Br⁻ counterion.
- C5**: A tricyclic NHC catalyst with a BF₄⁻ counterion.
- Oxidant B**: 2,6-di*t*Bu-4,4,4,4-tetraoxo-2,3-dihydro-1H-1,3-benzodioxole.

Table S1 Data:

entry	NHC catalyst	yield 3a /[% ^b]	ee 3a ^[c]	yield 4a /[% ^b]	ee 4a ^[c]	yield 5a /[% ^b]	ee 5a ^[c]
1	C5	74	-	n.d.	-	15	-
2	C1	54	17	n.d.	-	27	78
3	C2	46	-3	n.d.	-	31	50
4	C3	57	5	n.d.	-	29	80
5	C4	54	26	n.d.	-	29	80

[a] All reactions were performed by using **1a** (0.1 mmol), **2a** (0.15 mmol), catalyst (20 mol%), base (50 mol%), oxidant **B** (0.15 mmol) and solvent (2 mL) at rt for 24 h. [b] Isolated yield. [c] Determined by chiral-phase stationary supercritical fluid chromatography (SFC).

Preliminary screening gave product **5a** with high enantioselectivity and product **3a** with near racemic.

Table S2. Screening of solvents.

The reaction scheme shows the condensation of aldehyde **2a** (bearing a PMP group) and alcohol **1a** (bearing an O₂N group) in THF at room temperature for 24 h. The reaction is catalyzed by **C4** (20 mol%), Oxi B (1.5 equiv.), and the indicated base (0.5 equiv.). Three products are formed: **3a**, **4a**, and **5a**. **3a** is a near-racemic mixture, while **4a** and **5a** show significant enantioselectivity. The structure of catalyst **C4** is highlighted in a dashed box.

Table S2 Data:

entry	base	yield 3a /[% ^b]	ee 3a ^[c]	yield 4a /[% ^b]	ee 4a ^[c]	yield 5a /[% ^b]	ee 5a ^[c]
1	Cs₂CO₃	54	26	n.d.	-	29	80
2	CsF	<5	-	n.d.	-	<5	
3	CH ₃ COOCs	64	-13	n.d.	-	23	79
4	DBU	55	7	n.d.	-	17	79
5	DPEA	<5	-	n.d.	-	<5	
6	NEt ₃	<5	-	n.d.	-	<5	
7	DMAP	36	2	n.d.	-	18	76
8	DABCO	47	-9	n.d.	-	15	71
9	LiOH	<5	-	n.d.	-	<5	
10	NaHCO ₃	<5	-	n.d.	-	<5	
11	K ₂ CO ₃	55	-10	n.d.	-	16	77
12	K ₃ PO ₄	50	-11	n.d.	-	25	78
13	'BuOK	60	-7	n.d.	-	21	77

[a] All reactions were performed by using **1a** (0.1 mmol), **2a** (0.15 mmol), catalyst (20 mol%), base (50 mol%), oxidant **B** (0.15 mmol) and solvent (2 mL) at rt for 24 h. [b] Isolated yield. [c] Determined by chiral-phase stationary supercritical fluid chromatography (SFC).

Base screening shows that inorganic strong bases are better than organic bases in yields and have similar stereoselectivity.

Table S3. Screening of solvent.

entry	solvent	yield 3a/% ^b	ee 3a ^c	yield 4a/% ^b	ee 4a ^c	yield 5a/% ^b	ee 5a ^c
1	THF	54	26	n.d.	-	29	80
2	Dioxane	42	16	n.d.	-	18	80
3	Et ₂ O	30	-30	n.d.	-	24	80
4	CH ₃ CN	46	42	n.d.	-	17	69
5	EA	59	6	n.d.	-	30	80
6	Toluene	35	-4	n.d.	-	15	72
7	PhCF ₃	35	45	n.d.	-	16	82
8	DMF	33	16	n.d.	-	14	59
9	‘BuOH	30	-25	n.d.	-	21	72
10	Acetone	59	26	n.d.	-	15	79
11	DCM	64	56	n.d.	-	28	81
12	DCE	62	57	n.d.	-	28	80
13	CHCl ₃	46	26	n.d.	-	26	71

[a] All reactions were performed by using **1a** (0.1 mmol), **2a** (0.15 mmol), catalyst (20 mol%), base (80 mol%), oxidant **B** (0.15 mmol) and solvent (2 mL) at rt for 24 h. [b] Isolated yield. [c] Determined by chiral-phase stationary supercritical fluid chromatography (SFC).

Solvent screening shows that chlorinated solvent (DCM/DCE) could improve the enantioselectivity of product **3a**.

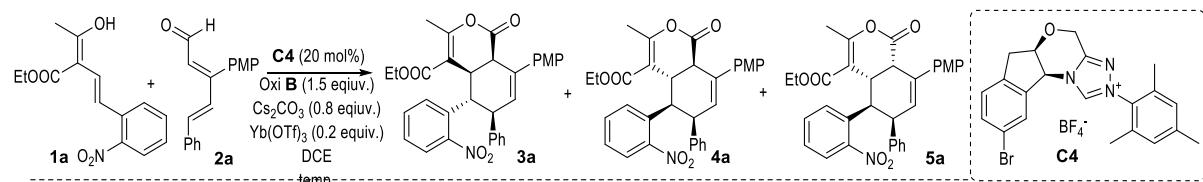
Table S4. Screening of additives.

entry	additive	yield 3a/% ^b	ee 3a ^c	yield 4a/% ^b	ee 4a ^c	yield 5a/% ^b	ee 5a ^c
1	None	62	57	n.d.	-	26	71
2 ^[d]	Sc(OTf) ₃	50	78	<10	98	<10	37
3	‘BuOH	50	40	n.d.	-	29	80
4	1,3-bis(3,5-bis(trifluoromethyl)phenyl)thiourea	50	74	n.d.	-	11	77
5 ^[d]	Yb(OTf)₃	51	79	12	99	n.d.	-
6	LiCl	45	66	n.d.	-	19	82

[a] All reactions were performed by using **1a** (0.1 mmol), **2a** (0.15 mmol), catalyst (20 mol%), base (50 mol%), additive (20 mol%), oxidant **B** (0.15 mmol) and solvent (2 mL) at rt for 24 h. [b] Isolated yield. [c] Determined by chiral-phase stationary supercritical fluid chromatography (SFC). [d] 0.8 equiv. base.

Additive screening shows that Lewis acid could give another product **4a** instead of **5a** and improve the enantioselectivity of **3a**. When use Lewis acid as additive, the use of 0.5 equiv. of base prevents the reaction from proceeding. It is necessary to increase the amount of base to 0.8 equiv. to react normally.

Table S5. Screening of temperature and time.

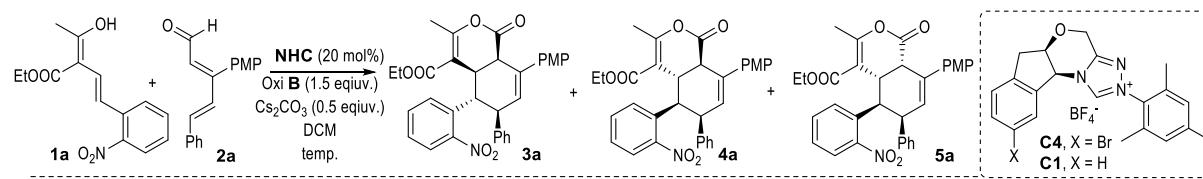


entry	temperature/°C	time/h	yield 3a /% ^[b]	ee 3a ^[c]	yield 4a /% ^[b]	ee 4a ^[c]	yield 5a /% ^[b]	ee 5a ^[c]
1	rt	16	19	91	<10	99	n.d.	-
2	0	8	59	94	9	30	20	99
3	45	4	65	61	<10	66	<10	99
4	60	4	61	54	n.d.	-	<10	70
5	80	4	61	56	n.d.	-	<10	63
6 ^[d]	0	9	65	90	20	99	n.d.	-
7 ^[e]	0	6	52	93	17	99	n.d.	-
8 ^[f]	0	3	39	92	<10	99	n.d.	-

[a] All reactions were performed by using **1a** (0.1 mmol), **2a** (0.15 mmol), catalyst (20 mol%), base (80 mol%), additive (20 mol%), oxidant **B** (0.15 mmol) and DCE (2 mL). [b] Isolated yield. [c] Determined by chiral-phase stationary supercritical fluid chromatography (SFC). [d] 3 mL DCE. [e] 5 mL DCE. [f] 10 mL DCE.

Lower the temperature of reduce the reaction time could obtain product **4a** instead of **5a** and improve the enantioselectivity of **3a**, which means **4a** is an intermediate product.

Table S6. Screening of the amount of base.

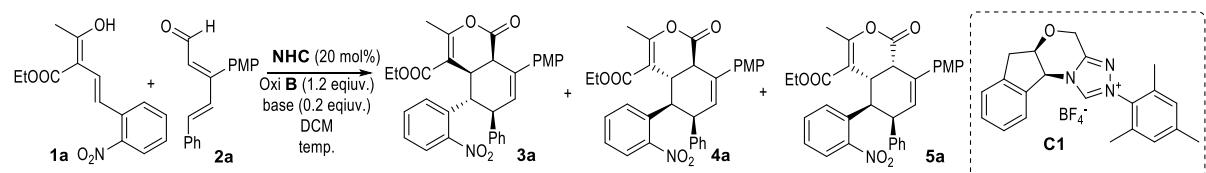


entry	NHC	temperature/°C	time/h	yield 3a /% ^[b]	ee 3a ^[c]	yield 4a /% ^[b]	ee 4a ^[c]	yield 5a /% ^[b]	ee 5a ^[c]
1	C4	rt	5	21	76	<10	99	<5	40
2	C4	0	9	21	91	<10	99	<5	20
3	C4	0	12	28	90	11	99	<5	26
4	C4	-10	10	24	91	8	99	<5	30
5	C1	-10	10	49	96	12	99	<5	51
6 ^[d]	C1	-20	24	49	99	15	99	<5	14
7	C1	-20	24	51	96	9	99	<5	-14
8 ^[e]	C1	-20	24	59	95	11	99	<5	8
9 ^[f]	C1	-20	24	53	96	10	99	10	76
10 ^[g]	C1	-20	24	36	90	n.d.	-	7	80

[a] All reactions were performed by using **1a** (0.1 mmol), **2a** (0.12 mmol), catalyst (20 mol%), base (50 mol%), oxidant **B** (0.15 mmol) and DCM (3 mL). [b] Isolated yield. [c] Determined by chiral-phase stationary supercritical fluid chromatography (SFC). [d] 0.2 equiv. base. [e] 0.8 equiv. base. [f] 1 equiv. base. [g] 2 equiv. base.

Under low temperature conditions, catalyst **C1** could give higher yields than **C4** with similar stereoselectivity control. Even though under -20 °C, the increase of the amount of base reduced the stereoselectivity of product **3a** and transforms **4a** to **5a**.

Table S7. Adjust the dosage ratio of substrates. ^{[a][b][c]}



entry	base	temperature/°C	time/h	yield 3a/% ^[b]	ee 3a ^[c]	yield 4a/% ^[b]	ee 4a ^[c]	yield 5a/% ^[b]	ee 5a ^[c]
1	Cs ₂ CO ₃	-20	48	41	99	11	99	n.d.	-
2	Cs ₂ CO ₃	-5	24	45	98	15	98	n.d.	-
3	Cs ₂ CO ₃	0	24	48	96	17	99	n.d.	-
4	K₃PO₄	0	24	51	96	19	99	n.d.	-

[a] All reactions were performed by using **1a** (0.12 mmol), **2a** (0.1 mmol), catalyst (20 mol%), base (20 mol%), oxidant **B** (0.12 mmol) and DCM (3 mL). [b] Isolated yield. [c] Determined by chiral-phase stationary supercritical fluid chromatography (SFC).

Since **1a** would disappear completely and **2a** would remain a little after most reactions, we changed the proportion of substrates to **2a** with 0.1 mmol and **1a** with 1.2 equivalent. After reducing the amount of base to 0.2 equiv., the reaction enantioselectivity could remain while increasing the reaction temperature to 0 °C. Replace the base from Cs₂CO₃ to K₃PO₄ slightly increases the product yields.

VI. Some ineffective results

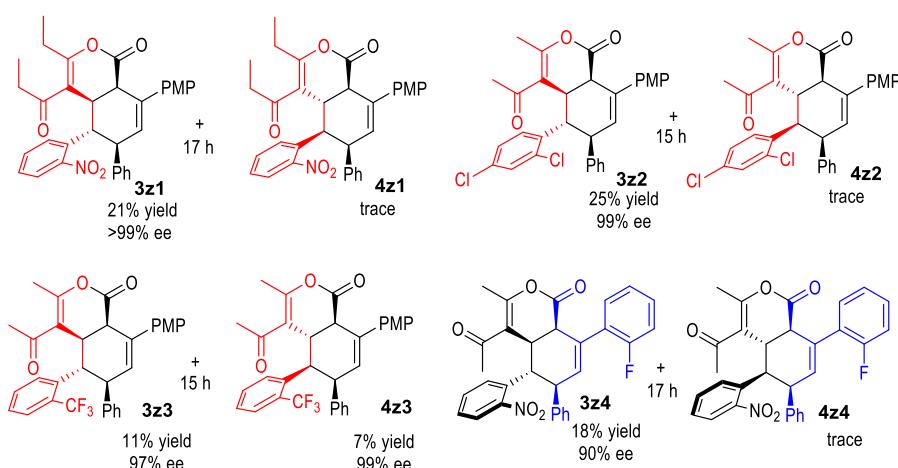


Figure S1. The ineffective results under standard reaction conditions

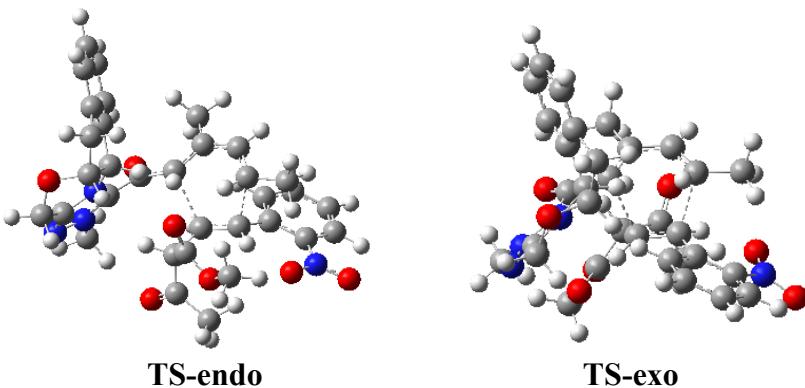
Some ineffective results were also presented in **Figure S1**. Replacing the methyl to a more steric-hindered ethyl resulted in a low yield (**3z1**). Changing ortho-NO₂ to ortho-CF₃ phenyl ring (**3z3**), or 2,4-Cl (**3z2**), also gave ineffective results. These might be due to the weaker electron-withdrawing effect than -NO₂. Introducing ortho-F (**3z4**) led to the addition being blocked so that undesirable yield.

VII. Calculation details

I. Electrophilicity indexes, equilibrium geometries and transition states calculations

All data in this study were calculated with the Gaussian 16 Revision A.03 software package [8] and were optimized at the B3LYP-D3 level of density functional theory (DFT) [9]. The basis set 3-21G was selected for all atoms. Vibrational frequency analysis was computed to ensure the saddle points have only one corresponding imaginary frequency. VMD and Multiwfn were used for drawing the transition state model. [10,11] To take the solvent and temperature effects into account, solvation-corrected single-point energy calculations were computed with M062x/def2TZVPP based on the previous optimized geometries at 273 K. In order to reduce computational complexity, the ethyl group was replaced with methyl, and aromatic groups in diene substrate were simplified with methyl. [12] The relative free Gibbs energies (273 K, in kcal/mol) are used for the following discussion.

II. Cartesian coordinates of optimized structures and transition state



TS-endo: -2023.8092338 Hartree

Number of imaginary frequencies: 1

C	-2.61769444	0.09785071	-0.24600640
C	-1.63597128	-0.41684070	0.63975797
C	-0.99363103	-1.78136194	0.41695425
C	-1.82094844	-2.81769109	1.25052747
O	-1.38517167	-3.07142880	2.35836412
C	-3.11089817	-3.40203432	0.73822385
C	-3.75740147	0.87746111	0.32579378
C	-5.08079797	0.82550153	-0.16995419
C	-6.11054363	1.60644700	0.36208387
C	-5.85705952	2.45479636	1.43255919
C	-4.56297979	2.52934635	1.94929835
C	-3.54055387	1.76163883	1.39671657
H	-1.68237481	-0.14536652	1.68938319
H	-2.91020858	-0.59635206	-1.02358919
C	-0.67667255	-2.07248299	-1.04651188
O	0.36496076	-1.69921797	-1.56297625
O	-1.62983898	-2.74012911	-1.69259034
C	-1.38747059	-3.04031823	-3.09001500
N	-5.48250575	-0.06045153	-1.27950520
O	-4.87132810	-1.12459304	-1.44784411
O	-6.42343576	0.29986117	-1.97734400
C	1.04790554	0.77455792	1.58312125
C	0.25478372	1.05440096	0.40617439
C	-0.34531783	2.32428946	0.16390965
C	-1.27467492	2.42057893	-0.85943801

C	-1.73984408	1.30705494	-1.60856005
O	1.02171691	1.37387578	2.65709015
C	1.98895699	-0.42745778	1.52923806
N	2.12376671	-1.33525174	2.50193342
N	3.06544229	-2.26929645	2.19594846
C	3.52734441	-1.90198928	1.02668572
N	2.89449118	-0.77060714	0.57808187
C	4.56056964	-2.59067809	0.18415404
O	5.11925190	-1.67972665	-0.73565460
C	4.14950246	-1.02607645	-1.55429088
C	3.25553004	-0.08977478	-0.68949177
C	4.92034498	-0.04781874	-2.45391707
C	5.02811421	1.19724004	-1.59848622
C	4.09865717	1.16603169	-0.55076380
C	5.89108324	2.28079924	-1.73554524
C	5.82151712	3.32374969	-0.80732122
C	4.90145703	3.28210389	0.24405943
C	4.02512234	2.20077392	0.37853629
H	-3.65914872	-3.82670326	1.58155438
H	-2.89063310	-4.20007890	0.02031948
H	-3.72154464	-2.66412002	0.21038960
H	-7.10030829	1.52625552	-0.07042791
H	-6.66039139	3.04796291	1.85698188
H	-4.34530392	3.18887858	2.78414105
H	-2.54088111	1.85293302	1.80650609
H	-1.23829155	-2.11635983	-3.65199546
H	-0.50488137	-3.67573897	-3.18594022
H	0.42146201	0.39653034	-0.43660424
H	-1.81066086	3.36112268	-0.97175159
H	-1.00220345	0.54952682	-1.86329880
H	4.09272124	-3.44775946	-0.32770684
H	5.36985153	-2.96783061	0.81338995
H	3.55218426	-1.76824758	-2.10042874
H	2.31915952	0.09652574	-1.22080489
H	4.35647130	0.14097422	-3.37717463
H	5.88591127	-0.47670380	-2.73785648
H	6.61935523	2.31059325	-2.54133418
H	6.49708443	4.16924681	-0.89745952
H	4.86873160	4.09222463	0.96634174
H	3.32747586	2.17738785	1.21100283
H	-2.28224649	-3.56008293	-3.42753884
C	1.35397557	-1.48035876	3.74705561
H	1.09547671	-0.48723770	4.10875451
H	0.45995060	-2.07719454	3.54348061
H	2.00045283	-1.99582098	4.45622874
C	-2.75561598	1.54258435	-2.70344515
H	-2.26417813	2.03282674	-3.55374222
H	-3.19026336	0.60861663	-3.07078125
H	-3.56690931	2.19887122	-2.37293782
C	-0.08922960	3.52254680	1.04887455
H	-0.49176126	3.37860487	2.05656046
H	0.98265479	3.70913963	1.17263719
H	-0.54694745	4.41569166	0.61490186
H	-0.02276300	-1.79742225	0.91136532

TS-exo: -2023.8088667 Hartree

Number of imaginary frequencies: 1

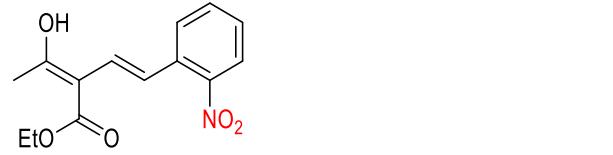
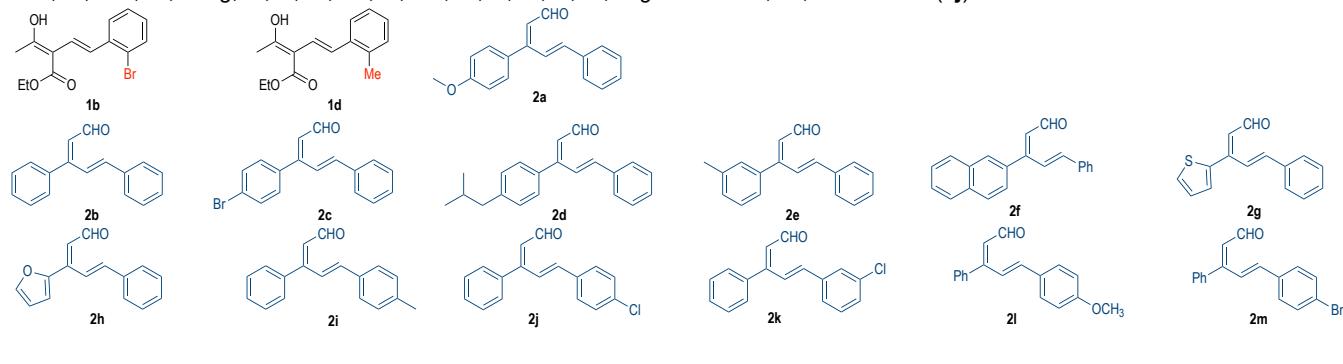
C	-2.68411217	0.55924565	0.62677244
C	-2.21385147	-0.51265244	-0.18955469
C	-2.52104568	-1.91389788	0.18603981
C	-4.10087489	-2.14380474	0.17848250
O	-4.74832117	-1.66534511	1.07558546
C	-4.69330739	-2.93676725	-0.95975195
C	-2.86894692	1.87237872	-0.09532881
C	-4.08532405	2.58162139	-0.17389434
C	-4.21703184	3.75816476	-0.91465218
C	-3.11955365	4.28565992	-1.58488788
C	-1.90116347	3.60886289	-1.53281089
C	-1.79187502	2.42405531	-0.80814236
H	-1.86221897	-0.34211877	-1.20150922
H	-3.54910285	0.26083214	1.21797824
C	-1.79777050	-2.92086054	-0.69197636
O	-1.43743448	-2.69703309	-1.83254051
O	-1.64954577	-4.08639725	-0.05934808
C	-0.94994568	-5.12939939	-0.78068857
N	-5.31915620	2.12381271	0.50124360
O	-5.21331702	1.51821181	1.56940642
O	-6.38497633	2.40007637	-0.03922526
C	1.17057777	-1.41350542	-0.23822929
C	0.34748769	-0.64894594	0.64683892
C	-0.07771267	-1.07532454	1.93978005
C	-1.00566670	-0.30730679	2.62030314
C	-1.59566897	0.90075563	2.09618561
O	1.38080515	-2.63248904	-0.24680891
C	1.91302240	-0.63177018	-1.32088363
N	1.97618592	-0.94986078	-2.61533272
N	2.80863023	-0.11738101	-3.30679332
C	3.26905849	0.70982370	-2.40484023
N	2.74916417	0.42837578	-1.16447918
C	4.18110578	1.88487955	-2.60214946
O	4.82692612	2.20626314	-1.38955343
C	3.93683715	2.43546656	-0.29861945
C	3.18999745	1.11975611	0.07191900
C	4.81485381	2.70287005	0.93317892
C	5.11168243	1.31053992	1.44940784
C	4.20681212	0.37835303	0.92506465
C	6.11684364	0.89692593	2.31875697
C	6.21189737	-0.45859071	2.64686708
C	5.31426305	-1.38637586	2.11095303
C	4.29592282	-0.97434047	1.24553929
H	-5.78173506	-2.87691347	-0.91275075
H	-4.38544927	-3.98636111	-0.87757216
H	-4.32824011	-2.56402933	-1.92317169
H	-5.18555391	4.24192870	-0.95173494
H	-3.21901421	5.20908417	-2.14600237
H	-1.03662988	4.00095594	-2.06070193
H	-0.84160696	1.89712718	-0.80150401
H	-1.42163445	-5.29898708	-1.75072711
H	0.09316126	-4.83631498	-0.91190749
H	0.26545648	0.40430661	0.40596551

H	-1.37243818	-0.66973733	3.57849608
H	-0.88916214	1.52985183	1.55041475
H	3.59505514	2.73683668	-2.98522645
H	4.95348022	1.64494267	-3.33606003
H	3.24379292	3.25324434	-0.54243613
H	2.28603373	1.35341466	0.64336198
H	4.26251430	3.30190243	1.66965447
H	5.70506204	3.27016004	0.64599555
H	6.82707492	1.61176232	2.72506625
H	6.99796054	-0.79533755	3.31625172
H	5.40902314	-2.43813348	2.36370704
H	3.61555579	-1.71043259	0.82796575
H	-1.02805950	-6.01383850	-0.15016846
C	1.34939423	-2.07040340	-3.32524181
H	2.06476930	-2.89426462	-3.37632083
H	0.44910743	-2.38252290	-2.79936779
H	1.11180437	-1.72065042	-4.32970035
C	-2.41986893	1.71340012	3.08183336
H	-1.75444436	2.10669061	3.85981818
H	-2.91754366	2.55940916	2.60478103
H	-3.18603682	1.10157929	3.56614909
C	0.41519352	-2.36975169	2.54580903
H	0.06861307	-3.23709199	1.97655911
H	1.50941990	-2.41476773	2.54814149
H	0.06311623	-2.46425740	3.57669544
H	-2.26636989	-2.07356311	1.24478753

VIII. Characterization of the substrates

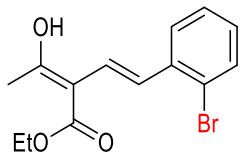
References for the known compounds:

- Guo, Y., Wu, L., Qiu, F. G., *Org. Lett.* 2022, **24**, 8370-8374. (**1b**, **1d**);
- Singha, R., Dhara, S., Ray, J. K., *Tetrahedron Lett.* 2013, **54**, 4841-4843. (**2a**, **2f**);
- Chatterjee, I., Bastida, D., Melchiorre, P., *Adv. Synth. Catal.* 2013, **355**, 3124-3130. (**2c**, **2k**);
- Zhu, T., Mou, C., Li, B., Smetankova, M., Song, B.-A., Chi, Y. R., *J. Am. Chem. Soc.* 2015, **137**, 5658-5661. (**2b**, **2d**, **2g**, **2h**, **2l**, **2m**);
- Xu, K., Li, W., Zhu, S., Zhu, T., *Angew. Chem. Int. Ed.* 2019, **58**, 17625-17630. (**2e**, **2i**);
- Li, Y., Luo, H., Tang, Z., Li, Y., Du, L., Xin, X., Li, S., Li, B., *Org. Lett.* 2021, **23**, 6450-6454. (**2j**).



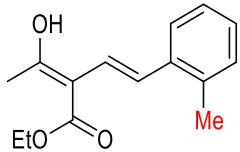
ethyl (E)-3-hydroxy-2-((E)-2-nitrostyryl) but-2-enoate (1a)

¹H NMR (400 MHz, CDCl₃) δ 13.60 (s, 1H), 7.92 (d, *J* = 8.2 Hz, 1H), 7.64 (d, *J* = 7.7 Hz, 1H), 7.56 (t, *J* = 7.5 Hz, 1H), 7.35 (t, *J* = 7.3 Hz, 1H), 7.18 (d, *J* = 16.1 Hz, 1H), 6.76 (d, *J* = 16.1 Hz, 1H), 4.33 (q, *J* = 7.1 Hz, 2H), 2.28 (s, 3H), 1.39 (t, *J* = 7.1 Hz, 3H).



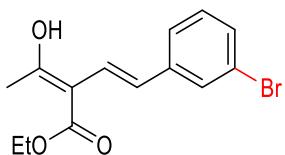
ethyl (E)-2-((E)-2-bromostyryl)-3-hydroxybut-2-enoate (1b)

¹H NMR (400 MHz, CDCl₃) δ 13.55 (s, 1H), 7.57 (m, J = 7.8, 3.5, 1.5 Hz, 2H), 7.32 (m, J = 7.5, 1.3 Hz, 1H), 7.11 (m, J = 7.5, 1.6 Hz, 1H), 7.05 (d, 1H), 6.71 (d, J = 16.2 Hz, 1H), 4.35 (q, J = 7.2 Hz, 2H), 2.29 (s, 3H), 1.42 (t, J = 7.1 Hz, 3H).



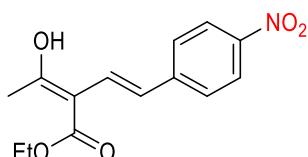
ethyl (E)-3-hydroxy-2-((E)-2-methylstyryl) but-2-enoate (1d)

¹H NMR (400 MHz, CDCl₃) δ 13.39 (s, 1H), 7.52 – 7.46 (m, 1H), 7.24 – 7.14 (m, 3H), 6.88 (d, J = 16.2 Hz, 1H), 6.62 (d, J = 16.1 Hz, 1H), 4.38 – 4.28 (m, 2H), 2.37 (s, J = 2.2 Hz, 3H), 2.25 (s, 3H), 1.38 (t, J = 7.1 Hz, 3H).



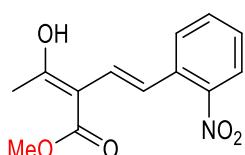
ethyl (E)-2-((E)-3-bromostyryl)-3-hydroxybut-2-enoate (1e)

¹H NMR (400 MHz, CDCl₃) δ 13.44 (s, 1H), 7.53 (t, J = 1.8 Hz, 1H), 7.35 – 7.28 (m, 2H), 7.18 (t, J = 7.8 Hz, 1H), 6.73 (d, J = 16.2 Hz, 1H), 6.55 (d, J = 16.3 Hz, 1H), 4.31 (q, J = 7.1 Hz, 2H), 2.23 (s, 3H), 1.37 (t, J = 7.1 Hz, 3H).



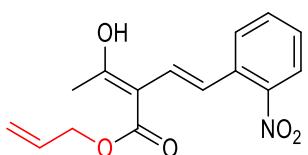
ethyl (E)-3-hydroxy-2-((E)-4-nitrostyryl) but-2-enoate (1f)

¹H NMR (400 MHz, CDCl₃) δ 13.69 (s, 1H), 8.21 (d, J = 8.6 Hz, 2H), 7.52 (d, J = 8.6 Hz, 2H), 6.98 (d, J = 16.3 Hz, 1H), 6.78 (d, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 2.31 (s, 3H), 1.42 (t, J = 7.1 Hz, 3H).



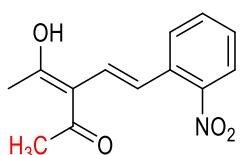
methyl (E)-3-hydroxy-2-((E)-2-nitrostyryl) but-2-enoate (1g)

¹H NMR (400 MHz, CDCl₃) δ 13.54 (s, 1H), 7.58 (dd, J = 6.7, 4.3 Hz, 2H), 7.32 (d, J = 7.5 Hz, 1H), 7.11 (d, J = 7.6 Hz, 1H), 7.06 (d, J = 16.4 Hz, 1H), 6.71 (d, J = 16.2 Hz, 1H), 4.35 (q, J = 7.1 Hz, 2H), 2.30 (s, 3H), 1.42 (t, J = 7.1 Hz, 3H).



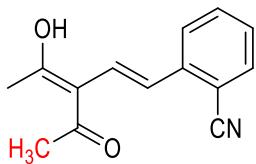
allyl (E)-3-hydroxy-2-((E)-2-nitrostyryl) but-2-enoate (1h)

¹H NMR (400 MHz, CDCl₃) δ 13.43 (s, 1H), 7.93 (d, J = 8.2 Hz, 1H), 7.64 (d, J = 7.6 Hz, 1H), 7.57 (t, J = 7.5 Hz, 1H), 7.36 (t, J = 7.6 Hz, 1H), 7.12 (d, J = 16.1 Hz, 1H), 6.76 (d, J = 16.1 Hz, 1H), 6.08 – 5.87 (m, 1H), 5.37 (d, J = 16.7 Hz, 1H), 5.29 (m, J = 10.4, 1.3 Hz, 1H), 4.77 (d, J = 5.7 Hz, 2H), 2.29 (s, 3H).



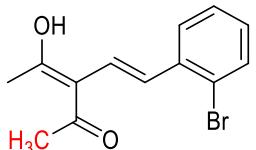
(E)-4-hydroxy-3-((E)-2-nitrostyryl) pent-3-en-2-one (1i)

¹H NMR (400 MHz, CDCl₃) δ 7.99 (d, J = 8.1 Hz, 1H), 7.70 – 7.57 (m, 2H), 7.43 (td, J = 8.4, 6.9, 1.9 Hz, 1H), 6.89 (d, J = 16.0 Hz, 1H), 6.73 (d, J = 16.0 Hz, 1H), 2.28 (s, 6H).



2-((1*E*,3*E*)-3-acetyl-4-hydroxypenta-1,3-dien-1-yl) benzonitrile (1j**)**

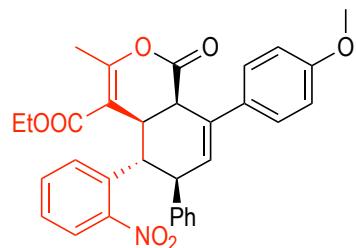
¹H NMR (400 MHz, CDCl₃) δ 7.66 (dd, *J* = 7.8, 2.5 Hz, 2H), 7.58 (t, *J* = 7.7 Hz, 1H), 7.34 (t, *J* = 7.6 Hz, 1H), 6.97 (d, *J* = 16.0 Hz, 1H), 6.77 (d, *J* = 16.1 Hz, 1H), 2.29 (s, 6H).



(E)-3-((E)-2-bromostyryl)-4-hydroxypent-3-en-2-one (1k**)**

¹H NMR (400 MHz, CDCl₃) δ 7.59 (t, *J* = 8.2 Hz, 2H), 7.36 – 7.31 (m, 1H), 7.15 (td, *J* = 7.7, 1.7 Hz, 1H), 6.80 (d, *J* = 16.0 Hz, 1H), 6.69 (d, *J* = 16.0 Hz, 1H), 2.29 (s, 6H).

IX. Characterization and SFC Chromatograms of Products



ethyl (4a*R*,5*S*,6*R*,8a*R*)-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-1-oxo-6-phenyl-4*a*,5,6,8*a*-tetrahydro-1*H*-isochromene-4-carboxylate (**3a**)

24h, light yellow solid; 51% yield, 96% ee;

HRMS (ESI+): calcd. for C₃₂H₂₉NO₇ +Na⁺ [M+Na⁺] 562.1836, found 562.1832;

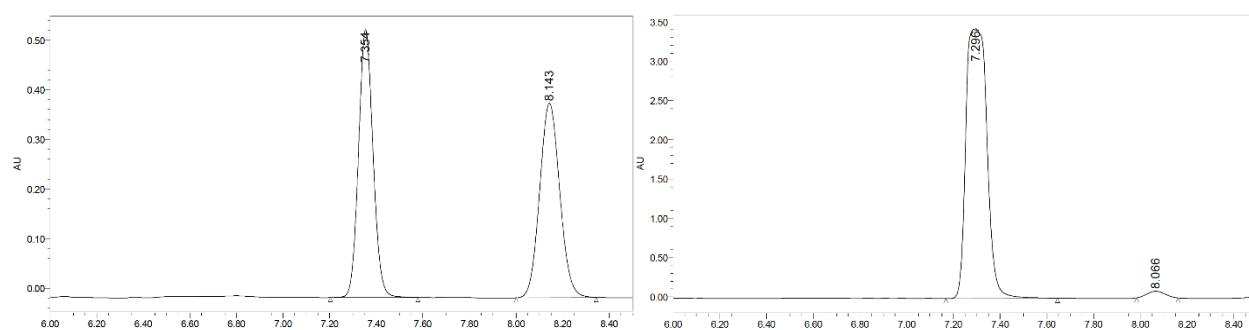
IR: ν_{max} (film, cm⁻¹): 3480, 2931, 1780, 1708, 1515, 1247, 699;

¹H NMR (400 MHz, CDCl₃): δ 7.70 (dd, *J* = 8.0, 1.4 Hz, 1H), 7.54 (td, *J* = 7.6, 1.4 Hz, 1H), 7.45 (dd, *J* = 8.1, 1.3 Hz, 1H), 7.42 – 7.37 (m, 2H), 7.30 – 7.26 (m, 1H), 7.17 – 7.10 (m, 3H), 6.92 – 6.83 (m, 4H), 6.21 (d, *J* = 2.9 Hz, 1H), 3.92 – 3.73 (m, 8H), 3.64 (dd, *J* = 12.0, 10.4 Hz, 1H), 2.24 (s, 3H), 1.11 (t, *J* = 7.1 Hz, 3H);

¹³C NMR (150 MHz, CDCl₃): δ 167.6, 166.1, 160.9, 159.4, 151.8, 141.3, 134.0, 133.4, 132.6, 131.3, 129.9, 128.8, 128.6, 128.1, 127.8, 127.2, 127.1, 123.8, 114.1, 109.6, 60.8, 55.5, 51.4, 44.1, 43.6, 38.5, 18.8, 14.2;

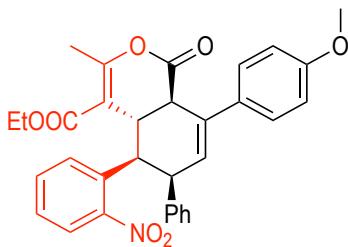
[α]_D²⁵ = -262.3 (c = 0.7 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 50:50 in the first 5 mins and maintaining in 50:50 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ= 254 nm), t_R (major) = 7.3 min, t_R (minor) = 8.1 min.



	RT	Area	% Area	Height	% Height
1	7.354	2342391	49.95	540083	57.95
2	8.143	2347126	50.05	391955	42.05

	RT	Area	% Area	Height	% Height
1	7.296	21426142	97.94	3433064	97.57
2	8.066	451120	2.06	85377	2.43



ethyl (4aS,5R,6R,8aR)-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-1-oxo-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromene-4-carboxylate (4a)

light yellow solid; 19% yield, >99% ee;

HRMS (ESI⁺): calcd. for $C_{32}H_{29}NO_7 + Na^+$ [M+Na⁺] 562.1836, found 562.1829;

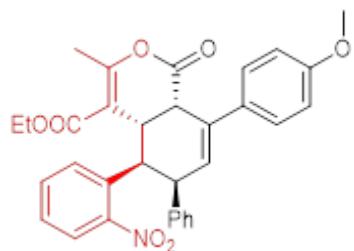
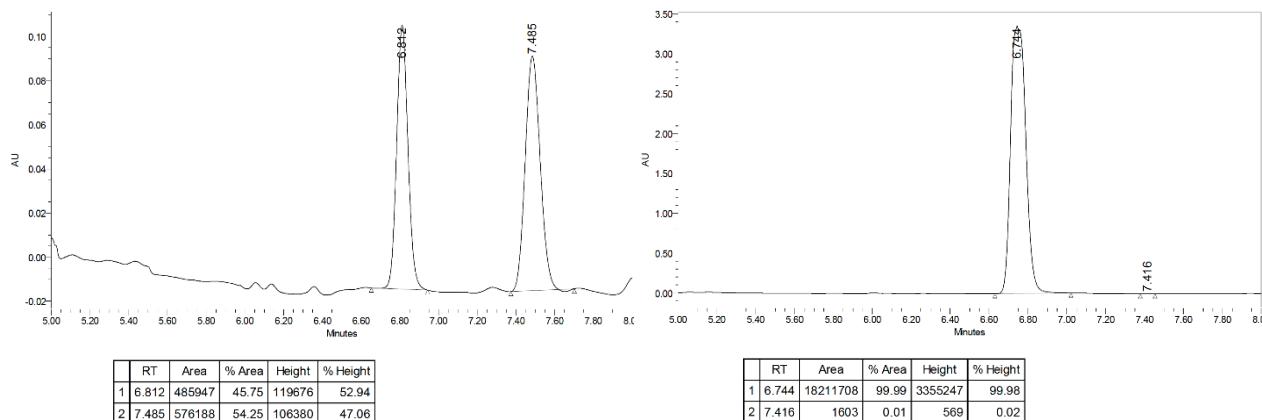
IR: ν_{max} (film, cm⁻¹): 3480, 2930, 1774, 1712, 1514, 1076, 780;

¹H NMR (400 MHz, CDCl₃): δ 7.87 (dd, J = 8.1, 1.4 Hz, 1H), 7.34 – 7.30 (m, 2H), 7.28 – 7.23 (m, 1H), 7.18 – 7.03 (m, 4H), 6.90 (d, J = 8.8 Hz, 2H), 6.77 – 6.63 (m, 2H), 6.31 (dd, J = 5.6, 2.0 Hz, 1H), 6.10 (dd, J = 8.0, 1.4 Hz, 1H), 4.38 – 4.33 (m, 2H), 4.01 – 3.93 (m, 1H), 3.83 (s, 3H), 3.60 – 3.45 (m, 3H), 1.98 (d, J = 1.9 Hz, 3H), 0.95 (t, J = 7.1 Hz, 3H);

¹³C NMR (150 MHz, CDCl₃): δ 167.7, 166.0, 159.1, 154.4, 150.8 138.0, 134.7, 133.4, 132.56, 131.62 130.5, 129.78, 129.5, 128.1, 127.4, 127.1, 126.4, 124.6, 114.9, 114.0, 61.1, 55.4, 46.2, 45.1, 39.7, 32.2, 17.2, 13.9;

$[\alpha]^{25}_D$ = -28.0 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 50:50 in the first 5 mins and maintaining in 50:50 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 6.7 min, t_R (minor) = 7.4 min.



ethyl (4aS,5R,6R,8aS)-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-1-oxo-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromene-4-carboxylate (5a)

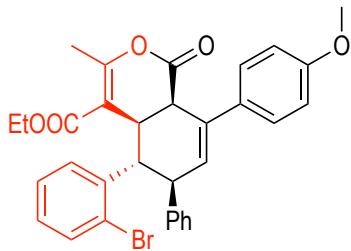
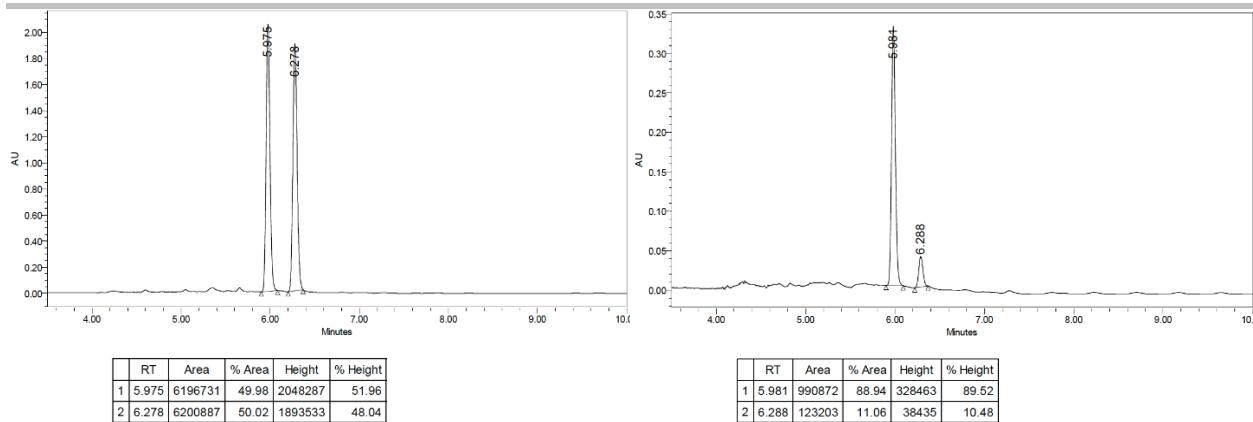
light yellow solid; 78% ee;

¹H NMR (400 MHz, CDCl₃): δ 7.63 (dd, J = 8.1, 1.4 Hz, 1H), 7.49 – 7.43 (m, 2H), 7.21 – 7.11 (m, 4H), 6.95 – 6.90 (m, 2H), 6.89 – 6.84 (m, 3H), 6.31 (d, J = 4.4 Hz, 1H), 6.10 (dd, J = 8.1, 1.3 Hz, 1H), 4.18 (dd, J = 6.3, 4.4 Hz, 1H), 3.99 (d, J = 5.2 Hz, 1H), 3.95 – 3.85 (m, 2H), 3.83 (s, 3H), 3.83 – 3.74 (m, 2H), 2.12 (s, 3H), 1.08 (t, J = 7.1 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 167.3, 166.8, 159.5, 159.1, 151.6, 139.5, 133.3, 133.0, 132.8, 131.6, 130.4, 129.9, 128.2, 127.7, 127.5, 127.4, 127.1, 123.6, 114.2, 110.5, 61.0, 55.5, 46.9, 44.0, 41.9, 32.7, 18.6, 14.1;

$[\alpha]^{25}_D$ = -80.0 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 50:50 in the first 5 mins and maintaining in 50:50 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 6.0 min, t_R (minor) = 6.3 min.



ethyl (4a*R*,5*S*,6*R*,8a*R*)-5-(2-bromophenyl)-8-(4-methoxyphenyl)-3-methyl-1-oxo-6-phenyl-4a,5,6,8a-tetrahydro-1*H*-isochromene-4-carboxylate (3b)

23 h, light yellow solid; 42% yield, 96% ee;

HRMS (ESI⁺): calcd. for C₃₂H₂₉BrO₅ + Na⁺ [M+Na⁺] 595.1091, found 595.1082;

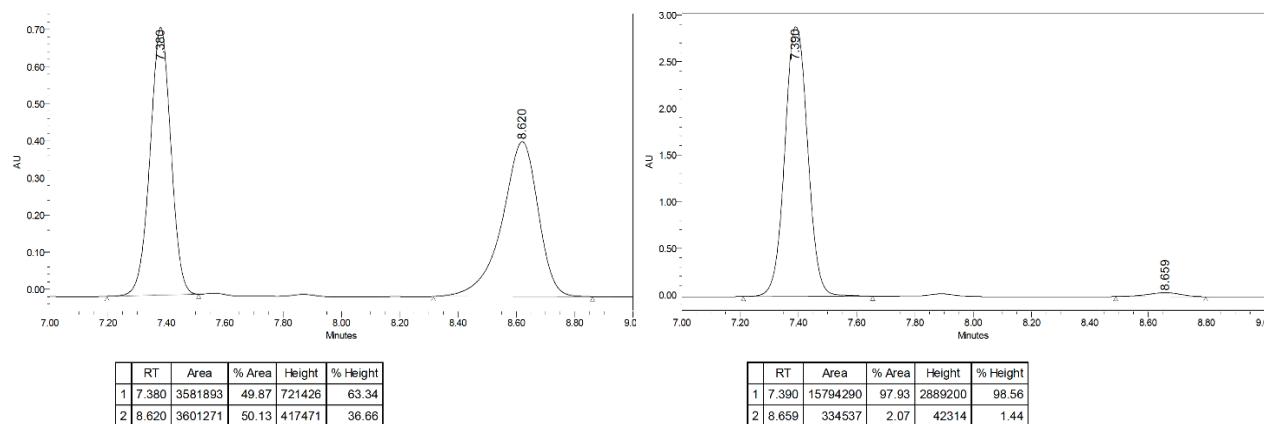
IR: ν_{max} (film, cm⁻¹): 2932, 1777, 1703, 1514, 1249, 1067, 700;

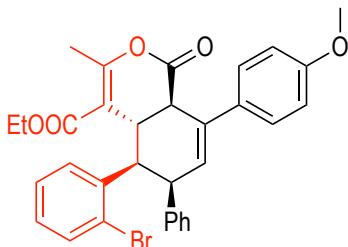
¹H NMR (400 MHz, CDCl₃): δ 7.47 (dd, *J* = 7.9, 1.7 Hz, 1H), 7.44 – 7.38 (m, 2H), 7.32 (dd, *J* = 8.0, 1.3 Hz, 1H), 7.28 – 7.24 (m, 1H), 7.18 – 7.10 (m, 3H), 7.02 – 6.95 (m, 3H), 6.93 – 6.87 (m, 2H), 6.25 (d, *J* = 2.8 Hz, 1H), 3.93 – 3.87 (m, 2H), 3.82 (s, 3H), 3.80 – 3.75 (m, 1H), 3.73 – 3.63 (m, 3H), 2.24 (s, 3H), 1.11 (t, *J* = 7.2 Hz, 3H);

¹³C NMR (150 MHz, CDCl₃): δ 168.3, 165.9, 156.0, 159.4, 141.8, 139.0, 133.1, 132.8, 132.7, 129.4, 129.3, 128.5, 128.4, 128.3, 127.6, 127.1, 126.9, 126.7, 114.1, 110.0, 60.6, 55.5, 50.7, 48.3, 43.6, 39.1, 18.6, 14.2;

$[\alpha]^{25}_D = -110.4$ (*c* = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 50:50 in the first 5 mins and maintaining in 50:50 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 7.4 min, t_R (minor) = 8.6 min.





ethyl (4aS,5R,6R,8aR)-5-(2-bromophenyl)-8-(4-methoxyphenyl)-3-methyl-1-oxo-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromene-4-carboxylate (4b)

light yellow solid; 10% yield, 99% ee;

HRMS (ESI+): calcd. for $C_{32}H_{29}BrO_5 + Na^+$ [M+Na⁺] 595.1091, found 595.1085;

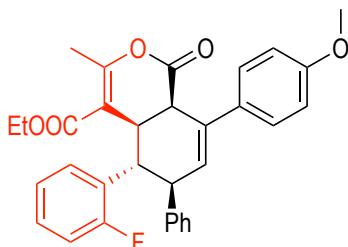
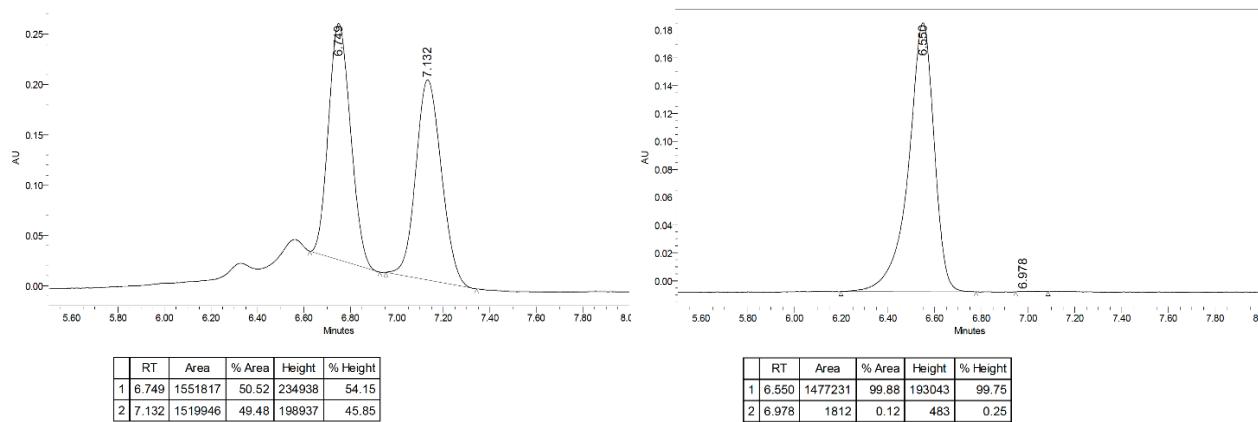
IR: ν_{max} (film, cm⁻¹): 2931, 1781, 1709, 1514, 1244, 1089, 699;

¹H NMR (400 MHz, CDCl₃) δ 7.47 (dd, $J = 7.9, 1.7$ Hz, 1H), 7.44 – 7.39 (m, 2H), 7.32 (dd, $J = 8.1, 1.3$ Hz, 1H), 7.28 – 7.24 (m, 1H), 7.16 – 7.11 (m, 3H), 7.01 – 6.95 (m, 3H), 6.92 – 6.87 (m, 2H), 6.25 (d, $J = 2.8$ Hz, 1H), 3.93 – 3.87 (m, 2H), 3.82 (s, 3H), 3.80 – 3.75 (m, 1H), 3.72 – 3.62 (m, 3H), 2.24 (s, 3H), 1.11 (t, $J = 7.2$ Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 168.3, 166.0, 160.0, 159.4, 141.8, 139.0, 133.1, 132.8, 132.7, 129.5, 129.3, 128.5, 128.4, 128.4, 127.6, 127.1, 126.9, 126.7, 114.1, 110.0, 60.6, 55.5, 50.7, 48.3, 43.6, 39.1, 18.7, 14.2.

$[\alpha]^{25}_D = -132.0$ ($c = 0.5$ in acetone, $\lambda = 589$ nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 80:20 in the first 5 mins and maintaining in 80:20 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, $\lambda = 254$ nm), t_R (major) = 6.5 min, t_R (minor) = 7.0 min.



ethyl (4aR,5S,6R,8aR)-5-(2-fluorophenyl)-8-(4-methoxyphenyl)-3-methyl-1-oxo-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromene-4-carboxylate (3c)

23 h, light yellow solid; 41% yield, 1.2:1 dr, 97% ee;

HRMS (ESI+): calcd. for $C_{32}H_{29}FO_5 + Na^+$ [M+Na⁺] 535.1891, found 535.1886;

IR: ν_{max} (film, cm⁻¹): 2989, 1777, 1704, 1516, 1261, 1066, 706;

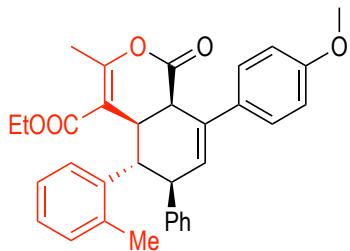
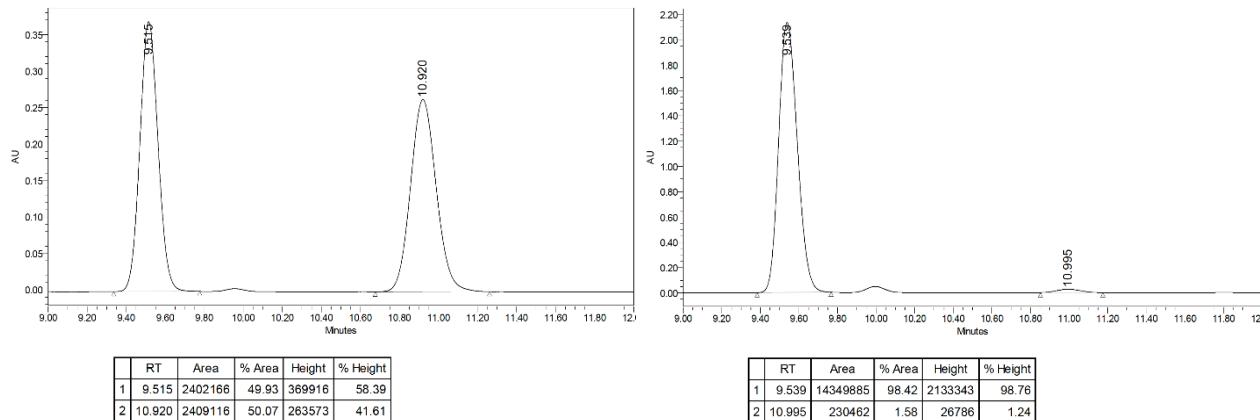
¹H NMR (400 MHz, CDCl₃): δ 7.47 (d, $J = 6.6$ Hz, 1H), 7.44 – 7.38 (m, 5H), 7.13 (dq, $J = 10.5, 5.9, 4.5$ Hz, 10H), 6.97 (s, 5H), 6.93 – 6.88 (m, 5H), 6.80 (d, $J = 8.8$ Hz, 2H), 6.49 (d, $J = 21.1$ Hz, 1H), 6.25 (d, $J = 3.0$ Hz, 2H), 4.24 (d, $J = 10.4$ Hz, 1H), 4.10 – 3.99 (m, 1H), 3.90 (dd, $J = 5.5, 1.2$ Hz, 4H), 3.82 (s, 8H), 3.67 (q, $J = 9.9, 8.1$ Hz, 4H), 3.35 (t, $J = 11.5$ Hz, 1H), 2.59 (t, $J = 11.4$ Hz, 1H), 2.23 (s, 7H), 1.13 (dt, $J = 22.9, 7.3$ Hz, 7H).

¹³C NMR (100 MHz, CDCl₃): δ 167.4(d, $J_{CF} = 262.6$ Hz), 167.1(d, $J_{CF} = 262.6$ Hz), 160.5, 159.7, 159.7 159.2, 133.3, 132.9(d, $J_{CF} = 7.07$ Hz), 132.7(d, $J_{CF} = 7.07$ Hz), 129.1, 128.8, 128.4, 128.2, 126.9, 126.8, 124.0, 123.3, 115.5(d, $J_{CF} = 29.3$ Hz), 115.3(d, $J_{CF} = 31.3$ Hz), 114.0, 111.2, 110.4, 60.6, 55.4, 51.7, 49.9, 47.6, 43.3, 41.3, 38.6, 36.7, 18.4, 14.0, 13.8.

¹⁹F NMR (377 MHz, CDCl₃) δ -111.4, -118.3.

$[\alpha]^{25}_D = -208.0$ ($c = 0.5$ in acetone, $\lambda = 589$ nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 80:20 in the first 5 mins and maintaining in 80:20 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ= 254 nm), t_R (major) = 9.5 min, t_R (minor) = 10.9 min.



ethyl (4aR,5S,6R,8aR)-8-(4-methoxyphenyl)-3-methyl-1-oxo-6-phenyl-5-(o-tolyl)-4a,5,6,8a-tetrahydro-1H-isochromene-4-carboxylate (3d)

18 h, light yellow solid; 32% yield, 94:6 dr, 96% ee;

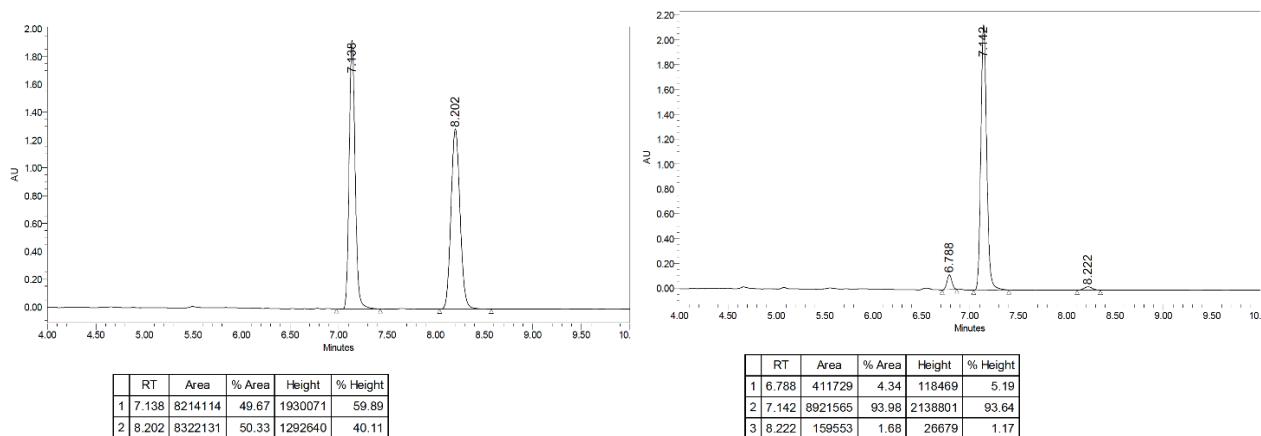
HRMS (ESI+): calcd. for C₃₃H₃₂O₅ +Na⁺ [M+Na⁺] 531.2142, found 531.2136;

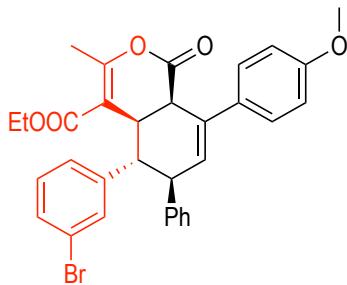
IR: ν_{max} (film, cm⁻¹): 2986, 1775, 1704, 1514, 1253, 1067, 706;

¹H NMR (400 MHz, CDCl₃): δ 7.47 – 7.42 (m, 2H), 7.39 (dd, J = 7.8, 1.3 Hz, 1H), 7.16 (td, J = 7.5, 1.4 Hz, 1H), 7.13 – 7.10 (m, 3H), 7.02 (td, J = 7.4, 1.3 Hz, 1H), 6.94 – 6.86 (m, 5H), 6.29 (d, J = 3.0 Hz, 1H), 3.91 (dd, J = 5.3, 1.2 Hz, 1H), 3.87 (dd, J = 10.3, 2.9 Hz, 1H), 3.83 (s, 3H), 3.81 – 3.65 (m, 3H), 3.09 (dd, J = 12.4, 10.2 Hz, 1H), 2.17 (s, 3H), 1.60 (s, 3H), 1.12 (t, J = 7.2 Hz, 3H);

¹³C NMR (100 MHz, CDCl₃): δ 168.8, 166.0, 159.3, 158.0, 142.6, 138.1, 137.8, 133.2, 132.8, 129.9, 129.3, 128.4, 128.3, 127.5, 127.0, 126.7, 126.6, 125.4, 114.1, 111.8, 60.7, 55.5, 51.5, 45.7, 43.6, 39.0, 19.6, 18.2, 14.1;

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ= 254 nm), t_R (major) = 7.1 min, t_R (minor) = 8.2 min.





ethyl (4aR,5S,6R,8aR)-5-(3-bromophenyl)-8-(4-methoxyphenyl)-3-methyl-1-oxo-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromene-4-carboxylate (3e)

23 h, light yellow solid; 38% yield, 96% ee;

HRMS (ESI+): calcd. for $C_{32}H_{29}BrO_5 + Na^+$ [M+Na⁺] 595.1091, found 595.1086;

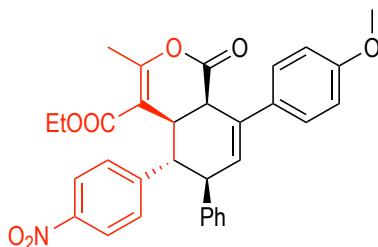
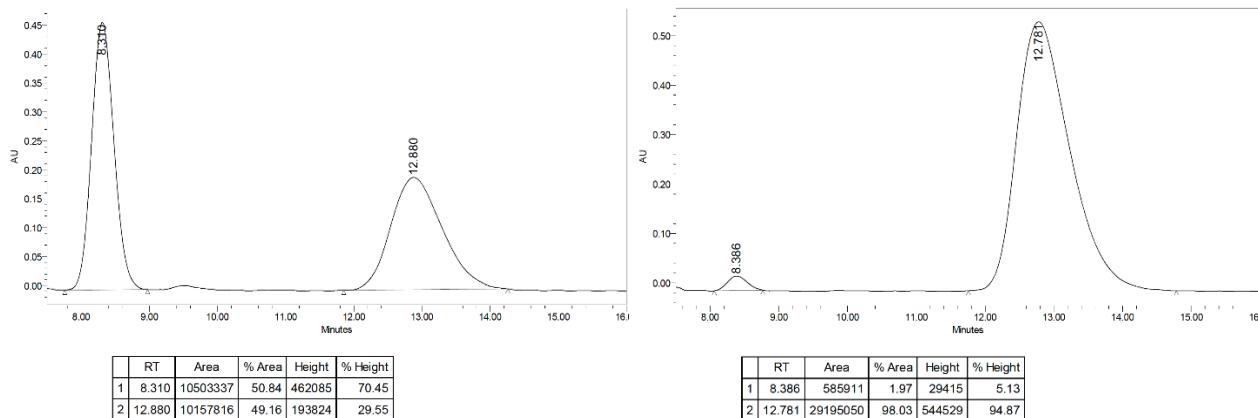
IR: ν_{max} (film, cm⁻¹): 2990, 1783, 1706, 1515, 1255, 1062, 706;

¹H NMR (400 MHz, CDCl₃): δ 7.43 – 7.38 (m, 2H), 7.31 – 7.27 (m, 1H), 7.20 – 7.08 (m, 4H), 7.03 – 6.96 (m, 1H), 6.96 – 6.85 (m, 5H), 6.23 (d, J = 3.0 Hz, 1H), 3.92 – 3.84 (m, 3H), 3.82 (s, 3H), 3.80 – 3.71 (m, 2H), 2.65 (dd, J = 12.3, 10.3 Hz, 1H), 2.23 (s, 3H), 1.19 (t, J = 7.1 Hz, 3H);

¹³C NMR (100 MHz, CDCl₃): δ 168.4, 165.8, 159.4, 159.0, 142.2, 142.2, 133.0, 132.6, 130.3, 129.6, 128.9, 128.6, 128.4, 127.0, 127.0, 122.0, 114.1, 111.3, 61.1, 55.5, 51.9, 50.8, 43.4, 38.8, 18.5, 14.2; (two aromatic carbon signals overlap with others in the 120–135 ppm region)

$[\alpha]^{25}_D$ = -80.8 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Chiralpak AD-3, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 12.8 min, t_R (minor) = 8.3 min.



ethyl (4aR,5S,6R,8aR)-8-(4-methoxyphenyl)-3-methyl-5-(4-nitrophenyl)-1-oxo-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromene-4-carboxylate (3f)

27 h, light yellow solid; 46% yield, 97:3 dr, 94% ee;

HRMS (ESI+): calcd. for $C_{32}H_{29}NO_7 + Na^+$ [M+Na⁺] 562.1836, found 562.1834;

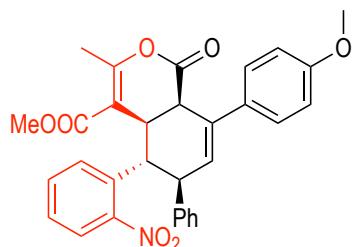
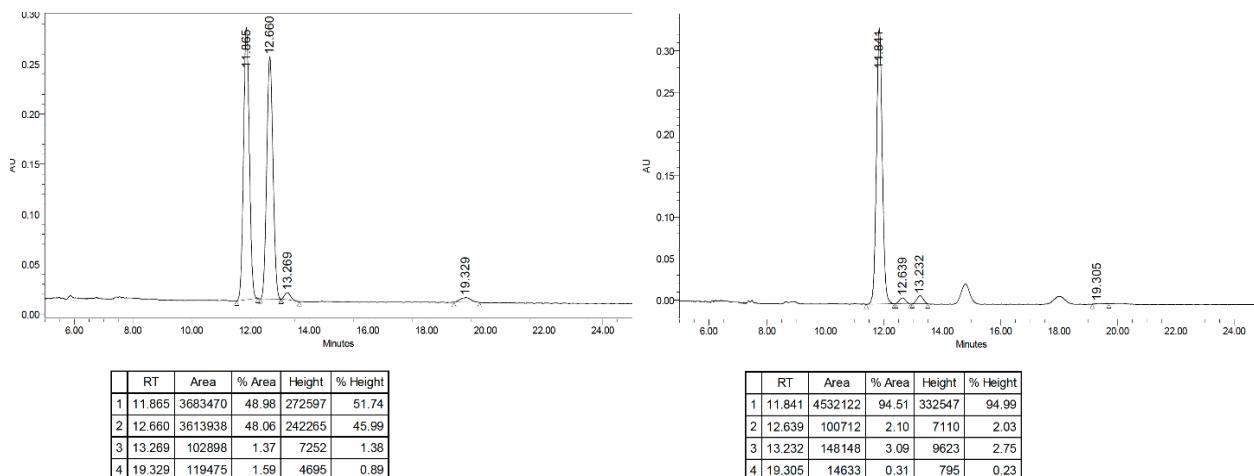
IR: ν_{max} (film, cm⁻¹): 2935, 1777, 1702, 1513, 1344, 1064, 701;

¹H NMR (600 MHz, CDCl₃): δ 8.03 (br, 2H), 7.41 – 7.38 (m, 2H), 7.16 – 7.11 (m, 4H), 6.91 – 6.88 (m, 5H), 6.23 (d, J = 2.9 Hz, 1H), 3.92 – 3.88 (m, 2H), 3.84 – 3.82 (m, 1H), 3.82 (s, 3H), 3.82 – 3.80 (m, 1H), 3.78 – 3.72 (m, 1H), 2.83 (dd, J = 12.3, 10.2 Hz, 1H), 2.23 (s, 3H), 1.10 (t, J = 7.1 Hz, 3H);

¹³C NMR (150 MHz, CDCl₃): δ 168.1, 165.8, 159.7, 159.6, 147.8, 147.2, 141.7, 133.2, 132.4, 128.8, 128.6, 128.3, 127.2, 127.1, 123.1, 114.2, 110.9, 61.1, 55.5, 52.3, 51.0, 43.4, 38.6, 18.7, 14.2; (one aromatic carbon signal overlaps with others in the 120–135 ppm region)

$[\alpha]^{25}_D$ = -41.6 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-2, gradient 100% CO₂ to CO₂/MeOH = 80:20 in the first 5 mins and maintaining in 80:20 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 11.8 min, t_R (minor) = 12.6 min.



methyl (4aR,5S,6R,8aR)-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-1-oxo-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromene-4-carboxylate (3g)

17 h, light yellow solid; 43% yield, 98% ee;

HRMS (ESI+): calcd. for C₃₁H₂₇NO₇ +Na⁺ [M+Na⁺] 548.1680, found 548.1671;

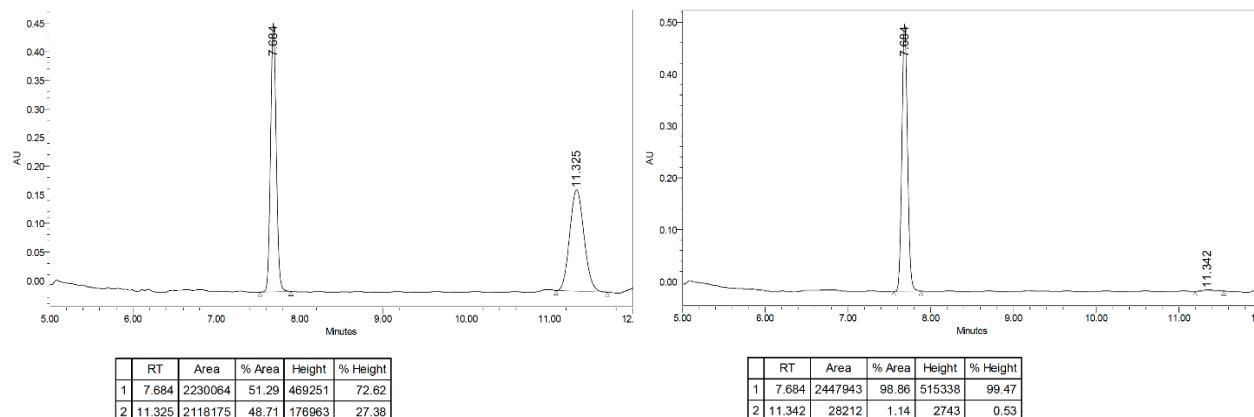
IR: ν_{max} (film, cm⁻¹): 2953, 1782, 1710, 1515, 1247, 1071, 700;

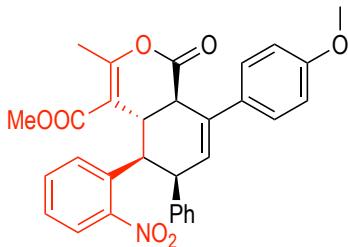
¹H NMR (400 MHz, CDCl₃) δ 7.69 (dd, *J* = 8.0, 1.4 Hz, 1H), 7.55 (td, *J* = 7.6, 1.4 Hz, 1H), 7.45 (dd, *J* = 8.2, 1.4 Hz, 1H), 7.42 – 7.36 (m, 2H), 7.31 – 7.26 (m, 1H), 7.18 – 7.11 (m, 3H), 6.93 – 6.84 (m, 4H), 6.20 (d, *J* = 2.9 Hz, 1H), 3.91 (dd, *J* = 5.1, 1.3 Hz, 1H), 3.89 – 3.84 (m, 1H), 3.82 (s, 3H), 3.79 (d, *J* = 5.3 Hz, 1H), 3.63 (dd, *J* = 12.0, 10.4 Hz, 1H), 3.37 (s, 3H), 2.24 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 167.5, 166.4, 161.4, 159.4, 151.7, 141.3, 133.9, 133.3, 132.6, 131.2, 129.9, 128.8, 128.7, 128.1, 127.9, 127.3, 127.1, 123.9, 114.1, 109.2, 55.5, 51.5, 51.3, 44.1, 43.5, 38.6, 18.7;

$[\alpha]^{25}_{D} = -69.6$ (*c* = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 50:50 in the first 5 mins and maintaining in 50:50 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 7.7 min, t_R (minor) = 11.3 min.





methyl (4aS,5R,6R,8aR)-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-1-oxo-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromene-4-carboxylate (4g)

light yellow solid; 17% yield, >99% ee;

HRMS (ESI+): calcd. for C₃₁H₂₇NO₇ + Na⁺ [M+Na⁺] 548.1680, found 548.1669;

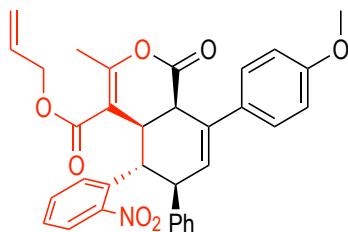
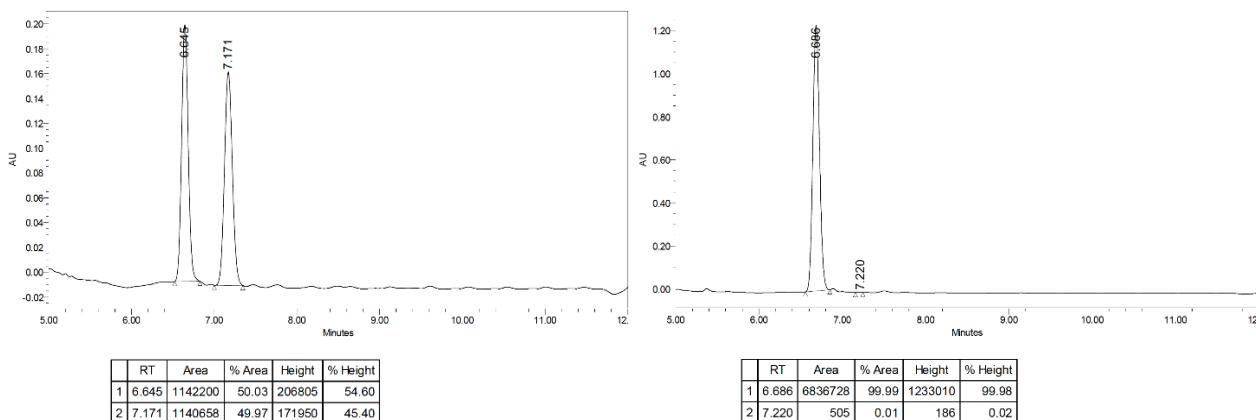
IR: ν_{max} (film, cm⁻¹): 2948, 1770, 1712, 1515, 1244, 1091, 706;

¹H NMR (400 MHz, CDCl₃) δ 7.86 (dd, J = 8.1, 1.4 Hz, 1H), 7.35 – 7.27 (m, 3H), 7.19 – 7.13 (m, 1H), 7.12 – 7.05 (m, 3H), 6.95 – 6.86 (m, 2H), 6.70 (br, 2H), 6.32 (dd, J = 6.3, 1.9 Hz, 1H), 6.08 (dd, J = 8.1, 1.3 Hz, 1H), 4.35 (t, J = 5.8 Hz, 1H), 4.27 (dd, J = 12.7, 5.2 Hz, 1H), 3.95 (dt, J = 13.1, 1.7 Hz, 1H), 3.83 (s, 3H), 3.54 (td, J = 12.8, 2.1 Hz, 1H), 3.11 (s, 3H), 1.99 (d, J = 1.9 Hz, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 167.6, 166.4, 159.1, 155.3, 150.7, 141.0, 137.8, 134.4, 133.3, 132.6, 131.8, 130.5, 129.7, 129.3, 128.1, 127.5, 126.4, 124.5, 114.4, 114.0, 55.4, 51.8, 46.2, 45.0, 39.9, 32.1, 17.3;

[α]²⁵_D = -20.8 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-2, gradient 100% CO₂ to CO₂/MeOH = 50:50 in the first 5 mins and maintaining in 50:50 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 6.7 min, t_R (minor) = 7.2 min.



allyl (4aR,5S,6R,8aR)-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-1-oxo-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromene-4-carboxylate (3h)

24 h, light yellow solid; 49% yield, 94:6 dr, 97% ee;

HRMS (ESI+): calcd. for C₃₃H₂₉NO₇ + Na⁺ [M+Na⁺] 574.1836, found 574.1827

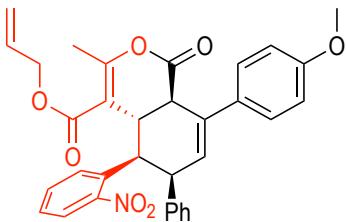
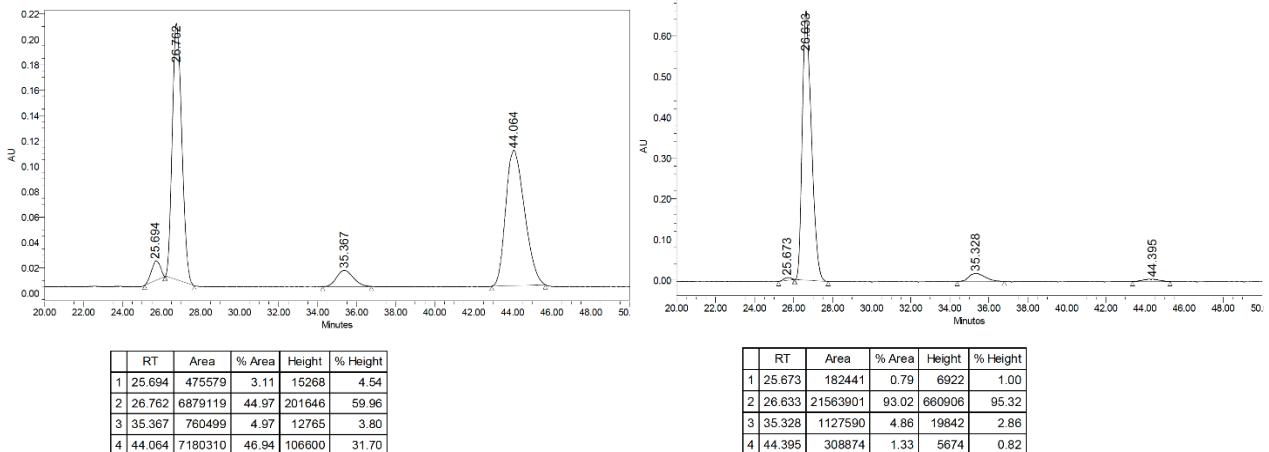
IR: ν_{max} (film, cm⁻¹): 3027, 1778, 1709, 1515, 1240, 1057, 700;

¹H NMR (400 MHz, CDCl₃) δ 7.69 (dd, J = 8.1, 1.4 Hz, 1H), 7.56 – 7.50 (m, 1H), 7.46 – 7.37 (m, 3H), 7.31 – 7.27 (m, 1H), 7.17 – 7.12 (m, 3H), 6.92 – 6.85 (m, 4H), 6.21 (d, J = 2.9 Hz, 1H), 5.76 (ddt, J = 16.6, 10.4, 5.8 Hz, 1H), 5.27 – 5.18 (m, 2H), 4.30 (ddt, J = 13.2, 5.6, 1.4 Hz, 1H), 4.20 – 4.12 (m, 1H), 3.92 (dd, J = 5.2, 1.3 Hz, 1H), 3.88 – 3.84 (m, 1H), 3.82 (s, 3H), 3.83 – 3.79 (m, 1H), 3.64 (dd, J = 12.1, 10.4 Hz, 1H), 2.26 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 167.5, 165.7, 161.4, 159.4, 151.7, 141.2, 133.8, 133.2, 132.5, 131.8, 131.4, 129.9, 128.8, 128.6, 128.1, 127.8, 127.2, 127.1, 123.8, 118.8, 114.1, 109.2, 65.5, 55.4, 51.3, 44.1, 43.5, 38.4, 18.8;

[α]²⁵_D = -80.8 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 90:10 in the first 5 mins and maintaining in 90:10 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 26.6 min, t_R (minor) = 44.4 min.



allyl (4aS,5R,6R,8aR)-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-1-oxo-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromene-4-carboxylate (4h)

light yellow solid; 12% yield, >99% ee;

HRMS (ESI+): calcd. for $C_{33}H_{29}NO_7 + Na^+$ [M+Na⁺] 574.1836, found 574.1829

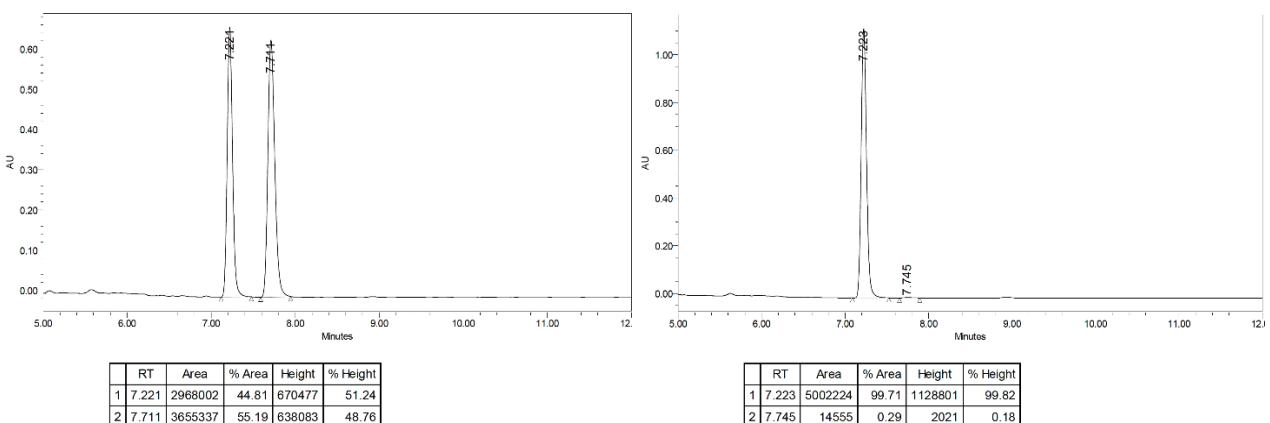
IR: ν_{max} (film, cm⁻¹): 2924, 1774, 1713, 1515, 1244, 1090, 703;

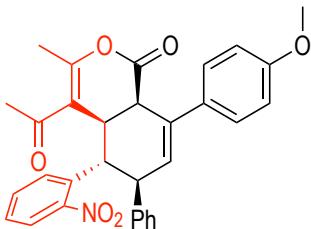
¹H NMR (400 MHz, CDCl₃) δ 7.88 (dd, $J = 8.2, 1.4$ Hz, 1H), 7.35 – 7.30 (m, 2H), 7.28 – 7.24 (m, 1H), 7.18 – 7.02 (m, 4H), 6.95 – 6.86 (m, 2H), 6.71 (s, 2H), 6.31 (dd, $J = 5.9, 2.1$ Hz, 1H), 6.10 (dd, $J = 8.0, 1.3$ Hz, 1H), 5.57 (ddt, $J = 17.4, 10.1, 6.0$ Hz, 1H), 5.17 – 5.08 (m, 2H), 4.41 – 4.31 (m, 2H), 4.01 – 3.93 (m, 2H), 3.91 – 3.85 (m, 1H), 3.83 (s, 3H), 3.54 (td, $J = 12.6, 2.1$ Hz, 1H), 1.99 (d, $J = 1.9$ Hz, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 167.6, 165.7, 159.1, 154.8, 150.6, 137.9, 134.6, 133.3, 132.5, 131.8, 131.3, 130.5, 129.8, 129.4, 128.1, 127.4, 126.4, 124.6, 119.4, 114.5, 114.0, 65.8, 55.4, 46.1, 45.0, 39.7, 32.2, 17.3;

$[\alpha]^{25}_D = -8.8$ ($c = 0.5$ in acetone, $\lambda = 589$ nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, $\lambda = 254$ nm), t_R (major) = 7.2 min, t_R (minor) = 7.7 min.





(4a*R*,5*S*,6*R*,8a*R*)-4-acetyl-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-6-phenyl-4*a*,5,6,8*a*-tetrahydro-1*H*-isochromen-1-one (3i)

13 h, light yellow solid; 72% yield, >99% ee;

HRMS (ESI+): calcd. for $C_{31}H_{27}NO_6 + Na^+$ [M+Na⁺] 532.1731, found 532.1727;

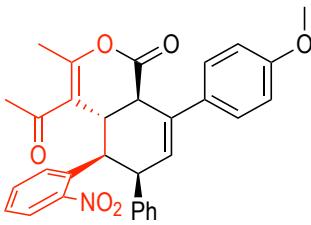
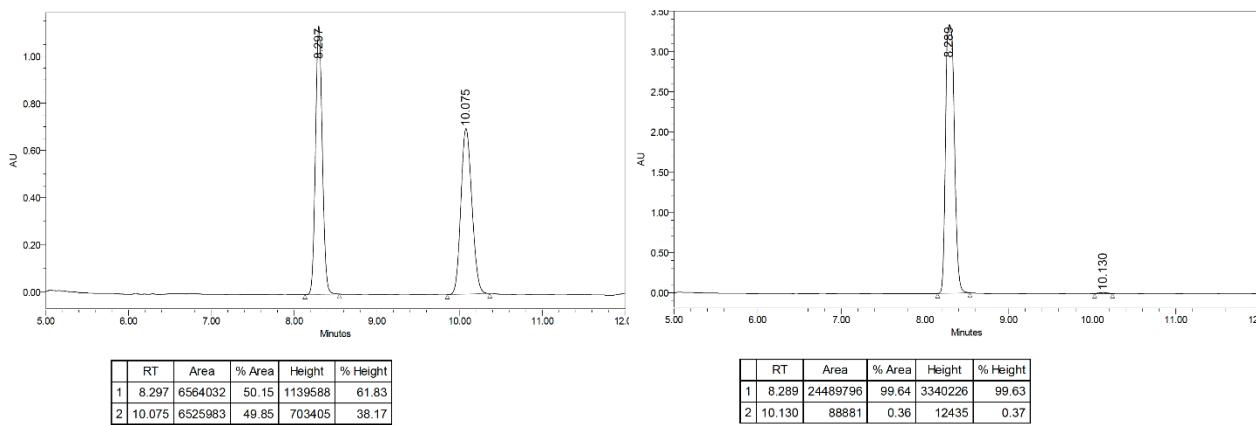
IR: ν_{max} (film, cm⁻¹): 2956, 1775, 1510, 1356, 1242, 1100, 702;

¹H NMR (400 MHz, CDCl₃) δ 7.60 – 7.53 (m, 2H), 7.42 – 7.35 (m, 3H), 7.32 – 7.26 (m, 1H), 7.19 – 7.12 (m, 3H), 6.94 – 6.83 (m, 4H), 6.19 (d, *J* = 2.8 Hz, 1H), 3.93 – 3.83 (m, 3H), 3.81 (s, 3H), 3.47 (t, *J* = 11.3 Hz, 1H), 2.18 (s, 3H), 1.73 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 198.7, 167.6, 159.4, 157.5, 151.8, 141.3, 134.1, 133.3, 132.6, 131.8, 130.1, 128.8, 128.6, 128.1, 128.0, 127.3, 127.1, 123.4, 119.9, 114.1, 55.5, 51.1, 44.2, 43.4, 39.2, 30.2, 19.3;

$[\alpha]^{25}_D = -66.4$ (*c* = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 50:50 in the first 5 mins and maintaining in 50:50 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), *t_R* (major) = 8.3 min, *t_R* (minor) = 10.1 min.



(4a*S*,5*R*,6*R*,8a*R*)-4-acetyl-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-6-phenyl-4*a*,5,6,8*a*-tetrahydro-1*H*-isochromen-1-one (4i)

light yellow solid; 14% yield, >99% ee;

HRMS (ESI+): calcd. for $C_{31}H_{27}NO_6 + Na^+$ [M+Na⁺] 532.1731, found 532.1724;

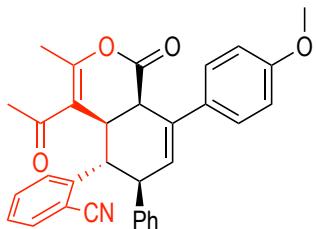
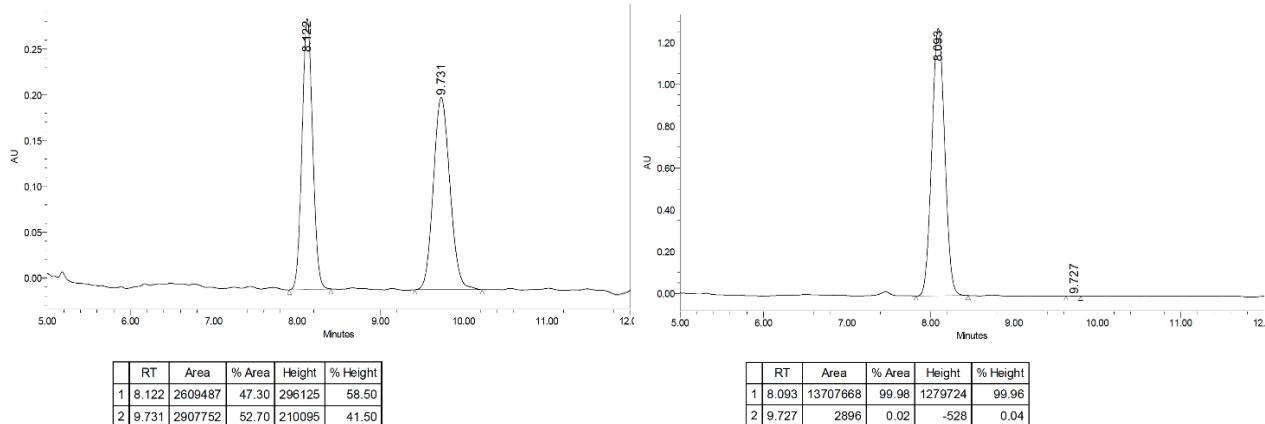
IR: ν_{max} (film, cm⁻¹): 2930, 1770, 1522, 1357, 1243, 1085, 702;

¹H NMR (400 MHz, CDCl₃) δ 7.85 (dd, *J* = 8.1, 1.4 Hz, 1H), 7.33 – 7.27 (m, 3H), 7.20 – 7.08 (m, 3H), 7.04 (td, *J* = 7.6, 1.4 Hz, 1H), 6.94 – 6.87 (m, 2H), 6.75 (br, 2H), 6.30 (dd, *J* = 6.2, 2.0 Hz, 1H), 6.04 (dd, *J* = 7.9, 1.3 Hz, 1H), 4.38 – 4.32 (m, 1H), 4.26 (dd, *J* = 12.7, 5.4 Hz, 1H), 3.93 (dt, *J* = 12.9, 1.7 Hz, 1H), 3.83 (s, 3H), 3.52 (td, *J* = 12.8, 2.0 Hz, 1H), 1.94 (d, *J* = 1.8 Hz, 3H), 1.54 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 201.3, 167.8, 159.1, 151.3, 150.9, 138.1, 135.1, 133.4, 132.6, 131.3, 130.6, 129.7, 129.6, 128.1, 127.7, 127.5, 126.5, 124.7, 123.6, 114.0, 55.4, 46.1, 45.3, 39.1, 33.8, 31.6, 17.4;

$[\alpha]^{25}_D = +45.6$ (*c* = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-2, gradient 100% CO₂ to CO₂/MeOH = 50:50 in the first 5 mins and maintaining in 50:50 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), *t_R* (major) = 8.1 min, *t_R* (minor) = 9.7 min.



2-((4aR,5S,6R,8aR)-4-acetyl-8-(4-methoxyphenyl)-3-methyl-1-oxo-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromen-5-yl)benzonitrile (3j)

13 h, light yellow solid; 67% yield, >99% ee;

HRMS (ESI+): calcd. for $C_{32}H_{27}NO_4 + Na^+$ [M+Na $^+$] 512.1832, found 512.1835

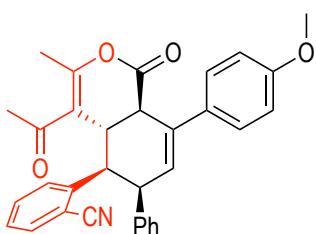
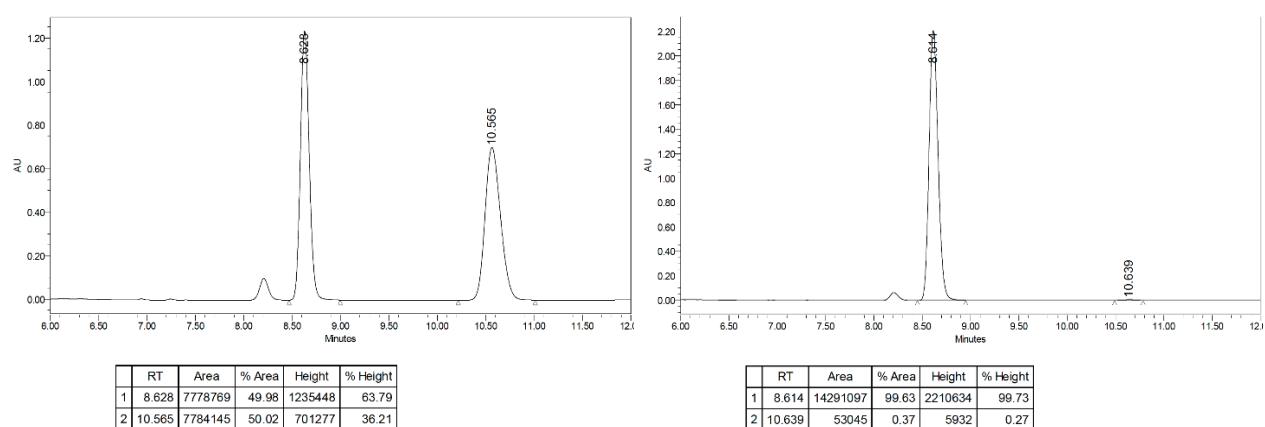
IR: ν_{max} (film, cm $^{-1}$): 2955, 2224, 1776, 1510, 1240, 1104, 702;

1H NMR (400 MHz, CDCl $_3$) δ 7.60 – 7.53 (m, 2H), 7.41 – 7.37 (m, 3H), 7.30 – 7.26 (m, 1H), 7.19 – 7.13 (m, 3H), 6.96 (dd, J = 7.7, 1.8 Hz, 2H), 6.91 – 6.87 (m, 2H), 6.22 (d, J = 2.9 Hz, 1H), 3.95 – 3.90 (m, 1H), 3.88 – 3.85 (m, 1H), 3.83 – 3.82 (m, 1H), 3.81 (s, 3H), 3.34 (dd, J = 11.8, 10.4 Hz, 1H), 2.22 (s, 3H), 1.71 (s, 3H);

^{13}C NMR (100 MHz, CDCl $_3$) δ 198.0, 167.4, 159.4, 157.6, 144.0, 141.4, 133.2, 132.6, 132.6, 132.5, 128.8, 128.7, 128.6, 128.2, 127.8, 127.2, 127.1, 119.7, 116.8, 114.9, 114.1, 114.0, 55.4, 50.4, 48.9, 43.3, 39.7, 30.1, 19.2;

$[\alpha]^{25}_D$ = -114.4 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO $_2$ to CO $_2$ /MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO $_2$ /MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 8.6 min, t_R (minor) = 10.6 min.



2-((4aS,5R,6R,8aR)-4-acetyl-8-(4-methoxyphenyl)-3-methyl-1-oxo-6-phenyl-4a,5,6,8a-tetrahydro-1*H*-isochromen-5-yl)benzonitrile (4j)

light yellow solid; 13% yield, >99% ee;

HRMS (ESI+): calcd. for $C_{32}H_{27}NO_4 + Na^+ [M+Na^+]$ 512.1832, found 512.1829

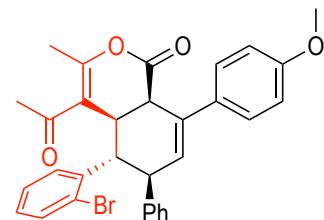
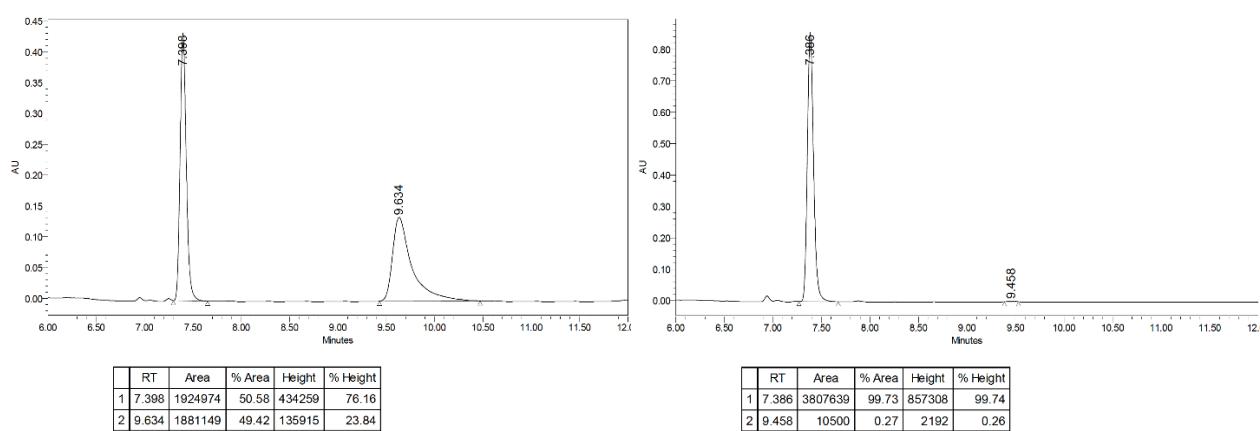
IR: ν_{max} (film, cm^{-1}): 2906, 2222, 1778, 1674, 1514, 1242, 1080, 706;

1H NMR (400 MHz, CDCl₃) δ 7.70 (dd, J = 7.8, 1.4 Hz, 1H), 7.35 – 7.29 (m, 2H), 7.23 (dd, J = 7.6, 1.1 Hz, 1H), 7.18 – 7.12 (m, 1H), 7.11 – 7.04 (m, 3H), 6.94 – 6.87 (m, 2H), 6.62 (br, 2H), 6.28 (dd, J = 6.3, 2.1 Hz, 1H), 6.05 (d, J = 7.9 Hz, 1H), 4.31 (dd, J = 12.9, 5.4 Hz, 1H), 4.06 – 3.98 (m, 2H), 3.83 (s, 3H), 3.53 (td, J = 13.0, 2.0 Hz, 1H), 1.97 (d, J = 1.9 Hz, 3H), 1.64 (s, 3H);

^{13}C NMR (100 MHz, CDCl₃) δ 201.0, 167.8, 159.1, 151.0, 144.2, 137.5, 133.4, 133.3, 132.9, 131.7, 130.3, 129.3, 128.1, 127.5, 127.4, 126.5, 123.5, 117.6, 114.1, 114.0, 55.4, 46.7, 45.2, 42.3, 33.1, 31.8, 17.4;

$[\alpha]^{25}_D$ = -132.0 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 7.4 min, t_R (minor) = 9.4 min.



(4a*R*,5*S*,6*R*,8a*R*)-4-acetyl-5-(2-bromophenyl)-8-(4-methoxyphenyl)-3-methyl-6-phenyl-4*a*,5,6,8*a*-tetrahydro-1*H*-isochromen-1-one (3k)

15 h, light yellow solid; 58% yield, 99% ee;

HRMS (ESI+): calcd. for $C_{31}H_{27}BrO_4 + Na^+ [M+Na^+]$ 565.0985, found 565.0981;

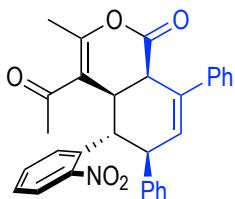
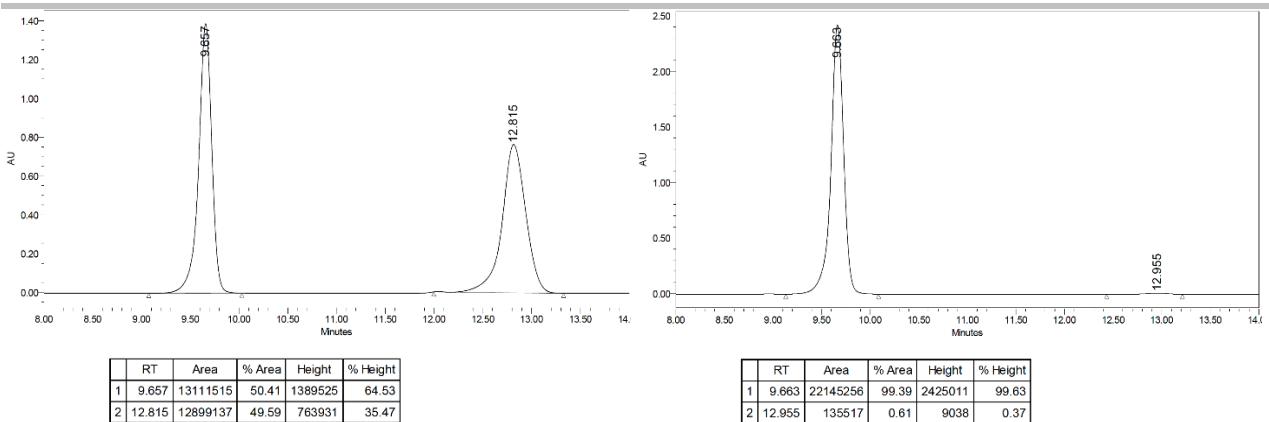
IR: ν_{max} (film, cm^{-1}): 2956, 1772, 1510, 1240, 1100, 1026, 765;

1H NMR (600 MHz, CDCl₃) δ 7.42 – 7.38 (m, 2H), 7.36 (dd, J = 7.9, 1.7 Hz, 1H), 7.33 – 7.27 (m, 2H), 7.17 – 7.10 (m, 3H), 7.02 (td, J = 7.6, 1.6 Hz, 1H), 6.98 – 6.94 (m, 2H), 6.90 (d, J = 8.7 Hz, 2H), 6.23 (d, J = 2.8 Hz, 1H), 3.88 – 3.83 (m, 2H), 3.82 (s, 3H), 3.76 (dd, J = 12.3, 5.3 Hz, 1H), 3.61 (dd, J = 12.2, 10.4 Hz, 1H), 2.15 (s, 3H), 1.64 (s, 3H);

^{13}C NMR (150 MHz, CDCl₃) δ 199.0, 168.3, 159.4, 155.9, 141.8, 139.6, 133.1, 132.8, 132.7, 129.7, 129.2, 128.6, 128.5, 128.4, 127.5, 127.5, 127.1, 126.9, 120.1, 114.1, 55.5, 50.6, 48.3, 43.5, 40.4, 29.9, 19.0;

$[\alpha]^{25}_D$ = -151.2 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 9.7 min, t_R (minor) = 12.9 min.



13 h, light yellow solid; 45% yield, 99% ee;

HRMS (ESI+): calcd. for $C_{30}H_{25}NO_5 + Na^+$ [M+Na⁺] 502.1625, found 502.1620;

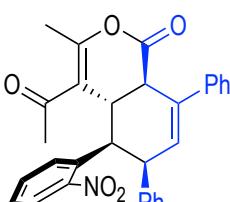
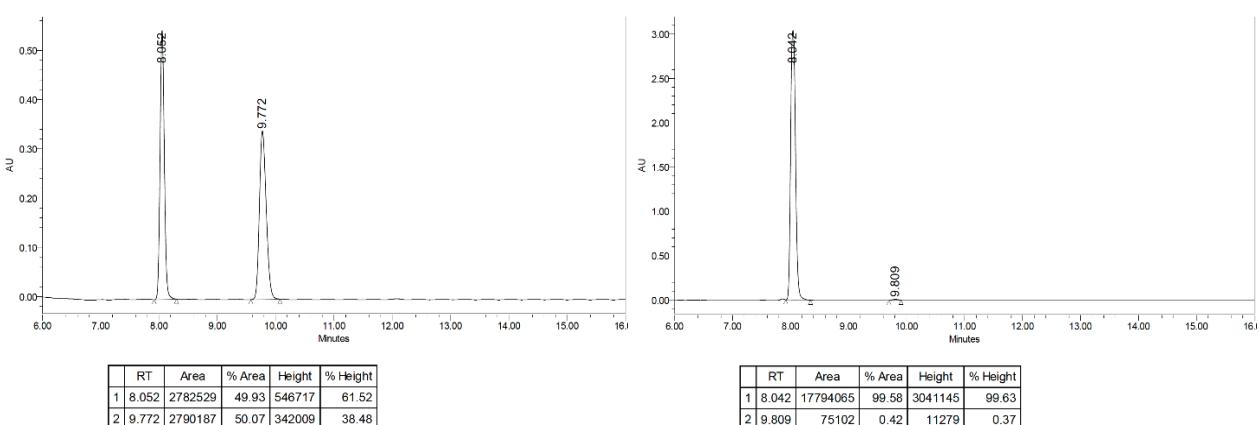
IR: ν_{max} (film, cm⁻¹): 3028, 1768, 1651, 1520, 1102, 750, 698;

¹H NMR (400 MHz, CDCl₃) δ 7.62 – 7.53 (m, 2H), 7.48 – 7.43 (m, 2H), 7.42 – 7.27 (m, 5H), 7.21 – 7.13 (m, 3H), 6.92 – 6.85 (m, 2H), 6.29 (d, $J = 2.8$ Hz, 1H), 3.96 – 3.84 (m, 3H), 3.49 (dd, $J = 11.8, 10.4$ Hz, 1H), 2.19 (s, 3H), 1.74 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 198.6, 167.6, 157.5, 151.8, 141.1, 140.0, 134.0, 134.0, 131.9, 130.2, 130.1, 128.8, 128.7, 128.1, 128.0, 127.9, 127.3, 126.0, 123.4, 119.9, 51.1, 44.2, 43.4, 39.2, 30.2, 19.3;

$[\alpha]^{25}_D = -83.2$ ($c = 0.5$ in acetone, $\lambda = 589$ nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, $\lambda = 254$ nm), t_R (major) = 8.0 min, t_R (minor) = 9.8 min.



light yellow solid; 10% yield, >99% ee;

HRMS (ESI+): calcd. for $C_{30}H_{25}NO_5 + Na^+$ [M+Na⁺] 502.1625, found 502.1625;

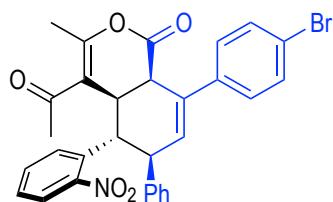
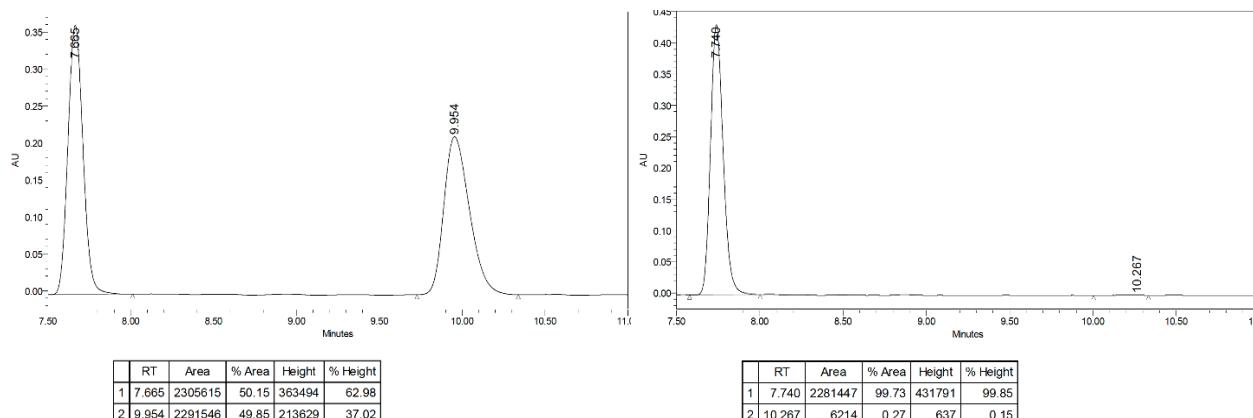
IR: ν_{max} (film, cm⁻¹): 3031, 1772, 1522, 1359, 1085, 976, 705;

¹H NMR (400 MHz, CDCl₃) δ 7.85 (dd, *J* = 8.1, 1.4 Hz, 1H), 7.41 – 7.33 (m, 4H), 7.32 – 7.27 (m, 2H), 7.21 – 7.09 (m, 3H), 7.04 (td, *J* = 7.6, 1.4 Hz, 1H), 6.76 (br, 2H), 6.37 (dd, *J* = 6.2, 2.0 Hz, 1H), 6.05 (dd, *J* = 8.1, 1.4 Hz, 1H), 4.40 – 4.34 (m, 1H), 4.27 (dd, *J* = 12.8, 5.4 Hz, 1H), 3.98 (dt, *J* = 12.9, 1.7 Hz, 1H), 3.53 (td, *J* = 12.9, 2.0 Hz, 1H), 1.95 (d, *J* = 1.9 Hz, 3H), 1.54 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 201.3, 167.7, 151.3, 151.0, 140.9, 138.0, 135.1, 133.2, 131.3, 131.1, 130.6, 129.6, 128.6, 128.2, 127.8, 127.5, 127.5, 125.4, 124.7, 123.5, 46.0, 45.3, 39.1, 33.8, 31.6, 17.4;

[α]²⁵_D = -2.4 (*c* = 0.5 in acetone, *λ* = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, *λ* = 254 nm), t_R (major) = 7.7 min, t_R (minor) = 10.3 min.



(4aR,5S,6R,8aR)-4-acetyl-8-(4-bromophenyl)-3-methyl-5-(2-nitrophenyl)-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (3m)

13 h, light yellow solid; 48% yield, >99% ee;

HRMS (ESI+): calcd. for C₃₀H₂₄BrNO₅ +Na⁺ [M+Na⁺] 580.0730, found 580.0724;

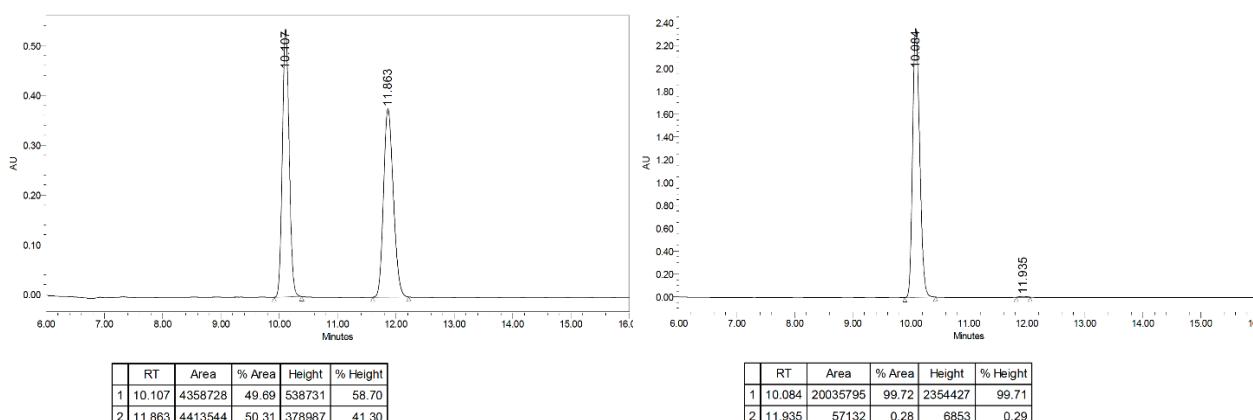
IR: ν_{max} (film, cm⁻¹): 3025, 1770, 1655, 1526, 1354, 1097, 700;

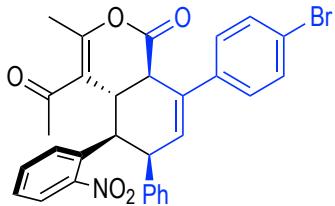
¹H NMR (400 MHz, CDCl₃) δ 7.60 – 7.54 (m, 2H), 7.50 – 7.45 (m, 2H), 7.43 – 7.37 (m, 1H), 7.35 – 7.28 (m, 3H), 7.19 – 7.12 (m, 3H), 6.88 – 6.82 (m, 2H), 6.26 (d, *J* = 2.8 Hz, 1H), 3.91 (dd, *J* = 12.1, 5.3 Hz, 1H), 3.87 – 3.81 (m, 2H), 3.49 (dd, *J* = 12.0, 10.4 Hz, 1H), 2.18 (s, 3H), 1.73 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 198.5, 167.4, 157.5, 151.8, 140.8, 139.0, 133.8, 133.2, 131.9, 131.8, 130.9, 123.0, 128.9, 128.1, 128.1, 127.7, 127.4, 123.5, 121.9, 119.8, 51.0, 44.0, 43.3, 39.1, 30.2, 19.3;

[α]²⁵_D = -71.2 (*c* = 0.5 in acetone, *λ* = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, *λ* = 254 nm), t_R (major) = 10.1 min, t_R (minor) = 11.9 min.





(4aS,5R,6R,8aR)-4-acetyl-8-(4-bromophenyl)-3-methyl-5-(2-nitrophenyl)-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (4m)

light yellow solid; 9% yield, >99% ee;

HRMS (ESI+): calcd. for $C_{30}H_{24}BrNO_5 + Na^+$ [M+Na⁺] 580.0730, found 580.0726;

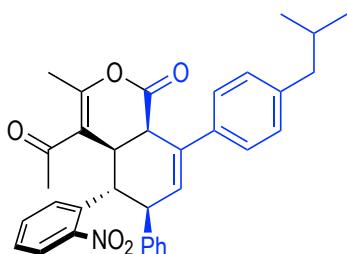
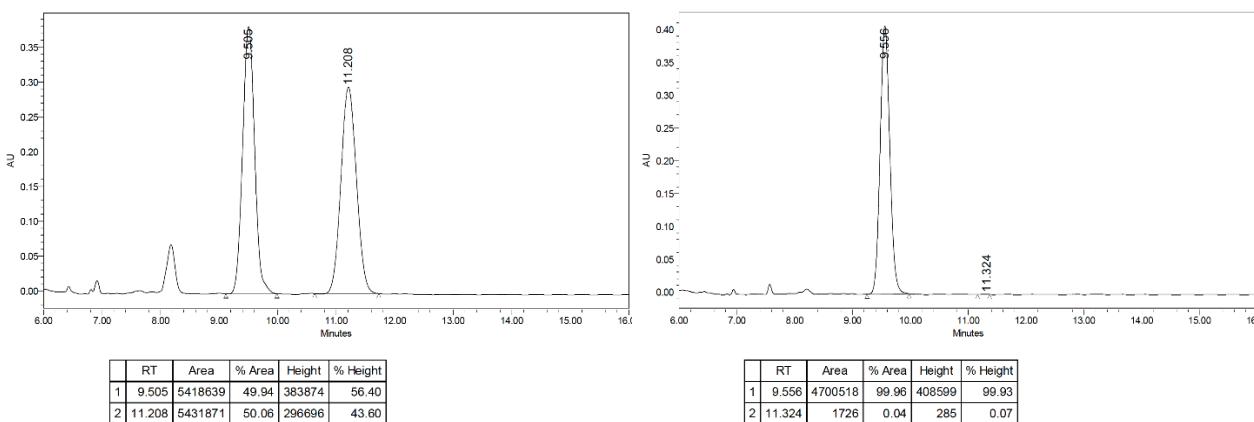
IR: ν_{max} (film, cm⁻¹): 3030, 1770, 1523, 1350, 1073, 975, 705;

¹H NMR (600 MHz, CDCl₃) δ 7.85 (d, $J = 8.1$ Hz, 1H), 7.48 (d, $J = 8.1$ Hz, 2H), 7.28 (d, $J = 8.0$ Hz, 1H), 7.24 (d, $J = 8.9$ Hz, 2H), 7.18 (t, $J = 7.3$ Hz, 1H), 7.12 (br, 2H), 7.04 (t, $J = 7.7$ Hz, 1H), 6.74 (br, 2H), 6.34 (d, $J = 6.1$ Hz, 1H), 6.05 (d, $J = 7.9$ Hz, 1H), 4.36 (t, $J = 5.9$ Hz, 1H), 4.26 (dd, $J = 12.9, 5.4$ Hz, 1H), 3.91 (d, $J = 12.9$ Hz, 1H), 3.52 (t, $J = 12.9$ Hz, 1H), 1.94 (s, 3H), 1.53 (s, 3H);

¹³C NMR (150 MHz, CDCl₃) δ 201.1, 167.6, 151.3, 151.0, 140.0, 137.8, 135.0, 132.4, 131.7, 131.7, 131.4, 129.6, 128.3, 127.8, 127.6, 127.2, 124.7, 123.5, 121.4, 46.0, 45.3, 39.1, 33.8, 31.5, 17.4;

$[\alpha]^{25}_D = +17.6$ ($c = 0.5$ in acetone, $\lambda = 589$ nm);

the enantioselectivity was determined by SFC (Trefoil CEL-2, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, $\lambda = 254$ nm), t_R (major) = 9.5 min, t_R (minor) = 11.3 min;



(4aR,5S,6R,8aR)-4-acetyl-8-(4-isobutylphenyl)-3-methyl-5-(2-nitrophenyl)-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (3n)

15 h, light yellow solid; 46% yield, 99% ee;

HRMS (ESI+): calcd. for $C_{34}H_{33}NO_5 + Na^+$ [M+Na⁺] 558.2251, found 558.2242;

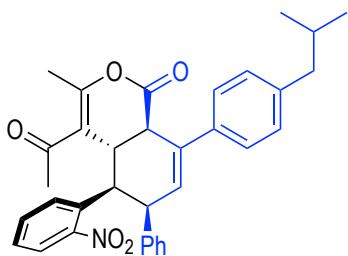
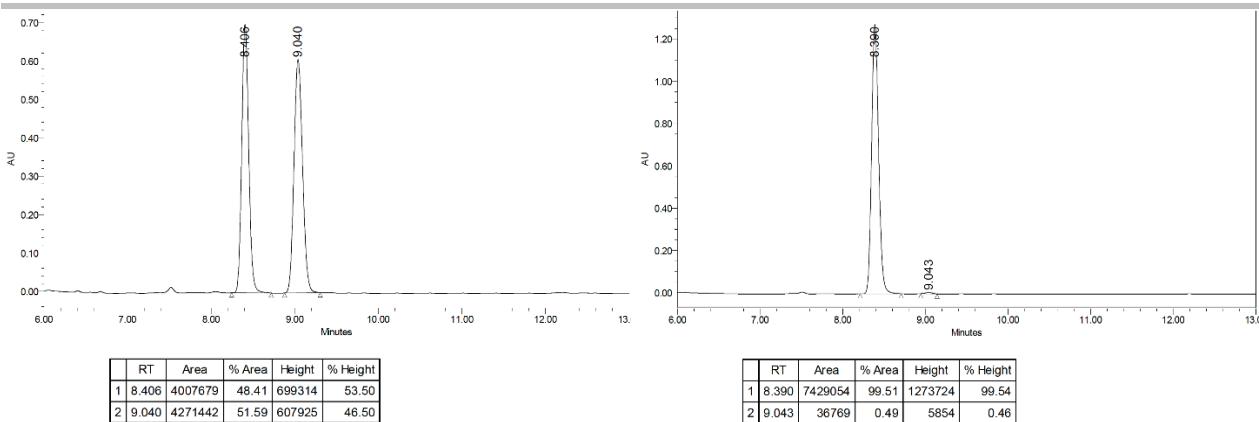
IR: ν_{max} (film, cm⁻¹): 2956, 1772, 1660, 1524, 1353, 1106, 701;

¹H NMR (400 MHz, CDCl₃) δ 7.61 – 7.52 (m, 2H), 7.42 – 7.34 (m, 3H), 7.30 (ddd, $J = 8.3, 6.7, 1.9$ Hz, 1H), 7.20 – 7.09 (m, 5H), 6.89 – 6.86 (m, 2H), 6.27 (d, $J = 2.8$ Hz, 1H), 3.94 – 3.84 (m, 3H), 3.49 (t, $J = 11.1$ Hz, 1H), 2.47 (d, $J = 7.2$ Hz, 2H), 2.18 (s, 3H), 1.86 (dt, $J = 13.5, 6.8$ Hz, 1H), 1.74 (s, 3H), 0.90 (d, $J = 6.6$ Hz, 6H);

¹³C NMR (100 MHz, CDCl₃) δ 198.6, 167.7, 157.5, 151.8, 141.6, 141.2, 137.3, 134.1, 133.8, 131.8, 130.1, 129.5, 129.3, 128.8, 128.1, 128.0, 127.3, 125.6, 123.4, 119.9, 51.1, 45.2, 44.2, 43.3, 39.2, 31.1, 30.3, 30.2, 22.5, 19.3;

$[\alpha]^{25}_D = -115.2$ ($c = 0.5$ in acetone, $\lambda = 589$ nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, $\lambda = 254$ nm), t_R (major) = 8.4 min, t_R (minor) = 9.0 min.



(4aS,5R,6R,8aR)-4-acetyl-8-(4-isobutylphenyl)-3-methyl-5-(2-nitrophenyl)-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (4n)

light yellow solid; 8% yield, 84:16 dr, >99% ee;

HRMS (ESI+): calcd. for $C_{34}H_{33}NO_5 + Na^+$ [M+Na⁺] 558.2251, found 558.2243;

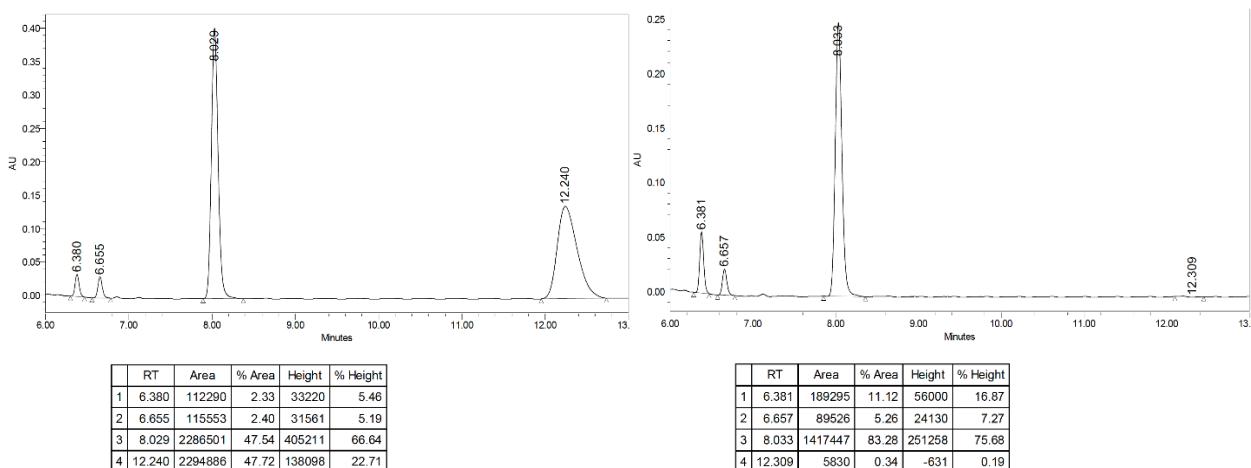
IR: ν_{max} (film, cm⁻¹): 2954, 1777, 1671, 1520, 1352, 1080, 969, 709;

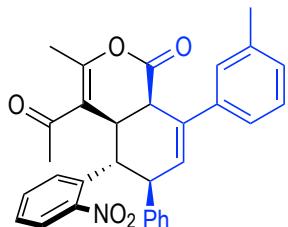
¹H NMR (400 MHz, CDCl₃) δ 7.85 (dd, $J = 8.1, 1.4$ Hz, 1H), 7.31 – 7.27 (m, 3H), 7.21 – 7.07 (m, 5H), 7.04 (td, $J = 7.6, 1.4$ Hz, 1H), 6.76 (br, 2H), 6.36 (dd, $J = 6.2, 2.0$ Hz, 1H), 6.04 (dd, $J = 8.0, 1.3$ Hz, 1H), 4.38 – 4.31 (m, 1H), 4.26 (dd, $J = 12.8, 5.4$ Hz, 1H), 3.97 (dt, $J = 13.0, 1.8$ Hz, 1H), 3.52 (td, $J = 12.8, 1.9$ Hz, 1H), 2.48 (d, $J = 7.2$ Hz, 2H), 1.94 (d, $J = 1.9$ Hz, 3H), 1.88 (dt, $J = 13.5, 6.8$ Hz, 1H), 1.54 (s, 3H), 0.91 (d, $J = 6.6$ Hz, 6H);

¹³C NMR (150 MHz, CDCl₃) δ 201.3, 167.8, 151.3, 151.0, 141.0, 138.1, 138.1, 135.1, 133.0, 131.3, 130.6, 130.3, 129.6, 129.3, 128.1, 127.7, 127.5, 125.0, 124.7, 123.6, 46.1, 45.3, 45.3, 39.1, 33.8, 31.6, 30.3, 22.6, 17.4;

[α]²⁵_D = -26.4 ($c = 0.5$ in acetone, $\lambda = 589$ nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, $\lambda = 254$ nm), t_R (major) = 8.0 min, t_R (minor) = 12.3 min.





(4aR,5S,6R,8aR)-4-acetyl-3-methyl-5-(2-nitrophenyl)-6-phenyl-8-(m-tolyl)-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (3o)

15 h, light yellow solid; 63% yield, >99% ee;

HRMS (ESI+): calcd. for $C_{31}H_{27}NO_5 + Na^+$ [M+Na⁺] 516.1781, found 516.1777;

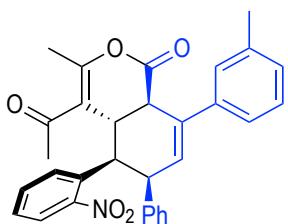
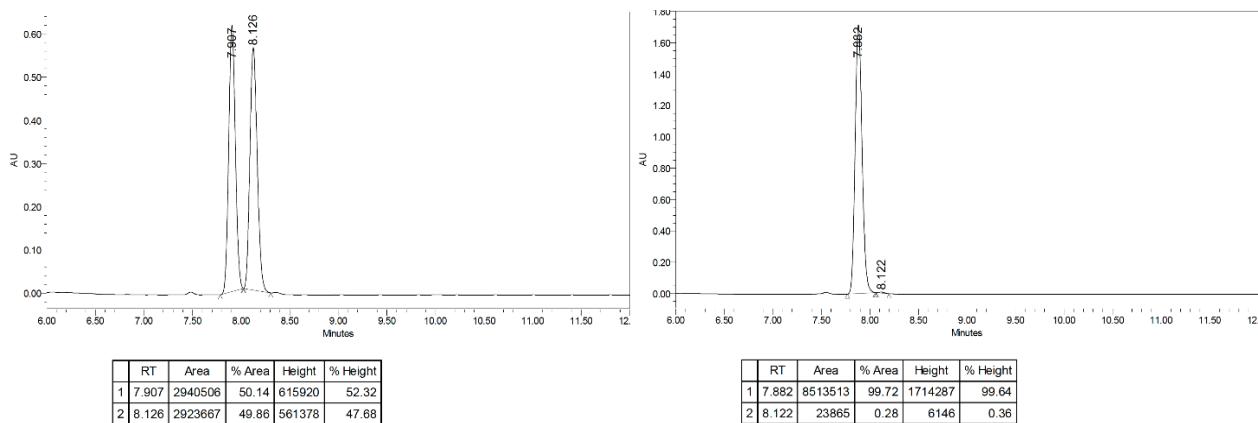
IR: ν_{max} (film, cm⁻¹): 3027, 1779, 1646, 1526, 1359, 1110, 702;

¹H NMR (400 MHz, CDCl₃) δ 7.61 – 7.53 (m, 2H), 7.42 – 7.38 (m, 1H), 7.30 (ddd, J = 8.3, 6.5, 2.1 Hz, 1H), 7.28 – 7.26 (m, 1H), 7.26 – 7.23 (m, 2H), 7.19 – 7.14 (m, 3H), 7.13 – 7.09 (m, 1H), 6.91 – 6.85 (m, 2H), 6.27 (d, J = 2.9 Hz, 1H), 3.95 – 3.81 (m, 3H), 3.49 (t, J = 11.0 Hz, 1H), 2.36 (s, 3H), 2.19 (s, 3H), 1.74 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 198.6, 167.6, 157.5, 151.8, 141.2, 139.9, 138.3, 134.0, 131.8, 130.1, 130.0, 128.8, 128.7, 128.6, 128.2, 128.0, 127.3, 126.7, 123.4, 123.1, 119.9, 51.1, 44.2, 43.3, 39.2, 30.2, 21.7, 19.3;

$[\alpha]^{25}_D$ = -85.6 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 7.9 min, t_R (minor) = 8.1 min.



(4aS,5R,6R,8aR)-4-acetyl-3-methyl-5-(2-nitrophenyl)-6-phenyl-8-(m-tolyl)-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (4o)

light yellow solid; 11% yield, 72:28 dr, >99% ee;

HRMS (ESI+): calcd. for $C_{31}H_{27}NO_5 + Na^+$ [M+Na⁺] 516.1781, found 516.1780;

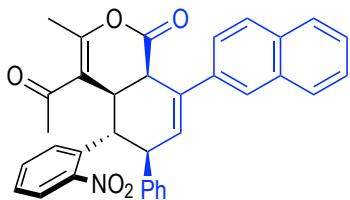
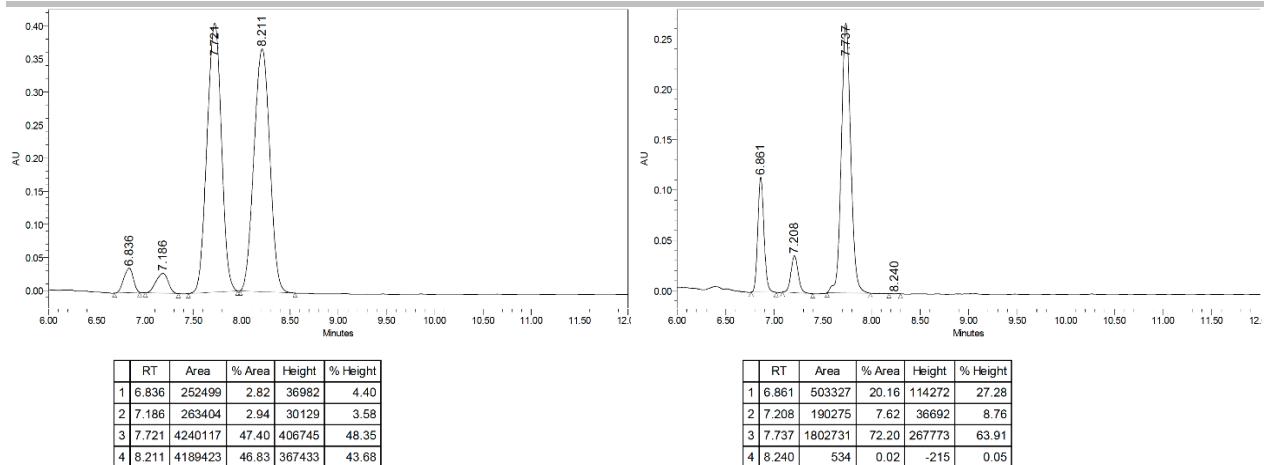
IR: ν_{max} (film, cm⁻¹): 2921, 1774, 1663, 1523, 1354, 1109, 699;

¹H NMR (600 MHz, CDCl₃) δ 7.85 (dd, J = 8.1, 1.4 Hz, 1H), 7.29 – 7.27 (m, 2H), 7.24 (d, J = 7.7 Hz, 1H), 7.21 – 7.09 (m, 5H), 7.04 (t, J = 7.7 Hz, 1H), 6.76 (br, 2H), 6.35 (dd, J = 6.2, 2.0 Hz, 1H), 6.05 (d, J = 7.9 Hz, 1H), 4.35 (t, J = 5.8 Hz, 1H), 4.26 (dd, J = 12.9, 5.4 Hz, 1H), 3.97 (dd, J = 12.4, 2.3 Hz, 1H), 3.56 – 3.49 (m, 1H), 2.37 (s, 3H), 1.95 (s, 3H), 1.54 (s, 3H);

¹³C NMR (150 MHz, CDCl₃) δ 201.3, 167.8, 151.3, 150.9, 140.9, 138.1, 138.0, 135.1, 133.2, 131.3, 130.9, 129.6, 128.4, 128.3, 128.2, 127.7, 127.5, 126.2, 124.7, 123.5, 122.5, 46.0, 45.3, 39.1, 33.8, 31.6, 21.7, 17.4;

$[\alpha]^{25}_D$ = -2.4 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-2, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 7.7 min, t_R (minor) = 8.2 min.



(4a*R*,5*S*,6*R*,8*aR*)-4-acetyl-3-methyl-8-(naphthalen-2-yl)-5-(2-nitrophenyl)-6-phenyl-4*a*,5,6,8*a*-tetrahydro-1*H*-isochromen-1-one (3p)

13 h, light yellow solid; 61% yield, 99% ee;

HRMS (ESI+): calcd. for $\text{C}_{34}\text{H}_{27}\text{NO}_5 + \text{Na}^+$ [M+Na $^+$] 552.1781, found 552.1779;

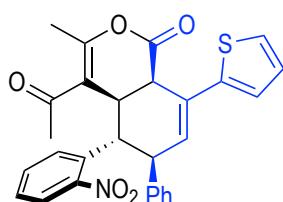
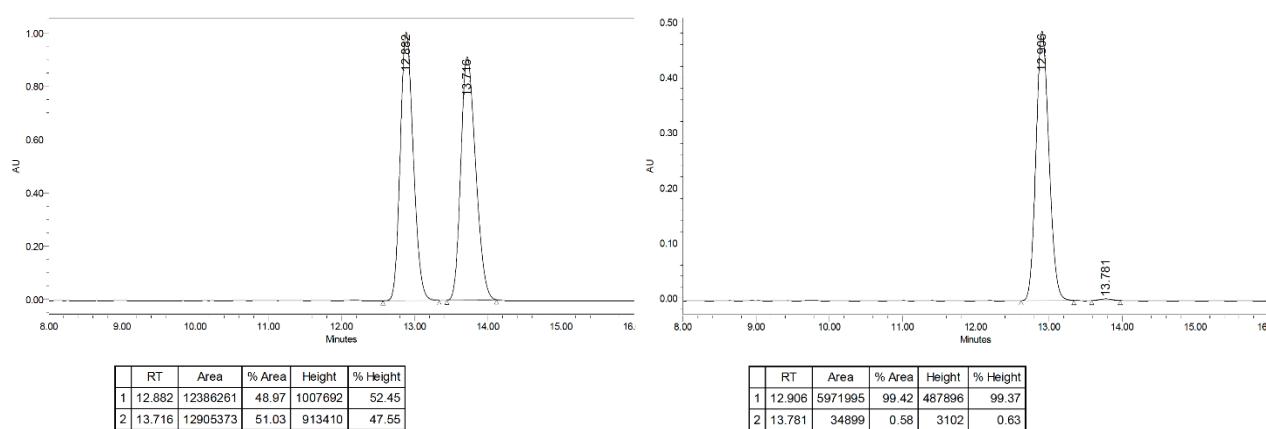
IR: ν_{max} (film, cm^{-1}): 3030, 1770, 1652, 1519, 1354, 1106, 704;

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.87 – 7.78 (m, 4H), 7.63 (dd, J = 8.7, 2.0 Hz, 1H), 7.61 – 7.54 (m, 2H), 7.50 – 7.44 (m, 2H), 7.41 (dd, J = 8.0, 1.3 Hz, 1H), 7.31 (ddd, J = 8.3, 6.8, 1.7 Hz, 1H), 7.22 – 7.15 (m, 3H), 6.95 – 6.88 (m, 2H), 6.43 (d, J = 2.9 Hz, 1H), 4.05 (dd, J = 5.4, 1.3 Hz, 1H), 3.98 (dd, J = 12.1, 5.3 Hz, 1H), 3.94 – 3.89 (m, 1H), 3.55 (dd, J = 12.1, 10.5 Hz, 1H), 2.21 (s, 3H), 1.76 (s, 3H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 198.6, 167.6, 157.6, 151.8, 141.1, 137.2, 134.0, 133.9, 133.5, 133.0, 131.9, 130.8, 130.0, 128.9, 128.4, 128.2, 128.1, 127.7, 127.4, 126.4, 126.2, 124.8, 124.2, 123.5, 119.9, 51.2, 44.2, 43.4, 39.3, 30.2, 19.3;

$[\alpha]^{25}_D$ = -74.4 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO_2 to CO_2/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO_2/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 12.9 min, t_R (minor) = 13.7 min.



(4aR,5S,6R,8aR)-4-acetyl-3-methyl-5-(2-nitrophenyl)-6-phenyl-8-(thiophen-2-yl)-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (3q)

15 h, light yellow solid; 36% yield, 98% ee;

HRMS (ESI+): calcd. for $C_{28}H_{23}NSO_5 + Na^+ [M+Na^+]$ 508.1189, found 508.1196;

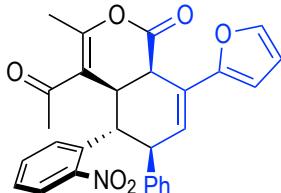
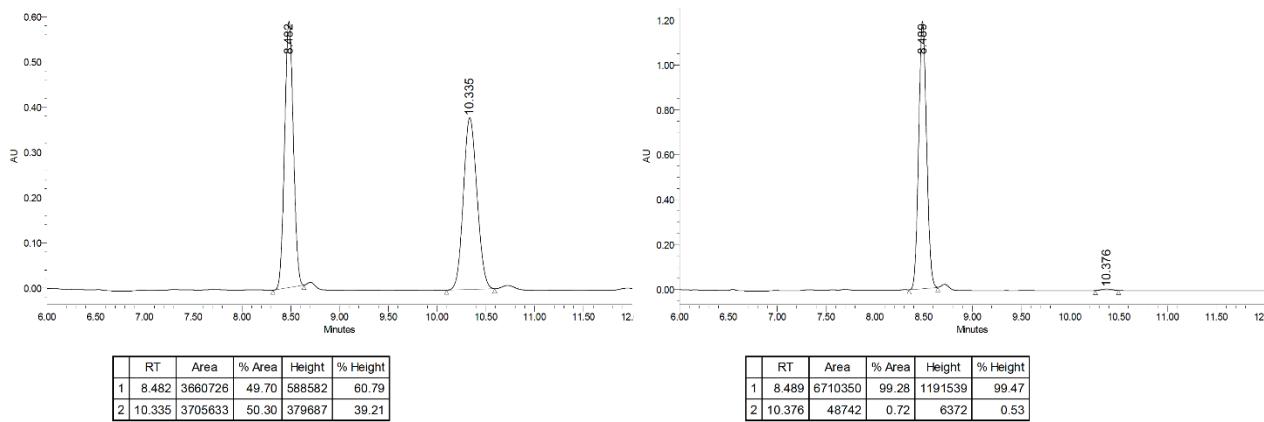
IR: ν_{max} (film, cm^{-1}): 3028, 1776, 1655, 1526, 1353, 1105, 701;

1H NMR (400 MHz, CDCl₃) δ 7.58 – 7.54 (m, 2H), 7.42 – 7.38 (m, 1H), 7.33 – 7.28 (m, 1H), 7.20 (dd, J = 4.4, 1.8 Hz, 1H), 7.17 – 7.13 (m, 3H), 7.02 – 6.98 (m, 2H), 6.87 – 6.82 (m, 2H), 6.34 (d, J = 2.9 Hz, 1H), 3.91 (dd, J = 12.1, 5.4 Hz, 1H), 3.88 – 3.82 (m, 2H), 3.48 (dd, J = 12.1, 10.4 Hz, 1H), 2.20 (s, 3H), 1.74 (s, 3H);

^{13}C NMR (100 MHz, CDCl₃) δ 198.4, 167.1, 157.5, 151.8, 144.1, 140.7, 133.7, 131.9, 130.0, 128.8, 128.6, 128.2, 128.1, 127.7, 127.4, 124.6, 123.7, 123.5, 119.7, 50.9, 44.2, 43.9, 38.9, 30.2, 19.3;

$[\alpha]^{25}_D$ = -180.0 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 8.5 min, t_R (minor) = 10.4 min.



(4aR,5S,6R,8aR)-4-acetyl-8-(furan-2-yl)-3-methyl-5-(2-nitrophenyl)-6-phenyl-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (3r)

15 h, light yellow solid; 34% yield, 98% ee;

HRMS (ESI+): calcd. for $C_{28}H_{23}NO_6 + Na^+ [M+Na^+]$ 492.1418, found 492.1422;

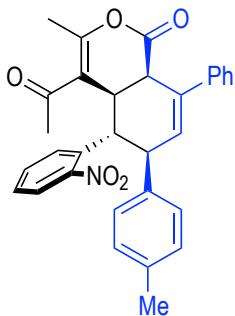
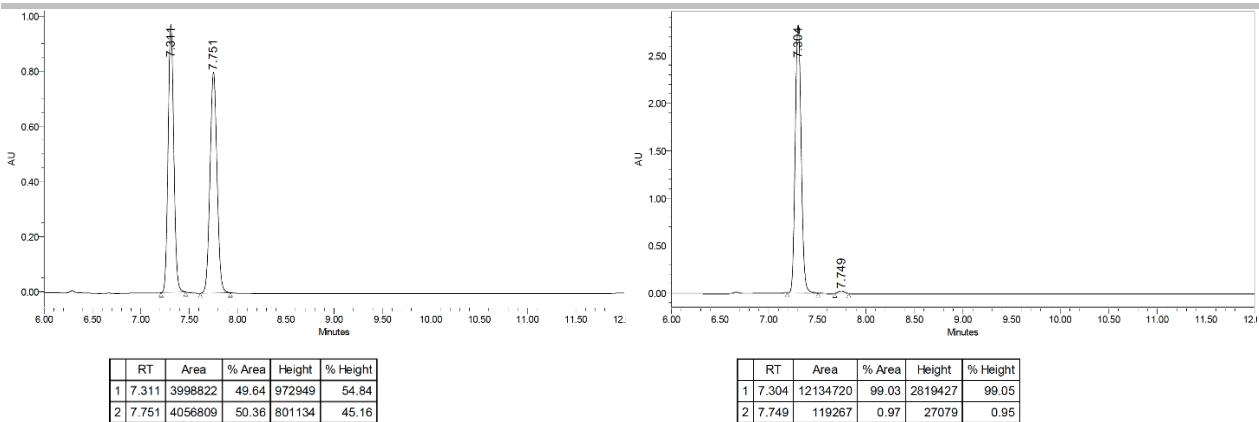
IR: ν_{max} (film, cm^{-1}): 3140, 1774, 1663, 1524, 1109, 759, 700;

1H NMR (400 MHz, CDCl₃) δ 7.55 (d, J = 4.1 Hz, 2H), 7.41 – 7.35 (m, 2H), 7.32 – 7.27 (m, 1H), 7.16 – 7.11 (m, 3H), 6.87 – 6.82 (m, 2H), 6.44 (d, J = 3.0 Hz, 1H), 6.42 (dd, J = 3.4, 1.8 Hz, 1H), 6.31 (d, J = 3.4 Hz, 1H), 3.90 – 3.84 (m, 2H), 3.81 (dd, J = 5.5, 1.4 Hz, 1H), 3.48 (dd, J = 12.0, 10.3 Hz, 1H), 2.19 (s, 3H), 1.73 (s, 3H);

^{13}C NMR (100 MHz, CDCl₃) δ 198.5, 167.0, 157.4, 153.2, 151.7, 142.1, 140.8, 133.8, 131.9, 130.0, 128.8, 128.1, 127.3, 126.8, 124.4, 123.5, 119.6, 111.6, 106.1, 50.6, 44.2, 41.4, 38.6, 30.2, 19.3;

$[\alpha]^{25}_D$ = -146.4 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 7.3 min, t_R (minor) = 7.7 min.



(4aR,5S,6R,8aR)-4-acetyl-3-methyl-5-(2-nitrophenyl)-8-phenyl-6-(*p*-tolyl)-4a,5,6,8a-tetrahydro-1*H*-isochromen-1-one (3s)

15 h, light yellow solid; 44% yield, 97:3 dr, 99% ee;

HRMS (ESI⁺): calcd. for $\text{C}_{31}\text{H}_{27}\text{NO}_5 + \text{Na}^+ [\text{M}+\text{Na}^+]$ 516.1781, found 516.1778;

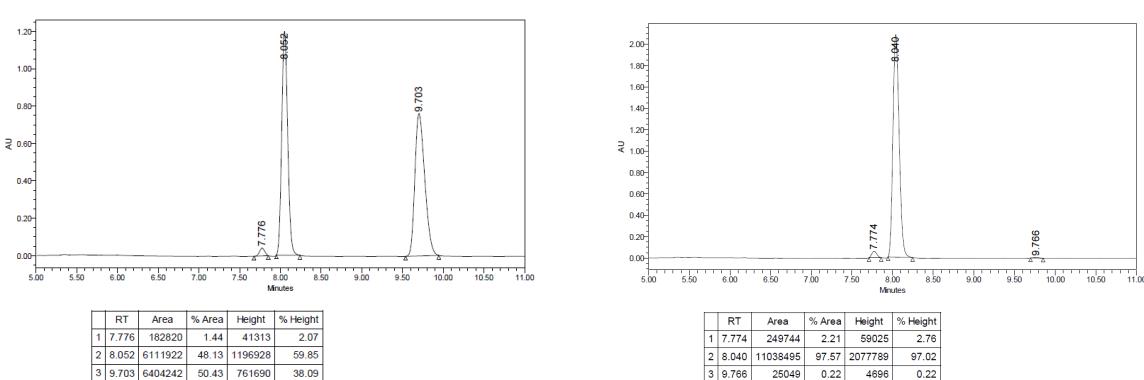
IR: ν_{max} (film, cm^{-1}): 3025, 1763, 1626, 1520, 1355, 1097, 742;

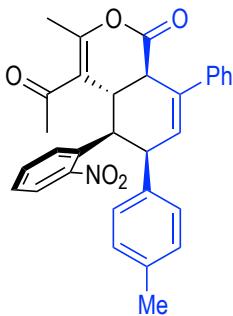
¹H NMR (400 MHz, CDCl₃) δ 7.60 – 7.52 (m, 2H), 7.47 – 7.40 (m, 3H), 7.38 – 7.27 (m, 4H), 6.99 – 6.94 (m, 2H), 6.81 – 6.75 (m, 2H), 6.26 (d, $J = 2.8$ Hz, 1H), 3.94 – 3.80 (m, 3H), 3.55 – 3.46 (m, 1H), 2.24 (s, 3H), 2.18 (s, 3H), 1.74 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 198.6, 167.6, 157.5, 151.8, 140.0, 138.0, 136.9, 134.2, 133.7, 131.9, 130.6, 130.0, 129.5, 128.7, 128.0, 127.8, 126.0, 123.5, 119.9, 50.6, 44.0, 43.4, 39.3, 30.2, 21.2, 19.3; (one aromatic carbon signal overlaps with others in the 120–135 ppm region)

$[\alpha]^{25}_{\text{D}} = -112.0$ ($c = 0.5$ in acetone, $\lambda = 589$ nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, $\lambda = 254$ nm), t_R (major) = 8.0 min, t_R (minor) = 9.7 min.





(4aS,5R,6R,8aR)-4-acetyl-3-methyl-5-(2-nitrophenyl)-8-phenyl-6-(p-tolyl)-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (4s)
light yellow solid; 7% yield, >99% ee;

HRMS (ESI+): calcd. for $C_{31}H_{27}NO_5 + Na^+$ [M+Na⁺] 516.1781, found 516.1779;

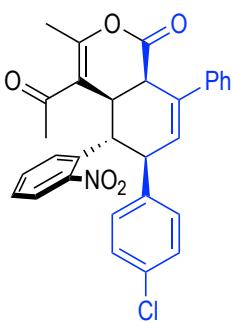
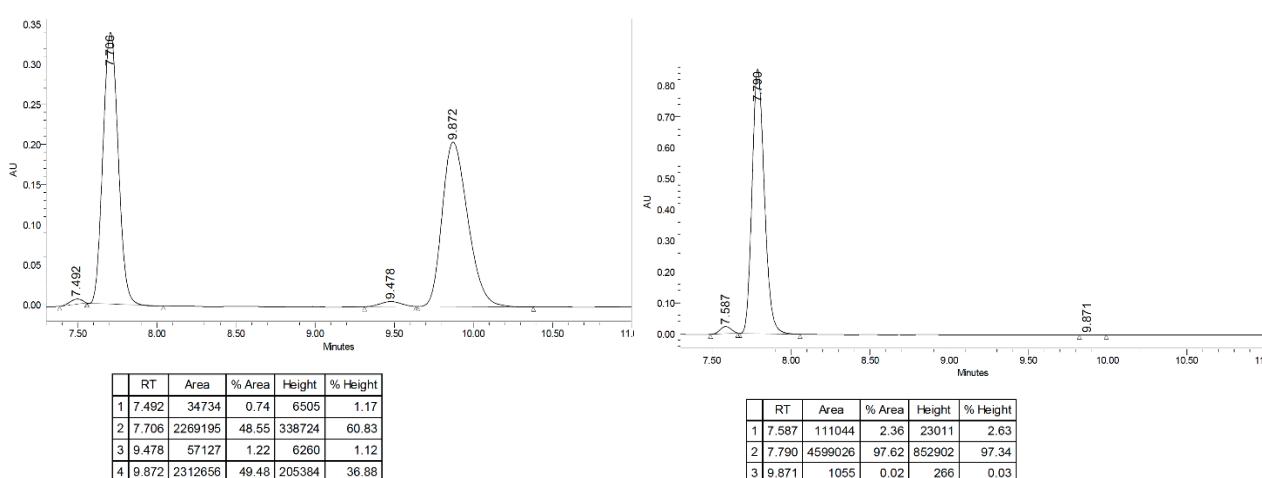
IR: ν_{max} (film, cm⁻¹): 2924, 1765, 1676, 1521, 1364, 1088, 726;

¹H NMR (400 MHz, CDCl₃) δ 7.85 (dd, J = 8.2, 1.4 Hz, 1H), 7.40 – 7.27 (m, 6H), 7.07 (td, J = 7.7, 1.4 Hz, 1H), 6.93 (d, J = 7.8 Hz, 2H), 6.64 (br, 2H), 6.35 (dd, J = 6.2, 2.0 Hz, 1H), 6.09 (d, J = 7.9 Hz, 1H), 4.31 (t, J = 5.5 Hz, 1H), 4.24 (dd, J = 12.7, 5.4 Hz, 1H), 3.97 (dt, J = 13.0, 1.7 Hz, 1H), 3.53 (td, J = 12.8, 2.0 Hz, 1H), 2.28 (s, 3H), 1.94 (d, J = 1.8 Hz, 3H), 1.54 (s, 3H);

¹³C NMR (150 MHz, CDCl₃) δ 201.3, 167.8, 151.3, 150.9, 141.0, 137.2, 135.2, 134.7, 132.9, 131.4, 129.8, 128.9, 128.6, 127.7, 127.4, 125.4, 124.6, 123.6, 45.7, 45.4, 39.2, 33.8, 31.6, 21.1, 17.4; (one aromatic carbon signal is overlap among 120–135 ppm)

$[\alpha]^{25}_D$ = -7.2 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 7.7 min, t_R (minor) = 9.9 min.



(4aR,5S,6R,8aR)-4-acetyl-6-(4-chlorophenyl)-3-methyl-5-(2-nitrophenyl)-8-phenyl-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (3t)
15 h, light yellow solid; 49% yield, 99% ee;

HRMS (ESI+): calcd. for $C_{30}H_{24}ClNO_5 + Na^+$ [M+Na⁺] 536.1235, found 536.1238;

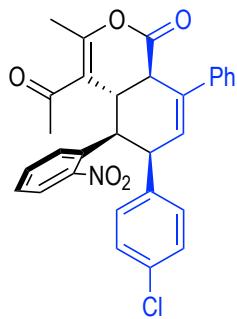
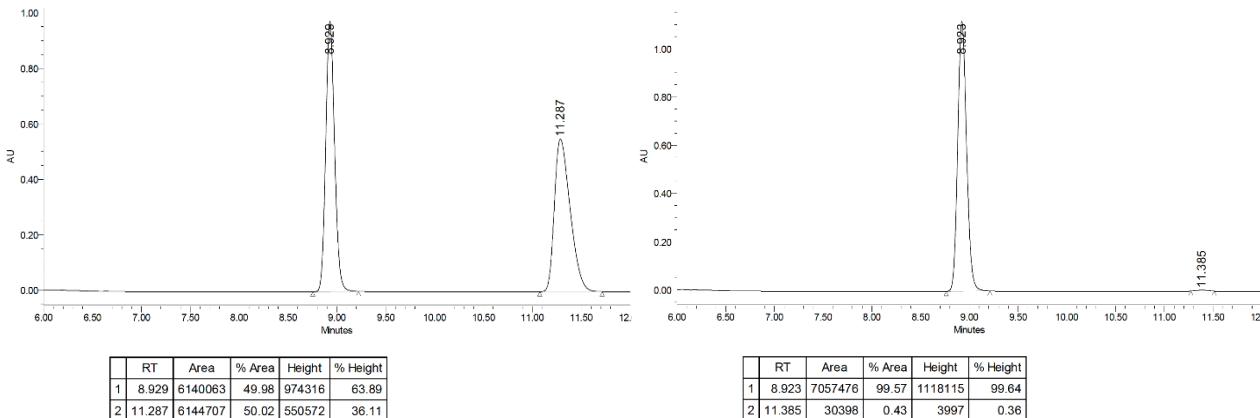
IR: ν_{max} (film, cm⁻¹): 3026, 1764, 1627, 1519, 1356, 1092, 743;

¹H NMR (400 MHz, CDCl₃) δ 7.56 (d, J = 3.9 Hz, 2H), 7.46 – 7.44 (m, 1H), 7.44 – 7.41 (m, 2H), 7.39 – 7.28 (m, 4H), 7.16 – 7.11 (m, 2H), 6.85 – 6.80 (m, 2H), 6.22 (d, J = 2.7 Hz, 1H), 3.91 (t, J = 6.1 Hz, 1H), 3.89 – 3.85 (m, 2H), 3.44 (dd, J = 11.7, 10.5 Hz, 1H), 2.19 (s, 3H), 1.74 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 198.4, 167.5, 157.8, 151.8, 139.8, 139.6, 134.5, 133.6, 133.2, 131.9, 129.9, 129.6, 129.5, 129.0, 128.8, 128.2, 128.0, 126.0, 123.6, 119.8, 50.3, 44.0, 43.4, 39.2, 30.2, 19.3;

$[\alpha]^{25}_D = -113.6$ ($c = 0.5$ in acetone, $\lambda = 589$ nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, $\lambda = 254$ nm), t_R (major) = 8.9 min, t_R (minor) = 11.3 min.



(4aS,5R,6R,8aR)-4-acetyl-6-(4-chlorophenyl)-3-methyl-5-(2-nitrophenyl)-8-phenyl-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (4t)

light yellow solid; 7% yield, >99% ee;

HRMS (ESI+): calcd. for C₃₀H₂₄CINO₅ +Na⁺ [M+Na⁺] 536.1235, found 536.1233;

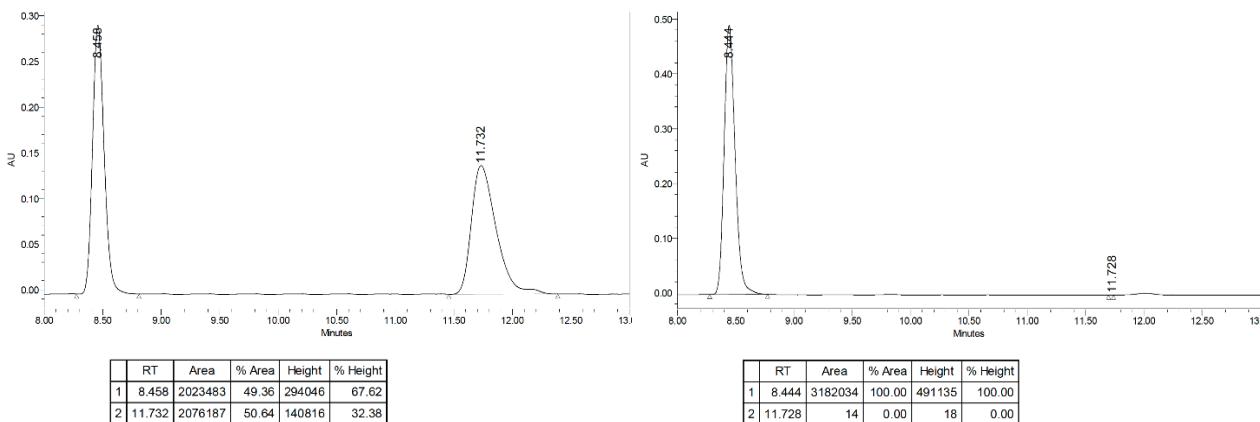
IR: ν_{max} (film, cm⁻¹): 3060, 1768, 1672, 1522, 1088, 728, 699;

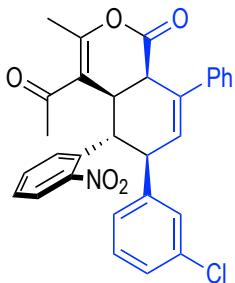
¹H NMR (400 MHz, CDCl₃) δ 7.87 (dd, $J = 8.1, 1.4$ Hz, 1H), 7.39 – 7.28 (m, 6H), 7.16 – 7.07 (m, 3H), 6.69 (br, 2H), 6.32 (dd, $J = 6.2, 2.0$ Hz, 1H), 6.12 (dd, $J = 8.0, 1.3$ Hz, 1H), 4.36 (td, $J = 5.9, 1.5$ Hz, 1H), 4.27 (dd, $J = 12.8, 5.4$ Hz, 1H), 3.97 (dt, $J = 13.0, 1.8$ Hz, 1H), 3.50 – 3.41 (m, 1H), 1.96 (d, $J = 1.8$ Hz, 3H), 1.55 (s, 3H);

¹³C NMR (150 MHz, CDCl₃) δ 201.2, 167.6, 151.2, 140.6, 136.6, 134.8, 133.7, 133.6, 131.6, 130.5, 129.5, 128.6, 128.3, 128.0, 127.7, 125.4, 124.9, 123.3, 45.4, 45.3, 39.0, 33.7, 31.6, 17.4;

$[\alpha]^{25}_D = -13.6$ ($c = 0.5$ in acetone, $\lambda = 589$ nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, $\lambda = 254$ nm), t_R (major) = 8.4 min, t_R (minor) = 11.7 min.





(4a*R*,5*S*,6*R*,8a*R*)-4-acetyl-6-(3-chlorophenyl)-3-methyl-5-(2-nitrophenyl)-8-phenyl-4a,5,6,8a-tetrahydro-1*H*-isochromen-1-one (3u)

15 h, light yellow solid; 46% yield, 99% ee;

HRMS (ESI+): calcd. for $C_{30}H_{24}ClNO_5 + Na^+$ [M+Na⁺] 536.1235, found 536.1232;

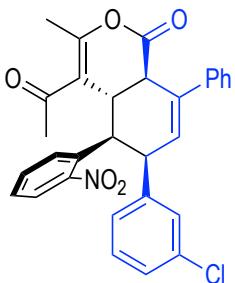
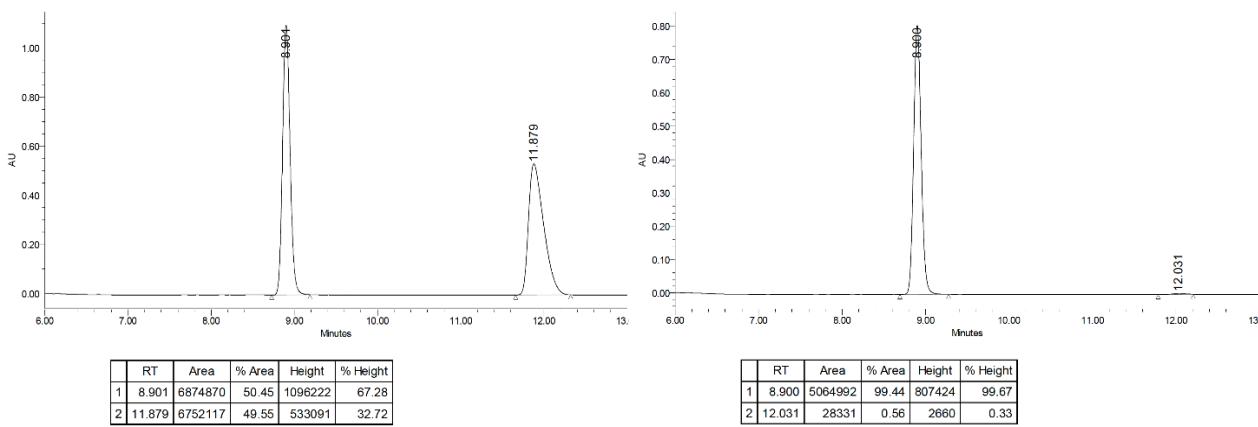
IR: ν_{max} (film, cm⁻¹): 3024, 1778, 1649, 1525, 1115, 763, 698;

¹H NMR (400 MHz, CDCl₃) δ 7.60 – 7.54 (m, 2H), 7.46 – 7.42 (m, 3H), 7.39 – 7.28 (m, 4H), 7.17 – 7.09 (m, 2H), 6.86 (td, J = 4.2, 1.7 Hz, 1H), 6.80 (d, J = 1.2 Hz, 1H), 6.23 (d, J = 2.7 Hz, 1H), 3.95 – 3.83 (m, 3H), 3.46 (t, J = 11.1 Hz, 1H), 2.19 (s, 3H), 1.75 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 198.3, 167.4, 157.8, 151.82, 143.1, 139.8, 134.6, 134.4, 133.4, 132.0, 130.3, 129.9, 129.3, 128.8, 128.3, 128.1, 127.6, 126.2, 126.0, 123.6, 119.7, 50.6, 43.8, 43.4, 39.2, 30.2, 19.3;

$[\alpha]^{25}_D$ = -131.2 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 8.9 min, t_R (minor) = 12.0 min.



(4a*S*,5*R*,6*R*,8a*R*)-4-acetyl-6-(3-chlorophenyl)-3-methyl-5-(2-nitrophenyl)-8-phenyl-4a,5,6,8a-tetrahydro-1*H*-isochromen-1-one (4u)

light yellow solid; 7% yield, 63:37 dr, >99% ee;

HRMS (ESI+): calcd. for $C_{30}H_{24}ClNO_5 + Na^+$ [M+Na⁺] 536.1235, found 536.1234;

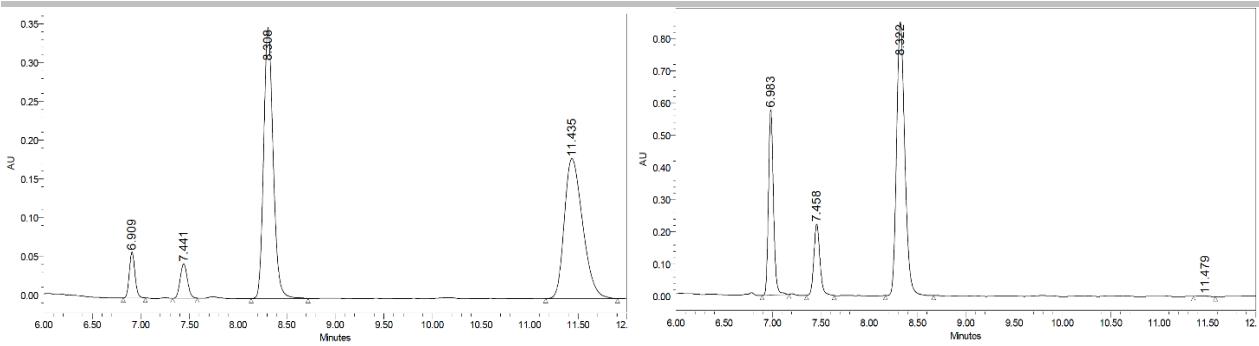
IR: ν_{max} (film, cm⁻¹): 2920, 1769, 1522, 1350, 1087, 979, 753;

¹H NMR (600 MHz, CDCl₃) δ 7.88 (d, J = 8.1 Hz, 1H), 7.38 – 7.35 (m, 3H), 7.32 (t, J = 7.6 Hz, 2H), 7.17 (d, J = 8.0 Hz, 1H), 7.14 – 7.06 (m, 2H), 6.78 – 6.54 (m, 3H), 6.32 (d, J = 6.0 Hz, 1H), 6.12 (d, J = 7.9 Hz, 1H), 4.35 (t, J = 6.0 Hz, 1H), 4.29 (dd, J = 13.0, 5.4 Hz, 1H), 3.97 (d, J = 13.0 Hz, 1H), 3.46 (t, J = 13.1 Hz, 1H), 1.95 (s, 3H), 1.55 (s, 3H);

¹³C NMR (150 MHz, CDCl₃) δ 201.2, 167.4, 151.3, 151.2, 140.7, 140.4, 134.8, 134.2, 134.0, 131.4, 130.2, 129.4, 129.4, 128.6, 128.1, 127.7, 127.7, 125.5, 124.9, 123.3, 45.8, 45.3, 39.1, 33.8, 31.6, 17.4;

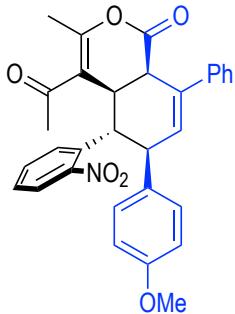
$[\alpha]^{25}_D$ = -3.2 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 8.3 min, t_R (minor) = 11.5 min.



	RT	Area	% Area	Height	% Height
1	6.909	236803	4.54	56989	9.31
2	7.441	229460	4.40	44524	7.03
3	8.308	2379980	45.60	349591	55.16
4	11.435	2373188	45.47	180683	28.51

	RT	Area	% Area	Height	% Height
1	6.983	2052260	25.51	576120	34.94
2	7.458	954317	11.86	221514	13.44
3	8.322	5027833	62.50	849506	51.53
4	11.479	9673	0.12	1512	0.09



(4a*R*,5*S*,6*R*,8a*R*)-4-acetyl-6-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-8-phenyl-4a,5,6,8a-tetrahydro-1*H*-isochromen-1-one (3v)

13 h, light yellow solid; 86% yield, >99% ee;

HRMS (ESI+): calcd. for $C_{31}H_{27}NO_6 + Na^+$ [M+Na⁺] 532.1731, found 532.1725;

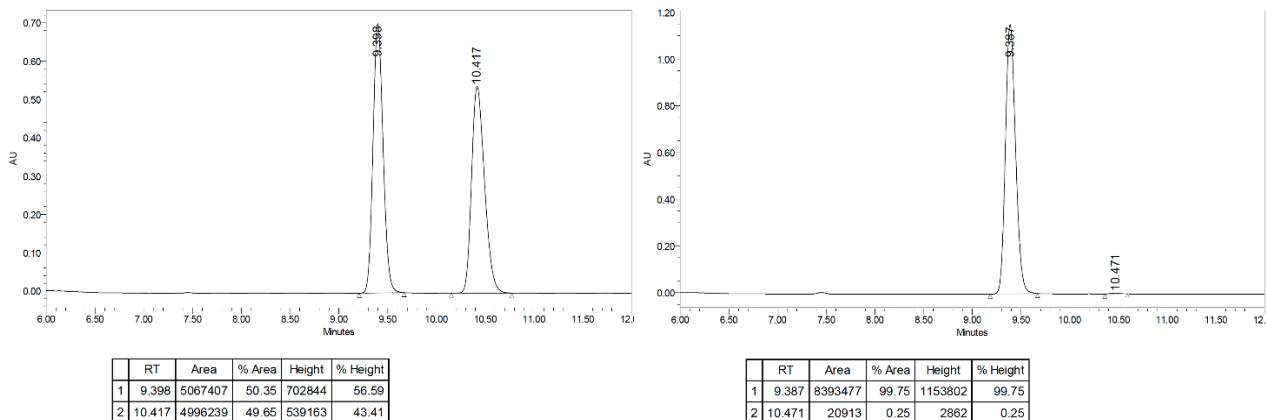
IR: ν_{max} (film, cm⁻¹): 2836, 1764, 1661, 1509, 1238, 1095, 759;

¹H NMR (400 MHz, CDCl₃) δ 7.59 – 7.52 (m, 2H), 7.47 – 7.40 (m, 3H), 7.38 – 7.27 (m, 4H), 6.83 – 6.77 (m, 2H), 6.72 – 6.66 (m, 2H), 6.26 (d, *J* = 2.9 Hz, 1H), 3.93 – 3.85 (m, 2H), 3.82 (dd, *J* = 10.3, 2.4 Hz, 1H), 3.72 (s, 3H), 3.48 (t, *J* = 10.8 Hz, 1H), 2.18 (s, 3H), 1.74 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 198.6, 167.6, 158.8, 157.5, 151.8, 140.0, 134.2, 133.7, 133.1, 131.9, 130.7, 130.0, 129.1, 128.7, 128.0, 127.8, 126.0, 123.5, 119.9, 114.2, 55.3, 50.2, 44.2, 43.4, 39.3, 30.2, 19.3;

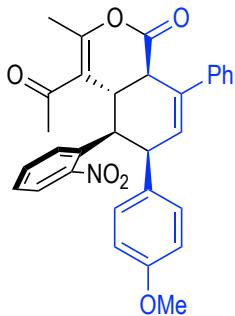
$[\alpha]^{25}_D$ = -108.0 (*c* = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), *t_R* (major) = 9.4 min, *t_R* (minor) = 10.5 min.



	RT	Area	% Area	Height	% Height
1	9.398	5067407	50.35	702844	56.59
2	10.417	4996239	49.65	539163	43.41

	RT	Area	% Area	Height	% Height
1	9.387	8393477	99.75	1153802	99.75
2	10.471	20913	0.25	2862	0.25



(4aS,5R,6R,8aR)-4-acetyl-6-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-8-phenyl-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (4v)

light yellow solid; 7% yield, >99% ee;

HRMS (ESI+): calcd. for $C_{31}H_{27}NO_6 + Na^+$ [M+Na⁺] 532.1731, found 532.1725;

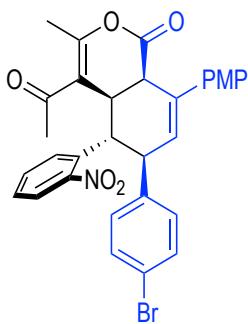
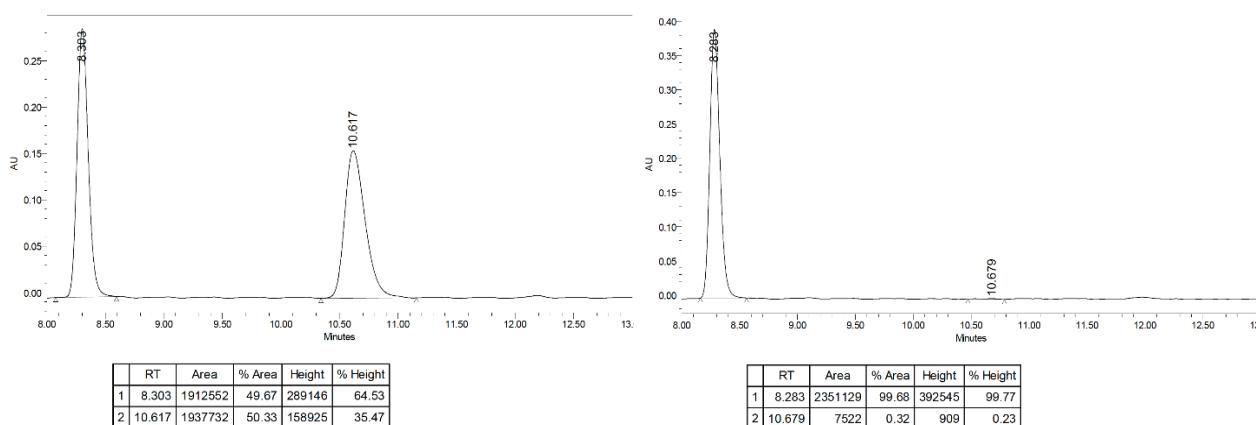
IR: ν_{max} (film, cm⁻¹): 3007, 1761, 1678, 1511, 1254, 1075, 728;

¹H NMR (400 MHz, CDCl₃) δ 7.85 (d, $J = 8.2$ Hz, 1H), 7.40 – 7.27 (m, 7H), 7.09 (t, $J = 7.6$ Hz, 1H), 6.67 (br, 3H), 6.35 (d, $J = 6.1$ Hz, 1H), 6.11 (d, $J = 7.9$ Hz, 1H), 4.30 (t, $J = 5.8$ Hz, 1H), 4.23 (dd, $J = 12.7, 5.3$ Hz, 1H), 4.03 – 3.90 (m, 1H), 3.76 (d, $J = 1.2$ Hz, 3H), 3.50 (t, $J = 12.8$ Hz, 1H), 1.94 (s, 3H), 1.55 (d, $J = 1.1$ Hz, 3H);

¹³C NMR (150 MHz, CDCl₃) δ 201.3, 167.8, 159.1, 151.3, 150.9, 140.9, 135.2, 132.8, 131.4, 131.4, 129.8, 128.6, 127.7, 127.5, 125.4, 124.6, 123.6, 113.5, 55.4, 45.3, 45.3, 39.2, 33.8, 31.6, 17.4;

$[\alpha]^{25}_D = -12.8$ ($c = 0.5$ in acetone, $\lambda = 589$ nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, $\lambda = 254$ nm), t_R (major) = 8.3 min, t_R (minor) = 10.7 min.



(4aR,5S,6R,8aR)-4-acetyl-6-(4-bromophenyl)-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (3w)

15 h, light yellow solid; 71% yield, 99% ee;

HRMS (ESI+): calcd. for $C_{31}H_{26}BrNO_6 + Na^+$ [M+Na⁺] 610.0836, found 610.0829;

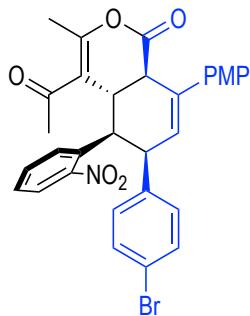
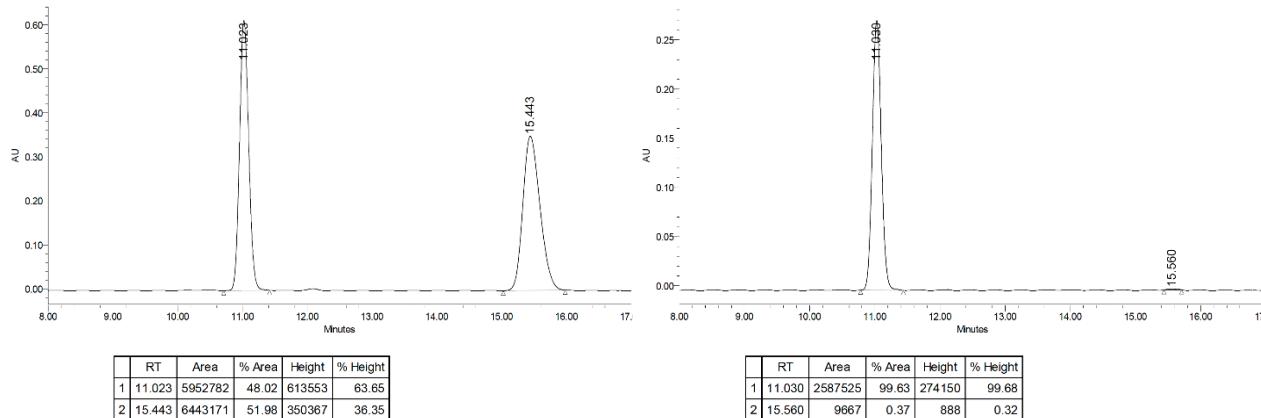
IR: ν_{max} (film, cm⁻¹): 2958, 1776, 1510, 1356, 1241, 1098, 823;

¹H NMR (400 MHz, CDCl₃) δ 7.59 – 7.52 (m, 2H), 7.43 (d, $J = 8.1$ Hz, 1H), 7.39 – 7.26 (m, 5H), 6.93 – 6.86 (m, 2H), 6.80 – 6.73 (m, 2H), 6.13 (d, $J = 2.8$ Hz, 1H), 3.91 – 3.82 (m, 3H), 3.81 (s, 3H), 3.41 (dd, $J = 12.0, 10.4$ Hz, 1H), 2.18 (s, 3H), 1.74 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 198.4, 167.6, 159.5, 157.7, 151.8, 140.4, 133.8, 133.6, 132.3, 131.9, 129.9, 129.8, 128.2, 127.9, 127.1, 123.6, 121.2, 119.8, 114.1, 55.5, 50.4, 43.9, 43.4, 39.2, 30.2, 19.3;

$[\alpha]^{25}_D = -99.2$ ($c = 0.5$ in acetone, $\lambda = 589$ nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 11.0 min, t_R (minor) = 15.5 min.



(4aS,5R,6R,8aR)-4-acetyl-6-(4-bromophenyl)-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-4a,5,6,8a-tetrahydro-1H-isochromen-1-one (4w)

light yellow solid; 6% yield, >99% ee;

HRMS (ESI+): calcd. for C₃₁H₂₆BrNO₆ +Na⁺ [M+Na⁺] 610.0836, found 610.0831;

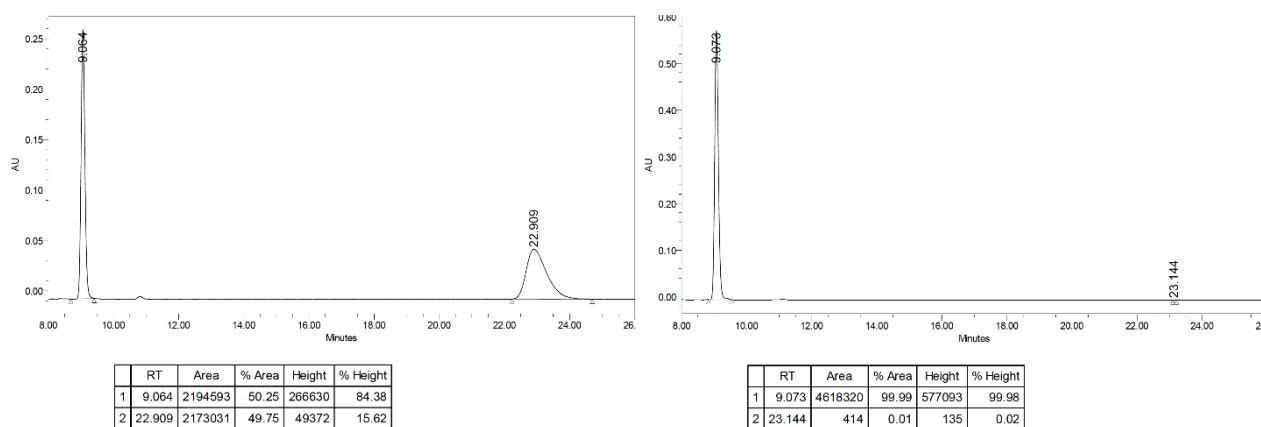
IR: ν_{max} (film, cm⁻¹): 2928, 1778, 1673, 1511, 1349, 1072, 822, 732;

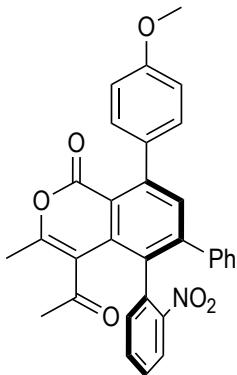
¹H NMR (400 MHz, CDCl₃) δ 7.86 (dd, J = 8.1, 1.4 Hz, 1H), 7.33 – 7.27 (m, 4H), 7.25 – 7.23 (m, 1H), 7.14 (td, J = 7.7, 1.4 Hz, 1H), 6.95 – 6.85 (m, 2H), 6.62 (br, 2H), 6.25 (dd, J = 6.1, 2.0 Hz, 1H), 6.11 (dd, J = 8.0, 1.3 Hz, 1H), 4.37 – 4.30 (m, 1H), 4.26 (dd, J = 12.7, 5.4 Hz, 1H), 3.92 (dt, J = 13.0, 1.7 Hz, 1H), 3.83 (s, 3H), 3.43 (ddd, J = 14.8, 11.9, 2.0 Hz, 1H), 1.95 (d, J = 1.8 Hz, 3H), 1.55 (s, 3H).

¹³C NMR (150 MHz, CDCl₃) δ 201.2, 167.7, 159.2, 151.3, 151.2, 137.2, 134.8, 133.1, 132.2, 131.6, 131.2, 129.5, 129.1, 128.0, 126.4, 124.8, 123.4, 121.6, 114.0, 55.4, 45.5, 45.3, 38.9, 33.8, 31.5, 17.4.

$[\alpha]^{25}_D$ = +27.2 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 9.1 min, t_R (minor) = 23.1 min.





(S)-4-acetyl-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-6-phenyl-1*H*-isochromen-1-one (S-6)

light yellow solid; 80% yield, 83:17 er;

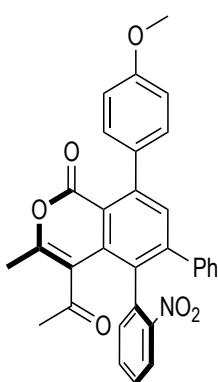
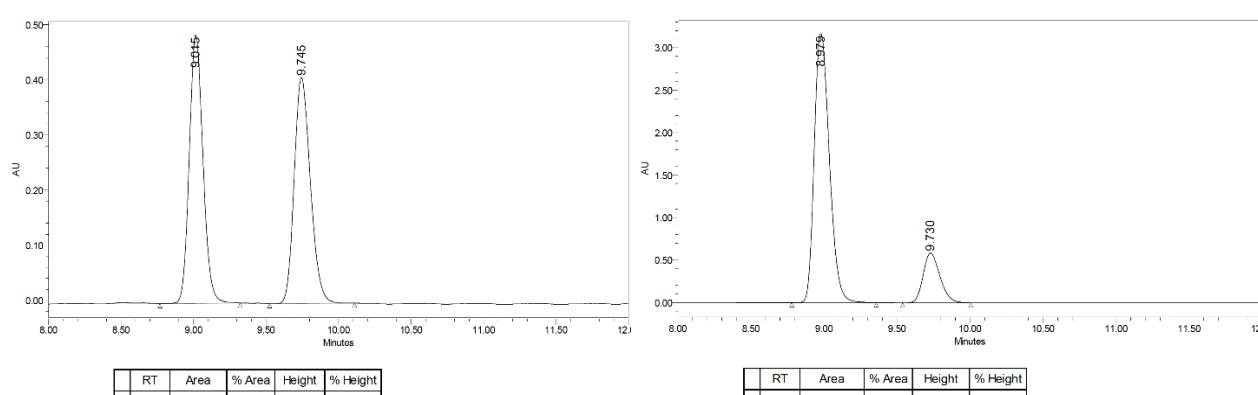
HRMS (ESI+): calcd. for $C_{31}H_{23}NO_6 + H^+$ [M+H⁺] 506.1598, found 506.1598;

IR: ν_{max} (film, cm⁻¹): 2840, 1738, 1692, 1515, 1337, 1040, 700;

¹H NMR (400 MHz, CDCl₃) δ 7.87 (dd, J = 8.2, 1.4 Hz, 1H), 7.57 (td, J = 7.5, 1.4 Hz, 1H), 7.51 – 7.45 (m, 1H), 7.42 (s, 1H), 7.39 – 7.32 (m, 3H), 7.20 – 7.08 (m, 3H), 7.00 – 6.95 (m, 2H), 6.89 – 6.85 (m, 2H), 3.87 (s, 3H), 2.17 (s, 3H), 1.83 (s, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 201.4, 159.5, 159.5, 153.0, 149.5, 147.5, 146.3, 139.1, 137.0, 134.8, 134.1, 133.6, 133.0, 132.6, 131.6, 130.3, 129.9, 129.1, 128.1, 127.8, 125.0, 118.9, 117.7, 113.6, 55.4, 32.0, 18.2;

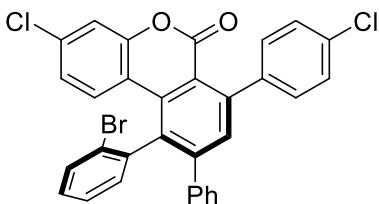
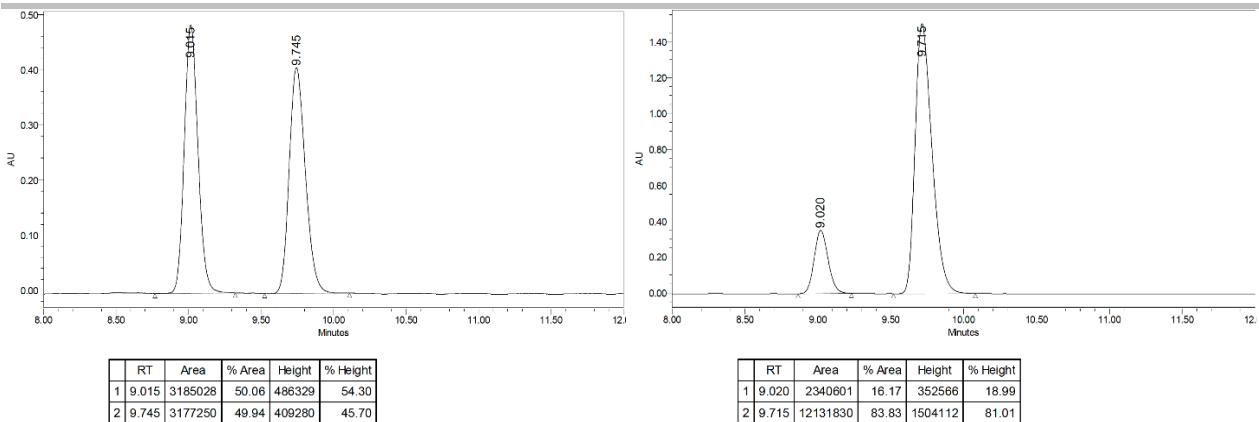
the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 9.0 min, t_R (minor) = 9.7 min.



(R)-4-acetyl-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-6-phenyl-1*H*-isochromen-1-one (R-6)

light yellow solid; 80% yield, 84:16 er;

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 9.7 min, t_R (minor) = 9.0 min.



(R)-10-(2-bromophenyl)-3-chloro-7-(4-chlorophenyl)-9-phenyl-6H-benzo[c]chromen-6-one (8)

yellow solid; 65% yield, 94% ee;

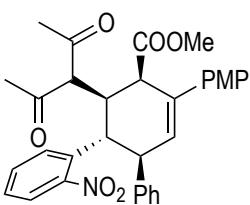
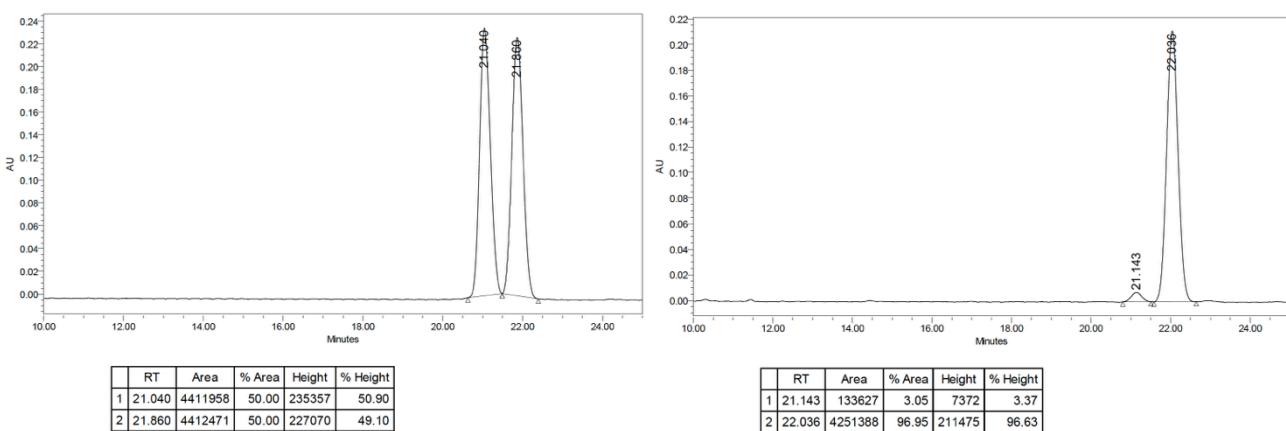
the enantioselectivity was determined by UPCC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 80:20 in the first 5 mins and maintaining in 80:20 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 22.0 min, t_R (minor) = 21.1 min; [α]_D²⁰ = -0.60 (c = 0.67 in acetone, λ = 589 nm);

HRMS (ESI+): calcd. for C₃₁H₁₇BrCl₂O₂+Na⁺ [M+Na⁺] 592.9681, found 592.9656;

IR: ν_{max} (film, cm⁻¹): 3061, 1734, 1604, 1397, 1088, 1067, 737, 705;

¹H NMR (400 MHz, CDCl₃) δ 7.56 (dd, J = 8.0, 1.3 Hz, 1H), 7.48 (s, 1H), 7.41 (d, J = 8.6 Hz, 2H), 7.38 – 7.34 (m, 2H), 7.34 – 7.31 (m, 1H), 7.27 (s, 1H), 7.23 – 7.13 (m, 5H), 7.11 – 7.06 (m, 2H), 6.78 (d, J = 1.9 Hz, 2H);

¹³C NMR (101 MHz, CDCl₃) δ 159.2, 151.8, 148.6, 145.5, 140.5, 140.4, 139.6, 135.9, 135.7, 134.6, 134.5, 133.8, 133.5, 132.7, 130.0, 129.8, 129.1, 128.4, 128.2, 127.9, 127.6, 125.0, 124.1, 118.8, 117.7, 117.5.



methyl (1'R,2'R,3'S,4'R)-3'-(2,4-dioxopentan-3-yl)-5'-(4-methoxyphenyl)-2''-nitro-1',2',3',4'-tetrahydro-[1,1':2',1"-terphenyl]-4'-carboxylate (9)

light yellow solid; 70% yield, 99% ee;

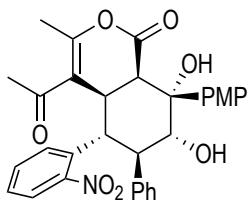
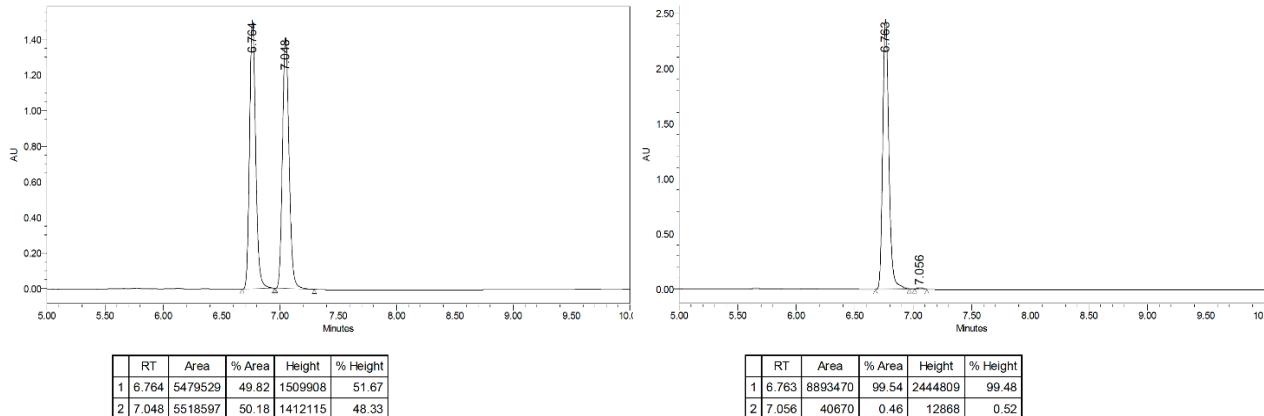
HRMS (ESI+): calcd. for $C_{32}H_{31}NO_7 + Na^+ [M+Na^+]$ 564.1993, found 564.1986;

IR: ν_{max} (film, cm^{-1}): 2955, 1733, 1536, 1233, 1159, 767, 708;

1H NMR (600 MHz, CDCl₃) δ 7.51 (t, J = 7.6 Hz, 1H), 7.46 – 7.43 (m, 1H), 7.35 (d, J = 8.1 Hz, 1H), 7.31 – 7.26 (m, 3H), 7.14 – 7.07 (m, 3H), 6.86 – 6.82 (m, 4H), 6.07 (d, J = 2.7 Hz, 1H), 4.55 (d, J = 10.4 Hz, 1H), 4.23 (d, J = 4.4 Hz, 1H), 3.85 (t, J = 11.0 Hz, 1H), 3.80 (s, 3H), 3.77 (s, 3H), 3.75 – 3.67 (m, 2H), 2.30 (s, 3H), 1.50 (s, 3H);

^{13}C NMR (150 MHz, CDCl₃) δ 202.6, 201.5, 172.8, 159.4, 152.5, 142.2, 134.7, 134.5, 132.9, 132.4, 131.7, 128.7, 128.5, 128.2, 128.0, 127.3, 127.1, 123.1, 114.0, 71.0, 55.4, 52.9, 52.4, 45.8, 43.3, 42.8, 31.7, 30.4;

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 6.8 min, t_R (minor) = 7.0 min.



(4aR,5R,6R,8R,8aR)-4-acetyl-7,8-dihydroxy-8-(4-methoxyphenyl)-3-methyl-5-(2-nitrophenyl)-6-phenyl-4a,5,6,7,8,8a-hexahydro-1H-isochromen-1-one (10)

light yellow solid; 78% yield, >99% ee;

HRMS (ESI+): calcd. for $C_{31}H_{29}NO_8 + Na^+ [M+Na^+]$ 566.1785, found 566.1778;

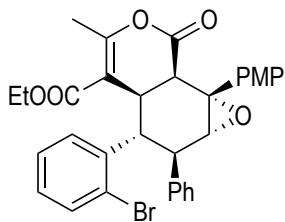
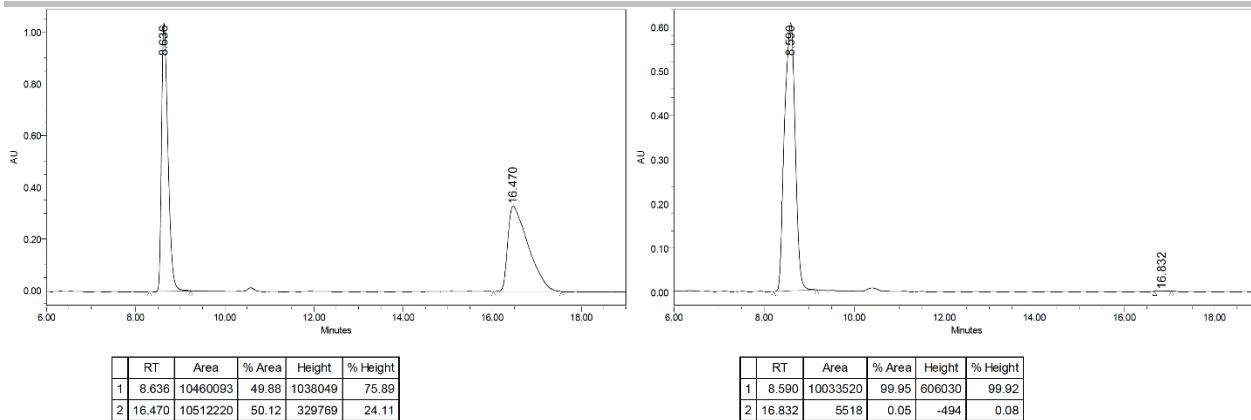
IR: ν_{max} (film, cm^{-1}): 3460, 2932, 1783, 1649, 1524, 1357, 1250, 1036, 699;

1H NMR (400 MHz, Acetone) δ 7.80 (dd, J = 8.0, 1.3 Hz, 1H), 7.69 – 7.64 (m, 2H), 7.55 (td, J = 7.7, 1.4 Hz, 1H), 7.47 (dd, J = 8.2, 1.4 Hz, 1H), 7.31 – 7.25 (m, 1H), 7.17 – 7.08 (m, 4H), 7.06 – 7.00 (m, 1H), 6.87 – 6.81 (m, 2H), 4.96 (dd, J = 9.6, 6.4 Hz, 1H), 4.30 (d, J = 1.3 Hz, 1H), 4.22 (dd, J = 11.8, 5.0 Hz, 1H), 3.90 (dd, J = 6.5, 1.7 Hz, 1H), 3.82 (d, J = 12.0 Hz, 1H), 3.77 (s, 3H), 3.65 (dd, J = 12.0, 9.6 Hz, 1H), 3.49 (d, J = 4.9 Hz, 1H), 2.82 (d, J = 1.5 Hz, 1H), 2.06 (s, 3H), 1.79 (s, 3H).

^{13}C NMR (100 MHz, Acetone) δ 197.1, 167.0, 159.5, 158.4, 151.8, 141.6, 137.5, 135.1, 133.0, 131.2, 130.2, 129.9, 128.9, 128.5, 127.2, 124.6, 120.2, 113.3, 75.5, 72.6, 55.4, 53.5, 52.9, 43.2, 38.2, 18.8.

$[\alpha]^{25}_D$ = +179.2 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 8.6 min, t_R (minor) = 16.8 min.



ethyl (1aR,2R,3R,3aR,7aR,7bR)-3-(2-bromophenyl)-7b-(4-methoxyphenyl)-5-methyl-7-oxo-2-phenyl-1a,3,3a,7,7a,7b-hexahydro-2H-oxireno[2,3-h]isochromene-4-carboxylate (11)

light yellow solid; 60% yield, 95% ee;

HRMS (ESI+): calcd. for $C_{32}H_{29}BrO_6 + Na^+$ [M+Na⁺] 611.1040, found 611.1030;

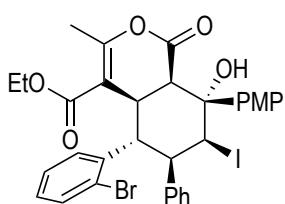
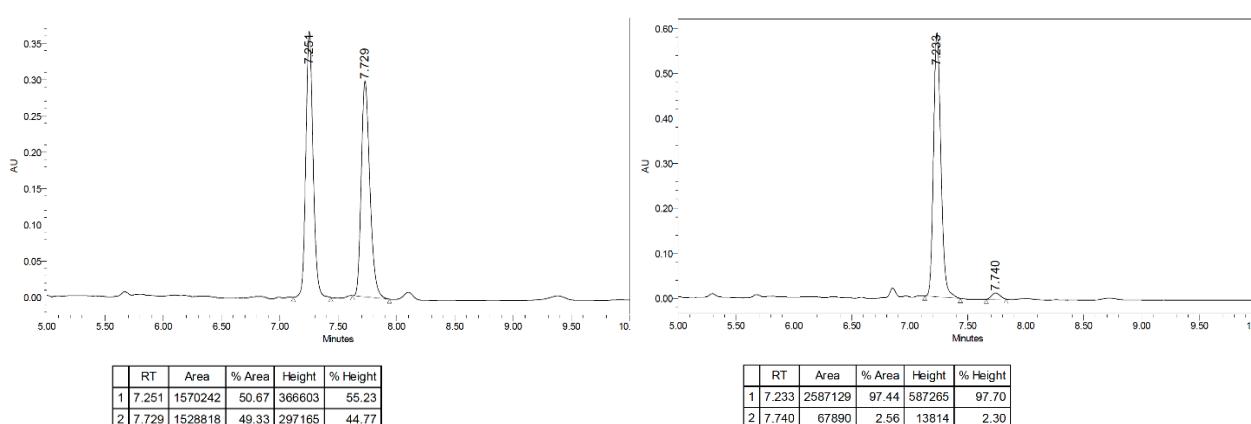
IR: ν_{max} (film, cm⁻¹): 2930, 1773, 1697, 1516, 1248, 1022, 759;

¹H NMR (400 MHz, Acetone) δ 7.64 (dd, J = 7.9, 1.6 Hz, 1H), 7.59 – 7.53 (m, 2H), 7.41 (td, J = 7.6, 1.3 Hz, 1H), 7.30 (dd, J = 8.1, 1.3 Hz, 1H), 7.27 – 7.17 (m, 3H), 7.13 – 7.05 (m, 3H), 6.96 – 6.91 (m, 2H), 4.17 (dd, J = 4.8, 0.8 Hz, 1H), 3.86 – 3.80 (m, 4H), 3.79 – 3.75 (m, 2H), 3.69 (d, J = 10.3 Hz, 1H), 3.64 – 3.56 (m, 1H), 3.43 (dd, J = 12.4, 10.4 Hz, 1H), 2.17 (s, 3H), 1.11 (t, J = 7.1 Hz, 3H);

¹³C NMR (100 MHz, Acetone) δ 168.1, 165.7, 161.3, 160.6, 141.9, 140.2, 133.0, 132.0, 131.3, 130.1, 129.5, 129.3, 129.2, 128.1, 127.8, 127.5, 114.3, 110.6, 62.3, 62.1, 61.0, 55.6, 49.8, 48.1, 45.2, 35.4, 18.4, 14.3;

$[\alpha]^{25}_D$ = -18.4 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 7.2 min, t_R (minor) = 7.7 min.



ethyl 2-((4a*R*,5*R*,6*R*,7*S*,8*R*,8a*R*)-5-(2-bromophenyl)-8-hydroxy-7-iodo-8-(4-methoxyphenyl)-3-methyl-1-oxo-6-phenyl-4a,5,6,7,8,8a-hexahydro-1*H*-isochromen-4-yl)-2-oxoacetate (12)

light yellow solid; 88% yield, 85:15 dr, 93% ee;

HRMS (ESI+): calcd. for $C_{32}H_{30}IO_6Br + Na^+$ [M+Na⁺] 739.0163, found 739.0141;

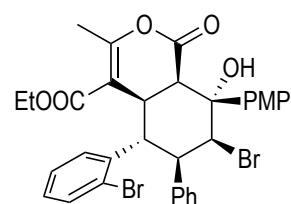
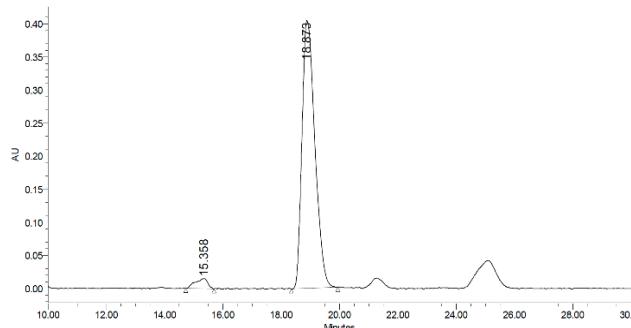
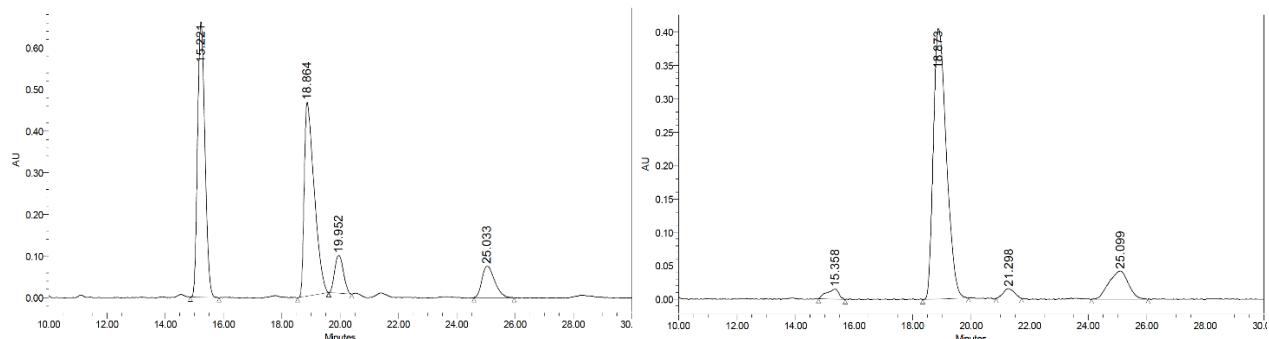
IR: ν_{max} (film, cm⁻¹): 3480, 2899, 1764, 1663, 1518, 1251, 1023, 758;

¹H NMR (400 MHz, Acetone) δ 7.65 – 7.59 (m, 2H), 7.45 (dd, J = 8.1, 1.3 Hz, 1H), 7.33 (dd, J = 7.9, 1.7 Hz, 1H), 7.17 – 7.08 (m, 5H), 7.07 – 7.02 (m, 1H), 6.98 – 6.92 (m, 1H), 6.90 – 6.84 (m, 2H), 5.19 (s, 1H), 5.10 (ddd, J = 9.6, 3.1, 1.4 Hz, 1H), 4.19 – 4.11 (m, 3H), 4.08 – 4.03 (m, 1H), 3.81 (s, 3H), 3.68 (dq, J = 10.9, 7.1 Hz, 1H), 3.47 (dq, J = 10.9, 7.1 Hz, 1H), 2.24 (s, 3H), 1.07 (t, J = 7.1 Hz, 3H).

¹³C NMR (100 MHz, Acetone) δ 166.2, 165.5, 160.7, 159.9, 143.3, 140.5, 136.9, 133.2, 131.5, 130.6, 128.8, 128.6, 128.2, 128.0, 127.1, 113.50, 110.2, 75.8, 60.7, 55.5, 49.0, 47.2, 46.9, 44.9, 44.8, 39.4, 18.8, 14.3.

$[\alpha]^{25}_D$ = +226.4 (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO₂ to CO₂/MeOH = 80:20 in the first 5 mins and maintaining in 80:20 CO₂/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_R (major) = 18.9 min, t_R (minor) = 15.3 min.



ethyl (4a*R*,5*R*,6*R*,7*S*,8*R*,8a*R*)-7-bromo-5-(2-bromophenyl)-8-hydroxy-8-(4-methoxyphenyl)-3-methyl-1-oxo-6-phenyl-4a,5,6,7,8,8a-hexahydro-1*H*-isochromene-4-carboxylate (13)

light yellow solid; 57% yield, 93% ee;

HRMS (ESI+): calcd. for $C_{32}H_{30}Br_2O_6 + Na^+$ [M+Na⁺] 691.0301, found 691.0294;

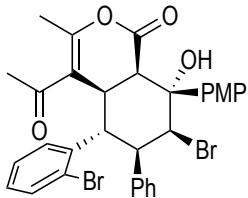
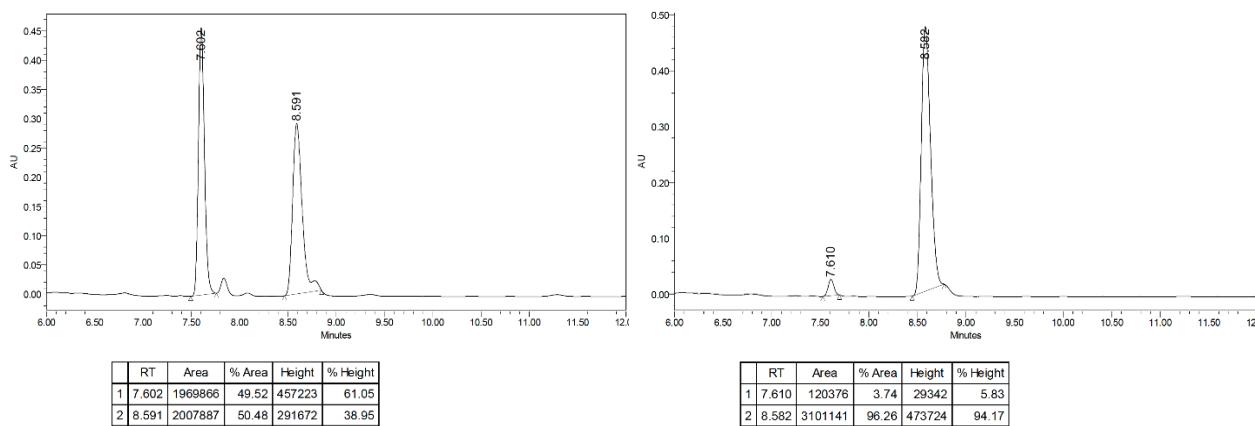
IR: ν_{max} (film, cm⁻¹): 3481, 2895, 1769, 1706, 1250, 1061, 1025, 760;

¹H NMR (400 MHz, Acetone) δ 7.65 – 7.60 (m, 2H), 7.43 (dd, J = 8.1, 1.3 Hz, 1H), 7.35 (dd, J = 7.9, 1.7 Hz, 1H), 7.24 – 7.19 (m, 2H), 7.15 – 7.08 (m, 3H), 7.07 – 7.01 (m, 1H), 6.95 (ddd, J = 8.8, 7.3, 1.7 Hz, 1H), 6.90 – 6.85 (m, 2H), 5.20 (s, 1H), 5.01 (d, J = 2.7 Hz,

1H), 4.59 (dd, J = 11.7, 2.9 Hz, 1H), 4.34 – 4.25 (m, 1H), 4.15 – 4.08 (m, 2H), 3.81 (s, 3H), 3.68 (dq, J = 10.9, 7.1 Hz, 1H), 3.46 (dq, J = 10.9, 7.1 Hz, 1H), 2.23 (s, 3H), 1.07 (t, J = 7.1 Hz, 3H);

^{13}C NMR (100 MHz, Acetone) δ 166.2, 165.1, 160.8, 159.9, 141.9, 140.4, 136.4, 133.2, 131.2, 130.7, 128.9, 128.5, 128.3, 128.2, 128.0, 127.1, 113.7, 110.3, 76.3, 62.8, 60.7, 55.5, 48.6, 47.3, 42.8, 39.1, 18.9, 14.3; $[\alpha]^{25}_{\text{D}} = +179.2$ (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO_2 to CO_2/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO_2/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_{R} (major) = 8.6 min, t_{R} (minor) = 7.6 min.



(4a*R*,5*R*,6*R*,7*S*,8*R*,8a*R*)-4-acetyl-7-bromo-5-(2-bromophenyl)-8-hydroxy-8-(4-methoxyphenyl)-3-methyl-6-phenyl-4a,5,6,7,8,8a-hexahydro-1*H*-isochromen-1-one (14)

light yellow solid; 86% yield, 98% ee;

HRMS (ESI+): calcd. for $\text{C}_{31}\text{H}_{28}\text{Br}_2\text{O}_6 + \text{Na}^+$ $[\text{M}+\text{Na}^+]$ 661.0196, found 661.0187;

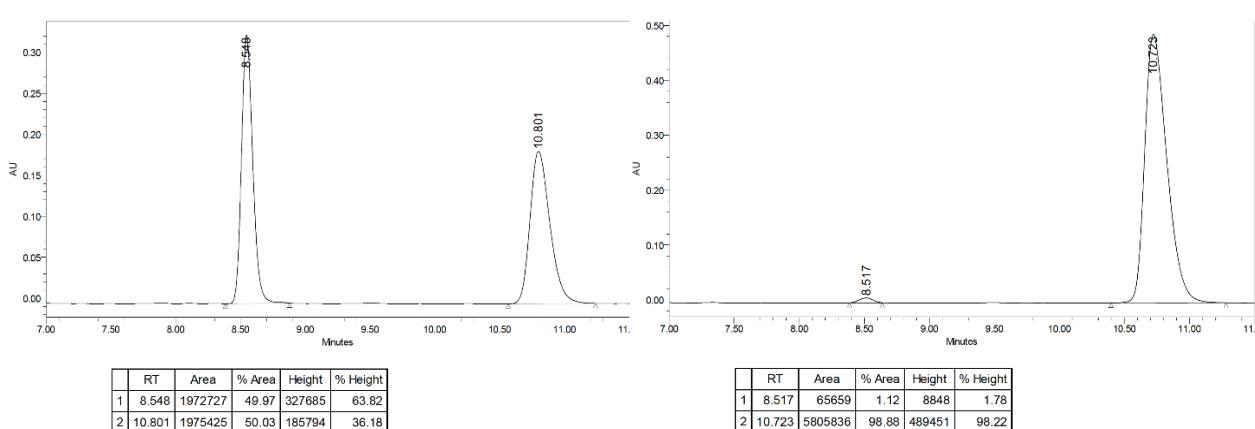
IR: ν_{max} (film, cm^{-1}): 3480, 2899, 1762, 1664, 1252, 1105, 1029, 759;

^1H NMR (600 MHz, MeOD) δ 7.62 – 7.57 (m, 2H), 7.42 (dd, J = 8.1, 1.3 Hz, 1H), 7.24 (dd, J = 7.9, 1.7 Hz, 1H), 7.18 – 7.15 (m, 2H), 7.09 (t, J = 7.6 Hz, 2H), 7.07 – 7.01 (m, 2H), 6.96 – 6.93 (m, 1H), 6.93 – 6.89 (m, 2H), 4.90 – 4.88 (br, 1H), 4.49 (dd, J = 11.7, 2.8 Hz, 1H), 4.26 (t, J = 11.6 Hz, 1H), 4.08 – 4.01 (m, 2H), 3.81 (s, 3H), 2.19 (s, 3H), 1.79 (s, 3H);

^{13}C NMR (100 MHz, MeOD) δ 199.8, 166.9, 160.4, 159.0, 141.9, 141.2, 136.5, 133.9, 131.2, 130.9, 129.4, 128.8, 128.7, 128.6, 128.6, 127.5, 120.1, 114.1, 76.5, 62.9, 55.7, 48.9, 47.7, 43.1, 40.6, 29.7, 19.3;

$[\alpha]^{25}_{\text{D}} = +51.2$ (c = 0.5 in acetone, λ = 589 nm);

the enantioselectivity was determined by SFC (Trefoil CEL-1, gradient 100% CO_2 to CO_2/MeOH = 60:40 in the first 5 mins and maintaining in 60:40 CO_2/MeOH in the rest of time, flow rate: 0.5 mL/min, λ = 254 nm), t_{R} (major) = 10.7 min, t_{R} (minor) = 8.5 min.



X. Absolute configuration determination

X-Ray Crystallographic Data

Absolute configurations of products **3** were assigned based on the crystal X-ray structures of **3i**, **3j**. Absolute configurations of products **4**, **5** were assigned based on the crystal X-ray structures of **4a**, **5a**. Absolute configurations of products **8** were assigned based on the crystal X-ray structures of *rac*-**9**, *rac*-**10**, and *rac*-**14**. A light yellow rodlike crystal of **3i**, **3j** were obtained by vaporization of *n*-hexane/DCM (3:1) solution of corresponding compounds. A light yellow rodlike crystal of **8** was obtained by vaporization of *n*-hexane/DCM/CH₃CN (3:2:1) solution of compound **8**. A light yellow needle-like crystal of **4a**, **5a** were obtained by vaporization of *n*-hexane/Acetone (3:1) solution of corresponding compounds. A light yellow block-shaped crystal of *rac*-**9**, *rac*-**10**, *rac*-**14** were obtained by vaporization of *n*-hexane/DCM (3:1) solution of corresponding compounds.

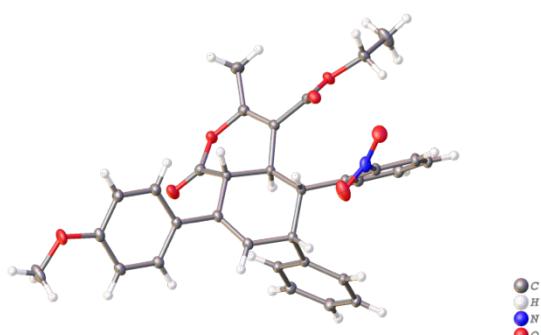


Figure S2. The crystal structure of **4a**.

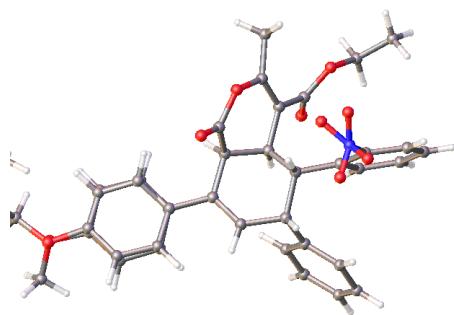


Figure S3. The crystal structure of **5a**

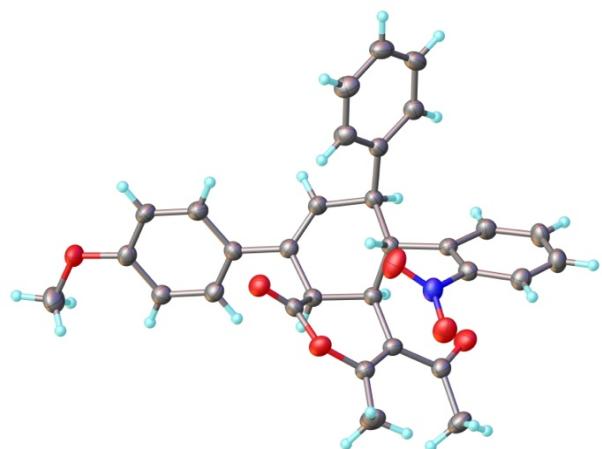


Figure S4. The crystal structure of **3i**.

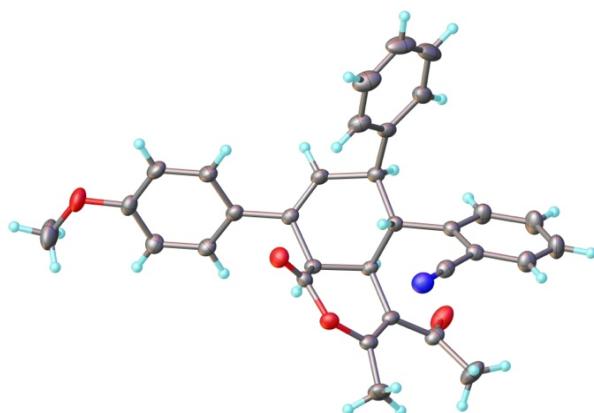


Figure S5. The crystal structure of **3j**.

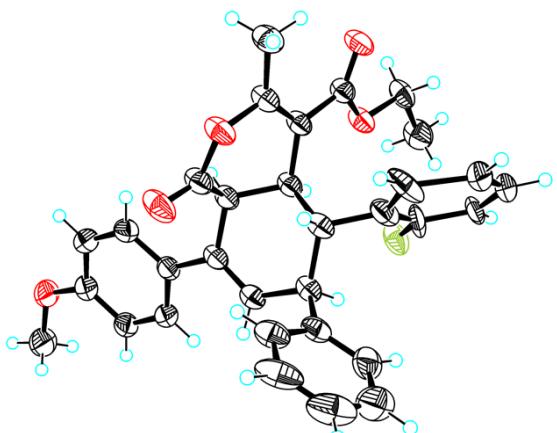


Figure S6. The crystal structure of *rac*-3c.

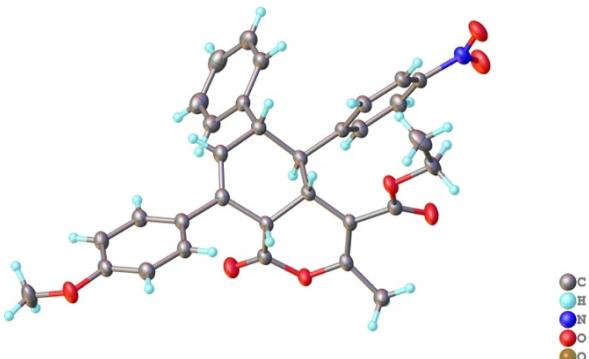


Figure S7. The crystal structure of *rac*-3f

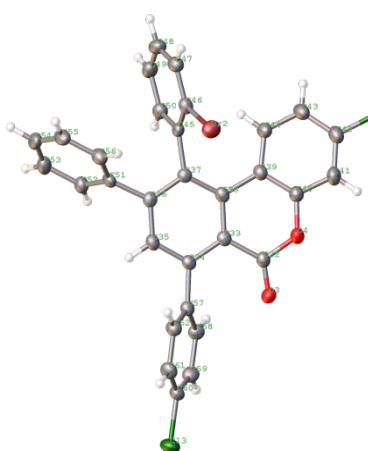


Figure S8. The crystal structure of (*R*)-8.

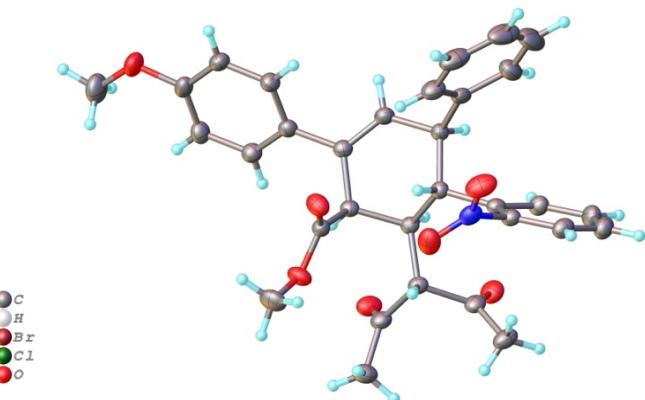


Figure S9. The crystal structure of *rac*-9

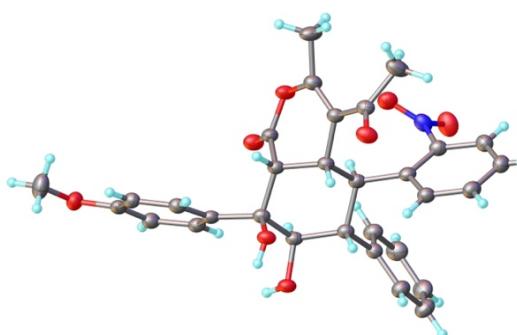


Figure S10. The crystal structure of *rac*-10.

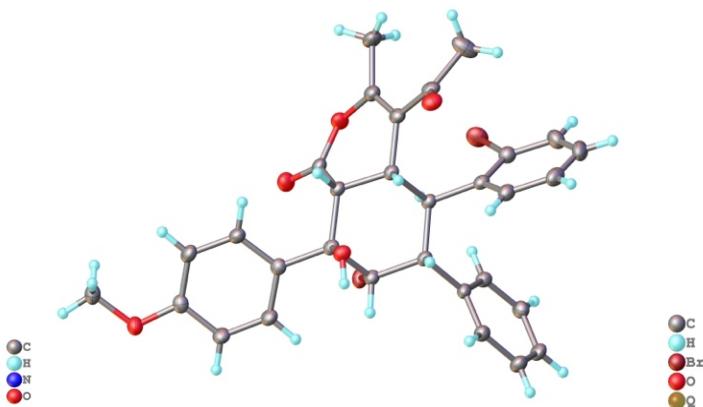


Figure S11. The crystal structure of *rac*-14.

Table S8. Crystal data and structure refinement of optical compounds.

Compound number	4a	5a	3i	3j	(R)-8
Deposition number	2246562	2377578	2244572	2244570	2235169
Empirical formula	C ₃₂ H ₂₉ NO ₇	C ₃₂ H ₂₉ NO ₇	C ₃₁ H ₂₇ NO ₆	C ₃₂ H ₂₇ NO ₄	C ₃₁ H ₁₇ BrCl ₂ O ₂
Formula weight	539.56	539.56	509.56	489.57	572.26
Temperature/K	100(2)	149.99(10)	150.00(10)	150.00(10)	149.99(10)
Crystal system	orthorhombic	tetragonal	monoclinic	triclinic	monoclinic
Space group	P2 ₁ 2 ₁ 2 ₁	P4 ₃	P2 ₁	P1	C2
a/Å	10.51673(10)	16.7397(3)	7.02860(10)	7.28960(10)	43.9803(4)
b/Å	11.34590(10)	16.7397(3)	19.9167(2)	9.99900(10)	8.17570(10)
c/Å	22.2044(2)	10.5216(3)	10.57480(10)	10.3560(2)	14.36051(10)
$\alpha/^\circ$	90	90	90	90.9030(10)	90
$\beta/^\circ$	90	90	103.5740(10)	104.2710(10)	105.4170(10)
$\gamma/^\circ$	90	90	90	97.5320(10)	90
Volume/Å ³	2649.47(4)	2948.33(12)	1438.98(3)	724.321(19)	4977.80(9)
Z	4	4	2	1	8
$\rho_{\text{calc}}/\text{g/cm}^3$	1.353	1.313	1.372	1.317	1.527
μ/mm^{-1}	0.096	0.742	2.415	2.327	4.450
F(000)	1136	1236	620	300	2304
Crystal size/mm ³	0.260 × 0.180 × 0.120	0.22 × 0.15 × 0.08	0.1 × 0.09 × 0.07	0.13 × 0.12 × 0.1	0.31 × 0.06 × 0.04
Radiation	Mo K α ($\lambda = 0.71073$)	Cu K α ($\lambda = 1.54184$)	Cu K α ($\lambda = 1.54184$)	Cu K α ($\lambda = 1.54184$)	CuK α ($\lambda = 1.54184$)
2 Θ range for data collection/ $^\circ$	5.592 to 59.992	7.468 to 152.312	8.602 to 148.556	8.822 to 148.958	6.384 to 148.378
Index ranges	-14 ≤ h ≤ 13, -15 ≤ k ≤ 15, -31 ≤ l ≤ 31	-19 ≤ h ≤ 19, -20 ≤ k ≤ 13, -12 ≤ l ≤ 12	-8 ≤ h ≤ 8, -24 ≤ k ≤ 24, -12 ≤ l ≤ 11	-9 ≤ h ≤ 9, -12 ≤ k ≤ 12, -12 ≤ l ≤ 12	-54 ≤ h ≤ 54, -10 ≤ k ≤ 9, -17 ≤ l ≤ 17
Reflections collected	48521	20352	15310	21601	37893
Independent reflections	7623 [$R_{\text{int}} = 0.0247$, $R_{\text{sigma}} = 0.0139$]	5846 [$R_{\text{int}} = 0.0265$, $R_{\text{sigma}} = 0.0251$]	5652 [$R_{\text{int}} = 0.0367$, $R_{\text{sigma}} = 0.0419$]	5516 [$R_{\text{int}} = 0.0259$, $R_{\text{sigma}} = 0.0210$]	9656 [$R_{\text{int}} = 0.0611$, $R_{\text{sigma}} = 0.0438$]
Data/restraints/parameters	7623/0/364	5846/60/539	5652/1/373	5516/3/364	9656/2/649
Goodness-of-fit on F^2	1.045	1.055	1.056	1.022	1.065
Final R indexes [$ I \geq 2\sigma(I)$]	$R_1 = 0.0297$, $wR_2 = 0.0791$	$R_1 = 0.0710$, $wR_2 = 0.2088$	$R_1 = 0.0364$, $wR_2 = 0.0926$	$R_1 = 0.0295$, $wR_2 = 0.0766$	$R_1 = 0.0386$, $wR_2 = 0.1032$
Final R indexes [all data]	$R_1 = 0.0303$, $wR_2 = 0.0798$	$R_1 = 0.0749$, $wR_2 = 0.2159$	$R_1 = 0.0395$, $wR_2 = 0.0945$	$R_1 = 0.0302$, $wR_2 = 0.0774$	$R_1 = 0.0399$, $wR_2 = 0.1051$
Largest diff.peak/hole/eÅ ⁻³	0.30/-0.17	0.80/-0.36	0.39/-0.37	0.20/-0.33	0.64/-0.93
Flack parameter	-0.06(12)	0.01(2)	-0.023(9)	0.005(4)	-0.016(9)

Table S9. Crystal data and structure refinement of racemic compounds.

Compound number	<i>rac-3c</i>	<i>rac-3f</i>	<i>rac-9</i>	<i>rac-10</i>	<i>rac-14</i>
Deposition number	2244575	2244573	2244569	2244568	2243102
Empirical formula	C ₃₂ H ₂₉ FO ₅	C ₃₂ H ₂₉ NO ₇	C ₃₂ H ₃₁ NO ₇	C ₃₁ H ₂₉ NO ₈	C ₃₁ H ₂₈ Br ₂ O ₅
Formula weight	512.55	539.56	541.58	543.6	640.35
Temperature/K	100.00(10)	149.99(10)	149.99(10)	100.00(10)	99.98(10)
Crystal system	monoclinic	triclinic	monoclinic	monoclinic	monoclinic
Space group	<i>P</i> 2 ₁ /c	<i>P</i> -1	<i>P</i> 2 ₁ /c	<i>P</i> 2 ₁ /n	<i>P</i> 2 ₁ /c
<i>a</i> /Å	10.7735(2)	10.3074(3)	8.2745(2)	22.0372(3)	14.45320(10)
<i>b</i> /Å	28.0445(5)	10.5620(4)	33.3961(7)	9.66930(10)	14.99130(10)
<i>c</i> /Å	8.8667(2)	12.7619(4)	10.3175(2)	26.7286(4)	12.68540(10)
<i>α</i> /°	90	102.425(3)	90	90	90
<i>β</i> /°	104.427(2)	99.324(3)	104.258(2)	110.983(2)	101.1850(10)
<i>γ</i> /°	90	101.043(3)	90	90	90
Volume/Å ³	2594.48(9)	1301.40(8)	2763.27(11)	5317.76(14)	2696.37(3)
<i>Z</i>	4	2	4	4	4
ρ _{calc} g/cm ³	1.312	1.377	1.302	1.358	1.577
μ/mm ⁻¹	0.758	0.799	0.752	0.814	4.143
<i>F</i> (000)	1080	568	1144	2288	1296.0
Crystal size/mm ³	0.11 × 0.1 × 0.07	0.25 × 0.1 × 0.1	0.1 × 0.1 × 0.1	0.23 × 0.12 × 0.04	0.18×0.15×0.13
Radiation	Cu <i>K</i> α (<i>λ</i> = 1.54184)	Cu <i>K</i> α (<i>λ</i> = 1.54184)			
2θ range for data collection/°	6.304 to 148.372	7.258 to 149.078	9.232 to 149.438	6.472 to 148.89	8.574 to 149.004
Index ranges	-13 ≤ <i>h</i> ≤ 12, -28 ≤ <i>k</i> ≤ 34, -9 ≤ <i>l</i> ≤ 11	-12 ≤ <i>h</i> ≤ 12, -13 ≤ <i>k</i> ≤ 13, -15 ≤ <i>l</i> ≤ 8	-9 ≤ <i>h</i> ≤ 10, -35 ≤ <i>k</i> ≤ 41, -12 ≤ <i>l</i> ≤ 11	-25 ≤ <i>h</i> ≤ 27, -12 ≤ <i>k</i> ≤ 11, -33 ≤ <i>l</i> ≤ 31	-9 ≤ <i>h</i> ≤ 17, -17 ≤ <i>k</i> ≤ 12, -15 ≤ <i>l</i> ≤ 15
Reflections collected	13083	12729	15386	30744	15346
Independent reflections	5108 [<i>R</i> _{int} = 0.0564, <i>R</i> _{sigma} = 0.0521]	5112 [<i>R</i> _{int} = 0.0274, <i>R</i> _{sigma} = 0.0388]	5479 [<i>R</i> _{int} = 0.0340, <i>R</i> _{sigma} = 0.0406]	10519 [<i>R</i> _{int} = 0.0591, <i>R</i> _{sigma} = 0.0484]	5330 [<i>R</i> _{int} = 0.0276, <i>R</i> _{sigma} = 0.0252]
Data/restraints/parameters	5108/62/401	5112/0/364	5479/0/365	10519/0/731	5330/0/347
Goodness-of-fit on <i>F</i> ²	1.12	1.068	1.072	1.06	1.068
Final <i>R</i> indexes [<i>I</i> >=2σ (<i>I</i>)]	<i>R</i> ₁ = 0.0634, <i>wR</i> ₂ = 0.1862	<i>R</i> ₁ = 0.0436, <i>wR</i> ₂ = 0.1115	<i>R</i> ₁ = 0.0417, <i>wR</i> ₂ = 0.1043	<i>R</i> ₁ = 0.0534, <i>wR</i> ₂ = 0.1479	<i>R</i> ₁ = 0.0309, <i>wR</i> ₂ = 0.0852
Final <i>R</i> indexes [all data]	<i>R</i> ₁ = 0.0794, <i>wR</i> ₂ = 0.1954	<i>R</i> ₁ = 0.0522, <i>wR</i> ₂ = 0.1160	<i>R</i> ₁ = 0.0514, <i>wR</i> ₂ = 0.1090	<i>R</i> ₁ = 0.0607, <i>wR</i> ₂ = 0.1560	<i>R</i> ₁ = 0.0320, <i>wR</i> ₂ = 0.0860
Largest diff.peak/hole/eÅ ⁻³	0.23/-0.23	0.24/-0.21	0.19/-0.23	0.32/-0.34	0.82/-0.81

ECD determination Data

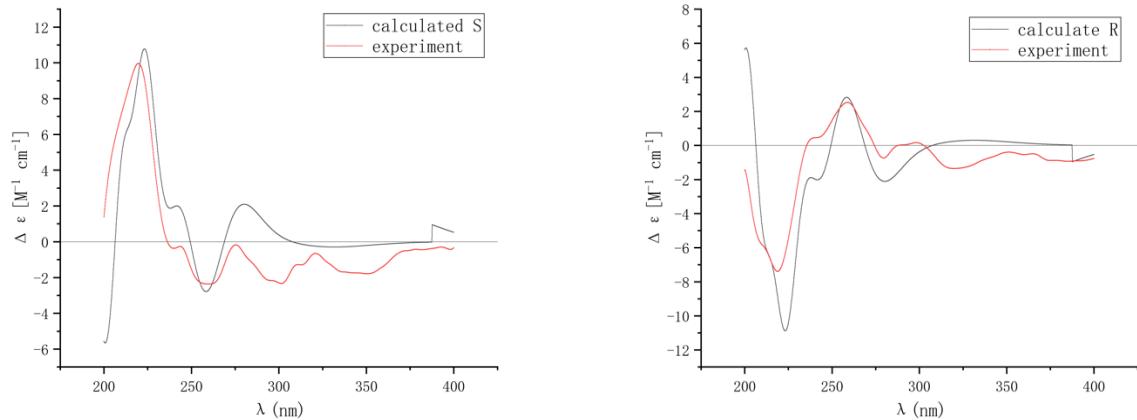
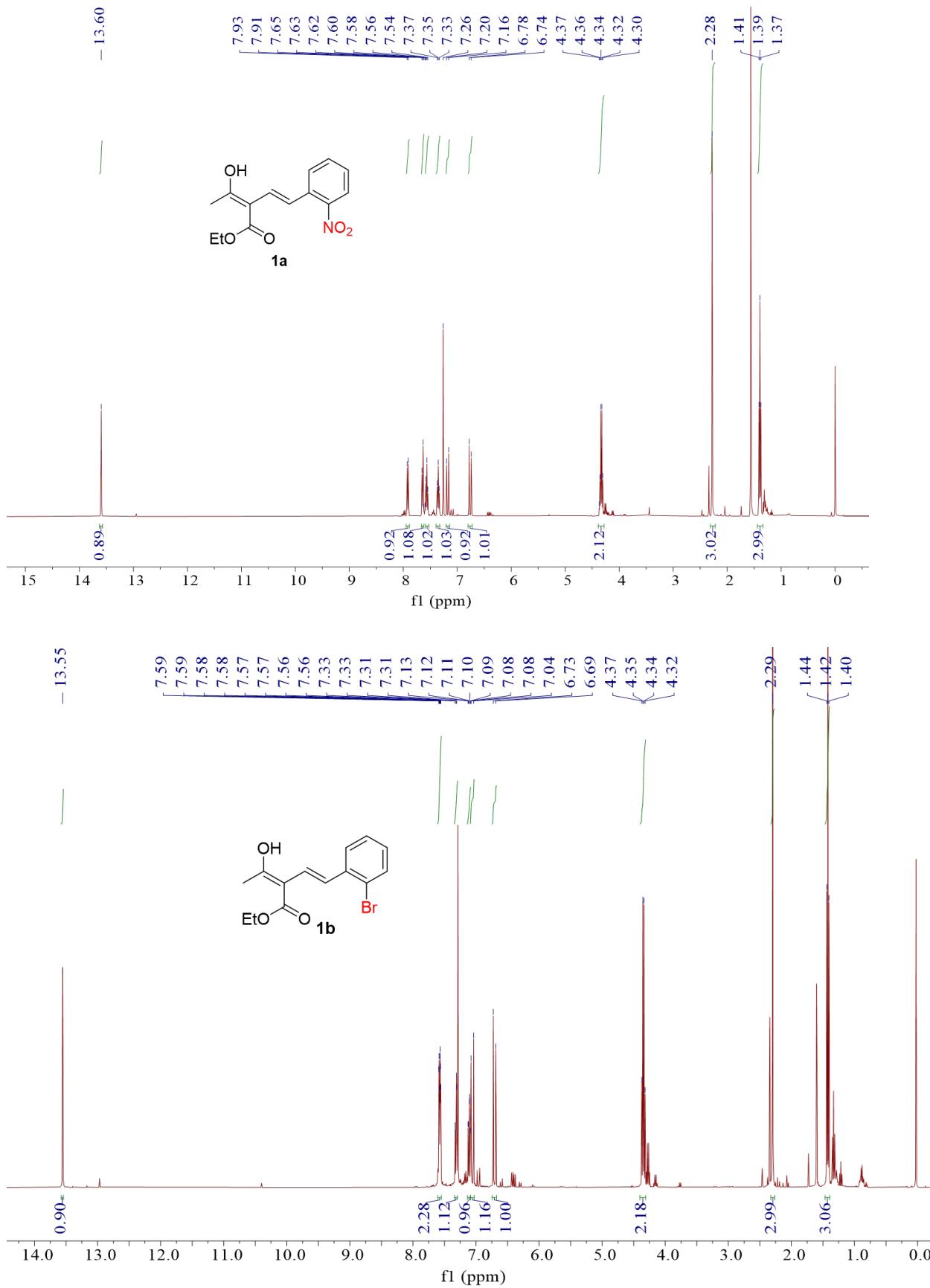


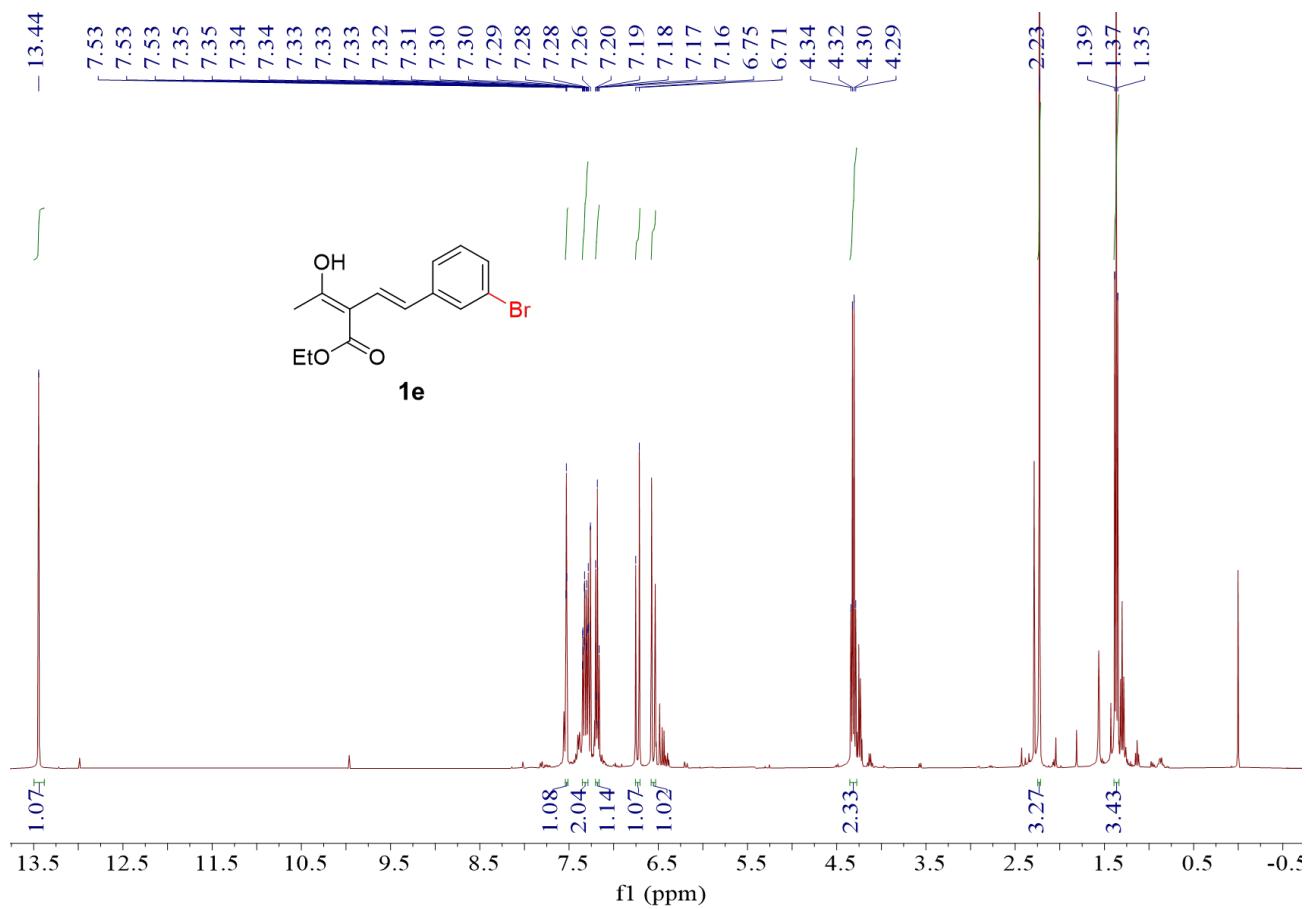
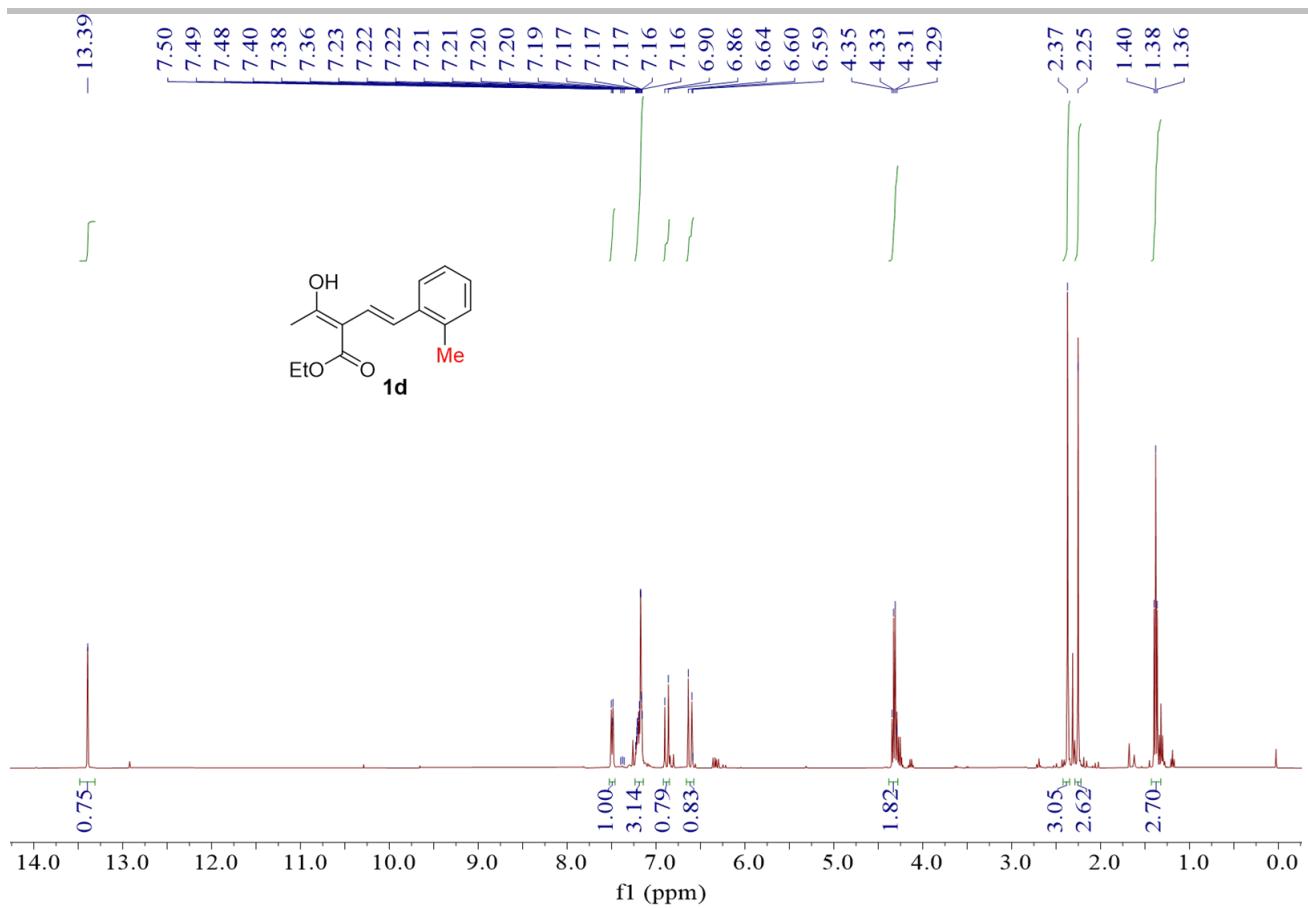
Figure S12. Comparsion of the calculated vs experimental ECD spectra in CH_3CN for compound **6** (left from **3i**, right from **4i**).

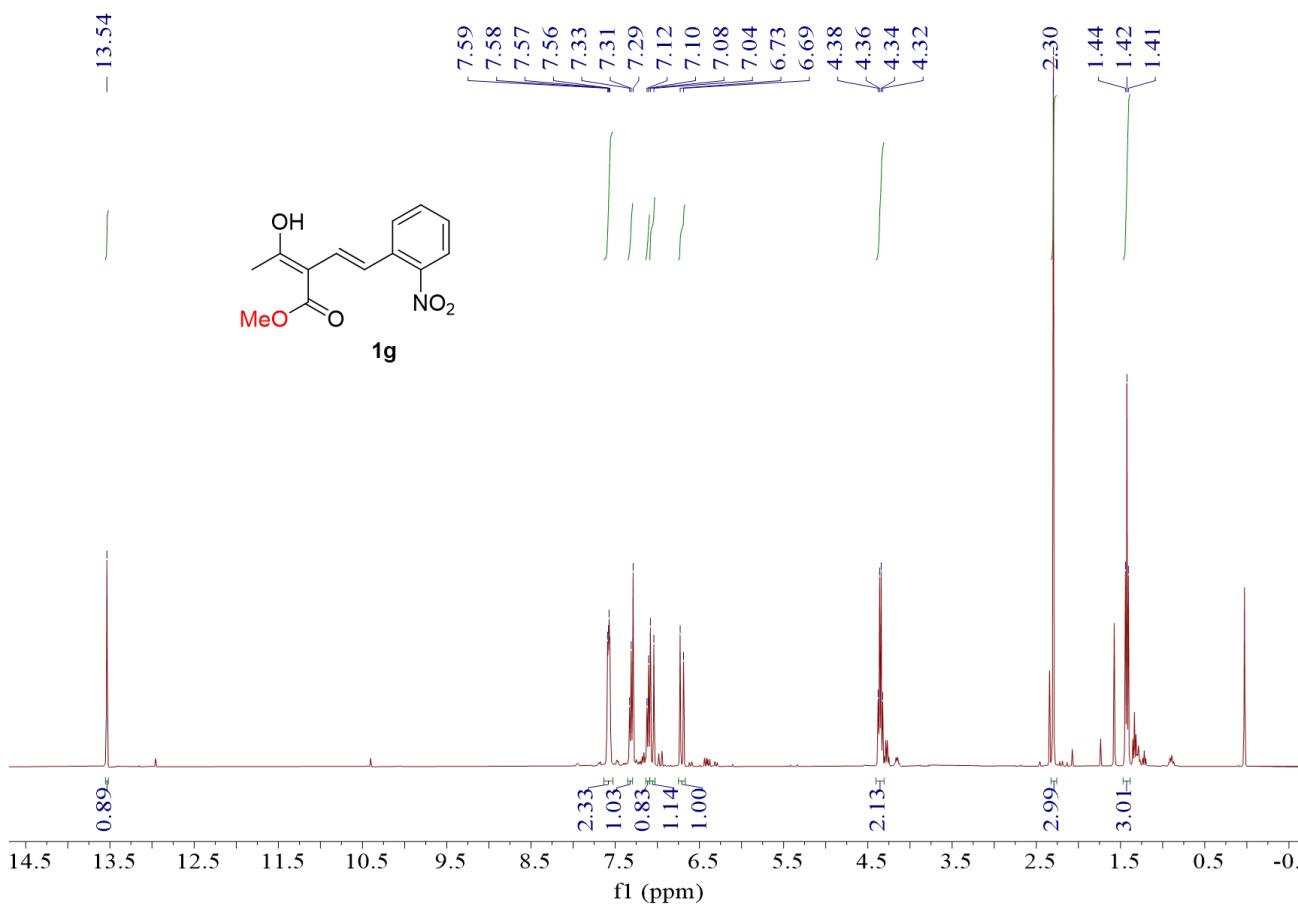
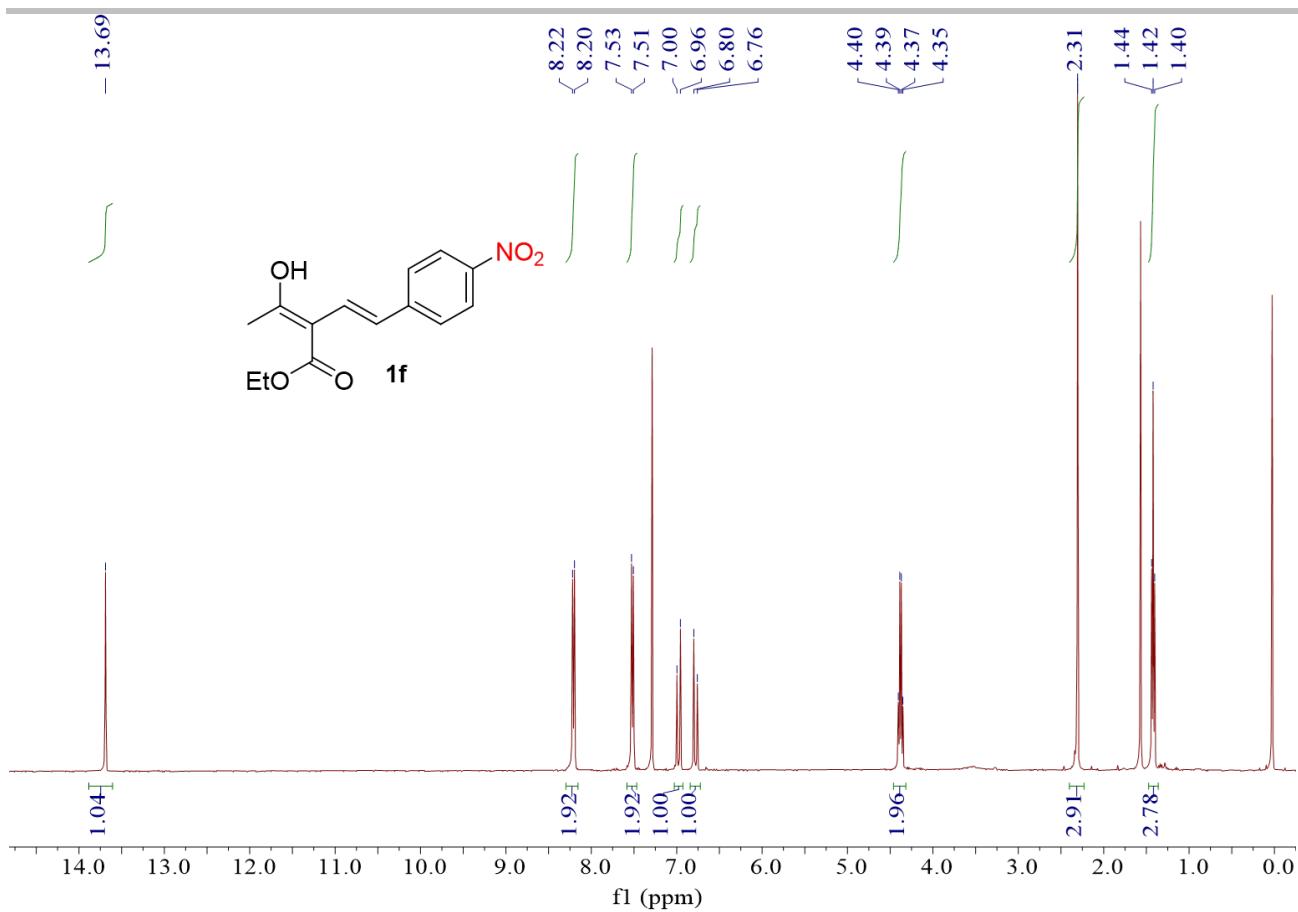
ECD calculation methods:

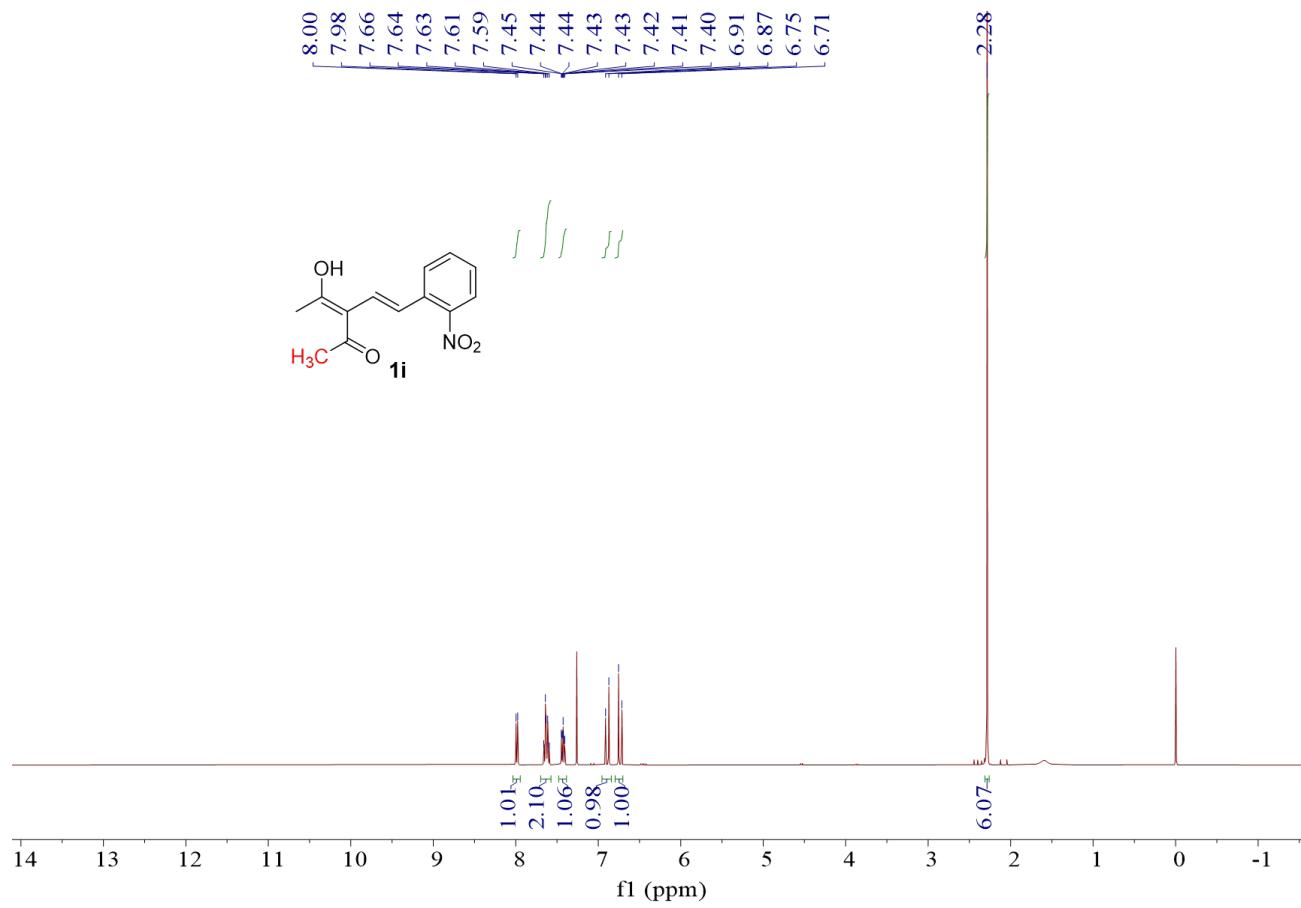
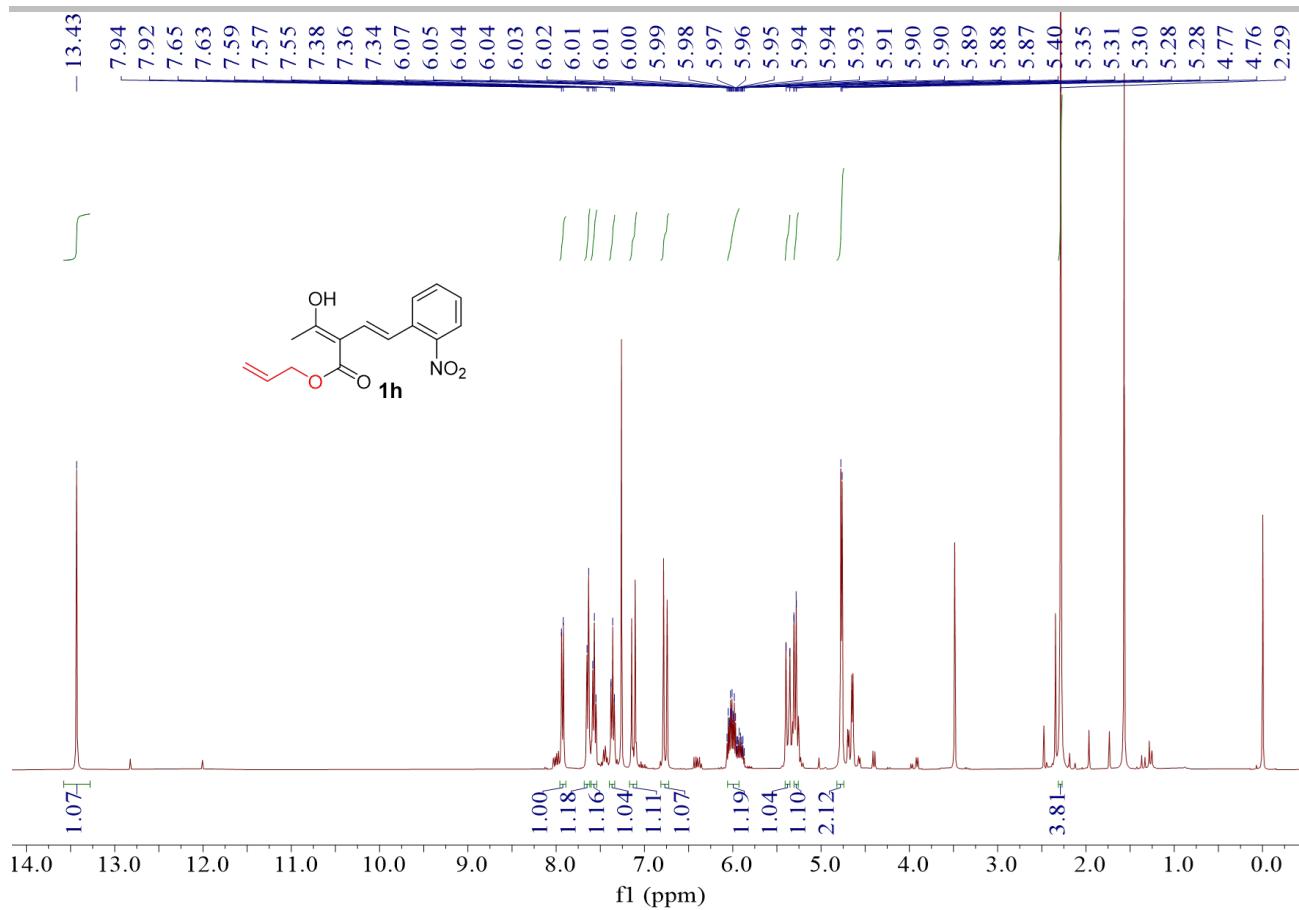
Monte Carlo conformational searches were carried out by means of the Spartan's 14 software using Merck Molecular Force Field (MMFF). The conformers with Boltzmann-population of over 5% were chosen for ECD calculations, and then the conformers were initially optimized at B3LYP/6-31g level in gas. The theoretical calculation of ECD was conducted in Acetonitrile using Time-dependent Density functional theory (TD-DFT) at the B3LYP/6-31+g (d, p) level for all conformers of compounds **6**. Rotatory strengths for a total of 30 excited states were calculated. ECD spectra were generated using the program SpecDis 1.6 (University of Würzburg, Würzburg, Germany) and GraphPad Prism 5 (University of California San Diego, USA) from dipole-length rotational strengths by applying Gaussian band shapes with sigma = 0.3 eV.

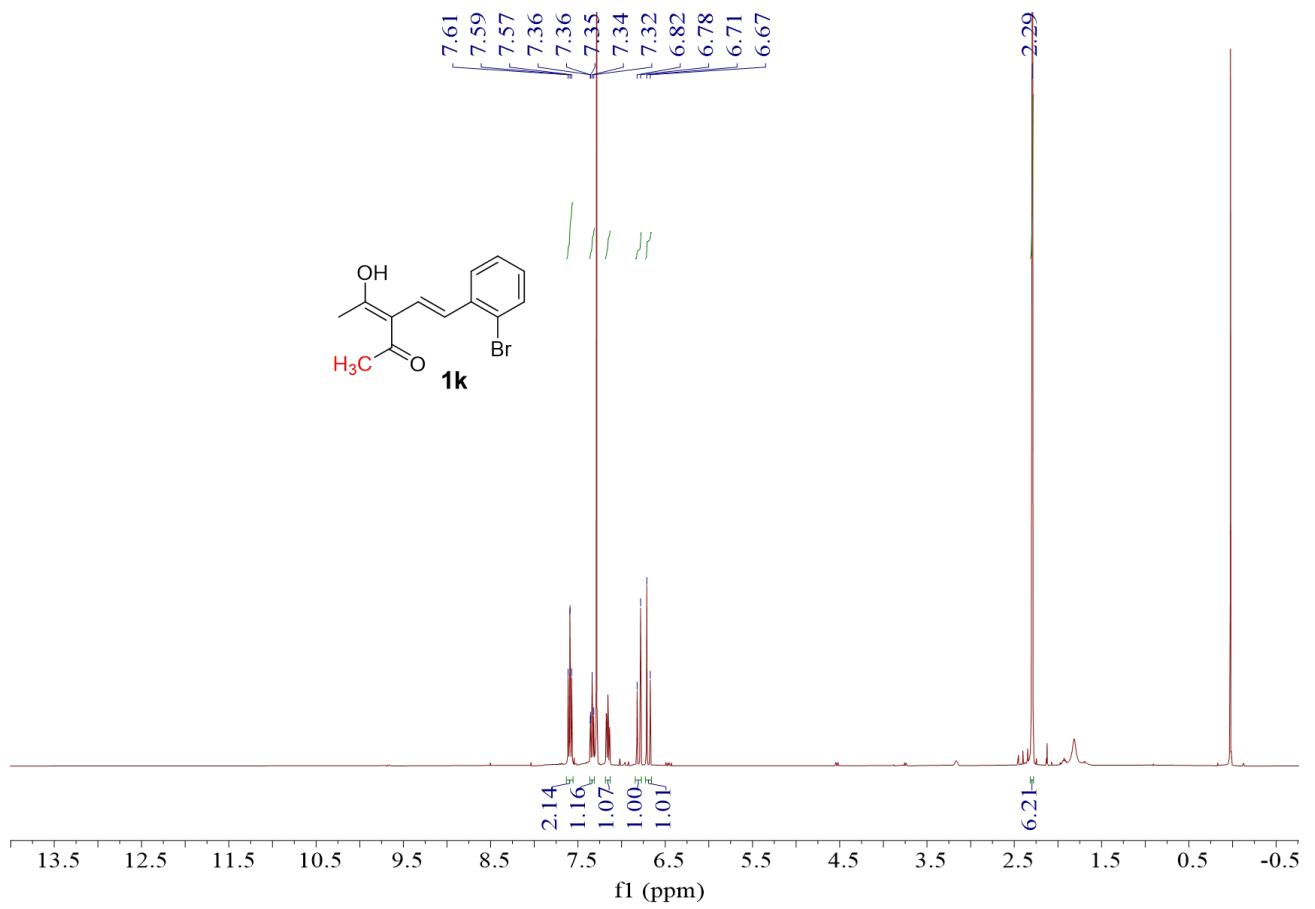
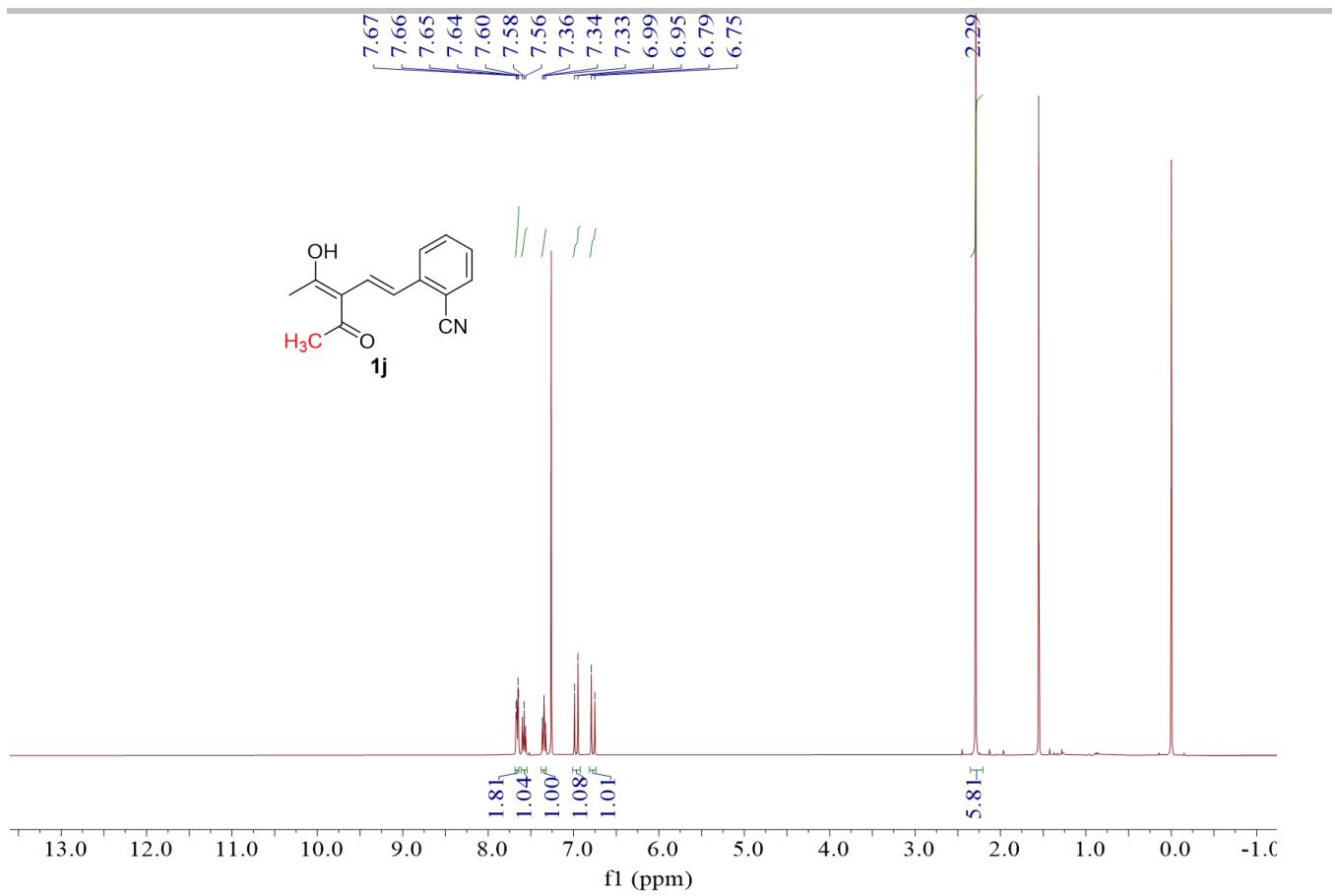
XI. Copies of ^1H NMR spectra of 1



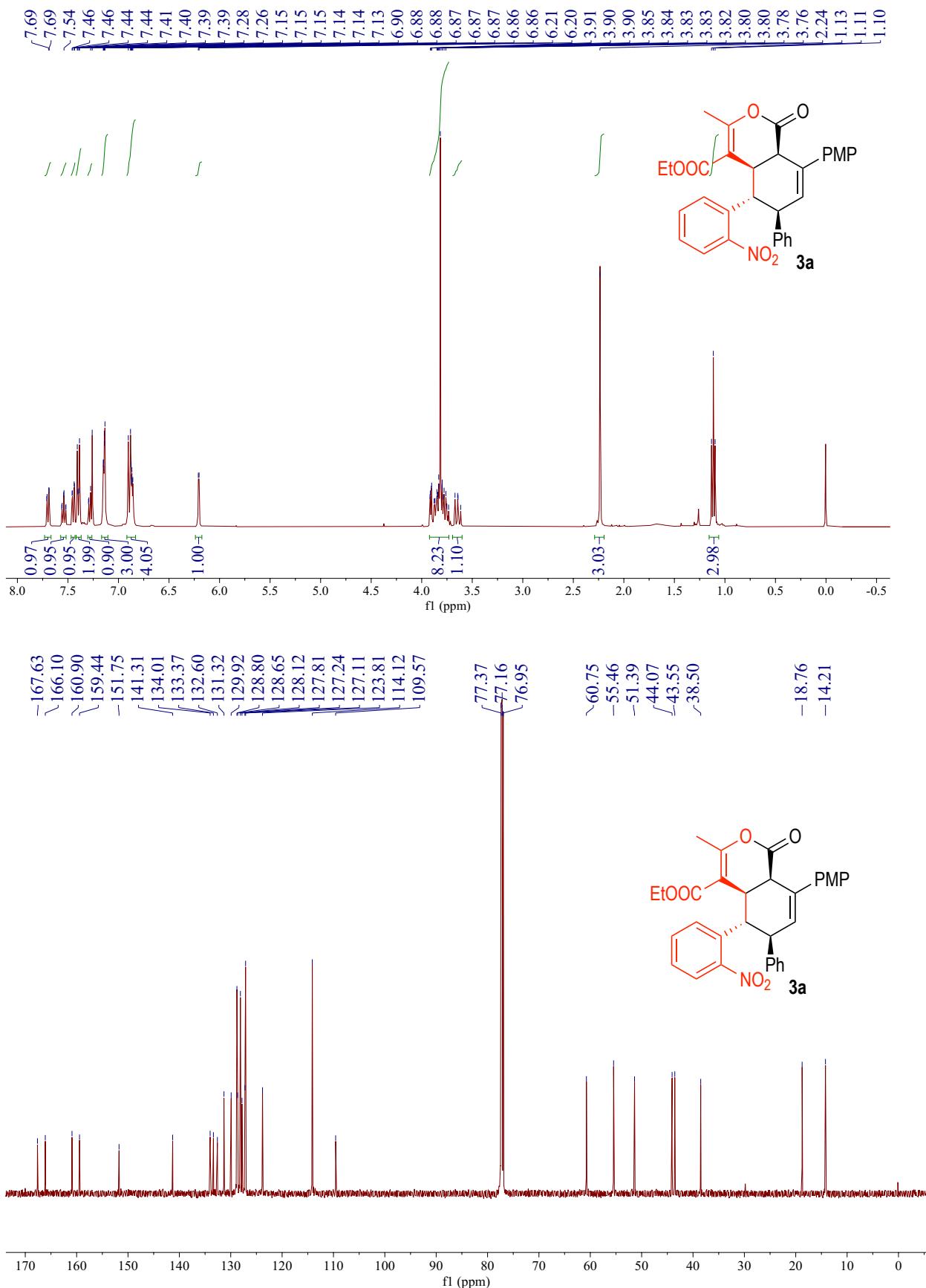


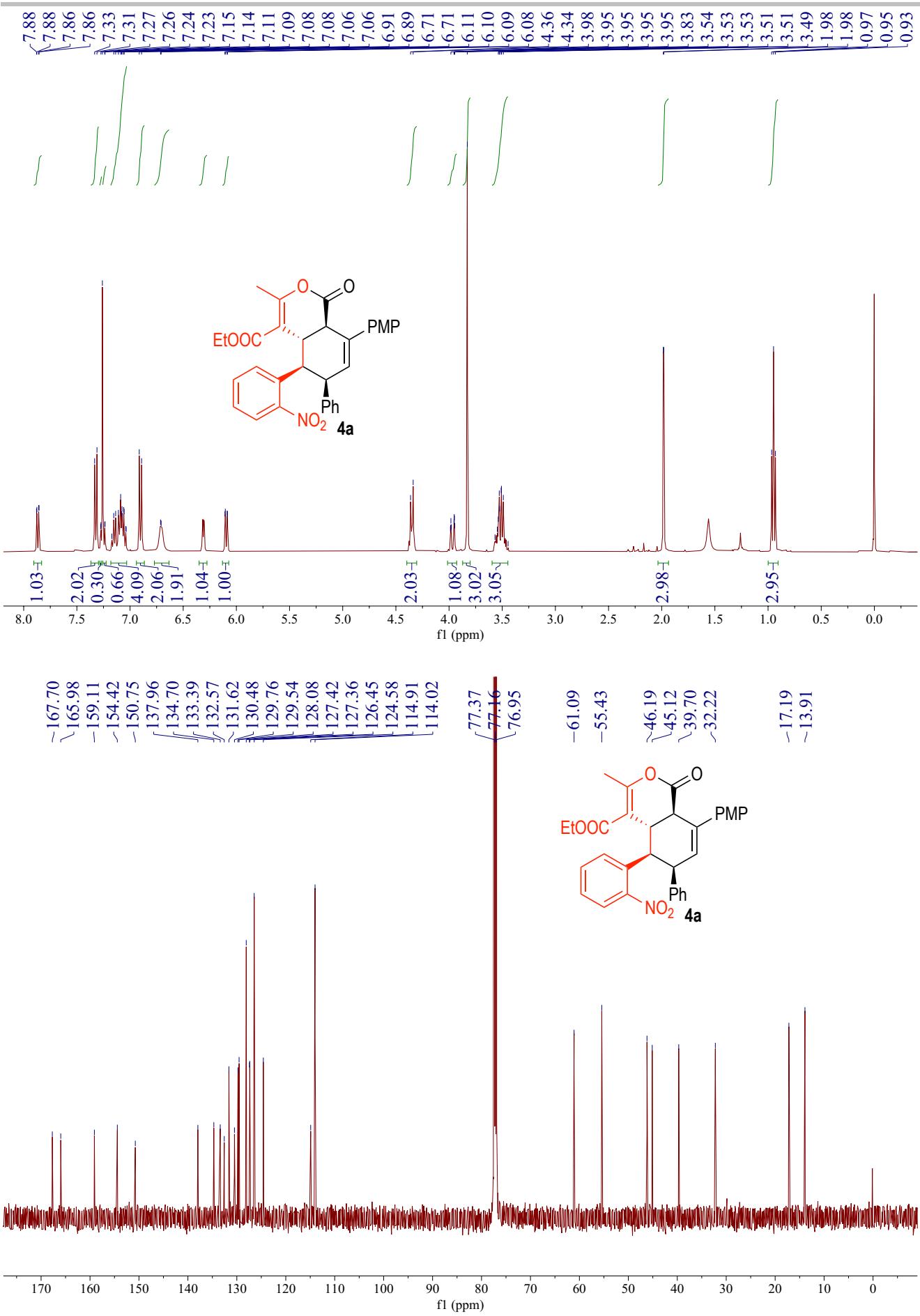


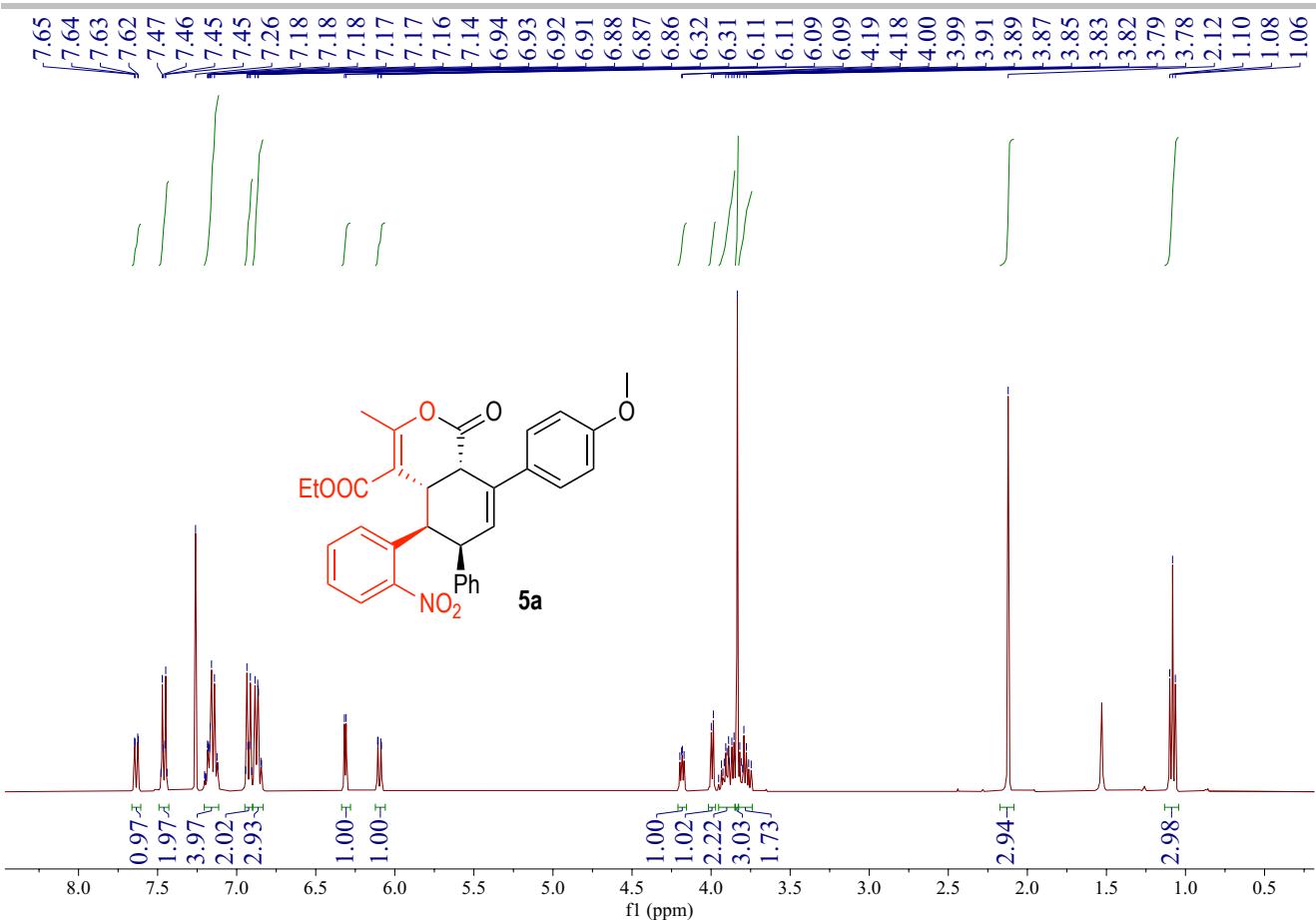


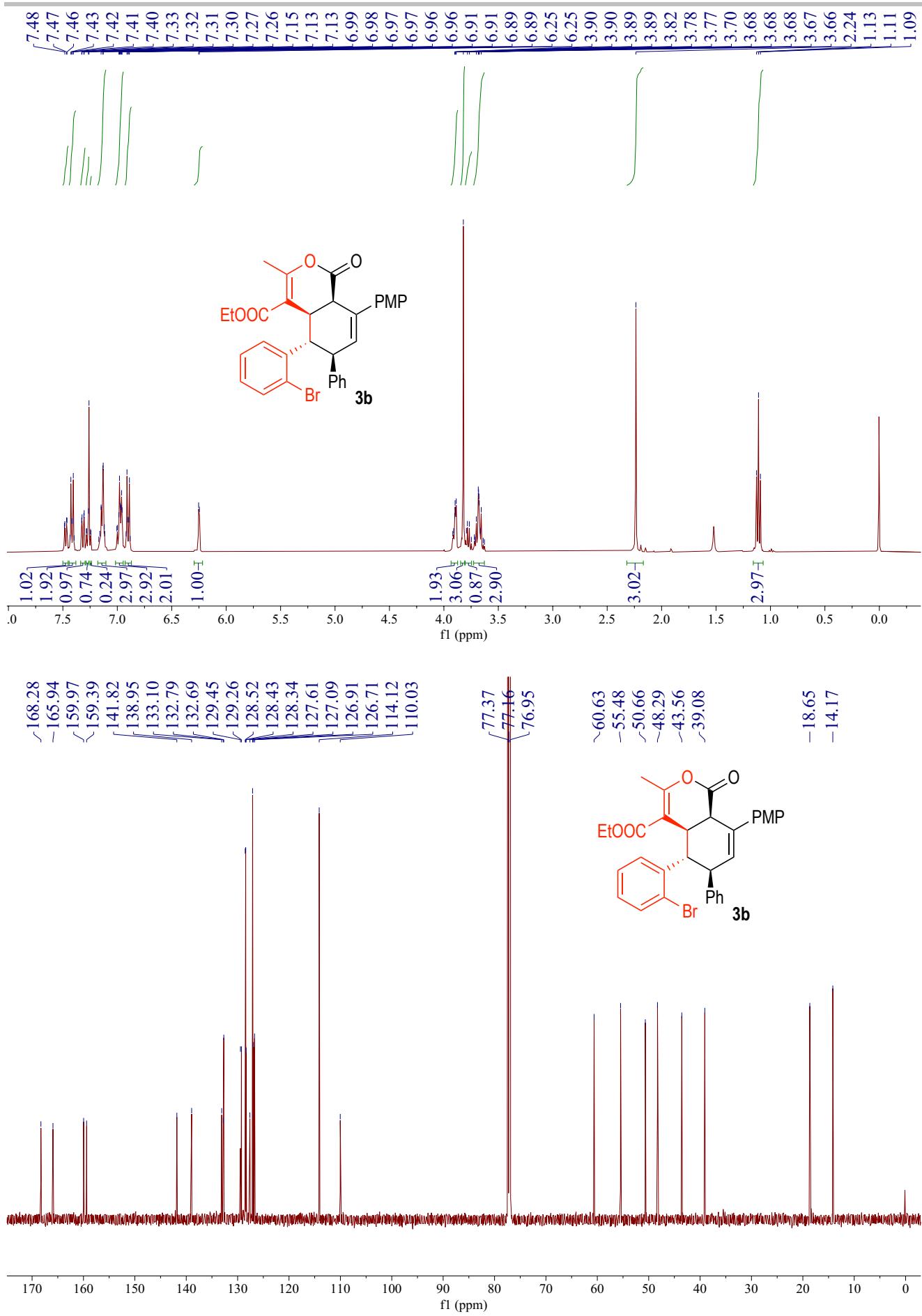


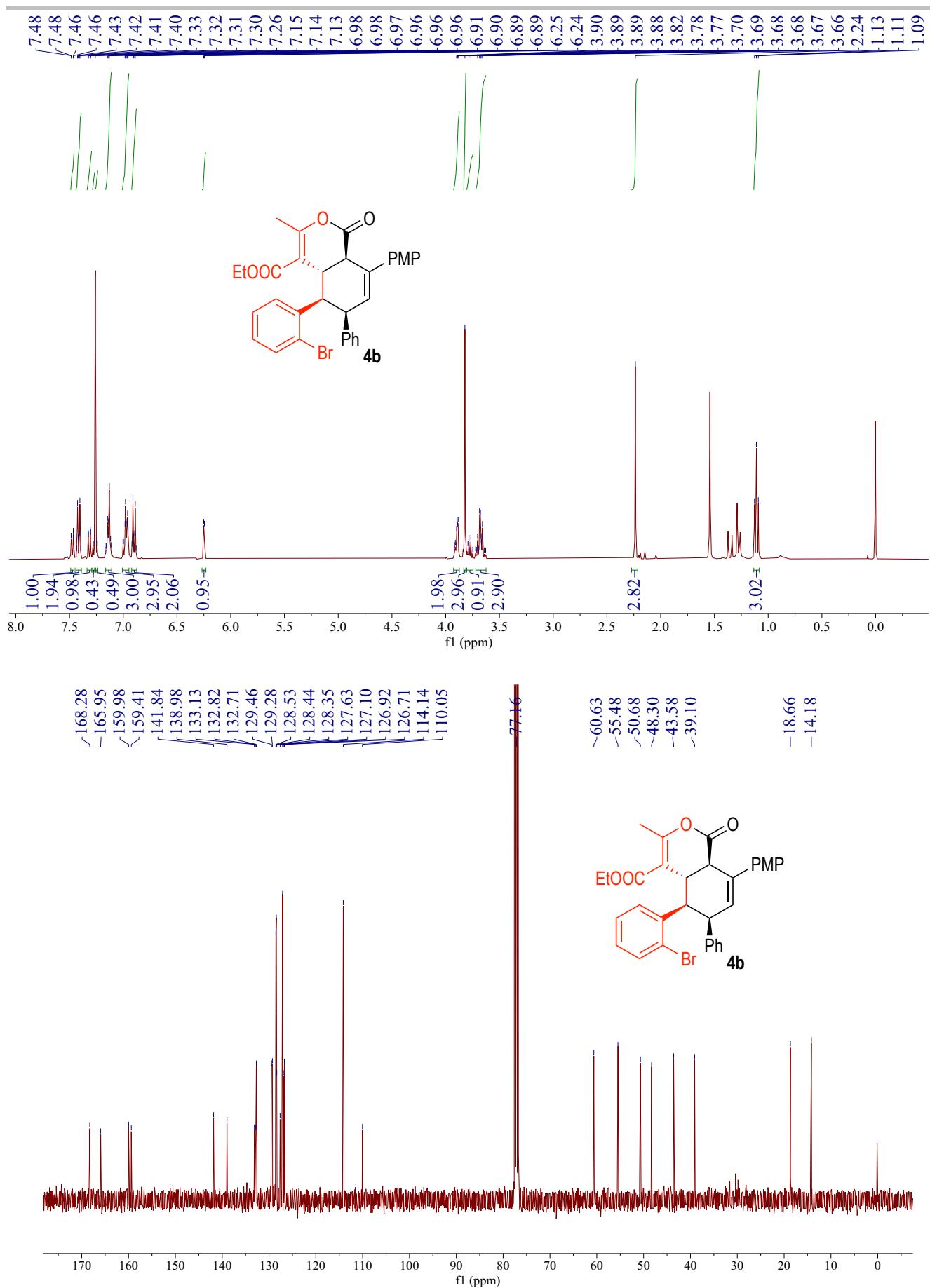
XII. Copies of ^1H NMR, ^{13}C NMR, ^{19}F NMR spectra of products

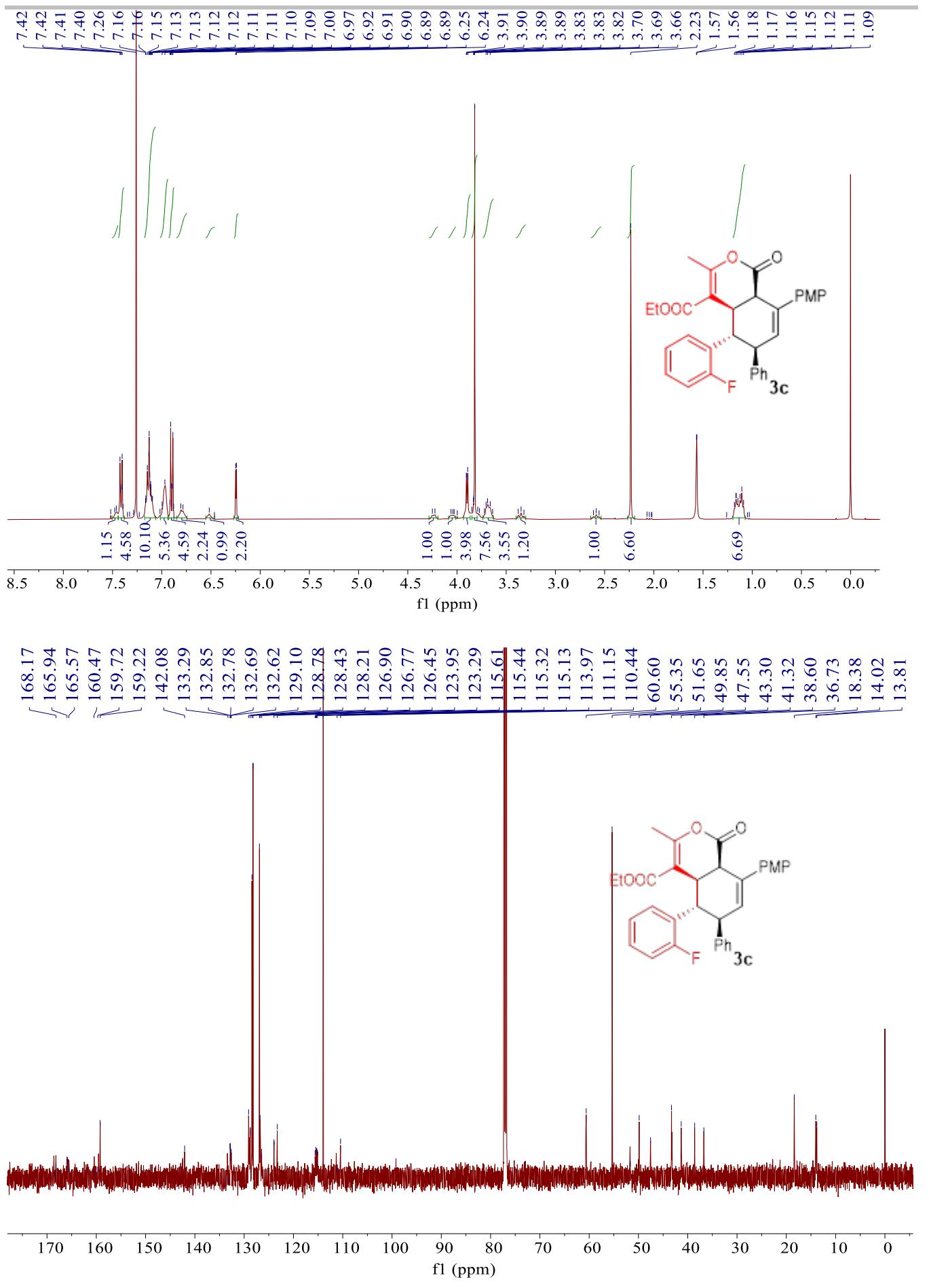


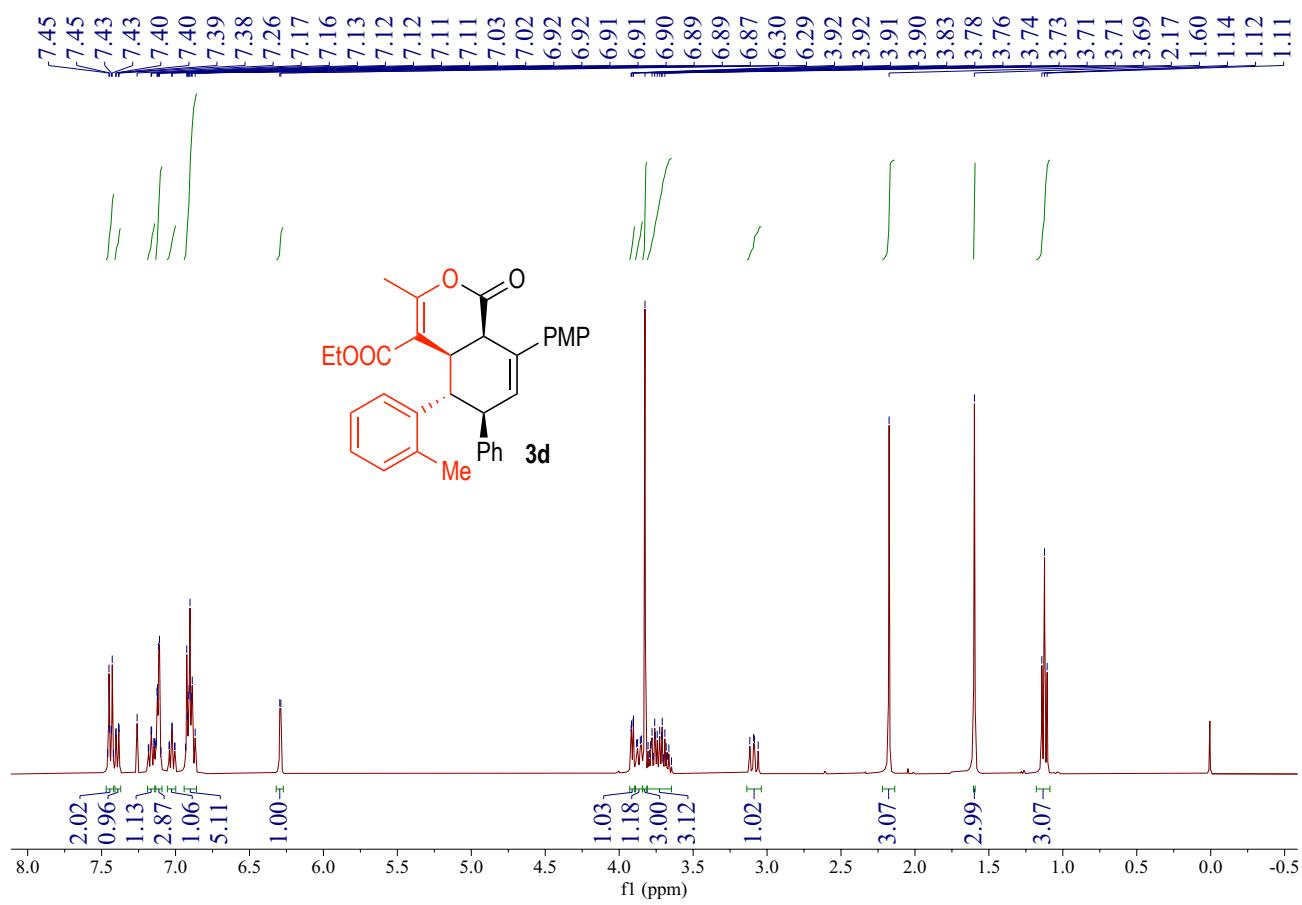
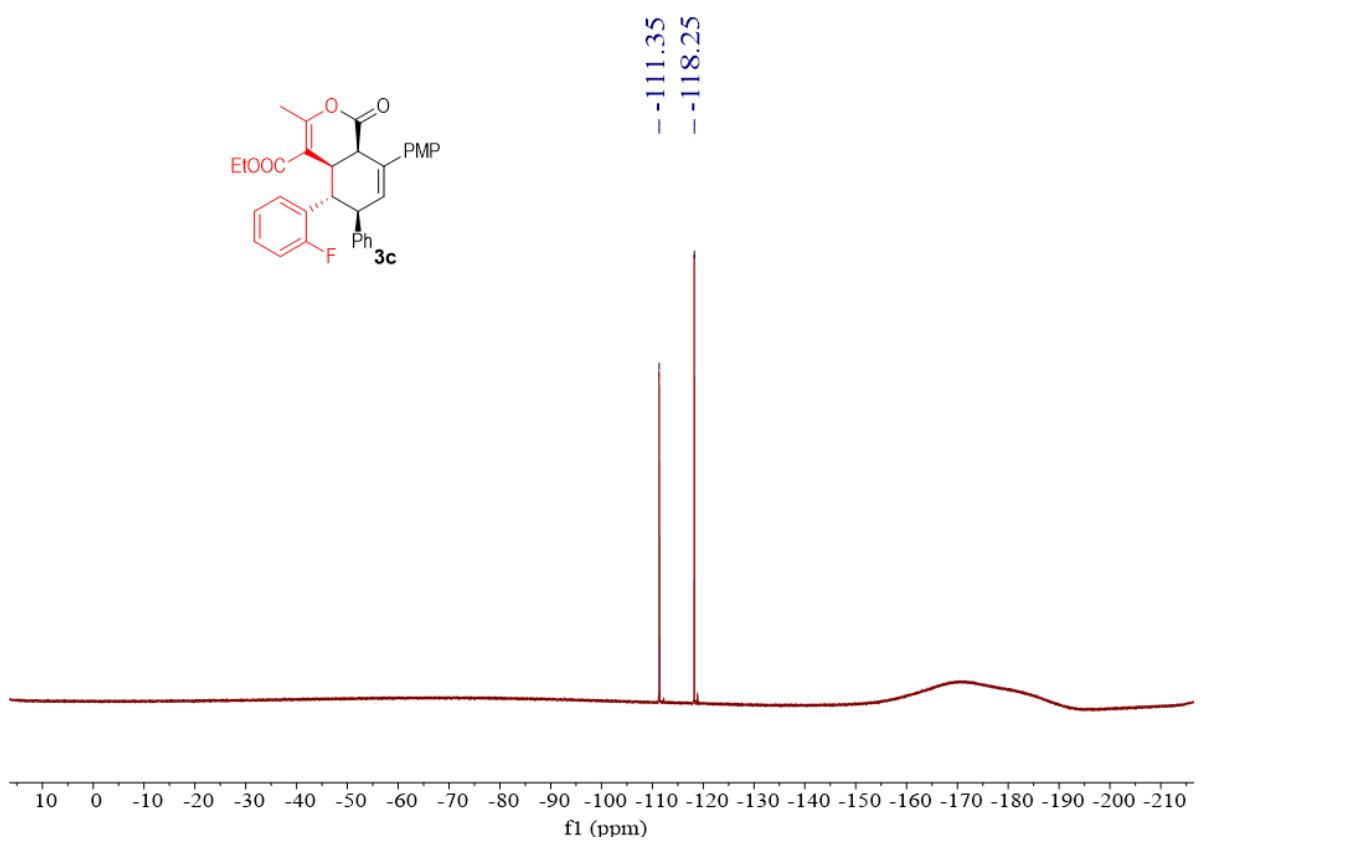


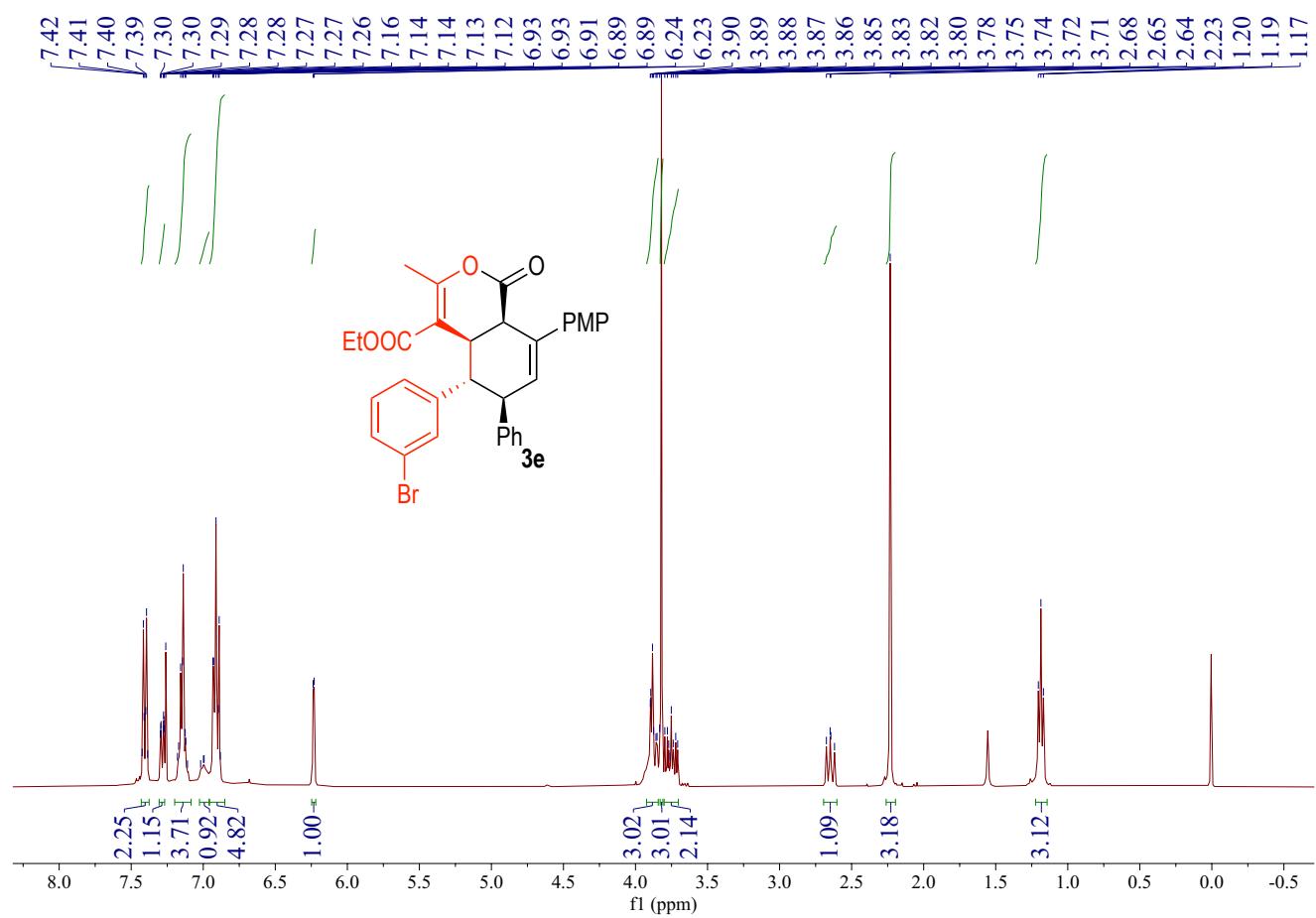
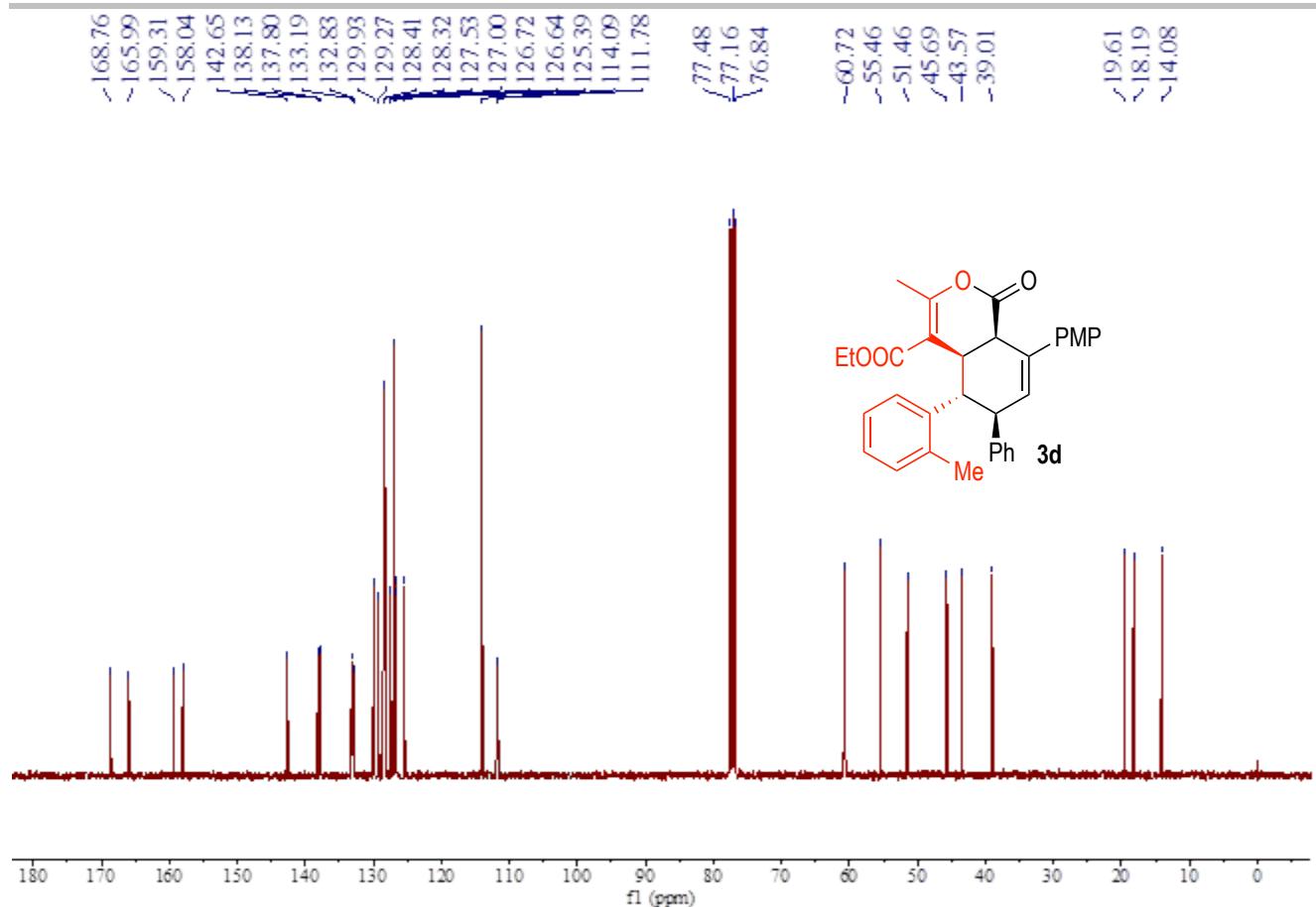


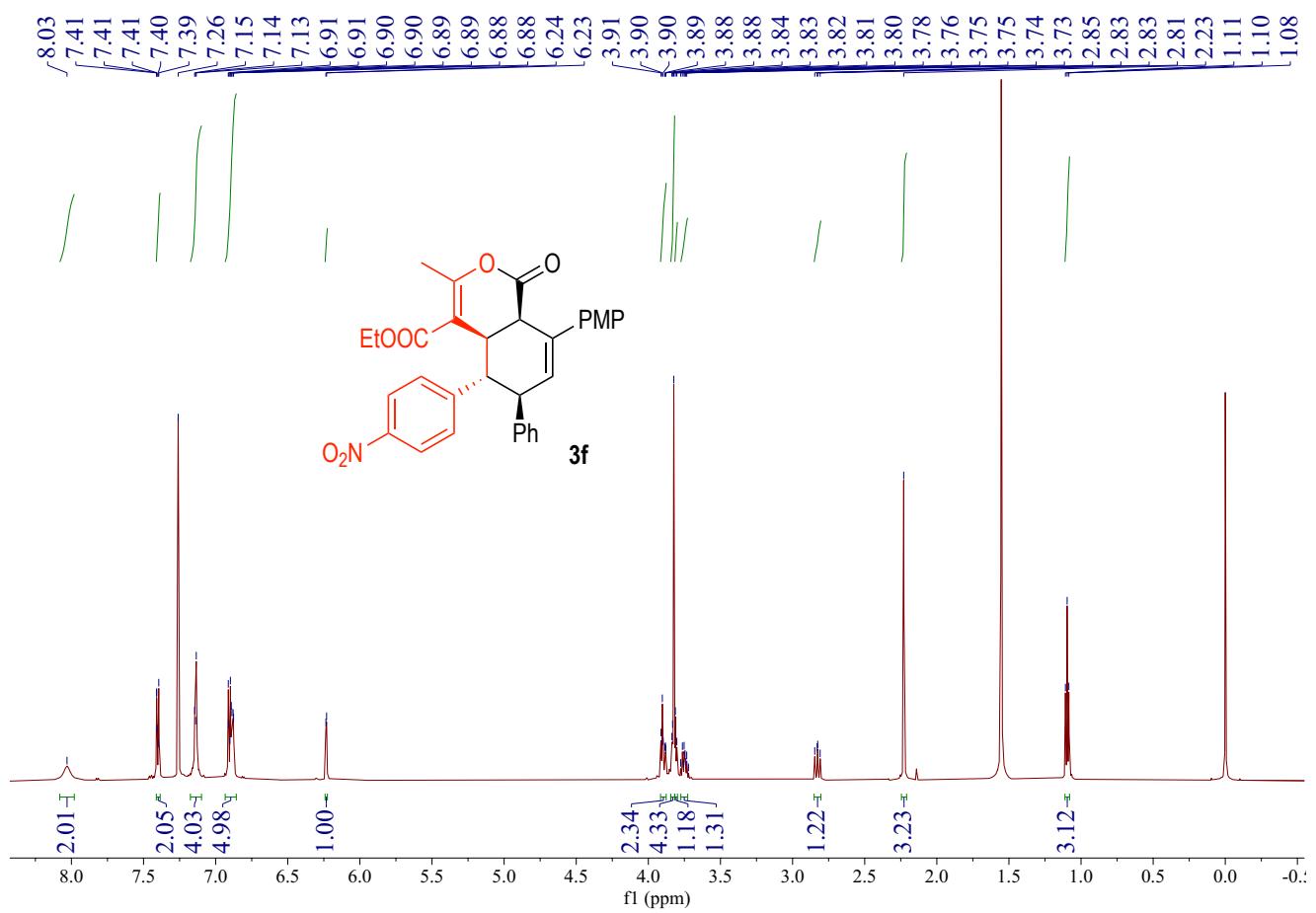
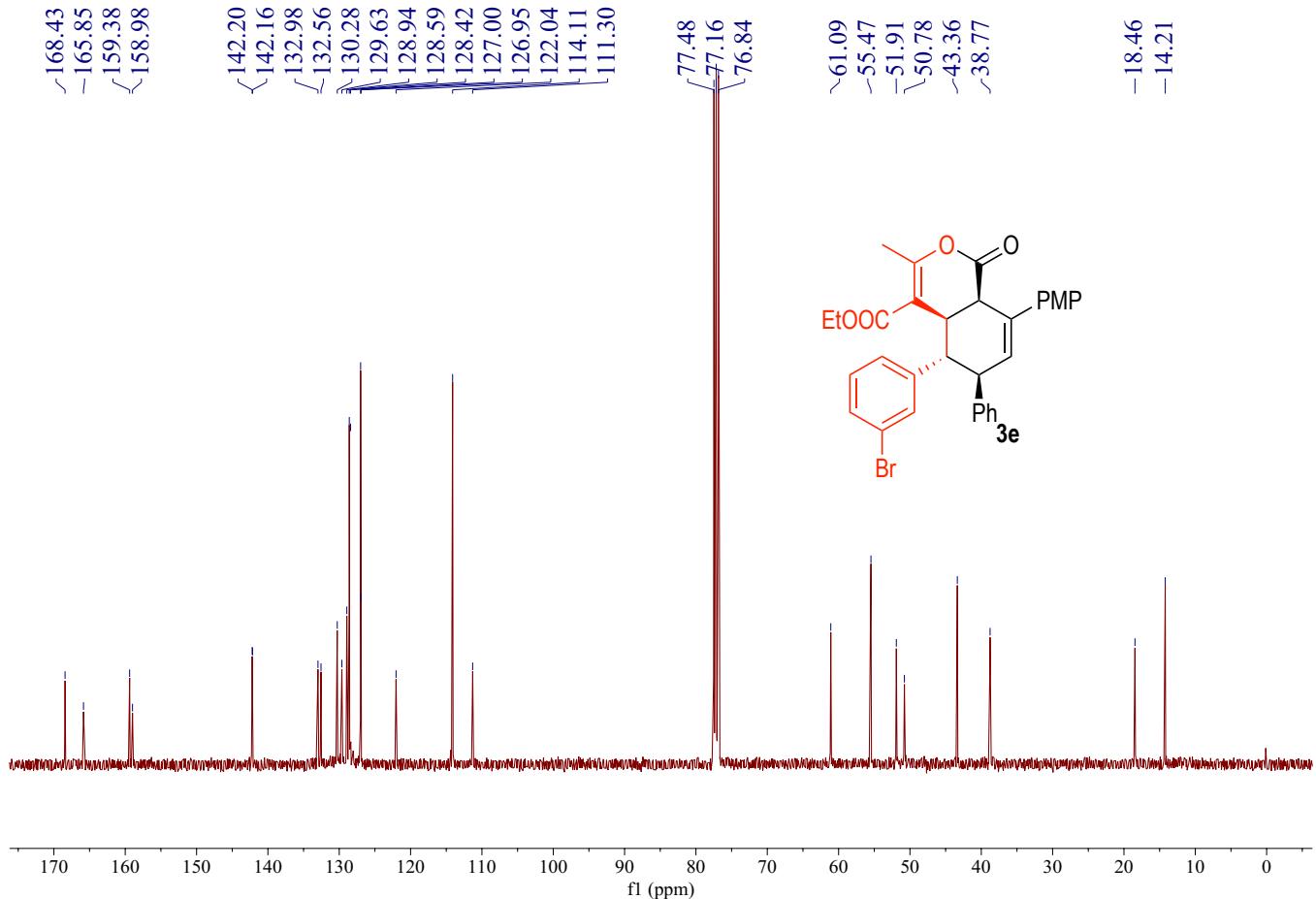


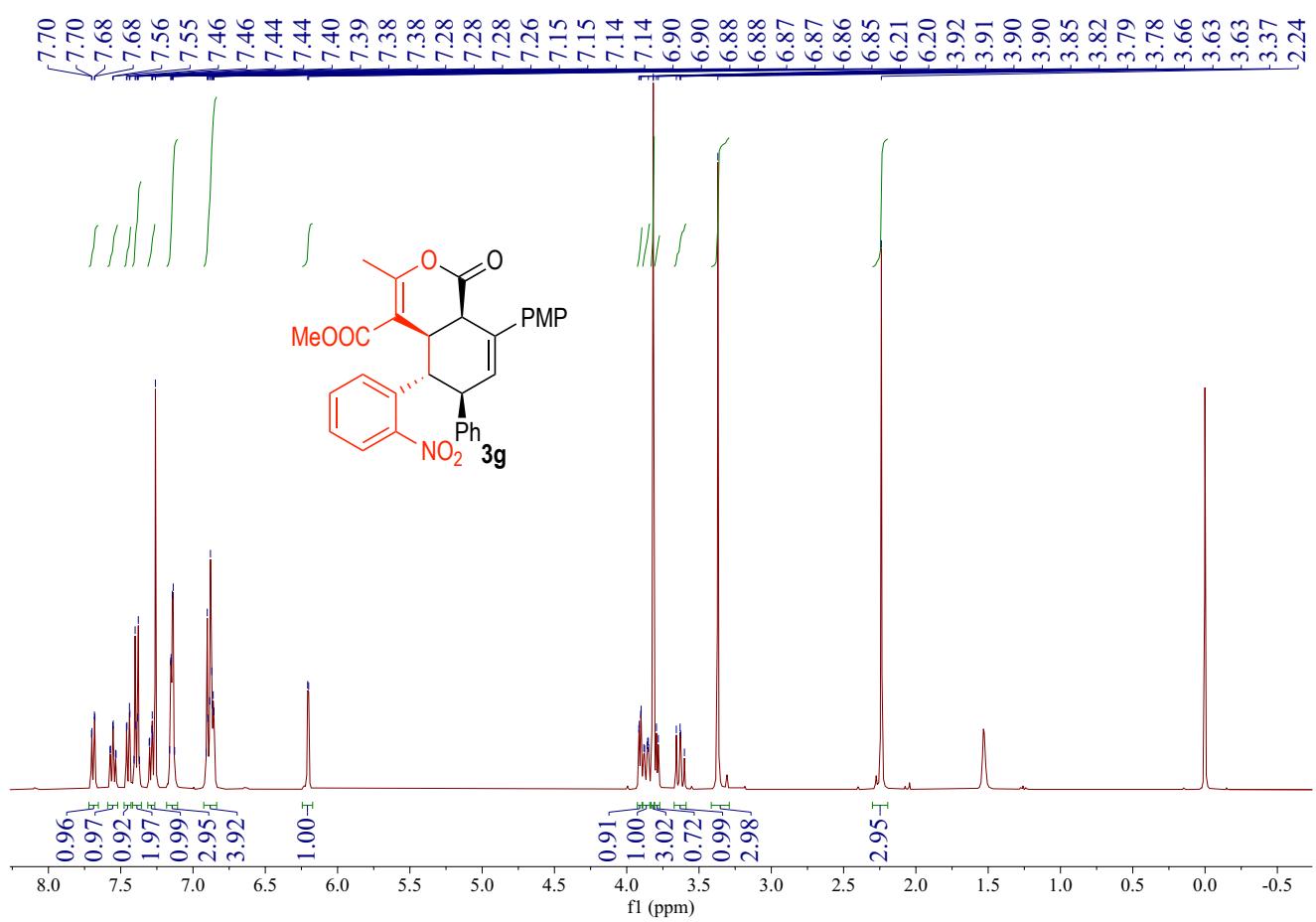
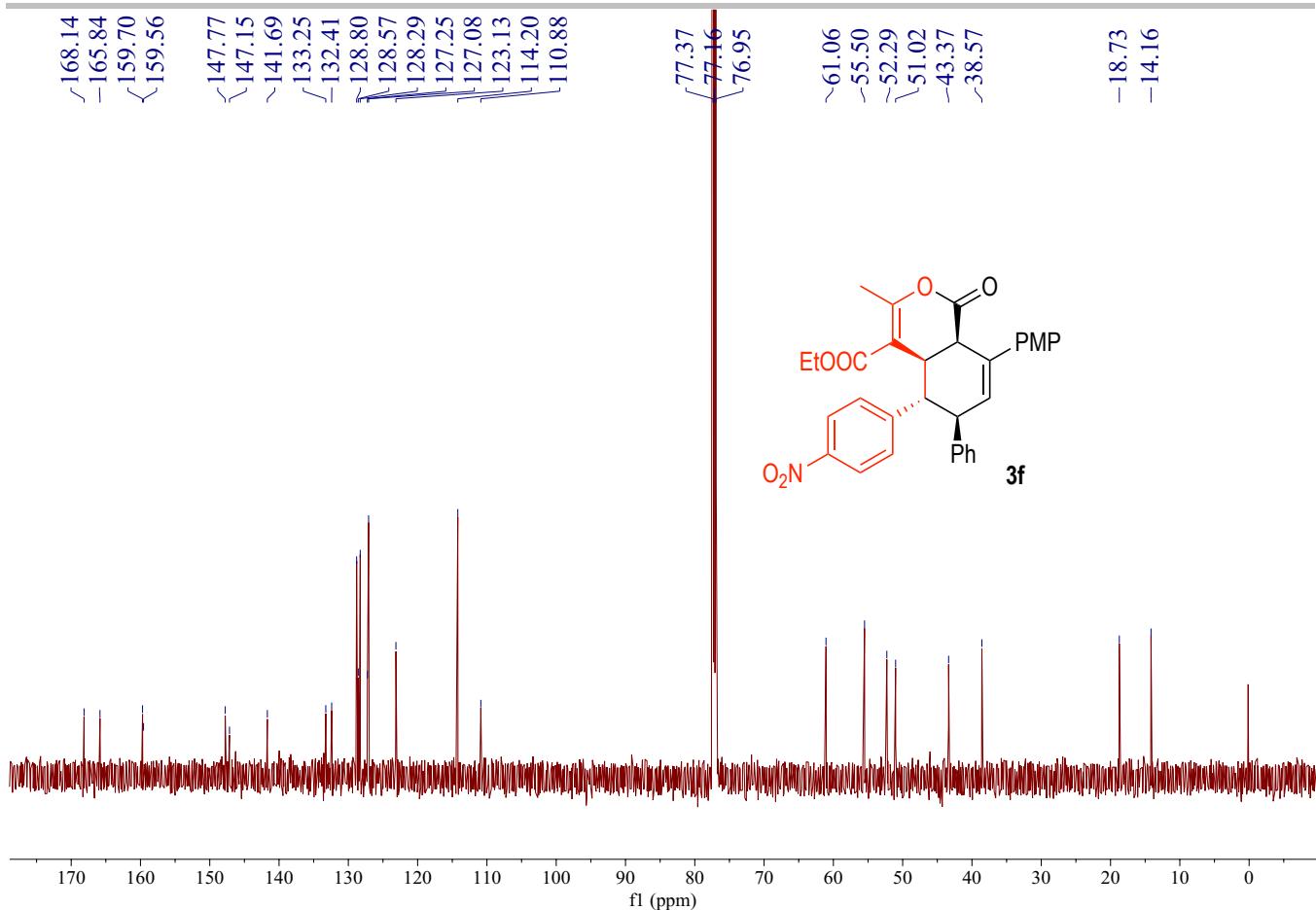


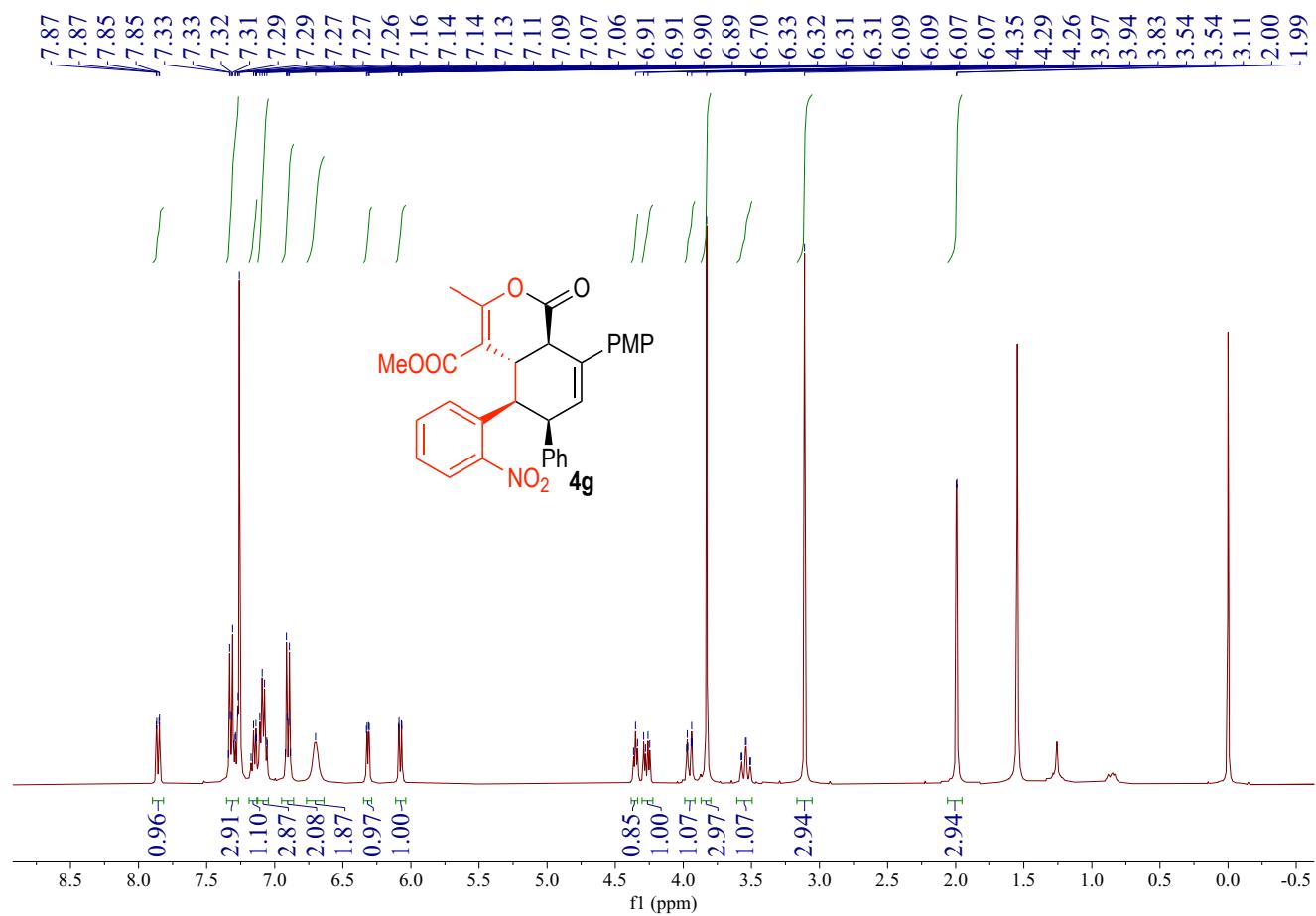
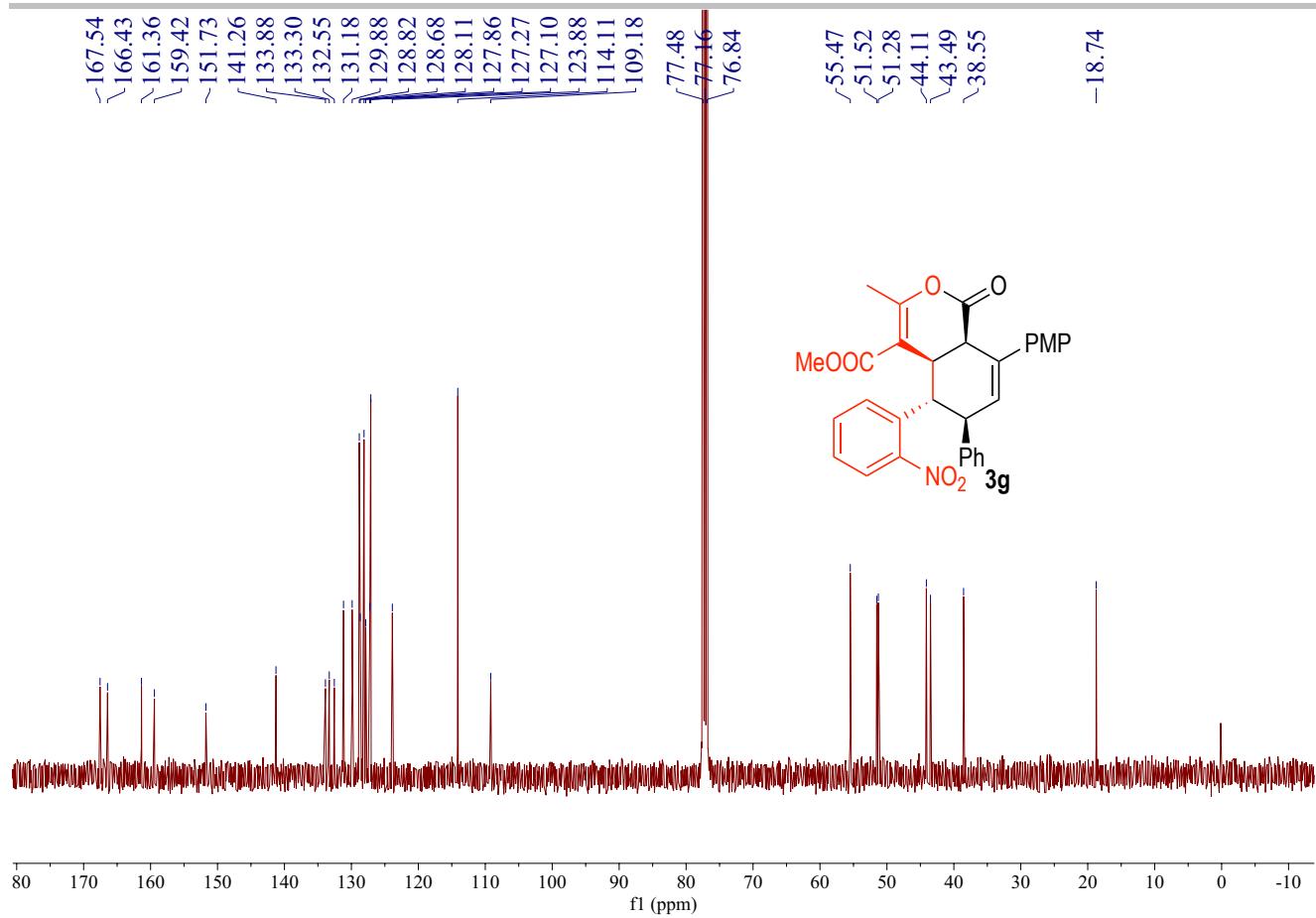


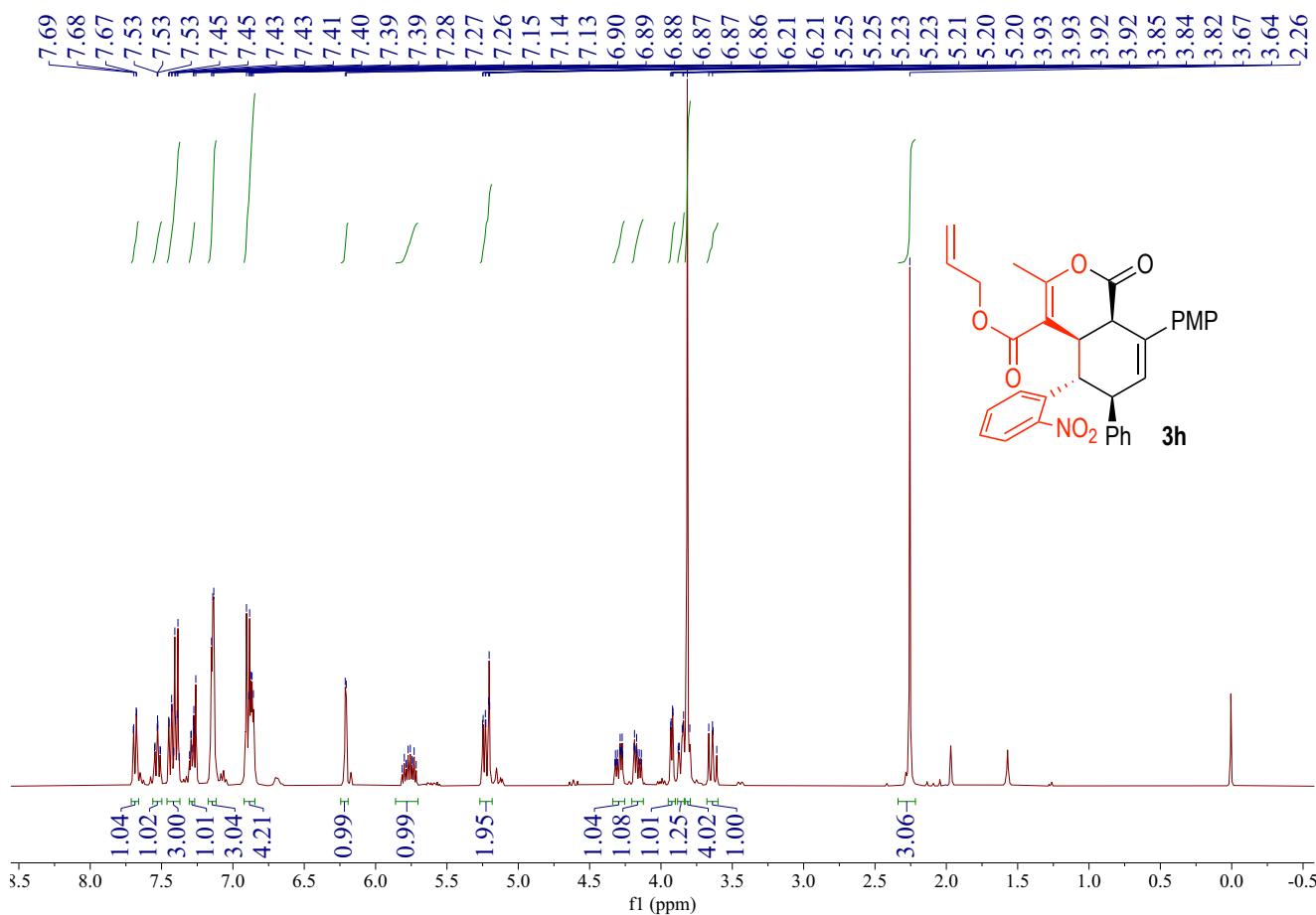
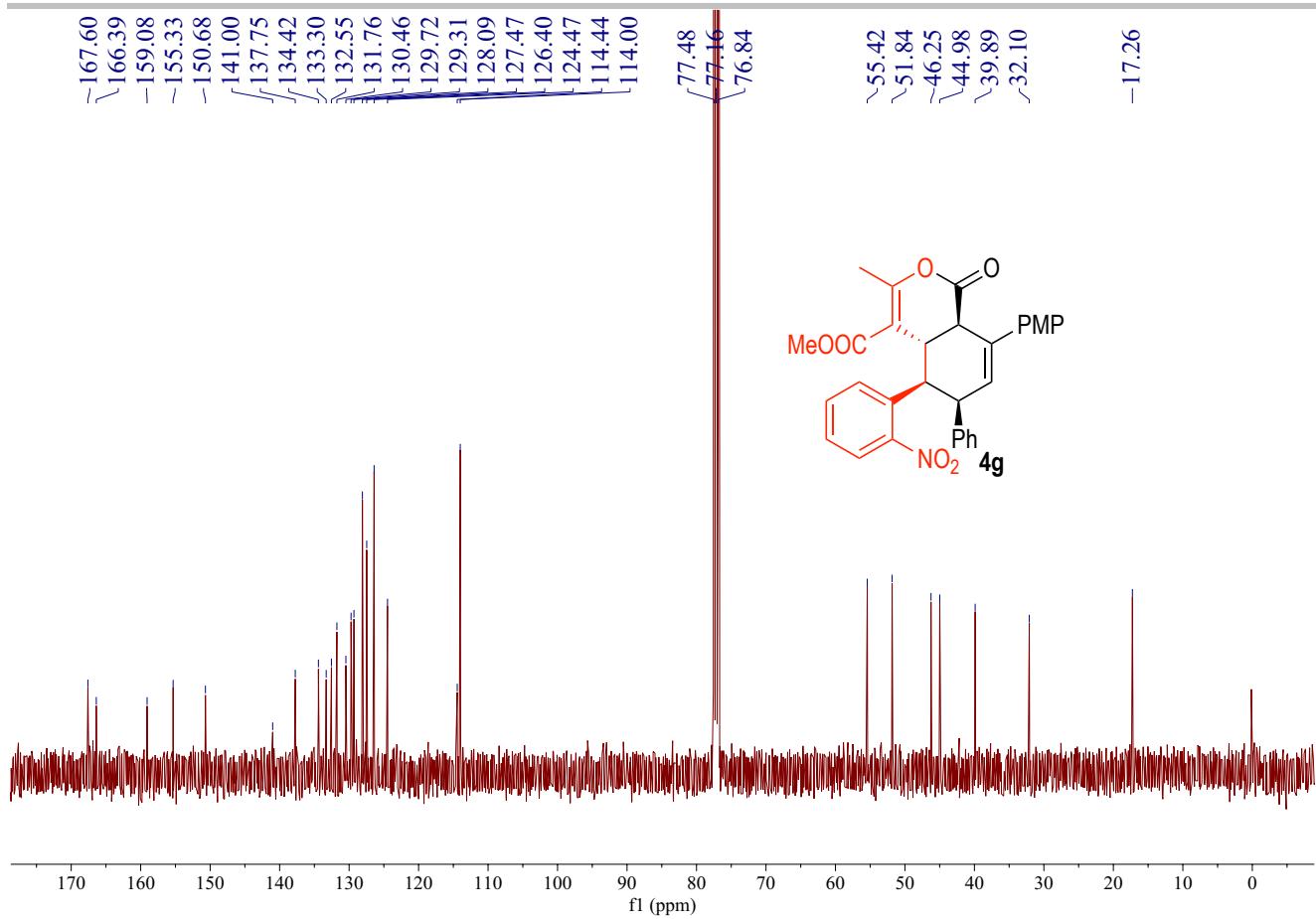


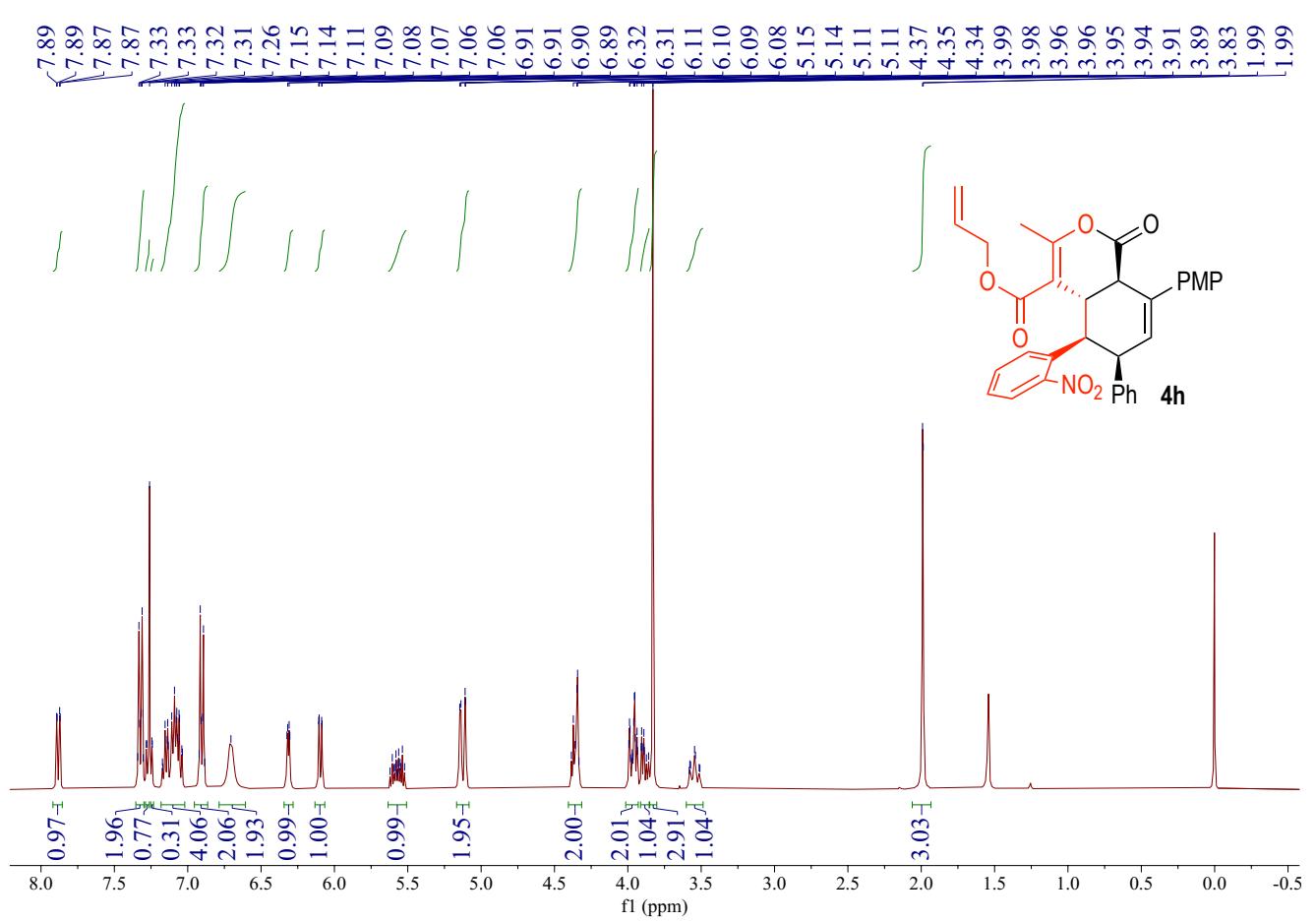
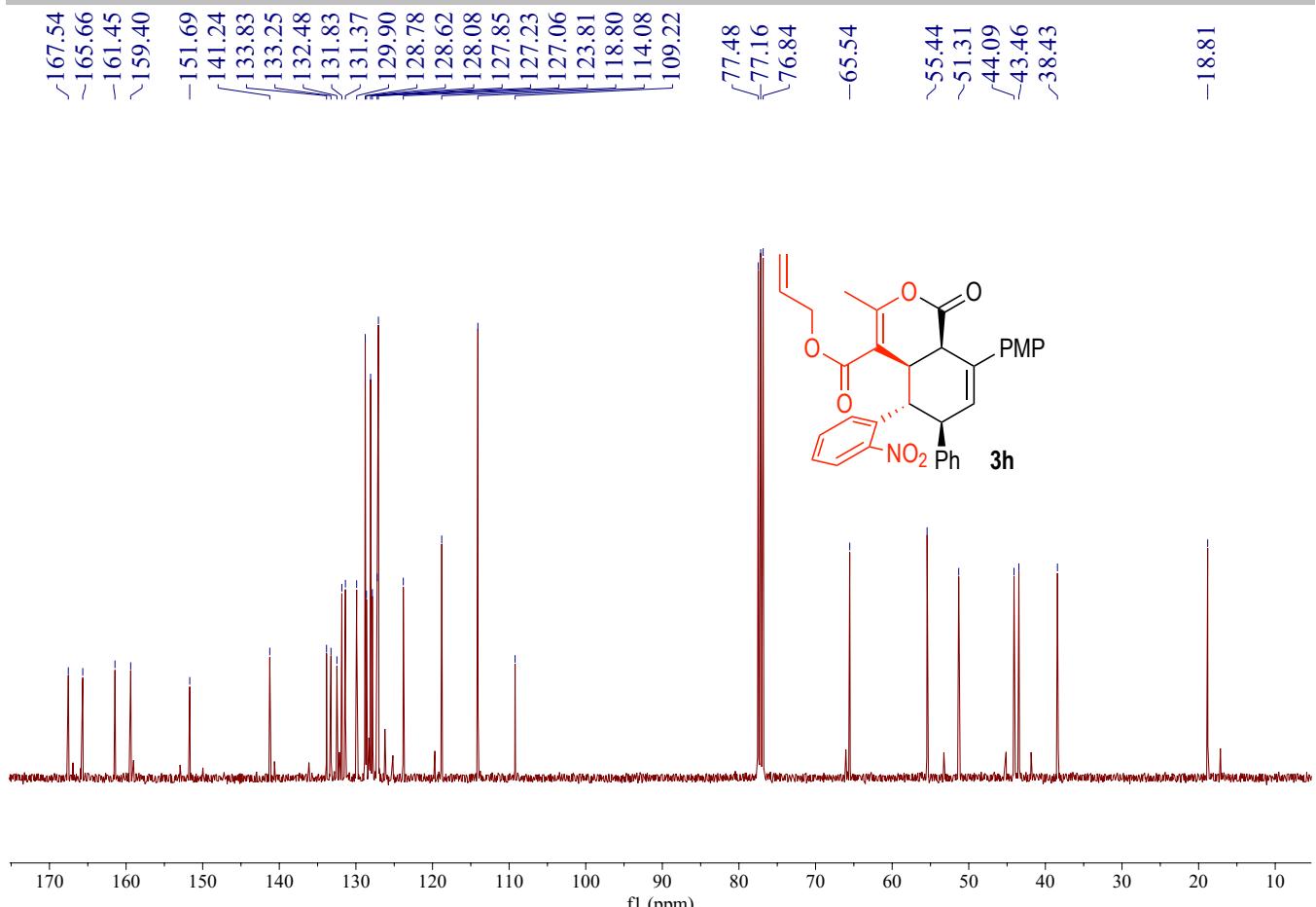


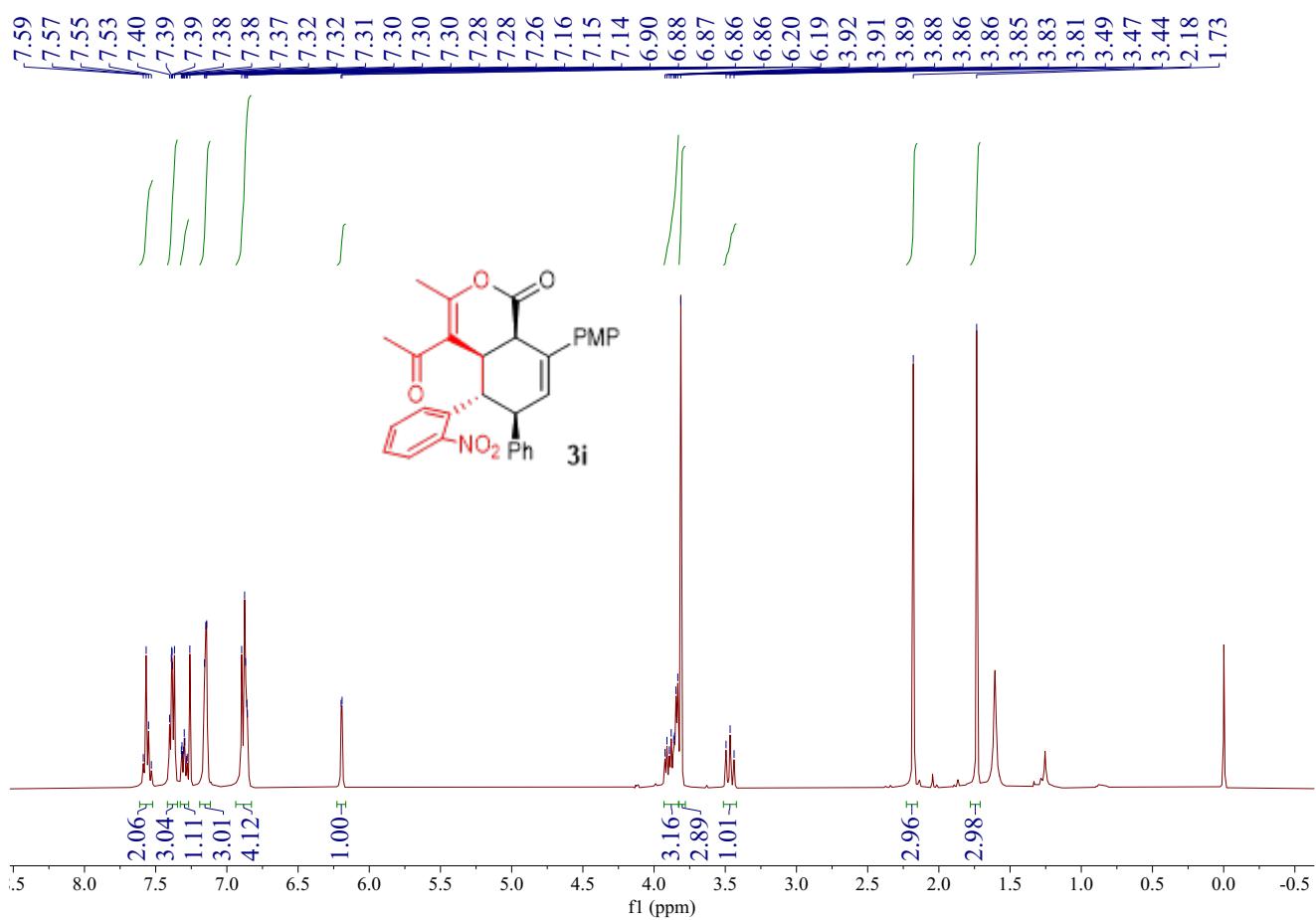
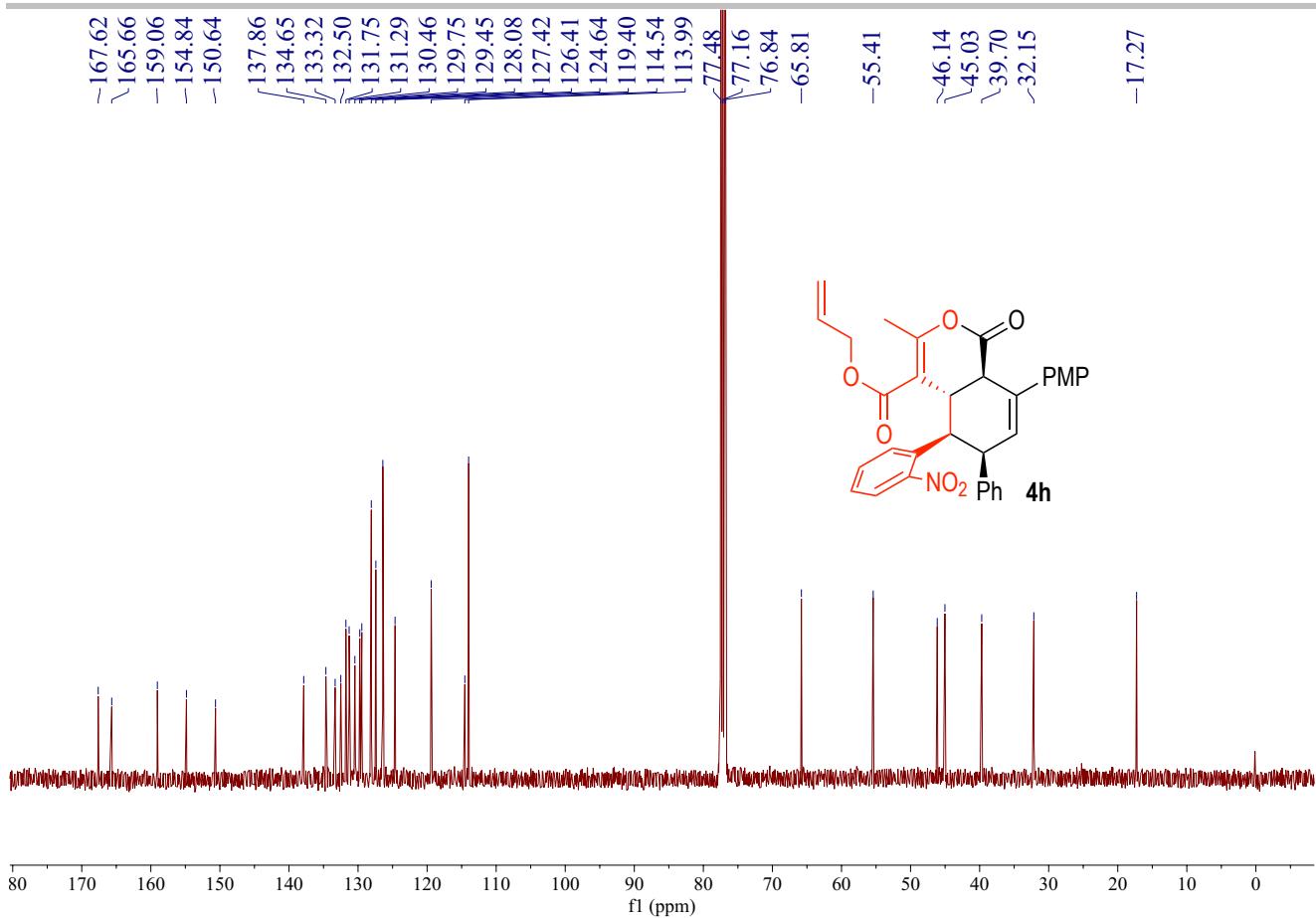


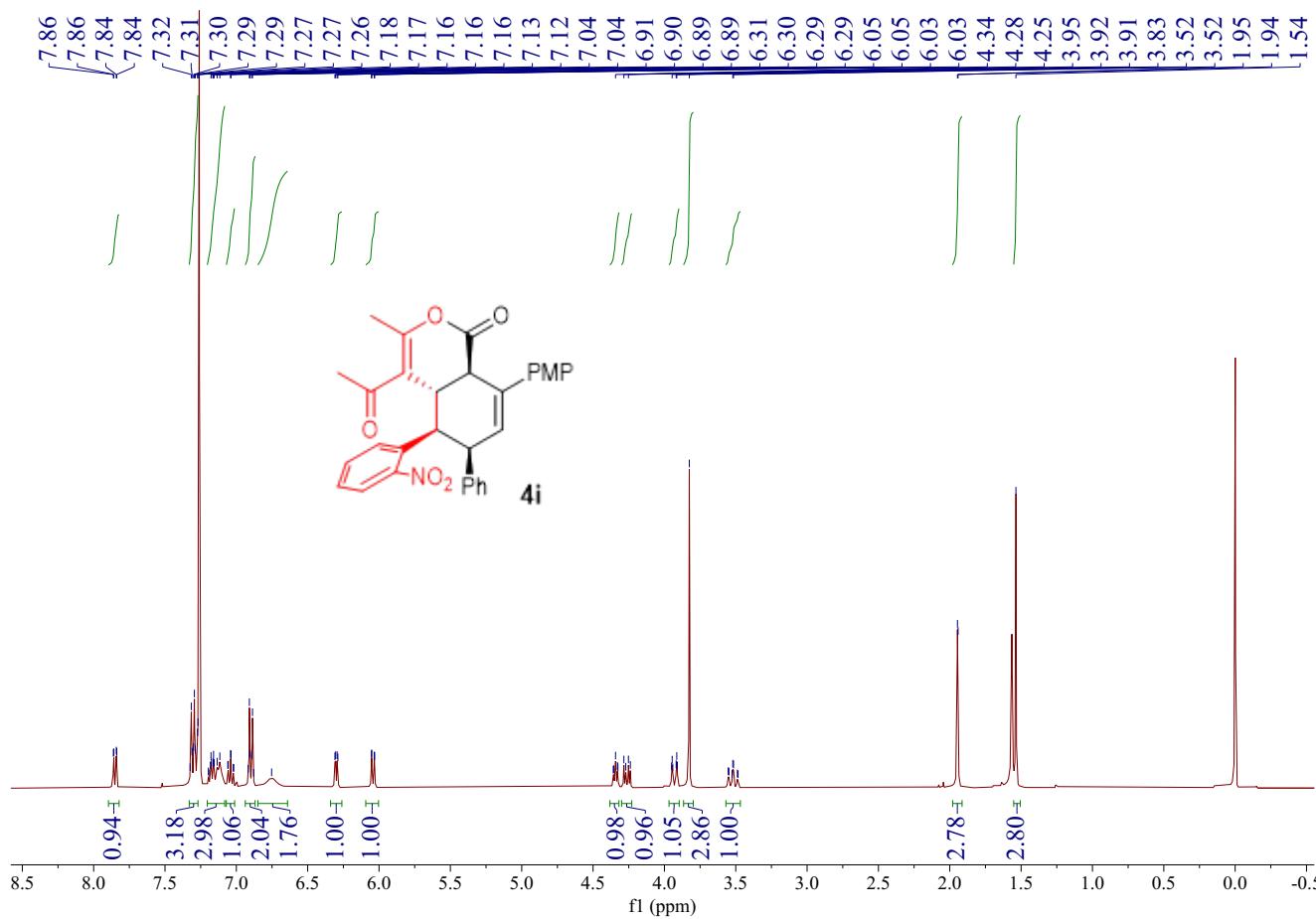
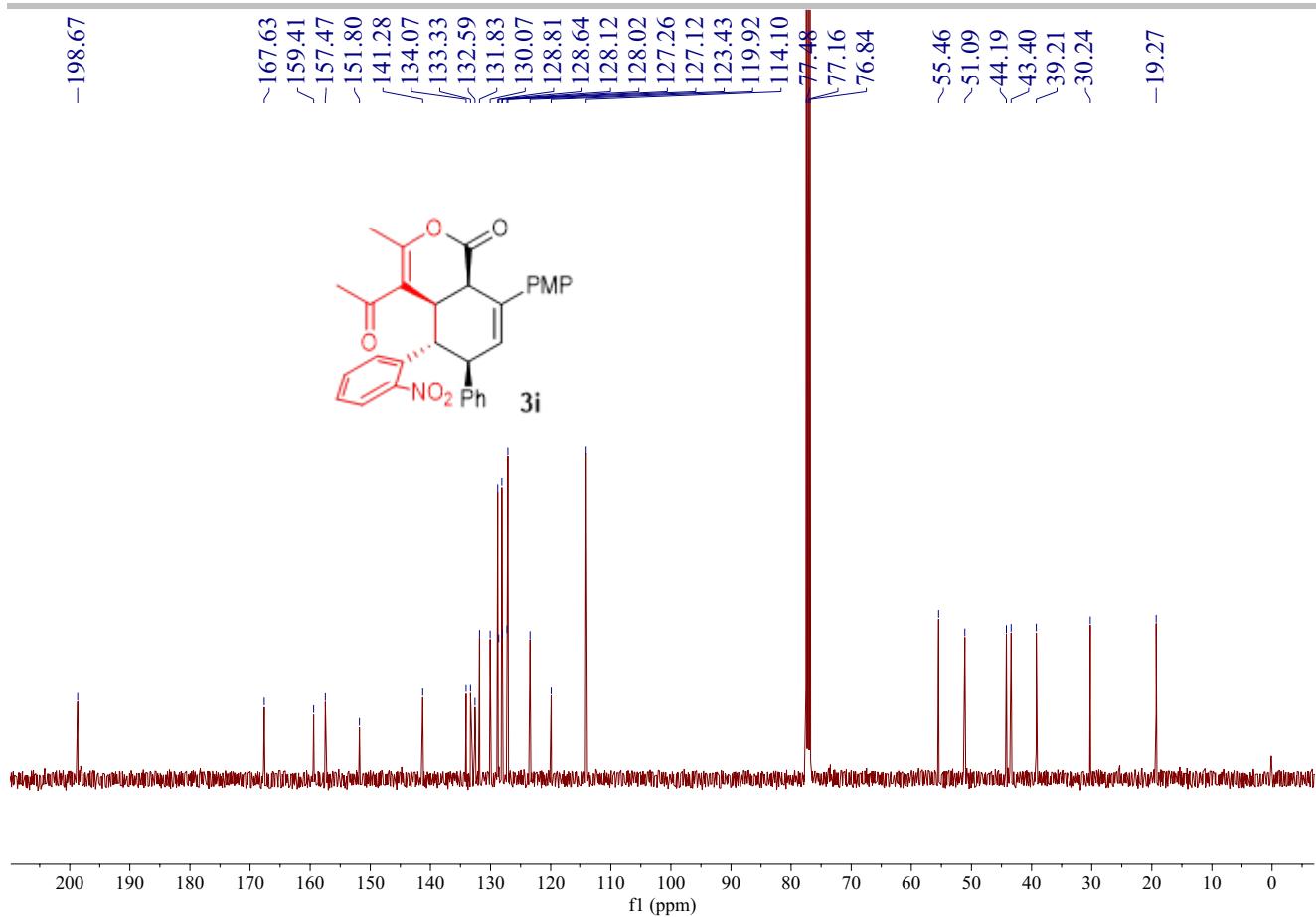


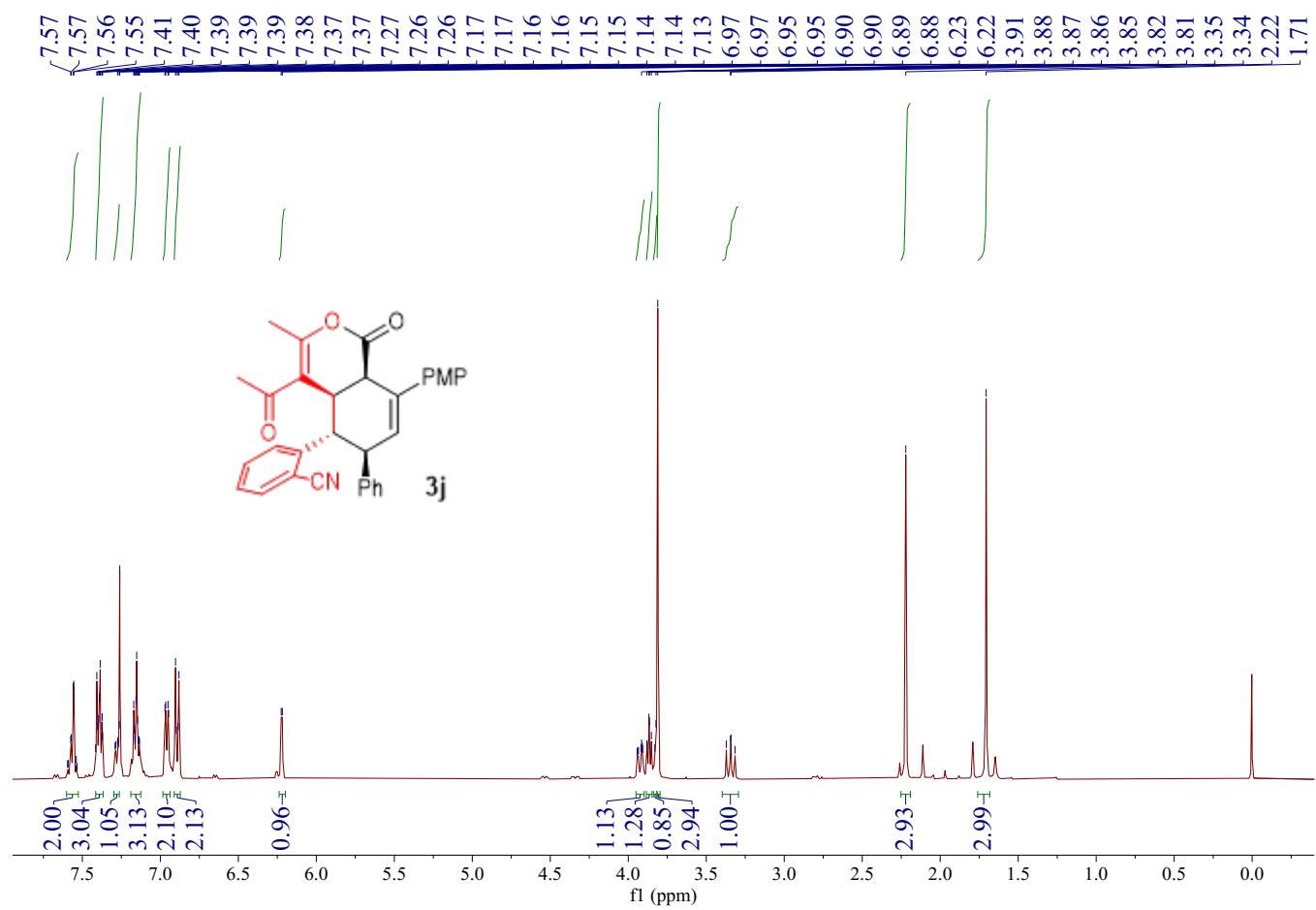
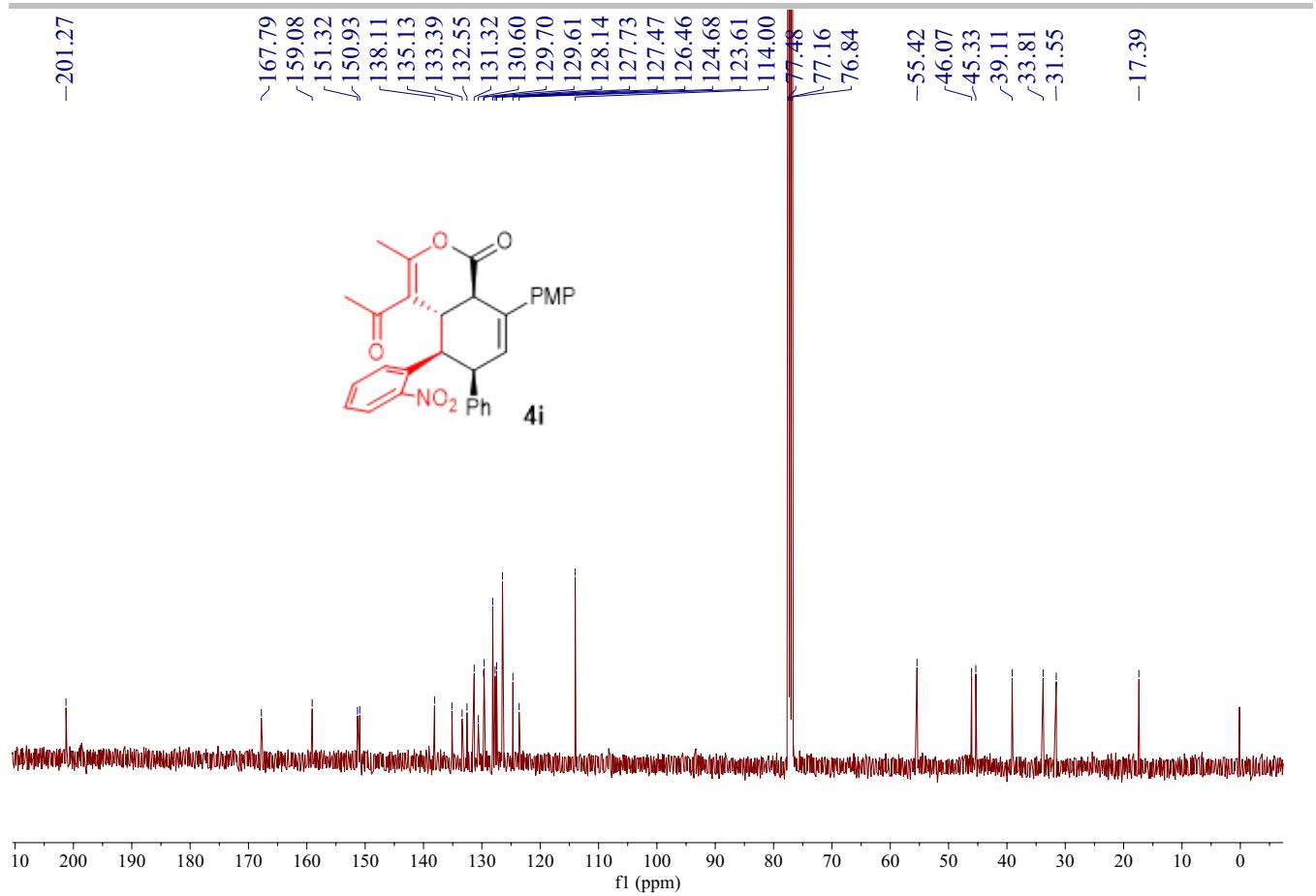


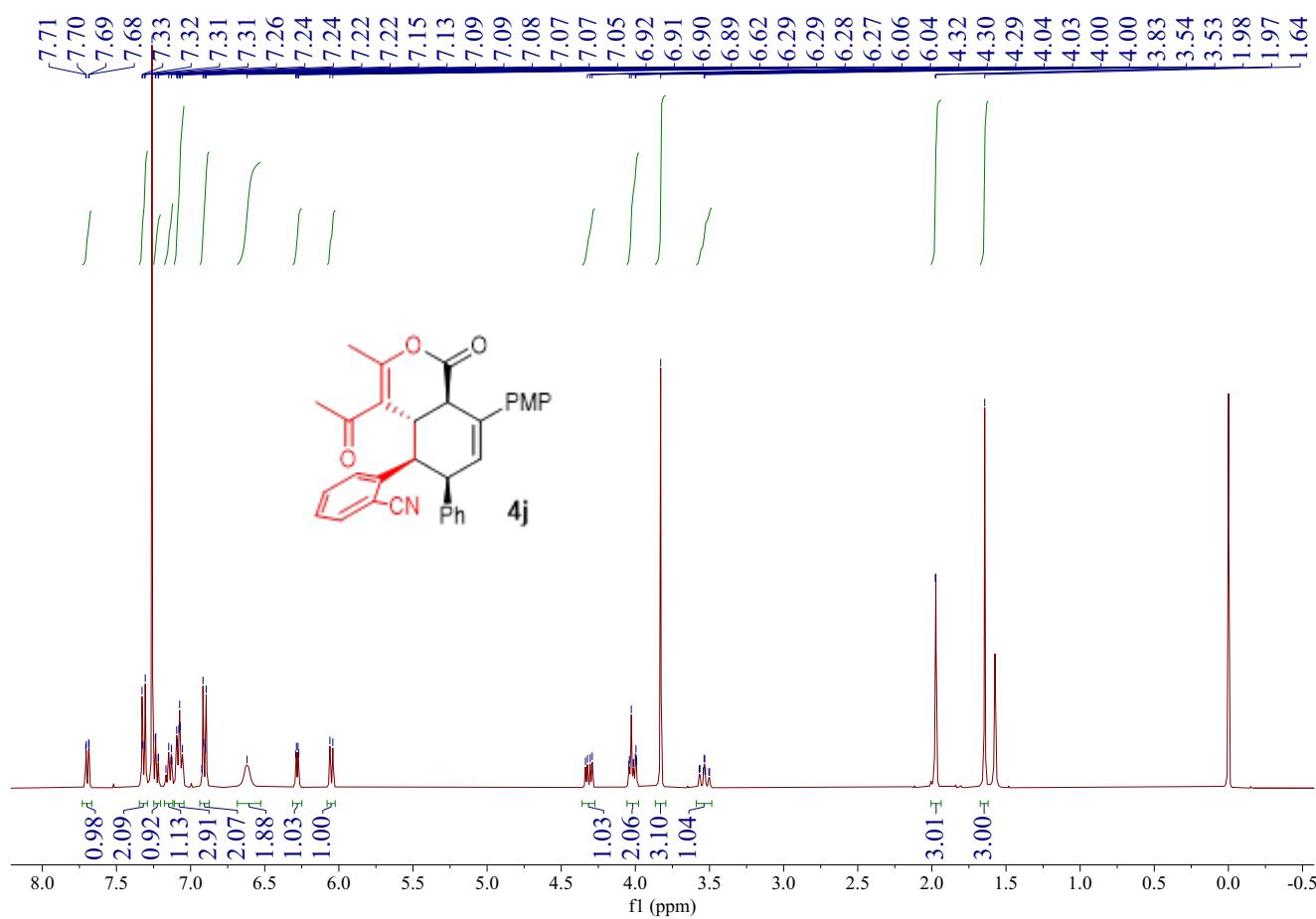
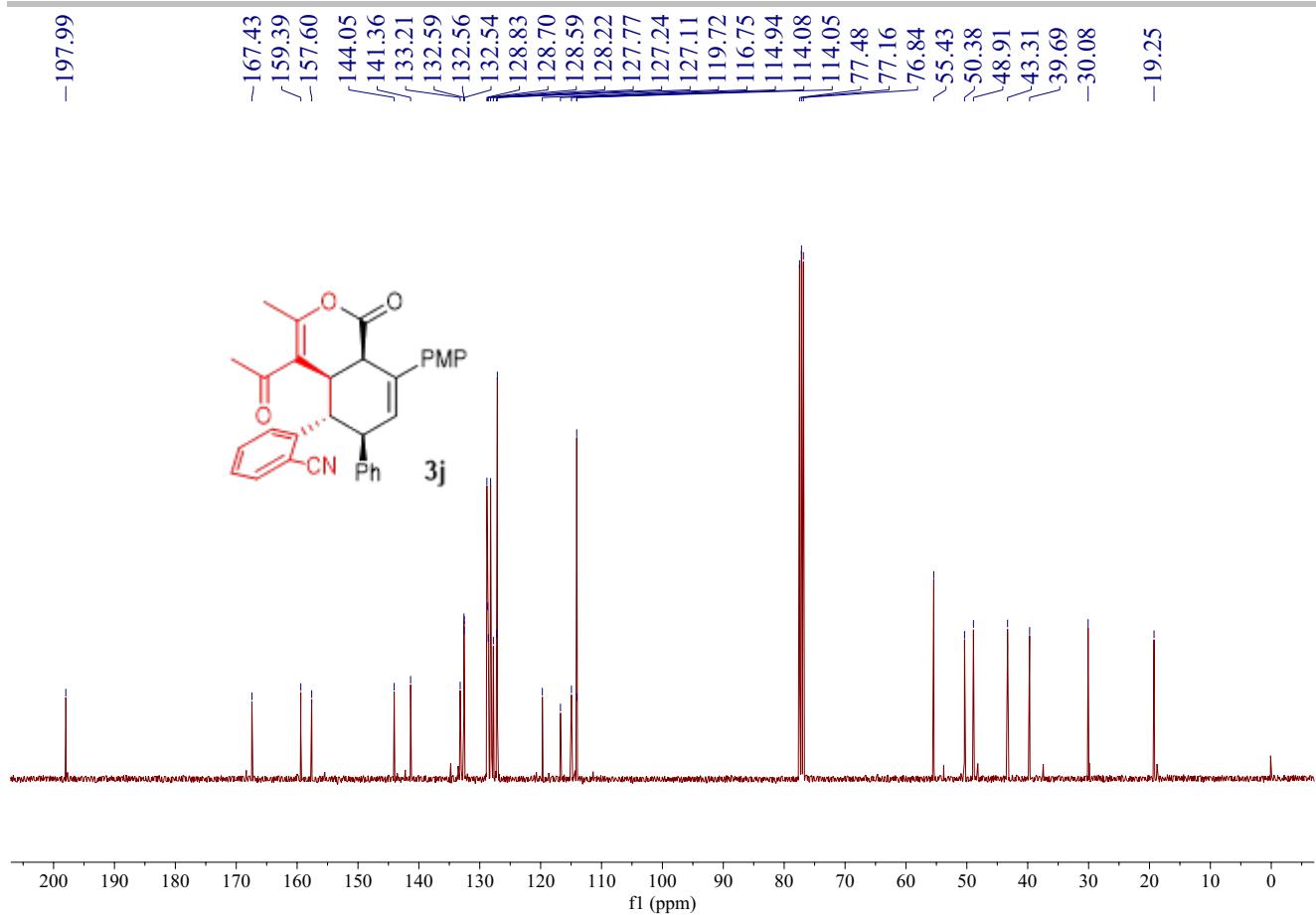


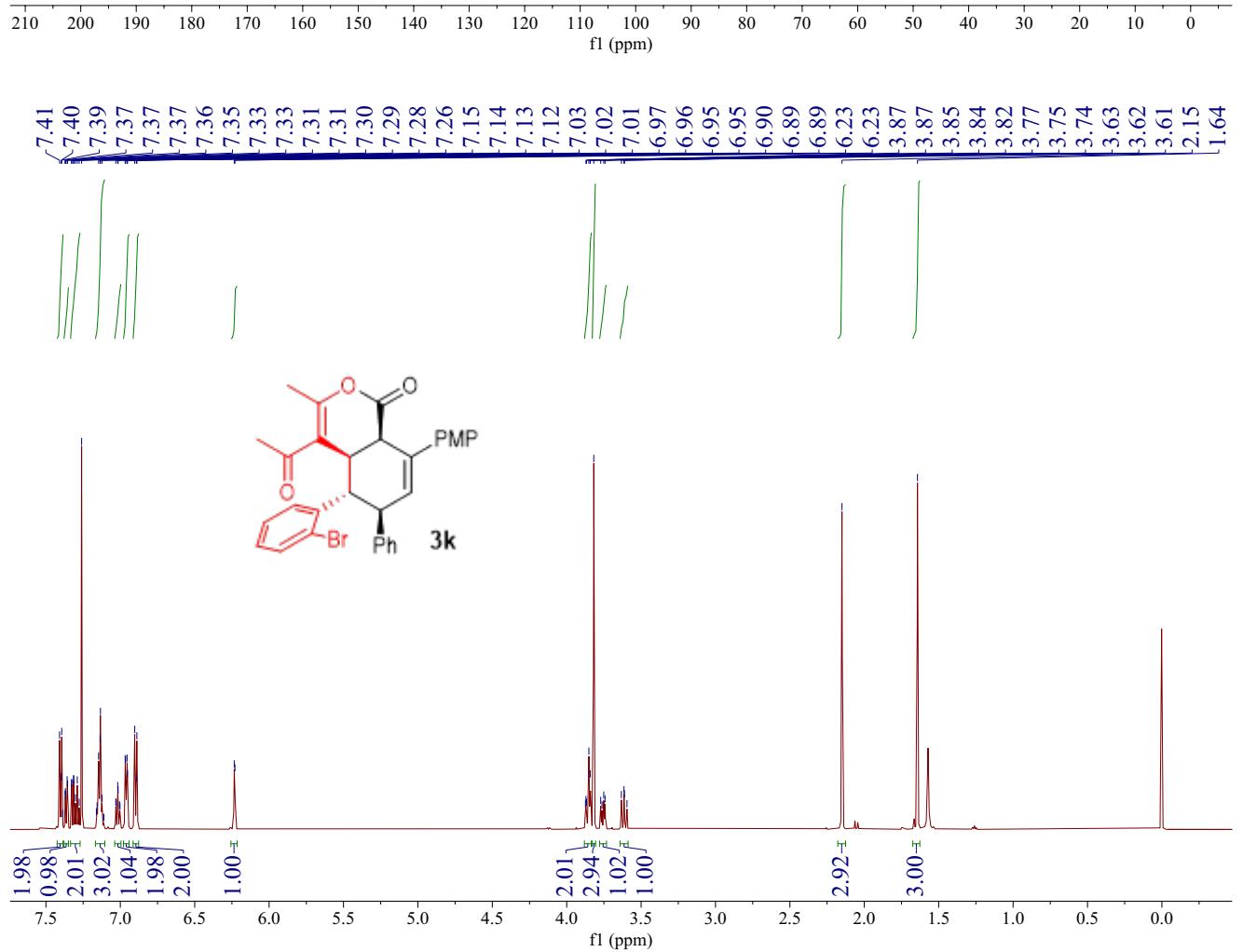
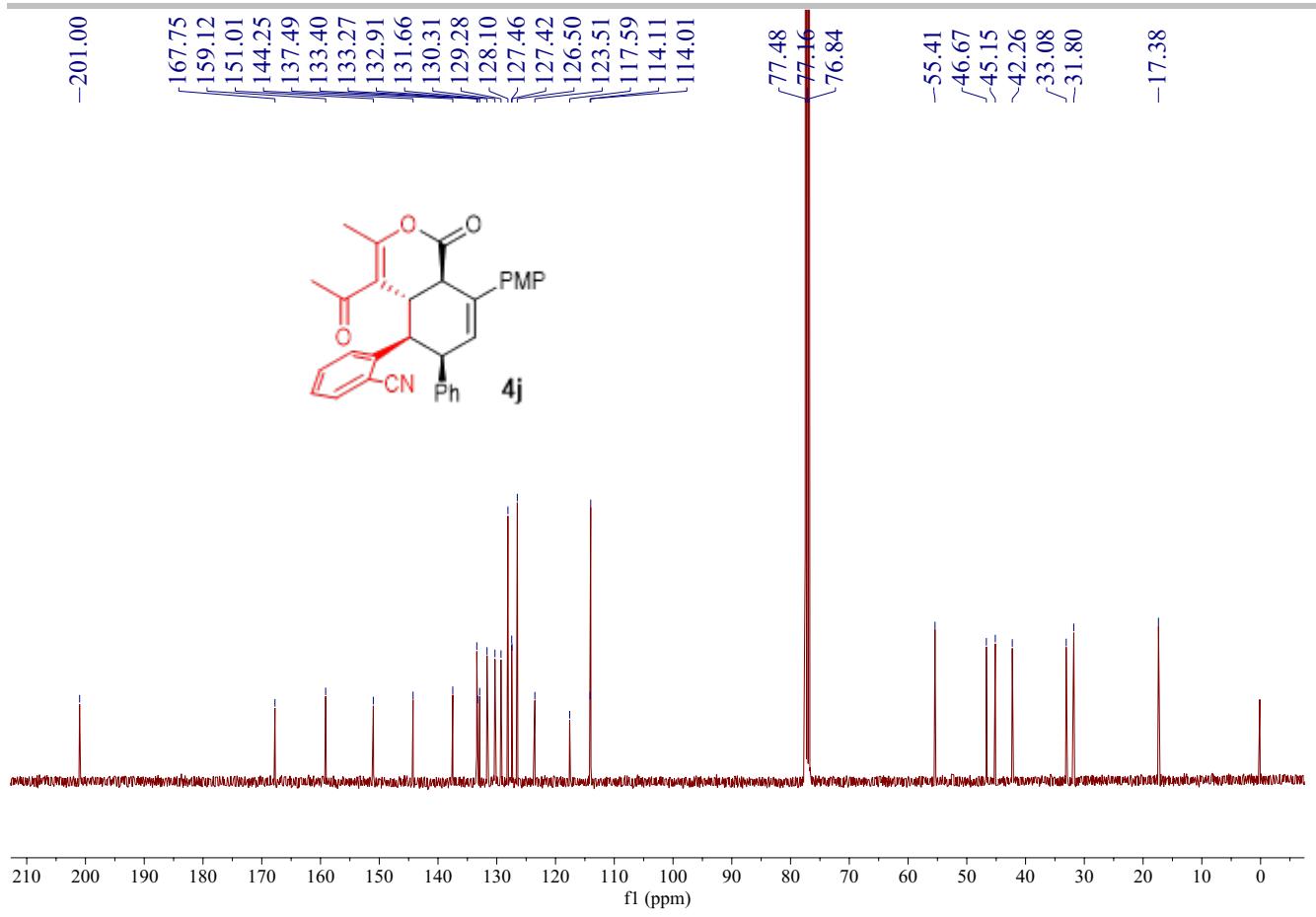


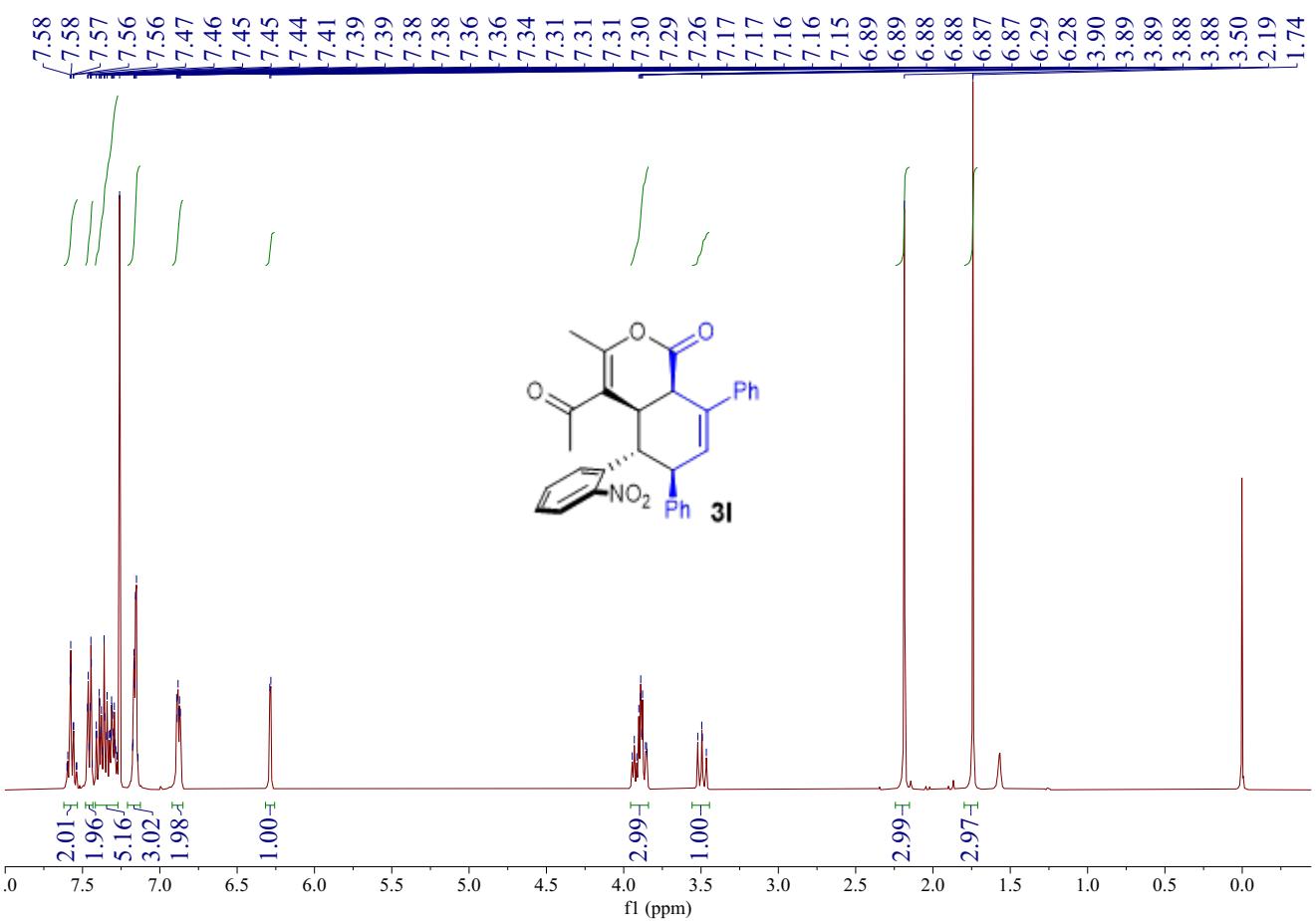
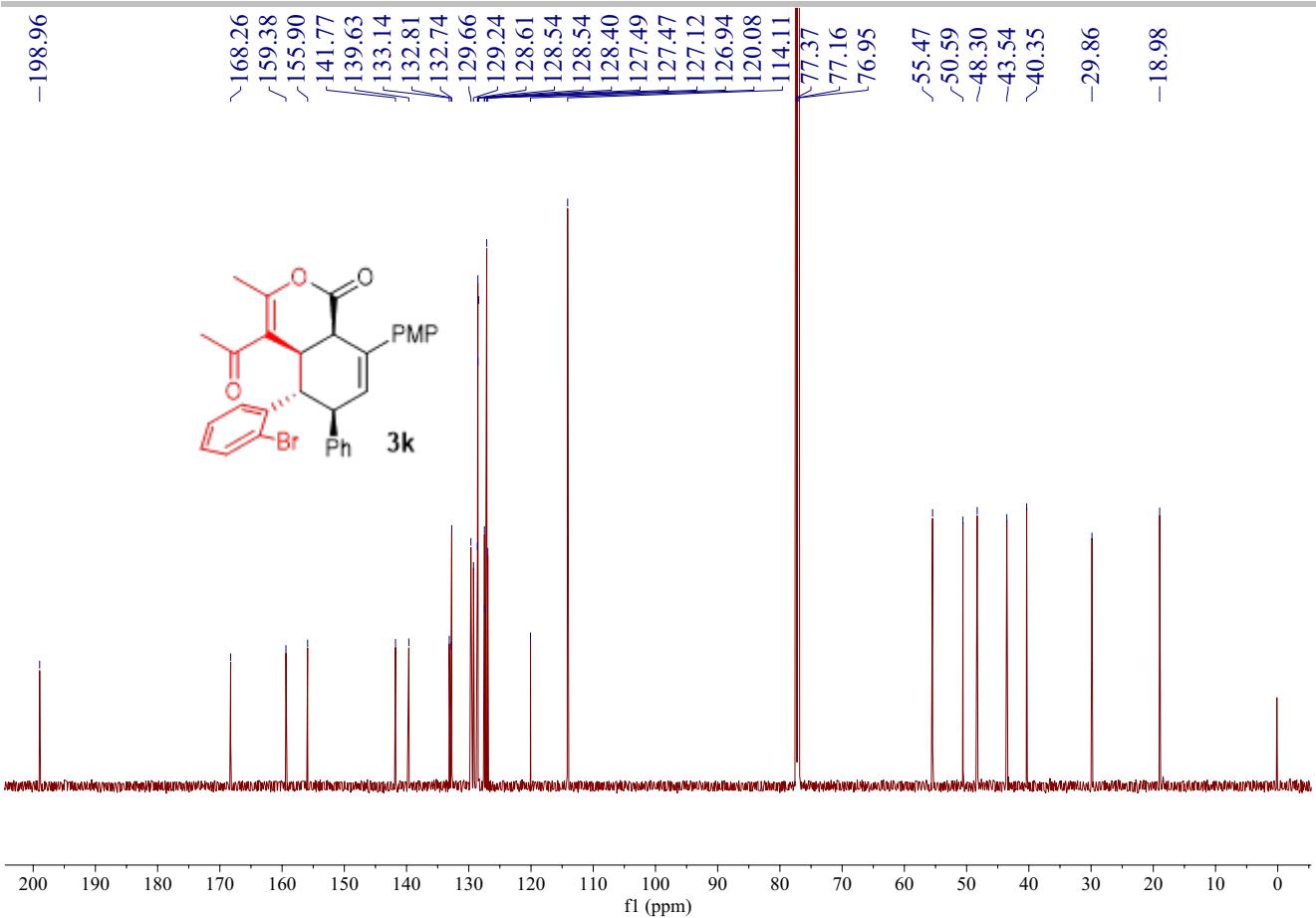


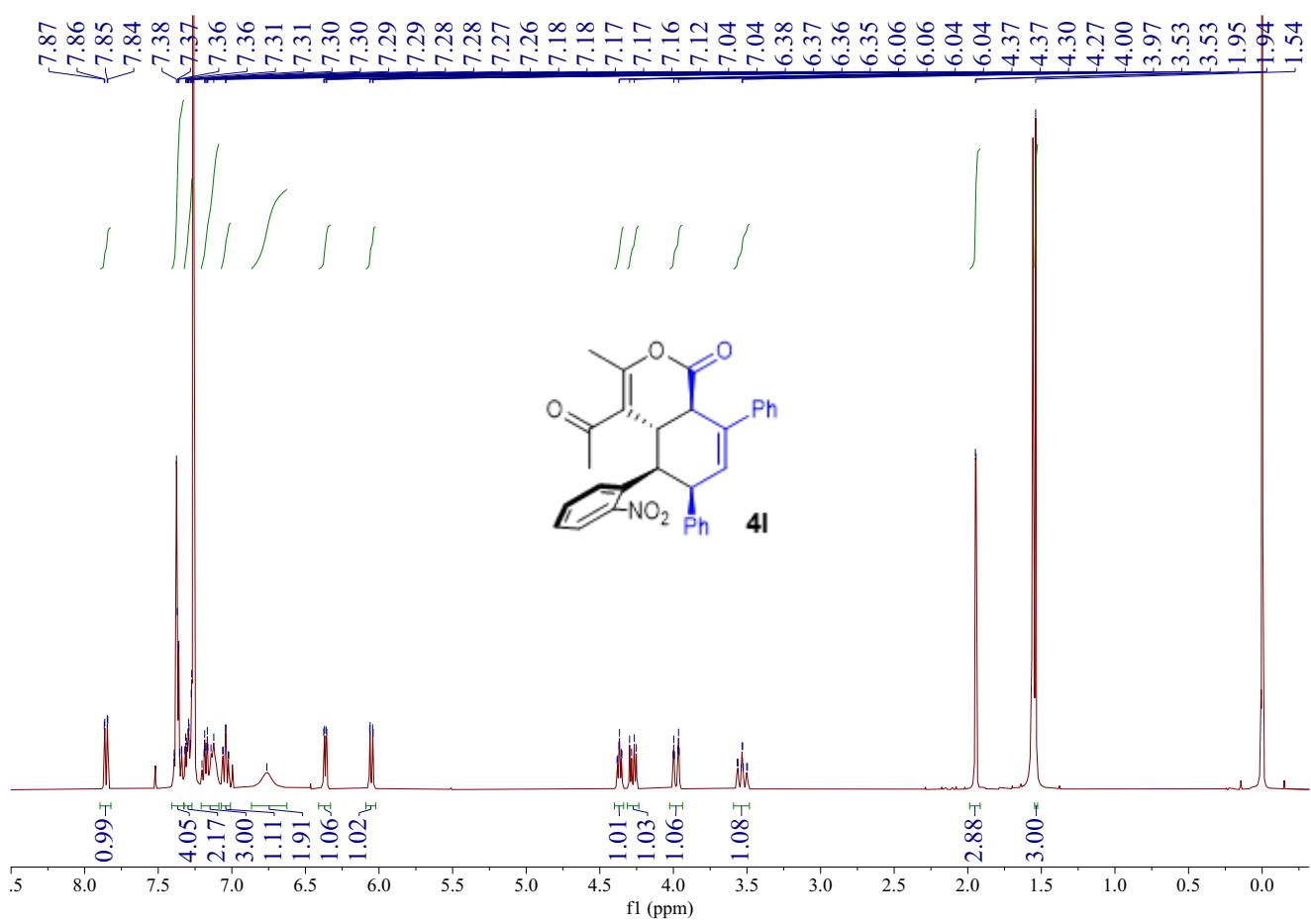
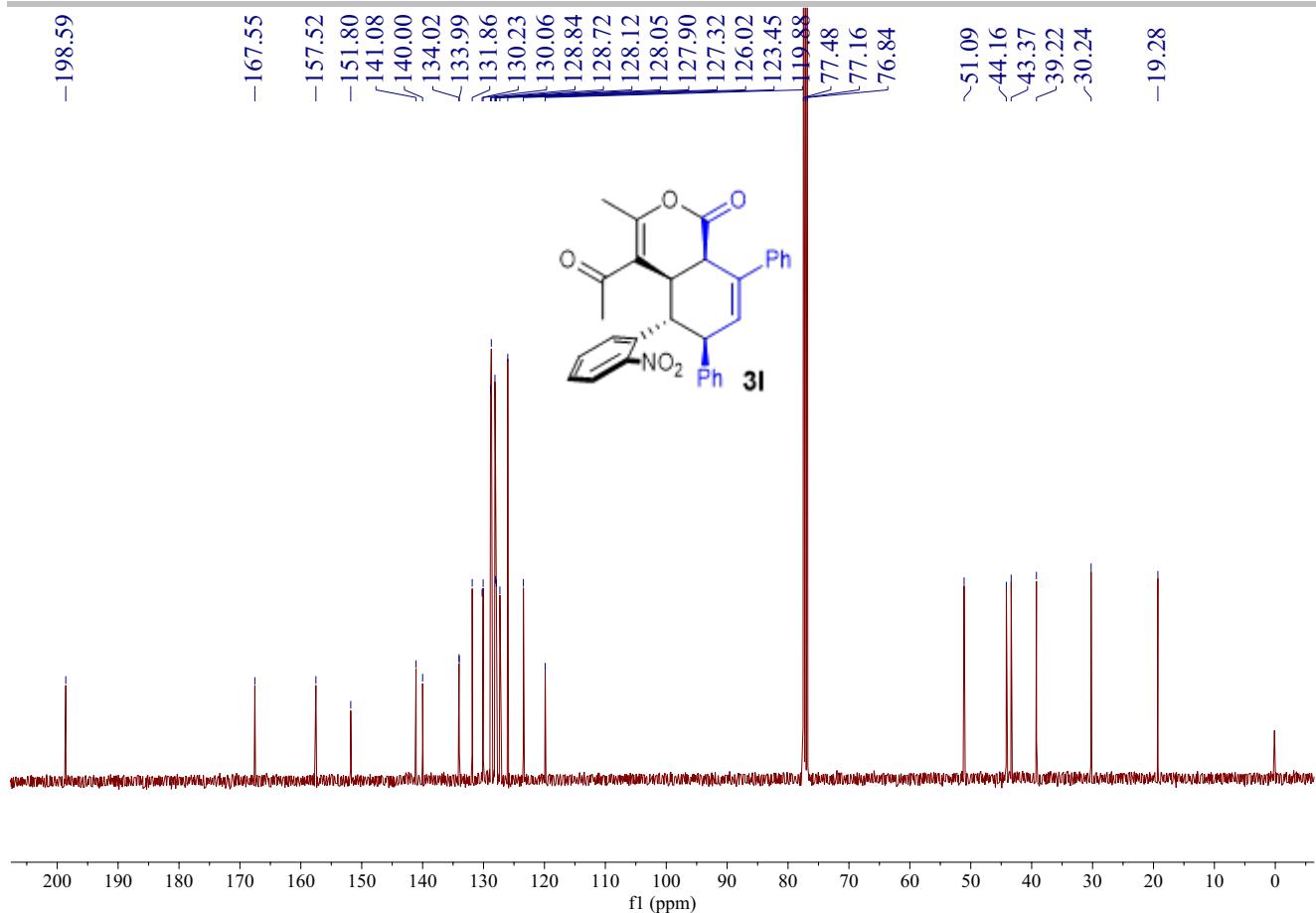


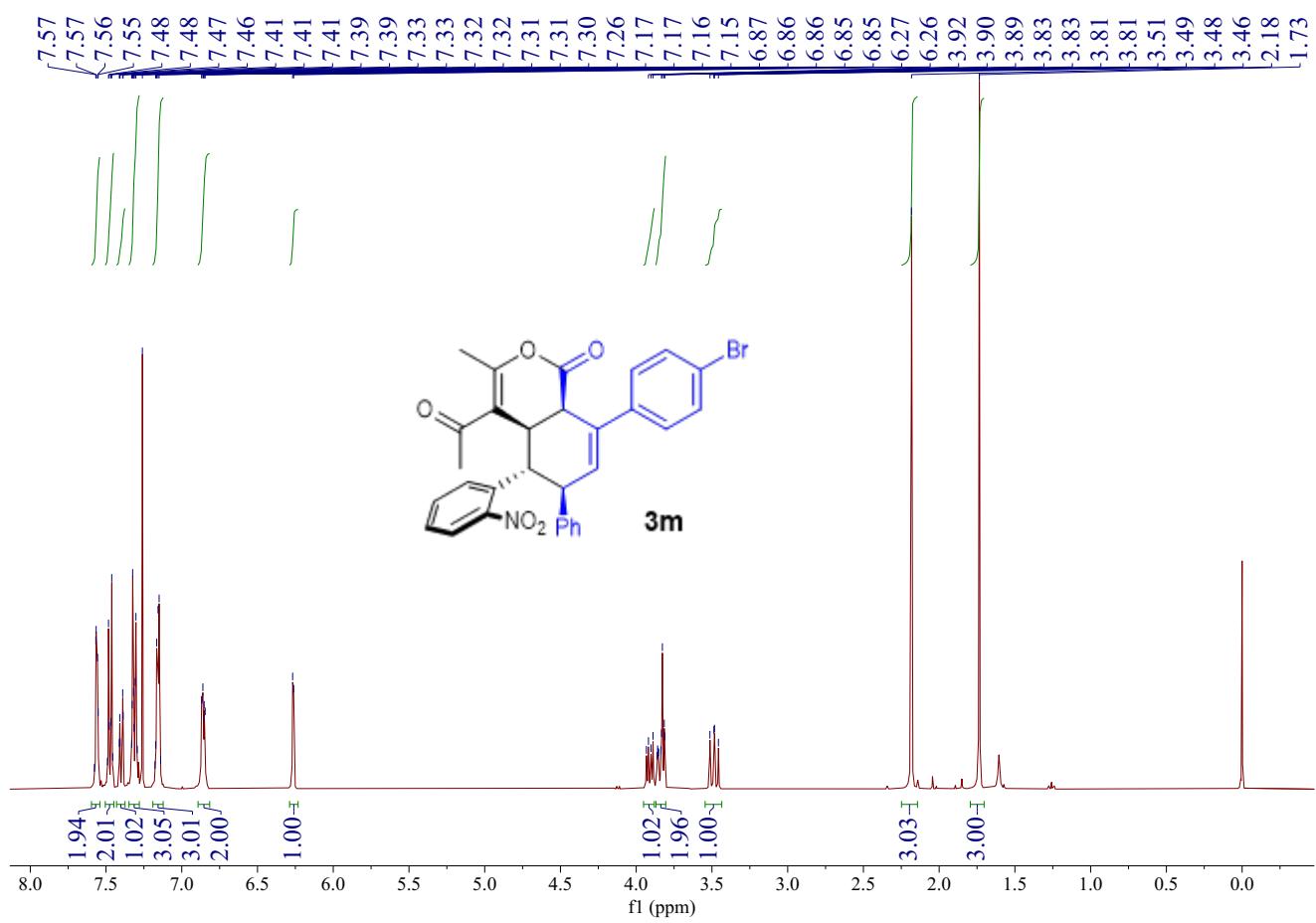
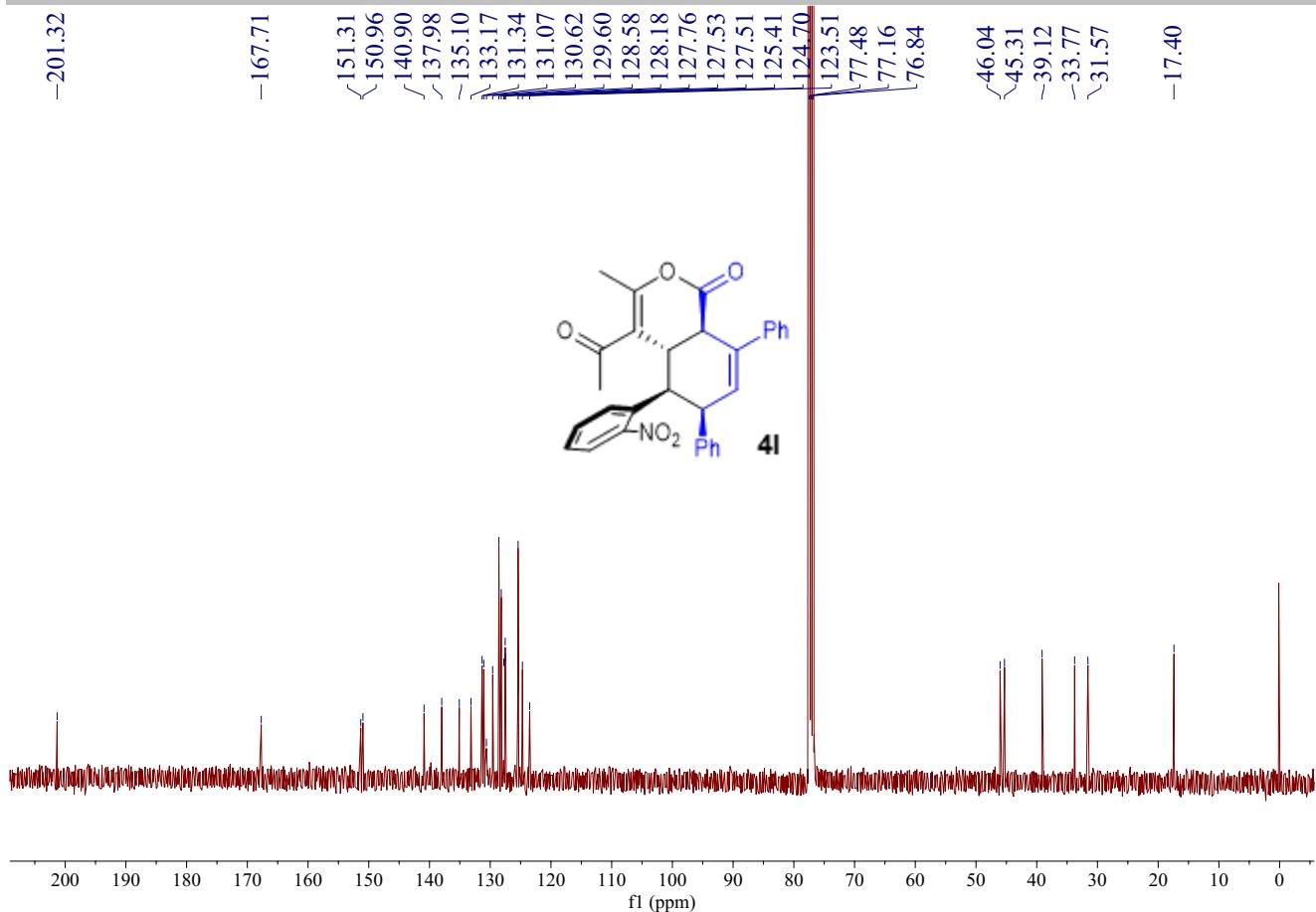


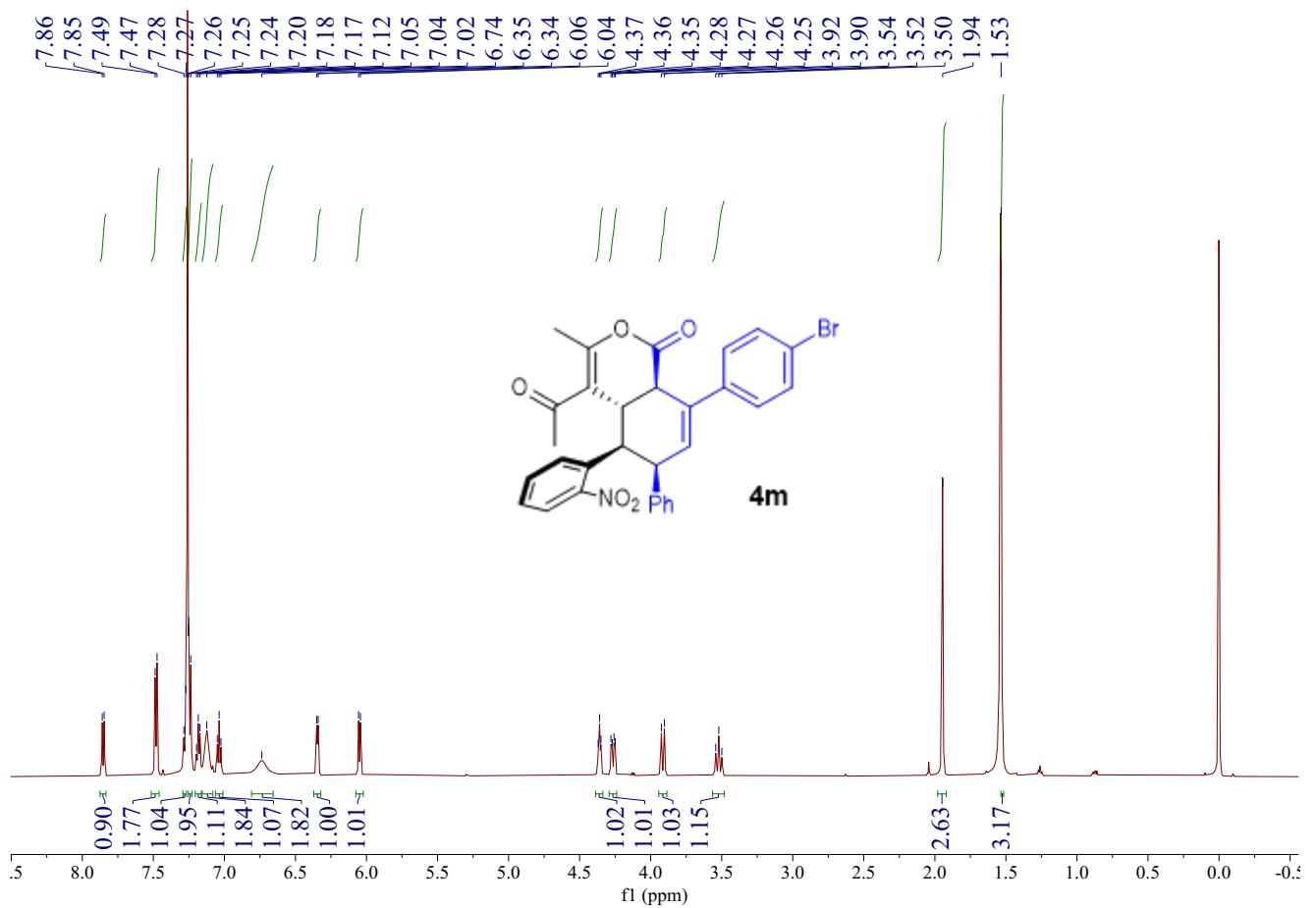
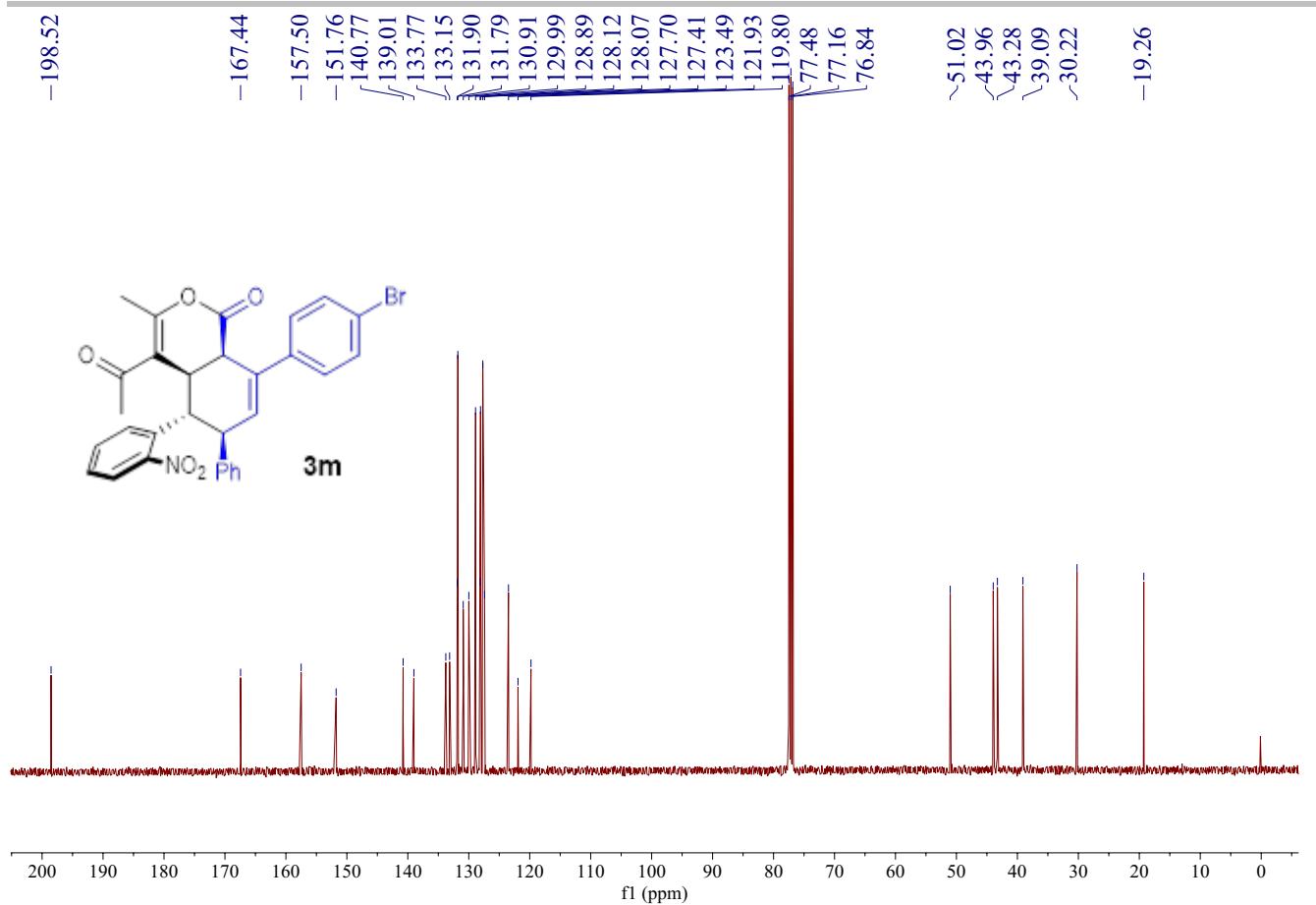


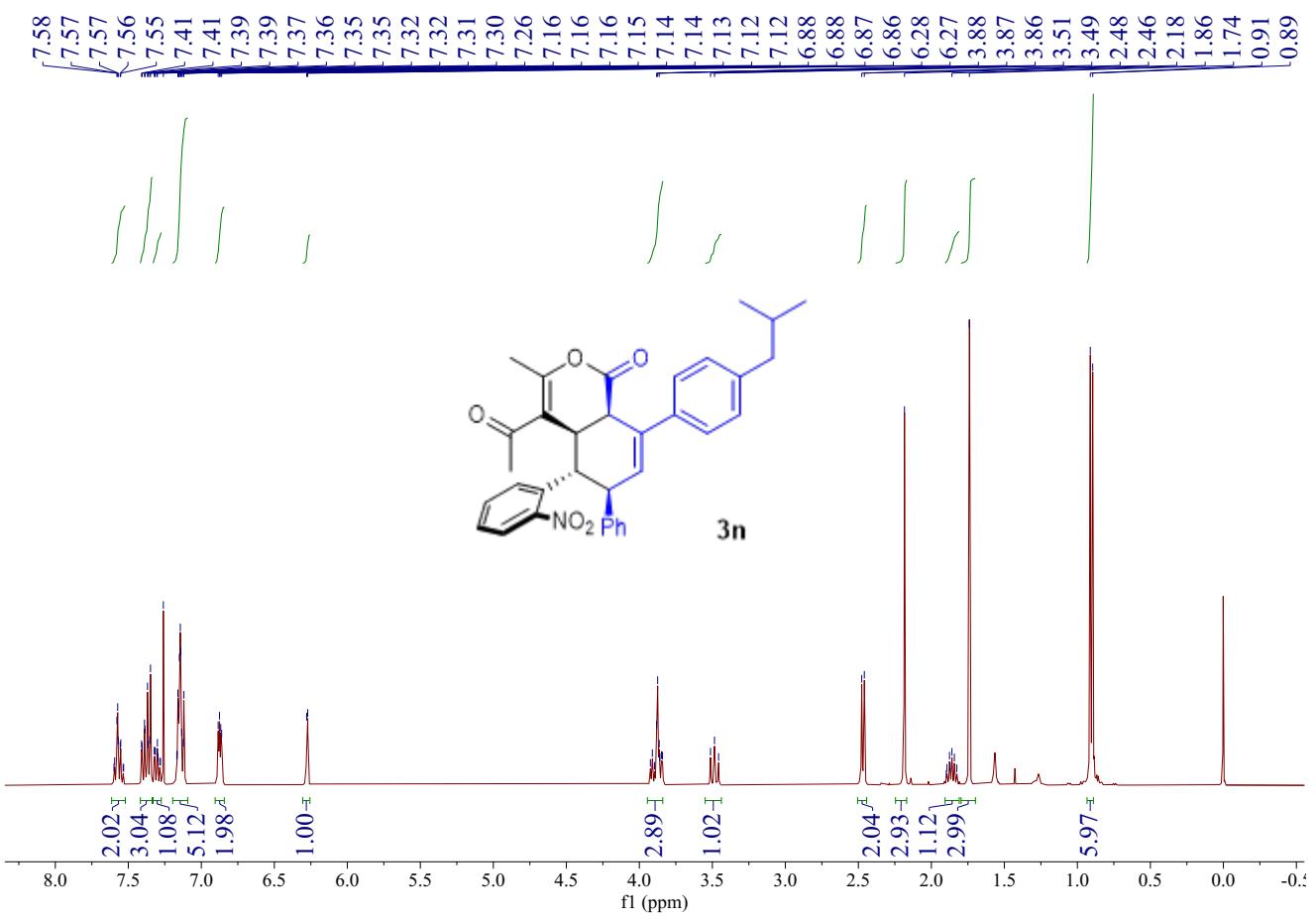
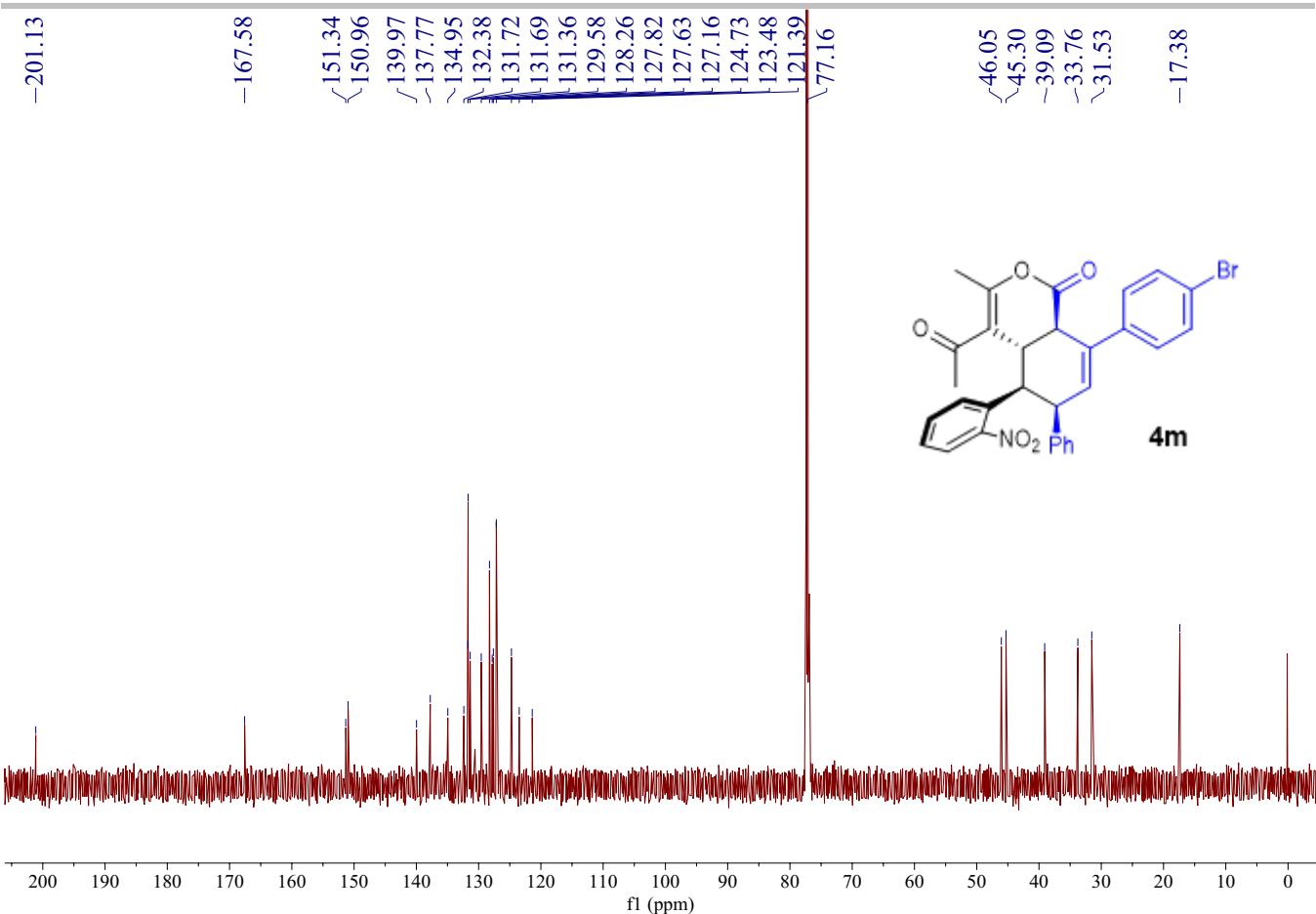


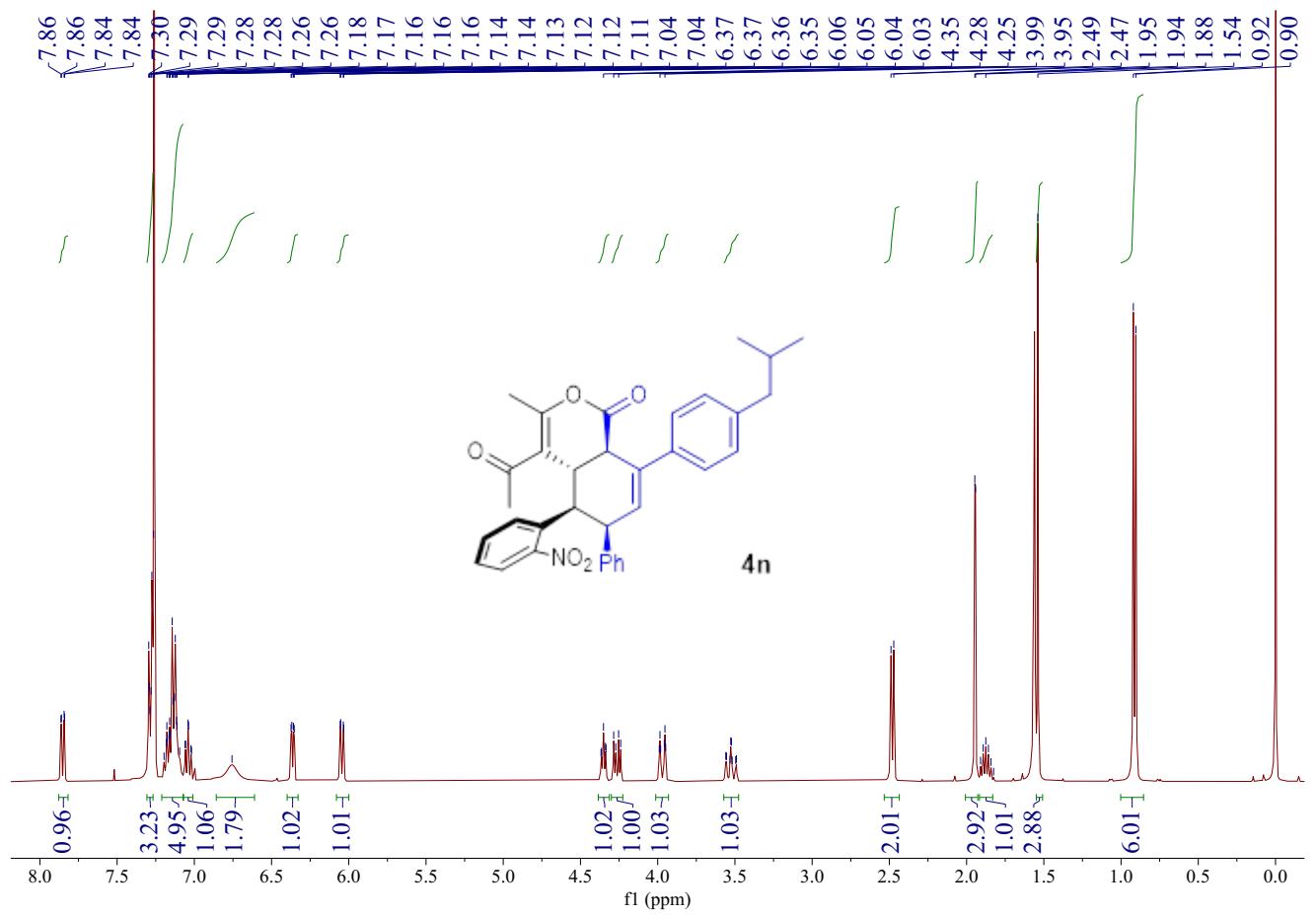
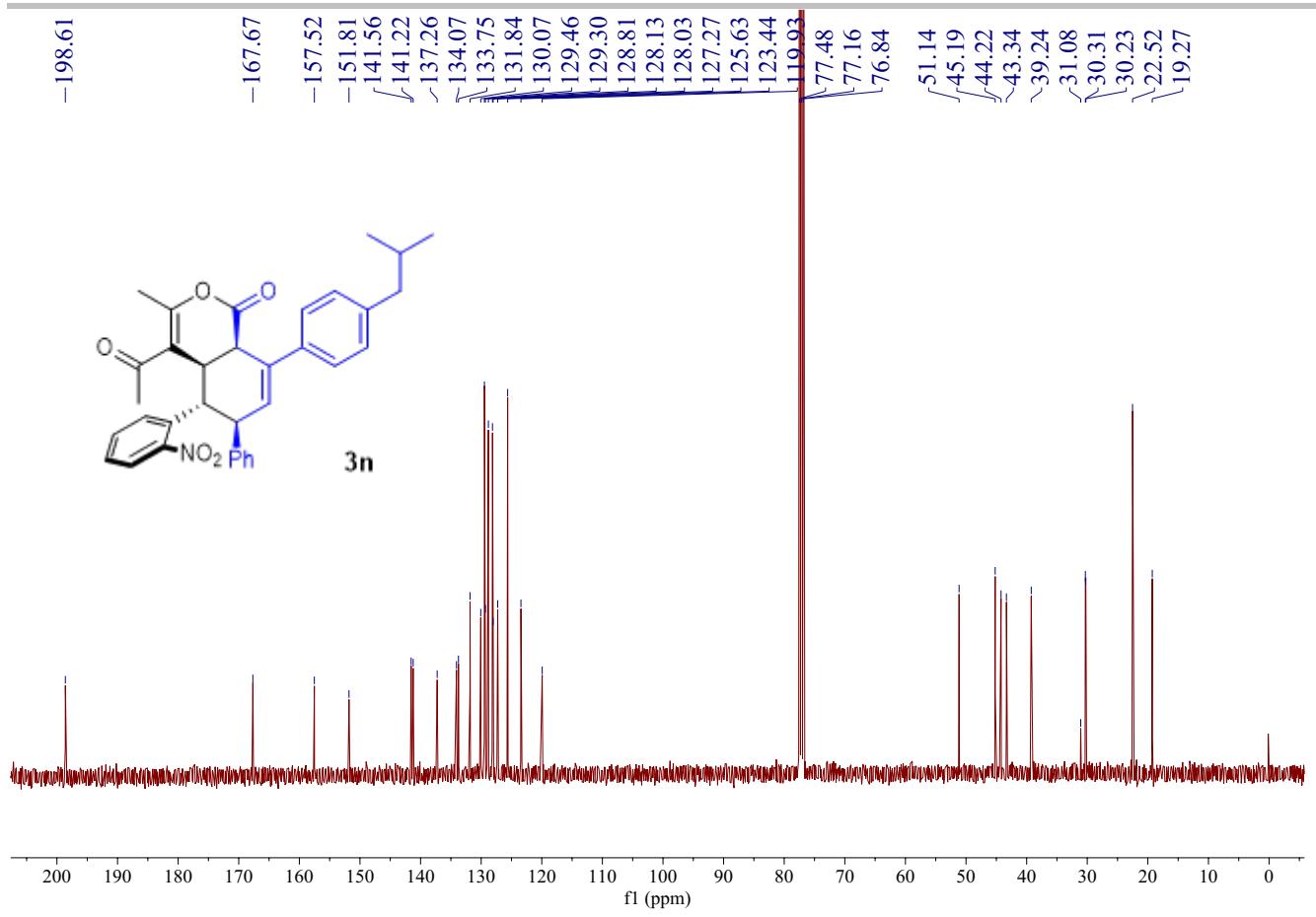


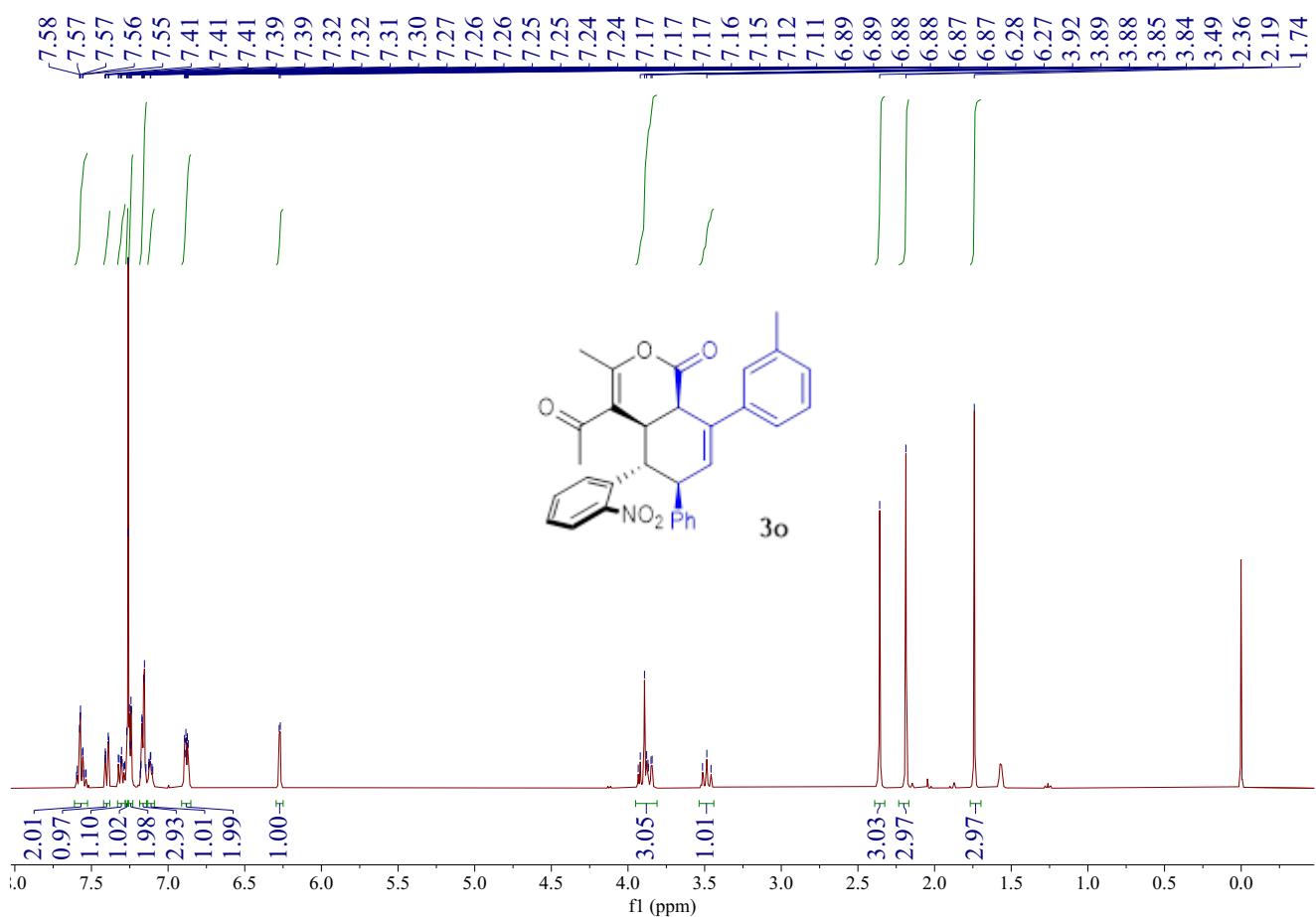
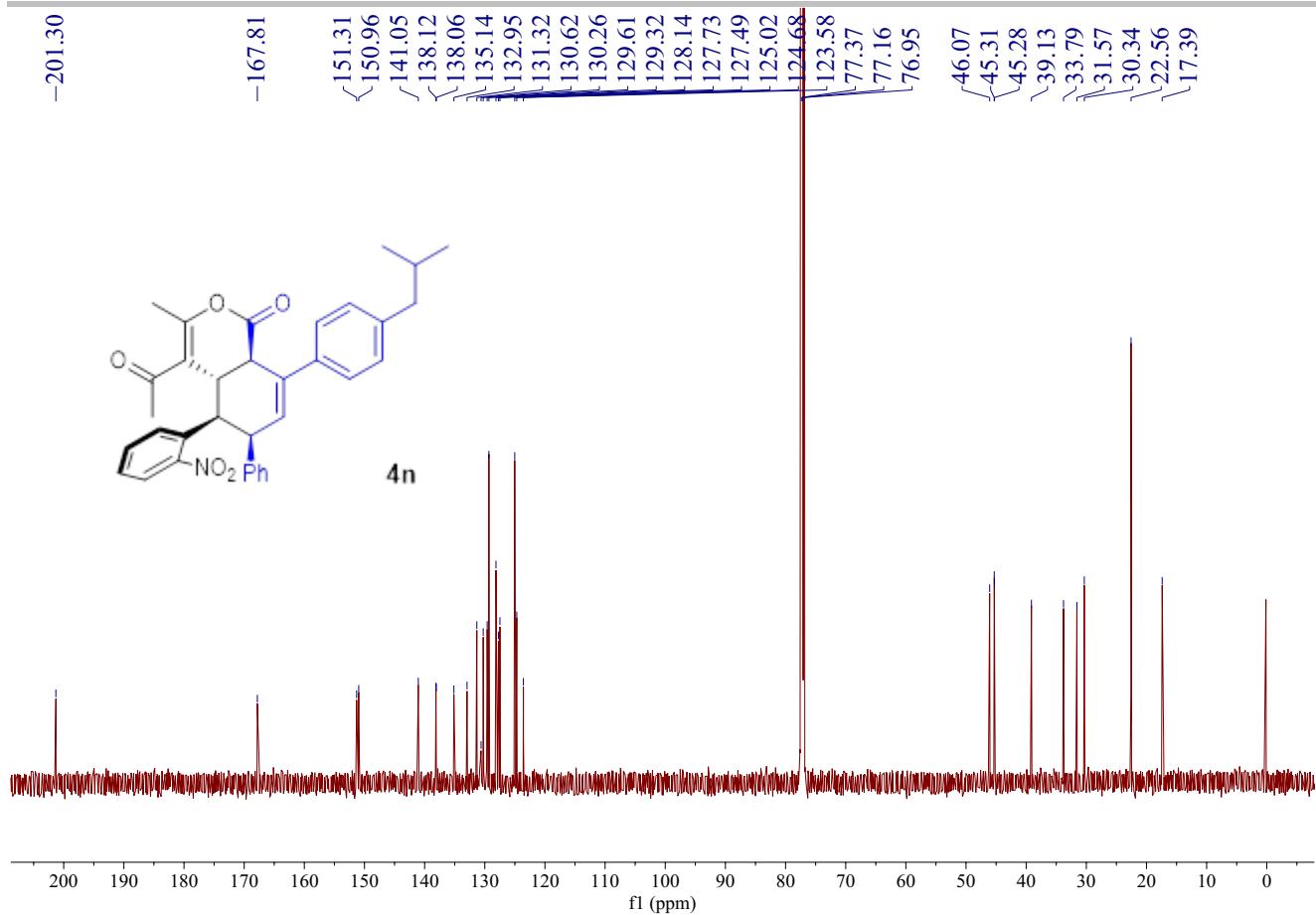


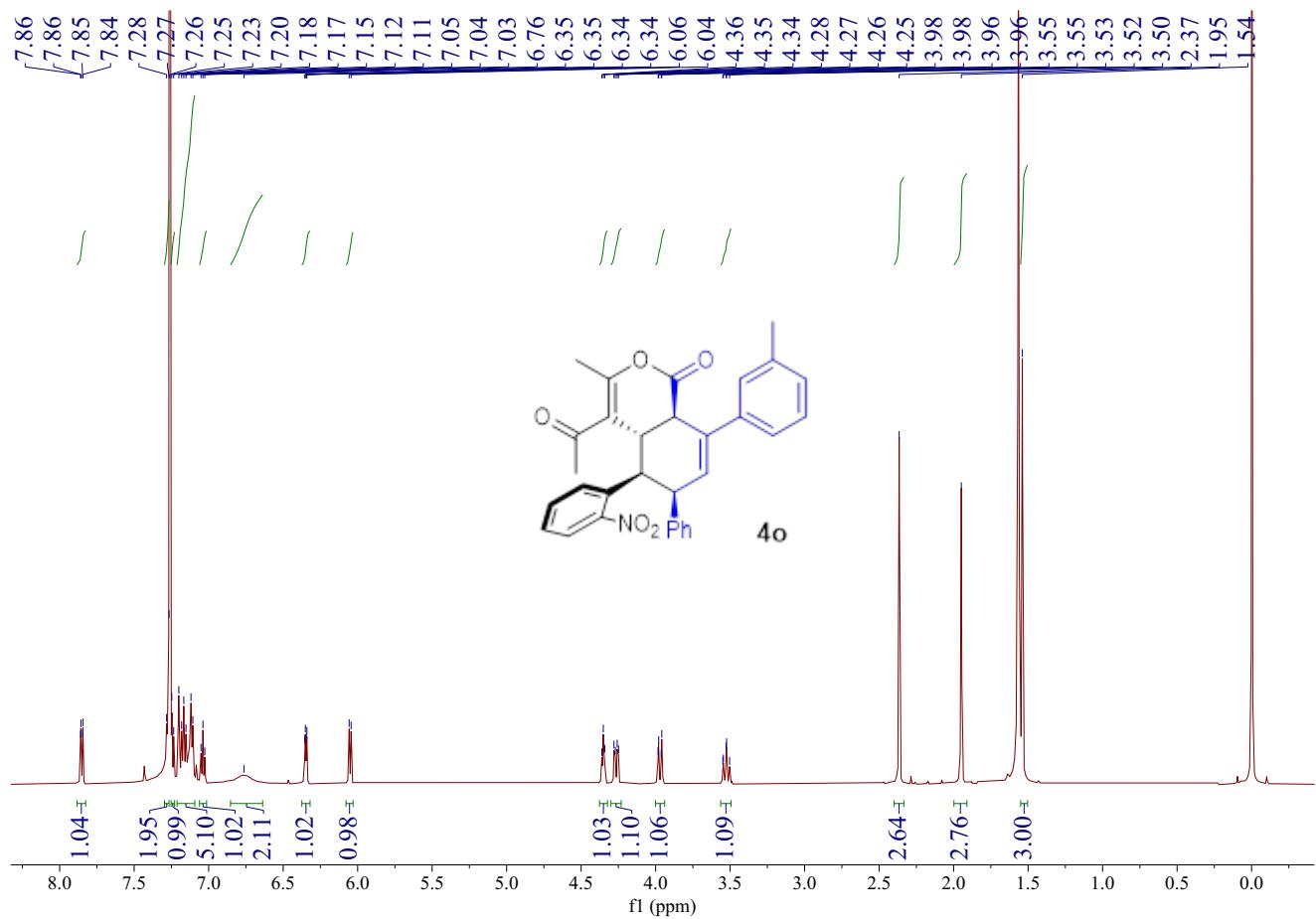
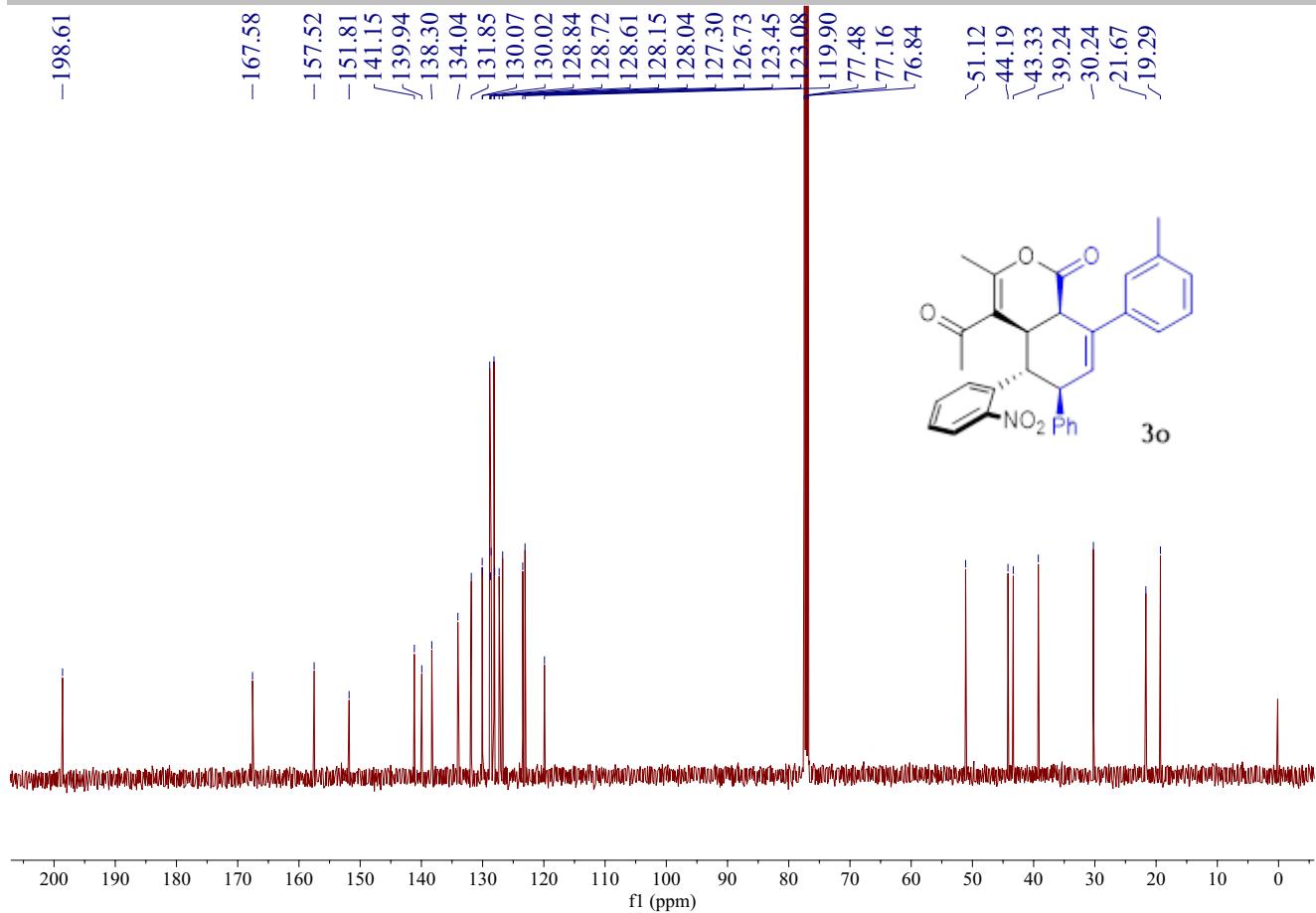


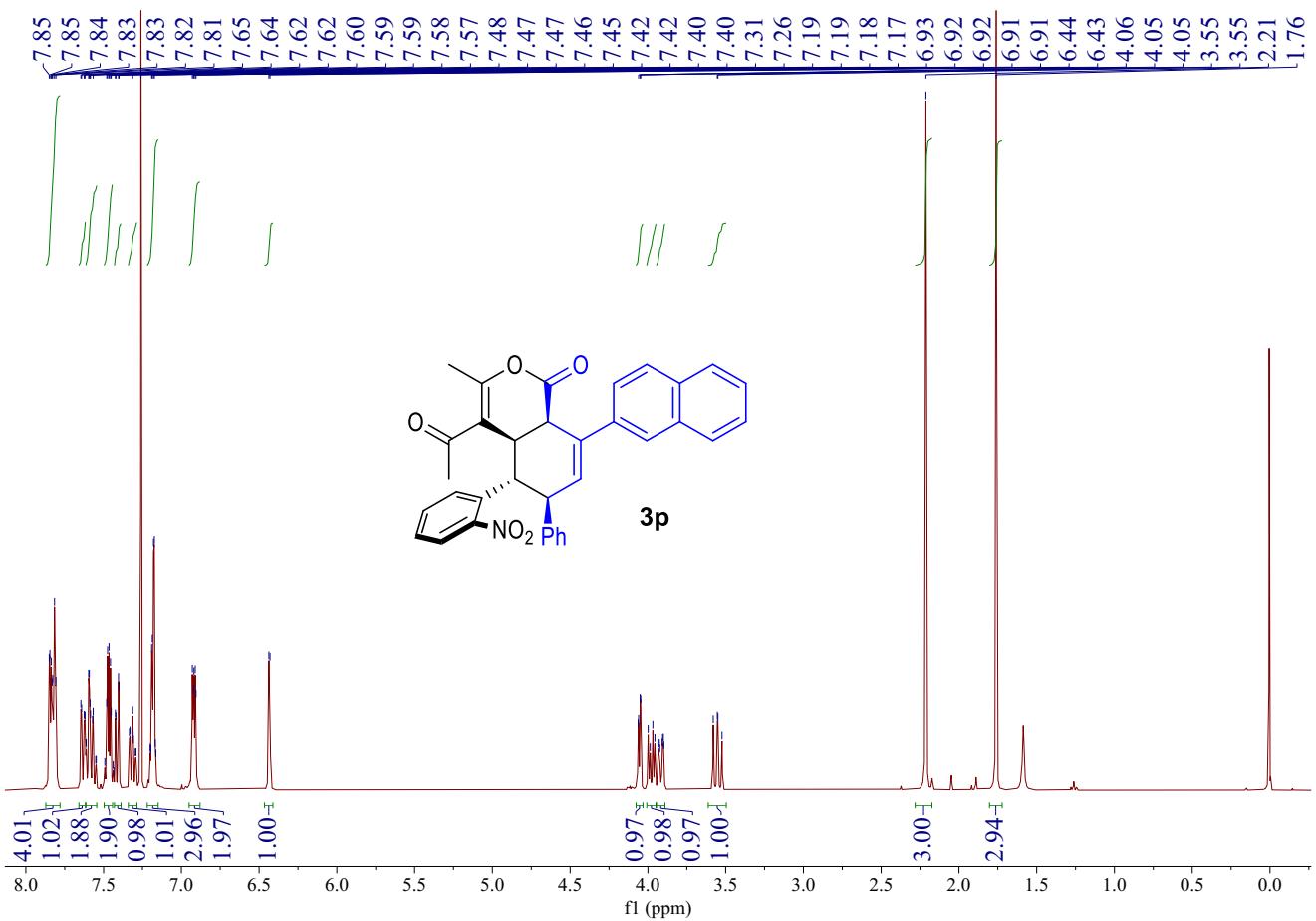
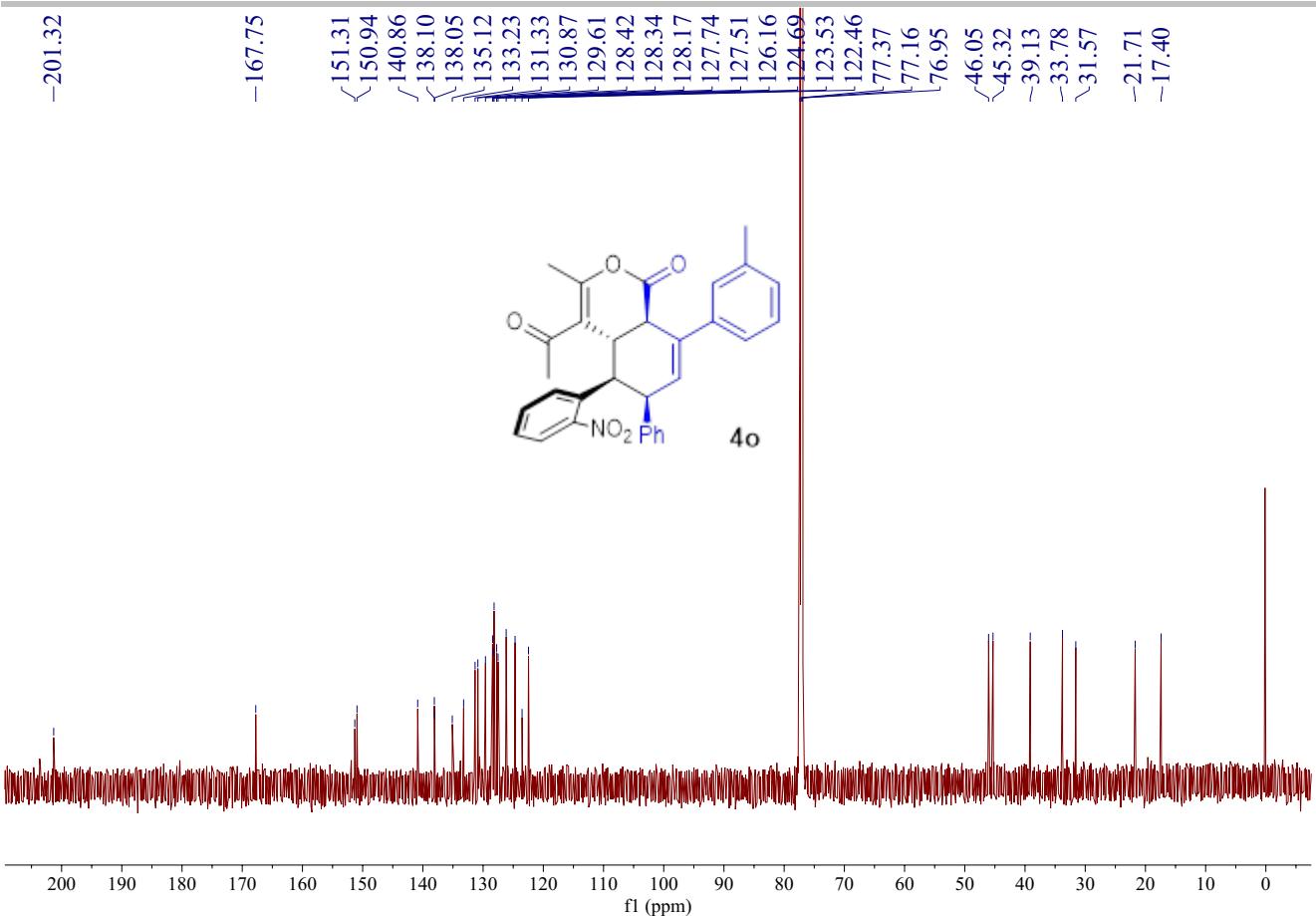


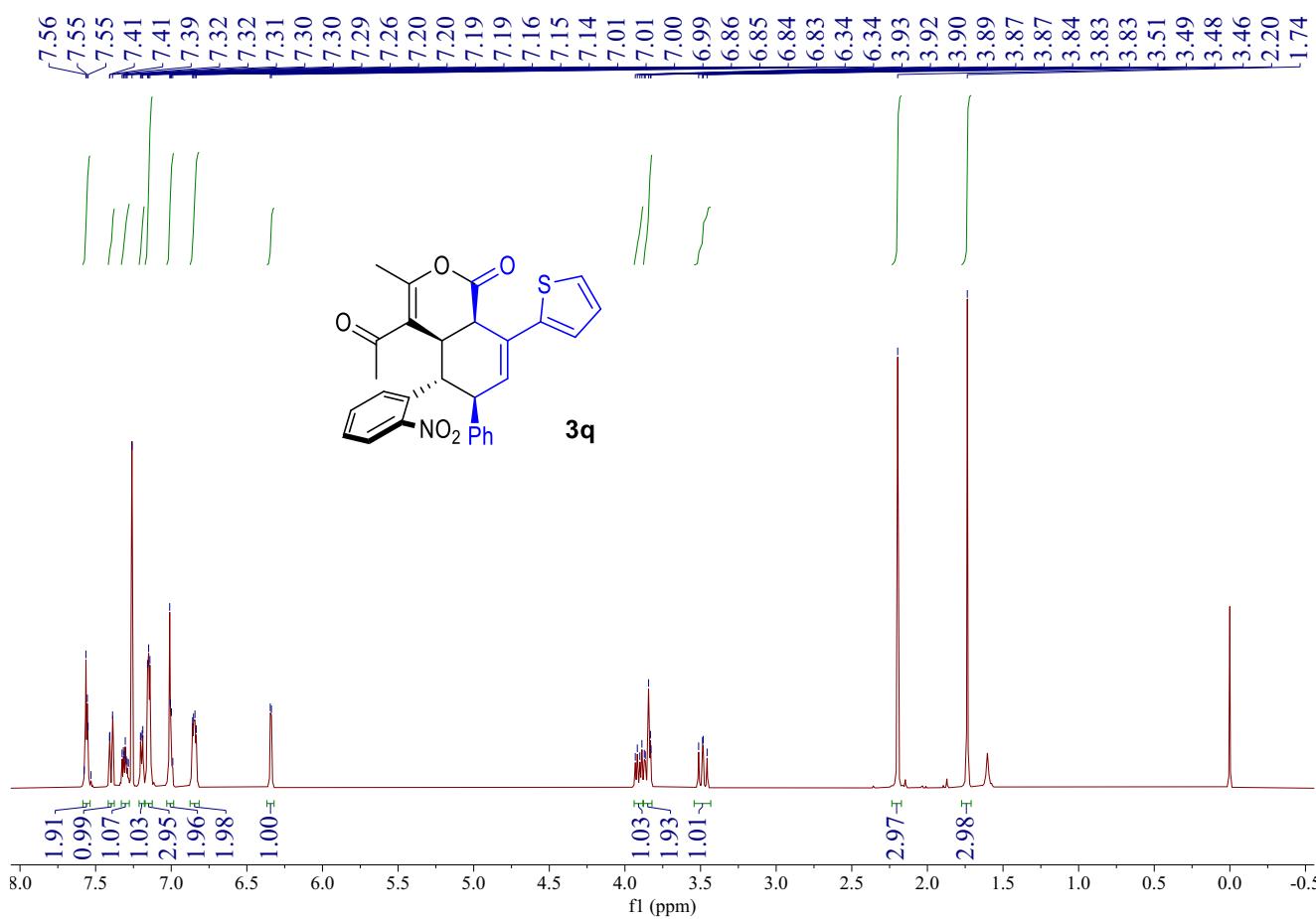
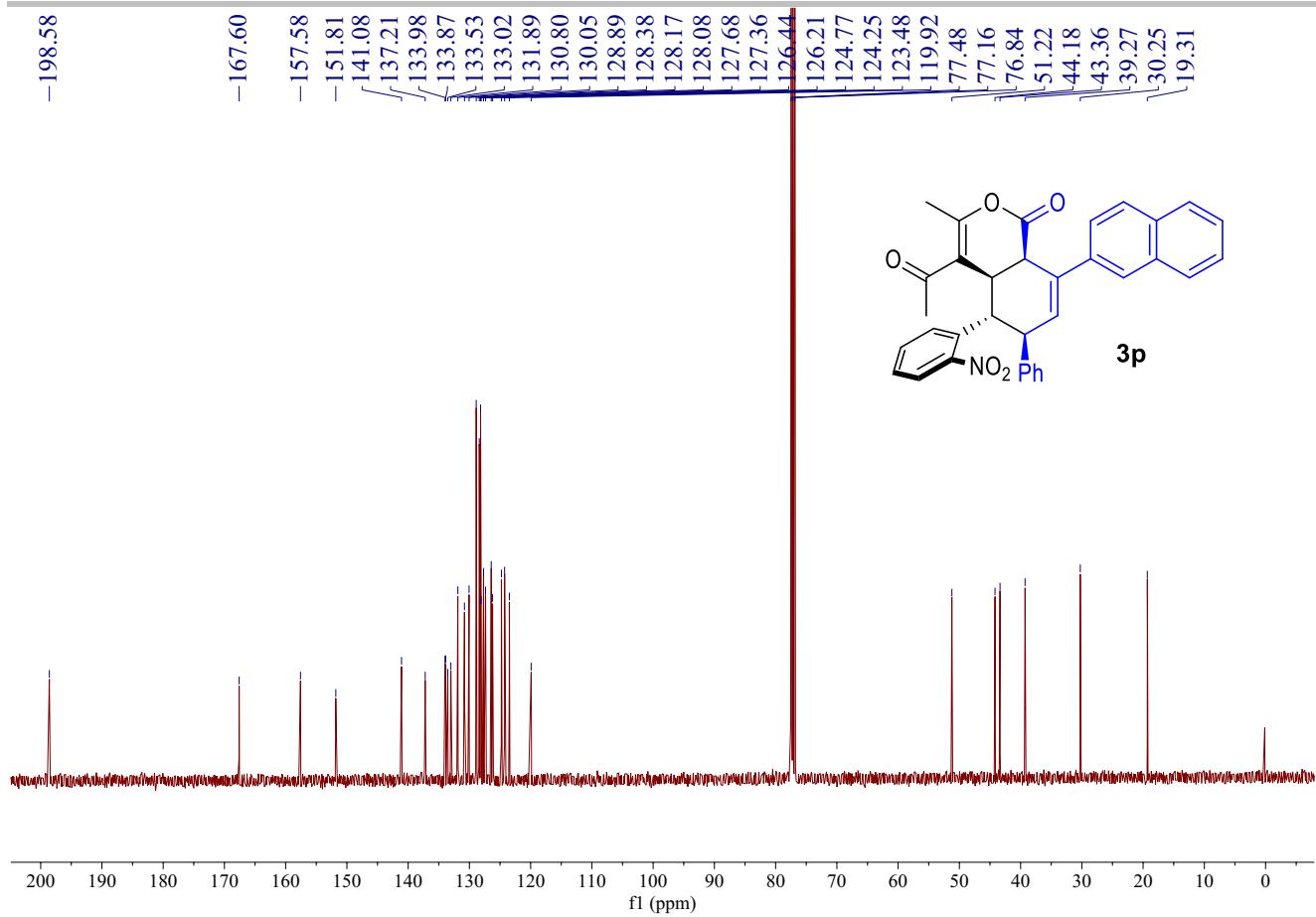


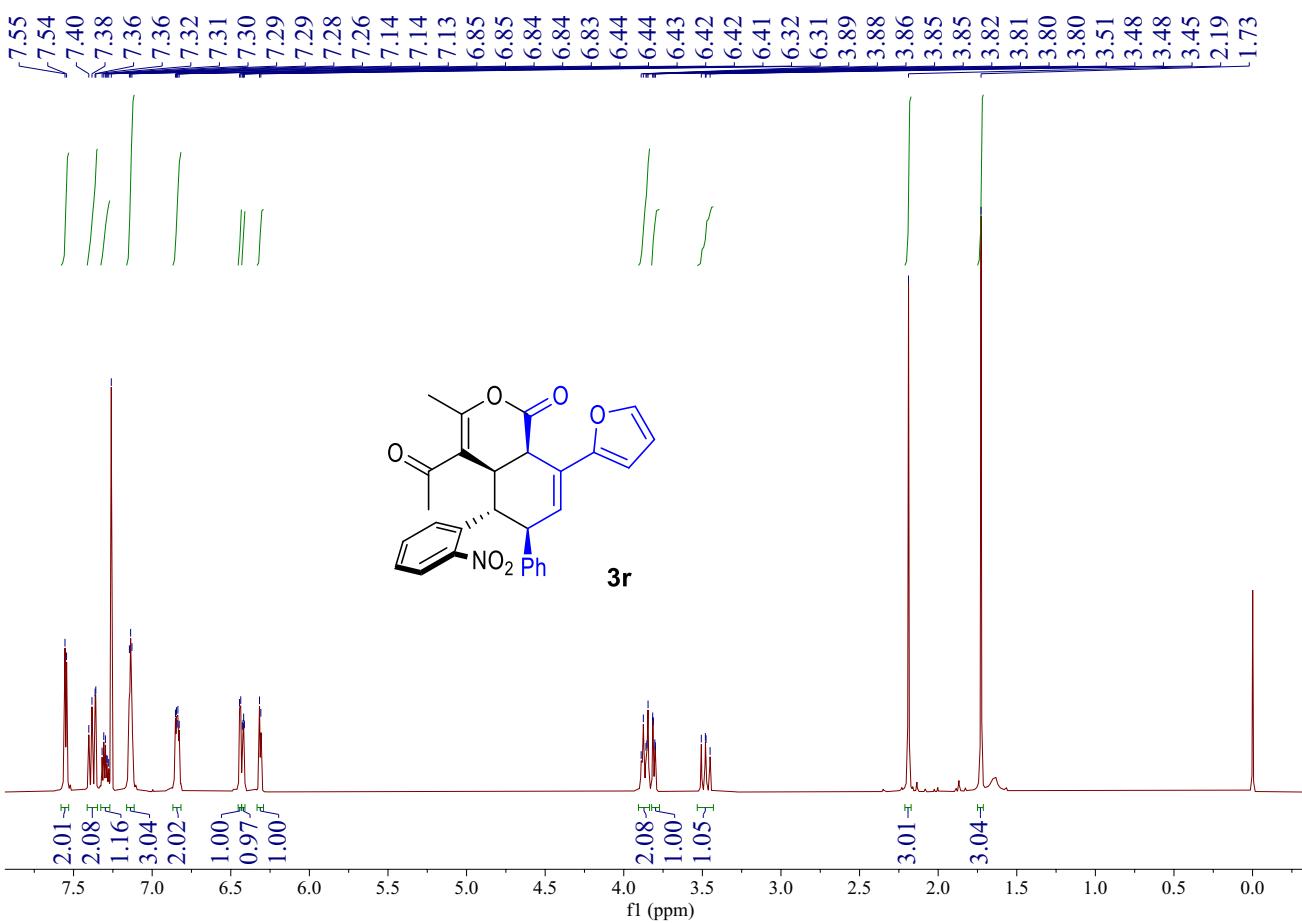
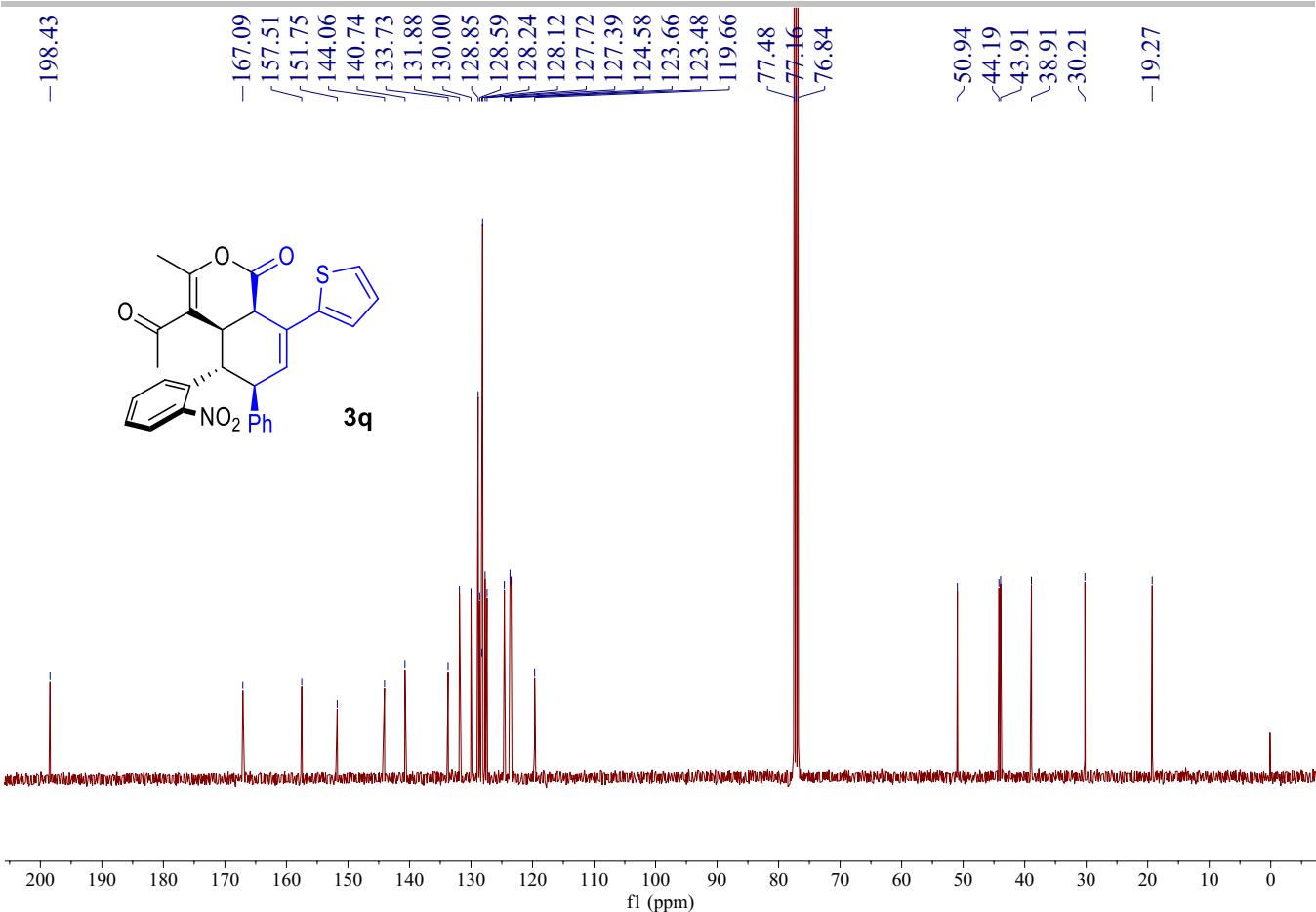


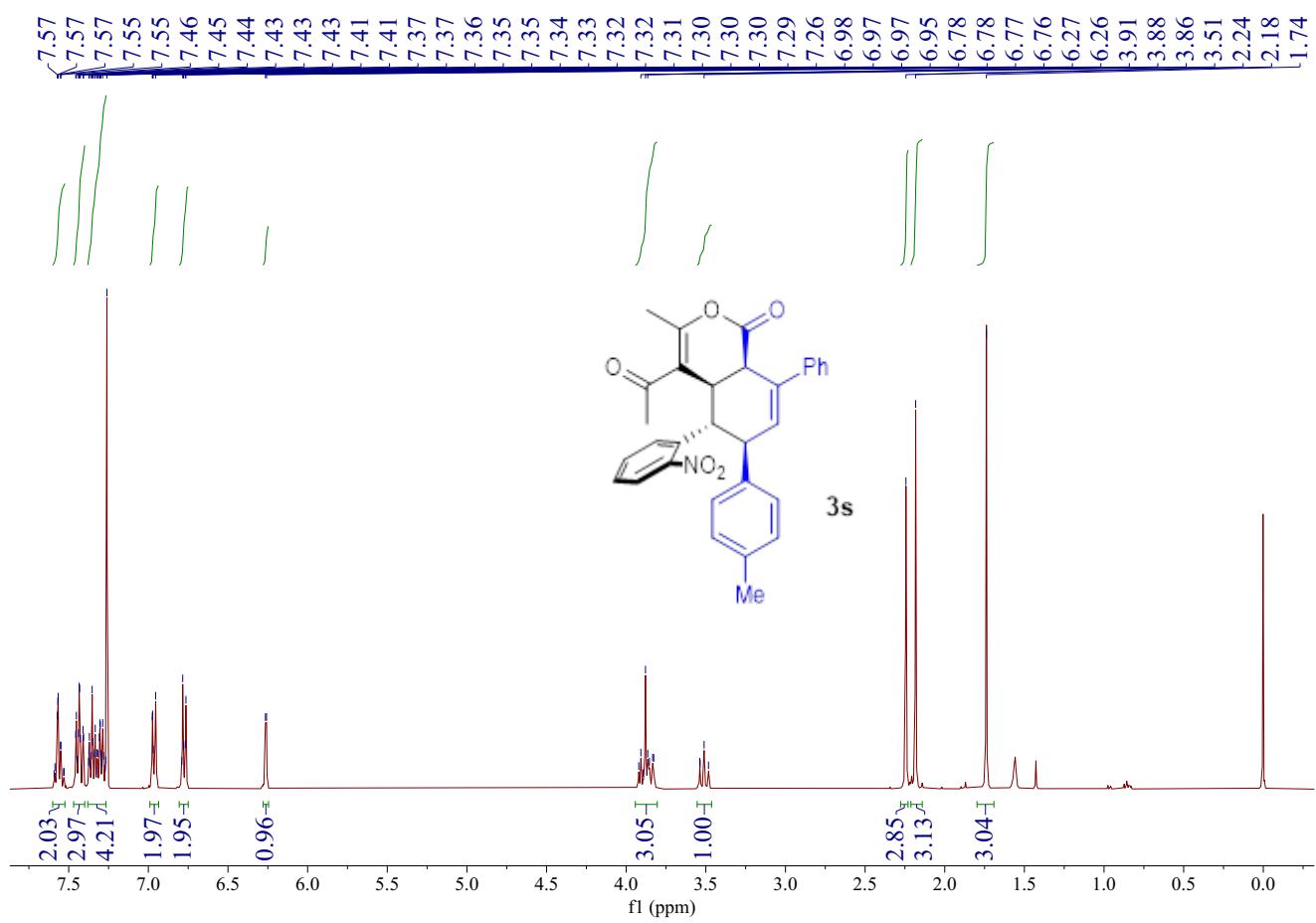
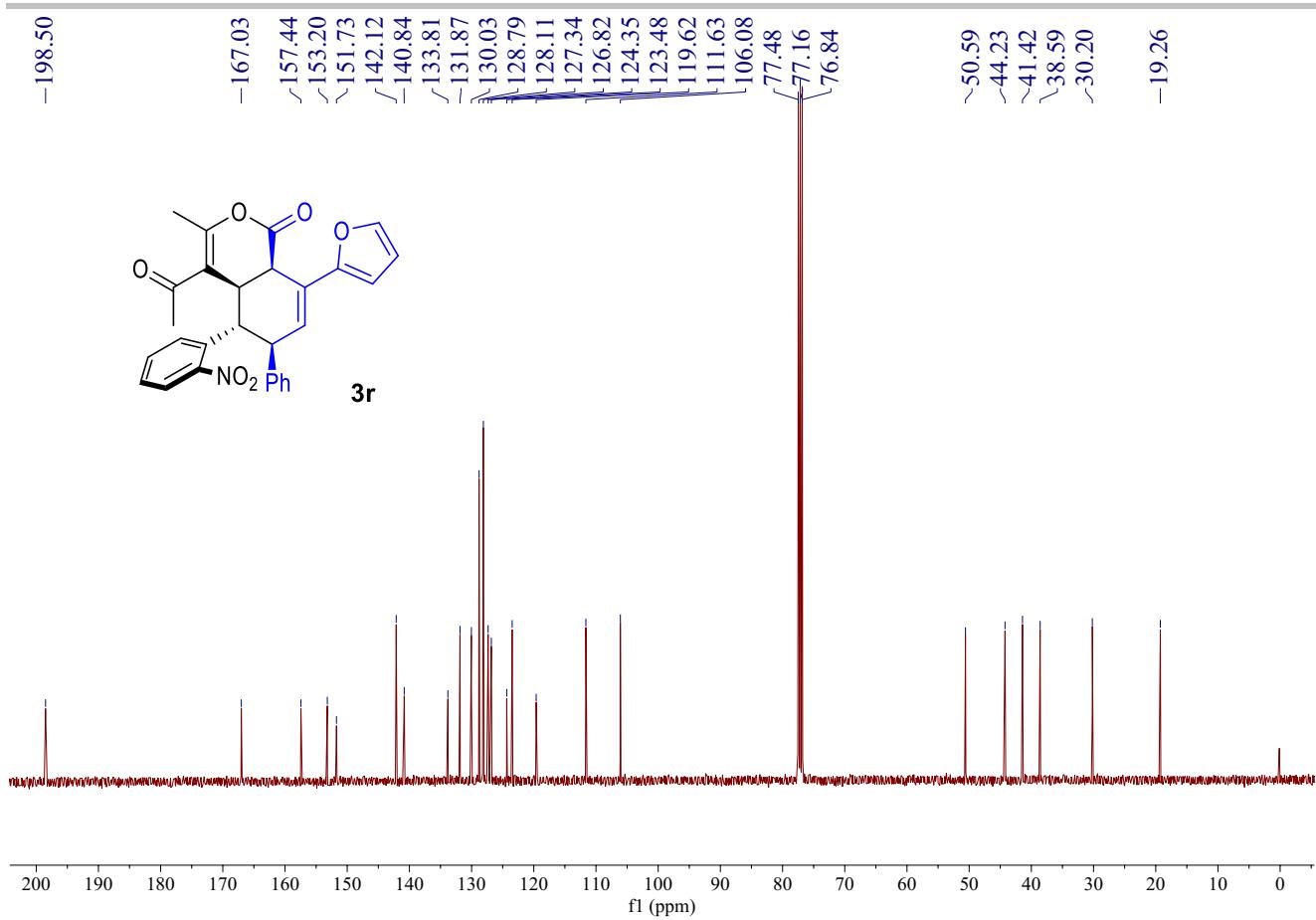


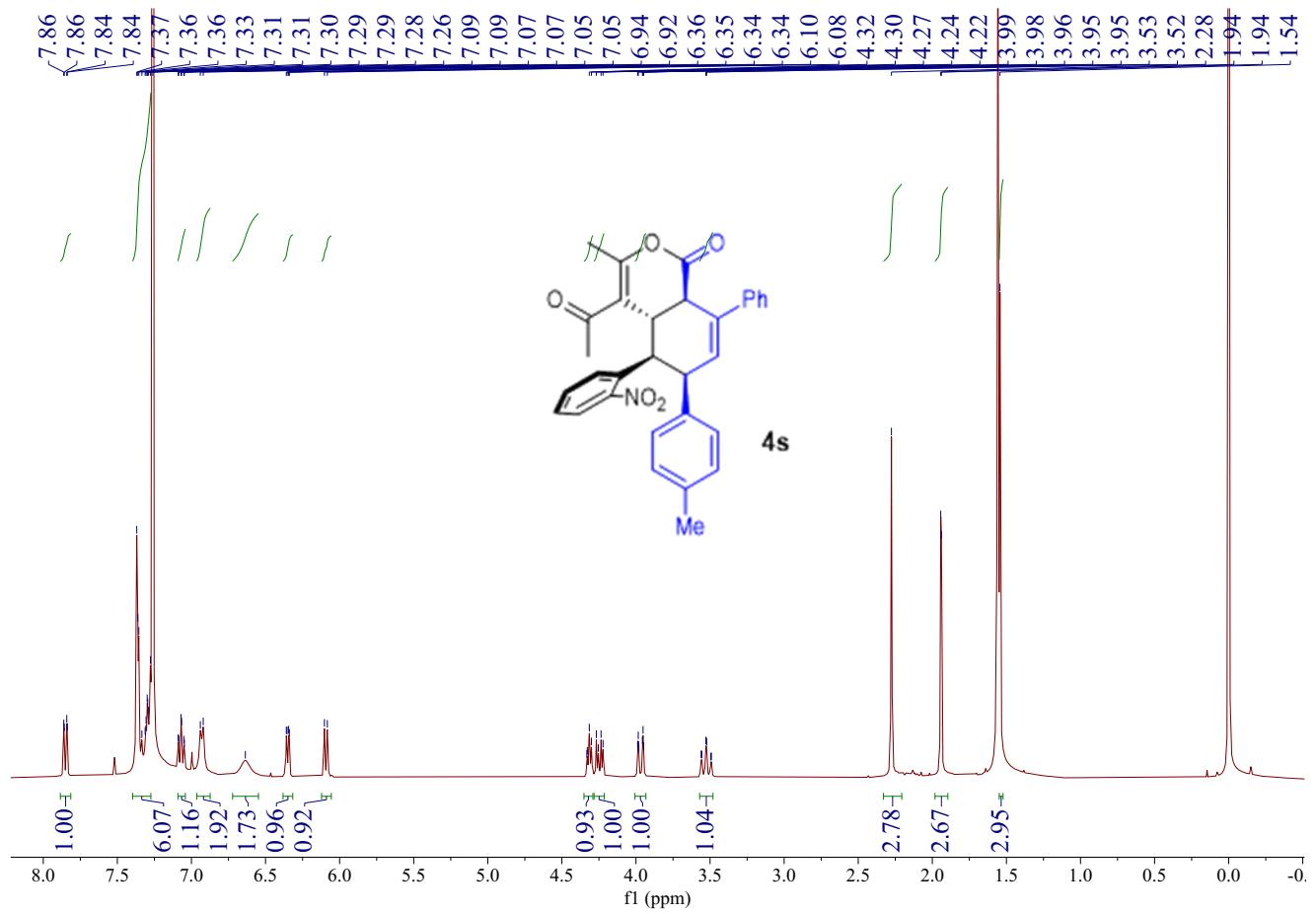
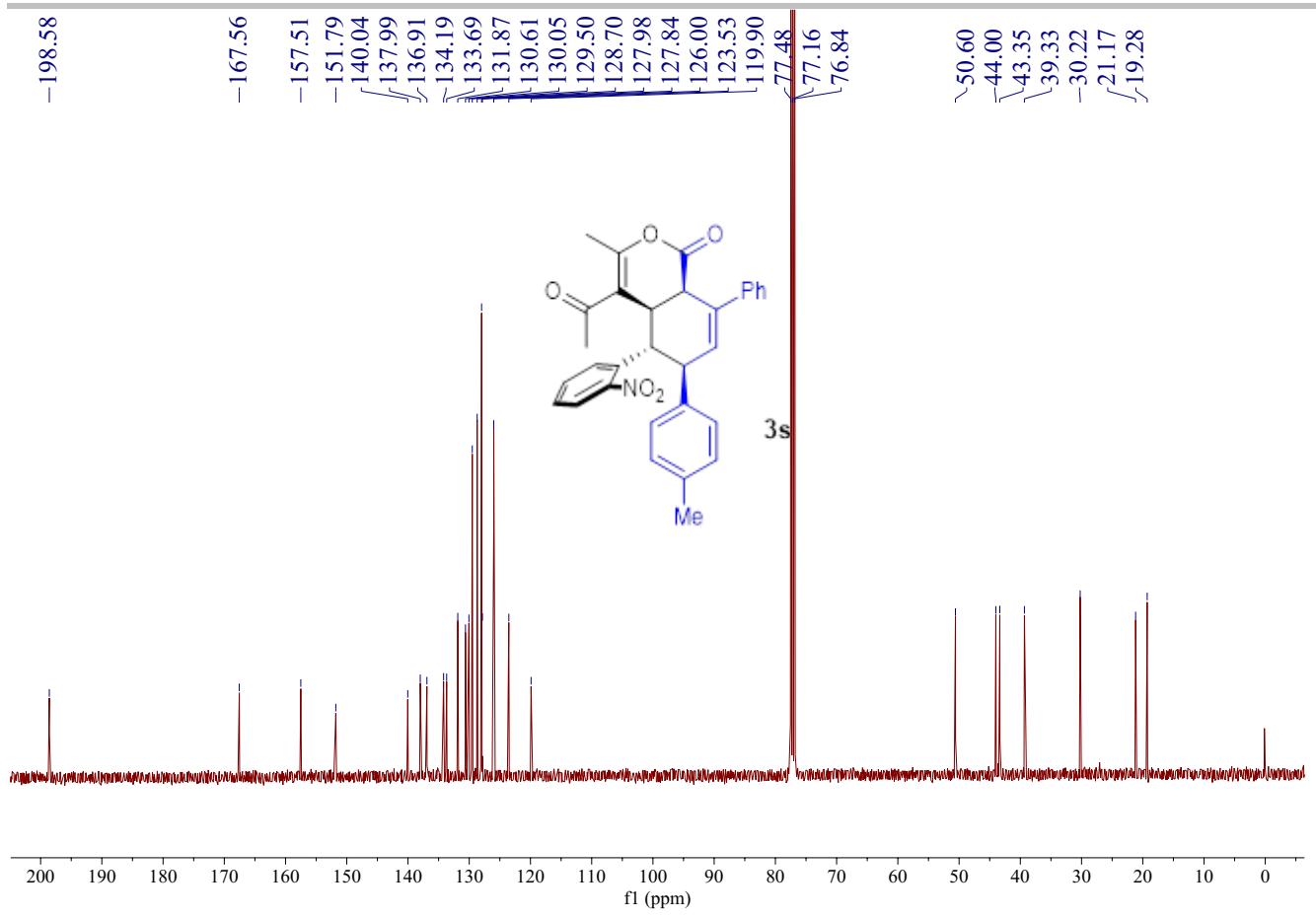


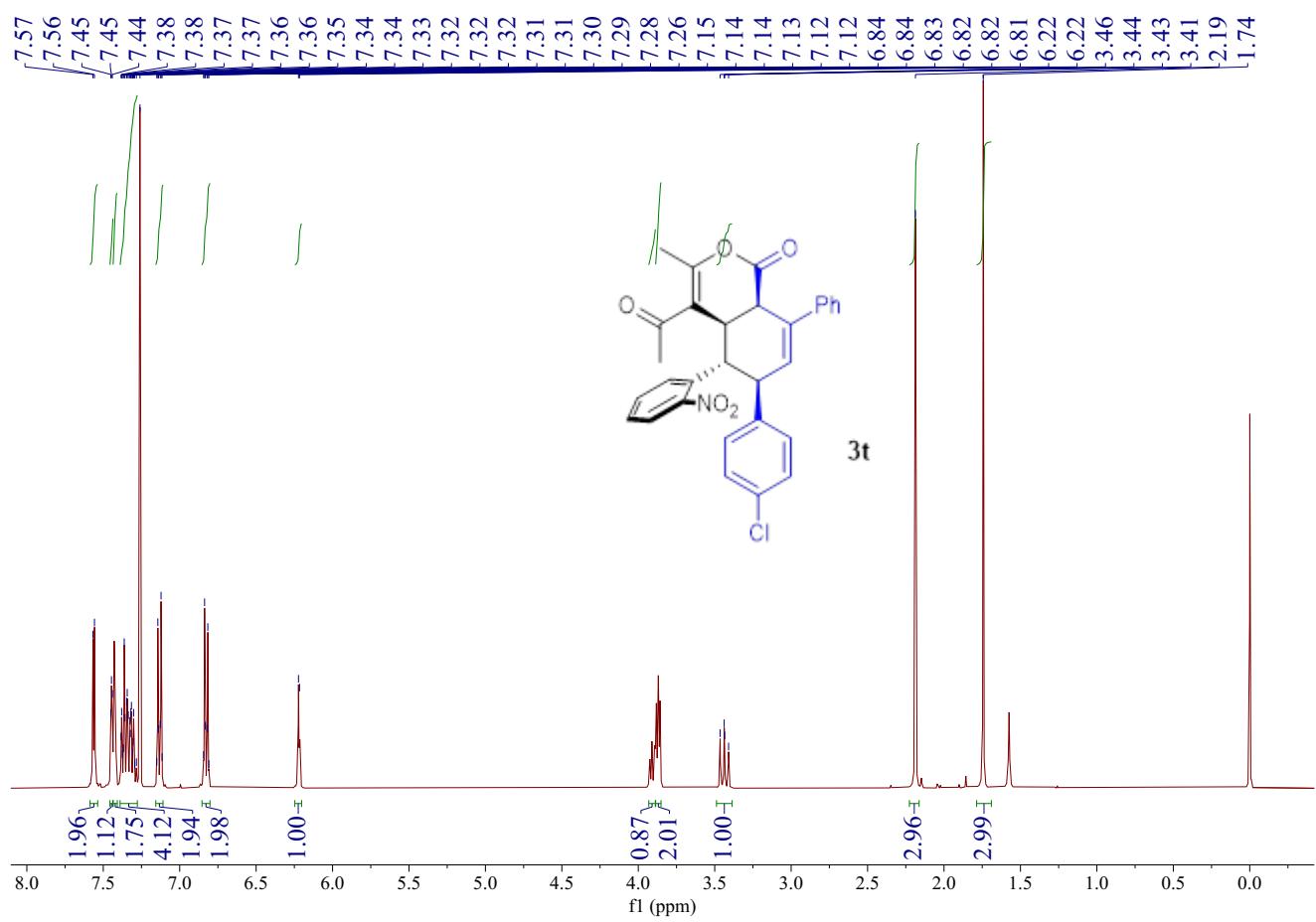
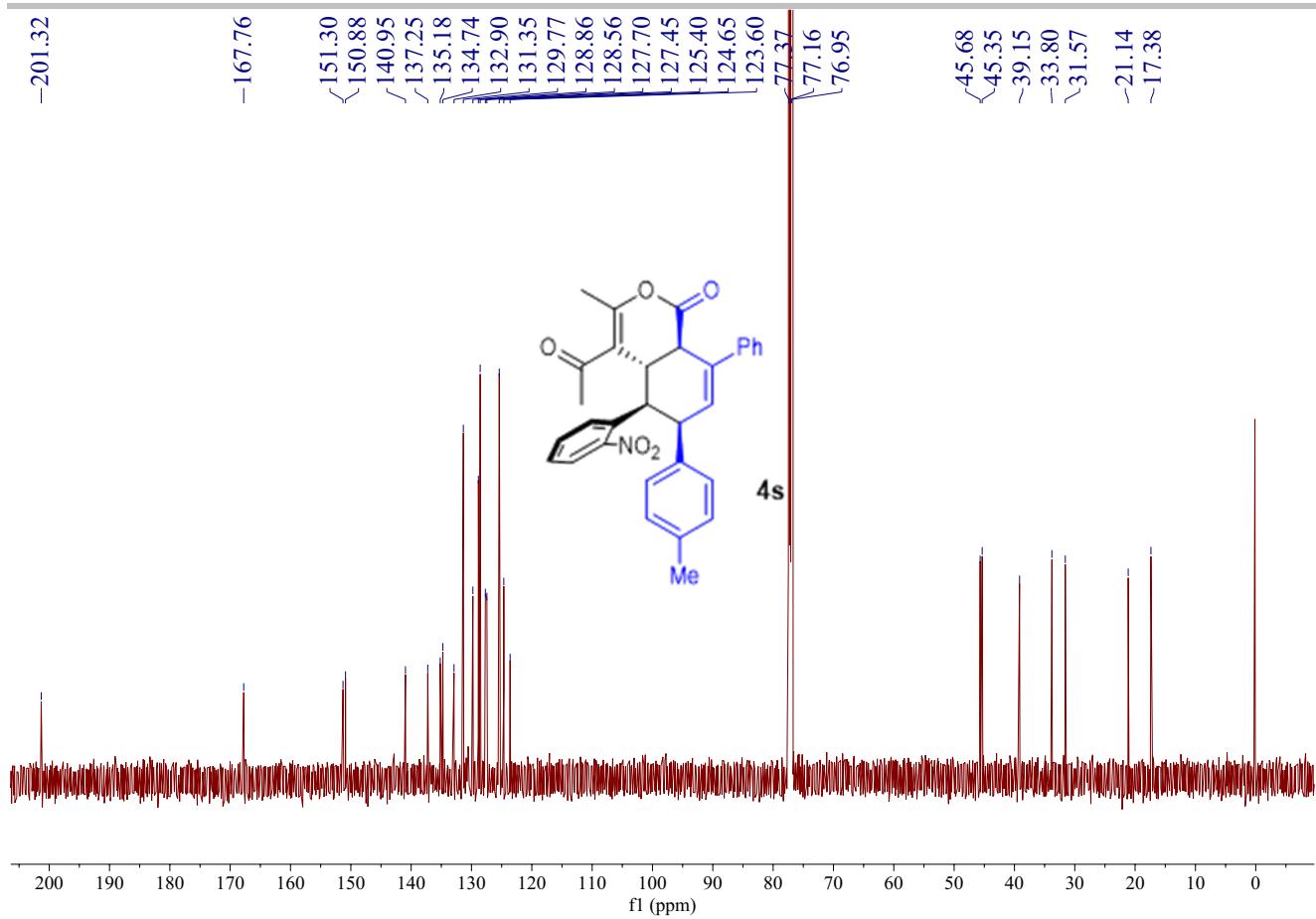


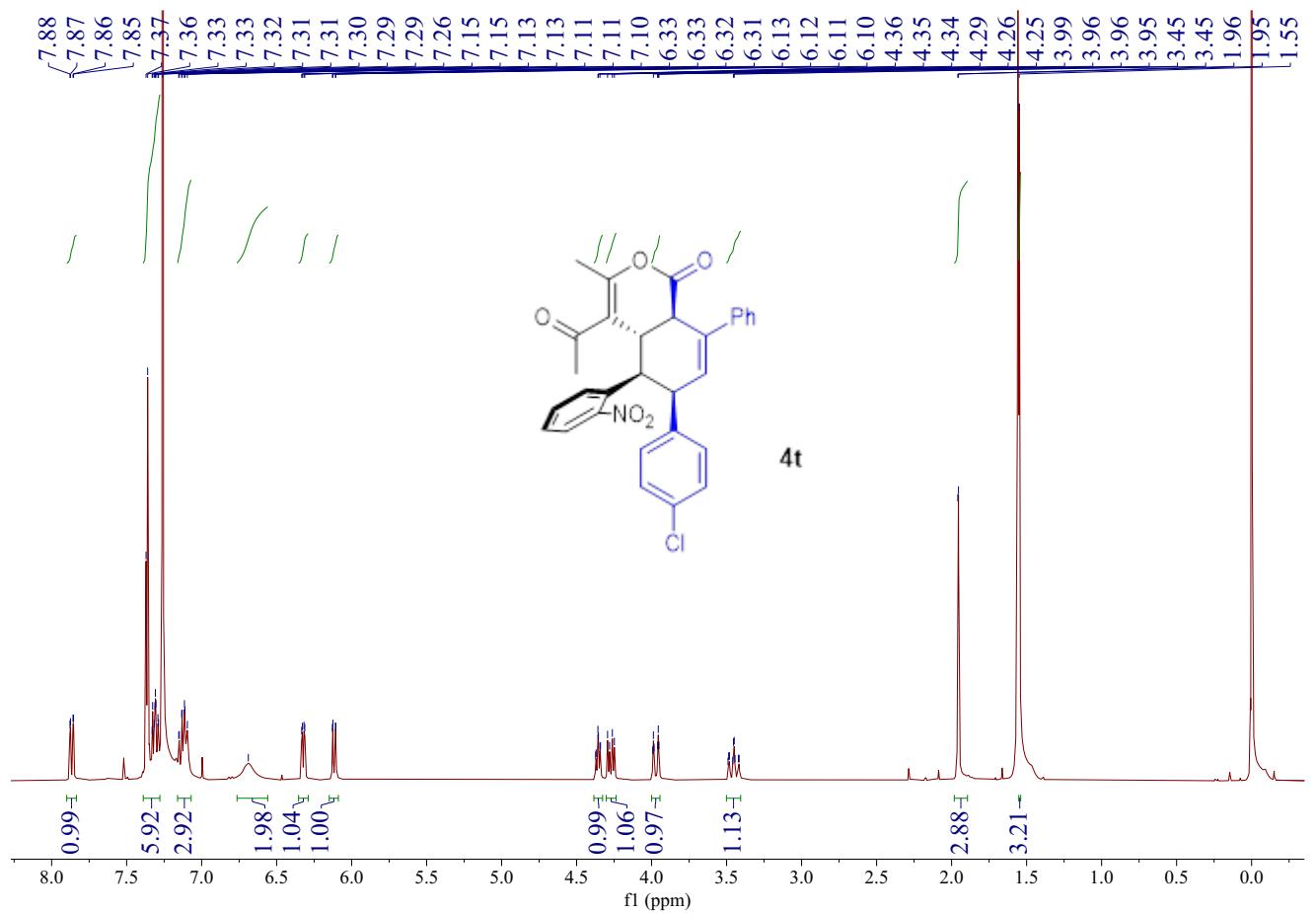
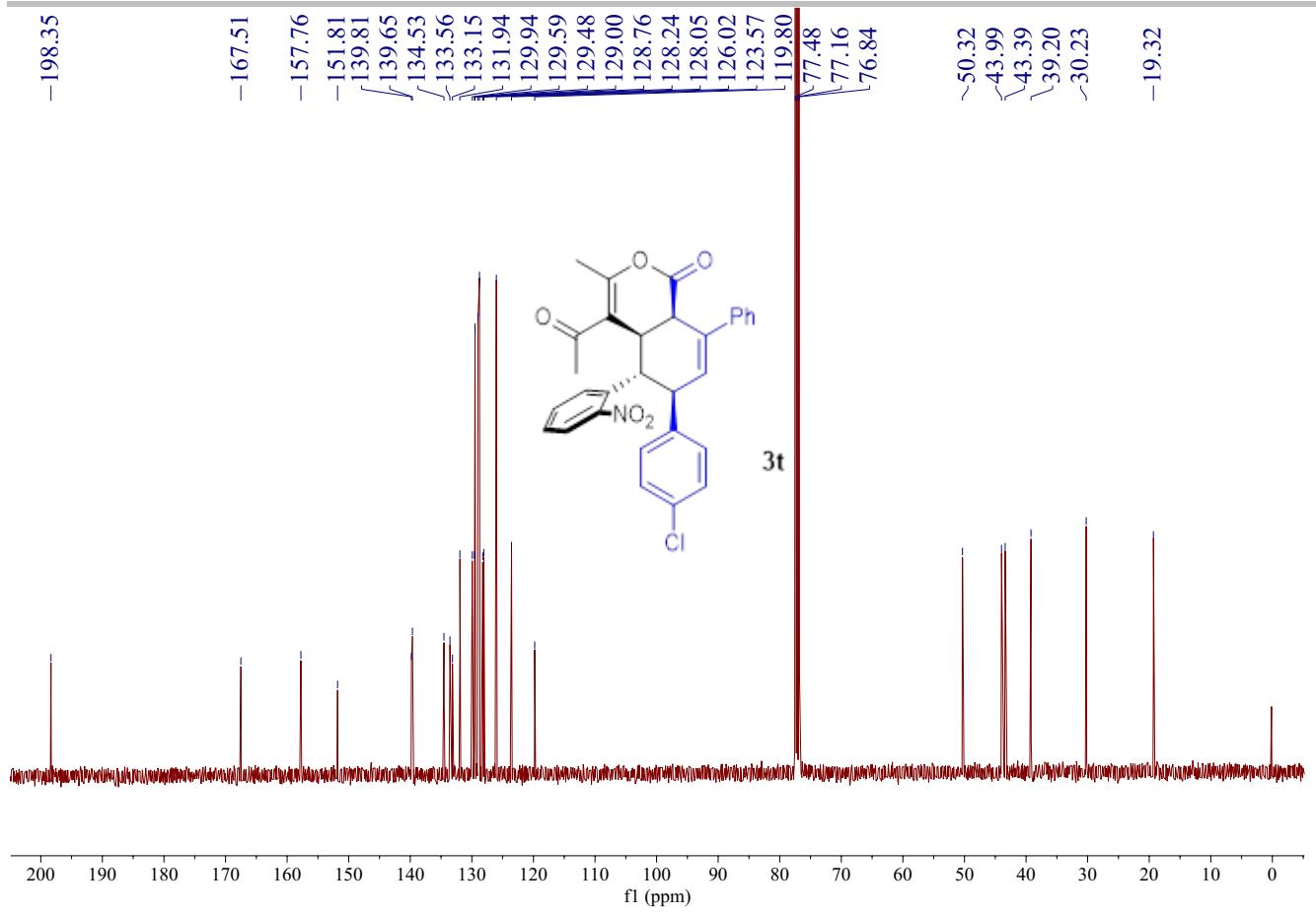


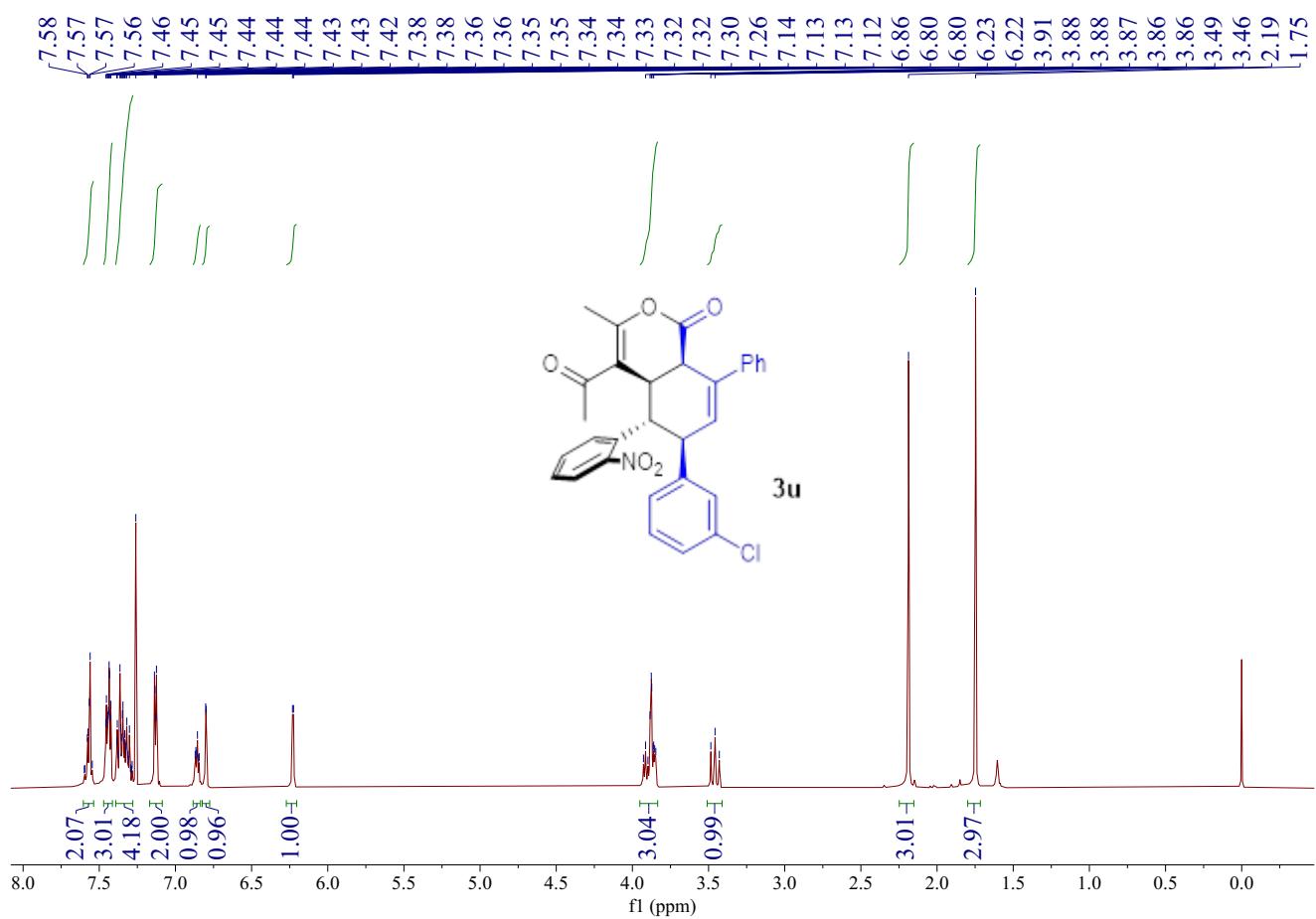
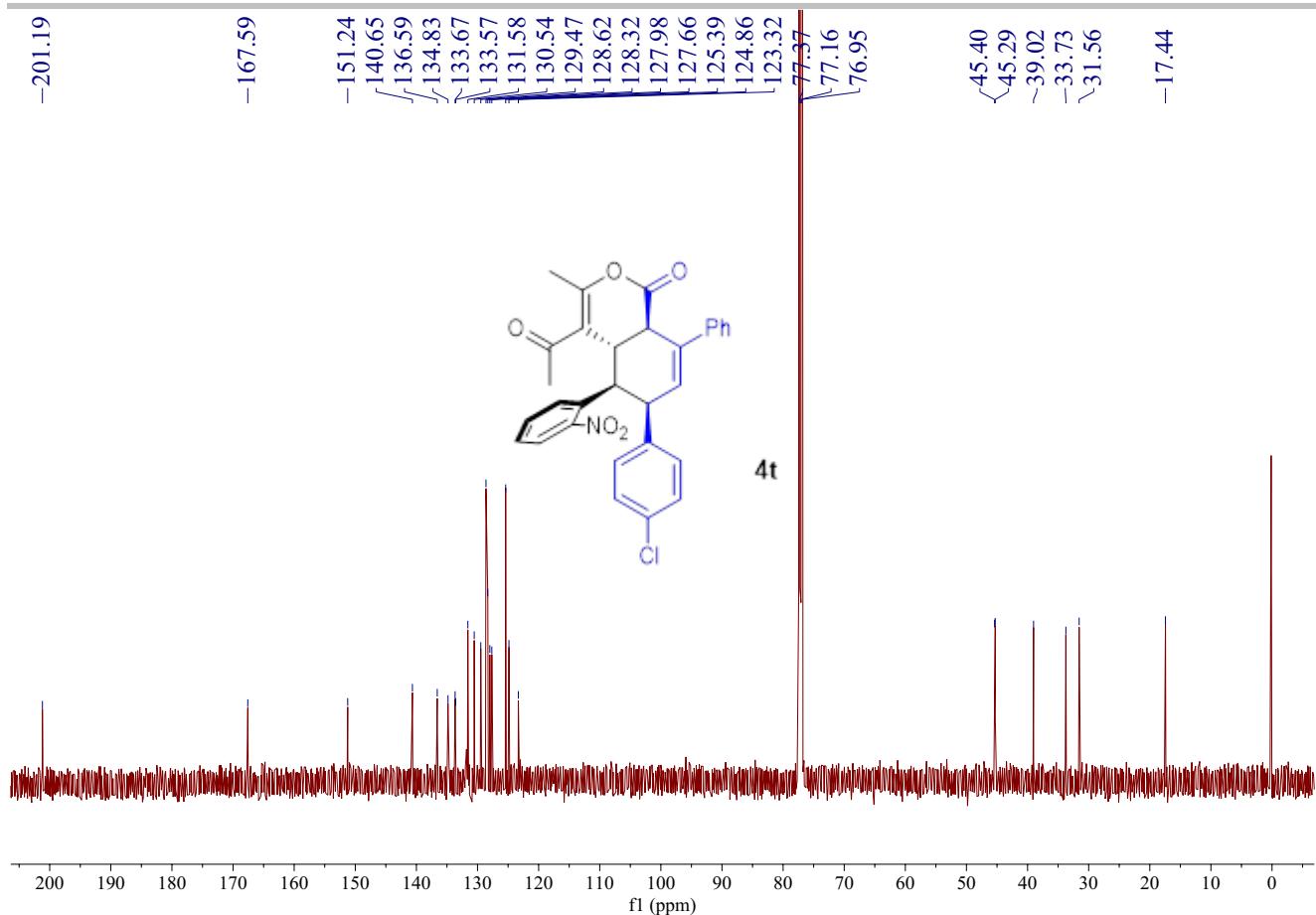


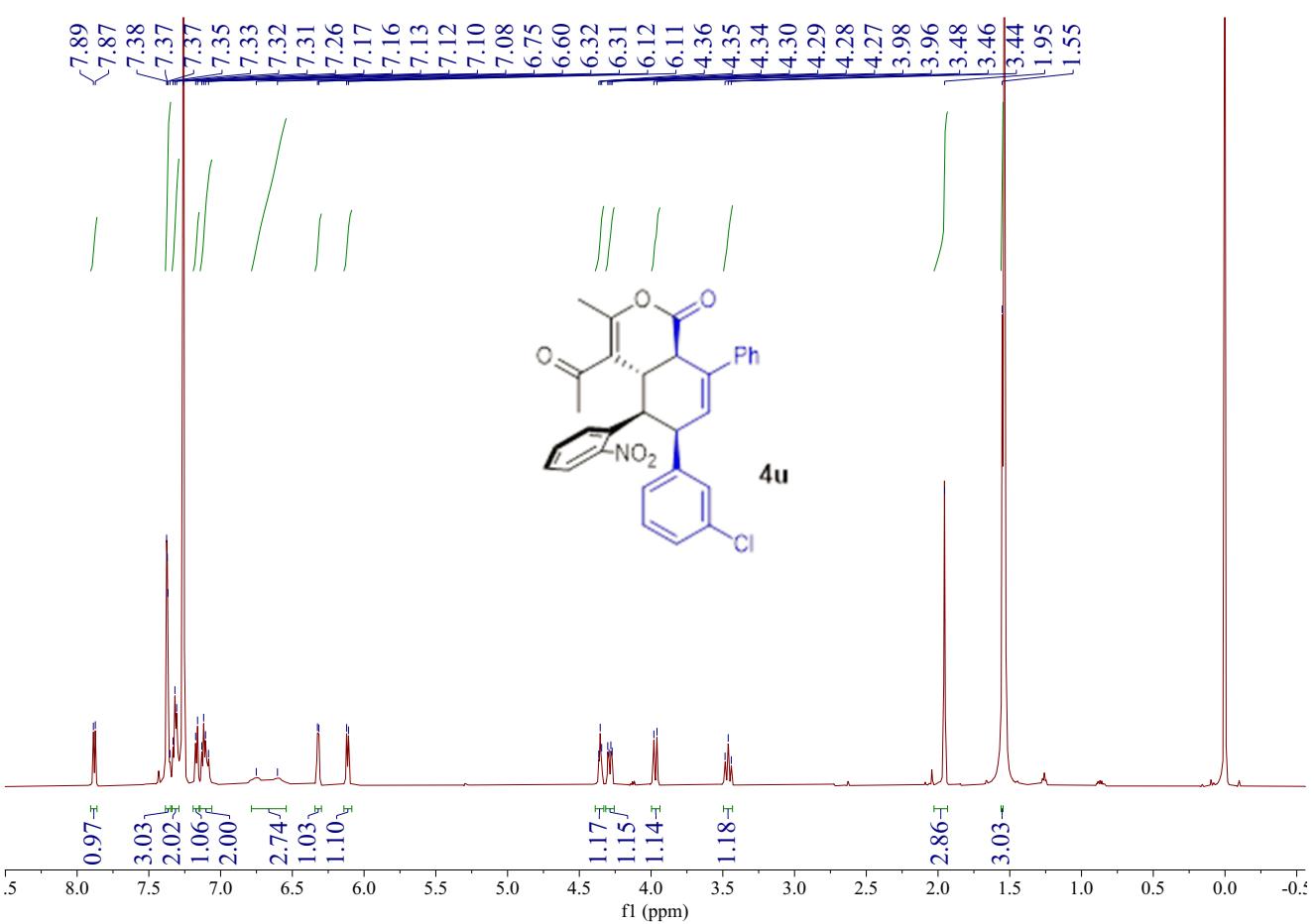
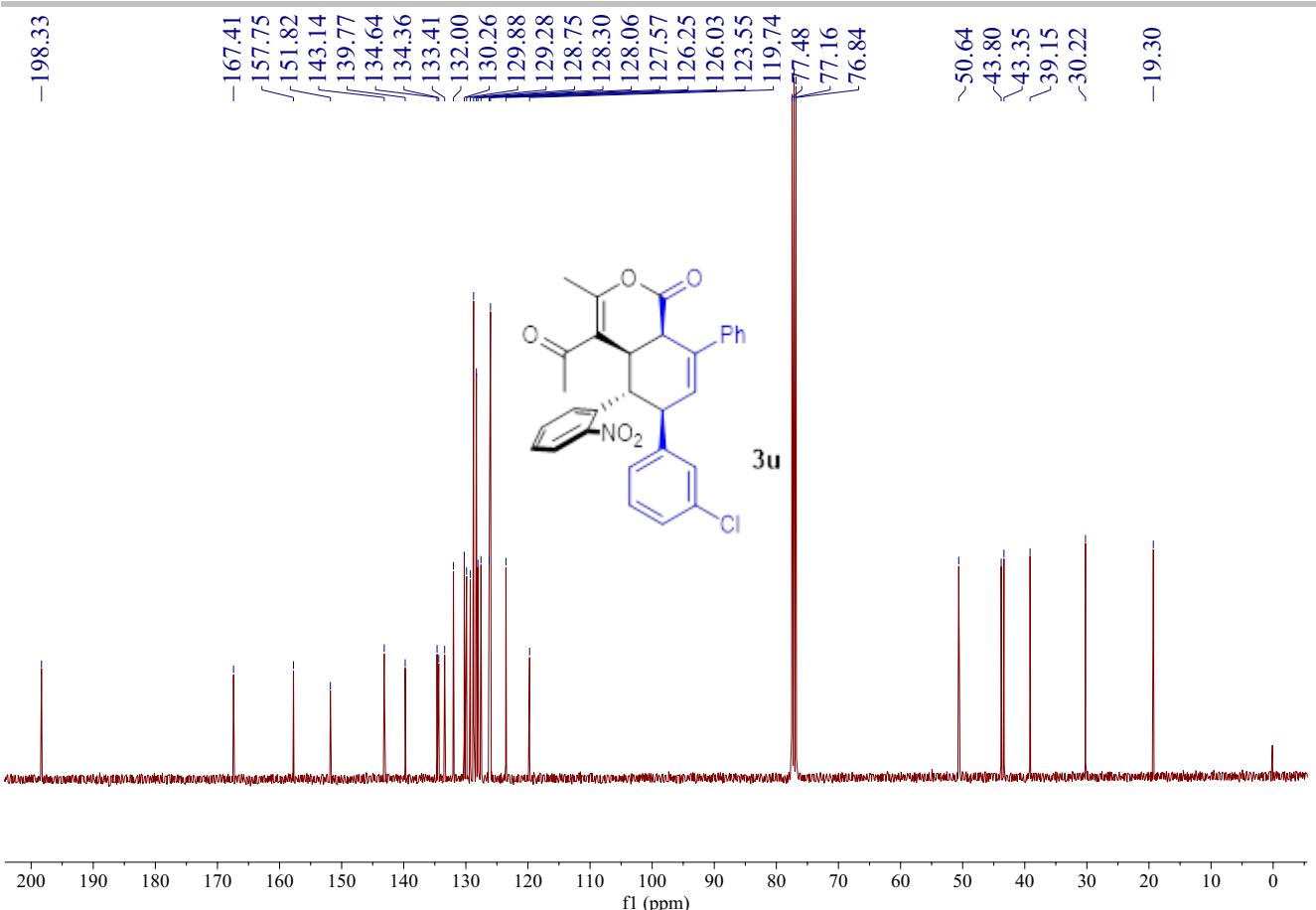


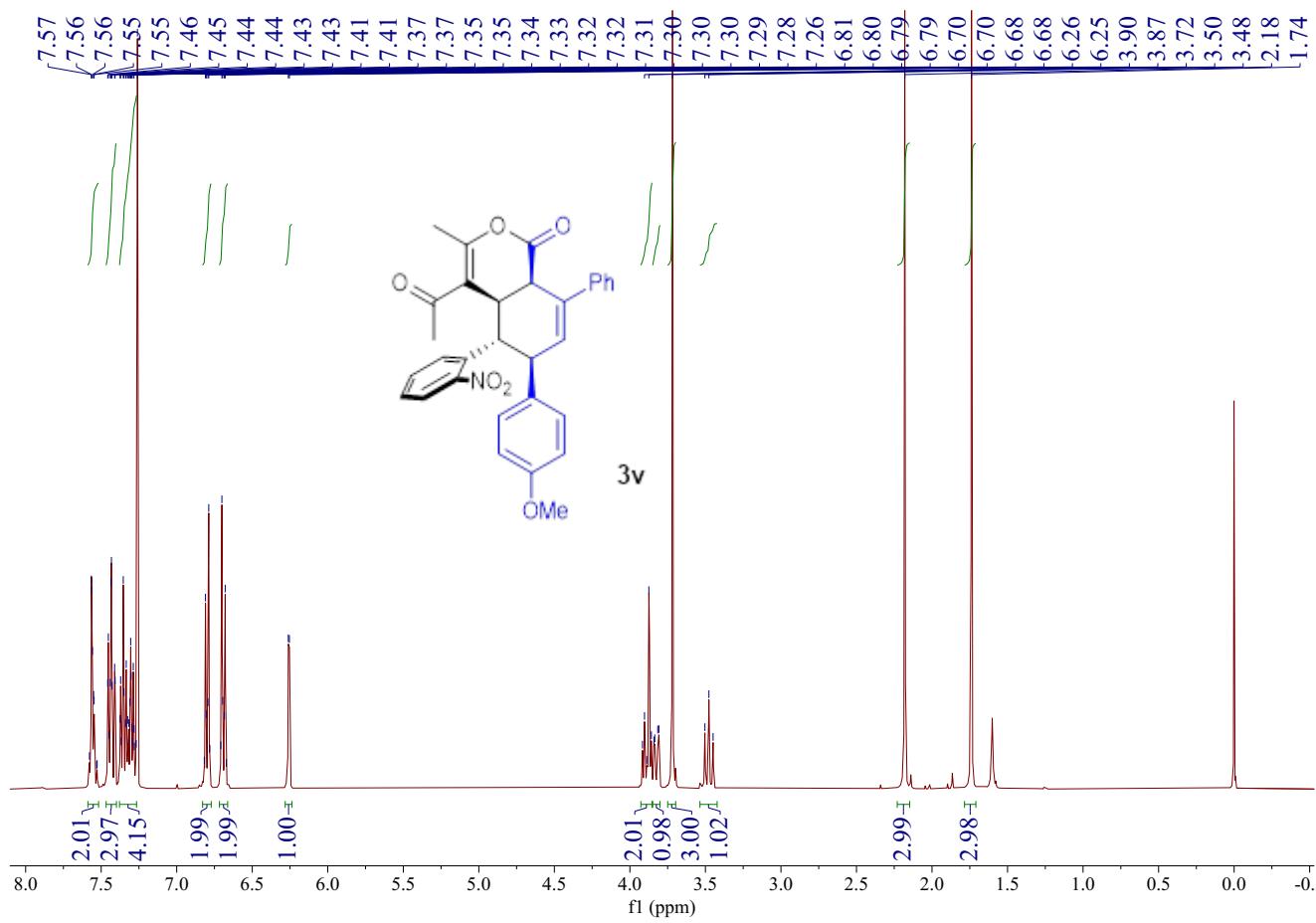
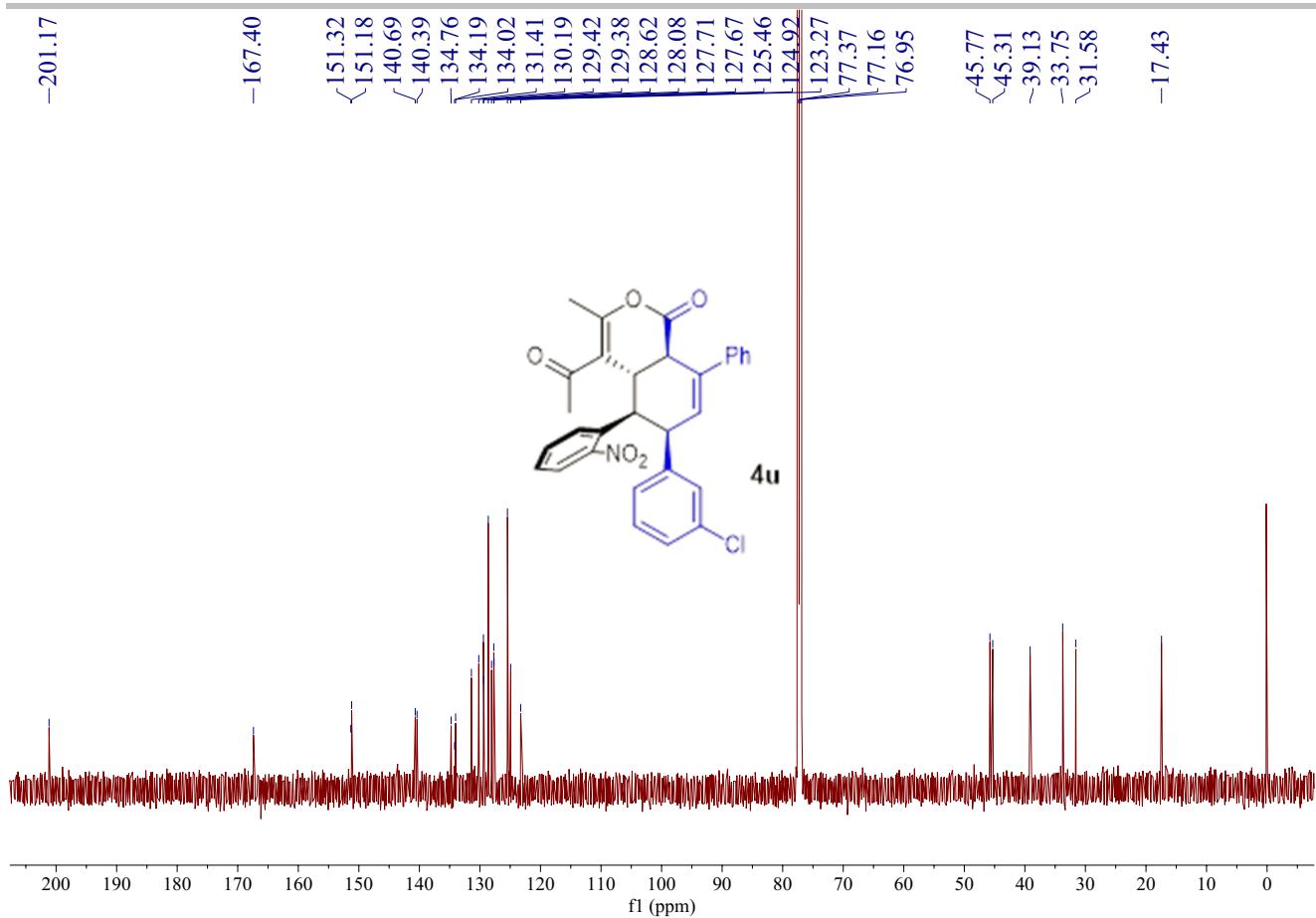


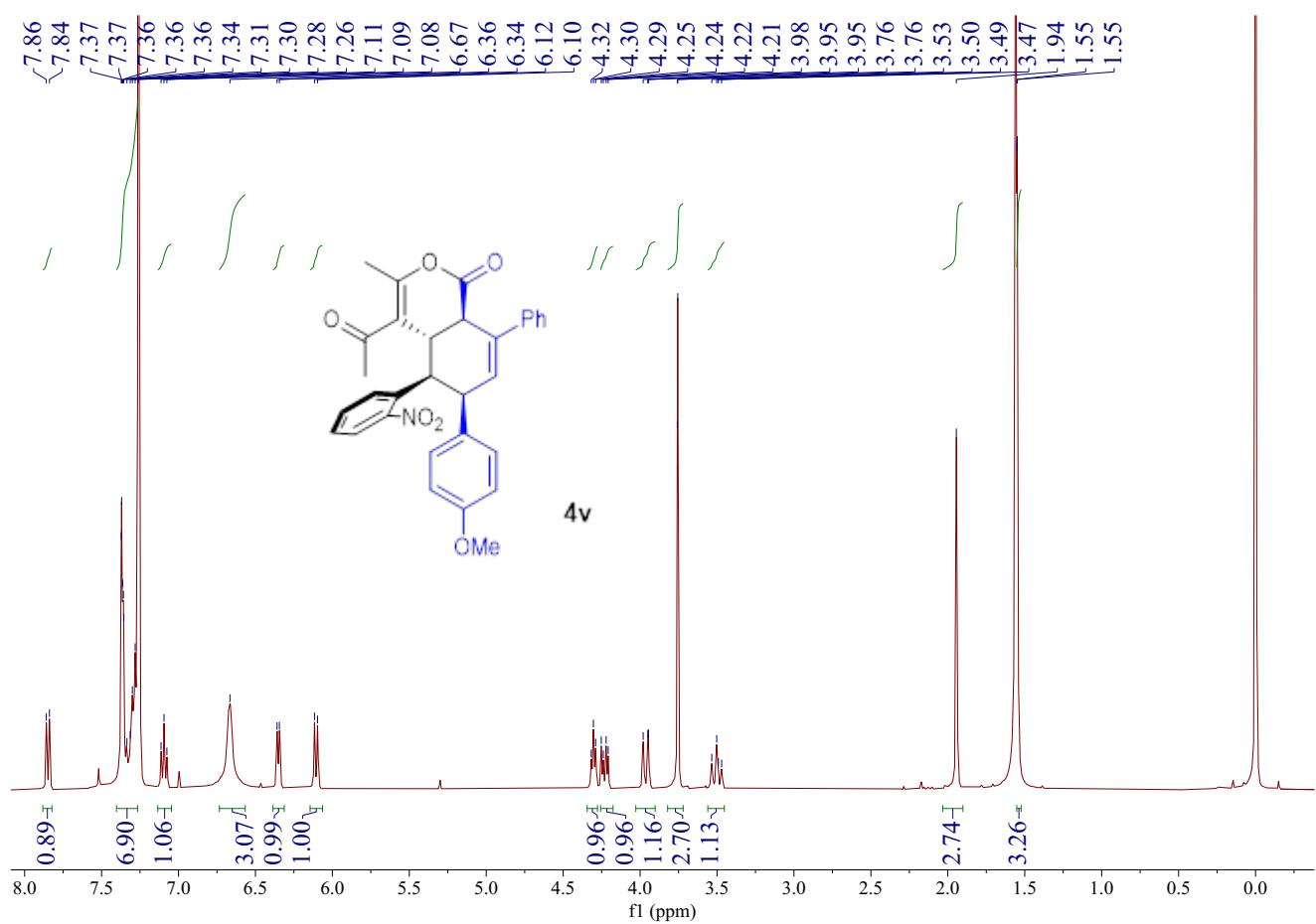
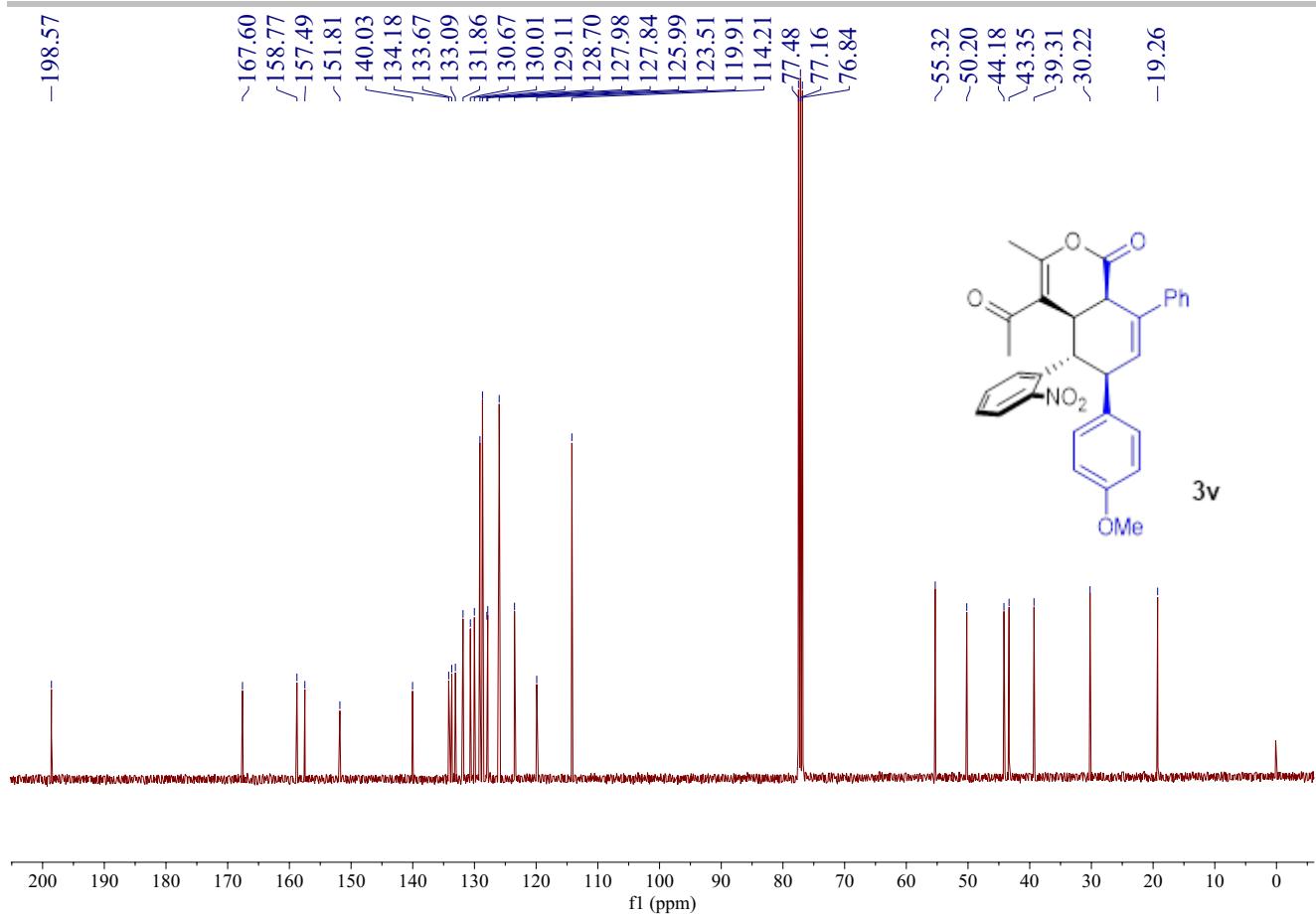


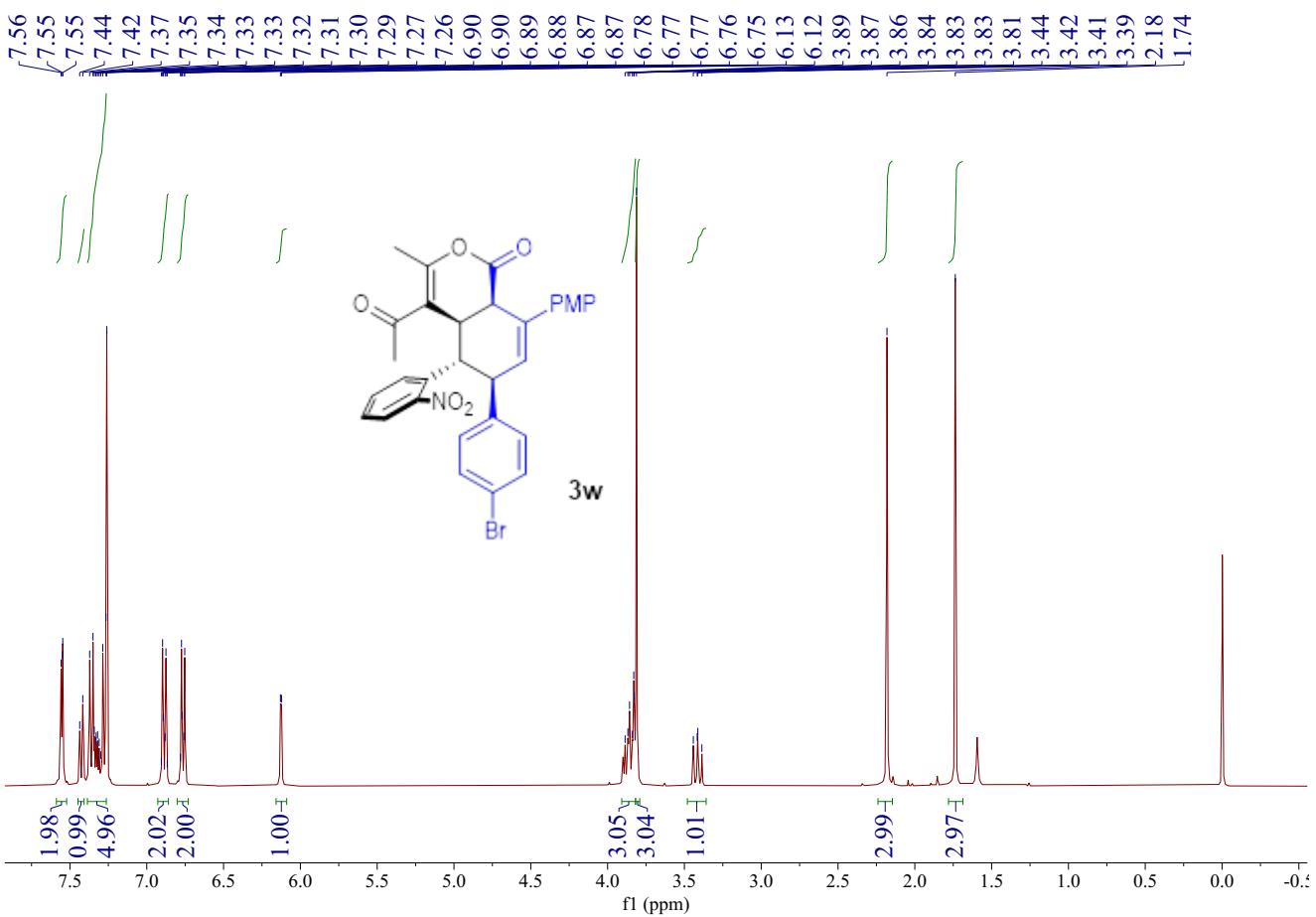
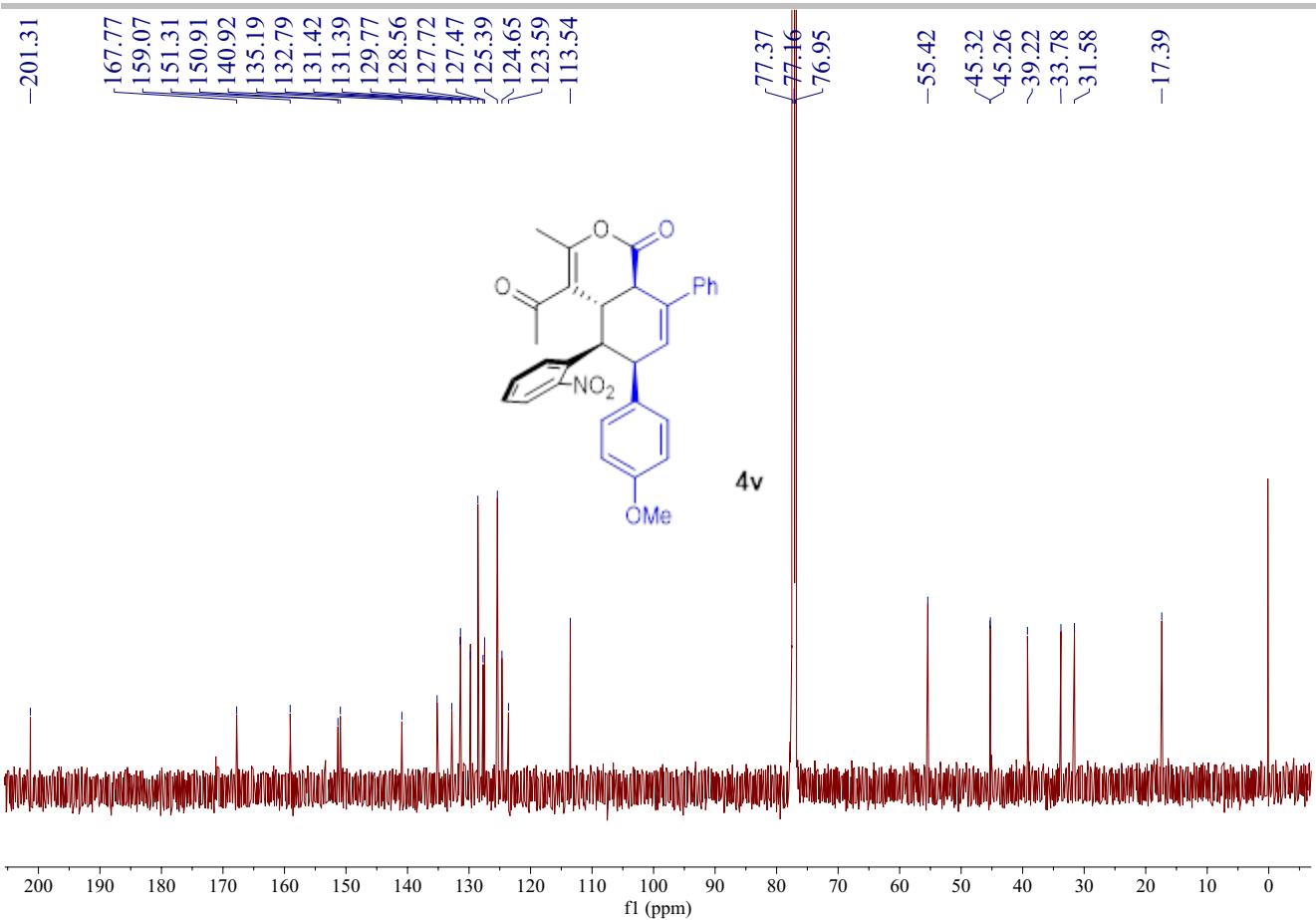


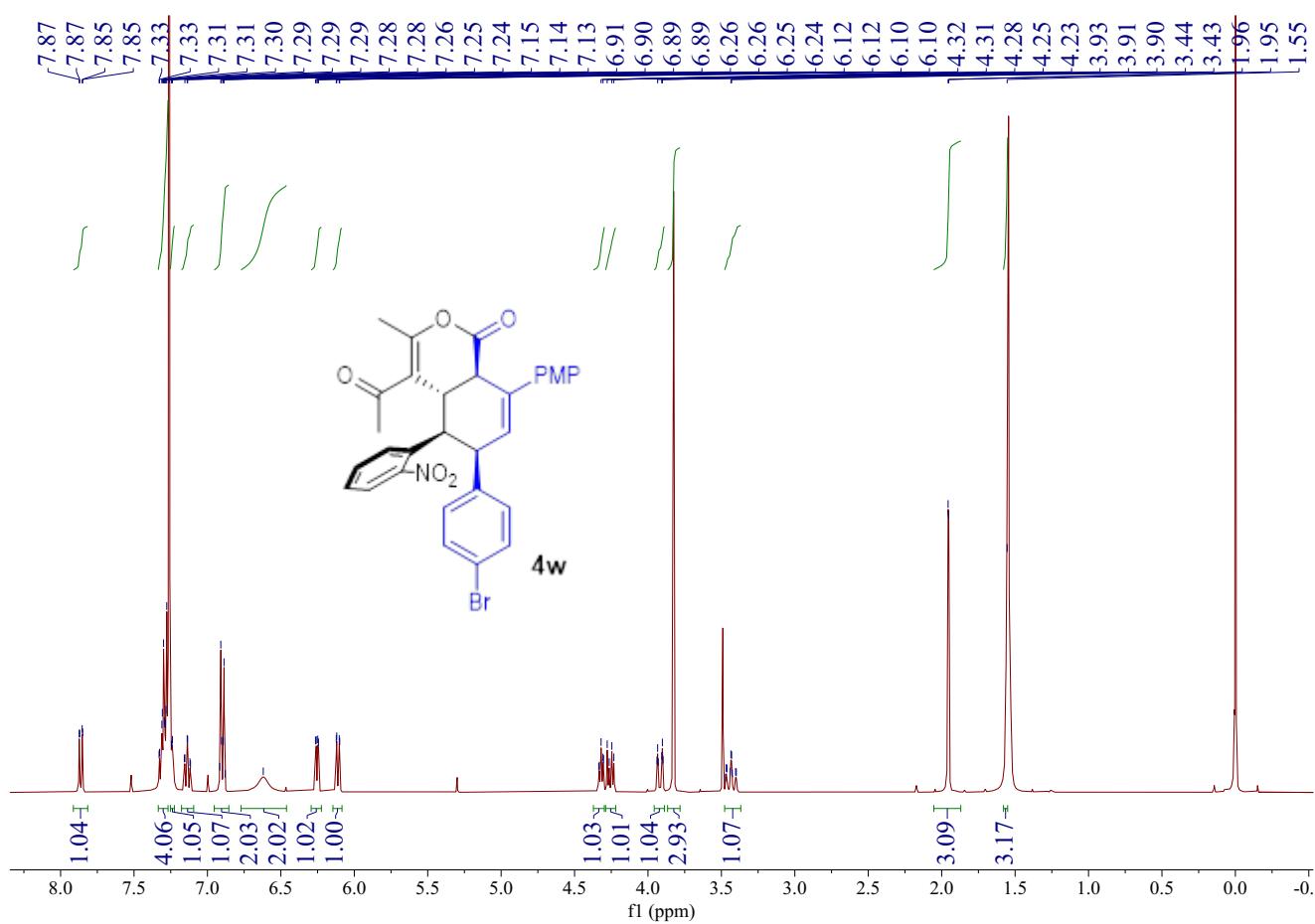
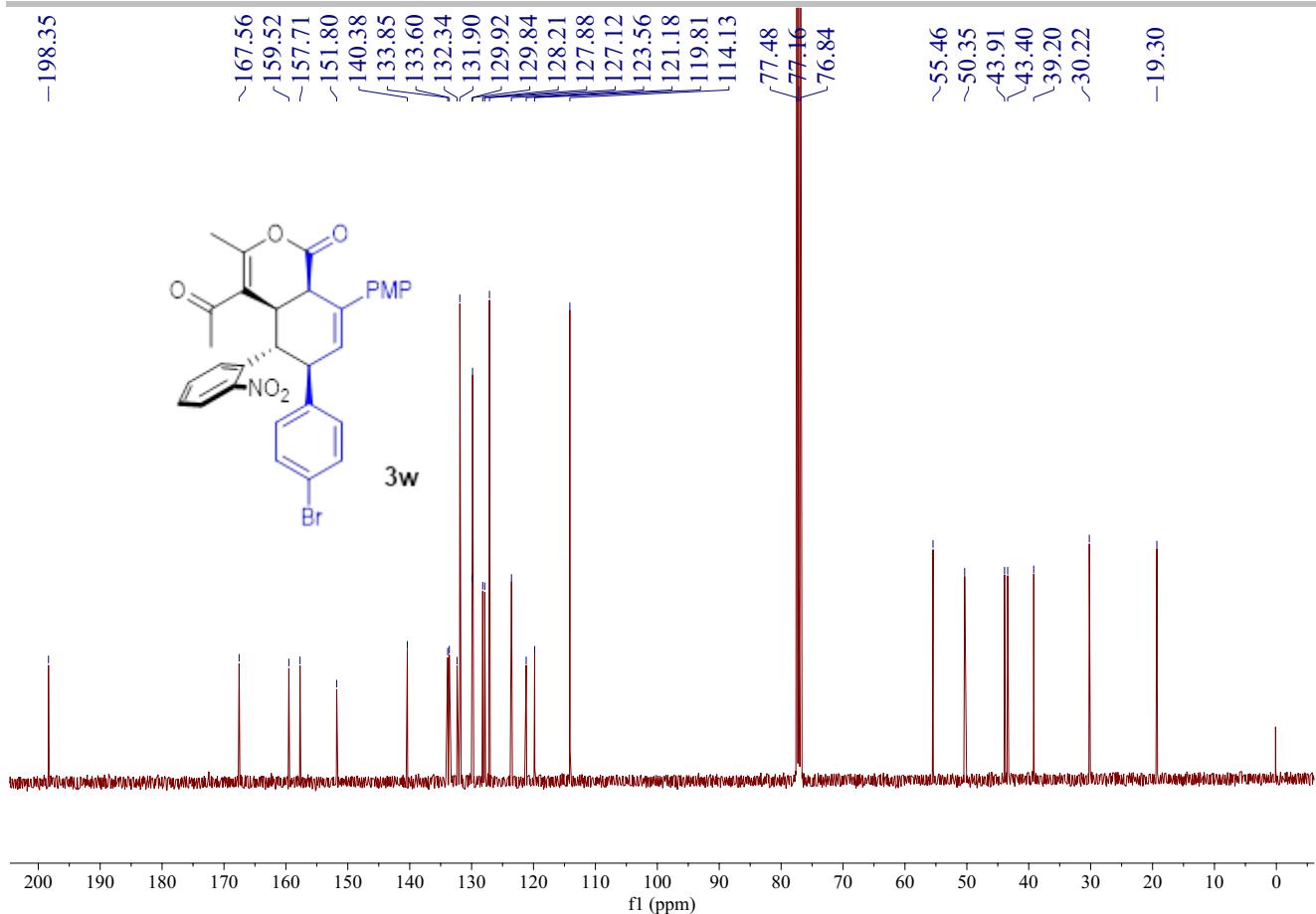


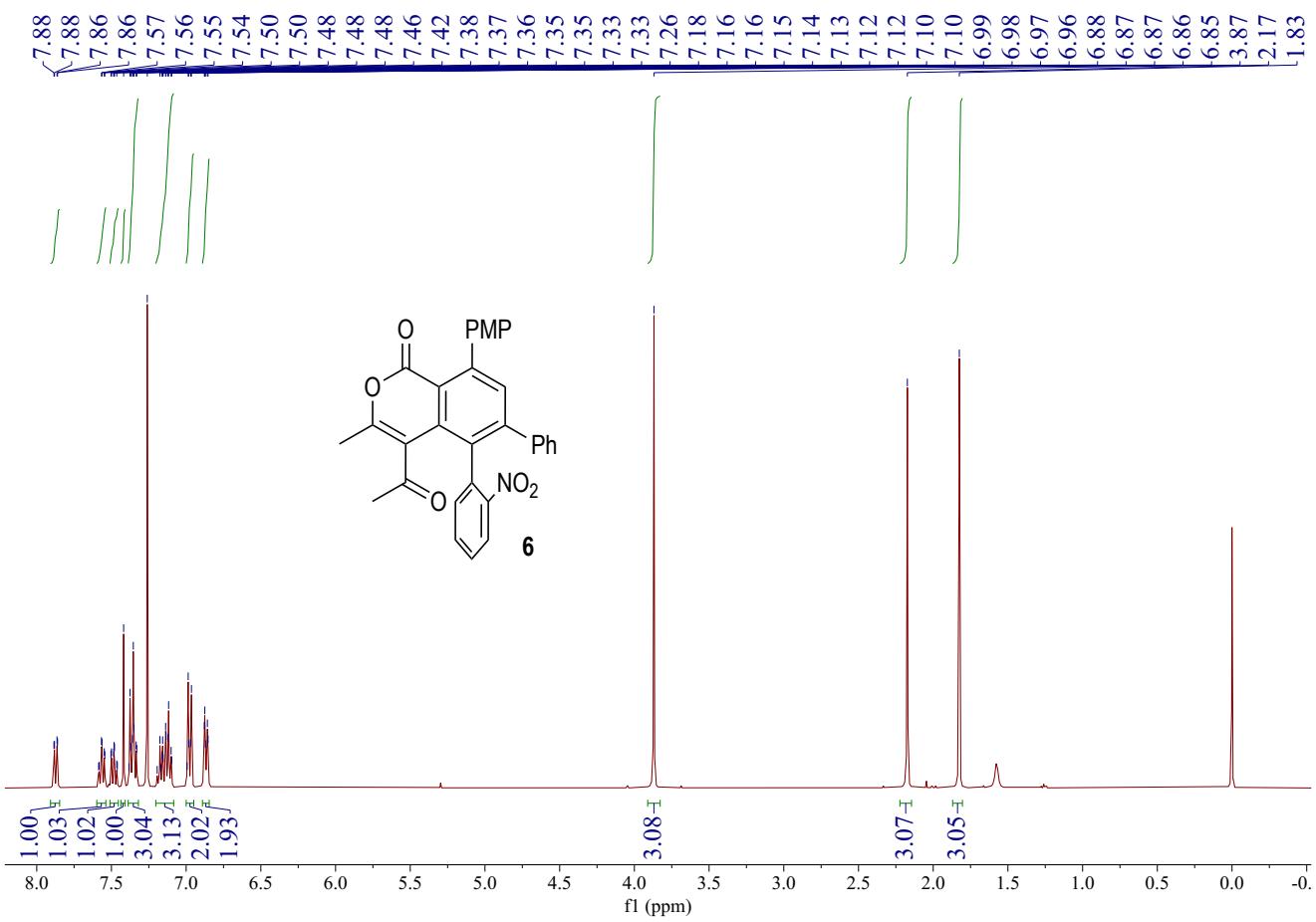
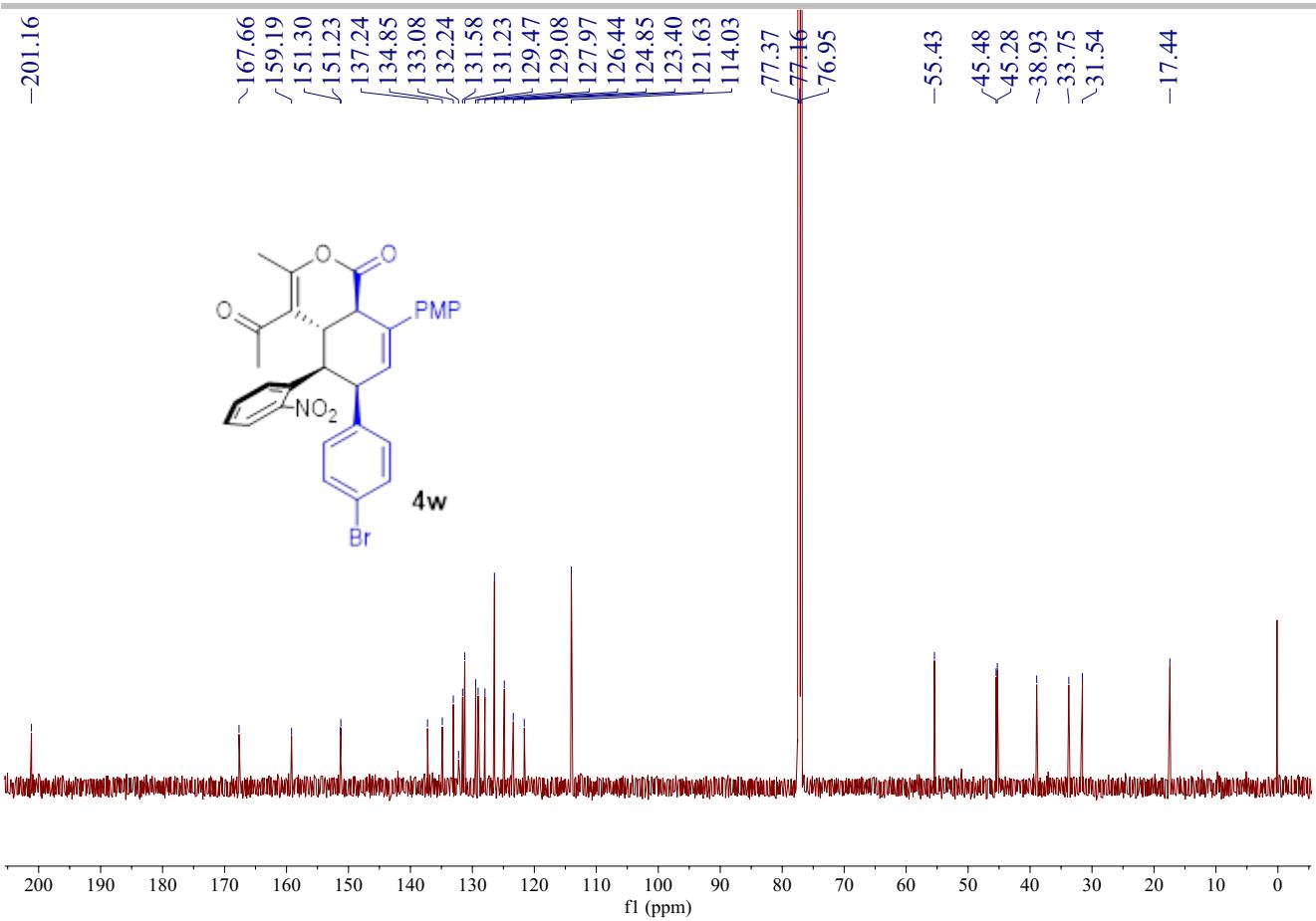


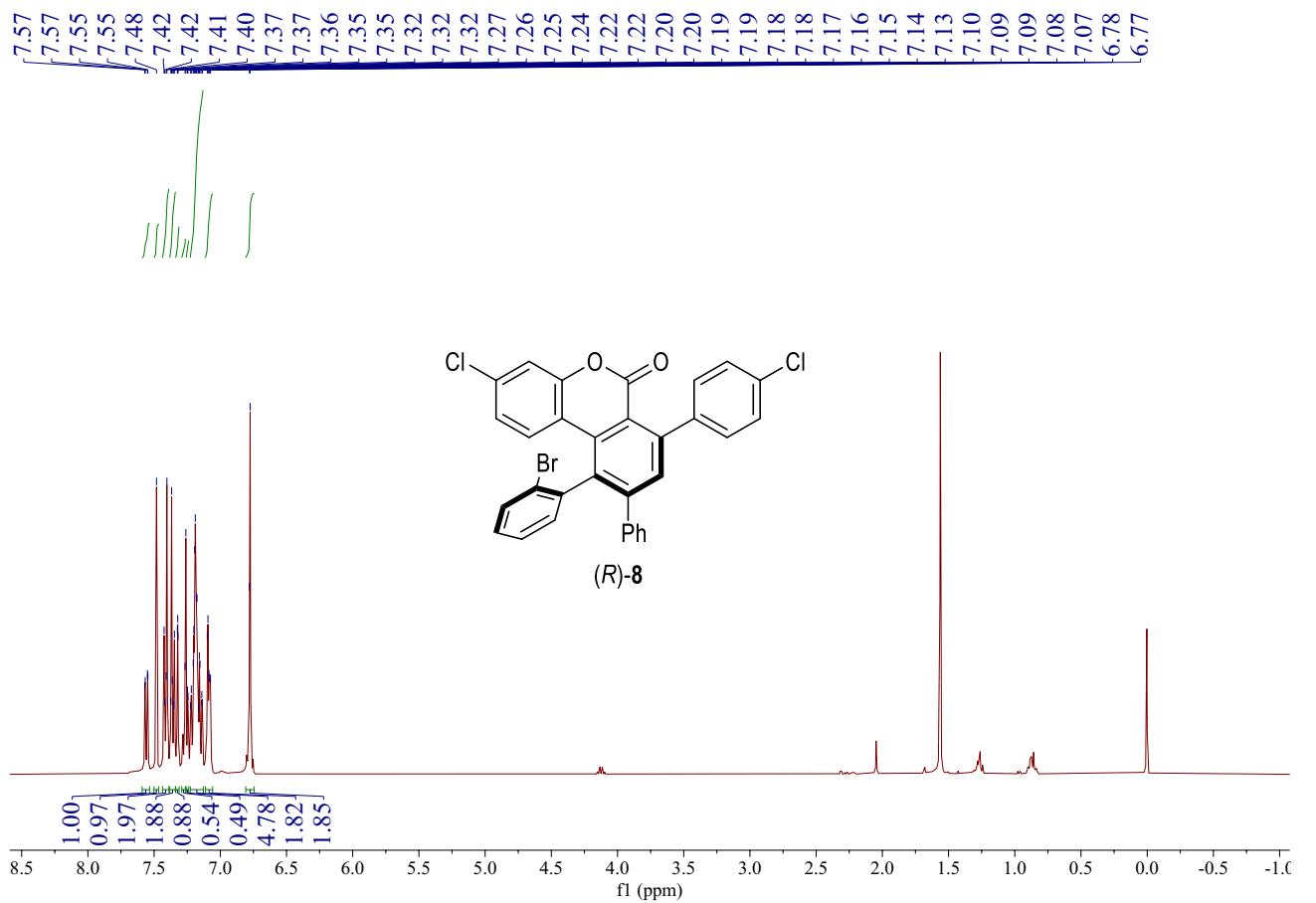
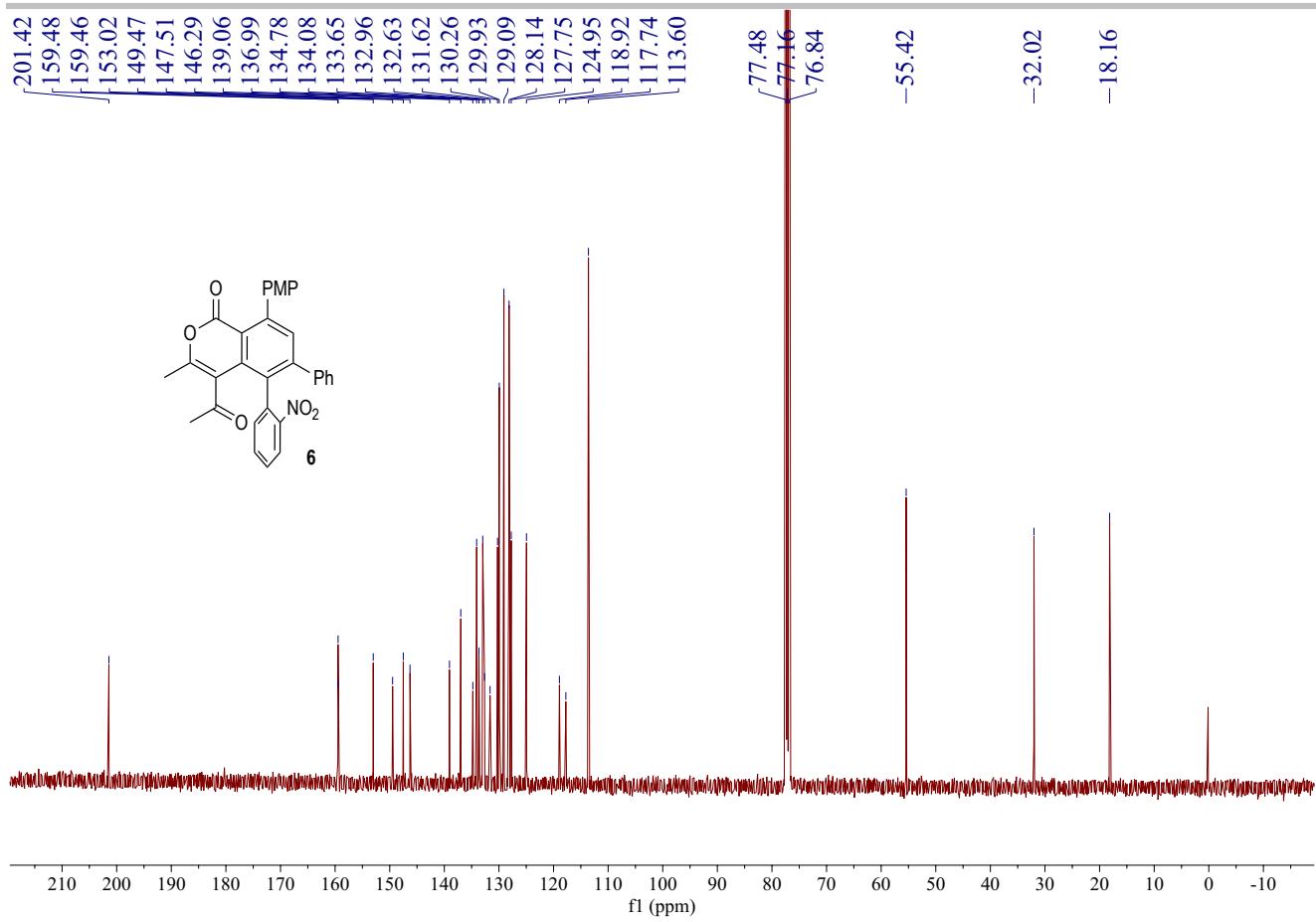


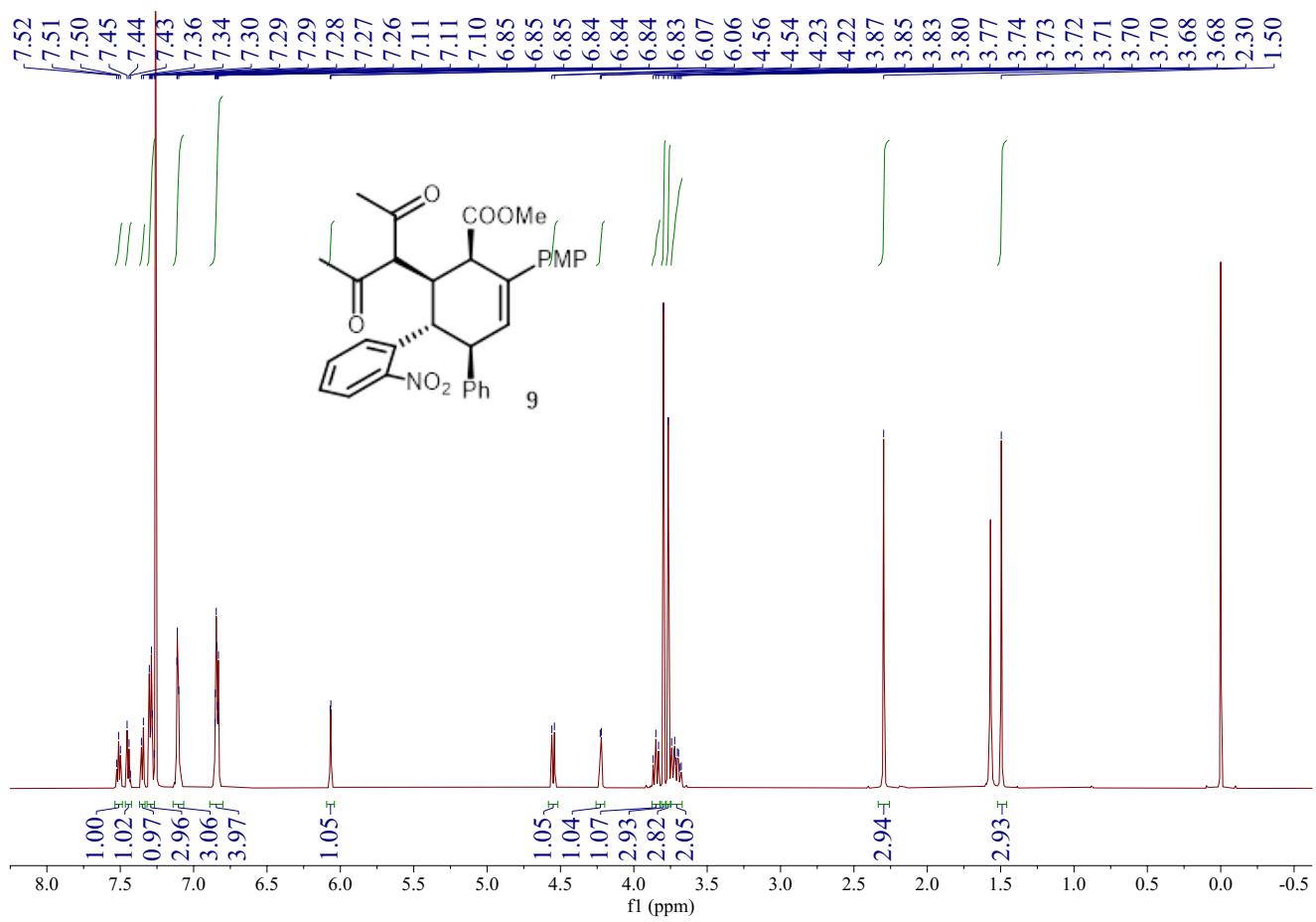
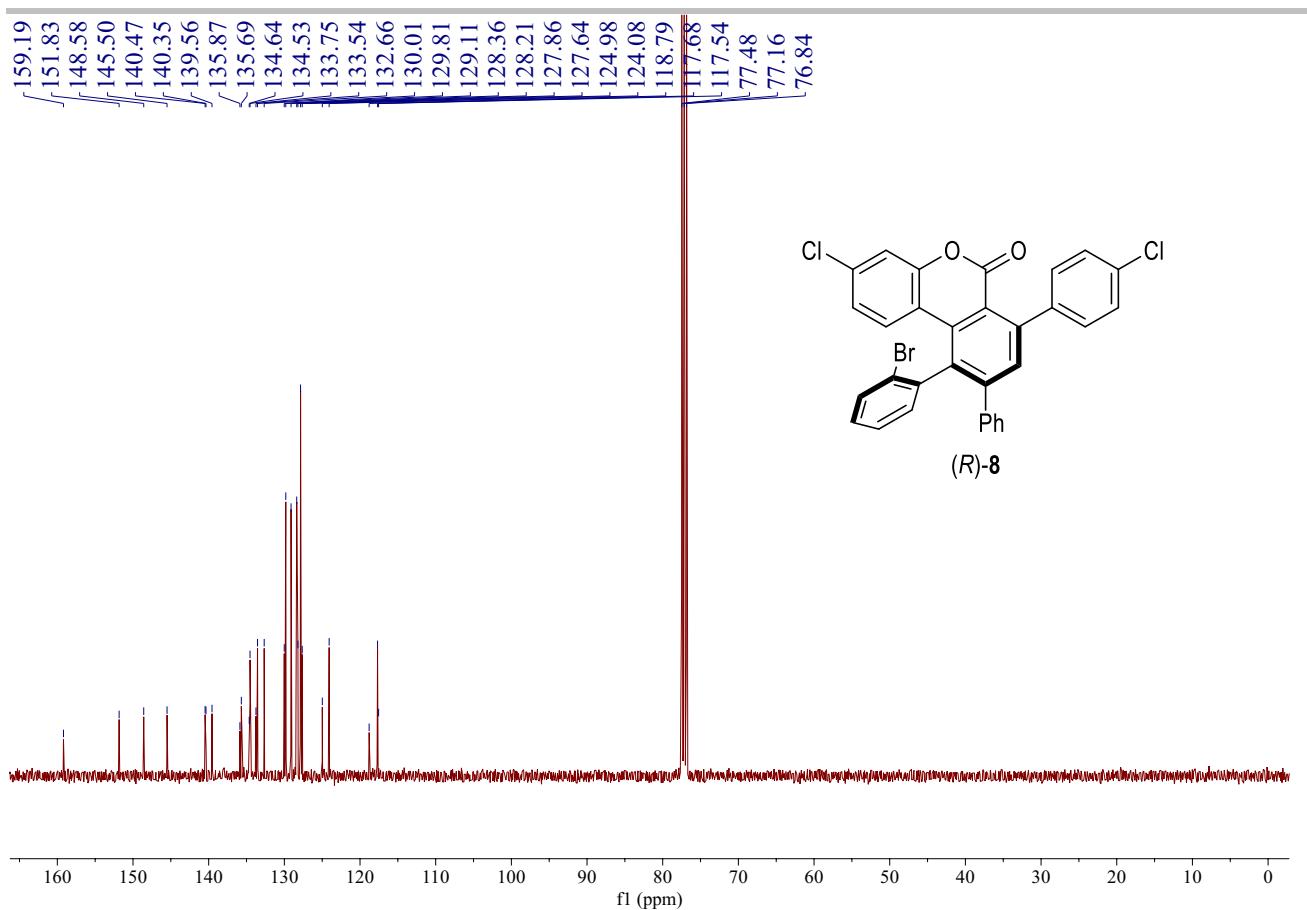


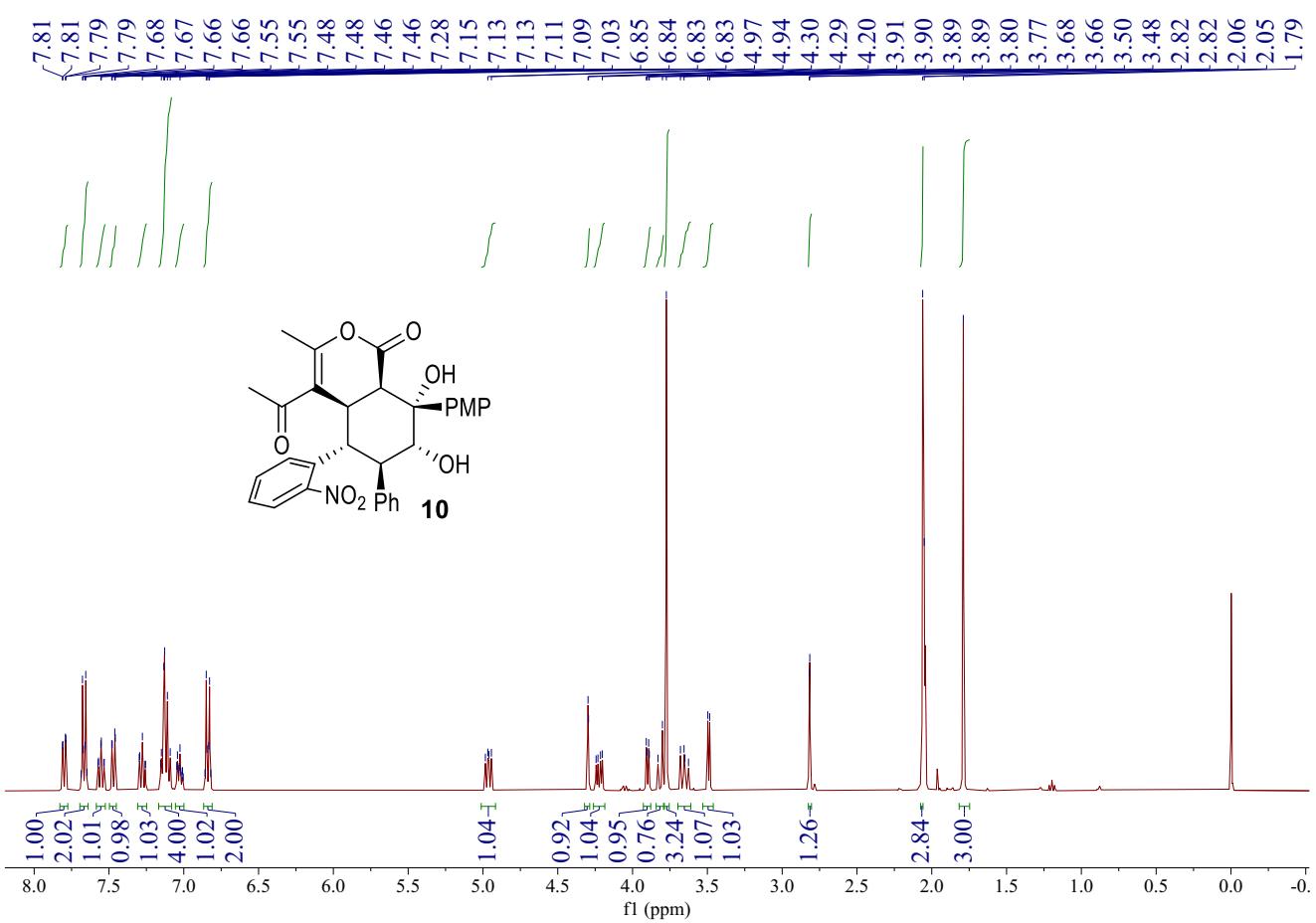
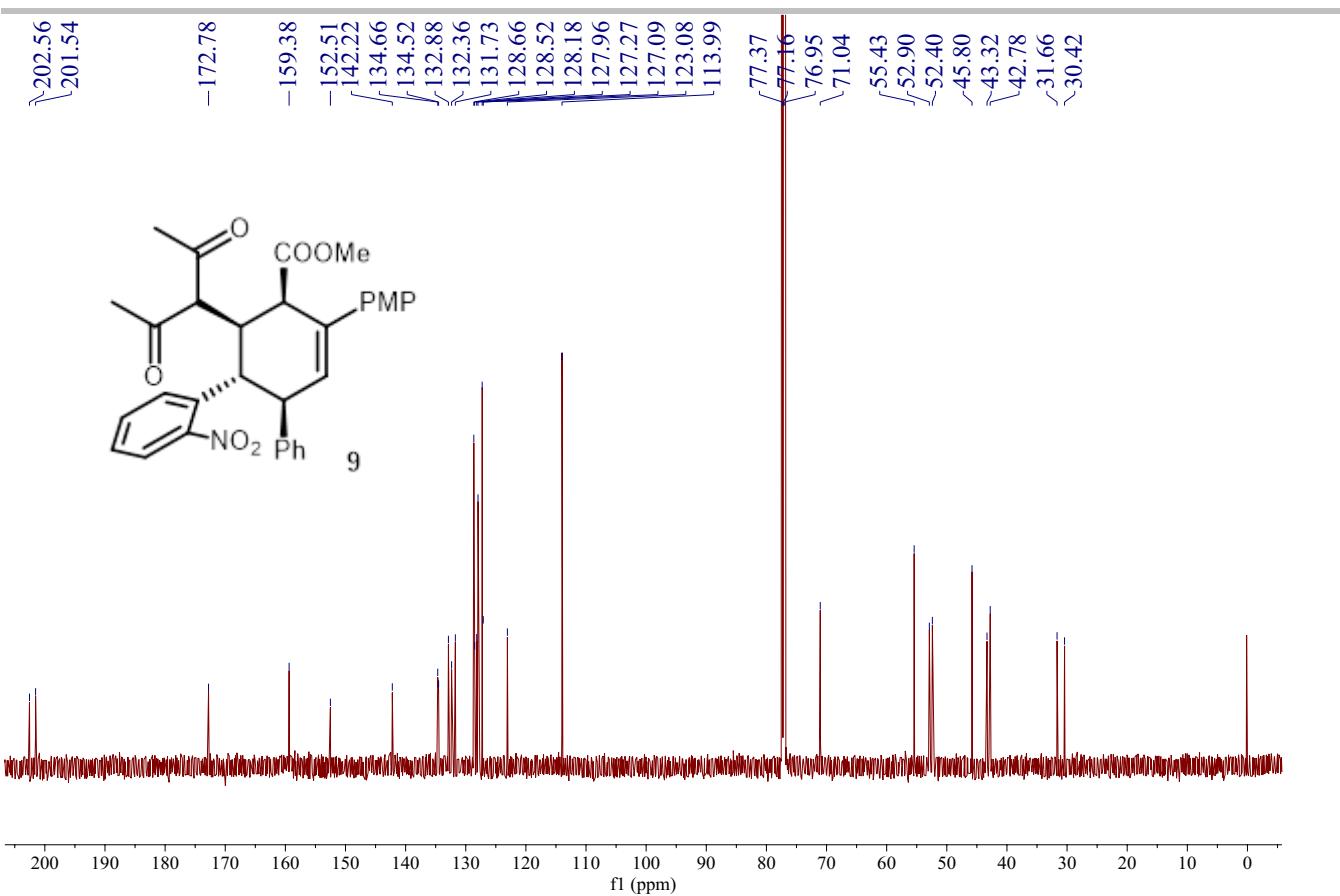


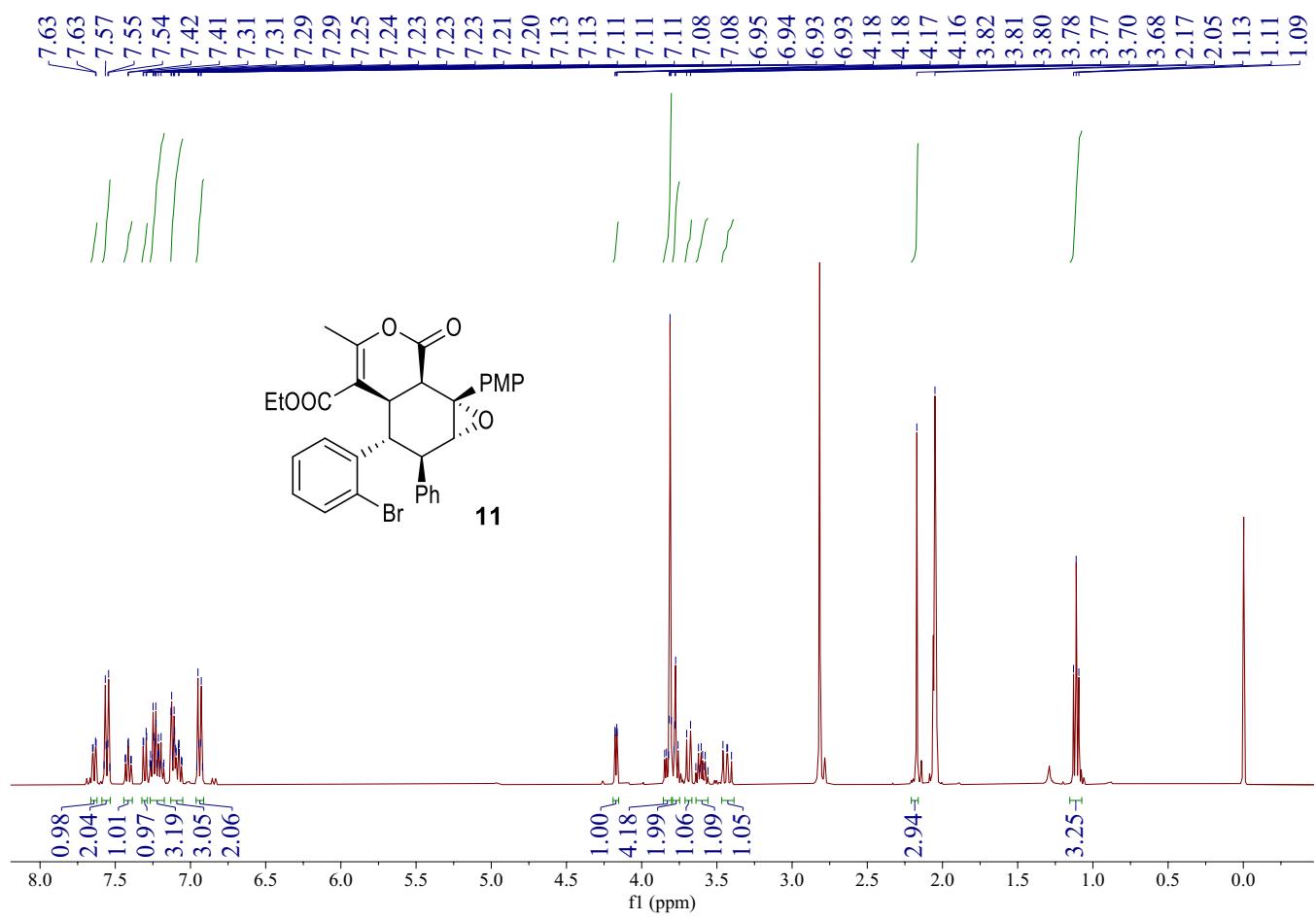
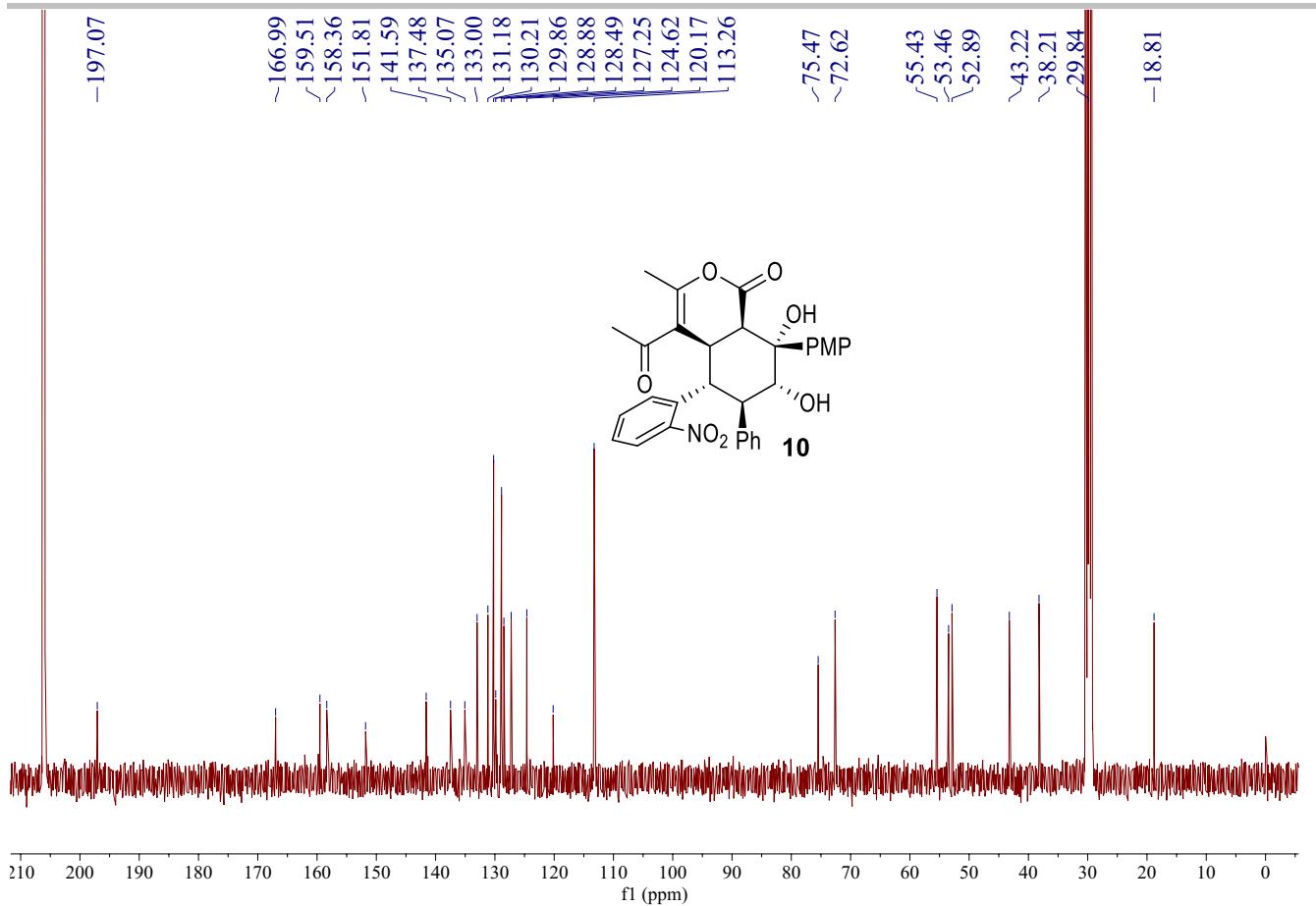


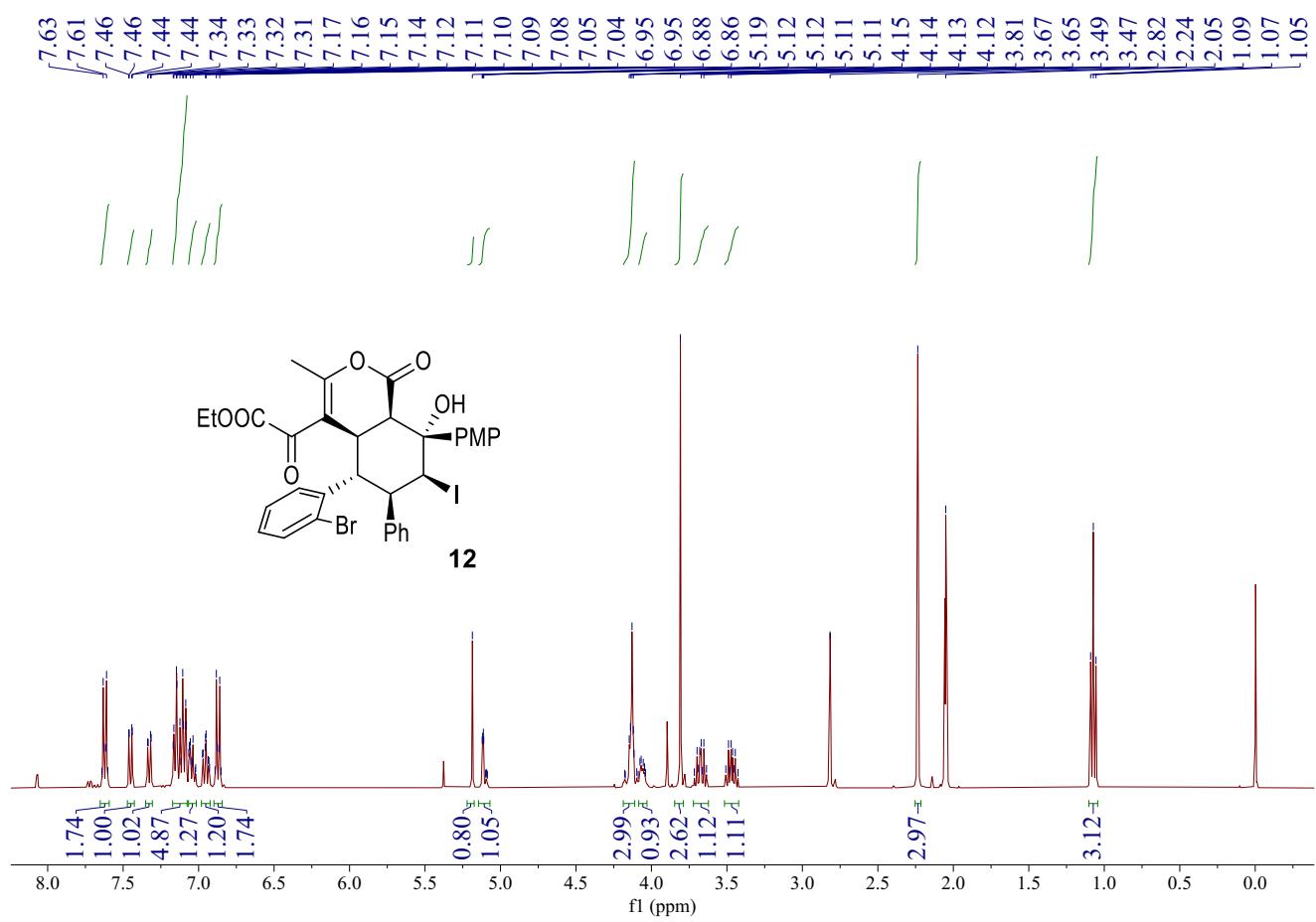
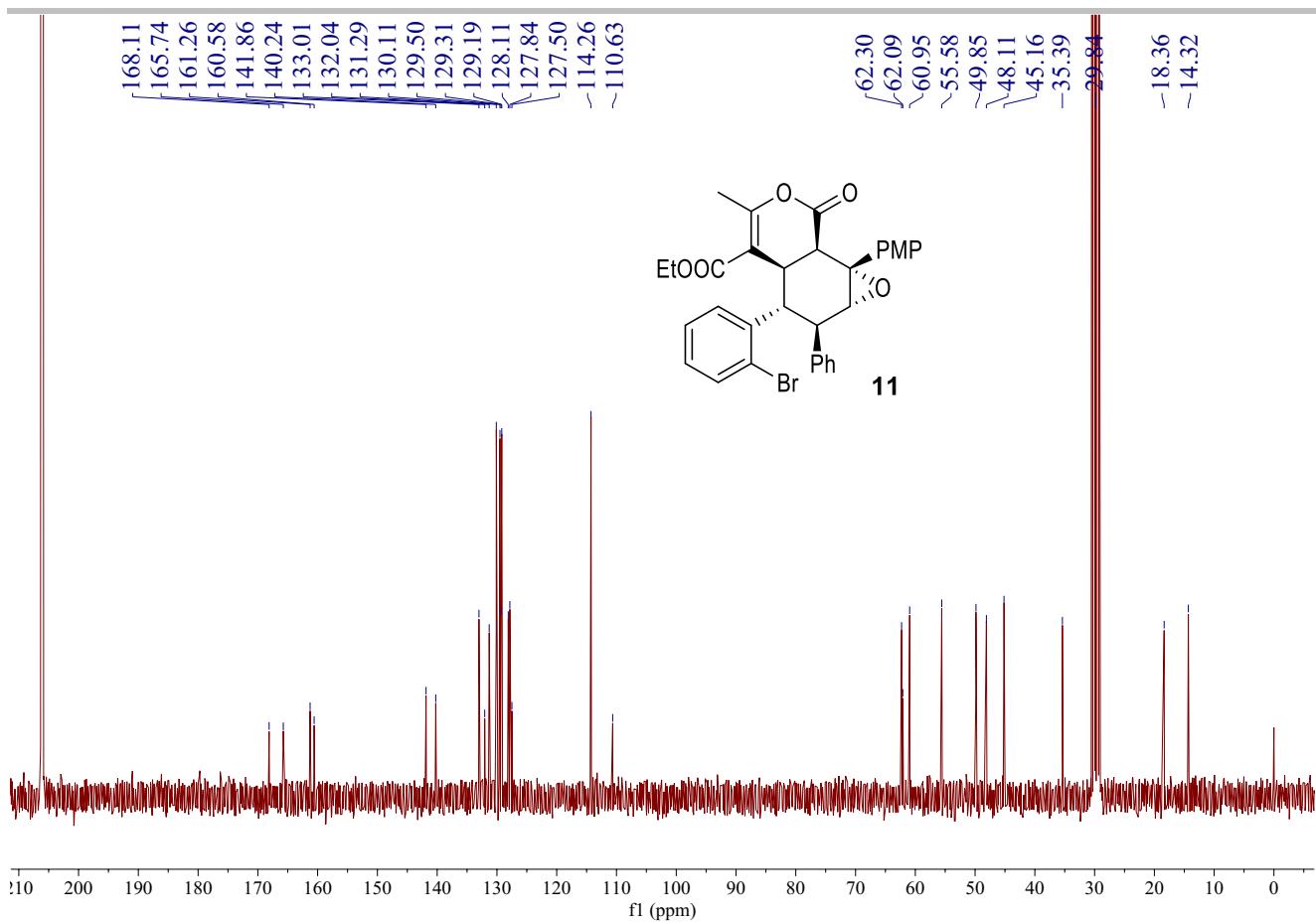


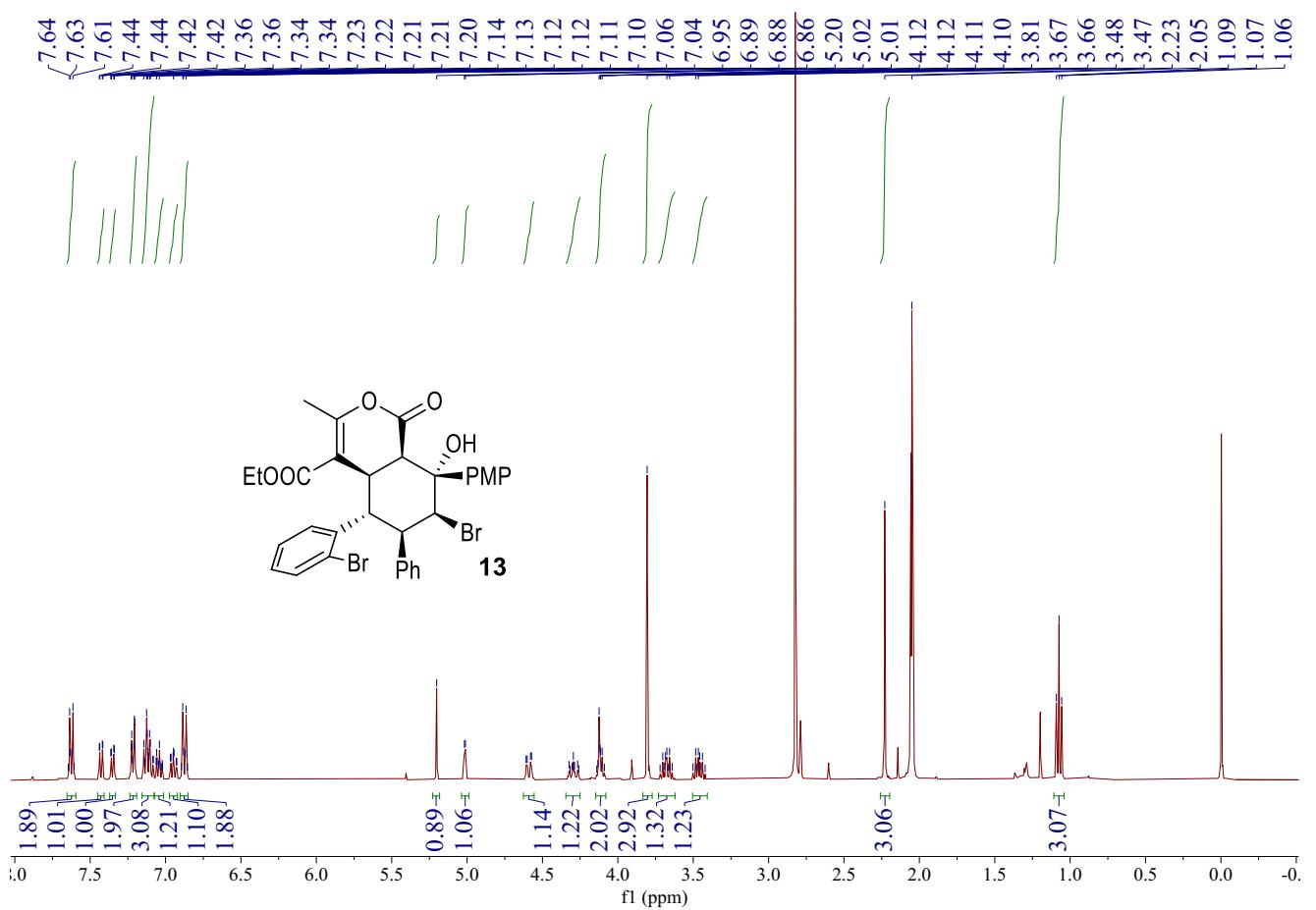
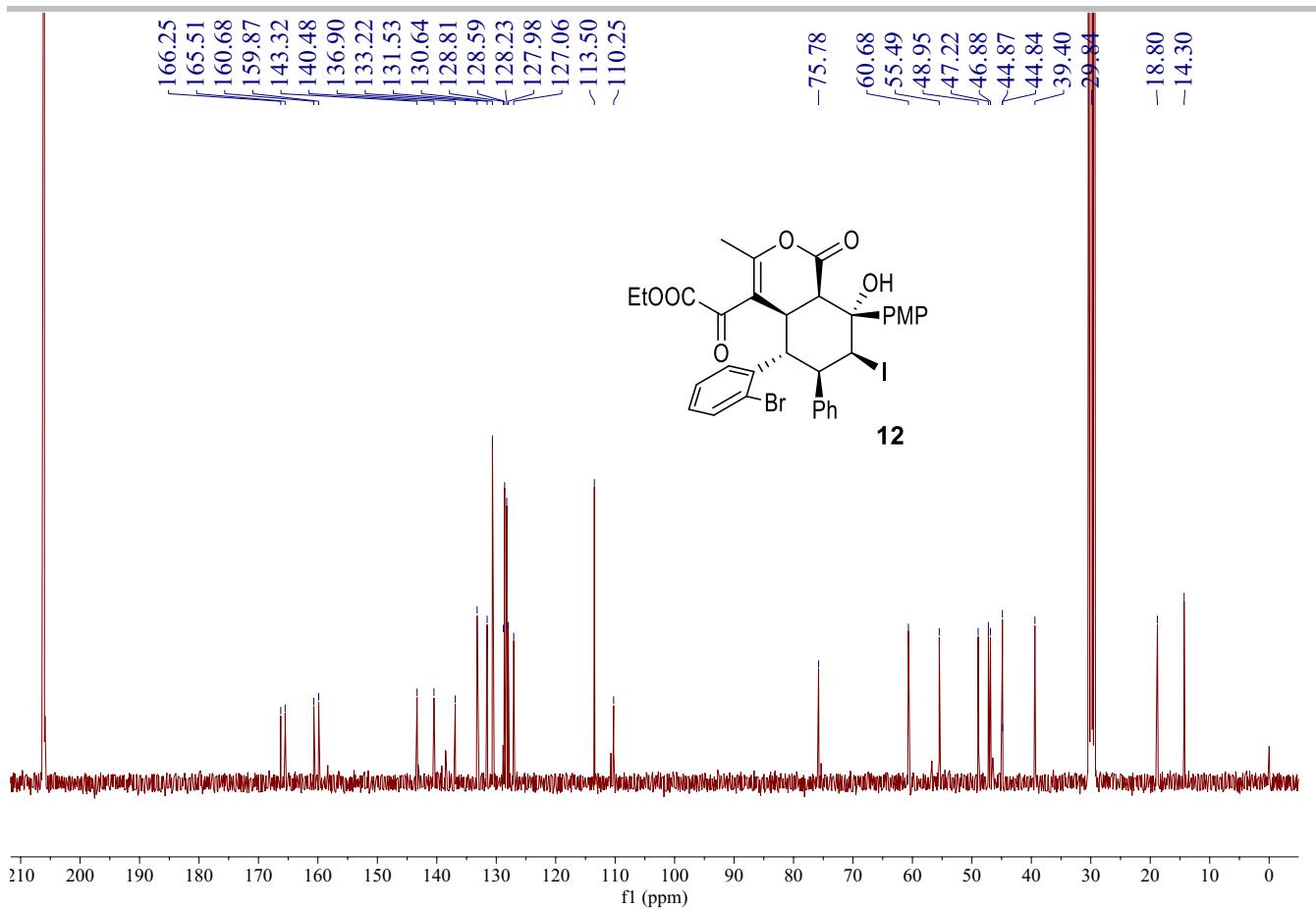


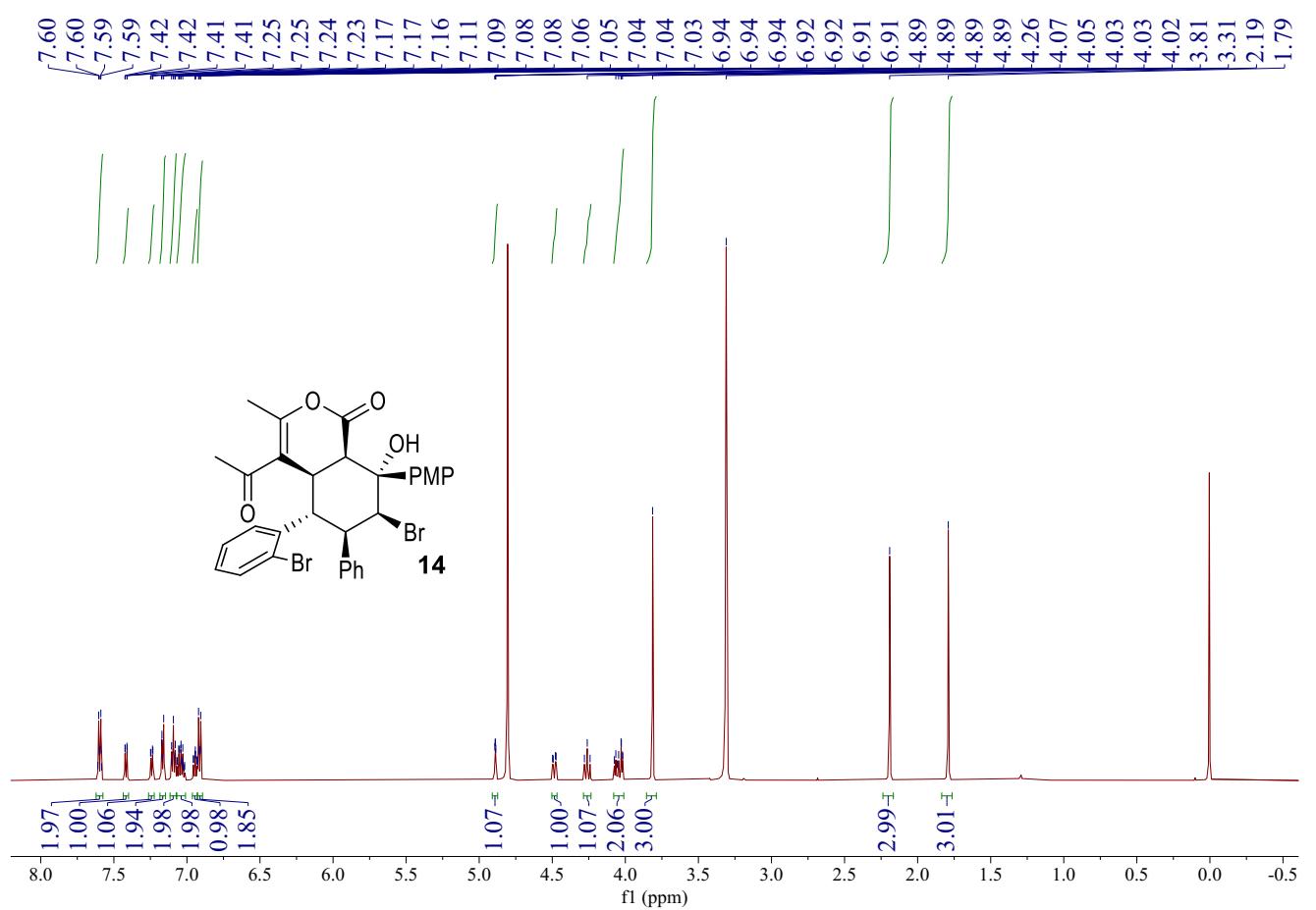
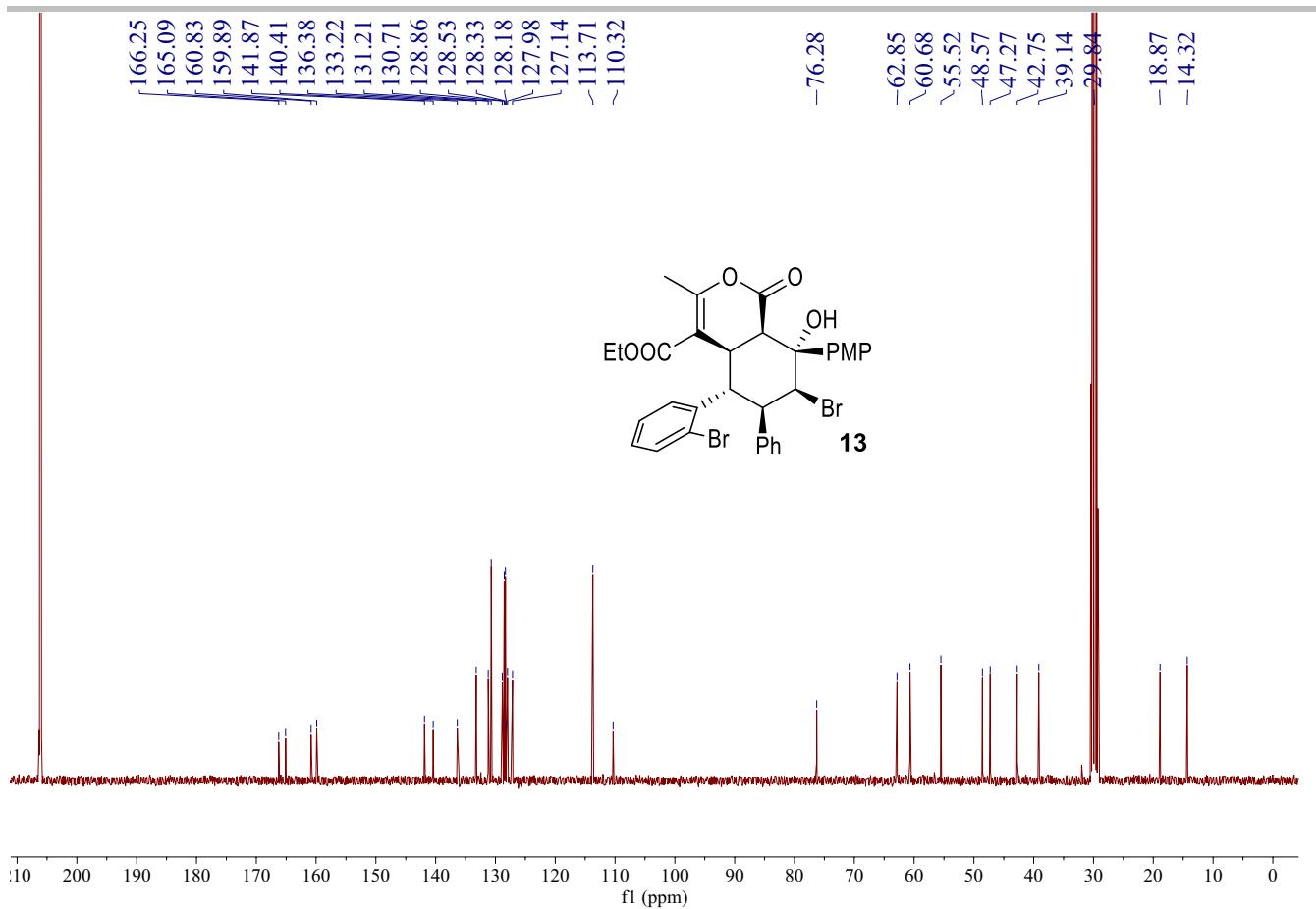




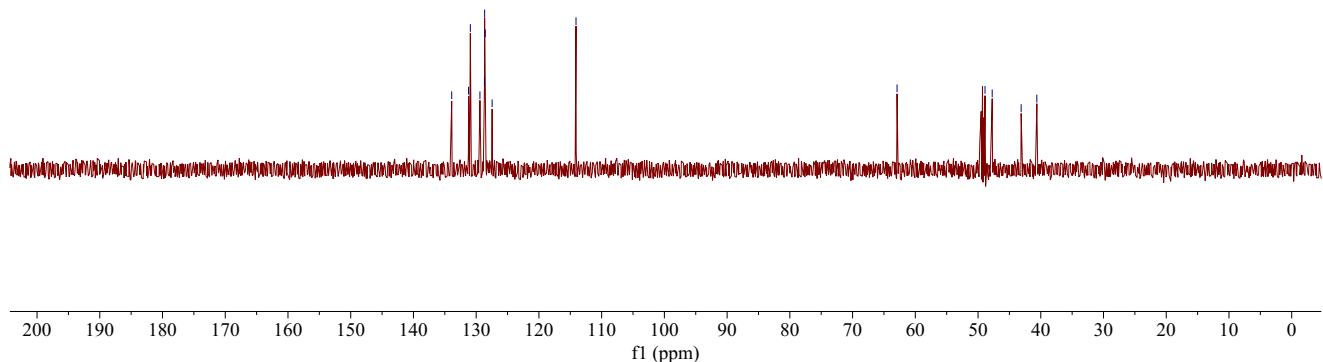
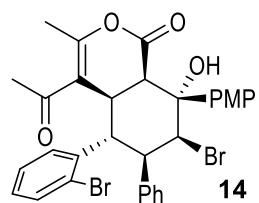
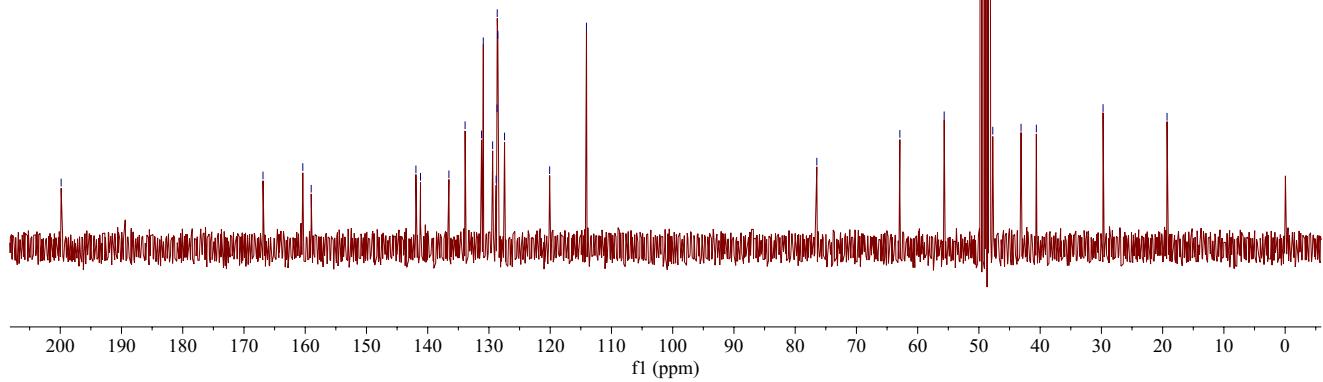
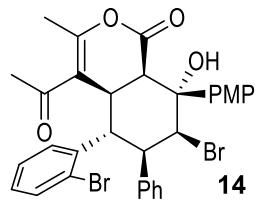
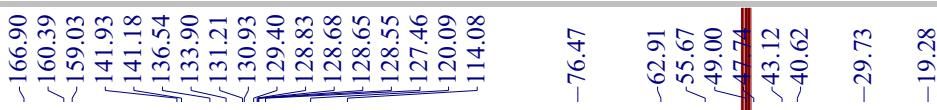








-199.85



XIII. References cited in the SI

- [1] (a) Liu, S., Nakajima, K., Nishibayashi, Y., *RSC Adv.* 2019, **9**, 18918-18922; (b) Hong, S., Shin, Y., Jung, M., Ha, M. W., Park, Y., Lee, Y.-J., Shin, J., Oh, K. B., Lee, S. K., Park, H.-g., *Eur. J. Med. Chem.* 2015, **96**, 218-230; (c) Guo, Y., Wu, L., Qiu, F. G., *Org. Lett.* 2022, **24**, 8370-8374.
- [2] (a) Yasmin, N., Ray, J. K., *Synlett* 2010, **6**, 924-930; (b) Singha, R., Dhara, S., Ray, J. K., *Tetrahedron Lett.* 2013, **54**, 4841-4843; (c) Zhu, T., Mou, C., Li, B., Smetankova, M., Song, B.-A., Chi, Y. R., *J. Am. Chem. Soc.* 2015, **137**, 5658-5661; (d) Chatterjee, I., Bastida, D., Melchiorre, P., *Adv. Synth. Catal.* 2013, **355**, 3124-3130; (e) Xu, K., Li, W., Zhu, S., Zhu, T., *Angew. Chem. Int. Ed.* 2019, **58**, 17625-17630; (f) Li, Y., Luo, H., Tang, Z., Li, Y., Du, L., Xin, X., Li, S., Li, B., *Org. Lett.* 2021, **23**, 6450-6454.
- [3] Ma, L.-J., Li, X.-X., Kusuyama, T., El-Sayed, I. E.-T., Inokuchi, T., *J. Org. Chem.* 2009, **74**, 9218-9221.
- [4] Zhang, M., Ruzi, R., Li, N., Xie, J., Zhu, C., *Org. Chem. Front.* 2018, **5**, 749-752.
- [5] Jackman, H., Marsden, S. P., Shapland, P., Barrett, S., *Org. Lett.* 2007, **9**, 5179-5182.
- [6] De Corso, R. D., Panunzi, B., Tingoli, M., *Tetrahedron Lett.* 2001, **42**, 7245-7247.
- [7] Wang, H., Toh, R. W., Shi, X., Wang, T., Cong, X., Wu, J., *Nat. Commun.* 2020, **11**, 4462.
- [8] Frisch, M. J., Trucks, G. W., Schlegel, H. B., Scuseria, G. E., Robb, M. A., Cheeseman, J. R., Scalmani, G., Barone, V., Petersson, G. A., Nakatsuji, H., Li, X., Caricato, M., Marenich, A. V., Bloino, J., Janesko, B. G., Gomperts, R., Mennucci, B., Hratchian, H. P., Ortiz, J. V., Izmaylov, A. F., Sonnenberg, J. L., Williams-Young, D., Ding, F., Lipparini, F., Egidi, F., Goings, J., Peng, B., Petrone, A., Henderson, T., Ranasinghe, D., Zakrzewski, V. G., Gao, J., Rega, N., Zheng, G., Liang, W., Hada, M., Ehara, M., Toyota, K., Fukuda, R., Hasegawa, J., Ishida, M., Nakajima, T., Honda, Y., Kitao, O., Nakai, H., Vreven, T., Throssell, K., Montgomery, J. A. Jr., Peralta, J. E., Ogliaro, F., Bearpark, M. J., Heyd, J. J., Brothers, E. N., Kudin, K. N., Staroverov, V. N., Keith, T. A., Kobayashi, R., Normand, J., Raghavachari, K., Rendell, A. P., Burant, J. C., Iyengar, S. S., Tomasi, J., Cossi, M., Millam, J. M., Klene, M., Adamo, C., Cammi, R., Ochterski, J. W., Martin, R. L., Morokuma, K., Farkas, O., Foresman, J. B., Fox, D. J., Gaussian, Inc., Wallingford CT 2016.
- [9] (a) Becke, A. D., *J. Chem. Phys.* 1993, **98**, 5648; (b) Becke, A. D., *J. Chem. Phys.* 1993, **98**, 5648-5652; (c) Lee, C., Yang, W., Parr, R. G., *Phys. Rev. B* 1988, **37**, 785-789; (d) Becke, A. D., *Phys. Rev. A* 1988, **38**, 3098-3100.
- [10] Lu, T., Chen, F., *J. Comput. Chem.* 2012, **33**, 580-592.
- [11] Humphrey, W., Dalke, A., Schulten, K., *J. Mol. Graphics* 1996, **14**, 33-38.
- [12] (a) Liu, W.-B., Zheng, C., Zhuo, C.-X., Dai, L.-X., You, S.-L., *J. Am. Chem. Soc.* 2012, **134**, 4812-4821; (b) Huang, Y., Yang, X., Lv, Z., Cai, C., Kai, C., Pei, Y., Feng, Y., *Angew. Chem. Int. Ed.* 2015, **54**, 7299-7302.