Supporting information for

Visible-Light-Driven Decarboxylative C(sp³)-H Alkylation of Glycine Derivatives via in situ formation of NHPI Esters

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1. Experimental Section

1.1 General Considerations

All manipulations were conducted with Schlenk tube. ¹H NMR spectra were recorded on Bruker AVIII-400 spectrometers. Chemical shifts (in ppm) were referenced to tetramethylsilane ($\delta = 0$ ppm) in CDCl₃ as an internal standard. Data were reported as follows: chemical shift (ppm), multiplicity (s = singlet, d = doublet, t = triplet, td = doublet of triplets, m = multiplet, br = broad signal), coupling constants (Hz), integration and assignment. ¹³C NMR spectra were obtained by using the same NMR spectrometers and were calibrated with CDCl₃ ($\delta = 77.00$ ppm). High resolution mass spectrometry (HRMS) data were obtained on a QTOF mass analyzer with electrospray ionization (ESI) through a Bruker Daltonicmior OTOF-QII. All anhydrous solvents were commercially supplied from Energy-Chemical. Substrates were purchased from Bidepharm, Aladdin, Energy, or synthesized according to the procedures outlined below. Unless otherwise noted, materials obtained from commercial suppliers were used without further purification.

1.2 Reaction setup

The optimization of reaction conditions was using parallel photoreactor (Purchasing from Shanghai 3STechnology Co., Ltd, AF2 type) irradiated with 12 W blue-light-emitting diodes (LEDs, 450-455 nm), 25 mL schlenk tubes were used for all 0.2 mmol scale reactions, the temperature was maintained at 30 °C.



(1)

(2)

(1) Reaction sealing tubes and the manual bottle capping tools for 0.2 mmol reactions.

(2) Parallel reaction setups under a photoreactor for 0.2 mmol reactions.

1.3 Starting Materials

(a) Preparation of glycine esters



According to literature reports,^[1] the following substrates **1a-1h** were prepared.

(b) Preparation of glycine dipeptides



According to literature reports,^[2] **1i-11** were prepared.

(c) Carboxylic acid

The substrates **2a-2u** are commercially available and were used as received.

2e: cyclobutanecarboxylic acid

2h: 2,3-dihydro-1H-indene-2-carboxylic acid

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2d: 4-(thiophen-2-yl)butanoic acid

2g: cycloheptanecarboxylic acid

2j: tetrahydro-2H-pyran-4-carboxylic acid 2k: tetrahydrofuran-2-carboxylic acid



2m: pivalic acid



2p: (9Z,12Z)-octadeca-9,12-dienoic acid



2r: (4R)-4-((8R,9S,10S,13R,14S,17R)-10,13-dimethyl-3,7,12-trioxohexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoic acid

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2t: 2-cyclopropylacetic acid

2n: 1-methylcyclohexane-1-carboxylic acid 2o: (3r,5r,7r)-adamantane-1-carboxylic acid COOH

Ме

2q: 5-(2,5-dimethylphenoxy)-2,2-dimethylpentanoic acid

ћ_{Ме}



2s: (4*R*)-4-((3*R*,8*R*,9*S*,10*S*,13*R*,14*S*,17*R*)-3-hydroxy-10,13dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoic acid

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2u: hept-6-enoic acid

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butanoic acid

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2f: cyclopentanecarboxylic acid

2i: 2-ethylhexanoic acid



2I: (tert-butoxycarbonyl)proline

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1.4 General procedure for the reaction



In an oven dried 25 mL Schlenk tube was charged with 1 (0.2 mmol, 1.0 equiv), 2 (0.3 mmol, 1.5 equiv), Xantphos (93 mg, 0.16 mmol, 0.8 equiv), ICH₂CH₂I (45 mg, 0.16 mmol, 0.8 equiv), NHPI (49 mg, 0.3 mmol, 1.5 equiv), DMAP (37 mg, 0.3 mmol, 1.5 equiv) and DABCO (34 mg, 0.3 mmol, 1.5 equiv). The tube was then evacuated and back-filled under argon flow (this sequence was repeated three times), anhydrous DMSO (2.0 mL) was added under Ar. The tube was screw capped and heated to 30 °C under irradiation of Blue LEDs (450-455 nm). After stirring for 24 h, t the reaction mixture was quenched by water and extracted with EtOAc three times. The combined organic phases was removed under vacuo. The residue was purified by silica gel column chromatography to afford the product **3** or **4**.

1.5 The effect of different reaction conditions

Table S1. The effect of different solvents a,b

Ph ^H OEt +	O Xan OH <u>I</u> P	utphos (0.8 equiv), ICH ₂ CH ₂ I (0.8 equ NHPI (1.5 equiv), DMAP (1.5 equiv), h-PTZ (5 mol%), DABCO (1.5 equiv) Solvent, 30 °C, 450-455 nm, 24 h.	hiv) Ph N OEt
1a	2a		3а
Entry		Solvents	Yield (%) ^b
1		DMSO	56
2		CH₃CN	11
3		DCE	15
4		1,4-Dioxane	21
5		Acetone	9
6		DMA	15
7		EA	10
8		PhCF ₃	5

^a Standard conditions: **1a** (0.2 mmol, 1.0 equiv), **2a** (0.3 mmol, 1.5 equiv), Xantphos (0.16 mmol, 0.8 equiv), ICH₂CH₂I (0.16 mmol, 0.8 equiv), NHPI (0.3 mmol, 1.5 equiv), DMAP (0.3 mmol, 1.5 equiv), Ph-PTZ (0.01 mmol, 5 mol%), DABCO (0.3 mmol, 1.5 equiv), Solvent (2 mL), 450-455 nm, at 30 °C for 24 h. ^b Isolated yield.

Ph ^{-N} OEt +	O O O O O H H O H H H I H I H H I H I H H I H I I H I I H I I H I I H I	$\xrightarrow{iv)} Ph' \xrightarrow{H} OEt$
1a	2a	3a
Entry	Bases	Yield (%) ^b
1	DABCO	56
2	BTMG	30
3	NEt ₃	43
6	DIPEA	10
9	DBU	43
4	K ₂ CO ₃	53
7	KHCO3	22
8	NaHCO ₃	21

Table S2. The effect of different bases a,b

^a Standard conditions: **1a** (0.2 mmol, 1.0 equiv), **2a** (0.3 mmol, 1.5 equiv), Xantphos (0.16 mmol, 0.8 equiv), ICH₂CH₂I (0.16 mmol, 0.8 equiv), NHPI (0.3 mmol, 1.5 equiv), DMAP (0.3 mmol, 1.5 equiv), Ph-PTZ (0.01 mmol, 5 mol%), Base (0.3 mmol, 1.5 equiv), DMSO (2 mL), 450-455 nm, at 30 °C for 24 h. ^{*b*} Isolated yield.

Ph ^N OEt +	P-Cat (x equiv), ICH ₂ CH ₂ I (0.8 equiv) NHPI (1.5 equiv), DMAP (1.5 equiv), Ph-PTZ (5 mol%), DABCO (1.5 equiv) DMSO, 30 °C, 450-455 nm, 24 h.	
Entry	Phosphines	Yield (%) ^b
1	Xantphos (0.8 equiv)	56
2	DPEphos (0.8 equiv)	55
3	Rac-BINAP (0.8 equiv)	0
4	dppp (0.8 equiv)	48
5	dppf (0.8 equiv)	0
6	PPh ₃ (1.6 equiv)	46
7	PCy ₃ (1.6 equiv)	23

Table S3. The effect of different phosphines *a,b*

^a Standard conditions: **1a** (0.2 mmol, 1.0 equiv), **2a** (0.3 mmol, 1.5 equiv), Phosphines (x mmol, x equiv), ICH₂CH₂I (0.16 mmol, 0.8 equiv), NHPI (0.3 mmol, 1.5 equiv), DMAP (0.3 mmol, 1.5 equiv), Ph-PTZ (0.01 mmol, 5 mol%), DABCO (0.3 mmol, 1.5 equiv), DMSO (2 mL), 450-455 nm, at 30 °C for 24 h. ^{*b*} Isolated yield.



	Table 54. The effect of unferent photocalaryst	
H U	Antphos (0.8 equiv), ICH ₂ CH ₂ I (0.8 equiv) + NHPI (1.5 equiv), DMAP (1.5 equiv),	
Ph ^N OEt	Photocatalyst (x mol%), DABCO (1.5 equiv) DMSO, 30 °C, 450-455 nm, 24 h.	\bigcirc
1a	2a	3a
Entry	Photocatalyst	Yield (%) ^b
1	Ph-PTZ (5 mol%)	56
2	4CzIPN (5 mol%)	32
3	Eosin Y (5 mol%)	44
4	Rhodamine 6G (5 mol%)	71
5	Methylene Blue (5 mol%)	20
6	Ir[dF(CF ₃)ppy] ₂ (dtbbpy)PF ₆ (2 mol%)	53
7	lr(ppy) ₃ (2 mol%)	54
8	[Ru(bpy) ₃]Cl ₂ (2 mol%)	48

Table S4. The effect of different photocatalyst *a,b*

 a Standard conditions: **1a** (0.2 mmol, 1.0 equiv), **2a** (0.3 mmol, 1.5 equiv), Xantphos (0.16 mmol, 0.8 equiv), ICH_2CH_2I (0.16 mmol, 0.8 equiv), NHPI (0.3 mmol, 1.5 equiv), DMAP (0.3 mmol, 1.5 equiv), Photocatalyst (x mmol, x mol%), DABCO (0.3 mmol, 1.5 equiv), DMSO (2 mL), 450-455 nm, at 30 $^\circ$ C for 24 h. b Isolated yield.









Rhodamine 6G

2+ 2CI

Ph-PTZ







Eosin Y



Methylene blue trihydrate

lr[dF(CF₃)ppy]₂(dtbbpy)PF₆

[Ru(bpy)3]Cl2

Ph ^N OEt +	ОН	Xantphos (0.8 equiv), ICH ₂ CH ₂ I (0.8 equiv) NHPI (1.5 equiv), DMAP (1.5 equiv), Rhodamine B (5 mol%), DABCO (1.5 equiv) DMSO, 30 °C, wave length, 24 h.	Ph ^H OEt
1a	2a		3a
Entry		wave lengths	Yield (%) ^b
1		520~525 nm	55
2		460~465 nm	65
3		450~455 nm	71
4		440~445 nm	62
5		420~425 nm	56
6		390~395 nm	34
7		360~365 nm	32
8		Dark	0

Table S5. The effect of different wave length a,b

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^a Standard conditions: **1a** (0.2 mmol, 1.0 equiv), **2a** (0.3 mmol, 1.5 equiv), Xantphos (0.16 mmol, 0.8 equiv), ICH₂CH₂I (0.16 mmol, 0.8 equiv), NHPI (0.3 mmol, 1.5 equiv), DMAP (0.3 mmol, 1.5 equiv), Rhodamine 6G (0.01 mmol, 5 mol%), DABCO (0.3 mmol, 1.5 equiv), DMSO (2 mL), wave length LED, at 30 °C for 24 h. ^b Isolated yield.

Table S6. The effect of different iodines a,b

H O	+OH _	Xantphos (0.8 equiv), Iodine (x equiv) NHPI (1.5 equiv), DMAP (1.5 equiv),	
Ph ¹ OEt	2a	DABCO (1.5 equiv) DMSO, 30 °C, 450-455 nm, 24 h.	Ja 3a
Entry		lodines	Yield (%) ^b
1	IC	CH ₂ CH ₂ I (0.8 equiv)	72
2		CH ₃ I (1.6 equiv)	0
3		<i>n-</i> Bu₄NI (1.6 equiv)	0
4		Nal (1.6 equiv)	0

^{*a*} Standard conditions: **1a** (0.2 mmol, 1.0 equiv), **2a** (0.3 mmol, 1.5 equiv), Xantphos (0.16 mmol, 0.8 equiv), Iodine (x mmol, x equiv), NHPI (0.3 mmol, 1.5 equiv), DMAP (0.3 mmol, 1.5 equiv), DABCO (0.3 mmol, 1.5 equiv), DMSO (2 mL), 450-455 nm, at 30 °C for 24 h. ^{*b*} Isolated yield.

Table S7. Control experiments *a*,*b*

Ph ^{-N} OEt	+ CH Xantphos (0.8 equiv), ICH ₂ CH ₂ I (0.8 equiv) + OH NHPI (1.5 equiv), DMAP (1.5 equiv), Rhodamine 6G (5 mol%), DABCO (1.5 equiv) DMSO, 30 °C, 450-455 nm, 24 h. 2a	Ph ^H OEt 3a
Entry	Variations from standard coniditions	Yield (%) ^b
1	without Rhodamine 6G	72
2	without Xantphos	0
3	without ICH2CH2I	0
4	without NHPI	0
5	without DMAP	0
6	without DABCO	0
7	Dark	0

^{*a*} Standard conditions: **1a** (0.2 mmol, 1.0 equiv), **2a** (0.3 mmol, 1.5 equiv), Xantphos (0.16 mmol, 0.8 equiv), ICH₂CH₂I (0.16 mmol, 0.8 equiv), NHPI (0.3 mmol, 1.5 equiv), DMAP (0.3 mmol, 1.5 equiv), Rhodamine 6G (0.01 mmol, 5 mol%), DABCO (0.3 mmol, 1.5 equiv), DMSO (2 mL), wave length LED, at 30 °C for 24 h. ^{*b*} Isolated yield.

1.6 Analytical data for compounds



ethyl 2-cyclohexyl-2-(phenylamino)acetate (3a): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and cyclohexanecarboxylic acid (2a, 38 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product 3a as a white solid (37.8 mg, 72% yield): Rf = 0.8 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.18-7.15 (m, 2H), 6.74-6.70 (m, 1H), 6.64-6.62 (m, 2H), 4.20-4.13 (m, 3H), 3.89-3.85 (m, 1H), 1.85-1.67 (m, 6H), 1.27-1.15 (m, 8H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.6, 147.4, 129.2, 118.0, 113.5, 62.0, 60.8, 41.3, 29.6, 29.2, 26.2, 26.1, 26.0, 14.3 ppm; These data are in agreement with literature.^[2]



ethyl 2-cyclohexyl-2-(*o*-tolylamino)acetate (3b): The general procedure was followed using ethyl *o*-tolylglycinate (1b, 39 mg, 0.2 mmol) and cyclohexanecarboxylic acid (2a, 38 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product 3b as a colorless oil (21.6 mg, 39% yield): Rf = 0.7 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.10-7.04 (m, 2H), 6.66 (t, *J* = 7.2 Hz, 1H), 6.56 (d, *J* = 8.0 Hz, 1H), 4.17 (q, *J* = 7.2 Hz, 2H), 4.05-4.03 (m, 1H), 3.92-3.89 (m,1H), 2.21 (s, 3H), 1.91-1.70 (m, 6H), 1.27-1.23 (m, 8H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.8, 145.4, 130.3, 127.0, 122.7, 117.6, 110.4, 61.9, 60.8, 41.4, 29.6, 29.3, 26.2, 26.13, 26.07, 17.5, 14.3 ppm; These data are in agreement with literature.^[7]



ethyl 2-cyclohexyl-2-(*m*-tolylamino)acetate (3c): The general procedure was followed using ethyl *m*-tolylglycinate (1c, 39 mg, 0.2 mmol) and cyclohexanecarboxylic acid (2a, 38 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product 3c as a white solid (38.6 mg, 70% yield): Rf = 0.7 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.05 (t, *J* = 7.6 Hz, 1H), 6.55 (d, *J* = 7.2 Hz, 1H), 6.47-6.43 (m, 2H), 4.21-4.15 (m, 2H), 4.09 (br, 1H), 3.86 (d, *J* = 6.0 Hz, 1H), 2.27 (s, 3H), 1.89-1.66 (m, 6H), 1.29-1.17 (m, 8H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.7, 147.5, 139.0, 129.1, 119.0, 114.4, 110.5, 62.0, 60.7, 41.3, 29.6, 29.2, 26.2, 26.1, 26.0, 21.6, 14.3 ppm; These data are in agreement with literature.^[2]



ethyl 2-cyclohexyl-2-(*p*-tolylamino)acetate (3d): The general procedure was followed using ethyl *p*-tolylglycinate (1d, 39 mg, 0.2 mmol) and cyclohexanecarboxylic acid (2a, 38 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product 3d as a colorless oil (41.3 mg, 75% yield): Rf = 0.7 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 6.98 (d, *J* = 7.6 Hz, 2H), 6.56 (d, *J* = 8.0 Hz, 2H), 4.17 (q, *J* = 7.2 Hz, 2H), 4.01 (br, 1H), 3.83 (d, *J* = 6.4 Hz, 1H), 2.23 (s, 3H), 1.88-1.66 (m, 6H), 1.30-1.14 (m, 8H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.8, 145.2, 129.7, 127.3, 113.7, 62.4, 60.7, 41.3, 29.6, 29.2, 26.2, 26.07, 26.03, 20.3, 14.3 ppm; These data are in agreement with literature.^[2]



ethyl 2-cyclohexyl-2-((4-methoxyphenyl)amino)acetate (3e): The general procedure was followed using ethyl (4-methoxyphenyl)glycinate (1e, 42 mg, 0.2 mmol) and cyclohexanecarboxylic acid (2a, 38 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 20:1) afforded product 3e as a colorless oil (32.6 mg, 56% yield): Rf = 0.5 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 6.77-6.73 (m, 2H), 6.62-6.58 (m, 2H), 4.15 (q, *J* = 7.2 Hz, 2H), 3.87 (br, 1H), 3.77-3.73 (m, 4H), 1.88-1.65 (m, 6H), 1.29-1.14 (m, 8H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 174.0, 152.6, 141.6, 115.2, 114.8, 63.4, 60.7, 55.7, 41.3, 29.6, 29.2, 26.2, 26.1, 26.0, 14.3 ppm; These data are in agreement with literature.^[3]



ethyl 2-((4-chlorophenyl)amino)-2-cyclohexylacetate (3f): The general procedure was followed using ethyl (4-chlorophenyl)glycinate (1f, 43 mg, 0.2 mmol) and cyclohexanecarboxylic acid (2a, 38 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product 3f as a colorless oil (19.9 mg, 34% yield): Rf = 0.6 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.11-7.09 (m, 2H), 6.56-6.52 (m, 2H), 4.19-4.13 (m, 3H), 3.80 (d, *J* = 6.4 Hz, 1H), 1.85-1.66 (m, 6H), 1.26-1.18 (m, 8H);

¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.4, 146.0, 129.1, 122.7, 114.6, 62.2, 60.9, 41.2, 29.7, 29.6, 29.1, 26.1, 26.03, 26.00, 14.3 ppm; These data are in agreement with literature.^[4]



ethyl 2-cyclohexyl-2-(naphthalen-2-ylamino)acetate (3g): The general procedure was followed using ethyl naphthalen-2-ylglycinate (1g, 46 mg, 0.2 mmol) and cyclohexanecarboxylic acid (2a, 38 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product 3g as a colorless oil (32.6 mg, 52% yield): Rf = 0.6 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.68-7.60 (m, 3H), 7.38-7.34 (m, 1H), 7.23-7.19 (m, 1H), 6.94 (dd, *J* = 8.8, 2.8 Hz, 1H), 6.83 (d, *J* = 2.4 Hz, 1H), 4.34 (br, 1H), 4.24-4.15 (m, 2H), 4.03 (d, *J* = 6.0 Hz, 1H), 1.95-1.69 (m, 6H), 1.32-1.20 (m, 8H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.6, 145.0, 135.0, 129.0, 127.8, 127.6, 126.3, 126.0, 122.2, 118.2, 105.5, 62.0, 60.9, 41.2, 29.6, 29.3, 26.2, 26.1, 26.0, 14.3 ppm; These data are in agreement with literature.^[2]



tert-butyl 2-cyclohexyl-2-(phenylamino)acetate (3h): The general procedure was followed using ethyl naphthalen-2-ylglycinate (1h, 58 mg, 0.2 mmol) and cyclohexanecarboxylic acid (2a, 38 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product 3h as a colorless oil (23.2 mg, 40% yield): Rf = 0.6 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.17-7.13 (m, 2H), 6.70 (t, *J* = 7.2, 1H), 6.62 (d, *J* = 8.0, 2H), 4.13-4.12 (m, 1H), 3.77-3.74 (m, 1H), 1.79-1.65 (m, 5H), 1.42 (s, 9H), 1.29-1.21 (m, 5H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 172.8, 147.7, 129.2, 117.8, 113.6, 81.4, 62.4, 41.3, 29.5, 29.2, 28.1, 26.3, 26.2, 26.1 ppm; These data are in agreement with literature.^[3]



N-benzyl-2-cyclohexyl-2-(phenylamino)acetamide (3i): The general procedure was followed using *N*-benzyl-2-(phenylamino)acetamide (1i, 48 mg, 0.2 mmol) and cyclohexanecarboxylic acid (2a, 38 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum

ether : ethyl acetate = 20:1 to 5:1) afforded product **3i** as a white solid (44.5 mg, 69% yield): Rf = 0.5 (petroleum ether : ethyl acetate = 5:1); ¹H NMR (400 MHz, CDCl₃): δ 7.26-7.15 (m, 6H), 7.08-7.05 (m, 1H), 6.80 (t, *J* = 7.2 Hz, 1H), 6.62 (d, *J* = 8.0 Hz, 2H), 4.51 (dd, *J* = 14.8, 6.0 Hz, 1H), 4.39 (dd, *J* = 14.8, 5.6 Hz, 1H), 3.94-3.93 (m, 1H), 3.65-3.63 (m, 1H), 2.07-2.03 (m, 1H), 1.81-1.69 (m, 5H), 1.32-1.21 (m, 5H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 172.6, 147.3, 138.2, 129.3, 128.5, 127.6, 127.3, 119.0, 113.7, 64.9, 43.2, 41.1, 30.3, 28.2, 26.21, 26.18, 26.1 ppm; These data are in agreement with literature.^[1]



2-cyclohexyl-1-morpholino-2-(phenylamino)ethan-1-one (3j): The general procedure was followed using 1-morpholino-2-(phenylamino)ethan-1-one (**1j**, 44 mg, 0.2 mmol) and cyclohexanecarboxylic acid (**2a**, 38 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product **3j** as a white solid (33.4 mg, 55% yield): Rf = 0.8 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (**400 MHz, CDCl₃**): δ 7.17-7.13 (m, 2H), 6.73-6.69 (m, 1H), 6.62 (d, *J* = 8.0 Hz, 2H), 4.46 (br, 1H), 4.09 (d, *J* = 6.0 Hz, 1H), 3.64-3.57 (m, 8H), 1.90-1.66 (m, 6H), 1.24-1.13 (m, 5H); ¹³C{¹H} NMR (**100 MHz, CDCl₃**): δ 171.8, 148.1, 129.3, 118.1, 113.9, 67.0, 66.6, 58.4, 46.3, 42.4, 42.0, 30.3, 29.7, 28.7, 26.2, 26.1 ppm; These data are in agreement with literature.^[1]



3k

methyl (2-cyclohexyl-2-(phenylamino)acetyl)glycinate (3k): The general procedure was followed using methyl phenylglycylglycinate (1k, 44 mg, 0.2 mmol) and cyclohexanecarboxylic acid (2a, 38 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 10:1 to 2:1) afforded product 3k as a yellow oil (34.7 mg, 57% yield): Rf = 0.5 (petroleum ether : ethyl acetate = 2:1); ¹H NMR (400 MHz, CDCl₃): δ 7.21-7.17 (m, 2H), 6.79 (t, *J* = 7.2 Hz, 1H), 6.63 (d, *J* = 8.0 Hz, 2H), 4.17 (dd, *J* = 18.4, 6.4 Hz, 1H), 3.96-3.87 (m, 2H), 3.71 (s, 3H), 3.62 (d, *J* = 4.0 Hz, 1H), 2.07-2.00 (m, 1H), 1.81-1.68 (m, 5H), 1.35-1.13 (m, 6H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.2, 170.1, 147.3, 129.4, 119.0, 113.7, 64.7, 52.2, 41.2, 40.8, 30.2, 28.0, 26.24, 26.20, 26.1 ppm; These data are in agreement with literature.^[4]



methyl (2-cyclohexyl-2-(phenylamino)acetyl)-*L***-phenylalaninate (31):** The general procedure was followed using methyl phenylglycyl-*L*-phenylalaninate (**11**, 62 mg, 0.2 mmol) and cyclohexanecarboxylic acid (**2a**, 38 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 10:1 to 6:1) afforded product **31** as a white solid (41.8 mg, 53% yield, 1:1 d.r.): Rf = 0.7 (petroleum ether : ethyl acetate = 2:1); ¹H NMR (400 MHz, CDCl₃): δ 7.25-7.00 (m, 7H), 6.84-6.75 (m, 2H), 6.61 (d, *J* = 8.0 Hz, 1H), 6.55 (d, *J* = 7.6 Hz, 1H), 5.00-4.97 (m, 0.5H), 4.94-4.89 (m, 0.5H), 3.91-3.82 (m, 1H), 3.70-3.49 (m, 4H), 3.23-2.91 (m, 2H), 1.79-1.65 (m, 4H), 1.32-1.12 (m, 7H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 172.6, 172.4, 171.8, 171.6, 147.4, 147.1, 136.1, 135.3, 129.4, 129.2, 129.14, 129.08, 128.5, 126.94, 126.91, 119.1, 118.9, 114.0, 113.5, 65.1, 64.4, 52.8, 52.2, 52.1, 41.1, 41.0, 38.0, 37.9, 30.2, 30.0, 28.0, 27.9, 26.24, 26.19, 26.16, 26.1, 26.0 ppm; These data are in agreement with literature.^[2]



ethyl 2-(phenylamino)nonanoate (4a): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and octanoic acid (2b, 43 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product 4a as a colorless oil (26.2 mg, 47% yield): Rf = 0.6 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.17 (t, *J* = 7.6 Hz, 2H), 6.73 (t, *J* = 7.6 Hz, 1H), 6.63 (d, *J* = 8.0 Hz, 2H), 4.18 (q, *J* = 7.2 Hz, 2H), 4.07-4.03 (m, 1H), 1.87-1.74 (m, 2H), 1.35-1.23 (m, 13H), 0.90-0.87 (m, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 174.3, 147.0, 129.3, 118.1, 113.4, 60.9, 56.6, 33.1, 31.7, 29.3, 29.1, 25.5, 22.6, 14.2, 14.0 ppm; These data are in agreement with literature.^[3]



ethyl 5-chloro-2-(phenylamino)pentanoate (4b): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and 4-chlorobutanoic acid (2c, 37 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 20:1) afforded product 4b as a colorless oil (25 mg, 49% yield): Rf = 0.5 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.24-7.20 (m, 2H), 6.71 (t, *J* = 7.2 Hz, 1H), 6.56-6.54 (m, 2H), 4.25-4.13 (m, 3H), 3.60-3.55 (m, 1H), 3.46-3.34 (m, 1H), 2.30-2.04 (m, 4H),

1.24 (t, J = 6.8 Hz, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 174.5, 146.7, 129.2, 116.6, 111.9, 60.9, 60.8, 48.2, 30.9, 23.8, 14.2 ppm; These data are in agreement with literature.^[3]



ethyl 2-(phenylamino)-5-(thiophen-2-yl)pentanoate (4c): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and 4-(thiophen-2-yl)butanoic acid (2d, 51 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 20:1) afforded product 4c as a colorless oil (41 mg, 68% yield): Rf = 0.6 (petroleum ether : ethyl acetate = 2:1); ¹H NMR (400 MHz, CDCl₃): δ 7.17 (t, *J* = 8.0 Hz, 2H), 7.12 (d, *J* = 5.2 Hz, 1H), 6.93-6.90 (m, 1H), 6.79 (s, 1H), 6.7 (t, *J* = 7.6 Hz, 1H), 6.62 (d, *J* = 8.0 Hz, 2H), 4.18 (q, *J* = 7.2 Hz, 2H), 4.11-4.08 (m, 2H), 2.96-2.86 (m, 2H), 1.93-1.86 (m, 4H), 1.24 (t, *J* = 7.6 Hz, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.9, 146.8, 144.5, 129.3, 126.7, 124.3, 123.1, 118.3, 113.5, 61.1, 56.4, 32.3, 29.5, 27.6, 14.2 ppm; These data are in agreement with literature. ^[8]



ethyl 2-cyclobutyl-2-(phenylamino)acetate (4d): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and cyclobutanecarboxylic acid (2e, 30 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 20:1) afforded product 4d as a colorless oil (28.4 mg, 61% yield): Rf = 0.5 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.17 (t, *J* = 7.6 Hz, 2H), 6.73 (t, *J* = 7.2 Hz, 1H), 6.63 (d, *J* = 8.0 Hz, 2H), 4.19-4.12 (m, 2H), 4.04 (br, 1H), 3.97 (d, *J* = 8.0 Hz, 1H), 2.75-2.64 (m, 1H), 2.08-1.86 (m, 6H), 1.23 (t, *J* = 7.2 Hz, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.4, 147.3, 129.2, 118.2, 113.5, 61.0, 60.8, 38.3, 25.3, 24.7, 18.0, 14.3 ppm; These data are in agreement with literature. ^[4]



ethyl 2-cyclopentyl-2-(phenylamino)acetate (4e): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and cyclopentanecarboxylic acid (2f, 34 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 20:1) afforded product 4e as a colorless oil (31.7 mg, 64% yield): Rf = 0.5 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.18-7.14 (m, 2H), 6.75-

6.71 (m, 1H), 6.65-6.63 (m, 2H), 4.17 (q, *J* = 7.2 Hz, 2H), 4.12-4.09 (m, 1H), 3.87 (t, *J* = 8.0 Hz, 1H), 2.28-2.20 (m, 1H), 1.86-1.81 (m, 1H), 1.77-1.64 (m, 3H), 1.62-1.56 (m, 2H), 1.52-1.43 (m, 2H), 1.26-1.22 (m, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 174.1, 147.3, 129.2, 118.2, 113.5, 60.84, 60.76, 43.2, 29.3, 29.0, 25.3, 25.1, 14.2 ppm; These data are in agreement with literature. ^[4]



ethyl 2-cycloheptyl-2-(phenylamino)acetate (4f): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and cycloheptanecarboxylic acid (2g, 43 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product 4f as a colorless oil (38 mg, 69% yield): Rf = 0.7 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.18-7.14 (m, 2H), 6.74-6.70 (m, 1H), 6.62 (d, *J* = 8.0 Hz, 2H), 4.20-4.14 (m, 3H), 3.91-3.87 (m, 1H), 1.96 (s, 1H), 1.86-1.68 (m, 4H), 1.60-1.41 (m, 8H), 1.26-1.23 (m, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.7, 147.3, 129.3, 118.1, 113.5, 62.3, 60.8, 42.7, 31.1, 30.0, 28.4, 27.9, 26.7, 26.6, 14.3 ppm; These data are in agreement with literature.^[5]



ethyl 2-(2,3-dihydro-1*H*-inden-2-yl)-2-(phenylamino)acetate (4g): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and 2,3-dihydro-1*H*-indene-2-carboxylic acid (2h, 49 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product 4g as a colorless oil (29.6 mg, 50% yield): Rf = 0.5 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.21-7.16 (m, 6H), 6.76 (t, *J* = 7.2 Hz, 1H), 6.66 (d, *J* = 8.0 Hz, 2H), 4.22-4.09 (m, 4H), 3.14 (dd, *J* = 15.6, 7.6 Hz, 1H), 3.06-2.88 (m, 4H), 1.24 (t, *J* = 7.2 Hz, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.5, 147.0, 142.2, 142.0, 129.3, 126.5, 124.4, 118.4, 113.6, 61.0, 60.2, 42.6, 35.9, 35.6, 14.2 ppm; These data are in agreement with literature.^[5]



4h

ethyl 3-ethyl-2-(phenylamino)heptanoate (4h): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and 2-ethylhexanoic acid (2i, 43 mg, 0.3 mmol).

Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product **4h** as a colorless oil (24.1 mg, 43% yield): Rf = 0.8 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.17 (t, *J* = 7.6 Hz, 2H), 6.73 (t, *J* = 7.2 Hz, 1H), 6.63 (d, *J* = 8.0 Hz, 2H), 4.23-4.13 (m, 2H), 4.08 (s, 2H), 1.80-1.74 (m, 1H), 1.46-1.23 (m, 11H), 0.98-0.88 (m, 6H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 174.0, 147.5, 147.4, 129.3, 118.1, 113.5, 60.8, 58.8, 42.7, 29.5, 29.3, 29.1, 23.1, 22.90, 22.87, 22.5, 14.3, 14.0, 11.6, 11.4 ppm; These data are in agreement with literature.^[3]



ethyl 2-(phenylamino)-2-(tetrahydro-2H-pyran-4-yl)acetate (4i): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and tetrahydro-2*H*-pyran-4-carboxylic acid (2j, 39 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 20:1 to 3:1) afforded product 4i as a colorless oil (40.5 mg, 77% yield): Rf = 0.3 (petroleum ether : ethyl acetate = 5:1); ¹H NMR (400 MHz, CDCl₃): δ 7.17 (t, *J* = 7.6 Hz, 2H), 6.74 (t, *J* = 7.2 Hz, 1H), 6.64 (d, *J* = 8.0 Hz, 2H), 4.21-4.14 (m, 3H), 4.04-3.98 (m, 2H), 3.92-3.90 (m, 1H), 3.42-3.35 (m, 2H), 2.05-1.95 (m, 1H), 1.79-1.74 (m, 1H), 1.60-1.53 (m, 3H), 1.25 (t, *J* = 7.2 Hz, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.0, 147.1, 129.3, 118.4, 113.6, 67.8, 67.5, 61.4, 61.0, 38.7, 29.32, 29.28, 14.3 ppm; These data are in agreement with literature.^[5]



ethyl 2-(phenylamino)-2-(tetrahydrofuran-2-yl)acetate (4j): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and tetrahydrofuran-2-carboxylic acid (2k, 35 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 20:1 to 5:1) afforded product 4j as a colorless oil (22 mg, 44% yield): Rf = 0.3 (petroleum ether : ethyl acetate = 5:1); ¹H NMR (400 MHz, CDCl₃): δ 7.19-7.15 (m, 2H), 6.77-6.69 (m, 2H), 6.64 (d, *J* = 8.0 Hz, 1H), 4.44-4.31 (m, 1H), 4.24-4.19 (m, 2H), 4.13-4.05 (m, 1H), 3.97-3.85 (m, 1H), 3.83-3.75 (m, 1H), 2.04-1.89 (m, 4H), 1.24 (t, *J* = 7.2 Hz, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 172.5, 172.1, 147.4, 146.9, 129.3, 129.2, 118.5, 118.4, 113.9, 113.7, 79.8, 79.3, 69.2, 68.7, 61.3, 61.2, 60.7, 59.8, 28.4, 27.9, 26.0, 25.6, 14.20, 14.16 ppm; These data are in agreement with literature.^[6]



tert-butyl 2-(2-ethoxy-2-oxo-1-(phenylamino)ethyl)pyrrolidine-1-carboxylate (4k): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and (*tert*-butoxycarbonyl)proline (2l, 65 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 20:1 to 10:1) afforded product 4k as a colorless oil (33.3 mg, 48% yield): Rf = 0.5 (petroleum ether : ethyl acetate = 5:1); ¹H NMR (400 MHz, CDCl₃): δ 7.18-7.10 (m, 2H), 6.72-6.63 (m, 3H), 5.36-5.28 (m, 0.31H), 4.75 (br, 0.33H), 4.66 (br, 0.27H), 4.55 (br, 0.52H), 4.40-4.36 (m, 0.44H), 4.32-4.29 (m, 0.37H), 4.25-4.10 (m, 2.76H), 3.52-3.13 (m, 2.09H), 2.01-1.74 (m, 4H), 1.60-1.48 (m, 9H), 1.28-1.24 (m, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 172.7, 172.5, 156.0, 155.0, 154.1, 147.9, 147.1, 129.3, 129.1, 118.0, 117.5, 113.6, 113.1, 112.9, 79.8, 79.6, 61.4, 61.2, 60.3, 59.8, 58.7, 58.0, 57.7, 47.1, 28.7, 28.5, 27.3, 26.6, 24.3, 23.7, 14.2, 14.1 ppm; These data are in agreement with literature.^[3]



ethyl 3,3-dimethyl-2-(phenylamino)butanoate (4l): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and pivalic acid (2m, 31 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 3s0:1) afforded product 4l as a colorless oil (29.7 mg, 63% yield): Rf = 0.7 (petroleum ether : ethyl acetate = 10:1). ¹H NMR (400 MHz, CDCl₃): δ 7.19-7.15 (m, 2H), 6.73 (t, *J* = 7.2 Hz, 1H), 6.67 (d, *J* = 8.0 Hz, 2H), 4.19-4.10 (m, 3H), 3.80-3.78 (m, 1H), 1.24 (t, *J* = 7.2 Hz, 3H), 1.08 (s, 9H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.3, 147.7, 129.2, 118.3, 113.8, 65.5, 60.5, 34.5, 26.8, 14.3 ppm; These data are in agreement with literature.^[3]



ethyl 2-(1-methylcyclohexyl)-2-(phenylamino)acetate (4m): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and 1-methylcyclohexane-1-carboxylic acid (2n, 43 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product 4m as a colorless oil (35.8 mg, 65% yield): Rf = 0.8 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ

7.16 (t, J = 8.0 Hz, 2H), 6.72 (t, J = 7.2 Hz, 1H), 6.67 (d, J = 8.0 Hz, 2H), 4.20-4.09 (m, 3H), 3.95 (s, 1H), 1.66-1.46 (m, 8H), 1.35-1.30 (m, 2H), 1.23 (t, J = 7.2 Hz, 3H), 1.05 (s, 3H); ¹³C{¹H} **NMR (100 MHz, CDCl₃):** δ 173.3, 147.8, 129.3, 118.2, 113.8, 64.5, 60.5, 37.1, 34.91, 34.87, 26.1, 21.8, 21.7, 20.3, 14.3 ppm; These data are in agreement with literature.^[3]



ethyl 2-((3*r*,5*r*,7*r*)-adamantan-1-yl)-2-(phenylamino)acetate (4n): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and (3*r*,5*r*,7*r*)-adamantane-1-carboxylic acid (2o, 54 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product 4n as a colorless oil (40.3 mg, 64% yield): Rf = 0.7 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.16 (t, *J* = 8.0 Hz, 2H), 6.72 (t, *J* = 7.2 Hz, 1H), 6.66 (d, *J* = 8.0 Hz, 2H), 4.19-4.10 (m, 3H), 3.66 (s, 1H), 2.04-2.01 (m, 3H), 1.83-1.57 (m, 12H), 1.24 (t, *J* = 7.2 Hz, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 172.8, 147.9, 129.2, 118.1, 113.8, 66.4, 60.5, 39.0, 36.9, 36.4, 28.4, 14.3 ppm; These data are in agreement with literature.^[3]



ethyl (10*Z*,13*Z*)-2-(phenylamino)nonadeca-10,13-dienoate (40): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and (9*Z*,12*Z*)-octadeca-9,12-dienoic acid (2p, 84 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product 4o as a colorless oil (39.2 mg, 47% yield): Rf = 0.7 (petroleum ether : ethyl acetate = 10:1). ¹H NMR (400 MHz, CDCl₃): δ 7.19-7.15 (m, 2H), 6.73 (t, *J* = 7.6 Hz, 1H), 6.62 (d, *J* = 7.6 Hz, 2H), 5.42-5.30 (m, 4H), 4.18 (q, *J* = 7.2 Hz, 2H), 4.06-4.03 (m, 1H), 2.79-2.76 (m, 2H), 2.08-2.02 (m, 4H), 1.84-1.71 (m, 2H), 1.32-1.23 (m, 19H), 0.91-0.88 (m, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 174.3, 147.0, 130.2, 130.0, 129.3, 128.0, 127.9, 118.2, 113.4, 61.0, 56.7, 33.1, 31.5, 29.7, 29.6, 29.34, 29.31, 29.2, 27.2, 25.6, 25.5, 22.7, 22.6, 14.2, 14.1 ppm. HRMS (ESI-TOF) m/z calcd for C₂₇H₄₄NO₂ (M + H)⁺: 414.3372, found 414.3388.



ethyl 6-(2,5-dimethylphenoxy)-3,3-dimethyl-2-(phenylamino)hexanoate (4p): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and 5-(2,5-dimethylphenoxy)-2,2-dimethylpentanoic acid (2q, 75 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 10:1 to 4:1) afforded product 4p as a colorless oil (52.3 mg, 68% yield): Rf = 0.8 (petroleum ether : ethyl acetate = 2:1). ¹H NMR (400 MHz, CDCl₃): δ 7.26-7.17 (m, 2H), 7.07-6.96 (m, 1H), 6.76 (t, *J* = 7.2 Hz, 1H), 6.71-6.67 (m, 3H), 6.63 (s, 1H), 4.20-4.15 (m, 3H), 3.96-3.92 (m, 3H), 2.33 (s, 3H), 2.19 (s, 3H), 1.97-1.79 (m, 2H), 1.64-1.57 (m, 2H), 1.27 (t, *J* = 7.2 Hz, 3H), 1.11 (d, *J* = 6.0 Hz, 6H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 173.2, 156.9, 147.6, 136.4, 130.3, 129.3, 123.5, 120.7, 118.4, 113.9, 112.0, 68.2, 64.1, 60.6, 36.8, 36.0, 24.1, 24.0, 23.5, 21.4, 15.7, 14.3 ppm. HRMS (ESI-TOF) m/z calcd for C₂₄H₃₂NO₃ (M - H)⁻: 382.2382, found 382.2365.



ethyl(5*R*)-5-((8*R*,9*S*,10*S*,13*R*,14*S*,17*R*)-10,13-dimethyl-3,7,12-trioxohexadecahydro-1*H*-cyclopenta[a]phenanthren-17-yl)-2-(phenylamino)hexanoate (4q): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and (4*R*)-4-((8*R*,9*S*,10*S*,13*R*,14*S*,17*R*)-10,13-dimethyl-3,7,12-trioxohexadecahydro-1*H*-

cyclopenta[*a*]phenanthren-17-yl) pentanoic acid (**2r**, 121 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 10:1 to 3:1) afforded product **4q** as a colorless oil (47.1 mg, 44% yield): Rf = 0.9 (petroleum ether : ethyl acetate = 1:1). ¹**H NMR (400 MHz, CDCl₃):** δ 7.15 (t, *J* = 7.6 Hz, 2H), 6.71 (t, *J* = 7.2 Hz, 1H), 6.60 (d, *J* = 8.0 Hz, 2H), 4.17-4.10 (m, 3H), 4.00 (br, 1H), 2.92-2.79 (m, 3H), 2.35-2.11 (m, 8H), 2.03-2.01 (m, 2H), 1.99 (br, 1H), 1.97-1.92 (m, 2H), 1.86-1.76 (m, 2H), 1.64-1.52 (m, 2H), 1.38 (s, 3H), 1.27-1.21 (m, 7H), 1.05 (d, *J* = 8.0 Hz, 3H), 0.87-0.83 (m, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 211.93, 211.91, 209.0, 208.7, 174.2, 174.1, 146.9, 146.8, 129.2, 118.2, 118.1, 113.42, 113.35, 61.0, 60.9, 60.3, 57.0, 56.8, 56.7, 51.7, 48.9, 46.8, 45.5, 45.4, 44.9, 42.7, 38.6, 36.4, 35.9, 35.7, 35.6, 35.2, 31.0, 30.9, 29.9, 29.8, 27.6, 25.1, 21.8, 18.9, 18.8, 14.2, 14.1, 11.8 ppm. HRMS (ESI-TOF) m/z calcd for C₃₃H₄₆NO₅ (M + H)⁺: 536.3376, found 536.3394.



ethyl(5R)-5-((3R,8R,9S,10S,13R,14S,17R)-3-hydroxy-10,13-dimethylhexadecahydro-1Hcyclopenta[a]phenanthren-17-yl)-2-(phenylamino)hexanoate (4r): The general procedure was followed using ethyl phenylglycinate (1a, 36 mg, 0.2 mmol) and (4R)-4-((3R,8R,9S,10S,13R,14S,17R)-3-hydroxy-10,13-dimethylhexadecahydro-1Hcyclopenta[a]phenanthren-17-yl) pentanoic acid (2s, 113 mg, 0.3 mmol). Purification of this material by chromatography on silica gel (petroleum ether : ethyl acetate = 10:1 to 2:1) afforded product 4r as a colorless oil (43.6 mg, 43% yield): Rf = 0.5 (petroleum ether : ethyl acetate = 2:1). ¹**H NMR (400 MHz, CDCl₃):** δ 7.16 (t, J = 7.6 Hz, 2H), 6.72 (t, J = 7.2 Hz, 1H), 6.62 (d, J = 8.0 Hz, 2H), 4.18 (q, J = 7.2 Hz, 2H), 4.01-3.98 (m, 1H), 3.65-3.58 (m, 1H), 1.97-1.93 (m, 1H), 1.85-1.64 (m, 7H), 1.57-1.48 (m, 3H), 1.40-0.99 (m, 21H), 0.92-0.91 (m, 6H), 0.63 (d, J = 5.6 Hz, 3H); ${}^{13}C{^{1}H}$ NMR (100 MHz, CDCl₃): δ 174.3, 174.2, 147.0, 146.9, 129.2, 118.12, 118.09, 113.43, 113.41, 71.8, 60.92, 60.90, 57.1, 57.0, 56.43, 56.42, 55.8, 42.7, 42.0, 40.4, 40.1, 36.4, 35.8, 35.5, 35.4, 35.3, 34.5, 31.5, 30.5, 29.62, 29.56, 28.14, 28.10, 27.2, 26.4, 24.1, 23.3, 20.8, 18.6, 18.5, 14.2, 12.0 ppm. HRMS (ESI-TOF) m/z calcd for C₃₃H₅₂NO₃ (M + H)⁺: 510.3947, found 510.3959.

1.7 Mechanistic studies

1.7.1 Radical-trapping experiments



Procedure of eq. 1: To an oven-dried 25 mL Schlenk tube was charged with 1a (36 mg, 0.2 mmol, 1 equiv), 2a (38 mg, 0.3 mmol, 1.5 equiv), Xantphos (93 mg, 0.16 mmol, 0.8 equiv), ICH₂CH₂I (45 mg, 0.16 mmol, 0.8 equiv), NHPI (49 mg, 0.3 mmol, 1.5 equiv), DMAP (37 mg, 0.3 mmol, 1.5 equiv), DABCO (34 mg, 0.3 mmol, 1.5 equiv) and TEMPO (94 mg, 0.6 mmol, 3.0 equiv). The tube was then evacuated and back-filled under argon flow (this sequence was repeated three times), anhydrous DMSO (2.0 mL) was added under Ar. The tube was screw capped and heated to 30 °C under irradiation of Blue LEDs (450-455 nm). After stirring for 24 h, the reaction mixture was quenched by water and extracted with EtOAc three times. The combined organic phases were removed under vacuo. The resulting mixture was analyzed by HRMS. TEMPO-adduct 5 and 6 were detected. TEMPO-adduct 5 HRMS (ESI-TOF) m/z calcd for $C_{19}H_{31}N_2O_3$ (M + H)⁺: 335.2335, found 335.2353.



Procedure of eq. 2: To an oven-dried 25 mL Schlenk tube was charged with 1a (36 mg, 0.2 mmol, 1 equiv), 2a (38 mg, 0.3 mmol, 1.5 equiv), Xantphos (93 mg, 0.16 mmol, 0.8 equiv), ICH₂CH₂I (45 mg, 0.16 mmol, 0.8 equiv), NHPI (49 mg, 0.3 mmol, 1.5 equiv), DMAP (37 mg, 0.3 mmol, 1.5 equiv) and DABCO (34 mg, 0.3 mmol, 1.5 equiv). The tube was then evacuated and back-filled under argon flow (this sequence was repeated three times), anhydrous DMSO (2.0 mL) and ethene-1,1-diyldibenzene (108 mg, 0.6 mmol, 3 equiv) were added in turn under Ar. The tube was screw capped and heated to 30 °C under irradiation of Blue LEDs (450-455 nm). After stirring for 24 h, the reaction mixture was quenched by water and extracted with EtOAc three times. The combined organic phases were removed under vacuo. 1,3,5-trimethoxybenzene (16.8 mg, 0.1 mmol) was added as an internal standard before the mixturewas transferred to an NMR tube and diluted with CDCl₃. ¹H NMR analysis showed the yield of 7 was 19%. These data are in agreement with literature.^[3] The resulting mixture was analyzed by HRMS. 7, 8 and 9 were detected. 7 HRMS (ESI-TOF) m/z calcd for C₂₀H₂₃ (M + H)⁺: 263.1800, found 263.1829. 9 HRMS (ESI-TOF) m/z calcd for C₂₀H₂₃ (M + H)⁺: 358.1807, found 358.1836.



¹H NMR spectra (400 MHz, CDCl₃) of 7

1.7.2 Radical clock experiments



Procedure of eq. 3: In an oven-dried 25 mL Schlenk tube was charged with 1a (36 mg, 0.2 mmol, 1.0 equiv), 2t (30 mg, 0.3 mmol, 1.5 equiv), Xantphos (93 mg, 0.16 mmol, 0.8 equiv), ICH₂CH₂I (45 mg, 0.16 mmol, 0.8 equiv), NHPI (49 mg, 0.3 mmol, 1.5 equiv), DMAP (37 mg, 0.3 mmol, 1.5 equiv) and DABCO (34 mg, 0.3 mmol, 1.5 equiv). The tube was then evacuated and back-filled under argon flow (this sequence was repeated three times), anhydrous DMSO (2.0 mL) was added under Ar. The tube was screw capped and heated to 30 °C under irradiation of Blue LEDs (450-455 nm). After stirring for 24 h, the reaction mixture was quenched by water and extracted with EtOAc three times. The combined organic phases was removed under vacuo. The residue was purified by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 10:1) afforded product 10 as a colorless oil (30.6 mg, 66% yield): Rf = 0.6 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.19-7.15 (m, 2H), 6.76-6.72 (m, 1H), 6.64-6.62 (m, 2H), 5.88-5.78 (m, 1H), 5.10-5.01 (m, 2H), 4.19 (q, *J* = 7.2 Hz, 2H), 4.08 (t, *J* = 6.4 Hz, 1H), 2.24-2.18 (m, 2H), 1.98-1.80 (m, 2H), 1.27-1.23 (m, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 174.0, 146.9, 137.2, 129.3, 118.3, 115.7, 113.5, 61.0, 56.1, 32.2, 29.7, 14.2 ppm. These data are in agreement with literature.^[4]



 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of **10**



Procedure of eq. 4: In an oven-dried 25 mL Schlenk tube was charged with 1a (36 mg, 0.2 mmol, 1.0 equiv), 2u (38 mg, 0.3 mmol, 1.5 equiv), Xantphos (93 mg, 0.16 mmol, 0.8 equiv), ICH₂CH₂I (45 mg, 0.16 mmol, 0.8 equiv), NHPI (49 mg, 0.3 mmol, 1.5 equiv), DMAP (37 mg, 0.3 mmol, 1.5 equiv) and DABCO (34 mg, 0.3 mmol, 1.5 equiv). The tube was then evacuated and back-filled under argon flow (this sequence was repeated three times), anhydrous DMSO (2.0 mL) was added under Ar. The tube was screw capped and heated to 30 °C under irradiation of Blue LEDs (450-455 nm). After stirring for 24 h, the reaction mixture was quenched by water and extracted with EtOAc three times. The combined organic phases was removed under vacuo. The residue was purified by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 10:1) afforded product 11 as a colorless oil (28.7 mg, 55% yield): Rf = 0.6 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.17 (t, *J* = 7.6 Hz, 2H), 6.73 (t, *J* = 7.2 Hz, 1H), 6.63 (d, *J* = 8.0 Hz, 2H), 4.21-4.13 (m, 2H), 4.06-4.04 (m, 1H), 1.99-1.74 (m, 5H), 1.67-1.48 (m, 4H), 1.27-1.22 (m, 5H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 174.6, 147.0, 129.3, 118.2, 113.4, 60.9, 56.4, 39.6, 36.7, 32.8, 32.6, 25.1, 24.9, 14.2 ppm. These data are in agreement with literature.^[4]







1.7.3 Control experiments



Procedure of eq. 5: In an oven-dried 25 mL Schlenk tube was charged with 1a (36 mg, 0.2 mmol, 1.0 equiv), 2a (38 mg, 0.3 mmol, 1.5 equiv), Xantphos (93 mg, 0.16 mmol, 0.8 equiv), ICH₂CH₂I (45 mg, 0.16 mmol, 0.8 equiv), NHPI (49 mg, 0.3 mmol, 1.5 equiv), DMAP (37 mg, 0.3 mmol, 1.5 equiv) and DABCO (34 mg, 0.3 mmol, 1.5 equiv). The tube was then evacuated and backfilled under argon flow (this sequence was repeated three times), anhydrous DMSO (2.0 mL) was added under Ar. The tube was screw capped and heated to 30 °C. After stirring for 24 h, the reaction mixture was quenched by water and extracted with EtOAc three times. The combined organic phases was removed under vacuo. The residue was purified by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 10:1) afforded product **3a** as a white solid (37.8 mg, 72% yield): Rf = 0.8 (petroleum ether : ethyl acetate = 10:1) and 12 as a white solid (5.7 mg, 8%) yield): Rf = 0.2 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.18-7.14 (m, 4H), 6.74-6.70 (m, 2H), 6.63 (d, J = 8.0 Hz, 4H), 4.20-4.14 (m, 4H), 3.88-3.85 (m, 2H), 1.26-1.25 (m, 6H); These data are in agreement with literature.^[7] HRMS (ESI-TOF) m/z calcd for $C_{20}H_{25}N_2O_4 (M + H)^+$: 357.1814, found 357.1826.



¹H NMR spectra (400 MHz, CDCl₃) of **12**



Procedure of eq. 6: In an oven-dried 25 mL Schlenk tube was charged with 1a (36 mg, 0.2 mmol, 1.0 equiv), 2a (38 mg, 0.3 mmol, 1.5 equiv), Xantphos (93 mg, 0.16 mmol, 0.8 equiv), ICH₂CH₂I (45 mg, 0.16 mmol, 0.8 equiv), NHPI (49 mg, 0.3 mmol, 1.5 equiv), DMAP (37 mg, 0.3 mmol, 1.5 equiv) and DABCO (34 mg, 0.3 mmol, 1.5 equiv). The tube was then evacuated and back-filled under argon flow (this sequence was repeated three times), anhydrous DMSO (2.0 mL) was added under Ar. The tube was screw capped and heated to 30 °C in dark. After stirring for 24 h, the reaction mixture was quenched by water and extracted with EtOAc three times. The combined organic phases was removed under vacuo. The residue was purified by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 10:1) afforded product 13 as a white solid (27.3 mg, 50% yield): Rf = 0.4 (petroleum ether : ethyl acetate = 10:1); ¹H NMR (400 MHz, CDCl₃): δ 7.89-7.85 (m, 2H), 7.80-7.75 (m, 2H), 2.77-2.69 (m, 1H), 2.12-2.06 (m, 2H), 1.86-1.80 (m, 2H), 1.70-1.60 (m, 3H), 1.42-1.27 (m, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 171.8, 162.1, 134.6, 129.0, 123.9, 40.4, 28.8, 25.4, 25.0 ppm. These data are in agreement with literature.^[10]







Procedure of eq. 7: In an oven-dried 25 mL Schlenk tube was charged with 1a (36 mg, 0.2 mmol, 1.0 equiv), 13 (82 mg, 0.3 mmol, 1.5 equiv), Xantphos (93 mg, 0.16 mmol, 0.8 equiv), ICH₂CH₂I (45 mg, 0.16 mmol, 0.8 equiv), NHPI (49 mg, 0.3 mmol, 1.5 equiv), DMAP (37 mg, 0.3 mmol, 1.5 equiv) and DABCO (34 mg, 0.3 mmol, 1.5 equiv). The tube was then evacuated and back-filled under argon flow (this sequence was repeated three times), anhydrous DMSO (2.0 mL) was added under Ar. The tube was screw capped and heated to 30 °C. After stirring for 24 h, the reaction mixture was quenched by water and extracted with EtOAc three times. The combined organic phases was removed under vacuo. The residue was purified by chromatography on silica gel (petroleum ether : ethyl acetate = 40:1 to 30:1) afforded product **3a** as a white solid (28.8 mg, 55% yield): Rf = 0.8 (petroleum ether : ethyl acetate = 10:1).

1.7.4 Kinetic monitoring experiment

Ph ^N OE	standar conditio	d ⊵ns F		OEt +						
1a		2a				3a		N	HPI este	r 13
Time/yield %	0 h	0.5h	1h	2h	4h	8h	12h	16h	20h	24h
3 a	0	5	15	22	37	42	52	56	66	73
NHPI ester 13	0	26	29	25	8	0	0	0	0	0

In an oven-dried 25 mL Schlenk tube was charged with **1a** (36 mg, 0.2 mmol, 1.0 equiv), **2a** (38 mg, 0.3 mmol, 1.5 equiv), Xantphos (93 mg, 0.16 mmol, 0.8 equiv), ICH₂CH₂I (45 mg, 0.16 mmol, 0.8 equiv), NHPI (49 mg, 0.3 mmol, 1.5 equiv), DMAP (37 mg, 0.3 mmol, 1.5 equiv) and DABCO (34 mg, 0.3 mmol, 1.5 equiv). The tube was then evacuated and back-filled under argon flow (this sequence was repeated three times), anhydrous DMSO (2.0 mL) was added under Ar. The tube was screw capped and heated to 30 °C under irradiation of Blue LEDs (450-455 nm). 100 μ L of reaction mixture was taken by a syringe at indicated time (0.5 h, 1 h, 2 h, 4 h, 8 h, 12 h, 16 h, 20h, 24h), the resulting mixture was quenched by water and extracted with EtOAc three times. The combined organic phases was removed under vacuo. ¹H NMR of the crude mixture using 1,3,5-trimethylbenzene as internal standard (Figure S1).



Figure S1. Kinetic monitoring experiment

1.7.5 UV-Vis absorption spectra

UV-Vis absorption spectra of NHPI ester **13**, **1a**, ICH₂CH₂I, Xantphos, a mixture of Xantphos and ICH₂CH₂I, a mixture of **1a**, ICH₂CH₂I and Xantphos, a mixture of NHPI ester **13**, ICH₂CH₂I and Xantphos, a mixture of NHPI ester **13**, ICH₂CH₂I and Xantphos, a mixture of NHPI ester **13**, ICH₂CH₂I and Xantphos, a mixture of NHPI ester **13**, ICH₂CH₂I and Xantphos, a mixture of NHPI ester **13**, ICH₂CH₂I and Xantphos were provided respectively. The UV-vis absorption spectra of DMSO solutions of NHPI ester **13** (1×10⁻³ M), **1a** (1×10⁻³ M), Xantphos (1×10⁻⁴ M), ICH₂CH₂I (1×10⁻⁴ M), a mixture of Xantphos (1×10⁻⁴ M) and ICH₂CH₂I (1×10⁻⁴ M), a mixture of **1a** (1×10⁻³ M), ICH₂CH₂I (1×10⁻⁴ M) and ICH₂CH₂I (1×10⁻⁴ M), a mixture of NHPI ester **13** (1×10⁻³ M), Xantphos (1×10⁻⁴ M) and ICH₂CH₂I (1×10⁻⁴ M), a mixture of NHPI ester **13** (1×10⁻³ M), **1a** (1×10⁻³ M), Xantphos (1×10⁻⁴ M) and ICH₂CH₂I (1×10⁻⁴ M), a mixture of NHPI ester **13** (1×10⁻³ M), **1a** (1×10⁻³ M), Xantphos (1×10⁻⁴ M) and ICH₂CH₂I (1×10⁻⁴ M) and ICH₂CH₂I (1×10⁻⁴ M) and ICH₂CH₂I (1×10⁻⁴ M) and ICH₂CH₂I (1×10⁻⁴ M) in DMSO, visible red shift was observed in UV/vis absorption spectrum. This indicates the formation of an electron donor-acceptor (EDA) complex between the NHPI ester **13**, Xantphos and ICH₂CH₂I.



Figure S2. UV-Vis studies about the EDA complex

1.8 References

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2. NMR spectra of products







¹³C{¹H} NMR spectra (100 MHz, CDCl₃) of 3b







 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of 3d



 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of 3e







 $^{13}\mathrm{C}\{^{1}\mathrm{H}\}$ NMR spectra (100 MHz, CDCl_3) of 3g



 $^{13}\mathrm{C}\{^{1}\mathrm{H}\}$ NMR spectra (100 MHz, CDCl₃) of 3h



¹³C{¹H} NMR spectra (100 MHz, CDCl₃) of **3i**











 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of **3**l



¹³C{¹H} NMR spectra (100 MHz, CDCl₃) of 4a



 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of 4b



¹³C{¹H} NMR spectra (100 MHz, CDCl₃) of **4c**



 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of 4d



¹³C{¹H} NMR spectra (100 MHz, CDCl₃) of **4e**



¹³C{¹H} NMR spectra (100 MHz, CDCl₃) of **4f**



 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of 4g



 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of **4h**



 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of **4i**











 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of 4k







 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of **4m**







 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of 4o















 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of 4q





 $^{13}C\{^{1}H\}$ NMR spectra (100 MHz, CDCl₃) of 4r